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(54) Title: METHOD FOR DIAGNOSIS OF LYME ARTHRITIS, METHOD FOR DIFFERENTIAL DIAGNOSIS OF LYME ARTHRITIS, LYSOPHOSPHATIDYLETHANOLAMINE FOR USE AS BIOMARKER, KIT FOR DIAGNOSIS OF LYME ARTHRITIS AND KIT FOR DIFFERENTIAL DIAGNOSIS OF LYME ARTHRITIS

(57) Abstract: The subject matter of the invention relates to a method for *in vitro* diagnosis of Lyme disease and a method for *in vitro* differential diagnosis of Lyme arthritis versus rheumatoid arthritis, in which methods, in a sample from a subject, the level of lysophosphatidylethanolamine comprising myristic acid (LysoPE(14:0)) is determined and such determined level of lysophosphatidylethanolamine is compared with the level of lysophosphatidylethanolamine comprising myristic acid in a reference sample; wherein the level of lysophosphatidylethanolamine comprising myristic acid which is higher than the level in the said reference sample indicates that the subject suffers from Lyme disease. The subject matter of the invention further relates to lysophosphatidylethanolamine comprising myristic for use as a biomarker of Lyme disease, as a biomarker of Lyme arthritis, as a biomarker for differential diagnosis of Lyme arthritis versus rheumatoid arthritis, as a biomarker of neuroborreliosis. The subject matter of the invention also relates to a kit for *in vitro* diagnosis of Lyme disease and a kit for *in vitro* differential diagnosis of Lyme arthritis, which kits comprise a means for determining the level of lysophosphatidylethanolamine comprising myristic acid and instructions for carrying out the methods for diagnosis according to the invention.



METHOD FOR DIAGNOSIS OF LYME ARTHRITIS, METHOD FOR DIFFERENTIAL DIAGNOSIS OF LYME ARTHRITIS, LYSOPHOSPHATIDYLETHANOLAMINE FOR USE AS BIOMARKER, KIT FOR DIAGNOSIS OF LYME ARTHRITIS AND KIT FOR DIFFERENTIAL DIAGNOSIS OF LYME ARTHRITIS

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The subject matter of the invention relates to a method for *in vitro* diagnosis of Lyme disease, a method for *in vitro* differential diagnosis of Lyme arthritis, lysophosphatidylethanolamine containing myristic acid for use as a biomarker of Lyme disease, as a biomarker of Lyme arthritis and neuroborreliosis, and also as a biomarker for differential diagnosis of (tick-borne) Lyme arthritis and further a kit for *in vitro* diagnosis of Lyme disease and a kit for *in vitro* differential diagnosis of Lyme arthritis.

#### **Technical Field**

15 The present invention relates generally to the field of medicine, more specifically to the diagnostics of Lyme disease, especially Lyme arthritis and neuroborreliosis, and in particular to differential diagnosis of Lyme arthritis versus reumathoid arthritis, with the use of a specific biological marker, i.e. a lysophospholipid comprising myristic acid, as a biomarker of Lyme disease, and in particular a biomarker of Lyme arthritis and neuroborreliosis, and also as a biomarker for differential diagnosis of (tick-borne) Lyme arthritis, and a kit for *in vitro* diagnosis of Lyme disease and a kit for *in vitro* differential diagnosis of Lyme arthritis.

#### **Background Art**

25 Lyme disease, also known as Lyme borreliosis or a disease caused by Lyme disease spirochetes, is an infectious disease which results from an infection caused by the bacterium *Borrelia burgdorferi* transmitted to an organism by the bite of an infected tick. Lyme disease occurs in human subjects, in particular from regions endemic for Lyme disease, or persons exposed to the risk of getting the disease, for example foresters and woodcutters.

30 In recent years, a significant increase in the incidence of Lyme disease in human subjects has been observed. Presently, the disease has a global spread and thus 250,000 cases of the disease are reported yearly in the USA, whereas in Europe the incidence is 200,000 cases, of which 15,000 cases of the disease are reported in Poland. In a human subject the

disease is caused by several species of Lyme disease spirochete, presently described as the burgdorferi group or *Borrelia burgdorferi* sensu lato, which comprises *Borrelia burgdorferi* sensu stricto, *B. garinii* and *B. afzelii*.

5 The problem of Lyme disease infections concerns also pets and domestic animals, which may contribute to significant economic losses in those sectors.

Lyme disease develops through three distinct stages, from an early stage of the infection until a late stage. In the early stage (Stage 1) the disease may be asymptomatic or may have flu-like symptoms. In 50-80% of cases a skin inflammatory rash can be observed several days after the tick bite, which has a very characteristic appearance and is called  
10 *erythema migrans* (EM). If left untreated, the disease gives various pathological symptoms, including articular, neurological, dermatological and heart symptoms. After several weeks from the infection, the disease transforms into Stage 2, which is manifested, among others, in Lyme arthritis (LA) and neurological disorders (neuroborreliosis). After several months or years from the infection, the disease evolves into a chronic atrophic form, the so-called  
15 Stage 3, which may involve the development of e.g. encephalopathy and/or encephalomyelitis. Similarity between the symptoms of Lyme disease and those of other unrelated diseases, as well as variability of those symptoms, make the clinical diagnosis difficult. The early stage of the disease may be asymptomatic until the disease develops into very advanced clinical stages, such as (tick-borne) Lyme arthritis or neuroborreliosis. In late  
20 stages of the disease additional, other, non-specific symptoms may appear which are difficult to diagnose using presently available diagnostic means.

Presently, the diagnosis of Lyme disease is based on immunological tests detecting the presence of anti-*Borrelia burgdorferi* antibodies in blood. These tests are usually performed by ELISA technique and then confirmed by Western blott technique. However,  
25 *Borrelia burgdorferi* sensu lato expresses various surface proteins by adaptation to various environments, which results in the genetic diversity and varied expression of *Borrelia burgdorferi* genes. This situation has a significant impact on the possibility of developing specific and sensitive kits for the diagnosis of Lyme disease. Anti-*Borrelia burgdorferi* IgM antibodies usually appear within several days or weeks from the beginning of the infection  
30 and may remain in the course of the disease. IgG antibodies appear later, in the majority of patients – about one month from the beginning of active infection, and they may remain in the organism for many years from the infection and elimination of symptoms. The assessment of serological response is complicated considering the multitude of species causing the disease and due to cross-species diversity for the main immunodominant

antigens, as well as genetic diversity. The results of serological tests with those antigens in terms of specificity and sensitivity are highly variable. Therefore, such tests are characterized by insufficient specificity due to possible cross-reactivity with antibodies connected with different pathogens, such as *Treponema pallidum*, other spirochetes, 5 *Rickettsia* or *Helicobacter pylori*. For this reason, it is necessary to carry out additional Western blot analyses in the case of positive samples in immunological tests in order to confirm the initial results obtained by ELISA technique. It is actually considered that an elevated level of antibodies in diseased persons is an insufficient diagnostic factor; therefore there is a need to develop an unambiguous biomarker of Lyme disease, including Lyme 10 arthritis and neuroborreliosis. Moreover, there are still reported cases of the disease, despite negative results of tests for antibodies.

The diagnosis of Lyme arthritis is additionally made more complicated by the fact that there is a disease which has practically the same clinical symptoms, i.e. rheumatoid arthritis (RA). Both diseases, despite almost identical symptoms involving *inter alia* joint 15 pains, require a totally different treatment, allowing for a prognosis of a positive outcome thereof. Therefore, there is a need for specific and sensitive means for differential diagnosis of Lyme arthritis versus rheumatoid arthritis, which will make it possible to apply adequate treatment and cure the disease.

Similarly, the diagnosis of neuroborreliosis is made more complicated by the fact 20 that there is another disease entity which has very similar symptoms, i.e. tick-borne encephalitis (TBE). Unlike neuroborreliosis, tick-borne encephalitis is caused by a virus and not bacterium. Thus, both diseases, despite almost identical symptoms, require a totally different treatment. Therefore, there is a need for specific means and diagnostic methods allowing unambiguous clinical diagnosis of those diseases, which will make it possible to 25 apply adequate treatment and curing of the disease.

Presently available diagnostic means and methods are therefore insufficient. There is therefore a need for specific and sensitive methods for *in vitro* diagnosis of Lyme disease, in particular Lyme arthritis and neuroborreliosis, and especially *in vitro* differential diagnosis of Lyme arthritis versus rheumatoid arthritis. There is also an urgent need for 30 providing specific means, including biomarkers and diagnostic kits, for specific diagnosis of Lyme disease, especially Lyme arthritis and neuroborreliosis, in particular for differential diagnosis versus other disease entities.

The object of the invention is therefore to provide methods and means for specific and sensitive diagnosis of Lyme disease, especially Lyme arthritis and neuroborreliosis, and

in particular differential diagnosis of Lyme arthritis versus rheumatoid arthritis, differential diagnosis of neuroborreliosis, such as specific and sensitive biomarkers as well as diagnostic methods and diagnostic kits using such biomarkers.

## 5 *Summary of Invention*

The above objects have been achieved by the solutions claimed in the attached patent claims. Preferable variants of the invention are defined in the dependent claims.

The subject matter of the invention relates to a method for *in vitro* diagnosis of Lyme disease in a subject, characterized in that:

10 a) in a sample from a subject, the level of lysophosphatidylethanolamine comprising myristic acid (LysoPE(14:0)) is determined, and

b) the level of lysophosphatidylethanolamine determined in a) is compared with the level of lysophosphatidylethanolamine comprising myristic acid in a reference sample; wherein the level of lysophosphatidylethanolamine comprising myristic acid which is higher  
15 than the level in the said reference sample indicates that the subject suffers from Lyme disease.

Preferably, in the method for *in vitro* diagnosis of Lyme disease according to the invention a sample selected from a group comprising whole blood, plasma, serum and cerebrospinal fluid is used as the sample from a subject.

20 More preferably, blood plasma is used as the sample from a subject.

Preferably, in the method for *in vitro* diagnosis of Lyme disease according to the invention the subject is a human subject.

Preferably, in the method for *in vitro* diagnosis of Lyme disease according to the invention the level of lysophosphatidylethanolamine comprising myristic acid is determined  
25 by liquid chromatography coupled with mass spectrometry (LC-MS) method, in particular LC-MS/MS method.

Preferably, in the method for *in vitro* diagnosis of Lyme disease according to the invention Lyme disease is Lyme arthritis or neuroborreliosis.

The subject matter of the invention further relates to a method for *in vitro*  
30 differential diagnosis of Lyme arthritis versus rheumatoid arthritis, characterized in that:

a) in a sample from a subject the level of lysophosphatidylethanolamine comprising myristic acid (LysoPE(14:0)) is determined, and

b) the level of lysophosphatidylethanolamine determined in a) is compared with the level in a reference sample; wherein the level of lysophosphatidylethanolamine comprising myristic acid which is higher than the level in the said reference sample indicates that the subject suffers from Lyme arthritis.

5 Preferably, in the method for *in vitro* differential diagnosis according to the invention a sample selected from a group comprising whole blood, plasma and serum is used as the sample from a subject.

More preferably, in the method for *in vitro* differential diagnosis according to the invention blood plasma is used as the sample from the subject.

10 Preferably, in the method for *in vitro* differential diagnosis according to the invention the subject is a human subject.

Preferably, in the method for *in vitro* differential diagnosis according to the invention the level of lysophosphatidylethanolamine comprising myristic acid is measured by liquid chromatography coupled with mass spectrometry (LC-MS) method, in particular  
15 LC-MS/MS method.

The subject matter of the invention also relates to lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of Lyme disease.

The subject matter of the invention also relates to lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of Lyme arthritis.

20 The subject matter of the invention also relates to lysophosphatidylethanolamine comprising myristic acid for use as a biomarker for differential diagnosis of Lyme arthritis versus rheumatoid arthritis.

The subject matter of the invention further relates to lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of neuroborreliosis.

25 The subject matter of the invention also relates to a kit for *in vitro* diagnosis of Lyme disease, characterised in that it comprises a means for determining the level of lysophosphatidylethanolamine comprising myristic acid and instructions for carrying out the method for *in vitro* diagnosis of Lyme disease as described above.

30 The subject matter of the invention also relates to a kit for *in vitro* differential diagnosis of Lyme arthritis, characterised in that it comprises a means for determining the level of lysophosphatidylethanolamine comprising myristic acid and instructions for carrying out the method for *in vitro* differential diagnosis of Lyme arthritis versus rheumatoid arthritis as described above.

### ***Detailed Description of Invention***

A biomarker, in other words a biological marker or a bioindicator, is a biological indicator which enables qualitative and/or quantitative assessment of various medical, pathological and disease conditions and/or biological phenomena or features. In modern medicine the biomarkers play invaluable role; they enable, among others, a quick, precise, specific and sensitive diagnosis of various diseases and disorders. Such biomarkers are defined as molecular, genetic and biochemical factors used for a precise and easy diagnosis of diseases, for example, chronic, genetic diseases, cancers, as well as for the assessment of progression of such diseases or monitoring the treatment thereof, or the assessment of the probability of occurrence of such kind of diseases in the examined subject.

A significant development in the field of research concerning changes in lipid metabolism has been seen over last decade. That is why an analysis of phospholipid profile has been suggested for the purpose of early clinical diagnosis of many illnesses, e.g. for the purpose of characterizing neoplasms or other diseases – see e.g. publications by Wymann M. P., and Schneider R. 2008. Lipid signalling in disease. *Nat. Rev. Mol. Cell Biol.* 9: 162–176 and Fernandis A. Z., and Wenk M. R. 2009. Lipid-based biomarkers for cancer. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* 877: 2830–2835. However, the analysis of phospholipids' profile has not been used so far for the diagnosis of Lyme disease, in particular Lyme arthritis and neuroborreliosis.

The present inventors have identified and developed a new biomarker which enables specific and sensitive diagnosis of Lyme disease, especially Lyme arthritis and neuroborreliosis, as well as differential diagnosis of Lyme arthritis and neuroborreliosis versus other diseases, such as rheumatoid arthritis (RA) or tick-borne encephalitis (TBE).

The biomarker identified by the present inventors is lysophosphatidylethanolamine comprising myristic acid (LPE (14:0) or 14:0 Lyso PE). It is a chemical compound having the molecular weight of 425.497 g/mol; the monoisotopic mass of 425.254 g/mol; the molecular formula:  $C_{19}H_{40}NO_7P$  and the structural formula as presented in Figure 1.

The subject matter of the invention thus relates to lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of Lyme disease, lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of Lyme arthritis, lysophosphatidylethanolamine comprising myristic acid for use as a biomarker for differential diagnosis of Lyme arthritis versus rheumatoid arthritis, lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of neuroborreliosis, as well as *in vitro* diagnostic methods and diagnostic kits using

lysophosphatidylethanolamine comprising myristic acid as a biomarker of Lyme disease, especially Lyme arthritis and neuroborreliosis, as well as differential diagnosis of Lyme arthritis. According to the invention it is also possible to make a differential diagnosis of neuroborreliosis versus tick-borne encephalitis (TBE).

5           Phosphatidylethanolamine comprising myristic acid is an exogenous compound that is synthesized in bacterial, fungal and plant organisms, but this compound is not synthesized in animal organisms, including human subjects. Therefore, the presence of trace amounts of compounds comprising this acid, including phospholipids, especially phosphatidylethanolamine, in animal organisms, including human subjects, is due to an  
10           exogenous factor, e.g. a diet including high amounts of fats of plant origin. Phospholipids constitute a basic structural element of each cell membrane. Among all identified phospholipids isolated from *Borrelia burgdorferi* cells, a significant proportion of lysophospholipids comprising a specific acid identified, that is myristic acid, has been reported. Lysophospholipids (LPLs) belong to a group phospholipids that are formed as a  
15           result of hydrolisys of phospholipids (PLs) with the participation of phospholipase A2 (PLA2) enzyme. It has been found that the said enzyme plays a key role in the immunological response of an organism to a bacterial infection. An increased PLA2 activity in the plasma of patients with Lyme arthritis has also been confirmed. Additionally, it has been demonstrated that bacteria have a specific protein that transports LPLs within their cell  
20           membrane, as a result of which they accumulate on the inner part of the membrane. See e.g. the following publications: Belisle, J. T., Brandt, M. E., Radolf, J. D., & Norgard, M. V. (1994). Fatty acids of *Treponema pallidum* and *Borrelia burgdorferi* lipoproteins. *Journal of bacteriology*, 176(8), 2151-2157; Ben-Menachem, G., Kubler-Kielb, J., Coxon, B., Yergey, A., & Schneerson, R. (2003). A newly discovered cholesteryl galactoside from *Borrelia burgdorferi*. *Proceedings of the National Academy of Sciences*, 100(13), 7913-7918; Fraser, C. M., Casjens, S., Huang, W. M., Sutton, G. G., Clayton, R., Lathigra, R., & Gwinn, M. (1997). Genomic sequence of a Lyme disease spirochaete, *Borrelia burgdorferi*. *Nature*, 390(6660), 580-586.  
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          However, so far lysophosphatidylethanolamine comprising myristic acid has neither  
30           been associated with the occurrence of any disease entity nor it has been used a biomarker for the diagnosis of any disease.

          As a result of studies carried out to assess changes in the phospholipid profile (unpublished data), the present inventors unexpectedly-found more than ten times higher concentration of lysophosphatidylethanolamine comprising myristic acid in body fluids,

such as plasma, from patients with Lyme disease, more specifically Lyme arthritis and neuroborreliosis, as compared to healthy persons as well as patients with RA and TBE but not affected by Lyme disease, Lyme arthritis or neuroborreliosis. The present inventors found the presence of lysophosphatidylethanolamine comprising myristic acid in the plasma  
5 of patients with rheumatoid arthritis (RA); however the content of that compound was not statistically different from the content in healthy persons. Similar results were obtained in the case of patients with neuroborreliosis.

Thus, the present invention provides lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of Lyme disease, especially Lyme arthritis and neuroborreliosis, and in particular for differential diagnosis of Lyme arthritis. The present  
10 invention also provides suitable *in vitro* diagnostic methods consisting in the determination of the said biomarker in a sample from a subject wherein an increased level or concentration of lysophosphatidylethanolamine comprising myristic acid in such a sample indicates the presence of Lyme disease, or Lyme arthritis or neuroborreliosis, in the examined subject.  
15 The examined subject is preferably a human subject, but it may also be an animal which needs a diagnosis in respect of Lyme disease, Lyme arthritis or neuroborreliosis. The reference sample according to the invention is a sample obtained from a control subject, which means a healthy subject without Lyme disease, without neuroborreliosis, without RA and without TBE. A sample from a subject with RA or TBE, but who simultaneously does not  
20 have Lyme disease and Lyme arthritis and neuroborreliosis, may also constitute such a reference sample.

The present invention also provides means, such as a biomarker and a diagnostic kit, and a method for differential diagnosis of Lyme disease versus other disease entities, in particular differential diagnosis of Lyme arthritis versus rheumatoid arthritis (RA) and  
25 neuroborreliosis versus tick-borne encephalitis (TBE).

The present inventors have found a highly specific diagnostic relationship between lysophosphatidylethanolamine comprising myristic acid and Lyme arthritis and neuroborreliosis. So far, lysophosphatidylethanolamine comprising myristic acid has not been used as a diagnostic biomarker and has not been associated with any disease entity, and it  
30 has not been used for diagnostic purposes either. The present inventors have demonstrated that lysophosphatidylethanolamine comprising myristic acid is a specific and sensitive biomarker of Lyme disease, especially Lyme arthritis and neuroborreliosis.

Determination of the level of lysophosphatidylethanolamine comprising myristic acid in the methods according to the invention may be conducted using standard methods

known in this field. Preferably, the level thereof is determined using a highly-specialized analytical technique i.e. liquid chromatography coupled with mass spectrometry (LC-MS), or more preferably by LC-MS/MS method, which methods as such are known in this field. It is also possible to determine the level of this compound by other analytical methods known  
5 in this field. For this purpose, antiphospholipid antibodies (APa), for example, can be used, which are directed against phospholipids and phospholipid-binding plasma proteins. Such an analytical method simplifies the procedure for determining the level of lysophosphatidylethanolamine comprising myristic acid in a sample from a subject and at the same time is characterized by a high specificity of determination.

10 The present invention thus provides a diagnostic kit for the diagnosis of Lyme disease comprising a means for determining lysophosphatidylethanolamine comprising myristic acid. The present invention also provides a diagnostic kit for differential diagnosis of Lyme arthritis comprising a means for determining lysophosphatidylethanolamine comprising myristic acid. The kits according to the invention may be used for a precise diagnosis of  
15 Lyme disease and Lyme arthritis in subjects exhibiting initial symptoms of the disease confirmed by presently used immunologic methods. The diagnostic kits according to the invention enable a simple and quick detection of the presence of lysophosphatidylethanolamine comprising myristic acid and its quantitative assessment in the examined sample from a subject, e.g. in plasma, and thus the kits are useful diagnostic  
20 tools which allow one to make a diagnosis of Lyme disease, and especially differential diagnosis of Lyme arthritis versus RA or alternatively neuroboreriosis versus TBE. The means for determining lysophosphatidylethanolamine comprising myristic acid in the kits according to the invention may be any means suitable for determining lysophosphatidylethanolamine comprising myristic acid in a sample from a subject, such as  
25 a sample of the body fluid from a subject, for example, cerebrospinal fluid or preferably plasma. For example, these may be antiphospholipid antibodies binding to this biomarker, i.e. LPE (14:0).

As a result of studies carried out by the present inventors to assess changes in the phospholipid profile in persons with Lyme arthritis versus healthy persons (unpublished  
30 data) and versus persons suffering from RA, it was unexpectedly found that the concentration of the lysophospholipid comprising specific acid, i.e. lysophosphatidylethanolamine comprising myristic acid, in the plasma of patients with Lyme arthritis was more than ten times higher compared to healthy persons. It was thus demonstrated that the presence of lysophosphatidylethanolamine comprising myristic acid

in body fluids, such as blood plasma, is a useful biomarker of Lyme disease, especially Lyme arthritis. Similarly, LPE (14:0) may be a useful biomarker of neuroborreliosis, which enables specific diagnosis of neuroborreliosis, especially differential diagnosis of neuroborreliosis versus TBE.

5           Lysophosphatidylethanolamine comprising myristic acid is present only in trace amounts in the plasma of healthy human subjects, which fact also applies to other phospholipids comprising myristic acid. This is also true for patients with RA. It results from the fact that myristic acid in the identified lysopholipid is an exogenous compound synthesized in bacterial, fungal or plant organisms, whereas it is not synthesized in the  
10 animal organism, including human organism. Therefore, the presence of trace amounts of compounds comprising myristic acid, including phospholipids, in the human organism is caused by a diet including large amounts of fats of plant origin. Phospholipids constitute a basic structural element of every cell membrane. Among all identified phospholipids isolated from *Borrelia burgdorferi* cells, a significant proportion of phospholipids  
15 comprising the identified specific acid has been reported. Lysophospholipids (LPLs) belong to a group of phospholipids formed as a result of hydrolysis of phospholipids (PLs) with the participation of phospholipase A2 (PLA2) enzyme. It has been found that the said enzyme plays a key role in the immunological response of an organism to a bacterial infection. Increased activity of PLA2 in the plasma of patients with Lyme arthritis has been also  
20 confirmed. Additionally, it has been demonstrated that bacteria have a specific protein that transports LPLs within their cell membrane, as a result of which they accumulate on the inner part of the membrane.

All the above information, when taken together, provides a basis for the conclusion that an increased content of the lysophospholipid comprising myristic acid in the samples  
25 from patients with Lyme disease, in particular with Lyme arthritis or neuroborreliosis, results from the presence of *Borrelia burgdorferi* in the subject's organism, which fact simultaneously indicates that the examined subject suffers from Lyme disease or Lyme arthritis or neuroborreliosis, respectively, and confirms the usefulness of this compound as a biomarker of these diseases as well as its usefulness in the methods for *in vitro* diagnosis  
30 of these diseases.

Additionally, in the case of Lyme arthritis, the diagnosis of which is additionally made more complicated by the presence of a disease which has practically identical clinical symptoms, i.e. rheumatoid arthritis (RA), the invention enables differential diagnosis of Lyme arthritis versus rheumatoid arthritis. Both diseases, despite almost identical symptoms,

require a totally different treatment. As a result of studies carried out by the present inventors to assess changes in the phospholipid profile in persons with rheumatoid arthritis versus healthy persons (unpublished data), the presence of lysophospholipid comprising myristic acid was reported in the plasma of patients with RA; however, the content of that compound was not statistically different from the content thereof in the healthy persons. Therefore, the present invention provides lysophosphatidylethanolamine comprising myristic acid as a biomarker of Lyme arthritis, and it further provides a method for *in vitro* diagnosis of this disease, which makes it possible to avoid diagnostic errors indicating RA. Preferably, the body fluid used in the diagnostic method according to the invention is plasma.

Similar results were obtained by the present inventors in the case of neuroborreliosis.

The present invention will now be illustrated in the example below, which however is not intended to limit in any way the scope of the invention defined in the patent claims. Unless indicated otherwise, all methods and parameters are as commonly used in the field to which this invention belongs, and the applied devices and reagents are used in a manner as recommended by the manufacturers thereof.

### ***Description of Figures***

**Figure 1** – shows the structural formula of lysophosphatidylethanolamine comprising myristic acid.

**Figure 2** – Fig. 2A shows a representative Total Ion Chromatogram (TIC) in negative ionisation mode for blood plasma of a patient with rheumatoid arthritis (RA) (the top chromatogram); Extracted Ion Chromatogram (EIC) for  $m/z$  424.6424 in negative ionisation mode (the bottom chromatogram); Fig. 2B – shows a representative mass spectrum MS of the peak at  $RT = 12.06$  min (the top spectrum); the fragmentation mass spectrum MS/MS of the ion having  $m/z$  424.2442 corresponding to the molecular ion of LPE (14:0) (the bottom spectrum) – the  $m/z$  227.2016 signal confirms the presence of myristic acid (14:0) in the structure of the fragmented compound.

**Figure 3** – shows a box-and-whisker plot which presents graphically changes in the relative content of LPE (14:0) in healthy persons, patients with rheumatoid arthritis and patients with Lyme arthritis. ( $p < 0.0001$ \*\*\*\*). LA – Lyme arthritis; RA – rheumatoid arthritis; C - control.

**Figure 4** – shows representative Total Ion Chromatograms (TICs), as a phospholipid profile in negative ionization mode: plasma from a patient with Lyme arthritis – panel A;

plasma from a healthy person – panel B; plasma from a patient with rheumatoid arthritis – panel C.

**Figure 5** – shows representative Extracted Ion Chromatograms (EICs) in negative ionization mode for  $m/z$  424.6424 corresponding to the molecular ion of LPE(14:0), retention time RT = 12 min: plasma from a patient with Lyme arthritis – panel A; plasma from a healthy person – panel B; plasma from a patient with rheumatoid arthritis – panel C.

**Figure 6** – shows mass spectrum MS of the peak at RT=12.06 min (the top panel); the fragmentation mass spectrum MS/MS of the ion having  $m/z$  424.2442 corresponding to the molecular ion of LPE (14:0) (the bottom panel) – the  $m/z$  227.2016 signal confirms the presence of myristic acid (14:0) in the structure of the fragmented compound.

**Figure 7** – shows representative overlaid-mode Extracted Ion Chromatograms (EICs) in negative ionization mode for  $m/z$  424.6424 corresponding to the molecular ion of LPE(14:0); retention time RT = 12 min: LA – plasma from a patient with Lyme arthritis; C – plasma from a healthy person; RA – plasma from a patient with rheumatoid arthritis.

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### **Example – Determination of the LPE (14:0) level**

#### **Test material**

The biological material used for the analyses was plasma obtained from the venous blood of persons suffering from rheumatoid arthritis (RA), persons suffering from Lyme arthritis (LA) and healthy persons. LA group consisted of 9 patients with diagnosed (tick-borne) Lyme arthritis (2 women and 7 men) aged from 22 to 81 years (the mean age was 49 years) hospitalized in the Department of Infectious Diseases and Neuroinfection of the University of Bialystok. The illness was diagnosed on the basis of the presence of anti-*Borrelia burgdorferi* specific IgM and/or IgG antibodies in blood serum, detected by ELISA method. RA group consisted of 9 patients with active RA (2 women and 7 men) aged 23 to 79 years (the mean age was 48 years). The assessment of activity of the disease was based on the four-variable DAS28-CRP parameter. The control group consisted of 9 healthy persons (2 women and 7 men) aged from 24 to 71 years (the mean age was 47 years). Venous blood was obtained from both the patients and healthy persons to heparinized test tubes. The samples were centrifuged at 2000 x g, in 4°C for 20 minutes in order to obtain plasma. After adding buthylhydroxytoluene (BHT), acting as an antioxidant, the samples were kept at -80°C until analysis. The study was started after obtaining the consent of the Bioethics

Committee of the Medical University of Bialystok and a written consent of each person participating in the study.

The compound was identified in both the plasma from patients and the plasma from healthy persons by a highly specialized analytical technique LC-MS using hydrophilic interaction liquid chromatography (HILIC), after previous isolation of lipid fraction by the solvent-solvent extraction method as described in detail below.

#### **Extraction of lipids from blood plasma**

Extraction was carried out in glass test tubes previously washed with chloroform (CHCl<sub>3</sub>) in order to remove any impurities. 200 µl of plasma and 1.5 ml of methanol (MeOH) cooled to -20°C was placed in each test tube. The mixture obtained was vortexed for 10 minutes. Then, 3 ml of CHCl<sub>3</sub> cooled to -20°C was added and was again vortexed for 3 minutes. In the next stage, extracted samples were incubated in ice for one hour. During incubation, from time to time, the content of the test tubes was vortexed for a moment. After incubation, in order to initiate separation of phases, 1.25 ml of ultrapure (Milli-Q) water was added and the samples were again left in ice for 10 minutes, their content being mixed in Vortex shaker from time to time. Then, the samples were centrifuged at 2500 x g for 10 minutes. After centrifuging, the bottom organic phase was transferred to a new glass test tube and another extraction was carried out by adding 2 ml of CHCl<sub>3</sub>/MeOH (2:1, v/v) mixture cooled to -20°C. The next obtained portion of chloroform layer was combined with the previously collected portion and evaporated to dry residue in a nitrogen atmosphere. The extracts obtained were kept in -80°C until analysis.

#### **Determination of the profile of phospholipids in blood plasma using the LC-MS-QTOF system**

To obtain the profile of phospholipids, the obtained extracts of blood plasma lipids from the patients with rheumatoid arthritis, patients with Lyme arthritis and healthy persons were analysed using LC-MS-QTOF in data dependent MS/MS mode (auto MS/MS). Separation of phospholipids classes was carried out with the use of hydrophilic interaction liquid chromatography (HILIC) using Ascentis Si HPLC Pore column, 15 cm x 1.0 mm, 3 mm; (Sigma–Aldrich) and gradient elution using a combination of two mobile phases A and B. Mobile phase A contained 25% water, 50% acetonitrile, and 25% v/v methanol and 10 mM addition of salt in the form of ammonium acetate. Mobile phase B contained 60%

acetonitrile, 40% methanol and 10 mM addition of ammonium acetate. The gradient applied at 40 ml/min mobile phase flow used a mixture having the following composition:

0% mobile phase A and 100% mobile phase B [0 min]

100% mobile phase A and 0% mobile phase B [20 min]

5 100% mobile phase A and 0% mobile phase B [35 min]

0% mobile phase A and 100% mobile phase B [45 min]

Mobile phase B was used as a solvent for samples of extracts comprising 20 µg of phospholipids. 5 µl of such prepared samples were injected into the chromatographic column.

10 Samples of the extracts were analysed in negative ionisation mode using electrospray ionisation (ESI) sources.

Agilent Technologies ultra-performance liquid chromatograph (UPLC), series 1290; Agilent Technologies QTOF mass detector, 6540, equipped with an electrospray ionization (ESI) source; Peak Scientific LC-MS 20 nitrogen generator; Ascentis Si HPLC Pore  
15 chromatographic column, 15 cm x 1.0 mm, 3 mm, Sigma–Aldrich were used for the diagnosis by the method according to the invention. However, the methods according to the invention may be carried out using any other devices, systems and analytical kits which enable quantitative determination of lysophosphatidylethanolamine comprising myristic acid.

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### ***Statistical analysis***

The results obtained were subjected to statistical analysis in the following way. Fold change of relative LPE (14:0) content in healthy persons, patients with rheumatoid arthritis and patients with Lyme arthritis was analysed. The statistical analysis was performed based  
25 on one-way ANOVA analysis. Relative amount of LPE (14:0) was calculated by dividing the peak area of LPE (14:0) obtained on the basis of Extracted Ion Chromatograms (EICs) by the peak area of PC (28:0) being an Internal Standard (ISTD). The results were expressed as arithmetical means and mean standard errors. The comparisons between the control group and the persons suffering from rheumatoid arthritis and Lyme arthritis were made using one-  
30 way ANOVA analysis and the post hoc test: Tukey's HSD test, with the use of the statistical package GraphPad Prism v.7.0, GraphPad Software, USA. The differences at  $p < 0.05$  were considered as statistically significant.

The results obtained are presented in Table 1 below and in Figures 2 to 7.

**Table 1.** Fold change of relative content of LPE (14:0) in healthy persons, patients with rheumatoid arthritis and patients with Lyme arthritis. Abbreviations: LA – Lyme arthritis; RA – rheumatoid arthritis; LPE - lysophosphatidylethanolamine; RT – retention time; ID – abbreviation of compound name; C – control.

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m/z	RT	ID	Log <sub>2</sub> (fold change)					
			LA versus C	Value p	RA versus C	Value p	LA versus RA	Value p
424.2464	12.38	LPE (14:0)	3.937	<0.0001	0.691	ns	-3.246	<0.0001

Figure 3 shows a box-and-whisker plot which presents graphically changes in the relative content of LPE (14:0) in healthy persons (C), patients with rheumatoid arthritis (RA) and patients with Lyme arthritis (LA). ( $p < 0.0001^{****}$ ). It has been demonstrated that the relative content of LPE (14:0) in patients with Lyme arthritis is statistically significantly higher compared to both healthy human subjects (control) and patients with RA.

Figure 4 shows representative exemplary Total Ion Chromatograms (TICs) as a phospholipid profile in negative ionisation mode: plasma from a patient with Lyme arthritis – panel A; plasma from a healthy person – panel B; plasma from a patient with rheumatoid arthritis – panel C.

Figure 5 shows representative exemplary Extracted Ion Chromatograms (EICs) in negative ionisation mode for m/z 424.6424 corresponding to the molecular ion of LPE (14:0), retention time RT=12 min: plasma for a patient with Lyme arthritis – panel A; plasma from a healthy person – panel B; plasma from a patient with rheumatoid arthritis – panel C. The difference in the obtained chromatogram profile of a patient with Lyme arthritis (panel A) can be clearly seen, which indicates the presence of LPE (14:0) and at the same time allows the diagnosis of the Lyme disease. A markedly higher peak of lysophospholipid comprising myristic acid, i.e. LPE (14:0) can be noticed in panel A as opposed to panels B and C.

Figure 6 shows MS mass spectrum of the peak at RT=12.06 min (top panel); fragmentation MS/MS mass spectrum of an ion having m/z 424.2442 corresponding the molecular ion of LPE (14:0) (bottom panel) – the m/z 227.2016 signal confirms the presence of myristic acid (14:0) in the structure of the fragmented compound.

Figure 7 shows representative overlaid mode Extracted Ion Chromatograms (EICs) in negative ionization mode for  $m/z$  424.6424 corresponding to the molecular ion of LPE (14:0), retention time  $RT=12$  min: plasma from a patient with Lyme arthritis - LA; plasma from a healthy person - C; plasma from a patient with rheumatoid arthritis - RA. It can be clearly seen that the highest peak was obtained for the patient with Lyme arthritis, which indicates that the method according to the invention using a biomarker in the form of lysophosphatidylethanolamine comprising myristic acid enables specific diagnosis of Lyme disease, in particular the diagnosis of Lyme arthritis versus a healthy person and differential diagnosis versus a person suffering from RA (but which person does not suffer from Lyme disease or Lyme arthritis or neuroborreliosis)

The data presented confirm that the methods according to the invention enable precise, sensitive and specific differentiation between a subject suffering from Lyme disease, especially Lyme arthritis, and both a healthy subject and a subject suffering from RA (but which subject does not suffer from Lyme disease or Lyme arthritis).

In summary, as it can be seen in the above Table 1 and in the presented Figures, the statistically significant differences in the content of LPE (14:0) were obtained in the studied samples from persons suffering from Lyme disease, especially Lyme arthritis versus the samples from healthy persons as well as persons suffering from rheumatoid arthritis (but without Lyme disease or Lyme arthritis or neuroborreliosis). In the samples of plasma obtained from patients with Lyme arthritis the relative content of LPE (14:0) was significantly higher than in the samples of plasma from healthy persons and persons with RA not suffering from Lyme disease/Lyme arthritis. The results obtained indicate that LPE (14:0) may be effectively used as a biomarker of Lyme disease, and in particular as a biomarker of Lyme arthritis, enabling specific diagnosis of Lyme arthritis, especially differential diagnosis of Lyme arthritis versus rheumatoid arthritis.

### Claims

1. A method for *in vitro* diagnosis of Lyme disease in a subject, characterized in that:
  - b) in a sample from a subject, the level of lysophosphatidylethanolamine comprising myristic acid (LysoPE(14:0)) is determined, and
  - b) the level of lysophosphatidylethanolamine determined in a) is compared with the level of lysophosphatidylethanolamine comprising myristic acid in a reference sample; wherein the level of lysophosphatidylethanolamine comprising myristic acid which is higher than the level in the said reference sample indicates that the subject suffers from Lyme disease.
2. The method for *in vitro* diagnosis of Lyme disease according to claim 1, characterized in that a sample selected from a group comprising whole blood, plasma, serum and cerebrospinal fluid is used as the sample from a subject.
3. The method for *in vitro* diagnosis of Lyme disease according to claim 2, characterized in that the sample is blood plasma.
4. The method for *in vitro* diagnosis of Lyme disease according to any one of claims 1 to 3, characterized in that the subject is a human subject.
5. The method for *in vitro* diagnosis of Lyme disease according to any one of claims 1 to 4, characterized in that the level of lysophosphatidylethanolamine comprising myristic acid is determined by liquid chromatography coupled with mass spectrometry (LC-MS) method, in particular LC-MS/MS method.
6. The method for *in vitro* diagnosis of Lyme disease according to any one of claims 1 to 5, characterized in that Lyme disease is Lyme arthritis or neuroborreliosis.
7. A method for *in vitro* differential diagnosis of Lyme arthritis versus rheumatoid arthritis, characterized in that:
  - a) in a sample from a subject, the level of lysophosphatidylethanolamine comprising myristic acid (LysoPE(14:0)) is determined, and

b) the level of lysophosphatidylethanolamine determined in a) is compared with the level in a reference sample; wherein the level of lysophosphatidylethanolamine comprising myristic acid which is higher than the level in the said reference sample indicates that the subject suffers from Lyme arthritis.

8. The method for *in vitro* differential diagnosis according to claim 7, characterized in that a sample selected from a group comprising whole blood, plasma and serum is used as the sample from a subject.

9. The method for *in vitro* differential diagnosis according to claim 8, characterized in that the sample is blood plasma.

10. The method for *in vitro* differential diagnosis according to any one of claims 7 to 9, characterized in that the subject is a human subject.

11. The method for *in vitro* differential diagnosis according to any one of claims 7 to 10, characterized in that the level of lysophosphatidylethanolamine comprising myristic acid is measured by liquid chromatography-mass spectrometry (LC-MS) method, in particular LC-MS/MS method.

12. Lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of Lyme disease.

13. Lysophosphatidylethanolamine comprising myristic acid for use as a biomarker of Lyme arthritis.

14. Lysophosphatidylethanolamine comprising myristic acid for use a biomarker for differential diagnosis of Lyme arthritis versus rheumatoid arthritis.

15. Lysophosphatidylethanolamine comprising myristic acid for use a biomarker of neuroborreliosis.

16. A kit for *in vitro* diagnosis of Lyme disease, characterised in that it comprises a means for determining the level of lysophosphatidylethanolamine comprising myristic acid and instructions for carrying out the method for *in vitro* diagnosis of Lyme disease according to any one of claims 1 to 6.

17. A kit for *in vitro* differential diagnosis of Lyme arthritis, characterised in that it comprises a means for determining the level lysophosphatidylethanolamine comprising myristic acid and instructions for carrying out the method for *in vitro* differential diagnosis of Lyme arthritis versus rheumatoid arthritis according to any one of claims 7 to 11.

Fig. 1

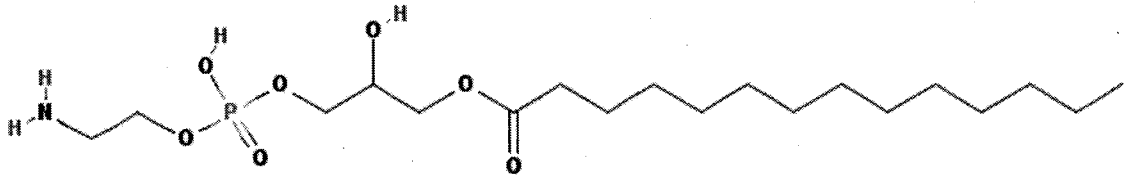


Fig. 2

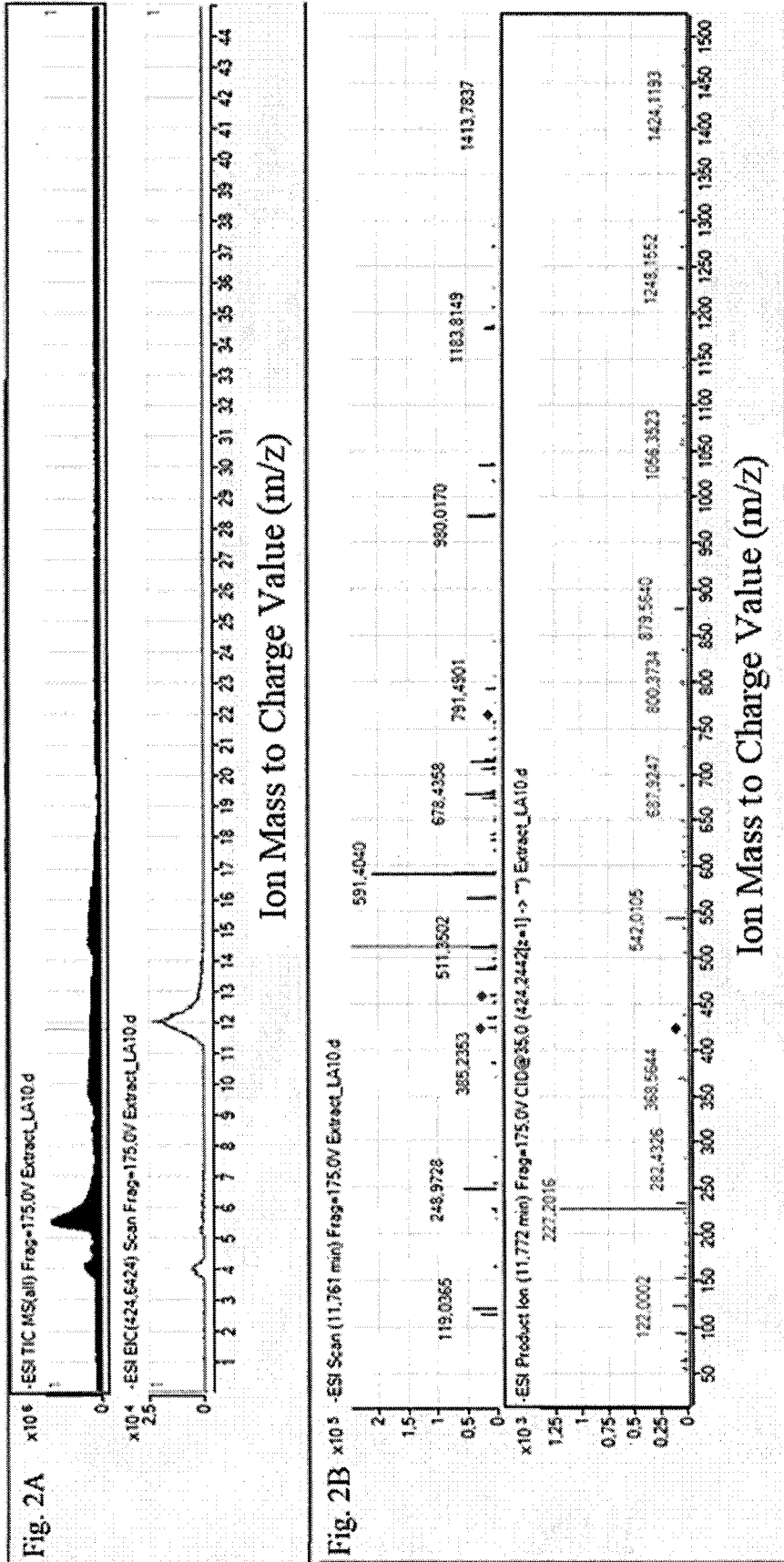


Fig. 3

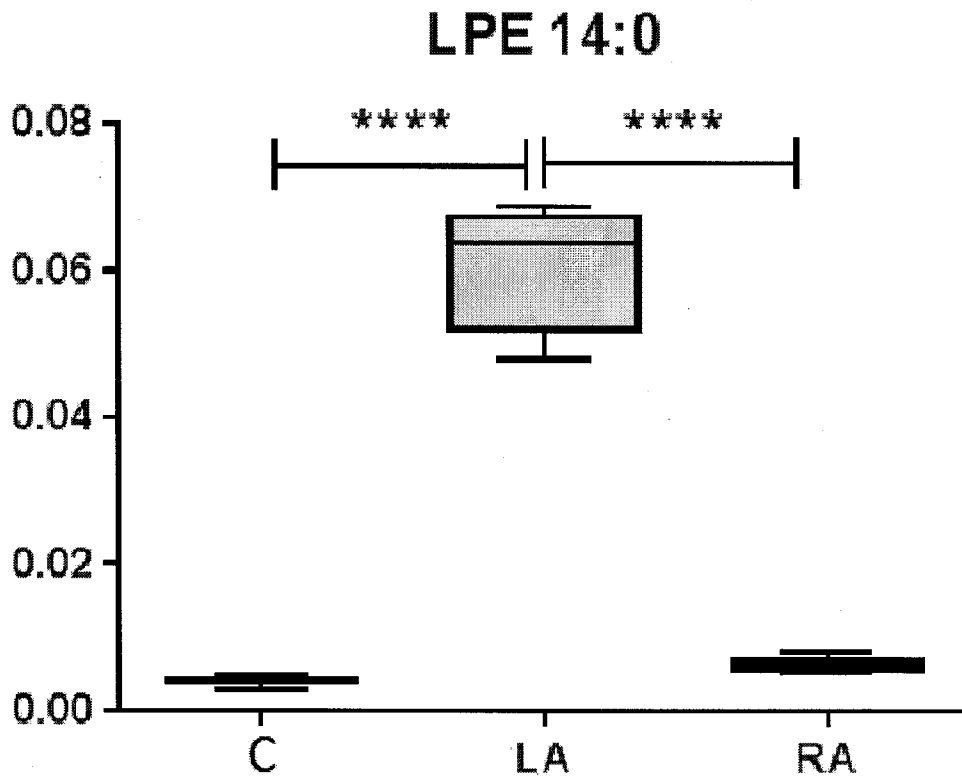
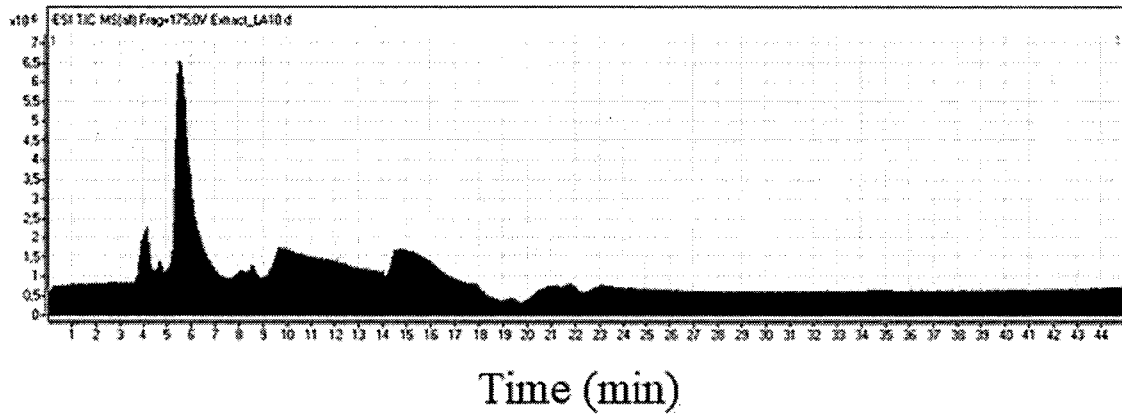
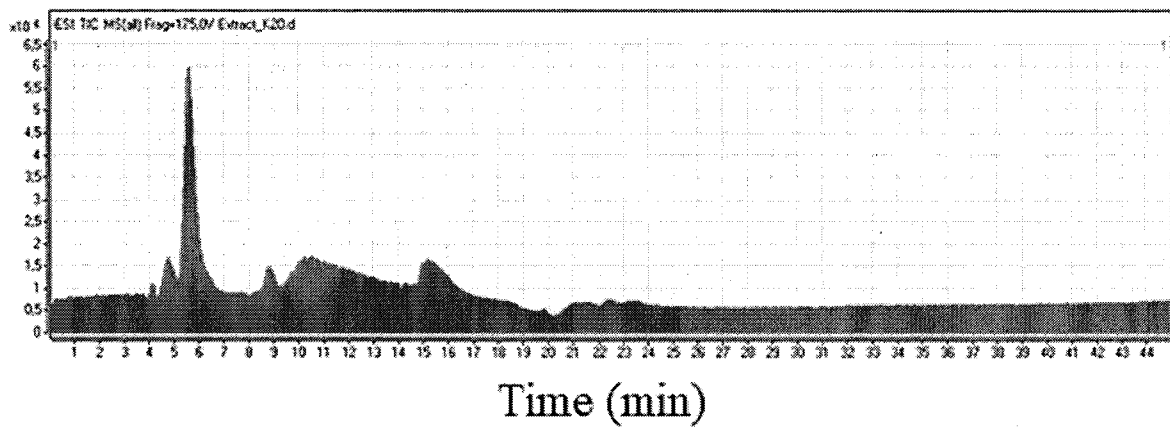


Fig. 4

Panel A



Panel B



Panel C

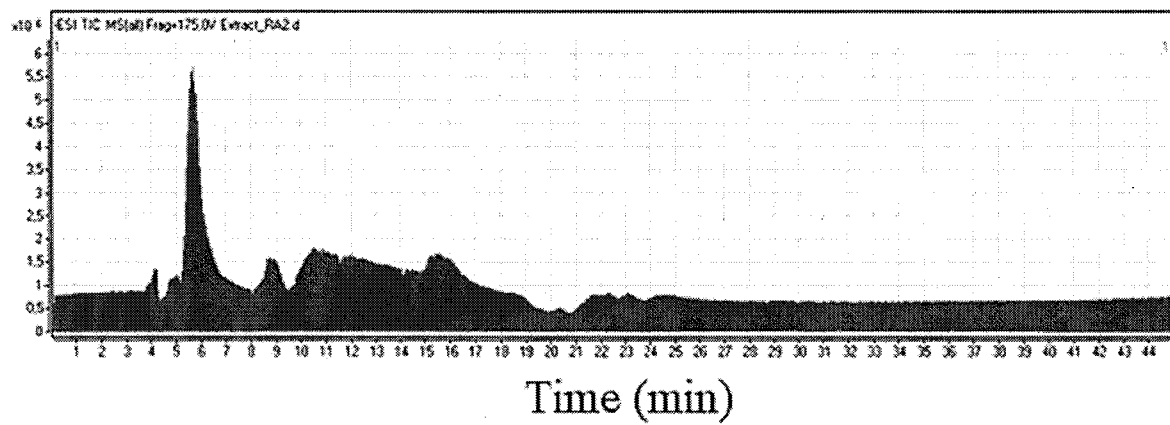
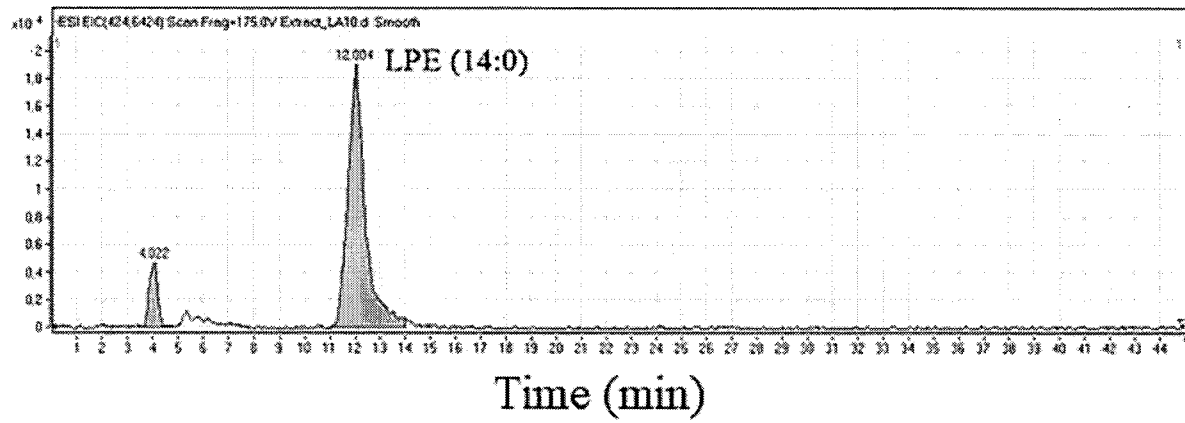
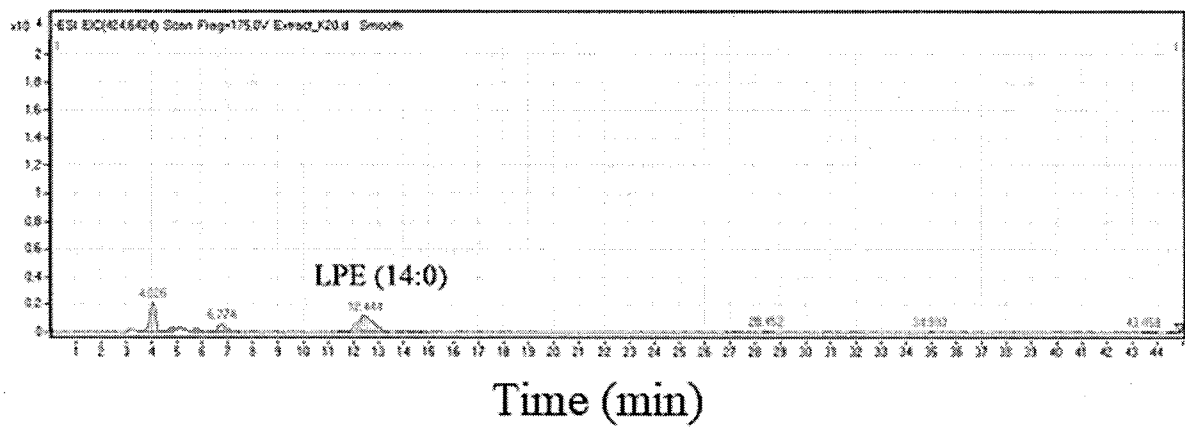


Fig. 5

Panel A



Panel B



Panel C

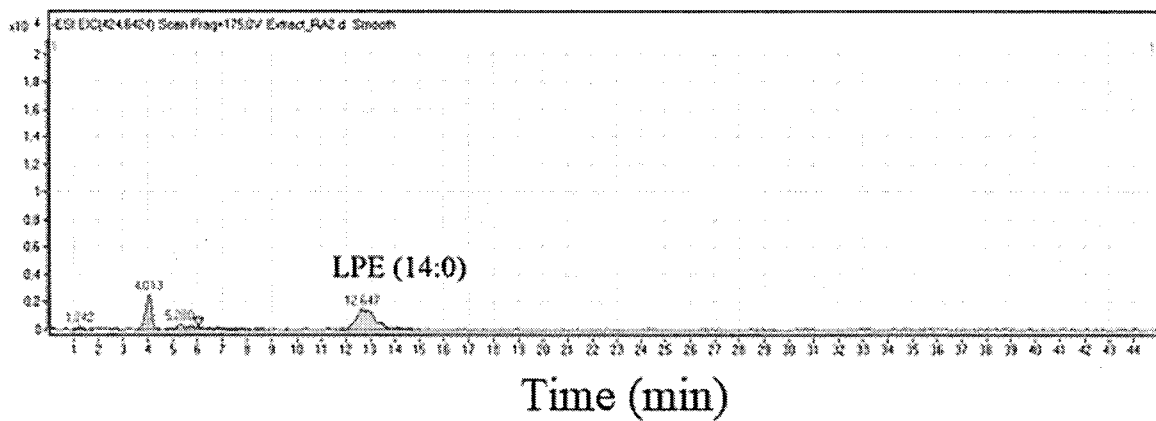


Fig. 6

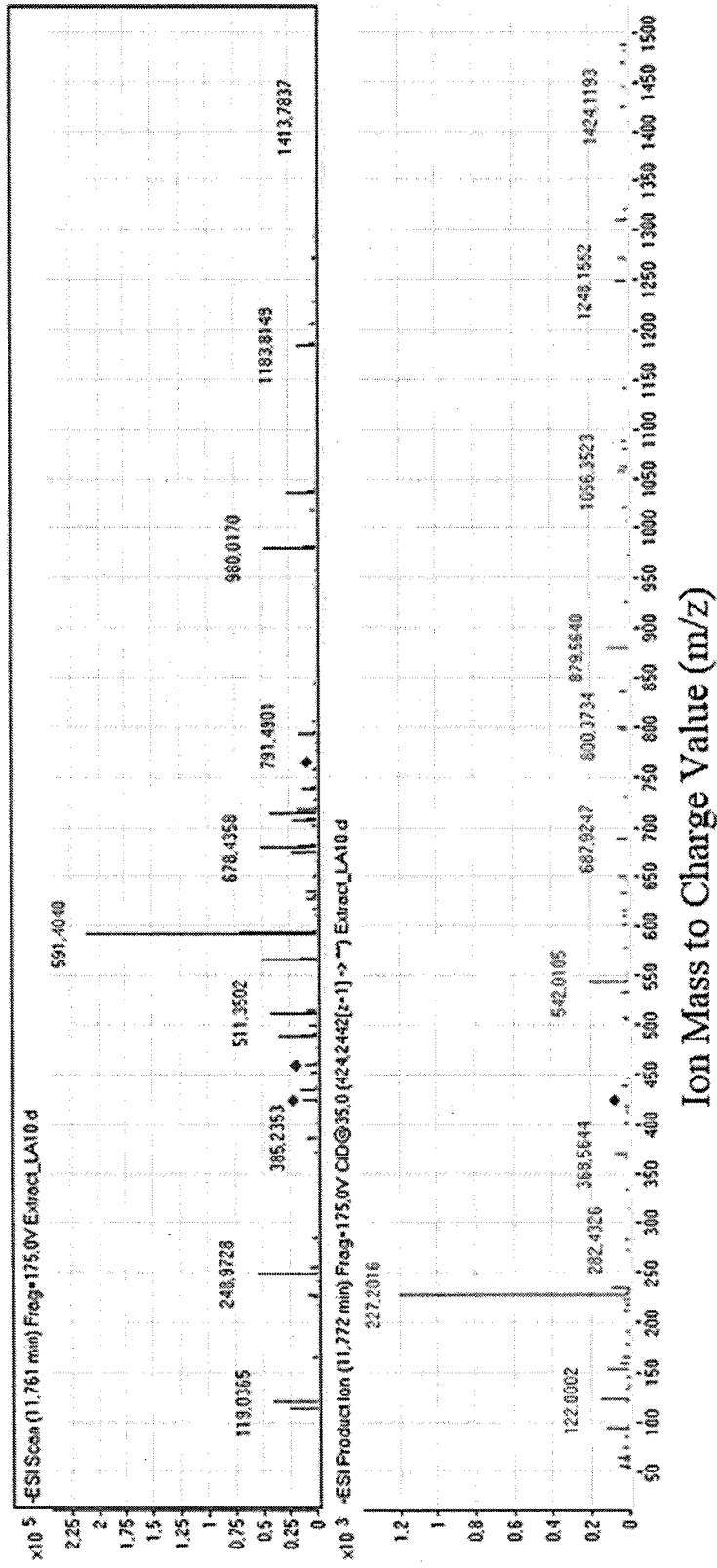
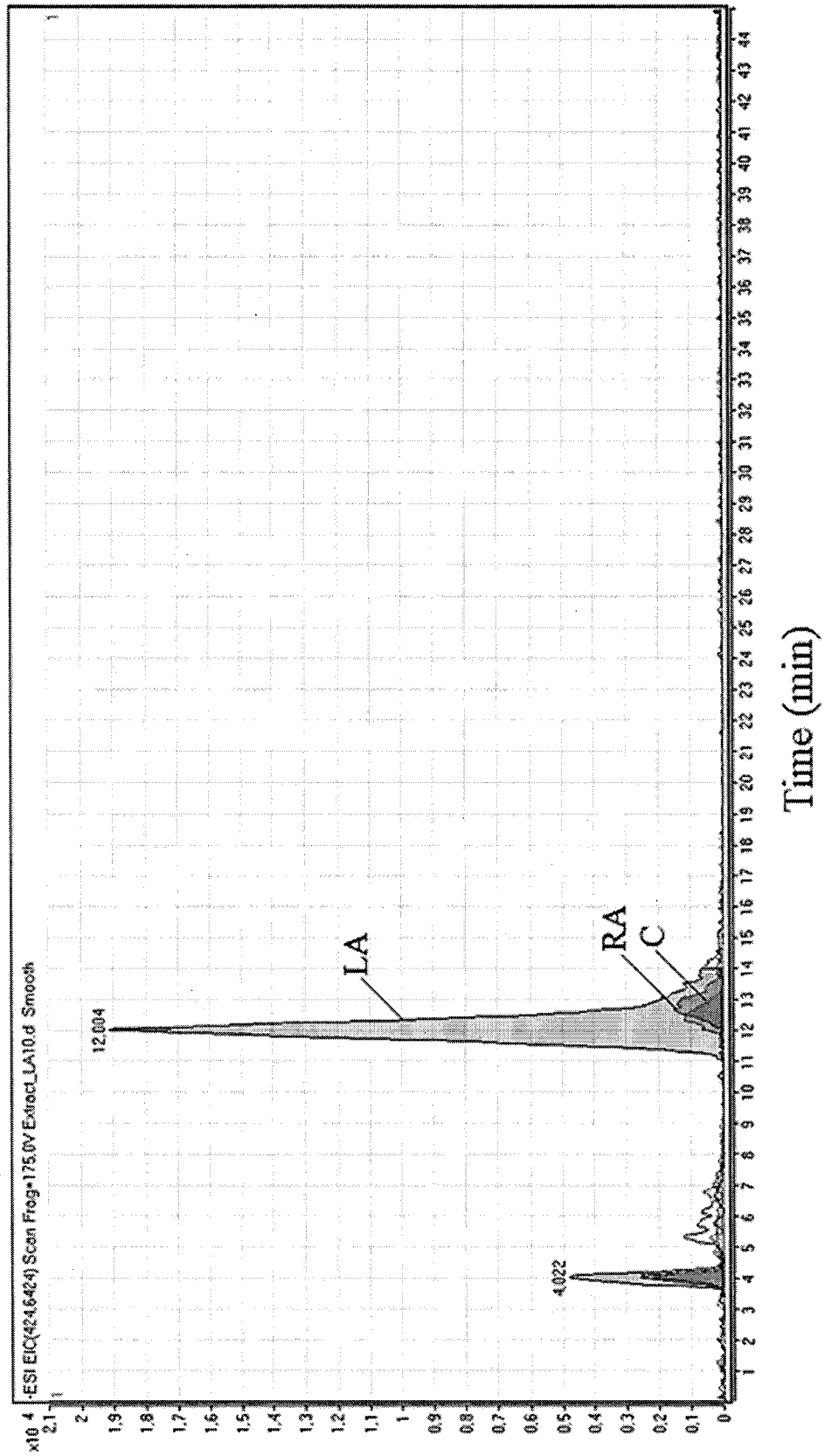


Fig. 7



**INTERNATIONAL SEARCH REPORT**

International application No  
PCT/PL2019/000047

**A. CLASSIFICATION OF SUBJECT MATTER**  
 INV. G01N33/569 G01N33/92  
 ADD.  
 According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**  
 Minimum documentation searched (classification system followed by classification symbols)  
 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)  
 EPO-Internal, WPI Data, BIOSIS, EMBASE

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2015/192597 A1 (KELAVKAR UDDHAV [US]) 9 July 2015 (2015-07-09) claims 1,4	12-15
X	LUCZAJ W ET AL: "Phospholipidomic Analysis Reveals Changes in Sphingomyelin and Lysophosphatidylcholine Profiles in Plasma from Patients with Neuroborreliosis", LIPIDS, SPRINGER, DE, vol. 52, no. 1, 10 November 2016 (2016-11-10), pages 93-98, XP036130392, ISSN: 0024-4201, DOI: 10.1007/S11745-016-4212-3 [retrieved on 2016-11-10] figures 2-5	1-6,16

Further documents are listed in the continuation of Box C.

See patent family annex.

\* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
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- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "&" document member of the same patent family

Date of the actual completion of the international search  21 October 2019	Date of mailing of the international search report  04/11/2019
Name and mailing address of the ISA/ European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Authorized officer  Wiesner, Martina

## INTERNATIONAL SEARCH REPORT

International application No

PCT/PL2019/000047

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT		
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>TAM VINCENT C ED - LAMBRIS DR JOHN D:  "Lipidomic profiling of bioactive lipids  by mass spectrometry during microbial  infections",  SEMINARS IN IMMUNOLOGY,  vol. 25, no. 3, 2013, pages 240-248,  XP028776088,  ISSN: 1044-5323, DOI:  10.1016/J.SMIM.2013.08.006  the whole document</p> <p style="text-align: center;">-----</p>	1-17
A	<p>VICTORIA A. BLAHO ET AL: "Lipidomic  Analysis of Dynamic Eicosanoid Responses  during the Induction and Resolution of  Lyme Arthritis",  JOURNAL OF BIOLOGICAL CHEMISTRY,  vol. 284, no. 32, 1 June 2009 (2009-06-01)  , pages 21599-21612, XP055634213,  ISSN: 0021-9258, DOI:  10.1074/jbc.M109.003822  the whole document</p> <p style="text-align: center;">-----</p>	1-17

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/PL2019/000047

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 2015192597 A1	09-07-2015	US 2015192597 A1 WO 2015105917 A1	09-07-2015 16-07-2015
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