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(54) Titre : POLYNUCLEOTIDES ET POLYPEPTIDES ISOLES ET PROCEDES POUR LES UTILISER POUR AUGMENTER LE RENDEMENT, LA BIOMASSE, LA VITESSE DE CROISSANCE, LA VIGUEUR, LA TENEUR EN HUILE, LA TOLERANCE AU STRESS ABIOTIQUE DE PLANTES ET L'EFFICACITE D'UTILISATION DE L'AZOTE
 (54) Title: ISOLATED POLYNUCLEOTIDES AND POLYPEPTIDES AND METHODS OF USING SAME FOR INCREASING PLANT YIELD, BIOMASS, GROWTH RATE, VIGOR, OIL CONTENT, ABIOTIC STRESS TOLERANCE OF PLANTS AND NITROGEN USE EFFICIENCY

(57) **Abrégé/Abstract:**

Provided are methods of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant by expressing within the plant an exogenous polynucleotide comprising a nucleic acid sequence at least 80 % identical to SEQ ID NO: 905, 882, 1-12, 15-105, 203-297, 299- 523, 845- 881, 883-904, 906-925 or 933; or an exogenous polynucleotide encoding a polypeptide at least 80 % identical to SEQ ID NO: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 or 932. Also provided isolated polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NOs:905, 882, 1- 13, 15-105, 203-523, 845- 881, 883-904, 906-925 and 933, which can be used to increase yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant.

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(54) Title: ISOLATED POLYNUCLEOTIDES AND POLYPEPTIDES AND METHODS OF USING SAME FOR INCREASING PLANT YIELD

(57) Abstract: Provided are methods of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant by expressing within the plant an exogenous polynucleotide comprising a nucleic acid sequence at least 80 % identical to SEQ ID NO: 905, 882, 1-12, 15-105, 203-297, 299-523, 845-881, 883-904, 906-925 or 933; or an exogenous polynucleotide encoding a polypeptide at least 80 % identical to SEQ ID NO: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 or 932. Also provided isolated polynucleotide comprising a nucleic acid sequence selected from the group consisting of SEQ ID NOs: 905, 882, 1-13, 15-105, 203-523, 845-881, 883-904, 906-925 and 933, which can be used to increase yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant.



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ISOLATED POLYNUCLEOTIDES AND POLYPEPTIDES AND METHODS OF
USING SAME FOR INCREASING PLANT YIELD, BIOMASS, GROWTH RATE,
VIGOR, OIL CONTENT, ABIOTIC STRESS TOLERANCE OF PLANTS AND
NITROGEN USE EFFICIENCY

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FIELD AND BACKGROUND OF THE INVENTION

The present invention, in some embodiments thereof, relates to polypeptides, polynucleotides encoding same, transgenic plants expressing same and methods of producing and using same, and, more particularly, but not exclusively, to methods of increasing plant yield, oil yield, seed yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency.

Abiotic stress conditions such as salinity, drought, flood, suboptimal temperature and toxic chemical pollution, cause substantial damage to agricultural plants. Most plants have evolved strategies to protect themselves against these conditions. However, if the severity and duration of the stress conditions are too great, the effects on plant development, growth and yield of most crop plants are profound. Furthermore, most of the crop plants are highly susceptible to abiotic stress (ABS) and thus necessitate optimal growth conditions for commercial crop yields. Continuous exposure to stress causes major alterations in the plant metabolism which ultimately leads to cell death and consequently yield losses.

The global shortage of water supply is one of the most severe agricultural problems affecting plant growth and crop yield and efforts are made to mitigate the harmful effects of desertification and salinization of the world's arable land. Thus, Agbiotech companies attempt to create new crop varieties which are tolerant to different abiotic stresses focusing mainly in developing new varieties that can tolerate water shortage for longer periods.

Suboptimal nutrient (macro and micro nutrient) affect plant growth and development through the whole plant life cycle. One of the essential macronutrients for the plant is Nitrogen. Nitrogen is responsible for biosynthesis of amino acids and nucleic acids, prosthetic groups, plant hormones, plant chemical defenses, and the like. Nitrogen is often the rate-limiting element in plant growth and all field crops have a fundamental dependence on inorganic nitrogenous fertilizer. Since fertilizer is rapidly

depleted from most soil types, it must be supplied to growing crops two or three times during the growing season. Additional important macronutrients are Phosphorous (P) and Potassium (K), which have a direct correlation to yield and general plant tolerance.

Vegetable or seed oils are the major source of energy and nutrition in human and animal diet. They are also used for the production of industrial products, such as paints, inks and lubricants. In addition, plant oils represent renewable sources of long-chain hydrocarbons which can be used as fuel. Since the currently used fossil fuels are finite resources and are gradually being depleted, fast growing biomass crops may be used as alternative fuels or for energy feedstocks and may reduce the dependence on fossil energy supplies. However, the major bottleneck for increasing consumption of plant oils as bio-fuel is the oil price, which is still higher than fossil fuel. In addition, the production rate of plant oil is limited by the availability of agricultural land and water. Thus, increasing plant oil yields from the same growing area can effectively overcome the shortage in production space and can decrease vegetable oil prices at the same time.

Studies aiming at increasing plant oil yields focus on the identification of genes involved in oil metabolism as well as in genes capable of increasing plant and seed yields in transgenic plants.

Genes known to be involved in increasing plant oil yields include those participating in fatty acid synthesis or sequestering such as desaturase [e.g., DELTA6, DELTA12 or acyl-ACP (Ssi2; Arabidopsis Information Resource (TAIR; TAIR No. AT2G43710)], OleosinA (TAIR No. AT3G01570) or FAD3 (TAIR No. AT2G29980), and various transcription factors and activators such as Lec1 [TAIR No. AT1G21970, Lotan *et al.* 1998. *Cell.* 26;93(7):1195-205], Lec2 [TAIR No. AT1G28300, Santos Mendoza *et al.* 2005, *FEBS Lett.* 579(21):4666-70], Fus3 (TAIR No. AT3G26790), AB13 [TAIR No. AT3G24650, Lara *et al.* 2003. *J Biol Chem.* 278(23): 21003-11] and Wri1 [TAIR No. AT3G54320, Cernac and Benning, 2004. *Plant J.* 40(4): 575-85].

Zabrouskov V., et al., 2002 (Physiol Plant. 116:172-185) describe an increase in the total lipid fraction by upregulation of endoplasmic reticulum (FAD3) and plastidal (FAD7) fatty acid desaturases in potato.

5 Wang HW et al., 2007 (Plant J. 52:716-29. Epub 2007 Sep 18) describe an increase in the content of total fatty acids and lipids in plant seeds by over-expressing the GmDof4 and GmDof11 transcription factors.

Vigeolas H, et al. [Plant Biotechnol J. 2007, 5(3):431-41] and U.S. Pat. Appl. No. 20060168684 discloses an increase in seed oil content in oil-seed rape (*Brassica napus* L.) by over-expression of a yeast glycerol-3-phosphate dehydrogenase under the control of a seed-specific promoter.

10 Katavic V, et al., 2000 (Biochem Soc Trans. 28:935-7) describe the use of the *Arabidopsis* FAE1 and yeast SLC1-1 genes for improvements in erucic acid and oil content in rapeseed.

U.S. Pat. Appl. No. 20080076179 discloses an isolated moss nucleic acid encoding a lipid metabolism protein (LMP) and transgenic plants expressing same with increased lipid levels.

U.S. Pat. Appl. No. 20060206961 discloses a method of increasing oil content in plants (e.g., in plant seeds), by expressing in the plant the Ypr140w polypeptide.

20 U.S. Pat. Appl. No. 20060174373 discloses a method of increasing oil content in plants by expressing a nucleic acid encoding a triacylglycerols (TAG) synthesis enhancing protein (TEP) in the plant.

U.S. Pat. Appl. Nos. 20070169219, 20070006345, 20070006346 and 20060195943, disclose transgenic plants with improved nitrogen use efficiency which can be used for the conversion into fuel or chemical feedstocks.

25 WO2004/104162 teaches polynucleotide sequences and methods of utilizing same for increasing the tolerance of a plant to abiotic stresses and/or increasing the biomass of a plant.

WO2007/020638 teaches polynucleotide sequences and methods of utilizing same for increasing the tolerance of a plant to abiotic stresses and/or increasing the biomass, vigor and/or yield of a plant.

30 WO2008/122890 teaches polynucleotide sequences and methods of utilizing same for increasing oil content, growth rate, biomass, yield and/or vigor of a plant.

SUMMARY OF THE INVENTION

According to an aspect of some embodiments of the present invention there is provided a method of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant, comprising expressing within
5 the plant an exogenous polynucleotide comprising a nucleic acid sequence at least 80 % identical to SEQ ID NO: 905, 882, 1-12, 15-105, 203-297, 299-523, 845-881, 883-904, 906-925 or 933, thereby increasing the yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of the plant.

10 According to an aspect of some embodiments of the present invention there is provided a method of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant, comprising expressing within the plant an exogenous polynucleotide comprising the nucleic acid sequence selected from the group consisting of SEQ ID NOs: 905, 882, 1-13, 15-105, 203-523, 845-881,
15 883-904, 906-925 and 933, thereby increasing the yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of the plant.

According to an aspect of some embodiments of the present invention there is provided a method of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant, comprising expressing within
20 the plant an exogenous polynucleotide comprising a nucleic acid sequence encoding a polypeptide at least 80 % identical to SEQ ID NO: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 or 932, thereby increasing the yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of the plant.

25 According to an aspect of some embodiments of the present invention there is provided a method of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant, comprising expressing within the plant an exogenous polynucleotide comprising a nucleic acid sequence encoding a polypeptide selected from the group consisting of SEQ ID NOs: 172, 146, 106-117,
30 120-145, 147-171, 173-202, 524-844 and 926-932, thereby increasing the yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of the plant.

According to an aspect of some embodiments of the present invention there is provided an isolated polynucleotide comprising a nucleic acid sequence at least 80 % identical to SEQ ID NO: 905, 882, 1-12, 15-105, 203-297, 299-523, 845-881, 883-904, 906-925 or 933, wherein said nucleic acid sequence is capable of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant.

According to an aspect of some embodiments of the present invention there is provided an isolated polynucleotide comprising the nucleic acid sequence selected from the group consisting of SEQ ID NOs: 905, 882, 1-13, 15-105, 203-523, 845-881, 883-904, 906-925 and 933.

According to an aspect of some embodiments of the present invention there is provided an isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises an amino acid sequence at least 80 % homologous to the amino acid sequence set forth in SEQ ID NO: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 or 932, wherein said amino acid sequence is capable of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant.

According to an aspect of some embodiments of the present invention there is provided an isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises the amino acid sequence selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-844 and 926-932.

According to an aspect of some embodiments of the present invention there is provided a nucleic acid construct comprising the isolated polynucleotide of the invention, and a promoter for directing transcription of said nucleic acid sequence in a host cell.

According to an aspect of some embodiments of the present invention there is provided an isolated polypeptide comprising an amino acid sequence at least 80 % homologous to SEQ ID NO: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 or 932, wherein said amino acid sequence is capable of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant.

According to an aspect of some embodiments of the present invention there is provided an isolated polypeptide comprising the amino acid sequence selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-844 and 926-932.

5 According to an aspect of some embodiments of the present invention there is provided a plant cell exogenously expressing the polynucleotide of the invention, or the nucleic acid construct of the invention.

According to an aspect of some embodiments of the present invention there is provided a plant cell exogenously expressing the polypeptide of the invention.

10 According to some embodiments of the invention, the nucleic acid sequence is as set forth in SEQ ID NO: 905, 882, 1-13, 15-105, 203-523, 845-881, 883-904, 906-925 or 933.

According to some embodiments of the invention, the polynucleotide consists of the nucleic acid sequence selected from the group consisting of SEQ ID NOs: 905, 15 882, 1-13, 15-105, 203-523, 845-881, 883-904, 906-925 and 933.

According to some embodiments of the invention, the nucleic acid sequence encodes an amino acid sequence at least 80 % homologous to SEQ ID NO: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 or 932.

20 According to some embodiments of the invention, the nucleic acid sequence encodes the amino acid sequence selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-844 and 926-932.

According to some embodiments of the invention, the plant cell forms part of a plant.

25 According to some embodiments of the invention, the method further comprising growing the plant expressing said exogenous polynucleotide under the abiotic stress.

30 According to some embodiments of the invention, the abiotic stress is selected from the group consisting of salinity, drought, water deprivation, flood, etiolation, low temperature, high temperature, heavy metal toxicity, anaerobiosis, nutrient deficiency, nutrient excess, atmospheric pollution and UV irradiation.

According to some embodiments of the invention, the yield comprises seed yield or oil yield.

Unless otherwise defined, all technical and/or scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which the invention pertains. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of embodiments of the invention, exemplary methods and/or materials are described below. In case of conflict, the patent specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and are not intended to be necessarily limiting.

10 BRIEF DESCRIPTION OF THE DRAWINGS

Some embodiments of the invention are herein described, by way of example only, with reference to the accompanying drawings. With specific reference now to the drawings in detail, it is stressed that the particulars shown are by way of example and for purposes of illustrative discussion of embodiments of the invention. In this regard, the description taken with the drawings makes apparent to those skilled in the art how embodiments of the invention may be practiced.

In the drawings:

FIG. 1 is a schematic illustration of the pGI binary plasmid used for expressing the isolated polynucleotide sequences of some embodiments of the invention. RB - T-DNA right border; LB - T-DNA left border; H- *HindIII* restriction enzyme; X - *XbaI* restriction enzyme; B - *BamHI* restriction enzyme; S - *SalI* restriction enzyme; Sm - *SmaI* restriction enzyme; R-I - *EcoRI* restriction enzyme; Sc - *SacI/SstI/Ecl136II*; (numbers) - Length in base-pairs; NOS pro = nopaline synthase promoter; NPT-II = neomycin phosphotransferase gene; NOS ter = nopaline synthase terminator; Poly-A signal (polyadenylation signal); GUSintron - the GUS reporter gene (coding sequence and intron) The isolated polynucleotide sequences of the invention were cloned into the vector while replacing the GUSintron reporter gene

FIG. 2 is a schematic illustration of the modified pGI binary plasmid used for expressing the isolated polynucleotide sequences of the invention. RB - T-DNA right border; LB - T-DNA left border; MCS - Multiple cloning site; RE - any restriction enzyme; (numbers) - Length in base-pairs; NOS pro = nopaline synthase promoter; NPT-II = neomycin phosphotransferase gene; NOS ter = nopaline synthase terminator;

Poly-A signal (polyadenylation signal); GUSintron – the GUS reporter gene (coding sequence and intron) The isolated polynucleotide sequences of the invention were cloned into the vector while replacing the GUSintron reporter gene.

FIGs. 3A-F are images depicting visualization of root development of transgenic plants exogenously expressing the polynucleotide of some embodiments of the invention when grown in transparent agar plates under normal (Figures 3A-B), osmotic stress (15 % PEG; Figures 3C-D) or nitrogen-limiting (Figures 3E-F) conditions. The different transgenes were grown in transparent agar plates for 17 days (7 days nursery and 10 days after transplanting). The plates were photographed every 3-4 days starting at day 1 after transplanting. Figure 3A – An image of a photograph of plants taken following 10 after transplanting days on agar plates when grown under normal (standard) conditions. Figure 3B – An image of root analysis of the plants shown in Figure 3A in which the lengths of the roots measured are represented by arrows. Figure 3C – An image of a photograph of plants taken following 10 days after transplanting on agar plates, grown under high osmotic (PEG 15 %) conditions. Figure 3D – An image of root analysis of the plants shown in Figure 3C in which the lengths of the roots measured are represented by arrows. Figure 3E – An image of a photograph of plants taken following 10 days after transplanting on agar plates, grown under low nitrogen conditions. Figure 3F – An image of root analysis of the plants shown in Figure 3E in which the lengths of the roots measured are represented by arrows.

DESCRIPTION OF SPECIFIC EMBODIMENTS OF THE INVENTION

The present invention, in some embodiments thereof, relates to polypeptides, polynucleotides encoding same, nucleic acid constructs comprising same, transgenic plants expressing same and methods of producing and using same for increasing plant yield, oil yield, seed yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency.

Before explaining at least one embodiment of the invention in detail, it is to be understood that the invention is not necessarily limited in its application to the details set forth in the following description or exemplified by the Examples. The invention is capable of other embodiments or of being practiced or carried out in various ways.

While reducing the present invention to practice, the present inventors have identified novel polypeptides and polynucleotides which can be used to increase yield, growth rate, biomass, oil content, vigor, abiotic stress tolerance and/or nitrogen use efficiency of a plant.

5 Thus, as shown in the Examples section which follows, the present inventors have utilized bioinformatics tools and microarray analyses to identify polynucleotides which enhance yield (e.g., seed yield, oil yield, oil content), growth rate, biomass, vigor, abiotic stress tolerance and/or nitrogen use efficiency of a plant. Genes which affect the trait-of-interest [Table 15; Example 5 of the Examples section which follows; SEQ ID
10 NOs:1-105 (polynucleotides) and SEQ ID NOs:106-202 (polypeptides)] were identified based on correlation analyses between expression profiles of various genes in tissues, developmental stages, fertilizer limiting conditions, abiotic stress conditions or normal conditions across several Arabidopsis ecotypes, Sorghum Accessions and Tomato accessions and various parameters of yield, biomass, vigor, growth rate and/or oil
15 content (Examples 1, 2, 3, 4, 10 and 11 of the Examples section which follows; Tables 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 28 and 29). Homologous polypeptides and polynucleotides having the same function (activity) were also identified [Table 16, Example 6 of the Examples section which follows; SEQ ID NOs:203-523 (polynucleotides), and SEQ ID NOs:524-844 (polypeptides)]. The identified genes
20 were cloned in binary vectors (see for example, Tables 17, 18, 19 and 20; Example 7 of the Examples section which follows; SEQ ID NOs:47, 845-925 and 933) and transgenic plants over-expressing the identified genes of the invention were generated (see for example, Example 8 of the Examples section which follows). These plants which are transformed with the identified polynucleotides were found to exhibit increased yield,
25 biomass, growth rate, vigor, seed yield, oil content, oil yield, flowering, harvest index and rosette area (Tables 21-27; Example 9 of the Examples section which follows). These results suggest the use of the novel polynucleotides and polypeptides of the invention for increasing yield (including oil yield, seed yield and oil content), growth rate, biomass, vigor, abiotic stress tolerance and/or nitrogen use efficiency of a plant.

30 Thus, according to an aspect of some embodiments of the invention, there is provided method of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant. The method is effected by

expressing within the plant an exogenous polynucleotide comprising a nucleic acid sequence at least 80 % identical to SEQ ID NO: 905, 882, 1-12, 15-105, 203-297, 299-523, 845-881, 883-904, 906-925, or 933, thereby increasing the yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of the plant.

As used herein the phrase "plant yield" refers to the amount (as determined by weight or size) or quantity (numbers) of tissues or organs produced per plant or per growing season. Hence increased yield could affect the economic benefit one can obtain from the plant in a certain growing area and/or growing time.

It should be noted that a plant yield can be affected by various parameters including, but not limited to, plant biomass; plant vigor; growth rate; seed yield; seed or grain quantity; seed or grain quality; oil yield; content of oil, starch and/or protein in harvested organs (e.g., seeds or vegetative parts of the plant); number of flowers (florets) per panicle (expressed as a ratio of number of filled seeds over number of primary panicles); harvest index; number of plants grown per area; number and size of harvested organs per plant and per area; number of plants per growing area (density); number of harvested organs in field; total leaf area; carbon assimilation and carbon partitioning (the distribution/allocation of carbon within the plant); resistance to shade; number of harvestable organs (e.g. seeds), seeds per pod, weight per seed; and modified architecture [such as increase stalk diameter, thickness or improvement of physical properties (e.g. elasticity)] .

As used herein the phrase "seed yield" refers to the number or weight of the seeds per plant, seeds per pod, or per growing area or to the weight of a single seed, or to the oil extracted per seed. Hence seed yield can be affected by seed dimensions (e.g., length, width, perimeter, area and/or volume), number of (filled) seeds and seed filling rate and by seed oil content. Hence increase seed yield per plant could affect the economic benefit one can obtain from the plant in a certain growing area and/or growing time; and increase seed yield per growing area could be achieved by increasing seed yield per plant, and/or by increasing number of plants grown on the same given area.

The term "seed" (also referred to as "grain" or "kernel") as used herein refers to a small embryonic plant enclosed in a covering called the seed coat (usually with

some stored food), the product of the ripened ovule of gymnosperm and angiosperm plants which occurs after fertilization and some growth within the mother plant.

The phrase "oil content" as used herein refers to the amount of lipids in a given plant organ, either the seeds (seed oil content) or the vegetative portion of the plant (vegetative oil content) and is typically expressed as percentage of dry weight (10
5 % humidity of seeds) or wet weight (for vegetative portion).

It should be noted that oil content is affected by intrinsic oil production of a tissue (e.g., seed, vegetative portion), as well as the mass or size of the oil-producing tissue per plant or per growth period.

10 In one embodiment, increase in oil content of the plant can be achieved by increasing the size/mass of a plant's tissue(s) which comprise oil per growth period. Thus, increased oil content of a plant can be achieved by increasing the yield, growth rate, biomass and vigor of the plant.

As used herein the phrase "plant biomass" refers to the amount (e.g., measured
15 in grams of air-dry tissue) of a tissue produced from the plant in a growing season, which could also determine or affect the plant yield or the yield per growing area. An increase in plant biomass can be in the whole plant or in parts thereof such as aboveground (harvestable) parts, vegetative biomass, roots and seeds.

As used herein the phrase "growth rate" refers to the increase in plant
20 organ/tissue size per time (can be measured in cm^2 per day).

As used herein the phrase "plant vigor" refers to the amount (measured by weight) of tissue produced by the plant in a given time. Hence increased vigor could determine or affect the plant yield or the yield per growing time or growing area. In addition, early vigor (seed and/or seedling) results in improved field stand.

25 It should be noted that a plant yield can be determined under stress (e.g., abiotic stress, nitrogen-limiting conditions) and/or non-stress (normal) conditions.

As used herein, the phrase "non-stress conditions" refers to the growth conditions (e.g., water, temperature, light-dark cycles, humidity, salt concentration, fertilizer concentration in soil, nutrient supply such as nitrogen, phosphorous and/or
30 potassium), that do not significantly go beyond the everyday climatic and other abiotic conditions that plants may encounter, and which allow optimal growth, metabolism, reproduction and/or viability of a plant at any stage in its life cycle (e.g., in a crop plant

from seed to a mature plant and back to seed again). Persons skilled in the art are aware of normal soil conditions and climatic conditions for a given plant in a given geographic location. It should be noted that while the non-stress conditions may include some mild variations from the optimal conditions (which vary from one type/species of a plant to another), such variations do not cause the plant to cease growing without the capacity to resume growth.

The phrase "abiotic stress" as used herein refers to any adverse effect on metabolism, growth, reproduction and/or viability of a plant. Accordingly, abiotic stress can be induced by suboptimal environmental growth conditions such as, for example, salinity, water deprivation, flooding, freezing, low or high temperature, heavy metal toxicity, anaerobiosis, nutrient deficiency, atmospheric pollution or UV irradiation. The implications of abiotic stress are discussed in the Background section.

The phrase "abiotic stress tolerance" as used herein refers to the ability of a plant to endure an abiotic stress without suffering a substantial alteration in metabolism, growth, productivity and/or viability.

As used herein the phrase "water use efficiency (WUE)" refers to the level of organic matter produced per unit of water consumed by the plant, *i.e.*, the dry weight of a plant in relation to the plant's water use, *e.g.*, the biomass produced per unit transpiration.

As used herein the phrase "fertilizer use efficiency" refers to the metabolic process(es) which lead to an increase in the plant's yield, biomass, vigor, and growth rate per fertilizer unit applied. The metabolic process can be the uptake, spread, absorbent, accumulation, relocation (within the plant) and use of one or more of the minerals and organic moieties absorbed by the plant, such as nitrogen, phosphates and/or potassium.

As used herein the phrase "fertilizer-limiting conditions" refers to growth conditions which include a level (*e.g.*, concentration) of a fertilizer applied which is below the level needed for normal plant metabolism, growth, reproduction and/or viability.

As used herein the phrase "nitrogen use efficiency (NUE)" refers to the metabolic process(es) which lead to an increase in the plant's yield, biomass, vigor, and growth rate per nitrogen unit applied. The metabolic process can be the uptake, spread,

absorbent, accumulation, relocation (within the plant) and use of nitrogen absorbed by the plant.

As used herein the phrase “nitrogen-limiting conditions” refers to growth conditions which include a level (e.g., concentration) of nitrogen (e.g., ammonium or nitrate) applied which is below the level needed for normal plant metabolism, growth, reproduction and/or viability.

Improved plant NUE and FUE is translated in the field into either harvesting similar quantities of yield, while implementing less fertilizers, or increased yields gained by implementing the same levels of fertilizers. Thus, improved NUE or FUE has a direct effect on plant yield in the field. Thus, the polynucleotides and polypeptides of some embodiments of the invention positively affect plant yield, seed yield, and plant biomass. In addition, the benefit of improved plant NUE will certainly improve crop quality and biochemical constituents of the seed such as protein yield and oil yield.

As used herein the term "increasing" refers to at least about 2 %, at least about 3 %, at least about 4 %, at least about 5 %, at least about 10 %, at least about 15 %, at least about 20 %, at least about 30 %, at least about 40 %, at least about 50 %, at least about 60 %, at least about 70 %, at least about 80 %, increase in yield, growth rate, biomass, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant as compared to a native plant [*i.e.*, a plant not modified with the biomolecules (polynucleotide or polypeptides) of the invention, e.g., a non-transformed plant of the same species which is grown under the same growth conditions].

The phrase “expressing within the plant an exogenous polynucleotide” as used herein refers to upregulating the expression level of an exogenous polynucleotide within the plant by introducing the exogenous polynucleotide into a plant cell or plant and expressing by recombinant means, as further described herein below.

As used herein "expressing" refers to expression at the mRNA and optionally polypeptide level.

As used herein, the phrase "exogenous polynucleotide" refers to a heterologous nucleic acid sequence which may not be naturally expressed within the plant or which overexpression in the plant is desired. The exogenous polynucleotide may be introduced into the plant in a stable or transient manner, so as to produce a ribonucleic acid (RNA) molecule and/or a polypeptide molecule. It should be noted that

the exogenous polynucleotide may comprise a nucleic acid sequence which is identical or partially homologous to an endogenous nucleic acid sequence of the plant.

The term “endogenous” as used herein refers to any polynucleotide or polypeptide which is present and/or naturally expressed within a plant or a cell thereof.

5 According to some embodiments of the invention the exogenous polynucleotide comprises a nucleic acid sequence which is at least about 80 %, at least about 81 %, at least about 82 %, at least about 83 %, at least about 84 %, at least about 85 %, at least about 86 %, at least about 87 %, at least about 88 %, at least about 89 %, at least about 90 %, at least about 91 %, at least about 92 %, at least about 93 %, at least about 93 %, at least about 94 %, at least about 95 %, at least about 96 %, at least about 10 about 97 %, at least about 98 %, at least about 99 %, e.g., 100 % identical to the nucleic acid sequence selected from the group consisting of SEQ ID NOs: 905, 882, 1-12, 15-105, 203-297, 299-523, 845-881, 883-904, 906-925 and 933.

15 Identity (e.g., percent homology) can be determined using any homology comparison software, including for example, the BlastN software of the National Center of Biotechnology Information (NCBI) such as by using default parameters.

According to some embodiments of the invention the exogenous polynucleotide is at least about 80 %, at least about 81 %, at least about 82 %, at least about 83 %, at least about 84 %, at least about 85 %, at least about 86 %, at least about 20 87 %, at least about 88 %, at least about 89 %, at least about 90 %, at least about 91 %, at least about 92 %, at least about 93 %, at least about 93 %, at least about 94 %, at least about 95 %, at least about 96 %, at least about 97 %, at least about 98 %, at least about 99 %, e.g., 100 % identical to the polynucleotide selected from the group consisting of SEQ ID NOs: 905, 882, 1-12, 15-105, 203-297, 299-523, 845-881, 883-904, 906-925 25 and 933.

According to some embodiments of the invention the exogenous polynucleotide is set forth by SEQ ID NO: 905, 882, 1-12, 15-105, 203-297, 299-523, 845-881, 883-904, 906-925 or 933.

30 According to an aspect of some embodiments of the invention, there is provided a method of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant. The method is effected by expressing within the plant an exogenous polynucleotide comprising the nucleic acid

sequence selected from the group consisting of SEQ ID NOs: 905, 882, 1-13, 15-105, 203-523, 845-881, 883-904, 906-925 and 933, thereby increasing the yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of the plant.

According to some embodiments of the invention the exogenous polynucleotide is selected from the group consisting of SEQ ID NOs: 905, 882, 1-13, 15-105, 203-523, 845-881, 883-904, 906-925 and 933.

5 As used herein the term "polynucleotide" refers to a single or double stranded nucleic acid sequence which is isolated and provided in the form of an RNA sequence, a complementary polynucleotide sequence (cDNA), a genomic polynucleotide sequence and/or a composite polynucleotide sequences (e.g., a combination of the above).

The term "isolated" refers to at least partially separated from the natural
10 environment e.g., from a plant cell.

As used herein the phrase "complementary polynucleotide sequence" refers to a sequence, which results from reverse transcription of messenger RNA using a reverse transcriptase or any other RNA dependent DNA polymerase. Such a sequence can be subsequently amplified *in vivo* or *in vitro* using a DNA dependent DNA polymerase.

15 As used herein the phrase "genomic polynucleotide sequence" refers to a sequence derived (isolated) from a chromosome and thus it represents a contiguous portion of a chromosome.

As used herein the phrase "composite polynucleotide sequence" refers to a sequence, which is at least partially complementary and at least partially genomic. A
20 composite sequence can include some exonal sequences required to encode the polypeptide of the present invention, as well as some intronic sequences interposing therebetween. The intronic sequences can be of any source, including of other genes, and typically will include conserved splicing signal sequences. Such intronic sequences may further include cis acting expression regulatory elements.

25 According to some embodiments of the invention, the exogenous polynucleotide of the invention encodes a polypeptide having an amino acid sequence at least about 80 %, at least about 81 %, at least about 82 %, at least about 83 %, at least about 84 %, at least about 85 %, at least about 86 %, at least about 87 %, at least about 88 %, at least about 89 %, at least about 90 %, at least about 91 %, at least about 92 %, at least about 93 %, at least about 94 %, at least about 95 %, at least about 96 %, at least about 97 %, at least about 98 %, at least about 99 %, or at least about 100 % of the amino acid sequence of the polypeptide encoded by the genomic polynucleotide sequence.

at least about 93 %, at least about 94 %, at least about 95 %, at least about 96 %, at least about 97 %, at least about 98 %, at least about 99 %, or more say 100 % homologous to the amino acid sequence selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 and 932.

5 Homology (e.g., percent homology) can be determined using any homology comparison software, including for example, the BlastP or TBLASTN software of the National Center of Biotechnology Information (NCBI) such as by using default parameters, when starting from a polypeptide sequence; or the tBLASTX algorithm (available via the NCBI) such as by using default parameters, which compares the six-
10 frame conceptual translation products of a nucleotide query sequence (both strands) against a protein sequence database.

Homologous sequences include both orthologous and paralogous sequences. The term “paralogous” relates to gene-duplications within the genome of a species leading to paralogous genes. The term “orthologous” relates to homologous genes in
15 different organisms due to ancestral relationship.

One option to identify orthologues in monocot plant species is by performing a reciprocal blast search. This may be done by a first blast involving blasting the sequence-of-interest against any sequence database, such as the publicly available NCBI database. If orthologues in rice were sought, the sequence-of-interest would be blasted
20 against, for example, the 28,469 full-length cDNA clones from *Oryza sativa* Nipponbare available at NCBI. The blast results may be filtered. The full-length sequences of either the filtered results or the non-filtered results are then blasted back (second blast) against the sequences of the organism from which the sequence-of-interest is derived. The results of the first and second blasts are then compared. An orthologue is identified
25 when the sequence resulting in the highest score (best hit) in the first blast identifies in the second blast the query sequence (the original sequence-of-interest) as the best hit. Using the same rationale a paralogue (homolog to a gene in the same organism) is found. In case of large sequence families, the ClustalW program may be used, followed by a neighbor-joining tree which helps visualizing the clustering.

30 According to some embodiments of the invention, the exogenous polynucleotide encodes a polypeptide consisting of the amino acid sequence set forth by

SEQ ID NO: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 or 932.

According to an aspect of some embodiments of the invention, the method of increasing yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant is effected by expressing within the plant an exogenous polynucleotide comprising the nucleic acid sequence encoding a polypeptide selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-844 and 926-932, thereby increasing the yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance and/or nitrogen use efficiency of the plant.

Nucleic acid sequences encoding the polypeptides of the present invention may be optimized for expression. Examples of such sequence modifications include, but are not limited to, an altered G/C content to more closely approach that typically found in the plant species of interest, and the removal of codons atypically found in the plant species commonly referred to as codon optimization.

The phrase "codon optimization" refers to the selection of appropriate DNA nucleotides for use within a structural gene or fragment thereof that approaches codon usage within the plant of interest. Therefore, an optimized gene or nucleic acid sequence refers to a gene in which the nucleotide sequence of a native or naturally occurring gene has been modified in order to utilize statistically-preferred or statistically-favored codons within the plant. The nucleotide sequence typically is examined at the DNA level and the coding region optimized for expression in the plant species determined using any suitable procedure, for example as described in Sardana et al. (1996, Plant Cell Reports 15:677-681). In this method, the standard deviation of codon usage, a measure of codon usage bias, may be calculated by first finding the squared proportional deviation of usage of each codon of the native gene relative to that of highly expressed plant genes, followed by a calculation of the average squared deviation. The formula used is: $1 \text{ SDCU} = n = 1/N \sum [(X_n - Y_n) / Y_n]^2 / N$, where X_n refers to the frequency of usage of codon n in highly expressed plant genes, where Y_n to the frequency of usage of codon n in the gene of interest and N refers to the total number of codons in the gene of interest. A Table of codon usage from highly expressed genes of dicotyledonous plants is compiled using the data of Murray et al. (1989, Nuc Acids Res. 17:477-498).

One method of optimizing the nucleic acid sequence in accordance with the preferred codon usage for a particular plant cell type is based on the direct use, without performing any extra statistical calculations, of codon optimization Tables such as those provided on-line at the Codon Usage Database through the NIAS (National Institute of Agrobiological Sciences) DNA bank in Japan. The Codon Usage Database contains
5 codon usage tables for a number of different species, with each codon usage Table having been statistically determined based on the data present in Genbank.

By using the above Tables to determine the most preferred or most favored codons for each amino acid in a particular species (for example, rice), a naturally-
10 occurring nucleotide sequence encoding a protein of interest can be codon optimized for that particular plant species. This is effected by replacing codons that may have a low statistical incidence in the particular species genome with corresponding codons, in regard to an amino acid, that are statistically more favored. However, one or more less-favored codons may be selected to delete existing restriction sites, to create new ones at
15 potentially useful junctions (5' and 3' ends to add signal peptide or termination cassettes, internal sites that might be used to cut and splice segments together to produce a correct full-length sequence), or to eliminate nucleotide sequences that may negatively effect mRNA stability or expression.

The naturally-occurring encoding nucleotide sequence may already, in
20 advance of any modification, contain a number of codons that correspond to a statistically-favored codon in a particular plant species. Therefore, codon optimization of the native nucleotide sequence may comprise determining which codons, within the native nucleotide sequence, are not statistically-favored with regards to a particular plant, and modifying these codons in accordance with a codon usage table of the
25 particular plant to produce a codon optimized derivative. A modified nucleotide sequence may be fully or partially optimized for plant codon usage provided that the protein encoded by the modified nucleotide sequence is produced at a level higher than

the protein encoded by the corresponding naturally occurring or native gene. Construction of synthetic genes by altering the codon usage is described in for example PCT Patent Application 93/07278.

5 According to some embodiments of the invention, the exogenous polynucleotide is a non-coding RNA.

As used herein the phrase 'non-coding RNA' refers to an RNA molecule which does not encode an amino acid sequence (a polypeptide). Examples of such non-coding RNA molecules include, but are not limited to, an antisense RNA, a pre-miRNA (precursor of a microRNA), or a precursor of a Piwi-interacting RNA (piRNA).

10 A non-limiting example of a non-coding RNA polynucleotide is provided in SEQ ID NO:72 (BDL90).

Thus, the invention encompasses nucleic acid sequences described hereinabove; fragments thereof, sequences hybridizable therewith, sequences homologous thereto, sequences encoding similar polypeptides with different codon usage, altered sequences characterized by mutations, such as deletion, insertion or substitution of one or more nucleotides, either naturally occurring or man induced, either randomly or in a targeted fashion.

The invention provides an isolated polynucleotide comprising a nucleic acid sequence at least about 80 %, at least about 81 %, at least about 82 %, at least about 83
20 %, at least about 84 %, at least about 85 %, at least about 86 %, at least about 87 %, at least about 88 %, at least about 89 %, at least about 90 %, at least about 91 %, at least about 92 %, at least about 93 %, at least about 93 %, at least about 94 %, at least about 95 %, at least about 96 %, at least about 97 %, at least about 98 %, at least about 99 %, e.g., 100 % identical to the polynucleotide selected from the group consisting of SEQ ID
25 NOs: 905, 882, 1-12, 15-105, 203-297, 299-523, 845-881, 883-904, 906-925 and 933.

According to some embodiments of the invention the nucleic acid sequence is capable of increasing yield, growth rate, vigor, biomass, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant.

30 According to some embodiments of the invention the isolated polynucleotide comprising the nucleic acid sequence selected from the group consisting of SEQ ID NOs: 905, 882, 1-13, 15-105, 203-523, 845-881, 883-904, 906-925 and 933.

According to some embodiments of the invention the isolated polynucleotide is set forth by SEQ ID NO: 905, 882, 1-13, 15-105, 203-523, 845-881, 883-904, 906-925 or 933.

The invention provides an isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises an amino acid sequence at least about 80 %, at least about 81 %, at least about 82 %, at least about 83 %, at least about 84 %, at least about 85 %, at least about 86 %, at least about 87 %, at least about 88 %, at least about 89 %, at least about 90 %, at least about 91 %, at least about 92 %, at least about 93 %, at least about 93 %, at least about 94 %, at least about 95 %, at least about 96 %, at least about 97 %, at least about 98 %, at least about 99 %, or more say 100 % homologous to the amino acid sequence selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 and 932.

According to some embodiments of the invention the amino acid sequence is capable of increasing yield, growth rate, vigor, biomass, oil content, abiotic stress tolerance and/or nitrogen use efficiency of a plant.

The invention provides an isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide which comprises the amino acid sequence selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-844 and 926-932.

The invention provides an isolated polynucleotide comprising a nucleic acid sequence encoding a polypeptide selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-844 and 926-932.

The invention provides an isolated polypeptide comprising an amino acid sequence at least about 80 %, at least about 81 %, at least about 82 %, at least about 83 %, at least about 84 %, at least about 85 %, at least about 86 %, at least about 87 %, at least about 88 %, at least about 89 %, at least about 90 %, at least about 91 %, at least about 92 %, at least about 93 %, at least about 93 %, at least about 94 %, at least about 95 %, at least about 96 %, at least about 97 %, at least about 98 %, at least about 99 %, or more say 100 % homologous to an amino acid sequence selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-616, 621-844, 926-931 and 932.

According to some embodiments of the invention, the polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NOs: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-844 and 926-932.

5 According to some embodiments of the invention, the polypeptide is set forth by SEQ ID NO: 172, 146, 106-117, 120-145, 147-171, 173-202, 524-844, 926-931 or 932.

The invention also encompasses fragments of the above described polypeptides and polypeptides having mutations, such as deletions, insertions or substitutions of one or more amino acids, either naturally occurring or man induced,
10 either randomly or in a targeted fashion.

The term "plant" as used herein encompasses whole plants, ancestors and progeny of the plants and plant parts, including seeds, shoots, stems, roots (including tubers), and plant cells, tissues and organs. The plant may be in any form including suspension cultures, embryos, meristematic regions, callus tissue, leaves, gametophytes,
15 sporophytes, pollen, and microspores. Plants that are particularly useful in the methods of the invention include all plants which belong to the superfamily Viridiplantae, in particular monocotyledonous and dicotyledonous plants including a fodder or forage legume, ornamental plant, food crop, tree, or shrub selected from the list comprising
20 *Acacia* spp., *Acer* spp., *Actinidia* spp., *Aesculus* spp., *Agathis australis*, *Albizia amara*,
Alsophila tricolor, *Andropogon* spp., *Arachis* spp., *Areca catechu*, *Astelia fragrans*,
Astragalus cicer, *Baikiaea plurijuga*, *Betula* spp., *Brassica* spp., *Bruguiera gymnorrhiza*,
Burkea africana, *Butea frondosa*, *Cadaba farinosa*, *Calliandra* spp., *Camellia sinensis*,
Canna indica, *Capsicum* spp., *Cassia* spp., *Centroema pubescens*, *Chacoomeles* spp.,
25 *Cinnamomum cassia*, *Coffea arabica*, *Colophospermum mopane*, *Coronillia varia*,
Cotoneaster serotina, *Crataegus* spp., *Cucumis* spp., *Cupressus* spp., *Cyathea dealbata*,
Cydonia oblonga, *Cryptomeria japonica*, *Cymbopogon* spp., *Cynthea dealbata*, *Cydonia oblonga*,
Dalbergia monetaria, *Davallia divaricata*, *Desmodium* spp., *Dicksonia squarosa*,
Dibeteropogon amplexens, *Dioclea* spp., *Dolichos* spp., *Dorycnium rectum*,
Echinochloa pyramidalis, *Ehaffia* spp., *Eleusine coracana*, *Eragrestis* spp., *Erythrina*
30 spp., *Eucalypfus* spp., *Euclea schimperi*, *Eulalia villosa*, *Pagopyrum* spp., *Feijoa sellowiana*,
Fragaria spp., *Flemingia* spp., *Freycinetia banksli*, *Geranium thunbergii*,
GinAgo biloba, *Glycine javanica*, *Gliricidia* spp., *Gossypium hirsutum*, *Grevillea* spp.,

Guibourtia coleosperma, Hedysarum spp., Hemaffhia altissima, Heteropogon contoffus,
 Hordeum vulgare, Hyparrhenia rufa, Hypericum erectum, Hypeffhelia dissolute, Indigo
 incamata, Iris spp., Leptarrhenia pyrolifolia, Lespediza spp., Lettuca spp., Leucaena
 leucocephala, Loudetia simplex, Lotonus bainesli, Lotus spp., Macrotyloma axillare,
 5 Malus spp., Manihot esculenta, Medicago saliva, Metasequoia glyptostroboides, Musa
 sapientum, Nicotianum spp., Onobrychis spp., Ornithopus spp., Oryza spp.,
 Peltophorum africanum, Pennisetum spp., Persea gratissima, Petunia spp., Phaseolus
 spp., Phoenix canariensis, Phormium cookianum, Photinia spp., Picea glauca, Pinus
 spp., Pisum sativam, Podocarpus totara, Pogonarthria fleckii, Pogonaffhria squarrosa,
 10 Populus spp., Prosopis cineraria, Pseudotsuga menziesii, Pterolobium stellatum, Pyrus
 communis, Quercus spp., Rhaphiolepis umbellata, Rhopalostylis sapida, Rhus
 natalensis, Ribes grossularia, Ribes spp., Robinia pseudoacacia, Rosa spp., Rubus spp.,
 Salix spp., Schyzachyrium sanguineum, Sciadopitys vefficillata, Sequoia sempervirens,
 Sequoiadendron giganteum, Sorghum bicolor, Spinacia spp., Sporobolus fimbriatus,
 15 Stiburus alopecuroides, Stylosanthos humilis, Tadehagi spp, Taxodium distichum,
 Themeda triandra, Trifolium spp., Triticum spp., Tsuga heterophylla, Vaccinium spp.,
 Vicia spp., Vitis vinifera, Watsonia pyramidata, Zantedeschia aethiopica, Zea mays,
 amaranth, artichoke, asparagus, broccoli, Brussels sprouts, cabbage, canola, carrot,
 cauliflower, celery, collard greens, flax, kale, lentil, oilseed rape, okra, onion, potato,
 20 rice, soybean, straw, sugar beet, sugar cane, sunflower, tomato, squash tea, maize,
 wheat, barely, rye, oat, peanut, pea, lentil and alfalfa, cotton, rapeseed, canola, pepper,
 sunflower, tobacco, eggplant, eucalyptus, a tree, an ornamental plant, a perennial grass
 and a forage crop. Alternatively algae and other non-Viridiplantae can be used for the
 methods of the present invention.

25 According to some embodiments of the invention, the plant used by the
 method of the invention is a crop plant such as rice, maize, wheat, barley, peanut,
 potato, sesame, olive tree, palm oil, banana, soybean, sunflower, canola, sugarcane,
 alfalfa, millet, leguminosae (bean, pea), flax, lupinus, rapeseed, tobacco, poplar and
 cotton.

30 According to some embodiments of the invention, there is provided a plant
 cell exogenously expressing the polynucleotide of some embodiments of the invention,

the nucleic acid construct of some embodiments of the invention and/or the polypeptide of some embodiments of the invention.

According to some embodiments of the invention, expressing the exogenous polynucleotide of the invention within the plant is effected by transforming one or more cells of the plant with the exogenous polynucleotide, followed by generating a mature plant from the transformed cells and cultivating the mature plant under conditions suitable for expressing the exogenous polynucleotide within the mature plant.

According to some embodiments of the invention, the transformation is effected by introducing to the plant cell a nucleic acid construct which includes the exogenous polynucleotide of some embodiments of the invention and at least one promoter for directing transcription of the exogenous polynucleotide in a host cell (a plant cell). Further details of suitable transformation approaches are provided hereinbelow.

According to some embodiments of the invention, there is provided a nucleic acid construct comprising the isolated polynucleotide of the invention, and a promoter for directing transcription of the nucleic acid sequence of the isolated polynucleotide in a host cell.

According to some embodiments of the invention, the isolated polynucleotide is operably linked to the promoter sequence.

A coding nucleic acid sequence is “operably linked” to a regulatory sequence (e.g., promoter) if the regulatory sequence is capable of exerting a regulatory effect on the coding sequence linked thereto.

As used herein, the term “promoter” refers to a region of DNA which lies upstream of the transcriptional initiation site of a gene to which RNA polymerase binds to initiate transcription of RNA. The promoter controls where (e.g., which portion of a plant) and/or when (e.g., at which stage or condition in the lifetime of an organism) the gene is expressed.

Any suitable promoter sequence can be used by the nucleic acid construct of the present invention. Preferably the promoter is a constitutive promoter, a tissue-specific, or an abiotic stress-inducible promoter.

Suitable constitutive promoters include, for example, CaMV 35S promoter (SEQ ID NO:1184; Odell et al., Nature 313:810-812, 1985); Arabidopsis At6669

promoter (SEQ ID NO:1183; see PCT Publication No. WO04081173A2); maize Ubi 1 (Christensen et al., *Plant Sol. Biol.* 18:675-689, 1992); rice actin (McElroy et al., *Plant Cell* 2:163-171, 1990); pEMU (Last et al., *Theor. Appl. Genet.* 81:581-588, 1991); CaMV 19S (Nilsson et al., *Physiol. Plant* 100:456-462, 1997); GOS2 (de Pater et al, *Plant J* Nov;2(6):837-44, 1992); ubiquitin (Christensen et al, *Plant Mol. Biol.* 18: 675-689, 1992); Rice cyclophilin (Bucholz et al, *Plant Mol Biol.* 25(5):837-43, 1994); Maize H3 histone (Lepetit et al, *Mol. Gen. Genet.* 231: 276-285, 1992); Actin 2 (An et al, *Plant J.* 10(1):107-121, 1996) and Synthetic Super MAS (Ni et al., *The Plant Journal* 7: 661-76, 1995). Other constitutive promoters include those in U.S. Pat. Nos. 5,659,026, 5,608,149; 5,608,144; 5,604,121; 5,569,597; 5,466,785; 5,399,680; 5,268,463; and 5,608,142.

Suitable tissue-specific promoters include, but not limited to, leaf-specific promoters [such as described, for example, by Yamamoto et al., *Plant J.* 12:255-265, 1997; Kwon et al., *Plant Physiol.* 105:357-67, 1994; Yamamoto et al., *Plant Cell Physiol.* 35:773-778, 1994; Gotor et al., *Plant J.* 3:509-18, 1993; Orozco et al., *Plant Mol. Biol.* 23:1129-1138, 1993; and Matsuoka et al., *Proc. Natl. Acad. Sci. USA* 90:9586-9590, 1993], seed-preferred promoters [e.g., from seed specific genes (Simon, et al., *Plant Mol. Biol.* 5. 191, 1985; Scofield, et al., *J. Biol. Chem.* 262: 12202, 1987; Baszczyński, et al., *Plant Mol. Biol.* 14: 633, 1990), Brazil Nut albumin (Pearson' et al., *Plant Mol. Biol.* 18: 235- 245, 1992), legumin (Ellis, et al. *Plant Mol. Biol.* 10: 203-214, 1988), Glutelin (rice) (Takaiwa, et al., *Mol. Gen. Genet.* 208: 15-22, 1986; Takaiwa, et al., *FEBS Letts.* 221: 43-47, 1987), Zein (Matzke et al *Plant Mol Biol*, 143).323-32 1990), napA (Stalberg, et al, *Planta* 199: 515-519, 1996), Wheat SPA (Albanietal, *Plant Cell*, 9: 171- 184, 1997), sunflower oleosin (Cummins, etal., *Plant Mol. Biol.* 19: 873-876, 1992)], endosperm specific promoters [e.g., wheat LMW and HMW, glutenin-1 (*Mol Gen Genet* 216:81-90, 1989; *NAR* 17:461-2), wheat a, b and g gliadins (*EMBO3*:1409-15, 1984), Barley ltr1 promoter, barley B1, C, D hordein (*Theor Appl Gen* 98:1253-62, 1999; *Plant J* 4:343-55, 1993; *Mol Gen Genet* 250:750- 60, 1996), Barley DOF (Mena et al, *The Plant Journal*, 116(1): 53- 62, 1998), Biz2 (EP99106056.7), Synthetic promoter (Vicente-Carbajosa et al., *Plant J.* 13: 629-640, 1998), rice prolamin NRP33, rice -globulin Glb-1 (Wu et al, *Plant Cell Physiology* 39(8) 885- 889, 1998), rice alpha-globulin REB/OHP-1 (Nakase et al. *Plant Mol. Biol.*

33: 513-S22, 1997), rice ADP-glucose PP (Trans Res 6:157-68, 1997), maize ESR gene family (Plant J 12:235-46, 1997), sorghum gamma- kafirin (PMB 32:1029-35, 1996)], embryo specific promoters [e.g., rice OSH1 (Sato et al, Proc. Nati. Acad. Sci. USA, 93: 8117-8122), KNOX (Postma-Haarsma et al, Plant Mol. Biol. 39:257-71, 1999), rice oleosin (Wu et al, J. Biochem., 123:386, 1998)], and flower-specific promoters [e.g., AtPRP4, chalcone synthase (chsA) (Van der Meer, et al., Plant Mol. Biol. 15, 95-109, 1990), LAT52 (Twell et al Mol. Gen Genet. 217:240-245; 1989), apetala- 3].

Suitable abiotic stress-inducible promoters include, but not limited to, salt-inducible promoters such as RD29A (Yamaguchi-Shinozaki et al., Mol. Gen. Genet. 236:331-340, 1993); drought-inducible promoters such as maize rab17 gene promoter (Pla et. al., Plant Mol. Biol. 21:259-266, 1993), maize rab28 gene promoter (Busk et. al., Plant J. 11:1285-1295, 1997) and maize Ivr2 gene promoter (Pelleschi et. al., Plant Mol. Biol. 39:373-380, 1999); heat-inducible promoters such as heat tomato hsp80-promoter from tomato (U.S. Pat. No. 5,187,267).

The nucleic acid construct of some embodiments of the invention can further include an appropriate selectable marker and/or an origin of replication. According to some embodiments of the invention, the nucleic acid construct utilized is a shuttle vector, which can propagate both in *E. coli* (wherein the construct comprises an appropriate selectable marker and origin of replication) and be compatible with propagation in cells. The construct according to the present invention can be, for example, a plasmid, a bacmid, a phagemid, a cosmid, a phage, a virus or an artificial chromosome.

The nucleic acid construct of some embodiments of the invention can be utilized to stably or transiently transform plant cells. In stable transformation, the exogenous polynucleotide is integrated into the plant genome and as such it represents a stable and inherited trait. In transient transformation, the exogenous polynucleotide is expressed by the cell transformed but it is not integrated into the genome and as such it represents a transient trait.

There are various methods of introducing foreign genes into both monocotyledonous and dicotyledonous plants (Potrykus, I., Annu. Rev. Plant. Physiol., Plant. Mol. Biol. (1991) 42:205-225; Shimamoto et al., Nature (1989) 338:274-276).

The principle methods of causing stable integration of exogenous DNA into plant genomic DNA include two main approaches:

(i) Agrobacterium-mediated gene transfer: Klee et al. (1987) *Annu. Rev. Plant Physiol.* 38:467-486; Klee and Rogers in *Cell Culture and Somatic Cell Genetics of Plants*, Vol. 6, Molecular Biology of Plant Nuclear Genes, eds. Schell, J., and Vasil, L. K., Academic Publishers, San Diego, Calif. (1989) p. 2-25; Gatenby, in *Plant Biotechnology*, eds. Kung, S. and Arntzen, C. J., Butterworth Publishers, Boston, Mass. (1989) p. 93-112.

(ii) Direct DNA uptake: Paszkowski et al., in *Cell Culture and Somatic Cell Genetics of Plants*, Vol. 6, Molecular Biology of Plant Nuclear Genes eds. Schell, J., and Vasil, L. K., Academic Publishers, San Diego, Calif. (1989) p. 52-68; including methods for direct uptake of DNA into protoplasts, Toriyama, K. et al. (1988) *Bio/Technology* 6:1072-1074. DNA uptake induced by brief electric shock of plant cells: Zhang et al. *Plant Cell Rep.* (1988) 7:379-384. Fromm et al. *Nature* (1986) 319:791-793. DNA injection into plant cells or tissues by particle bombardment, Klein et al. *Bio/Technology* (1988) 6:559-563; McCabe et al. *Bio/Technology* (1988) 6:923-926; Sanford, *Physiol. Plant.* (1990) 79:206-209; by the use of micropipette systems: Neuhaus et al., *Theor. Appl. Genet.* (1987) 75:30-36; Neuhaus and Spangenberg, *Physiol. Plant.* (1990) 79:213-217; glass fibers or silicon carbide whisker transformation of cell cultures, embryos or callus tissue, U.S. Pat. No. 5,464,765 or by the direct incubation of DNA with germinating pollen, DeWet et al. in *Experimental Manipulation of Ovule Tissue*, eds. Chapman, G. P. and Mantell, S. H. and Daniels, W. Longman, London, (1985) p. 197-209; and Ohta, *Proc. Natl. Acad. Sci. USA* (1986) 83:715-719.

The Agrobacterium system includes the use of plasmid vectors that contain defined DNA segments that integrate into the plant genomic DNA. Methods of inoculation of the plant tissue vary depending upon the plant species and the Agrobacterium delivery system. A widely used approach is the leaf disc procedure which can be performed with any tissue explant that provides a good source for initiation of whole plant differentiation. See, e.g., Horsch et al. in *Plant Molecular Biology Manual A5*, Kluwer Academic Publishers, Dordrecht (1988) p. 1-9. A supplementary approach employs the Agrobacterium delivery system in combination

with vacuum infiltration. The Agrobacterium system is especially viable in the creation of transgenic dicotyledonous plants.

There are various methods of direct DNA transfer into plant cells. In electroporation, the protoplasts are briefly exposed to a strong electric field. In
5 microinjection, the DNA is mechanically injected directly into the cells using very small micropipettes. In microparticle bombardment, the DNA is adsorbed on microprojectiles such as magnesium sulfate crystals or tungsten particles, and the microprojectiles are physically accelerated into cells or plant tissues.

Following stable transformation plant propagation is exercised. The most
10 common method of plant propagation is by seed. Regeneration by seed propagation, however, has the deficiency that due to heterozygosity there is a lack of uniformity in the crop, since seeds are produced by plants according to the genetic variances governed by Mendelian rules. Basically, each seed is genetically different and each will grow with its own specific traits. Therefore, it is preferred that the transformed plant be
15 produced such that the regenerated plant has the identical traits and characteristics of the parent transgenic plant. Therefore, it is preferred that the transformed plant be regenerated by micropropagation which provides a rapid, consistent reproduction of the transformed plants.

Micropropagation is a process of growing new generation plants from a single
20 piece of tissue that has been excised from a selected parent plant or cultivar. This process permits the mass reproduction of plants having the preferred tissue expressing the fusion protein. The new generation plants which are produced are genetically identical to, and have all of the characteristics of, the original plant. Micropropagation allows mass production of quality plant material in a short period of time and offers a
25 rapid multiplication of selected cultivars in the preservation of the characteristics of the original transgenic or transformed plant. The advantages of cloning plants are the speed of plant multiplication and the quality and uniformity of plants produced.

Micropropagation is a multi-stage procedure that requires alteration of culture
medium or growth conditions between stages. Thus, the micropropagation process
30 involves four basic stages: Stage one, initial tissue culturing; stage two, tissue culture multiplication; stage three, differentiation and plant formation; and stage four, greenhouse culturing and hardening. During stage one, initial tissue culturing, the tissue

culture is established and certified contaminant-free. During stage two, the initial tissue culture is multiplied until a sufficient number of tissue samples are produced to meet production goals. During stage three, the tissue samples grown in stage two are divided and grown into individual plantlets. At stage four, the transformed plantlets are transferred to a greenhouse for hardening where the plants' tolerance to light is gradually increased so that it can be grown in the natural environment.

According to some embodiments of the invention, the transgenic plants are generated by transient transformation of leaf cells, meristematic cells or the whole plant.

Transient transformation can be effected by any of the direct DNA transfer methods described above or by viral infection using modified plant viruses.

Viruses that have been shown to be useful for the transformation of plant hosts include CaMV, Tobacco mosaic virus (TMV), brome mosaic virus (BMV) and Bean Common Mosaic Virus (BV or BCMV). Transformation of plants using plant viruses is described in U.S. Pat. No. 4,855,237 (bean golden mosaic virus; BGV), EP-A 67,553 (TMV), Japanese Published Application No. 63-14693 (TMV), EPA 194,809 (BV), EPA 278,667 (BV); and Gluzman, Y. et al., *Communications in Molecular Biology: Viral Vectors*, Cold Spring Harbor Laboratory, New York, pp. 172-189 (1988). Pseudovirus particles for use in expressing foreign DNA in many hosts, including plants are described in WO 87/06261.

According to some embodiments of the invention, the virus used for transient transformations is avirulent and thus is incapable of causing severe symptoms such as reduced growth rate, mosaic, ring spots, leaf roll, yellowing, streaking, pox formation, tumor formation and pitting. A suitable avirulent virus may be a naturally occurring avirulent virus or an artificially attenuated virus. Virus attenuation may be effected by using methods well known in the art including, but not limited to, sub-lethal heating, chemical treatment or by directed mutagenesis techniques such as described, for example, by Kurihara and Watanabe (*Molecular Plant Pathology* 4:259-269, 2003), Galon et al. (1992), Atreya et al. (1992) and Huet et al. (1994).

Suitable virus strains can be obtained from available sources such as, for example, the American Type Culture Collection (ATCC) or by isolation from infected plants. Isolation of viruses from infected plant tissues can be effected by techniques well known in the art such as described, for example by Foster and Tatlor, Eds. "Plant

Virology Protocols: From Virus Isolation to Transgenic Resistance (Methods in Molecular Biology (Humana Pr), Vol 81)", Humana Press, 1998. Briefly, tissues of an infected plant believed to contain a high concentration of a suitable virus, preferably young leaves and flower petals, are ground in a buffer solution (e.g., phosphate buffer solution) to produce a virus infected sap which can be used in subsequent inoculations.

5 Construction of plant RNA viruses for the introduction and expression of non-viral exogenous polynucleotide sequences in plants is demonstrated by the above references as well as by Dawson, W. O. et al., Virology (1989) 172:285-292; Takamatsu et al. EMBO J. (1987) 6:307-311; French et al. Science (1986) 231:1294-1297; 10 Takamatsu et al. FEBS Letters (1990) 269:73-76; and U.S. Pat. No. 5,316,931.

When the virus is a DNA virus, suitable modifications can be made to the virus itself. Alternatively, the virus can first be cloned into a bacterial plasmid for ease of constructing the desired viral vector with the foreign DNA. The virus can then be excised from the plasmid. If the virus is a DNA virus, a bacterial origin of replication 15 can be attached to the viral DNA, which is then replicated by the bacteria. Transcription and translation of this DNA will produce the coat protein which will encapsidate the viral DNA. If the virus is an RNA virus, the virus is generally cloned as a cDNA and inserted into a plasmid. The plasmid is then used to make all of the constructions. The RNA virus is then produced by transcribing the viral sequence of the 20 plasmid and translation of the viral genes to produce the coat protein(s) which encapsidate the viral RNA.

In one embodiment, a plant viral polynucleotide is provided in which the native coat protein coding sequence has been deleted from a viral polynucleotide, a non-native plant viral coat protein coding sequence and a non-native promoter, preferably 25 the subgenomic promoter of the non-native coat protein coding sequence, capable of expression in the plant host, packaging of the recombinant plant viral polynucleotide, and ensuring a systemic infection of the host by the recombinant plant viral polynucleotide, has been inserted. Alternatively, the coat protein gene may be inactivated by insertion of the non-native polynucleotide sequence within it, such that a 30 protein is produced. The recombinant plant viral polynucleotide may contain one or more additional non-native subgenomic promoters. Each non-native subgenomic promoter is capable of transcribing or expressing adjacent genes or polynucleotide

sequences in the plant host and incapable of recombination with each other and with native subgenomic promoters. Non-native (foreign) polynucleotide sequences may be inserted adjacent the native plant viral subgenomic promoter or the native and a non-native plant viral subgenomic promoters if more than one polynucleotide sequence is included. The non-native polynucleotide sequences are transcribed or expressed in the host plant under control of the subgenomic promoter to produce the desired products.

In a second embodiment, a recombinant plant viral polynucleotide is provided as in the first embodiment except that the native coat protein coding sequence is placed adjacent one of the non-native coat protein subgenomic promoters instead of a non-native coat protein coding sequence.

In a third embodiment, a recombinant plant viral polynucleotide is provided in which the native coat protein gene is adjacent its subgenomic promoter and one or more non-native subgenomic promoters have been inserted into the viral polynucleotide. The inserted non-native subgenomic promoters are capable of transcribing or expressing adjacent genes in a plant host and are incapable of recombination with each other and with native subgenomic promoters. Non-native polynucleotide sequences may be inserted adjacent the non-native subgenomic plant viral promoters such that the sequences are transcribed or expressed in the host plant under control of the subgenomic promoters to produce the desired product.

In a fourth embodiment, a recombinant plant viral polynucleotide is provided as in the third embodiment except that the native coat protein coding sequence is replaced by a non-native coat protein coding sequence.

The viral vectors are encapsidated by the coat proteins encoded by the recombinant plant viral polynucleotide to produce a recombinant plant virus. The recombinant plant viral polynucleotide or recombinant plant virus is used to infect appropriate host plants. The recombinant plant viral polynucleotide is capable of replication in the host, systemic spread in the host, and transcription or expression of foreign gene(s) (exogenous polynucleotide) in the host to produce the desired protein.

Techniques for inoculation of viruses to plants may be found in Foster and Taylor, eds. "Plant Virology Protocols: From Virus Isolation to Transgenic Resistance (Methods in Molecular Biology (Humana Pr), Vol 81)", Humana Press, 1998; Maramorosh and Koprowski, eds. "Methods in Virology" 7 vols, Academic Press, New

York 1967-1984; Hill, S.A. "Methods in Plant Virology", Blackwell, Oxford, 1984; Walkey, D.G.A. "Applied Plant Virology", Wiley, New York, 1985; and Kado and Agrawa, eds. "Principles and Techniques in Plant Virology", Van Nostrand-Reinhold, New York.

5 In addition to the above, the polynucleotide of the present invention can also be introduced into a chloroplast genome thereby enabling chloroplast expression.

 A technique for introducing exogenous polynucleotide sequences to the genome of the chloroplasts is known. This technique involves the following procedures. First, plant cells are chemically treated so as to reduce the number of chloroplasts per cell to about one. Then, the exogenous polynucleotide is introduced via
10 particle bombardment into the cells with the aim of introducing at least one exogenous polynucleotide molecule into the chloroplasts. The exogenous polynucleotides selected such that it is integratable into the chloroplast's genome via homologous recombination which is readily effected by enzymes inherent to the chloroplast. To this end, the
15 exogenous polynucleotide includes, in addition to a gene of interest, at least one polynucleotide stretch which is derived from the chloroplast's genome. In addition, the exogenous polynucleotide includes a selectable marker, which serves by sequential selection procedures to ascertain that all or substantially all of the copies of the chloroplast genomes following such selection will include the exogenous
20 polynucleotide. Further details relating to this technique are found in U.S. Pat. Nos. 4,945,050; and 5,693,507. A polypeptide can thus be produced by the protein expression system of the chloroplast and become integrated into the chloroplast's inner membrane.

 Since processes which increase yield, growth rate, biomass, vigor, oil content,
25 abiotic stress tolerance and/or nitrogen use efficiency of a plant can involve multiple genes acting additively or in synergy (see, for example, in Quesda et al., Plant Physiol. 130:951-063, 2002), the present invention also envisages expressing a plurality of exogenous polynucleotides in a single host plant to thereby achieve superior effect on the yield, growth rate, biomass, vigor, oil content, abiotic stress tolerance and/or
30 nitrogen use efficiency of the plant.

 Expressing a plurality of exogenous polynucleotides in a single host plant can be effected by co-introducing multiple nucleic acid constructs, each including a

different exogenous polynucleotide, into a single plant cell. The transformed cell can than be regenerated into a mature plant using the methods described hereinabove.

Alternatively, expressing a plurality of exogenous polynucleotides in a single host plant can be effected by co-introducing into a single plant-cell a single nucleic-acid
5 construct including a plurality of different exogenous polynucleotides. Such a construct can be designed with a single promoter sequence which can transcribe a polycistronic messenger RNA including all the different exogenous polynucleotide sequences. To enable co-translation of the different polypeptides encoded by the polycistronic messenger RNA, the polynucleotide sequences can be inter-linked via an internal
10 ribosome entry site (IRES) sequence which facilitates translation of polynucleotide sequences positioned downstream of the IRES sequence. In this case, a transcribed polycistronic RNA molecule encoding the different polypeptides described above will be translated from both the capped 5' end and the two internal IRES sequences of the polycistronic RNA molecule to thereby produce in the cell all different polypeptides.
15 Alternatively, the construct can include several promoter sequences each linked to a different exogenous polynucleotide sequence.

The plant cell transformed with the construct including a plurality of different exogenous polynucleotides, can be regenerated into a mature plant, using the methods described hereinabove.

20 Alternatively, expressing a plurality of exogenous polynucleotides in a single host plant can be effected by introducing different nucleic acid constructs, including different exogenous polynucleotides, into a plurality of plants. The regenerated transformed plants can then be cross-bred and resultant progeny selected for superior yield, growth rate, biomass, vigor, oil content, abiotic stress tolerance and/or nitrogen
25 use efficiency traits, using conventional plant breeding techniques.

According to some embodiments of the invention, the method further comprising growing the plant expressing the exogenous polynucleotide under the abiotic stress.

30 Non-limiting examples of abiotic stress conditions include, salinity, drought, water deprivation, excess of water (e.g., flood, waterlogging), etiolation, low temperature, high temperature, heavy metal toxicity, anaerobiosis, nutrient deficiency, nutrient excess, atmospheric pollution and UV irradiation.

Thus, the invention encompasses plants exogenously expressing the polynucleotide(s), the nucleic acid constructs and/or polypeptide(s) of the invention. Once expressed within the plant cell or the entire plant, the level of the polypeptide encoded by the exogenous polynucleotide can be determined by methods well known in the art such as, activity assays, Western blots using antibodies capable of specifically binding the polypeptide, Enzyme-Linked Immuno Sorbent Assay (ELISA), radio-immuno-assays (RIA), immunohistochemistry, immunocytochemistry, immunofluorescence and the like.

Methods of determining the level in the plant of the RNA transcribed from the exogenous polynucleotide are well known in the art and include, for example, Northern blot analysis, reverse transcription polymerase chain reaction (RT-PCR) analysis (including quantitative, semi-quantitative or real-time RT-PCR) and RNA-*in situ* hybridization.

In addition, the endogenous homolog of the exogenous polynucleotide or polypeptide of the invention, or a fragment of the endogenous homolog (e.g. introns or untranslated regions) in the plant can be used as a marker for marker assisted selection (MAS), in which a marker is used for indirect selection of a genetic determinant or determinants of a trait of interest (e.g., biomass, growth rate, oil content, yield, abiotic stress tolerance). These genes (DNA or RNA sequence) may contain or be linked to polymorphic sites or genetic markers on the genome such as restriction fragment length polymorphism (RFLP), microsatellites and single nucleotide polymorphism (SNP), DNA fingerprinting (DFP), amplified fragment length polymorphism (AFLP), expression level polymorphism, polymorphism of the encoded polypeptide and any other polymorphism at the DNA or RNA sequence.

Examples of marker assisted selections include, but are not limited to, selection for a morphological trait (e.g., a gene that affects form, coloration, male sterility or resistance such as the presence or absence of awn, leaf sheath coloration, height, grain color, aroma of rice); selection for a biochemical trait (e.g., a gene that encodes a protein that can be extracted and observed; for example, isozymes and storage proteins); selection for a biological trait (e.g., pathogen races or insect biotypes based on host pathogen or host parasite interaction can be used as a marker since the genetic constitution of an organism can affect its susceptibility to pathogens or parasites).

The polynucleotides and polypeptides described hereinabove can be used in a wide range of economical plants, in a safe and cost effective manner.

Plant lines exogenously expressing the polynucleotide or the polypeptide of the invention are screened to identify those that show the greatest increase of the desired
5 plant trait.

The effect of the transgene (the exogenous polynucleotide encoding the polypeptide) on abiotic stress tolerance can be determined using known methods such as detailed below and in the Examples section which follows.

Abiotic stress tolerance - Transformed (*i.e.*, expressing the transgene) and
10 non-transformed (wild type) plants are exposed to an abiotic stress condition, such as water deprivation, suboptimal temperature (low temperature, high temperature), nutrient deficiency, nutrient excess, a salt stress condition, osmotic stress, heavy metal toxicity, anaerobiosis, atmospheric pollution and UV irradiation.

Salinity tolerance assay - Transgenic plants with tolerance to high salt
15 concentrations are expected to exhibit better germination, seedling vigor or growth in high salt. Salt stress can be effected in many ways such as, for example, by irrigating the plants with a hyperosmotic solution, by cultivating the plants hydroponically in a hyperosmotic growth solution (e.g., Hoagland solution), or by culturing the plants in a hyperosmotic growth medium [e.g., 50 % Murashige-Skoog medium (MS medium)].
20 Since different plants vary considerably in their tolerance to salinity, the salt concentration in the irrigation water, growth solution, or growth medium can be adjusted according to the specific characteristics of the specific plant cultivar or variety, so as to inflict a mild or moderate effect on the physiology and/or morphology of the plants (for guidelines as to appropriate concentration see, Bernstein and Kafkafi, Root
25 Growth Under Salinity Stress In: Plant Roots, The Hidden Half 3rd ed. Waisel Y, Eshel A and Kafkafi U. (editors) Marcel Dekker Inc., New York, 2002, and reference therein).

For example, a salinity tolerance test can be performed by irrigating plants at different developmental stages with increasing concentrations of sodium chloride (for example 50 mM, 100 mM, 200 mM, 400 mM NaCl) applied from the bottom and from
30 above to ensure even dispersal of salt. Following exposure to the stress condition the plants are frequently monitored until substantial physiological and/or morphological effects appear in wild type plants. Thus, the external phenotypic appearance, degree of

wilting and overall success to reach maturity and yield progeny are compared between control and transgenic plants.

Quantitative parameters of tolerance measured include, but are not limited to, the average wet and dry weight, growth rate, leaf size, leaf coverage (overall leaf area), the weight of the seeds yielded, the average seed size and the number of seeds produced per plant. Transformed plants not exhibiting substantial physiological and/or morphological effects, or exhibiting higher biomass than wild-type plants, are identified as abiotic stress tolerant plants.

Osmotic tolerance test - Osmotic stress assays (including sodium chloride and mannitol assays) are conducted to determine if an osmotic stress phenotype was sodium chloride-specific or if it was a general osmotic stress related phenotype. Plants which are tolerant to osmotic stress may have more tolerance to drought and/or freezing. For salt and osmotic stress germination experiments, the medium is supplemented for example with 50 mM, 100 mM, 200 mM NaCl or 100 mM, 200 mM NaCl, 400 mM mannitol.

Drought tolerance assay/Osmoticum assay - Tolerance to drought is performed to identify the genes conferring better plant survival after acute water deprivation. To analyze whether the transgenic plants are more tolerant to drought, an osmotic stress produced by the non-ionic osmolyte sorbitol in the medium can be performed. Control and transgenic plants are germinated and grown in plant-agar plates for 4 days, after which they are transferred to plates containing 500 mM sorbitol. The treatment causes growth retardation, then both control and transgenic plants are compared, by measuring plant weight (wet and dry), yield, and by growth rates measured as time to flowering.

Conversely, soil-based drought screens are performed with plants overexpressing the polynucleotides detailed above. Seeds from control Arabidopsis plants, or other transgenic plants overexpressing the polypeptide of the invention are germinated and transferred to pots. Drought stress is obtained after irrigation is ceased accompanied by placing the pots on absorbent paper to enhance the soil-drying rate. Transgenic and control plants are compared to each other when the majority of the control plants develop severe wilting. Plants are re-watered after obtaining a significant fraction of the control plants displaying a severe wilting. Plants are ranked comparing to

controls for each of two criteria: tolerance to the drought conditions and recovery (survival) following re-watering.

Cold stress tolerance - To analyze cold stress, mature (25 day old) plants are transferred to 4 °C chambers for 1 or 2 weeks, with constitutive light. Later on plants are moved back to greenhouse. Two weeks later damages from chilling period, resulting in growth retardation and other phenotypes, are compared between both control and transgenic plants, by measuring plant weight (wet and dry), and by comparing growth rates measured as time to flowering, plant size, yield, and the like.

Heat stress tolerance - Heat stress tolerance is achieved by exposing the plants to temperatures above 34 °C for a certain period. Plant tolerance is examined after transferring the plants back to 22 °C for recovery and evaluation after 5 days relative to internal controls (non-transgenic plants) or plants not exposed to neither cold or heat stress.

Water use efficiency – can be determined as the biomass produced per unit transpiration. To analyze WUE, leaf relative water content can be measured in control and transgenic plants. Fresh weight (FW) is immediately recorded; then leaves are soaked for 8 hours in distilled water at room temperature in the dark, and the turgid weight (TW) is recorded. Total dry weight (DW) is recorded after drying the leaves at 60 °C to a constant weight. Relative water content (RWC) is calculated according to the following Formula I:

Formula I

$$\text{RWC} = [(\text{FW} - \text{DW}) / (\text{TW} - \text{DW})] \times 100$$

Fertilizer use efficiency - To analyze whether the transgenic plants are more responsive to fertilizers, plants are grown in agar plates or pots with a limited amount of fertilizer, as described, for example, in Yanagisawa et al (Proc Natl Acad Sci U S A. 2004; 101:7833-8). The plants are analyzed for their overall size, time to flowering, yield, protein content of shoot and/or grain. The parameters checked are the overall size of the mature plant, its wet and dry weight, the weight of the seeds yielded, the average seed size and the number of seeds produced per plant. Other parameters that may be tested are: the chlorophyll content of leaves (as nitrogen plant status and the degree of leaf verdure is highly correlated), amino acid and the total protein content of the seeds or other plant parts such as leaves or shoots, oil content, etc. Similarly, instead of

providing nitrogen at limiting amounts, phosphate or potassium can be added at increasing concentrations. Again, the same parameters measured are the same as listed above. In this way, nitrogen use efficiency (NUE), phosphate use efficiency (PUE) and potassium use efficiency (KUE) are assessed, checking the ability of the transgenic plants to thrive under nutrient restraining conditions.

Nitrogen use efficiency – To analyze whether the transgenic Arabidopsis plants are more responsive to nitrogen, plants are grown in 0.75- 1.5 mM (nitrogen deficient conditions) or 6-10 mM (optimal nitrogen concentration). Plants are allowed to grow for additional 20 days or until seed production. The plants are then analyzed for their overall size, time to flowering, yield, protein content of shoot and/or grain/seed production. The parameters checked can be the overall size of the plant, wet and dry weight, the weight of the seeds yielded, the average seed size and the number of seeds produced per plant. Other parameters that may be tested are: the chlorophyll content of leaves (as nitrogen plant status and the degree of leaf greenness is highly correlated), amino acid and the total protein content of the seeds or other plant parts such as leaves or shoots and oil content. Transformed plants not exhibiting substantial physiological and/or morphological effects, or exhibiting higher measured parameters levels than wild-type plants, are identified as nitrogen use efficient plants.

Nitrogen Use Efficiency assay using plantlets – The assay is done according to Yanagisawa-S. et al. with minor modifications (“Metabolic engineering with Dof1 transcription factor in plants: Improved nitrogen assimilation and growth under low-nitrogen conditions” *Proc. Natl. Acad. Sci. USA* 101, 7833-7838). Briefly, transgenic plants which are grown for 7-10 days in 0.5 x MS [Murashige-Skoog] supplemented with a selection agent are transferred to two nitrogen-limiting conditions: MS media in which the combined nitrogen concentration (NH_4NO_3 and KNO_3) was 0.2 mM or 0.05 mM. Plants are allowed to grow for additional 30-40 days and then photographed, individually removed from the Agar (the shoot without the roots) and immediately weighed (fresh weight) for later statistical analysis. Constructs for which only T1 seeds are available are sown on selective media and at least 25 seedlings (each one representing an independent transformation event) are carefully transferred to the nitrogen-limiting media. For constructs for which T2 seeds are available, different transformation events are analyzed. Usually, 25 randomly selected plants from each

event are transferred to the nitrogen-limiting media allowed to grow for 3-4 additional weeks and individually weighed at the end of that period. Transgenic plants are compared to control plants grown in parallel under the same conditions. Mock-transgenic plants expressing the uidA reporter gene (GUS) under the same promoter are used as control.

Nitrogen determination – The procedure for N (nitrogen) concentration determination in the structural parts of the plants involves the potassium persulfate digestion method to convert organic N to NO_3^- (Purcell and King 1996 Argon. J. 88:111-113, the modified Cd^+ mediated reduction of NO_3^- to NO_2^- (Vodovotz 1996 Biotechniques 20:390-394) and the measurement of nitrite by the Griess assay (Vodovotz 1996, supra). The absorbance values are measured at 550 nm against a standard curve of NaNO_2 . The procedure is described in details in Samonte et al. 2006 Agron. J. 98:168-176.

Germination tests - Germination tests compare the percentage of seeds from transgenic plants that could complete the germination process to the percentage of seeds from control plants that are treated in the same manner. Normal conditions are considered for example, incubations at 22 °C under 22-hour light 2-hour dark daily cycles. Evaluation of germination and seedling vigor is conducted between 4 and 14 days after planting. The basal media is 50 % MS medium (Murashige and Skoog, 1962 Plant Physiology 15, 473-497).

Germination is checked also at unfavorable conditions such as cold (incubating at temperatures lower than 10 °C instead of 22 °C) or using seed inhibition solutions that contain high concentrations of an osmolyte such as sorbitol (at concentrations of 50 mM, 100 mM, 200 mM, 300 mM, 500 mM, and up to 1000 mM) or applying increasing concentrations of salt (of 50 mM, 100 mM, 200 mM, 300 mM, 500 mM NaCl).

The effect of the transgene on plant's vigor, growth rate, biomass, yield and/or oil content can be determined using known methods.

Plant vigor - The plant vigor can be calculated by the increase in growth parameters such as leaf area, fiber length, rosette diameter, plant fresh weight and the like per time.

Growth rate - The growth rate can be measured using digital analysis of growing plants. For example, images of plants growing in greenhouse on plot basis can be captured every 3 days and the rosette area can be calculated by digital analysis. Rosette area growth is calculated using the difference of rosette area between days of sampling divided by the difference in days between samples.

Evaluation of growth rate can be done by measuring plant biomass produced, rosette area, leaf size or root length per time (can be measured in cm^2 per day of leaf area).

Relative growth rate area can be calculated using Formula II.

10

Formula II:

Relative growth area rate = Regression coefficient of area along time course.

Thus, the relative growth area rate is in units of 1/day and length growth rate is in units of 1/day.

Seed yield - Evaluation of the seed yield per plant can be done by measuring the amount (weight or size) or quantity (*i.e.*, number) of dry seeds produced and harvested from 8-16 plants and divided by the number of plants.

For example, the total seeds from 8-16 plants can be collected, weighted using e.g., an analytical balance and the total weight can be divided by the number of plants. Seed yield per growing area can be calculated in the same manner while taking into account the growing area given to a single plant. Increase seed yield per growing area could be achieved by increasing seed yield per plant, and/or by increasing number of plants capable of growing in a given area.

In addition, seed yield can be determined via the weight of 1000 seeds. The weight of 1000 seeds can be determined as follows: seeds are scattered on a glass tray and a picture is taken. Each sample is weighted and then using the digital analysis, the number of seeds in each sample is calculated.

The 1000 seeds weight can be calculated using formula III:

Formula III:

1000 Seed Weight = number of seed in sample/ sample weight X 1000

30

The Harvest Index can be calculated using Formula IV

Formula IV:

Harvest Index = Average seed yield per plant/ Average dry weight

Grain protein concentration - Grain protein content (g grain protein m⁻²) is estimated as the product of the mass of grain N (g grain N m⁻²) multiplied by the N/protein conversion ratio of k-5.13 (Mosse 1990, supra). The grain protein concentration is estimated as the ratio of grain protein content per unit mass of the grain
5 (g grain protein kg⁻¹ grain).

Fiber length - Fiber length can be measured using fibrograph. The fibrograph system was used to compute length in terms of "Upper Half Mean" length. The upper half mean (UHM) is the average length of longer half of the fiber distribution. The fibrograph measures length in span lengths at a given percentage point.

10 According to some embodiments of the invention, increased yield of corn may be manifested as one or more of the following: increase in the number of plants per growing area, increase in the number of ears per plant, increase in the number of rows per ear, number of kernels per ear row, kernel weight, thousand kernel weight (1000-weight), ear length/diameter, increase oil content per kernel and increase starch content
15 per kernel.

As mentioned, the increase of plant yield can be determined by various parameters. For example, increased yield of rice may be manifested by an increase in one or more of the following: number of plants per growing area, number of panicles per plant, number of spikelets per panicle, number of flowers per panicle, increase in the
20 seed filling rate, increase in thousand kernel weight (1000-weight), increase oil content per seed, increase starch content per seed, among others. An increase in yield may also result in modified architecture, or may occur because of modified architecture.

Similarly, increased yield of soybean may be manifested by an increase in one or more of the following: number of plants per growing area, number of pods per plant,
25 number of seeds per pod, increase in the seed filling rate, increase in thousand seed weight (1000-weight), reduce pod shattering, increase oil content per seed, increase protein content per seed, among others. An increase in yield may also result in modified architecture, or may occur because of modified architecture.

Increased yield of canola may be manifested by an increase in one or more of
30 the following: number of plants per growing area, number of pods per plant, number of

seeds per pod, increase in the seed filling rate, increase in thousand seed weight (1000-weight), reduce pod shattering, increase oil content per seed, among others. An increase in yield may also result in modified architecture, or may occur because of modified architecture.

5 Increased yield of cotton may be manifested by an increase in one or more of the following: number of plants per growing area, number of bolls per plant, number of seeds per boll, increase in the seed filling rate, increase in thousand seed weight (1000-weight), increase oil content per seed, improve fiber length, fiber strength, among others. An increase in yield may also result in modified architecture, or may occur
10 because of modified architecture.

Oil content - The oil content of a plant can be determined by extraction of the oil from the seed or the vegetative portion of the plant. Briefly, lipids (oil) can be removed from the plant (e.g., seed) by grinding the plant tissue in the presence of specific solvents (e.g., hexane or petroleum ether) and extracting the oil in a continuous
15 extractor. Indirect oil content analysis can be carried out using various known methods such as Nuclear Magnetic Resonance (NMR) Spectroscopy, which measures the resonance energy absorbed by hydrogen atoms in the liquid state of the sample [See for example, Conway TF. and Earle FR., 1963, Journal of the American Oil Chemists' Society; Springer Berlin / Heidelberg, ISSN: 0003-021X (Print) 1558-9331 (Online)];
20 the Near Infrared (NI) Spectroscopy, which utilizes the absorption of near infrared energy (1100-2500 nm) by the sample; and a method described in WO/2001/023884, which is based on extracting oil a solvent, evaporating the solvent in a gas stream which forms oil particles, and directing a light into the gas stream and oil particles which forms a detectable reflected light.

25 Thus, the present invention is of high agricultural value for promoting the yield of commercially desired crops (e.g., biomass of vegetative organ such as poplar wood, or reproductive organ such as number of seeds or seed biomass).

 Any of the transgenic plants described hereinabove or parts thereof may be processed to produce a feed, meal, protein or oil preparation, such as for ruminant
30 animals.

 The transgenic plants described hereinabove, which exhibit an increased oil content can be used to produce plant oil (by extracting the oil from the plant).

The plant oil (including the seed oil and/or the vegetative portion oil) produced according to the method of the invention may be combined with a variety of other ingredients. The specific ingredients included in a product are determined according to the intended use. Exemplary products include animal feed, raw material for chemical modification, biodegradable plastic, blended food product, edible oil, biofuel, cooking oil, lubricant, biodiesel, snack food, cosmetics, and fermentation process raw material. Exemplary products to be incorporated to the plant oil include animal feeds, human food products such as extruded snack foods, breads, as a food binding agent, aquaculture feeds, fermentable mixtures, food supplements, sport drinks, nutritional food bars, multi-vitamin supplements, diet drinks, and cereal foods.

According to some embodiments of the invention, the oil comprises a seed oil.

According to some embodiments of the invention, the oil comprises a vegetative portion oil.

According to some embodiments of the invention, the plant cell forms a part of a plant.

As used herein the term "about" refers to $\pm 10\%$.

The terms "comprises", "comprising", "includes", "including", "having" and their conjugates mean "including but not limited to".

The term "consisting of" means "including and limited to".

The term "consisting essentially of" means that the composition, method or structure may include additional ingredients, steps and/or parts, but only if the additional ingredients, steps and/or parts do not materially alter the basic and novel characteristics of the claimed composition, method or structure.

As used herein, the singular form "a", "an" and "the" include plural references unless the context clearly dictates otherwise. For example, the term "a compound" or "at least one compound" may include a plurality of compounds, including mixtures thereof.

Throughout this application, various embodiments of this invention may be presented in a range format. It should be understood that the description in range format is merely for convenience and brevity and should not be construed as an inflexible limitation on the scope of the invention. Accordingly, the description of a range should be considered to have specifically disclosed all the possible sub-ranges as well as

individual numerical values within that range. For example, description of a range such as from 1 to 6 should be considered to have specifically disclosed sub-ranges such as from 1 to 3, from 1 to 4, from 1 to 5, from 2 to 4, from 2 to 6, from 3 to 6 etc., as well as individual numbers within that range, for example, 1, 2, 3, 4, 5, and 6. This applies
5 regardless of the breadth of the range.

Whenever a numerical range is indicated herein, it is meant to include any cited numeral (fractional or integral) within the indicated range. The phrases “ranging/ranges between” a first indicate number and a second indicate number and “ranging/ranges from” a first indicate number “to” a second indicate number are used
10 herein interchangeably and are meant to include the first and second indicated numbers and all the fractional and integral numerals therebetween.

As used herein the term "method" refers to manners, means, techniques and procedures for accomplishing a given task including, but not limited to, those manners, means, techniques and procedures either known to, or readily developed from known
15 manners, means, techniques and procedures by practitioners of the chemical, pharmacological, biological, biochemical and medical arts.

It is appreciated that certain features of the invention, which are, for clarity, described in the context of separate embodiments, may also be provided in combination in a single embodiment. Conversely, various features of the invention, which are, for
20 brevity, described in the context of a single embodiment, may also be provided separately or in any suitable subcombination or as suitable in any other described embodiment of the invention. Certain features described in the context of various embodiments are not to be considered essential features of those embodiments, unless the embodiment is inoperative without those elements.

25 Various embodiments and aspects of the present invention as delineated hereinabove and as claimed in the claims section below find experimental support in the following examples.

Unless otherwise defined, all technical and/or scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which
30 the invention pertains. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of embodiments of the invention, exemplary methods and/or materials are described below. In case of conflict, the patent

specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and are not intended to be necessarily limiting.

EXAMPLES

Reference is now made to the following examples, which together with the
5 above descriptions illustrate some embodiments of the invention in a non limiting fashion.

Generally, the nomenclature used herein and the laboratory procedures utilized in the present invention include molecular, biochemical, microbiological and recombinant DNA techniques. Such techniques are thoroughly explained in the
10 literature. See, for example, "Molecular Cloning: A laboratory Manual" Sambrook et al., (1989); "Current Protocols in Molecular Biology" Volumes I-III Ausubel, R. M., ed. (1994); Ausubel et al., "Current Protocols in Molecular Biology", John Wiley and Sons, Baltimore, Maryland (1989); Perbal, "A Practical Guide to Molecular Cloning", John Wiley & Sons, New York (1988); Watson et al., "Recombinant DNA", Scientific
15 American Books, New York; Birren et al. (eds) "Genome Analysis: A Laboratory Manual Series", Vols. 1-4, Cold Spring Harbor Laboratory Press, New York (1998); methodologies as set forth in U.S. Pat. Nos. 4,666,828; 4,683,202; 4,801,531; 5,192,659 and 5,272,057; "Cell Biology: A Laboratory Handbook", Volumes I-III Cellis, J. E., ed. (1994); "Current Protocols in Immunology" Volumes I-III Coligan J. E., ed. (1994);
20 Stites et al. (eds), "Basic and Clinical Immunology" (8th Edition), Appleton & Lange, Norwalk, CT (1994); Mishell and Shiigi (eds), "Selected Methods in Cellular Immunology", W. H. Freeman and Co., New York (1980); available immunoassays are extensively described in the patent and scientific literature, see, for example, U.S. Pat. Nos. 3,791,932; 3,839,153; 3,850,752; 3,850,578; 3,853,987; 3,867,517; 3,879,262;
25 3,901,654; 3,935,074; 3,984,533; 3,996,345; 4,034,074; 4,098,876; 4,879,219; 5,011,771 and 5,281,521; "Oligonucleotide Synthesis" Gait, M. J., ed. (1984); "Nucleic Acid Hybridization" Hames, B. D., and Higgins S. J., eds. (1985); "Transcription and Translation" Hames, B. D., and Higgins S. J., Eds. (1984); "Animal Cell Culture" Freshney, R. I., ed. (1986); "Immobilized Cells and Enzymes" IRL Press, (1986); "A
30 Practical Guide to Molecular Cloning" Perbal, B., (1984) and "Methods in Enzymology" Vol. 1-317, Academic Press; "PCR Protocols: A Guide To Methods And Applications", Academic Press, San Diego, CA (1990); Marshak et al., "Strategies for

Protein Purification and Characterization - A Laboratory Course Manual" CSHL Press (1996). Other general references are provided throughout this document. The procedures therein are believed to be well known in the art and are provided for the convenience of the reader.

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EXAMPLE 1
GENE IDENTIFICATION AND GENE ROLE PREDICTION USING
BIOINFORMATICS TOOLS

10 The present inventors have identified polynucleotides which can increase plant yield, seed yield, oil yield, oil content, biomass, growth rate, abiotic stress tolerance, nitrogen use efficiency and/or vigor of a plant, as follows.

 The nucleotide sequence datasets used here were from publicly available databases or from sequences obtained using the Solexa technology (e.g. Barley and
15 Sorghum). Sequence data from 100 different plant species was introduced into a single, comprehensive database. Other information on gene expression, protein annotation, enzymes and pathways were also incorporated. Major databases used include:

Genomes

 Arabidopsis genome [TAIR genome version 6];
20 Rice genome [IRGSP build 4.0];
 Poplar [Populus trichocarpa release 1.1 from JGI (assembly release v1.0)];
 Brachypodium [JGI 4x assembly];
 Soybean [DOE-JGI SCP, version Glyma0];
 Grape [French-Italian Public Consortium for Grapevine Genome
25 Characterization grapevine genome];
 Castobean [TIGR/J Craig Venter Institute 4x assembly [communis];
 Sorghum [DOE-JGI SCP, version Sbi1];
 Partially assembled genome of Maize;

Expressed EST and mRNA sequences were extracted from the following
30 ***databases:***

 GenBank versions 154, 157, 160, 161, 164, 165, 166 and 168;
 RefSeq;

TAIR;

Protein and pathway databases

Uniprot.

AraCyc.

5 ENZYME.

Microarray datasets were downloaded from:

GEO

TAIR.

Proprietary microarray data (See WO2008/122980 and Example 3 below).

10 ***QTL and SNPs information***

Gramene.

Panzea.

15 ***Database Assembly*** - was performed to build a wide, rich, reliable annotated and easy to analyze database comprised of publicly available genomic mRNA, ESTs DNA sequences, data from various crops as well as gene expression, protein annotation and pathway data QTLs, and other relevant information.

20 Database assembly is comprised of a toolbox of gene refining, structuring, annotation and analysis tools enabling to construct a tailored database for each gene discovery project. Gene refining and structuring tools enable to reliably detect splice variants and antisense transcripts, generating understanding of various potential phenotypic outcomes of a single gene. The capabilities of the "LEADS" platform of Compugen LTD for analyzing human genome have been confirmed and accepted by the scientific community [see e.g., "Widespread Antisense Transcription", Yelin, et al. (2003) Nature Biotechnology 21, 379-85; "Splicing of Alu Sequences", Lev-Maor, et al. (2003) Science 300 (5623), 1288-91; "Computational analysis of alternative splicing using EST tissue information", Xie H et al. Genomics 2002], and have been proven most efficient in plant genomics as well.

30 ***EST clustering and gene assembly*** - For gene clustering and assembly of organisms with available genome sequence data (arabidopsis, rice, castorbean, grape, brachypodium, poplar, soybean, sorghum) the genomic LEADS version (GANG) was employed. This tool allows most accurate clustering of ESTs and mRNA sequences on

genome, and predicts gene structure as well as alternative splicing events and anti-sense transcription.

For organisms with no available full genome sequence data, "expressed LEADS" clustering software was applied.

5 **Gene annotation** - Predicted genes and proteins were annotated as follows:

Blast search against all plant UniProt sequences was performed. Open reading frames of each putative transcript were analyzed and longest ORF with higher number of homologues was selected as predicted protein of the transcript. The predicted proteins were analyzed by InterPro.

10 Blast against proteins from AraCyc and ENZYME databases was used to map the predicted transcripts to AraCyc pathways.

Predicted proteins from different species were compared using blast algorithm to validate the accuracy of the predicted protein sequence, and for efficient detection of orthologs.

15 **Gene expression profiling** - Several data sources were exploited for gene expression profiling which combined microarray data and digital expression profile (see below). According to gene expression profile, a correlation analysis was performed to identify genes which are co-regulated under different developmental stages and environmental conditions and which are associated with different phenotypes.

20 Publicly available microarray datasets were downloaded from TAIR and NCBI GEO sites, renormalized, and integrated into the database. Expression profiling is one of the most important resource data for identifying genes important for yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance of plants and nitrogen use efficiency.

25 A digital expression profile summary was compiled for each cluster according to all keywords included in the sequence records comprising the cluster. Digital expression, also known as electronic Northern Blot, is a tool that displays virtual expression profile based on the EST sequences forming the gene cluster. The tool provides the expression profile of a cluster in terms of plant anatomy (e.g., the
30 tissue/organ in which the gene is expressed), developmental stage (the developmental stages at which a gene can be found) and profile of treatment (provides the physiological conditions under which a gene is expressed such as drought, cold,

pathogen infection, etc). Given a random distribution of ESTs in the different clusters, the digital expression provides a probability value that describes the probability of a cluster having a total of N ESTs to contain X ESTs from a certain collection of libraries. For the probability calculations, the following is taken into consideration: a) the number
5 of ESTs in the cluster, b) the number of ESTs of the implicated and related libraries, c) the overall number of ESTs available representing the species. Thereby clusters with low probability values are highly enriched with ESTs from the group of libraries of interest indicating a specialized expression.

Recently, the accuracy of this system was demonstrated by Portnoy et al.,
10 2009 (Analysis Of The Melon Fruit Transcriptome Based On 454 Pyrosequencing) in: Plant & Animal Genomes XVII Conference, San Diego, CA. Transcriptomic analysis, based on relative EST abundance in data was performed by 454 pyrosequencing of cDNA representing mRNA of the melon fruit. Fourteen double strand cDNA samples obtained from two genotypes, two fruit tissues (flesh and rind) and four developmental
15 stages were sequenced. GS FLX pyrosequencing (Roche/454 Life Sciences) of non-normalized and purified cDNA samples yielded 1,150,657 expressed sequence tags that assembled into 67,477 unigenes (32,357 singletons and 35,120 contigs). Analysis of the data obtained against the Cucurbit Genomics Database confirmed the accuracy of the sequencing and assembly. Expression patterns of selected genes fitted well their qRT-
20 PCR data.

EXAMPLE 2

PRODUCTION OF ARABIDOPSIS TRANSCRIPTOM AND HIGH THROUGHPUT CORRELATION ANALYSIS OF YIELD, BIOMASS AND/OR 25 VIGOR RELATED PARAMETERS USING 44K ARABIDOPSIS FULL GENOME OLIGONUCLEOTIDE MICRO-ARRAY

To produce a high throughput correlation analysis, the present inventors utilized an Arabidopsis thaliana oligonucleotide micro-array, produced by Agilent Technologies. The array oligonucleotide represents about 40,000 *A. thaliana* genes and
30 transcripts designed based on data from the TIGR ATH1 v.5 database and Arabidopsis MPSS (University of Delaware) databases. To define correlations between the levels of RNA expression and yield, biomass components or vigor related parameters, various

plant characteristics of 15 different *Arabidopsis* ecotypes were analyzed. Among them, nine ecotypes encompassing the observed variance were selected for RNA expression analysis. The correlation between the RNA levels and the characterized parameters was analyzed using Pearson correlation test.

5

Experimental procedures

RNA extraction – Five tissues at different developmental stages including root, leaf, flower at anthesis, seed at 5 days after flowering (DAF) and seed at 12 DAF, representing different plant characteristics, were sampled and RNA was extracted using TRIzol Reagent from Invitrogen. For convenience, each micro-array expression information tissue type has received a Set ID as summarized in Table 1 below.

15

Table 1
Tissues used for Arabidopsis transcriptom expression sets

<i>Expression Set</i>	<i>Set ID</i>
Root	A
Leaf	B
Flower	C
Seed 5 DAF	D
Seed 12 DAF	E

Table 1: Provided are the identification (ID) letters of each of the *Arabidopsis* expression sets (A-E). DAF = days after flowering.

20

Approximately 30-50 mg of tissue was taken from samples. The weighed tissues were ground using pestle and mortar in liquid nitrogen and resuspended in 500 μ l of TRIzol Reagent. To the homogenized lysate, 100 μ l of chloroform was added followed by precipitation using isopropanol and two washes with 75 % ethanol. The RNA was eluted in 30 μ l of RNase-free water. RNA samples were cleaned up using Qiagen's RNeasy minikit clean-up protocol as per the manufacturer's protocol.

25

Yield components and vigor related parameters assessment - eight out of the nine *Arabidopsis* ecotypes were used in each of 5 repetitive blocks (named A, B, C, D and E), each containing 20 plants per plot. The plants were grown in a greenhouse at controlled conditions in 22 °C, and the N:P:K fertilizer (20:20:20; weight ratios)

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[nitrogen (N), phosphorus (P) and potassium (K)] was added. During this time data was collected, documented and analyzed. Additional data was collected through the seedling stage of plants grown in a tissue culture in vertical grown transparent agar plates. Most of chosen parameters were analyzed by digital imaging.

5 **Digital imaging in Tissue culture** - A laboratory image acquisition system was used for capturing images of plantlets grown in square agar plates. The image acquisition system consists of a digital reflex camera (Canon EOS 300D) attached to a 55 mm focal length lens (Canon EF-S series), mounted on a reproduction device (Kaiser RS), which included 4 light units (4x150 Watts light bulb) and located in a darkroom.

10 **Digital imaging in Greenhouse** - The image capturing process was repeated every 3-4 days starting at day 7 till day 30. The same camera attached to a 24 mm focal length lens (Canon EF series), placed in a custom made iron mount, was used for capturing images of larger plants grown in white tubs in an environmental controlled greenhouse. The white tubs were square shape with measurements of 36 x 26.2 cm and
15 7.5 cm deep. During the capture process, the tubs were placed beneath the iron mount, while avoiding direct sun light and casting of shadows. This process was repeated every 3-4 days for up to 30 days.

 An image analysis system was used, which consists of a personal desktop computer (Intel P4 3.0 GHz processor) and a public domain program - ImageJ 1.37,
20 Java based image processing program, which was developed at the U.S National Institutes of Health and is freely available. Images were captured in resolution of 6 Mega Pixels (3072 x 2048 pixels) and stored in a low compression JPEG (Joint Photographic Experts Group standard) format. Next, analyzed data was saved to text files and processed using the JMP statistical analysis software (SAS institute).

25 **Leaf analysis** - Using the digital analysis leaves data was calculated, including leaf number, area, perimeter, length and width. On day 30, 3-4 representative plants were chosen from each plot of blocks A, B and C. The plants were dissected, each leaf was separated and was introduced between two glass trays, a photo of each plant was taken and the various parameters (such as leaf total area, laminar length etc.) were
30 calculated from the images. The blade circularity was calculated as laminar width divided by laminar length.

Root analysis - During 17 days, the different ecotypes were grown in transparent agar plates. The plates were photographed every 3 days starting at day 7 in

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the photography room and the roots development was documented (see examples in Figures 3A-F). The growth rate of roots was calculated according to Formula V.

Formula V:

5 Relative growth rate of root coverage = Regression coefficient of root coverage along time course.

Vegetative growth rate analysis - was calculated according to Formula VI. The analysis was ended with the appearance of overlapping plants.

Formula VI

10 Relative vegetative growth rate area = Regression coefficient of vegetative area along time course.

For comparison between ecotypes the calculated rate was normalized using plant developmental stage as represented by the number of true leaves. In cases where plants with 8 leaves had been sampled twice (for example at day 10 and day 13), only the largest sample was chosen and added to the Anova comparison.

15 **Seeds in siliques analysis** - On day 70, 15-17 siliques were collected from each plot in blocks D and E. The chosen siliques were light brown color but still intact. The siliques were opened in the photography room and the seeds were scatter on a glass tray, a high resolution digital picture was taken for each plot. Using the images the number of seeds per silique was determined.

Seeds average weight - At the end of the experiment all seeds from plots of blocks A-C were collected. An average weight of 0.02 grams was measured from each sample, the seeds were scattered on a glass tray and a picture was taken. Using the digital analysis, the number of seeds in each sample was calculated.

25 **Oil percentage in seeds** - At the end of the experiment all seeds from plots of blocks A-C were collected. Columbia seeds from 3 plots were mixed grounded and then mounted onto the extraction chamber. 210 ml of n-Hexane (Cat No. 080951 Biolab Ltd.) were used as the solvent. The extraction was performed for 30 hours at medium heat 50 °C. Once the extraction has ended the n-Hexane was evaporated using the evaporator at 35 °C and vacuum conditions. The process was repeated twice. The information gained from the Soxhlet extractor (Soxhlet, F. Die gewichtsanalytische Bestimmung des Milchfettes, Polytechnisches J. (Dingler's) 1879, 232, 461) was used to

30

create a calibration curve for the Low Resonance NMR. The content of oil of all seed samples was determined using the Low Resonance NMR (MARAN Ultra– Oxford Instrument) and its MultiQuant software package.

Silique length analysis - On day 50 from sowing, 30 siliques from different plants in each plot were sampled in block A. The chosen siliques were green-yellow in color and were collected from the bottom parts of a grown plant's stem. A digital photograph was taken to determine silique's length.

Dry weight and seed yield - On day 80 from sowing, the plants from blocks A-C were harvested and left to dry at 30 °C in a drying chamber. The biomass and seed weight of each plot was separated, measured and divided by the number of plants. Dry weight = total weight of the vegetative portion above ground (excluding roots) after drying at 30 °C in a drying chamber; Seed yield per plant = total seed weight per plant (gr).

Oil yield - The oil yield was calculated using Formula VII.

Formula VII:

Seed Oil yield = Seed yield per plant (gr) * Oil % in seed

Harvest Index - The harvest index was calculated using Formula IV as described above [Harvest Index = Average seed yield per plant/ Average dry weight].

Experimental Results

Nine different *Arabidopsis* ecotypes were grown and characterized for 18 parameters (named as vectors). Data parameters are summarized in Table 2, below.

Table 2
***Arabidopsis* correlated parameters (vectors)**

<i>Correlated parameter with</i>	<i>Correlation ID</i>
Root length day 13 (cm)	1
Root length day 7 (cm)	2
Relative root growth (cm /day) day 13	3
Fresh weight per plant (gr) at bolting stage	4
Dry matter per plant (gr)	5
Vegetative growth rate (cm ² / day) till 8 true leaves	6
Blade circularity	7
Lamina width (cm)	8
Lamina length (cm)	9
Total leaf area per plant (cm)	10
1000 Seed weight (gr)	11

<i>Correlated parameter with</i>	<i>Correlation ID</i>
Oil % per seed	12
Seeds per silique	13
Silique length (cm)	14
Seed yield per plant (gr)	15
Oil yield per plant (mg)	16
Harvest Index	17
Leaf width/length	18

Table 2. Provided are the Arabidopsis correlated parameters (correlation ID Nos. 1-18). Abbreviations: Cm = centimeter(s); gr = gram(s); mg = milligram(s).

5 The characterized values are summarized in Tables 3 and 4 below.

Table 3
Measured parameters in Arabidopsis ecotypes

<i>Ecotype</i>	<i>Seed yield per plant (gr)</i>	<i>Oil yield per plant (mg)</i>	<i>Oil % per seed</i>	<i>1000 Seed weight (gr)</i>	<i>Dry matter per plant (gr)</i>	<i>Harvest Index</i>	<i>Total leaf area per plant (cm)</i>	<i>Seeds per silique</i>	<i>Silique length (cm)</i>
An-1	0.34	118.63	34.42	0.0203	0.64	0.53	46.86	45.44	1.06
Col-0	0.44	138.73	31.19	0.0230	1.27	0.35	109.89	53.47	1.26
Ct-1	0.59	224.06	38.05	0.0252	1.05	0.56	58.36	58.47	1.31
Cvi (N8580)	0.42	116.26	27.76	0.0344	1.28	0.33	56.80	35.27	1.47
Gr-6	0.61	218.27	35.49	0.0202	1.69	0.37	114.66	48.56	1.24
Kondara	0.43	142.11	32.91	0.0263	1.34	0.32	110.82	37.00	1.09
Ler-1	0.36	114.15	31.56	0.0205	0.81	0.45	88.49	39.38	1.18
Mt-0	0.62	190.06	30.79	0.0226	1.21	0.51	121.79	40.53	1.18
Shakdara	0.55	187.62	34.02	0.0235	1.35	0.41	93.04	25.53	1.00

10 Table 3. Provided are the values of each of the parameters measured in Arabidopsis ecotypes: Seed yield per plant (gram); oil yield per plant (mg); oil % per seed; 1000 seed weight (gr); dry matter per plant (gr); harvest index; total leaf area per plant (cm); seeds per silique; Silique length (cm).

Table 4
Additional measured parameters in Arabidopsis ecotypes

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<i>Ecotype</i>	<i>Veg. GR</i>	<i>Relat. root growth</i>	<i>Root length day 7</i>	<i>Root length day 13</i>	<i>Fresh weight per plant</i>	<i>Lam. Leng.</i>	<i>Lam. width</i>	<i>Leaf width/length</i>	<i>Blade circularity</i>
An-1	0.313	0.631	0.937	4.419	1.510	2.767	1.385	0.353	0.509
Col-0	0.378	0.664	1.759	8.530	3.607	3.544	1.697	0.288	0.481
Ct-1	0.484	1.176	0.701	5.621	1.935	3.274	1.460	0.316	0.450
Cvi (N8580)	0.474	1.089	0.728	4.834	2.082	3.785	1.374	0.258	0.370
Gr-6	0.425	0.907	0.991	5.957	3.556	3.690	1.828	0.356	0.501

Ecotype	Veg. GR	Relat. root growth	Root length day 7	Root length day 13	Fresh weight per plant	Lam. Leng.	Lam. width	Leaf width/length	Blade circularity
Kondara	0.645	0.774	1.163	6.372	4.338	4.597	1.650	0.273	0.376
Ler-1	0.430	0.606	1.284	5.649	3.467	3.877	1.510	0.305	0.394
Mt-0	0.384	0.701	1.414	7.060	3.479	3.717	1.817	0.335	0.491
Shakdara	0.471	0.782	1.251	7.041	3.710	4.149	1.668	0.307	0.409

Table 4. Provided are the values of each of the parameters measured in Arabidopsis ecotypes: Veg. GR = vegetative growth rate (cm²/day) until 8 true leaves; Relat. Root growth = relative root growth (cm/day); Root length day 7 (cm); Root length day 13 (cm); fresh weight per plant (gr) at bolting stage; Lam. Leng. = Lamina length (cm); Lam. Width = Lamina width (cm); Leaf width/length; Blade circularity.

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Tables 5-7, below, provide the selected genes, the characterized parameters (which are used as x axis for correlation) and the correlated tissue transcriptom along with the correlation value (R, calculated using Pearson correlation). When the correlation coefficient (R) between the levels of a gene's expression in a certain tissue and a phenotypic performance across ecotypes is high in absolute value (between 0.5-1), there is an association between the gene (specifically the expression level of this gene) and the phenotypic character. A positive correlation indicates that the expression of the gene in a certain tissue or developmental stage and the correlation vector (phenotype performance) are positively associated (both, expression and phenotypic performance increase or decrease simultaneously) while a negative correlation indicates a negative association (while the one is increasing the other is decreasing and vice versa).

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Table 5

Correlation between the expression level of selected genes in specific tissues or developmental stages and the phenotypic performance across Arabidopsis ecotypes

Gene Name	Corr. Vec.	Exp. Set	R	Corr. Vec.	Exp. Set	R	Corr. Vec.	Exp. Set	R
BDL117	1	C	-0.907	1	A	-0.809	13	D	0.961
BDL118	13	A	0.817	6	A	0.805	5	D	-0.821
BDL118	17	D	0.958	17	D	0.834	8	D	-0.833
BDL118	8	D	-0.845	10	D	-0.912	10	D	-0.975
BDL118	4	D	-0.904						
BDL126	5	B	0.942						
BDL138	16	C	0.841	15	C	0.813	16	B	0.878
BDL138	15	B	0.896	13	A	0.94			

<i>Gene Name</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>
BDL140	14	C	0.841	3	C	0.821	11	C	0.855
BDL140	14	B	0.836	11	B	0.855	7	E	-0.812
BDL140	6	F	0.889	13	D	0.826	14	D	0.862
BDL147	16	B	0.948	15	B	0.898			
BDL149	14	B	0.83	3	B	0.891	14	A	0.951
BDL152	16	D	0.969	3	D	0.855	15	D	0.987
BDL153	14	C	0.836	11	C	0.823	14	A	-0.862
BDL153	11	A	0.88	8	D	-0.81			
BDL154	11	C	0.874						
BDL155	16	B	0.829	16	A	0.86			
BDL156	3	C	0.923	14	B	0.901			
BDL157	3	B	0.854	2	B	-0.825	5	D	-0.803
BDL157	17	D	0.923	9	D	-0.809	8	D	-0.834
BDL157	10	D	-0.915	4	D	-0.89			
BDL158	8	B	0.953	10	B	0.945	4	B	0.899
BDL158	11	A	-0.833						
BDL160	7	C	0.82	18	C	0.972	8	A	0.918
BDL160	8	A	0.839	8	A	0.834	10	A	0.93
BDL160	10	A	0.93	4	A	0.862	1	E	0.864
BDL160	1	E	0.841	2	E	0.861	2	E	0.839
BDL160	8	D	0.867	8	D	0.811	10	D	0.824
BDL162	5	B	0.89						
BDL163	16	B	0.925	15	B	0.884	18	E	0.828
BDL165	8	B	0.952	10	B	0.902	4	B	0.821
BDL165	8	E	0.807	15	E	0.816	11	D	0.846
BDL167	16	B	0.899	15	B	0.946	17	D	0.859
BDL167	2	D	-0.806						
BDL168	16	C	0.97	15	C	0.929	8	B	0.931
BDL168	10	B	0.88	13	A	-0.835	5	D	-0.911
BDL168	8	D	-0.946	10	D	-0.849			
BDL169	14	B	-0.82	11	B	-0.848	12	A	0.901
BDL171	14	C	-0.842	2	B	-0.844	5	A	0.803
BDL171	1	A	-0.851	2	A	-0.821	5	D	-0.827
BDL171	17	D	0.958	17	D	0.853	17	D	0.825
BDL171	9	D	-0.82	8	D	-0.87	16	D	0.857
BDL171	10	D	-0.901	10	D	-0.948	4	D	-0.808
BDL171	4	D	-0.932	15	D	0.838			
BDL173	7	B	0.892	9	B	-0.874	18	B	0.816
BDL173	17	D	0.948	8	D	-0.888	10	D	-0.974
BDL173	4	D	-0.871						
BDL174	17	C	0.901	5	D	-0.917	8	D	-0.879
BDL174	10	D	-0.85						
BDL176	5	D	-0.907	8	D	-0.913	13	D	0.93

<i>Gene Name</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>
BDL177	17	C	0.919	17	B	0.91	17	D	0.82
BDL181	16	C	0.893	15	C	0.838	8	B	0.816
BDL181	12	A	0.931	16	A	0.823	18	F	0.819
BDL181	16	D	0.865	15	D	0.856			
BDL182	12	A	0.913	12	D	0.825			
BDL183	16	B	0.915	15	B	0.898			
BDL186	12	B	0.944	11	B	0.833	8	D	-0.807
BDL187	16	B	0.908	15	B	0.835	5	D	-0.803
BDL187	8	D	-0.892						
BDL188	14	B	0.904	12	D	0.964	3	D	0.857
BDL188	2	D	-0.886						
BDL189	16	B	0.951	15	B	0.907	6	E	-0.854
BDL189	8	D	-0.821	1	D	-0.938			
BDL190	16	B	0.857	15	B	0.91	7	E	-0.865
BDL192	7	B	0.907	9	B	-0.806	17	D	0.91
BDL192	10	D	-0.907	4	D	-0.94	2	D	-0.82
BDL193	8	C	0.846	12	B	0.904	11	B	0.876
BDL193	11	A	0.885	11	E	0.923	5	D	-0.801
BDL193	8	D	-0.802	8	D	-0.844	10	D	-0.806
BDL194	12	B	0.933	18	D	0.877			
BDL196	16	D	0.917	15	D	0.937			
BDL197	13	A	0.91	8	D	0.837			
BDL200	2	C	0.818	16	B	0.864	15	B	0.832
BDL200	14	A	0.917	1	E	0.815	3	D	0.851
BDL201	9	C	0.846	5	D	0.916	17	D	-0.906
BDL201	8	D	0.954	10	D	0.947	4	D	0.865
BDL203	10	B	0.926	4	B	0.893	14	A	-0.828
BDL219	1	A	0.879	2	A	0.821	9	E	-0.801
BDL220	8	B	0.822	10	B	0.844	4	B	0.839
BDL221	12	C	0.897	16	C	0.917	15	C	0.814
BDL221	3	B	0.936	9	D	-0.936	6	D	-0.897
BDL221	4	D	-0.808						
BDL222	1	B	0.829	2	B	0.887	1	A	0.875
BDL222	2	A	0.849	2	D	0.925			
BDL223	14	C	0.93	14	B	0.835	3	B	0.851
BDL223	14	A	0.831						
BDL224	5	E	-0.826	9	E	-0.872			
BDL225	7	C	0.833	15	B	-0.806	13	A	-0.836
BDL227	2	A	-0.931						
BDL229	10	B	0.858	2	D	0.832			
BDL231	16	B	0.911	15	B	0.869	11	D	0.88
BDL233	14	C	-0.885						
BDL235	11	E	0.808	12	D	0.818	2	D	-0.887

<i>Gene Name</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>	<i>Corr. Vec.</i>	<i>Exp. Set</i>	<i>R</i>
BDL240	1	D	-0.808						
BDL241	13	A	-0.88						
BDL242	11	B	0.889						
BDL243	11	B	0.929						
BDL245	3	A	-0.863	11	A	-0.832	16	D	-0.806
BDL247	16	B	0.816						
BDL248	13	A	-0.829						
BDL249	8	C	-0.84	16	E	-0.808	15	E	-0.892
BDL250	9	A	-0.805	2	E	-0.85	17	D	0.815
BDL250	10	D	-0.809	4	D	-0.804	1	D	-0.9
BDL251	18	C	0.802	7	D	-0.861			
BDL47	8	B	0.845						
BDL49	7	E	0.805	9	E	-0.883	6	E	-0.809
BDL62	18	A	0.862	6	E	-0.869	13	D	0.829
BDL75	11	C	0.816	5	B	0.945	8	B	0.823
BDL79	7	B	0.876	18	B	0.841	12	D	0.884
BDL79	2	D	-0.938						
BDL81	8	C	0.897	12	B	0.803	1	D	-0.81
BDL81	2	D	-0.86						
BDL83	3	C	-0.861	2	C	0.905			
BDL85	8	D	0.82						

Table 5. Provided are the correlations between the expression level of selected genes in specific tissues or developmental stages (expression sets) and the phenotypic performance (correlation vector) across Arabidopsis ecotypes. The phenotypic characters [correlation (Corr.) vector (Vec.)] include yield (seed yield, oil yield, oil content), biomass, growth rate and/or vigor components as described in Tables 2, 3 and 4. Exp. Set = expression set according to Table 1 hereinabove.

Table 6 hereinbelow provides data about the homologous of selected genes, the characterized parameters (which are used as x axis for correlation) and the correlated tissue transcriptom along with the correlation value (R, calculated using Pearson correlation).

Table 6
Correlation between the expression level of homologous of the selected genes in specific tissues or developmental stages and the phenotypic performance across Arabidopsis ecotypes

<i>Gene Name</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>
BDL155 H1	leaf	relative root growth	0.814
BDL155 H1	seed5daf	Silique length	-0.855
BDL171 H0	leaf	Leaf width/length	0.835
BDL171 H0	seed5daf	Dry matter per plant	-0.82
BDL171 H0	seed5daf	Lamina width	-0.813

<i>Gene Name</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>
BDL183 H0	seed12daf	Blade circularity	-0.878
BDL231 H0	flower	seed weight	0.825
BDL231 H0	leaf	Silique length	0.816
BDL231 H0	leaf	relative root growth	0.854
BDL231 H0	root	seed weight	0.92
BDL248 H0	seed5daf	relative root growth	0.851
BDL70 H0	seed12daf	Lamina length	-0.816
BDL70 H0	seed5daf	seed yield per plant	-0.82

5 Table 6. Provided are the correlations between the expression levels of homologues of selected Arabidopsis genes in various tissues or developmental stages (Expression sets) and the phenotypic performance in various yield (seed yield, oil yield, oil content), biomass, growth rate and/or vigor components [Correlation (Corr.) vector (Vec.)] Corr. Vec. = correlation vector specified in Tables 2, 3 and 4; Exp. Set = expression set specified in Table 1.

EXAMPLE 3

10 PRODUCTION OF ARABIDOPSIS TRANSCRIPTOM AND HIGH THROUGHPUT CORRELATION ANALYSIS OF NORMAL AND NITROGEN LIMITING CONDITIONS USING 44K ARABIDOPSIS OLIGONUCLEOTIDE MICRO-ARRAY

In order to produce a high throughput correlation analysis, the present inventors utilized a Arabidopsis oligonucleotide micro-array, produced by Agilent
 15 Technologies. The array oligonucleotide represents about 44,000 Arabidopsis genes and transcripts. To define correlations between the levels of RNA expression with NUE, yield components or vigor related parameters various plant characteristics of 14 different Arabidopsis ecotypes were analyzed. Among them, ten ecotypes encompassing the observed variance were selected for RNA expression analysis. The
 20 correlation between the RNA levels and the characterized parameters was analyzed using Pearson correlation test.

Experimental procedures

RNA extraction – Two tissues of plants [leaves and stems] growing at two different nitrogen fertilization levels (1.5 mM Nitrogen or 6 mM Nitrogen) were
 25 sampled and RNA was extracted using TRIzol Reagent from Invitrogen. For convenience, each micro-array expression information tissue type has received a Set ID as summarized in Table 7 below.

30 **Table 7**
Tissues used for Arabidopsis transcriptom expression sets

<i>Expression Set</i>	<i>Set ID</i>
Leaves at 1.5 mM Nitrogen fertilization	A
Leaves at 6 mM Nitrogen fertilization	B
Stems at 1.5 mM Nitrogen fertilization	C
Stem at 6 mM Nitrogen fertilization	D

Table 7: Provided are the identification (ID) letters of each of the Arabidopsis expression sets.

Approximately 30-50 mg of tissue was taken from samples. The weighed
 5 tissues were ground using pestle and mortar in liquid nitrogen and resuspended in 500 μ l
 of TRIzol Reagent. To the homogenized lysate, 100 μ l of chloroform was added
 followed by precipitation using isopropanol and two washes with 75 % ethanol. The
 RNA was eluted in 30 μ l of RNase-free water. RNA samples were cleaned up using
 Qiagen's RNeasy minikit clean-up protocol as per the manufacturer's protocol
 10 (QIAGEN Inc, CA USA).

*Assessment of Arabidopsis yield components and vigor related parameters
 under different nitrogen fertilization levels* – 10 Arabidopsis accessions in 2 repetitive
 plots each containing 8 plants per plot were grown at greenhouse. The growing protocol
 used was as follows: surface sterilized seeds were sown in Eppendorf tubes containing
 15 0.5 x Murashige-Skoog basal salt medium and grown at 23 °C under 12-hour light and
 12-hour dark daily cycles for 10 days. Then, seedlings of similar size were carefully
 transferred to pots filled with a mix of perlite and peat in a 1:1 ratio. Constant nitrogen
 limiting conditions were achieved by irrigating the plants with a solution containing 1.5
 mM inorganic nitrogen in the form of KNO₃, supplemented with 2 mM CaCl₂, 1.25 mM
 20 KH₂PO₄, 1.50 mM MgSO₄, 5 mM KCl, 0.01 mM H₃BO₃ and microelements, while
 normal irrigation conditions (Normal Nitrogen conditions) was achieved by applying a
 solution of 6 mM inorganic nitrogen also in the form of KNO₃, supplemented with 2
 mM CaCl₂, 1.25 mM KH₂PO₄, 1.50 mM MgSO₄, 0.01 mM H₃BO₃ and microelements.
 To follow plant growth, trays were photographed the day nitrogen limiting conditions
 25 were initiated and subsequently every 3 days for about 15 additional days. Rosette plant
 area was then determined from the digital pictures. ImageJ software was used for
 quantifying the plant size from the digital pictures utilizing proprietary scripts designed
 to analyze the size of rosette area from individual plants as a function of time. The
 image analysis system included a personal desktop computer (Intel P4 3.0 GHz

processor) and a public domain program - ImageJ 1.37 (Java based image processing program, which was developed at the U.S. National Institutes of Health and freely available. Next, analyzed data was saved to text files and processed using the JMP statistical analysis software (SAS institute).

5 Data parameters collected are summarized in Table 8, hereinbelow.

Table 8
Arabidopsis correlated parameters (vectors)

<i>Correlated parameter with</i>	<i>Correlation Id</i>
N 1.5 mM; Rosette Area at day 8 [cm ²]	1
N 1.5 mM; Rosette Area at day 10 [cm ²]	2
N 1.5 mM; Plot Coverage at day 8 [%]	3
N 1.5 mM; Plot Coverage at day 10 [%]	4
N 1.5 mM; Leaf Number at day 10	5
N 1.5 mM; Leaf Blade Area at day 10 [cm ²]	6
N 1.5 mM; RGR of Rosette Area at day 3 [cm ² /day]	7
N 1.5 mM; t50 Flowering [day]	8
N 1.5 mM; Dry Weight [gr/plant]	9
N 1.5 mM; Seed Yield [gr/plant]	10
N 1.5 mM; Harvest Index	11
N 1.5 mM; 1000 Seeds weight [gr]	12
N 1.5 mM; seed yield/ rosette area at day 10 [gr/cm ²]	13
N 1.5 mM; seed yield/leaf blade [gr/cm ²]	14
N 1.5 mM; % Seed yield reduction compared to N 6 mM	15
N 1.5 mM; % Biomass reduction compared to N 6 mM	16
N 1.5 mM; N level /DW [SPAD unit/gr]	17
N 1.5 mM; DW/ N level [gr/ SPAD unit]	18
N 1.5 mM; seed yield/ N level [gr/ SPAD unit]	19
N 6 mM; Rosette Area at day 8 [cm ²]	20
N 6 mM; Rosette Area at day 10 [cm ²]	21
N 6 mM; Plot Coverage at day 8 [%]	22
N 6 mM; Plot Coverage at day 10 [%]	23
N 6 mM; Leaf Number at day 10	24
N 6 mM; Leaf Blade Area at day 10	25
N 6 mM; RGR of Rosette Area at day 3 [cm ² /gr]	26
N 6 mM; t50 Flowering [day]	27
N 6 mM; Dry Weight [gr/plant]	28
N 6 mM; Seed Yield [gr/plant]	29
N 6 mM; Harvest Index	30
N 6 mM; 1000 Seeds weight [gr]	31
N 6 mM; seed yield/ rosette area day at day 10 [gr/cm ²]	32
N 6 mM; seed yield/leaf blade [gr/cm ²]	33
N 6 mM; N level / FW	34
N 6 mM; DW/ N level [gr/ SPAD unit]	35

<i>Correlated parameter with</i>	<i>Correlation Id</i>
N 6 mM; N level /DW (SPAD unit/gr plant)	36
N 6 mM; Seed yield/N unit [gr/ SPAD unit]	37

5 Table 8. Provided are the Arabidopsis correlated parameters (vectors). "N" = Nitrogen at the noted concentrations; "gr." = grams; "SPAD" = chlorophyll levels; "t50" = time where 50% of plants flowered; "gr/ SPAD unit" = plant biomass expressed in grams per unit of nitrogen in plant measured by SPAD. "DW" = Plant Dry Weight; "FW" = Plant Fresh weight; "N level /DW" = plant Nitrogen level measured in SPAD unit per plant biomass [gr]; "DW/ N level" = plant biomass per plant [gr]/SPAD unit; Rosette Area (measured using digital analysis); Plot Coverage at the indicated day [%] (calculated by the dividing the total plant area with the total plot area); Leaf Blade Area at the indicated day [cm²] (measured using digital analysis); RGR (relative growth rate) of Rosette Area at the indicated day [cm²/day] (calculated using Formula II); t50 Flowering [day] (the day in which 50% of plant flower); 10 seed yield/ rosette area at day 10 [gr/cm²] (calculated); seed yield/leaf blade [gr/cm²] (calculated); seed yield/ N level [gr/ SPAD unit] (calculated).

Assessment of NUE, yield components and vigor-related parameters - Ten Arabidopsis ecotypes were grown in trays, each containing 8 plants per plot, in a 15 greenhouse with controlled temperature conditions for about 12 weeks. Plants were irrigated with different nitrogen concentration as described above depending on the treatment applied. During this time, data was collected documented and analyzed. Most of chosen parameters were analyzed by digital imaging.

Digital imaging – Greenhouse assay

20 An image acquisition system, which consists of a digital reflex camera (Canon EOS 400D) attached with a 55 mm focal length lens (Canon EF-S series) placed in a custom made Aluminum mount, was used for capturing images of plants planted in containers within an environmental controlled greenhouse. The image capturing process is repeated every 2-3 days starting at day 9-12 till day 16-19 (respectively) from 25 transplanting.

An image processing system was used, which consists of a personal desktop computer (Intel P4 3.0 GHz processor) and a public domain program - ImageJ 1.37, Java based image processing software, which was developed at the U.S. National Institutes of Health and is freely available. Images were captured in resolution of 10 30 Mega Pixels (3888x2592 pixels) and stored in a low compression JPEG (Joint Photographic

Experts Group standard) format. Next, image processing output data was saved to text files and analyzed using the JMP statistical analysis software (SAS institute).

Leaf analysis - Using the digital analysis leaves data was calculated, including leaf number, leaf blade area, plot coverage, Rosette diameter and Rosette area.

5 **Relative growth rate area:** The relative growth rate of the rosette and the leaves was calculated according to Formula II as described above.

10 **Seed yield and 1000 seeds weight** - At the end of the experiment all seeds from all plots were collected and weighed in order to measure seed yield per plant in terms of total seed weight per plant (gr). For the calculation of 1000 seed weight, an average weight of 0.02 grams was measured from each sample, the seeds were scattered on a glass tray and a picture was taken. Using the digital analysis, the number of seeds in each sample was calculated.

15 **Dry weight and seed yield** - At the end of the experiment, plant were harvested and left to dry at 30 °C in a drying chamber. The biomass was separated from the seeds, weighed and divided by the number of plants. Dry weight = total weight of the vegetative portion above ground (excluding roots) after drying at 30 °C in a drying chamber.

Harvest Index - The harvest index was calculated using Formula IV as described above [Harvest Index = Average seed yield per plant/ Average dry weight].

20 **T₅₀ days to flowering** – Each of the repeats was monitored for flowering date. Days of flowering was calculated from sowing date till 50 % of the plots flowered.

25 **Plant nitrogen level** - The chlorophyll content of leaves is a good indicator of the nitrogen plant status since the degree of leaf greenness is highly correlated to this parameter. Chlorophyll content was determined using a Minolta SPAD 502 chlorophyll meter and measurement was performed at time of flowering. SPAD meter readings were done on young fully developed leaf. Three measurements per leaf were taken per plot. Based on this measurement, parameters such as the ratio between seed yield per nitrogen unit [seed yield/N level = seed yield per plant [gr]/SPAD unit], plant DW per nitrogen unit [DW/ N level= plant biomass per plant [g]/SPAD unit], and nitrogen level
30 per gram of biomass [N level/DW= SPAD unit/ plant biomass per plant (gr)] were calculated.

Percent of seed yield reduction- measures the amount of seeds obtained in plants when grown under nitrogen-limiting conditions compared to seed yield produced at normal nitrogen levels expressed in %.

Experimental Results

5 10 different Arabidopsis accessions (ecotypes) were grown and characterized for 37 parameters as described above. The average for each of the measured parameters was calculated using the JMP software and values are summarized in Table 9 below. Subsequent correlation analysis between the various transcriptom sets (Table 7) was conducted. Following are the results integrated to the database.

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Table 9

Correlation between the expression level of selected genes in tissues under limiting or normal nitrogen fertilization and the phenotypic performance across Arabidopsis ecotypes

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<i>Gene Name</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>
BDL117	D	31	-0.748	A	12	-0.784			
BDL118	C	11	-0.914	C	10	-0.723	C	14	-0.75
BDL118	C	8	0.74						
BDL118	D	31	-0.766	B	30	-0.752	D	30	-0.794
BDL118	D	29	-0.715	B	27	0.791	A	11	-0.747
BDL126	A	1	-0.719						
BDL126	D	25	0.801	A	6	-0.715	A	2	-0.711
BDL138	B	28	-0.797	B	27	-0.786	C	9	-0.754
BDL140	D	28	-0.761						
BDL147	D	30	-0.703	C	11	-0.7			
BDL149	C	9	-0.744	A	1	0.703			
BDL152	A	14	-0.705						
BDL154	B	26	0.896	B	32	0.76	B	33	0.861
BDL155	C	9	-0.719						
BDL155_H0	B	28	0.768	D	28	0.757	D	29	0.751
BDL155_H0	C	7	0.743						
BDL156	D	24	0.725	D	21	0.734			
BDL157	B	29	0.714						
BDL158	B	30	-0.722	B	27	0.708			
BDL160	A	12	-0.807	C	9	-0.771	C	5	-0.797
BDL160	C	2	-0.816						
BDL162	B	31	-0.792						
BDL165	C	11	-0.755	C	8	0.748			

<i>Gene Name</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>
BDL117	D	31	-0.748	A	12	-0.784			
BDL118	C	11	-0.914	C	10	-0.723	C	14	-0.75
BDL118	C	8	0.74						
BDL167	C	10	-0.898	C	14	-0.894	C	13	-0.874
BDL167	C	15	0.737						
BDL167	D	30	-0.75	C	11	-0.709	C	7	-0.821
BDL168	A	9	0.703	A	11	-0.786			
BDL168	B	30	-0.73	D	21	0.721	B	27	0.738
BDL169	A	11	-0.804	A	10	-0.746	A	14	-0.714
BDL169	A	13	-0.708	A	8	0.707			
BDL171	B	33	0.865	A	9	-0.735			
BDL171	D	25	0.879	B	26	0.891	D	21	0.765
BDL171	D	20	0.711	B	29	0.735	B	32	0.741
BDL171_I10	C	15	0.719	C	8	0.78			
BDL173	B	26	0.723	B	33	0.72	A	15	0.751
BDL174	C	16	0.7	C	9	-0.73			
BDL176	D	25	0.713	D	24	-0.713	D	26	0.765
BDL176	D	21	0.702	D	20	-0.719	D	32	0.806
BDL176	D	33	0.759	A	7	0.746			
BDL177	B	27	-0.713	A	11	0.792			
BDL181	C	10	-0.75	C	14	-0.795	C	13	-0.805
BDL181	C	15	0.72	A	8	0.794			
BDL181	D	31	-0.704	B	27	0.723	C	11	-0.725
BDL182	A	6	0.728	A	2	0.717	A	1	0.763
BDL183	A	12	-0.764						
BDL183_H0	A	2	-0.725						
BDL186	A	10	-0.754	A	13	-0.713	A	15	0.805
BDL186	A	8	0.865						
BDL186	B	31	-0.714	B	27	0.722	A	11	-0.816
BDL187	A	2	0.711	A	1	0.801	A	10	-0.775
BDL187	A	14	-0.843	A	13	-0.89	A	15	0.721
BDL189	A	13	-0.859	A	15	0.748	A	8	0.759
BDL189	B	30	-0.702	B	27	0.832	A	11	-0.71
BDL189	C	7	-0.714	A	10	-0.825	A	14	-0.82
BDL192	B	24	-0.788	B	21	-0.863	B	20	-0.857
BDL192	B	29	-0.701	B	32	0.769			
BDL192	D	28	-0.764	B	30	-0.785	B	25	-0.802
BDL193	A	6	-0.754	A	2	-0.781	A	1	-0.875
BDL193	A	14	-0.732						
BDL193	D	30	-0.721	D	29	-0.819	C	11	-0.7

<i>Gene Name</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>
BDL117	D	31	-0.748	A	12	-0.784			
BDL118	C	11	-0.914	C	10	-0.723	C	14	-0.75
BDL118	C	8	0.74						
BDL194	B	25	0.786	B	21	0.707			
BDL196	D	25	-0.715	D	21	-0.71	D	20	-0.715
BDL197	A	6	-0.701	A	5	-0.843	A	2	-0.89
BDL197	A	1	-0.827						
BDL201	A	10	-0.768	A	14	-0.84	A	13	-0.872
BDL201	A	15	0.746						
BDL201	B	29	-0.815	A	2	0.747	A	1	0.784
BDL220	C	11	-0.81						
BDL220	D	30	-0.707	B	29	-0.785	C	9	0.752
BDL221	A	2	0.768	C	2	0.763	A	1	0.817
BDL221	B	31	-0.81	D	25	-0.743	D	24	-0.896
BDL221	C	1	0.74						
BDL221	D	21	-0.826	D	20	-0.776	A	5	0.825
BDL222	D	27	-0.716	A	11	0.77	A	14	0.731
BDL223	C	11	-0.757	C	15	0.786	C	8	0.821
BDL223_ H0	C	11	-0.736						
BDL223_ H1	C	8	0.784						
BDL223_ H1	D	24	0.823	A	12	0.815	C	15	0.793
BDL229	C	11	-0.798	C	10	-0.723	C	14	-0.771
BDL229	C	13	-0.769	A	8	0.753	C	8	0.828
BDL229	D	30	-0.787	B	27	0.793	D	27	0.873
BDL231	A	11	-0.798	C	6	-0.704	A	10	-0.826
BDL231	A	14	-0.836	A	13	-0.843	A	15	0.823
BDL231	A	8	0.768						
BDL231_ H0	A	11	-0.779	A	10	-0.799	A	14	-0.841
BDL231_ H0	A	13	-0.835	A	15	0.706	A	8	0.721
BDL233	C	8	0.835						
BDL233	D	28	0.755	C	11	-0.768	C	15	0.725
BDL235	C	11	-0.87	C	10	-0.749	A	14	-0.741
BDL235	C	14	-0.726	A	13	-0.702	C	8	0.802
BDL240	A	8	0.742						
BDL240	D	24	0.794	B	27	0.737	A	11	-0.787
BDL241	B	27	0.752						
BDL242	A	5	-0.719	A	15	-0.722			
BDL245	D	31	-0.875						

<i>Gene Name</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>
BDL117	D	31	-0.748	A	12	-0.784			
BDL118	C	11	-0.914	C	10	-0.723	C	14	-0.75
BDL118	C	8	0.74						
BDL247	D	31	0.851						
BDL249	A	8	-0.707						
BDL249	C	12	0.707	A	11	0.82	A	10	0.727
BDL250	A	10	0.717	A	14	0.797	A	13	0.81
BDL252	C	9	-0.707	C	7	-0.714			
BDL49	B	30	0.866	B	26	0.85	B	29	0.933
BDL49	B	33	0.762						
BDL58	B	26	0.763	D	26	0.755	B	29	0.897
BDL58	C	14	0.816	C	13	0.855	C	15	-0.784
BDL58	C	8	-0.843						
BDL58	D	32	0.758	D	33	0.778	C	10	0.8
BDL62	C	11	-0.858	C	10	-0.739	C	14	-0.77
BDL62	C	13	-0.747	C	8	0.819			
BDL63	C	16	-0.704						
BDL64	B	31	-0.793	C	5	-0.739			
BDL70_110	C	8	0.757						
BDL75	C	11	-0.716	C	8	0.713			
BDL75	D	31	-0.776	D	28	0.75	D	30	-0.832
BDL79	B	31	-0.77						
BDL85	C	10	0.824	C	14	0.754	C	13	0.721
BDL85	C	15	-0.705						

Table 9. Provided are the correlations (R) between the expression levels of selected genes in tissues (leaves or stems) under limiting (1.5 mM Nitrogen) or normal (6 mM Nitrogen) conditions (Expression sets) and the phenotypic performance in various yield (seed yield, oil yield, oil content), biomass, growth rate and/or vigor components [Correlation (Corr.) vector (Vec.)] under limiting or normal Nitrogen conditions. Corr. Vec. = correlation vector according to Table 8 hereinabove; Exp. Set = expression set according to Table 7 hereinabove.

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EXAMPLE 4

PRODUCTION OF SORGHUM TRANSCRIPTOM AND HIGH THROUGHPUT CORRELATION ANALYSIS WITH ABST RELATED PARAMETRERS USING 44K SORGHUM OLIGONUCLEOTIDE MICRO-ARRAYS

In order to produce a high throughput correlation analysis, the present inventors utilized a Sorghum oligonucleotide micro-array, produced by Agilent Technologies. The array oligonucleotide represents about 44,000 Sorghum genes and transcripts. In

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order to define correlations between the levels of RNA expression with ABST and yield components or vigor related parameters, various plant characteristics of 17 different sorghum varieties were analyzed. Among them, 10 varieties encompassing the observed variance were selected for RNA expression analysis. The correlation between the RNA levels and the characterized parameters was analyzed using Pearson correlation test.

Correlation of Sorghum varieties across ecotype grown under severe drought conditions

Experimental procedures

17 Sorghum varieties were grown in 3 repetitive plots, in field. Briefly, the growing protocol was as follows: sorghum seeds were sown in soil and grown under normal condition until around 35 days from sowing, around V8 (Last leaf visible, but still rolled up, ear beginning to swell). At this point, irrigation was stopped, and severe drought stress was developed. In order to define correlations between the levels of RNA expression with drought, yield components or vigor related parameters, the 17 different sorghum varieties were analyzed. Among them, 10 varieties encompassing the observed variance were selected for RNA expression analysis. The correlation between the RNA levels and the characterized parameters was analyzed using Pearson correlation test.

RNA extraction – All 10 selected Sorghum varieties were sample per each treatment. Plant tissues [Flag leaf and Flower meristem] growing under severe drought stress and plants grown under Normal conditions were sampled and RNA was extracted using TRIzol Reagent from Invitrogen. For convenience, each micro-array expression information tissue type has received a Set ID as summarized in Table 10 below.

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Table 10
Sorghum transcriptom expression sets

<i>Expression Set</i>	<i>Set ID</i>
Drought Stress: Flag leaf	U
Normal conditions: Flag leaf	X
Normal conditions: Flower meristem	Y

5 Table 10: Provided are the sorghum transcriptom expression set U, X and Y. Flag leaf = the leaf below the flower; Flower meristem = Apical meristem following panicle initiation.

The collected data parameters were as follows:

Grain per Plant (gr.) - At the end of the experiment (Inflorescence were dry) all spikes from plots within blocks A-C were collected. 5 Inflorescence were separately
10 threshed and grains were weighted, all additional Inflorescence were threshed together and weighted as well. The average weight per Inflorescence was calculated by dividing the total grain weight by number of total Inflorescence per plot, or in case of 5 inflorescence, by weight by the total grain number by 5.

Plant height – Plants were characterized for height during growing period at 6
15 time points. In each measure, plants were measured for their height using a measuring tape. Height was measured from ground level to top of the longest leaf.

Inflorescence Weight (gr.) - At the end of the experiment (when Inflorescence were dry) five Inflorescence from plots within blocks A-C were collected. The Inflorescence were weighted (gr.).

20 ***SPAD*** - Chlorophyll content was determined using a Minolta SPAD 502 chlorophyll meter and measurement was performed at time of flowering. SPAD meter readings were done on young fully developed leaf. Three measurements per leaf were taken per plot.

Vegetative dry weight and Inflorescence - At the end of the experiment (when
25 Inflorescence were dry) all Inflorescence and vegetative material from plots within blocks A-C were collected. The biomass and Inflorescence weight of each plot was separated, measured and divided by the number of Inflorescence.

Dry weight = total weight of the vegetative portion above ground (excluding roots) after drying at 70 °C in oven for 48 hours;

30 ***Harvest Index (for sorghum)*** - The harvest index is calculated using Formula VIII.

Formula VIII:

Harvest Index = Average grain dry weight per Inflorescence / (Average vegetative dry weight per Inflorescence + Average Inflorescence dry weight)

Experimental Results

5 16 different sorghum varieties were grown and characterized for 7 parameters:
 "Seed/plant normal" = total seed weight per plant under normal conditions; "DW-5
 Inflorescence Normal" – dry weight of five complete inflorescences (seeds and rachis)
 under normal conditions; "DW all Normal" = dry weight of per plot under normal
 conditions; "Weight of seeds (5 heads) gr Normal" = dry weight of seeds from five
 10 inflorescences under normal conditions; "Plant Height 4 Drought" = plant height in the
 4th time point under drought conditions; "Plant Height 4 Normal" = plant height in the
 4th time point under normal conditions; "Plant Height 6 Normal" = plant height in the 6th
 time point under normal conditions. The average for each of the measured parameter
 was calculated using the JMP software and values are summarized in Table 11 below.
 15 Subsequent correlation analysis between the various transcriptom sets (Table 10) and
 the average parameters, was conducted (Tables 12). Results were then integrated to the
 database.

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Table 11
Measured parameters in Sorghum accessions

Seed ID	Seed/Plant Normal	DW-5 Inflorescence Normal	DW all Normal	Weight of seeds (5 heads) gr Normal	Plant Height 4 Drought	Plant Height 4 Normal	Plant Height 6 Normal
20	0.031	0.039	5.408	0.237	38.000	37.313	37.313
21	0.026	0.062	3.091	0.225	30.833		
22	0.019	0.033	21.341	0.142	110.833	47.750	47.750
24	0.038	0.030	5.329	0.352	42.833	40.083	40.083
25	0.027	0.074	20.600	0.161	49.583	45.938	45.938
26	0.046	0.049	21.685	0.332	49.750	41.438	41.438
27	0.048	0.046	11.205	0.317	46.875	44.875	44.875
28	0.031	0.033	9.045	0.222	41.917	42.125	42.125
29	0.040	0.048	10.293	0.283	46.125	41.000	41.000
30	0.038	0.038	9.139	0.300	50.167	42.500	42.063
31	0.032	0.023	9.549	0.227	43.583	41.875	41.875
32	0.033	0.039	10.424	0.277	50.833	43.375	43.375
33	0.033	0.049	10.150	0.353	42.417	39.813	39.813
34	0.052	0.050	8.783	0.351	45.500	40.625	40.625
35	0.036	0.038	10.259	0.270	50.375	44.375	44.375
36	0.038	0.042	12.005	0.299	48.833	43.250	43.250
37	0.042	0.063	15.681	0.263	49.833	41.000	41.000

Table 11: Provided are the values of each of the parameters (as described above) measured in Sorghum accessions (Seed ID) under normal and drought conditions. Growth conditions are specified in the experimental procedure section.

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Table 12

Correlation between the expression level of homologues of selected genes in various tissues and the phenotypic performance under normal or abiotic stress conditions across Sorghum accessions

Gene Name	Exp. Set	Corr. Vec.	R
BDL102_H47	flower	DW all Normal	-0.721
BDL102_H47	flower	FW-Inflorescence/Plant Normal	0.717
BDL102_H47	flower	Plant Height 4 Normal	-0.722
BDL102_H47	flower	Plant Height 6 Normal	-0.718
BDL83_H57	flag leaf	DW-5 Inflorescence Normal	-0.712
BDL83_H57	flag leaf	Seed/Plant Normal	-0.708
BDL83_H57	flower	Plant Height 4 Normal	0.858
BDL83_H57	flower	Plant Height 6 Normal	0.853
BDL83_H58	flag leaf	Plant Height 4 Drought	0.763
BDL83_H58	flower	Seed/Plant Low-N	0.759
BDL83_H58	flower	Weight of seeds (5 heads) gr Normal	-0.707
BDL83_H58	Flower meristem	Leaf_No 2 Normal	0.802
BDL88_H13	Flag leaf	Leaf_No 3 Drought	-0.893
BDL88_H13	Flag leaf	Leaf_TP 1 Drought	-0.929
BDL88_H13	Flag leaf	Plant Height 2 Drought	-0.796
BDL88_H13	Flag leaf	Plant Height 3 Drought	-0.837
BDL88_H13	Flag leaf	Seed/Plant Normal	0.73

Table 12. Provided are the correlations (R) between the expression levels of homologues of selected genes in tissues (Flag leaf or Flower meristem; Expression sets) and the phenotypic performance in various yield, biomass, growth rate and/or vigor components [Correlation (Corr.) vector (Vec.)] under abiotic stress conditions (drought) or normal conditions across Sorghum accessions.

Sorghum vigor related parameters under 100 mM NaCl and low temperature (8-10 °C) – Ten Sorghum varieties were grown in 3 repetitive plots, each containing 17 plants, at a net house under semi-hydroponics conditions. Briefly, the growing protocol was as follows: Sorghum seeds were sown in trays filled with a mix of vermiculite and peat in a 1:1 ratio. Following germination, the trays were transferred to the high salinity solution (100 mM NaCl in addition to the Full Hogland solution), low temperature (8-10 °C in the presence of Full Hogland solution) or at Normal growth solution [Full Hogland solution at 20-24 °C].

Full Hogland solution consists of: KNO₃ - 0.808 grams/liter, MgSO₄ - 0.12 grams/liter, KH₂ PO₄ - 0.172 grams/liter and 0.01 % (volume/volume) of 'Super coratin' micro elements (Iron-EDDHA [ethylenediamine-N,N'-bis(2-hydroxyphenylacetic acid)]- 40.5 grams/liter; Mn - 20.2 grams/liter; Zn 10.1 grams/liter; Co 1.5 grams/liter; and Mo 1.1 grams/liter), solution's pH should be 6.5 – 6.8].

RNA extraction – All 10 selected Sorghum varieties were sampled per each treatment. Two tissues [leaves and roots] growing at 100 mM NaCl, low temperature (8-10 °C) or under Normal conditions (full Hogland at a temperature between 20-24 °C) were sampled and RNA was extracted using TRIzol Reagent from Invitrogen.

5 **Experimental Results**

10 10 different Sorghum varieties were grown and characterized for the following parameters: "Leaf number Normal" = leaf number per plant under normal conditions (average of five plants); "Plant Height Normal" = plant height under normal conditions (average of five plants); "Root DW 100 mM NaCl" – root dry weight per plant under salinity conditions (average of five plants); The average for each of the measured parameter was calculated using the JMP software and values are summarized in Table 13 below. Subsequent correlation analysis between the various transcriptom sets and the average parameters was conducted (Table 14). Results were then integrated to the database.

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Table 13
Measured parameters in Sorghum accessions

<i>Seed ID</i>	<i>Plant Height T1 NaCl</i>	<i>Plant Height T1+8 NaCl</i>	<i>Plant Height T1+15 NaCl</i>	<i>Leaf number T1 NaCl</i>	<i>Leaf number T1+8 Normal</i>	<i>DW Root/Plant NaCl</i>
20	7.900	14.200	21.800	3.000	4.167	0.05
22	9.500	16.267	23.167	3.133	4.500	0.10447479
26	10.933	20.367	30.367	3.400	4.800	0.12370635
27	7.933	13.333	22.833	3.067	4.600	0.06880519
28	9.700	15.900	23.700	3.333	4.533	0.07568254
29	8.533	16.533	23.300	3.067	4.967	0.07517045
30	8.900	15.467	22.467	3.067	4.600	0.13539542
31	10.367	18.933	26.833	3.267	4.933	0.09546434
34	7.000	13.680	20.280	3.000	4.500	0.16491667
37	7.833	15.767	23.567	3.067	4.567	0.13888278

20 Table 13: Provided are the measured parameters of the Sorghum Accessions under normal conditions or high salt conditions at the indicated time points. T1 (first day of measurements); T1+8 (8 days following the first day); T1+15 (15 days following the first day). The exact conditions are detailed above in the experiment description section.

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Table 14

Correlation between the expression level of homologues of selected genes in roots and the phenotypic performance under normal or abiotic stress conditions across Sorghum accessions

<i>Gene Name</i>	<i>Exp. Set</i>	<i>Corr. Vec.</i>	<i>R</i>
<i>BDL83_H58</i>	<i>root</i>	<i>DW Root/Plant NaCl</i>	<i>0.897</i>
<i>BDL83_H58</i>	<i>root</i>	<i>Leaf number T1 NaCl</i>	<i>-0.75</i>
<i>BDL83_H58</i>	<i>root</i>	<i>Plant Height T1 NaCl</i>	<i>-0.82</i>
<i>BDL83_H58</i>	<i>root</i>	<i>Plant Height T1+8 NaCl</i>	<i>-0.9</i>
<i>BDL83_H58</i>	<i>root</i>	<i>Plant Height T1+15 NaCl</i>	<i>-0.77</i>
<i>BDL83_H58</i>	<i>root</i>	<i>Leaf number T1 Normal</i>	<i>0.779</i>

5 Table 14. Provided are the correlations (R) between the expression levels of homologues of selected genes in roots [Expression (Exp.) sets] and the phenotypic performance [yield, biomass, growth rate and/or vigor components (Correlation vector)] at the indicated time points under abiotic stress conditions (salinity) or normal conditions across Sorghum accessions. Corr. Vec. = correlation vector as described hereinabove (Table 13). T1 (first day of measurements); T1+8 (8 days following the first day);
10 T1+15 (15 days following the first day).

EXAMPLE 5

IDENTIFICATION OF GENES WHICH INCREASE YIELD, BIOMASS, 15 GROWTH RATE, VIGOR, OIL CONTENT, ABIOTIC STRESS TOLERANCE OF PLANTS AND NITROGEN USE EFFICIENCY

Based on the above described bioinformatics and experimental tools, the present inventors have identified 105 genes (89 distinct gene families) which have a
20 major impact on yield, seed yield, oil yield, biomass, growth rate, vigor, oil content, abiotic stress tolerance, and/or nitrogen use efficiency when expression thereof is increased in plants. The identified genes, their curated polynucleotide and polypeptide sequences, as well as their updated sequences according to Genbank database are summarized in Table 15, hereinbelow.

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Table 15

Identified polynucleotides which affect plant yield, seed yield, oil yield, oil content, biomass, growth rate, vigor, abiotic stress tolerance and/or nitrogen use efficiency of a plant

30

<i>Serial No</i>	<i>Gene Name</i>	<i>Cluster Name</i>	<i>Organism</i>	<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
1	BDI.47	arabidopsis gb165 AT4 G38660	arabidopsis	1	106

<i>Serial No</i>	<i>Gene Name</i>	<i>Cluster Name</i>	<i>Organism</i>	<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
2	BDL47	arabidopsislgb165lAT4 G38660	arabidopsis	1	194
3	BDI 48	arabidopsislgb165lAT2 G25270	arabidopsis	2	107
4	BDL62	arabidopsislgb165lAT1 G64390	arabidopsis	3	108
5	BDL75	arabidopsislgb165lAT3 G55420	arabidopsis	4	109
6	BDL79	arabidopsislgb165lAT3 G20010	arabidopsis	5	110
7	BDL81	arabidopsislgb165lAT1 G13030	arabidopsis	6	111
8	BDI 83	arabidopsislgb165lAT1 G43670	arabidopsis	7	112
9	BDL117	arabidopsislgb165lAT4 G33240	arabidopsis	8	113
10	BDL118	arabidopsislgb165lAT2 G22125	arabidopsis	9	114
11	BDL126	arabidopsislgb165lAT3 G59100	arabidopsis	10	115
12	BDL138	arabidopsislgb165lAT3 G12180	arabidopsis	11	116
13	BDL140	arabidopsislgb165lAT5 G02640	arabidopsis	12	117
14	BDL147	arabidopsislgb165lAT2 G47930	arabidopsis	13	118
15	BDL149	arabidopsislgb165lAT5 G15750	arabidopsis	14	119
16	BDL152	arabidopsislgb165lAT1 G70420	arabidopsis	15	120
17	BDL153	arabidopsislgb165lAT5 G47690	arabidopsis	16	121
18	BDL154	arabidopsislgb165lAT1 G62940	arabidopsis	17	122
19	BDL155	arabidopsislgb165lAT4 G11630	arabidopsis	18	123

<i>Serial No</i>	<i>Gene Name</i>	<i>Cluster Name</i>	<i>Organism</i>	<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
20	BDL156	arabidopsislgb165 AT1 G01570	arabidopsis	19	124
21	BDL157	arabidopsislgb165 AT1 G14560	arabidopsis	20	125
22	BDL158	arabidopsislgb165 AT1 G22030	arabidopsis	21	126
23	BDL160	arabidopsislgb165 AT1 G28960	arabidopsis	22	127
24	BDL162	arabidopsislgb165 AT1 G49360	arabidopsis	23	128
25	BDL163	arabidopsislgb165 AT1 G51430	arabidopsis	24	129
26	BDL165	arabidopsislgb165 AT1 G65010	arabidopsis	25	130
27	BDL167	arabidopsislgb165 AT1 G79630	arabidopsis	26	131
28	BDL168	arabidopsislgb165 AT2 G01910	arabidopsis	27	132
29	BDL169	arabidopsislgb165 AT2 G19120	arabidopsis	28	133
30	BDL171	arabidopsislgb165 AT2 G32320	arabidopsis	29	134
31	BDL173	arabidopsislgb165 AT2 G36720	arabidopsis	30	135
32	BDL174	arabidopsislgb165 AT2 G39580	arabidopsis	31	136
33	BDL176	arabidopsislgb165 AT3 G10160	arabidopsis	32	137
34	BDL177	arabidopsislgb165 AT3 G18610	arabidopsis	33	138
35	BDL181	arabidopsislgb165 AT4 G12640	arabidopsis	34	139
36	BDL182	arabidopsislgb165 AT4 G14605	arabidopsis	35	140
37	BDL183	arabidopsislgb165 AT4 G15620	arabidopsis	36	141

<i>Serial No</i>	<i>Gene Name</i>	<i>Cluster Name</i>	<i>Organism</i>	<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
38	BDL186	arabidopsislgb165 AT4 G32560	arabidopsis	37	142
39	BDL187	arabidopsislgb165 AT4 G35130	arabidopsis	38	143
40	BDL188	arabidopsislgb165 AT4 G35560	arabidopsis	39	144
41	BDL189	arabidopsislgb165 AT5 G05560	arabidopsis	40	145
42	BDL190	arabidopsislgb165 AT5 G06690	arabidopsis	41	146
43	BDL192	arabidopsislgb165 AT5 G09880	arabidopsis	42	147
44	BDL193	arabidopsislgb165 AT5 G12950	arabidopsis	43	148
45	BDL194	arabidopsislgb165 AT5 G16540	arabidopsis	44	149
46	BDL196	arabidopsislgb165 AT5 G38180	arabidopsis	45	150
47	BDL197	arabidopsislgb165 AT5 G39240	arabidopsis	46	151
48	BDL200	arabidopsislgb165 AT5 G51470	arabidopsis	47	152
49	BDL201	arabidopsislgb165 AT5 G58250	arabidopsis	48	153
50	BDL203	arabidopsislgb165 AT5 G66440	arabidopsis	49	154
51	BDL219	arabidopsislgb165 AT1 G36095	arabidopsis	50	155
52	BDL220	arabidopsislgb165 AT1 G61170	arabidopsis	51	156
53	BDL221	arabidopsislgb165 AT1 G75860	arabidopsis	52	157
54	BDL222	arabidopsislgb165 AT1 G77885	arabidopsis	53	158
55	BDL223	arabidopsislgb165 AT2 G37975	arabidopsis	54	159

<i>Serial No</i>	<i>Gene Name</i>	<i>Cluster Name</i>	<i>Organism</i>	<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
56	BDL229	arabidopsislgb165 AT5 G53830	arabidopsis	55	160
57	BDI.230	arabidopsislgb154 ATII PEARA	arabidopsis	56	161
58	BDL231	arabidopsislgb165 AT1 G16350	arabidopsis	57	162
59	BDL235	arabidopsislgb165 AT2 G38970	arabidopsis	58	163
60	BDL242	arabidopsislgb165 AT3 G22740	arabidopsis	59	164
61	BDL243	arabidopsislgb165 AT3 G31430	arabidopsis	60	165
62	BDI.245	arabidopsislgb165 AT4 G12690	arabidopsis	61	166
63	BDL247	arabidopsislgb165 AT4 G29510	arabidopsis	62	167
64	BDL248	arabidopsislgb165 AT4 G37360	arabidopsis	63	168
65	BDL250	arabidopsislgb165 AT4 G37400	arabidopsis	64	169
66	BDL49	arabidopsislgb165 AT3 G06180	arabidopsis	65	170
67	BDL58	arabidopsislgb165 AT1 G22160	arabidopsis	66	171
68	BDL63	arabidopsislgb165 AT2 G40550	arabidopsis	67	172
69	BDL64	arabidopsislgb165 AT5 G38020	arabidopsis	68	173
70	BDL70	cottonlgb164 AF150730	cotton	69	174
71	BDL85	arabidopsislgb165 AT1 G05340	arabidopsis	70	175
72	BDL88	ricelgb157.2 AA754446	rice	71	176
73	BDL90	ricelgb154 AK102950	rice	72	
74	BDL94	ricelgb157.2 BE228242	rice	73	177
75	BDL102	maizelgb164 AI600933	maize	74	178

<i>Serial No</i>	<i>Gene Name</i>	<i>Cluster Name</i>	<i>Organism</i>	<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
76	BDL224	arabidopsislgb165lAT4 G29780	arabidopsis	75	179
77	BDI.225	arabidopsislgb165lAT5 G39670	arabidopsis	76	180
78	BDL226	arabidopsislgb165lAT5 G65470	arabidopsis	77	181
79	BDL227	arabidopsislgb165lAT3 G56410	arabidopsis	78	182
80	BDL228	castorbeanlgb160lEE25 6201	castorbean	79	183
81	BDL232	arabidopsislgb165lAT1 G56100	arabidopsis	80	184
82	BDI.233	arabidopsislgb165lAT1 G66360	arabidopsis	81	185
83	BDL234	arabidopsislgb165lAT2 G05400	arabidopsis	82	186
84	BDL237	arabidopsislgb165lAT2 G45510	arabidopsis	83	187
85	BDL238	arabidopsislgb165lAT3 G04540	arabidopsis	84	188
86	BDL240	arabidopsislgb165lAT3 G18480	arabidopsis	85	189
87	BDL241	arabidopsislgb165lAT3 G20020	arabidopsis	86	190
88	BDL249	arabidopsislgb165lAT4 G37370	arabidopsis	87	191
89	BDL251	arabidopsislgb165lAT5 G08250	arabidopsis	88	192
90	BDL252	arabidopsislgb165lAT5 G18650	arabidopsis	89	193
91	BDL79	arabidopsislgb165lAT3 G20010	arabidopsis	90	110
92	BDL126	arabidopsislgb165lAT3 G59100	arabidopsis	91	115
93	BDL153	arabidopsislgb165lAT5 G47690	arabidopsis	92	195

<i>Serial No</i>	<i>Gene Name</i>	<i>Cluster Name</i>	<i>Organism</i>	<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
94	BDL156	arabidopsislgb165 AT1 G01570	arabidopsis	93	124
95	BDL167	arabidopsislgb165 AT1 G79630	arabidopsis	94	196
96	BDL169	arabidopsislgb165 AT2 G19120	arabidopsis	95	197
97	BDL171	arabidopsislgb165 AT2 G32320	arabidopsis	96	134
98	BDL174	arabidopsislgb165 AT2 G39580	arabidopsis	97	198
99	BDL187	arabidopsislgb165 AT4 G35130	arabidopsis	98	143
100	BDL189	arabidopsislgb165 AT5 G05560	arabidopsis	99	199
101	BDL197	arabidopsislgb165 AT5 G39240	arabidopsis	100	200
102	BDL200	arabidopsislgb165 AT5 G51470	arabidopsis	101	152
103	BDL231	arabidopsislgb165 AT1 G16350	arabidopsis	102	162
104	BDL248	arabidopsislgb165 AT4 G37360	arabidopsis	103	168
105	BDL70	cottonlgb164 AF150730	cotton	104	201
106	BDL238	arabidopsislgb165 AT3 G04540	arabidopsis	105	202

Table 15: Provided are the identified genes, their annotation, organism and polynucleotide and polypeptide sequence identifiers.

5

EXAMPLE 6

IDENTIFICATION OF HOMOLOGOUS SEQUENCES THAT INCREASE SEED YIELD, OIL YIELD, GROWTH RATE, OIL CONTENT, BIOMASS, VIGOR, ABST RESISTANCE AND/OR NUE OF A PLANT

The concepts of orthology and paralogy have recently been applied to
10 functional characterizations and classifications on the scale of whole-genome

comparisons. Orthologs and paralogs constitute two major types of homologs: The first evolved from a common ancestor by specialization, and the latter are related by duplication events. It is assumed that paralogs arising from ancient duplication events are likely to have diverged in function while true orthologs are more likely to retain identical function over evolutionary time.

To identify putative orthologs of the genes affecting plant yield, oil yield, oil content, seed yield, growth rate, vigor, biomass, abiotic stress tolerance and/or nitrogen use efficiency, all sequences were aligned using the BLAST (*Basic Local Alignment Search Tool*). Sequences sufficiently similar were tentatively grouped. These putative orthologs were further organized under a Phylogram - a branching diagram (tree) assumed to be a representation of the evolutionary relationships among the biological taxa. Putative ortholog groups were analyzed as to their agreement with the phylogram and in cases of disagreements these ortholog groups were broken accordingly.

Expression data was analyzed and the EST libraries were classified using a fixed vocabulary of custom terms such as developmental stages (e.g., genes showing similar expression profile through development with up regulation at specific stage, such as at the seed filling stage) and/or plant organ (e.g., genes showing similar expression profile across their organs with up regulation at specific organs such as seed). The annotations from all the ESTs clustered to a gene were analyzed statistically by comparing their frequency in the cluster versus their abundance in the database, allowing the construction of a numeric and graphic expression profile of that gene, which is termed "digital expression". The rationale of using these two complementary methods with methods of phenotypic association studies of QTLs, SNPs and phenotype expression correlation is based on the assumption that true orthologs are likely to retain identical function over evolutionary time. These methods provide different sets of indications on function similarities between two homologous genes, similarities in the sequence level - identical amino acids in the protein domains and similarity in expression profiles.

Methods for searching and identifying homologues of yield and improved agronomic traits such as ABS tolerance and NUE related polypeptides or polynucleotides are well within the realm of the skilled artisan. The search and identification of homologous genes involves the screening of sequence information

available, for example, in public databases such as the DNA Database of Japan (DDBJ), Genbank, and the European Molecular Biology Laboratory Nucleic Acid Sequence Database (EMBL) or versions thereof or the MIPS database. A number of different search algorithms have been developed, including but not limited to the suite of programs referred to as BLAST programs. There are five implementations of BLAST, three designed for nucleotide sequence queries (BLASTN, BLASTX, and TBLASTX) and two designed for protein sequence queries (BLASTP and TBLASTN) (Coulson, Trends in Biotechnology: 76-80, 1994; Birren et al., Genome Analysis, I: 543, 1997). Such methods involve alignment and comparison of sequences. The BLAST algorithm calculates percent sequence identity and performs a statistical analysis of the similarity between the two sequences. The software for performing BLAST analysis is publicly available through the National Centre for Biotechnology Information. Other such software or algorithms are GAP, BESTFIT, FASTA and TFASTA. GAP uses the algorithm of Needleman and Wunsch (J. Mol. Biol. 48: 443-453, 1970) to find the alignment of two complete sequences that maximizes the number of matches and minimizes the number of gaps.

The homologous genes may belong to the same gene family. The analysis of a gene family may be carried out using sequence similarity analysis. To perform this analysis one may use standard programs for multiple alignments e.g. Clustal W. A neighbour-joining tree of the proteins homologous to the genes in this invention may be used to provide an overview of structural and ancestral relationships. Sequence identity may be calculated using an alignment program as described above. It is expected that other plants will carry a similar functional gene (ortholog) or a family of similar genes and those genes will provide the same preferred phenotype as the genes presented here. Advantageously, these family members may be useful in the methods of the invention. Example of other plants are included here but not limited to, barley (*Hordeum vulgare*), Arabidopsis (*Arabidopsis thaliana*), maize (*Zea mays*), cotton (*Gossypium*), Oilseed rape (*Brassica napus*), Rice (*Oryza sativa*), Sugar cane (*Saccharum officinarum*), Sorghum (*Sorghum bicolor*), Soybean (*Glycine max*), Sunflower (*Helianthus annuus*), Tomato (*Lycopersicon esculentum*), Wheat (*Triticum aestivum*).

The above-mentioned analyses for sequence homology can be carried out on a full-length sequence, but may also be based on a comparison of certain regions such as

conserved domains. The identification of such domains, would also be well within the realm of the person skilled in the art and would involve, for example, a computer readable format of the nucleic acids of the present invention, the use of alignment software programs and the use of publicly available information on protein domains, conserved motifs and boxes. This information is available in the PRODOM, PIR or Pfam database. Sequence analysis programs designed for motif searching may be used for identification of fragments, regions and conserved domains as mentioned above. Preferred computer programs include, but are not limited to, MEME, SIGNALSCAN, and GENESCAN.

A person skilled in the art may use the homologous sequences provided herein to find similar sequences in other species and other organisms. Homologues of a protein encompass, peptides, oligopeptides, polypeptides, proteins and enzymes having amino acid substitutions, deletions and/or insertions relative to the unmodified protein in question and having similar biological and functional activity as the unmodified protein from which they are derived. To produce such homologues, amino acids of the protein may be replaced by other amino acids having similar properties (conservative changes, such as similar hydrophobicity, hydrophilicity, antigenicity, propensity to form or break α -helical structures or β -sheet structures). Conservative substitution tables are well known in the art (see for example Creighton (1984) Proteins. W.H. Freeman and Company). Homologues of a nucleic acid encompass nucleic acids having nucleotide substitutions, deletions and/or insertions relative to the unmodified nucleic acid in question and having similar biological and functional activity as the unmodified nucleic acid from which they are derived.

Polynucleotides and polypeptides with significant homology to the identified genes described in Table 15 above have been identified from the databases using BLAST software using the Blastp and tBlastn algorithms. The query nucleotide sequences were SEQ ID NOS: 1-106 and the identified homologues are provided in Table 16, below. These genes are expected to increase plant yield, seed yield, oil yield, oil content, growth rate, biomass, vigor, ABST and/or NUE of a plant.

Table 16
Homologous polynucleotides and polypeptides

<i>Polynuc. SEQ ID NO:</i>	<i>Gene Name</i>	<i>Organism/Cluster name</i>	<i>Polypep. SEQ ID NO:</i>	<i>Homol. to SEQ ID NO:</i>	<i>% Global identity</i>	<i>Algor.</i>
203	BDL47_I10	radish gb164 EW717854	524	194	85.15	tblastn
204	BDL48_H0	radish gb164 EW735131	525	107	82.7	blastp
205	BDL62_H0	canola gb161 CD835773	526	108	94.5	blastp
206	BDL62_H1	radish gb164 EV543949	527	108	94.36	tblastn
207	BDL75_H0	canola gb161 EE485008	528	109	84.1	blastp
208	BDL83_H0	apple gb171 CN494551	529	112	86.8	blastp
209	BDL83_H1	aquilegia gb157.3 DR916286	530	112	85.9	blastp
210	BDL83_H2	artemisia gb164 EY037407	531	112	86.2	blastp
211	BDL83_H3	b_juncea gb164 EVGN00808312601672	532	112	91.7	blastp
212	BDL83_H4	b_oleracea gb161 AM387331	533	112	94.7	blastp
213	BDL83_H5	b_rapal gb162 AY161288	534	112	94.4	blastp
214	BDL83_H6	banana gb167 AF130251	535	112	85	blastp
215	BDL83_H7	barley gb157.3 BE420760	536	112	83.4	blastp
216	BDL83_H8	barley gb157.3 BG365169	537	112	82.1	blastp
217	BDL83_H9	bean gb167 CA896765	538	112	87.1	blastp
218	BDL83_H10	bean gb167 CB539815	539	112	87.4	blastp
219	BDL83_H11	beet gb162 BE590341	540	112	85.9	blastp
220	BDL83_H12	brachypodium gb169 BE213261	541	112	85.4	blastp
221	BDL83_H13	brachypodium gb169 BE419504	542	112	82.7	blastp
222	BDL83_H14	canola gb161 BNU20179	543	112	94.13	tblastn
223	BDL83_H15	canola gb161 CD818253	544	112	94.4	blastp
224	BDL83_H16	cassava gb164 DV445162	545	112	88	blastp
225	BDL83_H17	castorbean gb160 EE256791	546	112	87.4	blastp
226	BDL83_H18	centaurea gb166 EH734375	547	112	87.7	blastp
227	BDL83_H19	cichorium gb171 EH680019	548	112	88	blastp
228	BDL83_H20	citrus gb166 CV885954	549	112	89.15	tblastn
229	BDL83_H21	coffee gb157.2 DV685589	550	112	87.1	tblastn
230	BDL83_H22	cotton gb164 AI725778	551	112	86.8	blastp
231	BDL83_H23	cotton gb164 CA993334	552	112	88.3	blastp

<i>Polynuc. SEQ ID NO:</i>	<i>Gene Name</i>	<i>Organism/Cluster name</i>	<i>Polypep. SEQ ID NO:</i>	<i>Homol. to SEQ ID NO:</i>	<i>% Global identity</i>	<i>Algor.</i>
232	BDL83_H2 4	cowpealgb166 FF537383	553	112	87.7	blastp
233	BDI.83_H2 5	cynaralgb167 GE584395	554	112	84.16	tblastn
234	BDL83_H2 6	dandelionlgb161 DY8069 19	555	112	87.1	blastp
235	BDL83_H2 7	eucalyptuslgb166 CB9676 49	556	112	88	blastp
236	BDL83_H2 8	fescuelgb161 DT696580	557	112	82.8	blastp
237	BDL83_H2 9	gingerlgb164 DY345757	558	112	86.5	blastp
238	BDL83_H3 0	grapelgb160 CD008185	559	112	86.8	blastp
239	BDL83_H3 1	iceplantlgb164 CA833792	560	112	87.1	blastp
240	BDL83_H3 2	ipomoealgb157.2 CJ7555 28	561	112	86.8	blastp
241	BDL83_H3 3	lettuce gb157.2 AF16220 6	562	112	87.7	blastp
242	BDI.83_H3 4	leymuslgb166 EG375854	563	112	84	blastp
243	BDL83_H3 5	lovegrasslgb167 DN4818 48	564	112	82.8	blastp
244	BDL83_H3 6	maizelgb170 BG355384	565	112	83	blastp
245	BDL83_H3 7	maizelgb170 LLAI603703	566	112	83.5	blastp
246	BDL83_H3 8	medicagolgb157.2 AL386 990	567	112	80.5	blastp
247	BDL83_H3 9	medicagolgb157.2 AW69 5293	568	112	84.8	blastp
248	BDL83_H4 0	papayalgb165 EX248440	569	112	84.5	blastp
249	BDL83_H4 1	peanutlgb171 EE126296	570	112	86.8	blastp
250	BDL83_H4 2	peanutlgb171 ES752783	571	112	85.9	blastp
251	BDI.83_H4 3	pepperlgb171 CO907209	572	112	86.8	blastp
252	BDL83_H4 4	pepperlgb171 GD064098	573	112	87.4	blastp
253	BDL83_H4 5	physcomitrellalgb157 BJ1 57670	574	112	82.4	blastp
254	BDL83_H4 6	physcomitrellalgb157 BJ1 71093	575	112	83.63	tblastn
255	BDL83_H4 7	pinelgb157.2 AW010114	576	112	82.7	blastp
256	BDL83_H4 8	pinelgb157.2 CO169305	577	112	86.2	blastp
257	BDL83_H4 9	poplarlgb170 BI068614	578	112	88	blastp

<i>Polynuc. SEQ ID NO:</i>	<i>Gene Name</i>	<i>Organism/Cluster name</i>	<i>Polypep. SEQ ID NO:</i>	<i>Homol. to SEQ ID NO:</i>	<i>% Global identity</i>	<i>Algor.</i>
258	BDL83_H5 0	poplar gb170 IBU878945	579	112	86.05	tblastn
259	BDI.83_H5 1	potato gb157.2 BF053889	580	112	86.2	blastp
260	BDL83_H5 2	prunus gb167 DW341878	581	112	88.9	blastp
261	BDL83_H5 3	radish gb164 EV537348	582	112	95	blastp
262	BDL83_H5 4	radish gb164 EV565372	583	112	94.7	blastp
263	BDL83_H5 5	rice gb170 OS01G64660	584	112	84.5	blastp
264	BDL83_H5 6	rice gb170 OS05G36270	585	112	83.3	blastp
265	BDL83_H5 7	sorghum gb161.crplAW5 66083	586	112	84.8	blastp
266	BDL83_H5 8	sorghum gb161.crplAW6 71091	587	112	83.6	blastp
267	BDL83_H5 9	soybean gb168 AL388391	588	112	81.3	blastp
268	BDI.83_H6 0	soybean gb168 BE941320	589	112	86.5	blastp
269	BDL83_H6 1	soybean gb168 BF636881	590	112	87.1	blastp
270	BDL83_H6 2	soybean gb168 CD394856	591	112	86.2	blastp
271	BDL83_H6 3	spikemoss gb165 FE4436 31	592	112	83.9	blastp
272	BDL83_H6 4	spruce gb162 CO227794	593	112	87.4	blastp
273	BDL83_H6 5	spruce gb162 CO238226	594	112	83.3	blastp
274	BDL83_H6 6	spurge gb161 DV121804	595	112	87.4	blastp
275	BDL83_H6 7	strawberry gb164 DY671 211	596	112	87.4	blastp
276	BDL83_H6 8	sugarcane gb157.3 BQ533 620	597	112	85.4	blastp
277	BDI.83_H6 9	sunflower gb162 BU6720 90	598	112	84.2	blastp
278	BDL83_H7 0	sunflower gb162 CD8477 11	599	112	87.7	blastp
279	BDL83_H7 1	switchgrass gb167 DN143 181	600	112	84.8	blastp
280	BDL83_H7 2	switchgrass gb167 DN148 413	601	112	82.4	tblastn
281	BDL83_H7 3	tomato gb164 AI486777	602	112	88.3	blastp
282	BDL83_H7 4	tomato gb164 BG123415	603	112	85.9	blastp
283	BDL83_H7 5	triphysaria gb164 EY1662 97	604	112	85.9	blastp

<i>Polynuc. SEQ ID NO:</i>	<i>Gene Name</i>	<i>Organism/Cluster name</i>	<i>Polypep. SEQ ID NO:</i>	<i>Homol. to SEQ ID NO:</i>	<i>% Global identity</i>	<i>Algor.</i>
284	BDL83_H7 6	triphysarialgb164IEY1697 17	605	112	87.4	blastp
285	BDI.83_H7 7	wheatlgb164IBE213261	606	112	82.8	blastp
286	BDL83_II7 8	wheatlgb164IBE418868	607	112	82.8	blastp
287	BDL83_H7 9	wheatlgb164IBF500460	608	112	82.8	blastp
288	BDL118_H 0	castorbeanlgb160IMDL29 877M000477	609	114	81.4	blastp
289	BDL118_H 1	poplarlgb170IAI164922	610	114	81.3	blastp
290	BDL118_H 2	poplarlgb170IBI138300	611	114	82	blastp
291	BDL118_H 3	soybeanlgb168IBE658051	612	114	80.1	blastp
292	BDL118_H 4	soybeanlgb168ICB540069	613	114	80	blastp
293	BDL138_H 0	canolalgb161ICD817345	614	116	92.5	blastp
294	BDI.138_H 1	radishlgb164IEV536083	615	116	91.8	blastp
295	BDL138_II 2	radishlgb164IEW725583	616	116	91.1	blastp
296	BDL147_H 0	thellungiellalgb167IBY81 8222	617	118	81.6	blastp
297	BDL149_H 0	b_rapalgb162IDY009670	618	119	87.9	blastp
298	BDL149_H 1	canolalgb161ICD818601	619	119	88.5	blastp
299	BDL149_H 2	cowpealgb166IFF400239	620	119	82.4	blastp
300	BDL154_H 0	b_rapalgb162IEX046206	621	122	90.77	tblastn
301	BDL154_H 1	canolalgb161ICD841052	622	122	92.25	tblastn
302	BDL155_H 0	arabidopsislgb165IAT1G2 4240	623	123	92.9	blastp
303	BDI.155_H 1	arabidopsislgb165IAT5G1 1750	624	123	91.7	blastp
304	BDL155_II 2	canolalgb161IEE462449	625	123	81.3	blastp
305	BDL155_H 3	radishlgb164IEV545009	626	123	80.34	tblastn
306	BDL155_H 4	radishlgb164IEV569478	627	123	80.4	blastp
307	BDL158_H 0	radishlgb164IEW715818	628	126	88.46	tblastn
308	BDL160_H 0	canolalgb161IEE418738	629	127	83.9	blastp
309	BDL160_H 1	radishlgb164IEV545765	630	127	85.1	blastp

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310	BDL167_H 0	radish gb164 EV535056	631	196	82.7	tblastn
311	BDL168_H 0	radish gb164 EV525399	632	132	90.5	blastp
312	BDL171_II 0	arabidopsis gb165 AT2G3 1580	633	134	84	blastp
313	BDL182_H 0	canola gb161 CX188804	634	140	87.5	blastp
314	BDL183_H 0	arabidopsis gb165 AT4G1 5630	635	141	81.6	blastp
315	BDL183_H 1	b_rapa gb162 EE526726	636	141	83.7	blastp
316	BDL183_H 2	canola gb161 CD811977	637	141	80	blastp
317	BDL183_H 3	canola gb161 CN735162	638	141	83.7	blastp
318	BDL183_H 4	canola gb161 CN828640	639	141	83.7	blastp
319	BDL183_H 5	radish gb164 EW722612	640	141	83.2	blastp
320	BDL183_H 6	radish gb164 EX753993	641	141	81.6	blastp
321	BDL183_II 7	thellungiella gb167 BY82 4379	642	141	80	blastp
322	BDL190_H 0	b_oleracea gb161 AM395 197	643	146	84.49	tblastn
323	BDL190_H 1	canola gb161 EE407003	644	146	87.2	blastp
324	BDL190_H 2	radish gb164 EV547395	645	146	86.6	blastp
325	BDL190_H 3	thellungiella gb167 BY82 7749	646	146	89.8	blastp
326	BDL192_H 0	canola gb161 CD827541	647	147	80.37	tblastn
327	BDL201_H 0	b_junca gb164 EVLGN00 584109703185	648	153	81.8	blastp
328	BDL201_H 1	b_oleracea gb161 DY014 784	649	153	80.4	blastp
329	BDL201_H 2	b_oleracea gb161 DY015 288	650	153	82.2	blastp
330	BDL201_II 3	b_rapa gb162 BQ791239	651	153	83.2	blastp
331	BDL201_H 4	canola gb161 BQ704590	652	153	81.7	blastp
332	BDL201_H 5	canola gb161 CD822183	653	153	80.8	blastp
333	BDL201_H 6	canola gb161 CD838597	654	153	81.8	blastp
334	BDL201_H 7	radish gb164 EV527287	655	153	81.6	blastp
335	BDL201_H 8	radish gb164 EV534907	656	153	81.8	blastp

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336	BDL201_H 9	radish gb164 EV543313	657	153	81.3	blastp
337	BDI.201_H 10	radish gb164 EV565783	658	153	81.9	blastp
338	BDL201_II 11	thellungiella gb167 BY81 4893	659	153	83.89	tblastn
339	BDL223_H 0	arabidopsis gb165 AT3G5 4080	660	159	82.05	tblastn
340	BDL223_H 1	arabidopsis gb165 AT3G5 4085	661	159	82.1	blastp
341	BDL223_H 2	b_juncea gb164 EVGN01 441114591370	662	159	96.15	tblastn
342	BDL223_H 3	b_juncea gb164 EVGN05 602102561055	663	159	85	blastp
343	BDL223_H 4	b_oleracea gb161 EE5349 59	664	159	96.2	blastp
344	BDL223_H 5	b_oleracea gb161 ES9476 77	665	159	94.9	blastp
345	BDL223_H 6	b_rapal gb162 EE527700	666	159	96.2	blastp
346	BDI.223_H 7	canola gb161 CD812251	667	159	96.2	blastp
347	BDL223_II 8	canola gb161 CN734091	668	159	96.15	tblastn
348	BDL223_H 9	canola gb161 DY007214	669	159	96.15	tblastn
349	BDL223_H 10	canola gb161 EG019597	670	159	97.4	blastp
350	BDL223_H 11	canola gb161 ES992154	671	159	97.4	blastp
351	BDL223_H 12	canola gb161 EV087632	672	159	97.4	blastp
352	BDL223_H 13	radish gb164 EV524950	673	159	96.15	tblastn
353	BDL223_H 14	radish gb164 EY899056	674	159	96.2	blastp
354	BDL229_H 0	b_rapal gb162 EX066757	675	160	83.5	blastp
355	BDI.229_H 1	radish gb164 EW725388	676	160	82.59	tblastn
356	BDL231_II 0	arabidopsis gb165 AT1G7 9470	677	162	84.5	blastp
357	BDL231_H 1	canola gb161 CD826885	678	162	84.7	blastp
358	BDL231_H 2	canola gb161 CD827559	679	162	95.42	tblastn
359	BDL242_H 0	radish gb164 EV539529	680	164	85.9	blastp
360	BDL247_H 0	b_oleracea gb161 DY026 136	681	167	90.82	tblastn
361	BDL247_H 1	b_rapal gb162 AY185359	682	167	91.1	blastp

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362	BDL247_H 2	canolalgb161ICD838112	683	167	89.1	blastp
363	BDL247_H 3	canolalgb161IEE455228	684	167	91.1	blastp
364	BDL247_H 4	maizelgb170ILLEU94083 3	685	167	92.8	blastp
365	BDL247_H 5	radishlgb164IEV566311	686	167	92.6	blastp
366	BDL247_H 6	thellungiellalgb167IDN77 3706	687	167	93.08	tblastn
367	BDL248_H 0	arabidopsislgb165IAT4G3 7340	688	168	80.6	blastp
368	BDL70_H0	arabidopsislgb165IAT4G2 5960	689	201	80.6	blastp
369	BDL70_H1	poplarlgb170ICN192983	690	201	82.8	blastp
370	BDL70_H2	soybeanlgb168IBE822547	691	201	81.9	blastp
371	BDL70_H3	soybeanlgb168ICD415929	692	201	81.8	blastp
372	BDL85_H0	b_oleracealgb161IAM385 973	693	175	83.3	blastp
373	BDL85_H1	b_rapalgb162ICV544456	694	175	83.3	blastp
374	BDL85_H2	b_rapalgb162IEE523194	695	175	84.7	blastp
375	BDL85_H3	canolalgb161ICD813661	696	175	83.3	blastp
376	BDL85_H4	canolalgb161ICD822271	697	175	84.7	blastp
377	BDL85_H5	canolalgb161ICX280350	698	175	83.3	blastp
378	BDL85_H6	radishlgb164IEV527759	699	175	82.2	blastp
379	BDL85_H7	thellungiellalgb167IDN77 9034	700	175	83.6	blastp
380	BDL88_H0	barleylgb157.3IBI950587	701	176	82.3	blastp
381	BDL88_H1	barleylgb157.3IBI955547	702	176	88.6	blastp
382	BDL88_H2	brachypodiumlgb169IBE4 25841	703	176	82.7	blastp
383	BDL88_H3	brachypodiumlgb169IBM 817094	704	176	90	blastp
384	BDL88_H4	fescuelgb161ICK802755	705	176	88.7	blastp
385	BDL88_H5	leymuslgb166IEG378145	706	176	83.2	blastp
386	BDL88_H6	maizelgb170IAI622555	707	176	89.9	blastp
387	BDL88_H7	maizelgb170IAI712236	708	176	89.2	blastp
388	BDL88_H8	maizelgb170IAI855432	709	176	80.7	blastp
389	BDL88_H9	pseudoroegneriaalgb167IF F342352	710	176	88	blastp
390	BDL88_H 0	pseudoroegneriaalgb167IF F342444	711	176	82.3	blastp
391	BDL88_H 1	ricelgb170IOS08G43320	712	176	84.5	blastp
392	BDL88_H 2	sorghumlgb161.crplAW5 62977	713	176	81.4	blastp
393	BDL88_H 3	sorghumlgb161.crplBE12 9901	714	176	91.1	blastp
394	BDL88_H 4	sugarcanelgb157.3ISCFM EMPRO	715	176	91.4	blastp
395	BDL88_H 5	switchgrasslgb167IDN144 780	716	176	91.7	blastp

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396	BDL88_H1 6	switchgrass gb167 FE629 824	717	176	92	blastp
397	BDL88_H1 7	wheat gb164 BE406463	718	176	88	blastp
398	BDL88_H1 8	wheat gb164 BE425841	719	176	82	blastp
399	BDL88_H1 9	wheat gb164 BF474328	720	176	81.2	blastp
400	BDL88_H2 0	wheat gb164 BQ484144	721	176	87.7	blastp
401	BDL88_H2 1	wheat gb164 CA599299	722	176	88	blastp
402	BDL94_H0	rice gb170 OS01G09220	723	177	96.7	blastp
403	BDL102_H 0	avocado gb164 CK76710 8	724	178	80.69	tblastn
404	BDL102_H 1	banana gb167 FL650176	725	178	82.8	blastp
405	BDL102_H 2	banana gb167 FL657720	726	178	80.4	blastp
406	BDL102_H 3	banana gb167 FL658383	727	178	83.4	blastp
407	BDL102_H 4	barley gb157.3 BE215743	728	178	84.83	tblastn
408	BDL102_H 5	barley gb157.3 BE411917	729	178	85.2	blastp
409	BDL102_H 6	barley gb157.3 BF627967	730	178	82.76	tblastn
410	BDL102_H 7	brachypodium gb169 BE3 98478	731	178	83.7	blastp
411	BDL102_H 8	brachypodium gb169 BE3 99017	732	178	87.6	blastp
412	BDL102_H 9	brachypodium gb169 BE4 23566	733	178	85.5	blastp
413	BDL102_H 10	cenchrus gb166 EB65550 9	734	178	89	blastp
414	BDL102_H 11	cichorium gb171 DT2119 67	735	178	80.1	blastp
415	BDL102_H 12	cichorium gb171 EH7006 64	736	178	80	blastp
416	BDL102_H 13	citrus gb166 BE208879	737	178	80	blastp
417	BDL102_H 14	clover gb162 BB906663	738	178	80	blastp
418	BDL102_H 15	cowpea gb166 FC462243	739	178	80.7	blastp
419	BDL102_H 16	dandelion gb161 DY8137 50	740	178	80.7	blastp
420	BDL102_H 17	fescue gb161 DT685902	741	178	85.6	blastp
421	BDL102_H 18	ginger gb164 DY369602	742	178	80.7	blastp
422	BDL102_H 19	ipomoea gb157.2 BJ5570 23	743	178	80	tblastn

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423	BDL102_H 20	ipomoealgb157.2 BU6907 07	744	178	80	tblastn
424	BDL102_H 21	lettucelgb157.2 DW04478 6	745	178	80	blastp
425	BDL102_H 22	lettucelgb157.2 DW05075 7	746	178	80	tblastn
426	BDL102_H 23	lettucelgb157.2 DW10880 9	747	178	81.38	tblastn
427	BDL102_H 24	lettucelgb157.2 DW10881 0	748	178	80	tblastn
428	BDL102_H 25	liquoricegb171 FS238664	749	178	80.7	blastp
429	BDL102_H 26	liquoricegb171 FS241892	750	178	80	tblastn
430	BDL102_H 27	liriodendrongb166 CK76 6596	751	178	83.4	blastp
431	BDL102_H 28	lotusgb157.2 AW720301	752	178	82.1	blastp
432	BDL102_H 29	lotusgb157.2 CN825142	753	178	81.5	blastp
433	BDL102_H 30	lovegrassgb167 EH18838 8	754	178	88.97	tblastn
434	BDL102_H 31	maizegb170 AI391795	755	178	95.2	blastp
435	BDL102_H 32	maizegb170 LLBE51025 4	756	178	95.2	blastp
436	BDL102_H 33	melongb165 DV632852	757	178	80	tblastn
437	BDL102_H 34	milletgb161 CD725312	758	178	85.7	blastp
438	BDL102_H 35	nuphargb166 CV003178	759	178	81.4	blastp
439	BDL102_H 36	oatgb164 CN815719	760	178	91.03	tblastn
440	BDL102_H 37	oil_palmgb166 EL68109 8	761	178	86.21	tblastn
441	BDL102_H 38	oil_palmgb166 EL68263 0	762	178	84.1	blastp
442	BDL102_H 39	oil_palmgb166 EL68411 9	763	178	82.1	blastp
443	BDL102_H 40	oniongb162 CF446214	764	178	80.4	blastp
444	BDL102_H 41	papayalgb165 EX258155	765	178	80.7	blastp
445	BDL102_H 42	pineapplegb157.2 DT336 701	766	178	84.8	blastp
446	BDL102_H 43	pineapplegb157.2 DT338 401	767	178	84.8	blastp
447	BDL102_H 44	poppygb166 FE964559	768	178	80	tblastn
448	BDL102_H 45	ricegb170 OS03G31090	769	178	93.8	blastp

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449	BDL102_H 46	ryelgb164 BE637001	770	178	85.2	blastp
450	BDL102_H 47	sorghum gb161.crplAI673 920	771	178	96.6	blastp
451	BDL102_H 48	sorghum gb161.crplAW7 47023	772	178	83.3	blastp
452	BDL102_H 49	soybean gb168 AA66103 6	773	178	80	blastp
453	BDL102_H 50	soybean gb168 AW72030 1	774	178	80	tblastn
454	BDL102_H 51	sugarcane gb157.3 CA076 952	775	178	96.6	blastp
455	BDL102_H 52	sugarcane gb157.3 CA087 422	776	178	96.6	blastp
456	BDL102_H 53	sugarcane gb157.3 CA103 720	777	178	97.2	blastp
457	BDL102_H 54	sugarcane gb157.3 CA113 942	778	178	84.25	tblastn
458	BDL102_H 55	sugarcane gb157.3 CA114 406	779	178	97.2	blastp
459	BDL102_H 56	sugarcane gb157.3 CA116 867	780	178	87.6	blastp
460	BDL102_H 57	sugarcane gb157.3 CA119 956	781	178	90.34	tblastn
461	BDL102_H 58	sugarcane gb157.3 CA128 680	782	178	86.9	blastp
462	BDL102_H 59	sugarcane gb157.3 CA139 681	783	178	84.3	blastp
463	BDL102_H 60	sugarcane gb157.3 CA151 882	784	178	93.1	tblastn
464	BDL102_H 61	sugarcane gb157.3 CA153 401	785	178	96.6	blastp
465	BDL102_H 62	sugarcane gb157.3 CA193 794	786	178	97.2	blastp
466	BDL102_H 63	sugarcane gb157.3 CA215 946	787	178	80	blastp
467	BDL102_H 64	sugarcane gb157.3 CA235 573	788	178	80.3	blastp
468	BDL102_H 65	sugarcane gb157.3 CA241 742	789	178	82.76	tblastn
469	BDL102_H 66	sugarcane gb157.3 CA289 186	790	178	95.2	blastp
470	BDL102_H 67	sugarcane gb157.3 CF571 967	791	178	93.79	tblastn
471	BDL102_H 68	sugarcane gb157.3 CF574 520	792	178	97.2	blastp
472	BDL102_H 69	sunflower gb162 CD8485 88	793	178	80	blastp
473	BDL102_H 70	sunflower gb162 CD8509 71	794	178	80.7	blastp
474	BDL102_H 71	switchgrass gb167 DN142 384	795	178	95.9	blastp

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475	BDL102_H 72	switchgrass gb167 FE598 349	796	178	96.6	blastp
476	BDL102_H 73	switchgrass gb167 FE608 943	797	178	96.6	blastp
477	BDL102_H 74	switchgrass gb167 FE642 870	798	178	96.6	blastp
478	BDL102_H 75	switchgrass gb167 FL710 664	799	178	95.2	blastp
479	BDL102_H 76	teal gb171 FE861343	800	178	80.14	tblastn
480	BDL102_H 77	walnut gb166 CV195103	801	178	80.7	blastp
481	BDL102_H 78	walnut gb166 CV196374	802	178	80.7	blastp
482	BDL102_H 79	wheat gb164 BE398202	803	178	85.2	blastp
483	BDL102_H 80	wheat gb164 BE406254	804	178	85.2	blastp
484	BDL102_H 81	wheat gb164 BE414893	805	178	85.2	blastp
485	BDL102_H 82	wheat gb164 CA485952	806	178	96.6	blastp
486	BDL102_H 83	wheat gb164 CA486424	807	178	93.8	blastp
487	BDL226_H 0	canola gb161 CD835072	808	181	89.5	blastp
488	BDL226_H 1	radish gb164 EV568139	809	181	91.7	blastp
489	BDL233_H 0	radish gb164 EV549343	810	185	84.5	blastp
490	BDL237_H 0	arabidopsis gb165 AT2G4 4890	811	187	86.3	blastp
491	BDL238_H 0	arabidopsis gb165 AT1G3 2763	812	202	81.5	blastp
492	BDL240_H 0	castorbean gb160 MDL30 174M008707	813	189	82.4	blastp
493	BDL240_H 1	cotton gb164 BG446934	814	189	81.4	blastp
494	BDL240_H 2	soybean gb168 AW32969 3	815	189	80.7	blastp
495	BDL240_H 3	soybean gb168 BE329627	816	189	80.1	blastp
496	BDL241_H 0	canola gb161 CN825973	817	190	85.3	blastp
497	BDL249_H 0	radish gb164 EW735252	818	191	86.7	blastp
498	BDL251_H 0	arabidopsis gb165 AT5G2 3190	819	192	80.07	tblastn
499	BDL252_H 0	apple gb171 CN489978	820	193	80.9	blastp
500	BDL252_H 1	apple gb171 CN883406	821	193	80.9	blastp

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501	BDL252_H 2	b_rapalgb162 EX036871	822	193	91	blastp
502	BDI.252_H 3	cacaolgb167 CU476642	823	193	84.3	blastp
503	BDL252_II 4	canolalgb161 CN732395	824	193	90.6	blastp
504	BDL252_H 5	canolalgb161 FS900668	825	193	86.2	blastp
505	BDL252_H 6	castorbeanlgb160 MDL29 726M003996	826	193	81.4	blastp
506	BDL252_H 7	citruslgb166 CB417408	827	193	82.8	blastp
507	BDL252_H 8	cottonlgb164 AI725830	828	193	82.8	blastp
508	BDL252_H 9	cottonlgb164 CO094656	829	193	82.5	blastp
509	BDL252_H 10	cowpealgb166 FC458375	830	193	80.5	blastp
510	BDL252_H 11	grapelgb160 CB920145	831	193	80.5	blastp
511	BDI.252_H 12	ipomoealgb157.2 AU2238 26	832	193	80.2	blastp
512	BDL252_II 13	lettucelgb157.2 DW11675 3	833	193	80.22	tblastn
513	BDL252_H 14	medicagolgb157.2 AL371 996	834	193	80.9	blastp
514	BDL252_H 15	peachlgb157.2 BU039377	835	193	82.4	blastp
515	BDL252_H 16	poplarlgb170 BU816161	836	193	80.7	blastp
516	BDL252_H 17	prunuslgb167 BU039377	837	193	82.4	blastp
517	BDL252_H 18	radishlgb164 EV535608	838	193	92.1	blastp
518	BDL252_H 19	radishlgb164 EW723334	839	193	91.8	blastp
519	BDL252_H 20	soybeanlgb168 BF636462	840	193	80.1	blastp
520	BDI.252_H 21	soybeanlgb168 BU546186	841	193	82.4	blastp
521	BDL252_II 22	soybeanlgb168 CD390334	842	193	82.4	blastp
522	BDL252_H 23	strawberylgb164 CX6613 79	843	193	82	blastp
523	BDL252_H 24	thellungiellalgb167 BY81 2142	844	193	92.1	blastp

Table 16: Provided are polynucleotides and polypeptides which are homologous to the identified polynucleotides or polypeptides of Table 15. Homol. = homologue; Algor. = Algorithm;

EXAMPLE 7**GENE CLONING AND GENERATION OF BINARY VECTORS FOR PLANT
EXPRESSION**

To validate their role in improving oil content, plant yield, seed yield,
5 biomass, growth rate, ABST, NUE and/or vigor, selected genes were over-expressed in
plants, as follows.

Cloning strategy

Genes listed in Examples 5 and 6 hereinabove were cloned into binary vectors
for the generation of transgenic plants. For cloning, the full-length open reading frame
10 (ORF) was first identified. In case of ORF-EST clusters and in some cases already
published mRNA sequences were analyzed to identify the entire open reading frame by
comparing the results of several translation algorithms to known proteins from other
plant species. To clone the full-length cDNAs, reverse transcription (RT) followed by
polymerase chain reaction (PCR; RT-PCR) was performed on total RNA extracted from
15 leaves, flowers, siliques or other plant tissues, growing under normal conditions. Total
RNA was extracted as described in Example 2 above. Production of cDNA and PCR
amplification was performed using standard protocols described elsewhere (Sambrook
J., E.F. Fritsch, and T. Maniatis. 1989. Molecular Cloning. A Laboratory Manual., 2nd
Ed. Cold Spring Harbor Laboratory Press, New York.) which are well known to those
20 skilled in the art. PCR products were purified using PCR purification kit (Qiagen). In
case where the entire coding sequence was not found, RACE kit from Ambion,
Clontech or Invitrogen (RACE = Rapid Access to cDNA Ends) was used to access the
full cDNA transcript of the gene from the RNA samples described above. The RACE
procedure was performed for the genes BDL197 (SEQ ID NO:46), BDL156 (SEQ ID
25 NO:19), BDL169 (SEQ ID NO:28), BDL189 (SEQ ID NO:40), BDL200 (SEQ ID
NO:47), BDL79 (SEQ ID NO:5), BDL231 (SEQ ID NO:57), BDL238 (SEQ ID NO:84)
and BDL248 (SEQ ID NO:103) using the primers sequences listed in Table 17, below.
RACE products were cloned into high copy vector followed by sequencing or directly
sequenced. RACE products were sequenced as described hereinbelow for the genes
30 specified in Table 17.

The information from the RACE procedure was used for cloning of the full
length ORF of the corresponding genes.

Table 17
RACE primers used for sequencing of the identified genes of the invention

5

<i>Gene Name</i>	<i>Primers used for amplification</i>	<i>High copy plasmid used for cloning of RACE products</i>
BDL197_5'Race	BDL197_NGSP1_R2 (SEQ ID NO:934) (CGCCTGAAGCTTCTCCGAGAAC)	TopoTA
BDL156_5'Race	BDL156_NGSP1_R (SEQ ID NO:937) (ACGGTGTTCAGAAATTCGCGAG)	
BDL156_3'Race	BDL156_NGSP2_F (SEQ ID NO:938) (CTGAATGTGACTGGGATATGTCCGG)	TopoTA
BDL169_5'Race	BDL169_NGSP1_R (SEQ ID NO:939) (TGCACTGGAGCGTGTAGGGACAG)	
BDL169_3'Race	BDL169_NGSP1_F (SEQ ID NO:940) (GGCAGAAACTGTTTTATGGAAATGG)	
BDL189_5'Race	BDL189_NGSP1_R (SEQ ID NO:941) (TTCCTGTCCTTCGACCAAGGTTG)	TopoTA
BDL189_3'Race	BDL189_GSP2_F (SEQ ID NO:942) (CAGTCAATCTTCTTAGCATCGCTGAG)	
BDL200_5'Race	BDL200_NGSP_R6 (SEQ ID NO:943) (CCAATGCCAATACGATGGTCCGG)	TopoTA
BDL200_3'Race	BDL200_NGSP2_F (SEQ ID NO:944) (GAGCTTTGGAGATAAGATTGGTGCAG)	
BDL79_5'Race	BDL79_GSP1_R (SEQ ID NO:945) (GTATCACTCGAGGCACCATGGG)	TopoTA
BDL231_3'Race	BDL231_NGSP_R (SEQ ID NO:946) (ATGTGGGATTCCGAGACAGTGTCC)	TopoTA
BDL238_5'Race	BDL238_NGSP_R (SEQ ID NO:947) (TTTACCGTCCCCAAACGTTGCCG)	
BDL238_3'Race	BDL238_NGSP_F (SEQ ID NO:948) (CTCATCCGGACGATGTCTTACTTCTCTCC)	
BDL248_5'Race	BDL248_NGSP_R (SEQ ID NO:949) (GAGGTGACCGATCACTGGTAACGC)	
BDL215_5'Race	BDL215_NGSP1_R (SEQ ID NO:950) (TGGCTTTGAAAACGTAACATGCC)	
BDL215_3'Race	BDL215_NGSP2_F (SEQ ID NO:951) (TATTGGGATTTTCGGATCGATGG)	TopoTA

Table 17. Provided are the PCR primers used for RACE sequencing. Fwd = forward primer; Rev = reverse primer;

10 In case genomic DNA was cloned, as in the case of BDL90 and BDL94 the genes were amplified by direct PCR on genomic DNA extracted from leaf tissue using the DNAeasy kit (Qiagen Cat. No. 69104).

Usually, 2 sets of primers were synthesized for the amplification of each gene from a cDNA or a genomic sequence; an external set of primers and an internal set (nested PCR primers). When needed (e.g., when the first PCR reaction did not result in

a satisfactory product for sequencing), an additional primer (or two) of the nested PCR primers were used. Table 18 below provides primers used for cloning of selected genes.

5

Table 18
The PCR primers used for cloning the genes of the invention

Gene Name	Restriction Enzymes used for cloning	Primers used for amplification
BDL102	SalI, XbaI	BDL102_NF_SalI(SEQ ID NO:952) AATGTCGACTACCTGCCTTTCTCTCGTCC
		BDL102_EF_SalI(SEQ ID NO:953) AAAGTCGACACTCTACCTGCCTTTCTCTCG
		BDL102_NR_XbaI(SEQ ID NO:954) ATTCTAGACTATGTAGCCATCTCAACAATCAAAC
		BDL102_ER_XbaI(SEQ ID NO:955) ATTCTAGAGGTTTTGATAAATAGGTACTCAGG
BDL117		BDL117_EF_SmaI(SEQ ID NO:956) CCCCGGTCTCGGAGGTATCTTATTCCAG
		BDL117_ER_SmaI(SEQ ID NO:957) CCCCGGGATGCCACACTTAAGCCAAG
BDL118	SmaI, SmaI	BDL118_NF_SmaI(SEQ ID NO:958) TCCCCGGTCTGGGTCTACTTTTGATTTGAG
		BDL118_NR_SmaI(SEQ ID NO:959) TCCCCGGTGAAGCAGAAGTTTCGATTTAAG
BDL138	SalI, XbaI	BDL138_NF_SalI(SEQ ID NO:960) AAAGTCGACCGAATCGTAATTGTTGAAGAGAG
		BDL138_EF_SalI(SEQ ID NO:961) ATTGTCGACTTTAAGGAGAAGAGTCGCAGTC
		BDL138_NR_XbaI(SEQ ID NO:962) ATTCTAGATTAGAGAGTGGTTGATAACGCAGAG
		BDL138_ER_XbaI(SEQ ID NO:963) ACTCTAGACTACCTGTCACAATTTTCTAAAACAC
BDL140	XbaI, SacI	BDL140_NF_XbaI(SEQ ID NO:964) TATCTAGATTGAGAATGAACTCAGTGTGTATC
		BDL140_EF_XbaI(SEQ ID NO:965) AATCTAGAAACTAAACATTGAGAATGAACTCAG
		BDL140_NR_SacI(SEQ ID NO:966) TGAGCTCTTAAGTCATTTAGTTTGGATCAACAAC
		BDL140_ER_SacI(SEQ ID NO:967) CGAGCTCAGACCATGCATTTAAGGATCAC
BDL147	SalI, XbaI	BDL147_NF_SalI(SEQ ID NO:968) TTAGTCGACCACAGTAACCATGICCGTCTC
		BDL147_NR_XbaI(SEQ ID NO:969) TATCTAGAGTGCTGCTTACTCGCTGTTC
BDL149	SalI, XbaI	BDL149_NF_SalI(SEQ ID NO:970) AAAGTCGACCTCAAACCCAAGAACCTCATC
		BDL149_NR_XbaI(SEQ ID NO:971) ATTCTAGATGCAATAGTAGTAGCAGTGAACC
BDL152	XbaI, SacI	BDL152_NF_XbaI(SEQ ID NO:972) TATCTAGATTCAGACAAAACAGAGAGAACT
		BDL152_NR_SacI(SEQ ID NO:973) TGAGCTCCTAAGATCGGTTTAATCAATAGGG
BDL153	SalI, SmaI	BDL153_NF_SalI(SEQ ID NO:974) AAAGTCGACAACAGCTTCGGTTTAAGAGTTC

<i>Gene Name</i>	<i>Restriction Enzymes used for cloning</i>	<i>Primers used for amplification</i>
		BDL153_NF_SmaI(SEQ ID NO:975) TCCCGGGTCTACATTACGGCATAACGGC
BDL154	SalI, SacI	BDL154_NF_SalI(SEQ ID NO:976) TTAGTCGACTTAAAAATGGAGACTCAAAAGC
		BDL154_NR_SacI(SEQ ID NO:977) TGAGCTCCTACTACTTCTTGTGATGCTGAGG
BDL155	SalI, XbaI	BDL155_NF_SalI(SEQ ID NO:978) AATGTCGACTCCTCTTGCAGAGAGATGC
		BDL155_NR_XbaI(SEQ ID NO:979) ATTCTAGATCTCCTTTTGAGAGAGTGCAAC
BDL156	SalI, XbaI	BDL156_NF_SalI(SEQ ID NO:980) AATGTCGACTCGTCGTCTTCTCATTTCG
		BDL156_ER_XbaI(SEQ ID NO:981) ATTCTAGACTAATACGATTGGTAACAAGAAAACG
BDL157	SalI, XbaI	BDL157_NF_SalI(SEQ ID NO:982) ATTGTCGACTTCAATAAGAAATCTGCGTCC
		BDL157_EF_SalI(SEQ ID NO:983) AAAGTCGACGAATCTGCTTTTAAGCTTCTCG
		BDL157_NR_XbaI(SEQ ID NO:984) ATTCTAGACTAAAGAGAGTGAAGGAACAAAGACC
		BDL157_ER_XbaI(SEQ ID NO:985) ATTCTAGACTATTTCTTCTGTCTTCTGTGTCTTC
		BDL158_EF_XbaI(SEQ ID NO:986) AATCTAGACCTCACTTCTCTCTCTCTCTTC
BDL158		BDL158_ER_SmaI(SEQ ID NO:987) CCCCGGGAGCCTAAAGCCTAACCCAAC
		BDL160_NF_SalI(SEQ ID NO:988) AAAGTCGACTGATCTACACAGAATCCATTTC
BDL160	SalI, XbaI	BDL160_NR_XbaI(SEQ ID NO:989) AATCTAGATCATTACGCCATTACATTTTAGG
		BDL162_NF_SalI(SEQ ID NO:990) TTAGTCGACCCTAATAATGGCTTGCAGAGC
BDL162	SalI, XbaI	BDL162_NR_XbaI(SEQ ID NO:991) TATCTAGAAAATCTTGAGACTAAATCAAGCTG
		BDL167_NF_SalI(SEQ ID NO:992) AAAGTCGACGCAAGAAAGGGACTAACCAAG
BDL167	SalI, XbaI	BDL167_NR_XbaI(SEQ ID NO:993) ACTCTAGACTATGTCCGCACTTAACCTTAGAATCAC
		BDL168_NF_XbaI(SEQ ID NO:994) AATCTAGACTTGCTTCAAGATTGAGTGAG
BDL168	XbaI, SmaI	BDL168_EF_XbaI(SEQ ID NO:995) AATCTAGAATTAACCACTTCTGTGAAG
		BDL168_NR_SmaI(SEQ ID NO:996) TCCCGGGCTACCTTCTTCTTCTTCACTTCC
		BDL168_ER_SmaI(SEQ ID NO:997) TCCCGGGTCTTAAAGTCAGTCACCTTCTG
		BDL169_NF_SalI(SEQ ID NO:998) AAAGTCGACGAAGGTGAAGTGATGGATTCTG
		BDL169_EF_SalI(SEQ ID NO:999) AATGTCGACTGTTACCGATAAGAAGGTGAAG
BDL169	SalI, XbaI	BDL169_NR_XbaI(SEQ ID NO:1000) ATTCTAGACTAACAGCTTCAACGTAATTTGGTG

<i>Gene Name</i>	<i>Restriction Enzymes used for cloning</i>	<i>Primers used for amplification</i>
		BDL169_ER_XbaI(SEQ ID NO:1001) ATTCTAGATCAACGTCATTTTGTGCATATC
BDL173	XbaI, SmaI	BDL173_FF_XbaI(SEQ ID NO:1002) ATTCTAGATTTTCCCGAATCTATTCATCAC
		BDL173_ER_SmaI(SEQ ID NO:1003) CCCCGGGAACGCTTCACCCTTTAATCC
BDL174	Sall, SacI	BDL174_NF_Sall(SEQ ID NO:1004) AAAGTCGACCCGAGGAAGATGACGACAC
		BDL174_NR_SacI(SEQ ID NO:1005) TGAGCTCCTAGTTTCAAGCAAGAGTGATTCC
BDL176	SmaI, SmaI	BDL176_NF_SmaI(SEQ ID NO:1006) TCCCGGGTATTGAGCAGCCGTGAAATC
		BDL176_NR_SmaI(SEQ ID NO:1007) TCCCGGGTGGACCAAAGAATCAAATAGTAAC
BDL177	Sall, SacI	BDL177_NF_Sall(SEQ ID NO:1008) AATGTGCGACTCAGGATCTATGGCAAGTC
		BDL177_FF_Sall(SEQ ID NO:1009) AAAGTCGACGCTTGTGATCAGGATCTATGG
		BDL177_NR_SacI(SEQ ID NO:1010) TGAGCTCCTAAACAGCTTTCTTCTACTCTTCATC
		BDL177_ER_SacI(SEQ ID NO:1011) CGAGCTCACAGAAAACAAAAGAAACTAGGC
BDL181	Sall, SacI	BDL181_NF_Sall(SEQ ID NO:1012) AAAGTCGACGCGATAGATCTGACGAATGC
		BDL181_NR_SacI(SEQ ID NO:1013) TGAGCTCCTAGATTTTCATACTCAGGAAGCCAC
BDL182	Sall, SacI	BDL182_NF_Sall(SEQ ID NO:1014) AACGTCGACTCTACCATCGACAACGAGAAAC
		BDL182_NR_SacI_new(SEQ ID NO:1015) TGAGCTCCTACATTCACAACAAACCACCACTAC
BDL183	Sall, XbaI	BDL183_NF_Sall(SEQ ID NO:1016) AATGTGCGACTTATTTTGATCTTCTCCTCACTTCTG
		BDL183_FF_Sall(SEQ ID NO:1017) AAAGTCGACGATCAATCTTTGTTATCTCTCACTC
		BDL183_NR_XbaI(SEQ ID NO:1018) ATTCTAGACTAATCACACAAAACGACAAGAACAG
		BDL183_ER_XbaI(SEQ ID NO:1019) ACTCTAGAACGATGTGATAAAACATTAGAAGC
BDL186	Sall, XbaI	BDL186_NF_Sall(SEQ ID NO:1020) AAAGTCGACCGAAGTAAAAGTCGTGATGG
		BDL186_FF_Sall(SEQ ID NO:1021) AAAGTCGACACGCAAACGTGATCCTAAAC
		BDL186_NR_XbaI(SEQ ID NO:1022) ATTCTAGACTCAAGGGGACGAGATATCAG
		BDL186_ER_XbaI(SEQ ID NO:1023) ACTCTAGAAGGTAGAGAGCATCAAGGAAGC
BDL187	Sall, SmaI	BDL187_NF_Sall(SEQ ID NO:1024) ATAGTCGACATTCTTTCAGTTTCCGGTG
		BDL187_FF_Sall(SEQ ID NO:1025) AAAGTCGACGCTTGAATTCCTTTCAGTTTCC
		BDL187_NR_SmaI(SEQ ID NO:1026) TCCCGGGCTATTTTCACCAGTAATTTCCACAC

<i>Gene Name</i>	<i>Restriction Enzymes used for cloning</i>	<i>Primers used for amplification</i>
		BDL187_ER_SmaI(SEQ ID NO:1027) TCCCGGGTTACTTTAGCCACAATCTGTGTTTC
BDL188	XbaI, SmaI	BDL188_NF_XbaI(SEQ ID NO:1028) AATCTAGAATTTTCATTGTTCGCTTCG BDL188_NR_SmaI(SEQ ID NO:1029) TCCCGGGCTAACAGTAGGTAATTTTGACATCCAG
BDL189		BDL189_NF_SmaI(SEQ ID NO:1030) ACCCGGGCATTGCCTGTTGGCTTCG BDL189_NR_SmaI(SEQ ID NO:1031) ACCCGGGTTACAATACAATTGTTTAATTCGAGG
BDL190	SalI, XbaI	BDL190_NF_SalI(SEQ ID NO:1032) TTAGTCGACAAAATAATGGCAGCTTTGGC BDL190_EF_SalI(SEQ ID NO:1033) TTAGTCGACTCTCGTCACATATCTTCATCGAC BDL190_NR_XbaI(SEQ ID NO:1034) TATCTAGACTACTAGACAAATTTGTTGATCAATTC BDL190_ER_XbaI(SEQ ID NO:1035) TATCTAGACTAAAGAGAGAACTAGACAAATTTGTTG
BDL192	SalI, XbaI	BDL192_NF_SalI(SEQ ID NO:1036) ATTGTGCACTTACGAAATACGCCGAATC BDL192_EF_SalI(SEQ ID NO:1037) AATGTGCACTTCGAAACCCTAACAAAAGC BDL192_NR_XbaI(SEQ ID NO:1038) ACTCTAGAATCTGCATAGCAGTTAGAACAAG BDL192_ER_XbaI(SEQ ID NO:1039) ATTCTAGAGAAAGGTCCTCATTTCATAATCC
BDL193	XbaI, SacI	BDL193_EF_XbaI(SEQ ID NO:1040) AATCTAGACTTCATATTCAAATCTCCTCTCC BDL193_ER_SacI (SEQ ID NO:1041) CGAGCTCATCACAAACAAACCTAAGAGGC
BDL194	EcoRV, EcoRV	BDL194_NF_EcoRV(SEQ ID NO:1042) AAGATATCAGCCATTGTTCTTCATCATCTC BDL194_NR_EcoRV(SEQ ID NO:1043) ACGATATCCTAACAGGGTTTTTCAGTGCTGTG
BDL196	SalI, XbaI	BDL196_NF_salI(SEQ ID NO:1044) TTAGTCGACAAGACATGAAGTTCATGACACTAATG BDL196_EF_SalI(SEQ ID NO:1045) TTAGTCGACAACCTGAAACAAAAGAAGAGTCATC BDL196_NR_XbaI(SEQ ID NO:1046) TATCTAGATTATGAGCTTTAACAACCTAGTATAAGGAAC BDL196_ER_XbaI(SEQ ID NO:1047) TATCTAGACACCACAATTTAAGCTTCAAC
BDL197	SalI, XbaI	BDL197_NF_SalI(SEQ ID NO:1048) AAAGTCGACTGTTCTTGTCTTCACGATGAG BDL197_EF_SalI(SEQ ID NO:1049) AAAGTCGACTCTAAATCCTATGTTCTTGTCTTC BDL197_NR_XbaI(SEQ ID NO:1050) AATCTAGAGGTTCAAAATACGTAACACATTG BDL197_ER_XbaI(SEQ ID NO:1051) AACTCTAGAACCATATTAGGTTCAAAATACGTAAC
BDL201	SalI, XbaI	BDL201_NF_SalI(SEQ ID NO:1052) AAAGTCGACCGATCAACAGACTCTAATCAGC

<i>Gene Name</i>	<i>Restriction Enzymes used for cloning</i>	<i>Primers used for amplification</i>
		BDL201_NF_XbaI(SEQ ID NO:1053) ATTCTAGATTAGTTCTATACTGCAGATTCTTGGG
BDL203	SalI, XbaI	BDL203_NF_SalI(SEQ ID NO:1054) AATGTCGACTTCTCTCTGTTCTTIGCACTCG
		BDL203_EF_SalI(SEQ ID NO:1055) AAAGTCGACCTTCTTCTTTTCTCAATCTTTC
		BDL203_NR_XbaI(SEQ ID NO:1056) ATTCTAGATCTGTATCATTAAAACTGAGGAAG
		BDL203_ER_XbaI(SEQ ID NO:1057) ATTCTAGAGTGGCGAGACAACATTTCTAC
		BDL220_NF_SalI(SEQ ID NO:1058) AAAGTCGACCTCTCTCTCTAATGGGTAATTG
BDL220	SalI, XbaI	BDL220_EF_SalI(SEQ ID NO:1059) AATGTCGACTCACCACACAACAACCAAG
		BDL220_NR_XbaI(SEQ ID NO:1060) ACTCTAGAAATCCAACGTCAAATGAGAAG
		BDL220_NF_SalI(SEQ ID NO:1061) AAAGTCGACCTCTCTCTCTAATGGGTAATTG
		BDL221_NF_SalI(SEQ ID NO:1062) AATGTCGACTTGGATCAGAGAAAATATGTCG
BDL221	SalI, XbaI	BDL221_EF_SalI(SEQ ID NO:1063) ATAGTCGACCTTGAATCTGAAGCTAATCTTGG
		BDL221_NR_XbaI(SEQ ID NO:1064) ATTCTAGATTAGCATTAGAACGGGACAGTATAAG
		BDL221_ER_XbaI(SEQ ID NO:1065) ACTCTAGATCAAAGAATCGAGCATTAGAACGG
		BDL222_NF_SalI(SEQ ID NO:1066) AAAGTCGACCTAAACCGTAAAAGATGTCG
BDL222	SalI, XbaI	BDL222_EF_SalI(SEQ ID NO:1067) AACGTCGACACTTTTGTGTTTGCCTTTTCCTC
		BDL222_NR_XbaI(SEQ ID NO:1068) ATTCTAGATCTTCTTCATCACTCAATCGC
		BDL222_ER_XbaI(SEQ ID NO:1069) ATTCTAGACTGTGGTATTTAGGGAATACATCC
		BDL223_NF_SalI(SEQ ID NO:1070) AAAGTCGACCACAGAGAAATCATGGGGTTC
		BDL223_EF_SalI(SEQ ID NO:1071) AACGTCGACAGATATCGTTGGCTTCGTCTC
BDL223	SalI, XbaI	BDL223_NR_XbaI(SEQ ID NO:1072) ATTCTAGATTAGGTTTGATCATTTTAACCAGAG
		BDL223_ER_XbaI(SEQ ID NO:1073) ATTCTAGACTATGCAGAAATGTTTGGATTGAG
		BDL224_NF_XbaI(SEQ ID NO:1074) AATCTAGACCACAAAATTCGTCAAAGCTC
		BDL224_EF_XbaI(SEQ ID NO:1075) ATTCTAGATTTTCAAACCACAAAATTCGTC
BDL224	XbaI, SmaI	BDL224_NR_SmaI(SEQ ID NO:1076) CCCCGGGATCCAACCAATCCCTAAAATG
		BDL224_ER_SmaI(SEQ ID NO:1077) CCCCGGGATCAGCCACTTCTACTCTCAATTC
		BDL225_NF_SalI(SEQ ID NO:1078) AATGTCGACTCCTTGTGATTCATTATTTGCG
BDL225	SalI, XbaI	

<i>Gene Name</i>	<i>Restriction Enzymes used for cloning</i>	<i>Primers used for amplification</i>
		BDL225_EF_SalI(SEQ ID NO:1079) AAAGTCGACCAACATCTCCTCCAAAACATTC
		BDL225_NR_XbaI(SEQ ID NO:1080) ACTCTAGATTAGCAAGAAGAAAAGAAGTGCAG
		BDL225_ER_XbaI(SEQ ID NO:1081) ATTCTAGATTAGTAGTTTATACAAGGTGCGGAGAC
BDL226	EcoRV, EcoRV	BDL226_NF_EcoRV(SEQ ID NO:1082) AAGATATCGAAACTGGATCTGGGTTTATCC
		BDL226_NR_EcoRV(SEQ ID NO:1083) ATGATATCCTAAACTAATCAAACATGGCACATAC
BDL227		BDL227_NF_SalI(SEQ ID NO:1084) AAAGTCGACAGAGTTAAGTCAATCACAAACC
		BDL227_NR_SmaI(SEQ ID NO:1085) TCCCGGGTTACCATCAAGTTTTCTTGCTGAAG
BDL228	SalI, XbaI	BDL228_NF_SalI(SEQ ID NO:1086) AAAGTCGACCCAACACTATATCATGGCTACTATC
		BDL228_NR_XbaI(SEQ ID NO:1087) ATTCTAGACTAACCTCACTTGATGCTCTTGC
BDL229	SalI, XbaI	BDL229_NR_XbaI(SEQ ID NO:1088) TATCTAGAGCTAAACAAAATCCGGAGATAG
		BDL229_ER_XbaI(SEQ ID NO:1089) TATCTAGACAGTCACTCCATAACTATGATCAAAC
		BDL229_F_SalI(SEQ ID NO:1090) TTAGTCGACCTCATTAATGGAAGTTTCAACATC
		BDL229_F_SalI(SEQ ID NO:1091) TTAGTCGACCTCATTAATGGAAGTTTCAACATC
BDL230	SmaI, SmaI	BDL230_NF_SmaI(SEQ ID NO:1092) TCCCGGGTAAGTTTGTGAGATGGAATTAGTG
		BDL230_NR_SmaI(SEQ ID NO:1093) TCCCGGGCTAATTGGTTGGTTACAAGATGC
BDL231	SalI, XbaI	BDL231_NF_SalI(SEQ ID NO:1094) AATGTCGACTGGACTGAAGATGTCAGGATTC
		BDL231_EF_SalI(SEQ ID NO:1095) ATAGTCGACATTTCTCTCTATTGGCATCGAC
		BDL231_NR_XbaI(SEQ ID NO:1096) AATTCTAGACTAAACTGGGGAAAGCTAAAACG
		BDL231_ER_XbaI(SEQ ID NO:1097) AATTCTAGATCATAACATAAGAAAGTAACTGGGG
BDL232	XbaI, SacI	BDL232_NF_XbaI(SEQ ID NO:1098) AATCTAGACACCCCCTCAAAGAAATATAAC
		BDL232_EF_XbaI(SEQ ID NO:1099) AATCTAGAAAGAAATTCACACCCCCTCAAAG
		BDL232_NR_SacI(SEQ ID NO:1100) TGAGCTCCTAAAGGTGGAGTAATTAGAAGCG
		BDL232_ER_SacI(SEQ ID NO:1101) TGAGCTCTGGTGAAGTGTTAAGTAATTGTCG
BDL233	SalI, SacI	BDL233_NF_SalI(SEQ ID NO:1102) AAAGTCGACGAAAGAGAGAAAATGGAGAATATG
		BDL233_EF_SalI(SEQ ID NO:1103) AAAGTCGACCGATCTAAAGAAAGAGAGAAAATG
		BDL233_NR_SacI(SEQ ID NO:1104) TGAGCTCCTAAGAGTCGATCTAGAAAAGCAACATC

<i>Gene Name</i>	<i>Restriction Enzymes used for cloning</i>	<i>Primers used for amplification</i>
		BDL233_ER_SacI(SEQ ID NO:1105) TGAGCTCGAATTAGTCCTTGTGGTTCTACTC
BDL234	SalI, SmaI	BDL234_NF_SalI(SEQ ID NO:1106) AATGTCGACTGATAAGAATGCTCCTGACTGG
		BDL234_EF_SalI(SEQ ID NO:1107) AATGTCGACTCTTTCTCTGTATCTCGACGTTC
		BDL234_NR_SmaI(SEQ ID NO:1108) TCCCGGGCTAAAATCCAAGTGCCCAAGAAC
		BDL234_ER_SmaI(SEQ ID NO:1109) TCCCGGGCTAGCAAAACATAAAATCCAAGTGC
		BDL235_NF_SalI(SEQ ID NO:1110) AATGTCGACTCTTACTCAATCCGAAGAATGG
BDL235	SalI, SmaI	BDL235_NR_SmaI(SEQ ID NO:1111) CCCCGGGACTTCGATTACATTTCTCCTC
		BDL237_NF_SalI(SEQ ID NO:1112) AAAGTCGACCGAAGTAAGAAAAGAAAATGGAG
BDL237	SalI, XbaI	BDL237_EF_SalI(SEQ ID NO:1113) AAAGTCGACCTTCCAAGTAAGAAAAGAAAATG
		BDL237_NR_XbaI(SEQ ID NO:1114) AATCTAGATCATACTCAAGTGCTTGTCCCTCGG
		BDL237_ER_XbaI(SEQ ID NO:1115) ATTCTAGAGTTATGGTGTCTTGTTCACC
		BDL238_NF_SalI(SEQ ID NO:1116) AAAGTCGACCAACGAGCAAGAGAAAATGG
BDL238	SalI, XbaI	BDL238_EF_SalI(SEQ ID NO:1117) AAAGTCGACGATACAACGAGCAAGAGAAAATG
		BDL238_NR_XbaI(SEQ ID NO:1118) AATTCTAGATGGTTCTAGCTATCACTAGGTGC
		BDL238_ER_XbaI(SEQ ID NO:1119) AATTCTAGAGGCAATACAACAAGAGAAAATCTC
		BDL240_NF_SalI(SEQ ID NO:1120) AAAGTCGACCGAGTACAATGGAGGTTTCCG
BDL240	SalI, XbaI	BDL240_NR_XbaI(SEQ ID NO:1121) ATTCTAGATCAAGCTTAAAGACCGTGAGGAAG
		BDL241_NF_XbaI(SEQ ID NO:1122) AATCTAGAACAGTCGTCGTCGTCGAAGC
BDL241	XbaI, SmaI	BDL241_NR_SmaI(SEQ ID NO:1123) TCCCGGGCTAAAGGTAAGGATGAATTGTCAGAG
		BDL242_NF_SalI(SEQ ID NO:1124) AATGTCGACTCAAATCAATATGGGATCTTTC
BDL242	SalI, XbaI	BDL242_NR_XbaI(SEQ ID NO:1125) ATTCTAGATTATTGACAAGTCTATTGCCCG
		BDL245_NF_SalI(SEQ ID NO:1126) AATGTCGACTTCGTTAAATTATGTCTTTGAGG
BDL245	SalI, SmaI	BDL245_EF_SalI(SEQ ID NO:1127) AAAGTCGACTGACTCAGAGATCAACAAAACC
		BDL245_NR_SmaI(SEQ ID NO:1128) ACCCGGGTTAGACTTACTCCAATTTCCAAGC
		BDL245_ER_SmaI(SEQ ID NO:1129) TCCCGGGTCAAGAGTCGGTCACACGC
		BDL247_NF_SmaI(SEQ ID NO:1130) TCCCGGGTCCTTCTTGTGTGAGACCGAG
BDL247	SmaI, SacI	

<i>Gene Name</i>	<i>Restriction Enzymes used for cloning</i>	<i>Primers used for amplification</i>
		BDL247_NF_SacI(SEQ ID NO:1131) TGAGCTCCTAAGAACTTTAACGCATTTTGTAGTG
BDL248	SalI, XbaI	BDL248_NF_SalI(SEQ ID NO:1132) AAAGTCGACAACGTGATCAATATGGAAGCTC
		BDL248_EF_SalI(SEQ ID NO:1133) AAAGTCGACACTCACCAAATCCAACGTG
		BDL248_NR_XbaI(SEQ ID NO:1134) AATTCTAGACTAAACTCAAGAGGAGTCGGGTAAG
		BDL248_ER_XbaI(SEQ ID NO:1135) AATTCTAGATTAATTTCGTTACCGTTGCTAAG
		BDL249_NF_SalI(SEQ ID NO:1136) AAAGTCGACACCAAATAGATCTAAAACATGG
BDL249	SalI, XbaI	BDL249_EF_SalI(SEQ ID NO:1137) AATGTCGACTTCACTCACCAAATAGATCTAAAAC
		BDL249_NR_XbaI(SEQ ID NO:1138) AATCTAGATCAACGTCAAACGGACTCGTTG
		BDL249_ER_XbaI(SEQ ID NO:1139) AATCTAGATCACCAAAGTCTAAACGTCAAACG
		BDL250_NF_SalI(SEQ ID NO:1140) TTAGTCGACCAAAGATGTTTACTATGTGATTGTC
BDL250	SalI, XbaI	BDL250_EF_SalI(SEQ ID NO:1141) TTAGTCGACAGGAAGAGAAAGGTCAAAGATG
		BDL250_NR_XbaI(SEQ ID NO:1142) TATCTAGATCAAATCTCACATCTCCATGCATAG
		BDL250_ER_XbaI(SEQ ID NO:1143) TATCTAGATCATGTTTCGCATTACACAAATATCC
		BDL252_NF_XbaI(SEQ ID NO:1144) ATTCTAGATTCTCTGTCTCTTTGGCTTTTC
		BDL252_EF_XbaI(SEQ ID NO:1145) ATTCTAGATAAACTCTCAGCTTCCCATTC
BDL252	XbaI, SmaI	BDL252_NR_SmaI(SEQ ID NO:1146) TCCCGGGCTATTGTCATTGAGGAAGAACAGG
		BDL252_ER_SmaI(SEQ ID NO:1147) TCCCGGGCTAAAAGTTCCTTGCTTGCTTCTG
		BDL48_NF_SalI(SEQ ID NO:1148) AAAGTCGACAGATTGCGTCACTGTAGTAGTAGTAG
		BDL48_EF_SalI(SEQ ID NO:1149) AAAGTCGACCTGCAACTCTTTCTCACTTTCAC
BDL48	SalI, XbaI	BDL48_NR_XbaI(SEQ ID NO:1150) AGTCTAGAAAACATTTTGCTTAAGATCTACAGAG
		BDL48_ER_XbaI(SEQ ID NO:1151) ACTCTAGAAGACATGAAAGCACAAATCAAG
		BDL49_NF_SmaI(SEQ ID NO:1152) ACCCGGGCTAACATGCTCCATCTCCTTC
		BDL49_NR_SacI(SEQ ID NO:1153) TGAGCTCTCAACCTGATCAGCGATGGTCG
BDL58	SalI, XbaI	BDL58_NF_SalI(SEQ ID NO:1154) AAAGTCGACCACACAAGACTAACGATGTTGC
		BDL58_NR_XbaI(SEQ ID NO:1155) AATTCTAGATTA AAAAGAGACCTACACGGCG
BDL62	SalI, SacI	BDL62_NF_SalI(SEQ ID NO:1156) AATGTCGACTCTGGTCTCCCTATATCAGC

<i>Gene Name</i>	<i>Restriction Enzymes used for cloning</i>	<i>Primers used for amplification</i>
		BDL62_NF_SacI(SEQ ID NO:1157) TGAGCTCCTATTTGATGTTGTTGTTGTTGTTGCTG
BDL63	SalI, XbaI	BDL63_NF_SalI(SEQ ID NO:1158) ATAGTCGACGTTTTGAGATATGGGAGGACC
		BDL63_EF_SalI(SEQ ID NO:1159) AAAGTCGACCTCTAGATTCTTGGCGATTCTC
		BDL63_NR_XbaI(SEQ ID NO:1160) ATTCTAGATTAGTGGTTTTACTTGAGCCTCTCC
		BDL63_ER_SacI(SEQ ID NO:1161) TGAGCTCCTATTGTTCTGTTACGGTGGTTTTAC
		BDL64_NF_SalI(SEQ ID NO:1162) AATGTCGACGGACTTTAAACATGGGTGTTTC
BDL64		BDL64_NR_XbaI(SEQ ID NO:1163) ATTCTAGACTACTCATAGGTTTGTACTTCCTTG
		BDL75_NF_SalI(SEQ ID NO:1164) AAAGTCGACAAGAAGAAAGAAACAGAGAATCG
BDL75	SalI, XbaI	BDL75_NR_XbaI(SEQ ID NO:1165) ATTCTAGACTAATTGTTCAAAGTTCAGTGAGCC
		BDL79_NF_XbaI(SEQ ID NO:1166) ATTCTAGAGGAGATTTTGTAAATGGATTCTGC
BDL79	XbaI, SmaI	BDL79_EF_XbaI(SEQ ID NO:1167) ATTCTAGAGAAGGAGATTTTGTAAATGGATTCTC
		BDL79_NR_SmaI(SEQ ID NO:1168) TCCCGGGTTACGCTTAGACCATAACAACGAGTAG
		BDL79_ER_SmaI(SEQ ID NO:1169) TCCCGGGTTATTTACTTATGGCCTGTTTC
		BDL81_EF_SalI(SEQ ID NO:1170) AAAGTCGACCAGGGGTTTAAGGATTTTCTC
		BDL81_ER_SacI(SEQ ID NO:1171) CGAGCTCAAATGGCTTTCTCTACCCCTTG
BDL83	SalI, XbaI	BDL83_NF_SalI(SEQ ID NO:1172) AATGTCGACTGGTAGGCTGAGAGAAAGAAAG
		BDL83_NR_XbaI(SEQ ID NO:1173) AGTCTAGATTAGAGAATAAAAGAAGAATGAGAAGC
BDL85	XbaI, SalI	BDL85_NF_SalI(SEQ ID NO:1174) AATGTCGACTTAATCGTTAGAAGATGAGCCAG
		BDL85_NR_XbaI(SEQ ID NO:1175) ATTCTAGATCAGGCTTAGAAGCAAATGTCCAG
BDL88	SalI, XbaI	BDL88_NF_SalI(SEQ ID NO:1176) AATGTCGACGAGGAGATGGCGAGCAAC
		BDL88_NR_XbaI(SEQ ID NO:1177) TATCTAGATTATTAGGTATTGCACTTCCACTTC
BDL90		BDL90_EF_SalI(SEQ ID NO:1178) AAAGTCGACCACCAGAAACAAAGAGAGAGTG
		BDL90_ER_XbaI(SEQ ID NO:1179) ATTCTAGATATCATGCAACCACAAACAATAG
BDL94	XbaI, SmaI	BDL94_EF_XbaI_new(SEQ ID NO:1180) AATCTAGAAAGTCCAAGTGACCAACCATC
		BDL94_ER_SmaI_new(SEQ ID NO:1181) TCCCGGGGGGATACAAGATTATGCAGGC

Table 18. Provided are the PCR primers used for cloning the genes of some embodiments of the invention. Fwd = forward primer; Rev = reverse primer; Nested = nested primer for PCR (internal primer); External = external primer for PCR.

To facilitate cloning of the cDNAs/ genomic sequences, a 8-12 bp extension was added to the 5' of each primer. The primer extension includes an endonuclease restriction site. The restriction sites were selected using two parameters: (a). The site did not exist in the cDNA sequence; and (b). The restriction sites in the forward and reverse primers were designed such that the digested cDNA is inserted in the sense formation into the binary vector utilized for transformation.

Each digested PCR product was inserted into a high copy vector pBlue-script KS plasmid vector [pBlue-script KS plasmid vector] or into plasmids originating from this vector. In cases where the pGXN high copy vector (originated from pBlue-script KS) was used, the PCR product was inserted upstream to the NOS terminator (SEQ ID NO:1182) originated from pBI 101.3 binary vector (GenBank Accession No. U12640, nucleotides 4356 to 4693) and downstream to the 35S promoter (Table 20 below). The digested products and the linearized plasmid vector were ligated using T4 DNA ligase enzyme (Roche, Switzerland).

Sequencing of the amplified PCR products was performed, using ABI 377 sequencer (Amersham Biosciences Inc). In all cases, after confirmation of the sequence of the cloned genes, the cloned cDNA accompanied or not with the NOS terminator was introduced into the pGI binary vector [pBXYN or pQXYN containing the 35S CaMV promoter] according to Table 19 hereinabove, via digestion with appropriate restriction endonucleases. In any case the insert was followed by single copy of the NOS terminator (SEQ ID NO:1182x).

High copy plasmids containing the cloned genes were digested with the restriction endonucleases (New England BioLabs Inc) according to the sites designed in the primers (Table 18, above) and cloned into binary vectors according to Table 19, below.

Binary vectors used for cloning: The plasmid pPI was constructed by inserting a synthetic poly-(A) signal sequence, originating from pGL3 basic plasmid vector (Promega, Acc No U47295; bp 4658-4811) into the *Hind*III restriction site of the binary vector pBI101.3 (Clontech, Acc. No. U12640). pGI (pBXYN) (Figure 1) is similar to pPI, but the original gene in the backbone, the GUS gene, was replaced by the

GUS-Intron gene followed by the NOS terminator (SEQ ID NO:1182) (Vancanneyt, G, *et al* MGG 220, 245-50, 1990). pGI was used to clone the polynucleotide sequences, initially under the control of 35S promoter [Odell, JT, *et al.* Nature 313, 810 - 812 (28 February 1985); SEQ ID NO:1184.

- 5 The modified pGI vector (pQXYN) is a modified version of the pGI vector in which the cassette is inverted between the left and right borders so the gene and its corresponding promoter are close to the right border and the NPTII gene is close to the left border.

10

Table 19
Restriction enzyme sites used to clone the identified genes into binary vector

<i>Gene name</i>	<i>Binary vector</i>	<i>Restriction enzymes used for cloning into binary vector- FORWARD</i>	<i>Restriction enzymes used for cloning into binary vector- REVERSE</i>	<i>Restriction enzymes used for digesting the binary vector</i>
BDL62	pQXYN	Sall	SacI	Sall, SacI
BDL75	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL79	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL81	pQXYN	Sall	SacI	Sall, SacI
BDL83	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL117	pQXYN	SmaI	SmaI	SmaI, Ecl136II
BDL118	pQXYN	SmaI	SmaI	SmaI, Ecl136II
BDL138	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL140	pQXYN	XbaI	SacI	XbaI, SacI
BDL147	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL149	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL152	pQXYN	XbaI	SacI	XbaI, SacI
BDL153	pQXYN	Sall	SmaI	Sall, Ecl136II
BDL154	pQXYN	Sall	SacI	Sall, SacI
BDL155	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL156	pQXYN	Sall	SacI	SacI, Sall
BDL157	pQXYN	Sall	SacI	Sall, SacI
BDL158	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL160	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL162	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL167	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL168	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL169	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL171	pQXYN	XbaI	SacI	XbaI, SacI
BDL173	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL174	pQXYN	Sall	SacI	Sall, SacI
BDL176	pQXYN	SmaI	SmaI	SmaI, Ecl136II
BDL177	pQXYN	Sall	SacI	Sall, SacI
BDL181	pQXYN	Sall	SacI	Sall, SacI
BDL182	pQXYN	Sall	SacI	Sall, SacI
BDL183	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL186	pQXYN	Sall	SacI	Sall, SacI
BDL187	pQXYN	Sall	SmaI	Sall, Ecl136II

<i>Gene name</i>	<i>Binary vector</i>	<i>Restriction enzymes used for cloning into binary vector- FORWARD</i>	<i>Restriction enzymes used for cloning into binary vector- REVERSE</i>	<i>Restriction enzymes used for digesting the binary vector</i>
BDL188	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL189	pQXYN	SmaI	BamHI	SmaI, Ecl136II
BDL190	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL192	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL193	pQXYN	XbaI	SacI	XbaI, SacI
BDL194	pQXYN	EcoRV	EcoRV	SmaI, Ecl136II
BDL196	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL197	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL201	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL203	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL220	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL221	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL222	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL223	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL229	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL230	pQXYN	SmaI	SmaI	SmaI, Ecl136II
BDL231	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL235	pQXYN	Sall	SmaI	Sall, Ecl136II
BDL242	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL245	pQXYN	Sall	SmaI	Sall, Ecl136II
BDL247	pQXYN	SmaI	SacI	SmaI, SacI
BDL248	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL250	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL49	pQXYN	EcoRV	SacI	SmaI, SacI
BDL58	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL63	pQXYN	Sall	SacI	Sall, SacI
BDL64	pQXYN	Sall	XbaI	Sall, Ecl136II
BDL85	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL88	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL90	pQXYN	XbaI	Sall	Sall, Ecl136II
BDL94	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL102	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL224	pQXYN	XbaI	EcoRI	XbaI, EcoRI
BDL225	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL226	pQXYN	EcoRV	EcoRV	SmaI, Ecl136II
BDL227	pQXYN	Sall	SmaI	Sall, Ecl136II
BDL228	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL232	pQXYN	XbaI	EcoRI	XbaI, EcoRI
BDL233	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL234	pQXYN	Sall	SmaI	Sall, Ecl136II
BDL237	pQXYN	Sall	SacI	Sall, SacI
BDL238	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL240	pQXYN	Sall	SacI	Sall, SacI
BDL241	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL249	pQXYN	Sall	EcoRI	Sall, EcoRI
BDL252	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL47	pQXYN	XbaI	SmaI	XbaI, Ecl136II
BDL48	pQXYN	Sall	EcoRI	Sall, EcoRI

Table 19.

Table 20
Genes cloned from cDNA libraries or genomic DNA in a High copy number plasmid

<i>Gene Name</i>	<i>High copy plasmid</i>	<i>Amplified from</i>		<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
		<i>Organism</i>	<i>Origin</i>		
BDL62	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	847	108
BDL75	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	848	109
BDL79	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	849	110
BDL81	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	850	111
BDL83	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	851	112
BDL117	Topo B	Arabidopsis thaliana ND	RNA	852	926
BDL118	Topo B	Arabidopsis thaliana ND	RNA	853	114
BDL138	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	854	116
BDL140	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	855	117
BDL147	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	856	118
BDL149	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	857	119
BDL152	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	858	120
BDL153	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	859	121
BDL154	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	860	122
BDL155	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	861	123
BDL156	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	862	124
BDL157	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	863	125
BDL158	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	864	126
BDL160	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	865	127
BDI.162	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	866	128
BDL167	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	867	196
BDL168	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	868	132
BDL169	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	869	133
BDL171	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	870	927
BDL173	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	871	135

<i>Gene Name</i>	<i>High copy plasmid</i>	<i>Amplified from</i>		<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
		<i>Organism</i>	<i>Origin</i>		
BDL174	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	872	136
BDL176	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	873	137
BDL177	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	874	138
BDL181	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	875	139
BDL182	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	876	140
BDL183	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	877	141
BDL186	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	878	928
BDL187	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	879	929
BDL188	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	880	144
BDL189	Topo B	Arabidopsis thaliana ND	RNA	881	145
BDL190	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	882	146
BDL192	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	883	147
BDL193	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	884	148
BDL194	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	885	149
BDL196	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	886	150
BDL197	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	887	151
BDL201	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	888	153
BDL203	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	889	154
BDL220	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	890	156
BDL221	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	891	157
BDL222	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	892	158
BDL223	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	893	159
BDL229	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	894	160
BDL230	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	895	161
BDL231	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	896	162
BDL235	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	897	163

<i>Gene Name</i>	<i>High copy plasmid</i>	<i>Amplified from</i>		<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
		<i>Organism</i>	<i>Origin</i>		
BDL242	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	898	164
BDL245	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	899	166
BDL247	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	900	167
BDI.248	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	901	168
BDL250	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	902	169
BDL49	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	903	170
BDL58	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	904	171
BDL63	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	905	172
BDL64	Topo B	Arabidopsis thaliana ND	RNA	906	173
BDL85	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	907	175
BDL88	pGXN (pKG+Nos+35S)	RICE Oryza sativa L. Japonica ND	RNA	908	176
BDL90	Topo B	RICE Oryza sativa L.Japonica ND	gDNA	909	Non Coding polynucleotide
BDL94	pGXN (pKG+Nos+35S)	Rice Japonica ND leaves	gDNA	910	930
BDL102	pGXN (pKG+Nos+35S)	MAIZE Zea mays L. ND	RNA	911	178
BDL224	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	912	179
BDL225	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	913	180
BDI.226	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	914	181
BDL227	Topo B	Arabidopsis thaliana ND	RNA	915	182
BDL228	pGXN (pKG+Nos+35S)	CASTOR BEAN Ricinus communis L. ND	RNA	916	931
BDL232	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	917	184
BDL233	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	918	932
BDL234	pKS(Pks_J)	Arabidopsis thaliana ND	RNA	919	186
BDL237	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	920	187
BDL238	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	921	188
BDI.240	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	922	189

<i>Gene Name</i>	<i>High copy plasmid</i>	<i>Amplified from</i>		<i>Polynucleotide SEQ ID NO:</i>	<i>Polypeptide SEQ ID NO:</i>
		<i>Organism</i>	<i>Origin</i>		
BDL241	pKS(Pks_I)	Arabidopsis thaliana ND	RNA	923	190
BDL249	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	924	191
BDL252	pKS(Pks_I)	Arabidopsis thaliana ND	RNA	925	193
BDL47	High copy plasmid pMA	Synthetic DNA		845	106
BDL48	pGXN (pKG+Nos+35S)	Arabidopsis thaliana ND	RNA	846	107
BDL200	High copy plasmid	Synthetic DNA		47	152

Table 20: Cloned and synthetic genes are provided along with the sequence identifiers of their polynucleotides and polypeptides. Also provided are the source organism, tissue and the cloning vectors. ND = not a determined ecotype.

5 Selected DNA sequences were synthesized by a commercial supplier GeneArt, GmbH. Synthetic DNA is designed in silico. Suitable restriction enzymes sites were added to the cloned sequences at the 5' end and at the 3' end to enable later cloning into the pBXYN/ pQXYN binary downstream of the CaMV 35S promoter (SEQ ID NO:1184). For example BDL47 (SEQ ID NO:1 was synthesized and the XbaI and SmaI
10 restriction enzymes were added to the synthetic sequence in order to facilitate cloning.

For 7 genes, namely BDL117, BDL171, BDL186, BDL187, BDL94, BDL228 and BDL233, the protein translation of the amplified cDNA sequence did not match the initial bioinformatics prediction of the protein sequences. The polypeptide sequences encoded by the cloned sequence were predicted and are provided in SEQ ID NO: 926-
15 932.

EXAMPLE 8

PRODUCING TRANSGENIC ARABIDOPSIS PLANTS EXPRESSING THE IDENTIFIED POLYNUCLEOTIDES OF THE INVENTION

20 **Materials and Experimental Methods**

Plant transformation - The *Arabidopsis thaliana* var Columbia (To plants) were transformed according to the Floral Dip procedure [Clough SJ, Bent AF. (1998) Floral dip: a simplified method for Agrobacterium-mediated transformation of *Arabidopsis thaliana*. Plant J. 16(6): 735-43; and Desfeux C, Clough SJ, Bent AF.

(2000) Female reproductive tissues are the primary targets of *Agrobacterium*-mediated transformation by the *Arabidopsis* floral-dip method. *Plant Physiol.* 123(3): 895-904] with minor modifications. Briefly, *Arabidopsis thaliana* Columbia (Col0) T₀ plants were sown in 250 ml pots filled with wet peat-based growth mix. The pots were covered with aluminum foil and a plastic dome, kept at 4 °C for 3–4 days, then uncovered and incubated in a growth chamber at 18–24 °C under 16/8 hours light/dark cycles. The T₀ plants were ready for transformation six days before anthesis.

Single colonies of *Agrobacterium* carrying the binary vectors harboring the seed oil genes were cultured in LB medium supplemented with kanamycin (50 mg/L) and gentamycin (50 mg/L). The cultures were incubated at 28 °C for 48 hours under vigorous shaking and centrifuged at 4000 rpm for 5 minutes. The pellets comprising *Agrobacterium* cells were resuspended in a transformation medium which contained half-strength (2.15 g/L) Murashige-Skoog (Duchefa); 0.044 μM benzylamino purine (Sigma); 112 μg/L B5 Gambourg vitamins (Sigma); 5 % sucrose; and 0.2 ml/L Silwet L-77 (OSI Specialists, CT) in double-distilled water, at pH of 5.7.

Transformation of T₀ plants was performed by inverting each plant into an *Agrobacterium* suspension such that the above ground plant tissue was submerged for 3-5 seconds. Each inoculated T₀ plant was immediately placed in a plastic tray, then covered with clear plastic dome to maintain humidity and was kept in the dark at room temperature for 18 hours to facilitate infection and transformation. Transformed (transgenic) plants were then uncovered and transferred to a greenhouse for recovery and maturation. The transgenic T₀ plants were grown in the greenhouse for 3-5 weeks until siliques were brown and dry, then seeds were harvested from plants and kept at room temperature until sowing.

For generating T₁ and T₂ transgenic plants harboring the genes, seeds collected from transgenic T₀ plants were surface-sterilized by soaking in 70 % ethanol for 1 minute, followed by soaking in 5 % sodium hypochlorite and 0.05 % triton for 5 minutes. The surface-sterilized seeds were thoroughly washed in sterile distilled water then placed on culture plates containing half-strength Murashige-Skoog (Duchefa); 2 % sucrose; 0.8 % plant agar; 50 mM kanamycin; and 200 mM carbenicillin (Duchefa). The culture plates were incubated at 4 °C for 48 hours then transferred to a growth room at 25 °C for an additional week of incubation. Vital T₁ *Arabidopsis* plants were

114

transferred to a fresh culture plates for another week of incubation. Following incubation the T₁ plants were removed from culture plates and planted in growth mix contained in 250 ml pots. The transgenic plants were allowed to grow in a greenhouse to maturity. Seeds harvested from T₁ plants were cultured and grown to maturity as T₂ plants under the same conditions as used for culturing and growing the T₁ plants.

EXAMPLE 9

IMPROVED TRANSGENIC PLANT PERFORMANCE

To analyze the effect of expression of the isolated polynucleotides in plants, plants were grown in pots with an adequate amount of nutrients and water. The plants were analyzed for their overall size, growth rate, time to inflorescence emergence (bolting) and flowering, seed yield, weight of 1,000 seeds, dry matter and harvest index [(HI) seed yield/ dry matter]. Transgenic plants performance was compared to control plants grown in parallel under the same conditions. Mock- transgenic plants with an empty vector or expressing the uidA reporter gene (GUS-Intron) under the same promoter were used as control.

Parameters were measured as described in Examples 2, 3 and 4 above.

Statistical analyses - Plant growth rate, plant area, time to bolt, time to flower, weight of 1,000 seeds, seed yield, oil yield, dry matter, and harvest index area data were analyzed using t-test. To identify outperforming genes and constructs, results from mix of transformation events or independent events were analyzed. For gene versus control analysis t- test was applied, using significance of $p < 0.1$. The JMP statistics software package was used (Version 5.2.1, SAS Institute Inc., Cary, NC, USA).

Experimental Results

Plants expressing the polynucleotides of the invention were assayed for a number of commercially desired traits. Results are presented in Tables 21 and 22.

Analysis of plants in tissue culture assay - Tables 21 and 22, hereinbelow, depict analyses of seed yield in plants overexpressing the polynucleotides of the invention under the regulation of the constitutive 35S (SEQ ID NO:1184) or At6669 (SEQ ID NO:1183) promoters. In cases where a certain event appears more than once, the event was tested in several independent experiments.

Table 21
Results obtained in a tissue culture assay

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TP1</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TP1</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TP1</i>
BDL102	10471.1	P		0.24		0.01	0.02	0.13	0.05
BDL102	10471.1	Av		1.15		1.59	1.39	1.23	2.34
BDL102	10471.3	P		0.6		0.03	0.01	0.09	0.01
BDL102	10471.3	Av		1.1		1.43	1.37	1.24	1.72
BDL102	10472.1	P				0.04	0.15		0.03
BDL102	10472.1	Av				1.27	1.1		1.54
BDL102	10474.2	P	<0.01	<0.01	<0.01		0.23	0.13	
BDL102	10474.2	Av	2.02	1.61	1.51		1.1	1.13	
BDL102	10474.6	P		0.13	0.29				
BDL102	10474.6	Av		1.21	1.14				
BDL118	10481.2	P	<0.01	0.01	0.01			0.46	
BDL118	10481.2	Av	2.02	1.91	2			1.1	
BDL118	10481.5	P		0.2	0.01				
BDL118	10481.5	Av		1.15	1.29				
BDL118	10484.3	P	<0.01	<0.01	<0.01				
BDL118	10484.3	Av	1.51	1.41	1.58				
BDL140	10421.3	P	0.03	0.01	0.05				0.39
BDL140	10421.3	Av	1.61	1.72	1.68				1.18
BDL140	10423.1	P	0.21	0.09	0.28	0.01	0.11		0.19
BDL140	10423.1	Av	1.24	1.31	1.21	1.38	1.21		1.28
BDL140	10424.4	P	<0.01	<0.01	0.01				
BDL140	10424.4	Av	1.43	1.48	1.62				
BDL152	10431.4	P	0.15	0.12	0.12				
BDL152	10431.4	Av	1.17	1.18	1.18				
BDL152	10432.5	P	0.37	0.01		0.07	0.14		0.35
BDL152	10432.5	Av	1.16	1.28		1.22	1.13		1.25
BDL152	10434.1	P		0.43					
BDL152	10434.1	Av		1.12					
BDL152	10434.4	P		0.08		0.04	0.01	<0.01	0.08
BDL152	10434.4	Av		1.18		1.52	1.48	1.31	2.33
BDL153	10142.2	P				<0.01	<0.01		0.03
BDL153	10142.2	Av				1.83	1.43		1.99

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL153	10144.1	P	<0.01	0.02	0.07		0.31	0.15	0.7
BDL153	10144.1	Av	1.3	1.31	1.38		1.13	1.12	1.1
BDL153	10144.4	P							0.03
BDL153	10144.4	Av							1.28
BDL154	10703.1	P				0.09	0.14	0.05	0.09
BDL154	10703.1	Av				1.36	1.14	1.2	1.77
BDL154	10703.1 1	P				0.01	0.17	0.3	0.5
BDL154	10703.1 1	Av				1.37	1.21	1.16	1.18
BDL154	10703.6	P							0.42
BDL154	10703.6	Av							1.23
BDL154	10703.1	P	<0.01	<0.01	<0.01		0.11	0.02	
BDL154	10703.1	Av	1.34	1.5	1.61		1.14	1.18	
BDL154	10703.5	P		0.46	0.36		0.02	0.02	0.21
BDL154	10703.5	Av		1.1	1.16		1.47	1.39	1.52
BDL154	10703.6	P				0.24	0.06	0.07	0.15
BDL154	10703.6	Av				1.19	1.33	1.24	1.59
BDL155	9994.3	P	0.22						
BDL155	9994.3	Av	1.15						
BDL156	10853.6	P			0.08				
BDL156	10853.6	Av			1.12				
BDL156	10852.6	P			0.15				
BDL156	10852.6	Av			1.15				
BDL156	10853.6	P		0.07	0.01			0.24	
BDL156	10853.6	Av		1.34	1.71			1.22	
BDL156	10854.4	P	0.11	0.11	<0.01			0.01	
BDL156	10854.4	Av	1.13	1.26	1.66			1.24	
BDL156	10855.3	P			0.01			0.1	
BDL156	10855.3	Av			1.35			1.2	
BDL158	9973.3	P				0.01	0.03	0.17	0.09
BDL158	9973.3	Av				1.21	1.27	1.1	1.21
BDL158	9971.3	P	0.42	0.32	0.27				
BDL158	9971.3	Av	1.13	1.13	1.11				
BDL158	9973.1	P				0.08	0.24	0.17	0.02

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TP1</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TP1</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TP1</i>
BDL158	9973.1	Av				1.27	1.16	1.13	1.37
BDL158	9973.3	P				<0.01	0.02	0.06	0.01
BDL158	9973.3	Av				1.66	1.39	1.17	2.06
BDL158	9974.2	P	<0.01	<0.01	0.09				
BDL158	9974.2	Av	1.68	1.58	1.23				
BDL158	9974.3	P	<0.01	<0.01	0.13				
BDL158	9974.3	Av	1.76	1.62	1.58				
BDL158	9971.3	P		0.07	0.15			0.02	
BDL158	9971.3	Av		1.22	1.2			1.15	
BDL158	9973.1	P	0.01	0.41	0.03	<0.01	<0.01	0.08	0.02
BDL158	9973.1	Av	1.28	1.11	1.35	1.49	1.35	1.26	1.8
BDL158	9973.3	P				<0.01	<0.01	<0.01	0.02
BDL158	9973.3	Av				1.36	1.52	1.39	1.51
BDL160	10011.5	P	0.01	0.22	0.15				
BDL160	10011.5	Av	1.41	1.2	1.26				
BDL160	10011.5	P							0.45
BDL160	10011.5	Av							1.25
BDL160	10011.7	P				0.04			0.01
BDL160	10011.7	Av				1.34			1.95
BDL160	10013.1	P			0.53				
BDL160	10013.1	Av			1.11				
BDL160	10014.9	P	0.2	0.39	0.61				
BDL160	10014.9	Av	1.17	1.15	1.11				
BDL160	10015.2	P		0.07	0.12				
BDL160	10015.2	Av		1.19	1.22				
BDL167	10042.3	P	<0.01						
BDL167	10042.3	Av	1.3						
BDL167	10043.1	P	<0.01	0.02	0.14				
BDL167	10043.1	Av	1.37	1.18	1.2				
BDL167	10043.2	P	0.06	0.13	0.03				
BDL167	10043.2	Av	1.61	1.29	1.23				
BDL167	10043.3	P	0.01	0.01	<0.01			0.01	
BDL167	10043.3	Av	1.93	1.58	1.61			1.25	
BDL167	10044.2	P	<0.01	0.01	0.02	0.05	0.04	0.03	
BDL167	10044.2	Av	1.63	1.43	1.43	1.28	1.23	1.23	

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL167	10043.1	P	<0.01	<0.01	0.15		<0.01	0.12	0.47
BDL167	10043.1	Av	1.56	1.53	1.43		1.22	1.1	1.1
BDL167	10044.2	P				<0.01	<0.01		<0.01
BDL167	10044.2	Av				1.45	1.18		2.16
BDL168	9881.3	P				0.15			0.04
BDL168	9881.3	Av				1.23			1.67
BDL168	9881.4	P				<0.01	<0.01	<0.01	0.02
BDL168	9881.4	Av				1.89	1.72	1.38	2.28
BDL168	9882.1	P				0.01	0.01	0.04	<0.01
BDL168	9882.1	Av				1.9	1.68	1.29	2.57
BDL168	9883.3	P							0.2
BDL168	9883.3	Av							1.38
BDL168	9884.1	P				<0.01	<0.01	0.02	0.02
BDL168	9884.1	Av				2.05	1.73	1.38	2.61
BDL169	10744.2	P				0.08	0.02	0.08	0.29
BDL169	10744.2	Av				1.24	1.23	1.19	1.11
BDL169	10747.1	P				0.03	0.09	0.13	
BDL169	10747.1	Av				1.15	1.12	1.11	
BDL169	10747.5	P				0.06	0.01	<0.01	0.12
BDL169	10747.5	Av				1.62	1.59	1.4	1.73
BDL171	10661.2	P		0.25	0.28	<0.01	<0.01	<0.01	0.03
BDL171	10661.2	Av		1.19	1.23	1.62	1.65	1.39	1.8
BDL171	10661.5	P	0.02	0.04	0.12				
BDL171	10661.5	Av	1.45	1.35	1.24				
BDL171	10664.1	P	0.31	0.3	0.25	0.15	0.27	0.31	0.44
BDL171	10664.1	Av	1.17	1.18	1.23	1.23	1.15	1.14	1.17
BDL173	9951.2	P		0.18			0.42		0.3
BDL173	9951.2	Av		1.17			1.11		1.17
BDL173	9952.1	P				<0.01	<0.01	<0.01	<0.01
BDL173	9952.1	Av				2.17	1.99	1.45	3.32
BDL173	9952.2	P		0.35		<0.01	<0.01	0.01	0.02
BDL173	9952.2	Av		1.12		1.72	1.63	1.24	2.07
BDL174	11082.1	P					0.2	0.06	0.42
BDL174	11082.1	Av					1.14	1.17	1.18
BDL174	11083.1	P	<0.01	<0.01	0.07	<0.01	<0.01	0.01	0.01

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL174	11083.1	Av	1.64	1.32	1.2	1.78	1.67	1.39	1.66
BDL174	11083.2	P	0.04	0.17	0.12		0.14	0.18	
BDL174	11083.2	Av	1.2	1.16	1.25		1.34	1.32	
BDL174	11084.1	P	<0.01	<0.01	<0.01		0.03	0.1	0.03
BDL174	11084.1	Av	2.56	2.29	2.1		1.31	1.31	1.68
BDL174	11085.1	P		0.41	0.07	<0.01	<0.01	<0.01	<0.01
BDL174	11085.1	Av		1.11	1.24	1.95	2.33	1.98	1.71
BDL174	11083.2	P	<0.01	<0.01	0.01	0.3			0.31
BDL174	11083.2	Av	1.41	1.35	1.21	1.16			1.27
BDL174	11084.1	P				0.15			<0.01
BDL174	11084.1	Av				1.22			2.13
BDL174	11085.1	P				<0.01	0.01	0.04	0.01
BDL174	11085.1	Av				2.04	1.36	1.16	2.33
BDL176	9891.4	P	0.1	0.05	0.06				
BDL176	9891.4	Av	1.1	1.17	1.36				
BDL176	9893.2	P	0.04	<0.01	0.27				
BDL176	9893.2	Av	1.28	1.26	1.22				
BDL176	9893.3	P				0.38			
BDL176	9893.3	Av				1.13			
BDL176	9893.2	P				0.15			
BDL176	9893.2	Av				1.2			
BDL176	9893.3	P	0.05	0.01		0.36			0.16
BDL176	9893.3	Av	1.42	1.34		1.16			1.42
BDL177	10521.3	P						0.23	
BDL177	10521.3	Av						1.16	
BDL177	10524.2	P				0.13			
BDL177	10524.2	Av				1.16			
BDL181	11293.6	P			0.32				
BDL181	11293.6	Av			1.18				
BDL181	11294.7	P			0.36				
BDL181	11294.7	Av			1.13				
BDL181	11293.1	P			0.6				
BDL181	11293.1	Av			1.11				
BDL181	11293.6	P		0.01	<0.01				
BDL181	11293.6	Av		1.2	1.27				

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TP1</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TP1</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TP1</i>
BDL181	11294.7	P					0.16	0.17	
BDL181	11294.7	Av					1.1	1.1	
BDL182	10691.8	P	0.03	0.07	0.16				
BDL182	10691.8	Av	1.32	1.24	1.18				
BDL182	10692.2	P	0.17	0.08	0.19				
BDL182	10692.2	Av	1.19	1.27	1.2				
BDL182	10693.3	P	0.01	0.07	0.1	0.17	0.1	0.11	
BDL182	10693.3	Av	1.55	1.44	1.31	1.17	1.17	1.1	
BDL182	10693.5	P	0.04	0.24	0.12	<0.01	<0.01	<0.01	0.48
BDL182	10693.5	Av	1.33	1.15	1.22	1.39	1.33	1.21	1.1
BDL183	9943.4	P				0.02	0.11		0.02
BDL183	9943.4	Av				1.26	1.17		1.61
BDL183	9944.4	P				0.01	<0.01	0.02	0.36
BDL183	9944.4	Av				1.24	1.36	1.21	1.26
BDL189	11351.2	P	0.05						
BDL189	11351.2	Av	1.17						
BDL189	11353.3	P	0.22	0.04	0.11				
BDL189	11353.3	Av	1.19	1.2	1.17				
BDL189	11353.5	P	0.26						
BDL189	11353.5	Av	1.1						
BDL189	11355.4	P	0.12	0.12	0.1				
BDL189	11355.4	Av	1.12	1.11	1.11				
BDL189	11356.7	P	0.27						
BDL189	11356.7	Av	1.11						
BDL196	10242.2	P	0.02	0.09		0.05	0.03	0.09	
BDL196	10242.2	Av	1.18	1.13		1.29	1.28	1.16	
BDL196	10243.4	P	0.09	0.25	0.24	0.13	0.05	0.17	<0.01
BDL196	10243.4	Av	1.16	1.23	1.24	1.11	1.25	1.15	1.74
BDL196	10244.1	P	<0.01	0.07			0.23		0.21
BDL196	10244.1	Av	1.21	1.19			1.22		1.33
BDL196	10243.3	P				0.02			0.68
BDL196	10243.3	Av				1.16			1.1
BDL196	10243.4	P		0.01	<0.01			0.23	
BDL196	10243.4	Av		1.24	1.49			1.12	
BDL197	11362.2	P			0.14				

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL197	11362.2	Av			1.17				
BDL197	11363.1	P						0.04	
BDL197	11363.1	Av						1.17	
BDL197	11363.6	P			0.15				
BDL197	11363.6	Av			1.17				
BDL197	11364.1	P		0.08	0.01			0.09	
BDL197	11364.1	Av		1.14	1.42			1.15	
BDL197	11364.5	P		0.09	0.04				
BDL197	11364.5	Av		1.12	1.25				
BDL197	11363.6	P		0.2	0.07				
BDL197	11363.6	Av		1.11	1.19				
BDL201	9961.2	P	0.02	0.18	0.39				
BDL201	9961.2	Av	1.41	1.18	1.18				
BDL201	9961.3	P	0.08	0.13					
BDL201	9961.3	Av	1.21	1.28					
BDL201	9961.4	P				0.06	0.15	0.43	0.02
BDL201	9961.4	Av				1.37	1.25	1.11	1.76
BDL201	9964.3	P				<0.01	0.01		0.02
BDL201	9964.3	Av				1.34	1.24		1.84
BDL220	10331.2	P	<0.01	<0.01	0.15				
BDL220	10331.2	Av	1.75	1.37	1.15				
BDL220	10331.5	P	0.01	0.02	0.05				
BDL220	10331.5	Av	1.48	1.64	1.75				
BDL220	10334.2	P	<0.01	<0.01	0.04				
BDL220	10334.2	Av	1.58	1.37	1.21				
BDL221	10341.3	P	0.13	0.13	0.06				
BDL221	10341.3	Av	1.26	1.19	1.44				
BDL221	10341.4	P	<0.01	0.01	<0.01				
BDL221	10341.4	Av	1.57	1.3	1.35				
BDL221	10343.3	P		0.11	0.06	0.01	<0.01	<0.01	0.13
BDL221	10343.3	Av		1.1	1.16	1.37	1.4	1.4	1.55
BDL221	10341.1	P	0.26	0.45	0.19				
BDL221	10341.1	Av	1.18	1.15	1.26				
BDL221	10342.1	P	<0.01	<0.01	0.02	<0.01	<0.01	<0.01	<0.01
BDL221	10342.1	Av	1.9	1.46	1.32	2.02	2.08	1.68	4.31

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL221	10343.1	P	0.18	0.05	0.07	0.26	<0.01	<0.01	<0.01
BDL221	10343.1	Av	1.3	1.51	1.58	1.12	1.35	1.32	1.67
BDL221	10343.3	P	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
BDL221	10343.3	Av	2.7	1.8	1.59	1.75	2.12	1.87	2.4
BDL221	10343.4	P	0.05	0.11	0.19	0.01	<0.01	0.05	0.02
BDL221	10343.4	Av	1.88	1.56	1.57	1.46	1.65	1.49	2.07
BDL221	10344.3	P	0.02	0.01	0.05		0.1	0.18	<0.01
BDL221	10344.3	Av	1.9	1.61	1.56		1.14	1.12	1.63
BDL223	10793.5	P						0.11	
BDL223	10793.5	Av						1.14	
BDL223	10793.8	P				0.21	0.17	0.08	
BDL223	10793.8	Av				1.18	1.17	1.18	
BDL223	10796.2	P	<0.01	0.06					
BDL223	10796.2	Av	1.22	1.14					
BDL223	10791.1	P			0.07		0.12	0.03	
BDL223	10791.1	Av			1.21		1.17	1.2	
BDL223	10793.3	P	0.01	0.04	0.02			0.14	
BDL223	10793.3	Av	1.33	1.3	1.55			1.18	
BDL223	10793.5	P					0.07	0.04	
BDL223	10793.5	Av					1.14	1.18	
BDL223	10793.8	P		0.08	0.08	<0.01	<0.01	<0.01	<0.01
BDL223	10793.8	Av		1.17	1.19	1.54	1.65	1.5	1.64
BDL223	10796.1	P			0.11				
BDL223	10796.1	Av			1.15				
BDL224	10451.3	P		0.29					
BDL224	10451.3	Av		1.11					
BDL224	10451.5	P			0.1				
BDL224	10451.5	Av			1.15				
BDL224	10451.7	P	<0.01	<0.01	0.01	0.03	0.07	0.23	0.16
BDL224	10451.7	Av	1.58	1.69	1.8	1.39	1.26	1.19	1.41
BDL226	10861.2	P						0.4	
BDL226	10861.2	Av						1.1	
BDL227	11491.1	P				<0.01	<0.01	0.03	0.01
BDL227	11491.1	Av				1.5	1.46	1.21	1.95
BDL227	11491.3	P	<0.01	<0.01	<0.01	0.01	<0.01	0.02	0.04

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL227	11491.3	Av	2.12	1.6	1.42	1.64	1.65	1.56	2.26
BDL227	11491.5	P	<0.01	<0.01	0.02	<0.01	<0.01	0.01	<0.01
BDL227	11491.5	Av	1.84	1.53	1.42	1.78	1.85	1.64	2.73
BDL227	11492.5	P	<0.01	<0.01	0.01	0.26	0.03	0.04	0.01
BDL227	11492.5	Av	2.13	1.91	1.72	1.17	1.48	1.32	1.58
BDL227	11493.5	P	<0.01	<0.01	0.02	0.33	<0.01	<0.01	0.02
BDL227	11493.5	Av	1.55	1.63	1.74	1.1	1.47	1.47	1.53
BDL230	10671.3	P	<0.01	0.06	0.01				
BDL230	10671.3	Av	1.36	1.44	1.81				
BDL230	10671.5	P				0.33	0.49	0.46	
BDL230	10671.5	Av				1.22	1.16	1.11	
BDL231	11111.1	P	<0.01	<0.01	<0.01				
BDL231	11111.1	Av	1.55	1.53	1.51				
BDL231	11111.2	P	0.11	0.15	0.04	0.19		0.04	
BDL231	11111.2	Av	1.12	1.12	1.28	1.15		1.19	
BDL231	11111.3	P	<0.01	0.04	0.05				
BDL231	11111.3	Av	1.35	1.26	1.37				
BDL231	11112.2	P	<0.01	0.01	0.03	<0.01	<0.01	<0.01	0.01
BDL231	11112.2	Av	1.72	1.43	1.38	2.08	1.68	1.48	2.34
BDL231	11116.5	P	<0.01	<0.01	<0.01	<0.01	0.01	0.03	<0.01
BDL231	11116.5	Av	1.82	1.77	1.76	1.67	1.38	1.31	1.92
BDL231	11111.1	P	<0.01	<0.01	<0.01		0.36		0.07
BDL231	11111.1	Av	1.88	1.78	1.49		1.11		1.43
BDL231	11111.2	P	0.01	0.03	0.01	0.01	0.1		0.05
BDL231	11111.2	Av	1.66	1.6	1.45	1.28	1.15		1.51
BDL231	11111.3	P				0.01	0.26		0.53
BDL231	11111.3	Av				1.31	1.1		1.1
BDL231	11112.2	P	0.24	0.17		<0.01	<0.01		<0.01
BDL231	11112.2	Av	1.12	1.12		1.51	1.3		1.97
BDL231	11116.5	P				0.11			0.24
BDL231	11116.5	Av				1.19			1.35
BDL232	10904.1	P		0.61	0.54		0.43	0.23	
BDL232	10904.1	Av		1.17	1.2		1.1	1.14	
BDL232	10905.1	P				0.02	<0.01	0.05	0.02
BDL232	10905.1	Av				1.5	1.46	1.25	1.4

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL232	10906.3	P		0.25	0.14		0.59	0.25	
BDL232	10906.3	Av		1.29	1.45		1.1	1.21	
BDL232	10902.2	P				<0.01	<0.01	0.06	0.01
BDL232	10902.2	Av				1.7	1.47	1.24	2.1
BDL232	10905.1	P				<0.01	0.01	0.09	0.03
BDL232	10905.1	Av				1.59	1.42	1.2	2.17
BDL233	10825.4	P				<0.01	<0.01	<0.01	<0.01
BDL233	10825.4	Av				1.47	1.45	1.35	2.01
BDL233	10822.4	P			0.14				
BDL233	10822.4	Av			1.23				
BDL233	10824.2	P						0.05	
BDL233	10824.2	Av						1.16	
BDL233	10825.3	P							0.26
BDL233	10825.3	Av							1.27
BDL233	10825.4	P			0.24	<0.01	<0.01	<0.01	0.19
BDL233	10825.4	Av			1.17	1.56	1.59	1.51	1.3
BDL235	11413.2	P				<0.01	0.01	0.03	0.18
BDL235	11413.2	Av				1.63	1.3	1.19	1.43
BDL235	11413.2	P				<0.01	<0.01	0.05	0.01
BDL235	11413.2	Av				1.57	1.45	1.29	2.12
BDL237	10892.2	P	0.16				0.28	0.12	0.09
BDL237	10892.2	Av	1.21				1.1	1.12	1.4
BDL237	10893.1	P	<0.01	0.02	0.14		0.19	0.28	
BDL237	10893.1	Av	1.96	1.39	1.24		1.13	1.14	
BDL237	10895.3	P	<0.01	<0.01	0.01	0.01	<0.01	<0.01	0.02
BDL237	10895.3	Av	2.2	1.99	1.9	1.84	1.97	1.79	2.09
BDL237	10896.1	P	0.19		0.49				
BDL237	10896.1	Av	1.29		1.1				
BDL238	10951.4	P				0.08			0.01
BDL238	10951.4	Av				1.2			1.69
BDL238	10952.3	P	0.05						
BDL238	10952.3	Av	1.2						
BDL238	10953.3	P							0.32
BDL238	10953.3	Av							1.5
BDL238	10954.2	P				0.05			0.59

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL238	10954.2	Av				1.24			1.13
BDL238	10954.3	P	0.09			0.03	0.17	0.26	0.17
BDL238	10954.3	Av	1.16			1.3	1.12	1.1	1.45
BDL238	10951.4	P				0.13	0.04	0.23	0.17
BDL238	10951.4	Av				1.22	1.24	1.11	1.39
BDL238	10952.3	P				0.02	0.01	0.02	0.11
BDL238	10952.3	Av				1.44	1.43	1.37	1.54
BDL238	10954.2	P				0.02	0.08	0.17	0.22
BDL238	10954.2	Av				1.26	1.2	1.17	1.2
BDL240	10802.2	P	<0.01	0.2	0.31		0.16	0.1	0.1
BDL240	10802.2	Av	1.57	1.23	1.17		1.17	1.19	1.25
BDL240	10803.5	P	0.11	0.4	0.31				
BDL240	10803.5	Av	1.4	1.11	1.16				
BDL240	10806.4	P	<0.01	0.18					
BDL240	10806.4	Av	1.29	1.16					
BDL240	10806.6	P	0.02	0.02	0.04	0.02	0.01	0.01	0.03
BDL240	10806.6	Av	2.29	1.87	1.52	1.59	1.63	1.49	3.75
BDL241	10873.1	P							0.51
BDL241	10873.1	Av							1.2
BDL241	10874.3	P							0.32
BDL241	10874.3	Av							1.42
BDL241	10875.1	P				0.04	0.01	0.13	<0.01
BDL241	10875.1	Av				1.4	1.34	1.21	2.29
BDL241	10874.3	P	<0.01	<0.01	0.02			0.1	
BDL241	10874.3	Av	1.34	1.34	1.43			1.15	
BDL241	10875.1	P				<0.01	0.03	0.3	0.03
BDL241	10875.1	Av				1.32	1.29	1.16	1.52
BDL242	10731.3	P		0.03			0.06	0.18	0.11
BDL242	10731.3	Av		1.12			1.14	1.13	1.36
BDL242	10731.5	P				0.4	0.11	0.18	0.32
BDL242	10731.5	Av				1.13	1.16	1.1	1.23
BDL242	10731.2	P	0.36						
BDL242	10731.2	Av	1.2						
BDL242	10731.3	P	<0.01	<0.01	<0.01		0.02	<0.01	0.03
BDL242	10731.3	Av	3.13	2.79	2.3		1.26	1.27	1.65

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL249	11401.2	Av	3.01	2.92	2.73	1.78	1.84	1.71	3.51
BDL249	11401.5	P	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
BDL249	11401.5	Av	2.17	1.83	1.52	1.66	1.64	1.41	3.31
BDL249	11402.4	P	0.01	0.01	0.03	0.18	<0.01	<0.01	0.04
BDL249	11402.4	Av	1.49	1.44	1.39	1.14	1.64	1.49	1.54
BDL249	11403.2	P			0.48		0.19	0.26	
BDL249	11403.2	Av			1.12		1.2	1.16	
BDL249	11404.3	P				0.09	0.07	0.02	0.12
BDL249	11404.3	Av				1.41	1.4	1.26	1.57
BDL249	11401.2	P				0.29	0.2		0.17
BDL249	11401.2	Av				1.11	1.1		1.21
BDL249	11401.5	P				0.17			0.46
BDL249	11401.5	Av				1.2			1.21
BDL249	11403.2	P				0.04	0.15		0.35
BDL249	11403.2	Av				1.3	1.13		1.23
BDL249	11404.3	P				0.21	0.04	0.12	0.35
BDL249	11404.3	Av				1.11	1.27	1.19	1.14
BDL250	10841.3	P				0.01	0.12	0.17	
BDL250	10841.3	Av				1.26	1.23	1.12	
BDL250	10842.3	P		0.05	0.23	<0.01	<0.01	<0.01	0.03
BDL250	10842.3	Av		1.16	1.12	1.55	1.5	1.38	1.39
BDL250	10846.2	P				<0.01	<0.01	<0.01	
BDL250	10846.2	Av				1.36	1.37	1.29	
BDL250	10846.3	P				0.09	<0.01	<0.01	0.12
BDL250	10846.3	Av				1.33	1.39	1.29	1.44
BDL250	10841.3	P	0.01	0.1	0.09	<0.01	<0.01	<0.01	0.01
BDL250	10841.3	Av	1.23	1.21	1.22	1.82	1.89	1.68	2.37
BDL250	10842.3	P	0.05	0.15	0.26	<0.01	0.23	0.48	0.02
BDL250	10842.3	Av	1.73	1.59	1.5	1.4	1.26	1.14	2.03
BDL250	10843.2	P	0.01	0.02	0.04	<0.01	0.01	0.01	<0.01
BDL250	10843.2	Av	1.97	1.67	1.61	1.37	1.44	1.37	2.33
BDL250	10846.2	P	<0.01	0.01	<0.01	0.11	0.02	<0.01	0.08
BDL250	10846.2	Av	2.3	1.59	1.69	1.21	1.33	1.36	1.39
BDL250	10846.3	P	<0.01	<0.01	<0.01	0.01	<0.01	<0.01	0.01
BDL250	10846.3	Av	1.92	1.69	1.78	1.66	1.98	1.79	2.83

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL252	10881.1	P	<0.01	<0.01	<0.01	0.13	0.01	<0.01	0.04
BDL252	10881.1	Av	1.96	1.94	1.72	1.29	1.48	1.42	1.52
BDL252	10882.1	P	<0.01	<0.01	<0.01	0.05	0.01	0.01	0.02
BDL252	10882.1	Av	2.97	2.36	2.12	1.65	1.88	1.77	3.43
BDL252	10882.2	P	<0.01	<0.01	<0.01	0.03	0.04	<0.01	0.03
BDL252	10882.2	Av	1.89	1.48	1.43	1.47	1.54	1.45	2.04
BDL252	10882.4	P	0.03	<0.01	0.11	<0.01	0.01	0.03	0.01
BDL252	10882.4	Av	1.64	1.4	1.3	1.7	1.48	1.31	2.99
BDL252	10884.1	P	<0.01	0.02	0.13				
BDL252	10884.1	Av	1.55	1.31	1.17				
BDL58	10281.5	P	0.23	0.48	0.16				
BDL58	10281.5	Av	1.25	1.18	1.3				
BDL58	10282.3	P	<0.01	<0.01	0.01	0.12	<0.01	<0.01	0.26
BDL58	10282.3	Av	1.79	1.72	1.77	1.27	1.62	1.62	1.43
BDL62	10682.1	P							0.53
BDL62	10682.1	Av							1.16
BDL62	10684.2	P				0.01			0.22
BDL62	10684.2	Av				1.43			1.3
BDL62	10682.1	P		0.37	0.56				
BDL62	10682.1	Av		1.17	1.16				
BDL62	10684.2	P						0.09	
BDL62	10684.2	Av						1.13	
BDL64	10651.5	P		0.08	0.27				
BDL64	10651.5	Av		1.26	1.56				
BDL64	10653.1	P	0.01	0.06	0.01				
BDL64	10653.1	Av	1.24	1.23	1.53				
BDL64	10654.3	P		0.32	0.3	0.2	0.04	0.09	
BDL64	10654.3	Av		1.21	1.38	1.11	1.4	1.4	
BDL64	10651.1	P		0.53					
BDL64	10651.1	Av		1.11					
BDL64	10653.1	P	0.11	0.02	0.05	0.01	<0.01	0.01	0.14
BDL64	10653.1	Av	1.34	1.57	1.61	1.34	1.42	1.28	1.34
BDL64	10654.3	P	0.01	<0.01	<0.01	<0.01	<0.01	0.01	0.29
BDL64	10654.3	Av	1.41	1.49	1.46	1.28	1.29	1.23	1.27
BDL64	10651.1	P	<0.01	<0.01		0.02	0.15		0.2

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL64	10651.1	Av	1.5	1.31		1.41	1.13		1.44
BDL64	10651.3	P				<0.01	<0.01	0.13	<0.01
BDL64	10651.3	Av				1.96	1.41	1.15	3.02
BDL64	10653.1	P				0.03	0.3		0.08
BDL64	10653.1	Av				1.64	1.18		1.64
BDL64	10654.3	P				<0.01	0.22		0.04
BDL64	10654.3	Av				1.82	1.23		2.34
BDL79	11041.1	P							0.62
BDL79	11041.1	Av							1.12
BDL79	11042.1	P							0.61
BDL79	11042.1	Av							1.13
BDL79	11043.1	P							0.08
BDL79	11043.1	Av							1.64
BDL79	11042.3	P						0.33	
BDL79	11042.3	Av						1.12	
BDL79	11043.1	P			0.04	0.17	<0.01	0.01	0.37
BDL79	11043.1	Av			1.23	1.19	1.36	1.29	1.19
BDL81	10372.2	P	<0.01	0.35	0.43				
BDL81	10372.2	Av	1.25	1.11	1.13				
BDL81	10374.1	P							0.3
BDL81	10374.1	Av							1.24
BDL85	10411.3	P					0.08	0.03	
BDL85	10411.3	Av					1.15	1.12	
BDL85	10414.1	P	0.2		0.04				
BDL85	10414.1	Av	1.16		1.26				
BDL85	10414.2	P	0.19	0.02	<0.01				
BDL85	10414.2	Av	1.1	1.2	1.35				
BDL88	10291.2	P	0.01	0.03	0.03			0.07	
BDL88	10291.2	Av	1.88	1.59	1.52			1.13	
BDL88	10291.4	P	0.02	<0.01	<0.01		0.03	<0.01	
BDL88	10291.4	Av	1.45	1.43	1.53		1.21	1.28	
BDL88	10291.5	P	0.12	0.15	0.37				0.67
BDL88	10291.5	Av	1.43	1.35	1.17				1.11
BDL88	10293.3	P	<0.01	<0.01	<0.01				0.57
BDL88	10293.3	Av	2.76	2.34	2.24				1.12

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Leaf Area TPI</i>	<i>Leaf Area TP2</i>	<i>Leaf Area TP3</i>	<i>Roots Length TPI</i>	<i>Roots Length TP2</i>	<i>Roots Length TP3</i>	<i>Roots Coverage TPI</i>
BDL88	10291.2	P	<0.01	<0.01	<0.01				
BDL88	10291.2	Av	1.64	1.57	1.58				
BDL88	10291.4	P	<0.01	<0.01	<0.01				
BDL88	10291.4	Av	1.67	1.5	1.53				
BDL88	10291.5	P	<0.01	<0.01	<0.01				
BDL88	10291.5	Av	1.8	1.7	1.84				
BDL88	10293.3	P	0.03	0.05	0.06				
BDL88	10293.3	Av	1.19	1.16	1.22				
BDL88	10294.2	P	<0.01	0.01	0.02				
BDL88	10294.2	Av	1.82	1.66	1.78				
BDL90	10924.2	P				0.05	0.01	<0.01	0.16
BDL90	10924.2	Av				1.43	1.55	1.48	1.9
BDL90	10925.4	P				<0.01	<0.01	<0.01	0.07
BDL90	10925.4	Av				1.95	1.91	1.62	2.22
BDL90	10924.2	P	0.12	0.2	0.09	0.02	0.01	0.01	0.04
BDL90	10924.2	Av	1.32	1.23	1.22	1.31	1.31	1.31	1.93
BDL90	10925.4	P	0.04	0.12		<0.01	<0.01	<0.01	0.02
BDL90	10925.4	Av	1.25	1.15		1.68	1.61	1.51	2.25
BDL90	10921.6	P			0.1				
BDL90	10921.6	Av			1.19				
BDL90	10924.2	P			0.38	<0.01	<0.01	<0.01	0.03
BDL90	10924.2	Av			1.13	1.48	1.6	1.51	1.41
BDL90	10925.4	P	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
BDL90	10925.4	Av	1.6	1.66	1.8	1.85	1.81	1.71	2.9
BDL94	11721.4	P	0.04	0.03	0.07				
BDL94	11721.4	Av	1.41	1.39	1.16				
BDL94	11725.2	P	0.04						
BDL94	11725.2	Av	1.16						
BDL94	11725.3	P	0.04						
BDL94	11725.3	Av	1.21						
BDL94	11725.5	P	0.14						
BDL94	11725.5	Av	1.1						
BDL94	11725.3	P	0.01						
BDL94	11725.3	Av	1.29						

Table 21. "P" = P-value; "Av" = ratio between the averages of event and control. Note that when the average ratio is higher than "1" the effect of exogenous expression of the gene is an increase of

the desired trait; "Par" = Parameter according to the measured parameters; "Ev" = event. TP1 = Time point 1; TP2 = Time point 2; TP3 = Time point 3.

Table 22
Results obtained in a tissue culture assay

5

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL102	10471.1	P	0.08	0.05		0.11	0.36		
BDL102	10471.1	Av	1.58	1.35		1.26	1.11		
BDL102	10471.3	P	0.08	0.15		0.02	0.15		
BDL102	10471.3	Av	1.72	1.58		1.57	1.18		
BDL102	10472.1	P	0.19	0.28		0.25		0.29	
BDL102	10472.1	Av	1.28	1.22		1.19		1.18	
BDL102	10474.2	P	0.33	0.16	0.01	0.02	0.07	0.21	0.26
BDL102	10474.2	Av	1.22	1.38	1.4	1.44	1.19	1.53	1.5
BDL102	10474.6	P		0.2	0.31	0.14			
BDL102	10474.6	Av		1.19	1.15	1.22			
BDL118	10481.2	P	0.13	0.24	<0.01	0.14	0.28	0.03	0.01
BDL118	10481.2	Av	1.2	1.19	1.99	1.23	1.15	2.14	1.99
BDL118	10481.5	P			0.01			0.02	0.01
BDL118	10481.5	Av			1.36			2.03	1.97
BDL118	10483.4	P						0.07	0.03
BDL118	10483.4	Av						1.93	1.65
BDL118	10484.3	P			<0.01			<0.01	<0.01
BDL118	10484.3	Av			1.6			1.98	2.01
BDL140	10421.3	P	0.14	0.06	<0.01	<0.01		0.03	0.13
BDL140	10421.3	Av	1.45	1.64	1.7	1.68		1.81	1.57
BDL140	10423.1	P	0.21	0.28	0.22	0.16		0.41	0.33
BDL140	10423.1	Av	1.31	1.25	1.21	1.25		1.17	1.16
BDL140	10424.4	P		0.18	<0.01	0.2		0.08	0.45
BDL140	10424.4	Av		1.13	1.66	1.18		1.46	1.2
BDL152	10431.4	P		0.21	0.24	0.13		0.23	0.02
BDL152	10431.4	Av		1.23	1.18	1.27		1.18	1.27
BDL152	10432.5	P	0.1	0.04		0.03			
BDL152	10432.5	Av	1.25	1.31		1.32			
BDL152	10434.4	P	0.02	<0.01		<0.01	0.03		
BDL152	10434.4	Av	1.76	1.51		1.44	1.23		
BDL153	10142.2	P	0.04	0.58					
BDL153	10142.2	Av	1.57	1.1					
BDL153	10144.1	P	0.09	0.12	0.02	0.09	0.12	0.36	
BDL153	10144.1	Av	1.51	1.32	1.42	1.35	1.18	1.18	
BDL154	10703.1	P	0.38	0.47		0.52	0.44		
BDL154	10703.1	Av	1.13	1.18		1.15	1.13		
BDL154	10703.3	P					0.34		
BDL154	10703.3	Av					1.17		
BDL154	10703.5	P						0.38	0.16
BDL154	10703.5	Av						1.11	1.16
BDL154	10703.1	P	0.03	0.05	<0.01	0.01	0.01	0.01	0.01
BDL154	10703.1	Av	1.48	1.55	1.71	1.7	1.37	1.36	1.57
BDL154	10703.5	P	0.1	0.04	0.11	0.01	<0.01	0.34	0.11
BDL154	10703.5	Av	1.75	1.7	1.29	1.73	1.59	1.24	1.38

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL154	10703.6	P	0.07	0.04		0.07	0.14		
BDL154	10703.6	Av	1.63	1.47		1.45	1.28		
BDL155	9991.2	P						0.22	0.45
BDL155	9991.2	Av						1.44	1.27
BDL155	9993.2	P						0.55	
BDL155	9993.2	Av						1.45	
BDL155	9994.4	P					0.19		
BDL155	9994.4	Av					1.13		
BDL156	10852.6	P	0.41	0.51		0.19	0.09		
BDL156	10852.6	Av	1.1	1.12		1.21	1.22		
BDL156	10852.7	P					0.35		
BDL156	10852.7	Av					1.13		
BDL156	10853.6	P	0.58	0.25	0.07	0.07	0.16	0.28	0.04
BDL156	10853.6	Av	1.13	1.24	1.21	1.33	1.19	1.2	1.26
BDL156	10852.6	P			0.05	0.59	0.33	0.35	0.21
BDL156	10852.6	Av			1.29	1.13	1.16	1.12	1.24
BDL156	10853.6	P	0.42	0.25	<0.01	0.05	0.06	<0.01	0.01
BDL156	10853.6	Av	1.29	1.69	1.97	1.83	1.43	1.78	2.26
BDL156	10854.4	P		0.03	<0.01	<0.01	<0.01	<0.01	<0.01
BDL156	10854.4	Av		1.59	1.87	1.76	1.53	1.61	1.94
BDL156	10855.3	P	0.32	<0.01	<0.01	<0.01	0.01	0.46	0.05
BDL156	10855.3	Av	1.17	1.78	1.49	1.99	1.44	1.18	1.42
BDL158	9973.3	P	0.26						
BDL158	9973.3	Av	1.22						
BDL158	9971.3	P		0.12	0.45	0.03	0.3		
BDL158	9971.3	Av		1.27	1.11	1.34	1.11		
BDL158	9973.1	P		0.04		0.1			
BDL158	9973.1	Av		1.25		1.24			
BDL158	9973.3	P	0.04	0.27		0.32			
BDL158	9973.3	Av	1.43	1.23		1.16			
BDL158	9974.2	P	0.01	0.03	0.34	<0.01			0.49
BDL158	9974.2	Av	1.48	1.41	1.14	1.45			1.1
BDL158	9974.3	P	0.08	0.05	0.02	<0.01		0.01	0.01
BDL158	9974.3	Av	1.5	1.49	1.54	1.54		1.5	1.54
BDL158	9971.3	P		0.33	0.06	0.13	<0.01		
BDL158	9971.3	Av		1.13	1.25	1.24	1.46		
BDL158	9973.1	P	0.09	0.25	0.02	0.45	0.29		0.16
BDL158	9973.1	Av	1.22	1.22	1.37	1.13	1.13		1.2
BDL158	9973.3	P	0.02	0.1		0.03	<0.01		
BDL158	9973.3	Av	1.62	1.43		1.42	1.4		
BDL158	9974.2	P	0.06	0.02		0.03			
BDL158	9974.2	Av	1.21	1.24		1.32			
BDL158	9974.3	P			0.51				
BDL158	9974.3	Av			1.11				
BDL160	10011.5	P			0.24			<0.01	0.53
BDL160	10011.5	Av			1.23			1.48	1.33
BDL160	10011.6	P						0.51	0.66
BDL160	10011.6	Av						1.18	1.1
BDL160	10011.7	P						0.56	
BDL160	10011.7	Av						1.28	
BDL160	10015.1	P						0.03	0.12

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL160	10015.1	Av						1.57	1.25
BDL160	10011.5	P	0.61	0.38		0.42			
BDL160	10011.5	Av	1.11	1.14		1.14			
BDL160	10013.1	P			0.32				
BDL160	10013.1	Av			1.16				
BDL160	10014.9	P		0.52		0.37			
BDL160	10014.9	Av		1.16		1.18			
BDL160	10015.2	P			0.05			0.1	0.11
BDL160	10015.2	Av			1.28			1.29	1.42
BDL167	10042.3	P					0.12		
BDL167	10042.3	Av					1.17		
BDL167	10043.1	P		0.58	0.13	0.21	0.23		
BDL167	10043.1	Av		1.17	1.16	1.25	1.18		
BDL167	10043.2	P			0.18	0.34	0.09		
BDL167	10043.2	Av			1.14	1.1	1.19		
BDL167	10043.3	P	0.1	<0.01	<0.01	<0.01	<0.01		
BDL167	10043.3	Av	1.27	1.67	1.54	1.79	1.46		
BDL167	10044.2	P	0.05	<0.01	<0.01	<0.01	0.08		0.61
BDL167	10044.2	Av	1.26	1.48	1.38	1.55	1.2		1.12
BDL167	10043.1	P	<0.01	0.17	0.04	0.16	0.14	0.06	
BDL167	10043.1	Av	1.63	1.27	1.4	1.3	1.17	1.6	
BDL167	10044.2	P	0.01						
BDL167	10044.2	Av	1.39						
BDL168	9881.4	P	<0.01	0.14		0.08	0.05		
BDL168	9881.4	Av	1.74	1.47		1.42	1.21		
BDL168	9882.1	P	0.08	0.69					
BDL168	9882.1	Av	1.85	1.11					
BDL168	9884.1	P	<0.01	0.11		0.34	0.22		
BDL168	9884.1	Av	1.84	1.29		1.2	1.15		
BDL169	10744.2	P	0.02	0.17		0.13	0.24		
BDL169	10744.2	Av	1.34	1.21		1.22	1.16		
BDL169	10747.1	P	0.39			0.4			
BDL169	10747.1	Av	1.1			1.11			
BDL169	10747.5	P	0.04	0.02		0.02	0.07		
BDL169	10747.5	Av	1.59	1.41		1.37	1.28		
BDL171	10661.2	P	0.03	0.01	0.19	<0.01	0.01	0.06	0.02
BDL171	10661.2	Av	2.26	2.17	1.28	2.22	1.3	1.73	2.47
BDL171	10661.5	P		0.14	0.32	0.04		0.02	<0.01
BDL171	10661.5	Av		1.33	1.19	1.38		1.35	2.02
BDL171	10662.3	P		0.32		0.19			0.56
BDL171	10662.3	Av		1.12		1.18			1.13
BDL171	10663.3	P				0.45			0.39
BDL171	10663.3	Av				1.14			1.19
BDL171	10664.1	P		0.26	0.25	0.1	0.46	0.03	0.03
BDL171	10664.1	Av		1.33	1.24	1.35	1.1	1.39	1.89
BDL173	9951.2	P	0.37					0.27	
BDL173	9951.2	Av	1.32					1.43	
BDL173	9952.1	P	0.04	0.02		0.01	0.08		
BDL173	9952.1	Av	2.68	1.72		1.61	1.21		
BDL173	9952.2	P	0.15	0.56					
BDL173	9952.2	Av	1.44	1.11					

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL173	9954.3	P						0.14	
BDL173	9954.3	Av						1.46	
BDL174	11082.1	P	0.29	0.39		0.43	0.05		
BDL174	11082.1	Av	1.24	1.15		1.14	1.26		
BDL174	11083.1	P	0.07	0.11	0.41	0.05	0.15	0.48	0.37
BDL174	11083.1	Av	1.76	1.43	1.13	1.42	1.21	1.1	1.2
BDL174	11083.2	P	0.17	0.25	0.13	0.09	0.04	0.2	0.4
BDL174	11083.2	Av	1.27	1.36	1.26	1.38	1.44	1.21	1.19
BDL174	11084.1	P	0.04	0.06	<0.01	<0.01	0.01	0.06	0.05
BDL174	11084.1	Av	2.13	1.94	2.01	1.96	1.42	2.06	2.27
BDL174	11085.1	P	<0.01	<0.01	0.1	<0.01	<0.01	0.2	0.24
BDL174	11085.1	Av	2.66	2.13	1.27	2.17	2	1.24	1.37
BDL174	11083.2	P			0.19			0.12	0.29
BDL174	11083.2	Av			1.15			1.2	1.14
BDL174	11084.1	P	0.6						
BDL174	11084.1	Av	1.1						
BDL174	11085.1	P	0.05	0.32					
BDL174	11085.1	Av	1.42	1.14					
BDL176	9891.4	P		0.67	0.01	0.47		0.06	
BDL176	9891.4	Av		1.11	1.45	1.17		2.02	
BDL176	9893.2	P	0.21	0.4	0.22	0.26		0.15	
BDL176	9893.2	Av	1.37	1.22	1.21	1.25		1.38	
BDL176	9893.3	P	0.47	0.23		0.12		0.14	0.48
BDL176	9893.3	Av	1.17	1.31		1.3		1.29	1.22
BDL177	10521.3	P	0.66	0.34		0.28	0.06		
BDL177	10521.3	Av	1.1	1.15		1.17	1.3		
BDL181	11293.6	P		0.59	0.1	0.37	0.45	0.39	0.37
BDL181	11293.6	Av		1.15	1.3	1.24	1.15	1.17	1.26
BDL181	11294.7	P			0.04		0.12	0.11	0.1
BDL181	11294.7	Av			1.36		1.28	1.26	1.38
BDL181	11293.1	P			0.25	0.4	0.02	0.05	0.02
BDL181	11293.1	Av			1.19	1.11	1.22	1.59	1.58
BDL181	11293.6	P	0.23	0.15	<0.01	0.09	0.03	<0.01	<0.01
BDL181	11293.6	Av	1.23	1.22	1.34	1.25	1.18	1.48	1.45
BDL181	11294.7	P	0.02	0.21	0.3	0.05	0.01		0.57
BDL181	11294.7	Av	1.55	1.26	1.11	1.3	1.23		1.1
BDL182	10691.8	P			0.42			0.06	0.01
BDL182	10691.8	Av			1.15			1.53	2.11
BDL182	10692.2	P	0.53	0.2	0.28	0.07		0.28	0.15
BDL182	10692.2	Av	1.12	1.21	1.21	1.28		1.47	1.93
BDL182	10692.3	P		0.63		0.3	0.43		
BDL182	10692.3	Av		1.13		1.21	1.1		
BDL182	10693.3	P	0.02	0.03	0.22	<0.01		0.17	0.01
BDL182	10693.3	Av	1.37	1.55	1.25	1.62		1.41	1.88
BDL182	10693.5	P	0.02	<0.01	0.32	<0.01	0.13	0.06	0.01
BDL182	10693.5	Av	1.47	1.72	1.19	1.8	1.13	1.38	1.91
BDL183	9943.4	P	0.19	0.3		0.48			
BDL183	9943.4	Av	1.3	1.16		1.13			
BDL183	9944.4	P	0.02	0.52		0.46	0.06	0.44	
BDL183	9944.4	Av	1.37	1.15		1.15	1.2	1.16	
BDL189	11353.3	P		0.17	0.18	0.18			0.46

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL189	11353.3	Av		1.18	1.16	1.2			1.1
BDL189	11351.2	P			0.43				0.4
BDL189	11351.2	Av			1.1				1.17
BDL189	11353.3	P		0.24	0.22	0.11			0.33
BDL189	11353.3	Av		1.15	1.11	1.22			1.1
BDL189	11353.5	P						0.65	
BDL189	11353.5	Av						1.1	
BDL189	11355.4	P			0.22			0.28	0.32
BDL189	11355.4	Av			1.11			1.36	1.13
BDL196	10242.2	P					0.28	0.49	
BDL196	10242.2	Av					1.13	1.15	
BDL196	10243.4	P	<0.01	0.12	0.12	0.13	0.17	0.57	
BDL196	10243.4	Av	1.45	1.36	1.27	1.34	1.17	1.11	
BDL196	10244.1	P	0.16	0.43		0.28		0.02	
BDL196	10244.1	Av	1.65	1.27		1.27		1.45	
BDL196	10242.2	P			0.47				
BDL196	10242.2	Av			1.1				
BDL196	10243.3	P			0.6			0.43	0.23
BDL196	10243.3	Av			1.1			1.13	1.23
BDL196	10243.4	P	0.51	0.15	<0.01	0.11	0.16	0.13	0.03
BDL196	10243.4	Av	1.12	1.23	1.65	1.27	1.18	1.42	1.69
BDL197	11362.2	P		0.33	0.03	0.18	0.14		0.38
BDL197	11362.2	Av		1.18	1.35	1.32	1.24		1.19
BDL197	11363.1	P		0.23		0.12	<0.01		
BDL197	11363.1	Av		1.25		1.38	1.53		
BDL197	11363.6	P	0.39	0.01	0.05	<0.01	0.04	0.02	0.04
BDL197	11363.6	Av	1.19	1.59	1.31	1.77	1.36	1.41	1.61
BDL197	11364.1	P		0.04	<0.01	<0.01	<0.01	0.08	<0.01
BDL197	11364.1	Av		1.67	1.63	1.85	1.48	1.44	1.91
BDL197	11364.5	P			0.02	0.47		0.11	0.04
BDL197	11364.5	Av			1.37	1.16		1.18	1.42
BDL197	11363.1	P					0.1	0.53	0.17
BDL197	11363.1	Av					1.12	1.1	1.23
BDL197	11363.6	P	0.11	0.03	0.01	<0.01	<0.01	0.06	0.04
BDL197	11363.6	Av	1.39	1.49	1.3	1.58	1.24	1.45	1.55
BDL197	11364.1	P	0.3			0.53		0.62	0.07
BDL197	11364.1	Av	1.18			1.11		1.16	1.75
BDL197	11364.5	P					0.12		0.49
BDL197	11364.5	Av					1.12		1.13
BDL201	9961.2	P			0.47			0.57	0.47
BDL201	9961.2	Av			1.13			1.1	1.15
BDL201	9961.3	P						0.13	
BDL201	9961.3	Av						1.39	
BDL201	9961.4	P	0.19	0.41					
BDL201	9961.4	Av	1.3	1.14					
BDL201	9964.3	P	0.17	0.05		0.27			
BDL201	9964.3	Av	1.25	1.21		1.15			
BDL203	9831.14	P						0.62	0.42
BDL203	9831.14	Av						1.13	1.27
BDL203	9831.7	P					0.11		
BDL203	9831.7	Av					1.22		

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL203	9833.6	P						0.71	
BDL203	9833.6	Av						1.14	
BDL203	9835.2	P					0.33		
BDL203	9835.2	Av					1.14		
BDL220	10331.2	P						<0.01	0.06
BDL220	10331.2	Av						1.51	1.25
BDL220	10331.5	P		0.22	<0.01	0.1	0.27	0.05	0.11
BDL220	10331.5	Av		1.22	1.81	1.28	1.11	1.97	1.6
BDL220	10333.5	P						0.47	0.41
BDL220	10333.5	Av						1.16	1.18
BDL220	10334.2	P			0.34			0.35	
BDL220	10334.2	Av			1.13			1.14	
BDL221	10341.1	P						0.19	
BDL221	10341.1	Av						1.3	
BDL221	10341.3	P			0.01			0.01	<0.01
BDL221	10341.3	Av			1.5			1.92	2.18
BDL221	10341.4	P			0.02			<0.01	0.01
BDL221	10341.4	Av			1.29			1.98	2.04
BDL221	10343.3	P	0.03	0.01	0.07	<0.01	<0.01		
BDL221	10343.3	Av	1.55	1.6	1.21	1.6	1.41		
BDL221	10344.3	P						0.41	
BDL221	10344.3	Av						1.11	
BDL221	10341.1	P			0.15				0.59
BDL221	10341.1	Av			1.27				1.16
BDL221	10342.1	P	<0.01	<0.01	0.17	<0.01	<0.01	0.25	0.42
BDL221	10342.1	Av	3.03	2.06	1.22	1.88	1.52	1.24	1.24
BDL221	10343.1	P	0.01	<0.01	0.01	<0.01	<0.01	0.15	0.33
BDL221	10343.1	Av	1.8	1.61	1.62	1.6	1.41	1.46	1.51
BDL221	10343.3	P	0.01	0.02	0.03	<0.01	<0.01		
BDL221	10343.3	Av	3.03	2.69	1.4	2.71	1.92		
BDL221	10343.4	P	0.03	0.08	0.07	<0.01	0.01	0.14	0.08
BDL221	10343.4	Av	2.15	2.07	1.51	2.07	1.49	1.44	1.53
BDL221	10344.3	P	0.01	0.07	0.02	0.02	0.09	0.17	0.12
BDL221	10344.3	Av	1.67	1.49	1.49	1.48	1.23	1.39	1.33
BDL223	10791.1	P					0.33		
BDL223	10791.1	Av					1.13		
BDL223	10793.5	P	0.1	0.01		0.01	0.02		
BDL223	10793.5	Av	1.21	1.32		1.38	1.33		
BDL223	10793.8	P					0.21		
BDL223	10793.8	Av					1.18		
BDL223	10796.1	P				0.58	0.41		
BDL223	10796.1	Av				1.1	1.12		
BDL223	10796.2	P	0.41				0.33	0.34	0.16
BDL223	10796.2	Av	1.11				1.13	1.14	1.4
BDL223	10791.1	P		0.03	0.02	0.05	0.06		0.3
BDL223	10791.1	Av		1.38	1.34	1.43	1.3		1.24
BDL223	10793.3	P		0.34	<0.01	0.08	0.01	0.24	0.16
BDL223	10793.3	Av		1.45	1.64	1.62	1.48	1.15	1.39
BDL223	10793.5	P	0.47	0.23	0.44	0.15	0.04		
BDL223	10793.5	Av	1.13	1.29	1.11	1.34	1.31		
BDL223	10793.8	P	<0.01	<0.01	0.07	<0.01	<0.01		0.21

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL223	10793.8	Av	1.75	1.71	1.26	1.72	1.47		1.26
BDL223	10796.1	P		0.01	0.03	0.01	0.07		0.3
BDL223	10796.1	Av		1.47	1.31	1.61	1.29		1.2
BDL224	10451.5	P			0.17			0.21	0.2
BDL224	10451.5	Av			1.19			1.28	1.25
BDL224	10451.7	P	0.11	0.16	<0.01	0.01	0.35	0.11	0.19
BDL224	10451.7	Av	1.48	1.67	1.85	1.69	1.13	1.48	1.29
BDL224	10451.8	P				0.5			
BDL224	10451.8	Av				1.13			
BDL226	10861.2	P		0.41	0.41	0.15	0.08		
BDL226	10861.2	Av		1.18	1.1	1.26	1.26		
BDL226	10861.4	P		0.18		0.03	0.04		0.23
BDL226	10861.4	Av		1.27		1.37	1.28		1.2
BDL226	10862.2	P		0.56		0.23	0.19		
BDL226	10862.2	Av		1.11		1.2	1.18		
BDL227	11491.1	P	<0.01	0.12		0.25			
BDL227	11491.1	Av	1.59	1.25		1.2			
BDL227	11491.3	P	0.01	0.05	0.06	<0.01	<0.01	0.42	0.35
BDL227	11491.3	Av	1.87	1.71	1.29	1.66	1.52	1.12	1.23
BDL227	11491.5	P	0.01	0.03	0.04	<0.01	<0.01	0.36	0.15
BDL227	11491.5	Av	2.24	2	1.34	1.94	1.58	1.24	1.34
BDL227	11492.5	P	<0.01	0.03	<0.01	<0.01	0.02	0.13	0.12
BDL227	11492.5	Av	2.09	1.71	1.64	1.72	1.39	2.07	1.86
BDL227	11493.5	P	<0.01	0.01	<0.01	<0.01	<0.01	0.16	0.08
BDL227	11493.5	Av	2.13	2.02	1.78	2.05	1.64	1.55	1.73
BDL230	10671.3	P			<0.01			<0.01	<0.01
BDL230	10671.3	Av			1.95			1.84	2.02
BDL231	11111.1	P	0.58	0.12	<0.01	0.1	0.51	0.14	0.55
BDL231	11111.1	Av	1.1	1.24	1.5	1.27	1.1	1.24	1.1
BDL231	11111.2	P	0.24	0.09	<0.01	0.09	0.16		
BDL231	11111.2	Av	1.18	1.24	1.31	1.27	1.2		
BDL231	11111.3	P	0.33	0.08	<0.01	0.03		0.31	0.35
BDL231	11111.3	Av	1.13	1.36	1.38	1.39		1.15	1.2
BDL231	11112.2	P	<0.01	0.01	0.01	<0.01	0.05		0.52
BDL231	11112.2	Av	1.89	1.61	1.3	1.55	1.27		1.21
BDL231	11116.5	P	<0.01	<0.01	<0.01	<0.01	0.23	0.21	0.05
BDL231	11116.5	Av	1.97	1.79	1.74	1.78	1.18	1.28	1.45
BDL231	11111.1	P	0.09	0.31	<0.01	0.38	0.51	0.1	0.16
BDL231	11111.1	Av	1.25	1.18	1.41	1.16	1.1	1.23	1.2
BDL231	11111.2	P	0.02	0.2	0.01	0.32		0.07	0.28
BDL231	11111.2	Av	1.31	1.19	1.4	1.17		1.46	1.24
BDL231	11112.2	P	0.1						
BDL231	11112.2	Av	1.42						
BDL232	10904.1	P	0.6	0.26	0.33	0.16	0.11	0.71	0.5
BDL232	10904.1	Av	1.25	1.36	1.29	1.44	1.29	1.14	1.37
BDL232	10905.1	P	0.27	0.24		0.3			
BDL232	10905.1	Av	1.18	1.26		1.23			
BDL232	10906.3	P	0.48	0.24	0.02	0.07	0.06	0.14	0.13
BDL232	10906.3	Av	1.37	1.63	1.61	1.75	1.44	1.48	1.77
BDL232	10902.2	P	0.02	0.22		0.43			
BDL232	10902.2	Av	1.65	1.24		1.12			

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL232	10905.1	P	0.18						
BDL232	10905.1	Av	1.28						
BDL232	10906.4	P					0.06		
BDL232	10906.4	Av					1.15		
BDL233	10822.4	P					0.28		
BDL233	10822.4	Av					1.13		
BDL233	10824.2	P					0.05		
BDL233	10824.2	Av					1.25		
BDL233	10825.3	P					0.1		
BDL233	10825.3	Av					1.21		
BDL233	10825.4	P	<0.01	0.01		<0.01	0.03		
BDL233	10825.4	Av	1.78	1.49		1.43	1.29		
BDL233	10822.4	P			0.04		0.19	0.26	0.37
BDL233	10822.4	Av			1.37		1.21	1.18	1.24
BDL233	10824.1	P			0.43		0.48		
BDL233	10824.1	Av			1.11		1.1		
BDL233	10824.2	P		0.38	0.3	0.19	<0.01		
BDL233	10824.2	Av		1.16	1.16	1.29	1.5		
BDL233	10825.4	P	0.22	0.02	0.18	0.04	0.01		0.42
BDL233	10825.4	Av	1.23	1.44	1.23	1.46	1.47		1.14
BDL235	11412.2	P		0.34		0.33	0.4		
BDL235	11412.2	Av		1.14		1.17	1.11		
BDL235	11413.2	P	0.07	0.06		0.03			
BDL235	11413.2	Av	1.43	1.39		1.38			
BDL235	11413.3	P						0.51	0.3
BDL235	11413.3	Av						1.15	1.18
BDL235	11411.2	P					0.08		0.62
BDL235	11411.2	Av					1.16		1.15
BDL235	11413.2	P	<0.01	0.08		0.12	0.15		
BDL235	11413.2	Av	1.61	1.33		1.22	1.15		
BDL235	11413.3	P					0.09		
BDL235	11413.3	Av					1.16		
BDL237	10892.1	P						0.05	0.14
BDL237	10892.1	Av						1.43	1.33
BDL237	10892.2	P	0.18	0.19		0.4	0.18		
BDL237	10892.2	Av	1.23	1.16		1.14	1.18		
BDL237	10893.1	P	0.23	0.29	0.51	0.28	0.09		
BDL237	10893.1	Av	1.2	1.17	1.11	1.19	1.25		
BDL237	10895.3	P	0.01	0.01	<0.01	<0.01	<0.01	0.09	0.05
BDL237	10895.3	Av	2.68	2.3	1.85	2.32	1.77	1.59	1.71
BDL237	10896.1	P		0.71		0.57	0.27		
BDL237	10896.1	Av		1.11		1.13	1.21		
BDL238	10951.4	P	0.51						
BDL238	10951.4	Av	1.13						
BDL238	10952.3	P	0.12	0.12		0.25	0.09		
BDL238	10952.3	Av	1.3	1.29		1.25	1.32		
BDL238	10953.1	P			0.37		0.37		
BDL238	10953.1	Av			1.14		1.15		
BDL238	10954.2	P		0.52		0.61	0.5		
BDL238	10954.2	Av		1.13		1.11	1.12		
BDL238	10954.3	P							0.67

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL238	10954.3	Av							1.12
BDL240	10802.2	P					0.47		
BDL240	10802.2	Av					1.1		
BDL240	10806.6	P	0.33	0.43		0.27	0.19		
BDL240	10806.6	Av	1.28	1.19		1.2	1.18		
BDL240	10802.2	P	0.38				0.1		
BDL240	10802.2	Av	1.17				1.24		
BDL240	10803.5	P			0.48		0.17		
BDL240	10803.5	Av			1.12		1.18		
BDL240	10806.6	P	0.01	0.03	0.05	<0.01	0.01	0.15	0.26
BDL240	10806.6	Av	2.85	2.11	1.38	1.97	1.44	1.32	1.34
BDL241	10875.1	P	0.1				0.46		
BDL241	10875.1	Av	1.25				1.13		
BDL241	10873.1	P					0.22		0.61
BDL241	10873.1	Av					1.19		1.11
BDL241	10874.2	P					0.45		
BDL241	10874.2	Av					1.11		
BDL241	10874.3	P	0.54	0.05	0.01	0.03	0.05	0.08	0.01
BDL241	10874.3	Av	1.1	1.43	1.47	1.53	1.32	1.21	1.55
BDL241	10875.1	P	0.09	0.53					
BDL241	10875.1	Av	1.36	1.13					
BDL242	10731.3	P	<0.01	0.2		0.19	0.12		0.2
BDL242	10731.3	Av	1.38	1.21		1.19	1.21		1.22
BDL242	10731.5	P	0.1	0.31		0.32			
BDL242	10731.5	Av	1.34	1.16		1.15			
BDL242	10737.1	P					0.32		
BDL242	10737.1	Av					1.14		
BDL242	10731.3	P	<0.01	0.01	<0.01	<0.01	<0.01	0.05	0.04
BDL242	10731.3	Av	1.75	1.62	2.15	1.62	1.39	2.12	2.23
BDL242	10731.5	P	0.06						
BDL242	10731.5	Av	1.29						
BDL242	10731.7	P	<0.01	<0.01	0.01	0.01	0.03	0.01	0.04
BDL242	10731.7	Av	1.85	1.49	1.45	1.47	1.28	1.64	1.86
BDL242	10737.2	P	0.47	0.23	<0.01	0.07	0.05	<0.01	0.01
BDL242	10737.2	Av	1.26	1.38	1.86	1.43	1.32	1.91	2.05
BDL245	10811.2	P	0.54	0.17		0.08	0.21		0.3
BDL245	10811.2	Av	1.1	1.22		1.28	1.16		1.14
BDL245	10813.3	P					0.09		
BDL245	10813.3	Av					1.22		
BDL245	10816.3	P					0.14		
BDL245	10816.3	Av					1.18		
BDL247	10912.1	P		0.08		0.02	0.01		
BDL247	10912.1	Av		1.31		1.4	1.37		
BDL247	10912.2	P					0.08		
BDL247	10912.2	Av					1.22		
BDL247	10915.1	P		0.45	0.22	0.14	0.02		0.53
BDL247	10915.1	Av		1.19	1.2	1.28	1.4		1.12
BDL247	10911.1	P	0.15	0.1	0.01	0.01	0.02	0.3	0.23
BDL247	10911.1	Av	1.96	1.8	1.6	1.79	1.45	1.38	1.5
BDL247	10912.1	P	0.32	0.4	0.24	0.25	0.13	0.32	0.14
BDL247	10912.1	Av	1.15	1.22	1.2	1.24	1.21	1.27	1.56

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL247	10912.2	P	0.04	0.01	0.03	<0.01	<0.01	0.09	0.3
BDL247	10912.2	Av	1.73	1.74	1.36	1.81	1.62	1.41	1.35
BDL247	10912.6	P		0.61		0.57	0.39	0.21	0.48
BDL247	10912.6	Av		1.11		1.11	1.13	1.37	1.3
BDL247	10915.1	P	0.23	0.14	<0.01	0.03	0.08	0.02	0.06
BDL247	10915.1	Av	1.47	1.52	1.81	1.54	1.3	1.74	1.58
BDL248	11051.2	P	<0.01	0.01	0.03	0.02	<0.01	0.21	
BDL248	11051.2	Av	1.61	1.45	1.36	1.43	1.42	1.22	
BDL248	11052.2	P	<0.01	<0.01	<0.01	<0.01	<0.01	0.04	0.01
BDL248	11052.2	Av	1.89	1.68	1.89	1.7	1.44	1.74	1.81
BDL248	11053.3	P	<0.01	<0.01	0.07	<0.01	0.03	0.46	0.65
BDL248	11053.3	Av	2.04	1.62	1.29	1.57	1.34	1.13	1.14
BDL248	11054.3	P	0.15	0.15		0.24	0.04		
BDL248	11054.3	Av	1.18	1.18		1.19	1.29		
BDL248	11051.2	P					0.18		
BDL248	11051.2	Av					1.1		
BDL248	11054.2	P					0.18		
BDL248	11054.2	Av					1.1		
BDL249	11401.2	P	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01	<0.01
BDL249	11401.2	Av	3.19	2.43	2.68	2.35	1.67	2.55	3.02
BDL249	11401.5	P	<0.01	0.01	0.02	<0.01	0.02	0.39	0.44
BDL249	11401.5	Av	2.63	1.94	1.4	1.83	1.3	1.19	1.23
BDL249	11402.4	P	<0.01	<0.01	0.03	<0.01	<0.01	0.03	0.1
BDL249	11402.4	Av	2.42	1.99	1.38	2.02	1.65	1.37	1.43
BDL249	11403.2	P		0.46	0.34	0.27	0.2	0.56	
BDL249	11403.2	Av		1.22	1.17	1.25	1.21	1.11	
BDL249	11404.3	P	0.01	0.07		0.23	0.22		
BDL249	11404.3	Av	1.43	1.22		1.19	1.19		
BDL249	11401.5	P		0.31		0.45			
BDL249	11401.5	Av		1.1		1.11			
BDL249	11404.3	P	0.03	0.22		0.2	0.04		
BDL249	11404.3	Av	1.33	1.18		1.18	1.23		
BDL250	10842.3	P	<0.01	0.01	0.21	<0.01	0.02		0.34
BDL250	10842.3	Av	1.69	1.54	1.15	1.56	1.29		1.24
BDL250	10846.2	P	0.01	<0.01	0.29	<0.01	0.05	0.4	0.21
BDL250	10846.2	Av	1.43	1.41	1.12	1.45	1.25	1.19	1.33
BDL250	10846.3	P	0.02	0.36			0.05		
BDL250	10846.3	Av	1.37	1.11			1.26		
BDL250	10841.3	P	0.01	0.02	0.17	<0.01	<0.01		
BDL250	10841.3	Av	1.99	1.66	1.22	1.61	1.62		
BDL250	10842.3	P	0.23	0.52	0.12	0.44		0.41	0.59
BDL250	10842.3	Av	1.57	1.28	1.45	1.22		1.6	1.27
BDL250	10843.2	P	0.03	0.07	0.01	0.01	0.01	0.02	0.1
BDL250	10843.2	Av	1.85	1.67	1.55	1.62	1.37	1.53	1.6
BDL250	10846.2	P	0.04	<0.01	<0.01	<0.01	<0.01	0.45	0.67
BDL250	10846.2	Av	1.46	1.68	1.58	1.7	1.43	1.22	1.15
BDL250	10846.3	P	<0.01	<0.01	<0.01	<0.01	<0.01	0.05	0.03
BDL250	10846.3	Av	2.79	2.32	1.76	2.28	1.84	1.8	1.87
BDL252	10881.1	P	0.01	<0.01	<0.01	<0.01	<0.01	0.09	0.19
BDL252	10881.1	Av	2.1	1.87	1.68	1.9	1.48	1.43	1.68
BDL252	10882.1	P	0.01	0.01	<0.01	<0.01	<0.01	0.18	0.18

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL252	10882.1	Av	3.11	2.51	1.97	2.43	1.82	1.4	1.51
BDL252	10882.2	P	0.05	<0.01	0.04	<0.01	0.01		0.62
BDL252	10882.2	Av	1.87	1.74	1.35	1.72	1.44		1.11
BDL252	10882.4	P	0.01	0.01	0.17	0.01	0.38		
BDL252	10882.4	Av	2.06	1.62	1.24	1.52	1.13		
BDL252	10884.1	P			0.5			0.24	0.24
BDL252	10884.1	Av			1.1			1.22	1.3
BDL58	10281.5	P			0.07	0.46	0.15	0.55	0.02
BDL58	10281.5	Av			1.32	1.15	1.18	1.14	1.37
BDL58	10282.3	P	<0.01	0.01	<0.01	<0.01	<0.01	<0.01	<0.01
BDL58	10282.3	Av	1.94	2.18	1.77	2.29	1.81	1.79	1.83
BDL58	10285.3	P					0.06		
BDL58	10285.3	Av					1.21		
BDL62	10682.1	P		0.31	0.43	0.12			0.4
BDL62	10682.1	Av		1.35	1.19	1.46			1.33
BDL62	10684.2	P		0.43		0.33	0.08		0.58
BDL62	10684.2	Av		1.17		1.23	1.26		1.11
BDL62	10684.5	P			0.41	0.64	0.31	0.67	0.64
BDL62	10684.5	Av			1.14	1.11	1.15	1.13	1.13
BDL64	10651.5	P			0.02	0.39	0.07	0.17	0.26
BDL64	10651.5	Av			1.77	1.14	1.19	1.54	1.52
BDL64	10653.1	P		0.14	<0.01	0.13		<0.01	0.03
BDL64	10653.1	Av		1.21	1.62	1.24		1.42	1.51
BDL64	10653.3	P					0.01		
BDL64	10653.3	Av					1.25		
BDL64	10654.3	P	0.15	0.23	0.06	0.02	<0.01	0.37	0.27
BDL64	10654.3	Av	1.53	1.6	1.5	1.71	1.55	1.45	1.44
BDL64	10651.1	P			0.5			0.31	0.16
BDL64	10651.1	Av			1.13			1.45	1.91
BDL64	10651.2	P							0.62
BDL64	10651.2	Av							1.13
BDL64	10651.5	P							0.48
BDL64	10651.5	Av							1.15
BDL64	10653.1	P	0.01	0.02	0.01	<0.01	0.02	0.05	0.03
BDL64	10653.1	Av	2.18	2.28	1.69	2.43	1.25	2.27	3.13
BDL64	10654.3	P	<0.01	<0.01	0.02	<0.01	0.03	0.01	<0.01
BDL64	10654.3	Av	1.78	1.9	1.46	1.97	1.2	1.7	2.53
BDL64	10651.1	P	0.11						
BDL64	10651.1	Av	1.27						
BDL64	10651.3	P	0.14	0.32		0.46			
BDL64	10651.3	Av	1.43	1.22		1.14			
BDL79	11042.3	P					0.5		
BDL79	11042.3	Av					1.1		
BDL79	11042.3	P		0.44		0.28	0.04		
BDL79	11042.3	Av		1.18		1.26	1.36		
BDL79	11042.7	P			0.4				0.55
BDL79	11042.7	Av			1.12				1.11
BDL79	11043.1	P		0.27	0.03	0.25	0.03	0.29	0.22
BDL79	11043.1	Av		1.28	1.35	1.3	1.35	1.18	1.22
BDL85	10411.1	P	0.74	0.55		0.42			
BDL85	10411.1	Av	1.1	1.14		1.16			

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL85	10411.3	P	0.14	0.01		0.03	0.17		
BDL85	10411.3	Av	1.24	1.28		1.32	1.13		
BDL85	10412.2	P			0.44			0.06	0.15
BDL85	10412.2	Av			1.1			1.22	1.24
BDL85	10414.1	P		0.01	0.05	0.02	0.08	0.38	0.47
BDL85	10414.1	Av		1.3	1.29	1.39	1.18	1.17	1.11
BDL85	10414.2	P		0.27	<0.01	0.07	0.07	<0.01	0.08
BDL85	10414.2	Av		1.15	1.42	1.28	1.17	1.55	1.5
BDL88	10291.2	P	0.31	0.15	0.02	0.08	0.08	0.06	0.11
BDL88	10291.2	Av	1.26	1.33	1.45	1.36	1.24	1.35	1.36
BDL88	10291.4	P	0.01	0.02	<0.01	<0.01	<0.01	0.23	0.25
BDL88	10291.4	Av	1.72	1.57	1.54	1.62	1.42	1.44	1.35
BDL88	10291.5	P	0.59	0.44	0.48	0.38			
BDL88	10291.5	Av	1.13	1.17	1.13	1.17			
BDL88	10293.3	P	0.14	0.28	<0.01	0.11	0.26	0.16	0.08
BDL88	10293.3	Av	1.34	1.35	2.15	1.36	1.19	1.58	1.97
BDL88	10291.2	P	0.16	0.05	<0.01	0.01	0.21	0.33	0.47
BDL88	10291.2	Av	1.32	1.45	1.56	1.48	1.18	1.17	1.16
BDL88	10291.4	P			<0.01				
BDL88	10291.4	Av			1.49				
BDL88	10291.5	P			<0.01			0.15	0.38
BDL88	10291.5	Av			1.84			1.25	1.22
BDL88	10293.3	P			0.02				0.47
BDL88	10293.3	Av			1.23				1.15
BDL88	10294.2	P			<0.01			0.3	0.55
BDL88	10294.2	Av			1.76			1.23	1.16
BDL90	10921.3	P					0.22		
BDL90	10921.3	Av					1.18		
BDL90	10921.6	P					0.09		
BDL90	10921.6	Av					1.28		
BDL90	10924.2	P	0.06	0.04		<0.01	<0.01		
BDL90	10924.2	Av	2	1.75		1.73	1.5		
BDL90	10924.4	P					0.33		
BDL90	10924.4	Av					1.12		
BDL90	10925.4	P	0.02	0.03		<0.01	<0.01		
BDL90	10925.4	Av	2.27	1.81		1.76	1.46		
BDL90	10923.4	P					0.27		
BDL90	10923.4	Av					1.17		
BDL90	10924.2	P	0.06	0.28	0.22	0.33	0.06		
BDL90	10924.2	Av	1.43	1.26	1.19	1.22	1.3		
BDL90	10925.4	P	0.02	0.04		0.03	0.01		
BDL90	10925.4	Av	1.7	1.58		1.54	1.43		
BDL90	10921.6	P		0.75	0.04	0.42	0.02		0.13
BDL90	10921.6	Av		1.11	1.32	1.24	1.41		1.28
BDL90	10923.4	P				0.51	0.12		0.58
BDL90	10923.4	Av				1.15	1.25		1.15
BDL90	10924.2	P	0.16	0.01	0.28	0.01	<0.01		
BDL90	10924.2	Av	1.4	1.65	1.18	1.7	1.54		
BDL90	10925.4	P	<0.01	<0.01	<0.01	<0.01	<0.01	0.05	0.03
BDL90	10925.4	Av	2.34	2.48	1.88	2.4	1.61	1.55	1.78
BDL94	11721.4	P			0.45			0.09	0.26

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>Roots Coverage TP2</i>	<i>Roots Coverage TP3</i>	<i>RGR Of Leaf Area</i>	<i>RGR Of Root Coverage</i>	<i>RGR Of Roots Length</i>	<i>Fresh Weight</i>	<i>Dry Weight</i>
BDL94	11721.4	Av			1.11			1.29	1.16
BDI.94	11725.3	P						0.17	
BDL94	11725.3	Av						1.2	
BDL94	11725.3	P						0.31	
BDL94	11725.3	Av						1.29	

Table 22. "P" = P-value; "Av" = ratio between the averages of event and control. Note that when the average ratio is higher than "1" the effect of exogenous expression of the gene is an increase of the desired trait; "Par" = Parameter according to the measured parameters; "Ev" = event. TP1 = Time point 1; TP2 = Time point 2; TP3 = Time point 3; RGR = relative growth rate.

5

Greenhouse assays - Table 23 specifies the parameters that were measured in the greenhouse assays and which are presented in Tables 24, 25, 26 and 27. In cases where a certain event appears more than once, the event was tested in several independent experiments. The parameters were measured as follows:

10 The plants were analyzed for their overall size, growth rate, flowering, seed yield, weight of 1,000 seeds, dry matter and harvest index (HI- seed yield/dry matter). Transgenic plants performance was compared to control plants grown in parallel under the same conditions. Mock- transgenic plants expressing the uidA reporter gene (GUS-Intron) or with no gene at all, under the same promoter were used as control.

15 The experiment was planned in nested randomized plot distribution. For each gene of the invention three to five independent transformation events were analyzed from each construct.

20 **Digital imaging** - A laboratory image acquisition system, which consists of a digital reflex camera (Canon EOS 300D) attached with a 55 mm focal length lens (Canon EF-S series), mounted on a reproduction device (Kaiser RS), which included 4 light units (4 x 150 Watts light bulb) is used for capturing images of plant samples.

25 The image capturing process was repeated every 2 days starting from day 1 after transplanting till day 16. Same camera, placed in a custom made iron mount, was used for capturing images of larger plants sawn in white tubs in an environmental controlled greenhouse. The tubs were square shape include 1.7 liter trays. During the capture process, the tubs were placed beneath the iron mount, while avoiding direct sun light and casting of shadows.

An image analysis system was used, which consists of a personal desktop computer (Intel P4 3.0 GHz processor) and a public domain program - ImageJ 1.39

(Java based image processing program which was developed at the U.S National Institutes of Health and is freely available. Images were captured in resolution of 10 Mega Pixels (3888 x 2592 pixels) and stored in a low compression JPEG (Joint Photographic Experts Group standard) format. Next, analyzed data was saved to text files and processed using the JMP statistical analysis software (SAS institute).

Leaf growth analysis - Using the digital analysis leaves data was calculated, including leaf number, rosette area, rosette diameter, leaf blade area, plot coverage, leaf petiole length.

The vegetative growth rate of the plant was defined by formulas IX, X, XI and XII.

Formula IX:

Relative growth rate of leaf blade area = Regression coefficient of leaf area along time course.

Formula X:

Relative growth rate of rosette area = Regression coefficient of rosette area along time course.

Formula XI

Relative growth rate of rosette diameter = Regression coefficient of rosette diameter along time course.

Formula XII

Relative growth rate of plot coverage = Regression coefficient of plot coverage along time course.

Seeds average weight (Seed weight or 1000 seed weight) - At the end of the experiment all seeds were collected. The seeds were scattered on a glass tray and a picture was taken. Using the digital analysis, the number of seeds in each sample was calculated.

Plant dry weight and seed yield - On about day 80 from sowing, the plants were harvested and left to dry at 30 °C in a drying chamber. The biomass and seed weight of each plot were measured and divided by the number of plants in each plot. Dry weight = total weight of the vegetative portion above ground (excluding roots) after drying at 30 °C in a drying chamber;

145

Seed yield per plant = total seed weight per plant (gr.).

1000 seed weight (the weight of 1000 seeds) (gr.).

The harvest index was calculated using Formula IV (Harvest Index = Average seed yield per plant/ Average dry weight) as described above.

5 ***Oil percentage in seeds*** - At the end of the experiment all seeds from plots A-C were collected. Columbia seeds from 3 plots were mixed grounded and then mounted onto the extraction chamber. 210 ml of n-Hexane (Cat No. 080951 Biolab Ltd.) were used as the solvent. The extraction was performed for 30 hours at medium heat 50 °C. Once the extraction has ended the n-Hexane was evaporated using the evaporator at 35
10 °C and vacuum conditions. The process was repeated twice. The information gained from the Soxhlet extractor (Soxhlet, F. Die gewichtsanalytische Bestimmung des Milchfettes, Polytechnisches J. (Dingler's) 1879, 232, 461) was used to create a calibration curve for the Low Resonance NMR. The content of oil of all seed samples was determined using the Low Resonance NMR (MARAN Ultra– Oxford Instrument)
15 and its MultiQuant software package.

Oil yield - The oil yield was calculated using Formula VII (described above).

Silique length analysis - On day 50 from sowing, 30 siliques from different plants in each plot were sampled in block A. The chosen siliques were green-yellow in color and were collected from the bottom parts of a grown plant's stem. A digital
20 photograph was taken to determine silique's length.

Statistical analyses - To identify genes conferring significantly improved tolerance to abiotic stresses, the results obtained from the transgenic plants were compared to those obtained from control plants. To identify outperforming genes and constructs, results from the independent transformation events tested were analyzed
25 separately. Data was analyzed using Student's t-test and results were considered significant if the p value was less than 0.1. The JMP statistics software package was used (Version 5.2.1, SAS Institute Inc., Cary, NC, USA).

30

Table 23
Parameters measured in greenhouse assays

<i>Parameter</i>	<i>Number</i>
Rosette Diameter TP2	1
Rosette Diameter TP3	2
Rosette Diameter TP4	3
Rosette Area TP2	4
Rosette Area TP3	5
Rosette Area TP4	6
Plot Coverage TP2	7
Plot Coverage TP3	8
Plot Coverage TP4	9
Leaf Number TP2	10
Leaf Number TP3	11
Leaf Number TP4	12
Leaf Blade Area TP2	13
Leaf Blade Area TP3	14
Leaf Blade Area TP4	15
Leaf Petiole Length TP2	16
Leaf Petiole Length TP3	17
Leaf Petiole Length TP4	18
Blade Relative Area TP2	19
Blade Relative Area TP3	20
Blade Relative Area TP4	21
Petiole Relative Area TP2	22
Petiole Relative Area TP3	23
Petiole Relative Area TP4	24
RGR Of Leaf Blade Area	25
RGR Of Leaf Number	26
RGR Of Rosette Area	27
RGR Of Rosette Diameter	28
RGR Of Plot Coverage	29
Dry Weight	30
Fresh Weight	31
Inflorescence Emergence	32
Flowering	33
Seed Yield	34
Harvest Index	35
Seeds Weight	36
Oil Content	37

5 Table 23. Provided are the parameters measured in greenhouse experiments which are presented in Tables 24-27 hereinbelow. TP1 = Time point 1; TP2 = Time point 2; TP3 = Time point 3; RGR = relative growth rate.

Table 24
Results from greenhouse experiments

10

<i>Gene</i>	<i>Ev.</i>	<i>Par:</i>	<i>1</i>	<i>2</i>	<i>3</i>	<i>4</i>	<i>5</i>	<i>6</i>	<i>7</i>	<i>8</i>	<i>9</i>	<i>10</i>
BDL102	10471.1	P	<0.01	<0.01		0.13	0.34	0.03	0.13	0.34	0.03	
BDL102	10471.1	Av	1.24	1.14		1.31	1.21	1.15	1.31	1.21	1.15	
BDL102	10472.1	P	0.13	0.11	0.33	0.25	0.02	0.32	0.25	0.02	0.32	
BDL102	10472.1	Av	1.26	1.19	1.14	1.37	1.26	1.26	1.37	1.26	1.26	
BDL102	10474.1	P	0.22	0.23	0.26	0.19	0.2	0.23	0.19	0.2	0.23	0.19

<i>Gene</i>	<i>Ev.</i>	<i>Par:</i>	<i>1</i>	<i>2</i>	<i>3</i>	<i>4</i>	<i>5</i>	<i>6</i>	<i>7</i>	<i>8</i>	<i>9</i>	<i>10</i>
BDL102	10474.1	Av	1.35	1.33	1.2	1.65	1.87	1.49	1.65	1.87	1.49	1.1
BDL102	10474.2	P	0.17	0.02	0.01	0.1	0.05	<0.01	0.1	0.05	<0.01	<0.01
BDL102	10474.2	Av	1.3	1.23	1.11	1.59	1.53	1.27	1.59	1.53	1.27	1.11
BDL102	10474.6	P	0.02			0.23			0.23			
BDL102	10474.6	Av	1.11			1.15			1.15			
BDL117	10071.2	P				0.62			0.59			
BDL117	10071.2	Av				1.14			1.16			
BDL117	10074.1	P	0.28	0.22	0.16	0.29	0.3	0.3	0.28	0.29	0.29	0.34
BDL117	10074.1	Av	1.26	1.23	1.24	1.58	1.47	1.44	1.6	1.49	1.46	1.15
BDL117	10074.4	P	0.35	0.34	0.41	0.38	0.39	0.41	0.38	0.37	0.39	0.28
BDL117	10074.4	Av	1.21	1.14	1.15	1.41	1.29	1.26	1.43	1.3	1.28	1.13
BDL117	10073.1	P				0.57	0.58	0.64				
BDL117	10073.1	Av				1.13	1.17	1.14				
BDL117	10073.2	P			0.45	0.7	0.55	0.48	0.7	0.55	0.48	
BDL117	10073.2	Av			1.14	1.1	1.21	1.22	1.1	1.21	1.22	
BDL117	10074.1	P	0.41	0.4	0.39	0.44	0.37	0.38	0.44	0.37	0.38	0.13
BDL117	10074.1	Av	1.1	1.15	1.15	1.17	1.32	1.29	1.17	1.32	1.29	1.15
BDL117	10074.4	P			0.1		0.4	0.34		0.4	0.34	
BDL117	10074.4	Av			1.09		1.1	1.13		1.1	1.13	
BDL138	9812.1	P	0.04									
BDL138	9812.1	Av	1.06									
BDL138	9812.3	P				0.06			0.04			
BDL138	9812.3	Av				1.18			1.19			
BDL140	10423.1	P	0.02			0.01	0.22		0.01	0.22		
BDL140	10423.1	Av	1.16			1.39	1.22		1.39	1.22		
BDL140	10424.4	P				0.62			0.62			0.1
BDL140	10424.4	Av				1.18			1.18			1.08
BDL147	10301.5	P										0.1
BDL147	10301.5	Av										1.08
BDL147	10303.1	P	<0.01	0.01		0.06	0.04	0.08	0.06	0.04	0.08	0.05
BDL147	10303.1	Av	1.23	1.21		1.65	1.36	1.24	1.65	1.36	1.24	1.11
BDL147	10303.6	P				0.63	0.4		0.63	0.4		
BDL147	10303.6	Av				1.14	1.11		1.14	1.11		
BDL147	10304.2	P				0.33			0.33			
BDL147	10304.2	Av				1.19			1.19			
BDL147	10304.2	P					0.44	0.36		0.44	0.36	
BDL147	10304.2	Av					1.17	1.11		1.17	1.11	
BDL149	9823.3	P	0.22	0.08		0.33	0.13	0.29		0.03		
BDL149	9823.3	Av	1.14	1.1		1.14	1.18	1.14		1.11		
BDL149	9824.4	P	0.01	<0.01	<0.01	0.08	<0.01	0.04	0.09	<0.01	0.04	0.03
BDL149	9824.4	Av	1.16	1.13	1.13	1.35	1.26	1.29	1.37	1.28	1.31	1.05
BDL149	9823.3	P				0.65	0.67	0.63	0.65	0.67	0.63	
BDL149	9823.3	Av				1.17	1.1	1.13	1.17	1.1	1.13	
BDL152	10431.1	P				0.28			0.28			
BDL152	10431.1	Av				1.13			1.13			
BDL152	10431.4	P					0.2	0.45		0.2	0.45	
BDL152	10431.4	Av					1.12	1.11		1.12	1.11	
BDL152	10434.4	P			0.22	0.72		0.41	0.72		0.41	0.09
BDL152	10434.4	Av			1.12	1.12		1.2	1.12		1.2	1.07

<i>Gene</i>	<i>Ev.</i>	<i>Par:</i>	<i>1</i>	<i>2</i>	<i>3</i>	<i>4</i>	<i>5</i>	<i>6</i>	<i>7</i>	<i>8</i>	<i>9</i>	<i>10</i>
BDL153	10141.3	P	<0.01	<0.01	<0.01	0.04	<0.01	<0.01	0.05	<0.01	<0.01	
BDL153	10141.3	Av	1.21	1.17	1.17	1.41	1.33	1.34	1.43	1.34	1.36	
BDL153	10142.2	P	0.21	0.41	0.38	0.33	0.38	0.38	0.32	0.37	0.38	0.49
BDL153	10142.2	Av	1.33	1.2	1.25	1.64	1.55	1.57	1.66	1.58	1.6	1.15
BDL153	10143.1	P	0.03			0.14	0.07	0.37	0.14	0.04	0.34	
BDL153	10143.1	Av	1.07			1.22	1.12	1.14	1.23	1.13	1.16	
BDL153	10144.1	P				0.08	0.08	0.1	0.03	0.03	0.04	
BDL153	10144.1	Av				1.11	1.1	1.11	1.13	1.11	1.13	
BDL153	10141.3	P	0.1	<0.01	0.01			0.01			0.01	
BDL153	10141.3	Av	1.14	1.15	1.06			1.14			1.14	
BDL153	10142.2	P	0.01	0.01	0.14	0.15	0.44	0.14	0.15	0.44	0.14	
BDL153	10142.2	Av	1.17	1.16	1.16	1.18	1.19	1.31	1.18	1.19	1.31	
BDL153	10142.3	P						0.4			0.4	
BDL153	10142.3	Av						1.15			1.15	
BDL153	10143.1	P										0.38
BDL153	10143.1	Av										1.12
BDL153	10143.2	P					0.25	0.11		0.25	0.11	
BDL153	10143.2	Av					1.11	1.12		1.11	1.12	
BDL153	10144.1	P		0.3	0.17		0.39	0.34		0.39	0.34	
BDL153	10144.1	Av		1.23	1.1		1.44	1.13		1.44	1.13	
BDL154	10703.8	P	<0.01	0.03		0.12	0.07	0.43	0.12	0.07	0.43	
BDL154	10703.8	Av	1.29	1.2		1.53	1.37	1.13	1.53	1.37	1.13	
BDL155	9991.5	P	0.22	0.07	0.1	0.31		0.31	0.31		0.31	0.03
BDL155	9991.5	Av	1.13	1.12	1.12	1.14		1.12	1.14		1.12	1.09
BDL155	9994.3	P	0.17	0.02	0.08	0.16	0.06	0.17	0.16	0.06	0.17	0.09
BDL155	9994.3	Av	1.15	1.17	1.12	1.17	1.27	1.21	1.17	1.27	1.21	1.07
BDL162	10492.2	P			0.32	0.16	0.25	0.11	0.16	0.25	0.11	0.59
BDL162	10492.2	Av			1.13	1.22	1.15	1.34	1.22	1.15	1.34	1.11
BDL167	10044.2	P					0.17			0.6		
BDL167	10044.2	Av					1.17			1.1		
BDL167	10043.3	P					0.39			0.39		
BDL167	10043.3	Av					1.1			1.1		
BDL167	10044.2	P				0.21	0.17	0.31	0.21	0.17	0.31	0.03
BDL167	10044.2	Av				1.14	1.15	1.2	1.14	1.15	1.2	1.09
BDL168	9881.3	P	0.1	0.11	0.09	0.18	0.14	0.09	0.18	0.14	0.1	0.22
BDL168	9881.3	Av	1.22	1.19	1.24	1.48	1.44	1.49	1.5	1.46	1.52	1.14
BDL168	9881.4	P	0.14	0.24	0.18	0.05	0.11	0.27	0.05	0.11	0.26	0.4
BDL168	9881.4	Av	1.23	1.17	1.19	1.43	1.38	1.34	1.45	1.4	1.35	1.1
BDL168	9882.1	P				0.04	0.05	0.12	0.01	0.02	0.05	
BDL168	9882.1	Av				1.14	1.12	1.1	1.15	1.13	1.11	
BDL168	9882.3	P	0.58	0.64	0.49	0.51	0.55	0.51	0.49	0.53	0.5	0.54
BDL168	9882.3	Av	1.1	1.11	1.17	1.3	1.24	1.33	1.32	1.26	1.35	1.1
BDL168	9884.4	P			0.72			0.76			0.74	
BDL168	9884.4	Av			1.11			1.13			1.15	
BDL168	9881.4	P		0.02				0.51			0.51	
BDL168	9881.4	Av		1.09				1.15			1.15	
BDL168	9882.1	P		0.04			0.15	0.22				
BDL168	9882.1	Av		1.07			1.12	1.13				
BDL168	9884.1	P			0.01							

<i>Gene</i>	<i>Ev.</i>	<i>Par:</i>	<i>1</i>	<i>2</i>	<i>3</i>	<i>4</i>	<i>5</i>	<i>6</i>	<i>7</i>	<i>8</i>	<i>9</i>	<i>10</i>
BDL168	9884.1	Av			1.1							
BDL169	10743.4	P				0.43	0.55	0.56	0.54	0.7	0.72	
BDL169	10743.4	Av				1.27	1.2	1.19	1.17	1.11	1.1	
BDL169	10744.1	P	0.41			0.37	0.55		0.3	0.45	0.69	0.43
BDL169	10744.1	Av	1.1			1.27	1.18		1.35	1.25	1.1	1.13
BDL169	10747.1	P	0.29			0.47	0.64		0.42	0.54		<0.01
BDL169	10747.1	Av	1.15			1.34	1.15		1.42	1.22		1.17
BDL169	10747.5	P		0.08		0.01	0.02	0.24	0.01	0.01	0.13	<0.01
BDL169	10747.5	Av		1.09		1.4	1.28	1.11	1.49	1.36	1.18	1.16
BDL169	10741.3	P	0.58			0.43			0.43			
BDL169	10741.3	Av	1.13			1.2			1.2			
BDL169	10744.2	P	0.43	0.25	0.18	0.34	0.31	0.33	0.34	0.31	0.33	0.44
BDL169	10744.2	Av	1.24	1.26	1.21	1.6	1.61	1.42	1.6	1.61	1.42	1.12
BDL169	10747.1	P	0.01	0.14		0.33	0.09	0.53	0.33	0.09	0.53	
BDL169	10747.1	Av	1.16	1.15		1.44	1.3	1.17	1.44	1.3	1.17	
BDL169	10747.5	P	0.05	0.1	0.01	0.16	0.19	0.05	0.16	0.19	0.05	
BDL169	10747.5	Av	1.13	1.08	1.09	1.28	1.24	1.12	1.28	1.24	1.12	
BDL171	10661.2	P	0.7		0.44	0.59	0.7	0.56	0.59	0.7	0.56	
BDL171	10661.2	Av	1.1		1.1	1.23	1.17	1.22	1.23	1.17	1.22	
BDL171	10662.3	P	0.48	0.44		0.35	0.53		0.35	0.53		
BDL171	10662.3	Av	1.12	1.13		1.26	1.19		1.26	1.19		
BDL171	10664.1	P	0.01	<0.01	0.24	<0.01	0.01	0.42	<0.01	0.01	0.42	<0.01
BDL171	10664.1	Av	1.36	1.31	1.23	1.72	1.56	1.49	1.72	1.56	1.49	1.21
BDL171	10664.3	P	0.16	0.16	0.38	0.24	0.15	0.42	0.24	0.15	0.42	0.03
BDL171	10664.3	Av	1.26	1.22	1.14	1.49	1.44	1.36	1.49	1.44	1.36	1.19
BDL171	10661.5	P							0.29	0.38		0.42
BDL171	10661.5	Av							1.15	1.1		1.12
BDL171	10662.2	P	0.01	0.07		0.18			0.11			0.15
BDL171	10662.2	Av	1.16	1.1		1.2			1.28			1.18
BDL171	10662.3	P	0.02	<0.01	<0.01	<0.01	<0.01	0.01	<0.01	<0.01	<0.01	<0.01
BDL171	10662.3	Av	1.28	1.29	1.2	1.57	1.52	1.33	1.67	1.61	1.41	1.25
BDL171	10663.3	P	<0.01	<0.01		<0.01	0.04		<0.01	0.02		0.03
BDL171	10663.3	Av	1.26	1.18		1.32	1.21		1.4	1.28		1.18
BDL171	10664.1	P				0.76	0.81		0.69	0.75	0.85	
BDL171	10664.1	Av				1.19	1.14		1.26	1.21	1.11	
BDL171	10664.3	P	0.01	0.12		0.14	0.17	0.58	0.11	0.12	0.47	0.07
BDL171	10664.3	Av	1.23	1.15		1.44	1.3	1.16	1.52	1.37	1.23	1.25
BDL173	9952.1	P						0.67			0.64	
BDL173	9952.1	Av						1.11			1.13	
BDL177	10521.3	P				0.56			0.56			
BDL177	10521.3	Av				1.15			1.15			
BDL177	10524.2	P				0.09		0.44	0.09		0.44	
BDL177	10524.2	Av				1.2		1.1	1.2		1.1	
BDL182	10691.2	P					0.27			0.27		0.46
BDL182	10691.2	Av					1.12			1.12		1.11
BDL182	10692.3	P	0.44			0.43	0.37		0.43	0.37		0.05

<i>Gene</i>	<i>Ev.</i>	<i>Par:</i>	<i>1</i>	<i>2</i>	<i>3</i>	<i>4</i>	<i>5</i>	<i>6</i>	<i>7</i>	<i>8</i>	<i>9</i>	<i>10</i>
BDL182	10692.3	Av	1.11			1.25	1.12		1.25	1.12		1.08
BDL182	10693.2	P					0.65			0.65		
BDL182	10693.2	Av					1.1			1.1		
BDL182	10693.3	P	0.03	0.08		0.16	0.06	0.38	0.16	0.06	0.38	
BDL182	10693.3	Av	1.19	1.15		1.22	1.18	1.12	1.22	1.18	1.12	
BDL182	10693.5	P				0.48			0.48			
BDL182	10693.5	Av				1.1			1.1			
BDL182	10691.4	P	0.51	0.43		0.39	0.44		0.47	0.6		0.32
BDL182	10691.4	Av	1.11	1.11		1.36	1.21		1.26	1.13		1.19
BDL182	10691.8	P							0.87			
BDL182	10691.8	Av							1.1			
BDL183	9941.1	P						0.21	0.16		0.16	
BDL183	9941.1	Av						1.11	1.11		1.13	
BDL183	9943.4	P						0.3				
BDL183	9943.4	Av						1.1				
BDL183	9944.1	P	0.09	0.03		0.1	0.15	0.22	0.1	0.15	0.21	
BDL183	9944.1	Av	1.16	1.12		1.27	1.23	1.22	1.29	1.25	1.24	
BDL183	9941.1	P						0.65			0.65	
BDL183	9941.1	Av						1.11			1.11	
BDL183	9942.1	P	0.02	0.02		0.16	0.33	0.38	0.16	0.33	0.38	
BDL183	9942.1	Av	1.16	1.1		1.12	1.17	1.13	1.12	1.17	1.13	
BDL186	10002.2	P			0.1		0.05	<0.01		0.02	<0.01	
BDL186	10002.2	Av			1.07		1.13	1.25		1.14	1.27	
BDL186	10004.3	P			0.6			0.69			0.66	
BDL186	10004.3	Av			1.11			1.15			1.17	
BDL186	10001.3	P										0.53
BDL186	10001.3	Av										1.11
BDL186	10004.6	P				0.43			0.43			0.1
BDL186	10004.6	Av				1.2			1.2			1.08
BDL187	10502.2	P				0.29			0.6			
BDL187	10502.2	Av				1.2			1.13			
BDL187	10503.1	P				0.09	0.27		0.09	0.27		
BDL187	10503.1	Av				1.26	1.1		1.26	1.1		
BDL187	10503.3	P				0.66			0.66			
BDL187	10503.3	Av				1.16			1.16			
BDL187	10503.5	P	0.26									
BDL187	10503.5	Av	1.1									
BDL188	10462.4	P				0.14	0.07		0.14	0.07		0.32
BDL188	10462.4	Av				1.13	1.09		1.13	1.09		1.13
BDL188	10462.1	P	0.02	0.11		0.04	0.11	0.4	0.04	0.11	0.4	
BDL188	10462.1	Av	1.2	1.14		1.3	1.27	1.21	1.3	1.27	1.21	
BDL188	10462.4	P	0.03	0.03	0.14		0.09	0.1		0.09	0.1	
BDL188	10462.4	Av	1.16	1.16	1.1		1.24	1.23		1.24	1.23	
BDL190	10232.2	P	0.01	0.01	<0.01	0.43	0.09	0.14	0.43	0.09	0.14	
BDL190	10232.2	Av	1.18	1.15	1.13	1.17	1.25	1.24	1.17	1.25	1.24	
BDL190	10233.2	P	0.03	<0.01	0.09	0.03	0.01	<0.01	0.03	0.01	<0.01	
BDL190	10233.2	Av	1.14	1.16	1.18	1.19	1.35	1.29	1.19	1.35	1.29	
BDL190	10233.4	P			0.11	0.18		<0.01	0.18		<0.01	
BDL190	10233.4	Av			1.11	1.11		1.17	1.11		1.17	

<i>Gene</i>	<i>Ev.</i>	<i>Par:</i>	<i>1</i>	<i>2</i>	<i>3</i>	<i>4</i>	<i>5</i>	<i>6</i>	<i>7</i>	<i>8</i>	<i>9</i>	<i>10</i>
BDL190	10234.2	P		0.09			0.05			0.05		
BDL190	10234.2	Av		1.06			1.18			1.18		
BDL192	9921.6	P				0.26	0.4	0.48	0.23	0.37	0.44	
BDL192	9921.6	Av				1.14	1.14	1.13	1.16	1.15	1.14	
BDL192	9922.1	P	0.11			0.35	0.33	0.28	0.33	0.3	0.22	
BDL192	9922.1	Av	1.1			1.21	1.14	1.1	1.23	1.15	1.12	
BDL192	9921.6	P				0.21	0.14	0.41	0.21	0.14	0.41	
BDL192	9921.6	Av				1.12	1.14	1.14	1.12	1.14	1.14	
BDL192	9922.5	P				0.35	0.33	0.3	0.65	0.62		
BDL192	9922.5	Av				1.18	1.2	1.13	1.11	1.13		
BDL193	10152.2	P				0.08			0.06	0.17		0.01
BDL193	10152.2	Av				1.15			1.16	1.1		1.06
BDL193	10153.2	P							0.6			
BDL193	10153.2	Av							1.11			
BDL193	10153.4	P			0.04	0.4	0.37	0.21	0.38	0.34	0.18	
BDL193	10153.4	Av			1.08	1.23	1.16	1.16	1.25	1.18	1.17	
BDL193	10153.3	P	0.41	0.14		0.4	0.01	0.4	0.4	0.01	0.4	0.33
BDL193	10153.3	Av	1.12	1.15		1.21	1.31	1.14	1.21	1.31	1.14	1.13
BDL193	10153.4	P	0.33	0.25		0.51	0.27	0.17	0.51	0.27	0.17	
BDL193	10153.4	Av	1.1	1.11		1.14	1.23	1.14	1.14	1.23	1.14	
BDL196	10243.1	P	0.61	0.66		0.6	0.66	0.66	0.6	0.66	0.66	
BDL196	10243.1	Av	1.14	1.1		1.2	1.16	1.15	1.2	1.16	1.15	
BDL196	10243.1	P			0.02							
BDL196	10243.1	Av			1.05							
BDL201	9961.3	P	0.26	0.22	0.38	0.11	0.23	0.44	0.11	0.23	0.44	0.09
BDL201	9961.3	Av	1.13	1.11	1.16	1.33	1.23	1.35	1.33	1.23	1.35	1.07
BDL220	10333.5	P			0.48							
BDL220	10333.5	Av			1.1							
BDL223	10793.5	P	0.09	0.01	0.01	0.39	0.12	0.26	0.39	0.12	0.26	0.02
BDL223	10793.5	Av	1.15	1.12	1.1	1.12	1.25	1.13	1.12	1.25	1.13	1.06
BDL223	10793.8	P	<0.01	0.01		0.2	0.19		0.2	0.19		
BDL223	10793.8	Av	1.17	1.1		1.31	1.18		1.31	1.18		
BDL224	10451.7	P				0.59			0.59			0.05
BDL224	10451.7	Av				1.12			1.12			1.08
BDL226	10861.2	P	0.05	0.05		0.01	0.07		0.01	0.04		0.03
BDL226	10861.2	Av	1.12	1.12		1.26	1.17		1.34	1.24		1.21
BDL226	10861.4	P							0.65			0.31
BDL226	10861.4	Av							1.13			1.12
BDL226	10864.2	P	0.01	0.01	0.09	<0.01	<0.01	0.07	<0.01	<0.01	0.04	0.22
BDL226	10864.2	Av	1.2	1.15	1.09	1.49	1.34	1.21	1.59	1.42	1.29	1.13
BDL227	11491.3	P	0.21	0.01	0.08	0.27	0.07		0.27	0.07		
BDL227	11491.3	Av	1.11	1.09	1.09	1.12	1.21		1.12	1.21		
BDL227	11492.3	P	<0.01	<0.01			0.01	0.05		0.01	0.05	
BDL227	11492.3	Av	1.17	1.12			1.23	1.12		1.23	1.12	
BDL233	10822.1	P					0.63			0.63		
BDL233	10822.1	Av					1.12			1.12		
BDL233	10825.4	P	0.4	0.4	0.37	0.63	0.35	0.48	0.63	0.35	0.48	
BDL233	10825.4	Av	1.16	1.16	1.16	1.14	1.39	1.25	1.14	1.39	1.25	

<i>Gene</i>	<i>Ev.</i>	<i>Par:</i>	<i>1</i>	<i>2</i>	<i>3</i>	<i>4</i>	<i>5</i>	<i>6</i>	<i>7</i>	<i>8</i>	<i>9</i>	<i>10</i>
BDL237	10893.1	P				0.09			0.09			
BDL237	10893.1	Av				1.12			1.12			
BDL237	10895.1	P	0.37	0.29		0.05	0.56	0.55	0.05	0.56	0.55	
BDI.237	10895.1	Av	1.13	1.1		1.13	1.15	1.11	1.13	1.15	1.11	
BDL237	10895.2	P				0.29			0.29			
BDL237	10895.2	Av				1.12			1.12			
BDL237	10895.3	P	0.55			0.56			0.56			
BDI.237	10895.3	Av	1.13			1.12			1.12			
BDL238	10951.4	P	0.35				0.1			0.1		
BDL238	10951.4	Av	1.11				1.14			1.14		
BDL238	10952.3	P					0.68			0.68		
BDL238	10952.3	Av					1.1			1.1		
BDL238	10954.2	P	0.04	<0.01	<0.01	0.08	<0.01	<0.01	0.08	<0.01	<0.01	<0.01
BDL238	10954.2	Av	1.33	1.28	1.23	1.73	1.76	1.52	1.73	1.76	1.52	1.13
BDL238	10954.3	P				0.74			0.74			
BDL238	10954.3	Av				1.13			1.13			
BDL240	10802.2	P	0.02	0.03	0.12	0.1	0.02	0.04	0.1	0.02	0.04	
BDL240	10802.2	Av	1.22	1.26	1.18	1.34	1.54	1.35	1.34	1.54	1.35	
BDL241	10873.1	P	0.02	0.13	0.11	<0.01	0.38	0.02	<0.01	0.38	0.02	
BDL241	10873.1	Av	1.19	1.18	1.15	1.41	1.3	1.34	1.41	1.3	1.34	
BDL242	10731.2	P	0.4	0.52		0.14	0.46	0.58	0.14	0.46	0.58	
BDL242	10731.2	Av	1.13	1.12		1.22	1.28	1.14	1.22	1.28	1.14	
BDL242	10731.5	P				0.1			0.1			
BDL242	10731.5	Av				1.11			1.11			
BDL242	10731.6	P	<0.01	<0.01	0.18	0.03	0.25	0.08	0.03	0.25	0.08	
BDL242	10731.6	Av	1.21	1.22	1.19	1.48	1.45	1.36	1.48	1.45	1.36	
BDL242	10731.7	P				0.06			0.06			
BDL242	10731.7	Av				1.12			1.12			
BDL245	10813.3	P		0.03								
BDL245	10813.3	Av		1.08								
BDL250	10841.3	P	0.02	0.18	0.26	0.06	0.22	0.32	0.06	0.22	0.32	0.05
BDL250	10841.3	Av	1.24	1.18	1.11	1.36	1.41	1.17	1.36	1.41	1.17	1.08
BDI.250	10846.2	P	0.48			0.64	0.41		0.64	0.41		
BDL250	10846.2	Av	1.1			1.12	1.16		1.12	1.16		
BDL250	10846.3	P	0.38	0.36	0.48	0.58	0.46	0.62	0.58	0.46	0.62	
BDL250	10846.3	Av	1.11	1.17	1.11	1.17	1.34	1.19	1.17	1.34	1.19	
BDL252	10882.1	P	0.3	0.41	0.44	0.32	0.26	0.34	0.32	0.26	0.34	<0.01
BDL252	10882.1	Av	1.19	1.19	1.17	1.38	1.42	1.24	1.38	1.42	1.24	1.1
BDL48	10274.4	P	0.23	0.02	0.02	0.81	0.09	0.08	0.81	0.09	0.08	
BDL48	10274.4	Av	1.14	1.15	1.2	1.1	1.23	1.36	1.1	1.23	1.36	
BDL48	10271.1	P		0.06	<0.01	0.27	0.03	<0.01	0.27	0.03	<0.01	
BDL48	10271.1	Av		1.08	1.16	1.11	1.22	1.27	1.11	1.22	1.27	
BDL48	10274.3	P	0.07	0.01	<0.01	0.02	0.01	<0.01	0.02	0.01	<0.01	
BDL48	10274.3	Av	1.21	1.19	1.09	1.25	1.35	1.22	1.25	1.35	1.22	
BDL48	10274.4	P	0.29	0.2	0.13	0.22	0.03	0.22	0.22	0.03	0.22	
BDL48	10274.4	Av	1.14	1.15	1.14	1.14	1.21	1.26	1.14	1.21	1.26	
BDL48	10274.5	P	0.04	0.28	0.45	0.26	0.49	0.44	0.26	0.49	0.44	
BDL48	10274.5	Av	1.11	1.13	1.11	1.14	1.22	1.22	1.14	1.22	1.22	

<i>Gene</i>	<i>Ev.</i>	<i>Par.</i>	<i>1</i>	<i>2</i>	<i>3</i>	<i>4</i>	<i>5</i>	<i>6</i>	<i>7</i>	<i>8</i>	<i>9</i>	<i>10</i>
BDL63	10381.1	P	0.12	0.02	<0.01	0.11	0.21	<0.01	0.11	0.21	<0.01	
BDL63	10381.1	Av	1.2	1.18	1.17	1.2	1.39	1.37	1.2	1.39	1.37	
BDL63	10381.2	P	0.03	0.25	0.37	0.31	0.6	0.42	0.31	0.6	0.42	
BDL63	10381.2	Av	1.13	1.15	1.19	1.19	1.26	1.32	1.19	1.26	1.32	
BDL63	10384.8	P		0.02	<0.01		0.06	<0.01		0.06	<0.01	
BDL63	10384.8	Av		1.09	1.14		1.2	1.24		1.2	1.24	
BDL79	11042.1	P		0.08			0.04	0.52		0.04	0.52	
BDL79	11042.1	Av		1.06			1.16	1.1		1.16	1.1	
BDL79	11044.3	P	<0.01	0.03			0.14			0.14		
BDL79	11044.3	Av	1.17	1.11			1.14			1.14		
BDL81	10371.8	P				0.34			0.34			
BDL81	10371.8	Av				1.12			1.12			
BDL81	10374.1	P				0.22			0.22			
BDL81	10374.1	Av				1.16			1.16			
BDL81	10371.5	P		0.02	0.17			0.21			0.21	
BDL81	10371.5	Av		1.08	1.11			1.13			1.13	
BDL81	10371.8	P			0.6		0.8	0.57		0.8	0.57	
BDL81	10371.8	Av			1.13		1.13	1.22		1.13	1.22	
BDL81	10374.1	P	0.26	0.31	0.44	0.16	0.19	0.32	0.16	0.19	0.32	0.26
BDL81	10374.1	Av	1.23	1.22	1.19	1.44	1.45	1.49	1.44	1.45	1.49	1.12
BDL85	10411.1	P	0.25	0.34	0.15	0.13	0.39	0.33	0.13	0.39	0.33	0.12
BDL85	10411.1	Av	1.2	1.16	1.14	1.39	1.29	1.28	1.39	1.29	1.28	1.16

Table 24. Results of the greenhouse experiments. Provided are the measured values of each parameter [parameters (Par.) 1-10 according to the parameters described in Table 23 above] in plants expressing the indicated polynucleotides. "Ev" = event; "P" = P-value; "Av" = ratio between the averages of event and control. Note that when the average ratio is higher than "1" the effect of exogenous expression of the gene is an increase of the desired trait;

5

Table 25
Results from greenhouse experiments

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL102	10471.1	P			0.1	0.27	0.03	0.04	0.29			
BDL102	10471.1	Av			1.35	1.23	1.18	1.27	1.1			
BDL102	10472.1	P			0.17	0.35	0.25	0.21	0.22			
BDL102	10472.1	Av			1.36	1.23	1.3	1.29	1.15			
BDL102	10474.1	P			0.18	0.19	0.16	0.17	0.3			0.02
BDL102	10474.1	Av			1.56	1.82	1.54	1.4	1.3			1.03
BDL102	10474.2	P			0.04	<0.01	0.01	0.15	<0.01			
BDL102	10474.2	Av			1.51	1.48	1.34	1.42	1.22			
BDL102	10474.6	P			0.23			0.11				
BDL102	10474.6	Av			1.19			1.12				
BDL117	10071.2	P										0.1
BDL117	10071.2	Av										1.03
BDL117	10073.2	P						0.02	0.15			
BDL117	10073.2	Av						1.1	1.1			
BDL117	10074.1	P		0.21	0.27	0.29	0.26	0.18	0.12	0.04		
BDL117	10074.1	Av		1.19	1.46	1.38	1.33	1.59	1.35	1.17		
BDL117	10074.4	P	0.05	0.37	0.38	0.49	0.3	0.25	0.31			0.03
BDL117	10074.4	Av	1.06	1.12	1.28	1.16	1.19	1.43	1.28			1.02

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL117	10073.1	P			0.62	0.52	0.63					
BDL117	10073.1	Av			1.1	1.16	1.15					
BDL117	10073.2	P				0.56	0.37	0.35	0.53	0.51		
BDL117	10073.2	Av				1.19	1.25	1.14	1.13	1.15		
BDL117	10074.1	P		0.01	0.53	0.37	0.36	0.33	0.42	0.43		
BDL117	10074.1	Av		1.09	1.11	1.29	1.24	1.21	1.17	1.2		
BDL117	10074.4	P				0.37	0.26		0.24	0.16		
BDL117	10074.4	Av				1.1	1.12		1.13	1.11		
BDL138	9811.4	P						0.06				
BDL138	9811.4	Av						1.09				
BDL138	9812.1	P						<0.01	0.02			
BDL138	9812.1	Av						1.16	1.11			
BDL138	9812.3	P	0.05		0.11			0.37	0.52		0.01	
BDL138	9812.3	Av	1.04		1.13			1.14	1.1		1.03	
BDL138	9811.1	P									0.02	<0.01
BDL138	9811.1	Av									1.04	1.04
BDL138	9813.4	P										<0.01
BDL138	9813.4	Av										1.04
BDL140	10423.1	P			0.01	0.22						
BDL140	10423.1	Av			1.29	1.19						
BDL140	10424.3	P										0.49
BDL140	10424.3	Av										1.27
BDL140	10424.4	P						0.64	0.68			
BDL140	10424.4	Av						1.14	1.17			
BDL140	10423.1	P									0.03	
BDL140	10423.1	Av									1.04	
BDL147	10303.1	P			0.11	0.07	0.11	0.12	0.01			
BDL147	10303.1	Av			1.49	1.31	1.18	1.28	1.43			
BDL147	10303.6	P						0.38	0.37			
BDL147	10303.6	Av						1.11	1.12			
BDL147	10304.2	P			0.33							0.05
BDL147	10304.2	Av			1.18							1.04
BDL147	10303.5	P				0.41						
BDL147	10303.5	Av				1.1						
BDL147	10304.2	P				0.48						
BDL147	10304.2	Av				1.17						
BDL149	9823.3	P			0.23			<0.01	0.01			
BDL149	9823.3	Av			1.13			1.38	1.21			
BDL149	9824.3	P		0.09								
BDL149	9824.3	Av		1.04								
BDL149	9824.4	P		0.02	0.12	0.03	0.17	<0.01	<0.01	0.02		
BDL149	9824.4	Av		1.1	1.28	1.12	1.2	1.23	1.22	1.11		
BDL149	9823.3	P			0.62	0.47	0.56					
BDL149	9823.3	Av			1.13	1.16	1.12					
BDL152	10431.1	P			0.31							
BDL152	10431.1	Av			1.1							
BDL152	10431.4	P				0.16	0.47	0.27		0.09		0.03
BDL152	10431.4	Av				1.1	1.1	1.18		1.18		1.02
BDL152	10432.5	P				0.18	0.62					
BDL152	10432.5	Av				1.1	1.16					
BDL152	10434.1	P							0.76			
BDL152	10434.1	Av							1.11			
BDL152	10434.4	P					0.32	0.25	0.04	0.16		

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL152	10434.4	Av					1.2	1.13	1.37	1.12		
BDL152	10434.1	P									0.03	
BDL152	10434.1	Av									1.04	
BDL152	10434.4	P									0.05	
BDL152	10434.4	Av									1.04	
BDL153	10141.3	P			<0.0 1	<0.01	<0.0 1	0.25	0.05	<0.0 1		
BDL153	10141.3	Av			1.32	1.24	1.27	1.33	1.26	1.15		
BDL153	10142.2	P	0.03	0.33	0.28	0.54	0.41	0.2	0.33	0.55		0.07
BDL153	10142.2	Av	1.15	1.16	1.44	1.28	1.41	1.54	1.34	1.12		1.02
BDL153	10143.1	P	0.01		0.21		0.32		0.05		0.01	<0.01
BDL153	10143.1	Av	1.12		1.2		1.14		1.11		1.02	1.02
BDL153	10144.1	P		<0.01	0.05	0.08	0.07					
BDL153	10144.1	Av		1.09	1.13	1.1	1.1					
BDL153	10141.3	P				0.06	0.02	0.02	0.24			
BDL153	10141.3	Av				1.14	1.1	1.26	1.19			
BDL153	10142.2	P		0.1	0.17	0.35	0.06	0.02	0.05	0.04		
BDL153	10142.2	Av		1.05	1.1	1.17	1.22	1.48	1.31	1.26		
BDL153	10142.3	P					0.36	0.23				
BDL153	10142.3	Av					1.14	1.1				
BDL153	10143.1	P						0.15				
BDL153	10143.1	Av						1.14				
BDL153	10143.2	P		0.04		0.15		0.07		<0.0 1		
BDL153	10143.2	Av		1.06		1.11		1.2		1.15		
BDL153	10144.1	P				0.4	0.48		0.24	0.08		
BDL153	10144.1	Av				1.43	1.11		1.34	1.13		
BDL153	10144.4	P							0.54			
BDL153	10144.4	Av							1.1			
BDL154	10703.6	P						0.66				
BDL154	10703.6	Av						1.1				
BDL154	10703.8	P			0.08	0.09	0.54	<0.01	0.09			
BDL154	10703.8	Av			1.47	1.41	1.13	1.47	1.22			
BDL155	9991.5	P			0.36		0.09	0.31	0.41			
BDL155	9991.5	Av			1.11		1.18	1.29	1.35			
BDL155	9991.9	P							0.27			
BDL155	9991.9	Av							1.17			
BDL155	9993.2	P							0.64			
BDL155	9993.2	Av							1.12			
BDL155	9994.3	P	0.16		0.31	0.08	0.12	0.12	0.09	0.41		
BDL155	9994.3	Av	1.14		1.1	1.16	1.17	1.43	1.44	1.12		
BDL155	9993.2	P									0.06	
BDL155	9993.2	Av									1.03	
BDL155	9994.3	P									0.03	
BDL155	9994.3	Av									1.04	
BDL157	9911.4	P										0.09
BDL157	9911.4	Av										1.03
BDL157	9914.2	P							0.12	0.09		
BDL157	9914.2	Av							1.11	1.06		
BDL162	10492.2	P					0.12		0.44			
BDL162	10492.2	Av					1.39		1.15			
BDL162	10492.4	P							0.82			
BDL162	10492.4	Av							1.13			

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL162	10494.1	P						0.1	0.13			
BDL162	10494.1	Av						1.21	1.21			
BDL167	10043.1	P									0.17	
BDL167	10043.1	Av									1.21	
BDL167	10043.2	P									<0.01	0.01
BDL167	10043.2	Av									1.04	1.03
BDL167	10044.2	P			0.05	0.07						0.03
BDL167	10044.2	Av			1.15	1.16						1.02
BDL167	10042.3	P		0.1						0.2		
BDL167	10042.3	Av		1.05						1.12		
BDL167	10043.3	P				0.35	0.19	0.38		0.41		
BDL167	10043.3	Av				1.11	1.1	1.14		1.11		
BDL167	10044.2	P			0.51	0.28	0.27	0.31	0.18			
BDL167	10044.2	Av			1.1	1.14	1.15	1.17	1.17			
BDL168	9881.3	P	0.08	<0.01	0.24	0.17	0.1	0.11	0.21	0.3		<0.01
BDL168	9881.3	Av	1.09	1.1	1.32	1.36	1.44	1.17	1.21	1.15		1.02
BDL168	9881.4	P		0.01	0.1	0.32	0.05	0.16	0.36	0.44		
BDL168	9881.4	Av		1.15	1.3	1.23	1.32	1.45	1.25	1.14		
BDL168	9882.3	P			0.61	0.54	0.49	0.66	0.71	0.66		
BDL168	9882.3	Av			1.15	1.2	1.3	1.2	1.13	1.1		
BDL168	9884.4	P					0.7					
BDL168	9884.4	Av					1.15					
BDL168	9881.4	P					0.57	0.57	0.37	0.4		
BDL168	9881.4	Av					1.11	1.1	1.17	1.14		
BDL168	9882.1	P				0.1	0.22	0.48		<0.01		
BDL168	9882.1	Av				1.12	1.11	1.14		1.16		
BDL168	9882.3	P						0.39	0.22			
BDL168	9882.3	Av						1.12	1.11			
BDL168	9884.1	P							0.11			
BDL168	9884.1	Av							1.12			
BDL169	10743.4	P			0.55	0.5	0.69	0.44				
BDL169	10743.4	Av			1.16	1.19	1.12	1.17				
BDL169	10744.1	P	0.41		0.38			0.29	0.31			
BDL169	10744.1	Av	1.15		1.19			1.4	1.24			
BDL169	10747.1	P	0.09		0.58			0.01				
BDL169	10747.1	Av	1.11		1.22			1.42				
BDL169	10747.5	P	<0.01		0.14	0.06						
BDL169	10747.5	Av	1.11		1.29	1.15						
BDL169	10741.3	P			0.42	0.68		0.53				
BDL169	10741.3	Av			1.17	1.11		1.16				
BDL169	10744.2	P	0.02		0.26	0.31	0.32	0.37	0.24	0.02		0.02
BDL169	10744.2	Av	1.04		1.48	1.56	1.44	1.37	1.26	1.23		1.03
BDL169	10747.1	P			0.34	<0.01	0.36	0.02	0.02			
BDL169	10747.1	Av			1.39	1.32	1.2	1.16	1.17			
BDL169	10747.3	P										0.45
BDL169	10747.3	Av										1.1
BDL169	10747.5	P			0.11	0.14	<0.01	<0.01				
BDL169	10747.5	Av			1.2	1.26	1.27	1.22				
BDL171	10661.2	P	0.6		0.63	0.74	0.35	0.84	0.55			
BDL171	10661.2	Av	1.11		1.17	1.1	1.19	1.1	1.33			
BDL171	10662.3	P			0.4	0.67		0.5	0.15			

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL171	10662.3	Av			1.21	1.11		1.18	1.46			
BDL171	10664.1	P	0.09		<0.0 1	<0.01	0.48	0.03	<0.01	0.01		
BDL171	10664.1	Av	1.15		1.56	1.44	1.42	1.57	1.6	1.29		
BDL171	10664.3	P	0.34	0.47	0.24	0.04	0.46	0.08	0.38	0.48		
BDL171	10664.3	Av	1.19	1.12	1.3	1.29	1.18	1.39	1.43	1.26		
BDL171	10661.5	P	0.51					0.76	0.68			
BDL171	10661.5	Av	1.1					1.11	1.12			
BDL171	10662.2	P			0.39			0.06	0.42			
BDL171	10662.2	Av			1.11			1.47	1.14			
BDL171	10662.3	P	0.09		<0.0 1	0.02	<0.0 1	0.24	0.17	<0.0 1		
BDL171	10662.3	Av	1.3		1.39	1.5	1.33	1.46	1.37	1.23		
BDL171	10663.3	P	<0.01		0.04	0.07		<0.01	0.08			
BDL171	10663.3	Av	1.2		1.2	1.17		1.54	1.24			
BDL171	10664.1	P			0.71	0.79						0.01
BDL171	10664.1	Av			1.18	1.13						1.03
BDL171	10664.3	P	0.08		0.18	0.22		0.01	0.33		0.09	
BDL171	10664.3	Av	1.18		1.29	1.22		1.54	1.23		1.03	
BDL173	9951.2	P									<0.01	
BDL173	9951.2	Av									1.03	
BDL173	9952.1	P					0.61					
BDL173	9952.1	Av					1.13					
BDL173	9954.3	P								<0.01	0.02	
BDL173	9954.3	Av								1.05	1.04	
BDL173	9953.4	P									<0.01	
BDL173	9953.4	Av									1.03	
BDL177	10521.3	P			0.37		0.2					
BDL177	10521.3	Av			1.22		1.16					
BDL177	10522.2	P									<0.01	
BDL177	10522.2	Av									1.05	
BDL177	10524.2	P			0.16			0.21	0.28			
BDL177	10524.2	Av			1.14			1.14	1.17			
BDL182	10691.2	P				0.14	0.59	0.08				
BDL182	10691.2	Av				1.11	1.12	1.2				
BDL182	10692.3	P			0.53	0.2			0.73			
BDL182	10692.3	Av			1.16	1.15			1.13			
BDL182	10693.2	P				0.34		0.18		0.67		
BDL182	10693.2	Av				1.13		1.24		1.11		
BDL182	10693.3	P			0.25	0.07		0.05	0.25			
BDL182	10693.3	Av			1.18	1.14		1.42	1.35			
BDL182	10693.5	P						0.15	0.05			
BDL182	10693.5	Av						1.18	1.29			
BDL182	10691.2	P									0.02	0.01
BDL182	10691.2	Av									1.06	1.04
BDL182	10691.4	P	<0.01	0.2	0.56	0.65		<0.01	0.13			
BDL182	10691.4	Av	1.25	1.15	1.19	1.12		1.48	1.29			
BDL182	10691.8	P			0.79			0.84				
BDL182	10691.8	Av			1.14			1.14				
BDL182	10693.2	P									0.02	
BDL182	10693.2	Av									1.05	
BDL182	10693.3	P						0.85				
BDL182	10693.3	Av						1.13				

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL183	9941.1	P		<0.01			0.26					
BDL183	9941.1	Av		1.09			1.13					
BDL183	9942.4	P						0.02	<0.01			
BDL183	9942.4	Av						1.17	1.16			
BDL183	9943.4	P				0.21	0.15					
BDL183	9943.4	Av				1.11	1.14					
BDL183	9944.1	P	0.05	<0.01	0.09	0.01	0.3	<0.01	0.02			
BDL183	9944.1	Av	1.04	1.1	1.19	1.15	1.13	1.31	1.21			
BDL183	9941.1	P					0.64			0.56		
BDL183	9941.1	Av					1.11			1.16		
BDL183	9942.1	P			0.12	0.31	0.46	0.02				
BDL183	9942.1	Av			1.13	1.19	1.12	1.26				
BDL186	10002.2	P				0.01	<0.01			0.1		
BDL186	10002.2	Av				1.14	1.21			1.12		
BDL186	10004.3	P				0.71	0.59				0.09	<0.01
BDL186	10004.3	Av				1.13	1.15				1.02	1.02
BDL186	10001.3	P						0.3				
BDL186	10001.3	Av						1.25				
BDL186	10004.6	P			0.54							
BDL186	10004.6	Av			1.16							
BDL187	10502.2	P			0.38							
BDL187	10502.2	Av			1.12							
BDL187	10502.4	P						0.59				
BDL187	10502.4	Av						1.1				
BDL187	10503.1	P			0.05				0.26			
BDL187	10503.1	Av			1.26				1.15			
BDL187	10503.3	P			0.29				0.3			
BDL187	10503.3	Av			1.25				1.14			
BDL187	10503.5	P						0.2	0.08			
BDL187	10503.5	Av						1.36	1.25			
BDL188	10462.4	P				0.09						
BDL188	10462.4	Av				1.06						
BDL188	10462.1	P			0.02	0.11	0.44	0.25	0.29			
BDL188	10462.1	Av			1.28	1.32	1.19	1.15	1.14			
BDL188	10462.4	P				0.02	0.09	0.01	0.25			
BDL188	10462.4	Av				1.19	1.22	1.36	1.24			
BDL188	10464.5	P						0.33				
BDL188	10464.5	Av						1.19				
BDL190	10234.1	P				0.5						
BDL190	10234.1	Av				1.11						
BDL190	10234.2	P				0.57					0.01	
BDL190	10234.2	Av				1.1					1.03	
BDL190	10231.1	P						0.21				
BDL190	10231.1	Av						1.13				
BDL190	10231.2	P						0.02				
BDL190	10231.2	Av						1.24				
BDL190	10232.2	P			0.44	0.08	0.16	0.1	0.39	0.25		
BDL190	10232.2	Av			1.18	1.23	1.18	1.29	1.16	1.16		
BDL190	10233.2	P			0.05	0.02	<0.01	0.02	0.05	0.05		0.04
BDL190	10233.2	Av			1.18	1.31	1.21	1.25	1.15	1.31		1.02

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL190	10233.4	P					0.02	0.45	0.5	<0.01		
BDL190	10233.4	Av					1.2	1.16	1.12	1.16		
BDL190	10234.2	P				0.07		0.05				
BDL190	10234.2	Av				1.17		1.19				
BDL192	9921.3	P						0.38				
BDL192	9921.3	Av						1.13				
BDL192	9921.6	P			0.12	0.36	0.55					
BDL192	9921.6	Av			1.15	1.13	1.11					
BDL192	9922.1	P			0.11	0.12		0.57	0.56			
BDL192	9922.1	Av			1.18	1.11		1.17	1.11			
BDL192	9921.6	P		0.09	0.19		0.51					
BDL192	9921.6	Av		1.07	1.1		1.13					
BDL192	9922.5	P		0.01	0.35	0.29				0.5		
BDL192	9922.5	Av		1.06	1.18	1.2				1.11		
BDL193	10152.2	P	0.05	0.23				0.05	0.14			
BDL193	10152.2	Av	1.04	1.1				1.17	1.11			
BDL193	10152.3	P										0.02
BDL193	10152.3	Av										1.02
BDL193	10153.2	P						0.32				
BDL193	10153.2	Av						1.1				
BDL193	10153.4	P			0.37		0.29		0.46	0.09	<0.01	
BDL193	10153.4	Av			1.14		1.15		1.1	1.09	1.02	
BDL193	10153.2	P				0.42					0.07	<0.01
BDL193	10153.2	Av				1.12					1.02	1.03
BDL193	10153.3	P	0.07		0.37	0.01		0.2	0.22	0.09		
BDL193	10153.3	Av	1.1		1.15	1.27		1.2	1.18	1.16		
BDL193	10153.4	P			0.41	0.24	0.16	0.63				
BDL193	10153.4	Av			1.13	1.16	1.1	1.11				
BDL196	10243.1	P			0.65	0.77		0.63	0.71			
BDL196	10243.1	Av			1.2	1.11		1.22	1.11			
BDL201	9961.3	P	0.22	0.04	0.18	0.29	0.43		0.46	0.07		0.09
BDL201	9961.3	Av	1.1	1.06	1.28	1.16	1.33		1.12	1.25		1.02
BDL220	10331.5	P										<0.01
BDL220	10331.5	Av										1.05
BDL220	10331.7	P										0.39
BDL220	10331.7	Av										1.1
BDL220	10333.5	P					0.49					
BDL220	10333.5	Av					1.18					
BDL223	10793.3	P					0.1					
BDL223	10793.3	Av					1.1					
BDL223	10793.5	P	<0.01		0.47	0.01	0.02	0.1	0.21	0.12		0.08
BDL223	10793.5	Av	1.08		1.1	1.23	1.15	1.22	1.16	1.12		1.02
BDL223	10793.8	P	0.08		0.08	0.09	0.06	0.1	0.23			
BDL223	10793.8	Av	1.03		1.28	1.2	1.11	1.21	1.1			
BDL224	10451.7	P						0.57				
BDL224	10451.7	Av						1.11				
BDL224	10453.3	P										0.5
BDL224	10453.3	Av										1.12
BDL225	10401.4	P	0.07									
BDL225	10401.4	Av	1.04									
BDL226	10861.2	P	0.03		0.04	0.08		0.25	0.08	0.29	0.01	
BDL226	10861.2	Av	1.19		1.17	1.14		1.37	1.31	1.12	1.06	

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL226	10864.2	P	<0.01	0.07	<0.01	0.01	0.11	0.06	<0.01			
BDL226	10864.2	Av	1.17	1.08	1.38	1.24	1.13	1.43	1.36			
BDL227	11491.3	P				<0.01		0.5	0.37			
BDL227	11491.3	Av				1.23		1.12	1.12			
BDL227	11492.3	P			0.04	<0.01	0.06	0.03	0.04	0.35		
BDI.227	11492.3	Av			1.16	1.25	1.14	1.21	1.19	1.14		
BDL233	10822.1	P							<0.01			
BDL233	10822.1	Av							1.21			
BDL233	10825.4	P				0.39	0.47	0.32	0.5	0.02		
BDL233	10825.4	Av				1.32	1.29	1.27	1.18	1.12		
BDL237	10893.1	P			0.07							
BDL237	10893.1	Av			1.17							
BDL237	10895.1	P			0.09	0.44		0.16	0.26			
BDL237	10895.1	Av			1.11	1.18		1.22	1.1			
BDL237	10895.2	P			0.2	0.66						
BDL237	10895.2	Av			1.13	1.12						
BDL237	10895.3	P			0.51			0.36	0.64			
BDL237	10895.3	Av			1.12			1.26	1.11			
BDL237	10896.1	P				0.11				0.52		
BDL237	10896.1	Av				1.13				1.1		
BDL238	10951.4	P				0.09		0.22	0.18			
BDL238	10951.4	Av				1.11		1.13	1.12			
BDL238	10952.3	P				0.69				0.7		
BDL238	10952.3	Av				1.1				1.1		
BDL238	10954.2	P			0.08	<0.01	<0.01	0.11	<0.01			
BDL238	10954.2	Av			1.59	1.7	1.46	1.34	1.29			
BDL238	10954.3	P			0.67			0.58				
BDL238	10954.3	Av			1.15			1.1				
BDL240	10802.2	P			0.05	0.04	<0.01	<0.01	0.08	0.05		
BDL240	10802.2	Av			1.34	1.61	1.3	1.22	1.27	1.17		
BDL240	10806.2	P				0.3						
BDL240	10806.2	Av				1.1						
BDL240	10806.6	P	0.01									0.09
BDL240	10806.6	Av	1.06									1.04
BDL241	10873.1	P			<0.01	0.52	0.1	0.09	0.06			
BDI.241	10873.1	Av			1.41	1.24	1.33	1.23	1.23			
BDI.242	10731.2	P			0.05	0.45	0.62	0.49	0.45			
BDL242	10731.2	Av			1.26	1.27	1.11	1.11	1.11			
BDL242	10731.5	P			0.05							
BDL242	10731.5	Av			1.13							
BDL242	10731.6	P			0.01	0.19	0.02	<0.01	0.09		0.03	
BDL242	10731.6	Av			1.46	1.43	1.37	1.29	1.21		1.02	
BDL242	10731.7	P			0.04			0.25				
BDL242	10731.7	Av			1.14			1.15				
BDL245	10813.3	P	0.04	0.02				0.27	0.08	0.2		
BDL245	10813.3	Av	1.07	1.08				1.14	1.2	1.15		
BDL245	10816.3	P										0.02
BDL245	10816.3	Av										1.03
BDL245	10812.3	P								0.07		

<i>Gene</i>	<i>Ev.</i>	<i>Pa r</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL245	10812.3	Av								1.09		
BDL245	10813.3	P						0.12	0.03	0.01		
BDL245	10813.3	Av						1.14	1.11	1.16		
BDL247	10911.4	P										0.01
BDL247	10911.4	Av										1.03
BDL247	10912.6	P										0.02
BDL247	10912.6	Av										1.02
BDL248	11051.1	P									0.04	
BDI 248	11051.1	Av									1.03	
BDL248	11054.1	P									0.06	
BDL248	11054.1	Av									1.04	
BDL250	10841.3	P			0.05	0.23	0.45	0.05	0.01	0.21		
BDL250	10841.3	Av			1.31	1.31	1.12	1.38	1.32	1.1		
BDL250	10842.3	P				0.17						
BDI 250	10842.3	Av				1.11						
BDL250	10846.2	P				0.47		0.32	0.41			
BDL250	10846.2	Av				1.13		1.2	1.11			
BDL250	10846.3	P			0.54	0.46	0.51		0.14	0.26		
BDL250	10846.3	Av			1.15	1.3	1.22		1.25	1.15		
BDL252	10882.1	P			0.26	0.25	0.24	0.33	0.51	0.54	0.01	
BDL252	10882.1	Av			1.35	1.36	1.22	1.35	1.28	1.21	1.02	
BDL252	10882.4	P			0.49							
BDL252	10882.4	Av			1.14							
BDL48	10274.4	P	0.04				0.01	0.06	0.01	0.13		
BDL48	10274.4	Av	1.11				1.33	1.4	1.49	1.14		
BDL48	10271.1	P			0.18	0.02	<0.0 1	0.29		0.18	0.02	0.06
BDL48	10271.1	Av			1.13	1.21	1.24	1.11		1.22	1.02	1.02
BDL48	10271.5	P						0.51				
BDL48	10271.5	Av						1.1				
BDL48	10274.3	P			0.01	<0.01	<0.0 1	<0.01	<0.01	<0.0 1		
BDL48	10274.3	Av			1.24	1.33	1.16	1.44	1.32	1.17		
BDL48	10274.4	P			0.11	0.02	0.15	0.53	0.15	0.3		
BDL48	10274.4	Av			1.21	1.23	1.2	1.18	1.25	1.21		
BDL48	10274.5	P			0.41	0.48	0.59	0.27	0.02	0.21		
BDL48	10274.5	Av			1.12	1.23	1.12	1.24	1.2	1.23		
BDL63	10381.2	P						0.48				
BDL63	10381.2	Av						1.29				
BDL63	10381.1	P	0.04	0.1	0.21	0.23	0.04	0.19	0.01	<0.0 1		
BDL63	10381.1	Av	1.1	1.05	1.22	1.34	1.29	1.34	1.26	1.25		
BDI 63	10381.2	P			0.21	0.5	0.48	0.17	0.01	0.23		
BDL63	10381.2	Av			1.21	1.28	1.25	1.26	1.22	1.21		
BDL63	10384.8	P		0.1		0.04	<0.0 1	0.33	0.14	<0.0 1		
BDL63	10384.8	Av		1.05		1.2	1.2	1.15	1.1	1.25		
BDL79	11042.1	P	<0.01			0.12		0.2	0.03			
BDL79	11042.1	Av	1.08			1.1		1.14	1.11			
BDL79	11044.3	P				0.12		<0.01	0.01	0.41		
BDL79	11044.3	Av				1.12		1.24	1.15	1.18		
BDL81	10371.8	P			0.18							
BDL81	10371.8	Av			1.15							

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>11</i>	<i>12</i>	<i>13</i>	<i>14</i>	<i>15</i>	<i>16</i>	<i>17</i>	<i>18</i>	<i>19</i>	<i>20</i>
BDL81	10374.1	P			0.24							
BDL81	10374.1	Av			1.13							
BDL81	10371.5	P				0.2	0.32	0.06		0.02		
BDL81	10371.5	Av				1.15	1.15	1.18		1.1		
BDL81	10371.8	P				0.72	0.62	0.68	0.57	0.49		
BDL81	10371.8	Av				1.16	1.19	1.15	1.11	1.23		
BDL81	10374.1	P			0.18	0.17	0.35	0.1	0.15	0.41		
BDL81	10374.1	Av			1.41	1.41	1.35	1.3	1.35	1.26		
BDL85	10411.1	P			0.16	0.34	0.26	0.23	0.62	<0.01		
BDL85	10411.1	Av			1.29	1.3	1.28	1.32	1.12	1.1		
BDL85	10414.1	P										0.07
BDL85	10414.1	Av										1.02
BDL85	10414.2	P			0.71							
BDL85	10414.2	Av			1.1							

Table 25. Results of the greenhouse experiments. Provided are the measured values of each parameter [parameters (Par.) 11-20 according to the parameters described in Table 23 above] in plants expressing the indicated polynucleotides. "Ev" = event; "P" = P-value; "Av" = ratio between the averages of event and control. Note that when the average ratio is higher than "1" the effect of exogenous expression of the gene is an increase of the desired trait;

5

Table 26
Results from greenhouse experiments

10

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL102	10471.1	P		0.18	0.42		0.39		0.46		0.46	
BDL102	10471.1	Av		1.15	1.82		1.16		1.13		1.13	
BDL102	10471.3	P			0.48							
BDL102	10471.3	Av			1.38							
BDL102	10472.1	P		0.01	0.54		0.17		0.22		0.22	0.43
BDL102	10472.1	Av		1.26	2.05		1.28		1.24		1.24	1.11
BDL102	10474.1	P		0.06		0.29	0.01		0.02	0.17	0.02	0.1
BDL102	10474.1	Av		1.15		1.43	1.54		1.48	1.14	1.48	1.29
BDL102	10474.2	P		0.03			0.1		0.18		0.18	
BDL102	10474.2	Av		1.14			1.32		1.25		1.25	
BDL102	10474.6	P	0.02		0.55							
BDL102	10474.6	Av	1.02		1.76							
BDL117	10071.2	P						0.24				
BDL117	10071.2	Av						1.15				
BDL117	10073.2	P		0.46	0.01	0.01						
BDL117	10073.2	Av		1.19	1.2	1.14						
BDL117	10074.1	P			0.53		0.08	0.08	0.03	0.11	0.02	
BDL117	10074.1	Av			1.13		1.28	1.22	1.42	1.18	1.44	
BDL117	10074.4	P					0.33		0.19		0.14	
BDL117	10074.4	Av					1.16		1.24		1.26	
BDL117	10071.2	P						0.21				
BDL117	10071.2	Av						1.16				
BDL117	10073.1	P					0.35		0.43			0.47
BDL117	10073.1	Av					1.17		1.14			1.16
BDL117	10073.2	P					0.09		0.16	0.08	0.16	
BDL117	10073.2	Av					1.3		1.25	1.2	1.25	
BDL117	10074.1	P					0.16	0.29	0.08	0.1	0.08	

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL117	10074.1	Av					1.24	1.12	1.31	1.17	1.31	
BDL117	10074.4	P		0.71			0.54		0.43	0.27	0.43	
BDL117	10074.4	Av		1.14			1.1		1.13	1.1	1.13	
BDL138	9811.1	P				0.15						
BDL138	9811.1	Av				1.13						
BDL138	9811.4	P		0.41	0.03	<0.01						
BDL138	9811.4	Av		1.21	1.07	1.26						
BDL138	9812.1	P		<0.01	0.42	<0.01						
BDL138	9812.1	Av		1.14	1.16	1.2						
BDL138	9812.3	P	0.07									
BDL138	9812.3	Av	1.02									
BDL138	9813.1	P						0.28				
BDL138	9813.1	Av						1.13				
BDL138	9813.3	P		0.06	0.55							
BDL138	9813.3	Av		1.3	1.12							
BDL138	9811.1	P	0.04									
BDL138	9811.1	Av	1.02									
BDL138	9813.1	P		0.56				0.1				
BDL138	9813.1	Av		1.19				1.18				
BDL138	9813.4	P		0.75								
BDL138	9813.4	Av		1.14								
BDL138	9811.4	P		0.29								
BDL138	9811.4	Av		1.1								
BDL138	9812.1	P		0.39								
BDL138	9812.1	Av		1.61								
BDL138	9813.1	P		0.14				0.29				
BDL138	9813.1	Av		1.33				1.12				
BDL138	9813.3	P		0.36	0.09							
BDL138	9813.3	Av		1.87	1.07							
BDL140	10421.3	P		0.84		0.37						
BDL140	10421.3	Av		1.16		1.18						
BDL140	10424.3	P			0.47							
BDL140	10424.3	Av			1.18							
BDL140	10421.2	P		0.02	0.22							
BDL140	10421.2	Av		1.14	1.12							
BDL140	10423.1	P	0.08									
BDL140	10423.1	Av	1.03									
BDL147	10301.3	P		0.54								
BDL147	10301.3	Av		1.17								
BDL147	10303.1	P	0.01				0.53		0.32		0.32	
BDL147	10303.1	Av	1.02				1.11		1.18		1.18	
BDL147	10303.6	P	0.07									
BDL147	10303.6	Av	1.02									
BDL147	10304.2	P	<0.01					0.55				
BDL147	10304.2	Av	1.04					1.1				
BDL147	10301.5	P		0.36								
BDL147	10301.5	Av		1.15								
BDL147	10301.6	P		0.02								
BDL147	10301.6	Av		1.14								
BDL147	10303.1	P		0.01								
BDL147	10303.1	Av		1.29								
BDL147	10303.5	P		0.02		0.43		0.31				0.08
BDL147	10303.5	Av		1.17		1.17		1.12				1.09
BDL147	10304.2	P						0.04	0.36		0.36	

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL147	10304.2	Av						1.23	1.14		1.14	
BDL149	9823.1	P		0.04	0.05	0.11						
BDL149	9823.1	Av		1.17	1.19	1.17						
BDL149	9823.3	P		<0.01	<0.01	0.23		0.34	0.42			
BDL149	9823.3	Av		1.17	1.14	1.12		1.12	1.14			
BDL149	9824.3	P						0.34				
BDL149	9824.3	Av						1.11				
BDL149	9824.4	P					0.28		0.11		0.08	<0.01
BDL149	9824.4	Av					1.17		1.28		1.3	1.15
BDL149	9823.3	P					0.48		0.48		0.48	0.1
BDL149	9823.3	Av					1.12		1.12		1.12	1.08
BDL149	9824.3	P						0.27				
BDL149	9824.3	Av						1.14				
BDL152	10431.1	P	0.01									
BDL152	10431.1	Av	1.02									
BDL152	10432.5	P					0.37					
BDL152	10432.5	Av					1.17					
BDL152	10434.1	P			0.05							
BDL152	10434.1	Av			1.16							
BDL152	10434.4	P		0.58	0.27		0.3		0.32	0.31	0.32	0.56
BDL152	10434.4	Av		1.14	1.17		1.19		1.19	1.15	1.19	1.1
BDL152	10431.1	P		0.43	0.58							
BDL152	10431.1	Av		1.18	1.1							
BDL152	10431.3	P				0.02						
BDL152	10431.3	Av				1.18						
BDL152	10434.1	P			0.42							
BDL152	10434.1	Av			1.1							
BDL152	10434.4	P	0.09									
BDL152	10434.4	Av	1.03									
BDL153	10141.3	P	0.09				0.12		0.07	0.23	0.05	
BDL153	10141.3	Av	1.02				1.25		1.33	1.13	1.35	
BDL153	10142.2	P					0.04	0.09	0.01	0.11	<0.01	
BDL153	10142.2	Av					1.37	1.22	1.56	1.2	1.58	
BDL153	10143.1	P					0.45		0.48		0.4	
BDL153	10143.1	Av					1.12		1.12		1.14	
BDL153	10144.1	P					0.52	0.25	0.53		0.45	
BDL153	10144.1	Av					1.1	1.13	1.11		1.12	
BDL153	10141.3	P		0.22	0.67		0.48		0.39		0.39	
BDL153	10141.3	Av		1.28	1.12		1.11		1.14		1.14	
BDL153	10142.2	P		0.05	<0.01	0.63	0.11		0.06	0.04	0.06	
BDL153	10142.2	Av		1.36	1.2	1.1	1.26		1.33	1.18	1.33	
BDL153	10142.3	P					0.26		0.29	0.2	0.29	
BDL153	10142.3	Av					1.19		1.18	1.12	1.18	
BDL153	10143.1	P		0.09								0.25
BDL153	10143.1	Av		1.09								1.11
BDL153	10143.2	P		0.05			0.47	0.22	0.36	0.23	0.36	0.23
BDL153	10143.2	Av		1.11			1.11	1.15	1.15	1.1	1.15	1.13
BDL153	10144.1	P					0.28		0.26	0.06	0.26	
BDL153	10144.1	Av					1.17		1.18	1.17	1.18	
BDL154	10703.1	P		0.11	0.64	0.67						
BDL154	10703.1	Av		1.1	1.2	1.27						
BDL154	10703.5	P		0.37	0.44	0.66						
BDL154	10703.5	Av		1.18	1.26	1.13						
BDL154	10703.6	P		0.25	0.59	0.59						

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL154	10703.6	Av		1.28	1.47	1.29						
BDL154	10703.8	P		0.29			0.58		0.58		0.58	
BDL154	10703.8	Av		1.17			1.1		1.1		1.1	
BDL155	9991.3	P				0.65						
BDL155	9991.3	Av				1.11						
BDL155	9991.5	P			0.5		0.34		0.57	0.4	0.57	0.32
BDL155	9991.5	Av			1.23		1.17		1.11	1.12	1.11	1.22
BDL155	9991.9	P			0.39							
BDL155	9991.9	Av			1.22							
BDL155	9994.3	P			0.49	0.56	0.4		0.31	0.49	0.31	
BDL155	9994.3	Av			1.1	1.14	1.15		1.19	1.1	1.19	
BDL155	9994.5	P						0.41				
BDL155	9994.5	Av						1.14				
BDL157	9911.3	P		0.25	0.14							
BDL157	9911.3	Av		1.54	1.11							
BDL157	9911.4	P		<0.01								
BDL157	9911.4	Av		1.26								
BDL157	9914.2	P		0.37								
BDL157	9914.2	Av		1.23								
BDL157	9911.3	P		0.08	0.28							
BDL157	9911.3	Av		1.15	1.25							
BDL157	9911.4	P		0.46								
BDL157	9911.4	Av		1.23								
BDL157	9913.3	P		0.37								
BDL157	9913.3	Av		1.21								
BDL157	9914.2	P										0.45
BDL157	9914.2	Av										1.16
BDL160	10011.5	P		0.34		0.54		0.35				
BDL160	10011.5	Av		1.59		1.26		1.15				
BDL160	10011.6	P		0.03	0.61	0.67						
BDL160	10011.6	Av		1.15	1.23	1.16						
BDL160	10011.7	P		0.02								
BDL160	10011.7	Av		1.16								
BDL160	10013.1	P		0.01		0.41						
BDL160	10013.1	Av		1.32		1.27						
BDL160	10015.1	P		0.09		0.09						
BDL160	10015.1	Av		1.1		1.14						
BDL162	10491.1	P		0.42								
BDL162	10491.1	Av		1.44								
BDL162	10492.2	P					0.04		0.09	0.27	0.09	0.05
BDL162	10492.2	Av					1.4		1.34	1.17	1.34	1.44
BDL162	10492.4	P		0.37	0.39			0.55				
BDL162	10492.4	Av		1.83	1.25			1.11				
BDL162	10494.1	P			0.63							
BDL162	10494.1	Av			1.13							
BDL167	10042.3	P		0.19								
BDL167	10042.3	Av		1.21								
BDL167	10042.3	P						0.24				
BDL167	10042.3	Av						1.15				
BDL167	10043.2	P		0.13								0.6
BDL167	10043.2	Av		1.13								1.14
BDL167	10043.3	P					0.44					0.27
BDL167	10043.3	Av					1.12					1.12
BDL167	10043.4	P		0.05		0.33						0.14

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL167	10043.4	Av		1.16		1.12						1.46
BDL167	10044.2	P		0.2			0.27		0.2		0.2	0.24
BDL167	10044.2	Av		1.18			1.18		1.21		1.21	1.19
BDL168	9881.3	P					0.01		0.01	0.04	0.01	
BDL168	9881.3	Av					1.44		1.48	1.23	1.5	
BDL168	9881.4	P			0.36		0.06	0.05	0.07	0.11	0.05	
BDL168	9881.4	Av			1.14		1.31	1.24	1.33	1.18	1.35	
BDL168	9882.1	P									0.51	
BDL168	9882.1	Av									1.11	
BDL168	9882.3	P					0.08		0.09	0.16	0.07	
BDL168	9882.3	Av					1.31		1.33	1.17	1.35	
BDL168	9884.4	P	<0.01				0.32		0.46	0.2	0.39	
BDL168	9884.4	Av	1.03				1.17		1.14	1.16	1.16	
BDL168	9881.3	P			0.66							<0.01
BDL168	9881.3	Av			1.1							1.19
BDL168	9881.4	P		0.12			0.44		0.34	0.14	0.34	
BDL168	9881.4	Av		1.17			1.13		1.16	1.13	1.16	
BDL168	9882.1	P					0.53		0.39			0.58
BDL168	9882.1	Av					1.1		1.14			1.13
BDL168	9882.3	P		0.01	0.42			0.35				0.18
BDL168	9882.3	Av		1.19	1.1			1.12				1.2
BDL168	9883.3	P			0.06							
BDL168	9883.3	Av			1.09							
BDL168	9884.1	P								0.07		
BDL168	9884.1	Av								1.17		
BDL169	10743.4	P					0.54		0.37		0.65	
BDL169	10743.4	Av					1.11		1.18		1.1	
BDL169	10744.1	P			0.28			0.3				
BDL169	10744.1	Av			1.12			1.13				
BDL169	10747.1	P		0.6	0.53							
BDL169	10747.1	Av		1.12	1.17							
BDL169	10747.5	P							0.61		0.44	
BDL169	10747.5	Av							1.1		1.16	
BDL169	10741.3	P		0.73								
BDL169	10741.3	Av		1.1								
BDL169	10744.2	P					0.04		0.05	0.07	0.05	0.84
BDL169	10744.2	Av					1.43		1.41	1.19	1.41	1.1
BDL169	10747.1	P			0.65	0.43	0.35		0.43		0.43	
BDL169	10747.1	Av			1.21	1.24	1.18		1.15		1.15	
BDL169	10747.3	P				0.4						
BDL169	10747.3	Av				2.16						
BDL169	10747.5	P					0.16		0.53		0.53	
BDL169	10747.5	Av					1.27		1.11		1.11	
BDL171	10661.2	P					0.3		0.31	0.54	0.31	0.34
BDL171	10661.2	Av					1.19		1.21	1.1	1.21	1.3
BDL171	10661.5	P			0.36							
BDL171	10661.5	Av			1.13							
BDL171	10664.1	P			0.02		0.07		0.03	0.28	0.03	0.67
BDL171	10664.1	Av			1.19		1.39		1.48	1.17	1.48	1.22
BDL171	10664.3	P					0.34		0.08	0.46	0.08	0.61
BDL171	10664.3	Av					1.17		1.36	1.11	1.36	1.25
BDL171	10662.2	P		0.09	0.07							
BDL171	10662.2	Av		1.45	1.45							
BDL171	10662.3	P			0.42		0.07		0.11	0.21	0.07	0.6

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL171	10662.3	Av			1.2		1.33		1.32	1.16	1.4	1.15
BDL171	10663.3	P		0.22	0.01	0.56						
BDL171	10663.3	Av		1.19	1.51	1.14						
BDL171	10664.1	P									0.68	
BDL171	10664.1	Av									1.1	
BDL171	10664.3	P	0.09		0.07				0.48		0.34	
BDL171	10664.3	Av	1.02		1.22				1.14		1.21	
BDL173	9952.1	P			0.18		0.39	0.22	0.51		0.44	
BDL173	9952.1	Av			1.13		1.14	1.15	1.12		1.13	
BDL173	9954.2	P				0.09		0.35				
BDL173	9954.2	Av				1.08		1.11				
BDL173	9953.4	P	0.03									0.23
BDL173	9953.4	Av	1.02									1.3
BDL173	9954.2	P	0.02									0.1
BDL173	9954.2	Av	1.02									1.11
BDL176	9891.2	P		0.6	0.58							
BDL176	9891.2	Av		1.24	1.19							
BDL176	9892.3	P		0.38								
BDL176	9892.3	Av		1.89								
BDL176	9893.2	P		0.73								
BDL176	9893.2	Av		1.11								
BDL176	9893.3	P	0.01									0.37
BDL176	9893.3	Av	1.02									1.12
BDL177	10521.3	P					0.41			0.53		
BDL177	10521.3	Av					1.15			1.1		
BDL177	10522.2	P		0.44								
BDL177	10522.2	Av		1.19								
BDL182	10691.2	P			0.45		0.35		0.57	0.52	0.57	
BDL182	10691.2	Av			1.11		1.17		1.11	1.1	1.11	
BDL182	10693.2	P		0.12								
BDL182	10693.2	Av		1.28								
BDL182	10693.3	P							0.55		0.55	
BDL182	10693.3	Av							1.11		1.11	
BDL182	10693.5	P			0.01	0.07						
BDL182	10693.5	Av			1.28	1.08						
BDL182	10691.2	P				0.58						
BDL182	10691.2	Av				1.12						
BDL182	10691.4	P	0.09		0.29			0.11				
BDL182	10691.4	Av	1.02		1.27			1.2				
BDL182	10691.8	P			0.61							
BDL182	10691.8	Av			1.11							
BDL182	10693.2	P			0.38	0.57		0.37				
BDL182	10693.2	Av			1.1	1.13		1.12				
BDL182	10693.3	P		0.69	0.43	0.46						
BDL182	10693.3	Av		1.25	1.59	1.25						
BDL183	9941.1	P					0.37	0.25	0.5		0.42	
BDL183	9941.1	Av					1.14	1.14	1.12		1.13	
BDL183	9942.1	P										0.55
BDL183	9942.1	Av										1.12
BDL183	9942.4	P		0.22	0.09	0.01		0.07			0.53	
BDL183	9942.4	Av		1.24	1.17	1.13		1.21			1.11	
BDL183	9943.4	P					0.31		0.53	0.37		
BDL183	9943.4	Av					1.16		1.11	1.1		
BDL183	9944.1	P					0.46		0.23		0.18	

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL183	9944.1	Av					1.12		1.21		1.23	
BDL183	9944.4	P	<0.01									
BDL183	9944.4	Av	1.03									
BDL183	9941.1	P					0.38		0.42	0.21	0.42	
BDL183	9941.1	Av					1.15		1.13	1.13	1.13	
BDL183	9942.1	P		0.17			0.48		0.42		0.42	
BDL183	9942.1	Av		1.15			1.11		1.13		1.13	
BDL183	9942.4	P		0.02	0.21							
BDL183	9942.4	Av		1.16	1.13							
BDL183	9943.4	P		0.35				0.33				0.12
BDL183	9943.4	Av		1.24				1.13				1.16
BDL183	9944.2	P		0.59	0.5							
BDL183	9944.2	Av		1.12	1.14							
BDL186	10002.2	P					0.15	0.23	0.14	0.24	0.1	
BDL186	10002.2	Av					1.23	1.14	1.26	1.13	1.28	
BDL186	10004.3	P					0.32		0.39	0.2	0.33	
BDL186	10004.3	Av					1.16		1.16	1.16	1.18	
BDL186	10001.3	P			0.46							
BDL186	10001.3	Av			1.1							
BDL186	10004.3	P		0.55								
BDL186	10004.3	Av		1.15								
BDL187	10502.2	P		0.43								
BDL187	10502.2	Av		1.21								
BDL187	10501.2	P	0.01	0.58								
BDL187	10501.2	Av	1.02	1.38								
BDL187	10502.4	P		0.66								
BDL187	10502.4	Av		1.1								
BDL187	10503.5	P		0.38	0.2	0.34						
BDL187	10503.5	Av		1.13	1.24	1.12						
BDL188	10462.1	P		0.46								
BDL188	10462.1	Av		1.41								
BDL188	10464.5	P						0.33				
BDL188	10464.5	Av						1.12				
BDL188	10462.1	P					0.38		0.35		0.35	
BDL188	10462.1	Av					1.16		1.18		1.18	
BDL188	10462.4	P	0.02	0.41			0.26		0.24		0.24	
BDL188	10462.4	Av	1.02	1.32			1.2		1.22		1.22	
BDL188	10464.3	P		0.66								
BDL188	10464.3	Av		1.24								
BDL188	10464.5	P		0.29	0.1							
BDL188	10464.5	Av		1.38	1.12							
BDL190	10232.2	P										0.6
BDL190	10232.2	Av										1.1
BDL190	10233.4	P										0.01
BDL190	10233.4	Av										1.2
BDL190	10234.1	P	0.01									0.4
BDL190	10234.1	Av	1.02									1.14
BDL190	10231.1	P		0.42	0.24							
BDL190	10231.1	Av		1.18	1.21							
BDL190	10231.2	P		0.64								0.5
BDL190	10231.2	Av		1.14								1.2
BDL190	10232.2	P		<0.01			0.26		0.14		0.14	
BDL190	10232.2	Av		1.29			1.18		1.25		1.25	
BDL190	10233.2	P					0.1	0.29	0.05	0.01	0.05	

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL190	10233.2	Av					1.25	1.13	1.32	1.24	1.32	
BDL190	10233.4	P					0.16		0.29	0.29	0.29	
BDL190	10233.4	Av					1.23		1.17	1.1	1.17	
BDL190	10234.2	P		0.04								0.02
BDL190	10234.2	Av		1.11								1.09
BDL192	9921.3	P		0.54	0.56			0.16				
BDL192	9921.3	Av		1.19	1.11			1.16				
BDL192	9921.6	P					0.51		0.47		0.4	
BDL192	9921.6	Av					1.1		1.12		1.14	
BDL192	9922.1	P									0.52	
BDL192	9922.1	Av									1.11	
BDL192	9922.2	P	<0.01									
BDL192	9922.2	Av	1.02									
BDL192	9921.6	P					0.49	0.03	0.45		0.45	
BDL192	9921.6	Av					1.15	1.25	1.16		1.16	
BDL192	9922.5	P							0.54			0.51
BDL192	9922.5	Av							1.13			1.15
BDL193	10152.2	P						0.15				
BDL193	10152.2	Av						1.17				
BDL193	10152.3	P				0.3		0.1				
BDL193	10152.3	Av				1.11		1.2				
BDL193	10153.2	P						0.24				
BDL193	10153.2	Av						1.14				
BDL193	10153.4	P	0.04				0.39		0.4		0.33	
BDL193	10153.4	Av	1.02				1.14		1.14		1.16	
BDL193	10152.2	P			0.7							
BDL193	10152.2	Av			1.12							
BDL193	10152.3	P						0.14				
BDL193	10152.3	Av						1.16				
BDL193	10153.2	P	0.01									
BDL193	10153.2	Av	1.02									
BDL193	10153.3	P		0.35				0.4	0.48		0.48	
BDL193	10153.3	Av		1.22				1.1	1.14		1.14	
BDL193	10153.4	P					0.63		0.5		0.5	
BDL193	10153.4	Av					1.1		1.14		1.14	
BDL196	10241.3	P		0.5								
BDL196	10241.3	Av		1.1								
BDL196	10243.1	P			0.01				0.56		0.56	
BDL196	10243.1	Av			1.08				1.14		1.14	
BDL196	10243.2	P		0.37								
BDL196	10243.2	Av		1.29								
BDL196	10244.1	P		0.02	0.22	0.82						0.02
BDL196	10244.1	Av		1.21	1.11	1.1						1.13
BDL201	9961.3	P					0.12		0.12	0.31	0.12	
BDL201	9961.3	Av					1.31		1.32	1.16	1.32	
BDL201	9961.4	P			0.62							
BDL201	9961.4	Av			1.14							
BDL201	9961.6	P	0.01	0.01								
BDL201	9961.6	Av	1.02	1.54								
BDL201	9963.6	P		0.46	0.37	0.21						
BDL201	9963.6	Av		1.29	1.13	1.15						
BDL220	10331.5	P		0.76								
BDL220	10331.5	Av		1.13								
BDL220	10331.7	P			0.62	0.42		0.29				

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL.220	10331.7	Av			1.21	1.17		1.18				
BDL.220	10333.2	P		0.62								
BDL.220	10333.2	Av		1.49								
BDL.220	10333.5	P				0.01	0.25			0.26		0.65
BDL.220	10333.5	Av				1.13	1.22			1.18		1.14
BDL.220	10334.1	P	<0.01		0.36	0.64						
BDL.220	10334.1	Av	1.04		1.31	1.11						
BDL.223	10791.1	P				0.51						
BDL.223	10791.1	Av				1.78						
BDL.223	10793.1	P		0.02	0.58							
BDL.223	10793.1	Av		1.16	1.34							
BDL.223	10793.3	P				0.7						
BDL.223	10793.3	Av				1.1						
BDL.223	10793.5	P		0.01		<0.01	0.42		0.48		0.48	0.18
BDL.223	10793.5	Av		1.18		1.41	1.15		1.13		1.13	1.17
BDL.223	10793.8	P			0.6							
BDL.223	10793.8	Av			1.36							
BDL.224	10451.3	P							0.61	0.5	0.61	
BDL.224	10451.3	Av							1.1	1.11	1.1	
BDL.224	10451.7	P										0.78
BDL.224	10451.7	Av										1.11
BDL.224	10451.8	P		0.03								0.54
BDL.224	10451.8	Av		1.78								1.11
BDL.224	10453.1	P										0.77
BDL.224	10453.1	Av										1.11
BDL.224	10453.3	P		0.62				0.39				
BDL.224	10453.3	Av		1.82				1.14				
BDL.225	10401.4	P						0.18				
BDL.225	10401.4	Av						1.17				
BDL.225	10402.2	P		0.29	0.74							
BDL.225	10402.2	Av		1.28	1.1							
BDL.225	10402.5	P				0.38						
BDL.225	10402.5	Av				1.13						
BDL.225	10401.4	P		0.5								0.65
BDL.225	10401.4	Av		1.12								1.15
BDL.225	10402.2	P				0.42						0.38
BDL.225	10402.2	Av				1.1						1.15
BDL.225	10402.6	P										0.38
BDL.225	10402.6	Av										1.15
BDL.225	10402.9	P			0.07	0.01						
BDL.225	10402.9	Av			1.14	1.15						
BDL.226	10861.2	P			0.53	0.54						
BDL.226	10861.2	Av			1.22	1.12						
BDL.226	10862.2	P	0.03									
BDL.226	10862.2	Av	1.03									
BDL.226	10864.2	P			0.44		0.58		0.32		0.21	
BDL.226	10864.2	Av			1.17		1.1		1.2		1.27	
BDL.226	10861.1	P			0.68	0.72						
BDL.226	10861.1	Av			1.27	1.21						
BDL.226	10861.4	P		0.04		0.41						
BDL.226	10861.4	Av		1.13		1.44						
BDL.226	10862.2	P				0.21						
BDL.226	10862.2	Av				1.51						
BDL.226	10863.4	P			0.53							

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL240	10806.2	Av		1.1								1.11
BDL241	10873.1	P			0.55		0.11		0.09	0.17	0.09	0.34
BDL241	10873.1	Av			1.86		1.32		1.33	1.13	1.33	1.25
BDL241	10873.4	P				0.44						
BDL241	10873.4	Av				1.73						
BDL241	10874.2	P	<0.01									
BDL241	10874.2	Av	1.03									
BDL241	10875.1	P				0.33						
BDL241	10875.1	Av				1.57						
BDL241	10875.2	P			0.58	0.09						
BDL241	10875.2	Av			1.48	1.19						
BDL242	10731.2	P					0.59		0.46		0.46	
BDL242	10731.2	Av					1.1		1.14		1.14	
BDL242	10731.3	P			<0.01							
BDL242	10731.3	Av			1.74							
BDL242	10731.5	P			0.7							
BDL242	10731.5	Av			1.18							
BDL242	10731.6	P			0.63		0.07		0.07	0.09	0.07	0.04
BDL242	10731.6	Av			1.48		1.36		1.35	1.17	1.35	1.16
BDL242	10731.7	P			0.18							
BDL242	10731.7	Av			2.28							
BDL242	10737.2	P				0.07						
BDL242	10737.2	Av				1.22						
BDL245	10811.3	P		0.22	0.05	<0.01		0.2				
BDL245	10811.3	Av		1.47	1.25	1.25		1.16				
BDL245	10813.3	P			0.28	0.02		0.22				
BDL245	10813.3	Av			1.17	1.33		1.15				
BDL245	10811.2	P		0.37		0.3						
BDL245	10811.2	Av		1.22		1.49						
BDL245	10811.3	P		0.21	0.12	0.5						
BDL245	10811.3	Av		1.18	1.44	1.79						
BDL245	10812.3	P			0.55	0.11						
BDL245	10812.3	Av			2.2	1.12						
BDL245	10813.3	P		0.37		0.45		0.04				
BDL245	10813.3	Av		1.19		1.1		1.26				
BDL245	10816.3	P			0.52	0.23						
BDL245	10816.3	Av			1.81	1.14						
BDL247	10911.4	P		0.42								
BDL247	10911.4	Av		1.1								
BDL247	10912.1	P			0.48	0.58						
BDL247	10912.1	Av			1.26	1.16						
BDL247	10912.6	P				0.6						
BDL247	10912.6	Av				1.33						
BDL248	11051.2	P		0.45								
BDL248	11051.2	Av		1.14								
BDL250	10841.3	P		0.2		0.13	0.58		0.36		0.36	
BDL250	10841.3	Av		1.14		1.16	1.1		1.17		1.17	
BDL250	10842.1	P		0.45		0.37						
BDL250	10842.1	Av		1.12		1.31						
BDL250	10842.3	P		0.02		0.46						
BDL250	10842.3	Av		1.15		1.14						
BDL250	10843.2	P				0.42						
BDL250	10843.2	Av				1.19						
BDL250	10846.2	P		0.01		0.06						

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>21</i>	<i>22</i>	<i>23</i>	<i>24</i>	<i>25</i>	<i>26</i>	<i>27</i>	<i>28</i>	<i>29</i>	<i>30</i>
BDL63	10384.3	Av										1.14
BDL63	10384.7	P		0.04								<0.01
BDL63	10384.7	Av		1.12								1.16
BDL63	10384.8	P					0.12		0.11	0.06	0.11	0.31
BDL63	10384.8	Av					1.25		1.26	1.17	1.26	1.12
BDL79	11041.1	P		0.13	0.49	0.77						
BDL79	11041.1	Av		1.11	2.06	1.1						
BDL79	11042.1	P		0.09				0.2	0.54		0.54	
BDL79	11042.1	Av		1.15				1.16	1.11		1.11	
BDL79	11042.3	P			0.46							
BDL79	11042.3	Av			1.37							
BDL79	11043.1	P			0.47							
BDL79	11043.1	Av			1.92							
BDL79	11044.3	P		0.01		0.15						
BDL79	11044.3	Av		1.21		1.23						
BDL81	10371.5	P			0.6							
BDL81	10371.5	Av			1.27							
BDL81	10371.8	P			0.51							
BDL81	10371.8	Av			2.2							
BDL81	10372.1	P			0.54							
BDL81	10372.1	Av			2.43							
BDL81	10372.2	P		0.06	0.66							
BDL81	10372.2	Av		1.12	1.28							
BDL81	10374.1	P			0.42							
BDL81	10374.1	Av			1.99							
BDL81	10371.5	P				0.01	0.25		0.37	0.14	0.37	0.07
BDL81	10371.5	Av				1.13	1.18		1.14	1.13	1.14	1.11
BDL81	10371.8	P		0.2	0.05	0.02	0.33		0.23	0.19	0.23	0.04
BDL81	10371.8	Av		1.12	1.1	1.1	1.19		1.24	1.18	1.24	1.07
BDL81	10372.1	P		0.05								
BDL81	10372.1	Av		1.1								
BDL81	10372.2	P		0.33	0.4							0.59
BDL81	10372.2	Av		1.13	1.29							1.18
BDL81	10373.2	P		0.01								
BDL81	10373.2	Av		1.19								
BDL81	10374.1	P					0.06		0.02	0.23	0.02	0.04
BDL81	10374.1	Av					1.35		1.48	1.14	1.48	1.15
BDL85	10411.1	P				0.01	0.12		0.12	0.26	0.12	0.07
BDL85	10411.1	Av				1.09	1.29		1.28	1.12	1.28	1.16
BDL85	10411.3	P		0.59								
BDL85	10411.3	Av		1.51								
BDL85	10412.2	P		0.62								
BDL85	10412.2	Av		1.6								
BDL85	10414.1	P	0.1									
BDL85	10414.1	Av	1.03									
BDL85	10414.2	P										0.62
BDL85	10414.2	Av										1.19

Table 26. Results of the greenhouse experiments. Provided are the measured values of each parameter [parameters (Par.) 21-30 according to the parameters described in Table 23 above] in plants expressing the indicated polynucleotides. "Ev" = event; "P" = P-value; "Av" = ratio between the averages of event and control. Note that when the average ratio is higher than "1" the effect of exogenous expression of the gene is an increase of the desired trait;

175

Table 27
Results from greenhouse experiments

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>31</i>	<i>32</i>	<i>33</i>	<i>34</i>	<i>35</i>	<i>36</i>	<i>37</i>
BDL102	10471.1	P	0.12						
BDL102	10471.1	Av	1.12						
BDL102	10474.1	P	0.26						
BDL102	10474.1	Av	1.35						
BDL117	10074.1	P				0.69			
BDL117	10074.1	Av				1.1			
BDL117	10074.4	P				0.45			
BDL117	10074.4	Av				1.14			
BDL138	9811.1	P		0.06	0.11		0.22		
BDL138	9811.1	Av		1.18	1.11		1.1		
BDL138	9811.4	P		<0.01	0.19		0.48		
BDL138	9811.4	Av		1.21	1.11		1.2		
BDL138	9813.1	P		0.1			0.02		
BDL138	9813.1	Av		1.17			1.05		
BDL138	9813.4	P						0.01	
BDL138	9813.4	Av						1.08	
BDL138	9811.1	P					0.26	0.58	
BDL138	9811.1	Av					1.15	1.13	
BDL138	9811.4	P		<0.01	0.02				
BDL138	9811.4	Av		1.18	1.18				
BDL138	9812.1	P		0.01	0.03		0.1		
BDL138	9812.1	Av		1.13	1.1		1.07		
BDL138	9813.1	P		0.02	0.09				
BDL138	9813.1	Av		1.09	1.08				
BDL138	9813.3	P		0.09	0.03				
BDL138	9813.3	Av		1.17	1.16				
BDL140	10423.1	P	0.71						
BDL140	10423.1	Av	1.11						
BDL147	10301.5	P		0.01					
BDL147	10301.5	Av		1.1					
BDL147	10301.6	P		0.06	0.02				
BDL147	10301.6	Av		1.14	1.09				
BDL147	10303.1	P		0.16				0.66	
BDL147	10303.1	Av		1.11				1.14	
BDL147	10303.5	P		0.01	0.01				
BDL147	10303.5	Av		1.13	1.11				
BDL147	10303.6	P						0.1	
BDL147	10303.6	Av						1.13	
BDL149	9824.4	P	0.01						
BDL149	9824.4	Av	1.19						
BDL152	10434.4	P	0.8						
BDL152	10434.4	Av	1.13						
BDL153	10143.1	P				0.65		0.52	
BDL153	10143.1	Av				1.1		1.19	
BDL153	10143.2	P				0.26			0.05
BDL153	10143.2	Av				1.1			1.02
BDL155	9991.5	P	0.56						
BDL155	9991.5	Av	1.23						
BDL155	9991.9	P	0.1						
BDL155	9991.9	Av	1.31						
BDL155	9993.2	P	0.62						

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>31</i>	<i>32</i>	<i>33</i>	<i>34</i>	<i>35</i>	<i>36</i>	<i>37</i>
BDL155	9993.2	Av	1.1						
BDL155	9994.3	P	0.47						
BDL155	9994.3	Av	1.14						
BDL157	9911.3	P		0.02	0.33				
BDL157	9911.3	Av		1.32	1.19				
BDL157	9911.4	P				0.64			
BDL157	9911.4	Av				1.14			
BDL157	9913.1	P		0.01	<0.01				
BDL157	9913.1	Av		1.19	1.08				
BDL157	9913.3	P		0.11					
BDL157	9913.3	Av		1.17					
BDL157	9914.2	P		0.13			0.4		
BDL157	9914.2	Av		1.18			1.1		
BDL157	9911.3	P		0.15					
BDL157	9911.3	Av		1.13					
BDL157	9913.1	P						0.36	
BDL157	9913.1	Av						1.15	
BDL157	9913.3	P						0.03	
BDL157	9913.3	Av						1.06	
BDL157	9914.2	P				0.22			
BDL157	9914.2	Av				1.18			
BDL162	10492.2	P	0.1						
BDL162	10492.2	Av	1.8						
BDL167	10042.3	P		0.14					
BDL167	10042.3	Av		1.15					
BDL167	10042.4	P		0.01	0.01	0.19	0.01		
BDL167	10042.4	Av		1.15	1.05	1.1	1.06		
BDL167	10043.1	P		0.01	0.04		<0.01		
BDL167	10043.1	Av		1.16	1.06		1.1		
BDL167	10043.3	P		0.04	0.08		0.01		
BDL167	10043.3	Av		1.15	1.05		1.06		
BDL167	10044.2	P					0.52		
BDL167	10044.2	Av					1.11		
BDL167	10043.2	P				0.44	0.07		
BDL167	10043.2	Av				1.22	1.08		
BDL167	10043.3	P				<0.01			
BDL167	10043.3	Av				1.2			
BDL167	10043.4	P				0.22		0.2	
BDL167	10043.4	Av				1.21		1.2	
BDL167	10044.2	P				0.1			0.05
BDL167	10044.2	Av				1.22			1.03
BDL168	9881.3	P				<0.01			
BDL168	9881.3	Av				1.27			
BDL168	9881.4	P					0.4	0.42	
BDL168	9881.4	Av					1.21	1.2	
BDL168	9882.1	P						0.11	
BDL168	9882.1	Av						1.39	
BDL168	9882.3	P						0.18	
BDL168	9882.3	Av						1.32	
BDL169	10744.2	P	0.57						
BDL169	10744.2	Av	1.16						
BDL169	10747.1	P	0.46						
BDL169	10747.1	Av	1.11						
BDL169	10747.5	P	0.28						

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>31</i>	<i>32</i>	<i>33</i>	<i>34</i>	<i>35</i>	<i>36</i>	<i>37</i>
BDL169	10747.5	Av	1.16						
BDL171	10661.2	P	0.7						
BDL171	10661.2	Av	1.23						
BDL171	10664.1	P	0.6						
BDL171	10664.1	Av	1.39						
BDL173	9951.2	P	0.66						
BDL173	9951.2	Av	1.13						
BDL173	9952.1	P				0.21	0.12		
BDL173	9952.1	Av				1.23	1.14		
BDL173	9952.2	P		0.14		0.64	0.34		0.04
BDL173	9952.2	Av		1.1		1.15	1.11		1.04
BDL173	9953.4	P		0.58		<0.01	0.47	0.06	
BDL173	9953.4	Av		1.13		1.41	1.1	1.1	
BDL173	9954.2	P		0.12	0.05	0.22			
BDL173	9954.2	Av		1.17	1.08	1.1			
BDL173	9954.5	P		<0.01	0.04	0.57			0.07
BDL173	9954.5	Av		1.21	1.09	1.13			1.03
BDL176	9891.2	P		0.01	<0.01	0.31	0.01		<0.01
BDL176	9891.2	Av		1.17	1.06	1.22	1.14		1.06
BDL176	9891.4	P		<0.01	0.05				
BDL176	9891.4	Av		1.23	1.11				
BDL176	9892.3	P		<0.01	<0.01				0.09
BDL176	9892.3	Av		1.34	1.17				1.03
BDL176	9893.2	P		0.08	0.16	0.25			
BDL176	9893.2	Av		1.23	1.15	1.13			
BDL176	9893.3	P		<0.01		0.01	0.12		
BDL176	9893.3	Av		1.2		1.36	1.22		
BDL177	10521.3	P	0.03						
BDL177	10521.3	Av	1.43						
BDL177	10524.2	P	0.22						
BDL177	10524.2	Av	1.28						
BDL183	9941.1	P					0.58		
BDL183	9941.1	Av					1.1		
BDL183	9942.1	P					0.08		
BDL183	9942.1	Av					1.11		
BDL183	9943.4	P				<0.01			
BDL183	9943.4	Av				1.25			
BDL183	9944.2	P						0.29	
BDL183	9944.2	Av						1.24	
BDL190	10232.2	P		0.27		0.54			
BDL190	10232.2	Av		1.14		1.13			
BDL190	10233.2	P		0.38					0.03
BDL190	10233.2	Av		1.13					1.04
BDL190	10233.4	P		0.03		<0.01	0.05		
BDL190	10233.4	Av		1.13		1.34	1.12		
BDL190	10234.1	P		0.21	0.03	0.53			
BDL190	10234.1	Av		1.16	1.03	1.19			
BDL190	10231.1	P				0.07	0.07	0.01	
BDL190	10231.1	Av				1.06	1.09	1.15	
BDL190	10231.2	P						0.33	
BDL190	10231.2	Av						1.4	
BDL190	10232.2	P				0.04			
BDL190	10232.2	Av				1.08			
BDL190	10233.2	P						0.25	

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>31</i>	<i>32</i>	<i>33</i>	<i>34</i>	<i>35</i>	<i>36</i>	<i>37</i>
BDL190	10233.2	Av						1.25	
BDL192	9921.1	P		0.22	0.11				0.07
BDL192	9921.1	Av		1.18	1.1				1.03
BDL192	9922.1	P		0.01	0.04				
BDL192	9922.1	Av		1.15	1.06				
BDL192	9922.2	P		0.02	0.04			<0.01	
BDL192	9922.2	Av		1.21	1.09			1.12	
BDL192	9922.5	P				0.68			
BDL192	9922.5	Av				1.1			
BDL193	10152.2	P							0.08
BDL193	10152.2	Av							1.04
BDL196	10243.2	P						0.01	
BDL196	10243.2	Av						1.14	
BDL201	9963.6	P	0.15						
BDL201	9963.6	Av	1.28						
BDL201	9964.3	P	0.15						
BDL201	9964.3	Av	1.36						
BDL220	10333.5	P	0.35						
BDL220	10333.5	Av	1.19						
BDL223	10793.5	P	0.05						
BDL223	10793.5	Av	1.15						
BDL224	10451.8	P	0.02						
BDL224	10451.8	Av	1.5						
BDL225	10401.1	P	0.08						
BDL225	10401.1	Av	1.32						
BDL225	10401.4	P	0.01						
BDL225	10401.4	Av	1.52						
BDL225	10402.2	P	0.03						
BDL225	10402.2	Av	1.43						
BDL225	10402.5	P	0.26						
BDL225	10402.5	Av	1.39						
BDL225	10402.6	P	0.08						
BDL225	10402.6	Av	1.57						
BDL225	10402.9	P	0.12						
BDL225	10402.9	Av	1.36						
BDL226	10861.2	P	0.35						
BDL226	10861.2	Av	1.1						
BDL227	11492.3	P	0.05						
BDL227	11492.3	Av	1.15						
BDL233	10825.4	P	0.53						
BDL233	10825.4	Av	1.13						
BDL238	10954.2	P	<0.01						
BDL238	10954.2	Av	1.34						
BDL240	10802.2	P	0.23						
BDL240	10802.2	Av	1.3						
BDL241	10873.1	P	0.07						
BDL241	10873.1	Av	1.29						
BDL242	10731.2	P	0.62						
BDL242	10731.2	Av	1.1						
BDL242	10731.6	P	0.04						
BDL242	10731.6	Av	1.21						
BDL250	10846.3	P	0.53						
BDL250	10846.3	Av	1.15						
BDL48	10271.1	P	0.2						

<i>Gene</i>	<i>Ev.</i>	<i>Par</i>	<i>31</i>	<i>32</i>	<i>33</i>	<i>34</i>	<i>35</i>	<i>36</i>	<i>37</i>
BDL48	10271.1	Av	1.23						
BDL48	10271.3	P	0.08						
BDL48	10271.3	Av	1.48						
BDL48	10273.2	P	0.58						
BDL48	10273.2	Av	1.12						
BDL48	10274.4	P	0.01						
BDL48	10274.4	Av	1.95						
BDL48	10271.3	P		0.01	0.01				
BDL48	10271.3	Av		1.12	1.1				
BDL48	10271.5	P		0.06	0.1				
BDL48	10271.5	Av		1.06	1.05				
BDL48	10274.3	P						<0.01	
BDL48	10274.3	Av						1.44	
BDL48	10274.4	P						0.08	
BDL48	10274.4	Av						1.05	
BDL58	10282.3	P	0.63						
BDL58	10282.3	Av	1.19						
BDL63	10384.3	P	0.26						
BDL63	10384.3	Av	1.27						
BDL63	10381.1	P				<0.01	0.06		
BDL63	10381.1	Av				1.26	1.08		
BDL63	10381.2	P				0.56			
BDL63	10381.2	Av				1.14			
BDL63	10384.2	P				<0.01	0.04		
BDL63	10384.2	Av				1.27	1.08		
BDL63	10384.3	P		0.01	0.02	0.3			0.01
BDL63	10384.3	Av		1.12	1.09	1.15			1.04
BDL63	10384.7	P		0.05	0.06	0.08			
BDL63	10384.7	Av		1.07	1.07	1.15			
BDL63	10384.8	P				0.51			
BDL63	10384.8	Av				1.13			
BDL81	10371.5	P		<0.01	0.01	<0.01		0.23	0.04
BDL81	10371.5	Av		1.13	1.1	1.15		1.14	1.02
BDL81	10371.8	P				0.03			
BDL81	10371.8	Av				1.08			
BDL81	10372.1	P		<0.01	0.01				
BDL81	10372.1	Av		1.17	1.1				
BDL81	10372.2	P		0.22	0.1	0.59			
BDL81	10372.2	Av		1.11	1.11	1.14			
BDL81	10373.2	P		0.05					
BDL81	10373.2	Av		1.07					
BDL81	10374.1	P				0.05			
BDL81	10374.1	Av				1.2			
BDL85	10411.1	P	0.1						
BDL85	10411.1	Av	1.21						

Table 27. Results of the greenhouse experiments. Provided are the measured values of each parameter [parameters (Par.) 31-37 according to the parameters described in Table 23 above] in plants expressing the indicated polynucleotides. "Ev" = event; "P" = P-value; "Av" = ratio between the averages of event and control. Note that when the average ratio is higher than "1" the effect of exogenous expression of the gene is an increase of the desired trait;

EXAMPLE 10**PRODUCTION OF TOMATO TRANSCRIPTOM AND HIGH THROUGHPUT
CORRELATION ANALYSIS OF YIELD AND/OR VIGOR RELATED****5 PARAMETERS USING 44K TOMATO OLIGONUCLEOTIDE MICRO-ARRAY:
TOMATO FIELD EXPERIMENTS**

In order to produce a high throughput correlation analysis, the present inventors utilized a Tomato oligonucleotide micro-array, produced by Agilent Technologies. The array oligonucleotide represents about 44,000 Tomato genes and transcripts. In order to define correlations between the levels of RNA expression with yield components, ABST or vigor related parameters various plant characteristics of 18 different Tomato varieties were analyzed. Among them, 10 varieties encompassing the observed variance were selected for RNA expression analysis. The correlation between the RNA levels and the characterized parameters in field experiments was analyzed using Pearson correlation test.

Experimental procedures

Growth procedure in tomato field experiments - Tomato varieties were grown under normal conditions (4-6 Liters/m² per day) until flower stage.

RNA extraction – Leaves at different developmental stages, representing different plant characteristics, were sampled and RNA was extracted using TRIzol Reagent from Invitrogen.

Approximately 30-50 mg of tissue was taken from samples. The weighted tissues were ground using pestle and mortar in liquid nitrogen and resuspended in 500 µl of TRIzol Reagent. To the homogenized lysate, 100 µl of chloroform was added followed by precipitation using isopropanol and two washes with 75 % ethanol. The RNA was eluted in 30 µl of RNase-free water. RNA samples were cleaned up using Qiagen's RNeasy minikit clean-up protocol as per the manufacturer's protocol (QIAGEN Inc, CA USA).

Ripe fruit average weight (grams) - At the end of the experiment [when 50 % of the fruit were ripe (red)] all fruits from plots within blocks A-C were collected. The total fruits were counted and weighted. The average fruits weight was calculated by dividing the total fruit weight by the number of fruits.

Experimental Results

10 different Tomato varieties were grown and characterized for ripe fruit average weight (grams) as described above and the measured parameter [Ripe fruit average weight (gr.) at Normal Irrigation] is presented in Table 28 below.

5

Table 28
Measured parameters Tomato accessions

<i>Variety</i>	<i>Normal Irrigation; Ripe fruit average weight (gr.)</i>
612	0.05
613	0.01
617	0.01
618	0.05
622	0.01
623	0.01
626	0.03
629	0.00
630	0.00
631	0.01

10 Table 28: Provided are the measured yield components (Ripe fruit average weight under normal irrigation) for the tomato accessions (Varieties).

Subsequent correlation analysis between the leaf transcriptom set of BDL83_H74 Gene (SEQ ID NO:282) and the ripe fruit average weight under normal irrigation conditions was conducted and the correlation coefficient (R) was found to be
15 0.734.

20

25

EXAMPLE 11**PRODUCTION OF TOMATO TRANSCRIPTOM AND HIGH THROUGHPUT
CORRELATION ANALYSIS OF VIGOR RELATED PARAMETERS USING 44K
TOMATO OLIGONUCLEOTIDE MICRO-ARRAY**

5

Experimental procedures***Growth conditions for Tomato experiments***

Correlation of early vigor traits across collection of Tomato ecotypes – Ten Tomato varieties were grown in 3 repetitive plots, each containing 17 plants, at a net house under semi-hydroponics conditions. Briefly, the growing protocol was as follows: Tomato seeds were sown in trays filled with a mix of vermiculite and peat in a 1:1 ratio. Following germination, the trays were transferred to normal growth solution [full Hogland; KNO₃ - 0.808 grams/liter, MgSO₄ - 0.12 grams/liter, KH₂ PO₄ - 0.172 grams/liter and 0.01 % (volume/volume) of 'Super coratin' micro elements (Iron-EDDHA [ethylenediamine-N,N'-bis(2-hydroxyphenylacetic acid)]- 40.5 grams/liter; Mn - 20.2 grams/liter; Zn 10.1 grams/liter; Co 1.5 grams/liter; and Mo 1.1 grams/liter), solution's pH should be 6.5 – 6.8] at a temperature of 20-24 °C.

RNA extraction – All 10 selected Tomato varieties were sampled. Leaves from plant under Normal conditions were sampled and RNA was extracted using TRIzol Reagent from Invitrogen.

Tomato vigor related parameters – following 5 weeks of growing, plant were harvested and analyzed for leaf number. The analyzed data was saved to text files and processed using the JMP statistical analysis software (SAS institute).

Experimental Results

10 different Tomato varieties were grown and characterized for leaf number as described above. The average leaf number was calculated using the JMP software and values are summarized in Tables 29 below.

30

Table 29
Measured parameters Tomato accessions

<i>Variety</i>	<i>Leaf number</i>
1139	6.56
2078	6.89
2958	7.33
5077	6.22
5080	6.33
5084	6.44
5085	5.89
5088	5.56
5089	6.11
5092	5.67

5 Table 29. Provided are the measured vigor related parameter (leaf number) for the tomato accessions (Varieties).

Subsequent correlation analysis between the leaf transcriptom set of BDL83_H73 gene (SEQ ID NO:281) with the average leaf number was conducted, the correlation coefficient (R) was -0.794.

10 The genes identified herein improve plant yield in general, and more specifically oil yield, seed yield, oil content, plant growth rate, plant biomass, root measurements, and plant vigor. The output of the bioinformatics method described herein is a set of genes highly predicted to improve yield (seed yield, oil yield and content, biomass) and/or other agronomic important yields by modifying their
15 expression. Although each gene is predicted to have its own impact, modifying the mode of expression of more than one gene is expected to provide an additive or synergistic effect on the plant yield, plant growth rate, root measurements, plant vigor and/or other agronomic important yields performance. Altering the expression of each gene described here alone or set of genes together increases the overall yield plant
20 growth rate, root measurements, plant vigor and/or

Although the invention has been described in conjunction with specific embodiments thereof, it is evident that many alternatives, modifications and variations will be apparent to those skilled in the art. Accordingly, it is intended to embrace all
25 such alternatives, modifications and variations that fall within the spirit and broad scope of the appended claims.

Citation or identification of any reference in this application shall not be construed as an admission that such reference is available as prior art to the present invention. To the extent that section headings are used, they should not be construed as necessarily limiting.

What is claimed is:

1. A method of increasing yield, biomass, and/or growth rate of a plant, comprising:
 - (a) expressing within the plant an exogenous polynucleotide comprising a nucleic acid sequence at least 80% identical to the full-length sequence set forth by SEQ ID NO: 868, wherein said nucleic acid sequence has the same biological activity as SEQ ID NO: 868, and
 - (b) selecting plants resulting from step (a) for an increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a non-transformed plant of the same species, which is grown under the same growth conditions, thereby increasing the yield, biomass, and/or growth rate of the plant.

2. A method of increasing yield, biomass, and/or growth rate of a plant, comprising:
 - (a) expressing within the plant an exogenous polynucleotide comprising a nucleic acid sequence at least 80% identical to the full-length sequence set forth by SEQ ID NO: 27, wherein said nucleic acid sequence has the same biological activity as SEQ ID NO: 27, and
 - (b) selecting plants resulting from step (a) for an increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a non-transformed plant of the same species, which is grown under the same growth conditions, thereby increasing the yield, biomass, and/or growth rate of the plant.

3. A method of increasing yield, biomass, and/or growth rate, of a plant, comprising:
 - (a) expressing within the plant an exogenous polynucleotide comprising the nucleic acid sequence set forth in SEQ ID NO: 868, and
 - (b) selecting plants resulting from step (a) for an increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a non-transformed plant of the same species, which is grown under the same growth conditions, thereby increasing the yield, biomass, and/or growth rate of the plant.

4. A method of increasing yield, biomass, and/or growth rate of a plant, comprising:
 - (a) expressing within the plant an exogenous polynucleotide comprising the nucleic acid sequence set forth in SEQ ID NO: 27, and

(b) selecting plants resulting from step (a) for an increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a non-transformed plant of the same species, which is grown under the same growth conditions, thereby increasing the yield, biomass, and/or growth rate of the plant.

5. A method of increasing yield, biomass, and/or growth rate of a plant, comprising:

(a) expressing within the plant an exogenous polynucleotide comprising a nucleic acid sequence encoding a polypeptide at least 80% identical to the full-length sequence set forth by SEQ ID NO: 132, wherein said polypeptide has the same biological activity as SEQ ID NO: 132, and

(b) selecting plants resulting from step (a) for an increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a non-transformed plant of the same species, which is grown under the same growth conditions, thereby increasing the yield, biomass, and/or growth rate of the plant.

6. A method of increasing yield, biomass, and/or growth rate of a plant, comprising:

(a) expressing within the plant an exogenous polynucleotide comprising a nucleic acid sequence encoding the polypeptide set forth in SEQ ID NO: 132, and

(b) selecting plants resulting from step (a) for an increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a non-transformed plant of the same species, which is grown under the same growth conditions, thereby increasing the yield, biomass, and/or growth rate of the plant.

7. The method of any one of claims 1-4, further comprising selecting the plant resulting from step (a) for an increased rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to a non-transformed plant of the same species, which is grown under the same growth conditions.

8. The method of any one of claims 5-6, further comprising selecting the plant resulting from step (a) for an increased rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to a non-transformed plant of the same species, which is grown under the same growth conditions.

9. A method of growing a plant having increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a native plant of the same species which is grown under the same growth conditions, the method comprising:

(a) providing plants over-expressing a polypeptide comprising an amino acid sequence at least 80% identical to the full-length amino acid sequence set forth in SEQ ID NO: 132 as compared to a native plant of the same species, wherein said polypeptide has the same biological activity as SEQ ID NO: 132, (b) selecting said plants over-expressing said polypeptide for an increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a native plant of the same species which is grown under the same growth conditions, to thereby obtain plants over-expressing said polypeptide and having said increased yield, biomass, and/or growth rate, and

(c) growing a crop of said plants over-expressing said polypeptide and having said increased yield, biomass, and/or growth rate,

thereby growing the plant having the increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to the native plant of the same species which is grown under the same growth conditions.

10. A method of growing a plant having increased rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to a native plant of the same species which is grown under the same growth conditions, the method comprising:

(a) providing plants over-expressing a polypeptide comprising an amino acid sequence at least 80% identical to the full-length amino acid sequence set forth in SEQ ID NO: 132 as compared to a native plant of the same species, wherein said polypeptide has the same biological activity as SEQ ID NO: 132,

(b) selecting said plants for increased rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to a native plant of the same species which is grown under the same growth conditions, to thereby obtain plants over-expressing said polypeptide and having said increased rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area, and

(c) growing a crop of said plants over-expressing said polypeptide and having said increased rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area,

thereby growing the plant having the increased rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to the native plant of the same species which is grown under the same growth conditions.

11. A method of generating a transgenic plant, comprising:

(a) transforming a cell of a plant with a nucleic acid construct comprising an exogenous polynucleotide comprising a nucleic acid sequence encoding a polypeptide at least 80% identical to the full-length amino acid sequence set forth in SEQ ID NO: 132 and generating a mature plant from said cell of said plant, wherein said polypeptide has the same biological activity as SEQ ID NO: 132, and

(b) selecting a plant resulting from step (a) for an increased yield, biomass, and/or growth rate under non-stress growth conditions as compared to a non-transformed plant of the same species which is grown under the same growth conditions,

thereby generating the transgenic plant.

12. A method of generating a transgenic plant, comprising:

(a) transforming a cell of a plant with a nucleic acid construct comprising an exogenous polynucleotide comprising a nucleic acid sequence encoding a polypeptide at least 80% identical to the full-length amino acid sequence set forth in SEQ ID NO: 132 and generating a mature plant from said cell of said plant, wherein said polypeptide has the same biological activity as SEQ ID NO: 132, and

(b) selecting a plant resulting from step (a) for an increase in a trait selected from the group consisting of: rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and growth rate of leaf blade area under non-stress growth conditions as compared to a non-transformed plant of the same species which is grown under the same growth conditions,

thereby generating the transgenic plant.

13. A method of producing seeds of a crop plant, comprising:

(a) selecting a parent plant transformed with an exogenous polynucleotide encoding a polypeptide at least 80% identical to the full-length amino acid sequence set forth in SEQ ID NO: 132 for an increase in one or more traits selected from the group consisting of: rosette diameter, growth rate of plot coverage, growth rate of rosette diameter, growth rate of rosette area, and growth rate of leaf blade under non-stress growth conditions as compared to a non-transformed plant of the same species which is grown under the same growth conditions, wherein said polypeptide has the same biological activity as SEQ ID NO: 132,

(b) growing a seed-producing plant from said parent plant resultant of step (a), wherein said seed-producing plant which comprises said exogenous polynucleotide exhibits an increase in said one or more selected traits, and

(c) producing seeds from said seed-producing plant resultant of step (b),
thereby producing seeds of the crop plant.

14. The method of any one of claims 1 to 6, 9, and 11, wherein the yield is seed yield.

15. The method of any one of claims 1 to 6, 9, and 11, wherein the yield is oil yield.

16. The method of any one of claims 5 and 8-12, wherein said polypeptide is at least 85% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.

17. The method of any one of claims 5 and 8-12, wherein said polypeptide is at least 90% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.

18. The method of any one of claims 5 and 8-12, wherein said polypeptide is at least 95% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.

19. The method of any one of claims 5 and 8-12, wherein said polypeptide is at least 98% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
20. The method of any one of claims 5 and 8-12, wherein said polypeptide is at least 99% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
21. The method of any one of claims 5 and 8-12, wherein said polypeptide comprises the amino acid sequence set forth by SEQ ID NO: 132.
22. The method of any one of claims 5 and 8-12, wherein said polypeptide comprises the amino acid sequence set forth by SEQ ID NO: 132 or SEQ ID NO:632.
23. The method of claim 13, wherein said polypeptide is at least 85% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
24. The method of claim 13, wherein said polypeptide is at least 90% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
25. The method of claim 13, wherein said polypeptide is at least 95% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
26. The method of claim 13, wherein said polypeptide is at least 98% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
27. The method of claim 13, wherein said polypeptide is at least 99% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
28. The method of claim 13, wherein said polypeptide comprises the amino acid sequence set forth by SEQ ID NO: 132.
29. The method of claim 13, wherein said polypeptide comprises the amino acid sequence set forth by SEQ ID NO: 132 or SEQ ID NO:632.

30. The method of claim 1 or 2 or 5, wherein said nucleic acid sequence is as set forth in SEQ ID NO: 868.

31. The method of claim 1 or 2 or 5, wherein said nucleic acid sequence is as set forth in SEQ ID NO: 27.

32. The method of claim 5, wherein said nucleic acid sequence is as set forth in SEQ ID NO: 311.

33. The method of any one of claims 1-4, further comprising selecting the plant resulting from step (a) for an increased rosette diameter, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to a non-transformed plant of the same species which is grown under the same growth conditions.

34. The method of any one of claims 5-6, further comprising selecting the plant resulting from step (a) for an increased rosette diameter, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to a non-transformed plant of the same species which is grown under the same growth conditions.

35. A method of growing a plant having increased rosette diameter, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to a native plant of the same species which is grown under the same growth conditions, the method comprising:

(a) providing plants over-expressing a polypeptide comprising an amino acid sequence at least 80% identical to the full-length amino acid sequence set forth in SEQ ID NO: 132 as compared to a native plant of the same species, wherein said polypeptide has the same biological activity as SEQ ID NO: 132,

(b) selecting said plants for increased rosette diameter, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to a native plant of the same species which is grown under the same

growth conditions, to thereby obtain plants over-expressing said polypeptide and having said increased rosette diameter, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area, and

(c) growing a crop of said plants over-expressing said polypeptide and having said increased rosette diameter, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area,

thereby growing the plant having the increased rosette diameter, growth rate of rosette diameter, growth rate of rosette area, and/or growth rate of leaf blade area under non-stress growth conditions as compared to the native plant of the same species which is grown under the same growth conditions.

36. A method of generating a transgenic plant, comprising:

(a) transforming a cell of a plant with a nucleic acid construct comprising an exogenous polynucleotide comprising a nucleic acid sequence encoding a polypeptide at least 80% identical to the full-length amino acid sequence set forth in SEQ ID NO: 132 and generating a mature plant from said cell of said plant, wherein said polypeptide has the same biological activity as SEQ ID NO: 132, and

(b) selecting a plant resulting from step (a) for an increase in a trait selected from the group consisting of: rosette diameter, growth rate of rosette diameter, growth rate of rosette area, and growth rate of leaf blade area under non-stress growth conditions as compared to a non-transformed plant of the same species which is grown under the same growth conditions, thereby generating the transgenic plant.

37. The method of any one of claims 34-36, wherein said polypeptide is at least 85% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.

38. The method of any one of claims 34-36, wherein said polypeptide is at least 90% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.

39. The method of any one of claims 34-36, wherein said polypeptide is at least 95% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.

40. The method of any one of claims 34-36, wherein said polypeptide is at least 98% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
41. The method of any one of claims 34-36, wherein said polypeptide is at least 99% identical to the full-length amino acid sequence set forth by SEQ ID NO: 132.
42. The method of any one of claims 34-36, wherein said polypeptide comprises the amino acid sequence set forth by SEQ ID NO: 132.
43. The method of any one of claims 34-36, wherein said polypeptide comprises the amino acid sequence set forth by SEQ ID NO: 132 or SEQ ID NO:632.
44. The method of any one of claims 16-22, wherein the yield is seed yield.
45. The method of any one of claims 16-22, wherein the yield is oil yield.

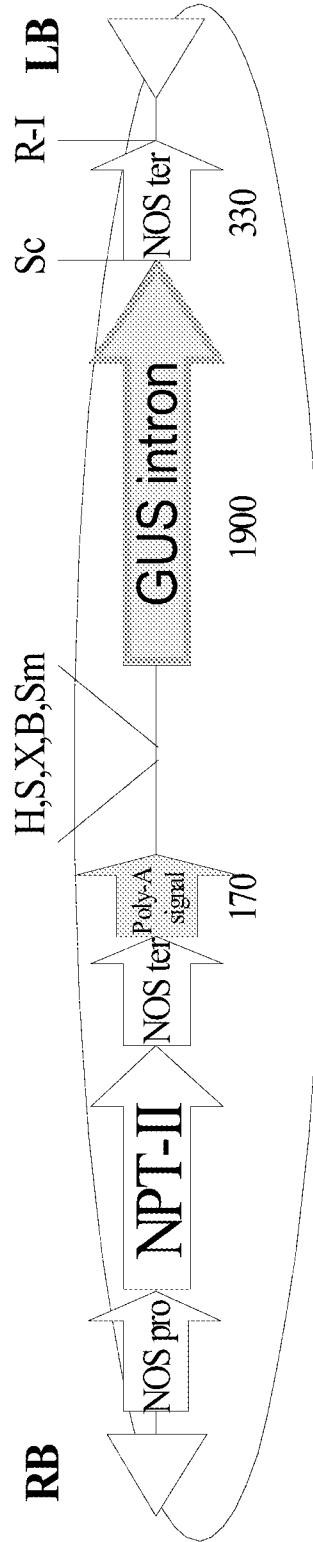


FIG. 1

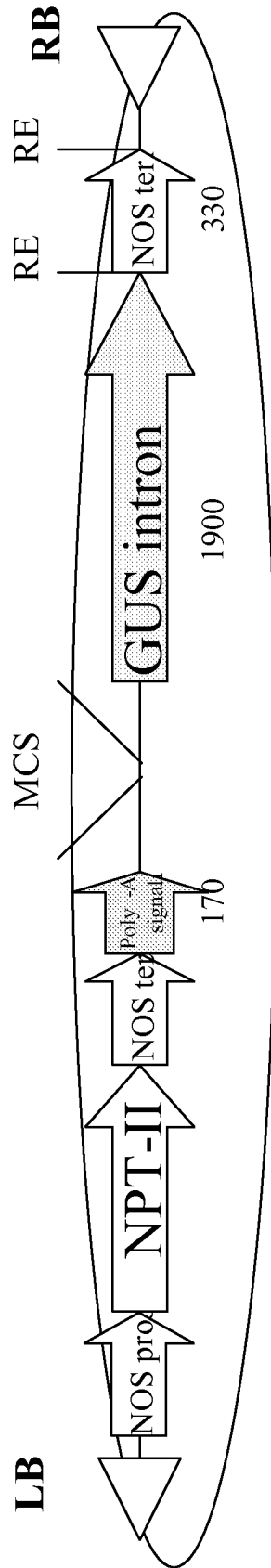


FIG. 2

Nitrogen limiting
conditions

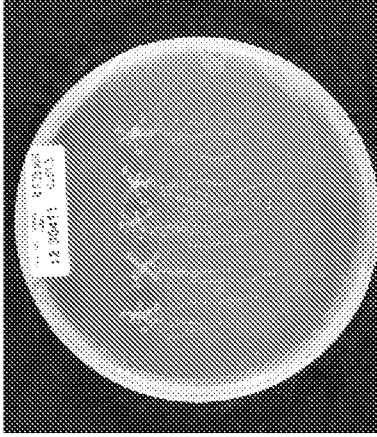


FIG. 3E

Osmotic stress
(15 % PEG)

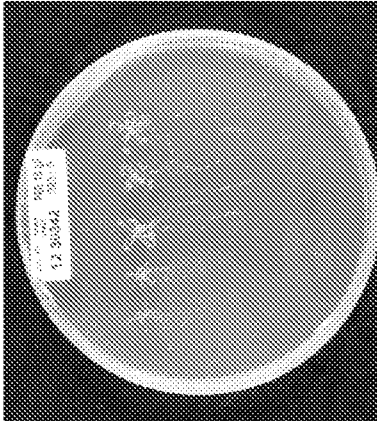


FIG. 3C

Normal conditions

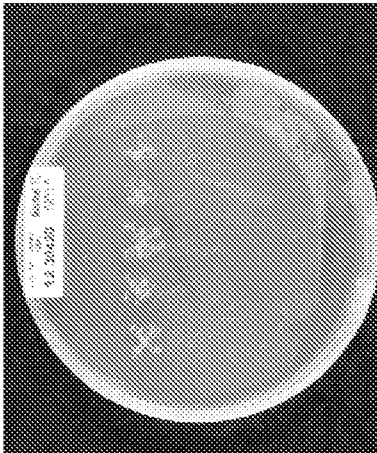


FIG. 3A

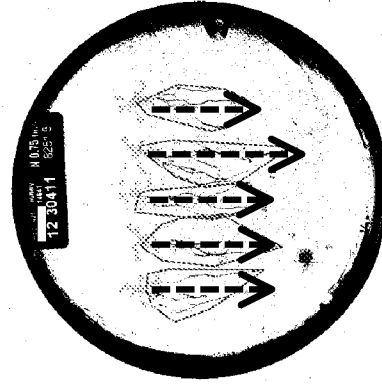


FIG. 3F

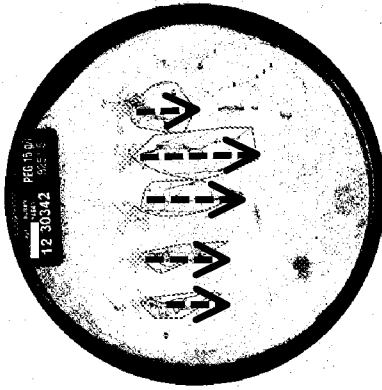


FIG. 3D

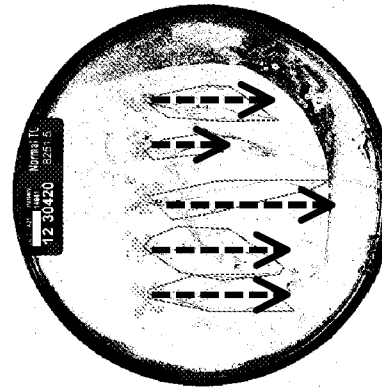


FIG. 3B