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(57) Abrégé/Abstract:

Provided are methods of treatment for skin disorders. In particular, treatment, the skin disorders are generally inflammatory skin disorders, including improper wound healing. Provided are methods of using of a cytokine molecule.



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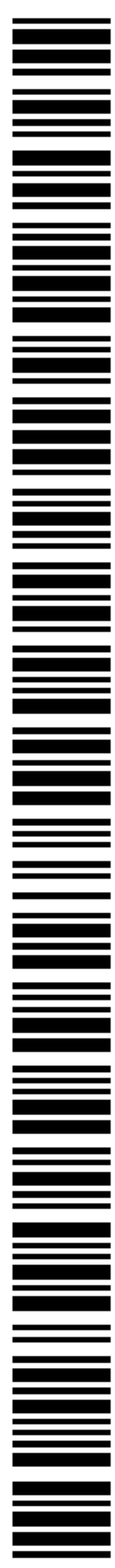
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**WO 2004/058178 A2**

## USES OF MAMMALIAN CYTOKINE; RELATED REAGENTS

5

## FIELD OF THE INVENTION

The present invention relates generally to uses of mammalian cytokine-like molecules and related reagents. More specifically, the invention relates to identification of mammalian cytokine-like proteins and inhibitors thereof that modulate skin or wound healing, e.g., inflammatory skin conditions.

10

## BACKGROUND OF THE INVENTION

Cytokines are small proteins that mediate signaling and communication between cells of the immune system, e.g., T cells, B cells, dendritic cells, and macrophages. These proteins mediate a number of cellular activities, including proliferation, growth, differentiation, migration, cell activation, and response to infection, foreign antigens, and wounds.

15

A particularly important family of cytokines is interleukin-6 (IL-6) family. These cytokines exhibit a wide range of often overlapping biological functions that are transmitted via multichain cell surface receptors, which are typically formed by high-affinity, cytokine-specific receptor chains and lower-affinity, signal-transducing chains. Receptor subunits are often shared among members of this cytokine subfamily.

20

Recently, a novel helical cytokine was identified that has structural homology to the IL-6 family of cytokines. This protein was designated p19, and was shown to be part of a novel composite factor consisting of a disulfide-bridged complex between p19 and the p40 subunit of IL-12. This novel p19p40 complex, also known as IL-23, is naturally expressed by activated mouse and human dendritic cells and has biological activities that are similar to but distinct from IL-12 (see, e.g., Oppmann *et al.* (2000) *Immunity* 13:715-725). The p19 subunit of IL-23 is also known as "IL-23p19."

25

The present invention identifies and provides IL-23, IL-23 agonists, and variants and derivatives thereof, as modulators of skin disorders, for example, for use in the treatment or diagnosis of skin conditions and disorders or wound healing, see, e.g., Fitzpatrick, *et al.* (eds.) (1993) *Dermatology in General Medicine 4th ed.*, McGraw-Hill, NY; Bos (ed.) (1989) *Skin Immune System*, CRC Press, Boca Raton, FL; Callen (1996) *General Practice Dermatology*, Appleton and Lange, Norwalk, CN; Rook, *et al.* (eds.) (1998) *Textbook of Dermatology*, Blackwell Publ., Malden, MA; Habifor and Habie (1995) *Clinical Dermatology: A Color Guide to Diagnosis and Therapy*, Mosby,

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Phila., PA.; Grob (ed.) (1997) *Epidemiology, Causes and Prevention of Skin Diseases*, Blackwell, Malden, MA; Hess and Salcido (2000) *Wound Care*, Springhouse Pub. Co., Springhouse, PA; Mani, et al. (1999) *Chronic Wound Healing: Clinical Measurement and Basic*, Balliere Tindall Ltd., London, UK; Wyngaarden and Smith (eds.) (1985) *Cecil's Textbook of Medicine*, W.B. Saunders Co., Phila., PA; Berkow (ed.) (1982) *The Merck Manual of Diagnosis and Therapy*, Merck Sharp & Dohme Research Laboratories, West Point, PA; Braunwald, et al. (eds.) (1991) *Harrison's Principles of Internal Medicine, 12th Ed.*, McGraw-Hill, Inc., NY, all of which are incorporated herein by reference.

The present invention provides methods and reagents for the treatment, prevention, and diagnosis of wounds and wound healing, e.g., burns, wounds of cartilage, nerves and spinal cord, muscle, soft tissues, blood vessels and angiogenesis, ulcers and pressure sores, bone fractures and osteoporosis, and for promoting skin growth, e.g., at harvested or donor sites used in skin grafting (see, e.g., Yamaguchi and Yoshikawa (2001) *J. Dermatol.* 28:521-534; Cairns, et al. (1993) *Arch. Surg.* 128:1246-1252; Hom, et al. (2002) *Facial Plast. Surg.* 18:41-52; Hackam and Ford (2002) *Surg. Infect.* (Larchmt.) 3(Suppl. 1):S23-S35; Oshima, et al. (2002) *Hum. Cell.* 15:118-128; Lal, et al. (2000) *Growth Horm. IGF Res.* 10 (Suppl. B):S39-S43; Rose and Herndon (1997) *Burns* 23:S19-S26; Schryvers, et al. (2000) *Arch. Phys. Med. Rehabil.* 81:1556-1562; Hidaka, et al. (2002) *Orthop. Clin. North Am.* 33:439-446; Dagum (1998) *J. Hand Ther.* 11:111-117; Coutts, et al. (2001) *Clin. Orthop.* 391 (Suppl.):S271-S279; Larsson (2002) *Scand. J. Surg.* 91:140-146; Goldstein (2000) *Clin. Orthop.* 379 (Suppl.):S113-119; Lieberman, et al. (2002) *Mol. Therapy* 6:141-147; Tuli, et al. (2003) *Arthritis Res. Ther.* 5:235-238; Li, et al. (2003) *Microsc. Res. Tech.* 60:107-114; van Hinsbergh, et al. (2001) 936:426-437; Conway, et al. (2001) *Cardiovasc. Res.* 49:507-521).

Skin wound healing involves a number of phases: inflammation, first with neutrophil and later monocyte/macrophage inflammation, new tissue formation, including matrix formation and differentiation of a neoepithelium, and finally remodeling and maturation. The initial inflammatory phase allows clot formation, controls infection, and promotes vascularization, and produces growth factors. If not controlled properly, the inflammation can lead to pathological healing, e.g., ulcers or scars.

Fibroblasts deposit provisional matrix or granulation tissue, while the newly formed provisional matrix is later degraded in a tissue remodeling process. Degradation of extracellular matrix is mediated by proteases, such as matrix metalloproteases (MMP), gelatinase, and collagenase, as well as protease inhibitors. An imbalance in matrix formation and degradation leads, at one extreme, to chronic ulcers and, on the other extreme, to fibrosis. For example, keloids, an "overhealed response," are fibrous tissue outgrowths (Michalik, et al. (2001) *J. Cell Biol.* 154:799-814; Okada, et al. (1997) *J. Cell Biol.* 137:67-77; Fedyk, et al. (2001) *J. Immunol.* 166:5749-5755;

Ravanti and Kahari (2000) *Int. J. Mol. Med.* 6:391-407; Peled, *et al.* (2000) *Clin. Past. Surg.* 27:489-500).

Matrix formation and reepithelialization depend on angiogenesis (Montesinos, *et al.* (1997) *J. Exp. Med.* 186:1615-1620; Malinda, *et al.* (1998) *J. Immunol.* 160:1001-1006).

5 Growth factors used in wound healing induce expression of anti-microbial factors, e.g., defensins, cathelicidins, secretory protease inhibitor, and gelatinase-associated lipocalin (from neutrophils) (Sorensen, *et al.* (2003) *J. Immunol.* 170:5583-5589).

In wound healing, cells such as platelets, monocyte/macrophages, T cells, and other immune cells, infiltrate the wound and produce factors that regulate growth of tissue. These factors include  
10 TGF, tumor necrosis factor (TNF), IL-1, IL-4, IL-6, oncostatin M, GRO-alpha, various angiogenic factors, and chemokines. In turn, these factors stimulate, for expression of, e.g., extracellular matrix and tissue inhibitor of metalloproteases (TIMP). (Ihn and Tamaki (2000) *J. Immunol.* 165:2149-2155; Feugate, *et al.* (2002) *J. Cell. Biol.* 156:161-172). Myofibroblasts, cells that are fibrogenic, are important for wound closure and contraction. Disease states characterized by accumulation of  
15 myofibroblasts include pulmonary fibrosis and scleroderma (Feugate, *et al.* (2002) *J. Cell Biol.* 156:161-172).

Wound healing of skin and other tissues is a complex process involving proliferation and migration of immune cells, endothelial cells, fibroblasts, stromal cells, myofibroblasts, smooth muscle cells, pericytes, and keratinocytes.

20 Parameters used to measure healing include rate of healing, breaking strength of healed wounds, degree of epithelialization, thickness of granulation tissue, and density of extracellular matrix (Matsuda, *et al.* (1998) *J. Exp. Med.* 187:297-306).

Ischemia or ischemia reperfusion, as occurs with traumatic injury and "muscle unloading"  
25 (chronic bedrest), results in neutrophil infiltration, where the neutrophils often produce tissue damage in excess to the damage caused by the ischemia. Consistent with this adverse effect of neutrophils on healing is that administration of cytokine antagonists, including antagonists to IL-1 or TNF, can also improve wound healing under certain conditions, even where the cytokine is ultimately required for normal repair (see, e.g., Ley (2003) *Am. J. Physiol. Regul. Integr. Comp. Physiol.* 285:R718-R719; Graves, *et al.* (2001) *J. Immunol.* 167:5316-5320). The present invention provides methods using an  
30 IL-23 antagonist to inhibit neutrophil-induced tissue injury, e.g., after trauma, wounding, or prolonged bedrest.

Improper wound healing, e.g., of cutaneous wounds, can result in chronic discomfort or disfigurement, and can lead to further complications, e.g., infections or dehydration. Thus, a need  
35 exists for effective treatment, both prophylactic and curative, to alleviate the symptoms of those

conditions. Alternatively, methods of diagnosis, e.g., of abnormal or modified health of those tissues will be useful. The present invention provides both.

#### SUMMARY OF THE INVENTION

5 The present invention is based, in part, upon the discovery that an IL-23 fusion protein, e.g., a fusion protein comprising the p19 subunit linked to the p40 subunit, enhanced wound healing response in various mouse models.

The invention provides a method of treating or improving healing comprising administering to a subject an effective amount of an agonist or antagonist of IL-23. Also provided is the above  
10 method, wherein the agonist or antagonist comprises a polypeptide of IL-23, or a derivative or variant thereof; a binding composition derived from an antibody that specifically binds to IL-23 or to IL-23R; or a nucleic acid encoding a polypeptide of IL-23, or a derivative or variant thereof. In addition, the invention provides the above method wherein the derivative or variant comprises an IL-23 hyperkine; wherein the agonist comprises a complex of a mature sequence of SEQ ID NO:10; and a  
15 mature sequence of SEQ ID NO:12; or the above method wherein the nucleic acid further comprises an expression vector.

In another aspect, the invention provides a method of treating or improving healing comprising administering to a subject an effective amount of an agonist or antagonist of IL-23, wherein the healing is of a skin or cutaneous wound; of an ulcer or graft; or is improper healing.  
20 Also provided is the above method wherein the treating or improving increases a pressure required to break a healed or healing wound; a stiffness of a healed or healing wound; a rate of healing of a wound; a granulation layer thickness of a healed or healing wound; recruitment of a cell to or towards a wound; or antimicrobial activity. In yet another aspect, the invention provides the above method wherein the cell is a CD11b<sup>+</sup>, MHC Class II<sup>+</sup> cell; a monocyte/macrophage; a CD31<sup>+</sup> endothelial cell;  
25 or an immune cell. Also provided is the above method wherein the recruitment is in or towards a granulation tissue; wherein the increased wound breaking pressure is about a 15% or about a 20% increase in wound breaking pressure; or the increased stiffness is about a 15% or about a 20% increase in stiffness. In another embodiment, the present invention provides the above method wherein the treating or improving  
30 comprises increased angiogenesis; or immune surveillance; or the above method wherein the increased angiogenesis is mediated by ICAM-1 or -2; or the increased immune surveillance is mediated by dendritic cells.

Yet another aspect of the above invention provides a method of treating or improving healing comprising administering to a subject an effective amount of an agonist or antagonist of IL-23,  
35 wherein the treating or improving comprises increased expression of a nucleic acid or protein of a cytokine in addition to IL-23; a signaling molecule; an anti-microbial molecule; a protease or

protease inhibitor; or a molecule of the extracellular matrix; or the above method wherein the cytokine nucleic acid or protein is IL-17, IL-6, IL-19, GRO-alpha, or GM-CSF; or wherein wherein the nucleic acid or protein is lactoferrin; DEC-205; CD50; nitric oxide synthase; or secretory leukoprotease inhibitor; or CD40L.

5 Also provided is the above method, wherein the antagonist comprises a nucleic acid; a blocking antibody to IL-23 or to IL-23R; or a soluble receptor derived from an extracellular part of IL-23R; the above method wherein the nucleic acid comprises an anti-sense nucleic acid; or interference RNA.

10 Yet another aspect of the present invention provides an agonist of IL-23 derived from the binding site of an antibody that specifically binds to an IL-23 receptor; the above agonist that is a polyclonal antibody; a monoclonal antibody; an Fab, Fv, or F(ab')<sub>2</sub> fragment; humanized; a peptide mimetic; or detectably labeled. In another embodiment, the present invention provides the above agonist comprising a complex of a polypeptide of the mature sequence of SEQ ID NO:10 and a polypeptide of the mature sequence of SEQ ID NO:12; the above agonist comprising a complex of  
15 two polypeptides of the mature sequence of SEQ ID NO:10 and two polypeptides of the mature sequence of SEQ ID NO:12. Moreover the invention provides the above agonist wherein contact of the agonist to a cell expressing hIL-23R and hIL-12beta1 results in an increase in proliferation of the cell. Also provided is a kit comprising the above agonist and a compartment; or instructions for use or disposal. Also provided is a nucleic acid encoding an agonist of IL-23 derived from the binding  
20 site of an antibody that specifically binds to an IL-23 receptor.

#### DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

25 All references cited herein are incorporated herein by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

As used herein, including the appended claims, the singular forms of words such as "a," "an," and "the," include their corresponding plural references unless the context clearly dictates otherwise.

30

#### I. General.

Interleukin-23 (IL-23) is a heterodimeric cytokine composed of a novel p19 subunit (a.k.a. IL-B30) and the p40 subunit of IL-12 (Oppmann, *et al.*, *supra*). The p19 subunit was identified during a computational search for members of the IL-6 helical cytokine family characterized by their  
35 unique four  $\alpha$ -helix bundle. Genetic analysis of the family, of which oncostatin-M, IL-11, cardiotrophin-1, and leukaemia inhibitory factor are members, reveals the closest evolutionary neighbor of p19 to be the p35 subunit of IL-12. Like p35, p19 requires co-expression of p40 for

biological activity (Wiekowski, *et al.* (2001) *J. Immunol.* 166:7563-7570). The IL-23 receptor (IL-23R) comprises a novel receptor subunit (IL-23R), that binds p19, and IL-12R $\beta$ 1, that binds p40 (Parham, *et al.* (2002) *J. Immunol.* 168:5699-5708). These two receptor subunits form the functional signaling complex and are expressed on CD4<sup>+</sup>CD45Rb<sup>lo</sup> memory T cells as well as interferon-  
5 gamma (IFN $\gamma$ ) activated bone marrow macrophages (Parham, *et al.*, *supra*).

Preliminary characterization of IL-23 suggests that it has potent effects on memory T cells from both humans and mice, as measured by proliferation and IFN $\gamma$  production. Consistent with the immunostimulatory properties of IL-23, mice in which haematopoietic cells constitutively express transgenic p19 have widespread multi-organ inflammation that results in premature death  
10 (Wiekowski, *et al.*, *supra*). The inflammatory disease is characterized by intense macrophage infiltration, neutrophilia, and elevated levels of proinflammatory monokines such as IL-1 and TNF, suggesting that IL-23 may also act on myeloid cells.

Recent studies analyzing the necessity of IL-12 in resistance to infectious diseases have yielded divergent results, depending on whether p35<sup>-/-</sup> or p40<sup>-/-</sup> mice are used. The former, which  
15 specifically lack IL-12 but express IL-23, are resistant to infection, whereas the latter, unable to express both IL-12 and IL-23, are more susceptible.

Transgenic mice deficient for the p19 subunit of IL-23 (IL-23p19) were resistant to EAE, a CNS autoimmune disease mediated by TH1 cells and inflammatory macrophages, while wild-type and heterozygous p19 control mice were highly susceptible. Mice deficient in the p40 subunit of IL-  
20 12 (IL-12p40 deficient mice) were also resistant to EAE, while mice deficient in the p35 subunit of IL-12 (IL-12p35 deficient mice) were highly susceptible to EAE. This is indicative of a role of IL-23 in the induction of EAE.

p19 deficient mice had a notable altered wound healing response following subcutaneous injection of an oil emulsion in mice. The p19 deficient mice were also defective in a variety of  
25 mouse disease models that required monocyte/macrophage activation. Monocytes/macrophages are known to stimulate wound repair, see, e.g., Schaffer and Nanney (1996) *Intl. Rev. Cytol* 169:151-181. In particular, it is shown below that delivery of IL-23 polypeptide into mouse skin could attract CD11b<sup>+</sup>/Class II<sup>+</sup> activated monocyte/macrophage populations.

## 30 II. Definitions.

“Agonist of IL-23” and “IL-23 agonist” encompasses an agonistic antibody that specifically binds to IL-23 receptor (IL-23R) and increases the signaling properties of IL-23R. Agonist of IL-23 also encompasses an agonistic antibody that specifically binds to the complex of IL-23R and IL-12R $\beta$ 1.

A "blocking antibody" encompasses, e.g., an antibody that specifically binds to IL-23 and prevents or impairs signaling mediated by IL-23 and IL-23R. A blocking antibody also encompasses an antibody that specifically binds to IL-23R and prevents or impairs signaling mediated by IL-23R, or mediated by IL-23 and IL-23R.

5 "Conservatively modified variants" applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refers to those nucleic acids which encode identical or essentially identical amino acid sequences, or where the nucleic acid does not encode an amino acid sequence, to essentially identical nucleic acid sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids  
10 may encode any given protein.

As to amino acid sequences, one of skill will recognize that an individual substitution to a nucleic acid, peptide, polypeptide, or protein sequence which substitutes an amino acid or a small percentage of amino acids in the encoded sequence for a conserved amino acid is a "conservatively modified variant." Conservative substitution tables providing functionally similar amino acids are  
15 well known in the art. An example of a conservative substitution is the exchange of an amino acid in one of the following groups for another amino acid of the same group (U.S. Pat. No. 5,767,063 issued to Lee, *et al.*; Kyte and Doolittle (1982) *J. Mol. Biol.* 157:105-132):

- (1) Hydrophobic: Norleucine, Ile, Val, Leu, Phe, Cys, or Met;
- (2) Neutral hydrophilic: Cys, Ser, Thr;
- 20 (3) Acidic: Asp, Glu;
- (4) Basic: Asn, Gln, His, Lys, Arg;
- (5) Residues that influence chain orientation: Gly, Pro;
- (6) Aromatic: Trp, Tyr, Phe;
- (7) Small amino acids: Gly, Ala, Ser.

25 The phrase "effective amount" means an amount sufficient to ameliorate a symptom or sign of the medical condition. Typical mammalian hosts will include mice, rats, cats, dogs, and primates, including humans. An effective amount for a particular patient may vary depending on factors such as the condition being treated, the overall health of the patient, the method route and dose of administration and the severity of side affects. When in combination, an effective amount is in ratio  
30 to a combination of components and the effect is not limited to individual components alone.

An "endpoint" for assessing or diagnosing improved healing, e.g., wound healing, includes, without limitation: wound breaking pressure; stiffness, looseness; immune surveillance; angiogenesis; wound-related anti-microbial activity; inflammation, e.g., by neutrophils; degree of expression of genes or polypeptides indicative of inflammation, angiogenesis, reepithelialization,  
35 anti-microbial action, and remodeling, e.g., matrix formation, matrix breakdown, or granulization.

As determined by a suitable endpoint, the present invention provides a method to improve healing by 10% or more, more generally by 15% or more, most generally by 20% or more, typically 25% or more, more typically by 30% or more, most typically by 35% or more, often by 40% or more, more often by 50% or more, most often by 60% or more, usually by 70% or more, more usually by 80% or more, most usually by 90% or more, ideally by 100% (i.e., 2-fold) or more, more ideally by 4-fold or more, and most ideally by 8-fold or more. As determined by a suitable endpoint, the present invention also provides a method to improve healing by about 10%, more generally by about 15%, most generally by about 20%, typically about 25%, more typically by about 30%, most typically by about 35%, often by about 40%, more often by about 50%, most often by about 60%, usually by about 70%, more usually by about 80%, most usually by about 90%, ideally by about 100% (i.e., 2-fold), more ideally by about 4-fold, and most ideally by about 8-fold.

“Improper wound healing” encompasses the absence or abnormally slow progression of the healing of a wound, e.g., delayed re-epithelialization. Improper wound healing can be found, e.g., in diabetic ulcers and abscesses, pressure ulcers, infected wounds, burns, advanced age, inadequate perfusion, and obesity. Improper wound healing also encompasses injuries leading to scars or to persistent infections, see, e.g., Singer and Clark (1999) *New Engl. J. Med.* 341:738-746; Calhoun, *et al.* (2002) *Adv. Skin Wound Care* 15:31-45; Rico, *et al.* (2002) *J. Surg. Res.* 102:193-197; Thomas (2001) *Cleve. Clin. J. Med.* 68:704-722; Ashcroft, *et al.* (2002) *Biogerontology* 3:337-345; Thomason (1999) *Home Care Provid.* 4:156-161; Gallagher (1997) *Ostomy. Wound Manage.* 43:18-27.

Parameters and endpoints used to assess wound healing and response to therapeutic, pharmacological, and diagnostic agents, include a number of histological, physiological, and biochemical parameters, e.g., infiltration, activation, or differentiation of neutrophils, monocytes, and macrophages, e.g., differentiation of monocytes to reparative macrophages, and appearance of new stroma, blood vessels, and nerves. Suitable parameters also include expression levels of signaling agents, e.g., transforming growth factor, interleukin-1, and insulin-like growth factor. Measures of epithelization, e.g., rate and thickness, migration of epidermal cells, granulation thickness, degradation and maturation of extracellular matrix, e.g., provisional matrix versus collagenous matrix, wound strength (breaking strength), and fibroblast proliferation rate and phenotype, are also suitable parameters. Increased granulation tissue thickness can resulting stronger healed wounds (see, e.g., Singer and Clark, *supra*, Werner and Grose (2002) *Physiol. Rev.* 83:835-870; Matsuda, *et al.* (1998) *J. Exp. Med.* 187:297-306; Wankell, *et al.* (2001) *EMBO J.* 20:5361-5372).

A composition that is “labeled” is detectable either directly or indirectly by, e.g., spectroscopic, photochemical, biochemical, metabolic, immunochemical, isotopic, or chemical methods. For example, useful labels include epitope tags, fluorettes, <sup>32</sup>P, <sup>33</sup>P, <sup>35</sup>S, <sup>14</sup>C, <sup>3</sup>H, <sup>125</sup>I, stable isotopes, fluorescent compounds, electron-dense reagents, substrates, or enzymes, e.g., as used in enzyme-linked immunoassays, see, e.g., Invitrogen (2002) *Catalogue*, Carlsbad, CA; Molecular

Probes (2002) *Catalogue*, Molecular Probes, Eugene, OR; Rozinov and Nolan (1998) *Chem. Biol.* 5:713-728.

Increase in "anti-microbial activity" encompasses an increase in expression, concentration, or level of a mediator of anti-microbial activity, both in the presence and absence of demonstrated  
5 reduction in biological activity, concentration, population, or number of a microbe in contact with a human or animal subject or host. The mediator of anti-microbial activity can be, e.g., an immune cell responsive to a bacterial, viral, fungal, or protozoal, or parasitic antigen, or an anti-microbial molecule, such as a defensin. "Anti-microbial activity" embraces, e.g., phagocytosis and any activity that is generally or usually associated with phagocytosis, e.g., exposure of a microbe to toxic oxygen.  
10 "Anti-microbial activity" also encompasses an change in expression, concentration, or level of a cell, gene, protein, or small molecule that is generally or usually associated with anti-microbial action, e.g., an increase in expression of a neutrophil gene. "Anti-microbial activity" is not limited to an increase in expression, i.e., it also encompasses a decrease in expression, where that decrease promotes anti-microbial activity.

15 "Proliferation" or "rate of proliferation" can be measured, e.g., by assessing the increase in cell number over a predetermined period or interval of time, or by the number or proportion of cells in S phase at any given point in time.

### III. Agonists and Antagonists.

20 The present invention provides methods of using IL-23 agonists including the full length cytokine protein (SEQ ID NO: 2 or 4). Also provided is a fusion protein, also known as "IL-23 hyperkine" (SEQ ID NO: 6 or 8), comprising p19 linked to p40 with a FLAG sequence as described for IL-6 in, e.g., Oppmann, *et al.*, *supra*; Fischer, *et al.* (1997) *Nature Biotechnol.* 15:142-145; Rakemann, *et al.* (1999) *J. Biol. Chem.* 274:1257-1266; and Peters, *et al.* (1998) *J. Immunol.*  
25 161:3575-3581, thereof. The invention also provides agonistic anti-IL-23R antibodies that are agonistic to the IL-23 receptor, e.g., antibodies that stimulate the IL-23 receptor in the absence or presence of IL-23.

Peptides of those sequences, or variants thereof, will be used to induce receptor signaling. Also contemplated are small molecules which also induce receptor signaling. Agonists of the present  
30 invention will be useful in the treatment of various inflammatory skin disorders, including but not limited to wound healing, skin disorders associated with impaired recruitment of myeloid/monocyte cells.

The invention provides IL-23 antagonists, e.g., a blocking antibody that binds to IL-23, a blocking antibody that binds to IL-23R, a soluble receptor based on the extracellular portion of IL-  
35 23R, and nucleic acids. The IL-23 antagonists of the present invention encompass nucleic acids that are anti-sense nucleic acids and RNA interference nucleic acids (see, e.g., Arenz and Schepers (2003)

*Naturwissenschaften* 90:345-359; Sazani and Kole (2003) *J. Clin. Invest.* 112:481-486; Pirollo, *et al.* (2003) *Pharmacol. Therapeutics* 99:55-77; Wang, *et al.* (2003) *Antisense Nucl. Acid Drug Devel.* 13:169-189).

5 III. Antibodies and Related Reagents.

Antibodies and binding compositions derived from an antigen-binding site of an antibody are provided. These include humanized antibodies, monoclonal antibodies, polyclonal antibodies, and binding fragments, such as Fab, F(ab)<sub>2</sub>, and Fv fragments, and engineered versions thereof. The antibody or binding composition may be agonistic or antagonistic. Antibodies that simultaneously  
10 bind to a ligand and receptor are contemplated. Monoclonal antibodies will usually bind with at least a K<sub>D</sub> of about 1 mM, more usually at least about 300 μM, typically at least about 100 μM, more typically at least about 30 μM, preferably at least about 10 μM, and more preferably at least about 3 μM or better.

Monoclonal, polyclonal, and humanized antibodies can be prepared. See, e.g., Sheperd and  
15 Dean (eds.) (2000) *Monoclonal Antibodies*, Oxford Univ. Press, New York, NY; Kontermann and Dubel (eds.) (2001) *Antibody Engineering*, Springer-Verlag, New York; Harlow and Lane (1988) *Antibodies A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, pp. 139-243; Carpenter, *et al.* (2000) *J. Immunol.* 165:6205; He, *et al.* (1998) *J. Immunol.* 160:1029; Tang, *et al.* (1999) *J. Biol. Chem.* 274:27371-27378).

20 Single chain antibodies, single domain antibodies, and bispecific antibodies are described, see, e.g., Malecki, *et al.* (2002) *Proc. Natl. Acad. Sci. USA* 99:213-218; Conrath, *et al.* (2001) *J. Biol. Chem.* 276:7346-7350; Desmyter, *et al.* (2001) *J. Biol. Chem.* 276:26285-26290, Kostelney, *et al.* (1992) *J. Immunol.* 148:1547-1553; U.S. Pat. Nos. 5,932, 448; 5,532,210; 6,129,914; 6,133,426; 4,946,778.

25 The invention also encompasses deamidated binding compositions, e.g., antibodies, and methods of using deamidated binding compositions (see, e.g., Zhang and Czupryn (2003) *J. Pharm. Biomed. Anal.* 30:1479-1490; Perkins, *et al.* (2000) *Pharm. Res.* 17:1110-1117; Lehrman, *et al.* (1992) *J. Protein Chem.* 11:657-663).

Antigen fragments may be joined to other materials, such as fused or covalently joined  
30 polypeptides, to be used as immunogens. An antigen and its fragments may be fused or covalently linked to a variety of immunogens, such as keyhole limpet hemocyanin, bovine serum albumin, or ovalbumin (Coligan, *et al.* (1994) *Current Protocols in Immunol.*, Vol. 2, 9.3-9.4, John Wiley and Sons, New York, NY). Peptides of suitable antigenicity can be selected from the polypeptide target, using an algorithm, such as those of Parker, *et al.* (1986) *Biochemistry* 25:5425-5432; Welling, *et al.*  
35 (1985) *FEBS Lett.* 188:215-218; Jameson and Wolf (1988) *Cabios* 4:181-186; or Hopp and Woods (1983) *Mol. Immunol.* 20:483-489.

Purification of antigen is not necessary for the generation of antibodies. Immunization can be performed by DNA vector immunization. See, e.g., Wang, *et al.* (1997) *Virology* 228:278-284. Alternatively, animals can be immunized with cells bearing the antigen of interest. Splenocytes can then be isolated from the immunized animals, and the splenocytes can fused with a myeloma cell line to produce a hybridoma. Resultant hybridomas can be screened for production of the desired antibody by functional assays or biological assays, that is, assays not dependent on possession of the purified antigen. Immunization with cells may prove superior for antibody generation than immunization with purified antigen (Meyaard, *et al.* (1997) *Immunity* 7:283-290; Wright, *et al.* (2000) *Immunity* 13:233-242; Preston, *et al.* (1997) *Eur. J. Immunol.* 27:1911-1918; Kaithamana, *et al.* (1999) *J. Immunol.* 163:5157-5164).

Antibody affinity, i.e., antibody to antigen binding properties can be measured, e.g., by surface plasmon resonance or enzyme linked immunosorbent assay (ELISA) (see, e.g., Maynard and Georgiou (2000) *Annu. Rev. Biomed. Eng.* 2:339-376; Karlsson, *et al.* (1991) *J. Immunol. Methods* 145:229-240; Neri, *et al.* (1997) *Nat. Biotechnol.* 15:1271-1275; Jonsson, *et al.* (1991) *Biotechniques* 11:620-627; Friguet, *et al.* (1985) *J. Immunol. Methods* 77:305-319; Hubble (1997) *Immunol. Today* 18:305-306).

Antibodies of the present invention will usually bind with at least a  $K_D$  of about  $10^{-3}$  M, more usually at least  $10^{-6}$  M, typically at least  $10^{-7}$  M, more typically at least  $10^{-8}$  M, preferably at least about  $10^{-9}$  M, and more preferably at least  $10^{-10}$  M, and most preferably at least  $10^{-11}$  M (see, e.g., Presta, *et al.* (2001) *Thromb. Haemost.* 85:379-389; Yang, *et al.* (2001) *Crit. Rev. Oncol. Hematol.* 38:17-23; Carnahan, *et al.* (2003) *Clin. Cancer Res.* (Suppl.) 9:3982s-3990s; Wilchek, *et al.* (1984) *Meth. Enzymol.* 104:3-55).

Antibodies to IL-23R, where the anti-IL-23R antibody has substantially the same nucleic acid and amino acid sequence as those recited herein, but possessing substitutions that do not substantially affect the functional aspects of the nucleic acid or amino acid sequence, are within the definition of the contemplated invention. Variants with truncations, deletions, additions, and substitutions of regions which do not substantially change the biological functions of these nucleic acids and polypeptides are also within the definition of the contemplated invention.

A humanized antibody encompasses a human antibody, antibody fragment, single chain antibody, and the like, that has one or more amino acid residues introduced into it from a source which is non-human (import antibody). The amino acids used for grafting may comprise the entire variable domain of the source, one or more of the complementary determining regions (CDRs) of the source, or all six of the CDRs of the source antibody. With grafting of the import amino acids or polypeptide regions on to the host antibody, the corresponding amino acids or regions of the host antibody are generally removed. A humanized antibody will comprise substantially all of at least one, and typically two, variable domains (Fab, Fab', F(ab')<sub>2</sub>, Fabc, Fv) in which all or substantially all

of the CDR regions correspond to those of a non-human immunoglobulin and all or substantially all of the framework regions are those of a human immunoglobulin consensus sequence. The framework regions and CDRs are highly conserved in sequence and conformation and can be accurately predicted, e.g., for use in grafting CDRs into an acceptor human antibody framework.

5 CDR regions can be grafted into a naturally occurring human acceptor framework, or in a consensus framework derived from many human antibodies. A number of human variable light (VL) and variable heavy (VH) consensus sequences have been identified. For humanization, a chain of the mouse antibody can be compared with the available human framework chains, where the human chain of closest homology is chosen for grafting (see, e.g., Maynard and Georgiou, *supra*; Li, *et al.* 10 (2002) *Immunol. Revs.* 190:53-68; Co, *et al.* (1991) *Proc. Natl. Acad. Sci. USA* 88:2869-2873; Sims, *et al.* (1993) *J. Immunol.* 151:2296-2308; Sato, *et al.* (1994) *Mol. Immunol.* 31:371-381; Morea, *et al.* (2000) *Methods* 20:267-279; Kabat *et al.* (1991) *Sequences of Proteins of Immunological Interest*, 5<sup>th</sup> ed., 4 vol., U.S. Department of Health Human Services, NIH, USA; U.S. Pat. No. 6,538,111, issued to Koike, *et al.*; U.S. Pat. No. 6,329,511, issued to Vasquez, *et al.*).

15 The humanized antibody of the present invention also encompasses substitutions, deletions, and/or insertions, using standard techniques of site-directed mutagenesis, e.g., those used for alanine scanning, see, e.g., Jin and Wells (1994) *Protein Sci.* 3:2351-2357; Cunningham and Wells (1997) *Curr. Opin. Struct. Biol.* 7:457-462; Jones, *et al.* (1998) *J. Biol. Chem.* 273:11667-11674; U.S. Pat. No. 4,816,567 issued to Cabilly, *et al.*

20 Embodiments of the present invention encompass fusion proteins, purification tags, and epitope tag, at an N-terminus, C-terminus, or positions within the polypeptide, e.g., FLAG tag and GSH-S transferase fusion protein. Amino acid changes can alter, add, or eliminate post-translational processes of the agonist anti-IL-23R antibody, e.g., sites for O- and N-glycosylation, and positions of cysteine residues used for disulfide formation, see, e.g., Wright and Morrison (1997) *Trends* 25 *Biotechnol.* 15:26-32; Kunkel, *et al.* (2000) *Biotechnol. Prog.* 16:462-470.

Binding properties of the humanized antibody can be improved by the following procedure, e.g., involving site-directed mutagenesis. Computer modeling allows visualization of which mouse framework amino acid residues are likely to interact with mouse CDRs. These "contacting" mouse framework amino acids are then superimposed on the homologous human framework. Where the 30 superimposition indicates that the mouse "contacting" framework amino acid is different from the corresponding human framework amino acid, human amino acid is changed to the corresponding mouse framework amino acid. "Contact" means interchain contact between a light chain and heavy chain, where, e.g., the amino acids are predicted to be within about 3 Angstroms of each other.

Site-directed mutagenesis can also be desirable where the amino acid of the human 35 framework is rare for that position and the corresponding amino acid in the mouse immunoglobulin is common for that position in human immunoglobulin sequences. Here, the human framework amino

acid can be mutated to the corresponding donor framework amino acid, see, e.g., U.S. Pat. No. 6,407,213, issued to Carter *et al.*; U.S. Pat. No. 6,180,370, issued to Queen, *et al.*, Jung, *et al.* (2001) *J. Mol. Biol.* 309:701-716.

The humanized antibody can comprise at least a portion of an immunoglobulin constant region (Fc), e.g., of a human immunoglobulin. The antibody can optionally include the CH1, hinge, CH2, CH3, and CH4 regions of the heavy chain. The humanized antibody can be selected from any class of immunoglobulins, including IgM, IgG, IgD, IgA and IgE, and any isotype, including IgG1, IgG2, IgG3 and IgG4. Standard methods can be used to improve, or remove, effector function. Effector function includes binding to FcRn, FcγR, and complement. Half-life can be improved, e.g., by using human IgG2 or IgG4 subclasses or by altering residues in the hinge region (see, e.g., Clark (2000) *Immunol. Today* 21:397-402; Presta, *et al.* (2002) *Biochem. Soc. Trans.* 30:487-490; Morea, *et al.* (2000) *Methods* 20:267-279).

The CDR and framework regions of the humanized antibody need not correspond precisely to the import or host sequences, e.g., these sequences can be mutagenized by substitution, insertion or deletion of at least one residue so that the residue at that site does not correspond to either the consensus or the import antibody. Such mutations, however, will not be extensive. Usually, at least 75% of the humanized antibody residues will correspond to those of the parental FR and CDR sequences, more often 90%, and most preferably greater than 95%.

Ordinarily, amino acid sequence variants of the humanized anti-IL-23R antibody will have an amino acid sequence having at least 75% amino acid sequence identity with the original humanized antibody amino acid sequences of either the heavy or the light chain (e.g. as in SEQ ID NOs:2 and 4), more preferably at least 80%, more preferably at least 85%, more preferably at least 90%, and most preferably at least 95%. Identity or homology with respect to this sequence is defined herein as the percentage of amino acid residues in the candidate sequence that are identical with the humanized anti-IL-23R residues, after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent sequence identity, and not considering any conservative substitutions as part of the sequence identity. None of N-terminal, C-terminal, or internal extensions, deletions, or insertions into the antibody sequence shall be construed as affecting sequence identity or homology.

An alternative to humanization is to use human antibody libraries displayed on phage or human antibody libraries contained in transgenic mice (see, e.g., Vaughan, *et al.* (1996) *Nat. Biotechnol.* 14:309-314; Barbas (1995) *Nature Med.* 1:837-839; de Haard, *et al.* (1999) *J. Biol. Chem.* 274:18218-18230; McCafferty *et al.* (1990) *Nature* 348:552-554; Clackson *et al.* (1991) *Nature* 352:624-628; Marks *et al.* (1991) *J. Mol. Biol.* 222:581-597; Mendez, *et al.* (1997) *Nature Genet.* 15:146-156; Hoogenboom and Chames (2000) *Immunol. Today* 21:371-377; Barbas, *et al.* (2001) *Phage Display: A Laboratory Manual*, Cold Spring Harbor Laboratory Press, Cold Spring

Harbor, New York; Kay, *et al.* (1996) *Phage Display of Peptides and Proteins: A Laboratory Manual*, Academic Press, San Diego, CA; de Bruin, *et al.* (1999) *Nat. Biotechnol.* 17:397-399).

#### IV. Nucleic Acids, Vectors, and Protein Purification.

5 An "expression vector" is a nucleic acid construct, generated recombinantly or synthetically, with one or more predetermined nucleic acid elements that permit transcription of a particular nucleic acid. Typically, the expression vector includes a nucleic acid to be transcribed operably linked to a promoter.

The light chain and heavy chain of the agonistic anti-IL-23R antibody can be encoded by one  
10 nucleic acid, where expression of the light chain is operably linked to a first promoter, and where expression of the heavy chain is operably linked to a second promoter. Alternatively, both light and heavy chains can be encoded by one nucleic acid, where expression of both chains is operably linked to one promoter. The nucleic acid or nucleic acids encoding the light chain and the heavy chain can be provided as one or as two vectors. The methods of the present invention encompass incorporation  
15 of the one or two vectors into the genome of a host cell (see, e.g., Chadd and Chamow (2001) *Curr. Opin. Biotechnol.* 12:188-194; Houdebine (2000) *Transgenic Res.* 9:305-320; Stoger, *et al.* (2002) *Curr. Opin. Biotechnol.* 13:161-166).

The nucleic acid encoding the light chain can further comprise a first vector, while the nucleic acid encoding the heavy chain can further comprise a second vector. Alternatively, one  
20 vector may comprise the nucleic acids encoding the light chain and the heavy chain.

For long-term or scaled-up expression of the agonistic anti-IL-23R antibody, one vector, containing the nucleic acids encoding both the light chain and the heavy chain, can be incorporated into the host genome, e.g., where incorporation is at one point or a plurality of points in the host genome. Coexpression of the light chain and heavy chain in a host cell produces a soluble antibody.  
25 The host cell can be, e.g., a mammalian, transformed or immortalized, insect, plant, yeast, or bacterial cell. The host cell may further comprise a transgenic animal. Combinations of the above embodiments are contemplated, e.g., where the light chain is simultaneously expressed by a vector that is incorporated in the host cell's genome and by a vector that is not incorporated in the genome.

Purification of an antibody, or fragments thereof, can involve ion exchange chromatography,  
30 immunoprecipitation, epitope tags, affinity chromatography, high pressure liquid chromatography, and use of stabilizing agents, detergents or emulsifiers (Dennison and Lovrien (1997) *Protein Expression Purif.* 11:149-161; Murby, *et al.* (1996) *Protein Expression Purif.* 7:129-136; Ausubel, *et al.* (2001) *Curr. Protocols Mol. Biol., Vol. 3*, John Wiley and Sons, New York, NY, pp. 17.0.1-17.23.8; Rajan, *et al.* (1998) *Protein Expression Purif.* 13:67-72; Amersham-Pharmacia (2001)  
35 *Catalogue*, Amersham-Pharmacia Biotech, Inc., pp. 543-567, 605-654; Gooding and Regnier (2002) *HPLC of Biological Molecules, 2<sup>nd</sup> ed.*, Marcel Dekker, NY).

## V. Kits.

This invention contemplates an agonistic anti-IL23R antibody, fragments thereof, nucleic acids encoding an agonistic anti-IL-23R antibody, or fragments thereof, in a diagnostic kit.

5 Encompassed is the use of binding compositions, including antibodies or antibody fragments, for the detection of IL-23R and metabolites and breakdown products thereof, and for the detection of IL-23R-dependent activities, e.g., biochemical or cellular activity. Conjugated antibodies are useful for diagnostic or kit purposes, and include antibodies coupled with a label or polypeptide, e.g., a dye, isotopes, enzyme, or metal, see, e.g., Le Doussal, *et al.* (1991) *J. Immunol.* 146:169-175; Gibellini, *et al.* (1998) *J. Immunol.* 160:3891-3898; Hsing and Bishop (1999) *J. Immunol.* 162:2804-2811; Everts, 10 *et al.* (2002) *J. Immunol.* 168:883-889.

The invention provides a kit, where the kit comprises a compartment containing an agonistic anti-IL-23R antibody, an antigenic fragment thereof, or a nucleic acid encoding an agonistic anti-IL-23R antibody, or a fragment thereof. In another embodiment the kit has a compartment, a nucleic acid, e.g., a probe, primer, or molecular beacon, see, e.g., Zammateo, *et al.* (2002) *Biotech. Annu. Rev.* 8:85-101; Klein (2002) *Trends Mol. Med.* 8:257-260. 15

The kit may comprise, e.g., a reagent and a compartment, a reagent and instructions for use, or a reagent with both a compartment and instructions for use. A kit for determining the binding of a test compound, e.g., acquired from a biological sample or from a chemical library, can comprise a 20 control compound, a labeled compound, and a method for separating free labeled compound from bound labeled compound. Diagnostic assays can be used with biological matrices such as live cells, cell extracts and lysates, fixed cells, cell cultures, bodily fluids, or forensic samples. Various assay formats exist, such as radioimmunoassays (RIA), ELISA, and lab on a chip (U.S. Pat. Nos. 6,176,962 and 6,517,234).

25 The method can further comprise contacting a sample from a control subject, normal subject, or normal tissue or fluid from the test subject, with the binding composition. Moreover, the method can additionally comprise comparing the specific binding of the composition to the test subject with the specific binding of the composition to the normal subject, control subject, or normal tissue or fluid from the test subject. Expression or activity of a test sample or test subject can be compared 30 with that from a control sample or control subject. A control sample can comprise, e.g., a sample of non-affected or non-inflamed tissue in a patient suffering from an immune disorder. Expression or activity from a control subject or control sample can be provided as a predetermined value, e.g., acquired from a statistically appropriate group of control subjects.

35 VI. Diagnostic uses; Therapeutic Compositions; Methods.

The present invention provides methods for the treatment and diagnosis of healing, improper healing, wound healing, and improper wound healing, e.g., of the skin. Provided are methods of improving normal wound healing, e.g., by improving the rate of healing, and of treating improper wound healing, e.g., wounds characterized by ulcers or excess fibrosis. Moreover, the invention provides methods of treating and preventing wound-related infections.

Gene therapy of skin disorders may be performed using a variety of methods. Delivery vehicles are well described in the art, see, e.g., Boulikas (1998) *Gene Therapy and Molecular Biology, Vol. 1*, Gene Therapy Press, Palo Alto, CA; Jolly, *et al.* (1994) *Cancer Gene Therapy* 1:51-64; Kimura, *et al.* (1994) *Human Gene Therapy* 5:845-852; and Kaplitt, *et al.* (1994) *Nat. Genetics* 6:148-153.

To prepare pharmaceutical or sterile compositions including a cytokine or a small molecule agonist, the entity is admixed with a pharmaceutically acceptable carrier or excipient which is preferably inert. Preparation of such pharmaceutical compositions is known in the art, see, e.g., *Remington's Pharmaceutical Sciences* and *U.S. Pharmacopeia: National Formulary*, Mack Publishing Company, Easton, PA (1984).

Cytokines are normally administered parentally, preferably intravenously. Since such proteins or peptides may be immunogenic they are preferably administered slowly, either by a conventional IV administration set or from a subcutaneous depot, e.g. as taught by Tomasi, *et al.*, U.S. patent 4,732,863. Means to minimize immunological reactions may be applied. Small molecule entities may be orally active. For treatment of skin disorders, the present invention may also be administered topically, see, e.g., Gilman, *et al.* (eds.) (1990) *Goodman and Gilman's: The Pharmacological Bases of Therapeutics, 8th ed.*, Pergamon Press; and *Remington's Pharmaceutical Sciences, 17th ed.* (1990), Mack Publishing Co., Easton, Penn.

Parenteral therapeutics may be administered in aqueous vehicles such as water, saline, or buffered vehicles with or without various additives and/or diluting agents. Alternatively, a suspension, such as a zinc suspension, can be prepared to include the peptide. Such a suspension can be useful for subcutaneous (SQ) or intramuscular (IM) injection, see, e.g., Avis, *et al.* (eds.) (1993) *Pharmaceutical Dosage Forms: Parenteral Medications 2d ed.*, Dekker, NY; Lieberman, *et al.* (eds. 1990) *Pharmaceutical Dosage Forms: Tablets 2d ed.*, Dekker, NY; Lieberman, *et al.* (eds. 1990) *Pharmaceutical Dosage Forms: Disperse Systems* Dekker, NY; Fodor, *et al.* (1991) *Science* 251:767-773, Coligan (ed.) *Current Protocols in Immunology*; Hood, *et al.* *Immunology Benjamin/Cummings*; Paul (ed.) *Fundamental Immunology*; Academic Press; Parce, *et al.* (1989) *Science* 246:243-247; Owicki, *et al.* (1990) *Proc. Natl. Acad. Sci. USA* 87:4007-4011; and Blundell and Johnson (1976) *Protein Crystallography*, Academic Press, New York.

Selecting an administration regimen for a therapeutic depends on several factors, including the serum or tissue turnover rate of the entity, the level of symptoms, the immunogenicity of the

entity, and the accessibility of the target cells, timing of administration, absorption through epithelial layers, etc. Preferably, an administration regimen maximizes the amount of therapeutic delivered to the patient consistent with an acceptable level of side effects. Accordingly, the amount of biologic delivered depends in part on the particular entity and the severity of the condition being treated.

5 Guidance in selecting appropriate doses of cytokine or small molecules are determined using standard methodologies.

Determination of the appropriate dose is made by the clinician, e.g., using parameters or factors known or suspected in the art to affect treatment or predicted to affect treatment. Generally, the dose begins with an amount somewhat less than the optimum dose and it is increased by small  
10 increments thereafter until the desired or optimum effect is achieved relative to any negative side effects. Important diagnostic measures include those of symptoms of, e.g., the inflammation or level of inflammatory cytokines produced. Preferably, a biologic that will be used is derived from the same species as the animal targeted for treatment, thereby minimizing a humoral response to the reagent.

15 Antibodies, antibody fragments, and cytokines can be provided by continuous infusion, or by doses at intervals of, e.g., one day, one week, or 1-7 times per week. Doses may be provided intravenously, subcutaneously, topically, orally, nasally, rectally, intramuscular, intracerebrally, or by inhalation. A preferred dose protocol is one involving the maximal dose or dose frequency that avoids significant undesirable side effects. A total weekly dose is generally at least 0.05  $\mu\text{g}/\text{kg}$  body  
20 weight, more generally at least 0.2  $\mu\text{g}/\text{kg}$ , most generally at least 0.5  $\mu\text{g}/\text{kg}$ , typically at least 1  $\mu\text{g}/\text{kg}$ , more typically at least 10  $\mu\text{g}/\text{kg}$ , most typically at least 100  $\mu\text{g}/\text{kg}$ , preferably at least 0.2 mg/kg, more preferably at least 1.0 mg/kg, most preferably at least 2.0 mg/kg, optimally at least 10 mg/kg, more optimally at least 25 mg/kg, and most optimally at least 50 mg/kg, see, e.g., Yang, *et al.* (2003) *New Engl. J. Med.* 349:427-434; Herold, *et al.* (2002) *New Engl. J. Med.* 346:1692-1698; Liu, *et al.* (1999)  
25 *J. Neurol. Neurosurg. Psych.* 67:451-456; Portielji, *et al.* (2003) *Cancer Immunol. Immunother.* 52:133-144. The desired dose of a small molecule therapeutic, e.g., a peptide mimetic, natural product, or organic chemical, is about the same as for an antibody or polypeptide, on a moles/kg basis.

The present invention also provides for administration of biologics in combination with  
30 known therapies, e.g., steroids, particularly glucocorticoids, which alleviate the symptoms, e.g., associated with inflammation, or antibiotics or anti-infectives. Daily dosages for glucocorticoids will range from at least about 1 mg, generally at least about 2 mg, and preferably at least about 5 mg per day. Generally, the dosage will be less than about 100 mg, typically less than about 50 mg, preferably less than about 20 mg, and more preferably at least about 10 mg per day. In general, the  
35 ranges will be from at least about 1 mg to about 100 mg, preferably from about 2 mg to 50 mg per

day. Suitable dose combinations with antibiotics, anti-infectives, or anti-inflammatories are also known.

The present invention provides agonists and antagonists of IL-23 for modulating genes relating to healing, e.g., wound healing. Also provided are methods of diagnosis of healing, e.g.,  
5 involving detecting expression or changes in expression of IL-23 modulated genes and gene products. These genes and gene products include, e.g., nitric oxide synthase 2 (NOS2), lactoferrin, IL-19, DEC-205, CD50 (ICAM-2), IL-25, TNFSF7 (CD27L), eosinophilic basic protein, and others.

The invention provides a method to modulate expression of MMP-7, e.g., for the treatment of wound healing. Matrix proteolysis is a hallmark of inflammation. Matrilysin, a metalloprotease, is  
10 used in wound repair (see, e.g., Parks, *et al.* (2001) *Chest* 120:36S-41S; Wilson, *et al.* (1999) *Science* 286:113-117).

The present invention provides methods for modulating activities and proteins relating to neutrophils, such as neutrophil chemoattractants and proteins and metabolites expressed by neutrophils. IL-23 stimulates IL-17 expression which, in turn, stimulates production of chemokines  
15 that attract neutrophils. Increased expression or activity of neutrophil response, lactoferrin, IL-17, IL-6, and nitric oxide, are found in a number of inflammatory conditions, and can play a role in modulating wound healing (see, e.g., Tsokos, *et al.* (2002) *Virchows Arch.* 441:494-499; Linden (2001) *Int. Arch. Allergy Immunol.* 126:179-184; Sheppard (2002) *Chest* 121:21S-25S; Redington (2000) *Monaldi Arch. Chest Dis.* 55:317-323; Vignola, *et al.* (2001) *Curr. Allergy Asthma Rep.*  
20 1:108-115).

Provided are methods of modulating expression of lactoferrin, a protein produced by neutrophils (see, e.g., Boyton, *et al.* (2002) *Brit. Medical Bull.* 61:1-12; Singh, *et al.* (2002) *Nature* 417:552-555; Gomez, *et al.* (2002) *Infect. Immun.* 70:7050-7053).

Provided are methods for modulating expression of neutrophil elastase, e.g., for modulating  
25 wound healing (see, e.g., Tkalcevic, *et al.* (2000) *Immunity* 12:201-210; Aprikyan, *et al.* (2001) *Curr. Opinion Immunol.* 13:535-538; Tremblay, *et al.* (2003) *Curr. Opin. Investig. Drugs* 4:556-565; Lee, *et al.* (2001) *Curr. Opinion Crit. Care* 7:1-7; Shapiro (2002) *Am. J. Respir. Cell Mol. Biol.* 26:266-268).

Also provided are methods to modulate neutrophil attractants for promoting wound healing,  
30 e.g., IL-17, nitric oxide, and GRO-alpha. IL-17 modulates neutrophil recruitment (see, e.g., Ye, *et al.* (2001) *J. Exp. Med.* 194:519-527; Ye, *et al.* (2001) *Am. J. Respir. Cell Mol. Biol.* 25:335-340). Nitric oxide, synthesized by nitric oxide synthase, can promote wound healing, e.g., by attracting monocytes and neutrophils to the wound, see, e.g., Schwentker, *et al.* (2002) *Nitric Oxide* 7:1-10; MacMicking, *et al.* (1997) *Annu. Rev. Immunol.* 15:323-350. CXCL-1 (a.k.a. GRO-alpha) promotes  
35 wound healing, e.g., by attracting neutrophils to wounds and stimulating keratinocyte proliferation

and angiogenesis (see, e.g., Gillitzer, *et al.* (2001) *J. Leukoc. Biol.* 69:513-521; Li and Thornhill (2000) *Cytokine* 12:1409-1413).

IL-6 promotes the healing of injuries, e.g., skin wounds, see, e.g., Gallucci, *et al.* (2001) *J. Interferon Cytokine Res.* 21:603-609; Sugawara, *et al.* (2001) *Cytokine* 15:328-336; Erdag, *et al.* (2002) *Ann. Surg.* 235:113-124; Nadeau, *et al.* (2002) *Microbes Infect.* 4:1379-1387; Imanishi, *et al.* (2000) *Prog. Retin. Eye Res.* 19:113-129; Gregory, *et al.* (1998) *J. Immunol.* 160:6056-6061.

Interferon-gamma (IFN $\gamma$ ) mediates proper wound healing, e.g., by modulating actin and collagen content, contractile capacity, and scar formation, see, e.g., Moulin, *et al.* (1998) *Exp. Cell Res.* 238:283-293; Ahdieh, *et al.* (2001) *Am. J. Physiol. Cell Physiol.* 281:C2029-C2038; Cornelissen, *et al.* (2000) *J. Dent. Res.* 79:1782-1788; Shtrichman, *et al.* (2001) *Curr. Opin. Microbiol.* 4:251-259; Ikeda, *et al.* (2002) *Cytokine Growth Factor Rev.* 13:95-109; Rottenberg, *et al.* (2002) *Curr. Opin. Immunol.* 14:444-451).

IFN $\gamma$  production is stimulated by CD27 (a.k.a. TNFRSF7). CD27 also stimulates cell proliferation and is implicated in the activation and development of T cells, and in T cell-dependent antibody production, including IgE production, by B cells (see, e.g., Takeda, *et al.* (2000) *J. Immunol.* 164:1741-1745; Nagumo, *et al.* (1998) *J. Immunol.* 161:6496-6502; Tomiyama, *et al.* (2002) *J. Immunol.* 168:5538-5550; Busse and Lemanske (2001) *New Engl. J. Med.* 344:350-362).

MUC5ac serves a number of biological functions, including wound healing, see, e.g., Dohrman, *et al.* (1998) *Biochim. Biophys. Acta* 1406:251-259; Rose, *et al.* (2000) *J. Aerosol. Med.* 13:245-261; Rogers (2000) *Monaldi Arch. Chest Dis.* 55:324-332; Enss, *et al.* (2000) *Inflamm. Res.* 49:162-169.

The broad scope of this invention is best understood with reference to the following examples, which are not intended to limit the inventions to the specific embodiments.

## EXAMPLES

## I. General Methods.

Standard methods of biochemistry and molecular biology are described or referenced, e.g., in  
 5 Maniatis, *et al.* (1982) *Molecular Cloning, A Laboratory Manual*, Cold Spring Harbor Press, Cold  
 Spring Harbor, NY; Sambrook and Russell (2001) *Molecular Cloning, 3<sup>rd</sup> ed.*, Cold Spring Harbor  
 Laboratory Press, Cold Spring Harbor, NY; Wu (1993) *Recombinant DNA, Vol. 217*, Academic  
 Press, San Diego, CA; Innis, *et al.* (eds.) (1990) *PCR Protocols: A Guide to Methods and*  
*Applications*, Academic Press, N.Y. Standard methods are also found in Ausbel, *et al.* (2001)  
 10 *Current Protocols in Molecular Biology, Vols. 1-4*, John Wiley and Sons, Inc. New York, NY, which  
 describes cloning in bacterial cells and DNA mutagenesis (Vol. 1), cloning in mammalian cells and  
 yeast (Vol. 2), glycoconjugates and protein expression (Vol. 3), and bioinformatics (Vol. 4).  
 Methods for producing fusion proteins are described. See, e.g., Invitrogen (2002) *Catalogue*,  
 Carlsbad, CA; Amersham Pharmacia Biotech (2002), *Catalogue*, Piscataway, NJ; Liu, *et al.* (2001)  
 15 *Curr. Protein Pept. Sci.* 2:107-121; Graddis, *et al.* (2002) *Curr. Pharm. Biotechnol.* 3:285-297.  
 Standard methods of histology are described (Carson (1997) *Histotechnology: A Self-Instructional*  
*Text, 2<sup>nd</sup> ed.*, Am. Soc. Clin. Pathol. Press, Chicago, IL; Bancroft and Gamble (eds.) (2002) *Theory*  
*and Practice of Histological Techniques, 5<sup>th</sup> ed.*, W.B. Saunders Co., Phila., PA).

Methods for flow cytometry, including fluorescence activated cell sorting (FACS), are  
 20 available, see, e.g., Owens, *et al.* (1994) *Flow Cytometry Principles for Clinical Laboratory Practice*,  
 John Wiley and Sons, Hoboken, NJ; Givan (2001) *Flow Cytometry, 2<sup>nd</sup> ed.*; Wiley-Liss, Hoboken,  
 NJ; Shapiro (2003) *Practical Flow Cytometry*, John Wiley and Sons, Hoboken, NJ.

Standard methods of histology of the immune system are described, see, e.g., Muller-  
 Harmelink (ed.) (1986) *Human Thymus: Histopathology and Pathology*, Springer Verlag, New York,  
 25 NY; Hiatt, *et al.* (2000) *Color Atlas of Histology*, Lippincott, Williams, and Wilkins, Phila, PA;  
 Louis, *et al.* (2002) *Basic Histology: Text and Atlas*, McGraw-Hill, New York, NY.

Methods for antibody production and modification are described in, e.g., Coligan, *et al.*  
 (2001) *Current Protocols in Immunology, Vol. 1*, John Wiley and Sons, Inc., New York; Harlow and  
 Lane (1999) *Using Antibodies, Cold Spring Harbor Laboratory Press*, Cold Spring Harbor, NY);  
 30 Harlow and Lane (1988) *Antibodies A Laboratory Manual*, Cold Spring Harbor Laboratory Press,  
 Cold Spring Harbor, NY; Einhauer, *et al.* (2001) *J. Biochem. Biophys. Methods* 49:455-465.  
 Methods for adenovirus engineering and transfection, e.g., into cells or mammals, are described  
 (Hurst, *et al.* (2002) *New Engl. J. Med.* 169:443-453; Danthinne and Imperiale (2000) *Gene Ther.*  
 7:1707-1714; Carlisle (2002) *Curr. Op. Mol. Ther.* 4:306-312.

35 Methods for protein purification such as immunoprecipitation, column chromatography,  
 electrophoresis, isoelectric focusing, centrifugation, and crystallization, are described (Coligan, *et al.*

(2000) *Current Protocols in Protein Science, Vol. 1*, John Wiley and Sons, Inc., New York). Chemical analysis, chemical modification, post-translational modification, and glycosylation of proteins is described. See, e.g., Coligan, *et al.* (2000) *Current Protocols in Protein Science, Vol. 2*, John Wiley and Sons, Inc., New York; Walker (ed.) (2002) *Protein Protocols Handbook*, Humana Press, Towota, NJ; Lundblad (1995) *Techniques in Protein Modification*, CRC Press, Boca Raton, FL. Techniques for characterizing binding interactions are described (Coligan, *et al.* (2001) *Current Protocols in Immunology, Vol. 4*, John Wiley and Sons, Inc., New York; Parker, *et al.* (2000) *J. Biomol. Screen.* 5: 77-88; Karlsson, *et al.* (1991) *J. Immunol. Methods* 145:229-240; Neri, *et al.* (1997) *Nat. Biotechnol.* 15:1271-1275; Jonsson, *et al.* (1991) *Biotechniques* 11:620-627; Friguet, *et al.* (1985) *J. Immunol. Methods* 77: 305-319; Hubble (1997) *Immunol. Today* 18:305-306; Shen, *et al.* (2001) *J. Biol. Chem.* 276:47311-47319).

Computer analysis is performed using software for determining, e.g., antigenic fragments, signal and leader sequences, protein folding, and functional domains, are available, see, e.g., Vector NTI® Suite (Informax, Inc., Bethesda, MD); GCG Wisconsin Package (Accelrys, Inc., San Diego, CA), and DeCypher® (TimeLogic Corp., Crystal Bay, Nevada); Menne, *et al.* (2000) *Bioinformatics* 16:741-742. Public sequence databases were also used, e.g., from GenBank and others.

## II. Wound Generation in p19 Deficient Mice.

Mice deficient in the p19 subunit of IL-23 (p19 knockout mice; p19KO mice) were injected with 1 mg of heat killed *Mycobacterium tuberculosis* (strain H37 RA) emulsified in incomplete Freund's adjuvant (IFA) over four dorsal-flank sites. After 14-18 days post exposure to bacterial antigen, the p19KO mice showed significant hair loss and skin lesions. These results suggest that in the absence of IL-23, mice have an abnormal immune response to bacterial antigen challenge leading to an impaired wound healing response.

## III. Incision Wound Generation and Assessment of Histological Responses.

Incision wounding (5 mm) was performed on the dorsal skin of wild-type (WT) and p19KO mice as described (see, e.g., Cohen and Mast (1990) *Advances in Understanding Trauma and Burn Injury* 30:S149-S155). Eighteen hours after linear incision wounding samples from skin surrounding the incision were harvested and subjected to confocal microscopy. Tissue sections were stained with anti IA-FITC and anti-CD11b-APC. p19KO mice showed delayed recruitment of CD11b+, MHC-class II positive cells, i.e., a delay in the recruitment of bone marrow derived monocytes during the wound healing process.

IL-23 was found to induce recruitment of activated monocytes/macrophages. C57BL/6 mice were injected with 10 micrograms of recombinant IL-23 (rIL-23) at a dorsal intradermal site. Skin samples were stained with anti-IA-FITC and anti-CD11b-APC and analyzed by confocal microscopy.

IL-23 induced recruitment of MHC-class II<sup>+</sup>, CD11b<sup>+</sup> monocytes to the site of cytokine injection. PBS treated skin samples appeared negative for recruitment of these monocytes. Taken together, the altered healing response of p19KO mice to injected *Mycobacterium tuberculosis* and the IL-23 recruitment of activated monocytes/macrophages suggest a role of IL-23 in the wound-healing response.

IL-23 also induced monocyte/macrophage recruitment in wound bed granulation tissue. Ten micrograms of rIL-23 was delivered intradermally to eight sites surrounding two 6 mm full thickness dorsal excisional wounds immediately post-wounding. On day 3, the two wounds and surrounding skin were excised, frozen in OCT, sectioned and stained with anti-CD11b. Counterstaining was done with hematoxylin (blue). IL-23 treated wound tissue exhibited higher monocyte/macrophage infiltrate than buffer control 3 days post-wounding. Similar results were found for CD31<sup>+</sup> endothelial cell migration.

Recombinant IL-23 was found to augment wound healing. Ten micrograms of rIL-23 was delivered intradermally to eight sites surrounding 6 mm full thickness excisional wounds on the backs of healing competent wild-type Balb/c mice immediately post wounding. On day 3 post wounding the two excisional wounds plus additional surrounding skin were excised. Wounds were fixed in formaldehyde, paraffin embedded and stained with hematoxylin and eosin. Histology slides were assessed for the depth of granulation tissue. IL-23 treated mice exhibited a greater thickness of the granulation tissue layer as compared to buffer control treated mice. Increase in the granulation tissue layer is imperative for late stage wound healing (see, e.g., Schaffer and Nanney, *supra*). Similar results were at 3, 6, and 10 days post wounding.

The same procedure was utilized to determine if IL-23 augments the normal re-epithelialization that occurs in wound healing. IL-23 treated mice exhibited thicker keratinocyte layers than buffer control mice.

#### IV. Incision Wound Healing and Assessment of Breaking Strength.

Twelve week old male C57Bl/6NT mice were shaved and hair was removed from the dorsum using a depilatory (Nair®, Church and Dwight Co., Princeton, NJ). Two 0.5 cm incisional wounds were created approximately 4 cm down from the nape of the neck and 1 cm away from the midline using surgical scissors. Each mouse received four 20 microliter intradermal injections with either saline or mIL-23 (10 micrograms per mouse) around the periphery of each wound to span the entire length of the incision on both sides. Each wound was closed using a medical adhesive (Mastisol®, Ferndale Labs., Ferndale, MI) and transparent dressing (OpSite 3000®, Smith and Nephew, Largo, FL). Three days later, mice were anesthetized with a ketamine/xylazine cocktail and wounds were analyzed *in vivo* using a biomechanical tissue characterizer (BTC-2000, SRLI Technologies, Nashville, TN), according to manufacturer's instructions. The BTC-2000 uses a high resolution

target laser and vacuum to measure skin deformation over a range of negative pressure and time. Negative pressure was applied at a rate of 10 mm Hg/second until wound rupture occurred.

Data were analyzed by plotting skin deformation as a function of negative pressure to give: 1) the stiffness of the healing wound (mmHg negative pressure required to deform the tissue) and 2) the total strength of the healing wound (i.e. mmHg negative pressure required to break the wound). Tests comparing saline-treated controls with mIL-23-treated experimentals demonstrated that 46% more negative pressure was required to break mIL-23-treated wounds (195.6 mm Hg) than to break to saline-treated wounds (134.1 mm Hg). Moreover, the healing wound following mIL-23 treatment (262.7 mm Hg/mm) was stiffer in comparison to saline-treated controls (198.4 mm Hg/mm).

Administration of an IL-23 agonist is contemplated to result in a desirable increase in stiffness (or reduction in looseness) of the healed wound, beyond which, an undesirable increase in stiffness (or undesirable reduction of looseness) occurs. Hence, an IL-23 antagonist is provided for reducing any undesirable stiffness (or undesired reduction of looseness) of a naturally healing wound or an IL-23 agonist treated wound. Alternatively, for example, an IL-23 agonist is contemplated to result in a decrease in undesirable stiffness, e.g., due to excess fibrosis. Hence, an IL-23 agonist is provided to reduce any undesirable stiffness.

#### V. IL-23 Treatment Increases Granulation Tissue.

The wound healing response in the mouse back excisional model is mediated by the combination of increased granulation tissue formation, re-epithelialization, and wound contraction. One phase of wound contraction in this model occurs very early and contributes significantly to the overall wound closure. To confirm that IL-23 wound healing promotion activities were also seen in a mouse model that did not have a prominent early wound contraction aspect to the healing response, the mouse head excisional model was chosen.

10 micrograms of recombinant IL-23 was delivered intradermally to 4 sites surrounding one 3 mm full thickness excisional wound on the crown of Balb/c mice's heads immediately post-wounding. On Day 3 post-wounding, the wound and surrounding skin were excised. Wounds were fixed in formaldehyde, paraffin embedded, and stained with hematoxylin and eosin (H/E). Histology slides from the center of the wound were assessed for the depth of granulation tissue.

The data at day 3 post wound infliction indicates that the increased granulation tissue activity resulting from IL-23 treatment was seen prior to the start of later-stage contraction and therefore is independent of early phase wound contraction. Note that a later-wound contraction phase also occurs in this model, but this occurs only after day 4-6.

#### VI. Gene Therapy.

IL-23 is delivered to the site of a cutaneous wound using gene therapy technology. Therapeutic delivery to the skin is accomplished by *ex vivo* or *in vivo* methods (see, e.g., Khavari (1997) *Mol. Med. Today* 3:533-538 and Khavari, *et al.* (2002) *J. Int. Med.* 252:1-10).

Recombinant adenoviral preparation were prepared by conventional techniques, see, e.g.,  
5 Srivastava in WO 93/09239, Samulski *et al.* (1989) *J Virol.* 63:3822-3828; Mendelson *et al.* (1988) *Virol.* 166:154-165; and Flotte *et al.* (1993) *Proc. Natl. Acad. Sci. USA* 90:10613-10617). Increasing number of adenoviral particles encoding mouse IL-23 hyperkine or green fluorescent protein (GFP) or saline diluent were delivered intradermally to 8 sites surrounding two 6 mm full thickness excisional wounds on the backs of Balb/c mice immediately post-wounding. On Day 3 post-  
10 wounding, the two wounds and surrounding skin were excised. Wounds were fixed in formaldehyde, paraffin embedded, and stained with hematoxylin and eosin (H/E). Histology slides from the center of the wound were assessed for the depth of granulation tissue. Delivery of mIL-23 hyperkine by this method resulted in an increased wound healing response as compared to vehicle alone.

#### 15 VII. Gene Expression with mIL-23 Hyperkine Treatment.

Excisional wounds were created on the backs of C57BI6/NT mice. Wounds were treated with 10 micrograms of mIL-23 hyperkine or saline control by intradermal (i.d.) injection around the periphery of the wound. Wounds were harvested at Day 1 and at Day 3 post-wounding. Skin samples distal to the wounded site were obtained as "non-wounded" controls. Taqman® real time  
20 PCR analysis (Applied Biosystems, Foster City, CA) was performed on the tissue samples. Expression data were relative to ubiquitin expression, where ubiquitin expression was set to one (1.0). Expression data were then compared according to the indicated pairs of data sets.

Expression was examined with and without IL-23 hyperkine treatment, in the absence of wounding (Table 1) and with and without IL-23 hyperkine with wounding (Table 2).. C57BI6/NT  
25 mice were treated with IL-23 hyperkine or saline, followed by determination of expression of the indicated gene by Taqman® real time PCR analysis (Table 1). Each mouse was injected intradermally, in the back, with either saline or with 10 micrograms IL-23 hyperkine. Tissue samples were taken and extracted at either 1, 3, or 7 days after injection, where the samples from the three dates were pooled, and then used for Taqman® analysis. The ratio of gene expression with and  
30 without IL-23 hyperkine treatment is shown (Table 1). IL-23 hyperkine provoked an increase in expression, of 2-fold or greater, of 15 of the 157 genes tested. These included IL-6 (33-fold), IL-19 (32-fold), and CXCL-1 (GRO-alpha) (11-fold) (Table 1).

Table 1. Ratio of [Gene expression with IL-23] / [Gene expression with saline]. Control (saline) and experimental (IL-23) data were acquired without wounding.

IL-6	33
IL-19	32
CXCL-1 (GRO-alpha)	11
IL-17	9
mMUC-5ac.fcgi	8
secretory leukoprotease inhibitor (SLPI)	5
granulocyte macrophage-colony stimulating factor (GM-CSF)	5
TNFSF5 (CD40L)	3
MAdCAM-1	3
interferon-gamma (IFN-gamma)	3
IL-9	3
12-lipoxygenase	2
tissue inhibitor of metalloproteinases-1 (TIMP-1)	2
IL-1alpha	2
IL-17RC	2

Expression data from Day 1 (1 day after wounding) and from Day 3 (3 days after wounding) are shown (Table 2). IL-23 with wounding stimulated increases in expression over that found with wounding only for a number of genes associated with healing or neutrophil response, including IL-17, nitric oxide synthase, lactoferrin, and matrix metalloproteinases (Table 2).

Table 2. Ratio of [Gene expression with IL-23, with wound] / [Gene expression with saline, with wound]

	Day 1	Day 3
IL-17	234	6.7
nitric oxide synthase 2 (NOS2)	17	--
lactoferrin	5.3	8.9
IL-12 (p35 subunit)	3.9	--
IL-12 (p40 subunit)	3.6	--
CD107 (LAMP-2; mac-3)	3.5	--
integrin beta7 chain	3.4	--
TNFSF11 (RANKL)	3.3	--
CD11a (LFA-1a chain)	3.0	--
IL-19	2.9	2.0
endoperoxidase synthase type II (COX-2)	2.8	--

IL-1RA	2.8	--
IL-17RA	2.7	--
CD29 (integrin b1)	2.6	--
leukotriene A4 hydrolase	2.5	--
MMP-13	2.4	--
MMP-8 (collagenase 2)	2.4	--
leukotriene B4 receptor	2.4	--
TNFRSF11a (RANK)	2.4	--
M-CSF1	2.3	--
TIMP-1	2.3	--
CD14	2.3	--
TNF-alpha converting enzyme (TACE)	2.3	--
Mac-1 alpha subunit (CD11b)	2.3	--
DEC-205	2.2	--
CD50 (ICAM-1)	2.2	--
CCL3 MIP-1alpha	2.2	--
CD86 (B7-2)	2.2	--
CD80 (B7-1)	2.1	--
IL-80	2.1	--
fibronectin	2.0	--
TNFRSF6a (FAS)	2.0	--
neutrophil elastase	0.1	13.7
interferon-gamma (IFN-gamma)	0.7	8.7
matrilysin (MMP-7)	0.1	5.3
TNFRSF7u (CD27)	0.8	4.9
IL-13Ra2	1.4	4.8
IL-12Rb1	1.6	3.8
myeloperoxidase	1.1	2.7
eosinophil major basic protein	0.4	2.7
proteinase 3	1.6	2.6
IL-15	1.4	2.5

IL-23 dependent trends in gene expression in common with two strains of mice (C57B1/6NT mice; Balb/c mice) were as follows. A panel of 158 genes was screened to determine which are regulated by IL-23 treatment during wound healing. Only five of these genes were upregulated by at

least 2-fold (Table 3), and only three of the genes were down regulated by 2-fold or greater (Table 3).

IL-17 has been shown to regulate genes important for remodeling in other tissue systems. IL-17 potentiates both MMP-3 and TIMP-1 expression and secretion stimulated by TNF and IL-1 in human colonic subepithelial myofibroblasts (Bamba, *et al.* (2003) *J Gastroenterol.* 38:548-554). In human osteoblasts, IL-17 synergizes with TNF-alpha, TGF-beta, and IFN-gamma to stimulate production of MMP-13 (Rifas and Arackal (2003) *Arthritis Rheum.* 48:993-1001). Given that IL-17 alone has minimal effects in these models, the major role of IL-17 may be to amplify the remodeling response. IL-19 is found in skin and has been associated with psoriasis (Ghoreschik, *et al.* (2003) *Nature Med.* 9:40-46; Gallagher, *et al.* (2000) *Genes Immunity* 1:442-450). Lactoferrin is an iron-binding protein present in mature neutrophil granules with a number of biological functions including anti-microbial properties and may thus protect against bacterial infection in the wound (Masson, *et al.* (1969) *J. Exp. Med.* 130:643-658; Farnaud and Evans (2003) *Mol Immunol.* 40:395-405). ICAM-2 is constitutively expressed on all vascular endothelial cells suggesting that IL-23 may promote angiogenesis in the wound (Yasuda, *et al.* (2002) *Am. J. Physiol.* 282:C917-C925; Sakurai, *et al.* (2003) *Invest. Ophthalmol. Vis. Sci.* 44:2743-2749; Moromizato, *et al.* (2000) *Am. J. Pathol.* 157:1277-1281). DEC-205 is a dendritic cell marker implicated in the dendritic cell maturation (Anjuere, *et al.* (1999) *Blood* 93:590-598; Bonifax, *et al.* (2002) *J. Exp. Med.* 196:1627-1638). Thus IL-23 of the present invention can promote immune surveillance in the skin.

Taken together, IL-23 treatment in wounds may have pleiotropic effects to accelerate the wound healing response. IL-23 treatment also reduced the expression of 3 genes in the wound environment, i.e., IL-25, TNSF7, and eosinophilic basic protein. Infusion of mice with IL-25 induced IL-4, IL-5, and IL-13 gene expression associated with lung remodeling pathology (Fort, *et al.* (2001) *Immunity* 15:985-995). Thus, IL-23 may modulate or reduce excess fibrosis in wound healing.

Relevant methodology was as follows. Hair was shaved and removed with Nair from the dorsum of 9 week old male C57Bl/6NT and 8 week old male Balb/c mice. Two 6 mm diameter excisional wounds were created approximately 3.5 cm down from the nape of the neck and 0.6 cm away from the midline using a punch biopsy tool. Each mouse was given mIL-23 hyperkine (10 micrograms per mouse) or saline in four 20 microliters intradermal injections around the periphery of each wound. A transparent dressing was applied and mice were allowed to recover and heal. One day later, mice were sacrificed and wounds were excised with a margin of approximately 1.5 mm. In addition, a sample of non-wounded skin from each mouse was obtained. All samples were snap frozen in liquid nitrogen and stored at -80 degrees Celsius for further processing. RNA was extracted from each sample, pooled together into appropriate groups, and analyzed by real time PCR for expression of a panel of 158 candidate genes.

Table 3. Modulation of gene expression, by at least 2-fold, in response to IL-23 in both C57B1/6NT mice and Balb/c mice.

Genes up-regulated.	C57B1/6NT mouse	Balb/c mouse
IL-17	234.5	207.9
Lactoferrin	5.3	3.3
IL-19	2.9	2.2
DEC-205	2.3	2.8
CD50 (ICAM-2)	2.3	2.1
Genes down-regulated.	C57B1/6NT mouse	Balb/c mouse
IL-25	5.88	5.56
TNSF7-CD27L	3.45	100
Eosinophilic basic protein	2.13	2.33

#### VIII. Agonist Anti-IL-23R Antibodies.

Mouse anti-IL-23R antibodies were prepared against human IL-23R using standard methods.

5 The resultant anti-IL-23R antibodies were screened for agonist activity using cells transfected with hIL-23R and hIL-12Rbeta1, where agonist activity was determined by increases in cell proliferation. Proliferation of transfectant Ba/F3 cells was measured by colorimetric methods using Alamar Blue, a growth indicator dye.

10 Cell proliferation was measured after culture in Roswell Park Memorial Institute (RPMI)-1640 medium, fetal calf serum (10%), 0.05 mM 2-mercaptoethanol, glutamine, penicillin, streptomycin, and mouse interleukin-3 (mIL-3) (10 ng/ml). Cells were incubated in the presence of one concentration of antibody, where the concentrations ranged from 0.01 to 10,000 ng/ml. Baseline cell proliferation was that in absence of antibody. Maximal proliferation with the antibodies tested occurred in the concentration range of 1000 to 10,000 ng/ml. "Detectable stimulation," e.g., of cell activity, cell proliferation, or a predetermined activity, refers, e.g., to a comparison of proliferation in the presence and absence of, e.g., an IL-23 agonist. "Detectable" may be a function of the context, e.g., of the reagents, instrumentation, or biological system.

15 Antibodies that stimulated expression included TC48-8B10.D5 (light chain, SEQ ID NOs:9 and 10; heavy chain, SEQ ID NOs:11 and 12), TC48-1H3.G5, TC48-2C9A5, and TC48-5B12.C9. 20 Of these antibodies, TC48-8B10.D5 showed the greatest agonist activity, in terms of antibody concentration provoking ½ maximal increase in cell proliferation. 100% cell proliferation means maximal increase in cell proliferation in response to the antibody, in titration curves (Table 4). The predicted cleavage point for the signal sequence of the light chain of TC48-8B10.D5 is between

amino acids 22 (Ala) and 23 (Glu) of SEQ ID NO:10, while the predicted cleavage point for the heavy chain is between amino acids 19 (Ser) and 20 (Gln) of SEQ ID NO:12.

Table 4. Proliferation of Ba/F3-2.2lo cells in response to agonistic antibodies.

Antibody	Maximal increase cell proliferation in response to antibody (units of OD <sub>570-600 nm</sub> )	Antibody concentration providing ½ maximal increase in cell proliferation.
TC48-8B10.D5 (SEQ ID NOs: 9-12)	0.026	10 ng/ml
TC48-2C9A5	0.021	20 ng/ml
TC48-1H3.G5	0.018	50 ng/ml
TC48-5B12.C9	0.026	100 ng/ml

5 The invention provides an agonistic murine antibody that specifically binds human IL-23R, TC48-8B10.D5. This antibody comprises a nucleic acid (SEQ ID NO:9) and polypeptide (SEQ ID NO:10) of the light chain, and the nucleic acid (SEQ ID NO:11) and polypeptide (SEQ ID NO:12) of the heavy chain of TC48-8B10.D5. Also provided are the nucleic acids and polypeptides comprising the hypervariable regions of SEQ ID NOs:9-12.

10 The hypervariable regions of the light chain are: ITSTDIDDDMI (amino acids 46-56); EGNTLRP (amino acids 72-78); and LQSDNMPLT (amino acids 111-119), of SEQ ID NO:10). The hypervariable regions of the heavy chain are: GYTFTSYWMN (amino acids 45-54); MIDPLDSETHYNQMFKD (amino acids 69-87); and GDNYYAMDY (amino acids 118-126), of SEQ ID NO:12.

15

#### IX. Rat Anti-Mouse IL-23R Antibodies and Wound Healing.

Antibodies against mouse IL-23R were raised in rats using standard methods, resulting in antibodies named 5C10, 29A5, and 10E11 (Table 5). An IL-23 hyperkine comprising a p19 subunit and p40 subunit covalently connected by way of an elasti-linker (InvivoGen, San Diego, CA) was prepared, and named "IL-23 elastikine." The anti-mouse IL-23R antibodies, IL-23 elastikine (control), 36E10 antibody (isotype control), and saline (control) were tested in a wound healing assay. The results demonstrated a stimulation of wound healing with the IL-23 elastikine, and lesser stimulation with the 5C10 antibody (Table 5):

25

Table 5. Wound healing; thickness of granulation tissue (micrometers).

Saline control	170 micrometers
IL-23 elastikine control	300
36E10 isotype control	170
5C10	205
29A5	170
10E11	125

The sequences in the Sequence Listing are summarized (Table 6):

Table6. Sequences in Sequence Listing.

SEQ ID NO:	Nucleic acid or Polypeptide
1	Mouse IL-19 nucleic acid
2	Mouse IL-19 polypeptide
3	Human IL-19 nucleic acid
4	Human IL-19 polypeptide
5	Mouse hyperkine nucleic acid
6	Mouse hyperkine polypeptide
7	Human hyperkine nucleic acid
8	Human hyperkine polypeptide
9	Mouse anti-human agonist Ab light chain nucleic acid
10	Mouse anti-human agonist Ab light chain polypeptide
11	Mouse anti-human agonist Ab heavy chain nucleic acid
12	Mouse anti-human agonist Ab heavy chain polypeptide

All citations herein are incorporated herein by reference to the same extent as if each  
 5 individual publication or patent application was specifically and individually indicated to be  
 incorporated by reference.

Many modifications and variations of this invention can be made without departing from its  
 spirit and scope, as will be apparent to those skilled in the art. The specific embodiments described  
 herein are offered by way of example only, and the invention is to be limited by the terms of the  
 10 appended claims, along with the full scope of equivalents to which such claims are entitled; and the  
 invention is not to be limited by the specific embodiments that have been presented herein by way of  
 example.

## SEQUENCE LISTING

&lt;110&gt; Schering Corporation

&lt;120&gt; Uses of mammalian cytokine; related reagents

&lt;130&gt; DX01578K

&lt;150&gt; US 60/436274

&lt;151&gt; 2002-12-23

&lt;160&gt; 12

&lt;170&gt; PatentIn version 3.1

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Ile Thr Val Lys Glu Phe Xaa Asp Ala Gly Gln Tyr Thr Cys His Lys  
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355 360 365	
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Pro Pro Lys Leu Leu Ile Ser Glu Gly Asn Thr Leu Arg Pro Gly Val  
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Val His Ser Gln Val Gln Leu Gln Gln Pro Gly Ala Glu Leu Val Arg

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	20		25		30														
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Thr Ser Tyr Trp Met Asn Trp Val Lys Gln Arg Pro Gly Gln Gly Leu	50		55		60														
Glu Trp Ile Gly Met Ile Asp Pro Leu Asp Ser Glu Thr His Tyr Asn	65		70		75														
Gln Met Phe Lys Asp Lys Ala Thr Leu Thr Val Asp Lys Ser Ser Ser	85		90		95														
Thr Ala Tyr Met Gln Leu Ser Ser Leu Thr Ser Glu Asp Tyr Ala Val	100		105		110														
Tyr Tyr Cys Ala Arg Gly Asp Asn Tyr Tyr Ala Met Asp Tyr Trp Gly	115		120		125														
Gln Gly Thr Ser Val Thr Val Ser Ser Ala Lys Thr Thr Pro Pro Ser	130		135		140														
Val Tyr Pro Leu Ala Pro Gly Ser Ala Ala Gln Thr Asn Ser Met Val	145		150		155														
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Ala Ser Ser Thr Lys Val Asp Lys Lys Ile Val Pro Arg Asp Cys Gly	225		230		235														
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Phe Ser Trp Phe Val Asp Asp Val Glu Val His Thr Ala Gln Thr Gln  
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Pro Arg Glu Glu Gln Phe Asn Ser Thr Phe Arg Ser Val Ser Glu Leu  
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Pro Ile Met His Gln Asp Trp Leu Asn Gly Lys Glu Phe Lys Cys Arg  
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Ala Gly Asn Thr Phe Thr Cys Ser Val Leu His Glu Gly Leu His Asn  
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His His Thr Glu Lys Ser Leu Ser His Ser Pro Gly Lys Ser Gln Ser  
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Pro Val Ala  
 465

**CLAIMS**

## WHAT IS CLAIMED IS:

1. A method of treating or improving healing comprising administering to a subject an effective amount of an agonist or antagonist of IL-23. 5
2. The method of Claim 1 wherein the agonist or antagonist comprises:
  - a) a polypeptide of IL-23, or a derivative or variant thereof;
  - b) a binding composition derived from an antibody that specifically binds to IL-23 or to IL-23R; or 10
  - c) a nucleic acid encoding a polypeptide of IL-23, or a derivative or variant thereof.
3. The method of Claim 2, wherein the derivative or variant comprises an IL-23 hyperkine.
4. The method of Claim 1, wherein the agonist comprises a complex of: 15
  - a) a mature sequence of SEQ ID NO:10; and
  - b) a mature sequence of SEQ ID NO:12.
5. The method of Claim 2, wherein the nucleic acid further comprises an expression vector. 20
6. The method of Claim 1, wherein the healing is:
  - a) of a skin or cutaneous wound;
  - b) of an ulcer or graft; or
  - c) improper healing. 25
7. The method of Claim 1, wherein the treating or improving increases:
  - a) a pressure required to break a healed or healing wound;
  - b) a stiffness of a healed or healing wound;
  - c) a rate of healing of a wound;
  - d) a granulation layer thickness of a healed or healing wound; 30
  - e) recruitment of a cell to or towards a wound; or
  - f) antimicrobial activity.
8. The method of Claim 7, wherein the cell is: 35
  - a) a CD11b+, MHC Class II+ cell;
  - b) a monocyte/macrophage; or
  - c) a CD31+ endothelial cell.

9. The method of Claim 7, wherein the recruitment is in or towards a granulation tissue.
10. The method of Claim 7, wherein:
- 5 a) the increased wound breaking pressure is about a 15% or about a 20% increase in wound breaking pressure; or
- b) the increased stiffness is about a 15% or about a 20% increase in stiffness.
11. The method of Claim 1, wherein the treating or improving comprises increased:
- 10 a) angiogenesis; or
- b) immune surveillance.
12. The method of Claim 11, wherein:
- a) the increased angiogenesis is mediated by ICAM-1 or -2; or
- 15 b) the increased immune surveillance is mediated by dendritic cells.
13. The method of Claim 1, wherein the treating or improving comprises increased expression of a nucleic acid or protein of:
- a) a cytokine in addition to IL-23;
- 20 b) a signaling molecule;
- c) an anti-microbial molecule;
- d) a protease or protease inhibitor; or
- e) a molecule of the extracellular matrix.
- 25 14. The method of Claim 13, wherein the cytokine nucleic acid or protein is IL-17, IL-6, IL-19, GRO-alpha, or GM-CSF.
15. The method of Claim 13, wherein the gene or protein is:
- a) lactoferrin;
- 30 b) DEC-205;
- c) CD50;
- d) nitric oxide synthase; or
- e) secretory leukoprotease inhibitor; or
- f) CD40L.
- 35 16. The method of Claim 1, wherein the antagonist comprises:
- a) a nucleic acid;

- b) a blocking antibody to IL-23 or to IL-23R; or
- c) a soluble receptor derived from an extracellular part of IL-23R.

17. The method of Claim 16, wherein the nucleic acid comprises:

- 5
- a) an anti-sense nucleic acid; or
  - b) interference RNA.

18. An agonist of IL-23 derived from the binding site of an antibody that specifically binds to an IL-23 receptor.

10

19. The agonist of Claim 18 that is:

- a) a polyclonal antibody;
- b) a monoclonal antibody;
- c) an Fab, Fv, or F(ab')<sub>2</sub> fragment;
- 15 d) humanized;
- e) a peptide mimetic; or
- f) detectably labeled.

20. The agonist of Claim 18, comprising a complex of:

- 20
- a) a polypeptide of the mature sequence of SEQ ID NO:10; and
  - b) a polypeptide of the mature sequence of SEQ ID NO:12.

21. The agonist of Claim 18, comprising a complex of:

- a) two polypeptides of the mature sequence of SEQ ID NO:10; and
- 25 b) two polypeptides of the mature sequence of SEQ ID NO:12.

22. The agonist of Claim 18, wherein contact of the agonist to a cell expressing hIL-23R and hIL-12beta1 results in an increase in proliferation of the cell.

30 23. A kit comprising the agonist of Claim 18 and:

- a) a compartment; or
- b) instructions for use or disposal.

35