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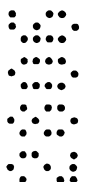
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ABSTRACT

The present invention relates to a substantially pure population of viable bile duct progenitor cells, and methods for isolating such cell. The present invention further concerns certain therapeutic uses for such progenitor cells, and their progeny.



THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. A substantially purified population of viable bile duct progenitor cells.
2. A cellular composition comprising, as the cellular component, a substantially purified population of viable non-hepatic bile duct progenitor cells, which progenitor cells are capable of proliferation in a culture medium.
3. The composition of claim 2, having fewer than 20% of lineage committed cells.
4. The composition of claim 2, which progenitor cells from a mammal.
5. The composition of claim 4, which mammal is a transgenic mammal.
6. The composition of claim 4, which mammal is a primate.
7. The composition of claim 6, which mammal is a human.
8. The composition of claim 4, which mammal is a miniature swine.
9. The composition of claim 2, which progenitor cells are capable of differentiation to tissue lineages of organs derived from the primitive gut.
10. The composition of claim 9, which progenitor cells are capable of differentiation to hepatic lineages.
11. The composition of claim 10, wherein the progenitor cells are characterized by expression of one or more of a hepatocyte nuclear factor (HNF); ATBF1; AFP; a LIM type homeobox gene; a forkhead transcription factor; a CCAAT-enhancer binding protein (C/EBP); OV-6; cytokeratin type 7; cytokeratin type 19; Span-1; OC.2; CC.3; or γ -glutamyl transpeptidase
12. The composition of claim 11, wherein the progenitor cells are characterized by expression of HNF3 β , alphafetoprotein (AFP), albumin, and ATBF-1.
13. The composition of claim 9, which progenitor cells are capable of differentiation to pancreatic lineages.

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ORIGINAL
COMPLETE SPECIFICATION

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Complete Specification for the invention entitled:

Bile duct progenitor cells and methods of use

The following statement is a full description of this invention, including the best
method of performing it known to us:

Bile Duct Progenitor Cells and Methods of Use

Background of the Invention

During the early stages of embryogenesis cells are totipotent and are capable of multidirectional differentiation. As development proceeds, the totipotent cells become determined and committed to differentiate into a given specialized cell type. Final differentiation is associated with the acquisition of specialized cell functions. Thus, the differentiated somatic cells maintain their specialized features throughout the life span of the organism, probably through sustained interactions between the genome and its microenvironment and cell-cell interactions (DiBerardino et al., 1984, *Science* 224:946-952; Wetts and Fraser, 1988, *Science* 239:1142-1144; Fisher, 1984, *PNAS* 81:4414-4418).

Because of the tremendous potential of progenitor cells to differentiate into distinct lineages, there has always existed a need for a continuous source of these isolated pluripotent progenitor cells. The pluripotent progenitor cells could be extremely useful in the treatment of different disorders that are characterized by insufficient or abnormal functioning of the fully differentiated cells in a given organ, as for example in the human pancreas or liver.

Summary of the Invention

The present invention relates to substantially pure preparations of viable progenitor cells, and methods for isolating such cells. The present invention further concerns certain uses for such progenitor cells, and their progeny.

In general, the invention features a cellular composition including, as the cellular component, a substantially pure population of viable bile duct progenitor cells which progenitor cells are capable of proliferation in a culture medium. In a preferred embodiment, the cellular composition has fewer than about 20%, more preferably fewer than about 10%, most preferably fewer than about 5% of lineage committed cells.

In general, the progenitor cells of the present invention are proliferative cells which can differentiate into cells making up the tissues of the gut, e.g., the liver, pancreas, gallbladder, intestines, etc. That is, the progenitor cells can give rise to differentiated cells of hepatic, pancreatic, gallbladder or intestinal lineages. However, as described in the pending examples, the subject method can be used to isolate populations of hematopoietic stem/progenitor cells. In preferred embodiments, the subject progenitor cells are pluripotent, e.g., the progenitor cells are capable of differentiating into two or more distinct lineages.

In one embodiment, the progenitor cells of the present invention are characterized by an ability for self-regeneration in a culture medium and differentiation to pancreatic lineages. In a preferred embodiment, the progenitor cells are inducible to differentiate into pancreatic islet cells, e.g., β islet cells, α islet cells, δ islet cells, or ϕ islet cells. Such
 5 pancreatic progenitor cells may be characterized in certain circumstances by the expression of one or more of: homeodomain type transcription factors such as STF-1; PAX gene(s) such as PAX6; PTF-1; hXBP-1; HNF genes(s); villin; tyrosine hydroxylase; insulin; glucagon; and/or neuropeptide Y.

In another embodiment, the invention features progenitor cells, such as may be
 10 obtained from a non-hepatic bile duct according to the present invention, which are characterized by an ability to differentiate (e.g., by induction) into hepatocytes when maintained in culture. Exemplary hepatic progenitor cells may be characterized by the expression of one or more of: a hepatocyte nuclear factor (HNF) transcription factor, e.g., HNF1 α , HNF1 β , HNF3 β , HNF3 γ , and/or HNF4; ATBF1; AFP; a LIM type homeobox
 15 gene, such as Islet-1; a "forkhead" transcription factor, such as fkh-1; a CCAAT-enhancer binding protein (C/EBP), such as C/EBP- β ; oval cell marker OV-6; cytokeratins (such as type 7 or 19), mucin Span-1; OC.2; OC.3; and/or γ -glutamyl transpeptidase.

In still another embodiment, the invention features hematopoietic progenitor cells which may differentiate into one or more of an erythrocyte, a megakaryocyte, a monocyte, a
 20 granulocyte, and/or an eosinophils, as well as fully differentiated lymphoid cells such as B lymphocytes and T lymphocytes. Certain of the subject hematopoietic progenitor cells may be characterized by expression of such markers as c-kit, CD34 and/or CD33, as well as OV-6, CAM5.2 and/or cytokeratin type 7 or 18.

In yet another embodiment, the invention features a pharmaceutical composition
 25 including as the cellular component, a substantially pure population of viable bile duct progenitor cells, which progenitor cells are capable of proliferation in a culture medium.

In general, the preferred progenitor cells will be of mammalian origin, e.g., cells isolated from a primate such as a human, from a miniature swine, or from a transgenic mammal, or are the cell culture progeny of such cells.

30 In preferred embodiments, the subject progenitor cells can be maintained in cell/tissue culture for at least about 7 days, more preferably for at least about 14 days, most preferably for at least about 21 days or longer.

In another aspect, the invention features a cellular composition comprising, as a cellular population, at least 75% (though more preferably at least 80, 90 or 95%) progenitor
 35 cells isolated from a bile duct and capable of self-regeneration in a culture medium.

In yet another aspect, the invention features, a cellular composition consisting essentially of, as the cellular population, viable non-hepatic duct progenitor cells capable of

self-regeneration in a culture medium and differentiation to members of the hepatic, pancreatic and gallbladder lineages. For instance, in certain embodiments the progenitor cells are isolated from cystic duct explants, pancreatic duct explants, common bile duct explants, or are the cell culture progeny of such cells.

5 Another aspect of the invention features a method for isolating progenitor cells from a bile duct. In general, the method provides for culturing an isolated population of cells having a microarchitecture of a mammalian bile duct, e.g. a micro-organ explant in which the original epithelial-mesenchymal microarchitecture is maintained, wherein the dimensions of the explant provide the isolated population of cells as maintainable in culture
10 for at least twenty-four hours, and includes in the population of cells at least one progenitor cell which can proliferate under such culture conditions. The cultured cell population is contacted with an agent, e.g., a mitogenic agent such as a growth factor, which agent causes proliferation of progenitor cells in the cultured population. Subsequently, progenitor cells from the explant that proliferate in response to the agent are isolated, such as by direct
15 mechanical separation of newly emerging buds from the rest of the explant or by dissolution of all or a portion of the explant and subsequent isolation of the progenitor cell population.

In another preferred embodiment, the agent is a growth factor, e.g., the growth factor is selected from a group consisting of IGF, TGF, FGF, EGF, HGF, or VEGF. In other embodiments, the growth factor is a member of the TGF β superfamily, preferably of the
20 DVR (dpp and vg1 related) family, e.g., BMP2 and/or BMP7.

In another preferred embodiment, the population of cells is cultured in a medium deficient in biological extracts, e.g., deficient in serum.

In a preferred embodiment, the bile duct is a common bile duct.

In another aspect, the invention features, a method for screening a compound for
25 ability to modulate one of growth, proliferation, and/or differentiation of progenitor cells obtained from a bile duct, including: (i) establishing an isolated population of cells having a microarchitecture of a mammalian bile duct, e.g., a micro-organ explant in which the original epithelial-mesenchymal microarchitecture is maintained, wherein the dimensions of the explant provide the isolated population of cells as maintainable in culture for at least
30 twenty-four hours, and includes at least one progenitor cell which has the ability to proliferate in the culture; (ii) contacting the population of cells with a test compound; and (iii) detecting one of growth, proliferation, and/or differentiation of the progenitor cells in the population, wherein a statistically significant change in the extent of one of growth, proliferation, and/or differentiation in the presence of the test compound relative to the
35 extent of one of growth, proliferation, and/or differentiation in the absence of the test compound indicates the ability of the test compound to modulate one of the growth, proliferation, and/or differentiation.

In another aspect, the invention features, a method for treating a disorder characterized by insufficient insulin activity, in a subject, including introducing into the subject a pharmaceutical composition including pancreatic progenitor cells, or differentiated cells arising therefrom, and a pharmaceutically acceptable carrier.

5 In a preferred embodiment the subject is a mammal, e.g., a primate, e.g., a human.

In another preferred embodiment the disorder is an insulin dependent diabetes, e.g., type I diabetes.

10 In yet another preferred embodiment, the pancreatic progenitor cells are induced to differentiate into pancreatic islet cells, e.g., β islet cells, α islet cells, δ islet cells, or ϕ islet cells, subsequent to being introduced into the subject. Preferably, the pancreatic progenitors cells are induced to differentiate into pancreatic islet, e.g., β islet cells, α islet cells, δ islet cells, or ϕ islet cells, in culture prior to introduction into the subject.

15 In another aspect, the invention features, a method for treating a disorder characterized by insufficient liver function, in a subject, comprising introducing into the subject a pharmaceutical composition including hepatic progenitor cells and a pharmaceutically acceptable carrier.

In a preferred embodiment, the subject is a mammal, e.g., a primate, e.g., a human.

20 In another preferred embodiment the disorder is selected from the group consisting of cirrhosis, hepatitis B, hepatitis C, sepsis, or ELAD. In yet another preferred embodiment, the hepatic progenitor cells are induced to differentiate into hepatocytes subsequent to being introduced into the subject. Preferably, the hepatic progenitors cells are induced to differentiate into hepatocytes in culture prior to introduction into the subject.

25 The practice of the present invention will employ, unless otherwise indicated, conventional techniques of cell biology, cell culture, molecular biology, transgenic biology, microbiology, recombinant DNA, and immunology, which are within the skill of the art. Such techniques are described in the literature. See, for example, *Molecular Cloning A Laboratory Manual*, 2nd Ed., ed. by Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory Press: 1989); *DNA Cloning*, Volumes I and II (D. N. Glover ed., 1985); *Oligonucleotide Synthesis* (M. J. Gait ed., 1984); Mullis et al. U.S. Patent No: 4,683,195; 30 *Nucleic Acid Hybridization* (B. D. Hames & S. J. Higgins eds. 1984); *Transcription And Translation* (B. D. Hames & S. J. Higgins eds. 1984); *Culture Of Animal Cells* (R. I. Freshney, Alan R. Liss, Inc., 1987); *Immobilized Cells And Enzymes* (IRL Press, 1986); B. Perbal, *A Practical Guide To Molecular Cloning* (1984); the treatise, *Methods In Enzymology* (Academic Press, Inc., N.Y.); *Gene Transfer Vectors For Mammalian Cells* (J. H. Miller and M. P. Calos eds., 1987, Cold Spring Harbor Laboratory); *Methods In Enzymology*, Vols. 154 and 155 (Wu et al. eds.), *Immunochemical Methods In Cell And*

Molecular Biology (Mayer and Walker, eds., Academic Press, London, 1987); *Handbook Of Experimental Immunology*, Volumes I-IV (D. M. Weir and C. C. Blackwell, eds., 1986); *Manipulating the Mouse Embryo*, (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1986).

- 5 Other features and advantages of the invention will be apparent from the following detailed description, and from the claims.

Detailed Description of the Drawings

- 10 Figure 1 is a graph depicting average number of BrdU positive nuclei in a common bile duct explant, 24 hours after administration of a growth factor. Growth factors, EGF, TGF- α , and basic FGF (bFGF) were administered in three doses: 1 ng/ml, 10 ng/ml and 100 ng/ml. DMEM minimal media was used as a control. Average fluorescence intensity was determined based on the number of positive nuclei.

- 15 Figure 2 is a graph depicting the percentage of BrdU labeled positive cells in a common bile duct explant, 24 hours after administration of a growth factor. Growth factors, EGF, TGF- α , and bFGF were administered and DMEM minimal media was used as a control.

- 20 Figure 3 is a micrographs depicting *in vitro* growth and expansion of the cells within a common bile duct explant in response to administration of growth factor bFGF.

- Figures 4A-4C a micrographs (120X) illustrating the types of cells expanded from bile duct microorgan cultures by IGF-1 stimulation. The addition of IGF-1 at 100ng/ml resulted in at least three types of responses; no growth or change in morphology (Figure 4A); the appearance of "spiny" colonies which are termed "N-type" colonies due to the neurite-like appearance of the colony extensions (Figure 4B); and the appearance of "L-type" colonies for liver-like, due to the epithelial blebs and formation of red blood cell foci (Figure 4C) which were identical in appearance to cultured embryonic liver (not shown).

- 25 Figure 5 is a graph demonstrating that ductal microorgan cultures on matrigel can give rise to multiple colony types.

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Detailed Description of the Invention

- The ability to isolate distinct populations of progenitor cells has been an important problem in modern biology. It can be easily envisioned that such isolated pluripotent progenitor cells could be very useful for treatment of various disorders associated with loss or abnormal functioning of fully differentiated cells in a given organ. For example, the ability to introduce isolated progenitor cells capable of subsequent differentiation, either in

culture or when introduced into a subject, into functional β islet cells, would have important implications for the treatment of insulin-dependent diabetes. In the same manner, the ability to deliver purified hepatic progenitor cells, having the ability to differentiate into mature hepatocytes, could be potentially useful in the process of liver regeneration or for treatment of disorders characterized by insufficient liver function. Prior to the instant invention, there was no apparently reliable source of purified populations of progenitor cells from gut tissue capable of further differentiation into distinct pancreatic, hepatic, gallbladder, intestinal or hematopoietic lineages, either in culture or when introduced into a subject.

Accordingly, certain aspects of the present invention relate to isolated populations of progenitor cells capable of subsequent differentiation to distinct pancreatic, hepatic, gallbladder, intestinal or hematopoietic lineages, methods for isolating such cells and therapeutic uses for such cells.

For convenience, certain terms employed in the specification, examples, and appended claims are collected here.

The term "culture medium" is recognized in the art, and refers generally to any substance or preparation used for the cultivation of living cells. Accordingly, a "tissue culture" refers to the maintenance or growth of tissue, e.g., explants of organ primordia or of an adult organ *in vitro* so as to preserve its architecture and function. A "cell culture" refers to a growth of cells *in vitro*; although the cells proliferate they do not organize into tissue *per se*.

Tissue and cell culture preparations of the subject micro-organ explants and amplified progenitor cell populations can take on a variety of formats. For instance, a "suspension culture" refers to a culture in which cells multiply while suspended in a suitable medium. Likewise, a "continuous flow culture" refers to the cultivation of cells or ductal explants in a continuous flow of fresh medium to maintain cell growth, e.g. viability. The term "conditioned media" refers to the supernatant, e.g. free of the cultured cells/tissue, resulting after a period of time in contact with the cultured cells such that the media has been altered to include certain paracrine and/or autocrine factors produced by the cells and secreted into the culture.

The terms "explant" and "micro-organ explant" refer to a portion of an organ taken from the body and grown in an artificial medium.

The term "tissue" refers to a group or layer of similarly specialized cells which together perform certain special functions.

The term "organ" refers to two or more adjacent layers of tissue, which layers of tissue maintain some form of cell-cell and/or cell-matrix interaction to form a microarchitecture.

The term "lineage committed cell" refers to a progenitor cell that is no longer pluripotent but has been induce to differentiate into a specific cell type, e.g., a pancreatic, hepatic or intestinal cell.

5 The term "progenitor cell" refers to an undifferentiated cell which is capable of proliferation and giving rise to more progenitor cells having the ability to generate a large number of mother cells that can in turn give rise to differentiated, or differentiable daughter cells. As used herein, the term "progenitor cell" is also intended to encompass a cell which is sometimes referred to in the art as a "stem cell". In a preferred embodiment, the term "progenitor cell" refers to a generalized mother cell whose descendants (progeny)
10 specialize, often in different directions, by differentiation, e.g., by acquiring completely individual characters, as occurs in progressive diversification of embryonic cells and tissues. A "bile duct progenitor cells" refers to progenitor cells arising in tissue of a bile duct and giving rise to such differentiated progeny as, for example, hepatic, pancreatic, intestinal, gallbladder or hematopoietic lineages.

15 As used herein the term "bile duct" refers to an intricate system of ducts, e.g., generally tubular structures used for secretion, either neonatal or adult. The term includes the hepatic duct, cystic duct, and pancreatic duct. The term "pancreatic duct" includes the accessory pancreatic duct, dorsal pancreatic duct, main pancreatic duct and ventral pancreatic duct. The term bile duct also encompasses the common bile duct. The main
20 function of the bile duct is to allow bile and other materials to drain from these organs and enter the gastrointestinal tract.

As used herein the term "common bile duct" refers to a region of the bile duct, either adult or neonatal, originating from the liver bile canaliculi and extending down to the papilla of Vater at the duodenal junction. The common bile duct is continuous with hepatic,
25 cystic and certain pancreatic ducts.

As used herein the term "non-hepatic bile duct" refers to that bile duct tissue which is not hepatic duct tissue, e.g., those portions of the common bile duct posterior to the hepatic duct. In addition, the term includes cystic and pancreatic ducts.

As used herein the term "substantially pure", with respect to progenitor cells, refers to
30 a population of progenitor cells that is at least about 75%, preferably at least about 85%, more preferably at least about 90%, and most preferably at least about 95% pure, with respect to progenitor cells making up a total cell population. Recast, the term "substantially pure" refers to a population of progenitor cell of the present invention that contain fewer than about 20%, more preferably fewer than about 10%, most preferably fewer than about
35 5%, of lineage committed cells in the original unamplified and isolated population prior to subsequent culturing and amplification.

As used herein, the term "cellular composition" refers to a preparation of cells, which preparation may include, in addition to the cells, non-cellular components such as cell culture media, e.g. proteins, amino acids, nucleic acids, nucleotides, co-enzyme, anti-oxidants, metals and the like. Furthermore, the cellular composition can have components which do not affect the growth or viability of the cellular component, but which are used to provide the cells in a particular format, e.g., as polymeric matrix for encapsulation or a pharmaceutical preparation.

As used herein the term "animal" refers to mammals, preferably mammals such as humans. Likewise, a "patient" or "subject" to be treated by the method of the invention can mean either a human or non-human animal.

As described below, in a preferred embodiment, the progenitor cells of the present invention are pancreatic or hepatic progenitor cells. The term "pancreas" is art recognized, and refers generally to a large, elongated, racemose gland situated transversely behind the stomach, between the spleen and duodenum. The pancreatic exocrine function, e.g., external secretion, provides a source of digestive enzymes. Indeed, "pancreatin" refers to a substance from the pancreas containing enzymes, principally amylase, protease, and lipase, which substance is used as a digestive aid. The exocrine portion is composed of several serous cells surrounding a lumen. These cells synthesize and secrete digestive enzymes such as trypsinogen, chymotrypsinogen, carboxypeptidase, ribonuclease, deoxyribonuclease, triacylglycerol lipase, phospholipase A₂, elastase, and amylase.

The endocrine portion of the pancreas is composed of the islets of Langerhans. The islets of Langerhans appear as rounded clusters of cells embedded within the exocrine pancreas. Four different types of cells- α , β , δ , and ϕ -have been identified in the islets. The α cells constitute about 20% of the cells found in pancreatic islets and produce the hormone glucagon. Glucagon acts on several tissues to make energy available in the intervals between feeding. In the liver, glucagon causes breakdown of glycogen and promotes gluconeogenesis from amino acid precursors. The δ cells produce somatostatin which acts in the pancreas to inhibit glucagon release and to decrease pancreatic exocrine secretion. The hormone pancreatic polypeptide is produced in the ϕ cells. This hormone inhibits pancreatic exocrine secretion of bicarbonate and enzymes, causes relaxation of the gallbladder, and decreases bile secretion. The most abundant cell in the islets, constituting 60-80% of the cells, is the β cell, which produces insulin. Insulin is known to cause the storage of excess nutrients arising during and shortly after feeding. The major target organs for insulin are the liver, muscle, and fat-organs specialized for storage of energy.

The term "pancreatic progenitor cell" refers to a cell which can differentiate into a cell of pancreatic lineage, e.g. a cell which can produce a hormone or enzyme normally produced by a pancreatic cell. For instance, a pancreatic progenitor cell may be caused to differentiate, at least partially, into α , β , δ , or ϕ islet cell, or a cell of exocrine fate. The

pancreatic progenitor cells of the invention can also be cultured prior to administration to a subject under conditions which promote cell proliferation and differentiation. These conditions include culturing the cells to allow proliferation and confluence *in vitro* at which time the cells can be made to form pseudo islet-like aggregates or clusters and secrete insulin, glucagon, and somatostatin.

The term "liver" refers to the large, dark-red gland in the upper part of the abdomen on the right side, just beneath the diaphragm. Its manifold functions include storage and filtration of blood, secretion of bile, conversion of sugars into glycogen, and many other metabolic activities.

The liver is a gland that supplies bile to intestine. In adult vertebrates, this function is a minor one, but the liver originally arose as a digestive gland in lower chordates. Throughout the liver, a network of tiny tubules collects bile--a solution of salts, bilirubin (made when hemoglobin from red blood cells is broken down in liver), and fatty acids. Bile accumulates in the gall bladder, which empties into the small intestine by way of a duct. Bile has two functions in the intestine. First, it acts as a detergent, breaking fat into small globules that can be attacked by digestive enzymes. Second, and more important, bile salts aid in the absorption of lipids from the intestine; removal of the gall bladder sometimes causes difficulty with lipid absorption.

Digested food molecules absorbed into the bloodstream from the intestine pass directly to the liver by way of the hepatic portal vein. Before these molecules pass on into the rest of the body, the liver may change their concentration and even their chemical structure. The liver performs a vital role in detoxifying otherwise poisonous substances. In addition, it stores food molecules that reach it from the intestine, converts them biochemically, and releases them back into the blood at a controlled rate. For instance, the liver removes glucose from the blood under the influence of the hormone insulin and stores it as glycogen. When the level of glucose in the blood falls, the hormone glucagon causes the liver to break down glycogen and release glucose into the blood.

The liver also synthesizes many of the blood proteins (e.g., albumins) and releases them into the blood when they are needed. In addition, the liver converts nitrogenous wastes into the form of urea for excretion by the kidneys. With the kidneys, the liver is vital in regulating what the blood contains when it reaches all the other organs of the body. Because the liver is the body's major organ for making all these biochemical adjustments, severe liver damage or loss of the liver is rapidly fatal.

The term "hepatic progenitor cell" as used herein refers to a cell which can differentiate in a cell of hepatic lineage, such a liver parenchymal cell, e.g., a hepatocyte. Hepatocytes are some of the most versatile cells in the body. Hepatocytes have both endocrine and exocrine functions, and synthesize and accumulate certain substance.

detoxify others, and secrete others to perform enzymatic, transport, or hormonal activities. The main activities of liver cells include bile secretion, regulation of carbohydrate, lipid, and protein metabolism, storage of substances important in metabolism, degradation and secretion of hormones, and transformation and excretion of drugs and toxins. The hepatic progenitor cells of the invention can also be cultured prior to administration to a subject under conditions which promote cell proliferation and differentiation.

The term "hematopoietic cells" herein refers to fully differentiated myeloid cells such as erythrocytes or red blood cells, megakaryocytes, monocytes, granulocytes, and eosinophils, as well as fully differentiated lymphoid cells such as B lymphocytes and T lymphocytes. Thus, a hematopoietic stem/progenitor cell includes the various hematopoietic precursor cells from which these differentiated cells develop, such as BFU-E (burst-forming units-erythroid), CFU-E (colony forming unit-erythroid), CFU-Meg (colony forming unit-megakaryocyte), CFU-GM (colony forming unit-granulocyte-monocyte), CFU-Eo (colony forming unit-eosinophil), and CFU-GEMM (colony forming unit-granulocyte-erythrocyte-megakaryocyte-monocyte).

Certain terms being set out above, it is noted that one aspect of the present invention features a method for isolating progenitor cells from micro-organ explants, e.g., ductal tissue explants. A salient feature of the subject method concerns the use of defined explants as sources from which discrete progenitor cell populations can be amplified. For instance, as described below, the progenitor source ductal tissue explants preferably are derived with dimensions that allow the explanted tissue to maintain its microarchitecture and biological function for prolonged periods of time in culture, e.g., the dimensions of the explant preserve the normal tissue architecture and at least a portion of the normal tissue function that is present *in vivo*. Such tissue explants can be maintained, for instance, in minimal culture media for extended periods of time (e.g., for 21 days or longer) and can be contacted with different factors. Accordingly, carefully defined conditions can be acquired in the culture so as selectively activate discrete populations of cells in the tissue explant. The progenitor cells of the present invention can be amplified, and subsequently isolated from the explant, based on a proliferative response upon, for example, addition of defined growth factors or biological extracts to the culture.

In general, the method of the present invention can be used to isolate progenitor cells from a bile duct explant by steps beginning with the culturing of an isolated population of cells having a microarchitecture of a mammalian bile duct, e.g. a micro-organ explant in which the original epithelial-mesenchymal microarchitecture of the originating duct is maintained, wherein the dimensions of the explant provide the isolated population of cells as maintainable in culture for at least twenty-four hours, and includes in the population of cells at least one progenitor cell which can proliferate under such culture conditions. The explant is contacted with an agent, e.g., a mitogenic agent such as a growth factor or other

biological extract, which agent causes proliferation of progenitor cells in the cultured population. Subsequently, progenitor cells from the explant that proliferate in response to the agent are isolated, such as by direct mechanical separation from the rest of the explant or by dissolution of all or a portion of the explant and subsequent isolation of the progenitor cell population.

In a illustrative embodiment, the size of the particular ductal explant will depend largely on (i) the availability of tissue, and (ii) a need for similar availability of nutrients to all cells in the tissue by diffusion. In a preferred embodiment, the ductal tissue explant is selected to provide diffusion of adequate nutrients and O₂ to every cell in a three dimensional organ. Accordingly, the size of the explant is determined by the requirement for a minimum level of accessibility to each cell absent specialized delivery structures or synthetic substrates.

A salient feature of the micro-organ cultures used to in the subject methods, according to the invention, is the ability to preserve the cellular microenvironment found *in vivo* for a ductal tissue. The invention is based, in part, upon the discovery that under certain circumstances growth of cells of both a stromal and epithelial layer, provided together in the same explant, will sustain active proliferation of cells of each layer. Moreover, the cell-cell and cell-matrix interactions provided in the explant itself are sufficient to support cellular homeostasis, e.g., maturation, differentiation and segregation of cells in the explant culture, thereby sustaining the microarchitecture and function of the explant for prolonged periods of time.

An example of physical contact between a cell and a noncellular substrate (matrix) is the physical contact between an epithelial cell and its basal lamina. An example of physical contact between a cell and another cell includes actual physical contact maintained by, for example, intercellular cell junctions such as gap junctions and tight junctions. Examples of functional contact between one cell and another cell includes electrical or chemical communication between cells. In addition, many cells communicate with other cells via chemical messages, e.g., hormones, which either diffuse locally (paracrine signalling and autocrine signalling), or are transported by the vascular system to more remote locations (endocrine signalling).

Not wishing to be bound by any particular theory, this microarchitecture of the ductal explants can be extremely important for the maintenance of the explant in minimal media, e.g., without exogenous sources of serum or growth factors, as the ductal explants can apparently be sustained in such minimal media by paracrine factors resulting from specific cellular interactions within the sample. Moreover, there is also a possibility that certain growth factors might act indirectly by activating cells, other than progenitor cells, to produce mitogenic factors that subsequently cause proliferation of progenitor cells within the explant. Accordingly, the ductal explants are derived such that they comprise both an

epithelial layer and a stromal layer, and maintain, *in vitro*, the physical and/or functional interaction between these two component of the explant.

However, the phrase "maintain, *in vitro*, the physical and/or functional interaction" is not intended to exclude an isolated population of cells in which at least one cell develops physical and/or functional contact with at least one cell or noncellular substance with which it is not in physical and/or functional contact *in vivo*. An example of such a development is of course proliferation of at least one cell of the isolated population of cells.

As emphasized through the present application, the micro-organ cultures used to prepare progenitor cells according to the invention preserve the normal tissue architecture that is present *in vivo*, e.g., the original epithelial-mesenchymal organization. In preferred embodiments, the populations of cells of the ductal explants are grouped in a manner that preserves the natural affinity of one cell to another, e.g., to preserve layers of different cells present in explant. Such an association facilitates intercellular communication. Many types of communication takes place among animal cells. This is particularly important in differentiating cells where induction is defined as the interaction between one (inducing) and another (responding) tissue or cell, as a result of which the responding cells undergo a change in the direction of differentiation. Moreover, inductive interactions occur in embryonic and adult cells and can act to establish and maintain morphogenetic patterns as well as induce differentiation (Gurdon (1992) Cell 68: 185-199). Accordingly, an exemplary micro-organ cultures prepared in accordance to use in the progenitor amplification method of the invention are described in Example I, and include a epithelial and mesenchymal cells grouped in a manner that includes a plurality of layers so as to preserve the natural affinity and interaction of one cell to another in and between each layer.

In addition to isolating a ductal explant which retains the cell-cell and cell-matrix architecture of the originating duct, the dimensions of the explant are important to the viability of the cells therein, e.g., where the micro-organ culture is intended to be sustained for prolonged periods of time, e.g., 7-21 days or longer. Accordingly, the dimensions of the explant are selected to provide diffusion of adequate nutrients and gases (e.g, O₂, CO₂, etc) to every cell in the three dimensional micro-organ explant, as well as diffusion of cellular waste out of the explant so as to minimize cellular toxicity and concomitant death due to localization of the waste in the micro-organ. Thus, in addition to the requirement of both epithelial and mesenchymal components, the size of the explant is determined by the requirement for a minimum level of accessibility to each cell in the absence specialized delivery structures or synthetic substrates. As described herein, this accessibility can be maintained if Aleph, an index calculated from the thickness and the width of the explant, is at least greater than approximately 1.5 mm⁻¹. As used herein, a surface to area index, "Aleph" is defined as $Aleph = \frac{1}{x} + \frac{1}{a} > 1.5mm^{-1}$; wherein x= radial thickness and a= the axial length of the duct explant in millimeters. Accordingly, the present invention provides

that the surface area to volume index of the tissue explant is maintained within a selected range. This selected range of surface area to volume indice provides the cells access to nutrients and to avenues of waste disposal by diffusion in a manner similar to cells of the *in vivo* organ from which the explant originated.

- 5 Examples of Aleph are provided in Table I wherein, for example, an explant having a thickness (x) of 0.1mm and a width (a) of 1mm would have an Aleph index of 11. In another instance, if $x = 0.3 \text{ mm}$ and $a = 4 \text{ mm}$, the Aleph is 3.48 mm^{-1} . To further illustrate, Applicant has observed that when x is varied and a is constant at 4mm, the proliferative activity of cells in a cultured explant is substantially reduced as the thickness of the explant increases. Accordingly, at $900 \mu\text{m}$ thickness, the number of proliferating cells in a micro-organ culture was found to about 10 fold less than in tissue from a similar source having a thickness of $300 \mu\text{m}$. The Aleph index for a tissue having a thickness of $900 \mu\text{m}$ is 1.36 mm^{-1} , below the minimum described herein whereas the Aleph index for tissue having a thickness of $300 \mu\text{m}$ is 3.58 mm^{-1} which is well within the range defined herein.

15

TABLE I: Different values for the surface area to volume ratio index "Aleph", as a function of a (width) and x (thickness) in mm^{-1}

		← Radial Length →				
↑ thickness ↓	x(mm)	a=1mm	a=2mm	a=3mm	a=4mm	a=5mm
	0.1	11	10.5	10.33	10.25	10.2
	0.2	6	5.5	5.33	5.25	5.2
	0.3	4.3	3.83	3.67	3.58	3.53
	0.4	3.5	3	2.83	2.75	2.7
	0.5	3	2.5	2.33	2.25	2.2
	0.6	2.66	2.16	2	1.91	1.87
	0.7	2.4	1.92	1.76	1.68	1.63
	0.8	2.25	1.75	1.58	1.5	1.45
	0.9	2.11	1.61	1.44	1.36	1.31
	1	2	1.5	1.33	1.25	1.2
	1.2	1.83	1.3	1.16	1.08	1.03
	1.3	1.77	1.26	1.1	1.02	0.96
	1.6	1.625	1.13	0.96	0.88	0.83
	2	1.5	1	0.83	0.75	0.7
			↑ Aleph ↓			

20

Again, not wishing to be bound by any particular theory, a number of factors provided by the three-dimensional culture system may contribute to its success in the subject method of activating progenitor cell populations:

(a) The appropriate choice of the explant size, vis-à-vis the use of the above Aleph calculations, three-dimensional matrix provides appropriate surface area to volume ratio for adequate diffusion of nutrients to all cells of the explant, and adequate diffusion of cellular waste away from all cells in the explant.

5 (b) Because of the three-dimensionality of the matrix, various cells continue to actively grow, in contrast to cells in monolayer cultures, which grow to confluence, exhibit contact inhibition, and cease to grow and divide. The elaboration of growth and regulatory factors by replicating cells of the explant may be partially responsible for stimulating proliferation and regulating differentiation of cells in culture.

10 (c) The three-dimensional matrix retains a spatial distribution of cellular elements, e.g., the epithelial-mesenchymal microarchitecture, which closely approximates that found in the counterpart tissue *in vivo*.

(d) The cell-cell and cell-matrix interactions may allow the establishment of localized microenvironments conducive to cellular induction and/or maturation. It has been
15 recognized that maintenance of a differentiated cellular phenotype requires not only growth/differentiation factors but also the appropriate cellular interactions. The present invention effectively mimics the microenvironment. Accordingly, the micro-organ preserves interactions which may be required to maintain the cells supporting the progenitor cells, the cells (if any) providing inductive signals to the progenitor cells, and the progenitor
20 cells themselves.

To further illustrate, the appended examples demonstrate that microorgan explants, in which the original epithelial-mesenchymal microarchitecture of the common bile duct are prepared by transverse sectioning of the duct every 300 μm , can be maintained in simple media (DMEM) for several weeks and can be used to isolate progenitor cells of the present
25 invention by growth induction upon contact with certain growth factors.

There are a large number of tissue culture media that exist for culturing tissue from animals. Some of these are complex and some are simple. While it is expected that the ductal explants may grow in complex media, it will generally be preferred that the explants be maintained in a simple medium, such as Dulbecco's Minimal Essential Media (DMEM),
30 in order to effect more precise control over the activation of certain progenitor populations in the explant. Furthermore, although the cultures may be grown in a media containing sera or other biological extracts such as pituitary extract, it has been demonstrated that neither sera nor any other biological extract is required for explants derived according to the above considerations (see U.S.S.N. 08/341,409.). Moreover, the explants can be maintained in the
35 absence of sera for extended periods of time. In preferred embodiments of the invention, the growth factors or other mitogenic agents are not included in the primary media for

maintenance of the cultures *in vitro*, but are used subsequently to cause proliferation of distinct populations of progenitor cells. See the appended examples.

The tissue explants may be maintained in any suitable culture vessel, such as a 12 or 24 well microplate, and may be maintained under typical culture conditions for cells isolated from the same animal, e.g., such as 37°C in 5% CO₂. The cultures may be shaken for improved aeration, the speed of shaking being, for example, 12 rpm.

In order to isolate progenitor cells from the ductal explants, it will generally be desirable to contact the explant with an agent which causes proliferation of one or more populations of progenitor cells in the explant. For instance, a mitogen, e.g., a substance that induces mitosis and cell transformation, can be used to detect a progenitor cell population in the explant, and where desirable, to cause the amplification of that population. To illustrate, a purified or semi-purified preparation of a growth factor can be applied to the culture. Induction of progenitor cells which respond to the applied growth factor can be detected by proliferation of the progenitor cells. However, as described below, amplification of the population need not occur to a large extent in order to use certain techniques for isolating the responsive population.

In yet other embodiments, the ductal explants and/or amplified progenitor cells can be cultured on feeder layers, e.g., layers of feeder cells which secrete inductive factors or polymeric layers containing inductive factors. For example, a matrigel layer can be used to induce hematopoietic progenitor cell expansion, as described in the appended examples. Matrigel (Collaborative Research, Inc., Bedford, Mass.) is a complex mixture of matrix and associated materials derived as an extract of murine basement membrane proteins, consisting predominantly of laminin, collagen IV, heparin sulfate proteoglycan, and nidogen and entactin was prepared from the EHS tumor as described Kleinman et al. "Basement Membrane Complexes with Biological Activity", Biochemistry, Vol. 25 (1986), pages 312-318. Other such matrixes can be provided, such as Humatrix. Likewise, natural and recombinantly engineered cells can be provided as feeder layers to the instant cultures.

Methods of measuring cell proliferation are well known in the art and most commonly include determining DNA synthesis characteristic of cell replication. There are numerous methods in the art for measuring DNA synthesis, any of which may be used according to the invention. In an embodiment of the invention, DNA synthesis has been determined using a radioactive label (³H-thymidine) or labeled nucleotide analogues (BrdU) for detection by immunofluorescence.

However, in addition to measuring DNA synthesis, morphological changes can be, and preferably will be, relied on as the basis for isolating responsive progenitor cell populations. For instance, as described in the appended examples, we have observed that certain growth factors cause amplification of progenitor cells in ductal explants so as to form structures that

can be easily detected by the naked eye or microscopy. In an exemplary embodiment, those progenitor cells which respond to growth factors by proliferation and subsequent formation of outgrowths from the explant, e.g., buds or blebs, can be easily detected. In another illustrative embodiment, other structural changes, e.g., changes in optical density of proliferating cells, can be detected via contrast microscopy.

Various techniques may be employed to isolate the activated progenitor cells of treated explant. Preferred isolation procedures for progenitor cells are the ones that result in as little cell death as possible. For example, the activated progenitor cells can be removed from the explant sample by mechanical means, e.g., mechanically sheared off with a pipette. In other instances, it will be possible to dissociate the progenitor cells from the entire explant, or sub-portion thereof, e.g., by enzymatic digestion of the explant, followed by isolation of the activated progenitor cell population based on specific cellular markers, e.g., using affinity separation techniques or fluorescence activated cell sorting (FACS).

To further illustrate, the examples below demonstrate that ductal explants contain growth factor responsive progenitor cell types. It is further demonstrated that different growth factors can induce/amplify distinct populations of progenitor cells within the ductal tissue explant to proliferate. This indicates the presence of specific growth factor receptors on the surface of distinct progenitor cell populations. This is important because the expression of these receptors marks the progenitor cell populations of interest. Monoclonal antibodies are particularly useful for identifying markers (surface membrane proteins, e.g., receptors) associated with particular cell lineages and/or stages of differentiation. Procedures for separation of the subject progenitor cell may include magnetic separation, using antibody coated magnetic beads, affinity chromatography, and "panning" with antibody attached to a solid matrix, e.g., plate, or other convenient technique. Techniques providing accurate separation include fluorescence activated cell sorting, which can have varying degrees of sophistication, e.g., a plurality of color channels, low angle and obtuse light scattering detecting channels, impedance channels, etc.

Conveniently, the antibodies may be conjugated with markers, such as magnetic beads, which allow for direct separation, biotin, which can be removed with avidin or streptavidin bound to a support, fluorochromes, which can be used with a fluorescence activated cell sorter, or the like, to allow for ease of separation of the particular cell type. Any technique may be employed which is not unduly detrimental to the viability of the cells.

In an illustrative embodiment, some of the antibodies for growth factor receptors that exist on the subject progenitor cells are commercially available (e.g., antibodies for EGF receptors, FGF receptors and/or TGF receptors), and for other growth factor receptors, antibodies can be made by methods well known to one skilled in the art. In addition to using antibodies to isolate progenitor cells of interest, one skilled in the art can also use the growth factors themselves to label the cells, for example, to permit "panning" processes.

Upon isolation, the progenitor cells of the present invention can be further characterized in the following manner: responsiveness to growth factors, specific gene expression, antigenic markers on the surface of such cells, and/or basic morphology.

For example, extent of growth factor responsivity, e.g., the concentration range of growth factor to which they will respond to, the maximal and minimal responses, and to what other growth factors and conditions to which they might respond, can be used to characterize the subject progenitor cells.

Furthermore, the isolated progenitor cells can be characterized by the expression of genes known to mark the developing (i.e., stem or progenitor) cells for liver, pancreas, gall bladder and intestine.

In an illustrative embodiment, the hepatocyte nuclear factor (HNF) transcription factor family, e.g., HNF1-4, are known to be expressed in various cell types at various times during liver and pancreas development. For example, the progenitor cell may express one or more HNF protein such as HNF1 α , HNF1 β , HNF3 β , HNF3 γ , and/or HNF4. ATBF1, a regulator of alphafetoprotein (AFP) gene expression (AFP is expressed early in liver development and is reexpressed in many liver carcinomas) is believed to be a transient marker of liver stem cells. The LIM type homeobox genes, such as Islet-1, are also known to be expressed during liver development. Glut2 is a marker for both early pancreatic and liver cells. Certain of the "forkhead" transcription factors, such as fkh-1 or the like, are understood to be markers in early gut tissue. Likewise, members of the CCAAT-enhancer binding protein (C/EBP) family, such as C/EBP- β , are markers for early liver development. Therefore, expression of one or more of these genes can be used to further characterize the hepatic progenitor cells.

In another illustrative embodiment, homeodomain type transcription factors such as STF-1 (also known as IPF-1, IDX-1 or PDX) have recently been shown to mark different populations of the developing pancreas. Some LIM genes have also been shown to regulate insulin gene expression and would also be markers for protodifferentiated β islet cells. Likewise, certain of the PAX genes, such as PAX6, are expressed during pancreas formation and may be used to characterize certain pancreatic progenitor cell populations. Other markers of pancreatic progenitor cells include the pancreas specific transcription factor PTF-1, and hXBP-1 and the like. Moreover, certain of the HNF proteins are expressed during early pancreas development and may be used as markers for pancreatic progenitor cells.

In the intestine, even though there are not very many lineage specific transcription factors that have been mapped to the gut, the exception is the Hox B (homeobox genes) gene family (and possibly others) which are regional type markers rather than cell type specific, but which can be used to characterize progenitor cells originating from the specific

region. e.g., the gut. Elastase is known to be any early marker of duodenal development and hence can be a candidate early marker for subject progenitor cells.

Certain of the subject hematopoietic progenitor cells may be characterized by expression of such markers as c-kit, CD34 and/or CD33, as well as OV-6, CAM5.2 and/or cytokeratin type 7 or 18.

The subject progenitor cells can also be characterized on the basis of specific antigenic markers or other markers that may be expressed on the cell surface, e.g., integrins, lectins, gangliosides, or transporters, or on the basis of specific cellular morphology. All of these techniques are known and available to the one skilled in the art. For example, certain of the subject hepatic and ductal progenitor cells may express the oval cell marker OV-6. Other progenitor cells isolated from the ductal microorgan cultures may additionally, or alternatively, express one or more markers including hematopoietic markers, cytokeratins (such as type 7 or 19), mucin Span-1, OC.2, OC.3 or γ -glutamyl transpeptidase. Progenitor cells giving rise to pancreatic cells may express such as markers as villin and/or tyrosine hydroxylase, as well as secrete such factors as insulin, glucagon and/or neuropeptide Y.

Once isolated and characterized, the subject progenitor cells can be cultured under conditions which allow further differentiation into specific cell lineages, e.g., hepatic, pancreatic, gallbladder, intestinal, or hematopoietic lineages. This can be achieved through a paradigm of induction that can be developed. For example, the subject progenitor cells can be recombined with the corresponding embryonic tissue to see if the embryonic tissue can instruct the adult cells to codevelop and codifferentiate. Alternatively, the progenitor cells can be contacted with one or more growth or differentiation factors which can induce differentiation of the cells. For instance, the cells can be treated with a TGF β , such as DVR sub-family member.

Furthermore, it has become apparent, from the prolonged viability of the explanted bile duct fragments in minimal media, that the tissues making up the ductal explants are themselves producing certain factors, e.g. paracrine and/or autocrine. Accordingly, such conditioned media generated by the explants in culture can be used to further maintain the progenitor cells in culture subsequent to isolation from the explant. The subject progenitor cells can be cultured in contact with the corresponding bile duct explant, or in the conditioned media produced by such explant.

In another preferred embodiment, the subject progenitor cells can be implanted into one of a number of regeneration models used in the art, e.g., partial pancreatectomy or streptozocin treatment of a host animal.

Accordingly, another aspect of the present invention pertains to the progeny of the subject progenitor cells, e.g. those cells which have been derived from the cells of the initial explant culture. Such progeny can include subsequent generations of progenitor cells, as

well as lineage committed cells. e.g., hepatic, pancreatic or gallbladder cells generated by inducing differentiation of the subject progenitor cells after their isolation from the explant, e.g., induced *in vitro*.

Yet another aspect of the present invention concerns cellular compositions which
 5 include, as a cellular component, substantially pure preparations of the subject progenitor cells, or the progeny thereof. Cellular compositions of the present invention include not only substantially pure populations of the progenitor cells, but can also include cell culture components, e.g., culture media including amino acids, metals, coenzyme factors, as well as
 10 small populations of non-progenitor cells, e.g., some of which may arise by subsequent differentiation of isolated progenitor cells of the invention. Furthermore, other non-cellular components include those which render the cellular component suitable for support under particular circumstances, e.g., implantation, e.g., continuous culture.

As common methods of administering the progenitor cells of the present invention to subjects, particularly human subjects, which are described in detail herein, include injection
 15 or implantation of the cells into target sites in the subjects, the cells of the invention can be inserted into a delivery device which facilitates introduction by, injection or implantation, of the cells into the subjects. Such delivery devices include tubes, e.g., catheters, for injecting cells and fluids into the body of a recipient subject. In a preferred embodiment, the tubes additionally have a needle, e.g., a syringe, through which the cells of the invention
 20 can be introduced into the subject at a desired location. The progenitor cells of the invention can be inserted into such a delivery device, e.g., a syringe, in different forms. For example, the cells can be suspended in a solution or embedded in a support matrix when contained in such a delivery device. As used herein, the term "solution" includes a pharmaceutically acceptable carrier or diluent in which the cells of the invention remain
 25 viable. Pharmaceutically acceptable carriers and diluents include saline, aqueous buffer solutions, solvents and/or dispersion media. The use of such carriers and diluents is well known in the art. The solution is preferably sterile and fluid to the extent that easy syringability exists. Preferably, the solution is stable under the conditions of manufacture and storage and preserved against the contaminating action of microorganisms such as
 30 bacteria and fungi through the use of, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. Solutions of the invention can be prepared by incorporating progenitor cells as described herein in a pharmaceutically acceptable carrier or diluent and, as required, other ingredients enumerated above, followed by filtered sterilization.

35 Support matrices in which the progenitor cells can be incorporated or embedded include matrices which are recipient-compatible and which degrade into products which are not harmful to the recipient. Natural and/or synthetic biodegradable matrices are examples of such matrices. Natural biodegradable matrices include plasma clots, e.g., derived from a

mammal. and collagen matrices. Synthetic biodegradable matrices include synthetic polymers such as polyanhydrides, polyorthoesters, and polylactic acid. Other examples of synthetic polymers and methods of incorporating or embedding cells into these matrices are known in the art. See e.g., U.S. Patent No. 4,298,002 and U.S. Patent No. 5,308,701.

- 5 These matrices provide support and protection for the fragile progenitor cells *in vivo* and are, therefore, the preferred form in which the progenitor cells are introduced into the recipient subjects.

The present invention also provides substantially pure progenitor cells which can be used therapeutically for treatment of various disorders associated with insufficient
10 functioning of the pancreas or liver.

To illustrate, the subject progenitor cells can be used in the treatment of a variety of pancreatic disorders, both exocrine and endocrine. For instance, the progenitor cells can be used to produce populations of differentiated pancreatic cells for repair subsequent to partial pancreatectomy, e.g., excision of a portion of the pancreas. Likewise, such cell populations
15 can be used to regenerate or replace pancreatic tissue loss due to, pancreatolysis, e.g., destruction of pancreatic tissue, such as pancreatitis, e.g., a condition due to autolysis of pancreatic tissue caused by escape of enzymes into the substance.

In an exemplary embodiment, the subject progenitor cells can be provided for patients suffering from any insulin-deficiency disorder. For instance, each year, over 728,000 new
20 cases of diabetes are diagnosed and 150,000 Americans die from the disease and its complications; the total yearly cost in the United States is over 20 billion dollars (Langer et al. (1993) *Science* 260:920-926). Diabetes is characterized by pancreatic islet destruction or dysfunction leading to loss of glucose control. Diabetes mellitus is a metabolic disorder defined by the presence of chronically elevated levels of blood glucose (hyperglycemia).
25 Insulin-dependent (Type 1) diabetes mellitus ("IDDM") results from an autoimmune-mediated destruction of the pancreatic β -cells with consequent loss of insulin production, which results in hyperglycemia. Type 1 diabetics require insulin replacement therapy to ensure survival. Non-insulin-dependent (Type 2) diabetes mellitus ("NIDDM") is initially characterized by hyperglycemia in the presence of higher-than-normal levels of plasma
30 insulin (hyperinsulinemia). In Type 2 diabetes, tissue processes which control carbohydrate metabolism are believed to have decreased sensitivity to insulin. Progression of the Type 2 diabetic state is associated with increasing concentrations of blood glucose, and coupled with a relative decrease in the rate of glucose-induced insulin secretion.

The primary aim of treatment in both forms of diabetes mellitus is the same, namely,
35 the reduction of blood glucose levels to as near normal as possible. Treatment of Type 1 diabetes involves administration of replacement doses of insulin. In contrast, treatment of Type 2 diabetes frequently does not require administration of insulin. For example, initial therapy of Type 2 diabetes may be based on diet and lifestyle changes augmented by

therapy with oral hypoglycemic agents such as sulfonylurea. Insulin therapy may be required, however, especially in the later stages of the disease, to produce control of hyperglycemia in an attempt to minimize complications of the disease, which may arise from islet exhaustion.

- 5 More recently, tissue-engineering approaches to treatment have focused on transplanting healthy pancreatic islets, usually encapsulated in a membrane to avoid immune rejection. Three general approaches have been tested in animal models. In the first, a tubular membrane is coiled in a housing that contained islets. The membrane is connected to a polymer graph that in turn connects the device to blood vessels. By
10 manipulation of the membrane permeability, so as to allow free diffusion of glucose and insulin back and forth through the membrane, yet block passage of antibodies and lymphocytes, normoglycemia was maintained in pancreatectomized animals treated with this device (Sullivan et al. (1991) *Science* 252:718).

- In a second approach, hollow fibers containing islet cells were immobilized in the
15 polysaccharide alginate. When the device was place intraperitoneally in diabetic animals, blood glucose levels were lowered and good tissue compatibility was observed (Lacey et al. (1991) *Science* 254:1782).

- Finally, islets have been placed in microcapsules composed of alginate or polyacrylates. In some cases, animals treated with these microcapsules maintained
20 normoglycemia for over two years (Lim et al. (1980) *Science* 210:908; O'Shea et al. (1984) *Biochim. Biochys. Acta.* 840:133; Sugamori et al. (1989) *Trans. Am. Soc. Artif. Intern. Organs* 35:791; Levesque et al. (1992) *Endocrinology* 130:644; and Lim et al. (1992) *Transplantation* 53:1180). However, all of these transplantation strategies require a large, reliable source of donor islets.

- 25 The pancreatic progenitor cells of the invention can be used for treatment of diabetes because they have the ability to differentiate into cells of pancreatic lineage, e.g., β islet cells. The progenitor cells of the invention can be cultured *in vitro* under conditions which can further induce these cells to differentiate into mature pancreatic cells, or they can undergo differentiation in vivo once introduced into a subject. Many methods for
30 encapsulating cells are known in the art. For example, a source of β islet cells producing insulin is encapsulated in implantable hollow fibers. Such fibers can be pre-spun and subsequently loaded with the β islet cells (Aebischer et al. U.S. Patent No. 4,892,538; Aebischer et al. U.S. Patent No. 5,106,627; Hoffman et al. (1990) *Expt. Neurobiol.* 110:39-44; Jaeger et al. (1990) *Prog. Brain Res.* 82:41-46; and Aebischer et al. (1991) *J. Biomech. Eng.* 113:178-183), or can be co-extruded with a polymer which acts to form a polymeric
35 coat about the β islet cells (Lim U.S. Patent No. 4,391,909; Sefton U.S. Patent No. 4,353,888; Sugamori et al. (1989) *Trans. Am. Artif. Intern. Organs* 35:791-799; Sefton et al.

(1987) *Biotechnol. Bioeng.* 29:1135-1143; and Aebischer et al. (1991) *Biomaterials* 12:50-55).

Moreover, in addition to providing a source of implantable cells, either in the form of the progenitor cell population of the differentiated progeny thereof, the subject cells can be
5 used to produce cultures of pancreatic cells for production and purification of secreted factors. For instance, cultured cells can be provided as a source of insulin. Likewise, exocrine cultures can be provided as a source for pancreatin.

The liver is an organ that is vulnerable to a wide variety of metabolic, circulatory, toxic, microbial, and neoplastic insults, and is, therefore, one of the most frequently injured
10 organs in the body. However, because the function of liver is very complex, synthetically recreating its function is practically impossible. One way of restoring liver function is by whole organ transplantation. Although transplantation of whole liver is often times successful it has plateaued at about 2200 transplants per year because of donor scarcity. Therefore, alternative treatments concentrate on manipulating the smallest functional unit of
15 the liver, the individual hepatocyte.

In yet another embodiment, the subject progenitor cells, or their progeny, can be used in the treatment various hepatic disorders. Vulnerable to a wide variety of metabolic, circulatory, toxic, microbial, and neoplastic insults, the liver is one of the most frequently injured organs in the body. In some instances, the disease is primarily localized in liver
20 cells. For example, primary liver diseases include hereditary disorders such as Gilbert's Syndrome, Crigler-Najjar Syndrome (either Type I or Type II), Dubin Johnson Syndrome, familial hypercholesterolemia (FH), ornithine transcarbamoylase (OTC) deficiency, hereditary emphysema and hemophilia; viral infections such as hepatitis A, B, and non-A, non-B hepatitis; and hepatic malignancies such as hepatocellular carcinoma. Robbins, S.L.
25 et al. (1984) *Pathologic Basis of Disease* (W.B. Saunders Company, Philadelphia) pp. 884-942. More often, the hepatic involvement is secondary, often to some of the most common diseases of man, such as cardiac decompensation, disseminated cancer, alcoholism, and extrahepatic infections. Robbins, S.L. et al. (1984) *Pathologic Basis of Disease* (W.B. Saunders Company, Philadelphia) pp. 884-942.

Whole liver transplantation, which is the current therapy for a variety of liver diseases, has been employed to successfully reconstitute LDL receptors in individuals with FH, thereby lowering serum cholesterol to normal levels. Whole liver transplantation, however, is limited by the scarcity of suitable donor organs. Li, Q. et al. (1993) *Human Gene Therapy* 4:403-409; Kay, M.A. et al. (1992) *Proc. Natl. Acad. Sci.* 89:89-93. In addition to
35 the difficulty in obtaining donor organs, the expense of liver transplantation, estimated at approximately \$200,000 to \$300,000 per procedure, prohibits its widespread application. Another unsolved problem is graft rejection. Foreign livers and liver cells are poorly tolerated by the recipient and are rapidly destroyed by the immune system in the absence of

immunosuppressive drugs. Li, Q. et al. (1993) *Human Gene Therapy* 4:403-409; Bumgardner, G.L. et al. (1992) *Transplantation* 53:857-862. While immunosuppressive drugs may be used to prevent rejection, they also block desirable immune responses such as those against bacterial and viral infections, thereby placing the recipient at risk of infection.

5 The hepatic progenitor cells of the invention can be used for treatment of many liver disorders because they have the ability to differentiate into cells of hepatic lineage, e.g., hepatocytes. The progenitor cells of the invention can be cultured *in vitro* under conditions which can further induce these cells to differentiate into mature hepatocytes, or they can
10 undergo differentiation *in vivo* once introduced into a subject. Many methods for encapsulating cells are known in the art, as has been described above for pancreatic cells. The hepatic progenitor cells can also be introduced directly into a subject, as they have the potential to induce liver regeneration.

15 Yet another aspect of the present invention provides methods for screening various compounds for their ability to modulate growth, proliferation or differentiation of distinct progenitor cell populations from bile duct tissue. A micro-organ explant that closely mimics the properties of a given set of tissue *in vivo* would have utility in screening assays in which compounds could be tested for their ability to modulate one of growth, proliferation or differentiation of progenitor cells in such tissue. Requirements of a
20 reproducible model for screening might include consistency in the micro-architecture, e.g. epithelial-mesenchymal interactions, and nutritional environment *in vitro*, as well as prolonged viability and proliferation of cultures beyond 24 hours to observe threshold effects of compounds being screened. This level of consistency cannot be achieved in the presence of undefined media supplements such as sera or tissue extracts that vary between
25 batches and cannot be adequately controlled. The dependence of a model on external growth supplements such as growth factors is also undesirable as growth factors or hormones may be included among the compounds to be tested.

30 In an illustrative embodiment, the ductal explants, which maintain their microarchitecture in culture, e.g., they preserve the normal epithelial-mesenchymal architecture that is present *in vivo*, can be used to screen various compounds or natural products. Such explants can be maintained in minimal culture media for extended periods of time (e.g., for 21 days or longer) and can be contacted with any compound, e.g., small molecule or natural product, e.g., growth factor, to determine the effect of such compound on one of cellular growth, proliferation or differentiation of progenitor cells in the explant. Detection and quantification of growth, proliferation or differentiation of these cells in
35 response to a given compound provides a means for determining the compound's efficacy at inducing one of the growth, proliferation or differentiation in a given ductal explant. Methods of measuring cell proliferation are well known in the art and most commonly include determining DNA synthesis characteristic of cell replication. There are numerous

methods in the art for measuring DNA synthesis, any of which may be used according to the invention. In an embodiment of the invention, DNA synthesis has been determined using a radioactive label (^3H -thymidine) or labeled nucleotide analogues (BrdU) for detection by immunofluorescence. The efficacy of the compound can be assessed by generating dose response curves from data obtained using various concentrations of the compound. A control assay can also be performed to provide a baseline for comparison. Identification of the progenitor cell population(s) amplified in response to a given test agent can be carried out according to such phenotyping as described above.

Exemplification

The invention now being generally described, it will be more readily understood by reference to the following examples which are included merely for purposes of illustration of certain aspects and embodiments of the present invention, and are not intended to limit the invention.

The common bile duct (CBD) is a structure whose developmental origins is poorly understood. Its primary function is the delivery of bile acids from the liver to the duodenum to aid in the emulsification of food in the digestive process. 80% of its length traverses through the pancreas and is connected into the pancreatic ductal system through a series of anastomoses. Through these anastomoses and into the CBD flow the digestive enzymes secreted by the exocrine pancreas. The relation of the CBD to the liver and pancreas, and its morphology, are therefore essential to both liver and pancreas normal function.

Since it contributes to both pancreatic and hepatic function, the question arises as to whether the CBD arises during development as primarily a hepatic structure, a pancreatic structure, or both. Little has been done to resolve this issue in the past due in part to a lack of early markers as well as interest. However, there are a number of reports in the literature citing circumstantial evidence that the adult animal retains within the gut system stem cells for both liver and pancreas.

Because the CBD serves a dual hepato-pancreatic function, it might be possible that it also possesses a dual identity, and might then retain within its structure stem cells for both the pancreas and liver. To test this hypothesis directly we isolated the CBD and its attendant main ducts, and cultured them in vitro as intact duct segments. The goal of this work was to study the duct as an intact physiological unit and to determine whether resident stem cells could be activated to give rise to either liver and/or pancreatic derivatives. This unique culture system allows us the ability to study the interaction of the mesenchyme with the epithelium, rather than the isolated culture systems of others, and is hence more representative of the in vivo situation.

Using our unique culture system we derived cultures of ductal fragments from the common bile duct of the adult rat in a combination of matrigel and IGF-1 in serum free

conditions induced the formation of liver specific cell types. The liver identity of the induced cells was determined by immunohistochemical detection of the expression of alphafetoprotein (AFP), albumin, and ATBF-1, a transcription factor shown to regulate AFP expression. In addition, the formation of red blood cell clusters was observed. The induction of albumin and AFP positive cell types was specific for the matrigel/IGF-1 combination; culture on plastic or collagen with IGF-1 failed to induce the appearance of these cell types. Likewise, the substitution of IGF-1 with either TGF α , EGF, IGF-II, PDGF, or FGF β , all failed to elicit liver formation, even in the presence of matrigel. However, in all of the matrigel conditions tried, we observed the formation of red blood cell clusters. Our results indicate that there exist both liver and hematopoietic stem cells resident in the CBD system, that red blood cell formation can be stimulated by a factor present in matrigel, and that the combination of matrigel with IGF-1 can induce the formation of liver specific cell types.

15 *Preparation of Ductal Explants*

6 week old female Spague/Dawley from Taconic were exsanguinated by CO₂. The common bile duct and associated pancreatic ducts was removed and placed in cold Dulbecco's Minimal Essential Medium (DMEM) supplemented with 2mM glutamine and penicillin/streptomycin (100units/ml). The duct was then cleaned of associated pancreatic tissue, liver tissue, fat, and blood vessels. The clean and intact duct was sectioned into approximately 250 μ m transverse slices, such that the original epithelial-mesenchymal microarchitecture was retained, and placed in medium on ice.

250 μ l of reduced growth factor Matrigel (Collaborative Research) was added to each well of a 24-well plate and incubated at 37°C for 30 minutes. The plates were then removed and allowed to stand at room temperature for 15 minutes. The CBD microorgan explants were placed onto the matrigel per well and stood at room temperature for 15 minutes to allow the ducts to adhere to the matrigel. 1 ml of DMEM (P/S/L-glut) with or without growth factor addition was added to the wells. Growth factors used in the studies described below (EGF, TGF α , PDGF, FGF β , FGF α , IGF-1, and IGF-II) were all obtained from PreproTech. Cultures were incubated at 37°C in a 5% CO₂ atmosphere. Cultures were fed once a week with fresh medium.

Immunohistochemistry

The cultures were fixed overnight at 4°C in 4% paraformaldehyde (PFA). The cultures were rinsed three times 10 minutes each with phosphate-buffered saline (PBS) prior to addition of 1% periodic acid to remove endogenous peroxidase activity. The cultures were incubated with 3% milk in PBS/0.1% Triton X-100 (PBST) for 30 minutes at room temperature. Primary antibody to alphafetoprotein and albumin (Accurate) were added at 1:250 for 1 hour at room temperature. ATBF-1 was used at 1:1000. The tissue was washed 3 times with PBST over a period of 2 hours. The secondary biotinylated antibody (Vector)

was added at 1:500 for 1 hour at room temperature. The tissue was then washed 3 times for 1 hour with PBST and incubated with horseradish peroxidase-conjugated to avidin (Vector) for 30 minutes at room temperature. Antibody-binding was detected by the addition of DAB/H₂O₂ (Gibco). After termination of the colorimetric reaction with H₂O, 80% glycerol in PBS was added to clear the cultures prior to photograph.

Growth Factors

Effectiveness of the growth factors in stimulating proliferation was judged by the incorporation of Bromodeoxyuridine (BrdU) into DNA by the responding cells. Antibodies to BrdU were used to visualize and characterize the short term responses (24-48 hr).

10 The long term response was judged by the ability of these populations of cells to be grown and expanded in cell culture as a result of specific growth factor addition.

Different growth factors (EGF, TGF- α , bFGF, aFGF, IGF-I, IGF-II, VEGF and HGF) were used. In addition to the results for EGF, TGF- α and bFGF described below, IGF-I, IGF-II, VEGF and HGF were demonstrated to cause expansion of certain subpopulations of the ductal explant.

1. Administration of EGF to the CBD Explant

EGF was administered in three different doses, 1 ng/ml, 10 ng/ml and 100 ng/ml to the CBD explant. Activation of proliferation as assessed by BrdU labeling occurred with administration of 10 ng/ml of growth factor EGF within a span of 24 hr (Figure 1 and 2). There was no difference observed between 10 and 100 ng/ml dose. Addition of EGF to the CBD tissue explant resulted in proliferation of distinct cells within the explant and resulted in clustering of these cells.

2. Administration of TGF α to the CBD Explant

TGF α was administered in the same doses as EGF. Activation of proliferation as assessed by BrdU labeling occurred with administration of 100 ng/ml of growth factor TGF- α within a span of 24 hr (Figure 1 and 2). Unlike EGF, administration of TGF- α to the CBD explant resulted in proliferation of cells throughout the explant.

3. Administration of FGF β to the CBD Explant

FGF β was administered in the same doses as the above described growth factors. Activation of proliferation as assessed by BrdU labeling occurred with administration of 10 ng/ml of growth factor FGF β within a span of 24 hr (Figure 1 and 2). Administration of 10 ng/ml of FGF β resulted in induction of distinct CBD structures to synchronously divide, implying organized regulation of proliferative potential and response. Figure 3 depicts *in vitro* growth and expansion of CBD explant, e.g., formation of outgrowths, e.g., blebs, in response to administration of FGF β .

Preliminary long term growth experiments indicate that there does exist a large proliferative potential within the CBD explant that can be maintained in culture for at least 21 days.

5 *Characterization of Expanded Progenitor Cell populations*

Antibodies that recognize transcription factors known to be expressed during pancreas and liver formation were used to characterize the population distribution in CBDs. STF-1, also known as IPF-1, has been shown to be critical in pancreas formation; Pax-6 has been shown to be critical in pancreatic endocrine cells; Islet-1 is expressed in early gut endoderm and in islet tissue; and HNF3 β is expressed in early gut endoderm and in liver precursor cells in the endoderm. ATBF-1, Lim1/2 were not detected in the CBD. Table 1 shows the average number of positive nuclei for each marker per 20X field, or approximately every 500 μ m duct length.

Table 1
Scoring of Marker Distribution in the CBD cryostat sections

Markers	# of nuclei	Average
STF-1	209	226
	170	
	300	
Pax-6	26	18
	10	
	18	
HNF3 β	55	50
	46	
	50	
Islet-1	33	38
	42	
	38	

HNF3 β is an early marker for endoderm formation and is one of the earliest markers expressed by the developing liver. Sections of CBD were stained for the early endoderm and liver marker HNF3 β . The common bile duct (CBD) contains cells that are immunopositive for HNF3 β . Its expression in the CBD is sporadic but specific to the epithelium, as expected. HNF3 β is expressed throughout the CBD and also in the main pancreatic ducts, although less frequently. Positive cells were often found in clusters of epithelial cells that were branching away from the main lumen of the CBD. HNF3 β -expressing cells were found uniformly throughout the CBD. Because of HNF3 β 's role as an early endoderm and liver marker in embryonic development, we hypothesized that these HNF3 β expressing cells might be liver stem cells resident in the adult hepatopancreatic duct system.

If the observed HNF3 β expressing cells are indeed liver precursors one might then use ductal cultures to attempt to *in vitro* activate the formation of liver-specific cell types and structures. Construction of such a culture system would then allow the study of the signals and interactions required to induce the liver development program. To this end we cultured
 5 duct fragments that had been transversely cut into approximate 250 μ m widths.

These were then cultured either on plastic, collagen, directly on feeder layers (STO, C3H10T1/2), or on matrigel (Collaborative Research, Inc., Bedford, Mass.). Matrigel was the only matrix permutation tried that gave liver formation. Furthermore, the formation of liver structures only occurred with the addition of IGF-1. Addition of FGF β , TGF α , EGF,
 10 IGF-II, PDGF-AA all failed to induce the formation of liver structures.

The addition of IGF-1 to duct cultures gave rise to three basic duct colonies; "Nonresponders" in which there was little observed growth or change in morphology (Figure 4A), "N-type" colonies which underwent dramatic changes in morphology to give rise to neuritelike processes which were in fact fibroblasts laying end on end (Figure 4B)
 15 and "L-type" colonies which took on a liver-like morphology, with epithelial blebs and the formation of red blood cell clusters (Figure 4C). This liver-like morphology resembles that of cultured embryonic liver (data not shown). The number and frequency of the formation of all three colony types is documented in the table of Figure 5.

Because of the morphological similarity between our L-type colonies and cultured embryonic liver, we decided to look for the expression of liver-specific markers in all three colony types. The markers used for this analysis were alphafetoprotein (AFP), albumin, and ATBF-1. All three have been shown to be specific for early liver. In all cases examined the only cultures which stained immunopositive for all three markers were the L-type colonies.
 20

For example, cultures were stained for the expression of the early liver marker AFP. AFP is expressed within 24 hours of liver formation in embryos, and is lost in adult hepatic tissue. Our data indicated extensive expression of AFP in IGF-1 treated duct fragments. We also saw loose cells that were highly positive for AFP. These tended to be large cells in diameter. We also observed the formation of duct-like structures that have arisen in the CBD explants. These duct-like structures were often surrounded by highly positive AFP
 25 staining cells. Some of the duct-like structures appear themselves to be AFP positive, probably due to the fact that AFP is a secreted factor.
 30

Cultures were also stained for the early liver marker ATBF-1. ATBF-1 has been shown to regulate expression of AFP during embryonic liver development. We observed staining of L-type colonies with the liver marker ATBF-1.

35

All of the above-cited references and publications are hereby incorporated by reference.

Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, many equivalents to the specific embodiments of the invention described herein. Such equivalents are intended to be encompassed by the following
5 claims.

Throughout this specification and the claims which follow, unless the context requires otherwise, the word "comprise", and variations such as "comprises" and "comprising", will be understood to imply the inclusion of a stated integer or step or group of integers or steps but not the exclusion of any other integer or step or group of integers or steps.

THE CLAIMS DEFINING THE INVENTION ARE AS FOLLOWS:

1. A substantially purified population of viable bile duct progenitor cells.
2. A cellular composition comprising, as the cellular component, a substantially purified population of viable non-hepatic bile duct progenitor cells, which progenitor cells are capable of proliferation in a culture medium.
3. The composition of claim 2, having fewer than 20% of lineage committed cells.
4. The composition of claim 2, which progenitor cells from a mammal.
5. The composition of claim 4, which mammal is a transgenic mammal.
6. The composition of claim 4, which mammal is a primate.
7. The composition of claim 6, which mammal is a human.
8. The composition of claim 4, which mammal is a miniature swine.
9. The composition of claim 2, which progenitor cells are capable of differentiation to tissue lineages of organs derived from the primitive gut.
10. The composition of claim 9, which progenitor cells are capable of differentiation to hepatic lineages.
11. The composition of claim 10, wherein the progenitor cells are characterized by expression of one or more of a hepatocyte nuclear factor (HNF); ATBF1; AFP; a LIM type homeobox gene; a forkhead transcription factor; a CCAAT-enhancer binding protein (C/EBP); OV-6; cytokeratin type 7; cytokeratin type 19; Span-1; OC.2; CC.3; or γ -glutamyl transpeptidase
12. The composition of claim 11, wherein the progenitor cells are characterized by expression of HNF3 β , alphafetoprotein (AFP), albumin, and ATBF-1.
13. The composition of claim 9, which progenitor cells are capable of differentiation to pancreatic lineages.

14. The composition of claim 13, wherein the progenitor cells are characterized by expression of one or more of STF-1; a PAX gene; PTF-1; hXBP-1; an HNF gene; villin; tyrosine hydroxylase; insulin; glucagon; or neuropeptide Y.
- 5 15. The composition of claim 14, wherein the progenitor cells are characterized by expression of STF-1 and PAX6.
16. The composition of claim 9, which progenitor cells are capable of differentiation to gallbladder lineages.
- 10 17. The composition of claim 9, which progenitor cells are capable of differentiation to intestinal lineages.
18. The composition of claim 2, which progenitor cells are capable of differentiation to hematopoietic lineages.
- 15 19. The composition of claim 17, wherein the progenitor cells are characterized by expression of one or more of c-kit, CD34, CD33, OV-6, CAM5.2 or cytokeratin type 7.
- 20 20. The composition of claim 2, which progenitor cells can be maintained in culture for at least about 7 days.
21. A cellular composition comprising, as a cellular population, at least 75% progenitor cells isolated from a bile duct and capable of self-regeneration in a culture medium.
- 25 22. A cellular composition consisting essentially of, as the cellular population, viable non-hepatic ductal progenitor cells capable of self-regeneration in a culture medium and differentiation to members of the hepatic, pancreatic and gallbladder lineages.
- 30 23. The composition of claim 22, which progenitor cells are isolated from a hepatic duct tissue, or are the progeny thereof.
24. The composition of claim 22, which progenitor cells are isolated from cystic duct tissue, or are the progeny thereof.
- 35 25. The composition of claim 22, which progenitor cells are isolated from pancreatic duct tissue, or are the progeny thereof.

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26. The composition of claim 22, which progenitor cells are isolated from common bile duct tissue, or are the progeny thereof.
27. The composition of claim 22, which progenitor cells are responsive to one or more growth factor selected from a group consisting of IGF, EGF, TGF, FGF, HGF and VEGF, or orthologous or paralogous factors thereof.
28. An isolated population of progenitor cells obtained from a non-hepatic bile duct the cells having the ability to differentiate into hepatocytes when maintained in culture.
29. The cells of claim 28, wherein the progenitor cells are characterized by expression of HNF3 β , alphafetoprotein (AFP), albumin and ATBF-1.
30. A pharmaceutical composition comprising the cellular composition of claim 2.
31. A pharmaceutical composition comprising the cellular composition of claim 21.
32. A pharmaceutical composition comprising the cellular composition of claim 22.
33. A pharmaceutical composition comprising the cellular composition of claim 28.
34. A method for isolating progenitor cells from a bile duct, comprising:
 - providing an explant culture of bile duct cells. which explant retains in culture the epithelial-mesenchymal microarchitecture of a mammalian bile duct wherein the dimensions of the explant provide the cells of the explant as maintainable in culture for at least twenty-four hours and the explant includes at least one progenitor cell which has the ability to proliferate;
 - contacting the explant with an agent which causes proliferation of progenitor cells in the explant; and
 - isolating, from the explant, progenitor cells that proliferate in response to the agent.

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35. The method of claim 34 wherein explant has a surface area to volume index of at least about 1.5 mm^{-1} .
36. The method of claim 34 wherein the agent is a growth factor.
37. The method of claim 36 wherein the growth factor is selected from a group consisting of IGF, EGF, TGF, FGF, HGF and VEGF, or orthologous or paralogous factors thereof.
38. The method of claim 34 wherein the progenitor cells are pancreatic progenitor cells.
39. The method of claim 34, wherein the progenitor cells are hepatic progenitor cells.
40. The method of claim 34, wherein the explant is cultured in medium deficient in biological extracts.
41. The method of claim 40, wherein the medium is deficient in serum.
42. The method of claim 34, wherein the bile duct is a common bile duct.
43. The method of claim 34 comprising the further step of passaging in culture the progenitor cells isolated from the explant for at least five passages.
44. The method of claim 34 comprising the further step of providing the progenitor cells isolated from the explant or the progeny thereof in a pharmaceutically acceptable carrier.
45. A method for treating a disorder characterized by insufficient liver function in a subject comprising introducing into the subject a pharmaceutical composition including hepatic progenitor cells and a pharmaceutically acceptable carrier.

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46. The method of claim 45 wherein the subject is a human.
47. The method of claim 45 wherein the disorder is selected from the group consisting of cirrhosis, hepatitis B, hepatitis C, sepsis or ELAD.
48. The method of claim 45, wherein the hepatic progenitor cells are induced to differentiate into hepatocytes subsequent to being introduced into the subject.
49. The method of claim 45 wherein the hepatic progenitors cells are induced to differentiate into hepatocytes in culture prior to introduction into the subject.
50. The method of claim 45, wherein the hepatic progenitors cells are characterized by expression of HNF3 β , alphafetoprotein (AFP), albumin, and ATBF-1.

DATED this 28th day of JUNE, 2000

ONTOGENY, INC

by **DAVIES COLLISON CAVE**
Patent Attorneys for the Applicants

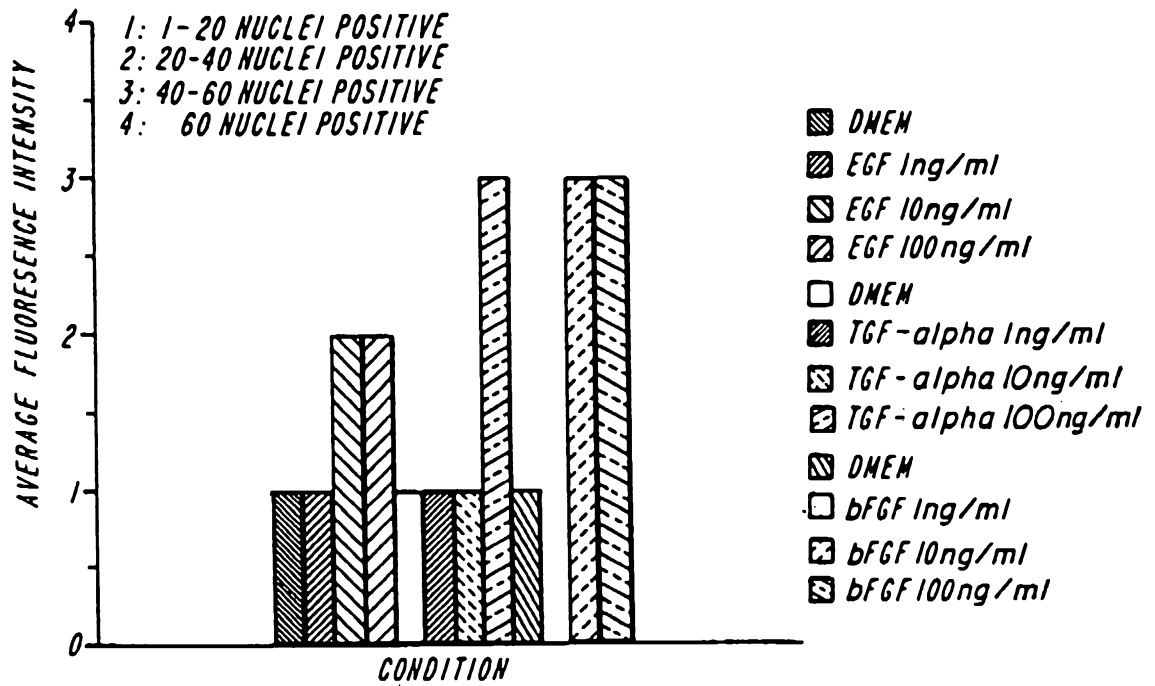


FIG. 1

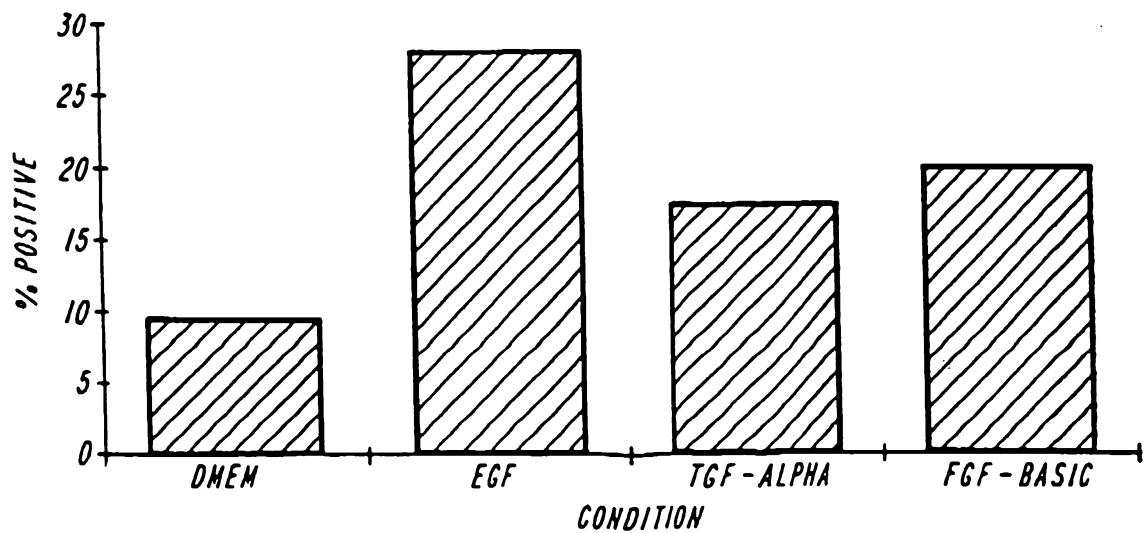


FIG. 2

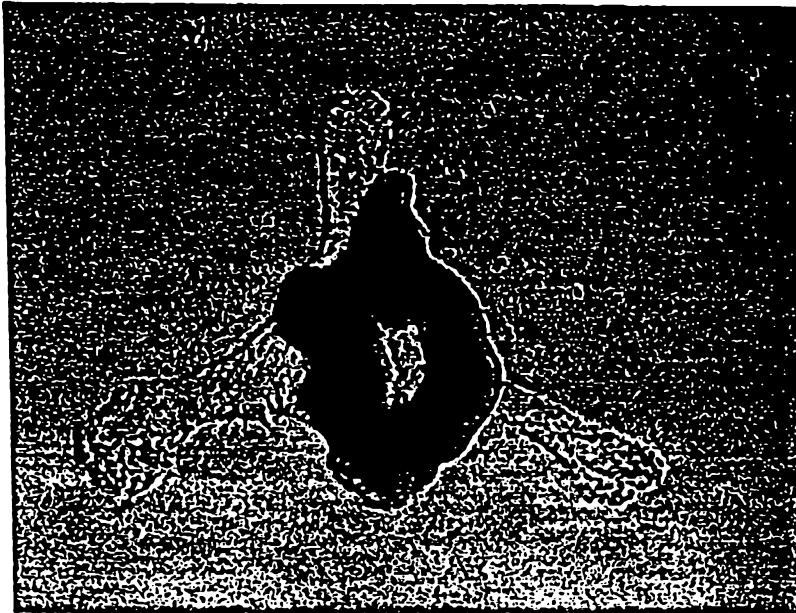


FIGURE 3

Figure 4A

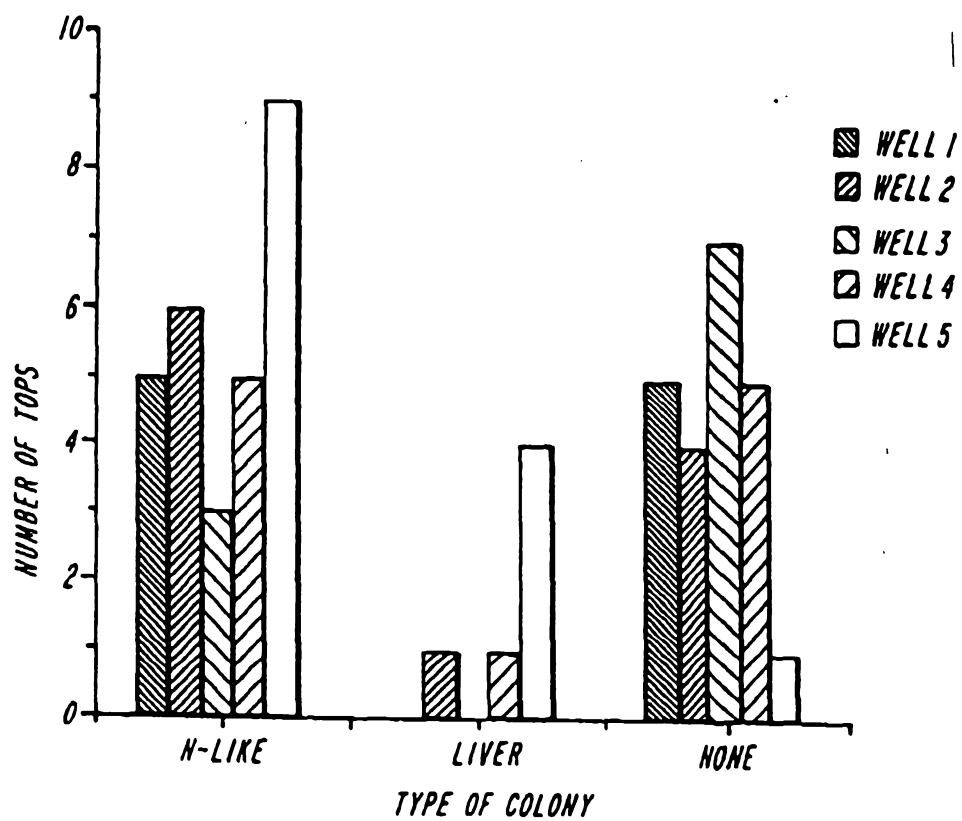


Figure 4B



Figure 4C



**FIG. 5**