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Antisense oligonucleotides which can inhibit the formation of capillary tubes by endothelial cells

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(71) Applicant(s)
Gene Signal International SA

(72) Inventor(s)
Al-Mahmood, Salman

(74) Agent/Attorney
Griffith Hack, 509 St Kilda Road, Melbourne, VIC, 3004

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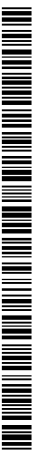


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- (71) Déposant et
(72) Inventeur : AL-MAHMOOD, Salman [FR/FR]; 2,
square Alice, F 75014 Paris (IQ).
- (74) Mandataires : BREESE, Pierre etc.; Breesé-Majerowicz,
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(54) Title: ANTISENSE OLIGONUCLEOTIDES WHICH CAN INHIBIT THE FORMATION OF CAPILLARY TUBES BY EN-
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(54) Titre : OLIGONUCLEOTIDES ANTISENS CAPABLES D'INHIBER LA FORMATION DES TUBES CAPILLAIRES PAR
DES CELLULES ENDOTHELIALES

(57) Abstract: The invention relates to pharmaceutical compositions which inhibit the formation of capillary tubes by endothelial cells, comprising at least one oligonucleotide which can inhibit the expression of the IRS-1 protein. According to the invention, the oligonucleotides are embodied as anti-angiogenesis agents. Said pharmaceutical compositions are particularly useful in treating angiogenesis-related pathologies.

(57) Abrégé : La présente invention a pour objet des compositions pharmaceutiques pour inhiber la formation de tubes capillaires par les cellules endothéliales, comprenant au moins un oligonucléotide capable d'inhiber l'expression de la protéine IRS-1. Les oligonucléotides selon l'invention sont indiqués comme agents anti-angiogéniques. Les compositions pharmaceutiques de l'invention sont particulièrement utiles pour le traitement de pathologies liées à l'angiogénèse.

**ANTISENSE OLIGONUCLEOTIDES CAPABLE OF INHIBITING THE
FORMATION OF CAPILLARY TUBES BY ENDOTHELIAL CELLS**

The present invention provides antisense oligonucleotides capable of inhibiting the expression of the protein IRS-1 and of inhibiting the formation of capillary tubes by endothelial cells. The oligonucleotides according to the invention are thus indicated as antiangiogenic agents. They are also indicated as anti-cell-multiplication agents, in particular as anti-tumor agents.

The invention also pertains to pharmaceutical compositions containing said oligonucleotides and the use of said oligonucleotides as analysis reagents.

All references, including any patents or patent applications, cited in this specification are hereby incorporated by reference. No admission is made that any reference constitutes prior art. The discussion of the references states what their authors assert, and the applicants reserve the right to challenge the accuracy and pertinency of the cited documents. It will be clearly understood that, although a number of prior art publications are referred to herein, this reference does not constitute an admission that any of these documents forms part of the common general knowledge in the art, in Australia or in any other country.

Angiogenesis is a fundamental process by means of which new blood vessels are formed. This process is essential in many normal physiological phenomena such as reproduction, development and even cicatrization. In these normal biological phenomena, angiogenesis is under strict control, i.e., it is triggered during a short period (several days) and then completely inhibited. However, many pathologies are linked to uncontrolled, invasive angiogenesis: arthritis, a pathology due to the damaging of cartilages by invasive neovessels; diabetic retinopathy or the invasion of the retina by neovessels leading to blindness of the patients; neovascularization

of the ocular apparatus presents the major cause of blindness and this neovascularization is involved in about twenty different eye diseases; and moreover the growth and metastasis of tumors which are linked directly to

5 neovascularization and are dependent on angiogenesis. The tumor stimulates the growth of neovessels by its own growth. Moreover, these neovessels are escape routes for tumors which thereby join up with the blood circulation and induce metastases in sites remote from the initial

10 tumor focus, such as the liver, lungs or bones.

Angiogenesis, the formation of neovessels by endothelial cells, involves the migration, growth and differentiation of endothelial cells. The regulation of these biological phenomena is directly linked to genetic

15 expression.

The research studies performed in the framework of the present invention made it possible to identify and prepare nucleic acid sequences involved in the regulation of angiogenesis.

20 Other studies pertaining to angiogenesis have shown a noteworthy expression and phosphorylation at the level of a tyrosine residue of an intracellular 180-kDa protein by endothelial cells cultured on a surface of type I collagen and stimulated by an angiogenic factor such as bFGF. The

25 noteworthy expression and phosphorylation at the level of the tyrosine residue of the intracellular 180-kDa protein accompanies the formation of capillary tubes by the endothelial cells.

This protein is already known as a substrate of the

30 insulin receptor (called IRS-1). It has been partially identified and investigated by certain diabetes researchers (Quon et al., J. Biol. Chem. (1994), 269 (45), 27920-27924).

These authors studied the role of IRS-1 in (i) the

35 translocation of GLUT 4 stimulated by insulin and (ii) the transport of glucose in rat adipose cells. In order to do this, they constructed a plasmid containing:

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- a double chain oligonucleotide obtained from the sense oligonucleotide of the following sequence SEQ ID NO. ID No. 1:5'-TCGATGTGAC GCTACTGATG AGTCCGTGAG GACGAAACTC TGGCCTAG-3'; and
- cDNA coding for human IRS-1, and transfected rat adipose cells with said plasmid.

The research performed in the framework of the present invention revealed that the expression of the protein IRS-1 is also induced in endothelial cells when said cells are stimulated by the angiogenic factor bFGF.

The invention thus pertains to a pharmaceutical composition active on angiogenesis phenomena comprising as active agent at least one substance selected from among: (i) a nucleic acid molecule of the gene coding for the protein IRS-1, a complementary sequence or a fragment thereof, (ii) a molecule capable of inhibiting the expression of a nucleic acid molecule according to (i).

In the framework of the invention, antisense oligonucleotides of the gene coding for this protein were prepared. These oligonucleotides present remarkable antiangiogenic and antitumor activities. They are therefore particularly useful in the treatment of diseases linked to invasive angiogenesis not controlled by gene therapy methods consisting of administering to an individual a composition containing at least one of these oligonucleotides.

Thus, an oligonucleotide according to the invention is constituted by the following nucleotide sequence of formula SEQ ID NO. 2:

5'-TATCCGGAGGGCTCGCCATGCTGCTGCGGAGCAGA-3',

a fragment thereof comprising at least 12 contiguous nucleotides or their derivative.

The invention pertains most particularly to an oligonucleotide constituted by one of the nucleotide sequences of formulas SEQ ID NO. 3 and 4 below:

5'-TATCCGGAGGGCTCGCCATGCTGCT-3'.

5'-TCGCCATGCTGCTGCGGAGCAGA-3',

a fragment of these comprising at least 12 contiguous nucleotides or their derivative.

The term derivative is understood to mean a sequence capable of hybridizing under strict conditions with one of the sequences SEQ ID NO. 2, 3 or 4, or with a fragment of these of at least 12 contiguous nucleotides.

The following sequences can be cited as examples of oligonucleotides according to the invention:

SEQ ID NO. 5: 5'-TATCCGGAGGGCCTGCCATGCTGCT-3',
SEQ ID NO. 6: 5'-TATCCGGAGG GCCTGCCATG CTGC-3',
SEQ ID NO. 7: 5'-TATCCGGAGG GCCTGCCATG CTG-3',
SEQ ID NO. 8: 5'-TATCCGGAGG GCCTGCCATG CT-3',
SEQ ID NO. 9: 5'-TATCCGGAGG GCCTGCCATG C-3',
SEQ ID NO. 10: 5'-TATCCGGAGG GCCTGCCATG-3',
SEQ ID NO. 11: 5'-TATCCGGAGG GCCTGCCAT-3',
SEQ ID NO. 12: 5'-TATCCGGAGG GCCTGCCA-3',
SEQ ID NO. 13: 5'-TATCCGGAGG GCCTGCC-3',
SEQ ID NO. 14: 5'-TATCCGGAGG GCCTGC-3',
SEQ ID NO. 15: 5'-TATCCGGAGG GCCTG-3',
SEQ ID NO. 16: 5'-TATCCGGAGG GCCT-3',
SEQ ID NO. 17: 5'-TATCCGGAGG GCC-3',
SEQ ID NO. 18: 5'-TATCCGGAGG GC-3',
SEQ ID NO. 19: 5'-CCGGAGG GCCTGCCATG CTGCT-3',
SEQ ID NO. 20: 5'-GAGG GCCTGCCATG CTGCT-3',
SEQ ID NO. 21: 5'-G GCCTGCCATG CTGCT-3',
SEQ ID NO. 22: 5'-CTGCCATG CTGCT-3',
SEQ ID NO. 23: 5'-TGCCATG CTGCT-3'.

All or part of the phosphodiester bonds of the invention are advantageously protected. This protection is generally implemented via the chemical route using classic methods that are well known by the expert in the field. The phosphodiester bonds can be protected, for example, by a thiol or amine functional group or by a phenyl group.

The 5'- and/or 3'- ends of the oligonucleotides of the invention are also advantageously protected, for example, using the technique described above for protecting the phosphodiester bonds.

The oligonucleotides of the invention can be synthesized using conventional techniques that are well known to the expert in the field, for example, using one of the DNA synthesizers marketed by various companies.

Although their mechanism of action has not been entirely elucidated, the oligonucleotides according to the invention inhibit the expression of the protein IRS-1 within endothelial cells. These oligonucleotides are capable of blocking the formation of neovessels by endothelial cells (i.e., they inhibit angiogenesis) and thus they inhibit the multiplication of tumor cells in mice.

The invention therefore also has as object a pharmaceutical composition for the inhibition of the gene coding for the protein IRS-1 comprising at least one oligonucleotide complementary of a part of said gene or of a transcript of said gene.

The molecule capable of inhibiting the expression of a nucleic acid molecule of the gene coding for the protein IRS-1 is preferably an antisense sequence of the region coding the sequence identified under the number SEQ ID NO. 28 in the attached sequence list.

Said antisense sequence advantageously comprises at least twelve contiguous nucleotides or their derivative.

More preferentially, the active agent capable of inhibiting the expression of a nucleic acid molecule coding for the protein IRS-1 of the composition of the invention is a nucleotide sequence selected from among the set of nucleotide sequences identified as SEQ ID NO. 2 to SEQ ID NO. 23 in the attached sequence list comprising at least twelve contiguous nucleotides or their derivative.

Such a composition advantageously comprises as active agent at least one oligonucleotide as defined above advantageously combined in said composition with an acceptable vehicle.

The analysis of the research performed in the framework of the invention made it possible to demonstrate that the protein IRS-1 represents a cellular constituent which is essential in the angiogenesis process. In fact, inhibition of the expression of the protein IRS-1 by said antisense oligonucleotides leads to the inhibition of the formation of capillary tubes by endothelial cells.

The oligonucleotides according to the invention and the compositions containing them are thus indicated as antiangiogenic agents. They are also indicated as anti-cell-multiplication agents, particularly as antitumor agents, and consequently are particularly useful for the treatment of tumors. Thus the present invention provides the use of said oligonucleotides for the preparation of a composition intended for the treatment or prevention of pathologies linked to invasive, uncontrolled angiogenesis such as, as nonlimitative example: the treatment of tumour vascularization, eye diseases linked to the neovascularization of the ocular apparatus such as retinopathies, rheumatoid arthritis, Crohn's disease, atherosclerosis, hyperstimulation of the ovary, psoriasis, endometritis associated with neovascularization, restenosis due to balloon angioplasty, tissue superproduction due to cicatrization, peripheral vascular disease, hypertension, vascular inflammation, Raynaud's disease and Raynaud's phenomena, aneurysm, arterial restenosis, thrombophlebitis, lymphangitis, lymphedema, tissue cicatrization and repair, ischemia, angina, myocardial infarction, chronic heart disease, cardiac insufficiencies such as congestive heart failure, age-related macular degeneration and osteoporosis.

The above pharmaceutical compositions are more particularly implemented in a manner such that they can be administered via the subcutaneous, intramuscular, intravenous or transdermal route. For such administration, use is made notably of aqueous suspensions, isotonic saline solutions or sterile,

Injectable solutions containing pharmacologically compatible dispersion agents and/or wetting agents such as, for example, propylene glycol or butylene glycol.

The usual unit dose to be administered contains from
5 0.001 mg to 50 mg of active principle.

The oligonucleotides of the invention are also useful as research reagents, notably for the *in vitro* study of signalization routes involving the 180-kDa protein, for example on tumor cells or non-tumor cells transfected by
10 said oligonucleotides. They are also useful for the *in vivo* study of signalization routes involving the 180-kDa protein in a large number of physiological and pathological phenomena such as angiogenesis or carcinogenesis essentially from the kinase/phosphatase
15 ratio.

Thus, the pharmaceutical compositions of the invention are particularly useful for the performance of tests for the diagnosis of pathologies linked to angiogenesis phenomena, notably for the diagnosis of retinopathies,
20 rheumatoid arthritis, Crohn's disease, atherosclerosis, hyperstimulation of the ovary, psoriasis, endometritis association with neovascularization, restenosis due to balloon angioplasty, tissue superproduction due to cicatrization, peripheral vascular disease, hypertension,
25 vascular inflammation, Raynaud's disease and Raynaud's phenomena, aneurysm, arterial restenosis, thrombophlebitis, lymphangitis, lymphedema, tissue cicatrization and repair, ischemia, angina, myocardial infarction, chronic heart disease, cardiac insufficiencies
30 such as congestive heart failure or age-linked macular degeneration and osteoporosis.

In the claims which follow and in the description of the invention, except where the context requires otherwise due to express language or necessary implication, the word
35 "comprise" or variations such as "comprises" or "comprising" is used in an inclusive sense, i.e. to specify the presence of the stated features but not to

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preclude the presence or addition of further features in various embodiments of the invention.

Other advantages and characteristics of the invention will become clear from the examples below in which the
5 term "oligonucleotide" is used to designate the oligonucleotide of SEQ ID NO.3 and which refer to the attached figures in which:

- figure 1A represents the Western Blot images obtained from supernatant samples stemming from unstimulated cells (track NS) and cells stimulated with bFGF (track S) developed with an anti-IRS-1 antibody,
- figure 1B shows the Western Blot images obtained after staining with silver nitrate obtained from the same supernatant samples stemming from unstimulated cells (track NS) and cells stimulated with bFGF (track S),
- figure 2 shows the Western Blot images obtained from supernatant cells stemming from unstimulated cells (track NS) and cells stimulated with bFGF (track B) when the membrane is incubated with an anti-phosphotyrosine monoclonal antibody and developed with an anti-isotope antibody tagged at the peroxidase as indicated in example 3,
- figures 3A to 3D show the images of the cultures on a type I collagen surface of the different lots of endothelial cells:
 - figure 3A shows the culture of untreated endothelial cells,
 - figure 3B shows the culture of endothelial cells stimulated with 3 ng/ml of bFGF,
 - figure 3C shows the culture of endothelial cells incubated with 100 µg/ml of oligonucleotide of SEQ ID NO. 3 for 4 hours and then stimulated with 3 ng/ml of bFGF,
 - figure 3D shows the culture of endothelial cells incubated with 100 µg/ml of oligonucleotide of SEQ ID NO. 3 for 4 hours,
- figures 4A to 4F illustrate the results of tests of the inhibition of corneal neovascularization in rats,
 - figure 4A shows the results obtained by subconjunctival injection of an antisense oligonucleotide at a concentration of 60 µM,
 - figure 4B shows the results obtained after subconjunctival injection of a sense oligonucleotide at a concentration of 60 µM,

- figure 4C shows the results obtained after topical application of an antisense oligonucleotide at a concentration of 200 μ m,
- figure 4D shows the results obtained after topical application of a sense oligonucleotide at a concentration of 200 μ m,
- figure 4E illustrates the state of the cornea in the absence of any treatment,
- figure 4F illustrates the state of the cornea when treated with subconjunctival injections of PBS,
- figures 5A to 5J illustrate the results of the inhibition of corneal neovascularization obtained in different groups of rats (described in section 6.6 Results) after de-epithelialization and limbic resection of the corneas of the rats on day 4 (figures 5A to 5E) and on day 9 (figures 5F to 5J). These are slit lamp photographs showing the comparison of the growth of the vessels in the various groups of rats. Enlargement x10.

Example 1: Demonstration of the induction of the expression of IRS-1 (the 180-kDa protein) in endothelial cells resulting from the stimulation of these cells with bFGF.

The 180-kDa protein was demonstrated in the following manner:

The endothelial cells were cultured in a 6-well microtitration plate previously covered with type I collagen as described in (Montesano et al., J. Cell. Biol., 1983, 83, 1648-1652). The culture medium was DMEM (Sigma) enriched with 10% of fetal calf serum, 4 mM glutamine, 500 U/ml penicillin and 100 μ g/ml streptomycin. After 3 to 4 days of culture, there resulted a semi-confluent layer of endothelial cells. The culture medium of six wells was aspirated and replaced by fresh culture medium. Three wells were enriched with 3 ng/ml of bFGF. After incubation for 48 hours, the wells were washed three times with a phosphate buffer and the cells were used to extract the messenger RNA (mRNA) according to protocols known by the expert in the field. The mRNAs were

reverse transcribed by a polymerization chain reaction (PCR) using each of four degenerated groups of oligo (dT) (T12MN) primers, M can be G, A or C; and N is G, A, T and C. Each group of primers is imposed by the base in position 3'(N) with a degeneration in the (M) position. Example: the set of primers in which N = G is constituted by:

SEQ ID NO. 24: 5'-TTTTTTTTTTTGG-3'

SEQ ID NO. 25: 5'-TTTTTTTTTTTAG-3'

SEQ ID NO. 26: 5'-TTTTTTTTTTTCG-3'

The cDNAs obtained in this manner were amplified and tagged by means of an arbitrary decamer in the presence of isotopically tagged ATP. The electrophoresis analysis of the cDNAs revealed the presence of an amplified 326-bp cDNA fragment in the sample stemming from the endothelial cells stimulated with bFGF, identified in the attached sequence listing as number SEQ ID NO. 27. However, this same fragment is weakly present or present in the trace state in the sample stemming from the endothelial cells that were not stimulated with bFGF. The sequencing of this fragment and the subsequent interrogation of the databases revealed that this fragment corresponds to a part of an already known gene, coding for the substrate of the insulin receptor (an intracellular 180-kDa protein).

Example 2: Demonstration of the induction of the expression of IRS-1 (the 180-kDa protein).

Endothelial cells cultured on a layer of type I collagen stimulated or not stimulated with bFGF (cf. example 1) were lysed in a cellular lyse buffer containing sodium orthovanadate. These solutions were then clarified by centrifugation at 14,000 g for 15 minutes. Supernatant samples stemming from unstimulated cells and cells stimulated with bFGF containing equivalent amounts of proteins were then taken up with an electrophoresis solution

containing 2% SDS and 15 mM of dithiothreitol, heated at 100°C for 5 minutes then deposited on polyacrylamide gel (gradient from 4 to 15% of acrylamide) under denatured conditions (in the presence of 2% SDS). After migration, the proteins were transferred onto a nitrocellulose membrane. The membrane was blocked by incubation at ambient temperature in a 5% milk solution in a PBS buffer. The membrane was then washed three times with a PBS buffer, incubated in a PBS buffer containing 1 µg/ml of anti-IRS-1 monoclonal antibody for 2 hours at ambient temperature and washed three times with a PBS buffer. The proteins were then developed with a secondary anti-isotope antibody coupled to peroxidase. The presence was noted of a protein of molecular weight 180 kDa recognized by the monoclonal anti-IRS-1 antibody in the preparations stemming from the endothelial cells stimulated with bFGF; this protein was weakly present in the preparation stemming from the endothelial cells not simulated with bFGF (figure 1).

Example 3: Demonstration of the induction of phosphorylation at the level of IRS-1 tyrosine (the 180-kDa protein).

Human endothelial cells cultured on a layer of type I collagen stimulated or not stimulated with bFGF were lysed in a cellular lyse buffer containing sodium orthovanadate. These solutions were then clarified by centrifugation at 14,000 g for 15 minutes (cf. example 2). The IRS-1 protein was extracted by means of an anti-IRS-1 monoclonal antibody. This extraction was performed after immunoprecipitation by means of an anti-IRS-1 monoclonal antibody (Sigma). After addition of the anti-IRS-1 antibody coupled to agarose, the suspension was incubated for 2 hours at ambient temperature then centrifuged at 4000 g for 15 minutes. The resultant precipitate was taken up with an electrophoresis solution containing 2% SDS and 15 mM of dithiothreitol, heated at 100°C for 5 minutes, then deposited on polyacrylamide gel (acrylamide gradient of 4 to 15%) under

denaturing conditions (in the presence of 2% SDS). After migration, the proteins were transferred onto a nitrocellulose membrane. The membrane was blocked by incubation at ambient temperature in a 5% milk solution in a PBS buffer. The membrane was then washed three times with a PBS buffer, incubated in a PBS buffer containing 1 µg/ml of anti-phosphotyrosine monoclonal antibody for 2 hours at ambient temperature, and then washed three times with a PBS buffer. The proteins were then developed by means of a secondary anti-isotope antibody coupled to peroxidase. It was found that the IRS-1 protein of molecular weight 180 kDa was phosphorylated at the level of the tyrosine residue in the preparations stemming from the endothelial cells stimulated with bFGF; this protein was very weakly phosphorylated at the level of the tyrosine residue in the preparation stemming from the endothelial cells not stimulated with bFGF (figure 2).

Example 4: Evaluation of the in vitro antiangiogenic activity of the oligonucleotide.

Human endothelial cells were cultured on a layer of type I collagen. The culture wells were divided into four lots on the seventh day of culture:

Lot 1: Wells corresponding to the culture of untreated endothelial cells (figure 3A).

Lot 2: Wells corresponding to the culture of endothelial cells stimulated with 3 ng/ml of bFGF (figure 3B).

Lot 3: Wells corresponding to the culture of endothelial cells incubated with 100 µg/ml of oligonucleotide of SEQ ID NO. 3 for 4 hours then stimulated with 3 ng/ml of bFGF (figure 3C).

Lot 4: Wells corresponding to the culture of endothelial cells incubated with 100 µg/ml of oligonucleotide of sequence SEQ ID NO. 3 for 4 hours (figure 3D).

The various wells were examined by means of an inverted phase optical microscope after 3 to 4 days of culture. Upon reading the results, it was found that the human endothelial cells in lot 2 formed capillary tubes following stimulation with bFGF. It was also found that the oligonucleotide inhibits the formation of neovessels by these same cells stimulated with bFGF in lot 3. Finally, it was found that the oligonucleotide does not modify in a pronounced manner the growth of the endothelial cells. In fact, the numbers of endothelial cells in the lot 1 wells and in the lot 4 wells were comparable.

Example 5: Evaluation of the in vivo activity of the oligonucleotide.

Three lots of naked mice were used. Each lot was constituted by 5 mice.

Lot no. 1: This lot was used as control. Each mouse was inoculated on day 0 with 200 µl of a suspension of B16 melanoma cells (provided by Institut Gustave Roussy, Villejuif) dispersed in PBS at the level of 10^6 cells/ml. These mice did not receive subsequent treatment.

Lot no. 2: Each mouse was inoculated subcutaneously on day 0 with 200 µl of a suspension of B16 melanoma cells dispersed in PBS at the level of 10^6 cells/ml. On day 1, day 2, day 3, day 4, day 5, day 6, day 7, day 8, day 9 and day 10 each mouse received a subcutaneous injection of 200 µl of an oligonucleotide solution diluted in PBS at a concentration of 500 µg/ml. The oligonucleotide injection was performed close to the cell injection site.

Lot no. 3: The mice of this lot were not inoculated with the B16 melanoma cells. However, each of the mice received an injection of 200 µl of an oligonucleotide solution in PBS at a concentration of 500 µg/ml; the injections were performed on day 1, day 2, day 3, day 4, day 5, day 6, day 7, day 8, day 9 and day 10.

The following results were obtained:

In the mice of lot no. 1, the tumor mass developed very rapidly after inoculation. In fact, the tumor mass reached a size of 1.6 to 2.5 cm in diameter after ten days in the mice of said lot no. 1 (untreated mice). The evolution of the tumor mass in the mice of lot no. 2 (mice treated after inoculation by injection of oligonucleotide on day 1, day 2 and day 3), exhibited a clearly lower increase in the volume of the tumor mass. The tumor mass in the mice of lot 2 did not exceed 0.8 cm in diameter on the tenth day. On the fourteenth day, the difference between the tumor mass of the mice of lot no. 2 and those of lot no. 1 was remarkable.

In the mice of lot no. 3 (mice not having received B16 melanoma cells but treated by injection of oligonucleotide for three days), an unexpected general effect was observed on the skin. It was identical to that observed on all of the mice treated with the oligonucleotide (lot 2). The skin had an aged, crumpled appearance. The emergence of hairs was also observed on all of the treated mice. There was a parallelism during the evolution between the regression of the cutaneous signs and the resumption of tumor growth.

Thus, it was found that the oligonucleotide inhibits the development and formation of neovessels by endothelial cells *in vitro*. The oligonucleotide also has a remarkable *in vivo* antitumor activity in the naked mouse.

Example 6: Evaluation of the antiangiogenic oligonucleotide on a corneal neovascularization model in the rat.

Based on the work published by Amano et al. (1998), the applicant employed, modified and analyzed a model of the formation of corneal neovessels in the rat after de-epithelialization and limbectomy (figures 5A to 5J). It is reproducible, allows direct slit-lamp examination and quantification of the neovessels. The details are described below. The model was then used for testing the efficacy of the antiangiogenic agents of the invention.

6.1 Animals and corneal neovascularization model

Male Wister rats (*Rattus norvegicus*), aged five weeks (Charles River France, St-Aubin les Elbeufs, France), free of specific pathogens, were fed and allowed to drink water freely, and maintained in the laboratory animal facility under fixed temperature and humidity conditions, with cycles of 12 hours of light/12 hours of darkness.

The rats were anesthetized with a mixture of ketamine (Kétamine 1000, UVA, Ivry-sur-Seine, France; 128 mg/kg) and chlorpromazine (Largactil 25 mg/ml; Specia Rhône Poulenc, Paris, France; 5 mg/kg), injected via the intramuscular route. A drop of oxybuprocaine (Novésine, Chibret, Clermont-Ferrand, France) was instilled in the right eye. Using an enlargement system (macroscope Wild MPS 51 S, LEICA, Heerbrugg, Switzerland), the corneal epithelium was removed by a micro sponge impregnated with 70% ethanol. A 1.5-mm band of conjunctiva, at the limbus, was excised with microsurgical scissors, and the eyelids were closed by a temporary blepharorrhaphy with a Vicryl 5.0 thread (Dacron, Alcon, Rueil-Malmaison, France). The eye was then rinsed abundantly with 1X PBS, an oxytetracycline cream was applied (Posicycline, Alcon, France) and the blepharorrhaphy was opened on the fourth day [8, 9].

6.2 Treatment by subconjunctival injections and topical applications of antiangiogenic oligonucleotide

The rats were divided into 6 groups:

Group A: model + subconjunctival injection of a 60- μ M antisense oligonucleotide solution in 1X PBS,

Group B: model + topical application of a 200- μ M antisense oligonucleotide solution in 1X PBS,

Group C: model + subconjunctival injection of a 60- μ M sense oligonucleotide solution in 1X PBS,

Group D: model + topical application of a 200- μ M sense oligonucleotide solution in 1X PBS,

Group E: model + subcutaneous injection of 1X PBS,

Group F: model without treatment.

All of the rats were subjected to de-epithelialization as described above; the treatment was performed every 24 hours starting on the fourth day and continuing until the ninth day. Neovascularization was examined at the beginning, in the middle and at the end of the protocol by slit-lamp examination; photographs were taken on day 0 and day 9.

6.3 Visualization and quantification of the neovascularization

The animals were euthanized 10 days after the de-epithelialization by lethal injection of pentobarbital (intraperitoneal injection). In order to fill the microvessels and quantify the corneal neovascularization, the upper part of the animals' bodies were perfused with fluorescein-dextran 2x1,000,000. The eyes were enucleated and immersed in paraformaldehyde/1X PBS 4% for 3 hours, then overnight in 1X PBS. The cornea was then isolated with 1 mm of limbus under surgical microscope and inserted in the flat state between plate and cover by means of 3 to 5 radial incisions. The flat corneas were then examined and photographed using fluorescence microscopy. After the whole corneas were reconstituted, they were scanned and the surfaces were measured by image analysis; a software program (NIH image) was used for the quantification of the neovascularization. For each photo, the total corneal surface was measured three times as was the neovascularized surface; the ratio of the means - neovascularized surface/total corneal surface - was used to obtain the percentage of neovascularization and to measure the inhibition obtained.

6.4 Statistical analysis

The results were expressed as means \pm SD. The percentages of neovascularized surface/total surface were compared with the nonparametric test of Mann-Whitney. Values of $P < 0.05$ were considered to be significant.

6.5 Dilution of the oligonucleotide

The oligonucleotide was diluted in 1X PBS at pH 7.2. Based on the data in the literature and the experiments performed with other oligonucleotides, it was decided to use a concentration of 60 μ M for the subconjunctival injections and a concentration of 200 μ M for the topical applications.

6.6 Results

Using the model of corneal neovessels, treatment was performed with the 5'-TATCCGGAGGGCTCGCCATGCTGCT-3' oligonucleotides identified under SEQ ID NO. 3 in the attached sequence listing modified in phosphorothioate form, daily, from day 4 to day 9, according to the following protocol:

Group A: subconjunctival injection of the antisense oligonucleotide at 60 μ M (AS 60),

Group B: topical application of the antisense oligonucleotide at 200 μ M (AS 200),

Group C: subconjunctival injection of the sense oligonucleotide at 60 μ M (S 60),

Group D: topical application of the sense oligonucleotide at 200 μ M (S 200),

Group E: subconjunctival injection of 1X PBS (PBS),

Group F: no treatment (0 Tt).

On the tenth day of the protocol, the rats were perfused with a solution of FITC/dextran and then euthanized. The corneas were collected and fixed in a 4% PAF solution. The corneas were then inserted in the flat state between plate

and cover in a glycerol solution. The fluorescent neovessels were observed and photographed using the fluorescence microscope. The photographs were scanned and the neovascularization percentages were measured for each animal.

The results observed are presented in table 1 below:

Table 1

	Group A AS 60	Group B AS 200	Group C S 60	Group D S 200	Group E PBS	Group F 0 Tt
Mean	0.6157	0.5058	0.9431	0.9392	0.9552	9.9170
SD	0.2194	0.1172	0.0964	0.0308	0.0481	0.0751
Number of measurements	15	15	15	12	9	9
SEM	0.0566	0.0303	0.0249	0.0089	0.0160	0.0250

The statistical analysis of the results using a nonparametric Mann-Whitney test yielded the following results:

The subconjunctival injections of 60- μ M of the antisense oligonucleotide (A) reduced neovascularization in relation to the control groups E and F (very significant results, $P < 0.0001$ and $P = 0.0011$); topical application of the antisense oligonucleotide at a concentration of 200 μ M (B) reduced neovascularization in relation to the control groups E and F (extremely significant results, $P < 0.0001$).

Compared to the subconjunctival administration of the sense oligonucleotide at 60 μ M (C) or the topical application of the sense oligonucleotide at 200 μ M (D), injection of the antisense oligonucleotide at 60 μ M (A) and topical application of the antisense oligonucleotide at 200 μ M (B) reduced neovascularization. These results were extremely significant ($P < 0.0001$) (figures 4A to 4F).

The inhibition of neovascularization was not significantly different depending on whether the antisense oligonucleotide was administered via the subconjunctival route (60 μ M) or applied topically (200 μ M). It was approximately 35% in relation to the controls (E and F).

The subconjunctival injection of the sense oligonucleotide at 60 μ M (C) and the topical application of the sense oligonucleotide at a concentration of 200 μ M (D) did not modify the neovascularization in relation to the control groups (E and F). In contrast, there was a small effect of the sense oligonucleotide in topical application (D) compared to the sense oligonucleotide in subconjunctival injections (C) ($P = 0.0117$).

Moreover, there was seen in the groups treated with the antisense oligonucleotide (A and B), a smaller diameter and density of the neovessels. Their distribution did not differ in relation to the control groups nor was any difference observed in relation to the level of inflammation (figure 4).

6.7 Secondary effects

No noteworthy secondary effects were seen in any of the groups during the two experimental series: after 6 days of treatment at the doses specified above, the skin of the rats was not crumpled, the fur was unchanged and the general condition of the animals was good; they fed normally until the last day and no suspicious mortality was observed. Although neither autopsies nor blood tests were performed, the general status of the animals at the end of the experiments did not suggest hepatic disorders. The only symptom observed was a transitory whitish deposit at the site of the conjunctival injections in 60% of the rats of group A, 60% of the rats of group C and 10% of the rats of group E. This deposit had been resorbed by the end of the experiments in all cases.

This example shows that — contrary to expectations — the subcutaneous injections of antisense oligonucleotide at a concentration of 60 μ M did not inhibit neovascularization to a greater extent than the topical application of the antisense oligonucleotide at a concentration of 200 μ M.

This can perhaps be explained by the difference in the concentrations employed; but this results suggests also a penetration of the oligonucleotide via the topical route rather than via the limbus. It also suggests the absence of prolonged release of the product from the injection site.

6.8 Conclusion

The application of the antisense oligonucleotide via the topical route or in subconjunctival injections reduces neovascularization in our model of corneal neovessels in the rat.

The purpose of this study was to test the efficacy of the antisense oligonucleotides stemming from the sequence of the gene IRS-1 on a previously developed model of corneal neovascularization in the rat.

This model is readily accessible, reproducible and quantifiable. This study moreover provided an initial evaluation of the concentration of oligonucleotide required *in vivo* to inhibit neovascularization.

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[pages 25 – 26 of French-language document]

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5. Pierga JY, Cammilleri S, Benyahia B, Magdelénat H. Applications of antisense oligonucleotides in cancer research. Bull Cancer 1994; 81: 1023-1042.

9. Hoang-Xuan T, Prisant O. Restoration of corneal epithelium from limbic stem cells. Med Sci 1998; 14: 1375-1377.

12. Berdugo Polak M. Iontophoresis administration of antisense oligonucleotides in the anterior segment of the eye: application to a corneal neovascularization model in the rat. DEA "Biology and Pathology of the Epithelia"; University of Paris VII, Feldmann G; Inserm U450, Director Courtois Y, under the direction of Behar Cohen F. 2000.

Key to figures

Sheet 1/5 = Figure 1

At top: N.S.: cells not stimulated with bFGF

S.: cells stimulated with bFGF

At bottom left: A - Development with anti-IRS-1 antibody

At bottom right B - Development with silver nitrate

Sheet 2/5 = Figure 2

At top: N.S.: cells not stimulated with bFGF

S.: cells stimulated with bFGF

At bottom - Development with anti-phosphotyrosine antibody

Sheet 4/5

Figure 4A. Subconjunctival injections of antisense oligonucleotides (60 μ M).

Figure 4B. Subconjunctival injections of sense oligonucleotides (60 μ M).

Figure 4C. Topical applications of antisense oligonucleotide (200 μ M).

Figure 4D. Topical applications of sense oligonucleotide (200 μ M).

Figure 4E. No treatment.

Figure 4F. Subconjunctival injections of PBS.

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Figure 5A. Day 4 AS [antisense] 60.

Figure 5B. Day 4 AS [antisense] 200.

Figure 5C. Day 4 S [sense] 200.

Figure 5D. Day 4 PBS.

Figure 5E. Day 4 No treatment.

Figure 5F. Day 9 AS [antisense] 60.

Figure 5G. Day 9 AS [antisense] 200.

Figure 5H. Day 9 S [sense] 200.

Figure 5I. Day 9 PBS.

Figure 5J. Day 9 No treatment.

THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. Use of a nucleic acid molecule, which is complementary to the nucleic acid encoding the IRS-1 protein, or a
5 fragment thereof, and which is capable of inhibiting the expression of a nucleic acid molecule encoding the IRS-1 protein, in the preparation of a medicament for the treatment or prevention of a pathology linked to angiogenesis, by inhibiting neovascularization.
10
2. Use according to claim 1, wherein said neovascularization inhibition is obtained by the inhibition of the formation of capillary tubes by endothelial cells.
15
3. Use according to claim 1 or claim 2, wherein said nucleic acid molecule has a sequence selected from the group consisting of SEQ ID NO.2 to SEQ ID NO.23, or a fragment thereof comprising at least twelve contiguous
20 nucleotides, and derivatives thereof.
4. Use according to any one of claims 1 to 3, wherein said nucleic acid molecule is antisense to at least twelve contiguous nucleotides of SEQ ID NO.28, or a derivative
25 thereof.
5. Use according to any one of claims 1 to 4, wherein said nucleic acid molecule is associated with an acceptable vehicle.
30
6. Use according to any one of claims 1 to 5, wherein said pathology linked to angiogenesis is selected in the group comprising retinopathies, rheumatoid arthritis, Crohn's disease, atherosclerosis, hyperstimulation of the
35 ovary, psoriasis, endometritis associated with neovascularization, restenosis due to balloon angioplasty, tissue superproduction due to cicatrization, peripheral

vascular disease, hypertension, vascular inflammation, Raynaud's disease and Raynaud's phenomena, aneurysm, arterial restenosis, thrombophlebitis, lymphangitis, lymphedema, tissue cicatrization and repair, ischemia, angina, myocardial infarction, chronic heart disease, cardiac insufficiencies such as congestive heart failure, age-related macular degeneration and osteoporosis.

7. A method for treating or preventing a pathology linked to angiogenesis, by inhibiting neovascularization, comprising the step of administering a nucleic acid molecule, which encodes the IRS-1 protein, or a complementary sequence or a fragment thereof and which is capable of inhibiting the expression of a nucleic acid molecule encoding the IRS-1 protein, to a patient.

8. The method according to claim 7, wherein said neovascularization inhibition is obtained by the inhibition of the formation of capillary tubes by endothelial cells.

9. The method according to claim 7 or claim 8, wherein said nucleic acid molecule has a sequence selected from the group consisting of SEQ ID NO.2 to SEQ ID NO.23, or a fragment thereof comprising at least twelve contiguous nucleotides, and derivatives thereof.

10. The method according to any one of claims 7 to 9, wherein said nucleic acid molecule is antisense to at least twelve contiguous nucleotides of SEQ ID NO.28, or a derivative thereof.

11. The method according to any one of claims 7 to 10, wherein said nucleic acid molecule is associated with an acceptable vehicle.

12. The method according to any one of claims 7 to 11,

wherein said nucleic acid molecule is administered via the subcutaneous, intramuscular, intravenous or transdermal route, in an amount from about 0.001 mg to about 50 mg.

- 5 13. The method according to any one of claims 7 to 12,
wherein said pathology linked to angiogenesis is selected
in the group comprising retinopathies, rheumatoid
arthritis, Crohn's disease, atherosclerosis,
hyperstimulation of the ovary, psoriasis, endometritis
10 associated with neovascularization, restenosis due to
balloon angioplasty, tissue superproduction due to
cicatriziation, peripheral vascular disease, hypertension,
vascular inflammation, Raynaud's disease and Raynaud's
phenomena, aneurysm, arterial restenosis,
15 thrombophlebitis, lymphangitis, lymphedema, tissue
cicatriziation and repair, ischemia, angina, myocardial
infarction, chronic heart disease, cardiac insufficiencies
such as congestive heart failure, age-related macular
degeneration and osteoporosis.

- 20 14. A use according to any one of claims 1 to 6,
substantially as herein described with reference to any
one of the examples and/or drawings.

- 25 15. A method according to any one of claims 7 to 13,
substantially as herein described with reference to any
one of the examples and/or drawings.

Dated this 4th day of January 2006

30 GENE SIGNAL

By their Patent Attorneys

GRIFFITH HACK

Fellows Institute of Patent and

Trade Mark Attorneys of Australia

35

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N.S : cellules non stimulées avec le bFGF
S : cellules stimulées avec le bFGF

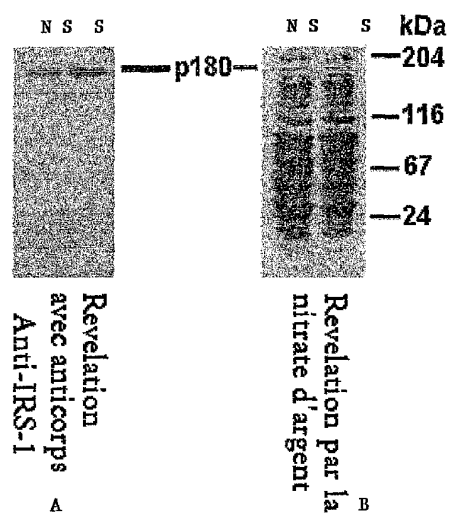
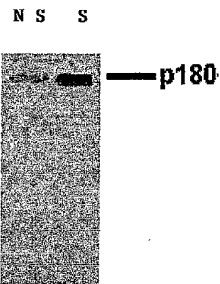


Fig.1

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N S : cellules non stimulées avec le bFGF
S : cellules stimulées avec le bFGF

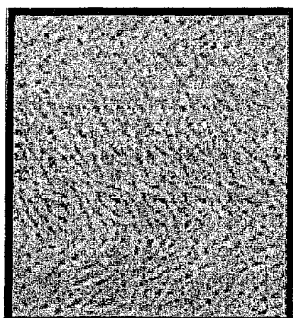


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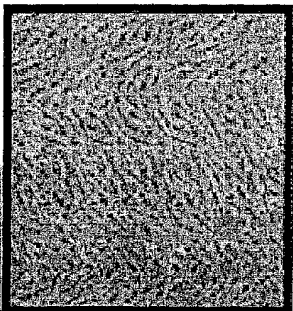
Fig.2

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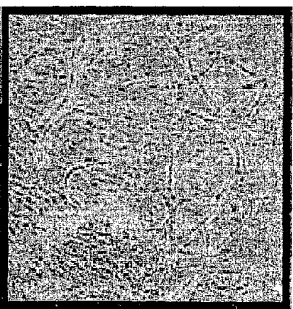
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LOT 3



LOT 2



LOT 1

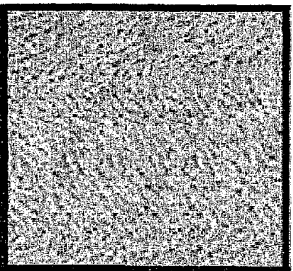


Fig. 3

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Figure 4A
Injections sous-conjonctivales
d'oligonucléotides antisens (60 μ M)

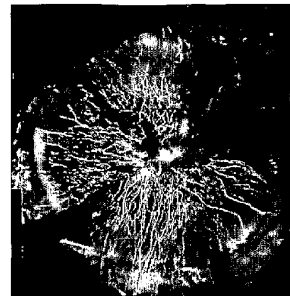


Figure 4B
Injections sous-conjonctivales
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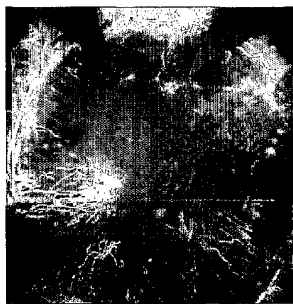


Figure 4C
Applications topiques
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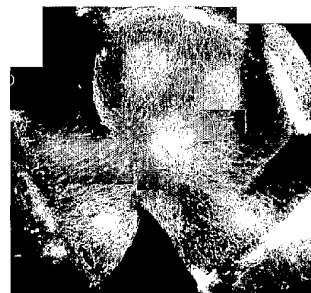


Figure 4D
Applications topiques
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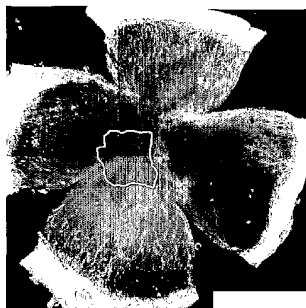


Figure 4E
Absence de traitement



Figure 4F
Injections sous-conjonctivales de PBS

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Figure 5A J4 AS60

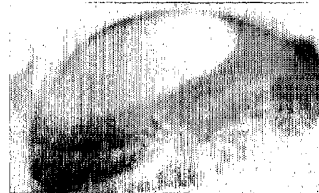


Figure 5B J4 AS200

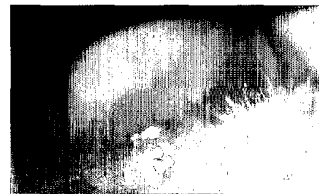


Figure 5C J4 S200

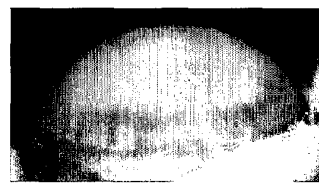


Figure 5D J4 PBS

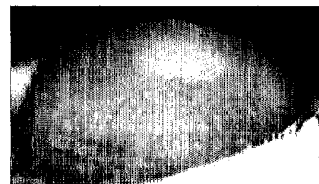


Figure 5E J4 OTt

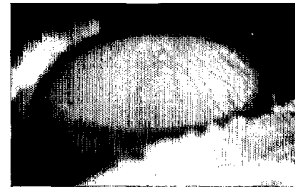


Figure 5F J9 AS60



Figure 5G J9 AS200



Figure 5H J9 S200

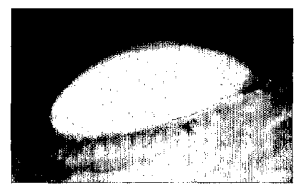


Figure 5I J9 PBS

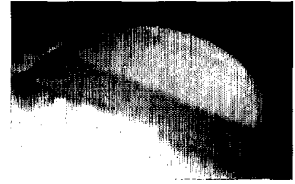


Figure 5J J9 OTt

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Gly Tyr Leu Arg Lys Pro Lys Ser Met His Lys Arg Phe Phe Val Leu
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Arg Ala Ala Ser Glu Ala Gly Gly Pro Ala Arg Leu Glu Tyr Tyr Glu
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Asn Glu Lys Lys Trp Arg His Lys Ser Ser Ala Pro Lys Arg Ser Ile
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Pro Leu Glu Ser Cys Phe Asn Ile Asn Lys Arg Ala Asp Ser Lys Asn
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Lys His Leu Val Ala Leu Tyr Thr Arg Asp Glu His Phe Ala Ile Ala
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gcg gac agc gag gcc gag caa gac agc tgg tac cag gct ctc cta cag 1357
Ala Asp Ser Glu Ala Glu Gln Asp Ser Trp Tyr Gln Ala Leu Leu Gln
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Leu His Asn Arg Ala Lys Gly His His Asp Gly Ala Ala Ala Leu Gly
115 120 125

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aga acc cac gcc cac cgg cat cgg ggc agc gcc cgg ctg cac ccc ccg Arg Thr His Ala His Arg His Arg Gly Ser Ala Arg Leu His Pro Pro 355 360 365	2125
ctc aac cac agc cgc tcc atc ccc atg ccg gct tcc cgc tgc tcg cct Leu Asn His Ser Arg Ser Ile Pro Met Pro Ala Ser Arg Cys Ser Pro 370 375 380 385 390	2173

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Ser Ala Thr Ser Pro Val Ser Leu Ser Ser Ser Ser Thr Ser Gly His			
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Ser Gly Ser Pro Ser Asp Gly Gly Phe Ile Ser Ser Asp Glu Tyr Gly			
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Ser Leu Gly His Thr Pro Pro Ala Arg Gly Glu Glu Glu Leu Ser Asn			
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<212> PRT
<213> Homo sapiens
<400> 29

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Arg Ala Ala Ser Glu Ala Gly Gly Pro Ala Arg Leu Glu Tyr Tyr Glu
          35          40          45

Asn Glu Lys Lys Trp Arg His Lys Ser Ser Ala Pro Lys Arg Ser Ile
          50          55          60

Pro Leu Glu Ser Cys Phe Asn Ile Asn Lys Arg Ala Asp Ser Lys Asn
          65          70          75          80

Lys His Leu Val Ala Leu Tyr Thr Arg Asp Glu His Phe Ala Ile Ala
          85          90          95

Ala Asp Ser Glu Ala Glu Gln Asp Ser Trp Tyr Gln Ala Leu Leu Gln
          100          105          110

Leu His Asn Arg Ala Lys Gly His His Asp Gly Ala Ala Ala Leu Gly
          115          120          125

Ala Gly Gly Gly Gly Gly Ser Cys Ser Gly Ser Ser Gly Leu Gly Glu
          130          135          140

Ala Gly Glu Asp Leu Ser Tyr Gly Asp Val Pro Pro Gly Pro Ala Phe
          145          150          155          160

Lys Glu Val Trp Gln Val Ile Leu Lys Pro Lys Gly Leu Gly Gln Thr
          165          170          175

Lys Asn Leu Ile Gly Ile Tyr Arg Leu Cys Leu Thr Ser Lys Thr Ile
          180          185          190

Ser Phe Val Lys Leu Asn Ser Glu Ala Ala Ala Val Val Leu Gln Leu
          195          200          205

Met Asn Ile Arg Arg Cys Gly His Ser Glu Asn Phe Phe Phe Ile Glu
          210          215          220

Val Gly Arg Ser Ala Val Thr Gly Pro Gly Glu Phe Trp Met Gln Val
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Asp Asp Ser Val Val Ala Gln Asn Met His Glu Thr Ile Leu Glu Ala
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Met Arg Ala Met Ser Asp Glu Phe Arg Pro Arg Ser Lys Ser Gln Ser

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Thr Glu Ser Ile Thr Ala Thr Ser Pro Ala Ser Met Val Gly Gly Lys		
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Pro Gly Ser Phe Arg Val Arg Ala Ser Ser Asp Gly Glu Gly Thr Met		
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Ser Arg Pro Ala Ser Val Asp Gly Ser Pro Val Ser Pro Ser Thr Asn		
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Arg Thr His Ala His Arg His Arg Gly Ser Ala Arg Leu His Pro Pro		
355	360	365
Leu Asn His Ser Arg Ser Ile Pro Met Pro Ala Ser Arg Cys Ser Pro		
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Ser Ala Thr Ser Pro Val Ser Leu Ser Ser Ser Ser Thr Ser Gly His		
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Gly Ser Thr Ser Asp Cys Leu Phe Pro Arg Arg Ser Ser Ala Ser Val		
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Ser Gly Ser Pro Ser Asp Gly Gly Phe Ile Ser Ser Asp Glu Tyr Gly		
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Ser Ser Pro Cys Asp Phe Arg Ser Ser Phe Arg Ser Val Thr Pro Asp		
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Ser Leu Gly His Thr Pro Pro Ala Arg Gly Glu Glu Glu Leu Ser Asn		
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Tyr Ile Cys Met Gly Gly Lys Gly Pro Ser Thr Leu Thr Ala Pro Asn		
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Val Pro Thr Arg Ser Tyr Pro Glu Glu Gly Leu Glu Met His Pro Leu		
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Glu Arg Arg Gly Gly His His Arg Pro Asp Ser Ser Thr Leu His Thr		

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Ser	Ala	Pro	Gln	Gln	Ile	Ile	Asn	Pro	Ile	Arg	Arg	His	Pro	Gln	Arg	
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Val	Asp	Pro	Asn	Gly	Tyr	Met	Met	Met	Ser	Pro	Ser	Gly	Gly	Cys	Ser	
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Pro	Asp	Ile	Gly	Gly	Gly	Pro	Ser	Ser	Ser	Ser	Ser	Ser	Ser	Asn	Ala	
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Val	Pro	Ser	Gly	Thr	Ser	Tyr	Gly	Lys	Leu	Trp	Thr	Asn	Gly	Val	Gly	
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Gly	His	His	Ser	His	Val	Leu	Pro	His	Pro	Lys	Pro	Pro	Val	Glu	Ser	
705				710					715					720		
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Pro	Val	Gly	Asp	Ser	Asn	Thr	Ser	Ser	Pro	Ser	Asp	Cys	Tyr	Tyr	Gly	
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Pro	Glu	Asp	Pro	Gln	His	Lys	Pro	Val	Leu	Ser	Tyr	Tyr	Ser	Leu	Pro	
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Arg	Ser	Phe	Lys	His	Thr	Gln	Arg	Pro	Gly	Glu	Pro	Glu	Glu	Gly	Ala	
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Arg	His	Gln	His	Leu	Arg	Leu	Ser	Thr	Ser	Ser	Gly	Arg	Leu	Leu	Tyr	
785				790					795					800		
Ala	Ala	Thr	Ala	Asp	Asp	Ser	Ser	Ser	Ser	Thr	Ser	Ser	Asp	Ser	Leu	
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Ala Ala Ser Ala Ser Pro Thr Gly Pro Gln Gly Ala Ala Glu Leu 1040	1045	1050
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