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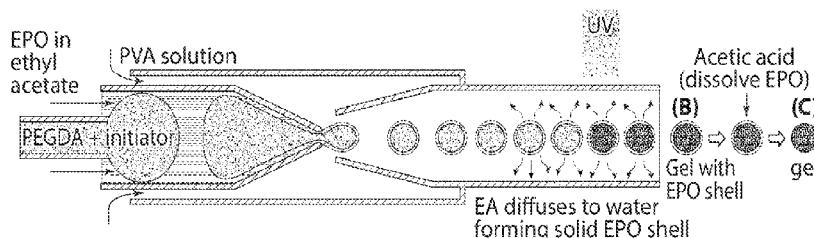


FIG. 1A

(57) Abstract: The present invention generally relates to microfluidic droplets and, in particular, to multiple emulsion microfluidic droplets. In certain aspects, particles such as gel particles can be prepared in an aqueous carrier from aqueous droplets (or a non-aqueous carrier from non-aqueous droplets). For example, in some embodiments, double-emulsion droplets of a first fluid, surrounded by a second fluid, contained in a carrier fluid may be prepared, where the first fluid forms a gel and the second fluid is removed. For instance, the second fluid may be dissolved in the carrier fluid, or the second fluid may be hardened, then removed, for example, due to a change in pH. Other embodiments of the present invention are generally directed to kits containing such microfluidic droplets, microfluidic devices for making such microfluidic droplets, or the like.

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SYSTEMS AND METHODS FOR MAKING AND USING GEL MICROSPHERES

RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Patent Application Serial No. 5 62/241,079, filed October 13, 2015, entitled “Systems and Methods for Making and Using Gel Microspheres,” by Weitz, *et al.*, incorporated herein by reference in its entirety.

GOVERNMENT FUNDING

This invention was made with government support under Grant No. NMR-1310266 awarded by the National Science Foundation. The government has certain rights in the 10 invention.

FIELD

The present invention generally relates to microfluidic droplets and, in particular, to multiple emulsion microfluidic droplets.

BACKGROUND

15 Double emulsions are drops containing at least one smaller drop that is composed of a second, substantially immiscible fluid. These core-shell structured fluids can be used, for instance, as templates to produce capsules; the outer drop contains the material that ultimately forms the shell of the capsule, whereas the inner drop constitutes the capsule interior core. These capsules can be used as vehicles for delivery of active ingredients in many fields, such as food, 20 pharmaceuticals, or cosmetics. However, successful application of these capsules may require good control over their permeability and mechanical stability, parameters that can be tuned with the composition and thickness of the capsule shell. This may involve control over the dimensions and composition of the double emulsions. This control is often difficult to achieve if double emulsions are produced by mechanical stirring or membrane emulsification, since these 25 conventional approaches typically yield double emulsion drops of different sizes that often contain multiple inner droplets.

SUMMARY

The present invention generally relates to microfluidic droplets and, in particular, to multiple emulsion microfluidic droplets. The subject matter of the present invention involves, in 30 some cases, interrelated products, alternative solutions to a particular problem, and/or a plurality of different uses of one or more systems and/or articles.

In one aspect, the present invention is generally directed to a method. In one set of embodiments, the method includes providing droplets of a first fluid comprising polymer, surrounded by a second fluid, contained in a carrier fluid; solidifying the second fluid; causing the polymer in the first fluid to form a gel; and removing the solidified second fluid, which may 5 form a suspension of the gel in the carrier fluid.

In another set of embodiments, the method includes acts of providing droplets comprising solvent and polymer contained in a carrier fluid, causing the polymer to form a gel, and removing the solvent from the gel, which may form a suspension of the gel in the carrier fluid.

In another aspect, the present invention encompasses methods of making one or more of 10 the embodiments described herein, for example, gel microspheres. In still another aspect, the present invention encompasses methods of using one or more of the embodiments described herein, for example, gel microspheres.

Other advantages and novel features of the present invention will become apparent from 15 the following detailed description of various non-limiting embodiments of the invention when considered in conjunction with the accompanying figures. In cases where the present specification and a document incorporated by reference include conflicting and/or inconsistent disclosure, the present specification shall control. If two or more documents incorporated by reference include conflicting and/or inconsistent disclosure with respect to each other, then the document having the later effective date shall control.

20 BRIEF DESCRIPTION OF THE DRAWINGS

Non-limiting embodiments of the present invention will be described by way of example with reference to the accompanying figures, which are schematic and are not intended to be drawn to scale. In the figures, each identical or nearly identical component illustrated is typically represented by a single numeral. For purposes of clarity, not every component is 25 labeled in every figure, nor is every component of each embodiment of the invention shown where illustration is not necessary to allow those of ordinary skill in the art to understand the invention. In the figures:

Figs. 1A-1E illustrate a method for producing gel droplets, in one embodiment of the invention;

30 Figs. 2A-2C illustrate a method for producing gel droplets, in accordance with another embodiment of the invention; and

Figs. 3A-3C illustrate a method for producing gel droplets, in another embodiment of the invention.

DETAILED DESCRIPTION

The present invention generally relates to microfluidic droplets and, in particular, to 5 multiple emulsion microfluidic droplets. In certain aspects, particles such as gel particles can be prepared in an aqueous carrier from aqueous droplets (or a non-aqueous carrier from non-aqueous droplets). For example, in some embodiments, double-emulsion droplets of a first fluid, surrounded by a second fluid, contained in a carrier fluid may be prepared, where the first fluid forms a gel and the second fluid is removed. For instance, the second fluid may be dissolved in 10 the carrier fluid, or the second fluid may be hardened, then removed, for example, due to a change in pH. Other embodiments of the present invention are generally directed to kits containing such microfluidic droplets, microfluidic devices for making such microfluidic droplets, or the like.

Certain aspects of the invention are generally directed to systems and methods for 15 making particles, such as gel particles, in an aqueous carrier. Although gel particles can be made from emulsions, e.g., where an aqueous droplet contained within a substantially immiscible oil carrier is then polymerized or gelled to form a particle, such gel particles have not typically been made within aqueous carriers.

Thus, in one aspect of the invention, particles such as gel particles can be prepared in an 20 aqueous carrier from aqueous droplets. In one set of embodiments, a double emulsion droplet may be prepared. For example, in Fig. 1C, a double emulsion droplet is prepared using a plurality of microfluidic channels, which may be concentrically positioned to produce double emulsion droplets, as is known to those of ordinary skill in the art. It should be understood, however, that Fig. 1C is merely one non-limiting illustrative example of a technique to produce 25 double emulsion droplets; a variety of other techniques to produce a double emulsion droplet could also be used. See, e.g., U.S. Pat. Nos. 7,776,927 or 9,039,273; Int. Pat. Appl. Pub. Nos. WO 2006/096571, WO 2008/121342, WO 2010/104604, WO 2011/028760, WO 2011/028764, or WO 2012/162296; U.S. Pat. Appl. Pub. Nos. 2009/0012187, 2009/0131543, 2012/0199226, 2012/0211084, or 2013/0046030; Int. Pat. Appl. Ser. No. PCT/US15/25921; or U.S. Pat. Appl. Ser. 30 Nos. 61/980,541 or 62/083,721, each of which is incorporated herein by reference.

In one set of embodiments, a double emulsion droplet may be prepared that comprises an inner (or first) fluid, surround by an outer (or second) fluid, contained in a carrier (or third) fluid. In some cases, the outer fluid may be substantially immiscible with the inner fluid and the carrier fluid, e.g., at least on the time scale of forming double emulsion droplets. The inner fluid may be 5 caused to form a gel, while the second fluid may be hardened, e.g., to form around the inner fluid. These may be performed in any order, e.g., simultaneously or sequentially. After the gel has been formed, the hardened second fluid may then be removed in some fashion, thereby leaving behind a gel in the carrier fluid.

For example, in one set of embodiments, after the double emulsion droplet has been 10 prepared, a polymer within the inner fluid may be polymerized or gelled, for example, by applying ultraviolet light or a change in temperature. For example, the inner fluid may contain agarose gel (which may be solidified, for example, with a change in temperature). For instance, the droplet may be formed at an elevated temperature (e.g., above room temperature, about 25 °C), then cooled (e.g., to room temperature or to a temperature below room temperature); or the 15 droplet may be formed at room temperature, then cooled to a temperature below room temperature, or the like, to cause polymerization or gelation to occur in the inner fluid.

As another example, the inner fluid may be polymerized or gelled chemically. For example, a chemical reaction or a cross-linking reaction may be induced in the inner fluid to cause polymerization or gelation to occur. In some embodiments, the polymerization is a free- 20 radical polymerization, e.g., which may be initiated by exposing the reactants to heat and/or light, such as ultraviolet (UV) light, and/or an initiator, such as a photoinitiator able to produce free radicals (e.g., via molecular cleavage) upon exposure to light. Examples include, but are not limited to, Irgacure 2559, 2-hydroxy-4-(3-triethoxysilylpropoxy)-diphenylketone, or tetramethylethylenediamine. Examples of such polymers include, but are not limited to 25 polyacrylamide, poly(*N*-isopropylacrylamide), or poly(ethylene glycol) diacrylate. Other species may be present as well.

The outer fluid may be hardened, for example, by inducing a chemical reaction (e.g., between the outer fluid and the carrier fluid), causing precipitation to occur (e.g., if a solvent in the outer fluid then enters the surrounding carrier fluid, leaving behind a polymer that solidifies), 30 changes in temperature, or the like. For example, in one set of embodiments, a first reactant within the fluidic droplet may be reacted with a second reactant within the liquid surrounding the

fluidic droplet to produce a solid, which may thus coat the fluidic droplet within a solid “shell” in some cases. A non-limiting example of a solidification reaction is a polymerization reaction involving production of a nylon (e.g., a polyamide), for example, from a diacyl chloride and a diamine. Other examples include polystyrene, polycaprolactone, polyisoprene, poly(lactic acid),
5 polystyrene (PS), polycaprolactone (PCL), polyisoprene (PIP), poly(lactic acid), polyethylene, polypropylene, polyacrylonitrile, polyimide, polyamide, and/or mixtures and/or co-polymers of these and/or other polymers.

As another example, the outer fluid may contain a methacrylate (which may be in polymerized form, i.e., as a polymethacrylate) contained in ethyl acetate, where the ethyl acetate
10 is soluble in water (e.g., in the carrier and/or inner fluids) and can be removed to leave behind the methacrylate (or polymethacrylate), which can solidify. Non-limiting examples of methacrylates include dimethylaminoethyl methacrylate, butyl methacrylate, ethyl methacrylate, methyl methacrylate, hydroxyethyl methacrylate, trimethylolpropane triacrylate, or the like (including the polymeric forms of these). As yet another example, the outer fluid may contain
15 agarose gel, and the temperature may be lowered (e.g., as discussed above) to cause the agarose gel to solidify to form a “shell” around the inner droplet.

It should be understood that these may happen in any order. For example, the inner fluid may be gelled before or after the outer fluid forms a shell, or both may occur simultaneously.

After the outer fluid hardens into a shell and the inner fluid forms a gel, the outer shell
20 may be removed, for example, mechanically, exposure to a suitable solvent, or other conditional changes (e.g., a change in pH). For example, if the outer shell comprises a polymer, the polymer may be degradable hydrolytically, enzymatically, photolytically, mechanically, etc. As another example, the polymer may be liquefied by heating the polymer above its melting temperature. For example, the outer shell may comprise agarose gel. In yet another example, a methacrylate
25 may be removed upon a change in pH, e.g., to acidic conditions (e.g., a pH of less than 7, less than 6, less than 5, less than 4, less than 3, etc.), thereby leaving behind the gel contained within the carrier fluid. The pH may be changed, for example, by applying a suitable acid or base. The pH may be changed, e.g., by at least about 2, 3, 4, or more pH units. Non-limiting examples of suitable acids include acetic acid, HCl, H₂SO₄, HNO₃, etc. at any suitable dilution or
30 concentration. Non-limiting examples of suitable bases include NaOH or KOH.

Accordingly, by removing the solidified second fluid, a suspension of gel particles contained in a carrier fluid may be obtained. As noted above, in one set of embodiments, the inner fluid forming the gel particles may be aqueous, yet the carrier fluid may also be aqueous instead of non-aqueous (e.g., an oil). (In addition, in another set of embodiments, both the 5 carrier fluid and the inner fluid may each be non-aqueous as well, while the outer fluid may be aqueous instead.)

It should be understood, however, that the invention is not limited to only double emulsion droplets. In another set of embodiments, particles can be made from single emulsion droplets contained within a carrier fluid, where both the fluid forming the particles and the 10 carrier fluid are aqueous (or alternatively, both may be non-aqueous). For instance, in one aspect, a single-emulsion droplet of a first phase (e.g., an oil phase) may be produced in a carrier phase (e.g., an aqueous or water phase), where the first phase is caused to form a polymer or a gel within the carrier phase. In some cases, the first phase may contain a solvent that is able to enter the surrounding carrier fluid, and/or the solvent may be subsequently removed, e.g., 15 resulting in a polymeric or gel particle contained within the carrier fluid.

The first fluid may be solidified to produce a particle. For example, the first fluid may be fluid polymerized or gelled chemically, e.g., as discussed above. For example, a temperature change, a chemical reaction or a cross-linking reaction may be induced in the first fluid to cause polymerization or gelation to occur. The first fluid may contain any of the materials discussed 20 herein, e.g., agarose, polyacrylamide, poly(*N*-isopropylacrylamide), or poly(ethylene glycol diacrylate), initiators such as Irgacure 2559, 2-hydroxy-4-(3-triethoxysilylpropoxy)-diphenylketone, or tetramethylmethylenediamine, etc.

In some cases, the first fluid may contain a solvent that can be removed, e.g., upon exposure to the surrounding fluid. The solvent may be removed, e.g., sequentially or 25 simultaneously with formation of particles. In some cases, removal of the solvent may cause gelation or polymerization in the first fluid to form particles. In certain embodiments, if the surrounding fluid is an aqueous fluid, the solvent may have relatively low solubility in water, such that initially a separate phase forms (e.g., the first fluid), but the solvent then gradually dissolves in the surrounding fluid. Examples of such solvents include, but are not limited to, 30 dichloromethane, 1,2-dichloroethane, 1-butanol, diethyl ether, ethyl acetate, methyl t-butyl ether (MTBE), or the like.

In some cases, the solvent may have a water solubility of less than about 1 g/ml, less than about 0.5 g/ml, less than about 0.4 g/ml, less than about 0.3 g/ml, less than about 0.2 g/ml, less than about 0.1 g/ml, less than about 0.05 g/ml, less than about 0.04 g/ml, less than about 0.03 g/ml, less than about 0.02 g/ml, less than about 0.01 g/ml, or less than about 0.005 g/ml. In 5 addition, in some embodiments, the solvent may be removed by exposing the solidified particles to a surrounding fluid that can at least partially remove the solvent, e.g., via extraction, dissolution, or other techniques. For example, particles may be collected from the surrounding carrier fluid, which may be replaced by another fluid (e.g., of the same or different composition) to at least partially remove the solvent from the solidified particles.

10 According to certain aspects, the systems and methods described herein can be used in a plurality of applications. For example, fields in which the particles and multiple emulsions described herein may be useful include, but are not limited to, food, beverage, health and beauty aids, paints and coatings, chemical separations, agricultural applications, and drugs and drug delivery. For instance, a precise quantity of a fluid, drug, pharmaceutical, or other species can be 15 contained in a droplet or particle designed to release its contents under particular conditions. In some instances, cells can be contained within a droplet or particle, and the cells can be stored and/or delivered, e.g., to a target medium, for example, within a subject. Other species that can be contained within a droplet or particle and delivered to a target medium include, for example, biochemical species such as nucleic acids such as siRNA, RNAi, RNA, and DNA, proteins, 20 peptides, or enzymes. Additional species that can be contained within a droplet or particle include, but are not limited to, colloidal particles, magnetic particles, nanoparticles, quantum dots, fragrances, proteins, indicators, dyes, fluorescent species, chemicals, or the like. The target medium may be any suitable medium, for example, water, saline, an aqueous medium, a hydrophobic medium, or the like.

25 For example, in one set of embodiments, a species may be contained within and/or on the surface of a particle. In some cases, for instance, the species may be bonded (for example, covalently) to the particle, and/or contained (e.g., physically) within the particle. The species may be, for example, DNA or any of other species described above. In some cases, the species may be present within the droplet (e.g., prior to formation of a particle), or added during or after 30 formation of the particle. For example, in one set of embodiments, DNA may be attached to a particle using EDC/NHS coupling (1-ethyl-3-(3-dimethylaminopropyl)carbodiimide/N-

hydroxysuccinimide) or other suitable coupling reactions. In other cases, however, DNA (or other species) may be physically contained within the particles.

In one aspect, the present invention is generally directed to a double (or higher) multiple emulsion. Generally, in a double emulsion, a first (or inner) fluidic droplet comprising a first fluid is surrounded by a second (or middle) fluidic droplet comprising a second fluid, which is contained within a continuous, carrier, or third fluid. Typically, a fluid is substantially immiscible with an adjacent fluid, although fluids that are not adjacent need not be immiscible, and may be miscible (or even identical) in some cases. Thus, for example, the first fluid may be immiscible with the second fluid, but may be miscible or immiscible with the third fluid.

10 Similarly, the second fluid may be immiscible with the third fluid. However, it should be understood that immiscibility is not necessarily required in all embodiments; in some cases, two adjacent fluids are not immiscible, but may retain separation in other ways, e.g., kinetically or through short exposure times.

Thus, as a non-limiting example, in a double emulsion droplet, the first fluid (innermost fluid) may be an aqueous or hydrophilic fluid (a “water” phase), the second fluid (middle fluid) may be a lipophilic or hydrophobic or “oil” phase that is substantially immiscible with the aqueous fluid, and the third (or carrier) fluid may be an aqueous fluid (a “water” fluid) that is substantially immiscible with the second fluid. This is sometimes generally referred to as a W/O/W double emulsion droplet (for water/oil/water), although it should be understood that this is mainly for the sake of convenience; for instance, the first fluid can be any suitable aqueous fluid, and it need not be pure water. For example, the aqueous fluid may be water, saline, an aqueous solution, ethanol, or the like, or any other fluid miscible in water. The oil, in contrast, may be immiscible in water, at least when left undisturbed under ambient conditions. In similar fashion, an O/W/O double emulsion droplet may be similarly defined. Furthermore, these principles may be extended to higher-order multiple emulsions droplets. In addition, it should be understood that other arrangements are also possible. For example, in one embodiment, the first fluid, the second fluid, and the third fluid may be all mutually immiscible.

As used herein, two fluids are immiscible, or not miscible, with each other when one is not soluble in the other to a level of at least 10% by weight at the temperature and under the conditions at which the emulsion is produced. For instance, two fluids may be selected to be immiscible within the time frame of the formation of the fluidic droplets. In some embodiments,

two fluids (e.g., the carrying fluid and the inner droplet fluid of a multiple emulsion) are compatible, or miscible, while the outer droplet fluid is incompatible or immiscible with one or both of the carrying and inner droplet fluids. In other embodiments, however, all three (or more) fluids may be mutually immiscible, and in certain cases, all of the fluids do not all necessarily 5 have to be water soluble. In still other embodiments, as mentioned, additional fourth, fifth, sixth, etc. fluids may be added to produce increasingly complex droplets within droplets, e.g., a carrying fluid may surround a first fluid, which may in turn surround a second fluid, which may in turn surround a third fluid, which in turn surround a fourth fluid, etc. In addition, the physical properties of each nesting layer of fluidic droplets may each be independently controlled, e.g., by 10 control over the composition of each nesting level.

The droplets may be microfluidic droplets, in some instances. For instance, the outer droplet may have a diameter of less than about 1 mm, less than about 500 micrometers, less than about 200 micrometers, less than about 100 micrometers, less than about 75 micrometers, less than about 50 micrometers, less than about 25 micrometers, less than about 10 micrometers, or 15 less than about 5 micrometers, or between about 50 micrometers and about 1 mm, between about 10 micrometers and about 500 micrometers, or between about 50 micrometers and about 100 micrometers in some cases. However, in some cases, the droplets may be larger. For example, the inner droplet (or a middle droplet) of a triple or other multiple emulsion droplet may have a diameter of less than about 1 mm, less than about 500 micrometers, less than about 200 micrometers, less than about 100 micrometers, less than about 75 micrometers, less than about 50 micrometers, less than about 25 micrometers, less than about 10 micrometers, or less than about 5 micrometers, or between about 50 micrometers and about 1 mm, between about 20 10 micrometers and about 500 micrometers, or between about 50 micrometers and about 100 micrometers in some cases.

25 The particles (e.g., gel particles) or droplets described herein may have any suitable average cross-sectional diameter. Those of ordinary skill in the art will be able to determine the average cross-sectional diameter of a single and/or a plurality of particles or droplets, for example, using laser light scattering, microscopic examination, or other known techniques. The average cross-sectional diameter of a single particle or droplet, in a non-spherical particle or droplet, is the diameter of a perfect sphere having the same volume as the non-spherical particle or droplet. The average cross-sectional diameter of a particle or droplet (and/or of a plurality or

series of particles or droplets) may be, for example, less than about 1 mm, less than about 500 micrometers, less than about 200 micrometers, less than about 100 micrometers, less than about 75 micrometers, less than about 50 micrometers, less than about 25 micrometers, less than about 10 micrometers, or less than about 5 micrometers, or between about 50 micrometers and about 1 mm, between about 10 micrometers and about 500 micrometers, or between about 5 micrometers and about 100 micrometers in some cases. The average cross-sectional diameter may also be at least about 1 micrometer, at least about 2 micrometers, at least about 3 micrometers, at least about 5 micrometers, at least about 10 micrometers, at least about 15 micrometers, or at least about 20 micrometers in certain cases. In some embodiments, at least 10 about 50%, at least about 75%, at least about 90%, at least about 95%, or at least about 99% of the particles or droplets within a plurality of particles or droplets has an average cross-sectional diameter within any of the ranges outlined in this paragraph.

The plurality of particles (e.g., gel particles) or droplets may have relatively uniform cross-sectional diameters in certain embodiments. The use of particles or droplets with relatively 15 uniform cross-sectional diameters can allow one to control viscosity, the amount of species delivered to a target, and/or other parameters of the delivery of fluid and/or species from the particles or droplets. In some embodiments, the particles or droplets of particles is monodisperse, or the plurality of particles or droplets has an overall average diameter and a distribution of diameters such that no more than about 5%, no more than about 2%, or no more 20 than about 1% of the particles or droplets have a diameter less than about 90% (or less than about 95%, or less than about 99%) and/or greater than about 110% (or greater than about 105%, or greater than about 101%) of the overall average diameter of the plurality of particles or droplets.

In some embodiments, the plurality of particles or droplets has an overall average diameter and a distribution of diameters such that the coefficient of variation of the cross-sectional diameters of the particles or droplets is less than about 10%, less than about 5%, less 25 than about 2%, between about 1% and about 10%, between about 1% and about 5%, or between about 1% and about 2%. The coefficient of variation can be determined by those of ordinary skill in the art, and may be defined as:

$$c_v = \frac{\sigma}{|\mu|}$$

30 wherein σ is the standard deviation and μ is the mean.

In certain aspects of the present invention, as discussed, multiple emulsions are formed by flowing fluids through one or more channels, e.g., as shown in Fig. 1C. The system may be a microfluidic system. “Microfluidic,” as used herein, refers to a device, apparatus, or system including at least one fluid channel having a cross-sectional dimension of less than about 1
5 millimeter (mm), and in some cases, a ratio of length to largest cross-sectional dimension of at least 3:1. One or more channels of the system may be a capillary tube. In some cases, multiple channels are provided, and in some embodiments, at least some are nested, as described herein. The channels may be in the microfluidic size range and may have, for example, average inner
10 diameters, or portions having an inner diameter, of less than about 1 millimeter, less than about 300 micrometers, less than about 100 micrometers, less than about 30 micrometers, less than about 10 micrometers, less than about 3 micrometers, or less than about 1 micrometer, thereby providing droplets having comparable average diameters. One or more of the channels may (but not necessarily), in cross-section, have a height that is substantially the same as a width at the same point. In cross-section, the channels may be rectangular or substantially non-rectangular,
15 such as circular or elliptical.

As used herein, the term “fluid” generally refers to a substance that tends to flow and to conform to the outline of its container, i.e., a liquid, a gas, a viscoelastic fluid, etc. In one embodiment, the fluid is a liquid. Typically, fluids are materials that are unable to withstand a static shear stress, and when a shear stress is applied, the fluid experiences a continuing and
20 permanent distortion. The fluid may have any suitable viscosity that permits flow. If two or more fluids are present, each fluid may be independently selected among essentially any fluids (liquids, gases, and the like) by those of ordinary skill in the art, by considering the relationship between the fluids.

A variety of materials and methods, according to certain aspects of the invention, can be
25 used to form articles or components such as those described herein, e.g., channels such as microfluidic channels, chambers, etc. For example, various articles or components can be formed from solid materials, in which the channels can be formed via micromachining, film deposition processes such as spin coating and chemical vapor deposition, laser fabrication, photolithographic techniques, etching methods including wet chemical or plasma processes, 3D printing, and the like. See, for example, *Scientific American*, 248:44-55, 1983 (Angell, *et al*).

In one set of embodiments, various structures or components of the articles described herein can be formed from glass or a polymer, for example, an elastomeric polymer such as polydimethylsiloxane (“PDMS”), polytetrafluoroethylene (“PTFE” or Teflon[®]), epoxy, norland optical adhesive, or the like. For instance, according to one embodiment, microfluidic channels 5 may be formed from glass tubes or capillaries. In addition, in some cases, a microfluidic channel may be implemented by fabricating the fluidic system separately using PDMS or other soft lithography techniques (details of soft lithography techniques suitable for this embodiment are discussed in the references entitled “Soft Lithography,” by Younan Xia and George M. Whitesides, published in the *Annual Review of Material Science*, 1998, Vol. 28, pages 153-184, 10 and “Soft Lithography in Biology and Biochemistry,” by George M. Whitesides, Emanuele Ostuni, Shuichi Takayama, Xingyu Jiang and Donald E. Ingber, published in the *Annual Review of Biomedical Engineering*, 2001, Vol. 3, pages 335-373; each of these references is incorporated herein by reference). In addition, in some embodiments, various structures or components of the articles described herein can be formed of a metal, for example, stainless steel.

15 Other examples of potentially suitable polymers include, but are not limited to, polyethylene terephthalate (PET), polyacrylate, polymethacrylate, polycarbonate, polystyrene, polyethylene, polypropylene, polyvinylchloride, cyclic olefin copolymer (COC), polytetrafluoroethylene, a fluorinated polymer, a silicone such as polydimethylsiloxane, polyvinylidene chloride, bis-benzocyclobutene (“BCB”), a polyimide, a fluorinated derivative of 20 a polyimide, or the like. Combinations, copolymers, or blends involving polymers including those described above are also envisioned. The device may also be formed from composite materials, for example, a composite of a polymer and a semiconductor material.

In some embodiments, various structures or components of the article are fabricated from 25 polymeric and/or flexible and/or elastomeric materials, and can be conveniently formed of a hardenable fluid, facilitating fabrication via molding (e.g. replica molding, injection molding, cast molding, etc.). The hardenable fluid can be essentially any fluid that can be induced to solidify, or that spontaneously solidifies, into a solid capable of containing and/or transporting fluids contemplated for use in and with the fluidic network. In one embodiment, the hardenable fluid comprises a polymeric liquid or a liquid polymeric precursor (i.e. a “prepolymer”).

30 Suitable polymeric liquids can include, for example, thermoplastic polymers, thermoset polymers, waxes, or mixtures or composites thereof heated above their melting point. As

another example, a suitable polymeric liquid may include a solution of one or more polymers in a suitable solvent, which solution forms a solid polymeric material upon removal of the solvent, for example, by evaporation. Such polymeric materials, which can be solidified from, for example, a melt state or by solvent evaporation, are well known to those of ordinary skill in the art. A variety of polymeric materials, many of which are elastomeric, are suitable, and are also suitable for forming molds or mold masters, for embodiments where one or both of the mold masters is composed of an elastomeric material. A non-limiting list of examples of such polymers includes polymers of the general classes of silicone polymers, epoxy polymers, and acrylate polymers. Epoxy polymers are characterized by the presence of a three-membered cyclic ether group commonly referred to as an epoxy group, 1,2-epoxide, or oxirane. For example, diglycidyl ethers of bisphenol A can be used, in addition to compounds based on aromatic amine, triazine, and cycloaliphatic backbones. Another example includes the well-known Novolac polymers. Non-limiting examples of silicone elastomers suitable for use according to the invention include those formed from precursors including the chlorosilanes such as methylchlorosilanes, ethylchlorosilanes, phenylchlorosilanes, dodecyltrichlorosilanes, etc.

Silicone polymers are used in certain embodiments, for example, the silicone elastomer polydimethylsiloxane. Non-limiting examples of PDMS polymers include those sold under the trademark Sylgard by Dow Chemical Co., Midland, MI, and particularly Sylgard 182, Sylgard 184, and Sylgard 186. Silicone polymers including PDMS have several beneficial properties simplifying fabrication of various structures of the invention. For instance, such materials are inexpensive, readily available, and can be solidified from a prepolymeric liquid via curing with heat. For example, PDMSs are typically curable by exposure of the prepolymeric liquid to temperatures of about, for example, about 65 °C to about 75 °C for exposure times of, for example, about an hour, about 3 hours, about 12 hours, etc. Also, silicone polymers, such as PDMS, can be elastomeric and thus may be useful for forming very small features with relatively high aspect ratios, necessary in certain embodiments of the invention. Flexible (e.g., elastomeric) molds or masters can be advantageous in this regard.

One advantage of forming structures such as microfluidic structures or channels from silicone polymers, such as PDMS, is the ability of such polymers to be oxidized, for example by exposure to an oxygen-containing plasma such as an air plasma, so that the oxidized structures contain, at their surface, chemical groups capable of cross-linking to other oxidized silicone

polymer surfaces or to the oxidized surfaces of a variety of other polymeric and non-polymeric materials. Thus, structures can be fabricated and then oxidized and essentially irreversibly sealed to other silicone polymer surfaces, or to the surfaces of other substrates reactive with the oxidized silicone polymer surfaces, without the need for separate adhesives or other sealing

5 means. In most cases, sealing can be completed simply by contacting an oxidized silicone surface to another surface without the need to apply auxiliary pressure to form the seal. That is, the pre-oxidized silicone surface acts as a contact adhesive against suitable mating surfaces.

10 Specifically, in addition to being irreversibly sealable or bonded to itself, oxidized silicone such as oxidized PDMS can also be sealed irreversibly to a range of oxidized materials other than itself including, for example, glass, silicon, silicon oxide, quartz, silicon nitride, polyethylene, polystyrene, glassy carbon, and epoxy polymers, which have been oxidized in a similar fashion to the PDMS surface (for example, via exposure to an oxygen-containing plasma). Oxidation and sealing methods useful in the context of the present invention, as well as overall molding techniques, are described in the art, for example, in an article entitled “Rapid Prototyping of

15 Microfluidic Systems and Polydimethylsiloxane,” *Anal. Chem.*, 70:474-480, 1998 (Duffy *et al.*), incorporated herein by reference.

Different components can be fabricated of different materials. For example, a base portion including a bottom wall and side walls can be fabricated from an opaque material such as silicon or PDMS, and a top portion can be fabricated from a transparent or at least partially transparent material, such as glass or a transparent polymer, for observation and/or control of the fluidic process. Components can be coated so as to expose a desired chemical functionality to fluids that contact interior channel walls, where the base supporting material does not have a precise, desired functionality. For example, components can be fabricated as illustrated, with interior channel walls coated with another material, e.g., as discussed herein. Material used to fabricate various components of the systems and devices of the invention, e.g., materials used to coat interior walls of fluid channels, may desirably be selected from among those materials that will not adversely affect or be affected by fluid flowing through the fluidic system, e.g., material(s) that is chemically inert in the presence of fluids to be used within the device. A non-limiting example of such a coating is disclosed below; additional examples are disclosed in Int. 25 Pat. Appl. Ser. No. PCT/US2009/000850, filed February 11, 2009, entitled “Surfaces, Including

Microfluidic Channels, With Controlled Wetting Properties," by Weitz, *et al.*, published as WO 2009/120254 on October 1, 2009, incorporated herein by reference.

In some embodiments, certain microfluidic structures of the invention (or interior, fluid-contacting surfaces) may be formed from certain oxidized silicone polymers. Such surfaces may be more hydrophilic than the surface of an elastomeric polymer. Such hydrophilic surfaces can thus be more easily filled and wetted with aqueous solutions.

In some embodiments, a bottom wall of a microfluidic device of the invention is formed of a material different from one or more side walls or a top wall, or other components. For example, in some embodiments, the interior surface of a bottom wall comprises the surface of a silicon wafer or microchip, or other substrate. Other components may, as described above, be sealed to such alternative substrates. Where it is desired to seal a component comprising a silicone polymer (e.g. PDMS) to a substrate (bottom wall) of different material, the substrate may be selected from the group of materials to which oxidized silicone polymer is able to irreversibly seal (e.g., glass, silicon, silicon oxide, quartz, silicon nitride, polyethylene, polystyrene, epoxy polymers, and glassy carbon surfaces which have been oxidized).

Alternatively, other sealing techniques may be used, as would be apparent to those of ordinary skill in the art, including, but not limited to, the use of separate adhesives, bonding, solvent bonding, ultrasonic welding, etc.

Thus, in certain embodiments, the design and/or fabrication of the article may be relatively simple, e.g., by using relatively well-known soft lithography and other techniques such as those described herein. In addition, in some embodiments, rapid and/or customized design of the article is possible, for example, in terms of geometry. In one set of embodiments, the article may be produced to be disposable, for example, in embodiments where the article is used with substances that are radioactive, toxic, poisonous, reactive, biohazardous, etc., and/or where the profile of the substance (e.g., the toxicology profile, the radioactivity profile, etc.) is unknown. Another advantage to forming channels or other structures (or interior, fluid-contacting surfaces) from oxidized silicone polymers is that these surfaces can be much more hydrophilic than the surfaces of typical elastomeric polymers (where a hydrophilic interior surface is desired). Such hydrophilic channel surfaces can thus be more easily filled and wetted with aqueous solutions than can structures comprised of typical, unoxidized elastomeric polymers or other hydrophobic materials.

In one set of embodiments, one or more of the channels within the device may be relatively hydrophobic or relatively hydrophilic, e.g. inherently, and/or by treating one or more of the surfaces or walls of the channel to render them more hydrophobic or hydrophilic. Generally, the fluids that are formed droplets in the device are substantially immiscible, at least 5 on the time scale of forming the droplets, and the fluids will often have different degrees of hydrophobicity or hydrophilicity. Thus, for example, a first fluid may be more hydrophilic (or more hydrophobic) relative to a second fluid, and the first and the second fluids may be substantially immiscible. Thus, the first fluid can form a discrete droplet within the second fluid, e.g., without substantial mixing of the first fluid and the second fluid (although some degree of 10 mixing may nevertheless occur under some conditions). Similarly, the second fluid may be more hydrophilic (or more hydrophobic) relative to a third fluid (which may be the same or different than the first fluid), and the second and third fluids may be substantially immiscible.

Accordingly, in some cases, a surface of a channel may be relatively hydrophobic or hydrophilic, depending on the fluid contained within the channel. In one set of embodiments, a 15 surface of the channel is hydrophobic or hydrophilic relative to other surfaces within the device. In addition, in some embodiments, a relatively hydrophobic surface may exhibit a water contact angle of greater than about 90°, and/or a relatively hydrophilic surface may exhibit a water contact angle of less than about 90°.

In some cases, relatively hydrophobic and/or hydrophilic surfaces may be used to 20 facilitate the flow of fluids within the channel, e.g., to maintain the nesting of multiple fluids within the channel in a particular order. Additional details of such coatings and other systems may be seen in U.S. Provisional Patent Application Serial No. 61/040,442, filed March 28, 2008, entitled “Surfaces, Including Microfluidic Channels, With Controlled Wetting Properties,” by Abate, *et al.*; and International Patent Application Serial No. PCT/US2009/000850, filed 25 February 11, 2009, entitled “Surfaces, Including Microfluidic Channels, With Controlled Wetting Properties,” by Abate, *et al.*, each incorporated herein by reference.

Certain aspects of the invention are generally directed to techniques for scaling up or “numbering up” devices such as those discussed herein. For example, in some cases, relatively 30 large numbers of devices may be used in parallel, for example at least about 10 devices, at least about 30 devices, at least about 50 devices, at least about 75 devices, at least about 100 devices, at least about 200 devices, at least about 300 devices, at least about 500 devices, at least about

750 devices, or at least about 1,000 devices or more may be operated in parallel. In some cases, an array of such devices may be formed by stacking the devices horizontally and/or vertically. The devices may be commonly controlled, or separately controlled, and can be provided with common or separate sources of various fluids, depending on the application.

5 Those of ordinary skill in the art will be aware of other techniques useful for scaling up or numbering up devices or articles such as those discussed herein. For example, in some embodiments, a fluid distributor can be used to distribute fluid from one or more inputs to a plurality of outputs, e.g., in one or more devices. For instance, a plurality of articles may be connected in three dimensions. In some cases, channel dimensions are chosen that allow
10 pressure variations within parallel devices to be substantially reduced. Other examples of suitable techniques include, but are not limited to, those disclosed in International Patent Application No. PCT/US2010/000753, filed March 12, 2010, entitled “Scale-up of Microfluidic Devices,” by Romanowsky, *et al.*, published as WO 2010/104597 on November 16, 2010, incorporated herein by reference in its entirety.

15 The following documents are incorporated herein by reference in their entirety for all purposes: U.S. Provisional Application Serial Number 61/980,541, filed April 16, 2014, entitled “Systems and methods for producing droplet emulsions with relatively thin shells”; International Patent Publication Number WO 2004/091763, filed April 9, 2004, entitled “Formation and Control of Fluidic Species,” by Link *et al.*; International Patent Publication Number WO
20 2004/002627, filed June 3, 2003, entitled “Method and Apparatus for Fluid Dispersion,” by Stone *et al.*; International Patent Publication Number WO 2006/096571, filed March 3, 2006, entitled “Method and Apparatus for Forming Multiple Emulsions,” by Weitz *et al.*; International Patent Publication Number WO 2005/021151, filed August 27, 2004, entitled “Electronic Control of Fluidic Species,” by Link *et al.*; International Patent Publication Number WO
25 2008/121342, filed March 28, 2008, entitled “Emulsions and Techniques for Formation,” by Chu *et al.*; International Patent Publication Number WO 2010/104604, filed March 12, 2010, entitled “Method for the Controlled Creation of Emulsions, Including Multiple Emulsions,” by Weitz *et al.*; International Patent Publication Number WO 2011/028760, filed September 1, 2010, entitled “Multiple Emulsions Created Using Junctions,” by Weitz *et al.*; International Patent Publication
30 Number WO 2011/028764, filed September 1, 2010, entitled “Multiple Emulsions Created Using Jetting and Other Techniques,” by Weitz *et al.*; International Patent Publication Number WO

2009/148598, filed June 4, 2009, entitled “Polymersomes, Phospholipids, and Other Species Associated with Droplets,” by Shum, *et al.*; International Patent Publication Number WO 2011/116154, filed March 16, 2011, entitled “Melt Emulsification,” by Shum, *et al.*; International Patent Publication Number WO 2009/148598, filed June 4, 2009, entitled 5 “Polymersomes, Colloidosomes, Liposomes, and other Species Associated with Fluidic Droplets,” by Shum, *et al.*; International Patent Publication Number WO 2012/162296, filed May 22, 2012, entitled “Control of Emulsions, Including Multiple Emulsions,” by Rotem, *et al.*; International Patent Publication Number WO 2013/006661, filed July 5, 2012, entitled “Multiple Emulsions and Techniques for the Formation of Multiple Emulsions,” by Kim, *et al.*; and 10 International Patent Publication Number WO 2013/032709, filed August 15, 2012, entitled “Systems and Methods for Shell Encapsulation,” by Weitz, *et al.*

In addition, U.S. Provisional Patent Application Serial No. 62/241,079, filed October 13, 2015, entitled “Systems and Methods for Making and Using Gel Microspheres,” by Weitz, *et al.*, is incorporated herein by reference in its entirety.

15 The following examples are intended to illustrate certain embodiments of the present invention, but do not exemplify the full scope of the invention.

EXAMPLE 1

This example illustrates a microfluidic approach to fabricate monodisperse hydrogel microspheres using a double emulsion as the template. Polymerizable precursors are added in 20 the inner phase and crosslinked after the generation of double emulsion drops. Depending on the precursor used, different types of middle phase may be used to temporarily separate the inner phase from the outer phase. In one example, the middle phase is a polymer solution in an organic solvent. After the formation of double emulsion drops, the solvent quickly diffuses into water with the polymer precipitating out to form a thin polymeric shell; then the polymer shell is 25 formed by adjusting the pH of the solution. In another example, double emulsion drops are prepared with an oil shell, and then the oil is removed by centrifugation. These methods allow microgels to be obtained conveniently and efficiently. Examples follow with fabrication of poly(ethylene glycol) diacrylate (PEGDA) and polyacrylamide (PAM) microgels.

Poly(ethylene glycol) diacrylate (PEGDA) microgels from double-emulsion templated 30 capsules. To fabricate monodisperse PEGDA microgels, water-in-oil-in-water (W/O/W) double emulsion drops are produced with a very thin oil layer using a glass-capillary based microfluidic

device as shown in Fig. 1A. The device has two tapered round capillaries co-axially assembled in a square capillary. Another thin capillary enters into the small-orifice tapered capillary as the inlet of inner phase (Fig. 1A). The middle phase is injected from the gap between the small-orifice tapered capillary and the thin capillary. The outer phase is injected from the gap between the small-orifice tapered capillary and the square capillary. The inner phase first breaks into liquid slugs at the tip of the thin capillary; each liquid slug then breaks into many monodisperse drops with a middle phase shell at the junction of the tapered capillaries. The inner phase was an aqueous solution of 20 wt.% PEGDA (Mw~700, Sigma-Aldrich), 2 wt% PVA (polyvinyl alcohol, Mw~13,000-23,000, 87-89% hydrolyzed, Sigma-Aldrich), 0.2 wt% Irgacure 2959 as the photo-initiator, with a small amount of FITC-dextran (Mw~10k, Sigma-Aldrich) to facilitate visualization. The middle phase was 10 wt.% Eudragit[®] EPO dissolved in ethyl acetate; Eudragit[®] EPO is a cationic copolymer that is soluble in acidic solution up to pH 5. The outer phase was 10 wt.% PVA solution. The pH of the inner and outer phases was adjusted to be 7-7.3 by adding BDH[®] buffer pH 11.

Fig. 1A shows a scheme for microfluidic generation of thin-shell microgel capsules. Fig. 1B shows confocal microscopy images of microgels with pH-responsive polymeric shells. Fig. 1C shows confocal microscopy images of clean microgels after removing the polymeric shells by adding acid. Fig. 1D illustrates that a microgel can be broken into small pieces after being pressed.

After the double emulsion drops were generated, ethyl acetate quickly diffused into water, with EPO precipitating to form the solid shell. In the meantime, UV light was illuminated on-chip to polymerize the PEGDA in the core. The resultant core-shell capsules were collected in 2 wt% PVA solution of pH 7-7.3. The EPO shell appeared to show wrinkles due to the volume shrinkage of PEGDA during polymerization, as shown in Fig. 1B. Depending on the shell thickness and capsule size, the shell may also crack under some conditions, as shown in Fig. 1E.

To remove the EPO shell, add diluted acetic acid of pH 4 was added to the gel suspension; the EPO was then dissolved by the acidic solution, resulting in PEGDA microgels, as shown in Fig. 1C. When the microgels were pressed between two glass slides, the gel broke into many small pieces without losing the fluorescent dye, indicating that PEGDA was fully polymerized (Fig. 1D).

EXAMPLE 2

This example illustrates an approach to fabricate polyacrylamide microgels from regular double emulsion. Monodisperse double emulsions were produced using a glass-capillary based microfluidic device as in Fig. 2A. The inner phase was the aqueous solution of 20 wt.% acrylamide, 0.2 wt.% N,N'-Methylenebisacrylamide, and 0.2 wt% Irgacure 2959 with 5 fluorescent microspheres (0.1 micrometers, green). The middle phase was HFE 7500 oil with 1 wt% krytox-PEO surfactant. The outer phase was 10 wt% PVA solution. UV light was applied to the double emulsion drops on-chip before collecting them in a vial. Double emulsion drops and single emulsion HFE drops were generated alternatively in the microfluidic channel in some 10 cases, resulting in a mixture of double emulsion and single emulsion drops, as shown in Fig. 2B.

To remove the HFE oil from the microgels, the sample was centrifuged at a speed of 1000 rpm. After centrifugation, the HFE oil drops settled down at the bottom. The 15 polyacrylamide microgels stayed above the HFE drops, and the top layer was the continuous water phase. The microgels were segregated using a pipette; the resultant microgels are shown in Fig. 2C. When press the microgel were pressed using a glass slide, the gel broke into several small pieces without losing any fluorescence, indicating the complete polymerization of acrylamide.

Fig. 2A shows a scheme of microfluidic generation of gel-oil double emulsion. Fig. 2B is a confocal microscopy image of polyacrylamide microgels with an oil shell. Fig. 2C is a 20 confocal microscopy image of clean polyacrylamide microgels after removing the oil shell.

EXAMPLE 3

This example shows PEGDA hydrogel microspheres prepared from single emulsions. To fabricate a microhydrogel, a glass capillary device was used. Poly(ethylene glycol) diacrylate (PEGDA) with different molecular weight (250, 575, and 700) in dichloromethane (DCM) with 25 volume ratio of 1 to 1 were chosen as inner phase liquid, and 10% PVA aqueous solution was chosen as outer phase liquid. The flow rates for inner and outer phase were 1000 microliters/h and 3500 microliters/h, respectively. Oil-in-water single emulsion droplets were steadily produced, and then UV light was used to crosslink PEGDA in the core to get stable monodisperse spherical drops, as shown in Fig. 3A.

30 The drops were washed three times with ethanol/water (v/v: 1/1) to remove the inside DCM, and then were immersed into pure water. The supernatant was exchanged with fresh pure

water for three times. Finally, PEG microhydrogel drops were suspended in pure deionized water, and were characterized using confocal microscope, as shown in bottom figures. Fig. 3B shows the initial status and Fig. 3C shows the dry status, optical images.

EXAMPLE 4

5 This example shows the preparation of poly(ethylene glycol) microhydrogel containing DNA from an O/W single emulsion.

Fabrication of glass capillary microfluidic device. All microhydrogel beads used in this example were prepared using glass capillary microfluidic device. The device was built on a glass slide, with a square glass tube and two cylindrical glass tubes. Briefly, two cylindrical 10 capillaries were pulled with a Flaming/Brown micropipette puller (Model P-97, Sutter Instrument Co., USA) to obtain tapered tips and then their tips were sanded to make a final diameters of 20 and 140 micrometers, respectively. The tapered capillary with small tip was then inserted into a second square capillary and was used as the injection capillary. Another tapered 15 capillary with large tip was inserted into the square capillary at the other side to confine the flow near the injection tip and was used as the collection capillary. Both capillaries were coaxially aligned within the square capillary, as shown in Fig. 4, which shows a schematic illustration of preparing PEG microhydrogel containing DNA.

Preparation of PEG microhydrogel beads. PEG microhydrogel beads were produced from O/W single emulsion droplets using the microfluidic device, followed by UV exposure to 20 polymerize PEG acrylate monomers. The oil phase was made of 20 % poly (ethylene glycol) methyl ether acrylate with molecular weight of 480, 5% poly(ethylene glycol) diacrylate with molecular weight of 575 as crosslinker, and 2.5% acrylic acid to bring in carboxyl groups in the hydrogel beads. The solvent was dichloromethane (DCM). 2,2-diethoxyacetophenone at a concentration of 4% was added to the oil phase as a photoinitiator. The aqueous phase was made 25 of 10% poly(vinyl alcohol) (PVA) in water. Both oil phase and aqueous phase were pumped into the glass capillary device with syringes using Harvard pumps (Harvard Apparatus Hollston, USA). The oil phase was injected into injection capillary with flow rate of 120 microliters/h, and the aqueous phase was injected through an end of square capillary and used as continuous phase with flow rate of 3000 microliters/h. O/W single emulsion drops were formed in the collection 30 capillary. As-formed drops were collected in a glass bottle with 3% PVA solution, followed by UV exposure to produce a crosslinked PEG hydrogel network. As-prepared microhydrogel

beads were washed with water and ethanol for several times to remove DCM in the microhydrogel. Finally, the microhydrogel beads were suspended in MES buffer with 1% Tween 20 for further use.

To bind DNA to the PEG microhydrogel, an EDC coupling method was used. 2 mM EDC (1-ethyl-3-(3-dimethylaminopropyl)carbodiimide) and 5 mM NHS (*N*-hydroxysuccinimide) were added to 1 ml of a microhydrogel bead suspension. After being placed on vortex for 10 min, 50 microliters of 1 mM NH₂-DNA were added into the suspension. After reacting for overnight at room temperature, the microhydrogel beads were washed with 1% Tween 20 solution three times; each time took about 30 min. Finally, microhydrogel beads were suspended in buffer solution for further testing.

To confirm successful incorporation of DNA into microhydrogel, the complementary DNA bearing FAM fluorophore solution was added to the microhydrogel beads suspension. After 45 min, the supernatant solution was removed and the microhydrogel beads were washed with 1% Tween 20 solution three times. Then, confocal fluorescence microscope was used to characterize the PEG microhydrogel with DNA. As a control, as-prepared PEG microhydrogel without DNA functionalization was chosen to be treated with the same process. Figs. 5A-5B show strong fluorescence signal from PEG microhydrogel functionalized with DNA, while Figs. 5C-5D show no detectable fluorescent signal for control sample without DNA. These results indicate successful incorporation of DNA with high loading efficiency.

Figs. 5A and 5B show confocal images of PEG microhydrogel with DNA after be treated with complementary DNA bearing FAM. Figs. 5C and 5D shows confocal images of PEG microhydrogel without DNA as control after being treated with complementary DNA bearing FAM. Figs. 5A and 5C are overlapping transmission and fluorescent images; Figs. 5B and 5D are fluorescent images.

25

While several embodiments of the present invention have been described and illustrated herein, those of ordinary skill in the art will readily envision a variety of other means and/or structures for performing the functions and/or obtaining the results and/or one or more of the advantages described herein, and each of such variations and/or modifications is deemed to be within the scope of the present invention. More generally, those skilled in the art will readily appreciate that all parameters, dimensions, materials, and configurations described herein are

meant to be exemplary and that the actual parameters, dimensions, materials, and/or configurations will depend upon the specific application or applications for which the teachings of the present invention is/are used. Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific 5 embodiments of the invention described herein. It is, therefore, to be understood that the foregoing embodiments are presented by way of example only and that, within the scope of the appended claims and equivalents thereto, the invention may be practiced otherwise than as specifically described and claimed. The present invention is directed to each individual feature, system, article, material, kit, and/or method described herein. In addition, any combination of 10 two or more such features, systems, articles, materials, kits, and/or methods, if such features, systems, articles, materials, kits, and/or methods are not mutually inconsistent, is included within the scope of the present invention.

15 All definitions, as defined and used herein, should be understood to control over dictionary definitions, definitions in documents incorporated by reference, and/or ordinary meanings of the defined terms.

The indefinite articles “a” and “an,” as used herein in the specification and in the claims, unless clearly indicated to the contrary, should be understood to mean “at least one.”

20 The phrase “and/or,” as used herein in the specification and in the claims, should be understood to mean “either or both” of the elements so conjoined, i.e., elements that are conjunctively present in some cases and disjunctively present in other cases. Multiple elements listed with “and/or” should be construed in the same fashion, i.e., “one or more” of the elements so conjoined. Other elements may optionally be present other than the elements specifically identified by the “and/or” clause, whether related or unrelated to those elements specifically identified. Thus, as a non-limiting example, a reference to “A and/or B”, when used in conjunction with open-ended language such as “comprising” can refer, in one embodiment, to A only (optionally including elements other than B); in another embodiment, to B only (optionally including elements other than A); in yet another embodiment, to both A and B (optionally including other elements); etc.

25 As used herein in the specification and in the claims, “or” should be understood to have the same meaning as “and/or” as defined above. For example, when separating items in a list, “or” or “and/or” shall be interpreted as being inclusive, i.e., the inclusion of at least one, but also

including more than one, of a number or list of elements, and, optionally, additional unlisted items. Only terms clearly indicated to the contrary, such as “only one of” or “exactly one of,” or, when used in the claims, “consisting of,” will refer to the inclusion of exactly one element of a number or list of elements. In general, the term “or” as used herein shall only be interpreted as 5 indicating exclusive alternatives (i.e. “one or the other but not both”) when preceded by terms of exclusivity, such as “either,” “one of,” “only one of,” or “exactly one of.” “Consisting essentially of,” when used in the claims, shall have its ordinary meaning as used in the field of patent law.

As used herein in the specification and in the claims, the phrase “at least one,” in 10 reference to a list of one or more elements, should be understood to mean at least one element selected from any one or more of the elements in the list of elements, but not necessarily including at least one of each and every element specifically listed within the list of elements and not excluding any combinations of elements in the list of elements. This definition also allows that elements may optionally be present other than the elements specifically identified within the 15 list of elements to which the phrase “at least one” refers, whether related or unrelated to those elements specifically identified. Thus, as a non-limiting example, “at least one of A and B” (or, equivalently, “at least one of A or B,” or, equivalently “at least one of A and/or B”) can refer, in one embodiment, to at least one, optionally including more than one, A, with no B present (and optionally including elements other than B); in another embodiment, to at least one, optionally 20 including more than one, B, with no A present (and optionally including elements other than A); in yet another embodiment, to at least one, optionally including more than one, A, and at least one, optionally including more than one, B (and optionally including other elements); etc.

When the word “about” is used herein in reference to a number, it should be understood 25 that still another embodiment of the invention includes that number not modified by the presence of the word “about.”

It should also be understood that, unless clearly indicated to the contrary, in any methods claimed herein that include more than one step or act, the order of the steps or acts of the method is not necessarily limited to the order in which the steps or acts of the method are recited.

In the claims, as well as in the specification above, all transitional phrases such as 30 “comprising,” “including,” “carrying,” “having,” “containing,” “involving,” “holding,” “composed of,” and the like are to be understood to be open-ended, i.e., to mean including but

not limited to. Only the transitional phrases “consisting of” and “consisting essentially of” shall be closed or semi-closed transitional phrases, respectively, as set forth in the United States Patent Office Manual of Patent Examining Procedures, Section 2111.03.

CLAIMS

What is claimed is:

1. A method, comprising:

providing droplets of a first fluid comprising polymer, surrounded by a second

5 fluid, contained in a carrier fluid;

solidifying the second fluid;

causing the polymer in the first fluid to form a gel; and

removing the solidified second fluid, thereby forming a suspension of the gel in
the carrier fluid.

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2. The method of claim 1, wherein causing the polymer to form a gel comprises applying ultraviolet light to cross-link the polymer to form the gel.

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3. The method of any one of claims 1 or 2, wherein causing the polymer to form a gel comprises applying TEMED to the carrier fluid, whereby the TEMED is able to diffuse into the first fluid.

20

4. The method of any one of claims 1-3, wherein the second fluid comprises a solvent and a second polymer, and solidifying the second fluid comprises causing the solvent to enter the carrier fluid such that the second polymer forms a solid.

25

5. The method of any one of claims 1-4, wherein solidifying the second fluid comprises solidifying the second fluid by altering the temperature of the second fluid.

6.

The method of any one of claims 1-5, wherein the solidified second fluid is a gel.

7. The method of any one of claims 1-6, wherein removing the solidified second fluid comprises reducing the pH of the carrier fluid such that the solidified second fluid dissolves in the carrier fluid.

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8. The method of claim 7, wherein altering the pH of the carrier fluid comprises reducing the pH of the carrier fluid by at least about 2 pH units.
9. The method of any one of claims 7 or 8, wherein altering the pH of the carrier fluid comprises reducing the pH of the carrier fluid to no more than 5.
10. The method of any one of claims 7-9, wherein altering the pH of the carrier fluid comprises adding an acid to the carrier fluid.
- 10 11. The method of any one of claims 1-10, wherein removing the solidified second fluid comprises mechanically removing the solidified second fluid.
12. The method of any one of claims 1-11, wherein the first fluid further comprises an initiator.
- 15 13. The method of claim 12, wherein the initiator comprises Irgacure 2559.
14. The method of any one of claims 1-13, wherein the polymer comprises poly(ethylene glycol) diacrylate.
- 20 15. The method of any one of claims 1-14, wherein the second fluid comprises an acrylate or a methacrylate.
16. The method of any one of claims 1-15, wherein the second fluid comprises dimethylaminoethyl methacrylate.
- 25 17. The method of any one of claims 1-16, wherein the second fluid comprises butyl methacrylate.
- 30 18. The method of any one of claims 1-17, wherein the second fluid comprises methyl methacrylate.

19. The method of any one of claims 1-18, wherein the droplets have an average diameter of no more than about 1 mm.
- 5 20. The method of any one of claims 1-19, wherein the droplets have a distribution of diameters such that at least 90% of the droplets have a diameter greater than 90% and less than 110% of the average diameter of the droplets.
- 10 21. The method of any one of claims 1-20, wherein the carrier fluid is aqueous.
22. The method of any one of claims 1-21, wherein the carrier fluid and the second fluid are substantially immiscible.
- 15 23. The method of any one of claims 1-22, wherein the first fluid and the second fluid are substantially immiscible.
24. The method of any one of claims 1-23, further comprising exposing a species to the gel to incorporate the species into the gel.
- 20 25. The method of claim 24, comprising bonding the species to the gel.
26. The method of claim 25, comprising covalently bonding the species to the gel.
27. The method of any one of claims 24-26, wherein the species comprises a nucleic acid.
- 25 28. The method of claim 27, wherein the species comprises DNA.
29. The method of claim 27 or 28, comprising bonding the DNA to the gel using an EDC coupling reaction.

30. A method, comprising:

providing droplets comprising solvent and polymer contained in a carrier fluid;

causing the polymer to form a gel; and

removing the solvent from the gel, thereby forming a suspension of the gel in the carrier fluid.

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31. The method of claim 30, wherein causing the polymer to form a gel comprises applying ultraviolet light to cross-link the polymer to form the gel.

10 32. The method of any one of claims 30 or 30, wherein causing the polymer to form a gel comprises applying TEMED to the carrier fluid, whereby the TEMED is able to diffuse into the first fluid.

15 33. The method of any one of claims 30-32, wherein the droplets further comprise an initiator.

34. The method of claim 33, wherein the initiator comprises Irgacure 2559.

20 35. The method of any one of claims 30-34, wherein the polymer comprises poly(ethylene glycol) diacrylate.

36. The method of any one of claims 30-35, wherein the solvent is able to form a separate phase when exposed to the carrier fluid.

25 37. The method of any one of claims 30-36, wherein removing the solvent from the gel comprises causing the solvent to enter the carrier fluid.

38. The method of any one of claims 30-37, wherein the carrier fluid is an aqueous fluid.

30 39. The method of any one of claims 30-38, wherein the carrier fluid is water.

40. The method of any one of claims 30-39, wherein the solvent has a water solubility of less than about 1 g/ml.
41. The method of any one of claims 30-40, further comprising exposing the gel to a species to the gel to incorporate the species into the gel.
5
42. The method of claim 41, comprising bonding the species to the gel.
43. The method of claim 42, comprising covalently bonding the species to the gel.
10
44. The method of any one of claims 30-43, wherein the species comprises a nucleic acid.
45. The method of claim 44, wherein the species comprises DNA.
15
46. The method of claim 44 or 45, comprising bonding the DNA to the gel using an EDC coupling reaction.

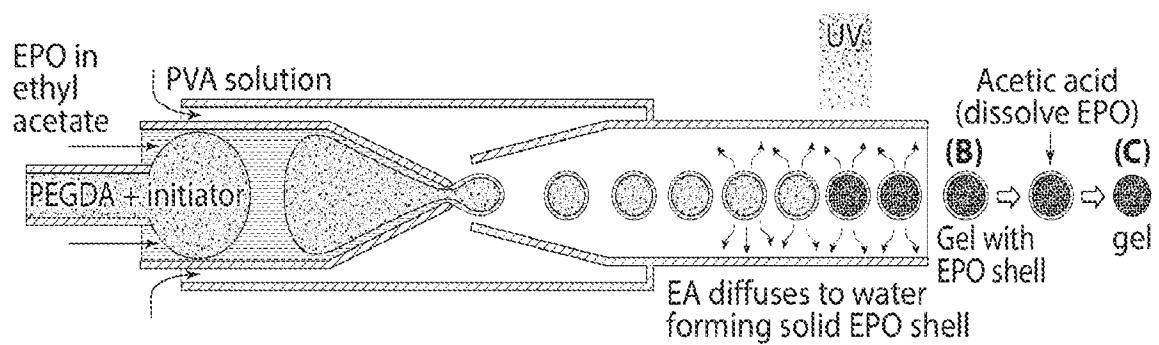


FIG. 1A

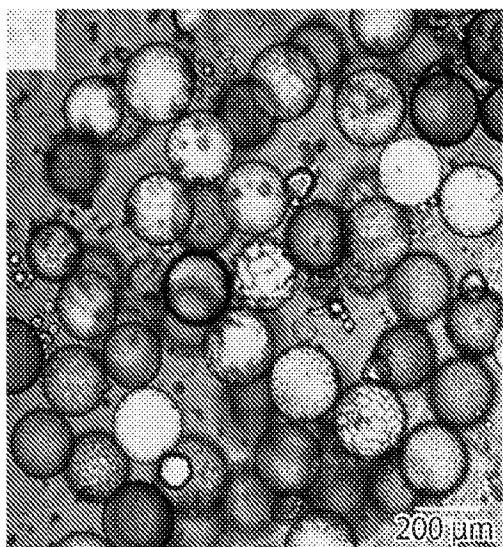


FIG. 1B

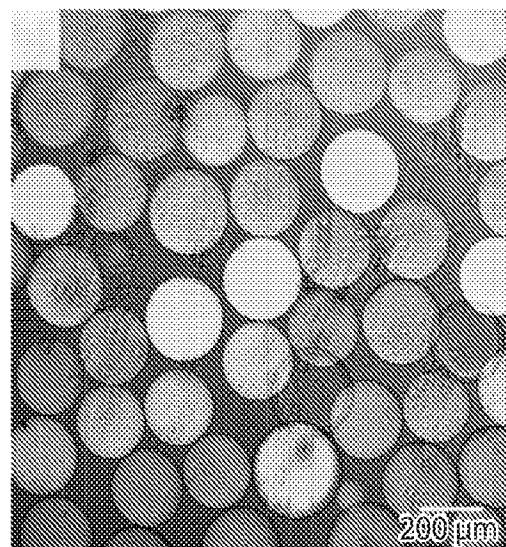


FIG. 1C

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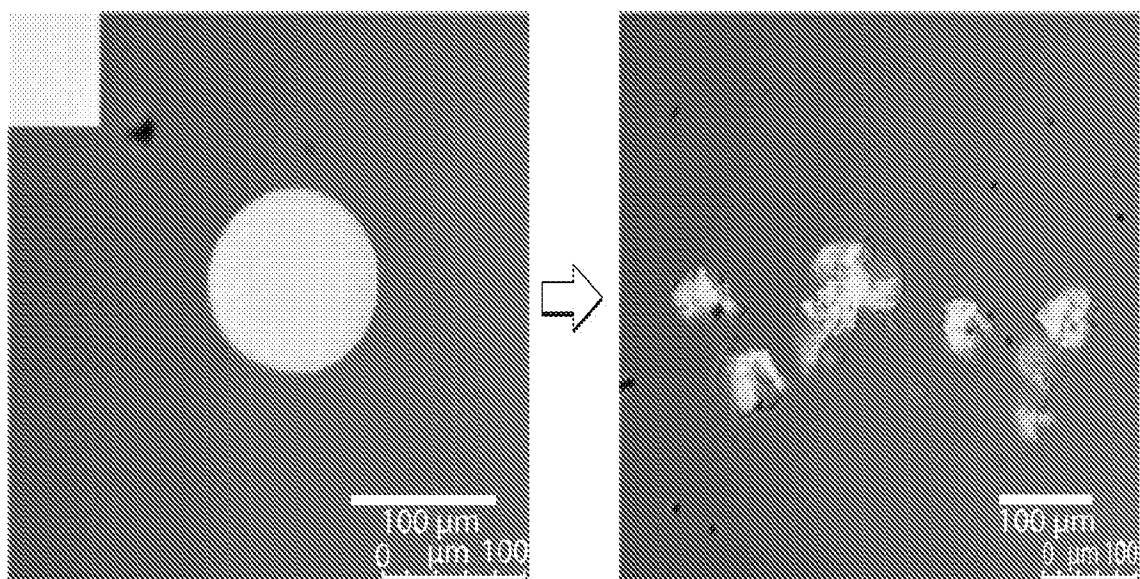


FIG. 1D

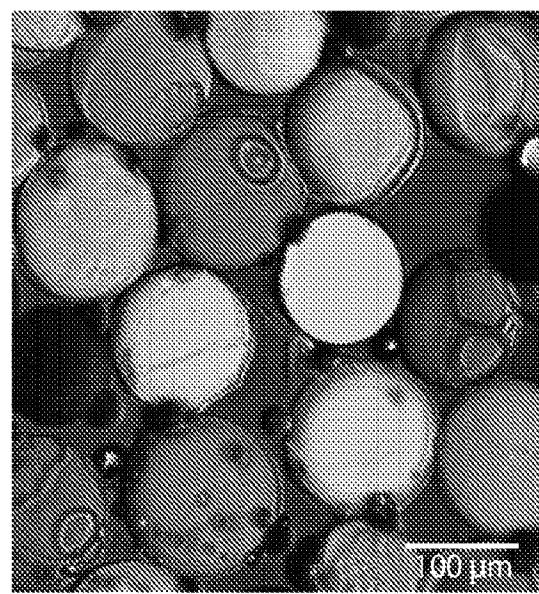


FIG. 1E

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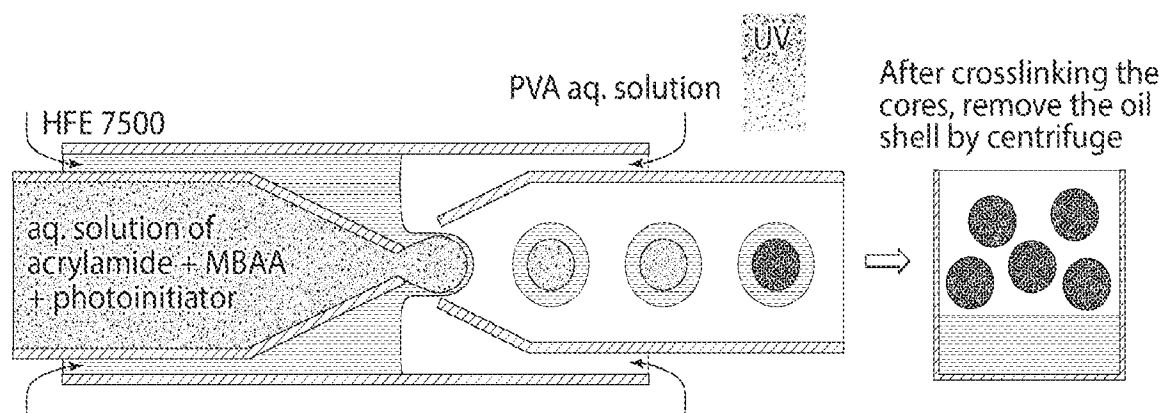


FIG. 2A

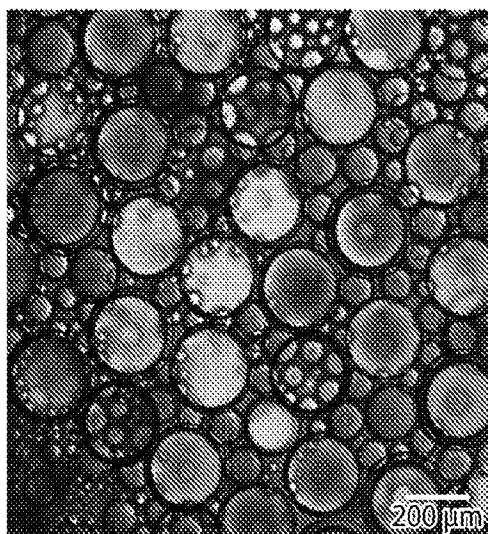


FIG. 2B

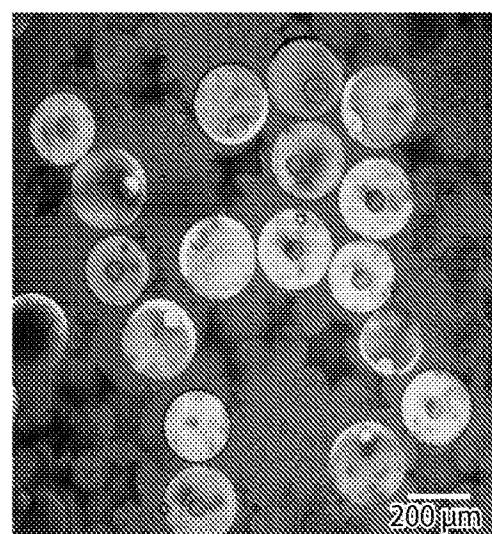


FIG. 2C

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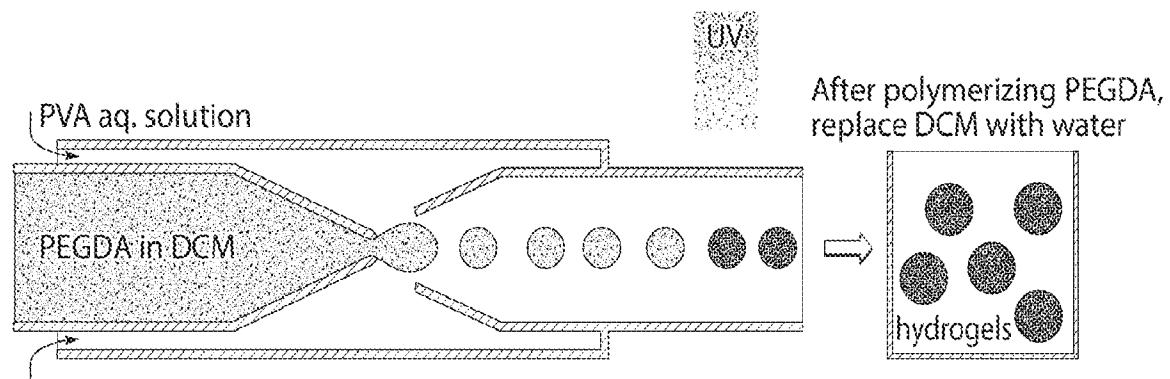


FIG. 3A

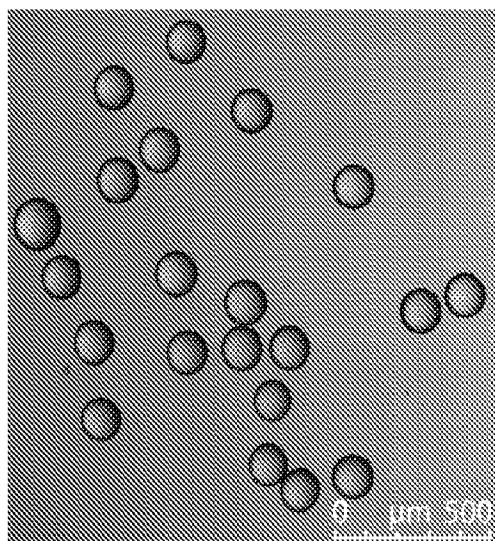


FIG. 3B

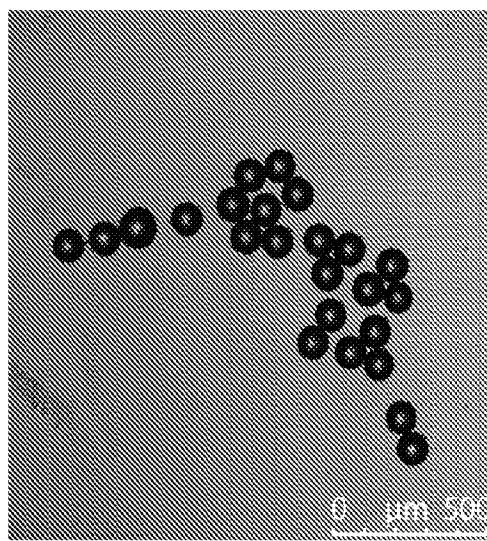


FIG. 3C

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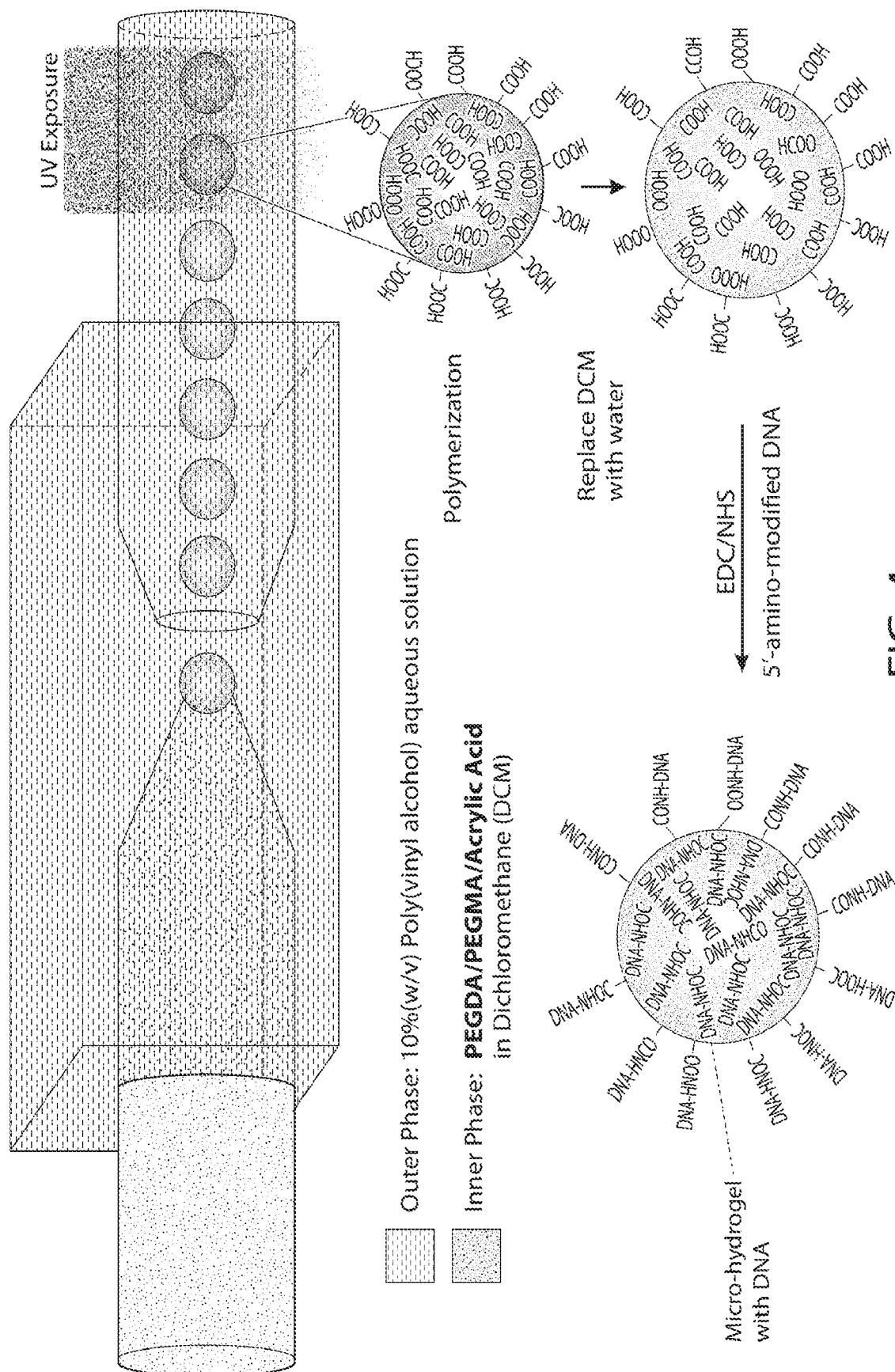


FIG. 4

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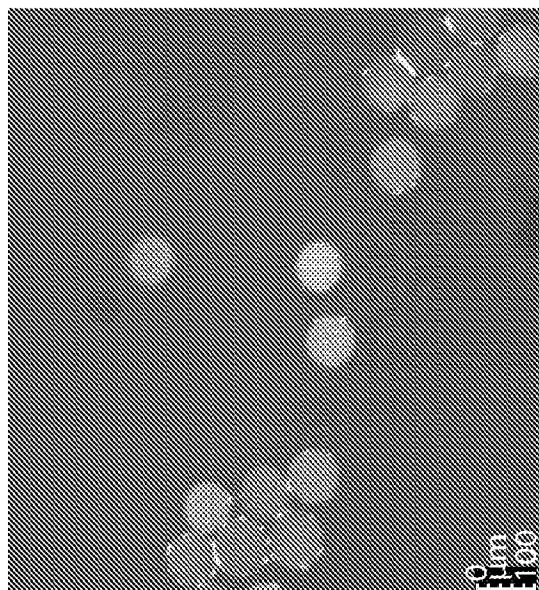


FIG. 5A

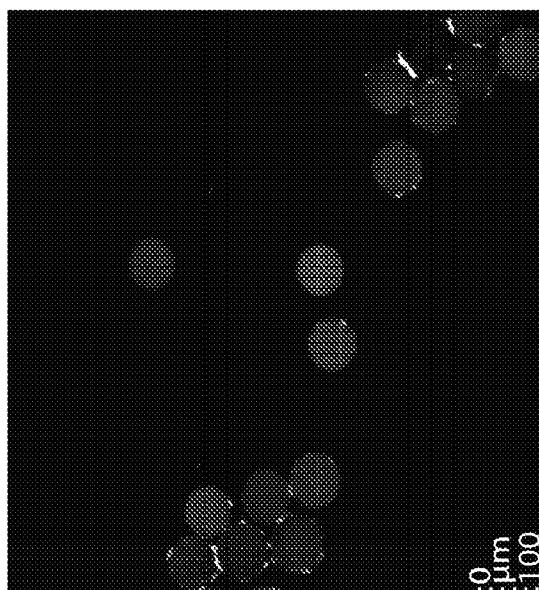


FIG. 5B

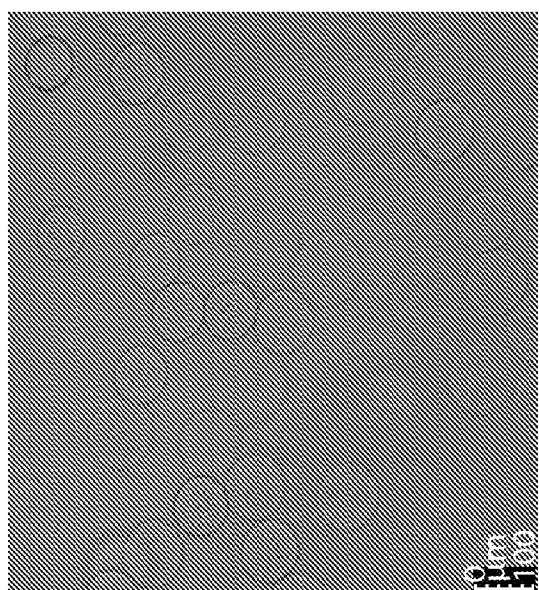


FIG. 5C



FIG. 5D