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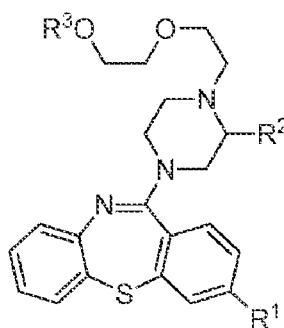
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(54) Title: HAPTENS OF QUETIAPINE FOR USE IN IMMUNOASSAYS



(I)

(57) **Abstract:** The invention relates to compounds of Formula I, wherein R^1 , R^2 , and R^3 are defined in the specification, useful for the synthesis of novel conjugates and immunogens derived from quetiapine. The invention also relates to conjugates of a quetiapine hapten and a protein.

HAPTENS OF QUETIAPINE FOR USE IN IMMUNOASSAYS

CROSS REFERENCE TO RELATED APPLICATIONS

This application claims priority of the benefits of the filing of U.S. Provisional Application Serial No. 61/691,462, filed August 21, 2012. The complete disclosures of the aforementioned related U.S. patent application is/are hereby incorporated herein by reference for all purposes.

FIELD OF THE INVENTION

The invention relates to the field of immunoassays for determining the presence of quetiapine in human biological fluids.

BACKGROUND OF THE INVENTION

Schizophrenia is a chronic and debilitating psychiatric disorder affecting approximately 0.45-1 % of the world's population (van Os, J.; Kapur, S. "Schizophrenia" Lancet 2009, 374, 635-645). The principal goals of treatment are to achieve sustained remission from psychotic symptoms, reduce the risk and consequences of relapse, and improve patient functioning and overall quality of life. While many patients with schizophrenia are able to achieve symptom stability with the available antipsychotic medications, poor adherence to medication is a common reason for relapse with daily administered oral medications. Several studies (Abdel-Baki, A.; Ouellet-Plamondon, C.; Malla, A. "Pharmacotherapy Challenges in Patients with First-Episode Psychosis" Journal of Affective Disorders 2012, 138, S3-S14) investigating the outcomes of non-compliance have shown that patients with schizophrenia who do not take their medication as prescribed have higher rates of relapse, hospital admission and suicide as well as increased mortality. It is estimated that 40 to 75% of patients with schizophrenia have difficulty adhering to a daily oral treatment regimen (Lieberman, J. A.; Stroup, T. S.; McEvoy, J. P.; Swartz, M. S.; Rosenheck, R. A.; Perkins, D. O.; Keefe, R. S. E.; Davis, S. M.; Davis, C. E.; Lebowitz, B. D.; Severe, J.; Hsiao, J. K. "Effectiveness of Antipsychotic Drugs in Patients with Chronic Schizophrenia" New England Journal of

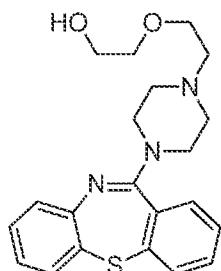
Medicine 2005, 353(12), 1209-1223). Therapeutic drug monitoring (TDM) is the quantification of serum or plasma concentrations of drugs, including anti-psychotic drugs, for treatment monitoring and optimization. Such monitoring permits, for example, the identification of patients that are not adhering to their medication regimen, that are not achieving therapeutic doses, that are non-responsive at therapeutic doses, that have suboptimal tolerability, that have pharmacokinetic drug-drug interactions, or that have abnormal metabolism resulting in inappropriate plasma concentrations. Considerable individual variability exists in the patient's ability to absorb, distribute, metabolize, and excrete anti-psychotic drugs. Such differences can be caused by concurrent disease, age, concomitant medication or genetic peculiarities. Different drug formulations can also influence the metabolism of anti-psychotic drugs. TDM permits dose optimization for individual patients, improving therapeutic and functional outcomes. TDM further permits a prescribing clinician to ensure compliance with prescribed dosages and achievement of effective serum concentrations.

To date, methods for determining the levels of serum or plasma concentrations of anti-psychotic drugs involve the use of liquid chromatography (LC) with UV or mass spectrometry detection, and radioimmunoassays (see, for example, Woestenborghs et al., 1990 "On the selectivity of some recently developed RIA's" in Methodological Surveys in Biochemistry and Analysis 20:241-246. Analysis of Drugs and Metabolites, Including Anti-infective Agents; Heykants et al., 1994 "The Pharmacokinetics of Risperidone in Humans: A Summary", J Clin Psychiatry 55/5, suppl:13-17; Huang et al., 1993 "Pharmacokinetics of the novel anti-psychotic agent risperidone and the prolactin response in healthy subjects", Clin Pharmacol Ther 54:257-268). Radioimmunoassays detect one or both of risperidone and paliperidone. Salamone et al. in US Patent No. 8,088,594 disclose a competitive immunoassay for risperidone using antibodies that detect both risperidone and paliperidone but not pharmacologically inactive metabolites. The antibodies used in the competitive immunoassay are developed against a particular immunogen. ID Labs Inc. (London, Ontario, Canada) markets an ELISA for olanzapine, another anti-psychotic drug, which also utilizes a competitive format. The Instructions For Use indicate that the assay is designed for screening purposes and intended for

forensic or research use, and is specifically not intended for therapeutic use. The Instructions recommend that all positive samples should be confirmed with gas chromatography/mass spectrometry (GC-MS), and indicate that the antibody used detects olanzapine and clozapine (see ID Labs Inc., "Instructions For Use Data Sheet IDEL-F083", Rev. Date Aug. 8, 2011). Some of these methods, namely HPLC and GC/MS, can be expensive and labor-intensive, and are generally only performed in large or specialty labs having the appropriate equipment.

A need exists for other methods for determining the levels of anti-psychotic drugs, particularly methods that can be performed in a prescribing clinician's office (where the treatment for an individual patient can be adjusted accordingly in a much more timely manner) and in other medical settings lacking LC or GC/MS equipment or requiring rapid test results.

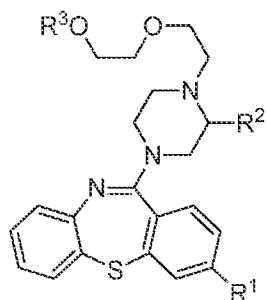
Quetiapine is:



SUMMARY OF THE INVENTION

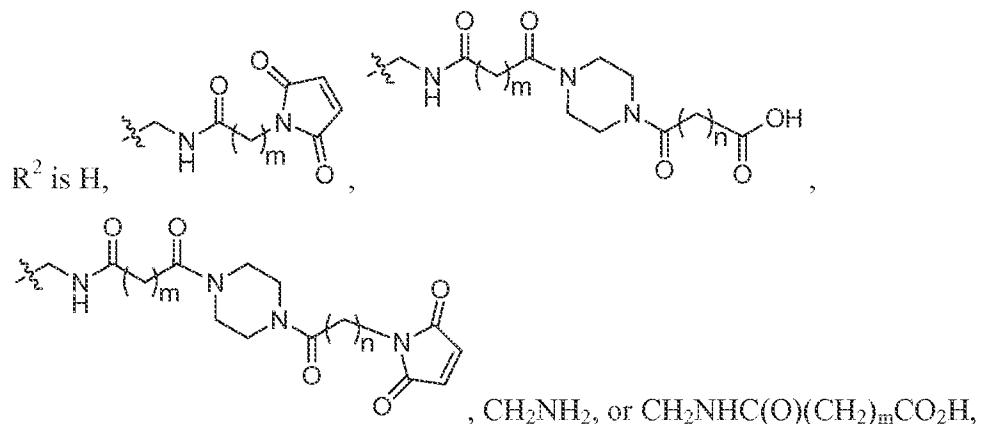
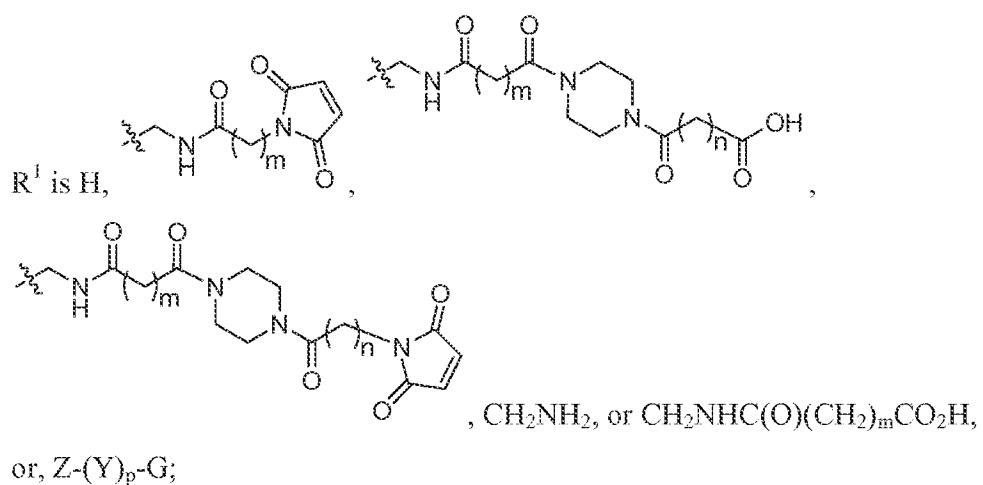
The subject invention provides compounds and conjugates that permit such an improved method for determining the levels of the anti-psychotic drug quetiapine.

The invention comprises compounds of Formula I



Formula I

wherein:

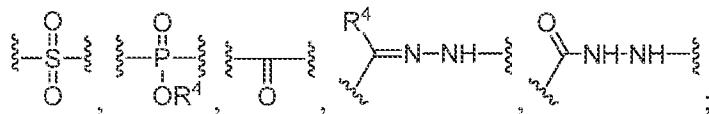


R^3 is H , or $W-(Y)_p-G$; provided that two of R^1 , R^2 , R^3 must be H , and further provided that R^1 , R^2 and R^3 may not all be H simultaneously;

wherein:

Z is selected from the group consisting of:

- $N(R^4)_2$ -, - $O-$ -, - $S-$ -, -alkyl-, -aminoalkyl-, -thioalkyl-, -heteroalkyl-, -alkylcarbonyl-,



R⁴ is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

wherein:

W is selected from the group consisting of:

-C(O)-, -alkyl-, -aminoalkyl-, -thioalkyl-, -heteroalkyl-, -alkylcarbonyl-;

Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5.

The invention comprises conjugates of compounds of the invention with immunogenic carriers such as proteins, and products produced by the process of contacting the compounds of the invention with immunogenic carriers.

BRIEF DESCRIPTION OF THE DRAWINGS

Figs. 1 and 2 show Competitive ELISA results generated with various hybridomas;

Fig. 3 shows the competitive immunoassay format used on a lateral flow assay device; and

Fig. 4 shows a typical dose response curve generated with quetiapine sub-clones 89-3, 89-13, and 89-5.

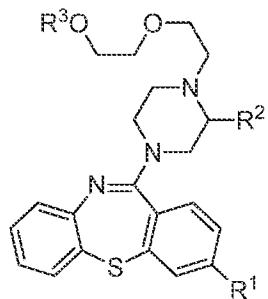
DETAILED DESCRIPTION OF THE INVENTION

The subject invention provides compounds and conjugates that permit the determination of levels of anti-psychotic drugs. Such methods will permit clinicians to evaluate objectively at an appointment how likely it is that the worsening of a patient's symptoms may be due to lack of adherence. Alternatively, if compliant, a clinician can consider a different treatment choice. Therapeutic drug monitoring, which is enabled by such methods, is key in identifying the most effective treatment options. Moreover, clinicians

believe that such TDM will help them to move into a very different relationship with their patients, i.e., to move from a hypothetical discussion on treatment non-adherence towards a more collaborative one by engaging patients to actively take ownership in optimizing their treatment regimen.

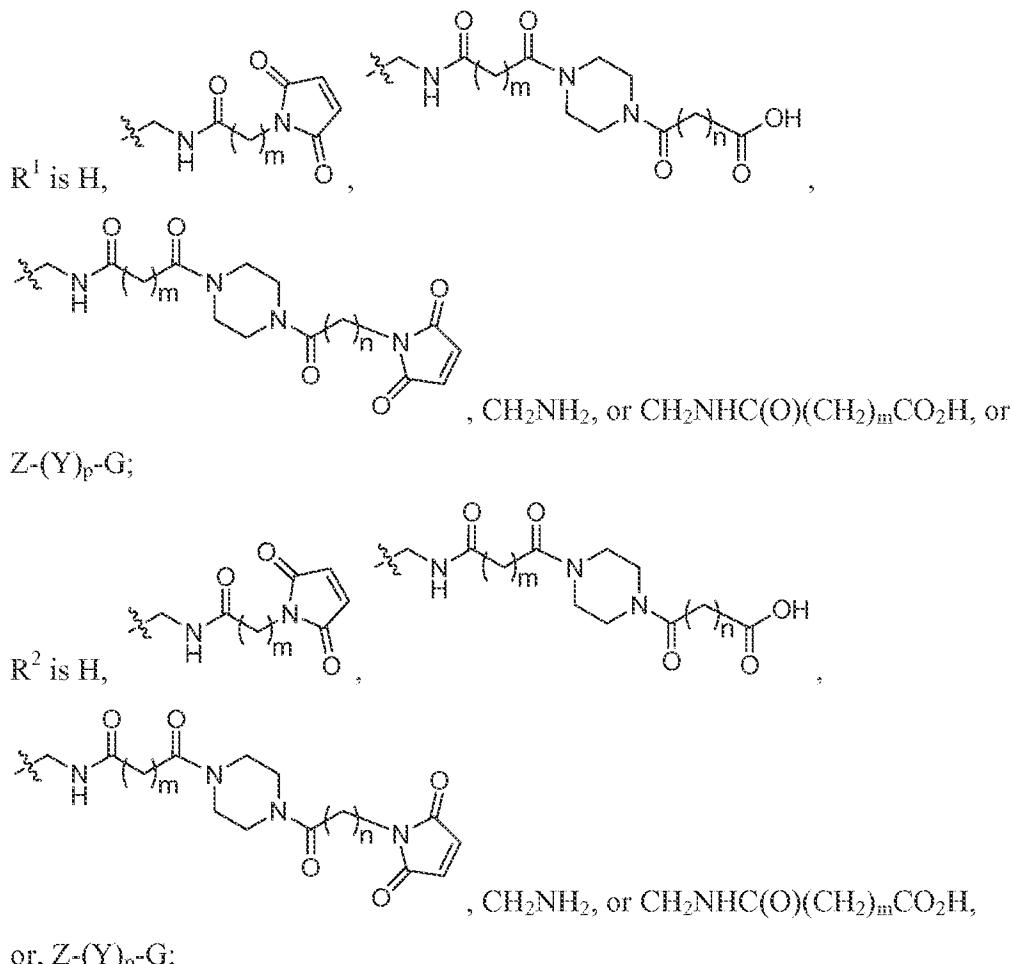
The development of the method requires first the synthesis of several immunogens, comprising a synthetic hapten linked to a protein. A hapten is a small molecule that can elicit an immune response when attached to a large carrier such as a protein. They are protein-free substances, of mostly low molecular weight, which are not capable of stimulating antibody formation alone, but which do react with antibodies. A hapten-protein conjugate is able to stimulate the production of antibodies. Specific antibody generation against small molecules is useful for immunoassay development (Pharm Res. 1992, 9(11):1375-9, Annali Dell'Istituto Superiore di Sanita. 1991, 27(1):167-74, Annali Dell'Istituto Superiore di Sanita. 1991, 27(1):149-54, Immunology Letters.1991, 28(1):79-83).

The invention comprises compounds of Formula I



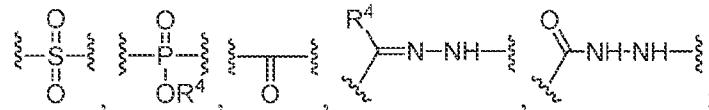
Formula I

wherein:



Z is selected from the group consisting of:

$-\text{N}(\text{R}^4)\text{-}$, $-\text{O-}$, $-\text{S-}$, $-\text{alkyl-}$, $-\text{aminoalkyl-}$, $-\text{thioalkyl-}$, $-\text{heteroalkyl-}$, $-\text{alkylcarbonyl-}$,



R^4 is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

wherein:

W is selected from the group consisting of:

$-\text{C(O)-}$, $-\text{alkyl-}$, $-\text{aminoalkyl-}$, $-\text{thioalkyl-}$, $-\text{heteroalkyl-}$, $-\text{alkylcarbonyl-}$;

Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

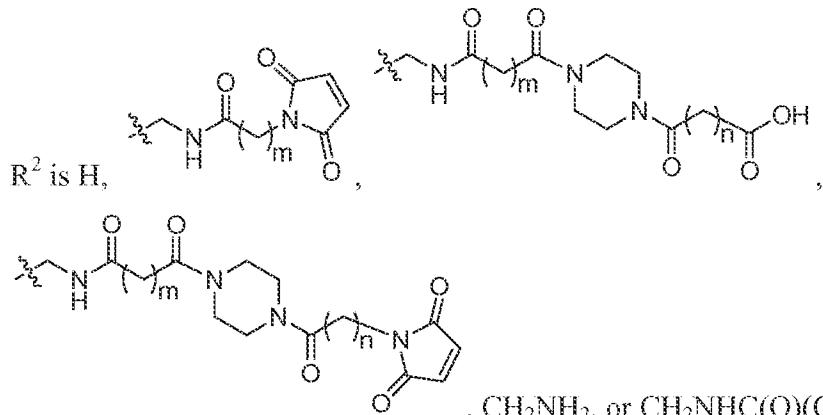
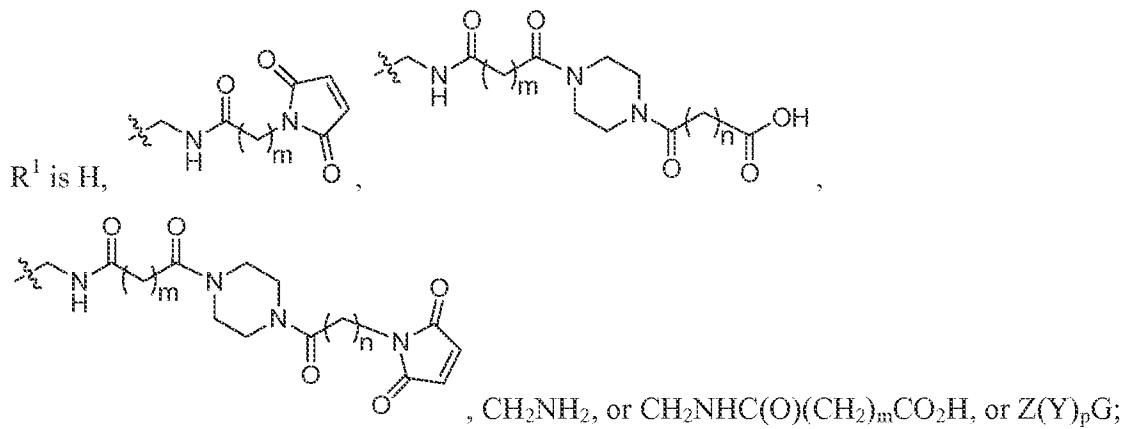
p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5.

The invention comprises compounds of Formula I

wherein:



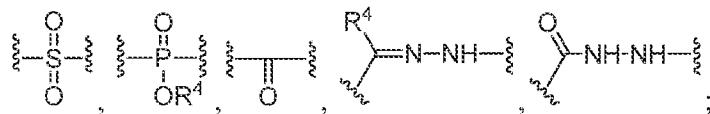
or, $Z \cdot (Y)_n \cdot G$;

R^3 is H, provided that either R^1 or R^2 must be H, and further provided that both R^1 and R^2 may not be H simultaneously;

wherein:

Z is selected from the group consisting of:

-N(R⁴)-, -O-, -S-, -alkyl-, -aminoalkyl-, -thioalkyl-, -heteroalkyl-, -alkylcarbonyl-,



R^4 is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5.

The invention comprises compounds of Formula I:

wherein:

R^1 is H, or $CH_2NH-(Y)_p-G$;

R^2 is H, or $CH_2NH-(Y)_p-G$; provided that either R^1 or R^2 must be H, and further provided that both R^1 and R^2 may not be H simultaneously;

R^3 is H;

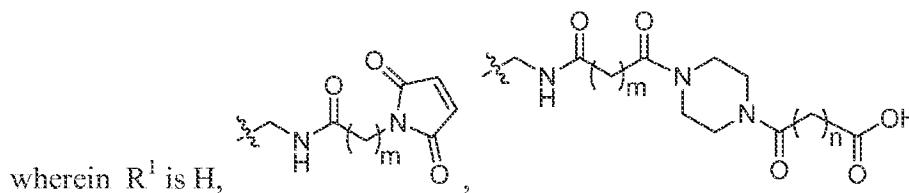
wherein:

Y is an organic spacer group;

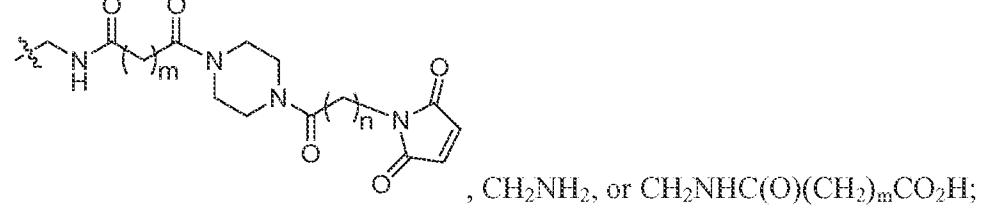
G is a functional linking group capable of binding to a carrier;

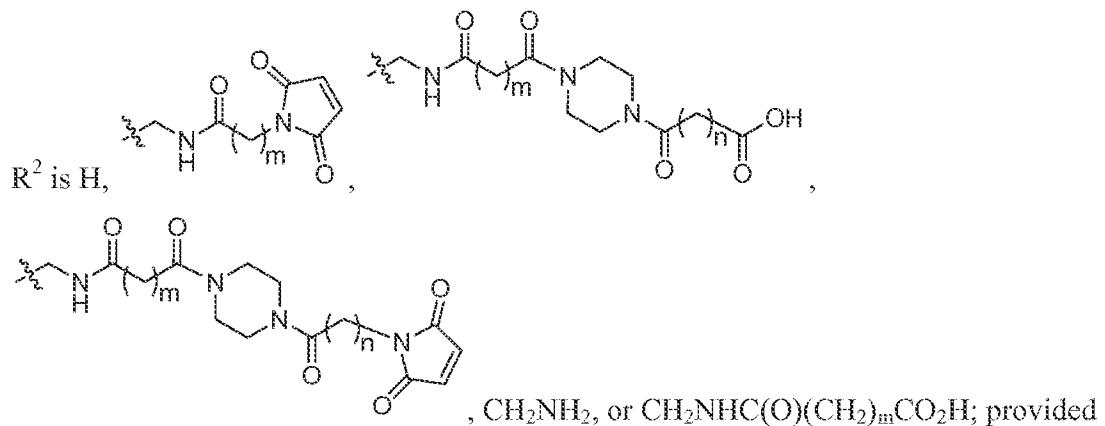
p is 1.

The invention comprises compounds of Formula I



wherein R^1 is H,



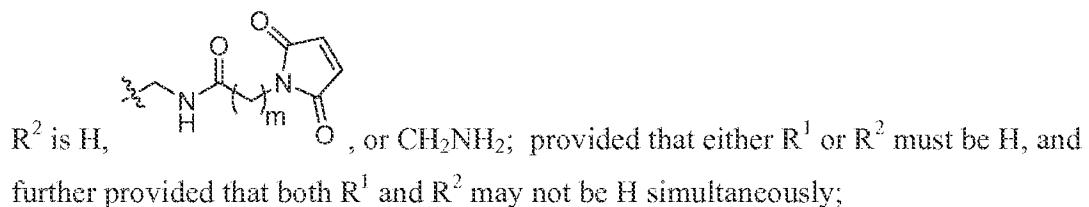
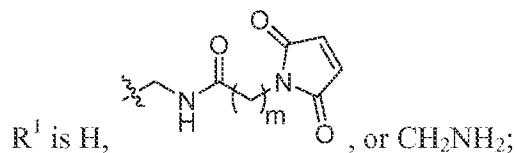


R^3 is H;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5.

In another embodiment of the invention:

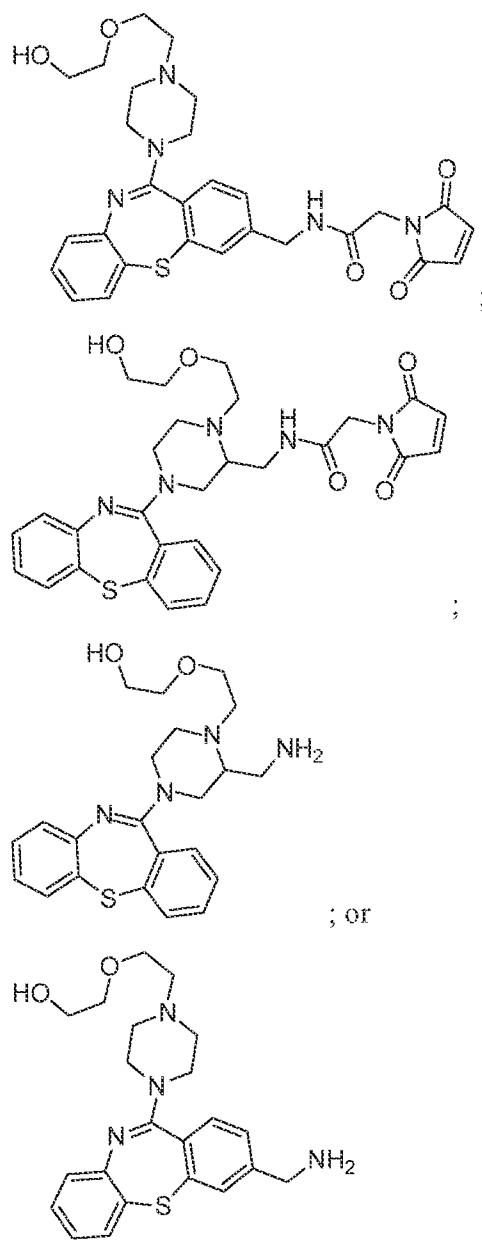


R^3 is H;

m is 1, 2, 3, 4, or 5;

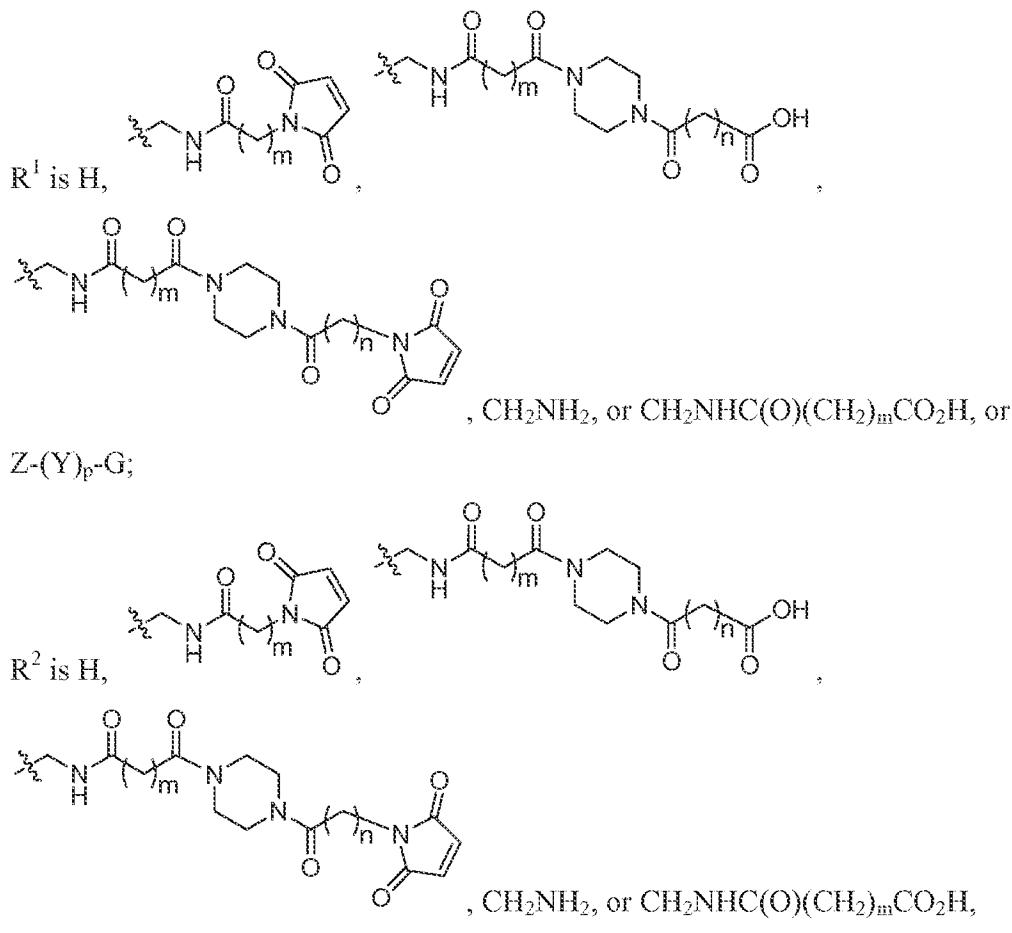
n is 1, 2, 3, 4, or 5.

Another embodiment of the invention is a compound of Formula I which is:



The invention further provides conjugates of the above compounds with an immunogenic carrier.

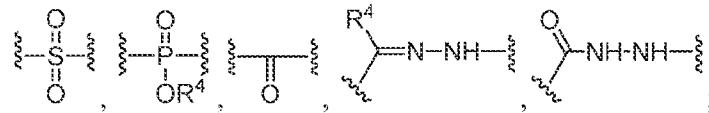
Another embodiment of the invention is thus a conjugate of a compound of Formula I wherein:



wherein:

Z is selected from the group consisting of:

$-\text{N}(\text{R}^4)\text{-}$, $-\text{O}\text{-}$, $-\text{S}\text{-}$, $-\text{alkyl-}$, $-\text{aminoalkyl-}$, $-\text{thioalkyl-}$, $-\text{heteroalkyl-}$, $-\text{alkylcarbonyl-}$,



R^4 is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

wherein:

W is selected from the group consisting of:

$-\text{C(O)-}$, $-\text{alkyl-}$, $-\text{aminoalkyl-}$, $-\text{thioalkyl-}$, $-\text{heteroalkyl-}$, $-\text{alkylcarbonyl-}$;

Y is an organic spacer group;

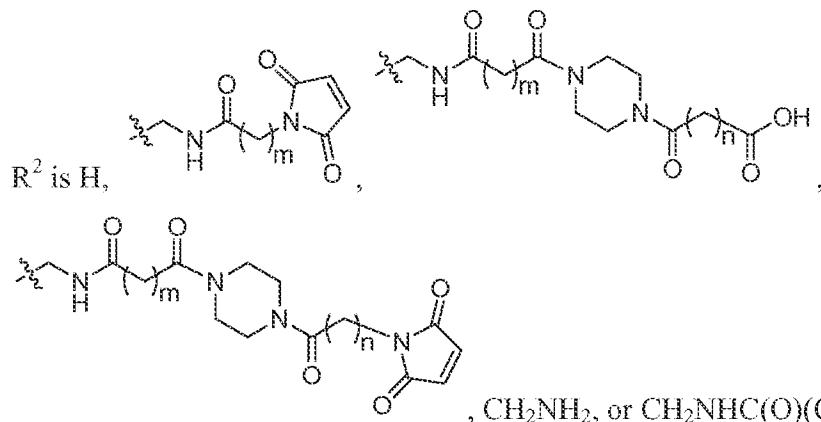
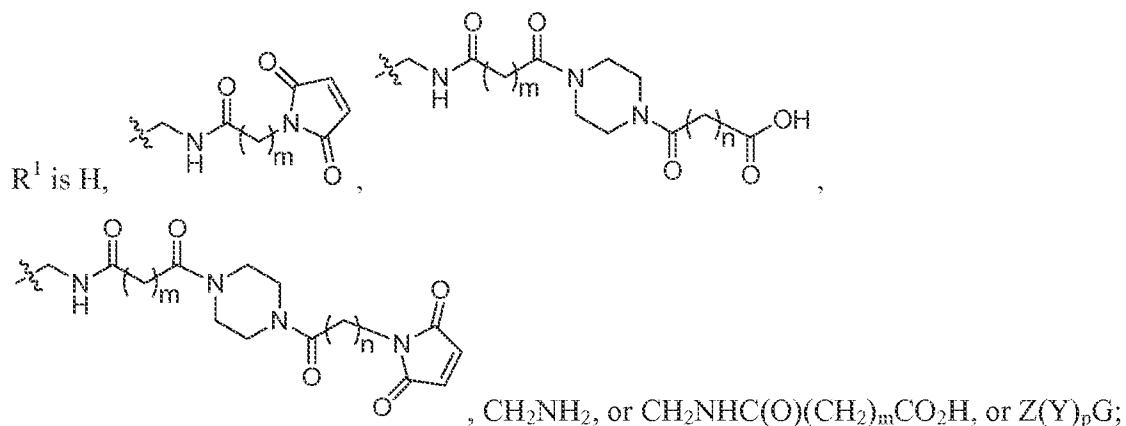
G is a functional linking group capable of binding to a carrier;

p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5; and an immunogenic carrier.

Another embodiment of the invention is a conjugate of a compound of Formula I wherein:



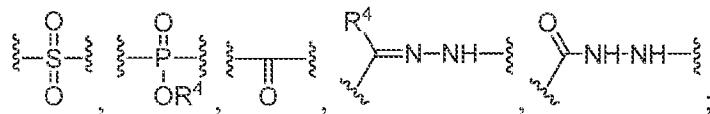
or, $\text{Z-(Y)}_p\text{-G}$;

R^3 is H, provided that either R^1 or R^2 must be H, and further provided that both R^1 and R^2 may not be H simultaneously;

wherein:

Z is selected from the group consisting of:

$-\text{N}(\text{R}^4)_2$, $-\text{O}-$, $-\text{S}-$, $-\text{alkyl-}$, $-\text{aminoalkyl-}$, $-\text{thioalkyl-}$, $-\text{heteroalkyl-}$, $-\text{alkylcarbonyl-}$,



R⁴ is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5; with an immunogenic carrier.

Another embodiment of the invention is a conjugate of a compound of Formula I wherein:

R¹ is H, or CH₂NH-(Y)_p-G;

R² is H, or CH₂NH-(Y)_p-G; provided that either R¹ or R² must be H, and further provided that both R¹ and R² may not be H simultaneously;

R³ is H;

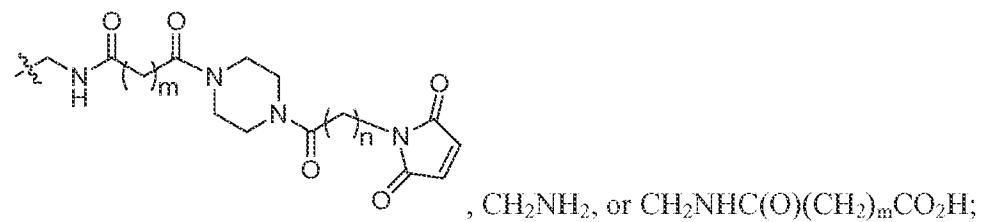
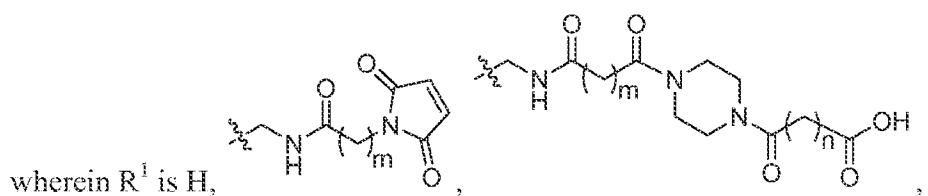
wherein:

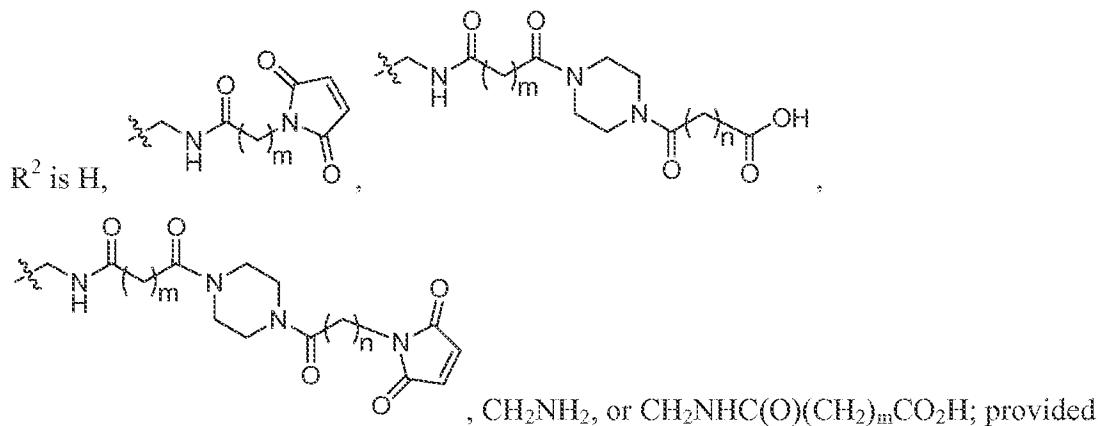
Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

p is 1; and an immunogenic carrier.

Another embodiment of the invention is a conjugate of a compound of Formula I





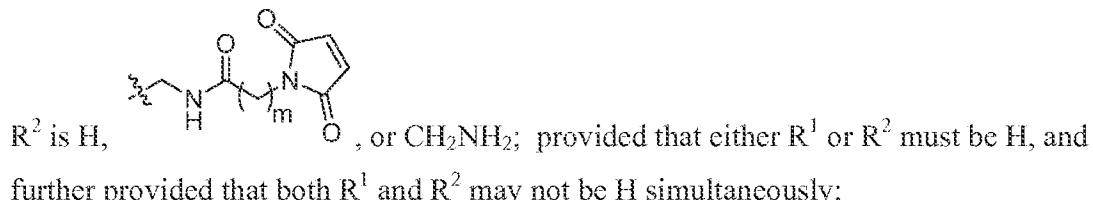
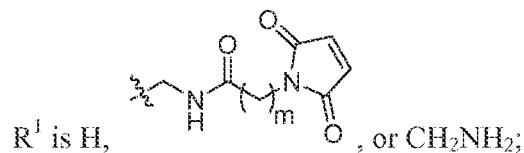
R^3 is H;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5; and an immunogenic carrier.

Another embodiment of the invention is a conjugate of a compound of Formula I

wherein:

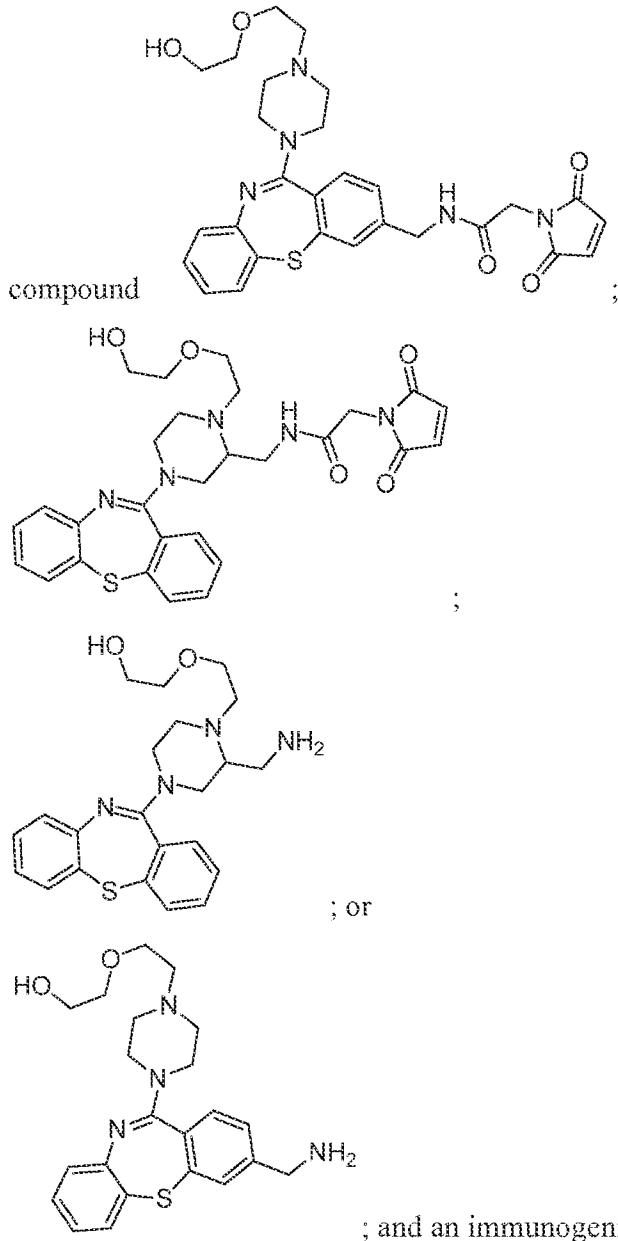


R^3 is H;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5; and an immunogenic carrier.

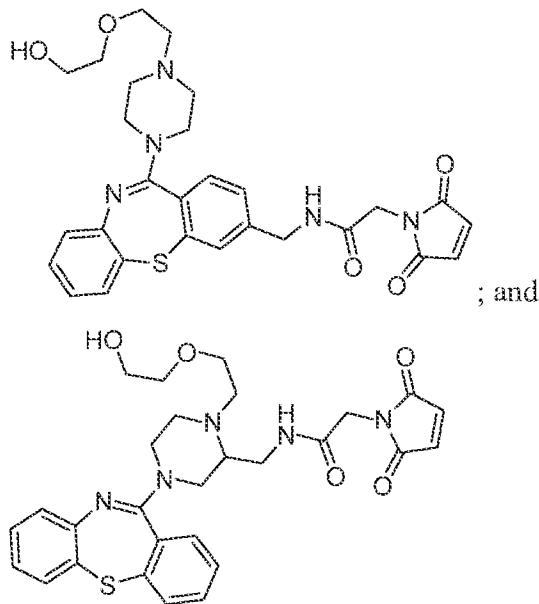
Another embodiment of the invention is a conjugate of the



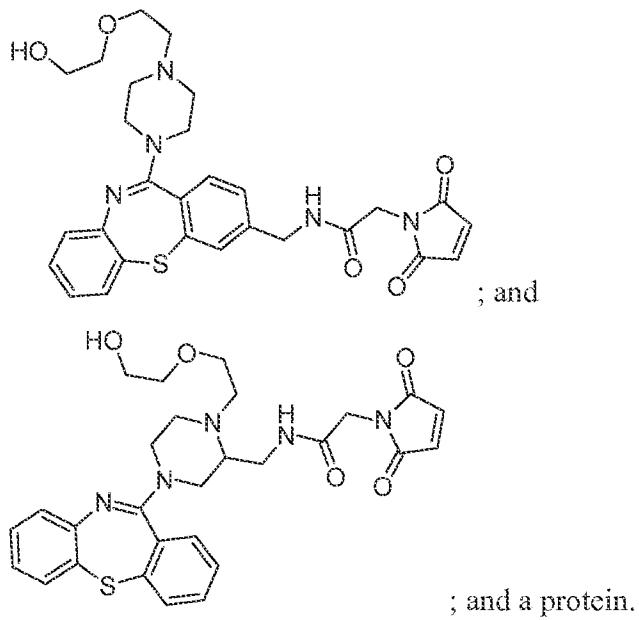
A preferred embodiment of the invention is any of the above conjugates wherein the immunogenic carrier is a protein.

A preferred embodiment of the invention is any of the above conjugates, wherein said protein is keyhole limpet hemocyanin, ovalbumin or bovine thyroglobulin.

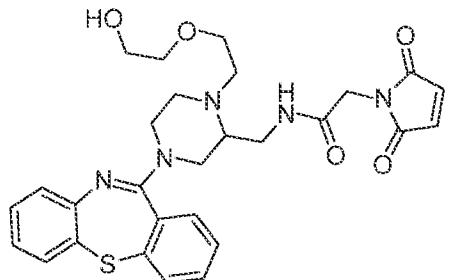
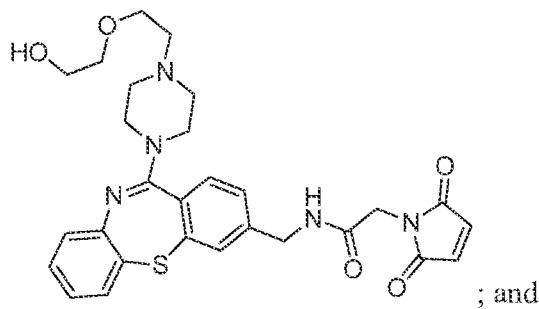
Another preferred embodiment of the invention is a conjugate of an immunogenic carrier and a compound selected from the group consisting of



Another preferred embodiment of the invention is a conjugate of a compound selected from the group consisting of:



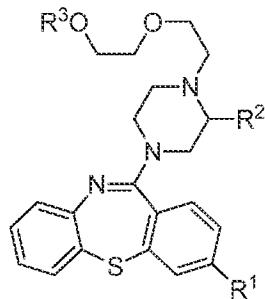
A more preferred embodiment of the invention is a conjugate of a protein and a compound selected from the group consisting of



; wherein said protein is keyhole limpet hemocyanin, ovalbumin or bovine thyroglobulin.

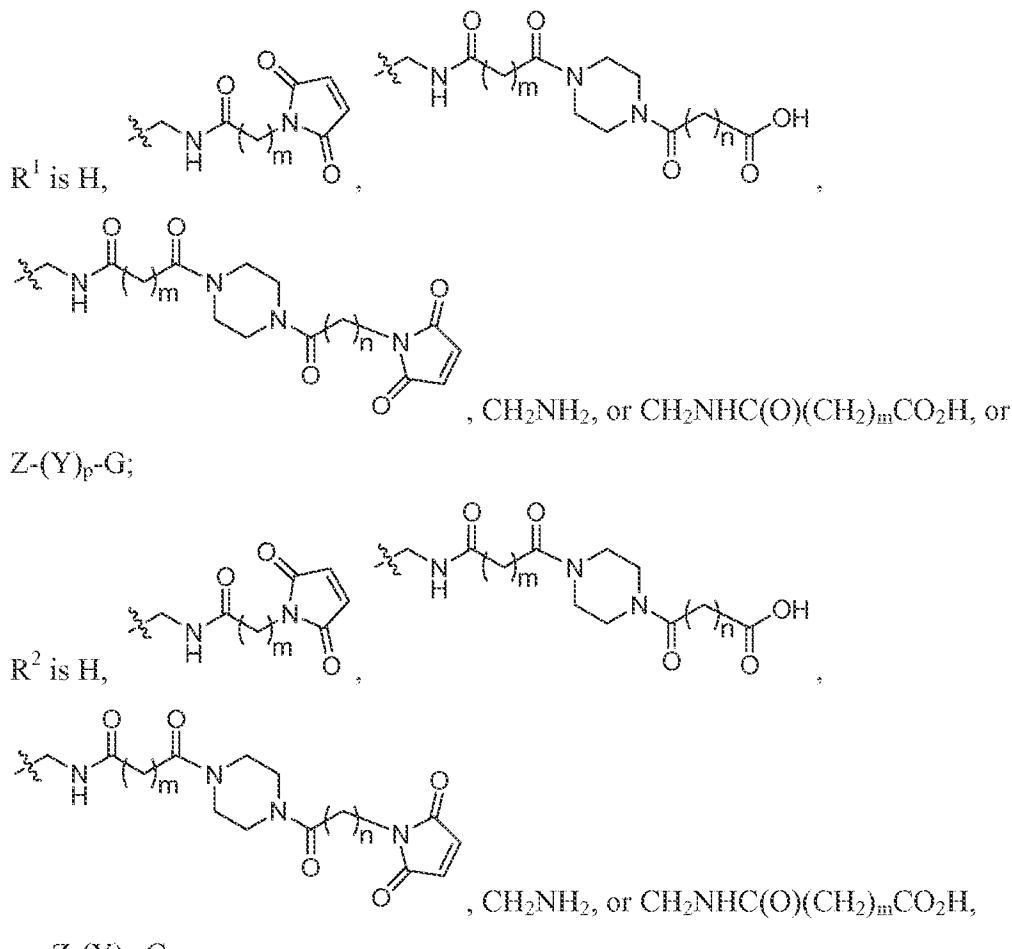
The invention also provides products formed from the process of contacting the above compounds with an immunogenic carrier.

Another embodiment of the invention is thus a product made by the process of contacting a compound of Formula I



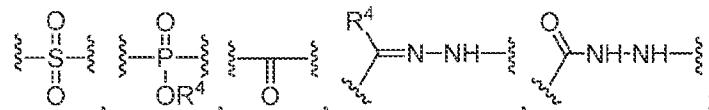
Formula I

wherein:



Z is selected from the group consisting of:

-N(R^4)-, -O-, -S-, -alkyl-, -aminoalkyl-, -thioalkyl-, -heteroalkyl-, -alkylcarbonyl-,



R^4 is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

wherein:

W is selected from the group consisting of:

-C(O)-, alkyl-, -aminoalkyl-, -thioalkyl-, -heteroalkyl-, -alkylcarbonyl-;

Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

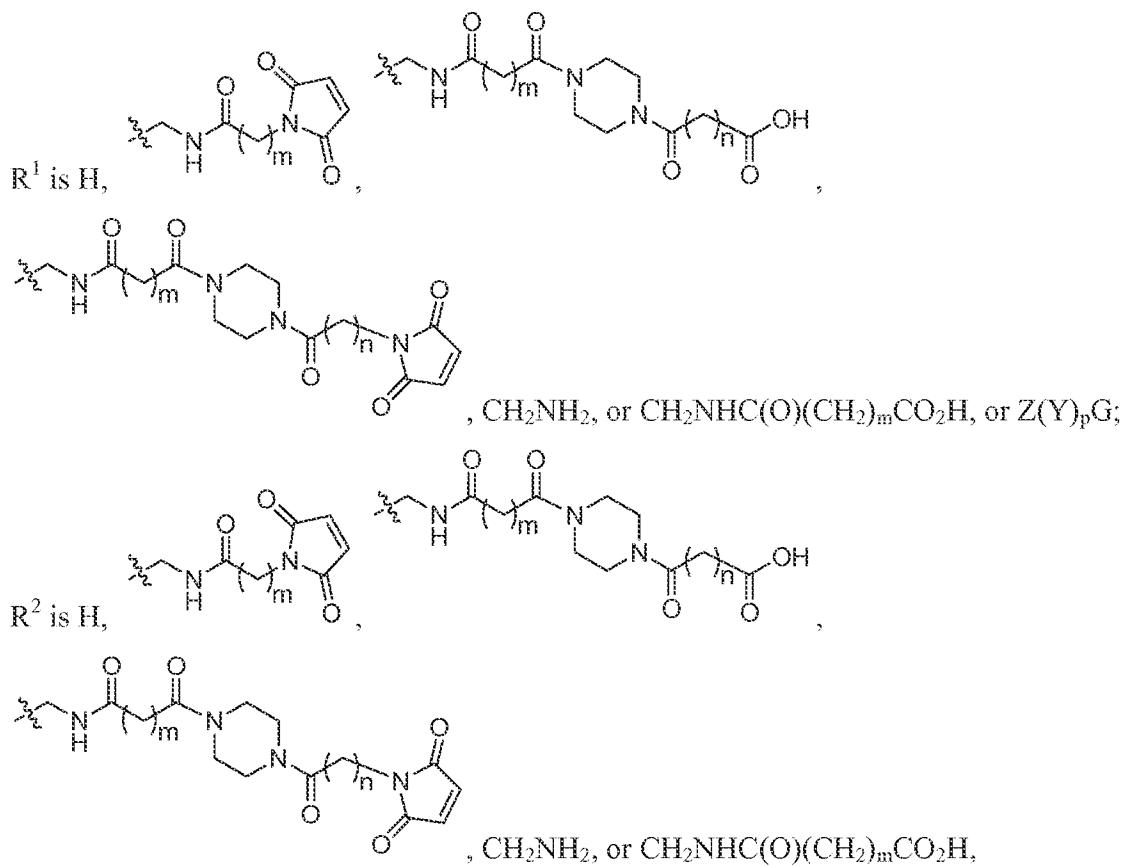
p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5; with an immunogenic carrier.

Another embodiment of the invention is a product made by the process of contacting a compound of Formula I

wherein:



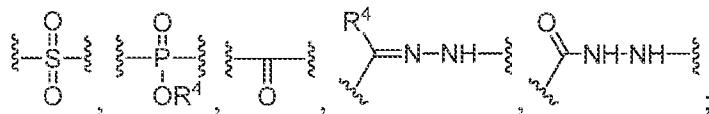
or, $\text{Z-(Y)}_p\text{-G}$;

R^3 is H, provided that either R^1 or R^2 must be H, and further provided that both R^1 and R^2 may not be H simultaneously;

wherein:

Z is selected from the group consisting of:

$-\text{N}(\text{R}^4)\text{-}$, $-\text{O-}$, $-\text{S-}$, $-\text{alkyl-}$, $-\text{aminoalkyl-}$, $-\text{thioalkyl-}$, $-\text{heteroalkyl-}$, $-\text{alkylcarbonyl-}$,



R^4 is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5; with an immunogenic carrier.

Another embodiment of the invention is a product made by the process of contacting a compound of Formula I

wherein:

R^1 is H, or $CH_2NH-(Y)_p-G$;

R^2 is H, or $CH_2NH-(Y)_p-G$; provided that either R^1 or R^2 must be H, and further provided that both R^1 and R^2 may not be H simultaneously;

R^3 is H;

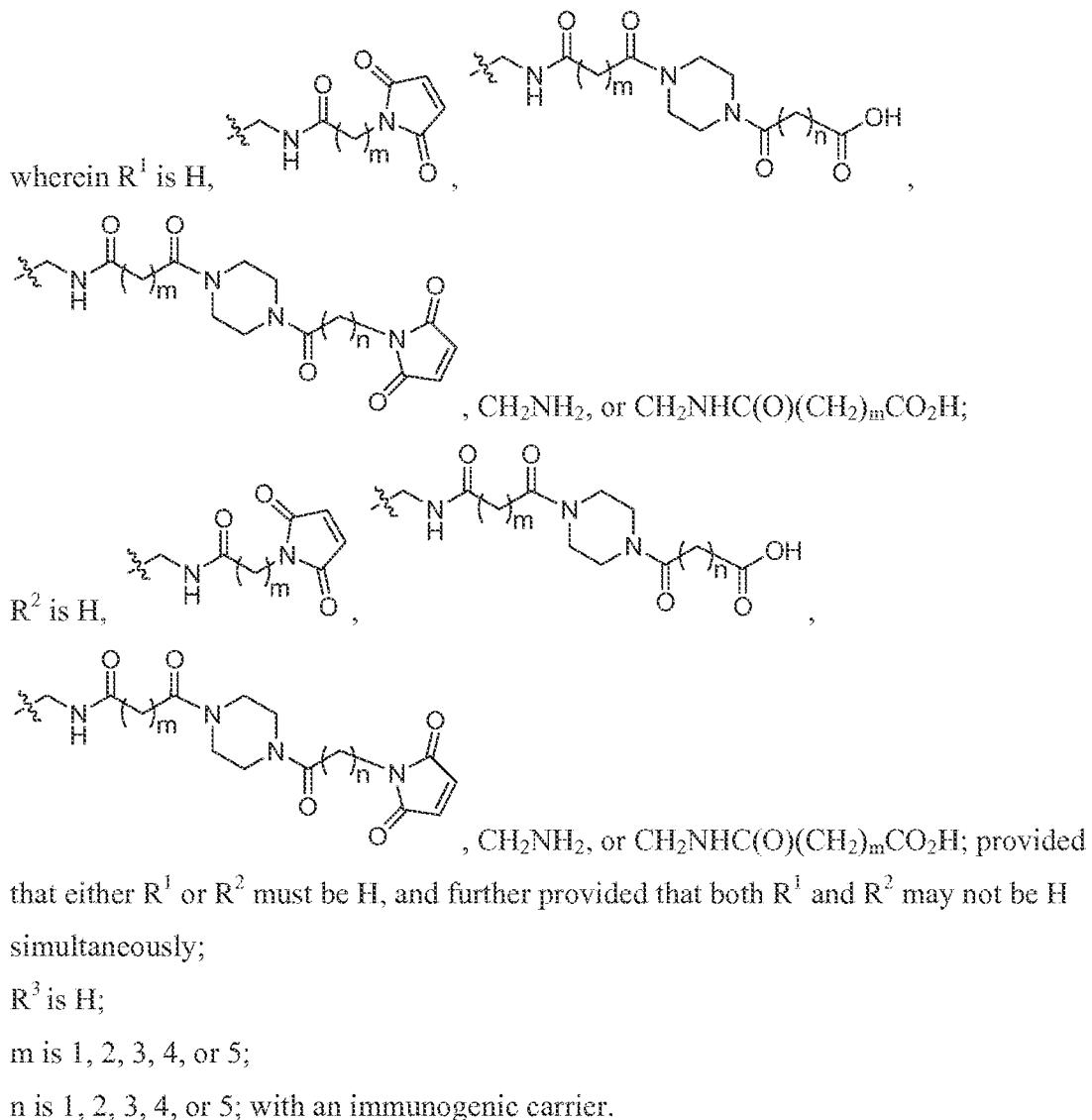
wherein:

Y is an organic spacer group;

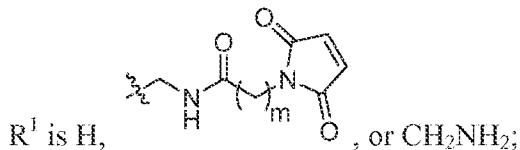
G is a functional linking group capable of binding to a carrier;

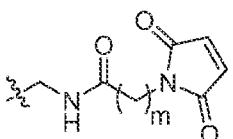
p is 1; with an immunogenic carrier.

Another embodiment of the invention is a product made by the process of contacting a compound of Formula I



Another embodiment of the invention is a product made by the process of contacting a compound of Formula I
wherein:





 R^2 is H, or CH_2NH_2 ; provided that either R^1 or R^2 must be H, and further provided that both R^1 and R^2 may not be H simultaneously;

 R^3 is H;

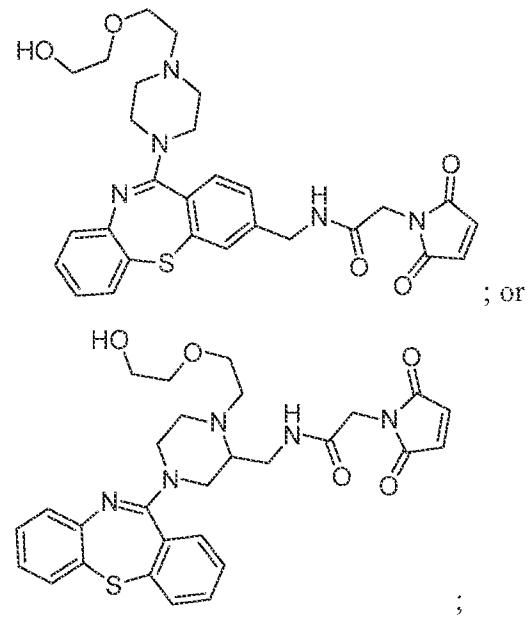
 m is 1, 2, 3, 4, or 5;

 n is 1, 2, 3, 4, or 5; with an immunogenic carrier.

A preferred embodiment of the invention is a product made by the process of contacting any of the above compounds with an immunogenic carrier wherein the immunogenic carrier is a protein.

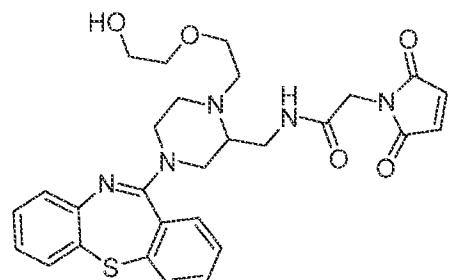
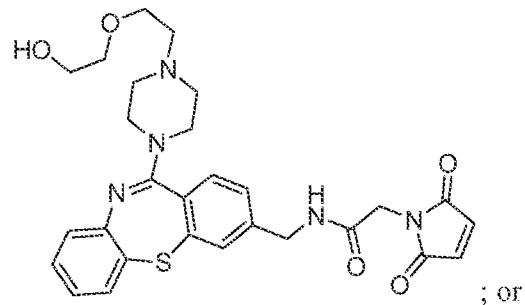
A more preferred embodiment of the invention is a product made by the process of contacting any of the above compounds with an immunogenic carrier wherein the immunogenic carrier is a protein and wherein said protein is keyhole limpet hemocyanin, ovalbumin or bovine thyroglobulin.

Another embodiment of the invention is a product made by the process of contacting a compound which is



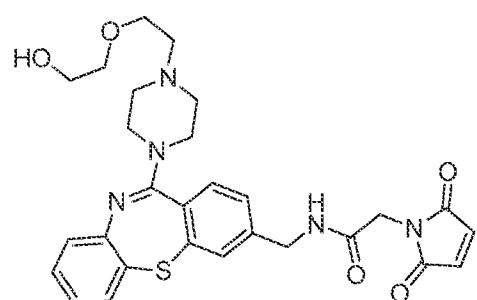
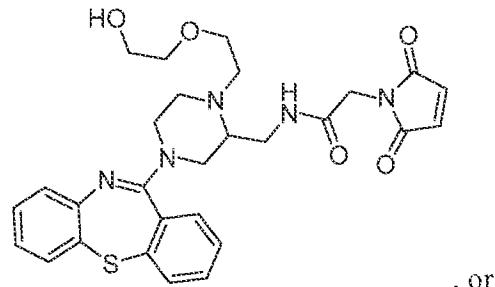
with an immunogenic carrier.

Another embodiment of the invention is a product made by the process of contacting a compound which is



with a protein.

Another embodiment of the invention is a product made by the process of contacting a compound which is



with a protein, wherein said protein is keyhole limpet hemocyanin, or bovine thyroglobulin.

ABBREVIATIONS

Herein and throughout the application, the following abbreviations may be used.

AMAS	N-(α -maleimidooxy) succinimide ester
BINAP	2,2'-bis(diphenylphosphino)-1,1'-binaphthyl
Boc or BOC	<i>tert</i> -butoxycarbonyl
BTG	bovine thyroglobulin
Bu ₃ N	tributylamine
DCC	dicyclohexylcarbodiimide
DCM	dichloromethane
DIEA	diisopropylethylamine
DMF	N,N-dimethylformamide
EDCI or EDC	1-(3-dimethylaminopropyl)-3-ethylcarbodiimide hydrochloride
EDTA	ethylenediaminetetraacetic acid
HOBT or HOBt	1-hydroxybenzotriazole hydrate
KLH	keyhole limpet hemocyanin
Pd ₂ (dba) ₃	tris(dibenzylideneacetone)dipalladium(0)
SATA	N-succinimidyl S-acetylthioacetate
TEA or Et ₃ N	triethylamine
THF	tetrahydrofuran
TFA	trifluoroacetic acid
r.t.	room temperature
DEAD	diethylazodicarboxylate
DIC	diisopropylcarbodiimide
NHS	N-hydroxysuccinimide
TFP	Tetrafluorophenyl
PNP	p-nitrophenyl
TBTU	O-(Benzotriazol-1-yl)-N,N,N',N'-tetramethyluronium tetrafluoroborate

DEPBT	3-(diethoxyphosphoryloxy)-1,2,3-benzotrazin-4(3H)-one
BOP-Cl	Bis(2-oxo-3-oxazolidinyl)phosphonic chloride
DTT	dithioerythritol

DEFINITIONS

The term "conjugate" refers to any substance formed from the joining together of separate parts. Representative conjugates in accordance with the present invention include those formed by the joining together of a small molecule, such as the compounds of Formula I, and a large molecule, such as a carrier or a polyamine polymer, particularly a protein. In the conjugate the small molecule maybe joined at one or more active sites on the large molecule.

The term "hapten" refers to a partial or incomplete antigen. A hapten is a protein-free substance, which is not capable of stimulating antibody formation, but which does react with antibodies. The antibodies are formed by coupling a hapten to a high molecular weight immunogenic carrier, and then injecting this coupled product, i.e., an immunogen, into a human or animal subject.

The term "immunogen" refers to a substance capable of eliciting, producing, or generating an immune response in an organism.

An "immunogenic carrier," as used herein, is an immunogenic substance, commonly a protein, that can join at one or more positions with haptens, thereby enabling the production of antibodies that can bind specifically with these haptens. Examples of immunogenic carrier substances include, but are not limited to, proteins, glycoproteins, complex polyamino-polysaccharides, particles, and nucleic acids that are recognized as foreign and thereby elicit an immunologic response from the host. The polyamino-polysaccharides may be prepared from polysaccharides using any of the conventional means known for this preparation.

Various protein types may be employed as immunogenic carriers, including without

limitation, albumins, serum proteins, lipoproteins, etc. Illustrative proteins include bovine serum albumin, keyhole limpet hemocyanin, egg ovalbumin, bovine thyroglobulin, fraction V human serum albumin, rabbit albumin, pumpkin seed globulin, diphtheria toxoid, tetanus toxoid, botulinus toxin, succinylated proteins, and synthetic poly(aminoacids) such as polylysine.

Immunogenic carriers can also include poly amino-polysaccharides, which are a high molecular weight polymer built up by repeated condensations of monosaccharides. Examples of polysaccharides are starches, glycogen, cellulose, carbohydrate gums such as gum arabic, agar, and so forth. The polysaccharide also contains poly(amino acid) residues and/or lipid residues.

The immunogenic carrier can also be a poly(nucleic acid) either alone or conjugated to one of the above mentioned poly(amino acids) or polysaccharides.

The immunogenic carrier can also include solid particles. The particles are generally at least about 0.02 microns (μm) and not more than about 100 μm , and usually about 0.05 μm to 10 μm in diameter. The particle can be organic or inorganic, swellable or non-swellable, porous or non-porous, optimally of a density approximating water, generally from about 0.7 to 1.5 g/mL, and composed of material that can be transparent, partially transparent, or opaque. The particles can be biological materials such as cells and microorganisms, including non-limiting examples such as erythrocytes, leukocytes, lymphocytes, hybridomas, *Streptococcus*, *Staphylococcus aureus*, *E. coli*, and viruses. The particles can also be comprised of organic and inorganic polymers, liposomes, latex, phospholipid vesicles, or lipoproteins.

The term “derivative” refers to a chemical compound or molecule made from a parent compound by one or more chemical reactions.

The term “analogue” of a chemical compound refers to a chemical compound that contains a chain of carbon atoms and the same particular functional groups as a reference

compound, but the carbon chain of the analogue is longer or shorter than that of the reference compound.

A “label,” “detector molecule,” or “reporter” is any molecule which produces, or can be induced to produce, a detectable signal. The label conjugated to an analyte, immunogen, antibody, or to another molecule such as a receptor or a molecule that can bind to a receptor such as a ligand, particularly a hapten or antibody. Non-limiting examples of labels include radioactive isotopes (e.g., ^{125}I), enzymes (e.g., β -galactosidase, peroxidase), enzyme fragments, enzyme substrates, enzyme inhibitors, coenzymes, catalysts, fluorophores (e.g., rhodamine, fluorescein isothiocyanate or FITC, or Dylight 649), dyes, chemiluminescers and luminescers (e.g., dioxetanes, luciferin), or sensitizers.

As used herein, a “spacer” refers to a portion of a chemical structure which connects two or more substructures such as haptens, carriers, immunogens, labels or binding partners through a functional linking group. These spacer groups are composed of the atoms typically present and assembled in ways typically found in organic compounds and so may be referred to as “organic spacing groups”. The chemical building blocks used to assemble the spacers will be described hereinafter in this application. Among the preferred spacers are straight or branched, saturated or unsaturated carbon chains. These carbon chains may also include one or more heteroatoms within the chain, one or more heteroatoms replacing one or more hydrogens of any carbon atom in the chain, or at the termini of the chains. By “heteroatoms” is meant atoms other than carbon which are chosen from the group consisting of oxygen, nitrogen, phosphorous and sulfur, wherein the nitrogen, phosphorous and sulfur atoms may exist in any oxidation state and may have carbon or other heteroatoms bonded to them. The spacer may also include cyclic or aromatic groups as part of the chain or as a substitution on one of the atoms in the chain.

The number of atoms in the spacing group is determined by counting the atoms other than hydrogen. The number of atoms in a chain within a spacing group is determined by counting the number of atoms other than hydrogen along the shortest route between the substructures being connected. Preferred chain lengths are between 1 to 20 atoms.

A “functional linking group” refers to a reactive group that is present on a hapten and may be used to provide an available reactive site through which the hapten portion may be coupled to another moiety through formation of a covalent chemical bond to produce a conjugate of a hapten with another moiety (such as a label or carrier). The hapten may be linked in this way to a moiety such as biotin to form a competitive binding partner for the hapten.

Spacer groups may be used to link the hapten to the carrier. Spacers of different lengths allow one to attach the hapten with differing distances from the carrier for presentation to the immune system of the animal or human being immunized for optimization of the antibody formation process. Attachment to different positions in the hapten molecule allows the opportunity to present specific sites on the hapten to the immune system to influence antibody recognition. The spacer may contain hydrophilic solubilizing groups to make the hapten derivative more soluble in aqueous media. Examples of hydrophilic solubilizing groups include but are not limited to polyoxyalkyloxy groups, for example, polyethylene glycol chains; hydroxyl, carboxylate and sulfonate groups.

The term “nucleophilic group” or “nucleophile” refers to a species that donates an electron-pair to form a chemical bond in a reaction. The term “electrophilic group” or “electrophile” refers to a species that accepts an electron-pair from a nucleophile to form a chemical bond in a reaction.

The term “substituted” refers to substitution of an atom or group of atoms in place of a hydrogen atom on a carbon atom in any position on the parent molecule. Non limiting examples of substituents include halogen atoms, amino, hydroxy, carboxy, alkyl, aryl, heteroalkyl, heteroaryl, cyano, alkoxy, nitro, aldehyde and ketone groups.

The term “alkyl” refers to both linear and branched chain radicals of up to 12 carbon atoms, unless otherwise indicated, and is specifically intended to include radicals having any degree or level of saturation. Alkyl includes, but is not limited to, methyl, ethyl,

propyl, isopropyl, butyl, isobutyl, sec-butyl, *tert*-butyl, pentyl, isopentyl, hexyl, isohexyl, heptyl, octyl, 2,2,4-trimethylpentyl, nonyl, decyl, undecyl and dodecyl.

The term “cycloalkyl” refers to a saturated or partially unsaturated monocyclic or bicyclic hydrocarbon ring radical composed of from 3 to 10 carbon atoms. Alkyl substituents may optionally be present on the ring. Examples include cyclopropyl, 1,1-dimethyl cyclobutyl, 1,2,3-trimethylcyclopentyl, cyclohexyl and cyclohexenyl.

The term “heteroatom” refers to a nitrogen atom, an oxygen atom, a phosphorous atom or a sulfur atom wherein the nitrogen, phosphorous and sulfur atoms can exist in any allowed oxidation states.

The term “heteroalkyl” refers to an alkyl group that includes one or more heteroatoms within the chain, one or more heteroatoms replacing one or more hydrogens of any carbon atom in the chain, or at termini of the chains.

The term “heterocyclyl” refers to a nonaromatic (*i.e.* saturated or partially unsaturated) ring composed of from 3 to 7 carbon atoms and at least one heteroatom selected from N, O or S. Alkyl substituents may optionally be present on the ring. Examples include tetrahydrofuryl, dihydropyranyl, piperidyl, 2,5-dimethylpiperidyl, morpholinyl, piperazinyl, thiomorpholinyl, pyrrolidinyl, pyrrolinyl, pyrazolidinyl, pyrazolinyl, imidazolidinyl and imidazolinyl.

The term “hydroxyalkyl” refers to at least one hydroxyl group bonded to any carbon atom along an alkyl chain.

The term “aminoalkyl” refers to at least one primary or secondary amino group bonded to any carbon atom along an alkyl chain.

The term “alkoxyalkyl” refers to at least one alkoxy group bonded to any carbon atom along an alkyl chain.

The term “polyalkoxyalkyl” refers to long-chain alkoxy compounds and includes polyethylene glycols of discreet or monodispersed sizes.

The term “thioalkyl” refers to at least one sulfur group bonded to any carbon atom along an alkyl chain. The sulfur group may be at any oxidation state and includes sulfoxides, sulfones and sulfates.

The term “carboxyalkyl” refers to at least one carboxylate group bonded to any carbon atom along an alkyl chain. The term “carboxylate group” includes carboxylic acids and alkyl, cycloalkyl, aryl or aralkyl carboxylate esters.

The term “alkylcarbonyl” refers to a group that has a carbonyl group bonded to any carbon atom along an alkyl chain. The term “heteroaryl” refers to 5- to 7-membered mono- or 8- to 10-membered bicyclic aromatic ring radicals, any ring of which may consist of from one to four heteroatoms selected from N, O or S where the nitrogen and sulfur atoms can exist in any allowed oxidation state. Examples include benzimidazolyl, benzothiazolyl, benzothienyl, benzoxazolyl, furyl, imidazolyl, isothiazolyl, isoxazolyl, oxazolyl, pyrazinyl, pyrazolyl, pyridyl, pyrimidinyl, pyrrolyl, quinolinyl, thiazolyl and thienyl.

The term “heteroaralkyl” refers to a C₁₋₆ alkyl group having a heteroaryl substituent. Examples include furylethyl and 2-quinolinylpropyl.

The term “alkoxy” refers to straight or branched chain radicals of up to 12 carbon atoms, unless otherwise indicated, bonded to an oxygen atom. Examples include methoxy, ethoxy, propoxy, isopropoxy and butoxy.

The term “aryl” refers to monocyclic or bicyclic aromatic ring radicals containing from 6 to 12 carbons in the ring. Alkyl substituents may optionally be present on the ring. Examples include phenyl, biphenyl and naphthalene.

The term “aralkyl” refers to a C₁₋₆ alkyl group containing an aryl substituent. Examples include benzyl, phenylethyl or 2-naphthylmethyl.

The term “heteroaralkyl” refers to a C₁₋₆ alkyl group containing a heteroaryl substituent. Examples include furylmethyl and pyridylpropyl.

The term “aryloxy” refers to an oxygen atom bound to an aryl substituent. Examples include phenoxy and benzyloxy.

The term “arylalkoxy” refers to an alkoxy group bound to an aryl substituent. Examples include phenylmethyl ether.

The term “acyl” refers to the group -C(O)R_a, where R_a is alkyl, aryl, aralkyl, heteroaryl and heteroaralkyl. An “acylating agent” adds the -C(O)R_a group to a molecule.

The term “sulfonyl” refers to the group -S(O)₂R_a, where R_a is hydrogen, alkyl, cycloalkyl, haloalkyl, aryl, aralkyl, heteroaryl and heteroaralkyl. A “sulfonylating agent” adds the -S(O)₂R_a group to a molecule.

Spacers bearing reactive functional linking groups for the attachment of haptens to carrier moieties may be prepared by a wide variety of methods. The spacer may be formed using a molecule that is differentially functionalized or activated with groups at either end to allow selective sequential reaction with the hapten and the carrier, but the same reactive moiety may also be used at both ends. The groups selected for reaction with the hapten and the functional linking group to be bound to the carrier are determined by the type of functionality on the hapten and the carrier that the hapten is to be bonded with. Spacers and methods of attachment to haptens and carriers include but are not limited to those described by Brinkley, M., A., *Bioconjugate Chem.* 1992, 3:2-13, Hermanson, Greg T., *Bioconjugate Techniques*, Academic Press, London, Amsterdam, Burlington, MA, USA, 2008 and *Thermo Scientific Pierce Crosslinking Technical Handbook*; available for

download or hard copy request from **Thermo Scientific** 3747 N Meridian Rd, Rockford, IL USA 61101, ph 800-874-3723 or at: <http://www.piercenet.com/> and references within. Many differentially activated molecules for formation of spacer groups are commercially available from vendors, for example **Thermo Scientific**.

For haptens bearing an amino group, modes of attachment of the spacer to the hapten include reaction of the amine on the hapten with a spacer building block bearing an acyl halide or active ester. "Active esters" are defined as esters that undergo reaction with a nucleophilic group, for example an amino group, under mild conditions to form a stable linkage. A stable linkage is defined as one that remains intact under conditions of further use, for example subsequent synthetic steps, use as an immunogen, or in a biochemical assay. A preferred example of a stable linkage is an amide bond. Active esters and methods of formation are described by Benoiton, N.L., in Houben-Weyl, *Methods of Organic Chemistry*, Thieme Stuttgart, New York, vol E22 section 3.2:443 and Benoiton, N.L., *Chemistry of Peptide Synthesis*, Taylor and Francis, NY, 2006. Preferred active esters include p-nitrophenyl ester (PNP), N-hydroxysuccinimide ester (NHS) and tetrafluorophenyl ester (TFP). Acyl halides may be prepared by many methods known to one skilled in the art for example, reaction of the carboxylic acid with thionyl chloride or oxalyl chloride, see: Fieser, L.F. and Fieser, M. *Reagents for Organic Synthesis*, John Wiley and Sons, NY, 1967 and references within. These may be converted to other active esters such as p-nitrophenyl esters (PNP) which may also be used in active bi-functional spacers as described by Wu et.al, *Organic Letters*, 2004, 6 (24):4407. N-hydroxysuccinimide (NHS) esters may be prepared by reaction of N,N-disuccinimidyl carbonate (CAS 74124-79-1) with the carboxylic acid of a compound in the presence of an organic base such as triethylamine or diisopropylethylamine in an aprotic solvent under anhydrous conditions as described in Example 35 of WO2012012595 or by using N-hydroxysuccinimide and dicyclohexylcarbodiimide (DCC) or other dehydrating agent, under anhydrous conditions. Tetrafluorophenyl esters (TFP) may be prepared by reaction of carboxylic acids with 2,3,5,6-tetrafluorophenyltrifluoroacetate in the presence of an organic base such as triethylamine or diisopropylethylamine in an aprotic solvent under anhydrous conditions as reported by Wilbur, et.al, *Bioconjugate Chem.*, 2004, 15(1):203.

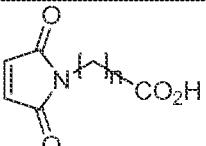
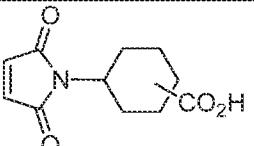
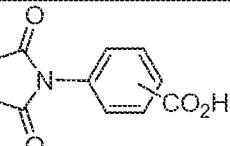
One skilled in the art will recognize that spacers shown in Table 1, among others, can be obtained using known methods and attached to amino-bearing haptens utilizing routine optimization of reaction conditions. These spacers allow attachment of the hapten to a thiol group on a carrier.

Table 1

	Reasonable values for m and n are between 1 and 10

Direct coupling of the amine on the hapten and a carboxylic acid functionality on the spacer building block in the presence of a coupling agent may also be used as a mode of attachment. Preferred reagents are those typically used in peptide synthesis. Peptide coupling reagents include but are not limited to O-(Benzotriazol-1-yl)-N,N,N',N'-tetramethyluronium tetrafluoroborate (TBTU, CAS #125700-67-6), see: Pruhs, S., *Org. Process. Res. Dev.* **2006**, 10:441; N-Hydroxybenzotriazole (HOBT, CAS #2592-95-2) with a carbodiimide dehydrating agent, for example N,N-dicyclohexylcarbodiimide (DCC), diisopropylcarbodiimide (DIC), or 1-ethyl-3-(3-dimethylaminopropyl)carbodiimidehydrochloride (EDC), see: König W., Geiger, R. *Chem. Ber.*, **1970**, 103 (3):788 ; 3-(diethoxyphosphoryloxy)-1,2,3-benzotrazin-4(3H)-one (DEPBT, CAS#165534-43-0), see: Liu, H. et.al., *Chinese Chemical Letters*, **2002**, 13(7):601; Bis(2-oxo-3-oxazolidinyl)phosphonic chloride; (BOP-Cl, CAS# 68641-49-6), see: Diago-Meseguer, J et.al. *Synthesis*, **1980**, 7:547-51 and others described in detail by Benoiton in *Chemistry of Peptide Synthesis*, CRC Press, Boca Raton, FL, **2005**, Chapter 2, and the technical bulletin provided by **Advanced Automated Peptide Protein Technologies** (aapptec), 6309 Shepardsville Rd., Louisville KY 40228, ph 888 692 9111; www.aapptec.com, and references within. These methods create a stable amide linkage attaching the hapten to the spacer. Examples of spacers that can be obtained using known methods and attached to amino-bearing haptens utilizing routine optimization of reaction conditions employing the methods described and cited above are shown, but not limited to those in Table 2. These spacers allow attachment of the hapten to a thiol group on a carrier.

Table 2

 reasonable range for n is between 1-10		
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Spacers may also be constructed in a step-wise fashion by sequential attachment of appropriate chemical groups to the hapten including the step of forming the functional linking group that is capable of binding to the carrier. See illustrative examples under General Reaction Schemes.

Additionally, when the hapten has a nucleophilic group, for example a thiol group, an amino group or a hydroxyl group which will become the point of attachment of the spacer, the spacer may also be constructed by alkylation of the thiol, amine or hydroxyl group. Any alkyl group that is appropriately substituted with a moiety capable of undergoing a substitution reaction, for example, an alkyl halide, or sulfonic acid ester such as p-Toluenesulfonate, may be used to attach the spacer. Many examples of alkylation reactions are known to one skilled in the art and specific examples may be found in the general chemical literature and optimized through routine experimentation. A discussion of alkylation reactions with many references can be found in Chapter 10 of *March's Advanced Organic Chemistry*, Smith, M.B., and March, J., John Wiley & sons, Inc. NY, 2001. Other linkages may also be employed such as reaction of the nucleophilic moiety, for example an amine, on the hapten with an isocyanate to form a urea or reaction with an isothiocyanate to form a thiourea linkage, see: Li, Z., et.al., *Phosphorus, Sulfur and Silicon and the Related Elements*, 2003, 178(2):293-297. Spacers may be attached to haptens bearing hydroxyl groups via reaction with isocyanate groups to form carbamate or urethane linkages. The spacer may be differentially activated with the isocyanate functional group on one end and a functional linking group capable of reacting with the carrier, see: Annunziato, M.E., Patel, U.S., Ranade, M. and Palumbo, P.S., *Bioconjugate Chem.*, 1993, 4:212-218.

For haptens bearing a carboxylic acid group, modes of attachment of a spacer portion to the hapten include activation of the carboxylic acid group as an acyl halide or active ester, examples of which are shown in Table 3, preparation of which are described previously, followed by reaction with an amino (-NH₂-), hydrazino (-NH-NH₂-), hydrazido (-C(O)-NH-NH₂-) or hydroxyl group (-OH) on the spacer portion to form an amide, hydrazide, diacylhydrazine or ester linkage, or direct coupling of the carboxylic

acid group with an amino group on the spacer portion or directly on the carrier with a peptide coupling reagent and/or carbodiimide dehydrating reagent, described previously, examples of which are shown in Tables 4 and 5. Procedures found in references cited previously for formation of activated esters and use of peptide coupling agents may be employed for attachment of carboxylic acid-bearing haptens to spacer building blocks and protein carriers with available amino groups utilizing routine optimization of reaction conditions.

Table 3

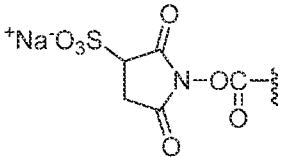
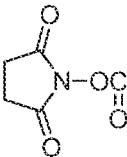
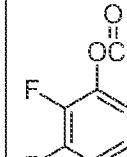
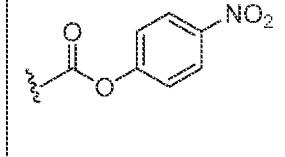
			
Sulfo NHS and NHS	TFP	Acyl chloride	PNP

Table 4

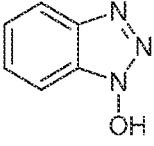
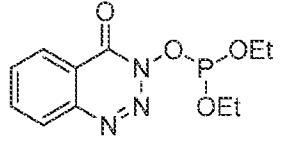
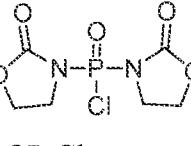
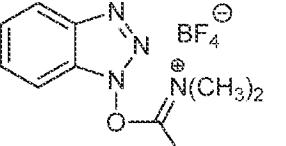
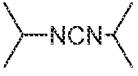
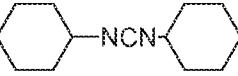
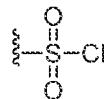
			
HOBT	DEPT	BOP-Cl	TBTU

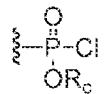
Table 5

		
diisopropylcarbodiimide (DIC)	Dicyclohexylcarbodiimide (DCC)	1-ethyl-3-(3-dimethylaminopropyl)carbodiimide.HCl (EDC)

Other electrophilic groups may be present on the hapten to attach the spacer, for example, a sulfonyl halide

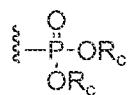


or electrophilic phosphorous group, for example:



See: Malachowski, William P., Coward, James K., *Journal of Organic Chemistry*, 1994, 59 (25):7616

or:

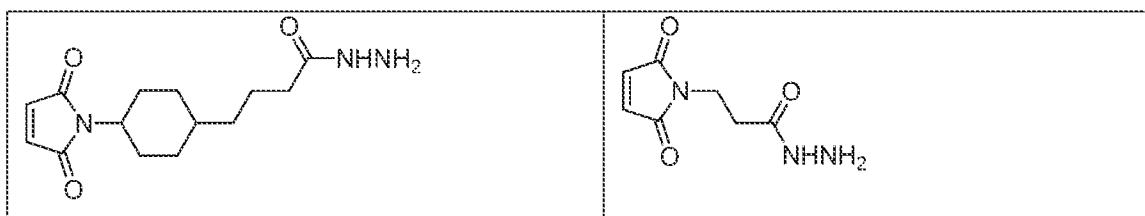


R_c is alkyl, cycloalkyl, aryl, substituted aryl, aralkyl.

See: Aliouane, L., et.al, *Tetrahedron Letters*, 2011, 52(28):8681.

Haptens that bear aldehyde or ketone groups may be attached to spacers using methods including but not limited to reaction with a hydrazide group H₂N-NH-C(O)- on the spacer to form an acylhydrazone, see: Chamow, S.M., Kogan, T.P., Peers, D.H., Hastings, R.C., Byrn, R.A. and Askenaszi, A., *J. Biol. Chem.*, 1992, 267(22): 15916. Examples of bifunctional hydrazide spacer groups that allow attachment to a thiol group on the carrier are shown in Table 6.

Table 6



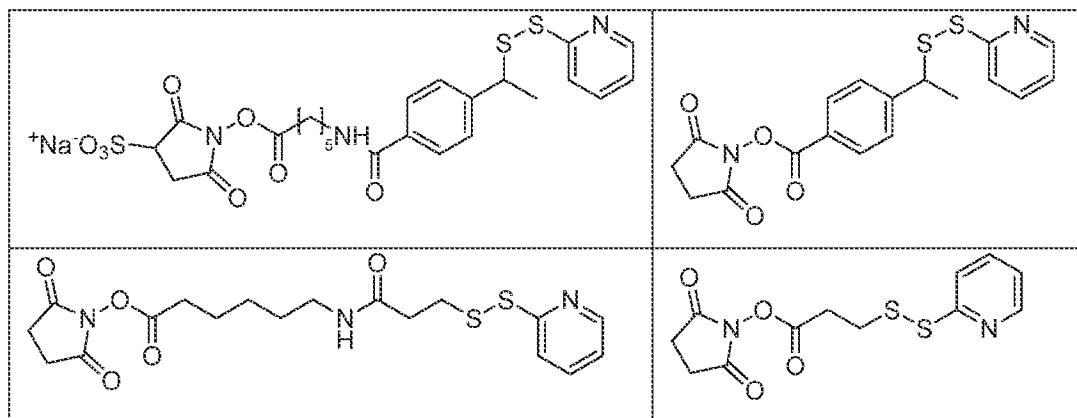
Haptens may also contain thiol groups which may be reacted with the carrier provided that the carrier has been modified to provide a group that may react with the thiol. Carrier groups may be modified by methods including but not limited to attachment of a group containing a maleimide functional group by reaction of an amino group on the carrier with N-Succinimidyl maleimidoacetate, (AMAS, CAS# 55750-61-3), Succinimidyl

iodoacetate (CAS# 151199-81-4), or any of the bifunctional spacer groups shown in Table 1 to introduce a group which may undergo a reaction resulting in attachment of the hapten to the carrier.

The functional linking group capable of forming a bond with the carrier may be any group capable of forming a stable linkage and may be reactive to a number of different groups on the carrier. The functional linking group may preferably react with an amino group, a carboxylic acid group or a thiol group on the carrier, or derivative thereof. Non-limiting examples of the functional linking group are a carboxylic acid group, acyl halide, active ester (as defined previously), isocyanate, isothiocyanate, alkyl halide, amino group, thiol group, maleimide group, acrylate group ($\text{H}_2\text{C}=\text{CH}-\text{C}(\text{O})-$) or vinyl sulfone group $\text{H}_2\text{C}=\text{CH}-\text{SO}_2^-$) See: Park, J.W., et.al., *Bioconjugate Chem.*, 2012, 23(3): 350. The functional linking group may be present as part of a differentially activated spacer building block that may be reacted stepwise with the hapten and the resulting hapten derivative may then be reacted with the carrier. Alternatively, the hapten may be derivatized with a spacer that bears a precursor group that may be transformed into the functional linking group by a subsequent reaction. When the functional linking group on the spacer is an amine or a carboxylic acid group, the coupling reaction with the carboxylic acid group or amine on the carrier may be carried out directly through the use of peptide coupling reagents according to procedures in the references cited above for these reagents.

Particular disulfide groups, for example, pyridyldisulfides, may be used as the functional linking group on the spacer which may undergo exchange with a thiol group on the carrier to form a mixed disulfide linkage, see: Ghetie, V., et al., *Bioconjugate Chem.*, 1990, 1:24-31. These spacers may be attached by reaction of the amine-bearing hapten with an active ester which is attached to a spacer bearing the pyridyldisulfide group, examples of which include but are not limited to those shown in Table 7.

Table 7



Most often the carrier is a protein and the ε-amino groups of the lysine residues may be used for attachment, either directly by reaction with an amine-reactive functional linking group or after derivitization with a thiol-containing group, including *N*-Succinimidyl S-Acetylthioacetate, (SATA, CAS 76931-93-6), or an analogue thereof, followed by cleavage of the acetate group with hydroxylamine to expose the thiol group for reaction with the functional linking group on the hapten. Thiol groups may also be introduced into the carrier by reduction of disulfide bonds within protein carriers with mild reducing reagents including but not limited to 2-mercaptoethylamine, see: Bilah, M., et.al., *Bioelectrochemistry*, 2010, 80(1):49, phosphine reagents, see: Kirley, T.L., *Analytical Biochemistry*, 1989, 180(2):231 or dithioerythritol (DTT, CAS 3483-12-3) Cleland, W., *Biochemistry*, 1964, 3:480–482.

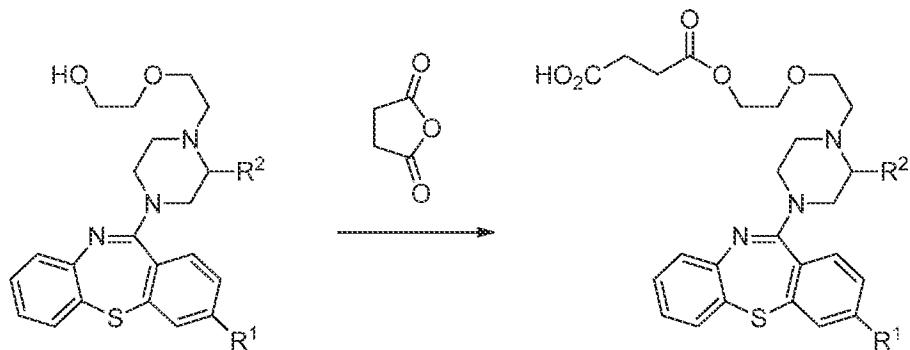
GENERAL REACTION SCHEMES

Representative compounds of the present invention can be synthesized in accordance with the general synthetic methods described below. Compounds of Formula (I) can be prepared by methods known to those who are skilled in the art. The following reaction schemes are only meant to represent examples of the invention and are in no way meant to be a limit of the invention.

Derivatives of quetiapine may be prepared by a number of methods. The primary hydroxyl group in quetiapine, the starting compound (R₁ and R₂ = H) shown in Scheme 1, may be acylated using, for example, succinic anhydride and the method described by

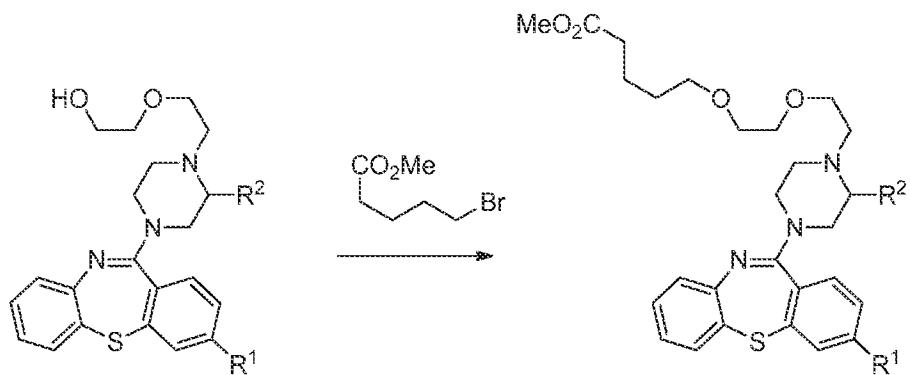
Fiedler, H., et.al., *Langmuir*, 1994, 10:3959. The resulting acid may be further functionalized as described elsewhere within this disclosure or attached directly to a carrier using any number of aforementioned methods including those shown in the subsequent examples.

Scheme 1

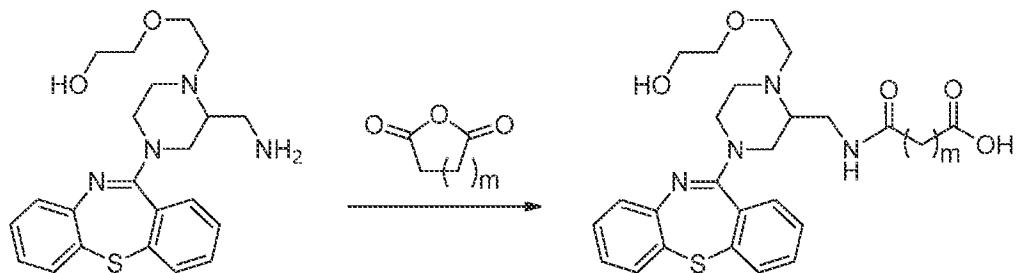


The Primary hydroxyl group of quetiapine may also be alkylated to form an ether according to the procedure of US20100069356, as shown in Scheme 2, using an alkyl halide or a sulfonate ester, such as 4-bromomethylpentanoate in the presence of tetrabutylammonium hydrogensulfate and aqueous sodium hydroxide to provide an acid which may be used as described above.

Scheme 2

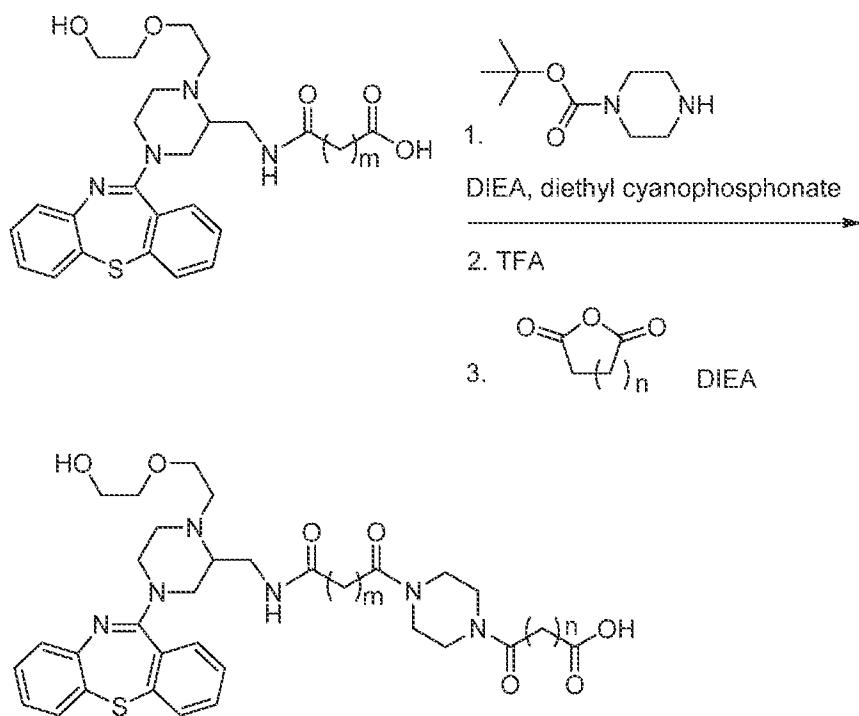


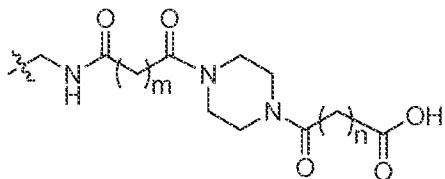
Scheme 3



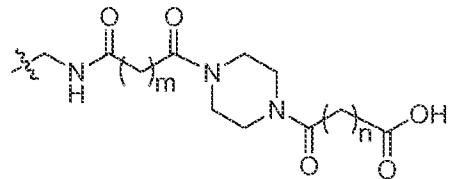
Compounds of Formula I where R² is CH₂NHC(O)(CH₂)_mCO₂H may be made according to Scheme 3. Reaction of 2-(2-(2-(aminomethyl)-4-(dibenzo[b,f][1,4]thiazepin-11-yl)piperazin-1-yl)ethoxy)ethanol, prepared as described in Example 1, proceeds with a cyclic anhydride compound, such as succinic anhydride or glutaric anhydride, in a solvent such as pyridine, at temperatures ranging from room temperature to 60 °C, for about 48 hours. Those skilled in the art will recognize that the same chemistry may be used to create compounds of Formula I where R¹ is CH₂NHC(O)(CH₂)_mCO₂H.

Scheme 4



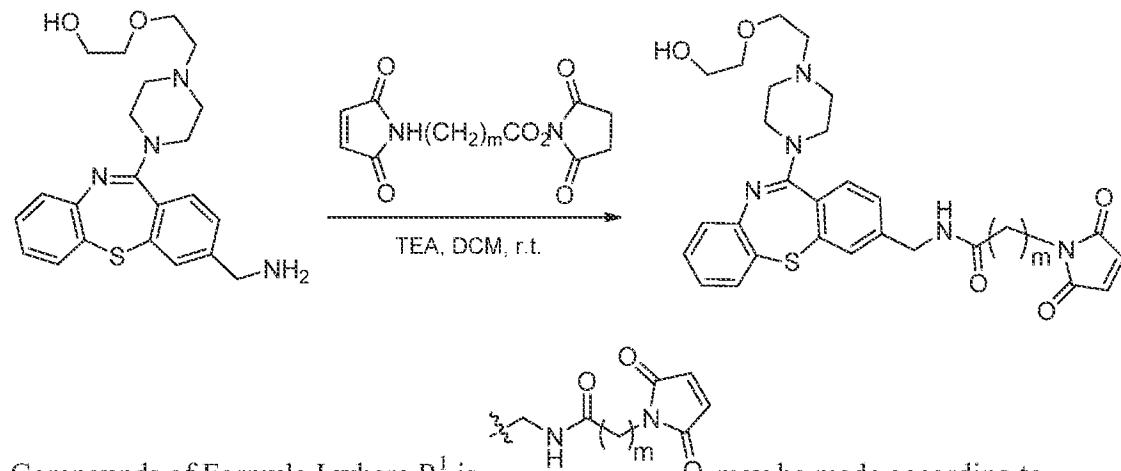


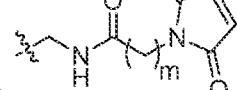
Compounds of Formula I where R^2 is  may be made according to Scheme 4. Compounds of Formula I, where R^2 is $CH_2NHC(O)(CH_2)_mCO_2H$, prepared as described in scheme 1, are treated with *N*-*t*-butoxycarbonylpiperazine, diethyl cyanophosphonate, and a base, such as diisopropylethylamine. The reaction is carried out in a solvent, such as dichloromethane, for about 2 hours at room temperature. Deprotection of the piperazinyl group is accomplished with trifluoroacetic anhydride as described in Scheme 4, followed by reaction with an appropriate anhydride, such as succinic anhydride or maleic anhydride, in the presence of a suitable base such as diisopropylethylamine. Those skilled in the art will recognize that the same chemistry may be used to create compounds of Formula I



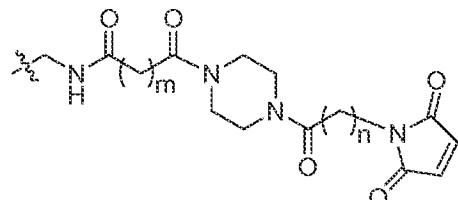
where R^1 is .

Scheme 5

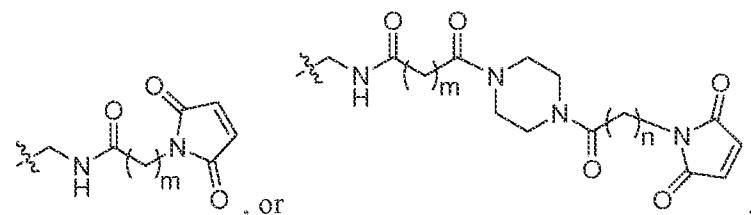


Compounds of Formula I where R^1 is  may be made according to Scheme 5. The maleimide may be introduced by any method known in the art. Maleimide functionalizing groups such as 2,5-dioxopyrrolidin-1-yl 2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)acetate where m is 1, may be used in a solvent such as DMF or

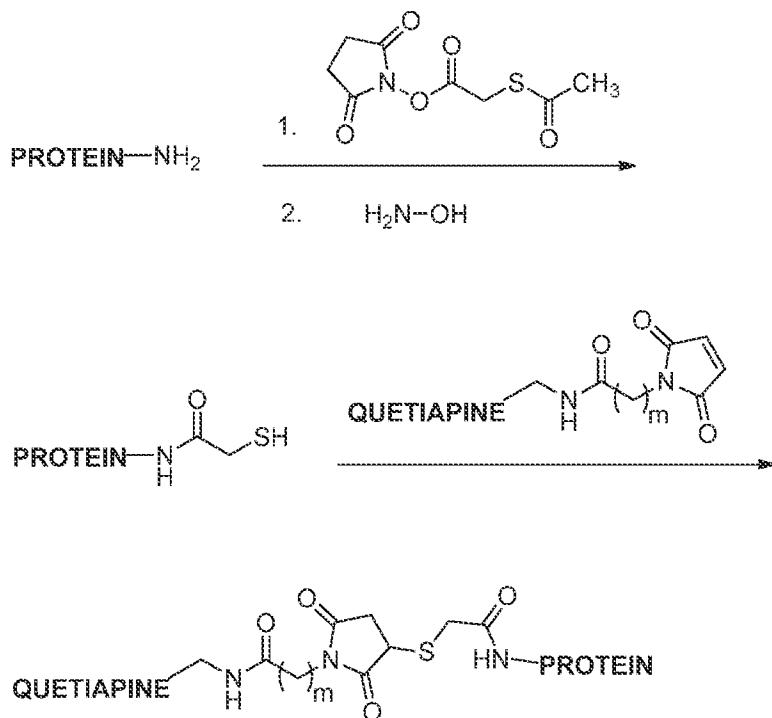
CH_2Cl_2 , and a base, such as tributylamine or triethylamine. Alternatively, the deprotected piperazinyl group described in Scheme 4 may be elaborated with a maleimide functionality, as described in Scheme 5 to give compounds of Formula I

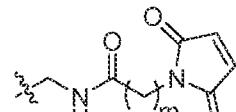


where R^1 is . Those skilled in the art will recognize that the same chemistry may be used to create compounds of Formula I where R^2 is

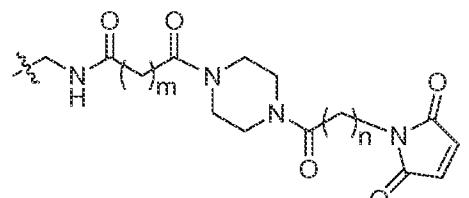


Scheme 6



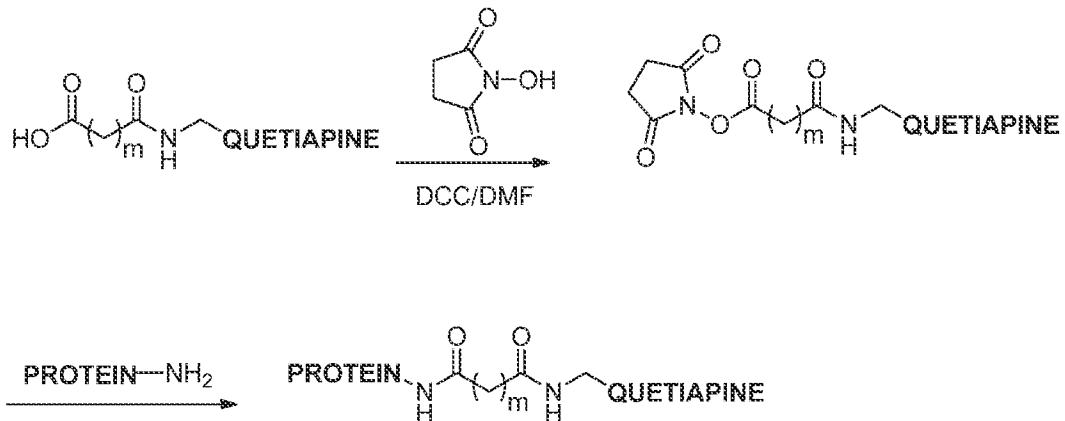


Maleimide functionalized haptens wherein R^1 or R^2 is conjugated to proteins according to the method shown in Scheme 6. Activation of protein lysine residues by acylation of the epsilon-nitrogen with N-succinimidyl S-acetylthioacetate (SATA), followed by subsequent hydrolysis of the S-acetyl group with hydroxylamine produces a nucleophilic sulphydryl group. Conjugation of the sulphydryl activated protein with the maleimide derivatized hapten (prepared as described in general scheme 3) proceeds via a Michael addition reaction. Suitable proteins are known to those skilled in the art and include keyhole limpet hemocyanin, bovine thyroglobulin, and ovalbumin. The same methodology may be used to conjugate proteins to maleimide



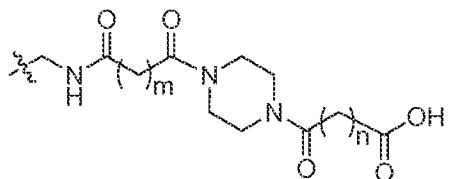
functionalized haptens where R^1 or R^2 is

Scheme 7:



Carboxylic acid functionalized haptens, wherein R^1 or R^2 is $CH_2NHC(O)(CH_2)_mCO_2H$, may be conjugated to proteins according to the method shown in Scheme 7. Reaction with N-hydroxysuccinimide and a suitable coupling agent, such as dicyclohexylcarbodiimide, and a base, such as tributyl amine, in a solvent such as DMF,

at a temperature of about 20 °C, for about 18 hrs activates the carboxylic acid with the hydroxypyrrolidine-2,5-dione leaving group. The activated linker and hapten may then be conjugated to a protein in a solvent, such as a pH 7.5 phosphate buffer, at about 20 °C, for about 2.5 hours. Suitable proteins are known to those skilled in the art and include keyhole limpet hemocyanin, bovine thyroglobulin, and ovalbumin. The same methodology may be used to conjugate proteins to carboxylic acid functionalized haptens



where R^1 or R^2 is

ANTIBODY PRODUCTION

The conjugates above are useful for the production of antibodies which bind the anti-psychotic drug to which they were generated (quetiapine). These antibodies can be used in assays to detect the presence and/or amount of the anti-psychotic drug in patient samples. Such detection permits therapeutic drug monitoring enabling all of the benefits thereof. Detection of levels of anti-psychotic drugs may be useful for many purposes, including: detection in combination with the detection of other anti-psychotic drugs, including those selected from the group consisting of risperidone, paliperidone, aripiprazole, olanzapine, and metabolites thereof, such detection permitting the simultaneous measurement of these anti-psychotic drugs; determination of patient adherence or compliance with prescribed therapy; use as a decision tool to determine whether a patient should be converted from an oral anti-psychotic regimen to a long-acting injectable anti-psychotic regimen; use as a decision tool to determine if the dose level or dosing interval of oral or injectable anti-psychotics should be increased or decreased to ensure attainment or maintenance of efficacious or safe drug levels; use as an aid in the initiation of anti-psychotic drug therapy by providing evidence of the attainment of minimum pK levels; use to determine bioequivalence of anti-psychotic drug in multiple formulations or from multiple sources; use to assess the impact of polypharmacy and potential drug-drug interactions; and use as an indication that a patient

should be excluded from or included in a clinical trial and as an aid in the subsequent monitoring of adherence to clinical trial medication requirements.

Having provided the conjugates of the subject invention, which comprise the compounds herein and an immunogenic carrier, antibodies can be generated, e.g., polyclonal, monoclonal, chimeric, and humanized antibodies, that bind to the anti-psychotic drug. Such antibodies that are particularly contemplated include monoclonal and polyclonal antibodies as well as fragments thereof, e.g., recombinant proteins, containing the antigen-binding domain and/or one or more complementarity determining regions of these antibodies. Preferably, the antibody will bind to the drug and any desired pharmacologically active metabolites. By altering the location of the attachment of an immunogenic carrier in a drug conjugate, selectivity and cross-reactivity with metabolites and/or related drugs can be engineered into the antibodies. For quetiapine, cross-reactivity with quetiapine metabolites such as N-desalkylquetiapine (norquetiapine), quetiapine sulfoxide, O-desalkylquetiapine or 7-hydroxy quetiapine may or may not be desirable. Antibodies may be generated that detect multiple ones of these drugs and/or metabolites, or antibodies may be generated that detect each separately (thus defining the antibody "specific binding" properties). An antibody specifically binds one or more compounds when its binding of the one or more compounds is equimolar or substantially equimolar.

Methods of producing such antibodies comprise inoculating a host with the conjugate (the compound and the immunogenic carrier being an immunogen) embodying features of the present invention. Suitable hosts include, but are not limited to, mice, rats, hamsters, guinea pigs, rabbits, chickens, donkeys, horses, monkeys, chimpanzees, orangutans, gorillas, humans, and any species capable of mounting a mature immune response. The immunization procedures are well established in the art and are set forth in numerous treatises and publications including "*The Immunoassay Handbook*", 2nd Edition, edited by David Wild (Nature Publishing Group, 2000) and the references cited therein.

Preferably, an immunogen embodying features of the present invention is administered to a host subject, e.g., an animal or human, in combination with an adjuvant. Suitable adjuvants include, but are not limited to, Freund's adjuvant, powdered aluminum hydroxide (alum), aluminum hydroxide together with *Bordetella pertussis*, and monophosphoryl lipid A-synthetic trehalose dicorynomycolate (MPL-TDM).

Polyclonal antibodies can be raised in a mammalian host by one or more injections of an immunogen which can optionally be administered together with an adjuvant. Typically, an immunogen or a combination of an immunogen and an adjuvant is injected into a mammalian host by one or multiple subcutaneous or intraperitoneal injections. Preferably, the immunization program is carried out over at least one week, and more preferably, over two or more weeks. Polyclonal antibodies produced in this manner can be isolated and purified utilizing methods well known in the art.

Monoclonal antibodies can be produced by the well-established hybridoma methods of Kohler and Milstein, e.g., *Nature* 256:495-497 (1975). Hybridoma methods typically involve immunizing a host or lymphocytes from a host, harvesting the monoclonal antibody secreting or having the potential to secrete lymphocytes, fusing the lymphocytes to immortalized cells, and selecting cells that secrete the desired monoclonal antibody.

A host can be immunized to elicit lymphocytes that produce or are capable of producing antibodies specific for an immunogen. Alternatively, the lymphocytes can be immunized *in vitro*. If human cells are desired, peripheral blood lymphocytes can be used, although spleen cells or lymphocytes from other mammalian sources are preferred.

The lymphocytes can be fused with an immortalized cell line to form hybridoma cells, a process which can be facilitated by the use of a fusing agent, e.g., polyethylene glycol. By way of illustration, mutant rodent, bovine, or human myeloma cells immortalized by transformation can be used. Substantially pure populations of hybridoma cells, as opposed to unfused immortalized cells, are preferred. Thus, following fusion, the cells can be grown in a suitable medium that inhibits the growth or survival of unfused,

immortalized cells, for example, by using mutant myeloma cells that lack the enzyme hypoxanthine guanine phosphoribosyl transferase (HGPRT). In such an instance, hypoxanthine, aminopterin, and thymidine can be added to the medium (HAT medium) to prevent the growth of HGPRT-deficient cells while permitting hybridomas to grow.

Preferably, immortalized cells fuse efficiently, can be isolated from mixed populations by selection in a medium such as HAT, and support stable and high-level expression of antibody following fusion. Preferred immortalized cell lines include myeloma cell lines available from the American Type Culture Collection, Manassas, VA.

Because hybridoma cells typically secrete antibody extracellularly, the culture media can be assayed for the presence of monoclonal antibodies specific for the anti-psychotic drug. Immunoprecipitation of *in vitro* binding assays, for example, radioimmunoassay (RIA) or enzyme-linked immunosorbent assay (ELISA), can be used to measure the binding specificity of monoclonal antibodies.

Monoclonal antibody-secreting hybridoma cells can be isolated as single clones by limiting dilution procedures and sub-cultured. Suitable culture media include, but are not limited to, Dulbecco's Modified Eagle's Medium, RPMI-1640, and polypeptide-free, polypeptide-reduced, or serum-free media, e.g., Ultra DOMA PF or HL-1, available from Biowhittaker, Walkersville, MD. Alternatively, the hybridoma cells can be grown *in vivo* as ascites.

Monoclonal antibodies can be isolated and/or purified from a culture medium or ascites fluid by conventional immunoglobulin (Ig) purification procedures including, but not limited to, polypeptide A-SEPHAROSE, hydroxylapatite chromatography, gel electrophoresis, dialysis, ammonium sulfate precipitation, and affinity chromatography.

Monoclonal antibodies can also be produced by recombinant methods such as are described in U.S. Patent No. 4,166,452. DNA encoding monoclonal antibodies can be isolated and sequenced using conventional procedures, e.g., using oligonucleotide probes

that specifically bind to murine heavy and light antibody chain genes, preferably to probe DNA isolated from monoclonal antibody hybridoma cells lines secreting antibodies specific for anti-psychotic drugs.

IMMUNOASSAYS

The antibodies thus produced can be used in immunoassays to recognize/bind to the anti-psychotic drug, thereby detecting the presence and/or amount of the drug in a patient sample. Preferably, the assay format is a competitive immunoassay format. Such an assay format and other assays are described, among other places, in Hampton et al. (*Serological Methods, A Laboratory Manual*, APS Press, St. Paul, MN 1990) and Maddox et al. (*J. Exp. Med.* 158:12111, 1983).

A reagent kit can also be provided comprising an antibody as described above. A representative reagent kit may comprise an antibody that binds to the anti-psychotic drug, quetiapine, a complex comprising an analog of an anti-psychotic drug or a derivative thereof coupled to a labeling moiety, and may optionally also comprise one or more calibrators comprising a known amount of an anti-psychotic drug or a related standard.

As noted above, reagent kits may comprise calibrators and/or control materials which comprise a known amount of the analyte to be measured. The concentration of the analyte can be calculated by comparing results obtained for a sample with results obtained for a standard. A calibration curve can be constructed and used for relating the sets of results and for determining the concentration of an analyte in a sample.

Any sample that is suspected of containing an analyte, e.g., an anti-psychotic drug, can be analyzed in accordance with the methods of the presently preferred embodiments. The sample can be pretreated if desired and can be prepared in any convenient medium that does not interfere with the assay. Preferably, the sample comprises an aqueous medium such as a body fluid from a host, most preferably plasma or serum.

Copending applications entitled "Haptens of Aripiprazole" (Attorney Docket No. PRD3265USPSP, first named inventor: Remmerie), "Haptens of Olanzapine" (Attorney Docket No. PRD3266USPSP, first named inventor: Remmerie), "Haptens of Paliperidone" (Attorney Docket No. PRD3267USPSP, first named inventor: Remmerie), "Haptens of Quetiapine" (Attorney Docket No. PRD3268USPSP, first named inventor: Remmerie), "Haptens of Risperidone and Paliperidone" (Attorney Docket No. PRD3269USPSP, first named inventor: Remmerie), "Antibodies to Aripiprazole Haptens and Use Thereof" (Attorney Docket No. CDS5128USPSP, first named inventor: Hryhorenko), "Antibodies to Olanzapine Haptens and Use Thereof" (Attorney Docket No. CDS5132USPSP, first named inventor: Hryhorenko), "Antibodies to Paliperidone Haptens and Use Thereof" (Attorney Docket No. CDS5126USPSP, first named inventor: Hryhorenko), "Antibodies to Quetiapine Haptens and Use Thereof" (Attorney Docket No. CDS5134USPSP, first named inventor: Hryhorenko), "Antibodies to Risperidone Haptens and Use Thereof" (Attorney Docket No. CDS5130USPSP, first named inventor: Hryhorenko), "Antibodies to Aripiprazole and Use Thereof" (Attorney Docket No. CDS5129USPSP, first named inventor: Hryhorenko), "Antibodies to Olanzapine and Use Thereof" (Attorney Docket No. CDS5133USPSP, first named inventor: Hryhorenko), "Antibodies to Paliperidone and Use Thereof" (Attorney Docket No. CDS5127USPSP, first named inventor: Hryhorenko), "Antibodies to Quetiapine and Use Thereof" (Attorney Docket No. CDS5135USPSP, first named inventor: Hryhorenko), "Antibodies to Risperidone and Use Thereof" (Attorney Docket No. CDS5131USPSP, first named inventor: Hryhorenko), all filed concurrently herewith are all incorporated herein by reference in their entireties.

EXAMPLES

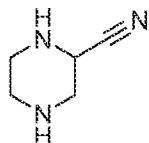
Representative compounds of the present invention can be synthesized in accordance with the general synthetic methods described below. Compounds of Formula (I) can be prepared by methods known to those who are skilled in the art. The following examples are only meant to represent examples of the invention and are in no way meant to be a limit of the invention.

Example 1

2-(2-(2-(aminomethyl)-4-(dibenzo[b,f][1,4]thiazepin-11-yl)piperazin-1-yl)ethoxy)ethanol

Step A

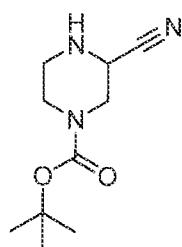
Piperazine-2-carbonitrile



A stirred solution of tetrahydrofuran (300 mL) and ethylenediamine (108.2 g) at 30 °C was treated dropwise with 2-chloroacrylonitrile (105.0 g) over a period of 2 hours and stirred for 6 additional hours at 30 °C. The reaction mixture was cooled to 20 °C and a precipitate formed. The reaction was filtered, and the filtrate was adjust pH to 4 by adding 35% hydrochloric acid. The resulting precipitate was collected by filtration. The combined precipitates were dissolved in 20% hydrochloric acid solution and then poured into THF solution to precipitate the title compound, which was dried under reduced pressure and used in the next reaction without additional purification. ¹H NMR: (D₂O, 400 MHz): δ (ppm) 5.00-4.97 (m, 1H), 3.79 (d, J=4.8 Hz, 2H), 3.62-3.44 (m, 4H).

Step B

tert-Butyl 3-cyanopiperazine-1-carboxylate

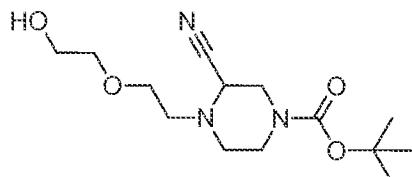


To a solution of compound piperazine-2-carbonitrile, prepared as described in the previous step, (90.6 g, 0.492 mol) was added triethylamine (206 mL, 1.476 mol) and Boc₂O (117 g, 0.542 mol). The reaction mixture was stirred at room temperature overnight, and then concentrated. The residue was purified by silica gel chromatography to provide the title compound.

¹H NMR: (CDCl₃, 400 MHz): δ (ppm) 4.06-3.91 (m, 3H), 3.28-2.83 (m, 4H), 1.47 (s, 9H).

Step C

tert-Butyl 3-cyano-4-(2-(2-hydroxyethoxy)ethyl)piperazine-1-carboxylate

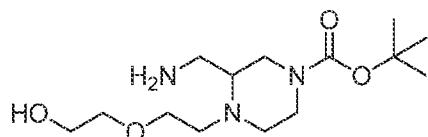


A solution of tert-butyl 3-cyanopiperazine-1-carboxylate, prepared as described in the previous step, (10 g, 0.047 mol) and 2-(2-hydroxyethoxy)acetaldehyde (14.8 g) (see: Bodin, A., *Contact Dermatitis*, 2001, 44:207) in dichloromethane was treated with formic acid (12.7 g), and the reaction mixture was stirred at room temperature overnight. Sodium cyanoborohydride (7.2 g, 0.118 mol) was added in portions. The reaction mixture was stirred at room temperature for 3 hours followed by the addition of water and extraction with dichloromethane. The organic layer was washed with brine, dried over sodium sulfate, filtered, and concentrated. The crude product was purified by column chromatography to provide the product.

¹H NMR: (CDCl₃, 400 MHz): δ (ppm) 4.15 (s, 1H), 3.69-3.63 (m, 4H), 3.58 (d, J=4.4 Hz, 2H), 3.47-3.44 (m, 4H), 2.61 (d, J=5.2 Hz, 2H), 2.51-2.48 (m, 4H), 1.43 (s, 9H).

Step D

tert-Butyl 3-(aminomethyl)-4-(2-(2-hydroxyethoxy)ethyl)piperazine-1-carboxylate



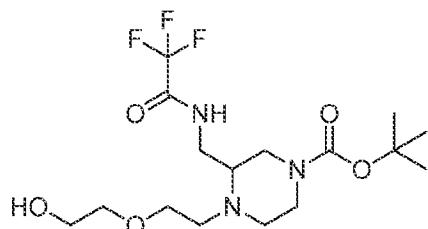
To a solution of tert-butyl 3-cyano-4-(2-(2-hydroxyethoxy)ethyl)piperazine-1-carboxylate, prepared as described in the previous step, (9.9 g, 33.1 mmol) in methanol (20 mL) was added Raney Ni (15 g). The reaction solution was stirred at room temperature overnight under hydrogen atmosphere (50 psi). The mixture was filtered and

concentrated to provide the product, which was used in the next step without additional purification.

ESI-MS (M+1): 304 calc. for C₁₄H₂₉N₃O₄ 303.

Step E

tert-Butyl 4-(2-(2-hydroxyethoxy)ethyl)-3-((2,2,2-trifluoroacetamido)methyl)piperazine-1-carboxylate

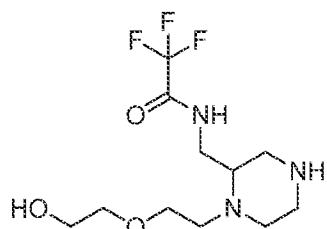


To a solution of tert-butyl 3-(aminomethyl)-4-(2-(2-hydroxyethoxy)ethyl)piperazine-1-carboxylate, prepared as described in the previous step (8.8 g) in dichloromethane (100 mL) was added triethylamine (8.8 g, 87.0 mmol) and trifluoroacetic anhydride (6.1 g, 29.0 mmol). The reaction mixture was stirred at room temperature for 12h, diluted with dichloromethane and washed with water. The organic layer was washed with brine, dried over sodium sulfate, filtered, and concentrated to give the crude product which was purified by column chromatography to provide the title compound.

ESI-MS (M+1): 400 calc. for C₁₆H₂₈F₃N₃O₅ 399.

Step F

2,2,2-Trifluoro-N-((1-(2-(2-hydroxyethoxy)ethyl)piperazin-2-yl)methyl)acetamide

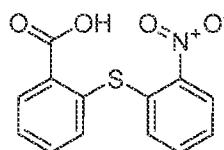


A solution of tert-butyl 4-(2-(2-hydroxyethoxy)ethyl)-3-((2,2,2-trifluoroacetamido)methyl)piperazine-1-carboxylate, prepared as described in the previous step, (8.6 g, crude) in methanolic hydrogen chloride (20 mL) was stirred at room

temperature for 1 hour, followed by concentration to provide the title compound which was used without further purification. ESI-MS (M+1): 300 calc. for $C_{11}H_{20}F_3N_3O_3$ 299.

Step G

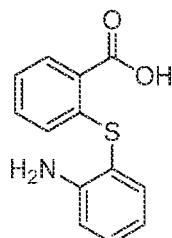
2-((2-Nitrophenyl)thio)benzoic acid



To a solution of 2-mercaptop-benzoic acid (30 g, 0.195 mol) in isopropanol (500 mL) at room temperature were added 1-fluoro-2-nitro-benzene (30.2 g, 0.214 mol), water (100 mL) and potassium hydroxide (31.1 g, 0.555 mol). The reaction mixture was stirred at room temperature overnight, quenched with water and diluted with ethyl acetate. The aqueous phase was extracted with ethyl acetate (3 x 400 mL) and the combined organic extracts were washed with saturated aqueous sodium chloride (500 mL), dried over magnesium sulfate, filtered, and concentrated. The crude residue was purified by flash column chromatography on silica gel to give the title compound. ESI-MS (M+1): 276 calc. for $C_{13}H_9NO_4S$ 275. 1H NMR: ($CDCl_3$, 400 MHz): δ (ppm) 8.12-8.07 (m, 2H), 7.54-7.43 (m, 2H), 7.42-7.39 (m, 2H), 7.35-7.31 (m, 1H), 7.12-7.09 (m, 1H).

Step H

2-((2-Aminophenyl)thio)benzoic acid

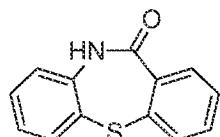


To a solution of 2-((2-nitrophenyl)thio)benzoic acid, prepared as described in the previous step, (43.3 g, 0.157 mol) in ethyl acetate (500 mL) was added Pd/C (8 g). The reaction solution was stirred at room temperature overnight under hydrogen gas atmosphere. The mixture was filtered and concentrated to provide the title compound. ESI-MS (M+1): 246 calc. for $C_{13}H_{11}NO_2S$ 245. 1H NMR: ($CDCl_3$, 400 MHz): δ (ppm)

8.20-8.17 (m, 1H), 7.51-7.48 (m, 1H), 7.36-7.30 (m, 2H), 7.21-7.17 (m, 1H), 6.88-6.80 (m, 3H).

Step I

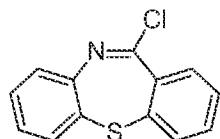
Dibenzo[b,f][1,4]thiazepin-11(10H)-one



To a solution of 2-((2-aminophenyl)thio)benzoic acid, prepared as described in the previous step, (30 g, 0.122 mol) in dichloromethane (300 mL) was added EDCI (35.2 g, 0.183 mol), triethylamine (51 mL, 0.366 mol) and HOBT (24.7 g, 0.183 mol). The reaction mixture was stirred at room temperature for 12 hours, washed with 1M aq. HCl, saturated aqueous sodium bicarbonate, saturated aqueous sodium chloride, and dried over MgSO₄. The solution was filtered, concentrated, and purified by column chromatography to provide the title compound. ESI-MS (M+1): 228 calc. for C₁₃H₉NOS 227. ¹H NMR: (CDCl₃, 400 MHz): δ (ppm) 7.70-7.67 (m, 1H), 7.58-7.52 (m, 2H), 7.50-7.42 (m, 2H), 7.39-7.35 (m, 1H), 7.24-7.22 (m, 1H), 7.17-7.13 (m, 1H).

Step J

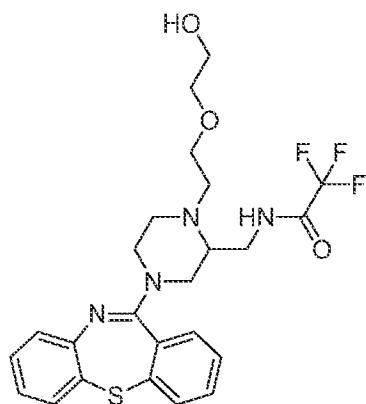
11-Chlorodibenzo[b,f][1,4]thiazepine



A solution of dibenzo[b,f][1,4]thiazepin-11(10H)-one, prepared as described in the previous step, (14.6 g, 64 mmol) in phosphorus oxychloride (20 mL) was heated to reflux for 2 hours. The mixture was concentrated to provide the crude product which was used directly without further purification. ESI-MS (M+1): 246 calc. for C₁₃H₈ClNS 245.

Step K

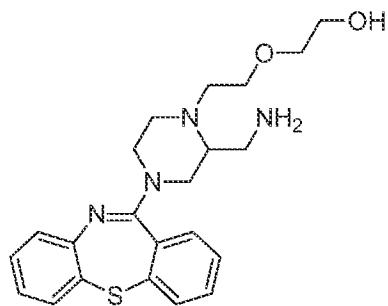
N-((4-(Dibenzo[b,f][1,4]thiazepin-11-yl)-1-(2-(2-hydroxyethoxy)ethyl)piperazin-2-yl)methyl)-2,2,2-trifluoroacetamide



To a solution of 11-chlorodibenzo[b,f][1,4]thiazepine, prepared as described in the previous step, (2 g, crude) in dioxane (20 mL) was added Pd₂(dba)₃ (327 mg, 0.357 mmol), BINAP (225 mg, 0.357 mmol), triethylamine (6 mL, 42.9 mmol) and 2,2,2-trifluoro-N-((1-(2-(2-hydroxyethoxy)ethyl)piperazin-2-yl)methyl)acetamide, prepared as described as Step F, (2.4 g, crude). The resulting mixture was heated to reflux overnight under nitrogen atmosphere, filtered through CELITE™, and concentrated. The residue was purified by silica gel chromatography to provide the title compound. ESI-MS (M+1): 509 calc. for C₂₄H₂₇F₃N₄O₃S 508.

Step L

2-(2-(2-(Aminomethyl)-4-(dibenzo[b,f][1,4]thiazepin-11-yl)piperazin-1-yl)ethoxy)ethanol

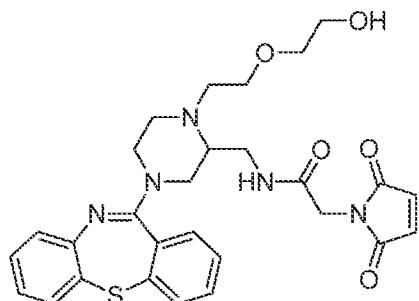


A mixture of N-((4-(dibenzo[b,f][1,4]thiazepin-11-yl)-1-(2-(2-hydroxyethoxy)ethyl)piperazin-2-yl)methyl)-2,2,2-trifluoroacetamide, prepared as described in the previous step, (2.0 g) and aqueous potassium carbonate (5%) (15 mL) in methanol (20 mL) was stirred at room temperature for 18 hours and extracted with ethyl acetate. The organic layers were washed with saturated aqueous sodium chloride, dried

over sodium sulfate, filtered, evaporated to give the crude product which was purified by column chromatography, and followed by prep-HPLC to provide the title compound as a yellow solid. ESI-MS (M+1): 413 calc. for C₂₂H₂₈N₄O₂S 412. ¹H NMR: (CDCl₃, 400 MHz): δ (ppm) 7.52-7.50 (m, 1H), 7.41-7.31 (m, 4H), 7.17-7.12 (m, 1H), 7.02-7.00 (m, 1H), 6.89-6.84 (m, 1H), 3.66-3.59 (m, 5H), 3.54-3.51 (m, 2H), 3.49-3.38 (m, 1H), 3.19-3.12 (m, 1H), 3.03-2.88 (m, 2H), 2.79-2.53 (m, 5H).

Example 2

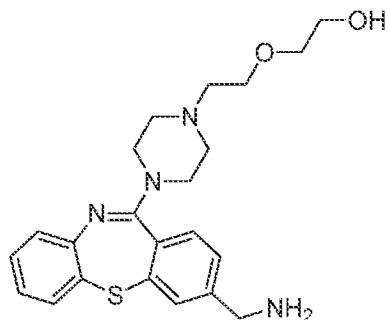
N-((4-(Dibenzo[b,f][1,4]thiazepin-11-yl)-1-(2-(2-hydroxyethoxy)ethyl)piperazin-2-yl)methyl)-2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)acetamide



To a solution of 2-(2-(2-(aminomethyl)-4-(dibenzo[b,f][1,4]thiazepin-11-yl)piperazin-1-yl)ethoxy)ethanol, prepared as described in Example 1, (7.8 mg, 19.0 μmoles) in 410 μL of DMF and 8.9 μL of tributylamine was added 480 μL of a DMF solution of N-(α-maleimidoacetoxy) succinimide ester (AMAS, 10 mg/mL, 4.8 mg, 19.0 μmoles). The resulting solution was allowed to stir for 60 minutes at 20 °C, then used as such in conjugation reaction with thiol-activated protein.

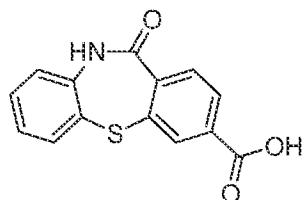
Example 3

2-{2-[4-(3-Aminomethyl-dibenzo[b,f][1,4]thiazepin-11-yl)-piperazin-1-yl]-ethoxy}-ethanol



Step A

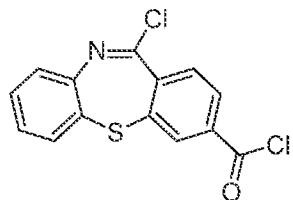
11-Oxo-10,11-dihydrodibenzo[b,f][1,4]thiazepine-3-carboxylic acid



A mixture of 2-amino-benzenethiol (1.34 mL, 12.5 mmol), 2-bromo-terephthalic acid (1.54 g, 6.3 mmol), cuprous oxide (0.50 g, 3.5 mmol), quinoline (6.3 mL), and pyridine (0.63 mL) was heated in a 180 °C oil bath under nitrogen for 20 hours, then cooled to room temperature. Concentrated hydrochloric acid (20 mL) was added slowly while cooling in cold water, with stirring. The resulting precipitate was filtered, washed with water, and dried to give crude title compound (2 g). LC-MS: m/z 270 (M-1).

Step B

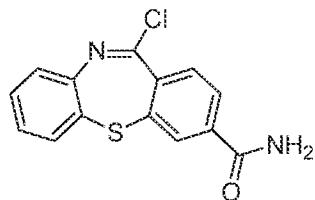
11-Chloro-dibenzo[b,f][1,4]thiazepine-3-carbonyl chloride



To a suspension of 11-oxo-10,11-dihydrodibenzo[b,f][1,4]thiazepine-3-carboxylic acid, prepared as described in the previous step, (0.41 g) in toluene (6.5 mL) was added DMF (0.125 mL) and thionyl chloride (6.5 mL). The mixture was heated in an 80 °C oil bath under nitrogen over night. The resulting solution was concentrated to dryness. The crude product was used for next step.

Step C

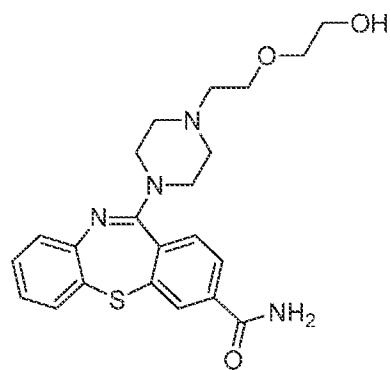
11-Chloro-dibenzo[b,f][1,4]thiazepine-3-carboxylic acid amide



A solution of 11-chloro-dibenzo[b,f][1,4]thiazepine-3-carbonyl chloride, prepared as described in the previous step, (ca 1.5 mmol) in dichloromethane (10 mL) was treated with a 1,4-dioxane solution of ammonia (0.5 M, 9 mL) under ice bath. The resulting suspension was stirred at room temperature for 1 hour, and the reaction was quenched with water (10 mL). The resulting precipitate was filtered, washed with water and dichloromethane, and dried. The organic layer of the filtrate was washed with saturated aqueous sodium bicarbonate solution and concentrated to additional off white product, which was used in the next step without additional purification. LC-MS: m/z 289 (M+1). ^1H NMR (DMSO-d₆, 400 MHz): δ (ppm) 8.19 (br, 1H), 8.00-7.96 (m, 2H), 7.90 (d, 1H), 7.64 (br, 1H), 7.56 (m, 1H), 7.47 (m, 1H), 7.31 (m, 2H).

Step D

11-{4-[2-(2-Hydroxy-ethoxy)-ethyl]-piperazin-1-yl}-dibenzo[b,f][1,4]thiazepine-3-carboxylic acid amide

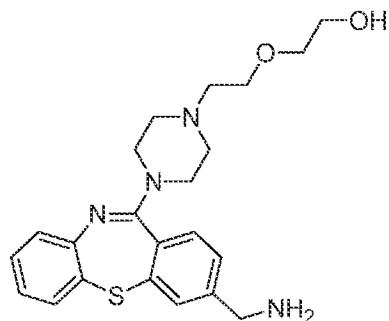


To a solution of 11-chloro-dibenzo[b,f][1,4]thiazepine-3-carboxylic acid amide, prepared as described in the previous step, (0.40 g) in DMF (1.5 mL) and toluene (1.5 mL) was added 2-(2-piperazin-1-yl-ethoxy)-ethanol (0. 50 g, 2.9 mmol). The solution was heated in a 110 °C oil bath under nitrogen for 5 hours, concentrated, and purified (silica gel, 2-

5% methanol-dichloromethane containing ammonia eluent) to give the title compound as an off white solid. LC-MS: m/z 427 (M+1).

Step E

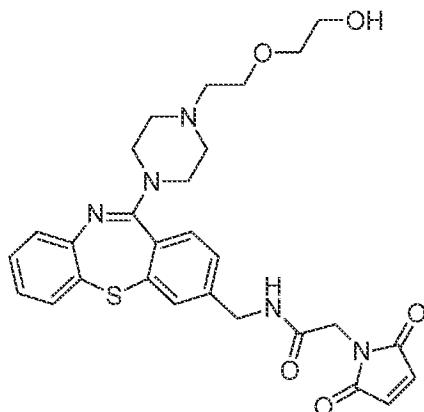
2-{2-[4-(3-Aminomethyl-dibenzo[b,f][1,4]thiazepin-11-yl)-piperazin-1-yl]-ethoxy}-ethanol



To a solution of 2-{2-[4-(3-aminomethyl-dibenzo[b,f][1,4]thiazepin-11-yl)-piperazin-1-yl]-ethoxy}-ethanol, prepared as described in the previous step, (0.24 g, 0.56 mmol) in THF (15 mL) was added 1 M lithium aluminum hydride THF solution (6 mL, 6 mmol). The white suspension was heated in a 70 °C oil bath under nitrogen for 2 hours. The reaction suspension was quenched with slow addition of saturated aqueous sodium sulfate solution under ice bath. The solution phase was separated, and solid was extracted with THF (5 X 10 mL). The combined organic phases were concentrated and purified (silica gel, 2-5% methanol-dichloromethane containing ammonia eluent) to give the title compound as an off white solid. LC-MS: m/z 413 (M+1). ¹H NMR (CDCl₃, 400 MHz) δ (ppm) 7.47 (s, 1H), 7.38 (m, 1H), 7.26 (m, 2H, overlapped with solvent), 7.17 (m, 1H), 7.06 (m, 1H), 6.88 (m, 1H), 3.85 (s, 2H), 3.76-3.46 (m, 11H, containing exchangeable protons), 2.66-2.57 (m, 8H).

Example 4

2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)-N-((11-(4-(2-(2-hydroxyethoxy)ethyl)piperazin-1-yl)dibenzo[b,f][1,4]thiazepin-3-yl)methyl)acetamide



To a solution of 2-{2-[4-(3-aminomethyl-dibenzo[b,f][1,4]thiazepin-11-yl)-piperazin-1-yl]-ethoxy}-ethanol, prepared as described in Example 3, (5.6 mg, 13.6 μ moles) in 295 μ L of DMF and 6.4 μ L of tributylamine was added 340 μ L of a DMF solution of N-(α -maleimidoacetoxy)succinimide ester (AMAS, 10 mg/mL, 3.4 mg, 13.6 μ moles). The resulting solution was allowed to stir for 60 minutes at 20 °C, then used as such in conjugation reaction with thiol-activated protein.

Example 5

N-((4-(dibenzo[b,f][1,4]thiazepin-11-yl)-1-(2-(2-hydroxyethoxy)ethyl)piperazin-2-yl)methyl)-2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)acetamide-bovine thyroglobulin-conjugate

Step A

Bovine Thyroglobulin (BTG) reaction with SATA:

To 3.0 mL of a solution of bovine thyroglobulin (BTG, 20.0 mg, 0.03 μ moles) in 100 mM phosphate buffer pH 7.5 was added 276.0 μ L of a DMF solution of N-succinimidyl-S-acetylthioacetate (SATA, 25 mg/mL, 6.9 mg, 30.0 μ moles). The resulting solution was incubated at 20 °C for 1 hour on a roller mixer. The reaction was purified on a Sephadex G-25 column using 100 mM phosphate buffer, 5 mM EDTA, at pH 6.0. To 6.0 mL of BTG-SATA (18.0 mg, 0.027 μ moles) was added 600 μ L of 2.5 M hydroxylamine, 50 mM EDTA, pH 7.0. The resulting solution was incubated at 20 °C for 1 hour on a roller mixer.

Step B

To an aliquot of BTG-SH solution, prepared as described in the previous step, 6.6mL, 0.027 μ moles) was added an aliquot of the solution prepared in Example 2 (898.9 μ L, 19.0 μ moles). The resulting cloudy mixture was incubated for 3 hours at 20 °C on a roller mixer. The reaction was filtered through a 0.45 μ m syringe filter, then purified on a Sephadex G-25 column using 100 mM phosphate buffer, 0.14M sodium chloride, at pH 7.4.

Example 6

2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)-N-((11-(4-(2-(2-hydroxyethoxy)ethyl)piperazin-1-yl)dibenzo[b,f][1,4]thiazepin-3-yl)methyl)acetamide-bovine thyroglobulin-conjugate

To an aliquot of BTG-SH solution, prepared as described in Example 5 Step A, (3.4mL, 0.014 μ moles) was added 641.4 μ L of 2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)-N-((11-(4-(2-(2-hydroxyethoxy)ethyl)piperazin-1-yl)dibenzo[b,f][1,4]thiazepin-3-yl)methyl)acetamide, prepared as described in Example 4, (13.6 μ moles). The resulting cloudy mixture was incubated for 3 hours at 20 °C on a roller mixer. The reaction was purified on a Sephadex G-25 column using 100 mM phosphate buffer, 0.14M sodium chloride, at pH 7.4.

Example 7

N-((4-(dibenzo[b,f][1,4]thiazepin-11-yl)-1-(2-(2-hydroxyethoxy)ethyl)piperazin-2-yl)methyl)-2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)acetamide-keyhole limpet hemocyanin-conjugate

Step A

Keyhole Limpet Hemocyanin (KLH) reaction with SATA

To a 3.18 mL solution of keyhole limpet hemocyanin (KLH, 15.6 mg, 0.156 μ moles) in 100 mM phosphate buffer, 0.46M sodium chloride, at pH 7.4 was added 72.1 μ L of a DMF solution of N-succinimidyl-S-acetylthioacetate (SATA, 25 mg/mL, 1.8 mg, 7.80 μ moles). The resulting solution was incubated at 20 °C for 1 hour on a roller mixer. The reaction was purified on a Sephadex G-25 column using 100 mM phosphate buffer, 0.46 M sodium chloride, 5 mM EDTA, at pH 6.0. To 6.27 mL of the resulting KLH-SATA solution (13.3 mg, 0.133 μ moles) was added 627 μ L of 2.5M hydroxylamine, 50 mM EDTA, at pH 7.0. The resulting solution was incubated at 20 °C for 1 hour on a roller mixer. The reaction was used as such in conjugation reaction with maleimide-activated hapten.

Step B

To an aliquot of KLH-SH solution, prepared as described in the previous step, (6.9mL, 0.133 μ moles) was added an aliquot of the solution prepared in Example 2, (624.3 μ L, 13.3 μ moles). The resulting cloudy mixture was incubated for 3 hours at 20 °C on a roller mixer. The reaction was filtered through a 0.45 μ m syringe filter then purified on a Sephadex G-25 column using 100 mM phosphate buffer, 0.46M sodium chloride, at pH 7.4.

Example 8

2-(2,5-dioxo-2,5-dihydro-1H-pyrrol-1-yl)-N-((11-(4-(2-(2-hydroxyethoxy)ethyl)piperazin-1-yl)dibenzo[b,f][1,4]thiazepin-3-yl)methyl)acetamide-keyhole limpet hemocyanin-conjugate

To an aliquot of the KLH-SH solution, prepared as described in Example 7 Step A (3.2mL, 0.061 μ moles) was added an aliquot of the solution prepared in Example 4 (283.0 μ L, 6.10 μ moles). The resulting cloudy mixture was incubated for 3 hours at 20 °C on a roller mixer. The reaction was purified on a Sephadex G-25 column using 100 mM phosphate buffer, 0.46M sodium chloride, at pH 7.4.

Example 9

Competitive Immunoassay for Quetiapine

Following a series of immunizations with quetiapine immunogens, mouse tail bleeds were tested for reactivity using an ELISA. Hybridoma supernatants were also tested, and the ELISA data shown in Tables 1 and 2 below shows reactivity of several hybridomas (fusion partner was NSO cells).

Table 1

Dilution	9	10	11	12	
400					
400					
1200					
1200					
3600	79	89	90	95	Cmpd # 9
3600					
10800					
10800					
Bl Sub	1.5858	1.3168	1.4302	0.0533	
	1.5111	1.0627	1.2186	0.0427	
	0.5578	0.4213	0.598	0.0219	
	0.554	0.4447	0.5353	0.0233	
	0.1932	0.1582	0.1868	0.0154	
	0.171	0.2111	0.1838	0.0132	
	0.0736	0.0722	0.0733	0.0107	
	0.0884	0.0774	0.086	0.0107	

Table 2

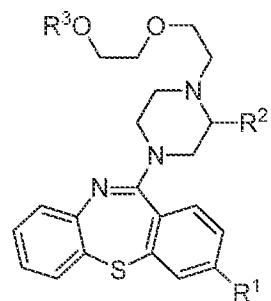
dilution	4C12	1A4	4G12	1F6
400	0.5467	0.2002	0.0144	0.1308
1200	0.1793	0.0619	0.01035	0.03905
3600	0.06655	0.026	0.00825	0.0192
10800	0.02755	0.0132	0.00765	0.01035
400	3.7296	0.24275	0.22585	0.00615
1200	2.4516	0.08695	0.0763	0.00685
3600	1.1575	0.0282	0.02875	0.00615
10800	0.4622	0.0147	0.0145	0.00645
dilution	5E9	2F2	3E2	

Supernatant was then tested by competition ELISA to determine if the signals were specific to quetiapine. Figs. 1 and 2 show the results from representative hybridomas. Data shows specific reactivity to quetiapine.

Fig. 3 shows the competitive immunoassay format used on a lateral flow assay device in which the capture antibody, a quetiapine clone, was deposited on a chip along with a detection conjugate consisting of quetiapine conjugated to a fluorophore. In this competitive format as shown in Fig. 3, a low level of analyte (quetiapine) results in high signal, whereas a high level of analyte (quetiapine) results in low signal. The amount of quetiapine in the sample can be calculated from the loss of fluorescence compared to a control sample with no drug present. A typical dose response curve generated with quetiapine sub-clones 89-3, 89-13, and 89-5 is shown in Fig. 4.

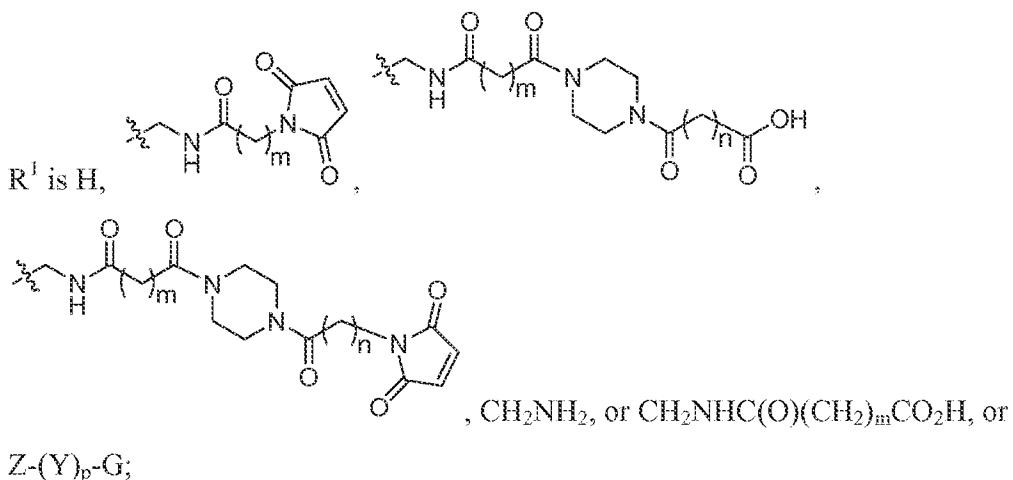
We claim:

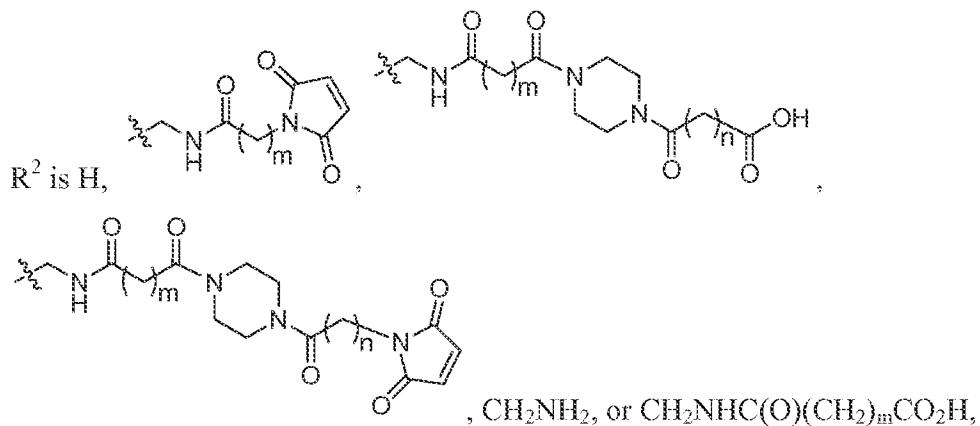
1. The compound of Formula I



Formula I

wherein:





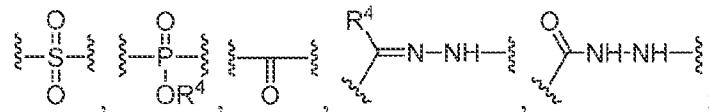
or, $Z-(Y)_p-G$;

R^3 is H, or $W-(Y)_p-G$; provided that two of R^1 , R^2 , R^3 must be H, and further provided that R^1 , R^2 and R^3 may not all be H simultaneously;

wherein:

Z is selected from the group consisting of:

$-\text{N}(\text{R}^4)-$, $-\text{O}-$, $-\text{S}-$, $-\text{alkyl}-$, $-\text{aminoalkyl}-$, $-\text{thioalkyl}-$, $-\text{heteroalkyl}-$, $-\text{alkylcarbonyl}-$,



R^4 is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

wherein:

W is selected from the group consisting of:

$-\text{C}(\text{O})-$, $-\text{alkyl}-$, $-\text{aminoalkyl}-$, $-\text{thioalkyl}-$, $-\text{heteroalkyl}-$, $-\text{alkylcarbonyl}-$;

Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

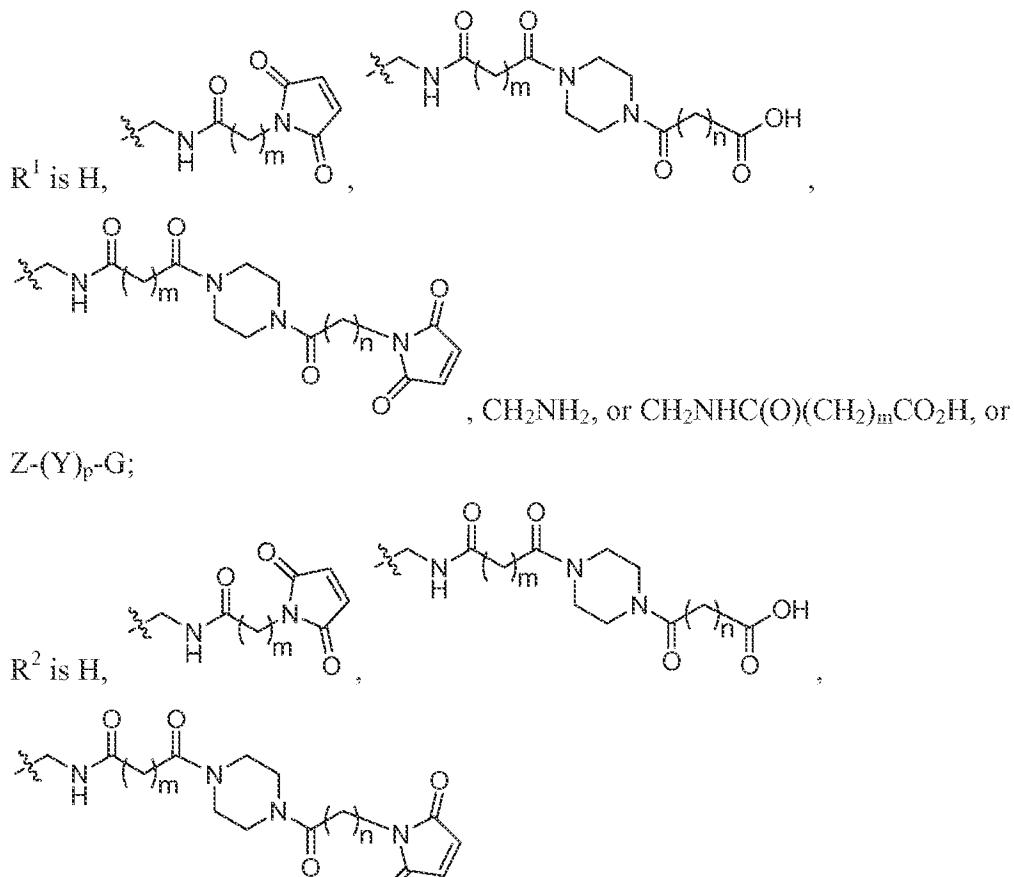
p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5.

2. The compound of Claim 1

wherein:



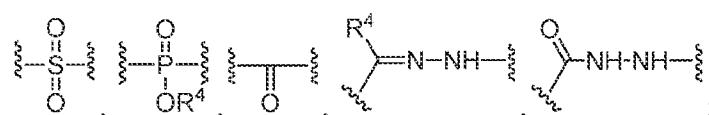
or, $Z-(Y)_p-G$;

R^3 is H, provided that either R^1 or R^2 must be H, and further provided that both R^1 and R^2 may not be H simultaneously;

wherein:

Z is selected from the group consisting of:

- $N(R^4)$ -, - O -, - S -, -alkyl-, -aminoalkyl-, -thioalkyl-, -heteroalkyl-, -alkylcarbonyl-,



R^4 is H, an alkyl group, cycloalkyl group, aralkyl group or substituted or unsubstituted aryl group;

Y is an organic spacer group;

G is a functional linking group capable of binding to a carrier;

p is 0, or 1;

m is 1, 2, 3, 4, or 5;

n is 1, 2, 3, 4, or 5.

3. The compound of Claim 2

wherein:

R¹ is H, or CH₂NH-(Y)_p-G;

R² is H, or CH₂NH-(Y)_p-G; provided that either R¹ or R² must be H, and further provided that both R¹ and R² may not be H simultaneously;

R³ is H;

wherein:

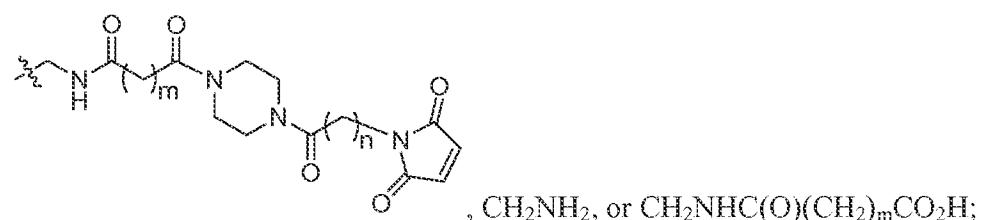
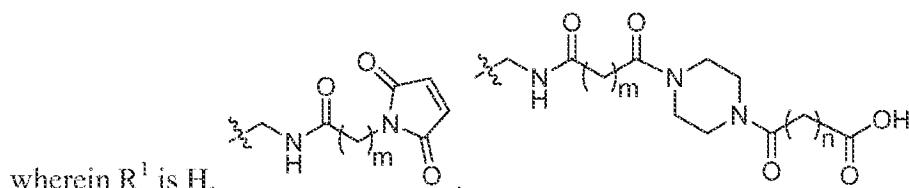
Y is an organic spacer group;

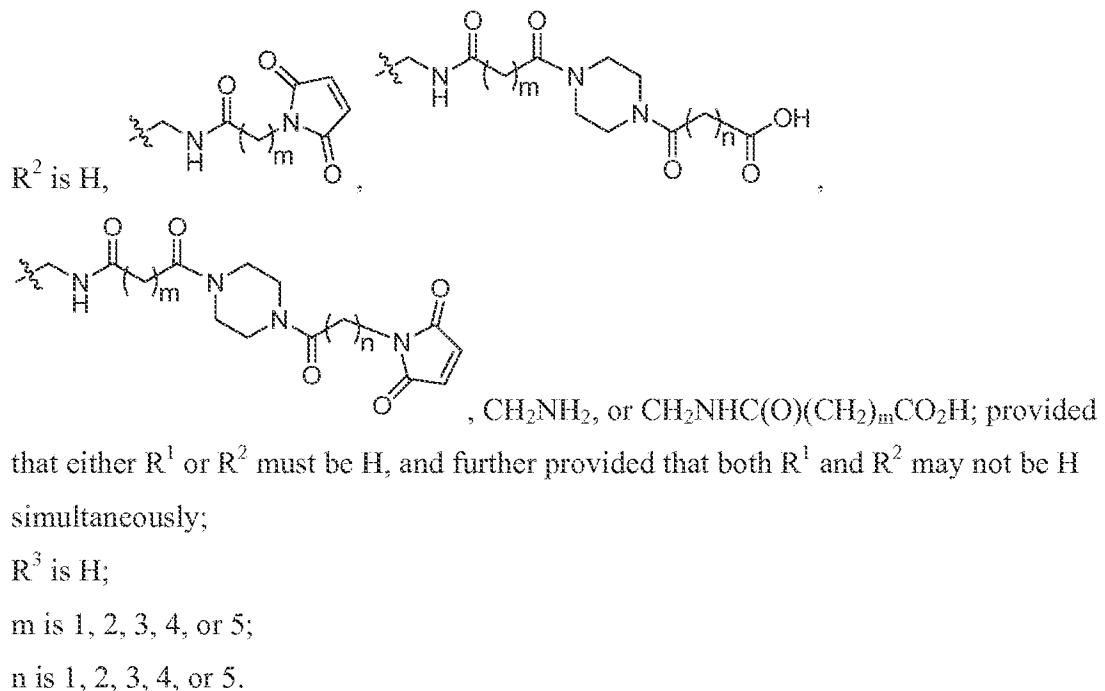
G is a functional linking group capable of binding to a carrier;

p is 1.

4. The compound of Claim 2

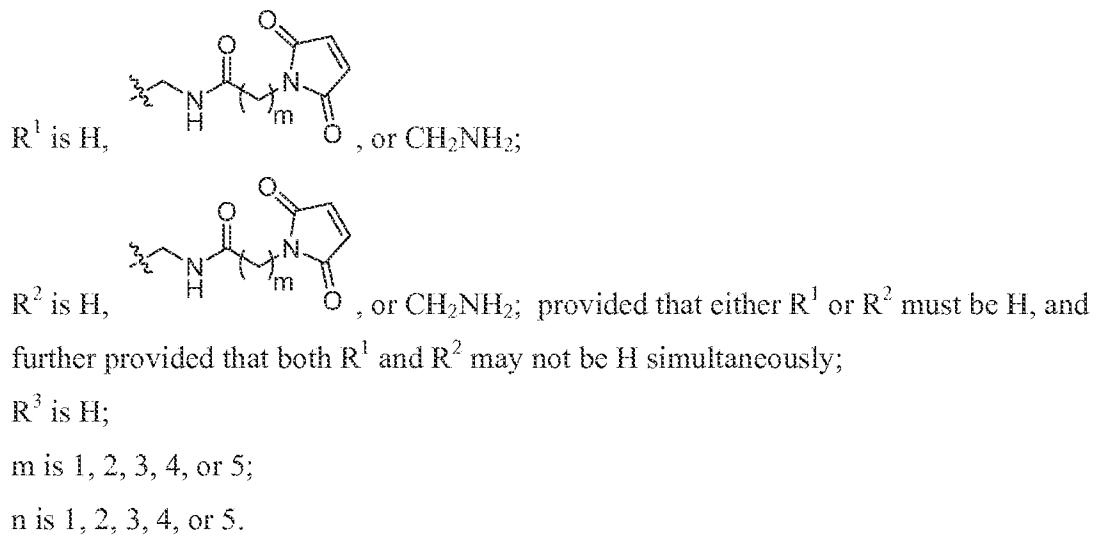
wherein:



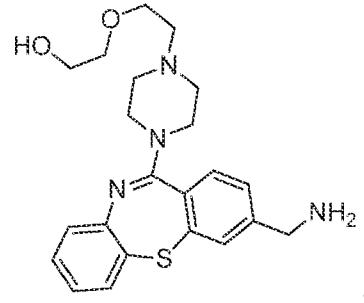
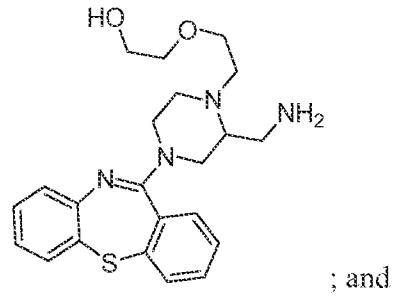
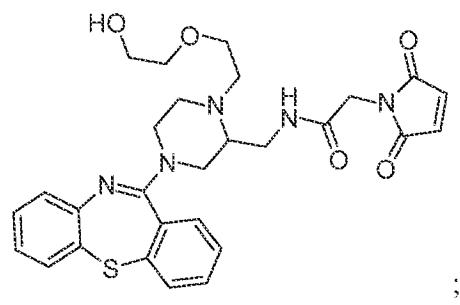
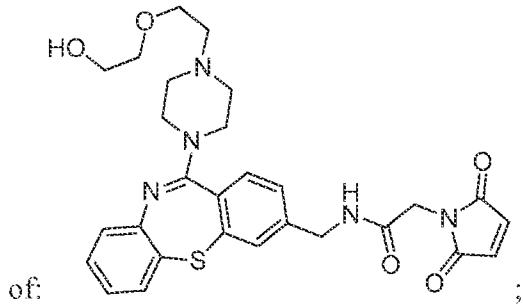


5. The compound of Claim 2

wherein:



6. The compound of Claim 1, selected from the group consisting



7. A conjugate of a compound of Claim 1 and an immunogenic carrier.

8. The conjugate of Claim 7, where said immunogenic carrier is a protein.

9. The conjugate of Claim 8, where said protein is keyhole limpet hemocyanin, ovalbumin or bovine thyroglobulin.

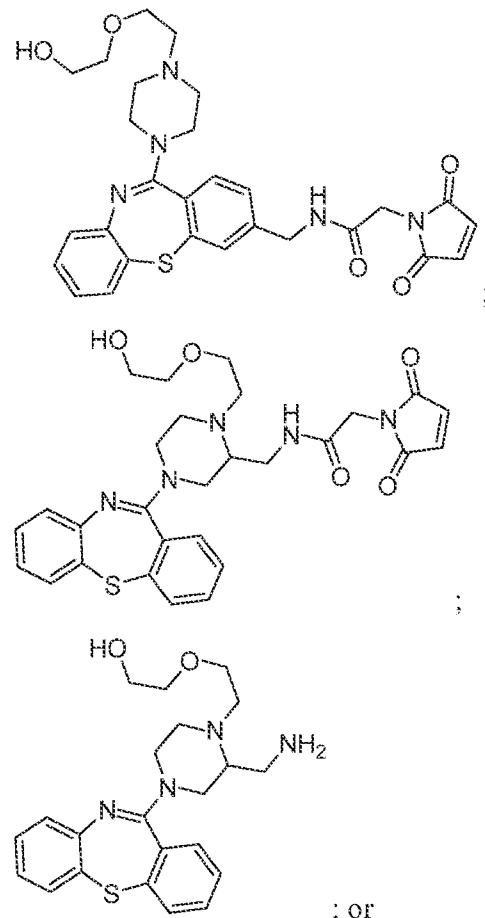
10. A conjugate of the compound of Claim 2 and an immunogenic carrier.

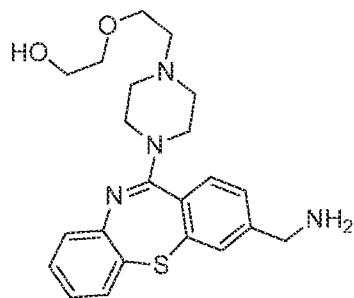
11. A conjugate of the compound of Claim 3 and an immunogenic carrier.

12. A conjugate of the compound of Claim 4 and an immunogenic carrier.

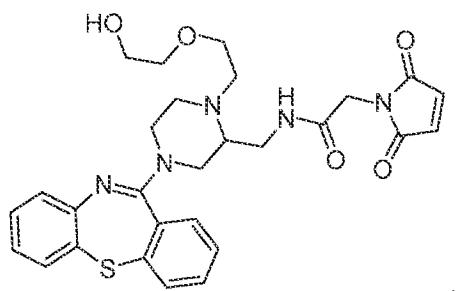
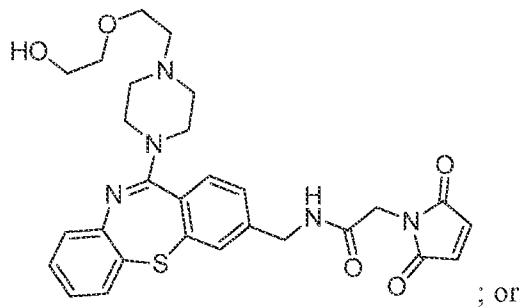
13. A conjugate of the compound of Claim 5 and an immunogenic carrier.

14. The conjugate of Claim 7 wherein the compound is





15. The conjugate of Claim 8 wherein the compound is:



16. The conjugate of Claim 15 wherein said protein is keyhole limpet hemocyanin, ovalbumin or bovine thyroglobulin.

17. A product made by the process of contacting a compound of Claim 1 with an immunogenic carrier.

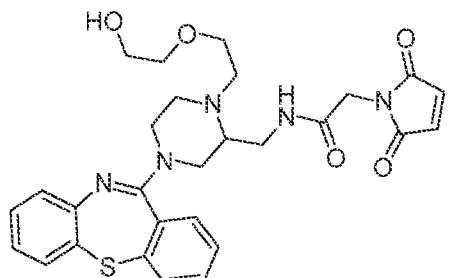
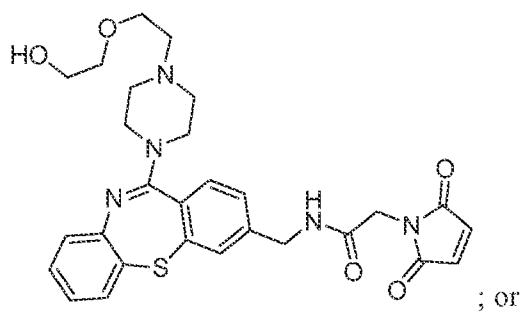
18. A product made by the process of contacting a compound of Claim 2 with an immunogenic carrier.

19. A product made by the process of contacting a compound of Claim 3 with an immunogenic carrier.

20. A product made by the process of contacting a compound of Claim 4 with an immunogenic carrier.

21. A product made by the process of contacting a compound of Claim 5 with an immunogenic carrier.

22. A product made by the process of contacting a compound which is



23. The product of Claim 22 wherein the immunogenic carrier is a protein

24. The product of Claim 23 wherein the protein is keyhole limpet hemocyanin, ovalbumin or bovine thyroglobulin.

FIGURE 1/4

Fig. 1

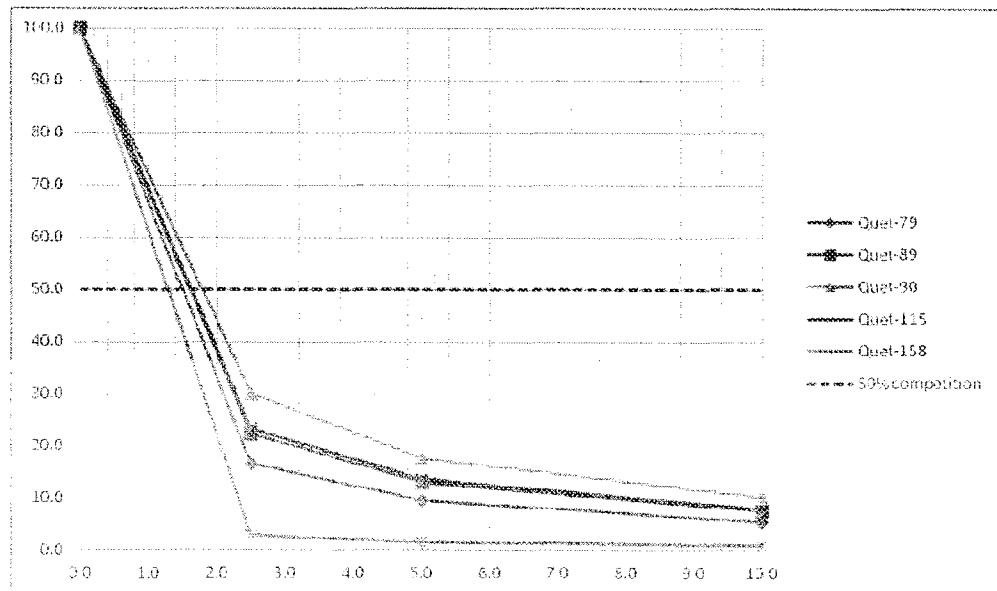


Fig. 2 FIGURE 2/4

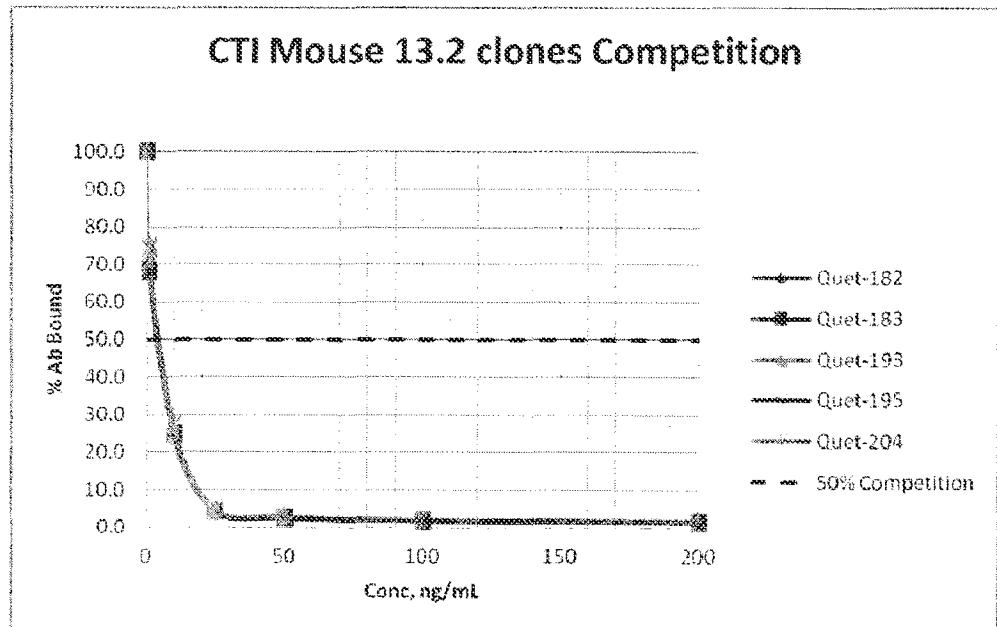


FIGURE 3/4

Fig. 3

Competitive Formats: Ab Down

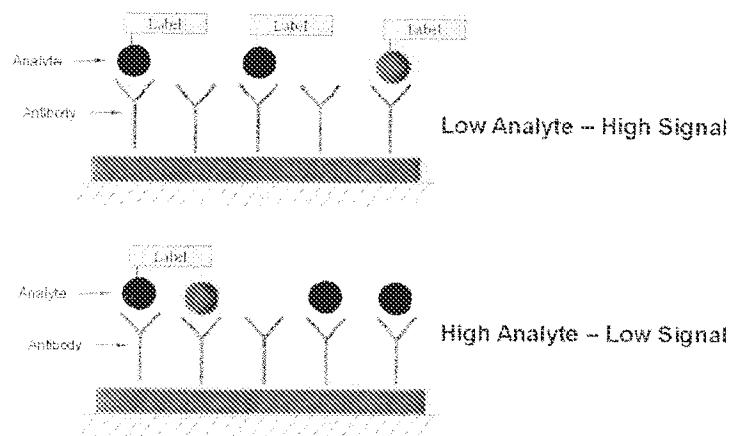
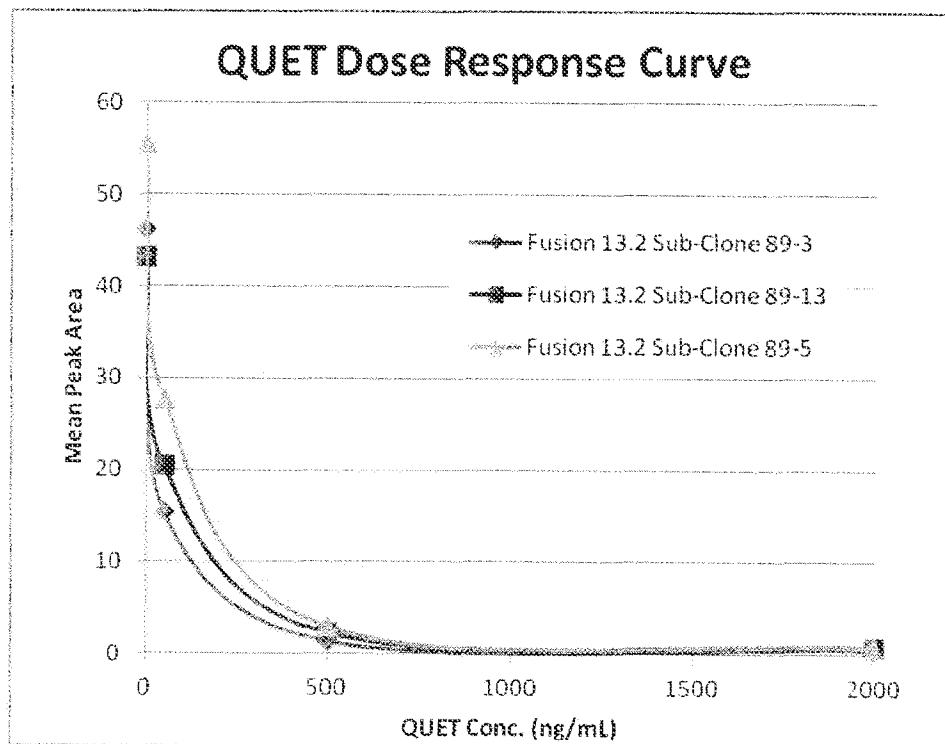


Fig. 4 FIGURE 4/4



INTERNATIONAL SEARCH REPORT

International application No

PCT/US2013/055724

A. CLASSIFICATION OF SUBJECT MATTER

INV. C07D417/12 C07D417/14 C07D281/16 G01N33/53 A61K47/48
A61K31/554 A61P25/00

ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

C07D G01N A61K A61P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

EPO-Internal, WPI Data, CHEM ABS Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2011/112657 A1 (KEMPHARM INC [US]; MICKLE TRAVIS [US]; GUENTHER SVEN [US]; BERA SANJIB) 15 September 2011 (2011-09-15) claims 1-170 examples (Tables I-VIII) -----	1
A	WO 2011/115733 A1 (SALADAX BIOMEDICAL INC [US]; SALAMONE SALVATORE J [US]; COURTNEY JODI) 22 September 2011 (2011-09-22) the whole document ----- -/-	2-24
A		1-24

Further documents are listed in the continuation of Box C.

See patent family annex.

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Date of the actual completion of the international search	Date of mailing of the international search report
17 September 2013	24/09/2013
Name and mailing address of the ISA/ European Patent Office, P.B. 5818 Patentlaan 2 NL-2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Authorized officer Marzi, Elena

INTERNATIONAL SEARCH REPORT

International application No
PCT/US2013/055724

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category ¹	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2011/082076 A1 (KEMPHARM INC [US]; MICKLE TRAVIS [US]; GUENTHER SVEN [US]; BERA SANJIB) 7 July 2011 (2011-07-07) claims 1-134 Compounds (I), (IV), (V), (VIII) figures 1-2, 3A, 3B paragraph [0048] - paragraph [0049] -----	1
A	Compounds (I), (IV), (V), (VIII) figures 1-2, 3A, 3B paragraph [0048] - paragraph [0049] -----	2-24
X	WO 2005/028458 A1 (FERMION OY [FI]; RUMMAKKO PETTERI [FI]; HUHTA SOINI [FI]; GRUMANN ARNE) 31 March 2005 (2005-03-31) Compounds of formula (III) , claim 1 -----	1
A	DAVIS P C ET AL: "Development and validation of an LC-MS/MS method for the determination of quetiapine and four related metabolites in human plasma", JOURNAL OF PHARMACEUTICAL AND BIOMEDICAL ANALYSIS, NEW YORK, NY, US, vol. 51, no. 5, 6 April 2010 (2010-04-06), pages 1113-1119, XP026822712, ISSN: 0731-7085 [retrieved on 2009-11-24] abstract page 1118, paragraph 4 -----	1-24
A	WO 2009/040409 A1 (NOVARTIS AG [CH]; ALBIENTZ PATRICK [FR]; GRENET JEAN-MICHE [FR]; HILLE) 2 April 2009 (2009-04-02) claims 1-20 -----	1-24

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Information on patent family members

International application No

PCT/US2013/055724

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