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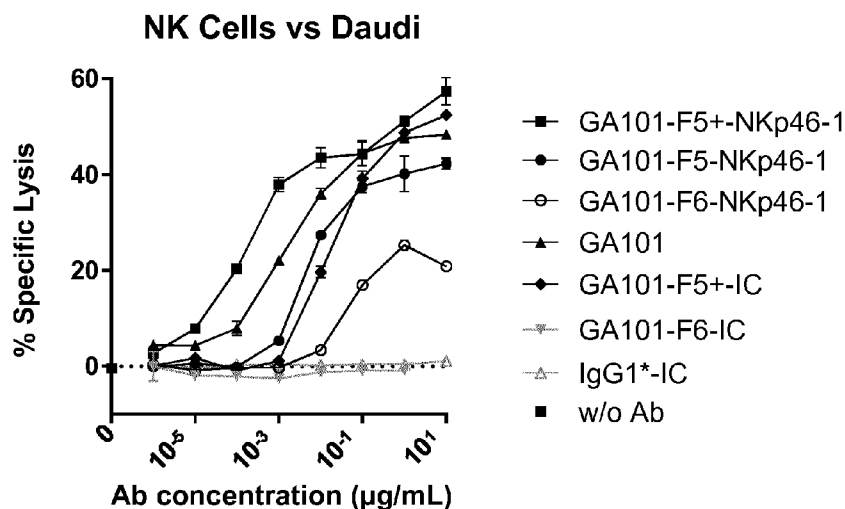
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[Continued on next page]

(54) Title: MULTISPECIFIC ANTIGEN BINDING PROTEINS

Figure 10



(57) Abstract: Multimeric multispecific proteins formed from dimerization between CH1 and CK domains and that bind two target antigens are provided. The proteins have advantages in production and in the treatment of disease, notably cancer or infectious disease.



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## MULTISPECIFIC ANTIGEN BINDING PROTEINS

### FIELD OF THE INVENTION

Multispecific proteins that bind and can be used to specifically redirect effector cells to lyse a target cell of interest are provided. The proteins formats have utility in the treatment of disease.

### CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims the benefit of U.S. Provisional Application No. 62/271,491 filed December 28, 2015 and PCT patent application No. PCT/EP2015/064070 filed 23 June 2015; both of which are incorporated herein by reference in their entirety; including any drawings and sequence listings.

### REFERENCE TO THE SEQUENCE LISTING

The present application is being filed along with a Sequence Listing in electronic format. The Sequence Listing is provided as a file entitled "BISP3 PCT\_ST25 txt", created June 21<sup>st</sup>, 2016, which is 301 KB in size. The information in the electronic format of the Sequence Listing is incorporated herein by reference in its entirety.

### BACKGROUND

Bispecific antibodies binding two different epitopes and offer opportunities for increasing specificity, broadening potency, and utilizing novel mechanisms of action that cannot be achieved with a traditional monoclonal antibody. A variety of formats for bispecific antibodies that bind to two targets simultaneously have been reported. Cross-linking two different receptors using a bispecific antibody to inhibit a signaling pathway has shown utility in a number of applications (see, e.g., Jackman, et al., (2010) J. Biol. Chem. 285:20850-20859). Bispecific antibodies have also been used to neutralize two different receptors. In other approaches, bispecific antibodies have been used to recruit immune effector cells, where T-cell activation is achieved in proximity to tumor cells by the bispecific antibody which binds receptors simultaneously on the two different cell types (see Baeuerle, P. A., et al, (2009) Cancer Res 69(12):4941-4). Most such approaches involve bispecific antibodies that link the CD3 complex on T cells to a tumor-associated antigen. The most well-studied bispecific antibody formats are "BiTe" antibodies and "DART" antibodies which do not comprise Fc domains. However these antibodies are known to be difficult to produce, require lengthy cell development, have low productions yields and/or cannot be produced (based on published literature) as a homogenous protein composition. Notably, in order to fully activate a T-cell, the T-cell and a cluster of BiTEs must interact on the surface of a target cell. Due to

the difficulties of finding antibody variable regions which are functional in the BiTE format, to date only a single immune cell receptor (CD3) has been targeted, in the CD19 x CD3 specific antibody blinatumamab. Bispecific antibodies developed to date also include those which link the CD3 complex on T cells to a tumor-associated antigen. In another example, a bispecific antibody having one arm which bound FcγRIII and another which bound to the HER2 receptor was developed for therapy of ovarian and breast tumors that overexpress the HER2 antigen.

However, despite the existence of a variety of formats for bispecific antibodies, there is therefore a need in the art for proteins with new and well-defined mechanisms of action that can bind two or more biological targets, and that have attractive properties for industrial development.

### SUMMARY OF THE INVENTION

The present invention arises from the discovery of a functional protein format that permits a wide range of antibody variable regions to be readily used, having advantages in manufacturing by being adapted to standard recombinant production techniques and without the need for development of product-specific folding or purification technique. While the new protein formats can be used to bind any desired antigens by incorporation the desired variable regions, advantageous examples are provided where multispecific proteins can bind to one (or optionally two or three) antigens of interest on target cells (e.g. cells to be eliminated or depleted), and one, two or three immune effector cell activating receptors on immune cells (e.g. lymphocytes, NK cells, T cells, etc.), optionally where one of the activating receptors is human CD16A. In some examples, the proteins possess one antigen binding domain (ABD) formed by immunoglobulin variable regions thereby binding to a target antigen, and a dimeric Fc domain that comprises N-linked glycosylation and binds the activating receptor CD16A. In some examples, the proteins possess two antigen binding domains (ABDs) each formed by immunoglobulin variable regions, thereby binding to two antigens (e.g. different antigens), and a dimeric Fc domain that comprises N-linked glycosylation and binds the activating receptor CD16A; when such protein includes an ABD that binds an effector cell activating receptor other than CD16A, the protein will therefore bind to two effector cell activating receptors (i.e., CD16A and the effector cell activating receptor other than CD16A), thereby providing advantageous immune enhancing activity. Other exemplary multispecific proteins can possess three antigen binding domains formed by immunoglobulin variable regions; such protein can for example have one or two ABDs that bind a different effector cell activating receptor (e.g. which may or may not include CD16A), and one or two ABDs that binds a cancer antigen. When such a protein with three



ABDs further include a dimeric Fc domain that comprises N-linked glycosylation and binds the activating receptor CD16A, the protein can for example have up to three ABDs that bind a cancer antigen, or one or two ABDs that bind a cancer antigen and one ABD that binds an effector cell activating receptor other than CD16A. Exemplary multispecific proteins can thus bind three antigens, wherein the antigens may be the same or different.

In one embodiment, provided is a multispecific protein that comprises: (i) a first antigen binding domain that binds to an activating receptor on an immune cell (e.g. effector cell), optionally an activating NK receptor, optionally wherein the receptor is a NK cell lectin-like receptor family member or an immunoglobulin superfamily member, optionally wherein the receptor is selected from a NKp46, NKp30, NKp44, CD137, CD3, CD8 and NKG2D polypeptide, (ii) a second antigen binding domain that binds to an antigen of interest expressed by a target cell, and a dimeric Fc domain that comprises N-linked glycosylation and binds human CD16A.

In another embodiment, provided is a multispecific protein that comprises: (i) a first antigen binding domain that binds to an activating receptor on an immune cell (e.g. effector cell), optionally an activating NK receptor, optionally wherein the receptor is a NK cell lectin-like receptor family member or an immunoglobulin superfamily member, optionally wherein the receptor is selected from a NKp46, NKp30, NKp44, CD137, CD3, CD8 and NKG2D polypeptide, (ii) a second antigen binding domain that binds to an antigen of interest expressed by a target cell, and (iii) a third antigen binding domain that binds to an activating receptor on an immune effector cell other than the activating receptor bound by the first antigen binding domain. Optionally, the protein further comprises a dimeric Fc domain that comprises N-linked glycosylation and binds human CD16A.

In one embodiment, the second antigen binding domain binds an antigen expressed on a cancer cell, an infected cell or a pro-inflammatory cell, e.g., a cancer antigen.

In one embodiment, the first and the third antigen binding domain each bind to a different activating NK receptor. In one embodiment, the first antigen binding domain that binds to an activating receptor on NK cell, optionally a natural cytotoxicity receptor (e.g. NKp46, NKp30, NKp44), and the third antigen binding domain binds to an activating receptor on a T cell (e.g. an effector T cell). Optionally, the third antigen binding domain binds human CD137, CD3, CD8 or NKG2D.

In one embodiment, the multispecific protein is designed to retain substantial FcγR (e.g. CD16) binding, e.g., compared to a conventional full-length human IgG1 antibody. Optionally the multispecific protein binds (e.g. via its Fc domain) to a human CD16, CD32A, CD32B and/or CD64 polypeptide. In some embodiments, the multispecific antibody is designed to have increased binding to a human CD16 polypeptide, e.g., compared to a

conventional full-length human IgG1 antibody, optionally wherein the Fc domain comprises one or more amino acid substitutions compared to a human IgG1 wild-type Fc domain.

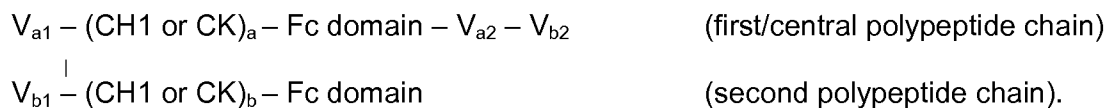
The proteins are made of different polypeptide chains that each comprise at least one heavy or light chain variable domain fused to a human CH1 or C $\kappa$  constant domain (a V-(CH1/C $\kappa$ ) unit), wherein the protein chains undergo CH1-C $\kappa$  dimerization and are bound to one another by non-covalent bonds and optionally further by disulfide bonds formed between respective CH1 and C $\kappa$  domain. Generally, two of the chains comprise Fc domains, such that a dimeric Fc domain is formed.

In one embodiment, provided is an isolated or purified heterodimeric or heterotrimeric protein that binds a first, second and optionally a third antigen, wherein the protein comprises two or three polypeptide chains each comprising a different V-(CH1/C $\kappa$ ) unit, optionally wherein two of chains further comprise an Fc domain fused to the C-terminus of the V-(CH1/C $\kappa$ ) unit, whereby the chains are bound to one another by non-covalent bonds and optionally further by disulfide bonds between CH1 and C $\kappa$  domains, optionally, whereby the chains are further bound by non-covalent bonds between respective variable regions, CH1 and C $\kappa$  domains, and optionally further, wherein two of the chains comprise Fc domain and are further bound by non-covalent bonds between CH3 domains of the Fc portion.

The variable and constant regions are selected and configured such that each chain will preferentially associate with its desired complementary partner chain. The resulting multimeric protein will therefore be simple to produce using conventional production methods using recombinant host cells. The choice of which VH, VL to associate with a CH1 and C $\kappa$  in a unit is based on affinity between the units to be paired so as to drive the formation of the desired multimer. The resulting multimer will be bound by non-covalent bonds between complementary VH and VL domains, by non-covalent bonds between complementary CH1 and C $\kappa$  domains, and optionally disulfide bonding between complementary CH1 and C $\kappa$  domains (and/or optionally further disulfide bonds between complementary hinge domains). VH-VL associations are stronger than VH-VH or VL-VL, consequently, as shown herein, one can place a VH or a VL next to either a CH1 or a C $\kappa$ , and the resulting V-C unit will partner preferably with its V-C counterpart. For example VH-C $\kappa$  will pair with VL-CH1 preferentially over VH-CH1. Additionally, by including an Fc domain, preferred chain pairing is further improved, as the two Fc-containing chains will be bound by non-covalent bonds between CH3 domains of the Fc domains. The different V-C combinations, optionally further combined with Fc pairing thereby provides tools to make heteromultimeric proteins.

In one example, a multispecific protein is provided that binds to three antigens, wherein one of the antigens is human CD16. In one embodiment, the protein comprises a

first and a second polypeptide chain each comprising a variable domain fused to a CH1 or C $\kappa$  domain (a V-(CH1/C $\kappa$ ) unit), in turn fused at its C-terminus to a human Fc domain, wherein the V-(CH1/C $\kappa$ ) unit of the first chain has undergone CH1-C $\kappa$  dimerization with the V-(CH1/C $\kappa$ ) unit of the second chain thereby forming a first antigen binding domain (ABD<sub>1</sub>) and a dimeric Fc domain, wherein one of the polypeptide chains further comprises an antigen binding domain that forms a second antigen binding domain (ABD<sub>2</sub>), and wherein the Fc domain binds to a human CD16 polypeptide. In one embodiment, the Fc domain comprises N-linked glycosylation at residue N297 (Kabat EU numbering). In one example, the protein has a domain arrangement:



In one example, a multispecific protein is provided that binds to three antigens, wherein one of the antigens is human CD16. In one embodiment, the protein comprises three polypeptide chains, each comprise a variable domain fused to a CH1 or C $\kappa$  domain (a V-(CH1/C $\kappa$ ) unit), wherein a first (central) chain comprises two V-(CH1/C $\kappa$ ) units and a human Fc domain interposed between the units, the second chain comprises one V-(CH1/C $\kappa$ ) unit and a human Fc domain, and the third chain comprises one V-(CH1/C $\kappa$ ) unit, wherein one of the V-(CH1/C $\kappa$ ) units of the central chain has undergone CH1-C $\kappa$  dimerization with the V-(CH1/C $\kappa$ ) unit of the second chain thereby forming a first antigen binding domain (ABD<sub>1</sub>) and a dimeric Fc domain, and wherein the other of the V-(CH1/C $\kappa$ ) units of the central chain has undergone CH1-C $\kappa$  dimerization with the V-(CH1/C $\kappa$ ) unit of the third chain thereby forming a second antigen binding domain (ABD<sub>2</sub>), and wherein the Fc domain binds to a human CD16 polypeptide. In one embodiment, the Fc domain comprises N-linked glycosylation at residue N297 (Kabat EU numbering). In one example, the protein has a domain arrangement:



In one embodiment, heterotrimer proteins are provided that have three antigen binding domains and that either lack binding to CD16 via an Fc domain, or that additionally comprise an Fc domain that binds to CD16. One example of such a protein is a trimer containing three polypeptide chains that each comprise a variable domain fused to a CH1 or C $\kappa$  domain (a V-(CH1/C $\kappa$ ) unit), wherein a first (central) chain comprises two V-(CH1/C $\kappa$ ) units and each of the second and third chains comprise one V-(CH1/C $\kappa$ ) unit, wherein one of

the V-(CH1/C $\kappa$ ) units of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the second chain thereby forming a first antigen binding domain (ABD<sub>1</sub>), wherein the other of the V-(CH1/C $\kappa$ ) units of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the third chain thereby forming a second antigen binding domain (ABD<sub>2</sub>), and wherein one of the polypeptide chains further comprises an antigen binding domain (e.g. a tandem variable domain, an scFv) that forms third antigen binding domain (ABD<sub>3</sub>). In one example, the protein has a domain arrangement:

V<sub>1</sub> – (CH1 or C $\kappa$ )<sub>c</sub> – Fc domain (second polypeptide)

V<sub>1</sub> – (CH1 or C $\kappa$ )<sub>a</sub> – Fc domain – V<sub>2</sub> – (CH1 or C $\kappa$ )<sub>b</sub> (first polypeptide)

V<sub>2</sub> – (CH1 or C $\kappa$ )<sub>d</sub> – V<sub>3</sub> – V<sub>3</sub> (third polypeptide).

When a protein having three antigen binding domains also comprises a dimeric Fc domain that binds to human CD16, the resulting protein will be capable of binding CD16 in addition to a first, second and third antigen.

In one embodiment, the central chain comprises an Fc domain (or portion) interposed between the two V-(CH1/C $\kappa$ ) units. In one embodiment, the second or third polypeptide comprises an Fc domain (or portion thereof), for example wherein the Fc domain is placed at the C-terminus of a V-(CH1/C $\kappa$ ) unit in the second or third chain, wherein the Fc domains (or portions) of the central chain and the Fc domain of the second or third chain associate within the heteromultimeric protein to form a dimeric Fc domain. In one embodiment, the dimeric Fc domain binds human FcRn and human CD16 polypeptide. In one embodiment, the Fc domain comprises N-linked glycosylation at residue N297 (Kabat EU numbering).

When a V-(CH1/C $\kappa$ ) unit of one chain has undergone dimerization with a V-(CH1/C $\kappa$ ) unit of another chain, the units will be bound by non-covalent bonds and optionally further by disulfide bond(s) between respective CH1 and C $\kappa$  domains (and further non-covalent bonds, as discussed above). The variable (V) domains and CH1/C $\kappa$  will be selected are configured such that each complementary pair of V-(CH1/C $\kappa$ ) units collectively comprises one VH, one VL, one CH1 and one C $\kappa$  domain.

In one embodiment, provided is a hetero-multimeric multispecific protein comprising: a first antigen binding domain (ABD<sub>1</sub>) that specifically binds to a first antigen of interest, a second antigen binding domain (ABD<sub>2</sub>) that specifically binds a second antigen of interest, wherein the first and second antigen are the same, and a third antigen binding domain (ABD<sub>3</sub>) that specifically binds a third antigen of interest, and at least a portion of a human Fc domain, wherein the Fc domain is interposed between the ABD<sub>1</sub> and ABD<sub>2</sub>. In one example, the first antigen is an antigen expressed by a target cell to be eliminated, the second antigen

is an antigen expressed by a target cell to be eliminated (either the same or different from the first antigen, or a different epitope on the same protein as the first antigen), and the third antigen is an antigen expressed by an immune effector cell (e.g. an NK cell and/or a T cell). In one example, the first and second antigen are the same antigen (optionally the same or a different epitope on the same antigen), such that the multispecific protein binds the antigen expressed by a target cell to be eliminated in bivalent manner, and binds the antigen expressed by an immune effector cell in monovalent manner. Such a multispecific protein may permit advantageous targeting of an antigen expressed by target cell by triggering an activating receptor on an effector cell in monovalent manner, thereby preventing or reducing agonist activity at the receptor on effector cells in the absence of target cells. In one example, the first antigen is an antigen expressed by a target cell to be eliminated, the second and the third antigen are each a different activating receptor expressed at the surface by an immune effector cell (e.g. an NK cell and/or a T cell); optionally one of the antigens is human CD137 and the other of the antigens is a different activating immune effector cell receptor, for example human NKp46, NKp30, NKp44, NKG2D, CD3 or CD8. In one embodiment, when the second and the third antigen are each a different activating receptor the multispecific protein binds to each activating receptor in monovalent manner.

In one aspect of any embodiment herein, the multispecific protein binds to an activating receptor on an immune effector cell in monovalent manner. In one embodiment, the multispecific protein is capable of mediating agonist activity (e.g. triggering signaling) of the activating receptor(s) bound by multispecific protein (or ABD thereof) in an immune effector cell expressing the activating receptor in the presence of a target cell (e.g. a cell to be eliminated that expresses an antigen bound by the multispecific protein). Optionally, the multispecific protein is capable of mediating agonist activity of the activating receptor(s) in an immune effector cell expressing the activating receptor and a target cell, yet does not substantially induce or mediate agonist activity of the activating receptor(s) in an immune effector cell expressing the activating receptor in the absence of a target cell. Agonist activity can be assessed by any suitable method, e.g. stimulation of activating-receptor dependent target cell lysis by an immune effector cell, activation and/or cytotoxicity markers on an immune cell, assessment of signaling or signaling pathways by the activating receptor, etc.

The multimeric polypeptide is composed of 2 or 3 different polypeptide chains in which 1 or 2 chains dimerize with a central chain based on CH1-CK heterodimerization. The multimer may be composed of a central (first) polypeptide chain comprising two immunoglobulin variable domains that are part of separate antigen binding domains (e.g., of different antigen specificities), with an Fc domain interposed between the two immunoglobulin variable domains on the polypeptide chain, and a CH1 or CK constant

domain placed on the polypeptide chain adjacent to one of, or each of, the variable domain. A second additional polypeptide chain will then be configured which will comprise a first immunoglobulin variable domain and a CH1 or CK constant region selected so as to permit CH1-CK heterodimerization with the central polypeptide chain; the immunoglobulin variable domain will be selected so as to complement the variable domain of the central chain that is adjacent to the CH1 or CK domain, whereby the complementary variable domains form an antigen binding domain for a first antigen of interest. The antigen binding domain for the second and third antigens of interest can then be formed according to several configurations. In one configuration, the central polypeptide chain comprises five immunoglobulin variable domains, wherein one variable domain is part of (together with the variable domain in the second polypeptide) the antigen binding domain for a first antigen of interest, the second and third variable domains are configured as tandem variable domains forming the antigen binding domain for the second antigen of interest (e.g. a heavy chain variable domain (VH) and a light chain (kappa) variable domain (VK), for example forming an scFv unit), and the fourth and fifth variable domains are configured as tandem variable domains forming the antigen binding domain for the second antigen of interest.

In a second configuration, the second polypeptide chain comprises (in addition to the first immunoglobulin variable domain and a CH1 or CK constant region) a second and third variable domain configured as a tandem variable domain forming the antigen binding domain for the third antigen, and the central polypeptide chain comprises three immunoglobulin variable domains, wherein one variable domain is part of (together with the variable domain in the second polypeptide) the antigen binding domain for a first antigen of interest, the second and third variable domains are configured as tandem variable domains forming the antigen binding domain for the second antigen of interest.

In a third configuration, the central polypeptide chain comprises two immunoglobulin variable domains each placed adjacent to a CH1 or CK constant domain, wherein a first of the variable domains is part of (together with the variable domain in the second polypeptide) the antigen binding domain for a first antigen of interest. A third polypeptide chain will then comprise (a) a first immunoglobulin variable domain adjacent to a CH1 or CK constant region selected so as to permit CH1-CK heterodimerization with the central polypeptide chain, whereby the second variable domain of the central chain and the first variable region of the third polypeptide form an antigen binding domain for the second antigen, and (b) a second and third variable domain configured as a tandem variable domain forming the antigen binding domain for the third antigen. In this configuration the central chain will comprise two V-(CH1/C $\kappa$ ) units with an interposed Fc domain: a first of the two V-(CH1/C $\kappa$ )

units will form a CH1-CK heterodimer with a V-(CH1/C $\kappa$ ) unit of the second chain, and the second of the two V-(CH1/C $\kappa$ ) unit will form a heterodimer with a V-(CH1/C $\kappa$ ) unit of the third chain. The immunoglobulin variable domain of the V-(CH1/C $\kappa$ ) unit of the third chain will be selected so as to complement the unpaired variable domain of the central chain, whereby the complementary variable domains form an antigen binding domain for a second antigen of interest.

Provided in one aspect are multimeric proteins that bind specifically to three antigens of interest (where the antigens may be the same or different), comprising a central (first) polypeptide chain comprising at least two variable domains that are part of different antigen binding domains, a CH1 or C $\kappa$  constant region fused to the C-terminus of one of the variable domains (thereby forming a V-(CH1/C $\kappa$ ) unit), and an Fc domain interposed between the two variable domains; and a second and/or third polypeptide chain that each comprise at least one V-(CH1/C $\kappa$ ) unit, wherein the variable domain and CH1 or C $\kappa$  constant region of the V-(CH1/C $\kappa$ ) unit of the second polypeptide chain (and, if present, third polypeptides) are complementary to the V and CH1 or C $\kappa$  constant region of the first polypeptide chain (but not to the V and CH1 or C $\kappa$  of the other of the second or third chain) such that the second (and, if present third polypeptide) chains preferentially form a CH1- C $\kappa$  heterodimer with the central chain, thereby forming a heterodimer (or heterotrimer). The CH1-C $\kappa$  heterodimers (or heterotrimers) will be characterized by non-covalent bonds and optionally further by disulfide bond(s) formed between respective CH1 and C $\kappa$  domains). When the second polypeptide comprises an Fc domain (and where the CH1/C $\kappa$  - Fc domain comprise hinge domains), the protein can optionally further be characterized by a disulfide bond formed between hinge domains.

In one advantageous format, provided are trimeric proteins that bind specifically to three antigens of interest (where the antigens may be the same or different) via three antigen binding domains (ABDs), comprising:

(i) a central (first) polypeptide chain comprising (e.g. from N- to C- terminus), a first V-(CH1/C $\kappa$ ) unit wherein the V domain forms part of a first ABD, an Fc domain or portion thereof, and a second V-(CH1/C $\kappa$ ) unit wherein the V domain forms part of a second ABD;

(ii) a second polypeptide chain comprising (e.g. from N- to C- terminus): a V-CH1/C $\kappa$  unit (and optionally an Fc domain or portion thereof), wherein the variable domain and CH1 or C $\kappa$  constant region are complementary to the CH1 or C $\kappa$  constant region the first (but not second) V-(CH1/C $\kappa$ ) unit of the central polypeptide chain (e.g., such that second chain

undergoes CH1/C $\kappa$  dimerization with the central chain and the V domain of the second chain forms the first ABD together with the V domain of the central chain); and

(iii) a third polypeptide chain comprising (e.g. from N- to C- terminus): a V-(CH1/C $\kappa$ ) unit and a tandem variable region (the tandem variable region forms a third ABD), and wherein the variable domain and CH1 or C $\kappa$  constant region are complementary to the variable domain and CH1 or C $\kappa$  constant region of the second (but not first) V-(CH1/C $\kappa$ ) unit of the central polypeptide chain (e.g., such that third chain undergoes CH1/C $\kappa$  dimerization with the central chain and the V domain of the V-(CH1/C $\kappa$ ) unit of the third chain forms the second ABD together with the V domain of the central chain). In one embodiment, the tandem variable region is an scFv (a V<sub>H</sub> fused to a V<sub>L</sub> via a peptide linker). Such trimeric protein can thus comprise two F(ab)-like structures and one tandem variable domain, providing advantageous binding properties.

In one embodiment, the multimeric, multispecific protein comprises a dimeric Fc domain that binds a human CD16A polypeptide.

In one example, the multispecific protein can specifically bind a first, second and a third antigen, wherein the first antigen is an antigen expressed by a target cell to be eliminated, the second antigen is an antigen expressed by a target cell to be eliminated, and the third antigen is an antigen expressed by an immune effector cell (e.g. an NK cell and/or a T cell), where the effector cells are directed to lyse the target cell, e.g. a cancer cell. In one embodiment, the first and second antigen are the same antigen, such that the multispecific protein binds the antigen expressed by a target cell to be eliminated in bivalent manner, and binds the antigen expressed by an immune effector cell in monovalent manner. Such a multispecific protein may permit advantageous targeting of an antigen expressed by target cell by triggering a selected activating receptor on an effector cell in monovalent manner, thereby preventing or reducing agonist activity at other receptor on effector cells (in the presence and/or absence of target cells).

In another example, the multispecific protein can specifically bind a first, second and a third antigen, wherein the first antigen is an antigen expressed by a target cell to be eliminated, the second antigen is an antigen expressed by an immune effector cell, and the third antigen is an antigen expressed by an immune effector cell (e.g. an NK cell and/or a T cell), where the effector cells are directed to the target cell, e.g. a cancer cell. The antigen on the effector cell can advantageously be an activating receptor. In one embodiment, the second and third antigen are different antigens, such that the multispecific protein binds the antigen expressed by a target cell to be eliminated in monovalent manner, and binds to two different antigens (e.g. activating receptors) expressed by immune effector cells in



monovalent manner. Such a multispecific protein may permit advantageous targeting of an antigen expressed by target cell by triggering multiple pathways on an effector cell and/or by causing redirection (to the target cell) of multiple populations of effector cells.

Furthermore, despite that the subject multispecific proteins are bound by CD16, unexpectedly they do not induce or increase down-modulation or internalization of the antigen of interest, even when targeting antigens of interest known to be susceptible to down-modulation or internalization when bound by conventional antibodies (such as full length human IgG1's). Based thereon, the subject multispecific proteins should be well suited for targeting antigens of interest expressed by target cells, e.g., tumor or infected cells, including antigens which are known to be capable of undergoing down-modulation or internalization when bound by conventional antibodies (e.g. antibodies with human IgG1 Fc domains that retain CD16 binding). This is a huge therapeutic benefit since it is known in the art that antigen internalization can substantially impede the ability of conventional human IgG1 antibodies to mediate ADCC against a target cell. Thus, in one embodiment, a multispecific protein (or an ABD thereof) binds an antigen expressed by target cell that is known to internalize upon binding to a conventional antibody (e.g. monoclonal monospecific human IgG1), wherein the multispecific protein causes less (or does not cause) induction or increase in internalization of the antigen compared to a conventional antibody.

In some embodiments, the multispecific antibody can be designed to bind to human CD16 and therefore can mediate target cell lysis via CD16, optionally in addition to other activating receptors on an effector cell.

In some embodiments (the proteins comprising three immunoglobulin ABDs), the multispecific antibody can be designed to lack binding to human CD16 and/or other FcγR, and it will not substantially activate effector cells via CD16, and the multispecific antibody will be selective for the particular effector cells of interest, as a function of the antigen bound by the multispecific antibody's hypervariable regions, and optionally avoid any unwanted FcγR-mediated cross-linking effect or toxicity (e.g. cytokine-mediated toxicity) and/or inhibitory-FcγR mediated inhibition of the effector cells targeted. The multispecific polypeptide is capable, for example, of directing target antigen-expressing effector cells to lyse a target cell expressing a target antigen, e.g. cancer antigen, viral antigen, etc. Where CD16 binding is not desired, the multimeric polypeptide can be designed to have a monomeric Fc domain or a dimeric Fc domain that does not bind CD16. For monomeric Fc domains, the Fc domain may comprise a CH3 domain having one or more amino acid mutations (e.g. substitutions) in the CH3 dimer interface to prevent CH3-CH3 dimerization. In another example of monomeric Fc domains, the Fc domain may comprise a tandem CH3 domain to prevent CH3-CH3 dimerization.

In one example, the first (central) polypeptide chain has the domain arrangement:

$V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK})$ ,

such that a hetero-multimeric polypeptide is formed having the domain arrangement:

$V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK})$  (first/central polypeptide chain)

5  $V_2 - (\text{CH1 or CK}) - V_3 - V_3$  (second polypeptide chain)

wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain. The  $V_1$  pair will form a first ABD, the  $V_2$  pair will form a second ABD, and the  $V_3$  will pair to form a third ABD. The Fc domain can be configured to as to avoid CH3 heterodimerization among central polypeptide chains, e.g., by including a tandem CH3 domain or by making amino acid modifications that decrease CH3-CH3 dimerization.

In another example, the first (central) polypeptide chain has the domain arrangement:

15  $V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK}) - V_3 - V_3$ , such that a hetero-multimeric polypeptide is formed having the domain arrangement:

$V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK}) - V_3 - V_3$  (first/central polypeptide chain)

20  $V_2 - (\text{CH1 or CK})$  (second polypeptide chain)

In another example, the first (central) polypeptide chain has the domain arrangement:

$V_1 - (\text{CH1 or CK})_a - \text{Fc domain} - V_2 - (\text{CH1 or CK})_b$ , such that a hetero-multimeric polypeptide is formed having the domain arrangement:

$V_1 - (\text{CH1 or CK})_c - \text{Fc domain}$  (second polypeptide)

25  $V_1 - (\text{CH1 or CK})_a - \text{Fc domain} - V_2 - (\text{CH1 or CK})_b$  (first polypeptide)

$V_2 - (\text{CH1 or CK})_d - V_3 - V_3$  (third polypeptide)

wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain. The  $V_1$  pair will form a first ABD, the  $V_2$  pair will form a second ABD, and the  $V_3$  will pair to form a third ABD.

In another example, the hetero-multimeric polypeptide formed has the domain arrangement:

35  $V_1 - (\text{CH1 or CK})_c$  (second polypeptide)

$V_1 - (\text{CH1 or CK})_a - \text{Fc domain} - V_2 - (\text{CH1 or CK})_b$  (first polypeptide)

$V_2 - (\text{CH1 or CK})_d - V_3 - V_3$  (third polypeptide).

In another example, the first (central) polypeptide chain has the domain arrangement:  $V_1 - (\text{CH1 or CK})_a - \text{Fc domain} - V_2 - (\text{CH1 or CK})_b - V_3 - V_3$ . The hetero-multimeric polypeptide formed can have the domain arrangement:

$V_1 - (\text{CH1 or CK})_c$  (second polypeptide)  
 $V_1 - (\text{CH1 or CK})_a - \text{Fc domain} - V_2 - (\text{CH1 or CK})_b - V_3 - V_3$  (first polypeptide)  
 $V_2 - (\text{CH1 or CK})_d$  (third polypeptide).

Optionally, in any embodiment, the second, and if present third, polypeptide chain of the multimeric protein can be characterized as being bound to the central/first chain by non-covalent bonds between complementary VH and VL domains, by non-covalent bonds between complementary CH1 and C $\kappa$  domains, and optionally disulfide bonding between complementary CH1 and C $\kappa$  domains (and/or optionally further disulfide bonds between complementary hinge domains, when present on both chains). Where the second or third chain is an Fc domain-containing chain, it can be characterized as being bound to the central/first chain non-covalent bonds between CH3 domains of the Fc domains.

Provided also is a purified or homogenous composition, wherein at least 90%, 95% or 99% of the proteins in the composition are a multimeric polypeptide of the disclosure, e.g. proteins comprised of the two or three polypeptide chains and having the domain structure indicated herein.

Optionally in any embodiment, each of the variable domains is a single immunoglobulin heavy or light chain variable domain. Optionally in any embodiment, one or more (of each of) the ABDs is a single non-immunoglobulin binding domain that binds an antigen, e.g. comprising a non-immunoglobulin scaffold.

Optionally in any embodiment, fusions or linkages on the same polypeptide chain between different domains (e.g., between two V domains placed in tandem, between V domains and CH1 or C $\kappa$  domains, between CH1 or C $\kappa$  domains and Fc domains, between Fc domains and V domains) may occur via intervening amino acid sequences, for example via a hinge region or linker peptide.

In another embodiment, particularly where agonist activity at a cell surface activating receptor is desired, multispecific proteins have a structure in which the freedom of motion (intrachain domain motion) or flexibility of one or more antigen binding domains (ABDs) is increased, e.g. compared to the ABDs of a conventional human IgG antibody. In one embodiment, multispecific proteins have a structure that permits the antigen binding site of the first antigen binding domain and the antigen binding site of the second antigen binding domain to be separated by a distance that enhances function, e.g., the ability of the

multispecific protein to induce signaling via a cell surface receptor and/or lysis of target cells, e.g., optionally a distance of less than 80 ångström (Å). Multispecific proteins wherein the ABDs possess greater flexibility and/or are separated by an optimized distance may enhance the formation of a lytic effector cell-target synapse, thereby potentiating activating receptor-mediated signaling.

In one embodiment, multispecific proteins having increased freedom of motion of the antigen binding domains (e.g. compared to the ABDs of a conventional human IgG antibody, e.g., a human IgG1 antibody). One example of such a protein is a monomeric or multimeric Fc domain-containing protein (e.g. a heterodimer or heterotrimer) in which an antigen binding domain (e.g., the ABD that binds an activating receptor on an immune cell or the ABD that bind the antigen of interest) is linked or fused to an Fc domain via a flexible linker. The linker can provide flexibility or freedom of motion of one or more ABDs by conferring the ability to bend thereby potentially decreasing the angle between the ABD and the Fc domain (or between the two ABDs) at the linker. Optionally, both antigen binding domains (and optionally more if additional ABDs are present in the multispecific protein) are linked or fused to the Fc domain via a linker, typically a flexible peptide linker. Optionally, other sequences or domains such as constant domains which optionally may be modified to alter (enhance or inhibit) one or more effector functions are placed between the Fc domain and an ABD, e.g. such that the ABD is fused to the Fc domain via a flexible linker and a constant region. Optionally, the protein with increased freedom of motion permits the protein to adopt a conformation in which the distance between the anti-activating receptor binding site and the target cell antigen of interest binding site is less than observed in proteins in which both binding domains were Fabs, or less than in full length antibodies.

An ABD can be connected to the Fc domain (or CH2 or CH3 domain thereof) via a flexible linker (optionally via intervening sequences such as constant region domains or portions thereof, e.g. CH1 or C $\kappa$ ). The linker can be a polypeptide linker, for example peptide linkers comprising a length of at least 5 residues, at least 10 residues, at least 15 residues, at least 20 residues, at least 25 residues, at least 30 residues or more. In other embodiments, the linkers comprises a length of between 2-4 residues, between 2-4 residues, between 2-6 residues, between 2-8 residues, between 2-10 residues, between 2-12 residues, between 2-14 residues, between 2-16 residues, between 2-18 residues, between 2-20 residues, between 2-30 residues, between 10-24 residues, between 10-26 residues, between 10-30 residues, or between 10-50 residues. Optionally a linker comprises an amino acid sequence derived from an antibody constant region, e.g., an N-terminal CH1 or hinge sequence. Optionally a linker comprises the amino acid sequence RTVA. Optionally a linker is a flexible linker predominantly or exclusively comprised of glycine and/or serine

residues, e.g., the amino acid sequence GEGTSTGS(G<sub>2</sub>S)<sub>2</sub>GGAD or the amino acid sequence (G<sub>4</sub>S)<sub>3</sub>.

Optionally in any embodiment, each antigen binding domain comprises the hypervariable regions, optionally the heavy and light chain CDRs, of an antibody. Optionally in any embodiment, a variable domain comprises framework residues from a human framework region, e.g., a variable domain comprises 1, 2 or 3 CDRs of human or non-human origin and framework residues of human origin.

Optionally in any embodiment, one or two of the antigens of interest is a cancer antigen, viral antigen or bacterial antigen, and one or two of the antigens of interest is a polypeptide expressed on the surface of an immune effector cell. Optionally in any embodiment, two of the antigens of interest are a cancer antigen (e.g. the same antigen or different antigens), and one of the antigens of interest is a polypeptide expressed on the surface of an immune effector cell. Optionally in any embodiment, one of the antigens of interest is a cancer antigen and two of the antigens of interest are a different activating receptor polypeptide expressed on the surface of an immune effector cell.

Optionally in any embodiment, all three of the antigens of interest are a cancer antigen, viral antigen or bacterial antigen, and the multimeric protein comprises a dimeric Fc domain capable of binding to human CD16. Optionally, all three of the antigens of interest are a different cancer antigen.

In one embodiment, provided is a protein that comprises (i) a first antigen binding domain that binds to an activating receptor on an immune effector cell (e.g. a NKp46, NKp30, NKp44, CD137, CD3, CD8, NKG2D or other polypeptide disclosed herein), (ii) a second antigen binding domain that binds to an antigen of interest expressed by a target cell and (iii) a third antigen binding domain that binds to an antigen of interest other than the activating receptor bound by the first antigen binding domain. For example, the third antigen binding domain may bind an antigen of interest expressed by a target cell, wherein the antigen of interest is the same or different from the antigen of interest bound by the second antigen binding domain. In one embodiment, the third antigen binding domain binds to the same antigen of interest as second antigen binding domain, optionally further wherein the third antigen binding domain binds to the same epitope or a different epitope on the antigen of interest as second antigen binding domain. In one embodiment, the second and third antigen binding domains bind a cancer antigen. In one embodiment, the second and third antigen binding domains bind a protein expressed (optionally over-expressed) at the surface of malignant immune cells, e.g. cells involved in a hematological malignancy, leukemia cells, lymphoma cells, a CD19 protein, a CD20 protein, etc. In one embodiment, the protein is used to treat a hematological malignancy, e.g., a leukemia or lymphoma cells. In another

embodiment, the second and/or third antigen binding domain binds a protein expressed (optionally over-expressed) on the surface of infected cells or by an infectious agent such as virally, bacterially or parasite infected cells. Optionally, the protein bind to the activating receptor in monovalent manner (the protein comprises a single antigen binding domain that binds the activating receptor). Optionally, the protein comprises an Fc domain that binds to human CD16A, optionally further wherein none of the antigen binding domains bind to CD16A. In one aspect, the protein has the features or domains arrangement of any embodiment disclosed herein.

In one embodiment, provided is a protein that comprises (i) a first antigen binding domain that binds to an activating receptor on an immune effector cell (e.g. a NKp46, NKp30, NKp44, CD137, CD3, CD8, NKG2D or other polypeptide disclosed herein), (ii) a second antigen binding domain that binds to an antigen of interest expressed by a target cell and (iii) a third antigen binding domain that binds to an activating receptor on an immune effector cell other than the activating receptor bound by the first antigen binding domain. In one embodiment, the first antigen binding domain binds human NKp46, the second antigen binding domain binds a cancer antigen, and the third antigen binding domain binds human CD137. In one embodiment, the second antigen binding domain binds a cancer antigen, optionally a protein expressed (optionally over-expressed) at the surface of malignant immune cells, e.g. cells involved in a hematological malignancy, leukemia cells, lymphoma cells, a CD19 protein, a CD20 protein, etc. In one embodiment, the protein is used to treat a hematological malignancy, e.g., a leukemia or lymphoma cells. In another embodiment, the second antigen binding domain binds a protein expressed (optionally over-expressed) on the surface of infected cells or by an infectious agent such as virally, bacterially or parasite infected cells. Optionally, the protein binds to each of the activating receptors in monovalent manner (the protein comprises a single antigen binding domain that binds to an activating receptor). Optionally, the protein comprises an Fc domain that binds to human CD16A, optionally further wherein none of the antigen binding domains bind to CD16A. In one aspect, the protein has the features or domains arrangement of any embodiment disclosed herein.

In one aspect of any of the embodiments herein, the multimeric protein has a great binding affinity (monovalent) for a cancer antigen (or a viral or bacterial antigen) than for an antigen expressed by an immune effector cell. Such antibodies will provide for advantageous pharmacological properties. In one aspect of any of the embodiments of the invention, the polypeptide has a  $K_d$  for binding (monovalent) to an antigen expressed by immune effector cell of less than  $10^{-7}$  M, preferably less than  $10^{-8}$  M, or preferably less than  $10^{-9}$  M for binding to an polypeptide expressed by an immune effector cell; optionally the polypeptide has a  $K_d$

for binding (monovalent) to a cancer, viral or bacterial antigen that is less than (i.e. has better binding affinity than) the  $K_d$  for binding (monovalent) to the antigen expressed by immune effector cell.

In one embodiment of any of the polypeptides herein, the multispecific protein is capable of directing effector cells (e.g. a T cell, an NK cell) expressing one of the antigens of interest to lyse a target cell expressing the other of the antigens of interest (e.g. a cancer cell, a virally infected cell, a bacterial cell, a pro-inflammatory cell, etc.).

In one embodiment of any of the polypeptides herein, the multispecific protein comprises a dimeric Fc domain capable of binding to human CD16, and the protein is capable of directing effector cells (e.g. a T cell, an NK cell) that express human CD16 to lyse a target cell expressing one or more of the antigens of interest (e.g. a cancer cell). In one embodiment, the multispecific protein causes lysis of the target cell at least in part by CD16-mediated antibody-dependent cell-mediated cytotoxicity ("ADCC"). In one embodiment, the multispecific protein causes lysis of the target cell by a combination of (a) enhancing or inducing signaling of an activating receptor on immune cells bound by an ABD of the multispecific protein, and (b) CD16-mediated antibody-dependent cell-mediated cytotoxicity ("ADCC").

In one aspect of any of the embodiments herein, provided is a recombinant nucleic acid encoding a first polypeptide chain, and/or a second polypeptide chain and/or a third polypeptide chain of any of the proteins of the disclosure. In one aspect of any of the embodiments herein, provided is a recombinant host cell comprising a nucleic acid encoding a first polypeptide chain, and/or a second polypeptide chain and/or a third polypeptide chain of any of the proteins of the disclosure, optionally wherein the host cell produces a protein of the disclosure with a yield (final productivity, following purification) of at least 1, 2, 3 or 4 mg/L. Also provided is a kit or set of nucleic acids comprising a recombinant nucleic acid encoding a first polypeptide chain of the disclosure, a recombinant nucleic acid encoding a second polypeptide chain of the disclosure, and, optionally, a recombinant nucleic acid encoding a third polypeptide chain of the disclosure. Also provided are methods of making monomeric, heterodimeric and heterotrimeric proteins of the disclosure.

In one embodiment, the invention provides methods of making a heterodimeric protein (e.g. any heterodimeric protein described herein), comprising:

a) providing a first nucleic acid encoding a first polypeptide chain described herein;

b) providing a second nucleic acid encoding a second polypeptide chain described herein;

c) optionally, providing a third nucleic acid encoding a third polypeptide chain described herein; and

d) expressing said first and second (and optionally third) nucleic acids in a host cell to produce a protein comprising said first and second polypeptide (and optionally third) chains, respectively; and recovering a heterodimeric (or optionally a heterotrimeric) protein. Optionally, the heterodimeric (or heterotrimeric) protein produced represents at least 20%, 25% or 30% of the total multispecific proteins obtained prior to purification. Optionally step (d) comprises loading the protein produced onto an affinity purification support, optionally an affinity exchange column, optionally a Protein-A support or column, and collecting the heterodimeric protein; and/or loading the protein produced (e.g., the protein collected following loading onto an affinity exchange or Protein A column) onto an ion exchange column; and collecting the heterodimeric fraction.

By virtue of their ability to be produced in standard cell lines and standardized methods with high yields, unlike BITE, DART and other multi-specific formats, the proteins of the disclosure also provide a convenient tool for screening for the most effective variable regions to incorporate into a multispecific protein. In one aspect, the present disclosure provides a method for identifying or evaluating candidate variable regions for use in a heterodimeric protein, comprising the steps of:

a) providing a plurality of nucleic acid pairs, wherein each pair includes one nucleic acid encoding a heavy chain candidate variable region and one nucleic acid encoding a light chain candidate variable region, for each of a plurality of heavy and light chain variable region pairs (e.g., obtained from different antibodies binding the same or different antigen(s) of interest);

b) for each of the plurality nucleic acid pairs, making a heterodimeric or trimeric protein by:

(i) producing a first nucleic acid encoding providing a first nucleic acid encoding a first polypeptide chain described herein;

(ii) providing a second nucleic acid encoding a second polypeptide chain described herein;

(iii) optionally, providing a third nucleic acid encoding a third polypeptide chain described herein;

wherein the nucleic acids encoding the heavy and light chain variable regions are independently positioned on the first, second or third nucleic acid such they form an antigen binding domain for the antigen of interest;

and

c) expressing said nucleic acids encoding the first and second (and optionally third) polypeptide chains in a host cell to produce a protein comprising said first and second (and



optionally third) polypeptide chains, respectively; and recovering a heterodimeric (or heterotrimeric) protein; and

d) evaluating the plurality of heterodimeric (or heterotrimeric) proteins produced for a biological activity of interest, e.g., an activity disclosed herein.

5 In one aspect, the present disclosure provides a library of at least 5, 10, 20, 30, 50 hetero-multimeric proteins of the disclosure, wherein the proteins share domain arrangements but differ in the amino acid sequence of the variable domains of one, two or three of their antigen binding domains.

10 In one aspect, the present disclosure provides a library of at least 2, 3, 4, 5 or 10 hetero-multimeric proteins of the disclosure, wherein the proteins share the amino acid sequence of the variable domain of one, two or three of their antigen binding domains, but differ in domain arrangements.

In one aspect, provided is a pharmaceutical composition comprising a compound or composition described herein, and a pharmaceutically acceptable carrier.

15 In one aspect provided is the use of a polypeptide or composition of any one of the above claims as a medicament for the treatment of disease.

In one aspect provided is a method of treating a disease in a subject comprising administering to the subject a compound or composition described herein.

In one embodiment, the disease is a cancer or an infectious disease.

20 Any of the methods can further be characterized as comprising any step described in the application, including notably in the "Detailed Description of the Invention"). The invention further relates to a protein obtainable by any of present methods. The disclosure further relates to pharmaceutical or diagnostic formulations of the antibodies of the present invention. The disclosure further relates to methods of using antibodies in methods of  
25 treatment or diagnosis.

These and additional advantageous aspects and features of the invention may be further described elsewhere herein.

#### BRIEF DESCRIPTION OF THE FIGURES

30 **Figure 1** shows that Anti-CD19-F1-Anti-CD3 does not cause T/B cell aggregation in the presence of B221 (CD19) or JURKAT (CD3) cell lines when separate, but it does cause aggregation of cells when both B221 and JURKAT cells are co-incubated.

**Figures 2A to 2E** show different domain arrangements of bispecific proteins produced. **Figure 2F** shows different domain arrangements proteins with three  
35 immunoglobulin ABDs.

**Figures 3A** and **3B** respectively demonstrate that bispecific F1 and F2 format proteins having NKp46 binding region based on NKp46-1, NKp46-2, NKp46-3 or NKp46-4 are able to direct resting NK cells to their CD19-positive Daudi tumor target cells, while isotype control antibody did not lead to the elimination of the Daudi cells. Rituximab (RTX) served as the positive control of ADCC, where the maximal response obtained with RTX (at 10 µg/ml in this assay) was 21.6% specific lysis.

**Figure 4A** shows that bispecific antibodies having NKp46 and CD19 binding regions in an F2 format protein do not activate resting NK cells in the absence of target cells; by contrast full length anti-NKp46 antibodies as well as positive control alemtuzumab did activate NK cells. **Figure 4B** shows that bispecific anti-NKp46 x anti-CD19 antibodies (including each of the NKp46-1, NKp46-2, NKp46-3 or NKp46-4 binding domains) activated resting NK cells in presence of Daudi target cells, while full-length anti-CD19 showed at best only very low activation of NK cells and neither full-length anti-NKp46 antibodies nor alemtuzumab elicited a substantial increase in activation beyond what was observed in the presence of NK cells alone. **Figure 4C** shows that in the presence of CD19-negative HUT78 cells, none of the bispecific anti-NKp46 x anti-CD19 antibodies (including each of the NKp46-1, NKp46-2, NKp46-3 or NKp46-4 variable regions) activated NK cells. However, the full-length anti-NKp46 antibodies and alemtuzumab resulted in detectable activation of NK cells, i.e., at a similar level observed in presence of NK cells alone. Isotype control antibody did not induce activation.

**Figures 5A** and **5B** shows that at low effector:target ratios of 1:1 each of the tested bispecific anti-NKp46 x anti-CD19 antibodies activated NK cells in the presence of Daudi cells, and that bispecific anti-NKp46 x anti-CD19 antibodies were far more potent (better elicited lysis of target cells) than a control anti-CD19 antibody as well as a full-length human IgG1 ADCC inducing antibody.

**Figures 6A** and **6B** show that each NKp46 x CD19 bispecific protein (Format F3, F5 and F6) induced specific lysis of Daudi (Figure 6A) or B221 (Figure 6B) cells by human KHYG-1 CD16-negative hNKp46-positive NK cell line, while rituximab and human IgG1 isotype control (IC) antibodies did not. **Figure 6C** shows that a NKp46 x KIR3DL2 bispecific protein (Format F6) induced specific lysis of HUT78 tumor cells via NKp46 binding (without CD16 binding) comparably to a conventional IgG1 antibody with the same anti-KIR3DL2 variable regions.

**Figure 7** shows a NKp46 x CD19 bispecific protein in F5 format whose Fc domain binds CD16 is far more potent in mediating Daudi target cell lysis than a full-length IgG1 anti-CD19 antibody or a F6 format bispecific protein. The figure also shows that a bispecific anti-CD19 in F6 format whose Fc domain does not bind CD16 was as potent in mediating NK cell

lysis of Daudi target cells as the full-length IgG1 anti-CD19 antibody, which is unexpected considering that the control IgG1 anti-CD19 antibody binds CD19 bivalently. At comparable levels of target cell lysis, CD19-F5-NKp46-3 was at least 1000 times more potent than the full-length anti-CD19 IgG1.

**Figure 8** shows the results of cytotoxicity assays using fresh NK cells (Daudi cell in the right hand panel and HUT78 cells in the left hand panel); the CD19-F6-NKp46-3 whose Fc domain does not bind CD16 due to a N297 substitution has as mode of action NKp46 triggering when NK cells encounter the target cell, while the CD19-F5-NKp46-3 bispecific protein demonstrated a far higher potency in mediating cytotoxicity toward Daudi cells. Neither the F5 nor F6 proteins mediated any NK cell cytotoxicity towards HUT78 cells.

**Figure 9** shows the results of flow cytometry staining of NK cell surface markers showed a strong upregulation of CD137 on the surface of NK cells by F5 proteins (Left-most panel: NK cells vs. Daudi; middle panel: NK cells vs. HUT78; right-most panel: NK cells alone). The full-length anti-CD19 IgG1 antibody that binds CD16 also showed CD137 upregulation, but to a far lesser extent than the CD19-F5-NKp46-3 protein. The CD19-F6-NKp46-3 which functions via NKp46 but not CD16 did not show any CD137 upregulation.

**Figure 10** shows the results of cytotoxicity assays which compared the ability of the GA101-F5<sup>+</sup>-NKp46-1 bispecific protein to a comparison antibody (GA101) containing the same variable regions to lyse Daudi cells. The results therein show that the GA101-F5<sup>+</sup>-NKp46-1 bispecific protein possesses far higher potency (approximately 10-fold increase in EC<sub>50</sub>) in mediating cytotoxicity toward Daudi cells than GA101.

## DETAILED DESCRIPTION OF THE INVENTION

### Definitions

As used in the specification, "a" or "an" may mean one or more. As used in the claim(s), when used in conjunction with the word "comprising", the words "a" or "an" may mean one or more than one.

Where "comprising" is used, this can optionally be replaced by "consisting essentially of", more optionally by "consisting of".

As used herein, the term "antigen binding domain" or "ABD" refers to a domain comprising a three-dimensional structure capable of immunospecifically binding to an epitope. Thus, in one embodiment, said domain can comprise a hypervariable region, optionally a VH and/or VL domain of an antibody chain, optionally at least a VH domain. In another embodiment, the binding domain may comprise at least one complementarity determining region (CDR) of an antibody chain. In another embodiment, the binding domain may comprise a polypeptide domain from a non-immunoglobulin scaffold.

The term "antibody" herein is used in the broadest sense and specifically includes full-length monoclonal antibodies, polyclonal antibodies, multispecific antibodies (e.g., bispecific antibodies), and antibody fragments and derivatives, so long as they exhibit the desired biological activity. Various techniques relevant to the production of antibodies are provided in, e.g., Harlow, et al., ANTIBODIES: A LABORATORY MANUAL, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., (1988). An "antibody fragment" comprises a portion of a full-length antibody, e.g. antigen-binding or variable regions thereof. Examples of antibody fragments include Fab, Fab', F(ab)<sub>2</sub>, F(ab')<sub>2</sub>, F(ab)<sub>3</sub>, Fv (typically the VL and VH domains of a single arm of an antibody), single-chain Fv (scFv), dsFv, Fd fragments (typically the VH and CH1 domain), and dAb (typically a VH domain) fragments; VH, VL, VhH, and V-NAR domains; minibodies, diabodies, triabodies, tetrabodies, and kappa bodies (see, e.g., Ill et al., Protein Eng 1997;10: 949-57); camel IgG; IgNAR; and multispecific antibody fragments formed from antibody fragments, and one or more isolated CDRs or a functional paratope, where isolated CDRs or antigen-binding residues or polypeptides can be associated or linked together so as to form a functional antibody fragment. Various types of antibody fragments have been described or reviewed in, e.g., Holliger and Hudson, Nat Biotechnol 2005; 23, 1126-1136; WO2005040219, and published U.S. Patent Applications 20050238646 and 20020161201.

The term "antibody derivative", as used herein, comprises a full-length antibody or a fragment of an antibody, e.g. comprising at least antigen-binding or variable regions thereof, wherein one or more of the amino acids are chemically modified, e.g., by alkylation, PEGylation, acylation, ester formation or amide formation or the like. This includes, but is not limited to, PEGylated antibodies, cysteine-PEGylated antibodies, and variants thereof.

The term "hypervariable region" when used herein refers to the amino acid residues of an antibody that are responsible for antigen binding. The hypervariable region generally comprises amino acid residues from a "complementarity-determining region" or "CDR" (e.g. residues 24-34 (L1), 50-56 (L2) and 89-97 (L3) in the light-chain variable domain and 31-35 (H1), 50-65 (H2) and 95-102 (H3) in the heavy-chain variable domain; Kabat et al. 1991) and/or those residues from a "hypervariable loop" (e.g. residues 26-32 (L1), 50-52 (L2) and 91-96 (L3) in the light-chain variable domain and 26-32 (H1), 53-55 (H2) and 96-101 (H3) in the heavy-chain variable domain; Chothia and Lesk, J. Mol. Biol 1987;196:901-917). Typically, the numbering of amino acid residues in this region is performed by the method described in Kabat et al., supra. Phrases such as "Kabat position", "variable domain residue numbering as in Kabat" and "according to Kabat" herein refer to this numbering system for heavy chain variable domains or light chain variable domains. Using the Kabat numbering system, the actual linear amino acid sequence of a peptide may contain fewer or additional

amino acids corresponding to a shortening of, or insertion into, a FR or CDR of the variable domain. For example, a heavy chain variable domain may include a single amino acid insert (residue 52a according to Kabat) after residue 52 of CDR H2 and inserted residues (e.g. residues 82a, 82b, and 82c, etc. according to Kabat) after heavy chain FR residue 82. The Kabat numbering of residues may be determined for a given antibody by alignment at regions of homology of the sequence of the antibody with a "standard" Kabat numbered sequence.

By "framework" or "FR" residues as used herein is meant the region of an antibody variable domain exclusive of those regions defined as CDRs. Each antibody variable domain framework can be further subdivided into the contiguous regions separated by the CDRs (FR1, FR2, FR3 and FR4).

By "constant region" as defined herein is meant an antibody-derived constant region that is encoded by one of the light or heavy chain immunoglobulin constant region genes. By "constant light chain" or "light chain constant region" as used herein is meant the region of an antibody encoded by the kappa ( $\kappa$ ) or lambda ( $\lambda$ ) light chains. The constant light chain typically comprises a single domain, and as defined herein refers to positions 108-214 of  $\kappa$ , or  $\lambda$ , wherein numbering is according to the EU index (Kabat et al., 1991, Sequences of Proteins of Immunological Interest, 5th Ed., United States Public Health Service, National Institutes of Health, Bethesda). By "constant heavy chain" or "heavy chain constant region" as used herein is meant the region of an antibody encoded by the mu, delta, gamma, alpha, or epsilon genes to define the antibody's isotype as IgM, IgD, IgG, IgA, or IgE, respectively. For full length IgG antibodies, the constant heavy chain, as defined herein, refers to the N-terminus of the CH1 domain to the C-terminus of the CH3 domain, thus comprising positions 118-447, wherein numbering is according to the EU index.

By "Fab" or "Fab region" as used herein is meant the polypeptide that comprises the VH, CH1, VL, and CL immunoglobulin domains. Fab may refer to this region in isolation, or this region in the context of a polypeptide, multispecific polypeptide or ABD, or any other embodiments as outlined herein.

By "single-chain Fv" or "scFv" as used herein are meant antibody fragments comprising the VH and VL domains of an antibody, wherein these domains are present in a single polypeptide chain. Generally, the Fv polypeptide further comprises a polypeptide linker between the VH and VL domains which enables the scFv to form the desired structure for antigen binding. Methods for producing scFvs are well known in the art. For a review of methods for producing scFvs see Pluckthun in The Pharmacology of Monoclonal Antibodies, vol. 113, Rosenberg and Moore eds. Springer-Verlag, New York, pp. 269-315 (1994).

By "Fv" or "Fv fragment" or "Fv region" as used herein is meant a polypeptide that comprises the VL and VH domains of a single antibody.

By "Fc" or "Fc region", as used herein is meant the polypeptide comprising the constant region of an antibody excluding the first constant region immunoglobulin domain. Thus Fc refers to the last two constant region immunoglobulin domains of IgA, IgD, and IgG, and the last three constant region immunoglobulin domains of IgE and IgM, and the flexible hinge N-terminal to these domains. For IgA and IgM, Fc may include the J chain. For IgG, Fc comprises immunoglobulin domains C $\gamma$ 2 (CH2) and C $\gamma$ 3 (CH3) and the hinge between C $\gamma$ 1 and C $\gamma$ 2. Although the boundaries of the Fc region may vary, the human IgG heavy chain Fc region is usually defined to comprise residues C226, P230 or A231 to its carboxyl-terminus, wherein the numbering is according to the EU index. Fc may refer to this region in isolation, or this region in the context of an Fc polypeptide, as described below. By "Fc polypeptide" or "Fc-derived polypeptide" as used herein is meant a polypeptide that comprises all or part of an Fc region. Fc polypeptides include but are not limited to antibodies, Fc fusions and Fc fragments. Also, Fc regions according to the invention include variants containing at least one modification that alters (enhances or diminishes) an Fc associated effector function. Also, Fc regions according to the invention include chimeric Fc regions comprising different portions or domains of different Fc regions, e.g., derived from antibodies of different isotype or species.

By "variable region" as used herein is meant the region of an antibody that comprises one or more Ig domains substantially encoded by any of the VL (including V $\kappa$  and V $\lambda$ ) and/or VH genes that make up the light chain (including  $\kappa$  and  $\lambda$ ) and heavy chain immunoglobulin genetic loci respectively. A light or heavy chain variable region (VL and VH) consists of a "framework" or "FR" region interrupted by three hypervariable regions referred to as "complementarity determining regions" or "CDRs". The extent of the framework region and CDRs have been precisely defined, for example as in Kabat (see "Sequences of Proteins of Immunological Interest," E. Kabat et al., U.S. Department of Health and Human Services, (1983)), and as in Chothia. The framework regions of an antibody, that is the combined framework regions of the constituent light and heavy chains, serves to position and align the CDRs, which are primarily responsible for binding to an antigen.

The term "specifically binds to" means that an antibody or polypeptide can bind preferably in a competitive binding assay to the binding partner, as assessed using either recombinant forms of the proteins, epitopes therein, or native proteins present on the surface of isolated target cells. Competitive binding assays and other methods for determining specific binding are further described below and are well known in the art.

The term "affinity", as used herein, means the strength of the binding of an antibody or polypeptide to an epitope. The affinity of an antibody is given by the dissociation constant  $K_D$ , defined as  $[Ab] \times [Ag] / [Ab-Ag]$ , where  $[Ab-Ag]$  is the molar concentration of the antibody-antigen complex,  $[Ab]$  is the molar concentration of the unbound antibody and  $[Ag]$  is the molar concentration of the unbound antigen. The affinity constant  $K_A$  is defined by  $1/K_D$ . Preferred methods for determining the affinity of mAbs can be found in Harlow, et al., Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1988), Coligan et al., eds., Current Protocols in Immunology, Greene Publishing Assoc. and Wiley Interscience, N.Y., (1992, 1993), and Muller, Meth. Enzymol. 92:589-601 (1983), which references are entirely incorporated herein by reference. One preferred and standard method well known in the art for determining the affinity of mAbs is the use of surface plasmon resonance (SPR) screening (such as by analysis with a BIAcore™ SPR analytical device).

By "amino acid modification" herein is meant an amino acid substitution, insertion, and/or deletion in a polypeptide sequence. An example of amino acid modification herein is a substitution. By "amino acid modification" herein is meant an amino acid substitution, insertion, and/or deletion in a polypeptide sequence. By "amino acid substitution" or "substitution" herein is meant the replacement of an amino acid at a given position in a protein sequence with another amino acid. For example, the substitution Y50W refers to a variant of a parent polypeptide, in which the tyrosine at position 50 is replaced with tryptophan. A "variant" of a polypeptide refers to a polypeptide having an amino acid sequence that is substantially identical to a reference polypeptide, typically a native or "parent" polypeptide. The polypeptide variant may possess one or more amino acid substitutions, deletions, and/or insertions at certain positions within the native amino acid sequence.

"Conservative" amino acid substitutions are those in which an amino acid residue is replaced with an amino acid residue having a side chain with similar physicochemical properties. Families of amino acid residues having similar side chains are known in the art, and include amino acids with basic side chains (e.g., lysine, arginine, histidine), acidic side chains (e.g., aspartic acid, glutamic acid), uncharged polar side chains (e.g., glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine, tryptophan), nonpolar side chains (e.g., alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine), beta-branched side chains (e.g., threonine, valine, isoleucine) and aromatic side chains (e.g., tyrosine, phenylalanine, tryptophan, histidine).

The term "identity" or "identical", when used in a relationship between the sequences of two or more polypeptides, refers to the degree of sequence relatedness between

polypeptides, as determined by the number of matches between strings of two or more amino acid residues. "Identity" measures the percent of identical matches between the smaller of two or more sequences with gap alignments (if any) addressed by a particular mathematical model or computer program (i.e., "algorithms"). Identity of related polypeptides can be readily calculated by known methods. Such methods include, but are not limited to, those described in Computational Molecular Biology, Lesk, A. M., ed., Oxford University Press, New York, 1988; Biocomputing: Informatics and Genome Projects, Smith, D. W., ed., Academic Press, New York, 1993; Computer Analysis of Sequence Data, Part 1, Griffin, A. M., and Griffin, H. G., eds., Humana Press, New Jersey, 1994; Sequence Analysis in Molecular Biology, von Heinje, G., Academic Press, 1987; Sequence Analysis Primer, Gribskov, M. and Devereux, J., eds., M. Stockton Press, New York, 1991; and Carillo et al., SIAM J. Applied Math. 48, 1073 (1988).

Preferred methods for determining identity are designed to give the largest match between the sequences tested. Methods of determining identity are described in publicly available computer programs. Preferred computer program methods for determining identity between two sequences include the GCG program package, including GAP (Devereux et al., Nucl. Acid. Res. 12, 387 (1984); Genetics Computer Group, University of Wisconsin, Madison, Wis.), BLASTP, BLASTN, and FASTA (Altschul et al., J. Mol. Biol. 215, 403-410 (1990)). The BLASTX program is publicly available from the National Center for Biotechnology Information (NCBI) and other sources (BLAST Manual, Altschul et al. NCB/NLM/NIH Bethesda, Md. 20894; Altschul et al., supra). The well-known Smith Waterman algorithm may also be used to determine identity.

An "isolated" molecule is a molecule that is the predominant species in the composition wherein it is found with respect to the class of molecules to which it belongs (i.e., it makes up at least about 50% of the type of molecule in the composition and typically will make up at least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, or more of the species of molecule, e.g., peptide, in the composition). Commonly, a composition of a polypeptide will exhibit 98%, 98%, or 99% homogeneity for polypeptides in the context of all present peptide species in the composition or at least with respect to substantially active peptide species in the context of proposed use.

In the context herein, "treatment" or "treating" refers to preventing, alleviating, managing, curing or reducing one or more symptoms or clinically relevant manifestations of a disease or disorder, unless contradicted by context. For example, "treatment" of a patient in whom no symptoms or clinically relevant manifestations of a disease or disorder have been identified is preventive or prophylactic therapy, whereas "treatment" of a patient in



whom symptoms or clinically relevant manifestations of a disease or disorder have been identified generally does not constitute preventive or prophylactic therapy.

The term "internalization", used interchangeably with "intracellular internalization", refers to the molecular, biochemical and cellular events associated with the process of translocating a molecule from the extracellular surface of a cell to the intracellular surface of a cell. The processes responsible for intracellular internalization of molecules are well-known and can involve, *inter alia*, the internalization of extracellular molecules (such as hormones, antibodies, and small organic molecules); membrane-associated molecules (such as cell-surface receptors); and complexes of membrane-associated molecules bound to extracellular molecules (for example, a ligand bound to a transmembrane receptor or an antibody bound to a membrane-associated molecule). Thus, "inducing and/or increasing internalization" refers to events wherein intracellular internalization is initiated and/or the rate and/or extent of intracellular internalization is increased.

As used herein, the phrase "NK cells" refers to a sub-population of lymphocytes that is involved in non-conventional immunity. NK cells can be identified by virtue of certain characteristics and biological properties, such as the expression of specific surface antigens including CD56 and/or NKp46 for human NK cells, the absence of the alpha/beta or gamma/delta TCR complex on the cell surface, the ability to bind to and kill cells that fail to express "self" MHC/HLA antigens by the activation of specific cytolytic machinery, the ability to kill tumor cells or other diseased cells that express a ligand for NK activating receptors, and the ability to release protein molecules called cytokines that stimulate or inhibit the immune response. Any of these characteristics and activities can be used to identify NK cells, using methods well known in the art. Any subpopulation of NK cells will also be encompassed by the term NK cells. Within the context herein "active" NK cells designate biologically active NK cells, including NK cells having the capacity of lysing target cells or enhancing the immune function of other cells. NK cells can be obtained by various techniques known in the art, such as isolation from blood samples, cytophoresis, tissue or cell collections, etc. Useful protocols for assays involving NK cells can be found in Natural Killer Cells Protocols (edited by Campbell KS and Colonna M). Humana Press. pp. 219-238 (2000).

As used herein, "T cells" refers to a sub-population of lymphocytes that mature in the thymus, and which display, among other molecules T cell receptors on their surface. T cells can be identified by virtue of certain characteristics and biological properties, such as the expression of specific surface antigens including the TCR, CD4 or CD8, the ability of certain T cells to kill tumor or infected cells, the ability of certain T cells to activate other cells of the immune system, and the ability to release protein molecules called cytokines that stimulate

or inhibit the immune response. Any of these characteristics and activities can be used to identify T cells, using methods well known in the art. Within the context herein, “active” or “activated” T cells designate biologically active T cells, more particularly T cells having the capacity of cytotoxicity or of stimulating an immune response by, e.g., secreting cytokines.

5 Active cells can be detected in any of a number of well-known methods, including functional assays and expression-based assays such as the expression of cytokines such as TNF-alpha.

As used herein, an agent that has “agonist” activity at a cell surface receptor (e.g. an activating receptor) is an agent that can cause or increase signalling by the receptor, e.g., an ability of the receptor to activate or transduce an intracellular signaling pathway. Changes in signaling activity can be measured, for example, by assays designed to measure changes in receptor signaling pathways, e.g. by monitoring phosphorylation of signal transduction components, assays to measure the association of certain signal transduction components with other proteins or intracellular structures, or in the biochemical activity of components

10 such as kinases, or assays designed to measure expression of reporter genes under control of receptor-sensitive promoters and enhancers, or indirectly by a downstream effect mediated by the receptor (e.g. activation of specific cytotoxic machinery in NK or T cells). Reporter genes can be naturally occurring genes (e.g. monitoring cytokine production) or they can be genes artificially introduced into a cell. Other genes can be placed under the control of such regulatory elements and thus serve to report the level of receptor signaling.

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### **Producing polypeptides**

The antigen binding domains (ABDs) described herein can be readily derived from any of a variety of immunoglobulin or non-immunoglobulin scaffolds, for example affibodies based on the Z-domain of staphylococcal protein A, engineered Kunitz domains,

25 monobodies or adnectins based on the 10th extracellular domain of human fibronectin III, anticalins derived from lipocalins, DARPins (designed ankyrin repeat domains, multimerized LDLR-A module, avimers or cysteine-rich knottin peptides. See, e.g., Gebauer and Skerra (2009) Current Opinion in Chemical Biology 13:245–255, the disclosure of which is incorporated herein by reference.

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Immunoglobulin ABDs can be obtained from variable domains derived from antibodies (from immunoglobulin chains), for example in the form of associated V<sub>L</sub> and V<sub>H</sub> domains found on two polypeptide chains, or a single chain antigen binding domain such as a scFv, a V<sub>H</sub> domain, a V<sub>L</sub> domain, a dAb, a V-NAR domain or a V<sub>H</sub>H domain. In certain advantageous protein formats disclosed herein that directly enable the use of a wide range

35 of variable regions from Fab or scFv without substantial further requirements for pairing

and/or folding, the an antigen binding domain (e.g., ABD<sub>1</sub> and ABD<sub>2</sub>) can also be readily derived from antibodies as a Fab or scFv.

Typically, antibodies are initially obtained by immunization of a non-human animal, e.g., a mouse, rat, guinea pig or rabbit, with an immunogen comprising a polypeptide, or a fragment or derivative thereof, typically an immunogenic fragment, for which it is desired to obtain antibodies (e.g. a human polypeptide). The step of immunizing a non-human mammal with an antigen may be carried out in any manner well known in the art for stimulating the production of antibodies in a mouse (see, for example, E. Harlow and D. Lane, *Antibodies: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY (1988), the entire disclosure of which is herein incorporated by reference). Human antibodies may also be produced by using, for immunization, transgenic animals that have been engineered to express a human antibody repertoire (Jakobovitz et al. *Nature* 362 (1993) 255), or by selection of antibody repertoires using phage display methods. For example, a XenoMouse (Abgenix, Fremont, CA) can be used for immunization. A XenoMouse is a murine host that has had its immunoglobulin genes replaced by functional human immunoglobulin genes. Thus, antibodies produced by this mouse or in hybridomas made from the B cells of this mouse, are already humanized. The XenoMouse is described in United States Patent No. 6,162,963, which is herein incorporated in its entirety by reference. Antibodies may also be produced by selection of combinatorial libraries of immunoglobulins, as disclosed for instance in (Ward et al. *Nature*, 341 (1989) p. 544, the entire disclosure of which is herein incorporated by reference). Phage display technology (McCafferty et al (1990) *Nature* 348:552-553) can be used to produce antibodies from immunoglobulin variable (V) domain gene repertoires from unimmunized donors. See, e.g., Griffith et al (1993) *EMBO J.* 12:725-734; US 5,565,332; US 5,573,905; US 5,567,610; and US 5,229,275). When combinatorial libraries comprise variable (V) domain gene repertoires of human origin, selection from combinatorial libraries will yield human antibodies.

Additionally, a wide range of antibodies are available in the scientific and patent literature, including DNA and/or amino acid sequences, or from commercial suppliers. Antibodies will typically be directed to a pre-determined antigen. Examples of antibodies include antibodies that recognize an antigen expressed by a target cell that is to be eliminated, for example a proliferating cell or a cell contributing to a disease pathology. Examples include antibodies that recognize tumor antigens, microbial (e.g. bacterial or parasite) antigens or viral antigens.

Variable domains and/or antigen binding domains can be selected based on the desired cellular target, and may include for example cancer antigens, bacterial or viral antigens, etc. As used herein, the term "bacterial antigen" includes, but is not limited to,

intact, attenuated or killed bacteria, any structural or functional bacterial protein or carbohydrate, or any peptide portion of a bacterial protein of sufficient length (typically about 8 amino acids or longer) to be antigenic. Examples include gram-positive bacterial antigens and gram-negative bacterial antigens. In some embodiments the bacterial antigen is derived from a bacterium selected from the group consisting of *Helicobacter* species, in particular *Helicobacter pylori*; *Borrelia* species, in particular *Borrelia burgdorferi*; *Legionella* species, in particular *Legionella pneumophila*; *Mycobacteria* species, in particular *M. tuberculosis*, *M. avium*, *M. intracellulare*, *M. kansasii*, *M. goodii*; *Staphylococcus* species, in particular *Staphylococcus aureus*; *Neisseria* species, in particular *N. gonorrhoeae*, *N. meningitidis*; *Listeria* species, in particular *Listeria monocytogenes*; *Streptococcus* species, in particular *S. pyogenes*, *S. agalactiae*; *S. faecalis*; *S. bovis*, *S. pneumoniae*; anaerobic *Streptococcus* species; pathogenic *Campylobacter* species; *Enterococcus* species; *Haemophilus* species, in particular *Haemophilus influenzae*; *Bacillus* species, in particular *Bacillus anthracis*; *Corynebacterium* species, in particular *Corynebacterium diphtheriae*; *Erysipelothrix* species, in particular *Erysipelothrix rhusiopathiae*; *Clostridium* species, in particular *C. perfringens*, *C. tetani*; *Enterobacter* species, in particular *Enterobacter aerogenes*, *Klebsiella* species, in particular *Klebsiella pneumoniae*, *Pasteurella* species, in particular *Pasteurella multocida*, *Bacteroides* species; *Fusobacterium* species, in particular *Fusobacterium nucleatum*; *Streptobacillus* species, in particular *Streptobacillus moniliformis*; *Treponema* species, in particular *Treponema pertense*; *Leptospira*; pathogenic *Escherichia* species; and *Actinomyces* species, in particular *Actinomyces israeli*.

As used herein, the term "viral antigen" includes, but is not limited to, intact, attenuated or killed whole virus, any structural or functional viral protein, or any peptide portion of a viral protein of sufficient length (typically about 8 amino acids or longer) to be antigenic. Sources of a viral antigen include, but are not limited to viruses from the families: Retroviridae (e.g., human immunodeficiency viruses, such as HIV-1 (also referred to as HTLV-III, LAV or HTLV-III/LAV, or HIV-III; and other isolates, such as HIV-LP; Picornaviridae (e.g., polio viruses, hepatitis A virus; enteroviruses, human Coxsackie viruses, rhinoviruses, echoviruses); Calciviridae (e.g., strains that cause gastroenteritis); Togaviridae (e.g., equine encephalitis viruses, rubella viruses); Flaviviridae (e.g., dengue viruses, encephalitis viruses, yellow fever viruses); Coronaviridae (e.g., coronaviruses); Rhabdoviridae (e.g., vesicular stomatitis viruses, rabies viruses); Filoviridae (e.g., Ebola viruses); Paramyxoviridae (e.g., parainfluenza viruses, mumps virus, measles virus, respiratory syncytial virus); Orthomyxoviridae (e.g., influenza viruses); Bunyaviridae (e.g., Hantaan viruses, bunya viruses, phleboviruses and Nairo viruses); Arenaviridae (hemorrhagic fever viruses); Reoviridae (e.g., reoviruses, orbiviruses and rotaviruses); Bornaviridae; Hepadnaviridae

(Hepatitis B virus); Parvoviridae (parvoviruses); Papovaviridae (papilloma viruses, polyoma viruses); Adenoviridae (most adenoviruses); Herpesviridae (herpes simplex virus (HSV) 1 and 2, varicella zoster virus, cytomegalovirus (CMV), herpes virus; Poxviridae (variola viruses, vaccinia viruses, pox viruses); and Iridoviridae (e.g., African swine fever virus); and unclassified viruses (e.g., the agent of delta hepatitis (thought to be a defective satellite of hepatitis B virus), Hepatitis C; Norwalk and related viruses, and astroviruses). Alternatively, a viral antigen may be produced recombinantly.

As used herein, the terms "cancer antigen" and "tumor antigen" are used interchangeably and refer to antigens that are differentially expressed by cancer cells or are expressed by non-tumoral cells (e.g. immune cells) having a pro-tumoral effect (e.g. an immunosuppressive effect), and can thereby be exploited in order to target cancer cells. Cancer antigens are antigens which can potentially stimulate apparently tumor-specific immune responses. Some of these antigens are encoded, although not necessarily expressed, or expressed at lower levels or less frequently, by normal cells. These antigens can be characterized as those which are normally silent (i.e., not expressed) in normal cells, those that are expressed only at certain stages of differentiation and those that are temporally expressed such as embryonic and fetal antigens. Other cancer antigens are encoded by mutant cellular genes, such as oncogenes (e.g., activated ras oncogene), suppressor genes (e.g., mutant p53), fusion proteins resulting from internal deletions or chromosomal translocations. Still other cancer antigens can be encoded by viral genes such as those carried on RNA and DNA tumor viruses. Still other cancer antigens can be expressed on immune cells capable of contributing to or mediating a pro-tumoral effect, e.g. cell that contributes to immune evasion, a monocyte or a macrophage, optionally a suppressor T cell, regulatory T cell, or myeloid-derived suppressor cell.

The cancer antigens are usually normal cell surface antigens which are either over-expressed or expressed at abnormal times, or are expressed by a targeted population of cells. Ideally the target antigen is expressed only on proliferative cells (e.g., tumor cells) or pro-tumoral cells (e.g. immune cells having an immunosuppressive effect), however this is rarely observed in practice. As a result, target antigens are in many cases selected on the basis of differential expression between proliferative/disease tissue and healthy tissue. Example of cancer antigens include: Receptor Tyrosine Kinase-like Orphan Receptor 1 (ROR1), Crypto, CD4, CD20, CD30, CD19, CD38, CD47, Glycoprotein NMB, CanAg, Her2 (ErbB2/Neu), a Siglec family member, for example CD22 (Siglec2) or CD33 (Siglec3), CD79, CD138, CD171, PSMA, L1-CAM, PSMA (prostate specific membrane antigen), BCMA, CD52, CD56, CD80, CD70, E-selectin, EphB2, Melanotransferrin, Mud 6 and TMEFF2. Examples of cancer antigens also include Immunoglobulin superfamily (IgSF) such as

cytokine receptors, Killer-Ig Like Receptor, CD28 family proteins, for example, Killer-Ig Like Receptor 3DL2 (KIR3DL2), B7-H3, B7-H4, B7-H6, PD-L1, IL-6 receptor. Examples also include MAGE, MART-1/Melan-A, gp100, major histocompatibility complex class I-related chain A and B polypeptides (MICA and MICB), adenosine deaminase-binding protein (ADAAbp), cyclophilin b, colorectal associated antigen (CRC)-C017-1A/GA733, protein tyrosine kinase 7 (PTK7), receptor protein tyrosine kinase 3 (TYRO-3), nectins (e.g. nectin-4), major histocompatibility complex class I-related chain A and B polypeptides (MICA and MICB), proteins of the UL16-binding protein (ULBP) family, proteins of the retinoic acid early transcript-1 (RAET1) family, carcinoembryonic antigen (CEA) and its immunogenic epitopes CAP-1 and CAP-2, etv6, aml1, prostate specific antigen (PSA), T-cell receptor/CD3-zeta chain, MAGE-family of tumor antigens, GAGE-family of tumor antigens, anti-Müllerian hormone Type II receptor, delta-like ligand 4 (DLL4), DR5, ROR1 (also known as Receptor Tyrosine Kinase-Like Orphan Receptor 1 or NTRKR1 (EC 2.7.10.1), BAGE, RAGE, LAGE-1, NAG, GnT-V, MUM-1, CDK4, MUC family, VEGF, VEGF receptors, Angiopoietin-2, PDGF, TGF- $\alpha$ , EGF, EGF receptor, members of the human EGF-like receptor family, e.g., HER-2/neu, HER-3, HER-4 or a heterodimeric receptor comprised of at least one HER subunit, gastrin releasing peptide receptor antigen, Muc-1, CA125, integrin receptors,  $\alpha\beta$ 3 integrins,  $\alpha$ 5 $\beta$ 1 integrins,  $\alpha$ 1b $\beta$ 3-integrins, PDGF beta receptor, SVE-cadherin, IL-8 receptor, hCG, IL-6 receptor, CSF1R (tumor-associated monocytes and macrophages),  $\alpha$ -fetoprotein, E-cadherin,  $\alpha$ -catenin,  $\beta$ -catenin and  $\gamma$ -catenin, p120ctn, PRAME, NY-ESO-1, cdc27, adenomatous polyposis coli protein (APC), fodrin, Connexin 37, Ig-idiotypic, p15, gp75, GM2 and GD2 gangliosides, viral products such as human papillomavirus proteins, imp-1, P1A, EBV-encoded nuclear antigen (EBNA)-1, brain glycogen phosphorylase, SSX-1, SSX-2 (HOM-MEL-40), SSX-1, SSX-4, SSX-5, SCP-1 and CT-7, and c-erbB-2, although this is not intended to be exhaustive. In one aspect, the antigen of interest is an antigen (e.g. any one of the antigens listed above) capable of undergoing intracellular internalization, for example when bound by a conventional human IgG1 antibody, either in the presence or absence of Fc $\gamma$  receptor cells. In one aspect, the antigen of interest is a CD19 or CD20 polypeptide; in one aspect, the multispecific protein comprises a VH and/or VL, or a scFv, or another ABD, that binds CD19 or CD20 comprising an amino acid sequence which is at least 60%, 70%, 80%, 85%, 90% or 95% identical to the sequence of the anti-CD19 or anti-CD20 respective VH, VL or scFv described in the Examples herein, or comprises the heavy and light chain CDR1, -2 and -3 of the anti-CD19 or anti-CD20 heavy and light chain variable regions disclosed herein. In one aspect, the multispecific protein competes for binding to a human

CD19 or CD20 polypeptide with an antibody, or a F5 or T6 protein, comprising the respective anti-CD19 or anti-CD20 VH, VL or scFv disclosed in the Examples herein.

In one embodiment, the ABD that binds an antigen of interest is derived from (e.g. comprises the hypervariable region of, or comprises one, two, three, four, five or six of the CDRs of) a parental antibody that binds an antigen of interest (e.g. a murine antibody, a human antibody) which, when bound to its antigenic target (the antigen of interest on cells), increases or induces down-modulation or intracellular internalization of the antigen of interest. In one embodiment, the antigen of interest is a cancer antigen, e.g. one of the cancer antigens listed above known to internalize (e.g. Immunoglobulin superfamily (IgSF) members, for example cytokine receptor  $\alpha$  or  $\beta$  chains, Killer-Ig Like Receptors, CD28 family proteins, B7-H3, B7-H4, B7-H6, KIR3DL2, PTK7, ROR1, L1-CAM, Siglec family members, EGF receptor and EGF-like receptor family members, EGFR, HER-2, integrins, anti-Müllerian hormone Type II receptor, CSF-1R, and others) In one embodiment, the antigen target is a polypeptide present on an immune cell capable of mediating a pro-tumoral effect, e.g. a monocyte or a macrophage, optionally a suppressor T cell, regulatory T cell, or myeloid-derived suppressor cell.

In exemplary embodiments, an ABD, variable domain or pair of complementary variable domains will bind an antigen expressed by a target cell that is to be eliminated (e.g., a tumor antigen, microbial (e.g. bacterial or parasitic) antigen, viral antigen, or antigen expressed on an immune cell that is contributing to inflammatory or autoimmune disease, and another ABD, variable domain or pair of complementary variable domains will bind to an antigen expressed on an immune cell, for example an immune effector cell, e.g. a cell surface receptor of an effector cells such as a T or NK cell. Examples of antigens expressed on immune cells, optionally immune effector cells, include antigens expressed on a member of the human lymphoid cell lineage, e.g. a human T cell, a human B cell or a human natural killer (NK) cell, a human monocyte, a human neutrophilic granulocyte or a human dendritic cell. Advantageously, such cells will have either a cytotoxic or an apoptotic effect on a target cell that is to be eliminated (e.g., that expresses a tumor antigen, microbial antigen, viral antigen, or antigen expressed on an immune cell that is contributing to inflammatory or autoimmune disease). Especially advantageously, the human lymphoid cell is a cytotoxic T cell or NK cell which, when activated, exerts a cytotoxic effect on the target cell. According to this embodiment, then, the cytotoxic activity of the human effector cells is recruited. According to another embodiment, the human effector cell is a member of the human myeloid lineage.

Antigens expressed on an immune cell to which antibodies or fragments that make up multispecific protein can bind also include NK and/or T cell receptors, e.g. any molecule

on the surface of NK cells or T cells, respectively, that can serve to direct the NK or T cell to the intended target cell to be eliminated, and preferably to permit the NK and/or T cell to mediate the elimination or lysis of the target cell. Examples include, e.g., members of the immunoglobulin superfamily, members of the killer-cell immunoglobulin-like receptor (KIR) family, the leukocyte immunoglobulin-like receptors (LILR) family, or the lectin family or the NK cell lectin-like receptor family. Activity can be measured for example by bringing target cells and effector cells into contact in presence of the multispecific polypeptide. Optionally the immune cell receptor is an immune effector cell activating receptor, e.g. an activating NK cell or T cell receptor. As used herein, the terms “activating NK cell receptor” and “activating T cell receptor” refers to any molecule on the surface of NK cells or T cells, respectively, that, when stimulated, causes a measurable increase in any property or activity known in the art as associated with NK cell or T cell activity, respectively, such as cytokine (for example IFN- $\gamma$  or TNF- $\alpha$ ) production, increases in intracellular free calcium levels, the ability to lyse target cells in a redirected killing assay as described, e.g. elsewhere in the present specification, or the ability to stimulate NK cell or T cell proliferation, respectively. The term “activating NK receptor” includes but is not limited to DNAX accessory molecule-1 (DNAM-1), 2B4, activating forms of KIR proteins (for example KIR2DS receptors, KIR2DS2, KIR2DS4), NKG2D, NKp30, CD137, CD69, NKp80, NKp44, NKp46, IL-2R, IL-12R, IL-15R, IL-18R and IL-21R. In one embodiment, the activating NK cell receptor is a receptor other than an Fc $\gamma$  receptor. In one embodiment, the activating NK cell receptor is a receptor other than NKp46.

Activation of cytotoxic T cells may occur via binding of the CD3 antigen as effector antigen on the surface of the cytotoxic T cell by a multispecific (e.g. bispecific) polypeptide of this embodiment. The human CD3 antigen is present on both helper T cells and cytotoxic T cells. Human CD3 denotes an antigen which is expressed on T cells as part of the multimolecular T cell complex and which comprises three different chains: CD3-epsilon, CD3-delta and CD3-gamma. Other effector cell antigens that can be bound by an ABD are the human CD8 antigen, the human CD2 antigen, the human CD28 antigen or the human CD25 antigen.

In one embodiment, the multispecific protein comprises one ABD that binds specifically to CD8, and one ABD that bind to CD3. In one embodiment, the multispecific protein comprises one ABD that binds specifically to an activating receptor present on effector NK cells, and one ABD that bind to an activating receptor present on effector T cells. In one embodiment, the multispecific comprises one ABD that binds to a cancer antigen, a viral antigen or a bacterial antigen.



The ABDs or variable domains which are incorporated into the polypeptides can be tested for any desired activity prior to inclusion in a polypeptide. Once appropriate antigen binding domains having desired specificity and/or activity are identified, DNA encoding each variable domain can be placed, in suitable arrangements, in an appropriate expression vector(s), together with DNA encoding any elements such as an enzymatic recognition tag, or CH2 and CH3 domains and any other optional elements (e.g. DNA encoding a linker or hinge region) for transfection into an appropriate host(s). The host is then used for the recombinant production of the polypeptide chains that make up the multispecific protein.

An ABD or variable region derived from an antibody will generally comprise at minimum a hypervariable region sufficient to confer binding activity when present in the multimeric polypeptide. It will be appreciated that an ABD or variable region may comprise other amino acids or functional domains as may be desired, including but not limited to linker elements (e.g. linker peptides, constant domain derived sequences, hinges, or fragments thereof, each of which can be placed between a variable domain and a CH1, C $\kappa$ , CH2 or CH3 domain, or between other domains as needed).

In any embodiment, ABDs or variable regions can be obtained from a humanized antibody in which residues from a complementary-determining region (CDR) of a human antibody are replaced by residues from a CDR of the original antibody (the parent or donor antibody, e.g. a murine or rat antibody) while maintaining the desired specificity, affinity, and capacity of the original antibody. The CDRs of the parent antibody, some or all of which are encoded by nucleic acids originating in a non-human organism, are grafted in whole or in part into the beta-sheet framework of a human antibody variable region to create an antibody, the specificity of which is determined by the engrafted CDRs. The creation of such antibodies is described in, e.g., WO 92/11018, Jones, 1986, Nature 321:522-525, Verhoeyen et al., 1988, Science 239:1534-1536. An antigen binding domain can thus have non-human hypervariable regions or CDRs and human frameworks region sequences (optionally with back mutations).

Polypeptide chains will be arranged in one or more expression vectors so as to produce the polypeptides having the desired domains operably linked to one another. The host cell may be of mammalian origin or may be selected from COS-1, COS-7, HEK293, BHK21, CHO, BSC-1, Hep G2, 653, SP2/0, 293, HeLa, myeloma, lymphoma, yeast, insect or plant cells, or any derivative, immortalized or transformed cell thereof.

The polypeptide can then be produced in an appropriate host cell or by any suitable synthetic process and brought into contact under appropriate conditions for the multimeric (e.g. dimer or trimer) polypeptide to form.

Polypeptide configurations

An isolated hetero-multimeric protein that binds a first, second and third antigen of interest can be prepared according to different configurations, in each case involving at least a central (first) polypeptide chain and a second polypeptide chain, and optionally a third polypeptide chain.

The first (central) polypeptide chain will provide one variable domain that will, together with a complementary variable domain on a second polypeptide chain, form an antigen binding domain specific for one (e.g. a first) antigen of interest. The first (central) polypeptide chain will also provide a second variable domain that will be paired with a complementary variable domain to form an antigen binding domain specific for another (e.g. a second) antigen of interest; the variable domain that is complementary to the second variable domain can be placed on the central polypeptide (e.g. adjacent to the second variable domain in a tandem variable domain construct such as an scFv), or can be placed on the second polypeptide chain, or can be placed on a third polypeptide chain. The second (and third, if present) polypeptide chains will associate with the central polypeptide chain by CH1-C $\kappa$  heterodimerization, forming non-covalent interactions and optionally further interchain disulfide bonds between respective hinge domains and between complementary CH1 and CK domains, with a single multimeric polypeptide being formed so long as CH/C $\kappa$  and VH/VK domains are chosen to give rise to a sole dimerization configuration. In a trimer, or when polypeptides are constructed for preparation of a trimer, there will generally be one polypeptide chain that comprises a non-naturally occurring VH-C $\kappa$  or VL-CH1 domain arrangement.

The first (central) polypeptide chain comprises a first variable domain (V) fused to a CH1 of CL constant region (e.g. the V domain is fused at its C-terminus to the N-terminus of a CH1 or CK constant region), a second variable domain, and an Fc domain (e.g. a full Fc domain or a portion thereof) interposed between the first and second variable domains may have the Examples of domain arrangement for the first polypeptide include but are not limited to:

scFv – Fc domain – VH – CH1
scFv – Fc domain – VK – CK
scFv – Fc domain – VK – CH1
scFv – Fc domain – VH – CK
(VH or VK) – Fc domain – VH – CH1
(VH or VK) – Fc domain – VK – CK
(VH or VK) – Fc domain – VK – CH1

(VH or VK) – Fc domain – VH – CK
(VH or VK) – CH1 – Fc domain – VH – CH1
(VH or VK) – CK – Fc domain – VK – CK
(VH or VK) – CK – Fc domain – VK – CH1
(VH or VK) – CH1 – Fc domain – VH – CK
(VH or VK) – CH1 – Fc domain – VK – CH1
(VH or VK) – CK – Fc domain – VH – CK
(VH or VK) – CK – Fc domain – VH – CH1
(VH or VK) – CH1 – Fc domain – VK – CK
VH - CH1– Fc domain – CH1 – (VH or VK)
VK – CK – Fc domain – CH1 – (VH or VK)
VH – CK – Fc domain – CH1 – (VH or VK)
VK - CH1– Fc domain – CH1 – (VH or VK)
VH - CH1– Fc domain – CK – (VH or VK)
VK – CK – Fc domain – CK – (VH or VK)
VH – CK – Fc domain – CK – (VH or VK)
VK - CH1– Fc domain – CK – (VH or VK) – scFV
VH - CH1– Fc domain – CH1 – (VH or VK) – scFV
VK – CK – Fc domain – CH1 – (VH or VK) – scFV
VH – CK – Fc domain – CH1 – (VH or VK) – scFV
VK - CH1– Fc domain – CH1 – (VH or VK) – scFV
VH - CH1– Fc domain – CK – (VH or VK) – scFV
VK – CK – Fc domain – CK – (VH or VK) – scFV
VH – CK – Fc domain – CK – (VH or VK) – scFV
VK - CH1– Fc domain – CK – (VH or VK) – scFV

A second polypeptide chain comprises a first variable domain (V) fused (e.g. at its C-terminus) to a CH1 or CL (e.g. CK) constant region selected to be complementary to the CH1 or CL constant region of the first polypeptide chain such that the first and second polypeptides form a CH1-CL (e.g., CH1-CK) heterodimer. The second polypeptide chain may further comprises an Fc domain (e.g. a full Fc domain or a portion thereof), e.g., fused to the C- terminus of the of the CH1 or CL domain or fused to the N-terminus of the variable domain. Examples of domain arrangement for the second polypeptide include but are not limited to:

(VH or VK) – (CH1)
(VH or VK) – (CK)
(VH or VK) – (CH1) – Fc domain
(VH or VK) – (CK) – Fc domain
(VH or VK) – (CH1 or (CK) – scFV

A third polypeptide chain, when present, can have the domain arrangement:

(VH or VK) – (CH1 or (CK)
(VH or VK) – (CH1 or (CK) – scFV

*Heterodimers with two ABDs and a dimeric Fc*

5           Examples of the domain arrangements (N- to C-terminal) of central polypeptide chains for use in such heterodimeric proteins include:

$V_{a1} - (CH1 \text{ or } CK)_a - \text{Fc domain} - V_{a2} - V_{b2};$

or

$V_{a2} - V_{b2} - \text{Fc domain} - V_{a1} - (CH1 \text{ or } CK)_a$

10          wherein  $V_{a1}$  is a light chain or heavy chain variable domain, and wherein one of  $V_{a2}$  and  $V_{b2}$  is a light chain variable domain and the other is a heavy chain variable domain.

The Fc domain of the central chain may be a full Fc domain (CH2-CH3) or a portion thereof sufficient to confer the desired functionality (e.g. CD16 and FcRn binding). A second polypeptide chain will then be configured which will comprise an immunoglobulin variable domain and a CH1 or CK constant region, e.g., a  $(CH1 \text{ or } CK)_b$  unit, selected so as to permit CH1-CK heterodimerization with the central polypeptide chain; the immunoglobulin variable domain will be selected so as to complement the variable domain of the central chain that is adjacent to the CH1 or CK domain, whereby the complementary variable domains form an antigen binding domain for a first antigen of interest.

20           For example, a second polypeptide chain can comprise a domain arrangement:

$V_{b1} - (CH1 \text{ or } CK)_b - \text{Fc domain}$

such that the  $(CH1 \text{ or } CK)_b$  dimerizes with the  $(CH1 \text{ or } CK)_a$  on the central chain, and the  $V_{b1}$  forms an antigen binding domain together with  $V_{a1}$  of the central chain. If  $V_{a1}$  of the central chain is a light chain variable domain,  $V_{b1}$  will be a heavy chain variable domain; and if  $V_{a1}$  of the central chain is a heavy chain variable domain,  $V_{b1}$  will be a light chain variable domain.

The antigen binding domain for the second antigen of interest can then be formed from  $V_{a2}$  and  $V_{b2}$  which are configured as tandem variable domains on the central or second

chain, thereby forming the antigen binding domain for the second antigen of interest (e.g. a heavy chain variable domain (VH) and a light chain (kappa) variable domain (VK), for example forming an scFv unit). The antigen binding domain for the second antigen of interest can also alternatively be formed from a single variable domain  $V_2$  present on the central chain.

The resulting heterodimer can in another example have the configuration as follows (see also Examples of such proteins shown as format 13 shown in Figure 2D):

$V_{a1} - (\text{CH1 or CK})_a - \text{Fc domain} - V_{a2} - V_{b2}$  (first/central polypeptide chain)

$V_{b1} - (\text{CH1 or CK})_b - \text{Fc domain}$  (second polypeptide chain)

wherein one of  $V_{a1}$  of the first polypeptide chain and  $V_{b1}$  of the second polypeptide chain is a light chain variable domain and the other is a heavy chain variable domain, and wherein one of  $V_{a2}$  and  $V_{b2}$  is a light chain variable domain and the other is a heavy chain variable domain.

In one embodiment, the heterodimeric bispecific Fc-derived polypeptide comprises a domain arrangement of one of the following, optionally wherein one or both hinge domains are replaced by a peptide linker, optionally wherein the Fc domain is fused via a peptide linker to an scFv that binds a polypeptide expressed by an immune effector cell (e.g. T cell, NK cell, etc.):

(VK-CK - hinge) – Fc domain – (anti-activating receptor ABD)

|  
(VH-CH1- hinge) – Fc domain

or

(VH-CH1- hinge) – Fc domain – (anti-activating receptor ABD)

|  
(VK-CK- hinge) – Fc domain

or

(VH-CH1- hinge) – Fc domain

|  
(VK-CK- hinge) – Fc domain – (anti-activating receptor ABD)

The anti-activating receptor ABD can for example be any ABD that binds to an activating receptor on an immune cell, e.g. activating NK cell receptor. The ABD may for example comprise an scFv.

Examples of domain arrangement for the dimeric polypeptide formed include but are not limited to those in the table below:

VK – VH – Fc domain – VH – (CH1)
Fc domain – VK – (CK)
VH – VK – Fc domain – VH – (CH1)

Fc domain – VK – (CK)
VK – VH – Fc domain – VK – (CH1)
Fc domain – VH – (CK)
VH – VK – Fc domain – VK – (CH1)
Fc domain – VH – (CK)
VH – (CH1) – Fc domain – VH – VK
VK – (CK) – Fc domain
VH – (CH1) – Fc domain – VK – VH
VK – (CK) – Fc domain
VK – (CH1) – Fc domain – VH – VK
VH – (CK) – Fc domain
VK – (CH1) – Fc domain – VK – VH
VH – (CK) – Fc domain

*Heterotrimers with two ABDs and a dimeric Fc*

Heterotrimeric proteins can for example be formed by using a central (first) polypeptide chain comprising a first variable domain (V) fused to a first CH1 or CK constant region, a second variable domain (V) fused to a second CH1 or CK constant region, and an Fc domain or portion thereof interposed between the first and second variable domains (i.e. the Fc domain is interposed between the first and second (V-(CH1/CK)) units. For example, a central polypeptide chain for use in a heterotrimeric protein can have the domain arrangements (N- to C- terminal) as follows:

$V_{a1} - (CH1 \text{ or } CK)_a - Fc \text{ domain} - V_{a2} - (CH1 \text{ or } CK)_b$ .

A second polypeptide chain can comprise a domain arrangement (N- to C- terminal):

$V_{b1} - (CH1 \text{ or } CK)_c - Fc \text{ domain}$

such that the  $(CH1 \text{ or } CK)_c$  dimerizes with the  $(CH1 \text{ or } CK)_a$  on the central chain, and the  $V_{a1}$  and  $V_{b1}$  form an antigen binding domain.

A third polypeptide chain can comprise a domain arrangement (N- to C- terminal):

$V_{b2} - (CH1 \text{ or } CK)_d$ ,

such that the  $(CH1 \text{ or } CK)_d$  dimerizes with the  $(CH1 \text{ or } CK)_b$  unit on the central chain, and the  $V_{a2}$  and  $V_{b2}$  form an antigen binding domain.

An example of a configuration of a resulting heterotrimer with a dimeric Fc domain (also shown as format 5 in Figure 2D) has a domain arrangement:

$V_{b1} - (\text{CH1 or CK})_c - \text{Fc domain}$  (second polypeptide)

$V_{a1} - (\overset{|}{\text{CH1}} \text{ or CK})_a - \text{Fc domain} - V_{a2} - (\text{CH1 or CK})_b$  (first polypeptide)

5  $V_{b2} - (\overset{|}{\text{CH1}} \text{ or CK})_d$  (third polypeptide)

Thus, in a configuration of a trimer polypeptide, the first polypeptide can have two variable domains that each form an antigen binding domain with a variable domain on a separate polypeptide chain (i.e. the variable domain of the second and third chains), the second polypeptide chain has one variable domain, and the third polypeptide has one variable domain, and wherein the trimer has a dimeric Fc domain that binds CD16.

Examples of domain arrangement for the trimeric bispecific polypeptide formed from include but are not limited to:

$\text{VK} - \text{CK} - \text{Fc domain}$	(second polypeptide)
$\text{VH} - \overset{ }{\text{CH1}} - \text{Fc domain} - \text{VH} - \text{CK}$	(first polypeptide)
$\text{VK} - \overset{ }{\text{CH1}}$	(third polypeptide)
$\text{VH} - \text{CH1} - \text{Fc domain}$	(second polypeptide)
$\text{VK} - \overset{ }{\text{CK}} - \text{Fc domain} - \text{VH} - \text{CK}$	(first polypeptide)
$\text{VK} - \overset{ }{\text{CH1}}$	(third polypeptide)
$\text{VH} - \text{CK} - \text{Fc domain}$	(second polypeptide)
$\text{VK} - \overset{ }{\text{CH1}} - \text{Fc domain} - \text{VH} - \text{CH1}$	(first polypeptide)
$\text{VK} - \overset{ }{\text{CK}}$	(third polypeptide)

#### 15 *Heterodimers with three ABDs (and optionally further a dimeric Fc)*

Examples of the domain arrangements (N- to C-terminal) of central polypeptide chains for use in such heterodimeric proteins include:

$V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK})_a$

wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain or heavy chain variable domain

or

$V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK})_a - V_3 - V_3$

wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain or heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain. The  $V_1$

pair will form a first ABD, the  $V_2$  pair will form a second ABD together with a complementary variable domain on a second polypeptide chain, and the two  $V_3$  domains will pair to form a third ABD. The Fc domain of the central chain may be a full Fc domain (CH2-CH3) or a portion thereof sufficient to confer the desired functionality (e.g. FcRn binding and/or CD16 binding). A second polypeptide chain will then be configured which will comprise an immunoglobulin variable domain and a CH1 or CK constant region, e.g., a (CH1 or CK)<sub>b</sub> unit, selected so as to permit CH1-CK heterodimerization with the central polypeptide chain; the immunoglobulin variable domain will be selected so as to complement the variable domain of the central chain that is adjacent to the CH1 or CK domain, whereby the complementary variable domains form an antigen binding domain for a first antigen of interest.

For example, a second polypeptide chain can comprise a domain arrangement:

(VH or VK) – (CH1 or (CK)<sub>b</sub>) – scFv

or

(VH or VK) – (CH1 or (CK)<sub>b</sub>).

such that the (CH1 or CK)<sub>b</sub> dimerizes with the (CH1 or CK)<sub>a</sub> on the central chain, and the V domain of the second chain forms an antigen binding domain together with  $V_2$  of the central chain. If  $V_2$  of the central chain is a light chain variable domain, the V of the second chain will be a heavy chain variable domain; and if  $V_2$  of the central chain is a heavy chain variable domain, the V of the second chain will be a light chain variable domain. If the central chain lacks the C-terminal scFv, then the second polypeptide will be chosen to comprise the C-terminal scFv.

When the central chain lacking the C-terminal scFv is employed, the resulting hetero-multimeric polypeptide is formed having the domain arrangement:

$V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK})$  (first/central polypeptide chain)

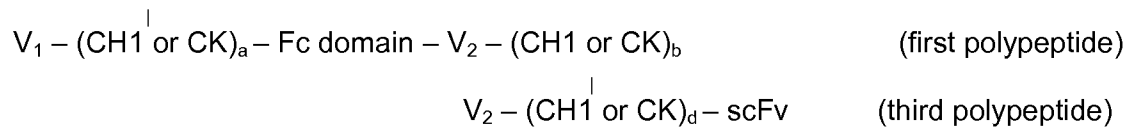
$V_2 - (\text{CH1 or CK}) - V_3 - V_3$  (second polypeptide chain)

wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain. The  $V_1$  pair will form a first ABD, the  $V_2$  pair will form a second ABD, and the  $V_3$  will pair to form a third ABD. The protein is also illustrated as format T11 in Figure 2F.

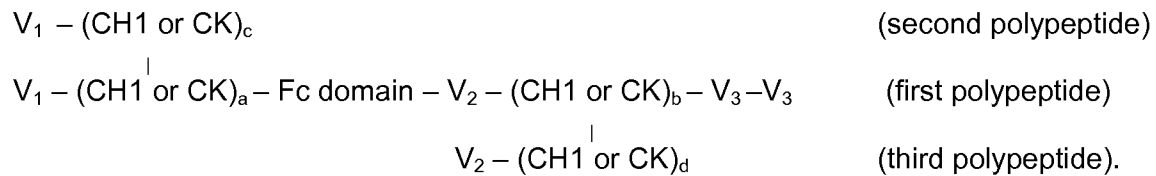
In another example, when the central chain lacking the C-terminal scFv is employed, the first (central) polypeptide chain has the domain arrangement:  $V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK}) - V_3 - V_3$ , such that a hetero-multimeric polypeptide is formed having the domain arrangement:



$$\begin{array}{ll} V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK}) - V_3 - V_3 & \text{(first/central polypeptide chain)} \\ \quad \quad \quad | & \\ \quad \quad \quad V_2 - (\text{CH1 or CK}) & \text{(second polypeptide chain)} \end{array}$$



In another example, the first (central) polypeptide chain has the domain arrangement:  $V_1 - (\text{CH1 or CK})_a - \text{Fc domain} - V_2 - (\text{CH1 or CK})_b - V_3 - V_3$ . The hetero-multimeric polypeptide formed can have the domain arrangement:



In any of the polypeptide chains herein, a hinge region will typically be present on a polypeptide chain between a CH1 domain and a CH2 domain of an Fc domain, and/or can be present between a CK domain and a CH2 domain. A hinge region can optionally be replaced for example by a suitable linker peptide.

The proteins domains described in the present disclosure can optionally be specified as being from N- to C- terminal. Protein arrangements of the disclosure for purposes of illustration are shown from N-terminus (on the left) to C-terminus. Domains can be referred to as fused to one another (e.g. a domain can be said to be fused to the C-terminus of the domain on its left, and/or a domain can be said to be fused to the N-terminus of the domain on its right).

The proteins domains described in the present disclosure can be fused to one another directly or via intervening amino acid sequences. For example, a CH1 or CK domain will be fused to an Fc domain (or CH2 or CH3 domain thereof) via a linker peptide, optionally a hinge region or a fragment thereof. In another example, a VH or VK domain will be fused to a CH3 domain via a linker peptide. VH and VL domains linked to another in tandem will be fused via a linker peptide (e.g. as an scFv). VH and VL domains linked to an Fc domain will be fused via a linker peptide. Two polypeptide chains will be bound to one another (indicated by “<sub>1</sub>”) by non-covalent bonds and optionally further by interchain disulfide bonds formed between cysteine residues within complementary CH1 and CK domains.

It will be appreciated that in any embodiment herein, a VK domain can be replaced by a Vλ variable domain.

In any of the domain arrangements, the Fc domain may comprise a CH2-CH3 unit (a full length CH2 and CH3 domain or a fragment thereof). In heterodimers or heterotrimers comprising two chains with Fc domains (a dimeric Fc domain), the CH3 domain will be

capable of CH3-CH3 dimerization (e.g. a wild-type CH3 domain). In heterodimers or heterotrimers comprising only one chain with an Fc domain (monomeric Fc domain), the Fc domain will be incapable of CH3-CH3 dimerization; for example the CH3 domain(s) will have amino acid modification(s) in the CH3 dimer interface or the Fc domain will comprise a tandem CH3 domain incapable of CH3-CH3 dimerization. In one embodiment of any aspect herein, a first CH3 domain is connected to a second CH3 domain by a linker. The tandem CH3 domain may have the domain arrangement, from N-terminus to C-terminus, as follows:

– CH3 – linker – CH3 –.

The linker in the tandem CH3 domain will be a flexible linker (e.g. peptide linker). In one embodiment the linker permits the CH3 domains to associate with one another by non-covalent interactions. In one embodiment, the linker is a peptide linker having 10-50 amino acid residues. In one embodiment, the linker has the formula (G<sub>4</sub>S)<sub>x</sub>. Optionally, x is 2, 3, 4, 5 or 6. In any of the embodiments, each CH3 domain is independently a full-length and/or native CH3 domain, or a fragment or modified CH3 domain which retains a functional CH3 dimerization interface.

An exemplary tandem CH3 with a flexible peptide linker (underlined) is shown below. An exemplary tandem CH3 domain can thus comprise an amino acid sequence of SEQ ID NO : 2, or a sequence at least 70%, 80%, 90%, 95% or 98% identical thereto:

GQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDG  
SFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPG GGGGSGGGSGGGGS GQP  
REPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFF  
LYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPG (SEQ ID NO : 2)

Tandem CH3 domains disclosed herein and CH3 domains with amino acid modification to prevent CH3-CH3 dimerization will retain partial FcRn binding (compared, e.g., to a wild type full length human IgG1 antibody). The examples of monomeric CH2-CH3 domains provided herein retain partial FcRn binding but have decreased human Fcγ receptor binding. Optionally the multimeric polypeptide is capable of binding to human FcRn with intermediate affinity, e.g. retains binding to FcRn but has decreased binding to a human FcRn receptor compared to a full-length wild type human IgG1 antibody. The Fc moiety may further comprise one or more amino acid modifications, e.g. in the CH2 domain, that decreases further (e.g. abolishes) binding to one or more Fcγ receptors.

The multimeric polypeptides with monomeric Fc domains can advantageously comprise a CH2 domain and a CH3 domain, wherein said CH3 domain comprises a modified CH3 dimer interface (e.g. a mutations in the CH3 dimer interface) to prevent dimerization with another Fc-derived polypeptide. In one embodiment a CH2-CH3 portion

comprising a CH3 domain modified to prevent homodimer formation comprises an amino acid sequence shown below, or a sequence at least 90, 95% or 98% identical thereto:

APELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKP  
REEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLP  
5 PSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSGDSFFLTSLKLTVD  
KSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPG, optionally further comprising a  
substitution at 1, 2, 3, 4, 5, 6 of residues 121, 136, 165, 175, 177 or 179 of the sequence.

In one embodiment of any of the polypeptides or methods herein, the CH3 domain comprises an amino acid substitution at 1, 2, 3, 4, 5, 6 or 7 of the positions L351, T366,  
10 L368, P395, F405, T407 (or Y407) and/or K409 (EU numbering as in Kabat).

When two variable regions that form an antigen binding domain are placed on the same polypeptide chain they are typically linked together by a linker of sufficient length to enable the ABD to fold in such a way as to permit binding to the antigen for which the ABD is intended to bind, e.g., they can form a scFv. Examples of linkers include, for example,  
15 linkers comprising glycine and serine residues, e.g., the amino acid sequence GEGTSTGS(G<sub>2</sub>S)<sub>2</sub>GGAD. In another specific embodiment, the VH domain and VL domains of an svFv are linked together by the amino acid sequence (G<sub>4</sub>S)<sub>3</sub>.

An ABD can be linked to a constant domain or Fc domain via a linker (e.g. a flexible polypeptide linker) that permits the ABD to be positioned such that it binds to its target  
20 antigen and exhibits the desired functionality, e.g. it possesses a sufficient range of motion relative to the rest of the multispecific protein (the Fc domain and/or other ABD) and thereby mediates signaling at a cell surface activating receptor. Examples of linkers include, for example, linkers derived from antibody hinge regions, an amino sequence RTVA, or linkers comprising glycine and serine residues, e.g., the amino acid sequence  
25 GEGTSTGS(G<sub>2</sub>S)<sub>2</sub>GGAD. In another specific embodiment, the V<sub>H</sub> domain and V<sub>L</sub> domains of a scFv are linked together by the amino acid sequence (G<sub>4</sub>S)<sub>3</sub>. Such linkers can be used particularly advantageously to link and ABD to a constant region or Fc domain when the ABD comprises two variable regions that are placed on the same polypeptide chain (e.g., an scFv)

Any of the peptide linkers contained in the subject multispecific proteins may comprise a length of at least 4 residues, at least 5 residues, at least 10 residues, at least 15 residues, at least 20 residues, at least 25 residues, at least 30 residues or more. In other  
30 embodiments, the linkers comprise a length of between 2-4 residues, between 2-4 residues, between 2-6 residues, between 2-8 residues, between 2-10 residues, between 2-12 residues, between 2-14 residues, between 2-16 residues, between 2-18 residues, between  
35 2- 20 residues, between 2-22 residues, between 2-24 residues, between 2-26 residues,

between 2-28 residues, between 2-30 residues, between 2 and 50 residues, or between 10 and 50 residues.

An ABD (e.g. an immunoglobulin variable region) can optionally be linked to a constant domain or Fc domain via a flexible linker (e.g. polypeptide linker) that leads to less structural rigidity or stiffness (e.g. between or amongst the ABD and Fc domain) compared to a conventional (e.g. wild-type full length human IgG) antibody. For example, the multispecific protein may have a structure or a flexible linker between the ABD and constant domain or Fc domain that permits an increased range of domain motion as compared to ABD in a conventional (e.g. wild-type full length human IgG) antibody. In particular, the structure or a flexible linker can be configured to confer on the antigen binding sites greater intrachain domain movement compared to antigen binding sites in a conventional human IgG1 antibody. Rigidity or domain motion/interchain domain movement can be determined, e.g., by computer modeling, electron microscopy, spectroscopy such as Nuclear Magnetic Resonance (NMR), X-ray crystallography (B-factors), or Sedimentation Velocity Analytical ultracentrifugation (AUC) to measure or compare the radius of gyration of proteins comprising the linker or hinge. A test protein or linker may have lower rigidity relative to a comparator protein if the test protein has a value obtained from one of the tests described in the previous sentence differs from the value of the comparator, e.g., an IgG1 antibody or a hinge, by at least 5%, 10%, 25%, 50%, 75%, or 100%. A person of skill in the art would be able to determine from the tests whether a test protein has at lower rigidity to that of another protein, respectively, by interpreting the results of these tests.

In one embodiment, the multispecific protein may have a structure or a flexible linker between the ABD and constant domain or Fc domain that permits two ABDs to have a spacing between said two ABDs comprising less than about 80 angstroms, less than about 60 angstroms or ranges from about 40-60 angstroms.

In one embodiment, a CH1 or CK domain is linked or fused to an Fc domain (e.g. to an CH2 or CH3 domain of an Fc domain) via a linker that comprises a fragment of a CH1 domain and/or hinge region. For example, a N-terminal amino acid sequence of CH1 can be fused to a variable domain in order to mimic as closely as possible the natural structure of a wild-type antibody. In one embodiment, the linker comprises an amino acid sequence from a hinge domain or an N-terminal CH1 amino acid. The sequence can be, for example, between 2-4 residues, between 2-4 residues, between 2-6 residues, between 2-8 residues, between 2-10 residues, between 2-12 residues, between 2-14 residues, between 2-16 residues, between 2-18 residues, between 2- 20 residues, between 2-22 residues, between

2-24 residues, between 2-26 residues, between 2-28 residues, or between 2-30 residues. In one embodiment linker comprises or consists of the amino acid sequence RTVA.

In one embodiment, a CH1 or C $\kappa$  domain is linked or fused to an Fc domain via a hinge region (or fragment thereof) derived from a hinge domain of a human IgG1 antibody. For example a hinge domain may comprise the amino acid sequence: T-H-T-C-S-S-C-P-A-P-E-L-L (one letter code), or an amino acid sequence at least 60%, 70%, 80% or 90% identical thereto, optionally wherein one or both cysteines are deleted or substituted by a different amino acid residue.

In one embodiment, the hinge region (or fragment thereof) is derived from a C $\mu$ 2-C $\mu$ 3 hinge domain of a human IgM antibody. For example a hinge domain may comprise the amino acid sequence: N-A-S-S-M-C-V-P-S-P-A-P-E-L-L (one letter code), or an amino acid sequence at least 60%, 70%, 80% or 90% identical thereto, optionally wherein one or both cysteines are deleted or substituted by a different amino acid residue.

Polypeptide chains that dimerize and associate with one another via non-covalent bonds may or may not additionally be bound by an interchain disulfide bond formed between respective CH1 and C $\kappa$  domains, and/or between respective hinge domains on the chains. CH1, C $\kappa$  and/or hinge domains (or other suitable linking amino acid sequences) can optionally be configured such that interchain disulfide bonds are formed between chains such that the desired pairing of chains is favored and undesired or incorrect disulfide bond formation is avoided. For example, when two polypeptide chains to be paired each possess a CH1 or C $\kappa$  adjacent to a hinge domain, the polypeptide chains can be configured such that the number of available cysteines for interchain disulfide bond formation between respective CH1/C $\kappa$ -hinge segments is reduced (or is entirely eliminated). For example, the amino acid sequences of respective CH1, C $\kappa$  and/or hinge domains can be modified to remove cysteine residues in both the CH1/C $\kappa$  and the hinge domain of a polypeptide; thereby the CH1 and C $\kappa$  domains of the two chains that dimerize will associate via non-covalent interaction(s).

In another example, the CH1 or C $\kappa$  domain adjacent (e.g., N-terminal to) a hinge domain comprises a cysteine capable of interchain disulfide bond formation, and the hinge domain which is placed at the C-terminus of the CH1 or C $\kappa$  comprises a deletion or substitution of one or both cysteines of the hinge (e.g. Cys 239 and Cys 242, as numbered for human IgG1 hinge according to Kabat). In one embodiment, the hinge region (or fragment thereof) comprise the amino acid sequence: T-H-T-S-P-P-S-P-A-P-E-L-L (one letter code), or an amino acid sequence at least 60%, 70%, 80% or 90% identical thereto.

In another example, the CH1 or C $\kappa$  domain adjacent (e.g., N-terminal to) a hinge domain comprises a deletion or substitution at a cysteine residue capable of interchain

disulfide bond formation, and the hinge domain placed at the C-terminus of the CH1 or C $\kappa$  comprises one or both cysteines of the hinge (e.g. Cys 239 and Cys 242, as numbered for human IgG1 hinge according to Kabat). In one embodiment, the hinge region (or fragment thereof) comprises the amino acid sequence: T-H-T-C-S-S-C-P-A-P-E-L-L (one letter code),  
5 or an amino acid sequence at least 60%, 70%, 80% or 90% identical thereto.

In another example, a hinge region is derived from an IgM antibody. In such embodiments, the CH1/CK pairing mimics the C $\mu$ 2 domain homodimerization in IgM antibodies. For example, the CH1 or C $\kappa$  domain adjacent (e.g., N-terminal to) a hinge domain comprises a deletion or substitution at a cysteine capable of interchain disulfide  
10 bond formation, and an IgM hinge domain which is placed at the C-terminus of the CH1 or C $\kappa$  comprises one or both cysteines of the hinge. In one embodiment, the hinge region (or fragment thereof) comprises the amino acid sequence: T-H-T-C-S-S-C-P-A-P-E-L-L (one letter code), or an amino acid sequence at least 60%, 70%, 80% or 90% identical thereto.

Constant region domains can be derived from any suitable human antibody,  
15 including, the constant heavy (CH1) and light (C $\kappa$ ) domains, hinge domains, CH2 and CH3 domains. "CH1" generally refers to positions 118-220 according to the EU index as in Kabat. "CH2" generally refers to positions 237-340 according to the EU index as in Kabat, and "CH3" generally refers to positions 341-447 according to the EU index as in Kabat.

A "hinge" or "hinge region" or "antibody hinge region" herein refers to the flexible  
20 polypeptide or linker between the first and second constant domains of an antibody. Structurally, the IgG CH1 domain ends at EU position 220, and the IgG CH2 domain begins at residue EU position 237. Thus for an IgG the hinge generally includes positions 221 (D221 in IgG1) to 236 (G236 in IgG1), wherein the numbering is according to the EU index as in Kabat. References to specific amino acid residues within constant region domains  
25 found within the polypeptides shall be, unless otherwise indicated or as otherwise dictated by context, be defined according to Kabat, in the context of an IgG antibody.

CH2 and CH3 domains which may be present in the subject antibodies or multispecific proteins can be derived from any suitable antibody. Such CH2 and CH3 domains can be used as wild-type domains or may serve as the basis for a modified CH2 or  
30 CH3 domain. Optionally the CH2 and/or CH3 domain is of human origin or may comprise that of another species (e.g., rodent, rabbit, non-human primate) or may comprise a modified or chimeric CH2 and/or CH3 domain, e.g., one comprising portions or residues from different CH2 or CH3 domains, e.g., from different antibody isotypes or species antibodies .

In embodiments where a multispecific is intended not to bind to human CD16A  
35 polypeptide, a CH2 and/or CH3 domain (or Fc domain comprising same) may comprise a

modification to decrease or abolish binding to FcγRIIIA (CD16). For example, CH2 mutations in a dimeric Fc domain proteins at residue N297 (Kabat numbering) can eliminate CD16A binding. However the person of skill in the art will appreciate that other configurations can be implemented. For example, substitutions into human IgG1 of IgG2 residues at positions 233-236 and IgG4 residues at positions 327, 330 and 331 were shown to greatly reduce binding to Fcγ receptors and thus ADCC and CDC. Furthermore, Idusogie *et al.* (2000) J Immunol. 164(8):4178-84 demonstrated that alanine substitution at different positions, including K322, significantly reduced complement activation.

In certain embodiments herein where binding to CD16A is desired, a CH2 and/or CH3 domain (or Fc domain comprising same) may be wild-type domains or may comprise one or more amino acid modifications (e.g. amino acid substitutions) which increase binding to human CD16 and optionally another receptor such as FcRn. Optionally, the modifications will not substantially decrease or abolish the ability of the Fc-derived polypeptide to bind to neonatal Fc receptor (FcRn), e.g. human FcRn. Typical modifications include modified human IgG1-derived constant regions comprising at least one amino acid modification (e.g. substitution, deletions, insertions), and/or altered types of glycosylation, e.g., hypofucosylation. Such modifications can affect interaction with Fc receptors: FcγRI (CD64), FcγRII (CD32), and FcγRIII (CD16). FcγRI (CD64), FcγRIIA (CD32A) and FcγRIII (CD 16) are activating (i.e., immune system enhancing) receptors while FcγRIIB (CD32B) is an inhibiting (i.e., immune system dampening) receptor. A modification may, for example, increase binding of the Fc domain to FcγRIIIa on effector (e.g. NK) cells and/or decrease binding to FcγRIIB. Examples of modifications are provided in PCT publication no. WO2014/044686, the disclosure of which is incorporated herein by reference. Specific mutations in IgG1 which affect (enhance) FcγRIIIa or FcRn binding are also set forth below.

Isotype	Species	Modification	Effector Function	Effect of Modification
IgG1	Human	T250Q/M428L	Increased binding to FcRn	Increased half-life
IgG1	Human	1M252Y/S254T/T256E + H433K/N434F	Increased binding to FcRn	Increased half-life
IgG1	Human	E333A	Increased binding to FcγRIIIa	Increased ADCC and CDC
IgG1	Human	S239D/A330L/I332E	Increased binding to FcγRIIIa	Increased ADCC
IgG1	Human	P257I/Q311	Increased	Unchanged



			binding to FcRn	half-life
IgG1	Human	S239D/I332E/G236A	Increased FcγRIIIa/FcγRIIb ratio	Increased macrophage phagocytosis

In some embodiments, the multispecific protein comprises a variant Fc region comprise at least one amino acid modification (for example, possessing 1, 2, 3, 4, 5, 6, 7, 8, 9, or more amino acid modifications) in the CH2 and/or CH3 domain of the Fc region, wherein the modification enhances binding to a human CD16 polypeptide. In other embodiments, the multispecific protein comprises at least one amino acid modification (for example, 1, 2, 3, 4, 5, 6, 7, 8, 9, or more amino acid modifications) in the CH2 domain of the Fc region from amino acids 237-341, or within the lower hinge-CH2 region that comprises residues 231-341. In some embodiments, the multispecific protein comprises at least two amino acid modifications (for example, 2, 3, 4, 5, 6, 7, 8, 9, or more amino acid modifications), wherein at least one of such modifications is within the CH3 region and at least one such modifications is within the CH2 region. Encompassed also are amino acid modifications in the hinge region. In one embodiment, encompassed are amino acid modifications in the CH1 domain, optionally in the upper hinge region that comprises residues 216-230 (Kabat EU numbering). Any suitable functional combination of Fc modifications can be made, for example any combination of the different Fc modifications which are disclosed in any of United States Patents Nos. US, 7,632,497; 7,521,542; 7,425,619; 7,416,727; 7,371,826; 7,355,008; 7,335,742; 7,332,581; 7,183,387; 7,122,637; 6,821,505 and 6,737,056; and/or in PCT Publications Nos. WO2011/109400; WO 2008/105886; WO 2008/002933; WO 2007/021841; WO 2007/106707; WO 06/088494; WO 05/115452; WO 05/110474; WO 04/1032269; WO 00/42072; WO 06/088494; WO 07/024249; WO 05/047327; WO 04/099249 and WO 04/063351; and/or in Lazar et al. (2006) *Proc. Nat. Acad. Sci. USA* 103(11): 405-410; Presta, L.G. et al. (2002) *Biochem. Soc. Trans.* 30(4):487-490; Shields, R.L. et al. (2002) *J. Biol. Chem.* 277(30):26733-26740 and Shields, R.L. et al. (2001) *J. Biol. Chem.* 276(9):6591-6604).

In some embodiments, the multispecific protein comprises an Fc domain comprising at least one amino acid modification (for example, 1, 2, 3, 4, 5, 6, 7, 8, 9, or more amino acid modifications) relative to a wild-type Fc region, such that the molecule has an enhanced binding affinity for human CD16 relative to the same molecule comprising a wild-type Fc region, optionally wherein the variant Fc region comprises a substitution at any one or more of positions 221, 239, 243, 247, 255, 256, 258, 267, 268, 269, 270, 272, 276, 278, 280, 283, 285, 286, 289, 290, 292, 293, 294, 295, 296, 298, 300, 301, 303, 305, 307, 308, 309, 310,

311, 312, 316, 320, 322, 326, 329, 330, 332, 331, 332, 333, 334, 335, 337, 338, 339, 340, 359, 360, 370, 373, 376, 378, 392, 396, 399, 402, 404, 416, 419, 421, 430, 434, 435, 437, 438 and/or 439 (Kabat EU numbering).

In one embodiment, the multispecific protein comprises an Fc domain comprising at least one amino acid modification (for example, 1, 2, 3, 4, 5, 6, 7, 8, 9, or more amino acid modifications) relative to a wild-type Fc region, such that the molecule has enhanced binding affinity for human CD16 relative to a molecule comprising a wild-type Fc region, optionally wherein the variant Fc region comprises a substitution at any one or more of positions 239, 298, 330, 332, 333 and/or 334 (e.g. S239D, S298A, A330L, I332E, E333A and/or K334A substitutions), optionally wherein the variant Fc region comprises a substitution at residues S239 and I332, e.g. a S239D and I332E substitution (Kabat EU numbering).

In some embodiments, the multispecific protein comprises an Fc domain comprising altered glycosylation patterns that increase binding affinity for human CD16. Such carbohydrate modifications can be accomplished by, for example, by expressing a nucleic acid encoding the multispecific protein in a host cell with altered glycosylation machinery. Cells with altered glycosylation machinery are known in the art and can be used as host cells in which to express recombinant antibodies to thereby produce an antibody with altered glycosylation. See, for example, Shields, R.L. et al. (2002) *J. Biol. Chem.* 277:26733-26740; Umana et al. (1999) *Nat. Biotech.* 17:176-1, as well as, European Patent No: EP 1,176,195; PCT Publications WO 06/133148; WO 03/035835; WO 99/54342, each of which is incorporated herein by reference in its entirety. In one aspect, the multispecific protein contains one or more hypofucosylated constant regions. Such multispecific protein may comprise an amino acid alteration or may not comprise an amino acid alteration and/or may be expressed or synthesized or treated under conditions that result in hypofucosylation. In one aspect, a multispecific protein composition comprises a multispecific protein described herein, wherein at least 20, 30, 40, 50, 60, 75, 85, 90, 95% or substantially all of the antibody species in the composition have a constant region comprising a core carbohydrate structure (e.g. complex, hybrid and high mannose structures) which lacks fucose. In one embodiment, provided is a multispecific protein composition which is free of antibodies comprising a core carbohydrate structure having fucose. The core carbohydrate will preferably be a sugar chain at Asn297.

Optionally, a multispecific protein comprising a dimeric Fc domain can be characterized by having a binding affinity to a human CD16 polypeptide that is within 1-log of that of a conventional human IgG1 antibody, e.g., as assessed by surface plasmon resonance.

In one embodiment, the multispecific protein comprising a dimeric Fc domain engineered to enhance Fc receptor binding can be characterized by having a binding affinity to a human CD16 polypeptide that is at least 1-log greater than that of a conventional or wild-type human IgG1 antibody, e.g., as assessed by surface plasmon resonance.

5        Optionally a multispecific protein comprising a dimeric Fc domain can be characterized by a  $K_d$  for binding (monovalent) to a human CD16 polypeptide of less than  $10^{-5}$  M (10  $\mu$ molar), optionally less than  $10^{-6}$  M (1  $\mu$ molar), as assessed by surface plasmon resonance (e.g. as in Example 13, SPR measurements performed on a Biacore T100 apparatus (Biacore GE Healthcare), with bispecific antibodies immobilized on a Sensor Chip  
10        CM5 and serial dilutions of soluble CD16 polypeptide injected over the immobilized bispecific antibodies.

In one embodiment, the disclosure provides methods of making a heterodimeric protein (e.g. any heterodimeric protein described herein), comprising:

a)        providing a first nucleic acid encoding a first polypeptide chain described  
15        herein (e.g., a polypeptide chain comprising a first variable domain (V) fused to a CH1 of CK constant region, a second variable domain (and optionally third variable domain, wherein the second and third variable domain form a first antigen binding domain), and an Fc domain or portion thereof interposed between the first and second variable domains);

b)        providing a second nucleic acid encoding a second polypeptide chain  
20        described herein (e.g., a polypeptide chain comprising a first variable domain (V) fused at its C-terminus to a CH1 or CK constant region selected to be complementary to the CH1 or CK constant region of the first polypeptide chain such that the first and second polypeptides form a CH1-CK heterodimer in which the first variable domain of the first polypeptide chain and the first variable domain of the second polypeptide form a second antigen binding  
25        domain, and an Fc domain or portion thereof); and

c)        expressing said first and second nucleic acids in a host cell to produce a protein comprising said first and second polypeptide chains, respectively; and recovering a heterodimeric protein comprising a dimeric Fc domain capable of binding human CD16. Optionally, the heterodimeric protein produced represents at least 20%, 25% or 30% of the  
30        total proteins (e.g. bispecific proteins) prior to purification. Optionally step (c) comprises loading the protein produced onto an affinity purification support, optionally an affinity exchange column, optionally a Protein-A support or column, and collecting the heterodimeric protein; and/or loading the protein produced (or the protein collected following loading onto an affinity exchange or Protein A column) onto an ion exchange column; and collecting the  
35        heterodimeric fraction. In one embodiment, the second variable domain (optionally together with the third variable domain) of the first polypeptide chain binds NKp46.

By virtue of their ability to be produced in standard cell lines and standardized methods with high yields, unlike BiTE™, DART™ and other bispecific formats, the proteins of the disclosure also provide a convenient tool for screening for the most effective variable regions to incorporate into a multispecific protein. In one aspect, the present disclosure provides a method for identifying or evaluating candidate variable regions for use in a heterodimeric protein, comprising the steps of:

a) providing a plurality of nucleic acid pairs, wherein each pair includes one nucleic acid encoding a heavy chain candidate variable region and one nucleic acid encoding a light chain candidate variable region, for each of a plurality of heavy and light chain variable region pairs (e.g., obtained from different antibodies binding the same or different antigen(s) of interest);

b) for each of the plurality nucleic acid pairs, making a heterodimeric protein by:

(i) producing a first nucleic acid encoding a first polypeptide chain comprising one of the heavy or light chain candidate variable domains (V) fused to a CH1 or CK constant region, a second variable domain (and optionally third variable domain, wherein the second and third variable domain form a first antigen binding domain), and an Fc domain or portion thereof interposed between the candidate and second variable domains);

(ii) producing a second nucleic acid encoding a second polypeptide chain comprising the other of the heavy or light chain candidate variable domains (V) fused at its C-terminus to a CH1 or CK constant region selected to be complementary to the CH1 or CK constant region of the first polypeptide chain such that the first and second polypeptides form a CH1-CK heterodimer in which the heavy and light chain candidate variable domains form a second antigen binding domain, and an Fc domain or portion thereof; and

(iii) expressing said nucleic acids encoding the first and second polypeptide chains in a host cell to produce a protein comprising said first and second polypeptide chains, respectively; and recovering a heterodimeric protein comprising a dimeric Fc domain capable of binding human CD16; and

c) evaluating the plurality of heterodimeric proteins produced for a biological activity of interest, e.g., an activity disclosed herein. In this method, one of the first or second antigen binding domains binds NKp46 and the other binds an antigen of interest. In one embodiment, the second variable domain (optionally together with the third variable domain) of the first polypeptide chain binds NKp46. Optionally, the heterodimeric protein produced represents at least 20%, 25% or 30% of the total proteins prior to purification. Optionally the recovering step in (iii) comprises loading the protein produced onto an affinity purification support, optionally an affinity exchange column, optionally a Protein-A support or column, and collecting the heterodimeric protein; and/or loading the protein produced (or the protein

collected following loading onto a affinity exchange or Protein A column) onto an ion exchange column; and collecting the heterodimeric fraction.

In one embodiment, the invention provides methods of making a heterotrimeric protein (e.g. any heterotrimeric protein described herein), comprising:

5 (a) providing a first nucleic acid encoding a first polypeptide chain described herein (e.g., a polypeptide chain comprising a first variable domain (V) fused to a first CH1 or CK constant region, a second variable domain fused to a second CH1 or CK constant region, and an Fc domain or portion thereof interposed between the first and second (V-CH1/CK) units);

10 (b) providing a second nucleic acid encoding a second polypeptide chain described herein (e.g., a polypeptide chain comprising a variable domain (V) fused at its C-terminus to a CH1 or CK constant region selected to be complementary to the first CH1 or CK constant region of the first polypeptide chain such that the first and second polypeptides form a CH1-CK heterodimer in which the first variable domain of the first polypeptide chain and the variable domain of the second polypeptide form an antigen binding domain, and an Fc domain or portion thereof);

15 (c) providing a third nucleic acid comprising a third polypeptide chain described herein (e.g., a polypeptide chain comprising a variable domain fused at its C-terminus to a CH1 or CK constant region, wherein the CH1 or CK constant region is selected to be complementary to the second variable domain and second CH1 or CK constant region of the first polypeptide chain such that the first and third polypeptides form a CH1-CK heterodimer in which the second variable domain of the first polypeptide and the variable domain of the third polypeptide form an antigen binding domain; and

20 (d) expressing said first, second and third nucleic acids in a host cell to produce a protein comprising said first, second and third polypeptide chains, respectively; and recovering a heterotrimeric protein comprising a dimeric Fc domain capable of binding human CD16. Optionally, the heterotrimeric protein produced represents at least 20%, 25% or 30% of the total proteins prior to purification. Optionally step (d) comprises loading the protein produced onto an affinity purification support, optionally an affinity exchange column, optionally a Protein-A support or column, and collecting the heterotrimeric protein; and/or loading the protein produced (e.g., the protein collected following loading onto an affinity exchange or Protein A column) onto an ion exchange column; and collecting the heterotrimeric fraction. In one embodiment, one of the antigen binding domains binds NKp46 and the other binds an antigen of interest. In one embodiment, the second or the third polypeptide further comprises an Fc domain or fragment thereof fused to the C-terminus of the CH1 or CK domain (e.g. via a hinge domain or linker).

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In one aspect, the present disclosure provides a method for identifying or evaluating candidate variable regions for use in a heterotrimeric protein, comprising the steps of:

a) providing a plurality of nucleic acid pairs, wherein each pair includes one nucleic acid encoding a heavy chain candidate variable region and one nucleic acid encoding a light chain candidate variable region, for each of a plurality of heavy and light chain variable region pairs (e.g., obtained from different antibodies binding the same or different antigen(s) of interest);

b) for each of the plurality nucleic acid pairs, making a heterotrimeric protein by:

(i) producing a first nucleic acid encoding a first polypeptide chain comprising one of the heavy or light chain candidate variable domains (V) fused to a first CH1 or CK constant region, a second variable domain fused to a second CH1 or CK constant region, and an Fc domain or portion thereof interposed between the first and second (V-CH1/CK) units);

(ii) producing a second nucleic acid encoding a second polypeptide chain comprising the other of the heavy or light chain candidate variable domains (V) fused at its C-terminus to a CH1 or CK constant region selected to be complementary to the first CH1 or CK constant region of the first polypeptide chain such that the first and second polypeptides form a CH1-CK heterodimer in which the heavy and light chain candidate variable domains form an antigen binding domain, and an Fc domain or portion thereof;

(iii) producing a third nucleic acid encoding a third polypeptide chain comprising a variable domain fused at its C-terminus to a CH1 or CK constant region, wherein the CH1 or CK constant region is selected to be complementary to the second variable domain and second CH1 or CK constant region of the first polypeptide chain such that the first and third polypeptides form a CH1-CK heterodimer in which the second variable domain of the first polypeptide and the variable domain of the third polypeptide form an antigen binding domain; and

(iv) expressing said nucleic acids encoding the first and second polypeptide chains in a host cell to produce said first and second polypeptide chains, respectively; and recovering a heterodimeric protein comprising a dimeric Fc domain capable of binding human CD16; and

c) evaluating the plurality of heterodimeric proteins produced for a biological activity of interest, e.g., an activity disclosed herein. In one embodiment, the second or the third polypeptide further comprises and Fc domain or fragment thereof fused to the C-terminus of the CH1 or CK domain (e.g. via a hinge domain or linker). Optionally, the heterotrimeric protein produced represents at least 20%, 25% or 30% of the total proteins prior to purification. Optionally the recovering step in (iii) loading the protein produced onto an affinity purification support, optionally an affinity exchange column, optionally a Protein-A support or

column, and collecting the heterotrimeric protein; and/or loading the protein produced (e.g., the protein collected following loading onto an affinity exchange or Protein A column) onto an ion exchange column; and collecting the heterotrimeric fraction.

In the methods for identifying or evaluating candidate variable regions, it will be appreciated that the candidate variable regions can be for example from an anti-activating receptor antibody or from an antigen that binds an antigen of interest. It will also be appreciated that the position of the respective ABDs for the candidate variable region pair and the other variable region pair can be inverted. For example, in a trimeric protein the methods can be modified such that the heavy and light chain candidate variable domains are formed by the second V region of the first polypeptide and the V region of the second polypeptide, and the other variable region pair are formed by the first V region of the first polypeptide and the V region of the third polypeptide.

In one embodiment, the second variable domain of the first polypeptide and the variable domain of the third polypeptide form an antigen binding domain that binds an activating receptor on an immune cell.

Furthermore, by providing a panel of different multispecific protein formats that all can be produced in standard cell lines and standardized methods with high yields, yet have different properties (e.g. conformational flexibility, spacing between two antigen binding domains, etc.) that can affect functional activity of the protein, the protein formats of the disclosure (e.g. any two or more of F1, F2, F3, F4, F5, F6, F7, F8, F9, F10, F11, T5, T6, T9, T11, and derivatives) can be used in a panel to screen proteins configurations or formats to identify the most effective configurations or formats for a given antigen of interest, or combination of first and second antigen of interest. Different protein formats may access or engage their antigen targets differently.

In one aspect, the present disclosure provides a method for identifying or evaluating candidate protein configurations for use in a heterodimeric protein, comprising the steps of: producing, separately (e.g. in separate containers), a plurality of multispecific proteins of the disclosure, wherein the proteins differ in their domain arrangements, and evaluating the plurality of multispecific proteins produced for a biological activity of interest, e.g., an activity disclosed herein. In one embodiment, the proteins having different domain arrangements share antigen binding domains (e.g. the same CDRs or variable domains) for an antigen of interest. In one embodiment 1, 2, 3, 4, 5, 6, 7 or more different proteins are produced and evaluated. In one embodiment, one or more of (or all of) the proteins are selected from the group of proteins having a domain arrangement disclosed herein, e.g. that of formats F5 or T5. In one embodiment the proteins are produced according to the methods disclosed herein.

In one aspect, the present disclosure provides a library of at least 5, 10, 20, 30, 50 hetero-multimeric proteins of the disclosure, wherein the proteins share domain arrangements but differ in the amino acid sequence of the variable domain of one or both of their antigen binding domains.

5 In one aspect, the present disclosure provides a library of at least 2, 3, 4, 5 or 10 hetero-multimeric proteins of the disclosure, wherein the proteins share the amino acid sequence of the variable domain of one or both of their antigen binding domains, but differ in domain arrangements.

10 In one aspect of the any of the embodiments herein, recovering a heterodimeric or heterotrimer protein can comprise introducing the protein to a solid phase so as to immobilize the protein. The immobilized protein can then subsequently be eluted. Generally, the solid support may be any suitable insoluble, functionalized material to which the proteins can be reversibly attached, either directly or indirectly, allowing them to be separated from unwanted materials, for example, excess reagents, contaminants, and solvents. Examples of  
15 solid supports include, for example, functionalized polymeric materials, e.g., agarose, or its bead form Sepharose®, dextran, polystyrene and polypropylene, or mixtures thereof; compact discs comprising microfluidic channel structures; protein array chips; pipet tips; membranes, e.g., nitrocellulose or PVDF membranes; and microparticles, e.g., paramagnetic or non-paramagnetic beads. In some embodiments, an affinity medium will be  
20 bound to the solid support and the protein will be indirectly attached to solid support via the affinity medium. In one aspect, the solid support comprises a protein A affinity medium or protein G affinity medium. A "protein A affinity medium" and a "protein G affinity medium" each refer to a solid phase onto which is bound a natural or synthetic protein comprising an Fc-binding domain of protein A or protein G, respectively, or a mutated variant or fragment of  
25 an Fc-binding domain of protein A or protein G, respectively, which variant or fragment retains the affinity for an Fc-portion of an antibody. Protein A and Protein G are bacterial cell wall proteins that have binding sites for the Fc portion of mammalian IgG. The capacity of these proteins for IgG varies with the species. In general, IgGs have a higher affinity for Protein G than for Protein A, and Protein G can bind IgG from a wider variety of species. The  
30 affinity of various IgG subclasses, especially from mouse and human, for Protein A varies more than for Protein G. Protein A can, therefore, be used to prepare isotypically pure IgG from some species. When covalently attached to a solid matrix, such as cross-linked agarose, these proteins can be used to capture and purify antigen-protein complexes from biochemical solutions. Commercially available products include, e.g., Protein G, A or L  
35 bonded to agarose or sepharose beads, for example EZview™ Red Protein G Affinity Gel is Protein G covalently bonded to 4% Agarose beads (Sigma Aldrich Co); or POROS® A, G,



and CaptureSelect® HPLC columns (Invitrogen Inc.). Affinity capture reagents are also described, for example, in the Antibody Purification Handbook, Biosciences, publication No. 18-1037-46, Edition AC, the disclosure of which is hereby incorporated by reference).

Once the multispecific protein is produced it can be assessed for biological activity. In one aspect of any embodiment herein, where a protein binds an antigen on a target cell to be eliminated and an activating receptor on an effector cell, a multispecific protein is capable of inducing activation of an immune effector cell (e.g. an NK cell, a T cell) when the protein is incubated in the presence of the effector cell and a target cell that expresses the antigen of interest). In one aspect of any embodiment herein, a multispecific protein is capable of inducing signaling at an immune effector cell activating receptor when the protein is incubated in the presence of the effector cell and a target cell that expresses the antigen of interest). Optionally, effector cell activation or signaling is characterized by increased expression of a cell surface marker of activation, e.g. CD107, CD69, etc. Activity can be measured for example by bringing target cells and effector cells into contact with one another, in presence of the multispecific polypeptide. In one example, aggregation of target cells and effector cells is measured. In another example, the multispecific protein may, for example, be assessed for the ability to cause a measurable increase in any property or activity known in the art as associated with NK cell activity, respectively, such as marker of cytotoxicity (CD107) or cytokine production (for example IFN- $\gamma$  or TNF- $\alpha$ ), increases in intracellular free calcium levels, the ability to lyse target cells in a redirected killing assay, etc. In one embodiment of any of the methods of identifying, evaluating or making a protein, the method comprises a step of evaluating the multispecific protein for its ability to induce or increase the activity immune cells that express an activating receptor bound by an ABD multispecific protein (e.g. a marker of activation or cytotoxicity, cytokine production, ability to lyse a target cell, etc.), when incubated in the presence of the immune cells and target cells expressing an antigen of interest bound by an ABD of the multispecific protein (e.g. the cancer antigen). In one embodiment, the immune cells express NKp46, CD16 and/or CD137. In one embodiment, the cells are NKp46<sup>+</sup> NK cells. In one embodiment, the immune cells are CD16<sup>+</sup> cells. In one embodiment, the immune cells are CD137<sup>+</sup> cells.

In the presence of target cells (target cells expressing the antigen of interest) and effector cells that express the activating receptor bound by the protein, the multispecific protein will be capable of causing an increase in a property or activity associated with effector (e.g. NK cell, T cell) cell activity (e.g. activation of NK cell cytotoxicity, CD107 expression, IFN $\gamma$  production) *in vitro*. For example, a multispecific protein of the disclosure can be selected for the ability to increase an NK or T cell activity by more than about 20%,

preferably with at least about 30%, at least about 40%, at least about 50%, or more compared to that achieved with the same effector: target cell ratio with the same NK or T cells and target cells that are not brought into contact with the multispecific protein, as measured by an assay of NK or T cell activity, e.g., a marker of activation of NK cell cytotoxicity, CD107 or CD69 expression, IFN $\gamma$  production, a classical *in vitro* chromium release test of cytotoxicity. Examples of protocols for activation and cytotoxicity assays are described in the Examples herein, as well as for example, in Pessino et al, J. Exp. Med, 1998, 188 (5): 953-960; Sivori et al, Eur J Immunol, 1999. 29:1656-1666; Brando et al, (2005) J. Leukoc. Biol. 78:359-371; El-Sherbiny et al, (2007) Cancer Research 67(18):8444-9; and Nolte-'t Hoen et al, (2007) Blood 109:670-673).

In one aspect of the any of the embodiments herein, evaluating heterodimeric or heterotrimeric proteins for a characteristic of interest comprises evaluating the proteins for one or more properties selected from the group consisting of: binding to an antigen of interest, binding to an antigen on an immune effector cell, binding to a tumor, viral or bacterial antigen, binding to an FcRn receptor, binding to human CD16 and/or other Fc-domain mediated effector function(s), agonistic or antagonistic activity at a polypeptide to which the multimeric proteins binds, ability to modulate the activity (e.g. cause the death of) a cell expressing the antigen of interest, ability to direct a lymphocyte to a cell expressing the antigen of interest, ability to activate a lymphocyte in the presence and/or absence of a cell expressing the antigen of interest, NK cell activation, stability or half-life *in vitro* or *in vivo*, production yield, purity within a composition, and susceptibility to aggregate in solution.

In one aspect, the present disclosure provides a method for identifying or evaluating a protein, comprising the steps of:

(a) providing nucleic acid(s) encoding a protein described herein ;

(b) expressing said nucleic acid(s) in a host cell to produce said protein, respectively; and recovering said protein; and

(c) evaluating the protein produced for a biological activity of interest, e.g., an activity disclosed herein, the ability to mediate the lysis of target cells (that express antigen of interest). In one embodiment, a plurality of different multispecific proteins are produced and evaluated.

In one embodiment, the step (c) comprises:

(i) testing the ability of the protein to cause effector cells (e.g. NK cells, T cells) that express an activating receptor bound by the protein to mediate the lysis of target cells, when incubated with such effector cells in the presence of target cells (that express antigen of

interest). Optionally, step (i) is followed by a step comprising: selecting a protein (e.g., for further development, for use as a medicament) that mediates the lysis of target cells.

In one aspect of any embodiment herein, a multispecific protein described herein that comprises an ABD that binds an activating receptor on an immune effector cell can for example be characterized by:

(a) being capable of inducing effector cells (e.g. T cell; NK cells) that express the activating receptor bound by the ABD of the multispecific protein to lyse target cells, when incubated in the presence of the effector cells and target cells; and

(b) lack of agonist activity at the activating receptor bound by the ABD when incubated with activating receptor-expressing effector cells in the absence of target cell; where the multispecific protein is capable of binding to CD16, the effector cells are CD16-negative cells, e.g. CD16<sup>-</sup> NK cells. Optionally, the effector cells are purified effector cells.

### Uses of compounds

In one aspect, provided is the use of any of the compounds defined herein, particularly the inventive multispecific proteins or antibodies and/or cells which express same for the manufacture of a pharmaceutical preparation for the treatment, prevention or diagnosis of a disease in a mammal in need thereof. Provided also are the use any of the compounds defined above as a medicament or an active component or active substance in a medicament. In a further aspect the invention provides methods for preparing a pharmaceutical composition containing a compound as defined herein, to provide a solid or a liquid formulation for administration orally, topically, or by injection. Such a method or process at least comprises the step of mixing the compound with a pharmaceutically acceptable carrier.

In one aspect, provided is a method to treat, prevent or more generally affect a predefined condition in an individual or to detect a certain condition by using or administering a multispecific protein described herein, or a (pharmaceutical) composition comprising same.

The polypeptides described herein can be used to prevent or treat disorders that can be treated with antibodies, such as cancers, solid and non-solid tumors, hematological malignancies, infections such as viral or microbial/bacterial infections, and inflammatory or autoimmune disorders.

In one embodiment, the antigen of interest expressed on the surface of a malignant cell of a type cancer selected from the group consisting of: carcinoma, including that of the bladder, head and neck, breast, colon, kidney, liver, lung, ovary, prostate, pancreas, stomach, cervix, thyroid and skin, including squamous cell carcinoma; hematopoietic tumors of lymphoid lineage, including leukemia, acute lymphocytic leukemia,

acute lymphoblastic leukemia, B-cell lymphoma, T-cell lymphoma, Hodgkin's lymphoma, non-Hodgkin's lymphoma, hairy cell lymphoma and Burkett's lymphoma; hematopoietic tumors of myeloid lineage, including acute and chronic myelogenous leukemias and promyelocytic leukemia; tumors of mesenchymal origin, including fibrosarcoma and rhabdomyosarcoma; other tumors, including neuroblastoma and glioma; tumors of the central and peripheral nervous system, including astrocytoma, neuroblastoma, glioma, and schwannomas; tumors of mesenchymal origin, including fibrosarcoma, rhabdomyosarcoma, and osteosarcoma; and other tumors, including melanoma, xeroderma pigmentosum, keratoacanthoma, seminoma, thyroid follicular cancer and teratocarcinoma, hematopoietic tumors of lymphoid lineage, for example T-cell and B-cell tumors, including but not limited to T-cell disorders such as T-prolymphocytic leukemia (T-PLL), including of the small cell and cerebriform cell type; large granular lymphocyte leukemia (LGL) preferably of the T-cell type; Sézary syndrome (SS); Adult T-cell leukemia lymphoma (ATLL); a/d T-NHL hepatosplenic lymphoma; peripheral/post-thymic T cell lymphoma (pleomorphic and immunoblastic subtypes); angio immunoblastic T-cell lymphoma; angiocentric (nasal) T-cell lymphoma; anaplastic (Ki 1+) large cell lymphoma; intestinal T-cell lymphoma; T-lymphoblastic; and lymphoma/leukemia (T-Lbly/T-ALL).

In one embodiment, the inventive multispecific polypeptides described herein can be used to prevent or treat a cancer selected from the group consisting of: carcinoma, including that of the bladder, head and neck, breast, colon, kidney, liver, lung, ovary, prostate, pancreas, stomach, cervix, thyroid and skin, including squamous cell carcinoma; hematopoietic tumors of lymphoid lineage, including leukemia, acute lymphocytic leukemia, acute lymphoblastic leukemia, B-cell lymphoma, T-cell lymphoma, Hodgkin's lymphoma, non-Hodgkin's lymphoma, hairy cell lymphoma and Burkett's lymphoma; hematopoietic tumors of myeloid lineage, including acute and chronic myelogenous leukemias and promyelocytic leukemia; tumors of mesenchymal origin, including fibrosarcoma and rhabdomyosarcoma; other tumors, including neuroblastoma and glioma; tumors of the central and peripheral nervous system, including astrocytoma, neuroblastoma, glioma, and schwannomas; tumors of mesenchymal origin, including fibrosarcoma, rhabdomyosarcoma, and osteosarcoma; and other tumors, including melanoma, xeroderma pigmentosum, keratoacanthoma, seminoma, thyroid follicular cancer and teratocarcinoma. Other exemplary disorders that can be treated according to the invention include hematopoietic tumors of lymphoid lineage, for example T-cell and B-cell tumors, including but not limited to T-cell disorders such as T-prolymphocytic leukemia (T-PLL), including of the small cell and cerebriform cell type; large granular lymphocyte leukemia (LGL) preferably of the T-cell type; Sézary syndrome (SS); Adult T-cell leukemia lymphoma (ATLL); a/d T-NHL hepatosplenic

lymphoma; peripheral/post-thymic T cell lymphoma (pleomorphic and immunoblastic subtypes); angio immunoblastic T-cell lymphoma; angiocentric (nasal) T-cell lymphoma; anaplastic (Ki 1+) large cell lymphoma; intestinal T-cell lymphoma; T-lymphoblastic; and lymphoma/leukaemia (T-Lbly/T-ALL).

5 In another aspect, the invention provides a method of restoring or potentiating the activity of immune effector cells (e.g. NK cells or T cells) in a patient in need thereof (e.g. a patient having a cancer, or a viral, parasite or bacterial infection), comprising the step of administering to the patient the multispecific protein. In one embodiment, this method is directed at increasing the activity of lymphocytes expressing the activating receptor bound  
10 by the multispecific protein.

In another embodiment the subject multispecific proteins may be used or administered in combination with immune cells, particularly NK cells, derived from a patient who is to be treated or from a different donor, and these NK cells administered to a patient in need thereof such as a patient having a disease in which increased lymphocyte (e.g. NK  
15 cell) activity is beneficial or which is caused or characterized by insufficient NK cell activity, such as a cancer, or a viral or microbial, e.g., bacterial or parasite infection. As NK cells (unlike CAR-T cells) do not express TCRs, these NK cells, even those derived from different donors will not induce a GVHD reaction. (See e.g., Glienke et al., "Advantages and applications of CAR-expressing natural killer cells", *Front. Pharmacol.* 6, Art. 21:1-6 (2015);  
20 Hermanson and Kaufman, *Front. Immunol.* 6, Art. 195:1-6 (2015))

In one aspect, the methods of treatment comprise administering to an individual a multispecific protein of the disclosure in a therapeutically effective amount. A therapeutically effective amount may be any amount that has a therapeutic effect in a patient having a disease or disorder (or promotes, enhances, and/or induces such an effect in at least a  
25 substantial proportion of patients with the disease or disorder and substantially similar characteristics as the patient).

The multispecific proteins of the disclosure can be included in kits. The kits may optionally further contain any number of polypeptides and/or other compounds, e.g., 1, 2, 3, 4, or any other number of multispecific proteins and/or other compounds. It will be  
30 appreciated that this description of the contents of the kits is not limiting in any way. For example, the kit may contain other types of therapeutic compounds. Optionally, the kits also include instructions for using the polypeptides, e.g., detailing the herein-described methods.

The invention also provides pharmaceutical compositions comprising the subject multispecific proteins and optionally other compounds as defined above. A multispecific  
35 protein and optionally another compound may be administered in purified form together with a pharmaceutical carrier as a pharmaceutical composition. The form depends on the

intended mode of administration and therapeutic or diagnostic application. The pharmaceutical carrier can be any compatible, nontoxic substance suitable to deliver the compounds to the patient. Pharmaceutically acceptable carriers are well known in the art and include, for example, aqueous solutions such as (sterile) water or physiologically buffered saline or other solvents or vehicles such as glycols, glycerol, oils such as olive oil or injectable organic esters, alcohol, fats, waxes, and inert solids. A pharmaceutically acceptable carrier may further contain physiologically acceptable compounds that act for example to stabilize or to increase the absorption of the compounds. Such physiologically acceptable compounds include, for example, carbohydrates, such as glucose, sucrose or dextrans, antioxidants, such as ascorbic acid or glutathione, chelating agents, low molecular weight proteins or other stabilizers or excipients. One skilled in the art would know that the choice of a pharmaceutically acceptable carrier, including a physiologically acceptable compound, depends, for example, on the route of administration of the composition. Pharmaceutically acceptable adjuvants, buffering agents, dispersing agents, and the like, may also be incorporated into the pharmaceutical compositions. Non-limiting examples of such adjuvants include by way of example inorganic and organic adjuvants such as alum, aluminum phosphate and aluminum hydroxide, squalene, liposomes, lipopolysaccharides, double stranded (ds) RNAs, single stranded (s-s) DNAs, and TLR agonists such as unmethylated CpG's.

Multispecific proteins according to the invention can be administered parenterally. Preparations of the compounds for parenteral administration must be sterile. Sterilization is readily accomplished by filtration through sterile filtration membranes, optionally prior to or following lyophilization and reconstitution. The parenteral route for administration of compounds is in accord with known methods, e.g. injection or infusion by intravenous, intraperitoneal, intramuscular, intraarterial, or intralesional routes. The compounds may be administered continuously by infusion or by bolus injection. A typical composition for intravenous infusion could be made up to contain 100 to 500 ml of sterile 0.9% NaCl or 5% glucose optionally supplemented with a 20% albumin solution and 1 mg to 10 g of the compound, depending on the particular type of compound and its required dosing regimen. Methods for preparing parenterally administrable compositions are well known in the art.

## EXAMPLES

### Example 1

#### Generation of anti-huNKp46 antibodies

Balb/c mice were immunized with a recombinant human NKp46 extracellular domain recombinant-Fc protein. Mice received one primo-immunization with an emulsion of 50 µg NKp46 protein and Complete Freund Adjuvant, intraperitoneally, a 2nd immunization with an emulsion of 50 µg NKp46 protein and Incomplete Freund Adjuvant, intraperitoneally, and finally a boost with 10 µg NKp46 protein, intravenously. Immune spleen cells were fused 3 days after the boost with X63.Ag8.653 immortalized B cells, and cultured in the presence of irradiated spleen cells.

Primary screen: Supernatant (SN) of growing clones were tested in a primary screen by flow cytometry using a cell line expressing the human NKp46 construct at the cell surface. Briefly, for FACS screening, the presence of reactive antibodies in supernatants was revealed by Goat anti-mouse polyclonal antibody (pAb) labeled with PE.

A panel of antibodies that bound NKp46 was selected, produced and their variable regions sequenced and these antibodies and derivatives thereof further evaluated for their activity in the context of a bispecific molecule.

### **Example 2:**

#### **Identification of a bispecific antibody format that binds FcRn but not FcγR for targeting effector cell receptors**

Experiments were conducted with the objective being the development of a new bispecific protein format that places an Fc domain on a polypeptide together with an anti-NKp46 binding domain and an anti-target antigen binding domain. Such bispecific proteins should bind to NKp46 monovalently via its anti-NKp46 binding domain. The monomeric Fc domain should retain at least partial binding to the human neonatal Fc receptor (FcRn), yet not substantially bind human CD16 and/or other human Fcγ receptors. Consequently, such bispecific proteins should not induce Fcγ-mediated (e.g. CD16-mediated) target cell lysis.

#### *Example 2-1 Construction and binding analysis of Anti-CD19-IgG1-Fcmono-Anti-CD3*

Since no anti-NKp46 bispecific antibody has been produced that could indicate whether such a protein could be functional, CD3 was used as a model antigen in place of NKp46 in order to investigate the possible functionality of a new monovalent bispecific protein format prior to targeting NK cells via NKp46.

A bispecific Fc-based on a scFv specific for tumor antigen CD19 (anti-CD19 scFv) and a scFv specific for activating receptor CD3 on a T cell (anti-CD3 scFv) was used to assess FcRn binding and CD19-binding functions of a new monomeric bispecific polypeptide format. The domain arrangement of the final polypeptide is referred to as the "F1" format

(the star in the CH2 domain indicates an optional N297S mutation, not included in the polypeptide tested here). (See **Figure 2**)

A bispecific monomeric Fc-containing polypeptide was constructed based on a scFv specific for the tumor antigen CD19 (anti-CD19 scFv) and a scFv specific for an activating receptor CD3 on a T cell (anti-CD3 scFv). The CH3 domain incorporated the mutations (EU numbering) L351K, T366S, P395V, F405R, T407A and K409Y. The polypeptide has domains arranged as follows: anti-CD19-CH2-CH3-anti-CD3. A DNA sequence coding for a CH3/VH linker peptide having the amino acid sequence STGS was also designed in order to insert a specific Sall restriction site at the CH3-VH junction.

This CH3 domain incorporated the mutations (EU numbering) L351K, T366S, P395V, F405R, T407A and K409Y. The selected CH2 domain was a wild-type CH2. DNA and amino acid sequences for the monomeric CH2-CH3 Fc portion and the anti-CD19 are shown below. The light chain and heavy chain DNA and amino acid sequences for the anti-CD19 scFv were as follows:

#### Anti-CD19-VK

GACATTCAGCTGACCCAATCTCCAGCTTCTTTGGCTGTGTCTCTAGGGCAGAGGGCCACCATCTCCTG  
CAAGGCCAGCCAAAGTGTTGATTATGATGGTGATAGTTATTTGAACTGGTACCAACAGATACCAGGAC  
AGCCACCCAAACTCCTCATCTATGATGCATCCAATCTAGTATCTGGGATTCCACCCAGGTTTAGTGGC  
AGTGGGTCTGGGACAGACTTCACCCTCAACATCCATCCTGTGGAGAAGGTGGATGCTGCAACCTATCA  
CTGTCAGCAAAGTACTGAGGACCCTTGGACGTTTCGGTGGAGGCACCAAGCTGGAAATCAAA (SEQ ID  
NO: 3)

#### Anti-CD19-VK

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGKLEIK (SEQ ID NO: 4)

#### Anti-CD19-VH

CAGGTTTCAGCTGCAGCAGTCTGGGGCTGAGCTGGTGCGGCCTGGGTCCTCAGTGAAGATTTCTGCAA  
AGCATCTGGCTACGCATTTCAGTAGCTACTGGATGAACTGGGTGAAGCAGAGGCCTGGACAGGGTCTTG  
AGTGGATTGGACAGATTTGGCCTGGAGATGGTGATACTAATAACGGAAGTTCAAGGGCAAGGCC  
ACACTGACTGCAGACGAATCCTCCAGCACAGCCTACATGCAGCTCAGCAGCCTGGCCTCTGAGGACTC  
TGCGGTCTATTTCTGTGCAAGACGAGAAACGACCACTGTCGGGCGTTATTACTATGCTATGGACTACT  
GGGGTCAAGGAACCACAGTCACCGTCTCCTCA (SEQ ID NO: 5)

#### Anti-CD19-VH



QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDNTNYNGKFKGKA  
TLTADSSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSS

(SEQ ID NO: 6)

5 The DNA and amino acid sequences for the monomeric CH2-CH3 Fc portion and final bispecific polypeptide were as follows:

IgG1-Fcmono (the last K was removed in that construct)

GCACCTGAACTCCTGGGGGGACCGTCAGTCTTCCTCTTCCCCCAAACCCAAGGACACCCTCATGAT  
CTCCCGGACCCCTGAGGTCACATGCGTGTTGGTGGACGTGAGCCACGAAGACCCTGAGGTCAAGTTCA  
10 ACTGGTACGTGGACGGCGTGAGGTGCATAATGCCAAGACAAAGCCGCGGGAGGAGCAGTACAACAGC  
ACGTACCGTGTGGTCAGCGTCCTCACCGTCTGCACCAGGACTGGCTGAATGGCAAGGAGTACAAGTG  
CAAGGTCTCCAACAAAGCCCTCCCAGCCCCCATCGAGAAAACCATCTCCAAAGCCAAAGGGCAGCCCC  
GAGAACCACAGGTGTACACCAAGCCCCCATCCCGGGAGGAGATGACCAAGAACCAGGTCAGCCTGTCC  
TGCTTGGTCAAAGGCTTCTATCCCAGCGACATCGCCGTGGAGTGGGAGAGCAATGGGCAGCCGGAGAA  
15 CAACTACAAGACCACGGTTCCCGTGCTGGACTCCGACGGCTCCTTCCGCCTCGCTAGCTACCTACCG  
TGGACAAGAGCAGGTGGCAGCAGGGGAACGTCTTCTCATGCTCCGTGATGCATGAGGCTCTGCACAAC  
CACTACACGCAGAAGAGCCTCTCCCTGTCCCCGGGG (SEQ ID NO : 7)

IgG1-Fcmono\* (\*the last K residue was removed in that construct)

20 APELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNS  
TYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTKPPSREEMTKNQVSL  
CLVKGFYPSDIAVEWESNGQPENNYKTTVPVLDSDGSFRLASYLTVDKSRWQQGNVFSCSVMHEALHN  
HYTQKSLSLSPG (SEQ ID NO: 8)

25 Anti-CD19-F1-Anti-CD3 Complete sequence (mature protein)

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGSGGGGSQVQLQQSGAE  
LVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDNTNYNGKFKGKATLTADSSST  
AYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSGGGSSAPELLGGPSVFLFPPKP  
30 KDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN  
GKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTKPPSREEMTKNQVSLCLVKGFYPSDIAVEWES  
NGQPENNYKTTVPVLDSDGSFRLASYLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGSTGSD  
IKLQQSGAELARPGASVKMSCKTSGYTFTRYTMHWVKQRPGQGLEWIGYINPSRGYTNYNQKFKDKAT  
LTTDKSSSTAYMQLSSLTSEDSAVYYCARYYDDHYCLDYWGQGTTLTVSSVEGGSGGSGGSGGSGVD  
35 DIQLTQSPAIMASAPGEKVTMTCRASSSVSYMNWYQQKSGTSPKRWIYDTSKVASGVPYRFSGSGSGT  
SYSLTISSMEAEDAATYYCQQWSSNPLTFGAGTKLELK (SEQ ID NO: 9)

*Cloning and the production of the recombinant proteins*

Coding sequences were generated by direct synthesis and/or by PCR. PCR was performed using the PrimeSTAR MAX DNA polymerase (Takara, #R045A) and PCR products were purified from 1% agarose gel using the NucleoSpin gel and PCR clean-up kit (Macherey-Nagel, #740609.250). Once purified the PCR products were quantified prior to the In-Fusion ligation reaction which was performed as described in the manufacturer's protocol (ClonTech, #ST0345). The plasmids were obtained after a miniprep preparation run on an EVO200 (Tecan) using the Nucleospin 96 plasmid kit (Macherey-Nagel, #740625.4). Plasmids were then sequenced for sequence confirmation before to transfecting the CHO cell line.

CHO cells were grown in the CD-CHO medium (Invitrogen) complemented with phenol red and 6 mM GlutaMax. The day before the transfection, cells were counted and seeded at 175,000 cells/ml. For the transfection, cells (200.000 cells/transfection) were prepared as described in the AMAXA SF cell line kit (AMAXA, #V4XC-2032) and nucleofected using the DS137 protocol with the Nucleofector 4D device. All the transfections were performed using 300 ng of verified plasmids. After transfection, cells were seeded into 24 well plates in pre-warmed culture medium. After 24 hours, hygromycin B was added in the culture medium (200 µg/ml). Protein expression was monitored after one week in culture. Cells expressing the proteins were then sub-cloned to obtain the best producers. Sub-cloning was performed using 96 flat-bottom well plates in which the cells are seeded at one cell per well into 200 µl of culture medium complemented with 200 µg/ml of hygromycin B. Cells were left for three weeks before testing the clone's productivity.

Recombinant proteins which contain an IgG1-Fc fragment were purified using Protein-A beads (- rProteinA Sepharose fast flow, GE Healthcare). Briefly, cell culture supernatants were concentrated, clarified by centrifugation and injected onto Protein-A columns to capture the recombinant Fc containing proteins. Proteins were eluted at acidic pH (citric acid 0.1M pH 3), and the eluate immediately neutralized using TRIS-HCL pH 8.5 and dialyzed against 1X PBS. Recombinant scFvs which contain a "six his" tag were purified by affinity chromatography using Cobalt resin. Other recombinant scFvs were purified by size exclusion chromatography (SEC).

*Example 2-2: Binding analysis of Anti-CD19-IgG1-Fcmono-Anti-CD3 to B221, JURKAT, HUT78 and CHO cell lines*

Cells were harvested and stained with the cell supernatant of the anti-CD19-F1-anti-CD3 producing cells during 1 h at 4°C. After two washes in staining buffer (PBS1X / BSA

0.2% / EDTA 2mM), cells were stained for 30 min at 4°C with goat anti-human (Fc)-PE antibody (IM0550 Beckman Coulter - 1/200). After two washes, stainings were conducted on a BD FACS Canto II and analyzed using the FlowJo software.

CD3 and CD19 expression were also controlled by flow cytometry: Cells were harvested and stained in PBS1X / BSA 0.2% / EDTA 2mM buffer during 30 min at 4°C using 5µl of the anti-CD3-APC and 5µl of the anti-CD19-FITC antibodies. After two washes, stainings were conducted on a BD FACS Canto II and analyzed using the FlowJo software.

The results of these experiments revealed that the Anti-CD19-F1-Anti-CD3 protein binds to CD3 cell lines (HUT78 and JURKAT cell lines) and to the CD19 cell line (B221 cell line) but not to the CHO cell line which was used as a negative control.

#### *Example 2-3:*

##### *T- and B- cell aggregation by purified Anti-CD19-F1-Anti-CD3*

Purified Anti-CD19-F1-Anti-CD3 was tested in a T/B cell aggregation assay to evaluate whether the antibody promotes the aggregation of CD19 and CD3 expressing cells.

The results of this assay are shown in **Figure 1**. The top panel shows that Anti-CD19-F1-Anti-CD3 does not cause aggregation in the presence of B221 (CD19) or JURKAT (CD3) cell lines, but it does cause aggregation of cells when both B221 and JURKAT cells are co-incubated, indicating that the bispecific antibody is functional. The lower panel shows the results of the control experiment conducted without antibody.

#### *Example 2-4:*

##### *Binding of bispecific monomeric Fc polypeptide to FcRn*

##### *Affinity study by Surface Plasmon Resonance (SPR)*

###### *Biacore T100 general procedure and reagents*

SPR measurements were performed on a Biacore T100 apparatus (Biacore GE Healthcare) at 25°C. In all Biacore experiments Acetate Buffer (50 mM Acetate pH5.6, 150 mM NaCl, 0.1% surfactant p20) and HBS-EP+ (Biacore GE Healthcare) were used as the running buffer and regeneration buffer respectively. Sensorgrams were analyzed with Biacore T100 Evaluation software. Recombinant mouse FcRn was purchased from R&D Systems.

###### *Immobilization of FcRn*

Recombinant FcRn proteins were immobilized covalently to carboxyl groups in the dextran layer on a Sensor Chip CM5. The chip surface was activated with EDC/NHS (N-

ethyl-N'-(3-dimethylaminopropyl) carbodiimidehydrochloride and N-hydroxysuccinimide (Biacore GE Healthcare)). FcRn proteins were diluted to 10 µg/ml in coupling buffer (10 mM acetate, pH 5.6) and injected until the appropriate immobilization level was reached (i.e. 2500 RU). Deactivation of the remaining activated groups was performed using 100 mM ethanolamine pH 8 (Biacore GE Healthcare).

#### *Affinity study*

Monovalent affinity study was conducted following the Single Cycle Kinetic (SCK) protocol. Five serial dilutions of soluble analytes (antibodies and bi-specific molecules) ranging from 41.5 to 660 nM were injected over the FcRn (without regeneration) and allowed to dissociate for 10 min before regeneration. For each analyte, the entire sensorgram was fitted using the 1:1 SCK binding model.

#### *Results*

Anti-CD19-F1-Anti-CD3 having its CH2-CH3 domains placed between two antigen binding domains, particularly two scFvs, was evaluated to assess whether such bispecific monomeric Fc protein could retain binding to FcRn and possess an improved *in vivo* half-life compared to conventional bispecific antibodies. The results of these experiments showed that FcRn binding was retained, the model suggesting a 1:1 ratio (1 FcRn for each monomeric Fc) instead of a 2:1 ratio (2 FcRn for each antibody) for a regular or wild-type IgG.

The binding affinity of this multispecific protein was evaluated using SPR, and was compared to a chimeric full length antibody containing intact human IgG1 constant regions. The monomeric Fc retained significant monomeric binding to FcRn (monomeric Fc: affinity of  $K_D=194$  nM; full length antibody with bivalent binding: avidity of  $K_D=15.4$  nM).

### **Example 3:**

#### **Construction of multimeric bispecific proteins with monomeric-Fc**

Activating receptors on effector cells such as NK cells can contribute to the activation of the effector cell and/or lysis of target cells, yet conventional antibodies can block the activating receptor, exemplified by anti-NKp46 antibodies which as full length IgG1 block NKp46 signalling. We therefore investigated whether the bispecific protein format could induce NKp46 triggering, without inducing NKp46 agonism in the absence of target cells, which could lead to inappropriate NK activation distant from the target and/or decreased overall activity toward target cells.

A new bispecific protein format was developed as a single chain protein which binds to FcRn but not FcγR. Additionally, multimeric proteins that comprise two or three polypeptide chains, wherein the Fc domain remains monomeric, were developed that are

compatible for use with antibody variable regions that do not maintain binding to their target when converted to scFv format. The latter formats can be used conveniently for antibody screening; by incorporating at least one binding region as a F(ab) structure, any anti-target (e.g. anti-tumor) antibody variable region can be directly expressed in a bispecific construct as the F(ab) format within the bispecific protein and tested, irrespective of whether the antibody would retain binding as an scFv, thereby simplifying screening and enhancing the number of antibodies available. These formats in which the Fc domain remains monomeric have the advantage of maintaining maximum conformational flexibility and as shown *infra* may permit optimal binding to activating receptor (e.g. NKp46) or target antigens.

Different constructs were made for use in the preparation of bispecific antibodies using the variable domains from the scFv specific for tumor antigen CD19 described in Example 2-1, and different variable regions from antibodies specific for the NKp46 receptor identified in Example 1.

In order for the Fc domain to remain monomeric in single chain polypeptides or in multimers in which only one chain had an Fc domain, CH3-CH3 dimerization was prevented through two different strategies: (1) through the use of CH3 domain incorporating specific mutations (EU numbering), i.e., L351K, T366S, P395V, F405R, T407A and K409Y; or (2) through the use of a tandem CH3 domain in which the tandem CH3 domains are separated by a flexible linker associated with one another, which prevents interchain CH3-CH3 dimerization. The DNA and amino acid sequences for the monomeric CH2-CH3 Fc portion containing the above-identified point mutations were the same as in Example 2-1. The DNA and amino acid sequences for the monomeric CH2-CH3-linker-CH3 Fc portion with tandem CH3 domains are shown in **Figures 2A-2D**.

The light chain and heavy chain DNA and amino acid sequences for the anti-CD19 scFv were also the same as in Example 2-1. Proteins were cloned, produced and purified as in Example 2-1. Shown below are an exemplary light chain and heavy chain DNA and amino acid sequences for an anti-NKp46 scFv referred to as "NKp46-3".

scFv	anti-	scFv sequence (VHVK) / - stop
NKp46		
NKp46-3		STGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGI SPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDYW GQGTTTLTVSSVEGGSGGSGGSGGVDDIVMTQSPATLSVTPGDRVSLSCRASQ SISDYLHWYQQKSHESPRLLIKYASQSISGIPSRFSGSGSGSDFTLSINSVEPED VGVYYCQNGHSFPLTFGAGTKLELK- (SEQ ID NO: 10)

*Format 1 (F1) (Anti-CD19-IgG1-Fcmono-Anti-NKp46 (scFv))*

The domain structure of Format 1 (F1) is shown in **Figure 2A**. A bispecific Fc-containing polypeptide was constructed based on a scFv specific for the tumor antigen CD19 (anti-CD19 scFv) and an scFV specific for the NKp46 receptor. The polypeptide is a single chain polypeptide having domains arranged (N- to C- termini) as follows:

(V<sub>K</sub>-V<sub>H</sub>)<sup>anti-CD19</sup> – CH2 – CH3 – (V<sub>H</sub>-V<sub>K</sub>)<sup>anti-NKp46</sup>

A DNA sequence coding for a CH3/VH linker peptide having the amino acid sequence STGS was designed in order to insert a specific Sall restriction site at the CH3-VH junction. The domain arrangement of the final polypeptide is shown in Figure 2 (star in the CH2 domain indicates an optional N297S mutation). The (V<sub>K</sub>-V<sub>H</sub>) units include a linker between the V<sub>H</sub> and V<sub>K</sub> domains. Proteins were cloned, produced and purified as in Example 2-1. The amino acid sequence is shown as follows:

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSGSGSGTDF  
TLNIHPVEKVDAAATYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGSGGGGSGVQLQQSGAELVRPGSSVKISCKA  
SGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKATLTADESSSTAYMQLSSLASEDSAVYFCARR  
ETTTVGRYYYAMDYWGQGTITVTVSSGGGSSAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKF  
NWIYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQVY  
KPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTVPVLDSDGSFRLASLYLTVDKSRWQQGNV  
FSCVMHEALHNYHTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGG  
ISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDDYWGQGTITLVSSVEGGSGGSG  
SGGGSGGVDDIVMTQSPATLSVTPGDRVSLSCRASQISDYLHWYQQKSHESPRLLIKYASQISGIPSRFSGSG  
SGSDFTLTSLINSVEPEDVGVYYCQNGHSFPLTFGAGTKLELK-

#### Format 2 (F2) : CD19-F2-NKp46-3

The domain structure of F2 polypeptides is shown in **Figure 2A**. The DNA and amino acid sequences for the monomeric CH2-CH3 Fc portion were as in Example 2-1 and it similarly contains CH3 domain mutations (the mutations (EU numbering) L351K, T366S, P395V, F405R, T407A and K409Y. The heterodimer is made up of:

(1) a first (central) polypeptide chain having domains arranged as follows (N- to C- termini):

(V<sub>K</sub>-V<sub>H</sub>)<sup>anti-CD19</sup> – CH2 – CH3 – V<sub>H</sub><sup>anti-NKp46</sup> – CH1

and

(2) a second polypeptide chain having domains arranged as follows (N- to C- termini): V<sub>K</sub><sup>anti-NKp46</sup> – CK.

The (V<sub>K</sub>-V<sub>H</sub>) unit was made up of a V<sub>H</sub> domain, a linker and a V<sub>K</sub> unit (i.e. an scFv). As with other formats of the bispecific polypeptides, the DNA sequence coded for a CH3/VH linker peptide having the amino acid sequence STGS designed in order to insert a specific Sall restriction site at the CH3-VH junction. Proteins were cloned, produced and purified as in Example 2-1. The amino acid sequences for the CD19-F2-NKp46-3 Polypeptide chain 1 is shown in SEQ ID NO: 11 and CD19-F2-NKp46-3 Polypeptide chain 2 in SEQ ID NO: 12.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
 SSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGSGGGGSSQVQLQQSGAE  
 LVRPGSSVKISCKASGYAFSSYWMNWVKQRPQGQLEWIGQIWPGDGDTNYNGKFKGKATLTADESSST  
 5 AYMLSSLASSEDNAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSGGGSSAPELLGGPSVFLFPPKP  
 KDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLN  
 GKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTKPPSREEMTKNQVSLVCLVKGFYPSDIAVEWES  
 NGQPENNYKTTVPVLDSDGSFRLASVLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGSTGSE  
 VQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKAT  
 10 LTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDYWGQGTITLVSSASTKGPSVFPLAPSSKSTSGG  
 TAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKP  
 SNTKVDKRVPEPKCDKTH- (SEQ ID NO: 11)

DIVMTQSPATLSVTPGDRVSLSCRASQSSISDYLHWYQQKSHESPRLLIKYASQSSISGIPSRFSGSGSG  
 15 SDFTLSINSVEPEDVGYYCQNGHSFPLTFGAGTKLELKRRTVAAPSVFIFPPSDEQLKSGTASVVCLL  
 NNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSP  
 VTKSFNRGEC- (SEQ ID NO: 12)

### Format 8 (F8)

20 The domain structure of F8 polypeptides is shown in **Figure 2B**. The DNA and amino  
 acid sequences for the monomeric CH2-CH3 Fc portion were as in Format F2 and it similarly  
 contains CH3 domain mutations (the mutations (EU numbering) L351K, T366S, P395V,  
 F405R, T407A and K409Y, as well as a N297S mutation which prevents N-linked  
 glycosylation and moreover abolishes FcγR binding. Three variants of F8 proteins were  
 25 produced: (a) one wherein the cysteine residues in the hinge region were left intact (wild-  
 type, referred to as F8A), (b) a second wherein the cysteine residues in the hinge region  
 were replaced by serine residues (F8B), and (c) a third including a linker sequence GGGSS  
 replacing residues DKTHTCPPCP in the hinge (F8C). Variants F8B and F8C provided  
 production advantages as these versions avoided the formation of homodimers of the central  
 30 chain. This heterotrimer is made up of;

(1) a first (central) polypeptide chain having domains arranged as follows (N- to C-  
 termini):

$$V_H^{\text{anti-CD19}} - CH1 - CH2 - CH3 - V_H^{\text{anti-NKp46}} - C_K$$

and

35 (2) a second polypeptide chain having domains arranged as follows (N- to C-  
 termini):  $V_K^{\text{anti-NKp46}} - CH1$

and

(3) a third polypeptide chain having domains arranged as follows (N- to C- termini):

$$V_K^{\text{anti-CD19}} - C_K.$$

40 Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins  
 was purified from cell culture supernatant by affinity chromatography using prot-A beads and

analysed and purified by SEC. The protein showed a high production yield of 3.7 mg/L (F8C) and with a simple SEC profile. The amino acid sequences for the three F8 protein chains for the F8 "C" variant are shown in SEQ ID NOS 13, 14 and 15.

5 DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRGEC- (SEQ ID NO: 13)

10 QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQT  
YICNVNHKPSNTKVDKRVKPKSCGGSSAPELLGGPSVFLFPPKPKDTLMI SRTPEVTCVVVDVSHED  
PEVKFNWYVDGVEVHNATKPREEQY**S**STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISK  
15 AKGQPREPQVYTKPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTVPVLDSDGSFRL  
ASYLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKT  
SGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSA  
VYYCARRGGSFDYWGQGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVCLLNNFYPREAKVQWKVD  
NALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC-  
20 (SEQ ID NO: 14)

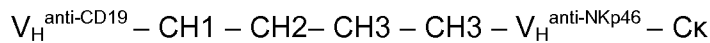
25 DIVMTQSPATLSVTPGDRVSLSCRASQSSISDYLHWYQQKSHESPRLLIKYASQSSISGIPSRFSGSGSG  
SDFTLSINSVEPEDVGVYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQTYICNVNHKPSNTKVDKR  
VEPKSCDKTH- (SEQ ID NO: 15)

#### *Format 9 (F9): CD19-F9-NKp46-3*

The F9 polypeptide is a trimeric polypeptide having a central polypeptide chain and  
30 two polypeptide chains each of which associate with the central chain via CH1-Ck  
dimerization. The domain structure of the trimeric F9 protein is shown in **Figure 2B**, wherein  
the bonds between the CH1 and Ck domains are interchain disulfide bonds. The two antigen  
binding domains have a F(ab) structure permitting the use of these antibodies irrespective of  
whether they remain functional in a scFv format. The DNA and amino acid sequences for the  
35 CH2-CH3 Fc portion comprise a tandem CH3 domain as in Format F4 and comprise a CH2  
domain comprising a N297S substitution. Three variants of F9 proteins were produced: (a) a  
first wherein the cysteine residues in the hinge region left intact (wild-type, referred to as  
F9A), (b) a second wherein the cysteine residues in the hinge region were replaced by  
serine residues (F9B), and (c) a third containing a linker sequence GGGSS which replaces  
40 residues DKTHTCPPCP in the hinge (F9C). Variants F9B and F9C provided advantages in  
production by avoiding the formation of homodimers of the central chain. The heterotrimer is  
made up of:



(1) a first (central) polypeptide chain having domains arranged as follows (N- to C-termini):

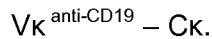


and

(2) a second polypeptide chain having domains arranged as follows (N- to C-termini):  $V_K^{\text{anti-NKp46}} - \text{CH1}$

and

(3) a third polypeptide chain having domains arranged as follows (N- to C-termini):



Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins was purified from cell culture supernatant by affinity chromatography using prot-A beads and analysed and purified by SEC. The protein showed a high production yield of 8.7 mg/L (F9A) and 3.0 mg/L (F9B), and with a simple SEC profile. The amino acid sequences for the three F9 protein chains for each of variants F9A, F9B and F9C are shown in the SEQ ID NOS listed in the table below.

Protein	SEQ ID NOS
F9A	16, 17, 18
F9B	19, 20, 21
F9C	22, 23, 24

#### F9A:

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRGEC- (SEQ ID NO: 16)

QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQT  
YICNVNHKPSNTKVDKRVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD  
VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDS  
GSFFLYSKLTVDKSRWQQGNVFCVMHEALHNHYTQKSLSLSPGSGGGGSGGGGSGGGGSGQPREPQV  
YTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSGSFFLYSKLTVDKSR  
WQQGNVFCVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTM  
HWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGS  
FDYWGQGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVCLLNNFYFPREAKVQWKVDNALQSGNSQ  
ESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC- (SEQ ID NO: 17)

DIVMTQSPATLSVTPGDRVSLSCRASQSSISDYLHWYQQKSHESPRLLIKYASQSSISGIPSRFSGSGSG  
SDFTLSINSVEPEDVGVYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV

KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKR  
VEPKSCDKTH- (SEQ ID NO: 18)

## F9B:

5 DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRGEC- (SEQ ID NO: 19)

10 QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQT  
YICNVNHKPSNTKVDRVEPKSCDKTHTSPSPAPPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDV  
15 VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD  
GSFFLYSKLTVDKSRWQQGNVVFSCSVMEALHNHYTQKSLSLSPGSGGGGSGGGGSGGGGSGQPREPQV  
YTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSR  
WQQGNVVFSCSVMEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTM  
20 HWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGS  
FDYWGQGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVCLLNNFYPREAKVQWKVDNALQSGNSQE  
SVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC- (SEQ ID NO: 20)

DIVMTQSPATLSVTPGDRVSLSCRASQSIDYLNWYQQKSHESPRLLIKYASQSIGIPSRFSGSGSG  
SDFTLSINSVEPEDVGYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
25 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDR  
VEPKSCDKTH- (SEQ ID NO: 21)

## F9C:

30 DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRGEC- (SEQ ID NO: 22)

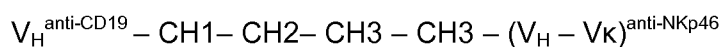
35 QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQT  
YICNVNHKPSNTKVDRVEPKSCGGGSSAPPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHED  
PEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISK  
40 AKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFL  
YSKLTVDKSRWQQGNVVFSCSVMEALHNHYTQKSLSLSPGSGGGGSGGGGSGGGGSGQPREPQVYTLPP  
SREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGN  
VFSCSVMEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQ  
SHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDYWG  
45 QGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQ  
DSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC- (SEQ ID NO: 23)

DIVMTQSPATLSVTPGDRVSLSCRASQSIDYLNWYQQKSHESPRLLIKYASQSIGIPSRFSGSGSG  
SDFTLSINSVEPEDVGYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
50 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDR  
VEPKSCDKTH- (SEQ ID NO: 24)

*Format 10 (F10): CD19-F10-NKp46-3*

The F10 polypeptide is a dimeric protein having a central polypeptide chain and a second polypeptide chain which associates with the central chain via CH1-Ck dimerization. The domain structure of the dimeric F10 protein is shown in **Figure 2B** wherein the bonds between the CH1 and Ck domains are interchain disulfide bonds. One of the two antigen binding domains has a Fab structure, and the other is a scFv. The DNA and amino acid sequences for the CH2-CH3 Fc portion comprise a tandem CH3 domain as shown in Format F4 and comprise a CH2 domain containing a N297S substitution. Three variants of F10 proteins were also produced: (a) a first wherein the cysteine residues in the hinge region were left intact (wild-type, referred to as F10A), (b) a second wherein the cysteine residues in the hinge region were replaced by serine residues (F10B), and (c) a third containing a linker sequence GGGSS replacing residues DKTHTCPPCP in the hinge (F10C). Variants F10B and F10C provided advantages in production as they avoid the formation of homodimers of the central chain. The (V<sub>K</sub>-V<sub>H</sub>) unit was made up of a V<sub>H</sub> domain, a linker and a V<sub>K</sub> unit (scFv). The heterodimer is made up of:

(1) a first (central) polypeptide chain having domains arranged as follows (N- to C-termini):



and

(2) a second polypeptide chain having domains arranged as follows (N- to C-termini): V<sub>K</sub><sup>anti-CD19</sup> - Ck.

Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins was purified from cell culture supernatant by affinity chromatography using prot-A beads and analysed and purified by SEC. The protein showed a good production yield of 2 mg/L (F10A) and with a simple SEC profile. The amino acid sequences for the three F9 protein chains for each of variants F10A, F10B and F10C are shown in the SEQ ID NOS listed in the table below.

Protein	SEQ ID NOS
F10A	25, 26
F10B	27, 28
F10C	29, 30

F10A:

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV

VCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRGEC- (SEQ ID NO: 25)

5 QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT  
YICNVNHKPSNTKVDKRVKPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVD  
VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
10 KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD  
GSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGSGGGGSGGGGSGGGGSGQPREPQV  
YTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSR  
WQQGNVFCFSVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTM  
HWVKQSHGKSLEWIGGISPNIIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGS  
15 FDYWGQGTTLTVSSVEGGSGGGSGGGSGGGVDDIVMTQSPATLSVTPGDRVSLSCRASQSIDYLHWY  
QQKSHESPRLLIKYASQSIGIPSRFSGSGSGSDFTLTINSVEPEDVGVYYCQNGHSFPLTFGAGTKL  
ELK- (SEQ ID NO: 26)

F10B:

- Frag1

20 DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGD SYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRGEC-(SEQ ID NO: 27)

25 QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT  
YICNVNHKPSNTKVDKRVKPKSCDKTHTSPPSPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVD  
VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
30 KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD  
GSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGSGGGGSGGGGSGGGGSGQPREPQV  
YTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSR  
WQQGNVFCFSVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTM  
HWVKQSHGKSLEWIGGISPNIIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGS  
35 FDYWGQGTTLTVSSVEGGSGGGSGGGSGGGVDDIVMTQSPATLSVTPGDRVSLSCRASQSIDYLHWY  
QQKSHESPRLLIKYASQSIGIPSRFSGSGSGSDFTLTINSVEPEDVGVYYCQNGHSFPLTFGAGTKL  
ELK- (SEQ ID NO: 28)

F10C:

40 DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGD SYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRGEC- (SEQ ID NO: 29)

45 QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT  
YICNVNHKPSNTKVDKRVKPKSCGGGSSPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSH  
DPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTIS  
50 KAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFF  
LYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGSGGGGSGGGGSGGGGSGQPREPQVYTL

PSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQG  
 NVFSCSVMEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVK  
 QSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDYW  
 GQGTTTLTVSSVEGGSGGSGGSGGVDDIVMTQSPATLSVTPGDRVSLSCRASQSSISDYLHWYQQKS  
 HESPRLLIKYASQSSISGIPSRFSGSGSGSDFTLTSLNSVEPEDVGVYYCQNGHSFPLTFGAGTKLELK-  
 (SEQ ID NO: 30)

*Format 11 (F11): CD19-F11-NKp46-3*

The domain structure of F11 polypeptides is shown in **Figure 2C**. The heterodimeric  
 protein is similar to F10 except that the structures of the antigen binding domains are  
 reversed. One of the two antigen binding domains has a Fab-like structure, and the other is  
 a scFv. The heterodimer is made up of

(1) a first (central) polypeptide chain having domains arranged as follows (N- to C-  
 termini):

$(V_K - V_H)^{\text{anti-CD19}} - \text{CH}_2 - \text{CH}_3 - \text{CH}_3 - V_H^{\text{anti-NKp46}} - C_K$

and

(2) a second polypeptide chain having domains arranged as follows (N- to C-  
 termini):  $V_K^{\text{anti-NKp46}} - \text{CH}_1$ .

Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins  
 was purified from cell culture supernatant by affinity chromatography using prot-A beads and  
 analysed and purified by SEC. The protein showed a good production yield of 2 mg/L and  
 with a simple SEC profile. The amino acid sequences for the two chains of the F11 protein  
 are shown in SEQ ID NOS 31 and 32.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGD SYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
 SSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGSGGGGSGVQLQQSGAE  
 LVRPGSSVKISCKASGYAFSSYWMNVKQRPQGQGLEWIGQIWPGDGDTNNGKFKGKATLTADSSST  
 AYMQLSSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTTVTVSSGGGSSAPELLGGPSVFLFPPKP  
 KDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLN  
 GKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWES  
 NGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQGNVFCVMEALHNHYTQKSLSLSPGSGGGS  
 GGGGSGGGGSGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPV  
 LDSDGSFFLYSKLTVDKSRWQQGNVFCVMEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGA  
 SVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELR  
 SLTSEDSAVYYCARRGGSFDYWGGQTTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVCLLNNFYPRE  
 AKVQWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNR  
 GEC- (SEQ ID NO: 31)

DIVMTQSPATLSVTPGDRVSLSCRASQSSISDYLHWYQQKSHESPRLLIKYASQSSISGIPSRFSGSGS  
 SDFTLTSLNSVEPEDVGVYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKR  
 VEPKSCDKTH- (SEQ ID NO: 32)

*Format 12 (F12): CD19-F12-NKp46-3*

The domain structure of the dimeric F12 polypeptides is shown in **Figure 2C**, wherein the bonds between the CH1 and Ck domains are disulfide bonds. The heterodimeric protein is similar to F11 but the CH1 and Ck domains within the F(ab) structure are inversed. The heterodimer is made up of:

(1) a first (central) polypeptide chain having domains arranged as follows (N- to C-termini):

$(V_k - V_H)^{\text{anti-CD19}} - \text{CH2} - \text{CH3} - \text{CH3} - V_H^{\text{anti-NKp46}} - \text{CH1}$

and

(2) a second polypeptide chain having domains arranged as follows (N- to C-termini):  $V_k^{\text{anti-NKp46}} - \text{Ck}$ .

Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins was purified from cell culture supernatant by affinity chromatography using prot-A beads and analysed and purified by SEC. The protein showed a good production yield of 2.8 mg/L and with a simple SEC profile. The amino acid sequences for the two chains of the F12 protein are shown in SEQ ID NOS: 33 and 34.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGD SYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGSGGGGSGVQLQQSGAE  
LVRPGSSVKISCKASGYAFSSYWMNWVKQRPQG LEWIGQIWPGDGDTNYNGKFKGKATLTADESSST  
AYMQLSSLAS EDSAVYFCARRETTT VGRYYYAMDYWGQGT TTVTVSSGGGSSAPELLGGPSVFLFPKP  
KDTLMISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLN  
GKEYKCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWES  
NGQPENNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNV FSCSV MHEALHNHYTQKSLSLSPGGGGGS  
GGGGSGGGGSGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVL  
DSDGSFFLYSKLTVDKSRWQQGNV FSCSV MHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGA  
SVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELR  
SLTSEDSAVYYCARRGGSFDYWGQGTTLTVSSASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEP  
VTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKRVEPKSCD  
KTH- (SEQ ID NO: 33)

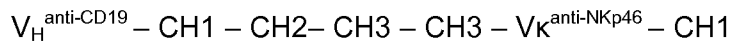
- Frag2

DIVMTQSPATLSVTPGDRVSLSCRASQSIDYLHWYQQKSHESPRLLIKYASQISGIPSRFSGSGSG  
SDFTLSINSVEPEDVGVIYCQNGHSFPLTFGAGTKLELKRVAAPSVFIFPPSDEQLKSGTASVVCLL  
NNFYPREAKVQWKVDNALQSGNSQESVTEQDSKSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSP  
VTKSFNRGEC- (SEQ ID NO: 34)

*Format 17 (F17): CD19-F17-NKp46-3*

The domain structure of the trimeric F17 polypeptides is shown in **Figure 2C**, wherein the bonds between the CH1 and Ck domains are disulfide bonds. The heterodimeric protein is similar to F9 but the  $V_H$  and  $V_k$  domains, and the CH1 and Ck, domains within the C-terminal F(ab) structure are each respectively inversed with their partner. The heterotrimer is made up of:

(1) a first (central) polypeptide chain having domains arranged as follows (N- to C-termini):

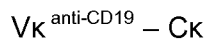


and

(2) a second polypeptide chain having domains arranged as follows (N- to C-termini):  $V_H^{\text{anti-NKp46}} - C_K$

and

(3) a third polypeptide chain having domains arranged as follows (N- to C-termini):



Additionally, three variants of F17 proteins were produced: (a) a first where the cysteine residues in the hinge region were left intact (wild-type, referred to as F17A), (b) a second wherein the cysteine residues in the hinge region were replaced by serine residues (F10B, and (c) a third containing a linker sequence GGGSS which replaces residues DKHTCPCPCP in the hinge (F17C). Proteins were cloned, produced and purified as in Example 2-1. The amino acid sequences for the three chains of the F17B protein chains are shown in SEQ ID NOS: 35, 36 and 37.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGD SYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRGEC-(SEQ ID NO: 35)

QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYAMDYWGQGTTVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT  
YICNVNHKPSNTKVDKRVEPKSCDKTHTSPPSPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD  
VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD  
GSFFLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGSGGGGSGGGGSGGGGSGQPREPQV  
YTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDKSR  
WQQGNVFCFSVMHEALHNHYTQKSLSLSPGSTGSDIVMTQSPATLSVTPGDRVSLSCRASQISIDYLH  
WYQQKSHESPRLLIKYASQISGIPSRFSGSGSGSDFTLTINSVEPEDVGVYYCQNGHSFPLTFGAGT  
KLELKASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLY  
SLSSVTVTPSSSLGTQTYICNVNHKPSNTKVDKRVEPKSCDKTH-(SEQ ID NO: 36)

EVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKA  
TLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFYWGQGTTTLTVSSRTVAAPSVFIFPPSDEQLKS  
GTASVVCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACE  
VTHQGLSSPVTKSFNRGEC-(SEQ ID NO: 37)

#### Example 4:

##### Bispecific NKp46 antibody formats with dimeric Fc domains

New protein constructions with dimeric Fc domains were developed that share many of the advantages of the monomeric Fc domain proteins of Example 3 but bind to FcRn with

greater affinity. Different protein formats were produced that either had low or substantially lack of binding to Fc $\gamma$ R (including CD16) or which had binding to Fc $\gamma$ Rs (including CD16), e.g. the binding affinity to human CD16 was within 1-log of that of wild-type human IgG1 antibodies, as assessed by SPR (e.g. see methods of Example 15. The different polypeptide

5 formats were tested and compared to investigate the functionality of heterodimeric proteins comprising a central chain with a (V<sub>H</sub>-(CH1/C $\kappa$ )-CH2-CH3-) unit or a (V $\kappa$ -(CH1 or C $\kappa$ )-CH2-CH3-) unit. One of both of the CH3 domains is fused, optionally via intervening amino acid sequences or domains, to a variable domain(s) (a single variable domain that associates with a variable domain on a separated polypeptide chain, a tandem variable domain (e.g., an

10 scFv), or a single variable domain that is capable of binding antigen as a single variable domain). The two chains associate by CH1-C $\kappa$  dimerization to form disulfide linked dimers, or if associated with a third chain, to form trimers.

Different constructs were made for use in the preparation of a bispecific antibody using the variable domains DNA and amino acid sequences derived from the scFv specific

15 for tumor antigen CD19 described in Example 2-1 and different variable regions from antibodies specific for NKp46 identified in Example 1. Proteins were cloned, produced and purified as in Example 2-1. Domains structures are shown in **Figures 2A-6D**.

*Format 5 (F5): CD19-F5-NKp46-3*

The domain structure of the trimeric F5 polypeptide is shown in **Figure 2D**, wherein

20 the interchain bonds between hinge domains (indicated in the figures between CH1/C $\kappa$  and CH2 domains on a chain) and interchain bonds between the CH1 and C $\kappa$  domains are interchain disulfide bonds. The heterotrimer is made up of:

(1) a first (central) polypeptide chain having domains arranged as follows (N- to C-termini):

25 V<sub>H</sub><sup>anti-CD19</sup> – CH1 – CH2 – CH3 – V<sub>H</sub><sup>anti-NKp46</sup> – C $\kappa$

and

(2) a second polypeptide chain having domains arranged as follows (N- to C-termini): V $\kappa$ <sup>anti-CD19</sup> – C $\kappa$  – CH2 – CH3

and

30 (3) a third polypeptide chain having domains arranged as follows (N- to C-termini):

V $\kappa$ <sup>anti-NKp46</sup> – CH1.

Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins was purified from cell culture supernatant by affinity chromatography using prot-A beads and analysed and purified by SEC. The protein showed a high production yield of 37 mg/L and

35 with a simple SEC profile. The amino acid sequences of the three chains are shown in SEQ ID NOS: 38, 39 and 40.



DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
 SSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFI FPPSDEQLKSGTASV  
 VCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQG  
 5 LSSPVTKSFNRGECDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVK  
 FNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQ  
 PREPQVYITLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKL  
 TVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGK- (SEQ ID NO: 38)

QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNNGKFKGKA  
 TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
 PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT  
 YICNVNHKPSNTKVDKRVPEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD  
 VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
 10 KTISKAKGQPREPQVYITLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD  
 GSFFLYSKLTVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVK  
 ISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLT  
 SEDSAVYYCARRGGSFDYWGQGTITVSSRTVAAPSVFI FPPSDEQLKSGTASVVCLLNNFYFPREAKV  
 15 QWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC  
 - (SEQ ID NO: 39)

DIVMTQSPATLSVTPGDRVSLSCRASQSIDYLHWYQQKSHESPRLLIKYASQSIGIPSRFSGSGSG  
 SDFTLINSVEPEDVGVIYCCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKR  
 20 VEPKSCDKTH- (SEQ ID NO: 40)

#### Format 6 (F6): CD19-F6-NKp46-3

The domain structure of heterotrimeric F6 polypeptides is shown in **Figure 2D**. The  
 F6 protein is the same as F5, but contains a N297S substitution to avoid N-linked  
 glycosylation. Proteins were cloned, produced and purified as in Example 2-1. Bispecific  
 30 proteins were purified from cell culture supernatant by affinity chromatography using prot-A  
 beads and analyzed and purified by SEC. The protein showed a high production yield of 12  
 mg/L and the purified proteins exhibited a simple SEC profile. The amino acid sequences of  
 the three chains of the F6 protein are shown in SEQ ID NOS: 41, 42 and 43.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
 SSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFI FPPSDEQLKSGTASV  
 VCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYSLSSTLTLSKADYEKHKVYACEVTHQG  
 40 LSSPVTKSFNRGECDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVK  
 FNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQ  
 PREPQVYITLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKL  
 TVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGK-(SEQ ID NO: 41)

QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNNGKFKGKA  
 TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
 PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT  
 YICNVNHKPSNTKVDKRVPEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVD  
 45 VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE

KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD  
 GSFFLYSKLTVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVK  
 ISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLT  
 SEDSAVYYCARRGGSFDYWGQGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKV  
 5 QWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC  
 - (SEQ ID NO: 42)

DIVMTQSPATLSVTPGDRVSLSCRASQISDYLHWYQQKSHESPRLLIKYASQSIGIPSRFSGSGSG  
 SDFTLSINSVEPEDVGYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
 10 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKR  
 VEPKSCDKTH- (SEQ ID NO: 43)

*Format 7 (F7) : CD19-F7-NKp46-3*

The domain structure of heterotrimeric F7 polypeptides is shown in **Figure 2D**. The  
 15 F7 protein is the same as F6, except for cysteine to serine substitutions in the CH1 and C<sub>k</sub>  
 domains that are linked at their C-termini to Fc domains, in order to prevent formation of a  
 minor population of dimeric species of the central chain with the V<sub>k</sub><sup>anti-NKp46</sup> – CH1 chain.  
 Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins were  
 purified from the cell culture supernatant by affinity chromatography using prot-A beads and  
 20 analyzed and purified by SEC. The protein showed a high production yield of 11 mg/L and  
 the purified proteins exhibited a simple SEC profile. The amino acid sequences of the three  
 chains of the 76 protein are shown in SEQ ID NOS: 44, 45 and 46.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
 25 SSGGTDFTLNIHPVEKVDAAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
 VCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
 LSSPVTKSFNRGECSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVK  
 FNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQ  
 PREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKL  
 30 TVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGK- (SEQ ID NO: 44)

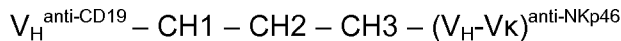
QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNNGKFKGKA  
 TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
 35 PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT  
 YICNVNHKPSNTKVDKRVEPKSSDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDV  
 VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
 KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD  
 GSFFLYSKLTVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVK  
 40 ISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLT  
 SEDSAVYYCARRGGSFDYWGQGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKV  
 QWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC  
 - (SEQ ID NO: 45)

DIVMTQSPATLSVTPGDRVSLSCRASQISDYLHWYQQKSHESPRLLIKYASQSIGIPSRFSGSGSG  
 45 SDFTLSINSVEPEDVGYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKR  
 VEPKSCDKTH- (SEQ ID NO: 46)

*Format 13 (F13): CD19-F13-NKp46-3*

The domain structure of the dimeric F13 polypeptide is shown in **Figure 2D**, wherein the interchain bonds between hinge domains (indicated between CH1/Ck and CH2 domains on a chain) and interchain bonds between the CH1 and Ck domains are interchain disulfide bonds. The heterodimer is made up of:

(1) a first (central) polypeptide chain having domains arranged as follows (N- to C-termini):



and

(2) a second polypeptide chain having domains arranged as follows (N- to C-termini):  $V_K^{\text{anti-CD19}} - \text{Ck} - \text{CH2} - \text{CH3}$ .

The  $(V_H - V_K)$  unit was made up of a  $V_H$  domain, a linker and a  $V_K$  unit (scFv).

Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins were purified from the cell culture supernatant by affinity chromatography using prot-A beads and analyzed and purified by SEC. The protein showed a high production yield of 6.4 mg/L and the purified proteins exhibited a simple SEC profile. The amino acid sequences of the two chains of the F13 protein are shown in SEQ ID NOS: 47 and 48.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGD SYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFFGGGKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNFFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSSTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRRGECDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVK  
FNWYVDGVEVHNAKTKPREEQY<sup>N</sup>STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQ  
PREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDSGSFLLYSKL  
TVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK- (SEQ ID NO: 47)

QVQLQQSGAELVLRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQT  
YICNVNHKPSNTKVDKRVPEPKSCKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVD  
VSHEDPEVKFNWYVDGVEVHNAKTKPREEQY<sup>N</sup>STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD  
GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVK  
ISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLT  
SEDSAVYYCARRGGSFDYWGQGTITLVSSVEGGSGGSGGSGGVDDIVMTQSPATLSVTPGDRVSL  
SCRASQSI SDYLHWYQQKSHESPRLLIKYASQSI SGIPSRFSGSGSGSDFTLSINSVEPEDVGYYCQ  
NGHSFPLTFGAGTKLELK- (SEQ ID NO: 48)

*Format 14 (F14): CD19-F14-NKp46-3*

The domain structure of the dimeric F14 polypeptide is shown in **Figure 2E**. The F14 polypeptide is a dimeric polypeptide which shares the structure of the F13 format, but instead of a wild-type Fc domain (CH2-CH3), the F14 bispecific format has CH2 domain mutations N297S to abolish N-linked glycosylation. Proteins were cloned, produced and

purified as in Example 2-1. Bispecific proteins were purified from cell culture supernatant by affinity chromatography using prot-A beads and analyzed and purified by SEC. The protein showed a high production yield of 2.4 mg/L and the purified proteins exhibited a simple SEC profile. The amino acid sequences of the two chains of the F14 protein are shown in SEQ ID NOS: 49 and 50.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
SGSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
VCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSSTYSLSSTLTLSKADYEKHKVYACEVTHQG  
LSSPVTKSFNRRGECDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVK  
FNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQ  
PREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKL  
TVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK-(SEQ ID NO: 49)

QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQT  
YICNVNHKPSNTKVDKRVKPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDV  
VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD  
GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVK  
ISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLT  
SEDSAVYYCARRGGSFYDWGQGTITLVSSVEGGSGGSGGSGGVDDIVMTQSPATLSVTPGDRVSL  
SCRASQSI SDYLHWYQQKSHESPRLLIKYASQSI SGIPSRFSGSGSGSDFTLSINSVEPEDVGVYYCQ  
NGHSFPLTFGAGTKLELK-(SEQ ID NO: 50)

#### *Format 15 (F15): CD19-F15-NKp46-3*

The domain structure of the trimeric F15 polypeptides is shown in **Figure 2E**. The F15 polypeptide is a dimeric polypeptide which shares the structure of the F6 format, but differs by inversion of the N-terminal V<sub>H</sub>-CH1 and V<sub>K</sub>-C<sub>K</sub> units between the central and second chains. Proteins were cloned, produced and purified as in Example 2-1. Bispecific proteins were purified from the cell culture supernatant by affinity chromatography using prot-A beads and analyzed and purified by SEC. The protein showed a good production yield of 0.9 mg/L and the purified proteins possessed a simple SEC profile. The amino acid sequences of the three chains of the F15 protein are shown in SEQ ID NOS: 51, 52 and 53.

QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSSASTKGPSVFPLA  
PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQT  
YICNVNHKPSNTKVDKRVKPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDV  
VSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSD  
GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK-(SEQ ID NO: 51)

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
 SSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
 VCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
 LSSPVTKSFNRGEC**CDK**THTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVK  
 5 FNWYVDGVEVHNAKTKPREEQSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQP  
 REPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLT  
 VDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTF  
 TEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCA  
 RRGGSFDYWGGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYFPREAKVQWKVDNALQS  
 10 GNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC- (SEQ ID  
 NO: 52)

DIVMTQSPATLSVTPGDRVSLSCRASQSISDYLHWYQQKSHESPRLLIKYASQSISGIPSRFSGSGSG  
 SDFTLINSVEPEDVGVYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
 15 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDR  
 VEPKSCDKTH-(SEQ ID NO: 53)

#### *Format 16 (F16): CD19-F16-NKp46-3*

The domain structure of the trimeric F16 polypeptide is shown in **Figure 2E**. The F16  
 20 polypeptide is a dimeric polypeptide which shares the structure of the F6 format, but differs  
 by inversion of the C-terminal V<sub>H</sub>-CK and V<sub>K</sub>-CH1 units between the central and second  
 chains. Proteins were cloned, produced and purified as in Example 2-1. The amino acid  
 sequences of the three chains of the F16 protein are shown in SEQ ID NOS: 54, 55 and 56.

DIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIYDASNLVSGIPPRFSG  
 25 SSGTDFTLNIHPVEKVDAATYHCQQSTEDPWTFGGGTKLEIKRTVAAPSVFIFPPSDEQLKSGTASV  
 VCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQG  
 LSSPVTKSFNRGEC**CDK**THTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVK  
 FNWYVDGVEVHNAKTKPREEQY**S**STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQ  
 PREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKL  
 30 TVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGK- (SEQ ID NO: 54)

QVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGQGLEWIGQIWPGDGDTNYNGKFKGKA  
 TLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQTTVTVSSASTKGPSVFPLA  
 35 PSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQT  
 YICNVNHKPSNTKVDRVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVD  
 VSHEDPEVKFNWYVDGVEVHNAKTKPREEQY**S**STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIE  
 KTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSD  
 GSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGSTGSDIVMTQSPATLSVTPGDRV  
 40 SLSCRASQSISDYLHWYQQKSHESPRLLIKYASQSISGIPSRFSGSGSGSDFTLINSVEPEDVGVYY  
 CQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLVKDYFPEPVTVSWNSGALT  
 GVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDRVEPKSCDKTH-(SEQ ID  
 NO: 55)

EVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKA  
 45 TLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDYWGGTTLTVSSRTVAAPSVFIFPPSDEQLKS  
 GTASVVCLLNNFYFPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACE  
 VTHQGLSSPVTKSFNRGEC-(SEQ ID NO: 56)

#### *Format T5 (T5)*

The domain structure of a trimeric T5 polypeptide is shown in Figure 2F. The T5 polypeptide is a trimeric polypeptide which shares the structure of the F5 format, but differs by fusion of an scFv unit at the C-terminus of the third chain (the chain lacking the Fc domain). This protein will therefore have two antigen binding domains for antigens of interest, and one for NKp46, and will bind CD16 via its Fc domain. Proteins were cloned, produced and purified as in Example 2-1. Two different T5 proteins were produced, as follows.

A first T5 protein had one antigen binding domain that binds human CD137, one antigen binding domain that binds human NKp46, and one antigen binding domain that binds CD19. The amino acid sequences of the three chains of the T5 CD137-T5-NKp46-CD19 protein are shown in below (anti-CD137 in bold and underlined, anti-CD19 underlined, anti-NKp46 in italics).

#### *CD137-T5-NKp46-CD19*

##### *Polypeptide 1:*

Residues 1-121 are anti-CD137 binding domain; 122-454 and 571-677 are T5 sequences; 455-570 are anti-NKp46 binding domain

**QVQLQQWGAGLLKPSETLSLTCAVYGGSFSGYYWSWIRQSPEKGLEWIGEINHGGYVTYNPSLESRVTI**  
**ISVDTSKNQFSLKLSSVTAADTAVYYCARDYGPNGYDWYFDLWGRGTLVTVSS**ASTKGPSVFPLAPSS  
 KSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVTPSSSLGTQTYIC  
 NVNHKPSNTKVDKRVPEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMI SRTPEVTCVVDVSH  
 EDPEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKT  
 SKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPVLDSDGSF  
 FLYSKLTVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISC  
 KTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSED  
 SAVYYCARRGGSFDYWGQGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVVCLLNMFYPREAKVQWK  
 VDNALQSGNSQESVTEQDSKDYSLSSLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC-  
 (SEQ ID NO: 57)

##### *Polypeptide 2:*

Residues 1-109 (anti-CD137); 110-443 are T5 sequences

**EIVLTQSPATLSLSPGERATLSCRASQSVSSYLAWYQQKPGQAPRLLIYDASNRAITGIPARFSGSGSG**  
**TDFTLTITSSLEPEDFAVYYCQQRSNWPPALTFCGGTKVEIK**RTVAAPSVFIFPPSDEQLKSGTASVVC  
 LLNMFYPREAKVQWKVDNALQSGNSQESVTEQDSKDYSLSSLTLSKADYEKHKVYACEVTHQGLS  
 SPVTKSFNRGEC DKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMI SRTPEVTCVVDVSHEDPEVKFN  
 WYVDGVEVHNAKTKPREEQYNSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPR  
 EPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPVLDSDGSFFLYSKLTV  
 DKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGK-(SEQ ID NO: 58)

##### *Polypeptide 3:*

Residues 1-107 are anti-NKp46 binding domain; 108-219 are T5 sequences; 220-469 are anti-CD19 binding domain

5 *DIVMTQSPATLSVTPGDRVSLSCRASQSSISDYLHWYQQKSHESPRLLIKYASQSSISGIPSRFSGSGSG*  
*SDFTLSINSVEPEDVGVIYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV*  
 KYDFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKR  
 VEPKSCDKTHGGSSSDIQLTQSPASLAVSLGQRATISCKASQSVDDYDGDSYLNWYQQIPGQPPKLLIY  
 10 DASNLVSGIIPRFSGSGSGTDFTLNIHPVEKVDAAITYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGS  
GGGGSQVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPGGLEWIGQIWPGDGDNTYNGK  
FKGKATLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSS-  
 (SEQ ID NO: 59)

A second T5 protein had two antigen binding domains that bind human CD20, originating from different antibodies (and binding to different epitopes on CD20). The first  
 15 anti-CD20 ABD contained the VH and VL of the parent antibody GA101 (GAZYVA®, Gazyvaro®, obinutuzumab, Roche Pharmaceuticals). The second anti-CD20 ABD contained the VH and VL of the parent antibody rituximab (Rituxan®, Mabthera®, Roche Pharmaceuticals). The third antigen binding domain binds human NKp46. The amino acid sequences of the three chains of the T5 protein are shown in below (Rituximab sequences in  
 20 bold and underlined, anti-GA101 sequences underlined, anti-NKp46 in italics).

#### *GA101-T5-Ritux-NKp46*

##### *Polypeptide 1:*

25 QVQLVQSGAEVKKPGSSVKVSCKASGYAFSSYWINWVRQAPGGGLEWMGRIFPGDGDTDYNGKFKGRV  
TITADKSTSTAYMELSSLRSEDTAVYYCARNVFDGYWLVYWGQGTLVTVSSASTKGPSVFPLAPSSKS  
 TSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNV  
 NHKPSNTKVDKRVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHED  
 30 PEVKFNWYVDGVEVHNAKTKPREEQYNSTYRVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISK  
 AKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFL  
 YSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGSTGS**QVQLQQPGAELVKPGASVKMSCKA**  
**SGYTF****TSYNMH****WKQTPGRGLEWIGAIYPNGDTSYNQKFKGKATLTADKSSSTAYMQLSSLTSEDSA**  
**VYYCARSTYYGGDWYFNVWGAGTTVTVSA****R****TVAAPS****VFI****FPPSDEQLKSGTASV****VCLLNNFY****P****REAKV**  
 35 **QWKVDNALQSGNSQESVTEQDSK****STYLS****SSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC**  
 -(SEQ ID NO: 60)

##### *Polypeptide 2:*

40 DIVMTQTPLSLPVTPGEPASISCRSSKSLHNSGITYLYWYLQKPGQSPQLLIYQMSNLVSGVPDRFS  
SGSGTDFTLKISRVEAEDVGVIYCAQNLELPYTFGGGTKVEIKRTVAAPSVFIFPPSDEQLKSGTAS  
VVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKSTYLSSSTLTLSKADYEKHKVYACEVTHQ  
 GLSSPVTKSFNRGE**CDK****HTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEV**  
 KFNWYVDGVEVHNAKTKPREEQYNSTYRVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKG  
 QPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSK  
 45 LTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGK- (SEQ ID NO: 61)

##### *Polypeptide 3:*

**QIVLSQSPAILSASPGEKVTMT****CRASSSVSYI****HWFQ****QKPGSSPKPWIYATSNLASGV****PVRFSGSGSGT**

**SYSLTISRVEAEDAATYYCQQWTSNPPTFGGGTKLEIK**ASTKGPSVFPLAPSSKSTSGGTAALGCLVK  
DYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKRV  
EPKSCDKTHGGSSSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPN  
IGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDYWGGTTLTVSSVEGGSG  
5 GSGGSGSGGVDDIVMTQSPATLSVTPGDRVSLSCRASQSIDYHLHWYQQKSHESPRLLIKYASQSID  
GIPSRFSGSGSGSDFTLINSVEPEDVGVYYCQNGHSFPLTFGAGTKLELK- (SEQ ID NO: 62)

### *Format T6 (T6)*

The domain structure of the trimeric T6 polypeptide is shown in Figure 2F. The T6  
10 polypeptide is a trimeric polypeptide which shares the structure of the F6 format, but differs  
by fusion of an scFv unit at the C-terminus of the third chain (the chain lacking the Fc  
domain). This protein has two antigen binding domains for antigens of interest, and one for  
NKp46, but does not bind CD16 via its Fc domain due to the N297 substitution. Proteins  
were cloned, produced and purified as in Example 2-1. Two different T6 proteins were  
15 produced. A first T6 protein had one antigen binding domain that binds human CD137, one  
antigen binding domain that binds human NKp46, and one antigen binding domain that binds  
CD19. The second T6 protein had two antigen binding domains that bind human CD20  
(CD137-T6-NKp46-CD19). The first anti-CD20 ABD contained the VH and VL of the parent  
antibody GA101 and the second anti-CD20 ABD contained the VH and VL of the parent  
20 antibody rituximab. The amino acid sequences of the three chains of the T6 proteins are  
shown in below.

### *CD137-T6-NKp46-CD19*

#### *Polypeptide 2:*

EIVLTQSPATLSLSPGERATLSCRASQSVSSYLAWYQQKPGQAPRLLIYDASNRATGIPARFSGSGSG  
TDFTLTITSSLEPEDFAVYYCQQRSNWPPALTFCGGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVC  
LLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLS  
SPVTKSFNRGEC**CD**KTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHEDPEVKFN  
30 WYVDGVEVHNAKTKPREEQY**S**STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPR  
EPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTV  
DKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGK- (SEQ ID NO: 63)

#### *Polypeptide 1:*

QVQLQQWAGLLKPSETLSLTCAVYGGSFSGYYWSWIRQSPEKGLEWIGEINHGGYVTYNPSLESRV  
ISVDTSKNQFSLKLSSVTAADTAVYYCARDYGPNGYDWYFDLWGRGTLTVSSASTKGPSVFPLAPSS  
KSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYIC  
NVNHKPSNTKVDKRVPEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSH  
EDPEVKFNWYVDGVEVHNAKTKPREEQY**S**STYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKT  
40 SKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSF  
FLYSKLTVDKSRWQQGNVFCFSVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISC  
KTSYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSED  
SAVYYCARRGGSFDYWGGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWK  
VDNALQSGNSQESVTEQDSKDSTYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC-  
45 (SEQ ID NO: 64)



*Polypeptide 3:*

DIVMTQSPATLSVTPGDRVSLSCRASQSISDYLHWYQQKSHESPRLLIKYASQSIGIPSRFSGSGSG  
 SDFTLINSINVEPEDVGVIYCAQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
 5 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKR  
 VEPKSCDKTHGGSSTDIQLTQSPASLAVSLGQRATISCKASQSVSDYDGD SYLNWYQQIPGQPPKLLIY  
 DASNLVSGIIPRFSGSGSGTDFTLNIHPVEKVDAAATYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGS  
 GGGGSQVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKRPGQGLEWIGQIWPGDGDTNYNGK  
 FK GKATLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGT TVTVSS-  
 10 (SEQ ID NO: 65)

*GA101-T6-Ritux-NKp46**Polypeptide 2:*

DIVMTQTPLSLPVTPGEPASISCRSSKSLLSNGITYLYWYLQKPGQSPQLLIYQMSNLVSGVPDRFS  
 GSGSGTDFTLKISRVEAEDVGVIYCAQNLLELPYTFGGGKVEIKRTVAAPSVFIFPPSDEQLKSGTAS  
 15 VVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQ  
 GLSSPVTKSFNRGEC DKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEV  
 KFNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKG  
 20 QPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSK  
 LTVDKSRWQQGNV FSCSV MHEALHNHYTQKSLSLSPGK-  
 (SEQ ID NO: 66)

*Polypeptide 1:*

QVQLVQSGAEVKKPGSSVKVSCKASGYAFSYSWINWVRQAPGQGLEWMGRIFPGDGD TDYNGKFKGRV  
 TITADKSTSTAYMELSSLRSEDTAVYYCARNVFDGYWL VYWGQGT LVT VSSASTKGPSVFPLAPSSKS  
 TSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNV  
 25 NHKPSNTKVDKRVEPKSCDKTHTCPPCPAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHED  
 PEVKFNWYVDGVEVHNAKTKPREEQYSSSTYRVVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISK  
 30 AKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFL  
 YSKLTVDKSRWQQGNV FSCSV MHEALHNHYTQKSLSLSPGSTGSQVQLQQPGAELVKPGASVKMSCKA  
 SGYTFSTSYNMHWVKQTPGRGLEWIGAIYPGNGDTSYNQKFKGKATLTADKSSSTAYMQLSSLTSEDSA  
 VYYCARSTYYGGDWYFNVWGAGTTVTVSARTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKV  
 QWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC  
 35 (SEQ ID NO: 67)

*Polypeptide 3:*

QIVLSQSPAILSASPGEKVTMTCRASSSVSYIHWFAQKPGSSPKPWYIYATSNLASGVPVRFSGSGSGT  
 SYSLTISRVEAEDAATYYCQQWTSNPPTFGGGTKLEIKASTKGPSVFPLAPSSKSTSGGTAALGCLVK  
 40 DYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKRV  
 EPKSCDKTHGGSSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPN  
 IGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDYWGQGTTLTVSSVEGGSG  
 GSGGSGSGGVDDIVMTQSPATLSVTPGDRVSLSCRASQSISDYLHWYQQKSHESPRLLIKYASQSIG  
 45 GIPSRFSGSGSGSDFTLINSINVEPEDVGVIYCAQNGHSFPLTFGAGTKLELK- (SEQ ID NO: 68)

**Format T9B (T9B)**

The domain structure of the trimeric T9B polypeptide is shown in Figure 2F. The T9B  
 polypeptide is a trimeric polypeptide which shares the structure of the F9 format (F9B  
 variant), but differs by fusion of an scFv unit at the C-terminus of the free CH1 domain (on  
 50 the third chain). This protein will therefore have two antigen binding domains for antigen of

interest, and one for NKp46, but will not bind CD16 via its Fc domain due to the monomeric Fc domain and/or the N297 substitution. Proteins were cloned, produced and purified as in Example 2-1. Two different T9B proteins were produced. A first T9B protein had one antigen binding domain that binds human CD137, one antigen binding domain that binds human NKp46, and one antigen binding domain that binds CD19 (CD137-T9B-NKp46-CD19). The second T9B protein had two antigen binding domains that bind human CD20. The first anti-CD20 ABD contained the VH and VL of the parent antibody GA101 and the second anti-CD20 ABD contained the VH and VL of the parent antibody rituximab. The amino acid sequences of the three chains of the T9B proteins are shown in below.

#### CD137-T9B-NKp46-CD19

##### Polypeptide 2:

EIVLTQSPATLSLSPGERATLSCRASQSVSSYLAWYQQKPGQAPRLLIYDASNRATGIPARFSGSGSGTDFTLTISSLEPEDFAVYYCQQRSNWPPALTFCGGTKVEIKRTVAAPSVFIFPPSDEQLKSGTASVVC LLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLS SPVTKSFNRGEC- (SEQ ID NO: 69)

##### Polypeptide 1:

QVQLQQWAGALLKPSETLSLTCAVYGGSFSGYYWSWIRQSPEKGLEWIGEINHGGYVTYNPSLESRVTI SVDTSKNQFSLKLSSVTAADTAVYYCARDYGPNGYDWYFDLWGRGTLTVTVSSASTKGPSVFPLAPSS KSTSGGTAALGCLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYIC NVNHKPSNTKVDKRVEPKSCDKTHTSPAPPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSH EDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVSVLTVLHQDWLNGKEYCKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSF FLYSKLTVDKSRWQQGNVFCSCVMHEALHNHYTQKSLSLSPGGGGSGGGGSGGGGSGQPREPQVYTL PPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTPPVLDSDGSFFLYSKLTVDKSRWQQ GNVFCSCVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWV KQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDY WGQGTTLTVSSRTVAAPSVFIFPPSDEQLKSGTASVVC LLNNFYPREAKVQWKVDNALQSGNSQESVT EQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC- (SEQ ID NO: 70)

##### Polypeptide 3:

DIVMTQSPATLSVTPGDRVSLSCRASQSI SDYLHWYQQKSHESPRLLIKYASQSI SGIPSRFSGSGSGSDFTLTINSVEPEDVGYYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKR VEPKSCDKTHGGSSSDIQLTQSPASLAVSLGQRATISCKASQSVDDYDGD SYLNWYQQIPGQPPKLLIY DASNLVSGIPPRFSGSGSGTDFTLNIHPVEKVDAAATYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGS GGGGSQVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPQG LEWIGQIWPGDGDTNYNGK FKGKATLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGT TVTVSS- (SEQ ID NO: 71)

#### GA101-T9B-Ritux-NKp46

##### Polypeptide 2:

DIVMTQTPLSLPVTPGEPASISCRSSKSLHNSGITYLYWYLQKPGQSPQLLIYQMSNLVSGVPDRFSGSGGTDFTLKISRVEADVGVYYCAQNLELPYTFGGGTKVEIKRTVAAPSVFIFPPSDEQLKSGTAS

VVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQ  
GLSSPVTKSFNREGC- (SEQ ID NO: 72)

*Polypeptide 1:*

5 QVQLVQSGAEVKKPGSSVKVSCKASGYAFSYSWINWVRQAPGGGLEWMGRIFPGDGD TDYNGKFKGRV  
TITADKSTSTAYMELSSLRSEDTAVYYCARNVFDGYWLVYWGQGT LVTVSSASTKGPSVFPLAPSSKS  
TSGGTAALGCLVKDYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNV  
NHKPSNTKVDKRV EPKSCDKTHT **SPPSP**APPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVDVSHED  
10 PEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVSVLT VLVHQLDNLNGKEYCKVSNKALPAPIEKTISK  
AKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFL  
YSKLTVDKSRWQQGNV FSCSV MHEALHNHYTQKSLSLSPGSGGGGSGGGGSGGGGSGQPREPQVYTLPP  
SREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGN  
VFSCSV MHEALHNHYTQKSLSLSPGSTGSQVQLQPGAE LVKPGASVKMSCKASGYTFTSYNMHWVKQ  
15 TPGRGLEWIGAIYPGNGDTSYNQKFKGKATLTADKSSSTAYMQLSSLTSEDSAVYYCARSTYYGGDWY  
FNVWGAGTTVTVSARTVAAPSVFIFPPSDEQLKSGTASVVCLLNNFYPREAKVQWKVDNALQSGNSQ  
SVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNREGC- (SEQ ID NO: 73)

*Polypeptide 3:*

20 QIVLSQSPAILSASPGEKVTMTCRASSSVSYIHW FQQKPGSSPKPWIYATSNLASGVPVRFSGSGSGT  
SYSLTISRVEAEDAATYYCQQWTSNPPTFGGGTKLEIKASTKGPSVFPLAPSSKSTSGGTAALGCLVK  
DYFPEPVT VSWNSGALTSGVHTFPAVLQSSGLYSLSSVVTVPSSSLGTQTYICNVNHKPSNTKVDKRV  
EPKSCDKTHGGSSSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPN  
IGGTSYNQKFKGKATLTVDKSSSTAYMELRLTSEDSAVYYCARRGGSFDYWGQGTTLTVSSVEGGSG  
25 GSGGSGSGGVDDIVMTQSPATLSVTPGDRVSLSCRASQSIDY LHWYQQKSHESPRLLIKYASQSID  
GIPSRFSGSGSGSDFTLSINSVEPEDVGVYYCQNGHSFPLTFGAGTKLELK- (SEQ ID NO: 74)

*Format T11 (T1): CD19-T11-NKp46-3*

The domain structure of the dimeric T11 polypeptide is shown in Figure 2F. The T11  
polypeptide is a trimeric polypeptide which shares the structure of the F11 format, but differs  
30 by fusion of an scFv unit at the C-terminus of the free CH1 domain. This protein therefore  
has antigen binding domains for antigen of interest, and one for NKp46, but does not bind  
CD16 via its Fc domain due to the monomeric Fc domain and/or the N297 substitution.  
Proteins were cloned, produced and purified as in Example 2-1. Two different T11B proteins  
were produced. A first T11B protein had one antigen binding domain that binds human  
35 CD137, one antigen binding domain that binds human NKp46, and one antigen binding  
domain that binds CD19 (CD137-T9B-NKp46-CD19). The second T9B protein had two  
antigen binding domains that bind human CD20. The first anti-CD20 ABD contained the VH  
and VL of the parent antibody GA101 and the second anti-CD20 ABD contained the VH and  
VL of the parent antibody rituximab. The amino acid sequences of the three chains of the  
40 T11 protein are shown in below.

*CD137-T11-NKp46-CD19*

*Polypeptide 1:*

EIVLTQSPATLSLSPGERATLSCRASQSVSSYLAWYQQKPGQAPRLLIYDASNRATGIPARFSGSGSG  
 TDFTLTISSELEPEDFAVYYCQQRSNWPPALTFCGGKVEIKGGGSGGGGSGGGGSGVQLQWAGLL  
 KPSETLSLTCAVYGGSFSGYYWSWIRQSPEKGLEWIGEINHGGYVTYNPSLESRVTISVDTSKNQFSL  
 KLSSVTAADTAVYYCARDYGPNGYDWFYFDLWGRGTLVTVSSGGGSSAPELLGGPSVFLFPPKPKDTLM  
 5 ISRTPEVTCVVVDVSHEDPEVKFNWYVDGVEVHNAKTKPREEQYSSTYRVSVLTVLHQDWLNGKEYK  
 CKVSNKALPAPIEKTISKAKGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPE  
 NNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGGGGSGGGGS  
 GGGSGQPREPQVYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGS  
 10 FFLYSKLTVDKSRWQQGNVFSCSVMHEALHNHYTQKSLSLSPGSTGSEVQLQQSGPELVKPGASVKIS  
 CKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPNIGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSE  
 DSAVYYCARRGGSFDYWGQGTTLTVSSRTVAAPSVFIFFPSDEQLKSGTASVCLLNNFYPREAKVQW  
 KVDNALQSGNSQESVTEQDSKDYSLSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC-  
 (SEQ ID NO: 75)

**Polypeptide 2:**

DIVMTQSPATLSVTPGDRVSLSCRASQSI SDYLHWYQQKSHESPRLLIKYASQSI SGIPSRFSGSGSG  
 SDFTLISINSVEPEDVGVIYCQNGHSFPLTFGAGTKLELKASTKGPSVFPLAPSSKSTSGGTAALGCLV  
 KDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKR  
 VEPKSCDKTHGGSSSDIQLTQSPASLAVSLGQRATISCKASQSVDDYDGD SYLNWYQQIPGQPPKLLIY  
 20 DASNLVSGIPPRFSGSGSGTDFTLNIHPVEKVDAAATYHCQQSTEDPWTFGGGTKLEIKGGGSGGGGS  
 GGGGSGVQLQQSGAELVRPGSSVKISCKASGYAFSSYWMNWVKQRPQGLEWIGQIWPGDGDTNYNGK  
 FKGKATLTADESSSTAYMQLSSLASEDSAVYFCARRETTTVGRYYYAMDYWGQGTITVTVSS-  
 (SEQ ID NO: 76)

**GA101-T11-Ritux-NKp46**

**Polypeptide 1:**

DIVMTQTPLSLPVTPGEPASISCRSSKSLHNSGITYLYWYLQKPGQSPQLLIYQMSNLVSGVPDRFS  
 GSGSGTDFTLKISRVEAEDVGVIYCAQNLLELPYTFGGGKVEIKGGGSGGGGSGGGGSGVQLVQSGA  
 30 EVKPKGSSSVKVSCKASGYAFSYSWINWVRQAPQGQGLEWMGRIFPGDGD TDYNGKFKGRVTITADKSTS  
 TAYMELSSLRSED TAVYYCARNVFDGYWL VYWGQGT LTVTVSSASTKGPSVFPLAPSSKSTSGGTAALG  
 CLVKDYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKV  
 DKRVEPKSCDKTHT **SPPS**PAPELLGGPSVFLFPPKPKDTLMISRTPEVTCVVVDVSHEDPEVKFNWYV  
 35 DGVEVHNAKTKPREEQYSSTYRVSVLTVLHQDWLNGKEYKCKVSNKALPAPIEKTISKAKGQPREPQ  
 VYTLPPSREEMTKNQVSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDKS  
 RWQQGNVFSCSVMHEALHNHYTQKSLSLSPGGGGSGGGGSGGGGSGQPREPQVYTLPPSREEMTKNQ  
 VSLTCLVKGFYPSDIAVEWESNGQPENNYKTTTPVLDSDGSFFLYSKLTVDKSRWQQGNVFSCSVMHE  
 ALHNHYTQKSLSLSPGSTGSQVQLQQPGAELVKPGASVKMSCKASGYTFTSYNMHWVKQTPGRGLEWI  
 40 GAIYPNGDTSYNQKFKGKATLTADKSSSTAYMQLSSLTSEDSAVYYCARSTYYGGDWYFNWVGAGTT  
 VTVSARTVAAPSVFIFFPSDEQLKSGTASVCLLNNFYPREAKVQWKVDNALQSGNSQESVTEQDSKD  
 STYLSSTLTLSKADYEKHKVYACEVTHQGLSSPVTKSFNRGEC- (SEQ ID NO: 77)

**Polypeptide 2:**

QIVLSQSPAILASAPGEKVTMTCRASSSVSYIHWYQQKPGSSPKPWIYATSNLASGVPVRFSGSGSGT  
 45 SYSLTISRVEAEDAATYYCQQWTSNPPTFGGGTKLEIKASTKGPSVFPLAPSSKSTSGGTAALGCLVK  
 DYFPEPVTVSWNSGALTSGVHTFPAVLQSSGLYSLSSVTVPSSSLGTQTYICNVNHKPSNTKVDKRV  
 EPKSCDKTHGGSSSEVQLQQSGPELVKPGASVKISCKTSGYTFTEYTMHWVKQSHGKSLEWIGGISPN  
 IGGTSYNQKFKGKATLTVDKSSSTAYMELRSLTSEDSAVYYCARRGGSFDYWGQGTTLTVSSVEGGSG  
 50 GSGGSGSGGVDIVMTQSPATLSVTPGDRVSLSCRASQSI SDYLHWYQQKSHESPRLLIKYASQSI  
 GIPSRFSGSGSGSDFTLISINSVEPEDVGVIYCQNGHSFPLTFGAGTKLELK- (SEQ ID NO: 78)

**Example 5:****NKp46 binding affinity by bispecific proteins detected by Surface Plasmon Resonance (SPR)***Biacore T100 general procedure and reagents*

5 SPR measurements were performed on a Biacore T100 apparatus (Biacore GE Healthcare) at 25°C. In all Biacore experiments HBS-EP+ (Biacore GE Healthcare) and NaOH 10mM served as running buffer and regeneration buffer respectively. Sensorgrams were analyzed with Biacore T100 Evaluation software. Protein-A was purchased from (GE Healthcare). Human NKp46 recombinant proteins were cloned, produced and purified at  
10 Innate Pharma.

*Immobilization of Protein-A*

Protein-A proteins were immobilized covalently to carboxyl groups in the dextran layer on a Sensor Chip CM5. The chip surface was activated with EDC/NHS (N-ethyl-N'-(3-dimethylaminopropyl) carbodiimidehydrochloride and N-hydroxysuccinimide (Biacore GE  
15 Healthcare)). Protein-A was diluted to 10 µg/ml in coupling buffer (10 mM acetate, pH 5.6) and injected until the appropriate immobilization level was reached (i.e. 2000 RU). Deactivation of the remaining activated groups was performed using 100 mM ethanolamine pH 8 (Biacore GE Healthcare).

*Binding study*

20 Antibodies were tested as different formats F5, F6, F9, F10, F11, F13, F14 and compared to the single chain format (F1), and an anti-NKp46 antibody as a full-length human IgG1.

Bispecific proteins at 1 µg/mL were captured onto Protein-A chip and recombinant human NKp46 proteins were injected at 5 µg/mL over captured bispecific antibodies. For  
25 blank subtraction, cycles were performed again replacing NKp46 proteins with running buffer.

*Affinity study*

Monovalent affinity study was conducted following a regular Capture-Kinetic protocol recommended by the manufacturer (Biacore GE Healthcare kinetic wizard). Seven  
30 serial dilutions of human NKp46 recombinant proteins, ranging from 6.25 to 400 nM were sequentially injected over the captured Bi-Specific antibodies and allowed to dissociate for 10 min before regeneration. The entire sensorgram sets were fitted using the 1:1 kinetic binding model.

*Results*

SPR showed that the bispecific polypeptides of formats F1, F5, F6, F9, F10, F11, F13, F14 retained binding to NKp46. Monovalent affinities and kinetic association and dissociation rate constants are shown below in the Table 3 below.

Table 3

Bispecific mAb	ka (1/Ms)	kd (1/s)	KD (M)
CD19-F1-NKp46-3	7.05E+04	6.44E-04	9.14E-09
CD19-F5-NKp46-3	7.555E+4	0.00510	67E-09
CD19-F6-NKp46-3	7.934E+4	0.00503	63E-09
CD19-F9A-NKp46-3	2.070E+5	0.00669	32E-09
CD19-F10A-NKp46-3	2.607E+5	0.00754	29E-09
CD19-F11A-NKp46-3	3.388E+5	0.01044	30E-09
CD19-F13-NKp46-3	8.300E+4	0.00565	68E-09
CD19-F14-NKp46-3	8.826E+4	0.00546	62E-09
NKp46-3 IgG1	2.224E+5	0.00433	20E-09

**Example 6:****Engagement of NK cells against Daudi tumor target with Fc-containing NKp46 x CD19 bispecific protein**

Bispecific antibodies having a monomeric Fc domain and a domain arrangement according to the single chain F1 or dimeric F2 formats described in Example 3, and a NKp46 binding region based on different anti-NKp46 variable domains (NKp46-1, NKp46-2, NKp46-3 or NKp46-4) were tested for functional ability to direct NK cells to lyse CD19-positive tumor target cells (Daudi, a well characterized B lymphoblast cell line). The F2 proteins additionally included variable regions of a further antibody (NKp46-9) which lost binding to NKp46 in the scFv format but which retained binding in the F(ab)-like format of F2.

Briefly, the cytolytic activity of each of (a) resting human NK cells, and (b) human NK cell line KHYG-1 transfected with human NKp46, was assessed in a classical 4-h  $^{51}\text{Cr}$ -release assay in U-bottom 96 well plates. Daudi cells were labelled with  $^{51}\text{Cr}$  (50  $\mu\text{Ci}$  (1.85 MBq)/ $1 \times 10^6$  cells), then mixed with KHYG-1 transfected with hNKp46 at an effector/target ratio equal to 50 for KHYG-1, and 10 (for F1 proteins) or 8.8 (for F2 proteins) for resting NK cells, in the presence of monomeric bi-specific antibodies at different concentrations. After brief centrifugation and 4 hours of incubation at  $37^\circ\text{C}$ , samples of supernatant were removed and transferred into a LumaPlate (Perkin Elmer Life Sciences, Boston, MA), and  $^{51}\text{Cr}$  release was measured with a TopCount NXT beta detector (PerkinElmer Life Sciences, Boston, MA). All experimental conditions were analyzed in triplicate, and the percentage of specific lysis was determined as follows:  $100 \times (\text{mean cpm experimental release} - \text{mean cpm spontaneous release}) / (\text{mean cpm total release} - \text{mean cpm spontaneous release})$ . Percentage of total release is obtained by lysis of target cells with 2% Triton X100 (Sigma) and spontaneous release corresponds to target cells in medium (without effectors or Abs).

**Results**

In the KHYG-1 hNKp46 NK experimental model, each bi-specific antibody NKp46-1, NKp46-2, NKp46-3, NKp46-4 or NKp46-9 induced specific lysis of Daudi cells by human KHYG-1 hNKp46 NK cell line compared to negative controls (Human IgG1 isotype control (IC) and CD19/CD3 bi-specific antibodies), thereby showing that these antibodies induce Daudi target cell lysis by KHYG-1 hNKp46 through CD19/NKp46 cross-linking.

When resting NK cells were used as effectors, each bi-specific antibody NKp46-1, NKp46-2, NKp46-3, NKp46-4 or NKp46-9 again induced specific lysis of Daudi cells by human NK cells compared to negative control (Human IgG1 isotype control (IC) antibody), thereby showing that these antibodies induce Daudi target cell lysis by human NK cells through CD19/NKp46 cross-linking. Rituximab (RTX, chimeric IgG1) was used as a positive control of ADCC (Antibody-Dependent Cell Cytotoxicity) by resting human NK cells. The

maximal response obtained with RTX (at 10 µg/ml in this assay) was 21.6% specific lysis illustrating that the bispecific antibodies have high target cell lysis activity. Results for experiments with resting NK cells are shown in **Figure 3A** for the single chain F1 proteins and **3B** for the dimeric F2 proteins.

5

#### Example 7:

##### **Comparison with full length anti-NKp46 mAbs and depleting anti-tumor mAbs: only NKp46 x CD19 bispecific proteins prevent non-specific NK activation**

10 In these experiments bispecific antibodies possessing a specific bispecific format were produced in order to assess whether such bispecific antibodies can mediate NKp46-mediated NK activation toward cancer target cells without triggering non-specific NK cell activation.

Particularly, NKp46 x CD19 bispecific proteins having an arrangement according to the F2 format described in Example 3 with anti-NKp46 variable domains from NKp46-1, 15 NKp46-2, NKp46-3, NKp46-4 or NKp46-9 were compared to:

- (a) full-length monospecific anti-NKp46 antibodies (NKp46-3 as human IgG1), and
- (b) the anti-CD19 antibody as a full-length human IgG1 as ADCC inducing antibody control comparator.

20 The experiments further included as controls: rituximab, an anti-CD20 ADCC inducing antibody control for a target antigen with high expression levels; anti-CD52 antibody alemtuzumab, a human IgG1, binds CD52 target present on both targets and NK cells; and negative control isotype control therapeutic antibody (a human IgG1 that does not bind a target present on the target cells (HUG1-IC).

25 The different proteins were tested in order to assess their relative functional effects on NK cell activation in the presence of CD19-positive tumor target cells (Daudi cells), in the presence of CD19-negative, CD16-positive target cells (HUT78 T-lymphoma cells), and in the absence of target cells.

30 Briefly, NK activation was tested by assessing CD69 and CD107 expression on NK cells by flow cytometry. The assay was carried out in 96 U well plates in completed RPMI, 150µL final/well. Effector cells were fresh NK cells purified from donors. Target cells were Daudi (CD19-positive), HUT78 (CD19-negative) or K562 (NK activation control cell line). In addition to K562 positive control, three conditions were tested, as follows:

- > NK cell alone
- > NK cells vs Daudi (CD19+)
- 35 > NK cells vs HUT78 (CD19-)



Effector :Target (E :T) ratio was 2.5 : 1 (50 000 E : 20 000 T), with an antibody dilution range starting to 10 µg/mL with 1/4 dilution (n=8 concentrations). Antibodies, target cells and effector cells were mixed; spun 1 min at 300g; incubated 4h at 37°C; spun 3 min at 500g; washed twice with Staining Buffer (SB); added 50µL of staining Ab mix; incubated 30 min at 300g; washed twice with SB resuspended pellet with CellFix ; stored overnight at 4°C; and fluorescence detected with Canto II (HTS).

### *Results*

#### 1. NK cells alone

Results of these experiments are shown in **Figure 4A**. In the absence of target-antigen expressing cells, none of the bispecific anti-NKp46 x anti-CD19 antibodies (including each of the NKp46-1, NKp46-2, NKp46-3, NKp46-4 and NKp46-9 variable regions) activated NK cells as assessed by CD69 or CD107 expression. The full-length anti-CD19 also did not activate NK cells. However, the full-length anti-NKp46 antibodies did cause detectable activation of NK cells. Alemtuzumab also induced activation of NK cells, at a very high level. The isotype control antibody did not induce activation.

#### 2. NK cells vs Daudi (CD19<sup>+</sup>)

Results of these experiments are shown in **Figure 4B**. In the presence of target-antigen expressing cells, each of the bispecific anti-NKp46 x anti-CD19 antibodies (including each of the NKp46-1, NKp46-2, NKp46-3, NKp46-4 and NKp46-9 binding domains) activated NK cells. The full-length anti-CD19 antibody showed at best only very low activation of NK cells. Neither full-length anti-NKp46 antibodies nor alemtuzumab showed a substantial increase in activation beyond what was observed in presence of NK cells alone. The data in **Figure 4** shows that full-length anti-NKp46 antibodies elicited a similar level of baseline activation as was observed in the presence of NK cells alone. Alemtuzumab also induced the activation of NK cells at a similar level of activation to what was observed in the presence of NK cells alone, and at higher antibody concentrations in this setting (ET 2.5 : 1) the activation was greater than with the bispecific anti-NKp46 x anti-CD19 antibody. The isotype control antibody did not induce activation.

#### 3. NK cells vs HUT78 (CD19<sup>-</sup>)

Results of these experiments are shown in **Figure 4C**. In the presence of target-antigen-negative HUT78 cells, none of the bispecific anti-NKp46 x anti-CD19 antibody (including each of the NKp46-1, NKp46-2, NKp46-3, NKp46-4 and NKp46-9 variable regions) activated NK cells. However, the full-length anti-NKp46 antibodies and alemtuzumab caused detectable activation of NK cells at a similar level observed in presence of NK cells alone. Isotype control antibody did not induce activation.

The foregoing results indicate that the inventive bispecific anti-NKp46 proteins are able to activate NK cells in a target-cell specific manner, unlike full-length monospecific anti-NKp46 antibodies and further unlike full-length antibodies of depleting IgG isotypes which also activate NK cells in the absence of target cells. The NK cell activation achieved with anti-NKp46 bispecific proteins remarkably was higher than that observed with full length anti-CD19 IgG1 antibodies. Therefore these bispecific antibodies should elicit less non-specific cytotoxicity and may be more potent when used in therapy.

#### Example 8:

##### **Comparative efficacy with depleting anti-tumor mAbs: NKp46 x CD19 bispecific proteins at low ET ratio**

These studies aimed to investigate whether bispecific antibodies can mediate NKp46-mediated NK cell activation toward cancer target cells at lower effector:target ratios. The ET ratio used in this Example was 1:1 which is believed to be closer to the setting that would be encountered in vivo than the 2.5:1 ET ratio used in Example 7 or the 10:1 ET ratio of Example 6.

NKp46 x CD19 bispecific proteins having an arrangement according to the F2 format described in Example 3 with anti-NKp46 variable domains from NKp46-1, NKp46-2, NKp46-3, NKp46-4 or NKp46-9 were compared to:

- (a) full-length monospecific anti-NKp46 antibodies (NKp46-3 as human IgG1), and
- (b) the anti-CD19 antibody as a full-length human IgG1 as ADCC inducing antibody control comparator.

The experiments further included as controls: rituximab (an anti-CD20 ADCC inducing antibody control for a target antigen with high expression levels); anti-CD52 antibody alemtuzumab (a human IgG1, binds CD52 target present on both targets and NK cells), and negative control isotype control therapeutic antibody (a human IgG1 that does not bind a target present on the target cells (HUG1-IC). The different proteins were tested for functional effect on NK cell activation as assessed by CD69 or CD107 expression in the presence of CD19-positive tumor target cells (Daudi cells), in the presence of CD19-negative, CD16-positive target cells (HUT78 T-lymphoma cells), and in the absence of target cells. The experiments were carried out as in Example 7 except that the ET ratio was 1:1.

#### *Results*

#### *Results*

The results of the above experiments are shown in **Figure 5 (5A: CD107 and 5B: CD69)**. In the presence of target-antigen expressing cells, each of the bispecific anti-NKp46

x anti-CD19 antibodies (respectively including NKp46-1, NKp46-2, NKp46-3, NKp46-4 or NKp46-9 variable regions) activated NK cells in the presence of Daudi cells.

The activation induced by bispecific anti-NKp46 x anti-CD19 antibody in the presence of Daudi cells was far more potent than that elicited by the full-length human IgG1 anti-CD19 antibody. This ADCC inducing antibody had low activity in this setting. Furthermore, in this low E:T ratio setting the activation induced by the bispecific anti-NKp46 x anti-CD19 antibody was as potent as the anti-CD20 antibody rituximab, with a difference being observed only at the highest concentrations that were 10 fold higher than concentrations in which differences were observed at the 2.5:1 ET ratio.

In the absence of target cells or in the presence of target antigen-negative HUT78 cells, full-length anti-NKp46 antibodies and alemtuzumab showed a similar level of baseline activation as was observed in the presence of Daudi cells. Anti-NKp46 x anti-CD19 antibody did not activate NK cells in presence of HUT78 cells.

The foregoing results indicate that the bispecific anti-NKp46 proteins of the invention are able to activate NK cells in a target-cell specific manner and at lower effector: target ratios and are more effective in mediating NK cell activation than traditional human IgG1 antibodies.

### Example 9:

#### Mechanism of action studies

NKp46 x CD19 bispecific proteins having an arrangement according to the F2, F3, F5 or F6 formats described in Examples 3 or 4 with anti-NKp46 variable domains from NKp46-3 were compared to rituximab (anti-CD20 ADCC inducing antibody), and to a human IgG1 isotype control antibody for their functional ability to direct CD16-/NKp46+ NK cell lines to lyse CD19-positive tumor target cells.

Briefly, the cytolytic activity of the CD16-/NKp46<sup>+</sup> human NK cell line KHYG-1 was assessed in a classical 4-h <sup>51</sup>Cr-release assay in U-bottom 96 well plates. Daudi or B221 cells were labelled with <sup>51</sup>Cr (50 µCi (1.85 MBq)/1 x 10<sup>6</sup> cells), then mixed with KHYG-1 at an effector/target ratio equal to 50:1, in the presence of test antibodies at dilution ranges starting from 10<sup>-7</sup> mol/L with 1/5 dilution (n=8 concentrations).

After a brief centrifugation and 4 hours of incubation at 37°C, 50µL of the supernatant were removed and transferred into a LumaPlate (Perkin Elmer Life Sciences, Boston, MA), and <sup>51</sup>Cr release was measured with a TopCount NXT beta detector (PerkinElmer Life Sciences, Boston, MA). All experimental conditions were analyzed in triplicate, and the percentage of specific lysis was determined as follows: 100 x (mean cpm experimental release - mean cpm spontaneous release)/ (mean cpm total release - mean cpm

spontaneous release). Percentage of total release is obtained by lysis of target cells with 2% Triton X100 (Sigma) and spontaneous release corresponds to target cells in medium (without effectors or Abs).

### Results

The results of the above experiments are shown in **Figure 6A** (KHYG-1 vs Daudi) and **6B** (KHYG-1 vs B221). In the KHYG-1 hNKp46 NK experimental model, each NKp46 x CD19 bispecific protein (Format F2, F3, F5 and F6) induced specific lysis of Daudi or B221 cells by human KHYG-1 hNKp46 NK cell line, while rituximab and the human IgG1 isotype control (IC) antibodies did not.

## Example 10

### Anti-KIR3DL2 bispecific proteins

Bispecific proteins targeting human KIR3DL2 (KIR3DL2 x NKp46 bispecific) were constructed as F6 formats and tested for activity. KIR3DL2 (CD158k; killer cell immunoglobulin like receptor, three Ig domains and long cytoplasmic tail 2) is a disulphide-linked homodimer of three-Ig domain molecules of about 140 kD, described in Pende et al. (1996) J. Exp. Med. 184: 505-518, the disclosure of which is incorporated herein by reference. Several allelic variants have been reported for KIR3DL2 polypeptides, each of these are encompassed by the term KIR3DL2. The amino acid sequence of the mature human KIR3DL2 (allele \*002) is shown in Genbank accession no. AAB52520. Briefly, the cytolytic activity of NK cells from Buffy coat from donors was assessed in a classic 4-h <sup>51</sup>Cr-release assay in U-bottom 96 well plates. HUT78 tumor cells (CTCL) that express KIR3DL2 were labelled with <sup>51</sup>Cr, then mixed with NK cells at an effector/target ratio equal to 10:1 (25 000:2500), in the presence of test antibodies at dilution ranges starting from 10 µg/mL (or 100 µg/mL) with 1/10 dilution (n=8). Assays were in cRPMI, 150 µL final/well, in triplicates.

Results are shown in **Figure 6C**. Despite its Fc domain not binding to CD16 in this format, the F6 protein structure produced as an NKp46 x KIR3DL2 bispecific protein surprisingly exhibited comparable ability to lyse target cells as a known anti-KIR3DL2 human IgG1 antibody that contained the same variable regions and which binds KIR3DL2 bivalently.

## Example 11

### Effect of intrachain domain motion within multimeric proteins

It was theorized by the inventors that the ability of NKp46 bispecific proteins to promote NKp46-mediated lysis of target cells may be affected by the distance between the two antigen binding domains in the bispecific protein which may impact the ability of one or both of the NKp46 antigen binding domain and the antigen binding domain which interacts

with an antigen of interest to interact with their respective targets. Also, it was further theorized that NKp46 mediated lysis of target cells may be impacted by the structure of the two antigen binding domains and/or their respective conformation, freedom of motion or flexibility which may be impacted by the structure of the two antigen binding domains as well as the manner by which they are associated with each other, e.g., by a linker peptide and its particular length and chemical composition. Particularly, it was theorized that a lytic NKp46-target cell synapse may vary as a function of the size and structure of the bispecific protein. Therefore, the inventors posited that bispecific proteins wherein the antigen binding domains are in a format whereby the antigen binding domains more closely mimics or approximates the conformation, spacing and flexibility of the antigen binding domains of

This was theorized because conformational flexibility, notably intrachain domain motion or movement, may for example affect the effective distance between NKp46 and antigen-of-interest binding sites, which in turn might have an effect on the NKp46-target cell synapse and the ability of a multimeric bispecific protein to mediate NKp46-mediated signaling and lysis. Based on these suppositions the inventors evaluated the lytic function of multimeric proteins of different bispecific protein formats and which comprise more or less freedom of motion of the antigen binding domains based on the structure of the antigen binding sites and the specific linkers separating these antigen binding sites.

Specifically, different NKp46 x tumor antigen bispecific proteins of different formats such as the F3, F4, F9, F10 and F11 format that bound different tumor antigens were evaluated for their relative ability to induce NKp46-mediated lysis of tumor target cells by KHYG-1 NK cells (NKp46<sup>+</sup>CD16<sup>-</sup>). F5 and F6 bispecific protein formats have distances between the NKp46 binding site and the antigen of interest binding site that are less than that of full-length antibodies. By contrast bispecific proteins targeting human CD19 (CD19 x NKp46 bispecific) in F9 format have binding sites that are spaced farther apart, similar to distances in the two binding sites in conventional full-length antibodies. Bispecific proteins were therefore constructed as F9 formats and compared to F10 and F11 formats. Structurally speaking, format F9, F10 and F11 are very close to one another, however formats F10 and F11 are characterized by one antigen binding domain with a Fab structure and the other antigen binding domain with a tandem variable domain structure (two variable domains separated by a flexible linker). F10 and F11 therefore have greater intrachain domain motion and/or less local steric hindrance, as well as possibly less distance between binding sites than in the F9 proteins.

The cytolytic activity of the CD16-/NKp46<sup>+</sup> human NK cell line KHYG-1 was assessed in a classical 4-h <sup>51</sup>Cr-release assay in U-bottom 96 well plates. Daudi or B221 cells were labelled with <sup>51</sup>Cr (50 µCi (1.85 MBq)/1 x 10<sup>6</sup> cells), then mixed with KHYG-1 at an

effector/target ratio equal to 50:1, in the presence of test antibodies at dilution range starting from  $10^{-7}$  mol/L with 1/5 dilution (n=8 concentrations). The results showed that formats F10 and F11 were both more potent than format F9 in inducing Daudi cell lysis by NK cells. As noted above F9 format proteins have distances between the NKp46 binding site and the antigen of interest binding site which is similar to full-length antibodies or about 80 Å, and the F10 and F11 proteins comprise a single chain domain connected to the Fc by a flexible linker and have substantially less than 80Å between the antigen binding sites (in the case of F10, about 55 Å).

Based thereon we studied the effects of even further shortened distances between the NKp46 and antigen of interest binding domains using other CD19 x NKp46 bispecific proteins. In these experiments F3, F4 protein formats were selected for comparison with protein formats F10 and F11. Each of these proteins have distances between antigen binding sites of less than 80Å, however, F3 and F4 are shorter than F10 and F11, and F3 and F4 have distances between antigen binding sites that are equivalent to F11 but 25Å less than that of F10. The results of these experiments indicated that the F3, F4, F10 and F11 formats did not significantly differ in their ability to induce Daudi cell lysis by NK cells. These results would suggest that there may be an optimal minimal spacing between the antigen binding domains that improves potency and/or that potency is affected by a combination of the spacing between the antigen binding domains and the flexibility and/or conformation of the antigen binding domains.

### Example 12:

#### Combining NKp46 and CD16 triggering

NKp46 x CD19 bispecific proteins that bind human CD16 having an arrangement according to the F5 format with anti-NKp46 variable domains from NKp46-3 were compared to the same bispecific antibody in a F6 format (which lacks CD16 binding), and to a human IgG1 isotype anti-CD19 antibody, as well as to a human IgG1 isotype control antibody for functional ability to direct purified NK cells to lyse CD19-positive Daudi tumor target cells.

Briefly, the cytolytic activity of fresh human purified NK cells from EFS Buffy Coat was assessed in a classical 4-h  $^{51}\text{Cr}$ -release assay in U-bottom 96 well plates. Daudi or HUT78 cells (negative control cells that do not express CD19) were labelled with  $^{51}\text{Cr}$  and then mixed with NK cells at an effector/target ratio equal to 10:1, in the presence of test antibodies at dilution range starting from 10 µg/ml with 1/10 dilution (n=8 concentrations).

After brief centrifugation and 4 hours of incubation at 37°C, 50µL of supernatant were removed and transferred into a LumaPlate (Perkin Elmer Life Sciences, Boston, MA), and  $^{51}\text{Cr}$  release was measured with a TopCount NXT beta detector (PerkinElmer Life Sciences,

Boston, MA). All experimental conditions were analyzed in triplicate, and the percentage of specific lysis was determined as follows:  $100 \times (\text{mean cpm experimental release} - \text{mean cpm spontaneous release}) / (\text{mean cpm total release} - \text{mean cpm spontaneous release})$ . Percentage of total release is obtained by lysis of target cells with 2% Triton X100 (Sigma) and spontaneous release corresponds to target cells in medium (without effectors or Abs).

The results of these experiments are shown in **Figure 7**. The CD19-F6-NKp46 (bispecific protein in F6 format) whose Fc domain does not bind CD16 due to a N297 substitution was as potent in mediating NK cell lysis of Daudi target cells as the full-length IgG1 anti-CD19 antibody. This result is remarkable especially considering that the control IgG1 anti-CD19 antibody binds CD19 bivalently and further since the anti-CD19 antibody is bound by CD16. The F6 protein was also compared to a protein CD19-F5-NKp46 that was identical to the CD19-F6-NKp46 protein with the exception of an asparagine at Kabat residue 297. Surprisingly, despite the strong NK activation mediated by CD16 triggering by the CD19-F5-NKp46 (F5 format protein) whose Fc domain binds CD16, the F5 format was far more potent in mediating Daudi target cell lysis than the full-length IgG1 anti-CD19 antibody or the F6 format bispecific protein. This would suggest that NKp46 can enhance target cell lysis even when CD16 is triggered. In fact, at comparable levels of target cell lysis, the CD19-F5-NKp46 was at least 1000 times more potent than the full-length anti-CD19 IgG1. These potency results would suggest that the inventive multispecific NKp46 antibodies should be well suited for use in human therapy, e.g., in treating cancer or infectious diseases.

### **Example 13:**

#### **Mechanisms of action of CD16-binding NKp46 x CD19 bispecific**

Lysis of Daudi cells by NKp46 x CD19 bispecific F5 and F6 were compared to a conventional human IgG1 antibody. As a control, lysis was also tested on HUT78 cells that lack CD19; positive control for HUT78 cell lysis was an anti-KIR3DL2 of human IgG1 isotype (HUT78 are KIR3DL2-positive). Cytotoxicity assays were carried out as in Example 10. Flow cytometry staining of NK cell surface markers was carried out as in Example 7.

Results for the cytotoxicity assays are shown in **Figure 8** (Daudi cell in the right hand panel and HUT78 cells in the left hand panel). the CD19-F6-NKp46-3 whose Fc domain does not bind CD16 due to a N297 substitution has as mode of action NKp46 triggering when NK cells encounter the target cell, while the CD19-F5-NKp46-3 bispecific protein demonstrated a far higher potency in mediating cytotoxicity toward Daudi cells. Neither the F5 nor the F6 protein mediated any NK cell cytotoxicity towards HUT78 cells.

The results of flow cytometry staining of NK cell surface markers showed a strong upregulation of CD137 on the surface of NK cells by F5 proteins. These results are shown in

**Figure 9** (Left-most panel: NK cells vs. Daudi; middle panel: NK cells vs. HUT78; right-most panel: NK cells alone). The CD19-F5-NKp46-3 whose Fc domain binds CD16 demonstrated the highest CD137 upregulation. The full-length anti-CD19 IgG1 antibody that binds CD16 also elicited CD137 upregulation, but to a far lesser extent than CD19-F5-NKp46-3. The CD19-F6-NKp46-3 which functions via NKp46 but not via CD16 did not elicit any detectable CD137 upregulation. It is hypothesized that the remarkable potency of the F5 format may arise from a particularly strong CD137 upregulation on NK cells which may be mediated by the dual targeting of NKp46 and CD16.

#### Example 14:

##### Fc-engineered CD16-binding NKp46 x CD20 bispecific

New bispecific proteins were further constructed in an attempt to generate an agent that could improve on the most potent new generation of Fc enhanced antibodies. In these experiments as the comparison antibody we selected the commercial antibody GA101 (GAZYVA®, Gazyvaro®, obinutuzumab, Roche Pharmaceuticals), which is an Fc-modified human IgG1 antibody having enhanced CD16A binding as a result of hypofucosylated N-linked glycosylation.

NKp46 x CD20 bispecific proteins were produced as proteins without CD16 binding (F6 format), with CD16 binding (F5 format), or as Fc-engineered format based on F5 but comprising two amino acid substitutions in the CH2 domain of the heavy chain that increase binding affinity for human CD16A (referred to as "F5+"). In these constructs the anti-CD20 ABDs comprise the V<sub>H</sub> and V<sub>L</sub> of GA101.

Lysis of Daudi cells by NKp46 x CD20 bispecific F5, F5+ and F6 antibodies were compared to the commercial antibody GA101 (GAZYVA®). Cytotoxicity assays were carried out as in Example 10.

Results for the cytotoxicity assays are shown in **Figure 10**. As shown therein the GA101-F5+-NKp46-1 bispecific protein demonstrated a far higher potency (approximately 10-fold increase in EC<sub>50</sub>) in mediating cytotoxicity toward Daudi cells that GA101.

Moreover, when ADCC optimized Fc are used for the bispecific format (F5<sup>+</sup>) a significant difference was observed between F5+-BS lacking the Nkp46 arm (GA101-F5+-IC; black diamond) and F5+-BS co-engaging CD16+NKp46 (GA101-F5<sup>+</sup>-NKp46-1; black square) confirming the contribution of NKp46 in GA101-F5<sup>+</sup>-NKp46-1 activity. Surprisingly, despite the high affinity of GA101-F5<sup>+</sup>-NKp46-1 for CD16 and the presumable maximum NK-cell mediated lysis, NKp46 nevertheless elicited a substantial further increase in cytotoxic activity. These results would suggest that agents capable of inducing ADCC via CD16, can be improved by further conferring on them the ability to induce NKp46-mediated



lysis, and also that the potency of bispecific anti-NKp46 agents can be improved by enhancing affinity for CD16 via Fc engineering.

#### Example 15:

##### Binding of different bispecific formats to FcRn

The affinity of different antibody formats for human FcRn was studied by Surface Plasmon Resonance (SPR) by immobilizing recombinant FcRn proteins covalently to carboxyl groups in the dextran layer on a Sensor Chip CM5, as described in Example 2-6.

A chimeric full length anti-CD19 antibody having intact human IgG1 constant regions and NKp46 x CD19 bispecific proteins having an arrangement according to the F3, F4, F5, F6, F9, F10, F11, F13 or F14 formats described in Examples 3 or 4 with anti-NKp46 variable domains from NKp46-3 (NKp46-2 for F2) were tested; for each analyte, the entire sensorgram was fitted using the steady state or 1:1 SCK binding model.

The results of these experiments are shown in Table 4 below. The bispecific proteins having dimeric Fc domains (formats F5, F6, F13, F14) bound to FcRn with affinity similar to that of the full-length IgG1 antibody. The bispecific proteins with monomeric Fc domains (F3, F4, F9, F10, F11) also displayed binding affinity to FcRn, however with lower affinity than the bispecific proteins having dimeric Fc domains.

Table 4

Antibody/Bispecific	SPR method	KD nM
Human IgG1/K Anti-CD19	SCK / Two state reaction	7.8
CD19-F5-NKp46-3	SCK / Two state reaction	2.6
CD19-F6- NKp46-3	SCK / Two state reaction	6.0
CD19-F13- NKp46-3	SCK / Two state reaction	15.2
CD19-F14- NKp46-3	SCK / Two state reaction	14.0
CD19-F3- NKp46-3	Steady State	474.4
CD19-F4- NKp46-3	Steady State	711.7
CD19-F9A- NKp46-3	Steady State	858.5
CD19-F10A- NKp46-3	Steady State	432.8
CD19-F11- NKp46-3	Steady State	595.5

#### Example 16:

##### Binding to FcγR

Different multimeric Fc proteins were evaluated to assess whether such bispecific monomeric Fc proteins could retain binding to Fc $\gamma$  receptors.

SPR measurements were performed on a Biacore T100 apparatus (Biacore GE Healthcare) at 25°C. In all Biacore experiments HBS-EP+ (Biacore GE Healthcare) and 10 mM NaOH, 500mM NaCl served as running buffer and regeneration buffer respectively. Sensorgrams were analyzed with Biacore T100 Evaluation software. Recombinant human FcR's (CD64, CD32a, CD32b, CD16a and CD16b) were cloned, produced and purified.

F5 and F6 bispecific antibodies CD19-F5-NKp46-3 or CD19-F6-NKp46-3 were immobilized covalently to carboxyl groups in the dextran layer on a Sensor Chip CM5. The chip surface was activated with EDC/NHS (N-ethyl-N'-(3-dimethylaminopropyl) carbodiimidehydrochloride and N-hydroxysuccinimide (Biacore GE Healthcare)). Bispecific antibodies were diluted to 10  $\mu$ g/ml in coupling buffer (10 mM acetate, pH 5.6) and injected until the appropriate immobilization level was reached (i.e. 800 to 900 RU). Deactivation of the remaining activated groups was performed using 100 mM ethanolamine pH 8 (Biacore GE Healthcare).

Monovalent affinity study was assessed following a classical kinetic wizard (as recommended by the manufacturer). Serial dilutions of soluble analytes (FcRs) ranging from 0.7 to 60 nM for CD64 and from 60 to 5000 nM for all the other FcRs were injected over the immobilized bispecific antibodies and allowed to dissociate for 10 min before regeneration. The entire sensorgram sets were fitted using the 1:1 kinetic binding model for CD64 and with the Steady State Affinity model for all the other FcRs.

The results showed that while full length wild type human IgG1 bound to all cynomolgus and human Fc $\gamma$  receptors, the CD19-F6-NKp46-3 bi-specific antibodies did not bind to any of the receptors. The CD19-F5-NKp46-3, on the other hand, bound to each of the human receptors CD64 (KD=0.7 nM), CD32a (KD=846 nM), CD32b (KD=1850 nM), CD16a (KD=1098 nM) and CD16b (KD=2426 nM). Conventional human anti-IgG1 antibodies have comparable binding to these Fc receptors (KD shown in the table below).

Human Fc $\gamma$ receptor	CD19-F5-NKp46-3 KD (nM)	Full length human IgG1 antibody KD (nM)
CD64	0.7	0.24
CD32a	846	379
CD32b	1850	1180
CD16a	1098	630
CD16b	2426	2410

**Example 17:****Improved product profile and yield of different bispecific formats compared to existing formats**

5 Blinatumomab and two bispecific antibodies having NKp46 and CD19 binding regions based on F1 to F17 formats and NKp46-3, and blinatumomab, respectively were cloned and produced under format 6 (F6), DART and BITE formats following the same protocol and using the same expression system. F6, DART and BITE bispecific proteins were purified from cell culture supernatant by affinity chromatography using prot-A beads for F6 or Ni-NTA beads for DART and BITE. Purified proteins were further analyzed and purified by SEC. BITE and DART showed a very low production yield compared to F6 and the purified proteins have a very complex SEC profile. DART and BITE are barely detectable by SDS-PAGE after Coomassie staining in the expected SEC fractions (3 and 4 for BITE and 4 and 5 for DART), whereas the F6 format showed a clear and simple SEC and SDS-PAGE profiles with a major peak (fraction 3) containing the multimeric bispecific proteins. The major peak for the F6 format corresponded to about 30% of the total proteins. These results are consistent for those seen with the F1 to F17 proteins (data not shown) indicating that the Fc domain (or Fc-derived domain) present in those formats facilitates the production and improves the quality and solubility of bispecific proteins.

20 Moreover, the Fc domains present in proteins F1 to F17 have the advantage of being suitable for usage in affinity chromatography without the need for the incorporation of peptide tags. This is desirable as such tags are undesirable in a therapeutic product as they may potentially elicit undesired immunogenicity. By contrast, BiTe and DART antibodies cannot be purified using protein A, whereas F1 to F17 antibodies are all bound by protein A. Table 6 below shows the productivity of different formats.

Format	SEC	SDS PAGE		Final « productivity » yield
		Reduced	Non Reduced	
F5	√	√	√	37mg/L
F6	√	√	√	12mg/L
F7	√	√	√	11mg/L
F8C	√	√	√	3,7mg/L
F9A	√	√	√	8,7mg/L
F9B	√	√	√	3,0mg/L
F10A	√	√	√	2,0mg/L
F11	√	√	√	2,0mg/L

F12	√	√	√	2,8mg/L
F13	√	√	√	6,4mg/L
F14	√	√	√	2,4mg/L
F15	√	√	√	0,9mg/L
BiTe	-	-	-	-
DART	-	-	-	-

All headings and sub-headings are used herein for convenience only and should not be construed as limiting the invention in any way. Any combination of the above-described elements in all possible variations thereof is encompassed by the invention unless otherwise indicated herein or otherwise clearly contradicted by context. Recitation of ranges of values herein are merely intended to serve as a shorthand method of referring individually to each separate value falling within the range, unless otherwise indicated herein, and each separate value is incorporated into the specification as if it were individually recited herein. Unless otherwise stated, all exact values provided herein are representative of corresponding approximate values (e. g., all exact exemplary values provided with respect to a particular factor or measurement can be considered to also provide a corresponding approximate measurement, modified by "about," where appropriate). All methods described herein can be performed in any suitable order unless otherwise indicated herein or otherwise clearly contradicted by context.

The use of any and all examples, or exemplary language (e.g., "such as") provided herein is intended merely to better illuminate the invention and does not pose a limitation on the scope of the invention unless otherwise indicated. No language in the specification should be construed as indicating any element is essential to the practice of the invention unless as much is explicitly stated.

The description herein of any aspect or embodiment of the invention using terms such as reference to an element or elements is intended to provide support for a similar aspect or embodiment of the invention that "consists of," "consists essentially of" or "substantially comprises" that particular element or elements, unless otherwise stated or clearly contradicted by context (e.g., a composition described herein as comprising a particular element should be understood as also describing a composition consisting of that element, unless otherwise stated or clearly contradicted by context).

This invention includes all modifications and equivalents of the subject matter recited in the aspects or claims presented herein to the maximum extent permitted by applicable law.

All publications and patent applications cited in this specification are herein incorporated by reference in their entireties as if each individual publication or patent application were specifically and individually indicated to be incorporated by reference.

5 Although the foregoing invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be readily apparent to one of ordinary skill in the art in light of the teachings of this invention that certain changes and modifications may be made thereto without departing from the spirit or scope of the appended claims.

## CLAIMS

1. A multispecific protein comprising a first and a second polypeptide chain each comprising a variable domain fused to a CH1 or C $\kappa$  domain (a V-(CH1/C $\kappa$ ) unit), in turn fused at its C-terminus to a human Fc domain, wherein the V-(CH1/C $\kappa$ ) unit of the first chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the second chain thereby forming a first antigen binding domain and a dimeric Fc domain, wherein one of the polypeptide chains further comprises an antigen binding domain that forms a second antigen binding domain, and wherein the Fc domain comprises N-linked glycosylation at residue N297 (Kabat EU numbering) and binds to a human CD16 polypeptide.

2. A multispecific protein comprising three polypeptide chains, each comprise a variable domain fused to a CH1 or C $\kappa$  domain (a V-(CH1/C $\kappa$ ) unit), wherein a first (central) chain comprises two V-(CH1/C $\kappa$ ) units and a human Fc domain interposed between the units, the second chain comprises one V-(CH1/C $\kappa$ ) unit and a human Fc domain, and the third chain comprises one V-(CH1/C $\kappa$ ) unit, wherein one of the V-(CH1/C $\kappa$ ) units of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the second chain thereby forming a first antigen binding domain and a dimeric Fc domain, and wherein the other of the V-(CH1/C $\kappa$ ) units of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the third chain thereby forming a second antigen binding domain, and wherein the Fc domain comprises N-linked glycosylation at residue N297 (Kabat EU numbering) and binds to a human CD16 polypeptide.

3. A multispecific protein that binds to three antigens of interest, the protein comprising three polypeptide chains that each comprise a variable domain fused to a CH1 or C $\kappa$  domain (a V-(CH1/C $\kappa$ ) unit), wherein a first (central) chain comprises two V-(CH1/C $\kappa$ ) units and each of the second and third chains comprise one V-(CH1/C $\kappa$ ) unit, wherein one of the V-(CH1/C $\kappa$ ) units of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the second chain thereby forming a first antigen binding domain, wherein the other of the V-(CH1/C $\kappa$ ) units of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the third chain thereby forming a second antigen binding domain, and wherein one of the polypeptide chains further comprises an antigen binding domain that forms a third antigen binding domain.

4. The protein of claim 3, wherein a first (central) chain comprises two V-(CH1/C $\kappa$ ) units and a human Fc domain interposed between the units, and the second chain comprises one V-(CH1/C $\kappa$ ) unit and a human Fc domain, wherein one of the V-(CH1/C $\kappa$ ) units of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the second chain thereby forming a first antigen binding domain and a dimeric Fc domain.

5. A multispecific protein that binds to three antigens of interest and to a human CD16 polypeptide, the protein comprising three polypeptide chains that each comprise a variable domain fused to a CH1 or C $\kappa$  domain (a V-(CH1/C $\kappa$ ) unit), wherein a first (central) chain comprises, from N- to C- terminus, a first V-(CH1/C $\kappa$ ) unit, a human Fc domain and a second V-(CH1/C $\kappa$ ) unit, and a second chain comprise from N- to C- terminus a first V-(CH1/C $\kappa$ ) unit and a human Fc domain, and a third chain comprises a V-(CH1/C $\kappa$ ) unit, and wherein the first V-(CH1/C $\kappa$ ) unit of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the second chain thereby forming a first antigen binding domain and a dimeric Fc domain, wherein the second V-(CH1/C $\kappa$ ) unit of the central chain is bound, by CH1-C $\kappa$  dimerization, to the V-(CH1/C $\kappa$ ) unit of the third chain thereby forming a second antigen binding domain, and wherein the third polypeptide chain further comprises an antigen binding domain that forms a third antigen binding domain.

6. The protein of any of claims 3-5, wherein the Fc domain comprises N-linked glycosylation at residue N297 (Kabat EU numbering) and binds to a human CD16 polypeptide.

7. The protein of any of the above claims, wherein the first variable domain of the first polypeptide chain and the first variable domain of the second polypeptide chain are derived from the same first parental antibody that specifically binds the first antigen, and the second variable domain of the first polypeptide is from a second parental antibody that specifically binds the second antigen.

8. The protein of claim 1, wherein the multispecific protein is a dimer with a dimeric Fc domain, having the domain arrangement:

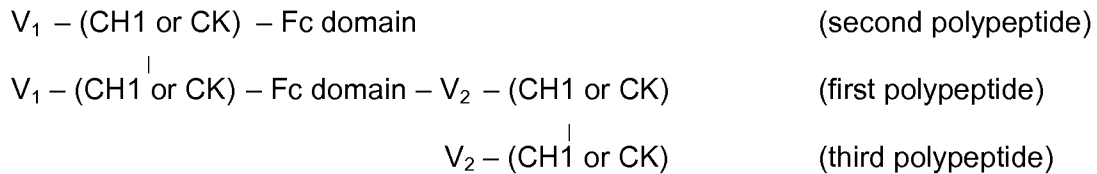
$$V_1 - (\text{CH1 or CK}) - \text{Fc domain} - V_2 - V_2$$

$$V_1 - (\text{CH1 or CK}) - \text{Fc domain}$$

wherein one V<sub>1</sub> is a light chain variable domain and the other V<sub>1</sub> is a heavy chain variable domain, wherein one V<sub>2</sub> is a light chain variable domain and the other V<sub>2</sub> is a heavy chain

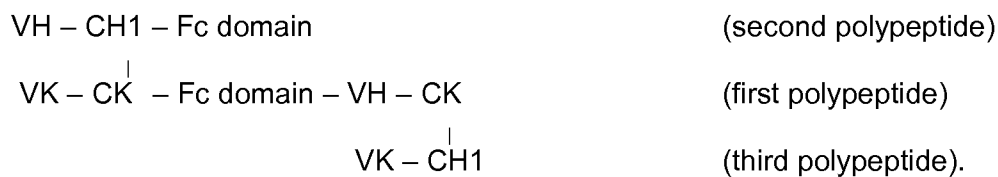
variable domain, wherein the  $V_1$  pair will form a first ABD, and the  $V_2$  pair will form a second ABD.

9. The protein of claims 2 or 6-7, wherein the multispecific protein is a trimer with a dimeric Fc domain, having the domain arrangement:

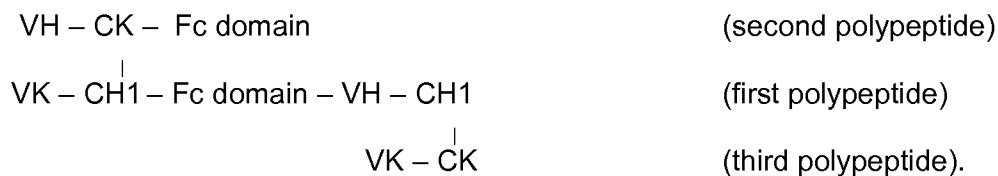


wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, wherein the  $V_1$  pair will form a first ABD, and the  $V_2$  pair will form a second ABD.

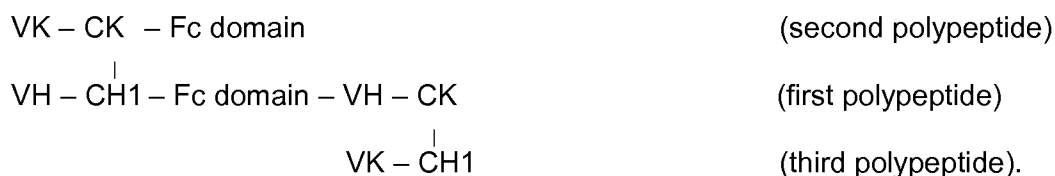
10. The protein of claim 9, having the domain arrangement:



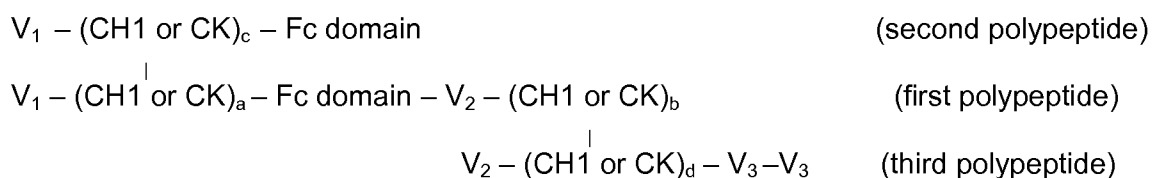
11. The protein of claim 9, having the domain arrangement:



12. The protein of claim 9, having the domain arrangement:



13. The protein of any of claims 3-7, having the domain arrangement:





wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain, wherein the  $V_1$  pair will form a first ABD, the  $V_2$  pair will form a second ABD, and the  $V_3$  will pair to form a third ABD.

14. The protein of claims 3-4 or 6-7, having the domain arrangement:

$V_1 - (\text{CH1 or CK})_c$  (second polypeptide)

$V_1 - (\text{CH1 or CK})_a - \text{Fc domain} - V_2 - (\text{CH1 or CK})_b$  (first polypeptide)

$V_2 - (\text{CH1 or CK})_d - V_3 - V_3$  (third polypeptide)

wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain, wherein the  $V_1$  pair will form a first ABD, the  $V_2$  pair will form a second ABD, and the  $V_3$  will pair to form a third ABD.

15. The protein of claim 3-4 or 6-7, having the domain arrangement:

$V_1 - (\text{CH1 or CK})_c$  (second polypeptide)

$V_1 - (\text{CH1 or CK})_a - \text{Fc domain} - V_2 - (\text{CH1 or CK})_b - V_3 - V_3$  (first polypeptide)

$V_2 - (\text{CH1 or CK})_d$  (third polypeptide).

wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain, wherein the  $V_1$  pair will form a first ABD, the  $V_2$  pair will form a second ABD, and the  $V_3$  will pair to form a third ABD.

16. A multimeric protein that comprises a first ABD that binds a first antigen of interest, a second ABD that binds a second antigen of interest, and a third ABD that binds a third antigen of interest, the protein having the domain arrangement:

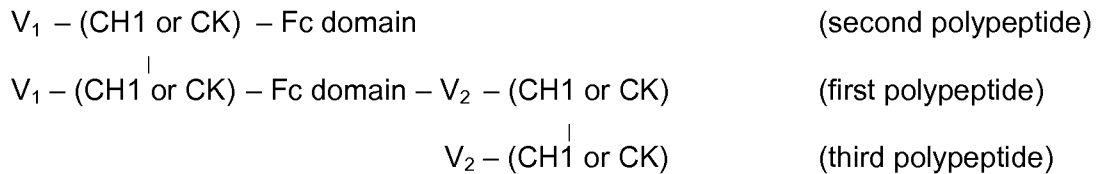
$V_1 - V_1 - \text{Fc domain} - V_2 - (\text{CH1 or CK})$  (first/central polypeptide chain)

$V_2 - (\text{CH1 or CK}) - V_3 - V_3$  (second polypeptide chain)

wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain.

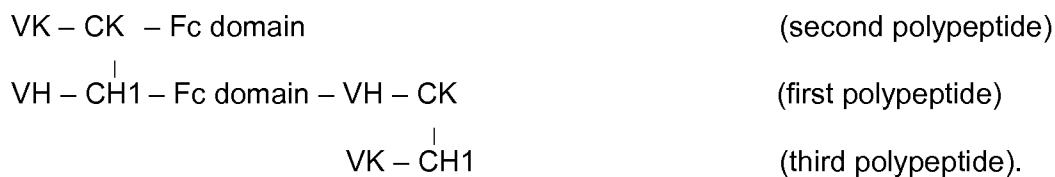
heavy chain variable domain, wherein the  $V_1$  pair will form a first ABD, the  $V_2$  pair will form a second ABD, and the  $V_3$  will pair to form a third ABD.

17. A multimeric protein that comprises a first ABD that binds a first antigen of interest and a second ABD that binds a second antigen of interest, and a dimeric Fc domain having N-linked glycosylation and that binds to human CD16, wherein the protein has the domain arrangement:



wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, wherein the  $V_1$  pair will form a first ABD, and the  $V_2$  pair will form a second ABD.

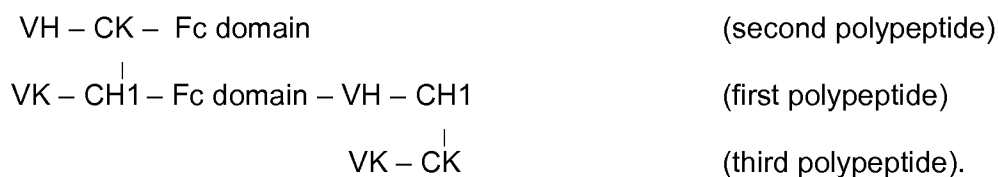
18. The protein of claim 17, having the domain arrangement:



19. The protein of claim 17, having the domain arrangement:



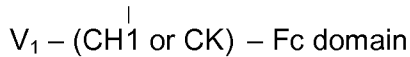
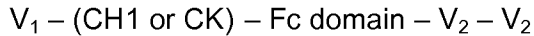
20. The protein of claim 17, having the domain arrangement:



21. The protein of any of the above claims, wherein an Fc domain is fused to an antigen binding domain, CH1 domain and/or CK domain via a hinge region comprising an

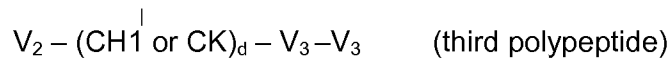
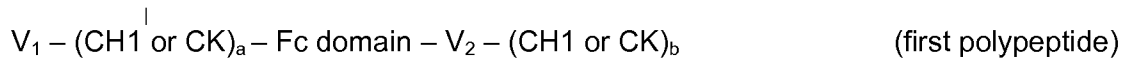
amino acid modification to substitute a cysteine residue in the hinge region by a non-cysteine residue.

22. A multimeric protein that comprises a first ABD that binds a first antigen of interest, a second ABD that binds a second antigen of interest, and a third ABD that binds a third antigen of interest, and a dimeric Fc domain having N-linked glycosylation and that binds to human CD16, wherein the protein has the domain arrangement:



wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, wherein the  $V_1$  pair will form a first ABD, and the  $V_2$  pair will form a second ABD.

23. A multimeric protein that comprises a first ABD that binds a first antigen of interest, a second ABD that binds a second antigen of interest, and a third ABD that binds a third antigen of interest, and a dimeric Fc domain having N-linked glycosylation and that binds to human CD16, wherein the protein has the domain arrangement:



wherein one  $V_1$  is a light chain variable domain and the other  $V_1$  is a heavy chain variable domain, wherein one  $V_2$  is a light chain variable domain and the other  $V_2$  is a heavy chain variable domain, and wherein one  $V_3$  is a light chain variable domain and the other  $V_3$  is a heavy chain variable domain, wherein the  $V_1$  pair will form a first ABD, the  $V_2$  pair will form a second ABD, and the  $V_3$  will pair to form a third ABD.

24. The protein of any one of the above claims, wherein each of the variable domains are single immunoglobulin heavy or light chain variable domains.

25. The protein of any one of the above claims, wherein an Fc domain is fused to an antigen binding domain, CH1 domain and/or CK domain via a hinge region or linker peptide.

26. The protein of any of claims 1-13 or 17-25, wherein the multispecific polypeptide binds to a human Fcγ receptor with an affinity for monovalent binding, as assessed by surface plasmon resonance, that is substantially equivalent to that of a full length wild type human IgG1 antibody.

27. The protein of any of claims 1-13 or 17-26, wherein the protein, immobilized on a surface, binds a soluble human CD16 with a KD for monovalent binding that is no more than 2000 nM, optionally 1300 nM, optionally, 1100 nM, as determined using surface plasmon resonance on Biacore.

28. The protein of any of the above claims, wherein the Fc domain(s) comprises a human CH2 domain comprising an amino acid substitution to increase binding to a human Fcγ receptor.

29. The protein of claims 14-16, wherein the Fc domains comprise a CH3 domain incapable of CH3-CH3 dimerization.

30. The protein of any of claims 14-16, wherein the protein substantially lacks binding to a human Fcγ receptor.

31. The protein of any of the above claims, wherein one ABD binds an activating receptor expressed at the surface of an effector cell, and one ABD binds a cancer, viral or bacterial antigen.

32. The protein of any of the above claims, wherein one ABD binds an activating receptor expressed at the surface of an effector cell, and two ABDs each bind a cancer, viral or bacterial antigen.

33. A protein that comprises: (i) a first antigen binding domain that binds to an activating receptor on an immune effector cell; (ii) a second antigen binding domain that binds to an antigen of interest expressed by a target cell; and (iii) a third antigen binding domain that binds to an antigen of interest other than the activating receptor bound by the first antigen binding domain.

34. The protein of any of the foregoing claims, wherein at least one of the antigen binding sites is configured to have greater intrachain domain movement compared to antigen binding sites in a conventional human IgG1 antibody.

35. The protein of any of the foregoing claims, wherein said protein comprises at least one polypeptide chain that comprises an antigen binding domain bound to a constant domain or Fc domain by a flexible linker, optionally a flexible polypeptide linker.

36. The protein of claims 33-35, wherein the protein is capable of directing an immune effector cell expressing the activating receptor to lyse a target cell expressing the cancer, viral or bacterial antigen, wherein said lysis of the target cell is mediated at least in part by signaling by the activating receptor.

37. The protein of claims 36, wherein the protein does not induce or increase signaling by the activating receptor in an immune effector cell expressing the activating receptor, in the absence of a target cell expressing the cancer, viral or bacterial antigen.

38. The protein of claim 33, wherein the third antigen binding domain binds an antigen of interest expressed by a target cell, wherein the antigen of interest is the same or different from the antigen of interest bound by the second antigen binding domain.

39. The protein of claim 33, wherein the third antigen binding domain binds to an activating receptor on an immune effector cell other than the activating receptor bound by the first antigen binding domain.

40. The protein of any one of claims 33-37, wherein the first antigen binding domain binds human NKp46 polypeptide, the second antigen binding domain binds a cancer antigen, and the third antigen binding domain binds human CD137.

41. The protein of any one of claims 33-40, wherein protein comprises an Fc domain that binds to human CD16A, optionally further wherein none of the antigen binding domains bind to CD16A. In one aspect, the protein has the features or domains arrangement of any embodiment disclosed herein.

42. The protein of any of the above claims, wherein the protein is a multimeric protein of any of claims 1-32.

43. The protein of any of the above claims, wherein at least one ABD binds an a cancer, viral or bacterial antigen that is known to be capable of undergoing inducing or increase in intracellular internalization upon being bound by a full-length human IgG1 antibody.

44. The protein of any of the above claims, wherein the protein does not substantially increase intracellular internalization of an antigen to which it binds on a target cell.

45. A pharmaceutical composition comprising a protein of any one of the above claims, and a pharmaceutically acceptable carrier.

46. Use of a protein or composition of any one of the above claims as a medicament for the treatment of disease.

47. A method of treating a disease in a subject comprising administering to the subject a composition of claims 1-45.

48. The method or use of claims 46-47, wherein the disease is a cancer or an infectious disease.

49. A method of making a heterodimeric or heterotrimeric protein, comprising:

- (a) providing a first nucleic acid encoding a first polypeptide chain according to claims 1-32;
- (b) providing a second nucleic acid encoding a second polypeptide chain according to claims 1-32;
- (c) optionally, providing a third nucleic acid comprising a third polypeptide chain according to claims 1-32; and
- (d) expressing said first and second (and optionally third) nucleic acids in a host cell to produce a protein comprising said first and second (and optionally third) polypeptide chains, respectively; loading the protein produced onto an affinity purification support, optionally a Protein-A support, and recovering a heterotrimeric protein.

50. A method for identifying or evaluating a multimeric polypeptide, comprising the steps of:

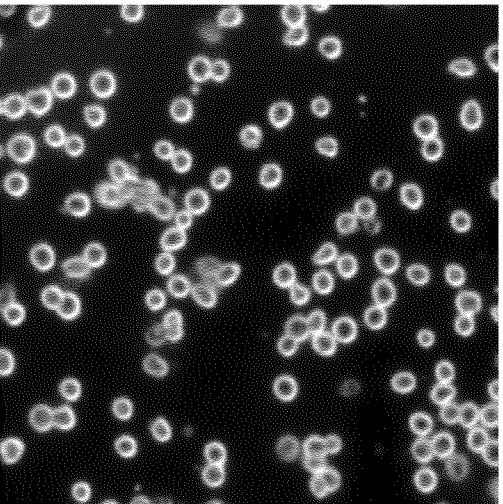
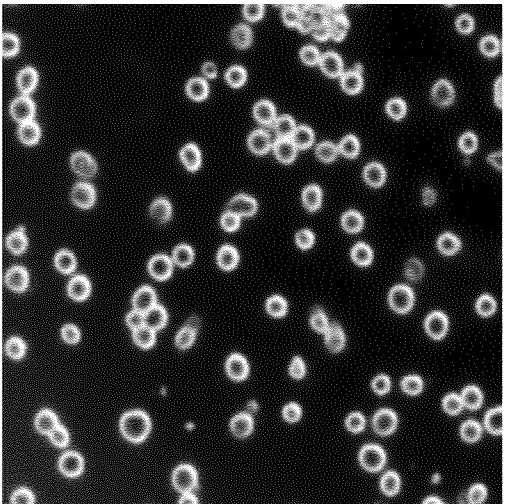
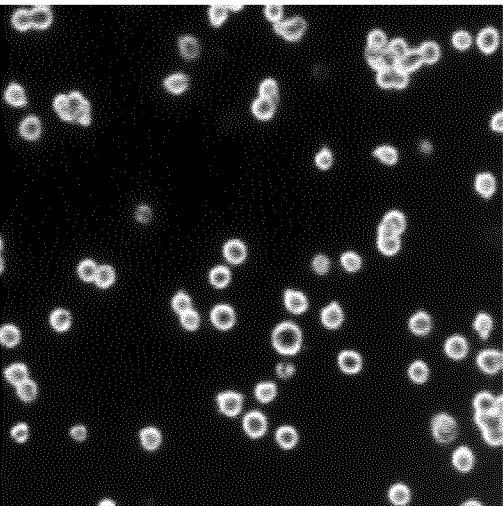
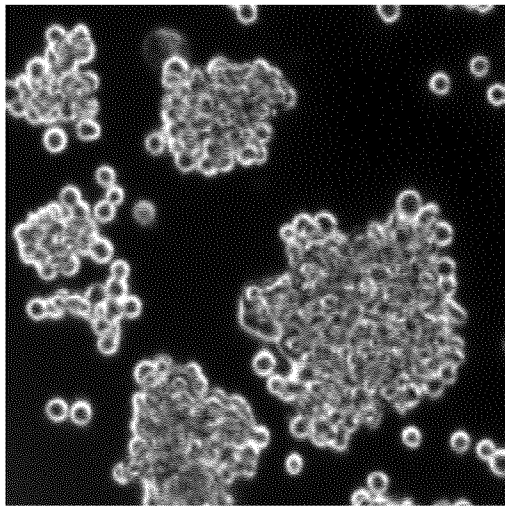
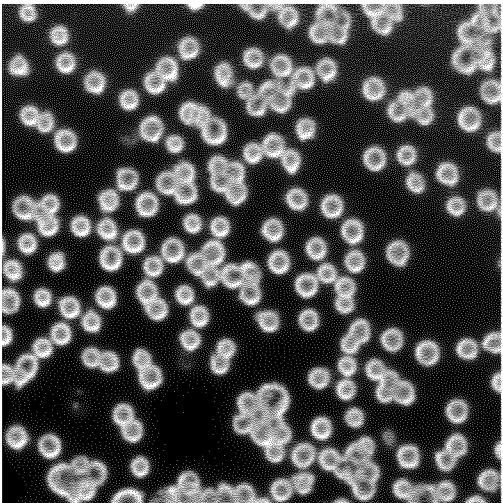
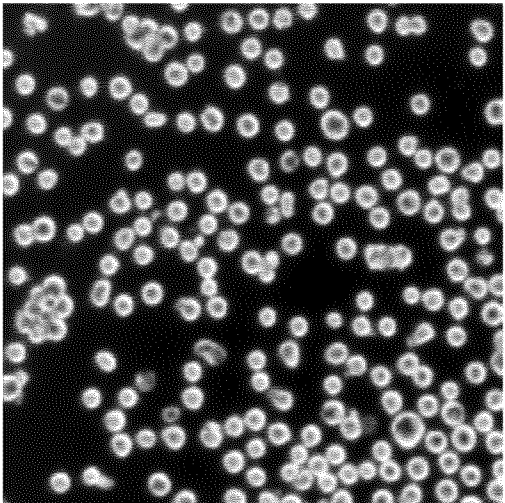
- (a) providing nucleic acids encoding the polypeptide chains of any of claims 1-32;
- (b) expressing said nucleic acids in a host cell to produce said polypeptide chains, respectively; and recovering a multimeric protein comprising said polypeptide chains; and
- (c) evaluating the polypeptide produced for a biological activity of interest.

Figure 1

Jurkat

B221 + Jurkat (50/50)

B221



Anti-CD19-  
monoFc-IgG1-  
Anti-CD3  
(1 µg/ml)

None

Figure 2A

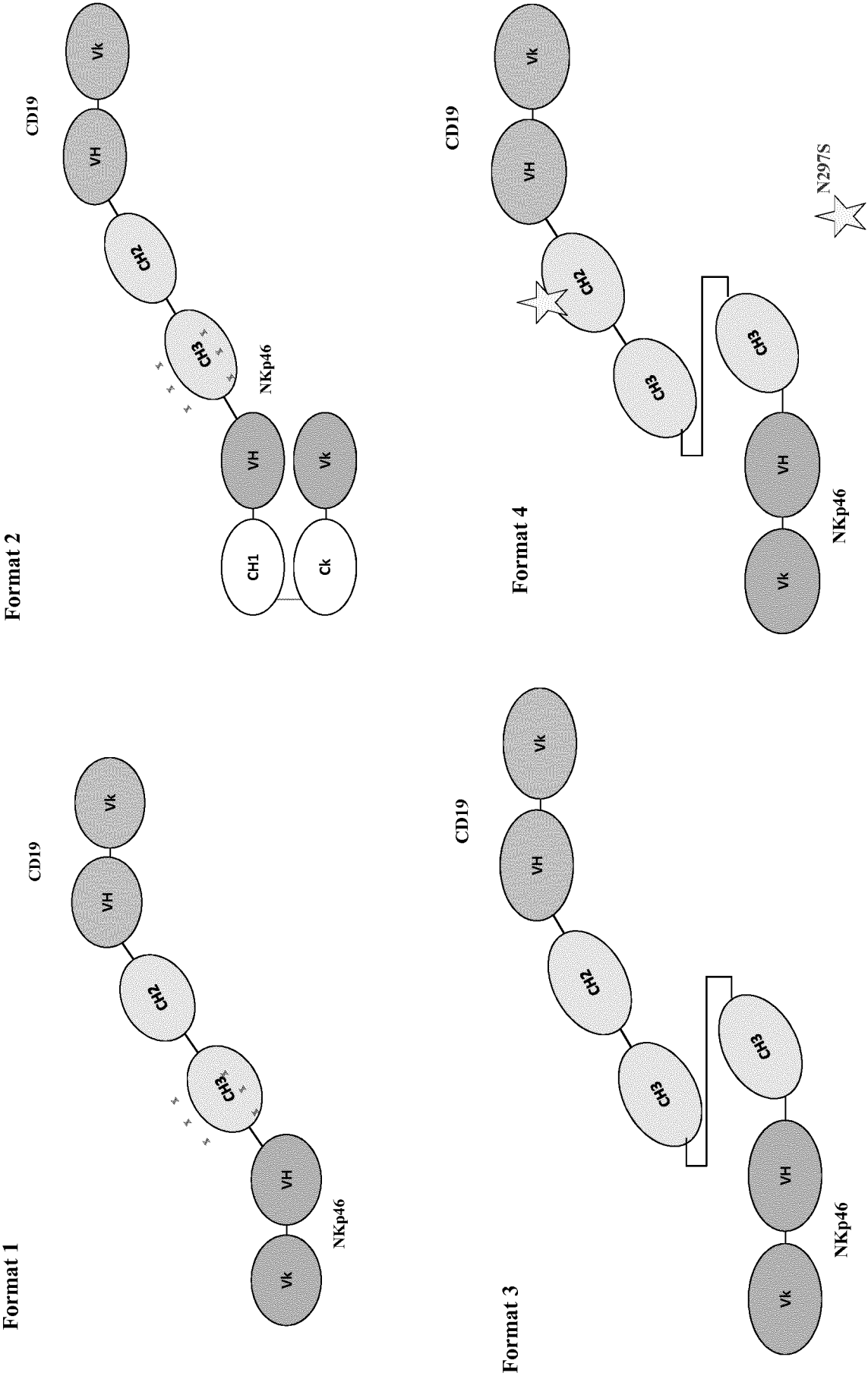
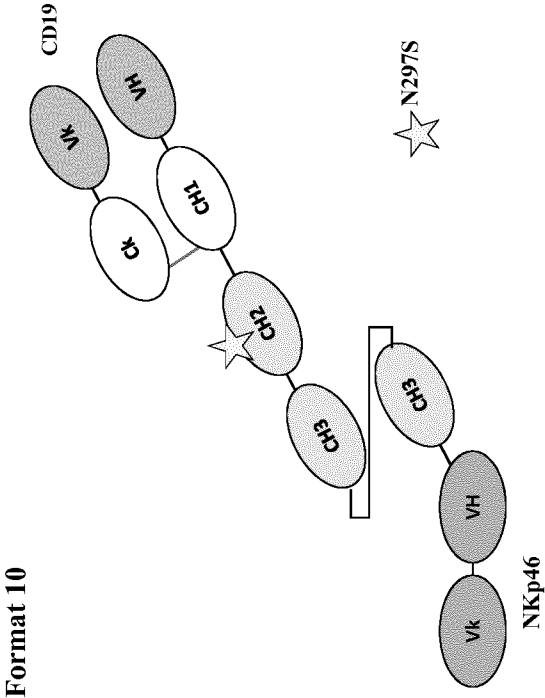
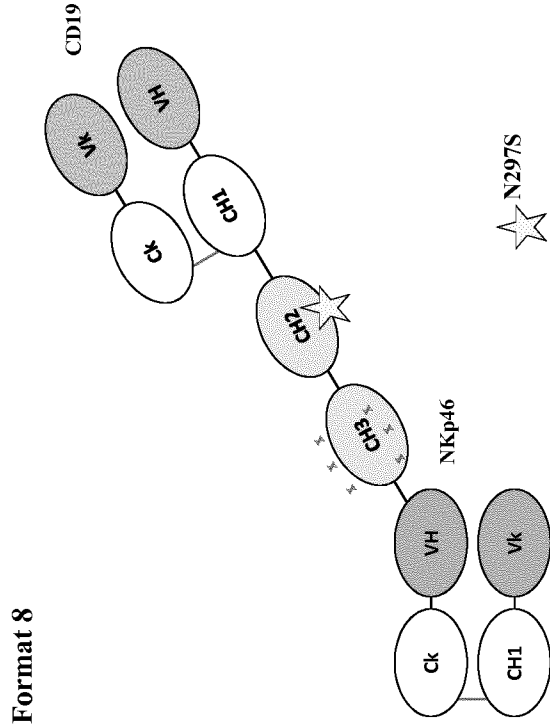
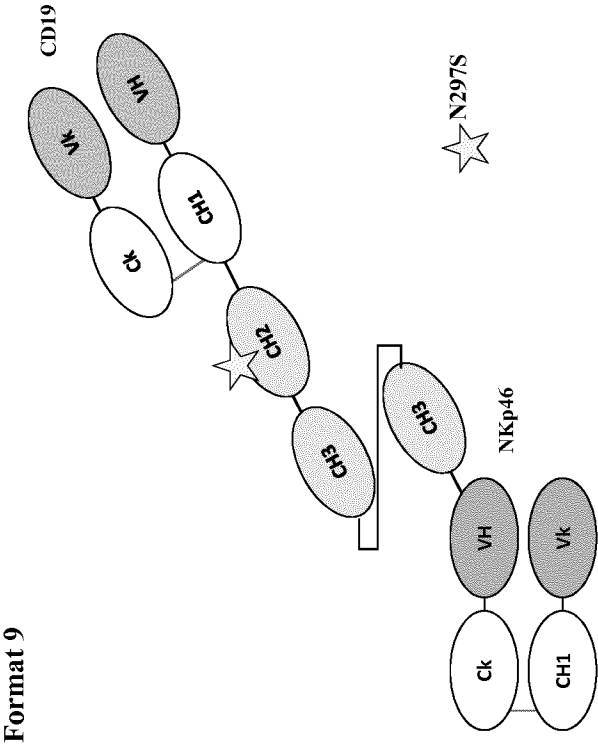




Figure 2B





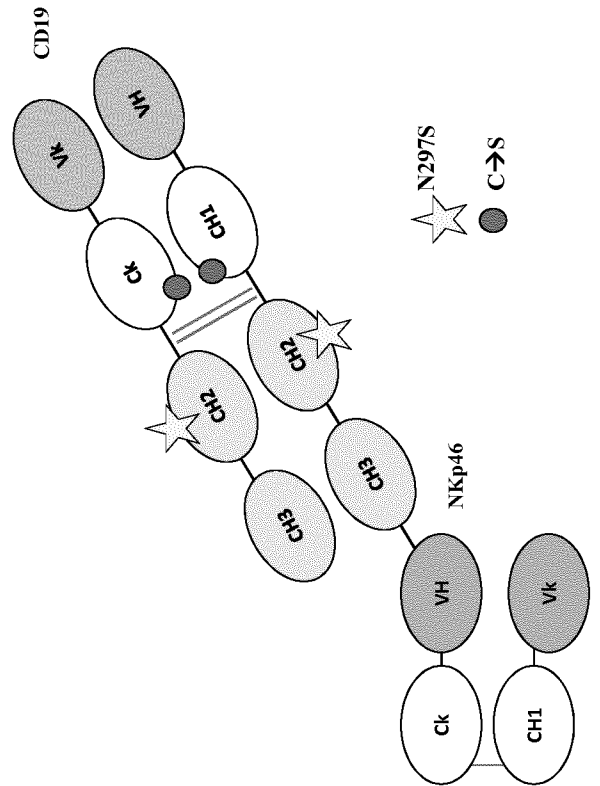
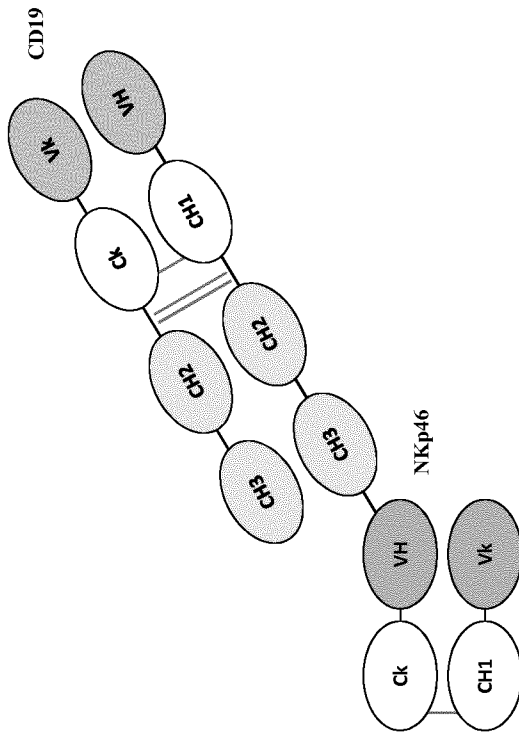
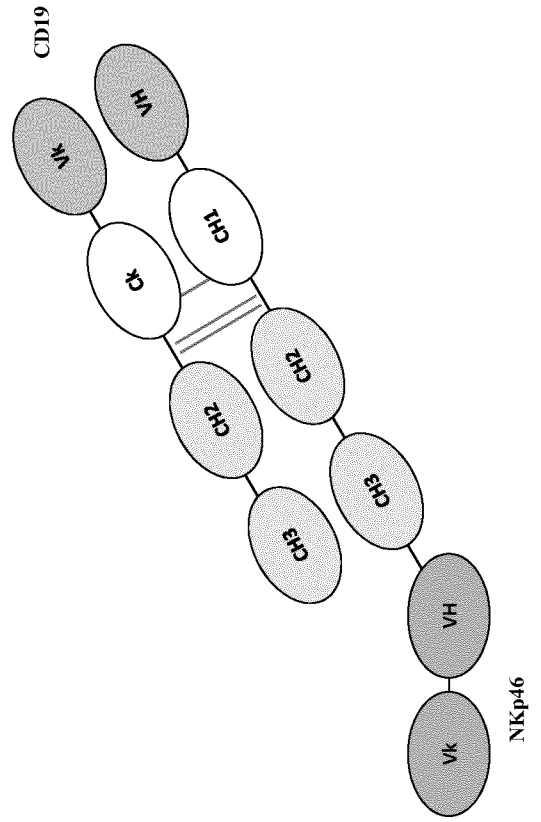
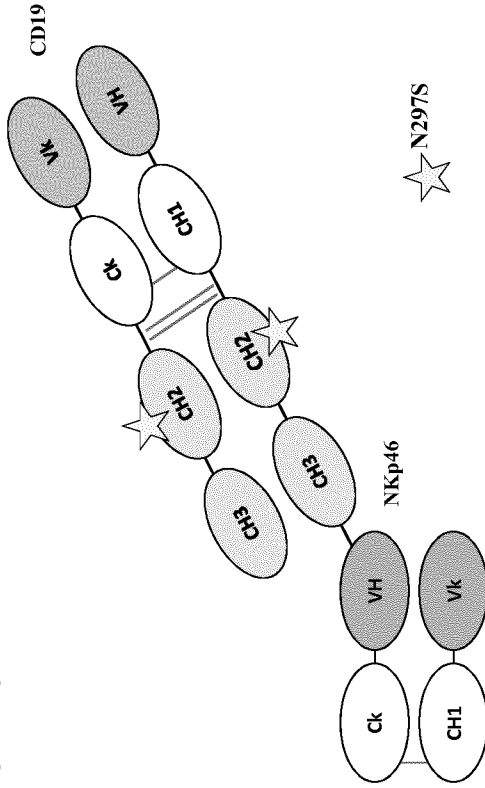
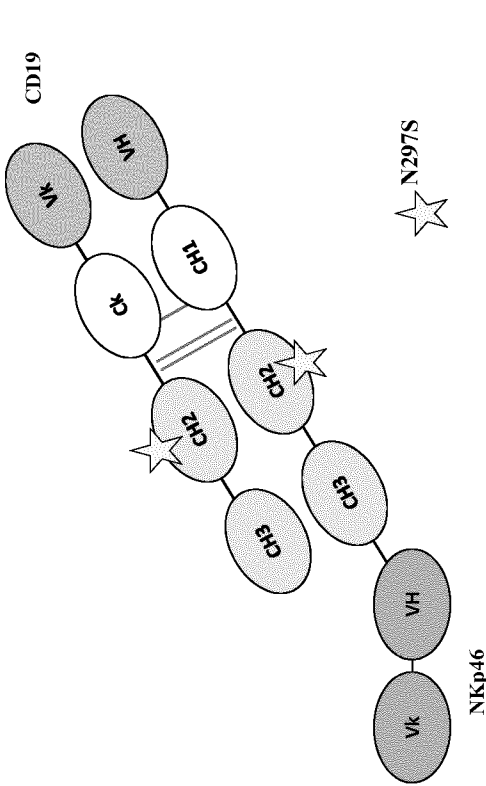
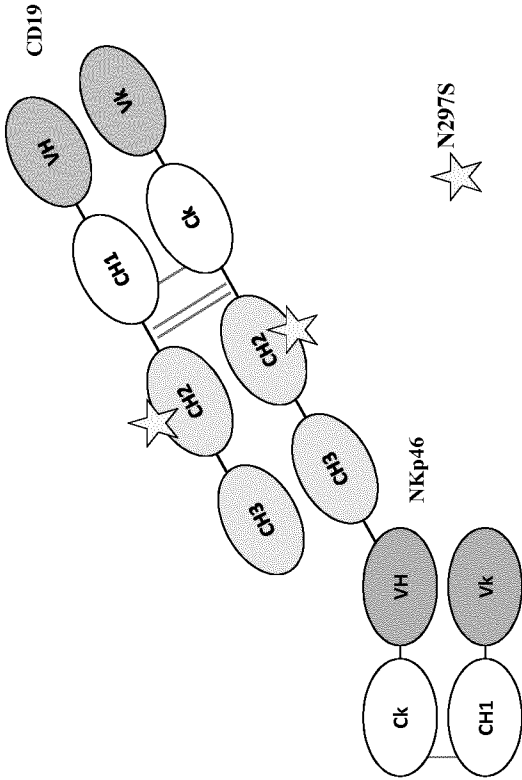


Figure 2E

Format 14



Format 15



Format 16

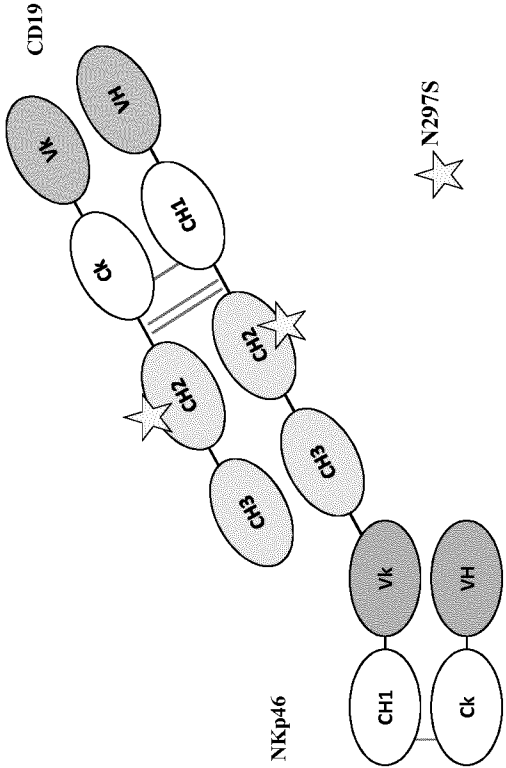


Figure 2F

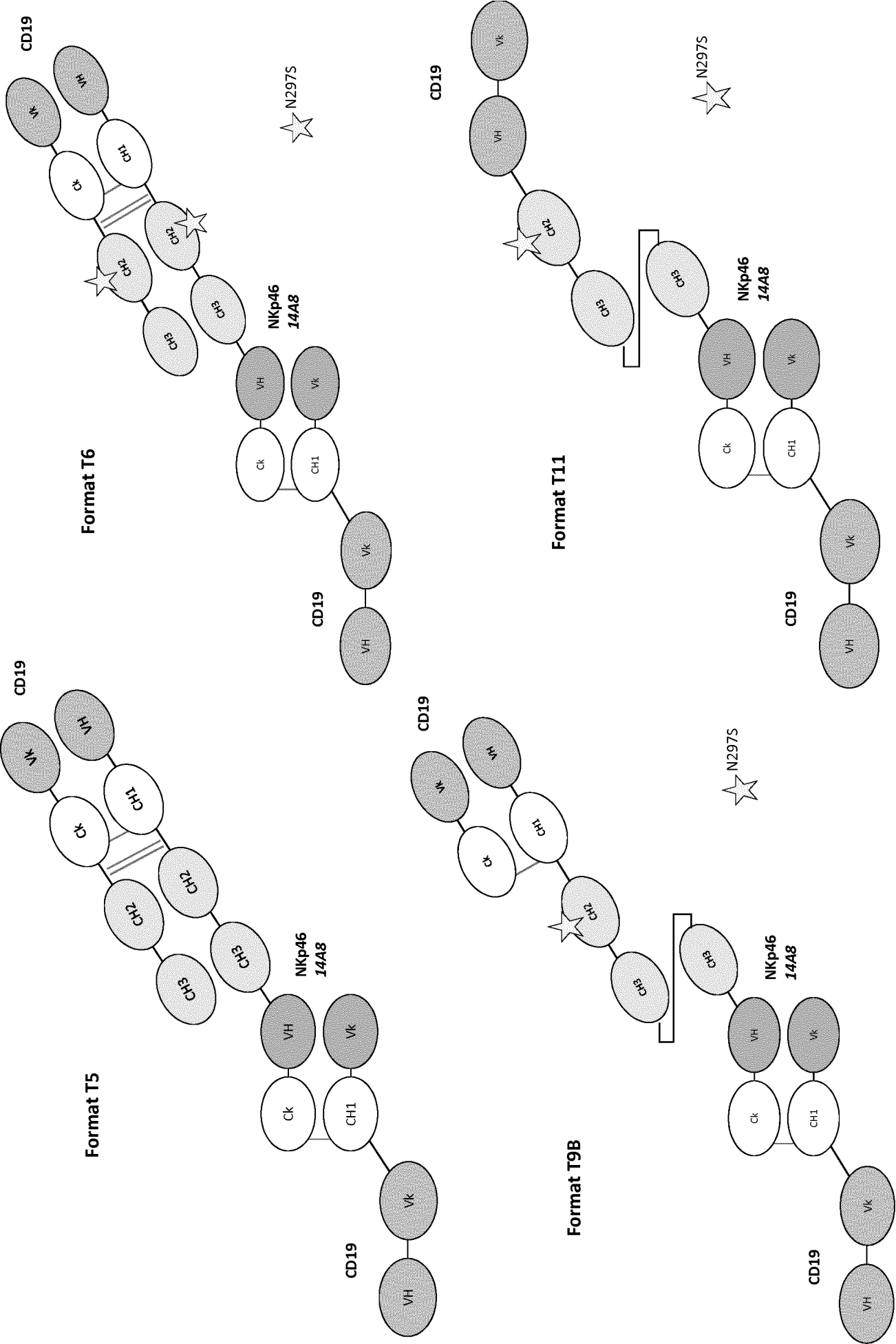


Figure 3A

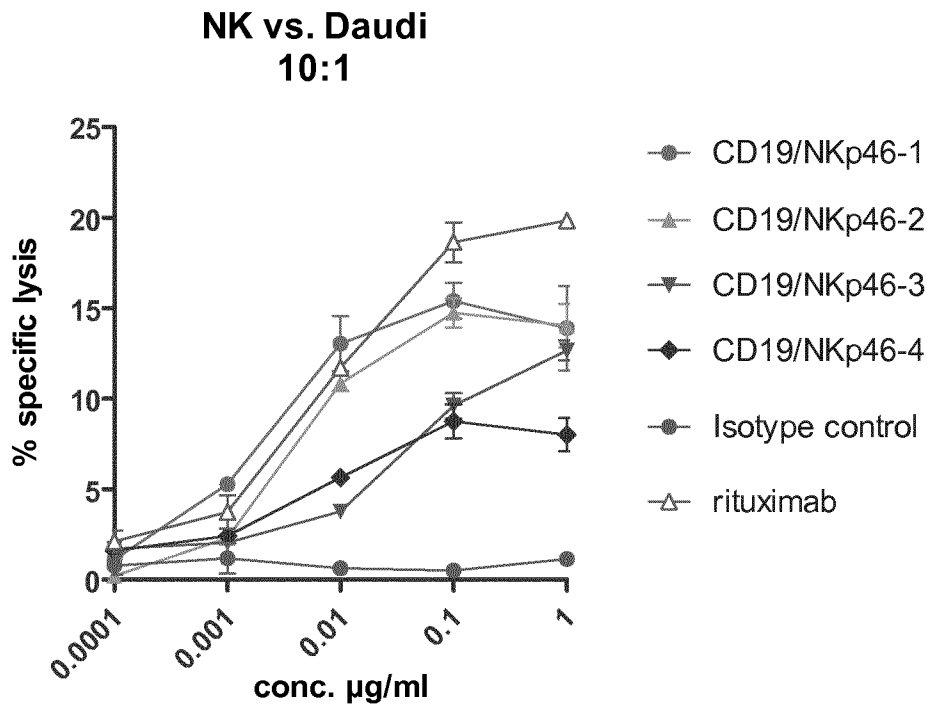


Figure 3B

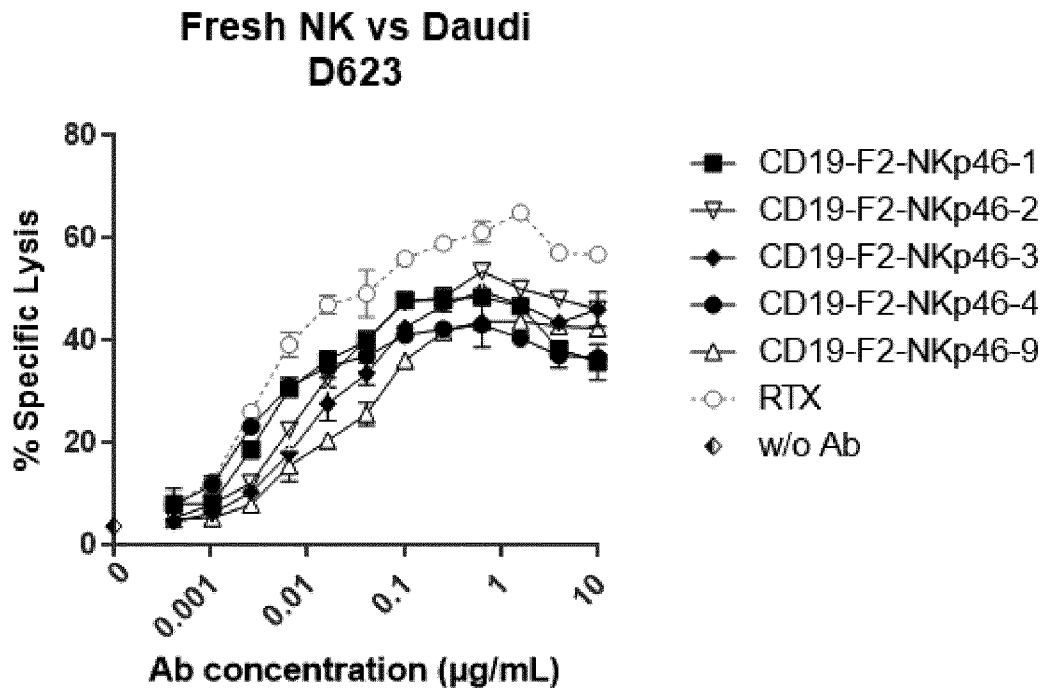


Figure 4A

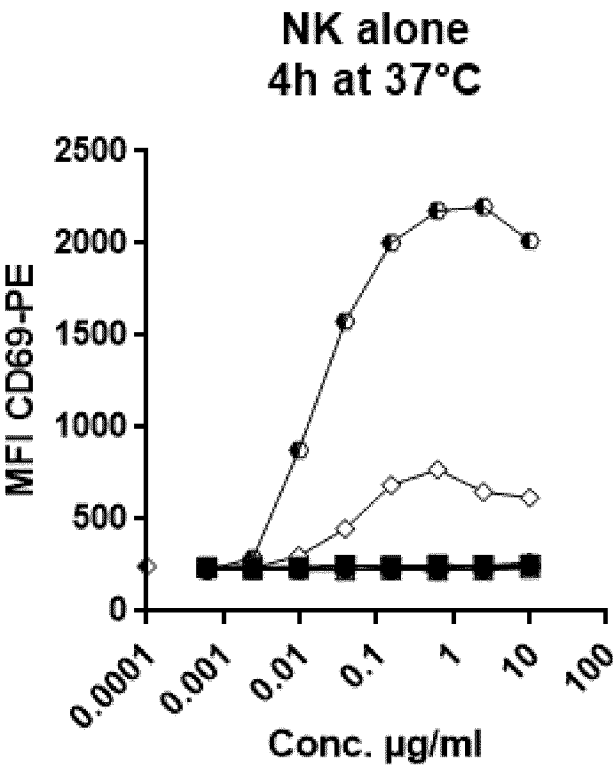
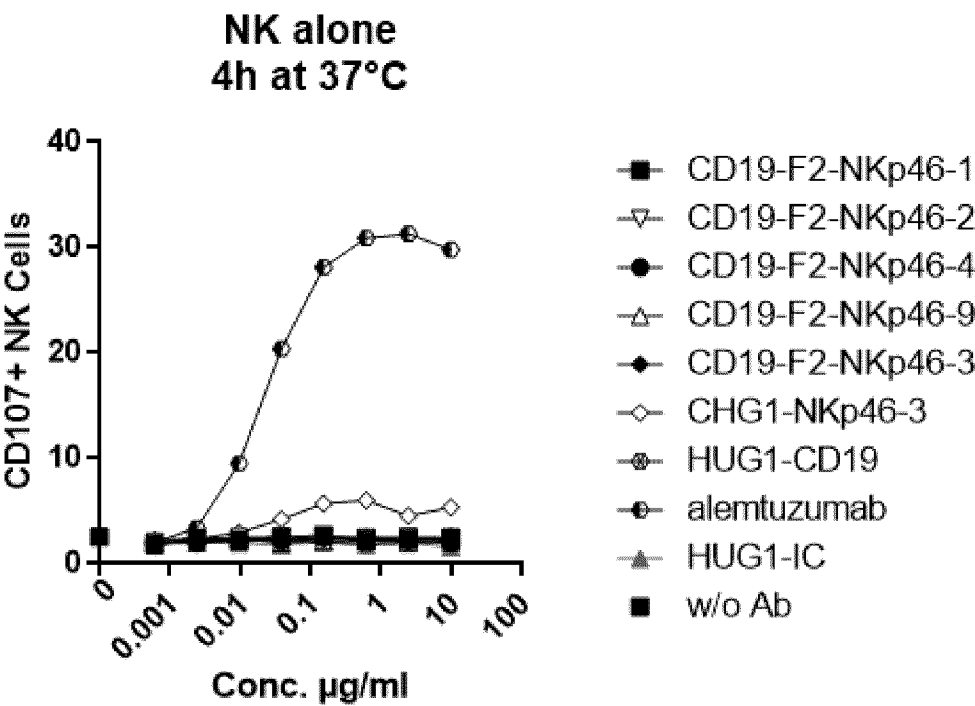


Figure 4B

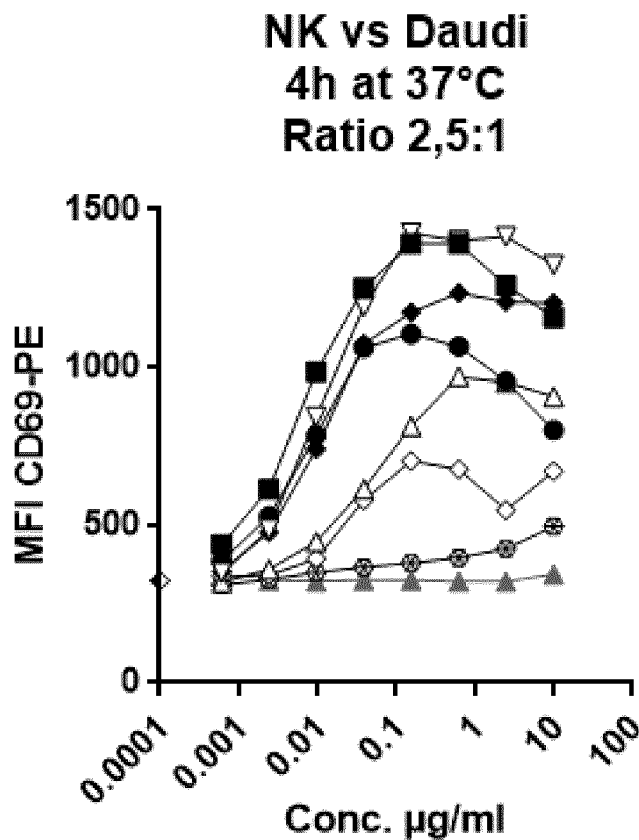
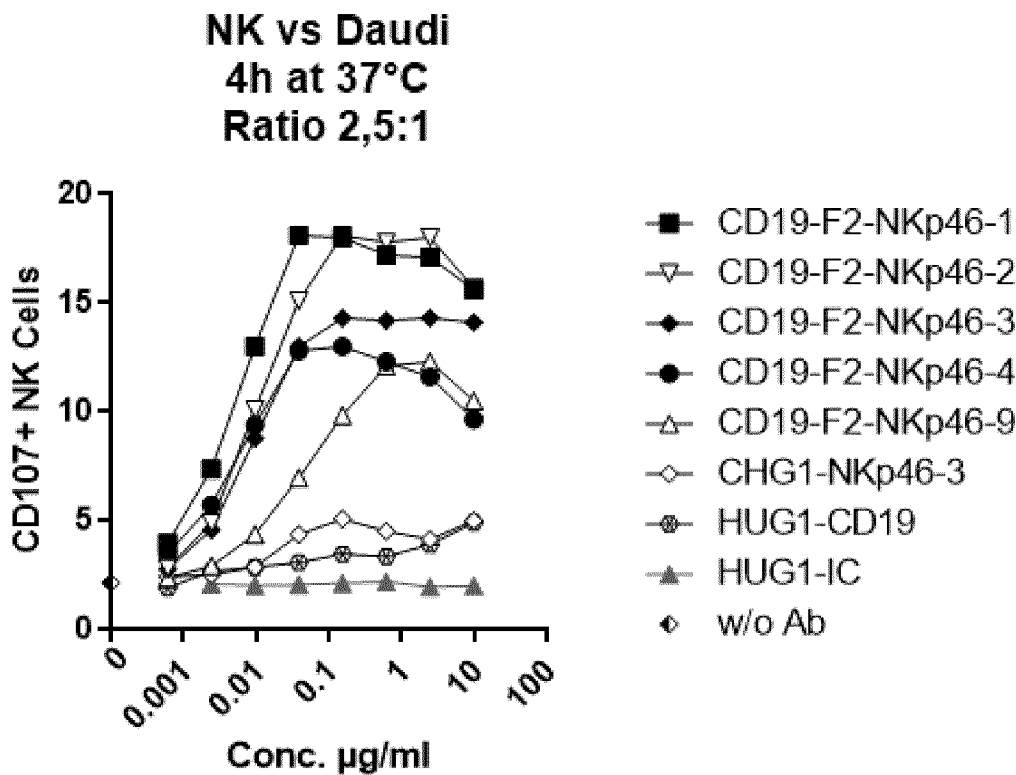






Figure 5A

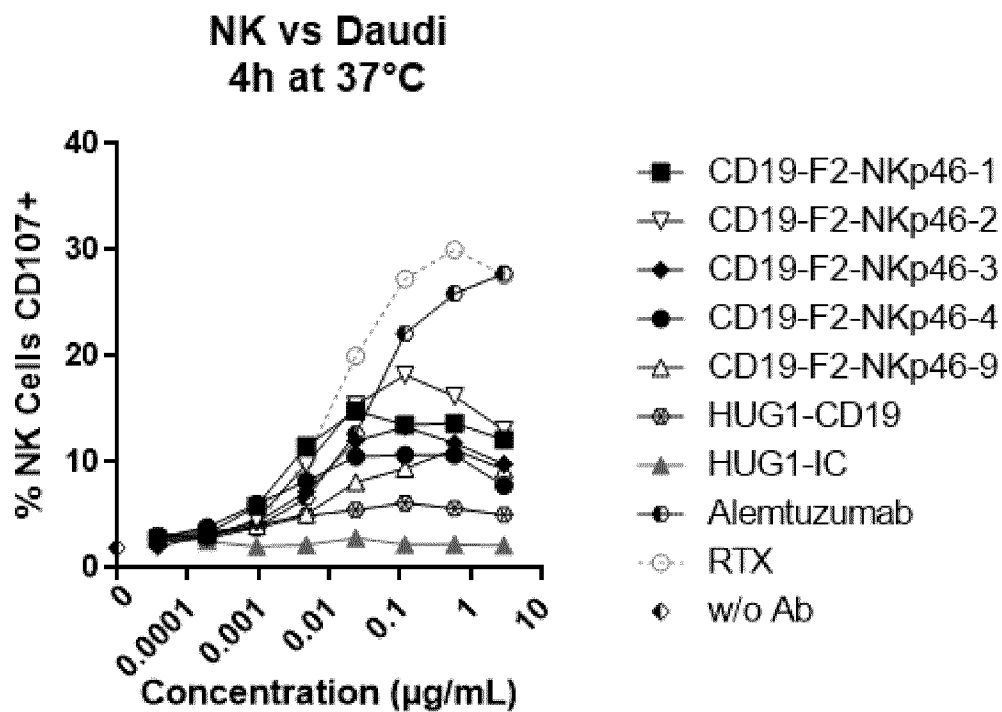


Figure 5B

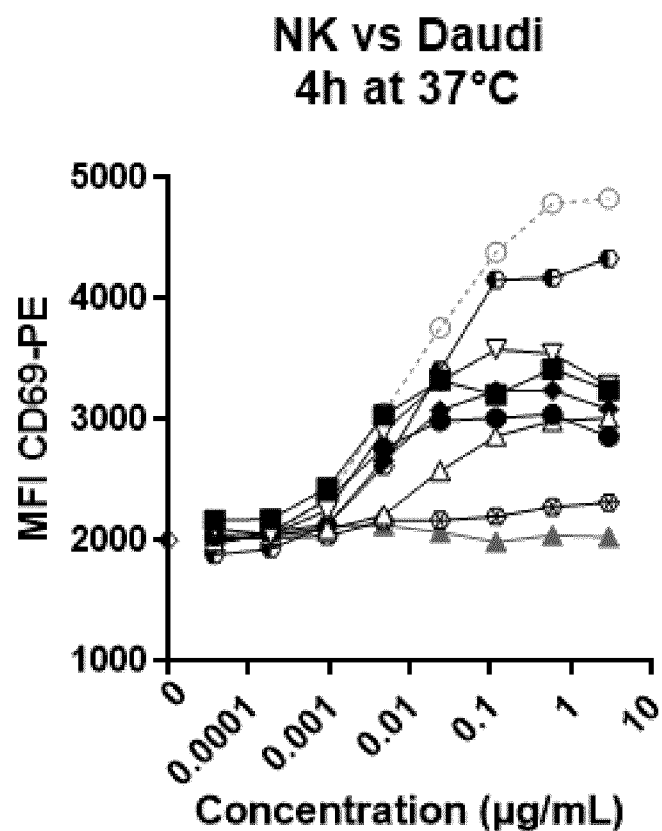


Figure 6A

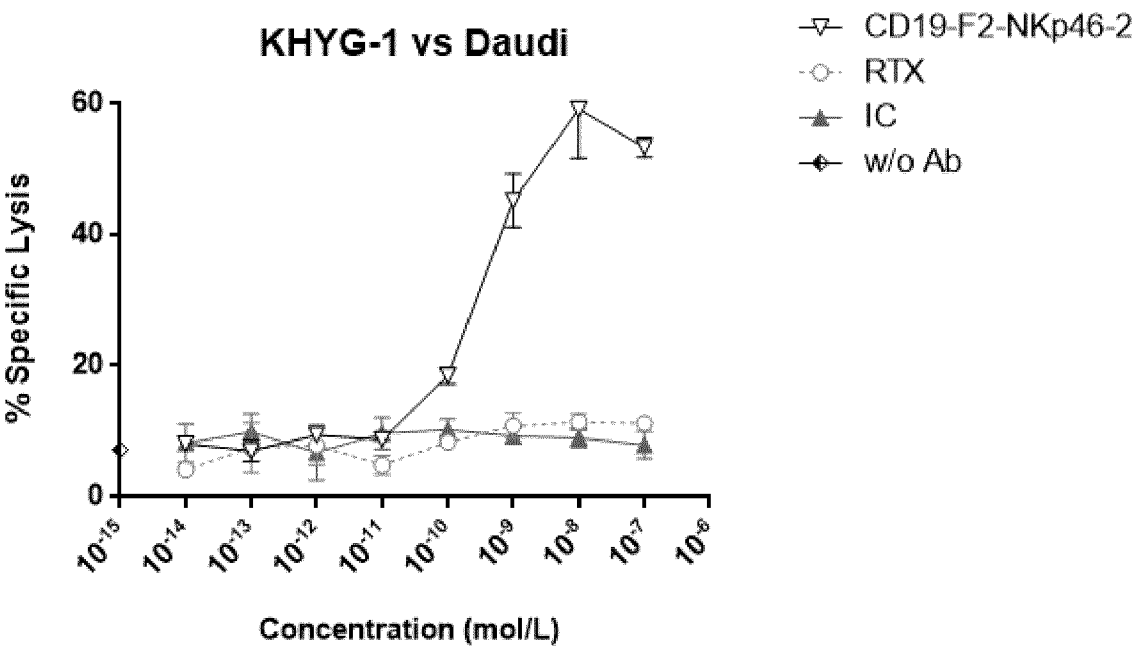


Figure 6B

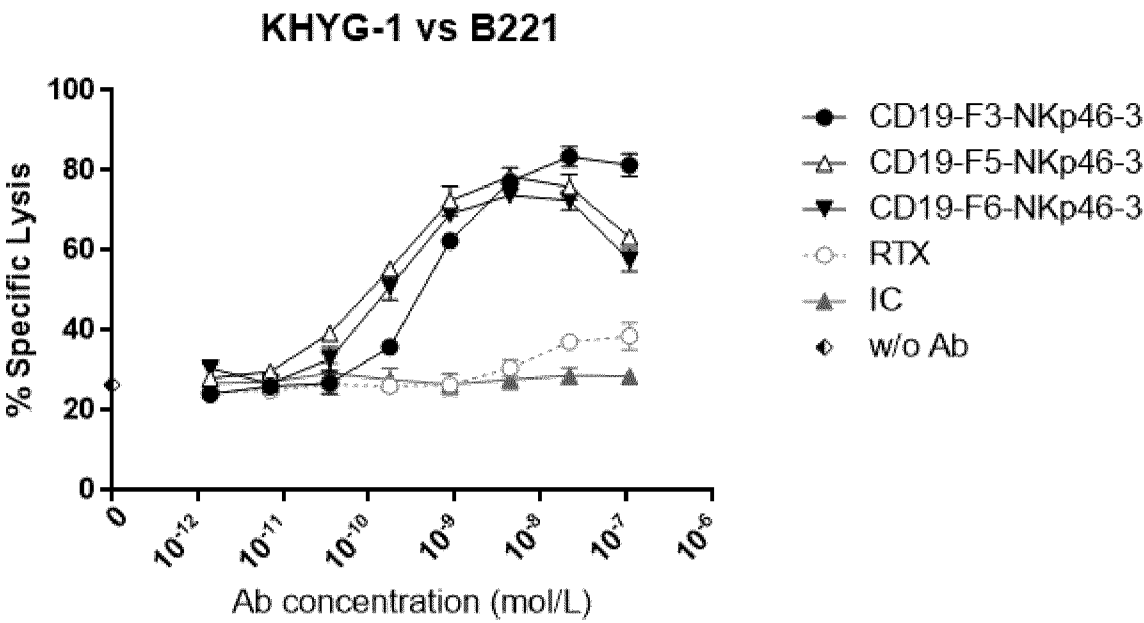


Figure 6C

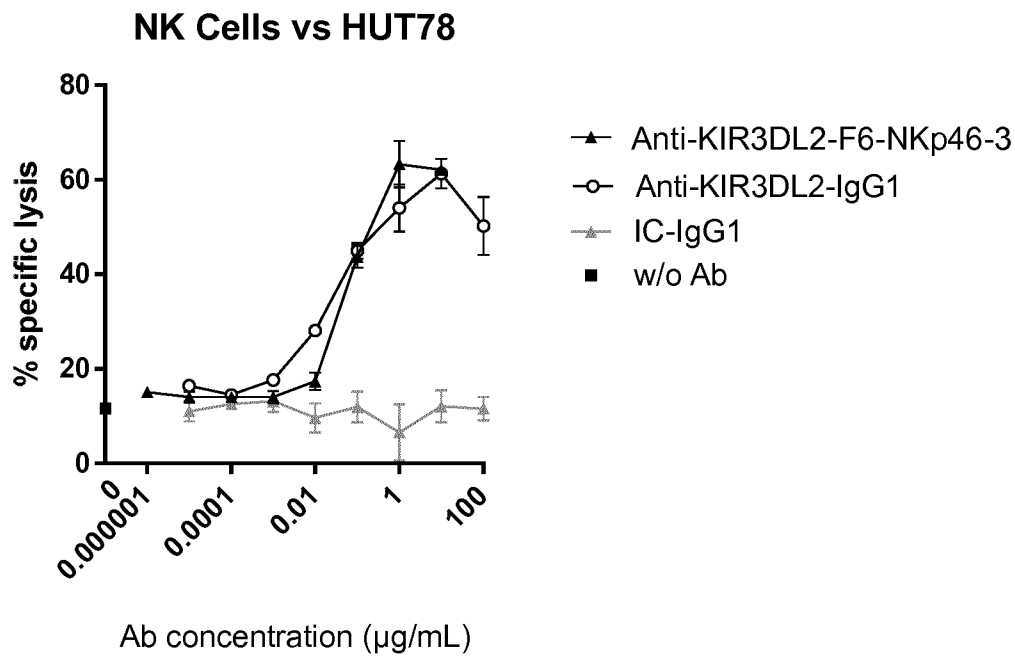


Figure 7

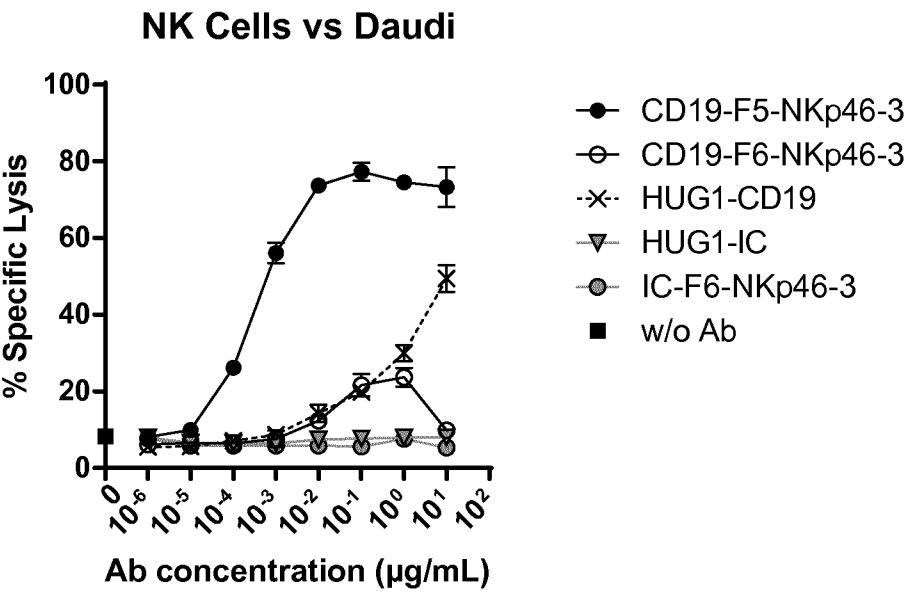


Figure 8

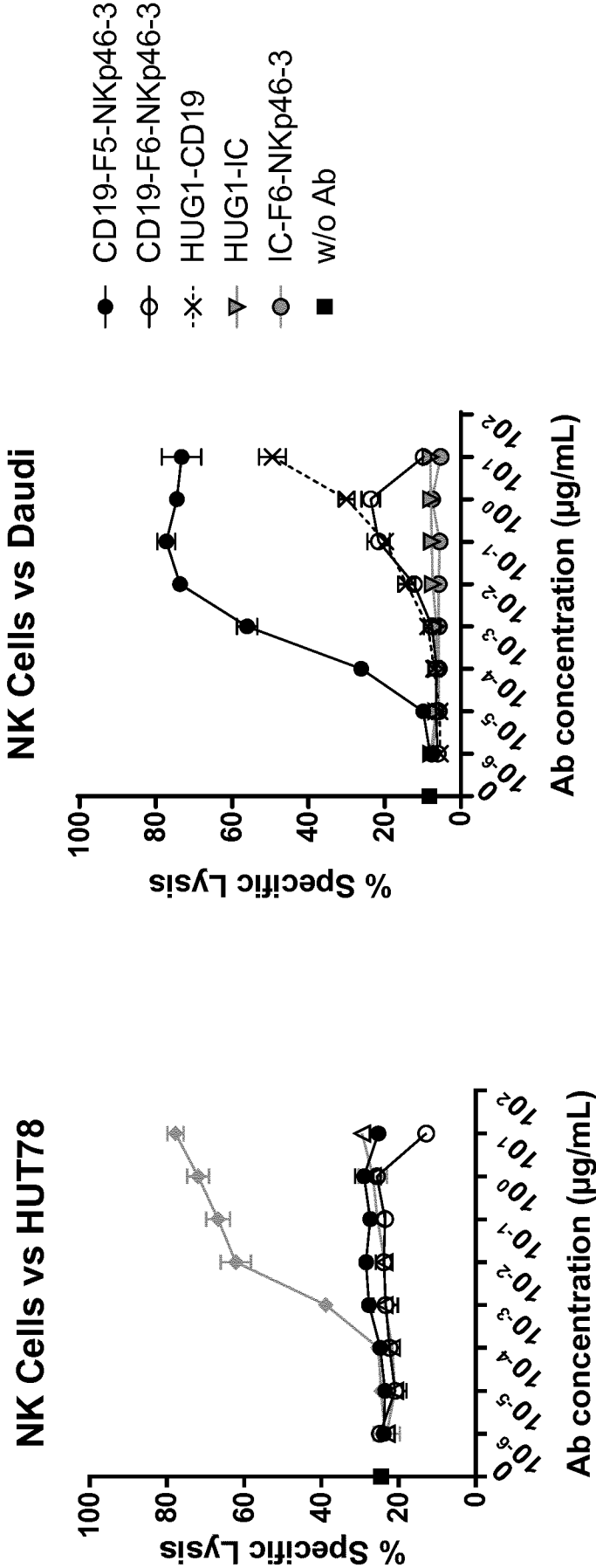
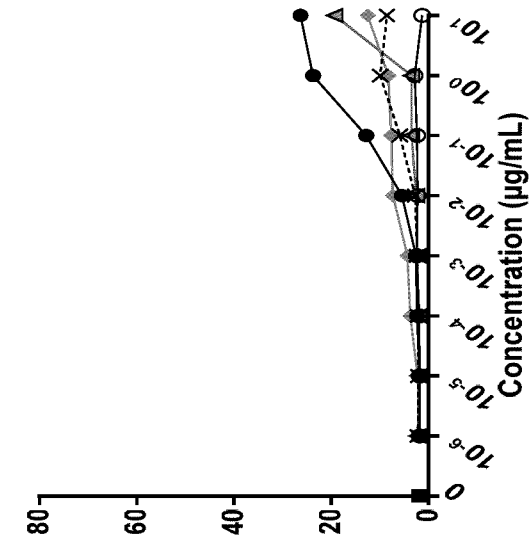


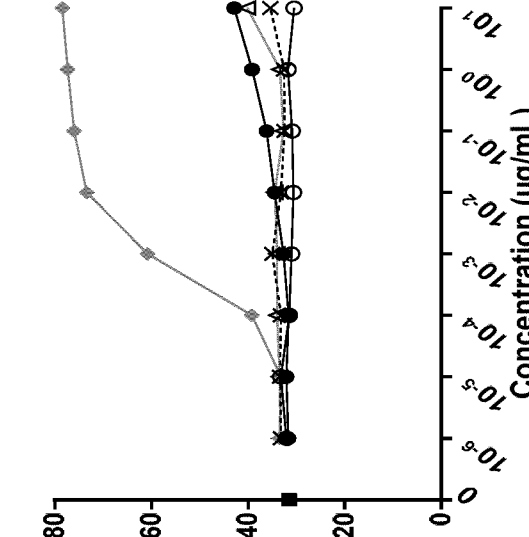
Figure 9

- CD19-F6-NKp46-3
- CD19-F5-NKp46-3
- x- Anti-CD19
- ◆ Anti-KIR3DL2
- △- IC
- w/o Ab

NK alone



NK vs HUT78



NK vs Daudi

