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(54) Title: CANINE ALLERGY THERAPEUTIC RECOMBINANT CHIMERIC ANTI-IGE MONOCLONAL ANTIBODY

(57) Abstract: The present invention provides methods and compositions for decreasing IgE levels in dogs. The methods and compositions are useful for treating allergic symptoms in dogs. The invention may comprise chimeric canine anti-IgE monoclonal antibody compositions and methods for using the compositions in the treatment of allergy. In preferred embodiments, the compositions of the present invention may act by binding soluble IgE in plasma, or by inhibiting IgE from binding to receptors on mast cells, B cells, and basophils.

CANINE ALLERGY THERAPEUTIC RECOMBINANT CHIMERIC ANTI-IGE
MONOCLONAL ANTIBODY

This application is a continuation of U.S. Application Serial No. 60/179,629, filed February 1, 2000.

FIELD OF THE INVENTION

The present invention provides compositions and methods for decreasing IgE levels and for alleviating allergic symptoms in canines. The compositions comprise chimeric canine anti-IgE mAbs and the methods are useful for treating allergies in canines.

BACKGROUND ART

It is estimated that up to 30% of all dogs suffer from allergies or allergy-related skin disorders. Specifically, allergic dermatitis has been estimated to affect between 3% and 15% of the entire canine population. Given the prevalence of allergies in dogs, there is a need to develop methods and compositions to properly diagnose and treat canine allergies.

The substances most likely to cause an allergic reaction vary from species to species. Common canine allergens include fleas, pollens, molds and dust. Allergy to fleas is believed to be the most common dog allergy.

Typically, a flea's saliva is the allergen, and a single fleabite can cause substantial itching. An additional form of allergy in dogs is termed atopy. Atopy is a condition where a dog is allergic to inhalants such as pollens, molds or microscopic mites found in house dust. Current treatments of canine allergies often focus on the use of steroids which cause undesirable side effects or allergen-mediated desensitization which requires a different treatment for each type of allergy.

In mammals, antibody molecules are classified into various isotypes referred to as IgA, IgD, IgE, IgG, and IgM. Antibody molecules consist of heavy and light chain components. The heavy chains of molecules of a given isotype have extensive regions of amino acid sequence homology, and conversely have regions of difference from antibodies belonging to other isotypes. The shared regions of the heavy chains provide members of each isotype with common abilities to bind to certain cell surface receptors or to other macromolecules such as complement, and therefore to activate particular immune effector functions. Accordingly, separation of antibody molecules into isotypes also serves to separate the antibodies according to a set of effector functions that they commonly activate. In humans

and dogs, Immunoglobulin E (IgE) is involved in allergy, and recognizes antigen in immediate hypersensitivity reactions.

Furthermore, IgE is the antibody type that is understood to be an important mediator of allergic response in mammals, including Type I immediate hypersensitivity. IgE molecules bind to receptors on mediator cells such as mast cells. This binding occurs when the Fc region of the IgE molecule is bound to Fc receptors on the mast cells. When such cell-bound IgE antibodies then bind to an allergen, the allergen cross-links multiple IgE antibodies on the mast cell surface. This cross-linking mediates Type I immediate hypersensitivity reactions and causes release of histamines and other molecules that produce symptoms associated with allergy.

In humans the serum level of total IgE is diagnostic of allergic disease. To explore the possibility that the serum level of IgE might also be diagnostic of allergy in dogs, DeBoer and Hill performed additional studies. (Hill and DeBoer, *Am. J. Vet. Res.* 55 (7):944-48 (1994)). They used monoclonal antibody ("mAb") D9 in an ELISA assay with the following configuration: D9 was bound to a substrate, antibodies were captured by D9 and then D9 having a marker was used to flag the captured antibody.

The Hill and DeBoer ELISA was used to establish the total amount of IgE in canine serum in an effort to diagnose canine allergy. However, it was found that quantifying IgE was not useful for diagnosing allergy in dogs. (See, e.g., Abstract and Discussion sections of Hill and DeBoer.) This finding was in direct contrast to the situation in human immunology. This result points out the difficulty of any attempt to correlate data between animals of two different genera.

This difficulty is further exemplified by the fact that dogs can be allergic to a different set of antigens than humans are. Allergies to fleas, for instance, are a severe problem for dogs but not humans. Furthermore, in instances where dogs and humans appear to be allergic to the same allergen extract, studies by doctors Esch and Greer of Greer Laboratories (Lenoir, NC), have indicated that the specific allergens in an allergen extract which produce canine disease are not necessarily the same allergens that produce disease in humans. For example, it is known that the immunodominant components of dust mite extracts are different in dogs than in humans.

Adding to the difficulty of study across genera of allergic mechanisms and response is that allergies in dogs are primarily expressed in the skin, while humans primarily

exhibit allergic symptoms in the respiratory system. Additionally, eosinophilia is correlated to allergies in humans, but not in dogs.

In considering the administration of a therapeutic composition to treat a physiological condition, when recombinant or chimeric molecules are administered *in vivo* to an animal, they may be quickly cleared from that animal. Recombinant IgG molecules have been used to increase the half-life of recombinant molecules when the recombinants are administered to an animal. *E.g.*, Capon, D., Chamow, S., et al., "Designing CD4 immunoadhesins," *Nature* 337:525 (1989); Byrn, R., Mordenti, C., et al., "Biological Properties of a CD4 Immunoadhesin," *Nature* 344:667 (1990); Haak-Frendscho, M., Ridgway, J., et al., "Human IgE Receptor Alpha-Chain IgG Chimera Blocks Passive Cutaneous Anaphylaxis Reaction *in vitro*," *Journal of Immunology* 151:351-53 (1993); U.S. Patent No. 5,116,964, issued May 26, 1992 to Capon, D. J., et al. entitled "Hybrid Immunoglobulins".

It is generally accepted that in humans IgG is the immunoglobulin isotype with the longest serum half-life. In dogs, the isotype with the longest half-life is not known. Although the sequences that are believed to correspond to a portion of exon 1 and 3 of at least two and possibly all four heavy chain canine IgG immunoglobulin sequences have

been reported in U.S. Patent No. 5,593,861 to Maeda et al., it is not known which of these heavy chain sequences is part of the IgG structure with the longest half-life in dogs.

IgE levels are elevated in human patients experiencing allergic disease, and IgE is believed to mediate allergic symptoms. Although the levels of serum IgE may not correlate with allergic disease in dogs, it may nevertheless be desirable to decrease IgE levels as a mechanism for alleviating allergic symptoms.

Furthermore, there is a need for compositions and methods for treatment of canine allergies which avoid the disadvantages of the conventional compositions and methods, yet provide effective treatment for canine allergies.

One object of the present invention is to provide compositions and methods for treatment of canine allergies, with substantially less side effects than those experienced with steroid treatments.

Another object of the invention is to provide compositions and methods of treatment for alleviating canine allergy symptoms that are effective independent of the type of allergen, and compositions and methods where treatment is based on the presence of an allergic response rather than a specific allergen.

Another object of the present invention is to provide compositions and methods of treatment for alleviating canine allergy symptoms by targeting IgE synthesis.

These and other objects will be apparent to those skilled in the art from the following disclosure and appended claims.

SUMMARY OF THE INVENTION

The present invention concerns compositions and methods for treating allergy in dogs. More particularly, the invention provides methods and compositions for administration to dogs, which compositions actually bind the dog's immunoglobulin E molecules so that the binding of free, serum IgE will inhibit this IgE from binding to the high affinity IgE receptor on mast cells and basophils. The compositions and methods provided may eliminate or reduce levels of free serum IgE. Lower free serum IgE levels may down regulate the synthesis and expression of the high affinity IgE receptor on basophils and mast cells. The result may be the reduction or elimination of free and/or total serum IgE and the reduction or elimination of the IgE response to allergen on skin mast cells. By "free serum IgE" is meant that IgE which is able to bind to the high affinity IgE receptor, and is unbound IgE in serum.

We have demonstrated that sustained elimination of detectable free and/or total serum IgE for 7 days and possibly a shorter time period results in a negative, feedback-loop that continues to suppress IgE synthesis. Sustained suppression of IgE synthesis will result in the elimination of a skin response to allergen.

In a preferred embodiment, the specificity and structure of a chimeric anti-IgE molecule of the present invention may allow for direct targeting of the IgE+ B cell. This binding may result in a reduction or elimination of IgE synthesis either through negative stimulation of the mature B cell or by destruction of the B cell by apoptosis or complement-mediated lysis.

The present invention may therefore comprise an IgE receptor molecule which comprises a chimera and which specifically binds to canine IgE. The receptor molecule may be an antibody molecule, preferably a monoclonal antibody ("mAb"), and the mAb may preferably have an affinity for exon 3 of canine IgE. The chimera may comprise canine and mouse immunoglobulin. The chimera may further comprise canine constant heavy and light domains fused to mouse heavy and light chain variable regions. The receptors and antibody molecules of the present invention may also comprise IgG heavy chain sequences.

The receptors and antibody molecules of the present invention may prevent binding of IgE to a second IgE receptor and the second IgE receptor may be located on one or more of a mast cell or basophil. The receptors and antibody molecules of the present invention may be comprised of protein, peptides, or other organic molecules.

The present invention also provides methods of treating canine allergies comprising administering to the canine a receptor or monoclonal antibody which comprises a chimera of the present invention and specifically binds to canine IgE. The methods of the present invention may result in a lowering of serum IgE levels in the treated canine, or in the binding of IgE on B cells and the subsequent elimination of clonal populations of B cells. The methods may also result in binding of serum IgE in plasma, or in an inhibition of IgE production in the treated canine. The lowering of serum IgE levels of the present methods may be caused by a disruption or blocking of interactions between IgE and receptors for IgE which may be located on mast cells or basophils.

The present invention further provides pharmaceutical formulations containing therapeutic amounts of the receptors of the present invention.

DESCRIPTION OF FIGURES

Figure 1 depicts the ability of chimeric 15A.2 to inhibit IgE from binding to the recombinant canine IgE receptor.

Figure 2(a & b) depict time course data for (free and total IgE) circulating IgE levels in 2 control and 3 experimental dogs following administrations of a recombinant chimeric anti-IgE mAb designated c15A.2. The mAb 15A.2 and its specificity is disclosed in the pending patent application Serial No. 09/281,760, filed on March 30, 1999.

Figure 3 depicts time course data for c15A.2 activity in dogs following a course of administration of chimeric antibody.

Figure 4 depicts time course data for circulating IgE complexed with c15A.2 in 3 dogs following administration of 8 courses of chimeric antibody.

Figure 5 depicts time course data for anti-chimeric 15A.2 activity observed in the serum of the experimental dogs over the course of administration of the chimeric antibody.

Figure 6 depicts flow cytometric data from a dog, double-stained with PE-labeled anti-exon 4 canine IgG mAb 14K.2 and FITC-labeled anti-canine IgE receptor mAb 9L.4 four days following a first course, three days following a

second course and 5 days following a 5th course of administration of chimeric anti-15A.2 mAb.

Figure 7 depicts time course data for Ragweed skin test reactivity in dogs prior to, and 3 days and 7 days following eight courses of administration of c15A.2 mAb.

Figure 8 depicts time course data for circulating free IgE levels in dogs 1001 and 1002 following administrations of a recombinant receptor-IgG antagonist designated cRcIg.

Figure 9 depicts free and total IgE in experimental dogs 1001 and 1002 over the time course of the experiment.

Figure 10 depicts the DNA sequence (SEQ. ID NO. 5) of the recombinant IgE receptor cRcIg with corresponding translation (SEQ. ID NO. 6). The four additional amino acids at the bottom of the figure that lack the corresponding nucleotides are SEQ ID NO 7.

DETAILED DESCRIPTION OF THE INVENTION

Definitions:

Affinity: an attractive force or binding strength between a ligand and its receptor or between two binding moieties. Affinity also keeps the binding pair bound in equilibrium.

Amino acids: Organic molecules containing an amino group that can be combined in linear arrays to form polypeptides, peptides or proteins. The 20 common amino

acids are alanine, arginine, asparagine, aspartic acid, cysteine, glutamic acid, glutamine, glycine, histidine, isoleucine, leucine, lysine, methionine, phenylalanine, proline, serine, threonine, tryptophan, tyrosine, and valine. It will be recognized by those skilled in the art that in embodiments of the invention with conservative variants, any of the common amino acids, as well as others not listed, may be used in the present invention.

Canine IgE Receptor: A recombinant or chimeric receptor as defined herein, which exhibits affinity for canine IgE, or otherwise has the result of removing IgE from canine sera, *in vivo* or *in vitro*.

cDNA clone: A duplex DNA sequence representing an RNA, carried in a cloning vector.

Chimeric mAb: An immunoglobulin molecule with a hybrid amino acid sequence that has resulted from combining amino acids from at least two different canine Ig sources. Generally, the amino acid sequences are not normally found together in nature. "mAb" indicates monoclonal antibody.

Cloning vector: A plasmid, phage DNA or other DNA sequence, able to replicate in a host cell and capable of carrying exogenously added DNA sequence for purposes of amplification or expression of the added DNA sequence.

Conservative variant: Conservative variants of nucleotide sequences include nucleotide substitutions that do not result in changes in the amino acid sequence, as well as nucleotide substitutions that result in conservative amino acid substitutions, or amino acid substitutions which do not substantially affect the character of the polypeptide translated from said nucleotides. For example, polypeptide character is not substantially affected if the substitutions do not preclude specific binding of the peptide to canine IgE receptor or other canine IgE ligands.

Conservative variants of amino acid sequences include amino acid substitutions or deletions that do not substantially affect the character of the variant polypeptide relative to the starting peptide. For example, polypeptide character is not substantially affected if the substitutions or deletions do not preclude specific binding of the variant peptide to a specific binding partner of the starting peptide. Included in this definition are glycosylated and other variants and derivatives that will be apparent to those skilled in the art and are considered to fall within the scope of this invention. Also included in this definition are amino acid insertions, substitutions, deletions and truncations that do not substantially affect the polypeptide character relative to the starting peptide.

Expression control sequence: A DNA sequence of nucleotides that controls and regulates expression of structural genes when operatively linked to those genes.

Exon: A contiguous region of DNA encoding a portion of a polypeptide. Reference to any exon, e.g. "DNA sequence of exon 6", refers to the complete exon or any portion thereof.

Free IgE or Serum IgE: IgE in circulation in a patient not complexed or bound with a native or administered receptor having affinity for IgE or other IgE molecules.

Genome: The entire DNA of a substance. It includes, among other things, the structural genes encoding for the polypeptides of the substance, as well as operator, promoter and ribosome binding and interaction sequences such as the Shine-Dalgarno sequences.

Specific binding: Binding of one substance to another at greater binding affinity than background binding. Two substances that exhibit specific binding are referred to as specific binding partners, or as a specific binding pair. An antibody and its antigen are one example of a specific binding pair.

Structural gene: A DNA sequence that encodes through its template or messenger RNA ("mRNA") a sequence of amino acids characteristic of a specific polypeptide.

Therapeutic amount: A "therapeutic amount" is an amount of mAb that decreases serum IgE levels or suppresses IgE production or activity in the treated animal and has the effect of ameliorating or preventing symptoms of a disorder or physiological condition.

Total IgE: By "total IgE" is meant total serum IgE, whether bound or not by another molecule.

As disclosed herein, a novel chimeric canine anti-IgE mAb was produced and administered to ragweed sensitized dogs. This chimeric molecule, designated c15A.2, consists of canine constant heavy and light domains fused to mouse heavy and light variable regions. In all cases, such administration led to a sustained decrease in circulating free IgE levels to below the level of detectability. In addition, total IgE, including that complexed with chimeric 15A.2 and still in circulation and detectable immediately after administration of c15A.2, was reduced over time. After 28 days of administration, free c15A.2 was detectable in serum, but both free and total serum IgE were undetectable. The use of a humanized anti-IgE monoclonal antibody as a therapeutic agent for human allergy was disclosed in U.S. Patent No. 5,593,861 to Maeda et al. It is believed that IgE removal by administration of

recombinant anti-IgE mAb has not previously been shown in dogs. The person of ordinary skill will realize that the present invention and the discussion herein pertaining to mAb is equally applicable to receptors which may comprise parts of antibody molecules. Therefore, the discussion herein is intended to relate to compositions of receptors, not solely antibodies.

The canine anti-IgE mAb compositions of the present invention may be recombinant or chimeric structures of canine and mouse immunoglobulins. The molecules may encompass chimera made from canine IgG constant heavy and light domains and murine immunoglobulin heavy and light variable region domains that have an affinity for exon 3 of canine IgE.

The terms anti-canine IgE mAb, chimeric mAb, recombinant mAb, and receptor as used herein encompass any and all conservative variants thereof and are considered to be within the scope of the present invention.

It is a feature of the present invention that administration of chimeric canine anti-IgE mAb or receptor is useful in the method of treatment of canine allergies. It is also a feature of the invention that administration of compositions of the present invention lowers serum IgE levels in canines.

The chimeric canine anti-IgE mAbs and receptors of the present invention are capable of disrupting or blocking the interaction between IgE and its receptors. Generally, the interference between IgE and its receptors is independent of the type of allergen causing or likely to cause allergic symptoms.

In embodiments of the present invention, chimeric canine anti-IgE mAbs and receptors of the present invention may act by blocking the binding of IgE to its receptors on mast cells or basophils by blocking the binding site on IgE molecules or otherwise interfering with the binding of IgE to its receptor. The chimeric canine anti-IgE mAbs and receptors of the present invention may also act by binding soluble IgE in plasma which complex is then removed from circulation by the body's normal mechanisms. In other embodiments of the present invention, the chimeric canine anti-IgE mAbs or receptors may act by binding IgE on B-cells and eliminating clonal populations of IgE+ B-cells. The chimeric canine anti-IgE mAbs or receptors of the present invention may also act by inhibiting IgE production. While not wanting to be bound by any particular theory, it is believed that the mAb or receptor may bind to B cells and induce a cross-linking event which may induce apoptosis of cells or lead to inhibitory signals which down-regulate or

eliminate IgE synthesis. Additionally, the binding of serum IgE, free IgE that is, may allow another regulatory molecule which may normally bind serum IgE in the exon 3 region to bind to IgE on the B cell and effect, subsequently, IgE synthesis through negative signals associated with and resulting from such binding.

In another embodiment, the chimeric canine anti-IgE mAbs or receptors of the present invention may comprise or be formulated with IgG heavy chain sequences to enhance the half-life of the molecules *in vivo* or increase their activity.

The method of treating dogs suffering allergic symptoms or of preventing allergic symptoms in dogs generally may comprise the administration of a therapeutic amount of a chimeric canine anti-IgE mAb or receptor to the treated animal. Precise dosages of chimeric canine anti-IgE mAb or receptor and administration parameters will be established in a manner consistent with that known to those of ordinary skill in the art. This may involve taking into account one or more of the following factors, although this list is intended to be representative and is not intended to be exclusive of other parameters which may be known or become known to those of ordinary skill in the art: the presence and severity of allergic symptoms, the species of dog, the

condition of the individual patient, the site of delivery, the method and length of administration, and other factors known to those of ordinary skill in the art or which may become known in the future.

Similarly, the dose of the chimeric canine anti-IgE mAb or receptor administered may be dependent on consideration of the properties of the IgE heavy chain isotypes used and other factors. For example, these considerations may include the binding activity and *in vivo* plasma half-life, the concentration of the chimeric canine anti-IgE mAb or receptor in the formulation, the administration route, the site and rate of dosage, and the clinical tolerance of the patient involved. This list is not intended to be limiting and the person of ordinary skill in the art will realize that other factors may also be relevant and advantageous to consider.

The therapeutic amount of the instant chimeric canine anti-IgE mAb or receptor may be administered in dosages and for a period of time sufficient to alleviate or suppress allergic symptoms and/or to decrease serum IgE levels and/or suppress IgE production or activity.

In general, the formulations of the present invention may contain other components in amounts that do not interfere with the preparation of stable and efficacious

forms of the canine anti-IgE mAb or receptor. Any additional components administered with the chimeric canine anti-IgE mAb or receptor may be present in amounts suitable for effective, safe pharmaceutical administration. Pharmaceutical excipients known to those of ordinary skill in the art may form a part of the subject compositions. For example, such excipients may include saline and other parenteral solutions, buffers and stabilizers, as well as any of the various suitable bulking agents, buffering agents, antioxidants, cosolvents and other ingredients known to those of ordinary skill in the art as being advantageous to include. In a preferred embodiment, the canine anti-IgE mAb or receptor may be formulated as a solution of protein in sterile PBS.

The chimeric canine anti-IgE mAb or receptor of the present invention may be administered in a way which is effective for disrupting, blocking or otherwise interfering with binding between IgE and its receptor or in a way which may enhance binding between the administered mAb or receptor and IgE. These methods will be apparent to those of ordinary skill in the art. In preferred embodiments, the chimeric canine anti-IgE mAb or receptor may be administered subcutaneously, intramuscularly or intravenously. Alternatively, the mAb or receptor may be formulated and

administered via suspensions, tablets, capsules or suppositories for oral, rectal or vaginal administration.

The following examples further illustrate the cloning, expression, and purification of the 15A.2 mAb and are not intended to be limiting.

Example I: Cloning of mouse 15A.2 variable region

Mouse monoclonal 15A.2 variable region was cloned by RT-PCR from 15A.2 hybridoma cells. A commercially available kit (Novagen, Madison, WI, Ig-Prime Kit) that consists of a set of degenerate PCR primers for the reverse transcription and amplification of IgVh and IgVl mRNA's from mouse hybridoma cell lines was used as a source of primers to clone the 15A.2 mRNA's encoding the light and heavy variable domains. The 5' and 3' primers for the IgVh domain was MuIgVh5'-B and MuIgMvh3'-1 respectively. MuIgVh5'-B is a mixture of two primers in one tube (provided by Novagene), having the sequences GGGAATTCATGRAATGSASCTGGGTYWTYCTCTT (SEQ. ID NO. 1) and ACTAGTCGACATGGACTCCAGGCTCAATTTAGTTTTTCCT (SEQ. ID NO. 2). MuIgMvh3'-1 has the sequence CCCAAGCTTACGAGGGGAAGACATTTGGGAA. The 5' and 3' primers for the IgVl domain was MuIgλV15'-A and MuIgλV13'-1 respectively. The mRNA encoding the 15A.2 IgVh and IgVl variable domains was reverse transcribed, amplified and cloned. MuIg(lambda)V15'-A has the sequence GGGAATTCATGGCCTGGAYTYCWCTYWTMYTCT. MuIg(lambda)V13'-1 has the sequence CCCAAGCTTAGCTCYTCWGWGGAIGGYGGRAA.

The PCR reactions were performed as follows:

Enzyme: Taq polymerase

- | | | | |
|----|-------|------------|-----------|
| 1) | 94 °C | 20 seconds | 35 cycles |
| | 60 °C | 58 seconds | |
| | 72 °C | 20 seconds | |
| 2) | 4 °C | Storage | |

Suitable clones of the 15A.2 IgVh and IgVl variable domains were identified using restriction endonuclease analysis and DNA sequence analysis. Comparison of these DNA sequences to known mouse IgVh and IgVl genes verified them as coding for the variable domains of a mouse monoclonal antibody.

Example II: Cloning of Canine IgG Constant Region

The canine immunoglobulin light and heavy constant regions were cloned by RT-PCR from dog lymphocyte cells. PCR primers based on the sequence of the constant domain of canine IgG was used for the reverse transcription and PCR amplification of mRNA encoding the immunoglobulin constant domains (disclosed in U.S. Patent No. 5,593,861 to Maeda et al.). The PCR reaction was the same as listed above. PCR products were cloned and subjected to DNA sequence analysis. Suitable clones of the immunoglobulin constant domains were identified using restriction endonuclease analysis and DNA sequence analysis. Comparison of these DNA sequences to known immunoglobulin genes verified them as coding for the constant domains of a canine immunoglobulin.

Example III: Cloning of full length mouse/canine chimeric 15A.2

The conventional process for the preparation of chimeric monoclonal antibodies was used to construct a full

length mouse/canine chimeric 15A.2 monoclonal antibody genes. The mouse 15A.2 variable region encoding sequence and the appropriate canine constant region coding sequence were linked together via PCR and cloned. After verification of the chimeric genes by DNA sequence analysis, a suitable mouse/canine chimeric 15A.2 light chain gene and mouse/canine 15A.2 heavy chain gene was selected for expression and protein production of the chimeric protein.

Identification of functional clones

Functional clones of the 15A.2 chimeric heavy and light chains were identified using a COS cell transient expression system. The full-length heavy chain and the full-length light chain were cloned downstream in correct orientation of the CMV promoter on the pcDNA3.1 vector (Invitrogen, Carlsbad, CA). The immunoglobulin leader signals on the proteins would cause the proteins to be secreted out of the cell into culture supernatant. Co-transfection of both DNAs into COS cells allowed transient gene expression, protein production and assembly of functional chimeric antibodies in the cell. IgE binding ELISA and anti-canine IgG ELISA were used to detect functional antibody activity in cell culture supernatants. The clones which gave the best binding activity were used to construct vectors for chimeric monoclonal antibody production in a baculovirus expression system. The sequences of both heavy chain and light chain are shown as follows:

Chimeric 15A.2 heavy chain DNA sequence (SEQ. ID NO. 3)

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ATGAAATGGAGCTGGGTTTTTCTCTTTCTCCTGTCAGTAACTGCGGGTGTGTTCT
CTGAGGTTTCAGCTGCAGCAGTCTGGACCTGAGCTGGTGAAGCCTGGGGCTTCAGTGAAGA
TATCCTGCAAGGCTTCTGGTTACTCATTACTGACTACTTTATGAACTGGGTGATGCAGA
GCCATGGAAAGAGCCTTGAGTGGATTGGTTCGTATTAATCCTTTCAATGGTGATCCTTTCT
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ACAACCAGAAGTTCAAGGGCAAGGCCACATTGACTGTAGACAAATCCTCTAGCACAGCCC
ACATGGAGCTCCGGAGCCTGGCATCTGAGGACTCTGCAGTCTATTATTGTGCAAGATTCT
ACTACGGACGTTACTATGCTATGGACTACTGGGGTCAAGGAACCTCAGTCACCGTCTCCT
CAGCCTCCACCACGGCCCCCTCGGTTTTCCCACTGGACCCCAGCTGCGGGTCCACTTCCG
GCTCCACGGTGGCCCTGGCCTGCCTGGTGTGAGGCTACTTCCCCGAGCCTGTAAGTGTGT
CCTGGAATTCCGGCTCCTTGACCAGCGGTGTGCACACCTTCCCGTCCGACCTGCAGTCTC
CAGGGCTCTACTCCCTCAGCAGCATGGTGACAGTGCCCTCCAGCAGGTGGTCCAGCGAGA
CCTTCACCTGCAACGTGGCCCACCCGGCCAGCAAACTAAAGTAGACAAGCCAGTGCCCA
AAAGAGAAAATGGAAGAGTTCCTCGCCCACCTGATTGTCCCAAATGCCAGCCCCCTGAAA
TGCTGGGAGGGCCTTCGGTCTTCATCTTTCCCCGAAACCCAAGGACACCCTCTTGATTG
CCCGAACACCTGAGGTACATGTGTGGTGGTGGATCTGGGACCAGAAGACCCTGAGGTGC
AGATCAGCTGGTTCGTGGACGGTAAGCAGATGCAAACAGCCAAGACTCAGCCTCGTGAGG
AGCAGTTC AATGGCACCTACCGTGTGGTCACTGTCTCCCCATTGGGCACCAGGACTGGC
TCAAGGGGAAGCAGTTCACGTGCAAAGTCAACAACAAAGCCCTCCCATCCCCGATCGAGA
GGACCATCTCCAAGGCCAGAGGGCAGGCCCATCAGCCCAGTGTGTATGTCTGCGCCCAT
CCCGGGAGGAGTTGAGCAAGAACACAGTCAGCTTGACATGCCTGATCAAAGACTTCTTCC
CACCTGACATTGATGTGGAGTGGCAGAGCAATGGACAGCAGGAGCCTGAGAGCAAGTACC
GCACGACCCCGCCCCAGCTGGACGAGGACGGGTCTACTTCTGTACAGCAAGCTCTCTG
TGGACAAGAGCCGCTGGCAGCGGGGAGACACCTTCATATGTGCGGTGATGCATGAAGCTC
TACACAACCACTACACACAGAAATCCCTCTCCCATTCTCCGGGTAAATGA

Chimeric 15A.2 light chain DNA sequence (SEQ. ID NO. 4)

ATGGCCTGGATTTCACTCTTATTCTCTCTCCTGGCTCTCAGCTCAGGGGCCATTT
CCCAGGCTGTTGTGACTCAGGAATCTGCACTCACCACATCACCTGGTCAAACAGTCACAC
TCACTTGTGCTCAAGTACTGGGGCTGTTACAAGTAACTATGCCAACTGGGTCCAAG
AAAACCAGATCATTATTCACTGGTCTAATAGGTGGTCCCAACAACCGAGCTCCAGGTG
TTCCTGCCAGATTCTCAGGCTCCCTGATTGGAGACAAGGCTGCCCTCACCATCACAGGGG
CACAGACTGAGGATGAGGCAATATATTTCTGTGCTCTATGGTACAGCAACCATTGGGTGT
TCGGTGGAGGAACCAAAGTACTGTCTAGGCCAGCCCAAGGCCTCCCCCTCGGTACAC
TCTTCCCGCCCTCCTCTGAGGAGCTCGGCGCAACAAGGCCACCCTGGTGTGCCTCATCA
GCGACTTCTACCCAGCGGCGTGACGGTGGCCTGGAAGGCAAGCGGCAGCCCCGTACCC
AGGGCGTGGAGACCACCAAGCCCTCCAAGCAGAGCAACAACAAGTACGCGGCCAGCAGCT

ACCTGAGCCTGACGCCTGACAAGTGGAAATCTCACAGCAGCTTCAGCTGCCTGGTCACGC
 ACGAGGGGAGCACCGTGGAGAAGAAGGTGGCCCCCGCAGAGTGCTCTTAG

Example IV: Expression of Chimeric 15A.2 in Insect Cells

A baculovirus expression system was used for larger scale production of the chimeric 15A.2 mouse/canine monoclonal antibody. Baculovirus expression is a common technique and the methods are well known to one of ordinary skill in the art. The 15A.2 heavy chain DNA was cloned into PharMingen's (San Diego, CA) pAc LIC baculovirus transfer vector for recombination into baculovirus. Chimeric 15A.2 light chain DNA was cloned into PharMingen's pAHis NT-A™ baculovirus transfer vector for recombination into baculovirus. Recombination and amplification of both virus constructs were amplified in insect sf-9 cells. The chimeric 15A.2 was expressed using insect High Five cells. The infection conditions were as following:

High Five cell density for infection:	1.5 x 10 ⁶ /ml
MOI for heavy chain virus infection:	10
MOI for light chain virus infection:	3
Time of protein expression:	72 hours

15A.2 was expressed and secreted into the cell culture media.

Purification of chimeric 15A.2 protein

The general purification scheme is as following:

1. Cell clarification:

The cell culture supernatant containing 15A.2 protein was harvested 72 hours after infection. The supernatant was

filtered through Millipore's (Bedford, MA) MILLIGUARD™ cartridge filtration system for cell clarification. Other filtration systems which have a pore size of 0.2 μm and are high capacity filters with low protein binding, and are able to withstand high pressure will also be suitable. The clarified supernatant was then concentrated for column chromatography.

2. Protein A column chromatography:

The concentrated sample was loaded onto a protein A column equilibrated with PBS buffer. The 15A.2 protein was eluted by pH gradient generated by mixing 4% glycerol, PBS, pH 7.2 and 4% glycerol, 25 mM sodium citrate buffer, pH 2.5. The protein was eluted at about pH 4.

3. Ion-exchange column chromatography:

The protein A purified 15A.2 was loaded onto the Q Sepharose™ (Pharmacia, Uppsala, Sweden) column equilibrated with 4% glycerol, PBS, pH 7.2 to remove contaminated proteins, DNA, RNA, virus, etc. Similar ion exchange resin which remove DNA, RNA, endotoxin and other negatively charged materials from purified proteins may also be utilized. The flow-through was collected.

4. Sterilization:

The Q flow-through 15A.2 was concentrated and sterilized by filtering through 0.2 μm filter unit. The final buffer component is as following:

0.7 x PBS
6.5 mM sodium citrate
4% glycerol
pH: - 7.0

The protein has at least 95% purity as indicated by SDS-PAGE protein gel and stored at - 80 °C for future use.

**Example V: Recombinant Canine anti-IgE mAb, (c15A.2):
Effects on IgE Activity *in vitro* and *in vivo***

Purified c15A.2 as obtained from Example IV was tested for its ability to prevent the binding of canine IgE to the canine IgE receptor *in vitro* (Figure 1). The amount of c15A.2 required to neutralize IgE in the sera of three dogs designated 2002, 2003 and 2103 was determined by the serum neutralization assay and is shown in Figure 8. The time course and events for the 40 day experimental period are summarized in Figures 2a and 2b. Dogs were sensitized to ragweed at birth at Lovelace Respiratory Research Institute (Albuquerque, NM) and used in studies to test the efficacy assessed by the dogs' ability to develop a subcutaneous skin response following administration of allergen.

The c15A.2 was administered to dogs 2002, 2003 and 2103 every five days for eight consecutive treatments. The amount of administered c15A.2 in the first treatment was equivalent to 10x the concentration of measurable free serum IgE concentration three days prior to the start of administration. Doses 1 and 2 received 10x free serum IgE concentration. All subsequent dosages were set at 5x free serum IgE concentration. The test article was diluted into

sterile PBS at the concentration required to deliver 10x or 5x serum IgE concentration by intravenous infusion over a 30 minute period.

Figure 2a follows the level of free and total serum IgE in each dog versus time, based upon dosages of c15A.2 mAb administered. Additional c15A.2 was then administered every five days for a total of 40 consecutive days. Serum IgE levels are assayed by standard ELISA techniques that are well known to those of ordinary skill in the art. IgE levels are generally considered to be undetectable in the ELISA technique below about 1 ng/ml.

Figure 2a also shows that free IgE in the experimental dogs dropped to undetectable levels within 60 minutes and did not return during the entire 40 day-treatment period. The levels of free serum IgE in control dogs 1101 and 1102 receiving only saline, did not change significantly during this period (Figure 2b). The assay used in these experiments consisted of an ELISA solid phase capture with mouse 15A.2 antibody and detection with another HRPO-conjugated mAb, 14K.2 which recognizes exon 4 of canine IgE. The assay detects all serum IgE not bound by c15A.2.

Total IgE bound by c15A.2 dropped more slowly than free IgE in experimental dogs presumably because it recirculated and was eventually cleared from circulation. The assay used

here consisted of an ELISA capture with mAb 14K.2 and detection with another anti-canine IgE mAb, which is not inhibited from binding by c15A.2. In dogs 2003 and 2103 free and total serum IgE has remained undetectable in assays for greater than 60 days following the last dose of administration of recombinant antibody. In dog 2002, 30 days after administration of chimeric antibody was halted, free serum IgE was detectable at about 200 ng/ml and remained at this level for the remaining 30 days of observation.

These data show the effects of a recombinant antibody of the present invention in: (1) clearing the serum of circulating IgE; and (2) providing a method for affecting the process by which IgE is replenished. These results are surprising given that serum IgE is understood to have a short half-life, probably on the order of about two days, and that recombinant or chimeric molecules may likewise exhibit relatively short half-lives *in vivo*. In untreated animals, circulating serum IgE is believed to be entirely replenished every 4 to 5 days, yet following administration of a canine anti-IgE mAb 15A.2 of the present invention, serum IgE levels remained suppressed below the level of detection 18 days after administration of the chimeric antibody.

Example VI: Clearance of the c15A.2

The presence of free chimeric 15A.2 in the serum of dogs 2002, 2003 and 2103, was determined by ELISA using a recombinant canine IgE solid phase with detection by a polyclonal goat anti-canine IgG conjugate. Figure 3 shows that the levels of c15A.2 in serum rise and fall consistently with the beginning and ending of each 5-day administration period. Free c15A.2 is still detectable (X µg/ml) in the serum of experimental dogs 24 days post-infusion.

Immune complexes of c15A.2 and canine IgE were measured by ELISA on a 14K.2 solid phase and detected by polyclonal anti-dog IgGfc conjugate. These data, summarized in Figure 4, show that complexed IgE is detected at high concentration early in the time course of treatment, but the level of complexes consistently falls over time. By day 28 no immune complexes are detected in dogs receiving c15A.2. This result suggests that the complexes are cleared from the circulating pool of serum immunoglobulin and that synthesis of new IgE in this time frame is reduced or eliminated. No changes were observed in control dogs. No immune response to the chimeric monoclonal antibody c15A.2 was observed in dogs 2003 and 2103. At 28 days post primary infusion an immune response to the test article was observed in dog 2002. Data in Figure 5 show that this response increases as more chimeric 15A.2 is administered and may play a role in the shorter serum half-life of c15A.2 in dog 2002 (Figure 3).

Example VII: Effect on Level of Expression of the High Affinity IgE Receptor on Basophils upon Administration of Canine anti-IgE mAb 15A.2 (c15A.2)

At five days, 14 days, and 29 days following the start of the c15A.2 administration 10 ml of whole blood was removed. Semi-purified populations of peripheral blood leukocytes were prepared by density gradient centrifugation and stained with reagents for two-dimensional flow cytometric analyses. The reagents used in these experiments were FITC-conjugated, anti-canine high-affinity IgE receptor mAb 9L.4 and PE-conjugated 14K.2. The cells double staining for these antibodies and resident in quadrant four (upper right quadrant) of the dot blot diagrams in Figure 6 are basophils.

Data in Figure 6 are for dog 2002 and are representative. The number of double staining cells in quadrant decreases over the time course of administration of c15A.2. The data suggest that the level of expression of high affinity IgE receptors in basophils is reduced by over 90% after 29 days of administration of c15A.2 to the experimental dog 2002. Elimination of serum IgE over time leads to a reduction in expression of receptors for IgE on basophils and may reflect a similar response in skin mast cells. A reduced mast cell expression of high affinity IgE

receptors will lead to a reduction or elimination of skin test reactivity to allergen.

Example VIII: Effect on Ragweed Skin Test Reactivity of Administration of Canine Anti-IgE mAb 15A.2

Ragweed sensitized dogs were infused with a 0.5% solution in PBS of Evans Blue Dye at 0.2 ml/kg and then challenged with allergen 10 minutes later. Five, 10-fold serial dilutions of ragweed allergen in PBS starting at 1000 PMU/ml were prepared and 100 μ l injected subcutaneously in a shaved section of the torso. Saline (PBS) served as a negative control and histamine (100 μ l of an 0.275 μ g/ml dilution of histamine in PBS) served as a positive control. Reactivity was measured 10 minutes later as the swelling and blue-dye diffusion in the skin compared with the histamine control. A rating of two was given to an allergen dilution in which the reaction was equivalent in size and color to the histamine reaction. A rating of 1 was given for a reaction that was half of the histamine control and a rating of zero was given when the reaction was equivalent to the saline negative control. Three experimental, 2002, 2003 and 2103 and two control, 1101 and 1102, dogs were skin tested as described above 1 week prior to administration of c15A.2, and then at 3 days and 7 days following the last

administration. Figure 7 shows that in control dogs the skin response to ragweed allergen was similar throughout the study. In control dog 1102 the ragweed skin response actually increased over time. In experimental dogs 2003 and 2103 the ragweed sensitivity after administration of c15A.2 for 40 days was completely absent for at least 7 days. Only dog 2002 showed any ragweed sensitivity at the highest ragweed concentration on day 3 post-infusion and that response was given a rating of 1 because of the spread of dye. It did not have the characteristic swelling seen in a 1 rating observed and recorded for the control dogs. At 7 days post-infusion for dog 2002 a response rating of 1 was given to the spots with 1000 PMU/ml, 100 PMU/ml and 10 PMU/ml. Again there was only color diffusion and no swelling. Given the fact that dog 2002 had no detectable free or total IgE during this skin-testing period, it seems likely that the skin reactions observed may not have been due to cross-linking of IgE on the mast cell high affinity IgE receptors. They may have been the result of some other skin reaction associated with the testing procedure.

Example IX: Chimeric Canine anti-IgE mAb Engineered for Long Serum Stability, Loss of Immunogenicity and for IgE+ B-cell Targeting

The person of ordinary skill in the art will realize that one may also provide the canine anti-IgE mAb of the present invention in a form which comprises a greater portion of the molecule as Ig taken from the dog sequence, and therefore should be expected to have greater serum stability, loss of immunogenicity and more effectively target IgE+ B cells.

Therefore, in another example a chimeric canine anti-IgE mAb that is engineered for long serum stability and an inability to induce an immune response against it is administered to a dog. This mAb binds to IgE in serum and to IgE on the surface of IgE-producing B-cells but not to IgE on mast cells or basophils. IgE synthesis is consequently reduced or eliminated. The resulting reduced IgE levels cause reduction in mast cell IgE receptor expression and reduction in associated allergic reactivity.

Example X: Recombinant Canine IgE Receptor Designated cRcIg
- Cloning and Expression in Insect Cells

The sequence that is believed to correspond to at least part of the α -subunit of the canine IgE receptor is set forth in the GENBANK database under Assession No. D16413.

The cRcIg receptor is comprised of the alpha domain of the receptor which binds IgE linked to exons 2 and 3 of

canine IgE. The IgG heavy chain sections of the receptor are the same as exons 2 and 3 of the c15A.2.

DNA encoding the chimeric IgE receptor cRcIg was introduced into a baculovirus genome using standard procedures. High five insect cell cultures were infected with baculovirus producing cRcIg. The cRcIg protein was purified from the cell culture medium by chromatography on protein A, followed by removal of contaminating proteins by ion exchange chromatography. The DNA sequence of the recombinant IgE receptor cRcIg with corresponding translation is shown in Figure 10.

The IgE receptor used in the study was a recombinant chimeric receptor-body, named cRcIg, composed of the soluble high affinity IgE receptor alpha unit fused with the canine IgG CH2 and CH3 domain. The soluble alpha subunit, which lacks the trans-membrane domain, was cloned by PCR using the information available in GeneBank. The IgG CH2 and CH3 domain was part of sequence of DE94 as claimed by the Chemo-Sero patent. The soluble receptor and DE94 CH2/CH3 were connected by PCR to form the full length cRcIg.

The full length cRcIg was cloned into PharMingen's baculovirus pAChistNA vector. The virus was amplified using sf-9 cells. The cRcIg protein was produced in a secret form and was expressed in High five cells for 48 hours with MOI

(multiplicity of infection) at 5. The protein was purified based on the CH2/CH3 domain's binding to Protein A column. Detailed purification scheme is the same as that of 15A.2 as illustrated in Example IV.

The purified cRcIg was concentrated to 2.9 mg/ml, and sterilized by filtering through 0.2 μ m filter unit. The final buffer component is as follows:

0.7 x PBS
6.5 mM sodium citrate
15% glycerol
pH: about 7.0

The protein has at least 95% purity as indicated by SDS-PAGE protein gel and stored at -80°C for future use.

Example XI: Recombinant Canine IgE Receptor Effects on IgE Activity *in vitro* and *in vivo*

Serum IgE level was determined by ELISA as described previously. Purified cRcIg as obtained from Example X was tested for its ability to prevent the binding of canine IgE to the canine IgE receptor *in vitro*. The amount of cRcIg required to neutralize serum IgE was measured by titration of purified cRcIg against dog sera *in vitro*. Purified cRcIg was then administered through I.V. injection as a bolus.

The recombinant canine IgE receptor cRcIg was therefore administered intravenously as a bolus to dogs 1001 and 1002. This amount of cRcIg was equivalent to 10x (141 mgs; 7.08 ml @ 2 mg/ml in dog 1002) or 20x (220 mgs; 11.04 ml @ 2 mgs/ml in dog 1001) the amount needed for 50% neutralization of recombinant receptor binding *in vitro*. Additional cRcIg was then administered at the dosage of 5 ml @ 2 mg/ml as a bolus to each dog for five consecutive days, on days 13-17. Figure 9 follows the level of free serum IgE in each dog versus time. In both dogs, IgE returned rapidly to the circulation after the initial dose of cRcIg on Day 0. Serum IgE levels are generally assayed by standard ELISA techniques well known to those of ordinary skill in the art. IgE levels are generally considered to be undetectable in the ELISA technique below about 1 ng/ml.

In dog 1001, serum IgE was undetectable by assay for greater than two months following the last dose of recombinant IgE receptor. In dog 1002, serum IgE levels returned to detectable levels over a period of days following the final administration of the recombinant receptor. It is to be noted however, that although serum IgE levels increased in dog 1002 they did not return to the elevated levels which were present prior to the administration of the recombinant receptor on day 0. Figure 10 shows total and free serum IgE levels in each of the dogs during the course of the experiment. These data show that circulating IgE was not immediately removed from serum. Free

serum IgE is unavailable but total IgE, presumably that complexed with cRcIg, was still in circulation.

These data document the effects of a recombinant receptor of the present invention in: (1) clearing the serum of circulating IgE; and (2) providing a method for affecting the process by which IgE is replenished.

Example XII: Peptides Which Bind to Canine IgE and Prevent Binding to the IgE Receptor and/or Bind to IgE on B cells and Affect Synthesis of IgE

The person of ordinary skill in the art will realize that peptides may be derived from combinatorial peptide libraries and comprise sequences of amino acids which when combined with IgE prevent binding to the IgE receptor. They are, then, peptides of any sequence of amino acids capable of binding to IgE, preferably exon 3 of IgE, and of preventing this IgE from binding to the IgE receptor. They may also bind to IgE on B cells.

Therefore, in another example peptides which bind to canine IgE are administered to a dog. The peptides bind IgE in serum and IgE on the surface of IgE-producing B-cells but not IgE on mast cells or basophils. IgE synthesis is reduced or eliminated. The resulting reduced IgE levels cause reduction in mast cell IgE receptor expression and reduction in associated allergic reactivity.

Example XIV: Small Molecules Which Bind to Canine IgE and Prevent Binding to the IgE Receptor and/or Bind to IgE on B Cells and Affect Synthesis of IgE

The person of ordinary skill in the art will realize that small organic molecules may be derived from combinatorial libraries and are screened in assays whereby the small organic molecules are isolated by their ability to inhibit IgE from binding to the IgE receptor. They, therefore may bind to IgE, preferably at a region within exon 3, and inhibit binding to the IgE receptor.

In another example a small molecule is administered to a dog. The small molecule binds IgE in serum and IgE on the surface of IgE-producing B-cells but not IgE on mast cells or basophils. IgE synthesis is consequently reduced or eliminated. The resulting reduced IgE levels cause reduction in mast cell IgE receptor expression and reduction in associated allergic reactivity.

Closing

It must be noted that as used herein and in the appended claims, the singular forms "a," "and," and "the" include plural referents unless the context clearly dictates otherwise. Thus, for example, reference to "a formulation" includes mixtures of different formulations and reference to "the method of treatment" includes reference to equivalent

steps and methods known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar to or equivalent to those described herein can be used in the practice or testing of the invention, the preferred methods and materials are described herein. All publications mentioned herein are fully incorporated herein by reference.

5 CLAIMS

1. An exogenous receptor molecule which specifically binds to canine IgE.
2. The receptor molecule of claim 1 wherein the receptor molecule is an antibody.
- 10 3. The antibody of claim 2 wherein the antibody is a monoclonal antibody.
4. The receptor molecule of claim 1 wherein the receptor molecule is a chimera of canine and mouse immunoglobulin.
- 15 5. The receptor molecule of claim 4 comprising a chimera of canine constant heavy and light domains fused to mouse heavy and light chain variable regions.
6. The receptor molecule of claim 1 wherein the
20 receptor molecule has an affinity for exon 3 of canine IgE.
7. The receptor molecule of claim 8 wherein the antibody is C15A.2.
8. The receptor molecule of claim 1, wherein the
25 receptor molecule prevents binding of IgE to a second receptor and wherein the second receptor is located on a cell selected from one or more of the group consisting of: B cells, mast cells, and basophils.
- 30 9. The receptor molecule of claim 1 wherein the receptor is an organic molecule.

- 5 10. The receptor molecule of claim 9 wherein the receptor is a protein or a peptide.
11. The receptor molecule of claim 10 wherein the receptor molecule further comprises at least a portion of an IgG heavy chain.
- 10 12. The receptor molecule of claim 1 wherein the receptor molecule comprises a chimeric monoclonal antibody which binds to IgE, and wherein the monoclonal antibody is a chimera of canine and mouse immunoglobulin.
- 15 13. The monoclonal antibody of claim 12 wherein the monoclonal antibody comprises canine constant heavy and light domains fused to mouse heavy and light chain variable regions.
14. The monoclonal antibody of claim 13 wherein the
20 monoclonal antibody has an affinity for exon 3 of canine IgE.
15. The monoclonal antibody of claim 14 wherein the monoclonal antibody prevents binding of IgE to a receptor and wherein the receptor is located on a
25 cell selected from one or more of the group consisting of: B cells, mast cells, and basophils.
16. A method of treating canine allergies comprising:
administering to the canine an exogenous
30 receptor molecule which specifically binds to IgE.

- 5 17. The method of claim 16 wherein the receptor molecule is a chimera of canine and mouse immunoglobulin.
18. The receptor molecule of claim 17 wherein the receptor is a monoclonal antibody.
- 10 19. The method of claim 18 wherein the receptor molecule comprises a chimera of canine constant heavy and light domains fused to mouse heavy and light chain variable regions.
- 15 20. The method of claim 16 wherein the receptor molecule has an affinity for exon 3 of canine IgE.
21. The method of claim 20 wherein the receptor molecule is C15A.2.
- 20 22. The method of claim 16 wherein the administration of the receptor molecule results in a lowering of serum IgE levels in the treated canine.
23. The method of claim 16 wherein the administration of the receptor molecule results in binding of IgE on B cells and the subsequent elimination of clonal populations of B cells.
- 25 24. The method of claim 16 wherein the administration of the receptor molecule results in the binding of IgE in plasma.
- 30 25. The method of claim 16 wherein the administration of the receptor molecule results in an inhibition of IgE production in the treated canine.

- 5 26. The method of claim 16 wherein the receptor molecule further comprises IgG heavy chain sequences.
27. The method of claim 22 wherein the lowering of serum IgE levels is caused by disruption or
10 blocking of interactions between IgE and receptors for IgE.
28. The method of claim 27 wherein the receptors for IgE are located on mast cells.
29. The method of claim 27 wherein the receptors for
15 IgE are located on basophils.
30. A pharmaceutical formulation for treating allergies in a canine, comprising:

a therapeutically effective amount of an exogenous receptor molecule, which exogenous
20 receptor molecule specifically binds to canine IgE.
31. The pharmaceutical formulation of claim 30 wherein the exogenous receptor molecule comprises a chimeric monoclonal antibody.
- 25 32. The pharmaceutical formulation of claim 31 wherein the chimeric monoclonal antibody comprises a chimera of canine constant heavy and light domains fused to mouse heavy and light chain variable regions and has an affinity for
30 exon 3 of canine IgE.

- 5 33. The pharmaceutical formulation of claim 30,
further optionally comprising one or more
components selected from the group consisting of:
saline or other parenteral solutions, buffering
agents, stabilizers, bulking agents,
10 antioxidants, and co-solvents.
34. The pharmaceutical formulation of claim 31 in a
form suitable for administration by one or more
of the following methods: intravenous
administration, subcutaneous administration, and
15 intramuscular administration.

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Neutralization of 0.1 ug/ml rIgE by chimeric 15A2

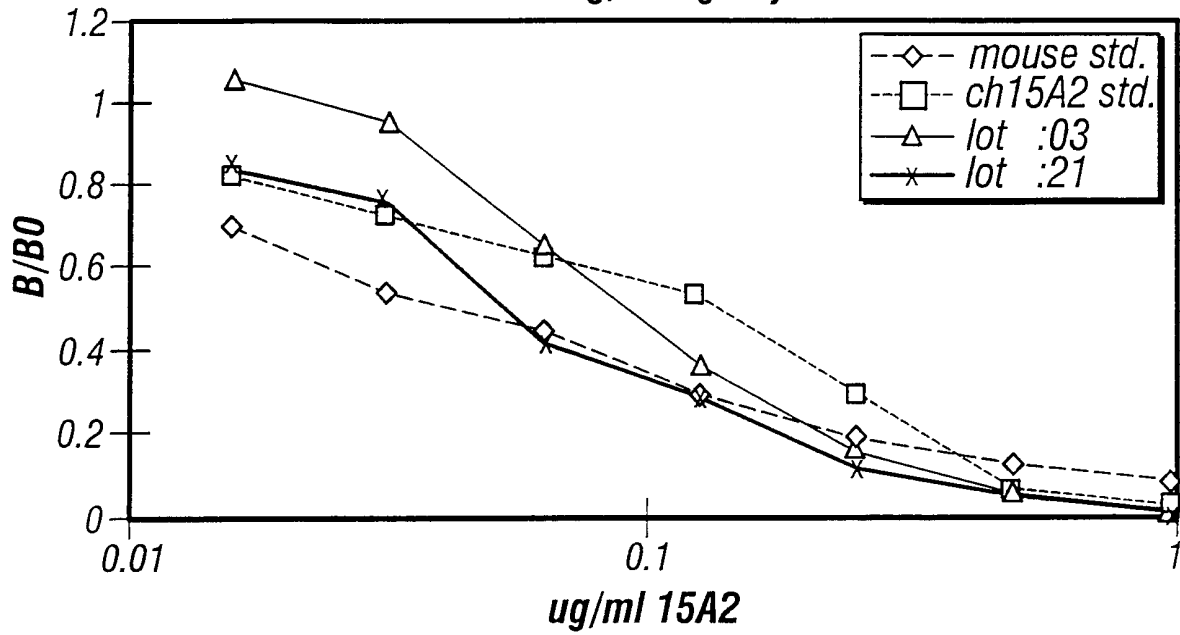


FIG. 1

Chimeric 15A2 Antibody Treated Dog 2001

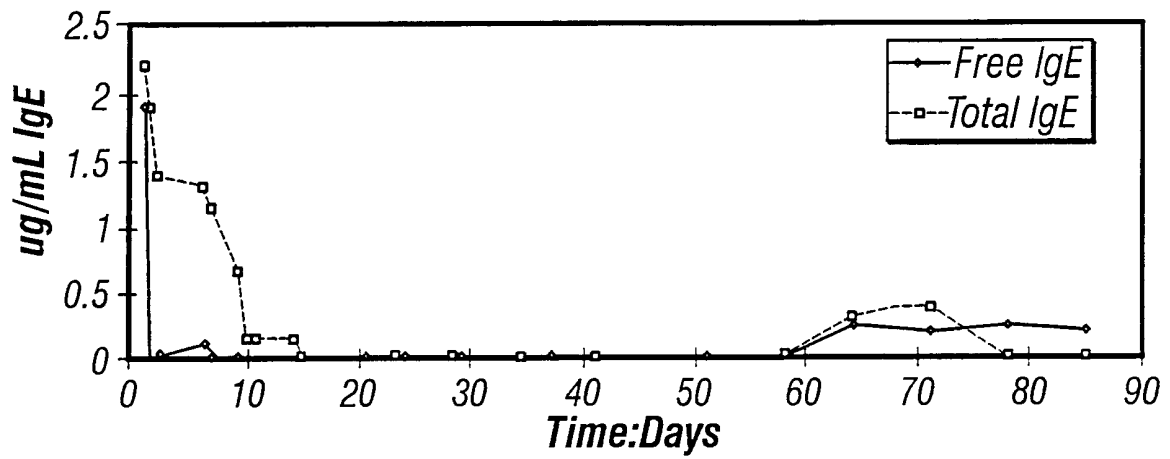


FIG. 2A-1

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Chimeric 15A2 Antibody Treated Dog 2002

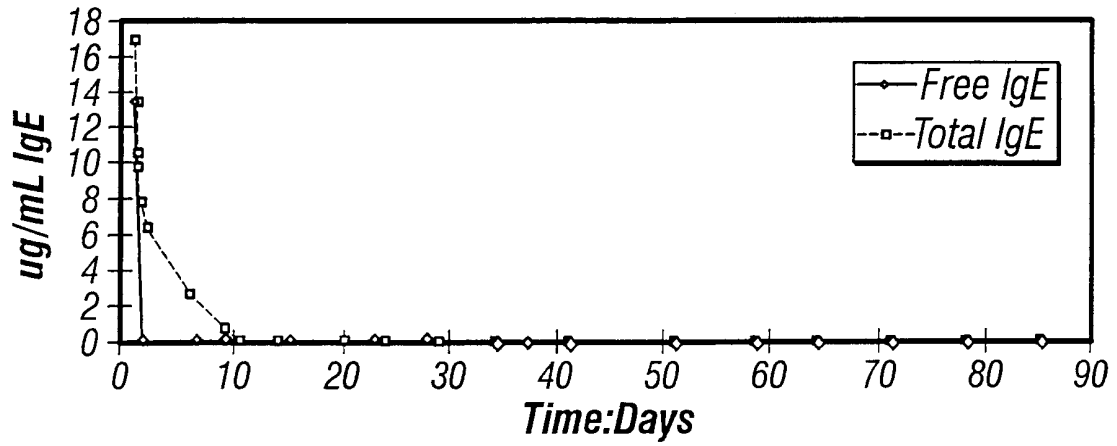


FIG. 2A-2

Chimeric 15A2 Antibody Treated Dog 2103

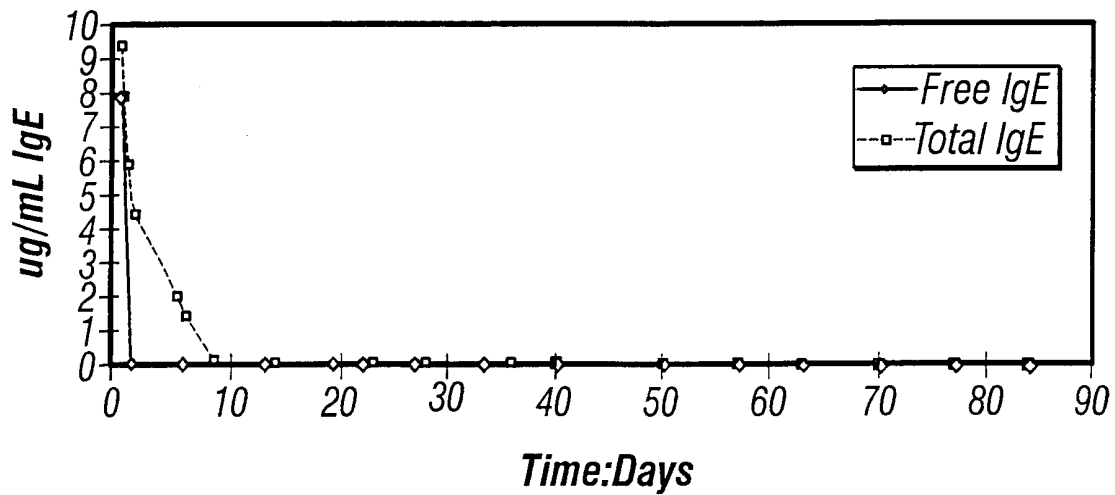


FIG. 2A-3

Free and Total Serum IgE in Control Dog 1102

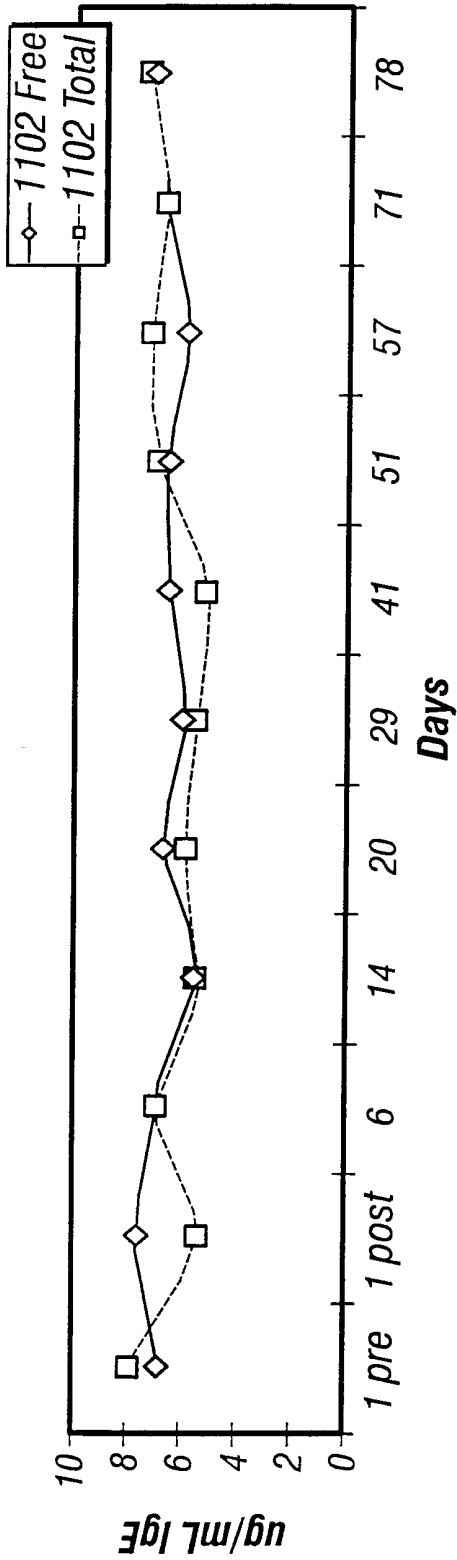


FIG. 2B-1

Free and Total Serum IgE in Control Dog 1101

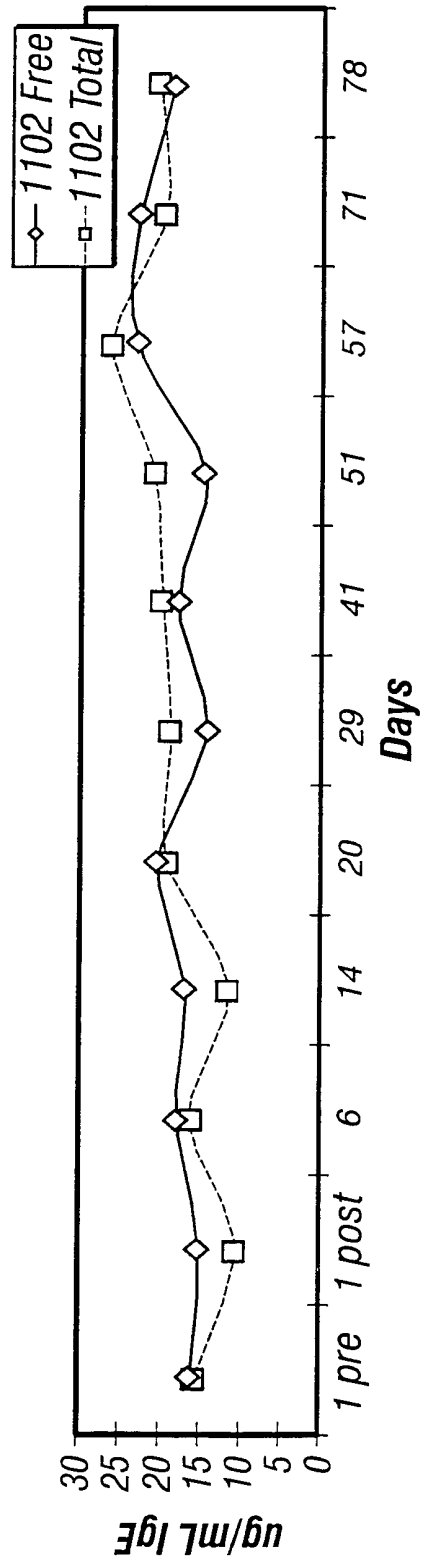


FIG. 2B-2

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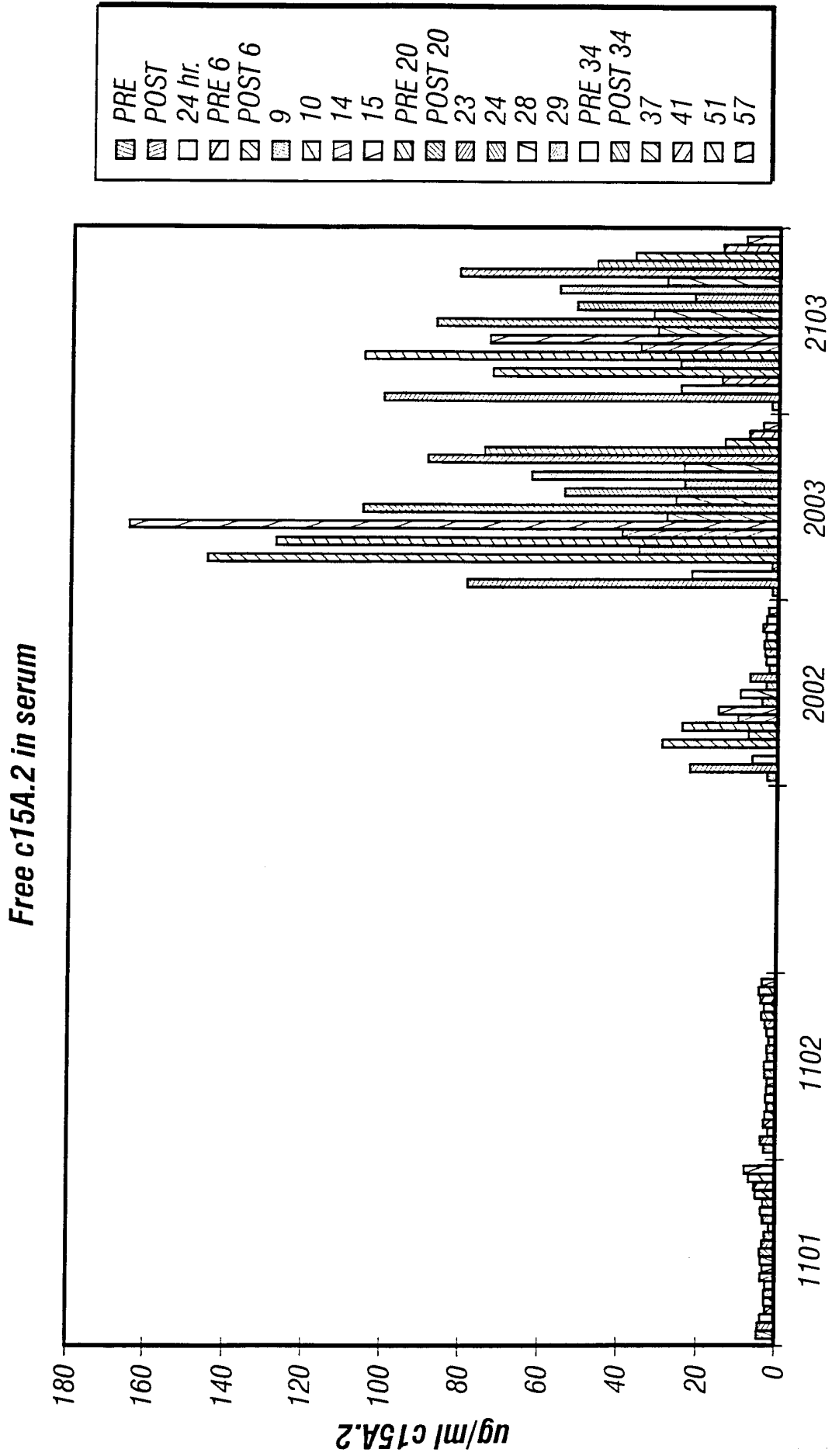
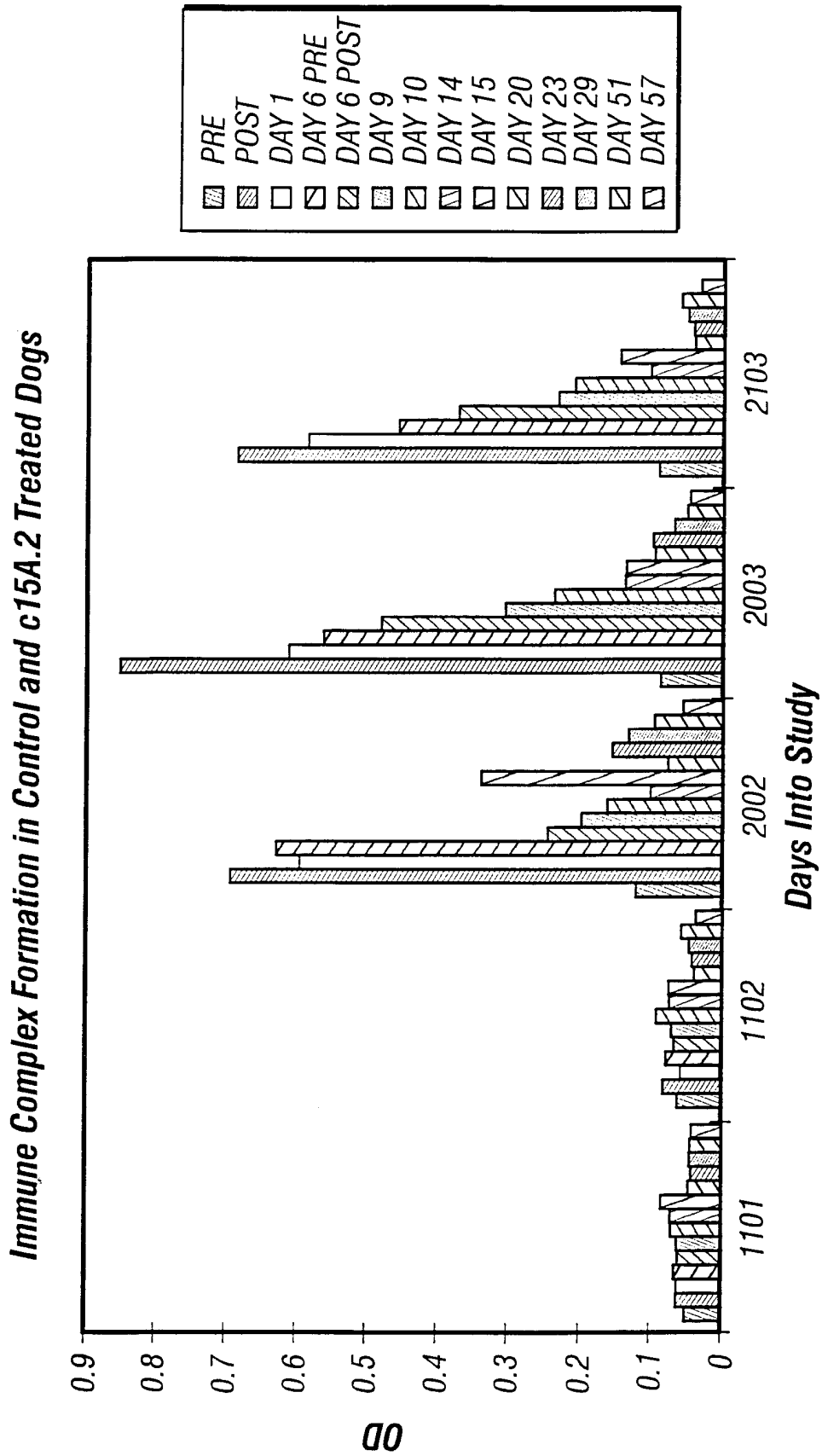


FIG. 3

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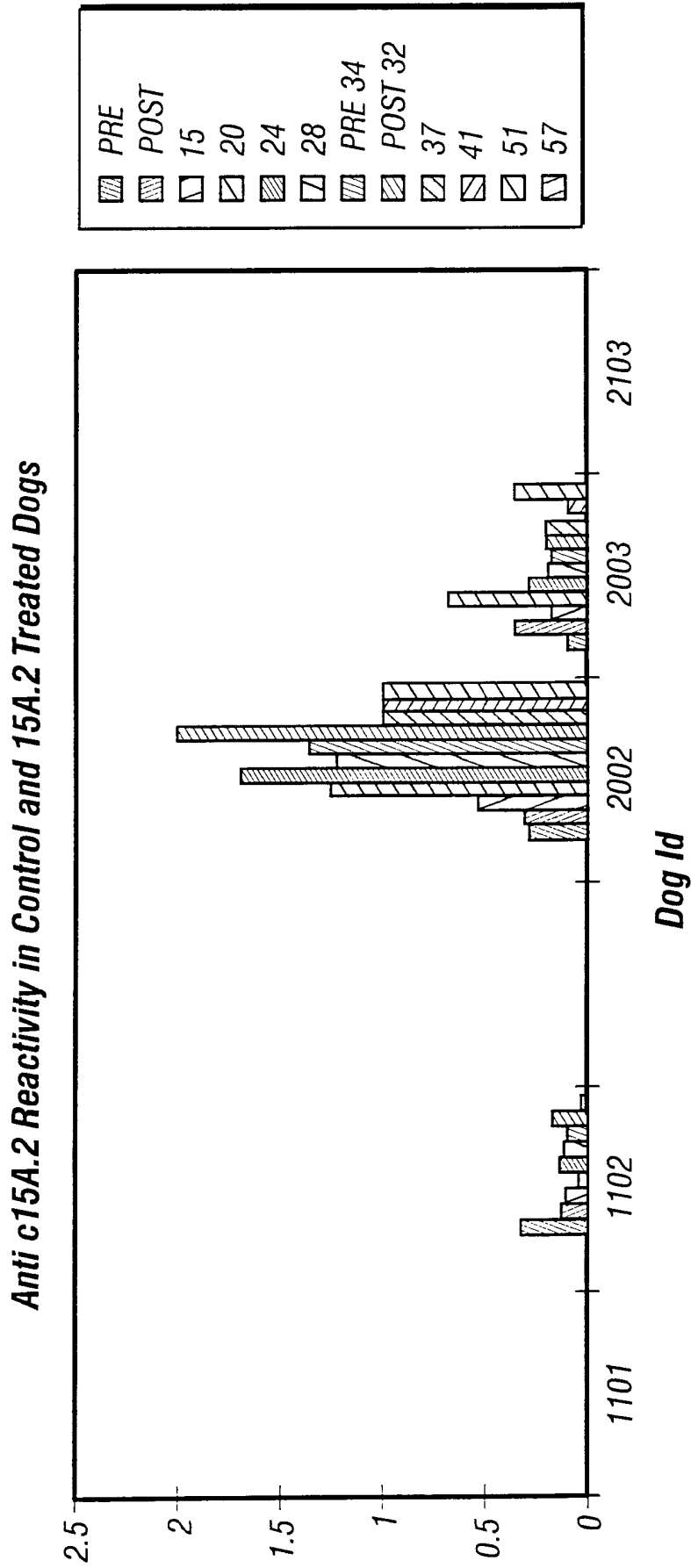


FIG. 5

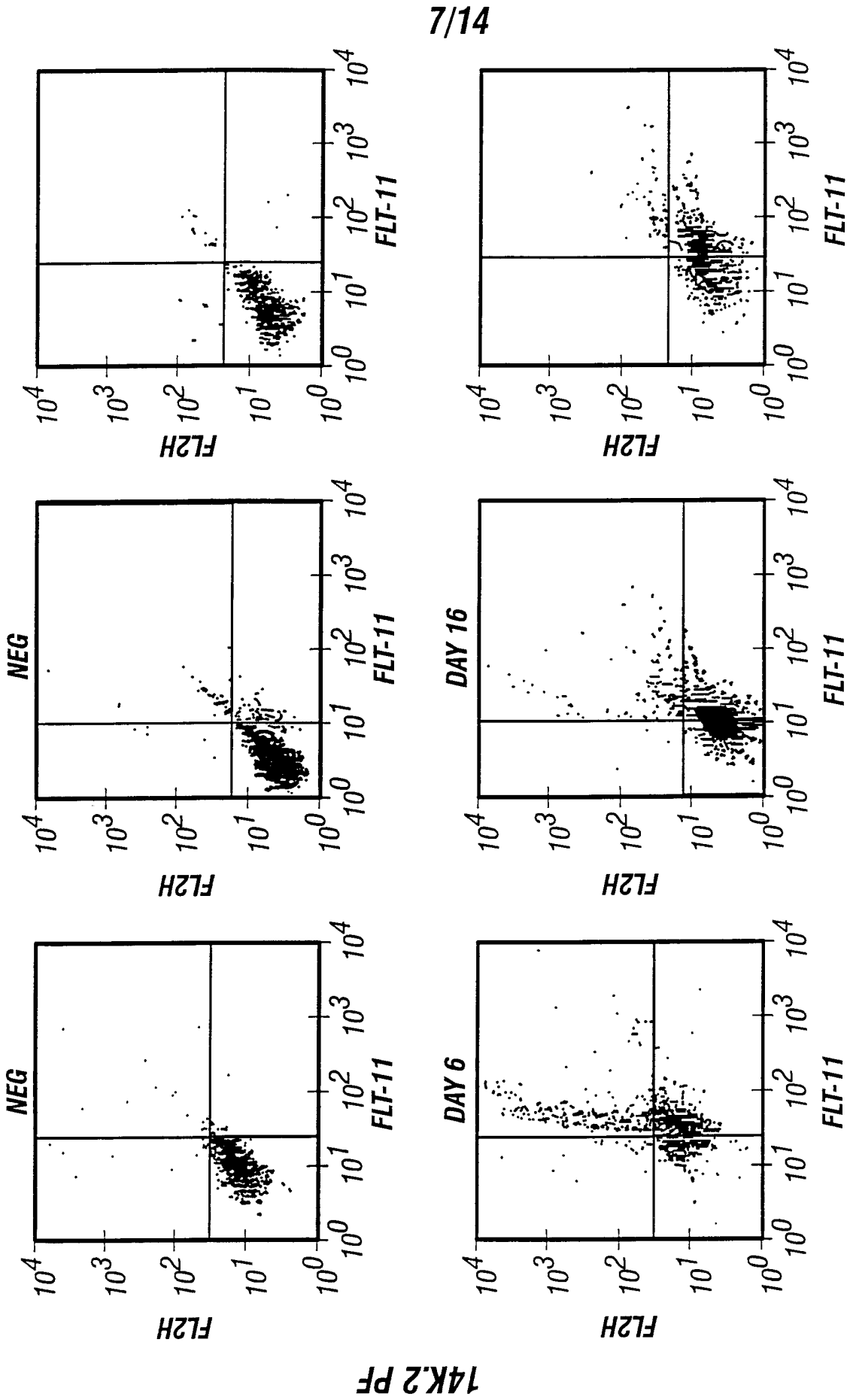


FIG. 6

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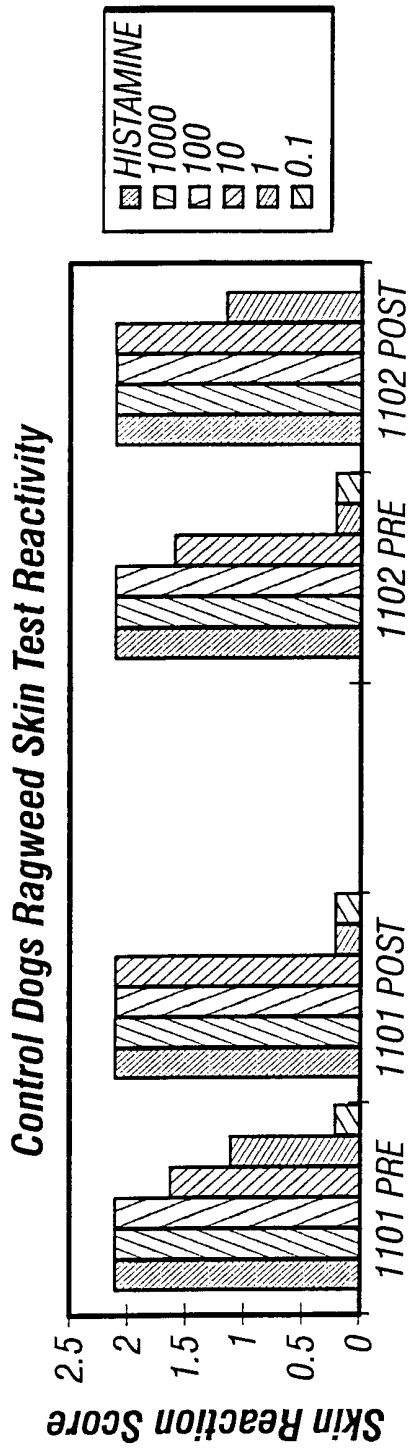


FIG. 7A

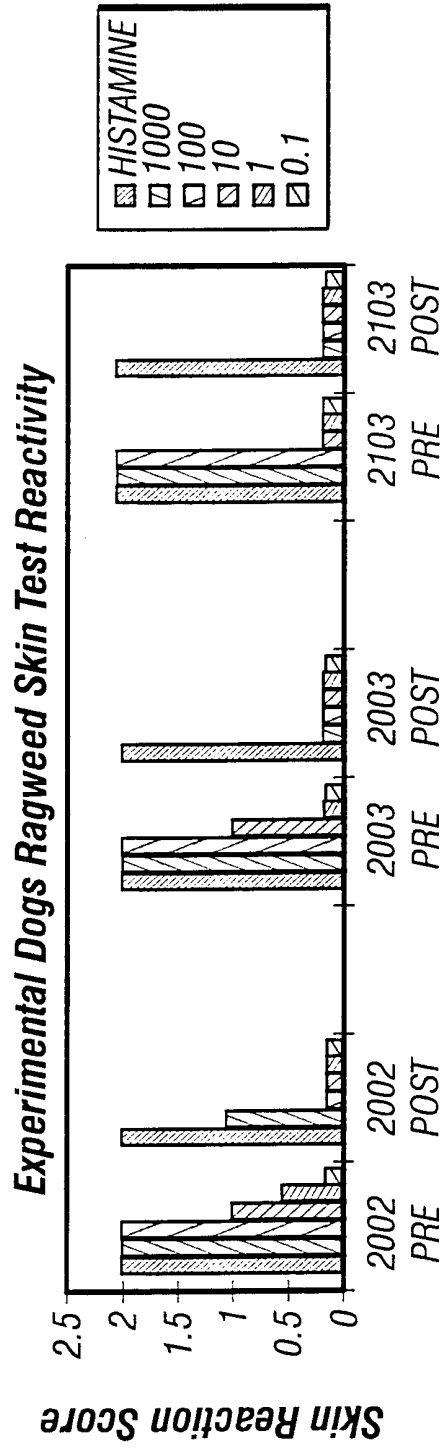


FIG. 7B

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DOG ID	WEIGHT (g)	BLOOD VOL. (ml)	IgE LEVEL (ug/ml)	TOTAL FREE IgE (mg)	10X c15A2 (mg)
2002	9,300g	558 ml	1.61	0.898mg	8.98mg
2003	12,200g	732ml	9.71	7.11mg	71.1mg
2103	8,000g	480ml	8.14	3.91mg	39.1mg
TOTAL					119.18mg

** DOG BLOOD VOLUME: 6% OF BODY WEIGHT

FIG. 8

Free IgE Level in cRclg - Treated Dogs

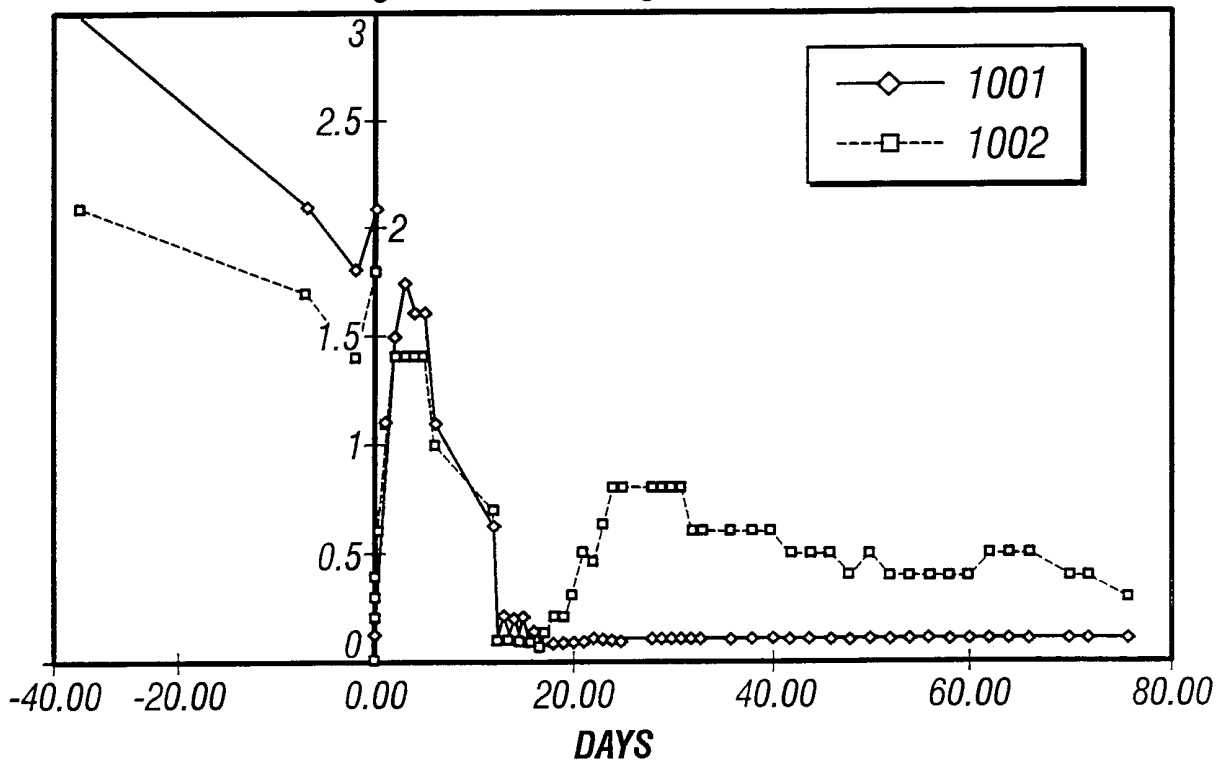


FIG. 9

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5401 CCGCGAGATG CCTGCTTCCA TGGGAGGCCCC TGCCCTGCTG TGGCTAGCGC
 M P A S M G G P A L L W L A
 GCGGCTCTAC GGACGAAGGT ACCCTCCGGG ACGGGACGAC ACCGATCGCG

L L L S S P G V M S S D T L K P T
5451 TGCTGCTCTC CTCTCCAGGT GTCATGTCAT CAGATACCTT GAAACCTACA
 ACGACGAGAG GAGAGGTCCA CAGTACAGTA GTCATGGAA CTTTGGATGT

V S M N P P W N T I L K D D S V T
5501 GTGTCCATGA ACCCGCCATG GAATACAATA TTGAAGGATG ACAGTGTGAC
 CACAGGTACT TGGCGGTAC CTTATGTTAT AACTTCCTAC TGTCACACTG

L T C T G N N S L E V D S A V W
5551 TCTTACATGT ACTGGGAACA ACTCCCTTGA AGTCGACTCT GCTGTGTGGC
 AGAATGTACA TGACCCCTTGT TGAGGGAACT TCAGCTGAGA CGACACACCCG

L H N N T T W Q E T T S R L D I N
5601 TCCACAACAA CACTACTTGG CAAGAGACGA CTTACCGTTT GGACATCAAT
 AGGTGTTGTT GTGATGAACC GTTCTCTGCT GAAGTGCAAA CCTGTAGTTA

K A Q I Q D S G E Y R C R E N R S
5651 AAAGCCCAA TCCAGGACAG TGGGGAGTAC AGGTGTCGGG AAAATAGATC
 TTTTCGGGTTT AGGTCCTGTC ACCCCTCATG TCCACAGCCC TTTTATCTAG

FIG. 10A

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I L S D P V Y L T V F T E W L I
5701 CATCCTGAGT GATCCTGTGT ACCTAACAGT CTTACACAGAG TGGCTGATCC
 GTAGGACTCA CTAGGACACA TGGATTGTCA GAAGTGTCTC ACCGACTAGG

 L Q A S A N V V M E G E S F L I R
5751 TTCAAGCCTC TGCCAACGTG GTGATGGAGG GTGAGAGCTT CCTCATCAGG
 AAGTTCGGAG ACGTTGCAC CACTACCTCC CACTCTCGAA GGAGTAGTCC

 C H S W K N L R L T K V T Y Y K D
5801 TGCCATAGTT GGAAGAATTT GAGGCTCACA AAGGTGACCT ACTACAAGGA
 ACGGTATCAA CCTTCTTAAA CTCCGAGTGT TTCCACTGGA TGATGTTCCCT

 G I P I R Y W Y E N F N I S I S
5851 TGGCATCCCC ATCAGGTAAT GGTACGAGAA CTTCAACATC TCCATTAGCA
 ACCGTAGGG TAGTCCATGA CCATGCTCTT GAAGTTGTAG AGGTAATCGT

 N V T T K N S G N Y S C S G Q I Q
5901 ACGTCACAAC CAAAAACAGC GGCAACTATT CCTGCTCAGG CCACATCCAG
 TGCAGTGTG GTTTTTGTG CCGTTGATAA GGACGAGTCC GGTCTAGGTC

 Q K G Y T S K V L N I I V K K E P
5951 CAGAAAGGCT ACACCTCTAA AGTCCCTCAAC ATTATTGTGA AAAAAAGAGCC
 GTCCTTCCGA TGTGGAGATT TCAGGAGTTG TAATAACACT TTTTTCCTCGG

FIG. 10B

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T K Q N K Y S G L H R P P D C P
6001 CACCAAGCAA AACAAAGTACT CCGGGCTACA CCGCCACCT GATTGTCCCA
 GTGGTTCGTT TTGTTTCATGA GGCCCGATGT GCGGGTGGA CTAACAGGGT

 K C P A P E M L G G P S V F I F P
6051 AATGCCCAGC CCCTGAAATG CTGGGAGGC CTTCCGGTCTT CATCTTCCCC
 TTACGGGTCG GGGACTTTAC GACCTCCCG GAAGCCAGAA GTAGAAAGGG

 P K P K D T L L I A R T P E V T C
6101 CCGAAACCCA AGGACACCCT CTTGATTGCC CGAACACCTG AGGTCACATG
 GGCTTTGGGT TCCTGTGGGA GAACTAACGG GCTTGTGGAC TCCAGTGTAC

 V V V D L D P E D P E V Q I S W
6151 TGTGGTGGTG GATCTGGACC CAGAAGACCC TGAGGTGCAG ATCAGCTGGT
 ACACCACCAC CTAGACCCTGG GTCTTCTGGG ACTCCACGTC TAGTCGACCA

 F V D G K Q M Q T A K T Q P R E E
6201 TCGTGGACGG TAAGCAGATG CAAACAGCCA AGACTCAGCC TCGTGAGGAG
 AGCACCTGCC ATTCGTCTAC GTTTGTCCGG TCTGAGTCGG AGCACTCCCTC

 Q F N G T Y R V V S D L P I G H Q
6251 CAGTCAATG GCACCTACCG TGTGGTFCAGT GACCTCCCA TTGGGCACCA
 GTC AAGTTAC CGTGGATGGC ACACCAGTCA CTGGAGGGGT AACCCGTGGT

FIG. 10C

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D W L K G K Q F T C K V N N K A
6301 GGACTGGCTC AAGGGGAAAGC AGTTCACCTG CAAAGTCAAC AACAAAGCCC
 CCTGACCGAG TTCCCCCTTCG TCAAGTGGAC GTTTCAGTTG TTGTTTCGGG

 L P S P I E R T I S K A R G L A I
6351 TCCCATCCCC GATCGAGAGG ACCATCTCCA AGGCCAGAGG GCTGGCCATA
 AGGTAGGGG CTAGCTCTCC TGGTAGAGGT TCCGGTCTCC CGACCGGTAT

SmaI, XmaI, Aval
 A S V Y V L P P S R E E L S K N T
6401 GCCAGTGTGT ATGTCCCTGCC GCCATCCCGG GAGGAGTTGA GCAAGAACAC
 CCGTCACACA TACAGGACGG CCGTAGGGCC CTCCTCAACT CGTTCTTGTG

 V S L T C L I K D F P P D I D
6451 AGTCAGCTTG ACATGCCTGA TCAAAGACTT CTTCCTCCCTT GACATTGATG
 TCAGTCGAAC TGTACGGACT AGTTTCTGAA GAAGGGGGGA CTGTAACCTAC

 V E W Q S N G Q Q E P E S K Y R T
6501 TGGAGTGGCA GAGCAATGGA CAGCAGGAGC CTGAGAGTAA GTACCGCACG
 ACCTCACCGT CTCGTTACCT GTCGTCCCTCG GACTCTCATT CATGGCGGTGC

 T L P Q L D E D G S Y F L Y S K L
6551 ACCCTGCCCC AGCTGGACGA GGACGGGTCC TACTTCTGT ACAGCAAGCT
 TGGGACGGGG TCGACCTGCT CCTGCCCCAGG ATGAAGGACA TGTCTGTTCCA

FIG. 10D

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S V D K S R W Q R G D T F I C A
6601 CTCGTGGAT AAGAGCCGCT GGCAGCGGG AGACACCTTC ATATGTGCGG
 GAGACACCTA TTCTCGGCGA CCGTCGCCCC TCTGTGGAAG TATACACGCC

 V M H E A L H N H Y T Q K S L S H
6651 TGATGCATGA AGCTCTACAC AACCACTACA CACAGAAATC CCTCTCCCAT
 ACTACGTACT TCGAGATGTG TTGGTGATGT GTGTCTTTAG GGAGAGGGTA

 S P G K

FIG. 10E

SEQUENCE LISTING

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<130> 036040036PC01

<140> TO BE ASSIGNED

<141> 2001-01-30

<150> 09/592,998

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cct ggg gct tca gtg aag ata tcc tgc aag gct tct ggt tac tca ttt	144
act gac tac ttt atg aac tgg gtg atg cag agc cat gga aag agc ctt	192
gag tgg att ggt cgt att aat cct ttc aat ggt gat cct ttc tac aac	240
cag aag ttc aag ggc aag gcc aca ttg act gta gac aaa tcc tct agc	288
aca gcc cac atg gag ctc cgg agc ctg gca tct gag gac tct gca gtc	336
tat tat tgt gca aga ttc tac tac gga cgt tac tat gct atg gac tac	384
tgg ggt caa gga acc tca gtc acc gtc tcc tca gcc tcc acc acg gcc	432
ccc tcg gtt ttc cca ctg gac ccc agc tgc ggg tcc act tcc ggc tcc	480
acg gtg gcc ctg gcc tgc ctg gtg tca ggc tac ttc ccc gag cct gta	528
act gtg tcc tgg aat tcc ggc tcc ttg acc agc ggt gtg cac acc ttc	576
ccg tcc gac ctg cag tcc tca ggg ctc tac tcc ctc agc agc atg gtg	624
aca gtg ccc tcc agc agg tgg tcc agc gag acc ttc acc tgc aac gtg	672
gcc cac ccg gcc agc aaa act aaa gta gac aag cca gtg ccc aaa aga	720
gaa aat gga aga gtt cct cgc cca cct gat tgt ccc aaa tgc cca gcc	768
cct gaa atg ctg gga ggg cct tcg gtc ttc atc ttt ccc ccg aaa ccc	816
aag gac acc ctc ttg att gcc cga aca cct gag gtc aca tgt gtg gtg	864
gtg gat ctg gga cca gaa gac cct gag gtg cag atc agc tgg ttc gtg	912
gac ggt aag cag atg caa aca gcc aag act cag cct cgt gag gag cag	960
ttc aat ggc acc tac cgt gtg gtc agt gtc ctc ccc att ggg cac cag	1008
gac tgg ctc aag ggg aag cag ttc acg tgc aaa gtc aac aac aaa gcc	1056
ctc cca tcc ccg atc gag agg acc atc tcc aag gcc aga ggg cag gcc	1104
cat cag ccc agt gtg tat gtc ctg ccg cca tcc cgg gag gag ttg agc	1152
aag aac aca gtc agc ttg aca tgc ctg atc aaa gac ttc ttc cca cct	1200
gac att gat gtg gag tgg cag agc aat gga cag cag gag cct gag agc	1248
aag tac cgc acg acc ccg ccc cag ctg gac gag gac ggg tcc tac ttc	1296
ctg tac agc aag ctc tct gtg gac aag agc cgc tgg cag ccg gga gac	1344
acc ttc ata tgt gcg gtg atg cat gaa gct cta cac aac cac tac aca	1392
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tgctcgtcaa gtactggggc tgttacaact agtaactatg ccaactgggt ccaagaaaaa	180
ccagatcatt tattcactgg tctaataagg ggtcccaaca accgagctcc aggtgttcct	240
gccagattct caggctccct gattggagac aaggctgcc tcaccatcac aggggcacag	300
actgaggatg aggcaatata tttctgtgct ctatggtaca gcaaccattg ggtgttcggt	360
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 agcctgacgc ctgacaagtg gaaatctcac agcagcttca gctgcctggg cagcagcagag 660
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ctg ctg ctc tcc tct cca ggt gtc atg tca tca gat acc ttg aaa cct 97
 Leu Leu Leu Ser Ser Pro Gly Val Met Ser Ser Asp Thr Leu Lys Pro
 15 20 25 30

aca gtg tcc atg aac ccg cca tgg aat aca ata ttg aag gat gac agt 145
 Thr Val Ser Met Asn Pro Pro Trp Asn Thr Ile Leu Lys Asp Asp Ser
 35 40 45

gtg act ctt aca tgt act cgg aac aac tcc ctt gaa gtc gac tct gct 193
 Val Thr Leu Thr Cys Thr Arg Asn Asn Ser Leu Glu Val Asp Ser Ala
 50 55 60

gtg tgg ctc cac aac aac act act tgg caa gag acc act tca cgt ttg 241
 Val Trp Leu His Asn Asn Thr Thr Trp Gln Glu Thr Thr Ser Arg Leu
 65 70 75

gac atc aat aaa gcc caa atc cag gac agt ggg gag tac agg tgt cgg 289
 Asp Ile Asn Lys Ala Gln Ile Gln Asp Ser Gly Glu Tyr Arg Cys Arg
 80 85 90

gaa aat aga tcc atc ctg agt gat cct gtg tac cta aca gtc ttc aca 337
 Glu Asn Arg Ser Ile Leu Ser Asp Pro Val Tyr Leu Thr Val Phe Thr
 95 100 105 110

gag tgg ctg atc ctt caa gcc tct gcc aac gtg gtg atg gag ggt gag 385
 Glu Trp Leu Ile Leu Gln Ala Ser Ala Asn Val Val Met Glu Gly Glu
 115 120 125

agc ttc ctc atc agg tgc cat agt tgg aag aat ttg agc ctc aca aag 433
 Ser Phe Leu Ile Arg Cys His Ser Trp Lys Asn Leu Ser Leu Thr Lys
 130 135 140

gtg acc tac tac aag gat ggc atc ccc atc agg tac tgg tac gag aac 481
 Val Thr Tyr Tyr Lys Asp Gly Ile Pro Ile Arg Tyr Trp Tyr Glu Asn
 145 150 155

ttc aac atc tcc att agc aac gtc aca acc aaa aac agc ggc aac tat 529
 Phe Asn Ile Ser Ile Ser Asn Val Thr Thr Lys Asn Ser Gly Asn Tyr

160	165	170	
tcc tgc tca ggc cag atc cag cag aaa ggc tac acc tct aaa gtc ctc			577
Ser Cys Ser Gly Gln Ile Gln Gln Lys Gly Tyr Thr Ser Lys Val Leu			
175	180	185	190
aac att att gtg aaa aaa gag ccc acc aag caa aac aag tac tcc ggg			625
Asn Ile Ile Val Lys Lys Glu Pro Thr Lys Gln Asn Lys Tyr Ser Gly			
	195	200	205
cta cac cgc cca cct gat tgt ccc aaa tgc cca gcc cct gaa atg ctg			673
Leu His Arg Pro Pro Asp Cys Pro Lys Cys Pro Ala Pro Glu Met Leu			
	210	215	220
gga ggg cct tcg gtc ttc atc ttt ccc ccg aaa ccc aag gac acc ctc			721
Gly Gly Pro Ser Val Phe Ile Phe Pro Pro Lys Pro Lys Asp Thr Leu			
	225	230	235
ttg att gcc cga aca cct gag gtc aca tgt gtg gtg gtg gat ctg gac			769
Leu Ile Ala Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Leu Asp			
	240	245	250
cca gaa gac cct gag gtg cag atc agc tgg ttc gtg gac ggt aag cag			817
Pro Glu Asp Pro Glu Val Gln Ile Ser Trp Phe Val Asp Gly Lys Gln			
255	260	265	270
atg caa aca gcc aag act cag cct cgt gag gag cag ttc aat ggc acc			865
Met Gln Thr Ala Lys Thr Gln Pro Arg Glu Glu Gln Phe Asn Gly Thr			
	275	280	285
tac cgt gtg gtc agt gac ctc ccc att ggg cac cag gac tgg ctc aag			913
Tyr Arg Val Val Ser Asp Leu Pro Ile Gly His Gln Asp Trp Leu Lys			
	290	295	300
ggg aag cag ttc acc tgc aaa gtc aac aac aaa gcc ctc cca tcc ccg			961
Gly Lys Gln Phe Thr Cys Lys Val Asn Asn Lys Ala Leu Pro Ser Pro			
	305	310	315
atc gag agg acc atc tcc aag gcc aga ggg ctg gcc ata gcc agt gtg			1009
Ile Glu Arg Thr Ile Ser Lys Ala Arg Gly Leu Ala Ile Ala Ser Val			
	320	325	330
tat gtc ctg ccg cca tcc cgg gag gag ttg agc aag aac aca gtc agc			1057
Tyr Val Leu Pro Pro Ser Arg Glu Glu Leu Ser Lys Asn Thr Val Ser			
335	340	345	350
ttg aca tgc ctg atc aaa gac ttc ttc ccc cct gac att gat gtg gag			1105
Leu Thr Cys Leu Ile Lys Asp Phe Phe Pro Pro Asp Ile Asp Val Glu			
	355	360	365
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Trp Gln Ser Asn Gly Gln Gln Glu Pro Glu Ser Lys Tyr Arg Thr Thr			
	370	375	380
ctg ccc cag ctg gac gag gac ggg tcc tac ttc ctg tac agc aag ctc			1201
Leu Pro Gln Leu Asp Glu Asp Gly Ser Tyr Phe Leu Tyr Ser Lys Leu			
	385	390	395

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tct gtg gat aag agc cgc tgg cag cgg gga gac acc ttc ata tgt gcg      1249
Ser Val Asp Lys Ser Arg Trp Gln Arg Gly Asp Thr Phe Ile Cys Ala
  400                               405                               410

gtg atg cat gaa gct cta cac aac cac tac aca cag aaa tcc ctc tcc      1297
Val Met His Glu Ala Leu His Asn His Tyr Thr Gln Lys Ser Leu Ser
  415                               420                               425                               430

cat                                                                1300
His
    
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<212> PRT
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  20                               25                               30
Ser Met Asn Pro Pro Trp Asn Thr Ile Leu Lys Asp Asp Ser Val Thr
  35                               40                               45
Leu Thr Cys Thr Arg Asn Asn Ser Leu Glu Val Asp Ser Ala Val Trp
  50                               55                               60
Leu His Asn Asn Thr Thr Trp Gln Glu Thr Thr Ser Arg Leu Asp Ile
  65                               70                               75                               80
Asn Lys Ala Gln Ile Gln Asp Ser Gly Glu Tyr Arg Cys Arg Glu Asn
  85                               90                               95
Arg Ser Ile Leu Ser Asp Pro Val Tyr Leu Thr Val Phe Thr Glu Trp
  100                              105                              110
Leu Ile Leu Gln Ala Ser Ala Asn Val Val Met Glu Gly Glu Ser Phe
  115                              120                              125
Leu Ile Arg Cys His Ser Trp Lys Asn Leu Ser Leu Thr Lys Val Thr
  130                              135                              140
Tyr Tyr Lys Asp Gly Ile Pro Ile Arg Tyr Trp Tyr Glu Asn Phe Asn
  145                              150                              155                              160
Ile Ser Ile Ser Asn Val Thr Thr Lys Asn Ser Gly Asn Tyr Ser Cys
  165                              170                              175
Ser Gly Gln Ile Gln Gln Lys Gly Tyr Thr Ser Lys Val Leu Asn Ile
  180                              185                              190
Ile Val Lys Lys Glu Pro Thr Lys Gln Asn Lys Tyr Ser Gly Leu His
  195                              200                              205
Arg Pro Pro Asp Cys Pro Lys Cys Pro Ala Pro Glu Met Leu Gly Gly
  210                              215                              220
Pro Ser Val Phe Ile Phe Pro Pro Lys Pro Lys Asp Thr Leu Leu Ile
  225                              230                              235                              240
Ala Arg Thr Pro Glu Val Thr Cys Val Val Val Asp Leu Asp Pro Glu
  245                              250                              255
Asp Pro Glu Val Gln Ile Ser Trp Phe Val Asp Gly Lys Gln Met Gln
  260                              265                              270
Thr Ala Lys Thr Gln Pro Arg Glu Glu Gln Phe Asn Gly Thr Tyr Arg
  275                              280                              285
Val Val Ser Asp Leu Pro Ile Gly His Gln Asp Trp Leu Lys Gly Lys
    
```


INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 01/02924

A. CLASSIFICATION OF SUBJECT MATTER
 IPC 7 C07K16/42 A61K39/395 A61P37/08

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
 IPC 7 C07K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, EMBL, BIOSIS, MEDLINE, EMBASE, PAJ, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	EP 0 957 111 A (IDEXX LAB INC) 17 November 1999 (1999-11-17) cited in the application page 4, line 38 -page 5, line 35 page 8, line 19 -page 11, line 45 claims 1-37	1-34
X	WO 97 20859 A (IDEXX LAB INC) 12 June 1997 (1997-06-12) page 5, line 22 -page 7, line 3; claims 1-3	1-3,8-10
X	US 5 945 294 A (FRANK GLENN R ET AL) 31 August 1999 (1999-08-31) abstract claims 35-77; examples 1,2	1,8-10
	-/--	

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

° Special categories of cited documents :

- *A* document defining the general state of the art which is not considered to be of particular relevance
- *E* earlier document but published on or after the international filing date
- *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- *O* document referring to an oral disclosure, use, exhibition or other means
- *P* document published prior to the international filing date but later than the priority date claimed

- *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- *X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- *Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- *&* document member of the same patent family

Date of the actual completion of the international search

22 June 2001

Date of mailing of the international search report

09/07/2001

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 Fax: (+31-70) 340-3016

Authorized officer
 Muller-Thomalla, K

INTERNATIONAL SEARCH REPORT

International Application No

PC1/US 01/02924

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98 45707 A (HESKA CORP) 15 October 1998 (1998-10-15) abstract page 35, line 1 -page 36; claims 5-8 -----	1,8-10
X	DEBOER D J ET AL: "Production and characterization of mouse monoclonal antibodies directed against canine IgE and IgG" VETERINARY IMMUNOLOGY AND IMMUNOPATHOLOGY, NL, AMSTERDAM, vol. 37, no. 3/04, 1 August 1993 (1993-08-01), pages 183-199, XP000647974 ISSN: 0165-2427 abstract page 186, paragraphs 3-5 page 192, last paragraph -page 195, last paragraph page 197, paragraph 3	1-3,8, 10,11, 15,30, 33,34
Y	----- -----	4-7,9, 12-14, 16-29, 31,32
Y	DATABASE EMBL 'Online! ID/AC: E03345, 8 October 1997 (1997-10-08) KURUMI ET AL.: "DNA sequence coding for unchangeable region of dog immunoglobulin gamma chain" XP002170410 abstract -----	4-7,9, 12-14, 16-29, 31,32

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: Part of claims 1,6,8-10,16,20,22-25,27-30,33,34

Present independent claims 1,16 and 30 relate to an extremely large number of possible compounds (and use of such compounds in claim 16). Support within the meaning of Article 6 PCT and/or disclosure within the meaning of Article 5 PCT is to be found, however, for only a very small proportion of the compounds claimed. In the present case, the claims so lack support, and the application so lacks disclosure, that a meaningful search over the whole of the claimed scope is impossible. Consequently, the search has been carried out for those parts of claims 1,6,8-10,16,20,22-25,27-30,33,34, which appear to be supported and disclosed, namely those parts relating to "antibodies" specific for IgE as defined in the various dependent claims which relate to said antibodies.

In this respect it should be noted that the expressions "exogenous" and "receptor" used in the present claims are not clear in the given context and are thus not considered to be limiting features with respect to the scope of the claims in which said expressions appear in. In the light of the present description, in particular table 3, the term "receptor" is interpreted as the IgE receptor cRcIg as reflected in said table, which is not considered to be equivalent to the standard definition of antibodies, e.g. monoclonals as in claim 3. The term "exogenous" in the given context is unclear and obscure. Furthermore in present claims 6,8,22-25,27-30,33 and 34 the claimed "molecule" is defined by reference to a desirable characteristic or property, leading to an objection for lack of clarity (Article 6 PCT). An attempt is made to define the product by reference to a result to be achieved.

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 01/02924

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP 0957111 A	17-11-1999	AU 4044900 A EP 1082346 A WO 0058365 A	16-10-2000 14-03-2001 05-10-2000
WO 9720859 A	12-06-1997	AU 1146897 A	27-06-1997
US 5945294 A	31-08-1999	AU 7411498 A EP 0943097 A WO 9823964 A	22-06-1998 22-09-1999 04-06-1998
WO 9845707 A	15-10-1998	US 6060326 A AU 6796498 A	09-05-2000 30-10-1998