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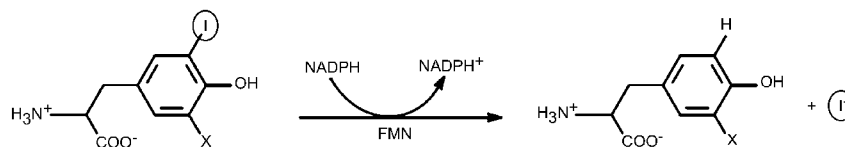
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(54) Title: COMPOSITIONS AND METHODS FOR CONTROLLING INSECT PROLIFERATION

FIG. 1



X = H (monoiodotyrosine, MIT), or I (diiodotyrosine, DIT)

(57) Abstract: The present invention relates to methods of controlling pest populations through inhibition of spermatogenesis.

COMPOSITIONS AND METHODS FOR CONTROLLING INSECT PROLIFERATION

RELATED APPLICATIONS

This application claims the benefit of priority under 35 U.S.C. § 119(e) to U.S. Provisional Application No. 62/029,863, filed on July 28, 2014, which is hereby incorporated herein by reference in its entirety.

STATEMENT AS TO FEDERALLY SPONSORED RESEARCH

This invention was made with government support under RO1 DK084186 awarded by the National Institute of Health (NIH). The government has certain rights in the invention.

BACKGROUND OF THE INVENTION

Many strategies exist for blocking insect reproduction. However, there is a pressing need to develop an approach to selectively control insect populations without damage to the environment.

SUMMARY OF THE INVENTION

The invention provides a method of preventing, reducing, or inhibiting insect reproduction comprising administering to the insect a composition comprising an agent that inhibits spermatogenesis (e.g., late stage spermatogenesis) in the insect, thereby preventing, reducing, or inhibiting reproduction in the insect. Preferably, the agent that inhibits spermatogenesis in the insect comprises an agent that inhibits expression or activity of iodotyrosine deiodinase (IYD) in the insect (i.e., in the male insect). For example, the agent inhibits expression or activity of IYD in a testis of the insect.

The methods described herein reduce a population size of the insect, i.e., the methods described herein reduce/control insect proliferation. For example, the methods described herein reduce an insect population, i.e., the methods reduce/decrease the number of insects in a defined environment by at least 1%, e.g., at least 5%, at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, or at least 99%.

The methods described herein also reduce fecundity and/or fertility of the insect. For example, the methods described herein reduce/decrease fecundity and/or fertility of the insect

by at least 1%, e.g., at least 5%, at least 10%, at least 15%, at least 20%, at least 25%, at least 30%, at least 35%, at least 40%, at least 45%, at least 50%, at least 55%, at least 60%, at least 65%, at least 70%, at least 75%, at least 80%, at least 85%, at least 90%, at least 95%, at least 99% or by 100%.

In some cases, the agent for inhibiting expression or activity of IYD comprises an antibody or fragment thereof, a binding protein, a polypeptide, or any combination thereof.

In other cases, the agent for inhibiting expression or activity of IYD comprises a small molecule. A small molecule is a compound that is less than 2000 Daltons in mass. The molecular mass of the small molecule is preferably less than 1000 Daltons, more preferably less than 600 Daltons, e.g., the compound is less than 500 Daltons, less than 400 Daltons, less than 300 Daltons, less than 200 Daltons, or less than 100 Daltons.

Small molecules are organic or inorganic. Exemplary organic small molecules include, but are not limited to, aliphatic hydrocarbons, alcohols, aldehydes, ketones, organic acids, esters, mono- and disaccharides, aromatic hydrocarbons, amino acids, and lipids. Exemplary inorganic small molecules comprise trace minerals, ions, free radicals, and metabolites. Alternatively, small molecules can be synthetically engineered to consist of a fragment, or small portion, or a longer amino acid chain to fill a binding pocket of an enzyme. Typically small molecules are less than one kilodalton.

Suitable small molecule inhibitors include 4-OH-2',3,4',5,6'-PCB; 4-OH-2',3,4',6'-TCB; 4-OH-2,2',5,5'-TCB; 4,4'-diOH-3,3',5,5'-TCB; 3-OH-2,2',5,5'-TCB; 4'-OH-BDE-17; 4-OH-BDE-42; 4'-OH-BDE-49; 4-OH-BDE90; 2-OH-BDE-15; 2'-OH-BDE-28; Bromoxynil; Oxyaclozanide; Tribromsalan; Closantel; Triclosan; Bithionol; Nitroxylin; Benzbromarone Rose Bengal; Erythrosine B; Phloxine B; D,L-3-(2-pyridon-5-yl)alanine; and D,L-3-(N-methyl-2-pyridon-5-yl)alanine; D,L-3-(N-ethyl-2-pyridon-5-yl)alanine; and D,L-3-(N-isopropyl-2-pyridon-5-yl)alanine. In a preferred aspect, the small molecule inhibitor is L-3-(N-methyl-2-pyridon-5-yl)alanine.

In other aspects, the agent comprises a nucleic acid molecule. For example, the nucleic acid molecule comprises double stranded ribonucleic acid (dsRNA), small hairpin RNA or short hairpin RNA (shRNA), or antisense RNA, or any portion thereof.

A variety of administration routes are available. For example, the agent is administered topically, orally, via inhalation, or via injection.

The composition is in the form of a gas, a liquid, a solid, or a semi-solid. In a preferred aspect, the composition comprises sugar water. In another example, the agent is

administered as a gaseous or liquid spray. In some cases, the agent is soluble in water. Preferably, the agent is nontoxic.

The effective amount of the agent is from 0.001 mg/kg to 250 mg/kg body weight, e.g., 0.001 mg/kg, 0.05 mg/kg, 0.1 mg/kg, 0.5 mg/kg, 1 mg/kg, 5 mg/kg, 10 mg/kg, 25 mg/kg, 50 mg/kg, 75 mg/kg, 100 mg/kg, 125 mg/kg, 150 mg/kg, 175 mg/kg, 200 mg/kg, 225 mg/kg, or 250 mg/kg body weight.

In some cases, the agent is administered at least once per day, at least once per week, or at least once per month. The agent is administered for a duration of one day, one week, one month, two months, three months, six months, 9 months, or one year. For example, the compositions described herein are placed in an insect “trap” as food for ingestion by the insects. The “traps” are replenished with fresh composition as needed, e.g., at least once per day, at least once per week, at least once per month, or at least once every three months.

The compositions and methods described herein are useful in inhibiting reproduction in any insect. For example, suitable insects include cockroaches (e.g., brown-banded cockroaches, German cockroaches, American cockroaches, Oriental cockroaches), beetles (woodworms, death watch beetles, fur beetles, varied carpet beetles, spider beetles, mealworm beetles, centipedes, red flower beetle, pine beetle), earwigs, flies (e.g., bottle flies, blue bottle flies, green bottle flies, house flies, fruit flies, horse flies, black flies, deer flies, house flies, tsetse flies), cicadas, hoppers, ants (e.g., Argentine ants, carpenter ants, fire ants, odorous house ants, pavement ants, pharaoh ants, thief ants, a leafcutter ant, a mountain fire ant, a Jerdon’s jumping ant), aphids, bed bugs, bees (e.g., a common eastern bumble bee, a bumble bee, a honeybee, an alfalfa leafcutter bee), wasps, mantids, earthworms, terrestrial crabs, firebrats, fleas, snails, slugs, crickets (e.g., house crickets), hornets, locusts, lice, moths (e.g., almond moths, Indianmeal moths, clothes moths, gypsy moths (common clothes moths), brown house moths), red spiders, silverfish, woollice, termites (e.g., dampwood termites, subterranean termites), ticks, wasps, mosquitos, gnats, millipedes, stink bugs, weevils, mites, silk worm, daphnia, fleas, Assassin bugs (kissing bugs), eye gnats, or rat fleas and woolly bears (e.g., hairy caterpillar).

The compositions and methods are also useful in inhibiting reproduction in the following insects: *Culex quinquefasciatus* (Southern house mosquito), *Drosophila willistoni* (Fruitfly), *Drosophila mojavensis* (Fruitfly), *Drosophila pseudoobscura* (Fruitfly), *Drosophila grimshawi* (Fruitfly), *Drosophila virilis* (Fruitfly), *Drosophila ananassae* (Fruitfly), *Ceratitis capitata* (Mediterranean fruitfly), *Drosophila sechellia* (Fruitfly),

Drosophila persimilis (Fruitfly), *Drosophila simulans* (Fruitfly), *Drosophila melanogaster* (Fruitfly), *Anopheles darling* (Mosquito), *Anopheles gambiae* (Mosquito), *Musca domestica* (Housfly), *Drosophila erecta* (Fruitfly), *Drosophila yakuba* (Fruitfly), *Aedes aegypti* (Mosquito, yellow fever), *Bombyx mori* (Silk worm), *Tribolium castaneum* (red flour beetle), *Nasonia vitripennis* (wasp), *Apis mellifera* (honeybee), *Apis dorsata* (honeybee), *Megachile rotundata* (alfalfa leafcutter bee), *Dendroctonus ponderosae* (mountain pine beetle), *Bombus impatiens* (common eastern bumble bee), *Bombus terrestris* (bumble bee), *Acromyrmex echinator* (leafcutter ants), *Solenopsis invicta* (fireant), *Apis florea* (honeybee), *Harpegnathos saltator* (Jerdon's jumping ant), *Cerapachys biroi* (ant), *Daphnia pulex* (daphnia), *Camponotus floridanus* (ant), and *Pediculus humanus corporis* (lice).

In some cases, the methods described herein are useful for inhibiting the reproduction of any invertebrate, such as a crab, a lobster, a snail, a clam, an octopus, a starfish, a sea-urchin or a worm.

In one aspect, the compositions are administered within a device, e.g., a “trap,” that attracts an insect to the device in order to administer the compositions described herein to inhibit spermatogenesis within the insect.

Definitions

By “agent” is meant any small molecule compound, antibody, nucleic acid molecule, or polypeptide, or fragments thereof.

By “alteration” is meant a change (increase or decrease) in the expression levels or activity of a gene or polypeptide as detected by standard art known methods such as those described herein. As used herein, an alteration includes a 10% change in expression levels, preferably a 25% change, more preferably a 40% change, and most preferably a 50% or greater change in expression levels.

“Controlling pests” as used in the present invention means killing pests, or preventing pests from developing, or from growing or preventing pests from infecting or infesting. Controlling pests as used herein also encompasses controlling insect progeny (development of eggs). Controlling pests as used herein also encompasses inhibiting viability, growth, development or reproduction of the insect, or decreasing pathogenicity or infectivity of the insect.

“Detect” refers to identifying the presence, absence, or amount of the nucleic acid (e.g., RNA) to be detected.

By “detectable label” is meant a composition that when linked to a molecule of interest renders the latter detectable, via, for example, spectroscopic, photochemical, biochemical, immunochemical, or chemical means. For example, useful labels may include radioactive isotopes, magnetic beads, metallic beads, colloidal particles, fluorescent dyes, electron-dense reagents, enzymes (for example, as commonly used in an ELISA), biotin, digoxigenin, or haptens.

By “effective amount” is meant the amount of an agent required to inhibit reproduction in an insect.

By “fragment” is meant a portion of a polypeptide or nucleic acid molecule. This portion contains, preferably, at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, or 90% of the entire length of the reference nucleic acid molecule or polypeptide. A fragment may contain 10, 20, 30, 40, 50, 60, 70, 80, 90, 100, 200, 300, 400, 500, 600, 700, 800, 900, or 1000 nucleotides or amino acids.

“Hybridization” means hydrogen bonding, which may be Watson-Crick, Hoogsteen or reversed Hoogsteen hydrogen bonding, between complementary nucleobases. For example, adenine and thymine are complementary nucleobases that pair through the formation of hydrogen bonds.

By “inhibitory nucleic acid” is meant a double-stranded RNA, siRNA, shRNA, or antisense RNA, or a portion thereof, or a mimetic thereof, that when administered to a mammalian cell results in a decrease (e.g., by 10%, 25%, 50%, 75%, or even 90-100%) in the expression of a target gene. Chitosan compositions are useful for the delivery of polynucleotides, such as inhibitory nucleic acid molecules. Typically, a nucleic acid inhibitor comprises at least a portion of a target nucleic acid molecule, or an ortholog thereof, or comprises at least a portion of the complementary strand of a target nucleic acid molecule. For example, an inhibitory nucleic acid molecule comprises at least a portion of any or all of the nucleic acids delineated herein.

By “isolated polynucleotide” is meant a nucleic acid (e.g., RNA, DNA, cDNA, etc.) that is free of the genes which, in the naturally-occurring genome of the organism from which the nucleic acid molecule of the invention is derived, flank the gene. The term therefore includes, for example, a recombinant DNA that is incorporated into a vector; into an autonomously replicating plasmid or virus; or into the genomic DNA of a prokaryote or eukaryote; or that exists as a separate molecule (for example, a cDNA or a genomic or cDNA fragment produced by PCR or restriction endonuclease digestion) independent of other

sequences. In addition, the term includes an RNA molecule that is transcribed from a DNA molecule, as well as a recombinant DNA that is part of a hybrid gene encoding additional polypeptide sequence.

By “ K_d ” is meant dissociation constant, a parameter of a drug that characterizes the specificity of interaction between a drug and its target.

By “ k_{cat} ” is meant the turnover number for an enzyme; the number of substrate molecules each enzyme site converts to product per unit time.

By “ K_m ” is meant Michaelis–Menten constant, which represents the substrate concentration at which an enzymatic reaction rate is half of V_{max} .

By “marker” is meant any protein or polynucleotide having an alteration in expression level or activity that is associated with a disease or disorder.

By “modulate” is meant alter (increase or decrease). Such alterations are detected by standard art known methods such as those described herein.

Nucleic acid molecules useful in the methods of the invention include any nucleic acid molecule that encodes a polypeptide of the invention or a fragment thereof. Such nucleic acid molecules need not be 100% identical with an endogenous nucleic acid sequence, but will typically exhibit substantial identity. Polynucleotides having “substantial identity” to an endogenous sequence are typically capable of hybridizing with at least one strand of a double-stranded nucleic acid molecule. Nucleic acid molecules useful in the methods of the invention include any nucleic acid molecule that encodes a polypeptide of the invention or a fragment thereof. Such nucleic acid molecules need not be 100% identical with an endogenous nucleic acid sequence, but will typically exhibit substantial identity. Polynucleotides having “substantial identity” to an endogenous sequence are typically capable of hybridizing with at least one strand of a double-stranded nucleic acid molecule. By “hybridize” is meant pair to form a double-stranded molecule between complementary polynucleotide sequences (e.g., a gene described herein), or portions thereof, under various conditions of stringency. (See, e.g., Wahl, G. M. and S. L. Berger (1987) *Methods Enzymol.* 152:399; Kimmel, A. R. (1987) *Methods Enzymol.* 152:507).

As used herein, “obtaining” as in “obtaining an agent” includes synthesizing, purchasing, or otherwise acquiring the agent.

As used herein, the terms “prevent,” “preventing,” and “prevention,” refer to reducing or inhibiting reproduction in an insect.

“Primer set” means a set of oligonucleotides that may be used, for example, for PCR. A primer set would consist of at least 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 30, 40, 50, 60, 80, 100, 200, 250, 300, 400, 500, 600, or more primers.

Ranges provided herein are understood to be shorthand for all of the values within the range. For example, a range of 1 to 50 is understood to include any number, combination of numbers, or sub-range from the group consisting 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, or 50 as well as all intervening decimal values between the aforementioned integers such as, for example, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, and 1.9. With respect to sub-ranges, “nested sub-ranges” that extend from either end point of the range are specifically contemplated. For example, a nested sub-range of an exemplary range of 1 to 50 may comprise 1 to 10, 1 to 20, 1 to 30, and 1 to 40 in one direction, or 50 to 40, 50 to 30, 50 to 20, and 50 to 10 in the other direction.

By “reduces” is meant a negative alteration of at least 10%, 25%, 50%, 75%, or 100%.

By “reference” is meant a standard or control condition.

A “reference sequence” is a defined sequence used as a basis for sequence comparison or a gene expression comparison. A reference sequence may be a subset of or the entirety of a specified sequence; for example, a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence. For polypeptides, the length of the reference polypeptide sequence will generally be at least about 16 amino acids, preferably at least about 20 amino acids, more preferably at least about 25 amino acids, and even more preferably about 35 amino acids, about 50 amino acids, or about 100 amino acids. For nucleic acids, the length of the reference nucleic acid sequence will generally be at least about 40 nucleotides, preferably at least about 60 nucleotides, more preferably at least about 75 nucleotides, and even more preferably about 100 nucleotides or about 300 or about 500 nucleotides or any integer thereabout or there between.

By “substantially identical” is meant a polypeptide or nucleic acid molecule exhibiting at least 50% identity to a reference amino acid sequence (for example, any one of the amino acid sequences described herein) or nucleic acid sequence (for example, any one of the nucleic acid sequences described herein). Preferably, such a sequence is at least 60%, more preferably 80% or 85%, and more preferably 90%, 95% or even 99% identical at the amino acid level or nucleic acid to the sequence used for comparison.

Sequence identity is typically measured using sequence analysis software (for example, Sequence Analysis Software Package of the Genetics Computer Group, University of Wisconsin Biotechnology Center, 1710 University Avenue, Madison, Wis. 53705, BLAST, BESTFIT, GAP, or PILEUP/PRETTYBOX programs). Such software matches identical or similar sequences by assigning degrees of homology to various substitutions, deletions, and/or other modifications. Conservative substitutions typically include substitutions within the following groups: glycine, alanine; valine, isoleucine, leucine; aspartic acid, glutamic acid, asparagine, glutamine; serine, threonine; lysine, arginine; and phenylalanine, tyrosine. In an exemplary approach to determining the degree of identity, a BLAST program may be used, with a probability score between e^{-3} and e^{-100} indicating a closely related sequence.

In some cases, by “subject” is meant a mammal, including, but not limited to, a human or non-human mammal, such as a bovine, equine, canine, ovine, or feline. Alternatively, by “subject” is meant an insect.

Unless specifically stated or obvious from context, as used herein, the term “or” is understood to be inclusive. Unless specifically stated or obvious from context, as used herein, the terms “a”, “an”, and “the” are understood to be singular or plural.

Unless specifically stated or obvious from context, as used herein, the term “about” is understood as within a range of normal tolerance in the art, for example within 2 standard deviations of the mean. About can be understood as within 10%, 9%, 8%, 7%, 6%, 5%, 4%, 3%, 2%, 1%, 0.5%, 0.1%, 0.05%, or 0.01% of the stated value. Unless otherwise clear from context, all numerical values provided herein are modified by the term about.

The transitional term “comprising,” which is synonymous with “including,” “containing,” or “characterized by,” is inclusive or open-ended and does not exclude additional, unrecited elements or method steps. By contrast, the transitional phrase “consisting of” excludes any element, step, or ingredient not specified in the claim. The transitional phrase “consisting essentially of” limits the scope of a claim to the specified materials or steps “and those that do not materially affect the basic and novel characteristic(s)” of the claimed invention.

The recitation of a listing of chemical groups in any definition of a variable herein includes definitions of that variable as any single group or combination of listed groups. The recitation of an embodiment for a variable or aspect herein includes that embodiment as any single embodiment or in combination with any other embodiments or portions thereof.

Any compositions or methods provided herein can be combined with one or more of any of the other compositions and methods provided herein.

Other features and advantages of the invention will be apparent from the following description of the preferred embodiments thereof, and from the claims. Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All published foreign patents and patent applications cited herein are incorporated herein by reference. Genbank and NCBI submissions indicated by accession number cited herein are incorporated herein by reference. All other published references, documents, manuscripts and scientific literature cited herein are incorporated herein by reference. In the case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a schematic showing the reductive deiodination of mono- and diiodotyrosine catalyzed by IYD (iodotyrosine deiodinase) and its bound cofactor, flavin mononucleotide (FMN and in its reduced form, FMNH₂).

Figure 2A and Figure 2B are schematics showing the co-crystal structure of mouse IYD-MIT (monoiodotyrosine). Figure 2A is a molecular model of the α_2 dimer of mmIYD-MIT (mouse iodotyrosine deiodinase). The monomers are depicted in green and purple cartoon representation. The FMN (Flavin mononucleotide) and MIT molecules are shown in stick representation and colored according to atom. Figure 2B is a molecular model of the α_2 dimer of mmIYD-MIT showing the residues involved in substrate recognition, Y157, E153 and K178. T235 provides an important hydrogen bond to the N5 of the FMN.

Figure 3 is a multiple sequence alignment showing that IYD homologs are present in diverse organisms. Determinants of IYD are outlined in red, residues interacting with FMN are marked with (+), residues interacting with substrate are marked with (*), residues marked in red are fully conserved, lid-forming sequences are boxed, and secondary structural elements derive from the structure of mmIYD-MIT. Numbering of the amino acids for each protein is indicated on the left and right of the alignment. Secondary structure elements are derived from the crystal structure of mmIYD bound to DIT (diiodotyrosine).

Figure 4 is a diagram depicting the phylogenetic analysis of catalytic domains. All proteins containing the signature residues (E153, Y157 and K178) promote deiodination of DIT. The closest structural analog (Blub) lacks these residues, and does not support deiodination of DIT (diiodotyrosine). The branch lengths are proportional to the number of amino acid substitutions per aligned residue.

Figure 5 is a graph showing the expression of IYD mRNA in *Drosophila*. Tissue specific mRNA expression profiling for *Drosophila melanogaster* (fly) indicated very high levels of IYD (gene CG6279 in flybase) in the adult male testes. Adult *Drosophila* are shown in red bars, and larva *Drosophila* are shown in blue bars.

Figure 6A and Figure 6B is a diagram and a bar graph, respectively, showing the RNAi knockdown of IYD in *Drosophila* testis. Figure 6A is a diagram showing the promoter dependent knockdown of IYD mRNA. Figure 6B is a bar graph showing the fertility and fecundity of *eya-GAL4*, *UAS-dmIYD-RNAi* and *eya-CAL1/UAS-dmIYD-RNAi*.

Figure 7A, Figure 7B, and Figure 7C is a diagram, a bar graph, and a photomicrograph, respectively, showing that late stage development of *Drosophila* is sensitive to IYD. Figure 7A is a diagram showing the stages of spermatogenesis in *Drosophila*. Figure 7B is a bar graph showing the abnormalities detected in the testis after RNAi degradation of IYD mRNA. Figure 7C is an image showing the phase contrast of *Drosophila* testes. Suppression of *dmIYD* disrupts late stage spermatogenesis.

Figure 8 is a bar graph showing the male germline cell specific expression profile of CG6279. Germline and somatic stem cells were located in niche, GB in gonioblast, S4 through S16 were spermatogonia (pre-differentiation), EC16 onwards were spermatocytes post-differentiation.

Figures 9A, Figure 9B, Figure 9C, and Figure 9D are photomicrographs showing testes of flies treated with digoxigenin-labeled riboprobes. Figure 9A is an image showing CG6279 isoform A specific antisense riboprobe. Figure 9B is an image showing CG6279 isoform A+B specific antisense riboprobe. Figure 9C is an image showing the control where isoform A specific sense riboprobes were used. Figure 9D is an image showing the control where isoform A + B specific sense riboprobes were used. (*) indicates early stages of spermatogenesis. (**) indicates later stages of spermatogenesis with strong staining for hybridization of riboprobes with CG6279 mRNA.

Figure 10 is a diagram showing the HPLC method schematic (left to right) for separation, detection and quantification of tyrosine produced from MIT. All gradients were

linear over the indicated time. A drop in the B mobile phase from 60% to 30% over 2 mins after the analytical time prevented impurities from affecting the internal standard peak by delaying their elution. The column was washed with 95% B and equilibrated with A prior to next injection of analyte.

DETAILED DESCRIPTION OF THE INVENTION

As described in detail below, described herein are compositions and methods that are useful for inhibiting spermatogenesis in insects, e.g., drosophila, by decreasing the expression or activity of iodotyrosine deiodinase (IYD). A putative IYD from *Drosophila* (dmIYD) was expressed and confirmed to catalyze deiodination. Its k_{cat}/K_m value was similar to those of all other homologues tested. Expression of dmIYD was primarily limited to the testis and its suppression by RNAi significantly disrupted spermatogenesis, particularly within the late stages of sperm development. The enzyme was critical for spermatogenesis in insects, but not mammals. Fertility and fecundity were greatly reduced when expression of this enzyme was suppressed, and hence, proliferation of insects is selectively reduced by inhibiting this enzyme. The molecular basis was consistent with iodotyrosine acting as a cellular signal and representing a progenitor of thyroxine. Prior to the invention described herein, a biological role had not been previously identified for this enzyme.

Iodotyrosine (IYD) deiodinase

IYD has been traditionally associated with iodide conservation by generating iodide (I⁻) from mono- and diiodotyrosine. IYD facilitates iodide salvage in thyroid tissue in vertebrates by catalyzing deiodination of mono- (I-Tyr) and diiodotyrosine (DIT, I₂-Tyr), that are formed as halogenated byproducts of thyroid hormone biosynthesis (often in surprisingly large quantities) (Goswami, A.; Rosenberg, I. N., *Endocrinology* 1977, 101, 331-341, Gnidehou, S. et al., *FASEB J.* 2004, 18, 1574-1576 and Rokita, S. E. et al., *Biochimie* 2010, 92, 1227-1235). Prior to the invention described herein, IYD had remained poorly characterized for more than 50 years despite its significant role in intrathyroidal iodine metabolism. Patients with deficient IYD suffer from goiter, enlarged thyroid, and other symptoms associated with hypothyroidism because iodotyrosine cannot be directly reutilized for thyroid hormone synthesis. The dehalogenation reaction catalyzed by IYD is unusual for aerobic organisms since the carbon-iodine bond is broken through a reductive process. More commonly, dehalogenation is accomplished by hydrolytic or oxidative pathways. Many of

the bacterial enzymes responsible for dehalogenation have emerged from existing enzyme superfamilies.

IYD has been associated with iodide homeostasis in humans since its mutation, first discovered over 50 years ago, was found to limit thyroid function. More recently, sequence homology and structural data identified this enzyme as the only known representative of the nitro-FMN reductase structural superfamily in humans. Members of this superfamily primarily derive from bacteria and their activities range from generating and chaperoning dihydroflavin, FADH₂ (Flavin Reductase Protein, FRP), to reducing nitroaromatic compounds (NOX) and degrading flavin for vitamin B₁₂ biosynthesis (BluB). All IYDs are indicated by four key residues, a threonine (Thr) for hydrogen bonding to the N5 position of FMN, a glutamic acid (Glu), tyrosine (Tyr) and lysine (Lys) for coordinating to the substrate iodotyrosine.

Functional IYDs have been confirmed in Chordata, Arthropoda, Cnidaria, Eubacteria and Archaeabacteria. Chordata require the tetraiodinated hormone, thyroxine, and rely on IYD to salvage iodide from mono- and diiodotyrosine formed as byproducts of its biosynthesis. Thyroxine is not produced by invertebrates and thus IYD is unlikely to provide iodide for its synthesis. Bacteria may use IYD in a catabolic role since this enzyme is capable of dehalogenating chloro-, bromo- and iodotyrosines. As described herein, the function of IYD in Arthropoda and Cnidaria is examined in the model Arthropoda, *Drosophila melanogaster*.

Drosophila IYD

As described in detail below, this *Drosophila* gene acts as an iodotyrosine deiodinase and is critical for sperm formation in insect testes. Prior to the invention described herein, no evidence in the literature had made a connection between the catalytic activity of this enzyme and spermatogenesis. As described herein, the deiodinase gene was suppressed, and a loss in *Drosophila* fertility was observed. This organism is a well-studied model for insects.

Two isoforms have been reported for *Drosophila* IYD (dmIYD) in the NCBI databases. These two are related by alternative splicing of the CG6279 gene. Isoform B resembles IYD orthologs from other organisms and includes a membrane domain, an intermediate domain and a catalytic domain (Phatarphekar, A. et al., *Mol. Biosys.* 2014, 10, 86-92 and Friedman, J. E. et al. *J. Biol. Chem.* 2006, 281, 2812-2819). Isoform A is unique to the *Drosophila* genus by the added presence of a large N-terminal domain (470 amino

acids) that does not coincide with any known protein domains listed in the conserved domain database (CDD).

The amino acid sequence of *Drosophila melanogaster* IYD (A Isoform) is shown below:

```

1 mepkhrhcyf krsassclvt pmhlhssris sshsqsaggi kpalasldpp ktpnsiektk
61 ksggflsflf gkrdrarnq setngnhisl edhdvrklir lysksvselr scleelrtlf
121 ereytiiera rrkcgeimsp sirryneflf lvphlnihfd kdldpasgtsp laagkllqnp
181 ianpemlcqk ltnftdfpla gfpgnlltkr pkkdpgdrpn kkikpipdqe nakqenpieq
241 eiqaekvad hevsveekis gledgeqkda lspseqkniq nskdktshhh snckmsshek
301 dlvtstnltcp ckpevessqs pvscpwiaen skceppcret ppqstrnsvs spqnsdises
361 skepahlmek epsggesans qqvqmlcdmi rtelttapdf iffdakwrii eilrevhkrq
421 lvhqkaipln ntrrpippqk rkpgepnqml knpkcmsdrs ahekggqgrit mdvdeliss
481 kllkhwpslf itlaliwivk rlffkgnrvl ktynldeqve eevehfadlg delqpaledk
541 phvpfvpqgn lnpngakrly elmrgrsir sfnsphkpdsl sviedciraa gtapsgahte
601 pwtycvvpqep elkrssireiv egeelvnysq rmhpqvwtdl rplqtnhvke ylteapylll
661 ifkqtyglse ngkrmrhyh neistsiaag illcalqaag laslvttpln cgpalrnlbg
721 rpvneklil lpvgypkdgc tvpdlarknl snimvtf

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(SEQ ID NO: 1) GenBank Accession No: NP_648433 (Version 1), incorporated

herein by reference.

The nucleic acid sequence of *Drosophila melanogaster* IYD (A Isoform) is shown

below:

```

1 aaaaaacttc acagtgcacac ttacattcac cccaaaaaaa gtaaaaaaaa aatagtcagg
61 attacaaaag ttatatattcc agataatggt acaaaatattt ttcgatttat aatgctgttg
121 ccattaaaga acccaaagac cgttgaaagc gaggtgaacc cgaatgtgac gctactgaag
181 cgccagatcg ccgaggatgg aaccaaaca caggcactgt tacttcaagc gttccgcttc
241 gagttgccta gtgacgcaa tgcattcca ctctccagg atcagttcca gccacagtca
301 gtcagcagga ggcattaagc ccgctttggc atcactagat ccacctaaaa cacccaatag
361 catcgagaaa acaaagaaat ccgggggatt tctctctttt ttgtttggca aacgagacag
421 gcgggagagg aatcagagtg agacaaatgg aaaccatata tcattggagg atcatgatgt
481 caggaagctt atacgtctct atagcaaaag cgtctcggaa ctgagatcct gtctggagga
541 gctaaggacg ctctttgagc gggaatacac cattatcgaa cgtgctcgaa ggaagtgtgg
601 agagataatg agtccttcaa ttcgactacta taatgaattt cttttcctgg tgccacatct
661 taatatacac ttcgacaagg atcttctctgc cagtggaaact agcccactcg ccgccgaaa
721 acttttgcaa aatccgatag ccaaccaga gatgttgtgc caaaagttga ccaatttcac
781 ggattttcct ctggccggtt ttcccgcaa tctactaacc aagaggccca aaaaagacc
841 ccaagaccgg ccaaacaaga agatcaaac aattcctgat caagaaaatg cgaagcaaga
901 aaatcccata gagcaggaaa ttccaggagg agaaaaagtc gctgatcatg aagtttcggt
961 agaagagaaa atcagcgggt tggaggatca ggaacagaaa gacgcactct ctccatcgga
1021 gcagaagaac atacaaaaca gtaaggataa aaccagtcac catcattcca attgcaagat
1081 gtccagccat gagaaggatt tagtatccac caatttgacc tgtccctgca agccggaagt
1141 ggaatcctca caaagcccag ttctctgccc ctggatagcc gaaaattcaa agtgtgaacc
1201 cccgtgtcgg gaaacaccac cacaatctac gcgaaattcc gtatccagtc cgcaaaattc
1261 agatatttct gaatccagca aagaaccggc tcaattgatg gagaaggaaac catccggtca
1321 ggaatccgag aatagccagc aagtgcagat gctttgcatg atgatacgta ccgagctaac
1381 cactgctccc gattttatat tcttcgatgc caagtggcgg atcatcgaga ttctgagaga
1441 ggtgcacaag cgccaattgg ttaccagaa ggccattcca ctcaacaata cacgccgacc
1501 gattccgccg caaaaacgaa aaccgggtga gcccaatcag atgcttaaga atccaaaatg
1561 catgtcggat aggagtcccc acgaaaaggg acaacagagg atcaccatgg atgtggacga
1621 actgatcagc agtccaagc tactcaagca ttggccaagt ttatttatca ctctggcgct

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1681 gatatggatc gtaaaaagat tgttttttaa agggaatcgc gtattaaaaa catataattt
1741 agatgaacaa gtcgaggagg aggttgaaca ctttgccgat cttggcgatg aactgcaacc
1801 ggctctggag gataagcccc atgtgccctt cgttcccgcg cagaatctca accctaattg
1861 agccaaacgc ttatatgaac tcatgcgtgg acgcgtagc attcgctcct ttaatagcca
1921 tcccaaaccg gatctcagcg tgatagagga ctgcattcgg gcggcgaggaa cagctccag
1981 tggcgcacac acagaaccct ggacgtactg cgtcgtagc gagccggaac tcaagcgatc
2041 cattcgggaa attgtggaac aggaggagct ggtcaactac agccagagga tgcattccga
2101 gtgggtcacc gatctgcgac ccctgcaaac caatcatgtc aaggagtatc tcaccgaggc
2161 gccgtacctt atactgatct tcaagcagac ctatggattg tcggagaatg gaaagcgaat
2221 gaggaggcat tactacaacg agatatccac ttcgattgcg gccggtatcc ttttgtgtgc
2281 ccttcaagcg gcaggactcg catcgctggg caccactcct ttgaactgcy gtccagcctt
2341 gagaaacctg ctcgggcgac ccgtcaatga aaaattactc atcctactgc ccgttggtta
2401 tccgaaagat ggatgcacag ttcccgactt ggcgagaaag aatttgtcaa atattatggt
2461 tactttttta ggtttattta cgtccgtgca gcttacgaaa tagttcaata aaaccaaact
2521 aaaatttatg gt

```

(SEQ ID NO: 2) GenBank Accession No: NM_140176 (Version 2), incorporated

herein by reference.

The amino acid sequence of *Drosophila melanogaster* IYD (B Isoform) is shown

below:

```

1 mdvdelisss kllkhwpslf itlaliwivk rlffkgnrvl ktynldeqve
eevehfadlg
61 delqpaledk phvpfvpqqn lnpngakrly elmrgrsir sfnshpkpdl sviedciraa
121 gtapsgahte pwtycvvqep elksireiv eqeelnysq rmhpqvwtdl rplqtnhvke
181 ylteapylil ifkqtyglse ngkrmrhy neistsiaag illcalqaag laslvttpln
241 cgpalrnllg rpvneklil lpgypkdg tvpdlarknl snimvtf

```

(SEQ ID NO: 3) GenBank Accession No: NP_001163414 (Version 1), incorporated

herein by reference.

The nucleic acid sequence of *Drosophila melanogaster* IYD (B Isoform) is shown

below:

```

1 atcgctatca gcaatagccg tgaccactt aaccaagtga taagccaatc cgagcgcaaa
61 agcaaagcca ctttctctcc gtcgcaaaag cagttacatc ttgaagcacc atggatgtgg
121 acgaactgat cagcagctcc aagctactca agcattggcc aagtttatct atcactctgg
181 cgctgatatg gatcgtaaaa agattgtttt ttaaaggga tgcggtatta aaaacatata
241 atttagatga acaagtcgag gaggaggttg aacactttgc cgatcttggc gatgaactgc
301 aaccggctct ggaggataag cccatgtgc cttcgttcc cggccagaat ctcaacccta
361 atggagccaa acgcttatat gaactcatgc gtggacgccg tagcattcgc tcctttaata
421 gccatcccaa accggatctc agcgtgatag aggactgcat tcgggcggcg ggaacagctc
481 ccagtggcgc acacacagaa ccctggacgt actgcgtcgt acaggagccg gaactcaagc
541 gatccattcg ggaaattgtg gaacaggagg agctgggtcaa ctacagccag aggatgcac
601 cgagtggtgt caccgatctg cgaccctgc aaaccaatca tgtcaaggag tatctcaccg
661 aggcgccgta cttatactg atcttcaagc agacctatgg attgtcggag aatggaaagc
721 gaatgaggag gcattactac aacgagatat ccacttcgat tgcggccggg atccttttgt
781 gtgcccttca agcggcagga ctgcgcatgc tggtcaccac tcctttgaac tgcgggtccg
841 ccttgagaaa cctgctcggg cgaccctgca atgaaaaatt actcatccta ctgccggttg
901 gttatccgaa agatggatgc acagttcccg acttggcgag aaagaatttg tcaaatatta
961 tggttacttt ttaaggttta ttacgtccg tgcagcttac gaaatagttc aataaaacca
1021 aactaaaatt tatggt

```

(SEQ ID NO: 4) GenBank Accession No: NM_001169943 (Version 1), incorporated herein by reference.

Human IYD

Iodotyrosine deiodinase (IYD) is a membrane bound enzyme in humans that is primarily expressed in thyroid tissue to recover iodide from mono- and diiodotyrosine (MIT, DIT) generated during thyroxine biosynthesis. This enzyme is highly unusual for its (i) ability to promote reductive dehalogenation and (ii) reliance on flavin to catalyze this process (Figure 1). *In vivo*, NADPH appears to act as the electron donor for this reaction but IYD is not directly reduced by NADPH. *In vitro*, dithionite serves as an alternative electron donor. IYD also effects debromination and dechlorination of the corresponding halotyrosines. A review on the enzyme iodotyrosine deiodinase focusing on its role in mammals is found in Rokita, S.E. et al., Biochimie 2010, 92, 1227-1235, incorporated herein by reference. A review on the rare occurrence of reductive dehalogenases in aerobic organisms is found in Rokita, S.E., Handbook in Flavoproteins, vol. 1 DeGruyter Berlin, 2013. P. 337-350, incorporated herein by reference.

The amino acid sequence of Human IYD is shown below:

```

1 myfltpilva ilcilvvwif knadrsmekk kgeprtraea rpwvdedlkd
ssdlhqaeed
61 adewqeseen vehipfshnh ypekemvkrs qefyellnkr rsvrfisneq vpmevidnvi
121 rtagtapsga htepwtfvvv kdpdvkhhkir kiieeeeein ymkrmghrwv tdlkklrtnw
181 ikeyldtapi lilifkqvvhg faangkkkvh yyneisvsia cgillaalqn aglvtvtttp
241 lncgprlrsl lgrpahekll mllpvgypsk eatvpdlkrk pldqimvtv

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(SEQ ID NO: 5) GenBank Accession No: NP_981932 (Version 1), incorporated herein by reference.

The nucleic acid sequence of Human IYD is shown below:

```

1 gaagagttct gcactttggt ccttcctgg cacacctgtg tctgcattcc
ttctatctcc
61 cggcattctc cactcctgtc tctgtgtggt taaaaaccgg tgtgggaagt gtgcacgcct
121 gtgacgtcag actccagacc atgtatttcc tgactcccat cttggtagcc attctctgca
181 ttttggttgt gtggatcttt aaaaatgccg acagaagcat ggagaaaaag aagggggagc
241 ctagaaccag ggccgaagct cgcccctggg tggatgaaga cttaaaagac agcagtgacc
301 tgcaccaagc agaagaagat gctgatgaat ggcaagaatc agaagaaaat gttgaacaca
361 tccccttctc tcataaccac tatcctgaga aggaaatggt taagaggtct caggaatttt
421 atgaacttct caataagaga cggtcagtca gggttcataag taatgagcaa gtcccaatgg
481 aagtcattga taatgtcatc agaacggcag gaacagcccc gagtggggct cacacagagc
541 cctggacctt cgtgggtgtg aaggaccag acgtgaagca caagattcga aagatcattg
601 aggaggaaag ggagatcaac tacatgaaaa ggatgggaca tcgctgggtc acagacctca
661 agaaactgag aaccaactgg attaaagagt acttggtatc tgcccctatt ttgattctca
721 ttttcaaaca agtacatggt ttcgccgcaa atggcaagaa aaaagtccac tactacaatg
781 agatcagtggt ttccatcgct tgtggcatcc tgctagctgc cctgcagaat gcaggtctgg
841 tgactgtcac taccactcct ctcaactgtg gccctcgact gaggggtgctc ctgggccgcc
901 ccgcacatga aaagctgctg atgctgctcc ccgtggggta cccagcaag gagggccacg

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961  tgcttgacct  caagcgcaaa  cctctggacc  agatcatggt  gacagtgtag  gcagggcccc
1021 ccaagggagt  ggcagggaga  tggcgccct  gcttttccct  gagcctctcg  cctgctcctc
1081 ttgggtctct  tgggtgctct  ttctccaggt  gtcaggtccc  ct cattgtct  ttctcaggtg
1141 gccacactat  gtcaagaagc  ctctccacac  tctgtggcac  ttccagtcct  ataaatcctg
1201 tttcttatcc  actttgaaa  tgcataaaca  ctttacaaag  aacatgcccg  ggtttttaca
1261 ttttaaaagt  tattctagac  aatcactatt  ggcttttttc  ttttattttt  aaaaaactca
1321 catagaggag  acaatcagaa  atttaccata  gtcccaagaa  tt cagctaca  tgatgactcg
1381 aatttaaaatt  tagattaatc  aaatggtctc  ctgttctctg  atttctggtg  gcttttagac
1441 actaattttt  gagaactact  tttttttttt  taccaaactt  cagggacact  ctgctagttt
1501 tgaataagta  accatcaaag  ttactaaaac  atggccaggc  gcagtggctc  atgctgttaa
1561 tcccagcact  ttgggaggcc  gaggtgggca  gattgcctga  gctcaagagt  tcgagaccag
1621 cctggccaac  atggtgaaac  cccatctcta  ctaaaaatac  aaaaaattag  ctgggcatgc
1681 acttgtaatt  ccagctactc  aggaggtctg  ggcaagagaa  ctgcttgaat  ccgggaggtg
1741 gagattgcag  tgagtcaaga  tgctgagatg  ccaccactgc  actccagcct  gggcaacaga
1801 gcgagaacct  gtctcaaaaa  aaacaaaaac  aaagttaact  aaatgtcacc  ttcacagaac
1861 aggacaggg  acccttgggt  cgacagggcc  tggctggcat  gtaaacggct  agttgactcg
1921 cctagtgggt  tggtagatct  gacttgtaag  cagttcatag  aagctgctca  atactcaaga
1981 atcttgcaag  acagttatta  aacattcata  gcttgaaatt  gcccatggtg  ggagagttta
2041 catcacaacc  actacaaatc  agcacttgct  ttcttcttct  taaagagagc  tgcttaccat
2101 acatagcacc  cacagtgtct  cccttctctt  ctacccaac  ccctgaaaca  cactgacccc
2161 tattctccag  atgacagcac  agctgtttgg  gactgatagg  gtcacagttg  ccaggttggg
2221 aatgctagat  gtagtcttca  cccatcacct  tctgtcgtcc  tcccaggggt  gtcataattt
2281 cagaggcaaa  ctcttacagc  ttggatgatt  tctcatcaag  caactctact  cacctctttt
2341 tcccccttcc  ctgtgcgttt  cttcttcttg  tctcattttt  ccttgagaaa  actctctcta
2401 taaaaaattt  ccatagagta  tgtggctgat  aaacaacaga  aatttgtttc  tcacagttct
2461 ggaggtgga  agtccaaggt  cggggtgcc  tcatggtcag  gtgctggtga  ggacctgat
2521 ctgggttgca  gactgctgac  ttcttattgt  gtctcgtgg  tagaaagagg  gttagagcct
2581 ctggcatgcc  ttttataagg  gtactaatcc  catcatgagg  cccgctcctt  atgacctcat
2641 cacctcccaa  aggacccct  cctaatacca  tcacattggg  ggtaggggtt  tcaacatatg
2701 aattgcgggg  gacacaaaca  ttccgtccat  ggcacctact  tcataggggt  tggcatgctg
2761 ctcggtacat  ggttaacatt  cataaatggt  gggggccctt  actgtggtag  aaaccattgt
2821 cactgcaagg  cagccctgct  tgggattcag  gtgaacaaat  gtaacgtggg  tgggtgaggg
2881 actccctcct  cttcagagaa  gccctcctgt  ggctcagggg  cagctgcagg  gccagatgga
2941 gttggcctgc  agggagttag  ccttacccca  tcagagacca  acttgaaaga  ttttttctc
3001 tgtgtagttc  gtgttatcaa  tctgatctgc  agccagggtt  tttacttctg  aagcattgga
3061 gttatcgatt  gccattgccc  catggttctc  actctttgca  ggcactcttc  aaatttttat
3121 tttaaaaatt  ataatgcaaa  catctggcca  tcaccttaac  caagtgacaa  agtcctagca
3181 caccattaat  ggtaggataa  ctgacattaa  gtgttcctgc  tgggatgaat  tatgaagtat
3241 tccttccaaa  gatgtcaatg  gaaactaatc  gagtcttaca  cctaattttc  agtttacagt
3301 aaatacagag  gataaagaag  ttaatggcat  ggtggaataa  ccaaaatgtg  tatcattcta
3361 caaaaaata  ggctggact  atttaaggag  tgagtgtcac  tgtgggaaga  ataagagggg
3421 gactgtcttg  tccttgctac  tcaaagagtg  gtctggggcc  tgtatcatca  ttgggaccac
3481 ctgggcgctt  gttagaatca  aagactctca  ggcctcacct  agacatccag  aatcaaaatc
3541 tgcattttaa  caagatgccc  aggtgatctg  catgcctgtt  aaagtctgag  ctctgttgca
3601 gataaccaga  gaccaacaaa  ccaaaggcaa  agcgtaagcc  ttgattggat  tttggttag
3661 gaaaaatctc  ataggtataa  aagacaactg  ggaatgttga  atgtggatct  ggtggtaggt
3721 gatattgtta  gtgatgtgag  tgatgcctgc  tgaagtactg  agagatgaag  catcatattg
3781 acaacttact  ccaggaaaaa  aggtgtgtgt  ataggtgtgt  gtttgagag  tgtgagcgag
3841 aactaacatg  gtaaaatgtt  aggtggagca  tatacagttc  tcattatact  cttattttta
3901 atttatattt  gaacacagtc  aaatctgaat  ggaaatacat  ttaaaataagc  atttctttca
3961 gtcataataa  taatttgctc  aaaaatgccc  tctaagtcca  tcattaggtt  aagcgattta
4021 ctgaaggtgc  agatatcatg  cttatttcat  tgggttgttt  tggaatgtcc  cccacttgga
4081 gagtttgagg  aatggaaatg  agaatcggag  gagaaataag  gccagaggct  cacattgctt
4141 gcagggaacc  gactgagtaa  tgaagaagag  gaaggaagcc  cttggaagga  acgtaaaatc

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4201 catttcagcc tgcgcgccct tggaatggag ggtcttctag ggaccagaaa caccagacac
 4261 ttcttctgcc ccaagcacac cccacactaa tttctgctgc agcatcacga tcaggggcaaa
 4321 tcagcacagc cccccagcca tcagtgggtg ctagaaaagt aaggataatt ttcttccttc
 4381 tcaactgcagc cccaagctct tattaacctt cttttatgtg ttcatataat gcagtcagtg
 4441 gacaactgtg ttcaaagtta gaaatagtg tggggaggc ttatagatcc tcttgctgtt
 4501 ctgaggcctt aaaacaaaaa aaataatgga atgattgcat cactgagagg aagtatactt
 4561 aattaagaaa gttgaaaatg gtttgtgtct tactaatagg actcacctga agactctact
 4621 ttgccaagag ggaattttta actcaaagtt gtgtcagcca gcacggggta gagccatggt
 4681 cctcactactt gctggcatcc ctcatctggt ggtctgttaa agcacagatt gctgggccgc
 4741 accctcaggg tgtgtgattg tgtgattcag tattgctggg tgggtgcccc atgctccagt
 4801 tctaattgtat tcccaggtgt tgctgatgct ggtgattggg taccacattt tgagaacct
 4861 ggggcatggt caatcaacaa atgcaatgct tgtgacttga cgtgataaaa gttcatttat
 4921 tgggcaggca aagtcctctg caggcctgga aacctccga gacaattact tgcattgggt
 4981 aaccagtaa tactgtggga tgcaacttct agggcatggc tgctccttc ctgaggcag
 5041 cagggggaga aaacactgga agaggtcacc ccaacaattc catcgtaact tggcctagga
 5101 gtgacctctt cagtctcaca accctttggc tagaactagt catgtgacct cacaggaggt
 5161 ggagcagtg cgtcaatggt atcatcccc tgcacctgca aggaaggagg cccagatgtg
 5221 tgtaggtgct ggatgtctct tccccaaaac atataactct ggatttggaa aacaaaactg
 5281 gatataacag aaaagaaaag gaggggagaa ggaaagtga agaaattggt gaaatcagaa
 5341 taggacagtg aaaggcataa agattttctt ctgaaacgct aagatcggtt gatatttgtc
 5401 taatacagct ttaaataagag tatcactttg ggccaaagaa aattgatata aacatacata
 5461 aagtggact aagattgact tcttctctaga catctctgcc tctgtccaca aaatggaaac
 5521 ttccatcaag gactactgga ggacagttca gtatttcttc ttatttgcct ctcctatga
 5581 aaggaagagt gacttctctg ctgcatgata ttgggggtaa atagttattg cacatttgct
 5641 caaaccagg caaagtgcac cccattttta gccaaattta caaacttaca aagatgatcc
 5701 accaatagtt ttctctattt actttattct tgcctgtgca cccaggctag aatgcagtg
 5761 tgtgattata gctcactgca gcctccaact tctggactca agtgatcctc ccaccttagc
 5821 ttcccaggta gctgggatac aggtgcacgc caccacaccc aggtagttt ttctaaatat
 5881 tggagtccc aaagagtatt caaactgatg tgatatttta aaagacattt tgtaagcatc
 5941 ttttaggaat atcagtaagt ggaagaaaaa cacatagtta tcagtggata ttatgccaga
 6001 gaggcagtg aatatataat ttattttgct aggtatcagc agagtgcct cttataattt
 6061 gtgtgaaatg gaaacaaagg taacatcgtg ttttcaattc acacataat acaagtaatg
 6121 agaggtccag aagaaatgtg gcttcagctc tgctgctact gtgcctccct tctcctgcc
 6181 cactcagccc acaaaatagg ctggacactc aaaaaacgtt gcgtttatct accttttaga
 6241 gaggtgaat agcagagaac tggaggtggg aatggtaagg aactcccagc aggttagtg
 6301 agggatggg ctgacgcac taaggctgat gccaggctct ctccctatct ggggtggcctc
 6361 agcaaatgac gtccagcaca tccaggggca ggctcaaggg agaacagccc ccaaagctaa
 6421 gatcctgcca agctaaatac agtagttcta atgaaatgtg agaggctata atccatttg
 6481 ggaaattcct aaaaagtcag gaggcagggg attggtttat gttattatca tgacctgaga
 6541 gtcattggctc agagccaaat gttcaggatt gaattcaaca gcatttaaat gcttttagag
 6601 caggatggaa atatgttagc aatgcctgca gagtgcgaag taaacgcaaa agccaatgag
 6661 atcataaagg aagttgttag ctaacctagg tggagtcgcc aacttccttc tactctaata
 6721 attaaaaata aaaataatac ttgggaggta actggaataa aggttctaaa atcaaaacc
 6781 tctgaagggt gaaaactggg agcctcctgt tcccatagta accacagcac tcagggcact
 6841 gtctcccagc gctggagtac tgtcttatga ccagagatcc taagcaacct ctgctcatct
 6901 gagttgtcca ccatattgtg ggcatgagtc cttgacaata gtaaatagca cctctgttcc
 6961 cttattgggt aaatgatttt ccaactctgg gaatgtgtag aattcattat ggaaataatg
 7021 caataattca aatccataat attgatactt tcatgttaag tttaggacta atcttgtgta
 7081 tgctccttaa gtgatttgaa tctttaaaaa gcttatgatt ccaatttgaa atgtgaaatt
 7141 gattttacgt ttgtgatttg aagttgaaag gtataagaat atttaactta gctcatgaaa
 7201 agtattagac tagatttact ataagtttaa tgtattagat ttacaagaga tgcttaata
 7261 tatgagaatg ttttgtctta attggttata atcttgtcat atcaatgatt tgaagtgtca
 7321 aaatagaaaa ttaaataatga taaattacac aagaagttta gaatgtttaa aagattttaa
 7381 taaacaaagc ctataactaa gaaaaaaaaa aaaaaaaaaa

(SEQ ID NO: 6) GenBank Accession No: NM_203395 (Version 2), incorporated herein by reference.

The amino acid sequence of mouse (*mus musculus*) IYD is shown below:

```

1 mfltltpvlva vvcilvvwvf knadrnlekk keeaqvqpww dedlkdstded lqveedaew
61 geaesvehi pfshtypeq emrmrsqefy ellnkrrsvr fissehvpme vienvikaag
121 tapsgahtep wtfvvvkdvd mkhkireiie eeeeinymkr mgkrwvtdlk klrtwnikey
181 ldtapvlili fkqvhgfaan gkkkvhyhne isvsiacll laalqnaglv tvtttplncg
241 prlrllgrp shekllvllp vgyprdatv pdlkrkaldq imvtv

```

(SEQ ID NO: 7) GenBank Accession No: NP_081667 (Version 1), incorporated herein by reference.

The nucleic acid sequence of mouse (*mus musculus*) IYD is shown below:

```

1 aggtcggact tcagaccatg tttctcctca cccagtcctt ggtagcagtt gtctgtatct
61 tgggtgtatg ggtcttttaa aatgccgaca ggaacctaga gaaaagaag gaggaggctc
121 aagttcagcc ctgggtggat gaagacttga aagacagcac agaagacctt caagtggaa
181 aagatgctga ggagtggcaa gaggcagagg agagtgtgga gcacatcccg ttctctcaca
241 cccgataccc agaacaggaa atgaggatga ggtcccagga attctatgag ctcctcaata
301 agagacgctc cgtcagggtt atcagcagcg agcacgtccc aatggaagtc attgaaaatg
361 tcatcaaagc agcaggaaca gtcceaagtg gggcccatc agagccctgg acctttgtgg
421 tgggtgaagga cccagacatg aagcataaga tcagagagat tatcgaagag gaggaagaaa
481 taaattacat gaaaaggatg ggaagcgat gggtcacaga cctgaagaaa ctgagaacca
541 actggattaa ggagtacttg gacaccgccc cagttctgat cctcatcttc aaacaagtcc
601 atggttttgc tgcaaatgga aagaagaagg tccactacta caacgagatc agtgtgtcca
661 tcgcctgtgg cctcctgctg gctgcaactg agaatgcagg gctagtgcag gtcactacca
721 ctccctcaa ctgtggtcct cgtctgaggg tgctcctggg ccgcccctca catgagaagc
781 tgttggtgct acttctctgt gggtagccca gcagagatgc cacagtgcct gacctcaagc
841 ggaaagctct ggaccagatc atggtgacag tatagccagc caacctaaag agatgctgga
901 atgctgggat gacccccact tctttcctcc cggggctgct ggcacctctt actctctgag
961 tcaggtcttc tgaaaccagt gtacagggaa ggccactttc cttgtagtga ttttttcat
1021 actttctaga actttcagtc cagtaagtta tttttaatgg actgttttag tgaatatatg
1081 gggcaagagg ccaaaaagca ggcataactt aatttctcta caaaattata aatgattttt
1141 tatttacata taaaagtc aaagaagtca cattagaaga tttgctatag ttcctaaagc
1201 tacctatgta atacttttta atttaaatat tgactaatca agtggtcctt ctgctctctg
1261 ctttctggtg gcattgagtc taagaattta ctactgagat ctattctgtc actaagcctc
1321 atagggatc tgccaatctt gaatgagtga cttttaacat aagcaaaata tgacactcac
1381 agaaccagag aggcattgtc aacctgccat ggatgtagct ggcaattatc tgaacatgag
1441 ttgcatggag tgtgccggag ctggcttgtg ctagcacatg gaagatgggt gttaaattat
1501 ctgaaattat acaacaaatt attaaaccat ttgtactttg

```

(SEQ ID NO: 8) GenBank Accession No: NM_027391 (Version 3), incorporated herein by reference.

The amino acid sequence of porcine (*Sus scrofa*) IYD is shown below:

```

1 mfltltpvlva vvcilmvwif knadgatkrr eeepraraea rpwvdedlkd stdvlqveed
61 adewqeseee vehvpfshtr ypekemvrrs qefyellnkr rsvrfisner vpmevidnvi
121 kaagtapsa htepwtfvvv kdpdvkhrir eimeeeekin ylkrmgprwv tdlkklrtnw
181 ikeywdtapv lilifkqvhg faangkkkih yyneisvsia cgiflaalqn aglvtvttt
241 fncgprlrvf lnrpanekll mflpvgyprsk eatvpdlprk pldqimvtv

```

(SEQ ID NO: 9) GenBank Accession No: NP_999581 (Version 1), incorporated herein by reference.

The nucleic acid sequence of porcine (*Sus scrofa*) IYD is shown below:

```

1  ccaccatggtt tcttctgact ccagtcttgg tagccgttgt ctgcattttg atggtgtgga
61  tcttttaaaa tgctgatgga gccaccaaga gaaggggaaga ggagcctcga gccagggctg
121 aggctcgacc ctgggtggat gaagacttga aagacagcac tgatgtcctc caggtggagg
181 aagatgctga tgaatggcaa gaggcagagg aagaggtgga acacgtcccg ttctctcata
241 cccgctatcc tgagaaggaa atggttaggc gatcccagga gttttatgag cttctcaata
301 agagacggtc ggtagattc ataagcaatg agcgagtcac aatggaagtc atcgataatg
361 tcatcaaggc agcaggaaca gcccgaagtg gggcccacac ggagcctcga acttttgtgg
421 ttgtgaaaga cccggacgtg aagcatagga ttcgggaaat catggaggag gaagagaaaa
481 taaactacct gaaaaggatg ggacctcgat gggtcacaga cctgaagaaa ctcaggacca
541 actggataaa agagtactgg gacacggccc ctgttttgat tctcatcttc aaacaagtac
601 atggttttgc tgcaaatggc aagaagaaga tccactacta caatgagatc agcgtttcca
661 tcgcttgccg catcttttta gctgccctgc agaacgcagg gctggtgact gttaccacca
721 ctcccttcaa ctgtggcccc cgactgaggg tgtttttgaa ccgcccctgcc aatgaaaagc
781 tgctgatggt tctcccgggt gggtacccca gcaaagaggc cacggtaccc gacctcccc
841 ggaaacccct ggaccagatc atggtgaccg tgtaggcagg atgtgggcag ggagatgctg
901 gaaggagggtg cccctcctcc cggtacctgc tcccctgggt atacaggctg tttttccct
961 ggggtgttgg tctccccagt gctttactgg gcgggggggtg gggggagaa ctaagaactt
1021 tgccgctgag aagccctgat tcccacagtg ggaatagctg gacgcacagg agaggtgaac
1081 cagcaggttt ttttccattt tttatccact ttggaaatgc attaaaaatt tgtaagcaa
1141 aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa

```

(SEQ ID NO: 10) GenBank Accession No: NM_214416 (Version 1), incorporated herein by reference.

The amino acid sequence of Honeybee IYD is shown below:

```

1  mltelfpfwt kywyhvliti vayiflklfy kilkkdaifi eknkqnfdis niikqlketn
61  qetedseesa lpkdlkhipy eykrpsemel fcqasefyki vtarrtirff ssdpvpkevi
121 yeiikaagta psgahtepwt fvavsnqkik eqiryivese eeinykkrmg vkwttdllpl
181 rtnwikeylt tapylllvfk qiygilpngk kkihyynems tciacgilit aiqyaglvlt
241 tstplncgpa irnllgrpsn eklvvllpvg ypakdatvpd lqrkslsdil veid

```

(SEQ ID NO: 11) GenBank Accession No: XP_397179 (Version 2), incorporated

herein by reference.

The nucleic acid sequence of Honeybee IYD is shown below:

```

1  ttaataaaaag tcattgtcta aagccagcgt cgatgaatta atttttttta
atagcctgag
61  tctttttttac gctgaataat taaatcgttt tcatgtattg cttaaagatt tactatatgt
121 ttcgatagat aatattctct ttttctaaga cagaaggtag aaaagacaac ttcaaatact
181 tgctaattat atatgataaa aattataaatt agtgcttttc acatttataa aattagaaac
241 ttattaagca tatcatgtat atgttaaaat acaagtactt aaatacttcg ttgtactgta
301 aatgattgat gaaaaattca tgaatatgta aatctatgta ttagtatata ataataatgg
361 gatacatcat ctgtttaacg cgataactta gatattgtag gtcagatata tcctactctt
421 gttgtaaacac atatcgttac ttacattgtc ttgaatcgtc aaaacaattg aatataatac
481 tcaatcttcg ttttcgaaat aaatatataa tttttattaa gaatgcttac tgaacttttt
541 ccatttttgg cgaaatactg gtaccacgtg ttaattacta ttgtagccta ttttttttta
601 aaattatttt ataaaatatt gaaaaaggac gctatattta ttgaaaaaaa taaacaaaat
661 tttgatattt caaatatcat aaagcaatta aaagaaacta atcaagaaac agaagatagt
721 gaagaatctg cattaccaaa agatttataa catattcctt atgaatataa aagaccttct
781 gaaatggagt tattctgtca agcttctgag ttttataaaa tagtaactgc tagaagaact
841 ataagatttt tcagttctga ccctgtacca aaagaagtca tatatgagat aataaaagct
901 gcaggcactg cgcctagtgg tgcacataca gaaccatgga cattcgtagc ggtgtcaaat
961 cagaagataa aagaacaaat acgttatatt gttgaaagcg aagaagaaat aaattataaa

```

1021 aaaagaatgg gagtaaagtg gacaactgac ttacttccat taagaacaaa ttggattaaa
 1081 gaatacctga ctaccgctcc ttatttgggtg cttgtattta aacaaattta tggaatttta
 1141 ccaaatggaa aaaagaaaat tcattattat aacgaaatga gcacatgtat tgcttgtggt
 1201 attctcatca ccgctataca gtatgcaggt ttggtaactc tgacttctac tcctttaaat
 1261 tgtggacctg ccatccgcaa ccttcttgga cgtccttcta atgaaaagct tgttgacta
 1321 ctgccagttg gttatccagc aaaggatgct acagtgcctg atcttcagcg aaaatctcta
 1381 tctgataatt tagtagaat tgattgacat atattataac aaattatggg aattaaagta
 1441 cctttgttta atattttatc agatacttgc ttgtaggaca atacaaaaat gagtacacat
 1501 aattatagat ttatataaat cctgtttttg taaaaaaact tatatttgtt ttgagta

(SEQ ID NO: 12) GenBank Accession No: XM_397179 (Version 5), incorporated

herein by reference.

The amino acid sequence of Mosquito (*Anopheles darling*) IYD is shown below:

1 masyleswvd slisssalvn ywnyifpmlv glylaislyd khryggrkkl
 riispptdst
 61 tdatrkldgm eyvgeldhdg eegyelspal eekphvpyrg atvtfakdps eaadkfyai
 121 nnrrsvrkfs qrpvdrdive rciaaagtgp sgahtepwtf clvsdrsvke sireiiesee
 181 linytqrmak qwvtdlrpiq tnhikeylse apylilvfkq tyshkadgtk kqhyneist
 241 siatgillca lqsaglsslv ttplncgpai rnllerppne kllvllpvgy aedctvpdl
 301 qrkpldeliv ry

(SEQ ID NO: 13) GenBank Accession No: ETN68090 (Version 1), incorporated

herein by reference.

The nucleic acid sequence of Mosquito (*Anopheles darling*) IYD is shown below:

1 aactgtttat gctattggtg tcacagagta ccatgcgtat gggtgcagag ctccactcat
 61 acacagctgt ttccgtgggtg tagtggttat cacatccgcc taacacgcgg aaggccccc
 121 gttcgatccc gggcggaac aggcgatatc tttttgcttc ctccaaacac ctcttgtgt
 181 gtggatcatc ttttttcttt tgcgaacca cgttcttttc gcgttctcaa gcgatccact
 241 atgcacctca cagaaacatt ctcaatttga aaatatcata cagcagcaac acgcgatatt
 301 taaggatagt tgattgttga caaataagga ccaatagatg gttttattga ttttagtta
 361 tgctcatcac taataacgaa agacttagat gagctttact tcctcttctt accggcaaat
 421 ttcttcttcg ttatcccgcc accaccacct cactgcgtt gtttactgat gaaacgcttc
 481 ttgctactcg cctgcacccg gtttttgcca ctaaaccgag actgctccca ggccatctgt
 541 ttggcggatg aattatcctg attgaaaact tcaccagcgg taccgtgcgc cccggcggat
 601 tgtaatttct gtttgtgctt gcctcccttt gctttcccg cggctccga cgatccaaca
 661 ttgcgtcaa gcagctccga gaagtcacg gccgcagcaa acagtgaagt catgtcggac
 721 gacttgagct tccgggcaaa gtctcgttca ctacgcggg tacgtaacat ctgtttcttc
 781 gccttcttcc tcaactggctc ctcatcttct tcttctgggt catcgtcac atcatcgtea
 841 tcatctccgg catcgaagtc atcatcttcc tcgagcgaga tgctaccacc atcactgaac
 901 tcttcatctt catcgtctcc accaaaacca cgtggttcat catcgtgtt atcgccctcc
 961 tcgtcgtctt catcatcatc ccccggtacg tcatcccaat catccaacgc accatcgtcg
 1021 tcctcttcat ctcatcatc agcatcctgc tgacccttct tcttaccctt ctcttgcctc
 1081 gctttacgct ccatgtcctg ttccagctcg gtcataagt caagctccga ccgatccata
 1141 ttggcaccat catcccgccc accaggaaca ccaaacgat ccaagtatgc atcgaactcg
 1201 tcgtcatcga gtgagtcaac gtcggaatca acctccgct cgtcttcttc ttcaccgac
 1261 ttctttcctt cggccttttag cttcagtgc tttttgcgt cctccgctcg gcggaatcga
 1321 tctcgtttct tctccagaaa tctgcagaaa ttagaaataa ttagttgtta ccacagtgga
 1381 atcgattcca cagagtaata caacagttgg acttactgga agatgtactg ctcatcctcg
 1441 gtgactggt ccttggtgag tgactcgacc ggtagagctc gtgagccggt aggctggtaa
 1501 tctcccttgc gttgtgccac accgagcgg cttggacggt ttacctcttg gccgctcgt
 1561 tcgtcgattg ccttttgggg tttcttcgga ttcttaaag caaatcgatc caggaaatgg

1621 ccgagtgaga aatcccgc aa tggatcacca tagtagctaa gtggggcatt gctaagaata
 1681 cattcggcga acttttgtac cgttggatga aagtattcga gataacgggc cagctcgtac
 1741 cggagcgtgt acttagcccc ggcaaaactcc gatgcacgat ggaaagcatc ataaccgcga
 1801 acaaccgatg aattgctaac agttcttccg gcacctttag gtggtttcac atcaagctct
 1861 ttctgaccc cgtcgtcatc gtcgtccttt ggtgccaaaga taggtccatc gcggtcgtcg
 1921 tccttctttg aagcggacag agctgtggcc tctgctggtg ccggtagacc atcgatcagc
 1981 agttgacgac gctttcggag cactttgctg atgatgatca gagggccaca aactcgagcc
 2041 ggtgggaaat aaaatgccac ctgtagcaga cgcttgatga aggcttgccg acgatcaggg
 2101 atcgatcgt tctgcatggc acggtgtata atgtagaaga acacggttga gatgcgtgga
 2161 ccgacggagg aaagctgagg atcaagcagc ttgcggtata gcgtattgta gaaacggttc
 2221 tgctcacgac cagggcgctc cgtcacttcc agcaacaacg agagcccttg gcaaccgatc
 2281 gcaatgtctg ccagatgcac cattcgatag ataacgttca gcagctcggg ctgcacgata
 2341 tcagctactg cgatgtcacg tttcacgcta ccgatcgctt tgcgtagaca ggccagtatg
 2401 gcctgcatcg tccgcgagtt gatctcacca cgctcggtca gaatcttgaa gaaggcaaaag
 2461 cacacgttaa tcagcttctg gcacgtggca aagtcaccgt acgaggcgat cgaggccagg
 2521 aacgagagcg cataatgctg cgctgtcgct gagacgttgt tgcgaaagag caatttctcc
 2581 gtttccgtgg caaccactag tgccatggcg ctgtgtttcc gcacgacctg caacagatga
 2641 tagagcgctt ttacggctac gcgcgctgac ggatcgccaa gtttgttgac gagcaccgtc
 2701 agcaaaaagg cttctttctc cggtatgtcc gcaaacagac gtgccgtgtg aacgatcgcc
 2761 ttgcacttgc tatcctcctg tcccagagtgc agtatcaccg atatgttggg cagaaagtta
 2821 aagtacagct cgcgtagctg atcctcgaag tgccagtgcg ccagcagtgg ttcgcgaatg
 2881 gatttcgaca gctcttgccg ctgtacgtta cgccaatcag cccacgaag tggaatcggg
 2941 attaaactttc tcgaggacgg aaggagtgat ttcagcatca gctcggttaa cacttccacc
 3001 acgtcaaggt ggcccttggt cgagggtttc accatcccga tcaccgtctc cagagcggac
 3061 agattgcaga acgggttcga ctgtacgagt agcgtcccg aactggctcg atcacgcgat
 3121 gttcccttct cgatcgcgct ctgcaaccag cgggcacgag atgggttgct cttcatgagg
 3181 acacgacatt cggcgctgaa ggccgcggcg cacagtctcc taagtctctg gatttcgggg
 3241 tctttcagct caaccacctc gccaccgacc gtattgtatc cgtcgaaatg ctctaccag
 3301 cgtttcacct tctcgtcgtc tcccggcagg ttgaaactac tgcccgtcg accgatttc
 3361 gcattcttcc ttttaccgcc agctgcctgc tccgtgcgtg tttcttcac cgcggctcg
 3421 tgttcgatgg gatcgctgat cggcacttcc tgttgttctc catcttcgt gaagacgatt
 3481 ttcttcggct tccggttggt atcgagcaca gcggtttcac ttgggttctc aaagtatcgt
 3541 ttcgagtcct ttttccgtac ttttctcctc tcgatagcca tcgtgaacac agaaccgcga
 3601 ttttttctgt ttatctcgaa acacgcgggtg cagtgcgttt atgctgtggc ggggtcgggt
 3661 tgacagcaca atcgggtgac cagataaaaa ataaacacgt agatgtcacg gacgggacaa
 3721 gcgcgctacg tctagccagt caaagcgact agtcagagt tagagaagt tagacaaaat
 3781 cccgcgaaaa gtgcaataat ttgttaaagt gaggtaaaact ctcaagtaaa atgcctccga
 3841 aacgaaaatc caccaccgca acggacgagg aagacgagga gtatcgcaag aagagggacc
 3901 gcaataatca ggtaacgagt taggtcgtcg atttcatcat cctgcatctt aacgatacgg
 3961 tttctttttg ctcaaaggcg gtgaaacgga gccgcgtaaa gtcgaagatg aaaacggaag
 4021 aaacgcagca gcgtgtgaac gaactacgag taaagaacca gctgctcgag gaaaagatcg
 4081 ataaccagca caaggagcta aagtttttga aggaactttt cctaaccagg gccaggcgga
 4141 aggccgacaa actggtcggg gtcaacctga acgatctttt gcgctgctcg gacgaggaaa
 4201 gcgacagtgc agatcaagg accagcaagc agaagaagaa cgagtcctca aaaggtgaag
 4261 gcagcagcgg gagcaaacgg tccagaaggt gagaggccac tcctggaatc cacatcctgc
 4321 atacgagtac taatttttaag caagcatttc cagacaattg aggaaagcat aaggcattaa
 4381 gatttttggg taaagttaag taagcaactc aacacgcaag tgtataaaat acttttaatt
 4441 gcatcatggt agtttttaata tatcagcgta agtttttaata cacctgtact attacacatc
 4501 attcaaccga tgttcaaacc ttctttgatg cttgttaaag tagaaagacg taaaggtaaa
 4561 atgagaaaaat actcatataa cggagtaatc catggatatt gtgtggaatg aattttgcgt
 4621 gcagccttgc tgaggccgtg tggcctaata gagaaggcgt cggacttcgg atccgaagat
 4681 tgcaggttcg agtctgtca cggctgtgct cgtgctgaga tgattttctt tatttatgta
 4741 caacgagcac atagactcct tttcaacaaa cgggtaaaat aaaacataga aatataggac
 4801 gtccagtagg tagtagtatc cagtgaacc ttttactctt tacttttccc attgtattaa

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4861 caatcgattt tgaaaaaaaa tccaacctca gtacgacaaa aatacaaaaag atttcttcca
4921 agcgaccgtg acaggactcg aacctgcaat cttcggatcc gaagtccgac gccttctcca
4981 ttaggccaca cgccagttg gcagggtgtca acttagatta taacttggaa tcattaacat
5041 taaaaatcaa attttttggtt attttccatc gaaacaatcc ttttttttcc attttactta
5101 cggtatgcta atctacaaat tttcagaatt gttttaaaga aaaaaaaaca aaatagccaa
5161 tcatgacagc cattctgggg atcaatgatt gccttaaagt tccaaagccg tttgctaaaa
5221 agcctcattg caatgtctgt ggccgcctta acattaaaac tatagtagcc atccgtttgc
5281 ttttcatcaa attttatctg gacattatth gcaaggtagc ctaggggagt ttacaacata
5341 tttaatgata tatttaagca tggcaccctt cgccagaatg gtcaccattg ctgtatthtt
5401 tatatthtatt ttataaaaaa ctccacaaat actcttcaaa agttgtagta tcgtttgcct
5461 tcaacaaaaa tataaaaaatc tactcagttc cagtacaaaa tcacatgaat gtgccattht
5521 ttgctaaatc acaccaaatc cttttctaag caccagttt tcaactggcc gtgtggccta
5581 atggagaagg cgtcggactt cggtaccgaa gattgcaggt tcgagtcctg tcacggtcgc
5641 tttgtgtgaa aattacttht tgttgtgtga tatatcaaac atctcttht tgctcttata
5701 ttaaatcaag tgataacatt tttatcagaa ttatcagaga atcgtgaaca caatgtatta
5761 caccgaata ttttgtgtgc gtgtgacaaa cttatcgttg tttacatggt tccggcggca
5821 gcagcacgtg gtgttatgca gttccccatc ccatcgctta gctccatcgt cgttatcgtc
5881 gttttatcaa acttatctaa aatcagcgcc cgactthttt tgcaaccgac cgtctggaca
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(SEQ ID NO: 14) GenBank Accession No: ADMH02000012 (Version 1),

incorporated herein by reference.

The amino acid sequence of Silk Worm (*Bombyx mori*) IYD is shown below:

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1 mniislvvwt lekncfivyt alivcfigft ifhknrldih yrnveikeek lartgpkkke
61 refpddgddd pelllpaape dtphipyrrp rksdeeilqk sreyyelmae rrtvrcfsae
121 pipqevldni vktagtapsg ahtepwtfvv vqdpqikeai reiveqeel nytqrmsrqr
181 vtdlkipfat pskpyleap alvlvfrqay swrsdgkkrm hyyseistai aagfllaaig
241 ycglvaltst plnanarlrd llsrpanerl elllpvgrph kettvpdldr kpleeimlki

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(SEQ ID NO: 15) GenBank Accession No: XP_004925281 (Version 1), incorporated

herein by reference.

The nucleic acid sequence of Silk Worm (*Bombyx mori*) IYD is shown below:

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1 tacattatgt atacaacagt cggctgaaga actcagccct gagtaaactg atcgagtaat
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121 tgtacacagc attaatagtt tgttttatag gcttcacgat attccacaag aacagattgg
181 atattcatta caggaatggt gaaataaagg aagaaaaatt ggcgcggaac ggcccaaaga
241 aaaaaagaac tgaattccct gatgatgggg acgatgacct agagttactc ctgccagcaa
301 tcccgaaga cactccgcac ataccttacc gacctccgag gaaatccgat gaagaaatcc
361 ttcagaagtc caggaatat tatgagctga tggccaaaag gcgaacgggt agatgtttta
421 gcgctgagcc tatactcag gaagtcttgg ataattattg gaagactgct ggcactgcac
481 catcaggcgc tcacactgaa cctggacct ttgtagtggc ccaagatcca caaatgaag
541 aagcaatccg cgagattgta gaacaagaag aggaattaaa ctacaccagc aggatgtcca
601 gacaatgggt gactgatctc aaaccttttg ccacaaagcc aagcaaacct tacctatctg
661 aagcaccagc tttagtcttg gtgttcagac aggcgtactc ctggagatca gatggaaaga
721 aacggatgca ttactatagt gagatcagca cggccatcgc agcaggattt ctgctagcag
781 ctattcagta ctgcggccta gtggcgetta catcgacacc tctaaatgcc aacgctcgtc
841 tccgggatct cctctcgaga ccagccaatg agagactgga actccttctg ccggtaggaa
901 ggccgcacaa ggaaacaact gtacctgatt tagatagaaa gccttttaga gagatcatgc
961 ttaagatctg aactgctgct tagttttaat agctcatcca aatgactgaa taaattaact
1021 atacaaaagt g

```

(SEQ ID NO: 16) GenBank Accession No: XM_004925224 (Version 2),

incorporated herein by reference.

The amino acid sequence of a wasp (*Nasonia vitripennis*) IYD is shown below:

```

1 mdtgqrypnn fsstmerads kiatgladfp ffakyghyvl atvvlvsivn lilgryskst
61 sivskepvas kelspdeefn eyetadged epylpkdlqh vkfeykrpse aeivkrslef
121 yeiadarrtl rffspdpvpr evirnivraa gtspsgahte pwtfvvsnp svkenireiv
181 ereeeinylk rmgkkttdl qplktdwke ylatapyll vfkqdygflp ngkrkihyk
241 emsvaiacgi litaiqyagl vltstplnc gpallrvlln pvneklvlll pvgypaeadt

```

301 vpglkrkdld evlvefe

(SEQ ID NO: 17) GenBank Accession No: XP_008213524 (Version 1),

incorporated herein by reference.

The nucleic acid sequence of a wasp (*Nasonia vitripennis*) IYD is shown below:

```

1 agtctatgga cacgggacag agatatccga ataatttttc atcgacgatg gagcggggccg
61 attcgaaaat cgcgaccggc ctcgacgcct tccccTTTTT cgccaagtac ggccactacg
121 tcctggcgac agtcgtcctg gtgtcgatcg tcaatctcat cctcggaagt tacagcaaga
181 gcacctcgat cgtctcgaaa gagcccgctg ccagcaagga attgagtcgg gacgaagagt
241 tcaacgagta cgagacagcc gagcgccccg aggacgagcc gtatttaccg aaggacctgc
301 agcacgtcaa gttcgagtac aagaggccgt ccgaggcgga gatcgttaag agatcgctgg
361 aattttacga gatagccgac gccagacgta cgctgcgctt cttcagtcct gaccagttc
421 cccgcgaagt aattcgcaac atcgtcagag ccgcgggaac ctccccagc ggagcccaca
481 ccgagccctg gactttcgtc gtggtgtcga atccctcggt gaaggagaac atccgcgaga
541 tcgtcgagcg cgaggaggag ataaactacc tcaagcgtat gggcaagaag tggacgaccg
601 atttgcagcc gctgaagacc gactggcgca aggagtacct cgcgacggcg ccttacctca
661 ttctcgtctt caagcaggat tacggctttc ttctaatgg aaagaggaag attcattact
721 acaaggagat gagcgtcgtt attgcctgcg gcatactcat cactgccatt cagtacgccg
781 gtttggtgac gctgacgtcg acgccgctga actgcggaac ggcactacgc gtcctgtcga
841 acaggccggt caacgagaag ctcgttctgc tgcgtcccggt tggttaccgg gccgaagatg
901 ccacggtacc tgggtctgaaa agaaaggacc ttgacgaggt gctcgtcgag ttcgagtgc
961 gctgaagcga cgtgagattc acaatcgaca aaagacatgt ttgactttta taaaaaatgt
1021 agatttttaa caaatacatt gttcgtaagt gagaaaacgc atctttatc tttctgggga
1081 gacagcttga tcggagaaat gcattgcgta tgcaggagga aaaacgcgct gaaatacagt
1141 ttgtataaac taatgtttat tcaaatttgc gtacattatt gttaaaaata agcatgtata
1201 aaaattatat ttaaggacgc atgttagaat caacggagac gcaaatcat tctgtgtgat
1261 acaattcaat tcacaatctt tatcatcggt tcttttataa ttcacctcta cgcactgata
1321 aatgatccgt atagctctct cggataccgt atgtatatat gaaaataaag gaaacgtgca
1381 tttcactttt ttacgcgag taccaaatat atatagtcgc caacgatagt gtgacgaagg
1441 agaaataaaa aaaataatca ttaagtacac gacgttccgt tcgttatacg cgtattatat
1501 tatatgtaaa atacaaaaat actgagctaa aacaaccaag gggcgttaga attgttagca
1561 cgcttttttt tcatcgttgc agaacttggc tttatcataa atggctttcg ttcttccttc
1621 gtttcgttat aatcttttat tactagactt acacgtatgc tgctaccgag agggactttt
1681 gtttctatg tcattaagtg tagaaatcag aaagtttcac tgtaacttaca gattaaaaaa
1741 cgttgttgct ctttttcttt tcattattta ctacattaga gcaattatth tatacgacac
1801 actaaacatt atgcacttgc aatcgaaaaat ggccaacaga atcaaggcaa aaatacagtt
1861 tagaaattca ttcggtgatt ttcgtctgta ataaatacta taaagaggca taaaataaat
1921 aatttaccag ctttttatcg cgattcgaca aggaatcgcg aattcatgag ggaacaaact
1981 ctttcgatg attctggtga cgagttctct ccgttattct ataaaattaa atcttcacgc
2041 ttttacacga ttccattgac gcctaacgca catttacact aatttttcac acgttcttca
2101 agaaaaaaat aaaaaaaaaa taaaaaaaaa cggactacat agaaaaataa actttatttc
2161 aaagttgctt gcttaaacgg gtgtcgaaat tttgaaattt caacaatgat gtacaaagtt
2221 acggaacta ggagagtatt tttctacaat aaaaaagga aatgctctgt ttcacgata
2281 ttaatctttt tatagttcag gctactttca gtcgtttggt taaaaactac cgcataatgc
2341 tattgtcaac ataatacact tagaattaca agaatttaca agaggtatga gcagatcaaa
2401 atctcgccca acgataggca agaagttaat tattgttata tatatatata tatactatat
2461 atagttacaa ttcttatttt accaagggtg agcccaatca gcttcgaatc tttcgtcgtt
2521 tcgcgaccga ggttctgaca atgtggaaca aactgtatag cgcagttact aatttaacaa
2581 cgcacacat ggaaatttca gtcctacagt ctagggttga aaaacgactt gttagtaaaa
2641 atcattaaaa aaaacctggt tgttttagta gtaaatcaaa aaaatcatcg tggtttagaa
2701 atatcacagc tgttttagatc attcatatat cacttctcca accgtatata aatttcccg
2761 atattcaata agatcgagtg caatgatcga ggtatattgt cagaaagaat agcgctaaaa
2821 cgtaagtcta cacaataaaa gaaacaaaaa aatctaaaaa gttcattcca catttcctta

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2881 tagcaacggtt aggcgcgaca acgattttcca acgactcaca agagaagggc tccaccttga
2941 aataaaaaata aaaaaaaaaat aaaaaatacgc tgcacgcaca cgccgagctt acttacagtc
3001 gcgagaataa acatcagaga tataccgatac ttatgcaacg cgccaaagtt acacatgtgc
3061 atccgatacgc cgcatacgcg gattgttcgc taataattga ggccagaata tatacctacg
3121 tcgacatcag acaaaaggag agaaacgcgc gcggcagact cattaaaaaa tacgcacaat
3181 tattagtcgt ctgagatcat aggatgcctg agaggataga tggatatata gacgtctgac
3241 aaacgttgca ccaatggttt ttcaactatt tcaactgagta cgtattttacg taatgttcaa
3301 caaactgtag ctttctgaga ttgaatgcga gagtacaaga atatatatat atatatatat
3361 ataattgtata tgcatacgaag aaggatgtga tatatgtata cgacagattg ttctttgtga
3421 atattataaag cgtcattgtt acgaccgcgc ctccggtgatg tgtgagcgta agtgtgtatc
3481 tgtgtgtgtg tgtgtgcacc aagtgttgcc ctgcataatc cgcgcgctgt ttggattatt
3541 ccagatcgaa gattatcgat tatattatat gtacaattat caaccctata gtctctataa
3601 gtaactagct gccacgactg aatttcgaag gatagatcga tgagattgat acagaagcac
3661 ccatcagcaa cactcgcgga ctacaagcag cagcagtcgc cctaaaagta cacaactgct
3721 tctgtaaacc ctacgctatt atttgttttc tgtttcttct ctctatatatt ata

```

(SEQ ID NO: 18) GenBank Accession No: XM_008215302 (Version 1),

incorporated herein by reference.

Another exemplary amino acid sequence of mosquito (*Anopheles gambiae*) IYD is shown below:

```

1 masyledlvd yflsssalvt ywnyifpvlv glylaislyd khrygrnkl ritepaatek
61 ngskldgiey vgdlehadyd plpaleekph vpyagatvlt aqdpleaaer fyaianrrs
121 vrkfssrpvd vaivercira agtapsgaht epwtfclvsd psikasvrei ieaeefvnyt
181 qrmakqvwtd lrpirtnhvk eylteaphlv lvfkqtygk vdgdatakkq hyneistsi
241 atgmllcafq caglsslvtt plncgpalt llerppnekl lvllpvgya ddctvpdlqr
301 kpledilvry

```

(SEQ ID NO: 19) GenBank Accession No: XP_3115442 (Version 4), incorporated

herein by reference.

Another exemplary nucleic acid sequence of mosquito (*Anopheles gambiae*) IYD is shown below:

```

1 tgttcttggc gcgcgtcggt gttgctcggt aacaccccat ccccatccgt
ctgcagagcc
61 atggcctcct atctggagga tttagtcgac tatttcctca gcagctcggc gctcggtacc
121 tactggaact acatcttccc cgtgctggtg ggactgtacc tggcgatctc cctgtacgac
181 aagcaccggt acggacggaa caagaagctg agaattacgg aaccggcggc aacagagaag
241 aatggcagca agctggacgg tatcgagat gttggagacc tggaacatgc tgactacgac
301 cactgcctg cactggagga gaaaccacac gtcccatacg ccggtgcaac ggtcacacta
361 gccaggacc cactagaggc agccgaacgg ttctacgcca tcgccaacaa tcggcgagcgt
421 gtgcgcaagt ttagctccc cccggtggat gtggccatcg tggagcgggt catccggggc
481 gccggtacgg cgccaagcgg tgccacacc gaaccgtgga cttctgtct cgtgtccgac
541 ctttcgatca aggcgagcgt gcgcgagatc atcgaggccg aggagtctgt caactacacg
601 cagcgaatgg ccaaacagt ggtgaccgat ctgcggccca ttcgtacgaa ccacgtgaag
661 gactacctga ccgaggcgcc catctggtg ctggtgttca agcagacgta cggctacaag
721 gtggatgggg acgccacggc caagaagcag cactactaca acgaaatatc gacctcgatc
781 gcgaccggca tgcgtctgtg cgcgttccag tgtgccgggt tgagctcgct cgttaccacg
841 ccgctcaact ggggccctgc cctgcgcacc ctgctcgagc ggccaccgaa cgagaagctg
901 ctggtgctgc tgcccgtcgg ctaccggcg gacgactgta ccgtgccgga tctgcagcgg
961 aaaccgctgg aagacatcct ggtgcgctat tga

```

(SEQ ID NO: 20) GenBank Accession No: XM_315442(Version 4), incorporated herein by reference.

The amino acid sequence of a house fly (*Musca domestica*) IYD is shown below:

```

1 meldeilets vflkywnvvl tilligilsk wlwnrkghkq ieredednei
ladssskaee
61 idyapaleek ehiaykgave tlpdgakefy kmmndrrsvr syrrepippi dvienciraa
121 gtspsgahte pwtfcvitdm eikqsirdii eseeelnyaq rmhkqwttld rplktnhvkp
181 yltdapylii vfkqtygvtk qgkrkqhyyn eisvsiavgi llcalqaagl sslvttplnc
241 gpalrallnr pvnekllvll pvgypaenck vpdlerkqld diivry

```

(SEQ ID NO: 21) GenBank Accession No: XP_005190343 (Version 1), incorporated herein by reference.

The nucleic acid sequence of a house fly (*Musca domestica*) IYD is shown below:

```

1 catttttgta ttttcattca acgaatatgg aacttgacga aattttggaa acttcggtat
61 tcctaaaata ctggaatggt gtactgacaa tattactcat tgggattttg agtaaattggc
121 tgtggaaccg aaaggggtcat aaacaaatcg aaagggaaga tgaagataat gaaattccttg
181 ccgattccag ttccaaagct gaagaaatag actatgcccc tgctctggaa gaaaaagaac
241 atattgcata taaaggggca gtggaaactt tacctgatgg tgctaaagaa ttctataaaa
301 tgatgaacga caggcgtagt gtctgctcat accgccgtga gccaatcca cccatcgatg
361 taattgaaaa ttgtatacga gctgctggta cctcaccgag tgggtgccat acagaaccct
421 ggactttttg tgttattacc gacatggaaa taaaacaaag tattcgagat attatcgaat
481 cggaagagga attaaattat gctcaacgta tgcacaaaca atggacaacg gatctgagac
541 cgtaaaaaac aaatcacgta aaaccatata taacagatgc tccctattta atattggtat
601 ttaaacaaac gtatggagtt accaaacaag gcaaacgtaa acaacattat tacaatgaaa
661 tatccgtatc catagcgggtg ggtattctac tgtgtgcctt gcaggcagct gggttgagtt
721 ccttggtcac cacaccatta aattgtggcc cagctttacg tgctttgcta aatagaccgg
781 ttaatgagaa attattggtt ctcttgccag tcggttatcc tgcggaataa tgtaaagtcc
841 ctgatttgga acgtaagcaa ttagatgata ttattgtcag atattag

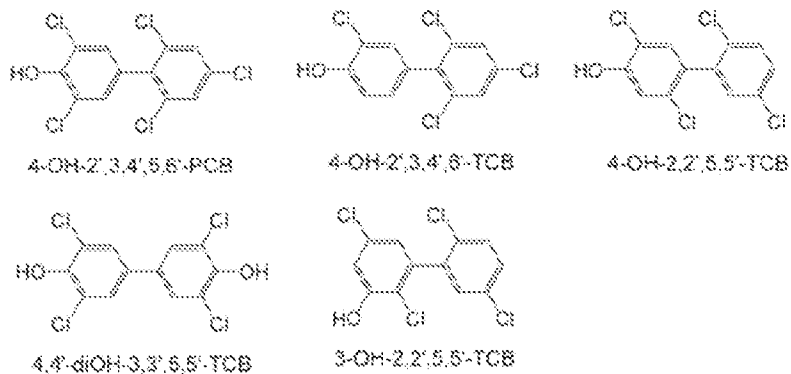
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(SEQ ID NO: 22) GenBank Accession No: XM_005190286 (Version 2), incorporated herein by reference.

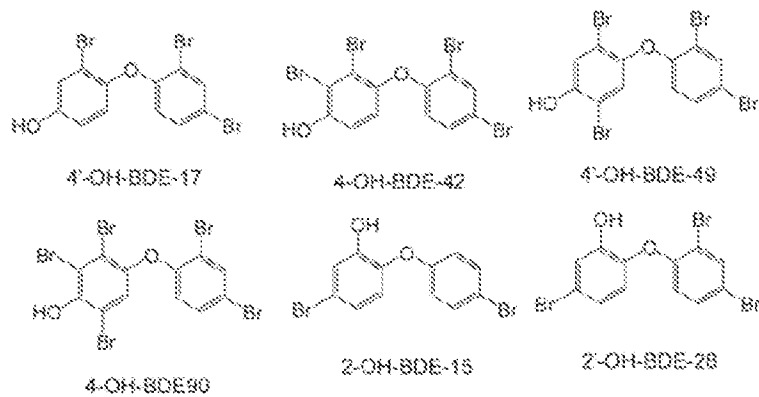
IYD Small molecule inhibitors

Several small molecule inhibitors have shown IYD-inhibitory activity. As such, these small molecules are useful as inhibitors of various IYDs. In humans, these inhibitors can disrupt thyroid hormone homeostasis by blocking iodide recycling through inhibition of IYD activity in the thyroid. Various classes of phenolic compounds comprise: PCBs (polychlorinated biphenyls), PBDEs (polybrominated diphenyl ethers), agrochemicals, antiparasitics, pharmaceuticals and food colorants.

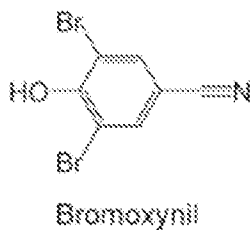
PCB examples include:

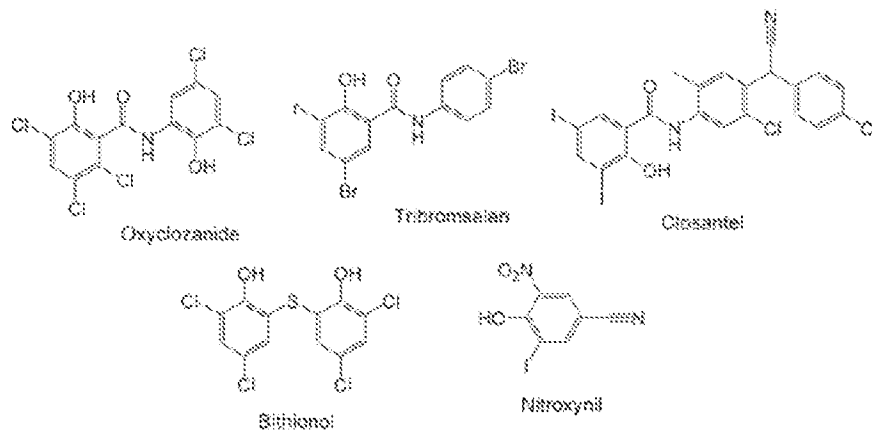
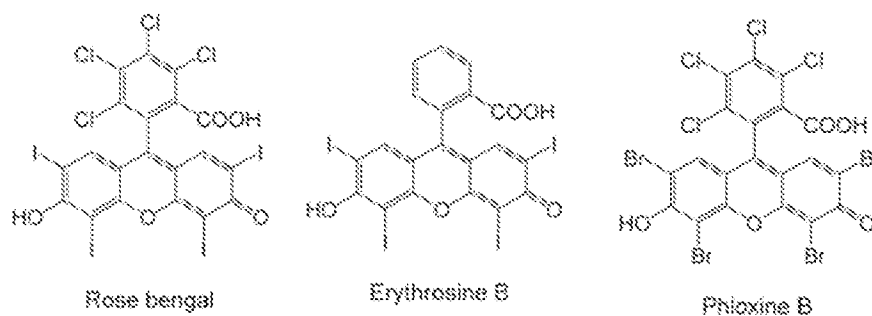


PBDE examples include:



Agrochemical examples include:

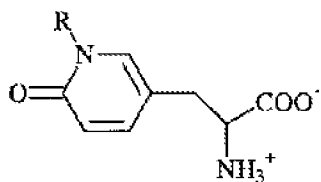


Antiparasitic examples include:Food colorant examples include:Competitive IYD small molecules

The pyridonyl amino acids (Table 6) were all observed to be reversible and competitive inhibitors of diiodotyrosine turnover under standard assay conditions (Kunishima, M., et al., *J. Am. Chem. Soc.* 1999, 121, 4722-4723, incorporated herein by reference).

Table 6: Kinetic and binding constants for iodotyrosine deiodinase.

inhibitor	R	K _I (nM)
1	H	1,100 (±100)
2	Me	24 (±8)
3	Et	87 (±13)
4	iPr	400 (±33)



Spermatogenesis

The method described herein comprise administering to the insect a composition comprising an agent that inhibits spermatogenesis (e.g., late stage spermatogenesis) in the insect, thereby preventing, reducing, or inhibiting reproduction in the insect.

Spermatogenesis is the process in which spermatozoa are produced from male primordial germ cells by way of mitosis and meiosis. The initial cells in this pathway are called spermatogonia, which yield primary spermatocytes by mitosis. The primary spermatocyte divides meiotically, Meiosis I, into two secondary spermatocytes; each secondary spermatocyte divides into two spermatids by Meiosis II. These develop into mature spermatozoa, also known as sperm cells. Thus, the primary spermatocyte gives rise to two cells, the secondary spermatocytes, and the two secondary spermatocytes by their subdivision produce four spermatozoa.

Spermatogenesis begins immediately after meiosis, when spermatids begin to undergo the morphological changes required for sperm development. Elongation of the spermatids occurs within a syncytial cyst that becomes polarized such that all the nuclei localize to one end and the growing ends of the sperm tails are found at the other end. Spermatid elongation is accompanied by dramatic changes in nuclear shape. Following meiosis, round spermatids (alternatively, onion stage spermatids), contain spherical nuclei that are approximately 5 μm in diameter. During sperm development, the nuclei become thinner and the chromatin condenses. Following elongation and nuclear shaping, the mature sperm are invested with their own membranes in a process called individualization. Individualization requires formation of an individualization complex (IC), composed of actin cones that form around the nuclei in a mature spermatid cyst. Following individualization, each group of mature sperm becomes coiled in a process that brings the entire length of the nearly 2mm long sperm bundle into the base of the testis. After coiling, the sperm are released into the testis lumen and are transferred to the seminal vesicle, where they are stored until needed for fertilization.

Common insects

Insects are a class of invertebrates within the arthropod phylum that have an achitinous exoskeleton, a three-part body (head, thorax and abdomen), three pairs of jointed legs, compound eyes and one pair of antennae. They are among the most diverse groups of animals on the planet, including more than a million described species and representing more than half of all known living organisms. The exact number of species is estimated between six and ten million, and potentially represents over 90% of the differing

animal life forms on Earth. Insects may be found in nearly all environments, although only a small number of species reside in the oceans, a habitat dominated by another arthropod group, crustaceans.

An arthropod is an invertebrate animal having an exoskeleton (external skeleton), a segmented body, and jointed appendages. Arthropods form the phylum Arthropoda, and include the insects, arachnids, myriapods, and crustaceans. Arthropods are characterized by their jointed limbs and cuticle made of chitin, often mineralised with calcium carbonate. The arthropod body consists of segments, each with a pair of appendages. The rigid cuticle inhibits growth, so arthropods replace it periodically by moulting. Their versatility has enabled them to become the most species-rich members of all ecological guilds in most environments. They have over a million described species, making up more than 80% of all described living animal species, some of which, unlike most animals, are very successful in dry environments. Like their exteriors, the internal organs of arthropods are generally built of repeated segments. Their nervous system is "ladder-like", with paired ventral nerve cords running through all segments and forming paired ganglia in each segment. Their heads are formed by fusion of varying numbers of segments, and their brains are formed by fusion of the ganglia of these segments and encircle the esophagus. The respiratory and excretory systems of arthropods vary, depending as much on their environment as on the subphylum to which they belong.

Extrapolation of the data herein indicates that inhibiting IYD enzyme in insects (e.g., mosquitos), may provide a strategy for population control. The compositions and methods described herein are useful for inhibiting spermatogenesis in any insect, e.g., cockroaches (e.g., brown-banded cockroaches, German cockroaches, American cockroaches, Oriental cockroaches), beetles (woodworms, death watch beetles, fur beetles, varied carpet beetles, spider beetles, mealworm beetles, centipedes, red flower beetle, pine beetle), earwigs, flies (e.g., bottle flies, blue bottle flies, green bottle flies, house flies, fruit flies, horse flies, black flies, deer flies, house flies, teste flies), cicadas, hoppers, ants (e.g., Argentine ants, carpenter ants, fire ants, odorous house ants, pavement ants, pharaoh ants, thief ants, a leafcutter ant, a mountain fireant, a Jerdon't jumping ant), aphids, bed bugs, bees (e.g., a common eastern bumble bee, a bumble bee, a honeybee, an alfalfa leafcutter bee), wasps, mantids, earthworms, terrestrial crabs, firebrats, fleas, snails, slugs, crickets (e.g., house crickets), hornets, locusts, lice, moths (e.g., almond moths, Indianmeal moths, cloths moths, gypsy moths (common clothes moths), brown house moths), red spiders, silverfish, woolice,

termites (e.g., dampwood termites, subterranean termites), ticks, wasps, mosquitos, gnats, millipedes, stink buds, weevils, mites, silk worm, daphnia, fleas, Assassin bugs (kissing bugs), eye gnats, or rat fleas and woolly bears (e.g., hairy caterpillar).

The compositions and methods are also useful in inhibiting reproduction in the following insects: *Culex quinquefasciatus* (Southern house mosquito), *Drosophila willistoni* (Fruitfly), *Drosophila mojavensis* (Fruitfly), *Drosophila pseudoobscura* (Fruitfly), *Drosophila grimshawi* (Fruitfly), *Drosophila virilis* (Fruitfly), *Drosophila ananassae* (Fruitfly), *Ceratitis capitata* (Mediterranean fruitfly), *Drosophila sechellia* (Fruitfly), *Drosophila persimilis* (Fruitfly), *Drosophila simulans* (Fruitfly), *Drosophila melanogaster* (Fruitfly), *Anopheles darling* (Mosquito), *Anopheles gambiae* (Mosquito), *Musca domestica* (Housfly), *Drosophila erecta* (Fruitfly), *Drosophila yakuba* (Fruitfly), *Aedes aegypti* (Mosquito, yellow fever), *Bombyx mori* (Silk worm), *Tribolium castaneum* (red flour beetle), *Nasonia vitripennis* (wasp), *Apis mellifera* (honeybee), *Apis dorsata* (honeybee), *Megachile rotundata* (alfalfa leafcutter bee), *Dendroctonus ponderosae* (mountain pine beetle), *Bombus impatiens* (common eastern bumble bee), *Bombus terrestris* (bumble bee), *Acromyrmex echinator* (leafcutter ants), *Solenopsis invicta* (fireant), *Apis florea* (honeybee), *Harpegnathos saltator* (Jerdon's jumping ant), *Cerapachys biroi* (ant), *Daphnia pulex* (daphnia), *Camponotus floridanus* (ant), and *Pediculus humanus corporis* (lice).

Blocking insect reproduction

The methods described herein offer a unique and selective approach for controlling insect population without damage to the environment. Inhibiting this enzyme in insects, (e.g., mosquitos) provides a new strategy for insect population control, i.e., reducing the number of insects.

Also provided is the identification of chemical, e.g., small molecule, inhibitors of IYD. The methods described herein involve selective application of an agent within the environment to selectively target insects (e.g., mosquitos) for controlling insect population, i.e., reducing the number of insects. Alternatively, the agent may be selectively placed such that the insect of interest is the only organism susceptible to population control.

One means of population control in insects is an insect growth regulator (IGR), which is a chemical substance that inhibits the life cycle of an insect, and is commonly used to control populations of harmful pests. IGRs prevent an insect from reaching maturity by interfering with its molting process (for example, azadirachtin, hydroprene, methoprene, pyriproxyfen and tritlumuron). This results in immature insects that cannot reproduce.

Hormonal IGRs mimic or inhibit one of the two major hormones involved in insect molting. Chitin synthesis inhibitors prevent the formation of chitin, a carbohydrate used to form the insect's exoskeleton. As described herein, in some cases, the compositions of the invention are administered in conjunction with IGRs.

Additionally, insecticides are substances that are used to kill insects, and are used in agriculture, medicine, industry, and by consumers. Two major groups include systemic and contact insecticides. Systemic insecticides become incorporated and distributed systemically throughout the whole plant. When insects feed on the plant, they ingest the insecticide. Systemic insecticides have activity pertaining to their residue which is called "residual activity" or long-term activity. Contact insecticides are toxic to insects upon direct contact. These can be inorganic insecticides, which are metals and include arsenates, copper and fluorine compounds, which are less commonly used, and the commonly used sulfur. Contact insecticides can be organic insecticides, i.e. organic chemical compounds, synthetically produced, and comprising the largest numbers of pesticides used today, or, they can be natural compounds like pyrethrum, neem oil etc. Contact insecticides usually have no residual activity. Some insecticides kill or harm other organisms in addition to those they are intended to kill. For example, birds may be poisoned when they eat food that was recently sprayed with insecticides or when they mistake an insecticide granule on the ground for food and eat it.

Dosages and frequency of administration of pest-controlling agent

The effective amount of the agent is from 0.001 mg/kg to 250 mg/kg body weight, e.g., 0.001 mg/kg, 0.005 mg/kg, 0.01 mg/kg, 0.05 mg/kg, 1 mg/kg, 5 mg/kg, 10 mg/kg, 25 mg/kg, 50 mg/kg, 75 mg/kg, 100 mg/kg, 125 mg/kg, 150 mg/kg, 175 mg/kg, 200 mg/kg, 225 mg/kg, or 250 mg/kg body weight. The agent is more effective with some insects than others. Amounts of some of the agents may be as low as 2.5 milligrams per kilogram of body weight, and possibly less. Even smaller amounts, for instance as little as 0.05 milligrams per kilogram of body weight are effective when the dosage is continued from day to day as may be desirable in some circumstances.

One treatment (dosage) may also be sufficient to alter such transient insects as mosquitoes and flies. Within a few weeks or months after treatment, it may become apparent that repeated dosages may be required to control insects, e.g., mosquitoes over a long period of time. As such, this type of insect control may be desired in particular circumstances over long periods of time. IYD inhibitors are not contact toxins, i.e., adult insects exposed to IYD

inhibitors will likely not die; however, their reproduction will be blocked. Male mosquitos live ~ 5 days and females can live for weeks, thus, the results of treatment may take days to weeks to detect in mosquitos. The effect depends on the lifetime of each target insect.

Insect traps

The In2Care® Mosquito Trap uses a unique mixture of bio-actives that target not only the mosquito and its larvae, but can also have an impact on the Dengue virus inside the mosquito. Trap activity is not limited to the trap itself, but extends to the surrounding area due to its mosquito-driven larvicide spreading features. The trap actives provide an environmentally-friendly alternative to synthetic pesticides and combines active ingredients that individually are used in other mosquito products which are WHO recommended and/or US-EPA approved.

Similar traps are utilized in the compositions and methods described herein. For example, the compositions are administered within a device, e.g., a “trap,” that attracts an insect to the device in order to administer the compositions described herein to the insect.

In one case, an egg-laying female is attracted to the trap, where it picks up biological agents within the trap to prevent, inhibit, or reduce insect reproduction in a male insect. In other cases, a male insect is attracted to the trap, where it picks up biological agents within the trap to prevent, inhibit, or reduce insect reproduction. Preferably, the agent inhibits spermatogenesis (e.g., late stage spermatogenesis). For example, the agent that inhibits spermatogenesis in the insect comprises an agent that inhibits expression or activity of IYD in the insect, e.g., the agent inhibits expression or activity of IYD in a testis of the insect.

EXAMPLES

Example 1: Protein co-crystal structure of mouse IYD in complex with moniodotyrosine (IYD-MIT) identified key amino acids

The structure of IYD from mouse and human identified key amino acids that were used to query databases of genomic information to identify the presence of the IYD gene in other organisms (Phatarphekar, A. et al., *Mol. Biosys.* 2014, 10, 86-92). The co-crystal structure of mouse IYD-MIT showed that E153, Y157 and K178 were key residues for substrate recognition (Figure 2A and Figure 2B). T235 provided an important hydrogen bond to the N5 of FMN (Flavin mononucleotide) (Figure 2B). These residues provided markers to identify IYD homologs through the tree of life. The key interacting residues, E153, Y157 and K178 were located on one monomer of the dimer, whereas the FMN molecule was

positioned between the monomers, and the adenine tail further extended into the adjacent monomer. Hydrophobic interactions between the phenyl of the MIT (monoiodotyrosine) and the flavin rings of FMN may contribute to the overall protein-ligand interaction. The structural results indicated that a small set of residues was sufficient to specify IYD activity.

Example 2: IYD Homologs are present in diverse organisms

A sequence alignment between several different organisms (human, mouse, zebrafish, lancelet, drosophila, daphnia, sea anemone, hydra, bacteria, archaea and bacterial BluB) showed the presence of IYD homologs (Figure 3). All vertebrates and invertebrates sequenced contain IYD. Several key residues were conserved among the organisms. For example, a tryptophan residue in the active site lid, which was responsible for substrate binding. Additionally, a glycine residue within the active site lid is also conserved among the organisms. Determinant IYD residues (glutamic acid, tyrosine, and lysine) were conserved among the organisms and are outlined in red in Figure 3.

Example 3: IYD Exhibits deiodinase activity

Selected orthologs were expressed in *E. coli* and were tested for deiodination activity. All of the orthologs displayed similar catalytic efficiency for deiodination of I₂-Tyr with k_{cat}/K_m values within an order of magnitude of each other (Table 1). However, prior to the invention described herein, the function of halogenated tyrosines and the biological role of IYD was unknown in organisms such as invertebrates that are not known to produce TH. See, Phatarphekar, A., et al., Mol. BioSyst. 2014, 10, 86-92, incorporated herein by reference, for a discussion linking iodotyrosine deiodinase genes with deiodinase activity in organisms not known to require iodide or respond to the iodide-containing hormone thyroxine (including the first insect, honey bee).

All the proteins containing the signature residues (E153, Y157 and K178) promoted deiodination of DIT (diiodotyrosine), with similar K_m , k_{cat} and k_{cat}/K_m values (Figure 4). For example, the k_{cat}/K_m in humans was $0.40 \pm 0.08 \mu\text{M}/\text{min}$, and $0.56 \pm 0.09 \mu\text{M}/\text{min}$ in drosophila. The closest structural homolog, BluB (vitamin B₁₂ biosynthesis), did not exhibit deiodinase activity (i.e., the K_m for diiodotyrosine was $\geq 133 \mu\text{M}$). BluB, however, does not contain the signature, key residues (E153, Y157 and K178). Steady-state kinetic characterization (K_m , k_{cat} and k_{cat}/K_m) of dehalogenation catalyzed by IYD homologs (intermediate and soluble domains) from representative organisms from diverse phyla is shown in Table 1. Kinetic constants were determined by quantification of ¹²⁵I- released from [¹²⁵I]-I₂-Tyr (Goswami, A.; Rosenberg, I. N., *Endocrinology* 1977, 101, 331-341, Gnidehou,

S. et al., *FASEB J.* 2004, 18, 1574-1576 and Rokita, S. E. et al., *Biochimie* 2010, 92, 1227-1235). Reductive deiodination of iodotyrosine was common to a diverse set of organisms.

Table 1: Kinetic characterization of IYD homologs (intermediate and soluble domains)

Organism	K_m (μM)	k_{cat} (min^{-1})	k_{cat}/K_m ($min^{-1}\mu M^{-1}$)
Human ^a	31 ± 6	12.5 ± 1.0	0.4 ± 0.10
Mouse ^b	9 ± 1	4.5 ± 0.7	0.49 ± 0.09
Zebrafish ^c	8 ± 1	4.1 ± 0.2	0.51 ± 0.07
Lancet ^c	6 ± 3	7.0 ± 1.0	0.14 ± 0.04
Drosophila ^d	13 ± 2	7.1 ± 0.2	0.55 ± 0.09
Honeybee ^c	29 ± 7	8.0 ± 1.0	0.28 ± 0.07
Daphnia ^c	7 ± 1	17.6 ± 0.8	2.50 ± 0.40
Sea Anemone ^c	105 ± 26	32.0 ± 4.0	0.30 ± 0.08
Hydra ^c	4 ± 1	13.0 ± 1.0	3.50 ± 0.90
Haliscomenobacter hydrossis ^c	6.6 ± 0.9	5.4 ± 0.3	0.8 ± 0.10
Archaea (60°C) ^d	1.8 ± 0.3	5.5 ± 0.2	3.10 ± 0.5
Archaea (25°C)	1 ± 0.1	0.44 ± 0.08	0.4 ± 0.1

^aSee, Hu, J.; et al., *J. Biol. Chem.* 2015, 290, 590-600. ^bSee, Thomas, S. R. et al., *J. Biol. Chem.* 2009, 284, 19659-19667. ^cSee, Phatarphekar, A. et al., *Mol Biosys.* 2014, 10, 86-92.

IYD orthologs were present in all 13 species of fruit fly including *Drosophila melanogaster* that have been sequenced to date. Orthologs of IYD were also found in most all genomes of other insects including the disease carrying vectors such as *Anopheles gambiae* (malaria) and *Aedes aegypti* (yellow fever). Using *Drosophila* as a genetic model, the biological role of IYD in invertebrates was investigated.

Example 4: Tissue distribution of IYD in *Drosophila*

The tissue expression of IYD mRNA in *Drosophila melanogaster* (fly) was disproportionately expressed in the testes as compared to other tissues (e.g., the male accessory gland, the ovaries, the trachea, the heart, and salivary glands) (Figure 5). Tissue specific mRNA expression profiling indicated very high levels of IYD (gene CG6279 in flybase) in adult male testes (Figure 5) (Chintapalli, V. R.; Wang, J.; *Nat. Genet.* 2007, 39, 715-720). The high IYD expression in the testes was observed in adult *Drosophila*, as compared to larva which are nearly devoid of IYD. High levels of expression in the testes was surprising, and

was further investigated (Figure 8). CG6279 was strongly expressed in testis during later stages of germline development, especially post differentiation (after meiosis).

IYD is a membrane bound enzyme in humans, and is primarily expressed in thyroid tissue. In humans, IYD assists in the recovery of iodide from mono- and diiodotyrosine (MIT, DIT) generated during thyroxine biosynthesis. IYD is critical for spermatogenesis in insects, but not mammals. Thus, the lack of IYD in mammalian testes makes IYD an attractive target for controlling insect populations without harming mammals, including humans.

Example 5: RNAi knockdown of IYD in *Drosophila* testis

Selective suppression of a chosen gene *in vivo* can be performed through applications of RNAi. Application of RNAi in *Drosophila* provided the first confirmation that IYD was directly involved in spermatogenesis. As described herein, suppression of this gene produced male sterility and developmental anomalies as evident when imaging the testes. Similar effects are expected in other Diptera (e.g., flies, mosquitos, etc.) since all insects sequenced to date contain a gene for IYD. In particular, IYD from *Drosophila* and *Anopheles* share 67% identity and 82% similarity.

RNAi-based suppression using double-stranded RNA (dsRNA)

A standard method for inducing RNAi-based suppression in *Anopheles* is to inject double-stranded RNA (dsRNA) directly into the body cavity. Thus, a series of dsRNAs complementary to the IYD gene are prepared and tested for their ability to suppress IYD *in vivo*. Not all dsRNA have the same efficiency to suppress the gene and hence multiple dsRNA are tested. If fertility does not diminish with this treatment, the expression of IYD in testes is measured with qPCR to confirm its presence. Additionally, direct injection may not necessarily suppress genes in the testes.

If necessary, dsRNA is also injected along with a fluorescent marker gene into embryos to generate transgenic species with the innate ability to suppress IYD expression. As described herein, a similar strategy was used in *Drosophila* and placed RNAi suppression of IYD under control of a series of promoters induced at different stages of spermatogenesis. Loss of IYD activity is observed *in vivo* as a loss of fertility and fecundity. Again, qPCR is used to correlate gene expression with the observed phenotype.

Promoter dependent knockdown of IYD

Expression of IYD mRNA was decreased using promotor dependent knockdown (Figure 6A, Figure 6B, and Figure 6C). Promoters used for IYD knockdown are shown in Table 2.

Table 2: Promoters used for IYD knockdown

Promoter	Cell Type	Expression Stage	Fertility, Fecundity affected?
nos	germ	early	no
bam	germ	late	no
c587	somatic	early	no
tj	somatic	early	no
eya	somatic	late	yes

As described herein, *Drosophila* IYD plays a key role in spermatogenesis, and disruption of IYD dramatically lowered *Drosophila* fertility and fecundity (Figure 6B). IYD was found to be crucial to sperm development, and suppression by RNAi disrupted the elongation of spermatocytes and blocked sperm formation. The average number of progeny in *eya*-GAL4/UAS-dmIYD-RNAi resulted in 7 sterile *Drosophila* samples, whereas the control *Drosophila* testes, *eya*-GAL4 and UAS-dmIYD-RNAi resulted in no sterile *Drosophila* (Figure 6B). The additional promoters evaluated, *nos*, *bam*, *c587*, and *tj* did not affect fertility and fecundity in *Drosophila*.

Iodotyrosine may generate as a signal during maturation, and thus IYD may be responsible for turning the signal off. These RNAi data provide an indication that insects may use iodination/deiodination of tyrosine for regulating certain developmental processes.

Example 6: Late stage development is sensitive to IYD

Spermatogenesis in *Drosophila* begins with mitotic division (alternatively, spermatogonia), of germ cells to somatic cells (Figure 7A). Before the transition to meiotic division, the expression of dmIYD (*Drosophila*IYD) is at its peak. Within meiotic division, a spermatocyte evolves into a round spermatide, at which point developmental abnormalities ensue. Elongated spermatids are generated following the formation of the round spermatids.

After RNAi degradation of IYD mRNA, abnormalities were detected in *Drosophila* testis (Figure 7B). In control testis, nearly 100% showed no phenotype, whereas in IYD RNAi samples, over 80% of the testis showed an abnormal phenotype.

Using phase contrast images of *Drosophila* testis, a considerable difference in control samples versus RNAi knockdown of IYD samples was highlighted (Figure 7C). In control

samples, a sperm reservoir was developed, whereas in the RNAi sample, no sperm reservoir was developed. The reservoir serves to ensure successful fertilization by providing the appropriate number of sperm in in the appropriate physiological state for fertilization. Development of the sperm reservoir is distinctive of late stage development. No sperm reservoir was developed in IYD RNAi drosophilatestis, indicating that suppression of dmIYD disrupted late stage spermatogenesis.

Example 7: Strong expression of isoforms A and B of CG6279 in late stages of spermatogenesis.

Two isoforms have been reported for *Drosophila* IYD (dmIYD) in the NCBI databases. These two are related by alternative splicing of the CG6279 gene. Isoform B resembles IYD orthologs from other organisms and includes a membrane domain, an intermediate domain and a catalytic domain (Phatarphekar, A. et al., *Mol. Biosys.* 2014, 10, 86-92 and Friedman, J. E. et al. *J. Biol. Chem.* 2006, 281, 2812-2819). Isoform A is unique to the *Drosophila* genus by the added presence of a large N-terminal domain (470 amino acids) that does not coincide with any known protein domains listed in the conserved domain database (CDD).

In situ hybridization experiments were conducted with male *Drosophila* testes, using sequence specific probes (riboprobes) targeting distinct regions of isoform A and B of CG6279 to identify the mRNA expression pattern as well as the stage of mRNA expression during spermatogenesis. *In situ* hybridization revealed strong expression during the later stages of spermatogenesis, and both riboprobes revealed similar expression patterns (Figures 9A, Figure 9B, Figure 9C, and Figure 9D).

Example 8: Binding affinities of dmIYD Isoform B using various dehalogenation ligands.

Isoform B of the drosophila enzyme (dmIYD) has been cloned, expressed and purified to homogeneity following standard protocols (Phatarphekar, A. et al., *Mol. Biosys.* 2014, 10, 86-92). This enzyme displayed similar binding affinities for iodo-, diiodo-, bromo- and chlorotyrosine (I-Tyr, I₂-Tyr, Br-Tyr and Cl-Tyr, respectively), but a 30-fold lower affinity for fluorotyrosine (F-Tyr), as shown in Table 3. Error represents the standard deviation from an average of 3 independent experiments.

Table 3: Dissociation constants (K_D) for halotyrosines with dmIYD

Ligand (dmIYD)	K _D (μM)
I ₂ -Tyr	0.55 ± 0.08
I-Tyr	0.62 ± 0.09

Br-Tyr	0.47 ± 0.06
Cl-Tyr	0.46 ± 0.08
F-Tyr	17.60 ± 0.80

The enzyme also catalytically dehalogenated all the halotyrosine ligands but F-Tyr. The catalytic efficiency (k_{cat}/K_m) for deiodination and debromination was similar and both were 3-4 fold more efficient than dechlorination, as shown in Table 4. (*) indicated I₂-Tyr dehalogenation measured by [¹²⁵I]-iodide released from [¹²⁵I]-I₂-Tyr.

Table 4: Steady-state kinetics of dehalogenation catalyzed by dmIYD

Ligand (dmIYD)	K _m (μM)	k _{cat} (min ⁻¹)	k _{cat} /K _m (min ⁻¹ μM ⁻¹)
I ₂ -Tyr*	13.2 ± 2.0	7.1 ± 0.2	0.55 ± 0.09
I-Tyr	14.4 ± 4	7.3 ± 0.6	0.50 ± 0.20
Br-Tyr	8 ± 2	4.2 ± 0.3	0.50 ± 0.10
Cl-Tyr	$21. \pm 6$	3.0 ± 0.3	0.14 ± 0.14
F-Tyr	N/A	<0.002	N/A

Substrate turnover measured by production of tyrosine using HPLC separation and detection

Catalytic deiodination of halotyrosines were measured by detecting and quantifying the tyrosine produced in presence of enzyme using reverse phase HPLC. The buffer used for reaction is the same as that used for the [¹²⁵I]-release assay as well as for equilibrium binding measurements (100 mM potassium phosphate, 200 mM KCl at pH 7.4). Each reaction (final volume of 1 ml) was initiated by addition of 0.5% dithionite (100 μl of 5% dithionite in 5% aqueous sodium bicarbonate) rather than the 1% dithionite used for the [¹²⁵I]-release assay. This change minimized interference between tyrosine and dithionite during HPLC separation. The total enzyme used per reaction was 0.08 μM and the duration of the reaction was 15-20 mins. M-cresol (30 μM) was added to each reaction tube as the internal standard (IS). The reaction was quenched with formic acid (5% final concentration) and the entire reaction mixture was injected manually onto a Microsorb MV 300-5 C18 column analytical HPLC column (Agilent) protected with a manually packed C18 silica guard column. The HPLC method used for analysis is provided in Figure 10.

The concentration of tyrosine produced by IYD was determined from a standard curve in which known concentrations of tyrosine were spiked into reaction mixtures not

containing the halotyrosine but otherwise identical. The ratio of the areas of tyrosine peak to the internal standard peak was plotted against the concentration of tyrosine spiked into the reaction and the best fit line was determined by linear regression. This equation of the line was used to determine the concentration of tyrosine produced in μM from a ratio of peak areas of tyrosine to the internal standard.

Example 9: Site-directed mutagenesis on dmIYD identified active site amino acid residues critical for catalysis

Site-directed mutagenesis studies on dmIYD were conducted to identify active site amino acid residues that are critical for catalysis. The E154Q mutation was most detrimental to catalytic efficiency of dmIYD, as shown in Table 5 and may provide a functional knockout mutation in continuing efforts to define the full role of dmIYD *in vivo*. Kinetic constants were determined by [^{125}I]-iodide released from [^{125}I]-I₂-Tyr.

Table 5: Steady-state kinetics for dehalogenation catalyzed by active site mutants of dmIYD.

Mutant	K _m (μM)	k _{cat} (min^{-1})	k _{cat} /K _m ($\text{min}^{-1} \mu\text{M}^{-1}$)
E154Q	3400 $\pm$ 500	2.4 $\pm$ 0.2	0.0007 $\pm$ 0.0001
K179Q	700 $\pm$ 100	7.7 $\pm$ 0.5	0.0110 $\pm$ 0.0020

Example 10: The significance of the CG6279 for spermatogenesis is confirmed using CRISPER-CAS9 and site-directed mutation studies

Genetic experiments are conducted using CRISPR-CAS9 technology to remove the entire CG6279 gene from the *Drosophila* genome. Additionally, a loss of function mutation with an E154Q mutation at the genomic level is generated. These experiments confirm the significance of the CG6279 gene for spermatogenesis.

Example 11: Feeding studies using halotyrosines are performed to determine their effects on fertility and spermatogenesis

Feeding studies are performed using halotyrosines added to the standard food for drosophila to determine their effects on fertility and spermatogenesis. This determines whether or not dietary supplements mimic the effect of inhibiting IYD since both allow an unnatural high concentration of halotyrosine to accumulate *in vivo*.

Example 12: Use of small molecule inhibitors in feeding traps to inhibit reproduction of insects

The feasibility of controlling insect (e.g., mosquito) population by an inhibitor of IYD is assessed through a combination of chemistry, biochemistry and biology. The most

efficient inhibitor of IYD requires a 3-step synthesis that is available from the literature. This compound is a pyridone analog of iodotyrosine and previously designed as a mimic of an intermediate formed during enzyme catalysis. Its high affinity was originally measured with porcine IYD. However, the active site structure and catalytic mechanism of all IYDs are similar enough that this inhibitor is effective against IYD from insects. This compound along with others available commercially is used for inhibition studies *in vitro* and *in vivo*.

The sequence of IYD from *Anopheles gambiae* is available, but has not yet been studied. Data assembled from other organisms strongly predict its ability to catalyze deiodination of I_n-Tyr (n= 1,2). The gene is synthesized commercially, expressed heterologously and characterized for its catalytic specificity. Additionally, the enzyme's sensitivity to the known inhibitors, nitrotyrosines and the pyridone derivative, is measured *in vitro* to confirm their potency before use as feeding supplements.

Feeding studies to measure the effectiveness of simple inhibitors to disrupt fertility

The ability to control mosquito fertility in the field is expedited by easy access to inhibitors of IYD already described in the literature. Nitro- and dinitrotyrosine were identified decades ago as potent inhibitors of IYD *in vitro*, and more recently a pyridone analog of iodotyrosine was found to maximize affinity to IYD based on the mechanism of catalysis common to all orthologs. These compounds are purchased and synthesized respectively for supplementing mosquito food and are even be tested by direct injection into the body cavity if necessary. Measuring a dose-dependent response establishes the minimum concentration necessary to observe the diminished fertility and fecundity. Since suppression of IYD and the presumed buildup of I- and I₂-Tyr are detrimental to *Drosophila*, mosquitos are also fed with media containing increasing amounts of I- and I₂-Tyr in an attempt to overwhelm the catalytic capacity of the native IYD. These treatments recapitulate the expected results of the RNAi investigation.

The most cost effective supplement that suppresses sperm development is then tested in the field. If disruption of I- and I₂-Tyr levels is a basis for the sterility, then inhibiting the initial formation of I and I₂-Tyr also block spermatogenesis. Biosynthesis of such halogenated amino acids is typically promoted by a peroxidase. Consequently, peroxidase inhibitors such as thiourea, methimazole and propylthiouracil are tested as dietary supplements to inhibit fertility as well. Already, methimazole and propylthiouracil are used as safe and effective drugs for controlling hyperthyroidism. They act by inhibiting the iodination step of thyroxine biosynthesis.

Utilizing commercially available female mosquito traps (e.g., In2Care®), an agent that reduces activity or expression of IYD is added to the media to inhibit reproduction of subsequent generations of mosquitos. Larvae eat much more than adults, so spiking a trap like those of "In2Care®" will reach the larvae. Male mosquitoes live on sugar water (e.g., rotting fruit), so in some cases, an agent that reduces activity or expression of IYD is added to a liquid composition to inhibit the reproduction of male mosquitoes.

OTHER EMBODIMENTS

While the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

The patent and scientific literature referred to herein establishes the knowledge that is available to those with skill in the art. All United States patents and published or unpublished United States patent applications cited herein are incorporated by reference. All published foreign patents and patent applications cited herein are hereby incorporated by reference. Genbank and NCBI submissions indicated by accession number cited herein are hereby incorporated by reference. All other published references, documents, manuscripts and scientific literature cited herein are hereby incorporated by reference.

While this invention has been particularly shown and described with references to preferred embodiments thereof, it will be understood by those skilled in the art that various changes in form and details may be made therein without departing from the scope of the invention encompassed by the appended claims.

What is claimed is:

1. A method of preventing, reducing, or inhibiting insect reproduction comprising:
administering to the insect a composition comprising an agent that inhibits spermatogenesis in the insect,
thereby preventing, reducing, or inhibiting reproduction in the insect.
2. The method of claim 1, wherein the agent that inhibits spermatogenesis in the insect comprises an agent that inhibits expression or activity of iodotyrosine deiodinase (IYD) in the insect.
3. The method of claim 2, wherein the agent inhibits expression or activity of IYD in a testis of the insect.
4. The method of claim 3, wherein the IYD inhibits late stage spermatogenesis.
5. The method of claim 1, wherein the method reduces a population size of the insect.
6. The method of claim 1, wherein fecundity of the insect is decreased.
7. The method of claim 1, wherein fertility of the insect is decreased.
8. The method of claim 1, wherein the agent for inhibiting expression or activity of IYD comprises an antibody or fragment thereof, a binding protein, a polypeptide, or any combination thereof.
9. The method of claim 1, wherein the agent for inhibiting expression or activity of IYD comprises a small molecule.
10. The method of claim 9, wherein the small molecule for inhibiting expression or activity of IYD comprises 4-OH-2',3,4',5,6'-PCB; 4-OH-2',3,4',6'-TCB; 4-OH-2,2',5,5'-TCB; 4,4'-diOH-3,3',5,5'-TCB; 3-OH-2,2',5,5'-TCB; 4'-OH-BDE-17; 4-OH-BDE-42; 4'-OH-BDE-49; 4-OH-BDE90; 2-OH-BDE-15; 2'-OH-BDE-28; Bromoxynil; Oxyaclozanide; Tribromsalan; Closantel; Triclosan; Bithionol; Nitroxylin; Benzbromarone

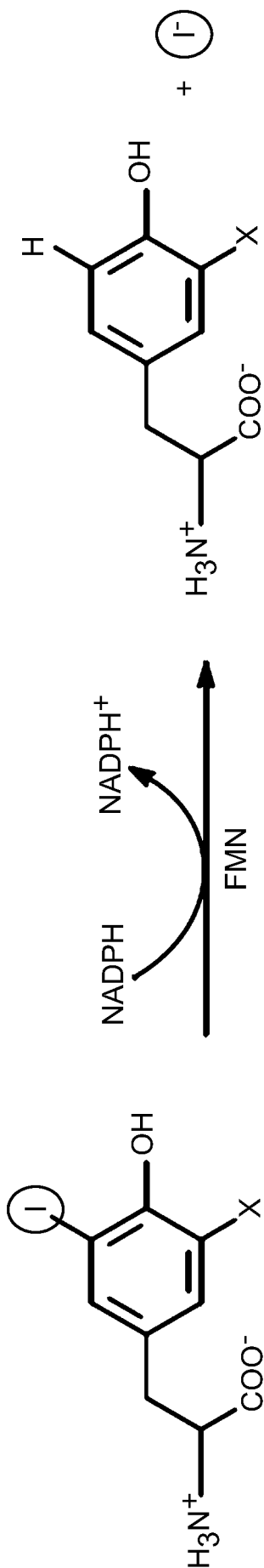
Rose Bengal; Erythrosine B; Phloxine B; D,L-3-(2-pyridon-5-yl)alanine; and D,L-3-(N-methyl-2-pyridon-5-yl)alanine; D,L-3-(N-ethyl-2-pyridon-5-yl)alanine; and D,L-3-(N-isopropyl-2-pyridon-5-yl)alanine.

11. The method of claim 1, wherein the agent comprises a nucleic acid molecule.
12. The method of claim 11, wherein the nucleic acid molecule comprises dsRNA, shRNA, or antisense RNA, or any portion thereof.
13. The method of claim 1, wherein the agent is administered as a spray.
14. The method of claim 1, wherein the agent is administered topically, orally, via inhalation, or via injection.
15. The method of claim 12, wherein the composition is in the form of a gas, a liquid, a solid, or a semi-solid.
16. The method of claim 1, wherein the agent is soluble in water.
17. The method of claim 1, wherein the agent is nontoxic.
18. The method of claim 14, wherein the composition comprises sugar water.
19. The method of claim 1, wherein the agent is administered at a concentration of 0.001 mg/kg to 250 mg/kg of body weight
20. The method of claim 1, wherein the agent is administered once per day.
21. The method of claim 1, wherein the insect comprises a cockroach, a beetle, an earwig, a fly, a cicada, a hopper, an ant, an aphid, a bed bug, a bee, a wasp, a mantid, an earthworm, a terrestrial crab, a firebrat, a flea, a snail, a slug, a cricket, a hornet, a locust, lice, a moth, a red spider, a silverfish, a woollice, a termite, a tick, a wasp, a mosquito, a gnat, a millipede, a

stink bug, a weevil, a mite, a silk worm, a daphnia, a flea, an assassin bug, an eye gnat, a rat flea or a woolly bear (hairy caterpillar).

22. The method of claim 1, wherein the insect comprises *Culex quinquefasciatus*, *Drosophila willistoni*, *Drosophila mojavensis*, *Drosophila pseudoobscura*, *Drosophila grimshawi*, *Drosophila virilis*, *Drosophila ananassae*, *Ceratitis capitata*, *Drosophila sechellia*, *Drosophila persimilis*, *Drosophila simulans*, *Drosophila melanogaster*, *Anopheles darling*, *Anopheles gambiae*, *Musca domestica*, *Drosophila erecta*, *Drosophila yakuba*, *Aedes aegypti*, *Bombyx mori*, *Tribolium castaneum*, *Nasonia vitripennis*, *Apis mellifera*, *Apis dorsata*, *Megachile rotundata*, *Dendroctonus ponderosae*, *Bombus impatiens*, *Bombus terrestris*, *Acromyrmex echinator*, *Solenopsis invicta*, *Apis florea*, *Harpegnathos saltator*, *Cerapachys biroi*, *Daphnia pulex*, *Camponotus floridanus*, or *Pediculus humanus corporis*.

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X = H (monoiodotyrosine, MIT), or I (diiodotyrosine, DIT)

FIG. 1

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FIG. 2A

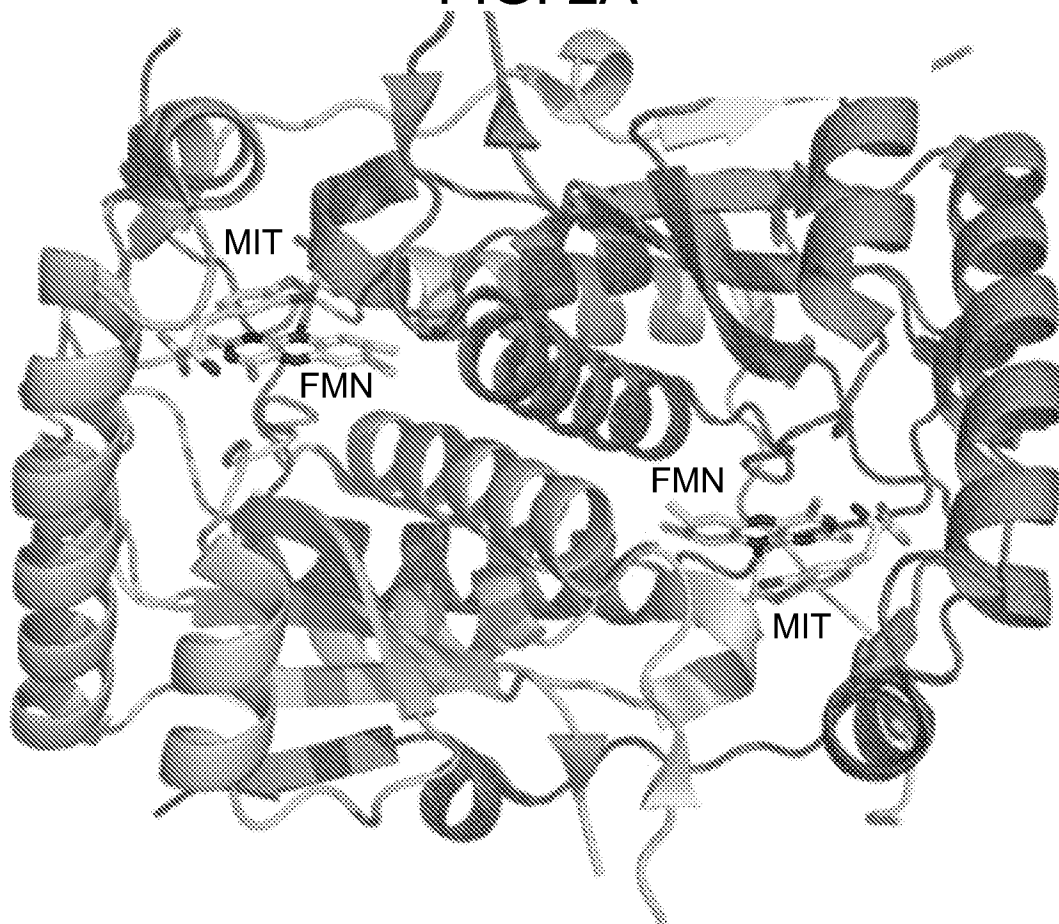


FIG. 2B

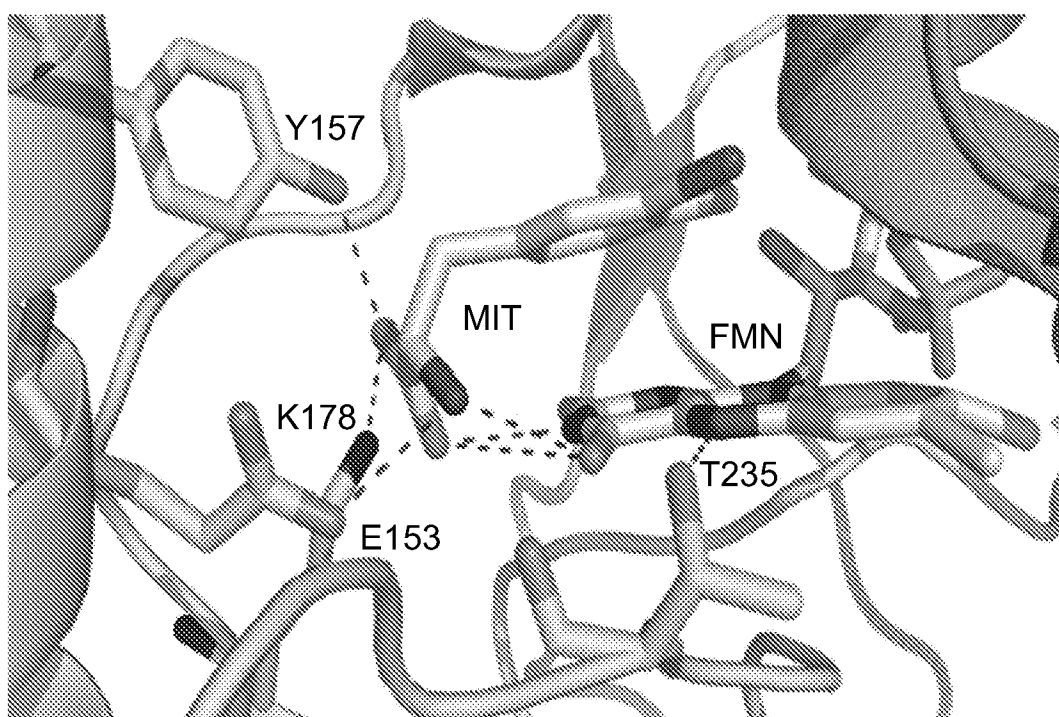


FIG. 3A

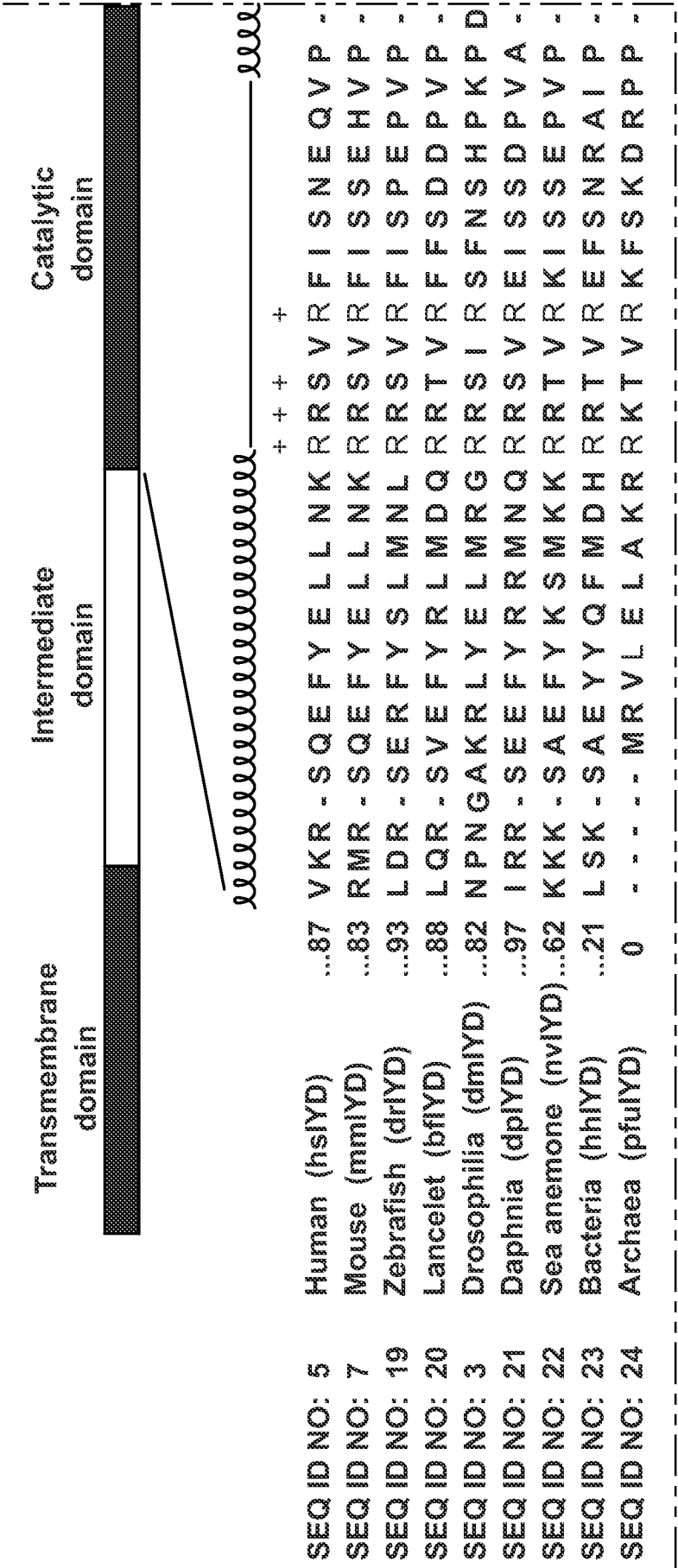
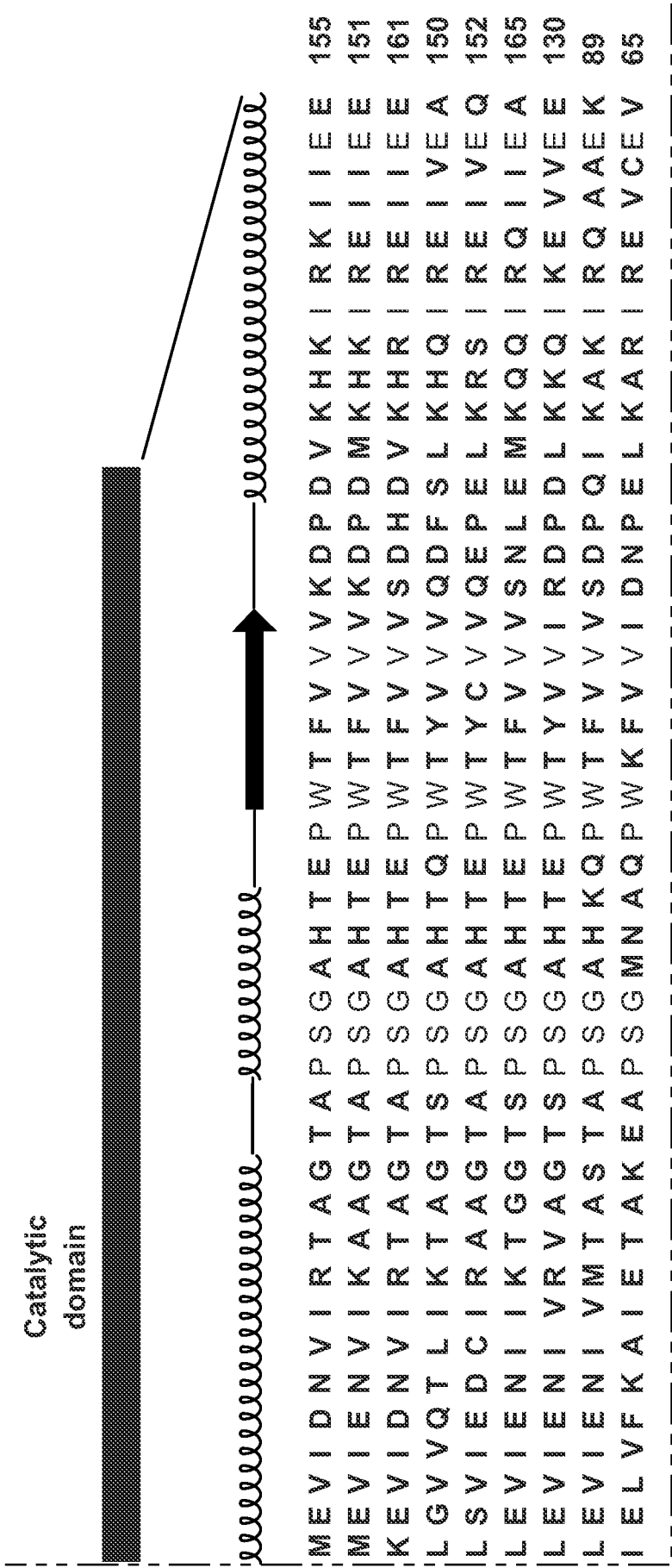


FIG. 3B



		*	*	active site lld - substrate binding
Human (hsIYD)	156	EE	EE	IN YMKRMG - - HRWVTDLKKLRTNW I
Mouse (mmIYD)	152	EE	EE	IN YMKRMG - - KRWVTDLKKLRTNW I
Zebrafish (drIYD)	162	EE	EE	IN YKQRMG - - NKWVQDLKRLRTNWV
Lancelet (blIYD)	157	EE	EE	IN YRKRM S - - ATWVKDLEKFR TTWE
Drosophila (dmIYD)	153	EE	EL	VNYSQRMH - - PQWVTDLRPLQTNHV
Daphnia (dpIYD)	166	EE	EE	IN YKQRMG - - DVWVQDLQPVGTTWV
Sea anemone (nvIYD)	131	EE	EQ	LN YARRMG - - EKWVQDLSI LKTIWS
Bacteria (hhIYD)	90	EE	EF	ES YNGRMS - - NEWLEDLQPF GTDWH
Archaea (pfuIYD)	66	EE	EK	FYERI KGELKEWLVE NEFTPE - -

FIG. 3D

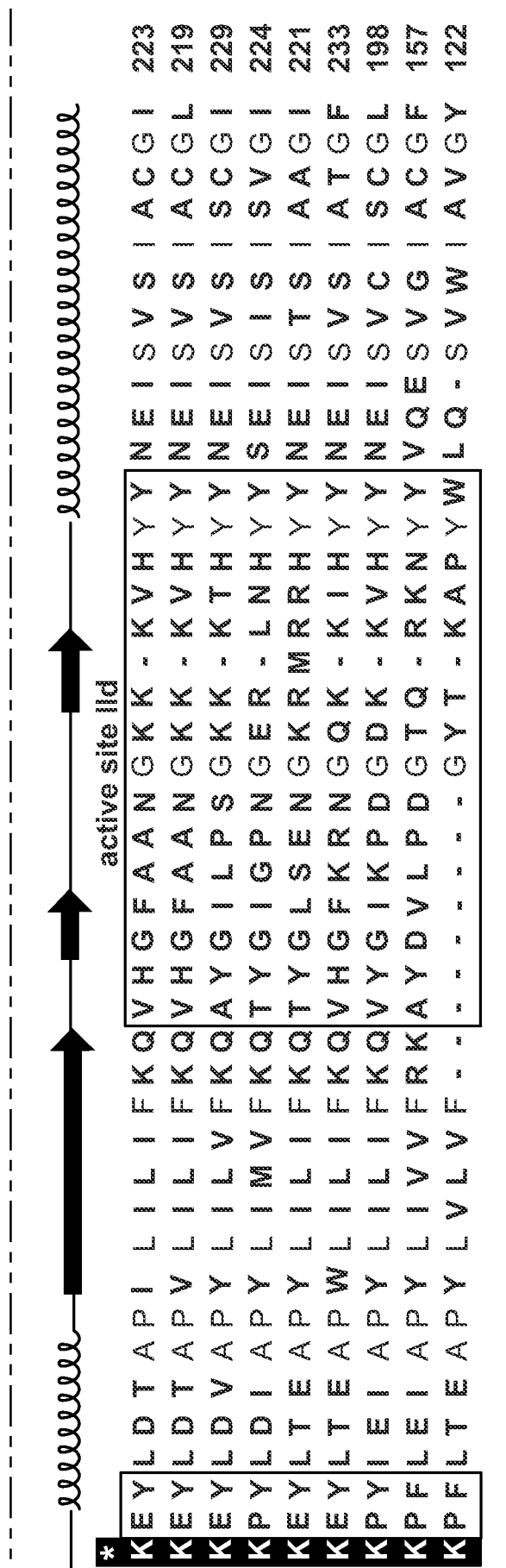


FIG. 3E

		
Human (hsiYD)	183 L L A A L Q N A G L V T V T T T T	+ P L N C G P R L R V L L
Mouse (mmiYD)	220 L L A A L Q N A G L V T V T T T T	P L N C G P R L R V L L
Zebrafish (driYD)	230 L L A A L Q N A G L V T V T T T T	P L N C G P Q L R S L L
Lancelet (bfiYD)	225 L L A A I Q N A G L V T V T S T T	P L N A G P A L R T L L
Drosophila (dmiYD)	222 L L C A L Q A A G L A S L V T T T	P L N C G P A L R N L L
Daphnia (dplYD)	234 L L A A I Q E A G L V T V T T T T	P L N C G P S I R V L L
Sea anemone (nviYD)	199 L L A A I Q N A G L V T V T S T T	P M N A G P R L R V L L
Bacteria (hhiYD)	158 L L A A I H Q A G L V A L T H T	P S P M N - F L Q K I L
Archaea (pfiYD)	123 F L L A L E E V E L G T V T Y T	P P N T R P - I E E L L

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FIG. 4

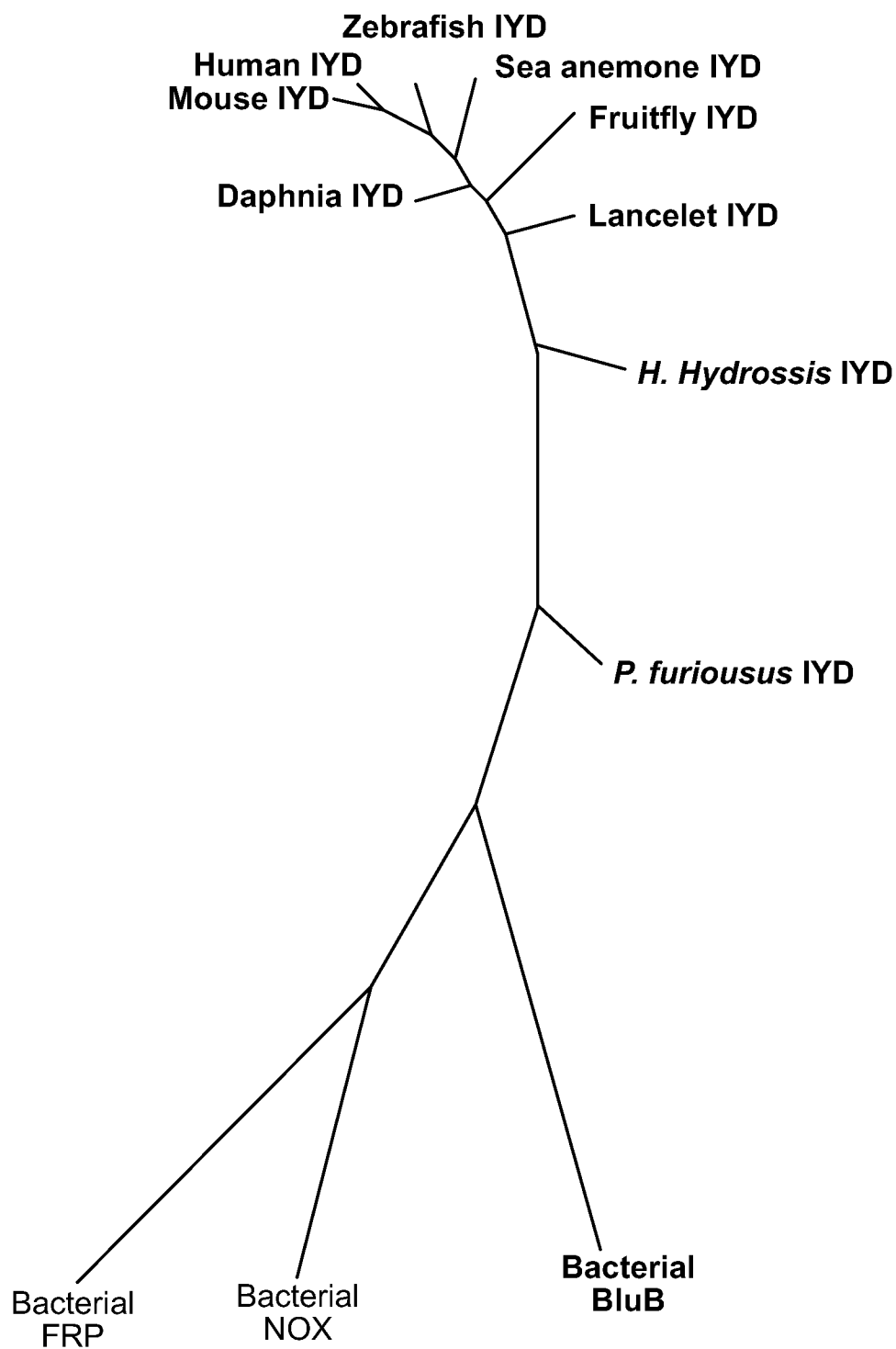
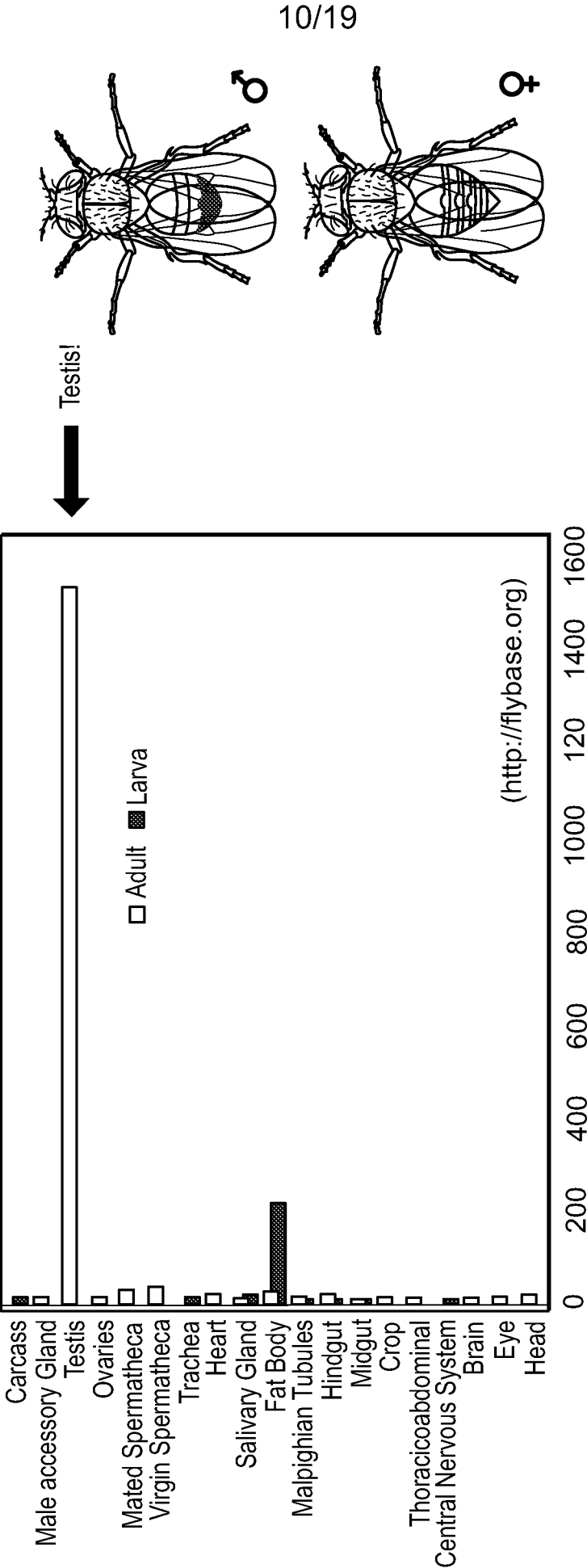


FIG. 5



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FIG. 6A

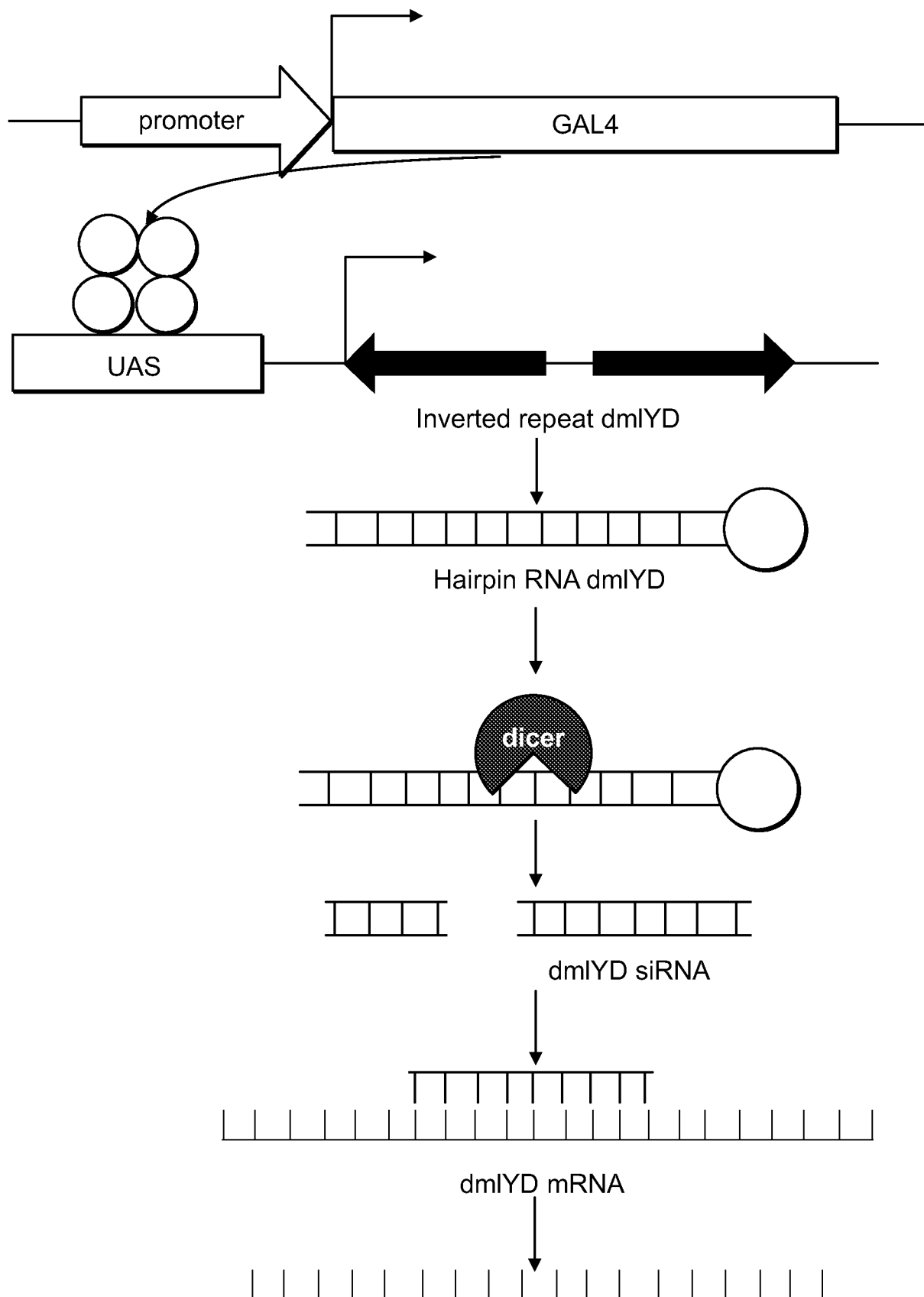


FIG. 6B

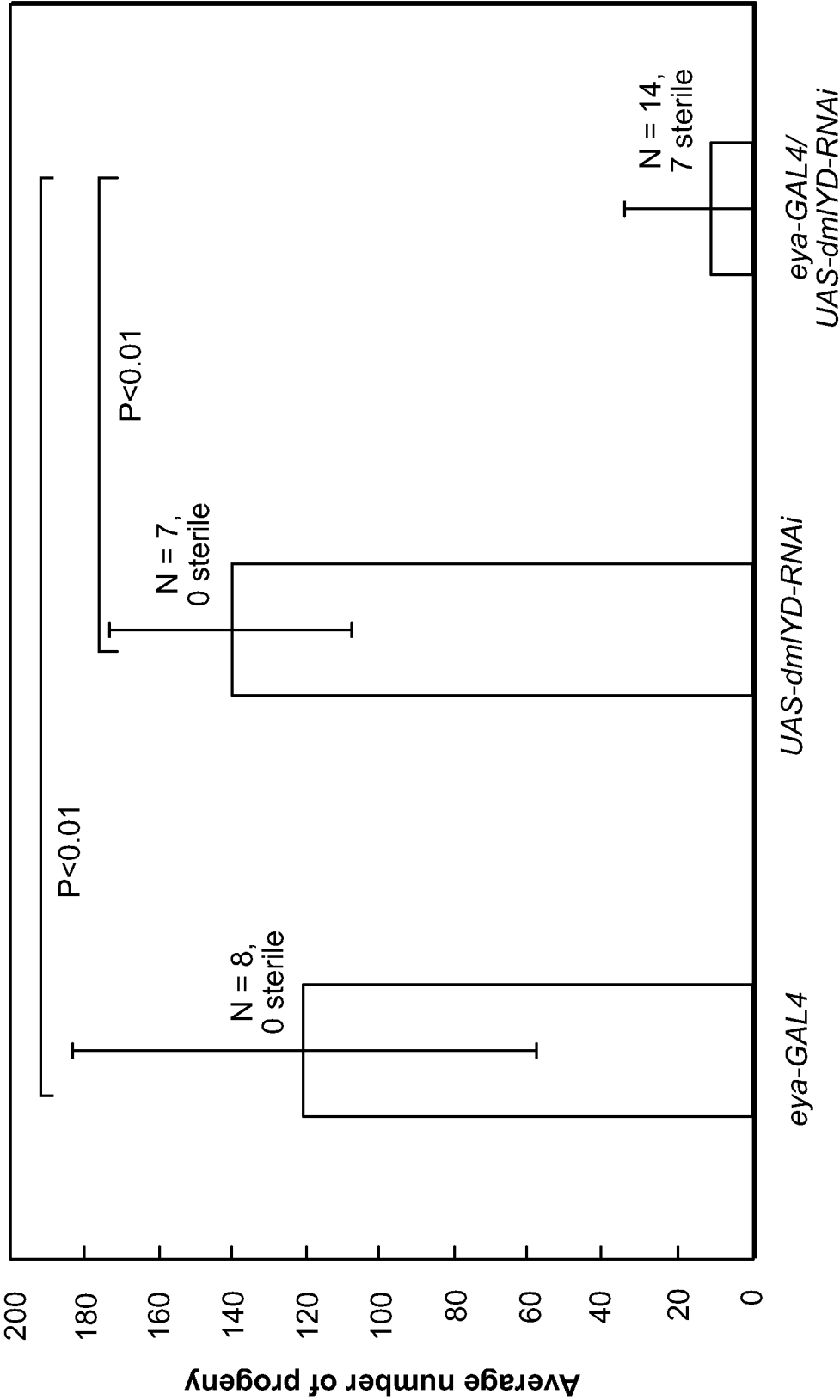
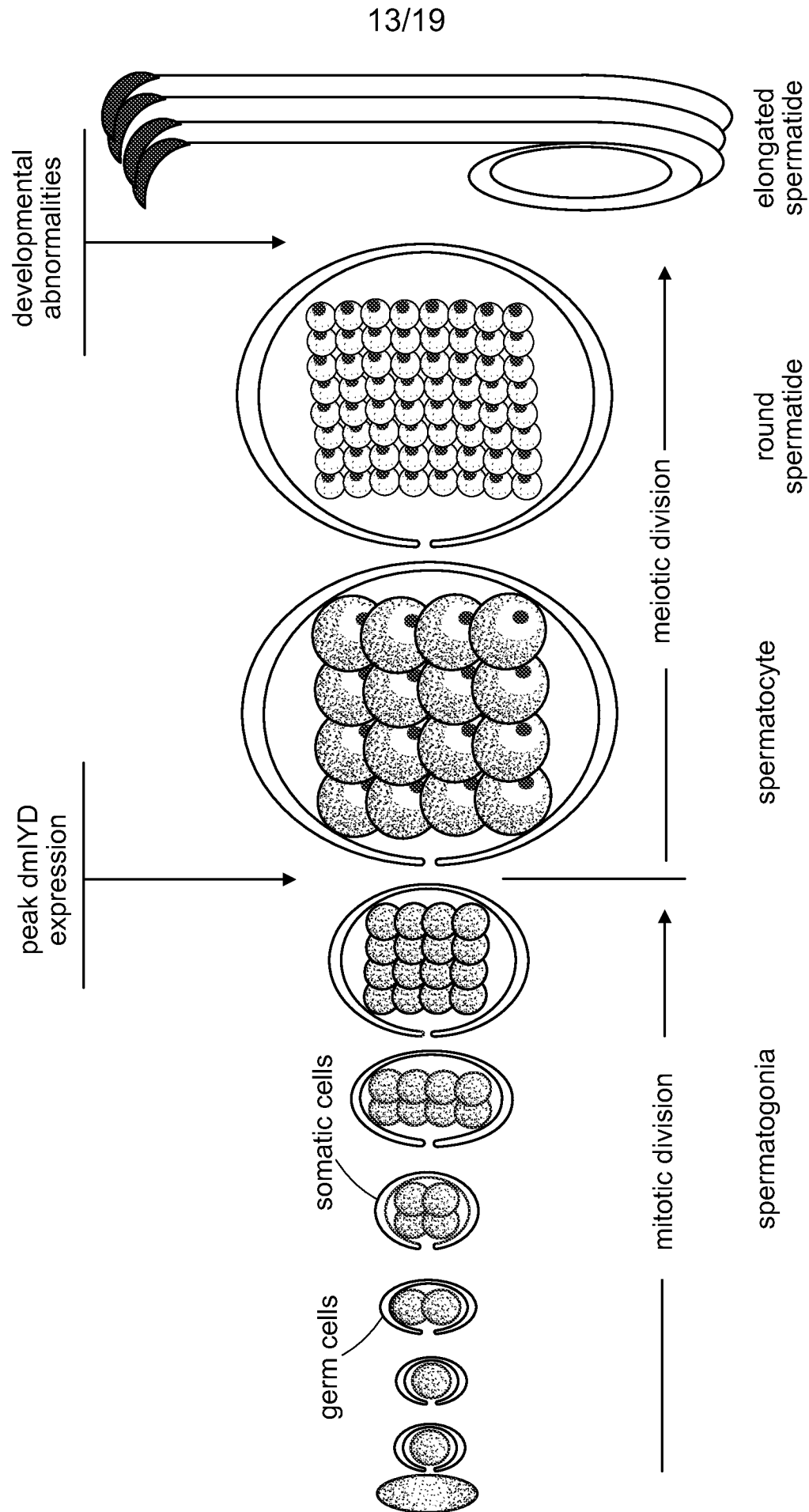


FIG. 7A



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FIG. 7B

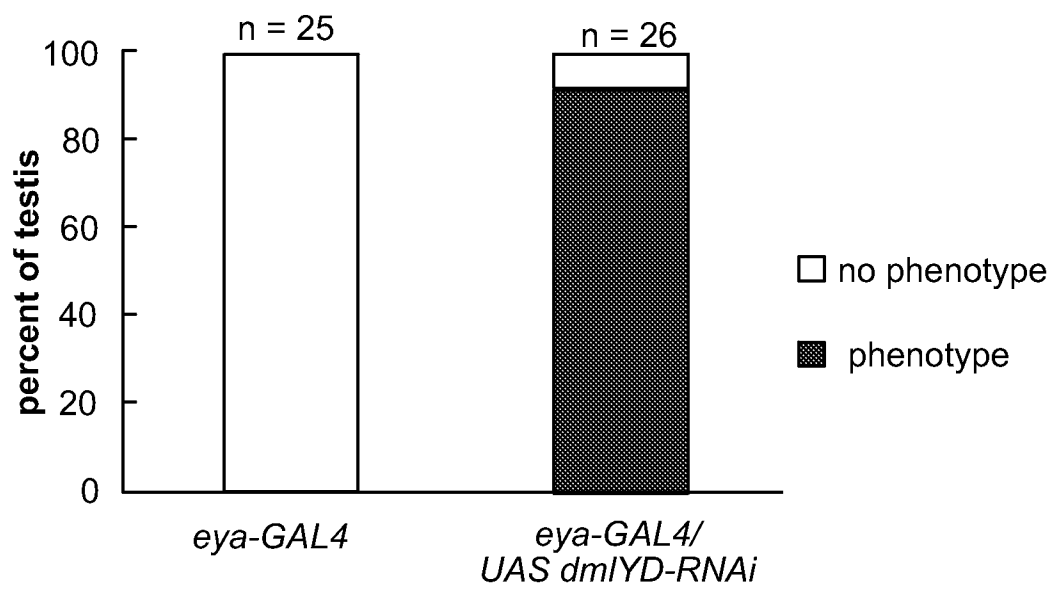


FIG. 7C

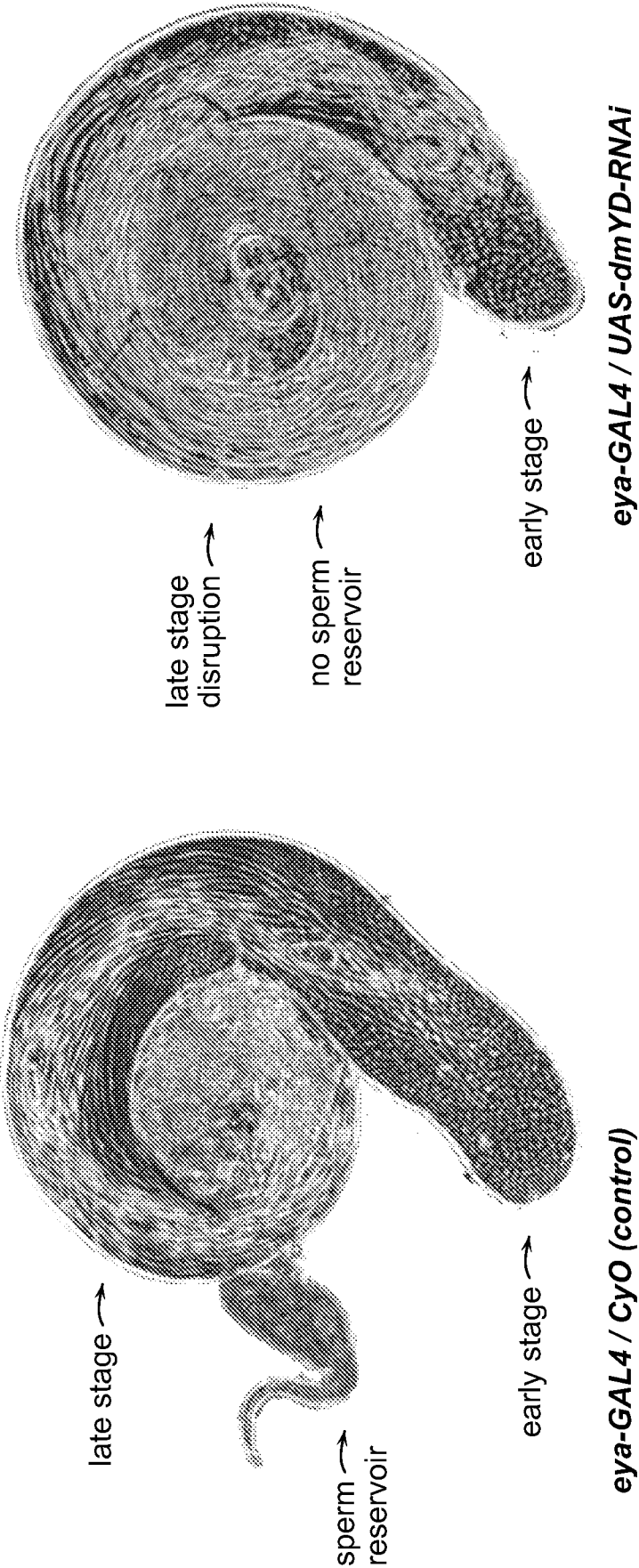
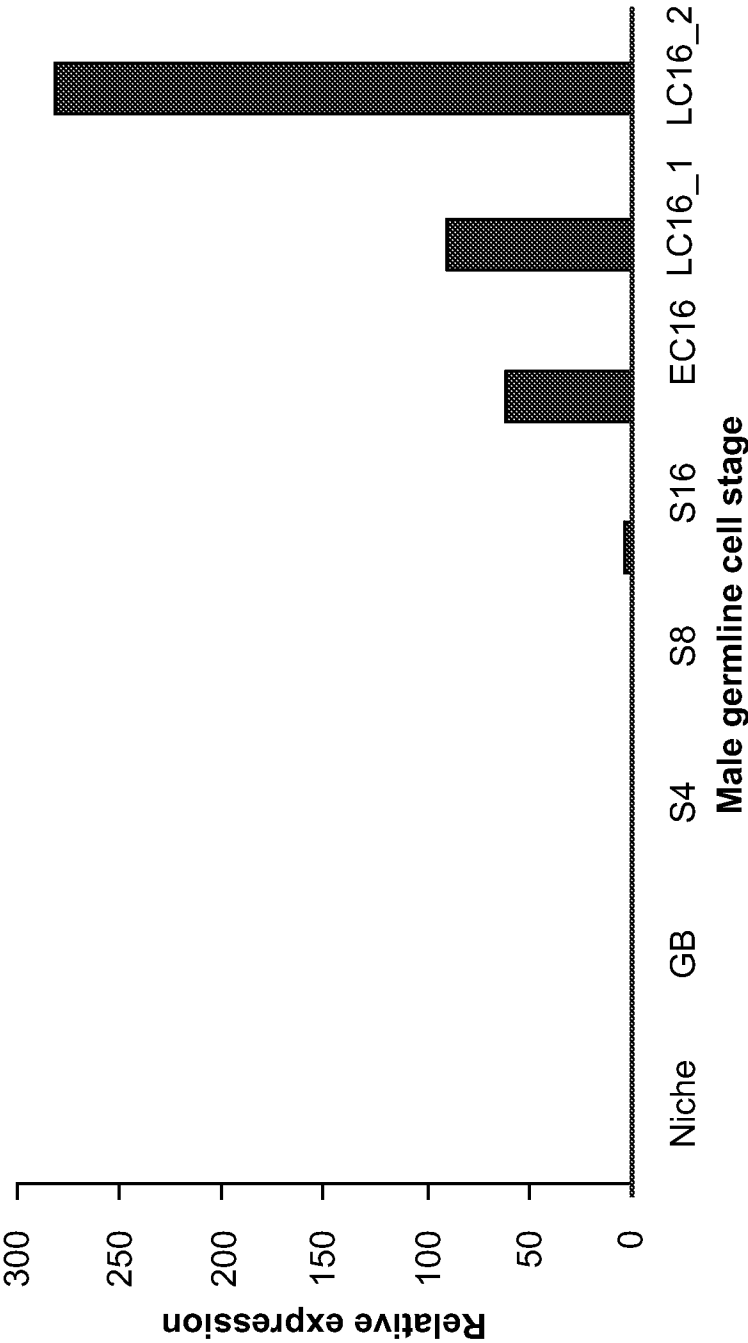


FIG. 8

Cell Stage Specific CG6279 expression in male testis



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FIG. 9A

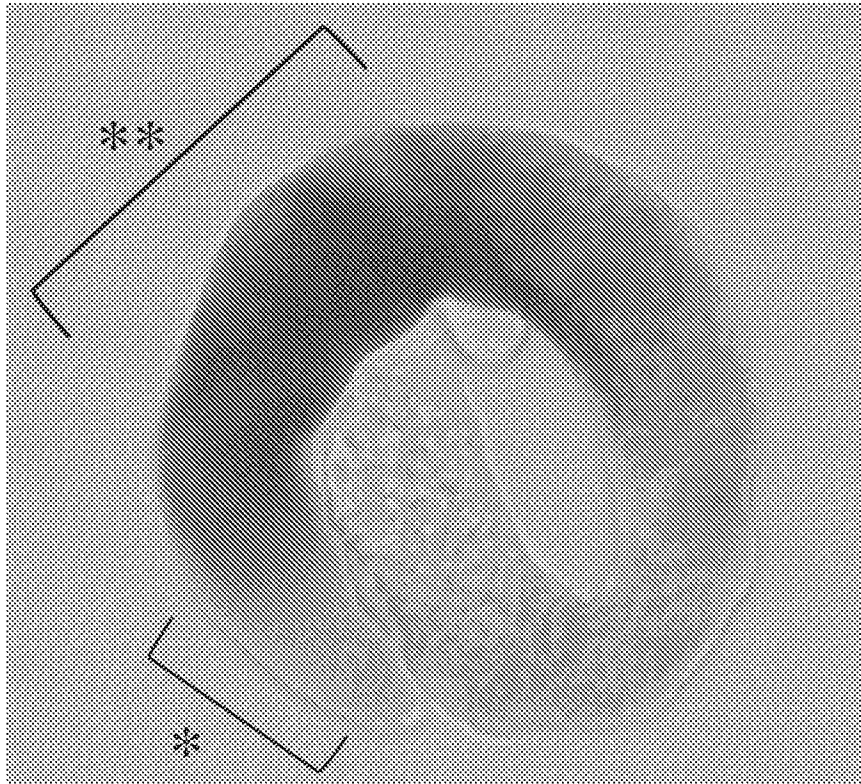
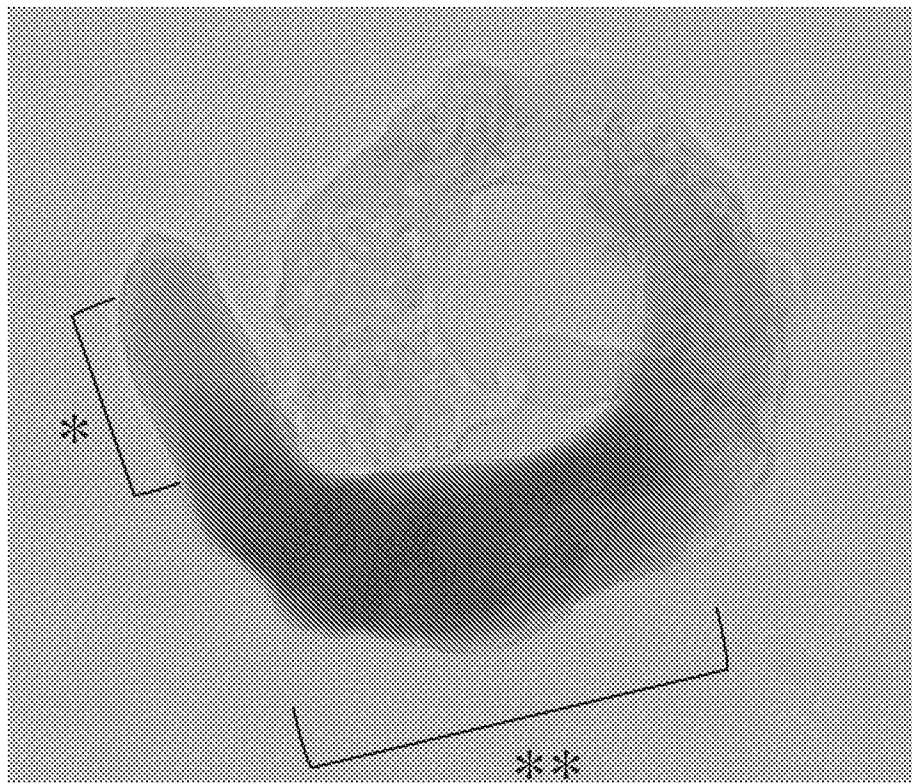


FIG. 9B



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FIG. 9C

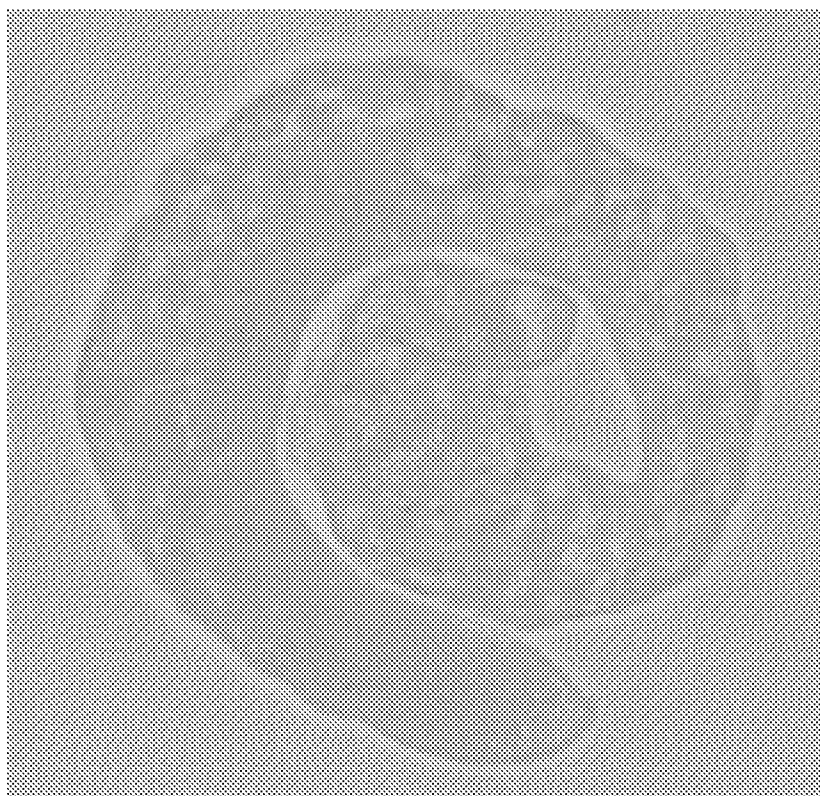


FIG. 9D

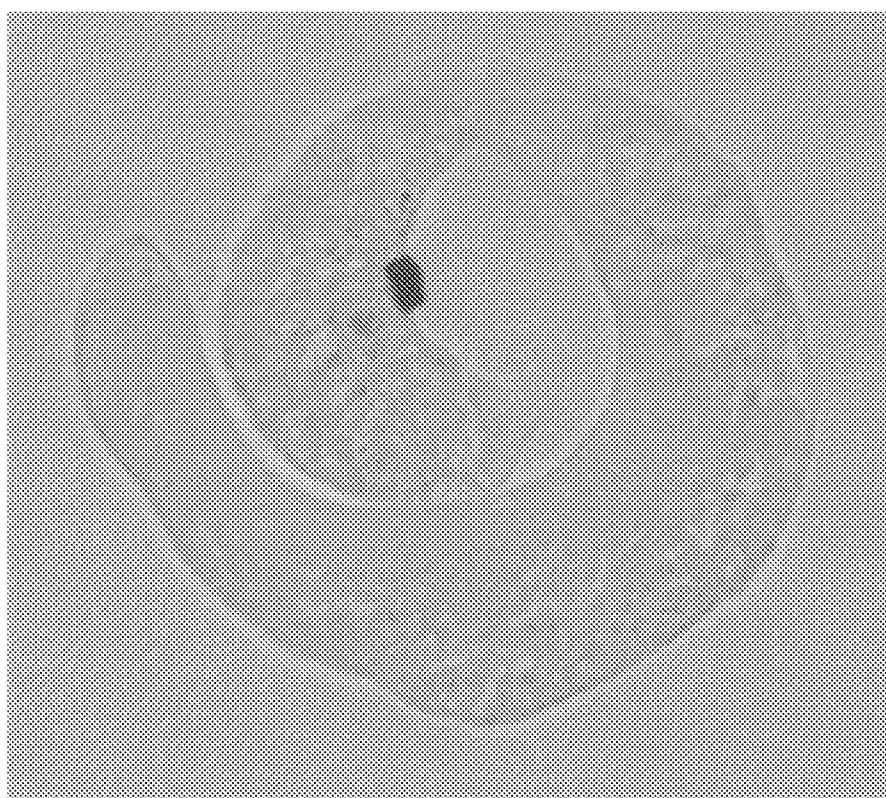
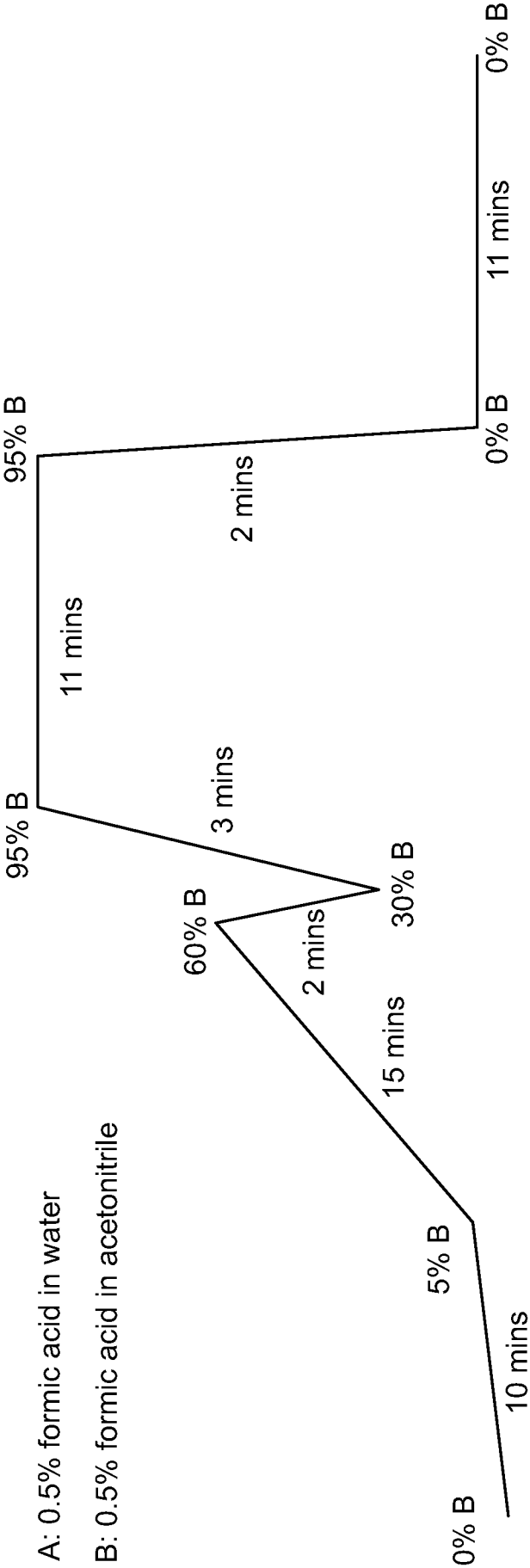


FIG. 10



INTERNATIONAL SEARCH REPORT

International application No.
PCT/US2015/042522**A. CLASSIFICATION OF SUBJECT MATTER****A01N 63/02(2006.01)i**

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

A01N 63/02; A01N 37/00; A01N 25/08; A01N 35/00; A01N 38/04; A01N 25/00; A01K 51/00; A01N 9/20

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Korean utility models and applications for utility models

Japanese utility models and applications for utility models

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

eKOMPASS(KIPO internal) & keywords: insect, reproduction, spermatogenesis, iodotyrosine deiodinase

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 4434180 A (VISSCHER) 28 February 1984 See claims 1-2 and column 1, lines 64-66.	1,5-7,11-22
A		2-4,8-10
X	US 3900567 A (LANCINI et al.) 19 August 1975 See claim 1.	1,5-7,11-22
X	US 2003-0044443 A1 (ERICKSON, JR. et al.) 06 March 2003 See claims 1, 15.	1,5-7,11-22
X	WO 2007-120461 A2 (HASS) 25 October 2007 See claims 1, 15.	1,5-7,11-22
X	US 5753615 A (THORPE et al.) 19 May 1998 See claim 14.	1,5-7,11-22



Further documents are listed in the continuation of Box C.



See patent family annex.

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"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

09 September 2015 (09.09.2015)

Date of mailing of the international search report

09 September 2015 (09.09.2015)

Name and mailing address of the ISA/KR

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INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.

PCT/US2015/042522

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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