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(54) Title: NOVOBIOCIN ANALOGUES HAVING MODIFIED SUGAR MOIEITIES

(57) Abstract: The disclosure provides novobiocin analogues with noviose replacements which are useful as Hsp90 inhibitors in the treatment of cancer.
NOVOBIOCIN ANALOGUES HAVING MODIFIED SUGAR MOIETIES

This application is being filed on 19 February 2010, as a PCT International Patent application in the name of the University of Kansas, a U.S. University, applicant for the designation of all countries except the US, and Huiping Zhao, a citizen of P.R. China, and Alison Catherine Donnelly, a citizen of the U.S., applicants for the designation of the US only, and claims priority to U.S. Utility patent application Serial Nos. 12/390,01 1, filed on February 20, 2009, and 12/390,175, filed on February 20, 2009.

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application is a continuation-in-part of U.S. Patent Application Serial No. 12/390,01 1, filed on February 20, 2009, and is also a continuation-in-part of U.S. Patent Application Serial No. 12/390,175, filed on February 20, 2009, each of which is incorporated herein by reference in its entirety.

STATEMENT REGARDING FEDERALLY SPONSORED RESEARCH OR DEVELOPMENT

[0002] The present disclosure was supported in part by National Institutes of Health COBRE in Protein Structure and Function Grant Nos. NIH 31207, R01CA120458, U01CA39610 (BSJB), and Institute for the Study of Aging and NIH Grant No. AG 12933 (MLM), and the Department of Defense Prostate Cancer Research Program (QH815179) and the government may have certain rights in the present disclosure.

BACKGROUND OF THE DISCLOSURE

[0003] The Hsp90 molecular chaperone has emerged as a promising target for the treatment of cancer. While most current therapies are directed at disrupting a single molecular function, Hsp90 is distinctive in its regulation of multiple oncogenic pathways. There are more than 100 client proteins that depend upon Hsp90 for their folding and conformational maintenance, many of which contribute to cancer cell proliferation.

[0004] Novobiocin, a member of the coumermycin family of antibiotics, was isolated from *streptomyces* and shown to manifest potent activity against Gram-positive bacteria.
Novobiocin elicits antimicrobial activity through binding the ATP-binding pocket of DNA gyrase and prohibiting ATP-hydrolysis.

[0005] The present disclosure is directed to novel novobiocin analogues having a modified sugar.

**BRIEF SUMMARY OF THE DISCLOSURE**

[0006] One embodiment of the present disclosure is directed to novobiocin analogues lacking the noviose sugar. One advantage of the present disclosure is that the noviose sugar found in the natural product is very complex and can only be synthesized through a time-consuming and expensive synthesis. Even the most efficient syntheses of noviose require more than 10 steps, starting from relatively expensive starting materials, and the overall yield is less than optimal.

[0007] Another embodiment of the disclosure is directed to methods of using the compounds of the present disclosure which are useful as Hsp90 inhibitors, and in particular as anti-cancer agents, as well as neuroprotective agents, and/or in the treatment of autoimmune disorders. Thus, the present disclosure is directed to the therapeutic use of such compounds in the treatment and/or prevention of cancer, autoimmune, or neurodegenerative disorders in a subject in need thereof.

[0008] In one aspect, the cancer is selected from the group consisting of breast cancer, colon cancer, pancreatic cancer, or prostate cancer. In another aspect, the cancer is selected from the group consisting of breast cancer, ovarian cancer, colon cancer, head and neck cancer, lung cancer, gynecological cancers, brain cancer, germ cell cancer, urothelial cancer, esophageal cancer, prostate cancer, bladder cancer, or pancreatic cancer. A patient in need of cancer treatment is administered a therapeutically effective amount of the compounds of the present disclosure.

[0009] In one aspect, the neurodegenerative disorder is a beta amyloid disorder, and is most preferably Alzheimer's disease.

[0010] In another aspect, the autoimmune disorder is mediated by a heat shock response which increases expression of nitric oxide synthase, cytokines, and chemokines. Administration of the Hsp90 inhibitors of the present disclosure leads to a heat shock response due to the dissociation of HSF-I from Hsp90.
[0011] In another aspect, the neurodegenerative disorder is an autoimmune disorder, such as multiple sclerosis.

[0012] In still another aspect, the compounds of the present disclosure exhibit neuroprotective effects by upregulation of Hsp70.

[0013] In another aspect, the compounds are not substrates for the P-glycoprotein efflux pumps and are capable of crossing the blood-brain barrier.

[0014] It is contemplated that one or more compounds of the present disclosure can be useful for modulating or inhibiting heat-shock protein 90 activity by administering one or more of the compounds of the present disclosure to a cell or subject and observing a decrease in the expression of a heat-shock protein 90 client protein.

[0015] Further in another aspect, the present disclosure is also directed to compounds having biaryl and heterocyclic amide side chains at R¹ that exploit hydrogen-bonding interactions with the binding pocket that typically binds the prenylated benzamide of novobiocin. See Burlison et al., Novobiocin Anallogues That Manifest Anti-proliferative Activity against Several Cancer Cell Lines, J. Org. Chem., 73(6) 2130 - 2137 (2008) (2/23/08 e-published); and Donnelly et al., The Design, Synthesis, and Evaluation of Coumarin Ring Derivatives of the Novobiocin Scaffold that Exhibit Antiproliferative Activity, J. Org. Chem. 73, 8901-8920 (2008), (e-published 10/22/08), both of which are incorporated by reference. Such compounds are also described in co-pending patent application Serial No 12/390,011, filed on February 20, 2009 by the present inventor, which is incorporated by reference.

[0016] In one aspect, the present disclosure is directed to compounds having modified sugar or other non-noviose moieties (X₉). In another aspect, compounds of the disclosure have modified amide side chains (R¹).

[0017] In one embodiment, the disclosure provides compounds according to Formula 1:

\[
\text{Formula 1:} \quad \text{X} \quad \text{Y} \quad \text{R}^1
\]

wherein R¹ is -NHCOR", where R" is a C₁-C₄ alkyl, aryl or heterocyclic group, each optionally substituted with one or more hydroxy, nitro, amino, alkyl, alkenyl, aryl, alkoxy or halo groups; X₉ is -O-alkyl, -O-alkylamino, -O-cycloalkyl, -O-(CO)-alkyl, -O-
(CO)-cycloalkyl, -O-(CH₂)n-pyridinyl, -O-(CH₂)n-piperidinyl, -O-(CH₂)n-pyrrolino, or -O-(CH₂)n-pyrrolidinyl (where n is 0, 1, 2 or 3), each optionally substituted with one or more amino, amido, alkyl, alkoxy, halo, pyrrolidinyl, or hydroxyl groups; or X⁹ is -O-mono-hydroxylated furanose, -O-dihydroxylated furanose, -O-mono-hydroxylated pyranose, -O-dihydroxylated pyranose, -O-trihydroxylated pyranose, -O-mono-hydroxylatedoxepinose, -O-dihydroxylated oxepinose, -O-azasugar, -O-acyl, ester, amino, amido, carbamate, phosphate ester, tosylate or mesylate; or X⁹ is -OH provided that R¹ is not an isoprenyl substituted benzamide, i.e., where R" is of the structure:

![Structure](image)

where R is an optional substituent; particularly H, C₁-C₄ alkyl, Ci-C₄ alkylamino, -(CO)-Ci-C₄ alkyl, or piperidinyl, each optionally substituted with Ci-C₄ alkyl; X is hydrogen, nitrile, halo, amino, amido, Ci-C₄ alkyl or alkoxy; and Y is hydrogen, amido, ester, amino, Ci-C₄ alkyl or alkoxy; or a pharmaceutically acceptable salt thereof.

[0018] According to one aspect, X⁹ is -O-alkyl, -O-alkylamino, -O-cycloalkyl, -O-(CO)-alkyl, -O-(CO)-cycloalkyl, -O-(CH₂)n-pyridinyl, -O-(CH₂)n-piperidinyl, -O-(CH₂)n-pyrrolino, or -O-(CH₂)n-pyrrolidinyl (where n is 0, 1, 2 or 3), each optionally substituted with one or more amino, amido alkyl, halo, alkoxy, or hydroxyl groups.

[0019] In another aspect, X⁹ is

![Structures](image)

Any of the above groups may be modified through the addition of one or more methylene groups to the ether linkage (i.e., -O-(CH₂)n-).
In another aspect, $X_9$ is $-O-(CH_2)_n-R$, where $R$ is selected from the group consisting of:

where $R''$ is H, alkoxy, C$_1$-C$_4$ alkyl, halo, amino or amido; and $n$ is as defined above.

In another aspect, $X_9$ is $-OR$, where $R$ is selected from the group consisting of: H, -COCH$_3$, mesylate, tosylate, -CONH$_2$, -CONHCH$_3$, -CON(CH$_3$)$_2$, -PO(OCH$_3$)$_2$, -COCH$_3$.

In one particular aspect, $R^1$ is -NHCOCH$_3$.

In other aspects, the disclosure provides compounds of formula (I), where $R^1$ is -NHCOR", and $R''$ is an aryl group selected from:
wherein $R_{24}$ and $R_{25}$ are independently hydrogen, $C_1-C_4$ alkyl, hydroxy or alkoxy; and $R_{33}$ is H; or $C_1-C_4$ alkyl, $C_1-C_4$ alkylamino, -(CO)-$C_1-C_4$ alkyl, or piperidinyl, each optionally substituted with $C_1-C_4$ alkyl; or $R''$ is a heterocyclic group:

wherein $R_{31}$ is hydrogen, halo, $C_1-C_4$ alkyl, hydroxy or alkoxy; and $R_{32}$ is hydrogen or $C_1-C_4$ alkyl.

[0024] In one specific aspect, $R''$ is an aryl group:

[0025] In this specific aspect, the disclosure provides compounds selected from the group consisting of :
In other aspects, the disclosure provides compounds of formula (I), where $R^1$ is -NHCOR", and $R^2$ is an aryl group:

$$\begin{align*}
\text{KU-131,} \\
\text{KU-133,} \\
\text{KU-135, and} \\
\text{KU-139.}
\end{align*}$$

[0026] In further specific aspects, the disclosure provides the following specific compounds:

- 4-(8-Methyl-7-(1-methylpiperidin-4-yloxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29a, KU-397);
- 4-(8-Methyl-7-(1-methylpiperidin-3-yloxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29c, KU-417);
- 4-(7-(2-(Dimethylamino)ethoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29e, KU-421);
4-(7-(3-(Dimethylamino)propoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29f, KU-406);

4-(8-methyl-2-oxo-7-(piperidin-4-yloxy)-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (30b, KU-415);

4-(8-methyl-2-oxo-7-(piperidin-3-yloxy)-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (30d, KU-419);

4-(8-methyl-7-(2-(methylamino)ethoxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (30g, KU-423);

4-Hydroxy-N-(8-methyl-7-(1-methylpiperidin-4-yloxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31a, KU-398);

4-Hydroxy-N-(8-methyl-2-oxo-7-(piperidin-4-yloxy)-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31b, KU-416);

4-Hydroxy-N-(8-methyl-7-(1-methylpiperidin-3-yloxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31c, KU-418);

4-Hydroxy-N-(8-methyl-2-oxo-7-(piperidin-3-yloxy)-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31d, KU-420);

N-(7-(2-(Dimethylamino)ethoxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (31e, KU-422);

N-(7-(3-(Dimethylamino)propoxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (31f, KU-407);

4-Hydroxy-N-(8-methyl-7-(2-(methylamino)ethoxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31g, KU-424);

N-(7-((2R,3R,4R)-3,4-dihydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (16a, KU-425);

N-(7-((2S,3R,4R)-3,4-dihydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (16b, KU-426);

4-Hydroxy-N-(7-((2R,3R)-3-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (17a, KU-247);

4-Hydroxy-N-(7-((2S,3R)-3-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (17b, KU-428);

4-Hydroxy-N-(7-((2S,4R)-4-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (18a, KU-429);
4-Hydroxy-N-(7-((2R,4R)-4-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (18b, KU-430);

4-(7-((2S,3S,4S)-3,4-Dihydroxytetrahydrofuran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylicarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (19, KU-431);

4-(7-((2S,4R)-4-Hydroxytetrahydrofuran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylicarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (20a, KU-432); and


[0028] In additional specific aspects, the disclosure provides the following compounds:

[0029] In other aspects, the disclosure provides compounds of formula (I), where R<sup>1</sup> is -NHCOR", and R" a heterocyclic group:
wherein $R^{31}$ is hydrogen, halo, isoprenyl, or alkoxy; and $R^{32}$ is hydrogen or $C_1$-$C_4$ alkyl.

[0030] In certain aspects, the disclosure provides the following compounds:

wherein $X$ is hydrogen or $-OCH_3$; $Y$ is $-CH_3$ or $-OCH_3$; $R^{31}$ is $H$, $Cl$, $-CH_2CH_2N(CH_3)_2$, $-CH_2CH_2CH_N(CH_3)_2$, $OCH_3$, or $H$; $R^{32}$ is $H$ or $-CH_3$; and $R$ is selected from the group consisting of:
[0031] In additional specific aspects, the disclosure provides the following compounds:

[0032] In a further aspect, the disclosure provides compounds of the structure:

wherein X is hydrogen or -OCH₃, Y is -CH₃ or -OCH₃; and R is selected from the group consisting of: H, -COCH₃, mesylate, tosylate, -CONH₂, -CONHCH₃, -CON(CH₃)₂, -PO(OCH₃)₂, -COCH₃,
and
[0033] In another specific aspect, the disclosure provides the following compounds:

[0034] In a further aspect, the following compound is provided:

3',6-dimethoxy-N-(8-methyl-2-oxo-7-(pyridin-3-ylmethoxy)-2H-chromen-3-yl)-[1',1''-biphenyl]-3-carboxamide (IC_{50} = 3.8 µM, PC-3, 24 hr.).
[0035] In another embodiment, the disclosure provides a pharmaceutical composition for the treatment of cancer in a patient in need thereof, the composition comprising a therapeutically effective amount of a compound of formula (I),

![Chemical Structure](image)

(I)

and a pharmaceutically acceptable carrier; wherein the substituents are described above.

[0036] In a further embodiment, the disclosure provides a method of treating cancer in a patient in need thereof, the method comprising administering a therapeutically effective amount of a compound of formula (I),

![Chemical Structure](image)

(I),

where the substituents are as described above.

[0037] In yet another aspect, R is -NR'COR", wherein R is hydrogen and R" is aryl according to:

![Chemical Structures](image)

or

wherein R is alkoxy; R is hydrogen, hydroxy, alkoxy, or aryloxy; and R is hydrogen, alkoxy, aryloxy, or amino; and X is ether or amino.

[0038] In still another aspect, R is -NR'COR", and R" is a heterocycle selected from the group consisting of pyridine, benzofuran, indole, and oxazole. In a preferred aspect, R is hydrogen and R" is a an indole according to:
In yet another aspect, \( R_1 \) is \(-NR'COR"\), where \( R' \) is hydrogen and \( R" \) is aryl or heterocycle according to:

\[
\begin{align*}
\text{or more preferably}
\end{align*}
\]

wherein \( R^{29} \) is hydrogen, alkoxy, or amino; and \( R^{30} \) is hydrogen, alkoxy, or aryloxy.

In yet a further aspect, the present disclosure is directed to compounds wherein \( R_1 \) is

\(-NR'COR"\), where \( R' \) is hydrogen and \( R" \) is a heterocycle according to:

\[
\begin{align*}
\end{align*}
\]

wherein \( R^{27} \) is hydrogen, hydroxy, alkoxy, or aryloxy; and \( R^{28} \) is hydrogen, alkoxy, aryloxy, or amino.

In yet a further aspect, the present disclosure is directed to compounds wherein \( R_1 \) is

\(-NR'COR"\), where \( R' \) is hydrogen and \( R" \) is a heterocycle according to:

\[
\begin{align*}
\end{align*}
\]

wherein \( X_n \) is a covalent bond, alkyl, alkenyl, alkynyl, or \(-OCH_2\); and \( R^{26} \) is aryl, amino, or hydroxy. Thus, the present disclosure encompasses compounds wherein \( R_1 \) is

\(-NR'COR"\), where \( R' \) is hydrogen and \( R" \) is aryl selected from:

\[
\begin{align*}
\end{align*}
\]

In yet a further aspect, the present disclosure is directed to compounds wherein \( R_1 \) is

\(-NR'COR"\), where \( R' \) is hydrogen and \( R" \) is a heterocycle selected from:
Additional aspects of the disclosure, together with the advantages and novel features appurtenant thereto, will be set forth in part in the description which follows, and in part will become apparent to those skilled in the art upon examination of the following, or may be learned from the practice of the disclosure. The objects and advantages of the disclosure may be realized and attained by means of the instrumentalities and combinations particularly pointed out in the appended claims.

**BRIEF DESCRIPTION OF THE DRAWINGS**

FIG. 1 shows the relative ratios of phospho-AKT by Western blot analyses when the compounds of Example 1 were tested for their ability to inhibit Hsp90 in Skbr3 breast cancer cells. Total protein concentration of each lysate was determined and equal amounts of protein were run in each lane of the gels. For the graphs shown in FIG. 1, the O.D.'s (optical density) of the Western bands for phospho-AKT were measured, as were the O.D.'s for actin probed as controls on the same blots. To obtain the graphed values, all specific O.D.'s (for Hsp90 clients) were normalized to the respective actin O.D.

FIG. 2 is a western blot analysis of Skbr3 cells treated with novobiocin analogue denominated herein as KU-3/A2 (2'-carbamate) and KU-1/A4 (diol) for 24 hours. After incubation, the cells were harvested, lysed, and equal amounts of the protein lysates loaded into SDS wells. After electrophoresis, the gel was probed with Her-2 and actin (control) antibodies. The specific decrease in Her-2 levels is a result of Hsp90 inhibition that leads to Her-2 degradation.

FIG. 3 (top panel) is a western blot analysis of prostate cancer LNCaP cells treated with KU-1/A4. The bottom panel is a western blot analysis of prostate cancer LAPC-4 cells incubated with KU-1/A4. Actin was used as a control in both assays.

FIG. 3 shows that KU-1/A4 induces Hsp90 at low concentrations in LNCaP cells. Western blot analysis of the effects of KU-1/A4 on Hsp90 client proteins in prostate
cancer LNCaP cells. Cells were treated with varying concentrations of KU-1/A4 for 24 hours and probed for the androgen receptor ("AR"), protein kinase β ("AKT"), Hsp90, and actin.

[0047] FIG. 4 shows the dose dependent effects of KU-1/A4 on Aβ-induced cell death in primary neurons. The compound was added two hours before the Aβ and the viability was determined at 48 hours. The data represents standard error of the means ("S.E.M.") from about 1500 cells from 3 preparations. *, p<0.05 for control vs. Aβ only. **, p<0.01. Aβ only vs. Aβ + KU-1/A4.

[0048] FIG. 5 shows the dose dependency of KU-1/A4 protection against Aβ toxicity. Neuronal cells were treated with vehicle only (clear), KU-1/A4 (100 nM, gray), or KU-1/A4 + Aβ (10 µM, black). The indicated concentrations of KU-1/A4 were added 2 hours before Aβ. Cell viability was determined 48 hours later as described in the methods section. *p<0.05 and **p<0.01, for Aβ alone vs. KU-1/A4 + Aβ. Data represent mean survival ± SE for three separate experiments with about 1500 cells per treatment condition. Aβ (10 µM) alone was used as 0% survival and DMSO control was used as 100% protection.

[0049] FIG. 6 shows that KU-1/A4 upregulates Hsp70 in neuronal cells. In FIG. 6A, primary cortical neurons were incubated with GA or KU-1/A4 for 48 hours and probed for Hsp70 and actin (control). In FIG. 6B, the ratio of Hsp70 to actin was determined for each treatment as described. *p<0.05 compared to DMSO control. Each bar represents the average of four separate experiments.

[0050] FIG. 7 shows the anti-proliferative and toxic effects of KU-1/A4 and GA. MCF-7 (FIG. 7A) or SkBr3 (FIG. 7B) cells were incubated with KU-1/A4 (closed circles) or GA (open circles) at varying concentrations. Viable cells were quantitated using the MTS/PMS assay as described herein. Values represent the mean ± SE for one representative experiment performed in triplicate. Assays were replicated three times and the IC50 of GA correlated well with previously published values (MCF-7 = 133 ± 2 and SkBr3 = 18 ± 5 nM). In FIG. 7C, neuronal cells were treated with DMSO (open bar), KU-1/A4, or GA at the indicated concentrations, and cell viability was determined 24 hours later as described in the methods sections. The data represent the mean percentage ± SE of surviving neurons for three separate experiments. *p<0.05 for control vs GA, and **p<0.001 for control vs GA.
FIG. 8 shows the efflux and transport of KU-I/A4 across bovine microvessel endothelial cells ("BMECs"). In FIG. 8A, BMECs were grown to confluency and incubated with rhodamine 123 (5 µM) alone or KU-1/A4 (indicated concentrations) plus Rhodamine 123 as described herein. Taxol (10 µM) was used as a positive p-glycoprotein substrate. In FIG. 8B, BMECs were grown to confluency on polycarbonate membranes and KU-1/A4 (10 µM) was added to the donor chamber. Aliquots from the receiver chamber were taken at the noted time points and analyzed by RP-HPLC for KU-1/A4 permeation.

FIG. 9 shows the results of a pilot study in which it was observed that following intravenous administration of 0.5 mg/kg of KU-32 commencing on day 0 (initial immunization with proteolipid protein ("PLP") for induction of experimental autoimmune encephalomyelitis ("EAE") and on day 7 (second booster immunization with PLP), the onset of EAE disease symptoms was delayed by several days and the overall severity of symptoms was reduced.

FIG. 10 shows the change in body weight of mice in the EAE study and supports the observations of clinical scores, where disease onset is delayed and the magnitude of weight loss is less in the KU-32 treated mice compared to the positive control mice. Weight loss is a good indicator of EAE disease onset in mice.

FIG. 11 shows the results of a second study in which KU-32 was administered on days 1, 3, 5, and 7. The early treatment regimen showed no change in disease onset or disease severity. In the late treatment regimen, KU-32 was injected IV on day 6, 8, 10, 12, and 14. This treatment resulted in an earlier onset of symptoms; however, the duration of symptoms was notably less than in the positive control group, and the clinical scores returned to baseline compared to the control group and the early treatment group in which the clinical score remained around 0.5.

FIG. 12 is a graph showing the corresponding body weight data for the second study described in FIG. 11.

FIG. 13 is a western blot analysis of DHNI and DHN2 after 24 hours incubation in SKBr3 breast cancer cells. Lane 1) 1% DMSO (control); lane 2) 0.01 µM DHNI; lane 3) 0.1 µM DHNI; lane 4) 1.0 µM DHNI; lane 5) 5 µM DHNI; lane 6) 10 µM DHNI; lane 7) DMSO (control); lane 8) 0.01 µM DHN2; lane 9) 0.1 µM DHN2; lane 10) 1.0 µM DHN2; lane 11) 5 µM DHN2; lane 12) 10 µM DHN2.
FIG. 14 are graphs showing the percent of cytotoxicity of DHNL and DHN2 on SKBr3 cells at varying concentrations and time.

FIG. 15 shows the SAR for biaryl novobiocin derivatives and Hsp90.

FIG. 16 shows the SAR for compound 46 and novobiocin.

FIG. 17 is a western blot analysis of Hsp90 client protein degradation assays against MCF-7 breast cancer cells. The concentration of compound 46 (in µM) is denoted above each lane. Geldamycin and DMSO were used as positive and negative controls, respectively.

DETAILED DESCRIPTION OF PREFERRED EMBODIMENT

Molecular terms, when used in this application, have their common meaning unless otherwise specified. It should be noted that the alphabetical letters used in the formulas of the present disclosure should be interpreted as the functional groups, moieties, or substituents as defined herein. Unless otherwise defined, the symbols have their ordinary and customary meaning to those skilled in the art.

As used herein, an "azasugar" refers to a sugar in which the ring-oxygen is replaced with an amino-group. The "azasugars" are preferably 1,3 or 1,4 azasugars, and the amino group may be either a secondary or tertiary amino group. Preferred tertiary amino groups are substituted with an alkyl or acyl group. In addition, the azasugar ring may be saturated or unsaturated.

As used herein, the term "sugar" refers to a sugar group in its cyclic form, for example, those derived from furanose (5-membered ring), pyranose (6-membered ring), or oxepanose (7-membered ring). Exemplary sugars are set forth in Yu et al., Synthesis of mono- and dihydroxylated furanoses, pyranoses, and an oxepanose for the preparation of natural product analogue libraries, J. Org. Chem. 70(14):5599-605 (2005), Harris et al., Syntheses of D- and L-Mannose, Galactose, and Talose via Diastereoselective and Enantioselective Dihydroxylation Reactions, J. Org. Chem. 1999 64(9), 2982-2983 (1999); Ahmed et al., De novo enantioselective syntheses of galacto-sugars and deoxy sugars via the iterative dihydroxylation of dideoxynucleoate, Org. Lett. 2005 7(4), 745-748 (2005); Haukaas et al., Enantioselective synthesis of 2-deoxy- and 2,3-dideoxyhexoses, Org. Lett. 2002 4(10), 1771-1774 (2002), all of which are incorporated by reference. Exemplary of sugar groups include threo furanosyl (from threose, a four-carbon sugar); ribofuranosyl (from ribose, a five-carbon sugar); arafuranosyl (also often referred to as
arabinofuranosyl; from arabinose, a five-carbon sugar); xylofuranosyl (from xylose, a five-carbon sugar), and lyxofuranosyl (from lyxose, a five-carbon sugar). The sugar may be mono-hydroxylated or poly-hydroxylated (e.g., di-hydroxylated, tri-hydroxylated).

The term "carbamate" refers to -COONHR, wherein R as used in this definition hydrogen, alkyl, aryl, or heteroaromatic.

The term "phosphate ester" refers to -PO 3RR", wherein R' and R" are independently hydrogen, alkyl, aryl, heteroaromatic.

The term "alcohol" indicates an optionally substituted hydrocarbon group having one or more hydroxy substituents. Exemplary alcohols include alkanols include those containing from about one up to twelve carbon atoms, with alkanols having one to up to six carbon atoms being most preferred. Exemplary of preferred aliphatic alcohols are: methanol, ethanol, 1-propanol, 2-propanol, 1-propen-2-ol, 1-butanol, 2-butanol, 2-methyl-2-propanol, 1-pentanol, 2-pentanol, 3-pentanol, 3-methyl-1-butanol, 1,2-ethandiol (ethylene glycol), 1,2,3-propanetriol (glycerol), i-1,2,3,4-butantetrol (i-erythritol), and 2,2-dihyroxymethyl-1,3-propandiol (pentaerythritol). When used as a sugar mimic at the 9-position, the alcohol is preferably poly-hydroxylated.

The terms "acyl" or "Ac" refers to -COR wherein R used in this definition is hydrogen, alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl. Most preferably, R is hydrogen, alkyl, aryl, or aralkyl.

The term "amido" indicates either a C-amido group such as -CONR'R" or an N-amido group such as -NRCOR" wherein R and R" as used in this definition are independently hydrogen, alkyl, alkenyl, alkynyl, alkoxy, carbocyclic, heterocyclic, aryl, or aralkyl. A "sulfoamido" group includes the -NR'-SO 2-R". Most preferably, R and R" are hydrogen, alkyl, aryl, or aralkyl.

The term "amino" signifies a primary, secondary or tertiary amino group of the formula -NR'R" wherein R and R" as used in this definition are independently hydrogen, alkyl, alkenyl, alkynyl, aralkyl, carbocyclic, heterocyclic, aralkyl, or other amino (in the case of hydrazide) or R' and R" together with the nitrogen atom to which they are attached, form a ring having 4 to 8 atoms. Thus, the term "amino," as used herein, includes unsubstituted, monosubstituted (e.g., monoalkylamino or monoarylalino), and disubstituted (e.g., dialkylamino or aralkylamino) amino groups. Amino groups include —NH2, methylamino, ethylamino, dimethylamino, diethylamino, methyl-ethylamino, pyrrolidin-1-yl, or piperidino, morpholino, etc. Other exemplary "amino" groups forming a ring include pyrrolyl, imidazolyl, pyrazolyl, isothiazolyl,
isoxazolyl, pyridyl, pyrazinyl, pyrimidinyl, pyridazinyl, indolizinyl, isoindolyl, indolyl, indazolyl, purinyl, ... having aryl groups attached to alkyl groups having one to six carbon atoms. Examples of such groups include benzyl, isopropoxy, tert-butoxy, hydroxyl... preferred "alkyl" groups herein contain 1 to 12 carbon atoms. Most preferred are "lower alkyl" which refer to an alkyl group of one to six, more preferably one to four, carbon atoms. The alkyl group may be optionally substituted with an amino, alkyl, halo, or hydroxyl group.

[0071] The term "alkoxy" denotes oxy-containing groups substituted with an alkyl, or cycloalkyl group. Examples include, without limitation, methoxy, ethoxy, tert-butoxy, and cyclohexyloxy. Most preferred are "lower alkoxy" groups having one to six carbon atoms. Examples of such groups include methoxy, ethoxy, propoxy, butoxy, isopropoxy, and tert-butoxy groups.

[0072] The terms "alkenyl" and "alkynyl" refer to unsaturated aliphatic groups analogous in length and possible substitution to the alkyls described above, but that contain at least one double bond or triple bond respectively.

[0073] The term "aryl" means a carbocyclic aromatic system containing one, two, or three rings wherein such rings may be attached together in a pendant manner or may be fused. The term "fused" means that a second ring is present (i.e., attached or formed) by having two adjacent atoms in common (i.e., shared) with the first ring. The term "fused" is equivalent to the term "condensed." The term "aryl" embraces aromatic groups such as phenyl, naphthyl, tetrahydronaphthyl, indane, and biphenyl. The aryl group may optionally be substituted with an amino, alkyl, halo, alkenyl, alkoxy, hydroxyl, carbocyclic, heterocyclic, or another aryl group. A preferred aryl is a pendant aryl according to:

![Diagram of aryl group]

[0074] The term "aralkyl" embraces aryl-substituted alkyl moieties. Preferable aralkyl groups are "lower aralkyl" groups having aryl groups attached to alkyl groups having one to six carbon atoms. Examples of such groups include benzyl,
diphenylmethyl, triphenylmethyl, phenylethyl, and diphenylethyl. The terms benzyl and phenylmethyl are interchangeable.

[0075] The term "aryloxy" embraces aryl groups, as defined above, attached to an oxygen atom. The aryloxy groups may optionally be substituted with a halo, hydroxyl, or alkyl group. Examples of such groups include phenoxy, 4-chloro-3-ethylphenoxy, 4-chloro-3-methylphenoxy, 3-chloro-4-ethylphenoxy, 3,4-dichlorophenoxy, 4-methylphenoxy, 3-trifluoromethoxyphenoxy, 3-trifluoromethylphenoxy, 4-fluorophenoxy, 3,4-dimethylphenoxy, 5-bromo-2-fluorophenoxy, 4-bromo-3-fluorophenoxy, 4-fluoro-3-methylphenoxy, 5,6,7,8-tetrahydronaphthoxy, 3-isopropylphenoxy, 3-cyclopropylphenoxy, 3-ethylphenoxy, 4-tert-butylphenoxy, 3-pentafluoroethylphenoxy, and 3-(1,1,2,2-tetrafluoroethoxy)phenoxy.

[0076] The term "aralkoxy" embraces oxy-containing aralkyl groups attached through an oxygen atom to other groups. "Lower aralkoxy" groups are those phenyl groups attached to lower alkoxy group as described above. Examples of such groups include benzylphenoxy, 1-phenylethoxy, 3-trifluoromethoxybenzylphenoxy, 3-trifluoromethylbenzylphenoxy, 3,5-difluorobenzylphenoxy, 3-bromobenzylphenoxy, 4-propylbenzylphenoxy, 2-fluoro-3-trifluoromethylbenzylphenoxy, and 2-phenylethoxy.

[0077] The term "carboxyl" refers to -R'C(=O)OR", wherein R' and R" as used in this definition are independently hydrogen, alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl or R' can additionally be a covalent bond. "Carboxyl" includes both carboxylic acids, and carboxylic acid esters. The term "carboxylic acid" refers to a carboxyl group in which R" is hydrogen. Such acids include formic, acetic, propionic, butyric, valeric acid, 2-methyl propionic acid, oxirane-carboxylic acid, and cyclopropane carboxylic acid. The term "carboxylic acid ester" or "ester" refers to a carboxyl group in which R" is alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl.

[0078] The term "carbocyclic" refers to a group that contains one or more covalently closed ring structures, and that the atoms forming the backbone of the ring are all carbon atoms. The ring structure may be saturated or unsaturated. The term thus distinguishes carbocyclic from heterocyclic rings in which the ring backbone contains at least one non-carbon atom. The term carbocyclic encompasses cycloalkyl ring systems.

[0079] The terms "cycloalkane" or "cyclic alkane" or "cycloalkyl" refer to a carbocyclic group in which the ring is a cyclic aliphatic hydrocarbon, for example, a cyclic alkyl group preferably with 3 to 12 ring carbons. "Cycloalkyl" includes, by way
of example, cyclopropyl, cyclobutyl, cyclopentyl, cyclohexyl, cycloheptyl, or cyclooctyl, and the like. The cycloalkyl group may be optionally substituted with an amino, alkyl, halo, or hydroxy 1 group.

[0080] The term "ether" refers to the group -R'-O-R" as used in this definition are independently hydrogen, alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl, and R’ can additionally be a covalent bond attached to a carbon.

[0081] The terms "halo" or "halogen" refer to fluoro, chloro, bromo, or iodo, usually regarding halo substitution for a hydrogen atom in an organic compound.

[0082] The terms "heterocyclic" or "heterocycle" means an optionally substituted, saturated or unsaturated, aromatic or non-aromatic cyclic hydrocarbon group with 4 to about 12 carbon atoms, preferably about 5 to about 6, wherein 1 to about 4 carbon atoms are replaced by nitrogen, oxygen or sulfur. Exemplary heterocyclic which are aromatic include groups pyridinyl, furanyl, benzofuranyl, isobenzofuranyl, pyrrolyl, thienyl, 1,2,3-triazolyl, 1,2,4-triazolyl, indoly, imidazolyl, thiazolyl, thiazolidinyl, pyrimidinyl, oxazolyl, triazinyl, and tetrazolyl. Exemplary heterocycles include benzimidazole, dihydrothiophene, dioxin, dioxane, dioxolane, dithiane, dithiazine, dithiole, furan, indole, 3-H indazole, 3-H-indole, imidazole, indolizine, isoindole, isothiazole, isoazole, morpholine, oxazole, oxadiazole, oxathiazole, oxathiazolidine, oxazine, oxadiazine, piperazine, piperidine, purine, pyran, pyrazine, pyrazole, pyridine, pyrimidine, pyridazine, pyrrole, pyrrolidine, tetrahydrofuran, tetrahydrothiophene, thiadiazine, thiaizole, thiatriazole, thiazole, thiomorpholine, thiophene, thiopyran, triazinyl, and triazole. The heterocycle may be optionally substituted with an amino, alkyl, alkenyl, alkynyl, halo, hydroxyl, carbocyclic, thio, other heterocyclic, or aryl group. Exemplary heterocyclic groups include 1-pyrrolyl, 2-pyrrolyl, 3-pyrrolyl, 1-indolyl, 2-indolyl, 3-indolyl, 1-pyridyl, 2-pyridyl, 3-pyridyl, 4-pyridyl, 1-imidazolyl, 2-imidazolyl, 3-imidazolyl, 4-imidazolyl, 1-pyrazolyl, 2-pyrazolyl, 3-pyrazolyl, A-pyrazolyl, 5-pyrazolyl, 1-pyrazinyl, 2-pyrazinyl, 1-pyrimidinyl, 2-pyrimidinyl, A-pyrimidinyl, 5-pyrimidinyl, 1-pyridazinyl, 2-pyridazinyl, 3-pyridazinyl, 4-pyridazinyl, 1-indolizinyl, 2-indolizinyl, 3-indolizinyl, 4-indolizinyl, 5-indolizinyl, 6-indolizinyl, 7-indolizinyl, 8-indolizinyl, 1-isooindolyl, 2-isooindolyl, 3-isooindolyl, 4-isooindolyl and 5-isooindolyl.

[0083] The term "hydroxy" or "hydroxyl" refers to the substituent -OH.

[0084] The term "oxo" shall refer to the substituent =0.
[0085] The term "nitro" means -NO₂.
[0086] The term "sulfanyl" refers to -SR’ where R’ as used in this definition is hydrogen, alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl.
[0087] The term "sulfonyl" refers to -SOR where R as used in this definition is hydrogen, alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl.
[0088] The term "sulfinyl" refers to -SOR’ where R’ as used in this definition is hydrogen, alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl.
[0089] "Optional" or "optionally" means that the subsequently described event or circumstance may or may not occur, and that the description includes instances where said event or circumstance occurs and instances in which it does not. "Optionally" is inclusive of embodiments in which the described conditions is present and embodiments in which the described condition is not present. For example, "optionally substituted phenyl" means that the phenyl may or may not be substituted, and that the description includes both unsubstituted phenyl and phenyl wherein there is substitution. "Optionally" is inclusive of embodiments in which the described conditions is present and embodiments in which the described condition is not present.
[0090] The compounds of the present disclosure can exist in tautomeric, geometric, or stereoisomeric forms. The present disclosure contemplates all such compounds, including cis- and trans-geometric isomers, E- and Z-geometric isomers, R- and S-enantiomers, diastereomers, d-isomers, l-isomers, the racemic mixtures thereof and other mixtures thereof, as falling within the scope of the disclosure.
[0091] Also included in the family of compounds of the present disclosure are the pharmaceutically acceptable salts, esters, and prodrugs thereof. The term "pharmaceutically-acceptable salts" embraces salts commonly used to form alkali metal salts and to form addition salts of free acids or free bases. The nature of the salt is not critical, provided that it is pharmaceutically acceptable. Suitable pharmaceutically acceptable acid addition salts of compounds of the present disclosure may be prepared from inorganic acid or from an organic acid. Examples of such inorganic acids are hydrochloric, hydrobromic, hydroiodic, nitric, carbonic, sulfuric, and phosphoric acid. Appropriate organic acids may be selected from aliphatic, cycloaliphatic, aromatic, araliphatic, heterocyclic, carboxylic and sulfonic classes of organic acids, examples of which are formic, acetic, propionic, succinic, glycolic, gluconic, lactic, malic, tartaric, citric, ascorbic, glucuronic, maleic, fumaric, pyruvic, aspartic, glutamic, benzoic, anthranilic, mesylic, salicylic, p-hydroxybenzoic, phenylacetic, mandelic, embonic
(pamoic), methanesulfonic, ethylsulfonic, benzenesulfonic, sulfanilic, stearic, cyclohexylaminosulfonic, algenic, and galacturonic acid. Suitable pharmaceutically-acceptable base addition salts of compounds of the present disclosure include metallic salts made from aluminum, calcium, lithium, magnesium, potassium, sodium and zinc or organic salts made from N,N'-dibenzylethlenendiamine, choline, chloroprocaine, diethanolamine, ethylenediamine, meglumine (N-methyl glucamine) and procain. All of these salts may be prepared by conventional means from the corresponding compounds of by reacting, for example, the appropriate acid or base with the compounds of the present disclosure.

As used herein, the term "pharmaceutically acceptable ester" refers to esters which hydrolyze in vivo and include, but are not limited to, those that break down readily in the human body to leave the parent compound or a salt thereof. Suitable ester groups include, for example, those derived from pharmaceutically acceptable aliphatic carboxylic acids, particularly alkanoic, alkenoic, cycloalkanoic and alkanedioic acids, in which each alkyl or alkenyl moiety advantageously has not more than 6 carbon atoms. Examples of particular esters include formates, acetates, propionates, butyrates, acrylates, and ethylsuccinates.

The term "pharmaceutically acceptable prodrugs" as used herein refers to those prodrugs of the compounds of the present disclosure which are, within the scope of sound medical judgment, suitable for use in contact with the tissues of humans and lower animals without undue toxicity, irritation, allergic response and the like, commensurate with a reasonable risk/benefit ratio, and effective for their intended use, where possible, of the compounds of the disclosure. The term "prodrug" refers to compounds that are rapidly transformed in vivo to yield the parent compound of the above formulae, for example, by hydrolysis in blood. A thorough discussion is provided in T. Higuchi and V. Stella, Prodrugs as Novel Delivery Systems, Vol. 14 of the A.C.S. Symposium Series and in Edward B. Roche, ed., Bioreversible Carriers in Drug Design, American Pharmaceutical Association and Pergamon Press, (1987), both of which are incorporated by reference herein.

The phrase "pharmaceutically acceptable" is employed herein to refer to those compounds, materials, compositions, and/or dosage forms which are, within the scope of sound medical judgment, suitable for use in contact with the tissues of human beings and animals without excessive toxicity, irritation, allergic response, or other problem or complication, commensurate with a reasonable benefit/risk ratio.
The phrase "pharmaceutically-acceptable carrier" as used herein means a pharmaceutically-acceptable material, composition or vehicle, such as a liquid or solid filler, diluent, excipient, solvent or encapsulating material, involved in carrying or transporting the subject novobiocin analogue or derivative from one organ, or portion of the body, to another organ, or portion of the body. Each carrier must be "acceptable" in the sense of being compatible with the other ingredients of the formulation and not injurious to the patient. Some examples of materials which may serve as pharmaceutically-acceptable carriers include: (1) sugars, such as lactose, glucose and sucrose; (2) starches, such as corn starch and potato starch; (3) cellulose, and its derivatives, such as sodium carboxymethyl cellulose, ethyl cellulose and cellulose acetate; (4) powdered tragacanth; (5) malt; (6) gelatin; (7) talc; (8) excipients, such as cocoa butter and suppository waxes; (9) oils, such as peanut oil, cottonseed oil, safflower oil, sesame oil, olive oil, corn oil and soybean oil; (10) glycols, such as propylene glycol; (11) polyols, such as glycerin, sorbitol, mannitol and polyethylene glycol; (12) esters, such as ethyl oleate and ethyl laurate; (13) agar; (14) buffering agents, such as magnesium hydroxide and aluminum hydroxide; (15) alginic acid; (16) pyrogen-free water; (17) isotonic saline; (18) Ringer's solution; (19) ethyl alcohol; (20) phosphate buffer solutions; and (21) other non-toxic compatible substances employed in pharmaceutical formulations.

The "patient" or "subject" to be treated with the compounds of the present disclosure can be any animal, e.g., dogs, cats, mice, monkeys, rats, rabbits, horses, cows, guinea pigs, sheep, and is preferably a mammal, such as a domesticated animal or a livestock animal. In another aspect, the patient is a human.

The term "autoimmune disorder" is intended to include disorders in which the immune system of a subject reacts to autoantigens, such that significant tissue or cell destruction occurs in the subject. The term "autoantigen" is intended to include any antigen of a subject that is recognized by the immune system of the subject. Autoimmune disorders include but are not limited to acute disseminated encephalomyelitis, Addison's disease, Alopecia universalis, ankylosing spondylitis, antiphospholipid antibody syndrome, aplastic anemia, autoimmune hepatitis autoimmune infertility, autoimmune thyroiditis, autoimmune neutropenia, Behçet's disease, bullous pemphigoid, Chagas' disease, cirrhosis, Coeliac disease, Crohn's disease, Chronic fatigue syndrome, chronic active hepatitis, dense deposit disease, discoid lupus, dermatitis, luten-sensitive enteropathy, dysautonomia, endometriosis, glomerulonephritis,
Goodpasture's disease, Graves' disease, Guillain-Barre syndrome, Hashimoto's disease, Hidradenitis suppurativa, idiopathic thrombocytopenia purpura, insulin dependent diabetes mellitus, interstitial cystitis, mixed connective tissue disease, multiple sclerosis, myasthenia gravis, neuromyotonia, opsoconus myoclonus syndrome, optic neuritis, Ord's thyroiditis, pemphigus vulgaris, pernicious anemia, polyarthritis, polymyositis, primary biliary cirrhosis, psoriasis, Reiter's syndrome, rheumatoid arthritis, sarcoidosis, scleroderma, Sjogren's syndrome, systemic lupus erythematosus, Takayasu's arteritis, temporal arteritis, ulcerative colitis, vitiligo, vulvodynia, warm autoimmune hemolytic anemia, or Wegener's granulomatosis. In a preferred aspect, the autoimmune disorder is multiple sclerosis or its animal model system termed experimental autoimmune encephalomyelitis ("EAE").

[0105] The term "neuroprotection" embraces to inhibition of progressive deterioration of neurons that leads to cell death.

[0106] The term "neurodegenerative disorder" embraces a disorder in which progressive loss of neurons occurs either in the peripheral nervous system or in the central nervous system. Examples of neurodegenerative disorders include, but are not limited to chronic neurodegenerative diseases such as diabetic peripheral neuropathy, Alzheimer's disease, Pick's disease, diffuse Lewy body disease, progressive supranuclear palsy (Steel-Richardson syndrome), multisystem degeneration (Shy-Drager syndrome), motor neuron diseases including amyotrophic lateral sclerosis ("ALS"), degenerative ataxias, cortical basal degeneration, ALS-Parkinson's-Dementia complex of Guam, subacute sclerosing panencephalitis, Huntington's disease, Parkinson's disease, multiple sclerosis, synucleinopathies, primary progressive aphasia, striatonigral degeneration, Machado-Joseph disease/spinocerebellar ataxia type 3 and olivopontocerebellar degenerations, Gilles De La Tourette's disease, bulbar and pseudobulbar palsy, spinal and spinobulbar muscular atrophy (Kennedy's disease), primary lateral sclerosis, familial spastic paraplegia, Wernicke-Korsakoff related dementia (alcohol induced dementia), Wolnym-Hoffmann disease, Kugelberg-Welander disease, Tay-Sach's disease, Sandhoff disease, familial spastic disease, Wohifart-Kugelberg-Welander disease, spastic paraparesis, progressive multifocal leukoencephalopathy, and prion diseases (including Creutzfeldt-Jakob, Gerstmann-Straussler-Scheinker disease, Kuru and fatal familial insomnia). Other conditions also included within the methods of the present disclosure include age-related dementia and other dementias, and conditions with memory loss including vascular dementia, diffuse white matter disease (Binswanger's disease),
dementia of endocrine or metabolic origin, dementia of head trauma and diffuse brain
damage, dementia pugilistica, and frontal lobe dementia. Also other neurodegenerative
disorders resulting from cerebral ischemia or infarction including embolic occlusion and
thrombotic occlusion as well as intracranial hemorrhage of any type (including, but not
limited to, epidural, subdural, subarachnoid, and intracerebral), and intracranial and
intravertebral lesions (including, but not limited to, contusion, penetration, shear,
compression, and laceration). Thus, the term also encompasses acute neurodegenerative
disorders such as those involving stroke, traumatic brain injury, schizophrenia, peripheral
nerve damage, hypoglycemia, spinal cord injury, epilepsy, and anoxia and hypoxia.

[0107] In one aspect, the neurodegenerative disorder is selected from Alzheimer's
disease, Parkinson's disease, amyotrophic lateral sclerosis, age-related memory loss,
senility and age-related dementia, most preferably, the neurodegenerative disorder is
Alzheimer's disease. Because, most preferably, the neurodegenerative disorder is
Alzheimer's disease, also characterized as an amyloidosis, other conditions within the
methods of the present disclosure include the treatment or prevention of other
amyloidosis disorders which share features including, but not limited to, hereditary
cerebral angiopathy, nonneuropathic hereditary amyloid, Down's syndrome,
macroglobulinemia, secondary familial Mediterranean fever, Muckle-Wells syndrome,
multiple myeloma, pancreatic- and cardiac-related amyloidosis, chronic hemodialysis
arthropathy, and Finnish and Iowa amyloidosis.

[0108] The term "inhibit" or "inhibiting" refers to a statistically significant and
measurable reduction in neurotoxicity, preferably as measured by one or more of the
assays discussed herein, preferably a reduction of at least about 10% versus control,
more preferably a reduction of about 50% or more, still more preferably a reduction of
about 60%, 70%, 80%, 90%, or more.

[0109] The term "preventing" as used herein means that the compounds of the
present disclosure are useful when administered to a patient who has not been diagnosed
as possibly having the disorder or disease at the time of administration, but who would
normally be expected to develop the disorder or disease or be at increased risk for the
disorder or disease. The compounds of the disclosure can slow the development of the
disorder or disease symptoms, delay the onset of the disorder or disease, or prevent the
individual from developing the disorder or disease at all. Preventing also includes
administration of the compounds of the disclosure to those individuals thought to be
predisposed to the disorder or disease due to age, familial history, genetic or
chromosomal abnormalities, and/or due to the presence of one or more biological markers for the disorder or disease.

[0110] The term "treating" as used herein generally means that the compounds of the disclosure can be used in humans or animals with at least a tentative diagnosis of the disorder or disease. The compounds of the disclosure can delay or slow the progression of the disorder or disease thereby giving the individual a more useful life span. The term "treatment" embraces at least an amelioration of the symptoms associated with the disorder or disease in the patient is achieved, where amelioration is used in a broad sense to refer to at least a reduction in the magnitude of a parameter, e.g., symptom, associated with the condition being treated. As such, "treatment" also includes situations where the diseased condition or disorder, or at least symptoms associated therewith, are completely inhibited, e.g., prevented from happening, or stopped, e.g., terminated, such that the patient no longer suffers from the condition or disorder, or at least the symptoms that characterize the condition or disorder.

[0111] A "therapeutically effective amount" is an amount of a compound of the present disclosure or a combination of two or more such compounds, which inhibits, totally or partially, the progression of the condition or alleviates, at least partially, one or more symptoms of the condition. A therapeutically effective amount can also be an amount that is prophylactically effective. The amount that is therapeutically effective will depend upon the patient's size and gender, the condition to be treated, the severity of the condition and the result sought. For a given patient and condition, a therapeutically effective amount can be determined by methods known to those of skill in the art. For example, in reference to the treatment of cancer using the compounds of the present disclosure, a therapeutically effective amount refers to that amount which has the effect of (1) reducing the size of the tumor, (2) inhibiting (that is, slowing to some extent, preferably stopping) tumor metastasis, (3) inhibiting to some extent (that is, slowing to some extent, preferably stopping) tumor growth, and/or, (4) relieving to some extent (or, preferably, eliminating) one or more symptoms associated with the cancer.

[0112] It is contemplated that several of the compounds of the present disclosure have been shown to modulate or inhibit Hsp90 \textit{in vitro}. As such, it is contemplated that therapeutically effective amounts of the compounds of the present disclosure can be useful as anti-cancer agents and/or neuroprotective agents.
In the context of cancer and neuroprotection, it is contemplated that some of the compounds of the present disclosure may be used with other Hsp90 inhibitors, chemotherapeutic agents, and/or neuroprotective agents.

The present disclosure is directed to the use of therapeutically effective amount of one or more of the compounds disclosed herein to treat and/or prevent a neurodegenerative disorder and/or to provide neuroprotection. Similarly, it is anticipated that the compounds of the present disclosure may be useful in the treatment of one or more types of cancer such as breast cancer, ovarian cancer, colon cancer, head and neck cancer, lung cancer, gynecological cancers, brain cancer, germ cell cancer, urothelial cancer, esophageal cancer, prostate cancer, bladder cancer, or pancreatic cancer.

Compositions of the Present Disclosure

According to another aspect, the present disclosure provides a pharmaceutical composition, which comprises a therapeutically-effective amount of one or more compounds of the present disclosure or a pharmaceutically-acceptable salt, ester or prodrug thereof, together with a pharmaceutically-acceptable diluent or carrier. The pharmaceutical compositions can be used to provide neuroprotection and used to treat and/or prevent cancer. The pharmaceutical compositions also can be used to treat neurodegenerative disorders.

The compositions may be formulated for any route of administration, in particular for oral, rectal, transdermal, subcutaneous, intravenous, intramuscular, intranasal, intraperitoneal, administration. The compositions may be formulated in any conventional form, for example, as tablets, capsules, caplets, solutions, suspensions, dispersions, syrups, sprays, gels, suppositories, patches, and emulsions.

Accordingly, the compounds of the present disclosure are useful in the treatment or alleviation of neurodegenerative disorders, such as Alzheimer's disease, Parkinson's disease, Lou Gehrig's disease, or multiple sclerosis, to name a few, not to mention central or peripheral nervous system damage, dysfunction, or complications involving same stemming from edema, injury, or trauma. Such damage, dysfunction, or complications may be characterized by an apparent neurological, neurodegenerative, physiological, psychological, or behavioral aberrations, the symptoms of which can be reduced by the administration of a therapeutically effective amount of the compounds of the present disclosure.

The following examples are provided to illustrate the present disclosure and are not intended to limit the scope thereof. Those skilled in the art will readily
understand that known variations of the conditions and processes of the following preparative procedures can be used to prepare these compounds.

EXAMPLE 1: SYNTHESIS OF NOVOBIOCIN ANALOGUES

[0120] In an effort to increase the affinity of novobiocin for the C-terminal ATP binding site, a library of novobiocin analogue compounds that contained both modified coumarin and sugar derivatives was prepared. The compounds were prepared as set forth in the scheme below along with a procedure recently developed for the synthesis of noviose. See Yu et al., Synthesis of (-)-Noviose from 2,3-O-Isopropylidene-D-erythronolactol, J. Org. Chem. 69, 7375-7378 (2004), which is incorporated by reference.

[0121] The novobiocin analogues prepared according to the scheme included modification of the coumarin ring by shortening of the amide side chain and removal of the 4-hydroxy substituent (A) (see Madhavan et al., Novel Coumarin Derivatives of Heterocyclic Compounds as Lipid Lowering Agents, Bioorg. Med. Chem. Lett. 13, 2547 (2003), which is incorporated by reference), removal of both the 4-hydroxy and amide linker (B), steric replacements of both the 4-hydroxy and benzamide ring (C), and 1,2-positional isomers of the noviosyl linkage (D and E).

[0122] These selected coumarin rings were coupled with trichloroacetimidate of noviose carbonate in the presence of boron trifluoride etherate as shown in scheme below. See Shen et al., Syntheses of Photolabile Novobiocin Analogues, Bioorg. Med. Chem. Lett. 14, 5903 (2004). The resulting cyclic carbonates (A1-E1) were treated with methanolic ammonia to provide 2’-carbamoyl (A2-E2), 3’-carbamoyl (A3-E3), and descarbamoyl products (A4-E4) in good yields. See also Yu et al., Hsp90 Inhibitors Identified from a Library of Novobiocin Analogues, J. Am. Chem. Soc. 127, 12778-12779 (2005), which is incorporated by reference.
wherein $R^1$ in the above scheme is hydrogen, amido, amino, or aryl; and
wherein $R^2$ in the above scheme is hydrogen, alkyl, or hydroxy.
Overall, the following twenty-three (23) analogues of novobiocin were prepared, which are set forth below:

N-(7-((3aR,4R,7R,7aR)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3aH-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-2-oxo-2H-chromen-3-yl)acetamide (Al). Noviose carbonate trichloroacetimidate (180 mg, 0.50 mmol) and 7-hydroxy-3-acetamino-coumarin A (133 mg, 0.60 mmol) were dissolved in CH₂Cl₂ (7 mL) before boron trifluoride etherate (30 µl, 0.03 mmol) was added to the suspension at 25 °C. The mixture was stirred at 25 °C for eight hours and quenched with Et₃N (0.4 mL, 2.8 mmol). The solvent was removed and the residue purified by chromatography (SiO₂, 5% acetone in CH₂Cl₂) to afford A1 (134 mg, 64%) as a colorless solid: [α]²⁵_D = - 71.0° (c, 0.1, CH₂Cl₂); ^1H NMR (CD₃Cl 400 MHz) δ 8.67 (s, 1H), 8.00 (br s, 1H), 7.46 (d, J = 8.6 Hz, IH), 7.05 (d, J = 2.3 Hz, IH), 7.00 (dd, J = 2.3, 8.6 Hz, IH), 5.82 (d, J = 1.5 Hz, IH), 5.02 (dd, J = 1.5, 7.8 Hz, IH), 4.94 (t, J = 7.8 Hz, IH), 3.62 (s, 3H), 3.30 (d, J = 7.8 Hz, IH), 2.26 (s, 3H), 1.37 (s, 3H), 1.21 (s, 3H); ^13C NMR (CD₃Cl 100 MHz) δ 169.7, 159.2, 157.4, 153.5, 151.4, 129.2, 123.9, 122.8, 115.1, 114.6, 104.1, 94.7, 83.4, 78.3, 77.6, 77.5, 61.1, 27.9, 25.2, 22.4; IR (film) ν_max 1819, 1764, 1615, 1560, 1507, 1375, 1300, 1212, 1168, 1107, 1072, 1034, 1002, 969 cm⁻¹, HRMS (FAB⁺) m/z 420.1285 (M + H⁺, C₂₀H₂₂NO₉ requires 420.1294).
Noviosylated coumarin A1 (20 mg, 0.047 mmol) was dissolved in methanolic ammonia (7.0 M, 2 mL) at 25 °C and stirred for 24 hours. The solvent was evaporated and the residue purified by preparative HPLC (SiO₂, 20% 2-propanol in hexanes) to afford A2 (4.2 mg, 22%), A3 (8.6 mg, 42%) and A4 (3.5 mg, 20%) as colorless solids.

**[0126]** A2: [α]D²⁵ = - 143.2 ° (c, 0.1, 50% MeOH in CH₂Cl₂); ¹H NMR (50% CD₃OD in CD₂Cl₂ 400 MHz) δ 8.58 (s, IH), 7.44 (d, J = 8.4 Hz, IH), 7.01 (s, IH), 6.97 (d, J = 8.4 Hz, IH), 5.59 (d, J = 2.0 Hz, IH), 5.03 (dd, J = 2.0, 3.6 Hz, IH), 4.25 (dd, J = 3.6, 9.7 Hz, IH), 3.57 (s, 3H), 3.30 (d, J = 9.7 Hz, IH), 2.19 (s, 3H), 1.31 (s, 3H), 1.13 (s, 3H); ¹³C NMR (50% CD₃OD in CD₂Cl₂ 100 MHz) δ 168.8, 157.2, 156.4, 155.5, 149.5, 126.9, 122.9, 120.4, 112.6, 112.3, 101.6, 94.8, 82.5, 77.0, 71.9, 64.7, 59.9, 27.0, 22.1, 20.6; IR (film) ν max 3473, 1716, 1689, 1610, 1540, 1528, 1505, 1375, 1240, cm⁻¹; HRMS (FAB⁺) m/z 437.1565 (M + H⁺, C₂₀H₂₅N₂O₉ requires 437.1560).

**[0127]** A3: [α]D²⁵ = - 116.2 ° (c, 0.24, 50% MeOH in CH₂Cl₂); ¹H NMR (CD₃OD 400 MHz) δ 8.59 (s, IH), 7.52 (d, J = 10.8 Hz, IH), 7.04 (s, IH), 7.03 (d, J = 10.8 Hz, IH), 5.56 (d, J = 2.4 Hz, IH), 5.25 (dd, J = 3.2, 9.8 Hz, IH), 4.20 (dd, J = 2.4, 3.2 Hz, IH), 3.58 (s, 3H), 3.35 (d, J = 9.8 Hz, IH), 2.22 (s, 3H), 1.27 (s, 3H), 1.18 (s, 3H); ¹³C NMR (CD₃OD 100 MHz) δ 171.6, 158.8, 158.7, 158.1, 151.8, 128.9, 125.6, 122.5, 114.4, 114.2, 103.1, 99.1, 81.6, 79.0, 71.8, 69.7, 60.1, 27.9, 22.9, 22.4; IR (film) ν max 3470, 1716, 1686, 1615, 1538, 1523, 1505, 1372, 1242, 1120 cm⁻¹; HRMS (FAB⁺) m/z 437.1576 (M + H⁺, C₂₀H₂₅N₂O₉ requires 437.1560).

**[0128]** A4: As shown in the scheme below, the coumarin ring (2) was constructed by the condensation of commercially available benzaldehyde 1 with glycine in the presence of acetic anhydride. See Madhavan et al., *Novel coumarin derivatives of heterocyclic compounds as lipid-Lowering agents*, Bioorg. Med. Chem. Lett. 13, 2547 (2003). After selective deprotection, the free phenol was coupled with the trichloroacetimidate of noviose carbonate (4) (Yu et al., *Synthesis of (-)-Noviose* from 2,3-O-Isopropylidene-D-erythronolactol, Org. Chem. 69, 7375-7380 (2004)) in the presence of catalytic boron trifluoride etherate (Shen et al., *Synthesis of Photolabile Novobiocin Analogues*, Bioorg. Med. Chem. Lett. 14, 5903-5907 (2004)). KU-1/A4 was furnished in excellent yield by
treatment of the cyclic carbonate 5 with triethylamine in methanol, resulting in solvolysis of the carbonate to afford the desired product.

![Chemical structure](image)

[0129] More specifically, triethylamine (0.2 mL) was added to a solution of noviosylated coumarin (45 mg, 0.10 mmol) in methanol (2 mL) at 25 °C. After stirring for 48 hours, the solvent was evaporated and the residue purified by preparative TLC (SiO₂, DCM-acetone; 4:1) to afford KU-1/A4 (35 mg, 0.086 mmol, 83%) as a white solid. N-(7-((2R,3R,4S,5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyl-tetrahydro-2H-pyran-2-yloxy)-2-oxo-2H-chromen-3-yl)acetamide (KU-1/A4). $[\alpha]_{25}^D = -351.6^\circ$ (c, 0.06, 50% MeOH in CH₂Cl₂); ¹HNMR (CD₃OD 400 MHz) $\delta$ 8.58 (s, 1H), 7.51 (d, $J = 8.3$ Hz, 1H), 7.03 (s, 1H), 7.02 (d, $J = 8.3$ Hz, 1H), 5.55 (d, $J = 2.3$ Hz, 1H), 4.10 (dd, $J = 3.3$, 9.6 Hz, 1H), 4.03 (dd, $J = 2.4$, 3.3 Hz, 1H), 3.60 (s, 3H), 3.38 (d, $J = 9.6$ Hz, 1H), 2.21 (s, 3H), 1.30 (s, 3H), 1.13 (s, 3H); ¹³CNMR (CD₃OD 100 MHz) $\delta$ 171.6, 158.9, 158.8, 151.8, 128.9, 125.7, 122.5, 114.3, 114.1, 103.1, 99.2, 84.2, 78.8, 71.5, 68.4, 61.1, 28.2, 22.9, 22.4; IR (film) $\nu_{max}$ 3326, 1714, 1674, 1613, 1558, 1553, 1108 cm⁻¹; HRMS (FAB+) $m/z$ 394.1492 (M + H⁺, Ci₉H₂₄O₈ requires 394.1502).
7-((3aR,4R,7R,7aR)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3aH-[1,3]dioxol[4,5-c]pyran-4-yloxy)-2H-chromen-2-one (B1). Noviose carbonate trichloroacetimidate (90 mg, 0.25 mmol) and 7-hydroxy-coumarin B (48 mg, 0.30 mmol) were dissolved in CH₂Cl₂ (2 mL) before boron trifluoride etherate (10 µL, 0.01 mmol) was added to the suspension at 25 °C. The mixture was stirred at 25 °C for eight hours and quenched with Et₃N (0.1 mL, 0.7 mmol). The solvent was removed and the residue purified by chromatography (SiO₂, 2% acetone in CH₂Cl₂) to afford B1 (66 mg, 73%) as a colorless solid: [α]D²⁵ = - 85.6° (c, 1.15, CH₂Cl₂); ¹H NMR (CDCl₃, 400 MHz) δ 7.69 (d, J = 9.5 Hz, IH), 7.43 (d, J = 8.6 Hz, IH), 7.05 (d, J = 2.3 Hz, IH), 6.95 (dd, J = 2.3, 8.6 Hz, IH), 6.34 (d, J = 9.5 Hz, IH), 5.84 (d, J = 1.3 Hz, IH), 5.03 (dd, J = 1.3, 7.7 Hz, IH), 4.94 (t, J = 7.7 Hz, IH), 3.62 (s, 3H), 3.30 (d, J = 7.7Hz, IH), 1.37 (s, 3H), 1.20 (s, 3H); ¹³CNMR (CDCl₃ 100 MHz) δ 161.2, 158.9, 155.9, 153.5, 143.5, 129.4, 114.7, 114.4, 113.7, 104.4, 94.6, 83.4, 78.3, 77.8, 77.5, 61.0, 27.9, 22.4; IR (film) νmax 1809, 1730, 1612, 1171, 1157, 1109 cm⁻¹; HRMS (FAB⁺) m/z 363.1083 (M + H⁺, C₇H₈O₈ requires 363.1080).

(3R,4S,5R,6R)-5-hydroxy-3-methoxy-2,2-dimethyl-6-(2-oxo-2H-chromen-7-yloxy)-tetrahydro-2H-pyran-4-yl carbamate (B2), (2R,3R,4R,5R)-4-hydroxy-5-methoxy-6,6-dimethyl-2-(2-oxo-2H-chromen-7-yloxy)-tetrahydro-2H-pyran-3-yl carbamate (B3) and 7-((2R,3R,4S,5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyl-tetrahydro-2H-pyran-2-yloxy)-2H-chromen-2-one (B4). Noviosylated coumarin B1 (25 mg, 0.07 mmol) was dissolved in methanolic ammonia (7.0 M, 2 mL) at 25 °C and stirred for 24 hrs. The solvent was evaporated and the residue purified by preparative TLC (SiO₂, 25% acetone in methylene chloride) to afford B2 (4.3 mg, 16%), B3 (14.5 mg, 52%) and B4 (4.0 mg, 17%) as colorless solids.

B2: [α]D²⁵ = -85.1° (c, 0.71, 50% MeOH in CH₂Cl₂); ¹H NMR (CD₃OD 400 MHz) δ 7.91 (d, J = 9.5 Hz, IH), 7.58 (dd, J = 1.3, 9.0 Hz, IH), 7.04 (s, IH), 7.03 (d, J = 9.0 Hz, IH), 6.30 (d, J = 9.5 Hz, IH), 5.65 (d, J = 2.1 Hz, IH), 5.04 (dd, J = 2.6, 3.4 Hz, IH), 4.28 (dd, J = 3.4, 9.9 Hz, IH), 3.62 (s, 3H), 3.39 (d, J = 9.5 Hz, IH), 1.35 (s, 3H), 1.15 (s, 3H); ¹³CNMR (CD₃OD 100 MHZ) δ 161.7, 159.7, 157.5, 155.3, 144.1, 129.1, 113.6, 113.4, 112.8, 103.0, 96.4, 83.9, 78.5, 73.4, 66.2, 60.8, 28.0, 21.8; IR (film) νmax 3438, 2982, 2932, 1731, 1616, 1403, 1338, 1280, 1117, 1002, 963 cm⁻¹; HRMS (FAB⁺) m/z 380.1333 (M + H⁺, C₇H₁₂O₂ requires 380.1345).

B3: [α]D²⁵ = -111.8° (c, 0.18, 50% MeOH in CH₂Cl₂); ¹H NMR (CD₃OD 400 MHz) δ 7.91 (d, J = 9.5 Hz, IH), 7.58 (d, J = 8.3 Hz, IH), 7.05 (s, IH), 7.04 (d, J = 8.3 Hz, IH), 7.04 (s, IH), 6.95 (d, J = 8.3 Hz, IH), 6.34 (d, J = 9.5 Hz, IH), 5.84 (d, J = 1.3 Hz, IH), 5.03 (dd, J = 1.3, 7.7 Hz, IH), 4.94 (t, J = 7.7 Hz, IH), 3.62 (s, 3H), 3.30 (d, J = 7.7Hz, IH), 1.15 (s, 3H); ¹³CNMR (CD₃OD 100 MHZ) δ 161.7, 159.7, 157.5, 155.3, 144.1, 129.1, 113.6, 113.4, 112.8, 103.0, 96.4, 83.9, 78.5, 73.4, 66.2, 60.8, 28.0, 21.8; IR (film) νmax 3438, 2982, 2932, 1731, 1616, 1403, 1338, 1280, 1117, 1002, 963 cm⁻¹; HRMS (FAB⁺) m/z 380.1333 (M + H⁺, C₇H₁₂O₂ requires 380.1345).
Hz, IH), 6.30 (d, J = 9.9 Hz, IH), 5.59 (d, J = 2.4 Hz, IH), 5.25 (dd, J = 3.2, 9.8 Hz, IH), 4.20 (dd, J = 2.4, 3.2 Hz, IH), 3.59 (d, J = 9.5 Hz, IH), 3.57 (s, 3H), 1.36 (s, 3H), 1.17 (s, 3H); $^1$C NMR (CD$_3$OD 100 MHz) $\delta$ 161.7, 159.9, 157.7, 155.3, 144.2, 129.1, 113.6, 113.5, 112.7, 102.9, 98.6, 81.1, 78.6, 71.4, 69.3, 60.6, 27.5, 22.0; IR (film) $\nu$$_{max}$ 3359, 2979, 2937, 1710, 1615, 1317, 1120, 1092, 995 cm$^{-1}$; HRMS (FAB$^+$) $m/z$ 380.1327 (M + H+, C$_{17}$H$_{21}$O$_7$ requires 380.1345).

B4: [$\alpha$]$^D_{25}$ = - 129.4 ° (c, 0.18, 50% MeOH in CH$_2$Cl$_2$); $^1$HNMR (CD$_3$OD 400 MHz) $\delta$ 7.91 (d, J = 9.5 Hz, IH), 7.57 (dd, J = 2.4, 10.4 Hz, IH), 7.02 (m, 2H), 6.27 (dd, J = 4.5, 9.5 Hz, IH), 5.57 (d, J = 2.4 Hz, IH), 4.11 (dd, J = 3.3, 9.5 Hz, IH), 4.03 (dd, J = 2.4, 3.3 Hz, IH), 3.60 (s, 3H), 3.39 (d, J = 9.5 Hz, IH), 1.35 (s, 3H), 1.12 (s, 3H); $^1$C NMR (CD$_3$OD 100 MHz) $\delta$ 161.7, 160.9, 155.4, 144.2, 129.0, 113.5, 113.4, 112.6, 102.9, 98.8, 83.7, 78.4, 71.1, 67.9, 60.7, 27.7, 22.0; IR (film) $\nu$$_{max}$ 3415, 2984, 2934, 1730, 1718, 1707, 1615, 1118, 999, 957 cm$^{-1}$; HRMS (FAB$^+$) $m/z$ 337.1 1279 (M + H+, C$_{17}$H$_{21}$O$_7$ requires 337.1287).

[0134] 7-((3R,4R,7R,7aR)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3aH-[1,3]dioxolo[4,5-c]pyran-4-yl)oxy)-4-methyl-3-phenyl-2H-chromen-2-one (Cl). Noviose carbonate trichloroacetimidate (90 mg, 0.25 mmol) and 7-hydroxy-4-methyl-3-phenyl-coumarin C (76 mg, 0.30 mmol) were dissolved in CH$_2$Cl$_2$ (2 mL) before boron trifluoride etherate (10 µL, 0.01 mmol) was added to the suspension at 25 °C. The mixture was stirred at 25 °C for eight hours and quenched with Et$_3$N (0.1 mL, 0.7 mmol). The solvent was removed and the residue purified by chromatography (SiO$_2$, 1% acetone in CH$_2$Cl$_2$) to afford Cl (92 mg, 73%) as a colorless solid: [$\alpha$]$^D_{25}$ = - 75.8 ° (c, 1.41, CH$_2$Cl$_2$); $^1$HNMR (CDCl$_3$ 400 MHz) $\delta$ 7.80 (d, J = 9.6 Hz, IH), 7.44 (m, 3H), 7.33 (m, 2H), 7.09 (d, J = 2.4 Hz, IH), 7.01 (dd, J = 2.4, 5.2 Hz, IH), 5.84 (d, J = 1.3 Hz, IH), 5.03 (dd, J = 1.3, 7.7 Hz, IH), 4.94 (t, J = 7.7 Hz, IH), 3.62 (s, 3H), 3.30 (d, J = 7.7Hz, IH), 2.31 (s, 3H), 1.37 (s, 3H), 1.20 (s, 3H); $^1$C NMR (CDCl$_3$ 100 MHz) $\delta$ 161.0, 158.0, 153.9, 153.0, 147.4, 134.3, 130.0 (2C), 128.3 (2C), 128.0, 126.2, 125.2, 115.6, 113.0, 103.7, 94.1, 82.9, 77.8, 76.7, 76.5, 60.5, 27.4, 22.0, 16.5; IR (film) $\nu$$_{max}$ 1874, 1715, 1612, 1564, 1507, 1383, 1262, 1167, 1130, 1113, 1070, 1033, 1006, 968, 936 cm$^{-1}$; HRMS(FAB$^+$) $m/z$ 453.1554 (M + H+, C$_{25}$H$_{23}$O$_8$ requires 453.1549).

[0135] (3R,4S,5R,6R)-5-hydroxy-3-methoxy-2,2-dimethyl-6-(4-methyl-2-oxo-3-phenyl-2H-chromen-7-yloxy)-tetrahydro-2H-pyran-4-yl carbamate (C2), (2R,3R,4R,5R)-4-hydroxy-5-methoxy-6,6-dimethyl-2-(4-methyl-2-oxo-3-phenyl-2H-chromen-7-yloxy)-tetrahydro-2H-pyran-3-yl carbamate (C3) and 7-((2R,3R,4S,5R)-3,4-dihydroxy-5-
methoxy-6,6-dimethyl-tetrahydro-2H-pyran-2-yl)-4-methyl-3-phenyl-2H-chromen-2-one (C4). Noviosylated coumarin C1 (25 mg, 0.055 mmol) was dissolved in methanolic ammonia (7.0 M, 2 mL) at 25 °C and stirred for 24 hours. The solvent was evaporated and the residue purified by preparative TLC (SiO2, 25% acetone in methylene chloride) to afford C2 (6.3 mg, 25%), C3 (13.7 mg, 53%) and C4 (3.0 mg, 13%) as colorless solids.

[0136] C2: [α]D 25° = -72.9 ° (c, 0.19, 50% MeOH in CH2Cl2); 1HNMR (CD3OD 400 MHz) δ 7.80 (d, J = 9.0 Hz, IH), 7.43 (m, 3H), 7.32 (m, 2H), 7.10 (m, 2H), 5.69 (d, J = 1.8 Hz, IH), 5.06 (dd, J = 2.1, 3.2 Hz, IH), 4.30 (dd, J = 3.2, 9.7 Hz, IH), 3.63 (s, 3H), 3.40 (d, J = 9.7 Hz, IH), 2.31 (s, 3H), 1.36 (s, 3H), 1.18 (s, 3H); 13CNMR (CD3OD 100 MHz) δ 162.2, 159.7, 158.0, 154.2, 149.2, 135.1, 130.3 (2C), 128.4 (2C), 128.1, 127.0, 124.7, 115.3, 113.7, 103.2, 96.8, 84.4, 78.9, 73.8, 66.7, 61.3, 28.4, 22.3, 15.8; IR (film) νmax 3474, 2986, 2924, 1713, 1605, 1382, 1355, 1263, 1124, 1001, 967 cm⁻¹; HRMS (FAB +) m/z 470.1821 (M + H+, C25H28NO8 requires 470.1815).

[0137] C3: [α]D 25° = -92.3 ° (c, 0.28, 50% MeOH in CH2Cl2); 1HNMR (CD3OD 400 MHz) δ 7.75 (d, J = 9.5 Hz, IH), 7.45 (m, 3H), 7.34 (m, 2H), 7.06 (m, 2H), 5.63 (d, J = 2.4 Hz, IH), 5.18 (dd, J = 3.2, 9.6 Hz, IH), 4.18 (dd, J = 2.4, 3.2 Hz, IH), 3.54 (s, 3H), 3.40 (d, J = 9.5 Hz, IH), 2.27 (s, 3H), 1.35 (s, 3H), 1.16 (s, 3H); 13CNMR (CD3CN 125 MHz) δ 160.7, 159.0, 156.0, 153.8, 148.0, 145.2, 130.1 (2C), 128.1 (2C), 127.7, 126.7, 124.4, 114.9, 113.1, 103.1, 98.2, 81.0, 78.4, 71.3, 69.0, 60.7, 27.7, 22.4, 15.8; IR (film) νmax 3459, 3331, 2981, 2925, 1714, 1606, 1379, 1335, 1263, 1124, 1072 cm⁻¹; HRMS (FAB +) m/z 470.181 (M + H+, C25H28NO8 requires 470.1815).

[0138] C4: [α]D 25° = -86.0 ° (c, 0.12, 50% MeOH in CH2Cl2); 1HNMR (CD3OD 400 MHz) δ 7.80 (d, J = 9.6 Hz, IH), 7.44 (m, 3H), 7.33 (m, 2H), 7.09 (m, 2H), 5.60 (d, J = 1.9 Hz, IH), 4.12 (dd, J = 3.3, 9.5 Hz, IH), 4.05 (dd, J = 2.4, 3.1 Hz, IH), 3.61 (s, 3H), 3.40 (d, J = 9.5 Hz, IH), 2.32 (s, 3H), 1.37 (s, 3H), 1.15 (s, 3H); 13CNMR (CD3OD 100 MHz) δ 161.9, 159.6, 153.8, 149.1, 134.7, 129.9 (2C), 127.9 (2C), 127.7, 126.5, 124.1, 114.7, 113.4, 102.7, 98.8, 83.8, 78.4, 71.1, 68.0, 60.7, 27.8, 22.0, 15.4; IR (film) νmax 3403, 2977, 2924, 1717, 1607, 1558, 1505, 1381, 1260, 1124, 992 cm⁻¹; HRMS (FAB +) m/z 427.1750 (M + H+, C24H27O7 requires 427.1757).

[0139] 8-(7-Methoxy-6,6-dimethyl-2-oxo-tetrahydro-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-chromen-2-one (Di) Noviose carbonate trichloroacetimidate (176 mg, 0.49 mmol) and 8-hydroxy-coumarin D (95 mg, 0.59 mmol) were dissolved in CH2Cl2 (5 mL). Boron trifluoride etherate (20 µL, 0.08 mmol) was added to the suspension at
25 °C. The resulting slurry was stirred at 25 °C for 10 hours before the solvent was removed and the residue purified by chromatography (SiO₂, 1% MeOH in CHCl₃) to afford D₁ (85 mg, 40%) as a colorless solid: [α]D²¹ = -57 ° (c = 0.1, 50% MeOH in CH₂Cl₂); ¹H NMR (CDCl₃, 500 MHz) δ 7.69 (d, J = 9.6 Hz, IH), 7.31 (t, J = 9.1 Hz, IH), 7.23 (dd, J = 2.8 Hz, 9.0 Hz, IH), 7.16 (d, J = 2.8 Hz, IH), 6.47 (d, J = 9.6 Hz, IH), 5.77 (d, J = 1.0 Hz, IH), 5.03 (dd, J = 1.2 Hz, 7.8 Hz, IH), 4.95 (t, J = 7.7 Hz, IH), 3.62 (s, 3H), 3.30 (d, J = 7.7 Hz, IH), 1.37 (s, 3H), 1.20 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 160.6, 153.1, 152.1, 149.4, 142.9, 120.8, 119.3, 118.0, 117.4, 113.3, 94.5, 82.9, 77.9, 77.2, 76.5, 60.5, 27.5, 22.0; IR (film) vₘₐₓ 3054, 2987, 1817, 1730, 1572, 1422, 1166, 1112, 1040, 896, 739 cm⁻¹; HRMS (FAB⁺) m/z 363.1088 (M + H⁺, C₁₂H₂₂NO₈ requires m/z 363.1080).

[0140] Carbamic acid 4-hydroxy-5-methoxy-6,6-dimethyl-2-(2-oxo-2 H-chromen-8-yloxy)-tetrahydro-pyran-3-yl ester (D₂), carbamic acid 5-hydroxy-3-methoxy-2,2-dimethyl-6-(2-oxo-2 H-chromen-8-yloxy)-tetrahydro-pyran-4-yl ester (D₃), 8-(3,4-Dihydroxy-5-methoxy-6,6-dimethyl-tetrahydro-pyran-2-yloxy)-chromen-2-one (D₄) D₁ (17 mg, 0.047 mmol) was dissolved in methanolic ammonia (2.0 M, 5 mL, 10 mmol) at 25 °C and stirred for five hours before the solvent was removed. The residue was purified by preparative TLC (SiO₂, 25% acetone in CH₂Cl₂) to afford D₂ (3.8 mg, 21%), D₃ (5.5 mg, 31%), and D₄ (7.2 mg, 46%) as colorless solids.

[0141] D₂: [α]D²¹ = -19 ° (c = 0.1, 50% MeOH in CH₂Cl₂); ¹H NMR (CD₃OD in CD₂Cl₂, 500 MHz) δ 7.79 (d, J = 9.6 Hz, IH), 7.26 (m, 3H), 6.43 (d, J = 9.6 Hz, IH), 5.59 (d, J = 2.0 Hz, IH), 5.05 (dd, J = 2.1 Hz, 3.4 Hz, IH), 4.28 (m, 2H), 3.61 (s, 3H), 3.32 (m, IH), 1.34 (s, 3H), 1.18 (s, 3H); ¹³C NMR (CD₃OD in CDCl₃, 100 MHz) δ 162.1, 158.0, 153.6, 149.3, 144.6, 121.1, 119.9, 117.7, 116.6, 113.6, 97.1, 84.5, 78.8, 74.1, 66.7, 61.4, 28.6, 22.4; IR (film) vₘₐₓ 3054, 2987, 1729, 1422, 896, 739, 705 cm⁻¹; HRMS (ESI⁺) m/z 380.1356 (M + H⁺, C₁₈H₂₆NO₈ requires m/z 380.1345).

[0142] D₃: [α]D²¹ = -69 ° (c = 0.1, 50% MeOH in CH₂Cl₂); ¹H NMR (CD₃OD in CD₂Cl₂, 500 MHz) δ 7.84 (d, J = 9.6 Hz, IH), 7.30 (m, 3H), 6.44 (d, J = 9.5 Hz, IH), 5.51 (d, J = 2.3 Hz, IH), 5.28 (dd, J = 3.2 Hz, 9.8 Hz, IH), 4.21 (m, IH), 3.56 (s, IH), 3.55 (s, 3H), 1.35 (s, 3H), 1.20 (s, 3H); ¹³C NMR (CD₃OD in CDCl₃, 125 MHz) δ 161.8, 157.4, 153.3, 148.7, 144.2, 120.8, 119.3, 117.4, 116.2, 113.2, 98.9, 81.3, 78.6, 71.5, 69.5, 60.8, 27.9, 22.3; IR (film) vₘₐₓ 3054, 2987, 1732, 1422, 896, 742 cm⁻¹; HRMS (ESI⁺) m/z 380.1348 (M + H⁺, C₁₈H₂₆NO₈ requires m/z 380.1345).
[0143] D4: [α]_D^{31} = - 91 ° (c = 0.1, 50% MeOH in CH₂Cl₂); ^1H NMR (CD₃OD in CD₂Cl₂, 500 MHz) δ 7.82 (d, J = 9.5 Hz, IH), 7.26 (m, 3H), 6.43 (d, J = 9.5 Hz, IH), 5.50 (d, J = 2.3 Hz, IH), 4.12 (dd, J = 3.4 Hz, 9.3 Hz, IH), 4.05 (d, J = 2.4 Hz, IH), 3.59 (s, 3H), 3.33 (m, IH), 1.35 (s, 3H), 1.15 (s, 3H); ^13C NMR (CD₃OD in CDCl₃, 125 MHz) δ 161.7, 153.4, 148.6, 144.2, 120.7, 119.3, 117.3, 116.1, 113.1, 98.9, 83.8, 78.3, 71.1, 68.0, 60.9, 28.0, 22.2; IR (film) v_max 3455, 3053, 2988, 1704, 1568, 1112, 738 cm⁻¹; HRMS (FAB⁺) m/z 337.1267 (M + H⁺, C₇H₂O₇ requires m/z 337.1287).

[0144] 6-(7-Methoxy-6,6-dimethyl-2-oxo-tetrahydro-[1,3]dioxolol[4,5-c]pyran-4-yloxy)-chromen-2-one (E1) Noviose carbonate trichloroacetimidate (150 mg, 0.42 mmol) and 6-hydroxycoumarin E (67 mg, 0.42 mmol) were dissolved in CH₂Cl₂ (4 mL). Boron trifluoride etherate (20 μL, 0.06 mmol) was added to the suspension at 25 °C. The resulting slurry was stirred at 25 °C for 10 hours before the solvent was removed and the residue purified by chromatography (SiO₂, 1% MeOH in CHCl₃) to afford E1 (63 mg, 42%) as a colorless solid: [α]_D^{31} = - 59 ° (c = 0.1, 50% MeOH in CH₂Cl₂); ^1H NMR (CDCl₃, 500 MHz) δ 7.69 (d, J = 9.6 Hz, IH), 7.30 (d, J = 9.0 Hz, IH), 7.23 (dd, J = 2.7 Hz, 9.0 Hz, IH), 7.16 (d, J = 2.7 Hz, IH), 6.47 (d, J = 9.6 Hz, IH), 5.77 (m, IH), 5.02 (dd, J = 1.0 Hz, 7.8 Hz, IH), 4.95 (d, J = 7.7 Hz, IH), 3.61 (s, 3H), 3.30 (d, J = 7.7 Hz, IH), 1.37 (s, 3H), 1.23 (s, 3H); ^13C NMR (CDCl₃, 125 MHz) δ 160.6, 153.1, 152.1, 149.4, 142.9, 130.8, 119.3, 118.0, 117.4, 113.3, 94.5, 82.9, 77.9, 72.7, 76.5, 60.5, 27.5, 22.0; IR (film) v_max 3054, 2987, 1818, 1730, 1422, 896, 739, 705 cm⁻¹; HRMS (FAB⁺) m/z 363.109 (M + H⁺, C₁₈H₂O₈ requires m/z 363.1080).

[0145] Carbamic acid 5-hydroxy-3-methoxy-2,2-dimethyl-6-(2-oxo-2 H-chromen-6-yloxy)-tetrahydro-pyran-4-y1 ester (E2), carbamic acid 4-hydroxy-5-methoxy-6,6-dimethyl-2-(2-oxo-2 H-chromen-6-yloxy)-tetrahydro-pyran-3-y1 ester (E3), 6-(3,4-Dihydroxy-5-methoxy-6,6-dimethyl-tetrahydro-pyran-2-yloxy)-chromen-2-one (E4) E1 (17 mg, 0.047 mmol) was dissolved in methanolic ammonia (7.0 M, 5 mL, 35 mmol) at 25 °C and stirred for five hours before the solvent was removed. The residue was purified by preparative TLC (SiO₂, 25% acetone in CH₂Cl₂) to afford compound E2 (7.8 mg, 34%), E3 (9.9 mg, 43%), and E4 (4.7 mg, 23%) as colorless solids.

[0146] E2: [α]_D^{31} = - 45 ° (c = 0.1, 50% MeOH in CH₂Cb)- ^1H NMR (CD₃OD in CD₂Cl₂, 500 MHz) δ 7.82 (d, J = 9.6 Hz, IH), 7.27 (m, 3H), 6.44 (d, J = 9.5 Hz, IH), 5.50 (d, J = 2.0Hz, IH), 5.05 (dd, J = 2.0 Hz, 3.4 Hz, IH), 4.28 (m, IH), 3.61 (s, 3H), 3.32 (m, IH), 1.34 (s, 3H), 1.18 (s, 3H); ^13C NMR (CD₃OD in CD₂Cl₂, 125 MHz) δ 161.6, 157.2, 153.1, 148.8, 143.9, 120.8, 119.3, 117.4, 116.4, 113.2, 96.7, 84.1, 78.4, 40
73.7, 66.3, 61.3, 28.4, 22.2; IR (film) ν_max 3054, 2987, 1729, 1422, 896, 738, 705 cm⁻¹; HRMS (ESI⁺) m/z 380.1327 (M + H⁺, C₈H₂₂NO₈ requires m/z 380.1345).

[0147] E3: [α]D⁰ -80 ° (c = 0.1, 50% MeOH in CH₂Cl₂); 1H NMR (CD₃OD in CD₂Cl₂, 500 MHz) δ 7.79 (d, J = 9.5 Hz, IH), 7.28 (d, J = 2.3 Hz, 2H), 7.25 (s, IH), 6.43 (d, J = 9.5 Hz, IH), 5.50 (d, J = 2.3 Hz, IH), 5.26 (dd, J = 3.2 Hz, 9.8 Hz, IH), 4.21 (t, J = 2.7 Hz, IH), 3.56 (m, IH), 3.55 (s, 3H), 1.35 (s, 3H), 1.19 (s, 3H); 13C NMR (CD₃OD in CD₂Cl₂, 125 MHz) δ 159.5, 155.0, 151.1, 146.8, 141.8, 118.7, 117.3, 115.4, 114.4, 111.2, 96.7, 79.3, 76.6, 69.6, 67.4, 59.0, 26.0, 20.4; IR (film) ν_max 3054, 2987, 1731, 1422, 1265, 896, 742 cm⁻¹; HRMS (ESI⁺) m/z 380.1324 (M + H⁺, C₈H₂₂NO₈ requires m/z 380.1345).

[0148] E4: [α]D⁰ -89 ° (c = 0.05, 50% MeOH in CH₂Cl₂); 1H NMR (CD₃OD in CD₂Cl₂, 400 MHz) δ 7.83 (d, J = 9.6 Hz, IH), 7.26 (m, 3H), 6.44 (d, J = 9.5 Hz, IH), 5.50 (d, J = 2.3 Hz, IH), 4.12 (dd, J = 3.4 Hz, 9.3 Hz, IH), 4.05 (d, J = 2.4 Hz, IH), 3.59 (s, 3H), 3.33 (m, IH), 1.34 (s, 3H), 1.14 (s, 3H); 13C NMR (CD₃OD in CD₂Cl₂, 125 MHz) δ 162.1, 153.8, 149.2, 144.5, 121.3, 119.8, 117.8, 116.8, 113.5, 99.3, 84.4, 78.8, 71.6, 68.5, 61.6, 28.6, 22.8; IR (film) ν_max 3454, 3054, 2987, 1705, 1568, 1422, 1111, 896, 738 cm⁻¹; HRMS (FAB⁺) m/z 337.1275 (M + H⁺, C₈H₂₂O₂ requires m/z 337.1287).

[0149] As discussed more fully below, these compounds were then tested for biological activity with respect to Hsp90 inhibition. Based on the results, various additional modifications to the side chains at R¹ and R² in the above scheme are proposed, as well as modifications to the coumarin ring and sugar moiety.

**EXAMPLE 2: DEGRADATION OF PHOSPHO-AKT**

[0150] Inhibition of Hsp90 results in the degradation of Hsp90-dependent clients via ubiquitination of the unfolded client followed by proteasome-mediated hydrolysis. To test whether Hsp90 client proteins were degraded in the presence of these novobiocin analogues, each member of the library from Example 1 was incubated with SKBr3 breast cancer cells at a concentration of 100 μM. Western blot analysis of the protein lysates demonstrated that several of the compounds were capable of causing the degradation of the Hsp90-dependent oncogenic client protein, phospho-AKT as represented in FIG. 1. Phospho-AKT was chosen as a client protein for this assay because of previous reports indicating that phospho-AKT is a more sensitive indicator of Hsp90 inhibition than
AKT. Geldanamycin (GDA, 0.5 µM) was used as a positive control for Hsp90 inhibition.

[0151] As can be seen from FIG. 1, A4/KU-1 (dil) and A3/KU-2 (3′-carbamate) were the most potent novobiocin analogues identified, based on their ability to inhibit Hsp90 and cause the degradation of phosphorylated AKT. As shown in FIG. 1, the most active compound identified in this assay was A4/KU-1 from the scheme above, which contains an N-acetyl side chain in lieu of the benzamide, lacks the 4-hydroxy of the coumarin moiety, and has an unmodified diol. Structure-activity relationships for these compounds suggests that attachment of the noviose moiety to the 7-position of the coumarin ring is preferred for biological activity (B vs. D and E). Further, incorporation of the amide linker (A) resulted in greater inhibitory activity than the unsubstituted derivative, B. It is likely that the diol (4) mimics the ribose ring in the normal substrate (ATP) and may explain why replacement with a cyclic carbonate (1) or 2′-carbamate (2) resulted in decrease of activity.

EXAMPLE 3: DEGRADATION OF HER-2

[0152] The IC50 for Hsp90 inhibitors is sometimes determined as the concentration of inhibitor required to produce 50% degradation of Her-2, another therapeutically important Hsp90 client protein involved in breast cancer. When KU-1/A4 was incubated with Skbr3 breast cancer cells at concentrations of 100 nM, 1 µM, and 10 µM, a rapid decrease in Her-2 was observed between 100 nM and 1 µM, as shown in the Western blot of FIG. 2. These data are normalized against actin, a non-Hsp90 client protein, used as a control for non-specific degradation. These data suggest the IC50 of KU-1/A4 is in the low micromolar range, whereas novobiocin in the same assay produces an IC50 of 700 µM.

EXAMPLE 4A: PROSTATE CANCER

[0153] The steroid hormone receptors are also dependent upon the Hsp90 protein folding machinery for activation and hormone binding. To determine whether KU-1/A4 had similar effects on the androgen receptor, KU-1/A4 was tested in both a mutated androgen receptor-dependent prostate cancer cell line (LNCaP) and a wild type androgen receptor prostate cancer cell line (LAPC-4). More specifically, the prostate cancer cells were grown in RPMI with 10% fetal calf serum in a standard fashion. Once the cells had
reached near confluence, they were treated with vehicle (DMSO) or varying concentrations of KU-1/A4 ranging from 10 nM to 100 µM for 24 hours. Cells were harvested and cell lysates prepared. Western blot analysis was then performed on the cell lysate utilizing commercially available antibodies against the androgen receptor, AKT, HIF-1α, Her2, and Hsp90. Actin was used as the control. More specifically, Western Blot analysis protein concentrations in serum samples were determined by the Pierce BCA protein assay kit according to the manufacturer's protocol. Western blot analysis (100 mg total protein/lane to start) was electrophoresed under reducing conditions on a SDS-PAGE gel. The separated proteins were transferred to a polyvinylidene difluoride membrane (Millipore, Bedford, MA) for 40 minutes at 80 V. The membranes were blocked for two hours at room temperature in Tris-buffered saline (pH 7.5) containing 0.2% I-block (Tropix, Bedford, MA), 1% milk, and 0.1% Tween-20 (TBS-T). The membranes were subsequently be incubated with a primary antibody to the above mentioned proteins (all of which have commercially available antibodies) overnight at 4 °C. The next day the membrane was washed three times in TBS-T followed by one hour incubation with an appropriate horseradish peroxidase labeled secondary antibody in blocking buffer (TBS-T). The membranes were again washed in TBS-T and Tris-buffered saline and developed in SuperSignal West Pico Chemiluminescent Substrate (Pierce, Rockford, IL) according to manufacturer's instructions. The blots were visualized by exposing the enhanced chemiluminescence-reacted blot to X-ray film.

[0154] In the LNCaP prostate cancer cell line, KU-1/A4 induced Hsp90 at the lowest concentrations tested (10 nM, FIG. 3). In contrast, degradation of the Hsp90 client proteins AR and AKT did not occur until 1 µM, providing a therapeutic window of more than 200-fold, suggesting increased levels of Hsps do not correlate directly with client protein degradation for inhibitors of the C-terminal ATP-binding pocket. In addition, KU-1/A4 drastically reduced levels of the androgen receptor at lower concentrations in the wild type androgen receptor prostate cancer cell line (LAPC-4).

[0155] To verify that KU-1/A4 was not affecting other transcriptional or translational processes that could account for decreased protein, Hsp90 levels were determined. Under normal conditions, Hsp90 binds heat shock factor 1 (HSF-1), but in the presence of Hsp90 inhibitors this interaction is lost and HSF-1 is able to induce the expression of Hsp90. As can be seen in FIG. 3 Hsp90 levels are significantly increased in a manner dependent on the concentration of KU-1/A4 consistent with similar results previously
obtained by incubation with geldanamycin and radicicol. Both of these data are in contrast to actin, which is not an Hsp90 client protein and thus remains unaffected by Hsp90 inhibitors.

**EXAMPLE 4B: AMIDE SIDE CHAIN MODIFICATIONS**

[0156] Since KU-1/A4 was shown to be the most potent C-terminal inhibitor of Hsp90 identified in Example 1, additional derivatives of the KU-1/A4 scaffold can be prepared. Modifications of the amide side chain allow for an in depth study of the hydrophobic cavity that binds to this portion of KU-1/A4 and the analogous benzamide of novobiocin. As such, analogues of KU-1/A4 that have increasingly larger hydrophobic groups by the use of different commercially available or readily synthesized anhydrides, such as those anhydrides shown in the scheme below. See Khoo, L.E., *Synthesis of Substituted 3-Aminocoumarins from Ethyl N-2-Hydroxyarylideneglycinates*, Syn. Comm. 29, 2533-2538 (1999), which is incorporated by reference.

![Scheme](image)

wherein in the scheme R is hydrogen, alkyl, alkenyl, alkynyl, aryl, carbocyclic, heterocyclic, aryl, or aralkyl, (and most preferably R is hydrogen, alkyl, aryl, and aralkyl);

and wherein R is hydrogen or CONH₂.

[0157] As part of this example, the amide linkage can also be reversed to determine the optimal profile of this functionality. As set forth in the scheme below, the 7-hydroxy-3-ethyl ester coumarin can be hydrolyzed to afford the corresponding acid, which can be coupled with amines that mimic the same side chains used in the KU-1/A4 amide studies for direct comparison of biological activity. Once coupled, the free phenols can be noviosylated as described earlier to afford the cyclic carbonate products. Treatment of the carbonate with methanolic ammonia can give the diol, 2- and 3-carbamoyl products as shown in the scheme below. See Shen et al., *Synthesis of*

wherein in the scheme R is hydrogen, alkyl, alkenyl, alkynyl, aryl, carbocyclic, heterocyclic, aryl, or aralkyl; and R' is hydrogen or CONH₂.

[0158] Most preferably, the R in the amide side chain is hydrogen, alkyl, aryl, and alkaryl, and the amines used in the above scheme are NH₃, methylamine, ethylamine, propylamine, n-butylamine, and phenylamine. However, it will be appreciated to those skilled in the art that other derivatives can be prepared in accordance with the above scheme, in addition to the KU-1/A4 analogues shown. That is, the amide side chain, coumarin ring, and sugar may be modified in accordance with the other examples shown herein.

EXAMPLE 5: ISOCOUMARIN DERIVATIVES

[0159] To determine the most favorable interaction of the coumarin lactone with Hsp90, the isocoumarin derivative of the compounds of the present disclosure is prepared. For example, with respect to KU-1/A4, the isocoumarin is prepared from the 4-benzyloxy lactone shown in the scheme below. Treatment of the lactone with sodium cyanide, followed by HCl/pyridine is known to produce similar isocoumarins. See Wells et al., Facile synthesis of 3-acylaminoisocoumarins, J. Org. Chem. 36, 1503-1506 (1971), which is incorporated by reference. Acylation of the amine followed by removal of the benzyl-protecting group provides the phenol, which can be coupled with noviose trichloroacetimidate to afford the cyclic carbonate precursor. Ammonolysis of the cyclic carbonate can afford both the diol and 3'-carbamoyl products.
It is appreciated to those skilled in the art that other isocoumarin derivatives can be prepared in accordance with the above scheme, in addition to the KU-1/A4 analogue shown. That is, the amide side chain, coumarin ring, and sugar is modified in accordance with the other examples shown herein.

EXAMPLE 6: DES(DIMETHYL) AND DESMETHOXY SUGARANALOGUES

Modifications to the gem-dimethyl groups and the methyl ether on the noviose moiety can be prepared. In this example, the des(dimethyl) and desmethoxy sugar analogues can be prepared. Using KU-1/A4 as an example in the scheme below, 2,3-O-isopropylidene-L-erythronolactol can be converted to the corresponding alkene by Wittig olefination. Dihydroxylation can afford the syn diol as noted in the earlier synthesis of noviose. See Yu et al., Synthesis of (-)-Noviose from 2,3-O-Isopropylidene-D-erythronolactol, J. Org. Chem. 69 7375-7378 (2004). Protection of the primary alcohol, followed by alkylation of the secondary alcohol can afford the orthogonally protected molecule. Selective removal of the benzyl group and oxidation of the resultant alcohol can give the aldehyde. Treatment of this aldehyde with aqueous sulfuric acid can remove the acid-labile protecting groups while simultaneously promoting cyclization.

Similarly, the desmethoxy compound can be prepared from the appropriately functionalized lactone (Stewart et al., 2-Deoxy-L-Ribose from L-Arabinose-1, 5-lactone, Tetrahedron Assym. 13, 2667-2672 (2002)) by the addition of excess methyl Grignard to provide the primary and tertiary alcohol product. Oxidation of the primary alcohol can give the lactone, which can be reduced to the lactol before deprotection with aqueous sulfuric acid to yield the desmethoxy product. Once obtained, these sugars can be treated with carbonyl diimidazole to furnish the cyclic carbonates before coupling with

wherein preferably R is lower alkyl; and
wherein R’ is preferably hydrogen or -CONH₂.

[0163] It will be appreciated that other demethylated and/or dealkoxylated derivatives can be prepared in accordance with the above scheme, in addition to the modified KU-1/A4 derivative shown above. That is, the amide side chain, coumarin ring, and sugar may be modified in accordance with the other examples shown herein.

**EXAMPLE 7: MODIFIED NOVOBIOCIN DERIVATIVES**

[0164] This example involves the modification to of the compounds of the present disclosure to complement the hydrogen bonding capabilities of the nucleotide bases (adenine and guanine) with those of the coumarin ring system as shown below. As an example, these analogues contain conformationally restricted hydrogen bond donors/acceptors of KU-1/A4 (F and G) and strategically placed hydrogen bond acceptors/donors to complement those found in guanine (H-L). In all cases, the hydrophobic pocket that accommodates the m-substituted benzamide ring of novobiocin can be probed by alteration of the side chain constituents. Although the schemes below are directed to preparing modifications of KU-1/A4, it will be appreciated to those skilled in the art that the same modifications could be made in conjunction with other analogues described herein, such as the A-E compounds of Example 1.
EXAMPLE 7F: HETEROCYCLIC MODIFICATIONS TO OUINOLONE

[0165] In this example, the coumarin ring is modified to create F analogues that resemble guanine and contain a conformationally biased hydrogen-bond donor/acceptor. The synthesis begins with commercially available 4-hydroxy-2-nitrobenzaldehyde following the procedure of Meanwell, et al., *Inhibitors of Blood Platelet cAMP Phosphodiesterase.* 2. Structure-Activity Relationships Associated with 1,3-Dihydro-2H-imidazo[4,5-b]quinolin-2-ones Substituted with Functionalized Side Chains, J. Med. Chem. 35 2672-2687 (1992). The phenol can be protected as the benzylether, followed by treatment with hydantoin phosphonate to give the corresponding olefin. See Meanwell et al., *Diethyl 2,4-dioxoimidazolidine-5-phosphonate: A Wadsworth-Emmons Reagent for the Mild and Efficient Preparation of C-5 Unsaturated Hydantoins,* J. Org. Chem. 56 6897-6904 (1991). Reduction of the benzylether, nitro, and olefin functionalities can provide the appropriate amine for subsequent addition to the carbonyl
upon treatment with iodine. See Meanwell et al., *Inhibitors of Blood Platelet cAMP Phosphodiesterase, Structure-Activity Relationships Associated with 1,3-Dihydro-2H-imidazo[4,5-b]quinolin-2-ones Substituted with Functionalized Side Chains*, J. Med. Chem. 35 2672-2687 (1992). As depicted earlier, the unmasked phenol can be coupled with the trichloroacetimidate of noviose carbonate, followed by removal of the carbonate moiety to furnish analogue F.

![Chemical Structure Diagram]

[0166] It will be readily appreciated to those skilled in the art that the foregoing scheme for the F analogues can be readily modified to prepare the following compounds, in addition to the oxidized imidazole attached to the quinolone shown above, by using commercially available or readily synthesized bases. Thus, the present disclosure encompasses novobiocin derivatives according to the formula:

![Chemical Structure Diagram]

wherein $X_4, X_5, X_6, X_8$ are preferably each -CH-; and wherein $R^a, R^b,$ and $R^c$ are independently hydrogen, alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl; or wherein $R^b$ is oxidized to form the carbonyl according to the formula:
EXAMPLE 7G: HETEROCYCLIC MODIFICATIONS

[0167] In this example, coumarin G is prepared from 7-benzyloxy-4-hydroxy-3-nitrocoumarin, according to the scheme below. See Buckle et al., Aryloxyalkyloxy- and aralkyloxy-4-hydroxy-3-nitro coumarins which inhibit histamine release in the rat and also antagonize the effects of a slow reacting substance of anaphylaxis, J. Med. Chem. 22 158-168 (1979). Treatment of the 4-hydroxyl group with phosphorous oxychloride (POCl₃) affords the corresponding 4-amino derivative upon subsequent exposure to ammonia. See Rassochandran et al., Mild method for the preparation of 4-chloro-3-nitro coumarins, Indian. J. Chem. 25B 328-329 (1986). Reduction of the nitro group, followed by reaction with triethyl orthoformate in the presence of acid affords the desired compound. See Trkovnik et al., Synthesis of new heterocyclocoumarins from 3,4-diamino- and 4-chloro-3-nitrocoumarins, Prep. Proced. Int. 19 450-455 (1987). Treatment of this 3,4-diamine with other commercially or readily available orthoesters (see McElvain et al., Ketene acetals. XVI. Phenylketene diethyl- and dimethylacetals from the pyrolysis of the corresponding orthoesters, J. Am. Chem. Soc. 68 1917-1921 (1946)) can provide a direct method for exploration of the hydrophobic pocket surrounding this moiety. The orthoesters readily condense with 1,2-diamines to produce the corresponding heterocyclic compounds. Once prepared, these compounds can be coupled with noviose carbonate in analogous fashion to that shown in above to afford the corresponding G analogues of KU-I/A4.
It will be readily appreciated to those skilled in the art that the foregoing scheme can be readily modified to prepare the following compounds, in addition to the imidazole shown above by using different orthoesters.

wherein $X_5$, $X_6$, $X_8$ are preferably each -CH$_2$; and

wherein $R^a$, $R^b$, and $R^c$ are independently hydrogen, alkyl, alkenyl, alkynyl, carbocyclic, heterocyclic, aryl, or aralkyl; or wherein $R^b$ is oxidized to form the carbonyl according to the formula:
EXAMPLE 7H

[0169] The nitrogen-containing H variants of the coumarin ring can be prepared from 2-methyl-3,5-pyridinediol, by bromination of the benzylic methyl group, followed by hydrolysis and oxidation to the corresponding aldehyde as set forth in the scheme below. See Morisawa et al., Anticoccidal agents. IV. Modification at the 5-position of 4-deoxypyridoxol and a4-norpyridoxol, Agric. Biol. Chem. 39 1275-1281 (1975). Using conditions previously employed for the syntheses of other coumarin derivatives by us, the aldehyde can be treated with glycine under basic conditions to yield the azacoumarin ring system. See Billeret et al., Convenient synthesis of 5-azacoumarins, J. Hetero. Chem. 30 671-674 (1993). Acylation of the amine with various anhydrides can furnish the acylated 7-hydroxyl and 4-amino derivatives, of which the 7-phenolic ester can be readily cleaved by subsequent treatment with potassium carbonate in methanol. The resulting phenol can be coupled with noviose carbonate as described earlier.

\[ \text{Example 7H} \]

[0170] While the scheme above illustrates the modified coumarin of KU-1/A4 with a limited number of amide side chain substitutions, it will be appreciated to those skilled in the art that other derivatives can be prepared in accordance with the above scheme, in addition to the KU-1/A4 analogues shown. That is, the amide side chain, coumarin ring, and sugar may be modified in accordance with the other examples shown herein.

EXAMPLE 71: COUMARIN SIDE CHAINS

[0171] The I analogues are directed to other side-chains extending from the coumarin ring. As an example, the KU-1/A4 coumarin ring can be prepared from 2,4-dihydroxy-5-nitrobenzaldehyde (see Chandrashekhar et al., g-substitution in the resorcinol nucleus, VI. Formylation of4-nitro and 2-nitro resorcinols, Proc. Ind. Acad. Sci. 29A 227-230 (1949)) and 2,4-dihydroxy-5-methoxybenzaldehyde (see Demyttenaere et al., Synthesis of6-methoxy-4H-l-benzopyran-7-ol, a character donating component of the fragrance of Wisteria sinensis, Tetrahedron 58 2163-2166 (2002))
according to the procedure of Khoo et al., *Synthesis of substituted 3-aminocoumarins from ethyl N-2-Hydroxyarylideneglycinates*, Syn. Commun. 29 2533-2538 (1999), as generally set forth in the scheme below. The o-hydroxybenzaldehyde can be treated with ethyl glycine under acidic conditions to afford the corresponding free amine upon basic workup. Both the amino and hydroxyl functionalities can be acylated with the same anhydrides as shown above. Subsequent hydrolysis of the phenolic ester can provide the coumarin amide, which can be coupled directly with noviose carbonate as described previously.

wherein in the scheme R is hydrogen, alkyl, alkenyl, alkynyl, aryl, carbocyclic, heterocyclic, aryl, or aralkyl;

wherein X is alkyl, alkenyl, alkynyl, aryl, aralkyl, alkoxy, halogen, or nitro.

[0172] Again, while the scheme above illustrates the modified coumarin ring of KU-1/A4 with a limited number of amide side chain substitutions, it will be appreciated to those skilled in the art that other derivatives can be prepared in accordance with the above scheme, in addition to the KU-1/A4 analogues shown. That is, the amide side chain, coumarin ring, and sugar may be modified in accordance with the other examples shown herein.

**EXAMPLE 7J: HETEROCYCLICS**

[0173] The J analogues can be prepared from 4-chloro-2-hydroxy-5-nitrobenzaldehyde (see Pal et al., *New arylsulfonylhydrazones of substituted benzaldehyde as anticancer agents*, Neoplasms 30 551-556 (1983)) by treatment with glycine, acetic anhydride, and sodium acetate as mentioned previously for the preparation of other coumarin derivatives as set forth in the following scheme. See Khoo et al., *Synthesis of substituted 3-aminocoumarins from ethyl N-2-Hydroxyarylideneglycinates*, Syn. Commun. 29 2533-2538 (1999). The chloro substituent can undergo nucleophilic aromatic displacement with ammonia as a consequence of the electron withdrawing \( \alpha \)-lactone and o-nitro group. Upon formation
of the 7-amino-6-nitrocoumarin, the nitro group can be reduced and immediately treated with triethyl orthoformate to produce the imidazole ring that resembles guanine. See Buckle et al., *Aryloxyalkyloxy- and aralkyloxy-4-hydroxy-3-nitro coumarins which inhibit hisamine release in the rat and also antagonize the effects of a slow reacting substance of anaphylaxis*, J. Med. Chem. 22 158-168 (1979). Subsequent treatment with lithium diisopropylsilylamide and trimethylsilyl trifluorosulfonic acid can provide the TMS-protected diaza compound. See Vorbruggen et al., *Organic Reactions*, Volume 55, John Wiley and Sons, NY pp 12-14 (2000) and references therein. The trichloroacetimidate of noviose carbonate can be added to a solution of this TMS-protected coumarin followed by addition of trifluoroacetic acid to afford the coupled product. Upon exposure of the cyclic carbonate to triethylamine in methanol, the resulting diol can be produced in a similar fashion as was used to make KU-1/A4 directly from the corresponding cyclic carbonate.

![Chemical structure](image)

wherein R is hydrogen, alkyl, alkenyl, alkynyl, aryl, carbocyclic, heterocyclic, aryl, or aralkyl.

**EXAMPLE 7K**

[0174] The K analogues of the KU-1/A4 coumarin moiety can be prepared from 5-methoxy-2-methylbenzonitrile as set forth in the scheme below. See Tomita et al., *Schmidt reaction with benzocycloalkenones*, J. Chem. Soc. C: Organic 2 183-188 (1969). Bromination of the benzylic methyl group, followed by displacement with potassium cyanide can furnish the dinitrile product, which is a substrate for acid catalyzed cyclization to form the corresponding 2-bromoisoquinoline. See Johnson et al., *The cyclization of dinitriles by anhydrous halogen acids. A new synthesis of isoquinolines*, J. Org. Chem. 27 3953-3958. Acylation of the free amine with the anhydrides shown in
Scheme 4 can furnish the amide products, which can be treated with dilute hydrochloric acid to produce the isoquinolone. As before, the free phenol can be coupled with noviose carbonate trichloroacetimidate, followed by removal of the cyclic carbonate to furnish K and its acylated (R) derivatives.

wherein R is hydrogen, alkyl, alkenyl, alkynyl, aryl, carbocyclic, heterocyclic, aryl, or aralkyl.

[0175] Again, while the scheme above illustrates the modified coumarin ring of KU-1/A4 with a limited number of amide side chain substitutions, it will be appreciated to those skilled in the art that other derivatives can be prepared in accordance with the above scheme, in addition to the KU-1/A4 analogues shown. That is, the amide side chain, coumarin ring, and sugar may be modified in accordance with the other examples shown herein.

**EXAMPLE 7L: QUINOLINES**

wherein \( R \) is hydrogen, alkyl, alkenyl, alkynyl, aryl, carbocyclic, heterocyclic, aryl, or aralkyl.

[0177] Again, while the scheme above illustrates the modified coumarin ring of KU-1/A4 with a limited number of amide side chain substitutions, it will be appreciated to those skilled in the art that other derivatives can be prepared in accordance with the above scheme, in addition to the KU-1/A4 analogues shown. That is, the amide side chain, coumarin ring, and sugar may be modified in accordance with the other examples shown herein.

**EXAMPLE 8: CHLOROBIOCIN ANALOGUES**

[0178] This example involves the modification of the carbohydrate reside. More specifically, analogues similar to that of novobiocin's chlorinated pyrrolic ester, chlorobiocin, can be prepared.

![Chemical structure of chlorobiocin](image)

**Chlorobiocin**

[0179] As an example, compound KU-1/A4 can be prepared, and then coupled with a variety of acids to selectively afford the equatorial acylated alcohols. Selective acylation is based upon previous studies aimed at the preparation of photolabile derivatives of novobiocin. See Shen et al., *Synthesis of Photolabile Novobiocin Analogues*, Bioorg. Med. Chem. Lett. 14 5903-5906 (2004), which is incorporated by reference. These acids can include the pyrrolic acid found in chlorobiocin as well as several other that are shown in the scheme below. Exemplary acids include pyrrolic
acids, indolic acids, pyridinic acids, benzoic acids, salicylic acid, para-hydrobenzoic acid, thiobenzoic acid, and pyrazolic acid. In one aspect, the sugar can be modified to include a functional group according to the formula \(-\text{R'}-\text{OR}''\), wherein \(\text{R}'\) is a covalent bond or alkyl, and \(\text{R}''\) is an acyl group. Most preferably, the acyl derivative comprises the group \(-\text{COR}\) wherein \(\text{R}\) is alkyl, aryl, aralkyl, or an aromatic heterocyclic group.

Alkylated, aralkylated, thiolated, halogenated, and hydroxylated pyroles, indoles, pyridines, and pyrazoles are attached to the sugar ring as shown in the scheme below.

In another aspect, various substituents can be added to the amine of the carbamate side chain. As an example, carbonate KU-9/A1 can be prepared and amines added to provide the 3'-carbamoyl products as generally set forth in the scheme below. Thus, in one aspect the sugar can be modified to include a functional group according to the formula \(-\text{R}-\text{OR}'\)', wherein \(\text{R}\) is a covalent bond or alkyl, and \(\text{R}'\) is C-amido. Most preferably, the C-amido group is \(-\text{CONRR}''\) wherein \(\text{R}=\text{H},\) and \(\text{R}'\) is alkyl, aralkyl, or an aromatic heterocyclic group. Pyroles, halogenated benzyls and pyridines, and alkyl groups are shown as the modified side chain of the sugar in the scheme below.

![Chemical structures](image-url)

wherein \(X\) is alkyl, alkenyl, alkynyl, hydroxyl, halo, and \(n\) is an integer, preferably 0, 1, 2, 3, or 4.

### EXAMPLES 9-11:

**FURANOSE AND PYRANOSE NOVOBIOCIN DERIVATIVES**

In this example, new pyranose and furanose derivatives can be prepared that have affinity with the sugar of GTP and phosphate binding region of Hsp90. These selected compounds are shown in below and include ester, amide, sulfonic ester, phosphonic ester, carbamoyl, sulfonamide, and hydroxyl derivatives. Initial compounds can be coupled with the coumarin ring present in KU-1/A4, but when a more potent
analogue is obtained, the best sugar derivative from these studies can be placed onto the optimized ring system.

EXAMPLES 9 AND 10: SYNTHESIS OF FURANOSE DERIVATIVES

[0182] The o-acetyl derivative can be prepared from ribose (9.1, Scheme 9). Treatment of the ribose hemiacetal with benzyl alcohol and hydrochloric gas can provide the benzyloxyacetal, 9.2. See Pigro et al., Readily available carbohydrate-derived imines and amides as chiral ligands for asymmetric catalysis, Tetrahedron 58 5459-5466 (2002).

[0183] Subsequent reaction with carbonyl diimidazole can furnish the 2,3-cyclic carbonate (9.3), (See Peixoto et al., Synthesis of Isothiochroman 2,2-dioxide and 1,2-benzoxathiin 2,2-dioxide Gyrase B Inhibitors, Tetrahedron Lett. 41 1741-1745 (2000)) allowing the primary alcohol to react with acetyl chloride in the following step. Debenzylation, followed by conversion to the trichloroacetimidate 9.5 (See Peixoto et al., Synthesis of Isothiochroman 2,2-dioxide and 1,2-benzoxathiin 2,2-dioxide Gyrase B Inhibitors, Tetrahedron Lett. 41 1741-1745 (2000)) can furnish a suitable substrate for coupling with the KU-1/A4 coumarin ring system. As noted in previous work, coupling of trichloroacetimidates with phenols in the presence of catalytic boron trifluoride affords one stereoisomer (9.6), which results from attack of the intermediate oxonium species away from the sterically crowded cyclic carbonate. See Shen et al., Synthesis of Photolabile Novobiocin Analogues, Bioorg. Med. Chem. Lett. 14 5903-5906 (2004).
has been previously observed that treatment of similar cyclic carbonates with methanolic triethylamine readily provides the corresponding diol products (9.7) in high yields (greater than 80%).

[0184] The remaining furanose derivatives can be prepared from benzyl-protected ribose carbonate (9.3, Scheme 10). Both the sulfonamide and N-acetyl analogues can be furnished by conversion of primary alcohol (9.3) to the corresponding azide by a Mitsunobu reaction with bis(azido)zinc pyridine complex. See Viaud et al., Zinc azide mediated Mitsunobu substitution, An expedient method for the one-pot azidation of alcohols, Synthesis 130-132 (1990). The resulting azide (10.1) can be reduced, and the primary amine converted to the sulfonamide and N-acetyl functionalities, 10.2 and 10.3, respectively. See Hansson et al., Synthesis of Beta-benzyl N-(tert-butoxycarbonyl)-L-erythro-Beta-(benzyloxy)aspartate from (R,R)-(+) -tartaric acid, J. Org. Chem. 51 4490-4492 (1986). To prepare methyl ester 10.4, the free alcohol can be oxidized directly to the acid, followed by methylation. Carbamate 10.5 can also be prepared from the same alcohol, simply by treatment with trichloro acetyl isocyanate according to the procedure of Kocovsky, Carbazates: a method of synthesis and some synthetic applications, Tetrahedron Lett. 27 5521-5524 (1986). Both the sulfonic ester and the phosphonic ester can be prepared by conversion of 9.3 to iodide 10.6, followed by generation of the requisite enolate to displace the halide. See Callant et al., An efficient preparation and the intramolecular cyclopropanation of Beta-diazo-Beta-ketophosphonates and Beta-diazo phosphonoacetates, Syn. Commun. 14 155-161 (1984). Subsequent treatment with palladium (0) and an amine can lead to allyl removal followed by decarboxylation to form 10.10 and 10.8. See Guibe, Allyl esters and their use in complex natural product syntheses, Tetrahedron 54 2967-3041 (1998).
EXAMPLE 11: SYNTHESIS OF PYRANOSE DERIVATIVES.

[0185] The pyranose derivatives, which resemble noviose and a ring-expanded ribose ring, can be prepared by our recently reported synthesis of 11.1. See Yu et al., Synthesis of Mono- and dihydroxylated furanoses, pyranoses, and an oxepanose for the Preparation of Natural Product Analogue Libraries, J. Org. Chem. 70 5599-5605 (2005), which is incorporated by reference in its entirety. The pyranose derivatives can be prepared in a similar manner from the known dihydropyrones (See Ahmed et al., Total synthesis of the microtubule stabilizing antitumor agent laulimalide and some nonnatural analogues: The power of Sharpless 'Asymmetric Epoxidation, J. Org. Chem. 68 3026-3042 (2003)), which is available in four steps from commercially available triacetyl D-glucal (Roth et al., Synthesis of a chiral synthon for the lactone portion of compactin and mevinolin, Tetrahedron Lett. 29 1255-12158 (1988)). The pyranose can be furnished by Sharpless asymmetric DI hydroxylation (SAD) of the olefin to give the product in high diastereomer excess (KoIb et al., Catalytic Asymmetric Dihydroxylation, Chem. Rev. 94 2483-2547 (1994)), which can be converted to the cyclic carbonate at a later time.
Reduction of the lactone with diisobutyl aluminum hydride can give lactol 11.2, which upon treatment with benzyl alcohol and hydrochloric gas can give the benzyloxyacetal 11.3. Similar studies have been used to prepare noviose from arabinose using an identical sequence of steps. See Peixoto et al., Synthesis of Isothiochroman 2,2-dioxide and 1,2-benzoxathiin 2,2-dioxide Gyrase B Inhibitors, Tetrahedron Lett. 41 1741-1745 (2000). The corresponding diol can be treated with carbonyl diimidazole to yield cyclic carbonate 11.4. The primary alcohol can be converted to the same functionalities as shown in the scheme above, using the chemistry depicted for the furanose derivatives.

Once the benzyl protected pyranose derivatives are prepared, they can undergo hydrogenolysis to afford the hemiacetal. Treatment of the lactol with trichloroacetonitrile can furnish the corresponding trichloroacetimidate for subsequent coupling with the requisite coumarin/coumarin analogue. The procedure outlined herein illustrates the success of coupling such compounds with the coumarin phenol and this procedure can be used to prepare the corresponding analogues as described herein.

Using the foregoing schemes, the syntheses of eight protected pyranose analogues that include mono- and dihydroxylated variants of both ring-expanded and ring contracted analogues. All eight of these compounds were orthogonally protected, such that the hemiacetal could be coupled directly to the coumarin phenol as used similarly for the construction of A4. Subsequent removal of the protecting group(s) or treatment of the cyclic carbonate with ammonia, can afford the corresponding diol or carbamate products as demonstrated earlier.
EXAMPLE 12: PREPARATION OF 4-DESHYDROXY
AND 8-DESMETHYL ANALOGUES

[0189] In this example, the 4-deshydroxy and 8-desmethyl variants of novobiocin can be prepared along with the 8-methyl and 4-hydroxy analogues of KU-2/A3 (3'carbamate) as shown below. Not only can the 3'-carbamoyl derivatives of these compounds be prepared, but also the corresponding diols for direct comparison to KU-1/A4 (diol).

The 7-hydroxyl can undergo selective noviosylation and the diazine can be reduced. The corresponding amine can be coupled with the known carboxylic acid and the carbonate opened with methanolic ammonia to give both 3-carbamoyl and diol derivatives. 4-Deshydroxy-8-desmethylnovobiocin can be constructed from 3-amino-7-hydroxy coumarin in analogous fashion as depicted in the scheme below. The KU-1/A4 and KU-2/A3 analogues incorporating the same coumarin functionalities can be prepared by an identical method (see Khoo, *Synthesis of Substituted 3-Aminocoumarins from Ethyl N-2-Hydroxyarylideneglycinates*, Syn. Comm. 29 2533-2538 (1999)) using acetic anhydride in lieu of the prenylated 4-hydroxybenzoic acid. Des(carbamoyl) derivatives of these compounds can also be prepared by removal of the cyclic carbonate with triethylamine in methanol, which affords similar products in stoichiometric yields.
EXAMPLE 12A: PREPARATION OF 4-DESHYDROXY AND 8-DESMETHYL ANALOGUES

The substituted benzamide side chain of novobiocin was prepared from methyl 3-allyl-4-hydroxybenzoate, 5 in the scheme below. Attempts to perform cross-metathesis on this substrate failed as complexation with the Grubbs' catalyst appeared to have occurred with the orthophenol substrate. Therefore, the phenol was temporarily masked as the acetate, which allowed for a productive cross-metathesis reaction between 2-methyl-2-butene and the allyl appendage in excellent yield to provide the prenylated benzoic ester, 7. The ester product (7) was then hydrolyzed and the phenol reprotected as the acetate to prevent subsequent ester formation. Attempts to couple the unprotected phenol as well as the benzoic acid directly with the coumarin amine resulted in the formation of a complex mixture of products that produced only trace amounts of the desired amide. Therefore, acid 9 was converted to the corresponding acid chloride (10) in high yield following standard conditions.
Preparation of the 4-deshydroxy coumarin ring was achieved by the condensation of 2-methylresorcinol (11) with the CBz-protected vinylagous carbamate 12, which produced the desired coumarin 13, in modest yield in the scheme below. The phenol was then noviosylated with the trichloroacetimidate of noviose (14) in the presence of catalytic amounts of boron trifluoride etherate to generate 15 in good yield. Hydrogenolysis of the benzyl carbonate afforded the amine 16, which was readily coupled with the acid chloride 10 to give 17 in good yield. Both the acetate and the cyclic carbonate were removed and modified, respectively, to give the desired 3'-carbamoyl product, 4-deshydroxynovobiocin (DHN1) in good yield. Alternatively, the acetate and cyclic carbonate could be readily hydrolyzed to yield the desired 3'-descarbamoyl-4-deshydroxynovobiocin product (DHN2) in a single step upon treatment with methanolic triethylamine.
Methyl 3-allyl-4-hydroxybenzoate (5). A mixture of methyl-4-allyloxybenzoate (4.74 g, 24.7 mmol) in JV,iV-diethylaniline (10 mL) was heated at reflux for 48 hours and cooled to room temperature. The mixture was diluted with diethyl ether (50 mL), washed with aqueous HCl (10% v/v, 3 x 20 mL), dried (Na₂SO₄), filtered and concentrated. The residue was purified by chromatography (10:1→5:1, hexanes:EtOAc) to afford 5 (3.55 g, 75%) as an off-white solid: ¹H NMR (CDCl₃, 400 MHz) δ 7.86 - 7.84 (m, 2H), 6.85 (dd, J = 2.8, 7.7 Hz, 1H), 6.07 - 5.95 (m, 1H), 5.68 (s, 1H), 5.21 - 5.15 (m, 2H), 3.88 (s, 3H), 3.45 (d, J = 6.3 Hz, 2H).

Methyl 4-acetoxy-3-allylbenzoate (6). Acetic anhydride (200 µL, 218 mg, 2.13 mmol) was added dropwise to a solution of phenol 5 (315 mg, 1.64 mmol) in pyridine (1.5 mL) at room temperature. The mixture was stirred for 14 hours before the solvent was removed. The residue was purified by chromatography (10:1, hexanes:EtOAc) to afford 6 (337 mg, 88%) as a colorless oil: ¹H NMR (CDCl₃, 400 MHz)
MHz) δ 7.95 - 7.90 (m, 2H), 7.11 (d, J = 8.2 Hz, IH), 5.92 - 5.83 (m, IH), 5.12 - 5.03 (m, 2H), 3.88 (s, 3H), 3.33 (d, J = 6.5 Hz, 2H), 2.29 (s, 3H); 13C NMR (CDCl₃, 100 MHz) δ 169.2, 166.8, 153.0, 135.6, 132.7, 132.4, 129.4, 128.4, 123.0, 117.2, 52.6, 35.0, 21.3; IR (neat) νₘₘₚ 3080, 3005, 2980, 2953, 2845, 1765, 1722, 1639, 1609, 1589, 1493, 1437, 1369, 1285, 1263, 1190, 1163, 1121 cm⁻¹; HRMS (ESI+) m/z 235.1076 (M + H⁺, C₃H₅O₄ requires m/z 235.0970).

[0196] Methyl 4-acetoxy-3-(3-methylbut-2-enyl)benzoate (7). Grubbs’ second generation catalyst (11 mg, 0.0130 mmol, 1 mol%) was added to a solution of acetate 6 (305 mg, 1.30 mmol) in a 1/10 solution of DCM/2-methyl-2-buten (5.5 mL). The mixture was stirred 14 hours and was concentrated. The residue was purified by chromatography (10:1, hexanes:EtOAc) to afford 7 (223 mg, 51%) as a white solid: 1H NMR (CDCl₃, 300 MHz) δ 7.92 (d, J = 1.9 Hz, IH), 7.88 (dd, J = 1.9, 8.4 Hz, IH), 7.07 (d, J = 8.4 Hz, IH), 5.22 (td, J = 1.3, 7.1 Hz, IH), 3.87 (s, 3H), 3.25 (d, J = 7.1 Hz, 2H), 2.24 (s, 3H), 1.72 (s, 3H), 1.68 (s, 3H); 13C NMR (CDCl₃, 100 MHz) δ 169.2, 166.9, 153.0, 134.3, 134.1, 132.3, 129.0, 128.3, 122.9, 121.4, 52.5, 29.2, 26.1, 21.2, 18.2; IR (neat) νₘₘₚ 3070, 3005, 2980, 2953, 2845, 1765, 1722, 1639, 1609, 1589, 1493, 1437, 1369, 1285, 1263, 1204, 1192, 1165, 1111 cm⁻¹; HRMS (ESI+) m/z 263.1296 (M + H⁺, C₃H₅O₄ requires m/z 263.1283).

[0197] 4-Hydroxy-3-(3-methylbut-2-enyl)benzoic acid (8). Lithium hydroxide (85 mg, 2.02 mmol) was added to a mixture of methyl ester 7 (106 mg, 0.405 mmol) in 0.5 mL of a 3/1/1 THFZMeOH/Z₂O solution. The reaction mixture was stirred at reflux for 14 hours, cooled to room temperature, and diluted with THF (1 mL). The solution was acidified the solution to pH = 3 by the dropwise addition of 6 M HCl. The layers were separated and the organic phase was dried (Na₂SO₄), filtered, and concentrated to afford acid 8 (63 mg, 75%) as a red oil that was suitable for use without further purification.

[0198] 4-Acetoxy-3-(3-methylbut-2-enyl)benzoic acid (9). Acetic anhydride (1 mL) was added dropwise to a solution of acid 8 (178 mg, 2.00 mmol) in pyridine (3 mL) at room temperature. After stirring for 48 hours, the mixture was poured into water (6 mL) and acidified to pH = 2 by the dropwise addition of 6 M HCl. The suspension was extracted with EtOAc (2 x 10 mL), and the combined organic fractions were dried (Na₂SO₄), filtered and concentrated. The residue was purified by chromatography (5:1, hexanes:EtOAc) to afford acid 9 (223 mg, 51%) as a white solid: 1H NMR (CDCl₃,
400 MHz) δ 8.01 - 7.96 (m, 2H), 7.15 (dd, J = 3.5, 8.2 Hz, IH), 5.24 (tt, J = 1.3, 7.2 Hz, IH), 3.30 (d, J = 7.2 Hz, 2H), 2.34 (s, 3H), 1.97 (s, 3H), 1.76 (s, 3H).

Benzy1 7-hydroxy-8-methyl-2-oxo-2 H-chromen-3-yl carbamate (13).

2-Methyl rescorcinol (1.20 g, 9.71 mmol) was added to a solution of vinyl carbamate 12 (2.7 g, 9.71 mmol) in acetic acid (50 mL). The mixture was stirred at reflux for 48 hours, cooled to room temperature, and filtered. The solid was recrystallized from methanol and H2O to afford 13 (1.30 g, 41%) as a yellow solid: 1H NMR (DMSO, 400 MHz) δ 10.30 (s, IH), 9.10 (s, IH), 8.12 (s, IH), 7.46 - 7.30 (m, 6H), 6.85 (d, J = 8.4 Hz, IH), 5.17 (s, 2H), 2.16 (s, 3H).

Benzy1-7-((3ai?, AR, IR, 7ai?)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3aH-[1.3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl carbamate (15).

Boron trifluoride etherate (61 µL, 69 mg, 0.49 mmol, 30 mol%) was added dropwise to a solution of (3aiR, AS, IR, 7ai?)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3aH-[1.3]dioxolo[4,5-c]pyran-4-y] 2,2,2-trichloroacetimidate (14, 588 mg, 1.62 mmol) and benzy1 7-hydroxy-8-methyl-2-oxo-2 H-chromen-3-yl carbamate (13, 527 mg, 1.62 mmol) in DCM (16 mL). After the mixture stirred for 14 hours, three drops OfEt3N were added and the mixture concentrated. The residue was purified by chromatography (DCM → 100:1, CH2Cl2:acetone) to afford 15 (670 mg, 81%) as a yellow foam: [α]22° = -19.7° (c = 1.54, 20% MeOH in DCM); 1H NMR (CDCl3, 400 MHz) δ 8.27 (s, IH), 7.85 (s, IH), 7.55 - 7.35 (m, 5H), 7.29 (d, J = 2.9 Hz, IH), 7.11 (d, J = 8.7 Hz, IH), 5.77 (d, J = 1.9 Hz, IH), 5.23 (s, 2H), 5.05 (d, J = 1.9 Hz, IH), 4.95 (t, J = 7.7 Hz, IH), 3.59 (s, 3H), 3.30 (d, J = 7.6 Hz, IH), 2.27 (s, 3H), 1.34 (s, 3H), 1.19 (s, 3H); 13C NMR (CDCl3, 100 MHz) δ 159.0, 155.2, 153.6, 153.6, 149.2, 136.0, 129.1 (2C), 129.0, 128.7 (2C), 125.8, 122.6, 122.1, 115.2, 115.1, 111.6, 94.8, 83.3, 78.4, 77.6, 77.0, 67.9, 61.0, 27.9, 22.6, 8.8; IR (film) νmax 3402, 3319, 3063, 3034, 2984, 2939, 2839, 1817, 1709, 1634, 1609, 1587, 1522, 1456, 1383, 1366, 1331, 1296, 1263, 1229, 1205, 1175 cm⁻¹; HRMS (ESI+) m/z 526.1688 (M + H⁺, C27H28NO14 requires m/z 526.1713).

3-Amino-7-((3ai?, AR, IR, 7ai?)-7-methoxy-6,6-dimethyl-2-oxo oxotetrahydrod3aH-[1.3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2 H-chromen-2-one (16).

Palladium on carbon (10%, 67 mg) was added to a solution of carbamate 15 (670 mg, 1.31 mmol) in THF (13 mL). The suspension stirred for 6 hours under a hydrogen atmosphere and was filtered through a plug of silica gel. The solvent was removed and the residue purified by chromatography (100:1 → 50:1, CH2Cl2:acetone) to
afford 16 (425 mg, 83%) as a pale yellow foam: [α]23 D = -26.4° (c = 0.780, 20% MeOH in DCM); 1H NMR (CDCl3, 400 MHz) δ 7.10 (d, J = 8.6 Hz, IH), 7.05 (d, J = 8.6 Hz, IH), 6.68 (s, IH), 5.73 (d, J = 2.0 Hz, IH), 5.04 (dd, J = 2.0, 7.9 Hz, IH), 4.95 (t, J = 7.7 Hz, IH), 4.11 (s, 2H), 3.54 (s, 3H), 3.29 (d, J = 7.6 Hz, IH), 2.28 (s, 3H), 1.34 (s, 3H), 1.21 (s, 3H); 13C NMR (CDCl3, 200 MHz) δ 159.6, 153.3, 153.0, 148.1, 130.2, 122.7, 116.1, 114.8, 111.9, 111.0, 94.5, 83.0, 78.0, 77.3, 76.4, 60.6, 27.5, 22.2, 8.6; IR (film) υmax 3462, 3362, 2984, 2937, 2839, 1807, 1707, 1636, 1595, 1497, 1387, 1371, 1331, 1263, 1169, 1109, 1078, 1036 cm⁻¹; HRMS (ESI+) m/z 392.1357 (M + H+, C9H22N2O8 requires m/z 392.1346).

[0202] 4-((7-((3ai?, AR, IR, 7ai?)-7-Methoxy-6,6-dimethyl-2-oxotetrahydro-3a H-[1.3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (17). Oxaly chloride (15 mg, 119 µmol) was added to a solution of benzoic acid 9 (28 mg, 113 µmol) in CH2Cl2 (0.5 mL), followed by the addition of catalytic DMF. After stirring for 2.5 hours, the acid chloride (10) was concentrated. The yellow solid was redissolved in CH2Cl2 (0.5 mL) and added dropwise over three minutes to a stirred solution of aniline 16 (34 mg, 87 µmol) in pyridine (0.5 mL) at 0°C. The resulting solution was stirred at room temperature for 3.5 hours and concentrated. The residue was purified by preparative TLC (SiO2, 40:1, CH2Cl2:acetone) to afford 17 (31 mg, 57%) as a colorless solid: [α]22 D = -21.7° (c = 0.840, 20% MeOH in CH2Cl2); 1H NMR (CDCl3, 500 MHz) δ 8.72 (s, IH), 8.64 (s, IH), 7.73 (d, J = 2.5 Hz, IH), 7.69 (dd, J = 2.5, 8.0 Hz, IH), 7.29 (d, J = 6.8 Hz, IH) 7.11 (d, J = 8.0 Hz, IH), 7.07 (d, J = 9.0 Hz, IH), 5.72 (d, J = 2.0 Hz, IH), 5.18 - 5.14 (m, IH), 4.99 (dd, J = 1.5, 7.5 Hz, IH), 4.89 (t, J = 8.0 Hz, IH), 3.53 (s, 3H), 3.26 - 3.32 (m, 3H), 2.27 (s, 3H), 2.23 (s, 3H), 1.70 (s, 3H), 1.66 (s, 3H), 1.29 (s, 3H), 1.13 (s, 3H); 13C NMR (CDCl3, 125 MHz) δ 167.9, 164.4, 158.1, 154.1, 152.2, 151.1, 148.1, 133.6, 133.3, 131.0, 128.6, 124.9, 123.1, 122.0 (2C), 121.2, 119.6, 113.8, 113.7, 110.2, 93.3, 81.9, 76.9, 76.3, 75.6, 59.5, 27.8, 26.5, 24.7, 21.1, 19.9, 16.9, 7.4; IR (film) υmax 3400, 2982, 2935, 2856, 1811, 1763, 1715, 1674, 1634, 1607, 1526, 1489, 1437, 1369, 1250, 1202, 1175, 1111, 1090 cm⁻¹; HRMS (ESI+) m/z 622.2277 (M + H+, C33H36NOn requires m/z 622.2289).

[0203] (3R, AS, 5R, 6i?)-5-Hydroxy-6-(3-(4-hydroxy-3-(3-methylbut-2-enyl)benzylamino)-8-methyl-2-oxo-2 H-chromen-7-yloxy)-3-methoxy-2,2-dimethyltetrahydro-2 H-pyran-4-yl carbamate (DHN1). A solution of carbonate 17
(32 mg, 52 µmol) in 7 M methanolic ammonia (2 mL) was stirred for 14 hours. The solvent was removed and the residue purified by preparative TLC (SiO₂, 25:1, CH₂Cl₂:methanol, developed 7 times) to afford DHN2 (2.5 mg, 9%) and 4-deshydroxynovobiocin (DHN1, 17.5 mg, 57%) as colorless solids; DHN1: [α]D = -20.3° (c = 0.300, 10% MeOH in CH₂Cl₂); 1H NMR (CDCl₃, 400 MHz) δ 8.70 (s, IH), 7.62 (d, J = 2.3 Hz, IH), 7.56 (dd, J = 2.3, 8.4 Hz, IH), 7.29 (d, J = 8.8 Hz, IH), 7.13 (d, J = 8.8 Hz, IH), 6.81 (d, J = 8.4 Hz, IH), 5.50 (d, J = 2.3 Hz, IH), 5.34 - 5.25 (m, 2H), 4.25 (t, J = 2.6 Hz, IH), 3.53 - 3.51 (m, IH), 3.50 (s, 3H), 3.34 (dd, J = 3.2, 8.9 Hz, 2H), 2.99 (s, IH), 2.94 (s, IH), 2.27 (s, 3H), 1.74 (s, 3H), 1.71 (s, 3H), 1.33 (s, 3H), 1.13 (s, 3H); 13C NMR (CDCl₃, 125 MHz) δ 166.3, 159.6, 158.9, 156.8, 155.9, 149.0, 133.9, 129.0, 128.6, 126.4, 125.6, 124.6, 124.1, 121.8, 121.4, 114.9, 114.4, 114.1, 111.2, 98.3, 81.4, 78.9, 72.2, 69.6, 61.5, 29.7, 29.3, 27.2, 22.6, 17.9, 8.2; IR (film) ν max 3400, 3379, 3360, 2978, 2928, 2853, 1709, 1659, 1632, 1605, 1528, 1504, 1367, 1254, 1136, 1117, 1086 cm⁻¹; HRMS (ESI+) m/z 597.2434 (M + H⁺, C₃H₇N₂O₃ requires m/z 597.2448). The structure of DHN1 is:

![Structure of DHN1]

[0204] N-(1-((2R, 3R, 5i)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yl)-oxy)-8-methyl-2-oxo-2 H-chromen-3-yl)4-hydroxy-3-(3-methylbut-2-enyl)benzylamide (DHN2). A solution of carbonate 17 (12 mg, 19.3 µmol) in 10/1 methanol/Et₂N (220 µL) was stirred for 14 hours. The solvent was removed and the residue purified by preparative TLC (SiO₂, 10:1, CH₂Cl₂:methanol) to afford DHN2 (8 mg, 75%) as a colorless solid: [α]D = -12.9° (c = 0.310, 10% MeOH in DCM); 1H NMR (CDCl₃, 400 MHz) δ 8.78 (s, IH), 8.66 (s, IH), 7.71 (d, J = 2.2 Hz, IH), 7.68 (dd, J = 2.2, 8.3 Hz, IH), 7.33 (d, J = 8.8 Hz, IH), 7.19 (d, J = 8.8 Hz, IH), 6.90 (d, J = 8.3 Hz, IH), 6.05 (s, IH), 5.61 (d, J = 1.6 Hz, IH), 5.33 (t, J = 7.1 Hz, IH), 4.27 - 4.23 (m, 2H), 3.61 (s, 3H), 3.45 - 3.35 (m, 3H), 2.77 (s, IH), 2.67 (s, IH), 2.05 (s, 3H), 1.80 (s, 3H), 1.79 (s, 3H), 1.38 (s, 3H), 1.14 (s, 3H); 13C NMR (CDCl₃, 125 MHz) δ 165.9,
159.5, 158.3, 155.9, 149.0, 135.8, 129.5, 127.5, 126.9, 125.9, 125.8, 124.2, 122.0, 120.9, 125.9, 124.2, 115.9, 114.2, 114.1, 111.2, 97.7, 84.3, 78.6, 71.2, 68.6, 62.0, 29.6, 29.3, 25.8, 22.5, 18.0, 8.2; IR (film) \( \nu_{\text{max}} \) 3402, 2974, 2928, 2854, 1717, 1701, 1645, 1605, 1526, 1506, 1367, 1254, 1088 cm\(^{-1}\); HRMS (ESI+) \( m/z \) 554.2363 (M + H\(^+\), C\(_{30}\)H\(_{36}\)NO\(_9\) requires \( m/z \) 554.2390). The structure of DNH2 is:

![Structure of DNH2](image)

**EXAMPLE 13: PREPARATION OF DIMERS**

[0205] It is contemplated that the C-terminal nucleotide binding sites are in close proximity to the one another along the Hsp90 dimer interface, and therefore dimeric inhibitors of the compounds of the present disclosure should provide compounds with enhanced inhibitory activity. This is based on the fact that the dimeric compound, coumermycin A1, was shown to be approximately 10 times more active than the monomeric compound, novobiocin.

[0206] The present disclosure thus includes dimers of the compounds disclosed herein. In one aspect, a dimeric inhibitor of KU-1/A4 can be prepared. As set forth in the scheme below, the Cbz group can be removed to furnish the aniline for subsequent coupling with bifunctional linkers to prepare dimeric inhibitors. The dimer containing pyrazole linker found in Coumermycin A1 can be prepared following the procedure developed by Olson et al., Tetrahedron Letters, Volume Date (2003) 44(1), 61-63 (2002). The diacid can be coupled with two equivalents of the coumarin amine using O-(7-azabenzotriazol-1-yl)-\( \Lambda,\Lambda,\Lambda',\Lambda'\)-tetramethyluronium hexafluorophosphate ("HATU") to furnish the cyclic carbonate precursor to the KU-1/A4 dimer. The carbonate can be removed upon treatment with methanolic triethylamine to provide the tetraol product. See Yu et al., *Hsp90 Inhibitors Identified from a Library of Novobiocin Analogues*, *J. Am. Chem. Soc.* 127 12778-12779 (2005). Similar to this method, a number of dimeric linkers can be used to perturb the dimeric angle and to extend the dimeric tether in an effort to elucidate structure-activity relationships. As such, ortho, meta, and para dibenzoic acids can be used in lieu of the pyrole biscarboxylic acid to determine optimal
angles. Linker length can be probed by the use of about 3-10 carbon dicarboxylic acids. If the studies support that both angle and linker length are important, then combinations of these linkers can be prepared and coupled to furnish the conformationally biased, extended compounds such as that shown below.

**EXAMPLE 14: PROSTATE CANCER XENOGRAFT TUMOR MODEL**

This example involves the *in vivo* effect of the compounds of the present disclosure using a prostate cancer mouse model. More specifically, four to six week old BALB/c nu/nu nude mice can be obtained commercially and maintained in ventilated cages under Institutional Animal Care and Use Committee approval. Separate male mice can be inoculated subcutaneously with 10^6 LNCaP cells suspended in 0.25 mL of Matrigel (BD, Bioscience, Bedford MA). Stable serum testosterone levels can be maintained in the mice by the implantation of 12.5 mg 90-day sustained release testosterone pellets (Innovative Research, Sarasota FL) subcutaneously prior to inoculation with tumor. Tumor volume can be measured twice a week with vernier calipers with tumor volumes calculated using the formula \[ \text{length} \times \text{width} \times \text{height} \times 0.52 \]. Mice with established tumor volumes of 5 mm can be selected for KU-1/A4
administration. Utilizing the paradigm for administration of 17-AAG (another Hsp90 inhibitor), animals can be treated with both continuous and intermittent dosing schedules. A control animal can be treated with vehicle alone (DMSO). For the continuous dosing schedule, mice can receive intraperitoneal injections of vehicle or the test compounds (e.g., KU-1/A4) for 5 days per week for 3 weeks. The intermittent group can receive one 5 day cycle and then monitored for progression.

Differing doses of the test compound (e.g., KU-1/A4) can be utilized based on pharmacokinetic information obtained from toxicity studies. When progression occurs, as defined by an increase in tumor size, the mice can receive a second 5 day cycle of the test compound (e.g., KU-1/A4). Response to the test compound can be assessed by measuring tumor volume and serum PSA levels using the PSA Assay Kit (American Qualex Antibodies, San Clemente CA). Further response can be assessed by harvesting the tumor at euthanasia and performing immunohistochemistry and western blot analysis of the Hsp90's client proteins known to be involved in cancer cell survival mechanisms such as signal transduction (e.g., AKT, Her2, PKinase), angiogenesis (e.g., HIF-1α), and metastasis (AR, MMP2). Each dose and control can be repeated three times to confirm results.

Statistical analysis can be performed to compare the average tumor volume over time between the different doses of the test compound and the control animals. The null hypothesis which is that KU-1/A4 can cause no change in tumor volume over time can be tested by the squared difference between mean tumor volume summed over all time points. We can use a Wilcoxon sum-rank test to compare PSA levels in the treatment and control group. Immunohistochemistry results can be assessed qualitatively based on staining intensity graded on a scale of 1 to 5.

To investigate toxicity, four to six week old BALB/c nu/nu nude mice can be obtained commercially and maintained in ventilated cages under Institutional Animal Care and Use Committee approval. Intraperitoneal injections of the test compound (e.g., KU-1/A4) can be given to non-tumor bearing mice at ranges of 25 mg/kg to 200 mg/kg five days a week for three weeks based on similar concentrations used for 17AAG. Serum samples can be obtained on days 5, 10, and 15. Serum chemistry and liver function analysis can be performed. Serum concentrations of test compound (e.g., KU-1/A4) can be determined by high performance liquid chromatography (HPLC). At sacrifice by CO₂ euthanasia, a complete blood count, gross necropsy and liver and kidney histopathology can be performed on the animals to determine toxicity. The
maximal tolerated dose can be calculated using up/down toxicity studies that can be used as the upper limit of dose for treatment.

EXAMPLE 15: NEUROPROTECTIVE EFFECTS

[0211] Recently, low concentrations of the Hsp90 inhibitor GDA were reported to induce expression of both Hsp70 and Hsp90, with a concomitant reduction in phosphorylated Tau (Dou et al., (2003)). In this example, KU-1/A4, a novel C-terminal Hsp90 inhibitor, was tested for protective effects against Aβ toxicity in primary neurons. See protocols in Michaelis et al., B-Amyloid-induced neurodegeneration and protection by structurally diverse microtubule-stabilizing agents, J Pharmacol Exp Ther 312 659-668 (2005), which is incorporated by reference.

[0212] As is shown in FIG. 4, concentrations of KU-1/A4 as low as 5 nM protected the neurons against Aβ, and the drug alone produced no toxicity. GDA partially protected the neurons against Aβ, but the drug alone was toxic to the neurons at concentrations above 20 nM. Thus, although GDA can increase Hsp90 levels, the result may be the degradation of client proteins essential for neuronal survival. This lack of KUI toxicity in both proliferating and post-mitotic cells suggested that further exploration of its mechanism(s) of action is warranted.

EXAMPLE 16: NEUROPROTECTIVE EFFECTS OF KU-1/A4 AND KU-32

[0213] Treatment of brain and neuronal cell cultures with Aβ25-35 produces distinct morphological changes and eventual cell death (Pike et al., Structure-Activity Analyses of β-Amyloid Peptides: Contributions of the β25-35 Region to Aggregation and Neurotoxicity, J. Neurochem., 64 253-265 (1995)). Pretreatment with neuroprotective agents can reduce or abolish these effects. In this example, the neuroprotective effects of KU-1/A4 were determined in primary neurons derived from embryonic rat brain exposed to Aβ (10 μM) in the presence or absence of the test compound for 48 hours. The percentage of surviving neurons was determined by labeling with the fluorescent dyes calcein-AM and propidium iodide as previously described. See Michaelis et al., β-Amyloid-Induced Neurodegeneration and Protection by Structurally Diverse Microtubule-Stabilizing Agents, J. Pharmacol. Exp. Ther. 312 659-668 (2005); Michaelis et al., Protection Against β-Amyloid Toxicity in Primary Neurons by Paclitaxel (Taxol), J. Neurochem. 70 1623-1627 (1998). The numbers of calcein-labeled live cells and
propidium iodide-labeled dead neurons in several fields were visualized via fluorescence microscopy and counted as described.

[0214] More specifically, primary cortical neurons were recovered from embryonic day 18 rat brains as described previously (Michaelis et al., Immunological Localization And Kinetic Characterization Of A Na+/Ca2+ Exchanger In Neuronal And Nonneuronal Cells, Brain Res. 661 104-1 16 (1994)). Briefly, about 10 to 16 brains were removed from fetuses delivered by cesarean section, and the cortices dissected out. The neurons were isolated and suspended in DMEM/F12 supplemented with 10% FBS and plated on sterile dishes or cover slips coated with 10 μg/ml poly-D-lysine and 5 μg/ml mouse laminin. After 24 hours, the serum-containing medium was removed, and neurons were maintained in Neurobasal medium (Gibco) with 2% B-27 supplements (Gibco). Cells were maintained in culture at 37 °C in 5% CO2 and 97% humidity for 7 to 8 eight days in vitro (DIV) before use. Neurons were exposed to 0.004% DMSO or the indicated concentrations of GA or KU-1/A4 for two hours before the addition of 10 μM Aβ25-35 for either 24 or 48 hours as indicated. Stock solutions of Aβ were prepared in sterile water at a concentration of 1.3 mM and stored at -20 °C. Aliquots of the stock solutions were diluted to 1 mM in sterile 50 mM Tris-HCl, pH 7.4, and incubated at 37 °C for 24 hours to promote oligomerization of the peptides prior to treatment of the neurons. The effects of KU-1/A4, GA, Aβ, or the indicated combinations on neuronal viability were determined by monitoring cell survival using the Live-Dead assay as previously described. Following drug treatment in the presence or absence of Aβ peptides for either 24 or 48 hours as indicated, neurons were labeled with 20 μM propidium iodide and 150 nM calcein acetoxy-methylester and placed on a fluorescent microscope stage. Digital images from 6 fields per dish were captured, and the number of viable (green) and dead (red) cells counted. All experiments were conducted with duplicate dishes from at least two neuronal preparations for each treatment, with about 1,000 cells analyzed/treatment condition. The data are expressed as the fraction of viable cells calculated from the total number of neurons counted under each treatment condition. The significance of differences between cultures exposed to various treatment conditions was determined using Student's t-test.

[0215] In the studies, treatment of primary cortical neurons with Aβ alone (10 μM) represented the basal level for neuronal survival. Pretreatment of neuronal cells with KU-1/A4 prevented Aβ-induced toxicity in a dose-dependent fashion, with an EC50...
value of about 6 nM (FIG. 5). While there were minor neuroprotective effects associated with KU-1/A4 concentrations as low as 0.5 nM, significant protection was not demonstrated until 5 nM. Treatment of neuronal cells with KU-1/A4 alone at 2OX the EC50 value (100 nM) did not result in any observed neurotoxicity.

In a similar study, the pretreatment of neuronal cells with the 8-methyyl derivative of KU-1/A4 (denominated KU-32) also prevented Aβ-induced toxicity in a dose-dependent fashion, but was more potent. The EC50 value of KU-32 was about 0.9 nM (data not shown).

EXAMPLE 17: KU-1/A4 UP-REGULATES HSP70 IN NEURONAL CELLS.

Inhibition of Hsp90 induces overexpression of both Hsp90 and Hsp70 through dissociation of Hsp90-HSF-1 complexes and subsequent translocation of HSF-1 to the nucleus. Induction of both Hsp90 and Hsp70 by GA in cultured cells was reported to result in decreased levels of aggregated tau and increased levels of soluble tau, indicating that Hsp90 inhibitors can reduce toxic tau aggregates. Thus, in this example, an immunoblot analysis of neuronal cells was performed for Hsp70.

Primary cortical neurons were prepared as described herein. Cells were incubated (37 °C in 5% CO2) with the indicated concentrations of GA or KU-1/A4 for 24 or 48 hours. Cells were washed three times with phosphate buffered saline (PBS), followed by addition of a cell lysis buffer (50 mM Tris-HCL, 150 mM NaCl, 5 mM EDTA, 5 mM EGTA, 1% NP40, 10% Glycerol, 1 mM Na3VO4, 10 mM sodium molybdate, 40 mM NaF, and 10 ul/ml of Calbiochem protease inhibitor cocktail III). Cells were collected by scraping the dishes and aspirating all contents. The protein concentration of each sample was determined using a bicinechonic acid assay from Pierce Biotechnology, Inc. Aliquots containing 25 μg total protein in reducing sample buffer (50 mM Tris-HCl pH 6.8, 6.7% glycerol, 2.7% SDS, 0.05% Bromphenol Blue), resolved by SDS-PAGE, and transferred to PVDF membranes as previously described (Michaelis et al., Effects of reactive oxygen species on brain synaptic plasma membrane Ca(2+)-ATPase, Free Radic. Biol. Med. 27 810-821 (1999)). The membranes were probed with the indicated primary HSP70 antibody (1:500, Upstate Biotechnology) and
immunoreactive proteins detected using the Lumiglo Western Blot Protein Detector Kit (KPL). The immunoblots were scanned using a Kodak Image Station 2000R to determine the pixel density of the bands and imported into Photoshop and analyzed as described in Zaidi et al., Oxidative inactivation of purified plasma membrane Ca2+-ATPase by hydrogen peroxide and protection by calmodulin, Biochem. 42 12001-12010 (2003). All blots were re-probed with antibodies to β-actin I-19R (Santa Cruz) as a loading control and the data are presented as the ratio of Hsp70 to actin in each lane. Cells from at least three separate cultures for each treatment condition were examined, and the significance of differences in the immunoreactivities of the bands observed under the various treatment conditions was assessed by Student's t-test for unpaired samples.

As shown in FIGs. 6A and 6B, in the neuronal cultures of the present disclosure, KU-1/A4 significantly increased Hsp70 levels at a concentration of 0.2 µM. However, concentrations of KU-1/A4 as low as 1 nM also led to increases in Hsp70 levels, compared to DMSO controls after 48 hours incubation. Hsp70 induction at 0.2 µM was comparable to that seen upon treatment with GA at the same concentration, suggesting KU-1/A4 could potentially attenuate tau aggregation in a manner similar to that reported for GA. In addition, incubation with higher concentrations of KU-1/A4 (1 and 10 µM) did not significantly increase Hsp70 levels, suggesting that the maximal neuroprotective effects of KU-1/A4 can be elicited at concentrations significantly lower than any potential cytotoxic effects.

EXAMPLE 18: ANTIPROLIFERATIVE EFFECTS OF KU-1/A4.

For most Hsp90 inhibitors, efficacy in degrading client proteins, such as Her2 and AKT, in two distinct cancer cell lines, SkBr3 and MCF-7, correlates well with their anti-proliferative effects (Dai et al., HSP90: a rising star on the horizon of anticancer targets, Future Oncol. 1, 529-540 (2005)). The anti-proliferative effect of GA treatment in these cell lines is well established, and almost complete cytotoxicity with GA at 1 µM was confirmed. GA elicited dose-dependent anti-proliferation in both cell lines, and the IC50 values were comparable to those reported previously (18 and 133 nM, respectively). In contrast, KU-1/A4 demonstrated no anti-proliferative effects in either cell line up to 100 µM (FIG. 7A and 7B), a concentration well above that necessary for complete neuroprotection, suggesting that C-terminal inhibitors possess a mechanism of action distinct from GA and other inhibitors of the N-terminus. When the effects of GA and
KU-1/A4 alone were examined in neuronal cells, GA induced significant cytotoxicity at
10 μM after 24 hours (FIG. 7C). Lower concentrations of GA led to substantial
cytotoxicity after 72 hours of incubation (data not shown). In contrast, 10 μM of KU-
1/A4 caused no toxic effects even after incubation for 72 hours, clearly indicating a
novel utility for C-terminal inhibitors.

EXAMPLE 19: TRANSPORT OF KU-1/A4 ACROSS THE BLOOD-BRAIN
BARRIER ("BBB") VIA RHODAMINE ASSAY

[0221] The BBB expresses high levels of P-glycoprotein (P-gp), an efflux pump
responsible for the extrusion of numerous drugs and other xenobiotics from cells. The
rhodamine 123 assay is often used to predict whether a compound is a potential substrate
for P-gp. In this assay, rhodamine 123 is used as a surrogate P-gp substrate. If a test
compound (KU-1/A4) is a substrate for P-gp, then its addition can increase rhodamine
123 uptake relative to the negative control determined by monitoring intracellular
fluorescence. Taxol (paclitaxel), a microtubule-stabilizing agent that exhibits
neuroprotective effects both in vitro and in vivo, is hampered as a CNS therapeutic
because it is a known P-gp substrate. Thus, in this example, a rhodamine 123 uptake
assay was performed as described in Silverstein et al., Utilization of uptake studies for
evaluating activity of efflux transporters, Current Protocols in Pharmacology 7 7.1
(2003).

[0222] Used as a positive control, Taxol significantly increased rhodamine 123
uptake in BMECs, while addition of KU-1/A4 had no effect on uptake even up to 50 μM,
indicating it is not a substrate for P-gp (FIG. 8A).

EXAMPLE 20: TRANSPORT OF KU-1/A4 ACROSS THE BLOOD-BRAIN
BARRIER VIA BMEC TRANSPORT IN VITRO

[0223] The ability to partition across the blood-brain barrier is an essential property
of drugs that are designed to elicit their effects on neuronal cells of the central nervous
system (CNS). Transport across primary cultures of BMECs in vitro provides a strong
correlation to the BBB permeability of a compound in vivo.

[0224] In this example, BMECs were grown on 0.4 μm polycarbonate membranes in
a petri dish coated with rat tail collagen and fibronectin. Once cells had formed a
confluent monolayer as determined by light microscopy, the membranes were transferred

Briefly, each chamber was filled with 3 mL of PBSA and the donor chamber included KU-1/A4 (10 μM). A temperature of 37 °C was maintained with an external circulating water bath and chamber contents were stirred with Teflon coated magnetic stir bars driven by an external console. At various time points (5, 15, 30, 45, and 90 minutes), 100 μL aliquots were removed from the receiver side and replaced with 100 μL of blank PBSA warmed to 37 °C. Samples of the donor solution were also taken for analysis. All samples were analyzed for concentration by RP-HPLC analysis using a Shimadzu dual pump HPLC system equipped with an Alltech (C18, 4.6 mm x 150 mm) column. The solvent system consisted of H₂O (solvent A) and methanol (solvent B). Analytical samples were eluted with 20% solvent B over 10 minutes at a flow rate of 0.90 mL/min and UV detection at 208 nm. The concentration of KU-1/A4 in each sample was determined by direct comparison of the area under the curve ("AUC") for each injection with the average AUC (five injections) of a standard KU-1/A4 solution (10 μM). The integrity of the cell monolayer was tested post experiment by monitoring the permeability of [¹⁴C]-sucrose, a low permeability marker. Radioactive samples were analyzed by liquid scintillation counting.

[0225] Not only was KU-1/A4 not a P-gp substrate, it also exhibited time-dependent linear transport across BMECs for up to 90 minutes (FIG. 8B). The concentration of KU-1/A4 at 90 minutes in the receiving chamber (1.2 μM) was 200-fold greater than the concentration necessary (5 nM) for 50% neuroprotection from Aβ-induced toxicity. These data suggest that pharmacologically active amounts of KU-1/A4 should penetrate the BBB and avoid efflux via P-gp.

[0226] In sum, KU-1/A4 induces Hsp70 and provides complete protection against Aβ-induced toxicity at non-cytotoxic concentrations. In fact, no toxicity was observed in the assays even at 20,000 X the EC₅₀ (100 μM) in non-neuronal cells, a concentration at which GA is severely toxic. In addition, KU-1/A4 increases Hsp90 levels at concentrations about 200-fold less than those required for client protein degradation. This provides a large therapeutic window for treatment of several disorders in which chaperones provide a protective effect. These attributes make KU-1/A4, and related
compounds, an ideal compound for development as a novel chemotherapeutic for the treatment of AD and other neuron degenerative disorders. Further, the time-dependent linear transport of KU-1/A4 across BMECs in vitro provides strong evidence that significant concentrations of the drug can be available to the CNS in vivo. Within 15 minutes, the concentration of KU-1/A4 present in the abluminal chamber was enough to provide full neuroprotective effects as demonstrated in vitro, suggesting that in vivo neuroprotection could be fast-acting. KU-1/A4 is also not a substrate for P-gp and is not expected to be removed by active efflux from the CNS.

**EXAMPLE 21: MULTIPLE SCLEROSIS**

[0227] The neurodegenerative disorder multiple sclerosis is often studied in an animal model system termed experimental autoimmune encephalomyelitis ("EAE"). EAE is an inflammatory condition characterized by multifocal perivascular CNS inflammatory infiltrates that primarily include T cells and monocytes. Bar-Or et al, *Molecular pathogenesis of multiple sclerosis*, Journal of Neuroimmunol. 100 252-259 (1999). EAE can be induced in animals by injection of immunodominant peptides from myelin proteins such as myelin basic protein (MBP), proteolipid protein (PLP), and myelin oligodendrocyte glycoprotein (MOG), or by transfer of CD4+ MHC class II-restricted T-cells reactive with these peptides. See Mokhtarian et al., *Nature* 309 312-314 (1984); Zamvil et al., *T-cell clones specific for myelin basic protein induce chronic relapsing paralysis and demyelination*, Nature 317:355-358 (1985). The EAE models are frequently used to study the pathogenesis of MS and to test novel therapeutic strategies aimed at treating MS.

[0228] Heat shock response suppresses inflammatory gene expression for nitric oxide synthase, cytokines and chemokines, all of which have been implicated in the development of multiple sclerosis (MS). HSR can be induced by a variety of stresses, including hyperthermia, oxidative stress, heavy metals, viral infection, and UV irradiation. Administration of Hsp90 inhibitors also leads to a HSR due to the dissociation of HSF-I from Hsp90. Therefore, KU-32, an 8-methyl derivative of KU-I, a novel Hsp90 inhibitor was evaluated in a murine autoimmune disease model (Experimental Autoimmune Encephalomyelitis; EAE) to determine if disease severity and disease incidence is diminished.
To evaluate the in vivo activity, SJL/J female mice at 6 to 8 weeks of age were divided into three groups with ten mice in each group. The first group was the negative control. For the KU-I treated and positive control groups, at the initiation state (Day 0), all mice (10/group) were immunized by intradermal ("i.d.") inoculation with 200 µg of proteolipid protein ("PLP") in a 0.2 mL emulsion with equal volumes of phosphate buffered saline ("PBS") and complete Freund's adjuvant ("CFA") (Difco, Detroit, MI). The injection volume was 100 µL at each injection site. Then, each mouse in KU-32 treated group received intravenous ("i.v.") injections of 0.5 mg/kg of KU-32 on day 0 and on day 10.

Disease progression was evaluated using a clinical scoring scale ranging from 0 to 5 as shown in the Table below:

<table>
<thead>
<tr>
<th>Score</th>
<th>Gross Pathology</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>No clinical disease</td>
</tr>
<tr>
<td>0.5</td>
<td>Tail weakness</td>
</tr>
<tr>
<td>1</td>
<td>Tail completely flaccid</td>
</tr>
<tr>
<td>2</td>
<td>Paraparesis (weakness, incomplete paralysis of one or two hind limbs)</td>
</tr>
<tr>
<td>3</td>
<td>Paraplegia (complete paralysis of two hind limbs)</td>
</tr>
<tr>
<td>4</td>
<td>Paraplegia with forelimb weakness or paralysis</td>
</tr>
<tr>
<td>5</td>
<td>Moribund or death</td>
</tr>
</tbody>
</table>

All of the immunized mice were scored blindly for about 7 weeks by the same observer. Mean daily clinical scores were calculated by adding the grades of each mouse individually divided by the number of mice in each group. All animals were observed daily and, upon signs of paralysis and weakness, moistened food was provided to the animals to prevent dehydration.

The results are shown in FIGs. 9 and 10. The results from a pilot studies showed that the disease onset was delayed and disease severity was reduced following administration of KU-32. The negative control group gradually increased in weight as expected.

A second study demonstrated that KU-32 administration given early in the EAE immunization protocol reduced disease relapse, whereas KU-32 administration a week into the immunization protocol reduced disease severity and inhibited disease
relapse. The experiment was the same as the foregoing except that KU-32 was administered on days 1, 3, 5, and 7 for the early treatment regimen showed no change in disease onset or disease severity. FIG. 11 shows the results of this second study where KU-32 for the early treatment regimen showed no change in disease onset or disease severity. In the late treatment regimen, KU-32 was injected i.v. on days 6, 8, 10, 12, and 14. This treatment resulted in an earlier onset of symptoms; however the duration of symptoms was notably less than in the positive control group, and the clinical scores returned to baseline compared to the control group and the early treatment group where the clinical score remained around 0.5. FIG. 12 is a graph showing the corresponding body weight data for the second study described in FIG. 11.

**EXAMPLE 22: HSP90 INHIBITION WITH DHNL AND DHN2**

[0234] Previous studies have demonstrated that novobiocin manifests weak activity against the Hsp90 protein folding process as demonstrated by its ability to induce degradation of ErbB2 in SkBr3 breast cancer cells at about 700 µM concentration. In this example, cells were washed once with cold phosphate-buffered saline (pH 7.0), and lysed by scraping in TMNS (50 mM Tris-HCl, pH7.5, 20 mM Na₂MoO₄, 0.1% NP-40, 150 mM NaCl) supplemented with 20 µg/mL aprotinin, 20 µg/mL leupeptin, and 1 mM phenylmethanesulfonyl fluoride. Cell lysate was clarified by centrifugation at 14,000 rpm at 4 °C for 15 minutes, and protein concentration was determined by using the BCA method (Pierce, Rockford IL). 20 µg total protein from cell lysates were separated by 4-20% gradient SDS-PAGE (Bio-Rad, Hercules CA). Western-blotting for ErbB2 was performed as described previously. Blotting for α-tubulin was used to verify equal loading of lanes.

[0235] As shown in FIG. 13, both DHNL and DHN2 compounds exhibited improved activity as compared to novobiocin. Western blot analyses of Hsp90-dependent client proteins ErbB2 (Her2) and p53 were investigated as well as the related heat shock protein, Hsp70. As can be seen in lane 6, DHNL induced the degradation of both ErbB2 and p53 between 5 and 10 µM (lanes 5 and 6), whereas DHN2 induced the degradation of these clients between 0.1 and 1.0 µM (lanes 9-12), clearly indicating that DHN2 is more effective than DHNL, which itself is about 70 times more active than novobiocin. Levels of actin were unaffected by inhibitor concentration in these immunoblot assays (data not shown).
N-terminal inhibitors of Hsp90 may induce the degradation of client proteins at concentrations that mirror that needed for anti-proliferative activity. Therefore, both DHN1 and DHN2 were evaluated for their ability to inhibit the growth of SKBr3 breast cancer cell lines. Cell growth was monitored using methylthiazol-tetrazolium (MTT). Briefly, cells ($5 \times 10^3$) were plated in 96-well microtiter plates (Costar) in a volume of 0.1 mL DMEM containing 0.1% FBS. After 12 hours, cells were exposed to drugs (final volume 0.2 mL/well). At various times after drug addition, 20 µL of 5 mg/mL MTT solution in PBS was added to each well for four hours. After removal of medium, 0.1 mL of DMSO was added to each well to dissolve the formazan crystals. Absorbance at 562 nm was determined using an ELx 808 microplate reader (Bio-Tek, Winooski VT). Six wells were assayed at each concentration and the mean absorbance was determined. Absorbance at 562 nm is directly proportional to viable cell number.

As shown in FIG. 14, neither treatment with DHN1 nor DHN2 resulted in substantial cytotoxicity, suggesting that C-terminal inhibitors of Hsp90 exhibit a
mechanism of action that differs from N-terminal inhibitors or perhaps that a different set of client proteins that are responsible for cell growth are selectively targeted by inhibitors of the N-terminus.

**EXAMPLE 23: PREPARATION OF BENZAMIDE NOVOBIOCIN DERIVATIVES**

[0238] This example involved the preparation of novobiocin analogues with highly substituted benzamides. As shown in the scheme below, the derivatives were assembled from three components: noviose carbonate (see Shen et al., *Syntheses of Photolabile Novobiocin Analogues*, Bioorg. Med. Chem. Lett. 14 5903 (2004)), 8-methylcoumarin (Toplak et al., *The synthesis of methyl 2-(benzyloxy carbonyl)amino-3-dimethylaminopropenoate. The synthesis of trisubstituted pyrroles, 3-amino-2H-pyran-2-ones, fused 2H-pyran-2-ones and 4H-pyridin-4-ones*, J. Hetero. Chem. 36 225-235 (1999), and a series of substituted benzoic acids. Previously, it had been demonstrated that the trichloroacetimidate of noviose carbonate couples directly to the coumarin phenol to afford the α-anomer in excellent yield (see Shen et al., *Syntheses of Photolabile Novobiocin Analogues*, Bioorg. Med. Chem. Lett. 14 5903 (2004)). Since no co-crystal structure of Hsp90 bound to C-terminal inhibitors exists, commercially available benzoic acids were chosen that contained various functionalities in an effort to probe for steric and electronic interactions with the putative hydrophobic pocket that is believed to bind to this region of novobiocin. The following shows the retrosynthesis of the novobiocin analogues:
More specifically, novobiocin analogues were prepared by the condensation of \(N,N\)-dimethylformamide dimethyl acetal (2) with Cbz-protected glycine (1) to produce the vinylagous carbamate, 3 as set forth in the scheme below. See Robinson et al., *Highly enantioselective synthesis of alpha,beta-diaminopropanoic acid derivatives using a catalytic asymmetric hydrogenation approach*, J. Org. Chem. 66 4141-4147 (2001). The 8-methylcoumarin 5 was prepared by a modified Pechmann condensation of 2-methylresorcinol (4) with 3. Toplak et al., J. Hetero. Chem. 36 225-235 (1999). The resulting phenol was noviosylated with the trichloroacetimidate of noviose carbonate (6) (Yu et al., *Synthesis of (-)-Noviosefrom 2,3-O-Isopropylidene-D-erythronolactol*, J. Org. Chem., 69 7375 -7378 (2004)) in the presence of catalytic boron trifluoride etherate to give 7 in good yield. See Shen et al., *Synthesis of Photolabile Novobiocin Analogues*, Bioorg. Med. Chem. Lett. 14 5903-5906 (2004). The benzyl carbonate was removed via hydrogenolysis to produce aminocoumarin 7, which proved to be a versatile intermediate throughout this project. The amine was readily coupled to a preselected library of benzoic acids in the presence of \(N\)-(3-dimethylamino-propyl)-\(N''\)-ethylcarbodiimide hydrochloride (EDCI) and 4-DMAP. During the course of the investigation, it was determined that utilization of 4-DMAP led to bisacylation, which proved difficult to separate from the monoacylated product. Therefore pyridine was employed as the base and provided exclusively monoacylated products. With the desired benzamides in hand, the cyclic carbonates underwent solvolysis with triethylamine in methanol to give the
diol in excellent yield. To complete the small library of inhibitors, aryl nitro compounds (15-17) were subjected to hydrogenation to afford the corresponding anilines.

Synthesis of Photolabile Novobiocin Analogues

[0240] General EDCI coupling procedure A: N-(3-Dimethylamino-propyl)- N'-ethylcarbodiimide hydrochloride (3 eq) was added to a solution of aminocoumarin 7 (1 eq), benzoic acid (3 eq) and 4-DMAP (2.0 eq) in CH₂Cl₂ at room temperature. The solution was stirred for 14 hours, concentrated and the residue purified via preparative TLC or column chrotography (SiO₂, 40:1; CH₂Cl₂:acetone) to afford the benzamide.

[0241] 2-Methoxy-N-(7-((3α,4i?,4i?,7i?,7α?)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3 αβH-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)benzamide (9a). Colorless solid (66%): [α]D²⁴ = -29.6° (c = 0.61, 20% MeOH in CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ 8.77 (s, 1H), 8.18 (d, J = 7.8 Hz, 1H), 7.46 (t, J = 7.8 Hz, 1H), 7.27 (d, J = 8.3 Hz, 1H), 7.09 - 7.02 (m, 2H), 6.99 (d, J = 8.3 Hz, 1H),
5.71 (s, IH), 4.99 (d, J = 7.9 Hz, IH), 4.91 (t, J = 7.9 Hz, IH), 4.05 (s, 3H), 3.53 (s, 3H),
3.24 (d, J = 7.9 Hz, IH), 2.23 (s, 3H), 1.29 (s, 3H), 1.14 (s, 3H); $^{13}$C NMR (100 MHz, CDCl$_3$) $\delta$ 164.5, 159.6, 158.1, 155.3, 153.6, 149.5, 134.3, 132.6, 126.4, 124.4, 123.5, 121.9, 121.3, 115.5, 115.1, 112.0, 111.5, 94.8, 83.3, 78.4, 77.4, 77.1, 61.0, 56.6, 27.9,
22.6, 8.8; IR (film) $\nu_{\text{max}}$ 3308, 3055, 2986, 2939, 2930, 1817, 1807, 1707, 1707, 1655, 1603,
1533, 1481, 1466, 1367, 1263, 1232 cm$^{-1}$; HRMS (ESI+) $m/z$ 526.1691 (M + H$^+$, C$_{27}$H$_{28}$NO$_i$$_0$ requires $m/z$ 526.1713).

10a

[0242] 3-Methoxy- N-[[7-(3 $\alpha$?,4$\alpha$?,7$\alpha$?,7$\alpha$?)]-7-methoxy-6,6-dimethyl-2-
oxotetrahydro-3 $\alpha$H-[l,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 $\alpha$H-chromen-3-
yl]benzamide (10a). Colorless solid (43%): $[\alpha]_{D}^{25}$ = -27.1$^o$ (c = 1.22, 20% MeOH in 
CH$_2$Cl$_2$); $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 8.82 (s, IH), 8.76 (s, IH), 7.51 - 7.44 (m, 3H),
7.39 (d, J = 8.7 Hz, IH), 7.19 - 7.11 (m, 2H), 5.81 (d, J = 1.8 Hz, IH), 5.08 (dd, J = 1.8,
7.8 Hz, IH), 4.98 (t, J = 7.8 Hz, IH), 3.91 (s, 3H), 3.62 (s, 3H), 3.33 (d, J = 7.8 Hz, IH),
2.32 (s, 3H), 1.38 (s, 3H), 1.22 (s, 3H); $^{13}$C NMR (100 MHz, CDCl$_3$) $\delta$ 166.3, 160.5,
159.6, 155.6, 153.6, 149.5, 135.5, 130.4, 126.3, 124.5, 122.6, 119.3, 119.0, 115.3, 115.1,
112.9, 111.7, 94.8, 83.3, 78.3, 77.4, 77.1, 61.0, 55.9, 27.9, 22.6, 8.9; IR (film) $\nu_{\text{max}}$ 3402,
3057, 2988, 2939, 2839, 1819, 1809, 1709, 1676, 1609, 1526, 1487, 1369, 1263, 1175,
1153, 1097, 1076 cm$^{-1}$; HRMS (ESI+) $m/z$ 526.1695 (M + H$^+$, C$_{27}$H$_{28}$NO$_i$$_0$ requires $m/z$
526.1713).

11a

[0243] 4-Methoxy- N-[[7-(3 $\alpha$?,4$\alpha$?,7$\alpha$?,7$\alpha$?)]-7-methoxy-6,6-dimethyl-2-
oxotetrahydro-3 $\alpha$H-[l,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 $\alpha$H-chromen-3-
yl)benzamide (Ha). Colorless solid (35%): [α] D 24 = -25.8° (c = 0.69, 20% MeOH in CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ 8.81 (s, 1H), 8.71 (s, 1H), 7.91 (d, J = 8.8 Hz, 2H), 7.38 (d, J = 8.7 Hz, 1H), 7.16 (d, J = 8.7 Hz, 1H), 7.02 (d, J = 8.8 Hz, 2H), 5.81 (d, J = 1.8 Hz, 1H), 5.08 (dd, J = 1.9, 7.9 Hz, 1H), 4.98 (t, J = 7.8 Hz, 1H), 3.91 (s, 3H), 3.62 (s, 3H), 3.33 (d, J = 7.9 Hz, 1H), 2.35 (s, 3H), 1.38 (s, 3H), 1.23 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 165.9, 163.4, 159.7, 155.5, 153.6, 149.4, 129.5 (3C), 126.3, 126.2, 124.1, 122.8, 115.3, 114.5 (2C), 111.7, 94.8, 83.3, 78.3, 77.4, 77.1, 61.0, 55.9, 27.9, 22.6, 8.9; IR (film) v max 3406, 2984, 2937, 2839, 1811, 1709, 1670, 1607, 1529, 1506, 1367, 1246, 1175 cm⁻¹; HRMS (ESI+) m/z 526.1690 (M + H⁺, C₂₇H₂₈NO₁₀ requires m/z 526.1713).

[0244] N-7-(7-(3 α?,4i?,7i?,7α?)-7-Methoxy-6,6-dimethyl-2-oxotetrahydro-3 αH-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)biphenyl-2-carboxamide (12a). Colorless solid (35%): [α] D 26 = -22.8° (c = 0.15, 20% MeOH in CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃) δ 8.70 (s, 1H), 7.93 (s, 1H), 7.80 (d, J = 7.8 Hz, 1H), 7.61 - 7.36 (m, 8H), 7.33 (d, J = 8.7 Hz, 1H), 7.13 (d, J = 8.7 Hz, 1H), 5.79 (d, J = 1.8 Hz, 1H), 5.6 (dd, J = 1.8, 7.9 Hz, 1H), 4.97 (t, J = 7.9 Hz, 1H), 3.61 (s, 3H), 3.32 (d, J = 7.9 Hz, 1H), 2.28 (s, 3H), 1.37 (s, 3H), 1.21 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 168.8, 158.8, 155.4 (2C), 140.6, 140.0, 135.1, 131.5, 131.1, 129.4 (2C), 129.2 (2C), 129.1 (2C), 128.6, 128.2, 126.2, 124.0, 122.4, 115.2, 115.1, 111.5, 94.7, 83.3, 78.3, 77.4, 77.1, 61.0, 27.9, 22.6, 8.8; IR (film) v max 3375, 2984, 2935, 1815, 1715, 1672, 1609, 1516, 1367, 1256, 1171, 1111, 1094, 1076 cm⁻¹; HRMS (ESI+) m/z 572.1893 (M + H⁺, C₃₂H₃₀NO₉ requires m/z 572.1921).

13a
[0245] 7-(3α,R,4α,R,7α,R,7α,R)-7-Methoxy-6,6-dimethyl-2-oxotetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yl)oxy)-8-methyl-2-oxo-2 H-chromen-3-yl)biphenyl-3-carboxamide (13a). Colorless solid (58%): [α]D25 = -19.2° (c = 0.12, 20% MeOH in CH2Cl2); 1H NMR (400 MHz, CD2Cl2) δ 8.86 (s, 1H), 8.83 (s, 1H), 8.18 (t, J = 1.7 Hz, IH), 7.99 - 7.85 (m, 2H), 7.71 (dd, J = 1.4, 8.5 Hz, 2H), 7.64 (t, J = 7.8 Hz, IH), 7.54 (t, J = 7.4 Hz, 2H), 7.46 (dd, J = 1.7, 7.4 Hz, 2H), 7.21 (d, J = 8.7 Hz, IH), 5.86 (d, J = 2.2 Hz, IH), 5.11 (dd, J = 2.2, 7.8 Hz, IH), 5.02 (t, J = 7.8 Hz, IH), 3.62 (s, 3H), 3.39 (d, J = 7.8 Hz, IH), 2.35 (s, 3H), 1.40 (s, 3H), 1.25 (s, 3H); 13C NMR (100 MHz, CD2Cl2) δ 166.2, 159.4, 155.6, 153.6, 152.9, 152.2, 127.6 (2C), 126.3, 126.2, 124.2, 122.8, 115.2, 115.1, 111.7, 94.9, 83.2, 78.3, 77.5, 77.0, 60.8, 27.7, 22.4, 8.5; IR (film) νmax 3398, 3063, 2984, 2934, 1809, 1713, 1674, 1607, 1522, 1369, 1261, 1236, 1173, 1155, 1097, 1078 cm⁻¹; HRMS (ESI+) m/z 572.1901 (M + H⁺, C32H30NO9 requires m/z 572.1921).

14a

[0246] N-(7-((3α,R,4α,R,7α,R)-7-Methoxy-6,6-dimethyl-2-oxotetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yl)oxy)-8-methyl-2-oxo-2 H-chromen-3-yl)biphenyl-4-carboxamide (14a). Colorless solid (32%): [α]D25 = -17.3° (c = 0.08, 20% MeOH in CH2Cl2); 1H NMR (400 MHz, CDCl3) δ 8.86 (s, 1H), 8.83 (s, 1H), 8.02 (d, J = 8.4 Hz, 2H), 7.76 (d, J = 8.4 Hz, 2H), 7.67 (dd, J = 1.3, 7.8 Hz, 2H), 7.51 (t, J = 7.8 Hz, 2H), 7.44 (d, J = 7.3 Hz, IH), 7.41 (d, J = 8.4 Hz, IH), 7.18 (d, J = 8.4 Hz, IH), 5.82 (d, J = 1.8 Hz, IH), 5.09 (dd, J = 1.8, 7.9 Hz, IH), 4.99 (t, J = 7.9 Hz, IH), 3.62 (s, 3H), 3.34 (d, J = 7.6 Hz, IH), 2.39 (s, 3H), 1.39 (s, 3H), 1.23 (s, 3H); 13C NMR (100 MHz, CDCl3) δ 165.8, 159.5, 155.6, 153.6, 149.5, 145.4, 140.0, 132.8, 129.4 (2C), 128.8, 128.1 (2C), 127.8 (2C), 127.2 (2C), 126.2, 124.1, 122.8, 115.2, 115.1, 111.7, 94.8, 83.2, 78.3, 77.5, 77.0, 60.8, 27.7, 22.4, 8.6; IR (film) νmax 3400, 3032, 2986, 2935, 2851, 1811, 1713, 1672, 1609, 1529, 1512, 1367, 1248, 1173, 1095 cm⁻¹; HRMS (ESI+) m/z 572.1924 (M + H⁺, C32H30NO9 requires m/z 572.1921).
[0247] IV-(7<3αR,4αR,7αR,7αR)-7-Methoxy-6,6-dimethyl-2-oxotetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-2-nitrobenzamide (15a). Yellow solid (74%): [α]D = -19.5° (c = 0.55, 20% MeOH in CH2Cl2); 1H NMR (400 MHz, CDCl3) δ 8.79 (s, 1H), 8.41 (s, 1H), 8.17 (dd, J = 0.9, 8.1 Hz, 1H), 7.80 - 7.64 (m, 3H), 7.41 (d, J = 8.7 Hz, 1H), 7.17 (d, J = 8.7 Hz, 1H), 5.82 (d, J = 1.8 Hz, 1H), 5.09 (dd, J = 1.8, 7.9 Hz, 1H), 4.98 (t, J = 7.9 Hz, 1H), 3.61 (s, 3H), 3.34 (d, J = 7.9 Hz, 1H), 2.32 (s, 3H), 1.38 (s, 3H), 1.13 (s, 3H); 13C NMR (100 MHz, CDCl3) δ 165.4, 159.2, 155.9, 153.6, 149.7, 146.8, 134.4, 132.2, 131.7, 128.8, 126.5, 125.3, 122.2, 115.3, 114.8, 111.8, 94.7, 83.3, 78.3, 77.7, 77.5, 77.1, 61.0, 27.9, 22.6, 8.8; IR (film) νmax 3379, 3310, 2986, 2937, 2885, 2841, 1809, 1713, 1676, 1607, 1529, 1371, 1348, 1252, 1171, 1105, 1086, 1072, 1036, 1003 cm⁻¹; HRMS (ESI+) m/z 541.1441 (M + H⁺, C26H25N2O requires m/z 541.1458).

[0248] N-(7<3α?,4α?,7α?,7α?)-7-Methoxy-6,6-dimethyl-2-oxotetrahydro-3 αH-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-3-nitrobenzamide (16a). Yellow solid (71%): [α]D = -28.4° (c = 0.29, 20% MeOH in CH2Cl2); 1H NMR (400 MHz, CD2Cl2) δ 8.83 (s, 1H), 8.82 (s, 1H) 8.77 (t, J = 1.9 Hz, 1H), 8.46 (dt, J = 1.9, 8.2 Hz, 1H), 8.27 (d, J = 8.2 Hz, 1H), 7.78 (t, J = 8.2 Hz, 1H), 7.46 (d, J = 8.7 Hz, 1H), 7.21 (d, J = 8.7 Hz, 1H), 5.86 (d, J = 2.1 Hz, 1H), 5.12 (dd, J = 2.1, 8.0 Hz, 1H), 5.03 (t, J = 8.0 Hz, 1H), 3.62 (s, 3H), 3.39 (d, J = 8.0 Hz, 1H), 2.33 (s, 3H), 1.40 (s, 3H), 1.18 (s, 3H); 13C NMR (100 MHz, CD2Cl2) δ 163.9, 159.3, 155.9, 153.6, 149.7, 148.9, 135.8, 133.1, 130.6, 127.1, 126.4, 125.1, 122.8, 122.2, 115.3, 114.8, 111.7, 94.8, 83.2, 78.3,
77.4, 77.0, 60.8, 27.7, 22.4, 8.6; IR (film) v max 3516, 3389, 3088, 3065, 2986, 2939, 2837, 1809, 1713, 1674, 1607, 1529, 1371, 1135, 1249, 1173, 1109, 1090, 1036 cm⁻¹; HRMS (ESI+) m/z 563.1249 (M + Na⁺, C₂₆H₂₄N₂OnNa requires m/z 563.1278).

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\text{17a}
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[0249] \(N-(7-((3\,\alpha?,4\,\alpha?,7\,\alpha?,7\,\alpha?)-7-	ext{Methoxy-6,6-dimethyl-2-oxotetrahydro-3 \(\alpha\)H-}\[1,3\]dioxolo[4,5-c]pyran-4-yl oxy)-8-methyl-2-oxo-2 \(\alpha\)H-chromen-3-yl)-4-nitrobenzamide (17a). \) Yellow solid (95%): \(\alpha\)D²⁴ = -29.5° (c = 0.20, 20% MeOH in CH₂Cl₂); \(^1H\) NMR (400 MHz, CD₂Cl₂) δ 8.83 (d, J = 2.2 Hz, 2H), 8.39 (dd, J = 1.7, 8.3 Hz, 2H) 8.13 (dt, J = 2.2, 8.3 Hz, 2H), 7.47 (d, J = 8.8 Hz, IH), 7.22 (d, J = 8.8 Hz, IH), 5.86 (d, J = 2.1 Hz, IH), 5.12 (dd, J = 2.1, 7.8 Hz, IH), 5.03 (t, J = 7.8 Hz, IH), 3.62 (s, 3H), 3.39 (d, J = 7.8 Hz, IH), 2.34 (s, 3H), 1.40 (s, 3H), 1.24 (s, 3H); \(^{13}C\) NMR (100 MHz, CD₂Cl₂) δ 164.3, 159.3, 155.9, 153.6, 150.4, 149.7, 139.6, 128.8 (2C), 126.4, 125.1, 122.4 (2C), 122.2, 115.3, 114.8, 111.7, 94.8, 83.2, 78.3, 77.5, 77.0, 60.8, 27.7, 22.4, 8.5; IR (film) v max 3383, 3364, 3105, 2982, 2945, 2833, 1811, 1709, 1672, 1605, 1529, 1371, 1346, 1177, 1109, 1092, 1028 cm⁻¹; HRMS (ESI+) m/z 563.1273 (M + Na⁺, C₂₆H₂₄N₂OnNa requires m/z 563.1278).

[0250] General procedure for solvolysis of the cyclic carbonate: Et₃N (10% total volume) was added dropwise to a solution of cyclic carbonate in methanol. The resulting mixture was stirred for 14 hours, and then concentrated. The residue was purified via preparative TLC or column chromatography (SiO₂, 4:1; CH₂Cl₂:acetone).

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[0251] \(N-(7-((2\,\alpha?,3\,\alpha?,4\,\alpha?,5\,\alpha?)\cdot3,4\text{-Dihydroxy}-5\text{-methoxy-6,6-dimethyltetrahydro-2 \(\alpha\)H-pyran-2-yl oxy)-8-methyl-2-oxo-2 \(\alpha\)H-chromen-3-yl)benzamide (8). \) Colorless solid (83%): \(\alpha\)D²⁴ = -25.2° (c = 0.16, 20% MeOH in CH₂Cl₂); \(^1H\) NMR (400 MHz, 20%
CD$_3$OD in CD$_2$Cl$_2$ δ 8.77 (s, 1H), 7.89 (d, J = 7.4 Hz, 2H) 7.59 (t, J = 7.4 Hz, 1H), 7.53 (d, J = 6.3 Hz, 1H), 7.51 (d, J = 7.4 Hz, 1H), 7.39 (d, J = 8.7 Hz, 1H), 7.21 (d, J = 8.7 Hz, 1H), 5.55 (d, J = 2.2 Hz, 1H), 4.18 - 4.08 (m, 2H), 3.57 (s, 3H), 3.33 - 3.31 (m, 1H), 2.27 (s, 3H), 1.35 (s, 3H), 1.10 (s, 3H); $^{13}$C NMR (100 MHz, 20% CD$_3$OD in CD$_2$Cl$_2$) δ 164.5, 157.7, 154.6, 147.6, 132.1, 130.7, 127.2 (2C), 125.5 (2C), 124.2, 123.1, 120.0, 112.5, 112.1, 110.0, 96.7, 82.5, 76.9, 69.5, 66.8, 60.0, 27.0, 20.6, 6.3; IR (film) ν$_{max}$ 3400, 3088, 3065, 2978, 2926, 2853, 1713, 1668, 1607, 1526, 1493, 1369, 1252, 1092, 1080 cm$^{-1}$; HRMS (ESI+) m/z 470.1826 (M + H$, C_{25}$H$_{28}$NO$_8$ requires m/z 470.1815).

[0252] $N$-(7-((2i?,3i?,4S,5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 $H$-pyran-2-yloxy)-8-methyl-2-oxo-2 $H$-chromen-3-yl)-2-methoxybenzamide (9). Colorless solid (74%): [α]$^{25}_D$ = -16.1° (c = 0.16, 20% MeOH in CH$_2$Cl$_2$); $^1$H NMR (400 MHz, 20% CD$_3$OD in CD$_2$Cl$_2$) δ 8.79 (s, 1H), 8.15 (d, J = 7.9 Hz, 1H), 7.57 (t, J = 7.8 Hz, 1H), 7.38 (d, J = 8.7 Hz, 1H), 7.20 (d, J = 8.7 Hz, 1H), 7.16 - 7.10 (m, 2H), 5.55 (s, 1H) 4.20 - 4.14 (m, 2H), 4.11 (s, 3H), 3.58 (s, 2H), 3.35 (d, J = 8.0 Hz, 1H), 2.28 (s, 3H), 1.34 (s, 3H), 1.11 (s, 3H); $^{13}$C NMR (100 MHz, 20% CD$_3$OD in CD$_2$Cl$_2$) δ 164.9, 160.2, 158.4, 156.7, 149.8, 134.6, 132.4, 126.3, 125.3, 123.0, 122.9, 121.9, 121.2, 114.7, 112.5, 111.7, 99.1, 84.8, 79.2, 71.9, 69.0, 62.1, 56.8, 29.1, 22.9, 8.5; IR (film) ν$_{max}$ 3373, 2947, 2835, 2525, 1641, 1630, 1610, 1448, 1412, 1398 cm$^{-1}$; HRMS (ESI+) m/z 500.1893 (M + H$, C_{26}$H$_{30}$NO$_9$ requires m/z 500.1921).

[0253] $N$-(7-((2i?,3i?,4S,5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 $H$-pyran-2-yloxy)-8-methyl-2-oxo-2 $H$-chromen-3-yl)-3-methoxybenzamide (10). Colorless solid (59%): [α]$^{24}_D$ = -16.9° (c = 0.81, 20% MeOH in CH$_2$Cl$_2$); $^1$H NMR (400 MHz, 20% CD$_3$OD in CD$_2$Cl$_2$) δ 8.70 (s, 1H), 7.42 - 7.33 (m, 4H), 7.19 (d, J = 8.7 Hz, 1H), 7.13 (d,
$J = 8.7 \text{ Hz}, \text{IH})$, 5.52 (s, IH), 4.19 - 4.05 (m, 2H), 3.84 (s, 3H), 3.55 (s, 3H), 3.32 (d, $J = 7.8 \text{ Hz}, \text{IH}$), 2.24 (s, 3H), 1.30 (s, 3H), 1.07 (s, 3H); $^{13}$C NMR (100 MHz, 20% CD$_2$OD in CD$_2$Cl$_2$) $\delta$ 167.1, 160.8, 160.1, 157.1, 150.0, 135.9, 130.6, 126.5, 126.1, 122.2, 119.6, 118.9, 114.8, 114.4, 113.3, 111.9, 99.2, 84.8, 79.3, 71.9, 69.1, 62.1, 56.0, 29.1, 22.9, 8.4; IR (film) $\nu_{\text{max}}$ 3400, 3082, 2980, 2937, 2835, 1709, 1670, 1607, 1526, 1369, 1259, 1138, 1090 cm$^{-1}$; HRMS (ESI+) $m/z$ 500.1899 (M + H$^+$, C$_{26}$H$_{30}$NO$_9$ requires $m/z$ 500.1921).

[0254] $N$-(7-[(2i?,3i?,45’,5i?)]-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyrano-2-ylxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-4-methoxybenzamide (11). Colorless solid (75%): $[^{25}\text{D}]_D = -13.0^\circ$ (c = 0.10, 20% MeOH in CH$_2$Cl$_2$); $^1$H NMR (400 MHz, CD$_2$Cl$_2$) $\delta$ 8.78 (s, IH), 8.68 (s, IH), 7.91 (dt, $J = 2.6, 6.9 \text{ Hz}, 2\text{H}$) , 7.40 (d, $J = 8.7 \text{ Hz}, \text{IH}$), 7.22 (d, $J = 8.7 \text{ Hz}, \text{IH}$), 7.03 (dt, $J = 2.6, 8.9 \text{ Hz}, 2\text{H}$), 5.63 (d, $J = 2.0 \text{ Hz}, \text{IH}$), 4.30 - 4.23 (m, 2H), 3.91 (s, 3H), 3.62 (s, 3H), 3.38 (d, $J = 8.8 \text{ Hz}, \text{IH}$), 2.98 - 2.70 (m, 2H), 2.31 (s, 3H), 1.39 (s, 3H), 1.16 (s, 3H); $^{13}$C NMR (100 MHz, CD$_2$Cl$_2$) $\delta$ 165.7, 163.3, 159.7, 156.3, 149.4 (2C), 129.4, 126.3, 126.0, 124.2, 122.4, 114.5 (2C), 114.4 (2C), 111.5, 98.3, 84.6, 78.9, 71.5, 68.9, 62.1, 55.9, 29.2, 22.7, 8.5; IR (film) $\nu_{\text{max}}$ 3404, 2976, 2934, 2841, 1607, 1506, 1369, 1248, 1176, 1091 cm$^{-1}$; HRMS (ESI+) $m/z$ 500.1896 (M + H$^+$, C$_{26}$H$_{30}$NO$_9$ requires $m/z$ 500.1921).

[0255] $N$-(7-[(2i?,3i?,45’,5i?)]-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyrano-2-ylxy)-8-methyl-2-oxo-2 H-chromen-3-yl)biphenyl-2-carboxamide (12). Colorless solid (55%): $[^{24}\text{D}]_D = -7.2^\circ$ (c = 0.13, 20% MeOH in CH$_2$Cl$_2$); $^1$H NMR (400 MHz, CD$_2$Cl$_2$) $\delta$ 8.77 (s, IH), 7.99 (s, IH), 7.78 (d, $J = 8.0 \text{ Hz}, \text{IH}$), 7.65 - 7.33 (m, 9H), 7.20 (d, $J = 8.8 \text{ Hz}, \text{IH}$), 5.61 (d, $J = 1.8 \text{ Hz}, \text{IH}$), 4.28 - 4.21 (m, 2H), 3.61 (s, 3H), 3.37 (d, $J = 8.7 \text{ Hz}, \text{IH}$), 2.25 (s, 3H), 1.38 (s, 3H), 1.14 (s, 3H); $^{13}$C NMR (125 MHz,
N-[7-(2i?,3i?,45',5i?)]-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)biphenyl-3-carboxamide (13).

Colorless solid (75%): [a]D24 = -24.3° (c = 0.09, 20% MeOH in CH2Cl2); 1H NMR (400 MHz, 20% CD3OD in CD2Cl2) δ 8.84 (s, IH), 8.82 (s, IH), 8.17 (s, IH), 7.91 (d, J = 7.7 Hz, IH), 7.86 (t, J = 6.7 Hz, IH), 7.71 (dd, J = 1.3, 7.7 Hz, 2H), 7.63 (t, J = 7.7 Hz, IH), 7.53 (t, J = 7.7 Hz, 2H), 7.44 (t, J = 7.5 Hz, 2H), 7.25 (d, J = 8.7 Hz, IH), 5.64 (s, IH), 4.31 - 4.25 (m, 2H), 3.62 (s, 3H), 3.38 (d, J = 8.7 Hz, IH), 2.32 (s, 3H), 1.39 (s, 3H), 1.16 (s, 3H); 13C NMR (125 MHz, 20% CD3OD in CD2Cl2) δ 166.9, 159.8, 156.8, 149.7, 145.0, 142.4, 140.3, 134.8, 131.3, 129.7, 129.3 (2C), 128.2, 127.6 (2C), 126.2 (2C), 125.7, 121.9, 114.5, 114.1, 111.6, 98.9, 84.5, 78.9, 71.6, 68.7, 61.8, 28.8, 22.6, 8.6; IR (film) νmax 3458, 3400, 3060, 2982, 2930, 2854, 1713, 1668, 1628, 1607, 1526, 1367, 1265, 1238, 1095, 1082 cm⁻¹; HRMS (ESI+) m/z 546.2112 (M + H+, C3iH32NO8 requires m/z 546.2128).

N-[7-(2i?,3i?,4S,5i?)]-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)biphenyl-4-carboxamide (14).

Colorless solid (80%): [a]D26 = -7.3° (c = 0.06, 20% MeOH in CH2Cl2); 1H NMR (400 MHz, 20% CD3OD in CD2Cl2) δ 8.87 (s, IH), 8.78 (s, IH), 7.98 (d, J = 8.4 Hz, 2H), 7.75 (d, J = 8.4 Hz, 2H), 7.65 (dd, J = 1.3, 7.8 Hz, 2H), 7.47 (t, J = 7.4 Hz, 2H), 7.38 (d, J = 94
7.8 Hz, 2H), 7.21 (d, J = 8.4 Hz, IH), 5.55 (d, J = 2.1 Hz, IH), 4.18 - 4.08 (m, 2H), 3.80 (s, 3H), 3.35 - 3.30 (m, IH), 2.27 (s, 3H), 1.32 (s, 3H), 1.07 (s, 3H); $^{13}$C NMR (100 MHz, 20% CD$_3$OD in CD$_2$Cl$_2$) δ 159.9, 156.7, 148.9, 145.8, 138.7, 129.3 (2C), 128.6, 128.1 (3C), 127.8 (2C), 127.5 (3C), 126.2, 125.3, 122.8, 114.5, 114.2, 111.6, 98.8, 84.5, 78.9, 71.6, 68.8, 61.9, 28.9, 22.6, 8.3; IR (film) $\nu_{\text{max}}$ 3404, 3059, 3032, 2978, 2932, 2835, 1709, 1666, 1609, 1531, 1416, 1367, 1265, 1252, 1095, 1078 cm$^{-1}$; HRMS (ESI+) $m/z$ 546.2140 (M + H$^+$, C$_{37}$H$_{32}$NO$_8$ requires $m/z$ 546.2128).

[0258] $N$-((7-((2i?,3i?,4S,5R)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 $H$-pyran-2-yloxy)-8-methyl-2-oxo-2 $H$-chromen-3-yl)-2-nitrobenzamide (15). Prepared as described above with the exception that the product was purified by preparative TLC (SiO$_2$, 1:1 hexanes:ethyl acetate, developed 5 times) to afford 15 (44%) as a yellow solid: [c]$^{25}$D = -15.9° (c = 0.15, 20% MeOH in CH$_2$Cl$_2$); $^1$H NMR (400 MHz, 20% CD$_3$OD in CD$_2$Cl$_2$) δ 8.75 (s, IH), 8.15 (dd, J = 0.9, 8.1 Hz, IH), 7.79 - 7.61 (m, 3H), 7.37 (d, J = 8.8 Hz, IH), 7.20 (d, J = 8.8 Hz, IH), 5.56 (d, J = 2.0 Hz, IH), 4.19 - 4.08 (m, 2H), 3.59 (s, 3H), 3.35 (d, J = 9.1 Hz, IH), 2.25 (s, 3H), 1.34 (s, 3H), 1.09 (s, 3H); $^{13}$C NMR (125 MHz, CD$_2$Cl$_2$) δ 165.1, 159.2, 156.8, 149.8, 146.9, 134.4, 132.2, 131.8, 128.9, 126.4, 125.6, 125.2, 122.0, 114.6, 114.0, 111.6, 98.2, 84.6, 78.9, 71.6, 69.0, 62.1, 29.3, 22.6, 8.5; IR (film) $\nu_{\text{max}}$ 3441, 3387, 2984, 2934, 1713, 1674, 1607, 1529, 1371, 1348, 1256, 1105, 1084 cm$^{-1}$; HRMS (ESI+) $m/z$ 515.1669 (M + H$^+$, C$_{23}$H$_{27}$N$_2$O$_4$I requires $m/z$ 515.1666).

[0259] $N$-((7-((2i?,3i?,4S,5i?)3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 $H$-pyran-2-yloxy)-8-methyl-2-oxo-2 $H$-chromen-3-yl)-3-nitrobenzamide (16). Yellow solid (73%): [c]$^{25}$D = -15.7° (c = 0.26, 20% MeOH in CH$_2$Cl$_2$); $^1$H NMR (400 MHz, CDCl$_3$) δ
8.71 (s, 2H), 8.36 (d, J = 1.0, 8.2 Hz, IH) 8.20 (d, J = 1.0, 8.2 Hz, IH), 7.68 (t, J = 8.0 Hz, IH), 7.31 (d, J = 8.7 Hz, IH), 7.15 (d, J = 8.7 Hz, IH), 5.51 (d, J = 2.0 Hz, IH), 4.14 - 4.05 (m, 2H), 3.59 (s, 3H), 3.31 (d, J = 9.1 Hz, IH), 2.33 (s, 3H), 1.29 (s, 3H), 1.10 (s, 3H); 13C NMR (100 MHz, CDCl3) δ 164.3, 159.7, 157.0, 149.7, 148.8, 135.7, 133.2, 130.5, 127.1, 126.7, 126.4, 122.9, 121.4, 114.6, 113.8, 111.7, 98.7, 84.5, 79.0, 71.5, 68.8, 62.1, 29.2, 22.7, 8.5; IR (film) νmax 3362, 2986, 2949, 2837, 1705, 1645, 1635, 1605, 1554, 1531, 1371, 1346, 1253, 1136, 1117, 1003, 1080, 1018 cm⁻¹; HRMS (ESI⁺) m/z 537.1477 (M + Na⁺), C25H26N2O19Na requires m/z 537.1485.

![Structure Image]

[0260] N-(7-((2i?,3i?,45i?,5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-4-nitrobenzamide (17). Yellow solid (79%): [α]D²⁶ = -13.1° (c = 0.16, 20% MeOH in CH₂Cl₂); ¹H NMR (400 MHz, 20% CD₃OD in CD₂Cl₂) δ 9.18 (s, IH), 8.79 (dd, J = 1.9, 6.9 Hz, 2H) 8.56 (dd, J = 1.9, 6.9 Hz, 2H), 7.85 (d, J = 8.7 Hz, IH), 7.67 (d, J = 8.7 Hz, IH), 6.00 (d, J = 2.2 Hz, IH), 4.60 (dd, J = 3.4, 9.4 Hz, IH), 4.56 (t, J = 3.4 Hz, IH), 4.02 (s, 3H), 3.80 (d, J = 9.4 Hz, IH), 2.71 (s, 3H), 1.75 (s, 3H), 1.61 (s, 3H); 13C NMR (100 MHz, 20% CD₃OD in CD₂Cl₂) δ 165.0, 159.6, 157.1, 150.3, 149.9, 139.7, 129.0 (2C), 127.0, 126.4, 124.2 (2C), 121.5, 114.5, 113.8, 111.7, 98.9, 84.4, 79.0, 71.6, 68.7, 61.8, 28.8, 22.5, 8.1; IR (film) νmax 3381, 3053, 2947, 2835, 1699, 1666, 1603, 1524, 1373, 1346, 1252, 1113, 1086, 1018 cm⁻¹; HRMS (ESI+) m/z 537.1486 (M + Na⁺), C25H26N2O19Na requires m/z 537.1485.

[0261] General procedure for reduction of nitro group: Palladium on carbon (10%, 0.1 eq) was added to a solution of 15, 16 or 17 (1 eq) in THF at room temperature. The suspension was stirred for six hours under a hydrogen atmosphere, filtered through a plug of SiO₂, and eluted with THF. The eluent was concentrated and the residue purified by preparative TLC (SiO₂, 100:1 → 50:1; CH₂Cl₂:acetone) to afford the aniline.
2-Amino-ΛH7-((2iU^,4S,5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)benzamide (18). Colorless solid (90%): [α]23D = -17.6° (c = 0.09, 20% MeOH in CH2Cl2); 1H NMR (500 MHz, 20% CD3OD in CD2Cl2) δ 8.64 (s, 1H), 7.48 (dd, J = 1.0, 7.5 Hz, IH), 7.31 (d, J = 9.0 Hz, IH), 7.24 - 7.17 (m, IH), 7.14 (d, J = 8.5 Hz, IH), 6.70 (d, J = 8.5 Hz, IH), 6.66 (t, J = 7.5 Hz, IH), 5.48 (d, J = 2.0 Hz, IH), 4.09 (dd, J = 3.3, 9.5 Hz, IH), 4.04 (t, J = 3.3 Hz, IH), 3.50 (s, 3H), 3.28 - 3.25 (m, IH), 2.21 (s, 3H), 1.26 (s, 3H), 1.03 (s, 3H); 13C NMR (125 MHz, 20% CD3OD in CD2Cl2) δ 168.4, 159.9, 156.5, 149.6, 149.5, 133.6, 127.9, 126.0, 124.6, 122.1, 118.0, 117.2, 115.3, 114.5, 114.3, 111.6, 98.8, 84.5, 78.9, 71.6, 68.8, 61.9, 28.9, 22.6, 8.3; IR (film)νmax 3470, 3408, 3362, 2978, 2926, 2853, 1707, 1657, 1609, 1520, 1450, 1408, 1367, 1263, 1242 1088 cm⁻¹; HRMS (ESI+) m/z 485.1919 (M + H+, C25H29N2O8 requires m/z 485.1924).

3-Amino- N-(7-((2i?,3i?,4S,5i?) -3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)benzamide (19). Colorless solid (77%): [α]26D = -24.3° (c = 0.07, 20% MeOH in CH2Cl2); 1H NMR (400 MHz, 20% CD3OD in CD2Cl2) δ 8.79 (s, 1H), 8.76 (s, 1H), 7.41 (d, J = 8.7 Hz, IH), 7.32 - 7.20 (m, 4H), 6.92 (d, J = 6.3 Hz, IH), 5.58 (d, J = 1.6 Hz, IH), 4.20 - 4.11 (m, 2H), 3.60 (s, 3H), 3.35 (d, J = 9.5 Hz, IH), 2.30 (s, 3H), 1.35 (s, 3H), 1.17 (s, 3H); 13C NMR (200 MHz, 20% CD3OD in CD2Cl2) δ 166.8, 159.4, 156.3, 149.2, 147.7, 134.7, 129.6, 125.7, 124.8, 121.6, 118.9, 116.3, 114.1, 113.8, 113.2, 111.2, 98.4, 84.1, 78.5, 71.2, 68.4, 61.4, 28.5, 22.2, 7.8; IR (film)νmax 3404, 2986, 2949, 2843, 1634, 1607, 1520, 1367, 1261, 1111, 1016 cm⁻¹; HRMS (ESI+) m/z 507.1740 (M + Na+, C25H28N2O8Na requires m/z 507.1743).
4-Amino- N-(7-((2R,3S,4S,5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)benzamide (20). Yellow solid (77%): [α]$_D^{26}$ = -15.9° (c = 0.30, 20% MeOH in CH$_2$Cl$_2$); $^1$H NMR (400 MHz, CDCl$_3$) δ 8.76 (s, 1H), 8.61 (s, 1H), 7.75 (dd, J = 1.8, 6.8 Hz, 2H), 7.31 (d, J = 8.7 Hz, 1H), 7.17 (d, J = 8.7 Hz, 1H), 6.73 (td, J = 1.8, 6.8 Hz, 2H), 5.60 (d, J = 1.5 Hz, 1H), 4.28 - 4.24 (m, 2H), 4.16 (s, 2H), 3.62 (s, 3H), 3.39 (d, J = 8.8 Hz, 1H), 3.17 (s, 1H), 2.90 (s, 1H), 2.27 (s, 3H), 1.39 (s, 3H), 1.14 (s, 3H); $^1$C NMR (100 MHz, CDCl$_3$) δ 166.2, 159.9, 156.7, 150.9, 149.3, 129.5 (2C), 126.1, 124.2, 123.3, 122.4, 114.7 (2C), 114.6 (2C), 111.5, 98.3, 84.7, 79.0, 71.6, 69.0, 62.3, 29.6, 22.9, 8.8; IR (film) v$_{max}$ 3381, 2980, 2941, 2839, 1697, 1634, 1607, 1531, 1510, 1367, 1252, 1184, 1092, 1078, 1020, 993, 966 cm$^{-1}$; HRMS (ESI+) m/z 485.1940 (M + H$^+$, C$_{25}$H$_{29}$N$_2$O$_8$ requires m/z 485.1924).

**EXAMPLE 24: PREPARATION OF NOVOBIOCIN DERIVATIVES WITH VARIOUS LINKERS**

Simultaneous with the investigation of aryl substitutes, modification of the amide and tether functionalities was also explored as shown in the scheme below. Sulfonamide 21 was assembled by sulfonylation of amine 7 from the prior example with benzenesulfonyl chloride, the carbonate of which was subjected to solvolysis to provide the resulting diol. The CBz-containing product 24, was obtained by direct solvolysis of 5. Amides 22, 23, and 25 were prepared by coupling the appropriate acid with amine 7 in the presence of EDCI and pyridine, followed by solvolysis of the cyclic carbonate.
More specifically, compound 21 was prepared as follows:

[0267] \( N\)-(7-((2i?,3i?,4i?,5i?) 3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 \( H \)-pyran-2-yloxy)-8-methyl-2-oxo-2 \( H \)-chromen-3-yl)benzenesulfonamide (21).

Benzenesulfonyl chloride (8 \( \mu \)L, 10.7 mg, 60.2 \( \mu \)mol) was added to a solution of 21.4 mg (54.7 \( \mu \)mol) aniline 7 in 0.5 mL of pyridine at room temperature. The reaction mixture was stirred for 14 hours and then concentrated. The residue was purified via preparative TLC (40:1 \( CH_2Cl_2 \):acetone) to afford 20 mg (69%) of the sulfonamide as a glassy solid.

The cyclic carbonate (20 mg) was dissolved in 0.5 mL of methanol and 0.1 mL of Et\(_3\)N was added dropwise. The reaction mixture was stirred for 14 hours at room temperature before concentration. The residue was purified by preparative TLC (10:1; \( CH_2Cl_2 \):methanol) to afford 15 mg (79%) of 21 as a white solid. \([\alpha]^{24}_D = -5.1^\circ \) (c = 0.42, 20% MeOH in \( CH_2Cl_2 \)); \( ^1H \) NMR (400 MHz, \( CD_2Cl_2 \)) \( \delta \) 7.88 (dd, \( J = 0.8, 8.0 \) Hz, 2H), 7.75 (s, IH), 7.58 (t, \( J = 7.2 \) Hz, IH), 7.50 (d, \( J = 7.2 \) Hz, IH), 7.48 (d, \( J = 7.2 \) Hz, IH), 7.41 (s, IH), 7.31 (d, \( J = 8.8 \) Hz, IH), 7.17 (d, \( J = 8.8 \) Hz, IH), 5.57 (d, \( J = 2.0 \) Hz, IH), 4.24 - 4.17 (m, 2H), 3.57 (s, 3H), 3.34 (d, \( J = 8.8 \) Hz, IH), 2.53 (s, 2H), 2.18 (s, 3H),
1.34 (s, 3H), 1.08 (s, 3H); 13C NMR (100 MHz, CD2Cl2) δ 159.1, 156.9, 150.1, 139.2, 134.0, 129.7 (2C), 127.6 (2C), 126.0, 125.7, 120.9, 114.7, 113.5, 111.6, 98.3, 84.5, 78.9, 71.6, 69.0, 62.1, 29.2, 22.7, 8.4; IR (film) νmax 3439, 3429, 2982, 2930, 2853, 1713, 1630, 1609, 1499, 1464, 1448, 1369, 1327, 1285, 1261, 1167, 1113, 1088 cm⁻¹; HRMS (ESI+) m/z 506.1490 (M + H⁺, C24H28NO9S requires m/z 506.1485).

22a

[0268] N-[7-((3α?,4α?,7α?,7α?)-(x)-dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-2-phenylacetamide (22a). Prepared by [94x84]pyran-2-yloxy)-8-methyl-2-oxo-2 phenylacetamide (22a). N-(3-Dimethylamino-propyl)- N'-ethylcarbodiimide hydrochloride (34 mg, 72 μmol) was added to a solution of aminocoumarin 7 (28 mg, 72 μmol) and phenyl acetic acid (24 mg, 179 μmol) in CH2Cl2 at room temperature. The solution was stirred for 14 hours, concentrated and the residue purified via preparative TLC (SiO₂, 40:1; CH2Cl2:acetone) to afford the 24 mg (66%) of 22a as a colorless solid: [α]D²⁴ = -28.5° (c = 0.39, CH2Cl2); 1H NMR (800 MHz, CD2Cl2) δ 8.62 (s, 1H), 8.00 (s, 1H), 7.36 (t, J = 8.0 Hz, 2H), 7.33 - 7.26 (m, 4H), 7.08 (d, J = 8.8 Hz, 1H), 5.74 (d, J = 1.6 Hz, 1H), 5.01 (dd, J = 1.6, 8.0 Hz, 1H), 4.92 (t, J = 8.0 Hz, 1H), 3.72 (s, 2H), 3.53 (s, 3H), 3.30 (d, J = 8.0 Hz, 1H), 2.22 (s, 3H), 1.31 (s, 3H), 1.15 (s, 3H); 13C NMR (200 MHz, CD2Cl2) δ 171.8, 160.6, 157.0, 155.1, 151.0, 136.2, 131.3 (2C), 130.9 (2C), 129.4, 127.6, 125.4, 124.0, 116.6, 116.5, 113.1, 96.3, 84.7, 79.9, 79.0, 78.5, 62.3, 46.5, 29.2, 23.9, 10.0; IR (film) νmax 3333, 3088, 3063, 3030, 2984, 2935, 2851, 1809, 1717, 1684, 1609, 1522, 1369, 1261, 1173, 1111, 1088, 1036 cm⁻¹; HRMS (ESI+) m/z 510.1755 (M + H⁺, C27H28NO9 requires m/z 510.1764).

22

[0269] N-(7-((2α?,3α?,4S,5α?)-(α)-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-2-phenylacetamide (22). Prepared by

100
the general cyclic carbonate solvolysis procedure described above to afford 22 (87\%) as a colorless solid: \([\alpha]_D^{24} = -15.1^\circ\) (c = 0.37, CH\(_2\)Cl\(_2\)); \(^1\)H NMR (800 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 8.58 (s, IH), 8.04 (s, IH), 7.40 - 7.31 (m, 5H), 7.29 (d, \(J = 8.8\) Hz, IH), 7.15 (d, \(J = 8.8\) Hz, IH), 5.56 (d, \(J = 0.8\) Hz, IH), 4.24 - 4.20 (m, 2H), 3.76 (s, 2H), 3.57 (s, 3H), 3.34 (d, \(J = 8.8\) Hz, IH), 2.95 (s, IH), 2.83 (s, IH), 2.22 (s, 3H), 1.31 (s, 3H), 1.10 (s, 3H); \(^{13}\)C NMR (200 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 171.9, 160.7, 157.9, 151.0, 136.2, 131.3 (2C), 130.9 (2C), 129.4, 127.5, 125.9, 123.5, 116.0, 115.7, 113.0, 99.8, 86.1, 80.4, 73.1, 70.5, 63.6, 46.5, 30.8, 24.2, 10.0; IR (film) \(\nu_{\text{max}}\) 3367, 3339, 3086, 3063, 3030, 2980, 2932, 2853, 2831, 1715, 1684, 1607, 1526, 1369, 1263, 1113, 1084 cm\(^{-1}\); HRMS (ESI+) \(m/z\) 484.1982 (M + H\(^+\), C\(_{28}\)H\(_{30}\)NO\(_8\) requires \(m/z\) 484.1971).

\[23a\]

\[0270\] \(N\)-((7-((3 \alpha\?,4\alpha\?,7\alpha\?,\alpha\?)-7-Methoxy-6,6-dimethyl-2-oxotetrahydro-3 \alpha\H-1,3)dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2 \ H-chromen-3-yl)-3-phenylpropanamid (23a). Prepared by the general EDCI coupling procedure described above to afford 23a (21 mg, 58\%) as a colorless solid: \([\alpha]_D^{23} = -23.4^\circ\) (c = 0.32, 20\% MeOH in CH\(_2\)Cl\(_2\)); \(^1\)H NMR (400 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 8.65 (s, IH), 7.97 (s, IH), 7.36 (d, \(J = 8.7\) Hz, IH), 7.32 - 7.13 (m, 5H), 7.15 (d, \(J = 8.7\) Hz, IH), 5.81 (d, \(J = 2.1\) Hz, IH), 5.08 (dd, \(J = 2.1, 8.0\) Hz, IH), 4.98 (t, \(J = 8.0\) Hz, IH), 3.60 (s, 3H), 3.35 (d, \(J = 8.0\) Hz, IH), 3.04 (t, \(J = 7.7\) Hz, 2H), 2.75 (t, \(J = 7.7\) Hz, 2H), 2.28 (s, 3H), 1.36 (s, 3H), 1.20 (s, 3H); \(^{13}\)C NMR (125 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 170.4, 158.0, 154.3, 152.4, 148.3, 139.9, 128.1 (2C), 127.7 (2C), 125.4, 124.8, 122.7, 121.4, 113.9 (2C), 110.4, 93.6, 82.0, 77.2, 76.4, 75.8, 59.6, 38.7, 31.2, 28.6, 21.9, 7.3; IR (film) \(\nu_{\text{max}}\) 3327, 3086, 3063, 3026, 2982, 2930, 2851, 1811, 1717, 1684, 1607, 1522, 1371, 1259, 1173, 1111, 1086, 1036, 1005 cm\(^{-1}\); HRMS (ESI+) \(m/z\) 524.1912 (M + H\(^+\), C\(_{28}\)H\(_{30}\)NO\(_8\) requires \(m/z\) 524.1921).

\[23\]

101
N-7-((2\&3,4,5,6)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yl)-8-methyl-2-oxo-2H-chromen-3-yl)-3-phenylpropanamide (23). Prepared by the general cyclic carbonate solvolysis procedure described above to afford 23 (16 mg, 79%) as a colorless solid: \([\delta^{24}D = -14.6^\circ (c = 0.49, \text{CH}_2\text{Cl}_2)]\); \(^1H\) NMR (800 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 8.60 (s, IH), 7.95 (s, IH), 7.31 - 7.24 (m, 3H), 7.23 (d, \(J = 7.2\) Hz, 2H), 7.18 (t, \(J = 8.7\) Hz, IH), 7.15 (d, \(J = 8.8\) Hz, IH), 5.56 (d, \(J = 2.4\) Hz, IH), 4.23 - 4.17 (m, 2H), 3.56 (s, 3H), 3.33 (d, \(J = 8.8\) Hz, IH), 3.01 (t, \(J = 8.0\) Hz, 2H), 2.85 (s, IH), 2.76 (s, IH), 2.72 (t, \(J = 8.0\) Hz, 2H), 2.22 (s, 3H), 1.33 (s, 3H), 1.10 (s, 3H); \(^13C\) NMR (200 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 173.2, 160.8, 157.8, 150.9, 142.5, 130.4 (2C), 130.2 (2C), 128.2, 127.5, 125.8, 123.6, 116.0, 115.8, 113.0, 99.8, 86.1, 80.4, 73.1, 70.5, 63.6, 40.9, 33.0, 30.8, 24.2, 10.0; IR (film) \(\nu_{\text{max}}\) 3427, 3391, 3325, 3080, 3086, 3061, 2932, 2833, 1709, 1684, 1607, 1529, 1377, 1223, 1113, 1084 cm\(^{-1}\); HRMS (ESI+) \(m/z\) 498.2140 (M + H\(^+\), \(\text{C}_{27}\text{H}_{32}\text{NO}_{8}\) requires \(m/z\) 498.2128).

Benzyl 7-((2\&3,4,5,6)-3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yl)-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (24). Et\(_3\)N (10% total volume) was added dropwise to a solution of cyclic carbonate 7a (25 mg, 48 \(\mu\)mol) in methanol (0.6 mL) at room temperature. The resulting mixture was stirred for 14 hours and then concentrated. The residue was purified via preparative TLC (SiO\(_2\), 4:1; CH\(_2\)Cl\(_2\):acetone) to afford 24 (19 mg, 82%) as a white solid: \([\delta^{25}D = -11.3^\circ (c = 0.84, 20\% \text{MeOH in CH}_2\text{Cl}_2)]\); \(^1H\) NMR (500 MHz, 20% CD\(_3\)OD in CD\(_2\)Cl\(_2\)) \(\delta\) 8.17 (s, IH), 7.35 - 7.21 (m, 6H), 7.10 (d, \(J = 9.0\) Hz, IH), 5.46 (d, \(J = 2.5\) Hz, IH), 5.13 (s, 2H), 4.07 - 4.00 (m, 2H), 3.56 (s, 3H), 3.25 (d, \(J = 9.0\) Hz, IH), 2.17 (s, 3H), 1.24 (s, 3H), 1.01 (s, 3H); \(^13C\) NMR (125 MHz, 20% CD\(_3\)OD in CD\(_2\)Cl\(_2\)) \(\delta\) 157.1, 154.1, 151.7, 147.1, 134.2, 126.7 (2C), 126.5, 126.3 (2C), 123.5, 120.9, 120.0, 112.2, 112.0, 109.3, 96.7, 82.3, 76.7, 69.5, 66.6, 65.9, 59.7, 26.7, 20.4, 6.0; IR (film) \(\nu_{\text{max}}\) 3443, 3421, 2982, 2936, 2836, 2525, 1701, 1632, 1609, 1456, 1416, 1360, 1288, 1115, 1086 cm\(^{-1}\); HRMS (ESI+) \(m/z\) 522.1721 (M + Na\(^+\), \(\text{C}_{28}\text{H}_{38}\text{NO}_{9}\)Na requires \(m/z\) 522.1740).
N-7-(2i?,3i?,4S,5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)cinnamamide (25). N-(3-Dimethylamino-propyl)- N'-ethylcarbodiimide hydrochloride (44 mg, 227 µmol) was added to a solution of aminocoumarin 7 (36 mg, 91 µmol) and trans-cinnamic acid (27 mg, 182 µmol) in CH₂Cl₂ containing 30% pyridine at room temperature. The solution was stirred for 14 hours, concentrated and the residue purified via preparative TLC (SiO₂, 40:1; CH₂Cl₂:acetone) to afford the 30 mg (63%) of the amide as an off-white solid. The cyclic carbonate (30 mg) was dissolved in 0.8 mL of methanol and 0.1 mL OfEt₃N was added dropwise. The reaction mixture was stirred for 14 hours and then was concentrated. The residue was purified by preparative TLC (10:1; CH₂Cl₂:methanol) to afford 25 (29 mg, 86%) as a colorless solid: [α]D²⁶ = -41.7° (c = 0.18, DMSO); ¹H NMR (500 MHz, 20% CD₃OD in CD₂Cl₂) δ 8.69 (s, IH), 7.66 (d, J = 16.0 Hz, IH), 7.56 (d, J = 6.5 Hz, 2H), 7.38 - 7.30 (m, 4H), 7.17 (d, J = 8.5 Hz, IH), 6.80 (d, J = 16.0 Hz, IH), 5.51 (s, IH), 4.19 - 4.02 (m, 2H), 3.53 (s, 3H), 3.30 (d, J = 9.0 Hz, IH), 2.23 (s, 3H), 1.28 (s, 3H), 1.06 (s, 3H); ¹³C NMR (125 MHz, 20% CD₃OD in CD₂Cl₂) δ 164.8, 157.9, 155.7, 148.8, 140.8, 134.7, 131.9, 129.0 (2C), 127.8 (2C), 125.9, 124.0, 122.1, 122.0, 113.3, 112.8, 110.8, 98.5, 83.4, 77.8, 70.8, 67.8, 61.0, 28.5, 22.9, 8.1; IR (film) vmax 3447, 3412, 3385, 3071, 3059, 2924, 2853, 1701, 1609, 1412, 1373, 1262, 1180, 1113, 1082, 1059, 1022 cm⁻¹; HRMS (ESI+) m/z 496.1967 (M + H⁺, C₂₇H₃₀NO₈ requires m/z 496.1971).

EXAMPLE 25: ANTI PROLIFERATIVE ACTIVITIES OF NOVOBIOCIN ANALOGUES

Upon completion of synthesis of library of compounds in Examples 23 and 24, the compounds were evaluated for anti-proliferative activity against SkBr3 (Her2 overexpressing breast cancer cells), MCF-7 (estrogen receptor positive breast cancer cells), HCT-16 (colon cancer cells characterized by wild-type p53), PL45 (pancreatic cancer cells), LNCaP (androgen sensitive prostate cancer cells) and PC-3 (androgen...
independent prostate cancer cells) cell lines. As shown in Table 2, the simplified benzamide 8 manifested anti-proliferative activity. This is in stark contrast to KU-1/A4, which does not. The monosubstituted benzamide variants improved activity. In fact, the most potent anti-proliferative agents identified were the methoxy (9-11) and phenyl (12-14) derivatives that produced activities against most cell lines. Thus, the data suggests that α/β-hydrogen bond acceptor and an m-aryl side chain on the benzamide were most effective. In the nitro (15-17) and aniline (18-20) series, the o-derivatives in each respective series were more active than the corresponding regioisomers, which implicates hydrogen bond interactions may be important in this region of the molecule. Surprisingly, activity was abolished upon replacement of the amide with a sulfonamide (21, Table 3). This suggests that key hydrogen bonding interactions also exist between the amide and the protein target that are critical for manifesting anti-proliferative activity. Concerning the spatial requirements of the hydrophobic pocket, we found that a two carbon spacer between the amide and phenyl ring (23) results in compounds that are more potent than those that contain a methylene linker (22), a benzyl carbamate (24), or a simplified benzamide (8). Furthermore, as seen with trans-cinnamide 25, increasing the rigidity in the ethyl linker provided an additional about 3 fold increase in inhibitory activity versus the saturated derivative (23). These results suggest that the hydrophobic pocket into which the benzamide projects may accommodate larger aromatic systems that exhibit increased affinity.

Overall, several novobiocin analogues were identified that manifested anti-proliferative activity against multiple transformed cells, in particular, the drug resistant pancreatic ductal adenocarcinoma (PL45), a very aggressive cancer that is associated with high mortality rates in patients.

**Table 2. Anti-proliferation Activities of Novobiocin Analogues Reported in μM (n = 3)**

<table>
<thead>
<tr>
<th>Entry (IC&lt;sub&gt;50&lt;/sub&gt;)</th>
<th>SkBr3</th>
<th>MCF-7</th>
<th>HCT-16</th>
<th>PL45</th>
<th>LNCaP</th>
<th>PC-3</th>
</tr>
</thead>
<tbody>
<tr>
<td>8</td>
<td>21.5 ± 1.4</td>
<td>20.6 ± 0.4</td>
<td>13.0 ± 2.1</td>
<td>3.4 ± 0.6</td>
<td>72.0 ± 4.0</td>
<td>67.6 ± 9.7</td>
</tr>
<tr>
<td>9</td>
<td>&gt;100</td>
<td>5.3 ± 1.3</td>
<td>&gt;100</td>
<td>35.8 ± 3.4</td>
<td>N/T</td>
<td>9.1 ± 0.5</td>
</tr>
<tr>
<td>10</td>
<td>&gt;100</td>
<td>5.6 ± 2.5</td>
<td>1.9 ± 0.6</td>
<td>2.8 ± 0.8</td>
<td>21.7 ± 2.0</td>
<td>N/T</td>
</tr>
<tr>
<td>11</td>
<td>15.6 ± 4.2</td>
<td>10.3 ± 0.9</td>
<td>15.9 ± 1.9</td>
<td>5.9 ± 2.1</td>
<td>7.3 ± 0.9</td>
<td>17.1 ± 4.3</td>
</tr>
<tr>
<td>12</td>
<td>39.1 ± 4.1</td>
<td>18.9 ± 7.0</td>
<td>32.7 ± 1.6</td>
<td>14.4 ± 2.4</td>
<td>17.3 ± 5.2</td>
<td>65.3 ± 5.6</td>
</tr>
</tbody>
</table>
Table 3. Anti-proliferation Activities of Novobiocin Analogues Reported in µM (n = 3).

<table>
<thead>
<tr>
<th>Entry (IC&lt;sub&gt;50&lt;/sub&gt;)</th>
<th>SkBr3</th>
<th>MCF-7</th>
<th>HCT-1</th>
<th>PL45</th>
<th>LNCaP</th>
<th>PC-3</th>
</tr>
</thead>
<tbody>
<tr>
<td>21</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>&gt;100</td>
</tr>
<tr>
<td>22</td>
<td>21.4 ± 2.2</td>
<td>16.4 ± 0.4</td>
<td>13.2 ± 0.6</td>
<td>8.5 ± 1.7</td>
<td>10.4 ± 0.2</td>
<td>43.3 ± 13.4</td>
</tr>
<tr>
<td>23</td>
<td>10.2 ± 2.3</td>
<td>6.9 ± 0.3</td>
<td>5.4 ± 0.6</td>
<td>9.8 ± 0.2</td>
<td>92.8 ± 0.9</td>
<td>22.0 ± 5.8</td>
</tr>
<tr>
<td>24</td>
<td>17.8 ± 2.4</td>
<td>12.1 ± 0.1</td>
<td>13.0 ± 0.2</td>
<td>11.4 ± 0.6</td>
<td>N/T</td>
<td>N/T</td>
</tr>
<tr>
<td>25</td>
<td>2.6 ± 0.6</td>
<td>4.0 ± 0.3</td>
<td>3.2 ± 0.5</td>
<td>3.0 ± 1.0</td>
<td>4.5 ± 0.7</td>
<td>3.9 ± 0.05</td>
</tr>
</tbody>
</table>

EXAMPLE 26: SYNTHESIS OF BIARYL NOVOBIOCIN ANALOGUES

[0276] In an effort to incorporate the structure-activity relationships into more efficacious inhibitors, a small library of novobiocin derivatives was prepared. The library explored optimization of the benzamide that contained α/?-methoxy and a m-phenyl substituent. It was believed that this set of compounds could be expeditiously prepared by the coupling of a 3-iodo-4-methoxy benzoic acid with 7, to produce an intermediate (27) upon which the incorporation of additional phenyl substituents could be pursued for elucidation of structure-activity relationships as shown in the scheme below:
To prepare the library of compounds, the Suzuki precursor 27 was prepared by coupling aminocoumarin 7 with benzoic acid 26 in the presence of EDCI and pyridine as shown in the scheme below. The biaryl substituents also included various hydrogen bond acceptors and donors to further probe key binding interactions with Hsp90. After an extensive survey of experimental conditions, it was found that dichloro[l,r-bis(diphenylphosphino)ferrocene] palladium(II) dichloromethane [Pd(dppf)Cl₂] in the presence of substituted phenyl boronic acids and 2 M potassium carbonate in dioxane at 50 °C provided the most reproducible cross-coupling. See Greenfield et al., Convenient synthesis of functionalized terphenyls, Tetrahedron Lett. 44 2729-2732 (2003). To complete the synthesis, the carbonates were removed upon solvolysis with methanolic triethylamine.
More specifically, the following compounds were prepared:

3-Iodo-4-methoxybenzoic acid (26). Lithium hydroxide (72 mg, 1.71 mmol) was added to a mixture of methyl 3-iodo-4-methoxybenzoate (100 mg, 0.342 mmol) in 3.0 mL of a 3:1:1 THF-MeOH-water solution at room temperature. The mixture was stirred for eight hours in the dark and then diluted with H₂O (2 mL). The solution was acidified to pH = 2 by the dropwise addition of concentrated HCl. The solution was extracted twice with EtOAc (10 mL portions) and the combined organic layers dried (Na₂SO₄), filtered, and concentrated to afford acid 26 (95 mg, 100%) as a yellow solid that was suitable for use without further purification: ¹H NMR (400 MHz, DMSO) δ 8.24 (d, J = 2.0 Hz, IH), 7.92 (dd, J = 2.0, 8.7 Hz, IH), 7.07 (d, J = 8.7 Hz, IH), 3.88 (s, 3H).

3-Iodo-4-methoxy- N-(7-((3α,i?,4α,i?,7α,i?,7α,i?)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)benzamide (27). Prepared by the procedure used for compound 24 to afford 27 (96%) as a yellow solid: [α]²⁵D = -13.9° (c = 0.17, CH₂Cl₂); ¹H NMR (400 MHz, DMSO) δ 9.69 (s, IH), 8.47 (d, J = 2.0 Hz, IH), 8.37 (d, J = 2.0 Hz, IH), 8.00 (dd, J = 2.0, 8.6 Hz, IH), 7.60 (d, J = 8.6 Hz, IH), 7.17 (d, J = 8.8 Hz, IH), 7.12 (d, J = 8.8 Hz, IH), 6.08 (d, J = 3.3 Hz, IH), 5.20 - 5.11 (m, 2H), 3.91 (s, 3H), 3.83 - 3.75 (m, 3H), 3.49 (s, 3H), 2.23 (s, 3H), 1.29 (s, 3H), 1.12 (s, 3H); ¹³C NMR (100 MHz, DMSO) δ 164.9, 161.5, 158.8, 156.0, 154.4, 150.3, 139.3, 130.8, 129.6, 128.2, 126.9, 122.7, 114.8, 114.6, 112.1, 111.8, 94.2, 86.7, 82.3, 78.5, 77.2, 76.9, 60.7, 57.6, 27.7, 23.6, 9.0; IR (film) v max 3406, 3096, 3067, 2982, 2937, 2843, 1811, 1701, 1670, 1607, 1593, 1526, 1487, 1367, 1256.
171, 1095, 1078, 1038, 1007 \text{ cm}^{-1}; \text{HRMS (ESI+) } m/z 652.0691 (M + H\textsuperscript{+}, C\textsubscript{27}H\textsubscript{27}NO\textsubscript{i}O\textsubscript{I} requires m/z 652.0680).

[0281] General procedure for Suzuki coupling and solvolysis of the cyclic carbonate: Aryl iodide 27 (1.0 eq), 2 M K\textsubscript{2}CO\textsubscript{3}(aq) (3.0 eq) and the aryl boronic acid were dissolved in dioxane before PdCl\textsubscript{2}(dppf)-CHCl\textsubscript{3} (3 mol%) was added to the solution at room temperature. The resulting solution was stirred at room temperature for 30 minutes and then warmed to 55 °C for 3 to 16 hours. After which, the mixture was concentrated, filtered through a pad of silica gel (eluted with 40:1; CH\textsubscript{2}Cl\textsubscript{2}:acetone) and purified via preparative TLC (SiO\textsubscript{2}, 40:1; CH\textsubscript{2}Cl\textsubscript{2}:acetone). The resulting product was dissolved in methanol containing 10% Et\textsubscript{3}N and stirred for 14 hours before concentrating. The residue was purified by preparative TLC (4:1; CH\textsubscript{2}Cl\textsubscript{2}:acetone) to afford the corresponding diol.

\[ \text{28} \]

\text{[0282]} \text{N}-\text{(7-((2,3,4,5,6)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-6-methoxybiphenyl-3-carboxamide (28). Colorless solid (46%, 2 steps): } [\alpha]^{25}_D = -8.7^\circ (c = 0.23, \text{CH}_2\text{Cl}_2); \text{^1H NMR (800 MHz, CD}_2\text{Cl}_2) \delta 8.78 (s, IH), 8.69 (s, IH), 7.93 (dd, J = 4.4, 8.0 Hz, IH), 7.87 (d, J = 2.4 Hz, IH), 7.55 (d, J = 8.0 Hz, 2H), 7.45 (t, J = 8.0 Hz, 2H), 7.93 (dd, J = 4.4, 8.0 Hz, 2H), 7.21 (d, J = 8.8 Hz, IH), 7.10 (d, J = 8.8 Hz, 2H), 5.60 (s, IH), 4.27 - 4.20 (m, 2H), 3.90 (s, 3H), 3.59 (s, 3H), 3.36 (d, J = 9.6 Hz, IH), 2.73 (s, 2H), 2.28 (s, 3H), 1.36 (s, 3H), 1.13 (s, 3H); \text{^13C NMR (200 MHz, CD}_2\text{Cl}_2) \delta 167.2, 161.7, 161.2, 157.9, 151.0, 139.4, 133.0, 131.8, 131.5 (3C), 130.0 (2C), 129.4, 128.1, 127.6, 125.8, 124.0, 116.1, 116.0, 113.0 (2C), 99.7, 86.1, 80.4, 73.1, 70.5, 63.7, 57.8, 30.8, 24.2, 10.1; IR (film) v\textsubscript{max} 3402, 3086, 3055, 3028, 2974, 2934, 2849, 2837, 1709, 1670, 1607, 1526, 1504, 1489, 1367, 1265, 1231, 1095 \text{ cm}^{-1}; \text{HRMS (ESI+)} m/z 576.2231 (M + H\textsuperscript{+}, C\textsubscript{32}H\textsubscript{34}NO\textsubscript{9} requires m/z 576.2234).
\[ N-(7-((2\,i^?,3\,i^?,4\,S,5\,i^?)\)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 \textit{H}-pyran-2-yloxy)-8-methyl-2-oxo-2 \textit{H}-chromen-3-yl)-6-methoxy-2'-methylbiphenyl-3-carboxamide \] (29). Colorless solid (45%, 2 steps): \( [\alpha]_{D}^{24} = -17.0^\circ \) (c = 0.23, CH\(_2\)Cl\(_2\)); \( ^{1}\text{H} \) NMR (400 MHz, CD\(_2\)Cl\(_2\)) \( \delta \) 8.78 (s, 1H), 8.68 (s, 1H), 7.98 (dd, \( J = 2.4, 8.6 \) Hz, 1H), 7.72 (d, \( J = 2.4 \) Hz, 1H), 7.39 (d, \( J = 8.7 \) Hz, 1H), 7.33 - 7.16 (m, 5H), 7.10 (d, \( J = 8.7 \) Hz, 1H), 5.61 (d, \( J = 1.9 \) Hz, 1H), 4.28 - 4.22 (m, 2H), 3.96 (s, 3H), 3.59 (s, 3H), 3.36 (d, \( J = 8.8 \) Hz, 1H), 2.83 (s, 2H), 2.78 (s, 1H), 2.14 (s, 3H), 1.36 (s, 3H), 1.32 (s, 3H); \( ^{13}\text{C} \) NMR (125 MHz, CD\(_2\)Cl\(_2\)) \( \delta \) 165.6, 160.4, 159.6, 156.3, 149.4, 138.0, 137.3, 131.6, 130.3 (2C), 130.0, 128.8, 128.1, 126.3, 126.0 (2C), 124.1, 122.4, 114.5, 114.4, 111.5, 111.0, 98.2, 84.6, 78.8, 71.6, 69.8, 62.1, 56.1, 29.3, 22.7, 20.0, 8.5; IR (film) \( \nu_{max} \) 3404, 3057, 2974, 2937, 1713, 1672, 1607, 1526, 1501, 1487, 1367, 1265, 1231, 1094 cm\(^{-1}\); HRMS (ESI+) \( m/z \) 590.2390 (M + H\(^{+}\), C\(_{33}\)H\(_{36}\)NO\(_9\) requires \( m/z \) 590.2390).

\[ N-(7-((2\,i^?,3\,i^?,4\,S,5\,i^?)\)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 \textit{H}-pyran-2-yloxy)-8-methyl-2-oxo-2 \textit{H}-chromen-3-yl)-6-methoxy-3'-methylbiphenyl-3-carboxamide \] (30). Colorless solid (46%, 2 steps): \( [\alpha]_{D}^{24} = -14.1^\circ \) (c = 0.17, CH\(_2\)Cl\(_2\)); \( ^{1}\text{H} \) NMR (400 MHz, CD\(_2\)Cl\(_2\)) \( \delta \) 8.79 (s, 1H), 8.70 (s, 1H), 7.91 (dd, \( J = 2.4, 8.6 \) Hz, 1H), 7.86 (d, \( J = 2.4 \) Hz, 1H), 7.41 (d, \( J = 8.6 \) Hz, 1H), 7.36 - 7.30 (m, 3H), 7.21 (d, \( J = 6.0 \) Hz, 2H), 7.10 (d, \( J = 8.7 \) Hz, 1H), 5.61 (d, \( J = 1.9 \) Hz, 1H), 4.27 - 4.20 (m, 2H), 3.90 (s, 3H), 3.59 (s, 3H), 3.37 (d, \( J = 8.8 \) Hz, 1H), 2.80 (s, 2H), 2.43 (s, 3H), 2.29 (s, 3H), 1.35 (s, 3H), 1.14 (s, 3H); \( ^{13}\text{C} \) NMR (125 MHz, CD\(_2\)Cl\(_2\)) \( \delta \) 164.5, 159.1, 158.5, 155.2, 148.3, 137.1, 136.6, 130.4, 129.4, 129.1, 127.4, 127.2 (2C), 125.9, 125.3, 124.9, 123.0, 121.3, 109
113.4, 113.3, 110.3, 110.2, 97.1, 83.4, 77.7, 70.5, 67.8, 61.0, 55.0, 28.1, 21.5, 20.4, 7.3;
IR (film) ν_{max} 3402, 3084, 3051, 2974, 2930, 2839, 1713, 1668, 1607, 1526, 1502, 1367,
1267, 1238, 1092 cm^{-1}; HRMS (ESI+) m/z 590.2390 (M + H^+, C_{33}H_{36}NO_9 requires m/z
590.2390).

[0285] N-(7-((2R,3i?,4,5i?,5i?) -3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-
pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-6-methoxy-4'-methylbiphenyl-3-
carboxamide (31). Colorless solid (66%, 2 steps): [α]^24_D = -16.8° (c = 0.10, CH_2Cl_2); ^1H
NMR (500 MHz, CD_2Cl_2) δ 8.76 (s, IH), 8.67 (s, IH), 7.88 (dd, J = 2.0, 8.5 Hz, IH),
7.83 (d, J = 2.0 Hz, IH), 7.41 (d, J = 8.0 Hz, 2H), 7.36 (d, J = 8.5 Hz, IH), 7.24 (d, J =
8.0 Hz, 2H), 7.18 (d, J = 8.5 Hz, IH), 7.07 (d, J = 8.5 Hz, IH), 7.07 (d, J = 8.5 Hz, IH), 5.58 (s, IH), 4.25 - 4.18
(m, 2H), 3.87 (s, 3H), 3.56 (s, 3H), 3.33 (d, J = 9.0 Hz, IH), 2.70 (s, IH), 2.65 (s, IH),
2.39 (s, 3H), 2.26 (s, 3H), 1.34 (s, 3H), 1.1 1 (s, 3H); ^13C NMR (125 MHz, CD_2Cl_2) δ
165.7, 160.2, 159.6, 156.3, 149.5, 137.8, 134.9, 131.4, 130.1, 129.8 (2C), 129.2 (2C),
128.3, 126.5, 126.1, 124.1, 122.5, 114.5, 114.4, 111.5, 111.3, 98.2, 84.6, 78.8, 71.6, 69.0,
62.1, 56.2, 29.3, 22.7, 21.3, 8.5; IR (film) ν_{max} 3404, 3084, 2972, 2926, 2853, 2841,
1713, 1668, 1520, 1367, 1265, 1232, 1094 cm^{-1}; HRMS (ESI+) m/z 590.2388 (M +
H^+, C_{33}H_{36}NO_9 requires m/z 590.2390).

[0286] N-(7-((2i?,3R,45?,5i?,5i?) -3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-
pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-2',6-dimethoxybiphenyl-3-
carboxamide (32). Colorless solid (62%, 2 steps): [α]^25_D = -12.2° (c = 0.30, CH_2Cl_2); ^1H
NMR (500 MHz, CD_2Cl_2) δ 8.75 (s, IH), 8.66 (s, IH), 7.92 (d, J = 2.5, 9.0 Hz, IH),
7.76 (d, J = 2.5 Hz, IH), 7.39 - 7.33 (m, 2H), 7.23 (dd, J = 2.0, 7.5 Hz, IH), 7.18 (d, J =

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8.5 Hz, IH), 7.07 (d, J = 9.0 Hz, IH), 7.04 - 6.98 (m, 2H), 5.58 (d, J = 2.5 Hz, IH), 4.27 - 4.20 (m, 2H), 3.84 (s, 3H), 3.77 (s, 3H), 3.57 (s, 3H), 3.33 (d, J = 9.0 Hz, IH), 2.75 (d, J = 2.5 Hz, IH), 2.67 (d, J = 2.5 Hz, IH), 2.26 (s, 3H), 1.34 (s, 3H), 1.11 (s, 3H); \(^{13}\)C
NMR (125 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 164.5, 159.7, 158.4, 156.3, 155.1, 148.3, 130.5, 129.6, 128.4, 127.5 (2C), 125.8, 124.9 (2C), 122.8, 121.3, 119.5, 113.3 (2C), 110.3, 110.2, 110.0, 97.0, 83.4, 77.7, 70.5, 67.8, 61.0, 55.1, 54.8, 28.1, 21.5, 7.3; IR (film) \(\nu_{\text{max}}\) 3404, 3080, 2930, 2835, 1709, 1670, 1607, 1526, 1502, 1487, 1367, 1265, 1244, 1094 cm\(^{-1}\); HRMS (ESI+) \(m/z\) 606.2346 (M + H\(^+\), C\(_{33}\)H\(_{36}\)NO\(_{10}\) requires \(m/z\) 606.2339).

[0287] \(N\)-7-(2i?,3i?,4S,5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 \(H\)-pyran-2-yloxy)-8-methyl-2-oxo-2 \(H\)-chromen-3-yl)-3',6-dimethoxybiphenyl-3-
carboxamide (33). Colorless solid (52%, 2 steps): \([\alpha]^{25}_{D} = -15.8^\circ\) (c = 0.43, CH\(_2\)Cl\(_2\)); \(^1H\) NMR (500 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 8.76 (s, 1H), 8.68 (s, 1H), 7.91 (dd, J = 2.5, 8.5 Hz, IH), 7.87 (d, J = 2.5 Hz, IH), 7.39 - 7.31 (m, 2H), 7.19 (d, J = 8.5 Hz, IH), 7.13 - 7.05 (m, 3H), 6.92 (dd, J = 2.5, 8.5 Hz, IH), 5.59 (d, J = 2.0 Hz, IH), 4.28 - 4.20 (m, 2H), 3.89 (s, 3H), 3.84 (s, 3H), 3.58 (s, 3H), 3.35 (d, J = 9.0 Hz, IH), 2.84 (d, J = 1.5 Hz, IH), 2.74 (d, J = 3.0 Hz, IH), 2.27 (s, 3H), 1.35 (s, 3H), 1.12 (s, 3H); \(^{13}\)C NMR (125 MHz, CD\(_2\)Cl\(_2\)) \(\delta\) 164.5, 159.0, 158.6, 158.4, 155.2, 148.3, 138.1, 130.1, 129.1, 128.3, 127.4, 125.3, 124.9, 123.0, 121.3, 121.2, 114.5, 113.3, 113.2, 112.1, 110.3, 110.2, 97.1, 83.4, 77.7, 70.4, 67.8, 61.0, 55.1, 54.5, 28.1, 21.5, 7.3; IR (film) \(\nu_{\text{max}}\) 3404, 3078, 2934, 2835, 1709, 1670, 1607, 1526, 1502, 1487, 1367, 1265, 1244, 1094 cm\(^{-1}\); HRMS (ESI+) \(m/z\) 606.2346 (M + H\(^+\), C\(_{33}\)H\(_{36}\)NO\(_{10}\) requires \(m/z\) 606.2339).
[0288] \( \text{N}^-7^-\text{((2\text{i}?3\text{i}?4\text{S},5\text{i}?)-3,4-\text{Dihydroxy}-5-\text{methoxy}-6,6-\text{dimethyltetrahydro-2} \ H^-\text{pyran-2-ylxy)-8-methyl-2-oxo-2 \ H^-\text{chromen-3-yl})-4',6-\text{dimethoxybiphenyl-3-}
\text{carboxamide (34). Colorless solid (46%, 2 steps): } [\alpha]^{26}_{D} = -13.9^\circ \ (c = 0.49, \text{CH}_2\text{Cl}_2); \ \text{^1H NMR} \ (500 \text{ MHz, CD}_2\text{Cl}_2) \ \delta 8.74 \ (s, \text{IH}), \ 8.66 \ (s, \text{IH}), \ 7.86 \ (dd, J = 2.5, 8.5 \text{ Hz, IH}), \ 7.82 \ (d, J = 2.0 \text{ Hz, IH}), \ 7.46 \ (dd, J = 2.0, 6.5 \text{ Hz, 2H}), \ 7.35 \ (d, J = 8.5 \text{ Hz, IH}), \ 7.17 \ (d, J = 8.5 \text{ Hz, IH}), \ 7.05 \ (d, J = 8.5 \text{ Hz, 1H}), 6.96 \ (dd, J = 2.5, 6.5 \text{ Hz, IH}), \ 5.57 \ (d, J = 2.0 \text{ Hz, IH}), \ 4.24 - 4.17 \ (m, 2H), \ 3.86 \ (s, 3H), \ 3.83 \ (s, 3H), \ 3.56 \ (s, 3H), \ 3.33 \ (d, J = 9.0 \text{ Hz, IH}), \ 2.25 \ (s, 3H), \ 1.34 \ (s, 3H), \ 1.11 \ (s, 3H); \ ^{13}\text{C NMR} \ (125 \text{ MHz, CD}_2\text{Cl}_2) \ \delta 164.6, \ 159.0, \ 158.5, \ 158.4, \ 155.1, \ 148.3, \ 129.9 \ (3\text{C}), \ 128.9 \ (2\text{C}), \ 126.9, \ 125.3, \ 124.9, \ 122.9, \ 121.3, \ 113.3 \ (2\text{C}), \ 112.7 \ (2\text{C}), \ 110.3, \ 110.1, \ 97.0, \ 83.4, \ 77.7, \ 70.4, \ 67.8, \ 61.0, \ 55.0, \ 54.5, \ 28.1, \ 21.5, \ 7.3; \ \text{IR (film) } \nu_{\text{max}} 3404, \ 3082, \ 3057, \ 2934, \ 2837, \ 1709, \ 1668, \ 1607, \ 1518, \ 1493, \ 1265, \ 1248, \ 1232, \ 1118, \ 1111, \ 1094, \ 1080 \text{ cm}^{-1}; \ \text{HRMS (ESI+) } m/z 606.2328 \ (\text{M} + \text{H}^+, \text{C}_{33}\text{H}_{36}\text{NO}_{10} \text{ requires } m/z 606.2339). \)

![Chemical Structure](image)

[0289] \( \text{N}^-7^-\text{((2\text{i}?3\text{i}?4\text{S},5\text{i}?)-3,4-\text{Dihydroxy}-5-\text{methoxy}-6,6-\text{dimethyltetrahydro-2} \ H^-\text{pyran-2-ylxy)-8-methyl-2-oxo-2 \ H^-\text{chromen-3-yl})-2'-\text{hydroxy-6-methoxybiphenyl-3-}
\text{carboxamide (35). Colorless solid (59%, 2 steps): } [\alpha]^{24}_{D} = -13.4^\circ \ (c = 0.50, 20% \text{ MeOH in CH}_2\text{Cl}_2); \ \text{^1H NMR} \ (400 \text{ MHz, CD}_2\text{Cl}_2) \ \delta 8.73 \ (s, \text{IH}), \ 8.67 \ (s, \text{IH}), \ 7.98 \ (dd, J = 2.3, 8.6 \text{ Hz, IH}), \ 7.88 \ (d, J = 2.3 \text{ Hz, IH}), \ 7.39 - 7.25 \ (m, \text{3H}), \ 7.18 \ (d, J = 8.8 \text{ Hz, IH}), \ 7.15 \ (d, J = 8.8 \text{ Hz, IH}), \ 7.06 \ (dd, J = 1.1, 7.4 \text{ Hz, IH}), \ 7.04 \ (d, J = 8.1 \text{ Hz, IH}), \ 5.94 \ (s, \text{IH}), \ 5.57 \ (d, J = 2.1 \text{ Hz, IH}), \ 4.26 - 4.17 \ (m, 2H), \ 3.95 \ (s, 3H), \ 3.58 \ (s, 3H), \ 3.35 \ (d, J = 9.0 \text{ Hz, IH}), \ 2.85 \ (s, \text{IH}), \ 2.75 \ (s, \text{IH}), \ 2.27 \ (s, 3\text{H}), \ 1.36 \ (s, 3\text{H}), \ 1.10 \ (s, 3\text{H}); \ ^{13}\text{C NMR} \ (125 \text{ MHz, CD}_2\text{Cl}_2) \ \delta 164.2, \ 158.5, \ 158.4, \ 155.2, \ 153.0, \ 148.3, \ 130.6 \ (2\text{C}), \ 128.9, \ 128.0, \ 126.5, \ 126.1, \ 125.0, \ 124.7, \ 123.2, \ 121.0, \ 120.1, \ 116.2, \ 113.3, \ 113.1, \ 110.5, \ 110.3, \ 96.7, \ 83.4, \ 77.7, \ 70.4, \ 67.8, \ 60.9, \ 55.5, \ 28.3, \ 22.2, \ 8.5; \ \text{IR (film) } \nu_{\text{max}} 3400, \ 3391, \ 3090, \ 2984, \ 2928, \ 2849, \ 1709, \ 1661, \ 1651, \ 1605, \ 1526, \ 1495, \ 1452, \ 1367, \ 1267, \ 1238, \ 1217, \ 1180, \ 1140, \ 1094 \text{ cm}^{-1}; \ \text{HRMS (ESI+) } m/z 592.2183 \ (\text{M} + \text{H}^+, \text{C}_{32}\text{H}_{34}\text{NO}_{10} \text{ requires } m/z 592.2183). \)
[0290] *N*-((2i?,3i?,45',5i?)\-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 *H*-pyran-2-yloxy)-8-methyl-2-oxo-2 *H*-chromen-3-yl)-3'-hydroxy-6-methoxybiphenyl-3-carboxamide (36). Colorless solid (34%, 2 steps): [α]_D\textsuperscript{24} = -13.5° (c = 0.16, 20% MeOH in CH\textsubscript{2}Cl\textsubscript{2}); \textit{IH} NMR (500 MHz, 20% CD\textsubscript{3}OD in CD\textsubscript{2}Cl\textsubscript{2}) δ 8.78 (s, 1H), 8.76 (d, J = 2.4 Hz, IH), 7.40 (d, J = 8.7 Hz, IH), 7.29 - 7.19 (m, 2H), 7.10 (d, J = 8.7 Hz, IH), 7.02 (dd, J = 1.0, 7.6 Hz, IH), 6.99 (t, J = 2.0 Hz, IH), 6.83 (td, J = 1.0, 8.0 Hz, IH), 5.56 (d, J = 2.1 Hz, IH), 4.19 - 4.09 (m, 2H), 3.89 (s, 3H), 3.58 (s, 3H), 3.34 - 3.32 (m, IH), 2.28 (s, 3H), 1.33 (s, 3H), 1.10 (s, 3H); \textit{13C} NMR (125 MHz, 20% CD\textsubscript{3}OD in CD\textsubscript{2}Cl\textsubscript{2}) δ 165.2, 159.2, 158.8, 156.0, 155.6, 148.5, 138.1, 130.3, 129.1, 128.4, 127.4, 125.2, 125.1, 124.0, 121.0, 120.2, 115.7, 113.7, 113.4, 113.2, 110.6, 110.4, 97.7, 83.5, 77.8, 70.5, 67.7, 68.9, 55.0, 27.9, 21.6, 7.2; IR (film) \nu\textsubscript{max} 3400, 2922, 2851, 1707, 1647, 1630, 1605, 1528, 1501, 1369, 1250, 1095 cm\textsuperscript{-1}; HRMS (ESI+) m/z 592.2191 (M + H\textsuperscript{+}), C\textsubscript{32}H\textsubscript{34}NOi\textsubscript{2} requires m/z 592.2183.

[0291] *N*-((2i?,3i?,45',5i?)\-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 *H*-pyran-2-yloxy)-8-methyl-2-oxo-2 *H*-chromen-3-yl)-4'-hydroxy-6-methoxybiphenyl-3-carboxamide (37). Colorless solid (29%, 2 steps): [α]_D\textsuperscript{24} = -21.7° (c = 0.06, 20% MeOH in CH\textsubscript{2}Cl\textsubscript{2}); \textit{IH} NMR (500 MHz, 20% CD\textsubscript{3}OD in CD\textsubscript{2}Cl\textsubscript{2}) δ 8.80 (s, 1H), 8.74 (s, IH), 7.87 (dd, J = 2.3, 8.7 Hz, IH), 7.82 (d, J = 2.3 Hz, IH), 7.42 - 7.35 (m, 3H), 7.21 (d, J = 8.8 Hz, IH), 7.08 (d, J = 8.6 Hz, IH), 6.86 (d, J = 8.6 Hz, IH), 5.55 (d, J = 2.0 Hz, IH), 4.16 - 4.09 (m, 2H), 3.88 (s, 3H), 3.57 (s, 3H), 3.33 (d, J = 9.4 Hz, IH), 2.27 (s, 3H), 1.32 (s, 3H), 1.09 (s, 3H); \textit{13C} NMR (125 MHz, 20% CD\textsubscript{3}OD in CD\textsubscript{2}Cl\textsubscript{2}) δ 164.5, 158.4, 158.0, 155.0, 154.7, 147.6, 129.4, 129.1 (2C), 128.0, 127.0, 126.0, 124.3, 124.2, 123.2.
120.2, 113.3 (2C), 112.5, 112.3, 109.7, 109.4, 96.9, 82.6, 77.0, 69.7, 66.8, 59.9, 54.1, 27.0, 20.7, 6.3; IR (film) ν max 3402, 3394, 2997, 2922, 2851, 1707, 1653, 1605, 1520, 1495, 1369, 1265, 1234, 1095 cm⁻¹; HRMS (ESI⁺) m/z 592.2176 (M + H⁺, C₃₂H₃₄NO₁₀ requires m/z 592.2183).

[0292] N-(7-((2i?,3i?,45',5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-2 H-chromen-3-yl)-4-methoxy-3-(thianthren-1-yl)benzamide (38). Colorless solid (51%, 2 steps): [α]D²⁶ = -12.0° (c = 0.82, CH₂Cl₂);

¹H NMR (500 MHz, 20% CD₃OD in CD₂Cl₂) δ 8.81 (s, 1H), 8.73 (s, 1H), 8.08 (d, J = 7.6 Hz, 1H), 7.80 (s, 1H), 7.61 (d, J = 6.4 Hz, 1H), 7.56 (d, J = 6.4 Hz, 1H), 7.42 - 7.16 (m, 8H), 5.63 (s, 1H), 4.28 - 4.22 (m, 2H), 3.94 (s, 3H), 3.61 (s, 3H), 3.38 (d, J = 8.0 Hz, 1H), 2.95 (s, 1H), 2.87 (s, 1H), 2.30 (s, 3H), 1.39 (s, 3H), 1.16 (s, 3H); ¹³C NMR (125 MHz, 20% CD₃OD in CD₂Cl₂) δ 165.1, 160.1, 159.2, 155.9, 153.5, 149.1, 138.0, 136.3, 136.1, 135.8, 135.6, 129.9, 129.7, 129.6, 129.2, 128.8, 128.5, 127.8, 127.6, 127.4, 126.0, 125.7, 123.9, 122.0, 114.1, 114.0, 111.1, 110.7, 97.8, 84.2, 78.5, 71.2, 68.6, 61.7, 55.9, 28.9, 22.3, 8.1; IR (film) ν max 3400, 3055, 2976, 2932, 2837, 1709, 1670, 1605, 1526, 1497, 1439, 1367, 1261, 1234, 1111, 1094 cm⁻¹; HRMS (ESI⁺) m/z 714.1822 (M + H⁺, C₃₈H₃₆NO₉S₂ requires m/z 714.1832).

**EXAMPLE 27: ANTI-PROLIFERATIVE ACTIVITIES OF BIARYL COMPOUNDS**

[0293] Upon completion of the synthesis of the biaryl derivatives from Example 26, the compounds were evaluated for anti-proliferative activity against the same cell lines described above. Unfortunately, as presented in Table 4, combinations of the /?-methoxy and m-phenyl substituents on the benzamide (28) did not produce compounds that inhibit cell growth more effectively as originally proposed (28 compared to 11 and 13). Likewise, the toyl derivatives (29 - 31) also manifested lower growth inhibition than biaryl 28, and complete activity was lost against most cell lines upon incorporation of
dihydrothianthrene, 38. Dihydrodibenzofuran 39 lacked reasonable solubility in DMSO and was therefore not evaluated in the studies. However, as seen in the methoxy series (32-34) and the phenol series (35-37), activity increased as polarity and hydrogen bond donor/acceptor properties of the inhibitor increased. For example, introduction of an o-OH (32) improved inhibition about 2 fold when compared to 28, but the corresponding phenol (35) was about 8 times more effective. Key structure-function relationships observed for the biaryl benzamide novobiocin derivatives are summarized in FIG. 15.

Table 4: Anti-proliferation Activities of Novobiocin Biaryl Analogues
Reported in µM (n = 3)

<table>
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<tr>
<th>Entry</th>
<th>IC₅₀</th>
<th>SkBr3</th>
<th>MCF-7</th>
<th>HCT-1</th>
<th>PL45</th>
<th>LNCaP</th>
<th>PC-3</th>
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<td>19.5 ± 2.3</td>
<td>11.4 ± 0.0</td>
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<td>29</td>
<td>32.4 ± 5.1</td>
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<td>6.8 ± 1.5</td>
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<td>50.7 ± 4.3</td>
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</tr>
<tr>
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<td>16.7 ± 6.3</td>
<td>2.4 ± 1.0</td>
<td>1.4 ± 0.1</td>
<td>3.7 ± 0.9</td>
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<td>32</td>
<td>7.1 ± 0.6</td>
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<td>5.2 ± 2.1</td>
<td>3.8 ± 1.7</td>
<td>6.0 ± 1.0</td>
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<tr>
<td>33</td>
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<td>5.1 ± 1.1</td>
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<td>44.0 ± 20.3</td>
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<tr>
<td>35</td>
<td>1.5 ± 0.1</td>
<td>1.5 ± 0.1</td>
<td>4.7 ± 1.4</td>
<td>1.4 ± 0.2</td>
<td>2.6 ± 0.6</td>
<td>16.6 ± 4.4</td>
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<td>36</td>
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<td>4.5 ± 1.2</td>
<td>1.1 ± 0.0</td>
<td>2.5 ± 1.2</td>
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<td>37</td>
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<td>&gt;100</td>
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<td>1.0 ± 0.1</td>
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</table>

**EXAMPLE 28: SYNTHESIS OF HETEROCYCLE NOVOBIOCIN DERIVATIVES**

[0294] In an effort to incorporate the structure-activity relationships into more efficacious inhibitors, a small library of novobiocin derivatives was prepared. This library focused on the incorporation of heterocycles into the benzamide region in order to investigate hydrogen bond donor/acceptor interactions and the effects of rigidity as suggested by initial findings.

[0295] The novobiocin derivatives were prepared by coupling commercially available carboxylic acids with aminocoumarin 7 from Example 23 via treatment with
EDCI and pyridine, the carbonates of the resulting molecules were then solvolyzed with methanolic triethylamine to afford the requisite diols, 40-47 as shown in the scheme below.

[0296] General EDCI coupling procedure B: N-(3-Dimethylamino-propyl)-\(N^{5}\)-ethylcarbodiimide hydrochloride (2.5 eq) was added to a solution of aminocoumarin 7 (1.0 eq), carboxylic acid (2.0 eq) in \(\text{CH}_2\text{Cl}_2\) containing 30% pyridine at room temperature. The solution was stirred for 14 hours, concentrated and the residue purified via preparative TLC (\(\text{SiO}_2, 40:1; \text{CH}_2\text{Cl}_2\):acetone) to afford the amide. The resulting product was dissolved in methanol containing 10% \(\text{Et}_3\text{N}\) and stirred for 14 hours at room temperature. The mixture was concentrated and the residue was purified by preparative TLC (10:1; \(\text{CH}_2\text{Cl}_2\):methanol or 4:1; \(\text{CH}_2\text{Cl}_2\):acetone) to afford the corresponding diol.

[0297] \(N^\prime-(7-((2i?,3i?,4,S,5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 \text{H}-pyran-2-yloxy)-8-methyl-2-oxo-2 \text{H}-chromen-3-yl)\)picolinamide (40). Yellow solid (56%, 2 steps): \([\alpha]^{25}_D = -18.8^\circ\) (c = 0.48, 20% MeOH in \(\text{CH}_2\text{Cl}_2\)); \(^1\text{H}\) NMR (500 MHz, 20% \(\text{CD}_3\text{OD}\) in \(\text{CD}_2\text{Cl}_2\)) 6 8.76 (s, IH), 8.66 (d, \(J = 4.5\) Hz, IH), 8.18 (d, \(J = 7.5\) Hz, IH), 7.93 (dt, \(J = 1.5, 7.5\) Hz, IH), 7.54 - 7.49 (m, IH), 7.37 (d, \(J = 8.5\) Hz, IH), 7.19 (d,
\[ J = 8.5 \text{ Hz, IH} \), 5.53 (d, \( J = 2.0 \text{ Hz, IH} \), 4.14 (dd, \( J = 3.5, 9.5 \text{ Hz, IH} \), 4.09 (t, \( J = 3.5 \text{ Hz, IH} \), 3.55 (s, 3H), 3.32 (d, \( J = 9.5 \text{ Hz, IH} \), 2.27 (s, 3H), 1.31 (s, 3H), 1.08 (s, 3H); \]

\[ ^{13}C \text{ NMR (125 MHz, 20\% CD}_3\text{OD in CD}_2\text{Cl}_2) \delta 165.0, 160.8, 158.1, 151.1, 150.6, 150.3, 139.5, 128.7, 127.4, 126.4, 123.8, 123.0, 115.9, 115.5, 112.9, 100.2, 85.8, 80.2, 72.9, 70.1, 63.1, 30.2, 23.9, 9.5; IR (film) \nu_{\text{max}} 3421, 3065, 2982, 2932, 2837, 2476, 1717, 1682, 1626, 1607, 1522, 1458, 1414, 1375, 1265, 1173, 1134, 1097, 1080 \text{ cm}^{-1}; \]

HRMS (ESI+) \( m/z \) 471.1751 (M + H\(^+\), \( C_{24}H_{27}N_2O_8 \) requires \( m/z \) All. 1767).

\[ \text{[0298]} \ N-(7-((2\text{i},3\text{i},4\text{S,5i})-3,4-\text{Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 \text{H-}} \text{pyran-2-yloxy)-8-methyl-2-oxo-2} H \text{-chromen-3-yl})\text{nicotinamide} \ (\text{41}). \text{ Colorless solid (52\%,} \text{ 2 steps): [} \alpha]^{25}_D = -16.1^\circ \ (c = 0.31, 20\% \text{ MeOH in CH}_2\text{Cl}_2); \]

\[ ^1H \text{ NMR (400 MHz, CD}_2\text{Cl}_2) \delta 9.13 (s, 1H), 8.80 - 8.75 (m, 1H), 8.77 (s, 1H), 8.73 (s, 1H), 8.20 (dd, \( J = 1.6, 7.2 \text{ Hz, IH} \), 7.47 (dd, \( J = 4.8, 7.6 \text{ Hz, IH} \), 7.38 (d, \( J = 8.4 \text{ Hz, IH} \), 7.21 (d, \( J = 8.4 \text{ Hz, IH} \), 5.60 (d, \( J = 2.0 \text{ Hz, IH} \), 4.27 - 4.17 (m, 2H), 3.58 (s, 3H), 3.35 (d, \( J = 8.4 \text{ Hz, IH} \), 2.75 (s, 2H), 2.27 (s, 3H), 1.36 (s, 3H), 1.12 (s, 3H); \]

\[ ^{13}C \text{ NMR (125 MHz, 20\% CD}_3\text{OD in CD}_2\text{Cl}_2) \delta 166.0, 160.9, 158.3, 154.0, 151.1, 149.8, 137.2, 131.7, 127.6 (2C), 125.6, 123.0, 115.9, 115.3, 113.0, 100.1, 85.8, 80.2, 72.9, 70.1, 63.2, 30.2, 23.9, 9.5; IR (film) \nu_{\text{max}} 3394, 3092, 3065, 2980, 2928, 2854, 2833, 1711, 1672, 1605, 1531, 1371, 1258, 1132, 1113, 1095, 1082 \text{ cm}^{-1}; \text{ HRMS (ESI+) } m/z 471.1763 \ (M + H\(^+\), \( C_{24}H_{27}N_2O_8 \) requires \( m/z \) All. 1161). \]

\[ \text{[0299]} \ N-(7-((2\text{i},3\text{i},4\text{S,5i})-3,4-\text{Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 \text{H-}} \text{pyran-2-yloxy)-8-methyl-2-oxo-2} H \text{-chromen-3-yl})\text{isonicotinamide} \ (\text{42}). \text{ Yellow solid (52\%,} \text{ 2 steps): [} \alpha]^{25}_D = -14.3^\circ \ (c = 0.67, 20\% \text{ MeOH in CH}_2\text{Cl}_2); \]

\[ ^1H \text{ NMR (800 MHz, 20\% CD}_3\text{OD in CD}_2\text{Cl}_2) \delta 8.83 - 8.74 (m, 3H), 7.87 - 7.80 (m, 2H), 7.42 (d, \( J = 8.0 \text{ Hz,} \]

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IH), 7.24 (d, J = 8.0 Hz, IH), 5.58 (d, J = 2.4 Hz, IH), 4.20 - 4.11 (m, 2H), 3.60 (s, 3H), 3.36 (d, J = 9.6 Hz, IH), 2.28 (s, 3H), 1.35 (s, 3H), 1.12 (s, 3H); \(^{13}\)C NMR (200 MHz, 20% CD\(_3\)OD in CD\(_2\)Cl\(_2\)) \(\delta\) 164.2, 159.1, 156.7, 150.2 (3C), 149.5, 141.5, 126.4, 126.0, 121.3, 121.0, 114.1, 113.4, 111.3, 98.5, 84.1, 78.6, 71.2, 68.4, 61.4, 28.5, 22.2, 7.8; IR (film) \(v_{\text{max}}\) 3393, 3055, 2982, 2932, 2835, 1709, 1674, 1628, 1607, 1529, 1373, 1258, 1134, 1111, 1095, 1080 cm\(^{-1}\); HRMS (ESI+) \(m/z\) 471.1748 (M + H\(^+\), C\(_{24}\)H\(_{27}\)N\(_2\)O\(_8\)) requires \(m/z\) 471.1767).

\[ \text{[0300]} \quad N^\text{I}-(7-((2i?,3i?,45',5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)benzofuran-2-carboxamide \quad (43), \]

Colorless solid (63%, 2 steps): \([\rho]^{24}_D = -16.8^\circ \) (c = 0.19, 20% MeOH in CH\(_2\)Cl\(_2\)); \(^1\)H NMR (400 MHz, 20% CD\(_3\)OD in CD\(_2\)Cl\(_2\)) \(\delta\) 8.70 (s, 1H), 7.68 (dd, J = 0.8, 8.8 Hz, 1H), 7.59 (dd, J = 0.8, 8.8 Hz, 1H), 7.58 (s, 1H), 7.45 (dt, J = 0.8, 7.6 Hz, 1H), 7.37 (d, J = 8.8 Hz, 1H), 7.29 (dt, J = 0.8, 7.6 Hz, 1H), 7.19 (d, J = 8.8 Hz, 1H), 5.52 (d, J = 2.4 Hz, 1H), 4.12 (dd, J = 3.4, 9.2 Hz, 1H), 4.07 (dd, J = 3.4, 9.2 Hz, 1H), 3.54 (s, 3H), 3.30 (d, J = 9.2 Hz, 1H), 2.25 (s, 3H), 1.29 (s, 3H), 1.06 (s, 3H); \(^{13}\)C NMR (125 MHz, 20% CD\(_3\)OD in CD\(_2\)Cl\(_2\)) \(\delta\) 161.5, 159.8, 159.0, 157.5, 151.8, 150.0, 130.1, 129.9, 128.3, 127.8, 126.4, 125.2, 123.3, 116.6, 116.0, 114.4, 114.2, 113.7, 100.9, 86.5, 81.0, 73.6, 70.8, 63.8, 30.8, 24.6, 10.2; IR (film) \(v_{\text{max}}\) 3404, 3385, 2985, 2935, 2511, 1717, 1670, 1626, 1607, 1576, 1548, 1418, 1377, 1265, 1115, 1092 cm\(^{-1}\); HRMS (ESI+) \(m/z\) 532.1566 (M + Na\(^+\), C\(_{27}\)H\(_{27}\)NO\(_9\)Na requires \(m/z\) 532.1583).

\[ \text{[0301]} \quad N^\text{I}-(7-((2i?,3i?,4S,5i?)-3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)benzo[ \text{h} ]thiophene-2-carboxamide \]

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(44). Colorless solid (42%, 2 steps): [α]D25 = -24.3° (c = 0.23, 20% MeOH in CH2Cl2);
1H NMR (400 MHz, 20% CD3OD in CD2Cl2) δ 8.66 (s, 1H), 8.00 (s, IH), 7.88 (t, J = 7.4 Hz, 2H), 7.45 - 7.33 (m, 3H), 7.19 (d, J = 8.8 Hz, IH), 5.53 (d, J = 2.0 Hz, IH), 4.12 (d, J = 9.2 Hz, IH), 4.09 - 4.05 (m, IH), 3.54 (s, 3H), 3.31 (d, J = 9.2 Hz, IH), 2.25 (s, 3H), 1.30 (s, 3H), 1.07 (s, 3H); 13C NMR (125 MHz, 20% CD3OD in CD2Cl2) δ 161.6, 156.9, 149.0, 145.3, 141.8, 139.5, 138.0, 127.2, 126.6, 126.2, 126.0, 125.8, 125.5, 123.0, 121.3, 114.7, 114.2, 111.7, 98.9, 84.5, 78.9, 71.6, 68.8, 61.7, 28.8, 22.6, 8.1; IR (film) νmax
3414, 3381, 2932, 1607, 1528, 1510, 1260, 1084 cm⁻¹; HRMS (ESI+) m/z 526.1532 (M + H⁺, C25H27NO10 requires m/z 526.1536).

(45). Colorless solid (39%, 2 steps): [α]D26 = -15.9° (c = 0.27, 20% MeOH in CH2Cl2);
1H NMR (500 MHz, 20% CD3OD in CD2Cl2) δ 8.64 (s, 1H), 7.39 (d, J = 8.0 Hz, IH), 7.31 (s, IH), 7.30 (d, J = 8.0 Hz, IH), 7.14 (d, J = 8.5 Hz, IH), 6.84 (d, J = 8.5 Hz, IH), 5.99 (s, 2H), 5.48 (s, IH), 4.08 (d, J = 9.5 Hz, IH), 4.04 (d, J = 2.0 Hz, IH), 3.48 (s, 3H), 3.25 (d, J = 9.5 Hz, IH), 2.20 (s, 3H), 1.22 (s, 3H), 1.12 (s, 3H); 13C NMR (125 MHz, 20% CD3OD in CD2Cl2) δ 164.7, 158.7, 155.7, 150.6, 148.4, 147.7, 127.2, 125.1, 123.9, 121.5, 121.0, 113.5, 113.2, 110.6, 107.5, 106.8, 101.5, 97.7, 83.5, 77.9, 70.5, 67.8, 60.8, 27.9, 21.6, 7.2; IR (film) νmax
3404, 3111, 3035, 2980, 2926, 2853, 1701, 1607, 1528, 1485, 1444, 1406, 1369, 1254, 1132, 1113, 1086, 1038 cm⁻¹; HRMS (ESI+) m/z
514.1708 (M + H⁺, C26H26N2O10 requires m/z 514.1713).
[0303] 3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-1  H-indole-2-carboxamide (46).

Yellow solid (31%, 2 steps): [α]D 25 = -18.2° (c = 0.22, 20% MeOH in CH2Cl2); 1H NMR (800 MHz, 20% CD3OD in CD2Cl2) δ 8.69 (s, IH), 7.66 (d, J = 8.0 Hz, IH), 7.46 (d, J = 8.0 Hz, IH), 7.36 (d, J = 9.0 Hz, IH), 7.27 (t, J = 7.5 Hz, IH), 7.20 (d, J = 9.0 Hz, IH), 7.17 (s, IH), 7.10 (t, J = 7.5 Hz, IH), 5.53 (d, J = 2.0 Hz, IH), 4.17 - 4.08 (m, 2H), 3.55 (s, 3H), 3.31 (d, J = 9.5 Hz, IH), 2.26 (s, 3H), 1.30 (s, 3H), 1.08 (s, 3H); 13C NMR (200 MHz, 20% CD3OD in CD2Cl2) δ 162.1, 161.0, 158.0, 150.9, 139.0, 131.8, 129.1, 127.3, 126.6 (2C), 123.7, 123.1, 122.3, 115.8, 115.5, 113.7, 112.9, 106.9, 100.2, 85.8, 80.2, 72.9, 70.1, 63.1, 30.1, 23.9, 9.4; IR (film) νmax 3443, 3421, 3404, 3003, 2986, 2935, 1609, 1541, 1364, 1263, 1105, 1082 cm−1; HRMS (ESI+) m/z 509.1916 (M + H+), C27H29N2O8 requires m/z 509.1924.

[0304] 3,4-Dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-1  H-indole-3-carboxamide (47).

Colorless solid (19%, 2 steps): [α]D 25 = -11.4° (c = 0.18, 20% MeOH in CH2Cl2); 1H NMR (500 MHz, 20% CD3OD in CD2Cl2) δ 8.75 (s, IH), 8.12 (dd, J = 2.0, 6.5 Hz, IH), 7.97 (s, IH), 7.49 (dd, J = 2.0, 6.5 Hz, IH), 7.38 (d, J = 8.5 Hz, IH), 7.31 - 7.24 (m, 2H), 7.20 (d, J = 8.5 Hz, IH), 5.55 (d, J = 2.5 Hz, IH), 4.16 (dd, J = 4.5, 9.5 Hz, IH), 4.12 (t, J = 4.5 Hz, IH), 3.57 (s, 3H), 3.33 - 3.31 (m, IH), 2.28 (s, 3H), 1.33 (s, 3H), 1.10 (s, 3H); 13C NMR (125 MHz, 20% CD3OD in CD2Cl2) δ 165.7, 161.3, 157.6, 150.6, 138.4, 131.0, 127.1, 126.4, 125.1, 124.5, 123.8, 123.5, 121.4, 115.8 (2C), 113.9, 112.8, 112.7, 100.0, 85.9, 80.1, 72.9, 70.1, 63.2, 30.2, 23.9, 9.6; IR (film) νmax 3439, 3418, 3394, 2957, 2924, 2853, 1636, 1529, 1437, 1379, 1261, 1180, 1128, 1082, 1020 cm−1; HRMS (ESI+) m/z 509.1924 (M + H+), C27H29N2O8 requires m/z 509.1924.
EXAMPLE 29: ANTIPROLIFERATIVE ACTIVITIES OF HETEROCYCLE NOVOBIOCIN DERIVATIVES

[0305] In this example, the inhibitory values were obtained by evaluation against the panel of cancer cell lines in Example 24. As shown in Table 5, in the nicotinic (40-42) series, the o-analogue was found to represent the most active regioisomer, consistent with the trend observed for the nitro (15-17) and aniline (18-20) derivatives. Likewise, benzofuran 43 and benzothiophene 44 were examined. It was postulated that because these structures contain a hydrogen bond acceptor in the same proximity as the other *ortho* derivatives they would therefore introduce a rigid two carbon spacer between the amide and the phenyl ring. Unfortunately, benzofuran (43) was less active than the *o*-OMe variant (9), and the bioisoelectronic benzothiophene (44) was even worse. The 2-indole was included for its potential to provide a hydrogen bond donor as was observed for o-anilines. Anti-proliferation activity was significantly increased against the majority of the cell lines indicating that the does indeed properly account for the activities observed for a *trans* two-carbon tether and an o-aniline. Substitution of the benzamide with a 2-indoleamide increased the activity greater than 500 fold against SkBr3 cells when compared to the natural product, novobiocin. Interestingly, the position of the nitrogen on the indole is critical for activity, as the 3-indoleamide (47) was approximately 3-10 times less effective than indoleamide compound 46. A summary of the observed trends for the indoleamine compound 46 is provided in FIG. 16.

Table 5. Anti-proliferation Activities of Novobiocin Heterocyclic Analogue

Reported in µM (n = 3).

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<th>PL45</th>
<th>LNCaP</th>
<th>PC-3</th>
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<td>28.4 ± 2.0</td>
<td>20.3 ± 4.1</td>
<td>23.3 ± 5.9</td>
<td>7.9 ± 0.7</td>
<td>44.9 ± 31.1</td>
<td>15.1 ± 2.1</td>
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<tr>
<td>41</td>
<td>65.9 ± 15.8</td>
<td>47.8 ± 4.2</td>
<td>37.3 ± 1.0</td>
<td>13.2 ± 1.0</td>
<td>50.1 ± 3.3</td>
<td>67.0 ± 20.2</td>
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<td>&gt;100</td>
<td>&gt;100</td>
<td>19.2 ± 2.5</td>
<td>34.8 ± 16.0</td>
<td>N/T</td>
<td></td>
</tr>
<tr>
<td>43</td>
<td>N/T</td>
<td>28.6 ± 4.1</td>
<td>9.6 ± 0.3</td>
<td>3.8 ± 0.4</td>
<td>7.9 ± 1.1</td>
<td>18.8 ± 4.4</td>
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<tr>
<td>44</td>
<td>N/T</td>
<td>96.3 ± 3.7</td>
<td>78.9 ± 3.5</td>
<td>33.2 ± 0.7</td>
<td>&gt;100</td>
<td>7.4 ± 2.1</td>
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<tr>
<td>45</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>5.2 ± 1.1</td>
<td>9.3 ± 0.6</td>
<td>10.9 ± 2.3</td>
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<tr>
<td>46</td>
<td>0.37 ± 0.06</td>
<td>0.57 ± 0.07</td>
<td>0.17 ± 0.01</td>
<td>0.47 ± 0.34</td>
<td>12.2 ± 0.0</td>
<td>22.3 ± 10.1</td>
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<tr>
<td>47</td>
<td>12.2 ± 1.5</td>
<td>5.3 ± 0.3</td>
<td>3.5 ± 0.5</td>
<td>1.8 ± 0.3</td>
<td>2.3 ± 0.1</td>
<td>4.8 ± 2.4</td>
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EXAMPLE 30: HSP90 INHIBITION OF 2-INDOLEAMIDE DERIVATIVE

In order to provide additional evidence that the growth inhibitory activity manifested by the 2-indoleamide compound 46 resulted from Hsp90 inhibition, 46 was evaluated by its ability to induce degradation of Hsp90-dependent client proteins. As seen in FIG. 4, Hsp90 client proteins such as Her2, Raf and Akt were degraded in a concentration-dependent manner while Hsp90 was induced in the presence of 46. Since non-Hsp90-dependent substrates such as actin remained unchanged, the inhibition of cell growth was directly correlated to the degradation of Hsp90-dependent client proteins.

EXAMPLE 31: SYNTHESES OF 5-, 6-, AND 8-ALKYL(OXY) RESORCINOL PRECURSORS FOR NOVOBIOCIN ANALOGUES.

In the following example, modifications at the 5-, 6-, and 8-position of the coumarin ring were made in order to mimic those at the 6-, 7-, and 3-position of guanine. Various resorcinol precursors were therefore made.

To generate the resorcinol precursors with substitutions at the 4-position, which result in coumarin ring systems with appendages at the 6-position, the phenols of benzaldehyde 1 were protected as the corresponding ethers as shown in the scheme below. The resulting benzaldehydes (2a-b) (Nabaei-Bidhendi et al., Convenient synthesis of polyhydroxyflavonoids, J. Indian Chem. Soc. 67 43-45 (1990)) were converted to their formate esters via Baeyer-Villiger oxidation, and then hydrolyzed to afford phenols 3a-b. See Horvath, R. F.; Chan, T. H. J. Org. Chem. 52 4489-4494 (1986); Miyake et al., Synthesis and Biological Activity of Arthrographol and Related Compounds, Heterocycles 43 665-674 (1996). O-alkylation with the requisite alkyl iodide proceeded in good yield and generated a series of protected 4-substituted resorcinolic ethers (4a-c). Ortho-lithidXion of 4a-c, followed by alkylation with methyl iodide provided the 2-methyl protected resorcinols, 5a-c. See Carreno, M. C.; Garcia Ruano, J. L.; Toledo, M. A.; Urbano, A. Tetrahedron: Asymmetry 8 913-921 (1997). Deprotection of the alkoxy ethers by exposure to acidic conditions gave resorcinols 6a-c. See Wang, Y.; Tan, W.; Li, W. Z.; Li, Y. J. Nat. Prod. 64 196-199 (2001).
wherein EOM = CH$_2$OEt.

[0309] 2,4-Bis(ethoxymethoxy)benzaldehyde (2a): N$_2$N-Diisopropylethylamine (25.3 mL, 145 mmol) was slowly added to 2,4-dihydroxybenzaldehyde (5.00 g, 36.2 mmol) in anhydrous N$_2$N-dimethylformamide (100 mL) over five minutes at room temperature. After 30 minutes, the solution was cooled to 0 °C and chloromethyl ethyl ether (14.2 mL, 145 mmol) was added and the mixture warmed to room temperature over 12 hours. The reaction was quenched by the addition of saturated aqueous NH$_4$Cl solution and extracted with EtOAc (3 x 50 mL). The combined organic fractions were washed with saturated aqueous NaCl, dried (Na$_2$SO$_4$), filtered, and concentrated. The residue was purified via column chromatography (SiO$_2$, 5:1 → 1:1 Hexane:EtOAc) to give 2a as a brown amorphous solid (9.10 g, 99%): $^1$H NMR (CDCl$_3$, 400 MHz) $\delta$ 10.34 (d, $J = 2.4$ Hz, IH), 7.81 (dd, $J = 8.7$, 2.8 Hz, IH), 6.89 (t, $J = 2.5$ Hz, IH), 6.74 (m, IH), 5.34 (d, $J = 2.8$, 2H), 5.28 (d, $J = 2.8$, 2H), 3.81-3.71 (m, 4H), 1.28-1.22 (m, 6H).

[0310] 2,4-Bis(ethoxymethoxy)phenol (3a): A solution of 2a (3.78 g, 12.0 mmol) in anhydrous CH$_2$Cl$_2$ (4.0 mL) was slowly added to mCPBA (70%) (3.26 g, 13.2 mmol) in anhydrous CH$_2$Cl$_2$ (16.3 mL) at 0 °C. The resulting solution was warmed to room temperature, then refluxed for 12 hours. After cooling to room temperature, the resulting solution was washed with saturated aqueous NaHCO$_3$ solution (3 x 20 mL) and 10% aqueous Na$_2$S$_2$O$_3$ (30 mL). Combined organic fractions were dried (Na$_2$SO$_4$), filtered, and concentrated. The residue was re-dissolved in MeOH (5 mL) and stirred with excess
10% aqueous NaOH for three hours at room temperature. The pH was adjusted to 2 with
6 M HCl and the solution was extracted with CH₂Cl₂ (3 x 10 mL). Combined organic
fractions were dried (Na₂SO₄), filtered, and concentrated to give 3a as an orange oil
(8.21 g, 94%): ¹H NMR (CDCl₃, 500 MHz) δ 6.89-6.85 (m, 2H), 6.67 (dd, J = 8.8, 2.7
Hz, 1H), 5.81 (d, J = 6.6 Hz, IH), 5.23 (s, 2H), 5.15 (s, 2H), 3.80-3.73 (m, 4H), 1.29-
1.24 (m, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 151.0, 145.0, 141.5, 115.2, 110.6, 106.0,
94.9, 94.2, 64.8, 64.1, 15.1, 15.1; IR (film) v_max 3362, 2887, 1460, 1286, 1162, 735 cm⁻¹;
HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₁₃H₂₀O₅, 279.1208; found, 279.1181.

[0311] 2,4-Bis(methoxymethoxy)phenol (3b): Benzaldehyde 2b (700 mg,
3.11 mmol) in CHCl₃ (1.80 mL) at 0 °C was treated with mCPBA (70% w/w, 1.61 g,
9.33 mmol). After 10 minutes, the solution was warmed to room temperature, then
refluxed for 12 hours. Upon cooling to room temperature, the solution was washed with
saturated aqueous NaHCO₃ (3 x 10 mL), saturated aqueous Na₂SO₃ (20 mL), saturated
aqueous NaCl, was dried (Na₂SO₄), filtered, and concentrated. The residue was
dissolved in MeOH (5 mL) and stirred with excess triethylamine for three hours at room
temperature. The solvent was concentrated and the residue purified by column
chromatography (SiO₂, 4:1 → 3:1 Hexane:EtOAc) to afford 3b as a yellow oil (320 mg,
50%): ¹H NMR (CDCl₃, 400 MHz) δ 6.87 (d, J = 8.9 Hz, IH), 6.86 (s, IH), 6.67 (dd, J =
11.5, 2.8 Hz, IH), 5.21 (s, 2H), 5.11 (s, 2H), 3.54 (s, 3H), 3.50 (s, 3H).

[0312] 2,4-Bis(ethoxymethoxy)-1-methoxybenzene (4a): Potassium carbonate
(14.3 g, 103 mmol) was added to 3a (2.50 g, 10.3 mmol) in N,N-dimethylformamide
(103 mL). After 10 minutes, methyl iodide (6.43 mL, 103 mmol) was added and the
solution was heated to reflux for 12 hours. Upon cooling to room temperature, the
solution was extracted with EtOAc (3 x 50 mL); combined organic fractions were
washed with saturated aqueous NaCl, dried (Na₂SO₄), and concentrated. The residue
was purified by column chromatography (SiO₂, 4:1 Hexane:EtOAc) to afford 4a as a yellow oil (2.40 g, 91%): ¹H NMR (CDCl₃, 500 MHz) δ 6.87 (d, J = 2.8 Hz, IH), 6.72
(d, J = 8.9 Hz, IH), 6.60 (dd, J = 13.3, 1.7 Hz, IH), 5.18 (s, 2H), 5.07 (s, 2H), 3.76 (s,
3H), 3.72-3.69 (m, 2H), 3.68-3.63 (m, 2H), 1.17-1.13 (m, 6H); ¹³C NMR (CDCl₃, 125
MHz) δ 150.7, 146.2, 143.9, 111.2, 107.9, 105.8, 93.2, 93.0, 63.3, 63.0, 55.4, 14.1, 14.0;
IR (film) v_max 2976, 2932, 2899, 2835, 1595, 1508, 1393, 1227, 1153, 1103, 1080, 1009,
847 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₁₃H₂₀O₅, 279.1208; found, 279.1181.
2,4-Bis(methoxymethoxy)-1-propoxybenzene (4b): Potassium carbonate (322 mg, 2.33 mmol) was added to 3b (50 mg, 0.233 mmol) in N,N-dimethylformamide (2.33 mL) at room temperature. After 10 minutes, iodopropane (226 µL, 2.33 mmol) was added and the solution was heated to reflux for 12 hours. Upon cooling to room temperature, the solution was extracted with EtOAc (3 x 10 mL); combined organic fractions were washed with saturated aqueous NaCl, dried (Na₂SO₄), filtered, and concentrated. The residue was purified by column chromatography (SiO₂, 5:1 Hexane:EtOAc) to afford 4b as a yellow oil (36.4 mg, 61%): ¹H NMR (CD₂Cl₂, 400 MHz) δ 6.87 (s, 1H), 6.84 (d, J = 2.9 Hz, 1H), 6.68 (dd, J = 11.7, 2.8 Hz, 1H), 5.19 (s, 2H), 5.12 (s, 2H), 3.93 (t, J = 6.6 Hz, 2H), 3.53 (s, 3H), 3.49 (s, 3H), 1.86-1.78 (m, 2H), 1.06 (t, J = 7.5 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 150.6, 146.5, 143.8, 113.7, 108.5, 106.6, 94.7, 94.2, 70.3, 55.2, 54.9, 21.6, 9.5; IR (film) νmax 2961, 2826, 1595, 1506, 1400, 1261, 1154, 1013, 1076, 924, 800 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₁₂H₁₀O₅, 257.1389; found, 257.1410; [M + Na]⁺ calced for C₁₂H₁₀O₅, 279.1208; found, 279.1165.

2,4-Bis(ethoxymethoxy)-1-isopropoxybenzene (4c): Potassium carbonate (2.85 g, 20.7 mmol) was added to 3a (500 mg, 2.07 mmol) in N,N-dimethylformamide (4.10 mL) at room temperature. After 10 minutes, 2-iodopropane (2.06 mL, 20.7 mmol) was added and the solution was heated to reflux for 12 hours. Upon cooling to room temperature, the solution was extracted with EtOAc (3 x 20 mL); combined organic fractions were washed with saturated aqueous NaCl, dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 5:1 → 1:1 Hexane:EtOAc) to afford 4c as a yellow oil (0.32 g, 55%): ¹H NMR (CD₂Cl₂, 400 MHz) δ 6.87 (s, 1H), 6.86 (d, J = 4.9 Hz, 1H), 6.66 (dd, J = 11.6, 3.4 Hz, 1H), 5.23 (s, 2H), 5.17 (s, 2H), 4.44-4.38 (m, 1H), 3.83-3.72 (m, 4H), 1.33 (s, 3H), 1.31 (s, 3H), 1.27-1.23 (m, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 152.4, 149.1, 143.2, 118.6, 109.5, 107.5, 94.4, 93.9, 72.8, 64.3, 64.1, 22.2 (2C), 15.1, 15.1; IR (film) νmax 2976, 1591, 1504, 1528, 1391, 1258, 1217, 1107, 1011, 847 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₁₃H₂₀O₅, 285.1702; found, 285.1746; [M + Na]⁺ calced for C₁₃H₂₀O₅, 307.1522; found, 307.1310.

1,3-Bis(ethoxymethoxy)-4-methoxy-2-methylbenzene (5a): A solution of 4a (632 mg, 2.27 mmol) in anhydrous THF (1.94 mL) was added dropwise to a solution of "BuLi (2.5 M in hexanes, 1.48 mL, 3.70 mmol) in anhydrous THF (1.62 mL) at room temperature. After one hour, the solution was cooled to -78 °C and methyl iodide
(620 µL, 9.87 mmol) was added. The resulting solution was warmed to room temperature over 12 hours, and the reaction was quenched by the addition of saturated aqueous NH₄Cl. Water (5 mL) was added and the solution was extracted with CH₂Cl₂ (3 x 10 mL). Combined organic fractions were dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 8:1 → 5:1 Hexane:EtOAc) to afford 5a as a yellow oil (353 mg, 53%): ¹H NMR (CDCl₃, 500 MHz) δ 6.74 (d, J = 9.0 Hz, IH), 6.60 (d, J = 9.0 Hz, IH), 5.10 (s, 2H), 5.05 (s, 2H), 3.78 (q, J = 7.1 Hz, 2H), 3.72 (s, 3H), 3.67 (q, J = 7.1 Hz, 2H), 2.14 (s, 3H), 1.18-1.15 (m, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 149.1, 146.5, 121.7, 109.0, 108.4, 96.2, 93.0, 64.3, 63.1, 55.1, 28.7, 14.2, 14.1, 8.8; IR (film) νmax 2918, 2359, 1487, 1260, 1248, 1082, 1055, 945, 798 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₁₄H₂₁O₅, 293.1365; found, 293.1357.

[0316] 1,3-Bis(methoxymethoxy)-2-methyl-4-propoxybenzene (5b): A solution of 4b (165 mg, 0.64 mmol) in anhydrous THF (520 µL) was added dropwise to a solution of "BuLi (2.5 M in hexanes, 390 µL, 0.97 mmol) in anhydrous THF (420 µL) at room temperature. After 1 hour, the solution was cooled to -78 °C and methyl iodide (160 µL, 2.58 mmol) was added. The resulting solution was warmed to room temperature over 12 hours, and the reaction was quenched by the addition of saturated aqueous NH₄Cl. Water (5 mL) was added and the solution was extracted with CH₂Cl₂ (3 x 10 mL). Combined organic fractions were dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 6:1 Hexane:EtOAc) to afford 5b as a yellow oil (166 mg, 95%): ¹H NMR (CDCl₃, 500 MHz) δ 6.66 (d, J = 9.0 Hz, IH), 6.60 (d, J = 9.0 Hz, IH), 5.02 (s, 2H), 5.00 (s, 2H), 3.80-3.77 (m, 2H), 3.49 (s, 3H), 3.47 (s, 3H), 2.14 (d, J = 7.1 Hz, 3H), 1.73-1.69 (m, 2H), 0.94 (t, J = 7.5 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 148.5, 148.5, 147.3, 145.6, 126.7, 123.0, 112.8, 110.8, 110.4, 99.2, 57.7, 57.6, 21.2, 10.9, 10.0; IR (film) νmax 2957, 2924, 2853, 1738, 1597, 1487, 1468, 1391, 1335, 1231, 1157, 974, 798 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₁₄H₂₁O₅, 271.1545; found, 271.1558.

[0317] 1,3-Bis(ethoxymethoxy)-4-isopropoxy-2-methylbenzene (5c): A solution of 4c (190 mg, 0.67 mmol) in anhydrous THF (530 µL) was added dropwise to a solution of "BuLi (2.5 M in hexanes, 410 µL, 1.00 mmol) in anhydrous THF (440 µL) at room temperature. After one hour, the solution was cooled to -78 °C and methyl iodide (170 µL, 2.67 mmol) was added. The resulting solution was warmed to room temperature over 12 hours, and the reaction was quenched by the addition of saturated aqueous NH₄Cl. Water (5 mL) was added and the solution was extracted with CH₂Cl₂ (3
Combined organic fractions were dried (Na$_2$SO$_4$), filtered, and concentrated. The residue was purified via column chromatography (SiO$_2$, 6:1 Hexane:EtOAc) to afford 5c as a yellow oil (157 mg, 79%): $^1$H NMR (CDCl$_3$, 500 MHz) $\delta$ 6.70 (d, $J = 9.0$ Hz, 1H), 6.61 (d, $J = 9.0$ Hz, IH), 5.10 (s, 2H), 5.08 (s, 2H), 4.34 (quintet, $J = 6.1$ Hz, IH), 3.78 (q, $J = 7.1$ Hz, 2H), 3.67 (q, $J = 7.1$ Hz, 2H), 2.13 (s, 3H), 1.23 (d, $J = 6.1$ Hz, 6H), 1.24-1.15 (m, 6H); $^{13}$C NMR (CDCl$_3$, 125 MHz) $\delta$ 150.3, 146.6, 145.3, 122.7, 113.7, 110.1, 97.3, 94.0, 71.5, 65.4, 64.2, 29.4, 22.2, 15.2, 15.2, 9.9; IR (film) $\nu_{\text{max}}$ 2924, 2853, 2359, 2339, 1951, 1483, 1113, 1057, 974 cm$^{-1}$; HRMS (ESI$^+$) m/z: [M + H]$^+$ calcd for C$_6$H$_{10}$O$_3$, 299.1858; found, 299.1909.

4-Methoxy-2-methylbenzene-1,3-diol (6a): A solution of 5a (910 mg, 3.37 mmol) in MeOH (28.0 mL) at room temperature was treated dropwise with 3 M HCl (9.00 mL, 26.9 mmol), then heated to reflux for one hour. Water (30 mL) was added and the solution was extracted with EtOAc (3 x 30 mL). Combined organic fractions were washed with saturated aqueous NaCl, dried (Na$_2$SO$_4$), filtered, and concentrated. The residue was purified via column chromatography (SiO$_2$, 6:1 Hexane:EtOAc) to afford 6a as a red amorphous solid (509 mg, 98%): $^1$H NMR (Acetone-$d_6$, 500 MHz) $\delta$ 7.68 (s, IH), 7.24 (s, IH), 6.60 (d, $J = 11$ Hz, IH), 6.29 (d, $J = 11$ Hz, IH), 3.74 (s, 3H), 2.09 (s, 3H); $^{13}$C NMR (CDCl$_3$, 125 MHz) $\delta$ 144.7, 142.1, 139.7, 115.6, 110.2, 108.6, 55.6, 7.6; IR (film) $\nu_{\text{max}}$ 3583, 2920, 2359, 1951, 1416, 1259, 1090, 1020, 798 cm$^{-1}$.

2-Methyl-4-propoxybenzene-1,3-diol (6b): A solution of 5b (580 mg, 2.15 mmol) in MeOH (17.9 mL) was treated dropwise with 3 M HCl (630 mL, 17.2 mmol), then heated to reflux for one hour. Water (20 mL) was added and the solution was extracted with EtOAc (3 x 20 mL). Combined organic fractions were washed with saturated aqueous NaCl, dried (Na$_2$SO$_4$), and concentrated to afford 6b as a red amorphous solid (387 mg, 99%). $^1$H NMR (CDCl$_3$, 500 MHz) $\delta$ 6.51 (d, $J = 8.7$ Hz, IH), 6.21 (d, $J = 8.6$ Hz, IH), 5.74 (s, IH), 4.36 (s, IH), 3.87-3.85 (m, 2H), 2.09 (s, 3H), 1.75-1.71 (m, 2H), 0.96 (t, $J = 7.5$ Hz, 3H); $^{13}$C NMR (CDCl$_3$, 125 MHz) $\delta$ 147.5, 143.8, 139.0, 109.8, 108.4, 103.8, 70.2, 21.6, 9.5, 7.3; IR (film) $\nu_{\text{max}}$ 3520, 3360, 2966, 2880, 2359, 2341, 1636, 1236, 1068, 785, 750 cm$^{-1}$.

4-Methoxybenzene-1,3-diol (6c): A solution of 5c (157 mg, 0.53 mmol) in MeOH (4.40 mL) at room temperature was treated dropwise with 3 M HCl (1.40 mL, 4.21 mmol), then heated to reflux for one hour. Water (5 mL) was added and the solution was extracted with EtOAc (3 x 10 mL). Combined organic fractions were...
washed with saturated aqueous NaCl, dried (Na₂SO₄), filtered, and concentrated to afford
6c as a red amorphous solid (95 mg, 99%): \(^1H\) NMR (CDCl₃, 500 MHz) \(\delta 6.54\) (d, \(J = 8.7\) Hz, IH), 6.21 (d, \(J = 8.7\) Hz, IH), 5.78 (s, IH), 4.37-4.32 (m, IH), 2.09 (s, 3H), 1.25 (d, \(J = 6.1\) Hz, 6H); \(^13C\) NMR (CDCl₃, 125 MHz) \(\delta 147.7, 144.9, 137.5, 110.8, 109.8, 104.0, 71.7, 21.3\) (2C); IR (film) \(v_{\text{max}}\) 3526, 2974, 2924, 2853, 1717, 1607, 1475, 1238, 1113, 1067, 928, 887, 791 cm\(^{-1}\).

[0321] To generate resorcinol precursors with substitutions at the 5-position, the phenols of 5-methoxy resorcinol 7 were once again protected as the corresponding alkoxy ethers, 8, as shown in the scheme below. Ortho-lithiation of 8, followed by treatment with methyl iodide, led to installation of a methyl group at the 2-position of 9. See Carreno, M. C.; Garcia Ruano, J. L.; Toledo, M. A.; Urbano, A. Tetrahedron: Asymmetry 8 913-921 (1997). Acidic deprotection was employed to afford resorcinol


[0322] 1-Methoxy-3,5-bis(methoxymethoxy)benzene (8): N,N-diisopropylethylamine (3.15 mL, 18.1 mmol) was added to 5-methoxybenzene-l,3-diol (634 mg, 4.52 mmol) in anhydrous \(\Lambda,\Lambda\)-dimethylformamide (12.6 mL) over five minutes at room temperature. After 30 minutes, the solution was cooled to 0 °C, methoxy methylchloride (3.02 mL, 18.1 mmol) was added, and the solution was warmed to room temperature over 12 hours. The reaction was quenched by the addition of saturated aqueous NaHCO₃ at 0 °C and extracted with EtOAc (3 x 10 mL). Combined organic fractions were washed with saturated aqueous NaCl, dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 6:1—>4:1 Hexane:EtOAc) to afford 8 as a yellow amorphous solid (441 mg, 43%): \(^1H\) NMR (CDCl₃, 500 MHz) \(\delta 6.29\) (t, \(J = 2.2\) Hz, IH), 6.21 (d, \(J = 2.2\) Hz, 2H), 5.07 (s, 4H), 3.69 (s, 3H), 3.40 (s, 6H); \(^13C\) NMR (CDCl₃, 125 MHz) \(\delta 161.394, 159.0\) (2C), 97.2, 96.2

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(2C), 94.5 (2C), 56.1, 55.4 (2C); IR (film) ν_{max} 2997, 2955, 2903, 2827, 1601, 1475, 1400, 1215, 1194, 1146, 1032, 991, 924, 829, 685 cm^{-1}; HRMS (ESI\(^{+}\)) m/z: [M + Na]\(^{+}\) calcd for C\(_{9}\)H\(_{6}\)O\(_{5}\), 251.0895; found, 251.0910.

[0323] 5-methoxy-1,3-bis(methoxymethoxy)-2-methylbenzene (9): A solution of 8 (441 mg, 1.93 mmol) in anhydrous THF (1.55 mL) was added dropwise to a solution of "BuLi (2.5 M in hexanes, 1.16 mL, 2.90 mmol) in anhydrous THF (1.26 mL) at room temperature. After one hour, the solution was cooled to -78 °C and methyl iodide (480 µL, 7.73 mmol) was added. The resulting solution was warmed to room temperature over 12 hours, and the reaction was quenched by the addition of saturated aqueous NH\(_{4}\)Cl. Water (5 mL) was added and the solution was extracted with CH\(_{2}\)Cl\(_{2}\) (3 x 10 mL). Combined organic fractions were dried (Na\(_{2}\)SO\(_{4}\)), filtered, and concentrated. The residue was purified via column chromatography (SiO\(_{2}\), 6:1 → 4:1; Hexane:EtOAc) to afford 9 as a yellow oil (314 mg, 67%): ^1\text{H} NMR (CDCl\(_{3}\), 500 MHz) δ 6.38 (d, J = 2.2 Hz, 1H), 6.24 (d, J = 2.1 Hz, 1H), 5.08 (d, J = 3.6 Hz, 2H), 5.06 (d, J = 2.6 Hz, 2H), 3.72 (s, 3H), 3.40 (s, 6H), 1.97 (s, 3H); ^13\text{C} NMR (CDCl\(_{3}\), 125 MHz) δ 160.3, 157.8, 155.4, 108.2, 93.9, 93.8, 93.7, 92.9, 55.0, 55.0, 54.6, 7.0; IR (film) ν_{max} 2953, 2934, 2905, 1597, 1497, 1396, 1215, 1144, 1126, 1074, 1059, 1028, 922, 822 cm^{-1}; HRMS (ESI\(^{+}\)) m/z: [M + H]\(^{+}\) calcd for C\(_{12}\)H\(_{18}\)O\(_{6}\), 243.1233; found, 243.1223.

[0324] 5-Methoxy-2-methylbenzene-1,3-diol (10): A solution of 9 (314 mg, 1.30 mmol) in MeOH (10.8 mL) at room temperature was treated dropwise with 3 M HCl (3.46 mL, 10.3 mmol), then heated to reflux for one hour. Water (11 mL) was added and the solution was extracted with EtOAc (3 x 15 mL). Combined organic fractions were washed with saturated aqueous NaCl, dried (Na\(_{2}\)SO\(_{4}\)), filtered, and concentrated to afford 10 as a red amorphous solid (177 mg, 99%): ^1\text{H} NMR (CDCl\(_{3}\), 500 MHz) δ 8.17 (s, 1H), 6.09 (d, J = 1.6 Hz, 1H), 6.04 (s, 1H), 3.67 (d, J = 9.9 Hz, 3H), 2.08 (d, J = 4.1 Hz, 3H); ^13\text{C} NMR (CDCl\(_{3}\), 125 MHz) δ 160.3, 157.8, 155.4, 108.4, 93.9, 93.8, 55.0, 7.0; IR (film) ν_{max} 3445, 2924, 2853, 2359, 2332, 1653, 1636, 1456, 1080, 1022, 798, 669 cm^{-1}; HRMS (ESI\(^{+}\)) m/z: [2M + H]\(^{+}\) calcd for C\(_{8}\)H\(_{16}\)O\(_{3}\), 309.1388 found, 309.1332.

[0325] To generate the resorcinol precursors with aryl substituents at the 2-position, the phenols of resorcinol 11 were protected as the corresponding alkoxy ethers, 12, as shown in the scheme below. Subsequent ort/zo-lithiation of 12, followed by the addition of benzyl bromide provided the benzyl derivative, 13. See Carreno, M. C.; Garcia Ruano, J. L.; Toledo, M. A.; Urbano, A. Tetrahedron: Asymmetry 8 913-921 (1997).

[0326] 1,3-Bis(methoxymethoxy)benzene (12): Sodium hydride (872 mg, 36.3 mmol) was added to resorcinol (1.00 g, 9.08 mmol) in anhydrous N,N-dimethylformamide (25.4 mL) at 0 °C. After 30 minutes, methoxy methylchlordride (2.76 mL, 36.3 mmol) was added and the resulting solution was warmed to room temperature over 12 hours. The reaction was cooled to 0 °C, quenched by the addition of saturated aqueous NaHCO3, and extracted with EtOAc (3 x 30 mL). Combined organic fractions were washed with saturated aqueous NaCl, dried (Na2SO4), filtered, and concentrated. The residue was purified via column chromatography (SiO2, 4:1 Hexane:EtOAc) to afford 12 as a yellow oil (1.75 g, 97%): 1H NMR (CDCl3, 400 MHz) δ 7.25-7.20 (m, 1H), 6.80 (d, J = 2.3 Hz, 1H), 6.75 (dd, J = 8.2, 2.4 Hz, 2H), 5.20 (s, 4H), 3.51 (s, 6H).
2-Benzyl-l,3-bis(methoxymethoxy)benzene (13): A solution of 12 (500 mg, 2.52 mmol) in anhydrous THF (2.02 mL) was added dropwise to a solution of "BuLi (2.5 M in hexanes, 1.51 mL, 3.78 mmol) in anhydrous THF (1.65 mL) at room temperature. After one hour, the solution was cooled to -40°C and benzyl bromide (1.22 mL, 10.10 mmol) was added. The resulting solution was warmed to room temperature over 12 hours, and the reaction was quenched by the addition of saturated aqueous NH₄Cl. Water (5 mL) was added and the solution was extracted with CH₂Cl₂ (3 x 10 mL). Combined organic fractions were dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 4:1; Hexane:EtOAc) to afford 13 as a yellow oil (214 mg, 30%): ¹H NMR (CDCl₃, 500 MHz) δ 7.17 (d, J = 7.9 Hz, 2H), 7.17-7.12 (m, 2H), 7.06-7.02 (m, 2H), 6.71 (d, J = 8.3 Hz, 2H), 5.09 (s, 4H), 4.00 (s, 2H), 3.29 (s, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 155.9 (2C), 141.6, 128.5 (2C), 128.0 (2C), 127.5, 125.4, 119.4, 107.7 (2C), 94.3 (2C), 56.0 (2C), 29.1; IR (Film) νₘₐₓ 2953, 2930, 1595, 1470, 1452, 1254, 1153, 1097, 1043, 941, 922, 727, 698 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₁₇H₂₀O₄, 311.1259; found, 311.1201.

2-Benzylbenzene-1,3-diol (14): A solution of 13 (214 mg, 0.74 mmol) in MeOH (6.20 mL) was treated dropwise with 3 M HCl (0.22 mL, 5.92 mmol), then heated to reflux for one hour. Water (10 mL) was added and the solution was extracted with EtOAc (3 x 15 mL). Combined organic fractions were washed with saturated aqueous NaCl, dried (Na₂SO₄), and concentrated to afford 14 as a red amorphous solid (149 mg, 99%). ¹H NMR (CDCl₃, 400 MHz) δ 7.31 (d, J = 6.6 Hz, 4H), 7.25-7.19 (m, 1H), 7.01 (t, J = 8.1 Hz, IH), 6.44 (d, J = 8.1 Hz, 2H), 4.82 (s, 2H), 4.09 (s, 2H).

2-Iodo-l,3-bis(methoxymethoxy)benzene (15): n-Butyllithium (2.5 M in hexanes, 0.22 mL, 0.56 mmol) was added to a solution of 12 (100 mg, 0.50 mmol) in anhydrous THF (790 µL) at 0°C. After 5 minutes, iodine (141 mg, 0.56 mmol) in anhydrous THF (320 µL) was added. After two hours at room temperature, the reaction was quenched via dropwise addition of MeOH and the solvent was concentrated. Water (5 mL) was added and the solution was extracted with EtOAc (3 x 10 mL). Combined organics were washed with saturated aqueous Na₂S₂O₃, saturated aqueous NaCl, dried (Na₂SO₄), filtered, and concentrated to afford 15 as a brown oil (129 mg, 79%): ¹H NMR (CDCl₃, 100 MHz) δ 7.25-7.18 (m, 1H), 6.79-6.71 (m, 2H), 5.27 (s, 2H), 5.18 (s, 2H), 3.54 (s, 3H), 3.50 (s, 3H); IR (film) νₘₐₓ 2953, 2924, 2853, 1458, 1377 cm⁻¹.

2,6-Bis(methoxymethoxy)biphenyl (16): Anhydrous toluene (2.0 mL) was added to a flask charged with Pd₂(dba)₃ (56.3 mg, 0.062 mmol), dicyclohexyl(2',6'-
dimethoxybiphenyl-2-yl)phosphine (50.5 mg, 0.12 mmol), phenylboronic acid (281 mg, 2.31 mmol), and potassium phosphate (979 mg, 4.61 mmol) at room temperature. After 15 minutes, a solution of 15 (500 mg, 1.54 mmol) in anhydrous toluene (1.0 mL) was added and the resulting solution was heated to reflux for 12 hours. Upon cooling to room temperature, ether was added, the solution was filtered through SiO₂ and concentrated to give 16 as a colorless amorphous solid (418 mg, 99%): ¹H NMR (CDCl₃, 500 MHz) δ 7.35-7.28 (m, 2H), 7.28-7.25 (m, 2H), 7.18-7.15 (m, 2H), 6.83 (d, J = 8.3 Hz, 2H), 4.96 (s, 4H), 3.24 (s, 6H); ¹³C NMR (CDCl₃, 125 MHz) δ 155.3, 155.0, 134.3, 130.8, 129.5, 128.7, 128.0, 127.6, 126.8, 122.6, 109.4 (2C), 94.9 (2C), 56.0 (2C); IR (film) νmax 2955, 2928, 2359, 2341, 1587, 1466, 1439, 1400, 1244, 1153, 1099, 1080, 1041, 922, 764, 733, 700 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₁₆H₁₈O₄Na, 297.1 103; found, 297.1052.

[0331] Biphenyl-2,6-diol (17): A solution of 16 (400 mg, 1.46 mmol) in MeOH (12.0 mL) at room temperature was treated dropwise with 3 M HCl (430 µL, 11.7 mmol), then heated to reflux for one hour. Water (15 mL) was added and the solution was extracted with EtOAc (3 x 20 mL). Combined organic fractions were washed with saturated aqueous NaCl, dried (Na₂SO₄), filtered, and concentrated to afford 17 as an orange amorphous solid (269 mg, 99%): ¹H NMR (CDCl₃, 400 MHz) δ 7.60 (d, J = 7.6 Hz, 2H), 7.53-7.49 (m, 1H), 7.46-7.44 (m, 2H), 7.18 (t, J = 8.2 Hz, 1H), 6.62 (d, J = 8.2 Hz, 2H), 4.84 (s, 1H), 4.83 (s, 1H).

[0332] To generate resorcinol precursors with alkyl substitutions at the 2-position, pyragallol (18) was O-alkylated with methyl iodide to generate 2-methoxy resorcinol amongst an inseparable mixture of regioisomers as shown in the scheme below. The mixture was subsequently subjected to coumarin formation and the corresponding products isolated. Preparation of 2-ethyl resorcinol (21) from 2,6-dihydroxyacetophenone (20) was accomplished according to published procedures. See Elliger, C. A. Synth. Commun. 15 1315-1324 (1985). The following scheme shows the synthesis of 2-methoxy resorcinol and 2-ethyl resorcinol.
2-Methoxybenzene-1,3-diol (19): Lithium carbonate (281 mg, 1.98 mmol) was added to pyrogallol (100 mg, 0.79 mmol) in N,N-dimethylformamide (3.0 mL) at room temperature. After five minutes, methyl iodide (130 µL, 1.98 mmol) was added and the resulting solution was heated to 50 °C for 48 hours. Upon cooling to room temperature, water (20 mL) was added and the solution was extracted with EtOAc (3 x 20 mL). Combined organic fractions were dried (Na₂ SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 5:1 → 1:1 Hexane:EtOAc) to afford 19 as a colorless amorphous solid (44.2 mg, 34%): ¹H NMR (CDCl₃, 400 MHz) δ 6.89 (td, J = 8.0, 0.9 Hz, H), 6.53 (dd, J = 8.2, 0.8 Hz, 2H), 5.83 (bs, 2H), 3.90 (s, 3H).

EXAMPLE 32: PREPARATION OF 5-, 6-,
AND 8-MODIFIED NOVOBIOCIN ANALOGUES

In this example, 5-, 6-, and 8-modified novobiocin analogues were prepared. More specifically, once resorcinols 6a-c, 10, 14, 17, 19, and 21 were obtained from Example 31, the corresponding coumarins 23a-h were synthesized through a modified Pechmann condensation with enecamine 22 as previously described. See Robinson, A. J.; Lim, C. Y.; He, L.; Ma, P.; Li, H.-Y. J. Org. Chem. 66 4141-4147 (2001); Toplak, R.; Svete, J.; Stanovnik, B.; Grdadolnik, S. G. J. Hetero. Chem. 36 225-235 (1999). The resulting coumarin phenols were noviosylated with the trichloroacetimidate of noviose cyclic carbonate (24) in the presence of catalytic boron trifluoride etherate to generate scaffolds 25a-h in good yield. See Shen, G.; Yu, X. M.; Blagg, B. S. J. Bioorg. Med. Chem. Lett. 14 5903-5906 (2004). The benzyl carbonate was removed via hydrogenolysis to produce the aminocoumarin, which was readily coupled with preselected benzoic acids in the presence of N-(3-dimethylamino-propyl)- N-ethylcarbodiimide hydrochloride (EDCI) and pyridine. Benzoic acids were chosen based on previously determined SAR trends reported by Burlison and co-workers. See
Burlison, J. A.; Avila, C.; Vielhauer, G.; Lubbers, D. J.; Holzbeierlein, J.; Blagg, B. S. J.
J. Org. Chem. 73 2130-2137 (2008). The cyclic carbonates were treated with triethylamine in methanol to give the solvolyzed products, \textit{26a-p} in moderate to good yield over three steps.

6a-c, 10, 14,

17, 19, 21

23a-23h

25a-25h

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25g n/a H H OMe
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26a biaryl H OMe Me
26b biaryl H OPr Me
26c biaryl H O'Pr Me
26d biaryl OMe H Me
26e biaryl H H Bn
26f biaryl H H Ph
26g biaryl H H OMe
26h biaryl H H Et
26i 2-indole H OMe Me
26j 2-indole H OPr Me
26k 2-indole H O'Pr Me
26l 2-indole OMe H Me
26m 2-indole H H Bn
26n 2-indole H H Ph
26o 2-indole H H OMe
26p 2-indole H H Et

wherein the 22, 24, biaryl, and 2-indole are:

[0335] Benzyl 7-hydroxy-6-methoxy-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (23a): A solution of 6a (183 mg, 1.19 mmol) and enamine 22 (331 mg, 1.19 mmol) in glacial acetic acid (7.40 mL) was heated to reflux for 40 hours. Upon cooling to room
temperature, the solution was extracted with EtOAc (3 x 20 mL); combined organic fractions were dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 100:1 CH₂Cl₂:Acetone) to afford **23a** as a yellow amorphous solid (195 mg, 46%): 1H NMR (CDCl₃, 400 MHz) δ 8.27 (s, IH), 7.54 (s, IH), 7.43-7.37 (m, 4H), 6.77 (s, IH), 6.07 (s, IH), 5.25 (s, 2H), 3.96 (s, 3H), 2.37 (s, 3H); 13C NMR (CDCl₃, 125 MHz) δ 159.0, 153.3, 144.1, 144.0, 135.7, 128.7, 128.5, 128.2 (2C), 122.5, 121.6, 111.6, 104.5, 67.4, 56.3, 8.2; IR (film) νmax 2910, 2359, 2339, 2159, 1693, 1537, 1354, 1209, 1078, 1024 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₁₇H₁₄NO₂, 378.0954; found, 378.0936.

**[0336]** Benzyl 7-hydroxy-8-methyl-2-oxo-6-propoxy-2H-chromen-3-ylcarbamate (23b): A solution of **6b** (390 mg, 2.14 mmol) and enamine **22** (596 mg, 2.14 mmol) in glacial acetic acid (13.4 mL) was heated to reflux for 36 hours. Upon cooling to room temperature, the precipitated yellow solid was collected by filtration, washed with water, recrystallized from MeOH/water, and extracted with EtOAc (3 x 20 mL). Combined organic fractions were washed with saturated aqueous NaCl, dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 100:1 CH₂Cl₂:Acetone) and recrystallized from MeOH/water to afford **23b** as a yellow amorphous solid (278 mg, 34%): 1H NMR (CD₂Cl₂, 400 MHz) δ 8.26 (s, IH), 7.56 (s, IH), 7.47-7.38 (m, 5H), 6.84 (s, IH), 6.28 (s, IH), 5.25 (s, 2H), 4.09 (t, J = 6.6 Hz, 2H), 2.36 (s, 3H), 1.93-1.88 (m, 2H), 1.10 (t, J = 7.4 Hz, 3H); 13C NMR (CDCl₃, 125 MHz) δ 158.0, 152.2, 144.9, 142.9, 142.3, 134.6, 129.0, 127.6, 127.5 (2C), 127.2 (2C), 121.5, 120.5, 110.9, 110.5, 104.4, 69.8, 66.3, 21.4, 9.4, 7.1; IR (film) νmax 2957, 2920, 2851, 2359, 2341, 1693, 1537, 1358, 1277, 1080, 1024, 910 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₁₆H₁₄NO₂, 384.1447; found, 384.1447.

**[0337]** Benzyl 7-hydroxy-6-isopropoxy-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (23c): A solution of **6c** (142 mg, 0.78 mmol) and enamine **22** (217 mg, 0.78 mmol) in glacial acetic acid (4.90 mL) was heated to reflux for 40 hours. Upon cooling to room temperature, the solution was extracted with EtOAc (3 x 10 mL); combined organic fractions were dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford **23c** as a yellow amorphous solid (159 mg, 53%): 1H NMR (CD₂Cl₂, 400 MHz) δ 8.26 (s, IH), 7.56 (s, IH), 7.44-7.38 (m, 5H), 6.85 (s, IH), 6.31 (s, IH), 5.25 (s, 2H), 4.66 (quintet, J = 6.1 Hz, IH), 2.35 (s, 3H), 1.42 (d, J = 6.0 Hz, 6H); 13C NMR (CDCl₃, 125 MHz) δ 159.1, 154.9, 146.7, 143.9 (2C), 142.0, 135.7, 128.7 (2C), 128.5 128.2 (2C), 122.6 (2C), 111.6, 107.0,
72.3, 67.4, 22.1 (2C), 8.2; IR (film) ν \(_{\text{max}}\) 3400, 2924, 2853, 2359, 1817, 1699, 1524, 1412, 1354, 1300, 1221, 1204, 1113, 1076, 1022, 824 cm\(^{-1}\); HRMS (ESI\(^+\)) \(m/z\): [M + H]\(^+\) calcd for \(C_2\text{H}_4\text{H}_2\text{NO}_6\), 384.1447; found, 384.1452.

**[0338]** Benzyl 7-hydroxy-5-methoxy-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (23d): A solution of 10 (251 mg, 1.63 mmol) and enamine 22 (680 mg, 2.44 mmol) in glacial acetic acid (10.2 mL) was heated to reflux for 40 hours. Upon cooling to room temperature, the solution was extracted with EtOAc (3 x 30 mL); combined organic fractions were dried (Na\(_2\)SO\(_4\)), filtered, and concentrated. The residue was purified via column chromatography (SiO\(_2\), 40:1 → 20:1; CH\(_2\)Cl\(_2\):Acetone) to afford 23d as a yellow amorphous solid (204 mg, 35%); \(^1\)H NMR (CD\(_2\)Cl\(_2\), 400 MHz) δ 8.48 (s, IH), 7.46-7.38 (m, 6H), 6.38 (s, IH), 5.25 (s, 2H), 5.15 (s, IH), 3.87 (s, 3H), 2.23 (s, 3H); \(^13\)C NMR (CDCl\(_3\), 125 MHz) δ 159.1, 155.8, 154.3, 153.2, 149.8, 137.0, 128.7, 128.6, 128.2, 128.2, 109.3, 109.0, 108.5, 105.6, 96.9, 70.8, 60.2, 7.3; IR (film) ν \(_{\text{max}}\) 3406, 2935, 2837, 1713, 1670, 1607, 1529, 1501, 1364, 1242, 1101, 1051, 991, 966, 735 cm\(^{-1}\); HRMS (ESI\(^+\)) \(m/z\): [M + Na]\(^+\) calcd for C\(_9\)H\(_9\)NO\(_6\), 378.0954; found, 378.0974.

**[0339]** Benzyl 8-benzyl-7-hydroxy-2-oxo-2H-chromen-3-ylcarbamate (23e): A solution of 14 (115 mg, 0.57 mmol) and enamine 22 (160 mg, 0.57 mmol) in glacial acetic acid (4.0 mL) was heated to reflux for 40 hours. Upon cooling to room temperature, the solution was extracted with EtOAc (3 x 10 mL); combined organic fractions were dried (Na\(_2\)SO\(_4\)), filtered, and concentrated. The residue was purified via column chromatography (SiO\(_2\), 100:1; CH\(_2\)Cl\(_2\):Acetone), followed by recrystallization from MeOH to afford 23e as an orange amorphous solid (296 mg, 48%); \(^1\)H NMR (CD\(_2\)Cl\(_2\), 400 MHz) δ 8.29 (s, IH), 7.53 (s, IH), 7.46-7.38 (m, 4H), 7.37-7.27 (m, 4H), 7.23-7.19 (m, 2H), 7.01 (t, J = 8.1 Hz, IH), 6.86 (d, J = 8.4 Hz, IH), 6.46 (d, J = 8.1 Hz, IH), 5.25 (s, 2H), 4.25 (s, 2H), 4.06 (s, IH); \(^13\)C NMR (CDCl\(_3\), 125 MHz) δ 157.7, 154.3, 153.9, 152.2, 148.0, 137.9, 134.5, 127.6, 127.6, 127.6, 127.6, 127.5, 127.3, 127.2, 127.4, 126.6, 125.4, 125.3, 121.4, 120.4, 114.0, 112.6, 66.5, 27.5; IR (film) ν \(_{\text{max}}\) 3381, 2957, 2928, 2359, 2341, 1693, 1607, 1526, 1466, 1454, 1383, 1366, 1219, 1204, 1076, 1045, 764, 737, 700 cm\(^{-1}\); HRMS (ESI\(^+\)) \(m/z\): [M + H]\(^+\) calcd for C\(_4\)H\(_9\)NO\(_5\), 402.1341; found, 402.1341.

**[0340]** Benzyl 7-hydroxy-2-oxo-8-phenyl-2H-chromen-3-ylcarbamate (23f): A solution of 17 (400 mg, 2.15 mmol) and enamine 22 (598 mg, 2.15 mmol) in glacial acetic acid (14.3 mL) was heated to reflux for 40 hours. Upon cooling to room temperature, the solution was extracted with EtOAc (3 x 30 mL); combined organic
fractions were dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 100:1; CH₂Cl₂:Acetone), then recrystallized from MeOH to afford 23f as an orange amorphous solid (264 mg, 27%): ¹H NMR (CDCl₃, 500 MHz) δ 8.25 (s, IH), 7.51-7.48 (m, 2H), 7.43-7.40 (m, 2H), 7.35-7.29 (m, 8H), 6.94 (d, J = 8.6 Hz, IH), 5.16 (s, 2H); ¹³C NMR (CDCl₃,125 MHz) δ 158.5 (2C), 154.3, 153.2, 147.7, 135.6, 130.9, 130.6, 130.5, 129.8, 129.4, 129.2 (2C), 128.7 (2C), 128.6, 128.3, 127.8, 122.2, 121.6, 113.3, 113.5, 67.5; IR (film) v_max 3391, 3339, 2964, 2870, 2357, 1732, 1682, 1620, 1547, 1497, 1480, 1460, 1420, 1383, 1366, 1307, 1284, 1215, 1045, 1047, 750 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]+ calcd for C₂₃H₁₇NO₅, 388.1185; found, 388.1214.

[0341] Benzyl 7-hydroxy-8-methoxy-2-oxo-2H-chromen-3-ylcarbamate (23g): A solution of 19 (1.10 g, 7.86 mmol) and enamine 22 (2.18 g, 7.86 mmol) in glacial acetic acid (60.0 mL) was heated to reflux for 90 hours. Upon cooling to room temperature, the solution was extracted with EtOAc (3 x 50 mL); combined organic fractions were washed with saturated aqueous NaCl, dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 11:1; Hexane:EtOAc → EtOAc) then recrystallized from MeOH/water to afford 23g as a colorless amorphous solid (207 mg, 7.7%): ¹H NMR (CDCl₃, 400 MHz) δ 8.30 (s, IH), 7.50 (s, IH), 7.43-7.36 (m, 5H), 7.13 (d, J = 8.6 Hz, IH), 6.97 (d, J = 7.9 Hz, IH), 6.04 (s, IH), 5.21 (s, 2H), 4.13 (s, 3H); ¹³C NMR (Acetone-ｄ₆, 100 MHz) δ 157.3, 153.3, 151.5 (2C), 144.1, 136.5, 134.4, 128.4 (2C), 128.1, 128.0 (2C), 122.7, 121.6, 113.6, 113.2, 66.7, 60.7; IR (film) v_max 2920, 2851, 2405, 2357, 1707, 1605, 1522, 1458, 1385, 1364, 1275, 1259, 1213, 1088, 1047, 750 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]+ calcd for C₁₄H₁₃NO₅, 364.0797; found, 364.0776.

[0342] Benzyl 8-ethyl-7-hydroxy-2-oxo-2H-chromen-3-ylcarbamate (23h): A solution of 21 (1.40 g, 10.1 mmol) and enamine 22 (2.80 g, 10.1 mmol) in glacial acetic acid (50.0 mL) was heated to reflux for 12 hours. Upon cooling to room temperature, the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 4:1 → 2:1; Hexane:EtOAc), then recrystallized from acetone/hexanes to afford 23h as a colorless amorphous solid (600 mg, 17%). ¹H NMR (DMSO-Ｊ₂, 400 MHz) δ 9.07 (s, IH), 8.12 (s, IH), 7.46-7.32 (m, 6H), 6.86 (d, J = 8.4 Hz, IH), 5.18 (s, 2H), 2.72 (q, J = 7.6 Hz, 2H), 1.11 (t, J = 7.6 Hz, 3H); ¹³C NMR (DMSO-Ｊ₂, 125 MsHz) δ 158.0, 157.1, 153.8 (2C), 149.5, 136.4, 128.4 (2C), 127.9, 127.8, 127.2, 125.8, 120.4, 116.6, 112.8, 111.4, 66.1, 15.7, 13.5; IR (film) v_max 3391, 3339, 2964, 2870, 2357, 1732, 1682, 1620,
1524, 1506, 1454, 1364, 1277, 1 188, 1097, 1024, 752, 698 cm⁻¹; HRMS (ESI⁺) m/z: [M + H⁺] calcd for C₁₉H₁₇N⁺, 340.1 185; found, 340.1 181.

[0343] Benzyl 6-methoxy-7-((3aR,4S,7R,7aR)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3aH-[1,3]dioxolo[4,5-c]pyran-4-yl)oxo-2H-chromen-3-ylcarbamate (25a): Boron trifluoride etherate (5.30 µL, 0.042 mmol) was added to 23a (50.0 mg, 0.14 mmol) and (3α?,45′,7i?,7α?)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (25a); After stirring at room temperature for 14 hours, triethylamine (150 µL) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to give 25a as a colorless foam (74.0 mg, 95%); ¹H NMR (CD₂Cl₂, 400 MHz) δ 8.29 (s, IH), 7.64 (s, IH), 7.47-7.39 (m, 5H), 6.91 (s, IH), 5.52 (d, J = 3.4 Hz, IH), 5.26 (s, 2H), 5.23 (dd, J = 8.4, 3.5 Hz, IH), 4.95 (t, J = 8.2 Hz, IH), 3.92 (s, 3H), 3.60 (s, 3H), 3.33 (d, J = 8.0 Hz, IH), 2.42 (s, 3H), 1.38 (s, 3H), 1.33 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 157.6, 152.7, 152.1, 148.1, 144.8, 141.8, 134.5, 127.8, 127.7, 127.5, 127.3, 122.3, 120.2, 119.8, 115.1, 109.6, 105.2, 98.3, 82.0, 77.1, 66.5, 65.5, 59.4, 57.4, 55.1, 26.0, 20.9, 8.9; IR (film) ν max 2961, 2939, 2906, 2854, 2828, 1709, 1579, 1522, 1464, 1389, 1371, 1205, 1174, 1111, 1072, 1034, 957, 800 cm⁻¹; HRMS (ESI⁺) m/z: [M + H⁺] calcd for C₂₈H₂₉N-Oct, 556.1819; found, 556.1822.

[0344] Benzyl 7-((3aR,4S,7R,7aR)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3aH-[1,3]dioxolo[4,5-c]pyran-4-yl)oxo-2H-chromen-3-ylcarbamate (25b): Boron trifluoride etherate (16.7 µL, 0.13 mmol) was added to 23b (170 mg, 0.44 mmol) and (3α?,45′,7i?,7α?)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetamidate (171 mg, 0.47 mmol) in anhydrous CH₂Cl₂ (3.00 mL). After stirring at room temperature for 14 hours, triethylamine (150 µL) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to give 25b as a colorless foam (246 mg, 95%); ¹H NMR (CDCl₃, 500 MHz) δ 8.17 (s, IH), 7.35-7.27 (m, 5H), 6.84 (s, IH), 5.96 (s, IH), 5.15 (s, 2H), 4.99 (d, J = 7.5 Hz, IH), 4.59 (d, J = 9.7 Hz, IH), 4.23 (d, J = 9.6 Hz, IH), 3.97 (t, J = 6.6 Hz, IH), 3.82-3.75 (m, 2H), 3.37 (s, 3H), 1.84-1.79 (m, 2H), 1.51 (s, 3H), 1.41 (s, 3H), 1.18 (s, 3H), 1.00 (t, J = 7.5 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 157.8, 154.3, 152.2, 151.8, 146.7, 144.2, 142.9, 134.6, 127.8, 127.6, 127.5, 127.2, 121.0, 120.9, 111.1, 105.3, 101.7, 91.6, 85.7, 82.8, 80.0, 69.8, 58.1, 54.8, 28.3, 28.2, 22.4, 21.3, 9.4; IR (film) ν max 2961, 2939, 2906,
2359, 2341, 1811, 1757, 1726, 1522, 1445, 1371, 1267, 1175, 1113, 1086, 825, 768 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₃₀H₃₃NO₃, 606.1952; found, 606.1950.

**0345** Benzyl 6-isopropoxy-7-((3aR,4S,7R,7aR)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3H-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (25c): Boron trifluoride etherate (1.30 µL, 0.010 mmol) was added to 23c (13.0 mg, 0.034 mmol) and (3α?,45',7i?,7α?)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetimidate (83.0 mg, 0.23 mmol) in anhydrous CH₂Cl₂ (1.30 mL). After stirring at room temperature for 1.5 hours, triethylamine (150 µL) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to give 25c as a colorless foam (19.0 mg, 95%): ¹H NMR (CDCl₃, 500 MHz) δ 8.17 (s, IH), 7.51 (s, IH), 7.35 (s, IH), 7.34-7.33 (m, 4H), 6.74 (s, IH), 5.54 (dd, J = 9.2, 1.2 Hz, IH), 5.16 (s, 2H), 4.87-4.84 (m, IH), 4.73 (dd, J = 7.9, 1.9 Hz, IH), 4.51 (quintet, J = 6.0 Hz, IH), 3.52 (s, 3H), 3.28 (d, J = 4.8 Hz, IH), 2.33 (s, 3H), 1.80-1.77 (m, 6H), 1.30 (s, 3H), 1.27 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 161.6, 158.6, 153.2, 153.1, 147.1, 146.8, 142.5, 135.5, 128.7, 128.6, 128.3, 123.3, 121.4, 121.1, 116.2, 108.6, 99.4, 83.1, 79.9, 76.1, 74.7, 72.2, 68.0, 60.5, 27.1, 25.6, 21.9, 21.6, 21.0, 10.1; IR (film) νmax 2955, 2922, 2853, 2359, 2339, 1819, 1711, 1520, 1464, 1375, 1171, 1111, 1034, 962, 822, 766 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₃₀H₃₃NO₃, 584.2132; found, 584.2111.

**0346** Benzyl 5-methoxy-7-((3aR,4i?,7i?,7α?)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (25d): Boron trifluoride etherate (18.5 µL, 0.15 mmol) was added to 23d (174 mg, 0.49 mmol) and (3α?,45',7i?,7α?)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3αH-[1,3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetimidate (621 mg, 1.71 mmol) in anhydrous CH₂Cl₂ (11.0 mL). After stirring at room temperature for 14 hours, triethylamine (150 µL) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to give 25d as a colorless foam (200 mg, 74%): ¹H NMR (CDCl₃, 500 MHz) δ 8.49 (s, IH), 7.34-7.27 (m, 5H), 6.67 (s, IH), 6.60 (s, IH), 5.69 (s, 2H), 5.16 (d, J = 5.3 Hz, IH), 4.89 (t, J = 7.8 Hz, IH), 4.63 (dd, J = 7.9, 2.4 Hz, IH), 3.83 (s, 3H), 3.37 (s, 3H), 3.15 (d, J = 8.0 Hz, IH), 2.16 (s, 3H), 2.16 (s, 3H), 2.12 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 158.9, 156.0, 155.2, 154.2, 153.1 (2C), 149.4, 135.7, 128.7 (2C), 128.5, 128.2 (2C), 120.8, 117.4, 106.6, 105.4, 94.6, 94.1, 82.9, 67.4, 60.6, 60.6, 56.1, 56.0, 22.2, 22.0, 7.9; IR
(film) \(v_{\text{max}}\) 2955, 2924, 2853, 1817, 1713, 1526, 1209, 1105, 1072, 1034, 976, 808 cm\(^{-1}\); HRMS (ESI\(^+\)) \(m/z\): [M + H]\(^+\) calcd for C\(_{2}\)H\(_{2}\)NOn, 556.1819; found, 556.1826.

[0347] Benzyl 8-benzyl-7-(3 α\(^?\),4\(^?\),7\(^?\),7\(^?\)α\(^?\))-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3 \(\text{ofH}\)-[1,3]dioxolo [4,5-c]pyran-4-yloxy)-2-oxo-2 \(H\)-chromen-3 -ylcarbamate (25e): Boron trifluoride ete erate (7.80 \(\mu\)L, 0.062 mmol) was added to 23e (80.0 mg, 0.21 mmol) and (3\(\alpha\(^?\),4\(^?\),7\(^?\),7\(^?\)α\(^?\))-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3 \(\text{ofH}\)-[1,3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetimidate (299 mg, 0.83 mmol) in anhydrous CH\(_{2}\)Cl\(_{2}\) (5.20 mL). After stirring at room temperature for 48 hours, triethylamine (150 \(\mu\)L) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO\(_{2}\), 40:1 CH\(_{2}\)Cl\(_{2}\):Acetone) to give 25e as a colorless foam (47.0 mg, 39%): \(\text{\(^1\))H NMR (CDCl}_3, 500 MHz) \(\delta\) 8.22 (s, IH), 7.46 (s, IH), 7.35-7.27 (m, 5H), 7.17-7.06 (m, 5H), 6.86 (d, \(J = 10\) Hz, IH), 6.01 (d, \(J = 10\) Hz, IH), 5.65 (d, \(J = 1.6\) Hz, IH), 5.23 (s, 2H), 5.16 (s, 2H), 4.77-4.70 (m, IH), 4.10 (s, IH), 3.50 (s, 3H), 3.28 (s, IH), 3.16 (d, \(J = 7.4\) Hz, IH), 1.25 (s, 3H), 1.18 (s, 3H); \(\text{\(^{13}\))C NMR (CDCl}_3, 125 MHz) \(\delta\) 155.1, 153.2 (2C), 153.1, 148.6 (2C), 139.6 (2C), 128.7, 128.6 (2C), 128.4 (2C), 128.3 (2C), 128.3 (2C), 126.5, 126.2, 123.1, 122.4, 121.7, 117.6, 114.9, 111.5, 94.7, 82.8, 67.6, 60.6 (2C), 29.7, 27.6, 21.9; IR (film) \(v_{\text{max}}\) 2926, 2854, 2359, 2341, 1811, 1709, 1607, 1522, 1456, 1381, 1366, 1259, 1209, 1171, 1078, 1049, 968, 766, 700 cm\(^{-1}\); HRMS (ESI\(^+\)) \(m/z\): [M + H]\(^+\) calcd for C\(_{3}\)H\(_{3}\)N\(_{1}\)O\(_{10}\), 602.2026; found, 602.2053.

[0348] Benzyl 7-(3 \(\alpha\(^?\),4\(^?\),7\(^?\),7\(^?\)α\(^?\))-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3 \(\text{ofH}\)-[1,3]dioxolo[4,5-c]pyran-4-yloxy)-2-oxo-8-phenyl-2 \(H\)-chromen-3-ylcarbamate (25f): Boron trifluoride ete erate (14.6 \(\mu\)L, 0.12 mmol) was added to 23f (155 mg, 0.39 mmol) and (3\(\alpha\(^?\),4\(^?\),7\(^?\),7\(^?\)α\(^?\))-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3 \(\text{ofH}\)-[1,3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetimidate (560 mg, 1.55 mmol) in anhydrous CH\(_{2}\)Cl\(_{2}\) (9.70 mL). After stirring at room temperature for 48 hours, triethylamine (150 \(\mu\)L) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO\(_{2}\), 100:1 → 40:1 CH\(_{2}\)Cl\(_{2}\):Acetone) to give 25f as a colorless foam (225 mg, 99%): \(\text{\(^1\))H NMR (CDCl}_2, 400 MHz) \(\delta\) 8.37 (s, IH), 7.75-7.73 (m, 2H), 7.60-7.36 (m, 10H), 7.32 (d, \(J = 8.8\) Hz, IH), 5.77 (d, \(J = 1.7\) Hz, IH), 5.26 (s, 2H), 4.76-4.68 (m, IH), 4.36-4.28 (m, IH), 3.56 (s, 3H), 3.28 (d, \(J = 7.2\) Hz, IH), 1.37 (s, 3H), 1.31 (s, 3H); \(\text{\(^{13}\))C NMR (CDCl}_3, 125 MHz) \(\delta\) 157.2, 153.0 (2C), 152.1 (2C), 152.1, 134.4, 129.9, 129.8, 129.4, 127.7 (2C), 127.5, 127.2 (2C), 127.1 (2C), 127.0, 126.3, 121.5 (2C), 120.3, 111.2 (2C), 93.9, 81.9, 66.5, 59.4 (3C), 20.9 (2C); IR (film) \(v_{\text{max}}\) 3400, 2959, 2926, 2853,
Benzy1 8-methoxy-7-(3 α?,4,5,7i?,7aR)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3 αH-[1,3]dioxolo[4,5-c]pyran-4-yl-2-oxo-2 H-chromen-3-ylcarbamate (25g): Boron trifluoride etherate (17.3 μL, 0.14 mmol) was added to 23g (157 mg, 0.46 mmol) and (3α?,4,5,7i?,7aR)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3 αH-[1,3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetimidate (665 mg, 1.83 mmol) in anhydrous CH₂Cl₂ (11.5 mL). After stirring at room temperature for 24 hours, triethylamine (150 μL) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 40:1 → 10:1 CH₂Cl₂:Acetone) to give 25g as a colorless foam (237 mg, 95%): ¹H NMR (CDCl₃, 500 MHz) δ 8.20 (s, IH), 7.48 (s, IH), 7.33-7.29 (m, 5H), 7.09 (dd, J = 14.2, 8.8 Hz, 2H), 5.72 (d, J = 1.8 Hz, IH), 5.16 (s, 2H), 5.02 (dd, J = 7.8, 1.8 Hz, IH), 4.89 (t, J = 7.8 Hz, IH), 3.88 (s, 3H), 3.52 (s, 3H), 3.21 (d, J = 7.8 Hz, IH), 1.27 (s, 3H) 1.17 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 158.0, 153.3, 153.2, 153.1, 149.8, 143.8, 137.1, 135.5, 128.7 (2C), 128.6, 128.3 (2C), 122.7, 122.2, 121.4, 116.1, 113.7, 95.3, 74.7, 72.9, 67.6, 61.9, 60.7, 60.6, 29.7, 29.4; IR (film) v_max 3400, 3319, 2984, 2935, 2359, 1815, 1715, 1609, 1526, 1464, 1383, 1364, 1285, 1213, 1175, 1111, 1063, 968, 764, 737, 700 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₂₇H₂₇NO₇, 564.1482; found, 564.1455.

Benzy1 8-ethyl-7-(3 α?,4,5,7i?,7aR)-7-methoxy-6,6-dimethyl-2-oxotetrahydro-3 αH-[1,3]dioxolo[4,5-c]pyran-4-yl-2-oxo-2 H-chromen-3-ylcarbamate (25h): Boron trifluoride etherate (19.0 μL, 0.15 mmol) was added to 23h (171 mg, 0.51 mmol) and (3α?,4,5,7i?,7aR)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3 αH-[1,3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetimidate (183 mg, 0.51 mmol) in anhydrous CH₂Cl₂ (11.0 mL). After stirring at room temperature for 24 hours, triethylamine (150 μL) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to give 25h as a colorless foam (138 mg, 51%): ¹H NMR (CD₂Cl₂, 400 MHz) δ 8.29 (s, IH), 7.62 (s, IH), 7.47-7.38 (m, 5H), 7.36 (d, J = 8.8 Hz, IH), 7.19 (d, J = 8.4 Hz, IH), 5.81 (d, J = 2.4 Hz, IH), 5.25 (s, 2H), 5.10 (dd, J = 8.0, 2.0 Hz, IH), 5.02 (t, J = 7.8 Hz, IH), 3.55 (s, 3H), 3.41 (d, J = 7.2 Hz, IH), 2.87 (q, J = 7.4 Hz, 2H), 1.42 (s, 3H), 1.28 (s, 3H), 1.21 (t, J = 7.4 Hz, 3H); ¹³C NMR (CD₂Cl₂, 100 MHz) δ 158.5, 154.8, 153.2, 153.1, 148.4, 136.0, 128.6 (2C), 128.4, 128.1, 125.6, 122.4, 121.5, 120.7, 114.8, 111.4, 94.8, 82.7, 77.9, 77.2,
N-(7-((2S,3R,4S,5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2H-pyran-2-yl oxo)-6-methoxy-8-methyl-2-oxo-2H-chromen-3-yl)-3',6-dimethoxybiphenyl-3-carboxamide (26a): Palladium on carbon (10%, 20.0 mg) was added to 25a (100 mg, 0.18 mmol) in anhydrous THF (5.00 mL) and the solution was placed under an atmosphere of H₂. After 6.5 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (56.0 mg, 75%). EDCI (21.4 mg, 0.11 mmol) and 3',6-dimethoxybiphenyl-3-carboxylic acid (23.1 mg, 0.089 mmol) were added to the amine (18.7 mg, 0.045 mmol) in 30% pyridine/CH₂Cl₂ (0.70 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford a colorless solid, which was used without further purification (10.5 mg, 36%). Triethylamine (150 µL) was added to the carbonate (10.4 mg, 0.016 mmol) in MeOH (2.50 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 20:1; CH₂Cl₂:MeOH) to afford 26a as a colorless amorphous solid (2.00 mg, 20%, 5% over 3 steps): ¹H NMR (CDCl₃, 500 MHz) δ 8.73 (s, IH), 8.70 (d, J = 5.4 Hz, IH), 7.84 (td, J = 6.2, 2.4 Hz, IH), 7.82 (s, IH), 7.30 (t, J = 8.0 Hz, IH), 7.06 (d, J = 7.8 Hz, IH), 7.03-7.00 (m, 2H), 6.88-6.86 (m, IH), 6.81 (s, IH), 4.99 (d, J = 6.6 Hz, IH), 4.24 (t, J = 4.2 Hz, IH), 4.00 (dd, J = 6.5, 3.7 Hz, IH), 3.90 (s, 3H), 3.86 (s, 3H), 3.83 (s, 3H), 3.80 (d, J = 7.4 Hz, IH), 3.45 (s, 3H), 3.08 (d, J = 4.7 Hz, IH), 2.67 (s, IH), 2.42 (s, 3H), 1.28 (d, J = 8.1 Hz, 3H), 1.18 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 164.6, 158.9, 158.3, 158.2, 148.2, 145.6, 142.5, 137.5, 130.1, 129.0, 128.2, 127.2, 124.9, 122.5, 122.2, 121.2, 121.0, 115.1, 144.2, 112.1, 110.0, 105.4, 101.3, 81.7, 76.8, 69.0, 68.0, 59.1, 55.3, 54.9, 54.3, 28.3, 28.2, 9.1; IR (film) νₘₜₕ max 2961, 2928, 1713, 1670, 1601, 1464, 1383, 1261, 1094, 1022, 798, 700 cm⁻¹; HRMS (ESI⁺) m/z: [M + H⁺]⁰ calcd for C₃₂H₇₉NO₆, 636.2445; found, 636.2477.
used without further purification (325 mg, 99%). EDCI (116 mg, 0.60 mmol) and 3',6-
dimethoxybiphenyl-3-carboxylic acid (125 mg, 0.4821 mmol) were added to the amine (108 mg, 0.2410 mmol) in 30% pyridine/CH₂Cl₂ (6.70 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 3:1 Hexane:Ether → 20:1 CH₂Cl₂:Acetone) to afford a colorless solid, which was used without further purification (51.0 mg, 31%). Triethylamine (150 µL) was added to the carbonate (51.0 mg, 0.074 mmol) in MeOH (2.50 mL). After 48 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford 26b as a colorless amorphous solid (22.8 mg, 47%, 14% over 3 steps): ¹H NMR (CD₂Cl₂, 400 MHz) δ 8.79 (s, IH), 8.78 (s, IH), 7.96 (dd, J = 8.6, 2.4 Hz, IH), 7.91 (d, J = 2.4 Hz, IH), 7.39 (t, J = 7.9 Hz, IH), 7.16-7.1 1 (m, 2H), 6.97-6.94 (m, 2H), 5.97 (s, IH), 5.14 (d, J = 6.5 Hz, IH), 4.31 (t, J = 3.5 Hz, IH), 4.12-4.06 (m, 2H), 4.03 (dd, J = 6.8, 1.8 Hz, IH), 3.93 (s, 3H), 3.88 (s, 3H), 3.65 (s, IH), 3.53 (s, 3H), 3.17 (d, J = 4.8 Hz, IH), 2.80 (s, IH), 2.48 (s, 3H), 1.95-1.90 (m, 2H), 1.37 (s, 3H), 1.35 (s, 3H), 1.1 1 (t, J = 7.4 Hz, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 165.0, 164.6, 158.8, 158.3, 147.7, 145.7, 142.3, 137.8, 137.5, 131.3, 129.0, 128.2, 127.2, 124.9, 122.3, 121.2, 121.0, 115.1 11, 114.2, 112.1, 110.0, 106.2, 101.1, 81.7, 70.0, 69.0, 68.0, 64.8, 59.1, 54.9, 54.3, 24.7, 24.0, 21.3, 9.5, 9.1; IR (film) ν max 3398, 3196, 2964, 2935, 2359, 2330, 1705, 1580, 1524, 1382, 1242, 124, 1194, 939, 808, 760, 735 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₃₆H₃₁INO₃, 664.2758; found, 664.2754.

[0353]  N-(7-((2S,3i?,4S,5i?)-3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-6-isopropoxy-8-methyl-2-oxo-2 H-chromen-3-yl)-3',6-
dimethoxybiphenyl-3-carboxamide (26c): Palladium on carbon (10%, 11 mg) was added to 25c (54.5 mg, 0.093 mmol) in anhydrous THF (600 µL) and the solution was placed under an atmosphere of H₂. After 12 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (42.0 mg, 99%). EDCI (14.9 mg, 0.078 mmol) and 3',6-dimethoxybiphenyl-3-carboxylic acid (16 mg, 0.062 mmol) were added to the amine (14.0 mg, 0.031 mmol) in 30% pyridine/CH₂Cl₂ (900 µL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 3:1 Hexane:Ether → 40:1 CH₂Cl₂:Acetone) to afford a colorless solid, which was used without further purification (17.5 mg, 82%). Triethylamine (150 µL) was added to the carbonate (17.5 mg, 0.025 mmol) in MeOH (2.50 mL) and CH₂Cl₂ (2.50 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography
(SiO₂, 10:1 CH₂Cl₂:Acetone) to afford 26c as a colorless amorphous solid (6.0 mg, 35%, 28% over 3 steps): ¹H NMR (CD₂Cl₂, 500 MHz) δ 8.69 (s, IH), 8.67 (s, IH), 7.84 (dd, J = 8.6, 2.4 Hz, IH), 7.80 (d, J = 2.4 Hz, IH), 7.29 (d, J = 8.0 Hz, IH), 7.25 (t, J = 7.9 Hz, IH), 7.03-7.01 (m, 2H), 6.87 (s, IH), 6.87-6.83 (m, IH), 4.96 (d, J = 6.8 Hz, IH), 4.61-4.56 (m, IH), 4.19 (t, J = 4.0 Hz, IH), 3.89 (dd, J = 6.8, 3.7 Hz, IH), 3.82 (s, 3H), 3.76 (s, 3H), 3.75 (s, IH), 3.41 (s, 3H), 3.34 (s, IH), 3.03 (d, J = 4.5 Hz, IH), 2.36 (s, 3H), 1.33 (t, J = 6.2 Hz, 6H), 1.25 (s, 3H), 1.23 (s, 3H); ¹³C NMR (CD₂Cl₂, 125 MHz) δ 164.6, 159.1, 158.1, 158.3, 146.4, 146.3, 142.5, 138.1, 130.2, 129.1, 128.3, 127.4, 125.2, 122.8, 122.2, 121.2, 112.1, 115.5, 144.5, 112.1, 110.2, 108.4, 101.4, 81.9, 77.0, 71.1, 69.2, 68.3, 59.1, 55.1, 54.5, 28.7, 28.6, 20.8, 20.8, 9.1; IR (film) νmax 2924, 2854, 2359, 2341, 1734, 1684, 1653, 1558, 1541, 1522, 1506, 1458, 1387, 1339, 1286, 1244, 1113, 912, 797 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₃₆H₄₁N₂O₄, 686.2578; found, 686.2610.

[0354]  N-(7-((2i?,3i?,45',5i?)-3,4-dihyrox-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-5-methoxy-8-methyl-2-oxo-2 H-chromen-3-yl)-3',6-dimethoxybiphenyl-3-carboxamide (26d): Palladium on carbon (10%, 40 mg) was added to 25d (200 mg, 0.36 mmol) in anhydrous THF (2.40 mL) and the solution was placed under an atmosphere of H₂. After 12 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (150 mg, 99%). EDCI (57.5 mg, 0.30 mmol) and 3',6-dimethoxybiphenyl-3-carboxylic acid (62 mg, 0.24 mmol) were added to the amine (50.6 mg, 0.12 mmol) in 30% pyridine/CH₂Cl₂ (3.30 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 3:1 Hexane:Ether → 40:1 → 10:1 CH₂Cl₂:Acetone) to afford a colorless solid, which was used without further purification (25.2 mg, 32%). Triethylamine (150 µL) was added to the carbonate (25.2 mg, 0.038 mmol) in MeOH (2.0 mL) and CH₂Cl₂ (2.0 mL). After 48 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford 26d as a colorless amorphous solid (17.0 mg, 70%, 22% over 3 steps): ¹H NMR (CD₂Cl₂, 400 MHz) δ 9.02 (s, IH), 8.97 (s, IH), 8.66 (s, IH), 7.96 (dd, J = 8.6, 2.4 Hz, IH), 7.91-7.90 (m, IH), 7.39 (t, J = 7.9 Hz, IH), 7.16-7.11 (m, 2H), 6.96 (dd, J = 8.3, 2.6 Hz), 6.85 (d, J = 5.5 Hz, IH), 5.70 (d, J = 2.1 Hz, IH), 4.36-4.33 (m, IH), 4.27 (m, IH), 3.99 (s, 3H), 3.93 (s, 3H), 3.88 (s, 3H), 3.62 (s, 3H), 3.41-3.38 (m, IH), 2.24 (s, 3H), 1.41 (s, 3H), 1.19 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 164.2, 158.7, 158.6, 158.3, 155.3, 153.5, 148.6, 137.6, 129.9, 128.9, 128.1,
127.1, 125.1, 121.0, 119.4, 118.9, 114.2, 114.2, 112.1, 110.0, 104.9, 103.7, 96.7, 92.9, 83.2, 70.1, 67.5, 60.9, 60.8, 54.8, 54.3, 21.9, 21.4, 6.8; IR (film) νmax 3405, 2986, 2934, 1713, 1609, 1528, 1383, 1250, 1213, 1053, 999, 914, 878, 737 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]+ calcd for C34H33NO1, 636.2445; found, 636.2482.

**[0355]** N-(8-benzyl-7-((2i?,3i?,4 S,5i?)3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-2-oxo-2 H-chromen-3-yl)-3',6-dimethoxybiphenyl-3-carboxamide (26e): Palladium on carbon (10%, 46 mg) was added to 25e (230 mg, 0.38 mmol) in anhydrous THF (2.50 mL) and the solution was placed under an atmosphere of H₂. After 12 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (177 mg, 99%). EDCI (61.5 mg, 0.32 mmol) and 3',6-dimethoxybiphenyl-3-carboxylic acid (66.3 mg, 0.26 mmol) were added to the amine (60.0 mg, 0.13 mmol) in 30% pyridine/CH₂Cl₂ (3.50 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford 26e as a colorless amorphous solid (6.00 mg, 51%, 7.1% over 3 steps): ¹H NMR (CD₂Cl₂, 400 MHz) δ 8.84 (s, IH), 8.72 (s, IH), 7.96 (dd, J = 10, 2.4 Hz, IH), 7.91 (d, J = 2.4 Hz, IH), 7.50 (d, J = 8.8 Hz, IH), 7.39 (t, J = 7.9 Hz, IH), 7.31 (d, J = 8.8 Hz, IH), 7.28-7.25 (m, 5H), 7.21-7.18 (m, IH), 7.15-7.11 (m, 2H), 6.97-6.94 (m, IH), 5.54 (d, J = 2.7 Hz, IH), 4.25 (t, J = 15.1 Hz, 2H), 4.17-4.11 (m, IH), 4.05 (d, J = 2.6 Hz, IH), 3.93 (s, 3H), 3.88 (s, 3H), 3.58 (s, 3H), 3.31 (d, J = 8.7 Hz, IH), 2.64 (s, IH), 2.04 (s, IH), 1.40 (s, 3H), 1.03 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 165.5, 159.8, 159.3, 159.3, 156.4, 148.9, 140.0 (2C), 138.6, 131.1, 130.0, 129.2, 128.5, 128.3, 128.2, 127.0, 126.2 (2C), 126.0 (2C), 124.1, 122.2, 122.0, 117.2, 115.2, 114.4, 113.2, 111.7, 111.0, 98.0, 70.6 (2C), 68.6, 61.6, 55.9, 55.4, 29.3, 28.9, 28.3; IR (film) νmax 3404, 2930, 2359, 2341, 1713, 1670, 1605, 1526, 1502, 1367, 1244, 1180, 1134, 1076, 1026, 960 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]+ calcd for C₃⁹H₃₉NO₂⁺, 682.2652; found, 682.2653.

**[0356]** N-(7-((2i?,3i?,45',5i?)3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-2-oxo-8-phenyl-2 H-chromen-3-yl)-3',6-dimethoxybiphenyl-3-carboxamide (26f): Palladium on carbon (10%, 14 mg) was added to 25f (68.0 mg,
0.12 mmol) in anhydrous THF (800 µL) and the solution was placed under an atmosphere of H₂. After 12 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (52.0 mg, 99%). EDCI (18.5 mg, 0.096 mmol) and 3',6-dimethoxybiphenyl-3-carboxylic acid (19.9 mg, 0.077 mmol) were added to the amine (17.5 mg, 0.039 mmol) in 30% pyridine/CH₂Cl₂ (1.10 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford a colorless solid, which was used without further purification (14.0 mg, 52%). Triethylamine (150 µL) was added to the carbonate (14.0 mg, 0.020 mmol) in MeOH (1.5 mL) and CH₂Cl₂ (1.5 mL). After 48 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford 26f as a colorless amorphous solid (5.20 mg, 39%, 20% over 3 steps): ¹H NMR (CD₂Cl₂, 500 MHz) δ 8.85 (s, IH), 8.65 (s, IH), 7.92 (d, J = 2.4 Hz, IH), 7.90 (d, J = 2.4 Hz, IH), 7.86 (d, J = 2.4 Hz, IH), 7.57-7.43 (m, 3H), 7.36-7.33 (m, 4H), 7.11-7.06 (m, 3H), 6.92 (d, J = 0.8 Hz, IH), 5.52 (d, J = 2.4 Hz, IH), 4.08 (q, J = 7.2, Hz, IH), 3.89 (s, 3H), 3.83 (s, 3H), 3.74 (dd, J = 9.0, 3.5 Hz, IH), 3.50 (s, 3H), 3.23 (d, J = 9.0 Hz, IH), 2.12 (s, IH), 2.00 (s, IH), 1.33 (s, 3H), 1.04 (s, 3H); ¹³C NMR (CD₂Cl₂, 125 MHz) δ 164.5, 159.0 (2C), 158.6, 158.1, 154.4, 147.2, 138.0 (2C), 130.7, 130.1, 129.7, 129.1, 128.7, 128.3, 127.4, 127.2, 127.0, 127.0, 125.2, 122.6, 121.6, 121.1, 118.6, 114.5, 113.8, 112.1, 111.3, 110.2, 97.5, 70.1 (2C), 67.4, 60.8, 55.0, 54.5, 21.9, 21.6; IR (film) νmax 3402, 2932, 2359, 2341, 1713, 1603, 1524, 1500, 1367, 1267, 1086, 1040, 964, 750 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₃₈H₃₇NOI⁺ 668.2496; found, 668.2485.

[0357] N-[(2,5,3i?,4,5i?)3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methoxy-2-oxo-2 H-chromen-3-yl)-3',6-dimethoxybiphenyl-3-carboxamide (26g): Palladium on carbon (10%, 47 mg) was added to 25g (237 mg, 0.44 mmol) in anhydrous THF (2.93 mL) and the solution was placed under an atmosphere of H₂. After 12 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (177 mg, 99%). EDCI (69.4 mg, 0.36 mmol) and 3',6-dimethoxybiphenyl-3-carboxylic acid (74.8 mg, 0.29 mmol) were added to the amine (59.0 mg, 0.14 mmol) in 30% pyridine/CH₂Cl₂ (4.00 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 3:1
Hexane:Ether → 40:1 \( \text{CH}_2\text{Cl}_2: \text{Acetone} \) to afford a colorless solid, which was used without further purification (26.0 mg, 28%).

[0358] Triethylamine (150 µL) was added to the carbonate (26.0 mg, 0.040 mmol) in MeOH (2.0 mL) and \( \text{CH}_2\text{Cl}_2 \) (2.0 mL). After 48 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 \( \text{CH}_2\text{Cl}_2: \text{Acetone} \)) to afford 26g as a colorless amorphous solid (15.7 mg, 63%, 18% over 3 steps): \(^1\text{H} \text{NMR} \) (\( \text{CD}_2\text{Cl}_2 \), 400 MHz) \( \delta \) 8.82 (s, IH), 8.73 (s, IH), 7.96 (dd, \( J = 8.6 \), 2.4 Hz, IH), 7.91 (d, \( J = 2.4 \) Hz, IH) 7.39 (t, \( J = 7.9 \) Hz, IH), 7.30 (s, 2H), 7.14 (d, \( J = 8.6 \) Hz, 2H), 7.12 (d, \( J = 2.2 \) Hz, IH), 6.96 (dd, \( J = 8.3 \), 2.5 Hz, IH), 5.61 (d, \( J = 2.4 \) Hz, IH), 4.29 (t, \( J = 4.0 \) Hz, IH), 4.27-4.25 (m, IH), 3.98 (s, 3H), 3.93 (s, 3H), 3.88 (s, 3H), 3.62 (s, 3H), 3.47 (s, IH), 3.37 (d, \( J = 8.8 \) Hz, IH), 2.62 (s, IH), 1.30 (s, 3H), 1.24 (s, 3H); \(^{13}\text{C} \text{NMR} \) (\( \text{CDCl}_3 \), 125 MHz) \( \delta \) 164.5, 158.8, 158.3, 157.8, 150.2 (2C), 142.9, 137.5, 135.6, 130.0, 128.9, 128.2, 127.2, 127.2, 124.9, 122.8, 121.6, 121.0, 114.3, 114.2, 112.3, 112.1, 110.0, 97.7, 70.0 (2C), 67.5, 60.8 (2C), 54.9, 54.3, 28.7, 28.3; IR (film) \( v_{\text{max}} \) 3402, 2961, 2928, 2853, 1713, 1672, 1607, 1504, 1462, 1367, 1263, 1248, 1086, 1040, 953, 798, 735, 700 cm⁻¹; HRMS (ESI⁺) \( m/z \): [M + H]⁺ calcd for \( \text{C}_{33}\text{H}_{35}\text{NO}_5 \), 622.2288; found, 622.2307.

[0359] \( \eta^\prime-(7-(\text{2i?},3\text{i?},4\text{S},5\text{i?})-3,4\text{-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-ethyl-2-oxo-2 H-chromen-3-yl)-3\text{'},6-dimethoxybiphenyl-3-carboxamide \) (26h): Palladium on carbon (10%, 12 mg) was added to 25h (121 mg, 0.22 mmol) in anhydrous THF (5.00 mL) and the solution was placed under an atmosphere of \( \text{H}_2 \). After 12 hours, the solution was filtered through SiO₂ (1:1 \( \text{CH}_2\text{Cl}_2: \text{Acetone} \)) and the eluent was concentrated to afford a yellow solid, which was used without further purification (90.0 mg, 99%). EDCI (46.2 mg, 0.24 mmol) and 3,6-dimethoxybiphenyl-3-carboxylic acid (43.9 mg, 0.19 mmol) were added to the amine (39.0 mg, 0.096 mmol) in 30% pyridine/\( \text{CH}_2\text{Cl}_2 \) (2.65 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 \( \text{CH}_2\text{Cl}_2: \text{Acetone} \)) to afford a colorless solid, which was used without further purification (39.1 mg, 66%).

Triethylamine (150 µL) was added to the carbonate (13.0 mg, 0.020 mmol) in MeOH (1.5 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 \( \text{CH}_2\text{Cl}_2: \text{Acetone} \)) to afford 26h as a colorless amorphous solid (4.10 mg, 33%, 22% over 3 steps): \(^1\text{H} \text{NMR} \) (\( \text{CD}_2\text{Cl}_2 \), 500 MHz) \( \delta \) 8.70 (s, IH), 8.61 (s, IH), 7.83 (dd, \( J = 8.5 \), 2.5 Hz, IH), 7.78 (d, \( J = 2.5 \) Hz, IH), 7.31 (d, \( J = 9.0 \) Hz, IH), 7.27 (t, \( J = 7.8 \) Hz, IH), 7.17 (d, \( J = 8.5 \) Hz, IH), 7.03-6.99 (m, 3H), 6.85-6.82 (m, IH), 5.49 (d, \( J = 1.5 \) Hz, IH), 4.15 (t, \( J = 8.5 \) Hz, IH), 4.14 (d, \( J = 8.5 \) Hz, IH),
3.90 (s, 2H), 3.87 (s, 3H), 3.81 (s, 3H), 3.28 (s, 3H), 3.27 (d, J = 8.5 Hz, IH), 2.76 (q, J = 4.5 Hz, 2H), 1.29 (s, 3H), 1.09 (t, J = 7.4 Hz, 3H), 1.08 (s, 3H); 13C NMR (CD2Cl2, 125 MHz) δ 164.5, 159.0, 158.6, 158.5, 155.1, 147.9, 138.1, 130.1, 129.1, 128.3, 127.4, 125.4, 125.2, 123.1, 121.4, 121.2, 119.4, 114.5, 113.5, 112.1, 110.8, 110.2, 97.6, 83.4, 77.7, 70.6, 67.8, 61.0, 55.1, 54.5, 28.2, 21.6, 15.6, 12.9; IR (film) νmax 3404, 2968, 2934, 2359, 2341, 1715, 1605, 1524, 1367, 1244, 1101, 1024, 995, 960, 800 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C14H17NOi 0.12; found, 0.13.

[0360] N-7-(7-((25',3i?,4'S',5i?)3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-6-methoxy-8-methyl-2-oxo-2 H-chromen-3-yl)-7 H-indole-2-carboxamide (26i): Palladium on carbon (10%, 15 mg) was added to 25a (74.0 mg, 0.13 mmol) in anhydrous THF (5.00 mL) and the solution was placed under an atmosphere of H2. After 12 hours, the solution was filtered through SiO2 (1:1 CH2Cl2:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (60.0 mg, 99%). EDCI (69.0 mg, 0.36 mmol) and iH-indole-2-carboxylic acid (46.4 mg, 0.29 mmol) were added to the amine (60.0 mg, 0.14 mmol) in 30% pyridine/C6H5Cl2 (3.50 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO2, 3:1 Hexane:Ether → 40:1 CH2Cl2:Acetone) to afford a colorless solid, which was used without further purification (68.0 mg, 85%).

Triethylamine (150 μL) was added to the carbonate (68.0 mg, 0.12 mmol) in MeOH (2.5 mL) and CH2Cl2 (2.50 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO2, 40:1 CH2Cl2:Acetone) to afford 26i as a colorless amorphous solid (12.6 mg, 19%, 16% over 3 steps): 1H NMR (CD2Cl2, 400 MHz) δ 8.29 (s, IH), 7.63 (s, IH), 7.45-7.39 (m, 3H), 6.92 (s, IH), 6.85 (s, IH), 6.19 (s, IH), 5.09 (d, J = 6.5 Hz, IH), 4.31-4.28 (m, 2H), 4.01-3.97 (m, 2H), 3.94 (s, 3H), 3.62 (s, IH), 3.56 (s, 3H), 3.15 (d, J = 4.9 Hz, IH), 2.46 (s, 3H), 2.36 (s, IH), 1.38 (d, J = 11.5 Hz, 3H), 1.32 (s, 3H); 13C NMR (CDCl3, 125 MHz) δ 157.6, 152.1, 148.2, 145.4, 142.2, 134.5, 129.9, 127.8, 127.7, 127.5, 127.3, 122.3, 121.2, 120.2, 115.0, 105.3, 105.1, 101.3, 81.7, 69.0, 68.0, 66.5, 59.1, 55.2, 28.7, 24.6, 24.1, 9.1; IR (film) νmax 2926, 1707, 1526, 1464, 1391, 1340, 1296, 1231, 1207, 1086, 1024, 943, 739, 700, 623 cm⁻¹; HRMS (ESI+) m/z: [M + Na]⁺ calcd for C28H30N2O9, 561.1849; found, 561.1781.

[0361] N-7-(7-((25',3i?,4'S',5i?)3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-8-methyl-2-oxo-6-propoxy-2 H-chromen-3-yl)-7 H-indole-2-carboxamide (26j): Palladium on carbon (10%, 85 mg) was added to 25b (425 mg, 0.729 mmol) in anhydrous THF (4.90 mL) and the solution was placed under an atmosphere of H2. After
12 hours, the solution was filtered through SiO\textsubscript{2} (1:1 CH\textsubscript{2}Cl\textsubscript{2}:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (325 mg, 99%). EDCI (116 mg, 0.6026 mmol) and \( \text{H-indole-2-carboxylic acid} \) (77.7 mg, 0.4821 mmol) were added to the amine (108 mg, 0.2410 mmol) in 30% pyridine/CH\textsubscript{2}Cl\textsubscript{2} (6.70 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO\textsubscript{2}, 3:1 Hexane:Ether → 40:1 CH\textsubscript{2}Cl\textsubscript{2}:Acetone) to afford a colorless solid, which was used without further purification (91.0 mg, 0.1536 mmol) in MeOH (2.5 mL) and CH\textsubscript{2}Cl\textsubscript{2} (2.50 mL). After 48 hours, the solvent was concentrated and the residue purified via column chromatography (SiO\textsubscript{2}, 40:1 CH\textsubscript{2}Cl\textsubscript{2}:Acetone) to afford 26j as a colorless amorphous solid (17.5 mg, 20%, 13% over 3 steps): \(^1\text{H} \) NMR (CD\textsubscript{2}Cl\textsubscript{2}, 400 MHz) \( \delta \) 9.32 (s, IH), 8.80 (s, IH), 8.76 (s, IH), 7.77 (d, \( J = 8.0 \) Hz, IH), 7.53 (d, \( J = 7.5 \) Hz, IH), 7.40-7.36 (m, IH), 7.24-7.20 (m, IH), 6.98 (s, IH), 6.01 (s, IH), 5.15 (d, \( J = 6.5 \) Hz, IH), 4.32-4.25 (m, IH), 4.11-4.04 (m, IH), 3.62-3.59 (m, 2H), 3.53 (s, 3H), 3.18-3.12 (m, IH), 2.64 (s, IH), 2.49 (s, 3H), 2.18 (s, IH), 1.95-1.91 (m, 2H), 1.36 (d, \( J = 9.6 \) Hz, 3H), 1.29 (d, \( J = 9.8 \) Hz, 3H), 1.12 (t, \( J = 7.4 \) Hz, 3H); \(^{13}\text{C} \) NMR (CDCl\textsubscript{3}, 125 MHz) \( \delta \) 160.1, 159.0, 148.8, 146.9, 143.4, 136.9, 129.8, 127.6, 125.5, 123.5, 123.0, 122.6, 122.3, 121.2, 116.0, 112.0, 107.3, 104.3, 102.2, 82.8, 71.0, 70.1, 69.1, 60.2, 59.7, 25.7, 23.1, 23.4, 10.5, 10.2; IR (film) \( \nu_{\text{max}} \) 3630, 3304, 2926, 2854, 2359, 2332, 1713, 1705, 1539, 1387, 1240, 1103, 947, 930, 822, 739 cm\(^{-1}\); HRMS (ESI\textsuperscript{+}) \textit{m/z}: [M + H\textsuperscript{+}] \textit{calc} for C\textsubscript{29}H\textsubscript{33}N\textsubscript{2}O\textsubscript{9}, 567.2342; found, 567.2367.

[0362] \( N^1-(7-((25',3i?,4S',5i?)\text{-3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-6-isopropoxy-8-methyl-2-oxo-2 H-chromen-3-yl})-7 H-indole-2-carboxamide \) (26k): Palladium on carbon (10%, 4 mg) was added to 25c (19.0 mg, 0.033 mmol) in anhydrous THF (220 \( \mu \)L) and the solution was placed under an atmosphere of H\textsubscript{2}. After 12 hours, the solution was filtered through SiO\textsubscript{2} (1:1 CH\textsubscript{2}Cl\textsubscript{2}:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (14.5 mg, 99%). EDCI (15.6 mg, 0.081 mmol) and \( \text{H-indole-2-carboxylic acid} \) (10.5 mg, 0.065 mmol) was added to the amine (14.5 mg, 0.033 mmol) in 30% pyridine/CH\textsubscript{2}Cl\textsubscript{2} (1.00 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO\textsubscript{2}, 40:1 CH\textsubscript{2}Cl\textsubscript{2}:Acetone) to afford a colorless solid, which was used without further purification (10.0 mg, 50%). Triethylamine (150 \( \mu \)L) was added to the carbonate (10.0 mg, 0.017 mmol) in MeOH (2.5 mL) and CH\textsubscript{2}Cl\textsubscript{2} (2.50 mL). After 12 hours, the solvent was
concentrated and the residue purified via column chromatography (SiO$_2$, 10:1 CH$_2$Cl$_2$:Acetone) to afford 26k as a colorless amorphous solid (6.00 mg, 46%, 23% over 3 steps): $^1$H NMR (CDCl$_3$, 500 MHz) $\delta$ 8.16 (s, IH), 7.52 (s, IH), 7.34-7.30 (m, 5H), 6.75 (s, IH), 4.93 (d, $J = 5.0$ Hz, IH), 4.56-4.51 (m, IH), 4.23 (t, $J = 4.0$ Hz, IH), 3.98-3.96 (m, IH), 3.76 (s, IH), 3.43 (s, 3H), 3.06 (d, $J = 4.3$ Hz, IH), 2.65 (s, IH), 2.38 (s, 3H), 1.33 (dd, $J = 11.2$, 6.1 Hz, 6H), 1.29 (s, 3H), 1.27 (s, 3H); $^{13}$C NMR (CDCl$_3$, 125 MHz) $\delta$ 157.6, 152.1, 146.6, 146.0, 142.1, 134.5, 127.7, 127.5, 127.2, 122.2, 121.3 (2C), 120.2 (2C), 115.0 (2C), 108.1 (2C), 101.2, 81.6, 71.2, 68.9, 68.1, 66.5, 59.0, 24.8, 23.6, 20.8 (2C), 9.1; IR (film) $\nu_{\text{max}}$ cm$^{-1}$: 3406, 2930, 2375, 1705, 1522, 1394, 1229, 1205, 1111, 1078, 1049, 933, 793, 739, 698; HRMS (ESI$^+$) m/z: [M + Na$^+$] calcd for C$_{28}$H$_{30}$N$_2$O$_9$, 589.2162; found, 589.21 11.

[N$^\ominus$-(7-((2i?,3i?,4S,5i?)3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-5-methoxy-8-methyl-2-oxo-2 H-chromen-3-yl)-7 H-indole-2-carboxamide (261): Palladium on carbon (10%, 40 mg) was added to 25d (200 mg, 0.36 mmol) in anhydrous THF (2.40 mL) and the solution was placed under an atmosphere of H$_2$. After 12 hours, the solution was filtered through SiO$_2$ (1:1 CH$_2$Cl$_2$:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (150 mg, 99%). EDCI (57.5 mg, 0.30 mmol) and iH-indole-2-carboxylic acid (38.7 mg, 0.24 mmol) were added to the amine (50.6 mg, 0.12 mmol) in 30% pyridine/C$_2$H$_5$Cl$_2$ (3.30 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO$_2$, 40:1 CH$_2$Cl$_2$:Acetone) to afford a colorless solid, which was used without further purification (26.3 mg, 39%). Triethylamine (150 µL) was added to the carbonate (26.3 mg, 0.047 mmol) in MeOH (2.00 mL) and CH$_2$Cl$_2$ (2.00 mL). After 48 hours, the solvent was concentrated and the residue purified via column chromatography (SiO$_2$, 40:1 CH$_2$Cl$_2$:Acetone) to afford 261 as a colorless amorphous solid (6.60 mg, 26%, 10% over 3 steps): $^1$H NMR (CD$_2$Cl$_2$, 400 MHz) $\delta$ 9.26 (s, IH), 8.96 (s, IH), 8.68 (s, IH), 7.74 (d, $J = 8.1$ Hz, IH), 7.52 (d, $J = 8.3$ Hz, IH), 7.38-7.34 (m, IH), 7.22-7.16 (m, IH), 6.84 (s, IH), 6.00 (s, IH), 5.65 (d, $J = 1.7$ Hz, IH), 4.26-4.21 (m, 2H), 3.96 (s, 3H), 3.59 (s, 3H), 3.35 (d, $J = 8.6$ Hz, IH), 2.25 (s, 3H), 1.53 (s, 3H), 1.20 (s, 3H); $^{13}$C NMR (CDCl$_3$, 125 MHz) $\delta$ 165.1, 157.4, 156.5, 149.7, 136.7, 134.7, 127.7, 125.3, 122.5, 121.1, 120.0, 111.9, 104.6, 103.8, 97.7, 94.0, 84.3, 84.2, 82.6, 69.6, 69.1, 66.1, 62.2, 62.0, 59.7, 23.1, 22.7, 14.2; IR (film) $\nu_{\text{max}}$ 3389, 2924, 2853, 1697, 1605, 1535, 1460, 1340, 1211, 1101, 1088, 962, 729 cm$^{-1}$; HRMS (ESI$^+$) m/z: [M + H$^+$] calcd for C$_{28}$H$_{30}$N$_2$O$_9$, 539.2030; found, 539.2056.
ΛH8-benzyl-7-((2iU^,4S,5i0-3,4-dihydroxy-5-methoxy-6,6-
dimethyltetrahydro-2 H-pyran-2-yloxy)-2-oxo-2 H-chromen-3-yl)-7 H-indole-2-carboxamide (26m): Palladium on carbon (10%, 46 mg) was added to 25e (230 mg, 0.38 mmol) in anhydrous THF (2.50 mL) and the solution was placed under an atmosphere of H₂. After 12 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (177 mg, 99%). EDCI (61.5 mg, 0.32 mmol) and IH-indole-2-carboxylic acid (41.4 mg, 0.26 mmol) were added to the amine (60.0 mg, 0.13 mmol) in 30% pyridine/CH₂Cl₂ (3.50 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 3:1 Hexane:Ether → 40:1 CH₂Cl₂:Acetone) to afford a yellow solid, which was used without further purification (66.2 mg, 85%). Triethylamine (150 µL) was added to the carbonate (66.2 mg, 0.11 mmol) in MeOH (2.50 mL) and CH₂Cl₂ (2.50 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford 26m as a colorless amorphous solid (6.50 mg, 10%, 8.4% over 3 steps): ¹H NMR (CD₂Cl₂, 500 MHz) δ 8.62 (s, IH), 8.62 (s, IH), 7.63-7.61 (m, IH), 7.47 (dd, J = 5.7, 3.3 Hz, IH), 7.16-7.10 (m, 4H), 7.10-7.04 (m, 4H), 6.99 (t, J = 8.2 Hz, IH), 6.74 (d, J = 8.0 Hz, IH), 6.43 (dd, J = 8.1, 0.7 Hz, IH), 5.31 (d, J = 2.9 Hz, IH), 4.17 (t, J = 6.8 Hz, IH), 4.01 (dd, J = 8.5, 2.9 Hz, IH), 3.90 (d, J = 13.1 Hz, 2H), 3.45 (s, 3H), 3.16 (d, J = 8.6 Hz, IH), 2.43 (s, IH), 2.21 (s, IH), 1.25 (s, 3H), 0.95 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 166.8, 155.4, 153.8, 140.2, 131.6, 130.2, 128.0, 127.6, 127.5 (2C), 127.4 (2C), 126.9, 125.1 (2C), 115.2, 108.3 (2C), 105.8 (2C), 97.1 (2C), 83.3 (2C), 77.2, 70.2, 67.9 (2C), 65.0, 60.6, 21.9, 13.1, 13.0; IR (film) νₘₐₓ 3333, 2961, 2926, 2854, 1717, 1601, 1466, 1261, 1090, 1076, 1041, 800, 750 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na⁺] calcld for C₃₃H₃₁N₂O₈, 607.2056; found, 607.2056.

Ν-(7-((2iU^,3i,45',5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yloxy)-2-oxo-8-phenyl-2 H-chromen-3-yl)-7 H-indole-2-carboxamide (26n): Palladium on carbon (10%, 14 mg) was added to 25f (68.0 mg, 0.12 mmol) in anhydrous THF (800 µL) and the solution was placed under an atmosphere of H₂. After 12 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (52.0 mg, 99%). EDCI (18.5 mg, 0.096 mmol) and iH-indole-2-carboxylic acid (12.4 mg, 0.077 mmol) were added to the amine (17.5 mg, 0.039 mmol) in 30% pyridine/CH₂Cl₂ (1.10 mL). After 12 hours, the solvent was concentrated and the
residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford a colorless solid, which was used without further purification (8.20 mg, 36%).

Triethylamine (150 µL) was added to the carbonate (8.2 mg, 0.014 mmol) in MeOH (1.00 mL) and CH₂Cl₂ (1.00 mL). After 48 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 CH₂Cl₂:Acetone) to afford 26n as a colorless amorphous solid (4.00 mg, 51%, 18% over 3 steps): ¹H NMR (CD₂Cl₂, 500 MHz) δ 9.23 (s, IH), 8.80 (s, IH), 8.67 (s, IH), 7.71 (dd, J = 8.0, 0.7 Hz, IH), 7.57 (d, J = 8.8 Hz, IH), 7.51-7.48 (m, 3H), 7.46-7.44 (m, IH), 7.37-7.32 (m, 4H), 7.19-7.17 (m, 2H), 5.53 (d, J = 2.4 Hz, IH), 3.86 (s, IH), 3.76-3.73 (m, 2H), 3.51 (s, 3H), 3.23 (d, J = 9.1 Hz, IH), 2.41 (s, IH), 1.34 (s, 3H), 1.05 (s, 3H); ¹³C NMR (CD₂Cl₂, 125 MHz) δ 159.1, 157.9, 154.5, 147.3, 136.0 (2C), 130.7 (2C), 129.8, 129.2, 127.3, 127.1, 127.0, 126.9, 124.5, 122.8, 121.6, 121.2, 120.3, 113.7, 111.4, 111.1, 103.1 (2C), 97.5, 83.2, 77.7, 70.1, 67.5, 60.8, 21.9, 21.7; IR (film) v max 3427, 2961, 2924, 2853, 2062, 1643, 1614, 1537, 1362, 1236, 1094, 1041, 962, 791, 739, 698 cm⁻¹; HRMS (ESI⁺) m/z: [M + Na]⁺ calcd for C₃₂H₅₀N₂O₈, 593.1900; found, 593.1890.

N-(7-(25',3i?,4,S,5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyltetrahydro-2 H-pyran-2-yl)oxy)-8-methoxy-2-oxo-2 H-chromen-3-yl)-7 H-indole-2-carboxamide (26o): Palladium on carbon (10%, 47 mg) was added to 25g (237 mg, 0.44 mmol) in anhydrous THF (2.93 mL) and the solution was placed under an atmosphere of H₂. After 12 hours, the solution was filtered through SiO₂ (1:1 CH₂Cl₂:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (177 mg, 99%). EDCI (69.4 mg, 0.36 mmol) and iH-indole-2-carboxylic acid (46.7 mg, 0.29 mmol) were added to the amine (59.0 mg, 0.14 mmol) in 30% pyridine/CH₂Cl₂ (4.00 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 3:1 Hexane:Ether → 40:1 CH₂Cl₂:Acetone) to afford a colorless solid, which was used without further purification (32.0 mg, 49%).

Triethylamine (150 µL) was added to the carbonate (32.0 mg, 0.071 mmol) in MeOH (2.00 mL) and CH₂Cl₂ (2.00 mL). After 48 hours, the solvent was concentrated and the residue purified via column chromatography (SiO₂, 3:1 CH₂Cl₂:Acetone) to afford 26o as a colorless amorphous solid (22.1 mg, 73%, 35% over 3 steps): ¹H NMR (CD₂Cl₂, 400 MHz) δ 9.28 (s, IH), 8.78 (s, IH), 7.77 (d, J = 8.1 Hz, IH), 7.53 (dd, J = 8.3, 0.8 Hz, IH), 7.38 (m, IH), 7.31 (s, 2H), 7.24 (d, J = 0.9 Hz, IH), 7.22-7.20 (m, IH), 6.02 (s, IH), 5.62 (d, J = 2.3 Hz, IH), 4.25 (t, J = 3.5 Hz, IH), 3.99 (s, 3H), 3.75 (dd, J = 9.0, 3.6
Hz, IH), 3.62 (s, 3H), 3.13 (d, $J = 3.6$ Hz, IH), 1.30 (s, 3H), 1.27 (s, 3H); $^{13}$C NMR (CDCl$_3$, 125 MHz) δ 163.8, 159.1, 157.7, 150.6, 143.3, 136.0, 135.8, 129.2, 126.9, 124.6, 122.8, 121.8, 121.6, 121.4, 120.3, 114.4, 112.5, 111.2, 103.1, 98.0, 83.2, 77.9, 74.0, 60.9, 58.7, 22.3, 21.8; IR (film) $v_{\text{max}}$ 3420, 2957, 2924, 2854, 2359, 1653, 1558, 1541, 1246, 1001, 798 cm$^{-1}$; HRMS (ESI$^+$) $m/z$: [M + H]$^+$ calcd for C$_{28}$H$_{30}$N$_2$O$_8$, 545.1900; found, 545.1909.


Palladium on carbon (10%, 12 mg) was added to 25h (121 mg, 0.22 mmol) in anhydrous THF (5.00 mL) and the solution was placed under an atmosphere of H$_2$. After 12 hours, the solution was filtered through SiO$_2$ (1:1 CH$_2$Cl$_2$:Acetone) and the eluent was concentrated to afford a yellow solid, which was used without further purification (90.0 mg, 99%). EDCI (28.3 mg, 0.15 mmol) and iH-indole-2-carboxylic acid (19.0 mg, 0.12 mmol) were added to the amine (24.0 mg, 0.059 mmol) in 30% pyridine/C$_2$H$_2$Cl$_2$ (1.63 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO$_2$, 40:1 CH$_2$Cl$_2$:Acetone) to afford an colorless foam, which was used without further purification (23.8 mg, 73%). Triethylamine (150 µL) was added to the carbonate (14.1 mg, 0.026 mmol) in MeOH (1.50 mL). After 12 hours, the solvent was concentrated and the residue purified via column chromatography (SiO$_2$, 40:1 CH$_2$Cl$_2$:Acetone) to afford 26p as a yellow amorphous solid (5.00 mg, 37%, 27% over 3 steps): $^1$H NMR (CD$_2$Cl$_2$, 500 MHz) δ 8.65 (s, IH), 7.62 (d, $J = 8.0$ Hz, IH), 7.41 (d, $J = 8.5$ Hz, IH), 7.33 (d, $J = 8.5$ Hz, IH), 7.23 (t, $J = 7.8$ Hz, IH), 7.20 (d, $J = 8.5$ Hz, 2H), 7.13 (s, 2H), 7.07 (t, $J = 7.5$ Hz, IH), 5.46 (d, $J = 2.0$ Hz, IH), 4.07 (dd, $J = 9.3$, 3.5 Hz, IH), 4.04 (t, $J = 3.5$ Hz, IH), 3.51 (s, 3H), 3.25 (d, $J = 7.2$ Hz, IH), 2.78 (q, $J = 7.0$ Hz, 2H), 1.27 (s, 3H), 1.10 (t, $J = 7.5$ Hz, 3H), 1.06 (s, 3H); $^{13}$C NMR (CD$_2$Cl$_2$, 125 MHz) δ 158.4, 157.3, 154.2, 146.8, 135.3, 128.1, 125.5, 124.0, 123.0, 123.0, 122.8, 120.1, 119.5, 118.7, 118.2, 112.0, 110.1, 109.7, 102.4, 97.0, 82.1, 76.5, 69.3, 66.4, 26.7, 20.3, 14.3, 11.4; IR (film) $v_{\text{max}}$ 3435, 3416, 2974, 2935, 2469, 2359, 2339, 1715, 1651, 1520, 1456, 1435, 1379, 1354, 1259, 1180, 1113, 1088, 1026, 997, 962, 798, 739 cm$^{-1}$; HRMS (ESI$^+$) $m/z$: [M + Na]$^+$ calcd for C$_{28}$H$_{30}$N$_2$O$_8$, 545.1900; found, 545.1909.
EXAMPLE 33: SYNTHESSES OF QUINOLINE-CONTAINING AND NAPHTHALENE-CONTAINING NOVOBIOCIN ANALOGUES

[0369] tert-Butyl quinolin-7-yl carbonate (28): Di-tert-butyl dicarbonate (7.40 g, 33.92 mmol) and 4-(dimethylamino)pyridine (222 mg, 1.81 mmol) were added in sequence to 7-hydroxyquinoline (2.00 g, 13.75 mmol) in anhydrous N,N-dimethylformamide (20.0 mL) at room temperature. After 18 hours, the reaction was diluted with EtOAc (250 mL). The organic layer was washed with 1.0 M NaOH (250 mL), water (3 x 250 mL), saturated aqueous NaCl solution (250 mL), dried (Na$_2$SO$_4$), filtered, and concentrated. The residue was purified via column chromatography (SiO$_2$, 1:1 Hexanes:EtOAc) to give 28 as a colorless amorphous solid (3.26 g, 97%): $^1$H NMR (CDCl$_3$, 500 MHz) $\delta$ 8.93 (dd, $J = 4.2$, 1.7 Hz, IH), 8.17 (bd, $J = 8.4$ Hz, IH), 7.90 (d, $J = 2.2$ Hz, IH), 7.84 (d, $J = 8.9$ Hz, IH), 7.43 (dd, $J = 8.8$, 2.3 Hz, IH), 7.40 (dd, $J = 8.3$, 4.3 Hz, IH), 1.60 (s, 9H); $^{13}$C NMR (CDCl$_3$, 125 MHz) $\delta$ 151.7, 151.7, 148.9, 135.9, 129.0, 126.3, 122.0, 121.0, 120.2, 84.2, 27.8 (3C); IR (film) $\nu_{\text{max}}$ 1759, 1277, 1240, 1142, 768, 750 cm$^{-1}$; HRMS (ESI$^+$) m/z: [M + H]$^+$ calc'd for C$_{14}$H$_{15}$N$_2$O$_3$, 246.1 130; found, 246.1 113.

[0370] N-(4-Methoxybenzyl)-7-(benzyloxy)quinolin-3-amine (31): Bromine (790 µl, 2.48 g, 15.38 mmol) was added to 28 (3.26 g, 13.30 mmol) in CCl$_4$ (30.0 mL) at room temperature. This solution was heated to reflux, anhydrous pyridine (1.20 mL, 1.17 g, 14.84 mmol) was added over 10 minutes, and the solution was stirred at reflux for 18 hours. The cooled reaction was diluted with EtOAc/MeOH (250 mL) and saturated aqueous NaHCO$_3$ (200 mL), then extracted with EtOAc (4 x 250 mL). Combined organic layers were dried (Na$_2$SO$_4$), filtered, and concentrated. The residue was purified via column chromatography (SiO$_2$, 4:1 Hexanes:EtOAc) to give 29 as a light yellow
solid containing greater than 80% 3-bromo isomer (2.00 g, 46%), which was used without further purification.

[0371] Hydrogen chloride was bubbled through 29 (2.00 g, 6.16 mmol) in anhydrous MeOH (44.0 mL) for three minutes at room temperature, then the solution was stirred at 50 °C for 5 minutes. The solvent was concentrated and the residue placed under high vacuum for six hours to ensure complete removal of MeOH. The yellow residue was dissolved in anhydrous dimethylformamide (44.0 mL) and cooled to 0 °C, then NaH (997 mg, 24.93 mmol) was added. After 15 minutes, BnBr (1.20 mL, 1.73 g, 10.09 mmol) was added and the reaction was warmed to room temperature over 18 hours. Reaction contents were partitioned between saturated aqueous NaHCO₃ (500 mL) and EtOAc (500 mL), then extracted with EtOAc (3 x 500 mL). The combined organic layers were washed with water (3 x 1 L), saturated aqueous NaCl solution (1 L), dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 9:1 Hexanes:EtOAc) to give 30 as a light orange solid containing greater than 80% 3-bromo isomer (1.70 g, 88%), which was used without further purification.

[0372] 4-Methoxy-benzylamine (1.77 mL, 1.87 g, 13.64 mmol) and anhydrous DMSO (2.90 mL) were added to a high-pressure flask charged with 30 (1.65 g, 5.26 mmol), K₂PO₄ (2.36 g, 11.10 mmol), CuI (167 mg, 0.88 mmol), and Z(-)-proline (141 mg, 1.22 mmol); the sealed flask was heated to 80 °C for 44 hours. After cooling to room temperature, reaction contents were partitioned between water (50 mL) and EtOAc (100 mL), then were extracted with EtOAc (2 x 100 mL). The combined organic layers were washed with saturated aqueous NaCl solution (200 mL), dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 1:1 Hexanes:EtOAc) to give 31 as a light yellow amorphous solid (1.43 g, 73%): ¹H NMR (CDCl₃, 500 MHz) δ 8.44 (d, J = 2.8 Hz, IH), 7.52 (d, J = 8.9 Hz, IH), 7.50 (m, 2H), 7.43-7.38 (m, 3H), 7.36-7.32 (m, 3H), 7.20 (dd, J = 8.9, 2.6 Hz, IH), 7.06 (d, J = 2.7 Hz, IH), 6.91 (m, 2H), 5.17 (s, 2H), 4.35 (d, J = 4.8 Hz, 2H), 4.17 (bt, J = 4.8 Hz, IH), 3.82 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 159.3, 156.9, 143.4, 143.3, 140.5, 137.0, 130.6, 129.1 (2C), 128.8 (2C), 128.2, 127.9 (2C), 127.3, 124.7, 120.4, 114.4 (2C), 112.0, 109.0, 70.3, 55.5, 48.0; IR (film) v max 3279, 2953, 2833, 1609, 1510, 1377, 1354, 1302, 1225, 1175, 1124, 1028, 995, 868, 818, 762, 708 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₂₄H₂₂N₂O₂, 371.1759; found, 371.1732.
N-(4-Methoxybenzyl)-N-(7-(benzyloxy)quinolin-3-yl)-4-methoxy-3-(3-methoxyphenyl)benzamide (32): Thionyl chloride (395 µl, 644 mg, 5.42 mmol) was added to 4-methoxy-3-(3-methoxyphenyl)-benzoic acid (464 mg, 1.80 mmol) in anhydrous THF (6.10 mL) at room temperature and the resulting solution was heated at reflux for 4.5 hours. The solvent was removed and the resulting acid chloride was used without further purification.

Sodium hydride (93 mg, 2.31 mmol) was added to 31 (508 mg, 1.37 mmol) in anhydrous THF (9.10 mL). After stirring for two hours at room temperature, a solution of the freshly-prepared acid chloride (1.80 mmol) in anhydrous THF (3.00 mL) was added and the reaction was heated to reflux for 18 hours. The reaction was cooled, partitioned between saturated aqueous NaHCO₃ (50 mL) and CH₂Cl₂ (50 mL), then extracted with CH₂Cl₂ (3 x 100 mL). Combined organic extracts were washed with saturated aqueous NaCl solution (200 mL), dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂: 1:1 EtOAc:Hexanes to 2:1 EtOAc:Hexanes) to give 32 as a pale yellow amorphous solid (684 mg, 82%): ¹H NMR (CDCl₃, 500 MHz) δ 8.35 (d, J = 2.3 Hz, IH), 7.67 (d, J = 2.4 Hz, IH), 7.57 (d, J = 9.0 Hz, IH), 7.48 (m, 2H), 7.44-7.34 (m, 5H), 7.29 (m, 2H), 7.23 (m, 2H), 7.13 (t, J = 7.9 Hz, IH), 6.81 (m, 2H), 6.78 (dd, J = 2.5, 1.0 Hz, IH), 6.74 (d, J = 8.6 Hz, IH), 6.64 (m, IH), 6.61 (m, IH), 5.19 (s, 2H), 5.15 (s, 2H), 3.77 (s, 3H), 3.72 (s, 3H), 3.66 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 170.2, 160.2, 159.3, 159.3, 157.9, 150.9, 147.9, 138.9, 136.4, 136.0, 132.6, 132.4, 130.5, 130.2 (2C), 130.1, 129.4, 129.0, 129.0 (2C), 128.5, 127.9 (2C), 127.4, 123.1, 122.1, 121.2, 114.8, 114.2 (2C), 113.3, 110.6, 108.6, 70.5, 55.7, 55.4, 55.3, 53.8; IR (film) ν max 3032, 3001, 2953, 2935, 2835, 1645, 1603, 1512, 1456, 1429, 1385, 1331, 1248, 1209, 1178, 1034, 1026, 818, 735, 698 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₃₉H₃₄N₂O₅, 611.2546; found, 611.2574.

N-(4-Methoxybenzyl)-N-(7-(hydroxy)quinolin-3-yl)-4-methoxy-3-(3-methoxyphenyl)benzamide (33): A solution of AlCl₃ (44 mg, 0.33 mmol) in anhydrous anisole (150 µl) was added to 32 (43 mg, 0.07 mmol) in anhydrous anisole (150 µl) and the resulting solution was stirred at room temperature for 18 hours. The reaction was diluted with MeOH (150 µl) and the solvent was concentrated. The residue was purified via column chromatography (SiO₂: 80:20:1 EtOAc:Hexanes:MeOH) to give 33 as a yellow amorphous solid in quantitative yield: ¹H NMR (CDCl₃, 500 MHz) δ 8.35 (s, IH), 7.74 (s, IH), 7.58 (d, J = 8.9 Hz, IH), 7.52 (bs, IH), 7.37 (m, IH), 7.31 (m, IH), 7.25-7.11 (m, 5H), 6.84-6.77 (m, 3H), 6.74 (d, J = 8.6 Hz, IH), 6.67 (d, J = 7.6 Hz, IH),
6.64 (s, IH), 5.16 (s, 2H), 3.76 (s, 3H), 3.70 (s, 3H), 3.68 (s, 3H); 13C NMR (CDCl₃, 125 MHz) δ 170.3, 159.4, 159.3, 158.0, 138.8, 135.5, 132.3, 130.4, 130.3, 130.1 (2C), 129.6, 129.1 (2C), 127.1, 123.0, 122.0, 114.9, 114.3 (2C), 113.3, 110.7, 55.8, 55.4, 53.9; IR (film) ν_max 2926, 1601, 5164-5173 (2005), 1302, 1296, 1250, 1230, 1200, 1149, 1143 (2C), 1133, 1107, 558, 554, 539; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₃₂H₂₈N₂O₅, 521.2076; found, 521.2030.

N-(7-Hydroxyquinolin-3-yl)-4-methoxy-3-(3-methoxyphenyl)-benzamide (34): Trifluromethanesulfonic acid (120 µl, 204 mg, 1.36 mmol) was added to 32 (184 mg, 0.30 mmol) in 1:1 CH₂C=O:TFA (1.36 mL) and the solution was stirred at room temperature for two hours. The resulting solution was diluted with CH₂Cl₂ (100 mL), washed with saturated aqueous NaHCO₃ (3 x 50 mL), saturated aqueous NaCl solution (2 x 50 mL), dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 100:1 EtOAc:MeOH to 4:1 EtOAc:MeOH) to give 34 as a yellow oil (51 mg, 43%); 1H NMR (CD₃OD, 500 MHz) δ 9.01 (bs, IH), 8.61 (bs, IH), 8.02 (dd, J = 8.5, 1.7 Hz, IH), 7.97 (d, J = 1.9 Hz, IH), 7.74 (d, J = 8.9 Hz, IH), 7.30 (t, J = 8.2 Hz, IH), 7.27 (d, J = 1.9 Hz, IH), 7.19 (dd, J = 8.9, 2.1 Hz, IH), 7.17 (d, J = 8.8 Hz, IH), 7.11-7.08 (m, 2H), 6.89 (m, IH), 3.88 (s, 3H), 3.81 (s, 3H); 13C NMR (CD₃OD, 125 MHz) δ 168.7, 161.2, 160.9, 160.2, 147.1, 146.2, 140.5, 132.0, 132.0, 131.5, 130.4, 130.2, 130.1, 128.5, 127.6, 124.2, 123.2, 121.5, 116.5, 113.8, 112.4, 109.7, 56.5, 55.9; IR (film) ν_max 3281, 3203, 2930, 2835, 1605, 1545, 1499, 1371, 1252, 1036, 764, 735 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₂₄H₂₆N₂O₄, 401.1501; found, 401.1481.


[0378] 2-(Benzyloxy)-6-bromonaphthalene (36): Sodium hydride (1.16 g, 29.12 mmol) was added to 6-bromo-2-naphthol (5.00 g, 22.41 mmol) in anhydrous N,N'-dimethylformamide (162 mL) at 0 °C. After 15 minutes, benzyl bromide (2.40 mL, 3.45 g, 20.18 mmol) was added and the reaction warmed to room temperature over 18 hours. The reaction was diluted with EtOAc (500 mL), saturated aqueous NaHCO₃ (200 mL) was added, and the solution was extracted with EtOAc (3 x 500 mL). The combined organic layers were washed with water (3 x 1 L), dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 5:1 Hexanes:CH₂Cl₂) to give 36 as a colorless amorphous solid (5.71 g, 90%): ¹H NMR (CDCl₃, 500 MHz) δ 7.94 (d, J = 1.9 Hz, IH), 7.68 (d, J = 9.0 Hz, IH), 7.61 (d, J = 8.7 Hz, IH), 7.53-7.47 (m, 3H), 7.46-7.40 (m, 2H), 7.37 (m, IH), 7.26 (dd, J = 9.0, 2.5 Hz, IH), 7.20 (d, J = 2.5 Hz, IH), 5.18 (s, 2H); ¹³C NMR (CDCl₃, 125 MHz) δ 157.2, 136.8, 133.2, 130.3, 129.9, 129.9, 128.9 (2C), 128.8, 128.7, 128.4, 127.8 (2C), 120.3, 117.4, 107.3, 70.3; IR (film) νmax 1585, 1452, 1256, 1219, 1204, 1165, 1065, 997, 924, 852, 820, 800, 733, 698, 476 cm⁻¹.

[0379] N-(4-Methoxybenzyl)-6-(benzyloxy)naphthalen-2-amine (37): 4-Methoxybenzylamine (3.40 mL, 3.59 g, 26.20 mmol) and anhydrous DMSO (5.20 mL) were added to a high-pressure flask charged with 36 (3.00 g, 9.58 mmol), K₃PO₄ (4.19 g, 19.73 mmol), CuI (290 mg, 1.52 mmol), and L-(-)-proline (255 mg, 2.21 mmol); the sealed flask was heated to 80 °C for 44 hours. After cooling to room temperature, reaction contents were partitioned between water (100 mL) and EtOAc (200 mL), then were extracted with EtOAc (2 x 200 mL). The combined organic layers were washed...
with saturated aqueous NaCl solution (500 mL), dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 7:1:1 Hexanes:CH₂Cl₂:EtOAc) to give 37 as a light orange amorphous solid (952 mg, 27%): ¹H NMR (CDCl₃, 500 MHz) δ 7.55 (d, J = 8.7 Hz, 2H), 7.49 (m, 2H), 7.41 (m, 2H), 7.37-7.32 (m, 3H), 7.15 (dd, J = 8.8, 2.6 Hz, IH), 7.13 (m, IH), 6.92 (dd, J = 8.8, 2.3 Hz, IH), 6.90 (d, J = 8.7 Hz, 2H), 6.87 (m, IH), 5.14 (s, 2H), 4.35 (s, 2H), 3.82 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 159.1, 154.6, 137.4, 130.7, 129.3 (2C), 128.8 (2C), 128.1, 128.0, 127.8 (2C), 119.4 (2C), 118.6, 114.3 (2C), 107.9, 70.3, 55.5; IR (film) ν max 3059, 3032, 2999, 2934, 2835, 1636, 1601, 1506, 1456, 1389, 1248, 1209, 1176, 1068, 1000, 700 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₂₂H₂₃NO₂, 370.1807; found, 370.1786.

[0380] N-(4-Methoxybenzyl)-N-(6-(benzyloxy)naphthalen-2-yl)-4-methoxy-3-(3-methoxyphenyl)-benzamide (38): Thionyl chloride (308 µl, 502 mg, 4.22 mmol) was added to 4-methoxy-3-(3-methoxyphenyl)-benzoic acid (365 mg, 1.41 mmol) in anhydrous THF (4.75 mL) at room temperature, and the resulting solution was heated at reflux for 4.5 hours. The solvent was removed and the resulting acid chloride was used without further purification.

Sodium hydride (85 mg, 2.12 mmol) was added to 37 (378 mg, 1.02 mmol) in anhydrous THF (7.50 mL). After stirring for two hours at room temperature, a solution of the freshly-prepared acid chloride (1.41 mmol) in anhydrous THF (2.40 mL) was added and the reaction was heated to reflux for 18 hours. The reaction was cooled, partitioned between saturated aqueous NaHCO₃ (50 mL) and CH₂Cl₂ (50 mL), then extracted with CH₂Cl₂ (3 x 100 mL). Combined organic extracts were washed with saturated aqueous NaCl solution (200 mL), dried (Na₂SO₄), filtered, and concentrated. The residue was purified via column chromatography (SiO₂, 4:1:1 Hexanes:EtOAc:CH₂Cl₂) to give 38 as a brown-yellow amorphous solid (515 mg, 83%): ¹H NMR (CDCl₃, 500 MHz) δ 7.58 (d, J = 9.0 Hz, IH), 7.55 (d, J = 8.7 Hz, IH), 7.50-7.45 (m, 3H), 7.44-7.40 (m, 2H), 7.38-7.34 (m, 2H), 7.29 (d, J = 2.2 Hz, IH), 7.25 (m, 2H), 7.21 (dd, J = 8.9, 2.6 Hz, IH), 7.15 (d, J = 2.4 Hz, IH), 7.09 (m, IH), 7.01 (dd, J = 8.7, 2.1 Hz, IH), 6.80 (m, 2H), 6.77 (dd, J = 8.2, 2.7, 0.9 Hz, IH), 6.74 (d, J = 8.7 Hz, IH), 6.59-6.56 (m, 2H), 5.16 (s, 2H), 5.14 (s, 2H), 3.78 (s, 3H), 3.72 (s, 3H), 3.62 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 170.0, 159.3, 159.0, 157.6, 157.3, 139.9, 139.1, 136.8, 133.0, 132.4, 130.5, 130.1, 130.1 (2C), 129.6, 129.5, 129.1, 128.9, 128.9 (2C), 128.4, 128.2, 127.9, 127.7 (2C), 127.3, 125.8, 122.1, 120.0, 114.7, 114.0 (2C), 113.2, 110.4, 107.1, 70.3, 55.7, 55.4, 55.2, 53.9; IR (film) ν max 3059, 3032, 2999, 2934, 2835, 1636, 1601, 1506, 1456, 1389, 1248, 1209,
1026, 854, 818, 735, 698 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₄₀H₃₅N₅O₅, 610.2593; found, 610.2567.

**[0381]** N-(4-Methoxybenzyl)-N-(6-hydroxynaphthalen-2-yl)-4-methoxy-3-(3-methoxyphenyl)benzamide (39): A solution of AlCl₃ (203 mg, 1.52 mmol) in anhydrous anisole (750 µl) was added to 38 (191 mg, 0.31 mmol) in anhydrous anisole (750 µl) and the resulting solution was stirred at room temperature for 18 hours. The reaction was diluted with MeOH (750 µl) and the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 1:1 EtOAc :Hexanes) to give 39 as a light yellow amorphous solid (126 mg, 77%): ¹H NMR (CDCl₃, 500 MHz) δ 7.56 (m, 1H), 7.50 (d, J = 8.7 Hz, 1H), 7.47 (dd, J = 8.6, 2.3 Hz, 1H), 7.36 (d, J = 2.0 Hz, 1H), 7.29 (d, J = 2.2 Hz, 1H), 7.25 (m, 2H), 7.12-7.06 (m, 3H), 7.00 (dd, J = 8.7, 2.1 Hz, 1H), 6.80 (m, 2H), 6.77 (m, 1H), 6.73 (d, J = 8.7 Hz, 1H), 6.59-6.55 (m, 2H), 5.43 (s, 1H), 5.15 (s, 2H), 3.77 (s, 3H), 3.71 (s, 3H), 3.62 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 170.2, 159.3, 159.0, 157.6, 154.2, 139.7, 139.0, 133.1, 132.4, 130.5, 130.1, 130.1 (2C), 130.0, 129.5, 128.9, 128.9, 128.0, 127.5, 127.3, 125.9, 122.1, 118.7, 114.7, 114.0 (2C), 113.2, 110.4, 109.5, 55.7, 55.4, 55.3, 54.0; IR (film) ν_max 3236, 2934, 2835, 1599, 1508, 1456, 1437, 1394, 1248, 1177, 1036 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₃₃H₂₉N₅O₅, 520.2124; found, 520.2120.

**[0382]** Phenols 33, 34, 39, and 40 were noviosylated with the trichloroacetimidate of noviose carbonate (24) in the presence of boron trifluoride etherate to provide noviose carbonate analogues 41a-d as shown in the scheme below. See Shen, G.; Yu, X. M.; Blagg, B. S. J. Bioorg. Med. Chem. Lett. 14 5903-5906 (2004). In particular, analogues 33 and 34 containing the quinoline nitrogen were both slow to react and low yielding, even when greater than stoichiometric boron trifluoride etherate was employed, suggesting chelation of the quinoline nitrogen to boron was problematic. Solvolysis of carbonates 41a-d with triethylamine in methanol/dichloromethane afforded diols 42a-d in moderate yields.
N-(4-Methoxybenzyl)-N-(7-((2R,3R,4S,5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyl-tetrahydro-2H-pyran-2-yl)-quinolin-3-yl)-4-methoxy-3-(3-methoxyphenyl)-benzamide (42a): Boron trifluoride etetherate (45 µl, 52 mg, 0.36 mmol) was added to 33 (160 mg, 0.31 mmol) and (3α?,45',7i?,76i?)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3c]pyran-4-yl 2,2,2-trichloroacetimidate (181 mg, 0.50 mmol) in anhydrous CH₂Cl₂ (5.00 mL). After stirring at room temperature for 40 hours, triethylamine (30 µl) was added and the solvent was concentrated. The residue was partially purified via column chromatography (SiO₂, 1:1 EtOAc:Hexanes) to provide 41a, which was used without further purification.

Carbonate 41a was added to MeOH (22.0 mL), CH₂Cl₂ (1.5 mL), and triethylamine (2.2 mL) and stirred for 18 hours at room temperature. The solvent was concentrated and the residue purified via column chromatography (SiO₂, 50:1 EtOAc:MeOH) to give 42a as a near-colorless amorphous solid (33 mg, 16% over two steps): 1H NMR (CDCl₃, 500 MHz) δ 8.38 (d, J = 2.4 Hz, IH), 7.67 (d, J = 2.4 Hz, IH), 7.65 (d, J = 2.3 Hz, IH), 7.54 (d, J = 9.1 Hz, IH), 7.41 (dd, J = 6.8, 2.3 Hz, IH), 7.26 (d, J = 2.3 Hz, IH), 7.22 (m, 2H), 7.17 (dd, J = 8.9, 2.4 Hz, IH), 7.10 (t, J = 7.8 Hz, IH), 6.80 (m, 2H), 6.75 (dd, J = 8.4, 1.0 Hz, IH), 6.74 (d, J = 8.7 Hz, IH), 6.62-6.57 (m, 2H), 5.69 (d, J = 2.3 Hz, IH), 5.15 (AB, J_AB = 14.6 Hz, IH), 5.15 (AB, J_AB = 14.6 Hz, IH), 4.23 (dd, J = 9.1, 3.4 Hz, IH), 4.15 (m, IH), 3.76 (s, 3H), 3.70 (s, 3H), 3.64 (s, 3H), 3.59 (s, 3H), 3.38 (d, J = 9.1 Hz, IH), 1.39 (s, 3H), 1.17 (s, 3H); 13C NMR (CDCl₃, 125 MHz) δ 170.3, 159.3, 159.2, 158.1, 157.9, 150.9, 147.4, 138.8, 136.1, 132.7, 132.4, 130.5, 130.1 (2C), 129.3, 129.0, 128.9, 127.3, 123.4, 122.0, 120.7, 114.8, 114.2 (2C), 113.2, 111.3, 110.7, 98.2, 84.5, 78.7, 71.1, 68.6, 62.0, 55.7, 55.4, 55.3, 53.8, 29.0, 23.0; IR (film) v_max 2937, 2833, 1601, 1246, 1209, 1117, 1033, 989, 964 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₄₀H₄₂N₂O₉, 695.2969; found, 695.2891.

N-(7-((2R,3R,4S,5R)-3,4-Dihydroxy-5-methoxy-6,6-dimethyl-tetrahydro-2H-pyran-2-yl)-quinolin-3-yl)-4-methoxy-3-(3-methoxyphenyl)-benzamide (42b): Boron trifluoride etetherate (46 µl, 53 mg, 0.37 mmol) was added to 34 (51 mg, 0.13 mmol) and (3α?,45',7i?,76i?)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3c]pyran-4-yl 2,2,2-trichloroacetimidate (180 mg, 0.50 mmol) in anhydrous CH₂Cl₂ (5.00 mL). After stirring at room temperature for 40 hours, triethylamine (30 µl) was added and the solvent was concentrated. The residue was purified via column chromatography (SiO₂, 1:1 EtOAc:Hexanes) to give 41b (29 mg, 37%) in a 10:1 ratio of anomers (αβ), which was used without further purification.
[0386] Carbonate 41b was added to MeOH (7.5 mL), CH₂Cl₂ (500 µL), and triethylamine (750 µL) and stirred for 18 hours at room temperature. The solvent was concentrated and the residue purified via column chromatography (SiO₂, 40:1 EtOAc:MeOH) to give 42b as a near-colorless amorphous solid (11 mg, 15% over two steps): ¹H NMR (CDCl₃, 500 MHz) δ 8.81 (bs, 2H), 7.98 (dd, J = 8.7, 2.4 Hz, IH), 7.89 (d, J = 2.3 Hz, IH), 7.72 (d, J = 9.0 Hz, IH), 7.70 (d, J = 1.9 Hz, IH), 7.37 (t, J = 8.0 Hz, IH), 7.23 (dd, J = 9.0, 2.3 Hz, IH), 7.13 (dt, J = 7.8, 1.2 Hz, IH), 7.10 (m, IH), 7.09 (d, J = 8.6 Hz, IH), 6.94 (ddd, J = 8.2, 2.5, 0.8 Hz, IH), 5.73 (d, J = 2.0 Hz, IH), 4.30-4.22 (m, 2H), 3.91 (s, 2H), 3.86 (s, 3H), 3.61 (s, 3H), 3.39 (d, J = 8.9 Hz, IH), 1.40 (s, 3H), 1.21 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 165.8, 159.9, 159.6, 157.1, 146.7, 144.7, 138.9, 131.1, 130.6, 129.9, 129.4, 129.0, 128.7, 126.6, 125.0, 124.0, 122.2, 120.5, 115.6, 113.2, 111.5, 111.4, 98.0, 84.6, 78.6, 71.4, 68.8, 62.1, 56.1, 55.6, 29.2, 23.0; IR (film) ν max 3288, 2976, 2934, 2835, 1373, 1250, 1117, 731 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₃₂H₃₄N₂O₈ 575.2393; found, 575.2368.

[0387] N-(4-Methoxybenzyl)-N-(6-((2R,3R,4S,5R)-3,4-dihydroxy-5-methoxy-6,6-dimethyl-tetrahydro-2H-pyran-2-yloxy)naphthalen-2-yl)-4-methoxy-3-(3-methoxyphenyl)-benzamide (42c): Boron trifluoride etherate (10 µL, 12 mg, 0.08 mmol) was added to 39 (77 mg, 0.15 mmol) and (3α?45,7i?,7α?)7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3 αH-[1.3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetimidate (214 mg, 0.59 mmol) in anhydrous CH₂Cl₂ (5.00 mL). After stirring at room temperature for 40 hours, triethylamine (10 µL) was added and the solvent was concentrated. The residue was partially purified via column chromatography (SiO₂, 1:1 EtOAc :Hexanes) to give 41c, which was used without further purification (69 mg, 64%).

[0388] Carbonate 41c (42 mg, 0.06 mmol) was added to MeOH (9.1 mL), CH₂Cl₂ (650 µL), and triethylamine (910 µL) and stirred for 18 hours at room temperature. The solvent was concentrated and the residue purified via column chromatography (SiO₂, 2:1 EtOAc:Hexanes to EtOAc) to give 42c as a colorless oil (27 mg, 66%, 42% over two steps): ¹H NMR (CDCl₃, 500 MHz) δ 7.58 (d, J = 8.8 Hz, IH), 7.55 (d, J = 9.0 Hz, IH), 7.48 (dd, J = 8.7, 2.2 Hz, IH), 7.37 (d, J = 2.3 Hz, IH), 7.35 (d, J = 2.0 Hz, IH), 7.28 (d, J = 2.3 Hz, IH), 7.25 (m, 2H), 7.14 (dd, J = 8.9, 2.4 Hz, IH), 7.09 (t, J = 7.9 Hz, IH), 7.04 (dd, J = 8.7, 2.1 Hz, IH), 6.80 (m, 2H), 6.76 (ddd, J = 8.2, 2.5, 0.9 Hz, IH), 6.73 (d, J = 8.7 Hz, IH), 6.58 (m, IH), 6.54 (brd, J = 7.7 Hz, IH), 5.66 (d, J = 2.1 Hz, IH), 5.14 (s, 2H), 4.25 (m, IH), 4.22 (m, IH), 3.77 (s, 3H), 3.70 (s, 3H), 3.63 (s, 3H), 3.61 (s, 3H), 3.38 (d, J = 9.2 Hz, IH), 2.84 (brs, IH), 2.71 (brs, IH), 1.39 (s, 3H), 1.20 (s, 3H); ¹³C
NMR (CDCl₃, 125 MHz) δ 170.2, 159.2, 159.0, 157.6, 155.1, 140.1, 139.0, 132.9, 132.4, 130.5, 130.1 (2C), 129.5, 129.5, 129.4, 128.9, 128.3, 128.1, 127.1, 125.8, 122.1, 119.5, 114.7, 114.0, 113.2 (2C), 110.4, 109.9, 97.9, 94.6, 78.6, 71.5, 68.7, 62.1, 55.7, 55.4, 55.3, 53.9, 29.4, 22.9; IR (film) v_max 3420, 2932, 2835, 1601, 1506, 1394, 1387, 1248, 1178, 1117, 1026, 993, 910, 733 cm⁻¹; HRMS (ESI⁺) m/z: [M + H]⁺ calcd for C₁₉H₂₃NO₅ 694.3016; found, 694.3010.

[0389] N-(6-((2R,3R,4S,5R)-3,4-Dihydroxy-5-methoxy-6,6-dimethyl-tetrahydro-2H-pyran-2-yloxy)naphthalen-2-yl)-4-methoxy-3-(3-methoxyphenyl)-benzamide (42d): Trifluoromethanesulfonic acid (130 µl, 221 mg, 1.47 mmol) was added to 38 (198 mg, 0.33 mmol) in 1:1 CH₂Cl₂:ITFA (1.48 mL) and the solution was stirred at room temperature for two hours. The resulting solution was diluted with CH₂Cl₂ (100 mL), washed with saturated aqueous NaHCO₃ (3 x 50 mL), saturated aqueous NaCl solution (2 x 50 mL), dried (Na₂SO₄), filtered, and concentrated. The residue was partially purified via column chromatography (SiO₂, 1:1:1 Hexanes:EtOAc:CH₂Cl₂) to give 40 as a purple solid, which was used without further purification (28 mg, 22%).

[0390] Boron trifluoride etherate (4 µl, 5 mg, 0.03 mmol) was added to 40 (28 mg, 0.07 mmol) and (3α,4β,5α,7α)-7-methoxy-6,6-dimethyl-2-oxo-tetrahydro-3-oxH-[1.3]dioxolo[4,5-c]pyran-4-yl 2,2,2-trichloroacetimidate (83 mg, 0.23 mmol) in anhydrous CH₂Cl₂ (1.85 mL). After stirring at room temperature for 40 hours, triethylamine (10 µl) was added and the solvent was concentrated. The residue was partially purified via column chromatography (SiO₂, 3:1:1 Hexanes:EtOAc:CH₂Cl₂) to provide 41d, which was used without further purification (30 mg).

[0391] Carbonate 41d was added to MeOH (10.0 mL), CH₂Cl₂ (1.0 mL), and triethylamine (1.0 mL) and stirred for 18 hours at room temperature. The solvent was concentrated and the residue purified via column chromatography (SiO₂, 2:1 EtOAc:Hexanes, then EtOAc) to give 42d as a colorless oil (12 mg, 6% in three steps):

¹H NMR (CDCl₃, 500 MHz) δ 8.28 (d, J = 1.9 Hz, IH), 7.96 (dd, J = 8.5, 2.4 Hz, IH), 7.90 (brs, IH), 7.86 (d, J = 2.4 Hz, IH), 7.75 (brs, IH), 7.73 (brs, IH), 7.55 (dd, J = 8.9, 2.2 Hz, IH), 7.42 (d, J = 2.3 Hz, IH), 7.38 (t, J = 8.0 Hz, IH), 7.18 (dd, J = 8.9, 2.5 Hz, IH), 7.15 (m, IH), 7.11 (m, IH), 7.09 (d, J = 8.7 Hz, IH), 6.94 (ddd, J = 8.2, 2.5, 0.8 Hz, IH), 5.69 (d, J = 2.1 Hz, IH), 4.28 (dt, J = 9.2, 3.6 Hz, IH), 4.25 (m, IH), 3.91 (s, 3H), 3.87 (s, 3H), 3.62 (s, 3H), 3.39 (d, J = 9.2 Hz, IH), 2.61 (d, J = 2.5 Hz, IH), 2.56 (d, J = 3.7 Hz, IH), 1.41 (s, 3H), 1.22 (s, 3H); ¹³C NMR (CDCl₃, 125 MHz) δ 165.4, 159.6 (2C), 154.2, 139.1, 134.2, 131.8, 131.0, 129.9, 129.7, 129.4, 129.4, 128.6, 128.2, 127.4,
122.2, 120.9, 119.5, 117.3, 115.5, 113.2, 111.3, 110.1, 97.9, 84.7, 78.5, 71.6, 68.8, 62.1, 56.1, 55.6, 29.4, 22.9; IR (film) v_{max} = 3400, 2970, 2930, 2835, 1605, 1535, 1342, 1250, 1178, 1117, 1024, 995, 966, 908, 733 cm^{-1}; HRMS (ESI+) m/z: [M + H]^+ calcd for C_{33}H_{35}NO_{6}, 574.2441; found, 574.2461.

**EXAMPLE 34: BIOLOGICAL ACTIVITY OF NOVOBIOCIN ANALOGUES**

[0392] Upon construction of the library of novobiocin analogues as set forth in Examples 31-33, the compounds were evaluated for anti-proliferative activity against SkBr3 (Her2 over-expressing breast cancer cells), MCF-7 (estrogen receptor positive breast cancer cells), LnCaP (androgen receptor sensitive prostate cancer cells), and PC-3 (androgen receptor insensitive prostate cancer cells) cell lines. Cells were maintained in a 1:1 mixture of Advanced DMEM/F12 (Gibco) supplemented with non-essential amino acids, L-glutamine (2 mM), streptomycin (500 µg/mL), penicillin (100 units/mL), and 10% FBS. Cells were grown to confluence in a humidified atmosphere (37 °C, 5% CO₂), seeded (2000/well, 100 µl) in 96-well plates, and allowed to attach overnight. Compound or GDA at varying concentrations in DMSO (1% DMSO final concentration) was added, and cells were returned to the incubator for 72 hours. At 72 hours, the number of viable cells was determined using an MTS/PMS cell proliferation kit (Promega) per the manufacturer’s instructions. Cells incubated in 1% DMSO were used at 100% proliferation, and values were adjusted accordingly. IC_{50} values were calculated from separate experiments performed in triplicate using GraphPad Prism.

[0393] As shown in Table 6, the 6-substituted analogues containing the biaryl sidechain (26a-26c) exhibited consistent activity against both breast cancer cell lines. However, 26a-26c were greater than 3-fold less active against MCF-7 cells and 7-fold less active against SKBr3 breast cancer cells than analogues containing a hydrogen at this position; steric bulk at the 6-position appears detrimental to activity against breast cancer cells. In contrast, 26a-26c were between 4- and 100-fold more active against prostate cancer cells than breast cancer cells, with compound 26b more active than 26a, which was more active than 26c. Against the androgen-insensitive PC-3 prostate cancer cell line, 26c exhibited similar activity to the 6-H derivative, while 26a and 26b respectively demonstrated 2- and 10-fold increases in anti-proliferation activity. However, against androgen-sensitive LnCaP cells, introduction of a 6-alkyloxy generally reduced activity, with 26c nearly 7 times less active than the corresponding 6-H and 26a.
and 26b, respectively, demonstrating about 50% and equivalent activities. The putative binding pocket for novobiocin does not appear to present hydrogen bond donors that can interact with this region of the coumarin ring system. Similarly, 6-position analogues containing the 2-indole sidechain (26i-26k) exhibited consistent activities against all cell lines, with between 2- and greater than 50-fold increased activity versus the corresponding biaryl analogues. Analogues 26i and 26k demonstrated similar activity to one another while 26j was between 10- and 3-fold more active against each cell line. Unlike biaryl-containing 26b, 26j was twice as active as the hydrogen analogue against LnCaP cells.

![Chemical Structure](image.png)

**Table 6.** Anti-proliferation activities of coumarin-derived novobiocin analogues.

<table>
<thead>
<tr>
<th>Compound (IC_{50}, \mu M)</th>
<th>R^1</th>
<th>R^2</th>
<th>R^3</th>
<th>R^4</th>
<th>MCF-7</th>
<th>SKBr3</th>
<th>PC-3</th>
<th>LnCaP</th>
</tr>
</thead>
<tbody>
<tr>
<td>26a</td>
<td>biaryl</td>
<td>H</td>
<td>OMe</td>
<td>Me</td>
<td>&gt; 100</td>
<td>58.8 ± 1.3</td>
<td>35.4</td>
<td>6.6</td>
</tr>
<tr>
<td>26b</td>
<td>biaryl</td>
<td>H</td>
<td>OPr</td>
<td>Me</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>5.6 ± 5.7</td>
<td>3.0 ± 0.6</td>
</tr>
<tr>
<td>26c</td>
<td>biaryl</td>
<td>H</td>
<td>OPr</td>
<td>Me</td>
<td>66.9 ± 3.1</td>
<td>58.6 ± 5.4</td>
<td>60.7 ± 9.1</td>
<td>14.4 ± 4.2</td>
</tr>
<tr>
<td>26d</td>
<td>biaryl</td>
<td>OMe</td>
<td>H</td>
<td>Me</td>
<td>82.8</td>
<td>55.7 ± 6.9</td>
<td>11.3 ± 2.0</td>
<td>2.0 ± 0.8</td>
</tr>
<tr>
<td>26e</td>
<td>biaryl</td>
<td>H</td>
<td>H</td>
<td>Bn</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>49.7 ± 25.0</td>
</tr>
<tr>
<td>26f</td>
<td>biaryl</td>
<td>H</td>
<td>H</td>
<td>Ph</td>
<td>&gt; 100</td>
<td>17.3 ± 3.4</td>
<td>&gt; 100</td>
<td>1.0 ± 0.1</td>
</tr>
<tr>
<td>26g</td>
<td>biaryl</td>
<td>H</td>
<td>H</td>
<td>OMe</td>
<td>9.0 ± 5.4</td>
<td>13.9 ± 1.2</td>
<td>2.3 ± 2.9</td>
<td>1.1 ± 0.1</td>
</tr>
<tr>
<td>26h</td>
<td>biaryl</td>
<td>H</td>
<td>H</td>
<td>Et</td>
<td>41.7 ± 14.0</td>
<td>28.6 ± 1.1</td>
<td>1.8 ± 0.6</td>
<td>1.6 ± 0.3</td>
</tr>
<tr>
<td>26i</td>
<td>2-indole</td>
<td>H</td>
<td>OMe</td>
<td>Me</td>
<td>24.4 ± 1.2</td>
<td>25.1 ± 7.7</td>
<td>20.2 ± 9.8</td>
<td>10.5 ± 0.3</td>
</tr>
<tr>
<td>26j</td>
<td>2-indole</td>
<td>H</td>
<td>OPr</td>
<td>Me</td>
<td>2.1 ± 0.1</td>
<td>2.1 ± 0.8</td>
<td>6.2 ± 1.8</td>
<td>1.8 ± 0.7</td>
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<tr>
<td>26k</td>
<td>2-indole</td>
<td>H</td>
<td>OPr</td>
<td>Me</td>
<td>20.0 ± 1.0</td>
<td>20.7 ± 0.4</td>
<td>11.9</td>
<td>11.4</td>
</tr>
<tr>
<td>26l</td>
<td>2-indole</td>
<td>OMe</td>
<td>H</td>
<td>Me</td>
<td>6.1 ± 1.7</td>
<td>9.0 ± 0.8</td>
<td>11.8 ± 1.3</td>
<td>12.9 ± 4.4</td>
</tr>
<tr>
<td>26m</td>
<td>2-indole</td>
<td>H</td>
<td>H</td>
<td>Bn</td>
<td>13.2 ± 0.6</td>
<td>38.0 ± 3.0</td>
<td>73.3 ± 3.7</td>
<td>67.6 ± 6.3</td>
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<tr>
<td>26n</td>
<td>2-indole</td>
<td>H</td>
<td>H</td>
<td>Ph</td>
<td>22.9 ± 2.1</td>
<td>38.8 ± 8.3</td>
<td>28.0 ± 12.1</td>
<td>27.6 ± 10.8</td>
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</table>
[0394] Incorporation of a hydrogen-bond acceptor at the 5-position (26d, 26f) produced a similar trend in breast cancer cells, exhibiting 4- and 15-fold reduced activity against both cell lines than the compound with a hydrogen at this position, respectively. See Burlison, J. A.; Avila, C.; Vielhauer, G.; Lubbers, D. J.; Holzbeierlein, J.; Blagg, B. S. J. J. Org. Chem. 73 2130-2137 (2008). However, this effect was reduced against both prostate cell lines, as both 26d and 26f exhibited comparable activities to their 6-H analogues. Similar to 6-alkyloxy substitution, inclusion of 5-methoxy functionality appears detrimental to anti-proliferation activity. It was previously demonstrated that 8-methyl analogues were about 10-fold more active than the corresponding 8-hydrogen derivatives. To further elucidate this trend, analogues containing alternate alkyl and aryl functionalities were evaluated. Against breast cancer cells, compound 26g exhibited 2-fold improved activity over its 8-methyl counterpart, and 5-fold increased activity over the similarly-sized 8-ethyl 26h. Replacing the hydrogen bonding group with steric bulk (26h to 26e, 26f), led to compounds that were less active against MCF-7 cells. However, against SKBr3 cells, 26f and 26g were only slightly less active than the 8-methyl derivative; 26e remained inactive. Steric bulk appears detrimental to inhibitory activity at this location, however, key interactions appear favored by incorporation of short alkoxy side chains. A similar trend was observed against prostate cancer cells. While 26e and 26f were inactive against PC-3 cells, 26g and 26h both exhibited 10-fold increased activity versus the 8-methyl derivative. However, in LnCaP cells, 26e was more than twice as active as the 8-methyl derivative, while 26f was essentially inactive. In contrast, 8-position analogues containing the 2-indole sidechain (26m-26p) exhibited varying trends against both prostate and breast cancer cells. Compounds 26m, 26n, and 26p are 23-fold, 40-fold, and 7-fold less active than the 8-methyl derivative against MCF-7 cells, respectively; unusually, 26o was inactive. A similar trend was observed against SKBr3 cells, with 26p exhibiting the most potent activity. Surprisingly, both 26o and 26p were inactive against both PC-3 and LnCaP cells, while 26m was about 4-fold less active than the 8-methyl analogue; 26n exhibited slightly reduced activity in

<table>
<thead>
<tr>
<th>26o</th>
<th>2-indole</th>
<th>H</th>
<th>H</th>
<th>OMe</th>
<th>&gt;100</th>
<th>9.7 ± 1.0</th>
<th>&gt;100</th>
<th>&gt;100</th>
</tr>
</thead>
<tbody>
<tr>
<td>26p</td>
<td>2-indole</td>
<td>H</td>
<td>H</td>
<td>Et</td>
<td>4.3 ± 2.5</td>
<td>4.3 ± 3.4</td>
<td>&gt;100</td>
<td>&gt;100</td>
</tr>
</tbody>
</table>

a Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.
comparison to the 8-methyl derivative. The selectivity of 26o and 26p for breast cancer cells versus prostate cancer cells is intriguing and requires further investigation.

Table 7. Anti-proliferation activities of quinoline-containing and naphthalene-containing novobiocin analogues.

<table>
<thead>
<tr>
<th>Compound</th>
<th>R¹</th>
<th>R⁷</th>
<th>X</th>
<th>MCF-7</th>
<th>SkBr3</th>
<th>PC-3</th>
<th>LnCaP</th>
</tr>
</thead>
<tbody>
<tr>
<td>42a</td>
<td>biaryl</td>
<td>4-OMe-Bn</td>
<td>N</td>
<td>&gt; 100&lt;sup&gt;a&lt;/sup&gt;</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
</tr>
<tr>
<td>42b</td>
<td>biaryl</td>
<td>H</td>
<td>N</td>
<td>13.1 ± 4.1</td>
<td>16.5 ± 6.2</td>
<td>17.6 ± 4.6</td>
<td>14.2 ± 0.4</td>
</tr>
<tr>
<td>42c</td>
<td>biaryl</td>
<td>4-OMe-Bn</td>
<td>CH</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
</tr>
<tr>
<td>42d</td>
<td>biaryl</td>
<td>H</td>
<td>CH</td>
<td>46.4 ± 5.3</td>
<td>38.9 ± 2.4</td>
<td>10.9 ± 0.7</td>
<td>19.6 ± 1.6</td>
</tr>
</tbody>
</table>

<sup>a</sup> Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.

As shown in Table 7, compounds 42a and 42c containing the/?-MeOBn-alkylated amides did not exhibit anti-proliferative activity against the cell lines tested. This is in contrast to analogues 42b and 42d lacking the/?-MeOBn functionality, which manifested modest antiproliferative activity. This stark difference suggests one of two scenarios regarding the role of the/?-MeOBn functionality; either the/?-MeOBn group of tertiary amides 42a and 42c is unable to occupy the same pocket as the 4-aryloxy substituted novobiocin analogues. (See Radanyi, C.; La Bras, G.; Messaoudi, S.; Couclier, C.; Peyrat, J.-F.; Brion, J.-D.; Marsaud, V.; Renoir, J.-M.; Alami, M. Bioorg. Med. Chem. Lett. 18 2495-2498 (2008); Le Bras, G.; Radanyi, C.; Peyrat, J.-F.; Brion, J.-D.; Alami, M.; Marsaud, V.; Stella, B.; Renoir, J.-M. J. Med. Chem. 50 6189-6200 (2007)) or more simply, the secondary amide is required for benzamide-containing novobiocin analogues to manifest anti-proliferative activity, an observation consistent with prior structure-activity trends. It is plausible that the steric congestion of amides 42a and 42c forces adoption of a more static conformation that disallows cis/trans isomerization of the amide, a feature that has been hypothesized to be essential for antiproliferative activity of novobiocin analogues against bacteria. Further evidence was gathered upon realization that the lack of reactivity for tertiary amides 42a and 42c to all but the harshest documented conditions for/?-MeOBn removal suggest these compounds

[0396] Against breast cancer cells, analogue 42b exhibited similar anti-proliferative activities to its corresponding 8-methylcoumarin analogue, while 42d was between 2- and 5-fold less active. See Burlison, J. A.; Avila, C.; Vielhauer, G.; Lubbers, D. J.; Holzbeierlein, J.; Blagg, B. S. J. J. Org. Chem. 73 2130-2137 (2008). In contrast, both 42b and 42d were significantly more active against PC-3 cells than the corresponding 8-methylcoumarin; 42b and 42d exhibited between 7- to 9-fold reduced activity against LnCap cells. Given that both 42b and 42d lack the 8-methyl feature that yields an increased activity of about 10-fold, it is reasonable to hypothesize that the quinoline- and naphthalene-derived analogues that include an 8-methyl substituent could exhibit anti-proliferative activities between 1-5 µM against breast cancer cells and 1-2 µM against prostate cancer cells, approximately an order of magnitude less than the novobiocin analogue containing a coumarin. These results suggest that the lactone moiety may provide helpful hydrogen-bonding interactions with the novobiocin binding pocket of Hsp90, but that these interactions are not essential for anti-proliferation activity. More importantly, these results implicate that continued optimization of the coumarin scaffold is likely to produce compounds with enhanced anti-proliferative activity.

[0397] In short, compound 26g and 26j demonstrated the most potent anti-proliferative activity against the cancer cell lines tested and represent scaffolds that can be further probed to improve activity. Derivatives 26f and 26o appear to represent compounds that exhibit differential selectivity for one cancer cell lines versus another, for reasons that remain unclear. Since these compounds demonstrated low micromolar activity against one cell line and are inactive against others, they may provide a tool for further exploration and perhaps unraveling of the complicated processes affected. The activities of analogues 42b and 42d, the first documented novobiocin analogues lacking the coumarin functionality, implicate that, while the coumarin ring may participate in hydrogen bonding interactions with Hsp90 that abrogate activity, these interactions are not essential for anti-proliferation activity. These analogues provide sufficient evidence to continue the search for optimal ring systems that bridge the benzamide and noviose functionalities.
EXAMPLE 35: HETEROCYCLE SIDE CHAIN DERIVATIVES (TRIAZOLES)

[0398] In this example, the novobiocin analogues having modified side chain derivatives can be prepared. In particular, triazole derivatives can be prepared. A preferred triazole derivative the amide is substituted according to

\[
\begin{align*}
\text{N} & \equiv \text{N} \quad \text{R}^{27} \\
\text{R}^{28} & \quad \text{N}
\end{align*}
\]

wherein \(R^{27}\) is hydrogen, hydroxyl, alkoxy, or aryloxy; and wherein \(R^{28}\) is hydrogen, alkoxy, aryloxy, or amino.

[0399] The coumarin amine can be acylated with popargylic acid to form the corresponding amide, the alkyne of which can be reacted with carefully chosen azides to afford the desired triazole products according to the following scheme. The aromatic azides can consist mainly of two substitution patterns, both of which are \textit{meta} to the azide and can allow for additional van Der Walls interactions with the binding pockets by the inclusion of alkylated amines and/or ethers. Based on our CoMFA model, several of these analogues are expected to produce IC50 values in the low to mid nanomolar range.

\[
\begin{align*}
\text{MeO} & \quad \text{O} \quad \text{Me} \quad \text{N} & \equiv \text{N} \quad \text{R}^{27} \\
\text{HO} & \quad \text{OH} & \quad \text{Aryl azide} & \quad \text{Cu (cat.)} \\
\text{N} & \equiv \text{N} & \quad \text{R}^{28} & \quad \text{N}
\end{align*}
\]

wherein \(R^{27}\) is hydrogen, hydroxyl, alkoxy, or aryloxy (most preferably hydroxyl, methoxy, ethoxy, propoxy, and phenoxy); and wherein \(R^{28}\) is hydrogen, alkoxy, aryloxy, or amino (most preferably hydrogen, methoxy, ethoxy, propoxy, phenoxy, -NH\(_2\), or -N(CH\(_3\))\(_2\)).
It will be appreciated that the aforementioned scheme illustrates the preferred location of the R\textsuperscript{27} and R\textsuperscript{28} substituents.

**EXAMPLE 36: HETEROCYCLE SIDE CHAIN DERIVATIVES**
(BIARYL AMINES AND BIARYL ETHERS)

In this example, the novobiocin analogues having modified side chain derivatives can be prepared. In particular, biarylamines and biarylethers that target the B subdomain can be prepared. In particular, amides having the following biaryl substitution can be prepared:

wherein X is ether or amino (most preferably -O—, -NH-, or -NCH\textsubscript{3}-);
wherein R\textsuperscript{24} is alkoxy (most preferably methoxy);
wherein R\textsuperscript{25} is hydrogen, hydroxyl, alkoxy, or aryloxy alkyl (most preferably hydroxyl, methoxy, propoxy, or phenoxy); and
wherein R\textsuperscript{26} is hydrogen, alkoxy, aryloxy, or amino (most preferably hydrogen, methoxy, ethoxy, propoxy, phenoxy, -NH\textsubscript{2}, or -N(CH\textsubscript{3})\textsubscript{2}).

The m-iodo benzamide shown in the scheme below can be utilized to perform cross coupling reactions with amines and phenols, allowing us to further diversify the second aromatic ring in an effort to achieve greater interactions with the binding pocket. Upon solvolysis of the cyclic carbonate with methanolic triethylamine, the corresponding product can be afforded. The anilines and phenols chosen for this study are predicted to produce compounds that manifest IC\textsubscript{50} values in the mid nanomolar range.
wherein X is ether or amino (most preferably -O—, -NH-, or -NCH3-);
wherein R25 is hydrogen, hydroxyl, alkoxy, or aryloxy alkyl (most preferably hydroxyl, methoxy, propoxy, or phenoxy); and
wherein R26 is hydrogen, alkoxy, aryloxy, or amino (most preferably hydrogen, methoxy, ethoxy, propoxy, phenoxy, -NH2, or -N(CH3)2).

It will be appreciated that the aforementioned scheme illustrates the preferred location of the R24, R25, and R26 substituents.

**EXAMPLE 37: HETEROCYCLE SIDE CHAIN DERIVATIVES**

**(OXAZOLES AND PYRIDINES)**

In this example, the novobiocin analogues having modified side chain derivatives can be prepared. In particular, benzoxazoles as indole mimics can be prepared. The R29 and R30 substituents of the benzoxazole core project into regions that have been observed as beneficial for Hsp90 inhibition. Thus, in one aspect, the amide side chain is define according to:

wherein R29 is hydrogen, alkoxy, or amino; and
wherein R30 is hydrogen, alkoxy, or aryloxy.

These side chains can be prepared by coupling a variety of commercially or readily available ortho aminophenols with oxalic acid monomethyl ester to form the requisite amide as set forth in the scheme below. Simply heating this compound in the
presence of acid or treatment with \( \text{P(O)Cl}_3 \) is known to furnish the benzoxazole ring system. In addition to the molecules drawn, corresponding pyridine analogues, especially the derivative that contains a nitrogen atom in lieu of the C-R functionality, can also be prepared

wherein \( R^{29} \) is hydrogen, alkoxy, amino (most preferably hydrogen, methoxy, ethoxy, propoxy, \(-\text{NHCH}_3\), or \(-\text{N(CH}_3)_2\)); and

wherein \( R^{30} \) is hydrogen, alkoxy, aryloxy; (most preferably hydrogen, methoxy, ethoxy, phenoxy; or phenoxy).

wherein \( R^{30} \) is hydrogen, alkoxy, or aryloxy.

[0406] It will be appreciated that the aforementioned scheme illustrates the preferred location of the \( R^{29} \) and \( R^{30} \) substituents.

**EXAMPLE 38: DESIGN SCAFFOLD FOR MODIFIED SUGAR ANALOGUES OF NOVOBIOCIN**

[0407] Modified sugar analogues of the novobiocin scaffold were designed and synthesized to elucidate structure-activity relationships for the noviose sugar of novobiocin. N-heterocycles are found in a wide variety of bioactive compounds, and polyhydroxyl azasugars mimic natural sugars found in the body and act as potent inhibitors of the enzyme glycosidase. The azasugar analogues, with nitrogen inserted at various positions within the ring structure, sought to probe the hydrogen-bonding interactions with the binding pocket as well as improve solubility. The corresponding cyclohexyl analogues were designed to examine whether it was simply a hydrophobic group that was necessary to fill the sugar binding pocket. Additionally, derivatives
having a single methyl group instead of the gem-dimethyl moiety present on the parent compound, as well as an unsubstituted derivative, were prepared. Finally, these analogues aimed to determine whether the diol was a necessary component for Hsp90 inhibition.

The biaryl benzamide side chain was selected were based upon previously obtained SAR for the amide side chain as described in Burlison, J. A., Avila, C., Vielhauer, G., Lubbers, D. J., Holzbeierlein, J., Blagg, B. S. J. J. Org. Chem., 73, 2130-2137 (2008). However, it will be appreciated that other side chains at the 3-position are well within the scope of the present disclosure.

[0409] The analogues were assembled in a modular fashion allowing sequential coupling of various sugars and the biaryl acid chloride with the desired scaffold. A Mitsunobu ether coupling reaction between the coumarin phenol 1 and a sugar mimic 2 yielded the desired sugars in good yields.

[0410] More specifically, as seen in the scheme below, the phenol functionality of 5 was quantitatively protected as the corresponding ester 6 using 30% acetic anhydride in pyridine. The free amine 7 was liberated through hydrogenolysis and coupled with the biaryl acyl chloride 9, which was generated from the corresponding biaryl acid 8. Finally, solvolysis of ester 50 by triethylamine in methanol afforded phenol 1 in good

**EXAMPLE 39: SYNTHESIS OF 1,3-AZASUGAR MIMICS**

[0411] In order to avoid synthetic complexity, only a single hydroxyl group was installed on the piperidine ring prior to Mitsunobu coupling and the amine was masked by a Boc group (16). As outlined in the scheme below, this compound was synthesized in three steps, starting with the regioselective butadiene monoxide ring opening with allylamine to yield 13 and 14 in a 3:1 ratio. Subsequent protection of the amine with Boc anhydride followed by RCM metathesis in the presence of Grubbs II catalyst afforded compound 16 in good yield.
With compound 1 and 16 in hand, compound 17 was synthesized with Mitsunobu conditions. Deprotection of 17 with either 10% TFA/CH₂Cl₂ or AcCl/MeOH produced amine 18 in quantitative yield. Treatment of 18 with 1 equivalent methyl iodine and excess potassium carbonate gave tertiary amine 19, which was either dihydroxylated using OSO₄/NMO to generate 20 or reduced using 10% Pd/C in THF to furnish 21. Compounds 23 and 25 were synthesized from compound 17 in good yield using the same conditions, and subsequently deprotected by 10% TFA/CF₄Cbt to afford compounds 24 and 26, respectively. Acylation of compound 18 with 30% acetic anhydride/pyridine solution yielded 27, which was either reduced to compound 28 or dihydroxylated to compound 29, as discussed previously.

EXAMPLE 40: SYNTHESIS OF 1,4-AZASUGAR MIMICS

To elucidate the effect of different substitution patterns on the piperidine ring, a Boc-protected 1,4-azasugar was coupled with phenol 1 to give compound 29.
Subsequent deprotection led to compound 30, which was then methylated or acetylated to afford compound 31 and 32 in good yield. The synthesis of these compounds is illustrated in the scheme below.

**EXAMPLE 41: CYCLOHEXYL SUGAR ANALOGUES**

[0414] Many of the simplified cyclohexyl sugar mimics were accessible using common procedures (Scheme 6), such as one-pot reductions with lithium aluminum hydride to produce compounds 34 and 35, or via Luche conditions to yield 36. Compounds containing a double bond were designed to be coupled to the scaffold and subsequently dihydroxylated to give the corresponding diols (37-39) as shown in the scheme below.
EXAMPLE 42: STRAIGHT CHAIN ANALOGUES

[0415] The cyclic azasugar and cyclohexyl sugar analogues allow only a limited range of conformations into which the sugar portion can orient itself. A small series of corresponding aliphatic chain sugar mimics were designed to allow more flexibility to explore the possibility of additional interactions outside of those allowed by the constrained ring structures. An aliphatic amine and dihydroxylated aliphatic chain were appended to the coumarin core 1 through standard Mitsunobu coupling of the Boc-protected amine and addition of allyl bromide followed by subsequent dihydroxylation, respectively (Scheme 7).

EXAMPLE 43: FURANOSE, PYRANOSE, AND OXEPANOSE DERIVATIVES

[0416] In addition to these simplified azasugars and corresponding cyclohexyl derivatives, a series of sugars of variable ring size and substitution was synthesized to further probe the binding pocket. Although noviose is a six-membered sugar, the ability of the pocket to better accommodate a sugar of a different size has never been explored or optimized. Thus, 5- and 7-membered sugars were synthesized and coupled to the same aforementioned scaffolds to probe the pocket dimensions and attempt to establish additional favorable interactions. Moreover, although noviose is appended to novobiocin as a single anomer, the β-anomer of alternate sugars may prove more active through properly orienting the sugar in the pocket. The α- and β-anomer of each sugar were separated and tested to identify the most potent in each pair. With these considerations, a
set of mono-, di- and trihydroxylated furanoses, pyranoses and oxepanose sugars E-L was designed as shown below. These sugars were synthesized by following the literature protocols previously reported by Blagg and coworkers. See Yu et al., *Synthesis of Mono- and dihydroxylated furanoses, pyranoses, and an oxepanose for the Preparation of Natural Product Analogue Libraries*, J. Org. Chem. 70, 5599-5605 (2005), which is incorporated by reference in its entirety.

![Diagram of sugars E-L](image)

The analogues were assembled as discussed previously, in a modular fashion allowing sequential coupling of various sugars and the biaryl acid chloride with the desired scaffold, as shown in the scheme below. A Mitsunobu ether coupling reaction between the coumarin phenol 1 and sugars E-L yielded the desired analogues in good yields.

![Diagram of coupling reaction](image)

**[0418]** Coumarin phenol 1 and protected sugar F were coupled under Mitsunobu conditions using PPh$_3$ and disopropylazadicarboxylate (DIAD) in THF at room temperature for one hour, yielding an inseparable mixture of diastereomers 10a and 10b in a 3:2 ratio (Scheme 9). The cyclic carbonates of the mixture of 10a and 10b were solvolyzed with 5 eq. of lithium hydroxide in MeOH: THF: H$_2$O (1:3:1) to afford a mixture of diols 11a and 11b in 64% yield. The two diastereomers 11a and 11b were
easily separated by flash column chromatography. The assignment of stereochemistry at the anomeric carbon was accomplished by NOESY experiments.

[0419] Compounds 12a and 12b and 14a and 14b were synthesized by coupling protected sugars G and H, respectively, with coumarin 1 under standard Mitsunobu conditions. Benzoyl (Bz) deprotection of 12a and 12b was accomplished using sodium methoxide in methanol at 0°C for 10 minutes to yield 13a and 13b in a 3:2 ratio. TIPS deprotection of 14a and 14b was achieved using TBAF in THF at room temperature for one hour to furnish 15a and 15b in a 3:2 ratio. Interestingly, a single diastereomer 16 was exclusively formed when coumarin 1 and protected sugar E were subjected to standard Mitsunobu conditions. Subsequent deprotection of the acetonide and benzyl protecting groups by treatment with a catalytic amount of PTSA in methanol and hydrogenolysis, respectively, afforded triol 17 in 80% overall yield.
As shown in the scheme, protected furanose sugars J-L were coupled to coumarin 1 using Mitsunobu conditions to afford 18, 20a and 20b and 22a and 22b in 54-76% yields. Exclusively diastereomer 18 was formed through the Mitsunobu coupling between protected sugar J and coumarin 1. Hydrolysis of cyclic carbonate in 18 was performed with 5 eq. of lithium hydroxide at room temperature over one hour to obtain diol 19 in 84% yield. The removal of silyl groups in compounds 20a and 20b and 22a and 22b was achieved with TBAF in THF over one hour to furnish compounds 21a and 21b and 23a and 23b, respectively.
The protected oxepanose (I) was coupled to coumarin 1 using Mitsunobu conditions to afford a diastereomeric mixture of compounds 24. The mixture was hydrolyzed with lithium hydroxide to afford separable diastereomers 25a and 25b in a 3:2 ratio with 68% overall yield as shown in the scheme above.

**EXAMPLE 44: PHOSPHATE AND CARBAMATE SUGAR ANALOGUES**

In addition to the sugar analogues synthesized, several simplified non-sugar molecules were appended in place of the noviose sugar. The various carbamates were installed through lewis acid catalysis with the corresponding reagents. These two carbamates allow exploration of hydrogen bonding with the nucleotide binding pocket while offering much smaller groups than the bulky noviose sugar. The phosphate ester was introduced through an esterification reaction. This phosphate ester is very different from the noviose sugar, while still offering the ability to interact with the pocket and explore the substituent tolerance of the region in which the noviose sugar resides. Moreover, a phosphate ester can increase the hydrophilicity of the inhibitor, an auspicious trait when considering its potential use as a drug.
EXAMPLE 45: MESYLATE, TOSYLATE AND DIMETHYL CARBAMATE ANALOGUES

As seen in Scheme 13, a series of mesylate, tosylate, and dimethyl carbamate analogues were synthesized starting once again from the various coumarins. Although the tosylate group has been explored by the Renoir group, the coumarin scaffolds are very dissimilar when comparing the analogues. While the mesylate and tosylate explore both the hydrogen bonding network and dimensions of the pocket, the dimethyl carbamate offers contrast with the previously discussed carbamates, specifically with the hydrogen bonding capabilities and space occupied. The desired functional group was installed on the phenol coumarin using the chloride of each corresponding group, in the presence of pyridine. Next, the benzyl carbonate was removed via hydrogenolysis to produce the aminocoumarin, which was readily coupled with the biaryl benzoic acid in the presence of $\mathcal{N}$-(3-dimethylamino-propyl)-$\mathcal{N}'$-ethylcarbodiimide hydrochloride (EDCI) and pyridine.

wherein $R$ is tosylate, mesylate, or carbamate.
EXAMPLE 46: OTHER ANALOGUES

[0424] Several additional analogues were synthesized to probe the importance of the 7-phenol of the coumarin ring, specifically. These analogues incorporate similar functionalities, but these groups are attached directly to the ring without the oxygen spacer. In addition to probing the interactions of this oxygen with the binding pocket, these analogues explore the possibility that the sugar or non-sugar side chain is cleaved upon entering the cell, exposing a free phenol to interaction with the target. In order to access these analogues, the benzaldehyde precursor was alkylated in the presence of HCl gas to yield the desired alkyl chloride in modest yield. The desired 3-position regioisomer had to be separated from the 5-position isomer through several purifications. An aqueous treatment of the alkyl chloride with calcium carbonate afforded the desired benzylic alcohol in good yield. Next, the resulting benzaldehyde was converted to its formate ester via Baeyer-Villiger oxidation, and then hydrolyzed to afford the corresponding phenol. The benzylic alcohol was subsequently oxidized to form the desired benzoic acid and then standard coumarin formation procedures yielded the desired coumarin scaffold. Next, the benzyl carbonate was removed via hydrogenolysis to produce the aminocoumarin, which was readily coupled with the biaryl benzoic acid in the presence of \( N- \) (3-dimethylamino-propyl)-\( N'' \)-ethylcarbodiimide hydrochloride (EDCI) and pyridine as shown in the scheme below.
As shown in the scheme below, the 7-benzoic acid coumarin was converted to the corresponding benzyl alcohol and methyl ether through hydride reduction and esterification, respectively. In addition, a benzamide was installed through a Curtius rearrangement, then subsequently acetylated under standard coupling conditions. Finally, through Sandmeyer conditions, the 7-position benzamide was converted to a 7-position iodide. Standard Suzuki coupling conditions allowed functionalization of this position with 4-pyridine. These analogues allow further probing of key interactions with the sugar binding pocket.
EXAMPLE 47: NOVOBIOCIN ANALOGUES WITH MONO- OR DIHYDROXYLATED FURANOSE OR PYRANOSE NOVIOSE REPLACEMENTS

[0426] The presence of sugar moieties in natural products is known to play a vital role in solubility, activity and bioavailability for these compounds. Furthermore, the ring size can impart significant affinity towards their target protein. With these considerations in mind, a series of mono- and di-hydroxylated furanose and pyranose sugars (1-5 shown below) were synthesized according to previously disclosed procedures for incorporation onto the novobiocin scaffold.

[0427] Incorporation of sugars 1-5 was envisioned to occur via a Mitsunobu coupling procedure between protected pyranoses and furanoses with phenol 6, as shown retrosynthetically below.

[0428] The preparation of intermediate 6 is described and illustrated below. The coumarin phenol 7 was converted to the methoxymethyl ether using methoxymethyl chloride and Hunig's base in Dimethylformamide. The free aniline, liberated through
hydrogenolysis with 10% Pd/C and hydrogen in tetrahydrofuran from 7, was coupled with acid chloride 9 to give benzamide 10. Subsequent cleavage of the methoxymethyl ether with 4N hydrochloride in dioxane provided phenol 6 in high yield.

Once prepared, the phenol of 6 was coupled with sugars 1-5 under Mitsunobu conditions to give an inseparable diastereomeric mixture of 11-13 and 15 (Scheme 3). In the case of compound 14, a single diastereomer was formed. Subsequent hydrolysis of the cyclic carbonates and acetyl esters of 11a-b and 14 with lithium hydroxide in THF/MeOH/H₂O (3:2:2, v/v) afforded a diastereomeric mixture of 16a-b and 19, respectively. At this stage, diastereomers 16a and 16b were separated by chromatography. The assignment of stereochemistry at the anomeric center was established through two-dimensional NMR studies utilizing NOESY. In a similar manner, hydrolysis of the benzoyl and acetyl ester of 12 upon treatment with NaOMe/MeOH yielded 17a and 17b, which could be separated by chromatography. The tri-isopropylsilyl of 13 and tert-butyldimethylsilyl groups of 15 were removed by the addition of tetrabutylammonium fluoride to give separable 18a and 18b, and 20a and 20b, respectively. Syntheses of novobiocin analogues containing mono- and di-hydroxylated furanoses and pyranoses are shown below.
[0430] Benzyl 7-(methoxymethoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (8).

To a solution of benzyl 7-hydroxy-8-methyl-2-oxo-2H-chromen-3-ylcarbamate 7 (975 mg, 3.0 mmol) in anhydrous N,N-dimethylformamide (15 mL) was added N,N-diisopropylethylamine (1.01 mL, 7.5 mmol) over 5 minutes at room temperature. The resulting solution was stirred for 30 minutes and cooled to 0 °C. To it was added methoxymethylchloride (1.25 mL, 7.5 mmol) dropwise and the resulting mixture was stirred at room temperature for 3 hours. The reaction was quenched by water and the precipitate was filtered, washed with diethyl ether and dried under vacuum to give compound 8 as a white amorphous solid (931 mg, 82%). $^1$H NMR (500 MHz, acetone-$d_6$) $\delta$ 8.26 (s, 1H), 8.10 (s, 1H), 7.42 (m, 6H), 7.15 (d, 1H, $J = 8.7$ Hz), 5.34 (s, 2H), 5.26 (s, 2H), 3.49 (s, 3H), 2.29 (s, 3H). $^{13}$C NMR (100 MHz, acetone-$d_6$) $\delta$ 158.8, 157.2,
154.2, 149.9, 137.4, 129.3, 129.0, 128.9, 126.2, 123.1, 112.8, 112.3, 114.9, 115.0, 95.6, 67.6, 56.4, 8.3. IR (KBr) ν max 3321, 2948, 2931, 28591, 1724, 1693, 1608, 1541, 1367, 1213, 1155, 898 cm−1. HRMS (ESI+) m/z [M+Na+] calcd for C20H19NNaO6 392.110, found 392.1 104.

[0431] 4-(7-(Methoxymethoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (10). Palladium on carbon (10%, 85.0mg) was added to a solution of compound 8 (930 mg, 2.52 mmol) in anhydrous THF (10 mL) and the mixture was stirred under an atmosphere of hydrogen for 1 hour and filtered through a celite pad. The filtrate was concentrated and dried under vacuum for 4 hours (573 mg, 90%) before redissolved in THF (3 mL). To it was added a solution of freshly prepared acid chloride 9 (965 mg, 3.63 mmol) in THF (3 mL) followed by pyridine (0.59 mL, 7.26 mmol). The mixture was stirred at room temperature for 12 hours and concentrated to dryness. The residue was purified by column chromatography on silica using hexane and ethyl acetate (3:1) as eluent to afford a white amorphous solid 10 (925 mg, 82%). 1H NMR (400 MHz, CDCl 3) δ 8.78 (s, IH), 8.70 (s, IH), 7.81 (d, IH, J = 2.2 Hz), 7.76 (dd, IH, J = 2.3, 8.4 Hz), 7.32 (d, IH, J = 8.7 Hz), 7.17 (d, IH, J = 8.3), 7.09 (d, IH, J = 8.7 Hz), 5.27 (s, 2H), 5.24 (dd, 2H, J = 3.6, 5.0 Hz), 3.50 (s, 3H), 3.32 (d, 2H, J = 7.2 Hz), 2.35 (s, 3H), 2.34 (s, 3H), 1.77 (s, 3H), 1.73 (s, 3H). 13C NMR (100 MHz, CDCl 3) δ 169.0, 165.5, 159.4, 156.8, 152.1, 149.2, 134.7, 134.4, 131.6, 129.4, 125.9, 125.8, 124.5, 123.0, 121.8, 120.7, 115.0, 114.1, 111.5, 94.7, 56.4, 28.9, 25.8, 21.0, 18.0, 8.4. IR (KBr) ν max 3406, 2954, 2914, 1762, 1703, 1666, 1604, 1535, 1369, 1245, 1066, 989 cm−1. HRMS (ESI+) m/z [M+Na+] calcd for C26H27NNaO7 488.1685; found 488.1704.

[0432] - 2-(7-Hydroxy-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (6). To neat compound 10 (900 mg, 1.93 mmol) at room temperature was added 4M HCl in dioxane (2.42 mL, 9.67 mmol) and stirred for 1 hour. The reaction was quenched with cold water (3 mL) and the precipitate was filtered, washed with ether and dried under vacuum to afford compound 6 as a pale yellow amorphous solid (757 mg, 93%). 1H NMR (500 MHz, CDCl3) δ 8.73 (s, IH), 8.61 (s, IH), 7.76 (d, IH, J = 2.2 Hz), 7.69 (dd, IH, J = 2.3, 8.3 Hz), 7.19 (d, IH, J = 8.4 Hz), 7.17 (s, IH), 6.72 (d, IH, J = 8.4 Hz), 6.72 (d, IH, J = 8.4 Hz), 6.18 (s, IH), 5.25 (t, IH, J = 7.2 Hz), 3.32 (d, 2H, J = 7.2 Hz), 2.37 (s, 3H), 2.32 (s, 3H), 1.78 (s, 3H), 1.74 (s, 3H). 13C NMR (125 MHz, CDCl 3) δ 169.5, 165.4, 159.7, 156.2, 152.2, 149.8, 134.8, 134.5, 131.6, 129.5, 126.0, 125.9, 125.1, 123.0, 121.2, 120.7, 113.3, 113.2, 112.1, 28.9, 25.8, 21.0, 18.1, 8.0. IR
(KBr) v max 3386, 3298, 2979, 2931, 1762, 1708, 1535, 1373, 1249, 1215, 1180 cm⁻¹.

HRMS (ESI⁺) m/z [M-H⁻] calcd for C24H22NO6 420.1447; found 420.1440.

[0433] General Procedure I for the Mitsunobu coupling reaction: To a solution of 1 equivalent of phenol 10, 1.2 equivalents of sugar derivatives 1-5 and 2 equivalents of triphenylphosphine in THF (3 mL) was added 2 equivalent of diisopropylazadicarboxylate at 0 °C. The reaction mixture was stirred at room temperature for 2 hours and then quenched with water and extracted with ethyl acetate (2 X 10 mL). The combined organic extracts were washed with brine, dried over anhydrous Na₂SO₄ and concentrated to dryness under vacuum. The residues were partially purified by column chromatography on silica to give compounds 11-15 as a mixture of diastereomers except 19 as a single diastereomer (54%-88% yields).

[0434] Hydrolysis of compound 11. To a solution of substrate 11 (68 mg, 0.130 mmol) in THF: MeOH: H₂O (1.5:1:1 mL) was added lithium hydroxide (27 mg, 0.65 mmol) at room temperature. The reaction mixture was stirred for 1 hour and neutralized with saturated ammonium chloride solution, extracted with ethyl acetate (3 X 5 mL), washed with brine. The combined organic layers were dried over anhydrous Na₂SO₄ and concentrated under vacuum. The residue was purified by column chromatography on silica using dichloromethane and methanol (96:4) as eluent to give diastereomers 16a (30 mg, 47%) and 16b (20 mg, 31%), both as white amorphous solids.

[0435] N-(7-((2R,3R,4R)-3,4-dihydroxytetrahydro-2H-pyran-2-yloxy)-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (16a). ¹H NMR (500 MHz, CD₂Cl₂) δ 8.70 (s, IH), 7.65 (d, IH, J = 2.5 Hz), 7.60 (dd, IH, J = 2.5, 8.3 Hz), 7.35 (s, IH), 7.10 (d, IH, J = 8.7 Hz), 6.85 (d, IH, J = 8.4 Hz), 5.36 (d, IH, J = Hz ), 5.34 (m, IH), 4.02 (m, 2H), 3.87 (t, IH, J = 3.1 Hz), 3.52 (m, IH), 3.35 (d, 2H, J = 7.2 Hz), 2.37 (s, 3H), 1.90 (m, 2H), 1.75 (s, 3H), 1.73 (s, 3H). ¹³C NMR (125 MHz, CD₂Cl₂) δ 166.7, 159.8, 159.5, 156.8, 149.4, 133.8, 129.3, 129.2, 126.8, 126.0, 124.9, 124.2, 122.6, 122.0, 115.7, 115.1, 115.0, 112.6, 99.2, 69.0, 67.7, 57.5, 30.7, 28.6, 25.7, 17.8, 8.4. IR (KBr) v max 3303, 2977, 2923, 2852, 1708, 1606, 1404, 1282, 1087, 973 cm⁻¹. HRMS (ESI⁺) m/z [M+Na⁺] calcd for C27H29NNaO8 518.1791, found 518.1786.

[0436] N-(7-(2S,3R,4R)-3,4-dihydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (16b). ¹H NMR (500 MHz, CD₂Cl₂) δ 8.73 (s, IH), 7.65 (d, IH, J = 2.3 Hz), 7.63 (dd, IH, J = 2.3, 8.3 Hz), 7.40 (d, IH, J = 8.7 Hz), 7.17 (d, IH, J = 8.7 Hz), 6.89 (d, IH, J = 8.4 Hz), 5.57 (d, IH, J = 3.3 Hz), 5.36 (m, IH), 4.20 (m, IH), 3.92 (t, IH, J = 3.3 Hz), 3.80 (m, 2H), 3.38
(d, 2H, J = 7.2 Hz), 2.36 (s, 3H), 1.99 (m, 2H), 1.79 (s, 3H), 1.76 (s, 3H). 13C NMR (125 MHz, CD2Cl2) δ 166.8, 159.8, 159.5, 156.3, 149.4, 133.7, 129.2, 129.2, 126.8, 126.0, 124.8, 124.4, 124.4, 122.0, 115.1, 115.1, 114.7, 112.1, 99.4, 69.7, 66.2, 60.3, 29.4, 28.5, 25.7, 17.7, 8.2. IR (KBr) v_max 3390, 2958, 2925, 2854, 2520, 1706, 1604, 1404, 1249, 1074, 970 cm⁻¹. HRMS (ESI⁺) m/z [M+Na⁺] calcd for C27H29NNaO7 518.1791, found 518.1788.

**[0437]** Hydrolysis of compound 12: A solution of the 12 (55 mg, 0.094 mmol) in methanol (3 mL) was treated with sodium methoxide (51 mg, 0.94 mmol) at 0 °C and the resulting yellow solution was stirred for 20 min at room temperature. The reaction mixture was neutralized with saturated ammonium chloride, extracted with ethyl acetate (3 x 5 mL). The combined organic layers were washed with brine, dried over anhydrous Na2SO4, filtered and concentrated under vacuum. The residue was purified by column chromatography on silica using dichloromethane and methanol (98:2) as eluent to give diastereomers 17a (33 mg, 73%) and 17a (8.2 mg, 18%), both as white amorphous solids.

**[0438]** 4-Hydroxy-N-(7-((2R,3R)-3-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (17a). ¹H NMR (400 MHz, CD3OD) δ 8.72 (s, IH), 8.70 (s, IH), 7.65 (d, IH, J = 2.3 Hz), 7.61 (dd, IH, J = 2.3, 8.3 Hz), 7.37 (d, IH, J = 8.6 Hz), 7.16 (d, IH, J = 8.7 Hz), 6.86 (d, IH, J = 8.4 Hz), 5.53 (d, IH J = 3.1 Hz), 5.33 (m, IH), 3.84 (m, IH), 3.61 (m, 2H), 3.35 (d, 2H, J = 7.2 Hz), 2.40 (s, 3H), 1.95 (m, IH), 1.76 (m, IH), 1.76 (s, 3H, J = 1.6), 1.73 (s, 3H). 13C NMR (125 MHz, CDCl3:MeOH-D4) δ 166.2, 159.5, 158.8, 155.9, 148.9, 134.1, 129.1, 126.5, 125.7, 124.8, 124.0, 122.0, 121.3, 115.0, 114.9, 114.5, 112.1, 97.3, 68.1, 60.3, 29.7, 28.5, 27.4, 25.8, 24.1, 17.8, 8.3. IR (KBr) v_max 3405, 2927, 2856, 2524, 1697, 1604, 1504, 1369, 1253, 1120, 983 cm⁻¹. HRMS (ESI⁺) m/z [M+Na⁺] calcd for C27H29NNaO7 502.1842, found 502.1830.

**[0439]** 4-Hydroxy-N-(7-((2S,3R)-3-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (17b). ¹H NMR (500 MHz, CDCl3) δ 8.72 (s, IH), 7.64 (d, IH, J = 2.4 Hz), 7.58 (dd, IH, J = 2.4, 8.3 Hz), 7.30 (d, IH, J = 8.7 Hz), 7.11 (d, IH, J = 8.7 Hz), 6.81 (d, IH, J = 8.4 Hz), 5.49 (d, IH, J = Hz), 5.31 (m, IH), 3.80 (m, IH), 3.58 (m, 3H), 3.33 (d, 2H, J = 6.8 Hz), 2.36 (s, 3H), 1.95 (s, 3H), 1.74 (m, 2H), 1.73 (s, 3H), 1.70 (s, 3H). 13C NMR (125 MHz, CD2Cl2) δ 166.0, 159.7, 159.6, 157.2, 144.9, 133.4, 129.9, 129.3, 127.4, 126.5, 126.0, 124.1, 123.3, 123.0, 115.7, 115.2, 115.0, 112.9, 101.6, 67.7, 63.6, 28.9, 28.9, 25.9, 22.6, 17.9, 8.4. IR (KBr)
v$_{max}$ 3407, 2929, 2858, 2522, 1699, 1606, 1520, 1358, 1253, 1109, 983 cm$^{-1}$. HRMS (ESI$^+$) m/z [M+Na$^+$] calcd for C27H29NNaO7 502.1842, found 502.1830.

[0440] Deprotection of silyl group and acetate hydrolysis of compound 13: To a solution of compound 13 (98 mg, 0.154 mmol) in THF (4 mL) was added a 1M solution of tetrabutylammonium fluoride in THF (0.31 mL, 0.31 mmol) at 0 °C. The resulting mixture was stirred at room temperature for 1 hour and quenched with a saturated ammonium chloride solution (2 mL), extracted with ethyl acetate (3 x 5 mL). The organic phases were combined, washed with brine, dried over anhydrous Na$_2$SO$_4$, filtered and concentrated under vacuum. The residue was purified by column chromatography on silica by using dichloromethane and methanol (98:2) as eluent to give diastereomers 18a (36 mg, 49%) and 18b (24 mg, 33%), both as white amorphous solids.

[0441] 4-Hydroxy-N-(7-((2S,4R)-4-hydroxytetrahydro-2H-pyran-2-yl-oxy)-7-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide 18a. $^1$H NMR (400 MHz, CD$_3$OD) δ 8.69 (s, IH), 7.65 (d, IH, J = 2.4 Hz), 7.61 (dd, IH, J = 2.4, 8.3 Hz), 7.37 (d, IH, J = 8.7 Hz), 7.09 (d, IH, J = 8.7 Hz), 6.86 (d, IH, J = 8.4 Hz), 5.35 (m, IH), 5.23 (dd, IH, J = 2.5, 7.5 Hz), 4.08 (d, IH, J = 12.0 Hz), 3.94 (s, IH), 3.56 (t, IH, J = 9.7 Hz), 3.35 (d, 2H, J = 7.1 Hz), 2.32 (s, 3H), 2.29 (m, IH), 1.90 (m, IH), 1.79 (m, IH), 1.76 (s, 3H), 1.73 (s, 3H), 1.63 (m, IH). $^{13}$C NMR (125 MHz, CD$_2$Cl$_2$) δ 169.0, 161.9, 161.6, 158.9, 151.4, 135.6, 131.2, 128.8, 127.9, 126.8, 126.6, 124.4, 124.1, 117.2, 117.0, 116.7, 114.4, 100.4, 67.5, 63.1, 41.6, 35.9, 30.5, 27.6, 19.6, 10.1. IR (KBr) v$_{max}$ 3396, 2956, 2925, 2864, 2518, 1708, 1604, 1504, 1446, 1367, 1249, 1110, 977 cm$^{-1}$. HRMS (ESI$^+$) m/z [M+H$^+$] calcd for C27H30NO7 480.2022, found 480.1658.

[0442] 4-Hydroxy-N-(7-((2R,4R)-4-hydroxytetrahydro-2H-pyran-2-yl-oxy)-7-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide 18b. $^1$H NMR (500 MHz, acetone-d6) δ 8.76 (s, IH), 8.71 (s, 2H), 7.77 (d, IH, J = 2.4 Hz), 7.71 (d, IH, J = 2.4, 8.4 Hz), 7.50 (d, 2H, J = 8.6 Hz), 7.21 (d, 2H, J = 8.7 Hz), 7.00 (d, 2H, J = 8.4 Hz), 5.86 (d, IH, J = 2.9 Hz), 5.39 (m, IH), 4.28 (m, IH), 3.99 (br s, OH), 3.77 (m, 2H), 3.40 (d, 3H, J = 7.1 Hz), 2.32 (s, 3H), 2.26 (m, IH), 1.96 (m, IH), 1.84 (m, IH), 1.77 (s, 6H), 1.62 (m, IH). $^{13}$C NMR (125 MHz, acetone-d6) δ 166.0, 159.7, 159.6, 156.9, 149.9, 133.4, 129.9, 129.3, 127.4, 126.5, 126.0, 124.2, 124.1, 123.2, 123.0, 115.7, 114.9, 112.6, 97.7, 63.5, 60.5, 40.3, 35.8, 28.9, 25.9, 17.9, 8.4. IR (KBr) v$_{max}$ 3400, 2960, 2929, 2885, 2522, 1706, 1604, 1502, 1367, 1253, 1085, 970 cm$^{-1}$. HRMS (ESI$^+$) m/z [M+H$^+$] calcd for C27H29NO7 480.1658, found 480.1658.

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[0443] 4-(7-((2S,3S,4S)-3,4-Dihydroxytetrahydrofuran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (19). To a solution of 14 (54 mg, 0.106 mmol) in THF: MeOH: H2O (1.5:1:1 mL) was added lithium hydroxide (27 mg, 0.53 mmol) at room temperature. The reaction mixture was stirred for 1 hour and neutralized with saturated ammonium chloride solution, extracted with ethyl acetate (3 X 5 mL), washed with brine. The combined organic layers were dried over anhydrous Na2SO4, and concentrated under vacuum. The residue was purified by column chromatography on silica using dichloromethane and methanol (97:3) as eluent to give 19 (35 mg, 68%) as a white amorphous solid. 1H NMR (500 MHz, acetone-d6) δ 9.14 (br s, 1H), 8.76 (s, IH), 8.71 (s, IH), 7.77 (d, IH, J = 1.9 Hz), 7.71 (dd, IH, J = 1.9, 8.4 Hz), 7.50 (d, IH, J = 8.6 Hz), 7.17 (d, IH, J = 8.7 Hz), 6.99 (d, IH, J = 8.4 Hz), 5.67 (d, IH, J = 1.7 Hz), 5.38 (m, IH), 4.53 (m, IH), 4.39 (m, IH), 4.17 (dd, IH, J = 5.3, 9.3 Hz), 3.89 (dd, IH, J = 3.8, 9.3 Hz), 3.40 (d, IH, J = 7.3 Hz), 2.26 (s, 3H), 1.75 (s, 6H). 13C NMR (125 MHz, CDCl3) δ 166.0, 159.5, 157.2, 149.9, 133.4, 129.9, 126.5, 126.0, 124.1, 124.0, 123.3, 122.9, 115.7, 115.2, 15.1, 113.0, 107.9, 79.2, 77.2, 73.9, 71.2, 28.8, 25.9, 17.9, 8.4. IR (KBr) \( \nu_{\text{max}} \) 3396, 2962, 1604, 1569, 1479, 1273, 1054, 981 cm\(^{-1}\). HRMS (ESI) m/z [M-H] calcd for C26H26NO8 480.1658, found 480.1658.

[0444] Silyl group deprotection and acetate hydrolysis of compound 13: To a solution of compound 15 (43 mg, 0.074 mmol) in THF (3 mL) was added a 1M solution of tetrabutylammonium fluoride in THF (0.15 mL, 0.15 mmol) at 0 °C. The resulting reaction mixture was stirred at rt for 1 h, quenched with saturated ammonium chloride (2 mL) and extracted with EtOAc (3 x 4 mL). The organic phases were combined, washed with brine, dried over anhydrous Na2SO4, filtered and concentrated under vacuum. The residue was purified by column chromatography on silica using dichloromethane and methanol (98:4) as eluent to give diastereomers 20a (16 mg, 47%) and 20b (11 mg, 31%), both as white amorphous solids.

[0445] 4-(7-((2S,4R)-4-Hydroxytetrahydrofuran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (20a). 1H NMR (400 MHz, acetone-d6) δ 9.13 (br s, IH), 8.77 (s, IH), 8.72 (s, IH), 7.77 (d, IH, J = 2.2 Hz), 7.71 (dd, IH, J = 2.2, 8.2 Hz), 7.50 (d, IH, J = 8.9 Hz), 7.25 (d, IH, J = 8.6 Hz), 6.99 (d, IH, J = 8.3 Hz), 6.07 (dd, IH, J = 2.7, 5.6 Hz), 5.39 (m, IH), 4.69 (m, IH), 4.20 (s, IH), 4.05 (dd, IH, J = 4.8, 9.6 Hz), 3.86 (d, IH, J = 9.7 Hz), 3.60 (s, IH), 3.39 (d, 2H, J = 7.3 Hz), 2.48 (m, IH), 2.35 (m, IH), 2.25 (s, 3H), 1.75 (s, 6H). 13C NMR (125 MHz, CDCl3 and MeOH-d4) δ 166.4, 159.6, 159.0, 156.3, 148.9, 133.7, 128.9, 128.7, 126.4, 125.6, 194
124.5, 124.1, 121.4, 115.1, 114.8, 114.4, 112.5, 103.3, 76.5, 70.4, 41.7, 29.6, 28.2, 25.7, 17.7, 8.4. IR (KBr) v max 3305, 2960, 2931, 2875, 1708, 1604, 1404, 1367, 1282, 1112, 1058, 974 cm⁻¹. HRMS (ESI) m/z [M-H⁻] calcd for C26H26NO7 464.1709, found 464.1706.

NOVOBIOCIN ANALOGUES WITH ALKYLAMINE AND HETEROCYCLIC NOVIOSE REPLACEMENTS

Although simplified sugar mimics were found to increase the anti-proliferative activity -200 times greater than novobiocin, more simplified analogues exhibiting enhanced solubility and activity were desired. N-Heterocycles are found in a variety of biologically active compounds, and in contrast to carbohydrates, are generally ionized at physiological pH. Upon review of the first set of studies, we proposed that the noviose appendage was responsible for solubilizing the predominately hydrophobic...
coumarin core and benzamide side chain. Thus, commercially available amines, 21-27, shown below, were selected as potential replacements for the noviose moiety. These alkylamines and heterocyclic analogues contain an ionizable amine located at various positions within the structure to afford potential hydrogen-bonding interactions while simultaneously enhancing solubility.

Originally, coupling of these amines with phenol 6 were expected to easily afford the desired analogues. However, the acetyl ester on the benzamide side chain was hydrolyzed under these conditions and resulted in an inseparable mixture of mono- or dialkylated products. To circumvent this issue, the amine was coupled with the coumarin ring and subsequently with the benzamide side chain to afford the desired analogues. The detailed synthesis is described as follows: tertiary amines or Boc-masked secondary amines were reacted with Cbz-protected coumarin in the presence of two equivalents of triphenylphosphine and diisopropylazodicarboxylate in tetrahydrofuran to give amine-derived coumarins, 28a-28f. The Cbz-protecting group was removed by hydrogenolysis to give the free amines, which were then coupled with acid chloride 9 to give compounds 29a-29g in good yield. Removal of the Boc protecting group with trifluoroacetic acid in methylene chloride afforded the secondary amine analogues 30b, 30d and 30g. Hydrolysis of the phenolic ester with 10% triethyl amine in methanol gave denoviosylated analogues, 31a-31g, in good to excellent yield. Synthesis of the amine containing analogues is shown below.
Benzyl 2-(8-methyl-7-(1-methylpiperidin-4-yloxy)-2-oxo-2H-chromen-3-yl amino)-2-oxoacetate (28a). General procedure II for Mitsunobu coupling: To a solution of 21 (0.64 g, 5.59 mmol), 7 (1.74 g, 5.59 mmol) and triphenylphosphine (2.93 g, 11.2 mmol) in THF (50 mL) was added diisopropylazodicarboxylate (2.26 g, 11.2 mmol) dropwise. The reaction mixture was stirred at room temperature for 4 hours and concentrated under vacuum. The residue was purified by column chromatography on silica using methylene chloride and methanol (10:1) as eluent to afford 28a as colorless foam (1.48 g, 63%). 

$^1$H MNR (400 MHz, CDCl$_3$) $\delta$ 8.28 (s, 1H), 7.55 (s, 1H), 7.45-7.38 (m, 5H), 7.29 (d, $J$ = 8.0, 1H), 6.87 (d, $J$ = 8.0, 1H), 5.25 (s, 2H), 4.49 (m, 1H), 2.72-2.68 (m, 2H), 2.52-2.49 (m, 2H), 2.38 (s, 3H), 2.35 (s, 1H), 2.11-2.05 (m, 2H), 1.96-1.92 (m, 2H). 

$^{13}$C MNR (100 MHz, CDCl$_3$) $\delta$ 159.1, 156.8, 153.4, 149.4, 135.8, 128.9, 128.7, 128.4, 125.3, 122.5, 121.6, 115.4, 113.5, 110.7, 77.4, 67.6, 52.4, 46.4, 30.8, 8.6. IR (film) $\nu_{max}$ 3406, 3319, 2939, 2849, 2791, 1711, 1609, 1524, 1366, 1271, 1227,
1204, 1103, 1038, 1024 cm$^{-1}$. HRMS (ESI$^+$) m/z Calc for [M+H$^+$] C24H27N2O5 423.1920, found 423.1920.

[0451] Benzyl 2-(8-methyl-7-(1-methylpiperidin-3-yl)oxy)-2-oxo-2H-chromen-3-ylamino)-2-oxoacetate (28c). Prepared from 22 (81 mg, 0.70 mmol) and 7 (215 mg, 0.70 mmol). 28c was obtained as colorless foam (64 mg, 22%). $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 8.22 (s, 1H), 7.76 (s, 1H), 7.39-7.33 (m, 5H), 7.22 (d, $J$ = 8.0, 1H), 6.88 (d, $J$ = 8.0, 1H), 5.21 (s, 2H), 4.39 (m, 1H), 2.97-2.95 (m, 1H), 2.64-2.62 (m, 1H), 2.30 (s, 3H), 2.25 (s, 1H), 2.21 (m, 1H), 2.16 (m, 1H), 2.07-2.05 (m, 2H), 1.85-1.82 (m, 1H), 1.64-1.62 (m, 1H), 1.48-1.46 (m, 1H). $^{13}$C NMR (100 MHz, CDCl$_3$) $\delta$ 158.8, 157.0, 153.3, 149.1, 135.7, 128.6, 128.4, 128.3, 125.1, 122.5, 121.4, 115.0, 111.3, 110.7, 74.1, 67.3, 59.7, 55.4, 46.3, 29.8, 23.1, 8.3. IR (film) $\nu_{max}$ 3408, 3302, 2978, 2926, 2853, 1713, 1609, 1522, 1464, 1375, 1290, 1236, 1107 cm$^{-1}$. HRMS (ESI$^+$) m/z Calc for [M+H$^+$] C24H27N2O5 423.1920, found: 423.1920.

[0452] Benzyl 7-(2-(dimethylamino)ethoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (28e). Prepared from 25 (40 mg, 0.50 mmol) and 7 (138 mg, 0.50 mmol). 28e was obtained as colorless oil (126mg, 64%). $^1$H NMR (400 MHz, CDCl$_3$) $\delta$ 8.24 (s, 1H), 7.58 (s, 1H), 7.40-7.34 (m, 5H), 7.25 (d, $J$ = 8.6, 1H), 6.83 (d, $J$ = 8.6, 1H), 5.22 (s, 2H), 4.14 (t, $J$ = 5.6, 2H), 2.80 (t, $J$ = 5.6, 2H), 2.37 (s, 6H), 2.29 (s, 3H). $^{13}$C NMR (100 MHz, CDCl$_3$) $\delta$ 159.0, 158.1, 153.3, 149.0, 135.8, 128.8, 128.6, 128.3, 125.3, 122.4, 121.4, 114.2, 113.4, 108.9, 67.6, 67.5, 58.3, 46.3, 21.9, 8.3. IR (film) $\nu_{max}$ 3406, 3302, 2980, 2939, 2878, 2824, 2773, 1713, 1609, 1524, 1456, 1383, 1227, 1111, 1024 cm$^{-1}$. HRMS (ESI$^+$) m/z Calc for [M+H$^+$] C22H25N2O5 397.1763, found: 397.1759.

[0453] Benzyl 7-(3-(dimethylamino)propoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamate (28f). Prepared from 26 (230 mg, 2.3 mmol) and 7 (703 mg, 2.3 mmol). 28f was obtained as colorless oil (670 mg, 72%). $^1$H NMR (400MHz, CDCl$_3$): $\delta$ 8.11 (s, 1H), 7.45 (s, 1H), 7.29-7.23 (m, 5H), 7.11 (d, $J$ = 8.6, 1H), 6.70 (d, $J$ = 8.6, 1H), 5.10 (s, 2H), 3.97 (t, $J$ = 5.8, 2H), 2.46 (t, $J$ = 5.8, 2H), 2.22 (s, 6H), 2.17 (s, 3H), 1.93 (m, 2H). $^{13}$C NMR (100 MHz, CDCl$_3$) $\delta$ 159.0, 158.2, 153.3, 149.0, 135.8, 128.8, 128.6, 128.4, 125.3, 122.5, 121.4, 114.0, 113.3, 108.9, 67.5, 66.8, 56.4, 45.4, 27.4, 8.2. IR (film) $\nu_{max}$ 3404, 3323, 2978, 2943, 2816, 2768, 1713, 1610, 1524, 1381, 1366, 1273, 1227, 1204, 1109, 1022 cm$^{-1}$. HRMS (ESI$^+$) m/z Calc for [M+H$^+$] C23H26N2O5 411.1920, found: 411.1918.

[0454] 4-(8-Methyl-7-(1-methylpiperidin-4-yl)oxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29a). General procedure III for the
Preparation of benzamide 29a-g: Palladium on carbon (10%, 10 mg) was added to a solution of 28a (85 mg, 0.2 mmol) in THF. The suspension was stirred overnight under a hydrogen atmosphere before it was filtered. The filtrate was concentrated and dried under vacuum for 4 hours before it was redissolved in THF (5 mL). To it was added freshly prepared acid chloride 9 (80 mg, 0.3 mmol) in THF (5 mL) and dry pyridine (32 mg, 0.4 mmol). The resulting mixture was stirred at room temperature overnight and concentrated to dryness. The residue was purified by column chromatography on silica using methylene chloride and methanol (10:1) as eluent to afford 29a as white amorphous solid (76 mg, 73%). 1H NMR (500MHz, CDCl₃), δ 8.79 (s, IH), 8.71 (s, IH), 7.81 (d, J = 2.1, IH), 7.77 (dd, J = 8.3, 2.1, IH), 7.34(d, J = 8.6, IH), 7.18 (d, J = 8.3, IH), 6.89 (d, J = 8.6, IH), 5.25 (m, IH), 4.52 (m, IH), 3.33 (d, J = 7.2, 2H), 2.76-2.70 (m, 2H), 2.55-2.45 (m, 2H), 2.40 (s, 3H), 2.35 (s, 6H), 2.15-2.09 (m, 2H), 1.97-1.92 (m, 2H), 1.78 (s, 3H), 1.74 (s, 3H). 13C NMR δ(125MHz, CDCl₃) 169.1, 165.6, 159.6, 157.1, 152.3, 149.7, 134.8, 134.5, 131.8, 129.5, 126.1, 125.8, 124.8, 123.1, 121.7, 120.8, 115.4, 113.5, 110.6, 77.4, 52.3, 46.2, 30.5, 29.0, 25.9, 21.1, 18.1, 8.6. IR (film) νmax 3400, 2922, 2851, 1765, 1711, 1672, 1607, 1526, 1493, 1369, 1248, 1202, 1175, 1099, 1040 cm⁻¹. HRMS (ESI⁺) m/z Calc for [M+H⁺] C30H35N2O6 519.2495, found: 519.2485.

[0455] 4-(8-Methyl-7-(1-methylpiperidin-3-yloxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-ynyl)phenyl acetate (29c). Compound 29c was obtained as a white amorphous solid (72 mg, 92%). 1H NMR (500MHz, CDCl₃) δ 8.77 (s, IH), 8.70 (s, IH), 7.81 (d, J = 2.1, IH), 7.77 (dd, J = 8.3, 2.3, IH), 7.32(d, J = 8.7, IH), 7.17 (d, J = 8.3, IH), 6.97 (d, J = 8.7, IH), 5.24 (m, IH), 4.54 (m, IH), 3.31 (d, J = 7.2, 2H), 3.18-3.16 (m, IH), 2.86-2.83 (m, IH), 2.41 (s, 3H), 2.34 (s, 3H), 2.31 (s, 3H), 2.29-2.12 (m, 3H), 1.91-1.86 (m, IH), 1.76 (s, 3H), 1.73 (s, 3H), 1.69-1.65 (m, IH), 1.54-1.47 (m, IH). 13C NMR δ(125MHz, CDCl₃) 169.0, 165.5, 159.6, 157.3, 152.2, 149.6, 134.7, 134.4, 131.8, 129.5, 126.0, 125.9, 124.7, 123.0, 121.6, 120.8, 115.4, 113.6, 110.9, 73.4, 59.2, 55.1, 45.8, 29.6, 28.9, 25.9, 22.7, 21.0, 18.1, 8.5. IR (film) νmax 3402, 2939, 2856, 2786, 1763, 1711, 1672, 1607, 1526, 1491, 1367, 1250, 1204, 1173, 1099 cm⁻¹. HRMS (ESI⁺) m/z Calc for [M+H⁺] C30H35N2O6 519.2495, found: 519.2493.

[0456] 4-(7-(2-(Dimethylamino)ethoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-ynyl)phenyl acetate (29e). Compound 29e (26 mg, 84%) was obtained as a light yellow, amorphous solid. 1H NMR (500MHz, CDCl₃) δ 8.79 (s, IH), 8.70 (s, IH), 7.81 (s, IH), 7.77(d, J = 8.3, IH), 7.34 (d, J = 8.7, IH), 7.18 (d, J =
8.3, (IH), 6.88 (d, J = 8.7, IH), 5.25 (m, IH), 4.22 (t, J = 5.5, 2H), 3.32 (d, J = 7.2, 2H),
2.91 (t, J = 5.5, 2H), 2.46 (s, 6H), 2.35 (s, 3H), 2.34 (s, 3H), 1.77 (s, 3H), 1.73 (s, 3H).
\[^{13}\text{C NMR} \text{ (125MHz, CDCl}_3 \text{) } \delta 169.1, 165.6, 159.7, 158.4, 152.3, 149.4, 134.8, 134.5,
131.8, 129.5, 126.1, 126.0, 124.8, 123.1, 121.7, 120.8, 114.4, 113.7, 109.1, 67.2, 58.2,
46.0, 29.0, 26.0, 21.1, 18.1, 8.4. IR (film) \nu_{\text{max}}^\text{IR} 3393, 2961, 2859, 1761, 1705, 1664,
1607, 1528, 1491, 1369, 1267, 1246, 1202, 1177, 1109 \text{ cm}^{-1}. \text{HRMS (ESI+)} m/z \text{Calc for}
[M+H+] C28H33N2O6 493.2339, found: 493.2336.

[0457] 4-(7-(3-(Dimethylamino)propoxy)-8-methyl-2-oxo-2H-chromen-3-
ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29f). Compound 29f (79 mg, 77%) was obtained as a white, light yellow, amorphous solid. \[^1\text{H NMR} \text{ (500MHz, CDCl}_3 \text{) } \delta
8.79 (s, IH), 8.70 (s, IH), 7.81 (s, IH), 7.77(d, J = 8.3, IH), 7.34 (d, J = 8.6, IH), 7.18
(d, J = 8.3, IH), 6.88 (d, J = 8.6, IH), 5.25 (m, IH), 4.15 (t, J = 6.0, 2H), 3.32 (d, J = 7.2,
2H), 2.77 (t, J = 7.4, 2H), 2.49 (s, 6H), 2.35 (s, 3H), 2.33 (s, 3H), 2.17 (m, 2H), 1.77 (s,
3H), 1.73 (s, 3H). \[^{13}\text{C NMR} \text{ (125MHz, CDCl}_3 \text{) } \delta 169.1, 165.6, 159.7, 158.4, 152.3,
149.4, 134.8, 134.5, 131.8, 129.6, 126.09, 126.05, 124.9, 123.1, 121.7, 120.9, 114.3,
113.6, 109.1, 66.7, 56.4, 45.0, 29.0, 26.8, 26.0, 21.1, 18.2, 8.4. IR (film) \nu_{\text{max}}^\text{IR} 3393, 2964,
2930, 1761, 1705, 1666, 1607, 1549, 1371, 1246, 1202, 1182, 1107 \text{ cm}^{-1}. \text{HRMS (ESI+)} m/z \text{Calc for}

[0458] 4-(8-methyl-2-oxo-7-(piperidin-4-oylo)-2H-chromen-3-ylcarbamoyl)-2-(3-
methylbut-2-enyl)phenyl acetate (30b). General procedure III for preparation of 30b, 30d
and 30g through Mitsunobu coupling, benzamide coupling and Boc deprotection: Boc
protected amine 23 (99 mg, 0.49 mmol) was coupled with 7 (152 mg, 0.49 mmol)
following general procedure II to afford partially purified 28b (160 mg), which went
through hydrogenolysis and coupled with acid chloride following the described general
procedure III to give compound 29b. Compound 29b was dissolved in 1% trifluoroacetic
acid in methylene chloride, stirred at room temperature for 2 hours and
concentrated. The residue was purified by column chromatography on silica by using
methylenec chloride and methanol (10:1) to give 30b as a light brown, amorphous solid
(106 mg, 3 steps, 43%). \[^1\text{H NMR} \text{ (400MHz, DMSO}_d^\text{6} \text{) }, 9.69 (s, IH), 8.50 (s, IH),
7.86 (s, IH), 7.84 (d, J = 8.3, IH), 7.61 (d, J = 8.4, IH), 7.25 (d, J = 8.3, IH), 7.18 (d, J =
8.4, IH), 5.20 (m, IH), 4.84 (m, IH), 3.28 (d, J = 7.2, 2H), 3.21 (m, 2H), 3.11 (m, 2H),
2.29 (s, 3H), 2.25 (s, 3H), 2.17-2.13 (m, 2H), 1.93-1.91 (m, 2H), 1.70 (s, 3H), 1.69 (s,
3H). \[^{13}\text{C NMR} \text{ (125MHz, DMSO}_d^\text{6} \text{) } \delta 168.9, 165.2, 158.1, 156.3, 151.5, 149.8, 133.6,
132.8, 131.4, 129.6, 129.4, 126.6, 126.3, 122.9, 121.4, 121.3, 113.5, 112.9, 110.6, 69.7,
40.2, 28.3, 27.1, 25.5, 20.7, 17.7, 8.1. IR (film) _ν_ max 3400, 2984, 2854, 1745, 1718, 1678, 1607, 1535, 1442, 1371, 1125, 1142, 1103 cm⁻¹. HRMS (ESI⁺) m/z Calc for [M+H⁺] C29H33N2O6 505.2339, found: 505.2340.

[0459] 4-(8-methyl-2-oxo-7-(piperidin-3-yloxy)-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (3Od). Compound 3Od was obtained as a light brown, amorphous solid (31 mg, 3 steps, 22%). ¹H NMR (500MHz, CDCl₃-CD₃OD), δ 8.68 (s, IH), 7.71 (s, IH), 7.68 (d, J = 8.4, IH), 7.30(d, J = 8.6, IH), 7.09 (d, J = 8.4, IH), 6.86 (d, J = 8.8, IH), 5.15 (m, IH), 4.56 (m, IH), 3.27-3.22 (m, 3H), 3.10-3.06 (m, IH), 2.99-2.97 (m, 2H), 2.27 (s, 3H), 2.26 (s, 3H), 2.03-1.82 (m, 4H), 1.68 (s, 3H), 1.64 (s, 3H). ¹³C NMR (125MHz, CDCl₃) δ 169.1, 165.6, 159.3, 155.8, 152.3, 149.5, 134.8, 134.5, 131.6, 129.6, 126.1, 125.9, 124.2, 123.1, 122.1, 120.8, 115.8, 114.4, 110.2, 69.4, 45.0, 43.9, 29.0, 27.6, 25.9, 25.7, 21.1, 18.1, 8.4. IR (film) _ν_ max 3402, 2968, 2935, 2860, 1763, 1701, 1676, 1607, 1528, 1493, 1439, 1369, 1252, 1204, 1178, 1140, 1099 cm⁻¹. HRMS (ESI⁺) m/z Calc for [M+H⁺] C29H33N2O6 505.2339, found: 505.2340.

[0460] 4-(8-methyl-7-(2-(methylamino)ethoxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (30g). Compound 30g was obtained as a light yellow, amorphous solid (53 mg, 3 steps, 34%). ¹H NMR (CDCl₃, 500MHz), δ 8.63 (s, IH), 7.66 (s, IH), 7.63 (dd, J = 8.3, 2.1, IH), 7.28(d, J = 8.6, IH), 7.05 (d, J = 8.3, IH), 6.82 (d, J = 8.6, IH), 5.10 (m, IH), 4.20 (t, J = 5.0, 2H), 3.26 (t, J = 5.0, 2H), 3.18 (d, J = 7.2, 2H), 2.62 (s, 3H), 2.21 (s, 6H), 1.62 (s, 3H), 1.59 (s, 3H). ¹³C NMR δ(CDCl₃, 125MHz) 169.4, 165.9, 159.4, 157.4, 152.1, 149.2, 134.7, 134.3, 131.4, 129.3, 126.0, 125.9, 125.1, 122.9, 121.6, 120.5, 114.3, 114.0, 109.1, 64.7, 49.5, 33.8, 28.7, 25.5, 20.7, 17.7, 7.8. IR (film) _ν_ max 3393, 2964, 2918, 2849, 1767, 1710, 1676, 1605, 1528, 1491, 1369, 1252, 1202, 1178, 1136, 1111 cm⁻¹. HRMS (ESI⁺) m/z Calc for [M+H⁺] C27H31N2O6 479.2182, found: 479.2181.

[0461] 4-Hydroxy-N-(8-methyl-7-(1-methylpiperidin-4-yloxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31a). General procedure IV for the preparation of compounds 31a-g: Compound 29a (52 mg, 0.1 mmol) was dissolved in 10% triethylamine/methanol (3mL). The solution was stirred at room temperature overnight and concentrated. The residue was purified by column chromatography on silica by using methylene chloride and methanol (10:1) to give 31a as a white, amorphous solid (37 mg, 73%). ¹H NMR (500MHz, DMSO) δ 9.22 (s, IH), 8.47 (s, IH), 7.68-7.66 (m, 2H), 7.53 (d, J = 8.7, IH), 7.11 (d, J = 8.9, IH), 6.90 (d, J = 8.7, IH), 5.30 (m, IH), 4.57 (m, IH), 3.27 (d, J = 7.3, 2H), 2.62-2.54 (m, 2H), 2.34-2.28 (m, 2H), 2.22 (s, 3H), 2.21 (s,
3H), 1.96-1.92 (m, 2H), 1.76-1.72 (m, 2H), 1.71 (s, 3H), 1.69 (s, 3H). $^{13}$C NMR (125MHz, DMSO$_d$$_6$) $\delta$ 165.3, 158.8, 158.4, 156.6, 149.5, 131.9, 129.2, 127.8, 127.7, 126.8, 126.0, 124.0, 122.3, 121.3, 114.6, 113.4, 112.7, 110.8, 72.1, 51.8, 45.7, 30.2, 27.9, 25.6, 17.7, 8.2. IR (film) $\nu_{\text{max}}$ 3421, 1703, 1666, 1601, 1528, 1504, 1366, 1248, 1178, 1150, 1094, 1040 cm$^{-1}$. HRMS (ESI$^+$) m/z Calc for [M+H$^+$] C28H33N2O5 477.2389, found: 477.2397.

[0462] 4-Hydroxy-N-(8-methyl-2-oxo-7-(piperidin-3-yloxy)-2H-chromen-3-yl)-3-(3-methylbut-2-ethyl)benzamide (31b). Compound 31b was obtained as a light brown, amorphous solid (13 mg, 56%). $^1$H NMR (500MHz, DMSO$_d$$_6$) $\delta$ 9.21 (s, 1H), 8.47 (s, IH), 7.67-7.65 (m, 2H), 7.53 (d, $J = 8.7$, IH), 7.12 (d, $J = 8.9$, IH), 6.90 (d, $J = 8.7$, IH), 5.31 (m, IH), 4.58 (m, IH), 3.27 (d, $J = 7.3$, 2H), 2.95-2.93 (m, 2H), 2.62-2.58 (m, 2H), 2.23 (s, 3H), 1.92-1.90 (m, 2H), 1.71 (s, 3H), 1.70 (s, 3H), 1.54-1.45 (m, 2H). $^{13}$C NMR (125MHz, DMSO$_d$$_6$) $\delta$ 1529, 1522, 140, 1367, 122.4, 122.4, 121.3, 114.6, 113.4, 113.4, 112.6, 110.9, 10.3, 43.3, 32.1, 32.0, 25.6, 17.7, 8.2. IR (film) $\nu_{\text{max}}$ 3401, 2959, 2927, 2872, 2858, 1724, 1693, 1643, 1532, 1529, 1447, 1367, 1261, 1117, 1072 cm$^{-1}$. HRMS (ESI$^+$) m/z Calc for [M+H$^+$] C27H31N2O5 463.2233, found: 463.2245.

[0463] 4-Hydroxy-N-(8-methyl-7-(1-methylpiperidin-3-yloxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31c). Compound 31c was obtained as a white, amorphous solid (19 mg, 69%). $^1$H NMR (400MHz, CDCl$_3$-CD$_3$OD) $\delta$ 8.69 (s, IH), 7.62 (d, $J = 2.1$, IH), 7.56 (dd, $J = 8.4$, 2.1, IH), 7.29 (d, $J = 8.7$, IH), 6.90 (d, $J = 8.7$, IH), 6.80 (d, $J = 8.3$, IH), 5.30 (m, IH), 4.42 (m, IH), 3.32 (d, $J = 7.1$, 2H), 3.05-3.02 (m, IH), 2.74-2.71 (m, IH), 2.32 (s, 3H), 2.27 (s, 3H), 2.25-2.06 (m, 3H), 1.85-1.81 (m, IH), 1.72 (s, 3H), 1.69 (s, 3H), 1.67-1.63 (m, IH), 1.50-1.40 (m, IH). $^{13}$C NMR (100MHz, CD$_2$Cl$_2$-CD$_3$OD) 5167.1, 160.3, 159.9, 157.6, 150.0, 133.9, 129.52, 129.47, 127.1, 126.3, 125.0, 122.8, 122.4, 122.3, 115.7, 115.3, 114.3, 111.3, 73.7, 59.6, 55.6, 46.1, 29.7, 28.8, 25.9, 22.8, 18.0, 8.4. IR (film) $\nu_{\text{max}}$ 3423, 1710, 1663, 1603, 1528, 1375, 1277, 1256, 1204, 1140 cm$^{-1}$. HRMS (ESI$^+$) m/z Calc for [M+H$^+$] C28H33N2O5 477.2389, found: 477.2391.

[0464] 4-Hydroxy-N-(8-methyl-2-oxo-7-(piperidin-3-yloxy)-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31d). Compound 31d was obtained as a light brown, amorphous solid (21 mg, 72%). $^1$H NMR (500MHz, CDCl$_3$-CD$_3$OD), $\delta$ 8.43 (s, IH), 7.38 (d, $J = 2.2$, IH), 7.34 (dd, $J = 8.3$, 2.2, IH), 7.11 (d, $J = 8.6$, IH), 6.71 (d, $J = 8.6$, IH), 6.60 (d, $J = 8.3$, IH), 5.07 (m, IH), 4.42 (m, IH), 3.08 (d, $J = 7.2$, 2H), 3.02 (m,
IH), 2.77-2.75 (m, 2H), 2.07 (s, 3H), 1.79-1.65 (m, 3H), 1.48 (s, 3H), 1.48-1.45 (m, IH), 1.45 (s, 3H). 13C NMR (125 MHz, CD2Cl2-CD3OD) δ 167.6, 163.2, 162.9, 160.2, 156.9, 150.2, 133.7, 129.7, 127.3, 126.7, 125.0, 122.6, 118.6, 116.3, 115.6, 115.4, 114.9, 111.2, 70.3, 54.0, 47.4, 44.6, 28.8, 27.4, 25.8, 19.3, 17.8, 8.3. IR (film) νmax 3402, 2959, 2928, 2872, 2858, 1726, 1668, 1605, 1526, 1502, 1454, 1366, 1259, 1120, 1072 cm⁻¹. HRMS (ESI⁺) m/z Calc for [M+H⁺] C27H31N2O5 463.2233, found: 463.2226.

[0465] N-(7-(2-(Dimethylamino)ethoxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (3le). Compound 31e was obtained as a white, amorphous foam (8.4 mg, 61%). 1H NMR (400MHz, CD2Cl2-CD3OD) δ 8.58 (s, IH), 7.60 (d, J = 2.4, IH), 7.54 (dd, J = 8.4, 2.4, IH), 7.33 (d, J = 8.7, IH), 6.90 (d, J = 8.7, IH), 6.82 (d, J = 8.4, IH), 5.31 (m, IH), 4.16 (t, J = 6.2, 2H), 3.32 (d, J = 7.2, 2H), 2.85 (t, J = 6.9, 2H), 2.39 (s, 6H), 2.26 (s, 3H), 1.74 (s, 3H), 1.71 (s, 3H). 13C NMR (125MHz, CD2Cl2-MeOD ) δ 167.3, 160.3, 160.1, 158.9, 149.9, 133.8, 129.6, 129.5, 127.1, 126.6, 125.4, 125.0, 122.5, 122.2, 115.3, 114.5, 114.3, 109.5, 67.1, 58.4, 45.8, 28.8, 25.8, 17.8, 8.2. IR (film) νmax 3408, 2968, 2930, 2883, 2862, 1757, 1705, 1664, 1605, 1501, 1367, 1263, 1178, 1109 cm⁻¹. HRMS (ESI⁺) m/z Calc for [M+H⁺] C26H31N2O5 451.2233, found: 451.2231.

[0466] N-(7-(3-(Dimethylamino)propoxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (3lf). Compound 31f was obtained as a white, amorphous solid (22 mg, 69%). 1H NMR (500MHz, DMSOδ) δ 9.24 (s, IH), 8.47 (s, IH), 7.68-7.66 (m, 2H), 7.56 (d, J = 8.7, IH), 7.07 (d, J = 8.7, IH), 6.89 (d, J = 8.7, IH), 5.31 (m, IH), 4.13 (t, J = 6.2, 2H), 3.27 (d, J = 7.2, 2H), 2.47 (t, J = 6.9, 2H), 2.22 (s, 3H), 2.21 (s, 6H), 1.92 (m, 2H), 1.71 (s, 3H), 1.70 (s, 3H). 13C NMR (125MHz, DMSOδ) δ 165.3, 158.7, 158.4, 158.1, 149.3, 131.8, 129.2, 128.1, 127.6, 126.8, 126.1, 124.0, 122.3, 121.2, 114.5, 112.7, 112.4, 109.1, 66.6, 55.5, 45.0, 40.1, 27.9, 26.6, 25.5, 17.7, 7.9. IR (film) νmax 3408, 2961, 2928, 1709, 1666, 1607, 1529, 1504, 1367, 1256, 1178, 1109 cm⁻¹. HRMS (ESI⁺) m/z Calc for [M+H⁺] C27H33N2O5 465.2389, found: 465.2388.

[0467] 4-Hydroxy-N-(8-methyl-7-(2-(methylamino)ethoxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (3lg). Compound 31g was obtained as a light brown, amorphous solid (11 mg, 46%). 1H NMR (400MHz, CDCl3-CD3OD) δ 8.42 (s, IH), 7.38 (d, J = 2.3, IH), 7.33 (dd, J = 8.4, 2.4, IH), 7.11 (d, J = 8.7, IH), 6.68 (d, J = 8.7, IH), 6.59 (d, J = 8.4, IH), 5.07 (m, IH), 3.94 (t, J = 5.0, 2H), 3.08 (d, J = 7.2, 2H), 2.88 (t, J = 5.0, 2H), 2.32 (s, 3H), 2.06 (s, 3H), 1.49 (s, 3H), 1.45 (s, 3H). 13C NMR
(100MHz, CDCl$_3$-MeOD) δ 166.5, 159.4, 159.1, 157.6, 148.8, 133.0, 128.6, 128.5, 126.1, 125.6, 124.5, 124.4, 123.9, 121.3, 114.4, 113.7, 113.5, 108.8, 66.2, 49.3, 34.6, 27.7, 25.1, 17.1, 7.4. IR (film) $\nu_{\text{max}}$ 3398, 2956, 2924, 2854, 1697, 1655, 1650, 1533, 1508, 1373, 1261, 1111 cm$^{-1}$. HRMS (ESI$^+$) m/z Calc for [M+H$^+$] C25H29N2O5 437.2076, found: 437.2076.

[0468] Biological evaluation of novobiocin analogues with amine substituents. Anti-proliferative activity manifested by these analogues was assessed against SKBr3 and MCF-7 cell lines. As shown in Table 19, the IC$_{50}$ values for the secondary and tertiary amines vary between 0.4-1.5 µM, making them 500 to 1500-fold more potent than novobiocin and -10-20 fold more efficacious than DHN2.

EXAMPLE 49: BIOLOGICAL EVALUATION OF NOVOBIOCIN ANALOGUES

[0469] Upon construction of the modified sugar and non-sugar analogues of novobiocin, the compounds were evaluated for anti-proliferative activity against SKBr3 (estrogen receptor negative, Her2 over-expressing breast cancer cells), MCF-7 (estrogen receptor positive breast cancer cells), LN3 (metastatic human prostate cancer cells), MM2 (metastatic mammary tumor cancer cells) and A549 (human alveolar basal epithelial cells) cell lines.

[0470] Cells were maintained in a 1:1 mixture of Advanced DMEM/F12 (Gibco) supplemented with non-essential amino acids, L-glutamine (2 mM), streptomycin (500 µg/mL), penicillin (100 units/mL), and 10% FBS. Cells were grown to confluence in a humidified atmosphere (37 °C, 5% CO$_2$), seeded (2000/well, 100 µL) in 96-well plates, and allowed to attach overnight. Compound or GDA at varying concentrations in DMSO (1% DMSO final concentration) was added, and cells were returned to the incubator for 72 hours. At 72 hours, the number of viable cells was determined using an MTS/PMS cell proliferation kit (Promega) per the manufacturer’s instructions. Cells incubated in 1% DMSO were used at 100% proliferation, and values were adjusted accordingly. IC$_{50}$ values were calculated from separate experiments performed in triplicate using GraphPad Prism.

[0471] The compounds in the following tables were synthesized. Biological data was collected on selected compounds, while biological data on the remaining compounds is in progress at the time of this filing ("TBD"). As shown in the tables below, the IC$_{50}$ values of secondary amines and tertiary amines vary between 1-2 µM, making them 300-
500-fold more active than novobiocin. While dihydroxylation on the piperidine ring decreases the anti-proliferative activity, inclusion of unsaturation on the piperidine does not affect activity. It was observed that acetylation of amine severely decreases the solubility, leading to inactive compounds. This insolubility can be overcome through installation of dihydroxy groups on the amide increases the solubility, but the resulting compound is still about 5-fold less active than secondary or tertiary amines.

Table 8. Biological evaluation of unsaturated azasugar analogues.

<table>
<thead>
<tr>
<th>Compound (IC₅₀, μM)</th>
<th>X</th>
<th>Y</th>
<th>Z</th>
<th>R¹</th>
<th>R²</th>
<th>MCF-7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM 2</th>
<th>A549</th>
</tr>
</thead>
<tbody>
<tr>
<td>37 KU-241</td>
<td>H</td>
<td>CH₃</td>
<td>C</td>
<td>H</td>
<td>H</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>3.63</td>
<td>&gt;100</td>
<td></td>
</tr>
<tr>
<td>38 KU-242</td>
<td>H</td>
<td>CH₃</td>
<td>C</td>
<td>CH₃</td>
<td>H</td>
<td>&gt;100</td>
<td>&gt;100</td>
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<td>2</td>
<td>&gt;100</td>
</tr>
<tr>
<td>39 KU-243</td>
<td>H</td>
<td>CH₃</td>
<td>C</td>
<td>CH₃</td>
<td>CH₃</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>7.25</td>
<td>3.87</td>
<td>0.95</td>
</tr>
<tr>
<td>18 KU-260</td>
<td>H</td>
<td>CH₃</td>
<td>N</td>
<td>H</td>
<td>-</td>
<td>1.73</td>
<td>1.61±0.1</td>
<td>4.23</td>
<td>4.36</td>
<td>1.42</td>
</tr>
<tr>
<td>22 KU-250</td>
<td>H</td>
<td>CH₃</td>
<td>N</td>
<td>CH₃</td>
<td>-</td>
<td>5.29±0.3</td>
<td>2.92±1,7</td>
<td>0.82</td>
<td>0.95</td>
<td>5.24</td>
</tr>
<tr>
<td>21 KU-255</td>
<td>H</td>
<td>CH₃</td>
<td>N</td>
<td>COCH₃</td>
<td>-</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>0.41</td>
<td>3.73</td>
<td>0.72</td>
</tr>
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<td>CH₃</td>
<td>C</td>
<td>H</td>
<td>H</td>
<td>TBD</td>
<td>TBD</td>
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<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>OCH₃</td>
<td>CH₃</td>
<td>C</td>
<td>CH₃</td>
<td>H</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>OCH₃</td>
<td>CH₃</td>
<td>C</td>
<td>CH₃</td>
<td>CH₃</td>
<td>TBD</td>
<td>TBD</td>
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<tr>
<td>KU-277</td>
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<td>H</td>
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<td>1.68±0.05</td>
<td>1.21±0.09</td>
<td>5.4</td>
<td>3.57</td>
<td>2.71</td>
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</table>
Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.

Table 9. Biological evaluation of azasugar analogues on 8-methyl coumarin.

<table>
<thead>
<tr>
<th>Compounds (IC₅₀, μM)</th>
<th>X</th>
<th>Y</th>
<th>Z</th>
<th>R¹</th>
<th>R²</th>
<th>MCF-7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM 2</th>
<th>A549</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>KU-280</strong></td>
<td>OCH₃</td>
<td>CH₃</td>
<td>N</td>
<td>CH₃</td>
<td>-</td>
<td>5.27 ± 0.28</td>
<td>1.96 ± 0.60</td>
<td>4.37</td>
<td>3.93</td>
<td>4.02</td>
</tr>
<tr>
<td><strong>KU-283</strong></td>
<td>OCH₃</td>
<td>CH₃</td>
<td>N</td>
<td>COCH₃</td>
<td>-</td>
<td>TBD</td>
<td>TBD</td>
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<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td><strong>H</strong></td>
<td>OCH₃</td>
<td>C</td>
<td>H</td>
<td>H</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td><strong>H</strong></td>
<td>OCH₃</td>
<td>C</td>
<td>CH₃</td>
<td>H</td>
<td>TBD</td>
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<tr>
<td><strong>KU-265</strong></td>
<td>H</td>
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<td>N</td>
<td>H</td>
<td>-</td>
<td>1.43 ± 0.46</td>
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<td>4.54</td>
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<td>OCH₃</td>
<td>N</td>
<td>CH₃</td>
<td>-</td>
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<td>3.42 ± 0.56</td>
<td>0.36</td>
<td>0.98</td>
<td>2.82</td>
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<tr>
<td><strong>KU-269</strong></td>
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<td>OCH₃</td>
<td>N</td>
<td>COCH₃</td>
<td>-</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>TBD</td>
<td>TBD</td>
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</tr>
</tbody>
</table>

*Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.*

![Chemical structure](image)

Table 9. Biological evaluation of azasugar analogues on 8-methyl coumarin.

<table>
<thead>
<tr>
<th>Cpd.</th>
<th>(IC₅₀, μM)</th>
<th>Z</th>
<th>W</th>
<th>R¹</th>
<th>R²</th>
<th>R³</th>
<th>R⁴</th>
<th>R⁵</th>
<th>MCF-7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM2</th>
<th>A549</th>
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<tbody>
<tr>
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<td>CH</td>
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<td>CH</td>
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<td>CO</td>
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<td>H</td>
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<td>C</td>
<td>H₃</td>
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<td>C</td>
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</table>

*Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.*

![Chemical structure](image)

**Table 10. Biological evaluation of dihydroxylated azasugar analogues on 6-methoxy coumarin.**

207
<table>
<thead>
<tr>
<th>Cpd. (IC&lt;sub&gt;50&lt;/sub&gt;, μM)</th>
<th>Z</th>
<th>W</th>
<th>R&lt;sup&gt;1&lt;/sup&gt;</th>
<th>R&lt;sup&gt;2&lt;/sup&gt;</th>
<th>R&lt;sup&gt;3&lt;/sup&gt;</th>
<th>R&lt;sup&gt;4&lt;/sup&gt;</th>
<th>R&lt;sup&gt;5&lt;/sup&gt;</th>
<th>MCF-7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM</th>
<th>A549</th>
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<tr>
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<td>C</td>
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<td>H</td>
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</tr>
<tr>
<td></td>
<td>C</td>
<td>C</td>
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<td>C</td>
<td>H</td>
<td>-</td>
<td>H</td>
<td>H</td>
<td>O H</td>
<td>2.90 ± 0.84</td>
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<td>C</td>
<td>H</td>
<td>-</td>
<td>H</td>
<td>O H</td>
<td>OH</td>
<td>3.34 ± 0.00</td>
<td>3.65 ± 0.28</td>
<td>9.45</td>
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<td>C</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>-</td>
<td>H</td>
<td>H</td>
<td>O H</td>
<td>1.41 ± 0.16</td>
<td>1.49 ± 0.07</td>
<td>6.63</td>
<td>4.47</td>
<td>3.19</td>
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<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
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<td>H</td>
<td>O H</td>
<td>OH</td>
<td>2.01 ± 0.57</td>
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<td>C</td>
<td>CO</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
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<td>H</td>
<td>O H</td>
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<td>KU-284</td>
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<td>C</td>
<td>CO</td>
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<td>&gt;50</td>
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<td>1.72</td>
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<td>C</td>
<td>H</td>
<td>H</td>
<td>1.79 ± 0.11</td>
<td>1.19 ± 0.21</td>
<td>4.07</td>
<td>2.84</td>
<td>1.99</td>
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<td>H</td>
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<td>H</td>
<td>H</td>
<td>2.82 ± 1.38</td>
<td>1.20 ± 0.10</td>
<td>5.27</td>
<td>4.02</td>
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<td>H</td>
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<td>O</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>18.15 ± 5.77</td>
<td>18.46 ± 11.82</td>
<td>7.36</td>
<td>13.7</td>
<td>30.8</td>
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<td>H</td>
<td>H</td>
<td>HC</td>
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<td>H</td>
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<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
</tbody>
</table>
Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.

Table 11. Biological evaluation of dihydroxylated azasugar analogues on 8-methoxy coumarin.

| Cpd. (IC\textsubscript{50}, µM) | Z | W | R\textsuperscript{1} | R\textsuperscript{2} | R\textsuperscript{3} | R\textsuperscript{4} | R\textsuperscript{5} | MCF-7 | SKBr3 | LN3 | MM2 | A549 |
|---|---|---|---|---|---|---|---|---|---|---|---|---|---|
| KU-271 | N | CH | H | - | H | H | H | 3.29 ± 0.52 | 2.61 ± 0.46 | 12.72 | 10.43 | TBD |
| KU-267 | N | CH | H | - | H | OH | OH | 10.17 ± 0.02 | 8.98 ± 0.55 | 12.19 | 14.43 | 16.84 |
| KU-272 | N | CH | CH\textsubscript{3} | - | H | H | H | 1.38 ± 0.17 | 1.40 ± 0.17 | 1.78 | 2.16 | 2.33 |
| KU-268 | N | CH | CH\textsubscript{3} | - | H | OH | OH | 11.02 ± 1.40 | 6.64 ± 0.68 | 1.5 | 3.52 | 8.58 |
| KU-270 | N | CH | COCH\textsubscript{3} | - | H | OH | OH | TBD | TBD | TBD | TBD | TBD |
| KU-273 | N | CH | COCH\textsubscript{3} | - | H | H | H | TBD | TBD | TBD | TBD | TBD |
Values represent mean ± standard deviation for a t least two separate experiments performed in triplicate.

\[ \text{Cpd. (IC}_{50}, \mu \text{M}) \quad Z \quad W \quad R^1 \quad R^2 \quad R^3 \quad R^4 \quad R^5 \quad \text{MCF-7} \quad \text{SKBr3} \quad \text{LN3} \quad \text{MM2} \quad A549 \]

<table>
<thead>
<tr>
<th>Cpd. (IC\text{50}, \mu\text{M})</th>
<th>Z</th>
<th>W</th>
<th>R^1</th>
<th>R^2</th>
<th>R^3</th>
<th>R^4</th>
<th>R^5</th>
<th>MCF-7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM2</th>
<th>A549</th>
</tr>
</thead>
<tbody>
<tr>
<td>KU-275</td>
<td>C</td>
<td>N</td>
<td>H</td>
<td>H</td>
<td>CH₃</td>
<td>H</td>
<td>H</td>
<td>1.39 ± 0.35</td>
<td>0.97 ± 0.09</td>
<td>4.75</td>
<td>3.71</td>
<td>3.06</td>
</tr>
<tr>
<td>KU-274</td>
<td>C</td>
<td>N</td>
<td>H</td>
<td>H</td>
<td>H</td>
<td>H</td>
<td>H</td>
<td>3.23 ± 0.20</td>
<td>1.79 ± 0.00</td>
<td>10.85</td>
<td>8.47</td>
<td>10.69</td>
</tr>
<tr>
<td>KU-276</td>
<td>C</td>
<td>N</td>
<td>H</td>
<td>H</td>
<td>COCH₃</td>
<td>H</td>
<td>H</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
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<tr>
<td></td>
<td>N</td>
<td>CH</td>
<td>H</td>
<td>HCl</td>
<td>H</td>
<td>H</td>
<td>H</td>
<td>TBD</td>
<td>TBD</td>
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<td>TBD</td>
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</tr>
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</table>

\(^a\) Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.

**Table 12. Biological evaluation of aliphatic chain analogues.**

<table>
<thead>
<tr>
<th>Compound (IC\text{50}, \mu\text{M})</th>
<th>X</th>
<th>Y</th>
<th>R</th>
<th>MCF-7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM2</th>
<th>A549</th>
</tr>
</thead>
<tbody>
<tr>
<td>56 KU-256</td>
<td>H</td>
<td>CH₃</td>
<td>[N]</td>
<td>5.69±1.5(^a)</td>
<td>&gt;50</td>
<td>1.59</td>
<td>4.02</td>
<td>TBD</td>
</tr>
<tr>
<td></td>
<td>OCH₃</td>
<td>CH₃</td>
<td>[N]</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td></td>
<td>H</td>
<td>OCH₃</td>
<td>[N]</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>KU-289</td>
<td>H</td>
<td>CH₃</td>
<td>CH₃NHMe</td>
<td>9.80 ± 1.14</td>
<td>5.36 ± 0.12</td>
<td>70.05</td>
<td>13.79</td>
<td>13.05</td>
</tr>
<tr>
<td>KU-290</td>
<td>H</td>
<td>CH₃</td>
<td>CH₃N(Me)₂</td>
<td>1.46 ± 0.12</td>
<td>1.02 ± 0.16</td>
<td>6.65</td>
<td>4.17</td>
<td>2.66</td>
</tr>
</tbody>
</table>

210
<p>| | | | | | | | |</p>
<table>
<thead>
<tr>
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</tr>
</thead>
<tbody>
<tr>
<td>KU-316</td>
<td>H</td>
<td>CH₃</td>
<td>CH₂CH₂NHMe</td>
<td>TBD</td>
<td>TBD</td>
<td>60.96</td>
<td>76.67</td>
</tr>
<tr>
<td>KU-317</td>
<td>H</td>
<td>CH₃</td>
<td>CH₂CH₂N(Me)₂</td>
<td>0.50 ± 0.04</td>
<td>0.60 ± 0.01</td>
<td>37.51</td>
<td>12.85</td>
</tr>
<tr>
<td>KU-293</td>
<td>OCH₃</td>
<td>CH₃</td>
<td>CH₂NHMe</td>
<td>6.23 ± 0.17</td>
<td>3.15 ± 0.67</td>
<td>16.31</td>
<td>11.77</td>
</tr>
<tr>
<td>KU-294</td>
<td>OCH₃</td>
<td>CH₃</td>
<td>CH₂N(Me)₂</td>
<td>1.29 ± 0.21</td>
<td>1.42 ± 0.26</td>
<td>4.71</td>
<td>4.87</td>
</tr>
<tr>
<td>KU-320</td>
<td>OCH₃</td>
<td>CH₃</td>
<td>CH₂CH₂NHMe</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>KU-321</td>
<td>OCH₃</td>
<td>CH₃</td>
<td>CH₂CH₂N(Me)₂</td>
<td>0.70 ± 0.19</td>
<td>0.49 ± 0.25</td>
<td>4.56</td>
<td>5.4</td>
</tr>
<tr>
<td>KU-291</td>
<td>H</td>
<td>OCH₃</td>
<td>CH₂NHMe</td>
<td>4.32 ± 0.40</td>
<td>3.60 ± 0.35</td>
<td>15.24</td>
<td>9.12</td>
</tr>
<tr>
<td>KU-292</td>
<td>H</td>
<td>OCH₃</td>
<td>CH₂N(Me)₂</td>
<td>3.08 ± 0.12</td>
<td>1.77 ± 0.15</td>
<td>5.22</td>
<td>6.2</td>
</tr>
<tr>
<td>KU-318</td>
<td>H</td>
<td>OCH₃</td>
<td>CH₂CH₂NHMe</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>KU-319</td>
<td>H</td>
<td>OCH₃</td>
<td>CH₂CH₂N(Me)₂</td>
<td>1.53 ± 0.18</td>
<td>0.91 ± 0.17</td>
<td>8.03</td>
<td>4.08</td>
</tr>
<tr>
<td>KU-322</td>
<td>H</td>
<td>CH₃</td>
<td>CHOHCH₂OH</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>&gt;100</td>
<td>0.19</td>
</tr>
<tr>
<td>KU-324</td>
<td>OCH₃</td>
<td>CH₃</td>
<td>CHOHCH₂OH</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>KU-323</td>
<td>H</td>
<td>OCH₃</td>
<td>CHOHCH₂OH</td>
<td>&gt;50</td>
<td>&gt;50</td>
<td>0.96</td>
<td>1.66</td>
</tr>
</tbody>
</table>

*a* Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.

As shown in the table, the six-membered sugar mimics (lla,b, 13b and 17) were found to be the most active compounds of the series. The compound lib manifested an IC₅₀ of 0.2 µM, making it 3500 times more active than novobiocin. The 6-membered sugar mimics with a 3'-hydroxy (13a and 13b) were not as active as the six-membered analogues with a 4'-hydroxy group. The 3'-hydroxy group appears to be essential for anti-proliferative activity.
Table 13. Biological evaluation of various sugars on 8-methyl coumarin. a

<table>
<thead>
<tr>
<th>Compound (IC&lt;sub&gt;50&lt;/sub&gt;, μM)</th>
<th>MCF7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM2</th>
<th>A549</th>
</tr>
</thead>
<tbody>
<tr>
<td>11a</td>
<td>2.56±0.1</td>
<td>6.23±0.6</td>
<td>3.59</td>
<td>11.78</td>
<td>2.27</td>
</tr>
<tr>
<td>11b</td>
<td>42.6±3.1</td>
<td>6.71±0.1</td>
<td>1.20</td>
<td>12.88</td>
<td>19.76</td>
</tr>
<tr>
<td>19</td>
<td>37.17±2.1</td>
<td>12.46±0.7</td>
<td>3.65</td>
<td>30.59</td>
<td>&gt;100</td>
</tr>
<tr>
<td>21a</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>5.34</td>
<td>13.69</td>
<td>21.88</td>
</tr>
<tr>
<td>21b</td>
<td>TBD</td>
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<td>15a</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>3.11</td>
<td>&gt;100</td>
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<tr>
<td>15b</td>
<td>11.20±0.6</td>
<td>9.28±0.1</td>
<td>8.56</td>
<td>0.39</td>
<td>TBD</td>
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<td>13a</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>3.75</td>
<td>10.06</td>
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<tr>
<td>13b</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>7.34</td>
<td>0.13</td>
<td>0.88</td>
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<tr>
<td>23a</td>
<td>&gt;100</td>
<td>35.24±2.2</td>
<td>3.72</td>
<td>10.96</td>
<td>0.99</td>
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<tr>
<td>23b</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>4.96</td>
<td>26.79</td>
<td>1.36</td>
</tr>
<tr>
<td>25a</td>
<td>&gt;100</td>
<td>2.38±0.1</td>
<td>26.55</td>
<td>&gt;100</td>
<td>0.18</td>
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<td>25b</td>
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</table>
Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.

Table 14. Biological evaluation of various sugars on 6-methoxy coumarin.

<table>
<thead>
<tr>
<th>Cpd. (IC_{50}, \mu M)</th>
<th>MCF7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM2</th>
<th>A549</th>
</tr>
</thead>
<tbody>
<tr>
<td>KU-343</td>
<td>9.498 ± 0.20</td>
<td>7.179 ± 0.25</td>
<td>12.76</td>
<td>24.03</td>
<td>3.34</td>
</tr>
<tr>
<td>KU-344</td>
<td>TBD</td>
<td>TBD</td>
<td>14.93</td>
<td>25.37</td>
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<tr>
<td>KU-345</td>
<td>&gt;100</td>
<td>&gt;100</td>
<td>1.49</td>
<td>1.234</td>
<td>1.9</td>
</tr>
<tr>
<td>KU-346</td>
<td>&gt;100</td>
<td>1.373 ± 0.18</td>
<td>3.33</td>
<td>2.35</td>
<td>3.64</td>
</tr>
</tbody>
</table>
Table 15. Biological evaluation of various sugars on 8-methoxy coumarin.

<table>
<thead>
<tr>
<th>Cpd. (IC50, μM)</th>
<th>MCF7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM2</th>
<th>A549</th>
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</tr>
<tr>
<td>KU-347</td>
<td>&gt;100</td>
<td>TBD</td>
<td>4.662</td>
<td>5.9</td>
<td>13.4</td>
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</tbody>
</table>

Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.
<table>
<thead>
<tr>
<th>Compound (IC50, μM)</th>
<th>MCF7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM2</th>
<th>A549</th>
</tr>
</thead>
<tbody>
<tr>
<td>KU-340</td>
<td>11.73 ± 0.98</td>
<td>7.57 ± 1.31</td>
<td>2.595</td>
<td>10.64</td>
<td>7.34</td>
</tr>
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<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>KU-337</td>
<td>95.77 ± 4.01</td>
<td>8.75 ± 0.61</td>
<td>11.5</td>
<td>TBD</td>
<td>TBD</td>
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<tr>
<td>KU-338</td>
<td>&gt;100</td>
<td>8.03 ± 0.16</td>
<td>19.48</td>
<td>TBD</td>
<td>47.85</td>
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<tr>
<td>KU-338</td>
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<td>8.03 ± 0.16</td>
<td>19.48</td>
<td>TBD</td>
<td>47.85</td>
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<tr>
<td>KU-337</td>
<td>95.77 ± 4.01</td>
<td>8.75 ± 0.61</td>
<td>11.5</td>
<td>TBD</td>
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</tr>
<tr>
<td>KU-339</td>
<td>14.34 ± 0.2</td>
<td>11.76 ± 0.08</td>
<td>6.782</td>
<td>17.94</td>
<td>13.75</td>
</tr>
</tbody>
</table>

αValues represent mean ± standard deviation for at least two separate experiments performed in triplicate.
Table 16. Anti-proliferation activities of non-sugar analogues.

<table>
<thead>
<tr>
<th>Compound (IC&lt;sub&gt;50&lt;/sub&gt;, µM)</th>
<th>X</th>
<th>Y</th>
<th>R</th>
<th>MCF-7</th>
<th>SKBr3</th>
<th>LN3</th>
<th>MM2</th>
<th>A549</th>
</tr>
</thead>
<tbody>
<tr>
<td>1a, KU-295</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>H</td>
<td>6.93 ± 0.6&lt;sup&gt;a&lt;/sup&gt;</td>
<td>8.88 ± 0.6</td>
<td>3.21</td>
<td>5.36</td>
<td>2.22</td>
</tr>
<tr>
<td>50a</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>COCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>1.4</td>
<td>0.98 ± 0.02</td>
<td>1.50</td>
<td>2.85</td>
<td>46.42</td>
</tr>
<tr>
<td>JB33</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>Noviose</td>
<td>18.7 ± 1.8</td>
<td>7.5 ± 1.0</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>KU-296</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>Ms</td>
<td>8.85 ± 1.1</td>
<td>7.33 ± 1.2</td>
<td>19.96</td>
<td>87.39</td>
<td>54.41</td>
</tr>
<tr>
<td>KU-297</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>Ts</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>0.58</td>
<td>0.09</td>
<td>33.46</td>
</tr>
<tr>
<td>KU-298</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CONH&lt;sub&gt;2&lt;/sub&gt;</td>
<td>1.16 ± 0.1</td>
<td>3.02 ± 0.7</td>
<td>2.61</td>
<td>6.37</td>
<td>2.36</td>
</tr>
<tr>
<td>KU-299</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CONMe</td>
<td>1.72 ± 0.2</td>
<td>2.40 ± 0.2</td>
<td>3.75</td>
<td>5.22</td>
<td>2.13</td>
</tr>
<tr>
<td>KU-300</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CONMe&lt;sub&gt;2&lt;/sub&gt;</td>
<td>74.35 ± 4.9</td>
<td>39.85 ± 0.6</td>
<td>5.40</td>
<td>42.23</td>
<td>0.42</td>
</tr>
<tr>
<td>KU-301</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>PO(OMe)&lt;sub&gt;2&lt;/sub&gt;</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>3.65</td>
<td>&gt; 100</td>
<td>4.57</td>
</tr>
<tr>
<td>1b, KU-179</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>H</td>
<td>11.7 ± 2.4&lt;sup&gt;a&lt;/sup&gt;</td>
<td>3.23 ± 1.5</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>50b, KU-135</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>COCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>1.5 ± 0.3</td>
<td>5.72 ± 0.03</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>51</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>Noviose</td>
<td>&gt; 100</td>
<td>58.8 ± 1.3</td>
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<td>TBD</td>
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<td>52, KU-302</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>Ms</td>
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<td>&gt; 100</td>
<td>28.52</td>
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<td>53, KU-303</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>Ts</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>&gt; 100</td>
<td>4.03</td>
<td>84.37</td>
</tr>
<tr>
<td>54, KU-304</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CONH&lt;sub&gt;2&lt;/sub&gt;</td>
<td>6.29 ± 1.1</td>
<td>6.83 ± 0.3</td>
<td>3.55</td>
<td>2.62</td>
<td>0.96</td>
</tr>
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<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CONMe</td>
<td>7.78 ± 2.1</td>
<td>1.53 ± 0.2</td>
<td>4.77</td>
<td>3.13</td>
<td>1.33</td>
</tr>
<tr>
<td>56, KU-178</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CONMe&lt;sub&gt;2&lt;/sub&gt;</td>
<td>34.4 ± 9.0</td>
<td>6.17 ± 2.1</td>
<td>12.53</td>
<td>4.04</td>
<td>TBD</td>
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<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>PO(OMe)&lt;sub&gt;2&lt;/sub&gt;</td>
<td>24.22 ± 3.0</td>
<td>26.48 ± 1.0</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
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<tr>
<td>Compound (IC$_{50}$, $\mu$M)</td>
<td>X</td>
<td>Y</td>
<td>R</td>
<td>MCF-7</td>
<td>SkBr3</td>
<td>LN3</td>
<td>MM2</td>
<td>A549</td>
</tr>
<tr>
<td>----------------------------</td>
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<td>--------</td>
<td>-------</td>
<td>------</td>
<td>------</td>
<td>------</td>
</tr>
<tr>
<td>1c, KU-308</td>
<td>H</td>
<td>OCH$_3$</td>
<td>H</td>
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<td>48.44</td>
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<tr>
<td>50c, KU-177</td>
<td>H</td>
<td>OCH$_3$</td>
<td>COCH$_3$</td>
<td>8.62 ± 2.5</td>
<td>20.5 ± 1.6</td>
<td>14.62</td>
<td>19.33</td>
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</tr>
<tr>
<td></td>
<td>H</td>
<td>OCH$_3$</td>
<td>Noviose</td>
<td>9.0 ± 5.4</td>
<td>13.9 ± 1.2</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
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<tr>
<td>KU-309</td>
<td>H</td>
<td>OCH$_3$</td>
<td>Ms</td>
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<td>&gt;100</td>
<td>0.87</td>
<td>1.14</td>
<td>0.79</td>
</tr>
<tr>
<td>KU-310</td>
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<td>OCH$_3$</td>
<td>Ts</td>
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<td>&gt;100</td>
<td>0.11</td>
<td>&gt; 100</td>
<td>26.09</td>
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<tr>
<td>KU-311</td>
<td>H</td>
<td>OCH$_3$</td>
<td>CONH$_2$</td>
<td>5.56 ± 0.3</td>
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<td>4.83</td>
<td>29.35</td>
<td>27.14</td>
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<tr>
<td>KU-312</td>
<td>H</td>
<td>OCH$_3$</td>
<td>CONHMe</td>
<td>6.54 ± 2.3</td>
<td>&gt;100</td>
<td>2.37</td>
<td>1.68</td>
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<td>KU-313</td>
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<td>CONMe$_2$</td>
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<td>&gt;100</td>
<td>15.73</td>
<td>11.57</td>
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<td>KU-314</td>
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<td>OCH$_3$</td>
<td>PO(OMe)$_2$</td>
<td>&gt;100</td>
<td>18.07 ± 3.6</td>
<td>6.58</td>
<td>11.46</td>
<td>17.21</td>
</tr>
</tbody>
</table>

$^a$ Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.

![Chemical structure](image)

Table 17. Anti-proliferation activities of modified scaffolds

<table>
<thead>
<tr>
<th>Compound (IC$_{50}$, $\mu$M)</th>
<th>X</th>
<th>Y</th>
<th>R</th>
<th>MCF-7</th>
<th>SkBr3</th>
<th>PC-3</th>
<th>LnCaP</th>
</tr>
</thead>
<tbody>
<tr>
<td>H</td>
<td>CH$_3$</td>
<td>COOH</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>CH$_3$</td>
<td>CH$_2$OH</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
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<tr>
<td>H</td>
<td>CH$_3$</td>
<td>COOCH$_3$</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>CH$_3$</td>
<td>NH$_2$</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>CH$_3$</td>
<td>NHCOCH$_3$</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>CH$_3$</td>
<td>4-pyridine</td>
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<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
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<td>45</td>
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<td>CH$_3$</td>
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<td>TBD</td>
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<td>CH$_2$OH</td>
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<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>44</td>
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<td>CH$_3$</td>
<td>COOCH$_3$</td>
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<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>46</td>
<td>OCH$_3$</td>
<td>CH$_3$</td>
<td>NH$_2$</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
</tbody>
</table>
Table 18. Anti-proliferation activities of novobiocin analogues that contain modified furanose or pyranose noviose replacements

<table>
<thead>
<tr>
<th>Compound (IC₅₀, μM)</th>
<th>X</th>
<th>Y</th>
<th>R</th>
<th>MCF-7</th>
<th>SkBr3</th>
<th>PC-3</th>
<th>LnCaP</th>
</tr>
</thead>
<tbody>
<tr>
<td>48</td>
<td>OCH₃</td>
<td>CH₃</td>
<td>NHCOCH₃</td>
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<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>49</td>
<td>OCH₃</td>
<td>CH₃</td>
<td>4-pyridine</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>OCH₃</td>
<td>COOH</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>OCH₃</td>
<td>CH₂OH</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>OCH₃</td>
<td>COOCH₃</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>OCH₃</td>
<td>NH₂</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
<tr>
<td>H</td>
<td>OCH₃</td>
<td>NHCOCH₃</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
<td>TBD</td>
</tr>
</tbody>
</table>

Values represent mean ± standard deviation for at least two separate experiments performed in triplicate.

Table 18. Anti-proliferation activities of novobiocin analogues that contain modified furanose or pyranose noviose replacements
<table>
<thead>
<tr>
<th>Compound (IC50, µM)</th>
<th>SKBr3</th>
<th>MCF7</th>
</tr>
</thead>
<tbody>
<tr>
<td>18a, KU-429</td>
<td>5.07±0.23</td>
<td>1.34±0.18</td>
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<tr>
<td>18b, KU-430</td>
<td>3.11±0.19</td>
<td>1.56±0.09</td>
</tr>
<tr>
<td>17a, KU-427</td>
<td>7.87±0.04</td>
<td>6.45±0.13</td>
</tr>
<tr>
<td>17b, KU-428</td>
<td>29.98±2.07</td>
<td>10.24±0.21</td>
</tr>
<tr>
<td>20a, KU-432</td>
<td>22.16±0.94</td>
<td>&gt;100</td>
</tr>
<tr>
<td>20b, KU-433</td>
<td>21.46±2.28</td>
<td>22.50±0.40</td>
</tr>
<tr>
<td>DHN2</td>
<td>10.86±0.47</td>
<td>11.29±0.41</td>
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</tbody>
</table>

![Diagram of DHN2 molecule]
Table 19. Biological evaluation of 7-oxyalkylamino and 7-oxypiperidinyl analogues of novobiocin.

<table>
<thead>
<tr>
<th>Compound (IC$_{50}$, μM)</th>
<th>X</th>
<th>Y</th>
<th>R$^{1}$</th>
<th>R$^{2}$</th>
<th>MCF-7</th>
<th>SKBr3</th>
</tr>
</thead>
<tbody>
<tr>
<td>29a, KU-397</td>
<td>H</td>
<td>CH$_3$</td>
<td>COCH$_3$</td>
<td><img src="image1" alt="Structure" /></td>
<td>0.58±0.0</td>
<td>1.18±0.20</td>
</tr>
<tr>
<td>29c, KU-417</td>
<td>H</td>
<td>CH$_3$</td>
<td>COCH$_3$</td>
<td><img src="image2" alt="Structure" /></td>
<td>1.42±0.0</td>
<td>1.57±0.05</td>
</tr>
<tr>
<td>29e, KU-421</td>
<td>H</td>
<td>CH$_3$</td>
<td>COCH$_3$</td>
<td><img src="image3" alt="Structure" /></td>
<td>1.32±0.0</td>
<td>4.76±0.52</td>
</tr>
<tr>
<td>29f, KU-406</td>
<td>H</td>
<td>CH$_3$</td>
<td>COCH$_3$</td>
<td><img src="image4" alt="Structure" /></td>
<td>0.46±0.1</td>
<td>1.18±0.03</td>
</tr>
<tr>
<td>30b, KU-415</td>
<td>H</td>
<td>CH$_3$</td>
<td>COCH$_3$</td>
<td><img src="image5" alt="Structure" /></td>
<td>0.77±0.2</td>
<td>1.53±0.14</td>
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<tr>
<td>30d, KU-419</td>
<td>H</td>
<td>CH$_3$</td>
<td>COCH$_3$</td>
<td><img src="image6" alt="Structure" /></td>
<td>1.23±0.0</td>
<td>1.54±0.11</td>
</tr>
<tr>
<td>30g, KU-423</td>
<td>H</td>
<td>CH$_3$</td>
<td>COCH$_3$</td>
<td><img src="image7" alt="Structure" /></td>
<td>0.91±0.2</td>
<td>2.08±0.13</td>
</tr>
<tr>
<td>31a, KU-398</td>
<td>H</td>
<td>CH$_3$</td>
<td>H</td>
<td><img src="image8" alt="Structure" /></td>
<td>0.76±0.1</td>
<td>1.09±0.10</td>
</tr>
<tr>
<td>31b, KU-416</td>
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<td>CH$_3$</td>
<td>H</td>
<td><img src="image9" alt="Structure" /></td>
<td>0.47±0.1</td>
<td>0.85±0.09</td>
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<td><img src="image10" alt="Structure" /></td>
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<td>H</td>
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<td>-----</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>31e, KU-422</td>
<td>H</td>
<td>CH₃</td>
<td>H</td>
<td>H₂C-(\text{N}^-)(\text{CH}_3)</td>
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<td>52.7</td>
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<td>31f, KU-407</td>
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<td>CH₃</td>
<td>H</td>
<td>H₂C-(\text{N}^-)(\text{CH}_3)</td>
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<td>1.35±0.38</td>
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<td>CH₃</td>
<td>H</td>
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<td>CH₃</td>
<td>(\text{H}_3\text{C}-\text{N}^-\text{CH}_3)</td>
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<tr>
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<td>(\text{H}_3\text{C}-\text{N}^-\text{CH}_3)</td>
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<td>TBD</td>
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<tr>
<td>KU-455</td>
<td>H</td>
<td>CH₃</td>
<td>(\text{H}_3\text{C}-\text{N}^-\text{CH}_3)</td>
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<tr>
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<td>(\text{H}_3\text{C}-\text{N}^-\text{CH}_3)</td>
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<tr>
<td>KU-457</td>
<td>H</td>
<td>CH₃</td>
<td>(\text{H}_3\text{C}-\text{N}^-\text{CH}_3)</td>
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<tr>
<td>KU-458</td>
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<td>CH₃</td>
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<td>OCH₃</td>
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<td>TBD</td>
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Table 20. Anti-proliferation activities of indole-2-carboxamide novobiocin analogues that contain noviose replacements.

<table>
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<tr>
<th>Compound (IC₅₀, µM)</th>
<th>X</th>
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<th>R</th>
<th>SKBr3</th>
<th>MCF-7</th>
<th>MM2</th>
<th>LN3</th>
<th>A549</th>
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<td>± 1.60</td>
<td>± 0.08</td>
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<tr>
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<td>CH₃</td>
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<td>CH₃</td>
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<td>TBD</td>
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<td>Compound (IC$_{50}$, μM)</td>
<td>X</td>
<td>Y</td>
<td>R</td>
<td>SKBr3</td>
<td>MCF-7</td>
<td>MM2</td>
<td>LN3</td>
<td>A549</td>
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<td>H</td>
<td>CH$_3$</td>
<td>[Chemical Structure]</td>
<td>0.48 ± 0.11</td>
<td>0.57 ± 0.04</td>
<td>11.40 ± 6.56</td>
<td>11.83 ± 0.68</td>
<td>3.91</td>
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<td>KU-362</td>
<td>H</td>
<td>OCH$_3$</td>
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<td>[Chemical Structure]</td>
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<td>0.87 ± 0.57</td>
<td>1.47 ± 0.66</td>
<td>0.59</td>
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<td>Not soluble</td>
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<tr>
<td>Compound (IC&lt;sub&gt;50&lt;/sub&gt;, μM)</td>
<td>X</td>
<td>Y</td>
<td>R</td>
<td>SKBr3</td>
<td>MCF-7</td>
<td>MM2</td>
<td>LN3</td>
<td>A549</td>
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<td>1.13 ±</td>
<td>5.23</td>
<td>13.59</td>
<td>18.35</td>
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<td>KU-371</td>
<td>H</td>
<td>OCH₃</td>
<td></td>
<td>1.50 ±</td>
<td>1.41</td>
<td>8.95</td>
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<td>7.06</td>
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<td>0.56</td>
<td>2.58</td>
<td>5.06</td>
<td>2.17</td>
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Table 21. Anti-proliferation activities of substituted indole-2-carboxamide novobiocin analogues that contain noviose replacements.

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<th>Compound (IC&lt;sub&gt;50&lt;/sub&gt;, µM)</th>
<th>X</th>
<th>Y</th>
<th>R</th>
<th>R&lt;sup&gt;31&lt;/sup&gt;</th>
<th>R&lt;sup&gt;32&lt;/sup&gt;</th>
<th>SKBr3</th>
<th>MCF-7</th>
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<tbody>
<tr>
<td>KU-379</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>H</td>
<td>0.82±0.21</td>
<td>1.59±0.04</td>
<td></td>
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<tr>
<td>KU-380</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td></td>
<td>Cl</td>
<td>H</td>
<td>0.35±0.21</td>
<td>0.09±0.03</td>
</tr>
<tr>
<td>KU-381</td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td></td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>1.04±0.28</td>
<td>0.09±0.03</td>
</tr>
<tr>
<td>KU-382</td>
<td>H</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>H</td>
<td>0.69±0.44</td>
<td>1.22±0.16</td>
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</tr>
<tr>
<td>KU-383</td>
<td>H</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
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<td>Cl</td>
<td>H</td>
<td>1.04±0.55</td>
<td>2.19±0.81</td>
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<td>KU-384</td>
<td>H</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td></td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>TBD</td>
<td>TBD</td>
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<tr>
<td>KU-385</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>H</td>
<td>11.40±0.70</td>
<td>67.68±2.63</td>
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<tr>
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<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td></td>
<td>Cl</td>
<td>H</td>
<td>0.51±0.11</td>
<td>4.95±0.45</td>
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<tr>
<td>KU-387</td>
<td>OCH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td></td>
<td>H</td>
<td>CH&lt;sub&gt;3&lt;/sub&gt;</td>
<td>TBD</td>
<td>TBD</td>
</tr>
</tbody>
</table>
Throughout this application, various publications are referenced. The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this pertains. The references disclosed are also individually and specifically incorporated by reference herein for the material contained in them that is discussed in the sentence in which the reference is relied upon.

(1) Powers, M. V., Workman, P. In Endocrine-Related Cancer 2006; Vol. 13, p S125-S135.


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[0520] From the foregoing it will be seen that this disclosure is one well adapted to attain all ends and objectives herein-above set forth, together with the other advantages which are obvious and which are inherent to the disclosure. Since many possible embodiments may be made of the disclosure without departing from the scope thereof, it is to be understood that all matters herein set forth or shown in the accompanying drawings are to be interpreted as illustrative, and not in a limiting sense. While specific embodiments have been shown and discussed, various modifications may of course be made, and the disclosure is not limited to the specific forms or arrangement of parts and steps described herein, except insofar as such limitations are included in the following claims. Further, it will be understood that certain features and subcombinations are of utility and may be employed without reference to other features and subcombinations. This is contemplated by and is within the scope of the claims.
CLAMS

What is claimed is:

1. A compound according to Formula I:

   ![Chemical Structure](image)

   wherein:

   $R^1$ is $-\text{NHCOR}^\prime$, where $R^\prime$ is a C1-C4 alkyl, aryl or heterocyclic group, each optionally substituted with one or more hydroxy, nitro, amino, alkyl, alkenyl, aryl, alkoxy or halo groups;

   $X^9$ is $-\text{O-alkyl}$, $-\text{O-alkylamino}$, $-\text{O-cycloalkyl}$, $-\text{O-(CO)-alkyl}$, $-\text{O-(CO)-cycloalkyl}$, $-\text{O-(CH}_2\text{)_n-pyridinyl}$, $-\text{O-(CH}_2\text{)_n-piperidinyl}$, $-\text{O-(CH}_2\text{)_n-pyrrolino}$, or $-\text{O-(CH}_2\text{)_n-pyrrolidinyl}$, each substituted with one or more amino, amido, alkyl, alkoxy, halo, pyrrolidinyl, or hydroxyl groups; and where $n$ is 0, 1 2 or 3; or $-\text{O-mono-hydroxylated furanose}$, $-\text{O-dihydroxylated furanose}$, $-\text{O-mono-hydroxylated pyranose}$, $-\text{O-dihydroxylated pyranose}$, $-\text{O-trihydroxylated pyranose}$, $-\text{O-mono-hydroxylated oxepinose}$, $-\text{O-dihydroxylated oxepinose}$, $-\text{O-azasugar}$, $-\text{O-acyl}$, ester, amino, amido, carbamate, phosphate ester, tosylate or mesylate; or $X^9$ is $-\text{OH}$ provided that $R^\prime$ is not of the structure:

   ![Chemical Structure](image)

   $X$ is H, nitrile, halo, amino, amido, C1-C4 alkyl or alkoxy; and

   $Y$ is H, amido, ester, amino, C1-C4 alkyl or alkoxy; or

   a pharmaceutically acceptable salt thereof.

2. The compound of claim 1 wherein $X_0$ is $-\text{O-alkyl}$, $-\text{O-alkylamino}$, $-\text{O-cycloalkyl}$, $-\text{O-cyclohexyl}$, $-\text{O-cyclopentyl}$, $-\text{O-cyclopropyl}$, $-\text{O-alkoxy}$, $-\text{O-halo}$, $-\text{O-amidino}$, ester, amino, amido, carbamate, phosphate ester, tosylate or mesylate; or $X_0$ is $-\text{OH}$ provided that $R^\prime$ is not of the structure:

   ![Chemical Structure](image)

   $X$ is H, nitrile, halo, amino, amido, C1-C4 alkyl or alkoxy; and

   $Y$ is H, amido, ester, amino, C1-C4 alkyl or alkoxy; or

   a pharmaceutically acceptable salt thereof.
-O-(CO)-alkyl, -O-(CO)-cycloalkyl, -O-(CH₂)ₙ-pyridinyl, -O-(CH₂)ₙ-piperidinyl, -O-(CH₂)ₙ-pyrrolidinyl, or -O-(CH₂)ₙ-pyrrolinyl, each optionally substituted with one or more amino, amido alkyl, halo, alkoxy, or hydroxyl groups.

3. The compound of claim 1 wherein X₀ is

![Structures](image1)

4. The compounds of claim 1 wherein R¹ is -NHCOCH₃.

5. The compound of claim 1 wherein Rᵣ is an aryl group selected from:

![Structures](image2)

wherein R²⁴ and R²⁵ are independently H, C₁-C₄ alkyl, hydroxy or alkoxy; and R³³ is H, C₁-C₄ alkyl, C₁-C₄ alkylamino, -(CO)-C₁-C₄ alkyl, or piperidinyl, each optionally substituted with C₁-C₄ alkyl; or Rᵣ is a heterocyclic group:

![Structures](image3)
wherein $R^{31}$ is H, halo, $C_1$-$C_4$ alkyl, hydroxy or alkoxy; and $R^{32}$ is H or $C_1$-$C_4$ alkyl.

6. The compound of claim 5 wherein $R''$ is an aryl group:

7. The compound of claim 6 in which the compound is selected from the group consisting of:

- KU-131,
- KU-133,
- KU-135, and
- KU-139.
8. The compound of claim 5 wherein R" is aryl according to:

$$\text{OR}^3$$

wherein $\text{R}^3$ is H, -CH$_3$, -COCH$_3$, -CH$_2$CH$_2$N(CH$_3$)$_2$, -CH$_2$CH$_2$CH$_2$N(CH$_3$)$_2$, or

9. The compound of claim 8 selected from the group consisting of:

- 4-(8-Methyl-7-(l-methylpiperidin-4-yloxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29a, KU-397);
- 4-(8-Methyl-7-(l-methylpiperidin-3-yloxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29c, KU-417);
- 4-(7-(2-(Dimethylamino)ethoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29e, KU-421);
- 4-(7-(3-(Dimethylamino)propoxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (29f, KU-406);
- 4-(8-methyl-2-oxo-7-(piperidin-4-yloxy)-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (30b, KU-415);
- 4-(8-methyl-2-oxo-7-(piperidin-3-yloxy)-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (30d, KU-419);
- 4-(8-methyl-7-(2-(dimethylamino)ethoxy)-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (30g, KU-423);
- 4-Hydroxy-N-(8-methyl-7-(l-methylpiperidin-4-yloxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31a, KU-398);
- 4-Hydroxy-N-(8-methyl-2-oxo-7-(piperidin-4-yloxy)-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31b, KU-416);
- 4-Hydroxy-N-(8-methyl-7-(l-methylpiperidin-3-yloxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31c, KU-418);
- 4-Hydroxy-N-(8-methyl-2-oxo-7-(piperidin-3-yloxy)-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (31d, KU-420);
N-(7-(2-(Dimethylamino)ethoxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (3Ie, KU-422);

N-(7-(3-(Dimethylamino)propoxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (3If, KU-407);

4-Hydroxy-N-(8-methyl-7-(2-(methylamino)ethoxy)-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (3Ig, KU-424);

N-(7-((2R,3R,4R)-3,4-dihydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (16a, KU-425);

N-(7-((2S,3R,4R)-3,4-dihydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-4-hydroxy-3-(3-methylbut-2-enyl)benzamide (16b, KU-426);

4-Hydroxy-N-(7-((2R,3R)-3-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (17a, KU-247);

4-Hydroxy-N-(7-((2S,3R)-3-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (17b, KU-428);

4-Hydroxy-N-(7-((2R,4R)-4-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (18a, KU-429);

4-Hydroxy-N-(7-((2R,4R)-4-hydroxytetrahydro-2H-pyran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-yl)-3-(3-methylbut-2-enyl)benzamide (18b, KU-430);

4-(7-((2S,3S,4S)-3,4-Dihydroxytetrahydrofuran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (19, KU-431);

4-(7-((2S,4R)-4-Hydroxytetrahydrofuran-2-yloxy)-8-methyl-2-oxo-2H-chromen-3-ylcarbamoyl)-2-(3-methylbut-2-enyl)phenyl acetate (20a, KU-432); and

10. The compound of claim 8:

![Chemical structure of KU-456](image)

![Chemical structure of KU-457](image)

![Chemical structure of KU-458](image)

![Chemical structure of KU-459](image)

11. The compound of claim 5 wherein $R^1$ is a heterocyclic group according to:

![Chemical structure](image)

wherein $R^{31}$ is H, halo, or alkoxy; and $R^{32}$ is H or alkyl.

12. The compound of claim 11:

![Chemical structure](image)

wherein
X is H or -OCH₃;
Y is -CH₃ or -OCH₃;

R₃¹ is H, Cl, -CH₂CH₂N(CH₃)₂, -CH₂CH₂CH₂N(CH₃)₂, -OCH₃, or H₃C⁻N⁺ (R₁ = H, CH₃, or CH₂CH₂CH₂N(CH₃)₂, and H₂C⁻N⁺ (R₃₂ = H or CH₃; and

R is selected from the group consisting of:

13. The compound of claim 12:
14. The compound of claim 6:

wherein

X is H or -OCH₃;

Y is -CH₃ or -OCH₃; and

R is selected from the group consisting of: H, -COCH₃, mesylate, tosylate, -CONH₂,

-CONHCH₃, -CON(CH₃)₂, -PO(OCH₃)₂, -COCH₃,
and
15. The compound of claim 14:

16. A pharmaceutical composition for the treatment of cancer in a patient in need thereof, the composition comprising a therapeutically effective amount of a compound of claim 1, and a pharmaceutically acceptable carrier.

17. A method of treating cancer in a patient in need thereof, the method comprising administering a therapeutically effective amount of a compound of claim 1.
### FIG. 1

![Bar Chart](image)

### FIG. 2

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<td>1 µM</td>
<td>10 µM</td>
</tr>
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<td>.1 µM</td>
<td>1 µM</td>
<td>10 µM</td>
</tr>
<tr>
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</table>

**FIG. 3**
FIG. 13
FIG. 14
FIG. 15

removal of 4-hydroxy

amide is critical for activity

polar substituents increase activity. Phenols are most active

H-bond acceptor

FIG. 16

rigid 2 carbon linker increases activity

removal of 4-hydroxy

H-bond donor increases activity
FIG. 17

<table>
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INTERNATIONAL SEARCH REPORT

International application No
PCT/US 10/24729

A CLASSIFICATION OF SUBJECT MATTER
IPC(8) - C07H 17/06 (201 0 0 1)
USPC - 536/13
According to International Patent Classification (IPC) or to both national classification and IPC

B FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
USPC 536/13

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
USPC 514/43, 250, 293, 536/271, 1, 544/65, 251, 549/230, 285, 288 (see search terms below)

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
PubWEST(USPT,PGPB,EPAB,JPAB), GoogleScholar, Freepatentsonline (structure search, species search, novobiocin substructure, coumarin substructure)
Search Hsp90, Novobiocin, coumermycin, coumarin, analogs, derivatives, cancer, benzamide

C DOCUMENTS CONSIDERED TO BE RELEVANT

<table>
<thead>
<tr>
<th>Category*</th>
<th>Citation of document, with indication, where appropriate, of the relevant passages</th>
<th>Relevant to claim No</th>
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<td>X---------</td>
<td>Lubbers Synthetic and computational efforts toward the understanding and development of novobiocin-de fixed inhibitors of Hsp90 B S , Saint Mary's College, Notre Dame, 2005, Title, Abstract, pg 7-8, pg 26, Fig 16, pg 30, Scheme 4, pg 31, Fig 17, pg 32, Fig 18 pg 30-31, pg 33, Scheme 5, pg 38, Scheme 8</td>
<td>1-3, 5-6, 8-9, 14, 16-17</td>
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<td>Donnelly et al Novobiocin and Additional Inhibitors of the Hsp90 C-Terminal Nucleotide-binding Pocket Curr Med Chem , 2008, Vol 15(26) 2702-2717, Title, pg 26, Fig 8, pg 28 Fig 10</td>
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Further documents are listed in the continuation of Box C

Date of the actual completion of the international search
5 April 2010 (05 04 2010)

Date of mailing of the international search report
13 APR 2010

Name and mailing address of the ISA/US
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