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(57) Abstract: Disclosed is the use of lipopeptides and lipoproteins as mucosal adjuvants for various vaccinations via mucous mem-
branes, particularly intranasally. Said lipopeptides represent peptides or proteins substituted with 2,3-diacetoxy(2R)-propyl at the
amino-terminal cystein of a peptide or protein, preferably S-(2,3-bisphosphatidylthio(2R)-propyl)cysteinyll peptides derived from my-
coplasmas. Said peptides are highly effective even in small doses, produce good immunization results, and increase the IgA level,
among others.

(57) Zusammenfassung: Es wird die Verwendung von Lipopeptiden und Lipoproteinen als mukosale Adjuvantien für verschie-
denste Vakzinierungen über die Schleimhäute vorgeschlagen, insbesondere auf intranasalem Wege. Bei den Lipopeptiden handelt
es sich um an der aminoterminalen Cystein eines Peptids oder Proteins 2,3-diacetoxy(2R)-propyl-substituierte Peptide oder Prote-
ine, bevorzugt aus Mykoplasmen abgeleitete S-(2,3-bisphosphatidylthio(2R)-propyl)cysteinyll-peptide. Die angegebenen Peptide sind
bereits in geringen Dosen hochwirksam und führen zu guten Immunisierungsergebnissen und Erhöhung u.a. des IgA Spiegels.

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**Use of a lipopeptide or lipoprotein as adjuvant in
therapeutic or prophylactic vaccination**

5 The invention relates to the use of a lipopeptide or
lipoprotein as adjuvant in therapeutic or prophylactic
vaccination, especially via the mucous membranes, i.e.
as mucosal adjuvant.

10 A third of all deaths each year in the world are even
now caused by infectious diseases, which are
additionally responsible for at least 15% of new
cancers. Infectious diseases are thought also to be
involved in the pathophysiology of various chronic
inflammatory, vascular or degenerative diseases.
15 Infectious diseases are the cause of high costs to the
general public through treatment costs and patients'
absence from work.

20 To combat infectious diseases, in general two routes
are followed, namely therapy and prophylaxis. In this
connection, immunizations have become the most
effective weapon against infectious diseases. However,
there are still many infectious diseases for which no
vaccine is yet available or adequate immunization
25 cannot be achieved. Many vaccines are inadequate
because of low efficiency, serious side effects, low
stability or high costs. There is thus a great need for
novel and improved immunizing substances (vaccines).

30 Vaccines have traditionally been used for prophylaxis
of infectious diseases and are satisfactory in this
area for many diseases. Recent findings suggest that
vaccinations additionally are a very suitable means for
the immunotherapy of other transmissible diseases for
35 which immunization has not yet been done, such as viral
hepatitis, Helicobacter pylori infections, herpes virus
infections etc. A further area of use is the
incorporation of vaccines in immunotherapies and
immunoprophylaxes for autoimmune diseases, inflammatory

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diseases, tumors, allergies and for contraception in humans and animals. In some cases (especially including the last case mentioned), the usability of vaccines appears to be linked to an efficient mucosal administration route and the generation, associated therewith, of a good mucosal immune response.

Most infections are either confined to the mucous membranes, or the pathogens must pass through the mucous membrane in early phases of the infection. The absolute aim with a vaccination must therefore be to obtain not only a systemic but in particular also a mucosal immune response in order thereby primarily to stop both the infection (colonization) and the development of the disease. A good mucosal immune response would distinctly reduce the risk of transmission.

The fact that the systemic and the mucosal immune system partly overlap but are not identical means that parenterally administered vaccines are less effective for protection against mucosal pathogens (McGhee, Mestecky et al., "The mucosal immune system: from fundamental concepts to vaccine development", Vaccine 10, 75-88 (1992)). In fact, parenterally administered vaccines essentially stimulate systemic immune responses, whereas vaccines administered by a mucosal route, i.e. via the mucous membranes, simulate the immune response elicited by natural infections. In this way, mucosal immunization leads to effective systemic and mucosal immune responses.

It is further to be expected that mucosal administration of vaccines will be associated with fewer side effects and be readily accepted by patients. Vaccines are easier to administer by a mucosal route and can be brought better into compliance with vaccination protocols. Provision thereof is associated

with lower costs.

The administration of antigens by a mucosal route has to date been associated with some considerable difficulties. A major problem is that the antigens administered in this way are frequently scarcely immunogenic. This derives from various mechanisms such as (i) increased rate of antigen elimination by non-specific clearance mechanisms of the host (for example siliary activity, peristalsis), (ii) antigen degradation by locally acting enzymes, (iii) antigen alteration and/or structural modification as a result of extreme pH values (acidic medium in the stomach, alkaline in the intestinal tract), (iv) low antigen permeability of the mucous membranes and (v) only very limited access to antigen-presenting cells.

Various strategies have been used to overcome these difficulties, e.g. inclusion or association of the antigens with particles (microparticles, nanoparticles, bacteria or bacterial fragments) as carriers, use of virus-like constructs; use of liposomes or ISCOMS (immunostimulatory complexes) or virosomes, use of transgenic plants, antigen production by attenuated viral or bacterial carriers, either as conventional vectors or as carriers of nucleic acid vaccines and/or administration of these aids with mucosal adjuvants. Despite intensive efforts in this area, to date virtually no sufficiently effective and at the same time well-tolerated mucosal adjuvants have achieved practical use.

Substances referred to as "adjuvants" are those which are added in an immunization to the actual antigen (i.e. the substance which provokes the desired immune response) in order to enhance the humoral and/or cell-mediated immune response ("Lexikon der Biochemie und Molekularbiologie", 1. Band, Spektrum, Akademischer

Verlag¹⁹⁹⁵). The use of many adjuvants is based solely on experience, and the effect can neither be accurately explained nor predicted. The following groups of adjuvants are traditionally used in particular:

5 aluminum hydroxide, emulsions of mineral oils, saponins, detergents, silicon compounds, thiourea, endotoxins of gram-negative bacteria, exotoxins of gram-positive bacteria, killed bacteria or parts thereof.

10

The use of optimal adjuvants plays a crucial role in vaccination. Antigens administered without adjuvant only rarely mediate an adequate immune response. In addition, not only the strength but also the quality of

15 the elicited immune response matters. Stimulation of an incorrect immunization pattern may lead to immunopathological reactions and exacerbation of the symptoms of infection. In this context, the adjuvant can help to assist the desired immune response.

20

Adjuvants approved for humans are limited. One of the few adjuvants accepted by approval authorities for humans is aluminum hydroxide. It is not possible to conclude from the fact that an adjuvant is active on

25 systemic administration, i.e. assists the effect of an antigen, that this also applies to other administration routes. A typical example is aluminum hydroxide which can assist the immunogenicity of a substance on intramuscular, subcutaneous, intraperitoneal or

30 intradermal administration but remains completely ineffective on mucosal administration.

There has been an intensive search in recent years for novel adjuvants, including those for the mucosal

35 administration route. Only a few substances have been found to be able to enhance mucosal responses. Among these, some act as carriers to which the antigens must be bound or fused thereto. Far fewer universally

employable "true" adjuvants which are admixed to the antigens have been found.

True mucosal adjuvants which have been discovered are
5 the heat-labile toxin from *Escherichia coli* and the
cholera toxin from *Vibrio cholerae*. Both have been
described as having activity as mucosal adjuvant
(Holmgren et al, "Cholera toxin and cholera B subunit
as oral-mucosal adjuvant and antigenvector system",
10 Vaccine 1179-1184, 1993 and Douce et al, "Mutants of
Escherichia coli heat-labile toxin lacking ADP-
ribosyltransferase activity act as nontoxic, mucosal
adjuvants", Proc. Natl. Acad. Sci. USA 92, 1644-1648).
However, their intrinsic toxicity and the potential
15 side effects are detrimental to their usability in
connection with human vaccinations. Although non-toxic
derivatives of these molecules have been produced by
genetic engineering, severe, intolerable side effects
are still reported, such as pathological changes in the
20 respiratory mucosa and penetration of the toxin into
the brain (N. Garçon, presentation at the "World Vaccine
Congress", Geneva, 26 to 28 September 1999; and:
van Ginkel, F.B. et al., "cutting edge: The Mucosal
Adjuvant Cholera Toxin Redirects Vaccine Proteins into
25 Olfactory Tissues", J. Immunol. 2000, 165, 4778-4782).

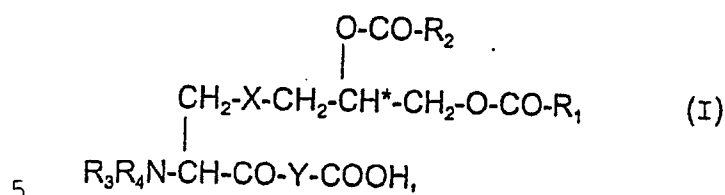
There is thus still a pressing need for novel tolerated
and effective mucosal adjuvants.

30 The invention was therefore based on the object of
developing a range of novel, highly active mucosal
adjuvants which are non-toxic for humans and which can
be employed with a wide variety of active ingredients
to be assisted in conventional or novel vaccines such
35 as, in particular, prophylactic or therapeutic vaccines
including cancer and DNA vaccines.

This object is achieved by providing the use of a

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lipopeptide or lipoprotein of the structure (I) as mucosal adjuvant in therapeutic or prophylactic vaccination via the mucous membranes,



where

R₁ and R₂, which may be identical or different, are C₇₋₂₅-alkyl, C₇₋₂₅-alkenyl or C₇₋₂₅-alkynyl,

10 X is S, O or CH₂,

R₃ and R₄ are independently of one another H or methyl and

Y is a physiologically tolerated amino acid sequence which consists of 1 to 25 amino acid residues and is
 15 not immunogenic per se in the species used,
 and the asymmetric carbon atom marked with * as the absolute R configuration when X is S (sulfur), according to the Cahn-Ingold-Prelog rule.

20 Although DE 19652586 A1 has already mentioned inter alia the use of a particular S-(2,3-dihydroxypropyl)cystein-peptide with two fatty acids linked in ester fashion to the dihydroxypropyl group as vaccine adjuvant, this was purely hypothetical and
 25 without reference to mucosal adjuvants, so that a conventional vaccination route is to be assumed.

The amino acid sequence (Y) linked carboxy-terminally to the 2,3-diacyloxypropyl-substituted amino acid is
 30 preferably selected from the following sequences:

- a) GQTNT,
- b) SKKKK,
- c) GNNDESNISFKEK or
- d) GQTDNNSSQSAAPGSGTTNT.

35

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Particularly preferred at present is an S-[2,3-bispalmitoyloxy-(2R)-propyl]cysteinyl-peptide where the peptide chain may in turn be a physiologically tolerated amino acid sequence which consists of 1 to 25
5 amino acids and is not immunogenic per se in the species used.

According to current knowledge, the carboxy-terminal peptide chain might have a function which controls and,
10 where appropriate, modifies the hydrophilicity or lipophilicity of the lipopeptide, so that it is possible in general to use all peptide or protein chains which - depending on the purpose for which the vaccine is employed - satisfy this criterion. The
15 peptide chain should be physiologically tolerated and, in particular, not itself immunogenic in the species used (i.e. the immunized (vaccinated) species, whether human or animal).

20 At present, the structure according to formula (I) is regarded as crucial for the activity as mucosal adjuvant, with the asymmetric center at the identified point appearing to have a central importance.

25 The mucosal adjuvant of the invention can be linked by all methods known to the skilled worker to the antigen or active molecule intended for the vaccination, be incorporated together with the latter in physical (e.g. microparticles, nanoparticles, liposomes, ISCOMS,
30 polymers) or biological particles (bacteria, bacterial parts) or virosomes or be mixed with the antigen. For binding to carriers it is also possible to provide transport molecules or transport proteins as carriers.

35 The lipoprotein or lipopeptide according to the abovementioned formula (I) which is used according to the invention is preferably present in a preparation with the active vaccination component (e.g. the

antigen) which is suitable and provided for intranasal, intra-NALT (nasal associated lymphoid tissue), aerosolized, oral, intrarectal, conjunctival, intravaginal, intraurethral administration or for
5 administration into the milk ducts of the female breast. Alternatively, the mucosal adjuvant of the invention can be present in a kit for coadministration with a vaccine by one of the aforementioned routes and be adapted therefore where appropriate.

10

The lipopeptide or lipoprotein of the invention is obtained in particular by synthesis. Lipopeptides of the invention might also be obtained by methods generally known in the art from a mycoplasma clone and
15 particularly advantageously from a *Mykoplasma fermentans* clone. The lipopeptides of the invention are also referred to as MALP, namely as "macrophage-activating lipopeptides". These include different variants, of which MALP-2 is a 2kDa lipopeptide
20 according to formula (I) with Y = GNNDESNISFKEK; R₃, R₄ = H and R₁, R₂ = palmitoyl(C15) (S-(2,3-bispalmitoyloxypropyl)-cysteinyl-GNNDESNISFKEK).

Synthesis of the lipopeptides and -proteins of the
25 invention can advantageously be carried out by the method in "Synthesis of N_α-Fmoc protected derivatives of S-(2,3-dihydroxypropyl)-cysteine and their application in peptide synthesis", J.W. Metzger, K.-H. Wiesmüller, G. Jung in: Int. J. Peptide Proteine Res. 38, 1991,
30 545-554". However, the natural (obtained for example from a mycoplasma clone) corresponding lipopeptides are erroneously referred as having the S configuration, although the same compounds ought according to the internationally valid Cahn-Ingold-Prelog nomenclature
35 be referred to as having the R configuration (see, for example, D. Chapman in "Introduction to Lipids", McGraw-Hill, London, 1969, p. 67 zu "Phosphoglycerides"; Burgos et al. J. Org. Chem. 1987,

52 4973-4977; Morr et al. Eur. J. Immunol. 2002, 32:3337-3347).

In a further development of the invention, the
5 lipopeptide or lipoprotein may be present in a preparation with at least one further adjuvant and/or antigen. It is possible in particular for it to be administered together with one or more anti-inflammatory, antiangiogenic, cytotoxic or
10 immunomodulatory substances or ligands (e.g. chemokines, cytokines, CD40 ligand) or with antibodies, or in a preparation therewith.

The lipopeptide or lipoprotein may also, as already
15 mentioned above, be associated or connected to a physical or biological carrier. It may furthermore be present and employed in a preparation with further additives and excipients, in particular preservatives or stabilizers.

20 The invention represents a great advance in the efforts to provide effective mucosal adjuvants: the lipopeptide can be modified within the framework of formula (I) and thus its qualitative and quantitative activity can be
25 varied and adapted to the desired use. It can be prepared at comparatively reasonable cost and is non toxic for humans. Admixture of comparatively small amounts suffices in most cases for distinct enhancement effects. The lipopeptides of the invention are well
30 characterized in terms of their chemical and biochemical properties and can be obtained sufficiently pure in particular by synthetic routes (Mühlradt, P.F., M. Kiess, H. Meyer, R. Sussmuth, G. Jung (1997), Isolation, structure, elucidation, and synthesis or a
35 macrophage stimulatory lipopeptide from Mycoplasma fermentans acting at picomolar concentration"; J. Exp. Med. 185: 1951; and Takeuchi, O., A. Kaufmann, K. Grote, T. Kawai, K. Hoshino, M. Morr, P.F. Mühlradt,

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and S. Akira (2000); "Cutting edge: Preferentially the R-stereoisomer of the mycoplasmal lipopeptide macrophage-activating lipopeptide-2 activates immune cells through a toll-like receptor 2-and MyD88-dependent signaling pathway"; J. Immunol. 164:554), and thus serious side effects are not according to current knowledge to be expected.

A further advantage of the invention is that on immunizations with admixture of the mucosal adjuvant of the invention there were found to be high concentrations of IgA in the mucosal secretions of treated experimental animals. This antibody species is particularly important for protecting the mucous membranes from infections.

Since at present absolutely no effective adjuvants are approved for intranasal immunization of human patients, the invention represents a significant medical advance in this area.

The lipopeptides or lipoproteins of the general structure (I) described in this invention can also be used generally, i.e. by other routes than mucosal, as adjuvants, including particular administrations for DNA vaccines and cancer vaccines, vaccines against non infectious diseases and the like, excluding the previously known MALP-2 peptides, namely S-(2,3-diacyloxypropyl)cystein-peptides of the sequence DhcGNNDESNI SFKEK, where N-terminally the amino acids at positions 2 and, where appropriate, 3 may be absent and/or C-terminally 1 to 2 amino acids may be deleted, for the conventional vaccination routes (intramuscular, subcutaneous, intradermal, intraperitoneal).

The adjuvants of the invention can be combined with a wide variety of antigens to give vaccines. Antigens which can be selected are, in particular, target

antigens for the prophylaxis and treatment of infectious diseases, tumors, autoimmune diseases, allergies, and chronic or acute inflammatory diseases. The selection may be made inter alia from the antigen pool of infective agents such as viruses, bacteria, parasites, rickettsia, mycoplasma, fungi and the like. An immunization also means a treatment with antigens for fertility control in human or animal populations.

10 The advantageous properties of the mucosal adjuvant of the invention which are evident from the examples cannot be inferred from any publication prior to this application.

15 **Methodological approach to the experiments/investigations**

At the outset, in vitro screening studies were initially carried out in order to be able to estimate the potential of the investigated lipopeptides in relation to the activation of antigen-presenting cells. The target cells used were dendritic cells derived from the bone marrow, which were obtained from precursors with the aid of GM-CSF. Entirely in contrast to what could be expected on the basis of the activity of MALP-2 in relation to macrophages, MALP-2 showed only weak activity on primary dendritic cells. Compared with the control samples, which had been incubated in the presence of E. coli lipopolysaccharides (LPS, 10 ng/ml), only a very weak activation of dendritic cells treated with 5 ng/ml MALP-2 was observed.

On the basis of these preliminary investigations it was not possible to expect MALP-2 and corresponding lipopeptides to be suitable as adjuvant, because a certain ability to activate dendritic cells is presumed for adjuvants. Dendritic cells are the principal group of antigen-presenting cells. They play a central part

in the primary immune response, where they (1.) represent the most efficient antigen-presenting cells, (2.) are the most important source of epitopes for specific T-cell clones, and (3.) are the most important
5 activators of resting T cells able to elicit primary immune responses *in vivo*. Whereas LPS-treated dendritic cells show a strong up regulation or multiplication of CD40 including CD80 and CD86, only a weak effect, or none, is noticeable in dendritic cells treated with
10 MALP-2.

This was to some extent to be expected and consistent with the results obtained on macrophages. It had been found with them that, after an initial up regulation
15 phase, as treatment with MALP-2 continued there was an increased turnover and reduced expression of MHC class II molecules which are essential for correct antigen presentation (M. Frisch et al., Eur. J. Immunol. (1996) 26, 1050-1057). The effect found for the adjuvant is
20 all the more surprising.

The results of the preliminary experiments were therefore initially strongly against a possible activity of the lipopeptides and -proteins investigated
25 here as adjuvants.

Although it was obvious from these *in vitro* studies that the lipopeptides in question promised no success in this regard, they were included as negative control
30 samples, namely as examples of macrophage-activating substances of low activity, in *in vivo* studies by the inventors, because the substances were available from earlier experiments.

35 Surprisingly, because in complete contrast to the *in vitro* experiments, it was found that, for example, MALP-2 was able on administration together with the model antigen β -galactosidase by either the intranasal

(i.n.) or intraperitoneal (i.p. (note: in the animal model)) route in a dose of only 0.5 µg per animal per dose, to increase the β-galactosidase-specific IgG serum titers by 675 to 3560 fold (i.n.) and 64 to 128 fold (i.p.). It is possible on use of the lipopeptides of the invention to elicit almost maximum IgG responses even after the first immunization, and these IgG titers correspond to those on administration of 10 µg (three-molar excess) of cholera toxin subunit B (CTB), a well-characterized mucosal adjuvant. Similar results are found on intradermal or subcutaneous administration of the lipopeptides and -proteins of the invention. It is therefore to be assumed that administration of the lipopeptides and -proteins of the invention can and ought to take place in a (molar) concentration which is at least 3 times lower than with conventional adjuvants.

It was also possible to show that administration of the lipopeptides of the invention by the intranasal route leads to an effective stimulation of the mucosal immune system overall. Thus, on administration of MALP-2 as adjuvant to the model antigen, respectively 36% and 23% antigen-specific IgA, based on the total IgA, were found in lung and vaginal lavages. This shows that the lipopeptides of the invention are not only able to elicit local mucosal immune responses, but the spread of IgA-producing cells to other remote areas of mucous membranes takes place to an extent leading to good mucosal immune responses. This effect is linked to the mucosal administration route.

Administration of the lipopeptides of the invention with the antigen also elicited stronger cellular immune responses than did CTB, both regionally in lymph nodes and in the spleen ($p < 0.05$). Analysis of β-galactosidase-specific IgG isotypes and the profiles of the cytokines secreted by cells stimulated *in vitro*

showed that administration with the lipopeptides induced a dominant Th2 response pattern. The results of the investigations therefore prove that the lipopeptides of the invention represent effective
5 adjuvants for mucosal administration of antigens in vaccines.

An important aspect which should be assessed here is that an incremental rise in the lipopeptide dose
10 (starting from the optimal doses mentioned herein) leads to a steady reduction in the immune response. This contrary effect is unexpected and could not have been inferred from the prior art.

15 EXAMPLE SECTION

General

The investigations described in the examples were
20 mainly carried out with a synthetic MALP-2, which substantially corresponds to a lipopeptide derived from mycoplasma, as mucosal adjuvant together with β -galactosidase as model antigen. This specific exemplary lipopeptide was selected on the basis of its previously
25 determined intrinsic properties (biochemical properties, macrophage-stimulating activity). Unless indicated otherwise, the synthetic lipopeptide S-[2,3-bispalmitoyloxypropyl]cysteinyl-GNNDESNISFKEK which was used in the examples is always referred to simply as
30 "MALP-2" hereinafter.

The MALP-2 doses were initially established in preliminary studies in which MALP-2 was administered subcutaneously, intradermally, intranasally or
35 intraperitoneally to mice. In all protocols, simultaneous or associated administration of MALP-2 with β -galactosidase led to a significant increase in the production of β -galactosidase-specific antibodies.

The induced immune responses obtained in the presence of MALP-2 after immunization by the intranasal or intraperitoneal route were then analyzed and compared with those obtained in comparative experiments with CTB as adjuvant.

The examples reveal that use of MALP-2 in a dose of only 0.5 µg per administration led to a significant increase both in the humoral and in the cellular β-galactosidase-specific responses. Even intraperitoneal administration led to an improved immune response, but the intranasal route proved to be far more effective. By comparison therewith, a three-fold molar excess of CTB was necessary to obtain comparable systemic or mucosal humoral responses. Concerning the cellular responses, spleen cells from mice immunized with MALP-2 showed distinctly higher proliferation ($p < 0.05$) than those from animals vaccinated with the three-fold molar excess of CTB.

The primary response obtained with MALP-2 as adjuvant by the intranasal route was characterized in terms of the kinetics of the β-galactosidase-specific antibody response through the presence of high antibody titers, which almost reached the maximum plateau even after the first immunization. Humoral and cellular responses were stronger after i.n. vaccination, indicating a differential local effect of MALP-2, which might be attributed either to its bioavailability or the spatial distribution of the specific receptors in the target cells. As already described, distinct immune responses were found even in mucosal tissues not directly reachable by the vaccination route, namely β-galactosidase-specific IgA in lung and vaginal lavages.

The investigations within the scope of the invention have therefore revealed that the lipopeptides of the invention are novel and potent mucosal adjuvants, it

being unnecessary, in contrast to other lipopeptides, to conjugate the target antigen with the active lipopeptide.

5 Throughout the experiments with the immunized animals, no serious side effects or signs of acute or chronic toxicity were observed. The relatively short peptide unit within the lipopeptides of the invention means that the immunogenicity is relatively low, thus
10 minimizing the risk of immune responses against the lipopeptides themselves. This represents a distinct advantage over proteins as adjuvants. No anti-MALP-2 antibodies were detectable after intranasal administration of MALP-2 as adjuvant. This represents a
15 distinct advantage over proteins as adjuvants, because an immune response against the adjuvant itself may impair a later immune response against another vaccine administered with the same adjuvant. The lipopeptides of the invention can be employed in vaccines against a
20 wide variety of pathogens and in cancer vaccines or vaccines for fertility control. Further advantages of the invention are the good stability of the lipopeptides during storage, greater purity of the lipopeptides which can be prepared by synthesis; and
25 predictable chemical and biochemical behavior.

Key to the figures:

30 Fig. 1. *In vitro* investigations with primary dendritic cells:

Primary dendritic cells from the bone marrow of BALB/c mice were obtained by *in vitro* maturation of precursors using recombinant GM-CSF (5×10^4 U/ml). The mature dendritic cells were stimulated with 10 ng/ml E.coli
35 lipopolysaccharide (LPS) or 5 ng/ml MALP2. The cells were then doubly labeled with CD11c-specific antibodies (dendritic cell markers) in combination with anti-CD40 or anti-CD80 or anti-CD86. Expression of CD40 or CD80

or CD86 in the CD11c-labeled cells was analyzed with the aid of flow cytometry. The results are expressed as a percentage of positive cells relative to the total CD11c-positive population.

5

Fig. 2. Analysis of dendritic cells after treatment with MAPL-2 with the aid of flow cytometry (FACScan):

Primary dendritic cells from the bone marrow of BALB/c mice were obtained by *in vitro* maturation of precursors using recombinant GM-CSF (5×10^4 U/ml). The mature dendritic cells were stimulated with 10 ng/ml E.coli lipopolysaccharide (LPS) or 5 ng/ml MALP2. The cells were then doubly labeled with CD11c-specific antibodies (dendritic cell markers) in combination with anti-CD40 or anti-CD80 or anti-CD86 and the cells were analyzed with the aid of flow cytometry. The gates were set on the basis of a labeling with unrelated control antibody isotopes.

20 Fig. 3. Humoral responses stimulated after immunization with MAPL-2 as adjuvant:

Mice were immunized by subcutaneous (s.c.), intraperitoneal (i.p.), intradermal (i.d.) and intranasal (i.n.) routes with either pure β -galactosidase (40 μ g) or β -galactosidase mixed with MALP-2 (0.5 μ g) on days 0, 7 and 14. On day 28 after the primary immunization, serum samples were removed and the titer of β -galactosidase-specific antibodies was determined by means of an ELISA. The results are shown as reciprocal $\log_2$ of the geometric mean of the endpoint titer. As a control, we included a group in which the animals were immunized by the route with β -galactosidase using aluminum hydroxide as adjuvant.

35 Fig. 4. Humoral responses stimulated after immunization using MALP-2 as adjuvant in a dose of 1 μ g per animal and immunization:

Mice were immunized by the intraperitoneal (i.p.) and

intranasal (i.n.) route with either pure β -galactosidase (40 μ g/dose) or β -galactosidase mixed with MALP-2 (1 μ g/dose) on days 0, 7 and 14. On day 28 after the primary immunization, serum samples were
5 taken and the titer of β -galactosidase-specific antibodies was determined by means of an ELISA. The results are shown as absorbances (OD 405 nm).

Fig. 5. Cellular responses stimulated after
10 immunization using different concentrations of MALP-2 as adjuvant.

β -Galactosidase-specific T-cell proliferation responses of mouse spleen cells immunized by the i.p. or i.n. routes with either pure β -galactosidase (40 μ g/dose) or
15 β -galactosidase mixed with MAPL-2 (1 or 0.5 μ g/dose) on days 0, 7 and 14. On day 28 after the primary immunization, the animals were sacrificed and the spleen cells were restimulated in vitro in the presence of 20 μ g/ml soluble β -galactosidase for four days. The
20 results are shown as stimulation indices (cpm of samples/cpm in unstimulated control cells).

Fig. 6. Kinetics of β -gal-specific IgG responses in sera of immunized mice:

25 Groups of animals (n=5) were immunized either i.n. (A) or i.p. (B) with 50 μ g of β -gal (), β -gal plus 10 μ g of CTB (), β -gal plus 0.5 μ g of MALP-2 () or pure buffer solution (). The days of the immunizations (day 0, 14 and 21) are marked by arrows. The results are shown as
30 the reciprocal $\log_2$ of the geometric endpoint titer, and the SEM (average standard deviation) is indicated by means of vertical lines.

Fig. 7. β -Galactosidase-specific IgA in lung and
35 vaginal lavages from mice immunized i.n.:

The results are shown as percentage of β -galactosidase-specific IgA in relation to the total IgA present. The SEM is indicated by vertical lines.

Fig. 8. Determination of the serum IgE level after immunization using MALP-2 as adjuvant:

Mice were immunized intraperitoneally (i.p.) and
5 intranasally (i.n.) either with pure β -galactosidase (40 μ g), exclusively MALP-2 or β -galactosidase mixed with MALP-2 (0.5 μ g) on days 0, 7 and 14. On day 28 after the first immunization, serum samples were obtained, and the IgE levels were determined by means
10 of a capture ELISA. The results are shown as IgE concentrations (ng/ml).

Fig. 9. β -Galactosidase-specific T-cell proliferation responses of the spleen (A and B) and regional lymph
15 node cells (C) of mice immunized i.p. or i.n.:

The cells were restimulated in vitro with various concentrations of soluble β -galactosidase for four days. The results are shown as average cpm subtracted from the background values for unstimulated cells from
20 groups of three. The SEM is depicted by vertical lines.

Fig. 10. Th profiles stimulated in immunized mice:

Serum- β -galactosidase-specific IgG isotypes and cytokines secreted by spleen cells stimulated in vitro
25 were determined for the immunized mice by means of an ELISA. The results are shown as ratios of IgG1/IgG2a and IL-10/IL-2 (the most commonly found cytokines) concentrations.

30 Fig. 11. Cytokines secreted by cells stimulated in vitro from immunized mice:

The cytokine production was measured by means of an ELISA in the liquid supernatant from cells which were cultured in the presence of β -gal (20 μ g/ml) for 48
35 (IL-2) or 96 hours (IFN γ , IL-4 and IL-10). The results are shown as the ratio between the amounts of cytokines found in the immunized groups compared with the unimmunized control mice.

Fig. 12: Humoral responses stimulated after vaccination with MALP-2 as adjuvant in a dose 2 μ g per animal per immunization. The mice were immunized orally either
5 with β -galactosidase alone (100 μ g/dose) or with β -galactosidase mixed with MALP-2 (2 μ g/dose) on days 0, 7, 14 and 31. On day 45 after the first immunization, serum samples were obtained, and the titers of the β -galactosidase-specific antibodies were determined with
10 an ELISA.

Example 1: In vitro stimulation by MALP-2 of primary dendritic cells obtained from the bone marrow of rodents.

15 *Experimental protocol:* Cultures of primary dendritic cells obtained from the bone marrow were obtained from BALB/c mice after in vitro maturation of precursors in the presence of recombinant GM-CSF (5×10^4 U/ml) by
20 the usual methods. The mature dendritic cells were stimulated with E. coli lipopolysaccharide (LPS) or 5 ng/ml MALP2. After stimulation for respectively 12 and 24 hours, the cells were analyzed with the aid of flow cytometry in order to establish the expression of
25 surface markers important for the antibody presentation ability.

In order to define compounds which have a potential as adjuvant for *in vivo* applications in the area of
30 vaccinations, a first *in vitro* investigation was carried out with primary cultures of dendritic cells derived from bone marrow. Dendritic cells were selected because they are the most efficient antigen-presenting cells and play a key part in the primary immune
35 response. In fact, they are the only cell type which is able *in vivo* to activate resting T cells to initiate a primary immune response. Accordingly, dendritic cells were treated with the investigated units or LPS, which

served as control. Samples were taken at various times, labeled with fluorescence-labeled antibodies which are specific for cellular markers which are decisive for the antigen-presenting properties of the dendritic cells, and analyzed by flow cytometry. The results found (Figs. 1 and 2) show that, differing from the positive control group, the expression of CD40 and the costimulating molecule CD86 in dendritic cells treated with MALP-2 was not increased. The effect on expression of the costimulating molecule CD80 was slight, if present at all. Costimulating molecules emit signals which in addition to the presentation of the epitopes involved within the context of the MHC molecules are essential for effective activation of T cells. It has been reported previously that the effect of proven mucous membrane adjuvants (mucosal adjuvants) such as, for example, cholera toxin is associated with a selective enhancement of the expression of costimulating molecules. Accordingly, the results found *in vitro* strongly indicate that MAPL-2 has little or no potential as mucosal adjuvant.

Example 2: Humoral responses stimulated after immunization by various routes using MALP-2 in various concentrations as adjuvant.

Experimental protocol: Six- to eight-week old female BALB/c (H-2d) mice were purchased from Harlan Winkelmann GmbH (Borchen, Germany) and treated in accordance with local and EU directives. Groups each of 5 mice were immunized on day 1, 7 and 14 either with 40 µg of pure β-galactosidase (Boehringer, Mannheim, Germany), or with admixture of 1 or 0.5 µg of synthetic MALP-2. For the intranasal administration (i.n.), 10 µl was administered into each nostril, whereas for i.p., s.c. and i.d. injection β-galactosidase was resuspended with or without MALP-2 in respectively 400, 100 and 100 µl of PBS. 28 days after the first immunization,

serum samples were taken and stored at -20°C until the β -gal-specific antibodies were determined. Nunc-Immuno MaxiSorp assay plates with 96 wells (Nunc, Roskilde, Denmark) were coated with $100\ \mu\text{l}$ of β -galactosidase (Boehringer, Mannheim, Germany) with $5\ \mu\text{g/ml}$ in $0.05\ \text{M}$ carbonate buffer ($\text{pH}\ 8.2$) per well. Serum dilutions with 1% BSA and 0.05% Tween-20 in PBS were added ($100\ \mu\text{g/well}$), and the plates were incubated at 37°C for 24 hours. After rinsing, biotinylated γ -chain specific goat anti-mouse IgG (Sigma Chemie, Deisenhofen, Germany) was added, and the plates were incubated at 37°C for a further hour. After rinsing four times, $100\ \mu\text{l}$ of peroxidase-conjugated streptavidin (Pharmingen) were added to the cells, and the plates were incubated at 37°C for 30 minutes. After rinsing four times, the reactions were developed by means of ABTS in $0.1\ \text{M}$ citrate-phosphate buffer ($\text{pH}\ 4.35$) which contained 0.01% H_2O_2 . The results have been shown either according to the absorption at $405\ \text{nm}$ or the endpoint titers (reciprocal $\log_2$ of the last dilution which, after incubation for 30 minutes, resulted in an optical density at $405\ \text{nm}$ of 0.1 units above the values for the negative control group).

Despite the disappointing results found in the *in vitro* investigation of primary dendritic cells using MALP-2, we decided to include in the secondary *in vivo* investigation groups of animals immunized with use of MALP-2 as adjuvant. Accordingly, the mice were immunized i.p., s.c., i.d. and i.n. either with the pure model antigen β -galactosidase or with the antigen plus MALP-2. In contrast to expectations, a strong adjuvant effect was found when the antigen was mixed with $0.5\ \mu\text{g}$ of MALP-2, independently of the mode of administration (Fig. 3). The strongest reactions were found with the i.p. and i.n. immunizations. The responses obtained were, however, always at least as strong as (i.d.) or stronger than on use of aluminum

hydroxide as standard adjuvant for s.c. injection (Fig. 3).

Since in preceding investigations using conventional lipopeptides considerably higher concentrations of the component used as adjuvant were given, the effect of higher doses of MALP-2 was to be investigated. For this purpose, animals were immunized with β -galactosidase by the two most effective immunization routes (i.p. and i.n.) with 1 μ g of MALP-2 as adjuvant. In contrast to expectations, increasing the dose of MALP-2 led to an abolition of the adjuvant effect (Fig. 4). This showed that we would not have been able to find an adjuvant effect at the level of the humoral responses in the *in vivo* investigations of MALP-2 on use of the standard concentrations indicated in the literature for other lipopeptides.

Example 3: Humoral responses stimulated after immunization by various routes using MALP-2 in various concentrations as adjuvant.

Experimental protocol: Six- to eight-week old female BALB/c (H-2d) mice were purchased from Harlan Winkelmann GmbH (Borchen, Germany) and treated in accordance with local and EU directives. Groups each of 5 mice were immunized on day 1, 7 and 14 either with 40 μ g of pure β -galactosidase (Boehringer, Mannheim, Germany), or with admixture of 1 or 0.5 μ g of synthetic MALP-2. For the intranasal administration (i.n.), 10 μ l was administered into each nostril, while for the i.p. injection β -galactosidase was resuspended with or without MALP-2 in 400 μ l of PBS. The spleens were removed and put together (pooled) for determination of the cellular immune responses. The cells were cultured in RPMI 1640 supplemented with 10% fetal calf serum, 100 U/ml penicillin, 50 μ g/ml streptomycin, 5×10^{-5} M 2-mercaptoethanol and 1 mM L-glutamine (GIBCO BRL,

Karlsruhe, Germany) at 37°C in a moist atmosphere with 5% CO₂. The spleen cell suspensions were adjusted to 5 x 10⁶ cells/ml in complete medium. They were introduced at 100 µl/well into a flat-bottomed 96-well microtiter plate (Nunc), and these plates were incubated with the addition of 20 µg/ml soluble β-galactosidase for 4 days. During the last 18 hours of culturing, 1 µCi of [³H]thymidine (Amersham International, Freiburg, Germany) were added to each well. The cells were then harvested on filter paper (Filtermat A; Wallac, Freiburg, Germany) with a cell harvester (Inotech, Wohlen, Switzerland), and the amount of [³H]thymidine incorporated into the DNA of the proliferated cells was determined with the aid of a γ scintillation counter (Wallac 1450, Micro-Trilux). The results are shown as the arithmetic mean of the [³H]thymidine uptake in cpm. The results are shown as stimulation indices (SI, cpm samples/cpm in unstimulated control cells).

20 Taking account of the surprising reduction in the adjuvant effect at the humoral level observed on use of MALP-2 in higher dosages, it was decided to find whether a similar effect can be observed at the level of the cellular immune responses. Accordingly, mice were immunized with either pure β-galactosidase or β-galactosidase mixed with 1 µg of MALP-2. Twenty-eight days after the immunization, the spleens were purified, restimulated *in vitro* with 20 µg/ml β-galactosidase, and their ability to proliferate was determined by measuring the incorporation of [³H]thymidine in their DNA by means of a γ scintillation counter. The results obtained (Fig. 5) confirmed that the use of MALP-2 in higher dosages not only leads to humoral responses no longer being detectable, but also the cell-mediated immunization is reduced.

Example 4: Joint intranasal and intraperitoneal administration of MALP-2 with a soluble antigen stimulates effective systemic humoral responses.

5 *Experimental protocol:* Six- to eight-week old female BALB/c (H-2d) mice were purchased from Harlan Winkelmann GmbH (Borchen, Germany) and treated in accordance with local and EU directives. Groups each of
10 5 mice were immunized on day 1, 14 and 21 either with 40 µg of pure β-galactosidase (Boehringer, Mannheim, Germany) with admixture of 0.5 µg of synthetic MALP-2 or 10 µg of cholera toxin B subunit (CTB; ICN Biomedicals Inc., Ohio) as standard adjuvant. For intranasal administration (i.n.), 10 µl were
15 administered into each nostril, while for the i.p. injection β-galactosidase was resuspended with or without MALP-2 in 400, 100 or 100 µl of PBS. Serum samples were taken at various times (day 0, 13, 20 and 30) and stored at -20°C until the β-galactosidase-
20 specific antibodies were determined. Nunc-Immuno MaxiSorp assay plates with 96 wells (Nunc, Roskilde, Denmark) were coated with 100 µl of β-gal (Boehringer, Mannheim, Germany) with 5 µg/ml in 0.05 M carbon buffer (pH 8.2) per well. Serial two-fold dilutions of the
25 sera or lavages in PBS with 1% BSA and 0.05% Tween-20 were added (100 µl/well) and the plates were incubated at 37°C for 24 hours. After rinsing, biotinylated γ chain-specific goat anti-mouse IgG (Sigma Chemie, Deisenhofen, Germany) was added, and the plates were
30 incubated at 37°C for a further hour. After rinsing four times, 100 µl of peroxidase-conjugated streptavidin (Pharmingen) were added to the cells, and the plates were incubated at 37°C for 30 minutes. After rinsing four times, the reactions were developed by
35 means of ABTS in 0.1 M citrate-phosphate buffer (pH 4.35) which contained 0.01% H₂O₂. The results have been shown as the reciprocal log₂ of the last dilution which, after incubation for 30 minutes, resulted in an

optimal density at 405 nm of 0.1 units compared with the values for the negative control group.

On the basis of the encouraging results of the preliminary studies, we decided to analyze in detail the immune responses found on use of MALP-2 as adjuvant by the two most effective immunization routes, namely i.p. and i.n., and compare them with those for a proven mucosal adjuvant. Accordingly, the ability of MALP-2 to elicit an effective humoral immune response was assessed by determining the serum titer of β -galactosidase-specific antibodies in immunized mice. As shown in Fig. 6A, administration of pure β -galactosidase (50 μ g/dose) resulted in induction of very low antibody titers, even after a second boost (endpoint titer about 1000). By comparison therewith, administration of β -galactosidase with use of MALP-2 i.n. induced, even with a single dose, to induction of very high titers ($> 60,000$) of specific IgG in all the mice, and the titers at the end of the immunization protocol were above 500,000 (Fig. 6). The kinetics and the overall efficacy of the antibody responses achieved with 0.5 μ g of MALP-2 were very similar to those achieved on administration of β -galactosidase with 10 μ g of CTB, a proven mucosal adjuvant, which was used as positive control substance.

A marked adjuvant effect was also observed on i.p. administration of MALP-2. In particular, coinjection of MALP-2 led to a 100-fold increase in the β -galactosidase-specific IgG titers compared with the titers in animals immunized with pure β -galactosidase (Fig. 6B). This difference was to be found even after the first immunization and was maintained after booster injections. Similar antibody titers were found on day 31 in animals immunized either i.n. or i.p. However, the primary responses after MALP-2 coinjection were more pronounced after the i.n. immunization.

Example 5: Intranasal coadministration of MALP-2 with a soluble antigen stimulates effective mucosal antibody responses.

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Experimental protocol: The mice were sacrificed on day 31, and the final samples were taken. Vaginal and lung lavages were obtained by rinsing the organs with 1 ml of PBS supplemented by 50 mM EDTA, 0.1% BSA and 10 mM PMSF. The lavages were then centrifuged in order to remove tissue detritus (10 min at 3000 x g) and the remaining liquid was stored at -20°C. In order to determine the total IgA concentration in the rinsings from lung and vagina, serial dilutions of the corresponding samples were incubated in microtiter plates, these plates having previously been coated with goat anti-mouse IgA (Sigma Chemie) as capture antibody (100 µl/well). Serial dilutions of purified mouse IgA (Sigma Chemie) were used to produce a standard plot.

20

In order to investigate the ability of MALP-2 to stimulate mucosal responses to antigens coadministered i.n., the production of β -galactosidase-specific IgA in lung and vaginal rinsings of immunized animals was. Whereas no production of β -galactosidase-specific IgA in detectable quantity in the lung rinsings took place after i.n. immunization with pure β -galactosidase, a significant increase in the level of antigen-specific IgA was found for the animals immunized with β -galactosidase and MALP-2 (Fig. 7). Coadministration of MALP-2 led to stimulation of an effective IgA production even in remote mucous membranes, and is demonstrated by the presence of significant concentrations of β -galactosidase-specific IgA in the vaginal rinsings (Fig. 7). No statistically significant differences were found in the levels of β -galactosidase-specific antibodies in the mucous membranes between animals immunized with 0.5 µg of

35

MALP-2 or 10 µg of CTB.

Example 6: Immunization with use of MALP-2 as adjuvant does not lead to more serum IgE.

5

Experimental protocol: To determine the total IgE concentration in the serum of immunized and control animals, serial dilutions of the appropriate samples were incubated in microtiter plates which had previously been coated with anti-mouse IgE (100 µl/well) as capture antibody. After blocking with PBS with 1% BSA and 0.05% Tween-20 at room temperature for two hours, a 1:100 dilution of the serum in PBS-Tween was added (100 µl/well) and the plates were incubated at 37°C for one hour. After rinsing, biotinylated anti-mouse IgE was added and the plates were incubated at 37°C for a further hour. After rinsing four times, 100 µl of peroxidase-conjugated streptavidin (Pharmingen) were put in the wells, and the plates were incubated at 37°C for 30 minutes. After rinsing four times, the reactions were developed by means of ABTS in 0.1 M citrate-phosphate buffer (pH 4.35) and a content of 0.01% H₂O₂. Serial dilutions of purified mouse IgE were used to produce a standard plot.

It is an acknowledged fact that the use of certain mucosal adjuvants may, because of an increase in IgE production, lead to allergic reactions. In order to check this hypothesis, we have started to examine the effect of MALP-2 administration on the serum IgE content. As is evident from Fig. 8, administration of MALP-2 by the parenteral (i.p.) route or via the mucous membranes does not lead to an increase in the serum IgE content. On the contrary, the presence of MALP-2 appears in fact to have a beneficial effect on the increase in the IgE content as observed on day 28 after i.p. immunization with pure β-galactosidase.

Example 7: MALP-2 stimulates efficient T-cell-mediated proliferation responses when it is administered together with a soluble antigen

5

Experimental protocol: Lymph nodes located below the lower jaw and the spleens were removed and put together for the analysis of the cellular immune responses. The cells were grown in RPMI 1640 supplemented by 10% fetal calf serum, 10 U/ml penicillin, 50 µg/ml streptomycin, 5 x 10⁻⁵ M 2-mercaptoethanol and 1 mM L-glutamine (GIBCO BRL, Karlsruhe, Germany) and stored at 37°C in a moist atmosphere with 5% CO₂. The suspensions of lymph node and spleen cells were adjusted to 5 x 10⁶ cells/ml in complete medium and put in a flat-bottomed 96-well microtiter plate (Nunc) with 100 µl/well, and the plates were incubated in the presence of various concentrations of soluble β-galactosidase for 4 days. Each concentration was tested in groups of three. During the last 18 hours of the incubation, 1 µCi of [³H]thymidine (Amersham International, Freiburg, Germany) was added to each well. The cells were then harvested on filter paper (Filtermat A; Wallac, Freiburg, Germany) using a cell harvester (Inotech, Wohlen, Switzerland), and the amount of incorporated [³H]thymidine in the DNA of the proliferated cells was determined with the aid of a γ scintillation counter (Wallac 1450, Micro-Trilux). The results have been shown as the arithmetic mean of the uptake of [³H]thymidine in cpm.

The immune responses of T cells were investigated on day 31 by determining the proliferation of the cells obtained from regional lymph nodes and spleens after *in vitro* restimulation with β-galactosidase. The spleen cells from animals immunized i.p. with pure β-galactosidase were used as positive control group and showed a significant proliferation response compared

with the unimmunized group (Fig. 9A). A further increase in proliferation was found in spleen cells derived from animals which had received coinjection of MALP-2 and antigen ($p < 0.05$). Whereas i.n. administration of pure β -galactosidase caused no detectable cell proliferation, coadministration of MALP-2 led to an efficient proliferation response both in regional (lymph node cells) and in systemic (spleen cells) (Figs. 9B and C). It is noteworthy that the greatest T-cell proliferation was observed for the spleen cells of mice to which MALP-2 and β -galactosidase had been administered i.n. (Fig. 9B). In all cases there was found to be a distinctly dose-dependent effect due to the increase in the β -galactosidase concentration during restimulation (5, 10, 20 $\mu\text{g/ml}$). Finally, use of MALP-2 (0.5 μg) as adjuvant led to a statistically significant ($p < 0.05$) increase in T-cell proliferation compared with i.n. immunization with (10 μg) plus β -galactosidase (Fig. 8B).

Example 8: Analysis of T-helper patterns elicited through the use of MALP-2 as adjuvant

Experimental protocol: Nunc-immuno MaxiSorp assay plates with 96 wells (Nunc, Roskilde, Denmark) were coated with 100 μl of β -galactosidase (Boehringer, Mannheim, Germany) with 5 $\mu\text{g/ml}$ in 0.05 M carbonate buffer (pH 8.2) per well. Serial two-fold dilutions of serum or rinsings in PBS with 1% BSA and 0.05% Tween 20 were added (100 $\mu\text{l/well}$), and the plates were incubated at 37°C for 2 hours. After rinsing, biotin-conjugated rat anti-mouse IgG1, IgG2a, IgG2b, or IgG3 (Pharmingen, Hamburg, Germany) was added in order to determine the Ig subclasses. The plates were incubated at 37°C for a further hour. After rinsing four times, 100 μl of peroxidase-conjugated streptavidin (Pharmingen) were added to the cells, and the plates were incubated at

- 37°C for 30 minutes. After rinsing four times, the reactions were developed with ABTS in 0.1 M citrate-phosphate buffer (pH 4.35) which contained 0.01% H₂O₂. In order to determine the serum concentration of the
- 5 IgG subclasses, standard plots were produced by coating the wells with an isotype-specific goat anti-mouse IgG and then incubating with purified mouse IgG1, IgG2a, IgG2b or IgG3 antibodies (Dianova, Hamburg, Germany).
- 10 Liquid supernatants from cultures of proliferating cells were removed on days 2 and 4 and stored at -70°C. Determination of IFN- γ , IL-2, IL-4 and IL-10 was carried out by means of an ELISA, using commercial antibodies from Pharmingen in accordance with the
- 15 manufacturer's instructions. Briefly, 96-well microtiter plates were coated overnight with purified rat anti-mouse IFN- γ , anti-IL-2, anti-IL-4 or anti-IL-10 mAbs (Pharmingen) at 4°C. After three rinsings, the plates were blocked and the liquid supernatants were
- 20 put in the wells. A standard plot was produced for each cytokine by using the corresponding recombinant rodent cytokines (Pharmingen). The plates were incubated at room temperature for a further 4 hours. After rinsing, biotinylated rat anti-mouse IFN- γ , IL-2, IL-4 or IL-10
- 25 mAbs (Pharmingen) was put in the wells, and the plates were incubated at room temperature for one hour. After rinsing six times, streptavidin-peroxidase conjugate was added and the plates were incubated at room temperature for 30 minutes. The plates were then
- 30 developed with ABTS as described above.

Firstly, the subclass distribution of the β -galactosidase-specific IgG which was present in the serum of the immunized mice was. As shown in Table 1,

35 the main type of β -galactosidase-specific IgG isotypes was IgG1, irrespective of the immunization protocol. This dominant Th2 response pattern was evident even after the first immunizing dose and was maintained

during the following boosters. IgG1 was the only isotype found on administration of pure β -galactosidase, whereas coadministration with CTB or MALP-2 led to the finding of further β -galactosidase-specific isotypes, namely IgG2a (Typ Th1), IgG2b (Typ Th2) and IgG3 (Typ Th1). Despite this, the IgG1/2a (Fig. 10), IgG1/2b or IgG1/3 ratio remained above 100.

Table 1. β -Galactosidase-specific IgG isotypes in the serum of immunized mice^a

Immunization group	IgG1	IgG2a	IgG2b	IgG3
β -gal (i.n.)	22.6 +/- 21.3	0.7 +/- 0.5	0.3 +/- 0.1	0.6 +/- 0.0
β -gal + MALP-2 (i.n.)	6439.0 +/- 1775	20.8 +/- 5.4	43.3 +/- 18.9	2.4 +/- 0.5
β -gal + CTB (i.n.)	4108.3 +/- 1437	31.9 +/- 9.5	49.2 +/- 17.3	2.4 +/- 0.6
β -gal (i.p.)	191.5 +/- 132.1	0.1 +/- 0.0	0.5 +/- 0.1	0.6 +/- 0.0
β -gal + MALP-2 (i.p.)	2829.7 +/- 1119	10.0 +/- 2.8	15.8 +/- 6.2	2.3 +/- 0.6
Control group	0.4 +/- 0.0	0.23 +/- 0.0	0.1 +/- 0.0	0.4 +/- 0.0

^a The results are shown as average (μ g/ml) $\pm$ SEM (5 mice per group)

In order to characterize further the type of Th response elicited by the immunization, the IFN- γ , IL-2, IL-4 and IL-10 content was measured in the liquid supernatants from spleen cells stimulated *in vitro* (Fig. 11A). It was found that, of these four cytokines, IL-10 was most prominent, indicating that a Th2 response pattern had been generated. The IL-10 content was significantly higher in mice immunized i.n. with MALP-2 than in the control group (2.2 ng/ml compared with 0.009 ng/ml, $p < 0.005$) and also by comparison with animals for which CTB had been used as mucosal adjuvant (0.6 ng/ml, $p < 0.05$). The observed response pattern with a dominant cytokine of the Th2 type was consistent with the finding of β -galactosidase-specific

IgG1 in the same animals (Fig. 10). In fact, the strong stimulation of IL-10 secretion is consistent with the part played by this cytokine in the inhibition of cytokine synthesis by Th1 cells, the improvement of B-cell proliferation and the stimulation of IgA production.

Despite the fact that the absolute levels of the cytokine concentration found in the cell culture media of cells obtained from regional lymph nodes were lower, the general pattern was the same as that found for spleen cell cultures (Fig. 10). It is of interest that although secretion of IL-10 and IL-4 was also stimulated in the cells from mice immunized i.p. with pure β -galactosidase, the Th1 cytokines IL-2 and IFN- γ remained below the limit of detection. In contrast thereto, IL-2 and IFN- γ were also found in the mice which had received the antigen mixed with CTB or MALP-2 (Fig. 10). These results are consistent with the IgG-isotype patterns found (Table 1) and confirm that although Th2 response types predominate, MALP-2 also assists stimulation of Th1 cells.

Example 9: Oral immunization with the model antigen β -galactosidase with use of MALP-2 as mucosal adjuvant.

Six- to eight-week old female BALB/c (H-2d) mice were purchased from Harlan Winkelmann GmbH (Borchen, Germany) and treated in accordance with local guidelines and directives of the European Community. Groups each of 5 mice were immunized on days 1, 14, 21 and 31 with 100 μ g of β -gal (Boehringer, Mannheim, Germany) either alone or with 2 μ g of synthetic MALP-2 as adjuvant by the oral route (dose 25 μ l). On day 45, serum samples were taken and stored at -20°C until β -gal-specific antibodies were determined. 96-well Nunc-Immuno MaxiSorp® assay plates (Nunc, Roskilde, Denmark) were coated with 100 μ l of β -gal at 5 μ g/ml in 0.05 M

carbonate buffer (pH 8.2) per well. Serial two-fold dilutions of the sera or lavages in PBS with 1% BSA and 0.05% Tween 20 were added (100 µg/well), and the plates were incubated at 37°C for 2 hours. After washing, 5 biotinylated γ chain-specific goat anti-mouse IgG (Sigma Chemie, Deisenhofen, Germany) was added, and the plates were incubated at 37°C for a further hour. After washing four times, 100 µl of peroxidase-conjugated streptavidin (Pharmingen) was added to the cells, and 10 the plates were incubated at 37°C for 30 minutes. After four washing steps, reactions were developed with ABTS in 0.1 M citrate-phosphate buffer (pH 4.35) which contained 0.01% H₂O₂. The final value titers have been expressed as the reciprocal logarithmic log₂ values of 15 the last dilution which resulted, after incubation for 30 minutes, in an optical density at 405 nm of 0.1 units above the values for the negative controls. Immune responses to β -gal were observed only in animals immunized with MALP-2 as mucosal adjuvant (see 20 Fig. 12).

It is to be expected that the immune responses elicited also with different dosages both in the body fluids and in the cells will be more effective than those elicited 25 on administration of the antigen alone. In addition, antigen-specific mucosal reactions are to be found at least locally in the intestines (i.e. the presence of antigen-specific secretory IgA in intestinal rinsings). It is additionally to be expected that secretory 30 responses can be found in remote mucous membranes.

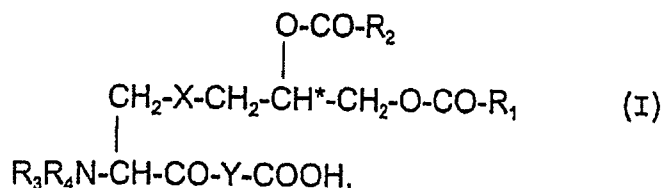
The reference in this specification to any prior publication (or information derived from it), or to any matter which is known, is not, and should not be taken as an acknowledgment or admission or any form of suggestion that that prior publication (or information derived from it) or known matter forms part of the common general knowledge in the field of endeavour to which this specification relates.

Throughout this specification, unless the context requires otherwise, the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated element or integer or group of elements or integers but not the exclusion of any other element or integer or group of elements or integers.

Claims:

1. The use of a lipopeptide or lipoprotein of the structure (I)

5



where

10 R_1 and R_2 , which may be identical or different, are C_{7-25} -alkyl, C_{7-25} -alkenyl or C_{7-25} -alkynyl,

X is S, O or CH_2 ,

R_3 and R_4 are independently of one another H or methyl and

15 Y is a physiologically tolerated amino acid sequence which consists of 1 to 25, preferably 12 to 25, amino acid residues and is not immunogenic per se in the species used,

20 and the asymmetric carbon atom marked with * as the absolute R configuration, according to the Cahn-Ingold-Prelog rule, when X is S (sulfur), as mucosal adjuvant in therapeutic or prophylactic vaccination via the mucous membranes.

- 25 2. The use as claimed in claim 1, characterized in that the amino acid sequence Y is preferably selected from.

a) GQTNT

b) SKKKK

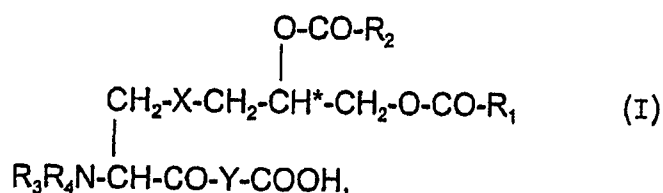
c) GNNDESNISFKEK

30 d) GQTDNNSSQSAAPGSGTTNT,

3. The use as claimed in claim 1 or 2, characterized in that the lipoprotein or lipopeptide of structure (I) is an S-[2,3-

bispalmitoyloxy(2*R*)propyl]cysteinyl-peptide, where the peptide is a physiologically tolerated amino acid sequence which consists of 1 to 25 amino acid residues and is preferably not immunogenic in the species used.

4. The use as claimed in any of claims 1 to 3, characterized in that the mucosal adjuvant is present in a preparation with the actual vaccine component which is intended for intranasal, intra-NALT, aerosolized oral, intrarectal, conjunctival, intravaginal or intraurethral administration or administration into the milk ducts of the female breast.
5. The use as claimed in any of claims 1 to 3, characterized in that the mucosal adjuvant is present in a kit for coadministration with a vaccine into the milk ducts of the female breast, by the intranasal, intra-NALT, aerosolized oral, intrarectal, conjunctival, intravaginal or intraurethral route.
6. The use of a lipopeptide or lipoprotein of the general structure (I)



where

- R_1 and R_2 , which may be identical or different, are C_{7-25} -alkyl, C_{7-25} -alkenyl or C_{7-25} -alkynyl, X is S , O or CH_2 , R_3 and R_4 are independently of one another H or methyl and

- 37 -

Y is a physiologically tolerated amino acid sequence which consists of 1 to 25, preferably 12 to 25, amino acid residues and is not immunogenic per se in the species used,
5 and the asymmetric carbon atom marked with * as the absolute R configuration, according to the Cahn-Ingold-Prelog rule, when X is S (sulfur), excepting an S-(2,3-diacyloxypropyl)cysteine-peptide of the sequence DhcGNNDESNISFKEK, where N-terminally the amino acids at positions 2 and, where appropriate, 3 are absent, and/or C-terminally 1 to 2 amino acids may be deleted,

as adjuvant in a non-mucosal vaccination.

15

7. The use as claimed in any of claims 1 to 6, characterized in that the lipopeptide or lipoprotein is present in a preparation with at least one further adjuvant and/or antigen.

20

8. The use as claimed in any of claims 1 to 7, characterized in that the lipopeptide or lipoprotein is associated or combined with a physical or biological carrier.

25

9. The use as claimed in any of claims 1 to 8, characterized in that the lipopeptide or lipoprotein is administered together with one or more anti-inflammatory, antiangiogenic, cytotoxic or immunomodulatory substances or ligands or with antibodies, or is present with these in a preparation.

30

10. The use as claimed in any of claims 1 to 9, characterized in that the lipopeptide or lipoprotein is present in a preparation which comprises further additives and excipients, in particular preservatives or stabilizers.

35

- 5 11. The use as claimed in any of claims 1 to 10, characterized in that the vaccine which is accompanied by the adjuvant, in the form of peptides, proteins, DNA, polysaccharides, glycolipids or glucoproteins.
12. The use according to anyone of claims 1 to 11, substantially as herein before described with reference to the figures and/or examples.

Figure 1

Expression of specific markers by murine DCs obtained from bone marrow after stimulation with MALP-2 or LPS

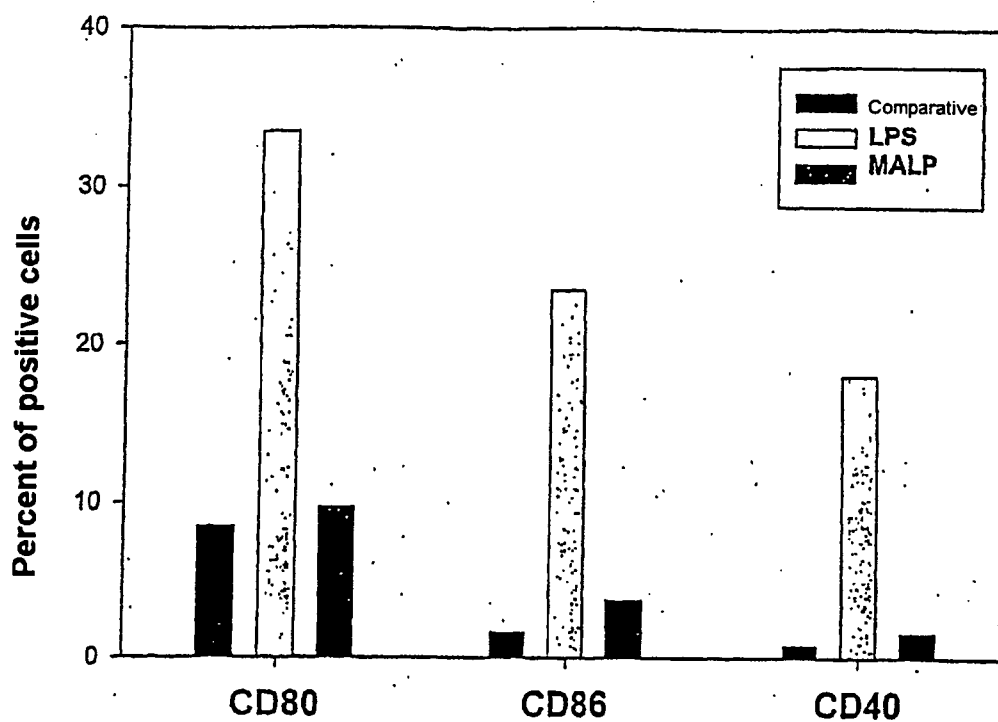


Figure 2

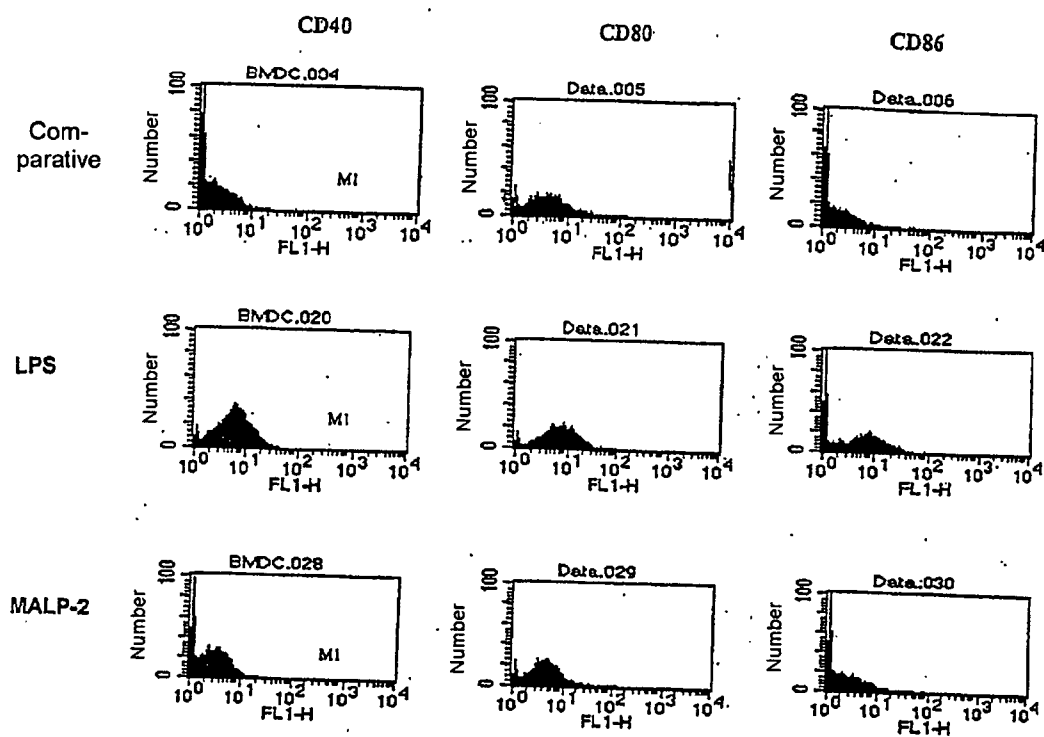


Figure 3

β -Galactoside-specific serum IgG response stimulated through use of MALP-2 as adjuvant

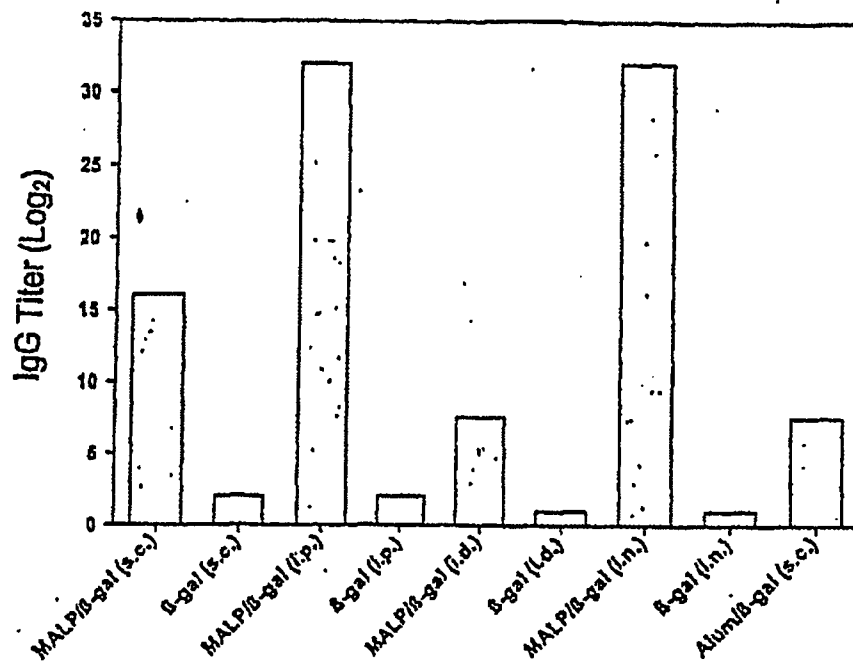


Figure 4

β -Galactosidase-specific serum IgG response stimulated through use of 1 μ g of MALP-2 as adjuvant

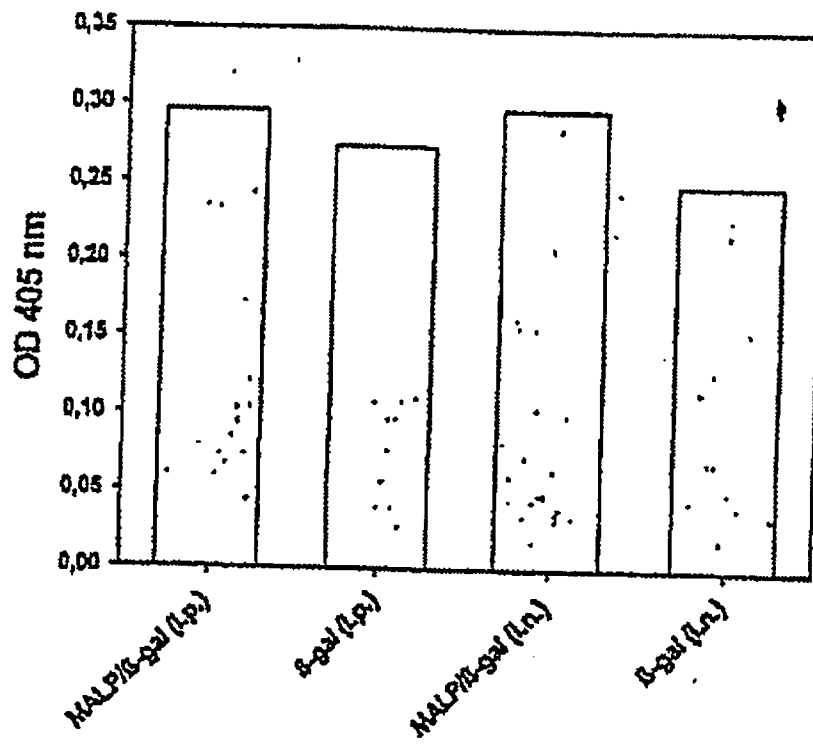


Figure 5

β -Galactosidase-specific proliferative responses
stimulated through use of MALP-2 as adjuvant

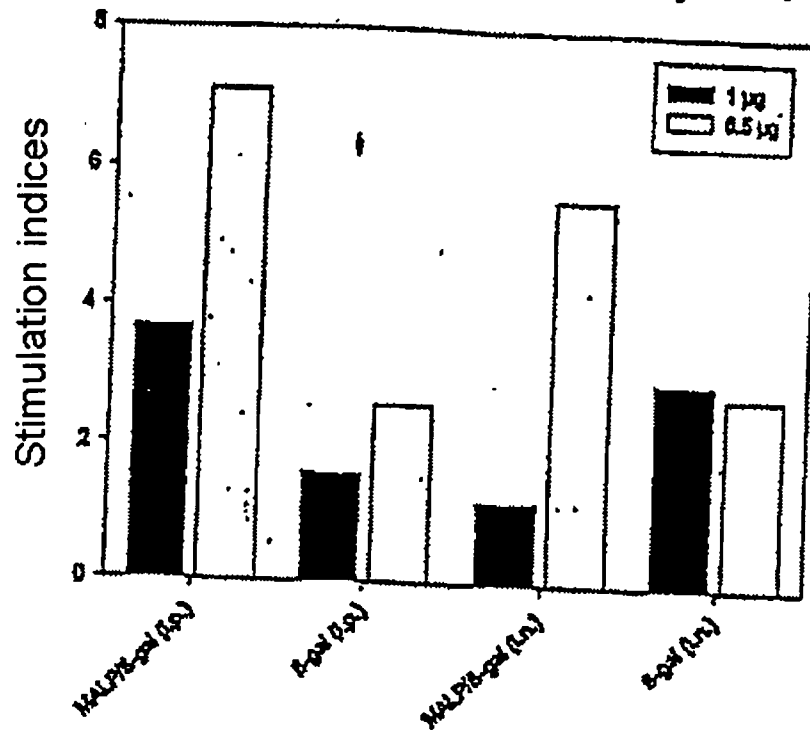


Figure 6

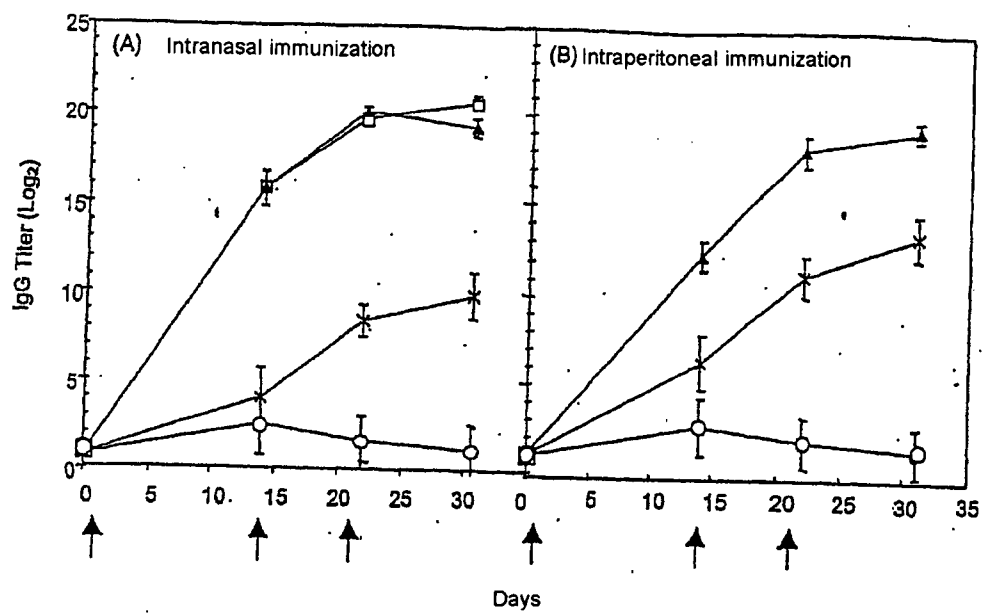


Figure 7

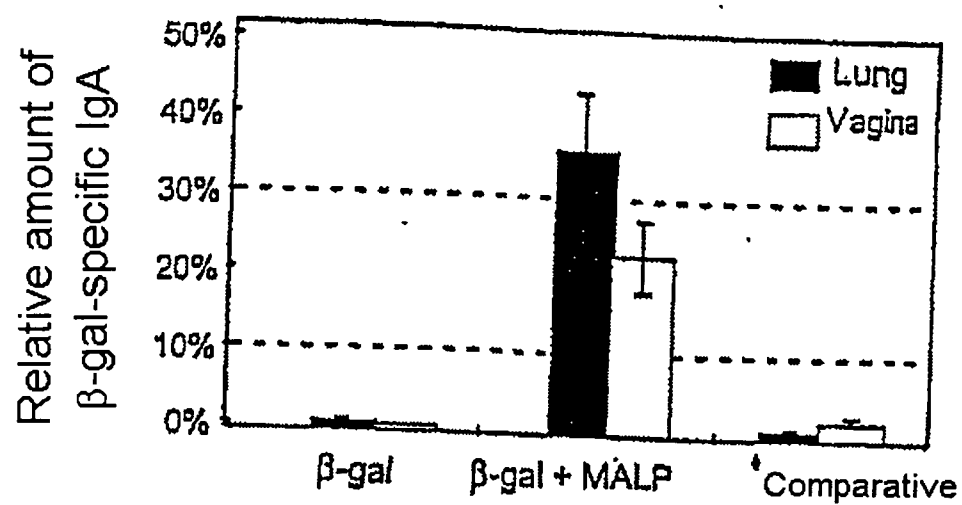


Figure 8

Determination of serum IgE in animals immunized with use of MALP-2 as adjuvant

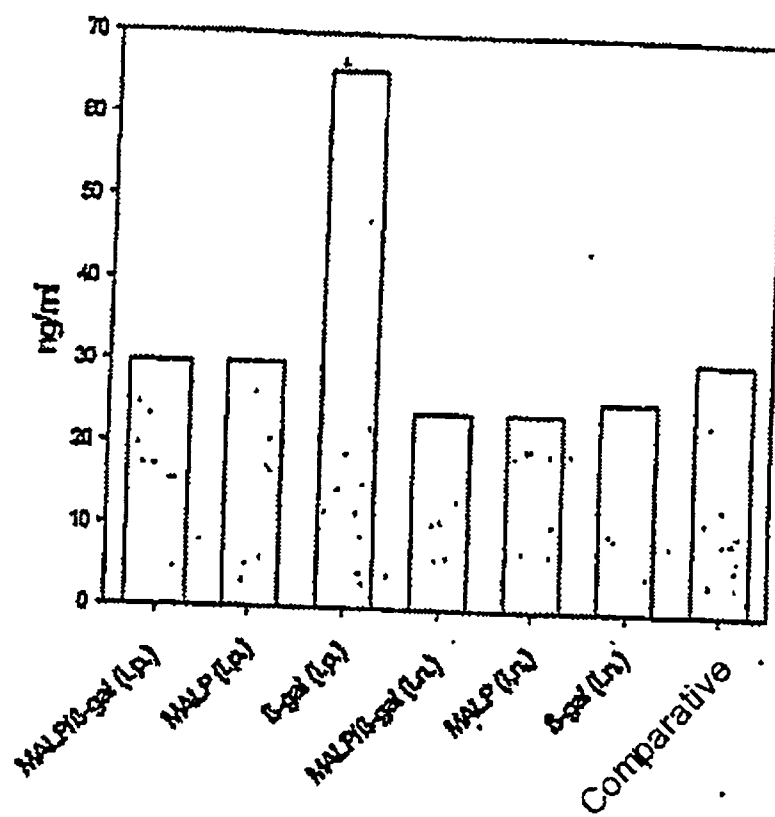


Figure 9

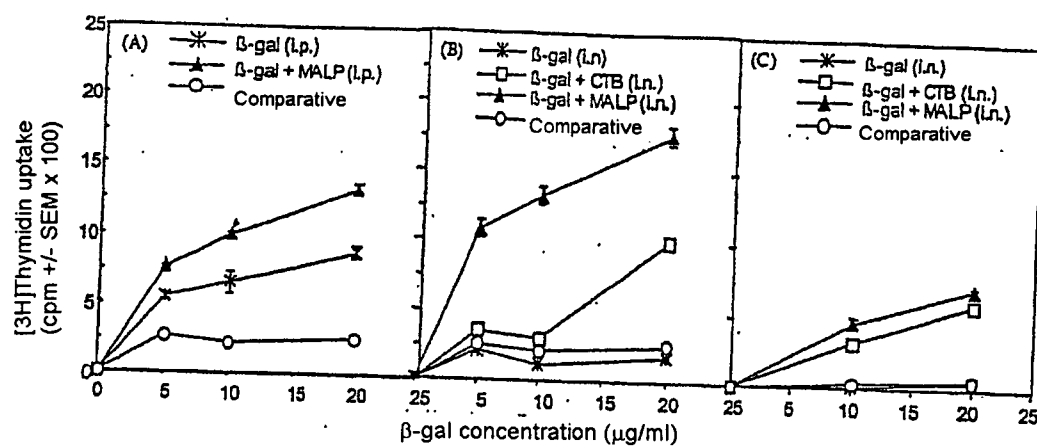


Figure 10

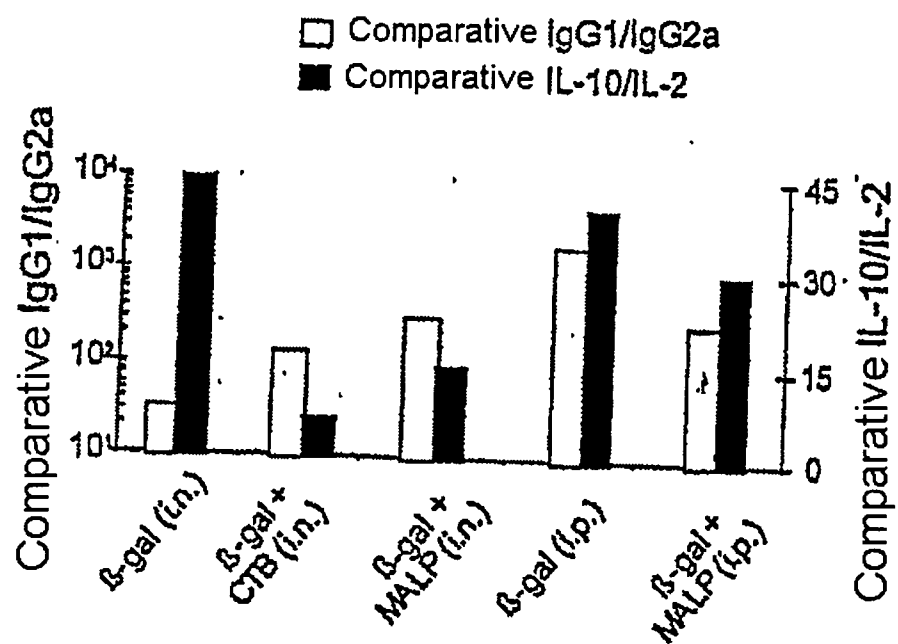


Figure 11

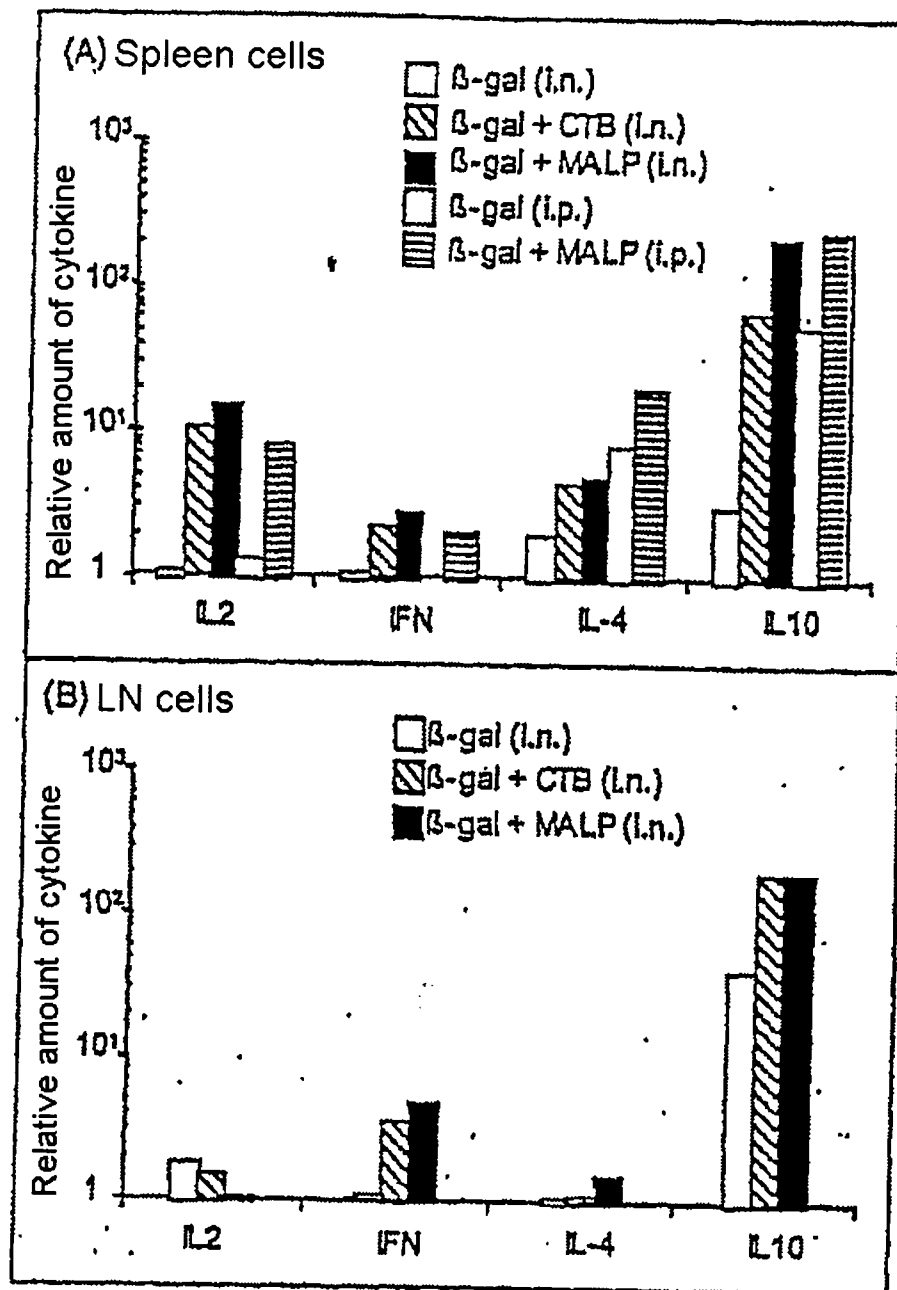


Figure 12

 β -Galactosidase + MALP-2 ORALAnti β -Gal-IgG (day 30) :