(21) International Application Number: PCT/US2004/031412

(22) International Filing Date: 24 September 2004 (24.09.2004)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
- 0322395.5 24 September 2003 (24.09.2003) GB
- 0404567.0 1 March 2004 (01.03.2004) GB

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(54) Title: 1,3-DIACETYLATED,26,27-ALKYL/HALOALKYL VITAMIN D₃ COMPOUNDS AND METHODS OF USE THEREOF

(57) Abstract: The invention provides (1,3)-diacetylated vitamin D₃ analogs of cholecalciferol, substituted at carbon (20) with methyl or cyclopropyl wherein carbon (16) is a single or double bond, and carbon (23) is a single, double, or triple bond. Various alkyl or haloalkyl substitutions are incorporated at carbon (25). The invention provides pharmaceutically acceptable esters, salts, and prodrugs thereof. Methods for using the compounds to treat vitamin D₃ associated states, and pharmaceutical compositions containing the compounds are also disclosed.
1,3-DIACYLATED, 26,27-ALKYL/HALOALKYL VITAMIN D₃ COMPOUNDS
AND METHODS OF USE THEREOF

Related Applications

This application claims priority to: U.S. provisional application Ser. No. 60/505,735, filed 24 September 2003; GB0322395.5, filed 24 September 2003; and GB0404567.0, filed 01 March 2004. Each of the aforementioned applications is incorporated herein in its entirety by this reference.

Background of the Invention

The importance of vitamin D (cholecalciferol) in the biological systems of higher animals has been recognized since its discovery by Mellanby in 1920 (Mellanby, E. (1921) Spec. Rep. Ser. Med. Res. Council (GB) SRS 61:4). It was in the interval of 1920-1930 that vitamin D officially became classified as a "vitamin" that was essential for the normal development of the skeleton and maintenance of calcium and phosphorous homeostasis.


Vitamin D₃ and its hormonally active forms are well-known regulators of calcium and phosphorous homeostasis. These compounds are known to stimulate, at least one of, intestinal absorption of calcium and phosphate, mobilization of bone mineral, and retention of calcium in the kidneys. Furthermore, the discovery of the presence of specific vitamin D receptors in more than 30 tissues has led to the identification of vitamin D₃ as a pluripotent regulator outside its classical role in calcium/bone homeostasis. A paracrine role for 1α,25(OH)₂D₃ has been suggested by the combined presence of enzymes capable of oxidizing vitamin D₃ into its active forms, e.g., 25-OHD-1α-hydroxylase, and specific receptors in several tissues such as bone, keratinocytes, placenta, and immune cells. Moreover, vitamin D₃ hormone and active metabolites have been found to be capable of regulating cell proliferation and differentiation of both normal and malignant cells (Reichel, H. et al. (1989) *Annu. Rev. Med.* 40:71-78).

Given the activities of vitamin D₃ and its metabolites, much attention has focused on the development of synthetic analogs of these compounds. A large number of these analogs involve structural modifications in the A ring, B ring, C/D rings, and, primarily, the side chain (Bouillon, R. et al., *Endocrine Reviews* 16(2):201-204).

Although a vast majority of the vitamin D₃ analogs developed to date involve structural modifications in the side chain, a few studies have reported the biological profile of A-ring diastereomers (Norman, A.W. et al. *J. Biol. Chem.* 268 (27): 20022-20030). Furthermore, biological esterification of steroids has been studied (Hochberg, R.B.,

Moreover, despite much effort in developing synthetic analogs, clinical applications of vitamin D and its structural analogs have been limited by the undesired side effects elicited by these compounds after administration to a subject for known indications/applications of vitamin D compounds.

**Brief Description of the Drawings**

The present invention is further described below with reference to the following non-limiting examples and with reference to the following figures, in which:

Figure 1 shows Percent Type 1 Diabetes Mellitus incidence for compound (2).

Figure 2 shows NOD mouse body weight (g) at two doses of compound (2).

Figure 3 shows the presence of vitamin D receptors (VDRs) on bladder cells.

Figure 4 shows calcitriol (the activated form of vitamin D₃) as effective in inhibiting the basal growth of bladder cells.

Figure 5 shows the evaluation of the effect of Vitamin D₃ analogue (2) on bladder function in an *in vivo* model –cyclophosphamide (CYP) induced chronic IC in rats.

**Summary of the Invention**

In one aspect, the invention provides a vitamin D₃ compound of formula I:
wherein: $A_1$ is single or double bond; $A_2$ is a single, double or triple bond; $X_1$ and $X_2$ are each independently $H_2$ or $=CH_2$, provided $X_1$ and $X_2$ are not both $=CH_2$; $R_1$ and $R_2$ are each independently $OC(O)C_1-C_4$ alkyl, $OC(O)$hydroxyalkyl, or $OC(O)$haloalkyl; $R_3$, $R_4$ and $R_5$ are each independently hydrogen, $C_1-C_4$ alkyl, hydroxyalkyl, or haloalkyl, with the understanding that $R_5$ is absent when $A_2$ is a triple bond, or $R_3$ and $R_4$ taken together with $C_{20}$ form $C_3-C_6$ cycloalkyl; $R_6$ and $R_7$ are each independently alkyl or haloalkyl; and $R_8$ is $H$, $C(O)C_1-C_4$ alkyl, $C(O)$hydroxyalkyl, or $C(O)$haloalkyl; provided that when $A_1$ is a single bond, $R_3$ is hydrogen and $R_4$ is methyl, then $A_2$ is a double or triple bond; and pharmaceutically acceptable esters, salts, and prodrugs thereof.

In a preferred embodiment, the invention provides vitamin D$_3$ compounds of formula I-a:

![Chemical structure](image)

wherein (in formula I above, $R_3$ is $H$, $R_4$ is methyl, $A_1$ is a double bond,) $R_5$ is $H$ (or absent if $A_2$ is a triple bond), and $A_2$, $X_1$, $X_2$, $R_1$, $R_2$, $R_6$, $R_7$, and $R_8$ are previously described.
In another preferred embodiment, the invention provides vitamin D₃ compounds of formula I-b:

![Chemical Structure](image)

wherein (in formula I above, R₃ and R₄ taken together with C-20 form cyclopropyl), R₅ is H (or absent if A₂ is a triple bond), and A₁, A₂, X₁, X₂, R₁, R₂, R₆, R₇, and R₈ are previously described.

In yet another aspect, the invention provides a pharmaceutical composition. The composition comprises an effective amount of a vitamin D₃ compound of formula I, and a pharmaceutically acceptable carrier.

In a further aspect, the method provides a method of ameliorating a deregulation of calcium and phosphate metabolism. The method includes administering to a subject a therapeutically effective amount of a vitamin D₃ compound of formula I, so as to ameliorate the deregulation of the calcium and phosphate metabolism.

In another aspect, the invention provides a method of modulating the expression of an immunoglobulin-like transcript 3 (ILT3) surface molecule in a cell. The method includes contacting the cell with a vitamin D₃ compound of formula I in an amount effective to modulate the expression of an immunoglobulin-like transcript 3 (ILT3) surface molecule in the cell.

In yet another aspect, the invention provides a method of treating an ILT3-associated disorder in a subject. The method includes administering to the subject a vitamin D₃ compound of formula I in an amount effective to modulate the expression of an ILT3 surface molecule, thereby treating the ILT3-associated disorder in the subject.

In still another aspect, the invention provides a method of inducing immunological tolerance in a subject. The method includes administering to the subject
a vitamin D₃ compound of formula I in an amount effective to modulate the expression
of an ILT3 surface molecule, thereby inducing immunological tolerance in the subject.

In a further aspect, the invention provides a method of inhibiting transplant
rejection in a subject. The method includes administering to the subject a vitamin D₃
compound of formula I in an amount effective to modulate the expression of an ILT3
surface molecule, thereby inhibiting transplant rejection in the subject.

In still another embodiment, the invention provides a method for preventing or
treating bladder dysfunction in a subject in need thereof by administering an effective
amount of a vitamin D₃ compound thereby to prevent or treat bladder dysfunction in
said subject.

In yet another aspect, the invention provides a packaged formulation for use in
the treatment of a vitamin D₃ associated state. The packaged formulation includes a
pharmaceutical composition comprising a vitamin D₃ compound of formula I and a
pharmaceutically-acceptable carrier, packaged with instructions for use in the treatment
of a vitamin D₃ associated state.

In another aspect, the invention provides a packaged formulation for use in the
treatment of an ILT-3 associated disorder. The packed formulation includes a
pharmaceutical composition comprising a vitamin D₃ compound of formula I and a
pharmaceutically-acceptable carrier, packaged with instructions for use in the treatment
of an ILT3-associated disorder.

In a further aspect, the invention provides a method for modulating
immunosuppressive activity by an antigen-presenting cell. The method includes
contacting an antigen-presenting cell with a vitamin D₃ compound of formula I in an
amount effective to modulate ILT3 surface molecule expression, thereby modulating the
immunosuppressive activity by the antigen-presenting cell.

**Detailed Description of the Invention**

1. **DEFINITIONS**

Before further description of the present invention, and in order that the
invention may be more readily understood, certain terms are first defined and collected
here for convenience.
The term "administration" or "administering" includes routes of introducing the vitamin D₃ compound(s) to a subject to perform their intended function. Examples of routes of administration which can be used include injection (subcutaneous, intravenous, parenterally, intraperitoneally, intrathecal), oral, inhalation, rectal and transdermal. The pharmaceutical preparations are, of course, given by forms suitable for each administration route. For example, these preparations are administered in tablets or capsule form, by injection, inhalation, eye lotion, ointment, suppository, etc. administration by injection, infusion or inhalation; topical by lotion or ointment; and rectal by suppositories. Oral administration is preferred. The injection can be bolus or can be continuous infusion. Depending on the route of administration, the vitamin D₃ compound can be coated with or disposed in a selected material to protect it from natural conditions which may detrimentally effect its ability to perform its intended function. The vitamin D₃ compound can be administered alone, or in conjunction with either another agent as described above or with a pharmaceutically-acceptable carrier, or both. The vitamin D₃ compound can be administered prior to the administration of the other agent, simultaneously with the agent, or after the administration of the agent. Furthermore, the vitamin D₃ compound can also be administered in a proform which is converted into its active metabolite, or more active metabolite in vivo.

The term "alkyl" refers to the radical of saturated aliphatic groups, including straight-chain alkyl groups, branched-chain alkyl groups, cycloalkyl (ycliclic) groups, alkyl substituted cycloalkyl groups, and cycloalkyl substituted alkyl groups. The term alkyl further includes alkyl groups, which can further include oxygen, nitrogen, sulfur or phosphorous atoms replacing one or more carbons of the hydrocarbon backbone, e.g., oxygen, nitrogen, sulfur or phosphorous atoms. In preferred embodiments, a straight chain or branched chain alkyl has 30 or fewer carbon atoms in its backbone (e.g., C₁-C₃₀ for straight chain, C₃-C₃₀ for branched chain), preferably 26 or fewer, and more preferably 20 or fewer, and still more preferably 4 or fewer. Likewise, preferred cycloalkyls have from 3-10 carbon atoms in their ring structure, and more preferably have 3, 4, 5, 6 or 7 carbons in the ring structure.

Moreover, the term alkyl as used throughout the specification and claims is intended to include both "unsubstituted alkyls" and "substituted alkyls," the latter of which refers to alkyl moieties having substituents replacing a hydrogen on one or more carbons of the hydrocarbon backbone. Such substituents can include, for example,
halogen, hydroxyl, alkylcarbonyloxy, arylcarbonyloxy, alkoxy carbonyloxy, aryloxy carbonyloxy, carboxylate, alkylcarbonyl, alkoxy carbonyl, aminocarbonyl, alkylthiocarbonyl, alkoxy, phosphate, phosphonato, phosphinato, cyano, amino (including alkyl amino, dialkylamino, arylamino, diarylamino, and alkylarylamino), acylamino (including alkylcarbonylamino, arylcarbonylamino, carbamoyl and ureido), amidino, imino, sulfhydryl, alkylthio, arythio, thiocarboxylate, sulfates, sulfonato, sulfamoyl, sulfonamido, nitro, trifluoromethyl, cyano, azido, heterocyclyl, alkylaryl, or an aromatic or heteroaromatic moiety. It will be understood by those skilled in the art that the moieties substituted on the hydrocarbon chain can themselves be substituted, if appropriate. Cycloalkyls can be further substituted, e.g., with the substituents described above. An "alkylaryl" moiety is an alkyl substituted with an aryl (e.g., phenylmethyl (benzyl)). The term "alkyl" also includes unsaturated aliphatic groups analogous in length and possible substitution to the alkyls described above, but that contain at least one double or triple bond respectively.

Unless the number of carbons is otherwise specified, "lower alkyl" as used herein means an alkyl group, as defined above, but having from one to ten carbons, more preferably from one to six, and most preferably from one to four carbon atoms in its backbone structure, which may be straight or branched-chain. Examples of lower alkyl groups include methyl, ethyl, n-propyl, i-propyl, tert-butyl, hexyl, heptyl, octyl and so forth. In preferred embodiment, the term "lower alkyl" includes a straight chain alkyl having 4 or fewer carbon atoms in its backbone, e.g., C1-C4 alkyl.

The terms "alkoxyalkyl," "polyaminoalkyl" and "thioalkoxyalkyl" refer to alkyl groups, as described above, which further include oxygen, nitrogen or sulfur atoms replacing one or more carbons of the hydrocarbon backbone, e.g., oxygen, nitrogen or sulfur atoms.

The terms "alkenyl" and "alkynyl" refer to unsaturated aliphatic groups analogous in length and possible substitution to the alkyls described above, but that contain at least one double or triple bond, respectively. For example, the invention contemplates cyano and propargyl groups.

The term "antigen" includes a substance which elicits an immune response. The antigens of the invention to which tolerance is induced may or may not be exogenously derived relative to the host. For example, the method of the invention may be used to induce tolerance to an "autoantigen." An autoantigen is a normal constituent of the
body that reacts with an autoantibody. The invention also includes inducing tolerance to an "alloantigen." Alloantigen refers to an antigen found only in some members of a species, for example the blood group substances. An allograft is a graft to a genetically different member of the same species. Allografts are rejected by virtue of the immunological response of T lymphocytes to histocompatibility antigens. The method of the invention also provides for inducing tolerance to a "xenoantigen." Xenoantigens are substances that cause an immune reaction due to differences between different species. Thus, a xenograft is a graft from a member of one species to a member of a different species. Xenografts are usually rejected within a few days by antibodies and cytotoxic T lymphocytes to histocompatibility antigens.

The language "antigen-presenting cell" or "APC" includes a cell that is able to present an antigen to, for example, a T helper cell. Antigen-presenting cells include B lymphocytes, accessory cells or non-lymphocytic cells, such as dendritic cells, Langerhans cells, and mononuclear phagocytes that help in the induction of an immune response by presenting antigen to helper T lymphocytes. The antigen-presenting cell of the present invention is preferably of myeloid origin, and includes, but is not limited to, dendritic cells, macrophages, monocytes. APCs of the present invention may be isolated from the bone marrow, blood, thymus, epidermis, liver, fetal liver, or the spleen.

The terms "antineoplastic agent" and "antiproliferative agent" are used interchangeably herein and includes agents that have the functional property of inhibiting the proliferation of a vitamin D3-responsive cells, e.g., inhibit the development or progression of a neoplasm having such a characteristic, particularly a hematopoietic neoplasm.

The term "aryl" as used herein, refers to the radical of aryl groups, including 5- and 6-membered single-ring aromatic groups that may include from zero to four heteroatoms, for example, benzene, pyrrole, furan, thiophene, imidazole, benzoxazole, benzothiazole, triazole, tetrazole, pyrazole, pyridine, pyrazine, pyridazine and pyrimidine, and the like. Aryl groups also include polycyclic fused aromatic groups such as naphthyl, quinolyl, indolyl, and the like. Those aryl groups having heteroatoms in the ring structure may also be referred to as "aryl heterocycles," "heteroaaryls" or "heteroaromatics." The aromatic ring can be substituted at one or more ring positions with such substituents as described above, as for example, halogen, hydroxyl, alkoxy, alkylcarbonyloxy, arylcarbonyloxy, alkoxy carbonyloxy, aryloxycarbonyloxy,
carboxylate, alkylcarbonyl, alkoxy carbonyl, aminocarbonyl, alkylthiocarbonyl, phosphate, phosphonato, phosphinato, cyano, amino (including alkyl amino, dialkylamino, arylamino, diarylamino, and alkylaryl amino), acylamino (including alkylcarbonylamino, arylcarbonylamino, carbamoyl and ureido), amidino, imino, sulphydryl, alkythio, arylthio, thiocarboxylate, sulfates, sulfonato, sulfamoyl, sulfonamido, nitro, trifluoromethyl, cyano, azido, heterocyclyl, alkylaryl, or an aromatic or heteroaromatic moiety. Aryl groups can also be fused or bridged with alicyclic or heterocyclic rings which are not aromatic so as to form a polycycle (e.g., tetralin).

The language "autoimmune disease" or "autoimmune disorder" refers to the condition where the immune system attacks the host's own tissue(s). In an autoimmune disease, the immune tolerance system of the patient fails to recognize self antigens and, as a consequence of this loss of tolerance, brings the force of the immune system to bear on tissues which express the antigen. Autoimmune disorders include, but are not limited to, type 1 insulin-dependent diabetes mellitus, adult respiratory distress syndrome, inflammatory bowel disease, dermatitis, meningitis, thrombotic thrombocytopenic purpura, Sjogren's syndrome, encephalitis, uveitis, uveoretinitis, leukocyte adhesion deficiency, rheumatoid arthritis, rheumatic fever, Reiter's syndrome, psoriatic arthritis, progressive systemic sclerosis, primary biliary cirrhosis, pemphigus, pemphigoid, necrotizing vasculitis, myasthenia gravis, multiple sclerosis, lupus erythematosus, polymyositis, sarcoidosis, granulomatosis, vasculitis, pernicious anemia, CNS inflammatory disorder, antigen-antibody complex mediated diseases, autoimmune haemolytic anemia, Hashimoto's thyroiditis, Graves disease, habitual spontaneous abortions, Reynard's syndrome, glomerulonephritis, dermatomyositis, chronic active hepatitis, celiac disease, autoimmune complications of AIDS, atrophic gastritis, ankylosing spondylitis and Addison's disease.

By “bladder dysfunction” is meant bladder conditions associated with overactivity of the detrusor muscle, for example, clinical BPH or overactive bladder. In the context of the present invention “bladder dysfunction” excludes bladder cancer.

The language "bone metabolism" includes direct or indirect effects in the formation or degeneration of bone structures, e.g., bone formation, bone resorption, etc., which may ultimately affect the concentrations in serum of calcium and phosphate. This term is also intended to include effects of compounds of the invention in bone cells, e.g., osteoclasts and osteoblasts, that may in turn result in bone formation and degeneration.

The language "calcium and phosphate homeostasis" refers to the careful balance of calcium and phosphate concentrations, intracellularly and extracellularly, triggered by fluctuations in the calcium and phosphate concentration in a cell, a tissue, an organ or a system. Fluctuations in calcium levels that result from direct or indirect responses to compounds of the invention are intended to be included by these terms.

The term "carcinoma" is art recognized and refers to malignancies of epithelial or endocrine tissues including respiratory system carcinomas, gastrointestinal system carcinomas, genitourinary system carcinomas, testicular carcinomas, breast carcinomas, prostatic carcinomas, endocrine system carcinomas, and melanomas. Exemplary carcinomas include those forming from tissue of the cervix, lung, prostate, bladder, breast, head and neck, colon and ovary. The term also includes carcinosarcomas, e.g., which include malignant tumors composed of carcinomatous and sarcomatous tissues. An "adenocarcinoma" refers to a carcinoma derived from glandular tissue or in which the tumor cells form recognizable glandular structures.

The term "chiral" refers to molecules which have the property of non-superimposability of the mirror image partner, while the term "achiral" refers to molecules which are superimposable on their mirror image partner.

The term "diastereomers" refers to stereoisomers with two or more centers of dissymmetry and whose molecules are not mirror images of one another.

The term “effective amount” includes an amount effective, at dosages and for periods of time necessary, to achieve the desired result, e.g., sufficient treat a vitamin D₃ associated state or to modulate ILT3 expression in a cell. An effective amount of vitamin D₃ compound may vary according to factors such as the disease state, age, and weight of the subject, and the ability of the vitamin D₃ compound to elicit a desired response in the subject. Dosage regimens may be adjusted to provide the optimum
therapeutic response. An effective amount is also one in which any toxic or detrimental effects (e.g., side effects) of the vitamin D₃ compound are outweighed by the therapeutically beneficial effects.

A therapeutically effective amount of vitamin D₃ compound (i.e., an effective dosage) may range from about 0.001 to 30 μg/kg body weight, preferably about 0.01 to 25 μg/kg body weight, more preferably about 0.1 to 20 μg/kg body weight, and even more preferably about 1 to 10 μg/kg, 2 to 9 μg/kg, 3 to 8 μg/kg, 4 to 7 μg/kg, or 5 to 6 μg/kg body weight. The skilled artisan will appreciate that certain factors may influence the dosage required to effectively treat a subject, including but not limited to the severity of the disease or disorder, previous treatments, the general health and/or age of the subject, and other diseases present. Moreover, treatment of a subject with a therapeutically effective amount of a vitamin D₃ compound can include a single treatment or, preferably, can include a series of treatments. In one example, a subject is treated with a vitamin D₃ compound in the range of between about 0.1 to 20 μg/kg body weight, one time per week for between about 1 to 10 weeks, preferably between 2 to 8 weeks, more preferably between about 3 to 7 weeks, and even more preferably for about 4, 5, or 6 weeks. It will also be appreciated that the effective dosage of a vitamin D₃ compound used for treatment may increase or decrease over the course of a particular treatment.

The term "enantiomers" refers to two stereoisomers of a compound which are non-superimposable mirror images of one another. An equimolar mixture of two enantiomers is called a "racemic mixture" or a "racemate."

The language "genomic" activities or effects of vitamin D₃ is intended to include those activities mediated by the nuclear receptor for 1α, 25(OH)₂D₃ (VD₃R), e.g.,

transcriptional activation of target genes.

The term "haloalkyl" is intended to include alkyl groups as defined above that are mono-, di- or polysubstituted by halogen, e.g., fluoromethyl and trifluoromethyl.

The term "halogen" designates -F, -Cl, -Br or -I.

The term "hydroxyl" means -OH.

The term "heteroatom" as used herein means an atom of any element other than carbon or hydrogen. Preferred heteroatoms are nitrogen, oxygen, sulfur and phosphorus.

The term "homeostasis" is art-recognized to mean maintenance of static, or constant, conditions in an internal environment.
The language "hormone secretion" is art-recognized and includes activities of vitamin D₃ compounds that control the transcription and processing responsible for secretion of a given hormone e.g., a parathyroid hormone (PTH) of a vitamin D₃ responsive cell (Bouillon, R. et al. (1995) Endocrine Reviews 16(2):235-237).

The language "hypercalcemia" or "hypercalcemic activity" is intended to have its accepted clinical meaning, namely, increases in calcium serum levels that are manifested in a subject by the following side effects, depression of central and peripheral nervous system, muscular weakness, constipation, abdominal pain, lack of appetite and, depressed relaxation of the heart during diastole. Symptomatic manifestations of hypercalcemia are triggered by a stimulation of at least one of the following activities, intestinal calcium transport, bone calcium metabolism and osteocalcin synthesis (reviewed in Bouillon, R. et al. (1995) Endocrinology Reviews 16(2): 200-257).

The terms "hyperproliferative" and "neoplastic" are used interchangeably, and include those cells having the capacity for autonomous growth, i.e., an abnormal state or condition characterized by rapidly proliferating cell growth. Hyperproliferative and neoplastic disease states may be categorized as pathologic, i.e., characterizing or constituting a disease state, or may be categorized as non-pathologic, i.e., a deviation from normal but not associated with a disease state. The term is meant to include all types of cancerous growths or oncogenic processes, metastatic tissues or malignantly transformed cells, tissues, or organs, irrespective of histopathologic type or stage of invasiveness. "Pathologic hyperproliferative" cells occur in disease states characterized by malignant tumor growth. Examples of non-pathologic hyperproliferative cells include proliferation of cells associated with wound repair.

The language "immunoglobulin-like transcript 3" or "ILT3" refers to a cell surface molecule of the immunoglobulin superfamily, which is expressed by antigen-presenting cells (APCs) such as monocytes, macrophages and dendritic cells. ILT3 is a member of the immunoglobulin-like transcript (ILT) family and displays a long cytoplasmic tail containing putative immunoreceptor tyrosine-based inhibitory motifs (ITIMs). ILT3 has been shown to behave as an inhibitory receptor when cross-linked to a stimulatory receptor. A cytoplasmic component of the ILT3-mediated signaling pathway is the SH2-containing phosphatase SHP-1, which becomes associated with ILT3 upon cross-linking. ILT3 is also internalized and ILT3 ligands are efficiently presented to specific T cells (see, e.g., Cella, M. et al. (1997) J. Exp. Med. 185:1743).
The determination of whether the candidate vitamin D₃ compound modulates the expression of the ILT3 surface molecule can be accomplished, for example, by comparison of ILT3 surface molecule expression to a control, by measuring mRNA expression, or by measuring protein expression.

An "ILT3-associated disorder" includes a disease, disorder or condition which is associated with an ILT3 molecule. ILT3 associated disorders include disorders in which ILT3 activity is aberrant or in which a non-ILT3 activity that would benefit from modulation of an ILT3 activity is aberrant. In one embodiment, the ILT3-associated disorder is an immune disorder, e.g., an autoimmune disorder, such as type 1 insulin-dependent diabetes mellitus, adult respiratory distress syndrome, inflammatory bowel disease, dermatitis, meningitis, thrombotic thrombocytopenic purpura, Sjogren's syndrome, encephalitis, uveitis, uveoretinitis, leukocyte adhesion deficiency, rheumatoid arthritis, rheumatic fever, Reiter's syndrome, psoriatic arthritis, progressive systemic sclerosis, primary biliary cirrhosis, pemphigus, pemphigoid, necrotizing vasculitis, myasthenia gravis, multiple sclerosis, lupus erythematosus, polymyositis, sarcoidosis, granulomatosis, vasculitis, pernicious anemia, CNS inflammatory disorder, antigen-antibody complex mediated diseases, autoimmune haemolytic anemia, Hashimoto's thyroiditis, Graves disease, habitual spontaneous abortions, Reynard's syndrome, glomerulonephritis, dermatomyositis, chronic active hepatitis, celiac disease, autoimmune complications of AIDS, atrophic gastritis, ankylosing spondylitis and Addison's disease; or transplant rejection, such as GVHD. In certain embodiments of the invention, the ILT3 associated disorder is an immune disorders, such as transplant rejections, graft versus host disease and autoimmune disorders.

The term "immune response" includes T and/or B cell responses, e.g., cellular and/or humoral immune responses. The claimed methods can be used to reduce both primary and secondary immune responses. The immune response of a subject can be determined by, for example, assaying antibody production, immune cell proliferation, the release of cytokines, the expression of cell surface markers, cytotoxicity, and the like.

The terms "immunological tolerance" or "tolerance to an antigen" or "immune tolerance" include unresponsiveness to an antigen without the induction of a prolonged generalized immune deficiency. Consequently, according to the invention, a tolerant host is capable of reacting to antigens other than the tolerizing antigen. Tolerance
represents an induced depression in the response of a subject that, had it not been subjected to the tolerance-inducing procedure, would be competent to mount an immune response to that antigen. In one embodiment of the invention, immunological tolerance is induced in an antigen-presenting cell, e.g., an antigen-presenting cell derived from the myeloid or lymphoid lineage, dendritic cells, monocytes and macrophages.

The language “immunosuppressive activity” refers to the process of inhibiting a normal immune response. Included in this response is when T and/or B clones of lymphocytes are depleted in size or suppressed in their reactivity, expansion or differentiation. Immunosuppressive activity may be in the form of inhibiting or blocking an immune response already in progress or may involve preventing the induction of an immune response. The functions of activated T cells may be inhibited by suppressing immune cell responses or by inducing specific tolerance, or both. Immunosuppression of T cell responses is generally an active, non-antigen-specific, process that requires continuous exposure of the T cells to the suppressive agent.

Tolerance, which involves inducing non-responsiveness or anergy in T cells, is distinguishable from immunosuppression in that it is generally antigen-specific and persists after exposure to the tolerizing agent has ceased. Operationally, tolerance can be demonstrated by the lack of a T cell response upon re-exposure to specific antigen in the absence of the tolerizing agent.

The language “improved biological properties” refers to any activity inherent in a compound of the invention that enhances its effectiveness in vivo. In a preferred embodiment, this term refers to any qualitative or quantitative improved therapeutic property of a vitamin D₃ compound, such as reduced toxicity, e.g., reduced hypercalcemic activity.

The language “inhibiting the growth” of the neoplasm includes the slowing, interrupting, arresting or stopping its growth and metastases and does not necessarily indicate a total elimination of the neoplastic growth.

The term "isomers" or "stereoisomers" refers to compounds which have identical chemical constitution, but differ with regard to the arrangement of the atoms or groups in space.

The term "leukemia" is intended to have its clinical meaning, namely, a neoplastic disease in which white corpuscle maturation is arrested at a primitive stage of cell development. The disease is characterized by an increased number of leukemic blast cells in the bone marrow, and by varying degrees of failure to produce normal hematopoietic cells. The condition may be either acute or chronic. Leukemia's are further typically categorized as being either lymphocytic i.e., being characterized by cells which have properties in common with normal lymphocytes, or myelocytic (or myelogenous), i.e., characterized by cells having some characteristics of normal granulocytic cells. Acute lymphocytic leukemia ("ALL") arises in lymphoid tissue, and ordinarily first manifests its presence in bone marrow. Acute myelocytic leukemia ("AML") arises from bone marrow hematopoietic stem cells or their progeny. The term acute myelocytic leukemia subsumes several subtypes of leukemia: myeloblastic leukemia, promyelocytic leukemia, and myelomonocytic leukemia. In addition, leukemias with erythroid or megakaryocytic properties are considered myelogenous leukemias as well.

The term "leukemic cancer" refers to all cancers or neoplasias of the hemopoietic and immune systems (blood and lymphatic system). The acute and chronic leukemias, together with the other types of tumors of the blood, bone marrow cells (myelomas), and lymph tissue (lymphomas), cause about 10% of all cancer deaths and about 50% of all cancer deaths in children and adults less than 30 years old. Chronic myelogenous leukemia (CML), also known as chronic granulocytic leukemia (CGL), is a neoplastic disorder of the hematopoietic stem cell. The term "leukemia" is art recognized and refers to a progressive, malignant disease of the blood-forming organs, marked by distorted proliferation and development of leukocytes and their precursors in the blood and bone marrow.

The term "modulate" refers to increases or decreases in the activity of a cell in response to exposure to a compound of the invention, e.g., the inhibition of proliferation and/or induction of differentiation of at least a sub-population of cells in an animal such that a desired end result is achieved, e.g., a therapeutic result. In preferred
embodiments, this phrase is intended to include hyperactive conditions that result in pathological disorders.

The common medical meaning of the term "neoplasia" refers to "new cell growth" that results as a loss of responsiveness to normal growth controls, e.g. to neoplastic cell growth. A "hyperplasia" refers to cells undergoing an abnormally high rate of growth. However, as used herein, the terms neoplasia and hyperplasia can be used interchangeably, as their context will reveal, referring to generally to cells experiencing abnormal cell growth rates. Neoplasias and hyperplasias include "tumors," which may be either benign, premalignant or malignant.


The phrases "parenteral administration" and "administered parenterally" as used herein means modes of administration other than enteral and topical administration, usually by injection, and includes, without limitation, intravenous, intramuscular, intraarterial, intrathecal, intracapsular, intraorbital, intracardiac, intradermal, intraperitoneal, transtracheal, subcutaneous, subcuticular, intracuticular, subcapsular, subarachnoid, intraspinal and intrasternal injection and infusion.
The terms "polycyclol" or "polycyclic radical" refer to the radical of two or more
cyclic rings (e.g., cycloalkyls, cycloalkenyls, cycloalkynyls, aryls and/or heterocycles) in
which two or more carbons are common to two adjoining rings, e.g., the rings are
"fused rings". Rings that are joined through non-adjacent atoms are termed "bridged"
rings. Each of the rings of the polycycle can be substituted with such substituents as
described above, as for example, halogen, hydroxyl, alkylcarbonyloxy, arylcarbonyloxy,
alkoxycarbonyloxy, arylxocarbonyloxy, carboxylate, alkylcarbonyl, alkoxyxycarbonyl,
aminocarbonyl, alkylthiocarbonyl, alkoxyl, phosphato, phosphonato, phosphinato,
cyano, amino (including alkyl amino, dialkylaminio, arylamino, diarylamino, and
alkylarylamino), acylamino (including alkylcarbonylamino, arylcarbonylamino,
carbamoyl and ureido), amidino, imino, sulfhydryl, alkylthio, arylthio, thiocarboxylate,
sulfates, sulfonato, sulfamoyl, sulfonamido, nitro, trifluoromethyl, cyano, azido,
heterocyclyl, alkyl, alkylaryl, or an aromatic or heteroaromatic moiety.

The term “prodrug” includes compounds with moieties which can be
metabolized in vivo. Generally, the prodrugs are metabolized in vivo by esterases or by
other mechanisms to active drugs. Examples of prodrugs and their uses are well known
The prodrugs can be prepared in situ during the final isolation and purification of the
compounds, or by separately reacting the purified compound in its free acid form or
hydroxyl with a suitable esterifying agent. Hydroxyl groups can be converted into esters
via treatment with a carboxylic acid. Examples of prodrug moieties include substituted
and unsubstituted, branch or unbranched lower alkyl ester moieties, (e.g., propionic
acid esters), lower alkenyl esters, di-lower alkyl-amino lower-alkyl esters (e.g.,
dimethylaminoethyl ester), acylamino lower alkyl esters (e.g., acetylloxymethyl ester),
acyloxy lower alkyl esters (e.g., pivaloyloxymethyl ester), ary1 esters (phenyl ester),
aryl-lower alkyl esters (e.g., benzyl ester), substituted (e.g., with methyl, halo, or
methoxy substituents) aryl and aryl-lower alkyl esters, amides, lower-alkyl amides, di-
lower alkyl amides, and hydroxy amides. Preferred prodrug moieties are propionic
acid esters and acyl esters. Prodrugs which are converted to active forms through other
mechanisms in vivo are also included.

The language "a prophylactically effective anti-neoplastic amount" of a
compound refers to an amount of a vitamin D₃ compound of the formula (I) or otherwise
described herein which is effective, upon single or multiple dose administration to the
patient, in preventing or delaying the occurrence of the onset of a neoplastic disease state.

The term "psoriasis" is intended to have its medical meaning, namely, a disease which afflicts primarily the skin and produces raised, thickened, scaling, nonscarring lesions. The lesions are usually sharply demarcated erythematous papules covered with overlapping shiny scales. The scales are typically silvery or slightly opalescent. Involvement of the nails frequently occurs resulting in pitting, separation of the nail, thickening and discoloration. Psoriasis is sometimes associated with arthritis, and it may be crippling.

The language "reduced toxicity" is intended to include a reduction in any undesired side effect elicited by a vitamin D$_3$ compound when administered in vivo, e.g., a reduction in the hypercalcemic activity.

The term "sarcoma" is art recognized and refers to malignant tumors of mesenchymal derivation.

The term "secosteroid" is art-recognized and includes compounds in which one of the cyclopentanoperhydro-phenanthrene rings of the steroid ring structure is broken. 1α,25(OH)$_2$D$_3$ and analogs thereof are hormonally active secosteroids. In the case of vitamin D$_3$, the 9-10 carbon-carbon bond of the B-ring is broken, generating a seco-B-steroid. The official IUPAC name for vitamin D$_3$ is 9,10-secocholesta-5,7,10(19)-trien-3B-ol. For convenience, a 6-s-trans conformer of 1α,25(OH)$_2$D$_3$ is illustrated herein having all carbon atoms numbered using standard steroid notation.
In the formulas presented herein, the various substituents on ring A are illustrated as joined to the steroid nucleus by one of these notations: a dotted line (---) or (\ldots) indicating a substituent which is in the β-orientation (i.e., above the plane of the ring), a wedged solid line (\(\rightleftharpoons\)) indicating a substituent which is in the α-orientation (i.e., below the plane of the molecule), or a wavy line (\(\sim\)) indicating that a substituent may be either above or below the plane of the ring. In regard to ring A, it should be understood that the stereochemical convention in the vitamin D field is opposite from the general chemical field, wherein a dotted line indicates a substituent on Ring A which is in an α-orientation (i.e., below the plane of the molecule), and a wedged solid line indicates a substituent on ring A which is in the β-orientation (i.e., above the plane of the ring). As shown, the A ring of the hormone 1α,25(OH)\(_2\)D\(_3\) contains two asymmetric centers at carbons 1 and 3, each one containing a hydroxyl group in well-characterized configurations, namely the 1α- and 3β-hydroxyl groups. In other words, carbons 1 and 3 of the A ring are said to be "chiral carbons" or "carbon centers."

Furthermore the indication of stereochemistry across a carbon-carbon double bond is also opposite from the general chemical field in that “Z” refers to what is often referred to as a “cis” (same side) conformation whereas “E” refers to what is often referred to as a “trans” (opposite side) conformation. Regardless, both configurations, cis/trans and/or Z/E are contemplated for the compounds for use in the present invention.

Also, throughout the patent literature, the A ring of a vitamin D compound is often depicted in generic formulae as any one of the following structures:

![Chemical Structure](image)

wherein \(X_1\) and \(X_2\) are defined as H (or H\(_2\)) or \(=\text{CH}_2\); or
wherein $X_1$ and $X_2$ are defined as $H_2$ or $CH_2$. Although there does not appear to be any set convention, it is clear that one of ordinary skill in the art understands either formula I or II to represent an A ring in which, for example, $X_1$ is $=CH_2$ and $X_2$ is defined as $H_2$, as follows:

For purposes of the instant invention, formula I will be used in all generic structures.

The term "sulfhydryl" or "thiol" means $-SH$.

The term "subject" includes organisms which are capable of suffering from a vitamin $D_3$ associated state or who could otherwise benefit from the administration of a vitamin $D_3$ compound of the invention, such as human and non-human animals. Preferred human animals include human patients suffering from or prone to suffering from a vitamin $D_3$ associated state, as described herein. The term "non-human animals" of the invention includes all vertebrates, e.g., mammals, e.g., rodents, e.g., mice, and non-mammals, such as non-human primates, sheep, dog, cow, chickens, amphibians, reptiles, etc.

The phrases "systemic administration," "administered systemically", "peripheral administration" and "administered peripherally" as used herein mean the administration
of a vitamin D₃ compound(s), drug or other material, such that it enters the patient's system and, thus, is subject to metabolism and other like processes, for example, subcutaneous administration.

The language "therapeutically effective anti-neoplastic amount" of a vitamin D₃ compound of the invention refers to an amount of an agent which is effective, upon single or multiple dose administration to the patient, in inhibiting the growth of a neoplastic vitamin D₃-responsive cells, or in prolonging the survivability of the patient with such neoplastic cells beyond that expected in the absence of such treatment.

The language "transplant rejection" refers to an immune reaction directed against a transplanted organ(s) from other human donors (allografts) or from other species such as sheep, pigs, or non-human primates (xenografts). Therefore, the method of the invention is useful for preventing an immune reaction to transplanted organs from other human donors (allografts) or from other species (xenografts). Such tissues for transplantation include, but are not limited to, heart, liver, kidney, lung, pancreas, pancreatic islets, bone marrow, brain tissue, cornea, bone, intestine, skin, and hematopoietic cells. Also included within this definition is "graft versus host disease" of "GVHD," which is a condition where the graft cells mount an immune response against the host. Therefore, the method of the invention is useful in preventing graft versus host disease in cases of mismatched bone marrow or lymphoid tissue transplanted for the treatment of acute leukemia, aplastic anemia, and enzyme or immune deficiencies, for example. The term "transplant rejection" also includes disease symptoms characterized by loss of organ function. For example, kidney rejection would be characterized by a rising creatinine level in blood. Heart rejection is characterized by an endomyocardial biopsy, while pancreas rejection is characterized by rising blood glucose levels. Liver rejection is characterized by the levels of transaminases of liver origin and bilirubin levels in blood. Intestine rejection is determined by biopsy, while lung rejection is determined by measurement of blood oxygenation.

The term "VDR" is intended to include members of the type II class of steroid/thyroid superfamily of receptors (Stunnenberg, H.G. (1993) Bio Essays 15(5):309-15), which are able to bind and transactivate through the vitamin D response element (VDRE) in the absence of a ligand (Damm et al. (1989) Nature 339:593-97; Sap et al. Nature 343:177-180).
The term "VDRE" refers to DNA sequences composed of half-sites arranged as direct repeats. It is known in the art that type II receptors do not bind to their respective binding site as homodimers but require an auxiliary factor, RXR (e.g. RXRα, RXRβ, RXRγ) for high affinity binding Yu et al. (1991) Cell 67:1251-1266; Bugge et al. (1992) EMBO J. 11:1409-1418; Kliewer et al. (1992) Nature 355:446-449; Leid et al. (1992) EMBO J. 11:1419-1435; Zhang et al. (1992) Nature 355:441-446.

The language “vitamin D₃ associated state” is a state which can be prevented, treated or otherwise ameliorated by administration of one or more compounds of the invention. Vitamin D₃ associated states include ILT3-associated disorders, disorders characterized by an aberrant activity of a vitamin D₃-responsive cell, disorders characterized by a deregulation of calcium and phosphate metabolism, and other disorders or states described herein.

The term "vitamin D₃-responsive cell" includes any cell which is is capable of responding to a vitamin D₃ compound having the formula I or otherwise described herein, or is associated with disorders involving an aberrant activity of hyperproliferative skin cells, parathyroid cells, neoplastic cells, immune cells, and bone cells. These cells can respond to vitamin D₃ activation by triggering genomic and/or non-genomic responses that ultimately result in the modulation of cell proliferation, differentiation survival, and/or other cellular activities such as hormone secretion. In a preferred embodiment, the ultimate responses of a cell are inhibition of cell proliferation and/or induction of differentiation-specific genes. Exemplary vitamin D₃ responsive cells include immune cells, bone cells, neuronal cells, endocrine cells, neoplastic cells, epidermal cells, endodermal cells, smooth muscle cells, among others.

With respect to the nomenclature of a chiral center, terms "d" and "l" configuration are as defined by the IUPAC Recommendations. As to the use of the terms, diastereomer, racemate, epimer and enantiomer will be used in their normal context to describe the stereochemistry of preparations.

2. VITAMIN D₃ COMPOUNDS OF THE INVENTION

A prominent feature of the vitamin D₃ compounds of the invention is acylation at the 1 and 3 positions on the A ring of the compounds. 1,3-diacyl vitamin D₃ compounds are described in U.S. Patent 5,976,784 to DeLuca et al. However, any compounds
specifically disclosed in U.S. Patent 5,976,784 to DeLuca et al. are excluded from the scope the appended claims.

The acylated vitamin D₃ compounds of formula I above exert a full spectrum of 1,25(OH)₂D₃ biological activities such as binding to the specific nuclear receptor VDR, suppression of the increased parathyroid hormone levels in 5,6-nephrectomized rats, suppression of INF-γ release in MLR cells, stimulation of HL-60 leukemia cell differentiation and inhibition of solid tumor cell proliferation. It is well known that in vivo and in cellular cultures 1,25-(OH)₂D₃ undergoes a cascade of metabolic modifications initiated by the influence of 24R-hydroxylase enzyme. First 24R-hydroxy metabolite is formed, which is oxidized to 24-keto intermediate, and then 23S-hydroxylation and fragmentation produce the fully inactive calcitriolic acid.

It has been discovered that 1,3-diacylated compounds of the invention have unexpected and/or superior properties as compared to corresponding 1,3-dihydroxy compounds. For example, 1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (2), 1,3-Di-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (4) and 1,3,25-Tri-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (5) have a significantly higher maximum tolerated dose and improved activity when compared with the corresponding dihydroxy compounds, 1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (1) and 1,25-dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (3).

Thus, in one aspect, the invention provides a vitamin D₃ compound of formula I:
wherein:

\[ A_1 \text{ is single or double bond;} \]
\[ A_2 \text{ is a single, double or triple bond;} \]
\[ X_1 \text{ and } X_2 \text{ are each independently } H_2 \text{ or } =CH_2, \text{ provided } X_1 \text{ and } X_2 \text{ are not both } =CH_2; \]
\[ R_1 \text{ and } R_2 \text{ are each independently } OC(O)C_1-C_4 \text{ alkyl, } OC(O)\text{hydroxyalkyl, or } \]
\[ OC(O)\text{haloalkyl;} \]
\[ R_3, R_4 \text{ and } R_5 \text{ are each independently hydrogen, } C_1-C_4 \text{ alkyl, hydroxyalkyl, or } \]
\[ \text{haloalkyl, with the understanding that } R_5 \text{ is absent when } A_2 \text{ is a triple bond, or } R_3 \text{ and } R_4 \]
\[ \text{taken together with } C_{20} \text{ form } C_3-C_6 \text{ cycloalkyl;} \]
\[ R_6 \text{ and } R_7 \text{ are each independently alkyl or haloalkyl; and } \]
\[ R_8 \text{ is } H, C(O)C_1-C_4 \text{ alkyl, } C(O)\text{hydroxyalkyl, or } C(O)\text{haloalkyl;} \]
\[ \text{provided that when } A_1 \text{ is single bond, } R_3 \text{ is hydrogen and } R_4 \text{ is methyl, then } A_2 \]
\[ \text{is a double or triple bond; and} \]
\[ \text{pharmacutically acceptable esters, salts, and prodrugs thereof.} \]

In one embodiment of the invention, \( X_1 \) is \( H_2 \) and \( X_2 \) is \( =CH_2 \). In another embodiment, \( X_1 \) and \( X_2 \) are \( H_2 \). In another embodiment, \( A_1 \) is a single bond. In another embodiment, \( A_1 \) is a double bond. In another embodiment, \( A_1 \) is a triple bond.

In a preferred embodiment, \( R_3 \) is hydrogen and \( R_4 \) is \( C_1-C_4 \) alkyl, preferably methyl. In another preferred embodiment, \( R_3 \) and \( R_4 \), taken together with \( C_{20} \), form \( C_3-C_6 \) cycloalkyl. In a preferred embodiment, \( R_3 \) and \( R_4 \), taken together with \( C_{20} \), form cyclopropyl.

In a preferred embodiment, \( R_1 \) and \( R_2 \) are each independently \( OC(O)C_1-C_4 \) alkyl, preferably \( OC(O)CH_3 \).

In a preferred embodiment, \( R_6 \) and \( R_7 \) are each independently alkyl or haloalkyl, preferably methyl, ethyl, or trifluoromethyl.

In a preferred embodiment, \( R_8 \) is \( H \) or \( C(O)C_1-C_4 \) alkyl.
Certain embodiments of the invention are directed to 1,3-acylated, 26,27-haloalkyl vitamin D₃ compounds. Such compounds are represented by the formula I-c:

\[
\begin{array}{c}
\text{I-c}
\end{array}
\]

wherein:

- \( A_1 \) is single or double bond;
- \( A_2 \) is a single, double or triple bond,
- \( X_1 \) and \( X_2 \) are each independently \( \text{H}_2 \) or \( \text{CH}_2 \), provided \( X_1 \) and \( X_2 \) are not both \( \text{CH}_2 \);
- \( R_1 \) and \( R_2 \) are each independently \( \text{OC(O)C}_1-\text{C}_4 \) alkyl, \( \text{OC(O)hydroxyalkyl} \), or \( \text{OC(O)haloalkyl} \);
- \( R_3 \), \( R_4 \) and \( R_5 \) are each independently hydrogen, \( \text{C}_1-\text{C}_4 \) alkyl, \( \text{hydroxyalkyl} \), or \( \text{haloalkyl} \), or \( R_3 \) and \( R_4 \) taken together with \( \text{C}_20 \) form \( \text{C}_3-\text{C}_6 \) cycloalkyl;
- \( R_6 \) and \( R_7 \) are each independently haloalkyl; and
- \( R_8 \) is \( \text{H}, \text{OC(O)C}_1-\text{C}_4 \) alkyl, \( \text{OC(O)hydroxyalkyl} \), or \( \text{OC(O)haloalkyl} \); and pharmaceutically acceptable esters, salts, and prodrugs thereof. In preferred embodiments, \( R_6 \) and \( R_7 \) are each independently trihaloalkyl, especially trifluoromethyl.

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In another embodiment of the invention, R₁ and R₂ are OC(O)CH₃, R₃ is H, R₄ is methyl, R₅ is H (or absent if A₂ is a triple bond), as shown in formula I-a.

\[
\begin{align*}
\text{H}_3\text{C(O)CO}^\text{-} & \text{OC(O)CH}_3 \\
A_1 & \quad A_2 \\
X_1 & \quad X_2 \\
R_6 & \quad R_7 \\
R_8 & \quad R_9
\end{align*}
\]

In a preferred embodiment, A₁ is a double bond, and X₁ is =CH₂ and X₂ is H₂.

When A₂ is a triple bond, it is preferred that R₈ is H or C(O)CH₃, and R₆ and R₇ are alkyl or haloalkyl. It is preferred that the alkyl group is methyl and the haloalkyl group is trifluoroalkyl, preferably trifluoromethyl. When A₂ is a double bond, it is preferred that R₈ is H, or C(O)CH₃, and R₆ and R₇ are alkyl, preferably methyl. It is also preferred that R₆ and R₇ are independently alkyl and haloalkyl. When A₂ is a single bond, it is preferred that R₈ is H or C(O)CH₃, and R₆ and R₇ are alkyl, preferably methyl.

In a preferred embodiment, A₁ is a double bond, and X₁ and X₂ are each H₂.

When A₂ is a triple bond, it is preferred that R₈ is H or C(O)CH₃, and R₆ and R₇ are alkyl or haloalkyl. It is preferred that the alkyl group is methyl or ethyl and the haloalkyl group is trifluoroalkyl, preferably trifluoromethyl. When A₂ is a double bond, it is preferred that R₈ is H or C(O)CH₃, and R₆ and R₇ are haloalkyl, preferably trifluoroalkyl, preferably trifluoromethyl. When A₂ is a single bond, it is preferred that R₈ is H or C(O)CH₃, and R₆ and R₇ are alkyl, preferably methyl.

In another embodiment of the invention of formula (I), R₁ and R₂ are OC(O)CH₃, A₁ is a single bond, and A₂ is a single, double or triple bond, except that when R₃ is H and R₄ is methyl, A₂ is a double or triple bond. In a preferred embodiment, R₃ is H, R₄ is methyl, R₅ is absent, R₆ is H or C(O)CH₃, and R₆ and R₇ are alkyl, preferably methyl.

Preferred compounds of the present invention are summarized in Table 1 and include the following: 1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (2), 1,3-Di-O-acetyl-1,25-Dihydroxy-16-ene-23-yn-26,27-hexafluoro-19-nor-cholecalciferol (4), 1,3,25-Tri-O-acetyl-1,25-Dihydroxy-16-ene-23,27-yn-26,27-hexafluoro-19-nor-cholecalciferol (5), 1,3-Di-O-acetyl-1,25-dihydroxy-16-
ene-23-ynocholecaciferol (7), 1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-
cholecaciferol (9), 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-cholecaciferol (11), 1,3,25-
Tri-O-acetyl-1,25-dihydroxy-16-ene-23-yno-26,27-hexafluoro-cholecaciferol (13), 1,3-
Di-O-acetyl-1,25-dihydroxy-16-ene-23-yno-26,27-hexafluoro-cholecaciferol (14), 1,3-
Di-O-acetyl-1,25-dihydroxy-16,23E-diene-25R,26-trifluoro-cholecaciferol (16), 1,3-Di-
O-acetyl-1,25-dihydroxy-16-ene-19-nor-cholecaciferol (18), 1,3-Di-O-Acetyl-1,25-
dihydroxy-16-ene-23-yno-19-nor-cholecaciferol (20), 1,3-Di-O-acetyl-1,25-dihydroxy-
16-ene-23-yno-26,27-bishomo-19-nor-cholecaciferol (22) and 1,3-Di-O-acetyl-1,25-
dihydroxy-23-yno-cholecaciferol (41).

### Table 1

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<tr>
<th>Compound</th>
<th>X₁</th>
<th>X₂</th>
<th>A₁</th>
<th>A₂</th>
<th>R₆</th>
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<tr>
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<td>⊙</td>
<td>CH₃</td>
<td>CH₃</td>
<td>H</td>
</tr>
</tbody>
</table>

* Z olefin
In another embodiment of the invention, R₁ and R₂ are each OC(O)CH₃, and R₃ and R₄ taken together with C₂₀ form cyclopropyl, and R₅ is H (or absent if A₂ is a triple bond), as shown in formula I-b.

In a preferred embodiment, X₁ is =CH₂ and X₂ is H₂. When A₁ is a single bond, and A₂ is a triple bond, it is preferred that R₈ is H or C(O)CH₃, and R₆ and R₇ are alkyl, preferably methyl. When A₁ is a single bond, and A₂ is a single bond, it is preferred that R₈ is H or C(O)CH₃, and R₆ and R₇ are alkyl, preferably methyl. When A₁ is a double bond, and A₂ is a single bond, it is preferable that R₈ is H or C(O)CH₃, and R₆ and R₇ are alkyl, preferably methyl.

In another preferred embodiment, X₁ and X₂ are each H₂. When A₁ is a single bond, and A₂ is a triple bond, it is preferred that R₈ is H or C(O)CH₃, and R₆ and R₇ are alkyl or haloalkyl. It is preferred that the alkyl group is methyl, and the haloalkyl group is trifluoroalkyl, preferably trifluoromethyl. When A₁ is a single bond, and A₂ is a double bond, it is preferred that R₈ is H or C(O)CH₃, R₆ and R₇ are haloalkyl, preferably trifluoroalkyl, preferably trifluoromethyl. When A₁ is a double bond, and A₂ is a single bond, it is preferred that R₈ is H or C(O)CH₃, R₆ and R₇ are alkyl, preferably methyl.

Preferred compounds of the present invention are summarized in Table 2 and include the following: 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-19-norcholecalciferol (24), 1,3,25-Tri-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-norcholecalciferol (26), 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-norcholecalciferol (28), 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-cholecalciferol (29), 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23E-ene-26,27-hexafluoro-19-norcholecalciferol (31), 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23Z-ene-26,27-hexafluoro-19-nor-
cholecalciferol (33), 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-cholecalciferol (35), 1,3-Di-O-acetyl-1α,25-dihydroxy-16-ene-20-cyclopropyl-19-nor-cholecalciferol (37), and 1,3-Di-O-acetyl-1α,25-hydroxy-16-ene-20-cyclopropyl-cholecalciferol (39).

Table 2

<table>
<thead>
<tr>
<th>Compound</th>
<th>X₁</th>
<th>X₂</th>
<th>A₁</th>
<th>A₂</th>
<th>R₆</th>
<th>R₇</th>
<th>R₈</th>
</tr>
</thead>
<tbody>
<tr>
<td>(24)</td>
<td>H₂</td>
<td>H₂</td>
<td></td>
<td></td>
<td>CH₃</td>
<td>CH₃</td>
<td>H</td>
</tr>
<tr>
<td>(27)</td>
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<td>H₂</td>
<td></td>
<td></td>
<td>CF₃</td>
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<td>H</td>
</tr>
<tr>
<td>(26)</td>
<td>H₂</td>
<td>H₂</td>
<td></td>
<td></td>
<td>CF₃</td>
<td>CF₃</td>
<td>C(O)CH₃</td>
</tr>
<tr>
<td>(29)</td>
<td>=CH₂</td>
<td>H₂</td>
<td></td>
<td></td>
<td>CH₃</td>
<td>CH₃</td>
<td>H</td>
</tr>
<tr>
<td>(31)</td>
<td>H₂</td>
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<td></td>
<td></td>
<td>CF₃</td>
<td>CF₃</td>
<td>H</td>
</tr>
<tr>
<td>(33)ᵃ</td>
<td>H₂</td>
<td>H₂</td>
<td></td>
<td></td>
<td>CF₃</td>
<td>CF₃</td>
<td>H</td>
</tr>
<tr>
<td>(35)</td>
<td>=CH₂</td>
<td>H₂</td>
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<td></td>
<td>CH₃</td>
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<td>H</td>
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<tr>
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<td>H</td>
</tr>
<tr>
<td>(39)</td>
<td>=CH₂</td>
<td>H₂</td>
<td></td>
<td></td>
<td>CH₃</td>
<td>CH₃</td>
<td>H</td>
</tr>
</tbody>
</table>

ᵃ Z olefin.

The structures of some of the compounds of the invention include asymmetric carbon atoms. Accordingly, the isomers arising from such asymmetry (e.g., all enantiomers and diastereomers) are included within the scope of the invention, unless indicated otherwise. Such isomers can be obtained in substantially pure form by classical separation techniques and/or by stereochemically controlled synthesis.

Naturally occurring or synthetic isomers can be separated in several ways known in the art. Methods for separating a racemic mixture of two enantiomers include
chromatography using a chiral stationary phase (see, e.g., "Chiral Liquid Chromatography," W.J. Lough, Ed. Chapman and Hall, New York (1989)). Enantiomers can also be separated by classical resolution techniques. For example, formation of diastereomeric salts and fractional crystallization can be used to separate enantiomers. For the separation of enantiomers of carboxylic acids, the diastereomeric salts can be formed by addition of enantiomerically pure chiral bases such as brucine, quinine, ephedrine, strychnine, and the like. Alternatively, diastereomeric esters can be formed with enantiomerically pure chiral alcohols such as menthol, followed by separation of the diastereomeric esters and hydrolysis to yield the free, enantiomerically enriched carboxylic acid. For separation of the optical isomers of amino compounds, addition of chiral carboxylic or sulfonic acids, such as camphorsulfonic acid, tartaric acid, mandelic acid, or lactic acid can result in formation of the diastereomeric salts.

3. USES OF THE VITAMIN D₃ COMPOUNDS OF THE INVENTION

In another embodiment, the invention also provides methods for treating a subject for a vitamin D₃ associated state, by administering to the subject an effective amount of a vitamin D₃ compound of formula I or otherwise described herein. Vitamin D₃ associated states include disorders involving an aberrant activity of a vitamin D₃-responsive cell, e.g., neoplastic cells, hyperproliferative skin cells, parathyroid cells, immune cells and bone cells, among others. Vitamin D₃ associated states also include ILT3-associated disorders. In certain embodiments, the subject is a mammal, e.g., a primate, e.g., a human.

In certain embodiments, the methods of the invention include administering to a subject a therapeutically effective amount of a vitamin D₃ compound in combination with another pharmaceutically active compound. Examples of pharmaceutically active compounds include compounds known to treat autoimmune disorders, e.g., immunosuppressant agents such as cyclosporin A, rapamycin, desoxyspergualine, FK 506, steroids, azathioprine, anti-T cell antibodies and monoclonal antibodies to T cell subpopulations. Other pharmaceutically active compounds that may be used can be found in Harrison's Principles of Internal Medicine, Thirteenth Edition, Eds. T.R. Harrison et al. McGraw-Hill N.Y., NY; and the Physicians Desk Reference 50th Edition 1997, Oradell New Jersey, Medical Economics Co., the complete contents of which are expressly incorporated herein by reference. The vitamin D₃ compound and the
pharmaceutically active compound may be administered to the subject in the same pharmaceutical composition or in different pharmaceutical compositions (at the same time or at different times).

A. Hyperproliferative Conditions

In another aspect, the present invention provides a method of treating a subject for a disorder characterized by aberrant activity of a vitamin D$_3$-responsive cell. The method involves administering to the subject an effective amount of a pharmaceutical composition of a vitamin D$_3$ compound of formula I or otherwise described herein such that the activity of the cell is modulated.

In certain embodiments, the cells to be treated are hyperproliferative cells. As described in greater detail below, the vitamin D$_3$ compounds of the invention can be used to inhibit the proliferation of a variety of hyperplastic and neoplastic tissues. In accordance with the present invention, vitamin D$_3$ compounds of the invention can be used in the treatment of both pathologic and non-pathologic proliferative conditions characterized by unwanted growth of vitamin D$_3$-responsive cells, e.g., hyperproliferative skin cells, immune cells, and tissue having transformed cells, e.g., such as carcinomas, sarcomas and leukemias. In other embodiments, the cells to be treated are aberrant secretory cells, e.g., parathyroid cells, immune cells.

The use of vitamin D compounds in treating hyperproliferative conditions has been limited because of their hypercalcemic effects. Thus, vitamin D$_3$ compounds of the invention can provide a less toxic alternative to current methods of treatment.

In one embodiment, the invention features a method for inhibiting the proliferation and/or inducing the differentiation of a hyperproliferative skin cell, e.g., an epidermal or an epithelial cell, e.g., a keratinocytes, by contacting the cells with a vitamin D$_3$ compound of the invention. In general, the method includes a step of contacting a pathological or non-pathological hyperproliferative cell with an effective amount of such vitamin D$_3$ compound to promote the differentiation of the hyperproliferative cells. The present method can be performed on cells in culture, e.g., in vitro or ex vivo, or can be performed on cells present in an animal subject, e.g., as part of an in vivo therapeutic protocol. The therapeutic regimen can be carried out on a human or any other animal subject.
The vitamin D₃ compounds of the present invention can be used to treat a hyperproliferative skin disorder. Exemplary disorders include, but are not limited to, psoriasis, basal cell carcinoma, keratinization disorders and keratosis. Additional examples of these disorders include eczema; lupus associated skin lesions; psoriatic arthritis; rheumatoid arthritis that involves hyperproliferation and inflammation of epithelial-related cells lining the joint capsule; dermatitides such as seborrheic dermatitis and solar dermatitis; keratoses such as seborrheic keratosis, senile keratosis, actinic keratosis. photo-induced keratosis, and keratosis follicularis; acne vulgaris; keloids and prophylaxis against keloid formation; nevi; warts including verruca, condyloma or condyloma acuminatum, and human papilloma viral (HPV) infections such as venereal warts; leukoplakia; lichen planus; and keratitis.

In an illustrative example, vitamin D₃ compounds of the invention can be used to inhibit the hyperproliferation of keratinocytes in treating diseases such as psoriasis by administering an effective amount of these compounds to a subject in need of treatment. The term "psoriasis" is intended to have its medical meaning, namely, a disease which afflicts primarily the skin and produces raised, thickened, scaling, nonscarring lesions. The lesions are usually sharply demarcated erythematous papules covered with overlapping shiny scales. The scales are typically silvery or slightly opalescent. Involvement of the nails frequently occurs resulting in pitting, separation of the nail, thickening and discoloration. Psoriasis is sometimes associated with arthritis, and it may be crippling. Hyperproliferation of keratinocytes is a key feature of psoriatic epidermal hyperplasia along with epidermal inflammation and reduced differentiation of keratinocytes. Multiple mechanisms have been invoked to explain the keratinocyte hyperproliferation that characterizes psoriasis. Disordered cellular immunity has also been implicated in the pathogenesis of psoriasis.

**B. Neoplasia**

The invention also features methods for inhibiting the proliferation and/or reversing the transformed phenotype of vitamin D₃-responsive hyperproliferative cells by contacting the cells with a vitamin D₃ compound of formula I or otherwise described herein. In general, the method includes a step of contacting pathological or non-pathological hyperproliferative cells with an effective amount of a vitamin D₃ compound of the invention for promoting the differentiation of the hyperproliferative cells. The
present method can be performed on cells in culture, e.g., in vitro or ex vivo, or can be performed on cells present in an animal subject, e.g., as part of an in vivo therapeutic protocol. The therapeutic regimen can be carried out on a human or other subject.


The subject method may also be used to inhibit the proliferation of hyperplastic/neoplastic cells of hematopoietic origin, e.g., arising from myeloid, lymphoid or erythroid lineages, or precursor cells thereof. For instance, the present invention contemplates the treatment of various myeloid disorders including, but not limited to, acute promyeloid leukemia (APML), acute myelogenous leukemia (AML) and chronic myelogenous leukemia (CML) (reviewed in Vaickus, L. (1991) Crit Rev. in Oncol./Hemotol. 11:267-97). Lymphoid malignancies which may be treated by the subject method include, but are not limited to acute lymphoblastic leukemia (ALL) which includes B-lineage ALL and T-lineage ALL, chronic lymphocytic leukemia (CLL), prolymphocytic leukemia (PLL), hairy cell leukemia (HLL) and Waldenstrom's macroglobulinemia (WM). Additional forms of malignant lymphomas contemplated by the treatment method of the present invention include, but are not limited to non-
Hodgkin lymphoma and variants thereof, peripheral T cell lymphomas, adult T cell leukemia/lymphoma (ATL), cutaneous T-cell lymphoma (CTCL), large granular lymphocytic leukemia (LGL) and Hodgkin's disease.

In certain embodiments, the vitamin D3 compounds of the invention can be used in combinatorial therapy with conventional cancer chemotherapeutics. Conventional treatment regimens for leukemia and for other tumors include radiation, drugs, or a combination of both. In addition to radiation, the following drugs, usually in combinations with each other, are often used to treat acute leukemias: vincristine, prednisone, methotrexate, mercaptopurine, cyclophosphamide, and cytarabine. In chronic leukemia, for example, busulfan, melphalan, and chlorambucil can be used in combination. All of the conventional anti-cancer drugs are highly toxic and tend to make patients quite ill while undergoing treatment. Vigorous therapy is based on the premise that unless every leukemic cell is destroyed, the residual cells will multiply and cause a relapse.

The subject method can also be useful in treating malignancies of the various organ systems, such as affecting lung, breast, lymphoid, gastrointestinal, and genitourinary tract as well as adenocarcinomas which include malignancies such as most colon cancers, renal-cell carcinoma, prostate cancer and/or testicular tumors, non-small cell carcinoma of the lung, cancer of the small intestine, cancer of the esophagus, and bladder cancer.

According to the general paradigm of vitamin D3 involvement in differentiation of transformed cells, exemplary solid tumors that can be treated according to the method of the present invention include vitamin D3-responsive phenotypes of sarcomas and carcinomas such as, but not limited to: fibrosarcoma, myxosarcoma, liposarcoma, chondrosarcoma, osteogenic sarcoma, chordoma, angiosarcoma, endotheliosarcoma, lymphangiosarcoma, lymphangioendotheliosarcoma, synovioma, mesothelioma, Ewing's tumor, leiomyosarcoma, rhabdomyosarcoma, colon carcinoma, pancreatic cancer, breast cancer, ovarian cancer, prostate cancer, bladder cancer, squamous cell carcinoma, basal cell carcinoma, adenocarcinoma, sweat gland carcinoma, sebaceous gland carcinoma, papillary carcinoma, papillary adenocarcinomas, cystadenocarcinoma, medullary carcinoma, bronchogenic carcinoma, renal cell carcinoma, hepatoma, bile duct carcinoma, chorionicarcinoma, seminoma, embryonal carcinoma, Wilms' tumor, cervical cancer, testicular tumor, lung carcinoma, small cell lung carcinoma, bladder...
carcinoma, epithelial carcinoma, glioma, astrocytoma, medulloblastoma, craniopharyngioma, ependymoma, pinealoma, hemangioblastoma, acoustic neuroma, oligodendroglialoma, menigioma, melanoma, neuroblastoma, and retinoblastoma.

Determination of a therapeutically effective anti-neoplastic amount or a prophylactically effective anti-neoplastic amount of the vitamin D₃ compound of the invention, can be readily made by the physician or veterinarian (the "attending clinician"), as one skilled in the art, by the use of known techniques and by observing results obtained under analogous circumstances. The dosages may be varied depending upon the requirements of the patient in the judgment of the attending clinician, the severity of the condition being treated and the particular compound being employed. In determining the therapeutically effective antineoplastic amount or dose, and the prophylactically effective antineoplastic amount or dose, a number of factors are considered by the attending clinician, including, but not limited to: the specific hyperplastic/neoplastic cell involved; pharmacodynamic characteristics of the particular agent and its mode and route of administration; the desirider time course of treatment; the species of mammal; its size, age, and general health; the specific disease involved; the degree of or involvement or the severity of the disease; the response of the individual patient; the particular compound administered; the mode of administration; the bioavailability characteristics of the preparation administered; the dose regimen selected; the kind of concurrent treatment (i.e., the interaction of the vitamin D₃ compounds of the invention with other co-administered therapeutics); and other relevant circumstances. U.S. Patent 5,427,916, for example, describes method for predicting the effectiveness of antineoplastic therapy in individual patients, and illustrates certain methods which can be used in conjunction with the treatment protocols of the instant invention.

Treatment can be initiated with smaller dosages which are less than the optimum dose of the compound. Thereafter, the dosage should be increased by small increments until the optimum effect under the circumstances is reached. For convenience, the total daily dosage may be divided and administered in portions during the day if desired. A therapeutically effective antineoplastic amount and a prophylactically effective antineoplastic amount of a vitamin D₃ compound of the invention is expected to vary from about 0.1 milligram per kilogram of body weight per day (mg/kg/day) to about 100 mg/kg/day.
Compounds which are determined to be effective for the prevention or treatment of tumors in animals, e.g., dogs, rodents, may also be useful in treatment of tumors in humans. Those skilled in the art of treating tumors in humans will know, based upon the data obtained in animal studies, the dosage and route of administration of the compound to humans. In general, the dosage and route of administration in humans is expected to be similar to that in animals.

The identification of those patients who are in need of prophylactic treatment for hyperplastic/neoplastic disease states is well within the ability and knowledge of one skilled in the art. Certain of the methods for identification of patients which are at risk of developing neoplastic disease states which can be treated by the subject method are appreciated in the medical arts, such as family history of the development of a particular disease state and the presence of risk factors associated with the development of that disease state in the subject patient. A clinician skilled in the art can readily identify such candidate patients, by the use of, for example, clinical tests, physical examination and medical/family history.

C. Immunological Activity

Healthy individuals protect themselves against foreign invaders using many different mechanisms, including physical barriers, phagocytic cells in the blood and tissues, a class of immune cells known as lymphocytes, and various blood-born molecules. All of these mechanisms participate in defending individuals from a potentially hostile environment. Some of these defense mechanisms, known as natural or innate immunity, are present in an individual prior to exposure to infectious microbes or other foreign macromolecules, are not enhanced by such exposures, and do not discriminate among most foreign substances. Other defense mechanisms, known as acquired or specific immunity, are induced or stimulated by exposure of foreign substances, are exquisitely specific for distinct macromolecules, and increase in magnitude and defensive capabilities with each successive exposure to a particular macromolecule. Substances that induce a specific immune response are known as antigens (see, e.g., Abbas, A. et al., *Cellular and Molecular Immunology*, W.B. Saunders Company, Philadelphia, 1991; Silverstein, A.M. A history of Immunology, San Diego, Academic Press, 1989; Unanue A. et al., *Textbook of Immunology*, 2nd ed. Williams and Wilkens, Baltimore, 1984).
One of the most remarkable properties of the immune system is its ability to distinguish between foreign antigens and self-antigens. Therefore, the lymphocytes in each individual are able to recognize and respond to many foreign antigens but are normally unresponsive to the potentially antigenic substances present in the individual.


Self-tolerance is an acquired process that has to be learned by the lymphocytes of each individual. It occurs in part because lymphocytes pass through a stage in their development when an encounter with antigen presented by antigen-presenting cells (APCs) leads to their death or inactivation in a process known as positive and negative selection (see, e.g., Debatin KM (2001) Ann. Hematol. 80 Suppl 3:B29; Abbas, A. (1991), supra). Thus, potentially self-recognizing lymphocytes come into contact with self-antigens at this stage of functional immaturity and are prevented from developing to a stage at which they would be able to respond to self-antigens. Autoimmunity arises when abnormalities in the induction or maintenance of self-tolerance occur that result in a loss of tolerance to a particular antigen(s) and a subsequent attack by the host’s immune system on the host’s tissues that express the antigen(s) (see, e.g., Boyton RJ et al. (2002) Clin. Exp. Immunol. 127:4; Hagiwara E. (2001) Ryumachi 41:888; Burt RK et al. (1992) Blood 99:768).

The ability of the immune system to distinguish between self and foreign antigens also plays a critical role in tissue transplantation. The success of a transplant depends on preventing the immune system of the host recipient from recognizing the transplant as foreign and, in some cases, preventing the graft from recognizing the host tissues as foreign. For example, when a host receives a bone marrow transplant, the transplanted bone marrow may recognize the new host as foreign, resulting in graft versus host disease (GVHD). Consequently, the survival of the host depends on preventing both the rejection of the donor marrow as well as rejection of the host by the graft immune reaction (see, e.g., Waldmann H et al. (2001) Int. Arch. Allergy Immunol. 126:11).

Currently, deleterious immune reactions that result in autoimmune diseases and transplant rejections are prevented or treated using agents such as steroids, azathioprine,
anti-T cell antibodies, and more recently, monoclonal antibodies to T cell subpopulations. Immunosuppressive drugs such as cyclosporin A (CsA), rapamycin, desoxyspergualine and FK-506 are also widely used.

Nonspecific immune suppression agents, such as steroids and antibodies to lymphocytes, put the host at increased risk for opportunistic infection and development of tumors. Moreover, many immunosuppressive drugs result in bone demineralization within the host (see, e.g., Chhajed PN et al. (2002) Indian J. Chest Dis. Allied 44:31; Wijdicks EF (2001) Liver Transpl. 7:937; Karamenic J et al. (2001) Med. Arh. 55:243; U.S. Patent No. 5,597,563 issued to Beschorner, WE and U.S. Patent No. 6,071,897 issued to DeLuca HF et al.). Because of the major drawbacks associated with existing immunosuppressive modalities, there is a need for a new approach for treating immune disorders, e.g., for inducing immune tolerance in a host.

Thus, in another aspect, the invention provides a method for modulating the activity of an immune cell by contacting the cell with a vitamin D₃ compound of formula I or otherwise described herein.

In one embodiment, the present invention provides a method for suppressing immune activity in an immune cell by contacting a pathological or non-pathological immune cell with an effective amount of a vitamin D₃ compound of the invention to thereby inhibit an immune response relative to the cell in the absence of the treatment.

The present method can be performed on cells in culture, e.g., in vitro or ex vivo, or can be performed on cells present in an animal subject, e.g., as part of an in vivo therapeutic protocol. In vivo treatment can be carried out on a human or other animal subject.


After identifying certain test compounds as effective suppressors of an immune response in vitro, these compounds can be used in vivo as part of a therapeutic protocol. Accordingly, another aspect of the invention provides a method of suppressing an immune response, comprising administering to a subject a pharmaceutical preparation of a vitamin D₃ compound of the invention, so as to inhibit immune reactions such as graft rejection, autoimmune disorders and inflammation.

In one embodiment, the invention provides a method for treating a subject for a vitamin D₃ associated state, wherein the vitamin D₃ associated state is an ILT3-associated disorder, by administering to the subject an effective amount of a vitamin D₃ compound of the invention. In one embodiment, the the ILT3-associated state is an immune disorder. In certain embodiments, the immune disorder is an autoimmune disorder. In a specific embodiment, the immune disorder is Type 1 diabetes mellitus. In other embodiments, the immune disorder is transplant rejection.

For example, the subject vitamin D₃ compound of the invention can be used to inhibit responses in clinical situations where it is desirable to downmodulate T cell responses. For example, in graft-versus-host disease, cases of transplantation, autoimmune diseases (including, for example, diabetes mellitus, arthritis (including rheumatoid arthritis, juvenile rheumatoid arthritis, osteoarthritis, psoriatic arthritis), multiple sclerosis, encephalomyelitis, diabetes, myasthenia gravis, systemic lupus erythematosus, autoimmune thyroiditis, dermatitis (including atopic dermatitis and eczematous dermatitis), psoriasis, Sjögren's Syndrome, including keratoconjunctivitis sicca secondary to Sjögren's Syndrome, alopecia areata, allergic responses due to arthropod bite reactions, Crohn's disease, aphthous ulcer, iritis, conjunctivitis,
keratoconjunctivitis, ulcerative colitis, asthma, allergic asthma, cutaneous lupus erythematosus, scleroderma, vaginitis, proctitis, drug eruptions, leprosy reversal reactions, erythema nodosum leprosum, autoimmune uveitis, allergic encephalomyelitis, acute necrotizing hemorrhagic encephalopathy, idiopathic bilateral progressive sensorineural hearing loss, aplastic anemia, pure red cell anemia, idiopathic thrombocytopenia, polychondritis, Wegener's granulomatosis, chronic active hepatitis, Stevens-Johnson syndrome, idiopathic sprue, lichen planus, Crohn's disease, Graves ophthalmopathy, sarcoidosis, primary biliary cirrhosis, uveitis posterior, and interstitial lung fibrosis. Downmodulation of immune activity will also be desirable in cases of allergy such as, atopic allergy.

Another aspect of the invention provides a method of modulating the expression of an immunoglobulin-like transcript 3 (ILT3) surface molecule in a cell. The method includes contacting the cell with a compound of formula I in an amount effective to modulate the expression of an immunoglobulin-like transcript 3 (ILT3) surface molecule in the cell. In one embodiment, cell is within a subject a subject. In another embodiment the moduliation is upregulation of expression. In other embodiment, the modulation is downregulation of expression.

Another related aspect of the invention provides a method of treating an ILT3-associated disorder in a subject. The method includes administering to the subject a compound of formula I in an amount effective to modulate the expression of an ILT3 surface molecule, thereby treating the ILT3-associated disorder in the subject.

In certain embodiments, the present invention provides methods and compositions for treating immune disorders, such as, for example, autoimmune disorders and transplant rejections, such as graft versus host disease (GVHD). These embodiments of the invention are based on the discovery that vitamin D₃ compounds of the invention are able to modulate the expression of immunoglobulin-like transcript 3 (ILT3) on cells, e.g., antigen-presenting cells.

Accordingly, another aspect of the invention provides a method for inhibiting transplant rejection in a subject. The method includes administering to the subject a compound of formula I in an amount effective to modulate the expression of an ILT3 surface molecule, thereby inhibiting transplant rejection in the subject. In one embodiment, the transplant is an organ transplant. In another embodiment, the
transplant is a pancreatic islet transplant. In yet another embodiment, the transplant is a bone marrow transplant.

As described before, determination of a therapeutically effective immunosuppressive amount can be readily made by the attending clinician, as one skilled in the art, by the use of known techniques and by observing results obtained under analogous circumstances. Compounds which are determined to be effective in animals, e.g., dogs, rodents, may be extrapolated accordingly to humans by those skilled in the art. Starting dose/regimen used in animals can be estimated based on prior studies. For example, doses of vitamin D₃ compounds of the invention to treat autoimmune disorders in rodents can be initially estimated in the range of 0.1 g/kg/day to 1 g/kg/day, administered orally or by injection.

Those skilled in the art will know based upon the data obtained in animal studies, the dosage and route of administration in humans is expected to be similar to that in animals. Exemplary dose ranges to be used in humans are from 0.25 to 10 µg/day, preferably 0.5 to 5 µg/day per adult (U.S. Pat. No. 4,341,774).

D. Calcium and Phosphate Homeostasis

The present invention also relates to a method of treating a subject a disorder characterized by deregulation of calcium metabolism. This method comprises contacting a pathological or non-pathological vitamin D₃ responsive cell with an effective amount of a vitamin D₃ compound of the invention to thereby directly or indirectly modulate calcium and phosphate homeostasis. Techniques for detecting calcium fluctuation in vivo or in vitro are known in the art.

from bone explants in vitro (Bouillon R. et al. (1992) J. Biol. Chem. 267:3044-3051), 2) measurement of serum osteocalcin levels [osteocalcin is an osteoblast-specific protein that after its synthesis is largely incorporated into the bone matrix, but partially released into the circulation (or tissue culture medium) and thus represents a good market of bone formation or turnover] (Bouillon R. et al. (1992) Clin. Chem. 38:2055-2060), or 3) bone ash content (Norman A.W. and Wong R.G. (1972) J. Nutr. 102:1709-1718). Only one kidney-oriented assay has been employed. In this assay, urinary Ca$^{2+}$ excretion is determined (Hartenbower D.L. et al. (1977) Walter de Gruyter, Berlin pp 587-589); this assay is dependent upon elevations in the serum Ca$^{2+}$ level and may reflect bone Ca$^{2+}$ mobilizing activity more than renal effects. Finally, there is a "soft tissue calcification" assay that can be used to detect the consequences of administration of a compound of the invention. In this assay a rat is administered an intraperitoneal dose of $^{45}$Ca$^{2+}$, followed by seven daily relative high doses of a compound of the invention; in the event of onset of a severe hypercalcemia, soft tissue calcification can be assessed by determination of the $^{45}$Ca$^{2+}$ level. In all these assays, vitamin D$_3$ compounds of the invention are administered to vitamin D-sufficient or -deficient animals, as a single dose or chronically (depending upon the assay protocol), at an appropriate time interval before the end point of the assay is quantified.

In certain embodiments, vitamin D$_3$ compounds of the invention can be used to modulate bone metabolism. The language "bone metabolism" is intended to include direct or indirect effects in the formation or degeneration of bone structures, e.g., bone formation, bone resorption, etc., which may ultimately affect the concentrations in serum of calcium and phosphate. This term is also intended to include effects of vitamin D$_3$ compounds in bone cells, e.g. osteoclasts and osteoblasts, that may in turn result in bone formation and degeneration. For example, it is known in the art, that vitamin D$_3$ compounds exert effects on the bone forming cells, the osteoblasts through genomic and non-genomic pathways (Walters M.R. et al. (1982) J. Biol. Chem. 257:7481-7484; Jurutka P.W. et al. (1993) Biochemistry 32:8184-8192; Mellon W.S. and DeLuca H.F. (1980) J. Biol. Chem. 255:4081-4086). Similarly, vitamin D$_3$ compounds are known in the art to support different activities of bone resorbing osteoclasts such as the stimulation of differentiation of monocytes and mononuclear phagocytes into osteoclasts (Abe E. et al. (1988) J. Bone Miner Res. 3:635-645; Takahashi N. et al. (1988) Endocrinology 123:1504-1510; Udagawa N. et al. (1990) Proc. Natl. Acad. Sci. USA
Accordingly, vitamin D₃ compounds of the invention that modulate the production of bone cells can influence bone formation and degeneration.

The present invention provides a method for modulating bone cell metabolism by contacting a pathological or a non-pathological bone cell with an effective amount of a vitamin D₃ compound of the invention to thereby modulate bone formation and degeneration. The present method can be performed on cells in culture, e.g., in vitro or ex vivo, or can be performed in cells present in an animal subject, e.g., cells in vivo. Exemplary culture systems that can be used include osteoblast cell lines, e.g., ROS 17/2.8 cell line, monocytes, bone marrow culture system (Suda T. et al. (1990) Med. Res. Rev. 7:333-366; Suda T. et al. (1992) J. Cell Biochem. 49:53-58) among others. Selected compounds can be further tested in vivo, for example, animal models of osteopetrosis and in human disease (Shapira F. (1993) Clin. Orthop. 294:34-44).

In a preferred embodiment, a method for treating osteoporosis is provided, comprising administering to a subject a pharmaceutical preparation of a vitamin D₃ compound of the invention to thereby ameliorate the condition relative to an untreated subject.

Vitamin D₃ compounds of the invention can be tested in ovariectomized animals, e.g., dogs, rodents, to assess the changes in bone mass and bone formation rates in both normal and estrogen-deficient animals. Clinical trials can be conducted in humans by attending clinicians to determine therapeutically effective amounts of the vitamin D₃ compounds of the invention in preventing and treating osteoporosis.

In other embodiments, therapeutic applications of the vitamin D₃ compounds of the invention include treatment of other diseases characterized by metabolic calcium and phosphate deficiencies. Exemplary of such diseases are the following: osteoporosis, osteodystrophy, osteomalacia, rickets, osteitis fibrosa cystica, renal osteodystrophy, osteosclerosis, anti-convulsant treatment, osteopenia, fibrogenesis-imperfecta ossium, secondary hyperparathyroidism, hypoparathyroidism, hyperparathyroidism, cirrhosis, obstructive jaundice, drug induced metabolism, medullary carcinoma, chronic renal disease, hypophosphatemic VDDR, vitamin D-dependent rickets, sarcoidosis, glucocorticoid antagonism, malabsorption syndrome, steatorrhea, tropical sprue, idiopathic hypercalcemia and milk fever.
E. Hormone Secretion

In yet another aspect, the present invention provides a method for modulating hormone secretion of a vitamin D₃-responsive cell, e.g., an endocrine cell. Hormone secretion includes both genomic and non-genomic activities of vitamin D₃ compounds of the invention that control the transcription and processing responsible for secretion of a given hormone e.g., parathyroid hormone (PTH), calcitonin, insulin, prolactin (PRL) and TRH in a vitamin D₃-responsive cell (Bouillon, R. et al. (1995) Endocrine Reviews 16(2):235-237).


In certain embodiments, the vitamin D₃ compounds of the present invention can be used to inhibit parathyroid hormone (PTH) processing, e.g., transcriptional, translational processing, and/or secretion of a parathyroid cell as part of a therapeutic protocol. Therapeutic methods using these compounds can be readily applied to all diseases, involving direct or indirect effects of PTH activity, e.g., primary or secondary responses.

Accordingly, therapeutic applications for the vitamin D₃ compounds of the invention include treating diseases such as secondary hyperparathyroidism of chronic renal failure (Slatopolsky E. et al. (1990) Kidney Int. 38:S41-S47; Brown A.J. et al. (1989) J. Clin. Invest. 84:728-732). Determination of therapeutically affective amounts and dose regimen can be performed by the skilled artisan using the data described in the art.

F. Protection Against Neuronal Loss

In yet another aspect, the present invention provides a method of protecting against neuronal loss by contacting a vitamin D₃ responsive cell, e.g., a neuronal cell, with a vitamin D₃ compound of the invention to prevent or retard neuron loss. The language "protecting against" is intended to include prevention, retardation, and/or termination of deterioration, impairment, or death of a neurons.

Neuron loss can be the result of any condition of a neuron in which its normal function is compromised. Neuron deterioration can be the result of any condition which compromises neuron function which is likely to lead to neuron loss. Neuron function can be compromised by, for example, altered biochemistry, physiology, or anatomy of a neuron. Deterioration of a neuron may include membrane, dendritic, or synaptic changes which are detrimental to normal neuronal functioning. The cause of the neuron deterioration, impairment, and/or death may be unknown. Alternatively, it may be the result of age- and/or disease-related changes which occur in the nervous system of a subject.

When neuron loss is described herein as "age-related", it is intended to include neuron loss resulting from known and unknown bodily changes of a subject which are associated with aging. When neuron loss is described herein as "disease-related", it is
intended to include neuron loss resulting from known and unknown bodily changes of a subject which are associated with disease. It should be understood, however, that these terms are not mutually exclusive and that, in fact, many conditions that result in the loss of neurons are both age- and disease-related.

Exemplary age-related diseases associated with neuron loss and changes in neuronal morphology include, for example, Alzheimer's Disease, Pick's Disease, Parkinson's Disease, Vascular Disease, Huntington's Disease, and Age-Associated Memory Impairment. In Alzheimer's Disease patients, neuron loss is most notable in the hippocampus, frontal, parietal, and anterior temporal cortices, amygdala, and the olfactory system. The most prominently affected zones of the hippocampus include the CA1 region, the subiculum, and the entorhinal cortex. Memory loss is considered the earliest and most representative cognitive change because the hippocampus is well known to play a crucial role in memory. Pick's Disease is characterized by severe neuronal degeneration in the neocortex of the frontal and anterior temporal lobes which is sometimes accompanied by death of neurons in the striatum. Parkinson's Disease can be identified by the loss of neurons in the substantia nigra and the locus ceruleus. Huntington's Disease is characterized by degeneration of the intrastriatal and cortical cholinergic neurons and GABA-ergic neurons. Parkinson's and Huntington's Diseases are usually associated with movement disorders, but often show cognitive impairment (memory loss) as well.

Age-Associated Memory Impairment (AAMI) is another age-associated disorder that is characterized by memory loss in healthy, elderly individuals in the later decades of life. Crook, T. et al. (1986) Devel. Neuropsych. 2(4):261-276. Presently, the neural basis for AAMI has not been precisely defined. However, neuron death with aging has been reported to occur in many species in brain regions implicated in memory, including cortex, hippocampus, amygdala, basal ganglia, cholinergic basal forebrain, locus ceruleus, raphe nuclei, and cerebellum. Crook, T. et al. (1986) Devel. Neuropsych. 2(4):261-276.

Vitamin D₃ compounds of the invention can protect against neuron loss by genomic or non-genomic mechanisms. Nuclear vitamin D₃ receptors are well known to exist in the periphery but have also been found in the brain, particularly in the hippocampus and neocortex. Non-genomic mechanisms may also prevent or retard neuron loss by regulating intraneuronal and/or peripheral calcium and phosphate levels.
Furthermore, vitamin D₃ compounds of the invention may protect against neuronal loss by acting indirectly, e.g., by modulating serum PTH levels. For example, a positive correlation has been demonstrated between serum PTH levels and cognitive decline in Alzheimer's Disease.

The present method can be performed on cells in culture, e.g. *in vitro* or *ex vivo*, or on cells present in an animal subject, e.g., *in vivo*. Vitamin D₃ compounds of the invention can be initially tested *in vitro* using neurons from embryonic rodent pups (See e.g. U.S. Patent No. 5,179,109-fetal rat tissue culture), or other mammalian (See e.g. U.S. Patent No. 5,089,517-fetal mouse tissue culture) or non-mammalian animal models. These culture systems have been used to characterize the protection of peripheral, as well as, central nervous system neurons in animal or tissue culture models of ischemia, stroke, trauma, nerve crush, Alzheimer's Disease, Pick's Disease, and Parkinson's Disease, among others. Examples of *in vitro* systems to study the prevention of destruction of neocortical neurons include using *in vitro* cultures of fetal mouse neurons and glial cells previously exposed to various glutamate agonists, such as kainate, NMDA, and α-amino-3-hydroxy-5-methyl-4-isoxazolepropionate (AMPA). U.S. Patent No. 5,089,517. See also U.S. Patent No. 5,170,109 (treatment of rat cortical/hippocampal neuron cultures with glutamate prior to treatment with neuroprotective compound); U.S. Patent Nos. 5,163,196 and 5,196,421 (neuroprotective excitatory amino acid receptor antagonists inhibit glycine, kainate, AMPA receptor binding in rats).

Alternatively, the effects of vitamin D₃ compounds of the invention can be characterized *in vivo* using animals models. Neuron deterioration in these model systems is often induced by experimental trauma or intervention (e.g. application of toxins, nerve crush, interruption of oxygen supply).

G. Smooth Muscle Cells

In yet another aspect, the present invention provides a method of modulating the activity of a vascular smooth muscle cell by contacting a vitamin D₃-responsive smooth muscle cell with a vitamin D₃ compound of the invention to activate or, preferably, inhibit the activity of the cell. The language "activity of a smooth muscle cell" is intended to include any activity of a smooth muscle cell, such as proliferation, migration, adhesion and/or metabolism.
In certain embodiments, the vitamin D₃ compounds of the invention can be used to treat diseases and conditions associated with aberrant activity of a vitamin D₃-responsive smooth muscle cell. For example, the present invention can be used in the treatment of hyperproliferative vascular diseases, such as hypertension induced vascular remodeling, vascular restenosis and atherosclerosis. In other embodiments, the compounds of the present invention can be used in treating disorders characterized by aberrant metabolism of a vitamin D₃-responsive smooth muscle cell, e.g., arterial hypertension.

The present method can be performed on cells in culture, e.g. in vitro or ex vivo, or on cells present in an animal subject, e.g., in vivo. Vitamin D₃ compounds of the invention can be initially tested in vitro as described in Catellot et al. (1982), J. Biol. Chem. 257(19): 11256.

4. SUPPRESSION OF RENIN EXPRESSION

The compounds of the present invention control blood pressure by the suppression of rennin expression and are useful as antihypertensive agents. Renin-angiotensin regulatory cascade plays a significant role in the regulation of blood pressure, electrolyte and volume homeostasis (Y.C. Li, Abstract, DeLuca Symposium on Vitamin D₃, Tauc, New Mexico, June 15 - June 19, 2002, p. 18). Thus, the invention provides a method of treating a subject for a vitamin D₃ associated state, wherein the vitamin D₃ associated state is a disorder characterized by an aberrant activity of a cell that expresses renin. The method includes administering to the subject an effective amount of a compound of formula I, such that rennin expression by the cell is suppressed, and the subject is thereby treated for hypertension.

5. BLADDER DYSFUNCTION

Morphological bladder changes, including a progressive de-nervation and hypertrophy of the bladder wall are frequent histological findings in patients with different bladder disorders leading to overactive bladder such as bladder disorders associated with, for example, clinical benign prostatic hyperplasia (BPH) and spinal cord injury.

The increase in tension and/or strain on the bladder observed in these conditions has been shown to be associated with cellular and molecular alterations,
e.g., in cytoskeletal and contractile proteins, in mitochondrial function, and in various enzyme activities of the smooth muscle cells. The hypertrophy of the bladder wall also involves alterations in its extracellular matrix and non-smooth muscle components.

These changes in the bladder are associated with the storage (irritative) symptoms, in particular frequency, urgency, urge incontinence and nocturia. These symptoms affect the social, psychological, domestic, occupational, physical and sexual lives of the patients leading to a profound negative impact on their quality of life.

At the present time, an ideal treatment of these symptoms has not been found. Each of the therapeutic options available (for example, anti-muscarinics or alpha-blockers) is associated with disadvantages relating to their mechanism of action, which is based only on the management of symptoms and not on the treatment of the etiology of the condition. In fact, the clinical utility of some of the available agents has been limited by poor efficacy and lack of universal patient acceptance due to a number of significant side effects.

As a consequence there is a need for new treatments that provide improved clinical effectiveness by targeting the underlying etiological factor, the abnormal growth and consequent dysfunction of bladder smooth muscle cells.

As described herein, it has now surprisingly been found that vitamin D analogues can treat and prevent bladder dysfunction in disorders associated with bladder hypertrophy, such as bladder overactivity and clinical BPH. Overactive bladder, also known as detrusor overactivity or detrusor instability, involves involuntary bladder spasms. A hyperactive detrusor muscle can cause overactive bladder. Although the underlying cause of overactive bladder can be neurological disease (e.g., multiple sclerosis, Parkinson's disease, stroke, spinal cord lesions), nerve damage caused by abdominal trauma, pelvic trauma, or surgery, stroke, multiple sclerosis, infection, bladder cancer, drug side effects or enlarged prostate (BPH), in many cases the cause is idiopathic, i.e. of unknown cause.

In addition, such vitamin D related compounds have an application in the treatment of irritative voiding symptoms associated with BPH. BPH is associated not only with enlargement of the gland leading to bladder outlet obstruction (BOO) and symptoms secondary to this, but also to morphological bladder changes, including a hypertrophy of the bladder wall and progressive de-nervation. These changes lead to
increased functional demands and disruption of the coordination within the bladder smooth muscle cells.

6. PHARMACEUTICAL COMPOSITIONS

The invention also provides a pharmaceutical composition, comprising an effective amount of a vitamin D3 compound of formula I or otherwise described herein and a pharmaceutically acceptable carrier. In a further embodiment, the effective amount is effective to treat a vitamin D3 associated state, as described previously.

In an embodiment, the vitamin D3 compound is administered to the subject using a pharmaceutically-acceptable formulation, e.g., a pharmaceutically-acceptable formulation that provides sustained delivery of the vitamin D3 compound to a subject for at least 12 hours, 24 hours, 36 hours, 48 hours, one week, two weeks, three weeks, or four weeks after the pharmaceutically-acceptable formulation is administered to the subject.

In certain embodiments, these pharmaceutical compositions are suitable for topical or oral administration to a subject. In other embodiments, as described in detail below, the pharmaceutical compositions of the present invention may be specially formulated for administration in solid or liquid form, including those adapted for the following: (1) oral administration, for example, drenches (aqueous or non-aqueous solutions or suspensions), tablets, boluses, powders, granules, pastes; (2) parenteral administration, for example, by subcutaneous, intramuscular or intravenous injection as, for example, a sterile solution or suspension; (3) topical application, for example, as a cream, ointment or spray applied to the skin; (4) intravaginally or intrarectally, for example, as a pessary, cream or foam; or (5) aerosol, for example, as an aqueous aerosol, liposomal preparation or solid particles containing the compound.

The phrase "pharmaceutically acceptable" refers to those vitamin D3 compounds of the present invention, compositions containing such compounds, and/or dosage forms which are, within the scope of sound medical judgment, suitable for use in contact with the tissues of human beings and animals without excessive toxicity, irritation, allergic response, or other problem or complication, commensurate with a reasonable benefit/risk ratio.

The phrase "pharmaceutically-acceptable carrier" includes pharmaceutically-acceptable material, composition or vehicle, such as a liquid or solid filler, diluent,
excipient, solvent or encapsulating material, involved in carrying or transporting the subject chemical from one organ, or portion of the body, to another organ, or portion of the body. Each carrier must be "acceptable" in the sense of being compatible with the other ingredients of the formulation and not injurious to the patient. Some examples of materials which can serve as pharmaceutically-acceptable carriers include: (1) sugars, such as lactose, glucose and sucrose; (2) starches, such as corn starch and potato starch; (3) cellulose, and its derivatives, such as sodium carboxymethyl cellulose, ethyl cellulose and cellulose acetate; (4) powdered tragacanth; (5) malt; (6) gelatin; (7) talc; (8) excipients, such as cocoa butter and suppository waxes; (9) oils, such as peanut oil, cottonseed oil, safflower oil, sesame oil, olive oil, corn oil and soybean oil; (10) glycols, such as propylene glycol; (11) polyols, such as glycerin, sorbitol, mannitol and polyethylene glycol; (12) esters, such as ethyl oleate and ethyl laurate; (13) agar; (14) buffering agents, such as magnesium hydroxide and aluminum hydroxide; (15) alginic acid; (16) pyrogen-free water; (17) isotonic saline; (18) Ringer's solution; (19) ethyl alcohol; (20) phosphate buffer solutions; and (21) other non-toxic compatible substances employed in pharmaceutical formulations.

Wetting agents, emulsifiers and lubricants, such as sodium lauryl sulfate and magnesium stearate, as well as coloring agents, release agents, coating agents, sweetening, flavoring and perfuming agents, preservatives and antioxidants can also be present in the compositions.

Examples of pharmaceutically-acceptable antioxidants include: (1) water soluble antioxidants, such as ascorbic acid, cysteine hydrochloride, sodium bisulfate, sodium metabisulfite, sodium sulfite and the like; (2) oil-soluble antioxidants, such as ascorbyl palmitate, butylated hydroxyanisole (BHA), butylated hydroxytoluene (BHT), lecithin, propyl gallate, alpha-tocopherol, and the like; and (3) metal chelating agents, such as citric acid, ethylenediamine tetraacetic acid (EDTA), sorbitol, tartaric acid, phosphoric acid, and the like.

Compositions containing a vitamin D3 compound(s) include those suitable for oral, nasal, topical (including buccal and sublingual), rectal, vaginal, aerosol and/or parenteral administration. The compositions may conveniently be presented in unit dosage form and may be prepared by any methods well known in the art of pharmacy. The amount of active ingredient which can be combined with a carrier material to produce a single dosage form will vary depending upon the host being treated, the
particular mode of administration. The amount of active ingredient which can be combined with a carrier material to produce a single dosage form will generally be that amount of the compound which produces a therapeutic effect. Generally, out of one hundred per cent, this amount will range from about 1 per cent to about ninety-nine percent of active ingredient, preferably from about 5 per cent to about 70 per cent, most preferably from about 10 per cent to about 30 per cent.

Methods of preparing these compositions include the step of bringing into association a vitamin D₃ compound(s) with the carrier and, optionally, one or more accessory ingredients. In general, the formulations are prepared by uniformly and intimately bringing into association a vitamin D₃ compound with liquid carriers, or finely divided solid carriers, or both, and then, if necessary, shaping the product.

Compositions of the invention suitable for oral administration may be in the form of capsules, cachets, pills, tablets, lozenges (using a flavored basis, usually sucrose and acacia or tragacanth), powders, granules, or as a solution or a suspension in an aqueous or non-aqueous liquid, or as an oil-in-water or water-in-oil liquid emulsion, or as an elixir or syrup, or as pastilles (using an inert base, such as gelatin and glycerin, or sucrose and acacia) and/or as mouth washes and the like, each containing a predetermined amount of a vitamin D₃ compound(s) as an active ingredient. A compound may also be administered as a bolus, ejectionary or paste.

In solid dosage forms of the invention for oral administration (capsules, tablets, pills, dragees, powders, granules and the like), the active ingredient is mixed with one or more pharmaceutically-acceptable carriers, such as sodium citrate or dicalcium phosphate, and/or any of the following: (1) fillers or extenders, such as starches, lactose, sucrose, glucose, mannitol, and/or silicic acid; (2) binders, such as, for example, carboxymethylcellulose, alginites, gelatin, polyvinyl pyrrolidone, sucrose and/or acacia; (3) humectants, such as glycerol; (4) disintegrating agents, such as agar-agar, calcium carbonate, potato or tapioca starch, alginic acid, certain silicates, and sodium carbonate; (5) solution retarding agents, such as paraffin; (6) absorption accelerators, such as quaternary ammonium compounds; (7) wetting agents, such as, for example, acetyl alcohol and glycerol monostearate; (8) absorbents, such as kaolin and bentonite clay; (9) lubricants, such as a talc, calcium stearate, magnesium stearate, solid polyethylene glycols, sodium lauryl sulfate, and mixtures thereof; and (10) coloring agents. In the case of capsules, tablets and pills, the pharmaceutical compositions may also comprise buffering.
agents. Solid compositions of a similar type may also be employed as fillers in soft and hard-filled gelatin capsules using such excipients as lactose or milk sugars, as well as high molecular weight polyethylene glycols and the like.

A tablet may be made by compression or molding, optionally with one or more accessory ingredients. Compressed tablets may be prepared using binder (for example, gelatin or hydroxypropylmethyl cellulose), lubricant, inert diluent, preservative, disintegrant (for example, sodium starch glycolate or cross-linked sodium carboxymethyl cellulose), surface-active or dispersing agent. Molded tablets may be made by molding in a suitable machine a mixture of the powdered active ingredient moistened with an inert liquid diluent.

The tablets, and other solid dosage forms of the pharmaceutical compositions of the present invention, such as dragees, capsules, pills and granules, may optionally be scored or prepared with coatings and shells, such as enteric coatings and other coatings well known in the pharmaceutical-formulating art. They may also be formulated so as to provide slow or controlled release of the active ingredient therein using, for example, hydroxypropylmethyl cellulose in varying proportions to provide the desired release profile, other polymer matrices, liposomes and/or microspheres. They may be sterilized by, for example, filtration through a bacteria-retaining filter, or by incorporating sterilizing agents in the form of sterile solid compositions which can be dissolved in sterile water, or some other sterile injectable medium immediately before use. These compositions may also optionally contain opacifying agents and may be of a composition that they release the active ingredient(s) only, or preferentially, in a certain portion of the gastrointestinal tract, optionally, in a delayed manner. Examples of embedding compositions which can be used include polymeric substances and waxes.

The active ingredient can also be in micro-encapsulated form, if appropriate, with one or more of the above-described excipients.

Liquid dosage forms for oral administration of the vitamin D₃ compound(s) include pharmaceutically-acceptable emulsions, microemulsions, solutions, suspensions, syrups and elixirs. In addition to the active ingredient, the liquid dosage forms may contain inert diluents commonly used in the art, such as, for example, water or other solvents, solubilizing agents and emulsifiers, such as ethyl alcohol, isopropyl alcohol, ethyl carbonate, ethyl acetate, benzyl alcohol, benzyl benzoate, propylene glycol, 1,3-butylene glycol, oils (in particular, cottonseed, groundnut, corn, germ, olive, castor and
sesame oils), glycerol, tetrahydrofurfuryl alcohol, polyethylene glycols and fatty acid esters of sorbitan, and mixtures thereof.

In addition to inert diluents, the oral compositions can include adjuvants such as wetting agents, emulsifying and suspending agents, sweetening, flavoring, coloring, perfuming and preservative agents.

Suspensions, in addition to the active vitamin D₃ compound(s) may contain suspending agents as, for example, ethoxylated isostearyl alcohols, polyoxyethylene sorbitol and sorbitan esters, microcrystalline cellulose, aluminum metahydroxide, bentonite, agar-agar and tragacanth, and mixtures thereof.

Pharmaceutical compositions of the invention for rectal or vaginal administration may be presented as a suppository, which may be prepared by mixing one or more vitamin D₃ compound(s) with one or more suitable nonirritating excipients or carriers comprising, for example, cocoa butter, polyethylene glycol, a suppository wax or a salicylate, and which is solid at room temperature, but liquid at body temperature and, therefore, will melt in the rectum or vaginal cavity and release the active agent.

Compositions of the present invention which are suitable for vaginal administration also include pessaries, tampons, creams, gels, pastes, foams or spray formulations containing such carriers as are known in the art to be appropriate.

Dosage forms for the topical or transdermal administration of a vitamin D₃ compound(s) include powders, sprays, ointments, pastes, creams, lotions, gels, solutions, patches and inhalants. The active vitamin D₃ compound(s) may be mixed under sterile conditions with a pharmaceutically-acceptable carrier, and with any preservatives, buffers, or propellants which may be required.

The ointments, pastes, creams and gels may contain, in addition to vitamin D₃ compound(s) of the present invention, excipients, such as animal and vegetable fats, oils, waxes, paraffins, starch, tragacanth, cellulose derivatives, polyethylene glycols, silicones, bentonites, silicic acid, talc and zinc oxide, or mixtures thereof.

Powders and sprays can contain, in addition to a vitamin D₃ compound(s), excipients such as lactose, talc, silicic acid, aluminum hydroxide, calcium silicates and polyamide powder, or mixtures of these substances. Sprays can additionally contain customary propellants, such as chlorofluorohydrocarbons and volatile unsubstituted hydrocarbons, such as butane and propane.
The vitamin D₃ compound(s) can be alternatively administered by aerosol. This is accomplished by preparing an aqueous aerosol, liposomal preparation or solid particles containing the compound. A nonaqueous (e.g., fluorocarbon propellant) suspension could be used. Sonic nebulizers are preferred because they minimize exposing the agent to shear, which can result in degradation of the compound.

Ordinarily, an aqueous aerosol is made by formulating an aqueous solution or suspension of the agent together with conventional pharmaceutically-acceptable carriers and stabilizers. The carriers and stabilizers vary with the requirements of the particular compound, but typically include nonionic surfactants (Tweens, Pluronics, or polyethylene glycol), innocuous proteins like serum albumin, sorbitan esters, oleic acid, lecithin, amino acids such as glycine, buffers, salts, sugars or sugar alcohols. Aerosols generally are prepared from isotonic solutions.

Transdermal patches have the added advantage of providing controlled delivery of a vitamin D₃ compound(s) to the body. Such dosage forms can be made by dissolving or dispersing the agent in the proper medium. Absorption enhancers can also be used to increase the flux of the active ingredient across the skin. The rate of such flux can be controlled by either providing a rate controlling membrane or dispersing the active ingredient in a polymer matrix or gel.

Ophthalmic formulations, eye ointments, powders, solutions and the like, are also contemplated as being within the scope of the invention.

Pharmaceutical compositions of the invention suitable for parenteral administration comprise one or more vitamin D₃ compound(s) in combination with one or more pharmaceutically-acceptable sterile isotonic aqueous or nonaqueous solutions, dispersions, suspensions or emulsions, or sterile powders which may be reconstituted into sterile injectable solutions or dispersions just prior to use, which may contain antioxidants, buffers, bacteriostats, solutes which render the formulation isotonic with the blood of the intended recipient or suspending or thickening agents.

Examples of suitable aqueous and nonaqueous carriers which may be employed in the pharmaceutical compositions of the invention include water, ethanol, polyols (such as glycerol, propylene glycol, polyethylene glycol, and the like), and suitable mixtures thereof, vegetable oils, such as olive oil, and injectable organic esters, such as ethyl oleate. Proper fluidity can be maintained, for example, by the use of coating
materials, such as lecithin, by the maintenance of the required particle size in the case of
dispersions, and by the use of surfactants.

These compositions may also contain adjuvants such as preservatives, wetting
agents, emulsifying agents and dispersing agents. Prevention of the action of
microorganisms may be ensured by the inclusion of various antibacterial and antifungal
agents, for example, paraben, chlorobutanol, phenol sorbic acid, and the like. It may also
be desirable to include isotonic agents, such as sugars, sodium chloride, and the like into
the compositions. In addition, prolonged absorption of the injectable pharmaceutical
form may be brought about by the inclusion of agents which delay absorption such as
aluminum monostearate and gelatin.

In some cases, in order to prolong the effect of a drug, it is desirable to slow the
absorption of the drug from subcutaneous or intramuscular injection. This may be
accomplished by the use of a liquid suspension of crystalline or amorphous material
having poor water solubility. The rate of absorption of the drug then depends upon its
rate of dissolution which, in turn, may depend upon crystal size and crystalline form.
Alternatively, delayed absorption of a parenterally-administered drug form is
accomplished by dissolving or suspending the drug in an oil vehicle.

Injectable depot forms are made by forming microencapsule matrices of vitamin
D$_3$ compound(s) in biodegradable polymers such as polylactide-polyglycolide.

Depending on the ratio of drug to polymer, and the nature of the particular polymer
employed, the rate of drug release can be controlled. Examples of other biodegradable
polymers include poly(orthoesters) and poly(anhydrides). Depot injectable formulations
are also prepared by entrapping the drug in liposomes or microemulsions which are
compatible with body tissue.

When the vitamin D$_3$ compound(s) are administered as pharmaceuticals, to
humans and animals, they can be given per se or as a pharmaceutical composition
containing, for example, 0.1 to 99.5% (more preferably, 0.5 to 90%) of active ingredient
in combination with a pharmaceutically-acceptable carrier.

Regardless of the route of administration selected, the vitamin D$_3$ compound(s),
which may be used in a suitable hydrated form, and/or the pharmaceutical compositions
of the present invention, are formulated into pharmaceutically-acceptable dosage forms
by conventional methods known to those of skill in the art.
Actual dosage levels and time course of administration of the active ingredients in the pharmaceutical compositions of the invention may be varied so as to obtain an amount of the active ingredient which is effective to achieve the desired therapeutic response for a particular patient, composition, and mode of administration, without being toxic to the patient. An exemplary dose range is from 0.1 to 10 mg per day.

A preferred dose of the vitamin D₃ compound for the present invention is the maximum that a patient can tolerate and not develop serious hypercalcemia. Preferably, the vitamin D₃ compound of the present invention is administered at a concentration of about 0.001 µg to about 100 µg per kilogram of body weight, about 0.001 – about 10 µg/kg or about 0.001 µg – about 100 µg/kg of body weight. Ranges intermediate to the above-recited values are also intended to be part of the invention.

**Exemplification of the Invention**

The invention is further illustrated by the following examples which should in no way should be construed as being further limiting.

**Synthesis of Compounds of the Invention**

*Experimental*

All operations involving vitamin D₃ analogs were conducted in amber-colored glassware in a nitrogen atmosphere. Tetrahydrofuran was distilled from sodium-benzophenone ketyl just prior to its use and solutions of solutes were dried with sodium sulfate. Melting points were determined on a Thomas-Hoover capillary apparatus and are uncorrected. Optical rotations were measured at 25 °C. ¹H NMR spectra were recorded at 400 MHz in CDCl₃ unless indicated otherwise. TLC was carried out on silica gel plates (Merck PF-254) with visualization under short-wavelength UV light or by spraying the plates with 10% phosphomolybdic acid in methanol followed by heating. Flash chromatography was carried out on 40-65 µm mesh silica gel.

Preparative HPLC was performed on a 5×50 cm column and 15-30 µm mesh silica gel at a flow rate of 100 ml/minute. The results are summarized in Table 1 for examples 1-10 and 19 (C₂₀-natural), and Table 2 for examples 11-18 (C₂₀-cyclopropyl).
EXAMPLE 1

Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (2)

![Chemical structure](image)

The starting material 1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (1) can be prepared as described in US Patent 5,428,029 to Doran et al.. 3 mg of 1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (1) was dissolved in 0.8 ml of pyridine, cooled to ice-bath temperature and 0.2 ml of acetic anhydride was added and maintained at that temperature for 16 h. Then the reaction mixture was diluted with 1 ml of water, stirred for 10 min in the ice bath and distributed between 5 ml of water and 20 ml of ethyl acetate. The organic layer was washed with 3 x 5 ml of water, once with 5 ml of saturated sodium hydrogen carbonate, once with 3 ml of brine then dried (sodium sulfate) and evaporated. The oily residue was taken up in 1:6 ethyl acetate – hexane and flash-chromatographed using a stepwise gradient of 1:6, 1:4 and 1:2 ethyl acetate - hexane. The column chromatography was monitored by TLC (1:4 ethyl acetate – hexane, spot visualization with phosphomolybdic acid spray), the appropriate fractions were pooled, evaporated, the residue taken up in methyl formate, filtered, then evaporated again to give 23.8 mg of the title compound (2) as a colorless syrup; 400 MHz $^1$H NMR $\delta$ 0.66 (3H, s), 0.90 (1H, m), 1.06 (3H, d, J=7.2 Hz), 1.51 (1H, m), 1.72-1.82 (3H,m), 1.9-2.1 (3H, m), 1.99 (3H, s) 2.04 (3H,s), 2.2-2.3 (3 m), 2.44-2.64 (6H, m), 2.78 (1H, m), 3.01 (1H, s), 5.10 (2H, m), 5.38 (1H, m), 5.43 (1H, d, J=12 Hz), 5.85 (1H, d, J=11.5 Hz), 5.97 (1H, dt, J=12 and 7.3 Hz), 6.25 (1H, d, J= 11.5 Hz).
EXAMPLE 2

Synthesis of 1,3-Di-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (4) and 1,3,25-Tri-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (5)

The starting material 1,25-dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (3) can be prepared as described in US Patents 5,451,574 and 5,612,328 to Baggiiolini et al. 314 mg (0.619 mmole) of 1,25-dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (3) was dissolved in 1.5 ml of pyridine, cooled to ice-bath temperature, and 0.4 ml of acetic anhydride was added. The reaction mixture was kept at room temperature for 7 hours and then for 23 hours in a refrigerator. It was then diluted with 10 ml water and extracted with 30 ml of ethyl acetate. The organic extract was washed with water and brine, dried over sodium sulfate and evaporated. The residue was FLASH chromatographed on a 10 x 140 mm column with 1:6 and 1:4 ethyl acetate-hexane as the mobile phase to give 126 mg of 1,3-Di-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (4), and 248 mg of 1,3,25-Tri-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (5).

EXAMPLE 3

Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-cholecalciferol (7)
A 10-mL round-bottom flask was charged with 40 mg of 1,25-dihydroxy-16-ene-23-yne-cholecalciferol (6). This material was dissolved in 1 mL of pyridine. This solution was cooled in an ice bath then 0.3 mL of acetic anhydride was added. The solution was stirred for 30 min, then refrigerated overnight, diluted with water and transferred to a separatory funnel with the aid of 10 mL of water and 40 mL of ethyl acetate. The organic layer was washed with 4 x 20 mL of water, 10 mL of brine passed through a plug of sodium sulfate and evaporated. The light brown, oily residue was taken up in 1:9 ethyl acetate - hexane then flash chromatographed on a 10x130 mm column using 1:9 ethyl acetate - hexane as mobile phase for fractions 1-5, 1:6 for fractions 6-13 and 1:4 ethyl acetate - hexane for fractions 14-20 (18 mL fractions). Fractions 14-19 contained the main band with Rf0.15 (TLC 1:4). Those fractions were pooled and evaporated to a colorless oil, 0.044 g. The material was taken up in methyl formate, filtered and evaporated to give a colorless, sticky foam, 0.0414 g of the title compound (7).

**EXAMPLE 4**

*Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-cholecalciferol (9)*

0.0468 g of 1,25-Dihydroxy-16,23E-diene-cholecalciferol (8) was dissolved in 1.5 mL of pyridine. This solution was cooled in an ice bath then refrigerated overnight, diluted with 10 mL of water while still immersed in the ice bath, stirred for 10 min and transferred to a separatory funnel with the aid of 10 mL of water and 40 mL of ethyl acetate. The organic layer was washed with 4x20 mL of water, 10 mL of brine passed through a plug of sodium sulfate and evaporated. The light brown, oily residue was taken up in 1:9 ethyl acetate - hexane then flash chromatographed on a 10x130 mm column using 1:9 ethyl acetate - hexane as mobile phase for fractions 1-3 (20 mL
fractions), 1:6 for fractions 6-8 and 1:4 ethyl acetate - hexane for fractions 9-17 (18 mL each). Fractions 11-14 contained the main band with Rf 0.09 (TLC 1:4). Those fractions were pooled and evaporated to a colorless oil, 0.0153 g. This material was taken up in methyl formate, filtered and evaporated, to give 0.014 g of the title compound (9).

EXAMPLE 5

_Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-cholecalciferol (11)_

0.0774 g of 1,25-Dihydroxy-16-ene-cholecalciferol (10) was dissolved in 1.5 mL of pyridine. This solution was cooled in an ice bath then 0.3 mL of acetic anhydride was added. The solution was stirred, refrigerated overnight then diluted with 1 mL of water, stirred for 1 h in the ice bath and diluted with 30 mL of ethyl acetate and 15 mL of water. The organic layer was washed with 4x15 mL of water, once with 5 mL of brine then dried (sodium sulfate) and evaporated. The light brown, oily residue was taken up in 1:9 ethyl acetate - hexane then flash chromatographed on a 10x130 mm column using 1:9 ethyl acetate - hexane as mobile phase for fraction 1 (20 mL fractions), 1:6 for fractions 2-7 and 1:4 ethyl acetate - hexane for fractions 8-13. Fractions 9-11 contained the main band with Rf 0.09 (TLC 1:4 ethyl acetate - hexane). Those fractions were pooled and evaporated to a colorless oil, 0.0354 g. This material was taken up in methyl formate, filtered and the solution evaporated, 0.027 g colorless film, the title compound (11).
EXAMPLE 6

Synthesis of 1,3,25-Tri-O-acetyl-1,25-dihydroxy-16-ene-23-yne-26,27-hexafluorocholecalciferol (13) and 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-26,27-hexafluorocholecalciferol (14)

0.0291 g of 1,25-dihydroxy-16-ene-23-yne-26,27-hexafluorocholecalciferol (12) was dissolved in 1.5 mL of pyridine. This solution was cooled in an ice bath then 0.25 mL of acetic anhydride was added. The solution was stirred for 20 min and kept in a freezer overnight. The cold solution was diluted with 15 mL of water, stirred for 10 min, and diluted with 30 mL of ethyl acetate. The organic layer was washed with 4x15 mL of water, once with 5 mL of brine then dried (sodium sulfate) and evaporated. The light brown, oily residue was taken up in 1:6 ethyl acetate - hexane then flash chromatographed on a 10x110 mm column using 1:6 ethyl acetate - hexane as mobile phase. Fractions 2-3 gave 72.3461 - 72.3285 = 0.0176 g. Evaporation of fractions 6-7 gave 0.0055 g. The residue of fractions 2 - 3 was taken up in methyl formate, filtered and evaporated to give 0.0107 g of the title triacetate (13). The residue of fractions 6-7 was taken up in methyl formate, filtered and evaporated to give 0.0049 g of diacetate (14).
EXAMPLE 7

_Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-25R,26-trifluorocholecalciferol (16)_

1.5 mL of 1,25-dihydroxy-16,23E-diene-25R,26-trifluorocholecalciferol (15) was dissolved in 1.5 mL of pyridine, cooled to ice-bath temperature and 0.4 mL of acetic anhydride was added. The mixture was then refrigerated. After two days the mixture was diluted with 1 mL of water, stirred for 10 min in the ice bath then distributed between 10 mL of water and 30 mL of ethyl acetate. The organic layer was washed with 4x15 mL of water, once with 5 mL of brine then dried (sodium sulfate) and evaporated. The light brown, oily residue was taken up in 1:6 ethyl acetate - hexane then flash chromatographed on a 10x130 mm column using 1:6 ethyl acetate - hexane as mobile phase. Fractions 4-6 (TLC, 1:4) contained the main band (see TLC) These fractions were evaporated and gave 0.0726 g. This residue was taken up in methyl formate, filtered and evaporated, to give 0.0649 g of colorless foam, the title compound (16).

EXAMPLE 8

_Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-19-nor-cholecalciferol (18)_

- 64 -
0.0535 g of 1,25-Dihydroxy-16-ene-19-nor-cholecalciferol (17) was dissolved in 1.5 mL of pyridine, cooled to ice-bath temperature and 0.3 mL of acetic anhydride was added and the mixture was refrigerated overnight. The solution was diluted with 1 mL of water, stirred for 10 min in the ice bath then distributed between 10 mL of water and 30 mL of ethyl acetate. The organic layer was washed with 4x15 mL of water, once with 5 mL of brine then dried (sodium sulfate) and evaporated. The nearly colorless, oily residue was taken up in 1:6 ethyl acetate - hexane as mobile phase for fractions 1-6 then 1:4 ethyl acetate - hexane was used. Fractions 9-19 (TLC, 1:4 ethyl acetate - hexane, Rf 0.09, see below) were pooled, evaporated, to give 0.0306 g, which was taken up in methyl formate, filtered, then evaporated. It gave 0.0376 of the title compound (18).

EXAMPLE 9

Synthesis of 1,3-Di-O-Acetyl-1,25-dihydroxy-16-ene-23-yne-19-nor-cholecalciferol (20)

50 mg of 1,25-dihydroxy-16-ene-23-yne-19-nor-cholecalciferol (19) was dissolved in 0.8 mL of pyridine, cooled to ice-bath temperature and 0.2 mL of acetic anhydride was added. The mixture was refrigerated for 3 days then diluted with 1 mL of water, stirred for 10 min in the ice bath and distributed between 5 mL of water and 20 mL of ethyl acetate. The organic layer was washed with 4x5 mL of water, once with 3 mL of brine then dried (sodium sulfate) and evaporated. The nearly colorless, oily residue was taken up in 1:6 ethyl acetate - hexane then flash chromatographed on a 15x120 mm column using 1:6 ethyl acetate - hexane as mobile phase for fractions 1-6, 1:4 for fractions 9-12, 1:3 for fractions 13-15 and 1:2 ethyl acetate - hexane for the remaining fractions. Fractions 11-16 (TLC, 1:4 ethyl acetate - hexane, Rf 0.09, see below) were pooled,
evaporated 76.1487 - 76.1260 = 0.0227 g, taken up in methyl formate, filtered, then evaporated. It gave 0.0186 g of the title compound (20).

EXAMPLE 10

*Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-26,27-bishomo-19-nor-cholecalciferol (22)*

0.0726 g of 1,25-dihydroxy-16-ene-23-yne-26,27-bishomo-19-nor-cholecalciferol (21) was dissolved in 0.8 mL of pyridine, cooled to ice-bath temperature and 0.2 mL of acetic anhydride was added. The solution was stirred in the ice-bath then refrigerated overnight. The solution was then diluted with 1 mL of water, stirred for 10 min in the ice bath and distributed between 10 mL of water and 25 mL of ethyl acetate. The organic layer was washed with 3x10 mL of water, once with 5 mL of saturated sodium hydrogen carbonate, once with 3 mL of brine then dried and evaporated, 33.5512 - 33.4654 = 0.0858 g of a tan oily residue that was flash-chromatographed on a 15x120 mm column using 1:6 as mobile phase. Fractions 7-11 (20 mL each) were pooled (TLC 1:4 ethyl acetate - hexane, Rf 0.14) and evaporated, 67.2834 - 67.2654 = 0.018 g. This residue was taken up in methyl formate, filtered and evaporated. It gave 0.0211 g of the title compound (22).
EXAMPLE 11

Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-19-nor-cholecalciferol (24)

0.282 g of 1,25-Dihydroxy-20-cyclopropyl-23-yne-19-nor-cholecalciferol (23) was dissolved in 0.8 mL of pyridine, cooled to ice-bath temperature and 0.2 mL of acetic anhydride was added and the mixture was refrigerated overnight, then diluted with 1 mL of water, stirred for 10 min in the ice bath and distributed between 5 mL of water and 20 mL of ethyl acetate. The organic layer was washed with 3x5 mL of water, once with 5 mL of saturated sodium hydrogen carbonate, once with 3 mL of brine then dried (sodium sulfate) and evaporated. The oily residue was taken up in 1:6 ethyl acetate - hexane then flash chromatographed on a 15x110 mm column using 1:6 ethyl acetate - hexane as mobile phase for fractions 1-4, 1:4 for fractions 5-12, 1:3 for fractions 13-15 ethyl acetate - hexane for the remaining fractions. Fractions 7-12 (TLC, 1:4 ethyl acetate - hexane, Rf 0.13) were pooled, evaporated, the residue taken up in methyl formate, filtered, then evaporated to give 0.023 g of the title compound (24).

EXAMPLE 12

Synthesis of 1,3,25-Tri-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (26) and 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (27)
0.1503 g of 1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-nor-
cholecalciferol (25) was dissolved in 0.8 mL of pyridine, cooled to ice-bath temperature
and 0.2 mL of acetic anhydride was added. The mixture was refrigerated overnight then
diluted with 1 mL of water, stirred for 10 min in the ice bath and distributed between 5
mL of water and 20 mL of ethyl acetate. The organic layer was washed with 3x5 mL of
water, once with 5 mL of saturated sodium hydrogen carbonate, once with 3 mL of brine
then dried (sodium sulfate) and evaporated. The oily residue was taken up in 1:6 ethyl
acetate - hexane then flash chromatographed on a 15x150 mm column using 1:6 ethyl
acetate - hexane as mobile phase for fractions 1-5, 1:4 for the remaining fractions.
Fractions 3-4 and 6-7 were pooled, evaporated, then taken up in methyl formate, filtered,
and evaporated to give 0.0476 g of the title triacetate (26) and 0.04670 g of the title
diacetate (27).

EXAMPLE 13

**Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-cholecalciferol (29)**

![Chemical Structure](image)

0.0369 g of 1,25-dihydroxy-20-cyclopropyl-23-yne-cholecalciferol (28) was dissolved
in 0.8 mL of pyridine, cooled to ice-bath temperature and 0.2 mL of acetic anhydride
was added and the mixture was refrigerated overnight, then diluted with 1 mL of water,
stirred for 10 min in the ice bath and distributed between 5 mL of water and 20 mL of
ethyl acetate. The organic layer was washed with 3x5 mL of water, once with 5 mL of
saturated sodium hydrogen carbonate, once with 3 mL of brine then dried (sodium
sulfate) and evaporated. The oily residue was taken up in 1:6 ethyl acetate - hexane then
flash-chromatographed on a 13x110 mm column using 1:6 ethyl acetate - hexane as
mobile phase for fractions 1-7, 1:4 ethyl acetate - hexane for the remaining fractions.
Fractions 9-11 (TLC, 1:4 ethyl acetate - hexane) were pooled, evaporated, taken up in methyl formate, filtered, then evaporated, to give 0.0099 g of the title compound (29).

**EXEMPLARY 14**

*Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23E-ene-26,27-hexafluoro-19-nor-cholecalciferol (31)*

![Chemical structure](image)

0.0328 g of 1,25-dihydroxy-20-cyclopropyl-23E-ene-26,27-hexafluoro-19-nor-cholecalciferol (30) was dissolved in 0.8 mL of pyridine, cooled to ice-bath temperature and 0.2 mL of acetic anhydride was added. The solution was refrigerated overnight. The solution was then diluted with 1 mL of water, stirred for 10 min in the ice bath and distributed between 5 mL of water and 20 mL of ethyl acetate. (Extraction of the aqueous layer gave no phosphomolybdic acid-detectable material). The organic layer was washed with 3x5 mL of water, once with 5 mL of saturated sodium hydrogen carbonate, once with 3 mL of brine then dried (sodium sulfate) and evaporated, the residue shows Rf 0.25 as the only spot. The oily residue was taken up in 1:6 ethyl acetate - hexane then flash-chromatographed on a 13.5x110 mm column using 1:6 ethyl acetate - hexane as mobile phase for fractions 1-10. Fractions 4-9 were pooled and evaporated, the residue taken up in methyl formate, filtered, then evaporated to give 0.0316 g of the title compound (31).
EXAMPLE 15

Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23Z-ene-26,27-hexafluoro-19-nor-cholecalciferol (33)

0.0429 g of 1,25-dihydroxy-20-cyclopropyl-23Z-ene-26,27-hexafluoro-19-nor-cholecalciferol (32) was dissolved in 0.8 mL of pyridine, cooled to ice-bath temperature and 0.2 mL of acetic anhydride was added. The solution was refrigerated overnight. The solution was then diluted with 1 mL of water, stirred for 10 min in the ice bath and distributed between 7 mL of water and 25 mL of ethyl acetate. The organic layer was washed with 3x5 mL of water, once with 5 mL of saturated sodium hydrogen carbonate, once with 3 mL of brine then dried (sodium sulfate, TLC (1:4 ethyl acetate - hexane shows mostly one spot) and evaporated, flash-chromatographed on a 15x120 mm column using 1:6 as mobile phase. Fractions 3-6 (20 mL each) were pooled and evaporated. The residue was taken up in methyl formate, filtered and evaporated, to give 0.0411 g of the title compound (33).

EXAMPLE 16

Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-cholecalciferol (35)

0.0797 g of 1,25-dihydroxy-20-cyclopropyl-cholecalciferol (34) was dissolved in 0.8 mL of pyridine, cooled to ice-bath temperature and 0.2 mL of acetic anhydride was
added. The solution was refrigerated overnight. The solution was then diluted with 1 mL of water, stirred for 10 min in the ice bath and distributed between 10 mL of water and 25 mL of ethyl acetate. The organic layer was washed with 3x10 mL of water, once with 5 mL of saturated sodium hydrogen carbonate, once with 3 mL of brine then dried and evaporated, to give 0.1061 g of a tan oily residue that was flash-chromatographed on a 15x120 mm column using 1:6 as mobile phase. Fractions 9-16 (20 mL each) were pooled (TLC 1:4 ethyl acetate - hexane, Rf 0.13) and evaporated. This residue was taken up in methyl formate, filtered and evaporated to give 0.0581 g of the title compound (35).

EXAMPLE 17

*Synthesis of 1,3-Di-O-acetyl-1α,25-dihydroxy-16-ene-20-cyclopropyl-19-nor-cholecalciferol (37)*

To the solution of 1α,25-Dihydroxy-16-ene-20-cyclopropyl-19-nor-cholecalciferol (36) (94 mg, 0.23 mmol) in pyridine (3 mL) at 0°C, acetic anhydride (0.5 mL, 5.3 mmol) was added. The mixture was stirred for 1h, refrigerated for 15h. and then was stirred for additional 8h. Water (10 mL) was added and after stirring for 15 min. the reaction mixture was extracted with AcOEt : Hexane 1:1 (25 mL), washed with water (4x 25 mL) and brine (20 mL), dried over Na2SO4. The residue (120 mg) after evaporation of the solvent was purified by FC (15 g, 30% AcOEt in hexane) to give the titled compound (37) (91 mg, 0.18 mmol, 80%). [α]$_{D}^{20}$ = +14.4 c 0.34, EtOH. UV λ max (EtOH): 242 nm (ε 34349), 250 nm (ε 40458), 260 nm (ε 27545); $^1$H NMR (CDCl$_3$): 6.25 (1H, d, J=11.1 Hz), 5.83 (1H, d, J=11.3 Hz), 5.35 (1H, m), 5.09 (2H, m), 2.82-1.98 (7H, m), 2.03 (3H, s), 1.98 (3H, s), 2.00-1.12 (15H, m), 1.18 (6H, s), 0.77 (3H, s), 0.80-0.36 (4H, m); $^{13}$C NMR (CDCl$_3$): 170.73(0), 170.65(0), 157.27(0), 142.55(0), 130.01(0), 125.06(1),...
123.84(1), 115.71(1), 71.32(0), 70.24(1), 69.99(1), 59.68(1), 50.40(0), 44.08(2),
41.40(2), 38.37(2), 35.96(2), 35.80(2), 32.93(2), 29.48(3), 29.31(2), 28.71(2), 23.71(2),
22.50(2), 21.56(3), 21.51(0), 21.44(3), 18.01(3), 12.93(2), 10.53(2); MS HRES.

EXAMPLE 18

Synthesis of 1,3-Di-O-acetyl-1α,25-hydroxy-16-ene-20-cyclopropyl-cholecalciferol (39)

To the solution of 1α,25-Dihydroxy-16-ene-20-cyclopropyl-cholecalciferol (38) (100
mg, 0.23 mmol) in pyridine (3 mL) at 0°C, acetic anhydride (0.5 mL, 5.3 mmol) was
added. The mixture was stirred for 2 h and then refrigerated for additional 15 h. Water
(10 mL) was added and after stirring for 15 min. the reaction mixture was extracted with
AcOEt: Hexane 1:1 (25 mL), washed with water (4x 25 mL), brine (20 mL) and dried
over Na₂SO₄. The residue (150 mg) after evaporation of the solvent was purified by FC
(15 g, 30% AcOEt in hexane) to give the titled compound (39) (92 mg, 0.18 mmol, 78
%). [α]ᵢ%= -14.9 c 0.37, EtOH. UV λmax (EtOH): 208 nm (ε 15949), 265 nm (ε
15745); °H NMR (CDCl₃): 6.34 (1H, d, J=11.3 Hz), 5.99 (1H, d, J=11.3 Hz), 5.47
(1H, m), 5.33 (1H, m), 5.31 (1H, s), 5.18 (1H, m), 5.04 (1H, m), 2.78 (1H, m), 2.64 (1H,
m), 2.40-1.10 (18H, m), 2.05 (3H, s), 2.01 (3H, s), 1.18 (6H, s), 0.76 (3H, s), 0.66-0.24
(4H, m); °C NMR (CDCl₃): 170.76(0), 170.22(0), 157.18(0), 143.02(0), 142.40(0),
131.94(0), 125.31(1), 125.10(1), 117.40(1), 115.22(2), 72.97(1), 71.32(0), 69.65(1),
59.71(1), 50.57(0), 44.07(2), 41.73(2), 38.36(2), 37.10(2), 35.80(2), 29.45(3), 29.35(2),
10.54(2); MS HRES Calculated for C₃₂H₄₆O₃ M+Na 533.3237. Observed M+Na 533.3236.
EXAMPLE 19

Synthesis of 1,3-Di-O-acetyl-1,25-dihydroxy-23-yne-cholecalciferol (41)

0.2007g of 40(0.486 mmol) was dissolved in 2 mL of pyridine. This solution was cooled in an ice bath and 0.6 mL of acetic anhydride was added. The solution was kept in an ice bath for 45 h then diluted with 10 mL of water, stirred for 10 min and equilibrated with 10 mL of water and 40 mL of ethyl acetate. The organic layer was washed with 4×20 mL of water, 10 mL of brine, dried (sodium sulfate) and evaporated. The brown, oily residue was flash chromatographed using 1:19, 1:9, and 1:4 ethyl acetate – hexane as stepwise gradient. The main band with Rf 0.16 (TLC 1:4 acetate –hexane) was evaporated to give 1,3-di-O-acetyl-1,25-dihydroxy-23-yne-cholecalciferol (41) a colorless foam, 0.0939 g.

Biological Assays and Data

As described in the following examples, the Inventors’ finding that calcitriol and Vitamin D₃ analogues can have an effect on the growth and function of bladder cells has been proven in in vitro models by culturing human stromal bladder cells and has been confirmed in a preclinical in vivo validated model.

EXAMPLE 20

Determination of Maximum Tolerated Dose (MTD)

The maximum tolerated dose of the vitamin D₃ compounds of the invention were determined in eight week-old female C57BL/6 mice (3 mice/group) dosed orally (0.1 ml/mouse) with various concentrations of Vitamin D₃ analogs daily for four days. Analogs were formulated in miglyol for a final concentration of 0.01, 0.03, 0.1 0.3, 1, 3, 10, 30, 100
and 300 μg/kg when given at 0.1 ml/mouse p.o. daily. Blood for serum calcium assay was drawn by tail bleed on day five, the final day of the study. Serum calcium levels were determined using a colorimetric assay (Sigma Diagnostics, procedure no. 597). The highest dose of analog tolerated without inducing hypercalcemia (serum calcium >10.7 mg/dl) was taken as the maximum tolerated dose (MTD). Table 3 shows the relative MTD for various vitamin D₃ compounds. Notably, compound (2) has an MTD that is more than 300 times greater than compound (1). Similarly, compounds (4) and (5) also have a MTD that is considerably greater than their parent compound (3).

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EXAMPLE 21

**Immunological Assay**

Immature dendritic cells (DC) were prepared as described in Romani, N. et al. (Romani, N. et al. (1996) *J. Immunol. Meth.* 196:137). IFN-γ production by allogeneic T cell activation in the mixed leukocyte response (MLR) was determined as described in Penna, G., et al., *J Immunol.*, 164: 2405-2411 (2000).

Briefly, peripheral blood mononuclear cells (PBMC) were separated from buffy coats by Ficoll gradient and the same number (3x10⁵) of allogeneic PBMC from 2 different donors were co-cultured in 96-well flat-bottom plates. The vitamin D₃ compounds were added to each of the cultures. After 5 days, IFN-γ production in the MLR assay was measured by ELISA and the results expressed as amount (nM) of test compound required to induce 50% inhibition of IFN-γ production (IC₅₀). The results are summarized in Table 3.

<table>
<thead>
<tr>
<th>Compound</th>
<th>MTD (mice) μg/kg</th>
<th>INF-γ IC₅₀ pM</th>
</tr>
</thead>
<tbody>
<tr>
<td>1,25(OH)₂D₃</td>
<td></td>
<td></td>
</tr>
<tr>
<td>1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-norcholecalciferol (1)</td>
<td>0.03</td>
<td>0.3</td>
</tr>
<tr>
<td>1,3-di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-norcholecalciferol (2)</td>
<td>0.1</td>
<td>722.0</td>
</tr>
<tr>
<td>1,25-dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-norcholecalciferol (3)</td>
<td>0.3</td>
<td>1.5</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-norcholecalciferol (4)</td>
<td>10</td>
<td>525.0</td>
</tr>
<tr>
<td>1,3,25-Tri-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-norcholecalciferol (5)</td>
<td>3</td>
<td>499.0</td>
</tr>
<tr>
<td>Compound</td>
<td>Concentration (µg/mL)</td>
<td></td>
</tr>
<tr>
<td>------------------------------------------------------------------------</td>
<td>-----------------------</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-cholecalciferol (7)</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-cholecalciferol (9)</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-cholecalciferol (11)</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>1,3,25-Tri-O-acetyl-1,25-dihydroxy-16-ene-26,27-hexafluoro-cholecalciferol (13)</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-26,27-hexafluoro-cholecalciferol (14)</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-25R,26-trifluoro-cholecalciferol (16)</td>
<td>0.3</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-19-norcholecalciferol (18)</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-Acetyl-1,25-dihydroxy-16-ene-19-norcholecalciferol (20)</td>
<td>30</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-26,27-bishomo-19-norcholecalciferol (22)</td>
<td>100</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-19-norcholecalciferol (24)</td>
<td>100</td>
<td></td>
</tr>
<tr>
<td>1,3,25-Tri-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-norcholecalciferol (26)</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-norcholecalciferol (27)</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-cholecalciferol (29)</td>
<td>30</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23E-ene-26,27-hexafluoro-19-norcholecalciferol (31)</td>
<td>0.3</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23Z-ene-26,27-hexafluoro-19-norcholecalciferol (33)</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-26-cyclopropylcholecalciferol (35)</td>
<td>30</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1α,25-dihydroxy-16-ene-20-cyclopropyl-19-norcholecalciferol (37)</td>
<td>10</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1α,25-hydroxy-16-ene-20-cyclopropylcholecalciferol (39)</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-23-yne-cholecalciferol (41)</td>
<td>1</td>
<td></td>
</tr>
</tbody>
</table>

**EXAMPLE 22**

**Proliferation Assay using Bladder Cancer Cell Lines**

Bladder cancer cell lines (T24, RT112, HT1376 and RT4 are human bladder cancer cell lines; NHEK are normal human keratinocytes) were obtained from the European Collection of Cell Cultures (Salisbury, UK). Cells were plated at 3 x 103 per
well, in flat bottomed 96-well plates in 100 μl of DMEM medium containing: 5% Fetal Clone I, 50 μg/l gentamicin, 1 mM sodium pyruvate and 1% non-essential amino acids. After culturing for 24 h at 37 °C in 5% CO2, to allow cells to adhere to the plates, VDR ligands (compounds (2), (4), (5) and other vitamin D₃ analogs as shown in Table 4) were added at concentrations ranging from 100 μM to 0.3 μM in 100 μl of above-mentioned complete medium. After a further 72 h of culture, cell proliferation was measured using a fluorescence-based proliferation assay kit (CyQuant Cell Proliferation Assay Kit, Molecular Probes, Eugene, OR, USA). The IC50 was calculated from the regression curve of the titration data. The results are shown in Table 4.

<table>
<thead>
<tr>
<th>Compound</th>
<th>ECV (μM)</th>
<th>RT112 (μM)</th>
<th>HT 1376 (μM)</th>
<th>RT4 (μM)</th>
<th>NHEK (μM)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1,25 dihydroxycholecalciferol</td>
<td>54.6</td>
<td>19</td>
<td>50</td>
<td>45</td>
<td>4.9</td>
</tr>
<tr>
<td>1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (1)</td>
<td>58.7</td>
<td>24</td>
<td>56</td>
<td>20</td>
<td>4.4</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (2)</td>
<td>55</td>
<td>1</td>
<td>100</td>
<td>20</td>
<td>9.8</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (4)</td>
<td>29</td>
<td>15</td>
<td>&gt;100</td>
<td>9</td>
<td>0.8</td>
</tr>
<tr>
<td>1,3,25-Tri-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (5)</td>
<td>32</td>
<td>13</td>
<td>&gt;100</td>
<td>7</td>
<td>25.6</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-cholecalciferol (7)</td>
<td>27.7</td>
<td>3</td>
<td>&gt;100</td>
<td>80</td>
<td>4.8</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-cholecalciferol (9)</td>
<td>58.7</td>
<td>14</td>
<td>&gt;100</td>
<td>82</td>
<td>5.1</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-cholecalciferol (11)</td>
<td>59.9</td>
<td>18</td>
<td>&gt;100</td>
<td>75</td>
<td>5</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-26,27-hexafluoro-cholecalciferol (14)</td>
<td>&gt;100</td>
<td>26</td>
<td>&gt;100</td>
<td>19</td>
<td>0.98</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-25R,26-trifluorocholecalciferol (16)</td>
<td>76.6</td>
<td>2</td>
<td>&gt;100</td>
<td>16</td>
<td>3.1</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-19-nor-cholecalciferol (18)</td>
<td>27.9</td>
<td>1</td>
<td>96</td>
<td>8</td>
<td>5.8</td>
</tr>
<tr>
<td>1,3-Di-O-Acetyl-1,25-dihydroxy-16-ene-19-nor-cholecalciferol (20)</td>
<td>26.7</td>
<td>1</td>
<td>&gt;100</td>
<td>9</td>
<td>6.7</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-26,27-bishomo-19-nor-cholecalciferol (22)</td>
<td>81.7</td>
<td>7</td>
<td>&gt;100</td>
<td>9</td>
<td>4.7</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-19-nor-cholecalciferol (24)</td>
<td>21.5</td>
<td>18</td>
<td>&gt;100</td>
<td>7</td>
<td>6.8</td>
</tr>
<tr>
<td>Compound</td>
<td>Inhibition (%)</td>
<td>VDR Expression</td>
<td>Cholecalciferol</td>
<td>HFF</td>
<td>Nor-cholecalciferol</td>
</tr>
<tr>
<td>----------</td>
<td>---------------</td>
<td>----------------</td>
<td>----------------</td>
<td>-----</td>
<td>-------------------</td>
</tr>
<tr>
<td>1,3,25-Tri-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (26)</td>
<td>63.5</td>
<td>15</td>
<td>&gt;100</td>
<td>5</td>
<td>32.8</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (27)</td>
<td>25.5</td>
<td>2/13</td>
<td>43</td>
<td>4/1,8</td>
<td>2.6</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (29)</td>
<td>49.6</td>
<td>8</td>
<td>80</td>
<td>13</td>
<td>5.9</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23E-ene-26,27-hexafluoro-19-nor-cholecalciferol (31)</td>
<td>24.3</td>
<td>7</td>
<td>68</td>
<td>9</td>
<td>0.9</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23Z-ene-26,27-hexafluoro-19-nor-cholecalciferol (33)</td>
<td>&gt;100</td>
<td>16</td>
<td>&gt;100</td>
<td>18</td>
<td>0.9</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-cholecalciferol (35)</td>
<td>52.1</td>
<td>9</td>
<td>67</td>
<td>10</td>
<td>4.7</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1α,25-dihydroxy-16-ene-20-cyclopropyl-19-nor-cholecalciferol (37)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>17.4</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1α25-hydroxy-16-ene-20-cyclopropyl-cholecalciferol (39)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>21.5</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-23-yne-cholecalciferol (41)</td>
<td>40</td>
<td>8</td>
<td>&gt;100</td>
<td>85</td>
<td>4.2</td>
</tr>
</tbody>
</table>

**EXAMPLE 23**

**Inhibition of type 1 diabetes development by VDR ligand administration**

The non-obese (NOD)/Lt mice used for the experiments were purchased from Charles River Laboratories (Calco, Italy). All mice were kept under specific pathogen-free conditions. Glucose levels in the tail venous blood were quantified using a EUROFlash (Lifescan, Issy les Moulineaux, France). A diagnosis of diabetes was after two sequential glucose measurements higher than 200 mg/dl.

1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (2) was dissolved in ethanol (1 mg/ml) and then diluted in miglyol 812. NOD/Lt female mice were dosed orally with vehicle (miglyol 812) alone or vehicle containing (2) (0.1 mg/Kg body weight or 0.2 mg/Kg body weight per os) 5x/week from 8 to 16 weeks of age and glycemia levels were monitored until 27 weeks of age. The incidence of disease was significantly lower in mice treated with compound (2) compared to controls, and the higher dose (0.2 mg/Kg) was the most effective as shown in Figure 1 and in Table 5. About 70% of vehicle-treated controls were diabetic by 27 weeks of age compared to only 30% (at the 0.1 mg/Kg dose) and 40% (at the 0.2 mg/Kg dose) of mice treated with compound (2) from 8 to 16 weeks of age. As shown in
Figure 2 and Table 6, compound (2), when given at the 0.1 mg/Kg or 0.2 mg/Kg dose, did not affect body weight, which suggests a lack of obvious toxic effects.

Table 5. Percent Type 1 Diabetes Mellitus incidence for compound (2).

<table>
<thead>
<tr>
<th>Weeks</th>
<th>Miglyol</th>
<th>Cmpd (2), 0.1 ug/kg p.o.</th>
<th>Cmpd (2), 0.2 ug/kg p.o.</th>
</tr>
</thead>
<tbody>
<tr>
<td>8</td>
<td>3</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>12</td>
<td>3</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>13</td>
<td>3</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>14</td>
<td>3</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>15</td>
<td>20</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>16</td>
<td>20</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>17</td>
<td>20</td>
<td>10</td>
<td>1</td>
</tr>
<tr>
<td>18</td>
<td>20</td>
<td>10</td>
<td>1</td>
</tr>
<tr>
<td>19</td>
<td>20</td>
<td>10</td>
<td>1</td>
</tr>
<tr>
<td>20</td>
<td>40</td>
<td>10</td>
<td>10</td>
</tr>
<tr>
<td>21</td>
<td>60</td>
<td>20</td>
<td>10</td>
</tr>
<tr>
<td>22</td>
<td>60</td>
<td>30</td>
<td>10</td>
</tr>
<tr>
<td>23</td>
<td>60</td>
<td>30</td>
<td>20</td>
</tr>
<tr>
<td>24</td>
<td>60</td>
<td>40</td>
<td>20</td>
</tr>
<tr>
<td>25</td>
<td>60</td>
<td>40</td>
<td>20</td>
</tr>
<tr>
<td>26</td>
<td>60</td>
<td>40</td>
<td>30</td>
</tr>
<tr>
<td>27</td>
<td>70</td>
<td>40</td>
<td>30</td>
</tr>
</tbody>
</table>

Table 6 NOD mouse body weight (g) at two doses of compound (2).

<table>
<thead>
<tr>
<th>Weeks</th>
<th>miglyol</th>
<th>Cmpd (2) 0.1 ug/kg p.o.</th>
<th>Cmpd 0.2 ug/kg p.o.</th>
</tr>
</thead>
<tbody>
<tr>
<td>8</td>
<td>21.49</td>
<td>22.07</td>
<td>21.25</td>
</tr>
<tr>
<td>9</td>
<td>21.92</td>
<td>22.98</td>
<td>21.75</td>
</tr>
<tr>
<td>10</td>
<td>22.23</td>
<td>22.77</td>
<td>22.06</td>
</tr>
<tr>
<td>11</td>
<td>23.3</td>
<td>23.4</td>
<td>22.99</td>
</tr>
<tr>
<td>12</td>
<td>24.24</td>
<td>24.01</td>
<td>24.21</td>
</tr>
<tr>
<td>13</td>
<td>24</td>
<td>23.69</td>
<td>24.4</td>
</tr>
<tr>
<td>14</td>
<td>24.35</td>
<td>24.1</td>
<td>24.34</td>
</tr>
<tr>
<td>15</td>
<td>25.17</td>
<td>24.6</td>
<td>24.53</td>
</tr>
<tr>
<td>16</td>
<td>24.82</td>
<td>25</td>
<td>25.01</td>
</tr>
<tr>
<td>17</td>
<td>25.07</td>
<td>24.6</td>
<td>24.33</td>
</tr>
<tr>
<td>18</td>
<td>24.54</td>
<td>25.1</td>
<td>24.89</td>
</tr>
<tr>
<td>19</td>
<td>24.57</td>
<td>25.6</td>
<td>25.29</td>
</tr>
<tr>
<td>20</td>
<td>24.66</td>
<td>25.2</td>
<td>24.9</td>
</tr>
<tr>
<td>21</td>
<td>26.2</td>
<td>25.6</td>
<td>25.41</td>
</tr>
</tbody>
</table>
EXEMPLARY 24

The activity of Calcitriol and Vitamin D₃ analogues on the growth and function of bladder cells

The Inventors’ finding that calcitriol and Vitamin D₃ analogues can have an effect on the growth and function of bladder cells has been proven in in vitro models by culturing human stromal bladder cells. The Inventors confirmed the presence of vitamin D receptors (VDRs), as previously reported in the literature, on these cells (see below in Figure 3).

In these models, calcitriol (the activated form of vitamin D₃) and other vitamin D₃ analogues have been shown to be effective in inhibiting the basal (Fig 4) growth of bladder cells. This activity, never reported before, is dose dependent with an IC₅₀ of 9.8 ±7×10⁻¹⁵ for calcitriol (1,25-dihydroxycholecalciferol) (on basal cells).

A similar investigation was performed on a number of other vitamin D compounds and the results (expressed as -Log IC₅₀) are shown in the table below. Data in the table refers to inhibitors effect of the compound on basal human bladder cell growth in cells which are not stimulated with testosterone or (in one case) are stimulated. The maximum tolerated dose (MTD) in rats is also listed for each compound (Table 7).

<table>
<thead>
<tr>
<th>Compound</th>
<th>-Log IC₅₀</th>
<th>MTD (ug/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-26,27-hexafluoro-19-nor-cholecalciferol (2)</td>
<td>7.4±0.57</td>
<td>0.1</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-cholecalciferol (7)*</td>
<td>10.3±0.26</td>
<td>10</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-cholecalciferol (9)*</td>
<td>11.38±0.39</td>
<td>3</td>
</tr>
<tr>
<td>Compound</td>
<td>IC₅₀ (μM) ± SEM</td>
<td>IC₅₀ (μM)</td>
</tr>
<tr>
<td>----------------------------------------------------</td>
<td>----------------</td>
<td>-----------</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-cholecalciferol (11)*</td>
<td>9.65±0.36</td>
<td>1</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-26,27-hexafluoro-cholecalciferol (14)*</td>
<td>8.7±0.27</td>
<td>10</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-19-norcholecalciferol (18)*</td>
<td>9.2±0.5</td>
<td>3</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-19-norcholecalciferol (20)</td>
<td>3.73±2.3</td>
<td>30</td>
</tr>
<tr>
<td>1,3,25-Tri-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-norcholecalciferol (26)</td>
<td>7.4±0.77</td>
<td>10</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-norcholecalciferol (27)*</td>
<td>8.92±0.29</td>
<td>10</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-ene-26,27-hexafluoro-19-norcholecalciferol (29)</td>
<td>&gt;2</td>
<td>30</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23E-ene-26,27-hexafluoro-19-norcholecalciferol (31)*</td>
<td>8.8±0.4</td>
<td>0.3</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23Z-ene-26,27-hexafluoro-19-norcholecalciferol (33)</td>
<td>6.7±0.36</td>
<td>10</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropylcholecalciferol (35)</td>
<td>6.4±1</td>
<td>30</td>
</tr>
<tr>
<td>1,3-Di-O-acetyl-1,25-dihydroxy-23-yne-cholecalciferol (41)</td>
<td>&gt;2</td>
<td>1</td>
</tr>
</tbody>
</table>

Compounds marked in the table with an asterisk (*) are those which are of particular interest in the context of the invention (these having the highest -LogIC₅₀ values for unstimulated cells).
EXAMPLE 25

*Evaluation of the effect of Vitamin D₃ analogues on bladder function in an in vivo model – cyclophosphamide (CYP) induced chronic IC in rats*

The rat model of chemical cystitis induced by intraperitoneal injection of CYP has been well accepted. CYP is used in clinical practice in the treatment of a number of malignant tumors. One of its metabolites, acrolein, is excreted in urine in large concentrations causing hemorrhagic cystitis associated with symptoms of urinary frequency, urgency and pelvic pain. The inflammatory process is characterized by changes in gross histology of bladder, increase in number and distribution of inflammatory cell infiltrates (mast cells, macrophage, PMNs), cyclo-oxygenase-2 expression and prostaglandin production, growth factor and cytokine production. The rat model of chemical cystitis closely resembles interstitial cystitis, a chronic, painful urinary bladder syndrome and has been used for the testing of therapeutic agents in the past.

This model was used to test the effects of 1,25-dihydroxyvitamin D₃ analogue in rats with CYP-induced cystitis. The effects of the treatment on the cystometric parameters in a conscious freely moving rat with CYP-induced cystitis were monitored. The following cystometric parameters were recorded in each animal: bladder capacity, filling pressure (pressure at the beginning of the bladder filling), threshold pressure (bladder pressure immediately prior to micturition), micturition pressure (the maximal bladder pressure during micturition), presence or absence of non-voiding bladder contractions (increases in bladder pressure of at least 10 cm H₂O without release of urine), and amplitude of non-voiding bladder contraction.

Animals: Wistar rats weighing 125-175g were used. Two groups of animals had a tube implanted into the urinary bladder for intravesical pressure recording. Following recovery all animals received three intraperitoneal injections of CYP and subsequently were divided into the treatment and sham control groups.

Treatment group: Rats treated with oral 1,25-dihydroxyvitamin D₃ analogue 1,3-di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (2) for 14 days (daily dose of 0.1µg/kg)
Control group: Rats treated with oral vehiculum (miglyol) in the dose identical to that delivered in the treatment group.

Cystometry was performed 24 hours following the last dose of the drug or vehiculum on awake freely moving animals. There were four Sham control animals and three Treated animals.

Methods

Implantation of the polyethylene tubing into the urinary bladder:

A lower midline abdominal incision was performed under general inhalation anesthesia (isoflurine with O₂) and polyethylene tubing (PE-50, Clay Adams, Parsippany, NJ) with the end flared by heat was inserted into the dome of the bladder and secured in place with a 6-0 prolene purse string suture. The distal end of the tubing was heat-sealed, tunneled subcutaneously and externalized at the back of the neck, out of the animal’s reach. Abdominal and neck incisions were closed with 4-0 nylon sutures.

Intraperitoneal injection of cyclophosphamide:

Following recovery (5 days) subject animals underwent three intraperitoneal injections of CYP (Sigma Chemical, St. Louis, MO; 75 mg/kg each, intraperitoneal) over the period of nine days. On the tenth day following the first CYP injection the sham control animals received the vehicle only, whereas the experimental group were treated with the 1,25-dihydroxyvitamin D₃ analogue 1,3-di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (delivered using gavage). Two weeks following the initiation of the treatment animals underwent a conscious cystometrogram to assess the function of the urinary bladder.
Cystometrogram:

An animal was placed unrestrained in a cage and the catheter was connected via a T-tube to a pressure transducer (Grass® Model PT300, West Warwick, RI) and microinjection pump (Harvard Apparatus 22, South Natick, MA). A 0.9% saline solution was infused at room temperature into the bladder at a rate of 10 ml/h. Intravesical pressure was recorded continuously using a Neurodata Acquisition System (Grass® Model 15, Astro-Med, Inc, West Warwick, RI). At least three reproducible micturition cycles were recorded after the initial stabilization period of 25 - 30 min.

Timeline of an experiment:

<table>
<thead>
<tr>
<th>Procedure</th>
<th>Days</th>
</tr>
</thead>
<tbody>
<tr>
<td>Acclimation period</td>
<td>1 – 5</td>
</tr>
<tr>
<td>Tube implantation + recovery period</td>
<td>6 – 10</td>
</tr>
<tr>
<td>CYP treatment (three doses of 75mg/kg i.p. every three days)</td>
<td>11 – 17</td>
</tr>
<tr>
<td>Treatment (sham or active)</td>
<td>18 – 31</td>
</tr>
<tr>
<td>Cystometric evaluation</td>
<td>32</td>
</tr>
</tbody>
</table>

Results

The data analysis is summarized in Tables 8 and 9 and Figure 5.

Table 8: Cystometric parameters for the control group

<table>
<thead>
<tr>
<th>Rat</th>
<th>Bl. Cap.</th>
<th>FP</th>
<th>TP</th>
<th>MP</th>
<th># of NVBC</th>
<th>Amplitude of NVBC</th>
</tr>
</thead>
<tbody>
<tr>
<td>RB 8</td>
<td>1,2</td>
<td>15</td>
<td>15</td>
<td>100</td>
<td>22</td>
<td>15</td>
</tr>
<tr>
<td></td>
<td>1,2</td>
<td>13</td>
<td>18</td>
<td>100</td>
<td>14</td>
<td>14</td>
</tr>
<tr>
<td></td>
<td>1,1</td>
<td>16</td>
<td>15</td>
<td>82</td>
<td>12</td>
<td>11</td>
</tr>
<tr>
<td>RB10</td>
<td>0,7</td>
<td>30</td>
<td>40</td>
<td>110</td>
<td>26</td>
<td>25</td>
</tr>
<tr>
<td></td>
<td>0,9</td>
<td>32</td>
<td>26</td>
<td>94</td>
<td>32</td>
<td>28</td>
</tr>
<tr>
<td></td>
<td>0,6</td>
<td>26</td>
<td>26</td>
<td>108</td>
<td>35</td>
<td>16</td>
</tr>
<tr>
<td>RB12</td>
<td>1,7</td>
<td>35</td>
<td>40</td>
<td>115</td>
<td>40</td>
<td>17</td>
</tr>
<tr>
<td>Rat</td>
<td>Bl. Cap.</td>
<td>FP</td>
<td>TP</td>
<td>MP</td>
<td># of NVBC</td>
<td>amplitude of NVBC</td>
</tr>
<tr>
<td>------</td>
<td>---------</td>
<td>-----</td>
<td>-----</td>
<td>-----</td>
<td>-----------</td>
<td>------------------</td>
</tr>
<tr>
<td>RB7</td>
<td>0.7</td>
<td>13</td>
<td>14</td>
<td>98</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>0.7</td>
<td>14</td>
<td>14</td>
<td>97</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>0.8</td>
<td>13</td>
<td>14</td>
<td>101</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>RB13</td>
<td>1.4</td>
<td>14</td>
<td>15</td>
<td>104</td>
<td>8</td>
<td>11</td>
</tr>
<tr>
<td></td>
<td>1.9</td>
<td>15</td>
<td>16</td>
<td>105</td>
<td>4</td>
<td>10</td>
</tr>
<tr>
<td></td>
<td>1.3</td>
<td>14</td>
<td>17</td>
<td>97</td>
<td>8</td>
<td>11</td>
</tr>
<tr>
<td>RB15</td>
<td>2.5</td>
<td>12</td>
<td>14</td>
<td>90</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>1.3</td>
<td>11</td>
<td>12</td>
<td>100</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td></td>
<td>1.5</td>
<td>10</td>
<td>11</td>
<td>108</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

Bl. Cap = bladder capacity (ml), FP = filling pressure (cmH$_2$O), TP = threshold pressure (cmH$_2$O), MP = micturition pressure (cmH$_2$O), # of NVBC = number of non-voiding bladder contractions, amplitude of NVBC = amplitude of non-voiding bladder contraction

Table 9: Cystometric parameters for the treatment group

Changes were noted in a number of cystometric parameters. Dramatic reductions in both the number and amplitude of non-voiding bladder contractions were observed in the drug treated animals. Less pronounced but still statistically significant reductions in the filling and threshold pressures were also recorded. The treatment did not result in a change of the bladder capacity.

Bladder overactivity associated with chronic cystitis manifests itself in frequent contractions of the bladder wall associated with irritative often painful urinary symptoms. The fact that non-voiding bladder contractions were reduced both in their frequency and amplitude strongly suggest that if the effects on the bladder function in patients with interstitial cystitis will be similar, treatment (e.g. oral treatment) with
vitamin D₃ analogues has a potential to relieve these debilitating symptoms. Reduction in filling and threshold pressures is significant from a clinical standpoint because the increased intravesical pressure associated with interstitial cystitis is a condition potentially jeopardizing the upper urinary tract.

This example provides a further demonstration that a vitamin D₃ analogue, 1,3-di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (2), has the ability to treat bladder dysfunction.

**Abbreviations**

<table>
<thead>
<tr>
<th>Abbreviation</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>T</td>
<td>testosterone</td>
</tr>
<tr>
<td>DHT</td>
<td>dihydrotestosterone</td>
</tr>
<tr>
<td>GF</td>
<td>growth factor</td>
</tr>
<tr>
<td>BPH</td>
<td>benign prostatic hyperplasia</td>
</tr>
<tr>
<td>BOO</td>
<td>Bladder Outlet Obstruction</td>
</tr>
<tr>
<td>AR</td>
<td>Androgen receptors</td>
</tr>
<tr>
<td>PSA</td>
<td>Prostate Specific Antigen</td>
</tr>
<tr>
<td>VDR</td>
<td>Vitamin D receptor</td>
</tr>
<tr>
<td>hBC</td>
<td>human bladder cells</td>
</tr>
<tr>
<td>KGF</td>
<td>keratinocyte growth factor</td>
</tr>
</tbody>
</table>

**EXAMPLE 26**

*Effects on BPH cells in vitro*

**MATERIALS AND METHODS**

**Materials**

- Minimum Essential Medium (MEM), DMEM-F12 1:1 mixture, Ham's F12 medium, phosphate buffered saline (PBS), bovine serum albumin (BSA) fraction V, glutamine, geneticine, collagenase type IV, calcitriol, testosterone (T), dihydrotestosterone (DHT), cyproterone acetate, and a kit for measuring calcemia were purchased from Sigma (St. Louis, MO). Plastic ware for cell cultures was purchased from Falcon (Oxnard, CA). Disposable filtration units for growth media preparation were purchased from PBI International (Milan, Italy).

**BPH cells**

- Human BPH cells, prepared, maintained and used as previously described in Crescioli C, *et al.* *Journal of Clinical and Endocrinology Metabolism* (2000) 85 p 2576-2583, were obtained from prostate tissues derived from 5 patients, who underwent
suprapubic adenomectomy for BPH, after informed consent and approval by the Local Ethical Committee. Patients did not receive any pharmacological treatment in the 3 months preceding surgery.

**BPH cell proliferation assay**

For all cell proliferation assays, 4x10^4 BPH cells were seeded onto 12-well plates in their growth medium, starved in red- and serum-free medium containing 0.1% BSA for 24 h, and then treated with specific stimuli for 48 h. Cells in phenol red- and serum-free medium containing 0.1% BSA were used as controls. Thereafter, cells were trypsinized, and each experimental point was derived from hemocytometer counting, averaging at least six different fields for each well, as previously reported (see Crescioli C, *et al.* *Journal of Clinical and Endocrinology Metabolism* (2000) 85 p 2576-2583).

Experiments were performed using increasing concentrations (10^{-18}-10^{-7}M) of calcitriol or vitamin-D analogs with or without a fixed concentration of T (10 nM), KGF or Des(1-3)IGF-I (10 ng/ml). Growth assays were also carried out using a fixed concentration of androgens (10 nM) with or without vitamin-D analogs (1 nM, 10 nM) or the anti-androgens finasteride (F, 1 nM) and cyproterone acetate (Cyp, 100 nM). Growth assays were also performed using a fixed concentration of T (10 nM) or GFs (10 ng/ml) with or without the vitamin-D analogs. (10 nM).

In the same experiment, each experimental point was repeated in triplicate or quadruplicate and experiments were performed 3 times. Results are expressed as % variation (mean±SEM) over the maximal T or GF-induced stimulation.

**Results**

BPH cell proliferation was significantly increased by testosterone (T). When cell growth was stimulated for 48 h with T the inhibitory effects of vitamin-D analogs were even more pronounced (Table 10).
Table 10. Inhibition of testosterone-induced human BPH cell proliferation by vitamin D analogs expressed as -LogIC50. The maximum tolerated dose (MTD) of each compound (i.e. the highest non-hypercalcemic dose) in mice is shown.

<table>
<thead>
<tr>
<th>Compound</th>
<th>-Log IC50</th>
<th>MTD in CD1 Mouse μg/kg</th>
</tr>
</thead>
<tbody>
<tr>
<td>Calcitriol</td>
<td>7.07 +/- 1.7</td>
<td>1</td>
</tr>
<tr>
<td>(7)</td>
<td>4.37 +/- 0.52</td>
<td>10</td>
</tr>
<tr>
<td>(9)</td>
<td>8.04 +/- 1.14</td>
<td>3</td>
</tr>
<tr>
<td>(11)</td>
<td>9.12 +/- 0.38</td>
<td>1</td>
</tr>
<tr>
<td>(14)</td>
<td>3.3 +/- 0.61</td>
<td>10</td>
</tr>
<tr>
<td>(18)</td>
<td>10.65 +/- 0.63</td>
<td>3</td>
</tr>
<tr>
<td>(20)</td>
<td>12.7 +/- 0.47</td>
<td>30</td>
</tr>
<tr>
<td>(26)</td>
<td>&gt;1</td>
<td>10</td>
</tr>
<tr>
<td>(27)</td>
<td>9.1 +/- 0.52</td>
<td>10</td>
</tr>
<tr>
<td>(29)</td>
<td>8.5 +/- 0.89</td>
<td>30</td>
</tr>
<tr>
<td>(35)</td>
<td>2.9 +/- 1.6</td>
<td>30</td>
</tr>
<tr>
<td>(41)</td>
<td>9.14 +/- 0.56</td>
<td>1</td>
</tr>
</tbody>
</table>

**EXAMPLE 27**

*Soft Gelatin Capsule Formulation I*

<table>
<thead>
<tr>
<th>Item</th>
<th>Ingredients</th>
<th>mg/Capsule</th>
</tr>
</thead>
<tbody>
<tr>
<td>10</td>
<td>1 Compound (2) from Example 1</td>
<td>10.001-0.02</td>
</tr>
<tr>
<td></td>
<td>2 Butylated Hydroxytoluene (BHT)</td>
<td>0.016</td>
</tr>
<tr>
<td></td>
<td>3 Butylated Hydroxyanisole (BHA)</td>
<td>0.016</td>
</tr>
<tr>
<td></td>
<td>4 Miglyol 812 gs.</td>
<td>160.0</td>
</tr>
</tbody>
</table>

15 Manufacturing Procedure:

1. BHT and BHA is suspended in Miglyol 812 and warmed to about 50 °C with stirring, until dissolved.

2. 1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-norcholecalciferol is dissolved in the solution from step 1 at 50 °C.

3. The solution from Step 2 is cooled at room temperature.

4. The solution from Step 3 is filled into soft gelatin capsules.
Note: All manufacturing steps are performed under a nitrogen atmosphere and protected from light.

EXAMPLE 28

Soft Gelatin Capsule Formulation II

<table>
<thead>
<tr>
<th>Item</th>
<th>Ingredients</th>
<th>mg/Capsule</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Compound(2) from Example 1</td>
<td>10.001-0.02</td>
</tr>
<tr>
<td>2</td>
<td>di-α-Tocopherol</td>
<td>0.016</td>
</tr>
<tr>
<td>3</td>
<td>Miglyol 812 q.s.</td>
<td>160.0</td>
</tr>
</tbody>
</table>

Manufacturing Procedure:
1. Di-α-Tocopherol is suspended in Miglyol 812 and warmed to about 50 °C with stirring, until dissolved.
2. 1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-norcholecalciferol is dissolved in the solution from step 1 at 50 °C.
3. The solution from Step 2 is cooled at room temperature.
4. The solution from Step 3 is filled into soft gelatin capsules.

Incorporation by Reference

The contents of all references (including literature references, issued patents, published patent applications, and co-pending patent applications) cited throughout this application are hereby expressly incorporated herein in their entireties by reference.

Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents of the specific embodiments of the invention described herein. Such equivalents are intended with be encompassed by the following claims.
CLAIMS

1. A vitamin D₃ compound of formula I:

\[
\begin{align*}
\text{A}_1 & \text{ is single or double bond;} \\
\text{A}_2 & \text{ is a single, double or triple bond;} \\
\text{X}_1 & \text{ and X}_2 \text{ are each independently H}_2 \text{ or } =\text{CH}_2, \text{ provided X}_1 \text{ and X}_2 \text{ are not both } =\text{CH}_2; \\
\text{R}_1 & \text{ and R}_2 \text{ are each independently OC(O)C}_1\text{-C}_4 \text{ alkyl, OC(O)hydroxyalkyl, or OC(O)haloalkyl;} \\
\text{R}_3, \text{ R}_4 \text{ and R}_5 & \text{ are each independently hydrogen, C}_1\text{-C}_4 \text{ alkyl, hydroxyalkyl, or haloalkyl, with the understanding that R}_5 \text{ is absent when A}_2 \text{ is a triple bond, or R}_3 \text{ and R}_4 \text{ taken together with } \text{C}_{20} \text{ form C}_3\text{-C}_6 \text{ cycloalkyl;} \\
\text{R}_6 & \text{ and R}_7 \text{ are each independently alkyl or haloalkyl; and} \\
\text{R}_8 & \text{ is H, C(O)C}_1\text{-C}_4 \text{ alkyl, C(O)hydroxyalkyl, or C(O)haloalkyl;} \\
& \text{ provided that when A}_1 \text{ is single bond, R}_3 \text{ is hydrogen and R}_4 \text{ is methyl, then A}_2 \text{ is a double or triple bond; and} \\
& \text{ pharmaceutically acceptable esters, salts, and prodrugs thereof.}
\end{align*}
\]

2. The compound of claim 1, wherein X₁ is H₂ and X₂(252,799),(286,809) is =CH₂.

3. The compound of claim 1, wherein X₁ and X₂ are H₂.
4. The compound of any preceding claim, wherein $A_1$ is a single bond.

5. The compound of any preceding claim, wherein $A_1$ is a double bond.

6. The compound of any preceding claim, wherein $A_2$ is a single bond.

7. The compound of any preceding claim, wherein $A_2$ is a double bond.

8. The compound of any preceding claim, wherein $A_2$ is triple bond.

9. The compound of any preceding claim, wherein $R_3$ is hydrogen.

10. The compound of any preceding claim, wherein $R_4$ is $C_1$-$C_4$ alkyl.

11. The compound of any preceding claim, wherein $R_3$ and $R_4$, taken together with $C_{20}$, form $C_3$-$C_6$ cycloalkyl.

12. The compound of any preceding claim, wherein $R_3$ and $R_4$, taken together with $C_{20}$, form cyclopropyl.

13. The compound of any preceding claim, wherein $R_1$ and $R_2$ are each independently $OC(O)C_1$-$C_4$ alkyl.

14. The compound of any preceding claim, wherein $R_1$ and $R_2$ are each $OC(O)CH_3$.

15. The compound of any preceding claim, wherein $R_6$ and $R_7$ are each independently alkyl or haloalkyl.

16. The compound of any preceding claim, wherein $R_6$ and $R_7$ are each independently methyl or trifluoromethyl.
17. The compound of any preceding claim, wherein \( R_6 \) and \( R_7 \) are each methyl.

18. The compound of any preceding claim, wherein \( R_6 \) and \( R_7 \) are each ethyl.

19. The compound of any preceding claim, wherein \( R_6 \) and \( R_7 \) are each trifluoromethyl.

20. The compound of claim 9, wherein \( R_6 \) is methyl and \( R_7 \) is trifluoromethyl.

21. The compound of any preceding claim, wherein \( R_8 \) is H or C(\( \text{O} \))C\( _1 \)-\( C_4 \) alkyl.

22. The compound of any preceding claim, wherein \( R_8 \) is H.

23. The compound of any preceding claim, wherein \( R_8 \) is C(\( \text{O} \))CH\( _3 \).

24. The compound of claim 4, wherein \( A_2 \) is a double bond.

25. The compound of claim 4, wherein \( A_2 \) is a triple bond.

26. The compound of any of claims 24-25, wherein \( R_3 \) is H and \( R_4 \) is C\( _1 \)-C\( _4 \) alkyl.

27. The compound of any of claims 24-26, wherein \( R_4 \) is methyl.
28. The compound of claim 1 having formula I-a

29. The compound of claim 28, wherein X₁ is =CH₂ and X₂ is H₂.

30. The compound of claim 28, wherein X₁ and X₂ are each H₂.

31. The compound of any of claims 28-30, wherein A₁ is a double bond.

32. The compound of any of claims 28-31, wherein A₂ is a single bond.

33. The compound of any of claims 28-31, wherein A₂ is a double bond.

34. The compound of any of claims 28-31, wherein A₂ is triple bond.

35. The compound of any of claims 28-30, wherein A₁ is a single bond and A₂ is a double bond.

36. The compound of any of claims 28-30, wherein A₁ is a single bond and A₂ is a triple bond.

37. The compound of any of claims 28-36, wherein R₈ is H or C(O)CH₃.

38. The compound of any of claims 28-37, wherein R₆ and R₇ are alkyl.
39. The compound of any preceding claim 28-38, wherein R₆ and R₇ are methyl.

40. The compound of any of claims 28-38, wherein R₆ and R₇ are ethyl.

41. The compound of any of claims 28-37, wherein R₆ and R₇ are haloalkyl.

42. The compound of claim 41, wherein R₆ and R₇ are trifluoroalkyl.

43. The compound of claim 41 or 42, wherein R₆ and R₇ are trifluoromethyl.

44. The compound of any of claims 28-37, wherein R₆ is trifluoromethyl and R₇ is methyl.

45. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol:

![Chemical Structure 1]

46. The compound of claim 28, wherein said compound is 1,3,25-Tri-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol:

![Chemical Structure 2]
47. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-19-nor-cholecalciferol:

48. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-19-nor-cholecalciferol:

49. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol:
50. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-26,27-bishomo-19-nor-cholecalciferol:

51. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yne-cholecalciferol:

52. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-cholecalciferol:
53. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-cholecalciferol:

54. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-23-yn-26,27-hexafluoro-cholecalciferol:

55. The compound of claim 1, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16,23E-diene-25R-26-trifluoro-cholecalciferol:
56. The compound of claim 28, wherein said compound is 1,3,25-Tri-O-acetyl-1,25-dihydroxy-16-ene-23-yn-26,27-hexafluoro-cholecalciferol:

57. The compound of claim 28, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-23-yn-cholecalciferol:

58. The compound of claim 1 having formula I-b

59. The compound of claim 58, wherein A₁ is a single bond.

60. The compound of claim 58, wherein A₁ is a double bond.
61. The compound of any claims 58-60, wherein and A₂ is a single bond.

62. The compound of any claims 58-60, wherein A₂ is double bond.

5

63. The compound of any claims 58-60, wherein A₂ is a triple bond.

64. The compound of any claims 58-63, wherein X₁ is =CH₂ and X₂ is H.

10 65. The compound of claims 58-63, wherein X₁ and X₂ are each H.

66. The compound of any claims 58-65, wherein R₈ is H or C(O)CH₃.

67. The compound of any claims 58-65 wherein R₈ is H.

15 68. The compound of any claims 58-67, wherein R₆ and R₇ are alkyl.

69. The compound of any claims 58-67, wherein R₆ and R₇ are methyl.

20 70. The compound of any claims 58-67, wherein R₆ and R₇ are haloalkyl.

71. The compound of any claims 58-67, wherein R₆ and R₇ are trifluoroalkyl.

25 72. The compound of any claims 58-67, wherein R₆ and R₇ are trifluoromethyl.

30
73. The compound of claim 58, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-19-nor-cholecalciferol:

![Chemical Structure]

74. The compound of claim 58, wherein said compound is 1,3,25-Tri-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-nor-cholecalciferol:

![Chemical Structure]

75. The compound of claim 58, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-26,27-hexafluoro-19-nor-cholecalciferol:

![Chemical Structure]
76. The compound of claim 58, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23-yne-cholecalciferol:

![Chemical Structure 1]

577. The compound of claim 58, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-cholecalciferol:

![Chemical Structure 2]

78. The compound of claim 58, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23E-ene-26,27-hexafluoro-19-nor-cholecalciferol:

![Chemical Structure 3]
79. The compound of claim 58, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-20-cyclopropyl-23Z-ene-26,27-hexafluoro-19-nor-cholecalciferol:

80. The compound of claim 58, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-20-cyclopropyl-19-nor-cholecalciferol:

81. The compound of claim 58, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16-ene-20-cyclopropyl-cholecalciferol:

82. A method for treating a subject for a vitamin D₃ associated state, comprising administering to said subject in need thereof an effective amount of a
vitamin D$_3$ compound of any one of claims 1-81, such that said subject is treated for said vitamin D$_3$ associated state.

83. The method of claim 82, wherein said vitamin D$_3$ associated state is an ILT3-associated disorder.

84. The method of claim 83, wherein said ILT3-associated disorder is an immune disorder.

85. The method of claim 84, wherein said immune disorder is an autoimmune disorder.

86. The method of claim 85, wherein said autoimmune disorder is selected from the group consisting of type 1 insulin-dependent diabetes mellitus, adult respiratory distress syndrome, inflammatory bowel disease, dermatitis, meningitis, thrombotic thrombocytopenic purpura, Sjogren's syndrome, encephalitis, uveitis, uveoretinitis, leukocyte adhesion deficiency, rheumatoid arthritis, rheumatic fever, Reiter's syndrome, psoriatic arthritis, progressive systemic sclerosis, primary biliary cirrhosis, pemphigus, pemphigoid, necrotizing vasculitis, myasthenia gravis, multiple sclerosis, lupus erythematosus, polymyositis, sarcoidosis, granulomatosis, vasculitis, pernicious anemia, CNS inflammatory disorder, antigen-antibody complex mediated diseases, autoimmune haemolytic anemia, Hashimoto's thyroiditis, Graves disease, habitual spontaneous abortions, Reynard's syndrome, glomerulonephritis, dermatomyositis, chronic active hepatitis, celiac disease, autoimmune complications of AIDS, atrophic gastritis, ankylosing spondylitis and Addison's disease.

87. The method of claim 84, wherein said immune disorder is transplant rejection.

88. The method of claim 86, wherein said autoimmune disorder is type I insulin dependent diabetes mellitus.

89. The method of claim 82, wherein said vitamin D$_3$ associated state is a disorder characterized by an aberrant activity of a vitamin D$_3$-responsive cell.
90. The method of claim 89, wherein said disorder comprises an aberrant activity of a hyperproliferative skin cell.

5 91. The method of claim 90, wherein said disorder is selected from psoriasis, basal cell carcinoma and keratosis.

92. The method of claim 89, wherein said disorder comprises an aberrant activity of an endocrine cell.

10 93. The method of claim 92, wherein said endocrine cell is a parathyroid cell and the aberrant activity is processing and/or secretion of parathyroid hormone.

94. The method of claim 93, wherein said disorder is secondary hyperparathyroidism.

95. The method of claim 89, wherein said disorder comprises an aberrant activity of a bone cell.

20 96. The method of claim 95, wherein said disorder is selected from osteoporosis, osteodystrophy, senile osteoporosis, osteomalacia, rickets, osteitis fibrosa cystica, and renal osteodystrophy.

97. The method of claim 89, wherein said disorder is cirrhosis or chronic renal disease.

98. The method of claim 82, wherein said vitamin D₃ compound is administered in combination with a pharmaceutically acceptable carrier.

30 99. A method of ameliorating a deregulation of calcium and phosphate metabolism, comprising administering to a subject a therapeutically effective amount of a compound of any one of claims 1 to 81, so as to ameliorate the deregulation of the calcium and phosphate metabolism.
100. The method of claim 99, wherein the deregulation of the calcium and phosphate metabolism leads to osteoporosis.

101. A method of modulating the expression of an immunoglobulin-like transcript 3 (ILT3) surface molecule in a cell, comprising contacting said cell with a compound of any one of claims 1-81 in an amount effective to modulate the expression of an immunoglobulin-like transcript 3 (ILT3) surface molecule in said cell.

102. The method of claim 101, wherein said cell is within a subject.

103. A method of treating an ILT3-associated disorder in a subject, comprising administering to said subject a compound of any one of claims 1-81 in an amount effective to modulate the expression of an ILT3 surface molecule, thereby treating said ILT3-associated disorder in said subject.

104. The method of claim 103, wherein said ILT3-associated disorder is an immune disorder.

105. The method of claim 104, wherein said immune disorder is an autoimmune disorder.

106. The method of claim 105, wherein said autoimmune disorder is type insulin dependent diabetes mellitus.

107. A method of inducing immunological tolerance in a subject, comprising administering to said subject a compound of any one of claims 1-81 in an amount effective to modulate the expression of an ILT3 surface molecule, thereby inducing immunological tolerance in said subject.

108. The method of claim 107, wherein said immunological tolerance is induced in an antigen-presenting cell.
109. The method of claim 108, wherein said antigen-presenting cell is selected from the group consisting of dendritic cells, monocytes, and macrophages.

110. A method of inhibiting transplant rejection in a subject comprising administering to said subject a compound of any one of claims 1-81 in an amount effective to modulate the expression of an ILT3 surface molecule, thereby inhibiting transplant rejection in said subject.

111. The method of claim 110, wherein said transplant is a solid organ transplant.

112. The method of claim 110, wherein said transplant is a pancreatic islet transplant.

113. The method of claim 110, wherein said transplant is a bone marrow transplant.

114. The method of any one of claims 99, 101, 103, 107, or 110, wherein said vitamin D₃ compound is administered to the subject using a pharmaceutically-acceptable formulation.

115. The method of claim 114, wherein said pharmaceutically-acceptable formulation provides sustained delivery of said vitamin D₃ compound to a subject for at least four weeks after the pharmaceutically-acceptable formulation is administered to the subject.

116. A method for modulating immunosuppressive activity by an antigen-presenting cell, comprising contacting an antigen-presenting cell with a compound of any one of claims 1-81 in an amount effective to modulate ILT3 surface molecule expression, thereby modulating said immunosuppressive activity by said antigen-presenting cell.
117. The method of any one of claims 99, 101, 103, 107, or 110, wherein said subject is a mammal.

118. The method of claims 101 or 116, wherein said cell is an antigen-presenting cell.

119. The method of claim 118, wherein said antigen-presenting cell is selected from the group consisting of dendritic cells, monocytes, and macrophages.

120. The method of any one of claims 101, 103, 107, or 110, wherein the expression of said immunoglobulin-like transcript 3 (ILT3) surface molecule is upregulated.

121. The method of any one of claims 82, 99, 101, 103, 107, or 110, wherein said compound is administered orally.

122. The method of any one of claims 82, 99, 101, 103, 107, or 110, wherein said compound is administered intravenously.

123. The method of any one of claims 82, 99, 101, 103, 107, or 110, wherein said compound is administered topically.

124. The method of any one of claims 82, 99, 101, 103, 107, or 110, wherein compound is administered parenterally.

125. The method of any one of claims 82, 99, 101, 103, 107, or 110, wherein said compound is administered at a concentration of 0.001 μg – 100 μg/kg of body weight.

126. The method of claim 125, wherein said compound is 1,3-Di-O-acetyl-1,25-dihydroxy-16,23Z-diene-26,27-hexafluoro-19-nor-cholecalciferol (2).
127. The method of claim 125, wherein said compound is 1,3-Di-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (4).

128. The method of claim 125, wherein said compound is 1,3,25-Tri-O-acetyl-1,25-Dihydroxy-16-ene-23-yne-26,27-hexafluoro-19-nor-cholecalciferol (5).

129. The method of claim 89, wherein the disorder is hypertension.

130. The method of claim 129, wherein the compound suppresses expression of renin, thereby treating the subject for hypertension.

131. The method of claim 89, wherein the disorder is benign prostate hypertrophy.

132. The method of claim 89, wherein the disorder is neoplastic disease.

133. The method of claim 132, wherein the neoplastic disease is selected from the group consisting of leukemia, lymphoma, melanoma, osteosarcoma, colon cancer, rectal cancer, prostate cancer, bladder cancer, and malignant tumors of the lung, breast, gastrointestinal tract, and genitourinary tract.

134. The method of claim 133, wherein the neoplastic disease is bladder cancer.

135. The method of claim 89, wherein the disorder is neuronal loss.

136. The method of claim 135, wherein the disorder is selected from the group consisting of Alzheimer's Disease, Pick's Disease, Parkinson's Disease, Vascular Disease, Huntington's Disease, and Age-Associated Memory Impairment.

137. The method of claim 89, wherein the disorder is characterized by an aberrant activity of a vitamin D₃-responsive smooth muscle cell.
138. The method of claim 137, wherein the disorder is hyperproliferative vascular disease selected from the group consisting of hypertension-induced vascular remodeling, vascular restenosis, and atherosclerosis.

139. The method of claim 137, wherein the disorder is arterial hypertension.

140. A method for preventing or treating bladder dysfunction in a subject in need thereof by administering an effective amount of a compound of any of claims 1-81 thereby to prevent or treat bladder dysfunction in said subject.

141. A method for preventing or treating bladder dysfunction in a subject in need thereof by administering an effective amount of a compound of formula I:

![Chemical Structure](image)

wherein:

A₁ is single or double bond;

A₂ is a single, double or triple bond;

X₁ and X₂ are each independently H₂ or =CH₂, provided X₁ and X₂ are not both

=CH₂;

R₁ and R₂ are each independently OC(O)C₁-C₄ alkyl, OC(O)hydroxyalkyl, or OC(O)haloalkyl;

R₃, R₄ and R₅ are each independently hydrogen, C₁-C₄ alkyl, hydroxyalkyl, or haloalkyl, with the understanding that R₃ is absent when A₂ is a triple bond, or R₃ and R₄ taken together with C₂₀ form C₃-C₆ cycloalkyl;

R₆ and R₇ are each independently alkyl or haloalkyl; and
R₈ is H, C(O)C₁₋₄ alkyl, C(O)hydroxyalkyl, or C(O)haloalkyl; and
pharmaceutically acceptable esters, salts, and prodrugs thereof;
thereby to prevent or treat bladder dysfunction in said subject.

142. The method of claim 140 or 141 wherein the compound is formulated in a
pharmaceutical composition together with a pharmaceutically acceptable diluent or
carrier.

143. The method of any one of claims 140-142, wherein said compound is a Vitamin
D receptor agonist.

144. The method of any one of claims 140-143, wherein said bladder dysfunction is
characterized by the presence of bladder hypertrophy.

145. The method of any one of claims 140-144, wherein said bladder dysfunction is
overactive bladder.

146. The method of any one of claims 140-145, wherein the subject is male.

147. The method of claim 140-146, wherein the male concurrently suffering from
BPH.

148. The method of any one of claims 140-147, wherein the subject is female.

149. The method of any of claims 82-147, wherein the subject is a mammal.

150. The method of any of claims 82-149, wherein the subject is human.

151. A pharmaceutical composition, comprising an effective amount of a
compound of any one of claims 1-81 and a pharmaceutically acceptable diluent or
carrier.

152. The pharmaceutical composition of claim 152, wherein said effective
amount is effective to treat a vitamin D₃ associated state.

153. The pharmaceutical composition of claim 151, wherein said vitamin D₃
associated state is an ILT3-associated disorder.
154. The pharmaceutical composition of claim 152, wherein said vitamin D₃ associated state is a disorder characterized by an aberrant activity of a vitamin D₃-responsive cell.

155. The pharmaceutical composition of claim 152, wherein said vitamin D₃ associated state is bladder dysfunction.

156. A packaged formulation for use in the treatment of a vitamin D₃ associated state, comprising a pharmaceutical composition comprising a compound of any one of claims 1-81 and instructions for use in the treatment of a vitamin D₃ associated state.

157. The package formulation of claim 156, wherein said vitamin D₃ associated state is an IILT3-associated disorder.

158. The packaged formulation of claim 156, wherein said vitamin D₃ associated state is a disorder characterized by an aberrant activity of a vitamin D₃-responsive cell.

159. The packaged formulation of claim 156, wherein said vitamin D₃ associated state is bladder dysfunction.
Figure 1

- Miglyol

(2) 0.1 ug/kg p.o.

(2) 0.2 ug/kg p.o.

% type 1 DM incidence

Weeks of age
Figure 2

- Miglyol
- (2) 0,1 ug/kg p.o.
- (2) 0,2 ug/kg p.o.

Body weight (g)

Weeks of age
INTERNATIONAL SEARCH REPORT

A. CLASSIFICATION OF SUBJECT MATTER

IPC(7) : A61K 31/59; C07C 401/00
US Cl. : 514/167; 552/653

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 514/167; 552/653

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

USPATFULL, USPREPUB, INTERNET VIA A9.com

C. DOCUMENTS CONSIDERED TO BE RELEVANT

<table>
<thead>
<tr>
<th>Category</th>
<th>Citation of document, with indication, where appropriate, of the relevant passages</th>
<th>Relevant to claim No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>X</td>
<td>WO 02/094247 A2 (ADORINI et al) 10 May 2002 (10.05.2002), see the entire document, especially lines 4-25 on page 3, lines 6-17 on page 4, examples, figures, and claims.</td>
<td>1-159</td>
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☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

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Date of the actual completion of the international search: 10 December 2004 (10.12.2004)

Date of mailing of the international search report: 18 JAN 2005

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Form PCT/ISA/210 (second sheet) (January 2004)