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(71) Applicant(s)
Gene Signal International SA

(72) Inventor(s)
Al-Mahmood, Salman

(74) Agent/Attorney
Griffith Hack, 509 St Kilda Road, Melbourne, VIC, 3004

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(71) Déposant et

(72) Inventeur : AL-MAHMOOD, Salman [FR/FR]; 2,
square Alice, F 75014 Paris (IQ).

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(74) Mandataires : BREESE, Pierre etc.; Breesé-Majerowicz,
3, avenue de l'Opéra, F 75001 Paris (FR).

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(54) Title: ANTISENSE OLIGONUCLEOTIDES WHICH CAN INHIBIT THE FORMATION OF CAPILLARY TUBES BY ENDOTHELIAL CELLS

(54) Titre : OLIGONUCLEOTIDES ANTISENS CAPABLES D'INHIBER LA FORMATION DES TUBES CAPILLAIRES PAR DES CELLULES ENDOTHELIALES

(57) Abstract: The invention relates to pharmaceutical compositions which inhibit the formation of capillary tubes by endothelial cells, comprising at least one oligonucleotide which can inhibit the expression of the IRS-1 protein. According to the invention, the oligonucleotides are embodied as anti-angiogenesis agents. Said pharmaceutical compositions are particularly useful in treating angiogenesis-related pathologies.

(57) Abrégé : La présente invention a pour objet des compositions pharmaceutiques pour inhiber la formation de tubes capillaires par les cellules endothéliales, comprenant au moins un oligonucléotide capable d'inhiber l'expression de la protéine IRS-1. Les oligonucléotides selon l'invention sont indiqués comme agents anti-angiogéniques. Les compositions pharmaceutiques de l'invention sont particulièrement utiles pour le traitement de pathologies liées à l'angiogénèse.

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**ANTISENSE OLIGONUCLEOTIDES CAPABLE OF INHIBITING THE
FORMATION OF CAPILLARY TUBES BY ENDOTHELIAL CELLS**

The present invention provides antisense oligonucleotides capable of inhibiting the expression of 5 the protein IRS-1 and of inhibiting the formation of capillary tubes by endothelial cells. The oligonucleotides according to the invention are thus indicated as antiangiogenic agents. They are also indicated as anti-cell-multiplication agents, in 10 particular as anti-tumor agents.

The invention also pertains to pharmaceutical compositions containing said oligonucleotides and the use of said oligonucleotides as analysis reagents.

All references, including any patents or patent 15 applications, cited in this specification are hereby incorporated by reference. No admission is made that any reference constitutes prior art. The discussion of the references states what their authors assert, and the applicants reserve the right to challenge the accuracy and 20 pertinency of the cited documents. It will be clearly understood that, although a number of prior art publications are referred to herein, this reference does not constitute an admission that any of these documents forms part of the common general knowledge in the art, in 25 Australia or in any other country.

Angiogenesis is a fundamental process by means of which new blood vessels are formed. This process is essential in many normal physiological phenomena such as reproduction, development and even cicatrization. In 30 these normal biological phenomena, angiogenesis is under strict control, i.e., it is triggered during a short period (several days) and then completely inhibited. However, many pathologies are linked to uncontrolled, 35 invasive angiogenesis: arthritis, a pathology due to the damaging of cartilages by invasive neovessels; diabetic retinopathy or the invasion of the retina by neovessels leading to blindness of the patients; neovascularization

of the ocular apparatus presents the major cause of blindness and this neovascularization is involved in about twenty different eye diseases; and moreover the growth and metastasis of tumors which are linked directly to 5 neovascularization and are dependent on angiogenesis. The tumor stimulates the growth of neovessels by its own growth. Moreover, these neovessels are escape routes for tumors which thereby join up with the blood circulation and induce metastases in sites remote from the initial 10 tumor focus, such as the liver, lungs or bones.

Angiogenesis, the formation of neovessels by 15 endothelial cells, involves the migration, growth and differentiation of endothelial cells. The regulation of these biological phenomena is directly linked to genetic expression.

The research studies performed in the framework of the present invention made it possible to identify and prepare nucleic acid sequences involved in the regulation of angiogenesis.

20 Other studies pertaining to angiogenesis have shown a noteworthy expression and phosphorylation at the level of a tyrosine residue of an intracellular 180-kDa protein by endothelial cells cultured on a surface of type I collagen and stimulated by an angiogenic factor such as bFGF. The 25 noteworthy expression and phosphorylation at the level of the tyrosine residue of the intracellular 180-kDa protein accompanies the formation of capillary tubes by the endothelial cells.

This protein is already known as a substrate of the 30 insulin receptor (called IRS-1). It has been partially identified and investigated by certain diabetes researchers (Quon et al., J. Biol. Chem. (1994), 269 (45), 27920-27924).

These authors studied the role of IRS-1 in (i) the 35 translocation of GLUT 4 stimulated by insulin and (ii) the transport of glucose in rat adipose cells. In order to do this, they constructed a plasmid containing:

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- a double chain oligonucleotide obtained from the sense oligonucleotide of the following sequence SEQ ID NO. ID No. 1:5'-TCGATGTGAC GCTACTGATG AGTCCGTGAG GACGAAACTC TGGCCTAG-3'; and
 - cDNA coding for human IRS-1, and transfected rat adipose cells with said plasmid.
- The research performed in the framework of the present invention revealed that the expression of the protein IRS-1 is also induced in endothelial cells when said cells are stimulated by the angiogenic factor bFGF.

The invention thus pertains to a pharmaceutical composition active on angiogenesis phenomena comprising as active agent at least one substance selected from among: (i) a nucleic acid molecule of the gene coding for the protein IRS-1, a complementary sequence or a fragment thereof, (ii) a molecule capable of inhibiting the expression of a nucleic acid molecule according to (i).

In the framework of the invention, antisense oligonucleotides of the gene coding for this protein were prepared. These oligonucleotides present remarkable antiangiogenic and antitumor activities. They are therefore particularly useful in the treatment of diseases linked to invasive angiogenesis not controlled by gene therapy methods consisting of administering to an individual a composition containing at least one of these oligonucleotides.

Thus, an oligonucleotide according to the invention is constituted by the following nucleotide sequence of formula SEQ ID NO. 2:

5'-TATCCGGAGGGCTGCCATGCTGCTGCGGAGCAGA-3',

a fragment thereof comprising at least 12 contiguous nucleotides or their derivative.

The invention pertains most particularly to an oligonucleotide constituted by one of the nucleotide sequences of formulas SEQ ID NO. 3 and 4 below:

5'-TATCCGGAGGGCTGCCATGCTGCT-3',

5'-TCGCCATGCTGCTGCGGAGCAGA-3',

a fragment of these comprising at least 12 contiguous nucleotides or their derivative.

The term derivative is understood to mean a sequence capable of hybridizing under strict conditions with one of the sequences SEQ ID NO. 2, 3 or 4, or with a fragment of these of at least 12 contiguous nucleotides.

The following sequences can be cited as examples of oligonucleotides according to the invention:

SEQ ID NO. 5: 5'-TATCCGGAGGGCCTGCCATGCTGCT-3',
SEQ ID NO. 6: 5'-TATCCGGAGG GCCTGCCATG CTGC-3',
SEQ ID NO. 7: 5'-TATCCGGAGG GCCTGCCATG CTG-3',
SEQ ID NO. 8: 5'-TATCCGGAGG GCCTGCCATG CT-3',
SEQ ID NO. 9: 5'-TATCCGGAGG GCCTGCCATG C-3',
SEQ ID NO. 10: 5'-TATCCGGAGG GCCTGCCATG-3',
SEQ ID NO. 11: 5'-TATCCGGAGG GCCTGCCAT-3',
SEQ ID NO. 12: 5'-TATCCGGAGG GCCTGCCA-3',
SEQ ID NO. 13: 5'-TATCCGGAGG GCCTGCC-3'.
SEQ ID NO. 14: 5'-TATCCGGAGG GCCTGC-3',
SEQ ID NO. 15: 5'-TATCCGGAGG GCCTG-3',
SEQ ID NO. 16: 5'-TATCCGGAGG GCCT-3',
SEQ ID NO. 17: 5'-TATCCGGAGG GCC-3',
SEQ ID NO. 18: 5'-TATCCGGAGG GC-3',
SEQ ID NO. 19: 5'-CCGGAGG GCCTGCCATG CTGCT-3',
SEQ ID NO. 20: 5'-GAGG GCCTGCCATG CTGCT-3',
SEQ ID NO. 21: 5'-G GCCTGCCATG CTGCT-3',
SEQ ID NO. 22: 5'-CTGCCATG CTGCT-3'.
SEQ ID NO. 23: 5'-TGCCATG CTGCT-3'.

All or part of the phosphodiester bonds of the invention are advantageously protected. This protection is generally implemented via the chemical route using classic methods that are well known by the expert in the field. The phosphodiester bonds can be protected, for example, by a thiol or amine functional group or by a phenyl group.

The 5'- and/or 3'- ends of the oligonucleotides of the invention are also advantageously protected, for example, using the technique described above for protecting the phosphodiester bonds.

The oligonucleotides of the invention can be synthesized using conventional techniques that are well known to the expert in the field, for example, using one of the DNA synthesizers marketed by various companies.

Although their mechanism of action has not been entirely elucidated, the oligonucleotides according to the invention inhibit the expression of the protein IRS-1 within endothelial cells. These oligonucleotides are capable of blocking the formation of neovessels by endothelial cells (i.e., they inhibit angiogenesis) and thus they inhibit the multiplication of tumor cells in mice.

The invention therefore also has as object a pharmaceutical composition for the inhibition of the gene coding for the protein IRS-1 comprising at least one oligonucleotide complementary of a part of said gene or of a transcript of said gene.

The molecule capable of inhibiting the expression of a nucleic acid molecule of the gene coding for the protein IRS-1 is preferably an antisense sequence of the region coding the sequence identified under the number SEQ ID NO. 28 in the attached sequence list.

Said antisense sequence advantageously comprises at least twelve contiguous nucleotides or their derivative.

More preferentially, the active agent capable of inhibiting the expression of a nucleic acid molecule coding for the protein IRS-1 of the composition of the invention is a nucleotide sequence selected from among the set of nucleotide sequences identified as SEQ ID NO. 2 to SEQ ID NO. 23 in the attached sequence list comprising at least twelve contiguous nucleotides or their derivative.

Such a composition advantageously comprises as active agent at least one oligonucleotide as defined above advantageously combined in said composition with an acceptable vehicle.

The analysis of the research performed in the framework of the invention made it possible to demonstrate that the protein IRS-1 represents a cellular constituent which is essential in the angiogenesis process. In fact, 5 inhibition of the expression of the protein IRS-1 by said antisense oligonucleotides leads to the inhibition of the formation of capillary tubes by endothelial cells.

The oligonucleotides according to the invention and the compositions containing them are thus indicated as 10 antiangiogenic agents. They are also indicated as anti-cell-multiplication agents, particularly as antitumor agents, and consequently are particularly useful for the treatment of tumors. Thus the present invention provides the use of said oligonucleotides for the preparation of a 15 composition intended for the treatment or prevention of pathologies linked to invasive, uncontrolled angiogenesis such as, as nonlimitative example: the treatment of tumour vascularization, eye diseases linked to the neovascularization of the ocular apparatus such as 20 retinopathies, rheumatoid arthritis, Crohn's disease, atherosclerosis, hyperstimulation of the ovary, psoriasis, endometritis associated with neovascularization, restenosis due to balloon angioplasty, tissue superproduction due to cicatrisation, peripheral vascular 25 disease, hypertension, vascular inflammation, Raynaud's disease and Raynaud's phenomena, aneurysm, arterial restenosis, thrombophlebitis, lymphangitis, lymphedema, tissue cicatrisation and repair, ischemia, angina, myocardial infarction, chronic heart disease, cardiac 30 insufficiencies such as congestive heart failure, age-related macular degeneration and osteoporosis.

The above pharmaceutical compositions are more particularly implemented in a manner such that they can be administered via the subcutaneous, intramuscular, 35 intravenous or transdermal route. For such administration, use is made notably of aqueous suspensions, isotonic saline solutions or sterile,

Injectable solutions containing pharmacologically compatible dispersion agents and/or wetting agents such as, for example, propylene glycol or butylene glycol.

The usual unit dose to be administered contains from 5 0.001 mg to 50 mg of active principle.

The oligonucleotides of the invention are also useful as research reagents, notably for the *in vitro* study of signalization routes involving the 180-kDa protein, for example on tumor cells or non-tumor cells transfected by 10 said oligonucleotides. They are also useful for the *in vivo* study of signalization routes involving the 180-kDa protein in a large number of physiological and pathological phenomena such as angiogenesis or carcinogenesis essentially from the kinase/phosphatase 15 ratio.

Thus, the pharmaceutical compositions of the invention are particularly useful for the performance of tests for the diagnosis of pathologies linked to angiogenesis phenomena, notably for the diagnosis of retinopathies, 20 rheumatoid arthritis, Crohn's disease, atherosclerosis, hyperstimulation of the ovary, psoriasis, endometritis association with neovascularization, restenosis due to balloon angioplasty, tissue superproduction due to cicatrization, peripheral vascular disease, hypertension, 25 vascular inflammation, Raynaud's disease and Raynaud's phenomena, aneurysm, arterial restenosis, thrombophlebitis, lymphangitis, lymphedema, tissue cicatrization and repair, ischemia, angina, myocardial infarction, chronic heart disease, cardiac insufficiencies 30 such as congestive heart failure or age-linked macular degeneration and osteoporosis.

In the claims which follow and in the description of the invention, except where the context requires otherwise due to express language or necessary implication, the word 35 "comprise" or variations such as "comprises" or "comprising" is used in an inclusive sense, i.e. to specify the presence of the stated features but not to

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preclude the presence or addition of further features in various embodiments of the invention.

Other advantages and characteristics of the invention will become clear from the examples below in which the

- 5 term "oligonucleotide" is used to designate the oligonucleotide of SEQ ID NO.3 and which refer to the attached figures in which:

- figure 1A represents the Western Blot images obtained from supernatant samples stemming from unstimulated cells (track NS) and cells stimulated with bFGF (track S) developed with an anti-IRS-1 antibody,
- figure 1B shows the Western Blot images obtained after staining with silver nitrate obtained from the same supernatant samples stemming from unstimulated cells (track NS) and cells stimulated with bFGF (track S),
- figure 2 shows the Western Blot images obtained from supernatant cells stemming from unstimulated cells (track NS) and cells stimulated with bFGF (track B) when the membrane is incubated with an anti-phosphotyrosine monoclonal antibody and developed with an anti-isotope antibody tagged at the peroxidase as indicated in example 3,
- figures 3A to 3D show the images of the cultures on a type I collagen surface of the different lots of endothelial cells:
 - figure 3A shows the culture of untreated endothelial cells,
 - figure 3B shows the culture of endothelial cells stimulated with 3 ng/ml of bFGF,
 - figure 3C shows the culture of endothelial cells incubated with 100 µg/ml of oligonucleotide of SEQ ID NO. 3 for 4 hours and then stimulated with 3 ng/ml of bFGF,
 - figure 3D shows the culture of endothelial cells incubated with 100 µg/ml of oligonucleotide of SEQ ID NO. 3 for 4 hours,
- figures 4A to 4F illustrate the results of tests of the inhibition of corneal neovascularization in rats,
 - figure 4A shows the results obtained by subconjunctival injection of an antisense oligonucleotide at a concentration of 60 µm,
 - figure 4B shows the results obtained after subconjunctival injection of a sense oligonucleotide at a concentration of 60 µm,

- figure 4C shows the results obtained after topical application of an antisense oligonucleotide at a concentration of 200 μ m,
- figure 4D shows the results obtained after topical application of a sense oligonucleotide at a concentration of 200 μ m,
- figure 4E illustrates the state of the cornea in the absence of any treatment,
- figure 4F illustrates the state of the cornea when treated with subconjunctival injections of PBS,
- figures 5A to 5J illustrate the results of the inhibition of corneal neovascularization obtained in different groups of rats (described in section 6.6 Results) after de-epithelialization and limbic resection of the corneas of the rats on day 4 (figures 5A to 5E) and on day 9 (figures 5F to 5J). These are slit lamp photographs showing the comparison of the growth of the vessels in the various groups of rats. Enlargement x10.

Example 1: Demonstration of the induction of the expression of IRS-1 (the 180-kDa protein) in endothelial cells resulting from the stimulation of these cells with bFGF.

The 180-kDa protein was demonstrated in the following manner:

The endothelial cells were cultured in a 6-well microtitration plate previously covered with type I collagen as described in (Montesano et al., J. Cell. Biol., 1983, 83, 1648-1652). The culture medium was DMEM (Sigma) enriched with 10% of fetal calf serum, 4 mM glutamine, 500 U/ml penicillin and 100 μ g/ml streptomycin. After 3 to 4 days of culture, there resulted a semi-confluent layer of endothelial cells. The culture medium of six wells was aspirated and replaced by fresh culture medium. Three wells were enriched with 3 ng/ml of bFGF. After incubation for 48 hours, the wells were washed three times with a phosphate buffer and the cells were used to extract the messenger RNA (mRNA) according to protocols known by the expert in the field. The mRNAs were

reverse transcribed by a polymerization chain reaction (PCR) using each of four degenerated groups of oligo (dT) (T12MN) primers, M can be G, A or C; and N is G, A, T and C. Each group of primers is imposed by the base in position 3'(N) with a degeneration in the (M) position. Example: the set of primers in which N = G is constituted by:

SEQ ID NO. 24: 5'-TTTTTTTTTTGG-3'

SEQ ID NO. 25: 5'-TTTTTTTTTTAG-3'

SEQ ID NO. 26: 5'-TTTTTTTTTTTCG-3'

The cDNAs obtained in this manner were amplified and tagged by means of an arbitrary decamer in the presence of isotopically tagged ATP. The electrophoresis analysis of the cDNAs revealed the presence of an amplified 326-bp cDNA fragment in the sample stemming from the endothelial cells stimulated with bFGF, identified in the attached sequence listing as number SEQ ID NO. 27. However, this same fragment is weakly present or present in the trace state in the sample stemming from the endothelial cells that were not stimulated with bFGF. The sequencing of this fragment and the subsequent interrogation of the databases revealed that this fragment corresponds to a part of an already known gene, coding for the substrate of the insulin receptor (an intracellular 180-kDa protein).

Example 2: Demonstration of the induction of the expression of IRS-1 (the 180-kDa protein).

Endothelial cells cultured on a layer of type I collagen stimulated or not stimulated with bFGF (cf. example 1) were lysed in a cellular lyse buffer containing sodium orthovanadate. These solutions were then clarified by centrifugation at 14,000 g for 15 minutes. Supernatant samples stemming from unstimulated cells and cells stimulated with bFGF containing equivalent amounts of proteins were then taken up with an electrophoresis solution

containing 2% SDS and 15 mM of dithiothreitol, heated at 100°C for 5 minutes then deposited on polyacrylamide gel (gradient from 4 to 15% of acrylamide) under denatured conditions (in the presence of 2% SDS). After migration, the proteins were transferred onto a nitrocellulose membrane. The membrane was blocked by incubation at ambient temperature in a 5% milk solution in a PBS buffer. The membrane was then washed three times with a PBS buffer, incubated in a PBS buffer containing 1 µg/ml of anti-IRS-1 monoclonal antibody for 2 hours at ambient temperature and washed three times with a PBS buffer. The proteins were then developed with a secondary anti-isotope antibody coupled to peroxidase. The presence was noted of a protein of molecular weight 180 kDa recognized by the monoclonal anti-IRS-1 antibody in the preparations stemming from the endothelial cells stimulated with bFGF; this protein was weakly present in the preparation stemming from the endothelial cells not simulated with bFGF (figure 1).

Example 3: Demonstration of the induction of phosphorylation at the level of IRS-1 tyrosine (the 180-kDa protein).

Human endothelial cells cultured on a layer of type I collagen stimulated or not stimulated with bFGF were lysed in a cellular lyse buffer containing sodium orthovanadate. These solutions were then clarified by centrifugation at 14,000 g for 15 minutes (cf. example 2). The IRS-1 protein was extracted by means of an anti-IRS-1 monoclonal antibody. This extraction was performed after immunoprecipitation by means of an anti-IRS-1 monoclonal antibody (Sigma). After addition of the anti-IRS-1 antibody coupled to agarose, the suspension was incubated for 2 hours at ambient temperature then centrifuged at 4000 g for 15 minutes. The resultant precipitate was taken up with an electrophoresis solution containing 2% SDS and 15 mM of dithiothreitol, heated at 100°C for 5 minutes, then deposited on polyacrylamide gel (acrylamide gradient of 4 to 15%) under

denaturing conditions (in the presence of 2% SDS). After migration, the proteins were transferred onto a nitrocellulose membrane. The membrane was blocked by incubation at ambient temperature in a 5% milk solution in a PBS buffer. The membrane was then washed three times with a PBS buffer, incubated in a PBS buffer containing 1 µg/ml of anti-phosphotyrosine monoclonal antibody for 2 hours at ambient temperature, and then washed three times with a PBS buffer. The proteins were then developed by means of a secondary anti-isotope antibody coupled to peroxidase. It was found that the IRS-1 protein of molecular weight 180 kDa was phosphorylated at the level of the tyrosine residue in the preparations stemming from the endothelial cells stimulated with bFGF; this protein was very weakly phosphorylated at the level of the tyrosine residue in the preparation stemming from the endothelial cells not stimulated with bFGF (figure 2).

Example 4: Evaluation of the in vitro antiangiogenic activity of the oligonucleotide.

Human endothelial cells were cultured on a layer of type I collagen. The culture wells were divided into four lots on the seventh day of culture:

Lot 1: Wells corresponding to the culture of untreated endothelial cells (figure 3A).

Lot 2: Wells corresponding to the culture of endothelial cells stimulated with 3 ng/ml of bFGF (figure 3B).

Lot 3: Wells corresponding to the culture of endothelial cells incubated with 100 µg/ml of oligonucleotide of SEQ ID NO. 3 for 4 hours then stimulated with 3 ng/ml of bFGF (figure 3C).

Lot 4: Wells corresponding to the culture of endothelial cells incubated with 100 µg/ml of oligonucleotide of sequence SEQ ID NO. 3 for 4 hours (figure 3D).

The various wells were examined by means of an inverted phase optical microscope after 3 to 4 days of culture. Upon reading the results, it was found that the human endothelial cells in lot 2 formed capillary tubes following stimulation with bFGF. It was also found that the oligonucleotide inhibits the formation of neovessels by these same cells stimulated with bFGF in lot 3. Finally, it was found that the oligonucleotide does not modify in a pronounced manner the growth of the endothelial cells. In fact, the numbers of endothelial cells in the lot 1 wells and in the lot 4 wells were comparable.

Example 5: Evaluation of the in vivo activity of the oligonucleotide.

Three lots of naked mice were used. Each lot was constituted by 5 mice.

Lot no. 1: This lot was used as control. Each mouse was inoculated on day 0 with 200 μ l of a suspension of B16 melanoma cells (provided by Institut Gustave Roussy, Villejuif) dispersed in PBS at the level of 10^6 cells/ml. These mice did not receive subsequent treatment.

Lot no. 2: Each mouse was inoculated subcutaneously on day 0 with 200 μ l of a suspension of B16 melanoma cells dispersed in PBS at the level of 10^6 cells/ml. On day 1, day 2, day 3, day 4, day 5, day 6, day 7, day 8, day 9 and day 10 each mouse received a subcutaneous injection of 200 μ l of an oligonucleotide solution diluted in PBS at a concentration of 500 μ g/ml. The oligonucleotide injection was performed close to the cell injection site.

Lot no. 3: The mice of this lot were not inoculated with the B16 melanoma cells. However, each of the mice received an injection of 200 μ l of an oligonucleotide solution in PBS at a concentration of 500 μ g/ml; the injections were performed on day 1, day 2, day 3, day 4, day 5, day 6, day 7, day 8, day 9 and day 10.

The following results were obtained:

In the mice of lot no. 1, the tumor mass developed very rapidly after inoculation. In fact, the tumor mass reached a size of 1.6 to 2.5 cm in diameter after ten days in the mice of said lot no. 1 (untreated mice). The evolution of the tumor mass in the mice of lot no. 2 (mice treated after inoculation by injection of oligonucleotide on day 1, day 2 and day 3), exhibited a clearly lower increase in the volume of the tumor mass. The tumor mass in the mice of lot 2 did not exceed 0.8 cm in diameter on the tenth day. On the fourteenth day, the difference between the tumor mass of the mice of lot no. 2 and those of lot no. 1 was remarkable.

In the mice of lot no. 3 (mice not having received B16 melanoma cells but treated by injection of oligonucleotide for three days), an unexpected general effect was observed on the skin. It was identical to that observed on all of the mice treated with the oligonucleotide (lot 2). The skin had an aged, crumpled appearance. The emergence of hairs was also observed on all of the treated mice. There was a parallelism during the evolution between the regression of the cutaneous signs and the resumption of tumor growth.

Thus, it was found that the oligonucleotide inhibits the development and formation of neovessels by endothelial cells *in vitro*. The oligonucleotide also has a remarkable *in vivo* antitumor activity in the naked mouse.

Example 6: Evaluation of the antiangiogenic oligonucleotide on a corneal neovascularization model in the rat.

Based on the work published by Amano et al. (1998), the applicant employed, modified and analyzed a model of the formation of corneal neovessels in the rat after de-epithelialization and limbectomy (figures 5A to 5J). It is reproducible, allows direct slit-lamp examination and quantification of the neovessels. The details are described below. The model was then used for testing the efficacy of the antiangiogenic agents of the invention.

6.1 Animals and corneal neovascularization model

Male Wister rats (*Rattus norvegicus*), aged five weeks (Charles River France, St-Aubin les Elbeufs, France), free of specific pathogens, were fed and allowed to drink water freely, and maintained in the laboratory animal facility under fixed temperature and humidity conditions, with cycles of 12 hours of light/12 hours of darkness.

The rats were anesthetized with a mixture of ketamine (Kétamine 1000, UVA, Ivry-sur-Seine, France; 128 mg/kg) and chlorpromazine (Largactil 25 mg/ml; Specia Rhône Poulen, Paris, France; 5 mg/kg), injected via the intramuscular route. A drop of oxybuprocaine (Novésine, Chibret, Clermont-Ferrand, France) was instilled in the right eye. Using an enlargement system (macroscope Wild MPS 51 S, LEICA, Heerbrugg, Switzerland), the corneal epithelium was removed by a microsponge impregnated with 70% ethanol. A 1.5-mm band of conjunctiva, at the limbus, was excised with microsurgical scissors, and the eyelids were closed by a temporary blepharorraphy with a Vicryl 5.0 thread (Dacron, Alcon, Rueil-Malmaison, France). The eye was then rinsed abundantly with 1X PBS, an oxytetracycline cream was applied (Posicycline, Alcon, France) and the blepharorraphy was opened on the fourth day [8, 9].

6.2 Treatment by subconjunctival injections and topical applications of antiangiogenic oligonucleotide

The rats were divided into 6 groups:

Group A: model + subconjunctival injection of a 60- μ M antisense oligonucleotide solution in 1X PBS,

Group B: model + topical application of a 200- μ M antisense oligonucleotide solution in 1X PBS,

Group C: model + subconjunctival injection of a 60- μ M sense oligonucleotide solution in 1X PBS,

Group D: model + topical application of a 200- μ M sense oligonucleotide solution in 1X PBS;

Group E: model + subcutaneous injection of 1X PBS;

Group F: model without treatment.

All of the rats were subjected to de-epithelialization as described above; the treatment was performed every 24 hours starting on the fourth day and continuing until the ninth day. Neovascularization was examined at the beginning, in the middle and at the end of the protocol by slit-lamp examination; photographs were taken on day 0 and day 9.

6.3 Visualization and quantification of the neovascularization

The animals were euthanized 10 days after the de-epithelialization by lethal injection of pentobarbital (intraperitoneal injection). In order to fill the microvessels and quantify the corneal neovascularization, the upper part of the animals' bodies were perfused with fluorescein-dextran 2x1,000,000. The eyes were enucleated and immersed in paraformaldehyde/1X PBS 4% for 3 hours, then overnight in 1X PBS. The cornea was then isolated with 1 mm of limbus under surgical microscope and inserted in the flat state between plate and cover by means of 3 to 5 radial incisions. The flat corneas were then examined and photographed using fluorescence microscopy. After the whole corneas were reconstituted, they were scanned and the surfaces were measured by image analysis; a software program (NIH image) was used for the quantification of the neovascularization. For each photo, the total corneal surface was measured three times as was the neovascularized surface; the ratio of the means - neovascularized surface/total corneal surface - was used to obtain the percentage of neovascularization and to measure the inhibition obtained.

6.4 Statistical analysis

The results were expressed as means \pm SD. The percentages of neovascularized surface/total surface were compared with the nonparametric test of Mann-Whitney. Values of $P < 0.05$ were considered to be significant.

6.5 Dilution of the oligonucleotide

The oligonucleotide was diluted in 1X PBS at pH 7.2. Based on the data in the literature and the experiments performed with other oligonucleotides, it was decided to use a concentration of 60 μ M for the subconjunctival injections and a concentration of 200 μ M for the topical applications.

6.6 Results

Using the model of corneal neovessels, treatment was performed with the 5'-TATCCGGAGGGCTGCCATGCTGCT-3' oligonucleotides identified under SEQ ID NO. 3 in the attached sequence listing modified in phosphorothioate form, daily, from day 4 to day 9, according to the following protocol:

Group A: subconjunctival injection of the antisense oligonucleotide at 60 μ M (AS 60),

Group B: topical application of the antisense oligonucleotide at 200 μ M (AS 200),

Group C: subconjunctival injection of the sense oligonucleotide at 60 μ M (S 60),

Group D: topical application of the sense oligonucleotide at 200 μ M (S 200),

Group E: subconjunctival injection of 1X PBS (PBS),

Group F: no treatment (0 Tt).

On the tenth day of the protocol, the rats were perfused with a solution of FITC/dextran and then euthanized. The corneas were collected and fixed in a 4% PAF solution. The corneas were then inserted in the flat state between plate

and cover in a glycerol solution. The fluorescent neovessels were observed and photographed using the fluorescence microscope. The photographs were scanned and the neovascularization percentages were measured for each animal.

The results observed are presented in table 1 below:

Table 1

	Group A AS 60	Group B AS 200	Group C S 60	Group D S 200	Group E PBS	Group F 0 Tt
Mean	0.6157	0.5058	0.9431	0.9392	0.9552	9.9170
SD	0.2194	0.1172	0.0964	0.0308	0.0481	0.0751
Number of measur ements s	15	15	15	12	9	9
SEM	0.0566	0.0303	0.0249	0.0089	0.0160	0.0250

The statistical analysis of the results using a nonparametric Mann-Whitney test yielded the following results:

The subconjunctival injections of 60- μ M of the antisense oligonucleotide (A) reduced neovascularization in relation to the control groups E and F (very significant results, $P < 0.0001$ and $P = 0.0011$); topical application of the antisense oligonucleotide at a concentration of 200 μ M (B) reduced neovascularization in relation to the control groups E and F (extremely significant results, $P < 0.0001$).

Compared to the subconjunctival administration of the sense oligonucleotide at 60 μ M (C) or the topical application of the sense oligonucleotide at 200 μ M (D), injection of the antisense oligonucleotide at 60 μ M (A) and topical application of the antisense oligonucleotide at 200 μ M (B) reduced neovascularization. These results were extremely significant ($P < 0.0001$) (figures 4A to 4F).

The inhibition of neovascularization was not significantly different depending on whether the antisense oligonucleotide was administered via the subconjunctival route (60 μ M) or applied topically (200 μ M). It was approximately 35% in relation to the controls (E and F).

The subconjunctival injection of the sense oligonucleotide at 60 μ M (C) and the topical application of the sense oligonucleotide at a concentration of 200 μ M (D) did not modify the neovascularization in relation to the control groups (E and F). In contrast, there was a small effect of the sense oligonucleotide in topical application (D) compared to the sense oligonucleotide in subconjunctival injections (C) ($P = 0.0117$).

Moreover, there was seen in the groups treated with the antisense oligonucleotide (A and B), a smaller diameter and density of the neovessels. Their distribution did not differ in relation to the control groups nor was any difference observed in relation to the level of inflammation (figure 4).

6.7 Secondary effects

No noteworthy secondary effects were seen in any of the groups during the two experimental series: after 6 days of treatment at the doses specified above, the skin of the rats was not crumpled, the fur was unchanged and the general condition of the animals was good; they fed normally until the last day and no suspicious mortality was observed. Although neither autopsies nor blood tests were performed, the general status of the animals at the end of the experiments did not suggest hepatic disorders. The only symptom observed was a transitory whitish deposit at the site of the conjunctival injections in 60% of the rats of group A, 60% of the rats of group C and 10% of the rats of group E. This deposit had been resorbed by the end of the experiments in all cases.

This example shows that — contrary to expectations — the subcutaneous injections of antisense oligonucleotide at a concentration of 60 μ M did not inhibit neovascularization to a greater extent than the topical application of the antisense oligonucleotide at a concentration of 200 μ M.

This can perhaps be explained by the difference in the concentrations employed; but this results suggests also a penetration of the oligonucleotide via the topical route rather than via the limbus. It also suggests the absence of prolonged release of the product from the injection site.

6.8 Conclusion

The application of the antisense oligonucleotide via the topical route or in subconjunctival injections reduces neovascularization in our model of corneal neovessels in the rat.

The purpose of this study was to test the efficacy of the antisense oligonucleotides stemming from the sequence of the gene IRS-1 on a previously developed model of corneal neovascularization in the rat.

This model is readily accessible, reproducible and quantifiable. This study moreover provided an initial evaluation of the concentration of oligonucleotide required *in vivo* to inhibit neovascularization.

BIBLIOGRAPHY

[pages 25 – 26 of French-language document]

All references are in English except:

5. Pierga JY, Cammilleri S, Benyahia B, Magdelénat H. Applications of antisense oligonucleotides in cancer research. *Bull Cancer* 1994; 81: 1023-1042.
9. Hoang-Xuan T, Prisant O. Restoration of corneal epithelium from limbic stem cells. *Med Sci* 1998; 14: 1375-1377.
12. Berdugo Polak M. Iontophoresis administration of antisense oligonucleotides in the anterior segment of the eye: application to a corneal neovascularization model in the rat. DEA "Biology and Pathology of the Epithelia"; University of Paris VII, Feldmann G; Inserm U450, Director Courtois Y, under the direction of Behar Cohen F. 2000.

Key to figures**Sheet 1/5 = Figure 1**

At top: N.S.: cells not stimulated with bFGF
S.: cells stimulated with bFGF
At bottom left: A - Development with anti-IRS-1 antibody
At bottom right B - Development with silver nitrate

Sheet 2/5 = Figure 2

At top: N.S.: cells not stimulated with bFGF
S.: cells stimulated with bFGF
At bottom - Development with anti-phosphotyrosine antibody

Sheet 4/5

Figure 4A. Subconjunctival injections of antisense oligonucleotides (60 μ M).
Figure 4B. Subconjunctival injections of sense oligonucleotides (60 μ M).
Figure 4C. Topical applications of antisense oligonucleotide (200 μ M).
Figure 4D. Topical applications of sense oligonucleotide (200 μ M).
Figure 4E. No treatment.
Figure 4F. Subconjunctival injections of PBS.

Sheet 5/5

Figure 5A. Day 4 AS [antisense] 60.
Figure 5B. Day 4 AS [antisense] 200.
Figure 5C. Day 4 S [sense] 200.
Figure 5D. Day 4 PBS.
Figure 5E. Day 4 No treatment.
Figure 5F. Day 9 AS [antisense] 60.
Figure 5G. Day 9 AS [antisense] 200.
Figure 5H. Day 9 S [sense] 200.
Figure 5I. Day 9 PBS.
Figure 5J. Day 9 No treatment.

THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. Use of a nucleic acid molecule, which is complementary to the nucleic acid encoding the IRS-1 protein, or a fragment thereof, and which is capable of inhibiting the expression of a nucleic acid molecule encoding the IRS-1 protein, in the preparation of a medicament for the treatment or prevention of a pathology linked to angiogenesis, by inhibiting neovascularization.
10
2. Use according to claim 1, wherein said neovascularization inhibition is obtained by the inhibition of the formation of capillary tubes by endothelial cells.
15
3. Use according to claim 1 or claim 2, wherein said nucleic acid molecule has a sequence selected from the group consisting of SEQ ID NO.2 to SEQ ID NO.23, or a fragment thereof comprising at least twelve contiguous nucleotides, and derivatives thereof.
20
4. Use according to any one of claims 1 to 3, wherein said nucleic acid molecule is antisense to at least twelve contiguous nucleotides of SEQ ID NO.28, or a derivative thereof.
25
5. Use according to any one of claims 1 to 4, wherein said nucleic acid molecule is associated with an acceptable vehicle.
30
6. Use according to any one of claims 1 to 5, wherein said pathology linked to angiogenesis is selected in the group comprising retinopathies, rheumatoid arthritis, Crohn's disease, atherosclerosis, hyperstimulation of the ovary, psoriasis, endometritis associated with neovascularization, restenosis due to balloon angioplasty, tissue superproduction due to cicatrization, peripheral
35

- vascular disease, hypertension, vascular inflammation, Raynaud's disease and Raynaud's phenomena, aneurysm, arterial restenosis, thrombophlebitis, lymphangitis, lymphedema, tissue cicatrization and repair, ischemia, 5 angina, myocardial infarction, chronic heart disease, cardiac insufficiencies such as congestive heart failure, age-related macular degeneration and osteoporosis.
7. A method for treating or preventing a pathology linked 10 to angiogenesis, by inhibiting neovascularization, comprising the step of administering a nucleic acid molecule, which encodes the IRS-1 protein, or a complementary sequence or a fragment thereof and which is capable of inhibiting the expression of a nucleic acid 15 molecule encoding the IRS-1 protein, to a patient.
8. The method according to claim 7, wherein said neovascularization inhibition is obtained by the inhibition of the formation of capillary tubes by 20 endothelial cells.
9. The method according to claim 7 or claim 8, wherein said nucleic acid molecule has a sequence selected from the group consisting of SEQ ID NO.2 to SEQ ID NO.23, or a 25 fragment thereof comprising at least twelve contiguous nucleotides, and derivatives thereof.
10. The method according to any one of claims 7 to 9, wherein said nucleic acid molecule is antisense to at 30 least twelve contiguous nucleotides of SEQ ID NO.28, or a derivative thereof.
11. The method according to any one of claims 7 to 10, wherein said nucleic acid molecule is associated with an 35 acceptable vehicle.
12. The method according to any one of claims 7 to 11,

wherein said nucleic acid molecule is administered via the subcutaneous, intramuscular, intravenous or transdermal route, in an amount from about 0.001 mg to about 50 mg.

- 5 13. The method according to any one of claims 7 to 12, wherein said pathology linked to angiogenesis is selected in the group comprising retinopathies, rheumatoid arthritis, Crohn's disease, atherosclerosis, hyperstimulation of the ovary, psoriasis, endometritis
- 10 14. The method according to any one of claims 7 to 12, associated with neovascularization, restenosis due to balloon angioplasty, tissue superproduction due to cicatrization, peripheral vascular disease, hypertension, vascular inflammation, Raynaud's disease and Raynaud's phenomena, aneurysm, arterial restenosis, thrombophlebitis, lymphangitis, lymphedema, tissue cicatrization and repair, ischemia, angina, myocardial infarction, chronic heart disease, cardiac insufficiencies such as congestive heart failure, age-related macular degeneration and osteoporosis.
- 15 15. A use according to any one of claims 1 to 6, substantially as herein described with reference to any one of the examples and/or drawings.
- 20 16. A method according to any one of claims 7 to 13, substantially as herein described with reference to any one of the examples and/or drawings.
- 25 17. A method according to any one of claims 7 to 13, substantially as herein described with reference to any one of the examples and/or drawings.

Dated this 4th day of January 2006

30 GENE SIGNAL
By their Patent Attorneys
GRIFFITH HACK
Fellows Institute of Patent and
Trade Mark Attorneys of Australia

35

n.s : cellules non stimulées avec le bFGF
s : cellules stimulées avec le bFGF

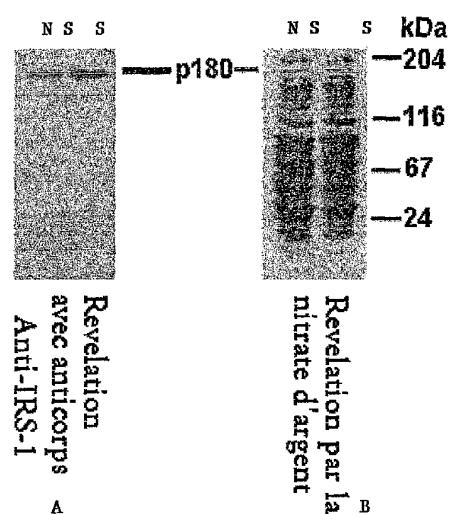
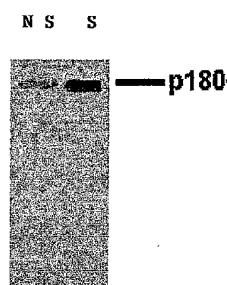


Fig.1

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_{NS}: cellules non stimulées avec le bFGF
_S: cellules stimulées avec le bFGF



Revelation avec anticorps anti-phosphotyrosine

Fig.2

3/ 5

LOT 1 LOT 2 LOT 3 LOT 4

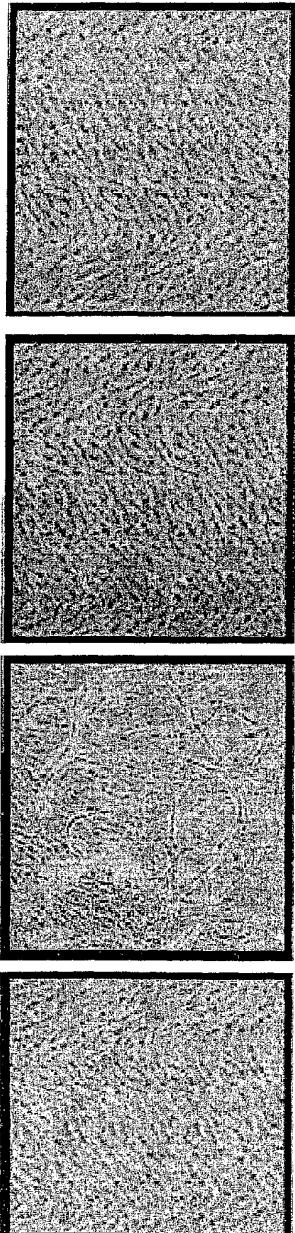


Fig.3



Figure 4A
Injections sous-conjonctivales
d'oligonucléotides antisens (60 μ M)

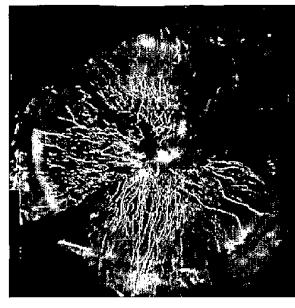


Figure 4B
Injections sous-conjonctivales
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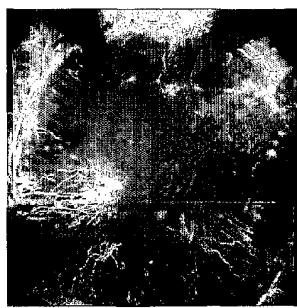


Figure 4C
Applications topiques
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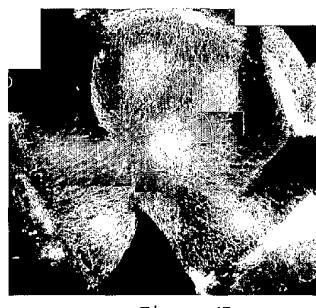


Figure 4D
Applications topiques
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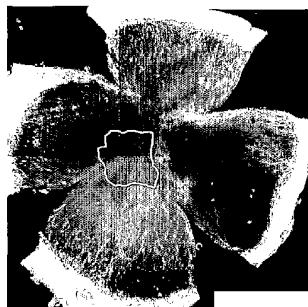


Figure 4E
Absence de traitement

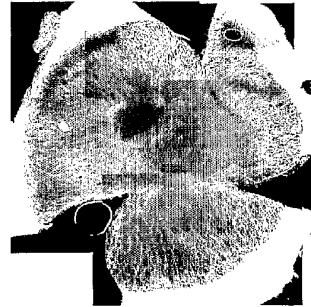


Figure 4F
Injections sous-conjonctivales de PBS

5/5



Figure 5A J4 AS60



Figure 5F J9 AS60

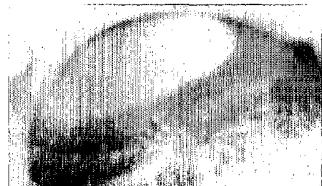


Figure 5B J4 AS200



Figure 5G J9 AS200

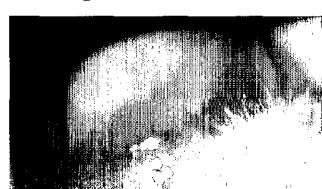


Figure 5C J4 S200



Figure 5H J9 S200

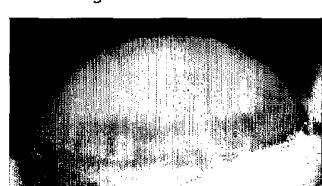


Figure 5D J4 PBS



Figure 5I J9 PBS

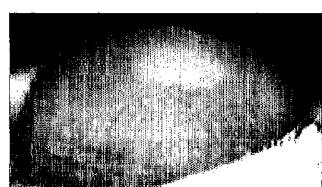


Figure 5E J4 OTt

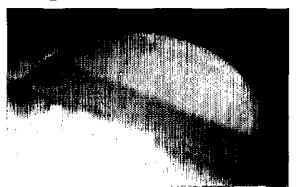


Figure 5J J9 OTt

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ccc ctt gag agc tgc ttc aac atc aac aag cgg gct gac tcc aag aac Pro Leu Glu Ser Cys Phe Asn Ile Asn Lys Arg Ala Asp Ser Lys Asn	1261
65 70 75 80	
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370

375

380

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Val Asp Pro Asn Gly Tyr Met Met Ser Pro Ser Gly Gly Cys Ser		
660 665 670		
cct gac att gga ggt ggc ccc agc agc agc agc agc aac gcc	3085	
Pro Asp Ile Gly Gly Pro Ser Ser Ser Ser Ser Asn Ala		
675 680 685		
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Val Pro Ser Gly Thr Ser Tyr Gly Lys Leu Trp Thr Asn Gly Val Gly		
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Gly His His Ser His Val Leu Pro His Pro Lys Pro Pro Val Glu Ser		
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Gly	Gly	Ser	Ser	Ser	Ser	Glu	Asp	Val	Lys	Arg	His	Ser	Ser	
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Asn Glu Lys Lys Trp Arg His Lys Ser Ser Ala Pro Lys Arg Ser Ile
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Pro Leu Glu Ser Cys Phe Asn Ile Asn Lys Arg Ala Asp Ser Lys Asn
 65 70 75 80

Lys His Leu Val Ala Leu Tyr Thr Arg Asp Glu His Phe Ala Ile Ala
 85 90 95

Ala Asp Ser Glu Ala Glu Gln Asp Ser Trp Tyr Gln Ala Leu Leu Gln
 100 105 110

Leu His Asn Arg Ala Lys Gly His His Asp Gly Ala Ala Ala Leu Gly
 115 120 125

Ala Gly Gly Gly Gly Ser Cys Ser Gly Ser Ser Gly Leu Gly Glu
 130 135 140

Ala Gly Glu Asp Leu Ser Tyr Gly Asp Val Pro Pro Gly Pro Ala Phe
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Lys Glu Val Trp Gln Val Ile Leu Lys Pro Lys Gly Leu Gln Thr
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Lys Asn Leu Ile Gly Ile Tyr Arg Leu Cys Leu Thr Ser Lys Thr Ile
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Ser Phe Val Lys Leu Asn Ser Glu Ala Ala Ala Val Val Leu Gln Leu
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Met Asn Ile Arg Arg Cys Gly His Ser Glu Asn Phe Phe Phe Ile Glu
 210 215 220

Val Gly Arg Ser Ala Val Thr Gly Pro Gly Glu Phe Trp Met Gln Val
 225 230 235 240

Asp Asp Ser Val Val Ala Gln Asn Met His Glu Thr Ile Leu Glu Ala
 245 250 255

Met Arg Ala Met Ser Asp Glu Phe Arg Pro Arg Ser Lys Ser Gln Ser

260 265 270

Ser Ser Asn Cys Ser Asn Pro Ile Ser Val Pro Leu Arg Arg His His
275 280 285

Leu Asn Asn Pro Pro Pro Ser Gln Val Gly Leu Thr Arg Arg Ser Arg
290 295 300

Thr Glu Ser Ile Thr Ala Thr Ser Pro Ala Ser Met Val Gly Gly Lys
305 310 315 320

Pro Gly Ser Phe Arg Val Arg Ala Ser Ser Asp Gly Glu Gly Thr Met
325 330 335

Ser Arg Pro Ala Ser Val Asp Gly Ser Pro Val Ser Pro Ser Thr Asn
340 345 350

Arg Thr His Ala His Arg His Arg Gly Ser Ala Arg Leu His Pro Pro
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Leu Asn His Ser Arg Ser Ile Pro Met Pro Ala Ser Arg Cys Ser Pro
370 375 380

Ser Ala Thr Ser Pro Val Ser Leu Ser Ser Ser Thr Ser Gly His
385 390 395 400

Gly Ser Thr Ser Asp Cys Leu Phe Pro Arg Arg Ser Ser Ala Ser Val
405 410 415

Ser Gly Ser Pro Ser Asp Gly Gly Phe Ile Ser Ser Asp Glu Tyr Gly
420 425 430

Ser Ser Pro Cys Asp Phe Arg Ser Ser Phe Arg Ser Val Thr Pro Asp
435 440 445

Ser Leu Gly His Thr Pro Pro Ala Arg Gly Glu Glu Leu Ser Asn
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Tyr Ile Cys Met Gly Gly Lys Gly Pro Ser Thr Leu Thr Ala Pro Asn
465 470 475 480

Gly His Tyr Ile Leu Ser Arg Gly Gly Asn Gly His Arg Cys Thr Pro
485 490 495

Gly Thr Gly Leu Gly Thr Ser Pro Ala Leu Ala Gly Asp Glu Ala Ala
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Ser Ala Ala Asp Leu Asp Asn Arg Phe Arg Lys Arg Thr His Ser Ala
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Gly Thr Ser Pro Thr Ile Thr His Gln Lys Thr Pro Ser Gln Ser Ser
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Val Ala Ser Ile Glu Glu Tyr Thr Glu Met Met Pro Ala Tyr Pro Pro
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Gly Gly Gly Ser Gly Gly Arg Leu Pro Gly His Arg His Ser Ala Phe
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Val Pro Thr Arg Ser Tyr Pro Glu Glu Gly Leu Glu Met His Pro Leu
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Glu Arg Arg Gly Gly His His Arg Pro Asp Ser Ser Thr Leu His Thr

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Gly Arg Lys Gly Ser Gly Asp Tyr Met Pro Met Ser Pro Lys Ser Val		
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Ser Ala Pro Gln Gln Ile Ile Asn Pro Ile Arg Arg His Pro Gln Arg		
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Val Asp Pro Asn Gly Tyr Met Met Ser Pro Ser Gly Gly Cys Ser		
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Pro Asp Ile Gly Gly Pro Ser Ser Ser Ser Ser Asn Ala		
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Val Pro Ser Gly Thr Ser Tyr Gly Lys Leu Trp Thr Asn Gly Val Gly		
690	695	700
Gly His His Ser His Val Leu Pro His Pro Lys Pro Pro Val Glu Ser		
705	710	715
Ser Gly Gly Lys Leu Leu Pro Cys Thr Gly Asp Tyr Met Asn Met Ser		
725	730	735
Pro Val Gly Asp Ser Asn Thr Ser Ser Pro Ser Asp Cys Tyr Tyr Gly		
740	745	750
Pro Glu Asp Pro Gln His Lys Pro Val Leu Ser Tyr Tyr Ser Leu Pro		
755	760	765
Arg Ser Phe Lys His Thr Gln Arg Pro Gly Glu Pro Glu Glu Gly Ala		
770	775	780
Arg His Gln His Leu Arg Leu Ser Thr Ser Ser Gly Arg Leu Leu Tyr		
785	790	795
Ala Ala Thr Ala Asp Asp Ser Ser Ser Thr Ser Ser Asp Ser Leu		
805	810	815
Gly Gly Gly Tyr Cys Gly Ala Arg Leu Glu Pro Ser Leu Pro His Pro		
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His His Gln Val Leu Gln Pro His Leu Pro Arg Lys Val Asp Thr Ala		
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Gln Gln Pro Leu Leu His Pro Pro Glu Pro Lys Ser Pro Gly Glu Tyr		
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Val Asn Ile Glu Phe Gly Ser Asp Gln Ser Gly Tyr Leu Ser Gly Pro		
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Val Ala Phe His Ser Ser Pro Ser Val Arg Cys Pro Ser Gln Leu Gln		
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Pro Ala Pro Arg Glu Glu Thr Gly Thr Glu Glu Tyr Met Lys Met		

930

935

940

Asp Leu Gly Pro Gly Arg Arg Ala Ala Trp Gln Glu Ser Thr Gly Val
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Glu Met Gly Arg Leu Gly Pro Ala Pro Pro Gly Ala Ala Ser Ile Cys
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Arg Pro Thr Arg Ala Val Pro Ser Ser Arg Gly Asp Tyr Met Thr Met
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