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(56) Related Art
Bachmann et al., Arch. Virol., 1975, vol. 48, no. 2, pp. 107-120
Randall et al., J. Gen. Virol., 1987, vol. 68, no. 11, pp. 2769-2780
Lin & Lamb, J. Virol., 2000, vol. 74, no. 19, pp. 9152-9166
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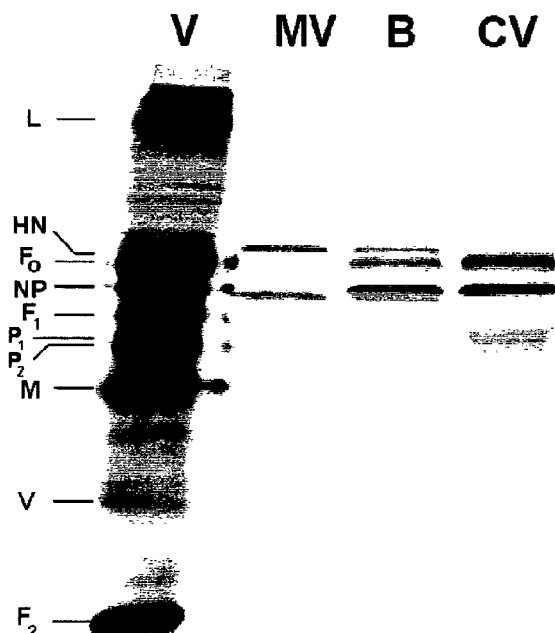
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(54) Title: A NOVEL VIRUS (CRYPTOVIRUS) WITHIN THE RUBULAVIRUS GENUS AND USES THEREFOR



(57) Abstract: The present invention is based on the discovery of a novel human virus that has been designated as a *Cryptovirus*, which falls within the *Rubulavirus* genus of the Paramyxoviridae. Disclosed are isolated *Cryptovirus*-specific nucleic acids, nucleic acid constructs containing the *Cryptovirus*-specific nucleic acids, *Cryptovirus*-specific proteins, isolated *Cryptovirus* particles, isolated anti-*Cryptovirus* antibodies, and compositions containing them. Uses for manufacturing a vaccine or a medicament are also disclosed. Also disclosed are inventive vectors containing the *Cryptovirus*-specific nucleic acids and host cells containing the vectors. Methods of isolating and propagating *Cryptovirus* are disclosed, as are methods of producing a mammalian cell line acutely or non productively infected with *Cryptovirus*; such cells are disclosed. Disclosed also are methods of detecting *Cryptovirus* particles, proteins or anti-*Cryptovirus* antibodies in a sample of a biological material. *In vitro* and *in vivo* screening methods for identifying potential antiviral therapeutic or prophylactic agents are disclosed. Methods of detecting a *Cryptovirus* infection in a mammal are disclosed. Methods of detecting *Cryptovirus* infection in a mammal are disclosed. Also disclosed is an animal model for human diseases, such as neurological, neurodegenerative,

and/or neuropsychiatric diseases, including epileptiform diseases (e.g., epilepsy, multiple sclerosis, chronic fatigue syndrome, and subacute sclerosing panencephalitis).

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NEUROVIRULENT VIRUS (*CRYPTOVIRUS*) WITHIN THE *RUBULAVIRUS* GENUS AND USES THEREFOR

Throughout the application various publications are referenced in parentheses. The disclosures of these publications in their entireties are hereby incorporated by reference in the application in order to more fully describe the state of the art to which this invention pertains.

BACKGROUND OF THE INVENTION

1. Field of the Invention

This invention relates to the medical arts and particularly to the field of virology, and, more particularly, to a novel human *Rubulavirus* which has been designated as a "*Cryptovirus*".

2. Discussion of the Related Art

The *Rubulavirus* genus of the Paramyxoviridae (see Fig. 1) are enveloped viruses characterized by a single minus-stranded RNA genome. There is substantial evidence that some rubulaviruses infect domestic animals and can cause neurological diseases in them. These neuropathic rubulaviruses include Canine Parainfluenza Virus Type 2, Porcine Rubulavirus (*aka* La Piedad-Michoacan-Mexico Virus), and Menangle Virus. The strong sequence homologies amongst these viruses imply that each of these viruses represent separate host-adapted species of a common ancestral Rubulavirus species.

Canine Parainfluenza Virus Type 2 (which is also known as CPiV) is associated with infectious tracheobronchitis in dogs. This is a non-lethal disease (usually) of the respiratory tract (Appel and Binn, Canine Infectious Tracheobronchitis. *In* Virus Infections of Carnivores. Elsevier Science Publishing Co., New York, N.Y., pp 201-211, 1987). However, the virus has also been found to be associated with posterior paresis (Evermann *et al.* *J.A.V.M.A.* 177:1132-1134, 1980), neurological dysfunction (Baumgartner *et al.* *Infect. Immun.* 31:1177-1183, 1981), encephalitis (Evermann *et al.* *Arch. Virol.* 68:165-172, 1981) and hydrocephalus (Baumgartner *et al.* *Vet. Pathol.* 19:79-92, 1982) in dogs.

Porcine Rubulavirus (which is also known as La Piedad-Michoacan-Mexico Virus or LPMV) is associated with Blue-Eye Disease (BED) of pigs. The symptoms of the disease include corneal opacity, extreme "nervousness" in young pigs and infertility in sows and boars (Ramirez-Mendoza *et al.*, *J. Comp. Pathol.* 117:237-252, 1997). The virus persists in the central nervous system of pigs after recovery from BED (Wiman *et al.* 1998; Hjertner *et al.* 1998). This virus also shares more than 95% of its nucleotide sequence with *Cryptovirus*.

Menangle virus is an emergent rubulavirus that was first identified in Australia in 1997 (Philbey *et al.*, *Emerging Infectious Diseases* 4:269-271, 1998). This virus is associated with severe degeneration of the brain and spinal cord of stillborn piglets (Philbey *et al.* 1998). Neurons of these animals contained Menangle virus inclusion bodies (nucleocapsids). Serological studies have also found neutralizing antibodies to the virus in pigs, fruit bats and at least two human piggery workers.

A number of human rubulaviruses are known to cause illness in humans. These include: (1) mumps virus (causes human mumps); (2) human parainfluenza virus type 2 (*aka* HPIV-2; associated with relatively mild upper respiratory, flu-like illness; (3) human parainfluenza virus types 4A and 4B (*aka* HPIV-4A, HPIV-4B; also associated with relatively mild upper respiratory, flu-like illnesses). With mumps there is also evidence of nervous system involvement in a significant number of patients, although this is virtually never life threatening.

In contrast, there are no published studies clearly demonstrating that another *Rubulavirus*, Simian Virus 5 (SV5), causes disease either in humans or in experimentally-infected animals, and there has been at least one published study demonstrating that SV5 does not cause disease in experimentally-infected mice, even mice with severe combined immunodeficiency (SCID mice) (Didcock *et al.*, *J. Virol.* 73:3125-3133, 1999).

In 1978, a report described the isolation of an "infectious agent" from the bone marrow of patients with multiple sclerosis (MS). (Mitchell, DN *et al.*, *Isolation of an infectious agent from bone marrow of patients with multiple sclerosis*, *Lancet* ii:387-391 [1978]). Subsequent reports described five separate "human SV5 isolates" derived from different MS patients. (Goswami, KKA *et al.*, *Does simian virus 5 infect humans?* *Journal of General Virology* 65:1295-1303 [1984]; Goswami, KKA *et al.*, *Evidence for the persistence of paramyxoviruses in human bone marrows*, *Journal of General Virology* 65:1881-1888 [1985]; Randall, RE *et al.*, *Isolation and characterization of monoclonal antibodies to simian virus 5 and their use in revealing antigenic differences between human, canine and simian isolates*, *Journal of General Virology* 68:2769-2780 [1987]). Nevertheless, a causal link between SV5 and MS remained speculative.

In 1987, Goswami *et al.* reported that the cerebrospinal fluid (CSF) of some MS patients contained antibodies to SV5. (Goswami KKA *et al.*, *Antibodies against the paramyxovirus SV5 in the cerebrospinal fluids of some multiple sclerosis patients*, *Nature* 327:244-247 [1987]). However, this report was controversial, since the results subsequently failed to be reproducible by other well respected paramyxovirologists. (Vandvik, B. and Norrby, E., *Paramyxovirus SV5 and multiple sclerosis*, *Nature* 338:769-771 [1989]; but see, Russell, WC and Randall, RE, *Multiple sclerosis and*

paramyxovirus, Nature 340:104 [1989]). Therefore, a clear causal link between SV5 and MS was not established in the art.

Multiple Sclerosis is a chronic degenerative central nervous system disease that most commonly affects young and early middle-aged adults (between 18 and 40 years of age). It is less commonly diagnosed in adolescents and even less so in children. Affecting 350,000 Americans, MS is one of the most frequent causes of neurologic disability except for traumatic injuries. (S.L. Hauser, *Multiple Sclerosis and other demyelinating diseases* In: *Harrison's Principles of Internal Medicine*, 13th ed., K.J. Isselbacher *et al.* (eds.), McGraw-Hill, pp.2287-95 [1994]). The onset, progression and outcome of the disease are highly variable with patients manifesting one of several patterns of illness. For example, for reasons that are unclear, MS affects twice as many females as males. Although the individual components that comprise the diagnostic, clinical tableau of MS have long been delineated, their sequence and severity of presentation from case to case are subject to great variation. (Hallpike J.F., Adams C.W.M., and Tourtellotte W.W., Eds, 1983, *Multiple Sclerosis: Pathology, Diagnosis and Management*, Chapman & Hall, London; McAlpine E. *et al.*, 1972, *Multiple Sclerosis: A Reappraisal*, Churchill Livingstone, Edinburgh; Rose A.S., 1972, *Multiple Sclerosis: A Clinical Interpretation*. In *Multiple Sclerosis: Immunology, Virology and Ultrastructure* Wolfgram F., Ellison G.W., Stevens J.G., and Andrews J.M., Eds., Academic Press, New York). It is fair to say that no two patients with MS are alike, and, consequently, there is contention as to what constitutes the stereotypic clinical history.

Most commonly, MS first presents as a series of neurological attacks followed by complete or partial remissions where symptoms lessen only to return after some period of stability (relapsing-remitting MS). In other patients, the disease is characterized by a gradual decline with no clear remissions but sometimes with brief plateaus or minor relief of symptoms (primary-progressive MS). In still other patients, there can be a relapsing and remitting course of illness in the early stages followed by progressive decline (secondary-progressive MS).

In general, the primary manifestations of chronic progressive and chronic relapsing MS do not vary greatly. Evidence for an insidious disease (apathy, depression, fatigue, loss of weight, muscle pains) often can be uncovered from the patient's chart before the first neurological manifestations. Among the first signs in about 50% of all definite MS cases are limb weakness, numbness, or tingling (parathesias) in one or more limbs, the extremities, or around the trunk. There is often discordance between signs and symptoms. Adams and Victor (1997, *Multiple sclerosis and allied demyelinating diseases*. In *Principles of Neurology*. Adams R.D. and Victor M., Eds., McGraw Hill, New York) mention that "it is a common aphorism that the patient with MS presents

with symptoms in one leg and signs (bilateral Babinski) in both.” Another common initial sign is a short-lived episode of retrobulbar neuritis affecting one or both eyes. Many MS patients will display papillitis (swelling of the optic nerve head), which depends on the proximity of the demyelinated plaque to the nerve head. There is considerable debate as to whether optic neuritis in a significant percentage of cases constitutes a separate disease or subclass of MS, but in about 50% of cases, the disease progresses to MS (Arnason *et al.*, *J. Neurol. Sci.* 22:419, 1974).

As diagnosis becomes established, a more regular group of clinical syndromes develops either progressively or in a remitting fashion. The majority of patients display a mixed or generalized type of disease involving optic nerves, brain stem, cerebellum, and spinal cord. About one third will exhibit a spinal form, and about 5% will display a cerebellar or pontobulbar-cerebellar form, and a similar percentage will have an amaurotic form. Adams and Victor (*supra*) estimate that at least 80% of their own clinical material comprised cerebrospinal and spinal forms of the disease.

Psychologic disturbances are frequently observed and can present as an inappropriate euphoric state, attributed by Adams and Victor (*supra*) probably to extensive white matter lesions in the frontal lobes. In a much higher percentage of MS patients depression and irritability are observed.

Until relatively recently, MS and epilepsy were considered discrete entities. The publication of numerous recent studies demonstrating an “overlap” between these disorders has corrected this misperception. Epidemiological and demographic studies conducted over the last decade have provided substantial evidence of concurrent epileptiform symptomology in a significant proportion of MS patients. While the concurrence of epileptiform symptoms is markedly higher in early onset MS (i.e. in children and adolescents), the overall prevalence of epilepsy in MS patients is many times higher than in the general population.

Human epilepsy is an enigmatic medical condition which, in fact, is not a specific disease – or even a single syndrome – but, rather a broad category of symptom complexes arising from any number of disordered brain functions that themselves can be secondary to a variety of pathologic processes. Today, a large number of clinical phenomena are recognized as epileptic seizures, some of which (*e.g.*, myoclonic and atonic seizures) are currently poorly understood and could, in fact, reflect neuronal mechanisms that are somewhat different from the pathophysiologic processes traditionally considered to be “epileptic.” Perhaps the best reflection of the enigmatic and complex nature of these illnesses is the simple fact that the etiology of the disease, in the overwhelming majority of the cases (greater than 70%), is either “cryptogenic” (*i.e.*, of obscure, indeterminate origin) or “idiopathic” (*i.e.*, of unknown cause).

Epilepsy is more than seizures. Epileptics typically exhibit a spectrum of responses, from little or no seizure activity, through mild activity (*petit mal* or “absence” seizures), to recurrent and intractable *grand mal* seizures (the occurrence of which is often misunderstood by the lay public to be the defining symptom of all forms of epilepsy; see *Epilepsy: A Comprehensive Textbook*, Engel, Jr. J. and Pedley, T.A., Eds., Lippincott-Raven, 1997). The condition in its entirety is comprised of many facets, different for each individual, that contribute to disability and impaired quality of life. While the physical spectrum of symptoms ranges from extremely subtle *petit mal* or “absence” seizures to profoundly disabling *grand mal* seizures, many patients experience other co-morbid processes (e.g., memory loss, confusion, lethargy, sleep disturbances, and clinical depression) which can be equally disabling. Treatment that focuses solely on seizures often does little to lessen disability. This is perhaps best illustrated by the patient who, having undergone successful surgical resection of epileptogenic brain tissue, becomes seizure-free but remains socially isolated and unemployed, with little evidence of an improved life. Therapeutic intervention can be optimal only when the multiple and interacting medical, psychological, and environmental factors that constitute epilepsy are addressed.

Another epileptiform disease is Subacute Sclerosing Panencephalitis (SSPE), a rare and fatal degenerative central nervous system disease of children and adolescents. (Sever and Zeman, editors, Measles Virus and Subacute Sclerosing Panencephalitis. *Neurology (Supplement 1)* 18:1-192, 1968; Payne and Baublis, *Perspectives in Virology* 7:179-195, 1971; Johannes and Sever, *Ann. Rev. Med.* 26:589-601, 1975; Meulen *et al.*, *Comp. Virol.* 18:105-159, 1983; Dyken, *Neurol. Clin.* 3:179-196, 1985). In its early stages it commonly presents as an affective or other behavioral disorder and progresses over a period of months to profound epileptiform neurological disease. Its later stages are characterized by intractable seizures, decerebrate rigidity, coma and death. At some time, virtually all SSPE patients are “misdiagnosed” with epilepsy.

Cases of SSPE have been described in both industrialized and developing countries throughout the world (Canal and Torck, *J. Neurol. Sci.* 1:380-389, 1964; Pettay *et al.*, *J. Infect. Dis.* 124:439-444, 1971; Haddad *et al.*, *Lancet* 2:1025, 1974; Soffer *et al.*, *Israeli J. Med. Sci.* 11:1-4, 1975; Naruszewicz-Lesiuk *et al.*, *Przeg Epidemiologiczna* 23:1-8, 1979; Moodie *et al.*, *South African Med. J.* 58:964-967, 1980). The frequency of the disease varies greatly, ranging from 0.06 and 0.10 cases per million total population per year (cpmpy) in Britain (Dick, *Brit. Med. J.* 3:359-360, 1973) and the United States (Jabbour *et al.*, *J. A. M. A.* 220:959-962, 1972) to 3.40 and 7.70 cpmpy in Israel (Soffer, *et al.*, *Israeli J. Med. Sci.* 11:1-4, 1975) and New Zealand (Baguley and Glasgow, *Lancet*

2:763-765, 1973). The factor(s) that are responsible for the ultimate etiopathogenesis of the disease are unclear.

Numerous studies have shown that the central nervous system tissues of SSPE patients are persistently-infected with measles virus. Substantial evidence indicates that the disease involves the
5 recrudescence of a persistent measles virus infection acquired earlier in life. Specific findings in SSPE patients which support this hypothesis include: (1) a history of childhood measles, (2) markedly elevated titers of measles virus-specific antibodies in serum, (3) the presence of measles virus-specific antibodies in cerebrospinal fluid, (4) the presence of measles virus antigens in CNS
10 tissues demonstrated by specific immunofluorescence, (5) intracellular inclusions of paramyxoviral nucleocapsids in oligodendroglial and neuronal cells, and (6) the isolation of infectious measles virus from brain and lymphatic tissues when co-cultivated with susceptible cells (Bouteille *et al.*, *Revue Neurologie* 113:454-458, 1965; Connolly *et al.*, *Lancet* 1:542-544, 1967; Legg, *Brit. Med. J.* 3:350-354, 1967; Payne *et al.*, *New Eng. J. Med.* 281:585-589, 1969; Horta-Barbosa *et al.*, *Nature* 221:974, 1969). Finally, and perhaps most convincingly, epidemiological evidence suggests that vaccination
15 against measles substantially reduces the risk of developing the disease (Modlin *et al.*, *Pediatrics* 59:505-512, 1977; Halsey *et al.*, *Am. J. Epidemiol.* 111:415-424, 1980; Dyken *et al.*, *Morb. Mortal. Weekly Report* 31:585-588, 1982).

Despite these findings, there are a number of anomalies that have been observed which are inconsistent with measles virus alone being the sole cause of the illness. These include the following.

20 First, neurovirulence. Clinical isolates of measles virus from patients with rubeola have not been shown to cause an SSPE-like illness in experimentally infected animals. Cell-associated SSPE-derived strains, however, have been shown to cause such disease in ferrets, marmosets and monkeys (Katz *et al.*, *J. Infect. Dis.* 121:188-195, 1970; Thormar *et al.*, *J. Infect. Dis.* 127:678-685, 1973; Ueda
et al., *Biken Journal* 18:179-181, 1975; Yamanouchi *et al.*, *Japan. J. Med. Sci. Biol.* 29:177-186, 1976; Thormar *et al.*, *J. Infect. Dis.* 136:229-238, 1977; Albrecht *et al.*, *Science* 195:64-66, 1977;
25 Thormar *et al.*, *J. Exp. Med.* 148:674-691, 1978; Ohuchi *et al.*, *Microbiol. Immunol.* 25:887-893 [1981]).

Second, distribution and morphology of virus inclusion bodies. Measles virus antigens in infected cells form large coalescing intracytoplasmic inclusions when examined by fluorescent
30 antibody techniques. When labeled with SSPE patient sera, virus antigens demonstrate distinctly different patterns in cell-associated SSPE-derived virus strains and in experimentally-infected animal CNS tissues. In such materials, intracellular inclusion bodies demonstrate a "peppery," particulate and/or "splattered" distribution (Doi *et al.*, *Japan. J. Med. Sci. Biol.* 25:321-333, 1972; Kimoto and

Baba, *Biken Journal* 18:123-133, 1975; de Felici *et al.*, *Annales Microbiologie* 126:523-538, 1975; Ohuchi *et al.*, *Microbiol. Immunol.* 23:877-888, 1979).

Third, ultrastructural morphology of virus nucleocapsids. In measles virus infected cells, cytoplasmic nucleocapsids are predominantly of a "fuzzy" or "granular" morphology (Tawara, *Virus* (Osaka) 14:85-88, 1965; Matsumoto, *Bull. Yamaguchi Med. School* 13: 167-189, 1966; Nakai *et al.*, *Virology* 38:50-67, 1969; Nakai and Imagawa, *J. Virol.* 3:187-197, 1969). In SSPE-derived CNS tissues, both fuzzy and smooth nucleocapsids have been consistently observed (Oyanagi *et al.*, *J. Virol.* 7:176-182, 1971; Dubois-Dalcq *et al.*, *Arch. Neurol.* 31:355-364, 1974). Smooth cytoplasmic nucleocapsids have also been observed in most cell-associated SSPE-derived cell lines (Doi *et al.*, *Japan. J. Med. Sci. Biol.* 25:321-333, 1972; Makino *et al.*, *Microbiol. Immunol.* 21:193-205, 1977; Ueda *et al.*, *Biken Journal* 18:113-122, 1975; Burnstein *et al.*, *Infect. Immun.* 10:1378-1382, 1974; Mirchamsy *et al.*, *Intervirology* 9:106-118, 1978; Schott *et al.*, *Revue Neurologie* 135:653-664, 1979) with some containing only such structures (Doi *et al.*, *Japan. J. Med. Sci. Biol.* 25:321-333, 1972; Thormar *et al.*, *J. Exp. Med.* 148:674-691, 1978).

Fourth, immunoreactivity of virus nucleocapsids. While fuzzy nucleocapsid aggregates are labeled with HRP-conjugated measles virus-specific antibody experimentally raised in animals, smooth virus nucleocapsids are not (Dubois-Dalcq *et al.*, *Lab. Invest.* 30:241-250, 1974; Brown *et al.*, *Acta Neuropathologica* 50:181-186, 1980). Most interestingly, both can readily be labeled with SSPE sera (Brown *et al.*, *Acta Neuropathologica* 50:181-186, 1980).

And fifth, epidemiology. The least understood feature of the measles virus theory of SSPE aetiopathogenesis is the extremely low incidence of the disease. Despite numerous investigations into the role of socioeconomic, demographic and genetic factors (Canal and Torck, *J. Neurol. Sci.* 1:380-389, 1964; Pettay *et al.*, *J. Infect. Dis.* 124:439-444, 1971; Haddad *et al.*, *Lancet* 2:1025, 1974; Soffer *et al.*, *Israeli J. Med. Sci.* 11:1-4, 1975; Naruszewicz-Lesiuk *et al.*, *Przeg Epidemiologiczna* 23:1-8, 1979; Moodie *et al.*, *South African Med. J.* 58:964-967, 1980; Dick, *Brit. Med. J.* 3:359-360, 1973; Jabbour *et al.*, *J. A. M. A.* 220:959-962, 1972; Baguley and Glasgow, *Lancet* 2:763-765, 1973; Modlin *et al.*, *Pediatrics* 59:505-512, 1977; Halsey *et al.*, *Am. J. Epidemiol.* 111:415-424, 1980; Dyken *et al.*, *Morb. Mortal. Weekly Report* 31:585-588, 1982), it has, until now, been completely unclear why SSPE is so rare when measles virus annually infects millions of children throughout the world.

A much more common idiopathic neurological and/or neuropsychiatric disease, which affects more than a half million Americans, is chronic fatigue syndrome (CFS), which frequently involves concurrent epileptiform symptomology. (P. H. Levine, *What we know about chronic fatigue*

syndrome and its relevance to the practicing physician, Am. J. Med. 105(3A):100S-03S [1998]). Chronic fatigue syndrome is characterized by a sudden onset of persistent, debilitating fatigue and energy loss that lasts at least six months and cannot be attributed to other medical or psychiatric conditions; symptoms include headache, cognitive and behavioral impairment (e.g., short-term memory loss), sore throat, pain in lymph nodes and joints, and low grade fever. (M. Terman *et al.*, *Chronic Fatigue Syndrome and Seasonal; Affective Disorder: Comorbidity, Diagnostic Overlap, and Implications for Treatment*, Am. J. Med. 105(3A):115S-24S [1998]). Depression and related symptoms are also common, including sleep disorders, anxiety, and worsening of premenstrual symptoms or other gynecological complications. (A.L. Komaroff and D. Buchwald, *Symptoms and signs of chronic fatigue syndrome*, Rev. Infect. Dis. 13:S8-S11 [1991]; B.L. Harlow *et al.*, *Reproductive correlates of chronic fatigue syndrome*, Am. J. Med. 105(3A):94S-99S [1998]). Other physiologic abnormalities are also associated with CFS in many patients, including neurally-mediated hypotension, hypocortisolism, and immunologic dysregulation. (P.H. Levine [1998]). A subgroup of CFS patients complain of exacerbated mood state, diminished ability to work and difficulty awakening during winter months, reminiscent of seasonal affective disorder. (M. Terman *et al.* [1998]).

The etiology of CFS has been unknown, and the heterogeneity of CFS symptoms has precluded the use of any particular diagnostic laboratory test. (P.H. Levine [1998]). Symptomatic parallels have been suggested between CFS and a number of other disease conditions, resulting from viral or bacterial infection, toxic exposure, orthostatic hypotension, and stress, but none of these has been shown to have a causal role in CFS. (E.g., I.R. Bell *et al.*, *Illness from low levels of environmental chemicals: relevance to chronic fatigue syndrome and fibromyalgia*, Am. J. Med. 105(3A):74S-82S [1998]; R.L. Bruno *et al.*, *Parallels between post-polio fatigue and chronic fatigue syndrome: a common pathophysiology?*, Am. J. Med. 105(3A):66S-73S [1998]; R. Glaser and J.K. Kiecolt-Glaser, *Stress-associated immune modulation: relevance to viral infections and chronic fatigue syndrome*, Am. J. Med. 105(3A):35S-42S [1998]; P.C. Rowe and H. Calkins, *Neurally mediated hypotension and chronic fatigue syndrome*, Am. J. Med. 105(3A):15S-21S [1998]; L.A. Jason *et al.*, *Estimating the prevalence of chronic fatigue syndrome among nurses*, Am. J. Med. 105(3A):91S-93S [1998]). Accordingly, there has been no known cause to which diagnosis and/or treatment of CSF could be directed. Consequently, the diagnosis and treatment of CFS have continued to be directed to symptoms, rather than to an underlying treatable cause. For example, the use of relaxin has been described for relaxing the involuntary muscles and thus relieve pain associated

with CFS. (S.K. Yue, *Method of treating myofascial pain syndrome with relaxin*, U.S. Patent No. 5,863,552).

There remains a need for an underlying causal factor for many idiopathic neurological, neurodegenerative, neuropsychological and neuropsychiatric disorders and primary tracheobronchial and/or lymphadenopathy-associated diseases, to which diagnostic testing, research and development, including screening of potential new antiviral drugs, and treatment can be directed. This and other benefits of the present invention are described herein.

SUMMARY OF THE INVENTION

The present invention is based on the discovery and isolation of a novel human virus that has been designated as a "*Cryptovirus*", which falls within the genus *Rubulavirus* of the family Paramyxoviridae. The genome of the isolated *Cryptovirus* of the present invention is a minus strand RNA having a nucleotide sequence entirely complementary to (SEQ ID NO:1).

The present invention relates to isolated nucleic acids that are *Cryptovirus*-specific. The inventive *Cryptovirus*-specific nucleic acids encompass: (A) nucleotide sequence of contiguous nucleotide positions 1-15246 of (SEQ ID NO:1), such as, but not limited, to plus strand RNAs (e.g., mRNAs) and cDNAs; or (B) a nucleotide sequence complementary to contiguous nucleotide positions 1-15246 of (SEQ ID NO:1), such as, but not limited to minus strand RNAs (e.g., genomic or cloned RNAs) and cDNAs; or (C) *Cryptovirus*-specific fragments of (A) or (B), such fragments being at least about five nucleotides long. The present invention encompasses both RNAs and DNAs, and thus it is understood by the skilled artisan that the present invention encompasses nucleic acids, i.e., RNAs, in which uracil residues ("U") replace the thymine residues ("T") in (SEQ ID NO:1). The inventive nucleic acids include useful *Cryptovirus*-specific probes and primers.

Inventive nucleic acid constructs, including cloning vectors and expression vectors, are provided that contain the inventive nucleic acid. Such inventive recombinant vectors are contained in a host cell of the present invention.

The present invention also relates to an isolated *Cryptovirus* protein encoded by a *Cryptovirus*-specific nucleic acid segment. The inventive *Cryptovirus* proteins include isolated *Cryptovirus* nucleocapsid and envelope proteins and chimeric proteins comprising a *Cryptovirus* protein moiety.

The invention relates to an isolated virion or other viral particle that contains the inventive *Cryptovirus* nucleic acid, such as a viral expression vector, or contains the inventive *Cryptovirus*

protein, such as an inventive pseudotyped virion or an inventive isolated *Cryptovirus* virion or other *Cryptovirus* particle.

5 Inventive compositions of matter that include the inventive *Cryptovirus* nucleic acid, *Cryptovirus* protein, or isolated virions and other viral particles, together with a carrier, are also included in the present invention.

Moreover, the present invention provides a method of isolating a *Cryptovirus* virion. The inventive method involves culturing a plurality of peripheral blood mononuclear cells (PBMNCs) that have been obtained from a human having a *Cryptovirus* infection. The PBMNCs are cultured in an artificial aqueous medium that includes an agent that increases cellular guanylyl cyclase activity, such as but not limited to cyclic GMP. The PBMNCs are then co-cultured in with a plurality of mammalian amnion cells in fresh aqueous medium including the agent, and the co-culture is passaged one or more times. Passaging is followed by co-cultivating a plurality of mammalian epithelial cells together with the PBMNCs and the mammalian amnion cells in fresh aqueous medium comprising the agent. This co-cultivation results in the production of *Cryptovirus* virions that are released into the aqueous medium. A supernatant of the aqueous medium is separated from the cells in the culture, to obtain the *Cryptovirus* virions, which are found in the supernatant. The inventive method facilitates the isolation from cellular material of *Cryptovirus* virions in great numbers. Virions isolated thereby can be further propagated by an inventive method of propagating a *Cryptovirus*, which the present invention provides.

20 The inventive method of propagating a *Cryptovirus* involves exposing a plurality of mammalian epithelial cells to a plurality of cell-free *Cryptovirus* virions, thus isolated, and further cultivating the *Cryptovirus* virion-exposed mammalian epithelial cells in an artificial aqueous medium comprising an agent that increases the activity of cellular guanylyl cyclase. Thus, a mammalian epithelial cell acutely infected with *Cryptovirus* is provided, which inventive cell is produced by the method.

25 The present invention also relates to a method of producing a mammalian cell line nonproductively infected with *Cryptovirus*. The method involves co-culturing PBMNCs that have been obtained from a human having a *Cryptovirus* infection, with mammalian amnion cells (e.g., rodent or primate amnion cells), in an artificial aqueous medium comprising an agent that increases cellular guanylyl cyclase activity, such that the mammalian amnion cells become nonproductively infected by *Cryptovirus*. After passaging the nonproductively infected mammalian amnion cells with the peripheral blood mononuclear cells, the co-culture becomes a monoculture of the nonproductively

infected mammalian amnion cells. The present invention also relates to a cell nonproductively infected with *Cryptovirus*, which cell is produced in accordance with the method.

Cryptovirus is associated with cryptogenic and idiopathic forms of human disease, e.g., epilepsy. *Cryptovirus* is also associated with other human neurological, neurodegenerative, and/or neuropsychiatric diseases where neural dysfunction and neuropathology are evident and where epileptiform symptomology is always concurrent (e.g. subacute sclerosing panencephalitis, SSPE) or is frequently concurrent (e.g., multiple sclerosis [MS] and chronic fatigue syndrome [CFS]). Thus, the inventive cell lines, viral particles and virions are particularly useful for screening potential antiviral agents to discover those that could be effective in treating mammals, including humans, infected with *Cryptovirus*.

In particular, useful in vitro methods of screening a potential antiviral therapeutic agent are provided. In accordance with the in vitro screening methods, the inventive *Cryptovirus*-infected cells are cultured, and then exposed to the potential antiviral therapeutic agent. If acutely infected mammalian epithelial cells are used, then the effect of the potential antiviral therapeutic agent on *Cryptovirus* replication and/or *Cryptovirus* virion assembly is measured (e.g., effect on *Cryptovirus* genomic replication, *Cryptovirus* transcription, and/or translation, i.e., protein synthesis, from *Cryptovirus* mRNAs, effect on numbers of *Cryptovirus* virions produced or completeness of *Cryptovirus* particles). Inhibition of *Cryptovirus* replication and/or *Cryptovirus* virion assembly, relative to a control not receiving the agent, indicates antiviral activity of the potential therapeutic agent. Alternatively, if nonproductively infected cells are used, measurement is made of the effect of the potential antiviral therapeutic agent on *Cryptovirus* replication, *Cryptovirus* genome replication, and/or *Cryptovirus*-specific transcription. Inhibition of *Cryptovirus* replication, *Cryptovirus* genome replication, and/or *Cryptovirus*-specific transcription, relative to a control not receiving the agent, indicates antiviral activity of the potential therapeutic agent. These inventive methods are useful for identifying, screening, or isolating promising new antiviral drugs. Once the potential of a chemical agent is identified by the inventive methods, then, further research can be done to ascertain its clinical usefulness. Thus, the inventive methods of screening a potential chemotherapeutic agent are of benefit in finding and developing pharmaceutical antiviral drugs aimed at treating *Cryptovirus*-related conditions and other conditions associated with other viruses of the Mononegavirales.

The present invention now also provides an animal model for the study of human diseases, for example a neurological, neurodegenerative, and/or neuropsychiatric disease (e.g., idiopathic epileptiform diseases, such as epilepsy, SSPE, MS, and CFS). The animal model involves a non-human mammal, which has been inoculated with an infectious cell-free *Cryptovirus* having a genome

comprising a single stranded RNA complementary to (SEQ ID NO:1), or has been inoculated with a cell nonproductively-infected with the *Cryptovirus*. The inoculated non-human mammal of the animal model exhibits at least one symptom characteristic of a human disease after being thus inoculated, which was not previously exhibited by the non-human mammal before inoculation.

5 The animal model is useful in an in vivo method of screening a potential antiviral therapeutic agent. The method involves administering the potential therapeutic agent to be screened, to the inventive animal model. Before administration of the potential therapeutic agent, the non-human mammal exhibits at least one symptom characteristic of a human disease. After administration of the potential therapeutic agent, the presence or absence of a beneficial antiviral effect is detected; the
10 presence of a beneficial antiviral effect, in comparison to a control animal not receiving the agent, indicates activity of the potential therapeutic agent.

 Employing an alternative embodiment of the inventive animal model, an in vivo method of screening a potential antiviral prophylactic agent is provided. The method involves administering a potential prophylactic agent to be screened to a non-human mammal, which does not have a symptom
15 of a human disease, for example a neurological, neurodegenerative, and/or neuropsychiatric disease. Then the animal is inoculated, as previously described, with an infectious cell-free *Cryptovirus* having a genome comprising a single stranded RNA complementary to (SEQ ID NO:1), or with a mammalian cell nonproductively-infected with the *Cryptovirus*. Subsequently, the presence or absence in the non-human mammal of a beneficial antiviral effect is detected, compared to a control
20 not receiving the potential prophylactic agent. The subsequent presence of a beneficial antiviral effect in the inoculated non-human mammal indicates activity of the potential prophylactic agent.

 The inventive nucleic acid constructs, *Cryptovirus* proteins, and particles and virions are also particularly useful in producing *Cryptovirus*-specific antibodies, and in the production or manufacture of vaccines, which antibodies and vaccines are directed specifically against *Cryptovirus* proteins,
25 such as the nucleocapsid or envelope proteins of *Cryptovirus*. These vaccines can include live attenuated virus; killed virus; recombinant chimeric viruses; proteins or other parts of virus; or one or more isolated or recombinantly expressed *Cryptovirus* proteins.

 The present invention relates also to an isolated antibody that specifically binds a *Cryptovirus* protein and the use of the inventive antibody in manufacturing a medicament for the treatment of
30 *Cryptovirus* infections. Also provided are compositions of matter comprising the antibody and a carrier.

In other aspects, the invention is usefully directed to methods and assays, e.g., for determining whether biological materials are contaminated with *Cryptovirus* or whether a mammal, including a human, is or has been infected with *Cryptovirus*.

5 In particular, the invention provides methods of detecting the presence or absence of a *Cryptovirus* protein, *Cryptovirus*-specific RNA, or *Cryptovirus*-specific antibody in a sample of a biological material, such as serum.

10 In the method of detecting *Cryptovirus* protein, the sample of the biological material is contacted with an inventive antibody that specifically binds a *Cryptovirus* protein; and if the presence of specific binding of the antibody to a constituent of the sample is detected, this indicates the presence of the *Cryptovirus* protein in the sample.

Similarly, in the method of detecting *Cryptovirus*-specific RNA in a sample of a biological material containing RNA, the sample is contacted with the inventive *Cryptovirus*-specific probe under at least moderately stringent hybridization conditions, and the formation of detectable hybridization products indicates the presence of the *Cryptovirus* RNA in the sample.

15 Alternatively, the sample containing RNA is subjected to an amplification of *Cryptovirus*-specific RNA in the sample, using at least one inventive *Cryptovirus*-specific primer in an amplification reaction mixture. By detecting the presence or absence of *Cryptovirus*-specific nucleic acid amplification products in the amplification reaction mixture, the presence or absence of *Cryptovirus*-specific RNA in the sample can be determined, with the presence of *Cryptovirus*-specific
20 amplification products in the reaction mixture indicating the presence of the *Cryptovirus*-specific RNA in the sample.

The present invention also provides a method of detecting the presence or absence of a *Cryptovirus*-specific antibody in a sample of an antibody-containing biological material, such as serum. The method involves contacting the sample of biological material with the inventive protein,
25 such as a *Cryptovirus* envelope protein, or alternatively, with the inventive virion or viral particle, under conditions allowing the formation of a specific protein-antibody complex, or antibody-bound virus complex, respectively. Detection of the presence of such specific protein-antibody complexes, or antibody-bound virus complexes, indicates the presence of the *Cryptovirus*-specific antibody in the sample. Inventive anti-*Cryptovirus* antibody detecting kits are also provided, which are useful for
30 practicing the method.

Thus, by practicing any of the foregoing inventive methods of detecting the presence or absence of a *Cryptovirus* protein, *Cryptovirus*-specific RNA, or *Cryptovirus*-specific antibody, with a sample of biological materials from a mammal, including a human, an inventive method of detecting

or diagnosing a *Cryptovirus* infection in the mammal is provided, as indicated by the presence in the sample of *Cryptovirus* protein, *Cryptovirus*-specific RNA, or *Cryptovirus*-specific antibody. These diagnostic methods are valuable because, regardless of the therapeutic strategy, it is advantageous to begin therapy at the time of "primary" infection
 5 (i.e., the first exposure to the virus) or as soon as possible thereafter (i.e., during development of the primary infection).

In a first aspect the invention provides an isolated nucleic acid, comprising:

- (A) contiguous nucleotide positions 1-15246 of (SEQ ID NO: 1);
- (B) a nucleotide sequence complementary to (A); or
- 10 (C) *Cryptovirus*-specific fragment of (A) or (B), comprising a nucleic acid segment selected from the group consisting of:
 - (i) contiguous nucleotide positions 152-1678 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (ii) contiguous nucleotide positions 1850-2515 of (SEQ ID NO:1), a
 15 complementary sequence, or a degenerate coding sequence;
 - (iii) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (iv) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1) combined with a further insertion of two guanine residues between nucleotide position
 20 2339 of (SEQ ID NO:1) and nucleotide position 2340 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (v) contiguous nucleotide positions 3141-4271 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (vi) contiguous nucleotide positions 4530-6182 of (SEQ ID NO:1), a
 25 complementary sequence, or a degenerate sequence;
 - (vii) contiguous nucleotide positions 4587-6182 of (SEQ ID NO:1), a complementary sequence, or a degenerate sequence;
 - (viii) contiguous nucleotide positions 4587-4835 of (SEQ ID NO:1), a complementary sequence, or a degenerate sequence;
 - 30 (ix) contiguous nucleotide positions 4836-6182 of (SEQ ID NO:1), a complementary sequence, or a degenerate sequence;
 - (x) contiguous nucleotide positions 4272-6515 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;

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(xi) contiguous nucleotide positions 6303-6434 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;

(xii) contiguous nucleotide positions 6584-8278 of (SEQ ID. NO:1), a complementary sequence, or a degenerate coding sequence; and

5 (xiii) contiguous nucleotide positions 8414-15178 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xiv) contiguous nucleotide positions 1684-1701 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

10 (xv) contiguous nucleotide positions 1700-1717 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xvi) contiguous nucleotide positions 4283-4300 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xvii) contiguous nucleotide positions 4299-4316 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

15 (xviii) contiguous nucleotide positions 4285-4302 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xix) contiguous nucleotide positions 4300-4317 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

20 (xx) contiguous nucleotide positions 4518-4535 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxi) contiguous nucleotide positions 4533-4550 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxii) contiguous nucleotide positions 6191-6208 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

25 (xxiii) contiguous nucleotide positions 6116-6133 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxiv) contiguous nucleotide positions 6192-6209 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

30 (xxv) contiguous nucleotide positions 7501-7518 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxvi) contiguous nucleotide positions 7517-7534 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxvii) contiguous nucleotide positions 4292-4549 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence.

14b

In a second aspect the invention provides a nucleic acid construct, comprising the nucleic acid of the first aspect.

In a third aspect the invention provides an expression vector, comprising the nucleic acid construct of the second aspect.

5 In a fourth aspect the invention provides a cloning vector, comprising the nucleic acid construct of the second aspect.

In a fifth aspect the invention provides a host cell, comprising the expression vector of the third aspect or the cloning vector of the fourth aspect.

10 In a sixth aspect the invention provides an isolated *Cryptovirus* protein encoded by a nucleic acid segment comprising:

(A) contiguous nucleotide positions 152-1678 of (SEQ ID NO: 1) or a degenerate sequence;

(B) contiguous nucleotide positions 1850-2515 of (SEQ ID NO: 1) or a degenerate sequence.

15 (C) contiguous nucleotide positions 1850-3023 of (SEQ ID NO: 1) combined with a further insertion of two guanine residues between nucleotide position 2339 of (SEQ ID NO: 1) and a nucleotide position 2340 of (SEQ ID NO: 1), or a degenerate sequence.

(D) contiguous nucleotide positions 3141-4271 of (SEQ ID NO: 1) or a degenerate sequence.

20 (E) contiguous nucleotide positions 4530-6182 of (SEQ ID NO: 1) or a degenerate sequence.

(F) contiguous nucleotide positions 4587-6182 of (SEQ ID NO: 1) or a degenerate sequence.

25 (G) contiguous nucleotide positions 4587-4835 (SEQ ID NO: 1) or a degenerate sequence.

(H) contiguous nucleotide positions 4836-6182 of (SEQ ID NO: 1) or a degenerate sequence.

(I) contiguous nucleotide positions 6303-6434 of (SEQ ID NO: 1) or a degenerate sequence.

30 (J) contiguous nucleotide positions 6584-8278 of (SEQ ID NO: 1) or a degenerate sequence.

(K) contiguous nucleotide positions 8414-15178 of (SEQ ID NO: 1) or a degenerate sequence.

14c

In a seventh aspect embodiment the invention provides a chimeric protein, comprising a *Cryptovirus* protein encoded by a nucleic acid segment comprising:

(A) contiguous nucleotide positions 152-1678 of (SEQ ID NO: 1) or a degenerate sequence.

5 (B) contiguous nucleotide positions 1850-2515 of (SEQ ID NO: 1) or a degenerate sequence.

(C) contiguous nucleotide positions 1850-3023 of (SEQ ID NO: 1) combined with a further insertion of two guanine residues into the nucleotide sequence between nucleotide position 2339 of (SEQ ID NO: 1) and a nucleotide position 2340 of (SEQ ID NO: 1), or a degenerate sequence.

10 (D) contiguous nucleotide positions 3141-4271 of (SEQ ID NO: 1) or a degenerate sequence.

(E) contiguous nucleotide positions 4530-6182 of (SEQ ID NO: 1) or a degenerate sequence.

15 (F) contiguous nucleotide positions 4587-6182 of (SEQ ID NO: 1) or a degenerate sequence.

(G) contiguous nucleotide positions 4587-4835 (SEQ ID NO: 1) or a degenerate sequence.

(H) contiguous nucleotide positions 4836-6182 of (SEQ ID NO: 1) or a degenerate sequence.

20 (I) contiguous nucleotide positions 6303-6434 of (SEQ ID NO: 1) or a degenerate sequence.

(J) contiguous nucleotide positions 6584-8278 of (SEQ ID NO: 1) or a degenerate sequence.

25 (K) contiguous nucleotide positions 8414-15178 of (SEQ ID NO: 1) or a degenerate sequence.

In one embodiment of the chimeric protein the *Cryptovirus* protein is a *Cryptovirus* envelope protein encoded by a nucleic acid segment comprising (E), (F), (G), (H), (I) or (J).

30 In an eighth aspect the invention provides use of the protein of the sixth or seventh aspect in producing a *Cryptovirus*-specific antibody.

In a ninth aspect the invention provides an isolated antibody that specifically binds the protein of the sixth or seventh aspect.

35 In one embodiment of the ninth aspect of the invention the antibody is polyclonal.

14d

In one embodiment of the ninth aspect of the invention the antibody is monoclonal.

In one embodiment of the ninth aspect of the invention the antibody is chimeric.

In a tenth aspect the invention provides use of the antibody of the ninth aspect in
5 manufacturing a medicament for the treatment of *Cryptovirus* infections.

In an eleventh aspect the invention provides an isolated viral particle comprising the nucleic acid of the first aspect.

In a twelfth aspect the invention provides an isolated *Cryptovirus* particle, comprising a genome having a nucleotide sequence entirely complementary to (SEQ ID
10 NO: 1).

In a thirteenth aspect the invention provides a composition of matter, comprising the nucleic acid of the first aspect, the protein of the sixth or seventh aspect, the antibody of the ninth aspect or the virion of the eleventh aspect, and a carrier.

In a fourteenth aspect the invention provides an isolated viral particle comprising
15 the protein of the sixth or seventh aspect.

In a fifteenth aspect the invention provides use of the nucleic acid of the first aspect, the nucleic acid construct of the second aspect, the protein of the sixth or seventh aspect, the viral particle of the ninth or eleventh aspect or the isolated *Cryptovirus* particle of the twelfth or fourteenth aspect in manufacturing a vaccine.

In one embodiment of the fifteenth aspect the viral particle is an attenuated
20 virion.

In one embodiment of the fifteenth aspect of the viral particle is a killed virion.

In a sixteenth aspect the invention provides an isolated *Cryptovirus* particle, wherein the *Cryptovirus* is Strain BBR.

In a seventeenth aspect the invention provides a probe or primer comprising the
25 nucleic acid of the first aspect.

In an eighteenth aspect the invention provides a method of detecting the presence or absence of a *Cryptovirus* protein in a sample of a biological material, comprising:

contacting the sample of the biological material with the antibody of the
30 invention; and

detecting specific binding of the antibody to a constituent of the sample, wherein the presence of specific binding indicates the presence of the *Cryptovirus* protein in the sample.

In a nineteenth aspect invention provides a method of detecting the presence or
35 absence of a *Cryptovirus*-specific RNA in a sample of a biological material, comprising:

obtaining a sample of a biological material comprising RNA;

contacting the sample with the probe of the invention under at least moderately stringent hybridization conditions, wherein the formation of detectable hybridization products indicates the presence of the *Cryptovirus*-specific RNA in the sample.

5 In a twentieth aspect the invention provides a method of detecting the presence or absence of a *Cryptovirus*-specific RNA in a sample of a biological material, comprising:

obtaining a sample of a biological material comprising RNA;

10 amplifying *Cryptovirus*-specific RNA in the sample using at least one primer of the invention in an amplification reaction mixture;

then detecting the presence or absence of *Cryptovirus*-specific nucleic acid amplification products in the amplification reaction mixture, wherein the presence of the amplification products in the reaction mixture indicates the presence of *Cryptovirus* RNA in the sample.

15 In one embodiment of the eighteenth, nineteenth or twentieth aspect the biological material is a cellular material.

In one embodiment of the eighteenth, nineteenth or twentieth aspect the biological material is blood or serum.

20 In one embodiment of the eighteenth, nineteenth or twentieth aspect the biological material is cerebrospinal fluid.

In one embodiment of the eighteenth, nineteenth or twentieth aspect the biological material is lymphoid tissue.

25 In one embodiment of the eighteenth, nineteenth or twentieth aspect the biological material is nervous tissue. In one embodiment the nervous tissue is brain tissue.

In a twentyfirst aspect the invention provides a method of detecting the presence or absence of a *Cryptovirus*-specific antibody in a sample of a biological material, comprising:

contacting the sample with the protein according to the invention;

30 allowing the formation of a specific protein-antibody complex;

detecting the presence of the specific protein-antibody complex, wherein the presence of a specific protein-antibody complex indicates the presence of the *Cryptovirus*-specific antibody in the sample.

In a twentysecond aspect the invention provides a method of detecting the presence or absence of a *Cryptovirus*-specific antibody in a sample of a biological material, comprising:

- contacting the sample with the protein of the invention;
- 5 allowing the formation of a specific protein-antibody complex;
- detecting the presence of the specific protein-antibody complex, wherein the presence of a specific protein-antibody complex indicates the presence of the *Cryptovirus*-specific antibody in the sample.

In a twentythird aspect the invention provides an assay method for detecting the presence or absence of an antibody that selective binds *Cryptovirus* in a sample of an antibody-containing biological material originating from a human, comprising:

- 10 contacting the sample, the sample originating from an individual suspected of having a *Cryptovirus* infection, with the envelope protein of the invention such that if antibody selectively binding *Cryptovirus* is present, an antibody-bound envelope protein
- 15 complex forms;

- contacting any antibody-bound protein complexes thus formed with an anti-human-antibody binding antibody, and allowing the formation of complexes of the antibody, with the antibody-bound envelope protein complexes; and

- 20 detecting the presence or absence of any antibody-bound envelope protein complexes thus formed, the presence of such complexes indicating the presence in the sample of antibody selectively binding *Cryptovirus*.

In a twentyfourth aspect the invention provides an assay method for detecting the presence or absence of antibody that selectively binds *Cryptovirus* antigen in a sample of an antibody-containing biological material originating from a human, the method

25 comprising:

- contacting the sample, the sample originating from an individual suspected of having a *Cryptovirus* infection, with the viral particle of the invention, such that, if antibody selectively binding *Cryptovirus* antigen is present, an antibody-bound virus
- 30 complex forms;

- contacting any antibody-bound virus complexes thus formed with anti-human antibody-binding antibody, and allowing the formation of complexes of the anti-human antibody-binding antibody with the antibody-bound virus complexes; and

- 35 detecting the presence or absence of any complexes formed, the presence of such complexes indicating the presence in the sample of antibody selectively binding *Cryptovirus* antigen.

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14g

In a twentyfifth aspect the invention provides a method of detecting a *Cryptovirus* infection in a mammal, comprising:

obtaining a sample of a biological material from the mammal; and
performing the method of the eighteenth, nineteenth, twentieth, twentyfirst,
5 twentysecond, twentythird or twentyfourth aspect using the sample, whereby detecting the
presence of the *Cryptovirus* protein, *Cryptovirus*-specific RNA, and/or *Cryptovirus*-
specific antibody in the sample indicates a *Cryptovirus* infection in the mammal.

In one embodiment of the twenty first, twentysecond, twentythird, twentyfourth
or twentyfifth aspect the biological material is a cellular material.

10 In one embodiment of the twenty first, twentysecond, twentythird, twentyfourth
or twentyfifth aspect the biological material is blood or serum.

In one embodiment of the twenty first, twentysecond, twentythird, twentyfourth
or twentyfifth aspect the biological material is cerebrospinal fluid.

In one embodiment of the twenty first, twentysecond, twentythird, twentyfourth
15 or twentyfifth aspect the biological material is lymphoid tissue.

In one embodiment of the twenty first, twentysecond, twentythird, twentyfourth
or twentyfifth aspect the biological material is nervous tissue. In one embodiment the
nervous tissue is brain tissue.

In one embodiment of the twentyfifth aspect the mammal is a human. In one
20 embodiment the human has a neurological, neurodegenerative, and/or neuropsychiatric
disease. In one embodiment the human has a primary tracheobronchial and/or
lymphadenopathy-associated illness.

In a twenty-sixth aspect the invention provides a method of isolating a
Cryptovirus virion, comprising:

25 (a) culturing a plurality of peripheral blood mononuclear cells that have
been obtained from a human having a *Cryptovirus* infection, in an artificial aqueous
medium comprising an agent that increases cellular guanylyl cyclase activity;

(b) co-culturing the plurality of peripheral blood mononuclear cells with a
plurality of mammalian amnion cells in fresh artificial aqueous medium comprising an
30 agent that increases cellular guanylyl cyclase activity;

14h

(c) passaging the peripheral blood mononuclear cells with the mammalian amnion cells in co-culture;

(d) co-cultivating a plurality of mammalian epithelial cells together with the peripheral blood mononuclear cells and the mammalian amnion cells in fresh artificial aqueous medium comprising an agent that increases cellular guanylyl cyclase activity; and

(e) separating a supernatant of the aqueous medium from the cells, to obtain a *Cryptovirus* virion in the supernatant.

In a twentyseventh aspect the invention provides a method of propagating a *Cryptovirus*, comprising:

(a) exposing a plurality of mammalian epithelial cells to a plurality of cell-free *Cryptovirus* virions, said *Cryptovirus* virions having been isolated by the method of the invention; and

(b) further cultivating the mammalian epithelial cells, thus virion-exposed, in an artificial aqueous medium comprising an agent that increases the activity of cellular guanylyl cyclase.

In a twentyeighth aspect the invention provides a method of producing a mammalian cell line non-productively infected with *Cryptovirus*, comprising:

(a) co-culturing peripheral blood mononuclear cells that have been obtained from a human having a *Cryptovirus* infection, with mammalian amnion cells, in an artificial aqueous medium comprising an agent that increases cellular guanylyl cyclase activity, such that the mammalian amnion cells become non-productively infected by *Cryptovirus*; and

(b) passaging the non-productively infected mammalian amnion cells with the peripheral blood mononuclear cells, whereby the co-culture becomes a monoculture of the non-productively infected mammalian amnion cells.

In one embodiment of the twentysixth, twentyseventh or twentyeighth aspect the mammalian amnion cells are human amnion cells. In one embodiment the human amnion cells are AV₃ cells.

In one embodiment of the twentysixth, twentyseventh or twentyeighth aspect the mammalian epithelial cells are simian epithelial cells selected from the group consisting of Vero or CV-1 cells. In one embodiment the CV-1 cells are subline CV-1 cells.

In one embodiment of the twentysixth, twentyseventh or twentyeighth aspect the agent that increases cellular guanylyl cyclase activity is cyclic GMP, insulin, zinc

dication, or a combination of any of these. In one embodiment the cyclic GMP is in a concentration of about 0.05 to about 5mM in the artificial aqueous medium.

In one embodiment of the twentyseventh, twentyeighth or twentyninth aspect the agent that increases cellular guanylyl cyclase activity is nitric oxide or a nitric oxide donor selected from the group consisting of organic nitrate compounds, iron nitrosyl compounds, S-nitrosothiol compounds, sydnonimine compounds, and nonoate compounds.

In one embodiment of the twentyseventh, twentyeighth or twentyninth aspect the aqueous medium further comprises glutamine.

In a thirtieth aspect the invention provides a method of producing a mammalian epithelial cell line acutely infected with *Cryptovirus*, comprising the method of the invention.

In a thirtyfirst aspect the invention provides a mammalian epithelial cell acutely infected with *Cryptovirus*, said cell being produced by the method of the invention.

In a thirtysecond aspect the invention provides a cell non-productively infected with *Cryptovirus*, wherein said cell is produced in accordance with a method of the invention.

In a thirtythird aspect the invention provides an *in vitro* method of screening a potential antiviral therapeutic agent, comprising:

- (a) culturing a mammalian epithelial cell acutely infected with *Cryptovirus* of the invention;
- (b) exposing the cells to the potential antiviral therapeutic agent; and
- (c) measuring the effect of the agent on *Cryptovirus* replication and/or *Cryptovirus* virion assembly, wherein inhibition of *Cryptovirus* replication and/or *Cryptovirus* virion assembly relative to a control indicates antiviral activity of the potential therapeutic agent.

In a thirtyfourth aspect the invention provides an *in vitro* method of screening a potential antiviral therapeutic agent, comprising:

- (a) culturing a cell non-productively infected with *Cryptovirus* of the invention;
- (b) exposing the cells to the potential antiviral therapeutic agent; and
- (c) measuring the effect of the agent on *Cryptovirus* replication, *Cryptovirus* genome replication, and/or *Cryptovirus*-specific wherein inhibition of *Cryptovirus* replication, *Cryptovirus* genome replication, and/or *Cryptovirus*-specific relative to a control, indicates antiviral activity of the potential therapeutic agent

14j

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In a thirtyfourth aspect the invention provides an animal model for the study of human diseases, comprising a non-human mammal, said non-human mammal having been artificially inoculated with an infectious cell-free *Cryptovirus* having a genome comprising a single stranded RNA complementary to (SEQ ID NO: 1), or having been
5 inoculated with a cell non-productively-infected with the *Cryptovirus*, whereby the non-human mammal exhibits at least one symptom characteristic of a human disease after being thus inoculated, said symptom not being previously exhibited by the non-human mammal.

10 In one embodiment of the thirtyfourth aspect the non-human mammal is a rodent or lagomorph.

In one embodiment of the thirtyfourth aspect the non-human mammal is a non-human primate.

In one embodiment of the thirtyfourth aspect the human disease is a neurological, neurodegenerative, and/neuropsychiatric disease.

15 In a thirtyfifth aspect the invention provides an in vivo method of screening a potential therapeutic agent, comprising:

(a) administering the potential therapeutic agent to be screened to the animal model of the invention, wherein the non-human mammal exhibits, before administration of the potential therapeutic agent, at least one symptom characteristic of a
20 human disease; and

(b) detecting the present or absence of a beneficial antiviral effect of the potential therapeutic agent, wherein the presence of a beneficial antiviral effect indicates activity of the potential therapeutic agent.

25 In a thirtysixth aspect the invention provides an in vivo method of screening a potential prophylactic agent, comprising:

(a) administering the potential prophylactic agent to be screened, to a non-human mammal not previously having a symptom of a human disease;

(b) inoculating the non-human mammal with an infectious cell-free *Cryptovirus* having a genome comprising a single stranded RNA complementary to (SEQ
30 ID NO: 1), or with a mammalian cell non-productively-infected with the *Cryptovirus*; and

(c) detecting the subsequent presence or absence in the non-human mammal of a beneficial antiviral effect, whereby the presence of a beneficial antiviral effect in the inoculated non-human mammal indicates activity of the potential prophylactic agent.

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In one embodiment of the thirtysixth aspect the potential prophylactic agent is an immunoprophylactic agent.

In one embodiment of the thirtyfifth or thirtysixth aspect the non-human mammal is a rodent or a lagomorph.

5 In one embodiment of the thirtyfifth or thirtysixth aspect the non-human mammal is a non-human primate.

In one embodiment of the thirtyfifth or thirtysixth aspect the human disease is a neurological, neurodegenerative, and/or neuropsychiatric disease.

10 In a thirtyseventh aspect the invention provides an anti-*Cryptovirus* antibody detecting kit, comprising:

the *Cryptovirus* particle of the invention; and
a labelled anti-human antibody-binding antibody.

In one embodiment of the thirtyseventh aspect the kit further comprises a solid matrix for supporting said *Cryptovirus* particle.

15 In thirtyeighth aspect the invention provides an anti-*Cryptovirus* antibody detecting kit, comprising:

the protein according to the invention; and
a labeled anti-human antibody-binding antibody.

20 In one embodiment of the thirtyeighth aspect the kit further comprises a solid matrix for supporting said protein.

Other features, objects, and advantages of the invention will be apparent from the accompanying drawings and the detailed description of the preferred embodiments hereinbelow.

25 BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a chart showing the taxonomic assignment of the human *Cryptovirus* of the present invention to the genus *Rubulavirus* of the Paramyxoviridae.

Fig. 2 is a phylogenetic tree, modified from the version appearing in Collins *et al.* (Chapter 41, page 1206, *Parainfluenza Viruses*, in *Virology*, 3rd Ed., Fields, Knipe, and Howley, Eds., Lippincott-Raven, Philadelphia, 1996). The modified tree emphasizes
30 the clustering of three *Rubulavirus* species (Porcine *Rubulavirus*; Canine Parainfluenza Virus Type 2; and the human *Cryptovirus* of the present invention) as distinct from the prototype *Rubulavirus* Simian Virus 5.

Fig. 3 is a representation of the genetic maps of typical members of each genus
35 of the family Paramyxoviridae. The gene size is drawn to scale. Vertical lines demark

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gene boundaries. The pneumovirus L gene transcription overlaps that of the 22K (M2) gene and is thus shown in a staggered format. This overlap configuration is seen in human and animal viruses, but not in other pneumoviruses (Lamb and Kolakofsky, Chapter 40, page 1181, *The Viruses and Their Replication*, in *Virology*, *supra*).

5 Fig. 4 is a representation of revised Rubulavirus genetic maps, which distinguish Simian Virus 5 from a cluster of related viruses that demonstrate neurotropism and encode an additional 22 amino acid "tail" at the carboxy terminus of their fusion proteins i. e., the "neurotropic species" of human *Cryptovirus*, Canine Parainfluenza Type 2, and Porcine *Rubulavirus*). The fusion (F) proteins of each neurotropic virus species are more
10 closely related to each other than they are to the fusion protein of Simian Virus 5 (see Fig. 10).

Fig. 5 shows a schematic and comparative autoradiograms of the [³⁵S]-methionine labeled proteins of gradient-purified human *Cryptovirus* (Strain BBR) and Simian Virus 5 (*NIH 21005-2WR* strain) following SDS-PAGE on 10% acrylamide
15 Laemmli slab gels under reducing conditions.

Fig. 6 is a collage of matched sets of fluorescent photomicrographs taken of various SSPE-derived nonproductively infected cell cultures following direct double labeling with rhodamine isothiocyanate-labeled goat anti-measles virus serum (Panels A, C, E, G and I) and rabbit anti-*Cryptovirus* serum, then followed with fluorescein isothiocyanate-labeled goat anti-rabbit IgG (Panels B, D, F, H and J). Panels A and B represent AV₃/SSPE/MV cells persistently-infected with *Cryptovirus* and also infected with the Edmonston strain of measles virus before being passaged onto coverslips for these immunofluorescent studies; Panels C and D represent the nonproductive SSPE-derived cell line designated "Kitaken" (Ueda *et al.*, *Biken Journal* 18:179-181, 1975); Panels E and F represent the nonproductive SSPE-derived cell line designated "Niigata" (Doi *et al.*, *Japan. J. Med. Sci. Biol.* 25:321-333, 1972); Panels G and H and I and J, respectively, represent the nonproductive SSPE-derived cell line designated "Biken" (Yamanouchi *et al.*, *Japan. J. Med. Sci. Biol.* 29:177-186, 1976; Ohuchi *et al.*, *Microbiol. Immunol.* 25:887-983, 1981).

Fig. 7 shows photographs of two male Colored mice, born of the same litter, two months after neonatal (two days after birth) intracerebral inoculation with plaque-purified *Cryptovirus* (strain BBR; Fig. 7A) or with the NIH 21005-2WR strain of Simian Virus 5 (SV5; Fig. 7B).

Fig. 8 shows photographs of two female Colored mice, born of the same litter, three months after neonatal (two days after birth) intracerebral inoculation with plaque-purified *Cryptovirus* (strain BBR; Fig. 8A) and six months after neonatal (two days after birth) intracerebral inoculation with plaque-purified *Cryptovirus* (Fig. 8B).

Fig. 9 is a comparison of the FASTA formatted (i.e., mRNA sense: 5' to 3') sequence of human *Cryptovirus Strain BBR* (SEQ ID NO:1) and Simian Virus 5 *Strain W3A* (SEQ ID NO:2). The number of variations from (SEQ ID NO:1) in each line of (SEQ. ID.NO:2) is tallied in the right-hand margin. The *Cryptovirus*-specific nucleotide positions that differ from the sequence of SV5 are in bold underlined type; if the difference is in a coding region, the relevant amino acid encoded is printed above the codon of the *Cryptovirus* nucleotide sequence, and if the different *Cryptovirus* nucleotide results in a codon encoding a different amino acid than the SV5 codon in the analogous position, an arrow leads from the SV5 amino acid to the different amino acid in the analogous *Cryptovirus* protein. Boxed nucleotides indicate known SV5 and analogous *Cryptovirus* start or stop sites, as indicated.

Fig. 10 is a comparison of Rubulavirus F Protein nucleotide (Fig. 10A; comparison of the FASTA formatted, i.e., mRNA sense: 5' to 3' sequence) and encoded amino acid (Fig. 10B) sequences. The first line (uppermost) represents an embodiment of the sequence of an inventive

Cryptovirus F protein ("CV" [Strain BBR]); the second line represents Canine Parainfluenza Virus Type 2 ("CPV" [Strain T1]; see Ito *et al.*, *J. Gen. Virol.* 81:719-727, 2000); the third line represents Porcine Rubulavirus ("PR"; Klenk and Klenk, *Direct Submission to EMBL / GenBank Databases*, September 2000, GenBank Accession AJ278916); the fourth line represents Simian Virus 5 ("W3A" [Strain W3A]; Paterson *et al.*, *Proc. Natl. Acad. Sci. USA* 81:6706-6710, 1984); and the fifth (bottom) line represents Simian Virus 5 ("WR" [Strain WR]; Ito *et al.*, *J. Virol.* 71:9855-9858, 1997). Amino acids that are bold and underlined denote amino acids that differ from those in the analogous sequence of the *Cryptovirus* F protein, and the tallies in the right margin are the number of differences for each sequence block.

Fig. 11 demonstrates expression of *Cryptovirus* proteins. Fig. 11 is a photograph of an autoradiogram of gradient-purified [³⁵S]-methionine-labeled *Cryptovirus* virions produced in acutely-infected Vero cells after SDS-PAGE under reducing conditions. The approximate molecular weights of the proteins indicated on the right side of Fig. 12A were calculated by comparing their migrations to marker proteins of known molecular weight (Sigma Biochemicals). L = the largest nucleocapsid associated protein, the major component of the virion-associated RNA dependent RNA polymerase; HN = the hemagglutinin protein, one of the envelope-associated glycoproteins; F₀ = the uncleaved fusion protein, a second envelope-associated glycoprotein; NP = the nucleocapsid protein, the major structural protein associated with the nucleocapsid; F₁ = the larger fragment of the cleaved fusion protein; P = the nucleocapsid associated phosphoprotein; M = the virion-associated matrix or membrane protein; V = a minor RNA binding protein thought to be a component of the viral polymerase; F₂ = the smaller fragment of the cleaved fusion protein. Note: the SH protein (about 5 kD), a small envelope-associated protein, ran off the gel and is not shown. Fig. 12 shows photographs of autoradiograms of typical radioimmunoassay profiles (RIPs) obtained by the precipitation and SDS-PAGE separation of [³⁵S]-methionine-labeled virus-specific proteins using the cerebrospinal fluids (CSFs) of a patient diagnosed with subacute sclerosing panencephalitis (Fig. 12A) and the CSFs of six randomly-selected neurology/ neurosurgery patients who had CSF taken for microbiological screening (Fig. 12B). Fig. 12A shows the RIPs resulting from the precipitation of [³⁵S]-methionine-labeled CV-1_C cells acutely-infected with the Edmonston strain of measles virus (Lane MV), identically-labeled CV-1_C cells acutely-infected with the BBR strain of *Cryptovirus* (Lane CV) or a mixture of both (Lane B) by the CSF of an 11 year male SSPE patient. Lane V represents a SDS-PAGE profile of [³⁵S]-methionine-labeled gradient purified *Cryptovirus* virions (see also Fig. 11). Fig. 12B shows the RIPs resulting from the precipitation of proteins from [³⁵S]-methionine-labeled CV-1_C cells acutely-infected with the BBR strain of *Cryptovirus* by the CSFs of

six neurology/neurosurgery patients. The patient whose RIP profile appears in Lane 2 was an adult male who had presented with ataxia, confusion and memory loss and had not been given a specific diagnosis. The patient whose RIP profile appears in Lane 4 was an infant female who presented with hydrocephalus and intractable seizures and subsequently died in *status epilepticus*. None of the CSFs from the patients in Fig. 12B precipitated any of the envelope proteins of measles virus (data not shown). Fig. 12A is the same as Fig. 23 (described below), but is reduced to the same scale as Fig. 12B for the purpose of comparison.

Fig. 13 shows a higher resolution autoradiogram of the radioimmunoassay profiles (RIPs) of the *Cryptovirus*-specific proteins precipitated from [³⁵S]-methionine-labeled CV-1_C cells acutely-infected with the BBR strain of *Cryptovirus* by two CSF specimens (Fig. 13A) and a schematic showing the migration of the major corresponding structural proteins of gradient-purified virions of the BBR strain of *Cryptovirus* (Fig. 13B Lane CV) and the NIH 21005-WR strain of SV5 (Fig. 13B Lane SV5). The RIPs in Fig. 13A represent CSF precipitates from patients assessed as *Cryptovirus*-negative (Lane “-”; i.e. not containing *Cryptovirus*-specific antibodies) and *Cryptovirus*-positive (Lane “+”; i.e. containing *Cryptovirus*-specific antibodies). Fig. 13B is a schematic showing the near co-migration of the F₀ and HN proteins of *Cryptovirus* and their separate migration in Simian Virus 5 (see also SDS-PAGE profiles in Fig. 5).

Fig. 14 shows an ELISA of matched serum and CSF specimens from four seropositive neurology/neurosurgery patients using gradient-purified *Cryptovirus* virions as the target. Control sera were rabbit antisera generated against mock-infected CV-1_C cells (column 1; “-”) and hyperimmune rabbit antisera generated against gradient-purified *Cryptovirus* virions (column 2; “+”). FN = infant female diagnosed with hydrocephalus and intractable seizures; SG = adult female diagnosed with idiopathic intracranial hypertension; WK = male child diagnosed with acute viral meningitis; JK = adult male having an undetermined diagnosis. Serum dilutions began at 1:20 (in the top rows) and proceeded by 2-fold serial dilution to the bottom. CSF dilutions began at 1:2, at the top, before proceeding likewise. Serum specimens were aliquoted from left to right while CSF specimens were aliquoted from right to left. Note that although all of the patients had *Cryptovirus*-specific antibodies in their serum, only the patient with a seizure disorder (FN) had such antibodies in her CSF.

Fig. 15 is a photograph of RIP assays using three sets of matched serum (S) and CSF (C) samples from patients diagnosed with Alzheimer’s disease.

Fig. 16 is a photograph of an autoradiogram following an RIP analysis using four CSF specimens from patients diagnosed with chronic fatigue syndrome (CFS). Lanes 1-3 were assessed as “*Cryptovirus* positive”; Lane 4 assessed as “*Cryptovirus* negative”.

Fig. 17 is a photograph of an autoradiogram following an RIP analysis using CSF samples obtained as “Collection 1” (see hereinbelow). The positive CSF precipitate in Lane 2 was subsequently found to have been obtained from a 55 year-old adult male who presented with ataxia, memory loss, blackouts, seizures, diplopia, and headaches.

Fig. 18 is a photograph of an autoradiogram following a RIP analysis using CSF samples obtained as “Collection 2” (see hereinbelow).

Fig. 19 is a photograph of an autoradiogram following an RIP assay conducted with serum samples obtained from 5 MS patients (out of the 38 samples obtained).

Fig. 20 is a photograph of an autoradiogram following an RIP assay conducted with serum samples from an 25 additional MS patients (out of the 38 samples obtained).

Fig. 21 is a photograph of an autoradiogram following an RIP assay conducted with 16 CSF specimens obtained from 16 MS patients.

Fig. 22 is a photograph of an autoradiogram obtained following creation of RIP profiles of the *Cryptovirus* NP protein (p63) precipitated from [³⁵S]-methionine-labeled AV₃/SSPE cells by the sera of six Australian SSPE patients (Lanes 1-6) and six control sera (Lanes 7-12; sera from pediatric patients without antibodies to the *Cryptovirus* major envelope proteins (F₀ and HN).

Fig. 23 is a photograph of an autoradiogram of RIP profiles of measles virus-specific proteins or *Cryptovirus*-specific proteins precipitated from [³⁵S]-methionine-labeled measles virus-infected CV-1_C cells (Lane MV), *Cryptovirus*-infected CV-1_C cells (Lane CV) or a mixture of both (Lane B) by CSF from an 11 year old male diagnosed with SSPE. Lane V = gradient-purified *Cryptovirus* virions from [³⁵S]-methionine-labeled *Cryptovirus*-infected CV-1_C cells. L = the largest nucleocapsid associated protein, the major component of the virion-associated RNA dependent RNA polymerase; HN = the hemagglutinin protein, one of the envelope-associated glycoproteins; F₀ = the uncleaved fusion protein, a second envelope-associated glycoprotein; NP = the nucleocapsid protein, the major structural protein associated with the nucleocapsid; F₁ = the larger fragment of the cleaved fusion protein; P = the nucleocapsid associated phosphoprotein; M = the virion-associated matrix or membrane protein; V = a minor RNA binding protein thought to be a component of the viral

polymerase; F₂ = the smaller fragment of the cleaved fusion protein. Note: the SH protein (about 5 kD), a small envelope-associated protein, ran off the gel and is not shown.

Fig. 24 shows photomicrographs of *Cryptovirus*-infected neurons. Fig. 24A demonstrates *Cryptovirus*-specific immunofluorescence in a single neuron in the brain of a Colored mouse inoculated when two days old with *Cryptovirus Strain BBR* (sacrificed 2 months post inoculation after presenting with seizures). Fig. 24B demonstrates cytoplasmic immunofluorescence in a single neuron from the brain of a guinea pig presenting with a subacute encephalopathy after inoculation with the Niigata-1 strain of SSPE-derived cell-associated virus (detected by an indirect fluorescent antibody technique using SSPE serum)(Doi *et al.*, *Japan. J. Med. Sci. Biol.* 25:321-333, 1972).

Fig. 25 shows photomicrographs of differential immunogold-labeling of the intracellular nucleocapsids of *Cryptovirus* and measles virus in persistently-infected AV₃/SSPE/MV cells. Fig. 25A shows labeling of the about 15-nm to about 17-nm "smooth" and narrow nucleocapsids of *Cryptovirus* with 10-nm gold beads. The etching technique used results in a loss of resolution of the fine structure of the smooth nucleocapsids making the herringbone pattern somewhat difficult to see. Fig. 25B shows labeling of the 25-nm "fuzzy" and wide nucleocapsids of measles virus with 15-nm gold beads. Magnification is approximately 500,000X.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The novel virus described herein has been designated a *Cryptovirus* (CV) on the basis of its inapparent, or "cryptic," cytopathology in cultured human cells and its slow and "encrypted" pathogenesis in experimentally infected animals. Given the nucleotide sequence present in the virus, and its structural, biological, and pathogenetic characteristics, *Cryptovirus* fits best within the *Rubulavirus* genus of the family Paramyxoviridae (Fig. 1). More specifically, this virus most closely resembles the viruses known as Canine Parainfluenza Virus Type 2 (which is also known as Canine Parainfluenza Virus, CPI and CPV) and Porcine Rubulavirus (which is also known as La-Piedad-Michoacan-Mexico Virus and LPMV). (see Fig. 2). The relationships between *Cryptovirus* and these two viruses can be seen in the relationships between their sequences and their structural, biological, and pathogenetic characteristics (see Fig. 3). CPV shares more than 95% of its nucleotide sequence with *Cryptovirus*. The extent of Menangle virus nucleotide sequence homology with *Cryptovirus* is presently unknown as the sequence of the Menangle virus genome has not yet been published.

There is also an apparent relationship between *Cryptovirus* and: (1) simian parainfluenza virus type 5 (which is also known as simian virus 5 and SV5; see Fig. 4, Fig. 5, Fig. 9, and Fig. 10; here, there is a relationship between the sequences and structural and immunological properties of the

viruses but little or no biological or pathogenetic similarity); (2) human mumps virus (here, there are certain structural, biological, and pathogenetic relationships); and (3) human measles virus (here again, there are certain structural, biological and pathogenetic relationships). These relationships help to classify *Cryptovirus* and to establish its novelty.

5 In addition to having a role in idiopathic and cryptogenic forms of epilepsy or epileptiform disease, i.e., an illness, disorder, or condition having epileptiform symptomology (e.g., CFS, MS, SSPE), *Cryptovirus* is also implicated in a spectrum of idiopathic disorders of the central nervous system (CNS) that present with compulsive or iterative physical, behavioral, or psychological symptoms. The manifestation of symptoms of these disorders as a consequence of *Cryptovirus*
10 infection is exclusively subacute or slow in nature taking weeks, months, or even years to develop. The spectrum of physical symptoms that have been presented by human patients that have been infected with *Cryptovirus* includes febrile response, ophthalmological disorders (photosensitivity, blurred vision, nystagmus, loss of vision) parathesias, paralysis, tremor, myoclonus, and *grand mal* and *petit mal* (absence) seizures. The spectrum of behavioral or psychological symptoms that have
15 been presented by patients includes repetitive movements and compulsive behaviors (characteristic of obsessive compulsive disorder), sleep disturbances, memory loss, and dysphoria, anorexia nervosa, autism, mental retardation, affective disorder, dysthymia (clinical depression), schizophrenia, and bipolar disorder.

While not essential features of the present invention, the portal of entry for *Cryptovirus*
20 infection can be the oral mucosa of the throat (i.e., the tracheo-bronchial epithelium), and the virus' incubation period can be of subacute duration (i.e., many days to weeks). Newly infected individuals can develop a febrile pharyngitis and lymphadenopathy of prolonged duration, not unlike infectious mononucleosis. Alternatively, it is thought that the portal of entry for the virus can also be transplacental, so that a mother carrying the virus can transmit it to her child *in utero*, and the child
25 can subsequently develop a neurological, neurodegenerative, and/or neuropsychiatric disease or other developmental disorder (e.g., autism, cerebral palsy, hydrocephalus, birth defect, partial paralysis). Such a child is frequently diagnosed or labelled as "retarded". Indeed, the incidence of epilepsy and seizures are dramatically higher in the severely "mentally-retarded" (as much as 50-fold higher than the general population).

30 The nucleotide sequence of the human *Cryptovirus* genome (15,246 contiguous nucleotides), in FASTA format (i.e., mRNA sense; 5' to 3'), is shown in Fig. 9 (SEQ ID NO:1). The actual genome of the virus is negative-stranded (antisense to mRNA), having a nucleotide sequence entirely complementary to (SEQ ID NO:1).

Accordingly, the present invention encompasses an isolated human negative-stranded RNA virus that, in FASTA format (*i.e.* in positive-stranded, mRNA-sense, the reverse and complementary sequence to the actual genome), has the sequence of SEQ ID NO:1. In Fig. 9, nucleotides that vary from those of the W3A strain of Simian Virus 5 are highlighted and the number of variations in each line is tallied in the right margin. The FASTA formatted sequence of human Cryptovirus *Strain BBR* was compared to Simian Virus 5 *Strain W3A* (SEQ ID NO:2; see Fig. 9). Comparisons between various Rubulavirus F Protein amino acid sequences have also been made (Fig 10).

A *Cryptovirus* "particle" is an entire *Cryptovirus* virion, as well as encompassing particles which are intermediates in virion formation (*e.g.*, nucleocapsids), or otherwise partial. *Cryptovirus* particles generally have one or more *Cryptovirus* proteins associated with the *Cryptovirus*-specific nucleic acid they contain. A preferred *Cryptovirus* particle or virion is *Cryptovirus* Strain BBR, which is deposited as ATCC Accession No. PTA-4745.

The present invention also relates to a composition of matter comprising the inventive *Cryptovirus* particle and a carrier.

As used herein a "carrier" can be an organic or an inorganic carrier or excipient, such as water or an aqueous solution, or an emulsion such as an oil/water or water/oil emulsion, and various types of wetting agents. The active ingredient, such as the inventive viral particle, nucleic acid construct, protein, or antibody, can optionally be compounded in a composition formulated, for example, with non-toxic, physiologically acceptable carriers for infusions, tablets, pellets, capsules, solutions, emulsions, suspensions, or in any other formulation suitable for its intended *in vitro* or *in vivo* use. Such carriers also include glucose, lactose, gum acacia, gelatin, mannitol, starch paste, magnesium trisilicate, talc, corn starch, keratin, colloidal silica, potato starch, urea, medium chain length triglycerides, dextrans, normal saline, phosphate buffered saline and other carriers suitable for use in manufacturing preparations, in solid, semisolid, or liquid form. In addition auxiliary, stabilizing, thickening and coloring agents and perfumes can be used as appropriate. Other examples of suitable carriers are described hereinbelow, but any suitable carrier known in the art is intended.

In the inventive method of isolating the *Cryptovirus* virion, as described hereinabove, the PBMNCs that have been obtained from a human having a *Cryptovirus* infection are cultured in an artificial aqueous medium that includes, importantly, an agent that increases cellular guanylyl cyclase activity.

For purposes of the present invention, the artificial aqueous medium is made by adding the agent that increases cellular guanylyl cyclase activity to a known minimal cell culture medium, such as IMEMZO, MEM, HYQPF Vero (Hyclone), or RPMI, buffered (*e.g.*, with HEPES) to pH 6.8-7.8

and most preferably to pH 6.8-7.2. The agent operates to permit and facilitate the isolation and/or propagation of Cryptovirus in accordance with the invention.

Optionally, fetal calf serum (about 2% v/v to about 10% v/v) is added to the medium. Antibiotics, such as penicillin or streptomycin, in conventional amounts, can also be added to the medium.

It is not essential to the present invention that cellular guanylyl cyclase activity actually be measured. In addition, the present invention is dependent neither upon any particular mechanism by which the agent may actually operate to increase cellular guanylyl cyclase activity (or not), nor upon any mechanism by which the agent operates to permit and/or facilitate the isolation and/or propagation of Cryptovirus in accordance with the invention.

Useful examples of the agent that increases cellular guanylyl cyclase activity include most preferably guanosine 3',5'-cyclic monophosphate ("cyclic GMP") (free acid, or preferably, a pharmaceutically acceptable salt thereof, such as a sodium, potassium, magnesium, calcium, or ammonium salt, or the like), insulin (preferably human insulin), zinc dication (preferably provided in a chloride, sulfate, carbonate, bicarbonate, nitrate, acetate, or other pharmaceutically acceptable salt thereof), or a combination of any or all of these.

Preferably, the cyclic GMP is used in a concentration of about 0.05 to about 5 mM in the artificial aqueous medium. More preferably, the cyclic GMP concentration in the medium is about 0.5 to about 2.5 mM, and most preferably about 0.75 mM to about 1.25 mM. A concentration above about 5 mM cyclic GMP is not optimally conducive to cultivating, propagating, or isolating *Cryptovirus*.

A preferred concentration range for insulin in the artificial aqueous medium is about 1 to about 10 mg/L, more preferably about 2 to about 6 mg/L, and most preferably about 3 to about 5 mg/L.

A preferred concentration range for zinc dication in the artificial aqueous medium is equivalent to about 0.05 to about 0.25 mg/L of $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$, more preferably equivalent to about 0.10 to about 0.20 mg/L of $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$, or most preferably equivalent to about 0.13 to about 0.15 mg/L of $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$.

Alternatively, in some embodiments, the agent that increases cellular guanylyl cyclase activity is nitric oxide or a nitric oxide donor. Nitric oxide gas is fully permeable across biological membranes. Inhalable nitric oxide gas can be administered to a mammalian subject by, for example, a mask in a controlled gas mixture as is known in the art. (E.g., Kieler-Jensen, N. *et al.*, *Inhaled nitric oxide in the evaluation of heart transplant candidates with elevated pulmonary vascular*

resistance, *J Heart Lung Transplant.* 13(3):366-75 [1994]; Rajek, A. *et al.*, *Inhaled nitric oxide reduces pulmonary vascular resistance more than prostaglandin E(1) during heart transplantation*, *Anesth Analg.* 90(3):523-30 [2000]; Solina, A. *et al.*, *A comparison of inhaled nitric oxide and milrinone for the treatment of pulmonary hypertension in adult cardiac surgery patients*, *J Cardiothorac Vasc. Anesth.* 14(1):12-17 [2000]; Fullerton, D.A. *et al.*, *Effective control of pulmonary vascular resistance with inhaled nitric oxide after cardiac operation*, *J Thorac Cardiovasc Surg* 111(4):753-62, discussion 762-3 [1996]). The concentration in the gas mixture of nitric oxide (NO) is preferably about 1 to 100 ppm NO, more preferably about 4 to 80 ppm NO, and most preferably about 20 to 40 ppm NO. The gas mixture also contains appropriate concentrations of oxygen and nitrogen and/or other inert gases, such as carbon dioxide, helium or argon.

Nitric oxide donors are compounds that produce NO-related physiological activity when applied to biological systems. Thus, NO-donors can mimic an endogenous NO-related response or substitute for an endogenous NO deficiency. The skilled artisan is aware that in biological systems there are at least three redox states of NO that can be released by various NO donors (NO^+ , NO^0 , or NO^-), all of which are encompassed by the terms "nitric oxide" or "NO" for purposes of the present invention. The redox state of NO makes a substantial difference to the NO donors reactivity towards other biomolecules, the profile of by-products, and the bioresponse (Feelisch, M., *The use of nitric oxide donors in pharmacological studies*, *Naunyn-Schmiedeberg's Arch. Pharmacol.* 358:113-22 [1998]). Some classes of NO donors require enzymatic catalysis, while others produce NO non-enzymatically; some NO donors require reduction, for example by thiols, and some oxidation, in order to release NO.

Preferred examples of nitric oxide donors include organic nitrate compounds, which are nitric acid esters of mono- and polyhydric alcohols. Typically, these have low water solubility, and stock solutions are prepared in ethanol or dimethyl sulfoxide (DMSO). Examples are glyceryl trinitrate (GTN) or nitroglycerin (NTG), pentaerythrityl tetranitrate (PETN), isosorbide dinitrate (ISDN), and isosorbide 5-mononitrate (IS-5-N). Administration of organic nitrates can be done intravenously, intraperitoneally, intramuscularly, transdermally, or in the case of PETN, ISDN, NTG, and IS-5-N, orally.

Other preferred examples of nitric oxide donors are S-nitrosothiol compounds, including S-nitroso-N-acetyl-D,L-penicillamine (SNAP), S-nitrosoglutathione (SNOG), S-nitrosoalbumin, S-nitrosocysteine. S-nitrosothiol compounds are particularly light-sensitive, but stock solutions kept on ice and in the dark are stable for several hours, and chelators such as EDTA can be added to stock

solutions to enhance stability. Administration is preferably by an intravenous or intra-arterial delivery route.

Other preferred examples of nitric oxide donors include sydnonimine compounds, such as molsidomine (N-ethoxycarbonyl-3-morpholino-sydnonimine), linsidomine (SIN-1; 3-morpholino-sydnonimine or 3-morpholinylsydnoneimine or 5-amino-3-morpholinyl-1,2,3-oxadiazolium, e.g., chloride salt), and pirsidomine (CAS 936). Stock solutions are typically prepared in DMSO or DMF, and are stable at 4°C to room temperature, if protected from light. Linsidomine is highly water soluble and stable in acidic solution in deoxygenated distilled water, adjusted to about pH 5, for an entire day. At physiological pH, SIN-1 undergoes rapid non-enzymatic hydrolysis to the open ring form SIN-1A, also a preferred nitric oxide donor, which is stable at pH 7.4 in the dark. Administration is preferably by an intravenous or intra-arterial delivery route.

Also useful as nitric oxide donors are iron nitrosyl compounds, such as sodium nitroprusside (SNP; sodium pentacyanonitrosyl ferrate(II)). Aqueous stock solutions are preferably made freshly in deoxygenated water before use and kept in the dark; stability of stock solutions is enhanced at pH 3-5. Inclusion in the delivery buffer of a physiologically compatible thiol, such as glutathione, can enhance release of NO. SNP is administered by intravenous infusion, and the skilled practitioner is aware that long-term use is precluded by the release of five equivalents of toxic CN-per mole SNP as NO is released.

A most preferred nitric oxide donor is chosen from among the so-called NONOate compounds. The NONOates are adducts of NO with nucleophilic residues (X⁻), such as an amine or sulfite group, in which an NO dimer is bound to the nucleophilic residue via a nitrogen atom to form a functional group of the structure X⁻-N(O)NO⁺. The NONOates typically release NO at predictable rates largely unaffected by biological reactants, and NO release is thought to be by acid-catalyzed dissociation with the regeneration of X⁻ and NO.

NONOates include most preferably diethylamine-NONOate (DEA/NO; N-Ethylethanamine:1,1-Diethyl-2-hydroxy-2-nitrosohydrazine (1:1) or 1-[N,N-diethylamino]diazene-1-ium-1,2-diolate). Other preferred NONOates include diethylene triamine-NONOate (DETA/NO; 2,2'-Hydroxynitrosohydrazino]bis-ethanamine), spermine-NONOate (SPER/NO; N-(4-[-1-(3-Aminopropyl)-2-hydroxy-2-nitrosohydrazino] butyl)-1,3-propanediamine), propylamino-propylamine-NONOate (PAPA/NO; 3-(2-Hydroxy-2-nitroso-1-propylhydrazino)-1-propanamine or (Z)-1-[N-(3-aminopropyl)-N-

(n-propyl)amino]diazene-1-ium-1,2-diolate), MAHMA-NONOate (MAHMA/NO; 6-(2-Hydroxy-1-methyl-2-nitrosohydrazino)-N-methyl-1-hexanamine), dipropyleneetriamine-NONOate (DPTA/NO; 3,3'-(Hydroxynitrosohydrazino)bis-1-propanamine), PIPERAZI/NO, proli-NONOate (PROLI/NO; 1-([2-carboxylato]pyrrolidin-1-yl)diazene-1-ium-1,2-diolate-methanol, e.g., disodium salt), SULFO-NONOate (SULFO/NO; hydroxydiazenesulfonic acid 1-oxide, e.g., diammonium salt), the sulfite NONOate (SULFI/NO), and Angelis salt (OXI/NO).

Almost all NONOate compounds are highly soluble in water, and aqueous stock solutions are prepared in cold deoxygenated 1 to 10 mM NaOH (preferably about pH 12) just prior to use. Alkaline stock solutions are stable for several hours if kept on ice in the dark. The characteristic UV absorbance of NONOates can be used for spectrophotometric quantification of NONOate in aqueous solutions. NONOates are preferably administered intravenously or intra-arterially.

Nitric oxide donors have different potencies (Ferraro, R. *et al.*, *Comparative effects of several nitric oxide donors on intracellular cyclic GMP levels in bovine chromaffin cells: correlation with nitric oxide production*, Br. J. Pharmacol. 127(3):779-87 [1999]). For example, DEA/NO is among the most potent nitric oxide donors, with a half-life of about 2 to 4 minutes; less potent are PAPA/NO ($t_{1/2}$ about 15 minutes), SPER/NO ($t_{1/2}$ about 34-40 minutes); even less potent are DETA/NO ($t_{1/2}$ about 20 hours) and SNAP ($t_{1/2}$ about 33 to 41 hours, although this can be shortened in the presence of a physiological reductant such as glutathione). SNP is also a potent NO donor. (See, Ferrero *et al.* [1999]; Salom, J.B. *et al.*, *Relaxant effects of sodium nitroprusside and NONOates in rabbit basilar artery*, Pharmacol. 57(2):79-87 [1998]; Salom, J.B. *et al.*, *Comparative relaxant effects of the NO donors sodium nitroprusside, DEA/NO and SPER/NO in rabbit carotid arteries*, Gen. Pharmacol. 32(1):75-79 [1999]; Salom, J.B. *et al.*, *Relaxant effects of sodium nitroprusside and NONOates in goat middle cerebral artery: delayed impairment by global ischemia-reperfusion*, Nitric Oxide 3(1):85-93 [1999]; Kimura, M. *et al.*, *Responses of human basilar and other isolated arteries to novel nitric oxide donors*, J. Cardiovasc. Pharmacol. 32(5):695-701 [1998]). Consequently, effective concentrations or doses of NONOates or other NO donors will vary, but can be determined by routine screening.

Stock solutions of NO donors are preferably made up freshly before use (at the appropriate pH for each particular NO donor), chilled on ice, and protected from light (e.g., by the use of darkened glass vials wrapped in aluminum foil), although organic nitrates can be stored for months to years if the vial is properly sealed. Preferably, immediately before administration to the subject, final dilutions are prepared in pharmaceutically acceptable buffer and the final pH of the NO

donor-containing buffer is checked for physiological suitability, especially when strongly acidic (e.g., hydrochloride salts) or alkaline (e.g., NONOates) stock solutions are used.

The product of NO exposure time and NO concentration largely determines the quality and magnitude of the biological response to exogenously supplied NO. Short-lived NO donors, such as DEA/NO, are most preferably administered by continuous infusion rather than by bolus to avoid delivering only a short burst of NO.

In accordance with the invention, the artificial aqueous medium preferably, but not necessarily, further includes glutamine at a preferred concentration of about 0.5 to about 5 mM concentration. A more preferred concentration of glutamine in the medium is about 1 to about 3 mM.

In the inventive methods of isolating a *Cryptovirus* virion and of producing a mammalian cell line nonproductively infected with *Cryptovirus*, the PBMNCs are co-cultured with mammalian amnion cells in the artificial aqueous medium, as described above.

Examples of useful mammalian cells include, but are not limited to, rodent, lagomorph, primate, ovine, bovine, canine, feline or porcine cells.

In accordance with the present invention, one preferred embodiment is a primate cell, i.e., a cell originating from a primate source. A primate is a member of the mammalian order Primates, including lemurs, tarsiers, monkeys (e.g., African Green Monkeys, colobus monkeys, and baboons), apes (e.g., chimpanzees, gorillas, orangutans, and gibbons), and humans.

An amnion cell is a cultured cell originally derived from an amniotic membrane or amniotic sac.

A preferred primate amnion cell is a human amnion cell, e.g., AV₃.

An example of the inventive cell nonproductively infected by the method is AV₃/SSPE, which is deposited as ATCC Accession No. PTA4746.

In these inventive methods, passaging of a co-culture of PBMNCs and mammalian amnion cells is done one or more times. Passaging of cultured cells into fresh culture medium (culture medium as described above), is typically done about twice per week. Preferably, at least about two to about 12 passages are done in accordance with the methods. Typically after about eight to twelve passages of the co-culture, virtually all mammalian amnion cells are nonproductively infected with *Cryptovirus*. Generally, within about two to about three passages, the PBMNCs have disappeared from the culture, leaving the mammalian amnion cells.

In accordance with the present invention, a mammalian epithelial cell is a cultured cell originally derived from a mammalian epithelial tissue. In one preferred embodiment, the mammalian epithelial cell is a rodent epithelial cell, such as baby hamster kidney (BHK) cells. In another preferred embodiment, the mammalian epithelial cell is a simian epithelial cell, for example a Vero or a CV-1 cell. Most preferably the CV-1 cell is subline CV-1_c.

In addition to the sequence information provided herein to identify the inventive *Cryptovirus* particle, the inventive *Cryptovirus* and its viral subcomponents can be, and have been, characterized by numerous virological, biochemical, and molecular techniques, including the following, by way of example:

5 *Plaque Titration Assay:* Formation of macroscopically visible plaques on monolayers of mammalian epithelial cells (e.g., BHK, Vero or CV-1c) can be used to quantitate preparations of infectious *Cryptovirus* (Robbins *et al.*, *J. Infect. Disease* 143:396-403, 1981).

10 *Neutralization Titration Assay:* Plaque formation can be inhibited by serial dilutions of clinical serum specimens and *Cryptovirus*-specific antisera generated in rabbits (*see e.g.*, Robbins *et al.*, *J. Infect. Disease* 143:396-403, 1981). Neutralization titration assays are routinely used in medical virological research to demonstrate that a given patient has neutralizing antibodies to a particular virus. A neutralization assay can be used diagnostically for the presence or absence of neutralizing antibody to *Cryptovirus*. In a typical example of a neutralization assay, serial dilutions of a biological material, such as a sample of serum or CSF to be tested, are typically incubated for
15 about one hour at 4°C with sufficient infectious virus to yield a net plating concentration of between about 100-200 plaque forming units of the virus per 0.2 mL of final diluent (including the diluted serum or CSF). After incubation, about 0.2 mL of the diluted virus-serum (or CSF) mixtures are then typically plated onto monolayers of susceptible cells (*e.g.* Vero or CV-1) and the cells are incubated at 37°C in a partial CO₂ atmosphere (*e.g.*, 5% v/v) (typically, with redistribution of the inoculum
20 every 15 minutes). At the end of the incubation period, inoculated monolayers are typically overlaid with sufficient volumes of a 2% (w/v) solution of carboxymethylcellulose in an artificial aqueous cell culture medium (*e.g.*, IMEMZO medium, buffered at pH between about 6.8 and about 7.4, typically containing fetal calf serum, and a suitable quantity of antibiotic, such as about 200 Units penicillin / mL and/or 100 µg streptomycin / mL) to last 10-12 days (*i.e.*, enough volume so that the
25 monolayers won't dry out). Optimally, the plates must not be moved during the incubation period. After 10-12 days, the overlay is aspirated and the cells are fixed with formalin fixative and stained with a protein stain (*e.g.*, Giemsa). The number of plaques formed on each plate is then enumerated and the PRD₅₀ calculated (PRD₅₀ = the Plaque Reduction Dilution; the dilution of serum or CSF at

which a 50% reduction in the number of plaques formed on controls [tubes containing virus and saline only] is observed).

Density Gradient Purification: Virions and intracellular nucleocapsids from productively- (e.g., Vero and CV-1_C) and nonproductively-infected (e.g., AV₃/SSPE) cells can be purified on sucrose-potassium tartrate gradients (virions) and CsCl gradients (nucleocapsids) (see Robbins *et al.*, *J. Infect. Disease* 143:396-403, 1981; Rapp and Robbins, *Intervirology* 16:160-167, 1981; Robbins and Rapp, *Arch. Virol.* 71:85-91, 1982; and Robbins and Abbott-Smith, *J. Virol. Meth.* 11:253-257, 1985).

Electron Microscopy: Electron microscopy, such as transmission or scanning electron microscopy are useful for examining the inventive *Cryptovirus* virion. When examined by electron microscopy, the *Cryptovirus* has been shown to have a morphology and ultrastructure consistent with other members of the Paramyxoviridae (*i.e.*, enveloped pleomorphic virions, about 100 nm to about 120 nm in diameter, containing helical nucleocapsids). Intracellular inclusions of the virus in thin sections of productively- (e.g., Vero and CV-1_C) and nonproductively-infected cells (e.g., AV₃/SSPE) have also been shown to be comprised of aggregates of filamentous structures with dimensions similar to the nucleocapsids of other members of the Paramyxoviridae (*i.e.*, helical herringbone-like structures, about 15 to about 17 nm in diameter) (see Robbins *et al.*, *J. Infect. Disease* 143:396-403, 1981 and Robbins and Rapp, *Arch. Virol.* 71:85-91, 1982).

Radioimmunoprecipitation (RIP) Assay: Extensive data has been generated by the comparative analysis of *Cryptovirus*-specific immunoprecipitates of [³⁵S]-methionine-labeled uninfected, nonproductively- and productively-infected mammalian cells by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE; see below).

SDS-PAGE: Purified virions and cytoplasmic nucleocapsids of the virus have been analyzed by SDS-PAGE under reducing and non-reducing conditions (see Fig. 11 and Fig. 12), an autoradiogram of gradient-purified [³⁵S]-methionine-labeled *Cryptovirus* virions produced in acutely-infected Vero cells after SDS-PAGE under reducing conditions. The approximate molecular weights of the proteins indicated on the right side of Fig. 11 were calculated by comparing their migrations to marker proteins of known molecular weights (Sigma Chemical Co., St. Louis, MO). The SH protein, a small envelope-associated protein having a MW of about 5 kD, is not shown in Fig. 11, because it has run off the gel.

Immuno-ultrastructural Analysis (Immunogold Analysis). The intracellular nucleocapsids of nonproductively and productively-infected mammalian cells are typically localized under the electron

microscope using *Cryptovirus*-specific or *Cryptovirus* nucleocapsid-specific antibodies, for example hyperimmune rabbit antibodies, and an indirect immunogold labeling technique (discussed hereinbelow).

The preceding are merely illustrative, and not an exhaustive list, of the known techniques typically useful for characterizing the isolated *Cryptovirus* virion of the present invention. Additional conventional techniques, or virological techniques yet to be discovered, can also be employed to further characterize the inventive *Cryptovirus* virion.

Additional characteristics of *Cryptovirus* include the following:

Latency and Persistence. *Cryptovirus* latently and persistently infects human peripheral blood mononuclear cells (PBMNCs). No other member of the Paramyxoviridae has been shown to do this. The evidence for infection includes: (1) detection of *Cryptovirus*-specific proteins by an indirect immunofluorescent antibody technique in PBMNCs following *in vitro* cultivation and induction with mitogens and/or cyclic GMP, (2) recovery of the virus from PBMNCs by serial cocultivation with mammalian cells (see Robbins *et al.*, *J. Infect. Dis.* 143:396-403, 1981) and (3) the ability to repeatedly recover the virus from PBMNCs drawn from an SSPE patient over a period of 18 months.

Defective Fusion Activity. Cell fusion, which is a hallmark of the Paramyxoviridae, is either defective or extremely limited in experimental *Cryptovirus* infections *in vitro* (*i.e.*, in dysgenic nonproductive infections of human amnion cells (AV₃) and productive infections of monkey kidney cells (e.g., Vero and CV-1c; see Robbins *et al.*, *J. Infect. Dis.* 143:396-403, 1981, and Robbins and Rapp, *Arch. Virol.* 71:85-91, 1982).

Restricted Expression in Latently-Infected Cells. *Cryptovirus*-specific protein expression is dysgenic in experimental nonproductive latent infections of mammalian amnion cells (e.g., human AV₃ cells). This restriction involves severely decreased expression, or non-expression, of the virus-encoded envelope proteins (F, HN and SH) (see Robbins and Rapp, *Arch. Virol.* 71:85-91, 1982).

B Cell Lymphotropism. *Cryptovirus* demonstrates a tropism for B cells, and can be harbored by such cells *in situ*. This has been demonstrated by successfully infecting EBV-transformed B cell lines from human donors with the virus (*i.e.*, by detecting the progressive formation of *Cryptovirus*-specific inclusion bodies in the cytoplasm of experimentally-infected EBV-transformed B cell lines by *Cryptovirus*-specific immunofluorescence). In contrast, *Cryptovirus*-specific proteins could not be detected in an experimentally-infected human T cell line, CCRF-CEM. Accordingly, *Cryptovirus* can reside in B cells in infected individuals.

Neurotropism. *Cryptovirus* also demonstrates a clear tropism for neurons in mice following intracerebral inoculation of neonatal animals (as detected by *Cryptovirus*-specific immunofluorescence). It is less clear whether other nervous system tissues are infected. While neurotropism, itself, may not be unique to *Cryptovirus* when compared and contrasted to other human members of the Paramyxoviridae (e.g., Measles Virus, Mumps Virus), some of the neuropathological consequences of *Cryptovirus* infection of CNS tissues (including neurons) appear to be (see below).

Hind Limb Atrophy and Paralysis. Hind limb paralysis and atrophy were seen in approximately 33% of Quackenbush mice intracerebrally-inoculated with *Cryptovirus* as newborns. In addition, hind limb atrophy and paralysis was observed in some of the offspring of adult female Quackenbush mice that had been inoculated with *Cryptovirus* as newborns but did not develop any overt symptomology. The frequency of the symptoms appearing in the latter situation was difficult to assess because the mothers tended to cannibalize the newborn animals that were born with, or subsequently developed, such characteristics.

Subacute/Slow Encephalopathic and Epileptogenic Potential. Approximately 30% of neonatal Colored mice that were inoculated with infectious *Cryptovirus* preparations went on to develop subacute/slow encephalopathic and/or epileptiform illness (the specific symptoms displayed by such animals are described below). The number of animals that actually developed encephalopathic and/or epileptiform illness was likely higher than 30%, because a number of previously asymptomatic animals were found dead in their cages in clonic postures (a symptom associated with death following or during intractable seizure). On at least two occasions, this occurred in animals after they had suffered from recurrent seizures the day before. The animals that developed such illnesses were predominantly male (approximately 2:1, male: female).

Slow Psychopathogenic Potential. Of the Colored mice that survived intracerebral inoculation with infectious *Cryptovirus* preparations as newborns and did not develop epileptiform illness as adolescent or young adult animals, approximately 30% went on to develop profound physical and behavioral abnormalities as adults. The abnormalities displayed by these animals are described hereinbelow. Sudden death was not seen in this group of animals. The animals which developed such symptoms were predominantly female (approximately 3:1, female: male).

The preceding are merely illustrative, and are not an exhaustive list, of some of the observable properties of the inventive *Cryptovirus* particle.

The present invention also relates to isolated nucleic acids and isolated proteins that are "*Cryptovirus*-specific," i.e., unique to *Cryptovirus*.

A *Cryptovirus*-specific nucleic acid segment or protein is determined by sequence similarity or homology to known sequences of bases or amino acids, respectively, for example other rubulavirus nucleic acid or protein sequences. Base and amino acid sequence homology is determined by conducting a sequence similarity search of a genomics/proteomics data base, such as the GenBank database of the National Center for Biotechnology Information (NCBI; www.ncbi.nlm.nih.gov/BLAST/), using a computerized algorithm, such as PowerBLAST, QBLAST, PSI-BLAST, PHI-BLAST, gapped or ungapped BLAST, or the "Align" program through the Baylor College of Medicine server (www.hgsc.bcm.tmc.edu/seq_data). (E.g., Altschul, S.F., *et al.*, *Gapped BLAST and PSI-BLAST: a new generation of protein database search programs*, Nucleic Acids Res. 25(17):3389-402 [1997]; Zhang, J., & Madden, T.L., *PowerBLAST: a new network BLAST application for interactive or automated sequence analysis and annotation*, Genome Res. 7(6):649-56 [1997]; Madden, T.L., *et al.*, *Applications of network BLAST server*, Methods Enzymol. 266:131-41 [1996]; Altschul, S.F., *et al.*, *Basic local alignment search tool*, J. Mol. Biol. 215(3):403-10 [1990]).

For purposes of the present invention the term "isolated" encompasses "purified". Thus, an isolated nucleic acid, protein, viral particle, or antibody that is further purified to a greater level of homogeneity, is also "isolated."

For purposes of the present invention the term "nucleic acid" includes a polynucleotide, of any length, either polymeric ribonucleotides (RNA) or polymeric deoxyribonucleotides (DNA), such as cDNA.

The term "isolated nucleic acid" refers to a *Cryptovirus* genomic RNA which is essentially free, i.e., contains less than about 50%, preferably less than about 70%, and even more preferably less than about 90% of the polypeptides with which the *Cryptovirus* genome is naturally associated. Alternatively, an "isolated" nucleic acid of the present invention is a *Cryptovirus*-specific "recombinant polynucleotide", which as used herein intends a polynucleotide of genomic RNA, sense RNA (i.e., mRNA sense), cDNA, semisynthetic, or synthetic origin, which, by virtue of its origin or manipulation: (1) is not associated with all or a portion of a polynucleotide with which it is associated in nature, (2) is linked to a polynucleotide other than that to which it is linked in nature, or (3) does not occur in nature. The inventive nucleic acid can be in a sense or antisense orientation.

As used herein, the "sense strand" of a nucleic acid contains the sequence that has sequence homology to that of mRNA. The "anti-sense strand" contains a sequence which is complementary to that of the "sense strand." Inventive nucleic acids also include double- and single-stranded DNA and RNA.

Techniques for purifying viral polynucleotides from viral particles are known in the art, and include for example, disruption of the particle with a chaotropic agent, differential extraction and separation of the polynucleotide(s) and polypeptides by ion-exchange chromatography, affinity chromatography, and sedimentation according to density.

5 Inventive nucleic acids also encompass polynucleotides with known types of modifications, for example, labels, methylation, "caps", substitution of one or more of the naturally occurring nucleotides with a nucleotide analog, internucleotide modifications such as, for example, polynucleotides with uncharged linkages (e.g., methyl phosphonates, phosphotriesters, phosphoamidates, carbamates, etc.) and with charged linkages (e.g., phosphorothioates, 10 phosphorodithioates, etc.), those containing pendant moieties, such as, for example proteins (including for e.g., nucleases, toxins, antibodies, signal peptides, poly-L-lysine, etc.), those with intercalators (e.g., acridine, psoralen, etc.), those containing chelators (e.g., metals, radioactive metals, boron, oxidative metals, etc.), those containing alkylators, those with modified linkages (e.g., alpha anomeric nucleic acids, etc.), as well as unmodified forms of the polynucleotide.

15 A "nucleic acid segment" is a polynucleotide subportion of a larger nucleic acid.

A nucleotide sequence complementary to an inventive *Cryptovirus*-specific nucleotide sequence, as used herein, is one binding specifically or hybridizing with a *Cryptovirus*-specific nucleotide base sequence. The phrase "binding specifically" or "hybridizing" encompasses the ability of a polynucleotide sequence to recognize a complementary base sequence and to form double-helical 20 segments therewith via the formation of hydrogen bonds between the complementary base pairs. Thus, a complementary sequence includes, for example, an antisense sequence with respect to a sense sequence or coding sequence. As known to those of skill in the art, the stability of hybrids is reflected in the melting temperature (T_m) of the hybrids. In general, the stability of a hybrid is a function of sodium ion concentration and temperature. Typically, the hybridization reaction is performed under 25 conditions of relatively low stringency, followed by washes of varying, but higher, stringency. Reference to hybridization stringency relates to such washing conditions.

As used herein, the phrase "moderately stringent hybridization" refers to conditions that permit target-RNA or DNA to bind a complementary nucleic acid that has at least about 95%, about 96%, about 97%, about 98%, about 99%, or about 100% identity to the target RNA segment or DNA 30 segment. Preferably, moderately stringent conditions are conditions approximately equivalent in stringency to hybridization in 50% formamide, 5 x Denhart's solution, 5 x SSPE, 0.2% SDS at 42°C, followed by washing in 0.2 x SSPE, 0.2% SDS, at 65°C.

The phrase "high stringency hybridization" refers to conditions that permit hybridization of only those nucleic acid sequences that form stable hybrids in 0.018 M NaCl at 65°C (i.e., if a hybrid is not stable in 0.018 M NaCl at 65°C, it will not be stable under high stringency conditions, as contemplated herein). High stringency conditions can be provided, for example, by hybridization in 50% formamide, 5X Denhart's solution, 5X SSPE, 0.2% SDS at 42°C, followed by washing in 0.1 x SSPE, and 0.1% SDS at 65°C.

The phrase "low stringency hybridization" typically refers to conditions equivalent to hybridization in 10% formamide, 5X Denhart's solution, 6 x SSPE, 0.2% SDS at 42°C, followed by washing in 1 x SSPE, 0.2% SDS, at 50°C. Denhart's solution and SSPE (see, e.g., Sambrook et al., Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press [1989]) are well known to those of skill in the art, as are other suitable hybridization buffers.

The inventive nucleic acids include a *Cryptovirus*-specific nucleic acid fragment at least about five contiguous nucleotides long, and up to 15245 contiguous nucleotides long, of SEQ ID NO:1, or a complementary sequence.

Thus, useful fragments include nucleic acid segments consisting of an open reading frame of the *Cryptovirus* genome or a complementary sequence. An "open reading frame" (ORF) is a region of a polynucleotide sequence which encodes a polypeptide; this region may represent a portion of a coding sequence or an entire coding sequence.

A "coding sequence" is a polynucleotide sequence which is transcribed into mRNA and/or translated into a polypeptide when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a translation start codon at the 5'-terminus and a translation stop codon at the 3'-terminus. A coding sequence can include, but is not limited to, mRNA, cDNA, and recombinant polynucleotide sequences.

Useful examples of the fragment include nucleic acid segments that encode *Cryptovirus* proteins, such as (i) contiguous nucleotide positions 152-1678 of (SEQ ID NO:1)(also designated [SEQ ID NO:3]); (ii) contiguous nucleotide positions 1850-2515 of (SEQ ID NO:1)(also designated [SEQ ID NO:5]); (iii) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1)(also designated [SEQ ID NO:33]); (iv) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1) combined with a further insertion of two guanine (G) residues between nucleotide position 2339 of (SEQ ID NO:1) and nucleotide position 2340 of (SEQ ID NO:1)(the combined sequence including the "GG" insertion being designated [SEQ ID NO:7]); (v) contiguous nucleotide positions 3141-4271 of (SEQ ID NO:1)(also designated [SEQ ID NO:9]); (vi) contiguous nucleotide positions 4530-6182 of (SEQ ID NO:1)(also designated [SEQ ID NO:11]); (vii) contiguous nucleotide positions 4587-6182 of (SEQ

ID NO:1)(also designated [SEQ ID NO:13]); (viii) contiguous nucleotide positions 4587-4835 of
 (SEQ ID NO:1)(also designated [SEQ ID NO:15]); (ix) contiguous nucleotide positions 4836-6182
 of (SEQ ID NO:1)(also designated [SEQ ID NO:17]); (x) contiguous nucleotide positions 4272-6515
 of (SEQ ID NO:1)(also designated [SEQ ID NO:34]); (xi) contiguous nucleotide positions 6303-6434
 5 of (SEQ ID NO:1)(also designated [SEQ ID NO:19]); (xii) contiguous nucleotide positions 6584-
 8278 of (SEQ ID NO:1)(also designated [SEQ ID NO:21]); or (xiii) contiguous nucleotide positions
 8414-15178 of (SEQ ID NO:1)(also designated [SEQ ID NO:23]). A nucleotide complementary to
 any of (SEQ ID NOS:3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 33, or 34), or a degenerate coding sequence
 of any of (SEQ ID NOS:3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 33, or 34) is also encompassed by the
 10 nucleic acid fragment.

As used herein, the term "degenerate coding sequence", or interchangeably, "degenerate
 sequence", refers to a protein-encoding nucleic acid sequence that has at least one codon that differs
 in at least one nucleotide position from any reference nucleic acid, e.g., any of SEQ ID NO:3, 5, 7, 9,
 11, 13, 15, 17, 19, 21, or 23, but which encodes the same amino acids as the reference nucleic acid.
 15 For example, codons specified by the triplets "UCU", "UCC", "UCA", and "UCG" are degenerate
 with respect to each other since all four of these codons encode the amino acid serine.

In some embodiments, the *Cryptovirus*-specific fragment is up to about 500 nucleotides long.
 In other embodiments, the fragment is up to about 50 nucleotides long. Other embodiments of the
 inventive nucleic acid fragment are about fifteen nucleotides to about 35 nucleotides long; for
 20 example, this is a preferred length for a *Cryptovirus*-specific primer of the present invention, which is
 a *Cryptovirus*-specific oligonucleotide for use in nucleic acid amplification reactions. Most
 preferably, the inventive *Cryptovirus*-specific primer is about 17 to about 22 nucleotides long, but
 primers as short as 7 contiguous nucleotides may be useful for some gene-specific sequences. (E.g.,
 Vincent, J., *et al.*, *Oligonucleonucleotides as short as 7-mers can be used for PCR amplification*,
 25 *DNA Cell Biol.* 13(1):75-82 [1994]).

The inventive probe is preferably about 7 to about 500 nucleotides long, most preferably
 about 15 to about 150 nucleotides long, and comprises, for at least part of its length, a *Cryptovirus*-
 specific nucleotide sequence at least 7 to 15 nucleotides long, such that the probe hybridizes to a
Cryptovirus-specific single stranded nucleic acid under suitably stringent hybridization conditions.
 30 For example, probes comprising the inventive oligonucleotide primer sequences described herein can
 be labeled for use as probes for detecting or analyzing *Cryptovirus*-specific nucleic acid, including
 nucleic acid amplification products.

Non-limiting examples of the *Cryptovirus*-specific fragments useful as primers or probes include nucleic acids comprising: contiguous nucleotide positions 1684-1701 of SEQ ID NO:1 (designated SEQ ID NO:35); contiguous nucleotide positions 1700-1717 of SEQ ID NO:1 (designated SEQ ID NO:36); contiguous nucleotide positions 4283-4300 of SEQ ID NO:1 (designated SEQ ID NO:37); contiguous nucleotide positions 4299-4316 of SEQ ID NO:1 (designated SEQ ID NO:38); contiguous nucleotide positions 4285-4302 of SEQ ID NO:1 (designated SEQ ID NO:39); contiguous nucleotide positions 4300-4317 of SEQ ID NO:1 (designated SEQ ID NO:40); contiguous nucleotide positions 4518-4535 of SEQ ID NO:1 (designated SEQ ID NO:41); contiguous nucleotide positions 4533-4550 of SEQ ID NO:1 (designated SEQ ID NO:42); contiguous nucleotide positions 6116-6133 of SEQ ID NO:1 (designated SEQ ID NO:44); contiguous nucleotide positions 6192-6209 of SEQ ID NO:1 (designated SEQ ID NO:45); contiguous nucleotide positions 6191-6208 of SEQ ID NO:1 (designated SEQ ID NO:43); contiguous nucleotide positions 7501-7518 of SEQ ID NO:1 (designated SEQ ID NO:46); contiguous nucleotide positions 7517-7534 of SEQ ID NO:1 (designated SEQ ID NO:47); or a nucleotide sequence complementary to any of the preceding SEQ ID NOS:35-47. A polynucleotide particularly useful as a probe, especially for probing nucleic acids in samples of biological materials originating from a human or amplification products derived therefrom, is a nucleic acid comprising contiguous nucleotide positions 4292-4549 of SEQ ID NO:1 (designated SEQ ID NO:48) or a complementary sequence. For probing nucleic acids derived from a sample of biological material from a human, even a large nucleic acid segment of SEQ ID NO:1 can be used, for example a nucleic acid comprising contiguous nucleotide positions 4272-6515 of SEQ ID NO:1 (designated SEQ ID NO:34) or a complementary sequence.

The primer is capable of acting as a point of initiation of synthesis of a polynucleotide strand when placed under appropriate conditions. The primer will be completely or substantially complementary to a region of the polynucleotide strand to be copied. Thus, under conditions conducive to hybridization, the primer will anneal to the complementary region of the analyte strand. Upon addition of suitable reactants, (e.g., a polymerase, nucleotide triphosphates, and the like), the primer is extended by the polymerizing agent to form a copy of the analyte strand. The primer may be single-stranded, or alternatively maybe partially or fully double-stranded.

The terms "analyte polynucleotide" and "analyte strand" refer to a single- or double-stranded nucleic acid molecule which is suspected of containing a *Cryptovirus*-specific target sequence, and which may be present in a sample of biological material.

The inventive probe is a structure comprised of a polynucleotide which forms a hybrid structure with a *Cryptovirus*-specific target sequence, due to complementarity of at least one nucleotide sequence in the probe with a sequence in the target region. The polynucleotide regions of probes may be composed of DNA, and/or RNA, and/or synthetic nucleotide analogs. "Target region" refers to a region of the nucleic acid which is to be amplified and/or detected. The term "target sequence" refers to a *Cryptovirus*-specific sequence with which a probe or primer will form a stable hybrid under desired conditions.

Included within probes are "capture probes" and "label probes". Preferably the probe does not contain a sequence complementary to sequence(s) used to prime a nucleic acid amplification reaction.

The term "capture probe" as used herein refers to a polynucleotide comprised of a single-stranded polynucleotide coupled to a binding partner. The single-stranded polynucleotide is comprised of a targeting polynucleotide sequence, which is complementary to a target sequence in a target region to be detected in the analyte polynucleotide. This complementary region is of sufficient length and complementarity to the target sequence to afford a duplex of stability which is sufficient to immobilize the analyte polynucleotide to a solid surface (via the binding partners). The binding partner is specific for a second binding partner; the second binding partner can be bound to the surface of a solid support, or may be linked indirectly via other structures or binding partners to a solid support.

The term "binding partner" as used herein refers to a molecule capable of binding a ligand molecule with high specificity, as for example an antigen and an antibody specific therefor. In general, the specific binding partners must bind with sufficient affinity to immobilize the analyte copy/complementary strand duplex (in the case of capture probes) under the isolation conditions. Specific binding partners are known in the art, and include, for example, biotin and avidin or streptavidin, IgG and protein A, the numerous known receptor-ligand couples, and complementary polynucleotide strands. In the case of complementary polynucleotide binding partners, the partners are generally at least about 15 bases in length, and may be at least 40 bases in length; in addition, they have a content of Gs and Cs of at least about 25% and as much as about 75%. The polynucleotides may be composed of DNA, RNA, or synthetic nucleotide analogs.

"Coupled" as used herein refers to attachment by covalent bonds or by strong non-covalent interactions (e.g., hydrophobic interactions, hydrogen bonds, etc.). Covalent bonds may be, for example, ester, ether, phosphoester, amide, peptide, imide, carbon-sulfur bonds, carbon-phosphorus bonds, and the like.

A "support" refers to any solid or semi-solid surface to which a desired binding partner may be anchored. Suitable supports include glass, plastic, metal, polymer gels, and the like, and may take the form of beads, wells, dipsticks, membranes, and the like.

As used herein, the term "label probe" refers to an oligonucleotide which is comprised of a targeting polynucleotide sequence, which is complementary to a target sequence to be detected in the analyte polynucleotide. This complementary region is of sufficient length and complementarity to the target sequence to afford a duplex comprised of the "label probe" and the "target sequence" to be detected by the label. The oligonucleotide is coupled to a label either directly, or indirectly via a set of ligand molecules with high specificity for each other.

A "label" includes any atom or moiety which can be used to provide a detectable (preferably quantifiable) signal, and which can be attached by conventional means to a polynucleotide or to polypeptide, such as an antibody. The label can be used alone or in conjunction with additional reagents. Such labels are themselves well-known in the art. The label can be a radioisotope, such as ^{14}C , ^{32}P , ^{35}S , ^3H , or ^{15}O , which is detected with suitable radiation detection means. Alternatively, the label can be a fluorescent labeling agent that chemically binds to antibodies or antigens without denaturation to form a fluorochrome (dye) that is a useful immunofluorescent tracer. A description of immunofluorescent analytic techniques is found in DeLuca, "Immunofluorescence Analysis", in *Antibody As a Tool*, Marchalonis et al., eds., John Wiley & Sons, Ltd., pp. 189-231 (1982). As well, any of diverse fluorescent dyes can optionally be used to label probes or primers or amplification products for ease of detection and/or analysis. Useful fluorescent dyes include, but are not limited to, rhodamine, fluorescein, SYBR Green I, Y10-PRO-1, thiazole orange, Hex (i.e., 6-carboxy-2',4',7',4,7-hexachlorofluorescein), pico green, edans, fluorescein, FAM (i.e., 6-carboxyfluorescein), or TET (i.e., 4,7,2',7'-tetrachloro-6-carboxyfluorescein). (E.g., J. Skeidsvoll and P.M. Ueland, *Analysis of double-stranded DNA by capillary electrophoresis with laser-induced fluorescence detection using the monomeric dye SYBR green I*, Anal. Biochem. 231(20):359-65 [1995]; H. Iwahana et al., *Multiple fluorescence-based PCR-SSCP analysis using internal fluorescent labeling of PCR products*, Biotechniques 21(30):510-14, 516-19 [1996]).

The inventive nucleic acid constructs include recombinant cloning and expression vectors (including plasmids and viral expression vectors, such as retroviral or adenoviral vectors), that contain the inventive nucleic acid. A "vector" is a replicon in which another polynucleotide segment is attached, so as to bring about the replication and/or expression of the attached segment. A "replicon" is any genetic element, e.g., a plasmid, a chromosome, a virus, a cosmid, etc., that behaves as an autonomous unit of polynucleotide replication within a cell, i.e., being capable of replication

under the replicon's own control. Inventive recombinant expression vectors contain one or more inventive nucleic acid segments and include at least a promoter region operatively linked to the inventive nucleic acid segment in a transcriptional unit. Preferred examples of inventive nucleic acid constructs are those that include one or more nucleic acid segments encoding a *Cryptovirus* protein, the *Cryptovirus* coding sequence(s) being thus suitably placed and operatively linked to suitable regulatory sequences within one or more transcriptional units within the construct.

"Control" or "regulatory" sequences, elements or regions refers to polynucleotide sequences which are necessary to effect the expression of coding sequences to which they are ligated. The nature of such control sequences differs depending upon the host organism; in prokaryotes, such control sequences generally include promoter, ribosomal binding site, and terminators; in eukaryotes, generally, such control sequences include promoters, terminators and, in some instances, enhancers. The term "control sequences" is intended to include, at a minimum, all components whose presence is necessary for expression, and may also include additional components whose presence is advantageous, for example, leader sequences.

As used herein, "expression" refers to the process by which genes are transcribed into mRNA, which is in turn translated into peptides, polypeptides, or proteins. With respect to a recombinant expression vector, a promoter region refers to a segment of nucleic acid that controls transcription of a coding sequence to which it is operatively linked. The promoter region includes specific sequences that are sufficient for RNA polymerase recognition, binding and transcription initiation. In addition, the promoter region includes sequences that modulate this recognition, binding and transcription initiation activity of RNA polymerase. These sequences may be *cis* acting or may be responsive to *trans* acting factors. Promoters, depending upon the nature of the regulation, may be constitutive or regulated developmentally or inducibly. Exemplary promoters contemplated for use in the practice of the present invention include the SV40 early promoter, the cytomegalovirus (CMV) promoter, the mouse mammary tumor virus (MMTV) steroid-inducible promoter, Moloney murine leukemia virus (MMLV) promoter, and the like.

For optimal expression of foreign genes in mammalian cells (e.g., the *Cryptovirus* genes of the present invention), the expression vector may also require terminator sequences and poly A addition sequences; enhancer sequences which increase expression may also be included, and sequences which cause amplification of the gene may also be desirable. Such sequences are known in the art.

As used herein, the term "operatively linked" refers to the functional relationship of nucleic acid with regulatory (control or effector) nucleotide sequences, such as promoters, enhancers,

transcriptional and translational stop sites, and other signal sequences. For example, operative linkage of DNA or RNA to a promoter refers to the physical and functional relationship between the DNA or RNA and the promoter such that the transcription of such DNA or RNA is initiated from the promoter by an RNA polymerase that specifically recognizes, binds to and transcribes the DNA or RNA, respectively. A regulatory sequence "operatively linked" to a coding sequence is ligated in such a way that expression of the coding sequence is achieved under conditions compatible with the regulatory sequences. Thus, for example, within a transcriptional unit, the promoter sequence, is located upstream (i.e., 5' in relation thereto) from the coding sequence and the coding sequence, is 3' to the promoter, or alternatively is in a sequence of genes or open reading frames 3' to the promoter and expression is coordinately regulated thereby. Both the promoter and coding sequences are oriented in a 5' to 3' manner, such that transcription can take place in vitro in the presence of all essential enzymes, transcription factors, co-factors, activators, and reactants, under favorable physical conditions, e.g., suitable pH and temperature. This does not mean that, in any particular cell, conditions will favor transcription. For example, transcription from a tissue-specific promoter is generally not favored in heterologous cell types from different tissues.

The inventive expression vector, comprising a *Cryptovirus*-specific nucleic acid, is used to transform a cell. "Transformation", as used herein, refers to the insertion of an exogenous polynucleotide into a host cell, irrespective of the method used for the insertion, for example, direct uptake, transfection, transduction, f-mating, microparticle bombardment, or electroporation. The exogenous polynucleotide may be maintained as a non-integrated vector, for example, a plasmid, or alternatively, may be integrated into the host genome. A "transformed" host cell refers to both the immediate cell that has undergone transformation and its progeny that maintain the originally exogenous polynucleotide.

Appropriate expression vectors are well known to those of skill in the art and include those that are replicable in eukaryotic cells and/or prokaryotic cells and those that remain episomal or those which integrate into the host cell genome.

Exemplary, eukaryotic expression vectors, include the cloned bovine papilloma virus genome, the cloned genomes of the murine retroviruses, and eukaryotic cassettes, such as the pSV-2 gpt system (described by Mulligan and Berg, 1979, Nature Vol. 277:108-114) the Okayama-Berg cloning system (Mol. Cell Biol. Vol. 2:161-170, 1982), pGAL4, pCI (e.g., pCI-neo), and the expression cloning vector described by Genetics Institute (Science Vol. 228:810-815, 1985), are available which provide substantial assurance of at least some expression of the protein of interest in the transformed mammalian cell.

Preferred are vectors which contain regulatory elements that can be linked to the inventive nucleic acids, for transfection or transduction of mammalian cells. Examples are cytomegalovirus (CMV) promoter-based vectors such as pcDNA1 (Invitrogen, San Diego, CA), MMTV promoter-based vectors such as pMAMNeo (Clontech, Palo Alto, CA) and pMSG (Pharmacia, Piscataway, NJ), and SV40 promoter-based vectors such as pSV β (Clontech, Palo Alto, CA). In one embodiment of the present invention, adenovirus-transferrin/polylysine-DNA (TfAdpl-DNA) vector complexes (Wagner et al., 1992, PNAS, USA, 89:6099-6103; Curiel et al., 1992, Hum. Gene Therapy, 3:147-154; Gao et al., 1993, Hum. Gene Ther., 4:14-24) are employed to transduce mammalian cells with heterologous *Cryptovirus*-specific nucleic acid. Any of the plasmid expression vectors described herein may be employed in a TfAdpl-DNA complex.

In addition, expression vectors may contain appropriate packaging signals that enable the vector to be packaged by a number of viral virions, e.g., retroviruses, such as human immunodeficiency virus, lentiviruses, mumps virus, herpes viruses, adenoviruses, resulting in the formation of a "viral vector." (See, e.g., Anderson, W.F., *Gene therapy scores against cancer*, Nat. Med. 6(8):862-63 [2000]). These viral vectors include, for example, Herpes simplex virus vectors (e.g., Geller et al., 1988, Science, 241:1667-1669), Vaccinia virus vectors (e.g., Piccini et al., 1987, Meth. in Enzymology, 153:545-563); Cytomegalovirus vectors (Mocarski et al., in *Viral Vectors*, Y. Gluzman and S.H. Hughes, Eds., Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y., 1988, pp. 78-84), Moloney murine leukemia virus vectors (Danos et al., 1980, PNAS, USA, 85:6469), adenovirus vectors (e.g., Logan et al., 1984, PNAS, USA, 81:3655-3659; Jones et al., 1979, Cell, 17:683-689; Berkner, 1988, Biotechniques, 6:616-626; Cotten et al., 1992, PNAS, USA, 89:6094-6098; Graham et al., 1991, Meth. Mol. Biol., 7:109-127), adeno-associated virus vectors, retrovirus vectors (see, e.g., U.S. Patent 5,252,479, WIPO publications WO 92/07573, WO 90/06997, WO 89/05345, WO 92/05266 and WO 92/14829, U.S. Patent Nos. 4,405,712 and 4,650,764; Shackleford et al., 1988, PNAS, USA, 85:9655-9659), and the like.

A preferred viral vector is Moloney murine leukemia virus and the pseudotyped retroviral vector derived from Moloney virus called vesicular-stomatitis-virus-glycoprotein (VSV-G)-Moloney murine leukemia virus. A most preferred viral vector is a pseudotyped (VSV-G) lentiviral vector derived from the HIV virus, which is used to transduce mammalian cells. (Naldini, L., et al., *In vivo gene delivery and stable transduction of nondividing cells by a lentiviral vector*, Science 272: 263-267 [1996]). This gene delivery system employs retroviral particles generated by a three-plasmid expression system. In this system a packaging construct contains the human cytomegalovirus (hCMV) immediate early promoter, driving the expression of all viral proteins. The construct's

design eliminates the cis-acting sequences crucial for viral packaging, reverse transcription and integration of these transcripts. The second plasmid encodes a heterologous envelope protein (*env*), namely the G glycoprotein of the vesicular stomatitis virus (VSV-G). The third plasmid, the transducing vector (pHR'), contains cis-acting sequences of human immunodeficiency virus (HIV) required for packaging, reverse transcription and integration, as well as unique restriction sites for cloning heterologous complementary DNAs (cDNAs). For example, a genetic selection marker, such as green fluorescent protein (GFP), enhanced green fluorescent protein (EGFP), blue fluorescent protein, yellow fluorescent protein, a fluorescent phycobiliprotein, β -galactosidase, and/or a gene encoding another preselected product is cloned downstream of the hCMV promoter in the HR' vector, and is operatively linked so as to form a transcriptional unit. A VSV-G pseudotyped retroviral vector system is capable of infecting a wide variety of cells including cells from different species and of integrating into the genome. Some retroviruses, i.e., lentiviruses, such as HIV, have the ability to infect non-dividing cells. Lentiviruses have a limited capacity for heterologous DNA sequences, the size limit for this vector being 7-7.5 kilobases (Verma, I.M. and Somia, N., *Gene Therapy – promises, problems and prospects*, Nature 389:239-242 [1997]). *In vivo* experiments with lentiviruses show that expression does not shut off like other retroviral vectors and that *in vivo* expression in brain, muscle, liver or pancreatic-islet cells, is sustained at least for over six months – the longest time tested so far (Verma and Somia [1997]; Anderson, WF., *Human Gene Therapy*, Nature (Suppl). 392:25-30 [1998]).

All of the above viruses may require modification to render them non-pathogenic or less antigenic. Other known viral vector systems, however, are also useful within the confines of the invention.

A particularly useful expression vector which is useful to express foreign cDNA and which may be used in vaccine preparation is *Vaccinia* virus. In this case, the heterologous cDNA is inserted into the *Vaccinia* genome. Techniques for the insertion of foreign cDNA into the vaccinia virus genome are known in the art, and utilize, for example, homologous recombination. The insertion of the heterologous cDNA is generally into a gene which is non-essential in nature, for example, the thymidine kinase gene (*tk*), which also provides a selectable marker. Plasmid vectors that greatly facilitate the construction of recombinant viruses have been described (see, for example, Mackett *et al.* (1984) in "*DNA Cloning*" Vol II IRL Press, p. 191, Chakrabarti *et al.* (1985), Mol. Cell Biol. 5:3403; Moss (1987) in "*Gene Transfer Vectors for Mammalian Cells*" (Miller and Calos, eds., p. 10). Expression of the desired polypeptides containing immunoreactive regions then occurs in cells and mammals that are infected and/or immunized with the live recombinant *Vaccinia* virus.

The inventive "nucleic acid construct" also encompasses a construct that is not contained in a vector, for example, a synthetic antisense oligonucleotide, such as a phosphorothioate oligodeoxynucleotide. Synthetic antisense oligonucleotides, or other antisense chemical structures designed to recognize and selectively bind to mRNA, are constructed to be complementary to portions of the *Cryptovirus* coding strand, for example, to coding sequences shown in SEQ ID NO:3, 5, 7, 9, 11, 13, 15, 17, 19, 21, or 23. When taken up by a mammalian cell, the antisense oligonucleotide prevents translational expression of at least part of the *Cryptovirus* coding region, the inventive antisense oligonucleotide is useful to prevent expression of a *Cryptovirus* protein. Antisense oligonucleotides inactivate target mRNA sequences by either binding thereto and inducing degradation of the mRNA by, for example, RNase I digestion, or inhibiting translation of mRNA target sequence by interfering with the binding of translation-regulating factors or ribosomes, or by inclusion of other chemical structures, such as ribozyme sequences or reactive chemical groups which either degrade or chemically modify the target mRNA. Gene-based therapy strategies employing antisense oligonucleotides are well known in the art. (E.g., Rait, A. et al., *3'-End conjugates of minimally phosphorothioate-protected oligonucleotides with 1-O-hexadecylglycerol: synthesis and anti-ras activity in radiation-resistant cells*, Bioconjug Chem., 11(2):153-60 [2000]; Stenton, G. R. et al., *Aerosolized syk antisense suppresses syk expression, mediator release from macrophages, and pulmonary inflammation*, J. Immunol., 164(7):3790-7 [2000]; Suzuki, J. et al., *Antisense Bcl-x oligonucleotide induces apoptosis and prevents arterial neointimal formation in murine cardiac allografts*, Cardiovas. Res., 45(3):783-7 [2000]; Kim, J. W. et al., *Antisense oligodeoxynucleotide of glyceraldehyde-3-phosphate dehydrogenase gene inhibits cell proliferation and induces apoptosis in human cervical carcinoma cell line*, Antisense Nucleic Acid Drug Dev., 9(6):507-13 [1999]; Han, D. C. et al., *Therapy with antisense TGF-beta1 oligodeoxynucleotides reduces kidney weight and matrix mRNAs in diabetic mice*, Am. J. Physiol. Renal Physiol., 278(4):F628-F634 [2000]; Scala, S. et al., *Adenovirus-mediated suppression of HMGI (Y) protein synthesis as potential therapy of human malignant neoplasias*, Proc. Natl. Acad. Sci. USA., 97(8):4256-4261 [2000]; Arteaga, C. L., et al., *Tissue-targeted antisense c-fos retroviral vector inhibits established breast cancer xenografts in nude mice*, Cancer Res., 56(5):1098-1103 [1996]; Muller, M. et al., *Antisense phosphorothioate oligodeoxynucleotide down-regulation of the insulin-like growth factor I receptor in ovarian cancer cells*, Int. J. Cancer, 77(4):567-71 [1998]; Engelhard, H. H., *Antisense Oligodeoxynucleotide Technology: Potential Use for the Treatment of Malignant Brain Tumors*, Cancer Control, 5(2):163-170 [1998]; Alvarez-Salas, L. M. et al., *Growth inhibition of cervical tumor cells by antisense oligodeoxynucleotides directed to the human papillomavirus type 16 E6 gene*, Antisense Nucleic Acid

Drug Dev., 9(5):441-50 [1999]; Im, S. A., et al., *Antiangiogenesis treatment for gliomas: transfer of antisense-vascular endothelial growth factor inhibits tumor growth in vivo*, Cancer Res., 59(4):895-900 [1999]; Maeshima, Y. et al., *Antisense oligonucleotides to proliferating cell nuclear antigen and Ki-67 inhibit human mesangial cell proliferation*, J. Am. Soc. Nephrol., 7(10):2219-29 [1996]; Chen, D. S. et al., *Retroviral Vector-mediated transfer of an antisense cyclin G1 construct inhibits osteosarcoma tumor growth in nude mice*, Hum. Gene Ther., 8(14):1667-74 [1997]; Hirao, T. et al., *Antisense epidermal growth factor receptor delivered by adenoviral vector blocks tumor growth in human gastric cancer*, Cancer Gene Ther., 6(5):423-7 [1999]; Wang, X. Y. et al., *Antisense inhibition of protein kinase Calpha reverses the transformed phenotype in human lung carcinoma cells*, Exp. Cell Res., 250(1):253-63 [1999]; Sacco, M.G. et al., *In vitro and in vivo antisense-mediated growth inhibition of a mammary adenocarcinoma from MMTV-neu transgenic mice*, Gene Ther., 5(3):388-93 [1998]; Leonetti, C. et al., *Antitumor effect of c-myc antisense phosphorothioate oligodeoxynucleotides on human melanoma cells in vitro and in mice*, J. Natl. Cancer Inst., 88(7):419-29 [1996]; Laird, A. D. et al., *Inhibition of tumor growth in liver epithelial cells transfected with a transforming growth factor alpha antisense gene*, Cancer Res. 54(15):4224-32 (Aug 1, 1994); Yazaki, T. et al., *Treatment of glioblastoma U-87 by systemic administration of an antisense protein kinase C-alpha phosphorothioate oligodeoxynucleotide*, Mol. Pharmacol., 50(2):236-42 [1996]; Ho, P. T. et al., *Antisense oligonucleotides as therapeutics for malignant diseases*, Semin. Oncol., 24(2):187-202 [1997]; Muller, M. et al., *Antisense phosphorothioate oligodeoxynucleotide down-regulation of the insulin-like growth factor I receptor in ovarian cancer cells*, Int. J. Cancer, 77(4):567-71 [1998]; Elez, R. et al., *Polo-like kinase1, a new target for antisense tumor therapy*, Biochem. Biophys. Res. Commun., 269(2):352-6 [2000]; Monia, B. P. et al., *Antitumor activity of a phosphorothioate antisense oligodeoxynucleotide targeted against C-raf kinase*, Nat. Med., 2(6):668-75 [1996]).

The present invention relates to an isolated *Cryptovirus* protein. The term "protein" refers to a polymer of amino acids of any length, i.e., a polypeptide, and does not refer to a specific length of the product; thus, "polypeptides", "peptides", and "oligopeptides", are included within the definition of "protein", and such terms are used interchangeably herein with "protein". The term "protein" also includes post-expression modifications of the polypeptide, for example, glycosylations, acetylations, phosphorylations and the like. Included within the definition of "protein" are, for example, polypeptides containing one or more analogs of an amino acid (including, for example, unnatural amino acids, etc.), polypeptides with substituted linkages, as well as other modifications known in the

art, both naturally occurring and non-naturally occurring. Methods of inserting analogs of amino acids into a peptide sequence are known in the art.

5 The phrase "isolated *Cryptovirus* protein" refers to a cryptoviral protein which is substantially free, i.e., contains less than about 50%, preferably less than about 70%, and even more preferably less than about 90%, of the cellular components and/or contaminants normally associated with a native in vivo environment. An isolated *Cryptovirus* protein of the present invention can also be further isolated from one or more other components of a *Cryptovirus* particle, for example, other *Cryptovirus* protein species, phospholipid components of the viral envelope, or the viral genome. In some useful embodiments the inventive isolated *Cryptovirus* protein is purified to homogeneity by
10 known virological and biochemical methods.

The inventive protein is encoded by a *Cryptovirus*-specific nucleic acid segment, which nucleic acid segment comprises:

- (i) contiguous nucleotide positions 152-1678 of (SEQ ID NO:1)(also designated SEQ ID NO:3), or a degenerate coding sequence, which encodes the *Cryptovirus* nucleocapsid (NP) protein;
- 15 (ii) contiguous nucleotide positions 1850-2515 of (SEQ ID NO:1)(also designated SEQ ID NO:5), or a degenerate coding sequence, which encodes a *Cryptovirus* RNA binding (V) protein thought to be a component of the viral RNA-dependent RNA polymerase;
- (iii) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1)(also designated SEQ ID NO:33) combined with a further insertion of two guanine (G) residues between nucleotide position
20 2339 of (SEQ ID NO:1) and nucleotide position 2340 of (SEQ ID NO:1)(the combined coding sequence including the "GG" insertion being designated [SEQ ID NO:7]), or a degenerate coding sequence thereof; this frameshift-causing insertion into the mRNA encoding the *Cryptovirus* nucleocapsid-associated phosphoprotein (P protein) occurs during processing of the mRNA and is not templated by the *Cryptovirus* minus stranded RNA genomic sequence;
- 25 (iv) contiguous nucleotide positions 3141-4271 of (SEQ ID NO:1)(also designated SEQ ID NO:9), or a degenerate coding sequence, which encodes the *Cryptovirus* virion-associated matrix or membrane (M) protein;
- (v) contiguous nucleotide positions 4530-6182 of (SEQ ID NO:1)(also designated SEQ ID NO:11) or a degenerate coding sequence, which encodes the *Cryptovirus* (uncleaved) fusion (F)
30 protein, which is a propeptide form of a major envelope-associated glycoprotein, and includes a 19-amino acid signal region at its amino terminus;
- (vi) contiguous nucleotide positions 4587-6182 of (SEQ ID NO:1)(also designated SEQ ID NO:13), or a degenerate coding sequence, which encodes the *Cryptovirus* (uncleaved) fusion (F₀)

protein, which is a propeptide form of a major envelope-associated glycoprotein, minus the 19-amino acid signal region at its amino terminus;

(vii) contiguous nucleotide positions 4587-4835 of (SEQ ID NO:1)(also designated SEQ ID NO:15), or a degenerate coding sequence, which encodes the cleaved F₂ protein;

(viii) contiguous nucleotide positions 4836-6182 of (SEQ ID NO:1)(also designated SEQ ID NO:17), or a degenerate coding sequence, which encodes the cleaved F₁ protein, including a 22-amino acid carboxy terminal peptide segment that is thought to be important to *Cryptovirus* infectivity;

(ix) contiguous nucleotide positions 6303-6434 of (SEQ ID NO:1)(also designated SEQ ID NO:19), or a degenerate coding sequence, which encodes the *Cryptovirus* SH protein, a small envelope-associated protein;

(x) contiguous nucleotide positions 6584-8278 of (SEQ ID NO:1)(also designated SEQ ID NO:21), or a degenerate coding sequence, which encodes the *Cryptovirus* hemagglutinin (HN) protein, another major envelope protein; or

(xi) contiguous nucleotide positions 8414-15178 of (SEQ ID NO:1)(also designated SEQ ID NO:23), or a degenerate coding sequence, which encodes the *Cryptovirus* largest nucleocapsid associated protein (L protein).

These are the amino acid sequences corresponding to the preceding inventive proteins in the same order:

(i) NP has the following amino acid sequence (SEQ ID NO:4):

Met	Ser	Ser	Val	Leu	Lys	Ala	Tyr	Glu	Arg	Phe	Thr	Leu	Thr	Gln	Glu
1				5				10						15	
Leu	Gln	Asp	Gln	Ser	Glu	Glu	Gly	Thr	Ile	Pro	Pro	Thr	Thr	Leu	Lys
			20					25						30	
Pro	Val	Ile	Arg	Val	Phe	Val	Leu	Thr	Ser	Asn	Asn	Pro	Glu	Leu	Arg
			35				40					45			
Ser	Arg	Leu	Leu	Leu	Phe	Cys	Leu	Arg	Ile	Val	Leu	Ser	Asn	Gly	Ala
			50			55					60				
Arg	Asp	Ser	His	Arg	Phe	Gly	Ala	Leu	Leu	Thr	Met	Phe	Ser	Leu	Pro
					70					75				80	
Ser	Ala	Thr	Met	Leu	Asn	His	Val	Lys	Leu	Ala	Asp	Gln	Ser	Pro	Glu
				85					90					95	
Ala	Asp	Ile	Glu	Arg	Val	Glu	Ile	Asp	Gly	Phe	Glu	Glu	Gly	Ser	Phe
			100					105					110		
Arg	Leu	Ile	Pro	Asn	Ala	Arg	Ser	Gly	Met	Ser	Arg	Gly	Glu	Ile	Asn
			115				120					125			
Ala	Tyr	Ala	Ala	Leu	Ala	Glu	Asp	Leu	Pro	Asp	Thr	Leu	Asn	His	Ala
			130			135					140				
Thr	Pro	Phe	Val	Asp	Ser	Glu	Val	Glu	Gly	Thr	Ala	Trp	Asp	Glu	Ile
					150					155				160	

Glu Thr Phe Leu Asp Met Cys Tyr Ser Val Leu Met Gln Ala Trp Ile
 165 170 175
 Val Thr Cys Lys Cys Met Thr Ala Pro Asp Gln Pro Ala Ala Ser Ile
 180 185 190
 5 Glu Lys Arg Leu Gln Lys Tyr Arg Gln Gln Gly Arg Ile Asn Pro Arg
 195 200 205
 Tyr Leu Leu Gln Pro Glu Ala Arg Arg Ile Ile Gln Asn Val Ile Arg
 210 215 220
 10 Lys Gly Met Val Val Arg His Phe Leu Thr Phe Glu Leu Gln Leu Ala
 225 230 235 240
 Arg Ala Gln Ser Leu Val Ser Asn Arg Tyr Tyr Ala Met Val Gly Asp
 245 250 255
 Val Gly Lys Tyr Ile Glu Asn Cys Gly Met Gly Gly Phe Phe Leu Thr
 260 265 270
 15 Leu Lys Tyr Ala Leu Gly Thr Arg Trp Pro Thr Leu Ala Leu Ala Ala
 275 280 285
 Phe Ser Gly Glu Leu Thr Lys Leu Lys Ser Leu Met Ala Leu Tyr Gln
 290 295 300
 20 Thr Leu Gly Glu Gln Ala Arg Tyr Leu Ala Leu Leu Glu Ser Pro His
 305 310 315 320
 Leu Met Asp Phe Ala Ala Ala Asn Tyr Pro Leu Leu Tyr Ser Tyr Ala
 325 330 335
 Met Gly Ile Gly Tyr Val Leu Asp Val Asn Met Arg Asn Tyr Ala Phe
 340 345 350
 25 Ser Arg Ser Tyr Met Asn Lys Thr Tyr Phe Gln Leu Gly Met Glu Thr
 355 360 365
 Ala Arg Lys Gln Gln Gly Ala Val Asp Met Arg Met Ala Glu Asp Leu
 370 375 380
 30 Gly Leu Thr Gln Ala Glu Arg Thr Glu Met Ala Asn Thr Leu Ala Lys
 385 390 395 400
 Leu Thr Thr Ala Asn Arg Gly Ala Asp Thr Arg Gly Gly Val Asn Pro
 405 410 415
 Phe Ser Ser Val Thr Gly Thr Thr Gln Met Pro Ala Ala Ala Thr Gly
 420 425 430
 35 Asp Thr Phe Glu Ser Tyr Met Ala Ala Asp Arg Leu Arg Gln Arg Tyr
 435 440 445
 Ala Asp Ala Gly Thr His Asp Asp Glu Met Pro Pro Leu Glu Glu Glu
 450 455 460
 40 Glu Glu Asp Asp Thr Ser Ala Gly Pro Arg Thr Glu Pro Thr Pro Glu
 465 470 475 480
 Gln Val Ala Leu Asp Ile Gln Ser Ala Ala Val Gly Ala Pro Ile His
 485 490 495
 Thr Asp Asp Leu Asn Ala Ala Leu Gly Asp Leu Asp Ile
 500 505 //(SEQ ID NO:4);
 45

(ii) V has the following amino acid sequence (SEQ ID NO:6):

Met Asp Pro Thr Asp Leu Ser Phe Ser Pro Asp Glu Ile Asn Lys Leu
 1 5 10 15
 Ile Glu Thr Gly Leu Asn Thr Val Glu Tyr Phe Thr Ser Gln Gln Val
 20 25 30
 50

Thr Gly Thr Ser Ser Leu Gly Lys Asn Thr Ile Pro Pro Gly Val Thr
 35 40 45
 Gly Leu Leu Thr Asn Ala Ala Gly Ala Lys Ile Gln Glu Ser Ile Asn
 50 55 60
 5 His Gln Lys Gly Ser Val Gly Gly Gly Thr Asn Pro Lys Lys Pro Arg
 65 70 75 80
 Ser Lys Ile Ala Ile Val Pro Ala Asp Asp Lys Thr Val Pro Glu Lys
 85 90 95
 10 Pro Ile Pro Asn Pro Leu Leu Gly Leu Asp Ser Thr Pro Ser Thr Gln
 100 105 110
 Thr Val Leu Asp Leu Ser Gly Lys Thr Leu Pro Ser Gly Ser Tyr Lys
 115 120 125
 Gly Val Lys Leu Ala Lys Phe Gly Lys Glu Asn Leu Met Thr Arg Phe
 130 135 140
 15 Ile Glu Glu Pro Arg Glu Asn Pro Ile Ala Thr Ser Ser Pro Ile Asp
 145 150 155 160
 Phe Lys Arg Gly Arg Asp Thr Gly Gly Phe His Arg Arg Glu Tyr Ser
 165 170 175
 20 Ile Gly Trp Val Gly Asp Glu Val Lys Val Thr Glu Trp Cys Asn Pro
 180 185 190
 Ser Cys Ser Pro Ile Thr Ala Ala Arg Arg Phe Lys Cys Thr Cys
 195 200 205
 His Gln Cys Pro Val Thr Cys Ser Glu Cys Glu Arg Asp Thr
 210 215 220 // (SEQ ID NO:6);

25

(iii) P has the following amino acid sequence (SEQ ID NO:8):

Met Asp Pro Thr Asp Leu Ser Phe Ser Pro Asp Glu Ile Asn Lys Leu
 1 5 10 15
 30 Ile Glu Thr Gly Leu Asn Thr Val Glu Tyr Phe Thr Ser Gln Gln Val
 20 25 30
 Thr Gly Thr Ser Ser Leu Gly Lys Asn Thr Ile Pro Pro Gly Val Thr
 35 40 45
 Gly Leu Leu Thr Asn Ala Ala Glu Ala Lys Ile Gln Glu Ser Ile Asn
 50 55 60
 35 His Gln Lys Gly Ser Val Gly Gly Gly Thr Asn Pro Lys Lys Pro Arg
 65 70 75 80
 Ser Lys Ile Ala Ile Val Pro Ala Asp Asp Lys Thr Val Pro Glu Lys
 85 90 95
 40 Pro Ile Pro Asn Pro Leu Leu Gly Leu Asp Ser Thr Pro Ser Thr Gln
 100 105 110
 Thr Val Leu Asp Leu Ser Gly Lys Thr Leu Pro Ser Gly Ser Tyr Lys
 115 120 125
 Gly Val Lys Leu Ala Lys Phe Gly Lys Glu Asn Leu Met Thr Arg Phe
 130 135 140
 45 Ile Glu Glu Pro Arg Glu Asn Pro Ile Ala Thr Ser Ser Pro Ile Asp
 145 150 155 160
 Phe Lys Arg Gly Ala Glu Ile Pro Val Gly Ser Ile Glu Gly Ser Thr
 165 170 175
 50 Gln Ser Asp Gly Trp Glu Met Lys Ser Arg Ser Leu Ser Gly Ala Ile
 180 185 190

His Pro Val Leu Gln Ser Pro Leu Gln Gln Gly Asp Leu Asn Ala Leu
 195 200 205
 Val Thr Asn Val Gln Ser Leu Ala Leu Asn Val Asn Glu Ile Leu Asn
 210 215 220
 5 Thr Val Arg Asn Leu Asp Ser Arg Met Asn Gln Leu Glu Thr Lys Val
 225 230 235 240
 Asp Arg Ile Leu Ser Ser Gln Ser Leu Ile Gln Thr Ile Lys Asn Asp
 245 250 255
 10 Ile Ile Gly Leu Lys Ala Gly Met Ala Thr Leu Glu Gly Met Ile Thr
 260 265 270
 Thr Val Lys Ile Met Asp Pro Gly Val Pro Ser Asn Val Thr Val Glu
 275 280 285
 Asp Val Arg Lys Lys Leu Ser Asn His Ala Val Val Val Pro Glu Ser
 290 295 300
 15 Phe Asn Asp Ser Phe Leu Thr Gln Ser Glu Asp Val Ile Ser Leu Asp
 305 310 315 320
 Glu Leu Ala Arg Pro Thr Ala Thr Ser Val Lys Lys Ile Val Arg Lys
 325 330 335
 20 Val Pro Pro Gln Lys Asp Leu Thr Gly Leu Lys Ile Thr Leu Glu Gln
 340 345 350
 Leu Ala Lys Asp Cys Ile Ser Lys Pro Lys Met Arg Glu Asp Tyr Leu
 355 360 365
 Leu Lys Ile Asn Gln Ala Ser Ser Glu Ala Gln Leu Ile Asp Leu Lys
 370 375 380
 25 Lys Ala Ile Ile Arg Ser Ala Ile
 385 390 //(SEQ ID NO:8);

(iv) M has the following amino acid sequence (SEQ ID NO:10):

30 Met Pro Ser Ile Ser Ile Pro Ala Asp Pro Thr Asn Pro Arg Gln Ser
 1 5 10 15
 Ile Lys Ala Phe Pro Ile Val Ile Asn Ser Asp Gly Gly Glu Lys Gly
 20 25 30
 Arg Leu Val Lys Gln Leu Arg Thr Thr Tyr Leu Asn Asp Leu Asp Thr
 35 35 40 45
 His Glu Pro Leu Val Thr Phe Val Asn Thr Tyr Gly Phe Ile Tyr Glu
 50 55 60
 Gln Asn Arg Gly Asn Ala Ile Val Gly Glu Asp Gln Leu Gly Lys Lys
 65 70 75 80
 40 Arg Glu Ala Val Thr Ala Ala Met Val Thr Leu Gly Cys Gly Pro Asn
 85 90 95
 Leu Pro Ser Leu Gly Asn Val Leu Arg Gln Leu Ser Glu Phe Gln Val
 100 105 110
 Ile Val Arg Lys Thr Ser Ser Lys Ala Glu Glu Met Val Phe Glu Ile
 115 120 125
 45 Val Lys Tyr Pro Arg Ile Phe Arg Gly His Thr Leu Ile Gln Lys Gly
 130 135 140
 Leu Val Cys Val Ser Ala Glu Lys Phe Val Lys Ser Pro Gly Lys Val
 145 150 155 160
 50 Gln Ser Gly Met Asp Tyr Leu Phe Ile Pro Thr Phe Leu Ser Val Thr
 165 170 175

Tyr Cys Pro Ala Ala Ile Lys Phe Gln Val Pro Gly Pro Met Leu Lys
 180 185 190
 Met Arg Ser Arg Tyr Thr Gln Ser Leu Gln Leu Glu Leu Met Ile Arg
 195 200 205
 5 Ile Leu Cys Lys Pro Asp Ser Pro Leu Met Lys Val His Ile Pro Asp
 210 215 220
 Lys Glu Gly Arg Gly Cys Leu Val Ser Val Trp Leu His Val Cys Asn
 225 230 235 240
 10 Ile Phe Lys Ser Gly Asn Lys Asn Gly Ser Glu Trp Gln Glu Tyr Trp
 245 250 255
 Met Arg Lys Cys Ala Asn Met Gln Leu Glu Val Ser Ile Ala Asp Met
 260 265 270
 Trp Gly Pro Thr Ile Ile Ile His Ala Arg Gly His Ile Pro Lys Ser
 275 280 285
 15 Ala Lys Leu Phe Phe Gly Lys Gly Gly Trp Ser Cys His Pro Leu His
 290 295 300
 Glu Ile Val Pro Ser Val Thr Lys Thr Leu Trp Ser Val Gly Cys Glu
 305 310 315 320
 20 Ile Thr Lys Ala Lys Ala Ile Ile Gln Glu Ser Ser Ile Ser Leu Leu
 325 330 335
 Val Glu Thr Thr Asp Ile Ile Ser Pro Lys Val Lys Ile Ser Ser Lys
 340 345 350
 His Arg Arg Phe Gly Lys Ser Asn Trp Gly Leu Phe Lys Lys Thr Lys
 355 360 365
 25 Ser Leu Pro Asn Leu Thr Glu Leu Glu
 370 375 //(SEQ ID NO:10);

(v) F has the following amino acid sequence (SEQ ID NO:12):

30 Met Ser Thr Ile Ile Gln Ser Leu Val Val Ser Cys Leu Leu Ala Gly
 1 5 10 15
 Ala Gly Ser Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro
 20 25 30
 Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe
 35 35 40 45
 35 Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys
 50 55 60
 Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu
 65 70 75 80
 40 Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro
 85 90 95
 Thr Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala
 100 105 110
 Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val
 115 120 125
 45 Lys Ala Asn Glu Asn Thr Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile
 130 135 140
 Gln Lys Thr Asn Ala Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser
 145 150 155 160
 50 Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Ile
 165 170 175

	Ser	Pro	Ala	Ile	Thr	Ala	Ala	Asn	Cys	Lys	Ala	Gln	Asp	Ala	Ile	Ile	
				180					185					190			
	Gly	Ser	Ile	Leu	Asn	Leu	Tyr	Leu	Thr	Glu	Leu	Thr	Thr	Ile	Phe	His	
			195					200					205				
5	Asn	Gln	Ile	Thr	Asn	Pro	Ala	Leu	Ser	Pro	Ile	Thr	Ile	Gln	Ala	Leu	
		210					215					220					
	Arg	Ile	Leu	Leu	Gly	Ser	Thr	Leu	Pro	Thr	Val	Val	Glu	Lys	Ser	Phe	
	225					230					235					240	
	Asn	Thr	Gln	Ile	Ser	Ala	Ala	Glu	Leu	Leu	Ser	Ser	Gly	Leu	Leu	Thr	
10					245					250					255		
	Gly	Gln	Ile	Val	Gly	Leu	Asp	Leu	Thr	Tyr	Met	Gln	Met	Val	Ile	Lys	
				260					265					270			
	Ile	Glu	Leu	Pro	Thr	Leu	Thr	Val	Gln	Pro	Ala	Thr	Gln	Ile	Ile	Asp	
				275				280					285				
15	Leu	Ala	Thr	Ile	Ser	Ala	Phe	Ile	Asn	Asn	Gln	Glu	Val	Met	Ala	Gln	
		290					295					300					
	Leu	Pro	Thr	Arg	Val	Ile	Val	Thr	Gly	Ser	Leu	Ile	Gln	Ala	Tyr	Pro	
	305					310					315					320	
	Ala	Ser	Gln	Cys	Thr	Ile	Thr	Pro	Asn	Thr	Val	Tyr	Cys	Arg	Tyr	Asn	
20					325					330					335		
	Asp	Ala	Gln	Val	Leu	Ser	Asp	Asp	Thr	Met	Ala	Cys	Leu	Gln	Gly	Asn	
				340					345					350			
	Leu	Thr	Arg	Cys	Thr	Phe	Ser	Pro	Val	Val	Gly	Ser	Phe	Leu	Thr	Arg	
			355					360					365				
25	Phe	Val	Leu	Phe	Asp	Gly	Ile	Val	Tyr	Ala	Asn	Cys	Arg	Ser	Met	Leu	
		370					375					380					
	Cys	Lys	Cys	Met	Gln	Pro	Ala	Ala	Val	Ile	Leu	Gln	Pro	Ser	Ser	Ser	
	385					390					395					400	
	Pro	Val	Thr	Val	Ile	Asp	Met	His	Lys	Cys	Val	Ser	Leu	Gln	Leu	Asp	
30					405					410					415		
	Asp	Leu	Arg	Phe	Thr	Ile	Thr	Gln	Leu	Ala	Asn	Val	Thr	Tyr	Asn	Ser	
				420					425					430			
	Thr	Ile	Lys	Leu	Glu	Thr	Ser	Gln	Ile	Leu	Pro	Ile	Asp	Pro	Leu	Asp	
			435					440					445				
35	Ile	Ser	Gln	Asn	Leu	Ala	Ala	Val	Asn	Lys	Ser	Leu	Ser	Asp	Ala	Leu	
		450					455					460					
	Gln	His	Leu	Ala	Gln	Ser	Asp	Thr	Tyr	Leu	Ser	Ala	Ile	Thr	Ser	Ala	
	465					470					475					480	
	Thr	Thr	Thr	Ser	Val	Leu	Ser	Ile	Ile	Ala	Ile	Cys	Leu	Gly	Ser	Leu	
40					485					490					495		
	Gly	Leu	Ile	Leu	Ile	Ile	Leu	Leu	Ser	Val	Val	Val	Trp	Lys	Leu	Leu	
				500					505					510			
	Thr	Ile	Val	Ala	Ala	Asn	Arg	Asn	Arg	Met	Glu	Asn	Phe	Val	Tyr	His	
			515					520					525				
45	Asn	Ser	Ala	Phe	His	His	Pro	Arg	Ser	Asp	Leu	Ser	Glu	Lys	Asn	Gln	
		530					535					540					
	Pro	Ala	Thr	Leu	Gly	Thr	Arg										
	545					550											

//(SEQ ID NO:12);

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(vi) F₀ has the following amino acid sequence (SEQ ID NO:14):

	Leu	Asp	Pro	Ala	Ala	Leu	Met	Gln	Ile	Gly	Val	Ile	Pro	Thr	Asn	Val
	1				5					10					15	
5	Arg	Gln	Leu	Met	Tyr	Tyr	Thr	Glu	Ala	Ser	Ser	Ala	Phe	Ile	Val	Val
			20					25					30			
	Lys	Leu	Met	Pro	Thr	Ile	Asp	Ser	Pro	Ile	Ser	Gly	Cys	Asn	Ile	Thr
		35					40					45				
	Ser	Ile	Ser	Ser	Tyr	Asn	Ala	Thr	Val	Thr	Lys	Leu	Leu	Gln	Pro	Ile
		50				55						60				
10	Gly	Glu	Asn	Leu	Glu	Thr	Ile	Arg	Asn	Gln	Leu	Ile	Pro	Thr	Arg	Arg
	65				70					75					80	
	Arg	Arg	Arg	Phe	Ala	Gly	Val	Val	Ile	Gly	Leu	Ala	Ala	Leu	Gly	Val
				85					90					95		
15	Ala	Thr	Ala	Ala	Gln	Val	Thr	Ala	Ala	Val	Ala	Leu	Val	Lys	Ala	Asn
				100				105						110		
	Glu	Asn	Thr	Ala	Ala	Ile	Leu	Asn	Leu	Lys	Asn	Ala	Ile	Gln	Lys	Thr
			115					120					125			
	Asn	Ala	Ala	Val	Ala	Asp	Val	Val	Gln	Ala	Thr	Gln	Ser	Leu	Gly	Thr
		130				135						140				
20	Ala	Val	Gln	Ala	Val	Gln	Asp	His	Ile	Asn	Ser	Val	Ile	Ser	Pro	Ala
	145					150					155				160	
	Ile	Thr	Ala	Ala	Asn	Cys	Lys	Ala	Gln	Asp	Ala	Ile	Ile	Gly	Ser	Ile
				165				170						175		
25	Leu	Asn	Leu	Tyr	Leu	Thr	Glu	Leu	Thr	Thr	Ile	Phe	His	Asn	Gln	Ile
			180					185						190		
	Thr	Asn	Pro	Ala	Leu	Ser	Pro	Ile	Thr	Ile	Gln	Ala	Leu	Arg	Ile	Leu
			195				200						205			
	Leu	Gly	Ser	Thr	Leu	Pro	Thr	Val	Val	Glu	Lys	Ser	Phe	Asn	Thr	Gln
		210				215						220				
30	Ile	Ser	Ala	Ala	Glu	Leu	Ser	Ser	Gly	Leu	Leu	Thr	Gly	Gln	Ile	
	225				230					235					240	
	Val	Gly	Leu	Asp	Leu	Thr	Tyr	Met	Gln	Met	Val	Ile	Lys	Ile	Glu	Leu
				245					250					255		
35	Pro	Thr	Leu	Thr	Val	Gln	Pro	Ala	Thr	Gln	Ile	Ile	Asp	Leu	Ala	Thr
				260				265						270		
	Ile	Ser	Ala	Phe	Ile	Asn	Asn	Gln	Glu	Val	Met	Ala	Gln	Leu	Pro	Thr
			275				280						285			
	Arg	Val	Ile	Val	Thr	Gly	Ser	Leu	Ile	Gln	Ala	Tyr	Pro	Ala	Ser	Gln
		290				295						300				
40	Cys	Thr	Ile	Thr	Pro	Asn	Thr	Val	Tyr	Cys	Arg	Tyr	Asn	Asp	Ala	Gln
	305				310						315				320	
	Val	Leu	Ser	Asp	Asp	Thr	Met	Ala	Cys	Leu	Gln	Gly	Asn	Leu	Thr	Arg
				325					330					335		
45	Cys	Thr	Phe	Ser	Pro	Val	Val	Gly	Ser	Phe	Leu	Thr	Arg	Phe	Val	Leu
				340				345						350		
	Phe	Asp	Gly	Ile	Val	Tyr	Ala	Asn	Cys	Arg	Ser	Met	Leu	Cys	Lys	Cys
		355					360						365			
	Met	Gln	Pro	Ala	Ala	Val	Ile	Leu	Gln	Pro	Ser	Ser	Pro	Val	Thr	
		370				375					380					
50	Val	Ile	Asp	Met	His	Lys	Cys	Val	Ser	Leu	Gln	Leu	Asp	Asp	Leu	Arg

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35

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- 52 -

100 105 110
 Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu Arg Ile Leu Leu Gly Ser
 115 120 125
 5 Thr Leu Pro Thr Val Val Glu Lys Ser Phe Asn Thr Gln Ile Ser Ala
 130 135 140
 Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr Gly Gln Ile Val Gly Leu
 145 150 155 160
 Asp Leu Thr Tyr Met Gln Met Val Ile Lys Ile Glu Leu Pro Thr Leu
 165 170 175
 10 Thr Val Gln Pro Ala Thr Gln Ile Ile Asp Leu Ala Thr Ile Ser Ala
 180 185 190
 Phe Ile Asn Asn Gln Glu Val Met Ala Gln Leu Pro Thr Arg Val Ile
 195 200 205
 Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro Ala Ser Gln Cys Thr Ile
 210 215 220
 15 Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn Asp Ala Gln Val Leu Ser
 225 230 235 240
 Asp Asp Thr Met Ala Cys Leu Gln Gly Asn Leu Thr Arg Cys Thr Phe
 245 250 255
 20 Ser Pro Val Val Gly Ser Phe Leu Thr Arg Phe Val Leu Phe Asp Gly
 260 265 270
 Ile Val Tyr Ala Asn Cys Arg Ser Met Leu Cys Lys Cys Met Gln Pro
 275 280 285
 25 Ala Ala Val Ile Leu Gln Pro Ser Ser Ser Pro Val Thr Val Ile Asp
 290 295 300
 Met His Lys Cys Val Ser Leu Gln Leu Asp Asp Leu Arg Phe Thr Ile
 305 310 315 320
 Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser Thr Ile Lys Leu Glu Thr
 325 330 335
 30 Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp Ile Ser Gln Asn Leu Ala
 340 345 350
 Ala Val Asn Lys Ser Leu Ser Asp Ala Leu Gln His Leu Ala Gln Ser
 355 360 365
 35 Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala Thr Thr Thr Ser Val Leu
 370 375 380
 Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu Gly Leu Ile Leu Ile Ile
 385 390 395 400
 Leu Leu Ser Val Val Val Trp Lys Leu Leu Thr Ile Val Ala Ala Asn
 405 410 415
 40 Arg Asn Arg Met Glu Asn Phe Val Tyr His Asn Ser Ala Phe His His
 420 425 430
 Pro Arg Ser Asp Leu Ser Glu Lys Asn Gln Pro Ala Thr Leu Gly Thr
 435 440 445
 Arg
 45 //(SEQ ID NO:18);

(ix) SH has the following amino acid sequence (SEQ ID NO:20):

Met Leu Pro Asp Pro Glu Asp Pro Glu Ser Lys Lys Ala Thr Arg Arg
 1 5 10 15

Thr Gly Asn Leu Ile Ile Cys Phe Leu Phe Ile Phe Phe Leu Phe Val
 20 25 30
 Thr Leu Ile Val Pro Thr Leu Arg His Leu Leu Ser
 35 40 // (SEQ ID NO:20);

5

(x) HN has the following amino acid sequence (SEQ ID NO:22):

Met Ile Ala Glu Asp Ala Pro Val Lys Gly Thr Cys Arg Val Leu Phe
 1 5 10 15
 Arg Thr Thr Thr Leu Ile Phe Leu Cys Thr Leu Leu Ala Leu Ser Ile
 10 20 25 30
 Ser Ile Leu Tyr Glu Ser Leu Ile Thr Gln Lys Gln Ile Met Ser Gln
 35 40 45
 Ala Gly Ser Thr Gly Ser Asn Ser Gly Leu Gly Gly Ile Thr Asp Leu
 50 55 60
 15 Leu Asn Asn Ile Leu Ser Val Ala Asn Gln Ile Ile Tyr Asn Ser Ala
 65 70 75 80
 Val Ala Leu Pro Leu Gln Leu Asp Thr Leu Glu Ser Thr Leu Leu Thr
 85 90 95
 20 Ala Ile Lys Ser Leu Gln Thr Ser Asp Lys Leu Glu Gln Asn Cys Ser
 100 105 110
 Trp Gly Ala Ala Leu Ile Asn Asp Asn Arg Tyr Ile Asn Gly Ile Asn
 115 120 125
 Gln Phe Tyr Phe Ser Ile Ala Glu Gly Arg Asn Leu Thr Leu Gly Pro
 130 135 140
 25 Leu Leu Asn Ile Pro Ser Phe Ile Pro Thr Ala Thr Thr Pro Glu Gly
 145 150 155 160
 Cys Thr Arg Ile Pro Ser Phe Ser Leu Thr Lys Thr His Trp Cys Tyr
 165 170 175
 30 Thr His Asn Val Ile Leu Asn Gly Cys Gln Asp His Val Ser Ser Asn
 180 185 190
 Gln Phe Val Ser Met Gly Ile Ile Glu Pro Thr Ser Ala Gly Phe Pro
 195 200 205
 Ser Phe Arg Thr Leu Lys Thr Leu Tyr Leu Ser Asp Gly Val Asn Arg
 210 215 220
 35 Lys Ser Cys Ser Ile Ser Thr Val Pro Gly Gly Cys Met Met Tyr Cys
 225 230 235 240
 Phe Val Ser Thr Gln Pro Glu Arg Asp Asp Tyr Phe Ser Thr Ala Pro
 245 250 255
 40 Pro Glu Gln Arg Ile Ile Ile Met Tyr Tyr Asn Asp Thr Ile Val Glu
 260 265 270
 Arg Ile Ile Asn Pro Pro Gly Val Leu Asp Val Trp Ala Thr Leu Asn
 275 280 285
 Pro Gly Thr Gly Ser Gly Val Tyr Tyr Leu Gly Trp Val Leu Phe Pro
 290 295 300
 45 Ile Tyr Gly Gly Val Ile Lys Asn Thr Ser Leu Trp Asn Asn Gln Ala
 305 310 315 320
 Asn Lys Tyr Phe Ile Pro Gln Met Val Ala Ala Leu Cys Ser Gln Asn
 325 330 335
 50 Gln Ala Thr Gln Val Gln Asn Ala Lys Ser Ser Tyr Tyr Ser Ser Trp
 340 345 350

Phe Gly Asn Arg Met Ile Gln Ser Gly Ile Leu Ala Cys Pro Leu Gln
 355 360 365
 Gln Asp Leu Thr Asn Glu Cys Leu Val Leu Pro Phe Ser Asn Asp Gln
 370 375 380
 5 Val Leu Met Gly Ala Glu Gly Arg Leu Tyr Met Tyr Gly Asp Ser Val
 385 390 395 400
 Tyr Tyr Tyr Gln Arg Ser Asn Ser Trp Trp Pro Met Thr Met Leu Tyr
 405 410 415
 10 Lys Val Thr Ile Thr Phe Thr Asn Gly Gln Pro Ser Ala Ile Ser Ala
 420 425 430
 Gln Asn Val Pro Thr Gln Gln Val Pro Arg Pro Gly Thr Gly Asp Cys
 435 440 445
 Phe Ala Thr Asn Arg Cys Pro Gly Phe Cys Leu Thr Gly Val Tyr Ala
 450 455 460
 15 Asp Ala Trp Leu Leu Thr Asn Pro Ser Ser Thr Ser Thr Phe Gly Ser
 465 470 475 480
 Glu Ala Thr Phe Thr Gly Ser Tyr Leu Asn Ala Ala Thr Gln Arg Ile
 485 490 495
 20 Asn Pro Thr Met Tyr Ile Ala Asn Asn Thr Gln Ile Ile Ser Ser Gln
 500 505 510
 Gln Phe Gly Ser Ser Gly Gln Glu Ala Ala Tyr Gly His Thr Thr Cys
 515 520 525
 Phe Arg Asp Thr Gly Ser Val Met Val Tyr Cys Ile Tyr Ile Ile Glu
 530 535 540
 25 Leu Ser Ser Ser Leu Leu Gly Gln Phe Gln Ile Val Pro Phe Ile Arg
 545 550 555 560
 Gln Val Thr Leu Ser
 565 //(SEQ ID NO:22);

30 and

(xi) L has the following amino acid sequence (SEQ ID NO:24):

Met Ala Gly Ser Arg Glu Ile Leu Leu Pro Glu Val His Leu Asn Ser
 1 5 10 15
 35 Pro Ile Val Lys His Lys Leu Tyr Tyr Tyr Ile Leu Leu Gly Asn Leu
 20 25 30
 Pro Asn Glu Ile Asp Ile Asp Asp Leu Gly Pro Leu His Asn Gln Asn
 35 40 45
 Trp Asn Gln Ile Ala His Glu Glu Ser Asn Leu Ala Gln Arg Leu Val
 50 55 60
 40 Asn Val Arg Asn Phe Leu Ile Thr His Ile Pro Asp Leu Arg Lys Gly
 65 70 75 80
 His Trp Gln Glu Tyr Val Asn Val Ile Leu Trp Pro Arg Ile Leu Pro
 85 90 95
 45 Leu Ile Pro Asp Phe Lys Ile Asn Asp Gln Leu Pro Leu Leu Lys Asn
 100 105 110
 Trp Asp Lys Leu Val Lys Glu Ser Cys Ser Val Ile Asn Ala Gly Thr
 115 120 125
 Ser Gln Cys Ile Gln Asn Leu Ser Tyr Gly Leu Thr Gly Arg Gly Asn
 130 135 140

	Leu	Phe	Thr	Arg	Ser	Arg	Glu	Leu	Ser	Gly	Asp	Arg	Arg	Asp	Ile	Asp
	145					150					155					160
	Leu	Lys	Thr	Val	Val	Ala	Ala	Trp	His	Asp	Ser	Asp	Trp	Lys	Arg	Ile
					165					170					175	
5	Ser	Asp	Phe	Trp	Ile	Met	Ile	Lys	Phe	Gln	Met	Arg	Gln	Leu	Ile	Val
				180					185					190		
	Arg	Gln	Thr	Asp	His	Asn	Asp	Pro	Asp	Leu	Ile	Thr	Tyr	Ile	Glu	Asn
			195					200					205			
10	Arg	Glu	Gly	Ile	Ile	Ile	Ile	Thr	Pro	Glu	Leu	Val	Ala	Leu	Phe	Asn
		210					215					220				
	Thr	Glu	Asn	His	Thr	Leu	Thr	Tyr	Met	Thr	Phe	Glu	Ile	Val	Leu	Met
	225					230					235					240
	Val	Ser	Asp	Met	Tyr	Glu	Gly	Arg	His	Asn	Ile	Leu	Ser	Leu	Cys	Thr
				245						250					255	
15	Val	Ser	Thr	Tyr	Leu	Asn	Pro	Leu	Lys	Lys	Arg	Ile	Thr	Tyr	Leu	Leu
				260					265					270		
	Ser	Leu	Val	Asp	Asn	Leu	Ala	Phe	Gln	Ile	Gly	Asp	Ala	Val	Tyr	Asn
			275					280					285			
20	Ile	Ile	Ala	Leu	Leu	Glu	Ser	Phe	Val	Tyr	Ala	Gln	Leu	Gln	Met	Ser
		290					295					300				
	Asp	Pro	Ile	Pro	Glu	Leu	Arg	Gly	Gln	Phe	His	Ala	Phe	Val	Cys	Ser
	305					310					315					320
	Glu	Ile	Leu	Asp	Ala	Leu	Arg	Gly	Thr	Asn	Ser	Phe	Thr	Gln	Asp	Glu
				325						330					335	
25	Leu	Arg	Thr	Val	Thr	Thr	Asn	Leu	Ile	Ser	Pro	Phe	Gln	Asp	Leu	Thr
				340					345					350		
	Pro	Asp	Leu	Thr	Ala	Glu	Leu	Leu	Cys	Ile	Met	Arg	Leu	Trp	Gly	His
			355					360					365			
30	Pro	Met	Leu	Thr	Ala	Ser	Gln	Ala	Ala	Gly	Lys	Val	Arg	Glu	Ser	Met
		370					375					380				
	Cys	Ala	Gly	Lys	Val	Leu	Asp	Phe	Pro	Thr	Ile	Met	Lys	Thr	Leu	Ala
	385					390					395					400
	Phe	Phe	His	Thr	Ile	Leu	Ile	Asn	Gly	Tyr	Arg	Arg	Lys	His	His	Gly
				405						410				415		
35	Val	Trp	Pro	Pro	Leu	Asn	Leu	Pro	Gly	Asn	Ala	Ser	Lys	Gly	Leu	Thr
				420					425					430		
	Glu	Leu	Met	Asn	Asp	Asn	Thr	Glu	Ile	Ser	Tyr	Glu	Phe	Thr	Leu	Lys
			435					440					445			
40	His	Trp	Lys	Glu	Ile	Ser	Leu	Ile	Lys	Phe	Lys	Lys	Cys	Phe	Asp	Ala
		450					455					460				
	Asp	Ala	Gly	Glu	Glu	Leu	Ser	Ile	Phe	Met	Lys	Asp	Lys	Ala	Ile	Ser
	465					470					475					480
	Ala	Pro	Lys	Gln	Asp	Trp	Met	Ser	Val	Phe	Arg	Arg	Ser	Leu	Ile	Lys
				485						490					495	
45	Gln	Arg	His	Gln	His	His	Gln	Val	Pro	Leu	Pro	Asn	Pro	Phe	Asn	Arg
				500					505					510		
	Arg	Leu	Leu	Leu	Asn	Phe	Leu	Gly	Asp	Asp	Lys	Phe	Asp	Pro	Asn	Val
			515					520					525			
50	Glu	Leu	Gln	Tyr	Val	Thr	Ser	Gly	Glu	Tyr	Leu	His	Asp	Asp	Thr	Phe
		530					535					540				
	Cys	Ala	Ser	Tyr	Ser	Leu	Lys	Glu	Lys	Glu	Ile	Lys	Pro	Asp	Gly	Arg

	545				550				555				560			
	Ile	Phe	Ala	Lys	Leu	Thr	Lys	Arg	Met	Arg	Ser	Cys	Gln	Val	Ile	Ala
					565					570					575	
5	Glu	Ser	Leu	Leu	Ala	Asn	His	Ala	Gly	Lys	Leu	Met	Lys	Glu	Asn	Gly
				580					585					590		
	Val	Val	Met	Asn	Gln	Leu	Ser	Leu	Thr	Lys	Ser	Leu	Leu	Thr	Met	Ser
			595					600				605				
	Gln	Ile	Gly	Ile	Ile	Ser	Glu	Arg	Ala	Arg	Lys	Ser	Thr	Arg	Asp	Asn
	610						615					620				
10	Ile	Asn	Arg	Pro	Gly	Phe	Gln	Asn	Ile	Gln	Arg	Asn	Lys	Ser	His	His
	625					630					635				640	
	Ser	Lys	Gln	Val	Asn	Gln	Arg	Asp	Pro	Ser	Asp	Asp	Phe	Glu	Leu	Ala
				645					650					655		
15	Ala	Ser	Phe	Leu	Thr	Thr	Asp	Leu	Lys	Lys	Tyr	Cys	Leu	Gln	Trp	Arg
				660					665					670		
	Tyr	Gln	Thr	Ile	Ile	Pro	Phe	Ala	Gln	Ser	Leu	Asn	Arg	Met	Tyr	Gly
			675					680					685			
	Tyr	Pro	His	Leu	Phe	Glu	Trp	Ile	His	Leu	Arg	Leu	Met	Arg	Ser	Thr
	690						695					700				
20	Leu	Tyr	Val	Gly	Asp	Pro	Phe	Asn	Pro	Pro	Ala	Asp	Thr	Ser	Gln	Phe
	705					710					715				720	
	Asp	Leu	Asp	Lys	Val	Ile	Asn	Gly	Asp	Ile	Phe	Ile	Val	Ser	Pro	Arg
				725						730				735		
25	Gly	Gly	Ile	Glu	Gly	Leu	Cys	Gln	Lys	Ala	Trp	Thr	Met	Ile	Ser	Ile
				740					745					750		
	Ser	Val	Ile	Ile	Leu	Ser	Ala	Thr	Glu	Ser	Gly	Thr	Arg	Val	Met	Ser
			755					760					765			
	Met	Val	Gln	Gly	Asp	Asn	Gln	Ala	Ile	Ala	Val	Thr	Thr	Arg	Val	Pro
	770						775					780				
30	Arg	Ser	Leu	Pro	Thr	Leu	Glu	Lys	Lys	Thr	Ile	Ala	Phe	Arg	Ser	Cys
	785					790					795				800	
	Asn	Leu	Phe	Phe	Glu	Arg	Leu	Lys	Cys	Asn	Asn	Phe	Gly	Leu	Gly	His
				805						810				815		
35	His	Leu	Lys	Glu	Gln	Glu	Thr	Ile	Ile	Ser	Ser	His	Phe	Phe	Val	Tyr
				820					825					830		
	Ser	Lys	Arg	Ile	Phe	Tyr	Gln	Gly	Arg	Ile	Leu	Thr	Gln	Ala	Leu	Lys
			835					840					845			
	Asn	Ala	Ser	Lys	Leu	Cys	Leu	Thr	Ala	Asp	Val	Leu	Gly	Glu	Cys	Thr
	850						855					860				
40	Gln	Ser	Ser	Cys	Ser	Asn	Leu	Ala	Thr	Thr	Val	Met	Arg	Leu	Thr	Glu
	865					870					875				880	
	Asn	Gly	Val	Glu	Lys	Asp	Ile	Cys	Phe	Tyr	Leu	Asn	Ile	Tyr	Met	Thr
				885						890				895		
45	Ile	Lys	Gln	Leu	Ser	Tyr	Asp	Ile	Ile	Phe	Pro	Gln	Val	Ser	Ile	Pro
				900					905					910		
	Gly	Asp	Gln	Ile	Thr	Leu	Glu	Tyr	Ile	Asn	Asn	Pro	His	Leu	Val	Ser
		915						920					925			
	Arg	Leu	Ala	Leu	Leu	Pro	Ser	Gln	Leu	Gly	Gly	Leu	Asn	Tyr	Leu	Ser
		930					935					940				
50	Cys	Ser	Arg	Leu	Phe	Asn	Arg	Asn	Ile	Gly	Asp	Pro	Val	Val	Ser	Ala
	945					950					955					960

	Val	Ala	Asp	Leu	Lys	Arg	Leu	Ile	Lys	Ser	Gly	Cys	Met	Asp	Tyr	Trp
				965						970					975	
	Ile	Leu	Tyr	Asn	Leu	Leu	Gly	Arg	Lys	Pro	Gly	Asn	Gly	Ser	Trp	Ala
			980						985					990		
5	Thr	Leu	Ala	Ala	Asp	Pro	Tyr	Ser	Ile	Asn	Ile	Glu	Tyr	Gln	Tyr	Pro
		995						1000					1005			
	Pro	Thr	Thr	Ala	Leu	Lys	Arg	His	Thr	Gln	Gln	Val	Leu	Met	Glu	Leu
		1010					1015					1020				
	Ser	Thr	Asn	Pro	Met	Leu	Arg	Gly	Ile	Phe	Ser	Asp	Asn	Ala	Gln	Ala
10		1025				1030					1035					1040
	Glu	Glu	Asn	Asn	Leu	Ala	Arg	Phe	Leu	Leu	Asp	Arg	Glu	Val	Ile	Phe
				1045						1050					1055	
	Pro	Arg	Val	Ala	His	Ile	Ile	Ile	Glu	Gln	Thr	Ser	Val	Gly	Arg	Arg
				1060					1065					1070		
15	Lys	Gln	Ile	Gln	Gly	Tyr	Leu	Asp	Ser	Thr	Arg	Ser	Ile	Met	Arg	Lys
		1075						1080					1085			
	Ser	Leu	Glu	Ile	Lys	Pro	Leu	Ser	Asn	Arg	Lys	Leu	Asn	Glu	Ile	Leu
		1090					1095					1100				
	Asp	Tyr	Asn	Ile	Asn	Tyr	Leu	Ala	Tyr	Asn	Leu	Ala	Leu	Leu	Lys	Asn
20		1105				1110					1115					1120
	Ala	Ile	Glu	Pro	Pro	Thr	Tyr	Leu	Lys	Ala	Met	Thr	Leu	Glu	Thr	Cys
				1125						1130					1135	
	Ser	Ile	Asp	Ile	Ala	Arg	Ser	Leu	Arg	Lys	Leu	Ser	Trp	Ala	Pro	Leu
				1140					1145					1150		
25	Leu	Gly	Gly	Arg	Asn	Leu	Glu	Gly	Leu	Glu	Thr	Pro	Asp	Pro	Ile	Glu
		1155						1160					1165			
	Ile	Thr	Ala	Gly	Ala	Leu	Ile	Val	Gly	Ser	Gly	Tyr	Cys	Glu	Gln	Cys
		1170					1175					1180				
	Ala	Ala	Gly	Asp	Asn	Arg	Phe	Thr	Trp	Phe	Phe	Leu	Pro	Ser	Gly	Ile
30		1185				1190					1195					1200
	Glu	Ile	Gly	Gly	Asp	Pro	Arg	Asp	Asn	Pro	Pro	Ile	Arg	Val	Pro	Tyr
				1205					1210					1215		
	Ile	Gly	Ser	Arg	Thr	Asp	Glu	Arg	Arg	Val	Ala	Ser	Met	Ala	Tyr	Ile
				1220				1225					1230			
35	Arg	Gly	Ala	Ser	Ser	Ser	Leu	Lys	Ala	Val	Leu	Arg	Leu	Ala	Gly	Val
		1235						1240					1245			
	Tyr	Ile	Trp	Ala	Phe	Gly	Asp	Thr	Leu	Glu	Asn	Trp	Ile	Asp	Ala	Leu
		1250					1255					1260				
	Asp	Leu	Ser	His	Thr	Arg	Val	Asn	Ile	Thr	Leu	Glu	Gln	Leu	Gln	Ser
40		1265				1270					1275					1280
	Leu	Thr	Pro	Leu	Pro	Thr	Ser	Ala	Asn	Leu	Thr	His	Arg	Leu	Asp	Asp
				1285						1290				1295		
	Gly	Thr	Thr	Thr	Leu	Lys	Phe	Thr	Pro	Ala	Ser	Ser	Tyr	Thr	Phe	Ser
				1300					1305					1310		
45	Ser	Phe	Thr	His	Ile	Ser	Asn	Asp	Glu	Gln	Tyr	Leu	Thr	Ile	Asn	Asp
		1315					1320						1325			
	Lys	Thr	Ala	Asp	Ser	Asn	Ile	Ile	Tyr	Gln	Gln	Leu	Met	Ile	Thr	Gly
		1330					1335					1340				
	Leu	Gly	Ile	Leu	Glu	Thr	Trp	Asn	Asn	Pro	Pro	Ile	Asn	Arg	Thr	Phe
50		1345				1350				1355						1360
	Glu	Glu	Ser	Thr	Leu	His	Leu	His	Thr	Gly	Ala	Ser	Cys	Cys	Val	Arg

				1365				1370				1375				
	Pro	Val	Asp	Ser	Cys	Ile	Ile	Ser	Glu	Ala	Leu	Thr	Val	Lys	Pro	His
				1380					1385				1390			
5	Ile	Thr	Val	Pro	Tyr	Ser	Asn	Lys	Phe	Val	Phe	Asp	Glu	Asp	Pro	Leu
			1395					1400					1405			
	Ser	Glu	Tyr	Glu	Thr	Ala	Lys	Leu	Glu	Ser	Leu	Ser	Phe	Gln	Ala	Gln
		1410					1415					1420				
	Leu	Gly	Asn	Ile	Asp	Ala	Val	Asp	Met	Thr	Gly	Lys	Leu	Thr	Leu	Leu
	1425				1430						1435				1440	
10	Ser	Gln	Phe	Thr	Ala	Arg	Gln	Ile	Ile	Asn	Ala	Ile	Thr	Gly	Leu	Asp
				1445						1450					1455	
	Glu	Ser	Val	Ser	Leu	Thr	Asn	Asp	Ala	Ile	Val	Ala	Ser	Asp	Tyr	Val
				1460					1465					1470		
15	Ser	Asn	Trp	Ile	Ser	Glu	Cys	Met	Tyr	Thr	Lys	Leu	Asp	Glu	Leu	Phe
		1475						1480						1485		
	Met	Tyr	Cys	Gly	Trp	Glu	Leu	Leu	Leu	Glu	Leu	Ser	Tyr	Gln	Met	Tyr
		1490					1495						1500			
	Tyr	Leu	Arg	Val	Val	Gly	Trp	Ser	Asn	Ile	Val	Asp	Tyr	Ser	Tyr	Met
	1505					1510						1515			1520	
20	Ile	Leu	Arg	Arg	Ile	Pro	Gly	Ala	Ala	Leu	Asn	Asn	Leu	Ala	Ser	Thr
					1525						1530				1535	
	Leu	Ser	His	Pro	Lys	Leu	Phe	Arg	Arg	Ala	Ile	Asn	Leu	Asp	Ile	Val
				1540					1545					1550		
25	Ala	Pro	Leu	Asn	Ala	Pro	His	Phe	Ala	Ser	Leu	Asp	Tyr	Ile	Lys	Met
		1555						1560					1565			
	Ser	Met	Asp	Ala	Ile	Leu	Trp	Gly	Cys	Lys	Arg	Val	Ile	Asn	Val	Leu
		1570					1575					1580				
	Ser	Asn	Gly	Gly	Asp	Leu	Glu	Leu	Val	Val	Thr	Ser	Glu	Asp	Ser	Leu
	1585				1590						1595				1600	
30	Ile	Leu	Ser	Asp	Arg	Ser	Met	Asn	Leu	Ile	Ala	Arg	Lys	Leu	Thr	Leu
				1605							1610				1615	
	Leu	Ser	Leu	Ile	His	His	Asn	Gly	Leu	Glu	Leu	Pro	Lys	Ile	Lys	Gly
				1620					1625					1630		
35	Phe	Ser	Pro	Asp	Glu	Lys	Cys	Phe	Ala	Leu	Thr	Glu	Phe	Leu	Arg	Lys
		1635						1640					1645			
	Val	Val	Asn	Ser	Gly	Leu	Ser	Ser	Ile	Glu	Asn	Leu	Ser	Asn	Phe	Met
		1650					1655					1660				
	Tyr	Asn	Val	Glu	Asn	Pro	Arg	Leu	Ala	Ala	Phe	Ala	Ser	Asn	Asn	Tyr
	1665				1670						1675				1680	
40	Tyr	Leu	Thr	Arg	Lys	Leu	Leu	Asn	Ser	Ile	Arg	Asp	Thr	Glu	Ser	Gly
				1685							1690				1695	
	Gln	Val	Ala	Val	Thr	Ser	Tyr	Tyr	Glu	Ser	Leu	Glu	Tyr	Ile	Asp	Ser
				1700					1705					1710		
45	Leu	Lys	Leu	Thr	Pro	His	Val	Pro	Gly	Thr	Ser	Cys	Ile	Glu	Asp	Asp
		1715						1720					1725			
	Ser	Leu	Cys	Thr	Asn	Asp	Tyr	Ile	Ile	Trp	Ile	Ile	Glu	Ser	Asn	Ala
		1730				1735						1740				
	Asn	Leu	Glu	Lys	Tyr	Pro	Ile	Pro	Asn	Ser	Pro	Glu	Asp	Asp	Ser	Asn
	1745				1750					1755					1760	
50	Phe	His	Asn	Phe	Lys	Leu	Asn	Ala	Pro	Ser	His	His	Thr	Leu	Arg	Pro
				1765						1770					1775	

	Leu	Gly	Leu	Ser	Ser	Thr	Ala	Trp	Tyr	Lys	Gly	Ile	Ser	Cys	Cys	Arg
				1780					1785					1790		
	Tyr	Leu	Glu	Arg	Leu	Lys	Leu	Pro	Gln	Gly	Asp	His	Leu	Tyr	Ile	Ala
			1795					1800					1805			
5	Glu	Gly	Ser	Gly	Ala	Ser	Met	Thr	Ile	Ile	Glu	Tyr	Leu	Phe	Pro	Gly
		1810					1815					1820				
	Arg	Lys	Ile	Tyr	Tyr	Asn	Ser	Leu	Phe	Ser	Ser	Gly	Asp	Asn	Pro	Pro
	1825					1830					1835					1840
10	Gln	Arg	Asn	Tyr	Ala	Pro	Met	Pro	Thr	Gln	Phe	Ile	Glu	Ser	Val	Pro
					1845						1850				1855	
	Tyr	Lys	Leu	Trp	Gln	Ala	His	Thr	Asp	Gln	Tyr	Pro	Glu	Ile	Phe	Glu
				1860					1865					1870		
	Asp	Phe	Ile	Pro	Leu	Trp	Asn	Gly	Asn	Ala	Ala	Met	Thr	Asp	Ile	Gly
			1875					1880					1885			
15	Met	Thr	Ala	Cys	Val	Glu	Phe	Ile	Ile	Asn	Arg	Val	Gly	Pro	Arg	Thr
		1890						1895					1900			
	Cys	Ser	Leu	Val	His	Val	Asp	Leu	Glu	Ser	Ser	Ala	Ser	Leu	Asn	Gln
	1905					1910					1915					1920
20	Gln	Cys	Leu	Ser	Lys	Pro	Ile	Ile	Asn	Ala	Ile	Ile	Thr	Ala	Thr	Thr
					1925					1930					1935	
	Val	Leu	Cys	Pro	His	Gly	Val	Leu	Ile	Leu	Lys	Tyr	Ser	Trp	Leu	Pro
				1940					1945					1950		
	Phe	Thr	Arg	Phe	Ser	Thr	Leu	Ile	Thr	Phe	Leu	Trp	Cys	Tyr	Phe	Glu
			1955					1960					1965			
25	Arg	Ile	Thr	Val	Leu	Arg	Ser	Thr	Tyr	Ser	Asp	Pro	Ala	Asn	His	Glu
		1970					1975					1980				
	Val	Tyr	Leu	Ile	Cys	Ile	Leu	Ala	Asn	Asn	Phe	Ala	Phe	Gln	Thr	Val
	1985					1990					1995					2000
30	Ser	Gln	Ala	Thr	Gly	Met	Ala	Met	Thr	Leu	Thr	Asp	Gln	Gly	Phe	Thr
					2005					2010					2015	
	Leu	Ile	Ser	Pro	Glu	Arg	Ile	Asn	Gln	Tyr	Trp	Asp	Gly	His	Leu	Lys
				2020					2025					2030		
	Gln	Glu	Arg	Ile	Val	Ala	Glu	Ala	Ile	Asp	Lys	Val	Val	Leu	Gly	Glu
				2035				2040					2045			
35	Asn	Ala	Leu	Phe	Asn	Ser	Ser	Asp	Asn	Glu	Leu	Ile	Leu	Lys	Cys	Gly
		2050					2055					2060				
	Gly	Thr	Pro	Asn	Ala	Arg	Asn	Leu	Ile	Asp	Ile	Glu	Pro	Val	Ala	Thr
	2065					2070					2075					2080
40	Phe	Ile	Glu	Phe	Glu	Gln	Leu	Ile	Cys	Thr	Met	Leu	Thr	Thr	His	Leu
					2085					2090					2095	
	Lys	Glu	Ile	Ile	Asp	Ile	Thr	Arg	Ser	Gly	Thr	Gln	Asp	Tyr	Glu	Ser
				2100					2105					2110		
	Leu	Leu	Leu	Thr	Pro	Tyr	Asn	Leu	Gly	Leu	Leu	Gly	Lys	Ile	Ser	Thr
				2115				2120					2125			
45	Ile	Val	Arg	Leu	Leu	Thr	Glu	Arg	Ile	Leu	Asn	His	Thr	Ile	Arg	Asn
		2130					2135					2140				
	Trp	Leu	Ile	Leu	Pro	Pro	Ser	Leu	Gln	Met	Ile	Val	Lys	Gln	Asp	Leu
	2145					2150					2155					2160
50	Glu	Phe	Gly	Ile	Phe	Arg	Ile	Thr	Ser	Ile	Leu	Asn	Ser	Asp	Arg	Phe
					2165					2170					2175	
	Leu	Lys	Leu	Ser	Pro	Asn	Arg	Lys	Tyr	Leu	Ile	Thr	Gln	Leu	Thr	Ala

The inventive protein also encompasses a polypeptide having substantially the same amino acid sequence as (SEQ ID NO:4), (SEQ ID NO:6) (SEQ ID NO:8), (SEQ ID NO:10), (SEQ ID NO:12), (SEQ ID NO:14), (SEQ ID NO:16), (SEQ ID NO:18), (SEQ ID NO:20), (SEQ ID NO:22), or (SEQ ID NO:24). As employed herein, the term "substantially the same amino acid sequence" refers to amino acid sequences having at least about 80%, still more preferably about 90% amino acid identity with respect to a reference amino acid sequence; with greater than about 95% amino acid sequence identity being especially preferred. It is recognized, however, that polypeptide containing less than the described levels of sequence identity arising as splice variants or that are modified by conservative amino acid substitutions are also encompassed within the scope of the present invention. The degree of sequence homology is determined by conducting an amino acid sequence similarity search of a protein data base, such as the database of the National Center for Biotechnology Information (NCBI; www.ncbi.nlm.nih.gov/BLAST/), using a computerized algorithm, such as PowerBLAST, QBLAST, PSI-BLAST, PHI-BLAST, gapped or ungapped BLAST, or the "Align" program through the Baylor College of Medicine server (www.hgsc.bcm.tmc.edu/seq_data). (E.g., Altschul, S.F., *et al.*, *Gapped BLAST and PSI-BLAST: a new generation of protein database search programs*, Nucleic Acids Res. 25(17):3389-402 [1997]; Zhang, J., & Madden, T.L., *PowerBLAST: a new network BLAST application for interactive or automated sequence analysis and annotation*, Genome Res. 7(6):649-56 [1997]; Madden, T.L., *et al.*, *Applications of network BLAST server*, Methods Enzymol. 266:131-41 [1996]; Altschul, S.F., *et al.*, *Basic local alignment search tool*, J. Mol. Biol. 215(3):403-10 [1990]).

Also encompassed by the term *Cryptovirus* protein, are biologically functional or active peptide analogs thereof. The term peptide "analog" includes any polypeptide having an amino acid residue sequence substantially identical to a sequence specifically shown herein in which one or more residues have been conservatively substituted with a functionally similar residue and which displays the ability to mimic the biological activity of a native *Cryptovirus* protein.

Examples of conservative substitutions include the substitution of one non-polar (hydrophobic) residue such as isoleucine, valine, leucine or methionine for another, the substitution of one polar (hydrophilic) residue for another such as between arginine and lysine, between glutamine and asparagine, between glycine and serine, the substitution of one basic residue such as lysine, arginine or histidine for another, or the substitution of one acidic residue, such as aspartic acid or glutamic acid for another.

The phrase "conservative substitution" also includes the use of a chemically derivatized residue in place of a non-derivatized residue, provided that such polypeptide displays the requisite biological activity.

"Chemical derivative" refers to a subject polypeptide having one or more residues chemically derivatized by reaction of a functional side group. Such derivatized molecules include, for example, those molecules in which free amino groups have been derivatized to form amine hydrochlorides, p-toluene sulfonyl groups, carbobenzoxy groups, t-butyloxycarbonyl groups, chloroacetyl groups or formyl groups. Free carboxyl groups may be derivatized to form salts, methyl and ethyl esters or other types of esters or hydrazides. Free hydroxyl groups may be derivatized to form O-acyl or O-alkyl derivatives. The imidazole nitrogen of histidine may be derivatized to form N-im-benzylhistidine. Also included as chemical derivatives are those peptides which contain one or more naturally occurring amino acid derivatives of the twenty standard amino acids. For example, 4-hydroxyproline may be substituted for proline; 5-hydroxylysine may be substituted for lysine; 3-methylhistidine may be substituted for histidine; homoserine may be substituted for serine; and ornithine may be substituted for lysine. The inventive polypeptide also includes any polypeptide having one or more additions and/or deletions of residues, relative to the sequence of an inventive polypeptide whose sequence is shown herein, so long as the requisite biological activity is maintained.

The present invention also encompasses a variant of a *Cryptovirus* protein designated by (SEQ ID NO:4), (SEQ ID NO:6) (SEQ ID NO:8), (SEQ ID NO:10), (SEQ ID NO:12), (SEQ ID NO:14), (SEQ ID NO:16), (SEQ ID NO:18), (SEQ ID NO:20), (SEQ ID NO:22), or (SEQ ID NO:24); a "variant" refers to a polypeptide in which the amino acid sequence of the designated *Cryptovirus* protein has been altered by the deletion, substitution, addition or rearrangement of one or more amino acids in the sequence. Methods by which variants occur (for example, by recombination) or are made (for example, by site directed mutagenesis) are known in the art.

The *Cryptovirus* protein can also include one or more labels, which are known to those of skill in the art.

The inventive proteins are isolated or purified by a variety of known biochemical means, including, for example, by the recombinant expression systems described herein, precipitation, gel filtration, ion-exchange, reverse-phase and affinity chromatography, electrophoresis, and the like. Other well-known methods are described in Deutscher *et al.*, *Guide to Protein Purification: Methods in Enzymology* Vol. 182, (Academic Press, [1990]).

The isolated *Cryptovirus* protein of the present invention can also be chemically synthesized. For example, synthetic polypeptide can be produced using Applied Biosystems, Inc. Model 430A or 431A automatic peptide synthesizer (Foster City, CA) employing the chemistry provided by the manufacturer and the amino acid sequences provided herein. Alternatively, the *Cryptovirus* protein can be isolated or purified from native cellular sources. Alternatively, the *Cryptovirus* protein can be isolated from the inventive chimeric proteins by the use of suitable proteases.

Alternatively, the *Cryptovirus* proteins can be recombinantly derived, for example, produced by eukaryotic or prokaryotic cells genetically modified to express *Cryptovirus* protein-encoding polynucleotides in accordance with the inventive technology as described herein. Recombinant methods are well known, as described, for example, in Sambrook *et al.*, *supra.*, 1989). An example of the means for preparing the inventive *Cryptovirus* protein is to express nucleic acids encoding the *Cryptovirus* protein of interest in a suitable host cell that contains the inventive expression vector, such as a bacterial cell, a yeast cell, an insect cell, an amphibian cell (i.e., oocyte), or a mammalian cell, using methods well known in the art, and recovering the expressed polypeptide, again using well-known methods.

"Recombinant host cells", "host cells", "cells", "cell lines", "cell cultures", and other such terms denoting prokaryotic or eukaryotic cell lines cultured as unicellular or monolayer entities, and refer to cells which can be, or have been, used as recipients for a recombinant expression vector or other foreign nucleic acids, such as DNA or RNA, and include the progeny of the original cell which has been transfected. It is understood that the progeny of a single parental cell may not necessarily be completely identical in morphology or in genomic or total DNA complement as the original parent, due to natural, accidental, or deliberate mutation.

The present invention includes chimeric proteins. The term "chimeric protein" generally refers to a polypeptide comprising an amino acid sequence drawn from two or more individual proteins that are not naturally so linked. In the present invention, "chimeric protein" is used to denote a polypeptide comprising a *Cryptovirus* protein, or a truncate or polypeptide variant thereof having an altered amino acid sequence, fused to a non-*Cryptovirus* protein or polypeptide moiety.

Chimeric proteins are most conveniently produced by expression of a fused gene, which encodes a portion of one polypeptide at the 5' end and a portion of a different polypeptide at the 3' end, where the different portions are joined in one reading frame which may be expressed in a suitable host cell. In some embodiments, the *Cryptovirus* protein is positioned at the carboxy terminus of the chimeric protein. In other embodiments the *Cryptovirus* protein is positioned at the amino terminus of the chimeric protein.

The non-*Cryptovirus* protein moiety, or "chimeric partner", of the inventive chimeric protein can be a functional enzyme fragment. Suitable functional enzyme fragments are those polypeptides which exhibit a quantifiable activity when expressed fused to the *Cryptovirus* protein. Exemplary enzymes include, without limitation, β -galactosidase (β -gal), β -lactamase, horseradish peroxidase (HRP), glucose oxidase (GO), human superoxide dismutase (hSOD), urease, and the like. These enzymes are convenient because the amount of chimeric protein produced can be quantified by means of simple colorimetric assays. Alternatively, one may employ antigenic proteins or fragments, e.g., human superoxide dismutase (hSOD), to permit simple detection and quantification of chimeric proteins using antibodies specific for the non-*Cryptovirus* protein chimeric partner. In chimeric proteins, useful chimeric partners include amino acid sequences that provide for secretion from a recombinant host, enhance the immunological reactivity of a *Cryptovirus* protein epitope(s), or facilitate the coupling of the *Cryptovirus* polypeptide to a support or a vaccine carrier. (See, e.g., EPO Pub. No. 116,201; U.S. Pat. No. 4,722,840; EPO Pub. No. 259,149; U.S. Pat. No. 4,629,783).

Embodiments of the inventive *Cryptovirus* protein are useful as immunoreactive polypeptides, including use for the production of a *Cryptovirus*-specific antibody. "Immunoreactive" refers to the ability of a polypeptide to bind immunologically to an antibody and/or to a lymphocyte antigen receptor due to antibody or receptor recognition of a specific epitope contained within the polypeptide; or the ability of a polypeptide to be immunogenic. An "immunogenic" protein is a polypeptide that elicits a cellular and/or humoral immune response, whether alone or linked to a carrier in the presence or absence of an adjuvant. The immunogenicity of various isolated *Cryptovirus* proteins, or *Cryptovirus* protein fragments, of interest is determined by routine screening. Immunological reactivity may be determined by antibody binding, more particularly by the kinetics of antibody binding, and/or by competition in binding using as competitor(s) a known polypeptide(s) containing an epitope against which the antibody is directed. The techniques for determining whether a polypeptide is immunologically reactive with an antibody are known in the art. Particularly useful examples of immunogenic *Cryptovirus* proteins are the envelope proteins, i.e., F, F₀, F₂, F₁, , HN, and SH proteins.

As used herein, "epitope" refers to an antigenic determinant of a polypeptide. An epitope can comprise 3 amino acids in a spatial conformation which is unique to the epitope. Epitopes typically are mapped to comprise at least about five amino acids, more usually at least about 8 amino acids to about 10 amino acids, or more. Methods of determining the spatial conformation of amino acids are known in the art, and include, for example, x-ray crystallography and two-dimensional nuclear magnetic resonance.

Immunogenic *Cryptovirus* proteins are useful for producing or manufacturing vaccines. As described above, *Cryptovirus* belongs to the Paramyxoviridae family of viruses. Numerous vaccines have been developed for humans and domestic animals against viruses in this virus family. For example, there are effective vaccines against measles virus, mumps virus, canine distemper virus, canine parainfluenza virus type 2, and Newcastle's disease virus (of fowl). These viruses were amenable to the development of effective vaccines because they have a narrow species tropism (*i.e.*, they infect only one, or only a few, species), they exist as only one, or only one predominant, serotype (making a vaccine universally protective against the virus), they cause significant morbidity in their host (*i.e.*, they are significant causes of human illness), and they are pandemic (*i.e.*, there is a worldwide concern).

Cryptovirus is amenable to vaccine development for the same reasons other family members have proven so. There is evidence for cross-neutralizability of hyper immune rabbit antiserum made against different sources of the virus, and very similar nucleotide sequences of the virus genome have been obtained from two sources of the virus (BBR strain and Niigata cell-associated strain).

In accordance with the invention, multivalent or monovalent vaccines can be prepared against one or more *Cryptovirus* proteins. In particular, vaccines are contemplated comprising one or more *Cryptovirus* proteins, such as, but not limited to, envelope proteins F, F₀, F₂, F₁, HN, and/or SH. Methods for manufacturing vaccines which contain an immunogenic polypeptide as an active ingredient, are known to one skilled in the art. Typically, such vaccines are prepared as injectable compositions comprising the *Cryptovirus* protein(s), either as liquid solutions or suspensions; solid forms suitable for solution in, or suspension in, liquid prior to injection may also be prepared. The preparation may also be emulsified, or the protein encapsulated in liposomes. The active immunogenic ingredients are typically mixed with excipients which are pharmaceutically acceptable and compatible with the active ingredient. Suitable excipients or carriers are, for example, water, saline, dextrose, glycerol, ethanol, or the like and combinations thereof. In addition, if desired, the vaccine may contain minor amounts of auxiliary substances such as wetting or emulsifying agents, pH buffering agents, and/or adjuvants which enhance the effectiveness of the vaccine.

Examples of adjuvants which may be effective include but are not limited to: aluminum hydroxide, N-acetyl-muramyl-L-threonyl-D-isoglutamine (thr-MDP), N-acetyl-nor-muramyl-L-alanyl-D-isoglutamine (CGP 11637, referred to as nor-MDP), N-acetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1-2-dipalmitoyl-sn-glycero-3-hydroxyphosphoryloxy)-ethylamine (CGP 19835A, referred to as MTP-PE), and RIBI, which contains three components extracted from bacteria, monophosphoryl lipid A, trehalose dimycolate and cell wall skeleton (MPL+TDM+CWS) in a 2% squalene/Tween 80 emulsion. The effectiveness of a particular adjuvant can be determined by measuring the amount of antibodies directed against an immunogenic *Cryptovirus* protein resulting from administration of this protein in vaccines which are also comprised of the adjuvant.

The proteins can be formulated into the vaccine as neutral or salt forms. Pharmaceutically acceptable salts include the acid addition salts (formed with free amino groups of the peptide) and which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or organic acids such as acetic, oxalic, tartaric, maleic, and the like. Salts formed with the free carboxyl groups may also be derived from inorganic bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, procaine, and the like.

The vaccines are conventionally administered parenterally by injection, for example, either subcutaneously or intramuscularly, but they can also be delivered intranasally, enterically, or by any other delivery route. Administration can be with or without adjuvants. Additional formulations of the vaccine composition that are suitable for other non-injection modes of administration include suppositories and, in some cases, oral formulations. For suppositories, traditional binders and carriers can include, for example, polyalkylene glycols or triglycerides; such suppositories may be formed from mixtures containing the active ingredient in the range of 0.5% to 10%, preferably 1%-2%. Oral formulations include such normally employed excipients as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate, and the like. These compositions can take the form of solutions, suspensions, emulsions, tablets, pills, capsules, caplets, sustained release formulations or powders and contain typically 10%-95% of active ingredient, preferably 25%-70%.

In addition to the above, it is also possible to manufacture live vaccines of attenuated microorganisms, e.g., weakened or avirulent virus particles, including mutated *Cryptovirus* particles, and other attenuated virus particles containing an inventive recombinant nucleic acid encoding one or more *Cryptovirus* proteins, or host cells that express recombinant *Cryptovirus* proteins encoded by

inventive expression vectors, as described herein. Suitable attenuated microorganisms are known in the art and include, for example, viruses (e.g., vaccinia virus) as well as bacteria.

Alternatively, killed *Cryptovirus* particles or virions, or killed pseudotyped viral particles or virions bearing *Cryptovirus* envelope proteins, are useful in the manufacture of a vaccine. Virions are killed for vaccine purposes by known methods.

The vaccines are administered in a manner compatible with the dosage formulation, and in such amount as will be prophylactically and/or therapeutically effective. The quantity to be administered, which is generally in the range of about 5 µg to about 250 µg of antigen per dose, depends on the subject to be vaccinated, capacity of the subject's immune system to synthesize antibodies, and the degree of protection desired. Precise amounts of active ingredient required to be administered depend on the judgment of the practitioner and can be particular to each vaccinated animal or human. Any suitable vertebrate animal can be vaccinated, particularly a member of a mammalian species, including rodents, lagomorphs, goats, pigs, cattle, sheep, and primates.

The vaccine may be given in a single dose schedule, or preferably in a multiple dose schedule. A multiple dose schedule is one in which a primary course of vaccination may be with 1-10 separate doses, followed by other doses given at subsequent time intervals required to maintain and/or reinforce the immune response, for example, at 1-4 months for a second dose, and if needed, a subsequent dose(s) after several months. The dosage regimen will also, at least in part, be determined by the immunological characteristics and needs of the individual animal or human to be vaccinated and must be dependent upon the judgment of the practitioner.

In addition, the vaccine containing the *Cryptovirus* proteins described above, can be administered in conjunction with other immunoregulatory agents, for example, immunoglobulins.

Compositions of the present invention can be administered to individual animals or humans to generate polyclonal antibodies (purified or isolated from serum using conventional techniques) which can then be used in a number of applications. For example, the polyclonal antibodies can be used to passively immunize an animal or human, or as immunochemical reagents, as described hereinbelow.

The present invention is also directed to an isolated antibody that specifically binds a *Cryptovirus* protein, such as the *Cryptovirus* NP, V, P, M, F, F₀, F₂, F₁, SH, HN, or L proteins. The term "antibody" refers to a polypeptide or group of polypeptides which are comprised of at least one antibody binding domain. A "binding domain" is formed from the folding of variable domains of an antibody molecule to form three-dimensional binding spaces with an internal surface shape and charge distribution complementary to the features of an epitope of an antigen, which allows an

immunological reaction with the antigen. An antibody binding domain can be formed from a heavy and/or a light chain domain (V_H and V_L , respectively), which form hypervariable loops which contribute to antigen binding. Typical vertebrate antibodies are tetramers or aggregates thereof, comprising light and heavy chains which are typically aggregated in a "Y" configuration and which may or may not have covalent linkages between the chains. In vertebrate antibodies, the amino acid sequences of all the chains of a particular antibody are homologous with the chains found in one antibody produced by the lymphocyte which produces that antibody in vivo, or in vitro (for example, in hybridomas).

An "isolated" antibody is an antibody, for instance polyclonal antibody, removed from the body of an animal or human that produced it. The inventive polyclonal antibody can be further purified from cellular material, e.g., in blood, lymph, or milk. A preferred embodiment is in the form of an antiserum directed against one or more *Cryptovirus* proteins. Alternatively, an "isolated" antibody of the present invention includes antibodies the production of which involves a manipulation or human intervention, for example, monoclonal antibody or chimeric antibody.

The inventive "antibody" includes any immunoglobulin, including IgG, IgM, IgA, IgD or IgE, or antibody fragment that binds a specific *Cryptovirus* epitope. Such antibodies can also be produced by hybridoma, chemical synthesis or recombinant methods known in the art. (E.g., Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (2 ed.), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, USA [1989]); Harlow and Lane, *Antibodies: A Laboratory Manual* (Cold Spring Harbor Laboratory [1988]). Both anti-*Cryptovirus* protein and anti-chimeric protein antibodies can be useful within the scope of the present invention. (See, e.g., Bahouth *et al.*, *Trends Pharmacol. Sci.* 12:338 [1991]; Ausubel *et al.*, *Current Protocols in Molecular Biology* (John Wiley and Sons, NY [1989]). Examples of chimeric antibodies are discussed in U.S. Pat. Nos. 4,816,397 and 4,816,567. Fluorescent-labeled antibodies, enzyme-conjugated antibodies, or antibodies otherwise labeled for facility of detection, as known in the art, are also included within "antibody."

"Antibody" also includes "chimeric antibody." Chimeric antibodies are antibodies in which the heavy and/or light chains are chimeric proteins. Typically the constant domain of the chains is from one particular species and/or class, and the variable domains are from a different species and/or class. Also included is any antibody in which either or both of the heavy or light chains are composed of combinations of sequences mimicking the sequences in antibodies of different sources, whether these sources be differing classes, or different species of origin, and whether or not the fusion point is at the variable/constant boundary. Thus, it is possible to produce antibodies in which neither the

constant nor the variable region mimic known antibody sequences. It then becomes possible, for example, to construct antibodies whose variable region has a higher specific affinity for a particular antigen, or whose constant region can elicit enhanced complement fixation, or to make other improvements in properties possessed by a particular constant region.

5 Included also within the definition of "antibody" are Fab and $F(ab')_2$ fragments of antibodies. The "Fab" region refers to those portions of the heavy and light chains which are roughly equivalent, or analogous, to the sequences which comprise the branch portion of the heavy and light chains, and which have been shown to exhibit immunological binding to a specified antigen, but which lack the effector F_c portion. A "Fab" fragment is an aggregate of one heavy and one light
10 chain. A $F(ab')_2$ fragment, which also lacks the effector F_c portion, is a tetramer containing the 2H and 2L chains, which are capable of selectively reacting with a designated antigen or antigen family. Methods of producing "Fab" and $F(ab')_2$ fragments of antibodies are known within the art and include, for example, proteolysis, and synthesis by recombinant techniques. Thus, the inventive anti-*Cryptovirus* antibodies can also be Fab or $F(ab')_2$ antibody fragments.

15 Also useful is a "single domain antibody" (dAb), which is an antibody which is comprised of a V_H domain, which reacts immunologically with a *Cryptovirus* antigen. A dAb does not contain a V_L domain, but may contain other antigen binding domains known to exist in antibodies, for example, the kappa and lambda domains. Methods for preparing dABs are known in the art. (See, e.g., Ward *et al.*, Nature 341: 544 [1989]).

20 Other preferred embodiments include altered antibodies such as humanized, CDR-grafted or bifunctional, i.e., divalent antibodies, all of which can also be produced by methods well known in the art. "Altered antibodies", which refers to antibodies in which the naturally occurring amino acid sequence in a vertebrate antibody has been varied. Utilizing recombinant DNA techniques, antibodies can be redesigned to obtain desired characteristics. The possible variations are many, and
25 range from the changing of one or more amino acids to the complete redesign of a region, for example, the constant region. Changes in the constant region, in general, to attain desired cellular process characteristics, e.g., changes in complement fixation, interaction with membranes, and other effector functions. Changes in the variable region may be made to alter antigen binding characteristics. The antibody may also be engineered to aid the specific delivery of a molecule or
30 substance to a specific cell or tissue site. The desired alterations may be made by known techniques in molecular biology, e.g., recombinant techniques, site directed mutagenesis, etc.

A preferred embodiment of the inventive antibody specifically binds a *Cryptovirus* envelope protein described hereinabove. Another preferred embodiment of the inventive antibody specifically

binds a *Cryptovirus* nucleocapsid protein, but antibodies that specifically bind any other *Cryptovirus* protein are also useful and preferred.

The inventive antibody can be used, inter alia, in diagnostic or assay methods and systems to detect *Cryptovirus* proteins present in a sample of biological material. With respect to the detection of such polypeptide, the antibodies can be used for in vitro diagnostic or assay methods, or in vivo imaging methods. Such antibodies can also be used for the immunoaffinity or affinity chromatography purification of the inventive *Cryptovirus* proteins.

The present invention includes an in vitro method of detecting the presence or absence of a *Cryptovirus* protein in a sample of a biological material. The method is particularly, but not exclusively, useful for testing clinical or experimental biological materials for diagnostic or pathology purposes.

The sample is contacted with the inventive antibody described herein; and known immunological procedures are employed for detecting specific binding of the antibody to a constituent of the sample. The presence of specific binding indicates the presence of the *Cryptovirus* protein in the sample.

Immunological procedures, useful for in vitro detection of target *Cryptovirus* proteins in a sample, include immunoassay systems that employ the inventive *Cryptovirus* protein-specific antibody in a detectable form.

Known protocols for such immunoassay techniques and systems are based, for example, upon competition, or direct reaction, or sandwich type assays. Such immunoassay techniques and systems include, for example, ELISA, immunoblotting, immunofluorescence assay (IFA), Pandex microfluorimetric assay, agglutination assays, flow cytometry, serum diagnostic assays and immunohistochemical staining procedures which are well known in the art.

An antibody can be made detectable by various means well known in the art. Assay systems that amplify the signals from a primary antibody-antigen complex are also known; examples of which are assays which utilize biotin and avidin, and enzyme-labeled and mediated immunoassays, such as ELISA assays. A detectable marker can be directly or indirectly attached to a primary or secondary antibody in the assay protocol. Useful markers include, for example, radionuclides, enzymes, fluorogens, chromogens and chemiluminescent labels. Embodiments can employ solid support matrices, or can involve immunoprecipitation.

These same immunoassay techniques and systems can be employed in the inventive method of detecting the presence or absence of a *Cryptovirus*-specific antibody in an antibody-containing biological material. Antibody-containing biological materials include, but are not limited to, whole blood and blood components, plasma, serum, spinal fluid, lymph fluid, the external sections of the respiratory, intestinal, and genitourinary tracts, tears, saliva, milk, white blood cells, and myelomas. Processed antibody-containing fractions or dilutions of any of these are also considered antibody-containing biological materials.

One preferred embodiment of the method of detecting the presence or absence of a *Cryptovirus*-specific antibody involves contacting the sample originating from an individual suspected of having a *Cryptovirus* infection with a viral virion or other viral particle containing one or more *Cryptovirus* envelope proteins, as described herein, such that, if antibody selectively binding *Cryptovirus* antigen is present, an antibody-bound complex forms. Then any antibody-bound *Cryptovirus* antigen complexes thus formed are contacted with anti-human antibody-binding immunoglobulin or anti-human antibody-binding fragments thereof, for example, Fab and/or F(ab')₂, fragments, and complexes of the immunoglobulin or the fragments thereof, are allowed to form with the antibody-bound *Cryptovirus* complexes. The presence or absence of any complexes formed is detected, by any known immunodetection means as described herein. The presence of such complexes indicates the presence in the sample of antibody that selectively binds *Cryptovirus* antigen.

Another more preferred embodiment involves contacting the sample originating from an individual suspected of having a *Cryptovirus* infection with the inventive *Cryptovirus* envelope protein, such that, if antibody selectively binding *Cryptovirus* is present, an antibody-bound envelope protein complex forms. Any antibody-bound envelope protein complexes thus formed are then contacted with anti-human antibody-binding immunoglobulin or anti-human antibody-binding fragments, such as Fab and F(ab')₂ fragments. The formation of complexes of the immunoglobulin or the Fab and F(ab')₂ fragments thereof is allowed with the antibody-bound envelope protein complexes; and the presence or absence of any antibody-bound envelope protein complexes thus formed is detected. The presence of such complexes indicating the presence in the sample of antibody selectively binding *Cryptovirus*.

The terms "selective" or "specific" binding of antibody to *Cryptovirus* proteins or *Cryptovirus* antigens therein, includes asymmetric cross-reactive binding with closely related rubulaviruses, such as SV5, but does not include non-specific binding to unrelated antigens or surfaces. The skilled artisan is aware of important controls that are preferably included in any

immunoassay system for the determination of non-specific antibody binding. Typically, for example in ELISA, a background level of non-specific binding is determined and used as a baseline.

“Complexed” means that a protein, such as an antibody, is a constituent or member of a complex, i.e., a mixture or adduct resulting from chemical binding or bonding between and/or among the other constituents. Chemical binding or bonding can have the nature of a covalent bond, ionic bond, hydrogen bond, hydrophobic bond, or any combination of these bonding types linking the constituents of the complex at any of their parts or moieties, of which a constituent can have one or a multiplicity of moieties of various sorts. Antibody-antigen binding is typically non-covalent. Not every constituent of a complex need be bound to every other constituent, but each constituent has at least one chemical bond with at least one other constituent of the complex. For example, a secondary antibody in the assay system may not be directly bound with the *Cryptovirus* antigen, or the viral particle or virion, yet it is “complexed” with it.

By way of further non-limiting illustration of a clinical diagnostic embodiment for detecting *Cryptovirus* infection in a human patient, a serum specimen from the patient is screened for *Cryptovirus*-specific antibodies to the major envelope proteins of the virus (F_0 and HN) by ELISA, radio-immunoprecipitation, immunoblotting techniques or any other immunological technique (e.g., direct or indirect fluorescent antibody techniques, immunobeads, etc.). Optionally, another blood sample is obtained from any seropositive individual. The virus can be detected in the PBMNCs of such samples by isolating the cells on a suitable gradient medium, culturing the cells in the presence of cyclic GMP and then either (1) screening the cells for intracellular *Cryptovirus*-specific inclusions with *Cryptovirus*-specific fluorescent antibodies or (2) PCR amplification of induced PBMNC with *Cryptovirus*-specific nucleotide primers.

While *Cryptovirus*-specific antibodies have been found in the serum of seropositive individuals, indicating current or former *Cryptovirus* infection, these antibodies are not necessarily indicative of epileptiform or encephalopathic disease. *Cryptovirus* appears to infect and be carried in the PBMNCs of a significant proportion of individuals without necessarily causing encephalopathic disease. These individuals can overtly appear to be asymptomatic. The neuropathological (e.g., epileptiform, encephalopathic, and other neurological, neurodegenerative, and/or neuropsychiatric disease) potential of the virus only appears to become manifest in individuals in whom the virus has infected nervous system tissues. Consequently, only *Cryptovirus*-specific antibodies (i.e. those directed against the major envelope proteins of the virus, F and HN) found in the cerebrospinal fluid (CSF) are fully indicative of neurological, neurodegenerative, and/or neuropsychiatric disease and then, they are virtually always indicative. By way of further example, ten human patients who were

previously diagnosed with chronic fatigue syndrome involving significant short-term memory loss, and for whom CSF samples could be obtained, were all found to have *Cryptovirus*-specific antibody in their CSF and electroencephalographic profiles consistent with a diagnosis of absence epilepsy (i.e., *petit mal* epilepsy).

5 The present invention also relates to an anti-*Cryptovirus* antibody detecting kit, useful for testing or assaying a biological sample, in particular an antibody-containing biological material. Thus, the inventive kits are beneficial for screening clinical supplies of human blood, serum, platelets, plasma, tissues and organs, to determine their safety for transfusion or transplantation purposes. Diagnostic applications of the inventive kits are also useful.

10 The inventive kit is particularly useful for practicing the inventive assay methods for detecting antibody that selectively binds *Cryptovirus* and its antigens and the inventive methods of detecting the presence or absence of a *Cryptovirus*-specific antibody. In some preferred embodiments, the kit contains an isolated *Cryptovirus* particle, comprising a genome having a nucleotide sequence entirely complementary to (SEQ ID NO:1). The kit also contains labeled anti-
15 human antibody-binding antibody, preferably anti-human immunoglobulin or labeled anti-human antibody-binding antibody fragments, such as Fab and/or F(ab')₂. A preferred embodiment of the kit further contains a solid matrix for supporting the *Cryptovirus* particle(s). In a preferred embodiment, the *Cryptovirus* particles in the anti-*Cryptovirus* antibody detecting kit are *Cryptovirus* virions, and in a preferred embodiment the *Cryptovirus* virions are produced from an acutely *Cryptovirus*-infected
20 cell line, such as an acutely infected baby hamster kidney (BHK) cell-derived cell line, a Vero-derived cell line, or a CV-1-derived cell line (e.g., a CV-1_c-derived cell line, described hereinbelow.).

Alternatively, in a most preferred embodiment of the inventive anti-*Cryptovirus* antibody detecting kit, which does not include *Cryptovirus* particles, the kit includes a plurality of one or more kinds of isolated *Cryptovirus* proteins and/or chimeric proteins comprising a *Cryptovirus* protein
25 moiety, as described hereinabove. In some preferred embodiments, the *Cryptovirus* protein or protein moiety is an envelope protein, as described herein. Such embodiments also contain labeled anti-human antibody-binding antibody, such as anti-human immunoglobulin or labeled anti-human antibody-binding antibody fragments, such as Fab and/or F(ab')₂ fragments. A preferred embodiment of such a kit further contains a solid matrix for supporting the *Cryptovirus* proteins.

30 Solid matrices, or supports, and methods for attaching or adsorbing viral particles and proteins to a solid matrix, are well known in the art. In accordance with the inventive kits, the term "solid matrix" includes any solid or semi-solid support or surface to which the viral particle or protein

can be anchored or adhered. Suitable matrices are made of glass, plastic, metal, polymer gels, and the like, and may take the form of beads, wells (e.g., single- or multi-well serum plates) slides, dipsticks, membranes, and the like.

5 As is known to the skilled artisan in using such kits, e.g., for ELISA, RIA, or sandwich-type assays, the biological sample, optimally solubilized or suspended and in an optimized dilution, is contacted with the viral particles or proteins, typically supported on the surface of the solid matrix (e.g., well of a serum plate), and after appropriate washes and incubations, is further contacted with the labeled anti-human antibody-binding immunoglobulin or labeled anti-human antibody-binding fragments. After further washes, commercially available plate readers and/or other accessory
10 detection equipment are typically employed in conjunction with the inventive kit, for detecting the formation of bound anti-human antibody complexes with human antibody that has bound to *Cryptovirus* particles or proteins.

Instructions for use are included in the kit. "Instructions for use" typically include a tangible expression describing the reagent concentration or at least one assay method parameter, such as the
15 relative amounts of reagent and sample to be admixed, maintenance time periods for reagent/sample admixtures, temperature, buffer conditions, incubations, washes, and the like, typically for an intended purpose, in particular the inventive assay methods as described herein.

Optionally, the kit also contains other useful components, such as, diluents, buffers, or other acceptable carriers, specimen containers, syringes, pipetting or measuring tools, paraphernalia for
20 concentrating, sedimenting, or fractionating samples, or the inventive antibodies for use in controls.

The materials or components assembled in the kit can be provided to the practitioner stored in any convenient and suitable ways that preserve their operability and utility. For example the components can be in dissolved, dehydrated, or lyophilized form; they can be provided at room, refrigerated or frozen temperatures.

25 The components are typically contained in suitable packaging material(s). As employed herein, the phrase "packaging material" refers to one or more physical structures used to house the contents of the kit. The packaging material is constructed by well known methods, preferably to provide a sterile, contaminant-free environment.

The packaging materials employed in the kit are those customarily utilized in virus- and
30 peptide-based systems. As used herein, the term "package" refers to a suitable solid matrix or material such as glass, plastic, paper, foil, and the like, capable of holding the individual kit components. Thus, for example, a package can be a glass vial used to contain suitable quantities of

an inventive composition containing viral or peptide components. The packaging material generally has an external label which indicates the contents, quantities, and/or purpose of the kit and/or its components.

Thus the present invention provides immunoassay methods and kits, useful for research, clinical diagnostics, and screening of blood, blood components or products, and tissue and organs intended for transfusion or transplantation. These applications are of great value and utility, because strong evidence shows that peripheral blood mononuclear cells (PBMNCs) can harbor *Cryptovirus* and can subsequently infect and cause disease in a patient who receives such contaminated blood, blood products, tissues, or cells.

The inventive methods of detecting the presence or absence of a *Cryptovirus*-specific RNA in a sample of a biological material, described hereinabove, are also particularly useful for these and other purposes. In accordance with the most preferred embodiments of the method, *Cryptovirus* RNAs are amplified by any of numerous known methods of amplifying nucleic acid segments, in the form of RNA or cDNA. Before amplification, it is preferable, but not necessary, to extract or separate RNA from DNA in the sample and to amplify nucleic acids remaining in that fraction of the sample separated from the DNA. The amplification products, if any, are then analyzed to detect the presence of *Cryptovirus*-specific amplification products. If *Cryptovirus*-specific amplification products are present, the findings are indicative of the presence of *Cryptovirus* RNA in the sample. For greater confidence in the interpretation of negatives (i.e., no detectable *Cryptovirus*-specific amplification products), analysis is optionally carried out following a control amplification of mRNAs specific for a cellular housekeeping gene, for example, a gene encoding β -actin, phosphofructokinase (PFK), glyceraldehyde 3-phosphate dehydrogenase, or phosphoglycerate kinase.

The RNAs in the sample, are amplified by a suitable amplification method. For example, in a preferred embodiment, a reverse transcriptase-mediated polymerase chain reaction (RT-PCR) is employed to amplify *Cryptovirus*-specific nucleic acids. Briefly, two enzymes are used in the amplification process, a reverse transcriptase to transcribe *Cryptovirus*-specific cDNA from a *Cryptovirus*-specific RNA template in the sample, a thermal resistant DNA polymerase (e.g., *Taq* polymerase), and *Cryptovirus*-specific primers to amplify the cDNA to produce *Cryptovirus*-specific amplification products. The use of limited cycle PCR yields semi-quantitative results. (E.g., Gelfand *et al.*, *Reverse transcription with thermostable DNA polymerase-high temperature reverse transcription*, U.S. Patent Nos. 5,310,652; 5,322,770; Gelfand *et al.*, *Unconventional nucleotide substitution in temperature selective RT-PCR*, U.S. Patent No. 5,618,703).

In another preferred embodiment of the inventive method, single enzyme RT-PCR is employed to amplify *Cryptovirus*-specific nucleic acids. Single enzymes now exist to perform both reverse transcription and polymerase functions, in a single reaction. For example, the Perkin Elmer recombinant *Thermus thermophilus* (rTth) enzyme (Roche Molecular), or other similar enzymes, are commercially available. Cycling instruments such as the Perkin Elmer ABI Prism 7700, the so-called Light Cycler (Roche Molecular), and/or other similar instruments are useful for carrying out RT-PCR. Optionally, single enzyme RT-PCR technology, for example, employing rTth enzyme, can be used in a PCR system.

By way of illustration only, RT-PCR-based testing is quite sensitive for detection of the virus. For example, a RT-PCR-priming technique has been used to confirm a detectable *Cryptovirus* carriage rate in PBMNCs nonproductively harboring the virus (without culturing, cyclic GMP induction, and passaging) on the order of 1:10⁵ PBMNC. In addition, numerous fragments of the *Cryptovirus* genome have been cloned from AV₃/SSPE cells using *Cryptovirus*-specific primers and a RT-PCR-based amplification technique.

Preferably, amplification and analysis are carried out in an automated fashion, with automated extraction of RNA from a sample, followed by PCR, and fluorescence detection of amplification products using probes, such as TaqMan or Molecular Beacon probes. Typically, the instrumentation includes software that provides quantitative analytical results during or directly following PCR without further amplification or analytical steps.

In another preferred embodiment, transcription-mediated amplification (TMA) is employed to amplify *Cryptovirus*-specific nucleic acids. (E.g., K. Kamisango *et al.*, *Quantitative detection of hepatitis B virus by transcription-mediated amplification and hybridization protection assay*, J. Clin. Microbiol. 37(2):310-14 [1999]; M. Hirose *et al.*, *New method to measure telomerase activity by transcription-mediated amplification and hybridization protection assay*, Clin. Chem. 44(12):2446-52 [1998]). Rather than employing RT-PCR for the amplification of a cDNA, TMA uses a probe that recognizes a *Cryptovirus*-specific (target sequence) RNA; in subsequent steps, from a promoter sequence built into the probe, an RNA polymerase repetitively transcribes a cDNA intermediate, in effect amplifying the original RNA transcripts and any new copies created, for a level of sensitivity approaching that of RT-PCR. The reaction takes place isothermally (one temperature), rather than cycling through different temperatures as in PCR.

Other useful amplification methods include a reverse transcriptase-mediated ligase chain reaction (RT-LCR), which has utility similar to RT-PCR. RT-LCR relies on reverse transcriptase to

generate cDNA from mRNA, then DNA ligase to join adjacent synthetic oligonucleotides after they have bound the target cDNA.

Most preferably, amplification of a *Cryptovirus*-specific nucleic acid segment in the sample can be achieved using *Cryptovirus*-specific oligonucleotide primers of the present invention, as described herein.

Optionally, high throughput analysis may be achieved by multiplexing techniques well known in the art, employing multiple primer sets, for example primers directed not only to *Cryptovirus*-specific nucleic acids, but to amplifying expression products of housekeeping genes (controls) or of other potential diagnostic markers, to yield additional diagnostic information. (E.g., Z. Lin *et al.*, *Multiplex genotype determination at a large number of gene loci*, Proc. Natl. Acad. Sci. USA 93(6):2582-87 [1996]; Demetriou *et al.*, *Method and probe for detection of gene associated with liver neoplastic disease*, U.S. Patent No. 5,866,329).

Hybridization analysis is a preferred method of analyzing the amplification products to detect the presence or absence of *Cryptovirus*-specific nucleic acid amplification products, employing one or more *Cryptovirus*-specific probe(s) that, under suitable conditions of stringency, hybridize(s) with single stranded *Cryptovirus*-specific nucleic acid amplification products comprising complementary nucleotide sequences. The amplification products are typically deposited on a substrate, such as a cellulose or nitrocellulose membrane, and then hybridized with labeled *Cryptovirus*-specific probe(s), optionally after an electrophoresis. Conventional dot blot, Southern, Northern, or fluorescence in situ (FISH) hybridization protocols, *in liquid* hybridization, hybridization protection assays, or other semi-quantitative or quantitative hybridization analysis methods are usefully employed along with the *Cryptovirus*-specific probes of the present invention. As is readily apparent to the skilled artisan, such analytical hybridization techniques and others (e.g., Northern blotting), are useful in accordance with other embodiments of the inventive method of detecting the presence or absence of a *Cryptovirus*-specific RNA in a sample of a biological material that do not involve any amplification step(s). In these embodiments, the inventive *Cryptovirus*-specific probes are contacted directly with RNA in the sample to perform hybridization analysis.

Alternatively, electrophoresis for analyzing or detecting amplification products is done rapidly and with high sensitivity by using any of various methods of conventional slab or capillary electrophoresis, with which the practitioner can optionally choose to employ any facilitating means of nucleic acid fragment detection, including, but not limited to, radionuclides, UV-absorbance or laser-induced fluorescence. (K. Keparnik *et al.*, *Fast detection of a (CA)18 microsatellite repeat in the IgE receptor gene by capillary electrophoresis with laser-induced fluorescence detection*, Electrophoresis

19(2);249-55 [1998]; H. Inoue *et al.*, *Enhanced separation of DNA sequencing products by capillary electrophoresis using a stepwise gradient of electric field strength*, J. Chromatogr. A. 802(1):179-84 [1998]; N.J. Dovichi, *DNA sequencing by capillary electrophoresis*, Electrophoresis 18(12-13):2393-99 [1997]; H. Arakawa *et al.*, *Analysis of single-strand conformation polymorphisms by capillary electrophoresis with laser induced fluorescence detection*, J. Pharm. Biomed. Anal. 15(9-10):1537-44 [1997]; Y. Baba, *Analysis of disease-causing genes and DNA-based drugs by capillary electrophoresis. Towards DNA diagnosis and gene therapy for human diseases*, J. Chromatogr. B. Biomed. Appl. 687(2):271-302 [1996]; K.C. Chan *et al.*, *High-speed electrophoretic separation of DNA fragments using a short capillary*, J. Chromatogr. B. Biomed. Sci. Appl. 695(1):13-15 [1997]).

Any biological material can be sampled for the purpose of practicing the inventive methods of detecting the presence or absence of a *Cryptovirus*-specific protein or *Cryptovirus*-specific RNA in a sample of a biological material. Preferred biological materials for sampling include blood or serum, lymphoid tissue, nervous tissue, including brain tissue. However, the biological material can be cerebrospinal fluid (CSF), lymph, plasma, feces, semen, prostatic fluid, tears, saliva, milk, gastric juice, mucus, synovial fluid, pleural effusion, peritoneal effusion, pericardial effusion, skin, vascular epithelium, oral epithelium, vaginal epithelium, cervical epithelium, uterine epithelium, intestinal epithelium, bronchial epithelium, esophageal epithelium, or mesothelium, or other biopsy sample of cellular material from any tissue. Cellular material includes any sample containing mammalian cells, including samples of tissues, expressed tissue fluids, tissue wash or tissue rinsate fluids, blood cells (e.g., peripheral blood mononuclear cells), tumors, organs, and also samples of in vitro cell culture constituents (including but not limited to conditioned medium resulting from the growth of cells in cell culture medium, putatively virally infected cells, recombinant cells, and cell components), or the like.

Tissue samples that can be collected include, but are not limited to, cell-containing material from the brain, blood, spleen, lymph node, vasculature, kidney, pituitary, ureter, bladder, urethra, thyroid, parotid gland, submaxillary gland, sublingual gland, bone, cartilage, lung, mediastinum, breast, uterus, ovary, testis, prostate, cervix uteri, endometrium, pancreas, liver, adrenal, esophagus, stomach, and/or intestine.

The sample is alternatively derived from cultured mammalian cells, cell-free extracts, or other specimens indirectly derived from a mammalian subject's body, as well as from substances taken directly from a subject's body. The samples can be stored before detection methods are applied (for example nucleic acid amplification and/or analysis, or immunochemical detection) by well known storage means that will preserve nucleic acids or proteins in a detectable and/or

analyzable condition, such as quick freezing, or a controlled freezing regime, in the presence of a cryoprotectant, for example, dimethyl sulfoxide (DMSO), glycerol, or propanediol-sucrose. Samples may also be pooled before or after storage for purposes of amplifying their *Cryptovirus*-specific nucleic acids for analysis and detection, or for purposes of detecting *Cryptovirus* protein.

5 The sample is optionally pre-treated by refrigerated or frozen storage overnight, by dilution, by phenol-chloroform extraction, or by other like means, to remove factors that may inhibit various amplification reactions that may be employed; such as heme-containing pigments or urinary factors. (E.g., J. Mahony *et al.*, *Urine specimens from pregnant and non-pregnant women inhibitory to amplification of Chlamydia trachomatis nucleic acid by PCR, ligase chain reaction, and*
10 *transcription-mediated amplification: identification of urinary substances associated with inhibition and removal of inhibitory activity*, J. Clin. Microbiol. 36(11):3122-26 [1998]).

 The present invention is also directed to an animal model for the study of human diseases, preferably, but not limited to, neurological, neurodegenerative, and/or neuropsychiatric diseases. The term "neurological diseases" refers to diseases of the nervous system, including neuropathies
15 manifested in the central nervous system and/or the peripheral nervous system. These include epileptiform diseases and non-epileptiform CNS diseases (e.g. Parkinsonism) and peripheral nervous system disease(s) (e.g. amyotrophic lateral sclerosis or "Lou Gehrig's Disease." "Neurodegenerative" diseases involve wasting or paralytic neurological diseases, which typically present with tremor, weakness and atrophy, for example Lou Gehrig's Disease or Alzheimer's
20 disease. The terms "neuropsychiatric" and "neuropsychological" are used interchangeably herein. "Neuropsychiatric diseases" are neurological diseases that also include behavioral symptoms that derive from the underlying neurophysiological processes. The animal model is a non-human mammal, such as, but not limited to, a rodent, a lagomorph, or a non-human primate.

 A rodent is any of the relatively small gnawing mammals of the order Rodentia, such as
25 mice, rats, hamsters, guinea pigs, squirrels, marmots, beavers, gophers, voles, porcupines, and agoutis. A lagomorph is any of various herbivorous mammals belonging to the order Lagomorpha, which includes rabbits, hares, and pikas.

 The animal is, or has been, inoculated with an infectious cell-free *Cryptovirus* virion of the present invention. Alternatively, the animal is, or has been, artificially inoculated with a cell
30 nonproductively-infected with *Cryptovirus*, such as AV₃/SSPE.

 Inoculation can be by a peripheral or an intracerebral delivery route. For the study of neurological, neurodegenerative, and/or neuropsychiatric diseases, intracerebral inoculation is

preferred, although for diseases presenting with involvement of the peripheral nervous system, peripheral inoculation can also be useful.

Intracerebral inoculation is by any suitable means, but preferably by direct injection into the brain, preferably into neural tissue, and most preferably by stereotactic injection means known in the art. Alternatively, intracerebral inoculation with *Cryptovirus* or nonproductitively infected cells can be by intraarterial (e.g., intracarotid) or intravenous injection or infusion, in conjunction with at least transient disruption of the blood brain barrier by physical or chemical means, delivered simultaneously with the *Cryptovirus* or nonproductively infected cells.

"Simultaneously" means that the physical or chemical means for disrupting the blood brain barrier are administered contemporaneously or concurrently with the *Cryptovirus* virions or nonproductively infected cells. "Simultaneously" also encompasses disrupting means being administered within about one hour after the *Cryptovirus* or nonproductively infected cells are last administered, preferably within about 30 minutes after, and most preferably, being administered simultaneously with the *Cryptovirus* or nonproductively infected cells. Alternatively, "simultaneously" means that the medicant is administered within about 30 minutes before, and preferably within about 15 minutes before the *Cryptovirus* or nonproductively infected cells are first administered.

Physical disruption of the blood brain barrier includes by means of "mechanical" injury or other physical trauma that breaches the blood brain barrier in at least one location of the brain's vasculature. Chemical disruption includes by an agent that transiently permeabilizes the blood-brain barrier and allows the *Cryptovirus* to enter the brain from the blood stream via the brain microvasculature. Such permeabilizing agents are known, for example, bradykinin and bradykinin analogs, and activators of calcium-dependent or ATP-dependent potassium channels. (e.g., B. Malfroy-Camine, *Method for increasing blood-brain barrier permeability by administering a bradykinin agonist of blood-brain barrier permeability*, U.S. Patent No. 5,112,596; J.W. Kozarich *et al.*, *Increasing blood brain barrier permeability with permeabilizer peptides*, U.S. Patent No. 5,268,164; Inamura, T. *et al.*, *Bradykinin selectively opens blood-tumor barrier in experimental brain tumors*, J. Cereb. Blood Flow Metab. 14(5):862-70 [1994]; K.L. Black, *Method for selective opening of abnormal brain tissue capillaries*, U.S. Patent Nos. 5,527,778 and 5,434,137; N.G. Rainov, *Selective uptake of viral and monocrystalline particles delivered intra-arterially to experimental brain neoplasms*, Hum. Gene. Ther. 6(12):1543-52 [1995]; N.G. Rainov *et al.*, *Long-term survival in a rodent brain tumor model by bradykinin-enhanced intra-arterial delivery of a therapeutic herpes simplex virus vector*, Cancer Gene Ther. 5(3):158-62 [1998]; F.H. Barnett *et al.*,

Selective delivery of herpes virus vectors to experimental brain tumors using RMP-7, Cancer Gene Ther. 6(1):14-20 [1999]; WO 01/54771 A2; and WO 01/54680 A2).

The inoculated non-human mammal exhibits at least one symptom characteristic of a human neurological, neurodegenerative, and/or neuropsychiatric disease after being thus inoculated, which was not previously exhibited by the non-human mammal before inoculation. Such symptoms include subacute symptoms and more slowly developing symptoms.

Generally, the subacute symptoms (developing from about 3 weeks to about 2 months post inoculation) associated with such experimental infections include: (1) cachexia/anorexia (*i.e.*, wasting or diminution of body mass and size); (2) degenerative neurologic wasting or paralysis; (3) atrophy of limb(s); (4) hindlimb paralysis; (5) photosensitivity or repetitive blinking; (6) hyperactivity or hyperesthesia (*e.g.*, nervousness, agitation, racing, jumpiness, extreme sensitivity to touch and sound); (7) ataxia (*i.e.*, loss of balance, wobbly gait); (8) hypesthesia; (9) withdrawal and isolation from other animals, closing of eyes, "hunched" posture; (10) stupor (*i.e.*, rigidity, semi-comatose, somnambulant motionlessness); (11) convulsions or seizures (*i.e.*, flaying of limbs, loss of consciousness, whirling, rolling and/or circling); (12) muscle spasms or myoclonus (*e.g.*, tremor, twitching of muscles, repetitive jerking of muscles); (13) corneal opacity (a clouding of the cornea) and (14) sudden death. Individual animals can present with one or more of the preceding subacute symptoms, but are generally observed displaying a complex of two or more symptoms. Subacute symptoms are more frequently observed in male animals compared to female animals.

More slowly developing symptoms (*i.e.*, those developing after about two months and sometimes not for about six months or more after inoculation) include: (1) obesity; (2) hypesthesia (*i.e.*, decreased sensitivity to sensory stimuli); (3) extreme lethargy and prolonged sleeping; (4) hyperactivity or hyperesthesia (*i.e.*, increased sensitivity to sensory stimuli); (5) aggressiveness (*e.g.*, jumping or biting); (6) obsessive compulsive behavior (*e.g.*, excessive and prolonged washing of the face or continual scratching); (7) self-mutilation (the extreme end of obsessive compulsive washing or scratching where the skin is damaged); (8) still-born fetuses and deformities in newborn animals (usually paralysis or limb atrophy) born to experimentally-infected females; and (9) infanticide (cannibalism of numerous newborns or entire litters). Individual animals can present with one or more of the preceding more slowly developing symptoms, but are generally observed displaying a complex of two or more symptoms. More slowly developing symptoms are more frequently observed in female animals compared to male animals.

The inventive animal model is an excellent model system for the study of neurodegenerative, wasting or paralytic neurological diseases which typically present with tremor, weakness and atrophy.

The inventive animal model is also, in particular, an excellent model system for the study of idiopathic epileptiform diseases because the infected animals present with virtually the entire spectrum of symptoms associated with epileptiform illnesses in humans. At present, most existing animal models of epilepsy (*e.g.*, induction of seizure by inoculation with the glutamate receptor agonist, kainite, or by partial suffocation) are contrived to produce seizures and the gross anatomical pathology associated with seizures without reference to the etiology of the actual symptomatic spectrum of the illnesses in humans. While these models are useful in developing therapeutics for seizure activity, there is little or no evidence that they are relevant to the ultimate aetiopathogenesis of epileptiform illnesses in humans or the actual spectrum of symptoms which occur.

In contrast, the animal model of the present invention is a truly homologous animal model; that is, one in which the actual factors/symptoms associated with the disease in humans are extant and can be specifically targeted by both therapeutic and prophylactic strategies. Thus, the inventive animal models disclosed herein can be used to screen antiviral medications or medicaments, including anti-epileptic and anti-psychotropic medicaments, as well as to test vaccines and other prophylactic remedies and to determine how to best coordinate and optimize any and all treatment strategies.

Cryptovirus is mildly cytopathic in cell culture but causes profound neuropathological disease in experimentally-infected animals. Any of the cytopathogenic and neuropathogenic traits of the virus can be used as markers in screens designed to identify and test potential antiviral therapeutic and/or prophylactic agents.

Accordingly, the present invention features *in vitro* and *in vivo* methods of screening potential antiviral therapeutic agents and/or antiviral prophylactic agents, including immunoprophylactic agents. A “potential” antiviral therapeutic or prophylactic agent is an agent that has not yet been clinically confirmed (*i.e.*, in phase III clinical trials) to have antiviral properties effective against *Cryptovirus*. Agents that have not been tested clinically against *Cryptovirus* infections or have been tested clinically against *Cryptovirus* infections only with respect to phase I and phase II clinical trials are also encompassed by “potential” antiviral therapeutic and/or prophylactic agents for purposes of the present invention.

In accordance with the inventive *in vitro* methods of screening a potential therapeutic or prophylactic agent, either acutely- or productively-infected mammalian cell cultures (*e.g.*, BHK, Vero, or CV-1c cells) or nonproductively infected carrier cultures (*e.g.*, AV₃/SSPE cells) can be used to evaluate the potential antiviral agent. While the acutely infected (productive) cellular system is preferentially useful for screening agents targeted at the processing and assembly of *Cryptovirus* envelope glycoproteins (*e.g.*, protease inhibition of F₀ cleavage activation), the nonproductively

infected cellular system (e.g., AV₃/SSPE cells) is preferred for screening for the efficacy of long-term treatment with transcriptional or other polymerase inhibitors (inhibiting the buildup of intracellular nucleocapsids and the eventual triggering of apoptotic cell death).

5 The inventive animal model is usefully employed in the *in vivo* method of screening a potential antiviral therapeutic agent. The method involves administering the potential therapeutic agent to be screened, to the inventive animal model after its inoculation with *Cryptovirus*, in accordance with the inventive animal model.

10 An alternative embodiment of the inventive animal model is employed in the *in vivo* method of screening a potential antiviral prophylactic agent. The method involves administering a potential prophylactic agent to be screened to a non-human mammal, which does not have a symptom of a human disease, such as but not limited to a neurological, neurodegenerative, and/or neuropsychiatric disease (e.g., an epileptiform disease). Then the animal is inoculated, as described herein, with the infectious cell-free *Cryptovirus* or with the mammalian cell nonproductively-infected with the *Cryptovirus*. The method is particularly, but not exclusively, useful for identifying potential anti-
15 epileptic or anti-psychotropic antiviral prophylactic agents.

Administration of the potential prophylactic agent or therapeutic agent is by any suitable delivery route, enteral (e.g., orally or by suppository) or parenteral (e.g., by injection, infusion, transmembrane, transdermal, or inhalation delivery route).

20 Examples of agents that can be evaluated, in accordance with the invention, include compounds or substances with known antiviral properties against viruses other than *Cryptovirus*; novel compounds or mixtures of compounds, such as cell, plant or animal extracts, with potential antiviral activity; and vaccines, as described hereinabove; or any combination of these.

25 Using the inventive *in vivo* method of screening, potential immunoprophylactic agents (*i.e.*, vaccines which stimulate the immune system to respond to, attack or inhibit virus replication, assembly or any other process associated with virus reproduction and spread) are also amenable to testing because non-human mammals can be inoculated with a putative prophylactic agent or vaccine (as mentioned above) and then challenged with infectious *Cryptovirus* to assess its utility in preventing the development of *Cryptovirus*-associated diseases. The use of such agents discovered in accordance with the invention may ultimately be necessary to control and eradicate *Cryptovirus*-
30 associated diseases in the human population much as measles and mumps vaccines have been used to bring these diseases under control in many countries.

In addition to those named above, one of ordinary skill in the art will recognize numerous potential antiviral chemo- and molecular-therapeutic agents that could be analyzed or evaluated using

the *in vitro* (cell culture) or *in vivo* methods of screening provided herein. These potential antiviral therapeutic and/or prophylactic agents can include existing antiviral agents known to affect viruses other than *Cryptovirus* (e.g., RibavirinTM, which is also known as VirazoleTM) and new potential antiviral agents. For example, molecular therapeutic agents (e.g., anti-sense nucleotides and ribozymes) or protease inhibitors can also be tested using the inventive *in vitro* and/or *in vivo* methods of screening. Agents that might inhibit the cleavage of the viral fusion protein (F₀) can be sought, and these could be particularly valuable, as there is evidence that cleavage of the fusion protein and its association with the viral hemagglutinin/neuraminidase protein (HN) are critical events in determining the pathogenicity of infections by other Paramyxoviridae (Yao *et al.*, *J. Virol.* **71**: 650-656, 1997). Further, these potential antiviral agents may be directed, for example, at *Cryptovirus* replication or assembly, or the expression or activity of *Cryptovirus* genes and proteins, such as, but not limited to, the *Cryptovirus*-encoded RNA-dependent RNA polymerase comprising the L protein and its companion P and V proteins. The inventive screening methods also can be employed to develop broad-spectrum antiviral agents, effective against viruses other than *Cryptovirus*.

Immunotherapeutic agents, such as those that attack, or stimulate the immune system to attack, infected cells or virus particles (by, e.g., passive antibody administration or introduction of *Cryptovirus*-specific monoclonal antibodies) are also amenable to testing because they can block or inhibit the assembly, release or cell-to-cell transfer of virions. However, administration of these agents may be of only limited value in "curing" persistent and chronic *Cryptovirus* infections because the virus appears to survive *in situ* by shutting off production of its envelope proteins and going into a "latent" or inapparent state, in which it appears to be undetectable by the immune system.

Appropriate amounts of potential prophylactic or therapeutic agents vary and are determined by routine screening.

In accordance with the inventive *in vivo* methods of screening a potential therapeutic agent or prophylactic agent, the agent is evaluated for an ability to induce, create, bring about, or result in a beneficial antiviral effect in the inventive animal model. A "beneficial antiviral effect" includes the prevention of infection with *Cryptovirus* or a reduction in the duration or severity of at least one symptom associated with *Cryptovirus* infection, in the animal subjected to the assay, compared to tissues in control animals. A "beneficial antiviral effect" also includes a prevention or reduction of cytopathic effect (CPE) in tissues sampled from the animal subjected to assay by the screening method. Also encompassed by a "beneficial antiviral effect" is an inhibitory effect on *Cryptovirus* replication and/or *Cryptovirus* virion assembly (e.g., inhibitory effect on *Cryptovirus* genomic replication, *Cryptovirus* transcription, and/or translation, i.e., protein synthesis, from *Cryptovirus*

mRNAs, or a diminution in the numbers of *Cryptovirus* virions produced or a relative lack of completeness of *Cryptovirus* particles, compared to a suitable control), which effect is measured by known means in cells or tissues sampled from the animal subjected to the assay.

Appropriate controls for use in the screening methods will be self-evident to the skilled artisan. Such controls can include: (1) animals administered with the same potential prophylactic or therapeutic agent and challenged with sterile artificial aqueous culture medium alone or the culture medium containing a strain of SV5; (2) animals mock-treated with saline (or the same carriers used in delivering the potential prophylactic or therapeutic agent) and challenged with *Cryptovirus*; and (3) animals mock-treated with saline (or the same carriers used in delivering the potential prophylactic or therapeutic agent) and challenged with sterile artificial aqueous culture medium alone or the culture medium containing a strain of SV5.

The practice of the present invention will employ, unless otherwise indicated, conventional or other known techniques of biochemistry, molecular biology, microbiology, virology, recombinant nucleic acid technology, and immunology, which are within the skill of the art. Such techniques are explained fully in the literature. (e.g., Maniatis *et al.*, *Molecular Cloning: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, USA (1982); *DNA Cloning*, Vols. I and II (D. N Glover ed. 1985); Sambrook *et al.*, *Molecular Cloning: A Laboratory Manual* (2 ed.), Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, USA (1989); Davis *et al.*, *Basic Methods in Molecular Biology*, Elsevier Science Publishing, Inc., New York, USA (1986); or *Methods in Enzymology: Guide to Molecular Cloning Techniques* Vol.152, S. L. Berger and A. R. Kimmerrl Eds., Academic Press Inc., San Diego, USA (1987); *Oligonucleotide Synthesis* (M. J. Gait ed, 1984); *Nucleic Acid Hybridization* (B. D. Hames & S. J. Higgins eds. 1984); *Transcription And Translation* (B. D. Hames & S. J. Higgins eds. 1984); *Animal Cell Culture* (R. I. Freshney ed. 1986); *Immobilized Cells And Enzymes* (IRL Press, 1986); B. Perbal, *A Practical Guide To Molecular Cloning* (1984); the series, *Methods In Enzymology* (Academic Press, Inc.); *Gene Transfer Vectors For Mammalian Cells* (J. H. Miller and M. P. Calos eds. 1987, Cold Spring Harbor Laboratory), *Methods in Enzymology* Vol. 154 and Vol. 155 (Wu and Grossman, and Wu, eds., respectively), Mayer and Walker, eds. (1987), *Immunochemical Methods In Cell And Molecular Biology* (Academic Press, London), Scopes, (1987), *Protein Purification: Principles And Practice*, Second Edition (Springer-Verlag, N.Y.), and *Handbook Of Experimental Immunology*, Volumes I-IV (D. M. Weir and C. C. Blackwell eds 1986).

The invention will now be described in greater detail by reference to the following non-limiting examples.

EXAMPLES

Example 1. Detection of *Cryptovirus* in Infected Cells and Isolation and Purification of *Cryptovirus* Viral Particles.

In accordance with the present invention, cell-free *Cryptovirus* particles were recovered from cells of the buffy coat (peripheral blood mononuclear cells; PBMNC) obtained from the peripheral blood of patients with SSPE. The technique used was modified from that of Robbins *et al.* (*J. Infect. Dis.* 143:396-403, 1981). Modifications included the addition of cyclic GMP (to a final concentration of 1 mM) to the aqueous culture medium, in accordance with the present invention, which medium was added to the initial PBMNC cultures and the primary cocultivate with human amnion cells (AV₃). Results were further optimized by using as the mammalian epithelial cell line in a co-cultivation step with the PBMNCs and amnion cells, a clonal subline of CV-1 cells (CV-1_c).

Successful isolation of the virus requires viable PBMNCs. Such PBMNCs were separated from other blood components by standard procedures on Ficoll-Hypaque™ gradient media. After centrifugation, the buffy coat cells banded at the interface of the media and were removed with a sterile pipette. They were then gently washed by dilution in 50 volumes of RPMI cell culture media containing 1-2% fetal calf serum and centrifuged in a table-top refrigerated centrifuge (1000 rpm for 5 minutes). The pelleted PBMNCs were then diluted to 2×10^5 cells per ml in RPMI media containing 10% fetal calf serum and 1 mM cyclic GMP (sodium salt) and incubated without disturbance at 37°C for 12-18 hours. Following this incubation, the cultures were seeded with sufficient AV₃ cells in Richter's Modified Minimal Essential Medium (IMEMZO) (supplemented with insulin, zinc and HEPES buffer, 2 mM L-glutamine, 200 Units penicillin/mL, 100 pg streptomycin/mL, 5-10% (v/v) fetal calf serum, pH between 6.8 and 7.0) to yield a net cell concentration of 2×10^5 cells per mL (for all cells in the culture) and were reincubated at 37°C. Once the cultures reached confluence (2-3 days), the monolayer was chelated with a solution of 0.02% w/v EDTA in CMF-PBS (calcium and magnesium free phosphate buffered saline), the cells were dispersed, and passaged at 2×10^5 cells/mL in IMEMZO as before.

The cultures were then blindly passaged in the same way when confluent every 3-4 days (roughly twice a week) for 2-3 weeks. After two weeks, a slide culture was prepared to examine the cells for the presence of *Cryptovirus*-specific inclusions in the cytoplasm using a *Cryptovirus*-specific indirect fluorescent antibody technique (exposure to hyperimmune rabbit anti-*Cryptovirus* antisera followed by labeling with fluorescein-conjugated goat anti-rabbit IgG).

When 5-10% of the cells were positive for *Cryptovirus*-specific inclusions, the cultures were ready for co-cultivation with the permissive CV-1_c cells mentioned above. This involved the 1:1

cocultivation of the passaged primary PBMNC/AV₃ cultures with CV-1_c cells in Richter's Modified Minimal Essential Medium diluted to yield a net concentration of 2×10^5 cells/mL. These cultures were then monitored for the development of subtle cytopathic effects (CPE; stellation and rounding of cells or the formation of multinucleated cells containing three or more nuclei) over the ensuing 4-5 days. If no CPE developed before the cultures become confluent, they were passaged and monitored again. If no CPE developed after three such passages, the cultures were discarded.

Once CPE was observed, the whole culture was frozen at -70°C, thawed, and the cells were dispersed and dispensed into 1.0 mL aliquots. These aliquots represented the putative primary isolation of the virus. The virus was then plaque-purified by titration on monolayers of CV-1_c cells overlaid with a semi-solid solution of 1% w/v sodium carboxymethylcellulose (NaCMC) containing 2% fetal calf serum, which was made up in Richter's Modified Minimal Essential Medium. The cultures were incubated at 37°C in a partial CO₂ atmosphere (5% v/v). Plaques formed in 8-12 days and were then picked and replaques, as above. Once triply plaque-purified, the virus was aliquoted onto CV-1_c monolayers (in IMEMZO media containing 5% fetal calf serum and supplements listed above) in 25-75 cm² tissue culture flasks. Once sufficient CPE developed, involving half or more of the cultured cells, the whole cultures were frozen, thawed, and the lysate was dispersed, re-aliquoted and refrozen at -70°C. Samples of the virus stock were then titrated for further use by the method of Robbins *et al.* (Robbins *et al.*, *J. Infect. Disease* 143:396-403, 1981).

Density Gradient Purification: Virions and intracellular nucleocapsids isolated from productively- (Vero and CV-1_c) and nonproductively-infected (AV₃/SSPE) cells were further purified on sucrose-potassium tartrate gradients (virions) and CsCl gradients (nucleocapsids) by the method of Robbins *et al.* (Robbins *et al.*, *J. Infect. Disease* 143:396-403, 1981; Rapp and Robbins, *Intervirology* 16:160-167, 1981; Robbins and Rapp, *Arch. Virol.* 71:85-91, 1982; and Robbins and Abbott-Smith, *J. Virol. Meth.* 11:253-257, 1985).

Example 2. Cryptovirus Propagation and Virion Isolation and Purification.

Once isolated, cell-free virus stocks were grown in simian epithelial cell lines (*e.g.* Vero or CV-1 cells). The *Cryptovirus* isolates used in the studies described herein were triply-plaque purified and grown in a clonal subline of CV-1 cells designated CV-1_c. Optimal production of infectious virus occurred when using IMEMZO supplemented with insulin, zinc and HEPES buffer, 2 mM L-glutamine, 200 Units penicillin/mL, 100 µg streptomycin/mL, 5-10% (v/v) fetal calf serum, at a pH between 6.8 and 7.0.

5 The presence of insulin, and optionally zinc dication, in the tissue culture medium was useful in obtaining viable titers of infectious virus. Independent attempts to grow the virus in CV-1 cells using standard media (e.g. MEM) produced very poor results. Conversely, the expression of *Cryptovirus* proteins and the productivity of *Cryptovirus* infections in primate cell cultures was dramatically enhanced (50- to 100-fold) by addition of cyclic GMP (1 mM; sodium salt) to standard media (specifically MEM). The enhancement obtained was very similar to the enhancement of measles virus replication published earlier (Robbins, *Intervirology* 32:204-208, 1991).

10 *Virion Isolation and Purification.* Virions were isolated and further purified from the supernatant tissue culture medium of acutely infected CV-1 cells 72 hours after infection. The procedure employed involved the separation of the virus particles by differential centrifugation.

15 The supernatant medium of infected cultures was decanted into a sterile plastic 50-mL Falcon centrifuge tube and clarified at 2000 rpm for 10 minutes. The supernatant was then transferred to an impact resistant glass centrifuge tube (Sorvall) and further clarified at 10,000 rpm for 10 minutes. All clarifications took place at 4° C in an RC2B Sorvall centrifuge. The supernatant fluid from the second clarification step was layered over a 60% w/v sucrose cushion (in 10 mM Tris, 5 mM EDTA, pH 7.2) and centrifuged at approximately 130,000 x G in a Beckman SW-28 rotor at 4° C for 90 minutes in a Beckman L70 ultracentrifuge. Materials were collected from the tissue culture medium-sucrose interface, pooled, diluted with tissue culture medium and recentrifuged onto another 60% sucrose cushion as described above. The materials at the interface were again removed, diluted with tissue culture medium, and centrifuged at 35,000 rpm (280,000 x G) for 60 minutes through a 30% w/v over 60% w/v discontinuous (i.e. layered) sucrose gradient prepared in the Tris EDTA buffer described above). The virions were collection from the 30%:60% sucrose interface, diluted with cold Tris EDTA buffer and pelleted in a Beckman SW41 rotor at 41,000 rpm and 4° C for 60 minutes. Pelleted virions were resuspended in a variable amount of the cold Tris EDTA buffer and frozen at – 70° C until further use. Total protein in each virion preparation was determined by the method of Lowry et al. (1951).

Example 3. Preparation of Antisera.

Antisera were raised in adult New Zealand White rabbits against sucrose-potassium tartrate gradient-purified virions of *Cryptovirus*, CsCl gradient-purified nucleocapsids (from infected CV-1_c cell cytoplasm), and against the major viral nucleocapsid protein, NP, eluted from polyacrylamide gels after SDS-PAGE. Rabbit antisera were also raised against the NIH 21005-2WR strain of SV5, and the Edmonston strain of measles virus.

Animals were inoculated by a pincushion technique which involved three series of three separate inoculations in each animal using a sterile 27 gauge needle and 1.0-mL syringe (one inoculation intradermally on the back; one inoculation intraperitoneally and one inoculation in a hind foot pad). The first series of inoculations were made using gradient purified and dialyzed virions (100 µg of virions in 0.3 mL of a 10 mM Tris 5 mM EDTA solution) mixed 1:1 with Freund's Complete Adjuvant and each inoculation contained approximately 200 µL of the virion:adjuvant mixture. The second series of inoculations were made two weeks later in the same locations but on the opposite side of each animal and consisted of the same amount of virions mixed with Freund's Incomplete Adjuvant. The third series of inoculations were made two weeks later in the same locations as the first inoculations but using only virions (diluted in 0.6 mL of the Tris-EDTA solution described above). Blood was harvested by intracardiac exsanguination of the animals two weeks after the final series of inoculations. The harvested blood was centrifuged (2000 rpm for 10 minutes) and allowed to clot on ice. The upper serum component was harvested and adsorbed against the pelleted component (2000 rpm for 10 minutes) of saline-washed freeze-dried acetone:methanol-extracted monkey kidney tissue (4° C for 1 hour with agitation every 15 minutes). The adsorbed serum was harvested by centrifugation (2,000 for 10 minutes) and stored in 1.0-mL aliquots at -20° C).

All of the anti-*Cryptovirus* antisera were strongly reactive with the corresponding *Cryptovirus*-specific materials from which they were generated when analyzed by (1) immunoprecipitation, (2) immunofluorescence, (3) immunoblotting, (4) ultrastructural immunolabelling techniques (immunogold), and (5) in the case of antisera generated against gradient-purified virus particles, neutralization titration assays. All the hyperimmune virus-specific antisera that were generated in the rabbits had homologous neutralization titers in excess of 1280 and, usually, in excess of 2560 (reciprocal dilution of PRD₅₀).

All experimentally-generated antisera were adsorbed against saline washed, freeze-dried, acetone:methanol extracts of monkey kidney tissue or similar extracts of AV₃ cells and/or CV-1_c cells.

While clinical sera were similarly adsorbed, CSF specimens were NOT preadsorbed due to the small volumes that were usually available and the requirement to retain aliquots for duplicate and parallel studies.

The precipitating "titers" of the experimental sera raised against purified nucleocapsids and purified viruses were not specifically determined although, routinely, 5-10 μ L were used in positive control immunoprecipitation reactions and 25 μ L of positive control antisera (diluted 1:10 or 1:20) for positive controls in ELISA assays.

Cryptovirus-specific antisera were also produced in mice experimentally inoculated with gradient-purified infectious *Cryptovirus* virions. These antisera were analyzed by immunoprecipitation, and were found to strongly precipitate all *Cryptovirus* envelope proteins.

There was clear *asymmetric* cross-reactivity between the antisera raised against *Cryptovirus* virions and antiserum raised against virions of the NIH 2WR-21005 strain of SV5. The asymmetry observed in this regard was always such that the heterologous reactions (i.e., *Cryptovirus*-specific antisera vs. SV5 materials and SVS-specific antiserum vs. *Cryptovirus* materials) were two- to four-fold weaker than the homologous reactions (i.e., *Cryptovirus*-specific antisera vs. *Cryptovirus* materials and SVS-specific antisera vs. SV5 materials). Another antiserum, which was independently prepared against NIH 21005-2WR strain of SV5 and kindly provided by Dr. Purnell Choppin, behaved in a similar asymmetric manner to the antiserum against SV5 described hereinabove.

Such cross reactivity is not surprising. Precisely the same sort of asymmetric cross-reactivity occurs when examining other paramyxovirus systems (e.g., there is also a two to four fold *asymmetric* cross-reactivity between measles virus antibodies when reacted with the closely related viruses of canine distemper and rinderpest and *vice versa*). There was also limited (i.e., much weaker) cross-reactivity between *Cryptovirus*-specific antibodies and other paramyxoviruses (e.g., measles virus).

Example 4. Characterization of isolated *Cryptovirus*.

Cryptovirus Neurovirulence and Neurotropism. As shown in Fig. 24A, *Cryptovirus* demonstrated a tropism for neurons. Intracranial inoculation of mice with infectious *Cryptovirus* or nonproductive virus-carrying cells (AV₃/SSPE) resulted in the subacute/slow development of a spectrum of neuropathological conditions that had epileptiform, neurological and/or neuropsychological components. These responses were similar to the "experimental SSPE" reported earlier in animals following inoculation with "cell-associated measles-like" virus such as Niigata, Kitaken and Biken (see Fig. 24B; Doi *et al.*, *Japan. J. Med. Sci. Biol.* 25:321-333, 1972; Ueda *et al.*,

Biken Journal 18:179-181, 1975; Yamanouchi *et al.*, *Japan. J. Med. Sci. Biol.* 29:177-186, 1976; Ohuchi *et al.*, *Microbiol. Immunol.* 25:887-983, 1981).

5 *Cryptovirus Presence in Neurovirulent SSPE-derived Virus-carrying Cell Lines.* Four virus-carrying cell lines derived from patients with SSPE were tested by immunofluorescence for the presence of measles virus- and/or *Cryptovirus* specific antigens. These were AV₃/SSPE/MV (an SSPE-derived cell line derived from PBMNC from an SSPE patient cocultivated with AV₃ cells which was experimentally-infected with Edmonston strain measles virus; Robbins, unpublished data);
10 the nonproductive SSPE-derived cell line designated "Kitaken" (Ueda *et al.*, *Biken Journal* 18:179-181, 1975); the nonproductive SSPE-derived cell line designated "Niigata" (Doi *et al.*, *Japan. J. Med. Sci. Biol.* 25:321-333, 1972); and the nonproductive SSPE-derived cell line designated "Biken" (Yamanouchi *et al.*, *Japan. J. Med. Sci. Biol.* 29:177-186, 1976; Ohuchi *et al.*, *Microbiol. Immunol.* 25:887-983, 1981). With the possible exception of the Niigata cell line, all of these virus-carrying cell
15 lines expressed both measles virus-specific and *Cryptovirus*-specific antigens when examined by virus-specific immunofluorescent techniques (shown in Fig. 6). Given that no cell-free clinical isolates of measles virus have ever been shown to cause SSPE-like illnesses in experimentally-infected animals, the presence of *Cryptovirus* in these cultures strongly suggests that the subacute/slow neuropathies seen in these animals are due to the presence of *Cryptovirus* in the cultures—not
20 measles virus.

Cryptovirus inclusion bodies (i.e. cytoplasmic nucleocapsids) displayed the same sort of "peppery" and or "splattered" distribution in both acutely-infected cells (CV-1_C) and nonproductively- and persistently-infected cells (AV₃/SSPE) as that previously described in CNS biopsy and autopsy materials from SSPE patients and in SSPE-derived nonproductive virus-carrying
25 cell lines (e.g., de Felici *et al.*, *Annales Microbiologie* 126:523-538 [1975]; Makino *et al.*, *Microbiology and Immunology* 21:193-205 [1977]; Brown *et al.*, *Acta Neuropathologica* 50:181-186 [1980]). This is most clearly evident in Panels B, D, F, H and J of Fig. 6. These characteristics were in sharp contrast to the discrete and "coalescing" distribution and morphology of intracellular measles virus inclusions bodies (see Panels A, C, E and G of Fig. 6).

30 *Neutralization Titration Assay.* Formation of macroscopically visible plaques on monolayers of mammalian cells (e.g., BHK, Vero and CV-1_C) can be used to quantitate preparations of infectious *Cryptovirus*. Plaque formation can be inhibited by serial dilutions of clinical serum specimens and

Cryptovirus-specific antisera generated in rabbits (see Robbins *et al.*, *J. Infect. Disease* 143:396-403, 1981). Plaque titration assays were conducted to determine the PRD₅₀ of isolated *Cryptovirus*.

Briefly, ten-fold serial dilutions of serum or CSF specimens to be tested were incubated for one hour at 4°C with sufficient infectious virus to yield a net plating concentration of between 100-200 plaque forming units of the virus / 0.2 mL of final diluent (including the diluted serum or CSF). After incubation, 0.2 mL of the diluted virus-serum (or virus-CSF) mixtures was then plated onto monolayers of susceptible cells (*e.g.* Vero or CV-1_c) and the cells were incubated at 37°C in a partial CO₂ atmosphere (5%v/v) (with redistribution of the inoculum every 15 minutes). At the end of the incubation period, inoculated monolayers were overlaid with sufficient volumes of a 2% (w/v) solution of carboxymethylcellulose (made up in IMEMZO medium containing 2% fetal calf serum, insulin, zinc, and HEPES buffer, 2 mM L-glutamine, 200 Units penicillin / mL, 100 µg streptomycin / mL, pH between 6.8 and 7.0) to last 10-12 days (*i.e.*, enough volume so that the monolayers won't dry out). The plates were not moved during the incubation period. After 10-12 days, the overlay was aspirated and the cells were fixed with formalin fixative and stained with a protein stain (*e.g.*, Giemsa). The number of plaques formed on each plate was then enumerated and the PRD₅₀ calculated.

In particular, cross neutralization assays involved the determination of the titer of antisera made against each species of virus which would neutralize 50% of the plaque forming units (PFUs) of each virus. Virus stocks of the BBR strain of *Cryptovirus* and the NIH 21005-2WR strain of SV5 were diluted in serum-free minimal essential medium (Eagle's MEM containing 2mM L-glutamine, 200 units of penicillin and 100 µg of streptomycin/ml with the pH adjusted to between 6.8 and 7.0 with NaHCO₃) to a titer of 1,000 PFUs per mL (resulting in 100 PFUs per well after dilution and plating). Antiserum raised in New Zealand White rabbits was serially-diluted in 10-fold increments in the same serum-free MEM. Aliquots (0.5 mL) of the diluted virus stocks were then mixed with 0.5 ml aliquots of each dilution of the antisera, gently mixed with a vortex and incubated on ice for one hour with gentle mixing every 15 minutes. Following this incubation period, the medium was aspirated from monolayers of CV-1 cells in 6-well cluster plates (NUNC), the monolayers were washed with warm saline, the saline was aspirated and 0.2 mL of each of the diluted antisera-virus incubates were plated onto two monolayers. The inoculated cluster plates were subsequently incubated at 37° C in a CO₂ incubator (containing 5% CO₂ v/v) for 1 hour with manual redistribution of the inocula every 15 minutes. Following this adsorption period, each well was overlaid with 10.0 mL of a semisolid overlay medium (1% w/v sodium carboxymethylcellulose in Eagle's MEM containing 2 mM L-glutamine, 200 units of penicillin and 100 µg of streptomycin/ml, 2% v/v fetal

calf serum with the pH adjusted to between 6.8 and 7.0 with NaHCO_3) and incubated for 10-12 days at 37° C in a CO_2 incubator (containing 5% CO_2 v/v) without being disturbed. Following this incubation period, the overlay was aspirated, the monolayers were gently washed with warm saline, and then fixed in formalin fixative (3.7% by weight formaldehyde gas in saline) for 1 hour or longer.

5 Following fixation, the fixative was aspirated and the fixed monolayers were gently washed with distilled water and stained with 1-2 mL per well of Giemsa stain (0.5 gm Giemsa powder dissolved in 42 mL of warmed [55° C] glycerin, 42 mL of absolute methanol, filtered and diluted 1:5 with formalin fixative immediately before use) for 1 hour at room temperature. The stain was subsequently decanted and the monolayers were washed under tap water and allowed to dry at room

10 temperature. Plaques on monolayers were illuminated on a light box, enumerated under a magnifying lens and recorded for each dilution, virus and antisera series. The neutralization titer of each antiserum virus series was calculated to be the reciprocal of the dilution of antisera resulting in a 50% decrease in the number of plaques formed.

The calculated neutralization titer for each crossed neutralization set (i.e. anti-*Cryptovirus* antiserum versus *Cryptovirus* and anti-*Cryptovirus* antiserum versus SV5; anti-SV5 antiserum versus *Cryptovirus* and anti-SV5 antisera versus SV5) was consistently 2-4 fold less for the heterologous mixtures (i.e. anti-*Cryptovirus* antisera versus SV5 and anti-SV5 antisera versus *Cryptovirus*) than for the homologous mixtures (anti-*Cryptovirus* antisera versus *Cryptovirus* and anti-SV5 antisera versus SV5). On no occasion did any heterologous mixture have less than a 2-fold difference when

20 compared to the homologous pair (in three separate trials).

Cryptovirus Ultrastructural and Immunoultrastructural Characterization. AV₃/SSPE/MV cells (AV₃/SSPE cells persistently and nonproductively infected with the Edmonston strain of measles virus), AV₃/SSPE cells, and CV-1_c cells acutely infected with the BBR strain of *Cryptovirus*

25 were fixed in situ (on glass cover slips) in 2% formaldehyde and picric acid and 3% glutaraldehyde in 0.1 M cacodylate buffer, pH 7.2, for 15 minutes at room temperature. Osmium tetroxide post-fixation was omitted for specimens that were to be treated with antibody (i.e. which were prepared for immunoultrastructural studies). Cover slips with fixed cells were washed in three changes of cacodylate buffer, dehydrated to 70% ethanol and embedded in LR White resin. Resin was

30 polymerized at 50°C for 24 hours. Ultrathin sections were cut and mounted on uncoated nickel grids.

Stained thin sections of CV-1_c cells acutely-infected with the BBR strain of *Cryptovirus* and AV₃/SSPE cells were examined by electron microscopy. In infected CV-1_c cells, pleomorphic virion particles, 100-120 nm in diameter, were seen budding from the surface of acutely-infected cells and

numerous accumulations of filamentous structures (helical nucleocapsids, 15-17 nm in diameter) were observed in the cell cytoplasm (data not shown). Both the virions and nucleocapsids were similar to those described for other members of the Paramyxoviridae. While virions were not observed budding from the surfaces of AV₃/SSPE cells, inclusions of intracellular nucleocapsids were seen in abundance and these were identical to those seen in the acutely-infected cells.

The intracellular nucleocapsids of nonproductively and productively-infected mammalian cells can readily be localized under the electron microscope using *Cryptovirus*-specific or *Cryptovirus* nucleocapsid-specific hyperimmune rabbit antibodies and an indirect immunogold labeling technique.

Immunolabelling was performed on sections of AV₃/SSPE/MV cells by floating sections mounted on nickel grids (processed as described without osmium tetroxide post-fixation) on drops of solution (see below) in a closed, humid chamber. Sections were etched according to the method of Ingram *et al.* (Parasitology Research 74:208-215, 1988). Non-specific labeling was reduced by incubation of the sections with 5% bovine serum albumin in modified Tris buffer (20 mM Tris, 0.5 M NaCl, 20 mM sodium azide and 0.05% Tween 20, pH 8.2) for 30 minutes at 37°C prior to immunolabeling. The modified Tris buffer was used for all dilutions and washes.

In single labeling experiments, sections were incubated with rabbit antisera (anti-Edmonston measles virus or anti-BBR strain of *Cryptovirus*) diluted 1:20 in modified Tris buffer for 2 hours at 37°C, washed in three changes of buffer, and incubated with a goat anti-rabbit IgG colloidal gold (10 or 15 nm particle size, 1:20 dilution, 1 hour at 37°C). After washing with two changes of buffer, followed by two changes of distilled water, sections were lightly contrasted with 2% uranyl acetate and led citrate, and examined in a JEOL 1200EX transmission electron microscope.

In double labeling experiments, sections were immunolabeled on one face, as described for single labeling, using rabbit anti-Edmonston measles virus and 15 nm colloidal gold particles, and ensuring that the reverse face of the section was not contaminated by labeling solutions. The labeled face was then coated with a thin film of Celloidin to reduce possible cross reaction of antibodies while the reverse face of the section was labeled. Immunolabeling of the reverse face of the sections was performed as described above, using the second antiserum (rabbit anti-BBR strain of *Cryptovirus*) and 10 nm colloidal gold particles. Examination of these double labeled sections allowed simultaneous comparison of labeling patterns of the two antisera.

The results of these studies were unequivocal and are shown in Fig. 25. The first labeling sequence (15 nm gold beads) labeled only the wider "fuzzy" measles virus nucleocapsids (as shown

in Fig. 25B), while the second labeling sequence (10 nm gold beads) labeled only the narrower smooth *Cryptovirus* nucleocapsids (as shown in Fig. 25A).

Example 5. Characterization of Isolated *Cryptovirus* Proteins

5 *Radioimmunoprecipitation (RIP) Assay:* Extensive data were generated by the comparative analysis of *Cryptovirus*-specific immunoprecipitates of [³⁵S]-methionine-labeled uninfected, nonproductively- and productively-infected human and primate cells by sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE; see below).

10 CV-1_c or Vero cell monolayers were infected with the BBR strain of *Cryptovirus*, the NIH 21005-2WR strain of SV5 or the Edmonston strain of measles virus at a multiplicity of infection of 1-2 PFU/cell using procedures as described elsewhere (Robbins and Rapp, *Virology* 106:317-326, 1981).

15 Labeling was accomplished twenty-four hours after infection by the following procedure. Tissue culture medium was removed from infected cell monolayers and the cells were washed with a serum and methionine-free Eagle's based MEM (starvation medium). The infected cultures were then supplemented with the starvation medium for 60-90 minutes and incubated at 37°C. Following the starvation period, cultures were labeled with the starvation medium containing 100 µCi/mL of [³⁵S]-methionine (Amersham). Labeling was carried out at 37°C in a 5% CO₂ atmosphere for 5-6 hours. Immunoprecipitations were carried out according to the procedure of Lamb *et al.*, *Virology* 20 91:60-78, 1978.

25 *SDS-PAGE:* Virions and immunoprecipitates were analyzed under denaturing and reducing conditions on 10% (or to detect the presence of very small peptide species [e.g., SH protein], 20%) polyacrylamide slab gels (Laemmli, 1970). After electrophoresis, gels were treated with a fluor solution (Amersham), were dried, and were then exposed to X-ray film.

30 Purified virions of the virus were analyzed by SDS-PAGE under reducing and non-reducing conditions (see Fig. 11, an autoradiogram of gradient-purified [³⁵S]-methionine-labeled *Cryptovirus* virions produced in acutely-infected Vero cells after SDS-PAGE under reducing conditions). The approximate molecular weights of the proteins indicated on the right side of the figure were calculated by comparing their migrations to marker proteins of known molecular weights (Sigma Chemical Co., St. Louis, MO). The annotations are defined in the brief description of the drawing.

The SH protein, a small envelope-associated protein having a MW of about 5 kD, is not shown because it has run off the gel (see below).

SH Protein. Due apparently to the small size of the SH protein and its relatively low methionine content, the SH protein was difficult to detect in radio-labeled virion preparations of both the BBR strain of *Cryptovirus* and the NIH 21005-2WR strain of SV5. When unlabeled purified virion preparations of both viruses were run on 20% polyacrylamide slab gels under denaturing and reducing conditions alongside of low molecular weight marker proteins (BioRad) and stained with a silver-staining technique (BioRad), a small protein with an M_r of approximately 5 kD, identified as the SH protein, was found in both *Cryptovirus* and SV5 virion preparations. There was no detectable difference between the migration of the SH protein from the BBR strain of *Cryptovirus* or the NIH 21005-2WR strain of SV5.

F₀ and HN Co-migration Anomaly. Although the major envelope proteins (F₀ and HN) of many Rubulaviruses (e.g., SV5) were readily discernible as separate bands on SDS-PAGE gels, the larger size of the *Cryptovirus* F₀ protein (i.e., +22 amino acids) resulted in a significantly slower rate of migration for this protein (M_r = 69 kD). As shown in Figs. 13A-B, close examination of such gels enabled one to discern both proteins, albeit with some difficulty. Figs. 13A and 13B show photographs of migration patterns of the major *Cryptovirus* envelope proteins, F₀ and HN. Fig. 13A illustrates the observed near co-migration of the major *Cryptovirus* envelope proteins, F₀ and HN. Enlargement of the RIP from the CSF-positive patient (right lane) in Fig. 13A shows the "bowed" or "crested" structure that resulted from the near co-migration of the F₀ and HN proteins of *Cryptovirus*. A diagrammatic interpretation of the near co-migration of the F₀ and HN proteins of *Cryptovirus* and the discrete migration of the analogous proteins of Simian virus 5 are shown in Fig. 13B.

Example 6. Experimental Infection of Mice: Creation of an Animal Model.

Infectious *Cryptovirus* stocks (prepared in CV-1_C cells) and live nonproductively-infected AV₃/SSPE cells were used to intracerebrally inoculate two strains of laboratory mice (Quackenbush and Colored, an outbred strain of C57 Black).

Briefly, neonatal mice (1-2 days old) were inoculated by injection with 0.025 mL (phosphate buffered saline, pH 7.4) containing either 5×10^4 PFU of cell-free *Cryptovirus* or 5×10^3 nonproductively infected human amnion cells (AV₃/SSPE). Following inoculation, the neonatal mice were returned to their mothers who were provided food and water *ad libitum*. Observations of

inoculated mice were made daily. Symptom of disease first appeared in affected animals after 21 days, and in others not until after more than 60 days. Observed symptoms included cachexia, muscle spasms, tremors, compulsive behaviors (e.g., extended periods of scratching, rubbing, or running in circles), hyperactivity/hyperesthesia, seizures and convulsions, and stupor. These results demonstrated that intracerebral inoculation with *Cryptovirus* results in subacute central nervous system (CNS) disease. Neurological, neurodegenerative, and/or neuropsychiatric disease presentation in mice is virtually indistinguishable from presentation in humans.

While all of the inoculated animals developed antibodies to the nucleocapsid protein of the virus (NP), not all of them developed antibodies to the envelope proteins (F, HN, and SH). More than 90% of the mice inoculated with *Cryptovirus* virions developed antibodies to the envelope proteins but only 33% of those inoculated with AV₃/SSPE cells did so.

Concurrently, while many of the animals inoculated with infectious *Cryptovirus* stocks developed profound neuropathological disease, fewer of the animals inoculated with nonproductively-infected AV₃/SSPE cells developed such illnesses, and there was a strong correlation between development of antibodies to the envelope proteins of the virus and the development of CNS symptoms. This suggests that the development of CNS disease depends on the establishment of an acute or subacute CNS infection by the virus and the expression of all of the virus' structural proteins in some cells or tissues.

More detailed examples follow:

Quackenbush mice. Two litters of 1-2 day old Quackenbush mice were intracerebrally-inoculated (in the right cerebral hemisphere) with 5×10^4 PFUs of either *Cryptovirus* (strain BBR)(10 individuals) or measles virus (Edmonston strain)(8 individuals), were returned to their mothers and were periodically observed over a period of three months. Two animals (one inoculated with *Cryptovirus* and one inoculated with measles virus) were found dead and partially consumed the next morning. Their deaths were attributed to "needle trauma" and/or maternal cannibalism. While none of the mice inoculated with measles virus developed any neurological, neurodegenerative, physiological or neuropsychiatric symptoms over the course of the study, two of the male mice inoculated with *Cryptovirus* developed atrophy and contralateral hindlimb paralysis (in their left hind legs) three to four weeks after inoculation. A third (female) mouse was observed dragging its left hind leg (unatrophied) approximately four weeks after inoculation but was found killed and partially eaten a day later. While 3 of 9 animals inoculated with *Cryptovirus* developed hind limb paralysis (33%), none of the animals showed overt signs of seizure, wasting or neuropsychiatric symptoms

over the course of the study. Hind limb paralysis was also seen in a number of the offspring of adult female Quackenbush mice that had been inoculated with *Cryptovirus* as newborns but that did not develop any overt symptomology. The frequency of this phenomenon was difficult to assess because the mothers tended to cannibalize the newborn animals that were born with, or subsequently developed, such characteristics.

Colored mice. Three litters of 1-2 day old Colored mice (comprising 26 individuals) were observed daily following intracerebral inoculation with 5×10^4 PFUs of *Cryptovirus* (strain BBR), Simian Virus 5 (NIH 21005-2WR strain) or measles virus (Edmonston strain) or mock-infected cells. Half of each litter was inoculated with *Cryptovirus* (13 animals) while the other half was apportioned into three groups and inoculated with either Simian Virus 5 (6 animals), measles virus (4 animals) or mock-infected CV-1_C lysate (3 animals). Each group was marked with phenol red stain on the upper skin of one foot to distinguish them (i.e. right front, left front, right rear, left rear). One animal inoculated with *Cryptovirus* died between 24 and 48 hours post-inoculation and this was attributed to “needle trauma/starvation” as it had stopped feeding (or was rejected) when it was returned to its mother. Between three and four weeks later, one male and one female animal were found dead in their cages in tonic-clonic posture—both having been inoculated with *Cryptovirus*. It was noted that both also appeared to be underweight when compared to their littermates. Two months after inoculation with *Cryptovirus*, a third mouse (male) was observed to have cachexia, anorectic wasting, tremors and seizures (Fig. 7A). Over the next month (approximately 11 weeks after inoculation), a fourth animal (also male) developed tremors and seizures although no wasting was observed (data not shown). A male littermate of the *Cryptovirus*-infected mouse shown in Fig 7A, which was inoculated with the NIH 21005-2WR strain of SV5, is shown in Fig. 7B. The same results were obtained when mice were inoculated with either the Edmonston strain of measles virus, mock-infected CV-1_C cells, or homogenized AV₃/SSPE cells (i.e. all remained healthy and none developed neurological, neurodegenerative or neuropsychiatric symptoms; data not shown).

Over the ensuing six months (observed up to nine months post inoculation), a significant number of the remaining animals (4 of the remaining 8) inoculated with *Cryptovirus* developed physiological, neurological and/or neuropsychiatric symptoms. Such late onset animals presented with symptoms that were in marked contrast to the overt seizure disorders observed in the subacute onset animals (i.e. those that developed symptoms in first three months post inoculation). These symptoms were dominated by physiological, neuropsychiatric and behavioral disturbances rather than more overt neurological symptoms and included: marked weight gain, extreme aggressive/passive

responses to stimuli, obsessive/compulsive behaviors, ataxia and tremor. Aggression was most frequently characterized in afflicted animals by physical agitation and a predisposition to biting when handled. Passive animals tended to eat excessively, gain weight, and sleep. Repetitive behavior was also sporadically observed and consisted primarily of endless pacing and/or facial washing so extreme as to result in the loss of fur on the head and neck and the development of abrasive wounds. One of the animals (a female shown in Fig. 8A) had abnormal cranial structure (microcephaly) and manifested a spectrum of physiological and behavioral symptoms at six months including obesity, tics and muscle twitching (along the back and left side) and obsessive/compulsive facial washing and scratching. Episodes of such obsessive/compulsive behavior were observed to last for an hour or more. A second female animal (shown in Fig. 8B) appeared overtly normal during the first five months post inoculation but between five and six months began displaying marked ataxia, tremors and aggression. While this animal maintained a normal body weight and appearance, it was prone to splaying its feet to maintain its balance when resting and stumbling when walking and biting and hissing/snarling when handled or disturbed. Neither animal shown in Fig. 8 developed overt (*grand mal*) seizures, in contrast to the animal shown in Fig. 7A. Two other animals (one male and one female) also developed mixtures of the slow onset symptomology (data not shown). Overt seizure activity was never observed in any of the late onset animals and none of the animals inoculated with SV5, measles virus or mock-infected cells developed any similar symptoms.

Of the 13 neonate animals inoculated with *Cryptovirus*, one died from needle trauma (sex uncertain), two died in tonic-clonic posture with signs of wasting (indicative of sudden death due to *status epilepticus* (one male; one female); two became wasted and developed grand mal seizures (both males); four developed a spectrum of slow onset neurological/neuropsychological symptoms (three females, one male); one committed infanticide after being bred (a female); and three never developed any symptoms (two females and one male). Removing the needle trauma death from the equation, 9 of 12 animals developed neurological, degenerative and/or neuropsychiatric symptoms (75%). Removing the two clonic-posture deaths as well, 7 of 10 animals developed symptoms (70%). This was highly significant compared to the combined results for the control inoculated mice (7 of 10 mice inoculated with *Cryptovirus* presenting with neurological symptoms versus 0 of 13 mice in the control groups; $P = 0.0005$, 2-sided Fisher exact test), and was also significant compared to just the SV5 inoculated mice (7 of 10 versus 0 of 6; $P = 0.01$, 2-sided Fisher exact test) and even when compared to just the measles inoculated mice (7 of 10 versus 0 of 4; $P = 0.035$, one-sided Fisher exact test). Statistical significance was not quite reached compared to the mice inoculated with uninfected cell lysate, because of the small number of mice in this group (7 of 10 versus 0 of 3, $P = 0.069$,

single-sided Fisher exact). However, when the two clonic-posture deaths were included even this small control group was significantly different (9 of 12 versus 0 of 3, $P = 0.044$, 2-sided Fisher exact). Thus, the disease(s) and symptoms resulting from *Cryptovirus* infection was profoundly significant.

5 *Infanticide.* Of the four Colored mice which were inoculated as neonates with *Cryptovirus* but did not develop subacute or slow onset symptoms over the first nine months of the study, three were female and were subsequently bred with uninfected males at nine months of age. While all of the offspring of two of the three resulting litters developed normally, all of the offspring in one of the litters (comprising 10 animals) were killed and wholly or partially cannibalized by their mother. Such
10 infanticide did not occur in litters to females that had been inoculated with SV5, measles or mock-infected cells. In a separate study (see below), one of the females which was inoculated with live AV3/SSPE cells—but did not develop any overt neurological symptoms—developed late physiological and behavioral symptoms and also committed infanticide of its whole litter after being bred with an uninfected male.

15 *Animal Model Employing Inoculation of Nonproductively Infected AV₃/SSPE cells.* In a separate study, two litters of neonatal mice (18 animals) were inoculated (1-2 days after birth) with either live or homogenized AV₃/SSPE cells (six animals each) or live or homogenized AV₃ cells (three animals each). There were no needle trauma deaths. While none of the animals inoculated
20 with the homogenized AV₃/SSPE cells, live AV₃ cells or homogenized AV₃ cells (12 animals) developed any subacute or slow onset symptoms whatsoever, one of the six animals inoculated with live AV₃/SSPE cells developed subacute degenerative and neurological symptoms (a male) and one developed slow onset symptoms (1 female). The animal that developed subacute symptomology began presenting with symptoms 24 days after intracerebral inoculation. Over a five day period, the
25 symptoms presented included cachexia, wasting, hunched posture, repetitive chirping and clicking, hyperesthesia, incontinence of urine, tremors, muscle spasms and coma. Overt seizures were not observed. It was sacrificed when coma developed. The animal that developed slow onset symptomology (a female) presented at five to six months (post-inoculation) with repetitive pacing, aggression and progressive obesity. The animal was bred at six months with an uninfected male.
30 Seven days after delivering a litter of eight offspring, it killed the whole litter and partially consumed each individual. No seizures or other signs of overt neurological symptoms were observed. None of the other females which were inoculated with homogenized AV₃/SSPE cells, AV₃ cells or homogenized AV₃ cells and bred (six animals in total) committed infanticide.

Example 7. Cryptovirus-Specific Antibodies Are Present In The Serum And Cerebrospinal Fluid (CSF) Of Human Patients Diagnosed With Neurological, Neurodegenerative And/Or Neuropsychiatric Diseases.

5 Evidence for the presence of *Cryptovirus*-specific antibodies to the major envelope proteins of the virus (F₀ and HN) in the serum and CSF of patients was determined by immunoprecipitation of [³⁵S]-methionine-labeled *Cryptovirus*-specific proteins produced in acutely-infected CV-1_c cells. Figs. 12A and 12B are photographs of autoradiograms, which serve as examples of RIP profiles of measles virus- or *Cryptovirus*-specific proteins precipitated from [³⁵S]-methionine-labeled acutely
10 infected CV-1_c cells by clinical CSF specimens followed by SDS-PAGE (reduced). In Fig. 12A, lane "V" contains gradient-purified *Cryptovirus* virions from acutely-infected, [³⁵S]methionine-labeled CV-1_c cells (BBR Strain). Lane "MV" contains proteins precipitated by the CSF of an 11-year old male SSPE patient from radiolabeled CV-1_c cells acutely-infected with measles virus (*Edmonston Strain*). Lane "B" contains proteins precipitated by the same CSF specimen from a 1:1
15 mixture of radiolabeled CV-1_c cells acutely-infected with either measles virus or *Cryptovirus*. Lane "CV" contains proteins precipitated by the same CSF specimen from radio-labeled CV-1_c cells acutely-infected with *Cryptovirus*. In Fig. 12B are shown RIP profiles of the *Cryptovirus*-specific proteins precipitated by the CSFs of six randomly-selected neurology/neurosurgery patients who had CSF taken for diagnostic screening. The patient whose sample appears in Lane 2 was an adult male
20 who had presented with ataxia, confusion and memory loss (tentatively diagnosed with *ataxic cerebellar syndrome*). The patient whose sample appears in Lane 4 was an infant female who presented with hydrocephalus and intractable seizures and who subsequently died in *status epilepticus*.

25 Example 8. Cryptovirus Is Implicated In The Aetiopathogenesis Of Disease in Patients Diagnosed with Idiopathic Human Neurological, Neurodegenerative, And/Or Neuropsychiatric Diseases.

Cryptovirus is implicated in the aetiopathogenesis of disease in patients diagnosed with idiopathic neurological, neurodegenerative, and/or neuropsychiatric diseases, including anorexia nervosa, multiple sclerosis (MS), epilepsy, subacute sclerosing panencephalitis (SSPE), autism,
30 mental retardation, affective disorder, dysthymia (clinical depression), schizophrenia, obsessive compulsive disorder, manic depression (bipolar disorder), chronic fatigue syndrome (CFS), hydrocephalus, ataxic cerebellar syndrome and atypical viral meningitis. Most patients who had *Cryptovirus*-specific antibodies in their CSF had been given multiple diagnoses. Thus, there is a

correlation between the presence of *Cryptovirus*-specific antibody to the major envelope proteins of the virus (F₀ and HN) in the CSF of neurology or neurosurgery patients and prior diagnosis of a condition with a significant “iterative” or compulsive component.

Although *Cryptovirus* seropositivity did not necessarily correlate to CSF positivity (*i.e.*, the presence of antibody to the *Cryptovirus* F₀ and HN proteins in the CSF) or a diagnosis of any neuropathological condition, CSF positivity strongly correlated with a prior diagnosis of a significant disorder of the central nervous system. These correlations were consistently found for patients with certain diagnoses (*e.g.*, SSPE, MS, CFS, and certain forms of idiopathic epilepsy) and incidentally found for specimens from patients with other diagnoses (*e.g.*, Alzheimer’s Disease).

Similar results were obtained for two CSF specimens analyzed by an immunoblotting technique (data not shown), or for serum and CSF specimens analyzed by an enzyme-linked-immunosorbent-assay (ELISA; see Fig. 14), although these assays were performed on only a proportion of CSF specimens due to the limited volumes available in some samples.

Although some patients presented with some of the above-mentioned symptoms and did not have antibody to the virus in their CSF specimens, in no instances were *Cryptovirus*-specific antibodies found in the CSF of patients that did not present with many of the symptoms and who had not been diagnosed with a significant neuropathological or neuropsychological disorder.

In addition, seropositive individuals (*i.e.*, those who have *Cryptovirus*-specific antibodies in their serum) harbor the virus in a nonproductive, inapparent but inducible state in their PBMNCs.

While the presence of the virus in an individual patient’s PBMNCs did not symmetrically correlate with the development of neuropathological disorder, these findings imply that the virus can gain entry into the CNS via a microvascular incident (*i.e.*, leakage of *Cryptovirus* carrying PBMNCs into the CNS) or by immune system responses to other CNS stimuli (*i.e.*, diapedesis of *Cryptovirus*-carrying lymphocytes into the CNS as part of an inflammatory response to another infection; a Trojan Horse phenomenon). Reference to a lack of symmetrical correlation means that, while all individuals whose PBMNCs were examined and had *Cryptovirus*-specific antibodies in their CSF carried the virus in those cells, not all individuals who were found to be carrying the virus in their PBMNCs were, at that time, suffering from any neurological, neurodegenerative, and/or neuropsychiatric disorder.

The following examples reveal more detail.

(a) *Alzheimer’s Disease*. As shown in Fig. 15, three matched sets of serum and CSF (provided by the National Neurological Research Specimen Bank (NNRSB) in Los Angeles, CA)

were examined by RIP analysis for the ability to precipitate the *Cryptovirus* F₀ and IIN proteins from radiolabeled acutely-infected CV-1_C cells. While all three had *Cryptovirus*-specific antibodies in their serum, only Patient 3 had these antibodies in his or her CSF. This implies that the illness Patient 3 was suffering, diagnosed as Alzheimer's disease, was complicated by concurrent *Cryptovirus* infection of the CNS tissues. Alternatively, Patient 3 could have been misdiagnosed, in which case he or she could actually be suffering from a *Cryptovirus*-related neuropathy.

Even though the sample size is small, it is interesting that all three of the Alzheimer's disease patients had been exposed to *Cryptovirus* and were probably carrying it in their lymphocytes. It appeared, unlikely, however, that *Cryptovirus* plays a role in the development of Alzheimer's disease, because Patients 1 and 2 did not appear to have the virus in their CNS tissues.

(b) *Ataxic Cerebellar Syndrome, Atypical Viral Meningitis, Hydrocephalus, Idiopathic Parasthesia and Status Epilepticus*. A blind screen (i.e., none of the diagnoses or medical histories pertaining to any of the specimens was provided prior to specimen screening) was conducted of 66 CSF specimens from neurology or neurosurgery patients who had CSF specimens taken for diagnostic screening by the Department of Clinical Microbiology at the Royal Brisbane and Royal Children's Hospitals, in Brisbane, Queensland, Australia. Of these CSF specimens, ten were *Cryptovirus*-positive (see Fig. 17). One of the ten *Cryptovirus*-positive CSF specimens was identified as being from an adult male patient who had been diagnosed with ataxic cerebellar syndrome. (see Fig. 12B). Another of the ten *Cryptovirus*-positive CSF specimens was identified as being from an adult female patient who had been diagnosed with atypical viral meningitis (data not shown). A third positive CSF specimen came from a 55 year old male that had presented with ataxia, memory loss, blackouts, seizures, diplopia and headache and had been diagnosed with hydrocephalus, chronic fatigue syndrome and possible epilepsy; a fourth positive CSF specimen was from an adult male who had been diagnosed with idiopathic parasthesia; and a fifth positive CSF specimen was from a female infant who presented with clonic hand movements and intractable seizures and was diagnosed with hydrocephalus and *status epilepticus* (see also c and d, below). Diagnoses and symptoms of the remaining five *Cryptovirus*-positive CSF specimens were unavailable.

(c) *Chronic Fatigue Syndrome (CFS)*. A number of adolescent and adult patients who presented with symptoms of CFS were subsequently found to have high titers of anti-*Cryptovirus* antibodies in their sera, demonstrating that primary infection with the virus can manifest as a chronic febrile tracheo-bronchial illness with associated chronic malaise and lymphadenopathy. This is not

unlike infectious mononucleosis in presentation (i.e., a sore throat and persistent "glandular fever"). There was no evidence of acute encephalitic (or encephalopathic) disease in such patients or in any other patient found to have *Cryptovirus*-specific antibodies in his or her serum or CSF. "Acute" is taken here to mean presenting with rapid onset and symptoms within seven days.

5 Fifty-six serum specimens from patients who had been diagnosed with CFS were provided for *Cryptovirus* screening by regional physicians (Brisbane and Southeast Queensland). Eleven matching CSF specimens were subsequently obtained. RIP analysis revealed that 54/56 (96.4%) of the serum samples and 10/11 (90.9%) of the CSF specimens contained *Cryptovirus*-specific antibodies (Fig. 16).

10 Including the patient who had been codiagnosed with hydrocephalus, epilepsy and CFS (see Fig. 17 and data for Epilepsy, below), a total of 12 CSF specimens from CFS patients were analyzed by RIP analysis and 11/12 (91.7%) had *Cryptovirus*-specific antibodies in them.

 Patients who had been diagnosed with CFS almost always had two, or more, concurrent diagnoses. These included: anorexia nervosa, MS, epilepsy, dysthymia (clinical depression),
15 schizophrenia, and manic depression (bipolar disorder). For example, one adolescent girl who was co-diagnosed with both anorexia nervosa and chronic fatigue syndrome (CFS) had *Cryptovirus*-specific antibodies in her CSF. It is of note that the etiology of virtually all of these disorders is idiopathic.

 While the symptoms presented by CFS patients cover a broad spectrum, the spectrum is, in
20 fact, fairly discrete and representative of the illness. This is perhaps best illustrated by examination of the medical records of five patients, presented below:

 Patient PR was an adult male, 55 years of age, who was suffering primarily from mental confusion, lethargy, memory loss, blurred vision, dysthymia, and *petit mal* seizures. EEG results were abnormal, which indicates epileptiform disease. Patient PR was ambulatory with progressively
25 deteriorating CNS symptoms.

 Patient DF was an adult male, 52 years of age, who was suffering primarily from mental confusion, lethargy, memory loss, dysthymia, and *petit mal* seizures. EEG results were abnormal, showing epileptiform responses in cortical and subcortical functions of the anterior hemispheres. Patient DF was ambulatory with progressively deteriorating CNS symptoms.

30 Patient NB was an adult female, 36 years of age, who was suffering primarily from mental confusion, lethargy and extreme fatigue, memory loss, dysthymia, ataxia, blurred vision, and parathesias, and had a history of glandular fever, recurrent sore throats of prolonged duration,

tremors, and *petit mal* seizures. NB's sister was diagnosed with anorexia and myoclonus. Patient NB was bedridden or partially ambulatory with progressively deteriorating CNS symptoms.

Patient KT was an adult female, 27 years of age, who was suffering primarily from mental confusion, loss of concentration, memory loss, anorexia, lethargy and extreme fatigue, and tremors, and had a history of recurrent febrile lymphadenopathy. Patient KT was stable but bedridden and only partially ambulatory.

Patient SS was an adult female, 23 years of age, who was suffering primarily from loss of concentration, memory loss, and lethargy, and had a history of dysthymia beginning at age 14 and EBV-negative glandular fever. Immediate family members (mother, father, two sisters, and one brother) were all seropositive. In addition, SS's mother had a 9 year history of dysthymia, and *Cryptovirus* antigens were detected in her cultured PBMNC. Two years after sampling, Patient SS was stable and ambulatory.

(d) *Epilepsy and Hydrocephalus*. RIP analysis was used to determine the presence of *Cryptovirus*-specific antibodies in two clinical collections of CSF specimens. The first collection included 66 specimens that were selected at random from those submitted to the Department of Clinical Microbiology at Royal Brisbane Hospital in Brisbane, Queensland by physicians in the Department of Neurology and Neurosurgery (see b above). None of the diagnoses or medical histories pertaining to any of the specimens was provided prior to specimen screening. Fig. 17 illustrates the results of RIP assays conducted with CSF from this collection. The positive CSF precipitate in Lane 2 was subsequently found to have come from a 55-year old adult male (RW) who presented with ataxia, memory loss, blackouts, seizures, diplopia, and headaches. He was determined to have a hydrocephalic condition and underwent surgery to insert a ventricular shunt to alleviate the condition. He had been diagnosed with hydrocephalus, epilepsy and Chronic Fatigue Syndrome (CFS).

Ten of the 66 CSF specimens in Collection 1 were found to contain *Cryptovirus*-specific antibody (15%). Diagnoses were obtained for five of these patients and included: (1) hydrocephalus and intractable seizures in an infant female who subsequently died (see Fig. 12B), (2) ataxic cerebellar syndrome in an adult male (see Fig. 12B), (3) atypical viral meningitis in a female child, (4) parathesia in an adult male, and (5) hydrocephalus, epilepsy and CFS in the patient described in connection with Fig. 17.

Diagnoses were obtained for only two of the 56 *Cryptovirus*-negative CSF specimens: one patient (WK, a male) was diagnosed with acute viral meningitis and one (SG, a female) was diagnosed with idiopathic intracranial hypertension.

The second collection (Collection 2) included 20 CSF specimens from children (<12 years old) that were collected by neurologists at Camperdown Children's Hospital in Sydney, New South Wales, Australia. Again, none of the diagnoses or medical histories pertaining to any of the specimens was provided prior to specimen screening. However, in this collection a request had been made to include an undisclosed number of CSF specimens from children who had either presented with epileptiform illness or had been diagnosed with some form of idiopathic epilepsy.

Fig. 18 illustrates the results of RIP assays conducted with CSF from Collection 2. The CSF precipitate analyzed in Lane 1 was from a newborn infant who developed intractable seizures and died in *status epilepticus* (Patient CT, below) and the precipitate analyzed in Lane 2 was from a child who had been given a diagnosis of Lennox-Gastaut/generalized epilepsy (Patient LB, below), respectively. The background noise in this autoradiogram was high as a result of the long-term exposure (30 days) required to see the bands.

Six of the 20 CSF specimens provided in the (biased) Collection 2 were found to have *Cryptovirus*-specific antibodies, and it was subsequently learned that this screening had identified 6 of the 7 specimens from patients who had been diagnosed with epilepsy or other forms of epileptiform illness and had been included in the collection. The six *Cryptovirus*-positive CSF specimens came from the following patients: (1) CT, a neonate with intractable fits and seizures, who died in *status epilepticus*, (2) LB, who was diagnosed with Lennox-Gastaut epilepsy and generalized epilepsy, (3) BM, who was diagnosed with severe retardation and epilepsy, (4) FZ, a two-month old child with intractable seizures who died in *status epilepticus*, (5) CN, who had hydrocephalus, cerebral palsy, and epileptiform seizures, and (6) LD, who had primary infantile spasms. Hydrocephalus was codiagnosed in 3 of 8 patients diagnosed with epilepsy or other epileptiform illness.

Although one of the 14 *Cryptovirus*-negative CSF specimens was obtained from a patient who had been diagnosed with epilepsy, diagnoses were not provided for the remaining specimens. They were simply characterized as pediatric neurology or neurosurgery specimens from asymptomatic patients (*i.e.*, patients who had not presented with epileptiform symptoms or been diagnosed with epileptiform illness).

(e) *Multiple Sclerosis (MS)*. Clinical specimens from patients with MS comprise one of the largest groups of materials screened (38 serum samples and 30 CSF samples including 30 matched sets of each). Eight of the serum samples came from MS patients in Brisbane, Queensland who had debilitating disease and were living in a nursing home run by the National Multiple Sclerosis Society of Australia. No CSF specimens were acquired from these patients. The 30 matched sets of serum and CSF were provided by the National Neurological Research Specimen Bank (NNRSB) in Los Angeles.

Fig. 19 illustrates the results of RIP assays conducted with serum samples of 5/30 MS patients provided by the NNRSB. The results obtained from an additional 25 serum specimens provided by the NNRSB are shown in Fig. 20, and the RIP results from 16/30 of the CSF specimens from MS patients provided by the NNRSB are shown in Fig. 21. RIPs performed using the remaining 8 specimens resulted in similar profiles (data not shown). As shown in Figs. 19-21, the results of these analyses demonstrated that all patients had high levels of *Cryptovirus*-specific antibodies in their serum (100%) and 29/30 had *Cryptovirus*-specific antibodies in their CSF (96.7%).

(f) *Subacute Sclerosing Panencephalitis (SSPE)*. The anomalies that have been observed which are inconsistent with measles virus alone being the sole cause of SSPE (see Discussion of Related Art) can be explained by the evidence that the aetiopathogenesis of SSPE involves dual infection of the CNS by measles and *Cryptovirus* (which was isolated from SSPE patients).

Sera from SSPE patients were found to precipitate the major nucleocapsid protein of the virus (NP, 63 kD) from nonproductively-infected AV₃/SSPE cells (see Fig. 22). Fig. 22 is a photograph of an autoradiogram obtained following creation of RIP profiles of the *Cryptovirus* NP protein (p63) precipitated from [³⁵S]-methionine-labeled AV₃/SSPE cells by the sera of six Australian SSPE patients (Lanes 1-6) and six control sera (Lanes 7-12; sera from pediatric patients without antibodies to the *Cryptovirus* major envelope proteins (F₀, HN).

Fig. 23 is a photograph of an autoradiogram of RIP profiles of measles virus-specific and *Cryptovirus*-specific proteins precipitated from [³⁵S]-methionine-labeled measles virus-infected CV-1_C cells (Lane MV), *Cryptovirus*-infected CV-1_C cells (Lane CV) or a mixture of both (Lane B) by CSF sampled from an 11-year old male diagnosed with SSPE. Lane V = gradient-purified *Cryptovirus* virions from [³⁵S]-methionine-labeled *Cryptovirus*-infected CV-1_C cells. Fig. 23 shows that CSF from this SSPE patient precipitated both *Cryptovirus* and measles virus proteins. The results of this assay demonstrate that SSPE CSF contains both measles virus-specific and *Cryptovirus*-specific antibodies (i.e. antibody to the HN protein of measles virus (Lane MV) and the

HN and F₀ proteins of *Cryptovirus* virus (Lane CV) and that both are present in nearly equal amounts. This was unique, since none of the other CSFs samples that precipitated *Cryptovirus* proteins (e.g., from MS, CFS, or epilepsy patients) also precipitated the measles virus HN protein.

The RIP profile of *Cryptovirus*-specific proteins precipitated by this CSF specimen is typical of those produced by the “*Cryptovirus*-positive” CSFs of MS patients, CFS patients and idiopathic epilepsy patients tested to date (compare Figs. 12A and 17). While there was considerable variation in the strength of the antibody response to the F₀ and HN proteins, there appeared to be little variation in the presence of antibody to one protein or the other. There was, however, a variable response to the F₁ and F₂ proteins (*i.e.*, in many patients, such responses appeared to be absent). This paradox may relate to the proteolytic cleavage of the protein *in situ* and corresponding immune responses (*i.e.*, the F₀ protein is efficiently cleaved by some patients, generating the F₁ and F₂ fragments and ultimately exposing their immune system to them); in other patients the protein may be cleaved less efficiently (or not at all) and, therefore, these patients do not generate as much of an antibody response to the fragments).

Example 9. Correlations Between Affected Human Patients and Experimentally-Infected Animals

Some of the examples herein highlight the epileptiform symptomology presented by many of the patients who have *Cryptovirus*-specific antibodies in their cerebrospinal fluid (CSF) or by mice experimentally-infected with the virus. This association is strong, but not all patients with *Cryptovirus*-specific antibodies in their CSF present with overt seizures or convulsions. There is instead a spectrum of responses, from little or no seizure activity, through mild activity (*petit mal* or “absence” seizures), to recurrent and intractable *grand mal* seizures (the occurrence of which is often misunderstood by the lay public to be the defining symptom of all forms of epilepsy; *see Epilepsy: A Comprehensive Textbook*, Engel, Jr. J. and Pedley, T.A., Eds., Lippincott-Raven, 1997).

With regard to the development of symptoms and manifestations of human *Cryptovirus*-infections, it is essential to be cognizant of the spatial (“where”), temporal (“when”), and quantum (“what else”) factors involved: (1) which cells, tissues, neural tracts and CNS structures become infected by the virus, (2) the developmental state of those systems at the time of infection, and (3) the role of environmental and host factors in development and progress of the infection, respectively.

For example, the data presented here establish a strong correlation between the development of epileptiform symptomology and *early* CNS infection with the virus (*i.e.*, in infancy, early childhood or adolescence and in experimentally-infected neonatal mice), this correlation is less strong

in adults (and adult mice who do not develop epileptiform symptoms) and the spectrum of CNS manifestations observed is much wider.

Generally, the characteristic most frequently and consistently presented by humans or animals that have been experimentally infected with the virus is the development of “iterative” or “compulsive” neuropathies and behaviors. This is most likely due to the selective loss of (or immunopathological damage done to) neurons (*e.g.* interneurons) or neuron tracts in different parts of the central nervous system (CNS) at different times or at different stages of CNS development.

When the medical records of patients who had *Cryptovirus*-specific antibodies in their CSF were examined, all of the patients had been diagnosed with one or more serious neurological disorders. These included, but were not limited to:

(1) subacute sclerosing panencephalitis (SSPE): 4 of 4 CSFs tested (100%), all adolescent patients);

(2) idiopathic / cryptogenic epilepsy: 6 of 7 CSFs tested (85.7%), from infants and children presenting with seizures and diagnosed with idiopathic or cryptogenic forms of epilepsy;

(3) multiple sclerosis (MS): 29 of 30 CSFs tested (96.7%) all adult specimens; and

(4) chronic fatigue syndrome (CFS) / clinical depression: 11 of 12 CSFs tested (91.7%), all adult specimens.

These results demonstrate a clear correlation between *Cryptovirus*-specific antibodies in the CSF and a narrow spectrum of CNS diseases. Although the diseases listed above have been defined as representatives of discrete pathognomonic entities, there is in reality substantial overlap between the symptoms presented by these patients and their diagnoses. For example, virtually every patient eventually diagnosed with SSPE is initially diagnosed with epilepsy. Similarly, early stage MS is extremely similar in presentation to CFS, and clinical depression is a common characteristic of both. Not surprisingly, another name for CFS is “atypical multiple sclerosis” (in Bell, *The Disease of a Thousand Names*, Pollard Publications, Lyndonville, NY [1991]).

There is a strong correlation between (1) the age of those patients who had severe epileptiform illness (SSPE and epilepsy, the majority of whom are infants, children, or adolescents) and (2) the age of those patients who had more diffuse or subtle neurological dysfunction (*e.g.*, MS and CFS patients who were all adults). Furthermore, SSPE, certain forms of idiopathic and cryptogenic epilepsy, and MS have many neuropathological characteristics in common. These include areas of discrete, focal or disseminated sclerosis (scar formation in CNS tissue), dysplastic lesions (either as the result of immunopathological processes or neuron tract loss), and perivascular cuffing of immune cells (evidence of inflammatory processes in the vicinity of lesions). Thus, each

of these diseases could represent a different pathological “complex” of spatial, temporal, and quantum factors that have *Cryptovirus* infection of CNS tissues as a shared characteristic. With respect to SSPE, previous data have established (and the data here confirm) that measles virus is also involved in this illness. SSPE is caused by widespread CNS infection by both viruses (resulting in inflammatory and disseminated sclerosis across the white matter of the brain) while certain forms of idiopathic epilepsy represent early infection of the CNS by *Cryptovirus* alone (resulting in the loss of susceptible interneurons and neuron tracts and the development of discrete dysplastic lesions). MS, occurring almost exclusively in adults, represents the pathological outcome of late and focal *Cryptovirus* infection of the CNS – due to the restriction of *Cryptovirus* replication in fully differentiated CNS tissues and the effective partitioning of brain by the mature glial architecture.

In summary, intracranial inoculation of mice with the virus, or with cells nonproductively-infected with the virus, results in the subacute development of neuropathological diseases in a significant proportion of the animals. These diseases are closely akin to the spectrum of human neuropathies seen in patients with *Cryptovirus*-specific antibodies in their cerebrospinal fluids.

Further, although patients with *Cryptovirus*-positive CSF had been diagnosed with a spectrum of illnesses, there was a clear partitioning of patients into two groups (1) infants, children and adolescents with illnesses dominated by subacute epileptiform physical symptoms which were often life-threatening and (2) young adults and adults with slowly-developing chronic illnesses which presented with less pronounced physical symptoms but significant neuropsychological components which were usually not life-threatening.

These findings support the conclusion that *Cryptovirus* is responsible for a neurological spectrum disorder whose ultimate manifestation depends on (1) the age of the individual when they contract the primary infection, (2) the mechanism by which the virus gains entry into the CNS tissue, (3) the extent of the infection at that time, (4) the stage of development of the CNS when it becomes infected, (5) the part of the CNS which becomes infected, (6) genetic factors (*e.g.*, immune system defects, neurological malformations, the presence for absence of virus receptors on CNS tissues, *etc.*) and (7) other environmental factors (*e.g.*, the occurrence of head trauma, neurosurgery of any kind, prior or concurrent infection of CNS tissue by other agents, exposure to drugs or toxic chemicals, *etc.*).

A number of embodiments of the invention have been described. Nevertheless, it will be understood that various modifications can be made without departing from the spirit and scope of the invention. Accordingly, other embodiments are within the scope of the following claims.

The claims defining the invention are as follows:

1. An isolated nucleic acid, comprising:
 - (A) contiguous nucleotide positions 1-15246 of (SEQ ID NO: 1);
 - (B) a nucleotide sequence complementary to (A); or
 - 5 (C) *Cryptovirus*-specific fragment of (A) or (B), comprising a nucleic acid segment selected from the group consisting of:
 - (i) contiguous nucleotide positions 152-1678 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (ii) contiguous nucleotide positions 1850-2515 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - 10 (iii) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (iv) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1) combined with a further insertion of two guanine residues between nucleotide position 2339 of (SEQ ID NO:1) and nucleotide position 2340 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (v) contiguous nucleotide positions 3141-4271 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (vi) contiguous nucleotide positions 4530-6182 of (SEQ ID NO:1), a complementary sequence, or a degenerate sequence;
 - 20 (vii) contiguous nucleotide positions 4587-6182 of (SEQ ID NO:1), a complementary sequence, or a degenerate sequence;
 - (viii) contiguous nucleotide positions 4587-4835 of (SEQ ID NO:1), a complementary sequence, or a degenerate sequence;
 - (ix) contiguous nucleotide positions 4836-6182 of (SEQ ID NO:1), a complementary sequence, or a degenerate sequence;
 - 25 (x) contiguous nucleotide positions 4272-6515 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - (xi) contiguous nucleotide positions 6303-6434 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence;
 - 30 (xii) contiguous nucleotide positions 6584-8278 of (SEQ ID. NO:1), a complementary sequence, or a degenerate coding sequence; and
 - (xiii) contiguous nucleotide positions 8414-15178 of (SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

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(xiv) contiguous nucleotide positions 1684-1701 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xv) contiguous nucleotide positions 1700-1717 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

5 (xvi) contiguous nucleotide positions 4283-4300 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xvii) contiguous nucleotide positions 4299-4316 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

10 (xviii) contiguous nucleotide positions 4285-4302 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xix) contiguous nucleotide positions 4300-4317 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xx) contiguous nucleotide positions 4518-4535 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

15 (xxi) contiguous nucleotide positions 4533-4550 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxii) contiguous nucleotide positions 6191-6208 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

20 (xxiii) contiguous nucleotide positions 6116-6133 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxiv) contiguous nucleotide positions 6192-6209 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxv) contiguous nucleotide positions 7501-7518 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

25 (xxvi) contiguous nucleotide positions 7517-7534 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence; and

(xxvii) contiguous nucleotide positions 4292-4549 (of SEQ ID NO:1), a complementary sequence, or a degenerate coding sequence.

2. The nucleic acid of Claim 1, wherein the nucleic acid is RNA.

30 3. The nucleic acid of Claim 1, wherein the nucleic acid is cDNA.

4. A nucleic acid construct, comprising the nucleic acid of any one of claims 1 to 3.

5. An expression vector, comprising the nucleic acid construct of Claim 4

6. A cloning vector, comprising the nucleic acid construct of Claim 4.

7. A host cell, comprising the expression vector of Claim 5 or the cloning vector of Claim 6.

8. The host cell of Claim 7, wherein the cell is a mammalian cell.

9. An isolated *Cryptovirus* protein encoded by a nucleic acid segment
5 comprising:

(A) contiguous nucleotide positions 152-1678 of (SEQ ID NO:1) or a degenerate sequence;

(B) contiguous nucleotide positions 1850-2515 of (SEQ ID NO:1) or a degenerate sequence;

10 (C) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1) combined with a further insertion of two guanine residues between nucleotide position 2339 of (SEQ ID NO:1) and nucleotide position 2340 of (SEQ ID NO:1), or a degenerate sequence;

(D) contiguous nucleotide positions 3141-4271 of (SEQ ID NO:1) or a
15 degenerate sequence;

(E) contiguous nucleotide positions 4530-6182 of (SEQ ID NO:1) or a degenerate sequence;

(F) contiguous nucleotide positions 4587-6182 of (SEQ ID NO:1) or a degenerate sequence;

20 (G) contiguous nucleotide positions 4587-4835 of (SEQ ID NO:1) or a degenerate sequence;

(H) contiguous nucleotide positions 4836-6182 of (SEQ ID NO:1) or a degenerate sequence;

(I) contiguous nucleotide positions 6303-6434 of (SEQ ID NO:1) or a
25 degenerate sequence;

(J) contiguous nucleotide positions 6584-8278 of (SEQ ID NO:1) or a degenerate sequence; or

(K) contiguous nucleotide positions 8414-15178 of (SEQ ID NO:1) or a degenerate sequence.

30 10. The protein of Claim 9, wherein the protein is a *Cryptovirus* envelope protein encoded by a nucleic acid segment comprising (E), (F), (G), (H), (I), or (J).

11. A chimeric protein, comprising a *Cryptovirus* protein encoded by a nucleic acid segment comprising:

(A) contiguous nucleotide positions 152-1678 of (SEQ ID NO:1) or a
35 degenerate sequence;

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(B) contiguous nucleotide positions 1850-2515 of (SEQ ID NO:1) or a degenerate sequence;

(C) contiguous nucleotide positions 1850-3023 of (SEQ ID NO:1) combined with a further insertion of two guanine residues into the nucleotide sequence between nucleotide position 2339 of (SEQ ID NO:1) and nucleotide position 2340 of (SEQ ID NO:1), or a degenerate sequence;

(D) contiguous nucleotide positions 3141-4271 of (SEQ ID NO:1) or a degenerate sequence;

(E) contiguous nucleotide positions 4530-6182 of (SEQ ID NO:1) or a degenerate sequence;

(F) contiguous nucleotide positions 4587-6182 of (SEQ ID NO:1) or a degenerate sequence;

(G) contiguous nucleotide positions 4587-4835 of (SEQ ID NO:1) or a degenerate sequence;

(H) contiguous nucleotide positions 4836-6182 of (SEQ ID NO:1) or a degenerate sequence;

(I) contiguous nucleotide positions 6303-6434 of (SEQ ID NO:1) or a degenerate sequence;

(J) contiguous nucleotide positions 6584-8278 of (SEQ ID NO:1) or a degenerate sequence; or

(K) contiguous nucleotide positions 8414-15178 of (SEQ ID NO:1) or a degenerate sequence.

12. The chimeric protein of claim 11 wherein the *Cryptovirus* protein is a *Cryptovirus* envelope protein encoded by a nucleic acid segment comprising (E), (F), (G), (H), (I) or (J).

13. Use of the protein of any one of claims 9 to 12 in producing a *Cryptovirus*-specific antibody.

14. An isolated antibody that specifically binds the protein of claim 9 or 10.

15. The antibody of claim 14, that specifically binds a *Cryptovirus* envelope protein encoded by:

(A) contiguous nucleotide positions 4530-6182 of (SEQ ID NO:1) or a degenerate sequence;

(B) contiguous nucleotide positions 4587-6182 of (SEQ ID NO:1) or a degenerate sequence;

(C) contiguous nucleotide positions 4587-4835 of (SEQ ID NO:1) or a degenerate sequence;

(D) contiguous nucleotide positions 4836-6182 of (SEQ ID NO:1) or a degenerate sequence;

5 (E) contiguous nucleotide positions 6303-6434 of (SEQ ID NO:1) or a degenerate sequence;

(F) contiguous nucleotide positions 6584-8278 of (SEQ ID NO:1) or a degenerate sequence.

16. The antibody of claim 14 or 15, wherein the antibody is a polyclonal.
- 10 17. The antibody of claim 14 or 15, wherein the antibody is monoclonal.
18. The antibody of claim 14 or 15, wherein the antibody is chimeric.
19. Use of the antibody of any one of claims 14 to 18 in manufacturing a medicament for the treatment of *Cryptovirus* infections.
20. An isolated viral particle comprising the nucleic acid of claim 1.
- 15 21. A composition of matter, comprising the nucleic acid of any one of claims 1 to 3, the protein of any one of claims 9 to 12, the antibody of any one of claims 14 to 18, or the virion of claim 20; and a carrier.
22. An isolated viral particle, comprising the protein of any one of claims 9 to 12.
- 20 23. An isolated *Cryptovirus* particle, comprising a genome having a nucleotide sequence entirely complementary to (SEQ ID NO:1).
24. Use of the nucleic acid of any one of claims 1 to 3, the nucleic acid construct of claim 4, the protein of any one of claims 9 to 12, the viral particle of claim 20 or 22, or *Cryptovirus* particle of claim 23, in manufacturing a vaccine.
- 25 25. The use according to claim 24, wherein the viral particle is an attenuated virion.
26. The use according to claim 24, wherein the viral particle is a killed virion.
27. An isolated *Cryptovirus* particle, wherein the *Cryptovirus* is Strain
30 BBR.
28. A probe or primer, comprising the nucleic acid of any one of claims 1 to 3.
29. A method of detecting the presence or absence of a *Cryptovirus* protein in a sample of a biological material, comprising:

contacting the sample of the biological material with the antibody of any one of claims 14 to 18; and

detecting specific binding of the antibody to a constituent of the sample, wherein the presence of specific binding indicates the presence of the *Cryptovirus* protein in the sample.

30. A method of detecting the presence or absence of a *Cryptovirus*-specific RNA in a sample of a biological material, comprising:

obtaining a sample of a biological material comprising RNA;

contacting the sample with the probe of claim 28 under at least moderately stringent hybridization conditions, wherein the formation of detectable hybridization product indicates the presence of the *Cryptovirus*-specific RNA in the sample.

31. A method of detecting the presence or absence of a *Cryptovirus*-specific RNA in a sample of a biological material, comprising:

obtaining a sample of a biological material comprising RNA;

amplifying *Cryptovirus*-specific RNA in the sample using at least one primer of claim 28 in an amplification reaction mixture;

then detecting the presence or absence of *Cryptovirus*-specific nucleic acid amplification products in the amplification reaction mixture, wherein the presence of the amplification products in the reaction mixture indicates the presence of the *Cryptovirus* RNA in the sample.

32. The method of any one of claims 29 to 31, wherein the biological material is a cellular material.

33. The method of any one of claims 29 to 31, wherein the biological material is blood or serum.

34. The method of any one of claims 29 to 31, wherein the biological material is cerebrospinal fluid.

35. The method of any one of claims 29 to 31, wherein the biological material is lymphoid tissue.

36. The method of any one of claims 29 to 31, wherein the biological material is nervous tissue.

37. The method of claim 36, wherein the nervous tissue is brain tissue.

38. A method of detecting the presence or absence of a *Cryptovirus*-specific antibody in a sample of a biological material, comprising:

contacting the sample with the protein of claim 9 or 10;

allowing the formation of a specific protein-antibody complex;

detecting the presence of the specific protein-antibody complex, wherein the presence of a specific protein-antibody complex indicates the presence of the *Cryptovirus*-specific antibody in the sample.

39. A method of detecting the presence of a *Cryptovirus*-specific antibody
5 in a sample of a biological material, comprising:

contacting the sample with the protein of claim 11 or 12;

allowing the formation of a specific protein-antibody complex;

detecting the presence of the specific protein-antibody complex, wherein the
presence of a specific protein-antibody complex indicates the presence of *Cryptovirus*-
10 specific antibody in the sample.

40. An assay method for detecting the presence or absence of an antibody
that selectively binds *Cryptovirus* in a sample of an antibody-containing biological
material originating from a human, comprising:

contacting the sample, the sample originating from an individual suspected of
15 having a *Cryptovirus* infection, with the envelope protein of claim 10, such that, if
antibody selectively binding *Cryptovirus* is present, an antibody-bound envelope protein
complex forms;

contacting any antibody-bound envelope protein complexes thus formed with
anti-human antibody-binding antibody, and allowing the formation of complexes of the
20 antibody, with the antibody-bound envelope protein complexes; and

detecting the presence or absence of any antibody-bound envelope protein
complexes thus formed, the presence of such complexes indicating the presence in the
sample of antibody selectively binding *Cryptovirus*.

41. An assay method for detecting the presence or absence of antibody that
25 selectively binds *Cryptovirus* antigen in a sample of an antibody-containing biological
material originating from a human, the method comprising:

contacting the sample, the sample originating from an individual suspected of
having a *Cryptovirus* infection, with the viral particle of claim 22, such that, if antibody
selectively binding *Cryptovirus* antigen is present, an antibody-bound virus complex
30 forms;

contacting any antibody-bound virus complexes thus formed with anti-human
antibody-binding antibody, and allowing the formation of complexes of the anti-human
antibody-binding antibody with the antibody-bound virus complexes; and

detecting the presence or absence of any complexes formed, the presence of such complexes indicating the presence in the sample of antibody selectively binding *Cryptovirus* antigen.

5 42. A method of detecting *Cryptovirus* infection in a mammal, comprising:
obtaining a sample of a biological material from the mammal; and
performing the method of any one of claims 29 to 41, using the sample, whereby
detecting the presence of the *Cryptovirus* protein, *Cryptovirus*-specific RNA, and/or
Cryptovirus-specific antibody in the sample indicates a *Cryptovirus* infection in the
mammal.

10 43. The method according to any one of claims 38 to 42, wherein the
biological material is cellular material.

44. The method according to any one of claims 38 to 42, wherein the
biological material is blood or serum.

15 45. The method according to any one of claims 38 to 42, wherein the
biological material is cerebrospinal fluid.

46. The method according to any one of claims 38 to 42, wherein the
biological material is lymphoid tissue.

47. The method according to any one of claims 38 to 42, wherein the
biological material is nervous tissue.

20 48. The method according to claim 47, wherein the nervous tissue is brain
tissue.

49. The method of claim 42, wherein the mammal is a human.

50. The method of claim 49, wherein the human has a neurological,
neurodegenerative, and/or neuropsychiatric disease.

25 51. The method of claim 49, wherein the human has a primary
tracheobronchial and/or lymphadenopathy-associated illness.

52. A method of isolating a *Cryptovirus* virion, comprising:

30 (a) culturing a plurality of peripheral blood mononuclear cells that have
been obtained from a human having a *Cryptovirus* infection, in an artificial aqueous
medium comprising an agent that increases cellular guanylyl cyclase activity.

(b) co-culturing the plurality of peripheral blood mononuclear cells with a
plurality of mammalian amnion cells in fresh artificial aqueous medium comprising an
agent that increases cellular guanylyl cyclase activity.

35 (c) passaging the peripheral blood mononuclear cells with the mammalian
amnion cells in co-culture.

(d) co-cultivating a plurality of mammalian epithelial cells together with the peripheral blood mononuclear cells and the mammalian amnion cells in fresh artificial aqueous medium comprising an agent that increases cellular guanylyl cyclase activity; and

(e) separating a supernatant of the aqueous medium from the cells, to obtain
5 a *Cryptovirus* virion in the supernatant.

53. A method of propagating a *Cryptovirus*, comprising:

(a) exposing a plurality of mammalian epithelial cells to a plurality of cell free *Cryptovirus* virions, said *Cryptovirus* virions having been isolated by the method of claim 52; and

10 (b) further cultivating the mammalian epithelial cells, thus virion-exposed, in an artificial aqueous medium comprising an agent that increases the activity of cellular guanylyl cyclase.

54. A method of producing a mammalian cell line non-productively infected with *Cryptovirus*, comprising:

15 (a) co-culturing peripheral blood mononuclear cells that have been obtained from a human having a *Cryptovirus* infection, with mammalian amnion cells, in an artificial aqueous medium comprising an agent that increases cellular guanylyl cyclase activity, such that the mammalian amnion cells become non-productively infected by *Cryptovirus*; and

20 (b) passaging the non-productively infected mammalian amnion cells with the peripheral blood mononuclear cells, whereby the co-culture becomes a monoculture of the non-productively infected mammalian amnion cells.

55. The method according to any one of claims 52 to 54, wherein the mammalian amnion cells are human amnion cells.

25 56. The method of claim 55, wherein the human amnion cells are AV₃ cells.

57. The method of any one of claims 52 to 54, wherein the mammalian epithelial cells are simian epithelial cells selected from the group consisting of Vero or CV-1 cells.

58. The method of claim 57, wherein the CV-1 cells are subline CV-1 cells.

30 59. The method of any one of claims 52 to 58, wherein the agent that increases cellular guanylyl cyclase activity is cyclic GMP, insulin, zinc dication, or a combination of any of these.

60. The method of claim 59 wherein the cyclic GMP is in a concentration of about 0.05 to about 5mM in the artificial aqueous medium.

61. The method of any one of claims 52 to 58, wherein the agent that increases cellular guanylyl cyclase activity is nitric oxide or a nitric oxide donor selected from the group consisting of organic nitrate compounds, iron nitrosyl compounds, S-nitrosothiol compounds, sydnonimine compounds, and nonoate compounds.

5 62. The method of any one of claims 52 to 61, wherein the aqueous medium further comprises glutamine.

63. A method of producing a mammalian epithelial cell line acutely infected with *Cryptovirus*, comprising the method of claim 53.

64. A mammalian epithelial cell acutely infected with *Cryptovirus*, said cell
10 being produced by the method of claim 53 or 63.

65. A cell non-productively infected with *Cryptovirus*, wherein said cell is produced in accordance with the method of claim 54.

66. An *in vitro* method of screening a potential antiviral therapeutic agent, comprising:

- 15 (a) culturing the cell of claim 64;
(b) exposing the cells to the potential antiviral therapeutic agent; and
(c) measuring the effect of the agent on *Cryptovirus* replication and/or *Cryptovirus* virion assembly, wherein inhibition of *Cryptovirus* replication and/or *Cryptovirus* virion assembly relative to a control indicates antiviral activity of the
20 potential therapeutic agent.

67. An *in vitro* method of screening a potential antiviral therapeutic agent, comprising:

- (a) culturing the cell of claim 65;
(b) exposing the cells to the potential antiviral therapeutic agent; and
25 (c) measuring the effect of the agent *Cryptovirus* replication, *Cryptovirus* genome replication, and/or *Cryptovirus*-specific transcription, wherein inhibition of *Cryptovirus* replication, *Cryptovirus* genome replication, and/or *Cryptovirus*-specific transcription, relative to a control, indicates antiviral activity of the potential therapeutic agent.

30 68. An animal model for the study of human diseases, comprising a non-human mammal, said non-human mammal having been artificially inoculated with an infectious cell-free *Cryptovirus* having a genome comprising a single stranded RNA complementary to (SEQ ID NO:1), or having been inoculated with a cell non-productively-infected with the *Cryptovirus*, whereby the non-human mammal exhibits at

least one symptom characteristic of a human disease after being thus inoculated, said symptom not being previously exhibited by the non-human mammal.

69. The animal model of claim 68, wherein the non-human mammal is a rodent or lagomorph.

70. The animal model of claim 68, wherein the non-human mammal is a non-human primate.

71. The animal model of any one of claims 68 to 70, wherein the human disease is a neurological, neurodegenerative, and/or neuropsychiatric disease.

72. An *in vivo* method of screening a potential therapeutic agent, comprising:

(a) administering the potential therapeutic agent to be screened to the animal model of any one of claims 68 to 71, wherein the non-human mammal exhibits, before administration of the potential therapeutic agent, at least one symptom characteristic of a human disease; and

(b) detecting the presence or absence of a beneficial antiviral effect of the potential therapeutic agent, wherein the presence of a beneficial antiviral effect indicates activity of the potential therapeutic agent.

73. An *in vivo* method of screening a potential prophylactic agent, comprising:

(a) administering the potential prophylactic agent to be screened, to a non-human mammal not previously having a symptom of a human disease;

(b) inoculating the non-human mammal with an infectious cell-free *Cryptovirus* having a genome comprising a single stranded RNA complementary to (SEQ ID NO:1), or with a mammalian cell non-productively-infected with the *Cryptovirus*; and

(c) detecting the subsequent presence or absence in the non-human mammal of a beneficial antiviral effect, whereby the presence of a beneficial antiviral effect in the inoculated non-human mammal indicates activity of the potential prophylactic agent.

74. The method of claim 73, wherein the potential prophylactic agent is an immunoprophylactic agent.

75. The method of claim 72 or 73, wherein the non-human mammal is a rodent or a lagomorph.

76. The method of claim 72 or 73, wherein the non-human mammal is a non-human primate.

77. The method of any one of claims 72 to 76, wherein the human disease is a neurological, neurodegenerative, and/or neuropsychiatric disease.

5 78. An anti-*Cryptovirus* antibody detecting kit, comprising:
the *Cryptovirus* particle of claim 22 or 23; and
a labelled anti-human antibody-binding antibody.

79. The detecting kit of claim 78, wherein the kit further comprises a solid matrix for supporting said *Cryptovirus* particle.

10 80. An anti-*Cryptovirus* antibody detecting kit, comprising:
the protein of any one of claims 9 to 12; and
a labelled anti-human antibody-binding antibody.

81. The detecting kit of claim 80, wherein the kit further comprises a solid matrix for supporting said protein.

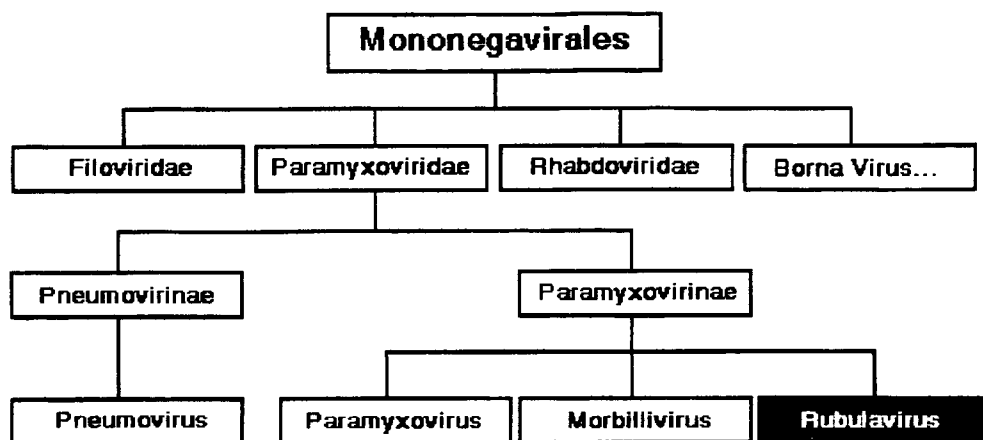


Fig. 1

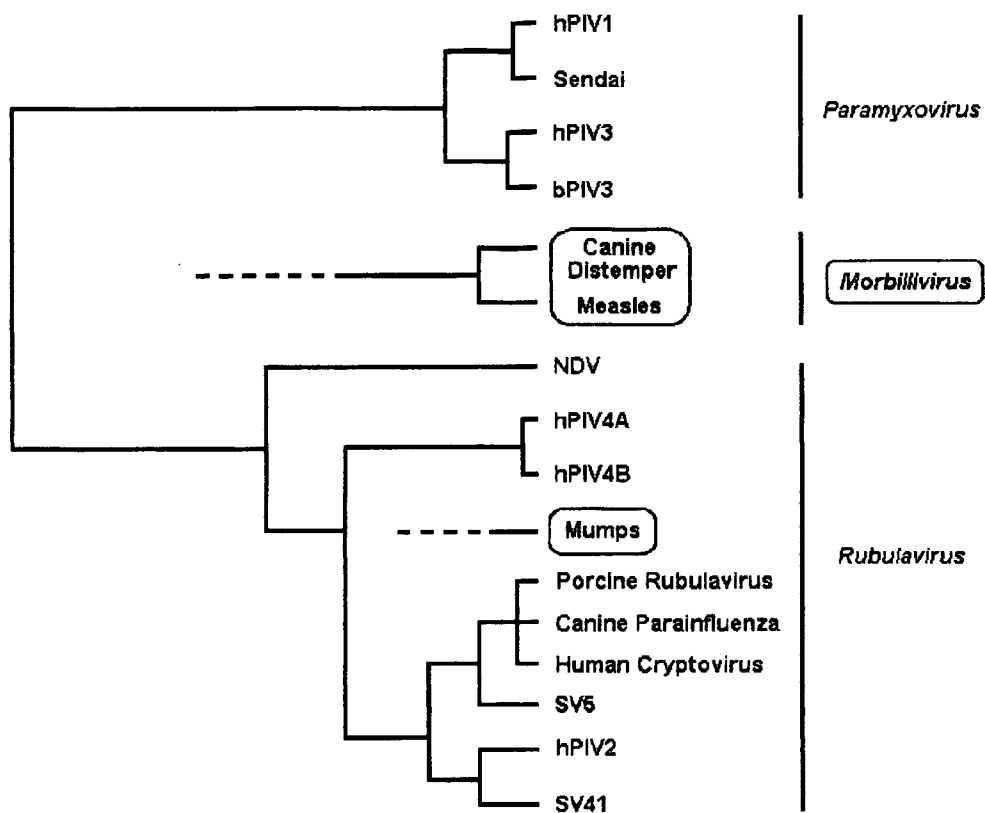
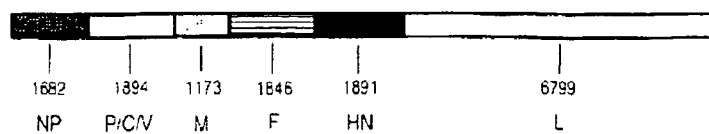
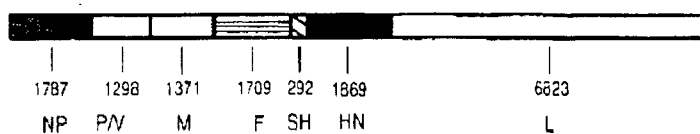
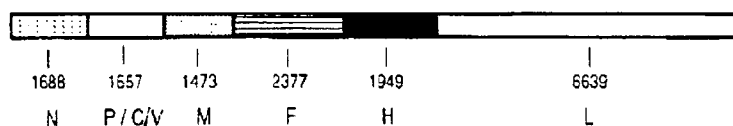
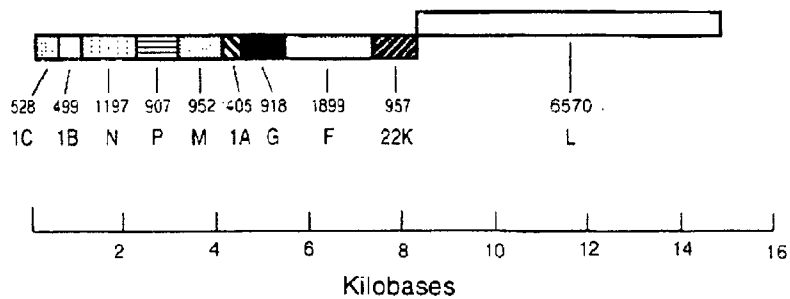


Fig. 2

Parainfluenza virus - Sendai virus**Rubula virus - SV5****Morbillivirus - Measles****Pneumovirus - Respiratory Syncytial Virus***Fig. 3*

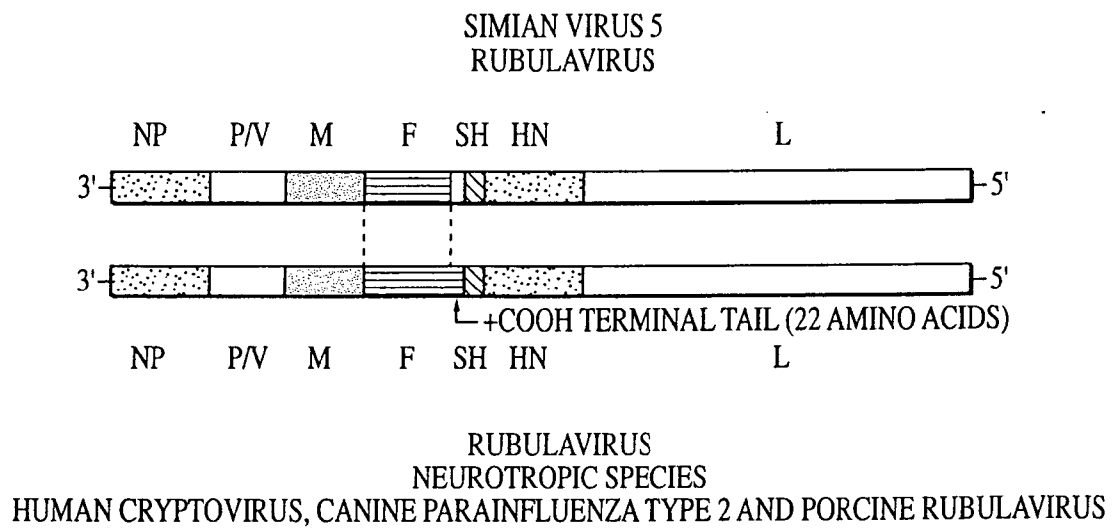


FIG. 4

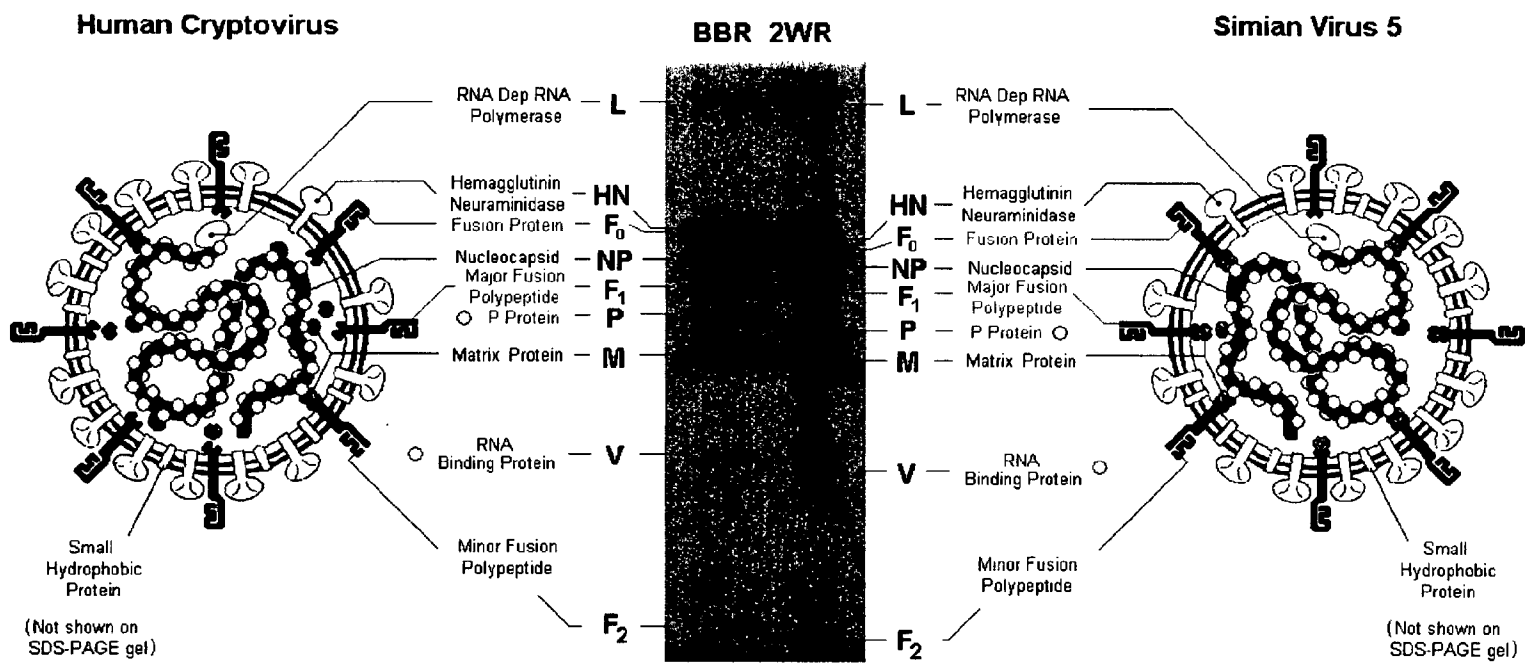


Fig. 5

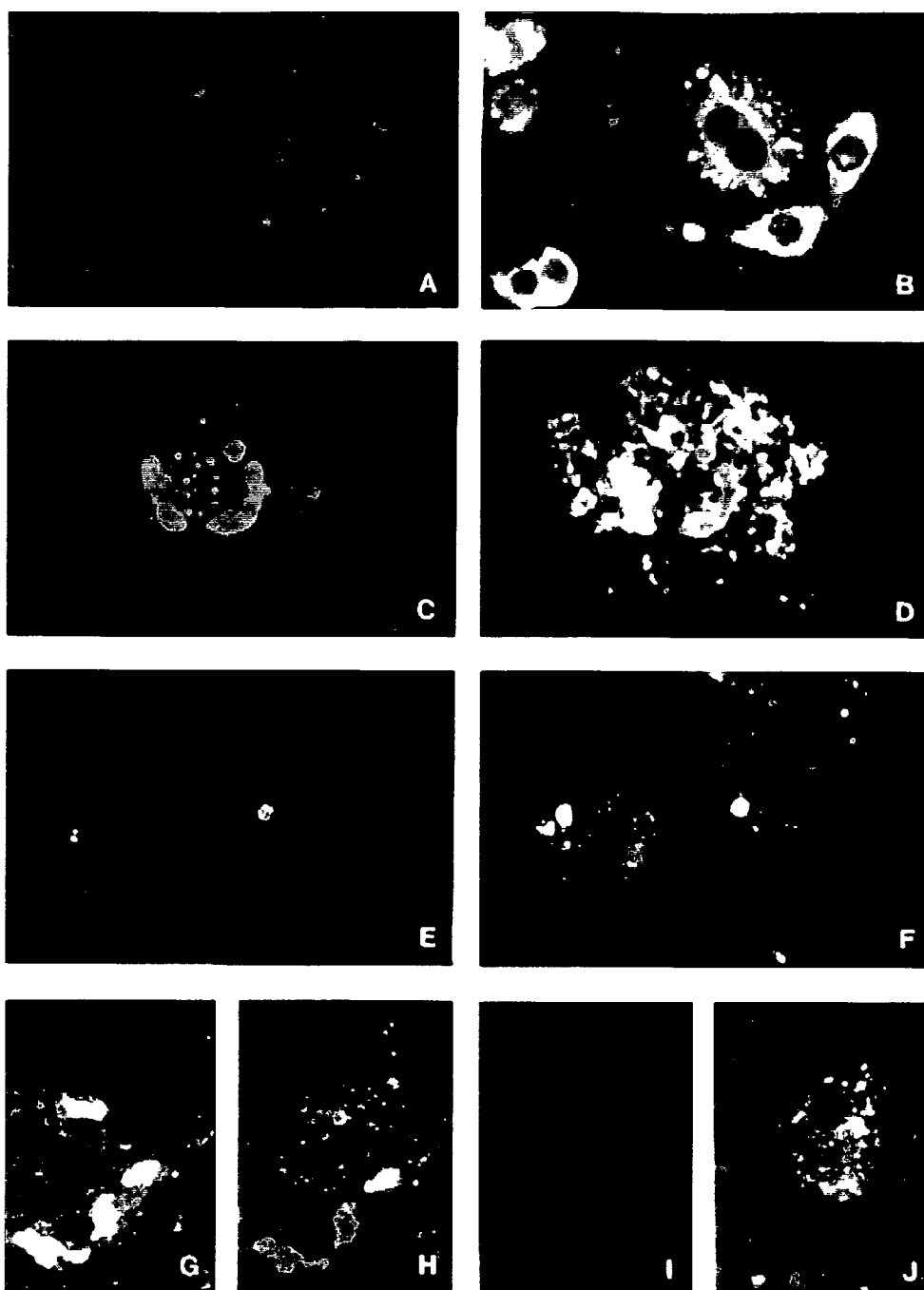


Fig. 6

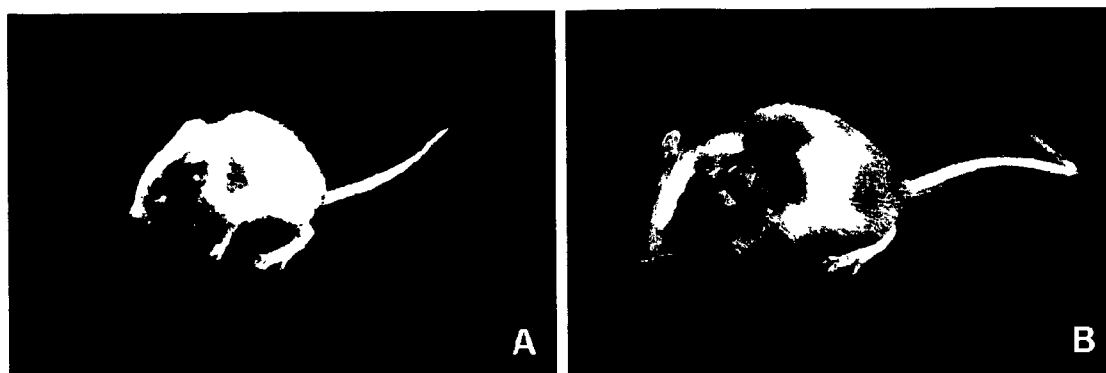


Fig. 7

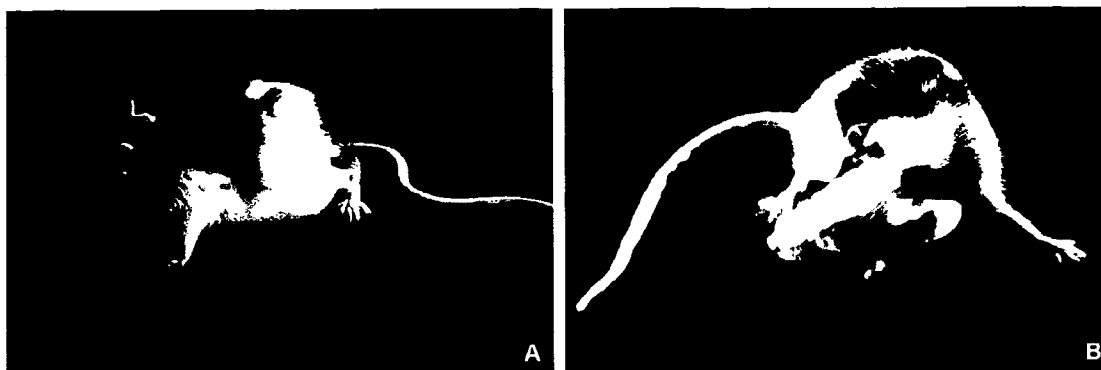


Fig. 8

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00001  ACCAAGGGGA AAATGAAGTG GTGACTCAA TCATCGAAGA CCCTCGAGAT TACATAGGTC
      ||||| ||||| ||||| ||||| ||||| |||||
00001  ACCAAGGGGA AAATGAAGTG GTGACTCAA TCATCGAAGA CCCTCGAGAT TACATAGGTC
                        Start of NP mRNA |→

00061  CGGAACCTAT GGCCTTCGTG ACCGACCTCG AGTCAGAGTA GTTCAATAAG GACCTATCAA
      ||||| ||||| ||||| ||||| ||||| |||||
00061  CGGAACCTAT GGCCTTCGTG ACCGACCTCG AGTCAGAGTA GTTCAATAAG GACCTATCAA

00121  GTTTGGGCAA TTTTTCGTCC CTGACACAAA AATGTCATCC GTGCTTAAAG CATATGAGAG 2
      ||||| ||||| ||||| ||||| ||||| |||||
00121  GTTTGGGCAA TTTTTCGTCC CCGACACAAA AATGTCATCC GTGCTTAAAG CATATGAGCG
                        |→ NP Protein Start

      Thr      Glu      Gly
00181  ATTCACACTC ACTCAAGAAC TGCAAGATCA GAGTGAAGAA GGACAATCC CACCTACAAC 3
      ||||| ||||| ||||| ||||| ||||| |||||
00181  ATTCACGCTC ACTCAAGAAC TGCAAGATCA GAGTGAGGAA GGTACAATCC CACCTACAAC

      Val      Ile→Val
00241  ACTAAAACCG GTTATCAGGG TATTTGTACT AACCTCTAAT AACCCAGAGC TAAGATCCCG 2
      ||||| ||||| ||||| ||||| ||||| |||||
00241  ACTAAAACCG GTAATCAGGG TATTTATACT AACCTCTAAT AACCCAGAGC TAAGATCCCG

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00301  GCTTCTTCTA TTCTGCCTAC GGATTGTTCT CAGTAATGGT GCAAGGGATT CCCATCGCTT
|||||
00301  GCTTCTTCTA TTCTGCCTAC GGATTGTTCT CAGTAATGGT GCAAGGGATT CCCATCGCTT

                                Leu                                Ala
00361  TGGAGCATTG CTTACAATGT TTTCGCTACC ATCAGCTACA ATGCTCAATC ATGTCAAATT 2
|||||
00361  TGGAGCATTG CTCACAATGT TTTCGCTACC ATCAGCCACA ATGCTCAATC ATGTCAAATT

00421  AGCTGACCAG TCACCAGAAG CTGATATCGA AAGGGTAGAG ATCGATGGCT TTGAGGAGGG
|||||
00421  AGCTGACCAG TCACCAGAAG CTGATATCGA AAGGGTAGAG ATCGATGGCT TTGAGGAGGG

                                Ile                                Ala Arg
00481  ATCATTCCGC TTAATTCCCA ATGCTCGCTC AGGTATGAGC CGTGGAGAGA TCAATGCCTA 3
|||||
00481  ATCATTCCGC TTAATCCCCA ATGCACGTC AGGTATGAGC CGTGGAGAGA TCAATGCCTA

00541  TGCTGCACTT GCAGAAGATC TACCTGACAC ACTAAACCAT GCAACACCTT TCGTTGATTG
|||||
00541  TGCTGCACTT GCAGAAGATC TACCTGACAC ACTAAACCAT GCAACACCTT TCGTTGATTG

                                Asp
00601  CGAAGTCGAG GGAAGTGCAT GGGACGGAGAT TGAGACTTTC TTAGATATGT GTTACAGTGT 1
|||||
00601  CGAAGTCGAG GGAAGTGCAT GGGATGAGAT TGAGACTTTC TTAGATATGT GTTACAGTGT

00661  CCTAATGCAG GCATGGATAG TGACTTGCAA GTGCATGACT GCGCCAGACC AACCTGCTGC
|||||
00661  CCTAATGCAG GCATGGATAG TGACTTGCAA GTGCATGACT GCGCCAGACC AACCTGCTGC

                                Pro
00721  TTCTATTGAG AAACGCCTGC AAAAATATCG TCAGCAAGGC AGGATCAACC CAAGATATCT 1
|||||
00721  TTCTATTGAG AAACGCCTGC AAAAATATCG TCAGCAAGGC AGGATCAACC CGAGATATCT

00781  CCTGCAACCG GAGGCTCGAC GAATAATCCA GAATGTAATC CGGAAGGGAA TGGTGGTCAG
|||||
00781  CCTGCAACCG GAGGCTCGAC GAATAATCCA GAATGTAATC CGGAAGGGAA TGGTGGTCAG

00841  ACATTTCCCTC ACCTTTGAAC TGCAGCTTGC CCGAGCACAA AGCCTTGAT CAAATAGGTA
|||||
00841  ACATTTCCCTC ACCTTTGAAC TGCAGCTTGC CCGAGCACAA AGCCTTGAT CAAATAGGTA

00901  TTATGCTATG GTAGGGGATG TTGGAAAGTA TATAGAGAAT TGTGGAATGG GAGGCTTCTT
|||||
00901  TTATGCTATG GTAGGGGATG TTGGAAAGTA TATAGAGAAT TGTGGAATGG GAGGCTTCTT

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Figure 9
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			Asp		Leu		
01621	AGTTGGAGCT	CCCATCCATA	CAGACGACCT	GAATGCCGCA	CTAGGTGATC	TTGACATCTA	2
01621	AGTTGGAGCT	CCCATCCATA	CAGATGACCT	GAATGCCGCA	CTGGGTGATC	TTGACATCTA	
					NP Protein Stop < 		
01681	GACAATTCAG	ATCCCAATCC	TAAATTGATA	TACCTAATTG	ATTAGTTAGA	TGGAACTACA	3
01681	GACAATTCAG	ATCCCAATCT	AAAATTGACA	TACCTAATTG	ATTAGTTAGA	TGGAACTACA	
01741	GTGGATTCCA	TAAGGTCCT	GCCTACCATC	GGCTTTTAAG	AAAAAAATAG	GCCCGGACGG	1
01741	GTGGATTCCA	TAAGGTCCT	GCCTACCATC	GGCTTTAAAG	AAAAAAATAG	GCCCGGACGG	
				NP mRNA End < > P / V mRNA Start			
01801	GTTAGCAACA	AGCGACTGCC	GATGCCAACA	GCGCAATCCA	CAATCTACAA	TGGATCCAC	1
01801	GTTAGCAACA	AGCGACTGCC	GGTGCCAACA	GCGCAATCCA	CAATCTACAA	TGGATCCAC	
					 >P / V Protein Start		
			Ile			Val	
01861	TGATCTGAGC	TTCTCCCCAG	ATGAGATTA	TAAGCTCATA	GAGACAGGCC	TGAATACTGT	1
01861	TGATCTGAGC	TTCTCCCCAG	ATGAGATCAA	TAAGCTCATA	GAGACAGGCC	TGAATACTGT	
01921	GGAGTATTTT	ACTTCCCAAC	AAGTCACAGG	AACATCCTCT	CTTGAAAGA	ATACAATACC	1
01921	AGAGTATTTT	ACTTCCCAAC	AAGTCACAGG	AACATCCTCT	CTTGAAAGA	ATACAATACC	
						Thr→Ile	
01981	ACCAGGGGTC	ACAGGACTAC	TAACCAATGC	TGCAGAGGCA	AAGATCCAAG	AGTCAATCAA	2
01981	ACCAGGGGTC	ACAGGACTAC	TAACCAATGC	TGCAGAGGCA	AAGATCCAAG	AGTCAACTAA	
		Gly		Ala→Thr Lys→Asn		Pro→Ser	
02041	CCATCAGAAG	GGTTCAGTTG	GTGGGGGTAC	AAACCCAAAG	AAACCGCGAT	CAAAAATTGC	4
02041	CCATCAGAAG	GGCTCAGTTG	GTGGGGGTGC	AAAACCAAG	AAACCGCGAC	CAAAAATTGC	
				Gly→Glu		Leu	
02101	CATTGTGCCA	GCAGATGACA	AAACAGTGCC	CGAAAAGCCG	ATCCCAAACC	CTCTACTAGG	2
02101	CATTGTGCCA	GCAGATGACA	AAACAGTGCC	CGGAAAGCCG	ATCCCAAACC	CTCTATTAGG	
			Thr				
02161	TCTGGACTCC	ACCCCGAGCA	CCCAAACCGT	GCTTGATCTA	AGTGGGAAAA	CATTACCATC	1
02161	TCTGGACTCC	ACCCCGAGCA	CCCAAACCTGT	GCTTGATCTA	AGTGGGAAAA	CATTACCATC	

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				Gly				
02221	AGGATCCTAT	AAGGGGGTTA	AGCTTGCGAA	ATTTGGGAAA	GAAAATCTGA	TGACACGGTT	1	
02221	AGGATCCTAT	AAGGGGGTTA	AGCTTGCGAA	ATTTGGAAAA	GAAAATCTGA	TGACACGGTT		
02281	CATCGAGGAA	CCCAGAGAGA	ATCCTATCGC	AACCAGTTCC	CCCATCGATT	TTAAGAGGGG		
02281	CATCGAGGAA	CCCAGAGAGA	ATCCTATCGC	AACCAGTTCC	CCCATCGATT	TTAAGAGGGG		
	Gly→Glu	Ala→Val						
02341	CAGAGATACC	GGTGGGTTC	ATAGAAGGGA	GTAACAATC	GGATGGGTGG	GAGATGAAGT	2	
02341	CAGGGATACC	GGCGGGTTC	ATAGAAGGGA	GTAACAATC	GGATGGGTGG	GAGATGAAGT		
02401	CAAGGTCACT	GAGTGGTGCA	ATCCATCCTG	TTCTCCAATC	ACCGCTGCAG	CAAGGCGATT		
02401	CAAGGTCACT	GAGTGGTGCA	ATCCATCCTG	TTCTCCAATC	ACCGCTGCAG	CAAGGCGATT		
	Leu	Ser→Asn						
02461	TAAATGCACT	TGTCACCAAT	GTCCAGTCAC	TTGCTCTGAA	TGTGAACGAG	ATACTTAAATA	2	
02461	TGAATGCACT	TGTCACCACT	GTCCAGTCAC	TTGCTCTGAA	TGTGAACGAG	ATACTTAAATA		
						V Protein Stop → 		
02521	CAGTGAGAAA	TTTGGACTCT	CGGATGAATC	AACTGGAGAC	AAAAGTAGAT	CGCATTCTCT		
02521	CAGTGAGAAA	TTTGGACTCT	CGGATGAATC	AACTGGAGAC	AAAAGTAGAT	CGCATTCTCT		
				Val→Ile				
02581	CATCTCAGTC	TCTAATCCAG	ACCATCAAGA	ATGACATAAT	TGGACTTAAA	GCAGGGATGG	1	
02581	CATCTCAGTC	TCTAATCCAG	ACCATCAAGA	ATGACATAGT	TGGACTTAAA	GCAGGGATGG		
02641	CTACTTTAGA	AGGAATGATT	ACAACGTGTA	AAATCATGGA	CCCGGGAGTT	CCCAGTAATG		
02641	CTACTTTAGA	AGGAATGATT	ACAACGTGTA	AAATCATGGA	CCCGGGAGTT	CCCAGTAATG		
			Thr→Lys					
02701	TTACTGTGGA	AGATGTACGC	AAGAAACTAA	GTAACCATGC	TGTTGTTGTG	CCAGAATCAT	1	
02701	TTACTGTGGA	AGATGTACGC	AAGACACTAA	GTAACCATGC	TGTTGTTGTG	CCAGAATCAT		
02761	TCAATGATAG	TTTCTTGACT	CAATCTGAAG	ATGTAATTC	ACTTGATGAG	TTGGCTCGAC		
02761	TCAATGATAG	TTTCTTGACT	CAATCTGAAG	ATGTAATTC	ACTTGATGAG	TTGGCTCGAC		
02821	CAACTGCAAC	AAGTGTTAAG	AAGATTGTCA	GGAAGGTTCC	TCCTCAGAAG	GATCTGACTG		
02821	CAACTGCAAC	AAGTGTTAAG	AAGATTGTCA	GGAAGGTTCC	TCCTCAGAAG	GATCTGACTG		

Ile
 02881 GATTGAAGAT CACACTAGAG CAATTGGCAA AGGATTGCAT CAGCAAACCG AAGATGAGGG 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 02881 GATTGAAGAT TACACTAGAG CAATTGGCAA AGGATTGCAT CAGCAAACCG AAGATGAGGG

Glu→Asp **Lys** **Ser**
 02941 AAGATTTATCT CCTCAAGATC AACCAGGCTT CTAGTGAGGC TCAGCTAATT GACCTCAAGA 3
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 02941 AAGAGTATCT CCTCAAAATC AACCAGGCTT CCAGTGAGGC TCAGCTAATT GACCTCAAGA

03001 AAGCAATCAT CCGCAGTGCA ATTTGATCAA GAAACACCCA ATTACACTAC ACTGGTATGA
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 03001 AAGCAATCAT CCGCAGTGCA ATTTGATCAA GAAACACCCA ATTACACTAC ACTGGTATGA
 P Protein Stop ←|

03061 CACTGTACTA ACCCTGAGGG TTTTAGAAAA AACGATTAAC GATAAATAAG CCCGAACACT
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 03061 CACTGTACTA ACCCTGAGGG TTTTAGAAAA AACGATTAAC GATAAATAAG CCCGAACACT
 P mRNA End ←| |→ M mRNA Start

03121 ACATACTACC TGAGGCAGCC ATGCCATCCA TCAGCATTCG CGCAGACCCC ACCAATCCAC 1
 ||| ||||| ||||| ||||| ||||| ||||| |||||
 03121 ACACACTACC TGAGGCAGCC ATGCCATCCA TCAGCATTCG CGCAGACCCC ACCAATCCAC
 |→ M Protein Start

Ile
 03181 GTCAATCAAT AAAAGCGTTC CCAATTGTGA TTAACAGTGA TGGGGGTGAG AAAGGCCGCT 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 03181 GTCAATCAAT AAAAGCGTTC CCAATTGTGA TCAACAGTGA TGGGGGTGAG AAAGGCCGCT

Arg
 03241 TGGTTAAACA ACTACGTACA ACCTACTTGA ATGACCTAGA TACTCATGAG CCACTGGTGA 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 03241 TGGTTAAACA ACTACGCACA ACCTACTTGA ATGACCTAGA TACTCATGAG CCACTGGTGA

Ile→Val **Asp→Asn** **Thr→Ala**
 03301 CATTCGTAAA TACCTATGGA TTCATCTACG AACAGAATCG GGGGAATGCC ATTGTCTGGAG 3
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 03301 CATTCATAAA TACCTATGGA TTCATCTACG AACAGGATCG GGGGAATACC ATTGTCTGGAG

Thr
 03361 AGGATCAACT TGGGAAGAAA AGAGAGGCTG TGACTTGCTGC AATGGTTACC CTTGGATGTG 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 03361 AGGATCAACT TGGGAAGAAA AGAGAGGCTG TGACCGCTGC AATGGTTACC CTTGGATGTG

Gly→Arg **Arg→Ser** **Gln**
 03421 GGCCTAATCT ACCATCATTA GGGAAATGTCC TGAGACAACT GAGTGAATTC CAAGTCAATTG 3
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 03421 GGCCTAATCT ACCATCATTA GGGAAATGTCC TGGGACAACT GAGGGAATTC CAGGTCACCTG

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03481  TTAGGAAGAC  ATCCAGCAAA  GCGGAAGAGA  TGGTCTTTGA  AATTGTAAAG  TATCCGAGAA
|||||||  |||||||  |||||||  |||||||  |||||||  |||||||
03481  TTAGGAAGAC  ATCCAGCAAA  GCGGAAGAGA  TGGTCTTTGA  AATTGTAAAG  TATCCGAGAA

03541  TATTTCTGGG  TCATACATTA  ATCCAGAAAG  GACTAGTCTG  TGTCTCCGCA  GAAAAATTTG
|||||||  |||||||  |||||||  |||||||  |||||||  |||||||
03541  TATTTCTGGG  TCATACATTA  ATCCAGAAAG  GACTAGTCTG  TGTCTCCGCA  GAAAAATTTG

                                Ile→Val
03601  TTAAGTCACC  AGGGAAAGTA  CAATCTGGAA  TGGACTATCT  CTCATTCCG  ACATTTCTGT  1
|||||||  |||||||  |||||||  |||||||  |||||||  |||||||
03601  TTAAGTCACC  AGGGAAAATA  CAATCTGGAA  TGGACTATCT  CTCATTCCG  ACATTTCTGT

                                Tyr
03661  CAGTGATCTTA  TGTGCCAGCT  GCAATCAAAT  TTCAGGTACC  TGGCCCCATG  TTGAAAATGA  1
|||||||  |||||||  |||||||  |||||||  |||||||  |||||||
03661  CAGTGATCTTA  CTGTGCCAGCT  GCAATCAAAT  TTCAGGTACC  TGGCCCCATG  TTGAAAATGA

                                Arg
03721  GGTCAAAGATA  CACTCAGAGC  TTACAACCTG  AACTAATGAT  AAGAATCCTG  TGTAAGCCCG  1
|  |||||||  |||||||  |||||||  |||||||  |||||||  |||||||
03721  GATCAAGATA  CACTCAGAGC  TTACAACCTG  AACTAATGAT  AAGAATCCTG  TGTAAGCCCG

                                Thr→Ile
03781  ATTCGCCACT  TATGAAGGTC  CATATCCCTG  ACAAGGAGGG  AAGAGGATGT  CTTGTATCAG  1
||||  ||||  ||||||  ||||  ||||  ||||||  ||||||
03781  ATTCGCCACT  TATGAAGGTC  CATACCCCTG  ACAAGGAGGG  AAGAGGATGT  CTTGTATCAG

03841  TATGGCTGCA  TGTATGCAAC  ATCTTCAAAT  CAGGAAACAA  GAATGGCAGT  GAGTGGCAGG
|||||||  |||||||  |||||||  |||||||  |||||||  |||||||
03841  TATGGCTGCA  TGTATGCAAC  ATCTTCAAAT  CAGGAAACAA  GAATGGCAGT  GAGTGGCAGG

03901  AATACTGGAT  GAGAAAGTGT  GCTAACATGC  AACTTGAAGT  GTCGATTGCA  GATATGTGGG
|||||||  |||||||  |||||||  |||||||  |||||||  |||||||
03901  AATACTGGAT  GAGAAAGTGT  GCTAACATGC  AACTTGAAGT  GTCGATTGCA  GATATGTGGG

03961  GACCAACTAT  CATAATTCAT  GCCAGAGGTC  ACATTCCCAA  AAGTGCTAAG  TTGTTTTTTG
|||||||  |||||||  |||||||  |||||||  |||||||  |||||||
03961  GACCAACTAT  CATAATTCAT  GCCAGAGGTC  ACATTCCCAA  AAGTGCTAAG  TTGTTTTTTG

                                Val→Ile
04021  GAAAGGGTGG  ATGGAGCTGC  CATCCACTTC  ACGAAATTGT  TCCAAGTGTC  ACTAAAACAC  1
|||||||  |||||||  |||||||  ||||  ||||  |||||||  |||||||
04021  GAAAGGGTGG  ATGGAGCTGC  CATCCACTTC  ACGAAGTTGT  TCCAAGTGTC  ACTAAAACAC

                                Val      Glu
04081  TATGGTCCGT  AGGTGTGAA  ATTACAAAGG  CGAAGGCAAT  AATACAAGAG  AGTAGCATCT  2
|||||||  |||||||  |||||||  |||||||  |||||||  |||||||
04081  TATGGTCCGT  GGGCTGTGAG  ATTACAAAGG  CGAAGGCAAT  AATACAAGAG  AGTAGCATCT

```

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Val

	Val→Gly				Leu		
04201	GCCGCTTTG <u>G</u>	GAAATCAAAT	TGGGGTCTGT	TCAAGAAAAC	TAAATCACT <u>A</u>	CCTAACCTAA	2
04201	GCCGCTTTGT	GAAATCAAAT	TGGGGTCTGT	TCAAGAAAAC	TAAATCACTG	CCTAACCTGA	

M Protein Stop ←

04381 GATTAGAGAG CTTAATTAAC TCTGTATTAA TAATAACACT ACTATTCCAA TAACTGGAAT 1
|||||
04381 GATTAGAGAG CTTAATTAGC TCTGTATTAA TAATAACACT ACTATTCCAA TAACTGGAAT

M mRNA End ←|

→ F mRNA Start

→ F Protein Start

04621 TTCCAACAAA TGTCCGGCAA CTTATGTATT ATACTGAGGC CTCATCAGCA TTCATTGTTG
|||||||
04621 TTCCAACAAA TGTCCGGCAA CTTATGTATT ATACTGAGGC CTCATCAGCA TTCATTGTTG

04681 TGAAGTTAAT GCCTACAATT GACTCGCCGA TTAGTGATG TAATATAACA TCAATTTCAA
 04681 TGAAGTTAAT GCCTACAATT GACTCGCCGA TTAGTGATG TAATATAACA TCAATTTCAA

04741 GCTATAATGC AACAGTGACA AAACCTCCTAC AGCCGATCGG TGAGAATTTG GAGACGATTA
||||||| ||||| ||||| ||||| ||||| |||||
04741 GCTATAATGC AACAGTGACA AAACCTCCTAC AGCCGATCGG TGAGAATTTG GAGACGATTA

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04801  GGAACCAGTT GATTCCAAC TCGGAGGAGAC GCCGGTTTGC AGGGGTGGTG ATTGGATTAG
|||||
04801  GGAACCAGTT GATTCCAAC TCGGAGGAGAC GCCGGTTTGC AGGGGTGGTG ATTGGATTAG

04861  CTGCATTAGG AGTAGCTACT GCCGCACAAG TCACTGCCGC AGTAGCACTA GTTAAAGGCAA 3
|||||
04861  CTGCATTAGG AGTAGCTACT GCCGCACAGG TCACTGCCGC AGTGGCACTA GTAAAGGCAA

04921  ATGAAAATAC TGC GGCTATA CTCAATCTCA AAAATGCAAT CCAAAAAACA AATGCAGCAG 1
|||||
04921  ATGAAAATGC TGC GGCTATA CTCAATCTCA AAAATGCAAT CCAAAAAACA AATGCAGCAG

04981  TTGCAGATGT GGTCCAGGCC ACACAATCAC TAGGAACGGC AGTTCAAGCA GTTCAAGATC
|||||
04981  TTGCAGATGT GGTCCAGGCC ACACAATCAC TAGGAACGGC AGTTCAAGCA GTTCAAGATC

05041  ACATAAACAG TGTGATAAGT CCAGCAATTA CAGCAGCCAA TTGTAAGGCC CAAGATGCTA 1
|||||
05041  ACATAAACAG TGTGGTAAGT CCAGCAATTA CAGCAGCCAA TTGTAAGGCC CAAGATGCTA

05101  TCATTGGCTC AATCCTCAAT CTCTATTTGA CCGAGTTGAC AACTTATCTTC CACAATCAAA 1
|||||
05101  TCATTGGCTC AATCCTCAAT CTCTATTTGA CCGAGTTGAC AACCATCTTC CACAATCAAA

05161  TTACAAACCC TGCATTGAGT CCTATTACAA TTCAAGCTTT AAGGATCCTA CTGGGGAGTA 1
|||||
05161  TTACAAACCC TGCATTGAGT CCCATTACAA TTCAAGCTTT AAGGATCCTA CTGGGGAGTA

05221  CCTTGCCGAC TGTGGTCGAA AAATCTTTCA ATACCCAGAT AAGTGCAGCT GAGCTTCTCT
|||||
05221  CCTTGCCGAC TGTGGTCGAA AAATCTTTCA ATACCCAGAT AAGTGCAGCT GAGCTTCTCT

05281  CATCAGGGTT GTTGACAGGC CAGATTGTGG GATTAGATT GACCTATATG CAGATGGTCA 1
|||||
05281  CATCAGGGTT ATTGACAGGC CAGATTGTGG GATTAGATT GACCTATATG CAGATGGTCA

05341  TAAAAATTGA GCTGCCAACT TTAAGTGTAC AACCTGCAAC CCAGATCATA GATCTGGCCA
|||||
05341  TAAAAATTGA GCTGCCAACT TTAAGTGTAC AACCTGCAAC CCAGATCATA GATCTGGCCA

05401  CCATTCTGCG ATTCATTAAC AATCAAGAAG TCATGGCCCA ATTACCAACA CGTGTATTG 1
|||||
05401  CCATTCTGCG ATTCATTAAC AATCAAGAAG TCATGGCCCA ATTACCAACA CGTGTATGG

```

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					Thr		
05461	TGACTGGCAG	CTTGATCCAA	GCCTATCCCG	CATCGCAATG	CAC <u>T</u> ATTACA	CCCAACACTG	1
05461	TGACTGGCAG	CTTGATCCAA	GCCTATCCCG	CATCGCAATG	CACCATTACA	CCCAACACTG	
					Thr		
05521	TGTACTCTAG	GTATAATGAT	GCCCAAGTAC	TCTCAGATGA	TAC <u>G</u> ATGGCT	TGCCTCCAAG	1
05521	TGTACTGTAG	GTATAATGAT	GCCCAAGTAC	TCTCAGATGA	TACTATGGCT	TGCCTCCAAG	
05581	GTAACCTGAC	AAGATGCACC	TTCTCTCCAG	TGGTTGGGAG	CTTTCTCACT	CGATTCTGTC	
05581	GTAACCTGAC	AAGATGCACC	TTCTCTCCAG	TGGTTGGGAG	CTTTCTCACT	CGATTCTGTC	
					Leu		
05641	TGTTTCGATGG	AATAGTTTAT	GCAAATTGCA	GGTCGATG <u>C</u> T	GTGCAAGTGC	ATGCAACCTG	1
05641	TGTTTCGATGG	AATAGTTTAT	GCAAATTGCA	GGTCGATGTT	GTGCAAGTGC	ATGCAACCTG	
	Ala					Tyr→His	
05701	CTG <u>C</u> CGTGAT	CCTACAGCCG	AGTTCATCCC	CTGTAACGTG	CATTGACATG	<u>C</u> ACAAATGTG	2
05701	CTGCTGTGAT	CCTACAGCCG	AGTTCATCCC	CTGTAACGTG	CATTGACATG	TACAAATGTG	
		Asn→Asp					
05761	TGAGTCTGCA	GCTTGAC <u>G</u> AT	CTCAGATTCA	CCATCACTCA	ATTGGCCAAT	GTAACCTACA	1
05761	TGAGTCTGCA	GCTTGACAAT	CTCAGATTCA	CCATCACTCA	ATTGGCCAAT	GTAACCTACA	
		Ser→Thr		Ser→Pro			
05821	ATAGCACCAT	CAAGCTTGAA	<u>A</u> CATCCCAGA	TCTTG <u>C</u> CTAT	TGATCCGTTG	GATATATCCC	2
05821	ATAGCACCAT	CAAGCTTGAA	TCATCCCAGA	TCTTGTCTAT	TGATCCGTTG	GATATATCCC	
	Gln	Leu					
05881	<u>A</u> GAAT <u>T</u> TAGC	TGCGGTGAAT	AAGAGTCTAA	GTGATGCACT	ACAACACTTA	GCACAAAGTG	2
05881	AAAATCTAGC	TGCGGTGAAT	AAGAGTCTAA	GTGATGCACT	ACAACACTTA	GCACAAAGTG	
	Tyr						
05941	ACACATAC <u>C</u> CT	TTCTGCAATC	ACATCAGCTA	CGACTACAAG	TGTATTATCC	ATAATAGCAA	1
05941	ACACATATCT	TTCTGCAATC	ACATCAGCTA	CGACTACAAG	TGTATTATCC	ATAATAGCAA	
06001	TCTGTCTTGG	ATCGTTAGGT	TTAATATTAA	TAATCTTGCT	CAGTGTAGTT	GTGTGGAAGT	
06001	TCTGTCTTGG	ATCGTTAGGT	TTAATATTAA	TAATCTTGCT	CAGTGTAGTT	GTGTGGAAGT	

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06661	ACTACTAGCA	TTAAGCATCT	CTATCCTTTA	TGAGAGTTTA	ATAACCCAAA	AGCAAATCAT	
06661	ACTACTAGCA	TTAAGCATCT	CTATCCTTTA	TGAGAGTTTA	ATAACCCAAA	AGCAAATCAT	
	Ser			Ser→Gly Ile			
06721	GAG <u>T</u> CAAGCA	GGCTCAACTG	GATCTAATTC	TGGATTAGGA	<u>G</u> GTAT <u>T</u> ACTG	ATCTTCTTAA	3
06721	GAGCCAAGCA	GGCTCAACTG	GATCTAATTC	TGGATTAGGA	AGTATCACTG	ATCTTCTTAA	
	Leu						
06781	TAATATTCT <u>T</u>	TCTGTCGCAA	ATCAGATTAT	ATATAACTCT	GCAGTCGCTC	TACCTCTACA	1
06781	TAATATTCTC	TCTGTCGCAA	ATCAGATTAT	ATATAACTCT	GCAGTCGCTC	TACCTCTACA	
06841	ATTGGACACT	CTTGAATCAA	CACTCCTTAC	AGCCATTAAG	TCTCTTCAAA	CCAGTGACAA	
06841	ATTGGACACT	CTTGAATCAA	CACTCCTTAC	AGCCATTAAG	TCTCTTCAAA	CCAGTGACAA	
			Ser→Gly				
06901	GCTAGAACAG	AACTGCTCGT	GG <u>G</u> TGCTGC	ACTGATTAAT	GATAATAGAT	ACATTAATGG	1
06901	GCTAGAACAG	AACTGCTCGT	GGAGTGCTGC	ACTGATTAAT	GATAATAGAT	ACATTAATGG	
	Phe						
06961	CATCAATCAG	TTCTATTT <u>C</u> T	CAATTGCTGA	GGGTCGCAAT	CTGACACTTG	GCCCACTTCT	1
06961	CATCAATCAG	TTCTATTTTT	CAATTGCTGA	GGGTCGCAAT	CTGACACTTG	GCCCACTTCT	
	Met→Ile						
07021	TAATAT <u>A</u> CCT	AGTTTCATTC	CAACTGCCAC	GACACCAGAG	GGCTGCACCA	GGATCCCATC	1
07021	TAATATGCCT	AGTTTCATTC	CAACTGCCAC	GACACCAGAG	GGCTGCACCA	GGATCCCATC	
			Thr				
07081	ATTCTCGCTC	ACTAAGACAC	ACTGGTGTTA	TAC <u>G</u> CACAAAT	GTTATCCTGA	ATGGATGCCA	1
07081	ATTCTCGCTC	ACTAAGACAC	ACTGGTGTTA	TACACACAAAT	GTTATCCTGA	ATGGATGCCA	
07141	GGATCATGTA	TCCTCAAATC	AATTTGTTTC	CATGGGAATC	ATTGAACCCA	CTTCTGCCGG	
07141	GGATCATGTA	TCCTCAAATC	AATTTGTTTC	CATGGGAATC	ATTGAACCCA	CTTCTGCCGG	
	Phe→Ser						
07201	GTTTCCAT <u>C</u> C	TTTCGAACCT	TAAAGACTCT	ATATCTCAGC	GATGGGGTCA	ATCGTAAGAG	1
07201	GTTTCCATTTC	TTTCGAACCC	TAAAGACTCT	ATATCTCAGC	GATGGGGTCA	ATCGTAAGAG	
				Val			
07261	CTGCTCTATC	AGTACAGTTC	CGGGGGGTTG	TATGATGTAC	TGTTTTGT <u>C</u> T	CTACTCAACC	1
07261	CTGCTCTATC	AGTACAGTTC	CGGGGGGTTG	TATGATGTAC	TGTTTTGTTT	CTACTCAACC	

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Ala→Thr

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Ser

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Val

08641 TCTTAGAAAG GGCCATTGGC AAGAGTATGT AAATGTAATA CTGTGGCCGC GAATTCTTCC 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 08641 TCTTAGAAAG GGCCATTGGC AAGAGTATGT CAATGTAATA CTGTGGCCGC GAATTCTTCC

08701 CTTGATCCCG GATTTTAAAA TCAATGACCA ATTGCCTCTG CTCAAAAATT GGGACAAGTT
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 08701 CTTGATCCCG GATTTTAAAA TCAATGACCA ATTGCCTCTG CTCAAAAATT GGGACAAGTT

Ala

08761 AGTTAAAGAA TCATGTTTCA TAATCAATGC GGGTACTTCC CAGTGCATTC AGAATCTCAG 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 08761 AGTTAAAGAA TCATGTTTCA TAATCAATGC AGGTACTTCC CAGTGCATTC AGAATCTCAG

08821 CTATGGACTG ACAGGTCGTG GGAACCTCTT TACACGATCA CGTGAACCTCT CTGGTGACCG
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 08821 CTATGGACTG ACAGGTCGTG GGAACCTCTT TACACGATCA CGTGAACCTCT CTGGTGACCG

Thr

08881 CAGGGATATT GATCTTAAGA CGGTGTGGC AGCATGGCAT GACTCAGACT GGAAAAGAAT 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 08881 CAGGGATATT GATCTTAAGA CAGTTGTGGC AGCATGGCAT GACTCAGACT GGAAAAGAAT

08941 AAGTGATTTT TGGATTATGA TCAAATCCA GATGAGACAA TTAATTGTGA GGCAAACAGA
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 08941 AAGTGATTTT TGGATTATGA TCAAATCCA GATGAGACAA TTAATTGTGA GGCAAACAGA

Ser→Pro

09001 TCATAATGAT CCTGATTTAA TCACGTATAT CGAAAATAGA GAAGGCATAA TCATCATAAC 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 09001 TCATAATGAT TCTGATTTAA TCACGTATAT CGAAAATAGA GAAGGCATAA TCATCATAAC

Asn

09061 CCCTGAACTG GTAGCATTAT TTAATACTGA GAATCATACA CTAACATACA TGACCTTTGA 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 09061 CCCTGAACTG GTAGCATTAT TTAACACTGA GAATCATACA CTAACATACA TGACCTTTGA

09121 AATTGTACTG ATGGTTTCAG ATATGTACGA AGGTCGTCAC AACATTTTAT CACTATGCAC
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 09121 AATTGTACTG ATGGTTTCAG ATATGTACGA AGGTCGTCAC AACATTTTAT CACTATGCAC

09181 AGTTAGCACT TACCTGAATC CTCTGAAGAA AAGAATAACA TATTTATTGA GCCTTG TAGA
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 09181 AGTTAGCACT TACCTGAATC CTCTGAAGAA AAGAATAACA TATTTATTGA GCCTTG TAGA

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09241  TAACTTAGCT  TTTCAGATAG  GTGATGCTGT  ATATAACATA  ATTGCTTTGC  TAGAATCCTT
      |||||
09241  TAACTTAGCT  TTTCAGATAG  GTGATGCTGT  ATATAACATA  ATTGCTTTGC  TAGAATCCTT

09301  TGTATATGCA  CAGTTGCAAA  TGTCAGATCC  CATCCCAGAA  CTCAGAGGAC  AATTCCATGC
      |||||
09301  TGTATATGCA  CAGTTGCAAA  TGTCAGATCC  CATCCCAGAA  CTCAGAGGAC  AATTCCATGC

09361  ATTCGTATGT  TCTGAGATTC  TTGATGCACT  Arg AAGGGAAGT  AATAGTTTCA  CCCAGGATGA  1
      |||||
09361  ATTCGTATGT  TCTGAGATTC  TTGATGCACT  AAGAGGAAGT  AATAGTTTCA  CCCAGGATGA

09421  ATTAAGAAGT  GTGACAAGTA  ATTTGATATC  CCCATTCCAA  GATCTGACCC  CAGATCTTAC
      |||||
09421  ATTAAGAAGT  GTGACAAGTA  ATTTGATATC  CCCATTCCAA  GATCTGACCC  CAGATCTTAC

09481  GGCTGAATTG  CTCTGTATAA  TGAGGCTTTG  GGGACACCCC  ATGCTCACTG  CCAGTCAAGC
      |||||
09481  GGCTGAATTG  CTCTGTATAA  TGAGGCTTTG  GGGACACCCC  ATGCTCACTG  CCAGTCAAGC

09541  Ala TGCGGGAAG  GTACGCGAGT  CTATGTGTGC  Gly TGGGAAAGTA  Leu CTGGACTTTC  Thr CCACTATTAT  5
      |||||
09541  TGCAGGAAAG  GTACGCGAGT  CTATGTGTGC  TGGAAAAGTA  TTAGACTTTC  CCACCATTAT

09601  GAAAACACTA  GCCTTTTTTC  ATACTATTCT  GATCAATGGA  TACAGGAGGA  AGCATCATGG
      |||||
09601  GAAAACACTA  GCCTTTTTTC  ATACTATTCT  GATCAATGGA  TACAGGAGGA  AGCATCATGG

09661  AGTATGGCCA  CCCTTAAACT  Pro TACCAAGTAA  TGCTTCAAAG  Thr GGTCTCACAG  AACTTATGAA  2
      |||||
09661  AGTATGGCCA  CCCTTAAACT  TACCGGGTAA  TGCTTCAAAG  GGTCTCACGG  AACTTATGAA

09721  Asn TGACAACACT  Glu GAGATAAGCT  ATGAATTCAC  ACTTAAGCAT  Val→Ile TGGAAGGAA  TCTCTCTTAT  3
      |||||
09721  TGACAATACT  GAAATAAGCT  ATGAATTCAC  ACTTAAGCAT  TGGAAGGAAG  TCTCTCTTAT

09781  AAAATTCAAG  AAATGTTTTG  ATGCAGACGC  AGGTGAGGAA  CTCAGTATAT  TTATGAAAGA
      |||||
09781  AAAATTCAAG  AAATGTTTTG  ATGCAGACGC  AGGTGAGGAA  CTCAGTATAT  TTATGAAAGA

09841  Lys TAAAGCAATT  AGTGCCCCAA  AACAAGACTG  GATGAGTGTG  TTTAGAAGAA  GCCTAATCAA  1
      |||||
09841  TAAGGCAATT  AGTGCCCCAA  AACAAGACTG  GATGAGTGTG  TTTAGAAGAA  GCCTAATCAA

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						Leu	
09901	ACAGCGCCAT	CAGCATCATC	AGGTCCCCCT	ACCAAATCCA	TTCAATCGAC	GGCT <u>A</u> TTGCT	1
09901	ACAGCGCCAT	CAGCATCATC	AGGTCCCCCT	ACCAAATCCA	TTCAATCGAC	GGCTGTTGCT	
09961	AAACTTTCTC	GGAGATGACA	AATTCGACCC	GAATGTGGAG	CTACAGTATG	TAACATCAGG	
09961	AAACTTTCTC	GGAGATGACA	AATTCGACCC	GAATGTGGAG	CTACAGTATG	TAACATCAGG	
10021	TGAGTATCTA	CATGATGACA	CGTTTTGTGC	ATCATATTCA	CTAAAAGAGA	AGGAAATTAA	
10021	TGAGTATCTA	CATGATGACA	CGTTTTGTGC	ATCATATTCA	CTAAAAGAGA	AGGAAATTAA	
10081	ACCTGATGGT	CGAATTTTTG	CAAAGTTGAC	TAAGAGAATG	AGATCATGTC	AAGTTATAGC	
10081	ACCTGATGGT	CGAATTTTTG	CAAAGTTGAC	TAAGAGAATG	AGATCATGTC	AAGTTATAGC	
10141	AGAATCTCTT	TTAGCGAACC	ATGCTGGGAA	GTTAATGAAA	GAGAATGGTG	TTGTGATGAA	
10141	AGAATCTCTT	TTAGCGAACC	ATGCTGGGAA	GTTAATGAAA	GAGAATGGTG	TTGTGATGAA	
						Lys→Arg	
10201	TCAGCTATCA	TTAACAAAAT	CACTATTAAC	AATGAGTCAG	ATTGGAATAA	TATCCGAGAG	1
10201	TCAGCTATCA	TTAACAAAAT	CACTATTAAC	AATGAGTCAG	ATTGGAATAA	TATCCGAGAA	
		Ser		Gln→Arg			
10261	AGCTAGAAAG	TCC <u>G</u> ACTCGAG	ATAACATAAA	TCC <u>G</u> ACCTGGT	TTCCAGAATA	TCCAGAGAAA	2
10261	AGCTAGAAAG	TCAACTCGAG	ATAACATAAA	TCAACCTGGT	TTCCAGAATA	TCCAGAGAAA	
10321	TAAATCACAT	CACTCCAAGC	AAGTCAATCA	GCGAGATCCA	AGTGATGACT	TTGAATTGGC	
10321	TAAATCACAT	CACTCCAAGC	AAGTCAATCA	GCGAGATCCA	AGTGATGACT	TTGAATTGGC	
					Asn		
10381	AGCATCTTTT	TTAACTACTG	ATCTCAAAAA	ATATTGTTTA	CAATGGAGGT	AT <u>C</u> CAGACAAT	1
10381	AGCATCTTTT	TTAACTACTG	ATCTCAAAAA	ATATTGTTTA	CAATGGAGGT	ACCAGACAAT	
		Leu			Phe		
10441	TATCCCATTT	GCTCAATC <u>A</u> C	TAAACAGAAT	GTATGGTTAT	CCTCATCTCT	T <u>C</u> GAGTGGAT	2
10441	TATCCCATTT	GCTCAATCAT	TAAACAGAAT	GTATGGTTAT	CCTCATCTCT	TTGAGTGGAT	

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	Leu			Asp			
10501	TCAC <u>T</u> TCGG	CTAATGCGTA	GTACACTTTA	CGTGGGGG <u>A</u> C	CCCTTCAACC	CACCAGCAGA	2
10501							
10501	TCAC <u>T</u> TACGG	CTAATGCGTA	GTACACTTTA	CGTGGGGGAT	CCCTTCAACC	CACCAGCAGA	
10561	TACCAGTCAA	TTTGATCTAG	ATAAAGTAAT	TAATGGAGAT	ATCTTCATTG	TATCACCCAG	
10561							
10561	TACCAGTCAA	TTTGATCTAG	ATAAAGTAAT	TAATGGAGAT	ATCTTCATTG	TATCACCCAG	
			Lys		Ala→Ser		
10621	AGGTGGAATT	GAAGGGCTGT	GTCAAAA <u>A</u> GC	TTGGACAATG	ATATCTATC <u>T</u>	CTGTGATAAT	2
10621							
10621	AGGTGGAATT	GAAGGGCTGT	GTCAAAAAGGC	TTGGACAATG	ATATCTATCG	CTGTGATAAT	
				Ser			
10681	TCTATCTGCC	ACAGAGTCTG	GCACACGAGT	AATGAG <u>C</u> ATG	GTGCAGGGAG	ATAATCAAGC	1
10681							
10681	TCTATCTGCC	ACAGAGTCTG	GCACACGAGT	AATGAGTATG	GTGCAGGGAG	ATAATCAAGC	
10741	AATTGCTGTC	ACCACACGAG	TACCAAGGAG	CCTGCCGACT	CTTGAGAAAA	AGACTATTGC	
10741							
10741	AATTGCTGTC	ACCACACGAG	TACCAAGGAG	CCTGCCGACT	CTTGAGAAAA	AGACTATTGC	
		Leu					
10801	TTT <u>T</u> AGATCT	TGTAAT <u>T</u> TAT	TCTTTGAGAG	GTTAAAAATGT	AATAATTTTG	GATTAGGTCA	1
10801							
10801	TTT <u>T</u> AGATCT	TGTAATCTAT	TCTTTGAGAG	GTTAAAAATGT	AATAATTTTG	GATTAGGTCA	
10861	CCATTGAAA	GAACAAGAGA	CTATCATTAG	TTCTCACTTC	TTTGTTTATA	GCAAGAGAAT	
10861							
10861	CCATTGAAA	GAACAAGAGA	CTATCATTAG	TTCTCACTTC	TTTGTTTATA	GCAAGAGAAT	
10921	ATTCTATCAG	GGGAGGATTC	TAACGCAAGC	CTTAAAAAAT	GCTAGTAAGC	TCTGCTTGAC	
10921							
10921	ATTCTATCAG	GGGAGGATTC	TAACGCAAGC	CTTAAAAAAT	GCTAGTAAGC	TCTGCTTGAC	
10981	AGCTGATGTC	CTAGGAGAAT	GCACCCAATC	ATCATGTTCT	AATCTTGCAA	CTACTGTCAT	
10981							
10981	AGCTGATGTC	CTAGGAGAAT	GCACCCAATC	ATCATGTTCT	AATCTTGCAA	CTACTGTCAT	
11041	GAGGTAACT	GAGAATGGTG	TTGAAAAAGA	TATCTGTTTC	TACTTGAATA	TCTATATGAC	
11041							
11041	GAGGTAACT	GAGAATGGTG	TTGAAAAAGA	TATCTGTTTC	TACTTGAATA	TCTATATGAC	
				Ser			
11101	CATCAAACAG	CTCTCCTATG	ATATCATCTT	CCCTCAAGTG	T <u>C</u> GATTCCTG	GAGATCAGAT	1
11101							
11101	CATCAAACAG	CTCTCCTATG	ATATCATCTT	CCCTCAAGTG	TCAATTCCTG	GAGATCAGAT	

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						Leu	
11161	CACATTAGAA	TACATAAATA	ATCCACACCT	GGTATCACGA	TTGGCTCTTT	T <u>A</u> CCATCCCA	1
11161	CACATTAGAA	TACATAAATA	ATCCACACCT	GGTATCACGA	TTGGCTCTTT	TGCCATCCCA	
			Leu			Gly	
11221	GTTAGGAGGT	CTAAACTACT	TGTCATGCAG	TAGGCTGTTC	AATCGAAACA	TAGG <u>T</u> GACCC	2
11221	GTTAGGAGGT	CTAAACTACC	TGTCATGCAG	TAGGCTGTTC	AATCGAAACA	TAGGCGACCC	
11281	GGTGGTTTCC	GCAGTTGCAG	ATCTTAAGAG	ATTAATTAAA	TCAGGATGTA	TGGATTACTG	
11281	GGTGGTTTCC	GCAGTTGCAG	ATCTTAAGAG	ATTAATTAAA	TCAGGATGTA	TGGATTACTG	
11341	GATCCTTTAT	AACTTATTAG	GGAGAAAACC	GGGAAACGGC	TCATGGGCTA	CTTTAGCAGC	
11341	GATCCTTTAT	AACTTATTAG	GGAGAAAACC	GGGAAACGGC	TCATGGGCTA	CTTTAGCAGC	
				Pro			
11401	TGACCCGTAC	TCAATCAATA	TAGAGTATCA	ATACCC <u>C</u> CCA	ACTACAGCTC	TTAAGAGGCA	1
11401	TGACCCGTAC	TCAATCAATA	TAGAGTATCA	ATACCCTCCA	ACTACAGCTC	TTAAGAGGCA	
		Ala→Val					
11461	CACCCAACAA	G <u>T</u> TCTGATGG	AACTCAGTAC	GAATCCAATG	TTACGTGGCA	TATTCTCTGA	1
11461	CACCCAACAA	GCTCTGATGG	AACTCAGTAC	GAATCCAATG	TTACGTGGCA	TATTCTCTGA	
			Asn				
11521	CAATGCACAG	GCAGAAGAAA	ATAA <u>T</u> CTTGC	TAGGTTTCTC	CTGGATAGGG	AGGTGATCTT	1
11521	CAATGCACAG	GCAGAAGAAA	ATAACCTTGC	TAGGTTTCTC	CTGGATAGGG	AGGTGATCTT	
					Lys		
11581	TCCGCGTGTA	GCTCACATCA	TCATTGAGCA	AACCAGTGTC	GGGAGGAGAA	A <u>G</u> CAGATTCA	1
11581	TCCGCGTGTA	GCTCACATCA	TCATTGAGCA	AACCAGTGTC	GGGAGGAGAA	AACAGATTCA	
				Val		Ile→Val	
11641	AGGATATTTG	GATTCAACTA	GATCGATAAT	GAGGAAATCA	CT <u>G</u> GAAATTA	AGCCCTT <u>G</u> TC	2
11641	AGGATATTTG	GATTCAACTA	GATCGATAAT	GAGGAAATCA	CTAGAAATTA	AGCCCTTATC	
				Leu			
11701	CAATAGGAAG	CTTAATGAAA	TACTGGATTA	CAACATCAAT	TAC <u>T</u> TAGCTT	ACAATTTGGC	1
11701	CAATAGGAAG	CTTAATGAAA	TACTGGATTA	CAACATCAAT	TACCTAGCTT	ACAATTTGGC	

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					Thr		
11761	ATTACTCAAG	AATGCTATTG	AACCTCCGAC	TTATTTGAAG	GCAATGAC <u>C</u> C	TTGAAACATG	1
11761	ATTACTCAAG	AATGCTATTG	AACCTCCGAC	TTATTTGAAG	GCAATGACAC	TTGAAACATG	
			Asn→Ser				
11821	TAGCATCGAC	ATTGCAAGGA	<u>G</u> CCTCCGGAA	GCTCTCCTGG	GCCCCACTCT	TGGGTGGGAG	1
11821	TAGCATCGAC	ATTGCAAGGA	ACCTCCGGAA	GCTCTCCTGG	GCCCCACTCT	TGGGTGGGAG	
		Leu		Ile			
11881	AAATCTTGAA	GGAC <u>T</u> AGAGA	CGCCAGATCC	CATTGAAAT <u>C</u>	ACTGCAGGAG	CATTAATTGT	2
11881	AAATCTTGAA	GGATTAGAGA	CGCCAGATCC	CATTGAAATT	ACTGCAGGAG	CATTAATTGT	
11941	TGGATCGGGC	TACTGTGAAC	AGTGTGCTGC	AGGAGACAAT	CGATTCACAT	GGTTTTTCTT	
11941	TGGATCGGGC	TACTGTGAAC	AGTGTGCTGC	AGGAGACAAT	CGATTCACAT	GGTTTTTCTT	
12001	GCCATCTGGT	ATCGAGATAG	GAGGGGATCC	CCGTGATAAT	CCTCCTATCC	GTGTACCGTA	
12001	GCCATCTGGT	ATCGAGATAG	GAGGGGATCC	CCGTGATAAT	CCTCCTATCC	GTGTACCGTA	
12061	CATTGGCTCC	AGGACTGATG	AGAGGAGGGT	AGCCTCAATG	GCATACATCA	GGGGTGCCTC	
12061	CATTGGCTCC	AGGACTGATG	AGAGGAGGGT	AGCCTCAATG	GCATACATCA	GGGGTGCCTC	
	Ser	Leu					
12121	<u>A</u> AGTAGC <u>C</u> TA	AAAGCAGTTC	TTAGACTGGC	GGGAGTGTAC	ATCTGGGCAT	TCGGAGATAC	2
12121	GAGTAGCTTA	AAAGCAGTTC	TTAGACTGGC	GGGAGTGTAC	ATCTGGGCAT	TCGGAGATAC	
12181	TCTGGAGAAT	TGGATAGATG	CACTGGATTT	GTCTCACACT	AGAGTTAACA	TCACACTTGA	
12181	TCTGGAGAAT	TGGATAGATG	CACTGGATTT	GTCTCACACT	AGAGTTAACA	TCACACTTGA	
		Leu					
12241	ACAG <u>T</u> TACAA	TCCCTCACCC	CACTTCCAAC	CTCTGCCAAT	CTAACCCATC	GGTTGGATGA	2
12241	ACAGCTGCAA	TCCCTCACCC	CACTTCCAAC	CTCTGCCAAT	CTAACCCATC	GGTTGGATGA	
			Ala				
12301	TGGCACAACT	ACCCTAAAGT	TTACTCCTGC	<u>A</u> AGCTCTTAC	ACCTTTTCAA	GTTTCACTCA	1
12301	TGGCACAACT	ACCCTAAAGT	TTACTCCTGC	GAGCTCTTAC	ACCTTTTCAA	GTTTCACTCA	
		Tyr					
12361	TATATCAAAT	GATGAGCAAT	A <u>T</u> CTGACAAT	TAATGACAAA	ACTGCAGATT	CAAATATAAT	1
12361	TATATCAAAT	GATGAGCAAT	ACCTGACAAT	TAATGACAAA	ACTGCAGATT	CAAATATAAT	

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				Gly			
12421	CTACCAACAG	TTAATGATCA	CTGGACTCGG	<u>G</u> ATCTTAGAA	ACATGGAATA	ATCCCCCAAT	1
12421	CTACCAACAG	TTAATGATCA	CTGGACTCGG	AATCTTAGAA	ACATGGAATA	ATCCCCCAAT	
12481	CAATAGAACA	TTCGAAGAAT	CTACCCTACA	TTTGCACACT	GGTGCATCAT	GTTGTGTCCG	
12481	CAATAGAACA	TTCGAAGAAT	CTACCCTACA	TTTGCACACT	GGTGCATCAT	GTTGTGTCCG	
			Leu→Ile				
12541	ACCTGTGGAC	TCCTGCATT <u>A</u>	TCTCAGAAGC	ATTAACAGTC	AAGCCACATA	TTACAGTACC	1
12541	ACCTGTGGAC	TCCTGCATT	TCTCAGAAGC	ATTAACAGTC	AAGCCACATA	TTACAGTACC	
			Glu		Glu		
12601	GTACAGCAAT	AAATTTGTAT	TTGATGA <u>A</u> GA	CCCGCTATCT	GAATATG <u>A</u> GA	CTGCAAAACT	2
12601	GTACAGCAAT	AAATTTGTAT	TTGATGAGGA	CCCGCTATCT	GAATATGAAA	CTGCAAAACT	
12661	GGAATCGTTA	TCATTCCAAG	CCCAATTAGG	CAACATTGAT	GCTGTAGATA	TGACAGGTAA	
12661	GGAATCGTTA	TCATTCCAAG	CCCAATTAGG	CAACATTGAT	GCTGTAGATA	TGACAGGTAA	
12721	ATTAACATTA	TTGTCCCAAT	TCACTGCAAG	GCAGATTATC	AATGCAATCA	CTGGACTCGA	
12721	ATTAACATTA	TTGTCCCAAT	TCACTGCAAG	GCAGATTATC	AATGCAATCA	CTGGACTCGA	
		Val					
12781	TGAGTCTGT <u>T</u>	TCTCTTACTA	ATGATGCCAT	TGTTGCATCA	GACTATGTCT	CCAATTGGAT	1
12781	TGAGTCTGTC	TCTCTTACTA	ATGATGCCAT	TGTTGCATCA	GACTATGTCT	CCAATTGGAT	
12841	TAGTGAATGC	ATGTATACCA	AATTAGATGA	ATTATTTATG	TATTGTGGGT	GGGAACTACT	
12841	TAGTGAATGC	ATGTATACCA	AATTAGATGA	ATTATTTATG	TATTGTGGGT	GGGAACTACT	
12901	ATTGGAAC TA	TCCTATCAAA	TGTATTATCT	GAGGGTAGTT	GGGTGGAGTA	ATATAGTGGA	
12901	ATTGGAAC TA	TCCTATCAAA	TGTATTATCT	GAGGGTAGTT	GGGTGGAGTA	ATATAGTGGA	
12961	TTATTCTTAC	ATGATCTTGA	GAAGAATCCC	GGGTGCAGCA	TTAAACAATC	TGGCATCTAC	
12961	TTATTCTTAC	ATGATCTTGA	GAAGAATCCC	GGGTGCAGCA	TTAAACAATC	TGGCATCTAC	
13021	ATTAAGTCAT	CCAAAAC TTT	TCCGACGAGC	TATCAACCTA	GATATAGTTG	CCCCCTTAAA	
13021	ATTAAGTCAT	CCAAAAC TTT	TCCGACGAGC	TATCAACCTA	GATATAGTTG	CCCCCTTAAA	

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13081 TGCTCCTCAT TTTGCATCTC TGGACTACAT CAAGATGAGT **Val→Met** ATGGATGCAA TACTCTGGGG 1
 ||||| ||||| ||||| ||||| ||||| |||||
 13081 TGCTCCTCAT TTTGCATCTC TGGACTACAT CAAGATGAGT GTGGATGCAA TACTCTGGGG

13141 CTGTAAAAGA GTCATCAATG TGCTCTCCAA TGGAGGGGAC TTAGAATTAG TTGTGACATC
 ||||| ||||| ||||| ||||| ||||| |||||
 13141 CTGTAAAAGA GTCATCAATG TGCTCTCCAA TGGAGGGGAC TTAGAATTAG TTGTGACATC

13201 TGAAGATAGC CTTATTCTCA GTGACCGATC CATGAATCTC ATTGCAAGGA AATTAACTTT
 ||||| ||||| ||||| ||||| ||||| |||||
 13201 TGAAGATAGC CTTATTCTCA GTGACCGATC CATGAATCTC ATTGCAAGGA AATTAACTTT

13261 ATTATCACTG ATTCACCATA ATGGTTTGA ACTACCAAAG ATTAAGGGGT TCTCTCCTGA
 ||||| ||||| ||||| ||||| ||||| |||||
 13261 ATTATCACTG ATTCACCATA ATGGTTTGA ACTACCAAAG ATTAAGGGGT TCTCTCCTGA

13321 TGAGAAGTGT TTCGCTTTGA CAGAAITTTT GAGGAAAGTG GTGAACTCAG GGTGAGTTC
 ||||| ||||| ||||| ||||| ||||| |||||
 13321 TGAGAAGTGT TTCGCTTTGA CAGAAITTTT GAGGAAAGTG GTGAACTCAG GGTGAGTTC

13381 AATAGAGAAC CTATCAAATT TTATGTACAA TGTGGAGAAC CCACGGCTTG CAGCATTCGC
 ||||| ||||| ||||| ||||| ||||| |||||
 13381 AATAGAGAAC CTATCAAATT TTATGTACAA TGTGGAGAAC CCACGGCTTG CAGCATTCGC

13441 CAGCAACAAT TACTACCTGA CCAGAAAATT ATTGAATTCA ATACGAGATA CTGAGTCSerAGG 1
 ||||| ||||| ||||| ||||| ||||| |||||
 13441 CAGCAACAAT TACTACCTGA CCAGAAAATT ATTGAATTCA ATACGAGATA CTGAGTCGGG

13501 TCAAGTAGCA GTCACCTCAT ATTATGAATC ATTAGAATAT ATTGATAGTC TTAAGCTAAC
 ||||| ||||| ||||| ||||| ||||| |||||
 13501 TCAAGTAGCA GTCACCTCAT ATTATGAATC ATTAGAATAT ATTGATAGTC TTAAGCTAAC

13561 CCCACATGTG CCTGGCACCT CATGCATTGA GGATGATAGT CTATGTACAA ATGATTACAT
 ||||| ||||| ||||| ||||| ||||| |||||
 13561 CCCACATGTG CCTGGCACCT CATGCATTGA GGATGATAGT CTATGTACAA ATGATTACAT

13621 AATCTGGATC ATAGAGTCTA ATGCAAACCT GGAGAAGTAT CCAATTCCAA ATAGCCCTGA
 ||||| ||||| ||||| ||||| ||||| |||||
 13621 AATCTGGATC ATAGAGTCTA ATGCAAACCT GGAGAAGTAT CCAATTCCAA ATAGCCCTGA

13681 GGATGATTCC AATTTCCATA ACTTTAAGTT GAATGCTCCA TCGCACCATA CCTTACGCCC
 ||||| ||||| ||||| ||||| ||||| |||||
 13681 GGATGATTCC AATTTCCATA ACTTTAAGTT GAATGCTCCA TCGCACCATA CCTTACGCCC

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Ser

13741 ATTAGGGTTG TCATCGACTG CTTGGTATAA GGGTATAAGC TGCTGCAGGT ACCTTGAGCG 1
 ||||| ||||| ||||| ||||| ||||| ||||| |||||
 13741 ATTAGGGTTG TCATCAACTG CTTGGTATAA GGGTATAAGC TGCTGCAGGT ACCTTGAGCG

13801 ATTAAAGCTA CCACAAGGTG ATCATTATA TATTGCAGAA GGTAGTGGTG CCAGTATGAC
 ||||| ||||| ||||| ||||| ||||| |||||
 13801 ATTAAAGCTA CCACAAGGTG ATCATTATA TATTGCAGAA GGTAGTGGTG CCAGTATGAC

13861 AATCATAGAA TACCTATTCC CAGGAAGAAA GATATATTAC AATTCTTTAT TTAGTAGTGG
 ||||| ||||| ||||| ||||| ||||| |||||
 13861 AATCATAGAA TACCTATTCC CAGGAAGAAA GATATATTAC AATTCTTTAT TTAGTAGTGG

13921 TGACAATCCC CCACAAAGAA ATTATGCACC AATGCCTACT CAGTTCATTG AGAGTGTCCC
 ||||| ||||| ||||| ||||| ||||| |||||
 13921 TGACAATCCC CCACAAAGAA ATTATGCACC AATGCCTACT CAGTTCATTG AGAGTGTCCC

13981 ATACAAGCTC TGGCAAGCAC ACACAGATCA ATATCCCGAG ATTTTGTAGG ACTTCATCCC
 ||||| ||||| ||||| ||||| ||||| |||||
 13981 ATACAAGCTC TGGCAAGCAC ACACAGATCA ATATCCCGAG ATTTTGTAGG ACTTCATCCC

14041 TCTATGGAAC GGAAACGCCG CCATGACTGA CATAGGAATG ACAGCTTGTG TAGAATTCAT
 ||||| ||||| ||||| ||||| ||||| |||||
 14041 TCTATGGAAC GGAAACGCCG CCATGACTGA CATAGGAATG ACAGCTTGTG TAGAATTCAT

Val

14101 CATCAATCGA GTTGGCCCAA GGACTTGCAG TTTAGTACAT GTAGATTGG AATCAAGTGC 1
 ||||| || ||||| ||||| ||||| ||||| |||||
 14101 CATCAATCGA GTCGGCCCAA GGACTTGCAG TTTAGTACAT GTAGATTGG AATCAAGTGC

14161 AAGCTTAAAT CAACAATGCC TGTCAAAGCC GATAATTAAT GCTATCATCA CTGCTACAAC
 ||||| ||||| ||||| ||||| ||||| |||||
 14161 AAGCTTAAAT CAACAATGCC TGTCAAAGCC GATAATTAAT GCTATCATCA CTGCTACAAC

14221 TGTTTGTGC CCTCATGGGG TGCTTATTCT GAAATATAGT TGGTTGCCAT TTAGTAGATT
 ||||| ||||| ||||| ||||| ||||| |||||
 14221 TGTTTGTGC CCTCATGGGG TGCTTATTCT GAAATATAGT TGGTTGCCAT TTAGTAGATT

14281 TAGTACTTTG ATCACTTTCT TATGGTGCTA CTTTGAGAGA ATCACTGTTC TTAGGAGCAC
 ||||| ||||| ||||| ||||| ||||| |||||
 14281 TAGTACTTTG ATCACTTTCT TATGGTGCTA CTTTGAGAGA ATCACTGTTC TTAGGAGCAC

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14341 ATATTCTGAT CCAGCTAATC ATGAGGTTTA TTTAATTGT ATCCTTGCCA ACAACTTTGC
 ||||| ||||| ||||| ||||| ||||| |||||

14341 ATATTCTGAT CCAGCTAATC ATGAGGTTTA TTTAATTGT ATCCTTGCCA ACAACTTTGC

14401 ATTCCAGACT GTCTCGCAGG CAACAGGAAT GGCGATGACT TTAACCGATC AAGGGTTTAC 1
 ||||| ||||| ||||| ||||| ||||| |||||

14401 ATTCCAGACT GTCTCGCAGG CAACAGGAAT GGCGATGACT TTAACCGATC AAGGGTTTAC

14461 **Thr** CTTGATATCA CCTGAAAGAA TAAATCAGTA TTGGGATGGT CACTTAAGC AAGAACGTAT 2
 ||||| ||||| ||||| ||||| ||||| |||||

14461 TTTGATATCA CCTGAAAGAA TAAATCAGTA TTGGGATGGT CACTTGAAGC AAGAACGTAT

14521 CGTAGCAGAA GCAATTGATA AGGTGGTTCT AGGAGAAAAT GCTCTATTTA ATTCGAGTGA
 ||||| ||||| ||||| ||||| ||||| |||||

14521 CGTAGCAGAA GCAATTGATA AGGTGGTTCT AGGAGAAAAT GCTCTATTTA ATTCGAGTGA

14581 TAATGAATTA ATTCTCAAAT GTGGAGGGAC ACCAAATGCA CGGAATCTTA TCGATATCGA 1
 ||||| ||||| ||||| ||||| ||||| |||||

14581 TAATGAATTA ATTCTCAAAT GTGGAGGGAC ACCAAATGCA CGGAATCTCA TCGATATCGA

14641 GCCAGTCGCA ACTTTCATAG AATTGGAACA **Leu** ACTAATCTGC ACAATGTTGA CAACCCACTT 2
 ||||| ||||| ||||| ||||| ||||| |||||

14641 GCCAGTCGCA ACTTTCATAG AATTGGAACA ATTGATCTGC ACAATGTTGA CAACCCACTT

14701 GAAGGAAATA ATTGATATAA CAAGGTCTGG AACCAGGAT TATGAAAGTT TATTACTCAC
 ||||| ||||| ||||| ||||| ||||| |||||

14701 GAAGGAAATA ATTGATATAA CAAGGTCTGG AACCAGGAT TATGAAAGTT TATTACTCAC

14761 TCCTTACAAT TTAGGTCTTC TTGGTAAAT CAGTACGATA GTGAGATTAT TAACAGAAAG
 ||||| ||||| ||||| ||||| ||||| |||||

14761 TCCTTACAAT TTAGGTCTTC TTGGTAAAT CAGTACGATA GTGAGATTAT TAACAGAAAG

14821 GATTCTAAAT CATACTATCA GGAATTGGTT GATCCTCCCA CCTTCGCTCC AGATGATCGT
 ||||| ||||| ||||| ||||| ||||| |||||

14821 GATTCTAAAT CATACTATCA GGAATTGGTT GATCCTCCCA CCTTCGCTCC GGATGATCGT

14881 GAAGCAGGAC TTGAATTCG GCATATTCAG GATTACTTCC ATCCTCAATT CTGATCGGTT
 ||||| ||||| ||||| ||||| ||||| |||||

14881 GAAGCAGGAC TTGAATTCG GCATATTCAG GATTACTTCC ATCCTCAATT CTGATCGGTT

14941 **Lys** CCTGAACTT TCTCCAAATA GGAAATACTT **Ala→Thr** GATTACACAA TTAAGTGCAG GCTACATTAG 2
 ||||| ||||| ||||| ||||| ||||| |||||

14941 CCTGAAGCTT TCTCCAAATA GGAAATACTT GATTGCACAA TTAAGTGCAG GCTACATTAG

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Cys

Tyr

L Protein Stop ←

L mRNA End ←

15241 CTTGGT // (SEQ ID No:2)

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CV-0001	ATGAGCACTA	TAATTCAATC	TCTGGTGGTC	TCCTGTCTAT	TGGCAGGAGC	AGGCAGCCTT	
CPV-0001	ATGGG <u>T</u> ACTA	<u>G</u> AATTCAAT <u>T</u>	TCTGGTGGTC	TCCTGTCTAT	TGGCAGGA <u>A</u> C	AGGCAGCCTT	5
PR-0001	ATGGG <u>T</u> ACTA	TAATTCAAT <u>T</u>	TCTGGTGGTC	TCCTGTCTAT	TGGCAGGAGC	AGGCAGCCTT	3
W3A-0001	ATGGG <u>T</u> ACTA	TAATTCAAT <u>T</u>	TCTGGTGGTC	TCCTGTCTAT	TGGCAGGAGC	AGGCAGCCTT	3
WR-0001	ATGGG <u>T</u> ACTA	<u>T</u> AATTCAAT <u>T</u>	TCTGGTGGTC	TCCTGTCTAT	TGGCAGGAGC	AGGCAGCCTT	4
CV-0061	GATCCAGCAG	CCCTCATGCA	AATCGGTGTC	ATTCCAACAA	ATGTCCGGCA	ACTTATGTAT	
CPV-0061	GATCCAGCAG	CCCTCATGCA	AATCGGTGTC	ATTCCAACAA	ATGTCCGGCA	ACTTATGTAT	
PR-0061	GATCCAGCAG	CCCTCATGCA	AATCGGTGTC	ATTCCAACAA	ATGTCCGGCA	ACTTATGTAT	
W3A-0061	GATCCAGCAG	CCCTCATGCA	AATCGGTGTC	ATTCCAACAA	ATGTCCGGCA	ACTTATGTAT	
WR-0061	GATC <u>T</u> AGCAG	CCCTCATGCA	AATCGGTGTC	ATTCCAACAA	ATGTCCGGCA	ACTTATGTAT	1
CV-0121	TATACTGAGG	CCTCATCAGC	ATTCATTGTT	GTGAAGTTAA	TGCCTACAAT	TGACTCGCCG	
CPV-0121	TATACTGAGG	<u>C</u> TTCATCAGC	ATTCATTGTT	GTGAAGTTAA	TGCCTACAAT	TGACTCGCCG	1
PR-0121	TATACTGAGG	CCTCATCAGC	ATTCATTGTT	GTGAAGTTAA	TGCCTACAAT	TGACTCGCCG	
W3A-0121	TATACTGAGG	CCTCATCAGC	ATTCATTGTT	GTGAAGTTAA	TGCCTACAAT	TGACTCGCCG	
WR-0121	TATACTGAGG	CCTCATC <u>G</u> GC	ATTCATTGTT	GTGAAGTTAA	TGCCTACAAT	TGACTCGCCG	1
CV-0181	ATTAGTGGAT	GTAATATAAC	ATCAATTTCA	AGCTATAATG	CAACAGTGAC	AAAACTCCTA	
CPV-0181	ATTAGTGG <u>G</u> T	GTAATATAAC	ATC <u>C</u> AATTTCA	AGCTATAATG	CAACA <u>A</u> TGAC	AAAAC <u>T</u> CTA	4
PR-0181	ATTAGTGGAT	GTAATATAAC	ATCAATTTCA	AGCTATAATG	CAACAGTGAC	AAAACTCCTA	
W3A-0181	ATTAGTGGAT	GTAATATAAC	ATCAATTTCA	AGCTATAATG	CAACAGTGAC	AAAACTCCTA	
WR-0181	ATTAGTGGAT	GTAATATAAC	ATCAATTTCA	AGCTATAATG	CAACAGTGAC	AAAACTCCTA	

Figure 10A
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CV-0241	CAGCCGATCG	GTGAGAATTT	GGAGACGATT	AGGAACCAGT	TGATTCCAAC	TCGGAGGAGA	
CPV-0241	CAGCCGATCG	GTGAGAATTT	<u>AGAGACGATT</u>	AGG <u>T</u> ACCAGT	TGATTCCAAC	TCGGAGGAGA	2
PR-0241	CAGCCGATCG	GTGAGAATTT	GGAA <u>A</u> CGATT	AGGAACCAGT	TGATTCCAAC	TCGGAGGAGA	1
W3A-0241	CAGCCGATCG	GTGAGAATTT	GGAGACGATT	AGGAACCAGT	TGATTCCAAC	TCGGAGGAGA	
WR-0241	CAGCCGATCG	GTGAGAATTT	GGAGACGATT	AGGAACCAGT	TGATTCCAAC	TCGGAG <u>A</u> AGA	1
CV-0301	CGCCGGTTTG	CAGGGGTGGT	GATTGGATTA	GCTGCATTAG	GAGTAGCTAC	TGCCGCACAA	
CPV-0301	CGCCGGTTTG	CAGGGGTGGT	GATTGGATTA	GC <u>C</u> GCATTAG	GAGTAGCTAC	TGC <u>A</u> GCACAG	3
PR-0301	CGCCGGTTTG	CAGGGGTGGT	GATTGGATTA	GCTGCATTAG	GAGTAGCTAC	TGCCGCACAG	1
W3A-0301	CGCCGGTTTG	CAGGGGTGGT	GATTGGATTA	GCTGCATTAG	GAGTAGCTAC	TGCCGCACAG	1
WR-0301	CGCCGGTTTG	CAGGGGTGGT	GATTGGATTA	GCTGCATTAG	GAGTAGCTAC	TGCCGCACAG	1
CV-0361	GTCAGTCCG	CAGTAGCACT	AGTTAAGGCA	AATGAAAATA	CTGCGGCTAT	ACTCAATCTC	
CPV-0361	GTCAGTCCG	CAGTAGCACT	AGT <u>AA</u> AGGCG	AAT <u>AAAAATG</u>	CTG <u>T</u> GGCTAT	ACTCAATCTC	5
PR 0361	GTCAGTCCG	CAGTAGCACT	AGT <u>AA</u> AGGCA	AAT <u>AAAAATG</u>	CTGCGGCTAT	ACTCAATCTC	3
W3A-0361	GTCAGTCCG	CAGT <u>G</u> GCACT	AGT <u>AA</u> AGGCA	AATGAAAAT <u>G</u>	CTGCGGCTAT	ACTCAATCTC	3
WR-0361	GTCAGTCCG	CAGTAGCACT	AGT <u>AA</u> AGGCA	AATGAAAAT <u>G</u>	CTGCGGCTAT	ACTCAATCTC	2
CV-0421	AAAAATGCAA	TCCAAAAAAC	AAATGCAGCA	GTTGCAGATG	TGGTCCAGGC	CACACAATCA	
CPV-0421	AAAAA <u>C</u> GCAA	TCCAAAAAAC	AAATGCAGCA	GTTGCAGAC <u>G</u>	TGGT <u>T</u> CAGGC	CACACAATCA	3
PR-0421	AAAAATGCAA	TCCAAAAAAC	AAAT <u>A</u> CAGCA	GTTGCAGATG	TGGTCCAGGC	CACACAATCA	1
W3A-0421	AAAAATGCAA	TCCAAAAAAC	AAATGCAGCA	GTTGCAGATG	TGGTCCAGGC	CACACAATCA	
WR-0421	AAAAATGCAA	TCCAAAAAAC	AAATGCAGCA	GTTGCAGATG	TGGTCCAGGC	CACACAATCA	
CV-0481	CTAGGAACGG	CAGTTCAAGC	AGTTCAAGAT	CACATAAACA	GTGTGATAAG	TCCAGCAATT	
CPV-0481	CTAGGAACGG	CAGTTCAAGC	AGTTCAAGAT	CACATAAA <u>TA</u>	GTGTG <u>G</u> TAAG	TCCAGCAATT	2
PR-0481	CTAGGAACGG	CAGTTCAAGC	AGTTCAAGAT	CACATAAACA	GTGTG <u>G</u> TAAG	TCCAGCAATT	1
W3A-0481	CTAGGAACGG	CAGTTCAAGC	AGTTCAAGAT	CACATAAACA	GTGTG <u>G</u> TAAG	TCCAGCAATT	1
WR-0481	CTAGGAACGG	CAGTTCAAGC	AGTTCAAGAT	CACATAAACA	GTGTG <u>G</u> TAAG	TCCAGCAATT	1

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CV-0541	ACAGCAGCCA	ATTGTAAGGC	CCAAGATGCT	ATCATTGGCT	CAATCCTCAA	TCTCTATTTG	
CPV-0541	ACAGCAGCCA	ATTG <u>C</u> AAAGC	CCAAGATGCT	ATCATTGGCT	CAAT <u>T</u> CTCAA	TCTCTATTTG	3
PR-0541	ACAGCAGCCA	ATTGTAAGGC	CCAAGATGCT	ATCATTGGCT	CAATCCTCAA	TCTCTATTTG	
W3A-0541	ACAGCAGCCA	ATTGTAAGGC	CCAAGATGCT	ATCATTGGCT	CAATCCTCAA	TCTCTATTTG	
WR-0541	ACAGCAGCCA	ATTGTAAGGC	CCAAGATGCT	ATCATTGGCT	CAATCCTCAA	TCTCTATTTG	
CV-0601	ACCGAGTTGA	CAACTATCTT	CCACAATCAA	ATTACAAACC	CTGCATTGAG	TCCTATTACA	
CPV-0601	ACCGAGTTGA	CAACTATCTT	CCACAATCAA	ATTACAAACC	<u>C</u> CGCATTGAG	TCCTATTACA	1
PR-0601	ACCGAGTTGA	CAACTATCTT	CCACAATCAA	ATTACAAACC	CTGCATTGAG	TCCTATTACA	
W3A-0601	ACCGAGTTGA	CAAC <u>C</u> ATCTT	CCACAATCAA	ATTACAAACC	CTGCATTGAG	TCC <u>C</u> ATTACA	2
WR-0601	ACCGAGTTGA	CAAC <u>C</u> ATCTT	CCACAATCAA	ATTACAAACC	CTGCATTGAG	TCC <u>C</u> ATTACA	2
CV-0661	ATTCAAGCTT	TAAGGATCCT	ACTGGGGAGT	ACCTTGCCGA	CTGTGGTCGA	AAAATCTTTC	
CPV-0661	ATTCAAGCTT	T <u>G</u> AGGATCCT	ACT <u>A</u> GGGAGT	ACCTTGCCGA	<u>C</u> CGTGGTCGA	AAAATCTTTC	3
PR-0661	ATTCAAGCTT	TAAGGATCCT	ACTGGGGAGT	ACCTTGCCGA	CTGTGGTCGA	AAAATCTTTC	
W3A-0661	ATTCAAGCTT	TAAGGATCCT	ACTGGGGAGT	ACCTTGCCGA	CTGTGGTCGA	AAAATCTTTC	
WR-0661	ATTCAAGCTT	TAAGGATCCT	ACTGGGGAGT	ACCTTGCCGA	CTGTGGTCGA	AAAATCTTTC	
CV-0721	AATACCCAGA	TAAGTGCAGC	TGAGCTTCTC	TCATCAGGGT	TGTTGACAGG	CCAGATTGTG	
CPV-0721	AATACCCAGA	TAAGTGCAGC	TGAGCTTCTC	TCATCAGGGT	<u>T</u> ATTGACAGG	CCAGATTGTG	1
PR-0721	AATACCCAGA	TAAGTGCAGC	TGAGCTTCTC	TCATCAGGGT	<u>T</u> ATTGACAGG	CCAGATTGTG	1
W3A-0721	AATACCCAGA	TAAGTGCAGC	TGAGCTTCTC	TCATCAGGGT	<u>T</u> ATTGACAGG	CCAGATTGTG	1
WR-0721	AATACCCAGA	TAAGTGCAGC	TGAGCTTCTC	TCATCAGGGT	<u>T</u> ATTGACAGG	CCAGATTGTG	1
CV-0781	GGATTAGATT	TGACCTATAT	GCAGATGGTC	ATAAAAAATTG	AGCTGCCAAC	TTTAACTGTA	
CPV-0781	GGATTAGATT	TGACCTA <u>C</u> AT	GCAGATGGTC	ATAAAAAATTG	AGCTGCCAAC	TTTAACTGTA	1
PR-0781	GGATTAGATT	TGACCTATAT	GCAGATGGTC	ATAAAAAATTG	AGCTGCCAAC	TTTAACTGTA	
W3A-0781	GGATTAGATT	TGACCTATAT	GCAGATGGTC	ATAAAAAATTG	AGCTGCCAAC	TTTAACTGTA	
WR-0781	GGATTAGATT	TGACCTATAT	GCAGATGGTC	ATAAAAAATTG	AGCTGCCAAC	TTTAACTGTA	

CV-0841	CAACCTGCAA	CCCAGATCAT	AGATCTGGCC	ACCATTTCTG	CATTCATTAA	CAATCAAGAA	
CPV-0841	CAACCTGCAA	CCCAGATCAT	AGATCTGG <u>T</u> C	ACCATTTCTG	CATTCATTAA	CAATCAAGAA	1
PR-0841	CAACCTGCAA	CCCAGATCAT	AGATCTGGCC	ACCATTTCTG	CATTCATTAA	CAATCAAGAA	
W3A-0841	CAACCTGCAA	CCCAGATCAT	AGATCTGGCC	ACCATTTCTG	CATTCATTAA	CAATCAAGAA	
WR-0841	CAACCTGCAA	CCCAGATCAT	AGATCTGGCC	ACCATTTCTG	CATTCATTAA	CAATCAAGAA	
CV-0901	GTCATGGCCC	AATTACCAAC	ACGTGTTATT	GTGACTGGCA	GCTTGATCCA	AGCCTATCCC	
CPV-0901	GT <u>T</u> ATGGCCC	AATTACCAAC	ACGTGTTATT	GTGACTGGCA	GCTTGATCCA	AGCCTATCCC	1
PR-0901	GTCATGGCCC	AATTACCAAC	ACGTGTTATT	GTGACTGGCA	GCTTGATCCA	AGCCTATCCC	
W3A-0901	GTCATGGCCC	AATTACCAAC	ACGTGTTAT <u>G</u>	GTGACTGGCA	GCTTGATCCA	AGCCTATCCC	1
WR-0901	GTCATGGCCC	AATTACCAAC	ACGTGTTAT <u>G</u>	GTGACTGGCA	GCTTGATCCA	AGCCTATCCC	1
CV-0961	GCATCGCAAT	GCACTATTAC	ACCCAACACT	GTGTAAGTGA	GGTATAATGA	TGCCCAAGTA	
CPV-0961	GCATCGCAAT	GCACTAT <u>C</u> AC	<u>C</u> CCCAACACT	GTGTAAGTGA	GGTATAATGA	TGCCCAAGTA	2
PR-0961	GCATCGCAAT	GCACTATTAC	ACCCAACACT	GTGTAAGTGA	GGTATAATGA	TGCCCAAGTA	
W3A-0961	GCATCGCAAT	GCAC <u>C</u> ATTAC	ACCCAACACT	GTGTAAGTGA	GGTATAATGA	TGCCCAAGTA	1
WR-0961	GCATCGCAAT	GCAC <u>C</u> ATTAC	ACCCAACACT	GTGTAAGTGA	GGTATAATGA	TGCCCAAGTA	1
CV-1021	CTCTCAGATG	ATACGATGGC	TTGCCTCCAA	GGTAACTTGA	CAAGATGCAC	CTTCTCTCCA	
CPV-1021	CTCTCAGATG	ATACGATGGC	TTGCCTCCAA	GGTAACTTGA	CAAGATGCAC	CTTCTCTCCA	
PR-1021	CTCTCAGATG	ATACGATGGC	TTGCCTCCAA	GGTAACTTGA	CAAGATGCAC	CTTCTCTCC <u>G</u>	1
W3A-1021	CTCTCAGATG	ATAC <u>T</u> ATGGC	TTGCCTCCAA	GGTAACTTGA	CAAGATGCAC	CTTCTCTCCA	1
WR-1021	CTCTCAGATG	ATAC <u>T</u> ATGGC	TTGCCTCCAA	GGTAACTTGA	CAAGATGCAC	CTTCTCTCCA	1
CV-1081	GTGGTTGGGA	GCTTTCTCAC	TCGATTCGTG	CTGTTTCGATG	GAATAGTTTA	TGCAAATTGC	
CPV-1081	GTGGTTGGGA	GCTTTCTCAC	TCGATTCGTG	CTGTT <u>T</u> GATG	GAATAGTTTA	TGCAAATTGC	1
PR-1081	GTGGTTGGGA	GCTTTCTCAC	TCGATTCATG	CTGTTTCGATG	GAATAGTTTA	TGCAAATTGC	1
W3A-1081	GTGGTTGGGA	GCTTTCTCAC	TCGATTCGTG	CTGTTTCGATG	GAATAGTTTA	TGCAAATTGC	
WR-1081	GTGGTTGGGA	GCTTTCTCAC	TCGATTCGTG	CTGTTTCGATG	GAATAGTTTA	TGCAAATTGC	

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CV-1141	AGGTCGATGC	TGTGCAAGTG	CATGCAACCT	GCTGCCGTGA	TCCTACAGCC	GAGTTCATCC	
CPV-1141	AGGTCGAT <u>G</u> T	TGTGCAAGTG	CATGCA <u>G</u> CCT	GCTGC <u>T</u> GT <u>T</u> A	TCCTACAGCC	GAG <u>C</u> TCATCC	5
PR-1141	AGGTCGAT <u>G</u> T	<u>T</u> ATGCAAGTG	CATGCA <u>G</u> CCT	GCTGC <u>T</u> GTGA	TCCTACAGCC	GAGTTCATCC	4
W3A-1141	AGGTCGAT <u>G</u> T	TGTGCAAGTG	CATGCAACCT	GCTGC <u>T</u> GTGA	TCCTACAGCC	GAGTTCATCC	2
WR-1141	AGGTCGAT <u>G</u> T	TGTGCAAGTG	CATGCAACCT	GCTGC <u>T</u> GTGA	TCCTACAGCC	GAGTTCATCC	2
CV-1201	CCTGTAACTG	TCATTGACAT	GCACAAATGT	GTGAGTCTGC	AGCTTGACGA	TCTCAGATTC	
CPV-1201	CCTGTAACTG	TCATTGACAT	<u>G</u> TACAAATGT	GTGAGTCTGC	AGCTTGAC <u>A</u> A	TCTCAGATTC	2
PR-1201	CCTGTAACTG	TCATTGACAT	<u>G</u> TACAAATGT	GTGAGTCTGC	AGCTTGAC <u>A</u> A	TCTCAGATTC	2
W3A-1201	CCTGTAACTG	TCATTGACAT	<u>G</u> TACAAATGT	GTGAGTCTGC	AGCTTGAC <u>A</u> A	TCTCAGATTC	2
WR-1201	CCTGTAACTG	TCATTGACAT	<u>G</u> TACAAATGT	GTGAGTCTGC	AGCTTGAC <u>A</u> A	TCTCAGATTC	2
CV-1261	ACCATCACTC	AATTGGCCAA	TGTAACCTAC	AATAGCACCA	TCAAGCTTGA	AACATCCCAG	
CPV-1261	ACCATCACTC	AATTGGCCAA	<u>T</u> ATAACCTAC	AATAGCACCA	TCAAGCTTGA	AACATCCCAG	1
PR-1261	ACCATCACTC	AATTGGCCAA	TGTAACCTAC	AATAGCACCA	TCAAGCTTGA	AACATCCCAG	
W3A-1261	ACCATCACTC	AATTGGCCAA	TGTAACCTAC	AATAGCACCA	TCAAGCTTGA	<u>A</u> TATCCCAG	1
WR-1261	ACCATCACTC	AATTGGCCAA	TGTAACCTAC	AATAGCACCA	TCAAGCTTGA	<u>A</u> TATCCCAG	1
CV-1321	ATCTTGCCTA	TTGATCCGTT	GGATATATCC	CAGAATTTAG	CTGCGGTGAA	TAAGAGTCTA	
CPV-1321	ATCTTGCCTA	<u>T</u> CGATCCGTT	GGATATATCC	CAGAAT <u>C</u> TAG	CTGCGGTGAA	TAAGAGTCTA	2
PR-1321	ATCTTGCCTA	TTGATCCGTT	GGATATATCC	CAGAAT <u>C</u> TAG	CTGCGGTGAA	TAAGAGTCTA	1
W3A-1321	ATCTT <u>G</u> TCTA	TTGATCCGTT	GGATATATCC	<u>C</u> AAAT <u>C</u> TAG	CTGCGGTGAA	TAAGAGTCTA	3
WR-1321	ATCTTGCCTA	TTGATCCGTT	GGATATATCC	CAGAAT <u>C</u> TAG	CTGCGGTGAA	TAAGAGTCTA	1
CV-1381	AGTGATGCAC	TACAACACTT	AGCACAAAGT	GACACATACC	TTTCTGCAAT	CACATCAGCT	
CPV-1381	AGTGATGCAC	TACAACACTT	AGCACAAAGT	GACACATACC	TTTCTGCAAT	CACATCAGCT	
PR-1381	AGTGATGCAC	TACAACACTT	AGCACAAAGT	GACACATACC	TTTCTGCAAT	CACATCAGCT	
W3A-1381	AGTGATGCAC	TACAACACTT	AGCACAAAGT	GACACATAT <u>C</u>	TTTCTGCAAT	CACATCAGCT	1
WR-1381	AGTGATGCAC	TACAACACTT	AGCACAAAGT	GACACATAT <u>C</u>	TTTCTGCAAT	CACATCAGCT	1

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CV-1441	ACGACTACAA	GTGTATTATC	CATAATAGCA	ATCTGTCTTG	GATCGTTAGG	TTTAATATTA	
CPV-1441	ACGACTACAA	GTGTATTATC	CATAATAGCA	ATCTGTCTTG	GATCGTTAGG	TTTAATATTA	
PR-1441	ACGACTACAA	GTGTATTATC	CATAAT <u>G</u> GCA	ATCTGTCTTG	GATCGTTAGG	TTTAATATTA	1
W3A-1441	ACGACTACAA	GTGTATTATC	CATAATAGCA	ATCTGTCTTG	GATCGTTAGG	TTTAATATTA	
WR-1441	ACGACTACAA	GTGTATTATC	CATAATAGCA	ATCTGTCTTG	GATCGTTAGG	TTTAATATTA	
CV-1501	ATAATCTTGC	TCAGTGTAGT	TGTGTGGAAG	TTATTGACCA	TTGTCCGCTGC	TAATCGAAAT	
CPV-1501	ATAATCTTGC	TCAGTGTAGT	TGTGTGGAAG	TTATTGACCA	TTGT <u>T</u> GCTGC	TAATCGAAAT	1
PR-1501	ATAATCTTGC	TCAGTGTAGT	TGTGTGGAAG	TTATTGACCA	TTGT <u>C</u> ACTGC	TAATCGAAAT	1
W3A-1501	ATAATCTTGC	TCAGTGTAGT	TGTGTGGAAG	TTATTGACCA	TTGTCCG <u>T</u> TGC	TAATCGAAAT	1
WR-1501	ATAATCTTGC	TCAGTGTAGT	TGTGTGGAAG	TTATTGACCA	TTGTCCGCTGC	TAATCGAAAT	
CV-1561	AGAATGGAGA	ATTTTGTTTA	TCATAATTCA	GCATTCCACC	ACCCACGATC	TGATCTCAGT	
CPV-1561	AGAATGGAGA	ATTTTGTTTA	TCATAATTCA	GCATTCCACC	AC <u>T</u> CACG <u>G</u> TC	TGATCTCAGT	2
PR-1561	AGAATGCAGA	ATTTTGTTTA	TCATAATTCA	GCATTCCACC	AC <u>T</u> CACGATC	TGATCTCAGT	1
W3A-1561	AGAATGGAGA	ATTTTGTTTA	TCATAA <u>ATAA</u>	GCATTCCACC	AC <u>T</u> CACGATC	TGATCTCAGT	3
WR-1561	AGAATGGAGA	ATTTTGTTTA	TCATAA <u>ATAA</u>	GCATTCCACC	AC <u>T</u> CACGATC	TGATCTCAGT	3

Termination Codon for W3A and WR (TAA) **J**

CV-1621	GAGAAAAATC	AACCTGCAAC	TCTTGGAACA	AGATAA	// (SEQ ID NO:49)
CPV-1621	GAGAAAAATC	AACCTGCAAC	TCTTGGAACA	AGATAA	// (SEQ ID NO:25)
PR-1621	GAGAAAAATC	AACCTGCAAC	TCTTGGAACA	AGATAA	// (SEQ ID NO:27)
W3A-1621	GAGAAAAATC	AACCTGCAAC	TCTTGGAACA	AGATAA	// (SEQ ID NO:29)
WR-1621	GAGAAAAATC	AACCTGCAAC	TCTTGGAACA	AGATAA	// (SEQ ID NO:31)

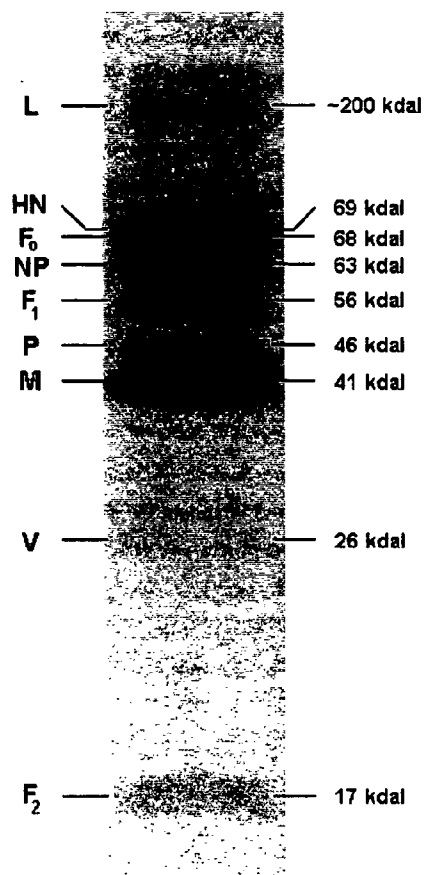
Termination Codon for CV, PR and CPV (TAA) **J**

CV	001	MSTIIQSLVV	SCLLAGAGSL	DPAALMQIGV	IPTNVRQLMY	YTEASSAFIV	VKLMPTIDSP	
CPV	001	<u>M</u> G <u>T</u> R <u>I</u> Q <u>F</u> LVV	SCLLAG <u>T</u> GSL	DPAALMQIGV	IPTNVRQLMY	YTEASSAFIV	VKLMPTIDSP	4
PR	001	<u>M</u> G <u>T</u> T <u>I</u> Q <u>F</u> LVV	SCLLAGAGSL	DPAALMQIGV	IPTNVRQLMY	YTEASSAFIV	VKLMPTIDSP	2
W3A	001	<u>M</u> G <u>T</u> T <u>I</u> Q <u>F</u> LVV	SCLLAGAGSL	DPAALMQIGV	IPTNVRQLMY	YTEASSAFIV	VKLMPTIDSP	2
WR	001	<u>M</u> G <u>T</u> T <u>I</u> Q <u>F</u> LVV	SCLLAGAGSL	<u>D</u> L <u>A</u> ALMQIGV	IPTNVRQLMY	YTEASSAFIV	VKLMPTIDSP	3
CV	061	ISGCNITSIS	SYNATVTKLL	QPIGENLETI	RNQLIPTRRR	RRFAGVVIGL	AALGVATAAQ	
CPV	061	ISGCNITSIS	SYNAT <u>M</u> TKLL	QPIGENLETI	<u>R</u> YQLIPTRRR	RRFAGVVIGL	AALGVATAAQ	2
PR	061	ISGCNITSIS	SYNATVTKLL	QPIGENLETI	RNQLIPTRRR	RRFAGVVIGL	AALGVATAAQ	
W3A	061	ISGCNITSIS	SYNATVTKLL	QPIGENLETI	RNQLIPTRRR	RRFAGVVIGL	AALGVATAAQ	
WR	061	ISGCNITSIS	SYNATVTKLL	QPIGENLETI	RNQLIPTRRR	RRFAGVVIGL	AALGVATAAQ	
CV	121	VTA A VALVKA	NENTAAILNL	KNAIQKTNA A	VADV V QATQS	LGTAVQAVQD	HINSVISPAI	
CPV	121	VTA A VALVKA	<u>N</u> KNA V AILNL	KNAIQKTNA A	VADV V QATQS	LGTAVQAVQD	HINSV <u>V</u> SPAI	4
PR	121	VTA A VALVKA	<u>N</u> KNA A AILNL	KNAIQKTNA T A	VADV V QATQS	LGTAVQAVQD	HINSV <u>V</u> SPAI	3
W3A	121	VTA A VALVKA	<u>N</u> EN A AAAILNL	KNAIQKTNA A	VADV V QATQS	LGTAVQAVQD	HINSV <u>V</u> SPAI	2
WR	121	VTA A VALVKA	<u>N</u> EN A AAAILNL	KNAIQKTNA A	VADV V QATQS	LGTAVQAVQD	HINSV <u>V</u> SPAI	2
CV	181	TAANCKAQDA	IIGSILNLYL	TELTTIFHNQ	ITNPALSPIT	IQALRILLGS	TLPTVVEKSF	
CPV	181	TAANCKAQDA	IIGSILNLYL	TELTTIFHNQ	ITNPALSPIT	IQALRILLGS	TLPTVVEKSF	
PR	181	TAANCKAQDA	IIGSILNLYL	TELTTIFHNQ	ITNPALSPIT	IQALRILLGS	TLPTVVEKSF	
W3A	181	TAANCKAQDA	IIGSILNLYL	TELTTIFHNQ	ITNPALSPIT	IQALRILLGS	TLPTVVEKSF	
WR	181	TAANCKAQDA	IIGSILNLYL	TELTTIFHNQ	ITNPALSPIT	IQALRILLGS	TLPTVVEKSF	
CV	241	NTQISAAELL	SSGLLTGQIV	GLDLTYMQMV	IKIELPTLTV	QPATQIIDLA	TISAFINNQE	
CPV	241	NTQISAAELL	SSGLLTGQIV	GLDLTYMQMV	IKIELPTLTV	QPATQIID <u>L</u> V	TISAFINNQE	1
PR	241	NTQISAAELL	SSGLLTGQIV	GLDLTYMQMV	IKIELPTLTV	QPATQIIDLA	TISAFINNQE	
W3A	241	NTQISAAELL	SSGLLTGQIV	GLDLTYMQMV	IKIELPTLTV	QPATQIIDLA	TISAFINNQE	
WR	241	NTQISAAELL	SSGLLTGQIV	GLDLTYMQMV	IKIELPTLTV	QPATQIIDLA	TISAFINNQE	
CV	301	VMAQLPTRVI	VTGSLIQAYP	ASQCTITPNT	VYCRYNDAQV	LSDDTMACLQ	GNLTRCTFSP	
CPV	301	VMAQLPTRVI	VTGSLIQAYP	ASQCTITPNT	VYCRYNDAQV	LSDDTMACLQ	GNLTRCTFSP	
PR	301	VMAQLPTRVI	VTGSLIQAYP	ASQCTITPNT	VYCRYNDAQV	LSDDTMACLQ	GNLTRCTFSP	
W3A	301	VMAQLPTRV <u>M</u>	VTGSLIQAYP	ASQCTITPNT	VYCRYNDAQV	LSDDTMACLQ	GNLTRCTFSP	1
WR	301	VMAQLPTRV <u>M</u>	VTGSLIQAYP	ASQCTITPNT	VYCRYNDAQV	LSDDTMACLQ	GNLTRCTFSP	1
CV	361	VVG S FLTRFV	LFDGIVYANC	RSMLCKCMQP	AAVILQPSSS	PVTVIDMHKC	VSLQLDDLRF	
CPV	361	VVG S FLTRFV	LFDGIVYANC	RSMLCKCMQP	AAVILQPSSS	PVTVID <u>M</u> YKC	VSLQLD <u>N</u> LRF	2
PR	361	VVG S FLTR <u>F</u> M	LFDGIVYANC	RSMLCKCMQP	AAVILQPSSS	PVTVID <u>M</u> YKC	VSLQLD <u>N</u> LRF	3
W3A	361	VVG S FLTRFV	LFDGIVYANC	RSMLCKCMQP	AAVILQPSSS	PVTVID <u>M</u> YKC	VSLQLD <u>N</u> LRF	2
WR	361	VVG S FLTRFV	LFDGIVYANC	RSMLCKCMQP	AAVILQPSSS	PVTVID <u>M</u> YKC	VSLQLD <u>N</u> LRF	2

Figure 10B
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CV	421	TITQLANVTY	NSTIKLETSQ	ILPIDPLDIS	QNLAAVNKSL	SDALQHLAQS	DTYLSAITS	
CPV	421	TITQLAN <u>I</u> TY	NSTIKLETSQ	ILPIDPLDIS	QNLAAVNKSL	SDALQHLAQS	DTYLSAITS	1
PR	421	TITQLANVTY	NSTIKLETSQ	ILPIDPLDIS	QNLAAVNKSL	SDALQHLAQS	DTYLSAITS	
W3A	421	TITQLANVTY	NSTIKLE <u>SS</u> Q	IL <u>S</u> IDPLDIS	QNLAAVNKSL	SDALQHLAQS	DTYLSAITS	2
WR	421	TITQLANVTY	NSTIKLE <u>SS</u> Q	ILPIDPLDIS	QNLAAVNKSL	SDALQHLAQS	DTYLSAITS	1
CV	481	TTTSVLSIIA	ICLGSGLLIL	IILLSV V VWK	LLTIVAANRN	RMENFVYHNS	AFHHPRSDLS	
CPV	481	TTTSVLSIIA	ICLGSGLLIL	IILLSV V VWK	LLTIVAANRN	RMENFVYHNS	AFHH <u>S</u> RS S DLS	1
PR	481	TTTSVLSI <u>M</u> A	ICLGSGLLIL	IILLSV V VWK	LLTIV <u>T</u> ANRN	RMENFVYHNS	AFHH <u>S</u> RS S DLS	3
W3A	481	TTTSVLSIIA	ICLGSGLLIL	IILLSV V VWK	LLTIV <u>V</u> ANRN	RMENFVYH <u>K</u>	← 529 AA	3
WR	481	TTTSVLSIIA	ICLGSGLLIL	IILLSV V VWK	LLTIVAANRN	RMENFVYH <u>K</u>	← 529 AA	2
CV	541	EKNQPATLGT	R	← 551 AA	<div style="display: flex; align-items: center;"> <div style="margin-right: 20px;"> (SEQ ID No:12) (SEQ ID No:26) (SEQ ID No:28) </div> <div style="margin-left: 20px;"> (SEQ ID No:30) (SEQ ID No:32) </div> </div>			
CPV	541	EKNQPATLGT	R	← 551 AA				
PR	541	EKNQPATLGT	R	← 551 AA				

*Fig. 11*

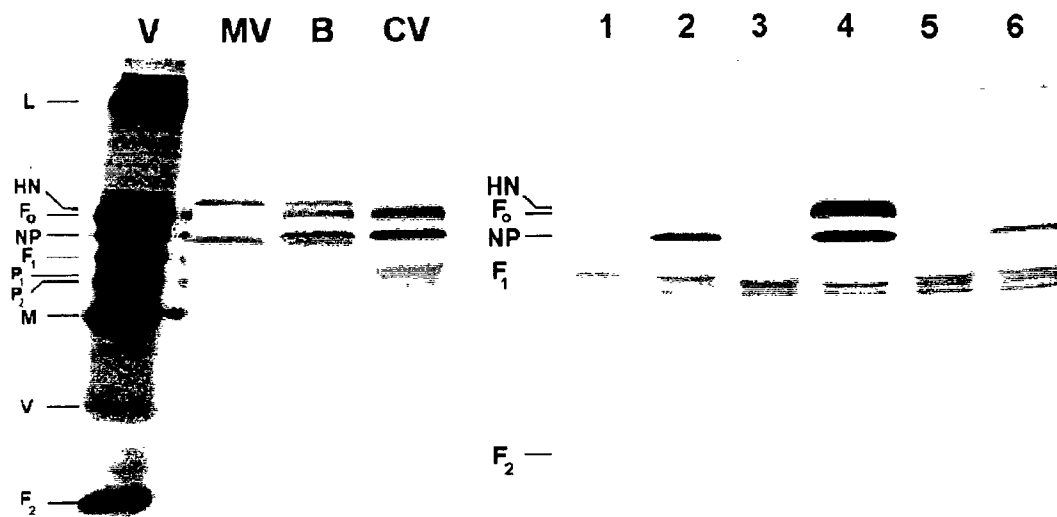


Fig. 12A

Fig. 12B

Fig. 12

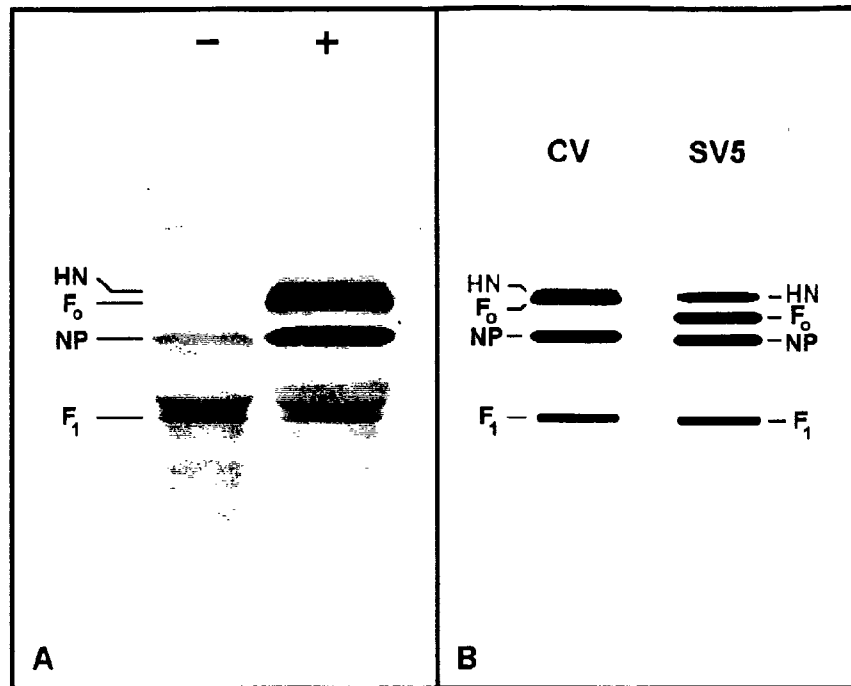


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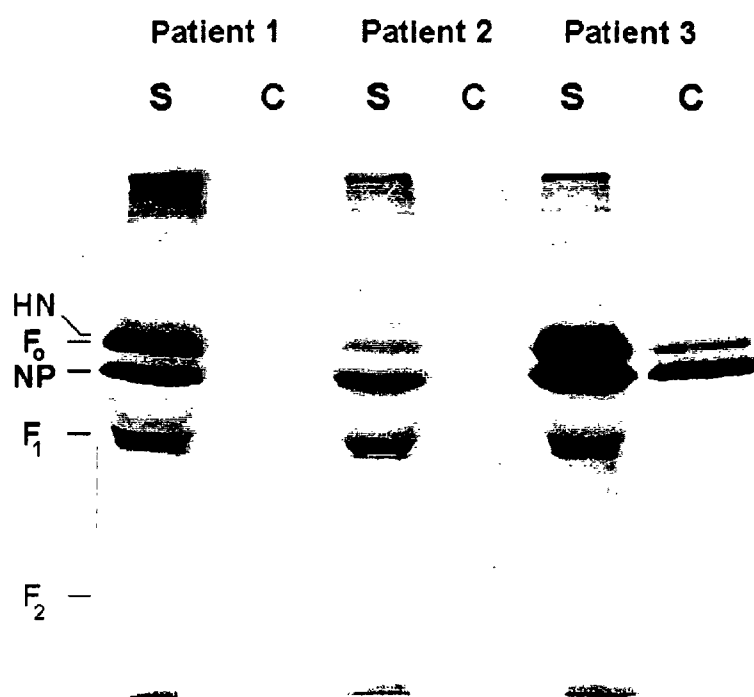


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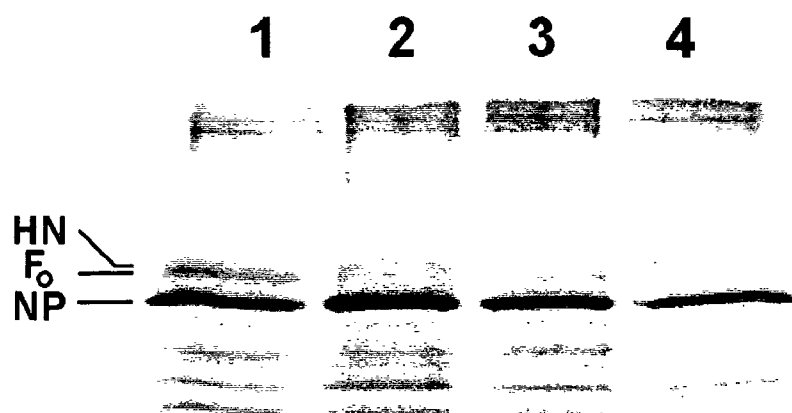


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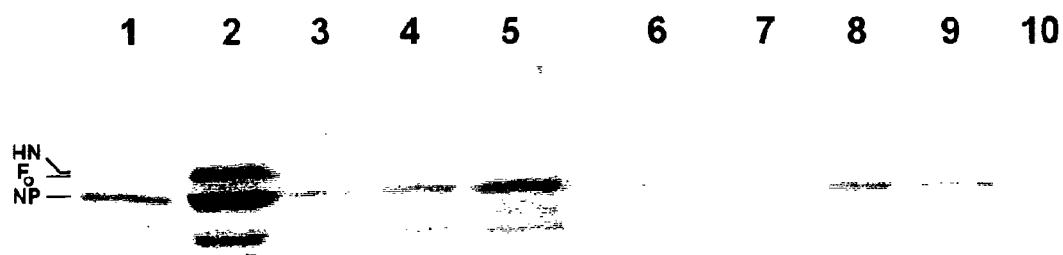


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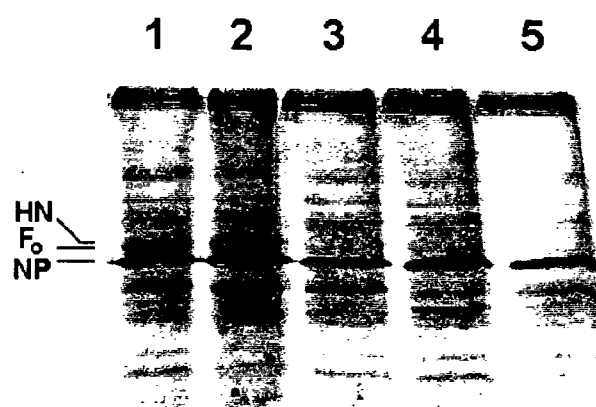


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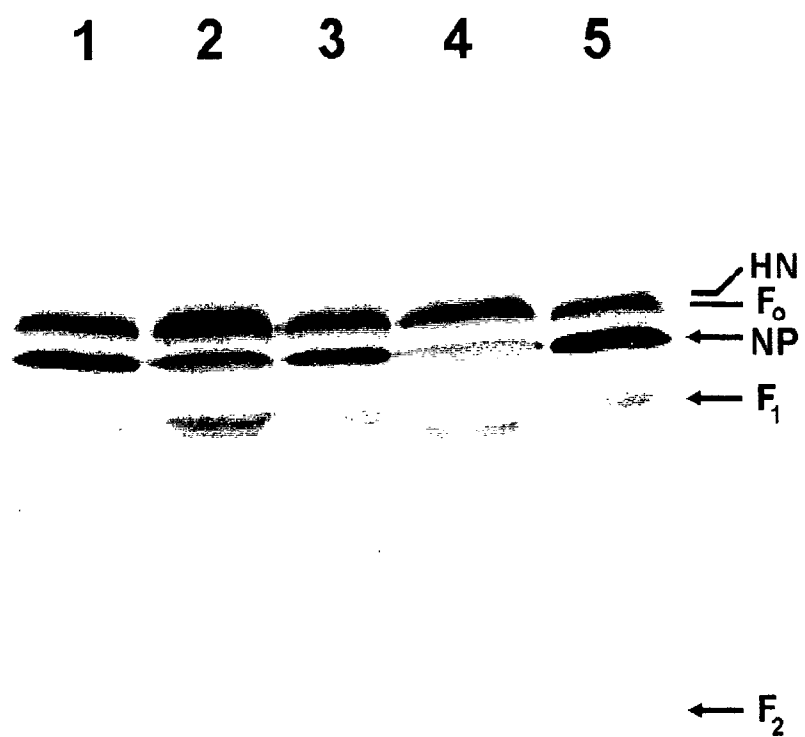


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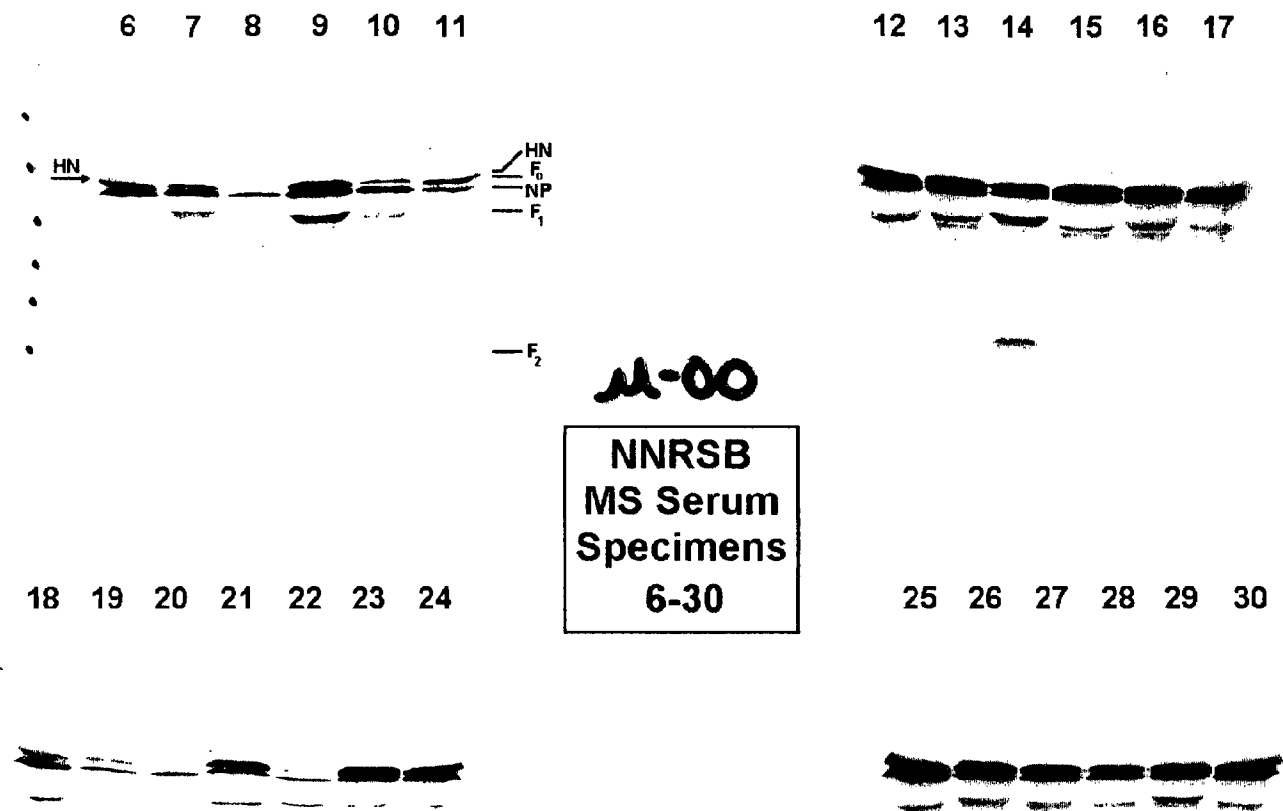
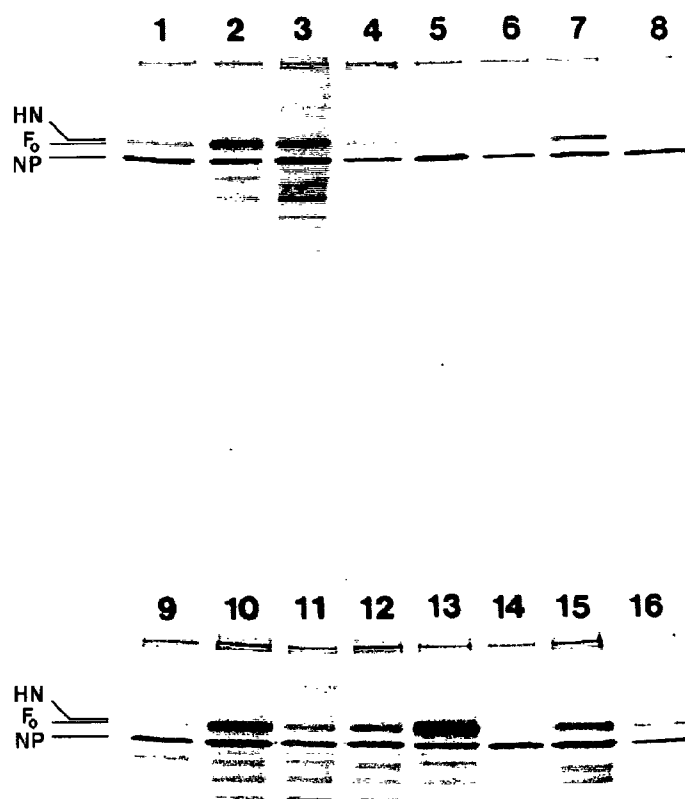


Fig. 20

*Fig. 21*

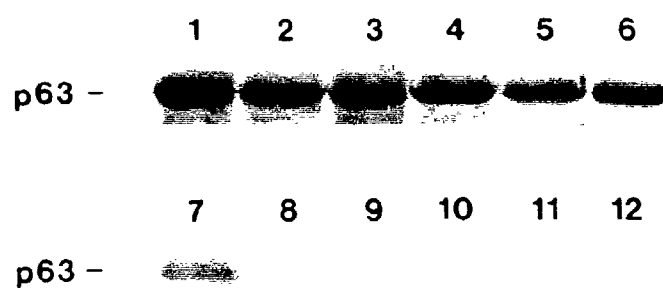


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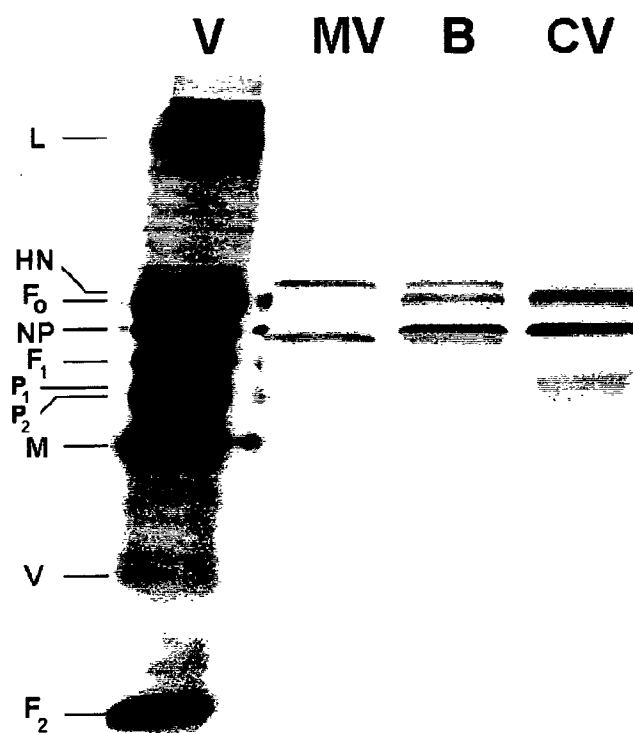


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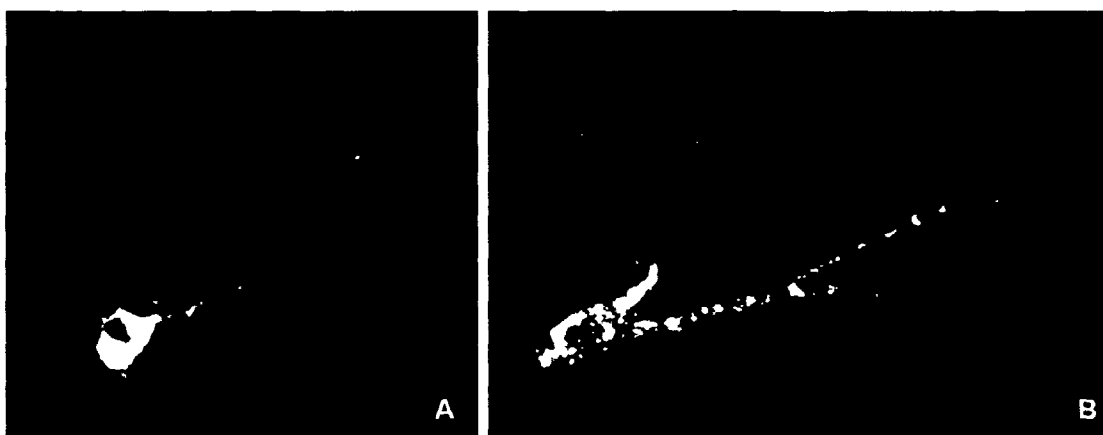
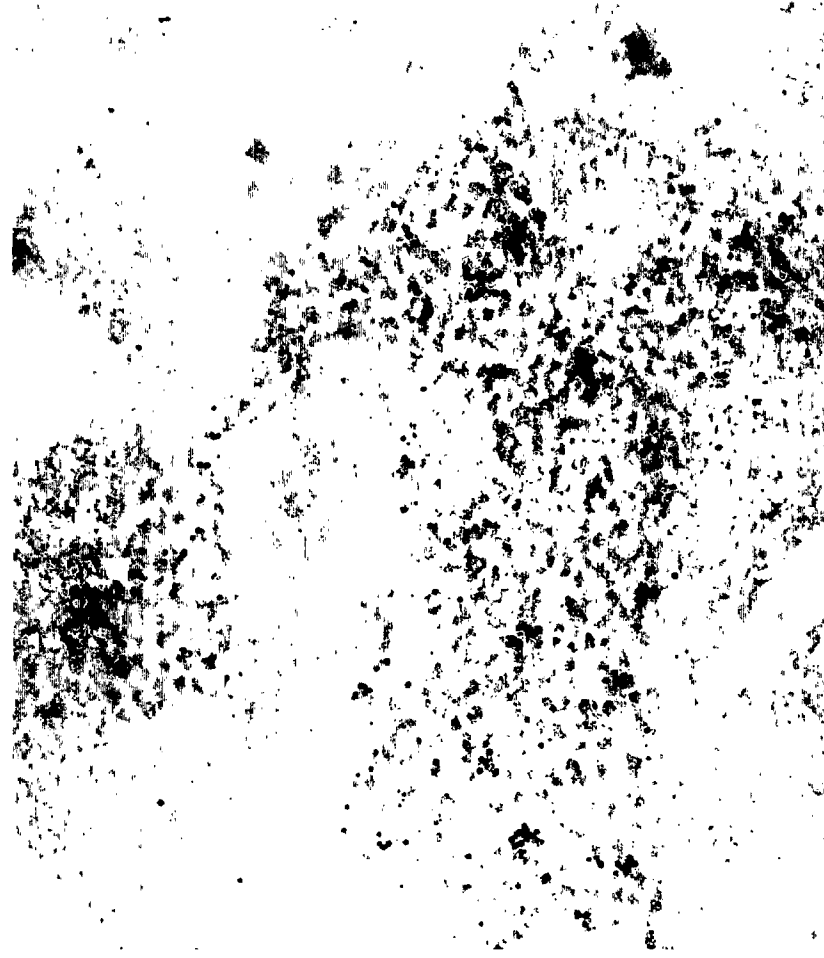


Fig. 24



SEQUENCE LISTING

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Steven J. Robbins (Inventor)

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Leu Gln Asp Gln Ser Glu Glu Gly Thr Ile Pro Pro Thr Thr Leu Lys
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ccg gtt atc agg gta ttt gta cta acc tct aat aac cca gag cta aga 144
Pro Val Ile Arg Val Phe Val Leu Thr Ser Asn Asn Pro Glu Leu Arg
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Arg Asp Ser His Arg Phe Gly Ala Leu Leu Thr Met Phe Ser Leu Pro
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Ser Ala Thr Met Leu Asn His Val Lys Leu Ala Asp Gln Ser Pro Glu
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gct gat atc gaa agg gta gag atc gat ggc ttt gag gag gga tca ttc 336
Ala Asp Ile Glu Arg Val Glu Ile Asp Gly Phe Glu Glu Gly Ser Phe
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cgc tta att ccc aat gct cgc tca ggt atg agc cgt gga gag atc aat 384
Arg Leu Ile Pro Asn Ala Arg Ser Gly Met Ser Arg Gly Glu Ile Asn
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gcc tat gct gca ctt gca gaa gat cta cct gac aca cta aac cat gca 432
Ala Tyr Ala Ala Leu Ala Glu Asp Leu Pro Asp Thr Leu Asn His Ala

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Arg Ala Gln Ser Leu Val Ser Asn Arg Tyr Tyr Ala Met Val Gly Asp			
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Val Gly Lys Tyr Ile Glu Asn Cys Gly Met Gly Gly Phe Phe Leu Thr			
	260	265	270
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Gly	Leu	Thr	Gln	Ala	Glu	Arg	Thr	Glu	Met	Ala	Asn	Thr	Leu	Ala
385				390						395				Lys
Leu	Thr	Thr	Ala	Asn	Arg	Gly	Ala	Asp	Thr	Arg	Gly	Gly	Val	Asn
			405					410						415
Phe	Ser	Ser	Val	Thr	Gly	Thr	Thr	Gln	Met	Pro	Ala	Ala	Ala	Thr
			420				425					430		Gly
Asp	Thr	Phe	Glu	Ser	Tyr	Met	Ala	Ala	Asp	Arg	Leu	Arg	Gln	Arg
	435						440					445		Tyr
Ala	Asp	Ala	Gly	Thr	His	Asp	Asp	Glu	Met	Pro	Pro	Leu	Glu	Glu
	450					455				460				Glu
Glu	Glu	Asp	Asp	Thr	Ser	Ala	Gly	Pro	Arg	Thr	Glu	Pro	Thr	Pro
465				470						475				480
Gln	Val	Ala	Leu	Asp	Ile	Gln	Ser	Ala	Ala	Val	Gly	Ala	Pro	Ile
			485					490						495
Thr	Asp	Asp	Leu	Asn	Ala	Ala	Leu	Gly	Asp	Leu	Asp	Ile		
			500					505						

<210> 5
 <211> 666
 <212> DNA
 <213> Cryptovirus

<220>
 <221> CDS
 <222> (1)...(666)
 <223> Cryptovirus V protein encoding sequence; cDNA in
 mRNA sense

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<400> 5
atg gat ccc act gat ctg agc ttc tcc cca gat gag att aat aag ctc 48
Met Asp Pro Thr Asp Leu Ser Phe Ser Pro Asp Glu Ile Asn Lys Leu
1 5 10 15

ata gag aca ggc ctg aat act gtg gag tat ttt act tcc caa caa gtc 96
Ile Glu Thr Gly Leu Asn Thr Val Glu Tyr Phe Thr Ser Gln Gln Val
20 25 30

aca gga aca tcc tct ctt gga aag aat aca ata cca cca ggg gtc aca 144
Thr Gly Thr Ser Ser Leu Gly Lys Asn Thr Ile Pro Pro Gly Val Thr
35 40 45

gga cta cta acc aat gct gca gag gca aag atc caa gag tca atc aac 192
Gly Leu Leu Thr Asn Ala Ala Glu Ala Lys Ile Gln Glu Ser Ile Asn
50 55 60

cat cag aag ggt tca gtt ggt ggg ggt aca aac cca aag aaa ccg cga 240
His Gln Lys Gly Ser Val Gly Gly Gly Thr Asn Pro Lys Lys Pro Arg
65 70 75 80

tca aaa att gcc att gtg cca gca gat gac aaa aca gtg ccc gaa aag 288
Ser Lys Ile Ala Ile Val Pro Ala Asp Asp Lys Thr Val Pro Glu Lys
85 90 95

ccg atc cca aac cct cta cta ggt ctg gac tcc acc ccg agc acc caa 336
Pro Ile Pro Asn Pro Leu Leu Gly Leu Asp Ser Thr Pro Ser Thr Gln
100 105 110

acc gtg ctt gat cta agt ggg aaa aca tta cca tca gga tcc tat aag 384
Thr Val Leu Asp Leu Ser Gly Lys Thr Leu Pro Ser Gly Ser Tyr Lys
115 120 125

ggg gtt aag ctt gcg aaa ttt ggg aaa gaa aat ctg atg aca cgg ttc 432
Gly Val Lys Leu Ala Lys Phe Gly Lys Glu Asn Leu Met Thr Arg Phe
130 135 140

atc gag gaa ccc aga gag aat cct atc gca acc agt tcc ccc atc gat 480
Ile Glu Glu Pro Arg Glu Asn Pro Ile Ala Thr Ser Ser Pro Ile Asp
145 150 155 160

ttt aag agg ggc aga gat acc ggt ggg ttc cat aga agg gag tac tca 528

```

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Phe Lys Arg Gly Arg Asp Thr Gly Gly Phe His Arg Arg Glu Tyr Ser
      165                      170                      175

atc gga tgg gtg gga gat gaa gtc aag gtc act gag tgg tgc aat cca 576
Ile Gly Trp Val Gly Asp Glu Val Lys Val Thr Glu Trp Cys Asn Pro
      180                      185                      190

tcc tgt tct cca atc acc gct gca gca agg cga ttt aaa tgc act tgt 624
Ser Cys Ser Pro Ile Thr Ala Ala Ala Arg Arg Phe Lys Cys Thr Cys
      195                      200                      205

cac caa tgt cca gtc act tgc tct gaa tgt gaa cga gat act 666
His Gln Cys Pro Val Thr Cys Ser Glu Cys Glu Arg Asp Thr
      210                      215                      220

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<210> 6
<211> 222
<212> PRT
<213> Cryptovirus V Protein

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<400> 6
Met Asp Pro Thr Asp Leu Ser Phe Ser Pro Asp Glu Ile Asn Lys Leu
 1      5      10      15
Ile Glu Thr Gly Leu Asn Thr Val Glu Tyr Phe Thr Ser Gln Gln Val
      20      25      30
Thr Gly Thr Ser Ser Leu Gly Lys Asn Thr Ile Pro Pro Gly Val Thr
      35      40      45
Gly Leu Leu Thr Asn Ala Ala Glu Ala Lys Ile Gln Glu Ser Ile Asn
      50      55      60
His Gln Lys Gly Ser Val Gly Gly Gly Thr Asn Pro Lys Lys Pro Arg
      65      70      75      80
Ser Lys Ile Ala Ile Val Pro Ala Asp Asp Lys Thr Val Pro Glu Lys
      85      90      95
Pro Ile Pro Asn Pro Leu Leu Gly Leu Asp Ser Thr Pro Ser Thr Gln
      100     105     110
Thr Val Leu Asp Leu Ser Gly Lys Thr Leu Pro Ser Gly Ser Tyr Lys
      115     120     125
Gly Val Lys Leu Ala Lys Phe Gly Lys Glu Asn Leu Met Thr Arg Phe
      130     135     140
Ile Glu Glu Pro Arg Glu Asn Pro Ile Ala Thr Ser Ser Pro Ile Asp
      145     150     155     160
Phe Lys Arg Gly Arg Asp Thr Gly Gly Phe His Arg Arg Glu Tyr Ser
      165     170     175
Ile Gly Trp Val Gly Asp Glu Val Lys Val Thr Glu Trp Cys Asn Pro
      180     185     190
Ser Cys Ser Pro Ile Thr Ala Ala Ala Arg Arg Phe Lys Cys Thr Cys
      195     200     205
His Gln Cys Pro Val Thr Cys Ser Glu Cys Glu Arg Asp Thr
      210     215     220

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```

<210> 7
<211> 1176

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<212> DNA

<213> Cryptovirus

<220>

<221> CDS

<222> (1)...(1176)

<223> Cryptovirus P protein encoding sequence; cDNA in mRNA sense

<400> 7

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atg gat ccc act gat ctg agc ttc tcc cca gat gag att aat aag ctc 48
Met Asp Pro Thr Asp Leu Ser Phe Ser Pro Asp Glu Ile Asn Lys Leu
 1          5          10          15

ata gag aca ggc ctg aat act gtg gag tat ttt act tcc caa caa gtc 96
Ile Glu Thr Gly Leu Asn Thr Val Glu Tyr Phe Thr Ser Gln Gln Val
          20          25          30

aca gga aca tcc tct ctt gga aag aat aca ata cca cca ggg gtc aca 144
Thr Gly Thr Ser Ser Leu Gly Lys Asn Thr Ile Pro Pro Gly Val Thr
          35          40          45

gga cta cta acc aat gct gca gag gca aag atc caa gag tca atc aac 192
Gly Leu Leu Thr Asn Ala Ala Glu Ala Lys Ile Gln Glu Ser Ile Asn
          50          55          60

cat cag aag ggt tca gtt ggt ggg ggt aca aac cca aag aaa ccg cga 240
His Gln Lys Gly Ser Val Gly Gly Gly Thr Asn Pro Lys Lys Pro Arg
          65          70          75          80

tca aaa att gcc att gtg cca gca gat gac aaa aca gtg ccc gaa aag 288
Ser Lys Ile Ala Ile Val Pro Ala Asp Asp Lys Thr Val Pro Glu Lys
          85          90          95

ccg atc cca aac cct cta cta ggt ctg gac tcc acc ccg agc acc caa 336
Pro Ile Pro Asn Pro Leu Leu Gly Leu Asp Ser Thr Pro Ser Thr Gln
          100          105          110

acc gtg ctt gat cta agt ggg aaa aca tta cca tca gga tcc tat aag 384
Thr Val Leu Asp Leu Ser Gly Lys Thr Leu Pro Ser Gly Ser Tyr Lys
          115          120          125

ggg gtt aag ctt gcg aaa ttt ggg aaa gaa aat ctg atg aca cgg ttc 432
Gly Val Lys Leu Ala Lys Phe Gly Lys Glu Asn Leu Met Thr Arg Phe
          130          135          140

atc gag gaa ccc aga gag aat cct atc gca acc agt tcc ccc atc gat 480
Ile Glu Glu Pro Arg Glu Asn Pro Ile Ala Thr Ser Ser Pro Ile Asp
          145          150          155          160

ttt aag agg ggg gca gag ata ccg gtg ggt tcc ata gaa ggg agt act 528
Phe Lys Arg Gly Ala Glu Ile Pro Val Gly Ser Ile Glu Gly Ser Thr
          165          170          175

caa tcg gat ggg tgg gag atg aag tca agg tca ctg agt ggt gca atc 576

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Gln Ser Asp Gly Trp Glu Met Lys Ser Arg Ser Leu Ser Gly Ala Ile	
180	185 190
cat cct gtt ctc caa tca ccg ctg cag caa ggc gat tta aat gca ctt	624
His Pro Val Leu Gln Ser Pro Leu Gln Gln Gly Asp Leu Asn Ala Leu	
195	200 205
gtc acc aat gtc cag tca ctt gct ctg aat gtg aac gag ata ctt aat	672
Val Thr Asn Val Gln Ser Leu Ala Leu Asn Val Asn Glu Ile Leu Asn	
210	215 220
aca gtg aga aat ttg gac tct cgg atg aat caa ctg gag aca aaa gta	720
Thr Val Arg Asn Leu Asp Ser Arg Met Asn Gln Leu Glu Thr Lys Val	
225	230 235 240
gat cgc att ctc tca tct cag tct cta atc cag acc atc aag aat gac	768
Asp Arg Ile Leu Ser Ser Gln Ser Leu Ile Gln Thr Ile Lys Asn Asp	
	245 250 255
ata att gga ctt aaa gca ggg atg gct act tta gaa gga atg att aca	816
Ile Ile Gly Leu Lys Ala Gly Met Ala Thr Leu Glu Gly Met Ile Thr	
	260 265 270
act gtg aaa atc atg gac ccg gga gtt ccc agt aat gtt act gtg gaa	864
Thr Val Lys Ile Met Asp Pro Gly Val Pro Ser Asn Val Thr Val Glu	
	275 280 285
gat gta cgc aag aaa cta agt aac cat gct gtt gtt gtg cca gaa tca	912
Asp Val Arg Lys Lys Leu Ser Asn His Ala Val Val Val Pro Glu Ser	
	290 295 300
ttc aat gat agt ttc ttg act caa tct gaa gat gta att tca ctt gat	960
Phe Asn Asp Ser Phe Leu Thr Gln Ser Glu Asp Val Ile Ser Leu Asp	
305	310 315 320
gag ttg gct cga cca act gca aca agt gtt aag aag att gtc agg aag	1008
Glu Leu Ala Arg Pro Thr Ala Thr Ser Val Lys Lys Ile Val Arg Lys	
	325 330 335
gtt cct cct cag aag gat ctg act gga ttg aag atc aca cta gag caa	1056
Val Pro Pro Gln Lys Asp Leu Thr Gly Leu Lys Ile Thr Leu Glu Gln	
	340 345 350
ttg gca aag gat tgc atc agc aaa ccg aag atg agg gaa gat tat ctc	1104
Leu Ala Lys Asp Cys Ile Ser Lys Pro Lys Met Arg Glu Asp Tyr Leu	
	355 360 365
ctc aag atc aac cag gct tct agt gag gct cag cta att gac ctc aag	1152
Leu Lys Ile Asn Gln Ala Ser Ser Glu Ala Gln Leu Ile Asp Leu Lys	
	370 375 380
aaa gca atc atc cgc agt gca att	1176
Lys Ala Ile Ile Arg Ser Ala Ile	
385	390

<210> 8

<211> 392

<212> PRT

<213> Cryptovirus P Protein

<400> 8

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Met Asp Pro Thr Asp Leu Ser Phe Ser Pro Asp Glu Ile Asn Lys Leu
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Ile Glu Thr Gly Leu Asn Thr Val Glu Tyr Phe Thr Ser Gln Gln Val
          20          25          30
Thr Gly Thr Ser Ser Leu Gly Lys Asn Thr Ile Pro Pro Gly Val Thr
          35          40          45
Gly Leu Leu Thr Asn Ala Ala Glu Ala Lys Ile Gln Glu Ser Ile Asn
          50          55          60
His Gln Lys Gly Ser Val Gly Gly Gly Thr Asn Pro Lys Lys Pro Arg
65          70          75          80
Ser Lys Ile Ala Ile Val Pro Ala Asp Asp Lys Thr Val Pro Glu Lys
          85          90          95
Pro Ile Pro Asn Pro Leu Leu Gly Leu Asp Ser Thr Pro Ser Thr Gln
          100          105          110
Thr Val Leu Asp Leu Ser Gly Lys Thr Leu Pro Ser Gly Ser Tyr Lys
          115          120          125
Gly Val Lys Leu Ala Lys Phe Gly Lys Glu Asn Leu Met Thr Arg Phe
          130          135          140
Ile Glu Glu Pro Arg Glu Asn Pro Ile Ala Thr Ser Ser Pro Ile Asp
145          150          155          160
Phe Lys Arg Gly Ala Glu Ile Pro Val Gly Ser Ile Glu Gly Ser Thr
          165          170          175
Gln Ser Asp Gly Trp Glu Met Lys Ser Arg Ser Leu Ser Gly Ala Ile
          180          185          190
His Pro Val Leu Gln Ser Pro Leu Gln Gln Gly Asp Leu Asn Ala Leu
          195          200          205
Val Thr Asn Val Gln Ser Leu Ala Leu Asn Val Asn Glu Ile Leu Asn
          210          215          220
Thr Val Arg Asn Leu Asp Ser Arg Met Asn Gln Leu Glu Thr Lys Val
225          230          235          240
Asp Arg Ile Leu Ser Ser Gln Ser Leu Ile Gln Thr Ile Lys Asn Asp
          245          250          255
Ile Ile Gly Leu Lys Ala Gly Met Ala Thr Leu Glu Gly Met Ile Thr
          260          265          270
Thr Val Lys Ile Met Asp Pro Gly Val Pro Ser Asn Val Thr Val Glu
          275          280          285
Asp Val Arg Lys Lys Leu Ser Asn His Ala Val Val Val Pro Glu Ser
          290          295          300
Phe Asn Asp Ser Phe Leu Thr Gln Ser Glu Asp Val Ile Ser Leu Asp
305          310          315          320
Glu Leu Ala Arg Pro Thr Ala Thr Ser Val Lys Lys Ile Val Arg Lys
          325          330          335
Val Pro Pro Gln Lys Asp Leu Thr Gly Leu Lys Ile Thr Leu Glu Gln
          340          345          350
Leu Ala Lys Asp Cys Ile Ser Lys Pro Lys Met Arg Glu Asp Tyr Leu
          355          360          365
Leu Lys Ile Asn Gln Ala Ser Ser Glu Ala Gln Leu Ile Asp Leu Lys
          370          375          380

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Lys Ala Ile Ile Arg Ser Ala Ile
385 390

<210> 9

<211> 1131

<212> DNA

<213> Cryptovirus

<220>

<221> CDS

<222> (1)...(1131)

<223> Cryptovirus M protein encoding sequence; cDNA in
mRNA sense

<400> 9

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Met Pro Ser Ile Ser Ile Pro Ala Asp Pro Thr Asn Pro Arg Gln Ser	
1 5 10 15	
ata aaa gcg ttc cca att gtg att aac agt gat ggg ggt gag aaa ggc	96
Ile Lys Ala Phe Pro Ile Val Ile Asn Ser Asp Gly Gly Glu Lys Gly	
20 25 30	
cgc ttg gtt aaa caa cta cgt aca acc tac ttg aat gac cta gat act	144
Arg Leu Val Lys Gln Leu Arg Thr Thr Tyr Leu Asn Asp Leu Asp Thr	
35 40 45	
cat gag cca ctg gtg aca ttc gta aat acc tat gga ttc atc tac gaa	192
His Glu Pro Leu Val Thr Phe Val Asn Thr Tyr Gly Phe Ile Tyr Glu	
50 55 60	
cag aat cgg ggg aat gcc att gtc gga gag gat caa ctt ggg aag aaa	240
Gln Asn Arg Gly Asn Ala Ile Val Gly Glu Asp Gln Leu Gly Lys Lys	
65 70 75 80	
aga gag gct gtg act gct gca atg gtt acc ctt gga tgt ggg cct aat	288
Arg Glu Ala Val Thr Ala Ala Met Val Thr Leu Gly Cys Gly Pro Asn	
85 90 95	
cta cca tca tta ggg aat gtc ctg aga caa ctg agt gaa ttc caa gtc	336
Leu Pro Ser Leu Gly Asn Val Leu Arg Gln Leu Ser Glu Phe Gln Val	
100 105 110	
att gtt agg aag aca tcc agc aaa gcg gaa gag atg gtc ttt gaa att	384
Ile Val Arg Lys Thr Ser Ser Lys Ala Glu Glu Met Val Phe Glu Ile	
115 120 125	
gtt aag tat ccg aga ata ttt cgg ggt cat aca tta atc cag aaa gga	432
Val Lys Tyr Pro Arg Ile Phe Arg Gly His Thr Leu Ile Gln Lys Gly	
130 135 140	
cta gtc tgt gtc tcc gca gaa aaa ttt gtt aag tca cca ggg aaa gta	480
Leu Val Cys Val Ser Ala Glu Lys Phe Val Lys Ser Pro Gly Lys Val	
145 150 155 160	

caa tct gga atg gac tat ctc ttc att ccg aca ttt ctg tca gtg act	528
Gln Ser Gly Met Asp Tyr Leu Phe Ile Pro Thr Phe Leu Ser Val Thr	
165 170 175	
tat tgt cca gct gca atc aaa ttt cag gta cct ggc ccc atg ttg aaa	576
Tyr Cys Pro Ala Ala Ile Lys Phe Gln Val Pro Gly Pro Met Leu Lys	
180 185 190	
atg agg tca aga tac act cag agc tta caa ctt gaa cta atg ata aga	624
Met Arg Ser Arg Tyr Thr Gln Ser Leu Gln Leu Glu Leu Met Ile Arg	
195 200 205	
atc ctg tgt aag ccc gat tcg cca ctt atg aag gtc cat atc cct gac	672
Ile Leu Cys Lys Pro Asp Ser Pro Leu Met Lys Val His Ile Pro Asp	
210 215 220	
aag gag gga aga gga tgt ctt gta tca gta tgg ctg cat gta tgc aac	720
Lys Glu Gly Arg Gly Cys Leu Val Ser Val Trp Leu His Val Cys Asn	
225 230 235 240	
atc ttc aaa tca gga aac aag aat ggc agt gag tgg cag gaa tac tgg	768
Ile Phe Lys Ser Gly Asn Lys Asn Gly Ser Glu Trp Gln Glu Tyr Trp	
245 250 255	
atg aga aag tgt gct aac atg caa ctt gaa gtg tcg att gca gat atg	816
Met Arg Lys Cys Ala Asn Met Gln Leu Glu Val Ser Ile Ala Asp Met	
260 265 270	
tgg gga cca act atc ata att cat gcc aga ggt cac att ccc aaa agt	864
Trp Gly Pro Thr Ile Ile Ile His Ala Arg Gly His Ile Pro Lys Ser	
275 280 285	
gct aag ttg ttt ttt gga aag ggt gga tgg agc tgc cat cca ctt cac	912
Ala Lys Leu Phe Phe Gly Lys Gly Gly Trp Ser Cys His Pro Leu His	
290 295 300	
gaa att gtt cca agt gtc act aaa aca cta tgg tcc gta ggt tgt gaa	960
Glu Ile Val Pro Ser Val Thr Lys Thr Leu Trp Ser Val Gly Cys Glu	
305 310 315 320	
att aca aag gcg aag gca ata ata caa gag agt agc atc tct ctt ctc	1008
Ile Thr Lys Ala Lys Ala Ile Ile Gln Glu Ser Ser Ile Ser Leu Leu	
325 330 335	
gtg gag act act gac atc ata agt cca aaa gtt aaa att tca tct aag	1056
Val Glu Thr Thr Asp Ile Ile Ser Pro Lys Val Lys Ile Ser Ser Lys	
340 345 350	
cat cgc cgc ttt ggg aaa tca aat tgg ggt ctg ttc aag aaa act aaa	1104
His Arg Arg Phe Gly Lys Ser Asn Trp Gly Leu Phe Lys Lys Thr Lys	
355 360 365	
tca cta cct aac cta acg gag ctg gaa	1131
Ser Leu Pro Asn Leu Thr Glu Leu Glu	

370

375

<210> 10
 <211> 377
 <212> PRT
 <213> Cryptovirus M Protein

<400> 10

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Ile	Lys	Ala	Phe	Pro	Ile	Val	Ile	Asn	Ser	Asp	Gly	Gly	Glu	Lys	Gly
		20						25					30		
Arg	Leu	Val	Lys	Gln	Leu	Arg	Thr	Thr	Tyr	Leu	Asn	Asp	Leu	Asp	Thr
		35					40					45			
His	Glu	Pro	Leu	Val	Thr	Phe	Val	Asn	Thr	Tyr	Gly	Phe	Ile	Tyr	Glu
	50					55				60					
Gln	Asn	Arg	Gly	Asn	Ala	Ile	Val	Gly	Glu	Asp	Gln	Leu	Gly	Lys	Lys
65				70					75					80	
Arg	Glu	Ala	Val	Thr	Ala	Ala	Met	Val	Thr	Leu	Gly	Cys	Gly	Pro	Asn
			85					90					95		
Leu	Pro	Ser	Leu	Gly	Asn	Val	Leu	Arg	Gln	Leu	Ser	Glu	Phe	Gln	Val
		100						105					110		
Ile	Val	Arg	Lys	Thr	Ser	Ser	Lys	Ala	Glu	Glu	Met	Val	Phe	Glu	Ile
		115					120					125			
Val	Lys	Tyr	Pro	Arg	Ile	Phe	Arg	Gly	His	Thr	Leu	Ile	Gln	Lys	Gly
	130					135					140				
Leu	Val	Cys	Val	Ser	Ala	Glu	Lys	Phe	Val	Lys	Ser	Pro	Gly	Lys	Val
145				150					155					160	
Gln	Ser	Gly	Met	Asp	Tyr	Leu	Phe	Ile	Pro	Thr	Phe	Leu	Ser	Val	Thr
			165					170					175		
Tyr	Cys	Pro	Ala	Ala	Ile	Lys	Phe	Gln	Val	Pro	Gly	Pro	Met	Leu	Lys
		180						185					190		
Met	Arg	Ser	Arg	Tyr	Thr	Gln	Ser	Leu	Gln	Leu	Glu	Leu	Met	Ile	Arg
	195					200					205				
Ile	Leu	Cys	Lys	Pro	Asp	Ser	Pro	Leu	Met	Lys	Val	His	Ile	Pro	Asp
	210					215					220				
Lys	Glu	Gly	Arg	Gly	Cys	Leu	Val	Ser	Val	Trp	Leu	His	Val	Cys	Asn
225				230					235					240	
Ile	Phe	Lys	Ser	Gly	Asn	Lys	Asn	Gly	Ser	Glu	Trp	Gln	Glu	Tyr	Trp
			245					250					255		
Met	Arg	Lys	Cys	Ala	Asn	Met	Gln	Leu	Glu	Val	Ser	Ile	Ala	Asp	Met
		260						265					270		
Trp	Gly	Pro	Thr	Ile	Ile	Ile	His	Ala	Arg	Gly	His	Ile	Pro	Lys	Ser
		275				280					285				
Ala	Lys	Leu	Phe	Phe	Gly	Lys	Gly	Gly	Trp	Ser	Cys	His	Pro	Leu	His
	290				295						300				
Glu	Ile	Val	Pro	Ser	Val	Thr	Lys	Thr	Leu	Trp	Ser	Val	Gly	Cys	Glu
305				310					315					320	
Ile	Thr	Lys	Ala	Lys	Ala	Ile	Ile	Gln	Glu	Ser	Ser	Ile	Ser	Leu	Leu
			325					330					335		
Val	Glu	Thr	Thr	Asp	Ile	Ile	Ser	Pro	Lys	Val	Lys	Ile	Ser	Ser	Lys
		340						345				350			
His	Arg	Arg	Phe	Gly	Lys	Ser	Asn	Trp	Gly	Leu	Phe	Lys	Lys	Thr	Lys
	355						360					365			

Ser Leu Pro Asn Leu Thr Glu Leu Glu
 370 375

<210> 11

<211> 1653

<212> DNA

<213> Cryptovirus

<220>

<221> CDS

<222> (1)...(1653)

<223> Cryptovirus F protein encoding sequence; cDNA in
 mRNA sense

<400> 11

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Met Ser Thr Ile Ile Gln Ser Leu Val Val Ser Cys Leu Leu Ala Gly	
1 5 10 15	
gca ggc agc ctt gat cca gca gcc ctg atg caa atc ggt gtc att cca	96
Ala Gly Ser Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro	
20 25 30	
aca aat gtc cgg caa ctt atg tat tat act gag gcc tca tca gca ttc	144
Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe	
35 40 45	
att gtt gtg aag tta atg cct aca att gac tgg ccg att agt gga tgt	192
Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys	
50 55 60	
aat ata aca tca att tca agc tat aat gca aca gtg aca aaa ctg cta	240
Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu	
65 70 75 80	
cag ccg atc ggt gag aat ttg gag acg att agg aac cag ttg att cca	288
Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro	
85 90 95	
act cgg agg aga cgc cgg ttt gca ggg gtg gtg att gga tta gct gca	336
Thr Arg Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala	
100 105 110	
tta gga gta gct act gcc gca caa gtc act gcc gca gta gca cta gtt	384
Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val	
115 120 125	
aag gca aat gaa aat act gcg gct ata ctg aat ctg aaa aat gca atc	432
Lys Ala Asn Glu Asn Thr Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile	
130 135 140	
caa aaa aca aat gca gca gtt gca gat gtg gtc cag gcc aca caa tca	480
Gln Lys Thr Asn Ala Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser	
145 150 155 160	

cta gga acg gca gtt caa gca gtt caa gat cac ata aac agt gtg ata	528
Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Ile	
165 170 175	
agt cca gca att aca gca gcc aat tgt aag gcc caa gat gct atc att	576
Ser Pro Ala Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile	
180 185 190	
ggc tca atc ctc aat ctc tat ttg acc gag ttg aca act atc ttc cac	624
Gly Ser Ile Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His	
195 200 205	
aat caa att aca aac cct gca ttg agt cct att aca att caa gct tta	672
Asn Gln Ile Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu	
210 215 220	
agg atc cta ctg ggg agt acc ttg ccg act gtg gtc gaa aaa tct ttc	720
Arg Ile Leu Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe	
225 230 235 240	
aat acc cag ata agt gca gct gag ctt ctc tca tca ggg ttg ttg aca	768
Asn Thr Gln Ile Ser Ala Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr	
245 250 255	
ggc cag att gtg gga tta gat ttg acc tat atg cag atg gtc ata aaa	816
Gly Gln Ile Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys	
260 265 270	
att gag ctg cca act tta act gta caa cct gca acc cag atc ata gat	864
Ile Glu Leu Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp	
275 280 285	
ctg gcc acc att tct gca ttc att aac aat caa gaa gtc atg gcc caa	912
Leu Ala Thr Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln	
290 295 300	
tta cca aca cgt gtt att gtg act ggc agc ttg atc caa gcc tat ccc	960
Leu Pro Thr Arg Val Ile Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro	
305 310 315 320	
gca tcg caa tgc act att aca ccc aac act gtg tac tgt agg tat aat	1008
Ala Ser Gln Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn	
325 330 335	
gat gcc caa gta ctc tca gat gat acg atg gct tgc ctc caa ggt aac	1056
Asp Ala Gln Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn	
340 345 350	
ttg aca aga tgc acc ttc tct cca gtg gtt ggg agc ttt ctc act cga	1104
Leu Thr Arg Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg	
355 360 365	
ttc gtg ctg ttc gat gga ata gtt tat gca aat tgc agg tcg atg ctg	1152
Phe Val Leu Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu	

370	375	380	
tgc aag tgc atg caa cct gct gcc gtg atc cta cag ccg agt tca tcc			1200
Cys Lys Cys Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser			
385	390	395	400
cct gta act gtc att gac atg cac aaa tgt gtg agt ctg cag ctt gac			1248
Pro Val Thr Val Ile Asp Met His Lys Cys Val Ser Leu Gln Leu Asp			
	405	410	415
gat ctc aga ttc acc atc act caa ttg gcc aat gta acc tac aat agc			1296
Asp Leu Arg Phe Thr Ile Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser			
	420	425	430
acc atc aag ctt gaa aca tcc cag atc ttg cct att gat ccg ttg gat			1344
Thr Ile Lys Leu Glu Thr Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp			
	435	440	445
ata tcc cag aat tta gct gcg gtg aat aag agt cta agt gat gca cta			1392
Ile Ser Gln Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu			
	450	455	460
caa cac tta gca caa agt gac aca tac ctt tct gca atc aca tca gct			1440
Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala			
	465	470	475
acg act aca agt gta tta tcc ata ata gca atc tgt ctt gga tgg tta			1488
Thr Thr Thr Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu			
	485	490	495
ggg tta ata tta ata atc ttg ctc agt gta gtt gtg tgg aag tta ttg			1536
Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu			
	500	505	510
acc att gtc gct gct aat cga aat aga atg gag aat ttt gtt tat cat			1584
Thr Ile Val Ala Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His			
	515	520	525
aat tca gca ttc cac cac cca cga tct gat ctc agt gag aaa aat caa			1632
Asn Ser Ala Phe His His Pro Arg Ser Asp Leu Ser Glu Lys Asn Gln			
	530	535	540
cct gca act ctt gga aca aga			1653
Pro Ala Thr Leu Gly Thr Arg			
	545	550	

<210> 12

<211> 551

<212> PRT

<213> Cryptovirus F protein

<400> 12

Met Ser Thr Ile Ile Gln Ser Leu Val Val Ser Cys Leu Leu Ala Gly

1

5

10

15

Ala Gly Ser Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro
 20 25 30
 Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe
 35 40 45
 Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys
 50 55 60
 Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu
 65 70 75 80
 Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro
 85 90 95
 Thr Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala
 100 105 110
 Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val
 115 120 125
 Lys Ala Asn Glu Asn Thr Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile
 130 135 140
 Gln Lys Thr Asn Ala Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser
 145 150 155 160
 Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Ile
 165 170 175
 Ser Pro Ala Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile
 180 185 190
 Gly Ser Ile Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His
 195 200 205
 Asn Gln Ile Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu
 210 215 220
 Arg Ile Leu Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe
 225 230 235 240
 Asn Thr Gln Ile Ser Ala Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr
 245 250 255
 Gly Gln Ile Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys
 260 265 270
 Ile Glu Leu Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp
 275 280 285
 Leu Ala Thr Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln
 290 295 300
 Leu Pro Thr Arg Val Ile Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro
 305 310 315 320
 Ala Ser Gln Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn
 325 330 335
 Asp Ala Gln Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn
 340 345 350
 Leu Thr Arg Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg
 355 360 365
 Phe Val Leu Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu
 370 375 380
 Cys Lys Cys Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser
 385 390 395 400
 Pro Val Thr Val Ile Asp Met His Lys Cys Val Ser Leu Gln Leu Asp
 405 410 415
 Asp Leu Arg Phe Thr Ile Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser
 420 425 430
 Thr Ile Lys Leu Glu Thr Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp
 435 440 445
 Ile Ser Gln Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu

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      450              455              460
Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala
465              470              475              480
Thr Thr Thr Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu
      485              490              495
Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu
      500              505              510
Thr Ile Val Ala Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His
      515              520              525
Asn Ser Ala Phe His His Pro Arg Ser Asp Leu Ser Glu Lys Asn Gln
      530              535              540
Pro Ala Thr Leu Gly Thr Arg
545              550

```

<210> 13

<211> 1596

<212> DNA

<213> Cryptovirus

<220>

<221> CDS

<222> (1)...(1596)

<223> Cryptovirus F0 protein encoding sequence; cDNA in mRNA sense

<400> 13

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ctt gat cca gca gcc ctc atg caa atc ggt gtc att cca aca aat gtc 48
Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro Thr Asn Val
 1              5              10              15

```

```

cgg caa ctt atg tat tat act gag gcc tca tca gca ttc att gtt gtg 96
Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe Ile Val Val
      20              25              30

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```

aag tta atg cct aca att gac tgc ccg att agt gga tgt aat ata aca 144
Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys Asn Ile Thr
      35              40              45

```

```

tca att tca agc tat aat gca aca gtg aca aaa ctc cta cag ccg atc 192
Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu Gln Pro Ile
      50              55              60

```

```

ggt gag aat ttg gag acg att agg aac cag ttg att cca act cgg agg 240
Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro Thr Arg Arg
      65              70              75              80

```

```

aga cgc cgg ttt gca ggg gtg gtg att gga tta gct gca tta gga gta 288
Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala Leu Gly Val
      85              90              95

```

```

gct act gcc gca caa gtc act gcc gca gta gca cta gtt aag gca aat 336
Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val Lys Ala Asn
      100              105              110

```


gaa aat act gcg gct ata ctc aat ctc aaa aat gca atc caa aaa aca	384
Glu Asn Thr Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile Gln Lys Thr	
115 120 125	
aat gca gca gtt gca gat gtg gtc cag gcc aca caa tca cta gga acg	432
Asn Ala Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser Leu Gly Thr	
130 135 140	
gca gtt caa gca gtt caa gat cac ata aac agt gtg ata agt cca gca	480
Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Ile Ser Pro Ala	
145 150 155 160	
att aca gca gcc aat tgt aag gcc caa gat gct atc att ggc tca atc	528
Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile Gly Ser Ile	
165 170 175	
ctc aat ctc tat ttg acc gag ttg aca act atc ttc cac aat caa att	576
Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His Asn Gln Ile	
180 185 190	
aca aac cct gca ttg agt cct att aca att caa gct tta agg atc cta	624
Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu Arg Ile Leu	
195 200 205	
ctg ggg agt acc ttg ccg act gtg gtc gaa aaa tct ttc aat acc cag	672
Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe Asn Thr Gln	
210 215 220	
ata agt gca gct gag ctt ctc tca tca ggg ttg ttg aca ggc cag att	720
Ile Ser Ala Ala Glu Leu Ser Ser Gly Leu Leu Thr Gly Gln Ile	
225 230 235 240	
gtg gga tta gat ttg acc tat atg cag atg gtc ata aaa att gag ctg	768
Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys Ile Glu Leu	
245 250 255	
cca act tta act gta caa cct gca acc cag atc ata gat ctg gcc acc	816
Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp Leu Ala Thr	
260 265 270	
att tct gca ttc att aac aat caa gaa gtc atg gcc caa tta cca aca	864
Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln Leu Pro Thr	
275 280 285	
cgt gtt att gtg act ggc agc ttg atc caa gcc tat ccc gca tcg caa	912
Arg Val Ile Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro Ala Ser Gln	
290 295 300	
tgc act att aca ccc aac act gtg tac tgt agg tat aat gat gcc caa	960
Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn Asp Ala Gln	
305 310 315 320	
gta ctc tca gat gat acg atg gct tgc ctc caa ggt aac ttg aca aga	1008
Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn Leu Thr Arg	
325 330 335	

tgc acc ttc tct cca gtg gtt ggg agc ttt ctc act cga ttc gtg ctg	1056
Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg Phe Val Leu	
340 345 350	
ttc gat gga ata gtt tat gca aat tgc agg tcg atg ctg tgc aag tgc	1104
Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu Cys Lys Cys	
355 360 365	
atg caa cct gct gcc gtg atc cta cag ccg agt tca tcc cct gta act	1152
Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser Pro Val Thr	
370 375 380	
gtc att gac atg cac aaa tgt gtg agt ctg cag ctt gac gat ctc aga	1200
Val Ile Asp Met His Lys Cys Val Ser Leu Gln Leu Asp Asp Leu Arg	
385 390 395 400	
ttc acc atc act caa ttg gcc aat gta acc tac aat agc acc atc aag	1248
Phe Thr Ile Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser Thr Ile Lys	
405 410 415	
ctt gaa aca tcc cag atc ttg cct att gat ccg ttg gat ata tcc cag	1296
Leu Glu Thr Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp Ile Ser Gln	
420 425 430	
aat tta gct gcg gtg aat aag agt cta agt gat gca cta caa cac tta	1344
Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu Gln His Leu	
435 440 445	
gca caa agt gac aca tac ctt tct gca atc aca tca gct acg act aca	1392
Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala Thr Thr Thr	
450 455 460	
agt gta tta tcc ata ata gca atc tgt ctt gga tcg tta ggt tta ata	1440
Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu Gly Leu Ile	
465 470 475 480	
tta ata atc ttg ctc agt gta gtt gtg tgg aag tta ttg acc att gtc	1488
Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu Thr Ile Val	
485 490 495	
gct gct aat cga aat aga atg gag aat ttt gtt tat cat aat tca gca	1536
Ala Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His Asn Ser Ala	
500 505 510	
ttc cac cac cca cga tct gat ctc agt gag aaa aat caa cct gca act	1584
Phe His His Pro Arg Ser Asp Leu Ser Glu Lys Asn Gln Pro Ala Thr	
515 520 525	
ctt gga aca aga	1596
Leu Gly Thr Arg	
530	

<210> 14

<211> 532

<212> PRT

<213> Cryptovirus F0 protein

<400> 14

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Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro Thr Asn Val
 1          5          10          15
Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe Ile Val Val
          20          25          30
Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys Asn Ile Thr
          35          40          45
Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu Gln Pro Ile
          50          55          60
Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro Thr Arg Arg
65          70          75          80
Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala Leu Gly Val
          85          90          95
Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val Lys Ala Asn
          100          105          110
Glu Asn Thr Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile Gln Lys Thr
          115          120          125
Asn Ala Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser Leu Gly Thr
130          135          140
Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Ile Ser Pro Ala
145          150          155          160
Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile Gly Ser Ile
          165          170          175
Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His Asn Gln Ile
          180          185          190
Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu Arg Ile Leu
          195          200          205
Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe Asn Thr Gln
          210          215          220
Ile Ser Ala Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr Gly Gln Ile
225          230          235          240
Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys Ile Glu Leu
          245          250          255
Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp Leu Ala Thr
          260          265          270
Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln Leu Pro Thr
          275          280          285
Arg Val Ile Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro Ala Ser Gln
          290          295          300
Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn Asp Ala Gln
305          310          315          320
Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn Leu Thr Arg
          325          330          335
Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg Phe Val Leu
          340          345          350
Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu Cys Lys Cys
          355          360          365
Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser Pro Val Thr
          370          375          380
Val Ile Asp Met His Lys Cys Val Ser Leu Gln Leu Asp Asp Leu Arg
385          390          395          400

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Phe Thr Ile Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser Thr Ile Lys
 405 410 415
 Leu Glu Thr Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp Ile Ser Gln
 420 425 430
 Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu Gln His Leu
 435 440 445
 Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala Thr Thr Thr
 450 455 460
 Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu Gly Leu Ile
 465 470 475 480
 Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu Thr Ile Val
 485 490 495
 Ala Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His Asn Ser Ala
 500 505 510
 Phe His His Pro Arg Ser Asp Leu Ser Glu Lys Asn Gln Pro Ala Thr
 515 520 525
 Leu Gly Thr Arg
 530

<210> 15

<211> 249

<212> DNA

<213> Cryptovirus

<220>

<221> CDS

<222> (1)...(249)

<223> Cryptovirus F2 coding sequence; cDNA in mRNA sense

<400> 15

ctt gat cca gca gcc ctc atg caa atc ggt gtc att cca aca aat gtc	48
Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro Thr Asn Val	
1 5 10 15	
cgg caa ctt atg tat tat act gag gcc tca tca gca ttc att gtt gtg	96
Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe Ile Val Val	
20 25 30	
aag tta atg cct aca att gac tgg ccg att agt gga tgt aat ata aca	144
Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys Asn Ile Thr	
35 40 45	
tca att tca agc tat aat gca aca gtg aca aaa ctc cta cag ccg atc	192
Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu Gln Pro Ile	
50 55 60	
ggt gag aat ttg gag acg att agg aac cag ttg att cca act cgg agg	240
Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro Thr Arg Arg	
65 70 75 80	
aga cgc cgg	249
Arg Arg Arg	

<210> 16
 <211> 83
 <212> PRT
 <213> Cryptovirus F2 protein

<400> 16
 Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro Thr Asn Val
 1 5 10 15
 Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe Ile Val Val
 20 25 30
 Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys Asn Ile Thr
 35 40 45
 Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu Gln Pro Ile
 50 55 60
 Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro Thr Arg Arg
 65 70 75 80
 Arg Arg Arg

<210> 17
 <211> 1347
 <212> DNA
 <213> Cryptovirus

<220>
 <221> CDS
 <222> (1)...(1347)
 <223> Cryptovirus F1 protein encoding sequence; cDNA in
 mRNA sense

<400> 17
 ttt gca ggg gtg gtg att gga tta gct gca tta gga gta gct act gcc 48
 Phe Ala Gly Val Val Ile Gly Leu Ala Ala Leu Gly Val Ala Thr Ala
 1 5 10 15
 gca caa gtc act gcc gca gta gca cta gtt aag gca aat gaa aat act 96
 Ala Gln Val Thr Ala Ala Val Ala Leu Val Lys Ala Asn Glu Asn Thr
 20 25 30
 gcg gct ata ctc aat ctc aaa aat gca atc caa aaa aca aat gca gca 144
 Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile Gln Lys Thr Asn Ala Ala
 35 40 45
 gtt gca gat gtg gtc cag gcc aca caa tca cta gga acg gca gtt caa 192
 Val Ala Asp Val Val Gln Ala Thr Gln Ser Leu Gly Thr Ala Val Gln
 50 55 60
 gca gtt caa gat cac ata aac agt gtg ata agt cca gca att aca gca 240
 Ala Val Gln Asp His Ile Asn Ser Val Ile Ser Pro Ala Ile Thr Ala
 65 70 75 80
 gcc aat tgt aag gcc caa gat gct atc att gcc tca atc ctc aat ctc 288
 Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile Gly Ser Ile Leu Asn Leu

	85	90	95	
tat ttg acc gag ttg aca act atc ttc cac aat caa att aca aac cct				336
Tyr Leu Thr Glu Leu Thr Thr Ile Phe His Asn Gln Ile Thr Asn Pro				
	100	105	110	
gca ttg agt cct att aca att caa gct tta agg atc cta ctg ggg agt				384
Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu Arg Ile Leu Leu Gly Ser				
	115	120	125	
acc ttg ccg act gtg gtc gaa aaa tct ttc aat acc cag ata agt gca				432
Thr Leu Pro Thr Val Val Glu Lys Ser Phe Asn Thr Gln Ile Ser Ala				
	130	135	140	
gct gag ctt ctc tca tca ggg ttg ttg aca ggc cag att gtg gga tta				480
Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr Gly Gln Ile Val Gly Leu				
	145	150	155	160
gat ttg acc tat atg cag atg gtc ata aaa att gag ctg cca act tta				528
Asp Leu Thr Tyr Met Gln Met Val Ile Lys Ile Glu Leu Pro Thr Leu				
	165	170	175	
act gta caa cct gca acc cag atc ata gat ctg gcc acc att tct gca				576
Thr Val Gln Pro Ala Thr Gln Ile Ile Asp Leu Ala Thr Ile Ser Ala				
	180	185	190	
ttc att aac aat caa gaa gtc atg gcc caa tta cca aca cgt gtt att				624
Phe Ile Asn Asn Gln Glu Val Met Ala Gln Leu Pro Thr Arg Val Ile				
	195	200	205	
gtg act ggc agc ttg atc caa gcc tat ccc gca tcg caa tgc act att				672
Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro Ala Ser Gln Cys Thr Ile				
	210	215	220	
aca ccc aac act gtg tac tgt agg tat aat gat gcc caa gta ctc tca				720
Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn Asp Ala Gln Val Leu Ser				
	225	230	235	240
gat gat acg atg gct tgc ctc caa ggt aac ttg aca aga tgc acc ttc				768
Asp Asp Thr Met Ala Cys Leu Gln Gly Asn Leu Thr Arg Cys Thr Phe				
	245	250	255	
tct cca gtg gtt ggg agc ttt ctc act cga ttc gtg ctg ttc gat gga				816
Ser Pro Val Val Gly Ser Phe Leu Thr Arg Phe Val Leu Phe Asp Gly				
	260	265	270	
ata gtt tat gca aat tgc agg tcg atg ctg tgc aag tgc atg caa cct				864
Ile Val Tyr Ala Asn Cys Arg Ser Met Leu Cys Lys Cys Met Gln Pro				
	275	280	285	
gct gcc gtg atc cta cag ccg agt tca tcc cct gta act gtc att gac				912
Ala Ala Val Ile Leu Gln Pro Ser Ser Ser Pro Val Thr Val Ile Asp				
	290	295	300	
atg cac aaa tgt gtg agt ctg cag ctt gac gat ctc aga ttc acc atc				960

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Met His Lys Cys Val Ser Leu Gln Leu Asp Asp Leu Arg Phe Thr Ile
305                      310                      315                      320

act caa ttg gcc aat gta acc tac aat agc acc atc aag ctt gaa aca 1008
Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser Thr Ile Lys Leu Glu Thr
                      325                      330                      335

tcc cag atc ttg cct att gat ccg ttg gat ata tcc cag aat tta gct 1056
Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp Ile Ser Gln Asn Leu Ala
                      340                      345                      350

gcg gtg aat aag agt cta agt gat gca cta caa cac tta gca caa agt 1104
Ala Val Asn Lys Ser Leu Ser Asp Ala Leu Gln His Leu Ala Gln Ser
                      355                      360                      365

gac aca tac ctt tct gca atc aca tca gct acg act aca agt gta tta 1152
Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala Thr Thr Ser Val Leu
                      370                      375                      380

tcc ata ata gca atc tgt ctt gga tcg tta ggt tta ata tta ata atc 1200
Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu Gly Leu Ile Leu Ile Ile
385                      390                      395                      400

ttg ctc agt gta gtt gtg tgg aag tta ttg acc att gtc gct gct aat 1248
Leu Leu Ser Val Val Val Trp Lys Leu Leu Thr Ile Val Ala Ala Asn
                      405                      410                      415

cga aat aga atg gag aat ttt gtt tat cat aat tca gca ttc cac cac 1296
Arg Asn Arg Met Glu Asn Phe Val Tyr His Asn Ser Ala Phe His His
                      420                      425                      430

cca cga tct gat ctc agt gag aaa aat caa cct gca act ctt gga aca 1344
Pro Arg Ser Asp Leu Ser Glu Lys Asn Gln Pro Ala Thr Leu Gly Thr
                      435                      440                      445

aga
Arg
1347

```

<210> 18
 <211> 449
 <212> PRT
 <213> Cryptovirus

```

<400> 18
Phe Ala Gly Val Val Ile Gly Leu Ala Ala Leu Gly Val Ala Thr Ala
1      5      10      15
Ala Gln Val Thr Ala Ala Val Ala Leu Val Lys Ala Asn Glu Asn Thr
20      25      30
Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile Gln Lys Thr Asn Ala Ala
35      40      45
Val Ala Asp Val Val Gln Ala Thr Gln Ser Leu Gly Thr Ala Val Gln
50      55      60
Ala Val Gln Asp His Ile Asn Ser Val Ile Ser Pro Ala Ile Thr Ala

```

65					70					75				80	
Ala	Asn	Cys	Lys	Ala	Gln	Asp	Ala	Ile	Ile	Gly	Ser	Ile	Leu	Asn	Leu
				85					90					95	
Tyr	Leu	Thr	Glu	Leu	Thr	Thr	Ile	Phe	His	Asn	Gln	Ile	Thr	Asn	Pro
			100					105					110		
Ala	Leu	Ser	Pro	Ile	Thr	Ile	Gln	Ala	Leu	Arg	Ile	Leu	Leu	Gly	Ser
			115					120					125		
Thr	Leu	Pro	Thr	Val	Val	Glu	Lys	Ser	Phe	Asn	Thr	Gln	Ile	Ser	Ala
			130				135					140			
Ala	Glu	Leu	Leu	Ser	Ser	Gly	Leu	Leu	Thr	Gly	Gln	Ile	Val	Gly	Leu
145					150					155					160
Asp	Leu	Thr	Tyr	Met	Gln	Met	Val	Ile	Lys	Ile	Glu	Leu	Pro	Thr	Leu
				165					170					175	
Thr	Val	Gln	Pro	Ala	Thr	Gln	Ile	Ile	Asp	Leu	Ala	Thr	Ile	Ser	Ala
			180					185					190		
Phe	Ile	Asn	Asn	Gln	Glu	Val	Met	Ala	Gln	Leu	Pro	Thr	Arg	Val	Ile
		195					200					205			
Val	Thr	Gly	Ser	Leu	Ile	Gln	Ala	Tyr	Pro	Ala	Ser	Gln	Cys	Thr	Ile
	210					215					220				
Thr	Pro	Asn	Thr	Val	Tyr	Cys	Arg	Tyr	Asn	Asp	Ala	Gln	Val	Leu	Ser
225				230						235					240
Asp	Asp	Thr	Met	Ala	Cys	Leu	Gln	Gly	Asn	Leu	Thr	Arg	Cys	Thr	Phe
			245						250					255	
Ser	Pro	Val	Val	Gly	Ser	Phe	Leu	Thr	Arg	Phe	Val	Leu	Phe	Asp	Gly
			260					265					270		
Ile	Val	Tyr	Ala	Asn	Cys	Arg	Ser	Met	Leu	Cys	Lys	Cys	Met	Gln	Pro
		275					280					285			
Ala	Ala	Val	Ile	Leu	Gln	Pro	Ser	Ser	Ser	Pro	Val	Thr	Val	Ile	Asp
	290					295					300				
Met	His	Lys	Cys	Val	Ser	Leu	Gln	Leu	Asp	Asp	Leu	Arg	Phe	Thr	Ile
305				310						315					320
Thr	Gln	Leu	Ala	Asn	Val	Thr	Tyr	Asn	Ser	Thr	Ile	Lys	Leu	Glu	Thr
			325						330					335	
Ser	Gln	Ile	Leu	Pro	Ile	Asp	Pro	Leu	Asp	Ile	Ser	Gln	Asn	Leu	Ala
			340					345					350		
Ala	Val	Asn	Lys	Ser	Leu	Ser	Asp	Ala	Leu	Gln	His	Leu	Ala	Gln	Ser
		355					360					365			
Asp	Thr	Tyr	Leu	Ser	Ala	Ile	Thr	Ser	Ala	Thr	Thr	Thr	Ser	Val	Leu
	370					375						380			
Ser	Ile	Ile	Ala	Ile	Cys	Leu	Gly	Ser	Leu	Gly	Leu	Ile	Leu	Ile	Ile
385				390						395					400
Leu	Leu	Ser	Val	Val	Val	Trp	Lys	Leu	Leu	Thr	Ile	Val	Ala	Ala	Asn
			405						410					415	
Arg	Asn	Arg	Met	Glu	Asn	Phe	Val	Tyr	His	Asn	Ser	Ala	Phe	His	His
			420					425					430		
Pro	Arg	Ser	Asp	Leu	Ser	Glu	Lys	Asn	Gln	Pro	Ala	Thr	Leu	Gly	Thr
		435					440					445			

Arg

<210> 19

<211> 132

<212> DNA

<213> Cryptovirus

<220>

<221> CDS

<222> (1)...(132)

<223> Cryptovirus SH protein encoding sequence; cDNA in mRNA sense

<400> 19

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atg ctg cct gat ccg gaa gat ccg gaa agc aaa aaa gct aca agg aga 48
Met Leu Pro Asp Pro Glu Asp Pro Glu Ser Lys Lys Ala Thr Arg Arg
 1           5           10           15

```

```

aca gga aac cta att atc tgc ttc cta ttc atc ttc ttt ctg ttt gta 96
Thr Gly Asn Leu Ile Ile Cys Phe Leu Phe Ile Phe Phe Leu Phe Val
          20           25           30

```

```

acc ctc att gtt cca act cta aga cac ttg cta tct 132
Thr Leu Ile Val Pro Thr Leu Arg His Leu Leu Ser
          35           40

```

<210> 20

<211> 44

<212> PRT

<213> Cryptovirus

<400> 20

```

Met Leu Pro Asp Pro Glu Asp Pro Glu Ser Lys Lys Ala Thr Arg Arg
 1           5           10           15

```

```

Thr Gly Asn Leu Ile Ile Cys Phe Leu Phe Ile Phe Phe Leu Phe Val
          20           25           30

```

```

Thr Leu Ile Val Pro Thr Leu Arg His Leu Leu Ser
          35           40

```

<210> 21

<211> 1695

<212> DNA

<213> Cryptovirus

<220>

<221> CDS

<222> (1)...(1695)

<223> Cryptovirus HN protein encoding sequence; cDNA in mRNA sense

<400> 21

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atg att gca gaa gat gcc cct gtt aag ggc act tgc cga gta tta ttt 48
Met Ile Ala Glu Asp Ala Pro Val Lys Gly Thr Cys Arg Val Leu Phe
 1           5           10           15

```

```

cgg aca aca act tta att ttt cta tgc aca cta cta gca tta agc atc 96
Arg Thr Thr Thr Leu Ile Phe Leu Cys Thr Leu Leu Ala Leu Ser Ile
          20           25           30

```

tct atc ctt tat gag agt tta ata acc caa aag caa atc atg agt caa	144
Ser Ile Leu Tyr Glu Ser Leu Ile Thr Gln Lys Gln Ile Met Ser Gln	
35 40 45	
gca ggc tca act gga tct aat tct gga tta gga ggt att act gat ctt	192
Ala Gly Ser Thr Gly Ser Asn Ser Gly Leu Gly Gly Ile Thr Asp Leu	
50 55 60	
ctt aat aat att ctt tct gtc gca aat cag att ata tat aac tct gca	240
Leu Asn Asn Ile Leu Ser Val Ala Asn Gln Ile Ile Tyr Asn Ser Ala	
65 70 75 80	
gtc gct cta cct cta caa ttg gac act ctt gaa tca aca ctc ctt aca	288
Val Ala Leu Pro Leu Gln Leu Asp Thr Leu Glu Ser Thr Leu Leu Thr	
85 90 95	
gcc att aag tct ctt caa acc agt gac aag cta gaa cag aac tgc tgc	336
Ala Ile Lys Ser Leu Gln Thr Ser Asp Lys Leu Glu Gln Asn Cys Ser	
100 105 110	
tgg ggt gct gca ctg att aat gat aat aga tac att aat ggc atc aat	384
Trp Gly Ala Ala Leu Ile Asn Asp Asn Arg Tyr Ile Asn Gly Ile Asn	
115 120 125	
cag ttc tat ttc tca att gct gag ggt cgc aat ctg aca ctt ggc cca	432
Gln Phe Tyr Phe Ser Ile Ala Glu Gly Arg Asn Leu Thr Leu Gly Pro	
130 135 140	
ctt ctt aat ata cct agt ttc att cca act gcc acg aca cca gag ggc	480
Leu Leu Asn Ile Pro Ser Phe Ile Pro Thr Ala Thr Thr Pro Glu Gly	
145 150 155 160	
tgc acc agg atc cca tca ttc tgc ctc act aag aca cac tgg tgt tat	528
Cys Thr Arg Ile Pro Ser Phe Ser Leu Thr Lys Thr His Trp Cys Tyr	
165 170 175	
acg cac aat gtt atc ctg aat gga tgc cag gat cat gta tcc tca aat	576
Thr His Asn Val Ile Leu Asn Gly Cys Gln Asp His Val Ser Ser Asn	
180 185 190	
caa ttt gtt tcc atg gga atc att gaa ccc act tct gcc ggg ttt cca	624
Gln Phe Val Ser Met Gly Ile Ile Glu Pro Thr Ser Ala Gly Phe Pro	
195 200 205	
tcc ttt cga acc tta aag act cta tat ctc agc gat ggg gtc aat cgt	672
Ser Phe Arg Thr Leu Lys Thr Leu Tyr Leu Ser Asp Gly Val Asn Arg	
210 215 220	
aag agc tgc tct atc agt aca gtt ccg ggg ggt tgt atg atg tac tgt	720
Lys Ser Cys Ser Ile Ser Thr Val Pro Gly Gly Cys Met Met Tyr Cys	
225 230 235 240	
ttt gtc tct act caa cca gag agg gat gac tac ttt tct acc gct cct	768
Phe Val Ser Thr Gln Pro Glu Arg Asp Asp Tyr Phe Ser Thr Ala Pro	
245 250 255	

cca gaa caa cga att att ata atg tac tat aat gat aca atc gtg gag	816
Pro Glu Gln Arg Ile Ile Ile Met Tyr Tyr Asn Asp Thr Ile Val Glu	
260 265 270	
cgc ata att aat cca ccc ggg gta cta gac gta tgg gca aca ttg aac	864
Arg Ile Ile Asn Pro Pro Gly Val Leu Asp Val Trp Ala Thr Leu Asn	
275 280 285	
cca gga aca gga agc ggg gta tat tat tta ggt tgg gtg ctc ttt cca	912
Pro Gly Thr Gly Ser Gly Val Tyr Tyr Leu Gly Trp Val Leu Phe Pro	
290 295 300	
ata tat ggc ggc gtg att aaa aat acg agt tta tgg aat aat caa gca	960
Ile Tyr Gly Gly Val Ile Lys Asn Thr Ser Leu Trp Asn Asn Gln Ala	
305 310 315 320	
aat aaa tac ttc att ccc cag atg gtt gct gct ctc tgc tca caa aac	1008
Asn Lys Tyr Phe Ile Pro Gln Met Val Ala Ala Leu Cys Ser Gln Asn	
325 330 335	
cag gca act caa gtc caa aat gct aag tca tca tac tat agc agc tgg	1056
Gln Ala Thr Gln Val Gln Asn Ala Lys Ser Ser Tyr Tyr Ser Ser Trp	
340 345 350	
ttt ggc aat cga atg att cag tct ggg atc ctg gca tgc cct ctt caa	1104
Phe Gly Asn Arg Met Ile Gln Ser Gly Ile Leu Ala Cys Pro Leu Gln	
355 360 365	
cag gat cta acc aat gag tgt tta gtt ctg ccc ttt tct aat gat cag	1152
Gln Asp Leu Thr Asn Glu Cys Leu Val Leu Pro Phe Ser Asn Asp Gln	
370 375 380	
gtg ctt atg ggt gct gaa ggg aga tta tat atg tat ggt gac tcg gtg	1200
Val Leu Met Gly Ala Glu Gly Arg Leu Tyr Met Tyr Gly Asp Ser Val	
385 390 395 400	
tat tac tac caa aga agc aat agt tgg tgg cct atg acc atg ctg tat	1248
Tyr Tyr Tyr Gln Arg Ser Asn Ser Trp Trp Pro Met Thr Met Leu Tyr	
405 410 415	
aag gta acc ata aca ttc act aat ggt cag cca tcc gct ata tca gct	1296
Lys Val Thr Ile Thr Phe Thr Asn Gly Gln Pro Ser Ala Ile Ser Ala	
420 425 430	
cag aat gtg ccc aca cag cag gtc cct aga cct ggg aca gga gac tgc	1344
Gln Asn Val Pro Thr Gln Gln Val Pro Arg Pro Gly Thr Gly Asp Cys	
435 440 445	
ttt gca acc aat aga tgt ccc ggt ttt tgc ttg aca gga gtg tat gct	1392
Phe Ala Thr Asn Arg Cys Pro Gly Phe Cys Leu Thr Gly Val Tyr Ala	
450 455 460	
gat gct tgg tta ctg acc aac cct tcg tct acc agt aca ttt gga tcg	1440
Asp Ala Trp Leu Leu Thr Asn Pro Ser Ser Thr Ser Thr Phe Gly Ser	

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465                               470                               475                               480
gaa gca acc ttc act ggt tct tat ctc aac gca gca act cag cgt atc 1488
Glu Ala Thr Phe Thr Gly Ser Tyr Leu Asn Ala Ala Thr Gln Arg Ile
                               485                               490                               495

aat ccg acg atg tat atc gcg aac aac aca cag atc ata agc tca cag 1536
Asn Pro Thr Met Tyr Ile Ala Asn Asn Thr Gln Ile Ile Ser Ser Gln
                               500                               505                               510

caa ttt gga tca agc ggt caa gaa gca gca tat ggc cac aca act tgt 1584
Gln Phe Gly Ser Ser Gly Gln Glu Ala Ala Tyr Gly His Thr Thr Cys
                               515                               520                               525

ttt agg gac aca ggc tct gtt atg gta tac tgt atc tat att att gaa 1632
Phe Arg Asp Thr Gly Ser Val Met Val Tyr Cys Ile Tyr Ile Ile Glu
                               530                               535                               540

ttg tcc tca tct ctc tta gga caa ttt cag att gtc cca ttt atc cgt 1680
Leu Ser Ser Ser Leu Leu Gly Gln Phe Gln Ile Val Pro Phe Ile Arg
545                               550                               555                               560

cag gtg aca cta tcc 1695
Gln Val Thr Leu Ser
                               565

```

<210> 22
 <211> 565
 <212> PRT
 <213> Cryptovirus

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<400> 22
Met Ile Ala Glu Asp Ala Pro Val Lys Gly Thr Cys Arg Val Leu Phe
 1                               5                               10                               15
Arg Thr Thr Thr Leu Ile Phe Leu Cys Thr Leu Leu Ala Leu Ser Ile
                20                               25                               30
Ser Ile Leu Tyr Glu Ser Leu Ile Thr Gln Lys Gln Ile Met Ser Gln
                35                               40                               45
Ala Gly Ser Thr Gly Ser Asn Ser Gly Leu Gly Gly Ile Thr Asp Leu
                50                               55                               60
Leu Asn Asn Ile Leu Ser Val Ala Asn Gln Ile Ile Tyr Asn Ser Ala
65                               70                               75                               80
Val Ala Leu Pro Leu Gln Leu Asp Thr Leu Glu Ser Thr Leu Leu Thr
                85                               90                               95
Ala Ile Lys Ser Leu Gln Thr Ser Asp Lys Leu Glu Gln Asn Cys Ser
                100                               105                               110
Trp Gly Ala Ala Leu Ile Asn Asp Asn Arg Tyr Ile Asn Gly Ile Asn
                115                               120                               125
Gln Phe Tyr Phe Ser Ile Ala Glu Gly Arg Asn Leu Thr Leu Gly Pro
                130                               135                               140
Leu Leu Asn Ile Pro Ser Phe Ile Pro Thr Ala Thr Thr Pro Glu Gly
145                               150                               155                               160
Cys Thr Arg Ile Pro Ser Phe Ser Leu Thr Lys Thr His Trp Cys Tyr
                165                               170                               175

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Thr His Asn Val Ile Leu Asn Gly Cys Gln Asp His Val Ser Ser Asn
      180      185      190
Gln Phe Val Ser Met Gly Ile Ile Glu Pro Thr Ser Ala Gly Phe Pro
      195      200      205
Ser Phe Arg Thr Leu Lys Thr Leu Tyr Leu Ser Asp Gly Val Asn Arg
      210      215      220
Lys Ser Cys Ser Ile Ser Thr Val Pro Gly Gly Cys Met Met Tyr Cys
      225      230      235      240
Phe Val Ser Thr Gln Pro Glu Arg Asp Asp Tyr Phe Ser Thr Ala Pro
      245      250      255
Pro Glu Gln Arg Ile Ile Ile Met Tyr Tyr Asn Asp Thr Ile Val Glu
      260      265      270
Arg Ile Ile Asn Pro Pro Gly Val Leu Asp Val Trp Ala Thr Leu Asn
      275      280      285
Pro Gly Thr Gly Ser Gly Val Tyr Tyr Leu Gly Trp Val Leu Phe Pro
      290      295      300
Ile Tyr Gly Gly Val Ile Lys Asn Thr Ser Leu Trp Asn Asn Gln Ala
      305      310      315      320
Asn Lys Tyr Phe Ile Pro Gln Met Val Ala Ala Leu Cys Ser Gln Asn
      325      330      335
Gln Ala Thr Gln Val Gln Asn Ala Lys Ser Ser Tyr Tyr Ser Ser Trp
      340      345      350
Phe Gly Asn Arg Met Ile Gln Ser Gly Ile Leu Ala Cys Pro Leu Gln
      355      360      365
Gln Asp Leu Thr Asn Glu Cys Leu Val Leu Pro Phe Ser Asn Asp Gln
      370      375      380
Val Leu Met Gly Ala Glu Gly Arg Leu Tyr Met Tyr Gly Asp Ser Val
      385      390      395      400
Tyr Tyr Tyr Gln Arg Ser Asn Ser Trp Trp Pro Met Thr Met Leu Tyr
      405      410      415
Lys Val Thr Ile Thr Phe Thr Asn Gly Gln Pro Ser Ala Ile Ser Ala
      420      425      430
Gln Asn Val Pro Thr Gln Gln Val Pro Arg Pro Gly Thr Gly Asp Cys
      435      440      445
Phe Ala Thr Asn Arg Cys Pro Gly Phe Cys Leu Thr Gly Val Tyr Ala
      450      455      460
Asp Ala Trp Leu Leu Thr Asn Pro Ser Ser Thr Ser Thr Phe Gly Ser
      465      470      475      480
Glu Ala Thr Phe Thr Gly Ser Tyr Leu Asn Ala Ala Thr Gln Arg Ile
      485      490      495
Asn Pro Thr Met Tyr Ile Ala Asn Asn Thr Gln Ile Ile Ser Ser Gln
      500      505      510
Gln Phe Gly Ser Ser Gly Gln Glu Ala Ala Tyr Gly His Thr Thr Cys
      515      520      525
Phe Arg Asp Thr Gly Ser Val Met Val Tyr Cys Ile Tyr Ile Ile Glu
      530      535      540
Leu Ser Ser Ser Leu Leu Gly Gln Phe Gln Ile Val Pro Phe Ile Arg
      545      550      555      560
Gln Val Thr Leu Ser
      565

```

<210> 23

<211> 6765

<212> DNA

<213> Cryptovirus

<220>

<221> CDS

<222> (1)...(6765)

<223> Cryptovirus L protein encoding sequence; cDNA in mRNA sense

<400> 23

atg gct ggg tct cgg gag ata tta ctc cct gaa gtc cat ctc aat tca	48
Met Ala Gly Ser Arg Glu Ile Leu Leu Pro Glu Val His Leu Asn Ser	
1 5 10 15	

cca att gta aag cat aag cta tac tat tac att cta ctt gga aac ctc	96
Pro Ile Val Lys His Lys Leu Tyr Tyr Tyr Ile Leu Leu Gly Asn Leu	
20 25 30	

cca aat gag atc gac att gac gat tta ggt cca tta cat aat caa aat	144
Pro Asn Glu Ile Asp Ile Asp Asp Leu Gly Pro Leu His Asn Gln Asn	
35 40 45	

tgg aat caa ata gca cat gaa gag tct aac tta gct caa cgc ttg gta	192
Trp Asn Gln Ile Ala His Glu Glu Ser Asn Leu Ala Gln Arg Leu Val	
50 55 60	

aat gta aga aat ttt cta att acc cac atc cct gat ctt aga aag ggc	240
Asn Val Arg Asn Phe Leu Ile Thr His Ile Pro Asp Leu Arg Lys Gly	
65 70 75 80	

cat tgg caa gag tat gta aat gta ata ctg tgg ccg cga att ctt ccc	288
His Trp Gln Glu Tyr Val Asn Val Ile Leu Trp Pro Arg Ile Leu Pro	
85 90 95	

ttg atc ccg gat ttt aaa atc aat gac caa ttg cct ctg ctc aaa aat	336
Leu Ile Pro Asp Phe Lys Ile Asn Asp Gln Leu Pro Leu Leu Lys Asn	
100 105 110	

tgg gac aag tta gtt aaa gaa tca tgt tca gta atc aat gcg ggt act	384
Trp Asp Lys Leu Val Lys Glu Ser Cys Ser Val Ile Asn Ala Gly Thr	
115 120 125	

tcc cag tgc att cag aat ctc agc tat gga ctg aca ggt cgt ggg aac	432
Ser Gln Cys Ile Gln Asn Leu Ser Tyr Gly Leu Thr Gly Arg Gly Asn	
130 135 140	

ctc ttt aca cga tca cgt gaa ctc tct ggt gac cgc agg gat att gat	480
Leu Phe Thr Arg Ser Arg Glu Leu Ser Gly Asp Arg Arg Asp Ile Asp	
145 150 155 160	

ctt aag acg gtt gtg gca gca tgg cat gac tca gac tgg aaa aga ata	528
Leu Lys Thr Val Val Ala Ala Trp His Asp Ser Asp Trp Lys Arg Ile	
165 170 175	

agt gat ttt tgg att atg atc aaa ttc cag atg aga caa tta att gtt	576
Ser Asp Phe Trp Ile Met Ile Lys Phe Gln Met Arg Gln Leu Ile Val	

180	185	190	
agg caa aca gat cat aat gat cct gat tta atc acg tat atc gaa aat			624
Arg Gln Thr Asp His Asn Asp Pro Asp Leu Ile Thr Tyr Ile Glu Asn			
195	200	205	
aga gaa ggc ata atc atc ata acc cct gaa ctg gta gca tta ttt aat			672
Arg Glu Gly Ile Ile Ile Ile Thr Pro Glu Leu Val Ala Leu Phe Asn			
210	215	220	
act gag aat cat aca cta aca tac atg acc ttt gaa att gta ctg atg			720
Thr Glu Asn His Thr Leu Thr Tyr Met Thr Phe Glu Ile Val Leu Met			
225	230	235	240
gtt tca gat atg tac gaa ggt cgt cac aac att tta tca cta tgc aca			768
Val Ser Asp Met Tyr Glu Gly Arg His Asn Ile Leu Ser Leu Cys Thr			
245	250	255	
gtt agc act tac ctg aat cct ctg aag aaa aga ata aca tat tta ttg			816
Val Ser Thr Tyr Leu Asn Pro Leu Lys Lys Arg Ile Thr Tyr Leu Leu			
260	265	270	
agc ctt gta gat aac tta gct ttt cag ata ggt gat gct gta tat aac			864
Ser Leu Val Asp Asn Leu Ala Phe Gln Ile Gly Asp Ala Val Tyr Asn			
275	280	285	
ata att gct ttg cta gaa tcc ttt gta tat gca cag ttg caa atg tca			912
Ile Ile Ala Leu Leu Glu Ser Phe Val Tyr Ala Gln Leu Gln Met Ser			
290	295	300	
gat ccc atc cca gaa ctc aga gga caa ttc cat gca ttc gta tgt tct			960
Asp Pro Ile Pro Glu Leu Arg Gly Gln Phe His Ala Phe Val Cys Ser			
305	310	315	320
gag att ctt gat gca cta agg gga act aat agt ttc acc cag gat gaa			1008
Glu Ile Leu Asp Ala Leu Arg Gly Thr Asn Ser Phe Thr Gln Asp Glu			
325	330	335	
tta aga act gtg aca act aat ttg ata tcc cca ttc caa gat ctg acc			1056
Leu Arg Thr Val Thr Thr Asn Leu Ile Ser Pro Phe Gln Asp Leu Thr			
340	345	350	
cca gat ctt acg gct gaa ttg ctc tgt ata atg agg ctt tgg gga cac			1104
Pro Asp Leu Thr Ala Glu Leu Leu Cys Ile Met Arg Leu Trp Gly His			
355	360	365	
ccc atg ctc act gcc agt caa gct gcg gga aag gta cgc gag tct atg			1152
Pro Met Leu Thr Ala Ser Gln Ala Ala Gly Lys Val Arg Glu Ser Met			
370	375	380	
tgt gct ggg aaa gta ctg gac ttt ccc act att atg aaa aca cta gcc			1200
Cys Ala Gly Lys Val Leu Asp Phe Pro Thr Ile Met Lys Thr Leu Ala			
385	390	395	400
ttt ttc cat act att ctg atc aat gga tac agg agg aag cat cat gga			1248

Phe	Phe	His	Thr	Ile	Leu	Ile	Asn	Gly	Tyr	Arg	Arg	Lys	His	His	Gly	
				405					410						415	
gta	tgg	cca	ccc	tta	aac	tta	cca	ggt	aat	gct	tca	aag	ggt	ctc	aca	1296
Val	Trp	Pro	Pro	Leu	Asn	Leu	Pro	Gly	Asn	Ala	Ser	Lys	Gly	Leu	Thr	
				420				425					430			
gaa	ctt	atg	aat	gac	aac	act	gag	ata	agc	tat	gaa	ttc	aca	ctt	aag	1344
Glu	Leu	Met	Asn	Asp	Asn	Thr	Glu	Ile	Ser	Tyr	Glu	Phe	Thr	Leu	Lys	
			435				440					445				
cat	tgg	aag	gaa	atc	tct	ctt	ata	aaa	ttc	aag	aaa	tgt	ttt	gat	gca	1392
His	Trp	Lys	Glu	Ile	Ser	Leu	Ile	Lys	Phe	Lys	Lys	Cys	Phe	Asp	Ala	
			450				455					460				
gac	gca	ggt	gag	gaa	ctc	agt	ata	ttt	atg	aaa	gat	aaa	gca	att	agt	1440
Asp	Ala	Gly	Glu	Glu	Leu	Ser	Ile	Phe	Met	Lys	Asp	Lys	Ala	Ile	Ser	
					470					475					480	
gcc	cca	aaa	caa	gac	tgg	atg	agt	gtg	ttt	aga	aga	agc	cta	atc	aaa	1488
Ala	Pro	Lys	Gln	Asp	Trp	Met	Ser	Val	Phe	Arg	Arg	Ser	Leu	Ile	Lys	
				485					490					495		
cag	cgc	cat	cag	cat	cat	cag	gtc	ccc	cta	cca	aat	cca	ttc	aat	cga	1536
Gln	Arg	His	Gln	His	His	Gln	Val	Pro	Leu	Pro	Asn	Pro	Phe	Asn	Arg	
			500					505					510			
cgg	cta	ttg	cta	aac	ttt	ctc	gga	gat	gac	aaa	ttc	gac	ccg	aat	gtg	1584
Arg	Leu	Leu	Leu	Asn	Phe	Leu	Gly	Asp	Asp	Lys	Phe	Asp	Pro	Asn	Val	
			515				520					525				
gag	cta	cag	tat	gta	aca	tca	ggt	gag	tat	cta	cat	gat	gac	acg	ttt	1632
Glu	Leu	Gln	Tyr	Val	Thr	Ser	Gly	Glu	Tyr	Leu	His	Asp	Asp	Thr	Phe	
			530				535					540				
tgt	gca	tca	tat	tca	cta	aaa	gag	aag	gaa	att	aaa	cct	gat	ggt	cga	1680
Cys	Ala	Ser	Tyr	Ser	Leu	Lys	Glu	Lys	Glu	Ile	Lys	Pro	Asp	Gly	Arg	
					550					555					560	
att	ttt	gca	aag	ttg	act	aag	aga	atg	aga	tca	tgt	caa	gtt	ata	gca	1728
Ile	Phe	Ala	Lys	Leu	Thr	Lys	Arg	Met	Arg	Ser	Cys	Gln	Val	Ile	Ala	
				565					570					575		
gaa	tct	ctt	tta	gcg	aac	cat	gct	ggg	aag	tta	atg	aaa	gag	aat	ggt	1776
Glu	Ser	Leu	Leu	Ala	Asn	His	Ala	Gly	Lys	Leu	Met	Lys	Glu	Asn	Gly	
			580					585					590			
gtt	gtg	atg	aat	cag	cta	tca	tta	aca	aaa	tca	cta	tta	aca	atg	agt	1824
Val	Val	Met	Asn	Gln	Leu	Ser	Leu	Thr	Lys	Ser	Leu	Leu	Thr	Met	Ser	
			595				600					605				
cag	att	gga	ata	ata	tcc	gag	aga	gct	aga	aag	tcg	act	cga	gat	aac	1872
Gln	Ile	Gly	Ile	Ile	Ser	Glu	Arg	Ala	Arg	Lys	Ser	Thr	Arg	Asp	Asn	
			610			615					620					

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Ile Asn Arg Pro Gly Phe Gln Asn Ile Gln Arg Asn Lys Ser His His	
625 630 635 640	
toc aag caa gtc aat cag cga gat cca agt gat gac ttt gaa ttg gca	1968
Ser Lys Gln Val Asn Gln Arg Asp Pro Ser Asp Asp Phe Glu Leu Ala	
645 650 655	
gca tct ttt tta act act gat ctc aaa aaa tat tgt tta caa tgg agg	2016
Ala Ser Phe Leu Thr Thr Asp Leu Lys Lys Tyr Cys Leu Gln Trp Arg	
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tat cag aca att atc cca ttt gct caa tca cta aac aga atg tat ggt	2064
Tyr Gln Thr Ile Ile Pro Phe Ala Gln Ser Leu Asn Arg Met Tyr Gly	
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Tyr Pro His Leu Phe Glu Trp Ile His Leu Arg Leu Met Arg Ser Thr	
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Leu Tyr Val Gly Asp Pro Phe Asn Pro Pro Ala Asp Thr Ser Gln Phe	
705 710 715 720	
gat cta gat aaa gta att aat gga gat atc ttc att gta tca ccc aga	2208
Asp Leu Asp Lys Val Ile Asn Gly Asp Ile Phe Ile Val Ser Pro Arg	
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Gly Gly Ile Glu Gly Leu Cys Gln Lys Ala Trp Thr Met Ile Ser Ile	
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Ser Val Ile Ile Leu Ser Ala Thr Glu Ser Gly Thr Arg Val Met Ser	
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Arg Ser Leu Pro Thr Leu Glu Lys Lys Thr Ile Ala Phe Arg Ser Cys	
785 790 795 800	
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Asn Leu Phe Phe Glu Arg Leu Lys Cys Asn Asn Phe Gly Leu Gly His	
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cat ttg aaa gaa caa gag act atc att agt tct cac ttc ttt gtt tat	2496
His Leu Lys Glu Gln Glu Thr Ile Ile Ser Ser His Phe Phe Val Tyr	
820 825 830	
agc aag aga ata ttc tat cag ggg agg att cta acg caa gcc tta aaa	2544
Ser Lys Arg Ile Phe Tyr Gln Gly Arg Ile Leu Thr Gln Ala Leu Lys	
835 840 845	

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Asn Ala Ser Lys Leu Cys Leu Thr Ala Asp Val Leu Gly Glu Cys Thr	
850 855 860	
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Gln Ser Ser Cys Ser Asn Leu Ala Thr Thr Val Met Arg Leu Thr Glu	
865 870 875 880	
aat ggt gtt gaa aaa gat atc tgt ttc tac ttg aat atc tat atg acc	2688
Asn Gly Val Glu Lys Asp Ile Cys Phe Tyr Leu Asn Ile Tyr Met Thr	
885 890 895	
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Ile Lys Gln Leu Ser Tyr Asp Ile Ile Phe Pro Gln Val Ser Ile Pro	
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gga gat cag atc aca tta gaa tac ata aat aat cca cac ctg gta tca	2784
Gly Asp Gln Ile Thr Leu Glu Tyr Ile Asn Asn Pro His Leu Val Ser	
915 920 925	
cga ttg gct ctt tta cca tcc cag tta gga ggt cta aac tac ttg tca	2832
Arg Leu Ala Leu Leu Pro Ser Gln Leu Gly Gly Leu Asn Tyr Leu Ser	
930 935 940	
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Cys Ser Arg Leu Phe Asn Arg Asn Ile Gly Asp Pro Val Val Ser Ala	
945 950 955 960	
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Val Ala Asp Leu Lys Arg Leu Ile Lys Ser Gly Cys Met Asp Tyr Trp	
965 970 975	
atc ctt tat aac tta tta ggg aga aaa ccg gga aac ggc tca tgg gct	2976
Ile Leu Tyr Asn Leu Leu Gly Arg Lys Pro Gly Asn Gly Ser Trp Ala	
980 985 990	
act tta gca gct gac ccg tac tca atc aat ata gag tat caa tac ccc	3024
Thr Leu Ala Ala Asp Pro Tyr Ser Ile Asn Ile Glu Tyr Gln Tyr Pro	
995 1000 1005	
cca act aca gct ctt aag agg cac acc caa caa gtt ctg atg gaa ctc	3072
Pro Thr Thr Ala Leu Lys Arg His Thr Gln Gln Val Leu Met Glu Leu	
1010 1015 1020	
agt acg aat cca atg tta cgt ggc ata ttc tct gac aat gca cag gca	3120
Ser Thr Asn Pro Met Leu Arg Gly Ile Phe Ser Asp Asn Ala Gln Ala	
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gaa gaa aat aat ctt gct agg ttt ctc ctg gat agg gag gtg atc ttt	3168
Glu Glu Asn Asn Leu Ala Arg Phe Leu Leu Asp Arg Glu Val Ile Phe	
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ccg cgt gta gct cac atc atc att gag caa acc agt gtc ggg agg aga	3216
Pro Arg Val Ala His Ile Ile Ile Glu Gln Thr Ser Val Gly Arg Arg	

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Lys Gln Ile Gln Gly Tyr Leu Asp Ser Thr Arg Ser Ile Met Arg Lys			
1075	1080	1085	
tca ctg gaa att aag ccc ttg tcc aat agg aag ctt aat gaa ata ctg			3312
Ser Leu Glu Ile Lys Pro Leu Ser Asn Arg Lys Leu Asn Glu Ile Leu			
1090	1095	1100	
gat tac aac atc aat tac tta gct tac aat ttg gca tta ctc aag aat			3360
Asp Tyr Asn Ile Asn Tyr Leu Ala Tyr Asn Leu Ala Leu Leu Lys Asn			
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gct att gaa cct ccg act tat ttg aag gca atg acc ctt gaa aca tgt			3408
Ala Ile Glu Pro Thr Tyr Leu Lys Ala Met Thr Leu Glu Thr Cys			
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Ser Ile Asp Ile Ala Arg Ser Leu Arg Lys Leu Ser Trp Ala Pro Leu			
1140	1145	1150	
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Leu Gly Gly Arg Asn Leu Glu Gly Leu Glu Thr Pro Asp Pro Ile Glu			
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atc act gca gga gca tta att gtt gga tcg ggc tac tgt gaa cag tgt			3552
Ile Thr Ala Gly Ala Leu Ile Val Gly Ser Gly Tyr Cys Glu Gln Cys			
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Ala Ala Gly Asp Asn Arg Phe Thr Trp Phe Phe Leu Pro Ser Gly Ile			
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Glu Ile Gly Gly Asp Pro Arg Asp Asn Pro Pro Ile Arg Val Pro Tyr			
1205	1210	1215	
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Ile Gly Ser Arg Thr Asp Glu Arg Arg Val Ala Ser Met Ala Tyr Ile			
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agg ggt gcc tca agt agc cta aaa gca gtt ctt aga ctg gcg gga gtg			3744
Arg Gly Ala Ser Ser Ser Leu Lys Ala Val Leu Arg Leu Ala Gly Val			
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Tyr Ile Trp Ala Phe Gly Asp Thr Leu Glu Asn Trp Ile Asp Ala Leu			
1250	1255	1260	
gat ttg tct cac act aga gtt aac atc aca ctt gaa cag tta caa tcc			3840
Asp Leu Ser His Thr Arg Val Asn Ile Thr Leu Glu Gln Leu Gln Ser			
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Leu Thr Pro Leu Pro Thr Ser Ala Asn Leu Thr His Arg Leu Asp Asp	
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Ser Phe Thr His Ile Ser Asn Asp Glu Gln Tyr Leu Thr Ile Asn Asp	
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aaa act gca gat tca aat ata atc tac caa cag tta atg atc act gga	4032
Lys Thr Ala Asp Ser Asn Ile Ile Tyr Gln Gln Leu Met Ile Thr Gly	
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ctc ggg atc tta gaa aca tgg aat aat ccc cca atc aat aga aca ttc	4080
Leu Gly Ile Leu Glu Thr Trp Asn Asn Pro Pro Ile Asn Arg Thr Phe	
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gaa gaa tct acc cta cat ttg cac act ggt gca tca tgt tgt gtc cga	4128
Glu Glu Ser Thr Leu His Leu His Thr Gly Ala Ser Cys Cys Val Arg	
1365 1370 1375	
cct gtg gac tcc tgc att atc tca gaa gca tta aca gtc aag cca cat	4176
Pro Val Asp Ser Cys Ile Ile Ser Glu Ala Leu Thr Val Lys Pro His	
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att aca gta ccg tac agc aat aaa ttt gta ttt gat gaa gac ccg cta	4224
Ile Thr Val Pro Tyr Ser Asn Lys Phe Val Phe Asp Glu Asp Pro Leu	
1395 1400 1405	
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Ser Glu Tyr Glu Thr Ala Lys Leu Glu Ser Leu Ser Phe Gln Ala Gln	
1410 1415 1420	
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Leu Gly Asn Ile Asp Ala Val Asp Met Thr Gly Lys Leu Thr Leu Leu	
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Ser Gln Phe Thr Ala Arg Gln Ile Ile Asn Ala Ile Thr Gly Leu Asp	
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gag tct gtt tct ctt act aat gat gcc att gtt gca tca gac tat gtc	4416
Glu Ser Val Ser Leu Thr Asn Asp Ala Ile Val Ala Ser Asp Tyr Val	
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Ser Asn Trp Ile Ser Glu Cys Met Tyr Thr Lys Leu Asp Glu Leu Phe	
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Met Tyr Cys Gly Trp Glu Leu Leu Leu Glu Leu Ser Tyr Gln Met Tyr	
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Tyr Leu Arg Val Val Gly Trp Ser Asn Ile Val Asp Tyr Ser Tyr Met	
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Ile Leu Arg Arg Ile Pro Gly Ala Ala Leu Asn Asn Leu Ala Ser Thr	
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tta agt cat cca aaa ctt ttc cga cga gct atc aac cta gat ata gtt	4656
Leu Ser His Pro Lys Leu Phe Arg Arg Ala Ile Asn Leu Asp Ile Val	
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gcc ccc tta aat gct cct cat ttt gca tct ctg gac tac atc aag atg	4704
Ala Pro Leu Asn Ala Pro His Phe Ala Ser Leu Asp Tyr Ile Lys Met	
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Leu Ser Leu Ile His His Asn Gly Leu Glu Leu Pro Lys Ile Lys Gly	
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Phe Ser Pro Asp Glu Lys Cys Phe Ala Leu Thr Glu Phe Leu Arg Lys	
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Val Val Asn Ser Gly Leu Ser Ser Ile Glu Asn Leu Ser Asn Phe Met	
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Leu Lys Leu Thr Pro His Val Pro Gly Thr Ser Cys Ile Glu Asp Asp	
1715	1720 1725

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 Ser Leu Cys Thr Asn Asp Tyr Ile Ile Trp Ile Ile Glu Ser Asn Ala
 1730 1735 1740

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 Asn Leu Glu Lys Tyr Pro Ile Pro Asn Ser Pro Glu Asp Asp Ser Asn
 1745 1750 1755 1760

ttc cat aac ttt aag ttg aat gct cca tcg cac cat acc tta cgc cca 5328
 Phe His Asn Phe Lys Leu Asn Ala Pro Ser His His Thr Leu Arg Pro
 1765 1770 1775

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 Leu Gly Leu Ser Ser Thr Ala Trp Tyr Lys Gly Ile Ser Cys Cys Arg
 1780 1785 1790

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 Tyr Leu Glu Arg Leu Lys Leu Pro Gln Gly Asp His Leu Tyr Ile Ala
 1795 1800 1805

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 Glu Gly Ser Gly Ala Ser Met Thr Ile Ile Glu Tyr Leu Phe Pro Gly
 1810 1815 1820

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 Arg Lys Ile Tyr Tyr Asn Ser Leu Phe Ser Ser Gly Asp Asn Pro Pro
 1825 1830 1835 1840

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 1845 1850 1855

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 1905 1910 1915 1920

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 1925 1930 1935

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Pro	Asn	Glu	Ile	Asp	Ile	Asp	Asp	Leu	Gly	Pro	Leu	His	Asn	Gln	Asn
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Trp	Asn	Gln	Ile	Ala	His	Glu	Glu	Ser	Asn	Leu	Ala	Gln	Arg	Leu	Val
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Leu	Phe	Thr	Arg	Ser	Arg	Glu	Leu	Ser	Gly	Asp	Arg	Arg	Asp	Ile	Asp
145					150					155				160	
Leu	Lys	Thr	Val	Val	Ala	Ala	Trp	His	Asp	Ser	Asp	Trp	Lys	Arg	Ile
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Ser	Asp	Phe	Trp	Ile	Met	Ile	Lys	Phe	Gln	Met	Arg	Gln	Leu	Ile	Val
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Val Ser Asp Met Tyr Glu Gly Arg His Asn Ile Leu Ser Leu Cys Thr
    245                250                255
Val Ser Thr Tyr Leu Asn Pro Leu Lys Lys Arg Ile Thr Tyr Leu Leu
    260                265                270
Ser Leu Val Asp Asn Leu Ala Phe Gln Ile Gly Asp Ala Val Tyr Asn
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Ile Ile Ala Leu Leu Glu Ser Phe Val Tyr Ala Gln Leu Gln Met Ser
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Asp Pro Ile Pro Glu Leu Arg Gly Gln Phe His Ala Phe Val Cys Ser
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Pro Asp Leu Thr Ala Glu Leu Leu Cys Ile Met Arg Leu Trp Gly His
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Pro Met Leu Thr Ala Ser Gln Ala Ala Gly Lys Val Arg Glu Ser Met
    370                375                380
Cys Ala Gly Lys Val Leu Asp Phe Pro Thr Ile Met Lys Thr Leu Ala
    385                390                395                400
Phe Phe His Thr Ile Leu Ile Asn Gly Tyr Arg Arg Lys His His Gly
    405                410                415
Val Trp Pro Pro Leu Asn Leu Pro Gly Asn Ala Ser Lys Gly Leu Thr
    420                425                430
Glu Leu Met Asn Asp Asn Thr Glu Ile Ser Tyr Glu Phe Thr Leu Lys
    435                440                445
His Trp Lys Glu Ile Ser Leu Ile Lys Phe Lys Lys Cys Phe Asp Ala
    450                455                460
Asp Ala Gly Glu Glu Leu Ser Ile Phe Met Lys Asp Lys Ala Ile Ser
    465                470                475                480
Ala Pro Lys Gln Asp Trp Met Ser Val Phe Arg Arg Ser Leu Ile Lys
    485                490                495
Gln Arg His Gln His His Gln Val Pro Leu Pro Asn Pro Phe Asn Arg
    500                505                510
Arg Leu Leu Leu Asn Phe Leu Gly Asp Asp Lys Phe Asp Pro Asn Val
    515                520                525
Glu Leu Gln Tyr Val Thr Ser Gly Glu Tyr Leu His Asp Asp Thr Phe
    530                535                540
Cys Ala Ser Tyr Ser Leu Lys Glu Lys Glu Ile Lys Pro Asp Gly Arg
    545                550                555                560
Ile Phe Ala Lys Leu Thr Lys Arg Met Arg Ser Cys Gln Val Ile Ala
    565                570                575
Glu Ser Leu Leu Ala Asn His Ala Gly Lys Leu Met Lys Glu Asn Gly
    580                585                590
Val Val Met Asn Gln Leu Ser Leu Thr Lys Ser Leu Leu Thr Met Ser
    595                600                605
Gln Ile Gly Ile Ile Ser Glu Arg Ala Arg Lys Ser Thr Arg Asp Asn
    610                615                620
Ile Asn Arg Pro Gly Phe Gln Asn Ile Gln Arg Asn Lys Ser His His

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625					630					635				640	
Ser	Lys	Gln	Val	Asn	Gln	Arg	Asp	Pro	Ser	Asp	Asp	Phe	Glu	Leu	Ala
				645						650				655	
Ala	Ser	Phe	Leu	Thr	Thr	Asp	Leu	Lys	Lys	Tyr	Cys	Leu	Gln	Trp	Arg
			660					665					670		
Tyr	Gln	Thr	Ile	Ile	Pro	Phe	Ala	Gln	Ser	Leu	Asn	Arg	Met	Tyr	Gly
		675					680					685			
Tyr	Pro	His	Leu	Phe	Glu	Trp	Ile	His	Leu	Arg	Leu	Met	Arg	Ser	Thr
	690					695					700				
Leu	Tyr	Val	Gly	Asp	Pro	Phe	Asn	Pro	Pro	Ala	Asp	Thr	Ser	Gln	Phe
705					710					715				720	
Asp	Leu	Asp	Lys	Val	Ile	Asn	Gly	Asp	Ile	Phe	Ile	Val	Ser	Pro	Arg
			725					730					735		
Gly	Gly	Ile	Glu	Gly	Leu	Cys	Gln	Lys	Ala	Trp	Thr	Met	Ile	Ser	Ile
		740					745					750			
Ser	Val	Ile	Ile	Leu	Ser	Ala	Thr	Glu	Ser	Gly	Thr	Arg	Val	Met	Ser
	755						760					765			
Met	Val	Gln	Gly	Asp	Asn	Gln	Ala	Ile	Ala	Val	Thr	Thr	Arg	Val	Pro
	770					775					780				
Arg	Ser	Leu	Pro	Thr	Leu	Glu	Lys	Lys	Thr	Ile	Ala	Phe	Arg	Ser	Cys
785					790					795				800	
Asn	Leu	Phe	Phe	Glu	Arg	Leu	Lys	Cys	Asn	Asn	Phe	Gly	Leu	Gly	His
			805					810					815		
His	Leu	Lys	Glu	Gln	Glu	Thr	Ile	Ile	Ser	Ser	His	Phe	Phe	Val	Tyr
		820					825						830		
Ser	Lys	Arg	Ile	Phe	Tyr	Gln	Gly	Arg	Ile	Leu	Thr	Gln	Ala	Leu	Lys
		835					840					845			
Asn	Ala	Ser	Lys	Leu	Cys	Leu	Thr	Ala	Asp	Val	Leu	Gly	Glu	Cys	Thr
	850					855				860					
Gln	Ser	Ser	Cys	Ser	Asn	Leu	Ala	Thr	Thr	Val	Met	Arg	Leu	Thr	Glu
865					870					875				880	
Asn	Gly	Val	Glu	Lys	Asp	Ile	Cys	Phe	Tyr	Leu	Asn	Ile	Tyr	Met	Thr
			885					890					895		
Ile	Lys	Gln	Leu	Ser	Tyr	Asp	Ile	Ile	Phe	Pro	Gln	Val	Ser	Ile	Pro
		900					905						910		
Gly	Asp	Gln	Ile	Thr	Leu	Glu	Tyr	Ile	Asn	Asn	Pro	His	Leu	Val	Ser
		915					920					925			
Arg	Leu	Ala	Leu	Leu	Pro	Ser	Gln	Leu	Gly	Gly	Leu	Asn	Tyr	Leu	Ser
	930					935					940				
Cys	Ser	Arg	Leu	Phe	Asn	Arg	Asn	Ile	Gly	Asp	Pro	Val	Val	Ser	Ala
945					950					955				960	
Val	Ala	Asp	Leu	Lys	Arg	Leu	Ile	Lys	Ser	Gly	Cys	Met	Asp	Tyr	Trp
			965					970					975		
Ile	Leu	Tyr	Asn	Leu	Leu	Gly	Arg	Lys	Pro	Gly	Asn	Gly	Ser	Trp	Ala
		980					985						990		
Thr	Leu	Ala	Ala	Asp	Pro	Tyr	Ser	Ile	Asn	Ile	Glu	Tyr	Gln	Tyr	Pro
		995					1000					1005			
Pro	Thr	Thr	Ala	Leu	Lys	Arg	His	Thr	Gln	Gln	Val	Leu	Met	Glu	Leu
	1010					1015					1020				
Ser	Thr	Asn	Pro	Met	Leu	Arg	Gly	Ile	Phe	Ser	Asp	Asn	Ala	Gln	Ala
1025					1030					1035				1040	
Glu	Glu	Asn	Asn	Leu	Ala	Arg	Phe	Leu	Leu	Asp	Arg	Glu	Val	Ile	Phe
			1045					1050					1055		
Pro	Arg	Val	Ala	His	Ile	Ile	Ile	Glu	Gln	Thr	Ser	Val	Gly	Arg	Arg
			1060					1065					1070		

Lys Gln Ile Gln Gly Tyr Leu Asp Ser Thr Arg Ser Ile Met Arg Lys
 1075 1080 1085
 Ser Leu Glu Ile Lys Pro Leu Ser Asn Arg Lys Leu Asn Glu Ile Leu
 1090 1095 1100
 Asp Tyr Asn Ile Asn Tyr Leu Ala Tyr Asn Leu Ala Leu Leu Lys Asn
 1105 1110 1115 1120
 Ala Ile Glu Pro Pro Thr Tyr Leu Lys Ala Met Thr Leu Glu Thr Cys
 1125 1130 1135
 Ser Ile Asp Ile Ala Arg Ser Leu Arg Lys Leu Ser Trp Ala Pro Leu
 1140 1145 1150
 Leu Gly Gly Arg Asn Leu Glu Gly Leu Glu Thr Pro Asp Pro Ile Glu
 1155 1160 1165
 Ile Thr Ala Gly Ala Leu Ile Val Gly Ser Gly Tyr Cys Glu Gln Cys
 1170 1175 1180
 Ala Ala Gly Asp Asn Arg Phe Thr Trp Phe Phe Leu Pro Ser Gly Ile
 1185 1190 1195 1200
 Glu Ile Gly Gly Asp Pro Arg Asp Asn Pro Pro Ile Arg Val Pro Tyr
 1205 1210 1215
 Ile Gly Ser Arg Thr Asp Glu Arg Arg Val Ala Ser Met Ala Tyr Ile
 1220 1225 1230
 Arg Gly Ala Ser Ser Ser Leu Lys Ala Val Leu Arg Leu Ala Gly Val
 1235 1240 1245
 Tyr Ile Trp Ala Phe Gly Asp Thr Leu Glu Asn Trp Ile Asp Ala Leu
 1250 1255 1260
 Asp Leu Ser His Thr Arg Val Asn Ile Thr Leu Glu Gln Leu Gln Ser
 1265 1270 1275 1280
 Leu Thr Pro Leu Pro Thr Ser Ala Asn Leu Thr His Arg Leu Asp Asp
 1285 1290 1295
 Gly Thr Thr Thr Leu Lys Phe Thr Pro Ala Ser Ser Tyr Thr Phe Ser
 1300 1305 1310
 Ser Phe Thr His Ile Ser Asn Asp Glu Gln Tyr Leu Thr Ile Asn Asp
 1315 1320 1325
 Lys Thr Ala Asp Ser Asn Ile Ile Tyr Gln Gln Leu Met Ile Thr Gly
 1330 1335 1340
 Leu Gly Ile Leu Glu Thr Trp Asn Asn Pro Pro Ile Asn Arg Thr Phe
 1345 1350 1355 1360
 Glu Glu Ser Thr Leu His Leu His Thr Gly Ala Ser Cys Cys Val Arg
 1365 1370 1375
 Pro Val Asp Ser Cys Ile Ile Ser Glu Ala Leu Thr Val Lys Pro His
 1380 1385 1390
 Ile Thr Val Pro Tyr Ser Asn Lys Phe Val Phe Asp Glu Asp Pro Leu
 1395 1400 1405
 Ser Glu Tyr Glu Thr Ala Lys Leu Glu Ser Leu Ser Phe Gln Ala Gln
 1410 1415 1420
 Leu Gly Asn Ile Asp Ala Val Asp Met Thr Gly Lys Leu Thr Leu Leu
 1425 1430 1435 1440
 Ser Gln Phe Thr Ala Arg Gln Ile Ile Asn Ala Ile Thr Gly Leu Asp
 1445 1450 1455
 Glu Ser Val Ser Leu Thr Asn Asp Ala Ile Val Ala Ser Asp Tyr Val
 1460 1465 1470
 Ser Asn Trp Ile Ser Glu Cys Met Tyr Thr Lys Leu Asp Glu Leu Phe
 1475 1480 1485
 Met Tyr Cys Gly Trp Glu Leu Leu Leu Glu Leu Ser Tyr Gln Met Tyr
 1490 1495 1500
 Tyr Leu Arg Val Val Gly Trp Ser Asn Ile Val Asp Tyr Ser Tyr Met

1505		1510		1515		1520
Ile Leu Arg Arg	Ile Pro Gly Ala	Leu Asn Asn Leu	Ala Ser Thr			
	1525		1530		1535	
Leu Ser His Pro	Lys Leu Phe Arg	Arg Ala Ile Asn Leu	Asp Ile Val			
	1540		1545		1550	
Ala Pro Leu Asn	Ala Pro His Phe	Ala Ser Leu Asp	Tyr Ile Lys Met			
	1555		1560		1565	
Ser Met Asp Ala	Ile Leu Trp Gly	Cys Lys Arg Val	Ile Asn Val Leu			
	1570		1575		1580	
Ser Asn Gly Gly	Asp Leu Glu Leu	Val Val Thr Ser	Glu Asp Ser Leu			
1585		1590		1595		1600
Ile Leu Ser Asp	Arg Ser Met Asn	Leu Ile Ala Arg	Lys Leu Thr Leu			
	1605		1610		1615	
Leu Ser Leu Ile	His His Asn Gly	Leu Glu Leu Pro	Lys Ile Lys Gly			
	1620		1625		1630	
Phe Ser Pro Asp	Glu Lys Cys Phe	Ala Leu Thr Glu	Phe Leu Arg Lys			
	1635		1640		1645	
Val Val Asn Ser	Gly Leu Ser Ser	Ile Glu Asn Leu	Ser Asn Phe Met			
	1650		1655		1660	
Tyr Asn Val Glu	Asn Pro Arg Leu	Ala Ala Phe Ala	Ser Asn Asn Tyr			
1665		1670		1675		1680
Tyr Leu Thr Arg	Lys Leu Leu Asn	Ser Ile Arg Asp	Thr Glu Ser Gly			
	1685		1690		1695	
Gln Val Ala Val	Thr Ser Tyr Tyr	Glu Ser Leu Glu	Tyr Ile Asp Ser			
	1700		1705		1710	
Leu Lys Leu Thr	Pro His Val Pro	Gly Thr Ser Cys	Ile Glu Asp Asp			
	1715		1720		1725	
Ser Leu Cys Thr	Asn Asp Tyr Ile	Ile Ile Trp Ile	Ile Glu Ser Asn	Ala		
	1730		1735		1740	
Asn Leu Glu Lys	Tyr Pro Ile Pro	Asn Ser Pro Glu	Asp Asp Ser Asn			
1745		1750		1755		1760
Phe His Asn Phe	Lys Leu Asn Ala	Pro Ser His His	Thr Leu Arg Pro			
	1765		1770		1775	
Leu Gly Leu Ser	Ser Thr Ala Trp	Tyr Lys Gly Ile	Ser Cys Cys Arg			
	1780		1785		1790	
Tyr Leu Glu Arg	Leu Lys Leu Pro	Gln Gly Asp His	Leu Tyr Ile Ala			
	1795		1800		1805	
Glu Gly Ser Gly	Ala Ser Met Thr	Ile Ile Glu Tyr	Leu Phe Pro Gly			
	1810		1815		1820	
Arg Lys Ile Tyr	Tyr Asn Ser Leu	Phe Ser Ser Gly	Asp Asn Pro Pro			
1825		1830		1835		1840
Gln Arg Asn Tyr	Ala Pro Met Pro	Thr Gln Phe Ile	Glu Ser Val Pro			
	1845		1850		1855	
Tyr Lys Leu Trp	Gln Ala His Thr	Asp Gln Tyr Pro	Glu Ile Phe Glu			
	1860		1865		1870	
Asp Phe Ile Pro	Leu Trp Asn Gly	Asn Ala Ala Met	Thr Asp Ile Gly			
	1875		1880		1885	
Met Thr Ala Cys	Val Glu Phe Ile	Ile Asn Arg Val	Gly Pro Arg Thr			
	1890		1895		1900	
Cys Ser Leu Val	His Val Asp Leu	Glu Ser Ser Ala	Ser Leu Asn Gln			
1905		1910		1915		1920
Gln Cys Leu Ser	Lys Pro Ile Ile	Asn Ala Ile Ile	Thr Ala Thr Thr			
	1925		1930		1935	
Val Leu Cys Pro	His Gly Val Leu	Ile Leu Lys Tyr	Ser Trp Leu Pro			
	1940		1945		1950	

Phe Thr Arg Phe Ser Thr Leu Ile Thr Phe Leu Trp Cys Tyr Phe Glu
 1955 1960 1965
 Arg Ile Thr Val Leu Arg Ser Thr Tyr Ser Asp Pro Ala Asn His Glu
 1970 1975 1980
 Val Tyr Leu Ile Cys Ile Leu Ala Asn Asn Phe Ala Phe Gln Thr Val
 1985 1990 1995 2000
 Ser Gln Ala Thr Gly Met Ala Met Thr Leu Thr Asp Gln Gly Phe Thr
 2005 2010 2015
 Leu Ile Ser Pro Glu Arg Ile Asn Gln Tyr Trp Asp Gly His Leu Lys
 2020 2025 2030
 Gln Glu Arg Ile Val Ala Glu Ala Ile Asp Lys Val Val Leu Gly Glu
 2035 2040 2045
 Asn Ala Leu Phe Asn Ser Ser Asp Asn Glu Leu Ile Leu Lys Cys Gly
 2050 2055 2060
 Gly Thr Pro Asn Ala Arg Asn Leu Ile Asp Ile Glu Pro Val Ala Thr
 2065 2070 2075 2080
 Phe Ile Glu Phe Glu Gln Leu Ile Cys Thr Met Leu Thr Thr His Leu
 2085 2090 2095
 Lys Glu Ile Ile Asp Ile Thr Arg Ser Gly Thr Gln Asp Tyr Glu Ser
 2100 2105 2110
 Leu Leu Leu Thr Pro Tyr Asn Leu Gly Leu Leu Gly Lys Ile Ser Thr
 2115 2120 2125
 Ile Val Arg Leu Leu Thr Glu Arg Ile Leu Asn His Thr Ile Arg Asn
 2130 2135 2140
 Trp Leu Ile Leu Pro Pro Ser Leu Gln Met Ile Val Lys Gln Asp Leu
 2145 2150 2155 2160
 Glu Phe Gly Ile Phe Arg Ile Thr Ser Ile Leu Asn Ser Asp Arg Phe
 2165 2170 2175
 Leu Lys Leu Ser Pro Asn Arg Lys Tyr Leu Ile Thr Gln Leu Thr Ala
 2180 2185 2190
 Gly Tyr Ile Arg Lys Leu Ile Glu Gly Asp Cys Asn Ile Asp Leu Thr
 2195 2200 2205
 Arg Pro Ile Gln Lys Gln Ile Trp Lys Ala Leu Gly Cys Val Val Tyr
 2210 2215 2220
 Cys His Asp Pro Met Asp Gln Arg Glu Ser Thr Glu Phe Ile Asp Ile
 2225 2230 2235 2240
 Asn Ile Asn Glu Glu Ile Asp Arg Gly Ile Asp Gly Glu Glu Ile
 2245 2250 2255

<210> 25

<211> 1656

<212> DNA

<213> Canine parainfluenza virus

<220>

<221> CDS

<222> (1)...(1656)

<223> Canine parainfluenza virus F protein encoding
 sequence with "TAA" termination codon; cDNA in
 mRNA sense

<400> 25

atg ggt act aga att caa ttt ctg gtg gtc tcc tgt cta ttg gca gga 48
 Met Gly Thr Arg Ile Gln Phe Leu Val Val Ser Cys Leu Leu Ala Gly

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aca ggc agc ctt gat cca gca gcc ctc atg caa atc ggt gtc att cca	96			
Thr Gly Ser Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro				
20 25 30				
aca aat gtc cgg caa ctt atg tat tat act gag gct tca tca gca ttc	144			
Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe				
35 40 45				
att gtt gtg aag tta atg cct aca att gac tcg ccg att agt ggg tgt	192			
Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys				
50 55 60				
aat ata aca tcc att tca agc tat aat gca aca atg aca aaa ctt cta	240			
Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Met Thr Lys Leu Leu				
65 70 75 80				
cag ccg atc ggt gag aat tta gag acg att agg tac cag ttg att cca	288			
Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Tyr Gln Leu Ile Pro				
85 90 95				
act cgg agg aga cgc cgg ttt gca ggg gtg gtg att gga tta gcc gca	336			
Thr Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala				
100 105 110				
tta gga gta gct act gca gca cag gtc act gcc gca gta gca cta gta	384			
Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val				
115 120 125				
aag gcg aat aaa aat gct gtg gct ata ctc aat ctc aaa aac gca atc	432			
Lys Ala Asn Lys Asn Ala Val Ala Ile Leu Asn Leu Lys Asn Ala Ile				
130 135 140				
caa aaa aca aat gca gca gtt gca gac gtg gtt cag gcc aca caa tca	480			
Gln Lys Thr Asn Ala Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser				
145 150 155 160				
cta gga acg gca gtt caa gca gtt caa gat cac ata aat agt gtg gta	528			
Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Val				
165 170 175				
agt cca gca att aca gca gcc aat tgc aaa gcc caa gat gct atc att	576			
Ser Pro Ala Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile				
180 185 190				
ggc tca att ctc aat ctc tat ttg acc gag ttg aca act atc ttc cac	624			
Gly Ser Ile Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His				
195 200 205				
aat caa att aca aac ccc gca ttg agt cct att aca att caa gct ttg	672			
Asn Gln Ile Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu				
210 215 220				
agg atc cta cta ggg agt acc ttg ccg acc gtg gtc gaa aaa tct ttc	720			

Arg	Ile	Leu	Leu	Gly	Ser	Thr	Leu	Pro	Thr	Val	Val	Glu	Lys	Ser	Phe	
225					230					235					240	
aat	acc	cag	ata	agt	gca	gct	gag	ctt	ctc	tca	tca	ggg	tta	ttg	aca	768
Asn	Thr	Gln	Ile	Ser	Ala	Ala	Glu	Leu	Leu	Ser	Ser	Gly	Leu	Leu	Thr	
				245				250						255		
ggc	cag	att	gtg	gga	tta	gat	ttg	acc	tac	atg	cag	atg	gtc	ata	aaa	816
Gly	Gln	Ile	Val	Gly	Leu	Asp	Leu	Thr	Tyr	Met	Gln	Met	Val	Ile	Lys	
			260					265					270			
att	gag	ctg	cca	act	tta	act	gta	caa	cct	gca	acc	cag	atc	ata	gat	864
Ile	Glu	Leu	Pro	Thr	Leu	Thr	Val	Gln	Pro	Ala	Thr	Gln	Ile	Ile	Asp	
		275					280					285				
ctg	gtc	acc	att	tct	gca	ttc	att	aac	aat	caa	gaa	gtt	atg	gcc	caa	912
Leu	Val	Thr	Ile	Ser	Ala	Phe	Ile	Asn	Asn	Gln	Glu	Val	Met	Ala	Gln	
	290					295				300						
tta	cca	aca	cgt	gtt	att	gtg	act	ggc	agc	ttg	atc	caa	gcc	tat	ccc	960
Leu	Pro	Thr	Arg	Val	Ile	Val	Thr	Gly	Ser	Leu	Ile	Gln	Ala	Tyr	Pro	
305					310					315					320	
gca	tgc	caa	tgc	act	atc	acc	ccc	aac	act	gtg	tac	tgt	agg	tat	aat	1008
Ala	Ser	Gln	Cys	Thr	Ile	Thr	Pro	Asn	Thr	Val	Tyr	Cys	Arg	Tyr	Asn	
				325					330					335		
gat	gcc	caa	gta	ctc	tca	gat	gat	acg	atg	gct	tgc	ctc	caa	ggg	aac	1056
Asp	Ala	Gln	Val	Leu	Ser	Asp	Asp	Thr	Met	Ala	Cys	Leu	Gln	Gly	Asn	
			340					345					350			
ttg	aca	aga	tgc	acc	ttc	tct	cca	gtg	gtt	ggg	agc	ttt	ctc	act	cga	1104
Leu	Thr	Arg	Cys	Thr	Phe	Ser	Pro	Val	Val	Gly	Ser	Phe	Leu	Thr	Arg	
		355					360					365				
ttc	gtg	ctg	ttt	gat	gga	ata	gtt	tat	gca	aat	tgc	agg	tgc	atg	ttg	1152
Phe	Val	Leu	Phe	Asp	Gly	Ile	Val	Tyr	Ala	Asn	Cys	Arg	Ser	Met	Leu	
	370					375					380					
tgc	aag	tgc	atg	cag	cct	gct	gct	gtt	atc	cta	cag	ccg	agc	tca	tcc	1200
Cys	Lys	Cys	Met	Gln	Pro	Ala	Ala	Val	Ile	Leu	Gln	Pro	Ser	Ser	Ser	
385					390					395					400	
cct	gta	act	gtc	att	gac	atg	tac	aaa	tgt	gtg	agt	ctg	cag	ctt	gac	1248
Pro	Val	Thr	Val	Ile	Asp	Met	Tyr	Lys	Cys	Val	Ser	Leu	Gln	Leu	Asp	
				405					410					415		
aat	ctc	aga	ttc	acc	atc	act	caa	ttg	gcc	aat	ata	acc	tac	aat	agc	1296
Asn	Leu	Arg	Phe	Thr	Ile	Thr	Gln	Leu	Ala	Asn	Ile	Thr	Tyr	Asn	Ser	
			420					425					430			
acc	atc	aag	ctt	gaa	aca	tcc	cag	atc	ttg	cct	atc	gat	ccg	ttg	gat	1344
Thr	Ile	Lys	Leu	Glu	Thr	Ser	Gln	Ile	Leu	Pro	Ile	Asp	Pro	Leu	Asp	
		435					440					445				

ata tcc cag aat cta gct gcg gtg aat aag agt cta agt gat gca cta 1392
 Ile Ser Gln Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu
 450 455 460

caa cac tta gca caa agt gac aca tac ctt tct gca atc aca tca gct 1440
 Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala
 465 470 475 480

acg act aca agt gta tta tcc ata ata gca atc tgt ctt gga tgg tta 1488
 Thr Thr Thr Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu
 485 490 495

ggt tta ata tta ata atc ttg ctc agt gta gtt gtg tgg aag tta ttg 1536
 Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu
 500 505 510

acc att gtt gct gct aat cga aat aga atg gag aat ttt gtt tat cat 1584
 Thr Ile Val Ala Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His
 515 520 525

aat tca gca ttc cac cac tca cgg tct gat ctc agt gag aaa aat caa 1632
 Asn Ser Ala Phe His His Ser Arg Ser Asp Leu Ser Glu Lys Asn Gln
 530 535 540

cct gca act ctt gga aca aga taa 1656
 Pro Ala Thr Leu Gly Thr Arg *
 545 550

<210> 26

<211> 551

<212> PRT

<213> Canine parainfluenza virus

<400> 26

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 Thr Gly Ser Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro
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 Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe
 35 40 45
 Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys
 50 55 60
 Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Met Thr Lys Leu Leu
 65 70 75 80
 Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Tyr Gln Leu Ile Pro
 85 90 95
 Thr Arg Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala
 100 105 110
 Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val
 115 120 125
 Lys Ala Asn Lys Asn Ala Val Ala Ile Leu Asn Leu Lys Asn Ala Ile
 130 135 140
 Gln Lys Thr Asn Ala Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser
 145 150 155 160


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Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Val
                               165                               170                               175
Ser Pro Ala Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile
                               180                               185                               190
Gly Ser Ile Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His
                               195                               200                               205
Asn Gln Ile Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu
                               210                               215                               220
Arg Ile Leu Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe
225                               230                               235                               240
Asn Thr Gln Ile Ser Ala Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr
                               245                               250                               255
Gly Gln Ile Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys
                               260                               265                               270
Ile Glu Leu Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp
                               275                               280                               285
Leu Val Thr Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln
                               290                               295                               300
Leu Pro Thr Arg Val Ile Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro
305                               310                               315                               320
Ala Ser Gln Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn
                               325                               330                               335
Asp Ala Gln Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn
                               340                               345                               350
Leu Thr Arg Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg
                               355                               360                               365
Phe Val Leu Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu
                               370                               375                               380
Cys Lys Cys Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser
385                               390                               395                               400
Pro Val Thr Val Ile Asp Met Tyr Lys Cys Val Ser Leu Gln Leu Asp
                               405                               410                               415
Asn Leu Arg Phe Thr Ile Thr Gln Leu Ala Asn Ile Thr Tyr Asn Ser
                               420                               425                               430
Thr Ile Lys Leu Glu Thr Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp
                               435                               440                               445
Ile Ser Gln Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu
                               450                               455                               460
Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala
465                               470                               475                               480
Thr Thr Thr Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu
                               485                               490                               495
Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu
                               500                               505                               510
Thr Ile Val Ala Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His
                               515                               520                               525
Asn Ser Ala Phe His His Ser Arg Ser Asp Leu Ser Glu Lys Asn Gln
                               530                               535                               540
Pro Ala Thr Leu Gly Thr Arg
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<210> 27

<211> 1656

<212> DNA

<213> Porcine Rubulavirus

<220>

<221> CDS

<222> (1)...(1656)

<223> Porcine Rubulavirus F protein encoding sequence
with "TAA" termination codon; cDNA in mRNA sense

<400> 27

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Met Gly Thr Ile Ile Gln Phe Leu Val Val Ser Cys Leu Leu Ala Gly	
1 5 10 15	
gca ggc agc ctt gat cca gca gcc ctc atg caa atc ggt gtc att cca	96
Ala Gly Ser Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro	
20 25 30	
aca aat gtc cgg caa ctt atg tat tat act gag gcc tca tca gca ttc	144
Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe	
35 40 45	
att gtt gtg aag tta atg cct aca att gac tcg ccg att agt gga tgt	192
Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys	
50 55 60	
aat ata aca tca att tca agc tat aat gca aca gtg aca aaa ctc cta	240
Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu	
65 70 75 80	
cag ccg atc ggt gag aat ttg gaa acg att agg aac cag ttg att cca	288
Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro	
85 90 95	
act cgg agg aga cgc cgg ttt gca ggg gtg gtg att gga tta gct gca	336
Thr Arg Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala	
100 105 110	
tta gga gta gct act gcc gca cag gtc act gcc gca gta gca cta gta	384
Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val	
115 120 125	
aag gca aat aaa aat gct gcg gct ata ctc aat ctc aaa aat gca atc	432
Lys Ala Asn Lys Asn Ala Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile	
130 135 140	
caa aaa aca aat aca gca gtt gca gat gtg gtc cag gcc aca caa tca	480
Gln Lys Thr Asn Thr Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser	
145 150 155 160	
cta gga acg gca gtt caa gca gtt caa gat cac ata aac agt gtg gta	528
Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Val	
165 170 175	
agt cca gca att aca gca gcc aat tgt aag gcc caa gat gct atc att	576
Ser Pro Ala Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile	

180	185	190	
ggc tca atc ctc aat ctc tat ttg acc gag ttg aca act atc ttc cac			624
Gly Ser Ile Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His			
195	200	205	
aat caa att aca aac cct gca ttg agt cct att aca att caa gct tta			672
Asn Gln Ile Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu			
210	215	220	
agg atc cta ctg ggg agt acc ttg ccg act gtg gtc gaa aaa tct ttc			720
Arg Ile Leu Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe			
225	230	235	240
aat acc cag ata agt gca gct gag ctt ctc tca tca ggg tta ttg aca			768
Asn Thr Gln Ile Ser Ala Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr			
245	250	255	
ggc cag att gtg gga tta gat ttg acc tat atg cag atg gtc ata aaa			816
Gly Gln Ile Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys			
260	265	270	
att gag ctg cca act tta act gta caa cct gca acc cag atc ata gat			864
Ile Glu Leu Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp			
275	280	285	
ctg gcc acc att tct gca ttc att aac aat caa gaa gtc atg gcc caa			912
Leu Ala Thr Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln			
290	295	300	
tta cca aca cgt gtt att gtg act ggc agc ttg atc caa gcc tat ccc			960
Leu Pro Thr Arg Val Ile Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro			
305	310	315	320
gca tcg caa tgc act att aca ccc aac act gtg tac tgt agg tat aat			1008
Ala Ser Gln Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn			
325	330	335	
gat gcc caa gta ctc tca gat gat acg atg gct tgc ctc caa ggt aac			1056
Asp Ala Gln Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn			
340	345	350	
ttg aca aga tgc acc ttc tct ccg gtg gtt ggg agc ttt ctc act cga			1104
Leu Thr Arg Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg			
355	360	365	
ttc atg ctg ttc gat gga ata gtt tat gca aat tgc agg tcg atg tta			1152
Phe Met Leu Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu			
370	375	380	
tgc aag tgc atg cag cct gct gct gtg atc cta cag ccg agt tca tcc			1200
Cys Lys Cys Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser			
385	390	395	400
cct gta act gtc att gac atg tac aaa tgt gtg agt ctg cag ctt gac			1248

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Pro Val Thr Val Ile Asp Met Tyr Lys Cys Val Ser Leu Gln Leu Asp
      405                      410                      415

aat ctc aga ttc acc atc act caa ttg gcc aat gta acc tac aat agc 1296
Asn Leu Arg Phe Thr Ile Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser
      420                      425                      430

acc atc aag ctt gaa aca tcc cag atc ttg cct att gat ccg ttg gat 1344
Thr Ile Lys Leu Glu Thr Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp
      435                      440                      445

ata tcc cag aat cta gct gcg gtg aat aag agt cta agt gat gca cta 1392
Ile Ser Gln Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu
      450                      455                      460

caa cac tta gca caa agt gac aca tac ctt tct gca atc aca tca gct 1440
Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala
465                      470                      475                      480

acg act aca agt gta tta tcc ata atg gca atc tgt ctt gga tcg tta 1488
Thr Thr Thr Ser Val Leu Ser Ile Met Ala Ile Cys Leu Gly Ser Leu
      485                      490                      495

ggg tta ata tta ata atc ttg ctc agt gta gtt gtg tgg aag tta ttg 1536
Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu
      500                      505                      510

acc att gtc act gct aat cga aat aga atg gag aat ttt gtt tat cat 1584
Thr Ile Val Thr Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His
      515                      520                      525

aat tca gca ttc cac cac tca cga tct gat ctc agt gag aaa aat caa 1632
Asn Ser Ala Phe His His Ser Arg Ser Asp Leu Ser Glu Lys Asn Gln
      530                      535                      540

cct gca act ctt gga aca aga taa
Pro Ala Thr Leu Gly Thr Arg *
545                      550

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<210> 28

<211> 551

<212> PRT

<213> Porcine Rubulavirus

<400> 28

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      20                      25                      30
Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe
      35                      40                      45
Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys
      50                      55                      60
Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu

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65					70					75				80	
Gln	Pro	Ile	Gly	Glu	Asn	Leu	Glu	Thr	Ile	Arg	Asn	Gln	Leu	Ile	Pro
				85					90					95	
Thr	Arg	Arg	Arg	Arg	Arg	Phe	Ala	Gly	Val	Val	Ile	Gly	Leu	Ala	Ala
			100					105					110		
Leu	Gly	Val	Ala	Thr	Ala	Ala	Gln	Val	Thr	Ala	Ala	Val	Ala	Leu	Val
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Lys	Ala	Asn	Lys	Asn	Ala	Ala	Ala	Ile	Leu	Asn	Leu	Lys	Asn	Ala	Ile
	130					135					140				
Gln	Lys	Thr	Asn	Thr	Ala	Val	Ala	Asp	Val	Val	Gln	Ala	Thr	Gln	Ser
145					150					155					160
Leu	Gly	Thr	Ala	Val	Gln	Ala	Val	Gln	Asp	His	Ile	Asn	Ser	Val	Val
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Ser	Pro	Ala	Ile	Thr	Ala	Ala	Asn	Cys	Lys	Ala	Gln	Asp	Ala	Ile	Ile
		180						185					190		
Gly	Ser	Ile	Leu	Asn	Leu	Tyr	Leu	Thr	Glu	Leu	Thr	Thr	Ile	Phe	His
	195					200						205			
Asn	Gln	Ile	Thr	Asn	Pro	Ala	Leu	Ser	Pro	Ile	Thr	Ile	Gln	Ala	Leu
	210					215					220				
Arg	Ile	Leu	Leu	Gly	Ser	Thr	Leu	Pro	Thr	Val	Val	Glu	Lys	Ser	Phe
225					230					235					240
Asn	Thr	Gln	Ile	Ser	Ala	Ala	Glu	Leu	Leu	Ser	Ser	Gly	Leu	Leu	Thr
			245					250						255	
Gly	Gln	Ile	Val	Gly	Leu	Asp	Leu	Thr	Tyr	Met	Gln	Met	Val	Ile	Lys
		260					265						270		
Ile	Glu	Leu	Pro	Thr	Leu	Thr	Val	Gln	Pro	Ala	Thr	Gln	Ile	Ile	Asp
	275					280						285			
Leu	Ala	Thr	Ile	Ser	Ala	Phe	Ile	Asn	Asn	Gln	Glu	Val	Met	Ala	Gln
	290					295					300				
Leu	Pro	Thr	Arg	Val	Ile	Val	Thr	Gly	Ser	Leu	Ile	Gln	Ala	Tyr	Pro
305					310					315					320
Ala	Ser	Gln	Cys	Thr	Ile	Thr	Pro	Asn	Thr	Val	Tyr	Cys	Arg	Tyr	Asn
			325					330						335	
Asp	Ala	Gln	Val	Leu	Ser	Asp	Asp	Thr	Met	Ala	Cys	Leu	Gln	Gly	Asn
		340						345					350		
Leu	Thr	Arg	Cys	Thr	Phe	Ser	Pro	Val	Val	Gly	Ser	Phe	Leu	Thr	Arg
	355						360					365			
Phe	Met	Leu	Phe	Asp	Gly	Ile	Val	Tyr	Ala	Asn	Cys	Arg	Ser	Met	Leu
	370					375					380				
Cys	Lys	Cys	Met	Gln	Pro	Ala	Ala	Val	Ile	Leu	Gln	Pro	Ser	Ser	Ser
385					390					395					400
Pro	Val	Thr	Val	Ile	Asp	Met	Tyr	Lys	Cys	Val	Ser	Leu	Gln	Leu	Asp
			405					410						415	
Asn	Leu	Arg	Phe	Thr	Ile	Thr	Gln	Leu	Ala	Asn	Val	Thr	Tyr	Asn	Ser
		420						425					430		
Thr	Ile	Lys	Leu	Glu	Thr	Ser	Gln	Ile	Leu	Pro	Ile	Asp	Pro	Leu	Asp
	435						440					445			
Ile	Ser	Gln	Asn	Leu	Ala	Ala	Val	Asn	Lys	Ser	Leu	Ser	Asp	Ala	Leu
	450					455					460				
Gln	His	Leu	Ala	Gln	Ser	Asp	Thr	Tyr	Leu	Ser	Ala	Ile	Thr	Ser	Ala
465					470					475					480
Thr	Thr	Thr	Ser	Val	Leu	Ser	Ile	Met	Ala	Ile	Cys	Leu	Gly	Ser	Leu
			485					490					495		
Gly	Leu	Ile	Leu	Ile	Ile	Leu	Leu	Ser	Val	Val	Val	Trp	Lys	Leu	Leu
			500					505					510		

Thr Ile Val Thr Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His
 515 520 525
 Asn Ser Ala Phe His His Ser Arg Ser Asp Leu Ser Glu Lys Asn Gln
 530 535 540
 Pro Ala Thr Leu Gly Thr Arg
 545 550

<210> 29

<211> 1656

<212> DNA

<213> Simian virus 5

<220>

<221> CDS

<222> (1)...(1590)

<223> Simian virus 5 W3A strain F protein encoding
 sequence; cDNA in mRNA sense

<400> 29

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Met Gly Thr Ile Ile Gln Phe Leu Val Val Ser Cys Leu Leu Ala Gly	
1 5 10 15	
gca ggc agc ctt gat cca gca gcc ctc atg caa atc ggt gtc att cca	96
Ala Gly Ser Leu Asp Pro Ala Ala Leu Met Gln Ile Gly Val Ile Pro	
20 25 30	
aca aat gtc cgg caa ctt atg tat tat act gag gcc tca tca gca ttc	144
Thr Asn Val Arg Gln Leu Met Tyr Thr Thr Glu Ala Ser Ser Ala Phe	
35 40 45	
att gtt gtg aag tta atg cct aca att gac tgg ccg att agt gga tgt	192
Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys	
50 55 60	
aat ata aca tca att tca agc tat aat gca aca gtg aca aaa ctc cta	240
Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu	
65 70 75 80	
cag ccg atc ggt gag aat ttg gag acg att agg aac cag ttg att cca	288
Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro	
85 90 95	
act cgg agg aga cgc cgg ttt gca ggg gtg gtg att gga tta gct gca	336
Thr Arg Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala	
100 105 110	
tta gga gta gct act gcc gca cag gtc act gcc gca gtg gca cta gta	384
Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val	
115 120 125	
aag gca aat gaa aat gct gcg gct ata ctc aat ctc aaa aat gca atc	432
Lys Ala Asn Glu Asn Ala Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile	
130 135 140	

caa aaa aca aat gca gca gtt gca gat gtg gtc cag gcc aca caa tca	480
Gln Lys Thr Asn Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser	
145 150 155 160	
cta gga acg gca gtt caa gca gtt caa gat cac ata aac agt gtg gta	528
Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Val	
165 170 175	
agt cca gca att aca gca gcc aat tgt aag gcc caa gat gct atc att	576
Ser Pro Ala Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile	
180 185 190	
ggc tca atc ctc aat ctc tat ttg acc gag ttg aca acc atc ttc cac	624
Gly Ser Ile Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His	
195 200 205	
aat caa att aca aac cct gca ttg agt ccc att aca att caa gct tta	672
Asn Gln Ile Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu	
210 215 220	
agg atc cta ctg ggg agt acc ttg ccg act gtg gtc gaa aaa tct ttc	720
Arg Ile Leu Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe	
225 230 235 240	
aat acc cag ata agt gca gct gag ctt ctc tca tca ggg tta ttg aca	768
Asn Thr Gln Ile Ser Ala Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr	
245 250 255	
ggc cag att gtg gga tta gat ttg acc tat atg cag atg gtc ata aaa	816
Gly Gln Ile Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys	
260 265 270	
att gag ctg cca act tta act gta caa cct gca acc cag atc ata gat	864
Ile Glu Leu Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp	
275 280 285	
ctg gcc acc att tct gca ttc att aac aat caa gaa gtc atg gcc caa	912
Leu Ala Thr Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln	
290 295 300	
tta cca aca cgt gtt atg gtg act ggc agc ttg atc caa gcc tat ccc	960
Leu Pro Thr Arg Val Met Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro	
305 310 315 320	
gca tcg caa tgc acc att aca ccc aac act gtg tac tgt agg tat aat	1008
Ala Ser Gln Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn	
325 330 335	
gat gcc caa gta ctc tca gat gat act atg gct tgc ctc caa ggt aac	1056
Asp Ala Gln Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn	
340 345 350	
ttg aca aga tgc acc ttc tct cca gtg gtt ggg agc ttt ctc act cga	1104
Leu Thr Arg Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg	

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Phe Val Leu Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu			
370	375	380	
tgc aag tgc atg caa cct gct gct gtg atc cta cag ccg agt tca tcc			1200
Cys Lys Cys Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser			
385	390	395	400
cct gta act gtc att gac atg tac aaa tgt gtg agt ctg cag ctt gac			1248
Pro Val Thr Val Ile Asp Met Tyr Lys Cys Val Ser Leu Gln Leu Asp			
	405	410	415
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Asn Leu Arg Phe Thr Ile Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser			
	420	425	430
acc atc aag ctt gaa tca tcc cag atc ttg tct att gat ccg ttg gat			1344
Thr Ile Lys Leu Glu Ser Ser Gln Ile Leu Ser Ile Asp Pro Leu Asp			
	435	440	445
ata tcc caa aat cta gct gcg gtg aat aag agt cta agt gat gca cta			1392
Ile Ser Gln Asn Leu Ala Val Ala Val Asn Lys Ser Leu Ser Asp Ala Leu			
	450	455	460
caa cac tta gca caa agt gac aca tat ctt tct gca atc aca tca gct			1440
Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala			
465	470	475	480
acg act aca agt gta tta tcc ata ata gca atc tgt ctt gga tcg tta			1488
Thr Thr Thr Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu			
	485	490	495
ggc tta ata tta ata atc ttg ctc agt gta gtt gtg tgg aag tta ttg			1536
Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu			
	500	505	510
acc att gtc gtt gct aat cga aat aga atg gag aat ttt gtt tat cat			1584
Thr Ile Val Val Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His			
	515	520	525
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Lys *			
tcttggaaca agataa			1656
<210> 30			
<211> 529			
<212> PRT			
<213> Simian virus 5			
<400> 30			
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	20	25	30
Thr Asn Val	Arg Gln Leu Met Tyr	Thr Glu Ala Ser	Ser Ala Phe
	35	40	45
Ile Val Val	Lys Leu Met Pro Thr	Ile Asp Ser Pro	Ile Ser Gly Cys
	50	55	60
Asn Ile Thr	Ser Ile Ser Ser Tyr	Asn Ala Thr Val	Thr Lys Leu Leu
	65	70	75
Gln Pro Ile	Gly Glu Asn Leu Glu Thr	Ile Arg Asn Gln	Leu Ile Pro
	85	90	95
Thr Arg Arg	Arg Arg Arg Phe Ala	Gly Val Val Ile	Gly Leu Ala Ala
	100	105	110
Leu Gly Val	Ala Thr Ala Ala Gln	Val Thr Ala Ala	Val Ala Leu Val
	115	120	125
Lys Ala Asn	Glu Asn Ala Ala Ala	Ile Leu Asn Leu	Lys Asn Ala Ile
	130	135	140
Gln Lys Thr	Asn Ala Ala Val Ala	Asp Val Val Gln	Ala Thr Gln Ser
	145	150	155
Leu Gly Thr	Ala Val Gln Ala Val	Gln Asp His Ile	Asn Ser Val Val
	165	170	175
Ser Pro Ala	Ile Thr Ala Ala Asn	Cys Lys Ala Gln	Asp Ala Ile Ile
	180	185	190
Gly Ser Ile	Leu Asn Leu Tyr Leu	Thr Glu Leu Thr	Thr Ile Phe His
	195	200	205
Asn Gln Ile	Thr Asn Pro Ala Leu	Ser Pro Ile Thr	Ile Gln Ala Leu
	210	215	220
Arg Ile Leu	Leu Gly Ser Thr Leu	Pro Thr Val Val	Glu Lys Ser Phe
	225	230	235
Asn Thr Gln	Ile Ser Ala Ala Glu	Leu Leu Ser Ser	Gly Leu Leu Thr
	245	250	255
Gly Gln Ile	Val Gly Leu Asp Leu	Thr Tyr Met Gln	Met Val Ile Lys
	260	265	270
Ile Glu Leu	Pro Thr Leu Thr Val	Gln Pro Ala Thr	Gln Ile Ile Asp
	275	280	285
Leu Ala Thr	Ile Ser Ala Phe Ile	Asn Asn Gln Glu	Val Met Ala Gln
	290	295	300
Leu Pro Thr	Arg Val Met Val Thr	Gly Ser Leu Ile	Gln Ala Tyr Pro
	305	310	315
Ala Ser Gln	Cys Thr Ile Thr Pro	Asn Thr Val Tyr	Cys Arg Tyr Asn
	325	330	335
Asp Ala Gln	Val Leu Ser Asp Asp	Thr Met Ala Cys	Leu Gln Gly Asn
	340	345	350
Leu Thr Arg	Cys Thr Phe Ser Pro	Val Val Gly Ser	Phe Leu Thr Arg
	355	360	365
Phe Val Leu	Phe Asp Gly Ile Val	Tyr Ala Asn Cys	Arg Ser Met Leu
	370	375	380
Cys Lys Cys	Met Gln Pro Ala Ala	Val Ile Leu Gln	Pro Ser Ser Ser
	385	390	395
Pro Val Thr	Val Ile Asp Met Tyr	Lys Cys Val Ser	Leu Gln Leu Asp
	405	410	415
Asn Leu Arg	Phe Thr Ile Thr Gln	Leu Ala Asn Val	Thr Tyr Asn Ser
	420	425	430
Thr Ile Lys	Leu Glu Ser Ser Gln	Ile Leu Ser Ile	Asp Pro Leu Asp
	435	440	445

Ile Ser Gln Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu
 450 455 460
 Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala
 465 470 475 480
 Thr Thr Thr Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu
 485 490 495
 Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu
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 Lys

<210> 31

<211> 1656

<212> DNA

<213> Simian virus 5

<220>

<221> CDS

<222> (1)...(1590)

<223> Simian virus 5 WR strain F protein encoding
 sequence; cDNA in mRNA sense

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gca ggc agc ctt gat cta gca gcc ctc atg caa atc ggt gtc att cca	96
Ala Gly Ser Leu Asp Leu Ala Ala Leu Met Gln Ile Gly Val Ile Pro	
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aca aat gtc cgg caa ctt atg tat tat act gag gcc tca tcg gca ttc	144
Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe	
35 40 45	
att gtt gtg aag tta atg cct aca att gac tcg ccg att agt gga tgt	192
Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys	
50 55 60	
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Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu	
65 70 75 80	
cag ccg atc ggt gag aat ttg gag acg att agg aac cag ttg att cca	288
Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro	
85 90 95	
act cgg aga aga cgc cgg ttt gca ggg gtg gtg att gga tta gct gca	336
Thr Arg Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala	
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tta gga gta gct act gcc gca cag gtc act gcc gca gta gca cta gta	384

Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val	
115 120 125	
aag gca aat gaa aat gct gcg gct ata ctc aat ctc aaa aat gca atc	432
Lys Ala Asn Glu Asn Ala Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile	
130 135 140	
caa aaa aca aat gca gca gtt gca gat gtg gtc cag gcc aca caa tca	480
Gln Lys Thr Asn Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser	
145 150 155 160	
cta gga acg gca gtt caa gca gtt caa gat cac ata aac agt gtg gta	528
Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Val	
165 170 175	
agt cca gca att aca gca gcc aat tgt aag gcc caa gat gct atc att	576
Ser Pro Ala Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile	
180 185 190	
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Gly Ser Ile Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His	
195 200 205	
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210 215 220	
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Arg Ile Leu Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe	
225 230 235 240	
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Asn Thr Gln Ile Ser Ala Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr	
245 250 255	
ggc cag att gtg gga tta gat ttg acc tat atg cag atg gtc ata aaa	816
Gly Gln Ile Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys	
260 265 270	
att gag ctg cca act tta act gta caa cct gca acc cag atc ata gat	864
Ile Glu Leu Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp	
275 280 285	
ctg gcc acc att tct gca ttc att aac aat caa gaa gtc atg gcc caa	912
Leu Ala Thr Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln	
290 295 300	
tta cca aca cgt gtt atg gtg act ggc agc ttg atc caa gcc tat ccc	960
Leu Pro Thr Arg Val Met Val Thr Gly Ser Leu Ile Gln Ala Tyr Pro	
305 310 315 320	
gca tcg caa tgc acc att aca ccc aac act gtg tac tgt agg tat aat	1008
Ala Ser Gln Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn	
325 330 335	

gat gcc caa gta ctc tca gat gat act atg gct tgc ctc caa ggt aac 1056
Asp Ala Gln Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn
340 345 350

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Leu Thr Arg Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg
355 360 365

ttc gtg ctg ttc gat gga ata gtt tat gca aat tgc agg tcg atg ttg 1152
Phe Val Leu Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu
370 375 380

tgc aag tgc atg caa cct gct gct gtg atc cta cag ccg agt tca tcc 1200
Cys Lys Cys Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser
385 390 395 400

cct gta act gtc att gac atg tac aaa tgt gtg agt ctg cag ctt gac 1248
Pro Val Thr Val Ile Asp Met Tyr Lys Cys Val Ser Leu Gln Leu Asp
405 410 415

aat ctc aga ttc acc atc act caa ttg gcc aat gta acc tac aat agc 1296
Asn Leu Arg Phe Thr Ile Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser
420 425 430

acc atc aag ctt gaa tca tcc cag atc ttg cct att gat ccg ttg gat 1344
Thr Ile Lys Leu Glu Ser Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp
435 440 445

ata tcc cag aat cta gct gcg gtg aat aag agt cta agt gat gca cta 1392
Ile Ser Gln Asn Leu Ala Val Asn Lys Ser Leu Ser Asp Ala Leu
450 455 460

caa cac tta gca caa agt gac aca tat ctt tct gca atc aca tca gct 1440
Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala
465 470 475 480

acg act aca agt gta tta tcc ata ata gca atc tgt ctt gga tcg tta 1488
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485 490 495

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Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu
500 505 510

acc att gtc gct gct aat cga aat aga atg gag aat ttt gtt tat cat 1584
Thr Ile Val Ala Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His
515 520 525

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Lys *

tcttggaaca agataa 1656

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<211> 529

<212> PRT

<213> Simian virus 5

<400> 32

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Thr Asn Val Arg Gln Leu Met Tyr Tyr Thr Glu Ala Ser Ser Ala Phe
      35           40           45
Ile Val Val Lys Leu Met Pro Thr Ile Asp Ser Pro Ile Ser Gly Cys
 50           55           60
Asn Ile Thr Ser Ile Ser Ser Tyr Asn Ala Thr Val Thr Lys Leu Leu
65           70           75           80
Gln Pro Ile Gly Glu Asn Leu Glu Thr Ile Arg Asn Gln Leu Ile Pro
      85           90           95
Thr Arg Arg Arg Arg Arg Phe Ala Gly Val Val Ile Gly Leu Ala Ala
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Leu Gly Val Ala Thr Ala Ala Gln Val Thr Ala Ala Val Ala Leu Val
      115          120          125
Lys Ala Asn Glu Asn Ala Ala Ala Ile Leu Asn Leu Lys Asn Ala Ile
130          135          140
Gln Lys Thr Asn Ala Ala Val Ala Asp Val Val Gln Ala Thr Gln Ser
145          150          155          160
Leu Gly Thr Ala Val Gln Ala Val Gln Asp His Ile Asn Ser Val Val
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Ser Pro Ala Ile Thr Ala Ala Asn Cys Lys Ala Gln Asp Ala Ile Ile
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Gly Ser Ile Leu Asn Leu Tyr Leu Thr Glu Leu Thr Thr Ile Phe His
      195          200          205
Asn Gln Ile Thr Asn Pro Ala Leu Ser Pro Ile Thr Ile Gln Ala Leu
210          215          220
Arg Ile Leu Leu Gly Ser Thr Leu Pro Thr Val Val Glu Lys Ser Phe
225          230          235          240
Asn Thr Gln Ile Ser Ala Ala Glu Leu Leu Ser Ser Gly Leu Leu Thr
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Gly Gln Ile Val Gly Leu Asp Leu Thr Tyr Met Gln Met Val Ile Lys
260          265          270
Ile Glu Leu Pro Thr Leu Thr Val Gln Pro Ala Thr Gln Ile Ile Asp
275          280          285
Leu Ala Thr Ile Ser Ala Phe Ile Asn Asn Gln Glu Val Met Ala Gln
290          295          300
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305          310          315          320
Ala Ser Gln Cys Thr Ile Thr Pro Asn Thr Val Tyr Cys Arg Tyr Asn
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Asp Ala Gln Val Leu Ser Asp Asp Thr Met Ala Cys Leu Gln Gly Asn
340          345          350
Leu Thr Arg Cys Thr Phe Ser Pro Val Val Gly Ser Phe Leu Thr Arg
355          360          365
Phe Val Leu Phe Asp Gly Ile Val Tyr Ala Asn Cys Arg Ser Met Leu
370          375          380
Cys Lys Cys Met Gln Pro Ala Ala Val Ile Leu Gln Pro Ser Ser Ser
385          390          395          400

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Pro Val Thr Val Ile Asp Met Tyr Lys Cys Val Ser Leu Gln Leu Asp
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 Asn Leu Arg Phe Thr Ile Thr Gln Leu Ala Asn Val Thr Tyr Asn Ser
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 Thr Ile Lys Leu Glu Ser Ser Gln Ile Leu Pro Ile Asp Pro Leu Asp
 435 440 445
 Ile Ser Gln Asn Leu Ala Ala Val Asn Lys Ser Leu Ser Asp Ala Leu
 450 455 460
 Gln His Leu Ala Gln Ser Asp Thr Tyr Leu Ser Ala Ile Thr Ser Ala
 465 470 475 480
 Thr Thr Thr Ser Val Leu Ser Ile Ile Ala Ile Cys Leu Gly Ser Leu
 485 490 495
 Gly Leu Ile Leu Ile Ile Leu Leu Ser Val Val Val Trp Lys Leu Leu
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 Thr Ile Val Ala Ala Asn Arg Asn Arg Met Glu Asn Phe Val Tyr His
 515 520 525
 Lys

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 <213> Cryptovirus

<220>
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 aatacaatac caccaggggt cacaggacta ctaaccaatg ctgcagaggc aaagatccaa 180
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<210> 34
 <211> 2244
 <212> DNA
 <213> Cryptovirus

<220>

<223> cDNA in mRNA sense

<400> 34

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<210> 35

<211> 18

<212> DNA

<213> Cryptovirus

<220>

<223> cDNA in mRNA sense

<400> 35

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18

<210> 36

<211> 18
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<220>
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<210> 37
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<210> 38
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<210> 39
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<220>
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<210> 46

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<220>
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<220>
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 aataacacta ctattccaat aactggaatt accagcttga tttatctcca aaatgattca 180
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<210> 49
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 <212> DNA
 <213> Cryptovirus

<220>
 <223> cDNA in mRNA sense

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