The invention provides an oligonucleotide comprising an inosine, and/or a nucleotide containing a base able to form a wobble base pair or a functional equivalent thereof, wherein the oligonucleotide, or a functional equivalent thereof, comprises a sequence which is complementary to at least part of a dystrophin pre-m RNA exon or at least part of a non-exon region of a dystrophin pre-m RNA said part being a contiguous stretch comprising at least 8 nucleotides. The invention further provides the use of said oligonucleotide for preventing or treating DMD or BMD.
Fig 1
Fig 2

<table>
<thead>
<tr>
<th>AON ID</th>
<th>Sequence (5' – 3')</th>
<th>SEQ ID NO</th>
</tr>
</thead>
<tbody>
<tr>
<td>PS220</td>
<td>UUUGGCCGCUGCCCAUGCCAUCUG</td>
<td>76</td>
</tr>
<tr>
<td>PS305</td>
<td>UUUGCCICUGCCCAUGCCAUCUG</td>
<td>557</td>
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<tr>
<td>PS309</td>
<td>UCAAUGUCUGACAACAGUUUGCCCI</td>
<td>based on 115</td>
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<tr>
<td>PS310</td>
<td>CAAUGUCUGACAACAGUUUGCCIC</td>
<td>based on 116</td>
</tr>
<tr>
<td>PS311</td>
<td>AAUGUUCUGACAACAGUUUGCCICU</td>
<td>based on 117</td>
</tr>
<tr>
<td>PS312</td>
<td>AUGUUCUGACAACAGUUUGCCICU</td>
<td>based on 118</td>
</tr>
<tr>
<td>PS313</td>
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<td>based on 119</td>
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<td>PS314</td>
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</tr>
<tr>
<td>PS316</td>
<td>UCUGACAACAGUUUGCCICUGCCCA</td>
<td>based on 122</td>
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</table>
OLIGONUCLEOTIDE COMPRISING AN INOSINE FOR TREATING DMD

FIELD OF THE INVENTION

[0001] The invention relates to the fields of molecular biology and medicine.

BACKGROUND OF THE INVENTION

[0002] A muscle disorder is a disease that usually has a significant impact on the life of an individual. A muscle disorder can have either a genetic cause or a non-genetic cause. An important group of muscle diseases with a genetic cause are Becker Muscular Dystrophy (BMD) and Duchenne Muscular Dystrophy (DMD). These disorders are caused by defects in a gene for a muscle protein.

[0003] Becker Muscular Dystrophy and Duchenne Muscular Dystrophy are genetic muscular dystrophies with a relatively high incidence. In both Duchenne and Becker muscular dystrophy the muscle protein dystrophin is affected. In Duchenne dystrophy is absent, whereas in Becker some dystrophin is present but its production is most often not sufficient and/or the dystrophin present is abnormally formed. Both diseases are associated with recessive X-linked inheritance. DMD results from a frameshift mutation in the DMD gene. The frameshift in the DMD gene’s transcript (mRNA) results in the production of a truncated non-functional dystrophin protein, resulting in progressive muscle wasting and weakness. BMD occurs as a mutation does not cause a frame-shift in the DMD transcript (mRNA). As in BMD some partly to largely functional dystrophin is present in contrast to DMD where dystrophin is absent, BMD has generally less severe symptoms than DMD. The onset of DMD is earlier than BMD. DMD usually manifests itself in early childhood, in BMD in the teens or in early adulthood. The progression of BMD is slower and less predictable than DMD. Patients with BMD can survive into mid to late adulthood. Patients with DMD rarely survive beyond their thirties.

[0004] Dystrophin plays an important structural role in the muscle fiber, connecting the extracellular matrix and the cytoskeleton. The N-terminal region binds actin, whereas the C-terminal end is part of the dystrophin glycoprotein complex (DGC), which spans the sarcolemma. In the absence of dystrophin, mechanical stress leads to sarcolemmal ruptures, causing an uncontrolled influx of calcium into the muscle fiber interior, thereby triggering calcium-activated proteases and fiber necrosis.

[0005] For most genetic muscular dystrophies no clinically applicable and effective therapies are currently available. Exon skipping techniques are nowadays explored in order to combat genetic muscular dystrophies. Promising results have recently been reported by us and others on a genetic therapy aimed at restoring the reading frame of the dystrophin pre-mRNA in cells from the mdx mouse, the GRMD dog (reference 59) and DMD patients1-11. By the targeted skipping of a specific exon, a DMD phenotype (lacking dystrophin) is converted into a milder BMD phenotype (partly to largely functional dystrophin). The skipping of an exon is preferably induced by the binding of antisense oligoribonucleotides (AONs) targeting either one or both of the splice sites, or exon-internal sequences. Since an exon will only be included in the mRNA when both the splice sites are recognised by the spliceosome complex, splice sites have been considered obvious targets for AONs. More preferably, one or more AONs are used which are specific for at least part of one or more exonic sequences involved in correct splicing of the exon. Using exon-internal AONs specific for an exon 46 sequence, we were previously able to modulate the splicing pattern in cultured myotubes from two different DMD patients with an exon 45 deletion11. Following AON treatment, exon 46 was skipped, which resulted in a restored reading frame and the induction of dystrophin synthesis in at least 75% of the cells. We have recently shown that exon skipping can also efficiently be induced in human control and patient muscle cells for 39 different DMD exons using exon-internal AONs11-15.

[0006] Hence, exon skipping techniques applied on the dystrophin gene result in the generation of at least partially functional—albeit shorter—dystrophin protein in DMD patients. Since DMD is caused by a dysfunctional dystrophin protein, it would be expected that the symptoms of DMD are sufficiently alleviated once a DMD patient has been provided with functional dystrophin protein. However, the present invention provides the insight that, even though exon skipping techniques are capable of inducing dystrophin synthesis, the oligonucleotide used for exon skipping technique can be improved any further by incorporating an inosine and/or a nucleotide containing a base able to form a wobble base pair in said oligonucleotide.

DESCRIPTION OF THE INVENTION

Oligonucleotide

[0007] In a first aspect, there is provided an oligonucleotide comprising an inosine and/or a nucleotide containing a base able to form a wobble base pair or a functional equivalent thereof, wherein the oligonucleotide, or a functional equivalent thereof, comprises a sequence which is complementary to at least part of a dystrophin pre-mRNA exon or at least part of a non-exon region of a dystrophin pre-mRNA said part being a contiguous stretch comprising at least 8 nucleotides.

[0008] The use of an inosine and/or a nucleotide containing a base able to form a wobble base pair in an oligonucleotide of the invention is very attractive as explained below. Inosine for example is a known modified base which can pair with three bases: uracil, adenine, and cytosine. Inosine is a nucleoside that is formed when hypoxanthine is attached to a ribose ring (also known as a ribofuranose) via a β-N9-glycosidic bond. Inosine is commonly found in tRNAs and is essential for proper translation of the genetic code in wobble base pairs. A wobble base pair is a G-U and I-U/A-I/C pair fundamental in RNA secondary structure. Its thermodynamic stability is comparable to that of the Watson-Crick base pair. Wobble base pairs are critical for the proper translation of the genetic code. The genetic code makes up for disparities in the number of amino acids (20) for triplet codons (64), by using modified base pairs in the first base of the anti-codon. Similarly, when designing primers for polymerase chain reaction, inosine is useful in that it will indiscriminately pair with adenine, thymine, or cytosine. A first advantage of using such a base allows one to design a primer that spans a single nucleotide polymorphism (SNP), without worry that the polymorphism will disrupt the primer’s annealing efficiency. Therefore in the invention, the use of such a base allows to design an oligonucleotide that may be used for an individual having a SNP within the dystrophin pre-mRNA stretch which is targeted by an oligonucleotide of the invention. A second advantage of using an inosine and/or a base able to form a
A fifth advantage of using an inosine and/or a base able to form a wobble base pair in an oligonucleotide of the invention is to allow to design an oligonucleotide with improved RNA binding kinetics and/or thermodynamic properties. The RNA binding kinetics and/or thermodynamic properties are at least in part determined by the melting temperature of an oligonucleotide (Tm; calculated with the oligonucleotide properties calculator (http://www.unc.edu/~cail/biotool/oligo/index.html) for single stranded RNA using the basic Tm and the nearest neighbour model), and/or the free energy of the AON-target exon complex (using RNA structure version 4.5). If a Tm is too high, the oligonucleotide is expected to be less specific. An acceptable Tm and free energy depend on the sequence of the oligonucleotide. Therefore, it is difficult to give preferred ranges for each of these parameters. An acceptable Tm may be ranged between 35 and 65°C and an acceptable free energy may be ranged between 15 and 45 kcal/mol.

The skilled person may therefore first choose an oligonucleotide as a potential therapeutic compound. In a second step, he may use the invention to further optimise said oligonucleotide by decreasing its immunogenicity and/or avoiding aggregation and/or quadruplex formation and/or by optimizing its Tm and/or free energy of the AON-target complex. He may try to introduce at least one inosine and/or a base able to form a wobble base pair in said oligonucleotide at a suitable position and assess how the immunogenicity and/or aggregation and/or quadruplex formation and/or Tm and/or free energy of the AON-target complex have been altered by the presence of said inosine and/or a base able to form a wobble base pair. If the alteration does not provide the desired alteration or decrease of immunogenicity and/or aggregation and/or quadruplex formation and/or its Tm and/or free energy of the AON-target complex he may choose to introduce a further inosine and/or a base able to form a wobble base pair in said oligonucleotide and/or to introduce a given inosine and/or a base able to form a wobble base pair at a distinct suitable position within said oligonucleotide.

An oligonucleotide comprising an inosine and/or a base able to form a wobble base pair may be defined as an oligonucleotide wherein at least one nucleotide has been substituted with an inosine and/or a base able to form a wobble base pair. The skilled person knows how to test whether a nucleotide contains a base able to form a wobble base pair. Since for example inosine can form a base pair with uracil, adenine, and/or cytosine, it means that at least one nucleotide able to form a base pair with uracil, adenine and/or cytosine has been substituted with inosine. However, in order to safeguard specificity, the inosine containing oligonucleotide preferably comprises the substitution of at least one, two, three, four nucleotide(s) able to form a base pair with uracil or adenine or cytosine as long as an acceptable level of a functional activity of said oligonucleotide is retained as defined later herein.

An oligonucleotide comprising an inosine and/or a base able to form a wobble base pair is preferably an oligonucleotide, which is still able to exhibit an acceptable level of a functional activity of a corresponding oligonucleotide not comprising an inosine and/or a base able to form a wobble base pair. A functional activity of said oligonucleotide is preferably to provide an individual with a functional dystrophin protein and/or mRNA and/or at least in part decreasing the production of an aberrant dystrophin protein and/or mRNA. Each of these features are later defined herein. An
acceptable level of such a functional activity is preferably at least 50%, 60%, 70%, 80%, 90%, 95% or 100% of the functional activity of the corresponding oligonucleotide which does not comprise an inosine and/or a base able to form a wobble base pair. Such functional activity may be as measured in a muscular tissue or in a muscular cell of an individual or in vitro in a cell by comparison to the functional activity of a corresponding oligonucleotide not comprising an inosine and/or a base able to form a wobble base pair. The assessment of the functionality may be carried out at the mRNA level, preferably using RT-PCR. The assessment of the functionality may be carried out at the protein level, preferably using western blot analysis or immunofluorescence analysis of cross-sections.

Within the context of the invention, an inosine and/or a base able to form a wobble base pair as present in an oligonucleotide is present in a part of said oligonucleotide which is complementary to at least part of a dystrophin pre-mRNA exon or at least part of a non-exon region of a dystrophin pre-mRNA said part being a contiguous stretch comprising at least 8 nucleotides. Therefore, in a preferred embodiment, an oligonucleotide comprising an inosine and/or a nucleotide containing a base able to form a wobble base pair or a functional equivalent thereof, wherein the oligonucleotide, or a functional equivalent thereof, comprises a sequence which is complementary to at least part of a dystrophin pre-mRNA exon or at least part of a non-exon region of a dystrophin pre-mRNA said part being a contiguous stretch comprising at least 8 nucleotides and wherein said inosine and/or a nucleotide containing a base able is present within the oligonucleotide sequence which is complementary to at least part of a dystrophin pre-mRNA as defined in previous sentence.

However, as later defined herein such inosine and/or a base able to form a wobble base pair may also be present in a linking moiety present in an oligonucleotide of the invention. Preferred linking moieties are later defined herein.

In a preferred embodiment, such oligonucleotide is preferably a medicament. More preferably, said medicament is for preventing or treating Duchenne Muscular Dystrophy or Becker Muscular Dystrophy in an individual or a patient. As defined herein a DMD pre-mRNA preferably means the pre-mRNA of a DMD gene of a DMD or BMD patient. A patient is preferably intended to mean a patient having DMD or BMD or a patient susceptible to develop DMD or BMD due to his or her genetic background. In the case of a DMD patient, an oligonucleotide used will preferably correct at least one of the DMD mutations as present in the DMD gene of said patient and therefore will preferably create a dystrophin that will look like a BMD dystrophin: said dystrophin will preferably be a functional dystrophin as later defined herein.

In the case of a BMD patient, an oligonucleotide as used will preferably correct at least one of the BMD mutations as present in the BMD gene of said patient and therefore will preferably create a, or more of a, dystrophin, which will be more functional than the dystrophin which was originally present in said BMD patient. Even more preferably, said medicament provides an individual with a functional or more (of) a functional dystrophin protein and/or mRNA and/or at least in part decreases the production of an aberrant dystrophin protein and/or mRNA.

Preferably, a method of the invention by inducing and/or promoting skipping of at least one exon of the DMD pre-mRNA as identified herein in one or more cells, preferably muscle cells of a patient, provides said patient with an increased production of a more of a functional dystrophin protein and/or mRNA and/or decreases the production of an aberrant or less functional dystrophin protein and/or mRNA in said patient.

Providing a patient with a more functional dystrophin protein and/or mRNA and/or decreasing the production of an aberrant dystrophin protein and/or mRNA in said patient is typically applied in a DMD patient. Increasing the production of a more functional or functional dystrophin and/or mRNA is typically applied in a BMD patient.

A preferred method is a method, wherein a patient or one or more cells of said patient is provided with an increased production of a more functional or functional dystrophin protein and/or mRNA and/or wherein the production of an aberrant dystrophin protein and/or mRNA in said patient is decreased, wherein the level of said aberrant or more functional dystrophin protein and/or mRNA is assessed by comparison to the level of said dystrophin and/or mRNA in said patient at the onset of the method.

As defined herein, a functional dystrophin is preferably a wild type dystrophin corresponding to a protein having the amino acid sequence as identified in SEQ ID NO: 1. A functional dystrophin is preferably a dystrophin, which has an actin binding domain in its N terminal part (first 240 amino acids at the N terminus), a cystein-rich domain (amino acid 3361 till 3685) and a C terminal domain (last 325 amino acids at the C terminus) each of these domains being present in a wild type dystrophin as known to the skilled person. The amino acids indicated herein correspond to amino acids of the wild type dystrophin being represented by SEQ ID NO: 1. In another embodiment, a functional dystrophin is a dystrophin, which exhibits to some extent an activity of a wild type dystrophin. “At least to some extent” preferably means at least 50%, 60%, 70%, 80%, 90%, 95% or 100% of a corresponding activity of a wild type functional dystrophin. In this context, an activity of a wild type dystrophin is preferably binding to actin and to the dystrophin-associated glycoprotein complex (DGC)\textsuperscript{56}. Binding of dystrophin to actin and to the DGC complex may be visualized by either co-immunoprecipitation using total protein extracts or immunofluorescence analysis of cross-sections, from a biopsy of a muscle suspected to be dystrophic, as known to the skilled person.

Individuals suffering from Duchenne muscular dystrophy typically have a mutation in the gene encoding dystrophin that prevents synthesis of the complete protein, i.e a premature stop prevents the synthesis of the C-terminus of the protein. In Becker muscular dystrophy the dystrophin gene also comprises a mutation compared to the wild type but the mutation does typically not include a premature stop and the C-terminus of the protein is typically synthesized. As a result a functional dystrophin protein is synthesized that has at least the same activity in kind as a wild type protein, although not necessarily the same amount of activity. In a preferred embodiment, a functional dystrophin protein means an in frame dystrophin gene. The genome of a BMD individual typically encodes a dystrophin protein comprising the N terminal part (first 240 amino acids at the N terminus), a cysteine-rich domain (amino acid 3361 till 3685) and a C terminal domain (last 325 amino acids at the C terminus) but its central rod shaped domain may be shorter than the one of a wild type dystrophin\textsuperscript{56}. Exon—skipping for the treatment of DMD is preferably but not exclusively directed to overcome a prema-
ture stop in the pre-mRNA by skipping an exon in the rod-domain shaped domain to correct the reading frame and allow synthesis of remainder of the dystrophin protein including the C-terminus, albeit that the protein is somewhat smaller as a result of a smaller rod domain. In a preferred embodiment, an individual having DMD and being treated using an oligonucleotide as defined herein will be provided a dystrophin, which exhibits at least to some extent an activity of a wild type dystrophin. More preferably, it said individual is a Duchenne patient or is suspected to be a Duchenne patient, a functional dystrophin is a dystrophin of an individual having BMD: preferably said dystrophin is able to interact with both actin and the DGC, but its central rod shaped domain may be shorter than the one of a wild type dystrophin (Aartsma-Rus et al (2006, ref 56). The central rod domain of wild type dystrophin comprises 24 spectrin-like repeats 56. For example, a central rod shaped domain of a dystrophin as provided herein may comprise 5 to 23, 10 to 22 or 12 to 18 spectrin-like repeats as long as it can bind to actin and to DGC. Decreasing the production of an aberrant dystrophin in said patient or in a cell of said patient may be assessed at the mRNA level and preferably means that 99%, 90%, 80%, 70%, 60%, 50%, 40%, 30%, 20%, 10%, 5% or less of the initial amount of aberrant dystrophin mRNA, is still detectable by RT PCR. An aberrant dystrophin mRNA or protein is also referred to herein as a non-functional or less to non-functional or semi-functional dystrophin mRNA or protein. A non-functional pre-mRNA dystrophin is preferably leads to an out of frame dystrophin protein, which means that no dystrophin protein will be produced and/or detected. A non functional dystrophin protein is preferably a dystrophin protein which is not able to bind actin and/or members of the DGC protein complex. A non-functional dystrophin protein or dystrophin mRNA does typically not have, or does not encode a dystrophin protein with an intact C-terminus of the protein.

Increasing the production of a functional dystrophin in said patient or in a cell of said patient may be assessed at the mRNA level (by RT-PCR analysis) and preferably means that a detectable amount of a functional or in frame dystrophin mRNA is detectable by RT PCR. In another embodiment, 1%, 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or more of the detectable dystrophin mRNA is a functional or in frame dystrophin mRNA.

Increasing the production of a functional dystrophin in said patient or in a cell of said patient may be assessed at the protein level (by immunofluorescence and western blot analyses) and preferably means that a detectable amount of a functional dystrophin protein is detectable by immunofluorescence or western blot analysis. In another embodiment, 1%, 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or more of the detectable dystrophin protein is a functional dystrophin protein.

An increase or a decrease is preferably assessed in a muscular tissue or in a muscular cell of an individual or a patient by comparison to the amount present in said individual or patient before treatment with said molecule or composition of the invention. Alternatively, the comparison can be made with a muscular tissue or cell of said individual or patient, which has not yet been treated with said oligonucleotide or composition in case the treatment is local.

In a preferred method, one or more symptom(s) from a DMD or a BMD patient is/are alleviated and/or one or more characteristic(s) of a muscle cell or tissue from a DMD or a BMD patient is/are alleviated using a molecule or a composition of the invention. Such symptoms may be assessed on the patient self. Such characteristics may be assessed at the cellular, tissue level of a given patient. An alleviation of one or more characteristics may be assessed by any of the following assays: prolongation of time to loss of walking, improvement of muscle strength, improvement of the ability to lift weight, improvement of the time taken to rise from the floor, improvement in the nine-meter walking time, improvement in the time taken for four-stairs climbing, improvement of the leg function grade, improvement of the pulmonary function, improvement of cardiac function, improvement of the quality of life. Each of these symptoms is known to the skilled person. As an example, the publication of Manzur et al (2008, ref 58) gives an extensive explanation of each one of these symptoms. For each of these symptoms, as soon as a detectable improvement or prolongation of parameter measured in an assay has been found, it will preferably mean that one or more symptoms of Duchenne Muscular Dystrophy or Becker Muscular Dystrophy has been alleviated in an individual using a molecule or composition of the invention. Detectable improvement or prolongation is preferably a statistically significant improvement or prolongation as described in Hodggets et al (2006, ref 57). Alternatively, the alleviation of one or more symptom(s) of Duchenne Muscular Dystrophy or Becker Muscular Dystrophy may be assessed by measuring an improvement of a muscle fiber function, integrity and/or survival as later defined herein.

An oligonucleotide as used herein preferably comprises an antisense oligonucleotide or antisense oligoribonucleotide. In a preferred embodiment an exon skipping technique is applied. Exon skipping interferes with the natural splicing processes occurring within a eukaryotic cell. In higher eukaryotes the genetic information for proteins in the DNA of the cell is encoded in exons which are separated from each other by intronic sequences. These introns are in some cases very long. The transcription machinery of eukaryotes generates a pre-mRNA which contains both exons and introns, while the splicing machinery, often already during the production of the pre-mRNA, generates the actual coding region for the protein by splicing together the exons present in the pre-mRNA.

Exon-skipping results in mature mRNA that lacks at least one skipped exon. Thus, when said exon codes for amino acids, exon skipping leads to the expression of an altered product. Technology for exon-skipping is currently directed towards the use of antisense oligonucleotides (AONs). Much of this work is done in the mdx mouse model for Duchenne muscular dystrophy. The mdx mouse carries a nonsense mutation in exon 23. Despite the mdx mutation, which should preclude the synthesis of a functional dystrophin protein, rare, naturally occurring dystrophin positive fibers have been
observed in mdx muscle tissue. These dystrophin-positive fibers are thought to have arisen from an apparently naturally occurring exon-skipping mechanism, either due to somatic mutations or through alternative splicing. AONs directed to, respectively, the 3' and/or 5' splice sites of introns 22 and 23 in dystrophin pre-mRNA, have been shown to interfere with factors normally involved in removal of intron 23 so that also exon 23 was removed from the mRNA.\(^5\), 6, 38, 40.

[0034] By the targeted skipping of a specific exon, a DMD phenotype is converted into a milder BMD phenotype. The skipping of an exon is preferably induced by the binding of AONs targeting either one or both of the splice sites, or exon-internal sequences. An oligonucleotide directed toward an exon internal sequence typically exhibits no overlap with non-exon sequences. It preferably does not overlap with the splice sites at least not insofar, as these are present in the intron. An oligonucleotide directed toward an exon internal sequence preferably does not contain a sequence complementary to an adjacent intron. Further provided is thus an oligonucleotide according to the invention, wherein said oligonucleotide, or a functional equivalent thereof, is for inhibiting inclusion of an exon of a dystrophin pre-mRNA into mRNA produced from splicing of said pre-mRNA. An exon skipping technique is preferably applied such that the absence of an exon from mRNA produced from dystrophin pre-mRNA generates a coding region for a more functional—albeit shorter—dystrophin protein. In this context, inhibiting inclusion of an exon preferably means that the detection of the original, aberrant dystrophin mRNA and/or protein is decreased as earlier defined herein.

[0035] Since an exon of a dystrophin pre-mRNA will only be included into the resulting mRNA when both the splice sites are recognised by the spliceosome complex, splice sites have been obvious targets for AONs. One embodiment therefore provides an oligonucleotide, or a functional equivalent thereof, comprising a sequence which is complementary to a non-exon region of a dystrophin pre-mRNA. In one embodiment an AON is used which is solely complementary to a non-exon region of a dystrophin pre-mRNA. This is however not necessary: it is also possible to use an AON which comprises an intron-specific sequence as well as exon-specific sequence. Such AON comprises a sequence which is complementary to a non-exon region of a dystrophin pre-mRNA, as well as a sequence which is complementary to an exon region of a dystrophin pre-mRNA. Of course, an AON is not necessarily complementary to the entire sequence of a dystrophin exon or intron. AONs, which are complementary to a part of such exon or intron are preferred. An AON is preferably complementary to at least part of a dystrophin exon and/or intron, said part having at least 8, 10, 13, 15, 20 nucleotides.

[0036] Splicing of a dystrophin pre-mRNA occurs via two sequential transaternification reactions. First, the 2'OH of a specific branch-point nucleotide within the intron that is defined during spliceosome assembly performs a nucleophilic attack on the first nucleotide of the intron at the 5' splice site forming the lariat intermediate. Second, the 3'OH of the released 5' exon then performs a nucleophilic attack at the last nucleotide of the intron at the 3' splice site thus joining the exons and releasing the intron lariat. The branch point and splice sites of an intron are thus involved in splicing event. Hence, an oligonucleotide comprising a sequence, which is complementary to such branch point and/or splice site is preferably used for exon skipping. Further provided is therefore an oligonucleotide, or a functional equivalent thereof, which comprises a sequence which is complementary to a splice site and/or branch point of a dystrophin pre-mRNA.

[0037] Since splice sites contain consensus sequences, the use of an oligonucleotide or a functional equivalent thereof (herein also called an AON) comprising a sequence which is complementary of a splice site involves the risk of promiscuous hybridization. Hybridization of AONs to other splice sites than the sites of the exon to be skipped could easily interfere with the accuracy of the splicing process. To overcome these and other potential problems related to the use of AONs which are complementary to an intron sequence, one preferred embodiment provides an oligonucleotide, or a functional equivalent thereof, comprising a sequence which is complementary to a dystrophin pre-mRNA exon. Preferably, said AON is capable of specifically inhibiting an exon inclusion signal of at least one exon in said dystrophin pre-mRNA. Interfering with an exon inclusion signal (EIS) has the advantage that such elements are located within the exon. By providing an AON for the interior of the exon to be skipped, it is possible to interfere with the exon inclusion signal thereby effectively masking the exon from the splicing apparatus. The failure of the splicing apparatus to recognize the exon to be skipped thus leads to exclusion of the exon from the final mRNA. This embodiment does not interfere directly with the enzymatic process of the splicing machinery (the joining of the exons). It is thought that this allows the method to be more specific and/or reliable. It is thought that an EIS is a particular structure of an exon that allows splice acceptor and donor to assume a particular spatial conformation. In this concept, it is the particular spatial conformation that enables the splicing machinery to recognize the exon. However, the invention is certainly not limited to this model. In a preferred embodiment, use is made of an oligonucleotide, which is capable of binding to an exon and is capable of inhibiting an EIS. An AON may specifically contact said exon at any point and still be able to specifically inhibit said EIS.

[0038] Within the context of the invention, a functional equivalent of an oligonucleotide preferably means an oligonucleotide as defined herein wherein one or more nucleotides have been substituted and wherein an activity of said functional equivalent is retained to at least some extent. Preferably, an activity of said functional equivalent is providing a functional dystrophin protein. Said activity of said functional equivalent is therefore preferably assessed by quantifying the amount of a functional dystrophin protein or by quantifying the amount of a functional dystrophin mRNA. A functional dystrophin protein (or a functional dystrophin mRNA) is herein preferably defined as being a dystrophin protein (or a dystrophin protein encoded by said mRNA) able to bind actin and members of the DGC protein. The assessment of said activity of an oligonucleotide is preferably done by RT-PCR (m-RNA) or by immunofluorescence or Western blot analyses (protein). Said activity is preferably retained at least some extent when it represents at least 50%, or at least 60%, or at least 70% or at least 80% or at least 90% or at least 95% or more of corresponding activity of said oligonucleotide the functional equivalent derives from. Such activity may be measured in a muscular tissue or in a muscular cell of an individual or in vitro in a cell by comparison to an activity of a corresponding oligonucleotide of said oligonucleotide the functional equivalent derives from. Throughout this application, when the word oligonucleotide is used it may be replaced by a functional equivalent thereof as defined herein.
Hence, an oligonucleotide, or a functional equivalent thereof, comprising or consisting of a sequence which is complementary to a dystrophin pre-mRNA exon provides good DMD therapeutic results. In one preferred embodiment an oligonucleotide, or a functional equivalent thereof, is used which comprises or consists of a sequence which is complementary to at least part of either dystrophin pre-mRNA exons 2 to 75 said part having or comprising at least 13 nucleotides. However, said part may also have at least 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50 nucleotides. A part of dystrophin pre-mRNA to which an oligonucleotide is complementary may also be called a contiguous stretch of dystrophin pre-mRNA.

Most preferably an AON is used which comprises or consists of a sequence which is complementary to at least part of dystrophin pre-mRNA exon 51, 45, 53, 44, 46, 52, 50, 43, 6, 7, 8, 55, 2, 11, 17, 19, 21, 57, 59, 62, 63, 65, 66, 69, and/or 75 said part having or comprising at least 13 nucleotides. However, said part may also have at least 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50 nucleotides. More preferred oligonucleotides are represented by a sequence that comprises or consists of each of the following sequences SEQ ID NO: 2 to SEQ ID NO:539 wherein at least one inosine and/or a base able to form a wobble base pair is present in said sequence. Preferably, an inosine has been introduced in one of these sequences to replace a guanosine, adenine, or a uracil. Accordingly, an even more preferred oligonucleotide as used herein is represented by a sequence that comprises or consists of SEQ ID NO:2 to SEQ ID NO:486 or SEQ ID NO:539, even more preferably SEQ ID NO:2 to NO:237 or SEQ ID NO:539, most preferably SEQ ID NO:76 wherein at least one inosine and/or a base able to form a wobble base pair is present in said sequence. Preferably, an inosine has been introduced in one of these sequences to replace a guanosine, adenine, or a uracil.

Accordingly, in another preferred embodiment, an oligonucleotide as used herein is represented by a sequence that comprises or consists of SEQ ID NO:540 to SEQ ID NO:576. More preferably, an oligonucleotide as used herein is represented by a sequence that comprises or consists of SEQ ID NO:557.

Said exons are listed in decreasing order of patient population applicability. Hence, the use of an AON comprising a sequence, which is complementary to at least part of dystrophin pre-mRNA exon 51 is suitable for use in a larger part of the DMD patient population as compared to an AON comprising a sequence which is complementary to dystrophin pre-mRNA exon 44 et cetera.

In a preferred embodiment, an oligonucleotide of the invention, which comprises a sequence that is complementary to part of dystrophin pre-mRNA is such that the complementary part is at least 50% of the length of the oligonucleotide of the invention, more preferably at least 60%, even more preferably at least 70%, even more preferably at least 80%, even more preferably at least 90% or even more preferably at least 95%, or even more preferably 98% or even more preferably at least 99%, or even more preferably 100%. In a most preferred embodiment, the oligonucleotide of the invention consists of a sequence that is complementary to part of dystrophin pre-mRNA as defined herein. As an example, an oligonucleotide may comprise a sequence that is complementary to part of dystrophin pre-mRNA as defined herein and additional flanking sequences. In a more preferred embodiment, the length of said complementary part of said oligonucleotide is of at least 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50 nucleotides. Preferably, additional flanking sequences are used to modify the binding of a protein to the oligonucleotide, to or modify a thermodynamic property of the oligonucleotide, more preferably to modify target RNA binding affinity. In a more preferred embodiment an oligonucleotide is capable of hybridising to the complementary part. In this context, “sufficiently” preferably means that using a
gel mobility shift assay as described in example 1 of EP 1 619 249, binding of an oligonucleotide is detectable. Optionally, said oligonucleotide may further be tested by transfection into muscle cells of patients. Skipping of the targeted exon may be assessed by RT-PCR (as described in EP 1 619 249). The complementary regions are preferably designed such that, when combined, they are specific for the exon in the pre-mRNA. Such specificity may be created with various lengths of complementary regions as this depends on the actual sequences in other (pre-)mRNA in the system. The risk that also one or more other pre-mRNA will be able to hybridise to the oligonucleotide decreases with increasing size of the oligonucleotide. It is clear that oligonucleotides comprising mismatches in the region of complementarity but that retain the capacity to hybridise to the targeted region(s) in the pre-mRNA, can be used in the present invention. However, preferably at least the complementary parts do not comprise such mismatches as these typically have a higher efficiency and a higher specificity, than oligonucleotides having such mismatches in one or more complementary regions. It is thought, that higher hybridisation strengths, (i.e. increasing number of interactions with the opposing strand) are favourable in increasing the efficiency of the process of interfering with the splicing machinery of the system. Preferably, the complementarity is between 90 and 100%. In general this allows for approximately 1 or 2 mismatch(es) in an oligonucleotide of around 20 nucleotides.

The secondary structure is best analysed in the context of the pre-mRNA wherein the exon resides. Such structures may be analysed in the actual RNA. However, it is currently possible to predict the secondary structure of an RNA molecule (at lowest energy costs) quite well using structure-modelling programs. A non-limiting example of a suitable program is RNA molfold version 3.1 server. A person skilled in the art will be able to predict, with suitable reproducibility, a likely structure of the exon, given the nucleotide sequence. Best predictions are obtained when providing such modelling programs with both the exon and flanking intron sequences. It is typically not necessary to model the structure of the entire pre-mRNA.

The open and closed structure to which the oligonucleotide is directed, are preferably adjacent to one another. It is thought, that in this way the annealing of the oligonucleotide to the open structure induces opening of the closed structure whereupon annealing progresses into this closed structure. Through this action the previously closed structure assumes a different conformation. The different conformation results in the disruption of the exon inclusion signal. However, when potential (cryptic) splice acceptor and/or donor sequences are present within the targeted exon, occasionally a new exon inclusion signal is generated defining a different (neo) exon, i.e. with a different 5' end, a different 3' end, or both. This type of activity is within the scope of the present invention as the targeted exon is excluded from the mRNA. The presence of a new exon, containing part of the targeted exon, in the mRNA does not alter the fact that the targeted exon, as such, is excluded. The inclusion of a neo-exon can be seen as a side effect, which occurs only occasionally. There are two possibilities when exon skipping is used to restore (part of) an open reading frame of dystrophin that is disrupted as a result of a mutation. One is that the neo-exon is functional in the restoration of the reading frame, whereas in the other case the reading frame is not restored. When selecting oligonucleotides for restoring dystrophin reading frames by means of exon-skipping it is of course clear that under these conditions only those oligonucleotides are selected that indeed result in exon-skipping that restores the dystrophin open reading frame, with or without a neo-exon.

Further provided is an oligonucleotide, or a functional equivalent thereof, comprising a sequence that is complementary to a binding site for a serine-arginine (SR) protein in RNA of an exon of a dystrophin pre-mRNA. In WO 2006/112705 we have disclosed the presence of a correlation between the effectiveness of an exon-internal antisense oligonucleotide (AOI) in inducing exon skipping and the presence of a (for example by ESE finder) predicted SR binding site in the target pre-mRNA site of said AON.

Therefore, in one embodiment an oligonucleotide is generated comprising determining a (putative) binding site for an SR (Ser-Arg) protein in RNA of a dystrophin exon and producing an oligonucleotide that is complementary to said RNA and that at least partly overlaps said (putative) binding site. The term “at least partly overlaps” is defined herein as to comprise an overlap of only a single nucleotide of an SR binding site as well as multiple nucleotides of said binding site as well as a complete overlap of said binding site. This embodiment preferably further comprises determining from a secondary structure of said RNA, a region that is hybridised to another part of said RNA (closed structure) and a region that is not hybridised in said structure (open structure), and subsequently generating an oligonucleotide that at least partly overlaps said (putative) binding site and that overlaps at least part of said closed structure and overlaps at least part of said open structure. In this way we increase the chance of obtaining an oligonucleotide that is capable of interfering with the exon inclusion from the pre-mRNA into mRNA. It is possible that a first selected SR-binding region does not have the requested open-closed structure in which case another (second) SR protein binding site is selected which is then subsequently tested for the presence of an open-closed structure. This process is continued until a sequence is identified which contains an SR protein binding site as well as an (partly overlapping) open-closed structure. This sequence is then used to design an oligonucleotide which is complementary to said sequence.

Such a method, for generating an oligonucleotide, is also performed by reversing the described order, i.e. first generating an oligonucleotide comprising determining, from a secondary structure of RNA from a dystrophin exon, a region that assumes a structure that is hybridised to another part of said RNA (closed structure) and a region that is not hybridised in said structure (open structure), and subsequently generating an oligonucleotide, of which at least a part of said oligonucleotide is complementary to said closed structure and of which at least another part of said oligonucleotide is complementary to said open structure. This is then followed by determining whether an SR protein binding site at least overlaps with said open/closed structure. In this way the method of WO 2004/083432 is improved. In yet another embodiment the selections are performed simultaneously.

Without wishing to be bound by any theory it is currently thought that use of an oligonucleotide directed to an SR protein binding site results in (at least partly) impairing the binding of an SR protein to the binding site of an SR protein which results in disrupted or impaired splicing.

Preferably, an open/closed structure and an SR protein binding site partly overlap and even more preferred an open/closed structure completely overlaps an SR protein...
binding site or an SR protein binding site completely overlaps an open/closed structure. This allows for an improved disruption of exon inclusion.

Besides consensus splice sites sequences, many (if not all) exons contain splicing regulatory sequences such as exonic splicing enhancer (ESE) sequences to facilitate the recognition of genuine splice sites by the spliceosome\[42,43\]. A subgroup of splicing factors, called the SR proteins, can bind to these ESEs and recruit other splicing factors, such as U1 and U2AF to (weakly defined) splice sites. The binding sites of the four most abundant SR proteins (SF2/ASF, SC35, SRp40 and SRp55) have been analyzed in detail and these results are implemented in ESE finder, a web source that predicts potential binding sites for these SR proteins\[42,43\]. There is a correlation between the effectiveness of an AON and the presence/absence of an SF2/ASF, SC35 and SRp40 binding site. In a preferred embodiment, the invention thus provides a combination as described above, wherein said SR protein is SF2/ASF or SC35 or SRp40.

In one embodiment an oligonucleotide, or a functional equivalent thereof is capable of specifically binding a regulatory RNA sequence which is required for the correct splicing of a dystrophin exon in a transcript. Several cis-acting RNA sequences are required for the correct splicing of exons in a transcript. In particular, supplementary elements such as intronic or exonic splicing enhancers (ISEs and ESEs) or silencers (ISSs and ESES) are identified to regulate specific and efficient splicing of constitutive and alternative exons. Using sequence-specific antisense oligonucleotides (AONs) that bind to the elements, their regulatory function is disturbed so that the exon is skipped, as shown for DMD. Hence, in one preferred embodiment an oligonucleotide or functional equivalent thereof is used which is complementary to an intronic splicing enhancer (ISE), an exonic splicing enhancer (ESE), an intronic splicing silencer (ISS) and/or an exonic splicing silencer (ESS). As already described herein before, a dystrophin exon is in one preferred embodiment skipped by an agent capable of specifically inhibiting an exon inclusion signal of said exon, so that said exon is not recognized by the splicing machinery as a part that needs to be included in the mRNA. As a result, a mRNA without said exon is formed.

An AON used herein is preferably complementary to a consecutive part or a contiguous stretch of between 8 and 50 nucleotides of dystrophin exon RNA or dystrophin intron RNA. In one embodiment an AON used herein is complementary to a consecutive part or a contiguous stretch of between 14 and 50 nucleotides of a dystrophin exon RNA or dystrophin intron RNA. Preferably, said AON is complementary to a consecutive part or contiguous stretch of between 14 and 25 nucleotides of said exon RNA. More preferably, an AON is used which comprises a sequence which is complementary to a consecutive part or a contiguous stretch of between 20 and 25 nucleotides of a dystrophin exon RNA or a dystrophin intron RNA.

Different types of nucleic acid may be used to generate an oligonucleotide. Preferably, said oligonucleotide comprises RNA, as RNA/RNA hybrids are very stable. Since one of the aims of the exon skipping technique is to directly splicing in subjects it is preferred that the oligonucleotide RNA comprises a modification providing the RNA with an additional property, for instance resistance to endonucleases, exonucleases, and RNaseH, additional hybridisation strength, increased stability (for instance in a bodily fluid), increased or decreased flexibility, reduced toxicity, increased intracellular transport, tissue-specificity, etc. Preferably, said modification comprises a 2’-O-methyl-phosphorothioate oligoribonucleotide modification. Preferably, said modification comprises a 2’-O-methyl-phosphorothioate oligodeoxyribonucleotide modification. One embodiment thus provides an oligonucleotide is used which comprises RNA which contains a modification, preferably a 2’-O-methyl modified ribose (RNA) or deoxyribose (DNA) modification.

In one embodiment the invention provides a hybrid oligonucleotide comprising an oligonucleotide comprising a 2’-O-methyl-phosphorothioate oligo(deoxy)ribonucleotide modification and locked nucleic acid. This particular oligonucleotide comprises better sequence specificity compared to an equivalent consisting of locked nucleic acid, and comprises increased effectiveness when compared with an oligonucleotide consisting of 2’-O-methyl-phosphorothioate oligo(deoxy)ribonucleotide modification.

With the advent of nucleic acid mimicking technology it has become possible to generate molecules that have a similar, preferably the same hybridisation characteristics in kind not necessarily in amount as nucleic acid itself. Such functional equivalents are of course also suitable for use in the invention. Preferred examples of functional equivalents of an oligonucleotide are peptide nucleic acid and/or locked nucleic acid. Most preferably, a morpholino phosphorodiamidate is used. Suitable but non-limiting examples of equivalents of oligonucleotides of the invention can be found in\[42,43\]. Hybridization between one or more of the equivalents among each other and/or together with nucleic acid are of course also suitable. In a preferred embodiment locked nucleic acid is used as a functional equivalent of an oligonucleotide, as locked nucleic acid displays a higher target affinity and reduced toxicity and therefore shows a higher efficiency of exon skipping.

In one embodiment an oligonucleotide, or a functional equivalent thereof, which is capable of inhibiting inclusion of a dystrophin exon into dystrophin mRNA is combined with at least one other oligonucleotide, or functional equivalent thereof, that is capable of inhibiting inclusion of another dystrophin exon into dystrophin mRNA. This way, inclusion of two or more exons of a dystrophin pre-mRNA produced from this pre-mRNA is prevented. This embodiment is further referred to as double- or multi-exon skipping. In most cases double-exon skipping results in the exclusion of only the two targeted exons from the dystrophin pre-mRNA. However, in other cases it was found that the targeted exons and the entire region in between said exons in said pre-mRNA were not present in the produced mRNA even when other exons (intervening exons) were present in such region. This multi-exon skipping was notably so for the combination of oligonucleotides derived from the DMD gene, wherein one oligonucleotide for exon 45 and one oligonucleotide for exon 51 was added to a cell transcribing the DMD gene. Such a set-up resulted in mRNA being produced that did not contain exons 45 to 51. Apparently, the structure of the pre-mRNA in the presence of the mentioned oligonucleotides was such that the splicing machinery was stimulated to connect exons 44 and 52 to each other. Other preferred examples of multi-exon skipping are:

the use of an oligonucleotide targeting exon 17, and a second one exon 48 which may result in the skipping of said exon 17 and exon 48.
[0063] the use of an oligonucleotide targeting exon 17, and a second one exon 51 which may result in the skipping of said both exons or of the entire region between exon 17 and exon 51.

[0064] the use of an oligonucleotide targeting exon 42, and a second one exon 55 which may result in the skipping of said both exons or of the entire region between exon 42 and exon 55.

[0065] the use of an oligonucleotide targeting exon 43, and a second one exon 51 which may result in the skipping of said both exons or of the entire region between exon 43 and exon 51.

[0066] the use of an oligonucleotide targeting exon 43, and a second one exon 55 which may result in the skipping of said both exons or of the entire region between exon 43 and exon 55.

[0067] the use of an oligonucleotide targeting exon 45, and a second one exon 55 which may result in the skipping of said both exons or of the entire region between exon 45 and exon 55.

[0068] the use of an oligonucleotide targeting exon 45, and a second one exon 59 which may result in the skipping of said both exons or of the entire region between exon 45 and exon 59.

[0069] the use of an oligonucleotide targeting exon 48, and a second one exon 59 which may result in the skipping of said both exons or of the entire region between exon 48 and exon 59.

[0070] the use of an oligonucleotide targeting exon 50, and a second one exon 51 which may result in the skipping of said both exons.

[0071] the use of an oligonucleotide targeting exon 51, and a second one exon 52 which may result in the skipping of said both exons.

[0072] Further provided is therefore an oligonucleotide which comprises at least 8, preferably between 16 to 80, consecutive nucleotides that are complementary to a first exon of a dystrophin pre-mRNA and wherein a nucleotide sequence is used which comprises at least 8, preferably between 16 to 80, consecutive nucleotides that are complementary to a second exon of said dystrophin pre-mRNA. Said first and said second exon may be the same.

[0073] In one preferred embodiment said first and said second exon are separated in said dystrophin pre-mRNA by at least one exon to which said oligonucleotide is not complementary. Alternatively, said first and said second exon are adjacent.

[0074] It is possible to specifically promote the skipping of also the intervening exons by providing a linkage between the two complementary oligonucleotides. Hence, in one embodiment stretches of nucleotides complementary to at least two dystrophin exons are separated by a linking moiety. The at least two stretches of nucleotides are thus linked in this embodiment so as to form a single molecule. Further provided is therefore an oligonucleotide, or functional equivalent thereof which is complementary to at least two exons in a dystrophin pre-mRNA, said oligonucleotide or functional equivalent comprising at least two parts wherein a first part comprises an oligonucleotide having at least 8, preferably between 16 to 80, consecutive nucleotides that are complementary to a first of said at least two exons and wherein a second part comprises an oligonucleotide having at least 8, preferably between 16 to 80, consecutive nucleotides that are complementary to a second exon in said dystrophin pre-mRNA. The linkage may be through any means, but is preferably accomplished through a nucleotide linkage. In the latter case, the number of nucleotides that do not contain an overlap between one or the other complementary exon can be zero, but is preferably between 4 to 40 nucleotides. The linking moiety can be any type of moiety capable of linking oligonucleotides. Preferably, said linking moiety comprises at least 4 uracil nucleotides. Currently, many different compounds are available that mimic hybridisation characteristics of oligonucleotides. Such a compound, called herein a functional equivalent of an oligonucleotide, is also suitable for the present invention if such equivalent comprises similar hybridisation characteristics in kind not necessarily in amount. Suitable functional equivalents are mentioned earlier in this description. As mentioned, oligonucleotides of the invention do not have to consist of only oligonucleotides that contribute to hybridisation to the targeted exon. There may be additional material and/or nucleotides added.

[0075] The DMD gene is a large gene, with many different exons. Considering that the gene is located on the X-chromosome, it is mostly boys that are affected, although girls can also be affected by the disease, as they may receive a bad copy of the gene from both parents, or are suffering from a particularly biased inactivation of the functional allele due to a particularly biased X chromosome inactivation in their muscle cells. The protein is encoded by a plurality of exons (79) over a range of at least 2.4 Mb. Deficits may occur in any part of the DMD gene. Skipping of a particular exon or particular exons can, very often, result in a restricted mRNA that encodes a shorter than normal but at least partially functional dystrophin protein. A practical problem in the development of a medicament based on exon-skipping technology is the plurality of mutations that may result in a deficiency in functional dystrophin protein in the cell. Despite the fact that already multiple different mutations can be corrected for by the skipping of a single exon, this plurality of mutations, requires the generation of a series of different pharmaceuticals as for different mutations different exons need to be skipped. An advantage of an oligonucleotide or of a composition comprising at least two distinct oligonucleotide as later defined herein capable of inducing skipping of two or more exons, is that more than one exon can be skipped with a single pharmaceutical. This property is not only useful but also useful in that only a limited number of pharmaceuticals need to be generated for treating many different DMD or particular, severe BMD mutations. Another option now open to the person skilled in the art is to select particularly functional reconstructed dystrophin proteins and produce compounds capable of generating these preferred dystrophin proteins. Such preferred end results are further referred to as mild phenotype dystrophins.

[0076] Dose ranges of oligonucleotide according to the invention are preferably designed on the basis of rising dose studies in clinical trials (in vivo use) for which rigorous protocol requirements exist. A molecule or an oligonucleotide as defined herein may be used at a dose which is ranged between 0.1 and 20 mg/kg, preferably 0.5 and 10 mg/kg.

[0077] In a preferred embodiment, a concentration of an oligonucleotide as defined herein, which is ranged between 0.1 nM and 1 pM is used. Preferably, this range is for in vitro use in a cellular model such as muscular cells or muscle tissue. More preferably, the concentration used is ranged between 0.3 to 400 nM, even more preferably between 1 to 200 nM. If several oligonucleotides are used, this concentra-
tion or dose may refer to the total concentration or dose of oligonucleotides or the concentration or dose of each oligonucleotide added.

[0078] The ranges of concentration or dose of oligonucleotide(s) as given above are preferred concentrations or doses for in vitro or ex vivo uses. The skilled person will understand that depending on the oligonucleotide(s) used, the target cell to be treated, the gene target and its expression levels, the medium used and the transfection and incubation conditions, the concentration or dose of oligonucleotide(s) used may further vary and may need to be optimised any further.

[0079] An oligonucleotide as defined herein for use according to the invention may be suitable for administration to a cell, tissue and/or an organ in vivo of individuals affected by or at risk of developing DMD or BMD, and may be administered in vivo, ex vivo or in vitro. Said oligonucleotide may be directly or indirectly administered to a cell, tissue and/or an organ in vivo of an individual affected by or at risk of developing DMD or BMD, and may be administered directly or indirectly in vivo, ex vivo or in vitro. As Duchenne and Becker muscular dystrophy have a pronounced phenotype in muscle cells, it is preferred that said cells are muscle cells, it is further preferred that said tissue is a muscular tissue and/or it is further preferred that said organ comprises or consists of a muscular tissue. A preferred organ is the heart. Preferably, said cells comprise a gene encoding a mutant dystrophin protein. Preferably, said cells are cells of an individual suffering from DMD or BMD.

[0080] An oligonucleotide of the invention may be indirectly administered using suitable means known in the art. An oligonucleotide may for example be provided to an individual or a cell, tissue or organ of said individual in the form of an expression vector wherein the expression vector encodes a transcript comprising said oligonucleotide. The expression vector is preferably introduced into a cell, tissue, organ or individual via a gene delivery vehicle. In a preferred embodiment, there is provided a viral-based expression vector comprising an expression cassette or a transcription cassette that drives expression or transcription of a molecule as identified herein. A preferred delivery vehicle is a viral vector such as an aden-associated virus vector (AAV), or a retroviral vector such as a lentivirus vector. Also, plasmids, artificial chromosomes, plasmids suitable for targeted homologous recombination and integration in the human genome of cells may be suitably applied for delivery of an oligonucleotide as defined herein. Preferred for the current invention are those vectors wherein transcription is driven from PolIII promoters, and/or wherein transcripts are in the form fusions with U1 or U7 transcripts, which yield good results for delivering small transcripts. It is within the skill of the artisan to design suitable transcripts. Preferred are PolIII driven transcripts. Preferably, in the form of a fusion transcript with an U1 or U7 transcript. Such fusions may be generated as described. The oligonucleotide may be delivered as is. However, the oligonucleotide may also be encoded by the viral vector. Typically, this is in the form of an RNA transcript that comprises the sequence of the oligonucleotide in a part of the transcript.

[0081] Improvements in means for providing an individual or a cell, tissue, organ of said individual with an oligonucleotide and/or an equivalent thereof, are anticipated considering the progress that has already thus far been achieved. Such future improvements may of course be incorporated to achieve the mentioned effect on restructuring of mRNA using a method of the invention. An oligonucleotide and/or an equivalent thereof can be delivered as is to an individual, a cell, tissue or organ of said individual. When administering an oligonucleotide and/or an equivalent thereof, it is preferred that an oligonucleotide and/or an equivalent thereof is dissolved in a solution that is compatible with the delivery method. For intravenous, subcutaneous, intramuscular, intrathecal and/or intraventricular administration it is preferred that the solution is a physiological salt solution. Particularly preferred in the invention is the use of an excipient that will aid in delivery of each of the constituents as defined herein to a cell and/or into a cell, preferably a muscle cell. Preferred are excipients capable of forming complexes, nanoparticles, micelles, vesicles and/or liposomes that deliver each constituent as defined herein, complexed or trapped in a vesicle or liposome through a cell membrane. Many of these excipients are known in the art. Suitable excipients comprise polyethyleneimine (PEI), or similar cationic polymers, including polypropyleneimine or polyethyleneimine copolymers (PECS) and derivatives, synthetic amphipols (SAINT-18), Lipofectin™, DOTAP and/or viral capsid proteins that are capable of self assembly into particles that can deliver each constituent as defined herein to a cell, preferably a muscle cell. Such excipients have been shown to efficiently deliver an oligonucleotide such as antisense nucleic acids to a wide variety of cultured cells, including muscle cells. Their high transfection potential is combined with an excepted low to moderate toxicity in terms of overall cell survival. The ease of structural modification can be used to allow further modifications and the analysis of their further (in vivo) nucleic acid transfer characteristics and toxicity.

[0082] Lipofectin represents an example of a liposomal transfection agent. It consists of two lipid components, a cationic lipid N-[1-(2,3-dioleoyloxy)propyl]-N,N,N,N-trimethylammonium chloride (DOTMA) (cp. DOTAP which is the methylsulfate salt) and a neutral lipid dioleoylphosphatidylethanolamine (DOPE). The neutral component mediates the intracellular release. Another group of delivery systems are polymeric nanoparticles.

[0083] Polycations such like diethylaminoethylaminoethyl (DEAE)-dextran, which are well known as DNA transfection reagent can be combined with butylexyanoacrylate (PBCA) and hexylexyanoacrylate (PHCA) to formate cationic nanoparticles that can deliver each constituent as defined herein, preferably an oligonucleotide across cell membranes into cells.

[0084] In addition to these common nanoparticle materials, the cationic peptide protamine offers an alternative approach to formulate an oligonucleotide with colloids. This colloidal nanoparticle system can form so called proticles, which can be prepared by a simple self-assembly process to package and mediate intracellular release of an oligonucleotide. The skilled person may select and adapt any of the above or other commercially available alternative excipients and delivery systems to package and deliver an oligonucleotide for use in the current invention to deliver it for the treatment of Duchenne Muscular Dystrophy or Becker Muscular Dystrophy in humans.

[0085] In addition, an oligonucleotide could be covalently or non-covalently linked to a targeting ligand specifically designed to facilitate the uptake in to the cell, cytoplasm and/or its nucleus. Such ligand could comprise (i) a compound (including but not limited to peptide(-like) structures)
recognising cell, tissue or organ specific elements facilitating cellular uptake and/or (ii) a chemical compound able to facilitate the uptake in to cells and/or the intracellular release of an oligonucleotide from vesicles, e.g. endosomes or lysosomes. Therefore, in a preferred embodiment, an oligonucleotide is formulated in a composition or a medicament or a composition, which is provided with at least one excipient and/or a targeting ligand for delivery and/or a delivery device thereof to a cell and/or enhancing its intracellular delivery. Accordingly, the invention also encompasses a pharmaceutically acceptable composition comprising an oligonucleotide and further comprising at least one excipient and/or a targeting ligand for delivery and/or a delivery device of said oligonucleotide to a cell and/or enhancing its intracellular delivery. It is to be understood that if a composition comprises an additional constituent such as an adjunct compound as later defined herein, each constituent of the composition may not be formulated in one single combination or composition or preparation. Depending on their identity, the skilled person will know which type of formulation is the most appropriate for each constituent as defined herein. In a preferred embodiment, the invention provides a composition or a preparation which is in the form of a kit of parts comprising an oligonucleotide and a further adjunct compound as later defined herein.

A preferred oligonucleotide is for preventing or treating Duchenne Muscular Dystrophy (DMD) or Becker Muscular Dystrophy (BMD) in an individual. An individual, which may be treated using an oligonucleotide of the invention may already have been diagnosed as having a DMD or a BMD. Alternatively, an individual which may be treated using an oligonucleotide of the invention may not have yet been diagnosed as having a DMD or a BMD but may be an individual having an increased risk of developing a DMD or a BMD in the future given his or her genetic background. A preferred individual is a human being.

Composition

In a further aspect, there is provided a composition comprising an oligonucleotide as defined herein. Preferably, said composition comprises at least two distinct oligonucleotides as defined herein. More preferably, these two distinct oligonucleotides are designed to skip distinct two or more exons as earlier defined herein for multi-exon skipping.

In a preferred embodiment, said composition being preferably a pharmaceutical composition said pharmaceutical composition comprising a pharmaceutically acceptable carrier, adjuvant, diluent and/or excipient. Such a pharmaceutical composition may comprise any pharmaceutically acceptable carrier, diluent, and preservative, adjuvant, solubilizer, diluent and/or excipient is also provided. Such pharmaceutically acceptable carrier, diluent, and/or excipient may for instance be found in Remington: The Science and Practice of Pharmacy, 20th Edition. Baltimore, Md.: Lippincott Williams & Wilkins, 2000. Each feature of said composition has earlier been defined herein.

If several oligonucleotides are used, concentration or dose already defined herein may refer to the total concentration or dose of all oligonucleotides used or the concentration or dose of each oligonucleotide used. Therefore in one embodiment, there is provided a composition wherein each or the total amount of oligonucleotide used is dosed in an amount ranged between 0.5 mg/kg and 10 mg/kg.

A preferred composition additionally comprises:

a) an adjunct compound for reducing inflammation, preferably for reducing muscle tissue inflammation, and/or
b) an adjunct compound for improving muscle fiber function, integrity and/or survival and/or
c) a compound exhibiting readthrough activity.

It has surprisingly been found that the skipping frequency of a dystrophin exon from a pre-mRNA comprising said exon, when using an oligonucleotide directed toward the exon or to one or both splice sites of said exon, is enhanced if cells expressing said pre-mRNA are also provided with an adjunct compound for reducing inflammation, preferably for reducing muscle tissue inflammation, and/or an adjunct compound for improving muscle fiber function, integrity and/or survival. The enhanced skipping frequency also increases the level of functional dystrophin protein produced in a muscle cell of a DMD or BMD individual.

According to the present invention, even when a dystrophin protein deficiency has been restored in a DMD patient by administering an oligonucleotide of the invention, the presence of tissue inflammation and damaged muscle cells still continues to contribute to the symptoms of DMD. Hence, even though the cause of DMD—i.e. a dysfunctional dystrophin protein—is alleviated, treatment of DMD is still further improved by additionally using an adjunct therapy according to the present invention. Furthermore, the present invention provides the insight that a reduction of inflammation does not result in significant reduction of AON uptake by muscle cells. This is surprising because, in general, inflammation enhances the trafficking of cells, blood and other compounds. As a result, AON uptake/delivery is also enhanced during inflammation. Hence, before the present invention it would have been expected that an adjunct therapy countering inflammation involves the risk of negatively influencing AON therapy. This, however, appears not to be the case.

An adjunct compound for reducing inflammation comprises any therapy which is capable of at least in part reducing inflammation, preferably inflammation caused by damaged muscle cells. Said adjunct compound is most preferably capable of reducing muscle tissue inflammation. Inflammation is preferably assessed by detecting an increase in the number of infiltrating immune cells such as neutrophils and/or mast cells and/or dendritic cells and/or lymphocytes in muscle tissue suspected to be dystrophic. This assessment is preferably carried out in cross-sections of a biopsy of muscle tissue suspected to be dystrophic after having specifically stained immune cells as identified above. The quantification is preferably carried out under the microscope. Reducing inflammation is therefore preferably assessed by detecting a decrease in the number of immune cells in a cross-section of muscle tissue suspected to be dystrophic. Detecting a decrease preferably means that the number of at least one sort of immune cells as identified above is decreased of at least 1%, 2%, 3%, 5%, 7%, 10%, 12%, 15%, 17%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or more compared to the number of a corresponding immune cell in a same individual before treatment. Most preferably, no infiltrating immune cells are detected in cross-sections of said biopsy.

An adjunct compound for improving muscle fiber function, integrity and/or survival comprises any therapy, which is capable of measurably enhancing muscle fiber function, integrity and/or survival as compared to an otherwise
similar situation wherein said adjunct compound is not present. The improvement of muscle fiber function, integrity and/or survival may be assessed using at least one of the following assays: a detectable decrease of creatine kinase in blood, a detectable decrease of necrosis of muscle fibers in a biopsy cross-section of a muscle suspected to be dystrophic, and/or a detectable increase of the homogeneity of the diameter of muscle fibers in a biopsy cross-section of a muscle suspected to be dystrophic. Each of these assays is known to the skilled person.

[0099] Creatine kinase may be detected in blood as described in 57. A detectable decrease in creatine kinase may mean a decrease of 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or more compared to the concentration of creatine kinase in a same individual before treatment.

[0100] A detectable decrease of necrosis of muscle fibers is preferably assessed in a muscle biopsy, more preferably as described in 57 using biopsy cross-sections. A detectable decrease of necrosis may be a decrease of 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or more of the area wherein necrosis has been identified using biopsy cross-sections. The decrease is measured by comparison to the necrosis as assessed in a same individual before treatment.

[0101] A detectable increase of the homogeneity of the diameter of a muscle fiber is preferably assessed in a muscle biopsy cross-section, more preferably as described in 57.

[0102] In one embodiment, an adjunct compound for increasing turnover of damaged muscle cells is used. An adjunct compound for increasing turnover of damaged muscle cells comprises any therapy, which is capable of at least in part inducing and/or increasing turnover of damaged muscle cells. Damaged muscle cells are muscle cells, which have significantly less clinically measurable functionality than a healthy, intact muscle cell. In the absence of dystrophin, mechanical stress leads to sarcornemal ruptures, causing an uncontrolled influx of calcium into the muscle fiber interior, thereby triggering calcium-activated proteases and fiber necrosis, resulting in damaged muscle cells. Increasing turnover of damaged muscle cells means that damaged muscle cells are more quickly broken down and/or removed as compared to a situation wherein turnover of damaged muscle cells is not increased. Turnover of damaged muscle cells is preferably assessed in a muscle biopsy, more preferably as described in 57 using a cross-section of a biopsy. A detectable increase of turnover may be an increase of 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or more of the area wherein turnover has been identified using a biopsy cross-section. The increase is measured by comparison to the turnover as assessed in a same individual before treatment.

[0103] Without wishing to be bound to theory, it is believed that increasing turnover of muscle cells is preferred because this reduces inflammatory responses.

[0104] According to the present invention, a composition of the invention further comprising an adjunct therapy for reducing inflammation, preferably for reducing muscle tissue inflammation in an individual, is particularly suitable for use as a medicament. Such composition is even better capable of alleviating one or more symptom(s) of Duchenne Muscular Dystrophy or Becker Muscular Dystrophy as compared to a combination not comprising said adjunct compound. This embodiment also enhances the skipping frequency of a dystrophin exon from a pre-mRNA comprising said exon, when using an oligonucleotide directed toward the exon or to one or both splice sites of said exon. The enhanced skipping frequency also increases the level of functional dystrophin protein produced in a muscle cell of a DMD or BMD individual.

[0105] Further provided is therefore a composition further comprising an adjunct compound for reducing inflammation, preferably for reducing muscle tissue inflammation in said individual, for use as a medicament, preferably for treating or preventing counteracting DMD. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein or altered or truncated dystrophin mRNA or protein is formed which is not sufficiently functional.

[0106] Preferred adjunct compound for reducing inflammation include a steroid, a TNFα inhibitor, a source of mGf1 and/or an antioxidant. However, any other compound able to reduce inflammation as defined herein is also encompassed within the present invention. Each of these compounds is later on exclusively presented. Each of the compounds extensively presented may be used separately or in combination with each other and/or in combination with one or more of the adjunct compounds used for improving muscle fiber function, integrity and/or survival.

[0107] Furthermore, a composition comprising an adjunct therapy for improving muscle fiber function, integrity and/or survival in an individual is particularly suitable for use as a medicament, preferably for treating or preventing DMD. Such composition is even better capable of alleviating one or more symptom(s) of Duchenne Muscular Dystrophy as compared to a composition not comprising said adjunct compound.

[0108] Preferred adjunct compounds for improving muscle fiber function, integrity and/or survival include an ion channel inhibitor, a protease inhibitor, L-arginine and/or an angiotensin II type I receptor blocker. However, any other compound able to improve muscle fiber function, integrity and/or survival as defined herein is also encompassed within the present invention. Each of these compounds is later on exclusively presented. Each of the compounds extensively presented may be used separately or in combination with each other and/or in combination with one or more of the adjunct compounds used for reducing inflammation.

[0109] In a particularly preferred embodiment, a composition further comprises a steroid. Such composition results in significant alleviation of DMD symptoms. This embodiment also enhances the skipping frequency of a dystrophin exon from a pre-mRNA comprising said exon, when using an oligonucleotide directed toward the exon or to one or both splice sites of said exon. The enhanced skipping frequency also increases the level of functional dystrophin protein produced in a muscle cell of a DMD or BMD individual.

[0110] In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional.

[0111] A steroid is a terpenoid lipid characterized by a carbon skeleton with four fused rings, generally arranged in a 6-6-6-5 fashion. Steroids vary by the functional groups attached to these rings and the oxidation state of the rings. Steroids include hormones and drugs, which are usually used to relieve swelling and inflammation, such as for instance prednisone, dexamethasone and vitamin D.

[0112] According to the present invention, supplemental effects of adjunct steroid therapy in DMD patients include reduction of tissue inflammation, suppression of cytotoxic cells, and improved calcium homeostasis. Most positive
results are obtained in younger boys. Preferably, the steroid is a corticosteroid, more preferably, a glucocorticosteroid. Preferably, prednisone steroids such as prednisone, prednizolone or delfazacort are used in a combination according to the invention\(^1\). Dose ranges of steroid or of a glucocorticosteroid to be used in the therapeutic applications as described herein are designed on the basis of rising dose studies in clinical trials for which rigorous protocol requirements exist. The usual doses are 0.5-1.0 mg/kg/day, preferably 0.75 mg/kg/day for prednisone, prednizolone and 0.4-1.4 mg/kg/day, preferably 0.9 mg/kg/day for delfazacort.

In one embodiment, a steroid is administered to said individual prior to administering a composition as earlier defined herein. In this embodiment, it is preferred that said steroid is administered at least one day, more preferably at least one week, more preferably at least two weeks, more preferred at least three weeks prior to administering said composition.

In another preferred embodiment, a combination further comprises a tumour necrosis factor-alpha (TNF\(\alpha\)) inhibitor. Tumour necrosis factor-alpha (TNF\(\alpha\)) is a pro-inflammatory cytokine that stimulates the inflammatory response. Pharmacological blockade of TNF\(\alpha\) activity with the neutralizing antibody infliximab (Remicade) is highly effective clinically at reducing symptoms of inflammatory diseases. In mdx mice, both infliximab and etanercept delay and reduce the necrosis of dystrophic muscle\(^2\),\(^5\),\(^7\) with additional physiological benefits on muscle strength, chloride channel function and reduced CK levels being demonstrated in chronically treated exercised adult mdx mice\(^2\). Such highly specific anti-inflammatory drugs designed for use in other clinical conditions, are attractive alternatives to the use of steroids for DMD. In one embodiment, the use of a TNF\(\alpha\) inhibitor is limited to periods of intensive muscle growth in boys when muscle damage and deterioration are especially pronounced.

In another preferred embodiment, a composition further comprising a TNF\(\alpha\) inhibitor for use as a medicament is also provided. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional. A preferred TNF\(\alpha\) inhibitor is a dimeric fusion protein consisting of the extracellular ligand-binding domain of the human p75 receptor of TNF\(\alpha\) linked to the Fc portion of human IgG1. A more preferred TNF\(\alpha\) inhibitor is etanercept (Amgen, America)\(^5\). The usual doses of etanercept are about 0.2 mg/kg, preferably about 0.5 mg/kg twice a week. The administration is preferably subcutaneous.

In another preferred embodiment, a composition of the invention further comprises a source of mIGF-1. As defined herein, a source of IGF-1 preferably encompasses mIGF-1 itself, a compound able of enhancing mIGF-1 expression and/or activity. Enhancing is herein synonymous with increasing. Expression of mIGF-1 is synergistic with amount of mIGF-1. mIGF-1 promotes regeneration of muscles through increase in satellite cell activity, and reduces inflammation and fibrosis\(^2\). Local injury of muscle results in increased mIGF-1 expression. In transgenic mice with extra IGF-1 genes, muscle hypertrophy and enlarged muscle fibers are observed\(^2\). Similarly, transgenic mdx mice show reduced muscle fiber degeneration\(^7\). Uregulation of the mIGF-1 gene and/or administration of extra amounts of mIGF-1 protein or a functional equivalent thereof (especially the mIGF-1 Es isoform as described in 27, human homolog IGF-1 isoform 4: SEQ ID NO: 577) thus promotes the effect of other, preferably genetic, therapies for DMD, including antisense-induced exon skipping. The additional mIGF-1 levels in the above mentioned transgenic mice do not induce cardiac problems nor promote cancer, and have no pathological side effects. As stated before, the amount of mIGF-1 is for instance increased by enhancing expression of the mIGF-1 gene and/or by administration of mIGF-1 protein and/or a functional equivalent thereof (especially the mIGF-1 Es isoform as described in 27, human homolog IGF-1 isoform 4: SEQ ID NO: 577). A composition of the invention further preferably comprises mIGF-1, a compound capable of enhancing mIGF-1 expression and/or an mIGF-1 activity, for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, such composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional.

Within the context of the invention, an increased amount of mIGF-1 may be reached by increasing the gene expression level of an IGF-1 gene, by increasing the amount of a corresponding IGF-1 protein and/or by increasing an activity of an IGF-1 protein. A preferred mIGF-1 protein has been earlier defined herein. An increase of an activity of said protein is herein understood to mean any detectable change in a biological activity exerted by said protein or in the steady state level of said protein as compared to said activity or steady-state in a individual who has not been treated. Increased amount or activity of mIGF-1 is preferably assessed by detection of increased expression of muscle hypertrophy biomarker GATA-2 (as described in 27).

Gene expression level is preferably assessed using classical molecular biology techniques such as (real time) PCR, arrays or Northern analysis. A steady state level of a protein is determined directly by quantifying the amount of a protein. Quantifying a protein amount may be carried out by any known technique such as Western blotting or immunoassay using an antibody raised against a protein. The skilled person will understand that alternatively or in combination with the quantification of a gene expression level and/or a corresponding protein, the quantification of a substrate of a corresponding protein or of any compound known to be associated with a function or activity of a corresponding protein or the quantification of said function or activity of a corresponding protein using a specific assay may be used to assess the alteration of an activity or steady state level of a protein.

In the invention, an activity or steady-state level of a said protein may be altered at the level of the protein itself, e.g., by providing a protein to a cell from an exogenous source.

Preferably, an increase or an up-regulation of the expression level of a said gene means an increase of at least 5% of the expression level of said gene using arrays. More preferably, an increase of the expression level of said gene means an increase of at least 10%, even more preferably at least 20%, at least 30%, at least 40%, at least 50%, at least 70%, at least 90%, at least 150% or more. In another preferred embodiment, an increase of the expression level of said protein means an increase of at least 5% of the expression level of said protein using Western blotting and/or using ELISA or a suitable assay. More preferably, an increase of the expression level of a protein means an increase of at least 10%, even more preferably at least 20%, at least 30%, at least 40%, at least 50%, at least 70%, at least 90%, at least 150% or more.
In another preferred embodiment, an increase of a polypeptide activity means an increase of at least 5% of a polypeptide activity using a suitable assay. More preferably, an increase of a polypeptide activity means an increase of at least 10%, even more preferably at least 20%, at least 30%, at least 40%, at least 50%, at least 70%, at least 90%, or at least 150% or more. The increase is preferably assessed by comparison to corresponding activity in the individual before treatment.

A preferred way of providing a source of mFGF is to introduce a transgene encoding mFGF, preferably an mFGF-1 Ea isoform (as described in 27, human homolog IGF-1 isoform 4; SEQ ID NO: 577), more preferably in an AAV vector as later defined herein. Such source of mFGF is specifically expressed in muscle tissue as described in mice in 27.

In another preferred embodiment, a composition further comprises an antioxidant. Oxidative stress is an important factor in the progression of DMD and promotes chronic inflammation and fibrosis. The most prevalent products of oxidative stress, the peroxidized lipids, are increased by an average of 35% in Duchenne boys. Increased levels of the enzymes superoxide dismutase and catalase reduce the excessive amount of free radicals causing these effects. In fact, a dietary supplement Protandim® (LifeVantage) was clinically tested and found to increase levels of superoxide dismutase (up to 30%) and catalase (up to 54%), which indeed significantly inhibited the peroxidation of lipids in 29 healthy persons. Such effective management of oxidative stress thus preserves muscle quality and so promotes the positive effect of DMD therapy. Idebeneone is another potent antioxidant with a chemical structure derived from natural coenzyme Q10. It protects mitochondria where adenosine triphosphate, ATP, is generated by oxidative phosphorylation. The absence of dystrophin in DMD negatively affects this process in the heart, and probably also in skeletal muscle. Idebeneone was recently applied in clinical trials in the US and Europe demonstrating efficacy on neurological aspects of Friedreich’s Ataxia. A phase-IIa double-blind, placebo-controlled randomized clinical trial with Idebeneone has recently been started in Belgium, including 21 Duchenne boys at 8 to 16 years of age. The primary objective of this study is to determine the effect of Idebeneone on heart muscle function. In addition, several different tests will be performed to detect the possible functional benefit on muscle strength in the patients. When effective, idebeneone is a preferred adjunct compound for use in a combination according to the present invention in order to enhance the therapeutic effect of DMD therapy, especially in the heart. A composition further comprising an antioxidant for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional. Depending on the identity of the antioxidant, the skilled person will know which quantities are preferably used. An antioxidant may include bacoside, silimarin, curcumin and/or a polyphenol. Preferably, a polyphenol is or comprises epigallocatechin-3-gallate (EGCG). Preferably, an antioxidant is a mixture of antioxidants as the dietary supplement Protandim® (LifeVantage). A daily capsule of 675 mg of Protandim® comprises 150 mg of B. monniera (45% bacosides), 225 mg of S. marianum (70-80% silimarin), 150 mg of W. somnifera powder, 75 mg green tea (98% polyphenols wherein 45% EGCG) and 75 mg tumeric (95% curcumin).

In another preferred embodiment, a composition further comprises an ion channel inhibitor. The presence of damaged muscle membranes in DMD disturbs the passage of calcium ions into the myofibers, and the consequently disrupted calcium homeostasis activates many enzymes, e.g. proteases, that cause additional damage and muscle necrosis. Ion channels that directly contribute to the pathological accumulation of calcium in dystrophic muscle are potential targets for adjunct compounds to treat DMD. There is evidence that some drugs, such as pentoxifylline, block exercise-sensitive calcium channels and antibiotics that block stretch activated channels reduce myofibre necrosis in mdx mice and CK levels in DMD boys. A composition further comprising an ion channel inhibitor for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional.

Preferably, an ion channel inhibitor of the class of xanthines is used. More preferably, said xanthines are derivatives of methylxanthines, and most preferably, said methylxanthine derivatives are chosen from the group consisting of pentoxifylline, furafylline, l-isoxylline, propentofylline, pentifylline, theophylline, theobromine, albutyryl, enprofylline and derivatives thereof. Most preferably is the use of pentoxifylline. Ion channel inhibitors of the class of xanthines enhance the skipping frequency of a dystrophin exon from a pre-mRNA comprising said exon, when using an oligonucleotide directed toward the exon or to one or both splice sites of said exon. The enhanced skipping frequency also increases the level of functional dystrophin protein produced in a muscle cell of a DMD or BMD individual.

Depending on the identity of the ion channel inhibitor, the skilled person will know which quantities are preferably used. Suitable dosages of pentoxifylline are between 1 mg/kg/day to 100 mg/kg/day, preferred dosages are between 10 mg/kg/day to 50 mg/kg/day. Typical dosages used in humans are 20 mg/kg/day.

In one embodiment, an ion channel inhibitor is administered to said individual prior to administering a composition comprising an oligonucleotide. In this embodiment, it is preferred that said ion channel inhibitor is administered at least one day, more preferred at least one week, more preferred at least two weeks, more preferred at least three weeks prior to administering a composition comprising an oligonucleotide.

In another preferred embodiment, a composition further comprises a protease inhibitor. Calpains are calcium-activated proteases that are increased in dystrophic muscle and account for myofiber degeneration. Calpain inhibitors such as calpastatin, leupeptin, calpeptin, calpain inhibitor III, or PD150606 are therefore applied to reduce the degeneration process. A new compound, BN 82270 (Ipsen) that has dual action as both a calpain inhibitor and an antioxidant increased muscle strength, decreased serum CK and reduced fibrosis of the mdx diaphragm, indicating a therapeutic effect with this new compound. Another compound of Leupeptin/Carminite (Myochr) has recently been proposed for clinical trials in DMD patients.

MG132 is another proteasomal inhibitor that has shown to reduce muscle membrane damage, and to amelio-
rate the histopathological signs of muscular dystrophy. MG-132 (CBZ-leucyl-leucyl-leucinal) is a cell-permeable, proteasomal inhibitor (Ki=4 nM), which inhibits NF kappa B activation by preventing Ikappa B degradation (IC50=3 μM). In addition, it is a peptide aldehyde that inhibits ubiquitin-mediated proteolysis by binding to and inactivating 20S and 26S proteasomes. MG-132 has shown to inhibit the proteasomal degradation of dystrophin-associated proteins in the dystrophic mdx mouse model. This compound is thus also suitable for use as an adjunct pharmacological compound for DMD. A composition further comprising a protease inhibitor for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, said combination is used in order to alleviate one or more symptom(s) of a severe form of DMD wherein a very short dystrophin protein is formed which is not sufficiently functional. Depending on the identity of the protease inhibitor, the skilled person will know which quantities are preferably used.

In another preferred embodiment, a composition further comprises L-arginine. Dystrophin-deficiency is associated with the loss of the DGC-complex at the fiber membranes, including neuronal nitric oxide synthase (nNOS). Expression of a nNOS transgene in mdx mice greatly reduced muscle membrane damage. Similarly, administration of L-arginine (the substrate for nitric oxide synthase) increased NO production and upregulated utrophin expression in mdx mice. Six weeks of L-arginine treatment improved muscle pathology and decreased serum CK in mdx mice. The use of L-arginine as a further constituent in a composition of the invention has not been disclosed.

A composition further comprising L-arginine for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional.

In another preferred embodiment, a composition further comprises angiotensin II type 1 receptor blocker Losartan, which normalizes muscle architecture, repair and function, as shown in the dystrophin-deficient mdx mouse model. A composition further comprising angiotensin II type 1 receptor blocker Losartan for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional. Depending on the identity of the angiotensin II type 1 receptor blocker, the skilled person will know which quantities are preferably used.

In another preferred embodiment, a composition further comprises an angiotensin-converting enzyme (ACE) inhibitor, preferably perindopril. ACE inhibitors are capable of lowering blood pressure. Early initiation of treatment with perindopril is associated with a lower mortality in DMD patients. A composition further comprising an ACE inhibitor, preferably perindopril for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional. The usual doses of an ACE inhibitor, preferably perindopril are about 2 to 4 mg/day. In a more preferred embodiment, an ACE inhibitor is combined with at least one of the previously identified adjunct compounds.

In another preferred embodiment, a composition further comprises a compound exhibiting a readthrough activity. A compound exhibiting a readthrough activity may be any compound, which is able to suppress a stop codon. For 20% of DMD patients, the mutation in the dystrophin gene is comprising a point mutation, of which 13% is a nonsense mutation. A compound exhibiting a readthrough activity or which is able to suppress a stop codon is a compound which is able to provide an increased amount of a functional dystrophin mRNA or protein and/or a decreased amount of an aberrant or truncated dystrophin mRNA or protein. Increased preferably means increased of at least 1%, 2%, 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 100% or more. Decreased preferably means decreased of at least 1%, 2%, 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 95%, 100% or more. An increase or a decrease of said protein is preferably assessed in a muscular tissue or in a muscular cell of an individual by comparison to the amount present in said individual before treatment with said compound exhibiting a readthrough activity. Alternatively, the comparison can be made with a muscular tissue or cell of said individual which has not yet been treated with said compound in case the treatment is local. The assessment of an amount at the protein level is preferably carried out using western blot analysis.

Preferred compounds exhibiting a readthrough activity comprise or consist of aminoglycosides, including, but not limited to, gentamicin (G418), paromomycin, gentamycin and/or 3-(5-(2-fluorophenyl)-1,2,4-oxadiazol-3-yl)benzoic acid, and derivatives thereof (references 64, 65).

A more preferred compound exhibiting a readthrough activity comprises or consists of PTC124™, and/or a functional equivalent thereof. PTC124™ is a registered trademark of PTC Therapeutics, Inc. South Plainfield, N.J. 3-(5-(2-fluorophenyl)-1,2,4-oxadiazol-3-yl)benzoic acid) also known as PTC124™ (references 16, 17) belongs to a new class of small molecules that mimics at lower concentrations the readthrough activity of gentamicin (reference 55). A functional equivalent of 34542-fluorophenyl)-1,2,4-oxadiazol-3-yl)benzoic acid) or of gentamicin is a compound which is able to exhibit a readthrough activity as earlier defined herein. Most preferably, a compound exhibiting a readthrough activity comprises or consists of gentamicin and/or 3-(5-(2-fluorophenyl)-1,2,4-oxadiazol-3-yl)benzoic acid) also known as PTC124™. A composition further comprising a compound exhibiting a readthrough activity, preferably comprising or consisting of gentamicin and/or 3-(5-(2-fluorophenyl)-1,2,4-oxadiazol-3-yl)benzoic acid) for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional. The usual doses of a compound exhibiting a readthrough activity, preferably 34542-fluorophenyl)-1,2,4-oxadiazol-3-yl)benzoic acid) or of gentamicin are ranged between 3 mg/kg/day to 200 mg/kg/day, preferred dosages are between 10 mg/kg to 50 mg/kg per day or twice a day.
In a more preferred embodiment, a compound exhibiting a readthrough activity is combined with at least one of the previously identified adjunct compounds.

In another preferred embodiment, a composition further comprises a compound, which is capable of enhancing exon skipping and/or inhibiting splicingosome assembly and/or splicing. Small chemical compounds, such as for instance specific indole derivatives, have been shown to selectively inhibit splicingosome assembly and splicing, for instance by interfering with the binding of serine- and arginine-rich (SR) proteins to their cognate splicing enhancers (ISEs or ISEs) and/or by interfering with the binding of splicing repressors to silencer sequences (ESSSs or ISSSs). These compounds are therefore suitable for applying as adjunct compounds that enhance exon skipping. A composition further comprising a compound for enhancing exon skipping and/or inhibiting splicingosome assembly and/or splicing for use as a medicament is also provided. Said medicament is preferably for alleviating one or more symptom(s) of DMD. In one embodiment, said composition is used in order to alleviate one or more symptom(s) of a severe form of BMD wherein a very short dystrophin protein is formed which is not sufficiently functional. Depending on the identity of the compound, which is capable of enhancing exon skipping and/or inhibiting splicingosome assembly and/or splicing, the skilled person will know which quantities are preferably used. In a more preferred embodiment, a compound for enhancing exon skipping and/or inhibiting splicingosome assembly and/or splicing is combined with an ACE inhibitor and/or with any adjunct compounds as identified earlier herein.

The invention thus provides a composition further comprising an adjunct compound, wherein said adjunct compound comprises a steroid, an ACE inhibitor (preferably perindopril), angiotensin II type 1 receptor blocker Losartan, a tumour necrosis factor-alpha (TNFac) inhibitor, a source of mGF-1, preferably mGF-1, a compound for enhancing mGF-1 activity, a compound for enhancing mGF-1 expression, a compound for enhancing mGF-1 activity, an antioxidant, an ion channel inhibitor, a protease inhibitor, L-arginine, a compound exhibiting a readthrough activity and/or inhibiting splicingosome assembly and/or splicing.

In one embodiment an individual is further provided with a functional dystrophin protein using a vector, preferably a viral vector, comprising a micro-mini-dystrophin gene. Most preferably, a recombiant adeno-associated viral (rAAV) vector is used. AAV is a single-stranded DNA parvovirus that is non-pathogenic and shews a helper-dependent life cycle. In contrast to other viruses (adenovirus, retrovirus, and herpes simplex virus), rAAV vectors have demonstrated to be very efficient in transducing mature skeletal muscle. Application of rAAV in classical DMD "gene addition" studies has been hindered by its restricted packaging limits (<5 kb). Therefore, rAAV is preferably applied for the efficient delivery of a much smaller micro- or mini-dystrophin gene. Administration of such micro- or mini-dystrophin gene results in the presence of an at least partially functional dystrophin protein. Reference is made to18-20.

Each constituent of a composition can be administered to an individual in any order. In one embodiment, each constituent is administered simultaneously (meaning that each constituent is administered within 10 hours, preferably within one hour). This is however not necessary. In one embodiment at least one adjunct compound is administered to an individual in need thereof before administration of an oligonucleotide. Alternatively, an oligonucleotide is administered to an individual in need thereof before administration of at least one adjunct compound.

Use

In a further aspect, there is provided the use of an oligonucleotide or of a composition as defined herein for the manufacture of a medicament for preventing or treating Duchenne Muscular Dystrophy or Becker Muscular Dystrophy in an individual. Each feature of said use has earlier been defined herein.

A treatment in a use or in a method according to the invention is at least one week, at least one month, at least several months, at least one year, at least 2, 3, 4, 5, 6 years or more. Each molecule or oligonucleotide or equivalent thereof as defined herein for use according to the invention may be suitable for direct administration to a cell, tissue and/or an organ in vivo of individuals affected by or at risk of developing DMD or BMD, and may be administered directly in vivo, ex vivo or in vitro. The frequency of administration of an oligonucleotide, composition, compound or adjunct compound of the invention may depend on several parameters such as the age of the patient, the mutation of the patient, the number of molecules (i.e. dose), the formulation of said molecule. The frequency may be ranged between at least once in two weeks, or three weeks or four weeks or five weeks or a longer time period.

Method

In a further aspect, there is provided a method for alleviating one or more symptom(s) of Duchenne Muscular Dystrophy or Becker Muscular Dystrophy in an individual or alleviate one or more characteristic(s) of a myogenic or muscle cell of said individual, the method comprising administering to said individual an oligonucleotide or a composition as defined herein.

There is further provided a method for enhancing, inducing or promoting skipping of an exon from a dystrophin pre-mRNA in a cell expressing said pre-mRNA in an individual suffering from Duchenne Muscular Dystrophy or Becker Muscular Dystrophy, the method comprising administering to said individual an oligonucleotide or a composition as defined herein. Further provided is a method for increasing the production of a functional dystrophin protein and/or decreasing the production of an aberrant dystrophin protein in a cell, said cell comprising a pre-mRNA of a dystrophin gene encoding an aberrant dystrophin protein, the method comprising providing said cell with an oligonucleotide or composition of the invention and allowing translation of mRNA produced from splicing of said pre-mRNA. In one embodiment, said method is performed in vitro, for instance using a cell culture. Preferably, said method is in vivo.

Unless otherwise indicated each embodiment as described herein may be combined with another embodiment as described herein.

In this document and in its claims, the verb "to comprise" and its conjugations is used in its non-limiting sense to mean that items following the word are included, but items not specifically mentioned are not excluded. In addition the verb "to consist" may be replaced by "to consist essentially of" meaning that a compound or adjunct compound as...
defined herein may comprise additional component(s) than the ones specifically identified, said additional component(s) not altering the unique characteristic of the invention.

[0149] In reference to an element by the indefinite article “a” or “an” does not exclude the possibility that more than one of the element is present, unless the context clearly requires that there be one and only one of the elements. The indefinite article “a” or “an” thus usually means “at least one”.

[0150] The word “approximately” or “about” when used in association with a numerical value (approximately 10, about 10) preferably means that the value may be the given value of 10 more or less 1% of the value.

[0151] All patent and literature references cited in the present specification are hereby incorporated by reference in their entirety. Each embodiment as identified herein may be combined together unless otherwise indicated.

[0152] The invention is further explained in the following examples. These examples do not limit the scope of the invention, but merely serve to clarify the invention.

BRIEF DESCRIPTION OF THE DRAWINGS

[0153] FIG. 1. In human control myotubes, PS220 and PS305 both targeting an identical sequence within exon 45, were directly compared for relative skipping efficiencies. PS220 reproducibly induced highest levels of exon 45 skipping (up to 73%), whereas with PS305 maximum exon 45 skipping levels of up to 46% were obtained. No exon 45 skipping was observed in non-treated cells. (M: DNA size marker; NT: non-treated cells)

[0154] FIG. 2. Graph showing relative exon 45 skipping levels of inosine-containing AONs as assessed by RT-PCR analysis. In human control myotubes, a series of new AONs, all targeting exon 45 and containing one inosine for guanosine substitution were tested for relative exon 45 skipping efficiencies when compared with PS220 and PS305 (see FIG. 1). All new inosine-containing AONs were effective, albeit at variable levels (between 4% and 25%). PS220 induced highest levels of exon 45 skipping (up to 72%), whereas with PS305 maximum exon 45 skipping levels of up to 63% were obtained. No exon 45 skipping was observed in non-treated cells. (M: DNA size marker; NT: non-treated cells).

EXAMPLES

Example 1

Materials and Methods

[0155] AON design was based on (partly) overlapping open secondary structures of the target exon RNA as predicted by the m-fold program, on (partly) overlapping putative SR-protein binding sites as predicted by the ESE-finder software. AONs were synthesized by Prosensa Therapeutics B.V. (Leiden, Netherlands), and contain 2’-O-methyl RNA and full-length phosphorothioate (PS) backbones.

Tissue Culturing, Transfection and RT-PCR Analysis

[0156] Myotube cultures derived from a healthy individual (“human control”) (examples 1, 3, and 4; exon 43, 50, 52 skipping) or a DMD patient carrying an exon 45 deletion (example 2; exon 46 skipping) were processed as described previously (Aartsma-Rus et al., Neuromuscul. Disord. 2002; 12: S71-77 and Hum Mol Genet. 2003; 12(8): 907-14). For the screening of AONs, myotube cultures were transfected with 200 nM for each AON (PS220 and PS305). Transfection reagent UNIFectylin (Prosensa Therapeutics BV, Netherlands) was used, with 2 μg UNIFectylin per μg AON. Exon skipping efficiencies were determined by nested RT-PCR analysis using primers in the exons flanking the targeted exon 45. PCR fragments were isolated from agarose gels for sequence verification. For quantification, the PCR products were analyzed using the DNA 1000 LabChip Kit on the Agilent 2100 bioanalyzer (Agilent Technologies, USA).

Results

[0157] DMD exon 45 skipping.

[0158] Two AONs, PS220 (SEQ ID NO: 76; 5’-UUUGCCUGCCCAUGCAUCCUG-3’) and PS305 (SEQ ID NO: 557; 5’-UUUGCCUGCCCAUGCAUCCUG-3’) both targeting an identical sequence within exon 45, were directly compared for relative skipping efficiencies in healthy control myotube cultures. Subsequent RT-PCR and sequence analysis of isolated RNA demonstrated that both AONs were indeed capable of inducing exon 45 skipping. PS220, consisting a GCCGCT stretch, reproducibly induced highest levels of exon 45 skipping (up to 73%), as shown in FIG. 1. However, PS305, which is identical to PS220 but containing an inosine for a G substitution at position 4 within that stretch is also effective and leading to exon 45 skipping levels of up to 46%. No exon 45 skipping was observed in non-treated cells (NT).

Example 2

Materials and Methods

[0159] AON design was based on (partly) overlapping open secondary structures of the target exon 45 RNA as predicted by the m-fold program, on (partly) overlapping putative SR-protein binding sites as predicted by the ESE-finder software. AONs were synthesized by Prosensa Therapeutics B.V. (Leiden, Netherlands), and contain 2’-O-methyl RNA, full-length phosphorothioate (PS) backbones and one inosine for guanosine substitution.

Tissue Culturing, Transfection and RT-PCR Analysis

[0160] Myotube cultures derived from a healthy individual (“human control”) were processed as described previously (Aartsma-Rus et al., Neuromuscul. Disord. 2002; 12: S71-77 and Hum Mol Genet. 2003; 12(8): 907-14). For the screening of AONs, myotube cultures were transfected with 200 nM for each AON. Transfection reagent UNIFectylin (Prosensa Therapeutics BV, Netherlands) was used, with 2 μg UNIFectylin per μg AON. Exon skipping efficiencies were determined by nested RT-PCR analysis using primers in the exons flanking the targeted exon 45. PCR fragments were isolated from agarose gels for sequence verification. For quantification, the PCR products were analyzed using the DNA 1000 LabChip Kit on the Agilent 2100 bioanalyzer (Agilent Technologies, USA).

Results

[0161] DMD exon 45 skipping.

[0162] An additional series of AONs targeting exon 45 and containing one inosine-substitution were tested in healthy control myotube cultures for exon 45 skipping efficiencies, and directly compared to PS220 (without inosine; SEQ ID NO: 76)) and PS305 (identical sequence as PS220 but with inosine substitution; SEQ ID NO: 557). Subsequent RT-PCR and sequence analysis of isolated RNA demonstrated that all
new AONs (PS309 to PS316) were capable of inducing exon 45 skipping between 4% (PS311) and 25% (PS310) as shown in FIG. 2. When compared to PS220 and PS305, PS220 induced highest levels of exon 45 skipping (up to 72%). Of the new inosine-containing AONs PS305 was most effective, showing exon 45 skipping levels of up to 63%. No exon 45 skipping was observed in non-treated cells (NT).

REFERENCES


[0223] 60. Dorn and Kippenberger, Curr Opin Mol Ther 2008 10(1) 10-20


[0228] 65. ... Nucleicman et al., 2006, Bioorg Med Chem Lett 16(24), 6310-5
Sequence listing

MDM gene amino acid sequence

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Human IGF-1 Isoform 4 amino acid sequence
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<400> SEQUENCE: 155

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<400> SEQUENCE: 161
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gcuucuuc uaccuuaagc uucag

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ccaucuc cccagcaaug uguu

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cuagcuucc agccauugug uugaa

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uaagcuucca gccauugugu ugsu

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gcuuccagcc cauugugugu auccu

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cuuccagcca uugugugas uccuu

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<220> FEATURE:
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<400> SEQUENCE: 181
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<400> SEQUENCE: 182

uuccagccau guguguauc ccuua

<210> SEQ ID NO 183
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<400> SEQUENCE: 183

cucgccau guguguauc uuuu

<210> SEQ ID NO 184
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<400> SEQUENCE: 184

cucgccau guguguauc uuuu

<210> SEQ ID NO 185
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<400> SEQUENCE: 185

cagc caugu guugu gauc uuuu

<210> SEQ ID NO 186
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<400> SEQUENCE: 186

cagc caugu guugu gauc uuuu

<210> SEQ ID NO 187
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<220> FEATURE:
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<400> SEQUENCE: 187

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cauugguuc aauccuuaa caaucu

ucagcucuc uuaagccacug

uucagcucu guaagccacu

uucagcucu guaagccacug g

uucagcucuc uuaagccacug a

uucagcucu guaagccacu ga

uucagcucu guaagccacu ga
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<400> SEQUENCE: 194
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<400> SEQUENCE: 195
ucagcuucuc guuagccacu ga

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<212> TYPE: RNA
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<400> SEQUENCE: 196
ucagcuucuc uuaagccacug au

<210> SEQ ID NO 197
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<400> SEQUENCE: 197
ucagcuucuc guuagccacu gau

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<400> SEQUENCE: 198
ucagcuucuc guuagccacu auu

<210> SEQ ID NO 199
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<400> SEQUENCE: 199
ucagcuucuc guuagccacu gauu

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<212> TYPE: RNA
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<400> SEQUENCE: 200
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<400> SEQUENCE: 201
ucagcuccug guuagccacu guua

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<400> SEQUENCE: 202
ucagcuccug uuuagccacug auua

<210> SEQ ID NO 203
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<400> SEQUENCE: 203
ucagcuccug uuagccacug auua

<210> SEQ ID NO 204
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<400> SEQUENCE: 204
ucagcuccug uuagccacug auua

<210> SEQ ID NO 205
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<212> TYPE: RNA
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<220> FEATURE:
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<400> SEQUENCE: 205
ucagcuccug uuagccacu guuua

<210> SEQ ID NO 206
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 206
caaguuccgu uagccacug

<210> SEQ ID NO 207
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agcuucugu agccacugau u

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cagcuucugu uagccacuga uua

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agcuucugu agccacugau uaa

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<223> OTHER INFORMATION: oligonucleotide
<400> SEQUENCE: 213

cagcuccug uagccacuga uuaa 24

<210> SEQ ID NO 214
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide
<400> SEQUENCE: 214

gcuucugu uagccacgaa uuaa 24

<210> SEQ ID NO 215
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<212> TYPE: RNA
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cagcuccug uagccacgaa uuaa 25

<210> SEQ ID NO 216
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gcuucugu uagccacgaa uuaa 24

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<213> ORGANISM: Artificial
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<400> SEQUENCE: 217

gcuucugu uagccacgau 20

<210> SEQ ID NO 218
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<400> SEQUENCE: 218

gcuucugu uagccacgau 20

<210> SEQ ID NO 219
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<220> FEATURE:
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<220> FEATURE:
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<400> SEQUENCE: 220
gcuucuguu gcuccacugau u

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<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 221
gcuucuguu gcuccacugau a

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<400> SEQUENCE: 222
gcuucugua gcuccacugau a

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<400> SEQUENCE: 223
gcuucuguu gcuccacugau aa

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<212> TYPE: RNA
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<400> SEQUENCE: 224
gcuucugua gcuccacugau aaa

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<212> TYPE: RNA
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<400> SEQUENCE: 225
gcuucuguu gcuccacugau uaaa

<210> SEQ ID NO 226
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<211> LENGTH: 23
<212> TYPE: RNA
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<220> FEATURE:
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<400> SEQUENCE: 226
gcuucuua gcuacuguauu aas

<210> SEQ ID NO 227
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<212> TYPE: RNA
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 227
ccauuuga uuaacaguucccc

<210> SEQ ID NO 228
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 228
agauacauu uguauuaagc

<210> SEQ ID NO 229
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<400> SEQUENCE: 229
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<212> TYPE: RNA
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<400> SEQUENCE: 230
gccauuuuc aacagaucu uca

<210> SEQ ID NO 231
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<212> TYPE: RNA
<213> ORGANISM: Artificial
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<400> SEQUENCE: 231
auucucagga auuugugucu uuc

<210> SEQ ID NO 232
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<212> TYPE: RNA
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<220> FEATURE:
OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 232
ucucaggaau uugugcuuuu c

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<212> TYPE: RNA
<213> ORGANISM: Artificial
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guccagcucc uguuaagcc

<210> SEQ ID NO 234
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cugauuaau ucuuuaauu c

<210> SEQ ID NO 235
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guuuuagca uguuccca

<210> SEQ ID NO 237
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<400> SEQUENCE: 237
caggaauug uguuucuc

<210> SEQ ID NO 238
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<400> SEQUENCE: 238
gcuuuucu uguugugc ucuu

<210> SEQ ID NO 239
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<212> TYPE: RNA
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<220> FEATURE:
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<400> SEQUENCE: 239
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uuuucuuu agugcugc cuuu

<210> SEQ ID NO 240
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<400> SEQUENCE: 240
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uucuuucu uguugcuc uuuc

<210> SEQ ID NO 241
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<400> SEQUENCE: 241
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uucuuucu uguugcuc uuuc

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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 242
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uucuuucu uguugcuc uuucc

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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 243
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uucuuucu uguugcuc uuuccag

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<212> TYPE: RNA
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 244
cuuuuug cguccuuuu ccaag 25

cuuuuug cguccuuuu ccaag
<211> LENGTH: 25
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<210> SEQ ID NO 246
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<212> TYPE: RNA
<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide
<400> SEQUENCE: 246

uuuagugcu gcuuuuuu caggu

<210> SEQ ID NO 247
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<212> TYPE: RNA
<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide
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uuagugcug cuuuuuuuc ggucu

<210> SEQ ID NO 248
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uagugcucu cuuuuuu cag guca

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agugcucu cuuuucag gucua

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<223> OTHER INFORMATION: oligonucleotide
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guagugcuc uuuucag guca

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<220> FEATURE:
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<400> SEQUENCE: 251

ugucugcuu uuccaggucu caagu

<210> SEQ ID NO: 252
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<212> TYPE: RNA
<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 252

ugucugcuu uuccaggucu aagug

<210> SEQ ID NO: 253
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<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 253

gcucucuu uuccaggucu cgagg

<210> SEQ ID NO: 254
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<212> TYPE: RNA
<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 254

cugucucuu cccaggucu gcagg

<210> SEQ ID NO: 255
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 255

ugucucuuu ccagguuca aggga

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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 256

gcucuuuuuc gcguuucag gcggc
cucuuuccg guucaagug ggaua

<210> SEQ ID NO 258
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<212> TYPE: RNA
<213> ORGANISM: Artificial
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SEQUENCE: 308

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SEQ ID NO 309
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TYPE: RNA
ORGANISM: Artificial
FEATURE:

OTHER INFORMATION: oligonucleotide

SEQUENCE: 309

gcuuuguu uuaaaauuu uggca

SEQ ID NO 310
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:

OTHER INFORMATION: oligonucleotide

SEQUENCE: 310

ucuuguuu ucaaaauuug ggcag

SEQ ID NO 311
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:

OTHER INFORMATION: oligonucleotide

SEQUENCE: 311

cuuguuuuc aaaaauuugg ggcag

SEQ ID NO 312
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:

OTHER INFORMATION: oligonucleotide

SEQUENCE: 312

uuguuuuuc aaaaauuugg gcggu

SEQ ID NO 313
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:

OTHER INFORMATION: oligonucleotide

SEQUENCE: 313

uguuuuuuu uuuugggca gcggu

SEQ ID NO 314
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:

OTHER INFORMATION: oligonucleotide

SEQUENCE: 314
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cacgguua uauguucu ccac 25

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<210> SEQ ID NO 337
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<210> SEQ ID NO 338
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<210> SEQ ID NO 339
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<400> SEQUENCE: 347

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<210> SEQ ID NO 348
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<210> SEQ ID NO 350
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<400> SEQUENCE: 350

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<210> SEQ ID NO 351
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<400> SEQUENCE: 351

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<210> SEQ ID NO 352
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<400> SEQUENCE: 352
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 353

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<210> SEQ ID NO 354
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<400> SEQUENCE: 354

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<400> SEQUENCE: 356

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<210> SEQ ID NO 357
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<210> SEQ ID NO 358
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<400> SEQUENCE: 359
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<400> SEQUENCE: 360

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<210> SEQ ID NO 361
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aaauugguca guccaggagc uaggu

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aaauugguca guccaggagc u

21

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<400> SEQUENCE: 365

uaguggcag uccaggacu agguc 25

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<400> SEQUENCE: 366

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guggucaguc caggagcag gugag 25

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<400> SEQUENCE: 368

uggucagucc caggagcuagg ucagg 25

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<400> SEQUENCE: 372

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<400> SEQUENCE: 373

aguccagg gcuaggucag cugcu

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guccagggc uaggucagc ugcuu

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ccaggacua ggucaggcug cuuug

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cagggacuag gcuaggcucu uuugc

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agaggcuagg ucaggcugcu ugc

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gagcuagguc aaggcuguagg cccu

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<400> SEQUENCE: 381
agcuagguc aaggcuguagc cccc

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<220> FEATURE:
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<400> SEQUENCE: 382
gcaggucag gcucuagucgc ccuc

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<400> SEQUENCE: 383
cucagucaag guaagcuauagc guuu

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cagcucuuga aguuaacggu uuacc
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gcucuugaag uuacgguu accgc
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cuagguaggg cuuucgcu cuacg
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uagguaggg guuugcuu uacgc
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<400> SEQUENCE: 388

aggcugggu gcuugcccu cuagcu
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<212> TYPE: RNA
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ggcuagcugu cuuucgcccc uacgc
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gucgcucgc uggccccca gcucu

<210> SEQ ID NO 391
<211> LENGTH: 25
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<213> ORGANISM: Artificial
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<400> SEQUENCE: 391

ucggcgcuc uggccucag cucuu

<210> SEQ ID NO 392
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 392

cagcgcucu ugccucaca gcucu

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<400> SEQUENCE: 393

agcgcucuuc gccccacgcu cuuga

<210> SEQ ID NO 394
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<212> TYPE: RNA
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<400> SEQUENCE: 394

ggcgcucuuc cccacacug ucgaa

<210> SEQ ID NO 395
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<212> TYPE: RNA
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<400> SEQUENCE: 395

gcugcucuc ccucagcucu ugaag

<210> SEQ ID NO 396
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<400> SEQUENCE: 396

cgcucuucgc cucagcucu gaag

<210> SEQ ID NO 397
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<212> TYPE: RNA
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<400> SEQUENCE: 397

ugcuuugccc ucacucuug aagua

25

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<400> SEQUENCE: 398

gcuuugccu cagcuuuga aguua

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<210> SEQ ID NO 399
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<212> TYPE: RNA
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<220> FEATURE:
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<400> SEQUENCE: 399

cuugccucc agcucuugaa guuua

25

<210> SEQ ID NO 400
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<400> SEQUENCE: 400

uugcuccua gcucuugaag uaaac

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<210> SEQ ID NO 401
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<212> TYPE: RNA
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<400> SEQUENCE: 401

uugccuccacg cucuugaagu aaacg

25

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<212> TYPE: RNA
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<220> FEATURE:
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<400> SEQUENCE: 402

ugccucacgc ucuugaagaa aacgg

25

<210> SEQ ID NO 403
<211> LENGTH: 25
<212> TYPE: RNA
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<400> SEQUENCE: 403

gccucagcu cuugaagaua acggu

<210> SEQ ID NO: 404
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
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<400> SEQUENCE: 404

ccucagcuc uugaagauaa cgguu

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<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 405

ccucagcu uugaagaas

<210> SEQ ID NO: 406
<211> LENGTH: 21
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 406

ccucagcuc uugaagaac g

<210> SEQ ID NO: 407
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 407

ccucagcucu gaugaacgc

<210> SEQ ID NO: 408
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 408

ccucagcucu ugaagaacs cgguu

<210> SEQ ID NO: 409
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 409
ucaacuucc uauuaacgg uuuac

<210> SEQ ID NO 410
<211> LENGTH: 25
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<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 410

agcuuuga guaasaggu uaceg

<210> SEQ ID NO 411
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 411

cucaugau aacgguuua ccgcc

<210> SEQ ID NO 412
<211> LENGTH: 26
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 412

ccacagcu ugcacuucuc aaugc

<210> SEQ ID NO 413
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 413

ccagggcu gucucuuugc augcu

<210> SEQ ID NO 414
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 414

acagggcu gucucuugaa ugcug

<210> SEQ ID NO 415
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 415

cagggcuuc gucucuuaau gcugc

<210> SEQ ID NO 416
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 416

aggcguguca cuuugcaaug cugcu 25

<210> SEQ ID NO: 417
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 417
gccguguca cuuugcaaug ucgug 25

<210> SEQ ID NO: 418
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 418
gccguguca cuuugcaaug gcugu 25

<210> SEQ ID NO: 419
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 419
cguugacauc ugcagaugcu cuguc 25

<210> SEQ ID NO: 420
<211> LENGTH: 23
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 420
cguugacauc ugcagaugcug cug 23

<210> SEQ ID NO: 421
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 421
guugacacuu guaugcguac ugucu 25

<210> SEQ ID NO: 422
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 422

uugccuuug caaugcugc gucuu

<210> SEQ ID NO 423
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 423

uugccuuugc aaugcugc gucuuc

<210> SEQ ID NO 424
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 424

gccuuugga auggcugc gucucu

<210> SEQ ID NO 425
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 425

cacuuuggca ucugcugc gucucu

<210> SEQ ID NO 426
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 426

cacuuuggca ucugcugc gucucu

<210> SEQ ID NO 427
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 427

cucuuggcau cuugcugc gucugc

<210> SEQ ID NO 428
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 428
uugcaaugc ugcugucuuc uugcu

<210> SEQ ID NO 429
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 429
gugcaaugc gcugucuuc ugcua

<210> SEQ ID NO 430
<211> LENGTH: 26
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 430
ugcaaugug cuugucuucu gcuau

<210> SEQ ID NO 431
<211> LENGTH: 26
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 431
gcaaugcug ugcuucuucg cuuag

<210> SEQ ID NO 432
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 432
cauaugcu guuucuucu guaug

<210> SEQ ID NO 433
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 433
aaugcuguc ucuucucug uuga

<210> SEQ ID NO 434
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 434
augucuguc guucucugua uggau

<210> SEQ ID NO 435
ugcuugucu ucuugcuau gaaua 25

<210> SEQ ID NO: 436
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<212> TYPE: RNA
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<400>SEQUENCE: 436

gcuucuguuc ucuugcuau gaaua 25

<210> SEQ ID NO: 437
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<222> OTHER INFORMATION: oligonucleotide

<400>SEQUENCE: 437

cucuguucuc ucucugcuau aaua 25

<210> SEQ ID NO: 438
<211> LENGTH: 25
<212> TYPE: RNA
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<220> FEATURE:
<222> OTHER INFORMATION: oligonucleotide

<400>SEQUENCE: 438

ugcuucuguuc ucuugcuau gaugu 25

<210> SEQ ID NO: 439
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<212> TYPE: RNA
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<220> FEATURE:
<222> OTHER INFORMATION: oligonucleotide

<400>SEQUENCE: 439

gcuucuguuc ucuugcuau auggu 25

<210> SEQ ID NO: 440
<211> LENGTH: 25
<212> TYPE: RNA
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<220> FEATURE:
<222> OTHER INFORMATION: oligonucleotide

<400>SEQUENCE: 440

cucuguucugu cuucugcuau auguc 25

<210> SEQ ID NO: 441
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<212> TYPE: RNA
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<220> FEATURE:
ugcuuucug cuaucaauaa uguca

<210> SEQ ID NO 442
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 442

ugcuuucugucaaucaauaa uguca

<210> SEQ ID NO 443
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 443

ucuucucugucaaucaauaa ucaau

<210> SEQ ID NO 444
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 444

cuucucucugucaaucaauaa ucaau

<210> SEQ ID NO 445
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 445

ucucucucucugucaaucaauaa ucaau

<210> SEQ ID NO 446
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 446

ucucucucucugucaaucaauaa ucaau

<210> SEQ ID NO 447
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 447

ucucucucucugucaaucaauaa ucaau
cuucauuga uuaugguca uccga

<210> SEQ ID NO 448
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 448

uugcuauaa uuaugcuau ccgac

<210> SEQ ID NO 449
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 449

ugcuauaau uaugcuau ccacc

<210> SEQ ID NO 450
<211> LENGTH: 26
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 450

gcuauaauu augcuauucc gaccu

<210> SEQ ID NO 451
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 451

cuaauguaau ugucauucc accug

<210> SEQ ID NO 452
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 452

uaugaaauu gucaauccgac ccuga

<210> SEQ ID NO 453
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
<223> OTHER INFORMATION: oligonucleotide

<400> SEQUENCE: 453

augaaauaug ucaauccgac cugag

<210> SEQ ID NO 454
ugaaauaugu caaucgacc uagcu

<210> SEQ ID NO 455
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 455
gaaauauguc aaucgaccu gagcu

<210> SEQ ID NO 456
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 456
aaauauguc aaucgaccu agcuu

<210> SEQ ID NO 457
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<212> TYPE: RNA
<213> ORGANISM: Artificial
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<400> SEQUENCE: 457
aaauauguc aaaucgaccu gcuu

<210> SEQ ID NO 458
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<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 459
uaauaugcau cagaccuug cuuug

<210> SEQ ID NO 459
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
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<400> SEQUENCE: 459
aaauaugcauc cgaccuugac uuugu

<210> SEQ ID NO 460
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 460

augucaucc gaccugacu uguu

SEQ ID NO 461
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 461
ugucauccg accugacu uguug

SEQ ID NO 462
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 462
gucauccgac ccugacuu uguu

SEQ ID NO 463
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 463
cuauccgac ccuacuu uguu

SEQ ID NO 464
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 464
cuauccgacc ugcuccu uguu

SEQ ID NO 465
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 465
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FEATURE:
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FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 518

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FEATURE:
OTHER INFORMATION: oligonucleotide

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ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 520

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FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 521

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ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 522

casuucuucc cacacagau u

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FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 537
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TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 538
cuuuaaugu cgugcgcgu

SEQ ID NO 539
LENGTH: 20
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

SEQUENCE: 539
ucaggaaga uggcaaunu

SEQ ID NO 540
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TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide

FEATURE:
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LOCATION: (5)...(5)
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FEATURE:
NAME/KEY: misc_feature
LOCATION: (5)...(5)
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 540
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LENGTH: 20
TYPE: RNA
ORGANISM: Artificial
FEATURE:
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FEATURE:
NAME/KEY: modified_base
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ucaggaaga uggcauuucu

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<u>ORGANISM: Artificial</u>
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uuuggcgcn cccaaugccga uccug

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OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 560

uwuccguc ucuaugccua ucucug

25

SEQ ID: 561
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
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OTHER INFORMATION: oligonucleotide
FEATURE:
NAME/KEY: modified_base
LOCATION: (25) ... (25)
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 561

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SEQ ID: 562
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide
FEATURE:
NAME/KEY: modified_base
LOCATION: (1) ... (1)
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 562

uwuccguc ucuaugccua ucucug

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SEQ ID: 563
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide
FEATURE:
NAME/KEY: modified_base
LOCATION: (2) ... (2)
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 563

uwuccguc ucuaugccua ucucug

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SEQ ID: 564
LENGTH: 25
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ORGANISM: Artificial
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OTHER INFORMATION: oligonucleotide
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<221> NAME/KEY: misc_feature
<222> LOCATION: (20) ... (20)
<223> OTHER INFORMATION: n is a, c, g, or u
<400> SEQUENCE: 567

uuugccgucgccauugcuaugg

<210> SEQ ID NO: 568
<211> LENGTH: 25
<212> TYPE: RNA
<213> ORGANISM: Artificial
<220> FEATURE:
OTHER INFORMATION: oligonucleotide
FEATURE:
- NAME/KEY: modified_base
- LOCATION: (4) (4)
- OTHER INFORMATION: n is a, c, g, or u

FEATURE:
- NAME/KEY: misc_feature
- LOCATION: (4) (4)
- OTHER INFORMATION: n is a, c, g, or u

SEQ ID NO: 568
uuuucenguc cccagugca ccuug

SEQ ID NO: 569
uuuguccguc cccagugca ccuug

SEQ ID NO: 570
uuugucgcgcuc cccagugca ccuug

SEQ ID NO: 571
uuugucgcgcgcuc cccagugca ccuug

SEQ ID NO: 572
uuugucgcgcgcgcuc cccagugca ccuug
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 571

uuugcgcug ccaauggca ucncng 25

SEQ ID NO 572
LENGTH: 25
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide
FEATURE:
NAME/KEY: modified_base
LOCATION: (14) (14)
OTHER INFORMATION: j
FEATURE:
NAME/KEY: misc_feature
LOCATION: (14) (14)
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 572

uuugcgcug ccaauggca ucncug 25

SEQ ID NO 573
LENGTH: 20
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide
FEATURE:
NAME/KEY: modified_base
LOCATION: (10) (10)
OTHER INFORMATION: j
FEATURE:
NAME/KEY: misc_feature
LOCATION: (10) (10)
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 573

ucagouucun uuaacccacug 20

SEQ ID NO 574
LENGTH: 20
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide
FEATURE:
NAME/KEY: modified_base
LOCATION: (14) (14)
OTHER INFORMATION: j
FEATURE:
NAME/KEY: misc_feature
LOCATION: (14) (14)
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 574

ucagouucug uuancccacug 20

SEQ ID NO 575
LENGTH: 20
TYPE: RNA
ORGANISM: Artificial
FEATURE:
OTHER INFORMATION: oligonucleotide
FEATURE:
NAME/KEY: modified_base
LOCATION: (4) (4)
OTHER INFORMATION: n is a, c, g, or u

SEQUENCE: 575
ucancuucug uaagccacucg

ucagcuucug uaagccacun

Met Gly Ile Ser Ser Leu Pro Thr Gln Leu Phe Lys Cys Cys Phe
1      5     10    15

Cys Asp Phe Leu Lys Val Lys Met His Thr Met Ser Ser Ser His Leu
20    25

Phe Tyr Leu Ala Leu Cys Leu Leu Thr Phe Thr Ser Ser Ala Thr Ala
35    40    45

Gly Pro Glu Thr Leu Cys Gly Ala Glu Leu Val Asp Ala Leu Gln Phe
50    55    60

Val Cys Gly Asp Arg Gly Phe Tyr Phe Asn Lys Pro Thr Gly Tyr Gly
65    70    75    80

Ser Ser Ser Arg Ala Pro Gln Thr Gly Ile Val Asp Glu Cys Cys
85    90    95

Phe Arg Ser Cys Asp Leu Arg Arg Leu Glu Met Tyr Cys Ala Pro Leu
100   105   110

Lys Pro Ala Lys Ser Ala Arg Ser Val Arg Ala Gln Arg His Thr Asp
115   120   125

Met Pro Lys Thr Gln Lys Lys Lys Gln His Leu Lys Asn Ala Ser Arg Gly
130   135   140

Ser Ala Gly Asn Lys Asn Tyr Arg Met
145   150
1. An isolated oligonucleotide comprising a sequence which is complementary to at least part of a dystrophin pre-mRNA exon or at least part of a non-exon region of a dystrophin pre-mRNA said part being a contiguous stretch comprising at least 8 nucleotides, wherein said oligonucleotide comprises one or both of an inosine nucleotide and a nucleotide containing a base able to form a wobble base pair with a complementary base to which it is paired.

2. An isolated oligonucleotide according to claim 1, wherein the contiguous stretch comprises between 13 and 50 nucleotides, of RNA of an exon of a dystrophin pre-mRNA.

3. An isolated oligonucleotide according to claim 2, wherein said exon comprises exon 51, 45, 53, 44, 46, 52, 50, 43, 6, 7, 8, 55, 2, 11, 17, 19, 21, 57, 59, 62, 63, 65, 66, 69, and/or 75.

4. (canceled)

5. An isolated oligonucleotide according to claim 1, wherein the oligonucleotide comprises a first part and a second part, wherein said first part comprises at least 8 consecutive nucleotides that are complementary to a first exon and wherein second part comprises at least 8 consecutive nucleotides that are complementary to a second exon in said dystrophin pre-mRNA.

6. An isolated oligonucleotide according to claim 5, wherein said first and said second exon are contiguous in said dystrophin pre-mRNA.

7. An oligonucleotide according to claim 5, wherein said first and said second exon are contiguous in said dystrophin pre-mRNA.

8. (canceled)

9. (canceled)

10. (canceled)

11. A composition comprising at least two distinct oligonucleotides as defined in claim 1.

12. A composition according to claim 11, wherein each said distinct oligonucleotide is dosed, independently, in an amount between 0.5 mg/kg and 10 mg/kg, inclusive.

13. A composition according to claim 11 in combination with one or more of:
   (a) an adjunct compound for reducing inflammation, preferably for reducing muscle tissue inflammation,  
   (b) an adjunct compound for improving muscle function, integrity and/or survival, and  
    (c) a compound exhibiting readthrough activity.

14. (canceled)

15. A method for alleviating one or more symptom(s) of Duchenne Muscular Dystrophy or Becker Muscular Dystrophy in an individual, the method comprising administering to said individual a composition as defined in claim 11.

16. An isolated oligonucleotide according to claim 2 wherein said contiguous stretch comprises between 14 and 25 nucleotides of RNA of an exon of a dystrophin pre-mRNA.

17. An isolated oligonucleotide according to claim 1, wherein the oligonucleotide comprises RNA.

18. An isolated oligonucleotide according to claim 17, wherein said RNA comprises a modified ribonucleotide.

19. An isolated oligonucleotide according to claim 18, wherein said modified ribonucleotide is a 2′-O-methyl modified ribose (RNA).

20. An isolated oligonucleotide according to claim 1, comprising a modified deoxyribose (DNA) base.

21. An isolated oligonucleotide according to claim 1, wherein said oligonucleotide comprises a peptide nucleic acid, a locked nucleic acid, a morpholino phosphorodiamidate, or a combination thereof.

22. An isolated oligonucleotide according to claim 21, comprising a morpholino phosphorodiamidate.

23. An isolated oligonucleotide according to claim 5, wherein said first and said second exon are contiguous in said dystrophin pre-mRNA.

24. The composition of claim 11, admixed with a pharmaceutically acceptable carrier, adjuvant, diluent and/or excipient.

25. The composition of claim 13, wherein said adjunct composition for reducing inflammation reduces tissue inflammation.

26. The method of claim 15, wherein the composition administered provides the individual with a functional dystrophin protein.

27. The method of claim 15, wherein the composition administered decreases the production of an aberrant dystrophin protein.

28. The method of claim 15, wherein the composition administered increases the production of a functional or a more functional dystrophin protein.

29. The method of claim 15, wherein the composition administered alleviates one or more symptom(s).

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