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(54) Title: METHOD, DEVICE AND COMPUTER PROGRAM PRODUCT FOR DIAGNOSING OF AT LEAST ONE EX-HAUST EMISSION CONTROL UNIT

(57) Abstract: The present invention relates to a kit comprising at least two compositions, wherein each of said compositions comprises at least one strain chosen from Lactobacillus. The Lactobacillus strains are chosen from the group consisting of Lactobacillus plantarum, lactobacillus rhamnosus, lactobacillus fermentum, lactobacillus paracasei and Lactobacillus gasseri.

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### KIT COMPRISING AT LEAST TWO LACTOBACILLUS COMPOSITIONS

#### Field of the invention

The present invention relates to a kit comprising at least two compositions, wherein each of said compositions comprises at least one strain chosen from Lactobacillus. The invention further relates to a method for administration of said kit, wherein each of said compositions of the kit is administered in a sequence or in an alternating sequence or in a combination thereof.

#### Background art

Probiotoic bacteria are defined as live microorganisms which when administered in adequate amounts beneficially affect the host. Lactobacilli and bifidobacteria are the most frequently used bacteria in probiotic products. These bacteria are generally safe, as are probiotics based on these organisms. The lack of pathogenicity extends across all age groups and to immunocompromised individuals. Intake of different probiotic bacteria has been shown to have clinical benefits in various physiologic or pathologic situations. The most clear cut effects have been shown in diarrhoea caused by antibiotic therapy or rotavirus infection. There are also studies showing positive clinical effects in inflammatory bowel diseases, atopic dermatitis and hypercholesterolemia after intake of probiotic bacteria. 25 The mechanism, by which probiotic bacteria contribute to these clinical improvements are not clear. In vitro human, as well as both in vivo and in vitro animal studies, have shown that different species of lactobacilli affects the innate and acquired immune system in many different ways. Clinical studies have 30 mainly shown stimulation of the innate cellular immune system and enhancement of humoral immune responses to

natural infections and systemic or oral immunisation.

Regarding effects of the innate immune system, increased

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phagocytic activity of polymorphonuclear cells (PMN) and increased NK cell tumor killing activity have been reported. To our knowledge, there are no clinical studies showing effects on the specific cellular immune system after intake of probiotic bacteria.

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In the present application the effects on the innate and acquired immune system following daily intake of lactobacilli or the Gram-negative bacteria P. lundensis have been thoroughly investigated. Interestingly, it has been observed an activation of the specific cellular immune system in subjects receiving L. plantarum and indications of such in subjects receiving L. paracasei. Moreover, immunity-enhancing effects on the innate immune system, such as expansion of the NKT cell population and increased phagocytoic activity were observed in subjects receiving different lactobacilli species. Intake of the Gram-negative bacteria P. lundensis had no effects, whatsoever, on the different immune parameters measured in this study.

20 Increased standard of living and improved sanitary conditions in the industrialized nations have increased life expectancy, foremost by reducing the burden of infection. However, at the same time, a number of diseases have, instead, increased. These include the three types of diseases that results from exaggerated or 25 dysregulated immunity, namely allergies, autoimmunity and inflammatory bowel disease. All these three types of diseases are more common in developed than in developing countries, more common in small than large families and 30 more common in urban than rural populations. This has led to the formulation of "the hygiene hypothesis", which states that the infant's developing immune system needs to obtain a certain degree of stimulation by infections, or perhaps commensal microbes, in order to mobilize 35 regulatory T cells so that proper tolerance mechanisms can develop.

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A number of measures to reduce allergies have been instituted. Parents have been given advice to avoid allergens, breast-feed and avoid smoking. Depite all attempts, allergies and other immunoregulatory diseases continue to increase globally.

In the present application a method to expand and/or increase the functional activity of regulatory T cells has been studied, thereby reducing the risk of developing immunoregulatory diseases such as allergy, autoimmune diseases (type 1-diabetes and multiple sclerosis) and inflammatory bowel disease. By intake of a sequence of lactobacillus strains, the immune system is activated in a manner that leads to formation of functionally active regulatory T cells. These cells can then down-regulate potentially harmful immune mediated diseases, such as allergy.

#### Summary of the invention

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An object of the present invention is a kit comprising at least two compositions, wherein each of said compositions comprises at least one strain chosen from Lactobacillus.

Another object of the present invention is a method for administration of above mentioned kit, wherein each of said compositions of the kit is administered in a sequence or in an alternating sequence or in a combination thereof.

### Brief description of the drawings

Figure 1 shows the numbers of volunteers reporting any minor adverse gastrointestinal effects during the trial.

Figure 2 shows base line numbers (day 0) of different lymphocytes per ml blood (mean ± (SEM))

Figure 3 shows base line (day 0) percentages or GMFI (mean  $\pm$  (SEM)) of lymphocytes positive for different cell activation and memory markers.

Figure 4. Subjects were randomly assigned to nine different study groups. The trial started with a wash out

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period of two weeks. Thereafter, the active study period followed. During this period, the subjects consumed one dose of study product per day for 14 (*L. Heal 19, L. fermentum, L. paracasei, L. gasseri, L. rhamnosus, P. lundensis* groups) or 35 days (*L. plantarum* and placebo group). Each dose contained 10<sup>10</sup> coloni forming units (CFU) (*lactobacilli* groups) or 10<sup>9</sup> CFU bacteria (*P. lundensis* group). Subjects were supplied with a list of probiotic products, which should not be consumed during the trial. A diary, in which each subject stated adverse effects, health conditions and confirmed intake of study product, was kept during the trial.

Figure 5. Percentages of lymphocytes expressing the activation phenotypes CD3CD8CD25, CD3CD8HLA-DR, CD3CD4CD25 and CD3CD4HLA-DR was analysed by flowcytometry. Group means (±SEM) based on individual ratios, day 14/day 0 and day 35/day 0 (for *L. plantarum* and placebo group only) is shown.

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Figure 6. Percentages of lymphocytes expressing the memory phenotypes CD3CD8CD45RO and CD3CD4CD45RO was analysed by flowcytometry. Group means (±SEM) based on individual ratios, day 14/day 0 and day 35/day 0 (for L. plantarum and placebo group only) is shown.

Figure 7. Percentages of lymphocytes positive for the NKT cell markers (CD56CD16CD3) was analysed by flowcytometry. Group calculations are based on individual ratios (day 14/day 0).

Figure 8. The phagocytic activity of neutrophils was analysed by incubating whole blood cells with FITC-labelled E. coli or S. aureus. The ratio between mean fluorescence values obtained at day 14 and day 0 was determined individually and group calculations are shown in this figure.

Figure 9 shows the ratio of lymphocytes expressing 35 the activation phenotypes CD4CD25 from experiment 2.

Figure 10 shows the ratio of lymphocytes expressing the activation phenotypes  ${\rm CD4}^{+}{\rm CD25}^{++}$  from experiment 2.

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Figure 11 shows the ratio of lymphocytes expressing the activation phenotypes  ${\rm CD8}^{\rm +}{\rm HLA-DR}^{\rm +}$  from experiment 2.

Figure 12 shows the ratio of lymphocytes expressing the activation phenotypes CD8+CD25+ from experiment 2.

Figure 13 shows the ratio of lymphocytes expressing the activation phenotypes CD4CD45RO from experiment 2. Detailed description of the invention

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The compositions comprised in the kit according to the invention and/or the compositions according to the invention may be independently selected from food compositions, dietary supplements, functional foods, medical foods and nutritional products.

The above mentioned food compositions may e.g. be beverages, yoghurts, juices, ice creams, breads, biscuits, cereals, health bars, spreads, or nutritional products.

Preferably, the kit according to the invention comprises at least 10 of the above mentioned compositions. More preferably the kit according to the invention comprises at least 7 of the above mentioned compositions. Most preferably the kit according to the invention comprises at least 5 of the above mentioned compositions.

The Lactobacillus used according to the invention may be selected from, but not limited to, the group consisting of Lactobacillus plantarum, Lactobacillus rhamnsosus, Lactobacillus fermentum, Lactobacillus paracasei and Lactobacillus gasseri.

The Lactobacillus plantarum used according to the
invention may be selected from, but not limited to, the
group consisting of Lactobacillus plantarum 299, DSM
6595, Lactobacillus plantarum 299v, DSM 9843,
Lactobacillus plantarum HEAL 9, DSM 15312, Lactobacillus
plantarum HEAL 19, DSM 15313, and Lactobacillus plantarum
HEAL 99, DSM 15316.

The Lactobacillus paracasei used according to the invention may be selected from, but not limited to, the

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group consisting of Lactobacillus paracasei 8700:2, DSM 13434, and Lactobacillus paracasei 02A, DSM13432.

The Lactobacillus gasseri used according to the invention is selected from, but not limited to, Lactobacillus gasseri VPG44, DSM 16737.

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Other probiotic bacterial strains, than the ones explicitly disclosed herein, may naturally be used in the present invention and are comprised of the invention.

Preferably, the compositions of the kit according to the invention and/or the composition according to the invention further comprise a carrier material. Said carrier material may independently be selected from the group consisting of oat meal gruel, lactic acid fermented foods, resistant starch, dietary fibres, carbohydrates, proteins, and glycosylated proteins.

Preferably, said at least one strain in the compositions of the kit according to the invention is present in an amount from about  $1 \times 10^6$  to about  $1 \times 10^{14}$  CFU. More preferred is that said at least one strain in the compositions of the kit according to the invention is present in an amount from  $1 \times 10^8$  to  $1 \times 10^{12}$ . Even more preferred is that that said at least one strain in the compositions of the kit according to the invention is present in an amount from  $1 \times 10^9$  to  $1 \times 10^{11}$ .

Preferably, said at least two strains in the composition according to the invention are each present in an amount from about  $1 \times 10^6$  to about  $1 \times 10^{14}$  CFU. More preferred is that said at least two strains in the composition according to the invention are each present in an amount from  $1 \times 10^8$  to  $1 \times 10^{12}$ . Even more preferred is that that said at least two strains in the composition according to the invention are each present in an amount from  $1 \times 10^9$  to  $1 \times 10^{11}$ . It is also possible that the at least two strains are present in a total amount from about  $1 \times 10^6$  to about  $1 \times 10^{14}$  CFU, preferably  $1 \times 10^8$  to  $1 \times 10^{12}$ , and more preferably  $1 \times 10^9$  to  $1 \times 10^{11}$ .

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The kit according to the invention may be administered in a sequence or in an alternating sequence or in a combination thereof. This means that the compositions comprising the respective strain or strains 5 could be administered alternating, e.g. strain A is administered first, then strain B is administered and then strain C is administered. Thereafter the sequence is repeated. It is also possible that first an alternating strain is administered (i.e. first A and then B and C) and thereafter only one certain strain is administered, 10 e.g. strain D, and thereafter another or the same sequence of bacterial strains are administered. The compositions of the kit may also be administered simultaneously.

15 Preferably, the kit is administered in a way such that each composition of the kit is administered at time intervals of between approximately 12 h and approximately 14 days, preferably approximately 24 h and approximately 7 days, more preferably between 24 h and 48 h. In one embodiment of the invention one composition comprising at least one probiotic bacterial strain is administered per day.

According to the invention it is possible to prevent virus infections by giving a kit comprising the compositions according to the invention to an individual. Treatable virus infections are i.e. those caused by a virus selected from the group consisting of common cold virus, rhinovirus, adenovirus, parainfluenza virus, respiratory syncytial virus, enterovirus and corona virus.

Thus, the use of a kit according to the invention may be very beneficial in the sense of being usable prophylactically, i.e. before the virus infection has developed. It is for instance very convenient for a normal healthy individual to take to composition of the invention prophylactically to stay healthy.

Examples

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#### Example 1

## Subjects and trial criteria

Fifty-seven apparently healthy volunteers within the age range 18-55 years (median, 26 years) were selected for this blind placebo controlled study. Subjects were 5 randomly assigned to eight groups, receiving either one of the following Gram-positive bacteria, L. plantarum 299v (n=7), L. plantarum Heal 19 (n=7), L. fermentum 35D(n=7), L. paracasei 8700:2 (n=7), L. gasseri VPG44 (n=7), L. rhamnosus 271 (n=7), or the Gram-negative bacteria, P. 10 lundensis (n=7) or placebo (n=10). The dose of bacteria was 10<sup>10</sup> bacteria/day for lactobacilli and 10<sup>9</sup> bacteria/day for P. lundensis. The control group took skim milk powder (1 q). Depending on the group, the study had a duration period of 6 or 9 weeks consisting of two 15 weeks wash out period, 2 or 5 weeks active study period and 2 weeks follow up period (Fig. 4). Each subject was supplied with a list of products containing probiotic products, which should not be consumed during the whole study period. Peripheral blood samples were withdrawn 20 from subjects by venipuncture at two or three time points, day 0, day 14 and day 35. A diary, in which each subject stated adverse effects, health conditions and confirmed intake of study product, was kept during the 25 trial.

#### Flow cytometry

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Phenotypic analysis of lymphocytes in whole blood was performed by flow cytometry. The following anti-human monoclonal antibodies were used as surface markers for different cell populations: CD3 FITC (SK7), CD4 APC (SK3), CD8 PerCP (SK1), CD19 PerCP (SJ25C1), CD56 PE (MY31), CD16 PE (B73.1), and CD5 FITC (L17F12). Following anti-human monoclonal antibodies were used for detection of different activation and memory markers: CD25 FITC (2A3), HLA-DR PE (L243), CD45RO PE (UCHL-1), CD38 PE (HB7), CD27 PE (L128), and CD11b PE (D12). All antibodies were purchased from Becton-Dickinson (Erembodegum,

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Belgium). Whole blood (100  $\mu$ l) was incubated with antibodies (10  $\mu$ l/antibody) for 30 min at 4° C in the dark. Thereafter, 2 ml of FACS lysing solution (Becton-Dickinson) was added and incubated for 15 min at 20° C in the dark. Cells were washed by adding 3 ml FACSFlow and centrifuged at 300 x g for 5 min. Washed cells were resuspended in 200 µl FACSFlow and analysed on a FacsCalibur (Becton-Dickinson) with CellQuest software.

#### Phagocytosis assay

The phagocytic activity of granulocytes and 10 monocytes were quantified with PHAGOTEST® (Orpegen Pharma, Heidelberg, Germany) according to manufacturers instruction with some modifications. Briefly, 20 x106 FITC labelled E. coli or FITC labelled S. aureus was added to pre-cooled whole blood (100  $\mu$ l). Blood cells and 15 bacteria were incubated on 37° C for 10 FacsCalibur with CellQuest software.

#### Calculations

Individual changes regarding different immune parameters were determined by calculating the ratio 20 between the individual values obtained at day 14 and day 0, or the values at day 35 and day 0. These ratios were used for all group calculations and statistics.

### Statistics

All statistical analyses were performed using Statview. Mann-Whitney U test were used to compare different groups.

#### Results

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### Clinical observations

Fifty-four out of fifty-seven volunteers completed 30 the study. Two persons were excluded due to infection and antibiotic treatment (one in the placebo group and one in the group receiving P. lundensis). One person was excluded day 16 due to pregnancy (placebo group). Only mild adverse gastrointestinal side effects were reported 35 following intake of study products (Fig. 1).

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## Intake of lactobacilli activates <u>T</u> cells

There were great baseline (day 0) individual variations regarding activation markers on CD4<sup>+</sup> and CD8<sup>+</sup> T cells. The baseline percentages of cells expressing 5 different cell surface markers are shown in Fig. 2. No significant differences were observed between different groups at this time point. Since huge inter-individual variations were observed, it was chosen to compare ratio values at day 14 and day 35 compared to day 0 for each individual. All calculations and comparisons were done on 10 these ratio values (day 14/day 0 and day 35/day 0). After 14 days of intake of study product containing L. plantarum 299v there was an approximately twofold increase of the expression of the activation marker CD25 on  $CD8^+$  T cells (p=0.01) (fig. 5). There was also a 15 strong, although not significant (p=0.12), indication of upregulation of HLA-DR on CD8+ cells following L. plantarum 299v intake. In addition, it was also observed a tendency towards activation of CD4 T cells after L. plantarum 299v intake. Intake of the other lactobacilli 20 species included in this study, as well as the Gramnegative bacteria P. lundensis activated neither CD8+ nor CD4 T cells. However, there was a tendency that intake of L. paracasei did increase the expression of HLA-DR on  $CD4^+$  T cells (p=0.18). 25

Intake of lactobacilli induces a memory phenotype of CD4+
T cells

Geometric means of the fluorescence intensity (GMFI) of the expression of CD45RO on CD4+ and CD8+ T cells were compared between groups receiving different study products. As above, group calculations based on individual ratio values (day 14/day 0 and day 35/day 0) were used for comparisons. After 35 days of intake of study product containing *L. plantarum 299v* the CD45RO GMFI on CD4+ T cells increased significantly (p=0.03). There was also a tendency towards increased CD45RO expression on CD8+ T cells following intake of *L*.

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plantarum (Fig. 6). Moreover, intake of L. paracasei seems to have a positive effect on upregulation of CD45RO on CD8+ T cells (p=0.10) (Fig. 6).

Effect on different cell populations following intake of study product

Following intake of *L. pararcasei* there was an increase in the percentage of lymphocytes being identified as NKT cells (P=0.06) (Fig 7). Relative increase/decrease compared to day 0 could not be detected regarding other cell populations, such as CD4+ T cells, CD8+ T cells, B cells, B-1 cells (CD19+CD5+), NK cells, granulocytes and monocytes.

## Phagocytic activity

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Granulocytes and monocytes were identified in the 15 FSC-SSC diagram. The ability of these cells to phagocytose FITC-labelled Gram-positive or Gram-negative bacteria was tested. As shown in fig. 8, granulocytes from volunteers given L. plantarum 299v (p=0.064), L. plantarum Heal 19 (p=0.064), L. fermentum (p=0.064) or L. 20 paracasei (p=0.05) were more efficient then granulocytes from placebo treated volunteers in phagocytosis of the Gram-negative bacteria E. coli. However, there was no difference between the groups in phagocytosis of the Gram-positive bacteria S. aureus. No differences in the 25 phagocytic activity of monocytes could be detected (data not shown).

#### Discussion

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The primary task of the immune system is to react rapidly and violently to micro-organisms thereby preventing and curing infections. The killing of microorganisms employs powerful mechanisms that also cause harm to our own tissues. Therefore, it is necessary that it neither reacts to our own tissues, nor to innocuous substances present in the environment. Therefore, the immune system develops and maintains tolerance both to the components of our own body, and to food and inhaled proteins. If this fails, a number of

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diseases may arise. Means to develop specific immune tolerance are an essential task of the immune system.

A central role in all immune reactions is played by the T helper cell. When a T helper cell becomes activated by its specific antigen, it becomes activated, divides, matures and produces a range of cytokines which direct the action of other types of cells in the immune system, such as cytotoxic T cells and B cells. Activation of T helper cells is necessary in order to produce most types of immune reactions, including production of antibodies. Conversely, if activation of T helper cells is prevented, most types of immune reactions are paralysed.

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There are several mechanisms by which activation of T helper cells and maintenance of tolerance is ensured. One mechanism is elimination in the thymus of T cells with capacity to recognize and react to own tissue. However, this elimination is not complete and, furthermore, we also need to develop specific immune tolerance to exogenous antigens. Otherwise we would react violently to all types of inhaled and ingested substances, leading to massive inflammation and wasted immune resources.

A cell type that is central for maintenance of tolerance is the regulatory T cell. This cell type can be recognized by certain markers, such as surface expression of CD4 and CD25, possession of intracellular CTLA-4, and transcription of the nuclear protein Foxp3. The regulatory T cells are capable of preventing other T cells to become activated when encountering harmless substances and, hence, prevent all types of unwanted immune reactions.

In the present context the symbol "+" in connection with a certain marker such as CD4+ and CD25+ means that the marker is expressed on a T cell. For instance CD4+CD25+ T cells are T cells that expresss both the CD4 marker and CD25 marker on its surface. However, nothing is said about the amount of the marker that is expressed,

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only that it is present. In the present context the symbol "++" in connection with a marker such as CD4++ or CD25++ means that there is a lot of marker expressed. The regulatory T cells are those cells with a lot of CD25 on the surface, i.e. CD4+CD25++ cells. On the other hand, CD4+CD25+ T cells are only activated T cells. Sometimes the specific symbols "+" and "++" are not used, e.g. CD4CD25 only, and this means that the cells are activated such CD4+CD25+ cells. Thus, CD4CD25 is the same as CD4+CD25+. When discussing regulatory T cells, it is always written as CD4+CD25++ cells.

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This blind placebo-controlled study is unique in that it is the first study comparing the influence of several immune parameters following intake of different Gram-positive lactobacilli or the Gram-negative bacteria 15 P. lundensis. Interestingly, intake of P. lundensis did not influence any of the measured parameters. In contrast, intake of lactobacilli affected different components of both the specific and innate immune system. A novel finding in this study was that intake of L. 20 plantarum had a pronounced positive effect on activation and induction of memory cells in the T cell populations. There was a significant upregulation of the IL-2 receptor  $\alpha$  chain (CD25) and a strong tendency towards upregulation of HLA-DR on cytotoxic T cells. A tendency towards 25 upregulation of these activation markers was also observed on helper T cells after intake of L. plantarum. Expression of activation markers indicates that the T cells have started to proliferate in response to antigen-30 specific or non-specific stimuli and that these cells more readily exert their effector functions compared to resting T cells. The mechanisms behind L. plantarum induced activation of T cells could be via antigen presenting cells that are activated by toll-like 35 receptors binding to microbial compounds. Activation of antigen presenting cells makes them more efficient in

presenting antigen to T cells. In addition, both helper

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and cytotoxic T cells have shown to have various expressions of toll-like receptors, which probably make these cells sensible for non-specific activation by microbial components and products.

5 In analogy to the helper T cell compartment, expression of CD45RO seems to mark a memory population also among cytotoxic T cells. There was found a significant increase in the expression of this memory cell marker on helper T cells, and a tendency towards 10 upregulation on cytotoxic T cells following 35 days intake of L. plantarum. In addition, intake of L. paracasei also showed a tendency towards upregulation of CD45RO on cytotoxic T cells. Relative to naïve T cells, CD45RO+ T cells can secrete a broad spectrum of 15 cytokines. Moreover, CD45RO+ T cells can proliferate and produce IL-2 when the CD3-TCR complex is stimulated under suboptimal conditions, whereas naïve T cells require a strong CD3-TCR stimulus to carry out these functions. The formation of memory T cells is important for induction of 20 an efficient immune response after infection and vaccination.

The innate cellular immune system was also affected by intake of probiotic bacteria. It was demonstrated that the natural killer T (NKT) cell population was expanded following intake of L. paracasei. NKT cells constitute a lymphocyte subpopulation that coexpress the NK cell marker CD56 and the T cell marker CD3-T cell receptor complex. Studies in both humans and mice have demonstrated that NKT cells play a central role in the regulation of autoimmune diseases, such as multiple sclerosis, type I diabetes, and systemic lupus. NKT cells also exert effector functions against tumour and virus infected cells. Thus, NKT cells are pleotropic in their functions. Other clinical studies evaluating the immunological effects of probiotic bacteria have shown that intake of L. rhamnosus HN001 and Bifidobacterium lactis HN019 enhance NK (including NKT) cell tumour

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killing activity of K562 cells. In this study it was also confirmed the observation by others that phagocytic activity of polymorphonuclear cells is increased after intake of different lactobacilli. The consequence of the observed effects on the different immune parameters in the present study is that one could speculate that the coincident activation of cytotoxic T cells and NKT cell expansion points to a strengthened immune defense against viral infections and/or tumours. The *in vitro* finding that lactobacilli induce mononuclear cells to secrete IL-12 and IL-18, supports the theory that intake of these bacteria stimulates cell-mediated activity.

In accordance with the present invention it has been concluded that intake of *L. plantarum* and *L. paracasei*15 has a profound effect on the specific and innate cellular immune system. However, the increase in immune function demonstrated herein is for the time being difficult to correlate to a proven health benefit in humans. In order to address this specific issue, further clinical trials in individuals suffering from e.g. viral infections or tumours need to be accomplished. In such studies, it would be of special interest to compare the effect of administration of *L. plantarum* and *L. paracasei* separately or in combination.

## 25 Example 2

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The goal of this example was to investigate the effect on the immune system by giving the same species of lactobacilli for a longer period of time compared to several lactobacilli (different species) administered in a sequence one after the other.

The volunteers were given a powder with freeze-dried bacteria during 14 or 35 days. As gram-positive bacteria the probiotic bacteria Lactobacillus plantarum 299v is used alone or in combination with L. rhamnosus, L.

fermentum, L. paracasei, and L. gasseri. As gram-negative bacteria Psedomonas lundensis is given.

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The following groups are studied:

- 1) Lactobacillus plantarum 35 days
- 2) L. plantarum 7d, L. rhamnosus 7d, L. fermentum 7d, L paracasei 7d, L. gasseri 7d. Totally 35 days. (Sequence)
- 5 3) A mixture of L. plantarum, L. rhamnosus, L. fermentum, L. paracasei, L. gasseri. Totally 14 days
  - 4) L. rhamnosus 14 days
  - 5) L. fermentum 14 days
  - 6) L. paracasei 14 days
- 10 7) *L. gasseri* 14 days
  - 8) Pseudomonas lundensis 14 days Control group 1) Placebo 35 days

Control group 2) Placebo 14 days

- Blood samples are taken at day 0, 14 and 35. The amount of helper T cells (CD4+) expressing high amounts of CD25 was defined in each group by flow cytometry as have been explained above in experiment 1.

  Results
- On day 14, there was a borderline significance of CD4+CD25++ T cells being expanded in individuals consuming the sequence of five different lactobacilli strains.

#### Discussion

Thelper cells (CD4+) expressing high density of the CD25 molecule (CD4+CD25++) have been shown to be important in order to protect against autoimmune diseases, allergies and inflammatory bowel diseases. The finding that these cells are expanded after intake of a sequence of different lactobacilli indicate that intake of these bacteria might be beneficial for the individual concerning the risk of developing the above mentioned diseases.

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#### CLAIMS

- 1. A kit comprising at least two compositions, wherein each of said compositions comprises at least one strain chosen from Lactobacillus.
- 2. A kit according to claim 1, wherein said composition is chosen from the group consisting of a food composition, a dietary supplement, a functional food, a medical food and a nutritional product.
- 3. A kit according to claim 2, wherein said food composition is chosen from the group consisting of beverages, yoghurts, juices, ice creams, breads, biscuits, cereals, health bars, spreads, and nutritional products.
- 4. A kit according to any one of claims 1-3, wherein said kit comprises at least 10 compositions, preferably at least 7 compositions, and most preferably at least 5 compositions.
- 5. A kit according to any one of claims 1-4, wherein said Lactobacillus strain is chosen from the group consisting of Lactobacillus plantarum, Lactobacillus rhamnsosus, Lactobacillus fermentum, Lactobacillus paracasei and Lactobacillus gasseri.
- 6. A kit according to claim 5, wherein said
  Lactobacillus plantarum is chosen from the group
  consisting of Lactobacillus plantarum 299, DSM 6595,
  Lactobacillus plantarum 299v, DSM 9843, Lactobacillus
  plantarum HEAL 9, DSM 15312, Lactobacillus plantarum HEAL
  19, DSM 15313, and Lactobacillus plantarum HEAL 99, DSM
  30 15316.
  - 7. A kit according to claim 5, wherein said Lactobacillus paracasei is chosen from the group consisting of Lactobacillus paracasei 8700:2, DSM 13434, and Lactobacillus paracasei 02A, DSM13432.
- 8. A kit according to claim 5, wherein said Lactobacillus gasseri is selected from Lactobacillus gasseri VPG44, DSM 16737.

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9. A kit according to any one of claims 1-8, wherein said at least two compositions comprise a carrier material.

- 10. A kit according to claim 9, wherein said carrier 5 material is chosen from the group consisting of oat meal gruel, lactic acid fermented foods, resistant starch, dietary fibres, carbohydrates, proteins, and glycosylated proteins.
- 11. A kit according to any one of claims 1-10, wherein each of said at least two strains in the composition is present in an amount from about  $1 \times 10^6$  to about  $1 \times 10^{14}$  CFU, preferably from  $1 \times 10^8$  to  $1 \times 10^{12}$ , and more preferably from  $1 \times 10^9$  to  $1 \times 10^{11}$ .
- 12. A method for administration of a kit according to any one of claims 1-11, wherein each of said compositions of the kit is administered in a sequence or in an alternating sequence or in a combination thereof.
- 13. A method according to claim 12, wherein each composition is administered at time intervals of between about 12 h and about 14 days, preferably about 24 h and about 7 days, more preferably between about 24 h and about 48 h.
- 14. A method for administration of a kit according to any one of claims 1-11, wherein said compositions are administered simultaneously.

Fig. 1

	Wash out period		Study period			Post study			
	(Week)		(Week)			period (Week)			
	-2	-1	1	2	3	4	5	+1	+2
L. plantarum	0/7	1/7	3/7	2/7	3/7	2/7	1/7	1/7	0/7
L. Heal 19	0/7	1/7	1/7	2/7				2/7	1/7
L. fermentum	0/7	0/7	0/7	0/7				1/7	0/7
L. paracasei	0/7	0/7	1/7	0/7				0/7	0/7
L. gasseri	0/7	0/7	3/7	1/7				4/7	0/7
L. rhamnosus	1/7	1/7	0/7	0/7				1/7	0/7
P. lundensis	1/6	1/6	1/6	1/6				0/6	0/6
Placebo	0/9	0/9	2/9	3/9	1/8	1/8	0/8	0/8	0/8

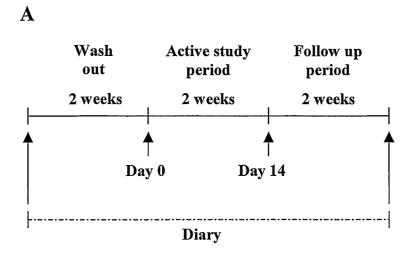
Fig. 2.

	CD4+ T cells	CD8+ T cells	NKT cells
	$(x 10^3)$	(x 10 <sup>3</sup> )	$(x 10^3)$
L. plantarum	647 (92)	318 (37)	64 (17)
L. Heal 19	817 (105)	328 (43)	56 (19)
L. fermentum	907 (82)	479 (51)	87 (21)
L. paracasei	794 (87)	321 (64)	98 (21)
L. gasseri	767 (54)	497 (110)	111 (39)
L. rhamnosus	775 (109)	387 (50)	109 (22)
P. lundensis	731 (65)	468 (84)	87 (29)
Placebo	650 (43)	300 (34)	107 (30)

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on CD8+ T cells GMFI CD45RO 27 (5,6) 38 (3,5) 23 (5,1) 50 (17) 45 (14) 40 (11) 61 (15) 36 (5,6) GMFI CD45RO on CD4+ T cells 110 (96) 126 (39) 53 (10) 65 (9,4) 39 (8,2) 71 (13) 83 (13) 80 (24) % CD8+HLA-DR+ of lymphocytes 5,0 (1,8) 6,1(1,6)6,8 (3,5) 6,6 (3,3) 7,0 (1,1) 9,4 (2,4) 5,2 (1,3) 5,2 (1,3) % CD4+HLA-DR+ of lymphocytes 4,2 (0,59) 3,0 (0,34) 4,3 (0,69) 4,4 (0,51) 4,4 (1,2) 8,5 (4,7) 3,3 (0,60) 10 (6,7) % CD8+CD25+ of lymphocytes 0,83 (0,19) 1,7 (0,73) 1,3 (0,13) 1,5 (0,40) 1,5 (0,31) 1,3 (0,26) 2,6 (1,4) 4,0 (2,3) % CD4+CD25+ of lymphocytes 14 (0,60) 10 (0,90) 15 (0,98) 17 (2,6) 15 (2,0) 13 (1,0) 17 (1,1) 18 (4,1) L. rhamnosus L. fermentum L. plantarum L. paracasei P. lundensis L. Heal 19 L. gasseri Placebo

Figure 3.



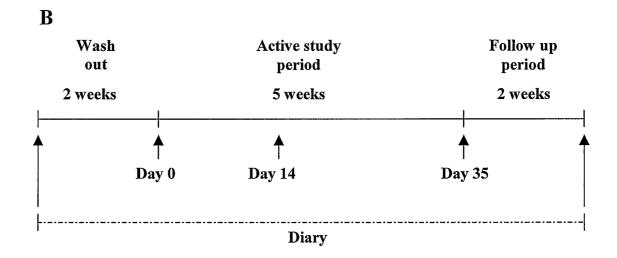
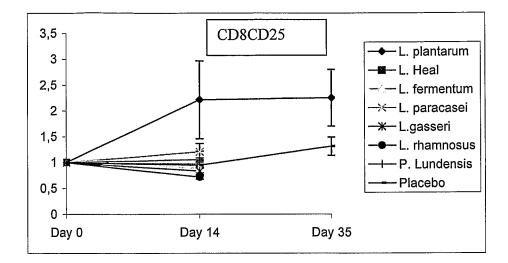
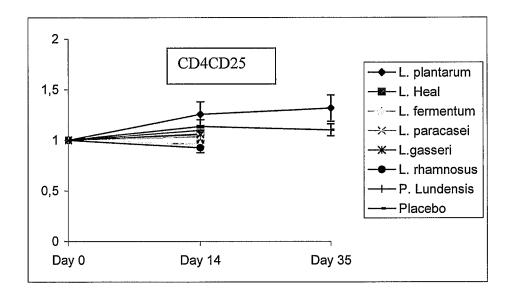


Fig. 4

Fig. 5





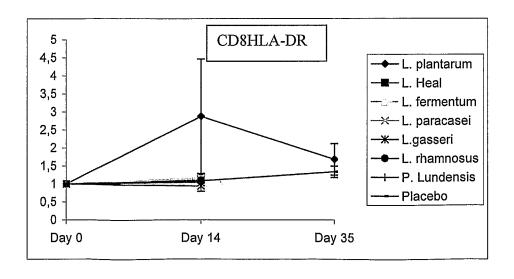


Fig. 5 (continued)

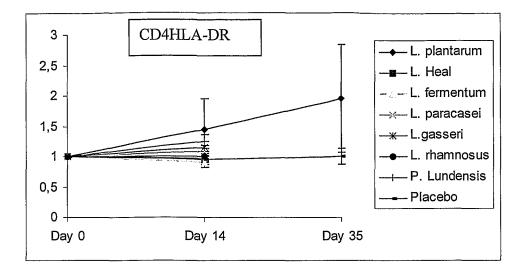
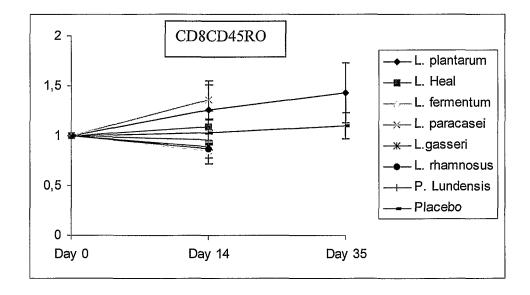
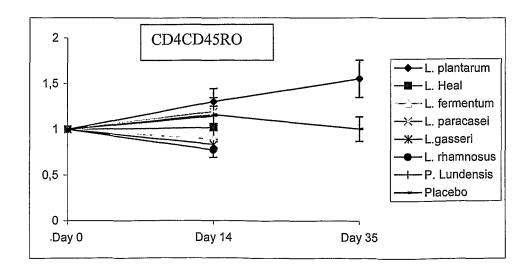


Fig. 6.





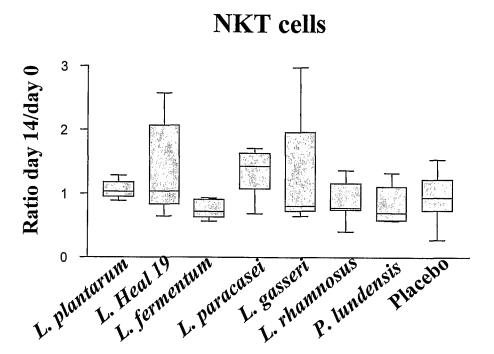


Fig. 7

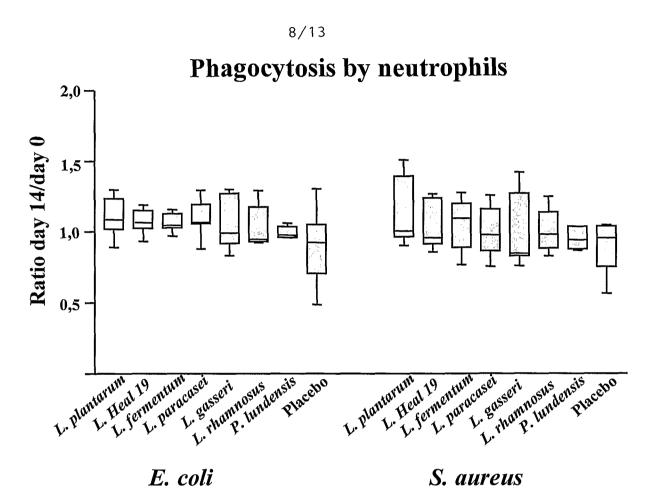
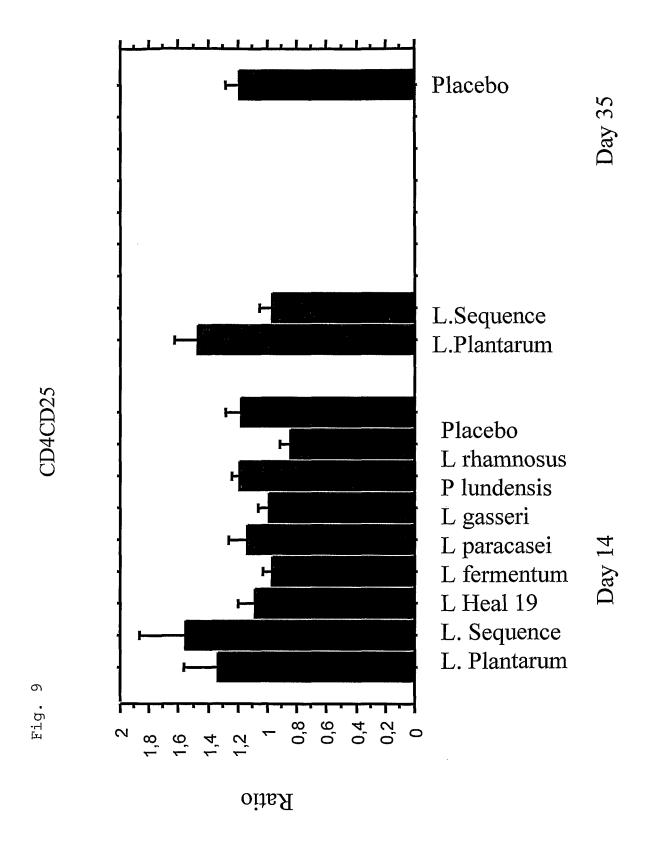
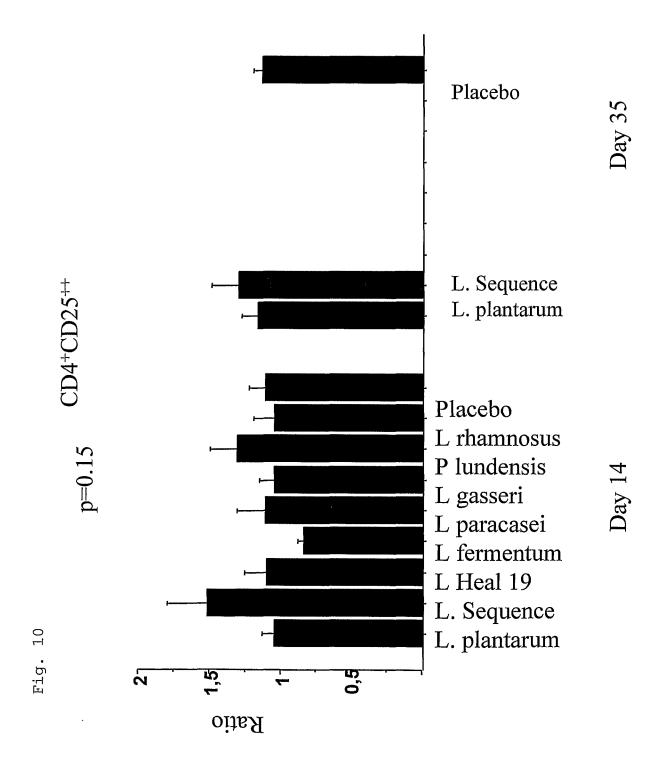


Fig. 8

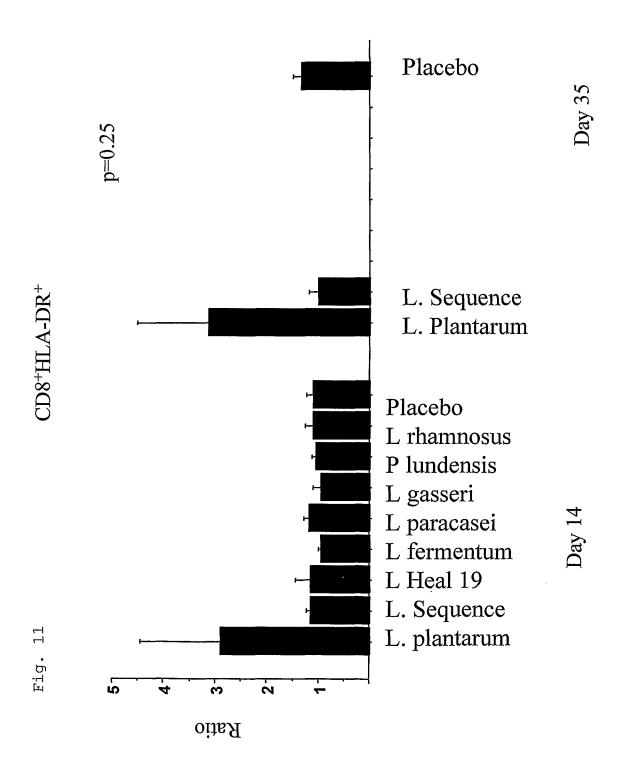


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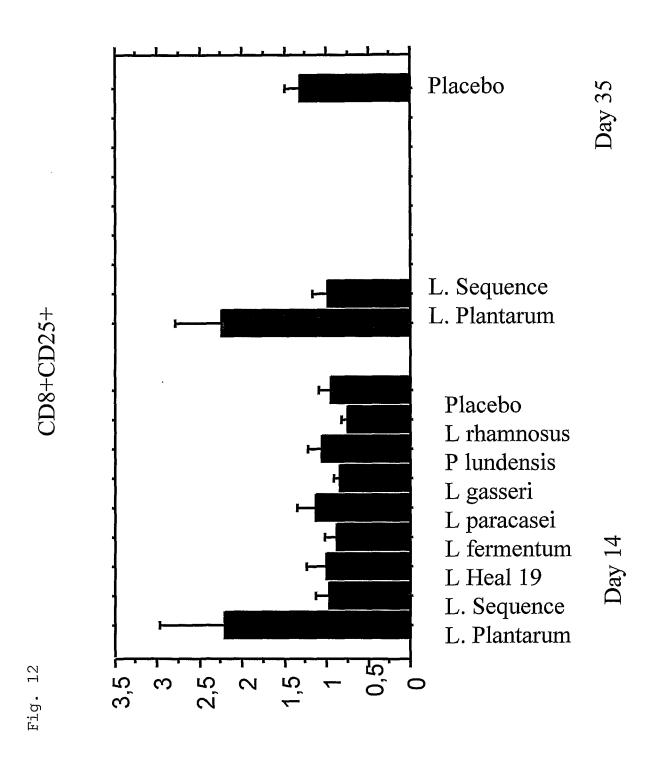


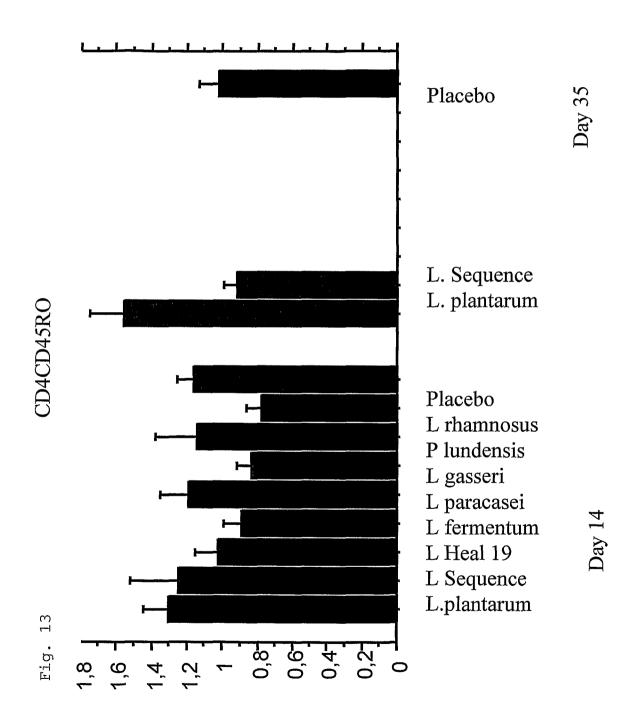
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# INDICATIONS RELATING TO DEPOSITED MICROORGANISM OR OTHER BIOLOGICAL MATERIAL

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A. The indications made below relate to the deposited microorganism or other biological material referred to in the description on page egg 5 , line 28 32 .				
B. IDENTIFICATION OF DEPOSIT	Further deposits are identified on an additional sheet X			
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Address of depositary institution (including postal code and country Mascheroder Weg 1 B D-3300 Braunschweig	y)			
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1995-03-16	DSM 9843			
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### A. CLASSIFICATION OF SUBJECT MATTER

IPC: see extra sheet
According to International Patent Classification (IPC) or to both national classification and IPC

#### B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC: A23L, A61K

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

#### SE, DK, FI, NO classes as above

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EPO-IN	TERNAL, WPI DATA, TXTE, BIOSIS, E	MBASE, MEDLINE			
C. DOCU	MENTS CONSIDERED TO BE RELEVANT				
Category*	Citation of document, with indication, where ap	propriate, of the relevant passages Relevant to claim			
Х	WO 0078322 A2 (MENDES S.R.L.), (28.12.2000), page 4, line abstract	28 December 2000 1 - line 18, claim 13,	1-5,9-14		
Υ			6-8		
Х	WO 2004103083 A1 (SYNBIOTICS AB), 2 December 2004 (02.12.2004)				
A			6-8		
1			'		
X Furth	er documents are listed in the continuation of Box	x C. X See patent family annex	۲.		
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Υ	National Library of Medicine (NLM), file Medline, Medline accession no. 10337020, Herias M V et al: "Immunomodulatory effects of colonizing the intestine of gnotobiotic rats"; & Clinical and experimental immunology, volume 116, no. 2, May 1999, page 283 - page 290	6	
Υ	National Library of Medicine (NLM), file Medline, Medline accession no. 16101937, Peng Guei-Cheng et al: "The efficacy and safety of heat-killed Lactobacillus for treatment of perennial allergic rhinitis induced by house-dust mite"; & Pediatric allergy and immunology: official publication of the European Society of Pediatric Allergy and Immunology, vol. 16, no. 5, August 2005, page 433 - page 438	7	
Y	National Library of Medicine (NLM), file Medline, Medline accession no. 12076036, Kitazawa Haruki et al: "A novel immunostimulating aspect of Lactobacillus gasseri: induction of "Gasserokine" as chemoattractants for macrophages"; & International journal of food microbiology, vol. 77, no. 1-2, 25 July 2002, page 29 - page 38	8	
A	EP 1020123 A1 (SITIA-YOMO S.P.A.), 19 July 2000 (19.07.2000), page 3, line 57; page 4, line 36, claim 2	1-14	
A	WO 2005077391 A1 (SYNBIOTICS AB), 25 August 2005 (25.08.2005)	1-14	
A	EP 0994183 A1 (RIKEN), 19 April 2000 (19.04.2000)	1-14	
A	WO 02060276 A1 (VALIO LTD), 8 August 2002 (08.08.2002)	1-14	
1			

#### International patent classification (IPC)

**A23L 1/03** (2006.01) A61K 35/74 (2006.01) A61P 31/16 (2006.01) C12R 1/20 (2006.01) C12R 1/225 (2006.01) C12R 1/25 (2006.01)

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International application No. PCT/SE2006/001137

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Thi	s inter	national search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
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25/11/2006

International application No. PCT/SE2006/001137

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