



(86) Date de dépôt PCT/PCT Filing Date: 2005/04/28  
(87) Date publication PCT/PCT Publication Date: 2006/08/17  
(45) Date de délivrance/Issue Date: 2009/06/30  
(85) Entrée phase nationale/National Entry: 2007/04/24  
(86) N° demande PCT/PCT Application No.: US 2005/014447  
(87) N° publication PCT/PCT Publication No.: 2006/085894  
(30) Priorités/Priorities: 2005/01/31 (US11/045,081);  
2005/04/04 (US11/097,161)

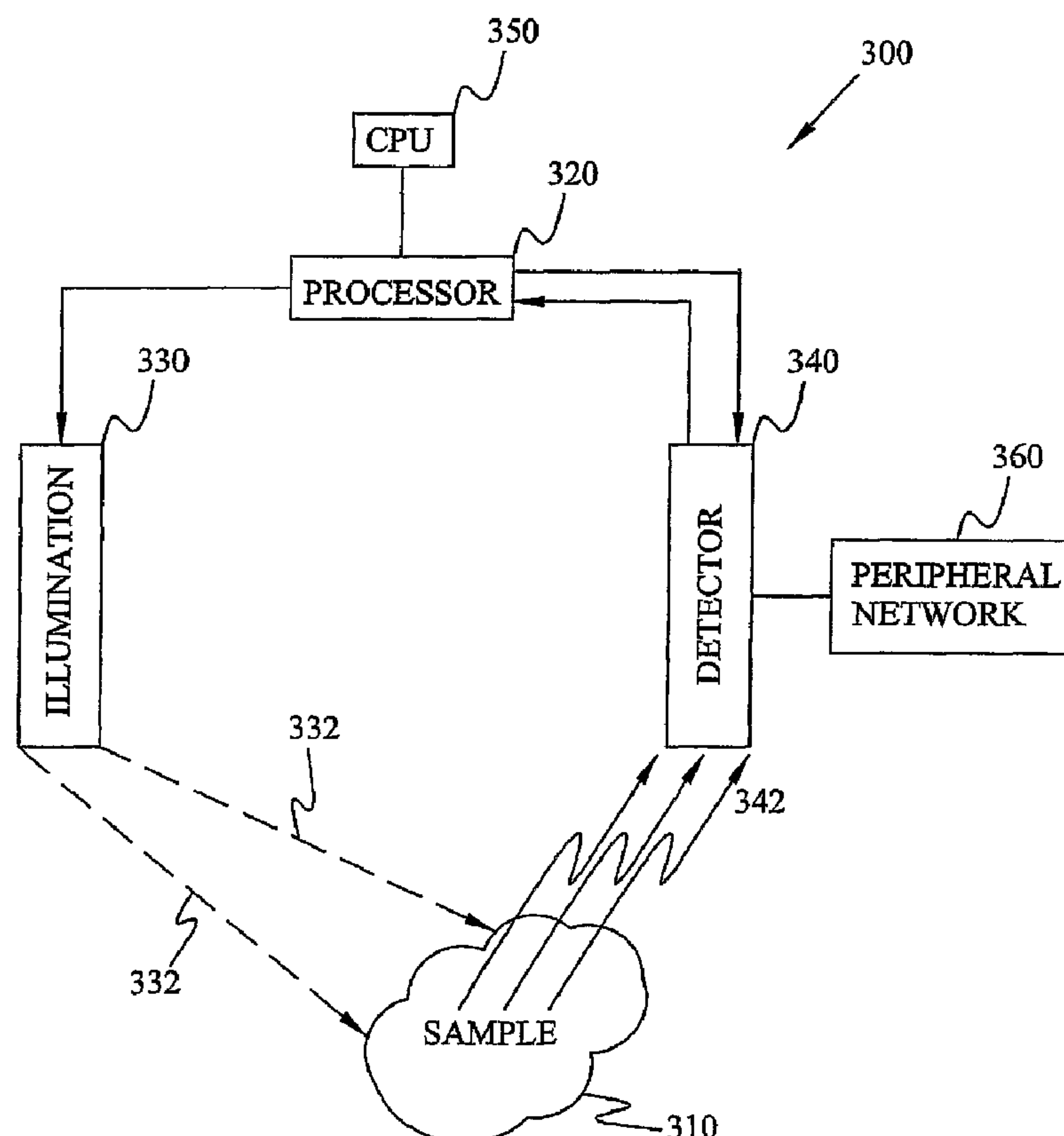
(51) Cl.Int./Int.Cl. *G01N 21/64* (2006.01),  
*G01N 1/30* (2006.01), *G01N 21/25* (2006.01),  
*G01N 21/65* (2006.01), *G01N 33/52* (2006.01)

(72) Inventeurs/Inventors:  
TUSCHEL, DAVID, US;  
MAIER, JOHN, US;  
DEMUTH, JOSEPH E., US

(73) Propriétaire/Owner:  
CHEMIMAGE CORPORATION, US

(74) Agent: DIMOCK STRATTON LLP

(54) Titre : APPAREIL ET PROCEDE D'IMAGERIE CHIMIQUE D'UN ECHANTILLON BIOLOGIQUE  
(54) Title: APPARATUS AND METHOD FOR CHEMICAL IMAGING OF A BIOLOGICAL SAMPLE



(57) Abrégé/Abstract:

In one embodiment, the disclosure relates to a method for determining illumination parameters for a stained sample, the method may include providing a stained sample and obtaining an absorption band of the sample; obtaining an emission band of the sample and determining the illumination parameters for the sample as a function of the absorption band and the emission band of the sample.



## (12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization  
International Bureau



(43) International Publication Date  
17 August 2006 (17.08.2006)

PCT

(10) International Publication Number  
**WO 2006/085894 A1**

(51) International Patent Classification:  
**G01N 15/06** (2006.01)

(74) Agent: **COMTOIS, Mark, C.**; Duane Morris LLP, 1667  
K Street, N.W., Suite 700, Washington, DC 20006 (US).

(21) International Application Number:  
PCT/US2005/014447

(22) International Filing Date: 28 April 2005 (28.04.2005)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:  
11/045,081 31 January 2005 (31.01.2005) US  
11/097,161 4 April 2005 (04.04.2005) US

(71) Applicant (for all designated States except US):  
**CHEMIMAGE CORPORATION** [US/US]; 7301  
Penn Avenue, Pittsburgh, PA 15208 (US).

(72) Inventors; and

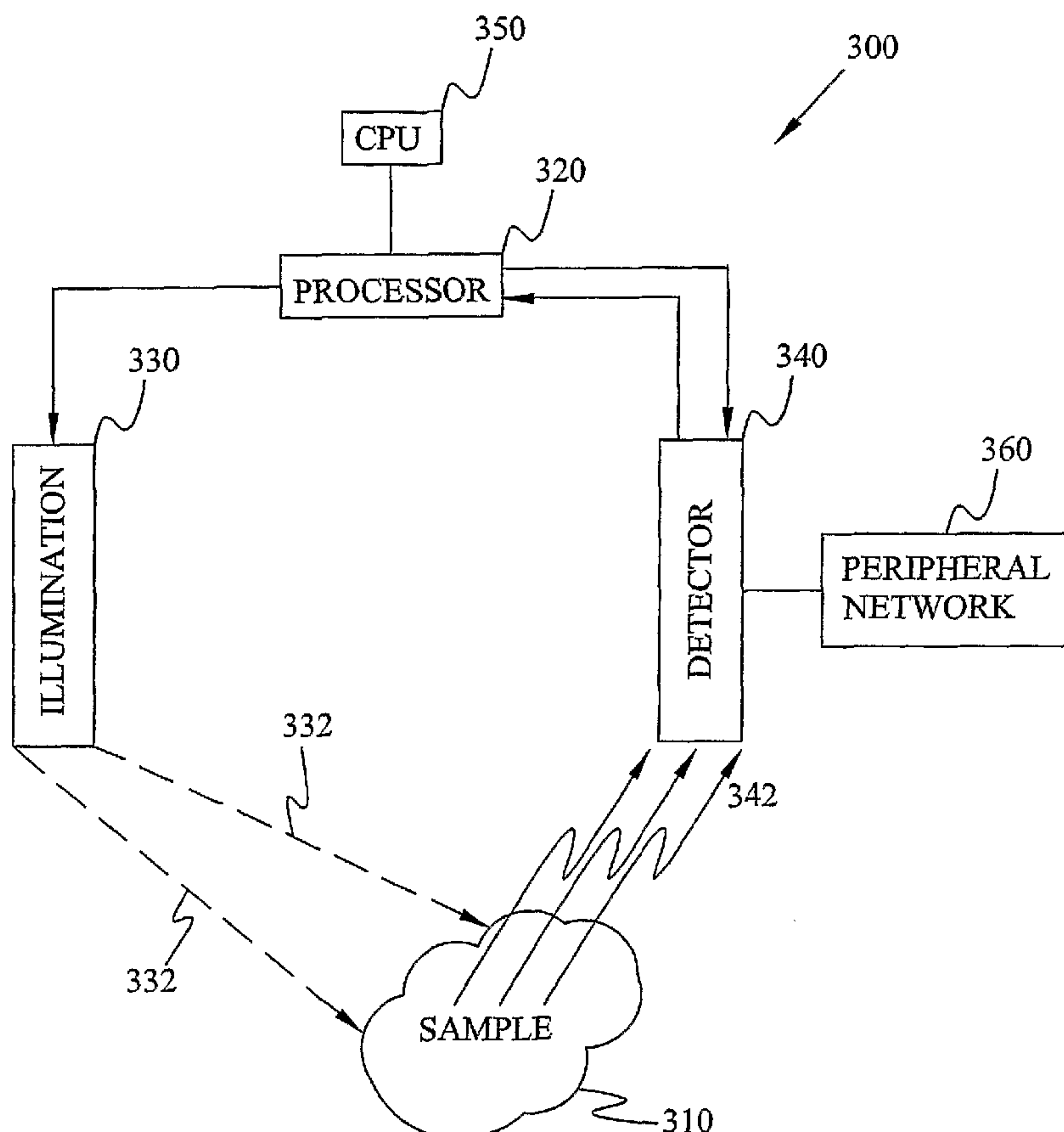
(75) Inventors/Applicants (for US only): **TUSCHEL, David**  
[US/US]; 915 Harvard Road, Monroeville, PA 15146 (US).  
**MAIER, John** [US/US]; 622 S. Lang Avenue, Pittsburgh,  
PA 15208 (US).

(81) Designated States (unless otherwise indicated, for every  
kind of national protection available): AE, AG, AL, AM,  
AT, AU, AZ, BA, BB, BG, BR, BW, BY, BZ, CA, CH, CN,  
CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, EG, ES, FI,  
GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE,  
KG, KM, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA,  
MD, MG, MK, MN, MW, MX, MZ, NA, NI, NO, NZ, OM,  
PG, PH, PL, PT, RO, RU, SC, SD, SE, SG, SK, SL, SM,  
SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN,  
YU, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every  
kind of regional protection available): ARIPO (BW, GH,  
GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM,  
ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM),  
European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI,  
FR, GB, GR, HU, IE, IS, IT, LT, LU, MC, NL, PL, PT, RO,  
SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN,  
GQ, GW, ML, MR, NE, SN, TD, TG).

[Continued on next page]

(54) Title: APPARATUS AND METHOD FOR CHEMICAL IMAGING OF A BIOLOGICAL SAMPLE



(57) Abstract: In one embodiment, the disclosure relates to a method for determining illumination parameters for a stained sample, the method may include providing a stained sample and obtaining an absorption band of the sample; obtaining an emission band of the sample and determining the illumination parameters for the sample as a function of the absorption band and the emission band of the sample.

**WO 2006/085894 A1**



**Published:**

— *with international search report*

*For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.*



## Apparatus and Method for Chemical Imaging of a Biological Sample

### Background

[0002] Spectroscopic imaging combines digital imaging and molecular spectroscopy techniques, which can include Raman scattering, fluorescence, photoluminescence, ultraviolet, visible and infrared absorption spectroscopies. When applied to the chemical analysis of materials, spectroscopic imaging is commonly referred to as chemical imaging. Instruments for performing spectroscopic (*i.e.*, chemical) imaging typically comprise image gathering optics, focal plane array imaging detectors and imaging spectrometers.

[0003] In general, the sample size determines the choice of image gathering optic. For example, a microscope is typically employed for the analysis of sub micron to millimeter spatial dimension samples. For larger objects, in the range of millimeter to meter dimensions, macro lens optics are appropriate. For samples located within relatively inaccessible environments, flexible fiberscopes or rigid borescopes can be employed. For very large scale objects, such as planetary objects, telescopes are appropriate image gathering optics.

[0004] Regardless of the type of optical equipment, a first step in any spectroscopic investigation is defining a suitable wavelength for illuminating the sample. The step of defining a suitable wavelength for illuminating the sample becomes even more important when simultaneous multiple images of the sample are sought.

Conventional methods suggest illuminating a sample with a first wavelength (*e.g.*, NIR or VIS) to obtain a first image, followed by illuminating the sample with a second wavelength to obtain a second image (*e.g.*, Raman or dispersive Raman). Consequently, the conventional process is time consuming and is not suited for simultaneous imaging of the sample. There is a need for an apparatus and method for determining illumination parameters of a sample *a priori* of illuminating the sample.

[0005] The current disclosure addresses the need described above. In one embodiment, the disclosure relates to a method for obtaining a chemical image of a biological sample by providing a biological sample labeled with a Fluorophore; irradiating the sample with photons having wavelength within the illumination wavelength range; obtaining a spectral image of the sample; and generating a chemical image from the spectral image. The chemical image may define at least two spectral images of the sample obtained simultaneously. The spectral images can include a Raman image and a fluorescent image.

[0006] In another embodiment, an apparatus for obtaining a spectral image of a biological sample comprising means for determining a range of illumination wavelengths, the illumination wavelength interacting with the sample to simultaneously provide a first and a second spectra of the sample; a photon source for directing photons with a wavelength within the range to the sample, the illuminating photons interacting with the sample to produce interacted photons; a tunable filter for receiving interacted photons and forming a spectral image of the sample.

[0007] In still another embodiment, the disclosure relates to a system for obtaining multiple spectra of a biological sample. The system can include a processor programmed with instructions to determine illumination parameters of the sample as a function of the emission bandwidth of said sample; an illumination source for directing photons having a wavelength within the illumination parameters of the sample, the illuminating photons

interacting with the sample to provide interacted photons; and a tunable filter for receiving the interacted photons from the sample and providing at least a first and a second spectra of the sample.

Brief Description of the Drawing

[0008] Fig. 1 graphically illustrates the relationship between intensity and wavelength of a sample;

[0009] Figs. 2A-2G each schematically illustrate spectral images of a sample receiving different excitation wavelengths; and

[0010] Fig. 3 is a functional diagram of a system according to one embodiment of the disclosure.



### Detailed Description

[0011] The disclosure generally relates to a method and apparatus for determining illumination parameters for a sample. Having an *a priori* knowledge of an optimal illumination parameters (e.g., optimal illumination wavelength range) for obtaining spectral images of a sample is particularly important in that the optimal illumination parameter enables simultaneous detection of more than one spectra of the sample. The optimal illumination parameters can also be used with different detection modes such as: wide field, Raman chemical imaging, multipoint, dispersive single point and dispersive line.

[0012] Fig. 1 graphically illustrates the relationship between intensity and wavelength of a sample. The method of obtaining absorption and emission bands are conventionally known. It is also known that the emission wavelength, associated with fluorescence imaging, is longer than the absorption wavelength. Thus, as a first step a sample may be illuminated with photons of different wavelengths (interchangeably, detection photons or illumination photons) to determine the sample's absorption and emission wavelengths.

[0013] In Fig. 1, line 125 represents the energy absorption relationship of a sample exposed to illumination photons. Peak 130 indicates the peak wavelength ( $\lambda_{\text{Abs, P}}$ ) for absorption spectrum of the sample; peak 150 indicates the peak wavelength for the emission spectrum ( $\lambda_{\text{m}}$ ) of the sample and the Raman scattering occurs at wavelengths shorter than that at peak 150. A range of wavelength where absorption energy of the sample can be detected is shown to extend from  $\lambda_{\text{abs-L}}$  to  $\lambda_{\text{abs-H}}$ . Similarly, a range of wavelength where emissive energy of the sample can be detect extends from  $\lambda_{\text{Em-L}}$  to  $\lambda_{\text{Em-H}}$ .

[0014] As will be discussed in greater detail, according to one embodiment of the disclosure, an optimal wavelength for multi-spectral imaging can occur at a wavelength

just longer than or about  $\lambda_{\text{abs-L}}$ . Thus, a method is disclosed for defining illumination parameters which includes (i) defining a range of absorption wavelengths for the sample; (ii) defining a range of emission wavelengths for the sample; and (iii) assessing suitable illumination parameters for the sample as a function of the absorption wavelength and the emission wavelength. These steps can be implemented sequentially or simultaneously. By way of example, the region shown as 155 in Fig. 1 shows a possible illumination wavelength such that it is shorter than the wavelength of a peak in the absorption and emission spectra. The illumination parameters may also be used to define an illumination laser line or a suitable Raman illumination wavelength. Since wavelength and frequency are inversely proportional, steps (i)-(iii) can be implemented and defined in view of a frequency band. That is, in view of a range of absorption wavelengths of a sample an equivalent frequency bandwidth for the sample can be defined.

[0015] In another embodiment of the disclosure, a method for determining illumination parameters for a sample includes: simultaneously illuminating the sample with illuminating photons. The illuminating photons can have several different wavelengths or define a broad range of wavelengths. Next, the emissive and absorption wavelengths for the sample can be defined. Alternatively, the sample's bandwidth for emission and absorption can be determined. The emission and the absorption bands can also define the peak intensity wavelength as well as the lower and the upper wavelength ranges for each band. Using the lower wavelength of the absorption band ( $\lambda_{\text{abs-L}}$ ) as a starting point, an optimal Raman wavelength detection wavelength for the sample can be defined as Raman scattered photons having wavelength at or longer than  $\lambda_{\text{abs-L}}$ . By way of example, one such region is shown as region 155 in Fig. 1. The illumination parameters thus obtained can be used to illuminate the sample with illuminating photons of different wavelengths to obtain simultaneous spectral images of the sample. The illuminating photons can be provided by a laser line, wide-field, Raman chemical



imaging, multipoint imaging, dispersive single point and dispersive lines specifically devised to be within the desired wavelength range.

[0016] Figs. 2A-2G each schematically illustrate spectral images of a sample receiving different excitation wavelengths. More specifically, Figs. 2A-2G depict absorption, emission and Raman spectra for a biological sample stained with a dye and a method for determining illumination parameters for the sample in view of absorption and emission spectra of the sample. In one embodiment of the disclosure the dye is a Fluorophore. Suitable Fluorophore stains include an immuno-fluorescent compound, a basophilic compound, an acidophilic compound, neutral stains and naturally occurring luminescent molecules. Once stained, the sample can be irradiated with photons having a wavelength within the illumination wavelength range in order to obtain the spectral images of the sample.

[0017] In Fig. 2A, peak 110 shows the emission peak for the stained sample. As is conventionally known, the emission bandwidth (or its equivalent range of wavelength) is a property of the material. In Fig. 2A, the emission range spans between  $\lambda_A$ -  $\lambda_B$ , with a peak emission wavelength occurring at  $\lambda_p$ . The illumination (excitation) wavelength is arbitrarily set at  $\lambda_{X1}$ . Raman peaks are identified as peaks 160. Raman peaks are shifted from excitation wavelength ( $\lambda_{X1}$ ) by a fixed wavelength which is commensurate with the energy lost due to Raman vibration. Increasing or decreasing the excitation wavelength will have a direct effect on the wavelength where the Raman peaks occur. This is schematically illustrated in Figs. 2A-2G where changing the excitation energy from the wavelength  $\lambda_{X1}$  to  $\lambda_{X7}$  results in shifting the wavelength where Raman peaks 160 occur. Referring again to Fig. 2A, the Raman peaks 160 occur at wavelength  $\lambda_{X1}-1/\psi$ ; where  $\psi$  is the Raman energy loss due to Raman excitation expressed in wavenumbers and can be quantified as

$\psi = (1/2\pi c) (k/\mu)^{1/2}$  ; where k is the chemical bond force constant, c is the speed of light, and  $\mu$  is the reduced mass of the molecular oscillator.

[0018] In Figs. 2B and 2C, peak 110 represents the absorption peak and peaks 160 represent Raman scattering peaks for the sample under study. Peak 130 illustrates the sample's Fluorescence emission spectrum. In Figs. 2B and 2C the excitation wavelength is set to  $\lambda_{x2}$  and  $\lambda_{x3}$ , respectively, such that the Fluorescent spectrum of the sample occurs at a wavelength near the excitation region as shown. As can be seen from Figs. 2B and 2C, the sample's Fluorescence spectrum overlaps with the Raman peaks 160, which as stated, occurs at a fixed wavelength from the excitation wavelength. The overlap makes spectral analysis difficult, if not impossible, as the Raman signals will become overwhelmed by the Fluorescent signals.

[0019] In contrast, the illumination parameter  $\lambda_{x4}$  in Fig. 2D is selected such that Raman peaks 160 occur just below the onset of Fluorescent spectrum 130. Here, each of the Raman 160, Fluorescence emission 130 and absorption 110 spectra are visible within a narrow range of wavelengths and the Raman and fluorescence emission signals can be detected substantially simultaneously with a single detection device.

[0020] In Fig. 2E, the sample has dual Fluorescence peaks 130 and 135. Fluorescence peak 135 defines a lower intensity peak as compared with Fluorescence peak 130. Signals from the Raman peaks 165 may be more easily detectable over Fluorescent signal indicating peak 135. In Fig. 2E, illumination wavelength  $\lambda_{x5}$  is selected such that Raman peaks 160 overlap with the Fluorescence peak 135. However, since the Raman signals have a higher intensity, Raman peaks 160 may be identified from emission spectrum 135.

[0021] In Fig. 2F the excitation wavelength is shifted to  $\lambda_{x6}$  and Raman Peaks 160 are no longer eclipsed by Fluorescent peaks 130 and 135. The Raman signals may



nonetheless be attenuated due to the absorption spectrum of peak 110. Since at least a portion of Raman peaks 160 falls between Fluorescent peak 135 and absorption peak 110, the Raman signal may be at least partially distinguishable. In Fig. 2G, excitation wavelength  $\lambda_{x7}$  is shifted to a lower wavelength resulting in the shifting of Raman peaks 160. Here, absorption peaks 110 overlap Raman peaks 160 thereby possibly attenuating the Raman signals, but not necessarily preventing detection of Raman scattering.

[0022] As can be seen from Figs. 2A – 2G, the excitation wavelength can be selected such that at least two spectra of the sample are simultaneously visible within a narrow range of wavelengths. According to one embodiment of the disclosure the excitation wavelength is selected such that Raman peaks appear at a wavelength substantially free from interference from other signals. According to another embodiment, the excitation wavelength can be selected such that signals from Raman peaks are distinguishable from signals depicting emission or Fluorescence spectra. According to still another embodiment of the disclosure, the excitation wavelength is selected such that an imaging device can simultaneously capture the Fluorescence as well as the Raman spectra for the sample. In still another embodiment of the disclosure, the excitation wavelength is selected such that an imaging device can simultaneously capture the Fluorescence emission as well as the Raman spectra for the sample.

[0023] In one embodiment of the disclosure, an apparatus is provided to assess the illumination parameter for a histologically labeled sample. The sample can be labeled with a conventional identifier, such as a Fluorophore substance. Next, an illumination parameter defining a suitable illumination wavelength can be selected such that both the emission peak and Raman scattering peaks can be detected with one imaging apparatus. The imaging apparatus may include gathering optics (*e.g.*, optical trains for collecting photons emitted, Raman scattered, transmitted, or reflected from the sample), one or more tunable filters (*e.g.*, liquid tunable crystal filter(LCTF), Acousto-optical Filter (AOTF) or fiber array spectral translator). A charged-coupled device or other suitable



camera or recording medium may be coupled to the imaging apparatus in order to capture the spectra.

[0024] In a system according to an embodiment of the disclosure, the illumination parameter for a sample includes one or more illumination sources, an optical train and a processor programmed with instructions to simultaneously illuminate the sample with illuminating photons and detect an emission band of the sample. The instructions can also include defining a lower wavelength range and an upper wavelength range for the band and determine the illumination parameters for the sample as a function of the absorption and the emission bands of the sample. Finally, the instructions may include defining a suitable Raman wavelength for the sample at a wavelength shorter than the lower wavelength range ( $\lambda_{EM, L}$ ) of the emission spectrum.

[0025] Fig. 3 is a functional diagram of a system according to one embodiment of the disclosure. In Fig. 3, system 300 is devised to obtain and analyze chemical images of sample 310. Sample 310 can be any biological, organic or inorganic sample suitable for histological studies. Illumination source 330 is positioned to provide excitation photons 332 to sample 310. Illumination source 330 can be positioned above, near or below the sample. Excitation photons 332 define excitation wavelengths which can cover a broad range of spectra (e.g.,  $\lambda_{X1} - \lambda_{X7}$ ). Moreover, illumination source 330 can be adapted to change excitation wavelengths based on instructions communicated by Processor 320. Detector 340 is positioned near sample 340 to receive interacted photons 342. Interacted photons 342 may include Fluorescence, reflection, transmission, emission and Raman photons. Interacted photons can be received and collected by Detector 340 which may include electro-optical devices suitable for gathering and analyzing photons of broad wavelengths. Detector 340 may include, for example, an optical train for gathering and focusing interacted photons; one or more optical filters for rejecting photons of undesired wavelengths; an LCTF for obtaining spectral images of Sample 310; and a charge-coupled device for devising a chemical image based on the spectral images of Sample

310. Detector 340 can communicate with peripheral network devices 360 such as printers, video recorders or internet communication systems.

[0026] Processor 320 can receive spectral images of the sample from Detector 340 and determine whether the illumination wavelength should be changed. For example, if Detector 340 devises a spectral image of a sample similar to Fig. 2C, processor 320 can determine that Raman signals from peaks 160 are indistinguishable from Fluorescent signal 130. Based on this determination, processor 320 can direct a change in excitation wavelength produced by illumination source 330 such that signal interference is reduced to that shown in Fig. 2D. Processor 350 can also communicate with CPU 350 to receive executable instructions or to access a database of related information such as the chemical identifier or the properties of the Fluorophore used on sample 310. CPU 350 can be used to communicate additional information to the processor.

[0027] After one or more iterations, Processor 320 can determine the optimal illumination parameter for the sample such that more than one spectral image can be detected simultaneously. The spectral image of the sample can be used to define one or more chemical images of the sample and to identify the sample under study. Such analysis can be implemented using a single detection and identification system and lends substantial efficiency to histological analysis.

[0028] While the principles of the disclosure have been disclosed in relation to specific exemplary embodiments, it is noted that the principles of the invention are not limited thereto and include all modification and variation to the specific embodiments disclosed herein.



**WHAT IS CLAIMED IS:**

1. A method for obtaining a chemical image of a biological sample comprising: providing a biological sample labeled with a Fluorophore; irradiating the sample with photons having wavelength within an illumination wavelength range; obtaining a spectral image of the sample; and generating a chemical image from the spectral image; wherein the chemical image defines at least two spectral images of the sample obtained simultaneously.
2. The method of claim 1, further comprising determining an illumination parameter for the labeled sample, the illumination wavelength range providing simultaneous spectral images of the sample.
3. The method of claim 2, wherein the step of determining an illumination wavelength further comprises defining a fluorescence emission peak for the labeled sample and selecting a wavelength that is less than the emission peak.
4. The method of claim 1, wherein the chemical image of the sample includes at least one of a fluorescent signal and a Raman signal.
5. The method of claim 1, wherein the chemical image of the spectral image of the sample is selected from the group consisting of Fluorescence, Reflection, Absorption, Transmission, Optical and Raman images.
6. The method of claim 1, wherein the Fluorophore further comprises an immuno-fluorescent compound, a basophilic compound, an acidophilic compound, a neutral stains and any naturally occurring luminescent molecules.
7. The method of claim 6, wherein the acidophilic stain is Eosin.
8. The method of claim 6, wherein the basophilic stain is Hematoxylin.
9. The method of claim 1, wherein the illumination wavelength overlaps with a region of an absorption bandwidth.



10. An apparatus for obtaining a spectral image of a biological sample comprising: means for determining a range of illumination wavelengths, the illumination wavelength interacting with the sample to simultaneously provide a first and a second spectra of the sample; a photon source for directing photons with a wavelength within the range to the sample, the illuminating photons interacting with the sample to produce interacted photons; a tunable filter for receiving interacted photons and forming a spectral image of the sample.
11. The apparatus of claim 10, wherein the first and the second spectra of the sample are formed from the same interacted photons.
12. The apparatus of claim 10, wherein the first spectrum is a Raman spectrum of the sample.
13. The apparatus of claim 10, wherein the second spectrum is a Fluorescent spectrum of the sample.
14. The apparatus of claim 10, the apparatus further comprising a rejection filter for filtering the photons from the photon source.
15. The apparatus of claim 10, the apparatus further comprising a rejection filter for filtering interacted photons.
16. The apparatus of claim 10, the apparatus further comprising an objective lens for collecting interacted photons and directing the interacted photons to the tunable filter.
17. The apparatus of claim 10, wherein the tunable filter is selected from the group consisting of LCTF, AOTF and fiber array spectral translator.
18. The apparatus of claim 10, wherein the spectral image of the sample includes at least one of a fluorescent signal and a Raman signal.

19. The apparatus of claim 10, wherein the sample further comprises a fluorophore substance.
20. The apparatus of claim 10, wherein the means for determining an illumination wavelength further comprises means for defining a fluorescence emission peak for selecting the sample and means for selecting a range of wavelengths shorter than that of the emission peak.
21. The apparatus of claim 10, wherein the means for determining an illumination wavelength further comprises means for defining an absorption bandwidth for the sample and means for selecting a range of wavelengths overlapping the absorption bandwidth.
22. The apparatus of claim 10, wherein the illumination wavelength is a range of wavelengths overlapping a region of an absorption bandwidth.
23. A system for obtaining multiple spectra of a biological sample comprising: a processor programmed with instructions to determine illumination parameters of the sample as a function of an emission bandwidth of said sample; an illumination source for directing photons having a wavelength within the illumination parameters of the sample, the illuminating photons interacting with the sample to provide interacted photons; and a tunable filter for receiving the interacted photons from the sample and providing at least a first and a second spectra of the sample.
24. The system of claim 23, wherein the first and the second spectra are detected from the sample substantially simultaneously.
25. The system of claim 23, wherein the first spectrum is a fluorescence spectrum of the sample.
26. The system of claim 23, wherein the first spectrum is an absorption spectrum of the sample.

27. The system of claim 23, wherein the second spectrum is a Raman spectrum of the sample.
28. The system of claim 23, wherein the illumination source further comprises a rejection filter.
29. The system of claim 23, the system further comprising an optical lens for collecting and directing the interacted photons to the tunable filter.
30. The system of claim 23, wherein the tunable filter is selected from the group consisting of LCTF and AOTF.
31. The system of claim 23, wherein the illumination parameters define a waveband where at least two spectra of the sample are inclusively detectable.



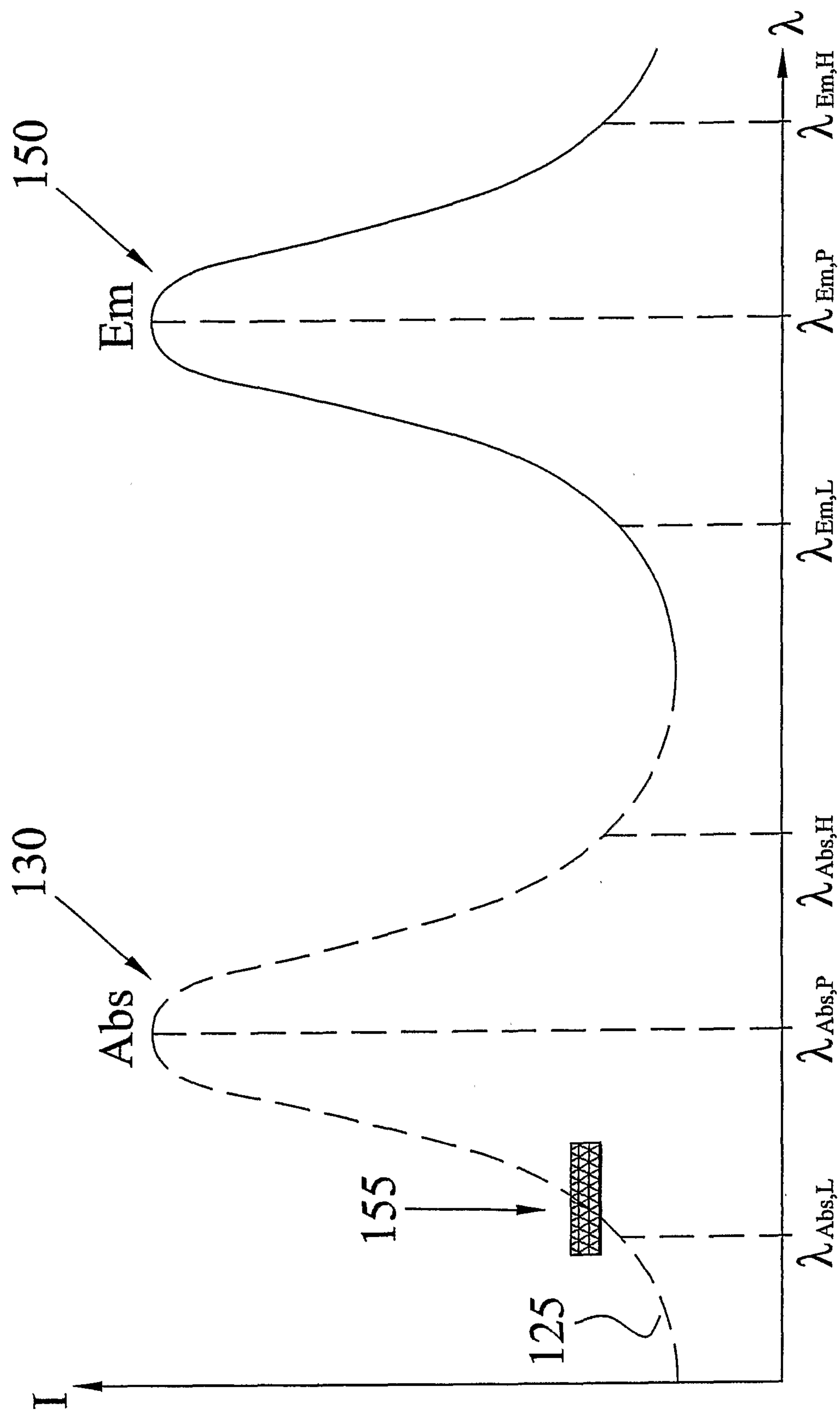


FIGURE 1

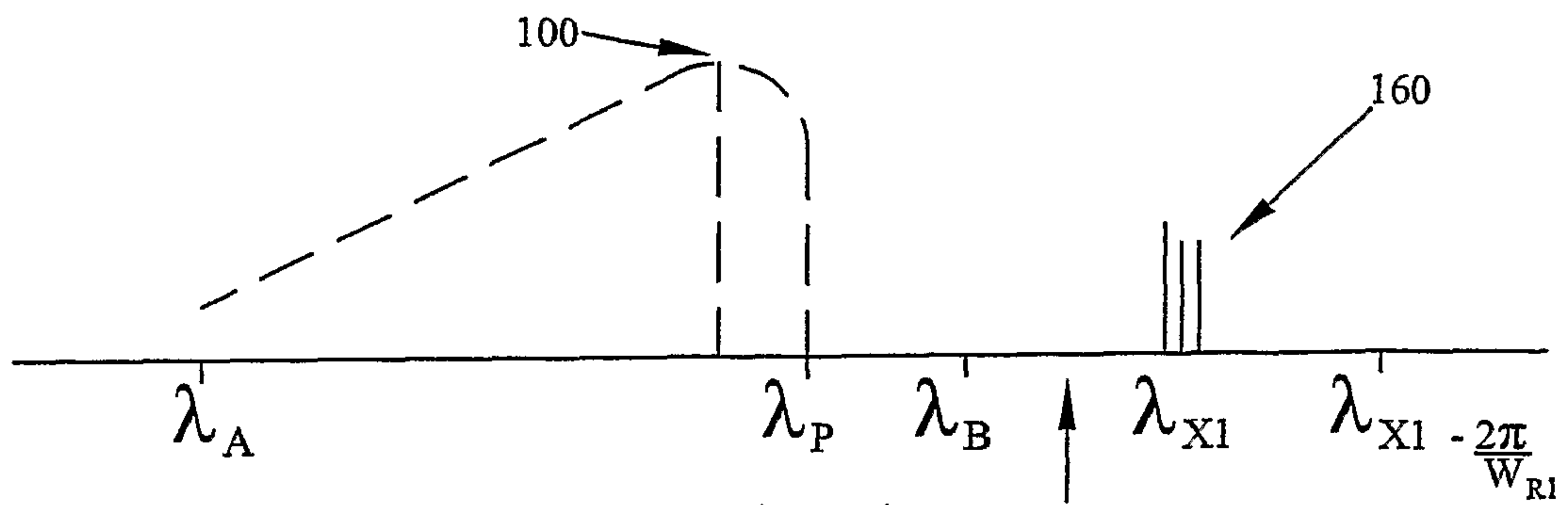


FIG. 2A

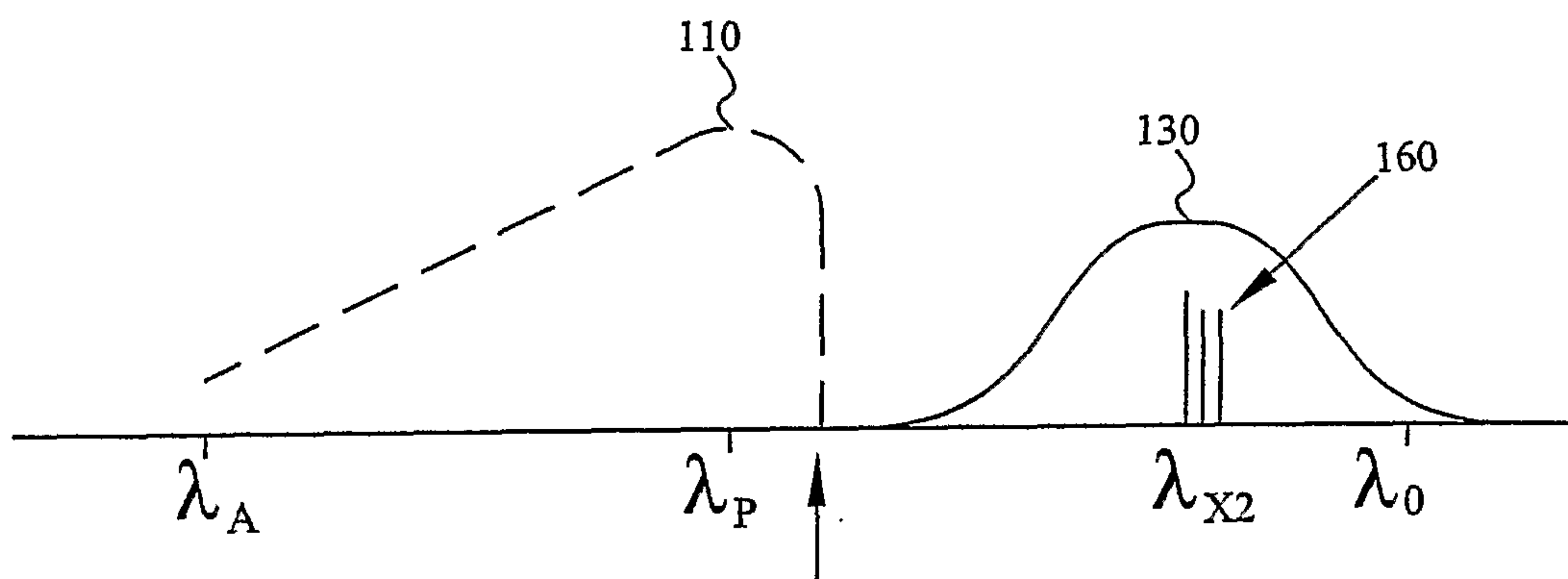


FIG. 2B

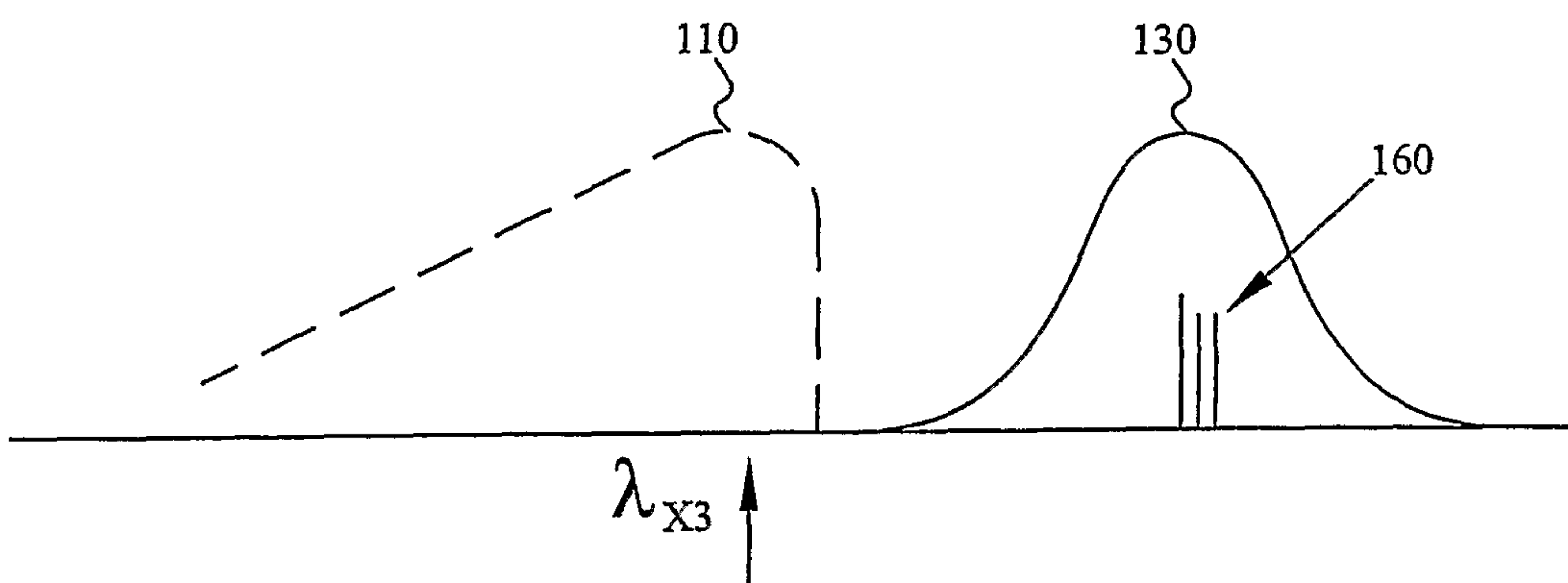


FIG. 2C

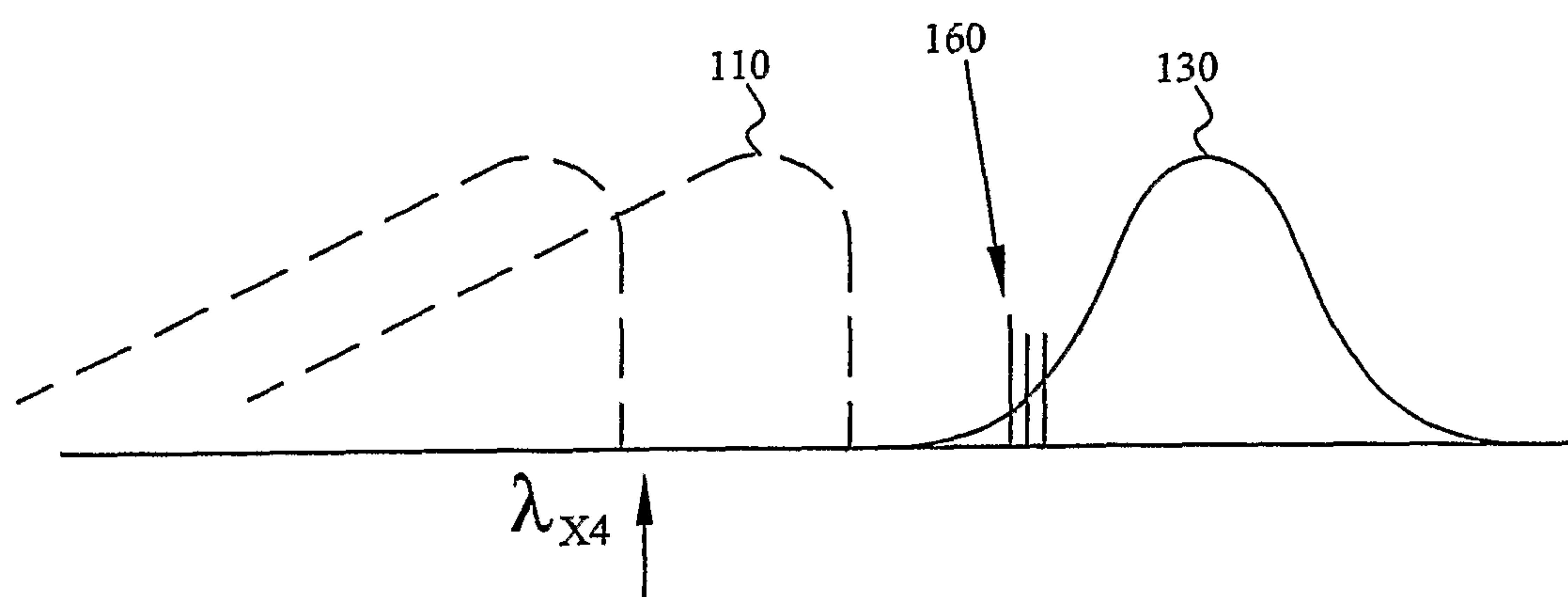


FIG. 2D

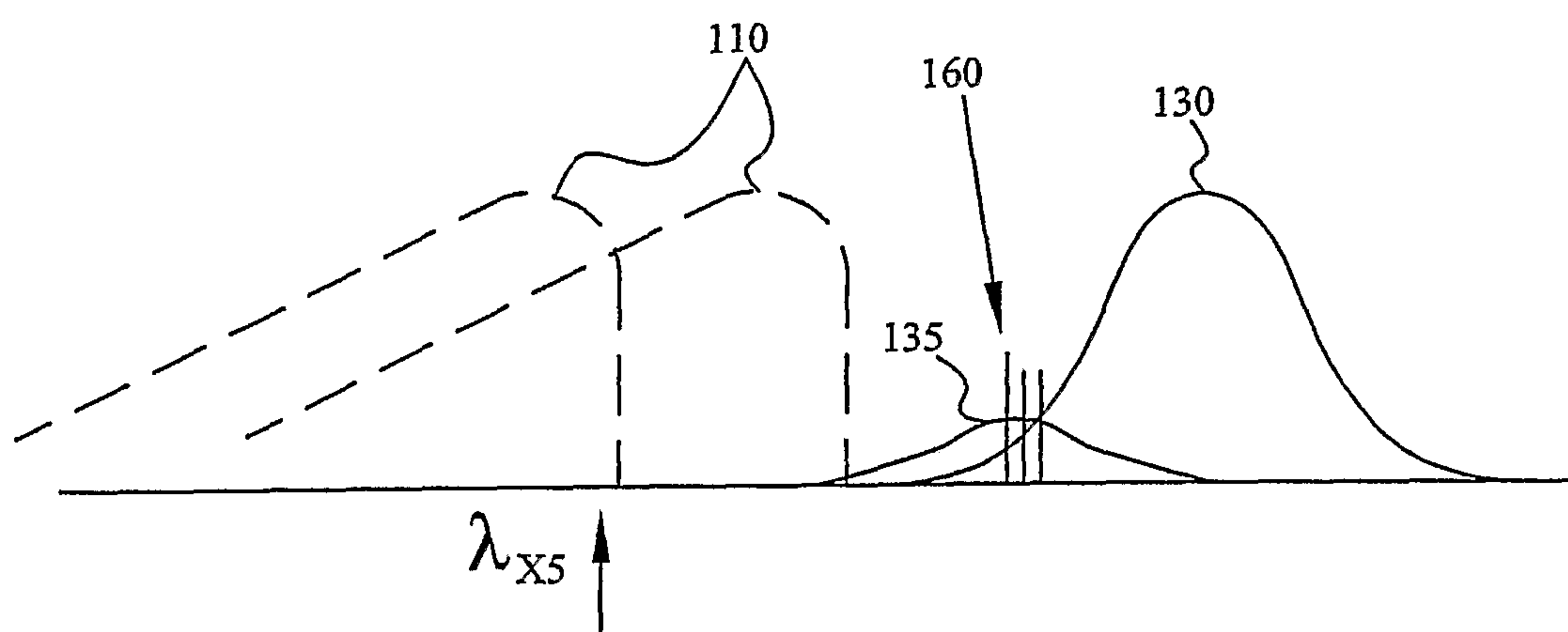


FIG. 2E



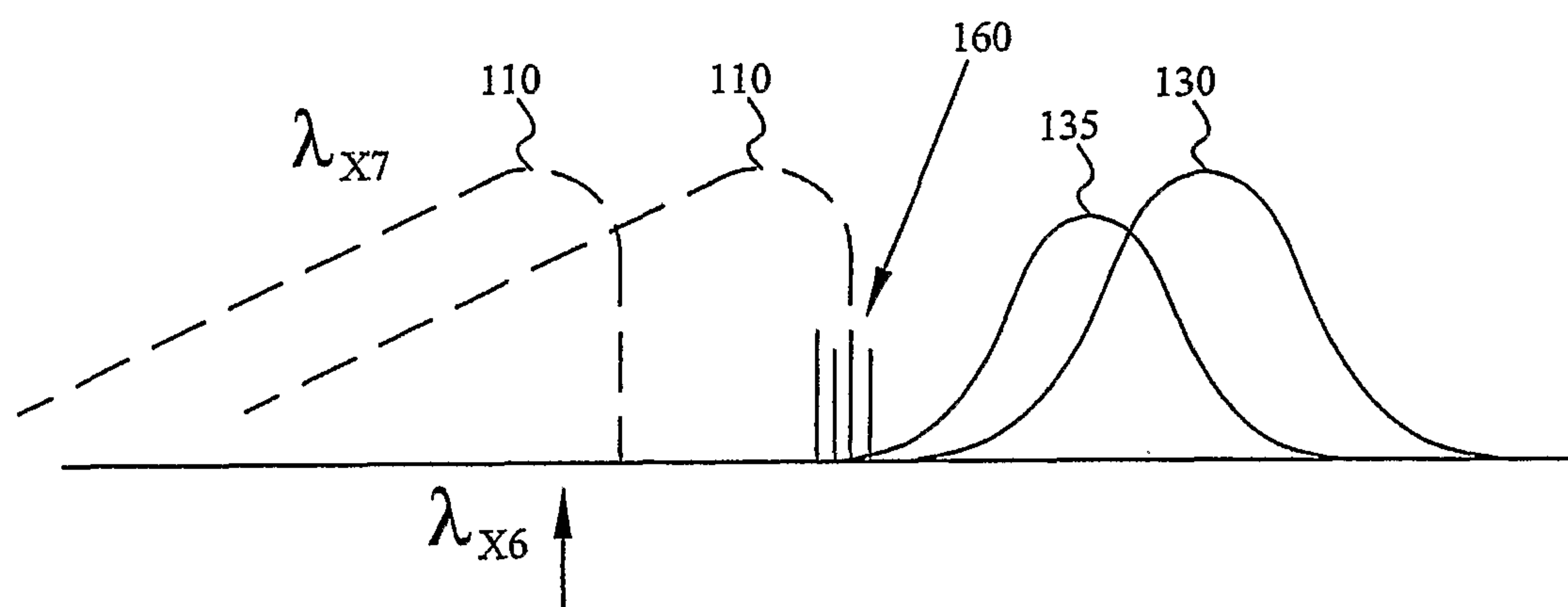


FIG. 2F

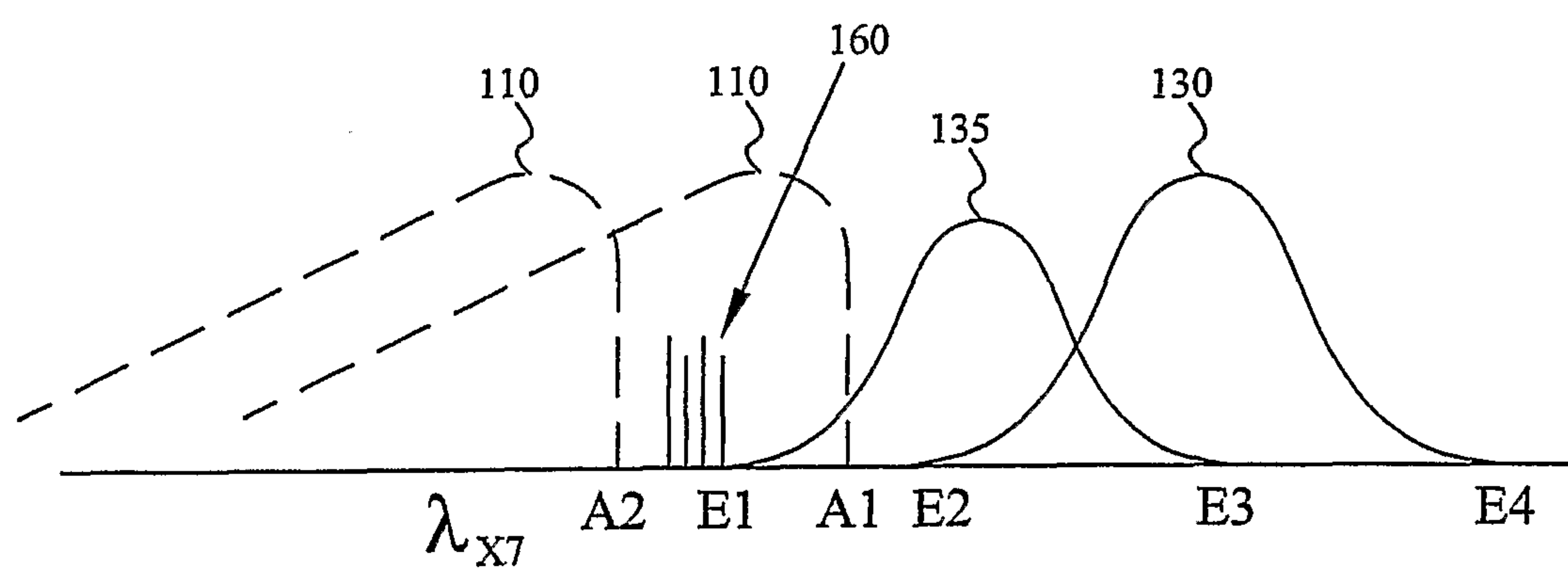


FIG. 2G

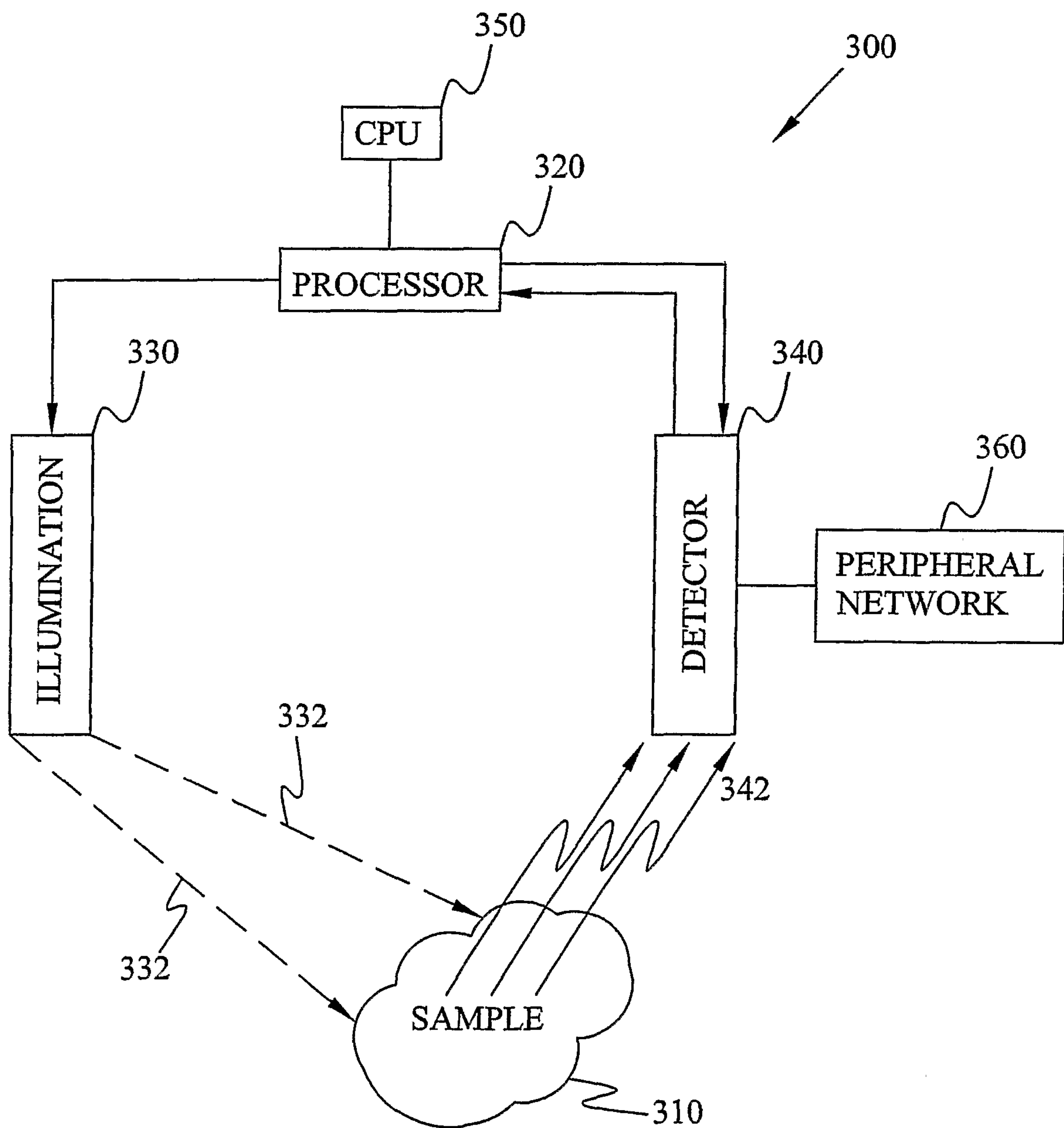


FIG.3



