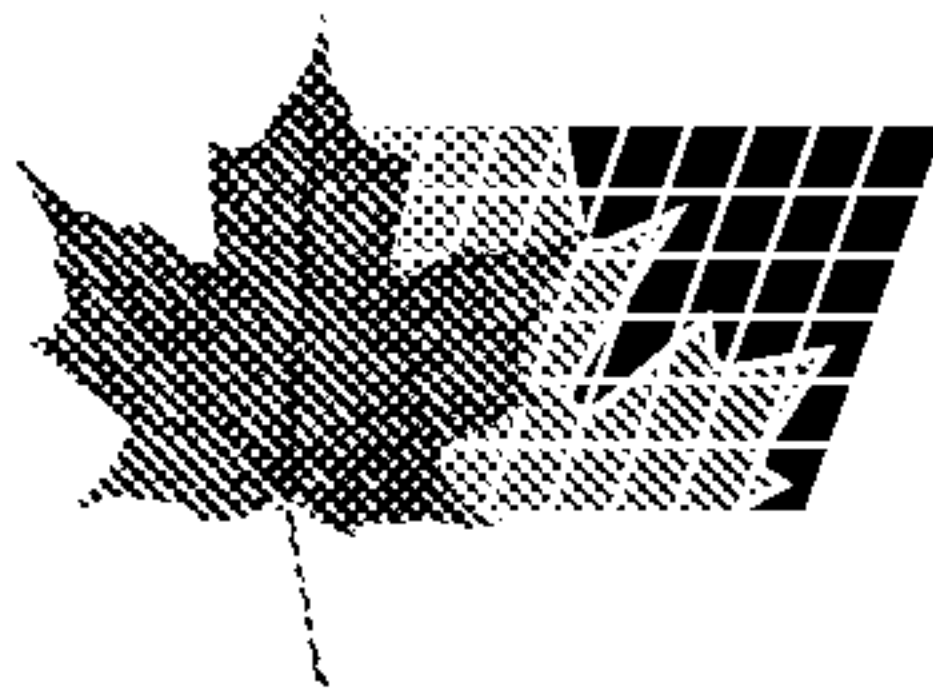




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(54) **SEQUENCES CARACTERISTIQUES DE LA TRANSCRIPTION
GENIQUE REGULEE PAR L'HYPOXEMIE**

(54) **SEQUENCES CHARACTERISTIC OF HYPOXIA-REGULATED
GENE TRANSCRIPTION**

(57) There are provided polynucleotide sequences for use in identifying genes which are modulated by hypoxic conditions.

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SEQUENCES CHARACTERISTIC OF
HYPOXIA-REGULATED GENE TRANSCRIPTION

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BACKGROUND OF THE INVENTION

TECHNICAL FIELD

The present invention relates to the identification of genes that are differentially expressed in hypoxia and use of the genes and gene products for diagnosis and therapeutic intervention. The invention further relates to identification of polynucleotide sequences that are differentially expressed in hypoxia and the use of the sequences for diagnosis and probes.

15

BACKGROUND ART

The level of tissue oxygenation plays an important role in normal development as well as in pathologic processes such as ischemia. Tissue oxygenation plays a significant regulatory/inducer role in both apoptosis and in angiogenesis (Bouck et al, 1996; Bunn et al, 1996; Dor et al, 1997; Carmeliet et al, 1998). Apoptosis (see Duke et al, 1996 for review) and growth arrest occur when cell growth and viability are reduced due to oxygen deprivation (hypoxia). Angiogenesis (i.e. blood vessel growth, vascularization) is stimulated when hypo-oxygenated cells secrete factors which stimulate proliferation and migration of endothelial cells in an attempt to restore oxygen homeostasis (for review see Hanahan et al, 1996).

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Hypoxia plays a critical role in the selection of mutations that contribute to more severe tumorigenic phenotypes (Graeber et al., 1996). Identifying activated or inactivated genes and gene products in hypoxia and ischemia is needed.

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Ischemic disease pathologies involve a decrease in the blood supply to a bodily organ, tissue or body part generally caused by constriction or obstruction of the blood vessels, as for example retinopathy, myocardial infarction and stroke. Therefore, apoptosis and/or angiogenesis as induced by the ischemic condition are also involved in these disease states. Neoangiogenesis is seen in some forms of retinopathy and in tumor growth. These processes are complex cascades of events controlled by many different genes reacting to the various stresses such as hypoxia.

The ability to monitor hypoxia-triggered activation of genes can provide a tool to identify not immediately evident ischemia in a patient. Identification of hypoxia-regulated genes permits the utilization of gene therapy or direct use of gene products, or alternatively inactivation of target genes for therapeutic intervention in treating the diseases and pathologies associated with hypoxia, ischemia and tumor growth.

SUMMARY OF THE INVENTION

The present invention provides purified, isolated and cloned polynucleotides (nucleic acid sequences) associated with hypoxia-regulated activity and having sequences designated as any one of SEQ ID NOS. 1-9, or having complementary or allelic variation sequences thereto.

The present invention provides a method of regulating angiogenesis or apoptosis in a patient in need of such treatment by administering to such patient a therapeutically effective amount of an antagonist of at least one protein as encoded by the nucleic acid sequences as set forth in any of SEQ ID NOS. 1-9.

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Also provided is the diagnostic tool for identifying genes modulated by hypoxic conditions having a detector for detecting the presence of a polynucleotide having a nucleic acid sequence according to any of SEQ. ID NOS. 1-9. A pharmaceutical composition for modulating hypoxia and ischemia having an effective amount of a polynucleotide having the nucleic acid sequence according to SEQ. ID NOS. 1-9 and a pharmaceutically acceptable carrier is also provided.

Also provided is a method of regulating hypoxia-associated pathologies by administering an effective amount of at least one antisense oligonucleotide against one of the nucleic acid sequences (SEQ. ID NOS. 1-9) or their proteins. There is provided a method of regulating hypoxia associated pathology by administering an effective amount of a protein encoded by the polynucleotides (SEQ. ID NOS. 1-9) as active ingredients in the pharmaceutically acceptable carrier.

Further, there are provided hypoxia response regulating genes.

BRIEF DESCRIPTION OF THE FIGURES

Other advantages of the present invention will be readily appreciated as the same becomes better understood by reference to the following detailed description when considered in connection with the accompanying drawings wherein:

Figure 1 shows the nucleic acid sequence of the 92 gene (Seq. I.D. No. 1);

Figure 2 shows the nucleic acid sequence of the 95 gene (Seq. I.D. No. 2);

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Figure 3 shows the nucleic acid sequence of the 98 gene (Seq. I.D. No. 3);

5 Figure 4 shows the nucleic acid sequence of the 60F6 gene (Seq. I.D. No. 4);

Figures 5 A-C show the nucleic acid sequence of the 648 gene (Seq. I.D. No. 5);
10

Figure 6 shows the nucleic acid sequence of the 24D4 gene (Seq. I.D. No. 6);

Figures 7 A and B show the nucleic acid sequence of
15 the 77H4 gene (Seq. I.D. No. 7);

Figure 8 shows the nucleic acid sequence of the 14G2 gene (Seq. I.D. No. 8); and

20 Figure 9 shows the nucleic acid sequence of the 29F3 gene (Seq. I.D. No. 9).

DESCRIPTION OF THE INVENTION

25 The present invention identifies polynucleotides (nucleic acid sequences) with sequences as set forth herein in SEQ. ID Nos. 1-9, that can be utilized diagnostically in hypoxia and ischemia and that can be used as targets for therapeutic intervention, or can be
30 used to identify genes that are regulated and respond to hypoxic conditions. SEQ ID Nos. 1-4 and 6-8 have not previously been identified. SEQ ID No. 5 was found to match sequences in data banks but has not been reported to be associated with hypoxia regulation.

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The present invention further provides candidate genes and gene products that can be utilized therapeutically and diagnostically in hypoxia and ischemia and that can regulate apoptosis or angiogenesis.

5 By regulate or modulate or control, it is meant that the process is either induced or inhibited to the degree necessary to effect a change in the process and the associated disease state in the patient. Whether induction or inhibition is being contemplated will be apparent from
10 the process and disease being treated and will be known to those skilled in the medical arts.

The present invention identifies genes for gene therapy, diagnostics and therapeutics that have direct causal relationships between a disease and its related
15 pathologies and up- or down-regulator (responder) genes. That is, the present invention is initiated by a physiological relationship between cause and effect.

The present invention also provides a method of regulating angiogenesis or apoptosis in a patient in need
20 of such treatment by administering to such patient a therapeutically effective amount of an antagonist of at least one protein as encoded by the nucleic acid sequences as set forth in any of SEQ ID NOS. 1-9.

As shown below, some sequences are partial gene
25 sequences which are markers/probes for genes that are upregulated under hypoxic conditions. These partial sequences can be designated "Expressed Sequence Tags" (ESTs) and are markers for the genes actually expressed in vivo and are ascertained as described herein in the
30 Examples or as is known in the art.

Generally, ESTs comprise DNA sequences corresponding to a portion of nuclear encoded mRNA. The EST has a length that allows for PCR (polymerase chain reaction), for use as a hybridization probe and is a

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unique designation for the gene with which it hybridizes (generally under conditions sufficiently stringent to require at least 95% base pairing). For a detailed description and review of ESTs and their functional
5 utility see, WO 93/00353 PCT Application which is incorporated herein in its entirety by reference, as well as the references by Zweiger et al, 1997; Okubo et al, 1997 and Braren et al, 1997.

The WO 93/00353 PCT application further
10 describes how the EST sequences can be used to identify the transcribed genes.

The present invention also provides a method of diagnosing the presence of ischemia or other hypoxia-associated pathologies in a patient including the steps of
15 analyzing a tissue sample from the patient for the presence of at least one expressed gene (up-regulated) identified by the sequences of the present invention utilized as probes. Methods of identification of hybridization can include immunohistochemical staining of
20 the tissue samples. Further, for identification of the gene, *in situ* hybridization, Southern blotting, single strand conformational polymorphism, restriction endonuclease fingerprinting (REF), PCR amplification and DNA-chip analysis using the nucleic acid sequences of the
25 present invention as probes/primers can be used.

Further, according to the present invention, purified, isolated and cloned hypoxia-responding genes identified by the probes/sequences hybridizing under stringent conditions with 95% homology set forth herein or
30 a complementary or allelic variation sequence and human homologies as needed thereto are disclosed. The present invention further provides proteins as encoded by the identified genes. The present invention further provides antibodies directed against these proteins. The present

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invention further provides transgenic animals and cell lines carrying at least one expressible gene identified by the present invention. The present invention further provides knock-out eucaryotic organisms in which at least
5 one nucleic acid sequences as identified by the probes of the present invention.

The present invention provides a method of regulating angiogenesis, apoptosis or other hypoxia-associated pathologies in a patient in need of such
10 treatment by administering to a patient a therapeutically effective amount of an antagonist of at least one protein as encoded by the nucleic acid sequences or sequences identified herein or by the probes of the present invention, or alternatively by a non-protein product of
15 the gene's activity, or inactivation of a gene by chemical compound. Alternatively, the present invention provides a method of regulating angiogenesis, apoptosis or other hypoxia-associated pathologies in a patient in need of such treatment by administering to a patient a
20 therapeutically effective amount of at least one antisense oligonucleotide against the nucleic acid sequences or dominant negative peptide directed against the sequences or their proteins.

The present invention further provides a method
25 of regulating angiogenesis or apoptosis in a patient in need of such treatment by administering to a patient a therapeutically effective amount of a protein encoded by the identified genes as active ingredients in a pharmaceutically acceptable carrier.

30 The present invention provides a method of providing an apoptosis-regulating gene, an angiogenesis-regulating gene or hypoxia-response regulating gene identified by the probes of the present invention, by administering directly to a patient in need of such
35 therapy an expressible vector comprising expression

control sequences operably linked to one of the identified genes and its human homolog if appropriate.

The proteins of the present invention can be produced recombinantly (see generally Marshak et al, 1996
5 "Strategies for Protein Purification and Characterization. A laboratory course manual." CSHL Press) and analogues can be due to post-translational processing. The term Analogue as used herein is defined as a nucleic acid sequence or protein which has some differences in their
10 amino acid/nucleotide sequences as compared to the native sequence of SEQ. ID NOS. 1-9. Ordinarily, the analogue will be generally at least 70% homologous over any portion that is functionally relevant. In more preferred embodiments the homology will be at least 80% and can
15 approach 95% homology to the protein/nucleotide sequence.

The amino acid or nucleotide sequence of an analogue can differ from that of the primary sequence when at least one residue is deleted, inserted or substituted, but the protein or nucleic acid molecule remains
20 functional. Differences in glycosylation can provide protein analogues.

Functionally relevant refers to the biological property of the molecule and in this context means an *in vivo* effector or antigenic function or activity that is
25 directly or indirectly performed by a naturally occurring protein or nucleic acid molecule. Effector functions include but are not limited to include receptor binding, any enzymatic activity or enzyme modulatory activity, any carrier binding activity, any hormonal activity, any
30 activity in promoting or inhibiting adhesion of cells to extracellular matrix or cell surface molecules, or any structural role as well as having the nucleic acid sequence encode functional protein and be expressible. The antigenic functions essentially mean the possession of an

epitope or antigenic site that is capable of cross-reacting with antibodies raised against a naturally occurring protein. Biologically active analogues share an effector function of the native which can, but need not, in addition possess an antigenic function.

The antibodies can be either monoclonal, polyclonal or recombinant and be used in immunoassays. Conveniently, the antibodies can be prepared against the immunogen or portion thereof for example a synthetic peptide based on the sequence, or prepared recombinantly by cloning techniques or the natural gene product and/or portions thereof can be isolated and used as the immunogen. Immunogens can be used to produce antibodies by standard antibody production technology well known to those skilled in the art as described generally in Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY, 1988 and Borrebaeck, *Antibody Engineering - A Practical Guide*, W.H. Freeman and Co., 1992. Antibody fragments can also be prepared from the antibodies and include Fab, F(ab')₂, and Fv by methods known to those skilled in the art.

For producing polyclonal antibodies a host, such as a rabbit or goat, is immunized with the immunogen or immunogen fragment, generally with an adjuvant and, if necessary, coupled to a carrier; antibodies to the immunogen are collected from the sera. Further, the polyclonal antibody can be absorbed such that it is monospecific. That is, the sera can be absorbed against related immunogens so that no cross-reactive antibodies remain in the sera rendering it monospecific.

For producing monoclonal antibodies the technique involves hyperimmunization of an appropriate donor with the immunogen, generally a mouse, and isolation of splenic antibody producing cells. These cells are

fused to a cell having immortality, such as a myeloma cell, to provide a fused cell hybrid which has immortality and secretes the required antibody. The cells are then cultured, in bulk, and the monoclonal antibodies harvested
5 from the culture media for use.

For producing recombinant antibody (see generally Huston et al, 1991; Johnson and Bird, 1991; Mernaugh and Mernaugh, 1995), messenger RNAs from antibody producing B-lymphocytes of animals, or hybridoma are
10 reverse-transcribed to obtain complimentary DNAs (cDNAs). Antibody cDNA, which can be full or partial length, is amplified and cloned into a phage or a plasmid. The cDNA can be a partial length of heavy and light chain cDNA, separated or connected by a linker. The antibody, or
15 antibody fragment, is expressed using a suitable expression system to obtain recombinant antibody. Antibody cDNA can also be obtained by screening pertinent expression libraries.

The antibody can be bound to a solid support
20 substrate or conjugated with a detectable moiety or be both bound and conjugated, as is well known in the art. (For a general discussion of conjugation of fluorescent or enzymatic moieties see Johnstone & Thorpe, *Immunochemistry in Practice*, Blackwell Scientific Publications, Oxford,
25 1982.) The binding of antibodies to a solid support substrate is also well known in the art. (See for a general discussion Harlow & Lane *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory Publications, New York, 1988 and Borrebaeck, *Antibody Engineering - A
30 Practical Guide*, W.H. Freeman and Co., 1992). The detectable moieties contemplated with the present invention can include, but are not limited to, fluorescent, metallic, enzymatic and radioactive markers such as biotin, gold, ferritin, alkaline phosphatase, β -

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galactosidase, peroxidase, urease, fluorescein, rhodamine, tritium, ¹⁴C and iodination.

These transgenics and knock-outs of the present invention are constructed using standard methods known in the art and as set forth in United States Patents 5,487,992, 5,464,764, 5,387,742, 5,360,735, 5,347,075, 5,298,422, 5,288,846, 5,221,778, 5,175,385, 5,175,384, 5,175,383, 4,736,866 as well as Burke and Olson (1991), Capecchi (1989), Davies et al. (1992), Dickinson et al. (1993), Duff and Lincoln (1995), Huxley et al. (1991), Jakobovits et al. (1993), Lamb et al. (1993), Pearson and Choi (1993), Rothstein (1991), Schedl et al. (1993), Strauss et al. (1993). Further, patent applications WO 94/23049, WO 93/14200, WO 94/06908, WO 94/28123 also provide information.

More specifically, any techniques known in the art can be used to introduce the transgene expressibly into animals to produce the parental lines of animals. Such techniques include, but are not limited to, pronuclear microinjection (U.S. patent 4,873,191); retrovirus mediated gene transfer into germ lines (Van der Putten et al., 1985); gene targeting in embryonic stem cells (Thompson et al., 1989; Mansour, 1990 and U.S. patent 5,614,396); electroporation of embryos (Lo, 1983); and sperm-mediated gene transfer (Lavitrano et al., 1989). For a review of such techniques see Gordon (1989).

Further, one parent strain instead of carrying a direct human transgene can have the homologous endogenous gene modified by gene targeting such that it approximates the transgene. That is, the endogenous gene has been "humanized" and/or mutated (Reaume et al, 1996). It should be noted that if the animal and human sequence are essentially homologous a "humanized" gene is not required. The transgenic parent can also carry an over expressed sequence, either the non-mutant or a mutant sequence and

humanized or not as required. The term transgene is therefore used to refer to all these possibilities.

Additionally, cells can be isolated from the offspring which carry a transgene from each transgenic parent and that are used to establish primary cell cultures or cell lines as is known in the art.

Where appropriate, a parent strain will be homozygous for the transgene. Additionally, where appropriate, the endogenous non-transgene in the genome that is homologous to the transgene will be non-expressive. By non-expressive is meant that the endogenous gene will not be expressed and that this non-expression is heritable in the offspring. For example, the endogenous homologous gene could be "knocked-out" by methods known in the art. Alternatively, the parental strain that receives one of the transgenes could carry a mutation at the endogenous homologous gene rendering it non-expressed.

The antagonist/regulating agent/active ingredient is dosed and delivered in a pharmaceutically acceptable carrier as described herein below. The term antagonist or antagonizing is used in its broadest sense. Antagonism can include any mechanism or treatment which results in inhibition, inactivation, blocking or reduction in gene activity or gene product. It should be noted that the inhibition of a gene or gene product can provide for an increase in a corresponding function that the gene or gene product was regulating. The antagonizing step can include blocking cellular receptors for the gene products and can include antisense treatment as discussed herein below. For example, a patient can be in need of inducing apoptosis in tumorigenic cells or angiogenesis in trauma situations where for example a limb must be reattached or in a transplant where revascularization is needed.

Many reviews have covered the main aspects of

antisense (AS) technology and its enormous therapeutic potential (Wright and Anazodo, 1995). There are reviews on the chemical (Crooke, 1995; Uhlmann et al, 1990), cellular (Wagner, 1994) and therapeutic (Hanania, et al, 5 1995; Scanlon, et al, 1995; Gewirtz, 1993) aspects of this rapidly developing technology. Within a relatively short time, ample information has accumulated about the *in vitro* use of AS nucleotide sequences in cultured primary cells and cell lines as well as for *in vivo* administration of 10 such nucleotide sequences for suppressing specific processes and changing body functions in a transient manner. Further, enough experience is now available *in vitro* and *in vivo* in animal models and human clinical trials to predict human efficacy.

15 Antisense intervention in the expression of specific genes can be achieved by the use of synthetic AS oligonucleotide sequences (for recent reports see Lefebvre-d'Hellencourt et al, 1995; Agrawal, 1996; Lev-Lehman et al, 1997). AS oligonucleotide sequences can be 20 short sequences of DNA, typically 15-30 mer but can be as small as 7 mer (Wagner et al, 1996), designed to complement a target mRNA of interest and form an RNA:AS duplex. This duplex formation can prevent processing, splicing, transport or translation of the relevant mRNA. 25 Moreover, certain AS nucleotide sequences can elicit cellular RNase H activity when hybridized with their target mRNA, resulting in mRNA degradation (Calabretta et al, 1996). In that case, RNase H will cleave the RNA component of the duplex and can potentially release the AS 30 to further hybridize with additional molecules of the target RNA. An additional mode of action results from the interaction of AS with genomic DNA to form a triple helix which can be transcriptionally inactive.

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The sequence target segment for the antisense oligonucleotide is selected such that the sequence exhibits suitable energy related characteristics important for oligonucleotide duplex formation with their complementary templates, and shows a low potential for self-dimerization or self-complementation [Anazodo et al., 1996]. For example, the computer program OLIGO (Primer Analysis Software, Version 3.4), can be used to determine antisense sequence melting temperature, free energy properties, and to estimate potential self-dimer formation and self-complimentary properties. The program allows the determination of a qualitative estimation of these two parameters (potential self-dimer formation and self-complimentary) and provides an indication of "no potential" or "some potential" or "essentially complete potential". Using this program target segments are generally selected that have estimates of no potential in these parameters. However, segments can be used that have "some potential" in one of the categories. A balance of the parameters is used in the selection as is known in the art. Further, the oligonucleotides are also selected as needed so that analogue substitution do not substantially affect function.

Phosphorothioate antisense oligonucleotides do not normally show significant toxicity at concentrations that are effective and exhibit sufficient pharmacodynamic half-lives in animals (Agarwal et al., 1996) and are nuclease resistant. Antisense induced loss-of-function phenotypes related with cellular development were shown for the glial fibrillary acidic protein (GFAP), for the establishment of tectal plate formation in chick (Galileo et al., 1991) and for the N-myc protein, responsible for the maintenance of cellular heterogeneity in neuroectodermal cultures (epithelial vs. neuroblastic

cells, which differ in their colony forming abilities, tumorigenicity and adherence) (Rosolen et al., 1990; Whitesell et al., 1991). Antisense oligonucleotide inhibition of basic fibroblast growth factor (bFgF),
5 having mitogenic and angiogenic properties, suppressed 80% of growth in glioma cells (Morrison, 1991) in a saturable and specific manner. Being hydrophobic, antisense oligonucleotides interact well with phospholipid membranes (Akhter et al., 1991). Following their interaction with
10 the cellular plasma membrane, they are actively (or passively) transported into living cells (Loke et al., 1989), in a saturable mechanism predicted to involve specific receptors (Yakubov et al., 1989).

Instead of an antisense sequence as discussed
15 herein above, ribozymes can be utilized. This is particularly necessary in cases where antisense therapy is limited by stoichiometric considerations (Sarver et al., 1990, Gene Regulation and Aids, pp. 305-325). Ribozymes can then be used that will target the same sequence.
20 Ribozymes are RNA molecules that possess RNA catalytic ability (see Cech for review) that cleave a specific site in a target RNA. The number of RNA molecules that are cleaved by a ribozyme is greater than the number predicted by stoichiometry. (Hampel and Tritz, 1989; Uhlenbeck,
25 1987).

Ribozymes catalyze the phosphodiester bond cleavage of RNA. Several ribozyme structural families have been identified including Group I introns, RNase P, the hepatitis delta virus ribozyme, hammerhead ribozymes
30 and the hairpin ribozyme originally derived from the negative strand of the tobacco ringspot virus satellite RNA (sTRSV) (Sullivan, 1994; U.S. Patent No. 5,225,347, columns 4-5). The latter two families are derived from viroids and virusoids, in which the ribozyme is believed

to separate monomers from oligomers created during rolling circle replication (Symons, 1989 and 1992). Hammerhead and hairpin ribozyme motifs are most commonly adapted for trans-cleavage of mRNAs for gene therapy (Sullivan, 1994).

5 The ribozyme type utilized in the present invention is selected as is known in the art. Hairpin ribozymes are now in clinical trial and are the preferred type. In general the ribozyme is from 30-100 nucleotides in length.

Modifications or analogues of nucleotides can be
10 introduced to improve the therapeutic properties of the nucleotides. Improved properties include increased nuclease resistance and/or increased ability to permeate cell membranes.

Nuclease resistance, where needed, is provided
15 by any method known in the art that does not interfere with biological activity of the antisense oligodeoxynucleotides, cDNA and/or ribozymes as needed for the method of use and delivery (Iyer et al., 1990; Eckstein, 1985; Spitzer and Eckstein, 1988; Woolf et al., 1990; Shaw
20 et al., 1991). Modifications that can be made to oligonucleotides in order to enhance nuclease resistance include modifying the phosphorous or oxygen heteroatom in the phosphate backbone. These include preparing methyl phosphonates, phosphorothioates, phosphorodithioates and
25 morpholino oligomers. In one embodiment it is provided by having phosphorothioate bonds linking between the four to six 3'-terminus nucleotide bases. Alternatively, phosphorothioate bonds link all the nucleotide bases. Other modifications known in the art can be used where the
30 biological activity is retained, but the stability to nucleases is substantially increased.

The present invention also includes all analogues of, or modifications to, an oligonucleotide of the invention that does not substantially affect the
35 function of the oligonucleotide. The nucleotides can be

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selected from naturally occurring or synthetic modified bases. Naturally occurring bases include adenine, guanine, cytosine, thymine and uracil. Modified bases of the oligonucleotides include xanthine, hypoxanthine, 2-aminoadenine, 6-methyl, 2-propyl and other alkyl adenines, 5-halo uracil, 5-halo cytosine, 6-aza cytosine and 6-aza thymine, pseudo uracil, 4-thiuracil, 8-halo adenine, 8-aminoadenine, 8-thiol adenine, 8-thiolalkyl adenines, 8-hydroxyl adenine and other 8-substituted adenines, 8-halo guanines, 8-amino guanine, 8-thiol guanine, 8-thioalkyl guanines, 8-hydroxyl guanine and other substituted guanines, other aza and deaza adenines, other aza and deaza guanines, 5-trifluoromethyl uracil and 5-trifluoro cytosine.

15 In addition, analogues of nucleotides can be prepared wherein the structure of the nucleotide is fundamentally altered and that are better suited as therapeutic or experimental reagents. An example of a nucleotide analogue is a peptide nucleic acid (PNA) 20 wherein the deoxyribose (or ribose) phosphate backbone in DNA (or RNA) is replaced with a polyamide backbone which is similar to that found in peptides. PNA analogues have been shown to be resistant to degradation by enzymes and to have extended lives *in vivo* and *in vitro*. Further, 25 PNAs have been shown to bind stronger to a complementary DNA sequence than a DNA molecule. This observation is attributed to the lack of charge repulsion between the PNA strand and the DNA strand. Other modifications that can be made to oligonucleotides include polymer backbones, cyclic 30 backbones, or acyclic backbones.

The active ingredients of the pharmaceutical composition can include oligonucleotides that are nuclease resistant needed for the practice of the invention or a fragment thereof shown to have the same effect targeted

against the appropriate sequence(s) and/or ribozymes. Combinations of active ingredients as disclosed in the present invention can be used including combinations of antisense sequences.

5 The antisense oligonucleotides (and/or ribozymes) and cDNA of the present invention can be synthesized by any method known in the art for ribonucleic or deoxyribonucleic nucleotides. For example, an Applied Biosystems 380B DNA synthesizer can be used. When
10 fragments are used, two or more such sequences can be synthesized and linked together for use in the present invention.

 The nucleotide sequences of the present invention can be delivered either directly or with viral
15 or non-viral vectors. When delivered directly the sequences are generally rendered nuclease resistant. Alternatively, the sequences can be incorporated into expression cassettes or constructs such that the sequence is expressed in the cell as discussed herein below.
20 Generally, the construct contains the proper regulatory sequence or promoter to allow the sequence to be expressed in the targeted cell.

 Negative dominant peptide refers to a partial cDNA sequence that encodes for a part of a protein, i.e. a
25 peptide (see Herskowitz, 1987). This peptide can have a different function from the protein it was derived from. It can interact with the full protein and inhibit its activity or it can interact with other proteins and inhibit their activity in response to the full protein.
30 Negative dominant means that the peptide is able to overcome the natural proteins and fully inhibit their activity to give the cell a different characteristics like resistance or sensitization to killing. For therapeutic intervention either the peptide itself is delivered as the
35 active ingredient of a pharmaceutical composition or the

cDNA can be delivered to the cell utilizing the same methods as for antisense delivery.

By gene therapy as used herein refers to the transfer of genetic material (e.g. DNA or RNA) of interest
5 into a host to treat or prevent a genetic or acquired disease or condition phenotype. The genetic material of interest encodes a product (e.g. a protein, polypeptide, peptide, functional RNA, antisense) whose production *in vivo* is desired. For example, the genetic material of
10 interest can encode a hormone, receptor, enzyme, polypeptide or peptide of therapeutic value. Alternatively, the genetic material of interest encodes a suicide gene. For a review see, in general, the text "Gene Therapy" (Advances in Pharmacology 40, Academic
15 Press, 1997).

Two basic approaches to gene therapy have evolved: (1) *ex vivo* and (2) *in vivo* gene therapy. In *ex vivo* gene therapy cells are removed from a patient, and while being cultured are treated *in vitro*. Generally, a
20 functional replacement gene is introduced into the cell via an appropriate gene delivery vehicle/method (transfection, transduction, homologous recombination, etc.) and an expression system as needed and then the modified cells are expanded in culture and returned to the
25 host/patient. These genetically reimplanted cells have been shown to express the transfected genetic material *in situ*.

In *in vivo* gene therapy, target cells are not removed from the subject rather the genetic material to be
30 transferred is introduced into the cells of the recipient organism *in situ*, that is within the recipient. In an alternative embodiment, if the host gene is defective, the gene is repaired *in situ* [Culver, 1998]. These

genetically altered cells have been shown to express the transfected genetic material *in situ*.

The gene expression vehicle is capable of delivery/transfer of heterologous nucleic acid into a host cell. The expression vehicle can include elements to control targeting, expression and transcription of the nucleic acid in a cell selective manner as is known in the art. It should be noted that often the 5' UTR and/or 3' UTR of the gene can be replaced by the 5' UTR and/or 3' UTR of the expression vehicle. Therefore as used herein the expression vehicle can, as needed, not include the 5' UTR and/or 3' UTR of the actual gene to be transferred and only include the specific amino acid coding region.

The expression vehicle can include a promoter for controlling transcription of the heterologous material and can be either a constitutive or inducible promoter to allow selective transcription. Enhancers that can be required to obtain necessary transcription levels can optionally be included. Enhancers are generally any non-translated DNA sequence which works contiguously with the coding sequence (in cis) to change the basal transcription level dictated by the promoter. The expression vehicle can also include a selection gene as described herein below.

Vectors can be introduced into cells or tissues by any one of a variety of known methods within the art. Such methods can be found generally described in Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Springs Harbor Laboratory, New York (1989, 1992), in Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley and Sons, Baltimore, Maryland (1989), Chang et al., *Somatic Gene Therapy*, CRC Press, Ann Arbor, MI (1995), Vega et al., *Gene Targeting*, CRC Press, Ann Arbor, MI (1995), *Vectors: A Survey of Molecular Cloning Vectors*

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and *Their Uses*, Butterworths, Boston MA (1988) and Gilboa et al (1986) and include, for example, stable or transient transfection, lipofection, electroporation and infection with recombinant viral vectors. In addition, see United States patent 4,866,042 for vectors involving the central nervous system and also United States patents 5,464,764 and 5,487,992 for positive-negative selection methods.

Introduction of nucleic acids by infection offers several advantages over the other listed methods. Higher efficiency can be obtained due to their infectious nature. Moreover, viruses are very specialized and typically infect and propagate in specific cell types. Thus, their natural specificity can be used to target the vectors to specific cell types *in vivo* or within a tissue or mixed culture of cells. Viral vectors can also be modified with specific receptors or ligands to alter target specificity through receptor mediated events.

A specific example of DNA viral vector for introducing and expressing recombinant sequences is the adenovirus derived vector Adenop53TK. This vector expresses a herpes virus thymidine kinase (TK) gene for either positive or negative selection and an expression cassette for desired recombinant sequences. This vector can be used to infect cells that have an adenovirus receptor which includes most cancers of epithelial origin as well as others. This vector as well as others that exhibit similar desired functions can be used to treat a mixed population of cells and can include, for example, an *in vitro* or *ex vivo* culture of cells, a tissue or a human subject.

Additional features can be added to the vector to ensure its safety and/or enhance its therapeutic efficacy. Such features include, for example, markers that can be used to negatively select against cells infected

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with the recombinant virus. An example of such a negative selection marker is the TK gene described above that confers sensitivity to the antibiotic gancyclovir. Negative selection is therefore a means by which infection
5 can be controlled because it provides inducible suicide through the addition of antibiotic. Such protection ensures that if, for example, mutations arise that produce altered forms of the viral vector or recombinant sequence, cellular transformation will not occur.

10 Features that limit expression to particular cell types can also be included. Such features include, for example, promoter and regulatory elements that are specific for the desired cell type.

In addition, recombinant viral vectors are
15 useful for *in vivo* expression of a desired nucleic acid because they offer advantages such as lateral infection and targeting specificity. Lateral infection is inherent in the life cycle of, for example, retrovirus and is the process by which a single infected cell produces many
20 progeny virions that bud off and infect neighboring cells. The result is that a large area becomes rapidly infected, most of which was not initially infected by the original viral particles. This is in contrast to vertical-type of infection in which the infectious agent spreads only
25 through daughter progeny. Viral vectors can also be produced that are unable to spread laterally. This characteristic can be useful if the desired purpose is to introduce a specified gene into only a localized number of targeted cells.

30 As described above, viruses are very specialized infectious agents that have evolved, in many cases, to elude host defense mechanisms. Typically, viruses infect and propagate in specific cell types. The targeting specificity of viral vectors utilizes its natural

specificity to specifically target predetermined cell types and thereby introduce a recombinant gene into the infected cell. The vector to be used in the methods of the invention will depend on desired cell type to be targeted and will be known to those skilled in the art. For example, if breast cancer is to be treated then a vector specific for such epithelial cells would be used. Likewise, if diseases or pathological conditions of the hematopoietic system are to be treated, then a viral vector that is specific for blood cells and their precursors, preferably for the specific type of hematopoietic cell, would be used.

Retroviral vectors can be constructed to function either as infectious particles or to undergo only a single initial round of infection. In the former case, the genome of the virus is modified so that it maintains all the necessary genes, regulatory sequences and packaging signals to synthesize new viral proteins and RNA. Once these molecules are synthesized, the host cell packages the RNA into new viral particles which are capable of undergoing further rounds of infection. The vector's genome is also engineered to encode and express the desired recombinant gene. In the case of non-infectious viral vectors, the vector genome is usually mutated to destroy the viral packaging signal that is required to encapsulate the RNA into viral particles. Without such a signal, any particles that are formed will not contain a genome and therefore cannot proceed through subsequent rounds of infection. The specific type of vector will depend upon the intended application. The actual vectors are also known and readily available within the art or can be constructed by one skilled in the art using well-known methodology.

The recombinant vector can be administered in several ways. If viral vectors are used, for example, the

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procedure can take advantage of their target specificity and consequently, do not have to be administered locally at the diseased site. However, local administration can provide a quicker and more effective treatment, 5 administration can also be performed by, for example, intravenous or subcutaneous injection into the subject. Injection of the viral vectors into a spinal fluid can also be used as a mode of administration, especially in the case of neuro-degenerative diseases. Following 10 injection, the viral vectors will circulate until they recognize host cells with the appropriate target specificity for infection.

DNA can also be administered using a gene gun. (Ziao & Bracksman, Nucleic Acids, Res. 24, 2630-2622 15 (1996)). The DNA is precipitated onto the surface of microscopic metal beads. The microprojectiles are accelerated with a shock wave or expanding helium gas, and penetrate tissues to a depth of several cell layers. For example, the Acal™ Gene Delivery Device manufactured by 20 Aegacetus, Inc., Middleton, WI, is suitable. Alternatively, nucleic DNA can pass through skin into the bloodstream simply by spotting the DNA onto skin with chemical or mechanical irritation (see WO 95/05853).

An alternate mode of administration can be by 25 direct inoculation locally at the site of the disease or pathological condition or by inoculation into the vascular system supplying the site with nutrients or into the spinal fluid. Local administration is advantageous because there is no dilution effect and, therefore, a 30 smaller dose is required to achieve expression in a majority of the targeted cells. Additionally, local inoculation can alleviate the targeting requirement required with other forms of administration since a vector can be used that infects all cells in the inoculated area.

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If expression is desired in only a specific subset of cells within the inoculated area, then promoter and regulatory elements that are specific for the desired subset can be used to accomplish this goal. Such non-
5 targeting vectors can be, for example, viral vectors, viral genome, plasmids, phagemids and the like. Transfection vehicles such as liposomes can also be used to introduce the non-viral vectors described above into recipient cells within the inoculated area. Such
10 transfection vehicles are known by one skilled within the art.

The pharmaceutical compositions containing the active ingredients of the present invention as described herein above are administered and dosed in accordance with
15 good medical practice, taking into account the clinical condition of the individual patient, the site and method of administration, scheduling of administration, patient age, sex, body weight and other factors known to medical practitioners. The pharmaceutically "effective amount"
20 for purposes herein is thus determined by such considerations as are known in the medical arts. The amount must be effective to achieve improvement including but not limited to improved survival rate or more rapid recovery, or improvement or elimination of symptoms and
25 other indicators as are selected as appropriate measures by those skilled in the medical arts. The pharmaceutical compositions can be combinations of the active ingredients but will include at least one active ingredient.

In the method of the present invention, the
30 pharmaceutical compositions of the present invention can be administered in various ways taking into account the nature of compounds in the pharmaceutical compositions. It should be noted that they can be administered as the compound or as pharmaceutically acceptable salt and can be
35 administered alone or as an active ingredient in

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combination with pharmaceutically acceptable carriers, diluents, adjuvants and vehicles. The compounds can be administered orally, subcutaneously or parenterally including intravenous, intra-arterial, intramuscular, 5 intra-peritoneally, and intra-nasal administration as well as intra-thecal and infusion techniques. Implants of the compounds are also useful. The patient being treated is a warm-blooded animal and, in particular, mammals including man. The pharmaceutically acceptable carriers, diluents, 10 adjuvants and vehicles as well as implant carriers generally refer to inert, non-toxic solid or liquid fillers, diluents or encapsulating material not reacting with the active ingredients of the invention.

It is noted that humans are treated generally 15 longer than the mice or other experimental animals exemplified herein which treatment has a length proportional to the length of the disease process and drug effectiveness. The doses can be single doses or multiple doses over a period of several days, but single doses are 20 preferred.

The doses can be single doses or multiple doses over a period of several days. The treatment generally has a length proportional to the length of the disease process and drug effectiveness and the patient species 25 being treated.

When administering the compound of the present invention parenterally, it will generally be formulated in a unit dosage injectable form (solution, suspension, emulsion). The pharmaceutical formulations suitable for 30 injection include sterile aqueous solutions or dispersions and sterile powders for reconstitution into sterile injectable solutions or dispersions. The carrier can be a solvent or dispersing medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, 35 liquid polyethylene glycol, and the like),

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suitable mixtures thereof, and vegetable oils.

Proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of
5 dispersion and by the use of surfactants. Non-aqueous vehicles such as cottonseed oil, sesame oil, olive oil, soybean oil, corn oil, sunflower oil, or peanut oil and esters, such as isopropyl myristate, can also be used as solvent systems for compound compositions. Additionally,
10 various additives which enhance the stability, sterility, and isotonicity of the compositions, including antimicrobial preservatives, antioxidants, chelating agents, and buffers, can be added. Prevention of the action of microorganisms can be ensured by various
15 antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, and the like. In many cases, it will be desirable to include isotonic agents, for example, sugars, sodium chloride, and the like. Prolonged absorption of the injectable
20 pharmaceutical form can be brought about by the use of agents delaying absorption, for example, aluminum monostearate and gelatin. According to the present invention, however, any vehicle, diluent, or additive used would have to be compatible with the compounds.

25 Sterile injectable solutions can be prepared by incorporating the compounds utilized in practicing the present invention in the required amount of the appropriate solvent with various of the other ingredients, as desired.

30 A pharmacological formulation of the present invention can be administered to the patient in an injectable formulation containing any compatible carrier, such as various vehicle, adjuvants, additives, and diluents; or the compounds utilized in the present
35 invention can be administered parenterally to the patient

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in the form of slow-release subcutaneous implants or targeted delivery systems such as monoclonal antibodies, vectored delivery, iontophoretic, polymer matrices, liposomes, and microspheres. Examples of delivery systems
5 useful in the present invention include: 5,225,182; 5,169,383; 5,167,616; 4,959,217; 4,925,678; 4,487,603; 4,486,194; 4,447,233; 4,447,224; 4,439,196; and 4,475,196.

Many other such implants, delivery systems, and modules are well-known to those skilled in the art.

10 A pharmacological formulation of the compound utilized in the present invention can be administered orally to the patient. Conventional methods such as administering the compounds in tablets, suspensions, solutions, emulsions, capsules, powders, syrups and the
15 like are usable. Known techniques which deliver it orally or intravenously and retain the biological activity are preferred.

In one embodiment, the compound of the present invention can be administered initially by intravenous
20 injection to bring blood levels to a suitable level. The patient's levels are then maintained by an oral dosage form, although other forms of administration, dependent upon the patient's condition and as indicated above, can be used. The quantity to be administered will vary for
25 the patient being treated and will vary from about 100 ng/kg of body weight to 100 mg/kg of body weight per day and preferably will be from 10 µg/kg to 10 mg/kg per day.

The present invention also provides a method of diagnosing the presence of ischemia in a patient including
30 the steps of analyzing a bodily fluid or tissue sample from the patient for the presence or gene product of at least one expressed gene (up-regulated) or their proteins and where ischemia is determined if the up-regulated gene or gene product is ascertained as described herein in the

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Example. The bodily fluids can include tears, serum, urine, sweat or other bodily fluid where secreted proteins from the tissue that is undergoing an ischemic event can be localized. Additional methods for identification of the gene or gene product are immunoassays, such as and ELISA or radioimmunoassays (RIA), can be used as are known to those in the art particularly to identify gene products in the samples. Immunohistochemical staining of tissue samples is also utilized for identification. Available immunoassays are extensively described in the patent and scientific literature. See, for example, United States patents 3,791,932; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; 4,098,876; 4,879,219; 5,011,771 and 5,281,521. Further for identification of the gene, *in situ* hybridization, Southern blotting, single strand conformational polymorphism, restriction endonuclease fingerprinting (REF), PCR amplification and DNA-chip analysis using nucleic acid sequence of the present invention as primers can be used.

The above discussion provides a factual basis for the use of the sequences of the present invention to identify hypoxia-regulated genes and provide diagnostic probes to identify ischemia. The methods used with and the utility of the present invention can be shown by the non-limiting examples herein.

GENERAL METHODS

Most of the techniques used in molecular biology are widely practiced in the art, and most practitioners are familiar with the standard resource materials which describe specific conditions and procedures. However, for convenience, the following paragraphs can serve as a

guideline.

General methods in molecular biology: Standard molecular biology techniques known in the art and not specifically described were generally followed as in
5 Sambrook et al., *Molecular Cloning: A Laboratory Manual*, Cold Springs Harbor Laboratory, New York (1989), and in Ausubel et al., *Current Protocols in Molecular Biology*, John Wiley and Sons, Baltimore, Maryland (1989) particularly for the Northern Analysis and In Situ
10 analysis and in Perbal, *A Practical Guide to Molecular Cloning*, John Wiley & Sons, New York (1988), and in Watson et al., *Recombinant DNA*, Scientific American Books, New York. Polymerase chain reaction (PCR) was carried out generally as in *PCR Protocols: A Guide To Methods And*
15 *Applications*, Academic Press, San Diego, CA (1990).

Reactions and manipulations involving other nucleic acid techniques, unless stated otherwise, were performed as generally described in Sambrook et al., 1989, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor
20 Laboratory Press, and methodology as set forth in United States patents 4,666,828; 4,683,202; 4,801,531; 5,192,659 and 5,272,057 and incorporated herein by reference.

Additionally, *in situ* (In cell) PCR in combination with flow cytometry can be used for detection
25 of cells containing specific DNA and mRNA sequences (Testoni et al, 1996, Blood 87:3822).

General methods in immunology: Standard methods in immunology known in the art and not specifically described are generally followed as in Stites et al.(eds),
30 *Basic and Clinical Immunology* (8th Edition), Appleton & Lange, Norwalk, CT (1994) and Mishell and Shiigi (eds), *Selected Methods in Cellular Immunology*, W.H. Freeman and Co., New York (1980). Available immunoassays are

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extensively described in the patent and scientific literature. See, for example, United States patents 3,791,932; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262; 3,901,654; 3,935,074; 3,984,533; 5 3,996,345; 4,034,074; 4,098,876; 4,879,219; 5,011,771 and 5,281,521 as well as Sambrook et al, *Molecular Cloning: A Laboratory Manual*, Cold Springs Harbor, New York, 1989.

10

GENERAL METHODS OF THE INVENTION

The general methods of the invention are generally as described in US patent application serial number USSN 09/309,862 of same applicant which is by 15 reference incorporated herein in its entirety.

In brief, USSN 09/309,862 provides methods for identifying genes regulated at the RNA level by cue-induced gene expression. It relates to the rapid isolation of differentially expressed or developmentally regulated 20 gene sequences through analysis of mRNAs obtained from specific cellular compartments and comparing the changes in the relative abundance of the mRNA in these compartments as a result of applying a cue to the tested biological samples. The cellular compartments include 25 polysomal and nonpolysomal fractions, nuclear fractions, cytoplasmic fractions and splicesomal fractions. The method includes the steps of exposing cells or tissue to a cue or stimulus such as mechanical, chemical, toxic, pharmaceutical or other stress, hormones, physiological 30 disorders or disease; fractionating the cells into compartments such as polysomes, nuclei, cytoplasm and splicesomes; extracting the mRNA from these fractions, and subjecting the mRNA to differential analysis using accepted methodologies, such as gene expression array

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(GEM) .

The method is designed for identifying and cloning genes which are either up- or down-regulated responsive to a specific pathology, stress, physiological
5 condition, and so on, and in general, to any factor that can influence cells or organisms to alter their gene expression.

Further in USSN 09/309,862, an example is provided which shows the use of RNA isolation from nuclei
10 for isolating genes whose steady state levels show only minor changes, but which show high differential expression when detected by nuclear RNA probe. Most such genes are regulated at the transcriptional level.

The specific mRNA of the invention is total
15 cellular mRNA, and regulation is specifically on the transcriptional level.

DNA Microarrays

Preparation of custom hypoxia-specific microarrays

20 The cell system consisted of the C6 or A172 glioma cell lines. The cells were exposed to hypoxia for 4 or 16 hours and compared to cells grown under normal conditions.

Subtracted libraries were made from the
25 following sample:

1. 16 hours hypoxia vs. normal (genes induced by hypoxia after 16 hours).
2. Normal vs. 16 hours hypoxia (genes reduced by
30 hypoxia).
3. 4 hours hypoxia vs. normal (genes induced by hypoxia after four hours).

Three enriched libraries from the three groups
35 above were made by the SSH method using the "PCR select

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cDNA subtraction kit" from Clontech. From library 1, 1,000 colonies were grown and the plasmids prepared in 96 well format. From libraries 2 and 3, 500 colonies were processed from each. Thus, a total of 2,000 individual
5 plasmids were prepared and used for the fabrication of a Gene Expression Microarray (GEM). For this, the inserts of each plasmid were amplified by PCR and robotically fabricated on the glass.

10 Preparation of probes for microarray hybridization

Isolated messenger RNA is labeled with fluorescent dNTP's using a reverse transcription reaction to generate a labeled cDNA probe. mRNA is extracted from either C6 or A172 cells cultured in normoxia conditions
15 and labeled with Cy3-dCTP (Amersham) and mRNA extracted from C6 or A172 cells cultured under hypoxia conditions is labeled with Cy5-dCTP (Amersham). The two labeled cDNA probes are then mixed and hybridized onto microarrays (Schena et al, 1996). Following hybridization the
20 microarray is scanned using a laser scanner and the amount of fluorescence of each of the fluorescence dyes is measured for each cDNA clone on the micro-array giving an indication of the level of mRNA in each of the original mRNA populations being tested. Comparison of the
25 fluorescence on each cDNA clone on the micro-array between the two different fluorescent dyes is a measure for the differential expression of the indicated genes between the two experimental conditions.

30 The following probes were made from C6 and A172 for screening the GEM:

1. Normoxia (Cy3 labeled) + 16 hours hypoxia (Cy5 labeled).

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2. Normoxia (Cy3 labeled) + 4 hours hypoxia (Cy5 labeled).

5 The following cDNA sequences were found to be induced under hypoxia conditions. The sequences are divided into three categories: 1. new genes; 2. known genes not known before this publication to be hypoxia-inducible; and 3. known genes known to be induced under hypoxia conditions.

10

In Situ Analysis:

In situ analysis is performed for the candidate genes identified by the differential response to exposure to hypoxia conditions as described above. The expression
15 is studied in normal tissues and in pathological models as described herein.

Utilizing microarray hybridization the sequences set forth herein were identified and cloned as being differentially expressed under hypoxia conditions (see
20 also Braren et al, 1997).

In parallel experiments Northern Analysis results and results obtained by the gene expression microarray analysis were found to coincide and either can be used to determine hypoxia-regulated response. As well
25 in other experiments, the results from *in situ* analysis showed a high degree of correlation with the Northern Analysis and microarray analysis.

The sequences are listed that were found, the sequences are identified by clone number. In some cases
30 either end of the clone has been sequenced for use or the entire clone sequence and protein sequence are provided.

Unigem1 (Syntheni) was utilized for screening of human glioma cell line A172 to identify genes whose expression is modified by hypoxia.

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A Retinopathy Model:

Three major biological processes occur in nervous tissues under hypoxic conditions:

1. apoptotic death of hypoxia-damaged cells;
2. angiogenesis induced by factors secreted by hypoxia-suffering cells (a feedback control of oxygen concentration in tissue); and
3. secretion of neurotrophic and neuroprotective factors.

Therefore, it was assumed that among novel genes transcriptionally regulated by hypoxia in C6 and A172 glioma cells, there are those with pro- and antiapoptotic function as well as secreted neurotrophic, neuroprotective and angiogenic factors. It is worth noting, that regulation of apoptosis and angiogenesis is closely linked to cancerogenesis.

As initial step of biological characterization, candidate genes were tested for their ability to induce/protect cells from apoptosis, for neurotrophic activity and for angiogenic/antiangiogenic activity.

The sequences of the invention, the methods used therewith and the utility of the present invention can be shown by the following non-limiting examples:

EXAMPLE 1: 92 (SEQ ID. No.: 1)Northern Blot Analysis:

Gene 92 is found upregulated after 16 hours of hypoxia. On Northern blots, it appears as a single 5 Kb transcript.

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Cloning:

Several partial human cDNA clones corresponding to gene 92 were isolated from human A172 cDNA library. The
5 length of available contig is 2212 bp and it contains an ORF potentially coding for a 437 amino acids (265-1576 bp) protein. The putative initiating ATG codon is preceded by in frame stop codon.

10 Bioinformatic Analysis:

Similarity search with 92 cDNA sequence against the public databases have shown 60% similarity to unknown Drosophila DNA sequence (AC004283) and mainly encompasses the 3' UTR and a part of the coding sequence. The search
15 against the protein public databases gave partial similarity to hypothetical C. elegans protein (1703624) (77% similarity and 46% Identity).

The 92 cDNA sequence contains a region of 55 nucleotides (336-390 bp) that is constituted of CGG
20 repeats. On the level of amino acids it appears as a GGD/SFGG repeated unit (aa 24-44). Two of the isolated cDNA clones contain a 30 nucleotides in frame deletion within this region, indicating that the amount of repeats can be variable. 44 of these nucleotides form a strong
25 stem and loop secondary structure. When 92 cDNA was in vitro translated, the obtained protein had much smaller size than expected (30 kD instead of 45 kD). This, means that the stem and loop structure formed right downstream to the putative initiation codon prevents the proper
30 progression of ribosome and the initiation actually starts from the next in frame ATG located at position 820 - 822.

EXAMPLE 2: 95 (SEQ ID. No.: 2)

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Northern Blot Analysis:

Expression of this gene is upregulated by hypoxia. In normoxic conditions it appears as a single
5 moderately expressed 3.9 Kb transcript. After four hours of hypoxia, an additional transcript of 4.3 Kb becomes evident. After 16 hours of hypoxia, an increase in the amount of the major 3.9 Kb transcript takes place and there appears the second additional transcript 4.5 Kb in
10 size.

Cloning:

A complete human 95 cDNA clone, corresponding to the major 3.9 Kb mRNA was isolated. The contig is 3,535 bp
15 long and it potentially codes for a 480 aa (ORF: nt 323-1763) protein. All the attempts to clone the gene 95 splice variants had thus far failed, although numerous different approaches were employed.

20 Bioinformatic And Literature Analysis:

Search against public databases revealed that gene 95 is similar to but not identical with a recently cloned human P53-responsive gene PA26 that codes for a nuclear protein (Oncogene, 18, 127 - 137, 1999). Like gene
25 95, the PA26 one is expressed in three splice variants. The major transcript is 2.6 Kb in size (T2) and is ubiquitously expressed, the second transcript is ~4 Kb (T1) and displays a certain tissue specificity of expression pattern. The third transcript is approximately
30 2.3 Kb (T3) and is very minor. Expression of 2.6 and 2.3 Kb transcripts is p53-regulated and is increased in response to irradiation, UV, doxorubicin etc. The 4 Kb transcript is not induced by p53. The PA26-specific transcripts share a set of C-terminal common exons, while
35 the first exon is individual for each splice form. Gene 95

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encoded putative protein shares the maximal amino acid
homology with the common region of PA26-specific proteins.
In its N-terminus, the putative protein 95 mostly
resembles the T2 (major p53-inducible variant) variant of
5 PA26.

PA26, maps to chromosome 6q21. Biologically it
shares properties common to the GADD family of Growth
Arrest and DNA Damage-inducible stress-response genes.

10 Domain Analysis:

A short sequence with features of second
paroxysmal targeting signal was found in protein 95
between amino acids 353-361. A putative coiled coil
region is found between amino acid positions 253-283.

15

Further Analysis Of Gene 95 Expression Pattern:

By analogy with gene PA26, it was assumed that
alternative splicing in gene 95 also occurs in the first
exons. To identify the mRNA species where the cDNA clone
20 was derived from, two 95-specific probes were synthesized:
(1) corresponding to the region homologous to the first
exon of PA26 transcript T2 (Probe 1), and (2)
corresponding to the region homologous to a common set of
PA26 exons (but encompassing the sequences that are mostly
25 diverged between the two genes) (Probe 2). On Northern
blots, both probes recognized the same major 3.9 Kb 95-
specific transcript that is up-regulated by hypoxia. No
additional transcripts were identified by Probe 2. The 95-
specific probe that was used for the initial set of
30 hybridizations did not recognize any additional
transcripts when hybridized to this newly prepared
Northern blot. Therefore, the occurrence of 95-specific
additional mRNA species seem to vary among different mRNA
preparations.

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The sizes of PA26 and gene 95 encoded transcripts are different. However, in order to exclude the possibility that the presence of alternative 95 RNA species stem from cross-hybridization, a PA26 specific
5 cDNA probe was synthesized originating from the common exons region. On the Northern blot used for the previous experiment, the PA26 hybridized to two typical mRNA species of 2.6 and 4 Kb. Their levels, unlike the level of 95 mRNA, were not affected by hypoxia.

10 There is contradictory data in the literature about the nature of p53 protein present in A172 cells: according to some sources, it is wild type, according to others, mutated. The experiments performed thus far, conclude that hypoxia-induced expression of gene 95 is
15 p53-independent.

The mammalian 95 expression vector was prepared and effects of this gene overexpression in mammalian cells were studied.

20 Summary:

95 is a novel hypoxia-induced human gene coding for a protein similar to p53-induced GADD protein PA26. However, the latter is inert to hypoxia stimulation. Therefore, there appears to exist two closely related
25 proteins encoded by different genes that are upregulated by different genotoxic stresses. Protein 95 contributes to hypoxia-induced growth regulation in a way similar to PA26, but in a p53 independent manner. It is known that hypoxia-induced growth arrest that can take place in the
30 core regions of p53-negative tumors is one of the obstacles for success of full chemotherapy. Therefore, inactivation of gene 95 can be a potent adjuvant for chemotherapeutic treatment of cancers.

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EXAMPLE 3: 98 (SEQ. ID. No.: 3)**Northern Blot Analysis:**

5 Expression of gene 98 is strongly up-regulated by hypoxia already after four hours of exposure. On Northern blots, it appears as a single mRNA species of 4.4. Kb.

Cloning:

10 A full-length 98 cDNA was cloned. It is 4138 bp long and contains an single ORF encompassing the nucleotides 204 - 1445. The putative protein is 414 amino acid a long.

15 **Bioinformatic Analysis:**

Search of the public databases revealed that 98 encoded protein is similar to two other human proteins: (1) a putative protein encoded by anonymous human 24945 mRNA sequence (AF131826) and (2) VDUP1 (protein induced in
20 HL-60 cells by dihydroxy vitamin D3 treatment) (S73591). No significant structural features were found by existing protein analysis tools within the 98 putative protein.

It was previously demonstrated that treatment with vitamin D3 can induce apoptosis in C6 rat glioma
25 cells (J. Neurosci Res, 46, 540 - 550). Therefore, the relationship between the vitamin D-induced cell killing and 98 gene expression and function in glioma cells was studied.

The mammalian 98 expression vector was then and
30 its effects studied.

EXAMPLE 4: 60F6 (SEQ. ID. No.: 4)

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Northern Blot Analysis:

Expression of this gene is moderately up-regulated after 16 hours of hypoxia. On Northern blot, it appears as a single 3.0 Kb species.

5

Cloning:

A complete 60F6 human cDNA clone was isolated from A172 cDNA library. The contig is 2675 bp long and contains a single ORF (bp 134 - 866) able to code for a putative protein of 244 amino acids.

10

Bioinformatic Analysis:

A similarity search against the public databases revealed that the N-terminal half of 60F6 sequence exactly corresponds to a human cDNA coding for RhoE/Rho8 small GTP-binding protein (P52199, HSRHO8GRN). The identity of gene 60F6 was not determined before, because the small sequenced fragment that was initially possessed, originated from the Rho8 long 3' UTR. All the sequence information available in public databases did not include the long 3' UTR of Rho8. Structurally, Rho8 belongs to a family of Ras-related GTPases that regulate the actin cytoskeleton. However, this protein is unique in that it is constitutively active: GTPase deficient and *in vivo* farnesylated (Mol Cell Biol. 1996 Jun; 16(6): 2689-99). Therefore, it is intriguing to find that this constitutively active G-protein is regulated on the level of transcription. Hypoxia regulation of Rho8 was not previously described.

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EXAMPLE 5: 648 (lysyl hydroxylase 2) (SEQ. ID. No.: 5)Northern analysis

Probe 648 has detected a single 3.8 Kb

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transcript on Northern blots. Expression was induced in C6 glioma cells already after 4 hours of hypoxia.

Cloning

5 After extension of initial cDNA probe by RACE it became evident that identified rat sequence is able to code for protein that represents a rat homologue of human lysyl hydroxylase 2 (PLOD2). The full-length open reading frames was cloned for both human and rat lysyl
10 hydroxylase 2 homologues (by PCR, using primers built on the basis of known sequence, for human variant, and degenerative primers, for rat variant). The encoded proteins have well defined signal peptides.

15 Bioinformatics data

 The cloned rat 648 cDNA contains an ORF coding for a putative protein that is 88% identical to the published human PLOD2 sequences. The least conserved sequences are within the signal peptide, however its
20 functional features are completely preserved. The cloned human cDNA is almost identical to published human PLOD2 sequence. The word "almost" in the previous sentence stems from the fact that both in human and in rat cDNA species cloned in QBI a stretch of amino acids between positions
25 501- 521 of published sequence PLOD2 sequence was absent. Therefore, QBI's PLOD2 variants are differentially spliced. Both rat and human homologues were amplified from RNA extracted from glioma cell lines cultured in hypoxic conditions.

30

Literature review

 Lysyl hydroxylases are the enzymes that catalyze the formation of hydroxylysine in collagens and other proteins with collagen-like amino-acid sequences, by the
35 hydroxylation of lysine residue in X-K-G sequences. The

hydroxyllysine residues have two important functions: (1) serve as sites of attachment of carbohydrate units, and (2) they are essential for the stability of the intermolecular collagen crosslinks. Congenital deficiency of lysyl hydroxylase in humans leads to increased solubility of collagens and, consequently, to numerous defects in organization of connective tissue in various organs. There are three known isoforms of lysyl hydroxylase, encoded by different genes. In humans, PLOD2 was found to be highly expressed in pancreas, skeletal muscle, heart and placenta (by Northern blot). Nothing is known either about the regulation of PLOD2 expression by hypoxia or about its involvement in angiogenesis and tumorigenesis. Induction of PLOD2 by hypoxia can probably account for hypoxia-induced tissue fibrosis. Indeed, specific lysyl hydroxylase inhibitor, minoxidil, was able to suppress both cellular collagen production and fibroblasts proliferation (J. Biol. Chem., 262, 11973 -8, 1987; Graefes Arch. Clin. Exp. Ophthalmol. 233, 347 -55, 1995). There were suggestions in literature to use modified lysyl hydroxylase inhibitor for treatment of vitreoretinopathy (Invest. Ophthalmol. Vis .Sci. 34, 567-75, 1993).

25 Analysis of alternatively spliced versions of gene 648

In order to establish whether the observed alternative splicing of PLOD2 is regulated by hypoxia, a set of PCR primers were synthesized that flank the alternatively spliced region. The expected sizes of RT-PCR products are: 216 bp, for published sequence and 156 bp, for the present sequence. Semi-quantitative RT-PCR was performed on RNA template extracted from human glioma A172 cells culture in either normoxia or in hypoxia for 4 and 16 hours. The obtained results clearly demonstrate that both PLOD2 forms are hypoxia regulated, but the form of

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the invention appears only in hypoxic conditions.

Testing potential pro- and antiapoptotic activity in transient transfection assays

5 pcDNA3-648 was transiently co-transfected together with pcDNA3-GFP in Hela and 293 cells. 24 and 48 hours later the cells were fixed and stained with DAPI. No apoptotic effect was observed in the transfected cells. In order to evaluate the anti-apoptotic properties of the 648
10 protein, a co-transfection assay was conducted using the pcDNA3-GFP and the FAS plasmids. No anti-apoptotic effect was observed.

Obtaining stable cell clones overexpressing 648 cDNA

15 C6 were stably transfected with 5 ug of the pcDNA3-648 plasmid. Following G418 selection the level of expression was measured using Northern blot analysis in comparison to its level in C6 cells after 16 hours under hypoxic conditions. Out of 18 independent clones from the
20 pcDNA3-648 transfection, no one was positive.

In situ hybridization analysis

Retinopathy model

25 Probe 648 demonstrates clear hybridization signal throughout the inner nuclear layer of "hypoxic" pup's retina while "normoxic" retina is negative for the expression. No hybridization signal was detected in adult retina.

30 In mouse embryo sections hybridization signal was detected in some apoptotic cells in the roof of the fourth brain ventricle and in developing retina ganglia, where expressing cells had no apoptotic features.

35 Multi-tissue block hybridization shows expression of 648 gene (rat PLOD2) in visceral smooth muscles in oviduct, uterus, stomach and intestine. Vascular smooth

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muscles do not display hybridization signal.

Most prominent cell type hybridizing to 648 probe in ovary are granulosa cells of larger secondary follicles. No hybridization signal is detected in
5 granulosa cells of primary and small secondary follicles. Significantly, hybridization signal is weakened in postovulatory follicles and completely disappears in corpora lutea. This shows that expression in granulosa cells is established at later stages of follicular
10 maturation and it is abruptly down-regulated upon ovulation and the onset of conversion into lutein cells. On the other hand, follicular involution is not accompanied by the changes in 648 expression since strong hybridization signal is preserved in granulosa cells of
15 atretic follicles.

Weak hybridization signal can be seen in some stromal cells surrounding large secondary follicles and corpora lutea as well as in cells of theca internal of secondary follicles. Prominent signal is found in
20 "interstitial glands". This shows distinct regulation of 648 expression in theca cells undergoing "luteinization" in different locations: it is down regulated in corpora lutea but preserved or even upregulated in interstitial glands.

25 As to the germinal cells, an oocyte that expresses 648 was found only in one primary follicle while many other primary and secondary follicles had no hybridization signal. This shows a transient expression of 648 in oocytes at some stage of their development.

30 Discrepancy in the hybridization patterns of human (published) and rat PLOD2 (648) genes is explained by different sensitivities of different detection methods (Northern blot vs. *in situ* hybridization). The rat probe used in the present invention does not span an
35 alternatively spliced region.

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EXAMPLE 6: 24D4 (SEQ. ID. No.: 6)5 Northern blot analysis

Expression of gene 24D4 is down-regulated after 16 hours of hypoxia. On Northern blots, it appears as a single 1.5 Kb mRNA species.

10 Cloning

A partial 24D4 human cDNA clone was isolated from A172 cDNA library. The available sequence is 1486 bp long and contains an N-terminal truncated ORF (bp 1-397).

15 Bioinformatic analysis

The sequence has no analogs in public databases. The available protein sequence contains three consequent Zn-finger motifs, all of C2H2 type (aa 52-72, 80-100 and 108-128). Zinc finger domains of this type are usually
20 found in nucleic acid-binding proteins.

EXAMPLE 7: 77H4 (SEQ. ID. No.: 7)Northern blot analysis

25 Expression of gene 77H4 is up-regulated after 16 hours of hypoxia. On Northern blots, it appears as a single mRNA species 0.6 -0.7 Kb in size.

Cloning

30 Several EST cDNA clones from public databases, corresponding to clone 77H4, were sequenced. All clones possess a poly A tail and a polyadenylation signal at their 3' end.

Bioinformatic analysis

EST clone 18E contains 580 bp, that include an ORF for a 120 aa protein (bp 92 - 452). The EST clone 3D contains 486 bp. Comparison of nucleotide sequences of clone 18E and 3D reveal that the latter has a out-of-frame deletion of 71 nucleotides between bp 279-339 of clone 18E (its putative coding region)(Fig.68). This raised the question whether 77H4 cDNA is at all coding. The sequence was analyzed by Genscan program, that predicts the potential coding sequences on the basis of codon usage. The only potentially coding segment was found in clone 18E between bp 93 and 235. An exhaustive search was performed of public databases for all 77H4-related sequences. Several independent contigs were identified in TIGR THC database. All of them are not completely identical to one another and contain nucleotide deletions of various length. This shows a certain variability in 77H4 nucleotide sequence.

Recently, a novel steroid receptor transcriptional coactivator, SRA, was found to be present as an RNA molecule in the transcription activating complex SRC-1 (Cell, 97, 17 - 27, 1999) . Although no similarity was found between clone 77H4 and SRA RNA on the sequence level, several characteristic features seem to be shared by both sequences:

- 1 both mRNAs, 77H4 and SRA, are approximately of the same size - 0.7 Kb;
- 2 sequencing multiple cDNA clones corresponding to either mRNA revealed extensive variability in certain regions;
- 3 hybridization signals of both mRNA, therefore, appear as fuzzy bands on Northern blots;
- 4 both mRNA do not exhibit characteristics of protein.

Therefore, the 77H4 cDNA clone has similar to

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SRA function and can serve a coactivator in some transcriptional complexes.

EXAMPLE 8: 14G2 (SEQ. ID. No.: 8)

5

Northern blot analysis

Expression of gene 14G2 is regulated within 16 hours of hypoxia. On Northern blots, it appears as a single mRNA species.

10

Cloning

A partial 14G2 human cDNA clone was isolated. The available sequence was then characterized and cloned as shown in Figure 8.

15

EXAMPLE 9: 29F3 (SEQ. ID. No.: 9)

Northern blot analysis

Expression of gene 29F3 is regulated within 16 hours of hypoxia. On Northern blots, it appears as a single mRNA species.

20

Cloning

A partial 29F3 human cDNA clone was isolated. The available sequence was then characterized and cloned as shown in Figure 9.

25

Throughout this application, various publications, including United States patents, are referenced by author and year and patents by number. Full citations for the publications are listed below. The disclosures of these publications and patents in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

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The invention has been described in an illustrative manner, and it is to be understood that the terminology which has been used is intended to be in the nature of words of description rather than of limitation.

5 Obviously, many modifications and variations of the present invention are possible in light of the above teachings. It is, therefore, to be understood that within the scope of the described invention, the invention can be practiced otherwise than as specifically described.

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AMENDED CLAIMS

[received by the International Bureau on 15 February 2000 (15.02.00);
original claims 1-12 replaced by new claims 1-12;
original claims 13-15 cancelled (2 pages)]

1. A polynucleotide selected from the group consisting of SEQ ID Nos: 1-9, and functional analogs and complements thereof, which is differentially expressed and/or modulated under hypoxic conditions.

2. The polynucleotide according to claim 1 having hypoxia-regulating activity.

3. A diagnostic tool for identifying genes modulated by hypoxic conditions comprising detection means for detecting the presence of the polynucleotide of claim 1 and functional analogs thereof.

4. A method of diagnosing the presence of a hypoxia-associated pathology by screening for the presence of the polynucleotide of claim 1 and functional analogs thereof.

5. A method of regulating hypoxia-associated pathologies by administering a pharmaceutically effective amount of the polynucleotide of claim 1 and functional analogs thereof and a pharmaceutically acceptable carrier.

6. A method of regulating hypoxia-associated pathologies by administering a pharmaceutically effective amount of a protein encoded by the polynucleotide of claim 1 and functional analogs thereof as active ingredients in a pharmaceutically acceptable carrier.

7. A method of regulating hypoxia-associated pathologies by administering an effective amount of a direct or indirect biologically active product of enzymatic activity of a protein encoded by the polynucleotide sequence according to claim 1 and functional analogs thereof as active ingredients in a pharmaceutically acceptable carrier.

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AMENDED SHEET (ARTICLE 19)

8. A method of regulating hypoxia-associated pathologies by administering a pharmaceutically effective amount of inhibitors of a direct or indirect biologically active product of enzymatic activity of a protein encoded by the polynucleotide sequence according to claim 1 and functional analogs thereof as active ingredients in a pharmaceutically acceptable carrier.

9. A method of regulating hypoxia-associated pathologies by inhibiting the enzymatic activity of a protein encoded by the polynucleotide sequence according to claim 1 and functional analogs thereof as active ingredients in a pharmaceutically acceptable carrier.

10. A pharmaceutical composition for modulating hypoxia and ischemia comprising an effective amount of a polynucleotide according to claim 1 and functional analogs thereof and a pharmaceutically acceptable carrier.

11. The pharmaceutical composition according to claim 10, wherein said pharmaceutical composition is used for gene therapy.

12. A pharmaceutical composition for modulating hypoxia and ischemia comprising a pharmaceutically effective amount of a protein encoded by the nucleic acid sequence according to SEQ ID NOS. 1-9 and functional analogs thereof and a pharmaceutically acceptable carrier.

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GCACNAGGTGTGTGGCAGCAANAGCCGCCAGTTCGGGACCNC CGCANCTGGGGTGGCAAC
GGCGCAGGAGGGGTGCGGGGAGGGAGTGGTGAGCGCAGGCGGCAGGGGTGTGGGAAAGA
CGAAGTCGCTATTTGCTGTCTGAGCGCGCTCGCAGCTCCTGGAAGTGTTGCCGCCTCTCG
GTTTCGCTCTCGCTCGCTGCGCTCCTAGAAGGGGGCGGCCGCTCCAGGACTGACCAGGGC
CAAGTGGCGCTCGGCGGGCACTACATGGCGGAGGGTGAAGGGTACTTCGCCATGTCTGAG
GACGAGCTGGCCTGCAGCCCCTACATCCCCCTAGGCGGGCGACTTCGGCGGGCGGGCGACTTC
GGCGGGCGGGCGACTTCGGCGGTGGCGGCAGCTTCGGTGGGCATTGCTTGGACTATTGCGAA
AGCCCTACGGCGCACTGCAATGTGCTGAACTGGGAGCAAGTGCAGCGGCTGGACGGCATC
CTGAGCGAGACCATTCCGATTCACGGGCGCGGCAACTTCCCCACGCTCGAGCTGCAGCCC
AGCCTGATCGTGAAGGTGGTGCGGGCGGCGCCTGGCCGAGAAGCGCATTGGCGTCCGCGAC
GTGCGCCTCAACGGCTCGGCAGCCAGCCATGTCTGCACCAGGACAGCGGCCTGGGCTAC
AAGGACCTGGACCTCATCTTCTGCGCCGACCTGCGCGGGGAAGGGGAGTTTCAGACTGTG
AAGGACGTCTGTGCTGGACTGCCTGTTGGACTTCTTACCCGAGGGGGTGAACAAAGAGAAG
ATCCACCAGTCACGCTCAAGGAAGCTTATGTGCAGAAAATGGTTAAAGTGTGCAATGAC
TCTGACCGATGGAGTCTTATATCCCTGTCAAACAACAGTGGCAAAAAATGTGGAAGTGA
TTTGTGGATTCCCTCCGGAGGCAGTTTGAATTCAGTGTAGATTCTTTTCAAATCAAATTA
GACTCTCTTCTGCTCTTTTATGAATGTTTCAAGAGAACCCAATGACTGAGACATTTTACCCC
ACAATAATCGGGGAGAGCGTCTATGGCGATTTCCAGGAAGCCTTTGATCACCTTTGTAAC
AAGATCATTGCCACCAGGAACCCAGAGGAAATCCGAGGGGGGAGGCCTGCTTAAGTACTGC
AACCTCTTGGTCAGGGGGCTTTAGGCCCGCCTCTGATGAAATCAAGACCCTTCAAAGGTAT
ATGTGTTCCAGGTTTTTTCATCGACTTCTCAGACATTGGAGAGCAGCAGAGAAAAGTGGAG
TCCTATTTGCAGAACCTCTTTGTGGGATTGGAAGCCCGCAAGTATGAGTATCTCATGACC
CTTCATGGAGTGGTAAATGAGAGCTCAGTGTGCCTGATGGGACATGAAAGAAGACAGACT
TTAAACCTTATCACCATGCTGGCTATCCGGGTGTTAGCTGACCAAAATGTCATTCTTAAT
GTGGCTAATGTCACCTTGATATTACCAGCCAGCCCCCTATGTAGCAGATGCCAACTTTAGC
AATTACTACATTGCACAGGTTTCAAGCCAGTATTCACGTGCCAGCAACAGACCTACTCCACT
TGGCTACCCTGCAATTAAGAATCATTTAAAAATGTCTGTGGGGGAAGCCATTTTCAGACAA
GACAGGAGAGAAAAAAAAAAAAAAAAAAAAAAAAAAAAA

Fig. 1

CTCGCGGCGGGGATGCTGAGGAGCGCTGGGTCCGGGAGCAGCCCTGGCCCCTGCGGACT
 TCCGAGGCCGTGAAAACCCCTGCGCTGCGGCCCTTCCCAGGCCCCCTATTCTTTTCGCCG
 TTCCCGAAGCCCGACTGGGGGAAGAGTCCAGCACCAAAGCGGGCCGTTCTCGGATTCCGGA
 GCGTTCTGGAGCCCCGAGAGACGCCCCGGGGTTCTAGAAGCTCCCCGGCGGGCGCCAGTC
 CCGGCTTCATTCGGGCGTCCCTCCGAAACCCACTCGGGTGCACGGGTGCTCGGCGAGCCG
 CGACCGGGTCCTGGCGCGCACCATGATCGTGGCGGACTCCGAGTGCCGCGCAGAGCTCAA
 GGACTACCTGCGGTTCCGCCCGGGCGGGCTCGGCGACTCGGGCCCCGGAGAGGAGCAGAG
 GGAGAGCCGGGCTCGGCGAGGCCCTCGAGGGCCCAGCGCCTTCATCCCCGTGGAGGAGGT
 CCTTCGGGAGGGGGCTGAGAGCCTCGAGCAGCACCTGGGGCTGGAGGCACTGATGTCCTC
 TGGGCGAGTAGACAACCTGGCAGTGGTGATGGGCCTGCACCCTGACTACTTTACCAGCTT
 CTGGCNCCTGCACTACCTGCTGCTGCACACGGATGGTCCCTTGGCCAGCTCCTGGCGCCA
 CTACATTGCCATCATGGCTGCCGCCCGCCATCAGTGTTCTTACCTGGTAGGCTCCCACAT
 GGCCGAGTTTCTGCAGACTGGTGGTGACCCTGAGTGGCTGCTGGGCCTCCACCGGGCCCC
 CGAGAAGCTGCGCAAACCTCAGCGAGATCAACAAGTTGCTGGCGCATCGGCCATGGCTCAT
 CACCAAGGAACACATCCAGGCCTTGCTGAAGACCGGGCGAGCACACTTGGTCCCTGGCCGA
 GCTCATTGAGGCTCTGGTCCTGCTCACCCACTGCCACTCGCTCTCCTCCTTCGTGTTTGG
 CTGTGGCATCCTCCCTGAGGGGGATGCAGATGGCAGCCCTGCCCCCAGGCACCTACACC
 CCCGAGTGAACAGAGCAGCCCCCAAGCAGGGACCCGTTGAACAACCTCTGGGGGCTTTGA
 GTCTGCCCGCGACGTGGAGGCGCTGATGGAGCGCATGCAGCAGCTGCAGGAGAGCCTGCT
 GCGGGATGAGGGGACGTCCCAGGAGGAGATGGAGAGCCGCTTTGAGCTGGAGAAGTCAGA
 GAGCCTGCTGGTGACCCCCCTCAGCTGACATCCTGGAGCCCTCTCCACACCCAGACATGCT
 GTGCTTTGTGGAAGACCCTACTTTCCGATATGAGGACTTCACTCGGAGAGGGGCTCAGGC
 ACCCCCTACCTTCCGGGGCCAGGATTATACCTGGGAAGACCATGGCTACTCGCTGATCCA
 GCGGCTTTACCCTGAGGGTGGGCAGCTGCTGGATGAGAAGTTCCAGGCAGCCTATAGCCT
 CACCTACAATACCATCGCCATGCACAGTGGTGTGGACACCTCCGTGCTCCGCAGGGCCAT
 CTGGAACCTATATCCACTGCGTCTTTGGCATCAGATATGATGACTATGATTATGGGGAGGT
 GAACCAGCTCCTGGAGCGGAACCTCAAGGTCTATATCAAGACAGTGGCCTGCTACCCAGA
 GAAGACCACCCGAAGAATGTACAACCTCTTCTGGAGGCACTTCCGCCACTCAGAGAAGGT
 CCACGTGAACTTGCTGCTCCTGGAGGCGCGCATGCAAGCCGCTCTGCTGTACGCCCTCCG
 TGCCATCACCCGCTACATGACCTGACTCCTGAGCAGGACCTGGGCCCCGGTTTCAGCTCCCC
 ACAAGGACTTCTCTGTCTGGAGACAGCCCCAGACCCTTTTGTGTCCCATGCCACCCCTCC
 CCACGCTGCAGTGGGCTTGTGTGTGATGTGCAGTCCCGAAGCCACACCCTCCCTTTTCT
 CACTGGAATGGACAGTTCATTGCACTGACTCTGGGATCTCAGCCCTGCTCCTGGGAGCTG
 GAAGAGCACTTGGAGATCCTAAGGGACCACACCCTTCTCCTTCCCCTGCCACAGAGGC
 AGAGGGCACAGGAAAGAAGCCGGGGCCAAGCTCGGAATTAATGTGCCACAAGTGTTGTGGC
 CTTCTGAACTGGGAAGTCCCTGGCTGGCCCCCGGGGGAGAGGGGCAAATGCCTCCGGGA
 CTGACACTCCAGGCAGCTTTGCCTTCTCTCCCCTGTCAATTTCCAGATTTTCATTACCTCCT
 ACTTGCCATTCACCCATCAATGTGAAAGTCAGGGTCACAGCTGGTCTGTGTGTCCAGTTC
 CCTAAAAGCCTGTTCTGTTGGGCAGCCTGAGGCTGTTGCCCGAATCCTAGTTCAGTTTTT
 TGACTTCCTTTGCCCTTTTTCCCTTTTCTCCATGCTTAATGGTGTGAGGCGTCAGGAGAG
 AGGCCAAGTACATAAAAAAAAAAAAAAGCAGATTATCTCTAGAGAGTTTGAGCCTTTGCT
 GGTCACATTGCCTTCTGAAGAGGAGGGAGTATTAGATTATAAATCCTCTTTATTTTGGTC
 CTTTATGCTTGAGGTTCCAACCTGGAGCCACAGTGTGTGANAGGAGGAGGAGAGGGAGAA
 TTCTGTTCTCCCANAGCTGCACCTGCCTCGCANAGGCCAGCACCCCACTCTCCTGCCTCC
 AGTGGCCCTGCCGCAGATGTCTCCCAAAAAGTTGAGCCTTTCTAGATGGCTTAGGTGGCA
 CCATGGCTCAGCAGGAGGGGCGGGAGGCACCAGGGTTCTTGTTTGGACCCTGCCCTGGG
 CCATGGCCAGGTGACCATGGCTACATTGCCAAACCTCTGACTGCCACAGCTGCAGACTGA
 GAGGGTGGGTCTGAGTCCCCACAATGTCTGAAGCTGCCCTGGGATTCTCAGGCCAACCT

Fig. 2a

GCCAACAGCAAGCGGATTTTCTTGCAAGATCAGGGACCCCATTTCTGCAGCCAGTGTCTC
CTGGGTGCCTTCTGAGGACTCCCACCCCATCCCAGTATCTCATCTGTCCCCTCTCCTGG
GGCTTAAGTGGGTTGCTTCCAGGCAGAAGCAGCCAAGGACCGATTCCAGGCACTTTCTGT
AGCAAATGACTGTGAATTACGACTTCTCTTGCCCTTCTTCTAGCAGTCTGTGCCTCCTCT
CTGACCAGTTTGGAGGGCACTGAAGAAAGGCAAGGGCCGTGCTGCTGCTGGGCGGGGCAG
GAGAGGAGCCTGGCCAGTGTGCCACATTAAATACCCGTGCAGGCGCGGAGAAGCAACCGG
CACCCCTTCCGGCCTGAAAGCCCTCCCTGCAAGAAGGTGTGCAGGAGAGAAGAGGCCCC
GGCATGGGGATCTGGGTTCTAGAGGGCATGTGATGACTGTAAATGTTCACTGGGTGGGTA
GGGAGTGGTATCCAGTGTTCAGTGCAGAAATCTTTGGCTTTGCTACCAGTTCCATATGA
TGAGAAATAAACGTTTCGCTGAGGTTTTGTTTCATAAAAAAAAAAAAAAAAAAAAAAAAAA
AAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA

Fig. 2b

GGCACGAGGACGAGCGGGAGCGCGGAGCAGCAGCCTCTGCTGCCCTGACTTTTTTAAGAAA
 TCTCAATGAACTATTTGTAGAGAATCACTGATCCGGCCTGCAAGCATTTTGCACGGGCAAA
 AATATCGATCAGTGTTAAGTGAAGATCACATTTTATATGCGATCTTGACTTTTTTTGTCTT
 ACATTATATTTTTATAGATTTTGTATAAACATGGTGCTGGGAAAGGTGAAGAGTTTGAC
 AATAAGCTTTGACTGTCTTAATGACAGCAATGTCCCTGTGTATTCTAGTGGGGATACCGT
 CTCAGGAAGGGTAAATTTAGAAGTTACTGGGGAAATCAGAGTAAAATCTCTTAAAATTCA
 TGCAAGAGGACATGCGAAAGTACGCTGGACTGAATCTAGAAACGCCGGCTCCAATACTGC
 CTATACACAGAATTACACTGAAGAAGTAGAGTATTTCAACCATAAAGACATCTTAATTGG
 GCACGAAAGAGATGATGATAATTCCGAAGAAGGCTTCCACACTATTCATTCAGGAAGGCA
 TGAATATGCATTCAGCTTCGAGCTTCCACAGACACCACTCGCTACCTCATTGGAAGGCCG
 ACATGGCAGTGTGCGCTATTGGGTGAAAGCCGAATTGCACAGGCCTTGGCTACTACAGT
 AAAATTAAAGAAGGAATTTACAGTCTTTGAGCATATAGATATCAACACTCCTTCATTACT
 GTCACCCCAAGCAGGCACAAAAGAAAAGACACTCTGTTGCTGGTTCTGTACCTCAGGCC
 AATATCCTTAAGTGCCAAAATTGAAAGGAAGGGCTATACCCAGGTGAATCAATTCAGAT
 ATTTGCTGAGATTGAGAACTGCTCTTCCCGAATGGTGGTGCCAAAGGCAGCCATTTACCA
 AACACAGGCCTTCTATGCCAAAGGGAAAATGAAGGAAGTAAAACAGCTTGTGGCTAACTT
 GCGTGGGGAATCCTTATCATCTGGAAGACAGAGACGTGGAATGGCAAGTTGCTGAAAAT
 TCCACCAGTTTCTCCCTCTATCCTCGACTGTAGTATAATCCGCGTGGAATATTCATAAT
 GGTATATGTGGATATTCCTGGAGCTATGGATTTATTTCTTAATTTGCCACTTGTATCGG
 TACCATTCTCTACATCCATTTGGTAGCAGAACCTCAAGTGTAAGCAGTCAGTGTAGCAT
 GAATATGAACTGGCTCAGTTTATCACTTCTGAAAGACCTGAAGCACCAACCCAGCTATGC
 AGAAGTGGTAACAGAGGAACAAAGGCGGAACAATCTTGCAACAGTGAGTGCTTGTGATGA
 CTTTGAGAGAGCCCTTCAAGGACCACTGTTTGCATATATCCAGGAGTTTCGATTCTTGCC
 TCCACCTCTTTATTCAGAGATTGATCCAAATCCTGATCAGTCAGCAGATGATAGACCATC
 CTGCCCCCTCTCGTTGAAGGAACACTTGGTTGAATCAAGTTGATGTGGGTTCCGAACCTGTA
 TCTCTTCCGGCTGAGGACAGAGAAGTATCTTGGAGACACGTTTCAGAGGAAGTGGAATTA
 CTTTTGCCCAGAAAAATGGCGAATACATGAAACAACCAGTGATCATGCTTTAGAAGCCTA
 CAGCAACATTCTGAGACTGCTCCAACATGCTTGAAGATCTAAGCTTTTCTCTTTTAAAC
 TGGCACATACTCAGAGCAGTCTTCTTAGCCTATGGTCGTACGTGTCAAGACATCACGTTG
 TAAAGAGGGGATGATTTCTTCTTTTGAATTTGAAAATTTGCACATGCTCAATGCTTACATT
 GTGCGGTTTCGACGTCACCTACAGCTTCTTTTTTTTTTTTTTTTTTTTTCTATTTTTGCCAGAC
 TCTTGATACTCTTAAACTTGTGTTGTGGTCAGCACACAAGGAACAAAACAAAGCTTTGA
 AAAAATTTAACATGAAAAACGCACTGACATTTTTTTTTTATTTAATATAGCCTGGACTT
 TACCTGCGTATGCACATGCTCAGAATTGTCTACTAGGCTGACTATGTATCACCTCTTCAG
 CTTGGATCCAATTGTGGATTTATTTACAAACATCAAATGCCTTCAAGCCAATCCTTTTTG
 CTGTATGTTTTGCAGCCTACTGTAGTAGATACGCAACAGATAATGTGGGAAAAAAGAGA
 TAAGAGGAGGAAGCTAATAAGAGACTGTCAAGATTGTATACCTTCTTGGTTTCTTTAAG
 AATTTGTTGCCTTTCTACTATTACAGCAAAGCAGCATTGTTGTTACTGACTGCCTAAAATC
 ACTTAATCTCAGGTGAACGCATCACTTGCCAAACTGTTGGAATGCTATTTGTGTTTTGTT
 GCACTGTTTTTTTCTGTTTGTGTTGTTGTTTATTTGGTTGGCTTTTTTGGAGAGGGAAATTT
 GGAAACGGGACATACAAAAAGTTACACACCCACATTCCCTTTTTTATCATGACATACAAG
 AAGAACTAGCAGAGCTAAGAATGGAGTGAAGAAAGGCAGTATGGCAGGCACCAGCAAAG
 AGTTGAGGGGCTGTTGCTCTTAAAAATTATTTTTTTTATTATTATTTTGAAGTATGGAAG
 TTTTCCATTCACTGGGGAAAGGAGGGAAAAGTGCATTTATTTTTATACAGAGTTACTTAA
 TTACCTCCAAAACACATATGTTGGAAATCGCTTTTGCTGGTGCAAAGTATATTAATGAGC
 AGGAATACATACATTGAGGTTATGAATAGAGAGCTCAATTTGTACCTTTGCTGTCTTGCT
 CAAGCTTGGTATGGCATGAAAACCTCGACTTTATTCCAAAAGTAACTTCAAATTTAAAT
 ACTAGAACGTTTGCTGCGATAAATCTTTTGGATTTTTGTGTTTTTCTAATGAGAATACTG
 TTTTTCATTACCTAAAGAACAATTTGCTAAACATGAGAAATCACTCACTTTGATTATGTA

Fig. 3a

CCTCCTCCCCTCCTCCCGCCCGCCGCCCCGCTCCCCGCCCCCGGSSGCCCCNGCGGAACG
GTTACGCCGCGACGAAGTAAGGGTGGGTTCTNAAGGAAAGCCCTTTGCCAATCTTGCAAG
ATTTGTAGACCAGCACTACAAAGATCGCATAGATCAAATAGGAAAAAAAAAATGTCGATTTT
TATTCAGTCTGATGGTTCTGTTCTTCATTGTGATTGTCATTAAAAAGTGGTAAATTGCTC
AATGTAATATTTTTGTGCGCTGTTTAGAAGTTGTGTGATTTTTTTGCCATCGTTGATAAAA
ATGCAAAGTCAAATAAAAGGTGTCTTGGTTTGATGTCATAGAATGATCCAAGGAGAGAAA
AAAGGTAGTTACTGTTTTACCAGAAAAGGTAATGAGTGAAGGAAAGAATAGTAGCAGAA
AGCACAGTTTGTGAGTAAAGCTGTCTGGAATTAAGTTACCAAAAATACAAAGCAAAAGGA
CTATTATTTTGGGTTGAAGCTCCAAAACCTGACAGCATCTGATAATCTGTTGGTTTATTTT
ACTTTTCATTAAATGAACATTGATGAGAGAAGATGCCACTTACCCAAGCTTTAGAGAATC
CCTAGTGGAAGATTATATGATAAACTTTTCAAGTCCTGACATAACACTAGGGCATTCTCTAGA
GTGTCATTGCTAAAACCTCACTGAACAGACGCAGCCAAGGTCTGTGTTTACGCACTTGGTC
TCTGTTGTTACGTAAAATAATAAGCATTATAAATAGTTTACAGATATTTTTTGACCAGTTC
CTTTTAGAGATTCTTTCAGAGAAGAAACCAGATCTGACCTGTTTATTGTTGGCGCTTGTT
GAAAACGAGCTTTCTTTCCCATGATAGTGCTTCGTTTTTGAAGTGTTGAAGCTGTGCTCC
CCTTAAATCGTGGCAGGAGAGATTAAGGTAATTACAACACTCAGTTCTATGTCTTACAAG
CACTTTGTCTTGTCTCTGCAAGAAAATTGATTCCAGTCATTTCCCATAAAATACAGACA
TTTTACCAACATAATATGCTTTGATTGATGCAGCATTATGCTTTGGGCAGTATTACAAAA
TAGCTGGCGAGTGCTTTCTGTATTTAAATATTGTAAAAATAAAATAAGTTATAACTGTTA
TAAAGCAGAACTTTTGTGTCATTTTTTAACTGTTGAAGTCNCTGTGTATGTTTGTGTTGG
TCAATGTTTCCNCAGTATTTATTAAACATACTTTTTTTTTTTCTTCAAATAAAAAAGTAA
CCATGTCTTTGTCTAAAAAAAAAAAAAAAAANANNNNNAAAAAAAA

Fig. 4

ATGGGGGGGATGCACGGTGAAGCCTGAGCTGCTGCTCCTGGCGCTCGTCCTCCACCCCTGG
AATCCCTGTCTGGGTGCGGACTCGGAGAAGCCCTCGAGCATCCCCACAGATAAATTATTA
GTCATAACTGTAGCAACAAAAGAAAGTGATGGATTCCATCGATTTATGCAGTCAGCCAAA
TATTTCAATTATACTGTGAAGGTCCTTGGTCAAGGAGAAGAATGGAGAGGTGGTGATGGA
ATTAATAGTATTGGAGGGGGCCAGAAAGTGAGATTAATGAAAGAAGTCATGGAACACTAT
GCTGATCAAGATGATCTGGTTGTCATGTTTACTGAATGCTTTGATGTCATATTTGCTGGT
GGTCCAGAAGAACTTCTAAAAAAATTCCAAAAGGCCAACCAAAAGTGGTCTTTGCAGCA
GATGGAATTTTGTGGCCAGATAAAAGACTAGCAGACAAGTATCCTGTTGTGCACATTGGG
AAACGCTATCTGAATTCAGGAGGATTTATTGGCTATGCTCCATATGTCAACCGTATAGTT
CAACAATGGAATCTCCAGGATAATGATGATGATCAGCTCTTTTACACTAAAGTTTACATT
GATCCACTGAAAAGGGGAAGCTATTAACATCACATTGGATCACAAATGCAAAATTTTCCAG
ACCTTAAATGGAGCTGTAGATGAAGTTGTTTTAAATTTGAAAATGGCAAAGCCAGAGCT
AAGAATACATTTTATGAAACATTACCAGTGGCAATTAATGGAAATGGACCCACCAAGATT
CTCCTGAATTATTTTGGAACTATGTACCCAATTCATGGACACAGGATAATGGCTGCACT
CTTTGTGAATTCGATACAGTCGACTTGTCTGCAGTAGATGTCCATCCAAACGTATCAATA
GGTGTTTTTATTGAGCAACCAACCCCTTTTCTACCTCGGTTTTCTGGACATATTGTTGACA
CTGGATTACCCAAAAGAAGCACTTAACTTTTTTATTCATAACAAAGAAGTTTATCATGAA
AAGGACATCAAGGTATTTTTTGTATAAAGCTAAGCATGAAATCAAACTATAAAAATAGTA
GGACCAGAAGAAAATCTAAGTCAAGCGGAAGCCAGAAACATGGGAATGGACTTTTGCCGT
CAGGATGAAAAGTGTGATTATTACTTTAGTGTGGATGCAGATGTTGTTTTGACAAATCCA
AGGACTTTAAAAATTTTGATTGAACAAAACAGAAAGATCATTGCTCCTCTTGTAACCTCGT
CATGGAAAGCTGTGGTCCAATTTCTGGGGAGCATTGAGTCCTGATGGATACTATGCACGA
TCTGAAGATTATGTGGATATTGTTCAAGGGAATAGAGTAGGAGTATGGAATGTCCCATAT
ATGGCTAATGTGTACTTAATTAAGGAAAGACACTCCGATCAGAGATGAATGAAAGGAAC
TATTTTGTTCGTGATAAACTGGATCCTGATATGGCTCTTTGCCGAAATGCTAGAGAAATG
ACTTTAVAAAGGGAAAAAGACTCCCCTACTCCGGAAACATTCCAAATGCTCAGCCCCCA
AAGGGTGTATTTATGTACATTTCTAATAGACATGAATTTGGAAGGCTATTATCCACTGCT
AATTACAATACTTCCCATTATAACAATGACCTCTGGCAGATTTTTGAAAATCCTGTGGAC
TGGAAGGAAAAGTATATAAACCGTGATTATTCAAAGATTTTCACTGAAAATATAGTTGAA
CAGCCCTGTCCAGATGTCTTTTGGTTCCCATATTTTCTGAAAAAGCCTGTGATGAATTG
GTAGAAGAAATGGAACATTACGGCAAATGGTCTGGGGGAAAACATCATGATAGCCGTATA
TCTGGTGGTTATGAAAATGTCCCAACTGATGATATCCACATGAAGCAAGTTGATCTGGAG
AATGTATGGCTTCATTTTATCCGGGAGTTCATTGCACCAGTTACACTGAAGGTCTTTGCA
GGCTATTATACGAAGGGATTTGCACTACTGAATTTTGTAGTAAAATACTCCCCTGAACGA
CAGCGTTCTCTTCGTCCTCATCATGATGCTTCTACATTTACCATAAACATTGCACTTAAT
AACGTGGGAGAAGACTTTCAGGGAGGTGGTTGCAAATTTCTAAGGTACAATTGCTCTATT
GAGTCACCACGAAAAGGCTGGAGCTTCATGCATCCTGGGAGACTCACACATTTGCATGAA
GGACTTCCTGTAAAAAATGGAACAAGATACATTGCAGTGTCAATTTATAGATCCCTAAGTT
ATTTACTTTTCATTGAATTGAAATTTATTTTGGATGAATGACTGGCATGAACACGTCCTT
GAAGTTGTGGCTGAGAAGATGAGAGGAATATTTAAATAACATCAACAGAACAACCTTCACT
TTGGGCCAAACATTTGAAAAACTTTTTATAAAAAATTGTTTGATATTTCTTAATGTCTGC
TCTGAGCCTTAAACACAG

Fig. 5a

ATGGGGGGGATGCACGGTGAAGCCTCAGCTGCTGCTCCTGGCGCTCGTCCTCCAGGGGTGG
 AATCCCTGTCTGGGTGCGGACTCGGAGAAGCCCTCGAGCATCCCCACAGATAAATTATTA
 GTCATAACTGTAGCAACAAAAGAAAGTGATGGATTCCATCGATTTATGCAGTCAGCCAAA
 TATTTCAATTATACTGTGAAGGTCCTTGGTCAAGGAGAAGAATGGAGAGGTGGTGATGGA
 ATTAATAGTATTGGAGGGGGGCCAGAAAGTGAGATTAATGAAAGAAGTCATGGAACACTAT
 GCTGATCAAGATGATCTGGTTGTCATGTTTACTGAATGCTTTGATGTCATATTTGCTGGT
 GGTCCAGAAGAAGTTCTAAAAAAATTCCAAAAGGCCAACCAAAAGTGGTCTTTGCAGCA
 GATGGAATTTTGTGGCCAGATAAAAGACTAGCAGACAAGTATCCTGTTGTGCACATTGGG
 AAACGCTATCTGAATTCAGGAGGATTTATTGGCTATGCTCCATATGTCAACCGTATAGTT
 CAACAATGGAATCTCCAGGATAATGATGATGATCAGCTCTTTTACACTAAAGTTTACATT
 GATCCACTGAAAAGGGAAGCTATTAACATCACATTGGATCACAAATGCAAAATTTTCCAG
 ACCTTAAATGGAGCTGTAGATGAAGTTGTTTTAAAATTTGAAAATGGCAAAGCCAGAGCT
 AAGAATACATTTTATGAAACATTACCAGTGGCAATTAATGGAAATGGACCCACCAAGATT
 CTCCTGAATTATTTTGGAACTATGTACCCAATTCATGGACACAGGATAATGGCTGCACT
 CTTTGTGAATTCGATACAGTCGACTTGTCTGCAGTAGATGTCCATCCAAACGTATCAATA
 GGTGTTTTTTATTGAGCAACCAACCCCTTTTCTACCTCGGTTTCTGGACATATTGTTGACA
 CTGGATTACCCAAAAGAAGCACTTAACTTTTTTATTCATAACAAAGAAGTTTATCATGAA
 AAGGACATCAAGGTATTTTTTGTATAAGCTAAGCATGAAATCAAACTATAAAAATAGTA
 GGACCAGAAGAAAATCTAAGTCAAGCGGAAGCCAGAAACATGGGAATGGACTTTTGCCGT
 CAGGATGAAAAGTGTGATTATTACTTTAGTGTGGATGCAGATGTTGTTTTGACAAATCCA
 AGGACTTTAAAAATTTTGATTGAACAAAACAGAAAGATCATTGCTCCTCTTGTAACCTCGT
 CATGGAAAGCTGTGGTCCAATTTCTGGGGAGCATTGAGTCCTGATGGATACTATGCACGA
 TCTGAAGATTATGTGGATATTGTTCAAGGGAATAGAGTAGGAGTATGGAATGTCCCATAT
 ATGGCTAATGTGTACTTAATTAAGGAAAGACACTCCGATCAGAGATGAATGAAAGGAAC
 TATTTTGTTCGTGATAAACTGGATCCTGATATGGCTCTTTGCCGAAATGCTAGAGAAATG
 GGTGTATTTATGTACATTTCTAATAGACATGAATTTGGAAGGCTATTATCCACTGCTAAT
 TACAATACTTCCCATTATAACAATGACCTCTGGCAGATTTTTGAAAATCCTGTGGACTGG
 AAGGAAAAGTATATAAACCGTGATTATTCAAAGATTTTCACTGAAAATATAGTTGAACAG
 CCCTGTCCAGATGTCTTTTGGTTCCCATATTTTCTGAAAAAGCCTGTGATGAATTGGTA
 GAAGAAATGGAACATTACGGCAAATGGTCTGGGGGAAACATCATGATAGCCGTATATCT
 GGTGGTTATGAAAATGTCCCAACTGATGATATCCACATGAAGCAAGTTGATCTGGAGAAT
 GTATGGCTTGATTTTATCCGGGAGTTCATTGCACCAGTTACACTGAAGGTCTTTGCAGGC
 TATTATACGAAGGGATTTGCACTACTGAATTTTGTAGTAAAATACTCCCTGAACGACAG
 CGTTCTCTTCGTCTCATCATGATGCTTCTACATTTACCATAAACATTGCACTTAATAAC
 GTGGGAGAAGACTTTCAGGGAGGTGGTTGCAAATTTCTAAGGTACAATTGCTCTATTGAG
 TCACCACGAAAAGGCTGGAGCTTCATGCATCCTGGGAGACTCACACATTTGCATGAAGGA
 CTTCTGTGTTAAAAATGGAACAAGATACATTGCAGTGTCATTTATAGATCCCTAAGTTATT
 TACTTTTCATTGAATTGAAATTTATTTTGGGTGAATGACTGGCATGAACACGTCTTTGAA
 GTTGTGGCTGAGAAGATGAGAGGAATATTTAAATAACATCAACAGAACAACCTTCACTTTG
 GGCCAAACATTTGAAAAACTTTTTATAAAAAATTGTTTGATATTTCTTAATGTCTGCTCT
 GAGCCTTAAACACAGATTGAAGAAGAAAAGAAAAGAAAACCTTAAATATTTATTTCTAT
 GCTTTGTTGCCCTCTGAGAATAATGACAATTTATGAATTTGTGTTTCAAATTGATAAAATA
 TTTAGGTACAAATAACAAGACTAATAATATTTTCTTATTTAAAAAAAGCATGGGAAGATT
 TTTATTTSTCSSSSTSTSGSGSSSTGTSGSCSSSSTGGSTSTSSSTGSSSSTTSCCSTG
 TTGTAAAACCTTGAAAATCAGATTCTAACTGATTGTATGCAACTAAGTATTTCTGAACAC
 CTATGCAGGTCTTATTTACAGTGTTACTAAGGGAACACACAAAGAATTACACAACGTTTT
 CCTCAAGAAAATGGTACAAAACACAACCGAGGAGCGTATACAGTTGAAAACATTTTTGTT
 TTGATTGGAAGGCAGATTATTTTATATTAGTATTAATAATCAAACCCTATGTTTCTTTCA
 GATGAATCTTCAAAGTGGATTATATTAAGCAGGTATTAGATTTAGAAAACCTTTCCATT
 TCTTAAAGTATTATCAAGTGTCAAGATCAGCAAGTGTCTTAAGTCAAATAGGTTTTTTT
 TTGTTGGTGGTTGTGCTTGCTTTTCTTTTTTAAAGTTCTAGAAAATAGGAAAACGAAA
 AATTCATTGAGATGAGTAGTGCATTTAATTATTTTTTAAAAAACTTTTTAAGTACTTGA

Fig. 5b-a

ATTTTATATCAGGAAAACAAAGTTGTTGAGCCTTGCTTCTTCCGTTTTGCCCTTTGTCTC
 GCTCCTTATTCTTTTTTTGGGGGGAGGGTTATTTGCTTTTTTATCTTCCTGGCATAATTTT
 CATTTTATTCTTCTGAGTGTCTATGTTAACTTCCCTCTATCCCGCTTATAAAAAAATTCT
 CCAACAAAAATACTTGTTGACTTGATGTTTTATCACTTCTCTAAGTAAGGTTGAAATATC
 CTTATTGTAGCTACTGTTTTTAATGTAAAGGTTAACTTGAAAAGAAATTCTTAATCACG
 GTGCCAAAATTCATTTTCTAACACCATGTGTTAGAAAATTATAAAAAATAAAATAATTTT
 AAAAAAAAAAAAAAAAAAAAAA

Fig. 5b-b

GGCTGCGAGAAGACGACAGAAGGGGTCCGTCTGCTCGGTGCGCTCGGGCTCCGCGCT
 AGTCCGCTCAGTGTTCTCCAATCGCTTTGGTACCCACGCAGTCCTCTCATCCGTCCTCCG
 CTGCCGTCCCGGGGCCCCACGTCTAACCCGGTGCTCTTCGGGGTCTCCGCGTCTCGCGAGA
 AGTCCTCGCCGCAGGCCTCGGGCTTTCGGGGCTTAGGGGGCGGATGGGGGACCGCGGAGTGA
 GGCTGGGGCTGCTGATGCCCATGCTCGCCCTGCTCTCCTGGGCGGGCTAGCCTGGGCGTAG
 CGGAGGAGACTCCCTCGCGCATCCACAGCAGATAAGTTATTAGTCATAACTGTAGCAACCA
 AAGAAAACGATGGATTCCACAGATTTATGAATTCAGCCAAGTATTTCAATTATACTGTGA
 AGGTTCTTGGTCAAGGGCAAGAGTGGAGAGGTGGTGATGGGATGAACAGTATTGGAGGGG
 GCCAGAAGGTGAGATTAATGAAAGAAGCCATGGAGCACTACGCCGGTCAGGACGATCTGG
 TCATCTTGTTTACTGAATGTTTTGATGTTATATTTGCTGGTGGGCCTGAAGAACTTCTTA
 AAAAGTTCCAAAAGACAAATCATAAAATCGTCTTTGCAGCGGATGCGCTGTTGTGGCCAG
 ATAAGCGGCTGGCAGACAAGTATCCTGGTGTGCACATTGGGAAACGCTACCTGAATTCTG
 GAGGCTTTATTGGCTATGCTCCCTACATCAGCCGTCTGGTCCAGCAGTGGGATCTGCAGG
 ATAATGATGACGACCAGCTCTTTTACACTAAAGTTTACATCGACCCGCTGAAAAGGGGAAG
 CTCTTAACATCACATTGGATCACAGATGCAAAATTTTCCAGGCCTTGAATGGAGCTACAG
 ACGAAGTTGTTTTAAAGTTTGAATGGTAAAAGCAGAGTGAAGAATACATTTTATGAAA
 CACTGCCAGTGGCCATCAATGGGAATGGGCCACCAAAATTCTCTTGAATTACTTTGGAA
 ACTATGTTCCAAATTCATGGACACAGGAAAATGGCTGTGCTCTTTGTGACTTTGACACAA
 TTGACCTGTCTACAGTAGATGTCTATCCGAAGGTAACACTAGGTGTTTTTATTGAACAAC
 CAACCCCTTTCTACCTCGGTTCTTGACTTACTGTTAACACTGGATTACCCTAAAGAAG
 CACTTCGACTCTTTGTCCATAATAAGAAGTTTATCATGAAAAGGACATCAAAGCGTTTG
 TTGATAAAGCTAAACACGACATCAGCTCTATAAAAATAGTAGGACCAGAGGAAAATCTAA
 GTCAAGCGGAAGCCAGAAACATGGGAATGGATTTCTGCCGTGAGGATGAAAAGTGTGATT
 ACTACTTTAGTGTGGATGCAGATGTTGTTTTGACAAACCCAAGAACTTTAAAAATTTTGA
 TTGAACAAAACAGGAAGATCATTGCCCTCTTGTGACACGTCATGGAAAGTTGTGGTCCA
 ACTTCTGGGGAGCCCTGAGTCCTGATGGATACTATGCTCGTTCTGAAGATTACGTAGATA
 TCGTTCAGGGAAACAGAGTAGGAATATGGAATGTCCCATACATGGCTAATGTGTACTTAA
 TTCAAGGGAAGACGCTGCGATCAGAGATGAGTGAAAGGAACTATTTTGTGCGTGATAAGT
 TGGATCCCGACATGTCTCTCTGCCGCAATGCTCGAGACATGACCTTACAAAGGGGAAAAAG
 ACTCCCCACTCCGGAAACATTCCAAATGCTCAGCCCCCAAAGGGTGTGTTTATGTACA
 TTTCTAACAGACATGAATTTGGACGGCTGATATCAACTGCTAATTACAACACTTCCCATC
 TCAACAATGACCTCTGGCAGATCTTTGAAAATCCCGTGGATTGGAAGGAAAAATATATAA
 ACCGTGACTATTCAAAGATTTTCACTGAAAATATAGTCGAGCAGCCCTGTCCAGATGTCT
 TCTGGTTTCCCATATTTTCTGAACGAGCCTGTGACGAGTTGGTAGAAGAAATGGAACATT
 ACGGCAAGTGGTCCGGGGGAAAGCATCATGACAGCCGTATATCTGGTGGCTATGAAAATC
 TCCAACGGATGACATTCATATGAAGCAGATTGACCTGGAGAACGTCTGGCTTCACTTTA
 TCCGAGAGTTTATCGCTCCAGTTACCCTGAAGGTCTTCGCGGGGATATTACACCAAGGGAT
 TTGCCCTGCTGAACTTCGTAGTGAAGTACTCGCCCGAAAGACAGCGCTCGCTCCGGCCTC
 ACCACGATGCGTCAACCTTCACCATCAACATTGCTCTAAATAATGTAGGAGAGGATTTTC
 AGGGAGGTGGATGCAAATTCCTAAGGTATAATTGCTCCATCGAATCCCCCGAAAAGGCT

Fig. 5c-a

GGAGCTTCATGCATCCTGGGAGGCTTACTCATCTACACGAAGGGCTTCCTGTCAAAAATG
 GAACAAGATACATTGCAGTCTCATTTATCGATCCCTAAGTTATTGACTGAACTTAAACTG
 AGTGGCTCTTTGAGATGGATGACTGGCGGGAACATGTCTCTGAAGTTGTACTTGAGAAGA
 CGAGAGGAATATTTAAATAATGTCACCAGAACACGTCACCTTTGGGCCAAGCATTGAAA
 ACTTTTTATATAAATTTGTTTTATGTTTCTTAACGTCTGCTCTGAGCCTTAAACACAGG
 TTGAAGAAGAAGAGAGAGGAAAAAAGTGAAAGTTGGTATTTATTTCTGTGCTTTAATTGT
 CTATGAAAATGATGACATTTTATAAAATGTTTAGGTACAAAGGCATGAATGATAATCAGT
 AAGCCTAATAATATTTTCTTATTTAAGGAGAACCTGAGAAGATTTTATTTTTCAGTGGGA
 GAAATATGGAAAATGGTTCTAAATGAGGGTCCGGCAGCTCTGGAAGCCCGGGATTCTGACG
 CGTACTGAATTTATGTGTAACCTTTTAAGCCATGCTGACCTCCGGGTAGATTGCTTTTCA
 GTGATAAGGAAGAAAACCCAAAGAAAATATTGCACAGAGGCTTTCTCAAGCAGCCTGGG
 CAGATGGCCAGTGGAAGCCCATCCACTGGAGATCCTCAGCTTGTGAGGCAGGTGCTCCTG
 TCCGTTGGAACTGGGCCCTGTGTGTCTCCAGGGCAAGCTCTCAGGGGAAGCTCACATC
 TGCCTGCTTTACAGAGTGCTTCAGGCGTCAGCTCCAAGTCAAACAGGATGTGTTTCCTTC
 TGTTTTTCCCCTCTAATTATAGAAAATAGTAAGGAAAAATATCAGTTTCATTGAGATTAG
 TAGTACATTTTACTATCTTCTTTTTTAACGATTAAGTACTTGAATTTTATATCAGGAAAA
 TAGTTTTTGAGCCTGTTCTTACCTTTGGCCGTAGTTGGTAGTTGGTCTCTTTGTTTTTCC
 TGGAGGAGGGGCATTTCTTTTCTCATATAAATACTTTCTCATTCTTAGTCTTGTTAT
 TACTTTTCTCTACCCCACTTTTTAAAAATTCCACAGCAAAATTTTATTTGAATTTT
 AATATTTCTCTGAATGAGGTTTAAATATCTTTATTAGAGCTACTGTTTTTAATTTAAAGG
 TTAACTTGAAGAAAGTCTTTATTCATGGTGCCAAAATGCATTTTCTAACTCTGTGTGT
 TAGAAAATAATGAAAAATAAAATAACTTACAATAAAAAAAAAAAAAAAAAAAAAAAAAA

Fig. 5c-b

CGGCCCTTCCCTTGCCACATCTGCGGCAAGGGCTTCATCACGCTCAGCAACCTCTCCAGG
 CACCTGAAGCTGCACCGGGGCATGGACTGACTGCCAGGCTGCGTGCGCCCTGCCCTCCAC
 CCAGCCTCCTGGACTCGGCCTGGACCAGGGGACCTCGGGACTGCGCGTGAGGCCCGGGCC
 CTCCAAATCCAAATCCAGACGCAGGCCCTGAAATGAGGGGACCCTGACTGGAGAGGTGGG
 GGCCACCAAAAACCCACAAAGGCCCGGAGCTGGGGGACCACAAACAGGGTCTTA
 GCTGGGGCAGGGGAGCCCAATCTAGGGAGAGACTCCTGAGCCTGAGGTCCCTGGAATGA
 GTGTGGGTAGCCGTAAGTCCCCAAGACATGGGGACTTTGCAGTGAGCAATGGGTCTCCAC
 AAGTACCTCTCATCTTGAGAGCCCTAATACTAAAAGATGGGCAACCCACCCCAACAGGG
 AAGACTGCCCCATTCCCTGAGAGCCATCATTCTAACGACCTTGATCTGGAGAATGTGGA
 GGGAGCATGTCCCTGAATTTTCTAGATCCCTCCAAATGCCACCCACCAGAGTCACTGGT
 GACCCAGAAAATGGATATAGCCGAAATCTGCCTTTCCCTTTTTTATTCCCTGTGCTGA
 AAGAGGGACCAGGGTAGATGCCCCCTGCCCTCGAATCCCCCTCCCGACTGTGGAATGG
 ATCGACCCTAACGATCTTCCCGCCCCCAACACTAGAATAGACTGGCCTGAAATCCCCTT
 GCCCAGTAGAATGGACTGATCTATGTGCACACACCCCATCACATGGAATGGGCTGGTCT
 AGGCTGTGGCCTGCCACCTTCTTAGAGTGAATAGGGGGGACACTCCTTTTTTTTTCTG
 TAGGGTGTGGGCCGGTCCACGCAATTTTTTATCCTGTGAACTCATTTGAGTGGGAGGTGG
 TGGACACCTGGGGTTTCTTNCCTNTTTCGGTAGCATCCGTTGGTNTTTTTTTCATNTN
 TGTTGGNTTGTNTGTTTNTGTTTTCTCCCNATCCCTAGGGGAAGGGGGCNTTTGGNTAG
 GGGGNGNCCCTGTNAGCCTNGNCCTTGCCCCCTGTNCCTTTNCCAGTGTTTNCAGGNC
 CCCCNATNAACCTTGTCCTGTCAGTNGNTGAAAAAAAAAAAAAAAAAAAAAAAAAAAAA
 AAAAAAA

Fig. 6

TTTCTTTTTTCTTTCGGCGTCTGCGGTGCTCGGAAGTGTGGTACTTCTCCTAGTTGCAG
TCAGGCTTCATACGCTATTGTCCTGCCCGTTAGAGCAGCCAGCGGGTACAGAATGGATTT
TGGAAGAGGGAGTCACCACTGGACCTCCAAGGAAGCCACGTGCAGACATCTACAACCTTC
GATCTCCTGACGAGTTTATTGTTGGCCAAAACCAGGCTTTGATTGAACCAGGATGAATGC
GGGTGTTGGAAGTAGAATATATATATACATATAAAATTGGTTGGGAGCCACGTGTACCAG
TGTGTGTTGATCTTGGCTTGATTCACTCTGCCTTGTAACAGAACTGGCGATGGAATATG
AGAGGAGCCCTCTGGAAAGAAAAGGACAGACCCTGTGCTTTCATGAAAGTGAAGATCTGG
CTGAACCAGTTCCACAAGGTTACTGTATACATAGCCTGAGTTTAAAAGGCTGTGCCCACT
TCAAGAATGTCATTGTTAGACTTTGAAATTTCTAACTGCCTACCTGCATAAAGAAAATAA
AATCTTTTAAATCAAAAAAAAAAAAAAAAAAAAAAAAAAAAAA

Fig. 7a

GGCAGAGGGAGTGTGGTACTTCTCCTAGTTGCAGTCAGGCTTTATACGCTGTTGTCCTG
CCCGTTAGAGCAGCCAGCGGGTACAGAATGGATTTTGGAAGAGGGAGTCACCACTGGACC
TCCAAGGAAGCCACGTGCAGACATCTACAACCTTCGATCTCCTGACGAGTTTATTGTTGG
CCAAAACCAGGCTTTGATTGAACCAGGATGAATGCGGGTGTGGAAGTAGAATATATATA
TACATATAAAATTGAACTGGCGATGGAATATGAGAGGAGCCCTCTGGAAAGAAAAGGAC
AGACCCTGTGCTTTCATGAAAGTGAAGATCTGGCTGAACCAGTTCCACAAGGTTACTGTA
TACATAGCCTGAGTTTAAAAGGCTGTGCCCACTTCAAGAATGTCATTGTTAGACTTTGAA
ATTTCTAACTGCCTACCTGCATAAAGAAAATAAAATCTTTTAAATCAAAAAAAAAAAAAAAAA
AAAAAA

Fig. 7b

GGCACGAGAGGCACAGCAGCAGCTCTATGCAGACAAGTGAAGTGTAGAACTGATTACTG
CTCCACCAAGAAGCCCCATAAGAGTGGTTATCCTGGACACAGAAGTGTGGAATTGAAAT
CCACAGAGCATTTTACAAGAGTTCTGACCTGGATGGGGTAAACCTCAGTGCACCTTCTTTT
CTGTTGGCCTCAGTATTACTGGATTGAAGAATTGCTGCTTCTTGTTAGGAGGTTCAATTC
ACTTATCATTACTTACAACCTTCATACTCAAAGCACTGAGAATTTCAAGTGGAGTATATTG
AAGTAGACTTCAGTTTCTTTGCATCATTCTGTATTCAATTTTTTTAATTATTTTCATAAC
CCTATTGAGTGTTTTTTAACTAAATTAACATGGCTCGAATGAACCGCCCAGCTCCTGTGG
AAGTCACATACAAGAACATGAGATTTCTTATTACACACAATCCAACCAATGCGACCTTAA
ACAAATTTATAGAGGAACTTAAGAAGTATGGAGTTACCACAATAGTAAGAGTATGTGAAG
CAACTTATGACACTACTCTTGTGGAGAAAGAAGGTATCCATGTTCTTGATTGGCCTTTTG
ATGATGGTGCACCACCATCCAACCAGATTGTTGATGACTGGGTAAAGTCTTGTGAAAATTA
AGTTTCGTGAAGAACCTGGTTGTTGTATTGCTGTTTATTGCGTTGCAGGCCTTGGGAGAG
CTCCAGTACTTGTTGCCCTAGCATTAAATTGAAGGTGGAATGAAATACGAAGATGCAGTAC
AATTCATAAGACAAAAGCGGCGTGGAGCTTTTAACAGCAAGCAACTTCTGTATTTGGAGA
AGTATCGTCCTAAAATGCGGCTGCGTTTCAAAGATTCCAACGGTCATAGAAACAACCTGTT
GCATTCAATAAAATTGGGGTGCCTAATGCTACTGGAAGTGGAAGTTGAGATAGGGCCTAA
TTTGTTATACATATTAGCCAACATGTTGGCTTAGTAAGTCTAATGAAGCTTCCATAGGAG
TATTGAAAGGCAGTTTTACCAGGCCTCAAGCTAGACAGATTTGGCAACCTCTGTATTTGG
GTTACAGTCAACCTATTTGGATACTTGGCAAAAGATTCTTGCTGTCAGCATATAAAATGT
GCTTGTCAATTTGTATCAATTGACCTTTCCCAAATCATGCAGTATTGAGTTATGACTTGT
TAAATCTATTCCCATGCCAGAATCTTATCAATACATAAGAAATTTAGGAAGATTAGGTGC
CAAAATACCCAGCACAACTTGTATATTTTTAGTACCATACAGAAGTAAAATCCCAGGA
ACTATGAACACTAGACCTTATGTGGTTTATTCCTTCAATCATTTCAAACATTGAAAGTAG
GGCCTACATGGTTATTTGCCTGCTCACTTTATGTTTACATCTCCACATTTCATACCAATA
TACGTCAGGTTTGCTTAACCATGATTTTTTTTTTTTTTTTACCAAGTCTTACAGTGATTAT
TTTACGTGTTTCCATGTATCTCACTTTGTGCTGTATTAAAAAAACCTCCATTTTGAAAT
CTACGTTGTACAGAAGCACATGTCTTTAATGTCTTCAGACAAAAAAGCCTTACATTAATT
TAATGTTTGCACCTCTGAGGTGCACTTAACAGGGAGGGCCTGAGAAAAGAATGGGAGGGG
GCTATTAATTATTTTTAGCAAAATGTTGCCTTTGTCTTGTCGCAACATGTAGAATATGCT
CTTTAATTTAGTAAAATATTTTTTTTAAAAGGTAGAGATGCTTTGGTATTGGTATCATAAA
CTTCCTGAAATTCTTGAATTTTTTTTCCCATACTATCAAGAAGTGTGTTTACCACTTATTT
TTGTTTGAAAGTGTGATTTTTTTTTTCTTCCCAACCTCTCCTTGCAAAAAAAGAAATGG
GTTTCTGCTAATGAATTGAGCAGACATCTAATATTTTATATGCCTTTTGAGCTGTGTAAC
TTAATATTTGGATACTTGACAATTTGTTTTATTATGTAATTGATAAAATGGTGATGTGTA
TTAATGTTAGTTCAACCATATATTTATACTGTCTGGGGATGTGTGGTTATAGTTCTGTGG
GAGAAATAATTTTGTGAGTGTTCCAGCCTTGTA AAAACTTAGTGCGAGAGCTGAAACAT
CTAAATAAATAATGACATGCATTTATCATCATTGAGATTGGTTTGCTTAAAATTAACCTA
TTTTGTAGAAGACAAAATGAATTGCACTTCACTTAATGTGTGTCCTCATCTTTTTTACAAA
TAAATGAAGGATTATAAATGATGTCAGCATTTTAGTAACTTTTAGACAAAATTTGTTAG
GGTCATTCATGAAAACCTTTAATACTAAAAGCACTTTCCATTATATACTTTTTTAAAGGTCT
AGATAATTTTGAACCAATTTATTATTGTGTACTGAGGAGAAATAATGTATAGTAGAGGAC
AGCCTTGGTTTGTAAGCTCAGTTCCACTAGTTTATGGTTTTGTGCAACTTTTGAGCCTC
AGTTTTCTCCTTTGCAAATTAATAATTACATACCTTTATAGATTTTGAAATTAATTTAAA
TATTAGTATTTGGACATGAAGGCTTAATGTTAAGTTTCCTTTAATGATCCACAATAATCC
CTTTGATCACGTTAATCTAAATCTAGATGTCTTTGTCTAATTTTTTTTTGAATAGCAGTTA
TAAATGTAAAGGACTCAAAGTTTAAGTAAAAAGTGATACTCCACCTTGTTGTTTCAAAGAA
TTTAGTTCCACCTCTTCATACCAGTTTAACTTAATATATTTTATTGGATTTTTAGACAG
GGCAAAAGGAAGAACAGGGGCTCTGGAGGCCCTTGGTTATTTAAATCTTGGATTATTTG
TGATAGTAATCACAAATTTTTGGCTAATTTTTAACCTGAGGTTTTGTTTTTTTTTTTAAA
GGAAATGCAGCCTAGTCTTGAGAACATAATTTTATATAATCAATTACTAAATGTTAACT
ATTACCACACAGCCCATAAAACAGCATTGCGTTTATTGAGAGAGAGGATGTGCCATCAT

Fig. 8a

GATTAATGAAAACCTATCTTTTGAGTTTGAAAAGAAATTAATTTGCAGTGTTTGGATTGTA
TATATGGTGCTAAAAATAAATTAATTTACTTTATAAACCTTATCTGTACATTATACGATG
TGATGAAATTTGCTTTTTTATCCAAATATTTTGTATCTTGTAATATGGCTAATTATAGGA
ATGCCTATAATACATCTTAGATTCCTTATATCTAATAAGAGTTCAAAGAGTTATGAGTTG
AAGTCTTGAATGCAGGAACTATCTGATAGTGTTCTAAAATTTGGTTACTTGGGTTTGGA
TACCCTTAGTGGGATGATGTAAATAGAGGCTAGCTACCTAGGCTTGTCTATAGCAACCAT
AATGTTGATGTAAGTAATGCGGTTACTGAATCATAAGAAAATGCCATCTCTTTTGTAGTTG
AAGGAAAACCTCTGGAAGTAGGTGCCATTGGTCATTCTGCAGTGCAGTGCACCATTTGTTT
CCCCTAGTGCCCTCTTTTCCCTAGGGCATTGCTCTCCTATTCCCACGCCCTTAACACAGCT
CTATACCTAGAAGCAGCCAGCCAGGCATGCAGTCACATTTAATCACATCCCCCTTCTAG
AGTGCTTCAAATGATGTAGTCCCTCAACTTGGCTAAAGAATCTCAATCTCTTGAAATTT
ATTTTTTTAATGTCATATTCATCTGGTAAATATCTACTGTTTGCCAGGCATTTAAGAATA
TGGCAAAGAACATAAAAGATGGTGTCAACAGATTTTGGTCACCAATGAGTACCCGACCCG
TTGCCATGATTAAGAGAGAATGCTTTCTATTGGAGTTTCAGGAAATATAATTTGAGAATA
CTTTAAAGGGAAGTGGAAGTATAAGTGAATGATATTTTCTTTTACATGTAAACAATGAA
GTTATTTCAAAGTTAAGTTTTAAACAAAATCCATGAAGTAGTGTCTGCCATACATGTTAA
TATTCTACATTCTTGCTTCCCTTAAATTAATATGTTTGTGTGTATATATGTGCCTCACAC
CTGAATTGAAAATTAAAGACTGGTTTAAAAGTGGTTTAAAAGTGACATTTAATGTTTCTC
CATTACGTTTGGGGTAACCAGCCTAAGTGGAATCTTGGAAGGAAAGTAAGGGAAAACTT
GTATTTGCCTTCAATGAATTAAACCAGTGATATGTTTAAACGTATGAATGAAAGGATTGAT
GGTGATTTTATAATTATATATATTGCCGCAGTAACCAGTTAATAAATTGATAGCTACCAT
TTAAAAAAAAAAAAAAAAAAAAA

Fig. 8b

GCCAAGAATTCGGGCACGAGGAGTGGGTATCTGAGTTAGTCAGTTTTTACCATTTATAAAAT
GTTTGGTGGAGGAAATTGAACTAATTGCTAAATTGTTAGTAACCAGTGCATTAAC TAGG
ACCCTACTGAGTGGACTGAAAGAATCGAAAATGTTTAACTGGTTGAGAGGCAATGATGTT
GCAAATGGGGTATTCTTCAAAGCTCCTTCTGGGGGGGAAATCTTCAAAGGCAATTATTCTG
AATGTAACTACAGACCAAATTGCAGTCTTCTGTAAGCATTTTCAGAGATTSCCTCAAATA
TTTTTTGATTAAAAAACTCTTCCGTGGTCTTTTGTGCTTCAGAACTACCCAGTACAACAG
GGTCTTCAGCCTGCTCAGGATCTCTAAAGAGAGCTAGCACACAGTCAGCCAAC TTTGGCT
GCTTCAACTCCTAGGAACAAGAAATGATGCTGAGATAATTTGTCTGGCAGGTATTATCAG
CCCACAATGACTGCTGTCATTTAGCCTCAAATGTTTATTTTTTTTTTTTACAATGCTGTA
TTTCTTTAGAACCTTCCTATTCCGAGTGTGGACCCTAGGCCAGCCCCATAGACTTCCCCT
GGGGACTTGTCAGAAATGCATAATTTTAGGCCCCACCCAGACCTGTTGGACCAGAATCT
TCATTTAACAAGATGCCCAGGTGATTCATTCATGTTTGAGAAGCTCTGCTTTAAATCACT
AAAGCAGTTACTGAGTAATTACTACCATCATGACTCTGAAGAGCTCCTATAGCCTTCAA
TGCACCTAACTCTACTCTAAAGGCAAATGTCCTCACTGGGAAATCTGATCTGCTGTTTCA
GAGAAGTGCAGGGCTACACAGTGTCTTACACTCCTATCTATTGATGTTTCTTGGTTTTGC
CTGGTAATCTGCTGCTTAAATGGATTATTTGATGACATATTGATATTA AACAGTCCTAT
TTTAGAAAAA

Fig. 9