

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
7 April 2011 (07.04.2011)

(10) International Publication Number
WO 2011/039634 A2

(51) International Patent Classification:
C12N 9/00 (2006.01)

(21) International Application Number:
PCT/IB2010/002589

(22) International Filing Date:
29 September 2010 (29.09.2010)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
61/246,847 29 September 2009 (29.09.2009) US

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(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PE, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

— without international search report and to be republished upon receipt of that report (Rule 48.2(g))



WO 2011/039634 A2

(54) Title: HYDROLYSIS OF MANNOSE-1-PHOSPHO-6-MANNOSE LINKAGE TO PHOSPHO-6-MANNOSE

(57) Abstract: Described herein are methods and genetically engineered cells useful for uncapping a mannose-6-phosphate residue on an oligosaccharide.

HYDROLYSIS OF MANNOSE-1-PHOSPHO-6-MANNOSE LINKAGE TO PHOSPHO-6-MANNOSE

CROSS-REFERENCE TO RELATED APPLICATIONS

This application claims priority to U.S. Application Serial No. 61/246,847, filed on September 29, 2009. The disclosure of the prior application is incorporated by reference in its entirety.

TECHNICAL FIELD

The invention relates to methods of hydrolyzing mannose-1-phospho-6-mannose linkages on glycoproteins, and more particularly, to using a mannosidase to hydrolyze mannose-1-phospho-6-mannose linkages to uncapped the phospho-6-mannose residues on the glycoprotein.

BACKGROUND

High performance expression systems are required to produce most biopharmaceuticals (e.g., recombinant proteins) currently under development. The biological activity of many of these biopharmaceuticals is dependent on their post-translational modification (e.g., phosphorylation or glycosylation). A yeast-based expression system combines the ease of genetic manipulation and fermentation of a microbial organism with the capability to secrete and to modify proteins. However, recombinant glycoproteins produced in yeast cells exhibit mainly heterogeneous high-mannose and hyper-mannose glycan structures, which can be detrimental to protein function, downstream processing, and subsequent therapeutic use, particularly where glycosylation plays a biologically significant role.

U.S. Application Serial No. 12/062,469 is incorporated by reference in its entirety.

SUMMARY

The present invention is based, at least in part, on the discovery of a mannosidase that is capable of hydrolyzing mannose-1-phospho-6-mannose linkages on glycoproteins. As such, the mannosidase can be used to obtain glycoproteins containing uncapped

terminal mannose-6-phosphate residues. *In vitro* and *in vivo* methods of obtaining such glycoproteins are described herein. Genetically engineered cells can be used in the methods to produce target molecules having uncapped terminal mannose-6-phosphate residues.

In one aspect, this document features a method for uncapping a mannose-6-phosphate residue on an oligosaccharide. The method includes providing the oligosaccharide having a mannose-1-phospho-6-mannose linkage; and contacting the oligosaccharide with a mannosidase capable of hydrolyzing the mannose-1-phospho-6-mannose linkage to phospho-6-mannose. The contacting step can be performed using a purified mannosidase, a recombinant mannosidase, a cell lysate containing the recombinant mannosidase, or a fungal cell containing the recombinant mannosidase. The mannosidase can include a targeting sequence. The oligosaccharide can be attached to a protein (e.g., a human protein expressed in a fungal organism).

In another aspect, this document features a method of producing a target protein having terminal phospho-6-mannose residues. The method includes providing a fungal cell genetically engineered to include a nucleic acid encoding a mannosidase, the mannosidase capable of hydrolyzing a mannose-1-phospho-6-mannose linkage to phospho-6-mannose; and introducing into the cell a nucleic acid encoding a target protein, wherein the cell produces the target protein comprising the terminal phospho-6-mannose residues.

This document also features a method of producing a target protein having terminal phospho-6-mannose residues in a fungal organism. The method includes providing a fungal cell genetically engineered to include a nucleic acid encoding a mannosidase capable of hydrolyzing a mannose-1-phospho-6-mannose linkage to phospho-6-mannose, wherein the fungal cell further includes a nucleic acid encoding a target protein; and isolating the target protein having the terminal phospho-6-mannose residues. The fungal cell further can include a nucleic acid encoding a polypeptide capable of promoting mannosyl phosphorylation and/or can be genetically engineered to be deficient in OCH1 activity.

This document also features an isolated fungal cell genetically engineered to produce glycoproteins comprising terminal phospho-6-mannose residues. The fungal cell

includes a nucleic acid encoding a mannosidase, wherein expression of the mannosidase in the fungal cell produces glycoproteins comprising the terminal phospho-6-mannose residues. The fungal cell further can include a nucleic acid encoding a target glycoprotein protein.

In another aspect, this document features a substantially pure culture of *Yarrowia lipolytica*, *Pichia pastoris*, *Hansenula polymorpha*, *Arxula adenivorans*, *Pichia methanolica*, *Oogataea minuta*, or *Aspergillus niger* cells, a substantial number of which are genetically engineered to produce glycoproteins comprising a terminal phospho-6-mannose residue, the cells comprising a nucleic acid encoding a mannosidase capable of hydrolyzing a mannose-1-phospho-6-mannose linkage to phospho-6-mannose.

In any of the embodiments described herein, the fungal organism can be *Yarrowia lipolytica* or *Arxula adenivorans*. The fungal organism can be a methylotrophic yeast such as *Pichia pastoris*, *Pichia methanolica*, *Oogataea minuta*, or *Hansenula polymorpha*. The fungal organism can be a filamentous fungus (e.g., a filamentous fungus selected from the group consisting of *Aspergillus caesiellus*, *Aspergillus candidus*, *Aspergillus carneus*, *Aspergillus clavatus*, *Aspergillus deflectus*, *Aspergillus flavus*, *Aspergillus fumigatus*, *Aspergillus glaucus*, *Aspergillus nidulans*, *Aspergillus niger*, *Aspergillus ochraceus*, *Aspergillus oryzae*, *Aspergillus parasiticus*, *Aspergillus penicilloides*, *Aspergillus restrictus*, *Aspergillus sojae*, *Aspergillus sydowi*, *Aspergillus tamari*, *Aspergillus terreus*, *Aspergillus ustus*, and *Aspergillus versicolor*).

In any of embodiments described herein, the protein can be a pathogen protein, a lysosomal protein, a growth factor, a cytokine, a chemokine, an antibody or antigen-binding fragment thereof, or a fusion protein. The lysosomal protein can be a lysosomal enzyme (e.g., a lysosomal enzyme associated with a lysosomal storage disorder (LSD) such as acid alpha glucosidase or alpha galactosidase). The LSD can be Fabry's disease, mucopolysaccharidosis I, Farber disease, Gaucher disease, GM1-gangliosidosis, Tay-Sachs disease, Sandhoff disease, GM2 activator disease, Krabbe disease, metachromatic leukodystrophy, Niemann-Pick disease, Scheie disease, Hunter disease, Sanfilippo disease, Morquio disease, Maroteaux-Lamy disease, hyaluronidase deficiency, aspartylglucosaminuria, fucosidosis, mannosidosis, Schindler disease, sialidosis type 1, Pompe disease, Pycnodysostosis, ceroid lipofuscinosis, cholesterol ester storage disease,

Wolman disease, Multiple sulfatase deficiency, galactosialidosis, mucopolipidosis, cystinosis, sialic acid storage disorder, chylomicron retention disease with Marinesco-Sjögren syndrome, Hermansky-Pudlak syndrome, Chediak-Higashi syndrome, Danon disease, or Geleophysic dysplasia. For example, the LSD can be Pompe disease or Fabry's disease.

In any of the embodiments described herein, for the mannosidase, the three dimensional protein coordinates of the atoms in the amino acid side chains located in the minimal catalytic center fall within 1.5 Å deviation of the coordinates of the equivalent atoms in FIG. 33.

In any of the embodiments described herein, the mannosidase can include an amino acid sequence having at least 90% identity (e.g., at least 95% or 98% identity) to the amino acid sequence set forth in residues 1 to 774 of SEQ ID NO:50 or to the amino acid sequence set forth in SEQ ID NO:50.

In any of the embodiments described herein, the mannosidase can include an amino acid sequence having (i) a GVGXXGXGG motif, where X is Gly, Ala, Ser, Thr, or Cys; (ii) a VRXE motif, where X is any amino acid other than Pro; (iii) an X₁YQGX₂ motif, where X₁ is Leu, Ile, Val, Ala, Phe, Tyr or Met, and X₂ is Thr, Ser, or Asn; or (iv) a GDXGN motif, where X can be any amino acid other than Pro.

In any of the embodiments described herein, the mannosidase can be a *C. cellulans*, *Streptomyces coelicolor*, or *Streptomyces lividans* mannosidase.

In any of the embodiments described herein, the fungal cell further can include a nucleic acid encoding a polypeptide capable of promoting mannosyl phosphorylation (e.g., a MNN4 polypeptide such as *Yarrowia lipolytica*, *S. cerevisiae*, *Ogataea minuta*, *Pichia pastoris*, or *C. albicans* polypeptide) and/or can be genetically engineered to be deficient in OCH1 activity. For example, the polypeptide capable of promoting mannosyl phosphorylation can be a *P. pastoris* PNO1 polypeptide.

In any of the embodiments described herein, the mannosidase can include a secretion signal and/or a targeting signal to target the mannosidase to an intracellular compartment. The target protein and the mannosidase can be co-secreted.

This document also features an isolated glycoprotein that includes terminal phospho-6-mannose residues, wherein the protein is produced by the methods described herein.

In yet another aspect, this document features a composition that includes a glycoprotein, wherein at least 47% of the N-glycans on the glycoprotein have terminal phospho-6-mannose residues. For example, at least 50%, 75%, 80%, 85%, or 90% of the N-glycans on the glycoprotein can have terminal phospho-6-mannose residues.

This document also features an isolated nucleic acid that includes a nucleotide sequence set forth in SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, or SEQ ID NO:14, or a nucleotide sequence that is at least 90% identical to SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, SEQ ID NO:14, or SEQ ID NO:20. This document also features a vector that includes a promoter operably linked to such a nucleic acid, wherein the nucleic acid encodes a mannosidase. The nucleic acid further can include a secretion signal or targeting sequence to target the mannosidase to an intracellular compartment.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the exemplary methods and materials are described below. All publications, patent applications, patents, Genbank® Accession Nos, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present application, including definitions, will control. The materials, methods, and examples are illustrative only and not intended to be limiting.

Other features and advantages of the invention will be apparent from the following detailed description, and from the claims.

Brief Description of the Drawings

FIG. 1 is a schematic of the pYLTmAX and pYLTmAXMnn4 constructs.

FIG. 2 is a series of electroferograms depicting sugar analysis of MTLY60 Δ och1 (1 wild type copy of Mnn4), MTLY60 Δ och1+Hp4dMnn4 (1WT +1 extra

copy of Mnn4) and MTLY60 Δ och1+Hp4dMnn4+TEFMnn4. P represents the monophosphorylated peak, PP represents the diphosphorylated peak, and Man8 represents the Man₈GlcNAc₂ peak.

FIG. 3 is a schematic of mammalian and yeast glycan phosphorylation pathways. The mammalian glycan phosphorylation pathway involves addition of a phospho-GlcNAc catalyzed by GlcNAc-phosphotransferase to Man₈GlcNAc₂ glycans followed by decapping of the GlcNAc to expose the phosphate by an uncovering enzyme. In contrast, yeast glycan phosphorylation involves addition of a phospho-mannose to Man₈GlcNAc₂ glycans, but no endogenous enzyme is present to uncap the mannose to expose the phosphate.

FIG. 4 is a series of electroferograms depicting N-glycans derived from strain MTLY60 Δ och1+Hp4dMnn4+TEFMnn4 treated for different time frames (7 hrs, 8 hrs, or overnight (ON)) with supernatants from *C. cellulans* medium.

FIG. 5 is a series of electroferograms depicting N-glycans derived from an MNN4 overexpressing strain treated with *C. cellulans* supernatant (SN) with and without phosphatase (CIP) incubation.

FIG. 6 is a graph of the absorbance units (mAU) of elution fractions at the indicated MW. Each elution fraction contained ~500 μ l.

FIG. 7 is a representation of a SDS-polyacrylamide gel after electrophoresis of elution fractions from silica-based gel filtration (250 μ l of each fraction was DOC/TCA precipitated). The boxed bands were cut out for peptide mass fingerprinting and de novo sequencing using tandem mass spectrometry (MS/MS).

FIG. 8A is the nucleotide sequence (SEQ ID NO:6) encoding CcMan1 (i.e., mannosidase candidate 1 from *C. Cellulans*) (on contig 1003), which was identified in the MS/MS de novo sequencing. FIG. 8B is the amino acid sequence (SEQ ID NO:7) of CcMan 1, including the signal sequence (in bold). The predicted molecular weight of the CcMan 1 polypeptide without the signal sequence is 92.6 kDa.

FIG. 9A is the nucleotide sequence (SEQ ID NO:8) encoding CcMan2 (on contig 774) and FIG. 9B is the amino acid sequence of CcMan2 (SEQ ID NO:9) with signal sequence (in bold). The predicted molecular weight of the CcMan2 polypeptide without the signal sequence is 121.6 kDa.

FIG. 10A is the nucleotide sequence (SEQ ID NO:10) encoding CcMan3 (on contig 774) and FIG. 10 B is the amino acid sequence of CcMan3 (SEQ ID NO:11) with signal sequence (in bold). The predicted molecular weight of the CcMan3 polypeptide without the signal sequence is 116 kDa.

FIG. 11A is the nucleotide sequence (SEQ ID NO:12) encoding CcMan4 (on contig 1237) and FIG. 11B is the amino acid sequence of CcMan4 (SEQ ID NO:13) with signal sequence (in bold). The predicted molecular weight of the CcMan4 polypeptide without the signal sequence is 184 kDa.

FIG. 12A is the nucleotide sequence (SEQ ID NO:14) encoding CcMan5 (on contig 896). FIG. 12B is the amino acid sequence of CcMan5 with signal sequence (in bold) (SEQ ID NO:15) and FIG. 12C is the amino acid sequence of CcMan5 without signal sequence (SEQ ID NO:50). The predicted molecular weight of the CcMan5 polypeptide without the signal sequence is 173 kDa.

FIG. 13 contains examples of expression plasmids for the expression of CcMan1-5 in the periplasm of *E. coli* (pET25-Man), as secreted proteins in *Yarrowia lipolytica* (pYLPSecCcMan1-5), as proteins targeted to the secretory pathway of *Yarrowia lipolytica*, tagged to the N-terminus (pYLPNtCcMan1-5) or tagged to the C-terminus (pYLPctCcMan1-5).

FIG. 14 is the nucleotide sequence of CcMan1 that has been codon optimized for expression in *E. coli* (SEQ ID NO:16).

FIG. 15 is the nucleotide sequence of CcMan2 that has been codon optimized for expression *E. coli* (SEQ ID NO:17).

FIG. 16 is the nucleotide sequence of CcMan3 that has been codon optimized for expression in *E. coli* (SEQ ID NO:18).

FIG. 17 is the nucleotide sequence of CcMan4 that has been codon optimized for expression in *E. coli* (SEQ ID NO:19).

FIG. 18 is the nucleotide sequence of CcMan5 that has been codon optimized for expression in *E. coli* (SEQ ID NO:20).

FIG. 19 is a schematic of the pLSAH36 and pLSH36 vectors and the cloning strategy for introducing the *C. cellulans* genes into the vectors.

FIG. 20 is a series of electroferograms depicting analysis of the periplasmic fraction of CcMan4 and CcMan5 expressing *E. coli* cells. Analysis was performed using DNA sequencer-assisted, fluorophore-assisted carbohydrate electrophoresis (DSA-FACE). The first and second panels represent the dextran ladder and the sugars from RNaseB, respectively. The third panel is the untreated Mnn4 sugars with “P” corresponding to the mono mannophosphorylated Man₈GlcNAc₂ peak, “PP” corresponding to the double mannophosphorylated Man₈GlcNAc₂ peak, and “Man8” corresponding to the Man₈GlcNAc₂ peak. Panels 4 to 9 are the results obtained with Mnn4 glycans incubated with the indicated periplasm, with or without a subsequent calf intestinal phosphatase (CIP) digest.

FIG. 21 is a schematic alignment of CcMan4 (1759 AA) and CcMan5 (1650 AA) with Bt3990 (744 AA) and Bt2199 (739 AA) mannosidases described in Zhu *et al.*, *Nat. Chem. Biol.*, 6(2):125-32. Epub 2009 Dec 27 (2010).

FIG. 22 is a series of electroferograms depicting the analysis of the CcMan4 and CcMan5 enzymes obtained from expressing *E. coli* cells. Analysis was performed using DSA-FACE using MNN4 overexpressing strain derived glycans (referred to as MNN4 glycans or MNN4 sugars) as a substrate. The first panel represents the dextran ladder and the second panel represents the untreated Mnn4 sugars. In the third through sixth panels, the sugars were incubated with the CcMan4domain periplasmic fraction not induced, induced overnight at 18°C with IPTG, the CcMan5domain periplasmic fraction not induced, and induced overnight at 18°C with IPTG, respectively. The last panel represents the sugars from RNaseB.

FIG. 23 is a ribbon representation of CcMan5₁₋₇₇₄. CcMan5₁₋₇₇₄ consists of a N-terminal β-sandwich domain (residues 8-271; light gray), an α-helical linker (residues 272-290; black) and a (αα)₆ barrel domain (residues 291-771; dark gray). The catalytic Ca²⁺ is shown as a sphere.

FIG. 24 is a ribbon representation of the CcMan5₁₋₇₇₄ protein backbone with side chains lining the substrate binding site shown in stick representation. Carbon, oxygen and nitrogen atoms are colored light gray, gray, and dark gray, respectively. The Ca²⁺ ion and waters W1, W2, W3 and W4 in the catalytic center are shown as spheres.

FIG. 25 is a ribbon representation of the CcMan5₁₋₇₇₄ protein backbone with side chains lining the substrate binding site and the modeled position of mannose-1-phospho-6-mannose (labeled Man-P-Man) shown in stick representation. Carbon, oxygen and nitrogen atoms are coloured light gray, gray, and dark gray, respectively. The Ca²⁺ ion and water molecules W1, W2, W3 and W4 in the catalytic center are shown as spheres (for comparison, the positions of W2 and W3 which will be displaced by the substrate O2 and O3 hydroxyl groups are still shown). Yellow, red and black dashed lines indicate coordination bonds with Ca²⁺, H-bonds with the proposed nucleophilic water (W4) and H-bonds with the -1 site mannose and phosphate, respectively. The -1 site mannose is modeled in its ground state chair conformation. During catalysis, its O2 hydroxyl will occupy a position nearer that seen for W2, in the equatorial coordination plane of Ca²⁺ ion, thereby leading to a distortion to a half-chair conformation in the mannose -1 ring and facilitating in line attack of the nucleophilic water (W4) on the C1 carbon (arrow).

FIG. 26A is the *Y. lipolytica* codon optimized nucleotide sequence encoding α -GalactosidaseA with lip2 pre sequence in bold and the Myc His tag underlined (SEQ ID NO:22). FIG. 26B is the amino acid sequence of α -GalactosidaseA with lip2 pre sequence in bold and the Myc His tag underlined (SEQ ID NO:23).

FIG. 27A is the codon optimized nucleotide sequence of human alpha glucosidase (GAA) with lip2 pre sequence in bold (SEQ ID NO:24). FIG. 27B is the amino acid sequence of human GAA with lip2 pre sequence in bold (SEQ ID NO:25), where the * represents the stop codon.

FIG. 28 is a schematic of a *Y. lipolytica* expression vector used for cloning of huGAA.

FIG. 29 is a series of electroferograms depicting analysis of treatment of huGAA with CcMan5 derived from the periplasmic fraction of *E. coli* cells. Analysis was performed using DSA-FACE.

FIG. 30 is a depiction of the minimal catalytic center of CcMan5. The numbering of equivalent residues in SEQ ID NO:50 is given in parenthesis. 1: Q (Q536); 2: N/D-E/Q (N588-Q589); 3: D/E (D355); 4: R (R405); 5: D/E-X-D/E (D660-X-D662); 6: G-G (G71-G72); and 7: T/S/G (T626).

FIG. 31 is an alignment of the amino acid sequence of CcMan5 (SEQ ID NO:50, the amino acid sequence set forth in SEQ ID NO:15 without the signal peptide) and 10 of its homologs using MUSCLE (Multiple Sequence Comparison by Log-Expectation). NP_630514 *Streptomyces*, SEQ ID NO:26; ZP_02866543 *Clostridium*, SEQ ID NO:27; NP_812442 *Bacteroides*, SEQ ID NO:28; YP_003584502 *Zunongwangia*, SEQ ID NO:29; YP_003120664 *Chitinophaga*, SEQ ID NO:30; AAK22560 *Caulobacter*, SEQ ID NO:31; ACL94075 *Caulobacter*, SEQ ID NO:32; ACT03290 *Paenibacillus*, SEQ ID NO:33; ACU59240 *Chitinophaga*, SEQ ID NO:34; ACU05553 *Pedobacter*, SEQ ID NO:35.

FIG. 32 is an alignment of the amino acid sequence of CcMan5 (SEQ ID NO:50) and 19 of its homologs using MUSCLE. *Streptomyces* NP_630514, SEQ ID NO:26; *Streptomyces* ZP_02866543, SEQ ID NO:36; ZP_06527366 *Streptomyces*, SEQ ID NO:37; YP_003013376 *Paenibacillus*, SEQ ID NO:38; NP_812442 *Bacteroides*, SEQ ID NO:28; ZP_04848482 *Bacteroides*, SEQ ID NO:39; ZP_03677957 *Bacteroides*, SEQ ID NO:40; YP_003584502 *Zunongwangia*, SEQ ID NO:29; ZP_01061975 *Leeuwenhoekiella*, SEQ ID NO:41; ZP_07083984 *Sphingobacterium*, SEQ ID NO:42; YP_003120664 *Chitinophaga*, SEQ ID NO:30; ZP_01885202 *Pedobacter*, SEQ ID NO:43; ZP_02866543 *Clostridium*, SEQ ID NO:27; XP_367221 *Magnaporthe*, SEQ ID NO:44; ZP_07042437 *Bacteroides*, SEQ ID NO:45; ZP_05759807 *Bacteroides*, SEQ ID NO:46; ZP_05287524 *Bacteroides*, SEQ ID NO:47; ZP_06076108 *Bacteroides*, SEQ ID NO:48; YP_001302992 *Parabacteroides*, SEQ ID NO:49.

FIG. 33 contains the structural coordinates of the residues surrounding the active site of CcMan5₁₋₇₇₄.

FIG. 34 contains the protein C alpha atoms and the catalytic Ca²⁺ atoms of the two CcMan5₁₋₇₇₄ molecules in the asymmetric unit in PDB entry 2xsg and describes the overall fold of the protein.

DETAILED DESCRIPTION

In general, this document provides methods and materials for hydrolyzing mannose-1-phospho-6-mannose linkages on glycoproteins to produce target molecules (e.g., target proteins) having uncapped phospho-6-mannose (M6P) residues. The

methods and materials described herein are particularly useful for producing agents for treating patients with lysosomal storage disorders (LSDs), a diverse group of hereditary metabolic disorders characterized by the accumulation of storage products in the lysosomes due to impaired activity of catabolic enzymes involved in their degradation. The build-up of storage products leads to cell dysfunction and progressive clinical manifestations. Deficiencies in catabolic enzymes can be corrected by enzyme replacement therapy (ERT), provided that the administered enzyme can be targeted to the lysosomes of the diseased cells. Lysosomal enzymes typically are glycoproteins that are synthesized in the endoplasmic reticulum (ER), transported via the secretory pathway to the Golgi, and then recruited to the lysosomes. One way in which lysosomal enzymes are delivered to the lysosome is via a cation-dependent (CD) mannose 6-phosphate receptor (MPR). M6P terminal glycans are recognized in the trans-Golgi network (TGN) by two MPRs that mediate the sorting of lysosomal enzymes from the secretory pathway and deliver the enzyme to the lysosome. Using the methods and materials described herein, a microbial based production process can be used to obtain therapeutic proteins with uncapped M6P glycans, which can be delivered to lysosomes by exploiting the same M6P dependent pathway. Thus, the methods and materials described herein are useful for preparing glycoproteins for the treatment of metabolic disorders such as LSDs.

Mannosidases

This document provides isolated nucleic acids encoding mannosidase polypeptides capable of hydrolyzing terminal mannose-1-phospho-6-mannose linkages on oligosaccharides, as well as isolated mannosidases capable of hydrolyzing terminal mannose-1-phospho-6-mannose linkages on oligosaccharides. The terms “nucleic acid” and “polynucleotide” are used interchangeably herein, and refer to both RNA and DNA, including cDNA, genomic DNA, synthetic DNA, and DNA (or RNA) containing nucleic acid analogs. Polynucleotides can have any three-dimensional structure. A nucleic acid can be double-stranded or single-stranded (i.e., a sense strand or an antisense strand). Non-limiting examples of polynucleotides include genes, gene fragments, exons, introns, messenger RNA (mRNA), transfer RNA, ribosomal RNA, siRNA, micro-RNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids,

vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes, and primers, as well as nucleic acid analogs.

“Polypeptide” and “protein” are used interchangeably herein and mean any peptide-linked chain of amino acids, regardless of length or post-translational modification. Typically, a polypeptide described herein (e.g., a mannosidase or a target protein having uncapped M6P residues) is isolated when it constitutes at least 60%, by weight, of the total protein in a preparation, e.g., 60% of the total protein in a sample. In some embodiments, a polypeptide described herein consists of at least 75%, at least 90%, or at least 99%, by weight, of the total protein in a preparation.

An “isolated nucleic acid” refers to a nucleic acid that is separated from other nucleic acid molecules that are present in a naturally-occurring genome, including nucleic acids that normally flank one or both sides of the nucleic acid in a naturally-occurring genome (e.g., a yeast genome). The term “isolated” as used herein with respect to nucleic acids also includes any non-naturally-occurring nucleic acid sequence, since such non-naturally-occurring sequences are not found in nature and do not have immediately contiguous sequences in a naturally-occurring genome.

An isolated nucleic acid can be, for example, a DNA molecule, provided one of the nucleic acid sequences normally found immediately flanking that DNA molecule in a naturally-occurring genome is removed or absent. Thus, an isolated nucleic acid includes, without limitation, a DNA molecule that exists as a separate molecule (e.g., a chemically synthesized nucleic acid, or a cDNA or genomic DNA fragment produced by PCR or restriction endonuclease treatment) independent of other sequences as well as DNA that is incorporated into a vector, an autonomously replicating plasmid, a virus (e.g., any paramyxovirus, retrovirus, lentivirus, adenovirus, or herpes virus), or into the genomic DNA of a prokaryote or eukaryote. In addition, an isolated nucleic acid can include an engineered nucleic acid such as a DNA molecule that is part of a hybrid or fusion nucleic acid. A nucleic acid existing among hundreds to millions of other nucleic acids within, for example, cDNA libraries or genomic libraries, or gel slices containing a genomic DNA restriction digest, is not considered an isolated nucleic acid.

The term “exogenous” as used herein with reference to nucleic acid and a particular host cell refers to any nucleic acid that does not occur in (and cannot be

obtained from) that particular cell as found in nature. Thus, a non-naturally-occurring nucleic acid is considered to be exogenous to a host cell once introduced into the host cell. It is important to note that non-naturally-occurring nucleic acids can contain nucleic acid subsequences or fragments of nucleic acid sequences that are found in nature provided that the nucleic acid as a whole does not exist in nature. For example, a nucleic acid molecule containing a genomic DNA sequence within an expression vector is non-naturally-occurring nucleic acid, and thus is exogenous to a host cell once introduced into the host cell, since that nucleic acid molecule as a whole (genomic DNA plus vector DNA) does not exist in nature. Thus, any vector, autonomously replicating plasmid, or virus (e.g., retrovirus, adenovirus, or herpes virus) that as a whole does not exist in nature is considered to be non-naturally-occurring nucleic acid. It follows that genomic DNA fragments produced by PCR or restriction endonuclease treatment as well as cDNAs are considered to be non-naturally-occurring nucleic acid since they exist as separate molecules not found in nature. It also follows that any nucleic acid containing a promoter sequence and polypeptide-encoding sequence (e.g., cDNA or genomic DNA) in an arrangement not found in nature is non-naturally-occurring nucleic acid. A nucleic acid that is naturally-occurring can be exogenous to a particular cell. For example, an entire chromosome isolated from a cell of yeast x is an exogenous nucleic acid with respect to a cell of yeast y once that chromosome is introduced into a cell of yeast.

A nucleic acid encoding a mannosidase can have at least 70% sequence identity (e.g., at least 80%, 85%, 90%, 95%, 97%, 98%, 99%, or 100% sequence identity) to a nucleotide sequence set forth in SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, or SEQ ID NO:14. In some embodiments, nucleic acids described herein can encode mannosidase polypeptides that have at least 70% (e.g., at least 75, 80, 85, 90, 95, 99, or 100 percent) identity to an amino acid sequence set forth in SEQ ID NOs: 7, 9, 11, 13, 15, 50. For example, a nucleic acid can encode a mannosidase having at least 90% (e.g., at least 95 or 98%) identity to the amino acid sequence set forth in SEQ ID NO:15 or SEQ ID NO:50, or a portion thereof. For example, a nucleic acid can encode a mannosidase having at least 90% identity to amino acid residues 1 to 774 of SEQ ID NO:50. The percent identity between a particular amino acid sequence and the amino acid sequence set forth in SEQ ID NO:7, SEQ ID NO:9, SEQ ID NO:11, SEQ ID NO:13,

SEQ ID NO:15, or SEQ ID NO:50 is determined as follows. First, the amino acid sequences are aligned using the BLAST 2 Sequences (Bl2seq) program from the stand-alone version of BLASTZ containing BLASTP version 2.0.14. This stand-alone version of BLASTZ can be obtained from Fish & Richardson's web site (e.g., www.fr.com/blast/) or the U.S. government's National Center for Biotechnology Information web site (www.ncbi.nlm.nih.gov). Instructions explaining how to use the Bl2seq program can be found in the readme file accompanying BLASTZ. Bl2seq performs a comparison between two amino acid sequences using the BLASTP algorithm. To compare two amino acid sequences, the options of Bl2seq are set as follows: -i is set to a file containing the first amino acid sequence to be compared (e.g., C:\seq1.txt); -j is set to a file containing the second amino acid sequence to be compared (e.g., C:\seq2.txt); -p is set to blastp; -o is set to any desired file name (e.g., C:\output.txt); and all other options are left at their default setting. For example, the following command can be used to generate an output file containing a comparison between two amino acid sequences: C:\Bl2seq -i c:\seq1.txt -j c:\seq2.txt -p blastp -o c:\output.txt. If the two compared sequences share homology, then the designated output file will present those regions of homology as aligned sequences. If the two compared sequences do not share homology, then the designated output file will not present aligned sequences. Similar procedures can be following for nucleic acid sequences except that blastn is used.

Once aligned, the number of matches is determined by counting the number of positions where an identical amino acid residue is presented in both sequences. The percent identity is determined by dividing the number of matches by the length of the full-length mannosidase polypeptide amino acid sequence followed by multiplying the resulting value by 100. For example, an amino acid sequence that has 700 matches when aligned with the sequence set forth in SEQ ID NO:7 is 77.8 percent identical to the sequence set forth in SEQ ID NO:7 (i.e., $700 \div 900 * 100 = 77.8$).

It is noted that the percent identity value is rounded to the nearest tenth. For example, 78.11, 78.12, 78.13, and 78.14 is rounded down to 78.1, while 78.15, 78.16, 78.17, 78.18, and 78.19 is rounded up to 78.2. It also is noted that the length value will always be an integer.

It will be appreciated that a number of nucleic acids can encode a polypeptide

having a particular amino acid sequence. The degeneracy of the genetic code is well known to the art; i.e., for many amino acids, there is more than one nucleotide triplet that serves as the codon for the amino acid. For example, codons in the coding sequence for a given mannosidase polypeptide can be modified such that optimal expression in a particular species (e.g., bacteria or fungus) is obtained, using appropriate codon bias tables for that species. For example, the nucleic acids set forth in SEQ ID NO:6, SEQ ID NO:8, SEQ ID NO:10, SEQ ID NO:12, or SEQ ID NO:14 can be codon optimized for *E. coli* expression as set forth in FIGs. 14-18 (see SEQ ID NOs:16-20).

Hybridization also can be used to assess homology between two nucleic acid sequences. A nucleic acid sequence described herein, or a fragment or variant thereof, can be used as a hybridization probe according to standard hybridization techniques. The hybridization of a probe of interest (e.g., a probe containing a portion of a CcMan5 nucleotide sequence) to DNA or RNA from a test source is an indication of the presence of DNA or RNA (e.g., a CcMan5 nucleotide sequence) corresponding to the probe in the test source. Hybridization conditions are known to those skilled in the art and can be found in Current Protocols in Molecular Biology, John Wiley & Sons, N.Y., 6.3.1-6.3.6, 1991. Moderate hybridization conditions are defined as equivalent to hybridization in 2X sodium chloride/sodium citrate (SSC) at 30°C, followed by a wash in 1 X SSC, 0.1% SDS at 50°C. Highly stringent conditions are defined as equivalent to hybridization in 6X sodium chloride/sodium citrate (SSC) at 45°C, followed by a wash in 0.2 X SSC, 0.1% SDS at 65°C.

Mannosidase polypeptides capable of hydrolyzing terminal mannose-1-phospho-6-mannose linkages on oligosaccharides also can be identified based on the three dimensional structure described herein for a portion of a mannosidase from *C. cellulans* (residues 1 to 774 of SEQ ID NO:50, also referred to as CcMan5₁₋₇₇₄). The three dimensional structure can be determined by, for example, X-ray diffraction of a crystal of CcMan5₁₋₇₇₄. Structural coordinates of CcMan5₁₋₇₇₄ (e.g., the coordinates of CcMan5₁₋₇₇₄ deposited with the Protein Data Bank (world wide web at PDB.org under PDB ID No. 2xs), the coordinates set forth in FIG. 33 for the catalytic center of CcMan5, or the coordinates set forth in FIG. 34 for the protein C alpha atoms and the catalytic Ca²⁺ atoms of the two CcMan5₁₋₇₇₄ molecules in the asymmetric unit in PDB entry 2xsg) are

useful for a number of applications, including, but not limited to, the characterization of a three dimensional structure of a mannosidase capable of hydrolyzing terminal mannose-1-phospho-6-mannose linkages on oligosaccharides, as well as the visualization, identification and characterization of regions of the mannosidase that are involved in acceptance of mannose-6-phosphate- α ,1-mannose (hereafter referred to as Man-P-Man) as a substrate, and conferring its ability to hydrolyse Man-P-Man to produce a terminal phospho-6-mannose. "Structural coordinates" are the Cartesian coordinates corresponding to an atom's spatial relationship to other atoms in a molecule or molecular complex. Structural coordinates can be obtained using x-ray crystallography techniques or NMR techniques, or can be derived using molecular replacement analysis or homology modeling. Various software programs allow for the graphical representation of a set of structural coordinates to obtain a three dimensional representation of a molecule or molecular complex. The structural coordinates of the structures described herein can be modified from the original set provided in FIG. 33 or FIG. 34 by mathematical manipulation, such as by inversion or integer additions or subtractions. As such, it is recognized that the structural coordinates of the present invention are relative, and are in no way specifically limited by the actual x, y, z coordinates of FIG. 33 or FIG. 34.

As set forth in Example 8, the structure of CcMan₅₁₋₇₇₄ consists of two domains, an N-terminal β -sandwich domain (residues 8 to 271 of SEQ ID NO:50) and a C-terminal ($\alpha\alpha$)6 barrel domain (residues 291 to 771 of SEQ ID NO:50), connected via an α -helical linker (residues 272 to 290 of SEQ ID NO:50). The interface between both domains gives shape to a shallow cavity that harbors a conserved catalytic Ca²⁺ ion and gives shape to the -1 substrate binding site (nomenclature as described by Davies *et al.*, *Biochem. J.* 321:557-9 (1997)) and the catalytic center. Residues 22, 25, 71, 72, 195, 196, 354, 405, 535, 536, 588, 589, 626, 658, 660, and 662 of SEQ ID NO: 50 form the substrate binding site.

The three dimensional structure of CcMan₅₁₋₇₇₄ can be characterized in part, or all, using the structural coordinates of PDB ID No. 2xs, or an extract of which that is presented in FIG. 33, comprising the residues surrounding the active site of CcMan₅₁₋₇₇₄, or an extract of which that is presented in FIG. 34, comprising the protein C alpha atoms and the catalytic Ca²⁺ atoms of the two CcMan₅₁₋₇₇₄ molecules in the asymmetric unit in

PDB entry 2xsg, and describing the overall fold of the protein. For example, the three-dimensional structure of CcMan5₁₋₇₇₄ can be characterized by the structural coordinates of amino acid residues 7 to 771 according to PDB ID No. 2xs, \pm a root mean square deviation from the conserved backbone atoms of said amino acids of not more than 2Å. In some embodiments, the three dimensional structure of CcMan5₁₋₇₇₄ comprises the complete structural coordinates of the amino acids according to PDB ID No. 2xs, \pm a root mean square deviation from the conserved backbone atoms of said amino acids of not more than 2Å (e.g., not more than 1.5 Å, 1.0Å or 0.5Å). As used herein, “root mean square deviation” is the square root of the arithmetic mean of the squares of the deviations from the mean, and is a way of expressing deviation or variation from the structural coordinates described herein. The present disclosure includes all embodiments comprising conservative substitutions of the noted amino acid residues resulting in same structural coordinates within the stated root mean square deviation.

The structural coordinates provided herein can be used to characterize a three dimensional structure of a mannosidase polypeptide. From such a structure, substrate binding sites, for example, can be computationally visualized, identified and characterized based on the surface structure of the molecule, surface charge, steric arrangement, the presence of reactive amino acids, regions of hydrophobicity or hydrophilicity, etc.

In order to use the structural coordinates generated for a structure described herein as set forth in FIGs. 33, FIG. 34, or PDB ID No. 2xs, the relevant coordinates can be displayed as, or converted to, a three dimensional shape or graphical representation. Software programs are commercially available that are capable of generating three dimensional graphical representations of molecules or portions thereof from a set of structural coordinates. Examples of commercially available software programs include, without limitation, the following: GRID (Oxford University, Oxford, UK) ; MCSS (Molecular Simulations, San Diego, CA); AUTODOCK (Scripps Research Institute, La Jolla, CA); DOCK (University of California, San Francisco, CA); Flo99 (Thistlesoft, Morris Township, NJ); Ludi (Molecular Simulations, San Diego, CA); QUANTA (Molecular Simulations, San Diego, CA); Insight (Molecular Simulations, San Diego,

CA); SYBYL (TRIPOS, Inc., St. Louis, MO); and LEAPFROG (TRIPOS, Inc., St. Louis, MO).

The structural coordinates described herein can be used with standard homology modeling techniques in order to determine the unknown three-dimensional structure of a molecule or molecular complex. Homology modeling involves constructing a model of an unknown structure using structural coordinates of one or more related protein molecules, molecular complexes or parts thereof. Homology modeling can be conducted by fitting common or homologous portions of the protein whose three dimensional structure is to be solved to the three dimensional structure of homologous structural elements in the known molecule, specifically using the relevant (i.e., homologous) structural coordinates provided by FIGs. 33 and 34 herein. Homology may be determined using amino acid sequence identity, homologous secondary structure elements, and/or homologous tertiary folds. Homology modeling can include rebuilding part or all of a three dimensional structure with replacement of amino acids (or other components) by those of the related structure to be solved. Accordingly, a three dimensional structure for the unknown molecule may be generated using the three dimensional structure of CcMan5₁₋₇₇₄ described herein, and refined using a number of techniques well known in the art.

Based on the three dimensional structure described herein, substitutions can be made in some of the atoms or side groups of CcMan5₁₋₇₇₄ or other mannosidases in order to improve or modify its selectivity. For example, CcMan5 contains a non-acidic residue at positions 536 and 588, which may allow the mannosidase to tolerate the phosphate linkage to the anomeric oxygen in Man-P-Man substrates. As such, corresponding residues in other mannosidases can be changed to non-acidic residues to increase the ability of the mannosidase to accept Man-P-Man substrates.

Other mannosidase polypeptide candidates suitable for use herein can be identified by analysis of nucleotide and polypeptide sequence alignments. For example, performing a query on a database of nucleotide or polypeptide sequences can identify homologs and/or orthologs of mannosidase polypeptides. Sequence analysis can involve BLAST, Reciprocal BLAST, or PSI-BLAST analysis of nonredundant databases using known mannosidase amino acid sequences. Those polypeptides in the database that have

greater than 40% sequence identity can be identified as candidates for further evaluation for suitability as a mannosidase polypeptide. Amino acid sequence similarity allows for conservative amino acid substitutions, such as substitution of one hydrophobic residue for another or substitution of one polar residue for another. If desired, manual inspection of such candidates can be carried out in order to narrow the number of candidates to be further evaluated. Manual inspection can be performed by selecting those candidates that appear to have domains suspected of being present in mannosidases capable of hydrolyzing terminal mannose-1-phospho-6-mannose linkages, e.g., one or more (e.g., 1, 2, 3, 4 or more) conserved domains or functional regions (e.g., substrate binding cavity). Such domains can include a glycine-rich motif GVGXXGXGG, where X is Gly, Ser, Thr, Val, Ala, Cys or Gln (or other amino acid with a small side chain). This motif is found at residues 69-77 of SEQ ID NO:50. This region makes a loop that provides essential hydrogen bonds to the -1 mannose and phosphate-binding subsite in the active site of the enzyme.

Another example of a conserved motif includes a VRXE motif, where Arg (R) makes a hydrogen bond to the -1 ring and possibly the +1 ring, Glu (E) is in a salt bridge to this R residue, probably shaping this motif; and X is Trp or any of the 20 amino acids except Pro. This motif is found at residues 404-407 of SEQ ID NO:50.

A suitable motif also can be an X₁YQGX₂ motif, where X₁ can be Leu, Ile, Val, Ala, Phe, Tyr or Met, and X₂ can be Thr, Ser or Asn. This motif is found at residues 534-538 of SEQ ID NO:50. The Gln (Q) in this motif is important as an E is present in mannosidases that do not have the ability to hydrolyze terminal mannose-1-phospho-6-mannose linkages on oligosaccharides. The Tyr (Y) in this motif also is thought to be important for the +1 site formation.

In addition, a region defined by residues 22, 25, 71, 72, 195, 196, 354, 405, 535, 536, 588, 589, 626, 658, 660, and 662 of SEQ ID NO:50 forms the substrate binding cavity of CcMan5. As a minimal requirement, G71, G72, D355, R405, Q536, N588, Q589, T626, D660, D662 form the catalytic center, where N588, Q589 and D660 are involved in coordinating the catalytic Ca²⁺ ion, D662 and D660 are involved in activating the nucleophilic water, Q536 stabilizes the anomeric oxygen during the transition state and G71, G72, D355, R405 and T626 are involved in substrate binding at

the -1 site. See FIG. 30 for a representation of this minimal catalytic center. As such, a mannosidase can be selected as a candidate mannosidase capable of hydrolyzing terminal mannose-1-phospho-6-mannose linkages when the three dimensional protein coordinates of the atoms in the amino acid side chains located in the minimal catalytic center (e.g., as set forth in FIG. 30) fall within 1.5 Å deviation of the coordinates of the equivalent atoms in FIG. 33.

A conserved motif also can be a GDXGN motif in the N-terminal domain of the protein, where X can be any amino acid except P. This motif is found at residues 21-25 of SEQ ID NO:50 and forms part of the substrate binding pocket of the enzyme as shown in FIG. 24. In particular, the side chains of the D and N line the substrate binding cavity and may shape an alternative subpocket to bind the +1 mannose.

As set forth in Example 14, performing a query on a database of polypeptide sequences identified homologs of CcMan5 in the following organisms: *Streptomyces coelicolor* (GenBank Accession No. NP_630514), *Streptomyces lividans* (GenBank Accession No. ZP_05522540); *Streptomyces lividans* (GenBank Accession No. ZP_06527366); *Clostridium spiroforme* (GenBank Accession No. ZP_02866543), *Bacteroides thetaiotaomicron* (GenBank Accession No. NP_812442), *Zunongwangia profunda* (GenBank Accession No. YP_003584502); *Chitinophaga pinensis* (GenBank Accession No. YP_003120664); *Paenibacillus sp* (GenBank Accession No. YP_003013376); *Bacteroides sp.* (GenBank Accession No. ZP_04848482); *Bacteroides cellulosilyticus* (GenBank Accession No. ZP_03677957); *Leeuwenhoekiella blandensis* (GenBank Accession No. ZP_01061975); *Sphingobacterium spiritivorum* (GenBank Accession No. ZP_07083984); and *Pedobacter sp.* (GenBank Accession No. ZP_01885202). The mannosidases from *Streptomyces coelicolor* and *Streptomyces lividans* are similar (66% sequence identity to the CcMan5 GH92 domain, with 501 identities over 765 aligned residues by BLASTP), not only in the above motifs but also in many the loops of the three dimensional structure.

Isolated nucleic acid molecules encoding mannosidase polypeptides can be produced by standard techniques. For example, polymerase chain reaction (PCR) techniques can be used to obtain an isolated nucleic acid containing a nucleotide sequence described herein. PCR can be used to amplify specific sequences from DNA as

well as RNA, including sequences from total genomic DNA or total cellular RNA. Generally, sequence information from the ends of the region of interest or beyond is employed to design oligonucleotide primers that are identical or similar in sequence to opposite strands of the template to be amplified. Various PCR strategies also are available by which site-specific nucleotide sequence modifications can be introduced into a template nucleic acid. Isolated nucleic acids also can be chemically synthesized, either as a single nucleic acid molecule (e.g., using automated DNA synthesis in the 3' to 5' direction using phosphoramidite technology) or as a series of oligonucleotides. For example, one or more pairs of long oligonucleotides (e.g., >100 nucleotides) can be synthesized that contain the desired sequence, with each pair containing a short segment of complementarity (e.g., about 15 nucleotides) such that a duplex is formed when the oligonucleotide pair is annealed. DNA polymerase is used to extend the oligonucleotides, resulting in a single, double-stranded nucleic acid molecule per oligonucleotide pair, which then can be ligated into a vector. Isolated nucleic acids of the invention also can be obtained by mutagenesis of, e.g., a naturally occurring DNA.

This document also provides (i) biologically active variants and (ii) biologically active fragments or biologically active variants thereof, of the mannosidases described herein. Biologically active variants of mannosidases can contain additions, deletions, or substitutions relative to the sequences set forth in SEQ ID NOs: 7, 9, 11, 13, 15, or 50. Proteins with substitutions will generally have not more than 50 (e.g., not more than one, two, three, four, five, six, seven, eight, nine, ten, 12, 15, 20, 25, 30, 35, 40, or 50) conservative amino acid substitutions. A conservative substitution is the substitution of one amino acid for another with similar characteristics. Conservative substitutions include substitutions within the following groups: valine, alanine and glycine; leucine, valine, and isoleucine; aspartic acid and glutamic acid; asparagine and glutamine; serine, cysteine, and threonine; lysine and arginine; and phenylalanine and tyrosine. The non-polar hydrophobic amino acids include alanine, leucine, isoleucine, valine, proline, phenylalanine, tryptophan and methionine. The polar neutral amino acids include glycine, serine, threonine, cysteine, tyrosine, asparagine and glutamine. The positively charged (basic) amino acids include arginine, lysine and histidine. The negatively charged (acidic) amino acids include aspartic acid and glutamic acid. Any substitution of

one member of the above-mentioned polar, basic or acidic groups by another member of the same group can be deemed a conservative substitution. By contrast, a non-conservative substitution is a substitution of one amino acid for another with dissimilar characteristics. The sequence alignments set forth in FIGs. 31 and 32 provide numerous examples of amino acid substitutions that can be made.

Deletion variants can lack one, two, three, four, five, six, seven, eight, nine, ten, 11, 12, 13, 14, 15, 16, 17, 18, 19, or 20 amino acid segments (of two or more amino acids) or non-contiguous single amino acids.

Additions (addition variants) include fusion proteins containing: (a) a mannosidase set forth in SEQ ID NOs: 7, 9, 11, 13, or 15, or a fragment thereof; and (b) internal or terminal (C or N) irrelevant or heterologous amino acid sequences. In the context of such fusion proteins, the term "heterologous amino acid sequences" refers to an amino acid sequence other than (a). A heterologous sequence can be, for example a sequence used for purification of the recombinant protein (e.g., FLAG, polyhistidine (e.g., hexahistidine), hemagglutinin (HA), glutathione-S-transferase (GST), or maltose-binding protein (MBP)). Heterologous sequences also can be proteins useful as diagnostic or detectable markers, for example, luciferase, green fluorescent protein (GFP), or chloramphenicol acetyl transferase (CAT). In some embodiments, the fusion protein contains a signal sequence from another protein. In certain host cells (e.g., yeast host cells), expression and/or secretion of the target protein can be increased through use of a heterologous signal sequence. In some embodiments, the fusion protein can contain a carrier (e.g., KLH) useful, e.g., in eliciting an immune response for antibody generation) or endoplasmic reticulum or Golgi apparatus retention signals. Heterologous sequences can be of varying length and in some cases can be a longer sequences than the full-length target proteins to which the heterologous sequences are attached.

Biologically active fragments or biologically active variants of the mannosidases have at least 40% (e.g., at least: 50%; 60%; 70%; 75%; 80%; 85%; 90%; 95%; 97%; 98%; 99%; 99.5%, or 100% or even greater) of the mannosidase activity (e.g., uncapping of M6P residues) of the wild-type, full-length, mature protein. For example, a biologically active fragment of a mannosidase can contain residues 1 to 774 of SEQ ID NO:50.

The mannosidases described herein can be used to produce molecules (e.g., target proteins) having uncapped terminal phospho-6-mannose (M6P) residues. The methods can be performed *in vitro* or *in vivo*.

In Vitro Methods of Uncapping M6P Residues

A mannosidase described herein can be recombinantly produced and used *in vitro* to uncap terminal M6P residues on oligosaccharides. To recombinantly produce a mannosidase, a vector is used that contains a promoter operably linked to nucleic acid encoding a mannosidase polypeptide. As used herein, a "promoter" refers to a DNA sequence that enables a gene to be transcribed. The promoter is recognized by RNA polymerase, which then initiates transcription. Thus, a promoter contains a DNA sequence that is either bound directly by, or is involved in the recruitment, of RNA polymerase. A promoter sequence can also include "enhancer regions," which are one or more regions of DNA that can be bound with proteins (namely, the trans-acting factors, much like a set of transcription factors) to enhance transcription levels of genes (hence the name) in a gene-cluster. The enhancer, while typically at the 5' end of a coding region, can also be separate from a promoter sequence and can be, e.g., within an intronic region of a gene or 3' to the coding region of the gene.

As used herein, "operably linked" means incorporated into a genetic construct (e.g., vector) so that expression control sequences effectively control expression of a coding sequence of interest.

Expression vectors can be introduced into host cells (e.g., by transformation or transfection) for expression of the encoded polypeptide, which then can be purified. Expression systems that can be used for small or large scale production of mannosidase polypeptides include, without limitation, microorganisms such as bacteria (e.g., *E. coli*) transformed with recombinant bacteriophage DNA, plasmid DNA, or cosmid DNA expression vectors containing the nucleic acid molecules, and fungal (e.g., *S. cerevisiae*, *Yarrowia lipolytica*, *Arxula adenivorans*, *Pichia pastoris*, *Hansenula polymorpha*, or *Aspergillus*) transformed with recombinant fungal expression vectors containing the nucleic acid molecules. Useful expression systems also include insect cell systems infected with recombinant virus expression vectors (e.g., baculovirus) containing the

nucleic acid molecules, and plant cell systems infected with recombinant virus expression vectors (e.g., tobacco mosaic virus) or transformed with recombinant plasmid expression vectors (e.g., Ti plasmid) containing the nucleic acid molecules. Mannosidase polypeptides also can be produced using mammalian expression systems, which include cells (e.g., immortalized cell lines such as COS cells, Chinese hamster ovary cells, HeLa cells, human embryonic kidney 293 cells, and 3T3 L1 cells) harboring recombinant expression constructs containing promoters derived from the genome of mammalian cells (e.g., the metallothionein promoter) or from mammalian viruses (e.g., the adenovirus late promoter and the cytomegalovirus promoter), along with the nucleic acids described herein.

Typically, recombinant mannosidase polypeptides are tagged with a heterologous amino acid sequence such FLAG, polyhistidine (e.g., hexahistidine), hemagglutinin (HA), glutathione-S-transferase (GST), or maltose-binding protein (MBP) to aid in purifying the protein. Other methods for purifying proteins include chromatographic techniques such as ion exchange, hydrophobic and reverse phase, size exclusion, affinity, hydrophobic charge-induction chromatography, and the like (see, e.g., Scopes, *Protein Purification: Principles and Practice*, third edition, Springer-Verlag, New York (1993); Burton and Harding, *J. Chromatogr. A* 814:71-81 (1998)).

To produce molecules having uncapped terminal M6P residues *in vitro*, a target molecule containing a mannose-1-phospho-6 mannose linkage is contacted under suitable conditions with a purified mannosidase or a cell lysate containing a recombinantly produced mannosidase. The cell lysate can be from any genetically engineered cell, including a fungal cell, a plant cell, or animal cell. Non-limiting examples of animal cells include nematode, insect, plant, bird, reptile, and mammals such as a mouse, rat, rabbit, hamster, gerbil, dog, cat, goat, pig, cow, horse, whale, monkey, or human. Upon contacting the target molecule (e.g., an oligosaccharide or glycoprotein) with the purified mannosidase or cell lysate, the mannosidase hydrolyzes the mannose-1-phospho-6 mannose linkage and produces a target molecule having one or more uncapped terminal M6P residues. The methods described in Example 2 can be used to determine if the terminal M6P residues have been uncapped. Following processing by the mannosidase, the target molecule having uncapped terminal M6P residues can be isolated.

Suitable methods for obtaining cell lysates that preserve the activity or integrity of the mannosidase activity in the lysate can include the use of appropriate buffers and/or inhibitors, including nuclease, protease and phosphatase inhibitors that preserve or minimize changes in N-glycosylation activities in the cell lysate. Such inhibitors include, for example, chelators such as ethylenediamine tetraacetic acid (EDTA), ethylene glycol bis(P-aminoethyl ether) N,N,N1,N1-tetraacetic acid (EGTA), protease inhibitors such as phenylmethylsulfonyl fluoride (PMSF), aprotinin, leupeptin, antipain and the like, and phosphatase inhibitors such as phosphate, sodium fluoride, vanadate and the like. Appropriate buffers and conditions for obtaining lysates containing enzymatic activities are described in, e.g., Ausubel et al. *Current Protocols in Molecular Biology* (Supplement 47), John Wiley & Sons, New York (1999); Harlow and Lane, *Antibodies: A Laboratory Manual* Cold Spring Harbor Laboratory Press (1988); Harlow and Lane, *Using Antibodies: A Laboratory Manual*, Cold Spring Harbor Press (1999); Tietz *Textbook of Clinical Chemistry*, 3rd ed. Burtis and Ashwood, eds. W.B. Saunders, Philadelphia, (1999).

A cell lysate can be further processed to eliminate or minimize the presence of interfering substances, as appropriate. If desired, a cell lysate can be fractionated by a variety of methods well known to those skilled in the art, including subcellular fractionation, and chromatographic techniques such as ion exchange, hydrophobic and reverse phase, size exclusion, affinity, hydrophobic charge-induction chromatography, and the like.

In some embodiments, a cell lysate can be prepared in which whole cellular organelles remain intact and/or functional. For example, a lysate can contain one or more of intact rough endoplasmic reticulum, intact smooth endoplasmic reticulum, or intact Golgi apparatus. Suitable methods for preparing lysates containing intact cellular organelles and testing for the functionality of the organelles are described in, e.g., Moreau *et al.* (1991) *J. Biol. Chem.* 266(7):4329-4333; Moreau *et al.* (1991) *J. Biol. Chem.* 266(7):4322-4328; Rexach *et al.* (1991) *J. Cell Biol.* 114(2):219-229; and Paulik *et al.* (1999) *Arch. Biochem. Biophys.* 367(2):265-273.

Target molecules, as used herein, refer to any molecule containing terminal mannose-1-phospho-6 mannose linkages or any molecule, when expressed in a cell of

fungal origin, that contains mannose-1-phospho-6 mannose linkages. Suitable target proteins include pathogen proteins such as tetanus toxoid or diphtheria toxoid; viral surface proteins such as cytomegalovirus (CMV) glycoproteins B, H and gCIII, human immunodeficiency virus 1 (HIV-1) envelope glycoproteins, Rous sarcoma virus (RSV) envelope glycoproteins, herpes simplex virus (HSV) envelope glycoproteins, Epstein Barr virus (EBV) envelope glycoproteins, varicella-zoster virus (VZV) envelope glycoproteins, human papilloma virus (HPV) envelope glycoproteins, Influenza virus glycoproteins, and Hepatitis family surface antigen; lysosomal proteins (e.g., acid alpha glucosidase, alpha galactosidase, glucocerebrosidase, cerebrosidase, or galactocerebrosidase); insulin; glucagons; growth factors; cytokines; chemokines; and antibodies or fragments thereof. Growth factors include, e.g., vascular endothelial growth factor (VEGF), Insulin-like growth factor (IGF), bone morphogenic protein (BMP), Granulocyte-colony stimulating factor (G-CSF), Granulocyte-macrophage colony stimulating factor (GM-CSF), Nerve growth factor (NGF); a Neurotrophin, Platelet-derived growth factor (PDGF), Erythropoietin (EPO), Thrombopoietin (TPO), Myostatin (GDF-8), Growth Differentiation factor-9 (GDF9), basic fibroblast growth factor (bFGF or FGF2), Epidermal growth factor (EGF), Hepatocyte growth factor (HGF). Cytokines include, for example, interleukins such as IL-1 to IL-33 (e.g., IL-1, IL-2, IL-3, IL-4, IL-5, IL-6, IL-7, IL-8, IL-9, IL-10, IL-12, IL-13, or IL-15)). Chemokines include, e.g., I-309, TCA-3, MCP-1, MIP-1 α , MIP-1 β , RANTES, C10, MRP-2, MARC, MCP-3, MCP-2, MRP-2, CCF18, MIP-1 γ , Eotaxin, MCP-5, MCP-4, NCC-1, Ck β 10, HCC-1, Leukotactin-1, LEC, NCC-4, TARC, PARC, or Eotaxin-2. Also included are tumor glycoproteins (e.g., tumor-associated antigens), for example, carcinoembryonic antigen (CEA), human mucins, HER-2/neu, and prostate-specific antigen (PSA) [Henderson and Finn, *Advances in Immunology*, 62, pp. 217-56 (1996)].

In some embodiments, the target protein can be one associated with a lysosomal storage disorder, which target proteins include, e.g., acid alpha glucosidase, alpha galactosidase, alpha-L-iduronidase, beta-D-galactosidase, beta-glucosidase, beta-hexosaminidase, beta-D-mannosidase, alpha-L-fucosidase, arylsulfatase B, arylsulfatase A, alpha-N-acetylgalactosaminidase, aspartylglucosaminidase, iduronate-2-sulfatase, alpha-glucosaminide-N-acetyltransferase, beta-D-glucuronidase, hyaluronidase, alpha-L-

mannosidase, alpha-neuraminidase, phosphotransferase, acid lipase, acid ceramidase, sphingomyelinase, thioesterase, cathepsin K, and lipoprotein lipase.

In some embodiments, the target proteins are fusion proteins in which the target protein is fused to another polypeptide sequence, or to a polymer, a carrier, an adjuvant, an immunotoxin, or a detectable (e.g., fluorescent, luminescent, or radioactive) moiety. For example, a target protein can be joined to a polymer such as polyethyleneglycol to increase the molecular weight of small proteins and/or increase circulation residence time.

In Vivo Methods of Uncapping M6P Residues

Genetically engineered cells described herein can be used to produce target molecules containing uncapped M6P residues. For example, a cell based method can include the steps of introducing into a fungal cell genetically engineered to include a nucleic acid encoding a mannosidase, a nucleic acid encoding a target molecule, wherein the cell produces the target molecule containing uncapped terminal M6P residues. In some embodiments, the nucleic acids encoding the mannosidase and target molecule contain a secretion sequence such that the mannosidase and target molecule are co-secreted.

Genetically engineered cells described herein contain a nucleic acid encoding a mannosidase and are useful for producing one or more target molecules having uncapped terminal M6P residues. Cells suitable for *in vivo* production of uncapped M6P residues can be of fungal origin, including *Yarrowia lipolytica*, *Arxula adeninivorans*, methylotrophic yeast (such as a methylotrophic yeast of the genus *Candida*, *Hansenula*, *Oogataea*, *Pichia* or *Torulopsis*) or filamentous fungi of the genus *Aspergillus*, *Trichoderma*, *Neurospora*, *Fusarium*, or *Chrysosporium*. Exemplary fungal species include, without limitation, *Pichia anomala*, *Pichia bovis*, *Pichia canadensis*, *Pichia carsonii*, *Pichia farinose*, *Pichia fermentans*, *Pichia fluxuum*, *Pichia membranaefaciens*, *Pichia membranaefaciens*, *Candida valida*, *Candida albicans*, *Candida ascalaphidarum*, *Candida amphixiae*, *Candida Antarctica*, *Candida atlantica*, *Candida atmosphaerica*, *Candida blattae*, *Candida carpophila*, *Candida cerambycidarum*, *Candida chauliodes*, *Candida corydalis*, *Candida dosseyi*, *Candida dubliniensis*, *Candida ergatensis*, *Candida*

fructus, Candida glabrata, Candida fermentati, Candida guilliermondii, Candida haemulonii, Candida insectamens, Candida insectorum, Candida intermedia, Candida jeffresii, Candida kefyri, Candida krusei, Candida lusitanae, Candida lyxosophila, Candida maltosa, Candida membranifaciens, Candida milleri, Candida oleophila, Candida oregonensis, Candida parapsilosis, Candida quercitrusa, Candida shehatei, Candida temnochilae, Candida tenuis, Candida tropicalis, Candida tsuchiyae, Candida sinolaborantium, Candida sojae, Candida viswanathii, Candida utilis, Oogataea minuta, Pichia membranaefaciens, Pichia silvestris, Pichia membranaefaciens, Pichia chodati, Pichia membranaefaciens, Pichia menbranaefaciens, Pichia minuscule, Pichia pastoris, Pichia pseudopolymorpha, Pichia quercuum, Pichia robertsii, Pichia saitoi, Pichia silvestrisi, Pichia strasburgensis, Pichia terricola, Pichia vanriji, Pseudozyma Antarctica, Rhodosporidium toruloides, Rhodotorula glutinis, Saccharomyces bayanus, Saccharomyces bayanus, Saccharomyces momdshuricus, Saccharomyces uvarum, Saccharomyces bayanus, Saccharomyces cerevisiae, Saccharomyces bisporus, Saccharomyces chevalieri, Saccharomyces delbrueckii, Saccharomyces exiguous, Saccharomyces fermentati, Saccharomyces fragilis, Saccharomyces marxianus, Saccharomyces mellis, Saccharomyces rosei, Saccharomyces rouxii, Saccharomyces uvarum, Saccharomyces willianus, Saccharomycodes ludwigii, Saccharomycopsis capsularis, Saccharomycopsis fibuligera, Saccharomycopsis fibuligera, Endomyces hordei, Endomycopsis fobuligera. Saturnispora saitoi, Schizosaccharomyces octosporus, Schizosaccharomyces pombe, Schwanniomyces occidentalis, Torulaspora delbrueckii, Torulaspora delbrueckii, Saccharomyces dairensis, Torulaspora delbrueckii, Torulaspora fermentati, Saccharomyces fermentati, Torulaspora delbrueckii, Torulaspora rosei, Saccharomyces rosei, Torulaspora delbrueckii, Saccharomyces rosei, Torulaspora delbrueckii, Saccharomyces delbrueckii, Torulaspora delbrueckii, Saccharomyces delbrueckii, Zygosaccharomyces mongolicus, Dorulaspora globosa, Debaryomyces globosus, Torulopsis globosa, Trichosporon cutaneum, Trigonopsis variabilis, Williopsis californica, Williopsis saturnus, Zygosaccharomyces bisporus, Zygosaccharomyces bisporus, Debaryomyces disporua. Saccharomyces bisporas, Zygosaccharomyces bisporus, Saccharomyces bisporus, Zygosaccharomyces mellis, Zygosaccharomyces priorianus, Zygosaccharomyces rouxiim, Zygosaccharomyces rouxii, Zygosaccharomyces

barkeri, *Saccharomyces rouxii*, *Zygosaccharomyces rouxii*, *Zygosaccharomyces major*, *Saccharomyces rousii*, *Pichia anomala*, *Pichia bovis*, *Pichia Canadensis*, *Pichia carsonii*, *Pichia farinose*, *Pichia fermentans*, *Pichia fluxuum*, *Pichia membranaefaciens*, *Pichia pseudopolymorpha*, *Pichia quercuum*, *Pichia robertsii*, *Pseudozyma Antarctica*, *Rhodospiridium toruloides*, *Rhodospiridium toruloides*, *Rhodotorula glutinis*, *Saccharomyces bayanus*, *Saccharomyces bayanus*, *Saccharomyces bisporus*, *Saccharomyces cerevisiae*, *Saccharomyces chevalieri*, *Saccharomyces delbrueckii*, *Saccharomyces fermentati*, *Saccharomyces fragilis*, *Saccharomyces ludwigii*, *Schizosaccharomyces pombe*, *Schwanniomyces occidentalis*, *Torulaspora delbrueckii*, *Torulaspora globosa*, *Trigonopsis variabilis*, *Williopsis californica*, *Williopsis saturnus*, *Zygosaccharomyces bisporus*, *Zygosaccharomyces mellis*, or *Zygosaccharomyces rouxii*. Exemplary filamentous fungi include various species of *Aspergillus* including, but not limited to, *Aspergillus caesiellus*, *Aspergillus candidus*, *Aspergillus carneus*, *Aspergillus clavatus*, *Aspergillus deflectus*, *Aspergillus flavus*, *Aspergillus fumigatus*, *Aspergillus glaucus*, *Aspergillus nidulans*, *Aspergillus niger*, *Aspergillus ochraceus*, *Aspergillus oryzae*, *Aspergillus parasiticus*, *Aspergillus penicilloides*, *Aspergillus restrictus*, *Aspergillus sojae*, *Aspergillus sydowi*, *Aspergillus tamari*, *Aspergillus terreus*, *Aspergillus ustus*, or *Aspergillus versicolor*. Such cells, prior to the genetic engineering as specified herein, can be obtained from a variety of commercial sources and research resource facilities, such as, for example, the American Type Culture Collection (Rockville, MD). Target molecules include proteins such as any of the target proteins described herein (see above).

Genetic engineering of a cell can include, in addition to an exogenous nucleic acid encoding a mannosidase, one or more genetic modifications such as: (i) deletion of an endogenous gene encoding an Outer CHain elongation (OCH1) protein; (ii) introduction of a recombinant nucleic acid encoding a polypeptide capable of promoting mannosyl phosphorylation (e.g, a MNN4 polypeptide from *Yarrowia lipolytica*, *S. cerevisiae*, *Ogataea minuta*, *Pichia pastoris*, or *C. albicans*, or PNO1 polypeptide from *P. pastoris*) to increasing phosphorylation of mannose residues; (iii) introduction or expression of an RNA molecule that interferes with the functional expression of an OCH1 protein; (iv) introduction of a recombinant nucleic acid encoding a wild-type (e.g., endogenous or

exogenous) protein having a N-glycosylation activity (i.e., expressing a protein having an N-glycosylation activity); (v) introduction of a recombinant nucleic acid encoding a target molecule described above; or (v) altering the promoter or enhancer elements of one or more endogenous genes encoding proteins having N-glycosylation activity to thus alter the expression of their encoded proteins. RNA molecules include, e.g., small-interfering RNA (siRNA), short hairpin RNA (shRNA), anti-sense RNA, or micro RNA (miRNA). Genetic engineering also includes altering an endogenous gene encoding a protein having an N-glycosylation activity to produce a protein having additions (e.g., a heterologous sequence), deletions, or substitutions (e.g., mutations such as point mutations; conservative or non-conservative mutations). Mutations can be introduced specifically (e.g., by site-directed mutagenesis or homologous recombination) or can be introduced randomly (for example, cells can be chemically mutagenized as described in, e.g., Newman and Ferro-Novick (1987) *J. Cell Biol.* 105(4):1587.

Genetic modifications described herein can result in one or more of (i) an increase in one or more activities in the genetically modified cell, (ii) a decrease in one or more activities in the genetically modified cell, or (iii) a change in the localization or intracellular distribution of one or more activities in the genetically modified cell. It is understood that an increase in the amount of a particular activity (e.g., promoting mannosyl phosphorylation) can be due to overexpressing one or more proteins capable of promoting mannosyl phosphorylation, an increase in copy number of an endogenous gene (e.g., gene duplication), or an alteration in the promoter or enhancer of an endogenous gene that stimulates an increase in expression of the protein encoded by the gene. A decrease in one or more particular activities can be due to overexpression of a mutant form (e.g., a dominant negative form), introduction or expression of one or more interfering RNA molecules that reduce the expression of one or more proteins having a particular activity, or deletion of one or more endogenous genes that encode a protein having the particular activity.

To disrupt a gene by homologous recombination, a “gene replacement” vector can be constructed in such a way to include a selectable marker gene. The selectable marker gene can be operably linked, at both 5' and 3' end, to portions of the gene of sufficient length to mediate homologous recombination. The selectable marker can be one of any

number of genes which either complement host cell auxotrophy or provide antibiotic resistance, including URA3, LEU2 and HIS3 genes. Other suitable selectable markers include the CAT gene, which confers chloramphenicol resistance to yeast cells, or the lacZ gene, which results in blue colonies due to the expression of β -galactosidase. Linearized DNA fragments of the gene replacement vector are then introduced into the cells using methods well known in the art (see below). Integration of the linear fragments into the genome and the disruption of the gene can be determined based on the selection marker and can be verified by, for example, Southern blot analysis. A selectable marker can be removed from the genome of the host cell by, e.g., Cre-loxP systems (see below).

Alternatively, a gene replacement vector can be constructed in such a way as to include a portion of the gene to be disrupted, which portion is devoid of any endogenous gene promoter sequence and encodes none or an inactive fragment of the coding sequence of the gene. An "inactive fragment" is a fragment of the gene that encodes a protein having, e.g., less than about 10% (e.g., less than about 9%, less than about 8%, less than about 7%, less than about 6%, less than about 5%, less than about 4%, less than about 3%, less than about 2%, less than about 1%, or 0%) of the activity of the protein produced from the full-length coding sequence of the gene. Such a portion of the gene is inserted in a vector in such a way that no known promoter sequence is operably linked to the gene sequence, but that a stop codon and a transcription termination sequence are operably linked to the portion of the gene sequence. This vector can be subsequently linearized in the portion of the gene sequence and transformed into a cell. By way of single homologous recombination, this linearized vector is then integrated in the endogenous counterpart of the gene.

Expression vectors can be autonomous or integrative. A recombinant nucleic acid (e.g., one encoding a mannosidase) can be introduced into the cell in the form of an expression vector such as a plasmid, phage, transposon, cosmid or virus particle. The recombinant nucleic acid can be maintained extrachromosomally or it can be integrated into the yeast cell chromosomal DNA. Expression vectors can contain selection marker genes encoding proteins required for cell viability under selected conditions (e.g., URA3, which encodes an enzyme necessary for uracil biosynthesis or TRP1, which encodes an enzyme required for tryptophan biosynthesis) to permit detection and/or selection of

those cells transformed with the desired nucleic acids (see, e.g., U.S. Pat. No. 4,704,362). Expression vectors can also include an autonomous replication sequence (ARS). For example, U.S. Pat. No. 4,837,148 describes autonomous replication sequences which provide a suitable means for maintaining plasmids in *Pichia pastoris*.

Integrative vectors are disclosed, e.g., in U.S. Pat. No. 4,882,279. Integrative vectors generally include a serially arranged sequence of at least a first insertable DNA fragment, a selectable marker gene, and a second insertable DNA fragment. The first and second insertable DNA fragments are each about 200 (e.g., about 250, about 300, about 350, about 400, about 450, about 500, or about 1000 or more) nucleotides in length and have nucleotide sequences which are homologous to portions of the genomic DNA of the species to be transformed. A nucleotide sequence containing a gene of interest (e.g., a gene encoding a protein having N-glycosylation activity) for expression is inserted in this vector between the first and second insertable DNA fragments whether before or after the marker gene. Integrative vectors can be linearized prior to yeast transformation to facilitate the integration of the nucleotide sequence of interest into the host cell genome.

An expression vector can feature a recombinant nucleic acid under the control of a yeast (e.g., *Yarrowia lipolytica*, *Arxula adenivorans*, *P. pastoris*, or other suitable fungal species) promoter, which enables them to be expressed in fungal cells. Suitable yeast promoters include, e.g., ADC1, TPI1, ADH2, hp4d, POX, and Gal10 (see, e.g., Guarente *et al.* (1982) *Proc. Natl. Acad. Sci. USA* 79(23):7410) promoters. Additional suitable promoters are described in, e.g., Zhu and Zhang (1999) *Bioinformatics* 15(7-8):608-611 and U.S. Patent No. 6,265,185.

A promoter can be constitutive or inducible (conditional). A constitutive promoter is understood to be a promoter whose expression is constant under the standard culturing conditions. Inducible promoters are promoters that are responsive to one or more induction cues. For example, an inducible promoter can be chemically regulated (e.g., a promoter whose transcriptional activity is regulated by the presence or absence of a chemical inducing agent such as an alcohol, tetracycline, a steroid, a metal, or other small molecule) or physically regulated (e.g., a promoter whose transcriptional activity is regulated by the presence or absence of a physical inducer such as light or high or low

temperatures). An inducible promoter can also be indirectly regulated by one or more transcription factors that are themselves directly regulated by chemical or physical cues.

It is understood that other genetically engineered modifications can also be conditional. For example, a gene can be conditionally deleted using, e.g., a site-specific DNA recombinase such as the Cre-loxP system (see, e.g., Gossen *et al.* (2002) *Ann. Rev. Genetics* 36:153-173 and U.S. Application Publication No. 20060014264).

A recombinant nucleic acid can be introduced into a cell described herein using a variety of methods such as the spheroplast technique or the whole-cell lithium chloride yeast transformation method. Other methods useful for transformation of plasmids or linear nucleic acid vectors into cells are described in, for example, U.S. Patent No. 4,929,555; Hinnen *et al.* (1978) *Proc. Nat. Acad. Sci. USA* 75:1929; Ito *et al.* (1983) *J. Bacteriol.* 153:163; U.S. Patent No. 4,879,231; and Sreekrishna *et al.* (1987) *Gene* 59:115, the disclosures of each of which are incorporated herein by reference in their entirety. Electroporation and PEG1000 whole cell transformation procedures may also be used, as described by Cregg and Russel, *Methods in Molecular Biology: Pichia Protocols*, Chapter 3, Humana Press, Totowa, N.J., pp. 27-39 (1998).

Transformed fungal cells can be selected for by using appropriate techniques including, but not limited to, culturing auxotrophic cells after transformation in the absence of the biochemical product required (due to the cell's auxotrophy), selection for and detection of a new phenotype, or culturing in the presence of an antibiotic which is toxic to the yeast in the absence of a resistance gene contained in the transformants. Transformants can also be selected and/or verified by integration of the expression cassette into the genome, which can be assessed by, e.g., Southern blot or PCR analysis.

Prior to introducing the vectors into a target cell of interest, the vectors can be grown (e.g., amplified) in bacterial cells such as *Escherichia coli* (*E. coli*) as described above. The vector DNA can be isolated from bacterial cells by any of the methods known in the art which result in the purification of vector DNA from the bacterial milieu. The purified vector DNA can be extracted extensively with phenol, chloroform, and ether, to ensure that no *E. coli* proteins are present in the plasmid DNA preparation, since these proteins can be toxic to mammalian cells.

In some embodiments, the genetically engineered fungal cell lacks the OCH1 gene or gene products (e.g., mRNA or protein) thereof, and is deficient in OCH1 activity. In some embodiments, the genetically engineered cell expresses a polypeptide capable of promoting mannosyl phosphorylation (e.g., a MNN4 polypeptide from *Yarrowia lipolytica*, *S. cerevisiae*, *Ogataea minuta*, *Pichia pastoris*, or *C. albicans*, or a PNO1 polypeptide from *P. pastoris*). For example, the fungal cell can express a MNN4 polypeptide from *Y. lipolytica* (Genbank® Accession Nos: XM_503217, Genolevures Ref: YALI0D24101g). In some embodiments, the genetically engineered cell is deficient in OCH1 activity and expresses a polypeptide capable of promoting mannosyl phosphorylation.

Following uncapping of the M6P residues, the target molecule can be isolated. In some embodiments, the target molecule is maintained within the yeast cell and released upon cell lysis. In some embodiments, the target molecule is secreted into the culture medium via a mechanism provided by a coding sequence (either native to the exogenous nucleic acid or engineered into the expression vector), which directs secretion of the molecule from the cell. The presence of the uncapped target molecule in the cell lysate or culture medium can be verified by a variety of standard protocols for detecting the presence of the molecule. For example, where the altered target molecule is a protein, such protocols can include, but are not limited to, immunoblotting or radioimmunoprecipitation with an antibody specific for the altered target protein (or the target protein itself), binding of a ligand specific for the altered target protein (or the target protein itself), or testing for a specific enzyme activity of the altered target protein (or the target protein itself).

In the target molecules produced using the methods described herein, at least 47% (e.g., at least 50, 55, 60, 65, 70, 75, 80, 85, or 90%) of the N-glycans on the glycoprotein have terminal phospho-6-mannose residues. The percentage of N-glycans having terminal phospho-6-mannose residues can be estimated from the peak areas in the DSA-FACE electropherograms. See Example 13.

In some embodiments, following isolation, the uncapped target molecule can be attached to a heterologous moiety, e.g., using enzymatic or chemical means. A “heterologous moiety” refers to any constituent that is joined (e.g., covalently or non-

covalently) to the altered target molecule, which constituent is different from a constituent originally present on the altered target molecule. Heterologous moieties include, e.g., polymers, carriers, adjuvants, immunotoxins, or detectable (e.g., fluorescent, luminescent, or radioactive) moieties. In some embodiments, an additional N-glycan can be added to the altered target molecule.

Methods for detecting glycosylation of a target molecule include DNA sequencer-assisted (DSA), fluorophore-assisted carbohydrate electrophoresis (FACE) or surface-enhanced laser desorption/ionization time-of-flight mass spectrometry (SELDI-TOF MS). For example, an analysis can utilize DSA-FACE in which, for example, glycoproteins are denatured followed by immobilization on, e.g., a membrane. The glycoproteins can then be reduced with a suitable reducing agent such as dithiothreitol (DTT) or β -mercaptoethanol. The sulfhydryl groups of the proteins can be carboxylated using an acid such as iodoacetic acid. Next, the N-glycans can be released from the protein using an enzyme such as N-glycosidase F. N-glycans, optionally, can be reconstituted and derivatized by reductive amination. The derivatized N-glycans can then be concentrated. Instrumentation suitable for N-glycan analysis includes, e.g., the ABI PRISM® 377 DNA sequencer (Applied Biosystems). Data analysis can be performed using, e.g., GENESCAN® 3.1 software (Applied Biosystems). Optionally, isolated mannoproteins can be further treated with one or more enzymes to confirm their N-glycan status. Additional methods of N-glycan analysis include, e.g., mass spectrometry (e.g., MALDI-TOF-MS), high-pressure liquid chromatography (HPLC) on normal phase, reversed phase and ion exchange chromatography (e.g., with pulsed amperometric detection when glycans are not labeled and with UV absorbance or fluorescence if glycans are appropriately labeled). See also Callewaert *et al.* (2001) *Glycobiology* 11(4):275-281 and Freire *et al.* (2006) *Bioconjug. Chem.* 17(2):559-564.

Cultures of Engineered Cells

This document also provides a substantially pure culture of any of the genetically engineered cells described herein. As used herein, a “substantially pure culture” of a genetically engineered cell is a culture of that cell in which less than about 40% (i.e., less than about : 35%; 30%; 25%; 20%; 15%; 10%; 5%; 2%; 1%; 0.5%; 0.25%; 0.1%; 0.01%;

0.001%; 0.0001%; or even less) of the total number of viable cells in the culture are viable cells other than the genetically engineered cell, e.g., bacterial, fungal (including yeast), mycoplasmal, or protozoan cells. The term "about" in this context means that the relevant percentage can be 15% percent of the specified percentage above or below the specified percentage. Thus, for example, about 20% can be 17% to 23%. Such a culture of genetically engineered cells includes the cells and a growth, storage, or transport medium. Media can be liquid, semi-solid (e.g., gelatinous media), or frozen. The culture includes the cells growing in the liquid or in/on the semi-solid medium or being stored or transported in a storage or transport medium, including a frozen storage or transport medium. The cultures are in a culture vessel or storage vessel or substrate (e.g., a culture dish, flask, or tube or a storage vial or tube).

The genetically engineered cells described herein can be stored, for example, as frozen cell suspensions, e.g., in buffer containing a cryoprotectant such as glycerol or sucrose, as lyophilized cells. Alternatively, they can be stored, for example, as dried cell preparations obtained, e.g., by fluidized bed drying or spray drying, or any other suitable drying method.

Metabolic Disorders

Molecules having uncapped terminal M6P residues can be used to treat a variety of metabolic disorders. A metabolic disorder is one that affects the production of energy within individual human (or animal) cells. Most metabolic disorders are genetic, though some can be "acquired" as a result of diet, toxins, infections, etc. Genetic metabolic disorders are also known as inborn errors of metabolism. In general, the genetic metabolic disorders are caused by genetic defects that result in missing or improperly constructed enzymes necessary for some step in the metabolic process of the cell. The largest classes of metabolic disorders are disorders of carbohydrate metabolism, disorders of amino acid metabolism, disorders of organic acid metabolism (organic acidurias), disorders of fatty acid oxidation and mitochondrial metabolism, disorders of porphyrin metabolism, disorders of purine or pyrimidine metabolism, disorders of steroid metabolism disorders of mitochondrial function, disorders of peroxisomal function, and lysosomal storage disorders (LSDs).

Examples of metabolic disorders that can be treated through the administration of one or more molecules having uncapped terminal M6P residues (or pharmaceutical compositions of the same) can include hereditary hemochromatosis, oculocutaneous albinism, protein C deficiency, type I hereditary angioedema, congenital sucrase-isomaltase deficiency, Crigler-Najjar type II, Laron syndrome, hereditary Myeloperoxidase, primary hypothyroidism, congenital long QT syndrome, tyroxine binding globulin deficiency, familial hypercholesterolemia, familial chylomicronemia, abeta-lipoproteinemia, low plasma lipoprotein A levels, hereditary emphysema with liver injury, congenital hypothyroidism, osteogenesis imperfecta, hereditary hypofibrinogenemia, alpha-1 antichymotrypsin deficiency, nephrogenic diabetes insipidus, neurohypophyseal diabetes insipidus, adenosine deaminase deficiency, Pelizaeus Merzbacher disease, von Willebrand disease type IIA, combined factors V and VIII deficiency, spondylo-epiphyseal dysplasia tarda, choroideremia, I cell disease, Batten disease, ataxia telangiectasias, ADPKD-autosomal dominant polycystic kidney disease, microvillus inclusion disease, tuberous sclerosis, oculocerebro-renal syndrome of Lowe, amyotrophic lateral sclerosis, myelodysplastic syndrome, Bare lymphocyte syndrome, Tangier disease, familial intrahepatic cholestasis, X-linked adrenoleukodystrophy, Scott syndrome, Hermansky-Pudlak syndrome types 1 and 2, Zellweger syndrome, rhizomelic chondrodysplasia puncta, autosomal recessive primary hyperoxaluria, Mohr Tranebjaerg syndrome, spinal and bullar muscular atrophy, primary ciliary dyskinesia (Kartagener's syndrome), gigantism and acromegaly, galactorrhea, Addison's disease, adrenal virilism, Cushing's syndrome, ketoacidosis, primary or secondary aldosteronism, Miller Dieker syndrome, lissencephaly, motor neuron disease, Usher's syndrome, Wiskott-Aldrich syndrome, Optiz syndrome, Huntington's disease, hereditary pancreatitis, anti-phospholipid syndrome, overlap connective tissue disease, Sjögren's syndrome, stiff-man syndrome, Brugada syndrome, congenital nephritic syndrome of the Finnish type, Dubin-Johnson syndrome, X-linked hypophosphatemia, Pendred syndrome, persistent hyperinsulinemic hypoglycemia of infancy, hereditary spherocytosis, aceruloplasminemia, infantile neuronal ceroid lipofuscinosis, pseudoachondroplasia and multiple epiphyseal, Stargardt-like macular dystrophy, X-linked Charcot-Marie-Tooth disease, autosomal dominant retinitis

pigmentosa, Wolcott-Rallison syndrome, Cushing's disease, limb-girdle muscular dystrophy, mucopolysaccharidosis type IV, hereditary familial amyloidosis of Finish, Anderson disease, sarcoma, chronic myelomonocytic leukemia, cardiomyopathy, faciogenital dysplasia, Torsion disease, Huntington and spinocerebellar ataxias, hereditary hyperhomocysteinemia, polyneuropathy, lower motor neuron disease, pigmented retinitis, seronegative polyarthritis, interstitial pulmonary fibrosis, Raynaud's phenomenon, Wegner's granulomatosis, preteinuria, CDG-Ia, CDG-Ib, CDG-Ic, CDG-Id, CDG-Ie, CDG-If, CDG-IIa, CDG-IIb, CDG-IIc, CDG-IId, Ehlers-Danlos syndrome, multiple exostoses, Griscelli syndrome (type 1 or type 2), or X-linked non-specific mental retardation. In addition, metabolic disorders can also include lysosomal storage disorders such as, but not limited to, Fabry disease, mucopolysaccharidosis I, Farber disease, Gaucher disease, GM₁-gangliosidosis, Tay-Sachs disease, Sandhoff disease, GM₂ activator disease, Krabbe disease, metachromatic leukodystrophy, Niemann-Pick disease (types A, B, and C), Scheie disease, Hunter disease, Sanfilippo disease, Morquio disease, Maroteaux-Lamy disease, hyaluronidase deficiency, aspartylglucosaminuria, fucosidosis, mannosidosis, Schindler disease, sialidosis type 1, Pompe disease, Pycnodysostosis, ceroid lipofuscinosis, cholesterol ester storage disease, Wolman disease, Multiple sulfatase deficiency, galactosialidosis, mucopolipidosis (types II, III, and IV), cystinosis, sialic acid storage disorder, chylomicron retention disease with Marinesco-Sjögren syndrome, Hermansky-Pudlak syndrome, Chediak-Higashi syndrome, Danon disease, or Geleophysic dysplasia.

Symptoms of a metabolic disorder are numerous and diverse and can include one or more of, e.g., anemia, fatigue, bruising easily, low blood platelets, liver enlargement, spleen enlargement, skeletal weakening, lung impairment, infections (e.g., chest infections or pneumonias), kidney impairment, progressive brain damage, seizures, extra thick meconium, coughing, wheezing, excess saliva or mucous production, shortness of breath, abdominal pain, occluded bowel or gut, fertility problems, polyps in the nose, clubbing of the finger/toe nails and skin, pain in the hands or feet, angiokeratoma, decreased perspiration, corneal and lenticular opacities, cataracts, mitral valve prolapse and/or regurgitation, cardiomegaly, temperature intolerance, difficulty walking, difficulty swallowing, progressive vision loss, progressive hearing loss, hypotonia, macroglossia,

areflexia, lower back pain, sleep apnea, orthopnea, somnolence, lordosis, or scoliosis. It is understood that due to the diverse nature of the defective or absent proteins and the resulting disease phenotypes (e.g., symptomatic presentation of a metabolic disorder), a given disorder will generally present only symptoms characteristic to that particular disorder. For example, a patient with Fabry disease can present a particular subset of the above-mentioned symptoms such as, but not limited to, temperature intolerance, corneal whirling, pain, skin rashes, nausea, or diarrhea. A patient with Gaucher syndrome can present with splenomegaly, cirrhosis, convulsions, hypertonia, apnea, osteoporosis, or skin discoloration.

In addition to the administration of one or more uncapped molecules described herein, a metabolic disorder can also be treated by proper nutrition and vitamins (e.g., cofactor therapy), physical therapy, and pain medications.

Depending on the specific nature of a given metabolic disorder, a patient can present these symptoms at any age. In many cases, symptoms can present in childhood or in early adulthood. For example, symptoms of Fabry disease can present at an early age, e.g., at 10 or 11 years of age.

As used herein, a subject “at risk of developing a metabolic disorder” is a subject that has a predisposition to develop a disorder, i.e., a genetic predisposition to develop metabolic disorder as a result of a mutation in an enzyme such as acid alpha glucosidase, alpha galactosidase, alpha-L-iduronidase, beta-D-galactosidase, beta-glucosidase, beta-hexosaminidase, beta-D-mannosidase, alpha-L-fucosidase, arylsulfatase B, arylsulfatase A, alpha-N-acetylgalactosaminidase, aspartylglucosaminidase, iduronate-2-sulfatase, alpha-glucosaminide-N-acetyltransferase, beta-D-glucuronidase, hyaluronidase, alpha-L-mannosidase, alpha-neuraminidase, phosphotransferase, acid lipase, acid ceramidase, sphingomyelinase, thioesterase, cathepsin K, or lipoprotein lipase. Clearly, subjects “at risk of developing a metabolic disorder” are not all the subjects within a species of interest.

A subject “suspected of having a disorder” is one having one or more symptoms of a metabolic disorder such as any of those described herein.

Pharmaceutical Compositions and Methods of Treatment

A target molecule having uncapped M6P residues can be incorporated into a pharmaceutical composition containing a therapeutically effective amount of the molecule and one or more adjuvants, excipients, carriers, and/or diluents. Acceptable diluents, carriers and excipients typically do not adversely affect a recipient's homeostasis (e.g., electrolyte balance). Acceptable carriers include biocompatible, inert or bioabsorbable salts, buffering agents, oligo- or polysaccharides, polymers, viscosity-improving agents, preservatives and the like. One exemplary carrier is physiologic saline (0.15 M NaCl, pH 7.0 to 7.4). Another exemplary carrier is 50 mM sodium phosphate, 100 mM sodium chloride. Further details on techniques for formulation and administration of pharmaceutical compositions can be found in, e.g., Remington's Pharmaceutical Sciences (Maack Publishing Co., Easton, Pa.). Supplementary active compounds can also be incorporated into the compositions.

Administration of a pharmaceutical composition containing molecules with uncapped M6P residues can be systemic or local. Pharmaceutical compositions can be formulated such that they are suitable for parenteral and/or non-parenteral administration. Specific administration modalities include subcutaneous, intravenous, intramuscular, intraperitoneal, transdermal, intrathecal, oral, rectal, buccal, topical, nasal, ophthalmic, intra-articular, intra-arterial, sub-arachnoid, bronchial, lymphatic, vaginal, and intra-uterine administration.

Administration can be by periodic injections of a bolus of the pharmaceutical composition or can be uninterrupted or continuous by intravenous or intraperitoneal administration from a reservoir which is external (e.g., an IV bag) or internal (e.g., a bioerodable implant, a bioartificial organ, or a colony of implanted altered N-glycosylation molecule production cells). See, e.g., U.S. Pat. Nos. 4,407,957, 5,798,113, and 5,800,828. Administration of a pharmaceutical composition can be achieved using suitable delivery means such as: a pump (see, e.g., *Annals of Pharmacotherapy*, 27:912 (1993); *Cancer*, 41:1270 (1993); *Cancer Research*, 44:1698 (1984); microencapsulation (see, e.g., U.S. Pat. Nos. 4,352,883; 4,353,888; and 5,084,350); continuous release polymer implants (see, e.g., Sabel, U.S. Pat. No. 4,883,666); macroencapsulation (see, e.g., U.S. Pat. Nos. 5,284,761, 5,158,881, 4,976,859 and 4,968,733 and published PCT patent applications WO92/19195, WO 95/05452); injection, either subcutaneously,

intravenously, intra-arterially, intramuscularly, or to other suitable site; or oral administration, in capsule, liquid, tablet, pill, or prolonged release formulation.

Examples of parenteral delivery systems include ethylene-vinyl acetate copolymer particles, osmotic pumps, implantable infusion systems, pump delivery, encapsulated cell delivery, liposomal delivery, needle-delivered injection, needle-less injection, nebulizer, aerosolizer, electroporation, and transdermal patch.

Formulations suitable for parenteral administration conveniently contain a sterile aqueous preparation of the altered N-glycosylation molecule, which preferably is isotonic with the blood of the recipient (*e.g.*, physiological saline solution). Formulations can be presented in unit-dose or multi-dose form.

Formulations suitable for oral administration can be presented as discrete units such as capsules, cachets, tablets, or lozenges, each containing a predetermined amount of the altered N-glycosylation molecule; or a suspension in an aqueous liquor or a non-aqueous liquid, such as a syrup, an elixir, an emulsion, or a draught.

A molecule having uncapped M6P residues suitable for topical administration can be administered to a mammal (*e.g.*, a human patient) as, *e.g.*, a cream, a spray, a foam, a gel, an ointment, a salve, or a dry rub. A dry rub can be rehydrated at the site of administration. Such molecules can also be infused directly into (*e.g.*, soaked into and dried) a bandage, gauze, or patch, which can then be applied topically. Such molecules can also be maintained in a semi-liquid, gelled, or fully-liquid state in a bandage, gauze, or patch for topical administration (*see, e.g.*, U.S. Patent No. 4,307,717).

Therapeutically effective amounts of a pharmaceutical composition can be administered to a subject in need thereof in a dosage regimen ascertainable by one of skill in the art. For example, a composition can be administered to the subject, *e.g.*, systemically at a dosage from 0.01 $\mu\text{g}/\text{kg}$ to 10,000 $\mu\text{g}/\text{kg}$ body weight of the subject, per dose. In another example, the dosage is from 1 $\mu\text{g}/\text{kg}$ to 100 $\mu\text{g}/\text{kg}$ body weight of the subject, per dose. In another example, the dosage is from 1 $\mu\text{g}/\text{kg}$ to 30 $\mu\text{g}/\text{kg}$ body weight of the subject, per dose, *e.g.*, from 3 $\mu\text{g}/\text{kg}$ to 10 $\mu\text{g}/\text{kg}$ body weight of the subject, per dose.

In order to optimize therapeutic efficacy, a molecule having uncapped M6P residues can be first administered at different dosing regimens. The unit dose and

regimen depend on factors that include, *e.g.*, the species of mammal, its immune status, the body weight of the mammal. Typically, levels of a such a molecule in a tissue can be monitored using appropriate screening assays as part of a clinical testing procedure, *e.g.*, to determine the efficacy of a given treatment regimen.

The frequency of dosing for a molecule having uncapped M6P residues is within the skills and clinical judgement of medical practitioners (*e.g.*, doctors or nurses). Typically, the administration regime is established by clinical trials which may establish optimal administration parameters. However, the practitioner may vary such administration regimes according to the subject's age, health, weight, sex and medical status. The frequency of dosing can be varied depending on whether the treatment is prophylactic or therapeutic.

Toxicity and therapeutic efficacy of such molecules or pharmaceutical compositions thereof can be determined by known pharmaceutical procedures in, for example, cell cultures or experimental animals. These procedures can be used, *e.g.*, for determining the LD50 (the dose lethal to 50% of the population) and the ED50 (the dose therapeutically effective in 50% of the population). The dose ratio between toxic and therapeutic effects is the therapeutic index and it can be expressed as the ratio LD50/ED50. Pharmaceutical compositions that exhibit high therapeutic indices are preferred. While pharmaceutical compositions that exhibit toxic side effects can be used, care should be taken to design a delivery system that targets such compounds to the site of affected tissue in order to minimize potential damage to normal cells (*e.g.*, non-target cells) and, thereby, reduce side effects.

The data obtained from the cell culture assays and animal studies can be used in formulating a range of dosage for use in appropriate subjects (*e.g.*, human patients). The dosage of such pharmaceutical compositions lies generally within a range of circulating concentrations that include the ED50 with little or no toxicity. The dosage may vary within this range depending upon the dosage form employed and the route of administration utilized. For a pharmaceutical composition used as described herein (*e.g.*, for treating a metabolic disorder in a subject), the therapeutically effective dose can be estimated initially from cell culture assays. A dose can be formulated in animal models to achieve a circulating plasma concentration range that includes the IC50 (*i.e.*, the

concentration of the pharmaceutical composition which achieves a half-maximal inhibition of symptoms) as determined in cell culture. Such information can be used to more accurately determine useful doses in humans. Levels in plasma can be measured, for example, by high performance liquid chromatography.

As defined herein, a “therapeutically effective amount” of a molecule having uncapped M6P residues is an amount of the molecule that is capable of producing a medically desirable result (e.g., amelioration of one or more symptoms of a metabolic disorder) in a treated subject. A therapeutically effective amount (i.e., an effective dosage) can include milligram or microgram amounts of the compound per kilogram of subject or sample weight (e.g., about 1 microgram per kilogram to about 500 milligrams per kilogram, about 100 micrograms per kilogram to about 5 milligrams per kilogram, or about 1 microgram per kilogram to about 50 micrograms per kilogram).

The subject can be any mammal, e.g., a human (e.g., a human patient) or a non-human primate (e.g., chimpanzee, baboon, or monkey), a mouse, a rat, a rabbit, a guinea pig, a gerbil, a hamster, a horse, a type of livestock (e.g., cow, pig, sheep, or goat), a dog, a cat, or a whale.

A molecule or pharmaceutical composition thereof described herein can be administered to a subject as a combination therapy with another treatment, e.g., a treatment for a metabolic disorder (e.g., a lysosomal storage disorder). For example, the combination therapy can include administering to the subject (e.g., a human patient) one or more additional agents that provide a therapeutic benefit to the subject who has, or is at risk of developing, (or suspected of having) a metabolic disorder (e.g., a lysosomal storage disorder). Thus, the compound or pharmaceutical composition and the one or more additional agents can be administered at the same time. Alternatively, the molecule can be administered first and the one or more additional agents administered second, or vice versa.

It will be appreciated that in instances where a previous therapy is particularly toxic (e.g., a treatment for a metabolic disorder with significant side-effect profiles), administration of a molecule described herein can be used to offset and/or lessen the amount of the previous therapy to a level sufficient to give the same or improved therapeutic benefit, but without the toxicity.

Any of the pharmaceutical compositions described herein can be included in a container, pack, or dispenser together with instructions for administration.

The following are examples of the practice of the invention. They are not to be construed as limiting the scope of the invention in any way.

EXAMPLES

EXAMPLE 1

Creation of a *Yarrowia lipolytica* strain with a higher degree of phosphorylated N-glycans

To upregulate the phosphorylation of glycans in *Y. lipolytica*, strain MTLY60 was transformed with 2 extra copies of the MNN4 gene, each in a separate expression vector. The MNN4 gene is involved in increasing glycan phosphorylation in yeast. FIG. 1 contains a schematic of the pYLTmAX plasmid into which the MNN4 gene was cloned to produce pYLTmAXMnn4, which contains the MNN4 open reading frame under control of the TEF promoter. A strain was made that contains two extra copies of the MNN4 gene, one under control of the hp4d promoter and one under control of the TEF1 promoter. N-glycans were prepared from strain MTLY60 Δ och1 (1 wild type copy of MNN4), strain MTLY60 Δ och1+Hp4dMNN4 (1WT +1 extra copy of MNN4) and strain MTLY60 Δ och1+Hp4dMNN4+TEFMNN4 (1WT +2 extra copies of Mnn4) and assayed by DNA sequencer-assisted (DSA), fluorophore-assisted carbohydrate electrophoresis (FACE). See, Callewaert *et al.*, *Glycobiology* 11(4):275-281 (2001). Based on the results in FIG. 2, it can be deduced that the mono phosphorylated peak is upregulated in the strain with 1 extra copy and that a peak of double phosphorylation appears. In the strain with 2 extra copies, the double phosphorylated peak was much higher and the peak of neutral Man₈GlcNAc₂ sugars was much lower.

EXAMPLE 2

Identification of a mannosidase activity that can uncap the capping mannose residue present on phosphorylated glycans of fungal origin.

The phosphorylation of sugars by yeast and filamentous fungi results in a mannose-phospho-mannose di-ester linkage (FIG. 3). To obtain a structure where the

phosphate is in a mono-ester linkage, a mannosidase is required that is able to hydrolyze the mannose-phosphate linkage, leaving the phosphate attached to the 6 position of a mannose of the high mannose glycan structure. Chiba *et al.*, *Glycobiology*, 12(12):821-8 (2002) indicate that a mannosidase from a *Cellulomonas* species is capable of decapping the mannose. However, Chiba *et al.* only partially purified the mannosidase protein and could not identify the gene encoding the protein.

A *Cellulosimicrobium cellulans* (also known as *Oerskovia xanthineolytica* and *Arthrobacter luteus*) isolate was obtained from the LMG bacteria collection and tested for production of mannosidase activity. The bacteria were grown at 30°C and in mannan containing medium to secrete the mannosidase in the medium. Bacterial supernatants (SN) were obtained from the cultures and tested for the desired mannosidase activity by incubating the SN with isolated N-glycans derived from the MNN4 overexpressing strain described in Example 1. After incubation, the glycans were assayed by DSA-FACE (FIG. 4).

After treatment with the SN, glycans gain an additional charge and migrate faster in the electric field and shift to the left hand side of the electroferogram. If these fast-running structures are indeed phosphomonoester-substituted high mannose glycans, they would be larger in size than the neutral products running at the same position. Treatment of such glycans with a phosphatase would result in neutral oligosaccharides that run much slower. As shown in FIG. 5, treatment with calf intestine phosphatase (CIP) resulted in the peaks displaying lower electrophoretic mobility, proving that the phosphates are terminal and that the mannose was decapped.

EXAMPLE 3

Partial purification and further identification of a mannosidase

To purify the mannosidase, *C. cellulans* was grown in 1L of medium B (Bagiyan *et al.*, *Eur. J. Biochem.* 249(1):286-92 (1997)) or medium A (Chiba *et al.*, 2002, *supra*). See Table 1. Thereafter, the medium was precipitated with 40% and 80% ammonium sulphate and the samples were analysed by SDS-PAGE. The ammonium sulphate fractions were dialyzed against 20mM Na-phosphate buffer pH 6.5 with 1 mM CaCl₂, and then tested for activity on oligosaccharides derived from a MNN4 overexpressing strain (Example 1).

TABLE 1
Medium components

Medium A (1 liter)	Medium B (1 liter)
2 g mannan	2 g mannan
0.5 g (NH ₄) ₂ SO ₄	2 g (NH ₄) ₂ SO ₄
0.4 g MgSO ₄ .7H ₂ O	0.02 g MgSO ₄ .7H ₂ O
20 mg FeSO ₄ .7H ₂ O	1 mg FeSO ₄ .7H ₂ O
60 mg CaCl ₂ .2H ₂ O	1 g yeast extract
1 g yeast extract	4.2 g KOH
7.54 g K ₂ HPO ₄	14 g KH ₂ PO ₄
2.32 g KH ₂ PO ₄	

Both cultivation conditions resulted in the production of the uncapping activity. Only the 40% ammonium sulphate fraction derived from medium B showed activity, whereas all fractions of the medium A supernatant displayed activity.

The 40% ammonium sulphate sample derived from the medium A cultivation was further purified over a silica-based gel filtration column (FIG. 6). This resulted in a peak with a shoulder around 670 kDa.

All elution fractions were incubated with oligosaccharides derived from a MNN4 overexpressing *Yarrowia lipolytica* strain (described in Example 1) (with or without a following CIP-digest) to test for the phosphate uncapping activity. The decapping and mannosidase activity was observed in all of the samples. Samples were also analyzed on SDS-PAGE (FIG. 7), which showed not just one protein band, but several protein bands. Several bands were cut out from the gel and portions of their sequence analyzed by de novo peptide sequencing using Mass Spectrometry.

The de novo sequencing results revealed several peptide sequences, which were compared against the sequences in the non redundant database using BLAST. Peptides with homology to the following proteins were identified: a phosphodiesterase, a hypothetical protein, a putative alpha-1,2 mannosidase (identified peptides shown in Table 2) (homology to a mannosidase from *Magnetospirillum*), and an aminopeptidase Y. The phosphodiesterase was a possible candidate, but here only 2 of the 6 peptides gave a hit. The mannosidase also was a candidate with 3/5 and 5/5 hits for 2 different mannosidases.

TABLE 2

Peptide Sequences

Peptide Sequence	SEQ ID NO
SAYQSFTTR	1
VWGFSHR	2
VEGGWLPR	3
TQGNNFALLPER	4
DVHAELTAMAR	5

EXAMPLE 4

Identification of mannosidases with the desired sequence
based on whole genome sequencing

To identify the mannosidase gene coding for the desired activity, the genome of *C. cellulans* was sequenced using a Titanium 454 sequencing (Eurofins MWG Operon). Due to the high GC content, the sequencing was only partial (1.96 Mbases) and of poor quality (with only a low average contig size). The high GC content of the genome that causes loop formation during the emulsion PCR (emPCR), resulting in deletions and very short sequences.

This problem was overcome using new sequencing chemistry for the emPCR that was made available in beta testing by Roche. This gave a much improved sequence (4.7 Mbases), allowing the identification of 5 mannosidase genes belonging to glycosyl hydrolase family 92, one of which (CcMan1, SEQ ID NO:6) corresponds to the sequence from which the peptides described in Example 3 were obtained. No mannosidases from family 38 or 47 were found. The start codon of each of CcMan1-CcMan4 was predicted by MetaGeneAnnotator (see the world wide web at metagene.cb.k.u-tokyo.ac.jp/metagene/) and compared to Blast results with known genes. The start codon of CcMan5 could not be predicted since it is missing from the sequence. The signal sequence of each gene was predicted with signal P (see the world wide web at cbs.dtu.dk/services/SignalP/) by two methods (neural networks and hidden markov models).

FIGs. 8-12 contain the nucleotide and encoded amino acid sequences of the 5 mannosidase genes from *C. cellulans*.

EXAMPLE 5

Heterologous expression of mannosidase for *in vitro* or *in vivo* mannose decapping.

In order to allow decapping of the yeast type phosphorylation by the mannosidase, it has to be expressed either heterologously in a different host or in the same fungal host from which the protein for therapeutic use is expressed. In the latter case it can be co-secreted or targeted to an intracellular compartment (e.g., Golgi apparatus or endoplasmic reticulum). This can be accomplished by cloning the gene (be it codon optimised for the target host or not) operably linked after a promoter in an expression vector. The mannosidase can be tagged with an epitope tag to allow easy detection and purification or expressed as such. It can be secreted in the periplasm of a bacterial cell or expressed intracellularly. In case of expression in the fungal host, the sequence can contain a secretion signal or a targeting signal to target the protein to an intracellular compartment or both. Table 3 contains a list of secretion and targeting signals for expression in fungal organisms. Examples of such expression vectors are presented in FIG. 13.

TABLE 3

Secretion and targeting signals for expression in fungal organisms

Secretion signals	Golgi targeting signal	
	N-terminal	C-terminal
<i>LIP2</i> prepro	<i>MNN2</i>	<i>KEX2</i>
<i>LIP2</i> pre	<i>MNN4</i>	
S.c. α mating factor	<i>MNN6</i>	
<i>XPR2</i> prepro	<i>MNN1</i>	
<i>XPR2</i> pre	<i>MNN9</i>	
	<i>OCH1</i>	
	<i>SEC12</i>	
	<i>KRE2</i>	

The CcMan1-Man5 genes were codon optimized for expression in *E. coli*. See FIGs. 14-18 for the codon optimized sequences. Table 4 contains the length of each

codon optimized nucleotide sequence and the predicted molecular weight of each polypeptide without the signal sequence.

TABLE 4
Codon Optimized Genes

	Length (bp)	SEQ ID NO	Size (kDa) of encoded product
CcMan1	2613	16	92.6
CcMan2	3483	17	121.6
CcMan3	3363	18	116
CcMan4	5283	19	184
CcMan5	4956	20	173

EXAMPLE 6

Cloning and Activity of *C. cellulans* Glycosyl Hydrolase (GH) Family 92 Enzymes

The CcMan1-CcMan5 codon optimized nucleic acids were cloned into *E. coli* vectors pLSH36, which contains a Spy signal sequence, and/or pLSAH36, which contains a DsbA signal sequence for periplasmic expression. Both pLSH36 and pLSAH36 result in the encoded polypeptide having a polyhistidine tag and a murine caspase-3 site, which can be used for the removal of the His6-tag during purification. FIG. 19 contains a schematic of the pLSH36 and pLSAH36 vectors as well as the cloning strategy for introducing the *C. cellulans* GH92 genes into the vectors. After cloning, the different mannosidases were transformed into *E. coli* BL21 + pICa2 expression strain. The transformed strains were grown to an optical density (OD) of 0.5 to 1 and induced with 1 mM isopropyl β -D-1-thiogalactopyranoside (IPTG). Different cell fractions (medium, periplasm, soluble and insoluble fraction) were isolated and analyzed by SDS PAGE and Western blotting with an anti-His6 antibody. For CcMan1, CcMan2, and CcMan3, expression was detected in all fractions. For CcMan4 and CcMan5, expression was the highest in the soluble fraction, but some expression also was detected in the other fractions.

To determine the activity of the CcMan1-CcMan5 proteins, activity tests were performed using methylumbelliferyl alpha mannoside (MUM) as set forth in Chiba *et al.*, 2002, *supra*. For CcMan1 and CcMan2, the medium and periplasm samples were able to hydrolyze MUM weakly, whereas CcMan3 and CcMan5 were not able to hydrolyze

MUM. The soluble fraction of CcMan4 gave the highest fluorescent signal, indicating that CcMan4 is the only mannosidase with α 1,2-mannosidase activity.

All medium and periplasmic samples of the 5 different *C. cellulans* mannosidases also were tested on sugars derived from the MNN4 overexpressing strain of Example 1 (referred to herein as MNN4 sugars) to see if they were able to degrade the sugars and uncap the mannose of the mannose-6-phosphate. The sugars were incubated overnight and analysed by DNA sequencer-assisted, fluorophore-assisted carbohydrate electrophoresis (DSA-FACE). The sugar profiles of the medium samples could not be analyzed due to fluorophoric molecules in the medium presentation resulting in irrelevant peaks in the electroferogram. The sugar profiles of the periplasm of CcMan1, CcMan2 and CcMan3 showed neither degradation nor decapping, CcMan4 showed degradation, and CcMan5 showed decapping activity (FIG. 20). A CIP-digest on the decapped sugars confirmed the decapping activity of CcMan5 as the dephosphorylated peaks moved to neutral Man8.

The active mannosidases CcMan4 and CcMan5 were aligned with Bt3990 (744 AA) and Bt2199 (739 AA), family 92 mannosidases with known structure (see Zhu *et al.*, *Nat. Chem. Biol.*, 6(2):125-32. Epub 2009 Dec 27 (2010)). See FIG. 21. Since only the first part of CcMan4 and CcMan5 aligned with Bt3990 and Bt2199, and because they are large proteins, it was decided to clone the first domain of each protein separately and test the activity. CcMan4domain (1-3357 bp, i.e., nucleotides 1-3357 of SEQ ID NO:20) and CcMan5domain (1-2322 bp, i.e., nucleotides 1-2322 of SEQ ID NO:20) were cloned into the pLSAH36 *E. coli* expression vector. See, FIG. 19 for a schematic of the pLSAH36 cloning vector. The expression vectors were transformed into the *E. coli* BL21 + pICa2 expression strain, which was grown to an OD of 0.5 to 1, and induced with 1 mM IPTG. Different cell fractions (medium, periplasm, soluble and insoluble fraction) were isolated and analyzed by SDS PAGE and Western blotting with an anti-His6 antibody. Expression was detected in all 4 cell fractions.

The activity of the domains was tested on Mnn4 sugars. Hereto, the periplasmic fraction of each of the CcMan4domain and CcMan5domain was incubated in the presence of Mnn4 sugars (FIG. 22) and analyzed by DSA-FACE. This experiment showed that the CcMan4domain lost its mannosidase activity since no degradation could

be detected (FIG. 22, panel 4). In contrast, the CcMan5 domain kept its uncapping activity (FIG. 22, panel 6).

EXAMPLE 7

Production and Purification of CcMan5 and its family 92 homologous domain

The recombinant CCman5 (nucleotides 1-4995 of SEQ ID NO:20 and CcMan5 domain (nucleotides 1-2322 of SEQ ID NO:20) were expressed in *E. coli* strain BL21codon + pICA2 that were transformed with the expression vectors pLSAHCcMan5 and pLSAHCcMan5domain. Expression was induced by IPTG under control of a λ pL-promotor (see WO 98/48025 and WO 04/074488). See Example 6 and FIG. 19 for a description of pLSAH. The transformed bacteria were grown in Luria Bertani (LB) medium supplemented with ampicillin (100 μ g/ml) and kanamycin (50 μ g/ml) overnight at 28 °C before 1/100 inoculation in a 20 liter fermenter provided with LB medium supplemented with ampicillin (100 μ g/ml) and 1 % glycerol. The initial stirring and airflow was 200 rpm and 1.5 l/min., respectively, and was automatically adapted to keep the pO₂ at 30 %. The temperature was kept at 28°C. The cells were grown to an optical density of $A_{600\text{ nm}} = 1.0$, transferred at 20°C, and expression was induced by addition of 1 mM IPTG overnight. Cells were then harvested and frozen at -20 °C. After thawing, the cells were gently resuspended at a concentration of 3 ml/g in 50 mM NaH₂PO₄ pH 8.0, 300 mM NaCl, 1 mM PMSF and 10 μ g/ml DNaseI. The periplasmic fraction was prepared by stirring the cell suspension for 1 h at 4 °C and was isolated by centrifugation at 18,000 x g for 30 min. All steps were conducted at 4 °C. The clear supernatant was applied to a 20 ml Ni-Sepharose 6 FF column (GE Healthcare), equilibrated with 20 mM NaH₂PO₄ pH 7.4, 300 mM NaCl, 20 mM imidazole, 0.1 % CHAPS. The column was eluted with 20 mM NaH₂PO₄ pH 7.4, 20 mM NaCl, 400 mM imidazole, 0.1 % CHAPS after an extra wash step with 50 mM of imidazole in the same buffer. The elution fraction was diluted 1/10 with 20 mM Tris pH 8.0, 0.1 % CHAPS and loaded on an 14 ml Source 15Q column (GE Healthcare) to remove contaminants. After equilibration, the protein of interest was eluted by a linear gradient over 10 column volumes of NaCl from 0 to 1 M in 20 mM Tris, 0.1 % CHAPS. The CcMan5 and CcMan5 domain containing fractions were further injected on a HiLoad 26/60 Superdex 200 prep grade with PBS as running

solution. The obtained fractions were analyzed by SDS-PAGE and western blotting with an anti-His6 antibody. Finally, the concentration was determined using the BCA assay (Pierce). The purified yield for the full-length CcMan5 protein was 5.7 mg, and for the CcMan5 family 92 domain, it was 110 mg from these 20 L fermentations, showing that the family 92 domain alone can be produced and purified in higher yield. The activity of the purified CcMan5 domain was tested on the Mnn4 isolated sugars as set forth in Example 6. A decapped sugar profile was obtained.

EXAMPLE 8

Structure of CcMan5domain

CcMan5₁₋₇₇₄ (residues 1 to 774 of SEQ ID NO:50, encoded by nucleotides 1-2322 of SEQ ID NO:20; corresponding to the mature protein after removal its natural leader sequence) was expressed in *E. coli* BL21 (DE3) periplasm as a fusion product starting with an N-terminal 6xHis tag followed by a 9 amino acid linker (VGPGSDEVD, SEQ ID NO:21) after the DsbA leader sequence. Cells were cultured in M9 medium containing 100 µg/ml of kanamycin and 100 µg/ml ampicillin at 28 C. At an OD₆₀₀ of 0.4, CcMan5₁₋₇₇₄ expression was induced by addition of 1mM IPTG and the culture was further grown overnight at 18°C. Cells from the overnight culture were harvested by centrifugation, washed and incubated for 20 min at 4°C with buffer containing 20 mM Tris/HCl pH 8.0, 20% sucrose, 5 mM EDTA, and 0.1 mg/ml lysozyme to make spheroplasts. Periplasmic proteins were isolated from spheroplasts by centrifugation at 20,000xg for 20 min.

CcMan5₁₋₇₇₄ was purified from the periplasmic extract by metal ion affinity chromatography (HisTrap HP, GE Healthcare, loading under a buffer containing 50 mM Tris-HCl pH 8.0, 150 mM NaCl, and eluted using an imidazole gradient up to 400 mM), ion exchange chromatography (HiTrap Q FF, GE Healthcare, buffer: 20 mM Tris-HCl pH 8.0, 40 mM NaCl and a NaCl gradient up to 1 M) and hydrophobic interaction chromatography (HiTrap Phenyl HP, GE Healthcare, loading buffer: 20 mM Tris-HCl pH 8.0, 10 mM NaCl, 1 M (NH₄)₂SO₄ and eluted using a (NH₄)₂SO₄ gradient up to 0 mM).

Purified CcMan5₁₋₇₇₄ was concentrated to 130 mg/ml in 10 mM Tris- HCl pH 8.0, 10mM NaCl and plate-like crystals (0.2x0.07x0.01 mm³) were grown by vapor diffusion using a crystallization solution containing 0.2 M Na fluoride, 0.1 M Bis-Tris propane pH

7.5 and 20% PEG 3350. Crystals were briefly transferred into a cryoprotecting solution containing the crystallization solution supplemented with 10%(v/v) glycerol and flash-cooled in liquid nitrogen. Single crystal diffraction data were collected at 100 K at the PXIII beamline at the Swiss Light Source (SLS, Villigen, Swiss) and beamline BM30A at the European Synchrotron Radiation Facility (ESRF, Grenoble, France). The structure of CcMan5₁₋₇₇₄ was solved using a KAuCl₄-soaked crystal for the calculation of experimental phases from a SAD experiment at 11.958 keV, corresponding to the Au L-III absorption edge. FIG. 33 contains the structural coordinates of the catalytic center. The CcMan5 model built from the experimental phases was refined by maximum likelihood methods against 2Å resolution data collected on a native crystal to a final R- and freeR-factor of 19.3 and 23.9 %, respectively. The final model contains 2 CcMan5₁₋₇₇₄ molecules per asymmetric unit (residues 8 to 771), comprising 11.513 protein atoms, 860 solvent atoms, 2 Ca²⁺ ions and 1 bis-tris-propane and glycerol molecule each.

Based on sequence similarity, CcMan5 falls within family 92 of glycosyl hydrolases (GH92), which are defined as exo-acting alpha-mannosidases. The X-ray structures for two GH92 family members with α 1, 2-mannosidase activity are available: Bt3990 and Bt2199 (PDB access codes 2WVX and 2WVY, respectively). The overall fold seen from the CcMan5₁₋₇₇₄ structure solved here, and deposited as PDB entry 2xsg, corresponds well with that seen in both Bt3990 and Bt2199, with r.m.s.d (root mean standard deviation) values of 1.99 Å and 2.12 Å over 624 and 621 matched C α atoms, respectively. CcMan5₁₋₇₇₄ consists of two domains, an N-terminal β -sandwich domain (residues 8 to 271) and a C-terminal ($\alpha\alpha$)₆ barrel domain (residues 291 to 771), connected via an α -helical linker (residues 272 to 290). The interface between both domains gives shape to a shallow cavity that harbors a conserved catalytic Ca²⁺ ion and gives shape to the -1 substrate binding site (nomenclature: Davies *et al.*, *Biochem. J.* 321:557-9 (1997)) and the catalytic center (FIGs. 23 and 24).

GH92 family glycosyl hydrolases are Ca²⁺-dependent alpha-mannosidases that catalyse glycosidic bond hydrolysis through a single displacement mechanism, leading to inversion of the anomeric configuration in the released mannose (Zhu *et al.*, 2010, *supra*). In CcMan5₁₋₇₇₄, the catalytic Ca²⁺ is octahedrally coordinated via the carbonyl oxygen of Asn 588, a carboxyl oxygen of Glu589 and Asp662 each, and three water

molecules (W1, W2, W3 – see FIG. 23) that lie in the equatorial coordination plane. An additional water molecule (W4) is present near the catalytic center, bound to the carboxyl groups of the conserved pair Asp 660 and Asp 662. The substrate binding cavity surrounding the catalytic Ca^{2+} is lined by the residues Asn 588, Gln 589, Thr 626, Thr 658, Asp 22, Asn 25, Gly 71, Gly 72, Phe 195, Tyr 196, Arg 405, Trp 354, Tyr 535, and Gln536 (FIG. 23).

CcMan5 sets itself apart from other alpha-mannosidases in the GH92 family because of its unique ability to accept mannose-alpha-1-phospho-6-mannose (Man-P-Man) as a substrate and a lack of alpha-1,2-, alpha-1,3-, alpha-1,4- or alpha-1,6-mannosidase activity. In order to obtain insight in the discriminating residues in the CcMan5 active site that give rise to this unique substrate specificity, Man-P-Man was modeled into the CcMan5₁₋₇₇₄ active site of molecule B of the asymmetric unit (FIG. 25). Positioning of the -1 mannose was based on the gross binding conformation observed in Bt3990 and guided by the positions of two water molecules (W2 and W3) and a glycerol molecule present in the apo active site. In this way, the O2, O3, O4 and O6 hydroxyl groups of the -1 mannose take equivalent positions to those observed for the water molecules W2, W3 and the O1 and O3 hydroxyl groups of the glycerol molecule, respectively. Thus, the mannose -1 O2 and O3 hydroxyl group position in the equatorial plane of the octahedral Ca^{2+} coordination sphere. O3 makes an additional hydrogen bond to the Asp 355 carboxyl group. The latter is further provides a H-bond to the O4 hydroxyl, which also comes within H-bonding distance of the Arg 405 guanidinium group. The O6 hydroxyl and O5 oxygen can be involved in H-bonding with the Gly 71 amide. For modeling, the -1 mannose was retained in its ground state chair conformation. As observed for Bt3990, positioning of the O2 hydroxyl group to come into idealized coordination with Ca^{2+} will lead to a distortion of the sugar ring to a half chair conformation (see FIG. 25). This is in line with the general acceptance that a distortion of the sugar ring during catalysis is required for the nucleophilic substitution at the acetal center in α -mannosides in order to break the 1,2-diaxial interaction of the incoming nucleophile with the O2 hydroxyl (Vocadlo *et al.*, *Curr. Opin. Chem. Biol.* 12:539-55 (2008)). The obtained model for substrate binding in the -1 site further shows that water molecule W4 lies in a good position to act as nucleophile for in line attack on the acetal

carbon. W4 is in H-bond interaction with the carboxyl groups of Asp 660 and Asp 662, which are conserved throughout GH92 enzymes and are proposed to form the base catalyst(s) for activation of the nucleophile. Therefore, the modeled substrate binding at the -1 site and the position of catalytic residues and nucleophile are consistent with the mechanistic requirements for nucleophilic substitution with inversion of the anomeric center in the released mannose. As discussed above, CcMan5 distinguishes itself by the ability to bind and hydrolyse Man-P-Man. The obtained model for Man-P-Man binding to the CcMan5 active site now provides a rationale for these observations. In known GH92 family members, the anomeric oxygen making the glycosidic bond, is in electrostatic interaction with the carboxyl group of a conserved glutamic acid residue (Glu 533 in Bt3990). The glutamic acid residue has been shown to serve as catalytic acid, stabilizing the transition intermediate by binding the anomeric oxygen and protonating the leaving group (Zhu *et. al.*, 2010, *supra*). In CcMan5, the equivalent residue to Bt3990 Glu 533 is mutated to glutamine, which is not able to serve as a proton donor and therefore explains the loss-of-function in CcMan5 for hydrolysis of mannosides. In Man-P-Man substrates, however, the phosphate bound to the anomeric oxygen constitutes a much stronger leaving group that would not require an acid catalyst to protonate the anomeric oxygen, explaining why enzymes like CcMan5 can retain catalytic activity for Man-P-Man substrates. Concomitant with substitution of the catalytic acid, the equivalent of Glu 585 in Bt3990 is replaced by Thr in CcMan5 (Thr 626). In Bt3990, Glu 585 interacts with Glu 533 and has been suggested to regulate the latter's pK_a and/or play a role in binding the leaving group in 2-linked mannosides (Zhu *et. al.*, 2010, *supra*).

It appears that mutation to non-acidic residues in the Gln 536 and Thr 626 pair alleviates part of the negative electrostatic potential in the binding site, thereby tolerating the phosphate linkage to the anomeric oxygen in Man-P-Man substrates. In CcMan5, the modeled phosphate binding site (P in FIG. 25) is shaped by Thr 626 and the amide of Gly 72, both of which appear able to donate a H-bond to the non-glycosidic oxygens in the phosphate.

Finally, based on the modeled binding of Man-p-Man in the CcMan5₁₋₇₇₄ active site, the reducing end mannose comes in the vicinity of two tyrosine residues, Tyr 535 and Tyr 196, suggesting the latter to residues form part of the +1 mannose binding site.

Both residues lay at the edge of a shallow cleft that could be involved in further interactions with glycans at the reducing end of the glycan tree.

EXAMPLE 9

Expression of α -GalactosidaseA in *Y. lipolytica*

A nucleic acid encoding human α -GalactosidaseA, without pre and pro sequence, was synthesized with codon optimization for *Y. lipolytica* and addition of a Myc-His tag. The obtained sequence was cloned in frame after the pre sequence of the lip2 gene. The nucleotide sequence of the codon optimized nucleotide sequence (SEQ ID NO:22) is set forth in FIG. 26A; amino acid sequence (SEQ ID NO:23) is presented in FIG. 26B.

Y. lipolytica MTLY60 with 2 extra copies of MNN4 and one copy of α -GalactosidaseA was induced in a larger culture to purify over a Ni-NTA column. Thus, they were grown in YTG and induced in oleic acid medium in 2 x 225 ml (2L shake flask) during 48 hours. The culture was centrifuged, followed by filtration of the medium over a 0.22 μ m filter. The filtered medium was desalted on a sephadex G25 XK50/100 column (GE Healthcare) to 20 mM NaH₂PO₄ pH 7.4, 0.5 M NaCl, 20 mM imidazole to remove non-protein disturbing contaminants before purification on Ni-sepharose 6 FF. The desalted protein fraction was loaded on a 4.3 ml Ni-sepharose 6 FF column (GE Healthcare), equilibrated with 20 mM NaH₂PO₄ pH 7.4, 0.5 M NaCl, 20 mM imidazole, washed with 50 mM imidazole in the same buffer and eluted with 20 mM NaH₂PO₄ pH 7.4, 20 mM NaCl, 400 mM imidazole. Samples 3-10 and 36-49 after the Ni-sepharose column were analysed on SDS-PAGE and Western blotting using an anti-His6 antibody. A protein band of around 50 kDa and of 65 kDa was present on coomassie in samples 40 and 41 was revealed by Coomassie blue staining of the SDS-PAGE gel. In the western blot, only a band of 50 kDa was detected and is most likely the α -GalactosidaseA. The estimated yield of the purified α -GalactosidaseA was 100-125 μ g/L culture medium.

The purified sample was used to determine the type of sugars on the recombinant α -GalactosidaseA. The sugars were removed in solution and afterwards labelled with APTS. After cleaning the sample by gel filtration, the sugars were analyzed on DSA-FACE. The expected sugars, the mono mannophosphorylated Man₈GlcNAc₂ peak (P)

and the double mannophosphorylated $\text{Man}_8\text{GlcNAc}_2$ peak (PP) were present as major peaks.

EXAMPLE 10

Expression of human alpha glucosidase in *Y. lipolytica*

Y. lipolytica strain OXY1589 was constructed that contained three copies of the human alpha glucosidase (also known as acid alpha glucosidase (GAA) or acid maltase EC3.2.1.3) and two copies of the *Y. lipolytica* MNN4 gene. The genotype of strain OXY1589 is as follows:

MatA, *leu2-958*, *ura3-302*, *xpr2-322*,
gut2-744, *ade2-844*

POX2-Lip2pre-huGAA:URA3Ex::zeta
POX2-Lip2pre-huGAA:LEU2Ex::zeta
POX2-Lip2pre-hGM-CSF:GUTEx::zeta
YIMNN4-POX2-hp4d-YLMNN4 :ADE2::PT targeted

All transformations were carried out according to well established protocols with modifications for the different selective markers. In all cases (unless otherwise specified), a huGAA integration fragment has been obtained by NotI restriction digestion in order to remove the kanamycin resistance gene from the expression plasmids. The resulting fragments were all separated by agarose gel electrophoresis followed by Qiagen column purification of the correct huGAA fragment. Strain OXY1589 was constructed by first cloning human GAA (huGAA) into a *Y. lipolytica* expression vector and constructing a *Y. lipolytica* MNN4 tandem expression vector. Three stable integrative transformations then were performed in order to obtain the final huGAA production strain OXY1589.

Y. lipolytica codon optimized huGAA expression vector: The nucleotide sequence encoding the 110 kDA human GAA (huGAA) precursor was chemically synthesized and codon optimized for *Y. lipolytica* expression. In the synthetic construct, the pre- and the pro- huGAA signal peptides were eliminated such that the protein starts at amino acid 57. The synthetic ORF of huGAA (FIG. 27A) is fused in frame at the 5' end to the 3' end of the *Y. lipolytica* LIP2 signal sequence (pre), followed by the coding sequence of two Xxx-Ala cleavage sites and flanked by BamHI and AvrII restriction sites

for cloning in expression vector. The construct is under the control of the inducible POX2 promoter. The complete amino acid sequence of the fusion construct is shown on FIG. 27B.

A general scheme of an expression vector is presented in FIG. 28. The bacterial moiety is derived from the plasmid pHSS6, and comprises a bacterial origin of replication (ori) and the kanamycin-resistant gene conferring resistance to kanamycin (KanR). The integration cassette comprises a) the selection marker for transformation to *Yarrowia lipolytica* (URA3; LEU2; GUT2), b) the expression cassette composed of a promoter, c) a multiple cloning site (MCS) to insert huGAA in frame with signal sequence and d) the terminator of the LIP2 gene. The integration cassette is flanked by zeta sequences for stable non-homologous integration into the *Y. lipolytica* genome. Two NotI restriction sites enable the isolation of the expression cassette before transformation. Plasmids pRAN034, pRAN036 and OXYP183 have been used to generate huGAA expression vectors pRAN058, pRAN059 and pRAN060, respectively, containing URA3, LEU2 and GUT2 transformation markers, respectively.

Tandem YIMNN4 expression vector: The YIMNN4 gene was cloned under control of the inducible pPOX2 promoter and the (semi)constitutive hp4d promoter. These two expression cassettes of YIMNN4 were subcloned in one vector as a tandem construct carrying flanking regions (PT) of the ADE2 gene for targeted integration into the ADE2 locus of the genome and the ADE2 gene as a selection marker.

Intermediate Strain OXYY1569: The first transformation was a co-transformation of the expression cassette purified from pRAN058 and pRAN059 vectors using URA3 and LEU2 marker to produce intermediate recombinant strain OXYY1569. OXYY1569 carries two expression constructs of huGAA under control of the pPOX2 promoter randomly integrated in the genome of strain G014.

OXYY1569 was selected as follows. PCR screening of genomic DNA was performed in order to confirm the integration of the foreign huGAA DNA into the genome of *Y. lipolytica*. Primers were designed to amplify a fragment of 2552bp from huGAA nucleotide sequence. Southern blot analysis of the genomic DNA also was performed in order to confirm the integration of at least 2 copies of huGAA DNA. In

particular, genomic DNA from OXY1569 clones were digested with Hind III and probed with huGAA DIG labeled specific probe.

In order to select a clone secreting high levels of huGAA, several randomly selected clones that were identified as positive in the PCR screening and Southern blot were grown in shake flasks under POX2 inducing conditions according to a standard procedure. In all cases, the culture supernatant was collected 72h post-induction and screened in a standard Western blot and enzyme activity assay analysis. N-Glycan analysis of OXY1569 indicated the predominant structure in OXY1569 is $\text{Man}_8\text{GlcNAc}_2$.

Intermediate Strain OXY1584: Recombinant strain OXY1569 was transformed in order to integrate two copies of the *Y. lipolytica* MNN4 gene into its genome to produce OXY1584. The transformation was performed with a SacII/XmaI derived expression cassette excised from plasmid OXP1479B. The expression cassette was designed for targeted integration into the ADE2 locus of *Y. lipolytica* genome. The recombinant strain was selected after Southern blotting and glycan analysis to evaluate the strain behavior with respect to the increased phosphorylation. Genomic DNA of several arbitrary chosen transformants was SpeI digested and probed with MNN4 specific DIG labeled probe. Correct targeted integration of MNN4 expression cassette into the ADE2 locus of *Y. lipolytica* genome should give 4207bp and 5683bp bands. Southern blot positive clones were grown in a standard shake flask procedure. N-glycan analysis of secreted proteins was performed in order to select the intermediate clone OXY1584. Compared to the parent strain OXY1569, the predominant structures after MNN4 over-expression are $\text{Man}_8\text{GlcNAc}_2(\text{PMan})_1$ and $\text{Man}_8\text{GlcNAc}_2(\text{PMan})_2$.

Production strain OXY1589: To generate the final prototrophic production strain OXY1589, a third copy of huGAA was integrated into the genome of recombinant OXY1584 strain. The transformation was performed with Not I excised expression cassette from pRAN069. Transformants were first screened by PCR on gDNA for presence of the additional copy of huGAA. To evaluate huGAA production arbitrary selected PCR positive clones were further analyzed for expression after a standard shake flask cultivation. The clone expressing the highest level of huGAA (OXY1589) was chosen after Western blot analysis and enzymatic activity assay. It also was reconfirmed

that the conversion levels of M8 to MP2-M8 and MP-M8 N-glycans was not influenced by the presence of the additional huGAA expression cassette.

EXAMPLE 11

Fed Batch Cultivation of Strain OXYY1589

To produce huGAA from strain OXYY1589 (Example 10), a fed batch process was established using a 10 L stirred tank, with a working volume of 6-8 liters. The process was divided in two phases:

- 1) Batch growth on glucose for biomass formation
- 2) Product formation by induction with help of a limited oleic acid feed.

Typically the batch phase was about 20 hours (h) and the production phase approximately 72 hours. At the end of the process, the culture broth was centrifuged and the supernatant was collected. The supernatant was used as starting material for the purification of the GAA (see Example 12).

The following parameters were controlled during the fermentation. Aeration was maintained at a constant value of 1.5 vvm air (volume per volume per minute). Dissolved oxygen (DO) was initially kept at 30%. The stirring was increased from 600 to 1200rpm depending on the DO levels. Once it reached the maximum of 1200 rpm, this speed was kept constant and the DO-setpoint was set to 10%. To maintain 10% DO, oxygen was spiked into the reactor with a maximal percentage of 50%. Foam evolution was controlled by a foam probe. In case of foam detection, antifoam was added to the bioreactor. The pH was controlled by adding 14% (v/v) ammonia (base) or 10% phosphoric acid to maintain a constant value of pH 6.8. The temperature was kept constant at 28°C throughout the whole process.

Biomass was monitored by measurement of optical density at 600 nm (OD600). The samples were diluted 2 – 1000 times in distilled water to obtain values in the linear range of the spectrophotometer. Product formation was detected by Western blot analysis and specific enzymatic activity tests.

EXAMPLE 12

Purification of recombinant huGAA (rhGAA)

The supernatant after cultivation (see Example 11) was clarified via depth filtration. The resulting material was then concentrated 20 times via TFF and diafiltered against 20 mM sodium phosphate pH 6 and 100 mM NaCl on a 10kDa MWCO membrane (Millipore).

Purification of rhGAA was start by adding ammonium sulphate up to a concentration of 1 M. After centrifugation, the supernatant was loaded on a Toyopearl-Phenyl 650M (Tosoh Biosciences) packed XK16/40 column. A linear gradient from 1 to 0 M ammonium sulphate was applied for elution. Those fractions that contain rhGAA were then pooled and subjected to a buffer exchange into 10 mM BIS-TRIS pH 6. Further purification was achieved via anion exchange chromatography on a source 30Q packed Tricorn 10/50 or XK25/20 column (GE Healthcare) using a linear salt gradient from 0 to 1 M NaCl. The resulting GAA-containing fractions were then concentrated before loading onto a final Hiload 16/60 superdex 200 gel filtration column (GE Healthcare) that was pre-equilibrated with 50 mM sodium phosphate pH 6 and 200 mM NaCl. Fractions were selected on the basis of specific activity and purity on Coomassie-stained SDS-PAGE gels and then combined and concentrated to a final concentration of 5-10 mg/ml. Protein concentration was done on 15 ml Amicon Ultra centrifugal devices (Millipore) with a MWCO of 10 kDa.

The reactions for the qualitative screening for rhGAA were started by adding the reaction buffer consisting of 0.35 mM 4-MUG, 0.1% BSA and 100 mM sodium acetate pH 4 in a 10:1 or 20:1 volume proportion to 10 or 5 µl of elution fraction. All reactions were done in 96-well flat-bottom microtiter plates. After an incubation period of 30 minutes to 1 hour at 37°C, an equal volume of 100 mM glycine pH11 was added to stop the reaction and the release of the fluorogenic reaction product 4-methylumbelliferone was observed under UV-light. Specific activities (units/mg protein) were determined using a colorimetric assay with the synthetic substrate p-nitrophenyl- α -D-glucopyranoside (PNPG) that measures the enzymatic release of the yellow coloured p-nitrophenolate reaction product. The reactions were started by mixing 10 µl of enzyme solution and 90 µl of substrate reaction buffer (2 mM PNPG in 150mM citrate-phosphate buffer pH4, 1% BSA) in reaction wells of a microtiterplate and were subsequently incubated at 37°C. After 1 to 2 hours an equal volume of stop buffer, 10% sodium

carbonate pH 12, was added to quench the reaction and bring the released p-nitrophenol (PNP) in its ionized state. Background-corrected absorbances and p-nitrophenolate standards were measured at a wavelength of 405 nm and specific activities were calculated. Protein concentrations were determined with the bicinchoninic acid (BCA) method. One unit was defined as the amount of enzyme that catalyzes the conversion of 1 nmol of PNPG to 1 nmol PNP and D-glucose per min at 37°C at a final substrate concentration of 2 mM in a citrate-phosphate buffer, pH 4.0.

EXAMPLE 13

Phosphate uncapping activity of heterologously expressed CcMan5 on glycoproteins expressed in a *Y. lipolytica* strain with a higher degree of phosphorylated N-glycans

The huGAA was expressed in *Y. lipolytica* strain OXY1589 to yield a glycoprotein with a high degree of phosphorylated N-glycan structures (see Example 10). The huGAA was purified as described in Example 12.

CcMan5 (1 and 5 μ l respectively at a concentration of 70 μ g/ml) was added to a solution of 4 μ g huGAA in 100 mM HEPES buffer pH 7.0 with 2 mM CaCl₂. The 20 μ l reaction mixture was incubated overnight at room temperature. The N-glycans were released with PNGaseF, labelled with APTS and subsequently analysed on DSA-FACE, essentially as described in Laroy W. *et al.*, *Nature Protocols*, 1: 397-405 (2006). The N-glycan profiles before and after CcMan5 treatment are shown in Figure 29. The N-glycan mixture released from purified huGAA is mainly composed of ManP-Man8GlcNAc₂ and (ManP)₂-Man8GlcNAc₂ (FIG. 29, panel B). A peak running slightly faster than ManP-Man8GlcNAc₂ can be assigned to ManP-Man7GlcNAc₂. Only very minor amounts of Man₈GlcNAc₂ and Man₇GlcNAc₂ are present. After incubation of huGAA with CcMan5 the conversion of ManP-Man8GlcNAc₂ and (ManP)₂-Man8GlcNAc₂ to P-Man8GlcNAc₂ and P₂-Man8GlcNAc₂ respectively is observed (FIG. 29, panel C and D). The peak in the electropherogram running between P-Man8GlcNAc₂ and P₂-Man8GlcNAc₂ corresponds to the partially uncapped bi-phosphorylated (ManP)₂-Man8GlcNAc₂ with a phosphodiester- and a phosphomonoester-linkage present ((MP)-M8-P in FIG. 29, panel

C and D). This product is further hydrolyzed to the fully uncapped P2-Man8GlcNAc₂ when using a higher concentration of CcMan5 or a longer incubation time.

The percentage of phosphorylated N-glycans versus neutral N-glycans was estimated from measuring the peak areas in the DSA-FACE electropherograms (FIG. 29). The figures related to the area under the curve are presented for the different N-glycans present on huGAA before (Panel B) and after CcMan5 treatment (Panel D). In huGAA (Panel B), (ManP)₂-Man8GlcNAc₂ (11597), ManP-Man6GlcNAc₂ (1261), ManP-Man7GlcNAc₂ (5901), ManP-Man8GlcNAc₂ (15576), Man6GlcNAc₂ (680), Man7GlcNAc₂ (1716), Man8GlcNAc₂ (1572) were present. Approximately 90 % of the N-glycans on recombinant huGAA were composed of mannose-phosphate containing structures.

After an overnight treatment of recombinant huGAA with CcMan5 (Panel D), P₂-Man8GlcNAc₂ (16182), (ManP)P-Man8GlcNAc₂ (1997), P-Man7GlcNAc₂ (8254), P-Man8GlcNAc₂ (17893), ManP-Man6GlcNAc₂ (500), ManP-Man7GlcNAc₂ (2495), ManP-Man8GlcNAc₂ (1326), Man6GlcNAc₂ (1097), Man7GlcNAc₂ (2143), Man8GlcNAc₂ (1599) were present. The N-glycans released from huGAA were composed of 83 % uncapped phosphorylated structures, 8 % is still mannose-phosphate capped and 9 % neutral N-glycans are present. The percentage of uncapped phosphorylated structures can be increased when using a higher concentration of CcMan5 or a longer incubation time.

EXAMPLE 14

Identification of Homologs Likely to Have Uncapping Activity

To identify other GH92 family members with similar predicted catalytic site topology and functionality, curated GH92 family members, as mined from the world wide web at cazy.org/GH92_all.html, were analyzed as were the top 500 hits obtained by Blastp search with the CcMan5 domain sequence on the Non Redundant Protein Sequences database at NCBI. Subsequently, these 392 sequences were used as the input for the multiple sequence alignment package MUSCLE (MULTiple Sequence Comparison by Log-Expectation), which also ranks the sequences in order of 'phylogenetic' distance (from closest related to furthest related).

Based on the curated GH92 family members from the Cazy database, MUSCLE alignment of all GH92 protein sequences (392) and the CcMan5 domain sequence identified the following as the closest homologs of CcMan5:

Streptomyces coelicolor CAA18915 (GenBank Accession No. NP_630514)

Clostridium spiroforme (GenBank Accession No. ZP_02866543)

Bacteroides thetaiotaomicron AAO78636 (GenBank Accession No. NP_812442)

Zunongwangia profunda ADF52306 (GenBank Accession No. YP_003584502)

Chitinophaga pinensis ACU58463 (GenBank Accession No. YP_003120664)

Their sequences and those of the next 5 closest homologs are aligned in FIG. 31.

Based on MUSCLE alignment of the 500 best scoring blastp protein hits versus the CcMan5 domain, the following were considered the closest homologs of CcMan5

Streptomyces coelicolor (GenBank Accession No. NP_630514)

Streptomyces lividans (GenBank Accession No. ZP_05522540)

Streptomyces lividans (GenBank Accession No. ZP_06527366)

Paenibacillus sp (GenBank Accession No. YP_003013376)

Bacteroides thetaiotaomicron (GenBank Accession No. NP_812442)

Bacteroides sp. (GenBank Accession No. ZP_04848482)

Bacteroides cellulosilyticus (GenBank Accession No. ZP_03677957)

Zunongwangia profunda (GenBank Accession No. YP_003584502)

Leeuwenhoekiella blandensis (GenBank Accession No. ZP_01061975)

Sphingobacterium spiritivorum (GenBank Accession No. ZP_07083984)

Chitinophaga pinensis (GenBank Accession No. YP_003120664)

Pedobacter sp. (GenBank Accession No. ZP_01885202)

Clostridium spiroforme (GenBank Accession No. ZP_02866543)

Alignment of these and the 5 next-best homologs can be found in FIG. 32. All 5 best hits from the annotated GH92 database are also found in these 13 best hits from the Blast search on the entire sequence database.

The top 5 hits in FIG. 31 and the top 13 hits in FIG. 32 uniquely share the following three motifs, which were shown in the crystal structure of Example 8 to be different from the alpha-1,2-mannosidase GH92 family members of which the structure was reported in Zhu *et. al.*, 2010, *supra*.

1) a glycine-rich motif GVGxxGxGG, with each X being G, S, T, V, A, C or Q (small side chains), numbering of crystal structure residues of CcMan5 domain: 69-77. This region makes a loop that provides essential hydrogen bonds to the -1 and phosphate-binding subsite in the active site of the enzyme.

2) a VRxE motif. The R makes a hydrogen bond to the -1 ring and possibly the +1 ring. E is in a salt bridge to this R residue, probably shaping this motif. x is W in the closest-related subfamily (top 3 homologs to CcMan5), or could be any of the 20 amino acids except P. This motif is found at residues 404-407 of SEQ ID NO:50.

3) a LYQGT motif, containing the Q which is an E in the mannosidases (proton donor), and which contains Y535, which is important for the +1 site formation. In some of the sequences, the L is A or Y and could reasonably be expected to also be I, V, A, F or M, and in some of them the T is N and can be expected to also tolerate S. Two *Caulobacter* sequences have an E instead of Q and would thus be predicted not to work on phosphorylated glycans.

4) a GDXGN motif. The D and N make part of the substrate binding cavity and may shape an alternative subpocket to bind the +1 mannose. X can be any amino acid other than P. This motif is found at residues 21-25 of SEQ ID NO:50.

Based on the above bioinformatics workflow and motif search based on the structure, it is thus possible to filter the GH92 sequences present in the non-redundant proteins sequence database (currently containing over 1220 sequences) for those rare family members that are good candidates for having the same substrate specificity to CcMan5, i.e., to be capable of uncapping Man-6-Pi-Man structures. In particular, the 3 sequences from *Streptomyces coelicolor* and *Streptomyces lividans* are similar to CcMan5, not only in the above motifs but also in many of the loops of the structure.

A search with Hidden Markov Models based on the sequence elements unique to CcMan5 and its closest homologs, reveals no further GH92 sequences which contain all of these elements, strongly indicating that no such GH92 members have obtained these elements through convergent evolution (these are the ones that would not be top-ranked in multiple-sequence alignments).

EXAMPLE 15

The presence of phosphate uncapping activity in GH92 glycosidases from *Bacteroides thetaiotaomicron*

An enzymatic analysis of 23 family GH92 α -mannosidases from *Bacteroides thetaiotaomicron* has been reported by Zhu, Y. *et al*, 2010, *supra*. Enzymes with α 1,2-, α 1,4-, α 1,3- or α 1,6-mannosidase activity are present in this group of enzymes, although some variants display very low activity. The three-dimensional structure of two α -1,2-mannosidases (Bt3990 and Bt2199) allowed to identify key amino-acid residues which seem to be a signature motif for α -1,2-mannosidase activity, i.e. His584-Glu585 and Trp99 in Bt3990. The activity on phosphorylated N-glycans (MNN4 sugars described in Example 1) of three GH92 enzymes from *B. thetaiotaomicron*, Bt3530 (Genbank nr AAO78636.1), Bt3965 (Genbank nr AAO79070.1) and Bt3994 (Genbank nr AAO79099.1) was tested. These enzymes display low α 1,4-mannosidase activity and lack the His-Glu and Pro-Trp motif.

Bt3530, Bt3965 and Bt3994 were expressed in *E.coli* and purified as described in Zhu *et al*, 2010, *supra*. Samples (1 μ l enzyme at a concentration of 0.1 mg/ml) were incubated with 7 μ l APTS-labeled MNN4 sugars dissolved in 10 mM HEPES buffer pH 7.0 with 2 mM CaCl₂ in an overnight assay at room temperature. A control assay with CcMan5 was included. To confirm the presence of a terminal phosphate the reaction mixture was incubated with CIP. An N-glycan preparation containing Man8GlcNAc₂ (M8) and the monophosphorylated ManP-Man8GlcNAc₂ (MP-M8) was used as substrate. No uncapping activity for Bt3530, Bt3965 and Bt3994 was detected under the above assay conditions. No shift in electrophoretic mobility of the peaks was observed compared to the CcMan5 control reaction (appearance of fast running P-M8 peak), followed by CIP treatment (disappearance of P-M8).

In an additional experiment, 1 μ l of enzyme, i.e. Bt3530 (0.1 mg/ml), Bt3965 (4.75 mg/ml) and Bt3994 (1.37 mg/ml) respectively, was incubated with MNN4 N-glycans at pH 7.0 (10 mM HEPES buffer pH 7.0 with 2 mM CaCl₂) and at pH 5.0 (10 mM Ammonium Acetate pH 5.0 with 2 mM CaCl₂) during 60 hours at room temperature. Very minor α 1,2-mannosidase activity was observed with Bt3530 at pH 7.0, as a small Man5GlcNAc₂ (M5) peak appears in the electropherogram. At pH 5.0, on

the other hand, no α 1,2-mannosidase activity is present, but a fast running peak at the left hand side of the electropherogram appears. This peak has the same electrophoretic mobility as P-Man8GlcNAc₂ (P-M8) and the terminal phosphate is hydrolyzed after incubation with CIP. CcMan5 (used at same concentration as Bt3530) is fully uncapping ManP-Man8GlcNAc₂ within 20 hours incubation at room temperature and at pH 7.0; therefore the observed activity of Bt3530 is rather low. After purification, the Bt3530 sample slowly precipitates when stored at 4 °C in 20 mM TRIS buffer, pH 8.0 with 300 mM NaCl. Therefore it is possible that instability of the Bt3530 protein influences the activity under the assay conditions used. Bt3965, which was used at a 40 times higher concentration, gave a similar result as Bt3530 at pH 7.0 (Panel G and H) and pH 5.0 (Panel I and J). No activity at all was observed with Bt3994 under the same reaction conditions (Panel K till N).

From these experiments can be concluded that phosphate uncapping activity is only a minor side activity of two of the three *B. thetaiotaomicron* GH 92 enzymes tested on MNN4 sugars.

OTHER EMBODIMENTS

While the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.

What is claimed is:

1. A method for uncapping a mannose-6-phosphate residue on an oligosaccharide, said method comprising
 - a) providing said oligosaccharide having a mannose-1-phospho-6-mannose linkage; and
 - b) contacting said oligosaccharide with a mannosidase capable of hydrolyzing said mannose-1-phospho-6-mannose linkage to phospho-6-mannose.
2. The method of claim 1, wherein said mannosidase comprises an amino acid sequence having at least 90% identity to the amino acid sequence set forth in residues 1 to 774 of SEQ ID NO:50 or to the amino acid sequence set forth in SEQ ID NO:50.
3. The method of claim 1, wherein said mannosidase comprises an amino acid sequence having at least 95% identity to the amino acid sequence set forth in residues 1 to 774 of SEQ ID NO:50 or to the amino acid sequence set forth in SEQ ID NO:50.
4. The method of claim 1, wherein said mannosidase comprises an amino acid sequence having at least 98% identity to the amino acid sequence set forth in residues 1 to 774 of SEQ ID NO:50 or to the amino acid sequence set forth in SEQ ID NO:50.
5. The method of claim 1, wherein for said mannosidase, the three dimensional protein coordinates of the atoms in the amino acid side chains located in the minimal catalytic center fall within 1.5 Å deviation of the coordinates of the equivalent atoms in FIG. 33.
6. The method of claim 1, wherein said mannosidase comprises an amino acid sequence having (i) a GVGXXGXGG motif, where X is Gly, Ala, Ser, Thr, or Cys; (ii) a VRXE motif, where X is any amino acid other than Pro; (iii) an X₁YQGX₂ motif, where X₁ is Leu, Ile, Val, Ala, Phe, Tyr or Met, and X₂ is Thr, Ser, or Asn; or (iv) GDXGN, where X can be any amino acid other than Pro.

7. The method of any one of claim 1-6, wherein said contacting step is performed using a purified mannosidase, a recombinant mannosidase, a cell lysate containing said recombinant mannosidase, or a fungal cell containing said recombinant mannosidase.
8. The method of any one of claims 1-7, wherein said oligosaccharide is attached to a protein.
9. The method of claim 7, wherein said protein is a human protein expressed in a fungal organism.
10. The method of claim 9, wherein said fungal organism is *Yarrowia lipolytica* or *Arxula adenivorans*.
11. The method of claim 9, wherein said fungal organism is a methylotrophic yeast.
12. The method of claim 11, wherein said methylotrophic yeast is *Pichia pastoris*, *Pichia methanolica*, *Oogataea minuta*, or *Hansenula polymorpha*.
13. The method of claim 9, wherein said fungal organism is a filamentous fungus.
14. The method of claim 13, wherein said filamentous fungus is selected from the group consisting of *Aspergillus caesiellus*, *Aspergillus candidus*, *Aspergillus carneus*, *Aspergillus clavatus*, *Aspergillus deflectus*, *Aspergillus flavus*, *Aspergillus fumigatus*, *Aspergillus glaucus*, *Aspergillus nidulans*, *Aspergillus niger*, *Aspergillus ochraceus*, *Aspergillus oryzae*, *Aspergillus parasiticus*, *Aspergillus penicilloides*, *Aspergillus restrictus*, *Aspergillus sojae*, *Aspergillus sydowi*, *Aspergillus tamari*, *Aspergillus terreus*, *Aspergillus ustus*, and *Aspergillus versicolor*.
15. The method of claim 8, wherein said protein is a pathogen protein, a lysosomal protein, a growth factor, a cytokine, a chemokine, an antibody or antigen-binding fragment thereof, or a fusion protein.

16. The method of claim 15, wherein said lysosomal protein is a lysosomal enzyme.
17. The method of claim 16, wherein said lysosomal enzyme is associated with a lysosomal storage disorder (LSD).
18. The method of claim 17, wherein said LSD is Fabry's disease, mucopolysaccharidosis I, Farber disease, Gaucher disease, GM1-gangliosidosis, Tay-Sachs disease, Sandhoff disease, GM2 activator disease, Krabbe disease, metachromatic leukodystrophy, Niemann-Pick disease, Scheie disease, Hunter disease, Sanfilippo disease, Morquio disease, Maroteaux-Lamy disease, hyaluronidase deficiency, aspartylglucosaminuria, fucosidosis, mannosidosis, Schindler disease, sialidosis type 1, Pompe disease, Pycnodysostosis, ceroid lipofuscinosis, cholesterol ester storage disease, Wolman disease, Multiple sulfatase deficiency, galactosialidosis, mucopolipidosis, cystinosis, sialic acid storage disorder, chylomicron retention disease with Marinesco-Sjögren syndrome, Hermansky-Pudlak syndrome, Chediak-Higashi syndrome, Danon disease, or Geleophysic dysplasia.
19. The method of claim 18, wherein said LSD is Pompe disease or Fabry's disease.
20. The method of any one of claims 1-6, wherein said mannosidase comprises a targeting sequence.
21. A method of producing a target protein having terminal phospho-6-mannose residues, said method comprising:
- providing a fungal cell genetically engineered to comprise a nucleic acid encoding a mannosidase, said mannosidase capable of hydrolyzing a mannose-1-phospho-6-mannose linkage to phospho-6-mannose; and
 - introducing into said cell a nucleic acid encoding a target protein, wherein said cell produces said target protein comprising said terminal phospho-6-mannose residues.

22. The method of claim 21, wherein for said mannosidase, the three dimensional protein coordinates of the atoms in the amino acid side chains located in the minimal catalytic center fall within 1.5 Å deviation of the coordinates of the equivalent atoms in FIG. 33.

23. The method of claim 21, wherein said mannosidase comprises a) an amino acid sequence having at least 90% sequence identity to amino acids 1 to 774 of SEQ ID NO:50 or to the amino acid sequence of SEQ ID NO:50; or (b) an amino acid sequence having (i) a GVGXXGXGG motif, where X is Gly, Ala, Ser, Thr, or Cys; (ii) a VRXE motif, where X is any amino acid other than Pro; (iii) an X₁YQGX₂ motif, where X₁ is Leu, Ile, Val, Ala, Phe, Tyr or Met, and X₂ is Thr, Ser, or Asn; or (iv) GDXGN, where X can be any amino acid other than Pro.

24. The method of any one of claims 21-23, wherein said fungal cell further comprises a nucleic acid encoding a polypeptide capable of promoting mannosyl phosphorylation.

25. The method of any one of claims 21-24, wherein said fungal cell is genetically engineered to be deficient in OCH1 activity.

26. The method of any one of claims 21-25, wherein said mannosidase is a *C. cellulans*, *Streptomyces coelicolor*, or *Streptomyces lividans* mannosidase.

27. A method of producing a target protein having terminal phospho-6-mannose residues in a fungal organism, said method comprising

a) providing a fungal cell genetically engineered to comprise a nucleic acid encoding a mannosidase capable of hydrolyzing a mannose-1-phospho-6-mannose linkage to phospho-6-mannose, and said fungal cell further comprising a nucleic acid encoding a target protein; and

b) isolating said target protein having said terminal phospho-6-mannose residues.

28. The method of claim 27, wherein for said mannosidase, the three dimensional protein coordinates of the atoms in the amino acid side chains located in the minimal catalytic center fall within 1.5 Å deviation of the coordinates of the equivalent atoms in FIG. 33.

29. The method of claim 27, wherein said mannosidase comprises (i) an amino acid sequence having at least 90% sequence identity to amino acids 1 to 774 of SEQ ID NO:50 or to the amino acid sequence set forth in SEQ ID NO:50; or (ii) an amino acid sequence having (i) a GVGXXGXGG motif, where X is Gly, Ala, Ser, Thr, or Cys; (ii) a VRXE motif, where X is any amino acid other than Pro; (iii) an X₁YQGX₂ motif, where X₁ is Leu, Ile, Val, Ala, Phe, Tyr or Met, and X₂ is Thr, Ser, or Asn; or (iv) GDXGN, where X can be any amino acid other than Pro.

30. The method of any one of claims 27-29, wherein said target protein and said mannosidase are co-secreted.

31. An isolated fungal cell genetically engineered to produce glycoproteins comprising terminal phospho-6-mannose residues, said fungal cell comprising a nucleic acid encoding a mannosidase, wherein expression of said mannosidase in said fungal cell produces glycoproteins comprising said terminal phospho-6-mannose residues.

32. The fungal cell of claim 31, wherein for said mannosidase, the three dimensional protein coordinates of the atoms in the amino acid side chains located in the minimal catalytic center fall within 1.5 Å deviation of the coordinates of the equivalent atoms in FIG. 33.

33. The fungal cell of claim 31, wherein said mannosidase comprises a) an amino acid sequence having at least 90% sequence identity to amino acids 1 to 774 of SEQ ID NO:50 or to the amino acid sequence of SEQ ID NO:50; or b) an amino acid sequence having (i) a GVGXXGXGG motif, where X is Gly, Ala, Ser, Thr, or Cys; (ii) a VRXE motif, where X is any amino acid other than Pro; (iii) an X₁YQGX₂ motif, where X₁ is

Leu, Ile, Val, Ala, Phe, Tyr or Met, and X₂ is Thr, Ser, or Asn; or (iv) GDXGN, where X can be any amino acid other than Pro.

34. The fungal cell of any one of claims 31-33, said fungal cell further comprising a nucleic acid encoding a polypeptide capable of promoting mannosyl phosphorylation.

35. The fungal cell of any one of claims 31-33, wherein said fungal cell is genetically engineered to be deficient in OCH1 activity.

36. The fungal cell of any one of claims 31-33, said fungal cell further comprising a nucleic acid encoding a polypeptide capable of promoting mannosyl phosphorylation, and wherein said fungal cell is genetically engineered to be deficient in OCH1 activity.

37. The fungal cell of any one of claims 31 to 36, said fungal cell further comprising a nucleic acid encoding a target protein, wherein said target protein is a glycoprotein.

38. The fungal cell of any one of claims 31 to 36, wherein said target protein is a human protein.

39. The fungal cell of claim 38, wherein said target protein is a pathogen protein, a lysosomal protein, a growth factor, a cytokine, a chemokine, an antibody or antigen-binding fragment thereof, or a fusion protein.

40. The fungal cell of claim 39, wherein said lysosomal protein is a lysosomal enzyme.

41. The fungal cell of claim 40, wherein said lysosomal enzyme is acid alpha glucosidase or alpha galactosidase.

42. The fungal cell of claim 38, wherein said target protein is a protein associated with a LSD.

43. The fungal cell of claim 42, wherein said LSD is Fabry's disease, mucopolysaccharidosis I, Farber disease, Gaucher disease, GM1-gangliosidosis, Tay-Sachs disease, Sandhoff disease, GM2 activator disease, Krabbe disease, metachromatic leukodystrophy, Niemann-Pick disease, Scheie disease, Hunter disease, Sanfilippo disease, Morquio disease, Maroteaux-Lamy disease, hyaluronidase deficiency, aspartylglucosaminuria, fucosidosis, mannosidosis, Schindler disease, sialidosis type 1, Pompe disease, Pycnodysostosis, ceroid lipofuscinosis, cholesterol ester storage disease, Wolman disease, Multiple sulfatase deficiency, galactosialidosis, mucopolipidosis, cystinosis, sialic acid storage disorder, chylomicron retention disease with Marinesco-Sjögren syndrome, Hermansky-Pudlak syndrome, Chediak-Higashi syndrome, Danon disease, or Geleophysic dysplasia.

44. The fungal cell of any one of claims 31 to 43, wherein said fungal cell is a *Yarrowia lipolytica* or *Arxula adenivorans* cell.

45. The fungal cell of claim 32, wherein said polypeptide capable of promoting mannosyl phosphorylation is a MNN4 polypeptide.

46. The fungal cell of claim 45, wherein said MNN4 polypeptide is a *Yarrowia lipolytica*, *S. cerevisiae*, *Ogataea minuta*, *Pichia pastoris*, or *C. albicans* polypeptide.

47. The fungal cell of claim 46, wherein said polypeptide capable of promoting mannosyl phosphorylation is a *P. pastoris* PNO1 polypeptide.

48. The fungal cell of any one of claims 31 to 47, wherein said mannosidase is a *C. cellulans*, *Streptomyces coelicolor*, or *Streptomyces lividans* mannosidase.

49. The fungal cell of any one of claims 31 to 47, wherein said mannosidase comprises a secretion signal.

50. The fungal cell of any one of claims 31 to 47, wherein said mannosidase comprises a targeting signal to target said mannosidase to an intracellular compartment.

51. The fungal cell of any one of claims 31 to 47, wherein said mannosidase comprises a secretion signal and a targeting signal to target said mannosidase to an intracellular compartment.

52. A substantially pure culture of *Yarrowia lipolytica*, *Pichia pastoris*, *Hansenula polymorpha*, *Arxula adenivorans*, *Pichia methanolica*, *Oogataea minuta*, or *Aspergillus niger* cells, a substantial number of which are genetically engineered to produce glycoproteins comprising a terminal phospho-6-mannose residue, said cells comprising a nucleic acid encoding a mannosidase capable of hydrolyzing a mannose-1-phospho-6-mannose linkage to phospho-6-mannose.

53. The culture of claim 52, wherein for said mannosidase, the three dimensional protein coordinates of the atoms in the amino acid side chains located in the minimal catalytic center fall within 1.5 Å deviation of the coordinates of the equivalent atoms in FIG. 33.

54. The culture of claim 52, wherein said mannosidase comprises a) an amino acid sequence having at least 90% sequence identity to amino acids 1 to 774 of SEQ ID NO:50 or to the amino acid sequence of SEQ ID NO:50; or b) an amino acid sequence having (i) a GVGXXGXGG motif, where X is Gly, Ala, Ser, Thr, or Cys; (ii) a VRXE motif, where X is any amino acid other than Pro; (iii) an X₁YQGX₂ motif, where X₁ is Leu, Ile, Val, Ala, Phe, Tyr or Met, and X₂ is Thr, Ser, or Asn; or (iv) GDXGN, where X can be any amino acid other than Pro.

55. The culture of any one of claims 52-54, wherein said cells further comprise a nucleic acid encoding a polypeptide capable of promoting mannosyl phosphorylation.

56. The culture of claim 55, wherein said cells are genetically engineered to be deficient in OCH1 activity.

57. An isolated glycoprotein comprising terminal phospho-6-mannose residues, wherein the protein is produced by the method of any one of claims 1-30.

58. A composition comprising a glycoprotein, wherein at least 47% of the N-glycans on said glycoprotein have terminal phospho-6-mannose residues.

59. The composition of claim 58, wherein at least 50% of the N-glycans on said glycoprotein have terminal phospho-6-mannose residues.

60. The composition of claim 58, wherein at least 75% of the N-glycans on said glycoprotein have terminal phospho-6-mannose residues.

61. The composition of claim 58, wherein at least 90% of the N-glycans on said glycoprotein have terminal phospho-6-mannose residues.

FIG. 1

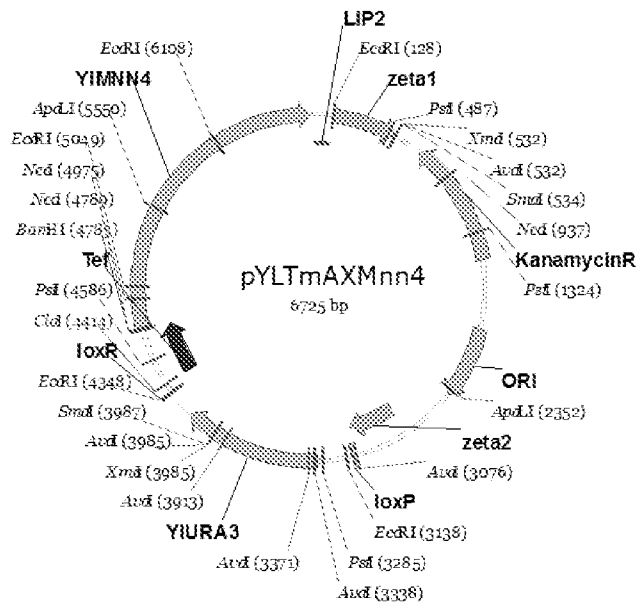
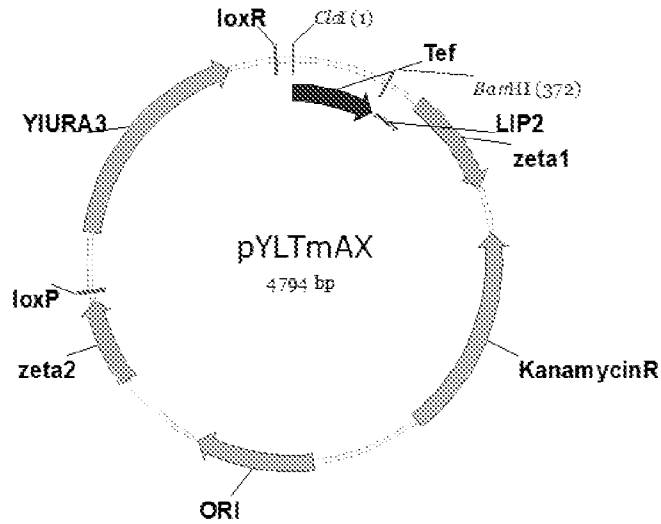


FIG. 2

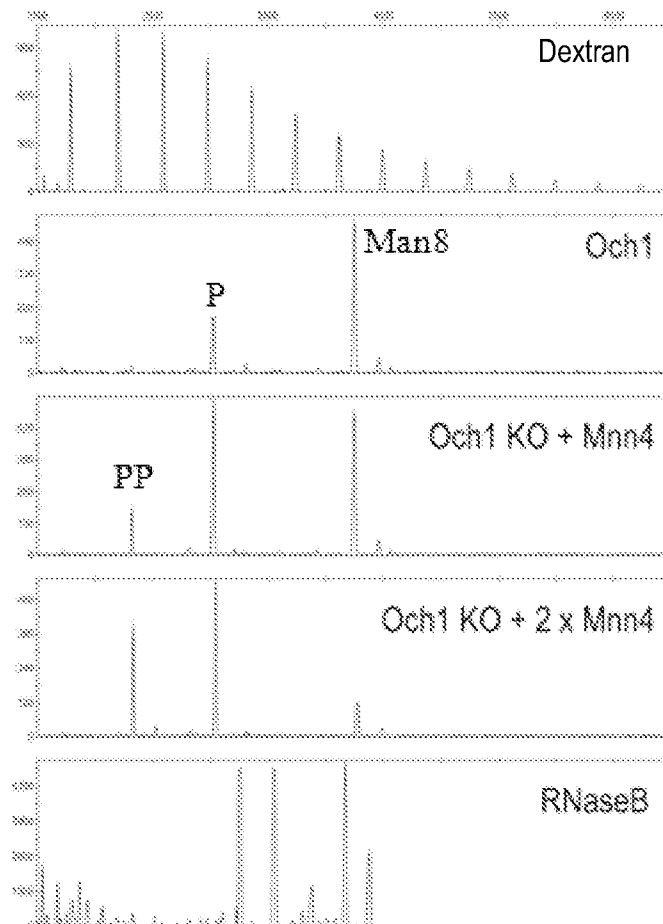


FIG. 3

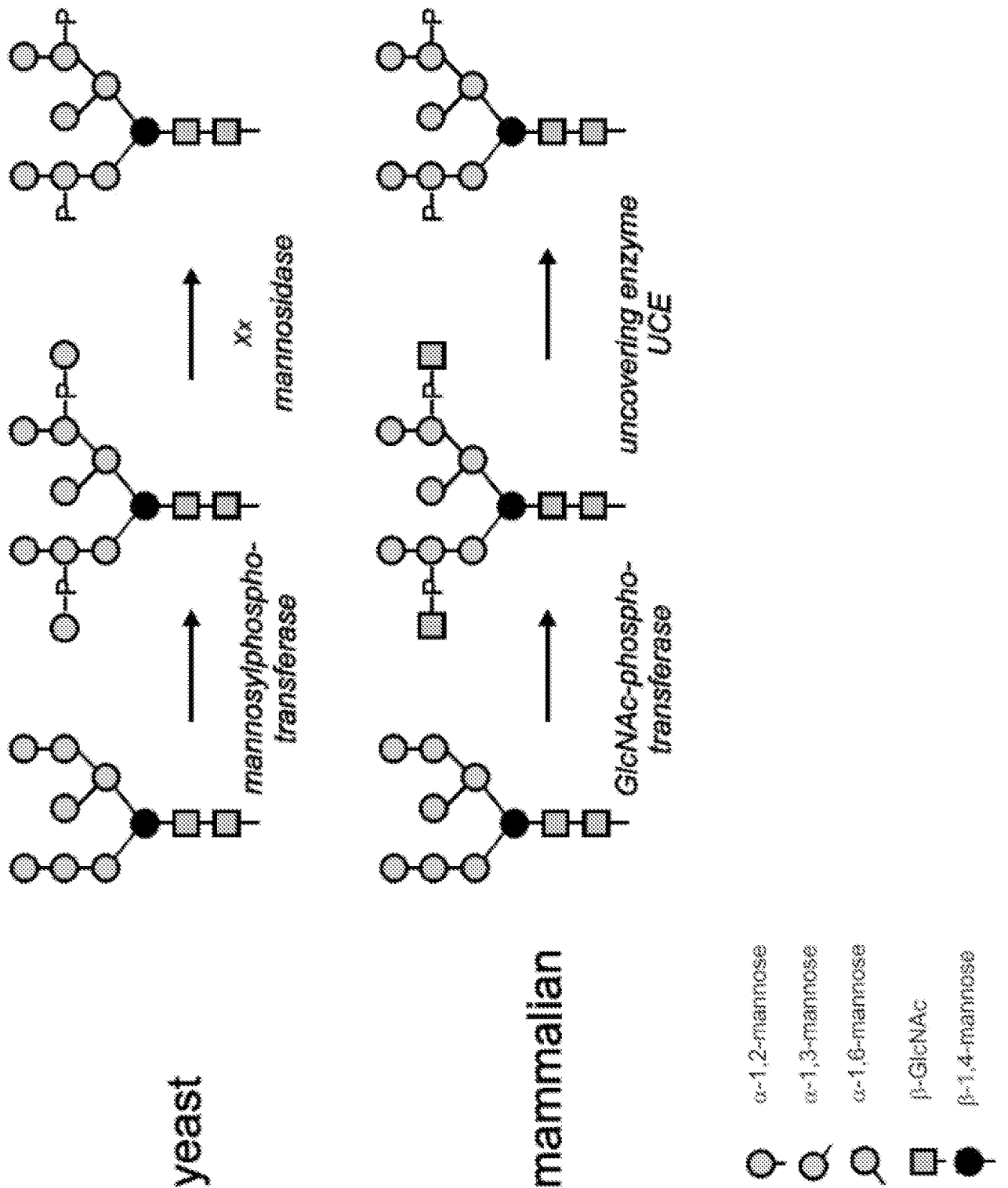


FIG. 4

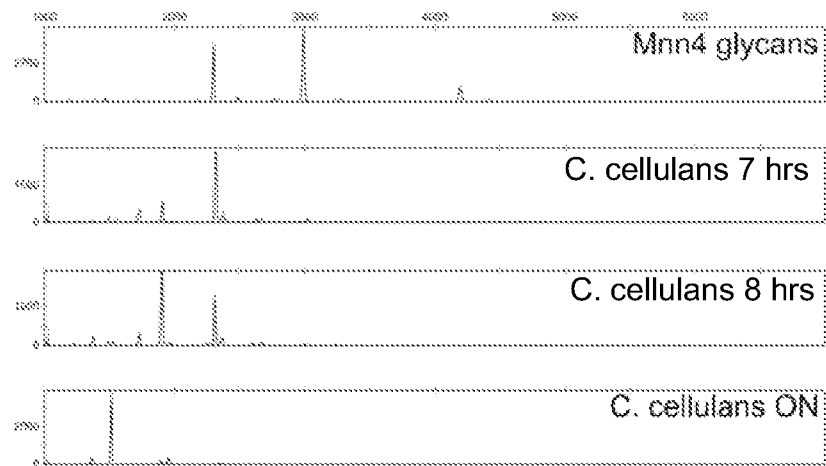


FIG. 5

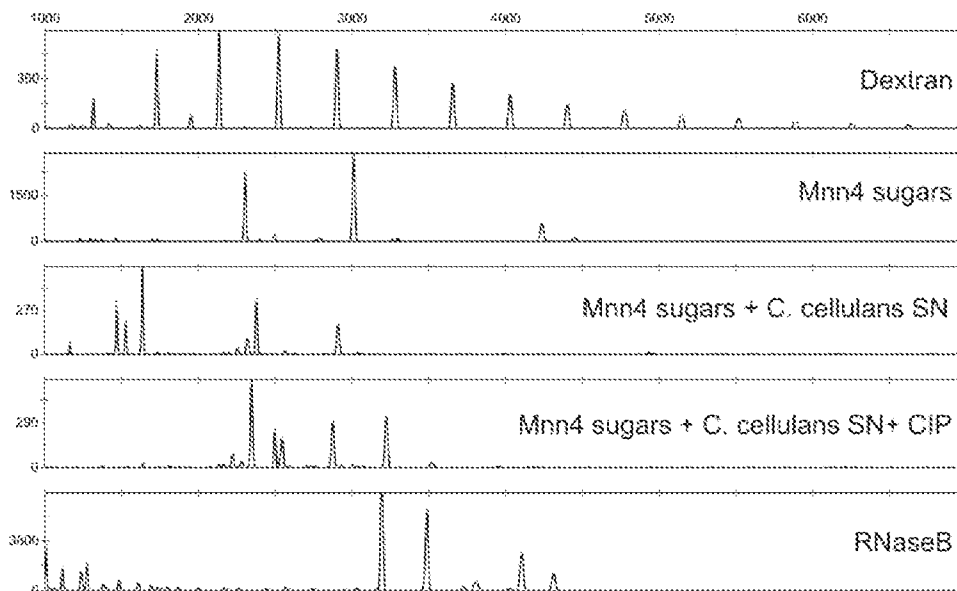


FIG. 6

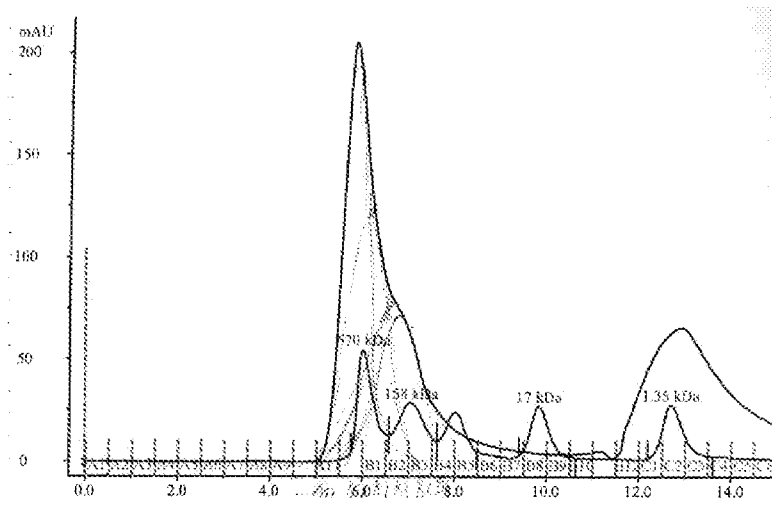


FIG. 7

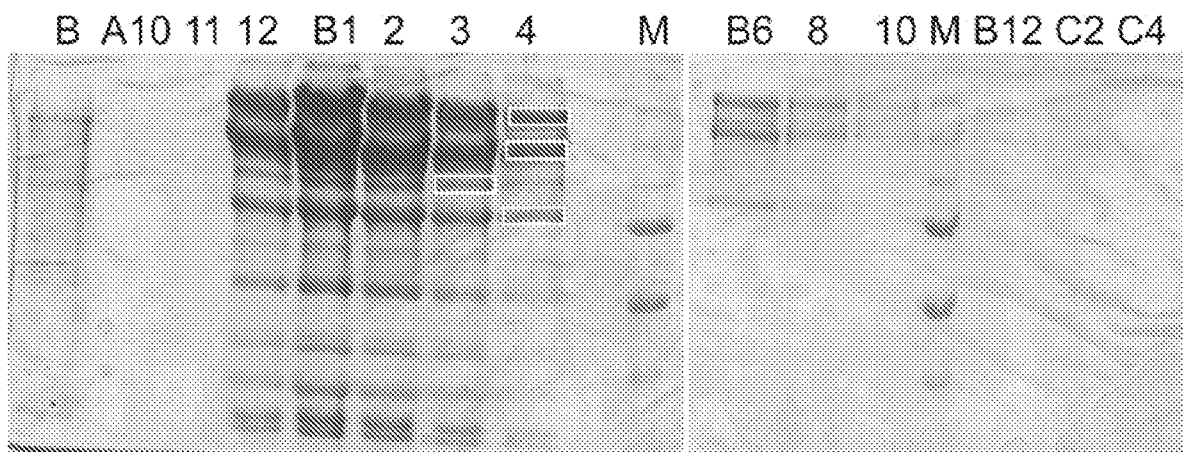


FIGURE 8A

ATGAGACGACCACGACTCGCCCTGCTCGCCGCGGGGCTCGCGCTCGCCGTGCGACCGGG
CACGCTGCTGCCCGTTCGCCGCGGGCGCCGCCCGCCGACGAGGGCACCCTCACCGCCG
CCGCGGGCGACGACCTCACGCTCGAGGTCAACCCGTTTCGTCGGCACCGAGAGCGAGGGC
AACGCCTACCCGGGCGCGACCGTGCCGTTTCGGCATGGTCCAGCTCAGCCCGGACAACAC
GAACTCCTACGCCTCGACGTGCTACAGCACGAACGCGGGGCGCGTGTGGGGCTTCAGCC
ACCGGCACGTGAACAGCGCGGGCTGCCCGCGGGCGGGCGAGCTGCTCGTCACGCCGAC
ACGAGCGCGACCCCGCGCACGTGCGGCTCCTTCATCGCCATCAAGGACCAGAAGAGCAC
CGAGCGCGCGTTCGGCCGGGTTCTACGAGGTGACCCTCGCGAACGACGTGCACGCCGAGC
TCACCGCGACCACGCGCGTTCGGCGCGCACCCGCTACACGTTCCCCGCCCTCGACGACGTG
CACCTGTGCTTCAACGTGGGCCAGACCCTGCGCGACGCCGGCGCGAGCTCGGTGACGTG
GGTCGACGACCGCACGCTCGAGGGCTGGGTGACAAACGGCGGGCTTCTGCGGCGGCACGC
CGGACAAGCAGCGGTACTTCTTCAGCGCGACGTTTCGACCGCCCGGTTCGCGTCGAGCGGC
ACGTGGGGGACCGATGCGCGCTACGTGCGGGCTCCACGACGAGCGAGGTTCGCGGGCGG
CAACAACGGCGCCGTTCGCGGTGTTTCGACACCACGACCGACCGCGACGTTCGAGGTGAGCG
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GACGAGGGCGGGCAGGTTCGCGTTCGACACCGTTCGCTGAGGAGGCCCGCGACGCGTGGAA
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CGCGGCATGGACCTCGAGGTCCACCAGGCCGACGGCTGGGACTACTACCAGAACTTCTC
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CGCGGAGAACTTCGCGCTCGGCACCGTCCCCGACGACATCGCGGACGAGCTGTGGGACT
ACCTCGTCGAGAACGCCACGACGACCCCGCCGGACGACGTTCGCGTCCGTTCGGGCGGCGC
AGCACCGAGTTCTACGCCGAGCACGGCCACGTGCCGTTCTACCCCGAGAACGAGGGCGG
CCTCGGCGGCCAGTTTCGAGGAGTACCGCCACGGCGGCTCGGCGACGCTCGAGCTCGCGC
TCGCCGACGCGAGCCTCGGCGCTGCGGCCGAGCGCACGGGTTCGCGAGGGCGGCCAGGCG
TTCTTCGACAAGGGTTCGCAACTGGCGCAACCTCTGGAACCCGGACGTTCGAGCTCTCGGG
TGGCTTCCAGGGCATGGTCAACGCGAAGCGCCCGACGGGCGAGTTTCGTCACGCTGCCCG
AGCTGACGGACGTTCACGCGCTCCGGCTTCACGAGGGCGTTCGGTGGCAGTACCAGTGG
ATGGTGCCGCGAGGACGTTCACGGGCTCCAGGAGGTTCATGGGCGGCGAGGACGGCTTCGT
CGAGCGTTCGACTACTACTTCGACCAGCCGGCGCTCGCCGCGAACCCCGGCGTCTCGC
CGAGCACGTGGGCCAAGGGCGGCAGCTCGTACTACACGACCATCCGCTACAACCCGGGC
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GAACGACGTCTTCGCCGCGAACCTCAACCGCTTCCCGGACACCCCGGGCGGCGGCGTTCG
GGAACGACGACCTCGGCACGCTTGCCGCTGGTACGTTCATGGCGTTCGCTCGGGTTCGAG
CCCGTTCATGCCGGGCTCGGGGATCCTCGCGCTCAACGCGCCGAAGGTGCAGGCCGCGAC
GCTCACGACCGATGCCGGGGCGACGCTGCGCATCGACGCGGGCGGCGGAACGAGAAGC
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GGGCTCACGTGGGGCACCGGCGCGGCCGACCGCATCCCGTTCGGTCTCCGCCGTTCGCC
CGCCCGCGCCGGTTCGAGGTTCGAGGCGAGCGCGCGTTCCTCGGCGGCCGGGCGTTCGTC
GCGGTCCGCGCGACCAGCACGGCCGACGCGCCGGTGGACGTGACTCTCACGACGCCGTT

CGGCGAGCGGACGGTCCGGCACGTGCAGCCGGGCAGGAGCGCCTACCAGTCGTTTCACGA
CGCGCACGACGTCCGTTCGAGGCCGGGACGGCGACCGTCACGGTCGTCGCCGCGGACGGC
ACGACGTCGACGGTCGACGCGGCGTACGAGGCGCTGGCCTGCGGC (SEQ ID NO: 6)

FIGURE 8B

MRRPRLALLAAGLALAVPGTLLPVAAGAAPADEGTVTAAAGDDLTLVNPFFVGTSEEG
NAYPGATVPFGMVQLSPDNTNSYASTSYSTNAGRVMWGFSHRHVNSAGCPAAGELLVTPD
TSATPRTSRSFIAIKDQKSTERASAGFYEVTLANDVHAELTATTRVGAHRYTFPASTTS
HLSFNVGQTLRDAGASSVTWVDDRTLEGWVDNNGGFCGGTPDKQRYFFSATFDRPVASSG
TWGTDARYVAGSTTSEVAGGNNGAVAVFDTTTTDRDVEVSVGVSVFVSVDGARANREAEAT
DEGGQVAFDFTVREEARDAWNAELGRAAIDASPDQRRIFYTQLYKTLLSPTIGSDVDGRY
RGMDLEVHQADGWDYYQNFSLWDTYRTQATLHALLPERAQDIVRSMYQHRVEGGWLPR
WSLGALETNIMAGDPVTPWLAENFALGTVPDDIADELWDYLVENATTPPDDVASVGRR
STEFYAEHGHVFPFYPENEGGLGGQFEEYRHGGSATLELALADASLGAAAERTGREGGQA
FLDKGRNWRNLWNPDELSSGGFQGMVNAKRPTGEFVTLPELTDVTRSGFHEGVPWQYQW
MVPQDVTGLQEVMGGEDGFVERLDYYFDQPALAANPGVSPSTWAKGGSSYYTTIRYNPG
NEPTIMNAWLYGYVGQPWKTNVLAANLNRFPDTPGGGVGNDDLGTAAWYVMASLGFE
PVMPSGILALNAPKVQAATLTTDAGATLRIDAAGANEKLPSYVAGLEVDGVAHTAAWL
DVAALQDGGTLDFDLSTAGLTWGTGAADRIPSVSAVAPPAPVEVEASARCLGGRAFV
AVRATSTADAPVDVTLTTPFGERTVRHVQPGRSAYQSFTTRTTSVEAGTATVTVVAADG
TTSTVDAAYEALACG (SEQ ID NO:7)

FIGURE 9A

GTGAGCCTCGCGCTCCCGCTGGCGGCGTACGCGGCGCCCGGGATCGGGGCGTCCGCCG
GACCGCCGCGGGACGGAGGCAGCGACGGGGTCCGATGCCGCGCCGTCGACGGCCCGC
TGGTTCGACTACGTCAACCCGTTTCATCGGGACCAAGGACGACGGCAACACCTACCCGGGC
GCTGCCGTGCCGTTCCGGCATGGTGCAACTCTCGCCGGACAACGGCCACAACGTCGGGTA
CGACTACGACCGCACGTCGGTGC GCGGGTTCTCGCTCGTGCACCTGTCCGGCGTCGGCT
GCGGCCTCGGCGGTCCGCTCCCGACCTGCCGACGACGGGCGCGATCACCTCGACCGAC
TACGGCCAGTACGCGCTCGGTTTCTCGCACGACGACGAGGAGGCCCTCGCCGGGGTACTA
CCGCGTGGGTCTCCAGGCGCCGGCGGGCACGATCGAGGCCGAGCTCACCGCGACCGAGC
GCACGGGCGTCCAGCGGTACACGTTCCCCGCGACGGCGCAGGCCAACGTCCTGCTCAAC
GCCGGCCAGGCGCTCAACCGGGTGACGGAGTCCGACGTGCGCGTCGTGGACGACCGCAC
GGTCGAGACGCGCATCACCGTCCGCGGGCTTCTGCCAGGACACCGAGCCGCGACGATCT
GGACCCGCACGACCTTCGACCGGCCGTTTCGTGCGCACGGCACGTGGGACGGCCAGGTC
GTCACCGCGGGCGCGGACGCCGCGTCCGGCGGGCAGGGCCGTCGCGGCGCGTACGTCAC
GTTTCGACACGACCGGCGGGCACCCTCGACGTGAGGCCGTCACCGCGATGAGCTACGTGG
GCGCCGACGGCGCCGCGGGCGAACCTCGCCGCGGAGGCCGGCACGTTTCGACGCCGTGCAC
GACGCCGCGCGCTCGGCCTGGGAGGAGCGGCTCGGCCTCGTGC GGGTTCGCGCAGGGCGA
CCCGGACGACCTGCGCACCTTCTACTCCTCGCTCTACCGCAGCTTCTTCGCGCCGAACG
TCGGCTCCGACGTGACGGGCGCTACCGCGGCTGGGACCAGGAGGTCCACGCCGCGGAA
CCGGACTTCACCTACTACCAGAACTACTCGCTCTGGGACACGTACCGCACCCAGCAGCA
GCTCCTGTACCTGCTCGCGCCCGACGAGTCCGGCCGACATGGCGCTCTCGCTCGTGCGCC
AGGGCCAGCAGGGCGGGTGGCTCCCGCGCTGGGGCTACGGCACGGTCGAGACGAACATC
ATGACCGGCGACCCGGCGACGCCGTTCTTCGTGAGCGCCTGGCGCCAGGGCTGCTCGC
GGGCCACGAGGAGGAGGCGTACGCGGTCCTGAGGGAGAACGCCGACGGCGTCCCGCCCG
CCGACTCGCCCTTCAACGGGCGCGCGGCGAACGTCGAGTACCTGCGCGACGGGTTTCGTC
CCGCACGAGCCGGCGCGCTCGGGCAAGCCCGGCGACTACGACCTCCAGCACGGCGCCTC
GGCGACCATGGAGTACGCCCTCGCCGACGCGATGCTCTCGACCATGGCGCGCGGCCTCG
GCCACGACGAGGACGCCGACCGGTACGCCGCCCGCGGCCAGAGCTACCGCAACGTGTTTC
GACCCGCGCACGGGCAACTTCCGGGCGCGTAACGCGGACGGCTTCTTCGTGGGCGACGC
GGACCCCGCGCACTCCGACGGGTTCCACGAGGGCACGGCGGTGCAGTACCAGTGGCTCG
TGCCCCAGGACGTGCCGGGCCTGTTTCGACCTCATGGGCGGCACCGACGCCGCGGTTCGAC
CGCCTCGATGCGTTCTTCGCGTACGACGAGCTCGTTCGCCGACCCCCGACGTCGCGAG
CGAGGTGTGGGTCAACGGCACGTACGACTACTACGGCTGGGAGACCTACAACCCGAACA
ACGAGCCCAACCTCCATGCGCCGTACGTCTACCTGTGGACCGGGCAGCCCTGGAAGACG
ACGGACGTGTCGTGCGCGCCGCGTTCGACCCCTCTTCACCGACGGCCCCGACGGCGTACGGG
CAACGACGACCTCGGCACGATGTCCGCGTGGCACGTGCTGTCGTGATCGGCCTGTACC
CGATCGTGC GGGCGCCGATCTGTGGGGCCTGACGACGCCGCTCTTCGACGACGTGACG
ATCACGCTCGACCCGGAGGTCTTCGGTTCGGGACTCCCTGCGCCTCACGGCGGACGGCGT
CGCGCCCGACACGCACTACACGCACTCCGTGTCGCTCGGCGGCGAGCCGCTCGATCGCG
CCTGGGTACGGGCGACGAGCTCACCGCGGCCGGCACGCTCGACGTGACCGTTCGGCACC
GAGCCGTCCGCGTGGGCGACCGACCCCGCGGCCTCGCCGGGCGCCGTCGTGCCTGCGGA
CGGCACGGTTCGAGCGCCTGTTTCGTTCGGCGCGACGCCGCGGCAGCCGTCCTCGCCCCG
GCGGGCGGACCGAGGTTCGAGTCCAGGTTCGTGCCCCAGGGCGCGGGGACGTCCAGCGGG
ACGCTCGAGGTGACGTCCGACGGCGCGGTACCGCGACGACCGACCTCGCCGAGTGGAC
CGCCGAGTCCGACGGCCTGCCGGCCACGGTTCGAGGGCACGGTGACGATCGAGGCTCCCG
CCGACGCCGAGCCGGGTCTGCACACGGTGC GGGCTCGTTCGTGCGCGACGCCGCGGGGACC

GAGGCGGTCCGCGAGGTCTCGGTCGTCGTGTCCGGGGAGTCGTGGATCGCCGACGCGTT
CGACAACGTCGGCATCGGCGACGCCGGGGCGGCCAACGCGAACCTCGACGGCTCGGGCG
CCTACCTCCTGCGCGACCTGCTCGCGGACCTCGGCGCCGTCCAGGGCCTGGAGCTCACC
GTGCCGGGCACGGACCTCACCTACACGCTCGGGGCCCCGCGGGCGGGCGCGCCCGACAA
CGTCGCCGCGAGCGGCGAGGTCTCGAGGTGCCCCGAGCACCTGCGCTCGGCCCGCCACC
TCTCGGTGGTCGGGACGAGCACGCACGGCACGCACGGGGGCGGCCTCGTCCTCGGGTTC
GCCGACGGCTCGTCGCAGACCGTCGACGTGCGCCTCAGCGACTGGTGCACGGGCTCGCC
CGAGCCCGGCAACATCACGGTCGCGAAGGCCGGGGCGCGCGGCGACCGCGAGAACGTGC
AGAAGATCGGCTGCGGCCTCTACGCCACCGCGCCCGTCGCGATCCCCGAGGGCAAGGTC
CTGACGTCGGTCACGCTGCCGAGCGACGAGCGGTTCCACGTGTTTCGCGATCGCGACCGA
CGCGACGGGGGACGTCCCCGCGCCGCAGGTCGAGGTCACGGCGCAGGCCCGCTGCCTCG
GCGGCAAGGCGTTTCGTGCGGGTGC GCGCGCTCAACACGGGCGAGCAGCCCGCCGCGATC
GAGCTCGCGACCCCGTACGGCTCCAAGCTCTTCGGTGACGTCGCTCCCGGGGCGAACGC
GTACCAGTCGTTTCGCCACCCGCGCCGCCCGTCGAGGCGGGCGAGGTCACGGTGACCG
TGACGACGCCCGACGGCGAGCCCCAGCAGGTCACGGCCGCGTACGACGCCCGCCGCTGC
TCC (SEQ ID NO:8)

FIGURE 9B

VSLALPLAAYAAPGIGASPATAAGTEAATGSDAAAVDGPLVDYVNPFIGTKDDGNTYPG
AAVPFGMVQLSPDNHNVGYDYDRTSVRGFSLVHLSGVGCGLGGPLPTLPTTGAITSTD
YGQYALGFSHDDEEASPGYYRVGLQAPAGTIEAELTATERTGVQRYTFPATAQANVLLN
AGQALNRVTESDVRVVDRTVETRITVRGFCQDTEPQTIWTRTTFDRPFVAHGTWDGQV
VTAGADAASGGEGRRGAYVTFDGTGGDLVEAVTAMS YVGADGAAANLAAEAGTFDAVH
DAARSAWEERLGLVRVAQGD PDDLRTFYSSLYRSFLAPNVGSDVDGRYRGWDQEVHAAE
PDFTYQNYSLWDTYRTQQQLLYLLAPDESADMALSLVRQGGGWLPRWGYGTVETNI
MTGDPATPFLVSAWRQGLLAGHEEEAYAVLRENADGVPPADSPFNGRAANVEYL RDGFV
PHEPARSGKPGDYDLQHGASATMEYALADAMLSTMARGLGHDEDADRYAARGQSYRNVF
DPRTGNFRARNADGFFVGDADPAHSDGFHEGTAVQYQWLVPQDVPGLFDLMGGTDAAVD
RLDAFFAYDELVADPPHVASEVWVNGTYDYYGWETYNPNNEPNLHAPYVYLWTGQPWKT
TDVVRAASTLFTDGPDGVTGNDDLGTMSAWHVLSSIGVYPIVPGADLWGLTTPLFDDVT
ITLDPEVFG RDSLRLTADGVAPDTHYTQSVSLGGEPLDRAWVTGDELTAAGTLDVTVGT
EPSAWATDPAASPGAVVPADGTVERL FVGATPRQPVLAPGGRTAVAVQVVAQGAGTSSG
TLEVTSDGAVTATTDLAEWTAESDGLPATVEGTVTIEAPADAEPGLHTVRLVVRDAAGT
EAVREVSVVVSGESWIADAFDNVGI GDAGAANANLDGSGAYLLRDL LADLGAVQGLELT
VPGTDLTYTLGAPRAGAPDNVAASGEVLEVPEHLRSARHLSVVGTS THGTHGGGLVLGF
ADGSSQTV DVRLSDWCTGSPEPGNITVAKAGARGDRENVQKIGCGLYATAPVAIPEGKV
LTSVTLPSDERFHVFAIATDATGDVPAPQVEVTAQARCLGGKAFVAVRALNTGEQPAAI
ELATPYGSKLFGDVAPGANAYQS FATRAAAVEAGEVTVTVTTPDGE PQQVTAAYDAAAC
S

FIGURE 10A

GTGCGGCGCTCCGTTCGCGGCGCTCTCTGCCACGGCGGTCTTGGCCCGCGGACTCTCGAT
CGCGCCCGCGTTCGGGCTCGCGGTCCCGGCGGTTCGCGGCCGCACCCGACCTCGTTGAGG
ACCCCGTCTCCTTCGTTCGACCCGTTTCGTTCGGGACCGGCCAGGCGACGGGCGTTCGTCGGG
GAGATCAACAACCTTCCCCGGGCCGTTCGATGCCGTTTCGGCATGATGCAGCTCTCGCCCGA
CACCCAGGTCTCCGTGGGCAACGGCGACAAGGCGTACGCGGGCTACCGCTACTCGCACC
AGGCGATCCGCGGCTTCTCCATGACGCACGCGGCCCGGGTGTGGATCTTCGGCGAC
GTCCCGATCCTCCCCGTGACGGGCGACGTTCGGGCGAGTACCCGTGGGACCGCAAGGAGGC
GTTTCAGCCACGACGCGGAGAGCGCCGAGGTTCGGCCGTTACGCGGTCACGCTCCAGTCGT
CGGGGATCGATGCGGAGGTGTTCGGCCGCGACCCGCTTCGGGCGGACTGACGTTTCGACTAC
CCCGAGGGCGGTTCGCGGTTCGACGAGTTCGTAACGCCGCGGGCTTCGCTTCGCGAGCGT
GCGCAACGCGACGGTTCGAGGTTCGAGGACGCGCGCACGGTTCACCGGCTTCGGTGACGAGCG
GCGGGTTCGCGGCAAGAACAACACGCACACGACGTAATTCGCGATCGAGCTTCGACCAG
GACGCGCAGGCGTTCGGCACGTGGCAGGGCTTCGACCGTCTCGCCCGGCGACCCGTCGGC
CGACGGCAACGCGCGGGCGCGTGGCTCACCTTCGCGCCCGGCGCGACGGTGCACGCGA
AGGTTCGGCATGTTCCTACGTGAGCGTTCGAGGGCGCGCGCCCAACCTTCGCGGCCGAGATC
CCGGGCTTCGACTTCGACGCCGTTCGGGACGCCAACCGCGCCGCTGGTCCGACCTGCT
CGGCAAGGTCCGCGTTCGCGGGGCGAGACGCCGACGACCTCACCATGTTCTACACGTCGC
TCTACCACTCGCTGCTGCACCCGAACACGTTTCACCGACGTGGACGGCCGGTACGTCGGG
TTCGACGGGGAGATCCACCAGGCCCGGAGGGGCGACGAGCGGTTCGCGAACTTCTCCGA
CTGGGACACGTACCGGTTCGCTTCGGCGCGTCCAGGCGTTCGTTGGCGCCCGACGAGGCGT
CGGACATGGCGCAGTCGCTTCGTCGAGGTCGCCGACGAGTCCGGTGGTTCGCGCGCTGG
CCCGTTCGCGAACCAGCACACGGGCCAGATGACCGGTGACTCCTTCGGTTCGCGCTCATCGC
GAGCATGTACGCGTTCGGGGCGCGCGACTTCGACGCGGAGTTCGGCGCTTCGCGCACATGG
TCAAGGGTTCGACGAGCGCCCGCCCGACCGCGAACGGTTCGTCGAGCGGCGCGGGATC
GAGACGTACCTTCGAGCGCGGTTCGCGCCCCAGACCGAGGAGTTCGGGGCGACCCCG
CGTTCGTCGGCGCGTTCGATCACGCTTCGAGTGGTTCGATTCGCCGACTTCGCGATTCGGG
TCGCGGCCGCGCTTCGGCCAGGACGACGTCGCCACCGAGTTCGCCCGCCCGCGGCCAGTGG
TGGCAGAACGTCCACGACCCCGTGACCCGCACGGCGGGCGCCCGGAACGACGACGGCAC
GTTTCGTTCGCGTTCGACGGGCGCGCGGGTTCGGGCGAGGAGGGTTCGACGAGGGCAACG
CCGAGCAGTACACGTTGGCTTCGTGCCGAGAACGTTCGCGGGGCTCACCGACGCGCTTCGGC
GGGCGTTCGAGGCCGTTCGCGGAGCGGCTTCGATGCCTTCACGGTTCAGCACAACGCCGGCCC
GAACGAGCCGTACCTGTGGATTCGGCAACGAGCCGAACTTCGGCGTCCCGTGGCTGTACG
ACTACGTGGGCCAGCCGTGGCGGACGAGCGAGCTTCGTTGGACGAGCTTACGTTCCACGCTG
TTCCGGCCCCGAGCCGAACGGCAAGCCCGGCAACGACGACCTTCGGCGCCCGAGGCCGGCTG
GTACGTGTGGGCCGCGATGGGCTGTACCCACCGCCGGGCGACGACGTCGTCGCGC
TCAACGCGCCGCGCTTCGACCGCGTTCGTTGGTTCGACCTTCGGCGAGGGCGACACCCCTGAC
CTGCGCGCCCCCGGCGCCTTCGACCGGCGCCCGTACATCAGCGGCGTTCACCATCGACGG
CGCAGCCTGGGACGGGACCTCCCTGCCGCGCCACGTCGCGCACGACGGCGGGCGTTCGTCG
AGCTTCGCGATGTCGACCGCACGCGACACGACGTTGGGGACCGCAGCCGAGGACGCCCCG
CCGTTCGTTGGCGCGACGGCGAGTCCGCCGTGGTTCGCCCGCGGACCCGGGCTTCGTCGAC
GGTTCGCCCCGGCGGGACCGCCGACGCGTTCGGTGGCCGTTCGAGCTCTTCGGGCGCCGACG
CCGCCGACGTCGCGGTTCGCGGTCGACGCGCCCGGGGGCATTCGGGGTTCGGTGAGCCCGCG
CTCGTTCGACGACGGCTTCGGGCCACCTCACCGGGACGGTCCCGTTCAGGTGGGTTCGCGG
CGTTCGCGTTCGGCTACACGACGCGCGCCTTCGTGCTTCGCGCCGGGGACGACGACGTCG
AGGTGCCCTCACCGTTCCTCGTTCGCCGCGCCCGGGTTCGCTTCGTTGCGGCCTACGACACG

GTCGGCACTGCGCCCGAGGCGAACC GCGGCGTTCGGGAACTTCGACGCGGCCGGCAACTC
GTTCTCGCGCGAGGCGCTCGCCGACGCGGGTCTCACGCCCGGGTTCGGCGCACGACGTTCG
ACGGCCTGGCGTTCACGTGGCCGTCCTCACCCGTGGGGCGCCCGGACTCGGTTCACGCTC
ACCGGCGAGACCGTGC GGCTCGACGCGCCGACGAGCCGGCTCGCGTTCGTGGGCGCCGC
GACCGACGGGACCCATCGCGGGACCGCGGTTCGTGACGTTTCGACGACGGCAGCACCCGCGA
CCACGACGATCGGCTTCGGCGACTGGGTGCTGCCGAGCGCGGACGGCTCGCCGGTTCGAG
GGCAACTCGGTTCGTTCGCGCAGATGAACCGGCGCAACGGCGACAAGGACAGCGCGTTCGT
GTTTCGCCACCGCCCCGTACACCGCGCCCGAGGACCGCCGCGTGGTTCGCGGTGAGGTTC
CCGACGTTCGACGACCTGCACGTCTTTGCGATCGCGACCGAGCCGGCCGCGGACGTGCAC
CTCGTGGACGTGACGGTCTCCCTGCGCTGCCTCGCCGGGACCCCGTACGTGGCGGTGCG
CGCGGCGAACGTCTCCGCCGGGGCCGTTCGACGTTCGACCTCACGACGGGCGTGGGCTCGC
GGTCCTTCACGGCCGTTCGCCCCGGCGCCAACGCCTACCAGTCGTTTCGCCGCCCGCGGC
GCGACCGGGAACGTTCGACGTTCACCGTTCACGGCCACGGGGGAGGAGGGGACGCAGACGGT
CGCGCGGACCGTTCGTTCGTCGCCGCTGCTCC (SEQ ID NO:10)

FIGURE 10B

VRRSVAALSATAVLAAGLSIAPAVGLAVPAVAAAPDLVEDPVSFVDPFVGTGQATGVVG
EINNFPGPSMPFGMMQLSPDTQVSVGNQDKAYAGYRYSHQAIRGFSMTHAAAGCWIFGD
VPILPVTGDVGQYPWDRKEAFSHDAESA EVGRYAVTLQSSGIDAEVSAATRSGGLTFDY
PEGGAASQVIVNAAGSLASVRNATVEVEDARTVTG SVTSGGFCGKNNHTTTYFAIELDQ
DAQAFGTWQGSTVSPGDPSADGNGAGAWLTFAPGATVHAKVGM SYVSVEGARANLAAEI
PGFDFDAVRDANRAAWS DLLGKVRVAGQDADDLTMFYTSLYHSL LHPNTFTD DVG RYVG
FDGEI HQAPEGHERYANFSDWDTYRSLGALQALLAPDQASDMAQSLVEVADQSGWLPRW
PVANQHTGQMTGDS SVPLIASMYAFGARDFDAESALAHMVKGATSAAPTANGYVQRRGI
ETYLERGYAPQTEEFRGDHRVVGASITLEWSIADFAIGQLAAALGQDDVATEYAARGQW
WQNVHDPVTRTAGARNDDGTFVRSQGGGGFGQEGFDEGNAEQYTWLVPQNVAGLTDALG
GREAVAERLDAFTVQHNAGPNEPYLWIGNEPNFGVPWLYDYVGQPWRTSELVDELSTL
FRPEPNGKPGNDDLGAQAGWYVWAAMGLYPTTPGTDVLALNAPRFDRVVVDLGEGLDLD
LRAPGASTGARYISGVTIDGAAWDGTS LPRHVAHDGGVVELAMSTARDTTWGTA AEDAP
PSWRDGESAVVAAADPGLVTVAPGGTADASVAVQLFGADAADVRVAVDAPGGIGVGEPA
LVDDGSGHLTGTVPVQVGAGVASGYHDARLVLSAGDDDVEVPLTVLVAAPGSLVAAAYDT
VGTAPEANRGVGNF DAAGNSFSREALADAGLTPGSAHDVDGLAFTWPSSPVGRPDSVTL
TGETVRLDAPTSRLAFVGAATDGTHRGTAVVTFDDGSTATTTIGFGDWLPSADGSPVE
GNSVVAQMNRRNGDKDSAFVFATAPYTAPEDRRVVAVRFPDVDDLHVFAIATEPAADVH
LVDVTVSLRCLAGTPYVAVRAANVSAGAVDVDLTTGVGSR SFTAVAPGANAYQSFAARG
ATGNVDVTVTATGEEGTQTVARTVVVPRCS (SEQ ID NO:11)

FIGURE 11A

ATGACCAGACCACTCCCGCCCGGACGCGCGGTTCGCGCGGTCCGGCAGCGGCCGCGCCCG
GCCCCTCGGCCTCGTGCTCGCCGCCCACTCGCCGTCCCGCTCGGGGTGCCTCTCGCGG
CCCCCGCGGGAGCCCTCGCTGCCGCGCCCGCCGCGGCCGCGGAGCCCGGCGACTTCTCG
TCCTCGTTCGAGTCCGGCGACCCGGCCGCGCTGCCACCACCGTGGCGGAGCGCGACGG
CGCGCCCTGGCAGGCGAACGTCGGCTCGTTCACGGCCGGCCTGCCCGGGAGCGTCTCG
GGCAGCTGAAGGGCGTCACGGCGAGCGCGCAGAACCTGCCCAACGAGGGCGCGGCGAAC
CTCGCCGACGGCAGCTCGGGACCAAGTGGCTCGCGTTCGCGTCGACCGGCTGGGTCCG
GTACGAGTTCGCCGAGCCCGTCTCGTTCGTTCGCGTACACGATGACCTCCGGCGACGACG
CCGCCGGTCGCGACCCGAAGACCTGGACGGTCGAGGGGTGCAACGACGGGTCCACGTGG
GCCGCGCTCGACCGCCGGACGGACGAGGACTTCCCGAACCCGACGACGCGCACGTT
CGAGCTCGAGGCGCCACC CGCGGCTACACGTACCTGCGCCTCAACGTACAGGCGAACT
CGGGCGACTCCATCGTCCAGCTCGCCGGGTGGGACCTCTCGGCCGACCTGAGCGCCGGC
CCGTCCGCGGCCCCCATGACGACGAAGGTCGGCACCGGGCCGCGCGTACGCTTACACAA
CAAGGCGGGCGTTCGGGTTCTCCGGCCTGCACTCGCTCCGGTACGACGGCTCGCACCTCG
CCGACGGCGAGACGTACGCGACGAACGTGCTCTACGACGACGTGGACGTCGTTCGTTCGGC
GAGGACACGCGCCTGAGCTACACGATCTTCCCGAGCTGCTCGACGATCTGCAGTACCC
GTCGACGTACGCGGCGGTGGACGTCCTGTTACCGACGGGACCTACCTGTCCGACCTCG
GCGCGCGGACGCGCACGAGACGGTCGCGACCCGCGCAGGCGCAGGGCGAGGGCAAGATC
CTCTACGCCGACCAGTGGAACTCGGTGCGGGTTCGACCTCGGCGACGTGCGCGAGGGCAA
GACCGTGGACCAGGTGCTGCTCGGGTACGACAACCCGGGCGGTACGCGGGGACGAAGT
TCGCGGGGCTGGCTCGACGACGTGAGATCACGGCGGAGCCGGCCACGATCGACGGGTTCG
AGCCTCGCCAACTACGTGGACACGCGCCGCGGCACGCTCGCGTTCGGGACGCTTCTCGCG
CGGGAACAACATCCCGCGACGGCGACGCCGAACGGGTTCAACTTCTGGACGCCGTACA
CGAACGCCTCCTCGCAGAGCTGGCTGTACGAGTACCACAAGGCCAACACGCCAACAAAC
AAGCCCGTCCCTCCAGGGCTTCGGGATCTCGCACGAGCCGAGCCCGTGGATGGGCGACCG
CAACCAGCTGACGTTCCCTCCCGTCGACGGCGTTCGGGGACGCCCGACGCCACGCTCTCGA
CGCGCGGCCTCGAGTTCGACCACGCGGACGAGACGGCGCGGCCGGACTACTACGGGGTC
ACGTTACCAACGGGTCCGCGATCGAGGCGACGCCACCAGCCACGGCGCGGTGCTCCG
CTTCAGCTACCCCGGAGCCAAGGGCCACGTGCTCGTGGACAAGGTGGACGGCTCCTCCA
AGCTCACGTACGACCAGGCCACGGGCACGATCTCCGGCTGGGTTCGAGAACGGCTCGGGC
CTGTCCGTGGGCCGCACGCGCATGTTTCGTTCGCGGCACCTTCGACCGTAGTCCGACGGC
GGTCGGGACGGCGGGCGGGCAACCGTTCGCGACGCGCGCTTCGCGACGTTTCGAGACGTCGT
CCGACAAGACGGTCGAGCTGCGCGTTCGCGACGTCGTTTCATCAGCCTCGACCAGGCGCGC
AAGAACCTCGACCTGGAGGTGACGGGCAAGACCTTCACGGAGGTCAAGGCCGCCGCCGCG
GCAGGCGTGGAACGACCGCCTGGGGGTTCATCGAGGTTCGAGGGCGCGAGCGAGGACCAGC
TCGTCACGCTGTACTCGAACCTTACC GCCTCAACCTGTACCCGAACCTCGCAGTTCGAG
AACACGGGCACGGCGCAGGAGCCGGTGTACAGGTACGCGAGCCCGGTCTCCGCGACCAC
GGGCTCCGCGACGGACACGCAGACCAACGCGAAGATCGTCGACGGCAAGATCTACGTGA
ACAACGGGTTCTGGGACACGTACCGCACGGCCTGGCCGGCGTACTCGCTCCTTACCCG
GAGCTCGCGGCCGAGCTGGTCGACGGGTTTCGTCCAGCAGTACC GCGACGGCGGGCTGGAT
CGCGCGCTGGTCTTCGCGGGGCTACGCCGACCTCATGACGGGCACGAGCTCCGACGTGG
CGTTCGCCGACGCGTACCTCAAGGGCTCGCTCCCCACGGGCACGGCGCTCGAGGCGTAC
GACGCCGCGCTGCGCAACGCGACCGTTCGCGCCCGGAGCAACGCCGTGGGCGCAAGGG
CCTGCAGACCTCGCCGTTCTTCGGGTTACGCGCGGAGTCCACGCACGAGTCCGTGTCGT

GGGGCCTGGAGGGCCTGGTCAACGACTTCGGCATCGGCAACATGGCCGCGCCCTCGCG
GAGGACCCGGCGACGCCGGAGGAGCGCCGCGAGACGCTGCGCGAGGAGTCCGCGTACTT
CCTCGAGCGGGCCACGCACTACGTTCGAGCTGTTTCGACCCCGAGGTTCGACTTCTTCGTGC
CGCGGCACGAGGACGGCACGTGGGCCGTCGACCCCGAGACGTACGACCCGGAGGCCTGG
GGCGGCGGGTACACCGAGACGAACGGCTGGAACCTTCGCGTTCACGCCCCGCAGGACGG
CCAGGGCCTCGCCAACCTCTACGGCGGCAAGCAGGGCCTCGAGGACAAGCTCGACGAGT
TCTTCTCCACGCCGGAGAAGGGCGCCGGCAACGGCGGCATCCACGAGCAGCGCGAGGCG
CGCGACGTCCGCATGGGCCAGTGGGGCATGAGCAACCAGGTGTCGCACCACATCCCGTG
GCTCTACGACGCCGCGGGCGCGCCGTCGAAGGCGCAGGAGAAGGTCCGCGAGGTACCC
GCCGCTGTTTCGTGGGCAGCGAGATCGGCCAGGGCTACCCGGGCGACGAGGACAACGGC
GAGATGTCGTCGTGGTGGATCTTCGCCTCGCTCGGCTTCTACCCGCTCCAGGTCCGGCTC
GGACCAGTACGCGGTTCGGTTCGCCGCTGTTTCGACAAGGCGACCGTGCACCTGCCGGACG
GCGACCTCGTCGTC AACGCCGAGAACA ACTCGGTTCGACAACGTCTACGTGCAGTCCCTC
GCGGTGGACGGCGAGGCCCGCACCTCGACGTCACTCTCCAGGCGGACCTCTCGGGCGG
CACGACTCTGGACTTCGTTCATGGGTCCGGAGCCGTTCGACTGGGGCACGGGCGAGGACG
ACGCGCCCGCGTTCGCTCACCGAGGGCGACGAGCCCCGACGCCGGTGCAGGACGCGACG
ACCGCGGGCCTCGGCACCACCACCGTTCGCCGACGGCGACGCCACCACGAGCGCCGCGGC
GCTCACGGACAACACGTCCGGGACGCGCACGACGTTTCGCCACCACGACGCCGTTCGATCA
CGTGGGCGGGCAACGGCATCCGCCCGACCGTTCGGGTTCGTACACGCTGACCTCCGGGGCG
AGCGGGACGGCGTTCACCGTCCGCATGGACTCTCGAGGGTTCGGACGACGGCGAGACGTG
GACGACGCTCGACGAGCGGTCCGGCGAGCAGTTCGGCTGGGCCCTGCAGACGCGGCCGT
TCACGGTTCGCGGAGCCGACGGCGTTCGCGCGGTACCGGGTTCACGGTTCACCGCGACGTCG
GGCTCCGGCGCGCTGTCGCTCGCCGAGGTTCGAGCTCCTCGCCGACCCGAAGGAGTTCGGG
GGCCGAGGAGTTCACCTCTCGGCCGCGCCGGACCGTTCGCGGCGTTCACGGGCGCGAGG
TCTCGGGCTCGTTCGCGACCTCACCGGGGTTCGAGGGCGACGTCGCGGGCGCTTCGACGTG
CAGGTTCGCGTTCGGCGACGGCTCCGAGCCGGTTCGCCGGGACGCTGCGGGCGGGCGCGTT
CGGCGGGTACGCGGTGGACGCCGCGCACACGTGGACCGCACCCGGCGTTCACCCCGTGA
CCGTTCACGGTTCGGGCGAGGGGATTCGAGACCGTTCGCGCCTCCTCGTACGTTCAGCGTC
TCGCTCCTGCGCGAGGGCTTCGCTGCTCGCCGCGTTCGACAACGTTCGATTCGGCGACGC
CGGGACGACGGTTCGGCTCGTTCGACGGCCAGGGCGTTCCTTCGACCCGGGCGCAGCTTCG
CGGCGAAGGGCTTCGTCCAGGGCGAGCGCGGACGGTTCGCCGGCACGGACCTTCGCGTTC
GACGTCCCGGCGGTCCCCGCCGGGACGCCGACAACGCCACGGGCGACGGGCGAGACCAT
CGAGCTTCGACGTCCCCGCGGACGCGGAGCAGCTTCGCGTTCGCGCACGGGCGACGGAGA
AGAACCAGCAGGCCACCGGCACGCTGACCTTCGACGACGGCTTCGACCCAGCCGATTCGAC
CTGAGCTTCGGCGACTGGTTCGGGCGCGGCCCGCAACCCCGTTCGGCAACATCCCGT
CGCGGTGACGGACAGCCGCTCCGCGGGCGGACGCCGAGACCCGGCACCCCGCCGCGGT
TCTTCGCGACGGCGCCGATCACCTCCCCGAGGGCAAGCGGCCCGTGAGCCTCACGCTC
CCGGACCAGCCGGGCGAGCTTCGCGCGACGGCCGCATCCACGTGGTTCGCGGTTCGCGCA
CGACGGCACGTTTCGCCGAGCACCCCGCGCTTCGAGGTTCACGGCCGCGGAGGGCGTTCGCG
TCGCCGTTCGGGCGAGACCTCGGACGTGGCGCTTCGCCAGGTGGCGGGGCGGCCGCGAGGGC
GCCGACCTCCGGGCGGCGGTTCAGTGGGGCGACGGCTCCGACGTCGCGGCCGGCGCGGT
GACCGACGGGTTCGGTTCGCGGCTTCGACGCTACACGGCGGCCGGGACGTACACGGCGT
ACGTTCGTTTCGACGACGGCTGGACCAGCCAGGTGGTTCGAGGTCCCGTTCACCGTTCGAC
GAGGCGGAGCCGGCCCTCGCCGTCGACGTCACGGTTCGACACACGCTGCCTTCGCCGGCAA
GGCGTACGTTCGCGGTTCGCGCGCGAGAACGGCGAGGACGTTCGCGCTTCGCGATCCGGCTC
TCACGCCGTTTCGGCACCAAGGAGGTTCGCGGCCGTCGCGCCGGGCGCAACGCCTACAG
TCGTTTCGCGACGCGGGTTCACGGCGGTTCGAGGCCGGCACCGTTCACCGTTCGAGGCGACGCG

CGGCACCGGCGACGAGGAGGTGACGGCGTCGATCCAGGCCGACTACGCCGCCGTGACCT
GCGGC (SEQ ID NO:12)

FIGURE 11B

MTRPLPPGRAVARSGSGRARPLGLVLAALAVPLGVPLAAPAGALAAAPAAAAEPGDFS
SSFESGDPAALPTTVAERDGPWQANVGSFTAGLPGSVLGQLKGV TASAQNL PNEGAAN
LADGSSGTKWLAFAS TGWVRYEFAEPVSVFVAYTMTSGDDAAGRDPKTWTVEGSNDGSTW
AALDRRTDEDFPNRQQTRTFELEAPTAAYTYLRLNVTANSGDSIVQLAGWDL SADLSAG
PSAAPMTTKVGTGPRV SFTNKAGVGFSGLHSLRYDGSHLADGETYATNVLYDDVDVVVG
EDTRLSYTI FPELLDDLQYPSTYAAVDVLF TDGTYLSDLGARDAHETVATAQAQGE GKI
LYADQWNSVRVDLGDVAEGKTV DQVLLGYDNPGGHAGTKFAGWLDDVEITAE PATIDGS
SLANYVDTRRGT LASGSFSRGNIPATATPNGFNFWTPYTNASSQSWLYEYHKANNANN
KPV LQGFGISHEPSPWMDRNL TFLPSTASGTPDATLSTRGLEFDHADETARPDY YGV
TFTNGSAIEATPTDHGAVLRFSYPGAKGHVLVDKVDGSSKLT YDQATGTISGWVENGSG
LSVGRTRMFVAGTFDRSPTAVGTAAGNRADARFATFETSSDKTVELRVATSFISLDQAR
KNLDLEVTGKTFTEVKAAAQA WNDRLGVIIEVEGASEDQLVTLYSNLYRLNLYPNSQFE
NTGTAQEPVYRYAS PVSATTGSATDTQTNAKIVDGKIYVNNGFWD TYRTAWPAYSLLYP
ELAAELVDG FVQQYRDGGWIARWSSPGYADLMTGTSSDVAFADAYLKGSLPTGTALEAY
DAALRNATVAPP SNAVGRKGLQTS PFLGFTPESTHESVSWGLEGLVND FGI GNMAAALA
EDPATPEERRETL REESAYFLERATHYVELFDPEVDFVPRHEDGTWAVDPETYDPEAW
GGGYTETNGWNFAFHAPQD GQGLANLYGGKQGLEDKLDEFFSTPEKGAGNGGIHEQREA
RDVRMGQWGM SNQVSHHI PWLYDAAGAPSKAQEKVREVT RRLFV GSEIGQGYPGDEDNG
EMSSW WIFASLGFYPLQVGS DQYAVGSP LFDKATVHL PDGDLVNAENNSVDNVYVQSL
AVDGEARTST SLSQADLSGGTTLDFVMGPEPSDWGTGEDDAPPSLTEGDEPPTPVQDAT
TAGLGT TTVADGDATTSAAALTDNTSGTRTTFATTTPSITWAGNGIRPTVGSYTLTSGA
SGTASPSAWTLEGSDDGETWTTLDERSGEQFRWALQTRPFTVAEPTAFARYRVTVTATS
GSGALSLAEVELLADPKESGAEELTLSAAPDRDGV TGREVSGSFATLTGVEGDVAALDV
QVAFGDGSEPVAGTLRAGAFGGYAVDAAHTWTAPGVYPVTVTVSGEGIETVSASSYVSV
SLLREGSLLAAYDNVICGDAGTTVGSCDGGVFFDRAQLAAKGFVQGERATVPGTDLAF
DVPVAVPAGQP DNATGDGQTIELDVPADAEQLSVIGTGTEKNQQATGTLTFDDGSTQPID
LSFGDWSGAARNPVFGNIPVAVTDSRLRGGSPQTGT PAAFFATAPITLPEGKRPVSLTL
PDQPGELSRDGR IHVVAVAHDGTFAEHPALEVTAAEGVTLAVGQTS DVALAQVAGGREG
ADLRAAVTWGDGSDVAAGAVTDG SVSGSHAYTAAGTYTAYVVVDDGWTSQVVEVPVTVT
EAEPALAVDVTVSTRCLAGKAYVAVRAENGEDVPLAIRLVTPFGTKEVA AVAPGANAYQ
SFATRVTA VEAGTVTVEATRGTGDEEVTASIQADYAAVTCG (SEQ ID NO:13)

FIGURE 12A

GCGCTCGCCGTTCGTTCGGCCTTCGCGCCCGCGACCGCCGCGAGCGCCGCCCGAGCCGCC
GTCGGCCGACTACGCGTCCCTGGTTCGACGTCTTCGTTCGGCACCAGGGCGACTTCGGCA
ACGACATGCCCCGCCGCGCAGGGCGCCGAACGGCCTTCGCGAAGGTCAACCCGCGCACGACC
CCGGGCCGCAACAACACCGGGTACGACTACGCGCAGTTCGAAGATCTCGGGCTTCACGCA
CACCAACCTCGACGGGGTTCGGGGGCTTCGGCGGGCGGTGGTTCGACCTCCTCGTGGTGCCGA
CGTCCGGGTTCGTACACGGCGCGCCCCGGCACGGGCACGTACGCGCACCCGTTCTCGCAC
GACGACGAGGACGCCGGACCGGGCTTCTACTCCGTTCGGGCTTCGGCAACGTTCGCGGGCAC
GGACGGCGCGATCACCGGCGCGCCGGGCACGATTCGAGGCCGAGGTTCGCGGGCGGCCACGC
GCTTCGGGCGTGCACCGCTACGCGTTCGCCGCGGGCTTCGACGCCGAGCCTTCGTTCGTGGAC
CTCGAGACGAACAACACGAGCCCGGGTTCGTTCCTTCGGTTCGAGGTTCGAGACGCGCGCGGA
CGGCACCGTGGAGCTGTCCGGACAGGTTCACGGGCTACTTCTACAACGCGGCCCTACACGC
TGTACTACACCGCGCGCACGCTCCAGCCCGCGACGGTTCGAGACGTGGGGCGACGACGAC
CGGCTTCGTTCGACGCCACGGCCCAGGACGGCGTTCGACACCGGCGCGATCCTCACGTTTCGA
CCCCGGCGGACGCCGGGGAGATTCGGGCTTCAGGTTCACCCTGTTCGCCGGTTCGAGCGTTCGAGC
AGGCGCGGATTCGACCAGCAGGTTCGAGCTTCGGCGACCTGTTCGTTCGACGCGATCCGTTCGAC
CGCACC CGCGCGGAGTGGAAACGCGACGCTTCGGGCGGGTTCGCGATTCGACGCCTTCGACGGC
GACGGACCCGACGGGCGAGCTCCAGCGGTCTTCTACACGCACCTCTACCGCATGTTTCG
CGATGCCGATGAACGCGACGAGCACCTTCGGGCACGTACCGCGGCGTTCGACGGGGCGGTTCG
CACGCCGCGCAGGGCTTCACGTACTACGACTTCGTGGGCCACGTGGGACGACTTCCGCAA
GTTCTCCGTTCATTCGCGTACATTCGACCCGGCGCTGTACCGGGACATGGTTCAGTTCGCTGG
TCTACCTGTTTCGCGGACGCCGAGGCGACGGGCACCGGCGGGCGGCCTTCGGCGGGTTCGTTCG
CACTTCGGTTCGCGGACGGTTCGCGCTGGGAGCGGTTCGTTCGGTTCGTGGTTCGCGGACGCGATTCG
CAAGGGCTTCGACGGGTTTCGACCGCCTTCGACGAGGCGTACCGGGCGCTTCAGCGGGCTTCG
TCGGGCGAGTACAGCGCGGACGAGCTCCGGCGCGGCTTCGTTGGCGGGCAACCCCGGCGCG
TCCGTTCGAGCGCGGCTTCGACCCAGTTCGGCCTTCGCGTTCGCGGACGAGCTTCGGCCT
GACCGAGGAGGCCGAGACGCTTCGCGCGAGCAGGGCGTTCGTGGCCGATTCGAGAAGCTTCACCA
AGCCGGGCGCGTTCGACCGCCCGCGGACGCGCAGGTTCGGCCTCCTCACCCCGCGCGCC
GCGGACGGGTCGTTCGAGAGCGCCGACCACGCGAAGTTCGAGGCCGCCGGCCTTCACCA
GGGCACGCTTCGCGAGTACCACTGGTTCGACGCGTTCGACATGGACGCGCTTCGTTCGAGG
CGATGGGCGGGCACGAGGCGGGCGCGCCTTCGGCATTCGCGCCACATGTTTCGGTTCGAGCAGCG
CCGGACGACGGCAAGGCCATGCTCCACTTCGAACGCCAACGAGATTCGACCTTCAGGGCGCC
GTACCTCTTCAACTACACGGGCGAGCCGAGCCTTCACGCGAAGTGGGCGCGCGCGATTC
ACACGAAGGAGACCTGGAACCGGTTCATTCGCGACCGGCTCCTCCAGCGCCGTTCGCGAGC
GGCGGCGGGGAGTTCACGCCGCCCTTGAAGACGAAGGTTCACCGGCTTCGACCCCGCGGG
GATGCTCCCCACGATGGACAACGACGCGGGCACGATGTCGACGATGTTTCGTTCGCGCGGG
CCGTTCGGGCTGTTCCCGGTTCACCGCGGGCTTCGTCCAGTTCAGGTTCGGGTCGCGCTTC
TTCGACTTCGACGACCATCACCTACGACGACGGCAGCGCCTTCACGGTTCACGGCCGACGG
CGTCTCCGAGGACGCGTTCATCGTTCAGTTCGCGACGCTTCGACGGCGCGACGTTTCGGCA
ACACGTGGGTCGACTACGCCACCGTGGTTCGGGGGAGCCGACCTTCGCGTTCGCGATGGGC
GAGCAGCCGAGCGACTGGGGCACGGACACCGCGCCCCGCTTCGATGAGTACCGCGAC
CGACGAGCCGGCCGAGGGACCGCGCGTTCAGCGCCGAACCGACACCGTTCGAGACCGGGC
ACGGCGGGCGCGCTTCGACGCGACCGTTCGACGCTTCACGCTTCGACGGCGGCCCGCCTTCGCGCG
CCCCGCCGCGCACGGACCTTCGTTCAGAGCGGGGCGGGCAGCGTTCGTTCGGGCTTCGCCGACGG
CGTTCACGGCGGGCGGTTCGAGGTTCGCGTTCGCGACCGCGCTTCACCGTTCCTTCGACGGGGA
CGGCGTTCGCGGACGCGCGCTTCCTTCGTTCGACCTTCGCGGACCGCCGCGCTTCGCCGACGGC

GTCGCCGCGGCGTCGCTCCAGGGACAGGGCGTCTCGGTGCGCTCGCCCCTGCGGCTGTC
CGTGGCGTCCGCCGAGCGCGACGCGCTCGCCGCGCTCGTTCGACGACGCCGTGCTCGTGC
GGCACGGGA ACTACTCCTCGGTGACGTTTCGACCGGTTCTCCACCGCGCTGACGAAGGCG
CAGGAGGCCCTCGGTGACGAGGCCGCGACGAGCATCGCGCTGCGGTTTCGCGGCCGACCG
GCTCGGTGCGGCGGCCGACGCGCTCGACCTCACGGGCGGCGGGTACCGCACGCTCGAGG
CCGAGCAGTCCGAGGCGTGGTTCGGGCGGGGAGCTGAAGAACGAGGGCAACAGCTCGTCC
GGCAACCTCGGCGGCGTTCGCTCCGGGTCGTGGGTGCAGTACCGCGACATGACCTTCGA
GACCGCCCGCGGGGACACCCCGCCGCGCTTCTCACGGTCCGGTACGACACGAGCTTCG
CCCCGACGGACACGCCGAGCACCGTTCGCGCTGCACGCGGGCGACGTGAGCGGCCCTGTG
GTCGCGACCGTTCGACCTGAAGGGCACGAGCGGCTGGGGCAAGTACACCGAGGTCACGGC
GGAGCTCGGCGACGTGCAGGCGCTCGTTCGACGCGCAGGTCGTTCAGTTCGAGCTGCTCG
CGCCGTCCGGGCGGAGCTGGGTTCGCAACTTCGACTGGTTCGGTTCAGCGCCGAGGAC
CCGGTTCGCCCCAGGTCAGCCGGGCGAGTCCCCGACGGTTCGACGATCGAGGCCGAGGACTG
GACCGCGAGCTCCGGTTCGCGGGCTCAAGAAGGAGTCTTCGACGTGGACGAGCGGTCCGG
TGACGAACGTCGGCGGCACCGCGGACGGCGACTGGATCGCCTACGGCGAGGTCGACCTG
GGTGAGCTCCCGCTCGGCGAGCTGTCGGTCCACTACGTGCACAACCTCAACCGGTCCGG
GAACAACCTCCGCGCTGTCGGTGTACCTCGACGCGTTCGACCCGGCGAACCCGGGCGAGC
CGTTCGTACCGTTCGCGCTGCCGACGACCGGGTTCGAGCTGGACCGCGGACGGGACCGCG
ACCGTTCGTCTGCCCCGAGACGGTGCAGGGGACGCACGAGGTGTTTCGTGCGCCTGTTCGAC
CGAGCCGTACGCCGACACCCGTACGTTCGCGAACCTTCGACAGCCTGACGTTTCGCGCCGG
GCGGCCCGACGTCGGTTCGTTCGTCGAGTCCGAGGCCCTGGACGTCGAACCTCCGGCCGCGGG
CTGAAGAACGAGAGCTCGACGTGGACGAGCGGTCCGGTTCGACGAACGTCGGCGGCACCGC
GGACGGCGACTGGCTTCGCTACGGCGAGATCGACCTCGGCTCCGCCGCGCTCGACACG
TCTCGGTCCACTACGTGCACAACCTCAACCGGTTCGGGCGGAACCTCCGCGCTGTTCGGT
TACCTCGACGCGTTCGACCCGGCGAACCCGGGCGAGCCGTTTCGTTCACCGTCCCGCTGGC
CAACACCGGGTTCGAGCTGGACGACGGACGGGACCGCCGTTCGTTCGACCTGCCGAGCACGG
TGCGCGGCAAGCACAGGTGTGGGTTCGCGCTTCACCGAGGCGTACGCCGACACCCG
TACGTCGCCAACCTTCGACAGCATGCGCTTCTTCACCGACGCGTACGACGTCGAGGTCCC
GCCGACCGACACCGCGGCGCTTCGCGGCGGTGGTTCGACGCGGCCGGGACGCCCGAGGCGG
AGATCGCGCGGTACGGCCGGATCGACGCGCGCGTCTTCACACGCGAGCTCGCGGCGGCA
CGGTCCGTGCTCGCCGACGCCGGCGCCACCCAGGCGCAGGCCGACGAGCGGGCGCGGCG
CCTCGGCCTGGCGACCGACAGCTCGTTCGCCGCGAGCGCCGTTCGGTTCGAGAACCTCG
TGGCGAGCGCCGAGGCCCTGACCGACGAGGGGTACAGCCCCGAGTCTTGGCAGGCCTTC
CGCACGGCTTCGCCGCGGCGACCGGGACGCTCGACGACGCGGCGGCGTCCGACGAGGC
GCTGCACGACGCGCGGCTTCGCGCTCCAGGGCGCCGTTCGACGCCCTGGAGGAGCCGGCCG
ACGTTCGTGCTTCGTCGAGGTCGAGGTCAGCCCGCGCTGCCTCGCCGGCAAGCCCTACGTC
GCGGTCCGCGCGGTGAACGTCCTCCGACGCGGCCGTTCGACGTCGAGCTGGCGTTCGTC
GGGCACGAGGTCGTTTCGTTCGGCGTTCGCGCCGGGGGCGAGCGCGTACCAGTTCGTTTCGCCG
CGCGGTCCGCGACGGGCGACCTGGACGTCACCGTTCACGGCGACGGGGCGGACGGCACC
CAGACGGTTCGAGCAGGTCGTCACCGTCCCGTTCCTGCTCC (SEQ ID NO:14)

FIGURE 12B

ALAVVGLAPATAASAAPEPPSADYASLVDVVFVGTGEGDFGNDMPAAQAPNGLAKVNPRTT
PGRNNTGYDYAQS KISGFTHTNLDGVGGSSGGGDLVVP TSGSYTARPGTGT YAHPFSH
DDEDAGPGFY SVGLGNVAGTDG AITGAPGTIEAEVAAATRS GVVHRYAF PAGSTPSLVVD
LETNNTSRRSSSVQVETRADGTVELSGQVTGYFYNAAYTLYYTARTLQPATVQ TWGDDD
RLVDATAQDGVDTGAILTFDPADAGEIGLQVTLSPVSV EQARIDQQVELGDLSFDAIRD
RTRAEWNATLGRVAIDASTATDPTGELQRLFYTHLYRMFAMP MNATSTSGTYRGVDGAV
HAAQGFTYYDSWATWDDFRKFSVIAYIDPALYRDMVQSLVYLFADAEATGTGGGLGGFV
HSVPTVRWERS SVVADAIKGFDFDRLEAYPALQRLV GQYSADELRRGYVAGNPGA
SVQRGYDQYGLSVIADDELGLTEEAETLREQASWPIEK LTKPGAWTAADGTQVGLLT PRA
ADGSWQSADHAKFEAAGLYQGT LWQYHWYDAYDMDALVEAMGGHEAARLGM RHMFGEGA
PDDGKAMLHSNANEIDLQAPYLFNYTGEPSLTQKWARAIYTKETWNR YIATGSSSAVPS
GGGEFTPPLKTKVYRLDPRGMLPTMDNDAGTMSTM FVAAAVGLFPVTAGSSQFQVGS PF
FDSTTITYDDGSAFTVTADGVSEDAFYVQSATLDGATFGNTWVDYATV VGGADLAFR MG
EQPSDWGTD TAPAFSMSTATDEPAEGPRVSAEPTTVQ TGDGGALDATVTLTLDGARLAA
PAGTDLVTSGAASV VGLPDGVTA AVTVASPTALT VSLTGTASADARFFVHLRDAALADG
VAAASLOGQGVSVRSPLRLSVASAERDALAALVDDAVLVRHGNYSSVTFDRFSTALTKA
QEALGDEAATSIALRFAADRLGAAADALDLTGGGYRTLEAEQSEAWSGGELKNEANSSS
GNLGGVRS GSWVQYRDMTFETAAGDTPPRFLT VRYDTSFAPTDT PSTVRVHAGDVSGPV
VATVDLKGTS GWGKYTEVT AELGDVQALVDAQVVT FELLAPSGRSWGNFDWFRFSAED
PAAPGQPGESPTVTIEAEDWTASSGRGLKKESSWTSGPVTNVGGTADGDW IAYGEVDL
GELPLGELSVHYVHNSNRSGNNSALSVYLDAFD PANPGE PFVTVPLPTTGSSWTADGTA
TVVLPETVQGTHEVFVRLSTEPYADHPYVANLDSLTFAPGGPTS SVVVESEAWTSNSGRG
LKNESSTWTSGPVTNVGGTADGDW LAYGEIDLGSAALDQLSVHYVHNSNRSGRNSALS V
YLDAFD PANPGE PFVTVPLANTGSSWTTDGTAVVDLPSTVVRGKHQVWVRLSTEAYADHP
YVANLDSMRFFTDAYDVEVPPTDTAALAAVVDAAGTPEAEIARYGRIDARVFTRELA AA
RSVLADAGATQAQADERARRLGLATDQLVPAERRRLENLVA SAEALTDEGYSPESWQAF
RTALAAATGTLDDAAASDEALHDARLALQGAVDAL EEPADVVLVEVEVSPRCLAGKPYV
AVRAVNVSDAAVDVELASSLGTRSFVGVAPGASAYQSFAARSATGDLDVTVTATGADGT
QTVEQVVTVPSCS (SEQ ID NO:15)

FIGURE 12C

APEPPSADYASLVDVFGTEGDFGNDMPAAQAPNGLAKVNPRTTPGRNNTGYDYAQSKI SGF
THTNLDGVGGSGGGDLLVVPTSGSYTARPGTGTIAHPFSDHDEEDAGPGFYSVGLGNVAGTD
GAITGAPGTIEAEVAAATRSGVHRYAFPAGSTPSLVVDLETNNTSRRSSSVQVETRADGTVE
LSGQVTGYFYNAAYTLYYTARTLQPATVQVTWGDRLVDDRLVDDATAQDGVDTGAILTFDPADAGEI
GLQVTLSPVSVQARIDQQVELGDLSDFAIRDRTRAEWNATLGRVAIDASTATDPTGELQRL
FYTHLYRMFAMPNATSTSGTYRGVDGAVHAAQGFYYDSWATWDDFRKFSVIAYIDPALYR
DMVQSLVYLFADAEATGTGGGLGGFVHVSPTVRWERSVSVVADAIKGFDFDRLDEAYPAL
QRLVGQYSADELRRGYVAGNPGASVQRGYDQYGLSVIADELGLTEEAETLREQASWPIEKLT
KPGAWTAADGTQVGLLTAPRAADGSWQSADHAKFEAAGLYQGTWQYHWYDAYDMDALVEAMG
GHEAARLGMRFHGFGEHAPDDGKAMLHNSANEIDLQAPYLFNYTGEPSLTQKWARAIYTKETW
NRYIATGSSSAVPSGGGEFTPPLKTKVYRLDPRGMLPTMDNDAGTMTMFVAAAAGLFPVTA
GSSQFQVGSPPFDSTTITYDDGSAFTVTADGVSEDAFYVQSATLDGATFGNTWVDYATVVGG
ADLAFRMGEQPSDWGTDTPAFSMSTATDEPAEGPRVSAEPTTVQTDGGALDATVTLTLDG
ARLAAPAGTDLVTSGAASVVGLPDGVTAAVTVASPTALTVSLTGTASADARFFVHLRDAALA
DGVAAAASLQGGVSVRSPLRLSVASAERDALAALVDDAVLVRHGNYSVTFDRFSTALTKAQ
EALGDEAATSIARFAADRLGAAADALDTGGGYRTLEAEQSEAWSSGELKNEANSSSGLNG
GVRSGSWVQYRDMTFETAAGDTPPRFLTTRYDTSFAPDTPSTVVRVHAGDVSQPVVATVDLK
GTSGWGKYTEVTAELGDVQALVDAQVVTPELLAPSGRSWVGNFDWFRFSAEDPAAPGQPGES
PTVTIEAEDWTASSGRGLKKESSWTSGPVTNVGGTADGDWIAEGEVDLDELPLGELSVHYV
HNSNRSGNNSALSVYLDADFDPANPGEFVTVPLPTTSSWTADGTATVVLPEVQGTHEVFFV
RLSTEPYADHPYVANLDSLTFAPGGPTSVVVESEAWTSNSGRGLKNESSWTSGPVTNVGGT
ADGDWLAYGEIDLGSAALDQLSVHYVHNSNRSGRNSALSVYLDADFDPANPGEFVTVPLANT
GSSWTTDGTAVVDLPSTVVRGKHQVWVRLSTEAYADHPYVANLDSMRFFTDAYDVEVPPTDTA
ALAAVVDAAGTPEAEIARYGRIDARVFTRELAARSVLADAGATQAQADERARRLGLATDQL
VPAERRRLENLVAEALTDGYSPEWQAFRTALAAATGTLDDAAASDEALHDARLALQGA
VDALEEPADVVLVEVEVSPRCLAGKPYVAVRAVNVSDAAVDVELASSLGTRSFVGVAPGASA
YQSFAARSATGDLDDVTATGADGTQTVQVTVPSCS (SEQ ID NO:50)

FIG. 13

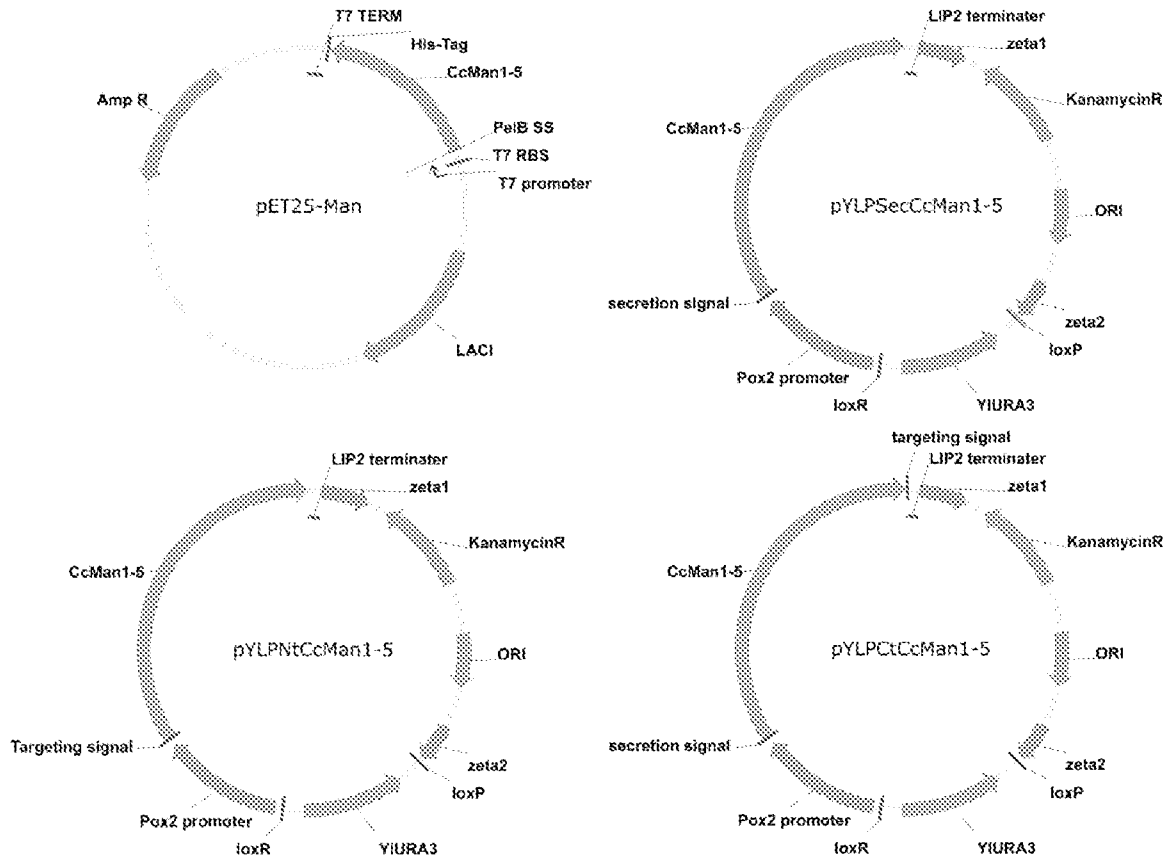


FIGURE 14

GCACCGGCAGATGAAGGCACCGTTACCGCAGCAGCCGGTGATGATCTGACCCTGGAAGTTAATCCGTTTGT
TGGCACCGAAAGCGAAGGTAATGCATATCCGGGTGCAACCGTTCCGTTTGGTATGGTTCAGCTGTCTCCGG
ATAATACCAATAGCTATGCCAGCACCAGCTATAGCACCAATGCAGGTTCGTGTTTGGGGTTTTAGCCATCGT
CATGTTAATAGCGCAGGTTGTCCGGCAGCCGGTGAAGTCTGGTTACACCGGATAACCAGCGCAACACCGCG
TACCAGCCGTAGCTTTATTGCCATCAAAGATCAGAAAAGCACCGAACGTGCAAGCGCAGGTTTTTATGAAG
TTACCCTGGCAAATGATGTTTCATGCAGAACTGACCGCAACCACCCGTGTTGGTGCACATCGTTATACCTTT
CCGGCAAGCACCACCTCTCATCTGAGCTTTAATGTTGGTGCAGACCCTGCGTGATGCCGGTGCAAGCAGCGT
TACCTGGGTTGATGATCGTACACTGGAAGGTTGGGTTGATAATGGTGGTTTTTGTGGTGGTACACCGGATA
AACAGCGCTATTTTTTTAGCGCAACCTTTGATCGTCCGGTTGCCAGCAGCGGTACATGGGGCACCGATGCA
CGTTATGTTGCAGGTAGCACCACAGTGAAGTTGCCGGTGGTAATAATGGTGCAGTTGCCGTTTTTGTATAC
CACCACCGATCGTGATGTTGAAGTTAGCGTTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGTGGT
GTGAAGCCGAAGCAACCGATGAAGGTGGTGCAGGTTGCATTTGATAACCGTTCGTGAAGAAGCACGCGACGCC
TGGAAATGCAGAACTGGGTCGTGCAGCAATTGATGCATCTCCGGATCAGCGTCGTATCTTTTATACCCAGCT
GTATAAAACCCTGCTGAGCCCGACCATTGGTCTGATGTTGATGGTTCGTATCGTGGTATGGATCTGGAAG
TTCATCAGGCAGATGGCTGGGATTATTATCAGAACTTTAGCCTGTGGGATACCTATCGTACCCAGGCAACC
CTGCATGCACCTGCTGCCGGAACGTGCACAGGATATTGTTTCGTAGCATGTATCAGCATCGTGTGAAGG
TGGTTGGCTGCCTCGTTGGTCTCTGGGTGCACCTGGAACCAATATCATGGCAGGCGATCCCGTTACCCCGT
GGCTGGCAGAAAATTTTGCACCTGGGCACCGTTCCGGATGATATTGCAGATGAAGTGGGATTATCTGGTT
GAAAATGCAACCACCACCCTCCGGATGATGTTGCCAGCGTTGGTTCGTTCGTAGCACCAGAAATTTATGCCGA
ACATGGTCATGTTCCGTTTTATCCGGAAAACGAAGGTGGCCTGGGTGGTTCAGTTTGAAGAATATCGTCATG
GTGGTAGCGCAACCCTGGAAGTGGCAGTGGCAGATGCAAGCCTGGGTGCCGAGCAGAACTACCCGGTCGT
GAAGGTGGCCAGGCATTTCTGGATAAAGGTTCGCAATTGGCGTAATCTGTGGAATCCGGATGTTGAACTGAG
CGGTGGTTTTTCAGGGTATGGTTAATGCAAAACGTCCGACCCGGTGAATTTGTTACCCTGCCGAACTGACCG
ATGTTACCCGTAGCGGTTTTTCATGAAGGTGTTCCGTGGCAGTATCAGTGGATGGTTCGCGAGGATGTTACC
GGTCTGCAAGAAGTTATGGGAGGCGAAGATGGTTTTGTGGAACGCCTGGATTATTATTTTATGATCAGCCTGC
ACTGGCAGCAAATCCGGGTGTTAGCCCGAGCACCTGGGCAAAAGGTGGTAGCAGCTATTATACCACCATTC
GCTATAATCCGGGTAATGAACCGACCATTATGAATGCATGGCTGTATGGTTATGTTGGTGCAGCCGTGGAAA
ACCAATGATGTTCTGGCAGCCAATCTGAATCGTTTTCCGGATACACCAGGTTGGTGGTGGTGGTGGTGGTGGT
TCTGGGCACCCTGGCAGCATGGTATGTTATGGCCAGCCTGGGTTTTGAACCGGTTATGCCTGGTAGCGGTA
TTCTGGCACTGAATGCACCGAAAGTTCAGGCAGCAACCCTGACCACCGATGCCGGTGCCACCCTGCGTATT
GATGCAGCCGGTGCAAATGAAAACTGCCGAGCTATGTTGCCGGTCTGGAAGTTGATGGTGTTCACATAC
CGCAGCATGGCTGGATGTTGCAGCACTGCAGGATGGTGGCACCCTGGATTTTATGATCTGAGCGGTACAAGCG
CAGGTCTGACATGGGGTACAGGTGCAGCAGATCGTATTCGAGCGTTAGCGCAGTTGCACCGCCTGCACCG
GTTGAAGTGGAAGCAAGCGCACGTTGTCTGGGTGGTTCGTGCATTTGTTGCAGTTTCGTGCAACCAGCACCGC
AGATGCACCGGTGGATGTTACACTGACCACACCGTTTTGGTGAACGTACCGTTTCGTATGTTTACCGCTGGT
GTAGCGCATATCAGAGCTTTACCACCCGTACCACCTCTGTTGAAGCAGGCACCAGCAACCGTTACCGTTGTT
GCAGCAGATGGCACCACCTCAACCGTTGATGCAGCATATGAAGCACTGGCATGTGGTTAATAA (SEQ ID
NO:16)

FIGURE 15

GCAGGCACCGAAGCAGCAACCGGTTCTGATGCAGCAGCAGTTGATGGTCCGCTGGTTGATTATGTGAATCC
GTTTATTGGCACCAAGATGATGGTAATACCTATCCGGGTGCAGCAGTTCCGTTTGGTATGGTTCAGCTGT
CTCCGGATAATGGTCATAATGTGGGCTATGATTATGATCGTACCAGCGTTCGTGGTTTTAGCCTGGTTCAT
CTGAGCGGTGTTGGTTGTGGTCTGGGTGGTCCGCTGCCGACACTGCCGACCACCGGTGCAATTACCAGCAC
CGATTATGGTCAGTATGCACTGGGTTTTAGCCATGATGATGAAGAAGCATCTCCGGGTTATTATCGTGTTG
GTCTGCAGGCACCTGCAGGAACCATTGAAGCAGAAGTACCAGCAACCGAACGTACCAGGTTTACAGCGTTAT
ACCTTTCCGGCAACCGCACAGGCAATGTTCTGCTGAATGCAGGTCAGGCACTGAATCGTGTTACCGAATC
TGATGTTTCGTGTTGTTGATGATCGTACCAGTTGAAACCCGTTTACCAGTGCCTGGTTTTTGTGAGGATACCG
AACCAGCAGACCTTTGGACCCGTTACCACCTTTGATCGTCCGTTTTGTTGCACATGGCACCTGGGATGGTCAG
GTTGTTACCAGCAGGCGCAGATGCAGCAAGCGGTGGTGAAGGTCGTGCTGGTGCATATGTTACCTTTGATAC
AACCAGGTTGGTATCTGGATGTTGAAGCAGTTACCAGCAATGAGCTATGTTGGTGCAGATGGTGCAGCAGCAA
ATCTGGCAGCAGAAGCAGGCACCTTTGACGCAGTTCATGATGCAGCACGTAGCGCATGGGAAGAACGTCTG
GGTCTGGTTTCGTGTTGCACAGGGTATCCGGATGATCTGCGTACCTTTTATAGCAGCCTGTATCGTAGCTT
TCTGGCACCGAATGTTGGTCTGATGTGGATGGTTCGTTATCGTGGTTGGGATCAGGAAGTTCACGCAGCAG
AACCAGGATTTTACCTATTATCAGAATTATAGCCTGTGGGATACCTATCGTACCCAACAGCAACTGCTGTAT
CTGCTGGCACCAGGATGAAAGCGCAGATATGGCATTGAGCCTGGTTTCGTGAGGGTGCAGAGGGTGGTTGGCT
GCCGTTGGGGTTATGGTACAGTGGAAACCAATATTATGACCCGGTATCCGGCAACCCCGTTTCTGGTTA
GCCGATGGCGTCAGGGTCTGCTGGCAGGTCATGAAGAAGAAGCATAACGCAGTCCGCTGAAAATGCAGAT
GGTGTTCCTCCGGCAGATAGCCCGTTAATGGTTCGTGCAGCAATGTTGAATATCTGCGTGGTGGTTTTGT
TCCGCATGAACCGGCACGTAGCGGTAACCGGGTGAATTATGATCTGCAGCATGGTGCAGCGCAACCATGG
AATATGCACTGGCAGATGCAATGCTGAGCACCATGGCACGTGGTCTGGGTGATGATGAAGATGCAGATCGT
TATGCAGCCCGTGGTGCAGAGCTATCGTAATGTTTTTGTATCCGCGTACCAGGTAATTTTTCGTGCACGTAATGC
CGATGGTTTTTTTTGTTGGTATGCAGATCCGGCACATTCTGATGGTTTTTTCATGAAGGCACCGCAGTTCAGT
ATCAGTGGCTGGTTCGGCAGGATGTTCCGGGTCTGTTTGTATCTGATGGGTGGCACCGATGCAGCCGTTGAT
CGTCTGGATGCATTTTTTGCCTATGATGAACTGGTTGCAGATCCTCCGCATGTTGCAAGCGAAGTTGGGT
TAATGGCACCTATGATTATTATGGCTGGGAAACCTATAATCCGAATAATGAACCGAATCTGCATGCACCGT
ATGTTTTATCTGTGGACCGGTGAGCCGTGGAAAACCCAGATGTTGTTTCGTGCAGCAAGCACCCCTGTTTACC
GATGGTCCGGATGGTGTACCAGGTAATGATGATCTGGGCACCATGAGCGCATGGCATGTTCTGAGCAGCAT
TGGTGTATATCCGATTGTTCCGGGTGCCGATCTGTGGGGTCTGACCACACCGCTGTTTGTATGATGTTACCA
TTACCCTGGACCCGGAAGTTTTTGGTTCGTGATAGCCTGCGTCTGACCAGCAGATGGTGTGGCACCGGATAACC
CATTATACCCAGAGCGTTAGCCTGGGTGGTGAACCGCTGGATCGTGCATGGGTTACAGGTGATGAACTGAC
CGCTGCAGGCACCCCTGGATGTTACCAGTTGGCACCGAACCAGCGCATGGGCAACCGATCCGGCAGCATCAC
CGGGTGCAGTTGTTCCGGCTGATGGCACCGTTGAACGTCTGTTTGTGGTGCACACCGCGTCAGCCGGTT
CTGGCACCAGGTTGGTACCGAAGTTGCAGTTACAGTTGAGTTGTTGCCAGGGTGCAGGCACCTCTAGCCGGCAC
CCTGGAAGTGACCTCTGATGGTGCAGTTACCAGCACCACCGATCTGGCAGAATGGACCCGAGAATCTGATG
GTTGCTGCTGCCACCGTTGAAGGAACCGTTACCATTGAAGCTCCGGCAGATGCCGAACCGGGTCTGCATACC
GTTGCTGCTGGTTGTTTCGTGATGCAGCCGGTACAGAAGCAGTTCGCGAAGTTAGCGTTGTTGTTAGCGGTGA
AAGCTGGATTGCAGATGCCTTTGATAATGTGGGTATTGGTGTATGCCGGTGCAGCAAAATGCAAAATCTGGATG
GTAGCGGTGCCTATCTGCTGCGTATCTGCTGGCCGATCTGGGTGCAGTTACAGGGTCTGGAACTGACCGTT
CCGGGTACTGATCTGACCTATACCCTGGGTGCACCGCGTGTGGTGCACCGGATAATGTTGCAGCCAGCGG
TGAAGTTCTGGAAGTTCCGGAACATCTGCGTAGCGCACGTATCTGAGCGTTGTGGGCACCAGCACCATG
GTACACATGGTGGTGGTCTGGTTCTGGGTTTTGCCGATGGTAGCAGCCAGACCGTTGATGTTTCGTCTGAGC
GATTGGTGTACCAGTTCTCCGGAACCGGGTAATATTACCAGTTGCAAAAAGCCGGTGCACGTGGTGTATCGTGA
AAATGTGCAGAAAATTGGCTGTGGTCTGTATGCAACCGCACCGGTGGCAATTCGGGAAGGTAAAGTTCTGA
CCAGCGTTACCCTGCCGTCTGATGAACGTTTTTTCATGTGTTTGCATTTGCAACCGATGCAACCGGTGATGTT
CCGGCACCGCAGGTTGAAGTTACCAGCACAGGCTCGTTGCTGGGTGGTAAAGCATTTGTTGCAGTTCGTGC
ACTGAATACCGGTGAACAGCCTGCAGCAATTGAACTGGCAACCCCGTATGGTAGCAAACTGTTTGGTGTATG
TTGCTCCGGGTGCAAAATGCATATCAGAGCTTTGCAACCCGTGCAGCAGCCGTTGAAGCCGGTGAAGTTACC
GTTACCAGTACCACACCGGATGGTGAACCGCAGCAGGTTACCAGCAGCATATGATGCAGCGGCATGTAGCTA
ATAA (SEQ ID NO:17)

FIGURE 16

GCAGGCACCGAAGCAGCAACCGGTTCTGATGCAGCAGCAGTTGATGGTCCGCTGGTTGATTATGTGAATCC
GTTTTATTGGCACCAAAGATGATGGTAATACCTATCCGGGTGCAGCAGTTCCGTTTGGTATGGTTCAGCTGT
CTCCGGATAATGGTCATAATGTGGGCTATGATTATGATCGTACCAGCGTTCGTGGTTTTAGCCTGGTTCAT
CTGAGCGGTGTTGGTGTGGTCTGGGTGGTCCGCTGCCGACACTGCCGACCACCGGTGCAATTACCAGCAC
CGATTATGGTCAGTATGCACTGGGTTTTAGCCATGATGATGAAGAAGCATCTCCGGTTATTATCGTGTTG
GTCTGCAGGCACCTGCAGGAACCATTGAAGCAGAAGTACCAGCAACCGAACGTACCAGTTCAGCGTTAT
ACCTTTCCGGCAACCGCACAGGCAAATGTTCTGCTGAATGCAGGTCAGGCACTGAATCGTGTTACCGAATC
TGATGTTTCGTGTTGTTGATGATCGTACCAGTTGAAACCCGTATTACCAGTCCGTTGGTTTTGTCAGGATACCG
AACCGCAGACCATTTGGACCCGTACCACCTTTGATCGTCCGTTTGGTGCACATGGCACCTGGGATGGTCAG
GTTGTTACCAGCAGGCGCAGATGCAGCAAGCGGTGGTGAAGGTCGTTCGTGGTGCATATGTTACCTTTGATAC
AACCGGTGGTGTATCTGGATGTTGAAGCAGTTACCAGCAATGAGCTATGTTGGTGCAGATGGTGCAGCAGCAA
ATCTGGCAGCAGAAGCAGGCACCTTTGACGCAGTTCATGATGCAGCACGTAGCGCATGGGAAGAACGTCTG
GGTCTGGTTCGTGTTGCACAGGGTGTATCCGGATGATCTGCGTACCTTTTATAGCAGCCTGTATCGTAGCTT
TCTGGCACCGAATGTTGGTCTGATGTGGATGGTTCGTTATCGTGGTTGGGATCAGGAAGTTCACGCAGCAG
AACCGGATTTTACCTATTATCAGAATTATAGCCTGTGGGATACCTATCGTACCCAACAGCAACTGCTGTAT
CTGCTGGCACCAGGATGAAAGCGCAGATATGGCATTGAGCCTGGTTTCGTTCAGGGTCAGCAGGGTGGTTGGCT
GCCGTTGGGGTTATGGTACAGTGGAAACCAATATTATGACCCGGTATCCGGCAACCCCGTTTCTGGTTA
GCGCATGGCGTCAGGGTCTGCTGGCAGGTCATGAAGAAGAAGCATAACGCAGTCCGTGAAAATGCAGAT
GGTGTTCCTCCGGCAGATAGCCCGTTAATGGTTCGTGCAGCAATGTTGAATATCTGCGTGTGGTTTTGT
TCCGCATGAACCGGCACGTAGCGGTAAACCGGGTGAATATGATCTGCAGCATGGTGCAGCAACCGAATGG
AATATGCACTGGCAGATGCAATGCTGAGCACCATGGCACGTGGTCTGGGTGATGATGAAGATGCAGATCGT
TATGCAGCCCGTGGTGCAGAGCTATCGTAATGTTTTGATCCGCGTACCAGTAATTTTCGTGCACGTAATGC
CGATGGTTTTTTTTGTTGGTGTATGCAGATCCGGCACATTCTGATGGTTTTTCATGAAGGCACCGCAGTTCAGT
ATCAGTGGCTGGTTCGCGCAGGATGTTCCGGGTCTGTTTGTATCTGATGGGTGGCACCGATGCAGCCGTTGAT
CGTCTGGATGCATTTTTTGCCTATGATGAACTGGTTGCAGATCCTCCGCATGTTGCAAGCGAAGTTGGGT
TAATGGCACCTATGATTATTATGGCTGGGAAACCTATAATCCGAATAATGAACCGAATCTGCATGCACCGT
ATGTTTTATCTGTGGACCGGTGAGCCGTGGAAAACCGATGTTGTTTCGTGCAGCAAGCACCCCTGTTTACC
GATGGTCCGGATGGTGTACCAGGTAATGATGATCTGGGCACCATGAGCGCATGGCATGTTCTGAGCAGCAT
TGGTGTATATCCGATTGTTCCGGGTGCCGATCTGTGGGGTCTGACCACACCGCTGTTTGTATGATGTTACCA
TTACCCTGGACCCGGAAGTTTTTGGTTCGTGATAGCCTGCGTCTGACCGCAGATGGTGTGGCACCGGATAACC
CATTATACCCAGAGCGTTAGCCTGGGTGGTGAACCGCTGGATCGTGCATGGGTTACAGGTGATGAACTGAC
CGCTGCAGGCACCCCTGGATGTTACCAGTTGGCACCGAACCGAGCGCATGGGCAACCGATCCGGCAGCATCAC
CGGGTGCAGTTGTTCCGGCTGATGGCACCGTTGAACGTCTGTTTGGTGGTGAACACCGCGTCAGCCGGTT
CTGGCACCAGGTTGGTCTGATGGTGCAGTTACCAGTTCAGGTTGTTGCCAGGGTGCAGGCACCTCTAGCCGCAC
CCTGGAAGTGACCTCTGATGGTGCAGTTACCAGTTCAGGTTGTTGCCAGGGTGCAGGCACCTCTAGCCGCAC
GTTGCTGCTGGTTGTTTCGTGATGCAGCCGGTACAGAAGCAGTTCGCGAAGTTAGCGTTGTTGTTAGCGGTGA
AAGCTGGATTGCAGATGCCTTTGATAATGTGGGTATTGGTGTATGCCGGTGCAGCAAAATGCAAAATCTGGATG
GTAGCGGTGCCTATCTGCTGCGTGTATCTGCTGGCCGATCTGGGTGCAGTTCAGGGTCTGGAACTGACCGTT
CCGGGTACTGATCTGACCTATAACCCTGGGTGCACCGCGTGTGGTGCACCGGATAATGTTGCAGCCAGCGG
TGAAGTTCTGGAAGTTCCGGAACATCTGCGTAGCGCACGTCTGAGCGTTGTGGGCACCAGCACCATG
GTACACATGGTGGTGGTCTGGTTCTGGGTTTTGCCGATGGTAGCAGCCAGACCGTTGATGTTTCGTCTGAGC
GATTGGTGTACCAGTTCTCCGGAACCGGGTAATATTACCAGTTGCAAAAAGCCGGTGCACGTGGTGTATCGTGA
AAATGTGCAGAAAATTGGCTGTGGTCTGTATGCAACCGCACCGGTGGCAATTCGGAAGGTAAGTTCTGA
CCAGCGTTACCCTGCCGTCTGATGAACGTTTTTCATGTGTTTGAATGCAACCGATGCAACCGGTGATGTT
CCGGCACCGCAGGTTGAAGTTACCAGCACAGGCTCGTTGCTGGGTGGTAAAGCATTTGTTGCAGTTCGTGC
ACTGAATACCGGTGAACAGCCTGCAGCAATTGAACTGGCAACCCCGTATGGTAGCAAACTGTTTGGTGTATG
TTGCTCCGGGTGCAAAATGCATATCAGAGCTTTGCAACCCGTGCAGCAGCCGTTGAAGCCGGTGAAGTTACC
GTTACCAGTACCACACCGGATGGTGAACCGCAGCAGGTTACCAGCAGCATATGATGCAGCGGCATGTAGCTA
ATAA (SEQ ID NO:18)

FIGURE 17

GCAGAACC GG GTGATTTTAGCAGCAGCTTTGAATCTGGCGATCCGGCAGCACTGCCGACCACCGTTGCAGA
ACGTGATGGTGCACCGTGGCAGGCAAATGTTGGTAGCTTTACCGCAGGTCTGCCTGGTAGCGTTCTGGGTC
AGCTGAAAGGTGTTACCGCAAGCGCACAGAATCTGCCGAATGAAGGTGCAGCAAATCTGGCAGATGGTAGC
AGCGGCACCAAATGGCTGGCATTGCAAGCACC GG TGGGTTTCGTTATGAATTTGCAGAACC GG TTAGCTT
TGTTGCATATAACCATGACCAGCGGTGATGATGCCGCAGGTCGTGATCCGAAAACCTGGACCGTTGAAGGTA
GCAATGATGGTCTACCTGGGCAGCACTGGATCGTTCGTACCGATGAAGATTTTCCGAATCGTCAGCAGACC
CGTACCTTTGAACTGGAAGCACC GACC GCAGCATATACTATCTGCGTCTGAATGTTACCGCAAATAGCGG
TGATAGCATTGTTTCAGCTGGCAGGTTGGGATCTGAGCGCAGATCTGTCTGCAGGTCCGAGCGCAGCACC GA
TGACCACCAAAGTTGGCACC GG TCCGCGTGT TAGCTTTACCAATAAAGCCGGTGTGGTTTTAGCGGTCTG
CATAGCCTGCGTTATGATGGTAGCCATCTGGCCGATGGTGAAACCTATGCAACCAATGTGCTGTATGATGA
TGTTGATGTTGTGGTTGGTGAAGATACCCGTCTGAGCTATAACATTTTTCCGGAACCTGCTGGATGATCTGC
AGTATCCGAGCACCTATGCAGCAGTTGATGTTCTGTTTACCGATGGCACCTATCTGAGCGATCTGGGTGCA
CGTGATGCACATGAAACCGTTGCAACCGCACAGGCACAGGGTGAAGGTA AAAATCTGTATGCCGATCAGTG
GAATAGCGTTCGTGTTGATCTGGGTGATGTTGCAGAAGGTA AAAACCGTTGATCAGGTTCTGCTGGGTTATG
ATAATCCGGTGGTTCATGCAGGCACCAAATTTGCAGGTTGGCTGGATGATGTTGAAATTACCGCAGAACC G
GCAACCATGATGGTAGCTCACTGGCAAATTTGTTGATACCCGTCTGGCACCCTGGCAAGCGGTAGCTT
TAGCCGTGTTAATAATATCCGGCAACCGCAACCCGCAATGGTTTTAATTTTTGGACCCCGTATACCAATG
CAAGCAGCCAGAGCTGGCTGTATGAATATCATAAAGCCAATAATGCGAATAATAAACCCGGTCTGCAGGGT
TTTTGGTATTAGCCATGAACCGAGCCCGTGGATGGGTGATCGTAATCAGCTGACCTTTCTGCCGAGCACC GC
AAGCGGTACACCGGATGCAACCCCTGAGCACCCGTGGTCTGGAATTTGATCATGCAGATGAAACCGCACGTC
CGGATTATTATGGTGTGACCTTTACCAATGGTAGCGCAATTGAAGCAACCCCGACCGATCATGGTGCAGTT
CTGCGTTTTAGCTATCCGGGTGCAAAAGGTTCATGTTCTGGTGGATAAAGTTGATGGTAGCAGTAAACTGAC
CTATGATCAGGCAACCGGCACCATTAGCGGTTGGGTTGAAAATGGTAGCGGTCTGAGCGTTGGTCTGATACC
GTATGTTTGTGTCAGGCACCTTTGATCGTAGCCCCGACCGCAGTTGGCACAGCAGCAGGTAATCGTGCAGAT
GCAGTTTTGCAACCTTTGAAACCAGCAGCGATAAAAACCGTGGAACTGCGTGTGCAACCAGCTTTATTAG
CCTGGATCAGGCACGTAAAAATCTGGATCTGGAAGTTACCGGTAAAACCTTTACCGAAGTTAAAGCAGCAG
CAGCACAGGCATGGAATGATCGTCTGGGTGTTATTGAAGTTGAAGGTGCAAGCGAAGATCAGCTGGTTACC
CTGTATAGCAATCTGTATCGCCTGAATCTGTATCCGAATAGCCAGTTTTGAAAATACCGGCACCGCACAGGA
ACCGGTTTTATCGTTACGCATCTCCGGTTAGCGCAACCACCGGTAGCGCAACCGATACCCAGACCAATGCCA
AAATTGTGGATGGCAAAATTTATGTGAATAATGGCTTTTTGGGATACCTATCGTACCGCATGGCCTGCATAT
AGCCTGCTGTATCCGGAACCTGGCAGCAGA ACTGGTTGATGGTTTTGTTTCAGCAGTATCGTGATGGTGGTTG
GATTGCACGTTGGAGCAGTCCGGGTTATGCAGATCTGATGACCGGTACAAGCTCTGATGTTGCATTTGCAG
ATGCCTATCTGAAAGGTAGCCTGCCGACCGGTACAGCACTGGAAGCATATGATGCAGCACTGCGTAATGCA
ACCGTTGCACCTCCGAGCAATGCAGTTGGTTCGTAAGGTTCTGCAGACAAGCCCGTTTTCTGGGTTTTACACC
GGAAAGCACCATTGAAAGCGTTAGCTGGGTTCTGGAAGGTTCTGGTTAATGATTTTTGGCAATATGG
TGCAGCACTGGCAGAAGATCCGGCAACACCGGAAGAAGCTCGTGAAACCCCTGCGTGAAGAAAGCCATAT
TTTTCTGGAACGTGCCACCCATTATGTTGAACTGTTTGTATCCGGAAGTGGATTTTTTTGTTCCGCGTCATGA
AGATGGTACATGGGCAGTTGATCCGGAAACCTATGATCCGGAAGCATGGGGTGGTGGTTATACCGAAAACA
ATGGCTGGAATTTTGCATTTTCATGCACCGCAGGATGGTCAGGGTCTGGCAAATCTGTATGGTGGTAAACAG
GGTCTGGAAGATAAACTGGATGAATTTTTTAGCACACCGGAAAAAGGTGCAGGTAATGGTGGTATTCATGA
ACAGCGTGAAGCACGTGATGTTTCGTATGGGTGAGTGGGTTATGAGCAATCAGGTTAGCCATCATATCCGT
GGCTGTATGATGCAGCCGGTCTCCGAGCAAAGCACAGGAAAAAGTTCCGGAAGTTACCCGTCTGTGTTT
GTTGGTAGCGAAATTTGGTCAGGGTTATCCGGGTGATGAAGATAATGGTGAAATGTCTCTGGTGGATTTT
TGCAAGCCTGGGTTTTTATCCGCTGCAGGTTGGTAGCGATCAGTATGCAGTTGGTTCTCCGCTGTTTGATA
AAGCAACCGTTCATCTGCCGGATGGTGTGTTGTTAATGCCGAAAATAATAGCGTGGATAATGTGTAT
GTTTCAGAGCCTGGCAGTTGATGGTGAAGCACGTACCAGCACAGCCTGAGCCAGGCAGATCTGAGCGGTGG
CACCACCCTGGATTTTTGTTATGGGTCCGGAACCGAGCGATTGGGGCACCGGTGAAGATGATGCACCTCCGT
CACTGACCGAAGGTGATGAACCTCCGACACCGGTTTCAGGATGCAACCACCGCAGGCCTGGGCACCACCACC
GTTGCCGATGGTGTGCCACCACCTCTGCAGCAGCCCTGACCGATAATACCAGCGGCACCCGTACCACCTT
TGCAACCACCACCCGAGCATTACATGGGCAGGTAATGGCATTTCGTCCGACCGTTGGTAGCTATAACCTGA
CCTCTGGTGAAGCGGCACCGCAAGCCCGTCTGCATGGACCCTGGAAGGTTCTGATGATGGCGAAAACCTGG
ACCACACTGGATGAACGTAGCGGTGAACAGTTTCGTTGGGCACCTGCAGACCCGTCCGTTTACCGTTGCCGA
ACCGACCGCATTTGCACGTTATCGTGTACC GTTACC GC AACCAGCGGTTCTGGTGC ACTGAGCCTGGCAG
AAGTTGAACTGCTGGCAGATCCGAAAGAAAGCGGTGCAGAAGAACTGACCCTGTCTGCAGCACC GG ATCGT

GATGGCGTTACCGGTCGTGAAGTTAGCGGTTCTTTTGCAACCCTGACCGGTGTTGAAGGTGATGTTGCCGC
ACTGGATGTTTCAGGTTGCATTTGGTGTGGTAGCGAACCAGGTTGCAGGTACACTGCGTGCCGGTGCATTTG
GTGGTTATGCAGTTGATGCAGCACATACTGGACCGCACCGGGTGTATCCGGTTACCGTGACCGTTAGC
GGTGAAGGTATTGAAACCGTTAGCGCAAGCAGCTATGTTAGCGTTAGCCTGCTGCGTGAAGGTTCTCTGCT
GGCAGCATATGATAATGTGTGCATTGGTGTGCAGGTACAACCGTTGGTTCCTGTGATGGTCAGGGCGTTT
TTTTTGATCGTGCACAGCTGGCAGCAAAAGGTTTTGTGCAGGGTGAACGTGCAACCGTTCCGGGTACAGAT
CTGGCATTGATGTTCCGGCAGTTCGGGCTGGTCAGCCTGATAATGCAACCGGTGATGGTCAGACCATTGA
ACTGGATGTTCCGGCTGATGCAGAACAGCTGAGCGTTATTGGCACCGGCACCGAAAAAATCAGCAGGCAA
CCGGTACACTGACCTTTGATGATGGTTCTACCCAGCCGATTGATCTGAGCTTTGGTGTATTGGAGCGGTGCA
GCACGTAATCCGGTGTGGTAATATTCCGGTTGCAGTTACCGATAGCCGTCTGCGTGGTGGTTCCTCCGCA
GACCGGTACACCGGCAGCATTTTTTGCCACCGCACCGATTACCCTGCCGGAAGGTAAACGTCCGGTTAGCC
TGACCCTGCCGGATCAGCCTGGTGAAGTGAAGTGAAGTGAAGTGAAGTGAAGTGAAGTGAAGTGAAGTGAAGT
GGCACCTTGCAGAACATCCTGCACTGGAAGTGAAGTGAAGTGAAGTGAAGTGAAGTGAAGTGAAGTGAAGT
CTCAGATGTTGCACTGGCACAGGTTGCCGGTGGTGTGAGGTTGCAGATCTGCGTGCCGAGTTACCTGGG
GTGATGGTTCTGATGTGGCAGCCGGTGCCGTTACCGATGGTAGCGTTAGCGGTAGCCATGCATATACCGCA
GCAGGCACCTATACCGCATATGTTGTTGTGGATGATGGTTGGACCAGCCAGGTTGTTGAAGTTCCGGTGAC
CGTTACAGAAGCCGAACCGGCACCTGGCCGTTGATGTACCGTTAGCACCCGTTGCCTGGCAGGTAAAGCAT
ATGTTGCAGTGCCTGCAGAAAATGGTGAAGATGTTCCGCTGGCAATTCGTCTGGTTACCCGTTTGGCACC
AAAGAAGTTCAGCAGTTGCTCCGGGAGCCAATGCATATCAGAGCTTTGCAACCCGTGTTACCGCAGTTGA
AGCAGGCACCGTTACCGTTGAAGCCACCCGTGGCACCGGTGATGAAGAAGTTACCGCCAGCATTACGGCAG
ATTATGCAGCCGTTACCTGCGGTTAATAA (SEQ ID NO:19)

GATGGCGATTGGCTGGCATATGGCGAAATTGATCTGGGCAGCGCAGCACTGGATCAGCTGTCTGTGCATTA
TGTTTCATAATTCTAATCGCTCTGGTCGTAATTCTGCACTGTCTGTGTATCTGGATGCCTTTGATCCGGCAA
ATCCGGGTGAACCGTTTGTGACAGTGCCGCTGGCAAATACCGGTAGCTCTTGGACCACCGATGGTACTGCA
GTTGTGGATCTGCCGTCTACCGTTCGTGGTAAACATCAGGTTTGGGTTTCGTCTGTCTACCGAAGCATATGC
CGATCATCCGTATGTGGCCAATCTGGATTCTATGCGCTTTTTTACCGATGCATATGATGTTGAAGTTCCTC
CGACCGATACAGCAGCACTGGCAGCCGTTGTTGATGCAGCAGGTACACCGGAAGCAGAAATTGCACGTTAT
GGTCGTATTGATGCCCCGTGTTTTTACCCGTGAACTGGCAGCAGCACGTAGCGTTCTGGCCGATGCCGGTGC
AACACAGGCACAGGCAGATGAACGTGCTCGTCTGTTGTTGATGCAGCAGGTACACCGGAAGCAGAAATTGCACGTTAT
GTCGTCTGCTGGAAAATCTGGTTGCCAGCGCAGAAGCACTGACCGACGAAGGTTATTCTCCGAAAGCTGG
CAGGCATTTTCGTACCGCACTGGCTGCTGCAACCGGCACCCTGGATGATGCAGCAGCATCTGATGAAGCACT
GCATGATGCACGTCTGGCGCTGCAGGGTGCAGTTGATGCACTGGAAGAACCGGCAGATGTTGTTCTGGTTG
AAGTTGAAGTTTCTCCGCGTTGTCTGGCAGGTAAACCGTATGTTGCCGTTTCGTGCAGTTAATGTTTCTGAT
GCAGCCGTTGATGTTGAACTGGCAAGCTCTCTGGGCACCCGTAGCTTTGTTGGTGTGGCACCCGGGTGCGAG
CGCATATCAGAGCTTTCAGCCCGTAGCGCAACCGGTGATCTGGATGTTACCGTGACCGCAACCGGTGCGAG
ATGGTACTCAGACCGTTGAACAGGTTGTGACCGTTCCGAGCTGTAGCTAATAA (SEQ ID NO:20)

FIGURE 20

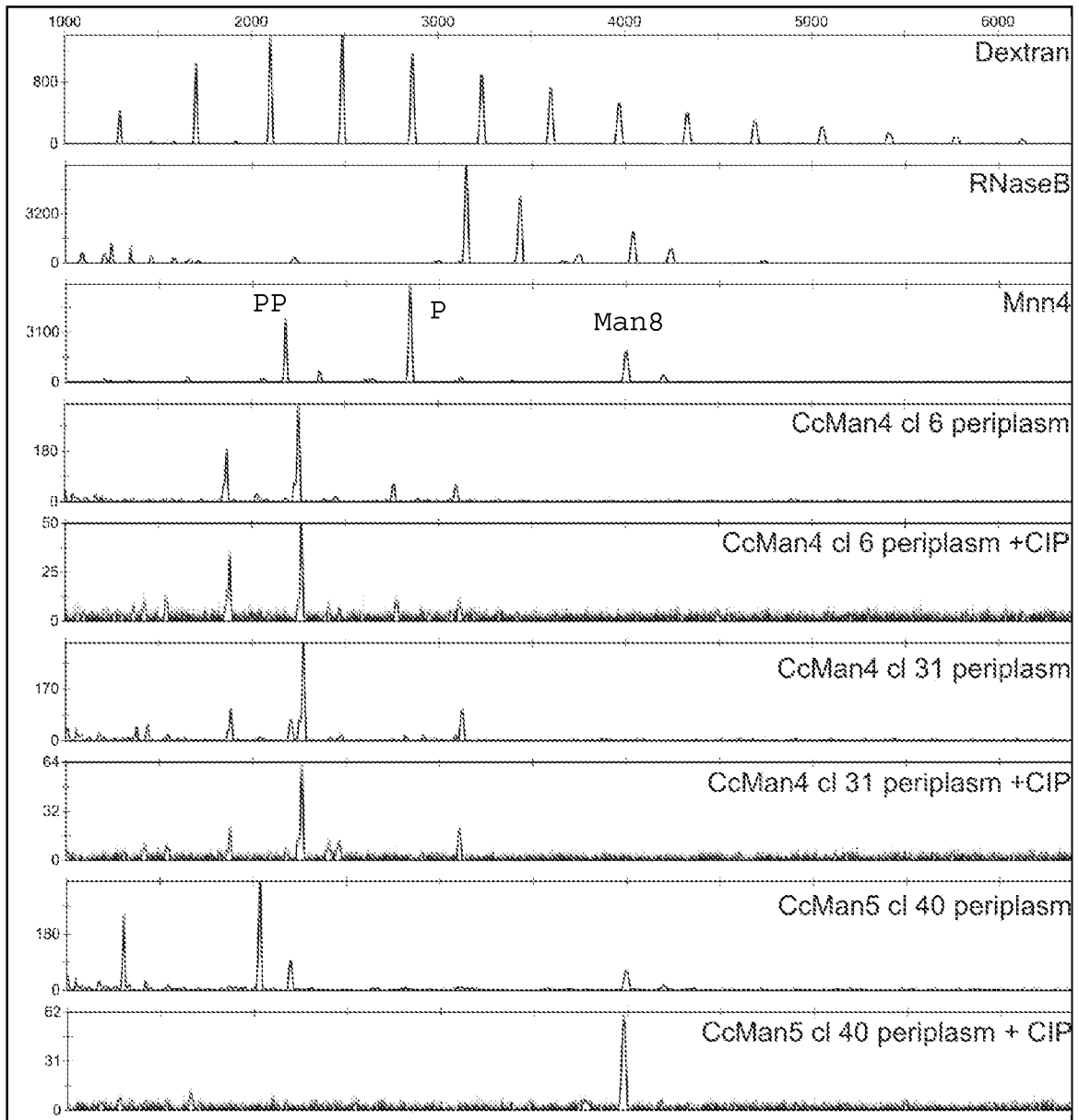


FIGURE 21

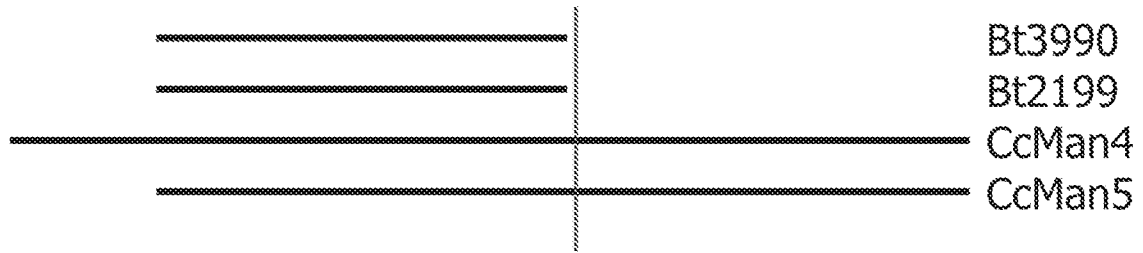


FIGURE 22

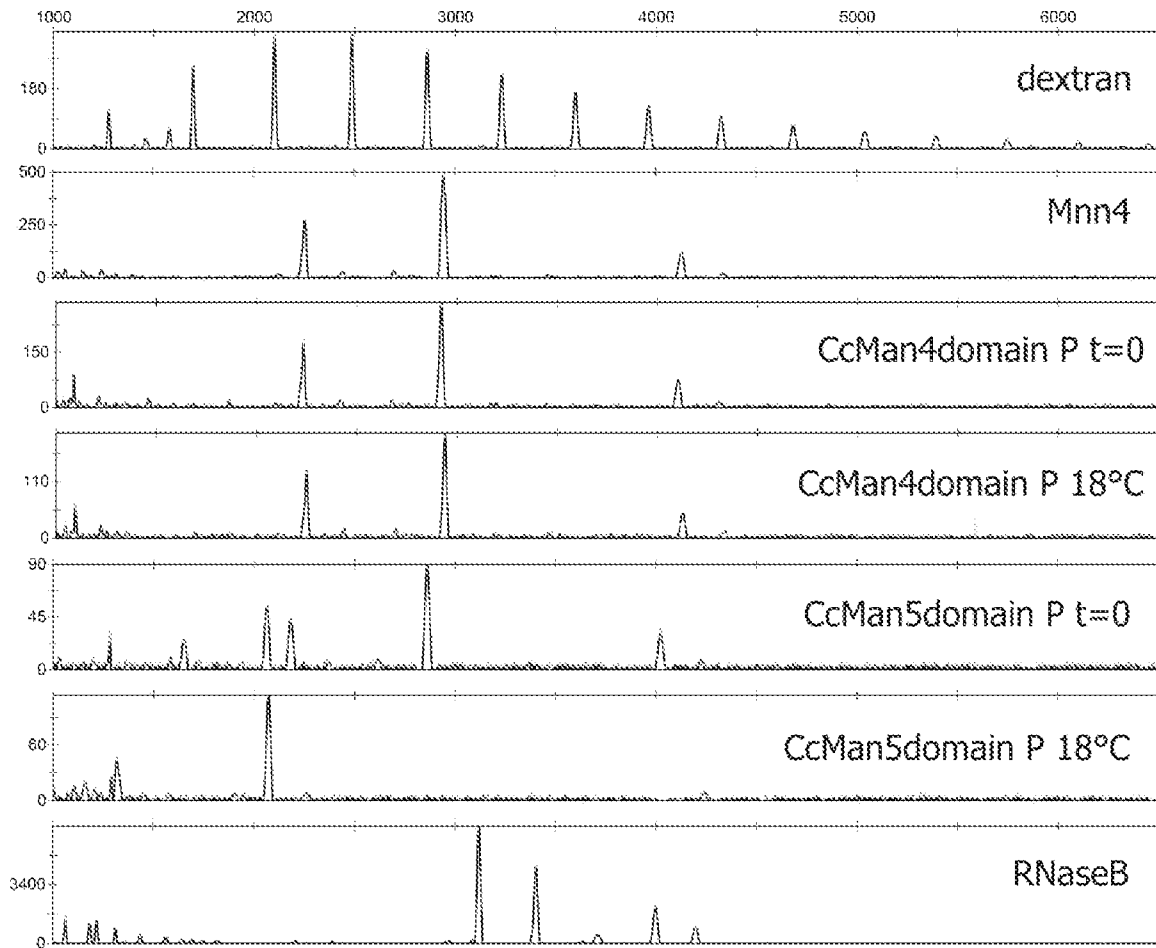


FIG. 23



FIG. 24

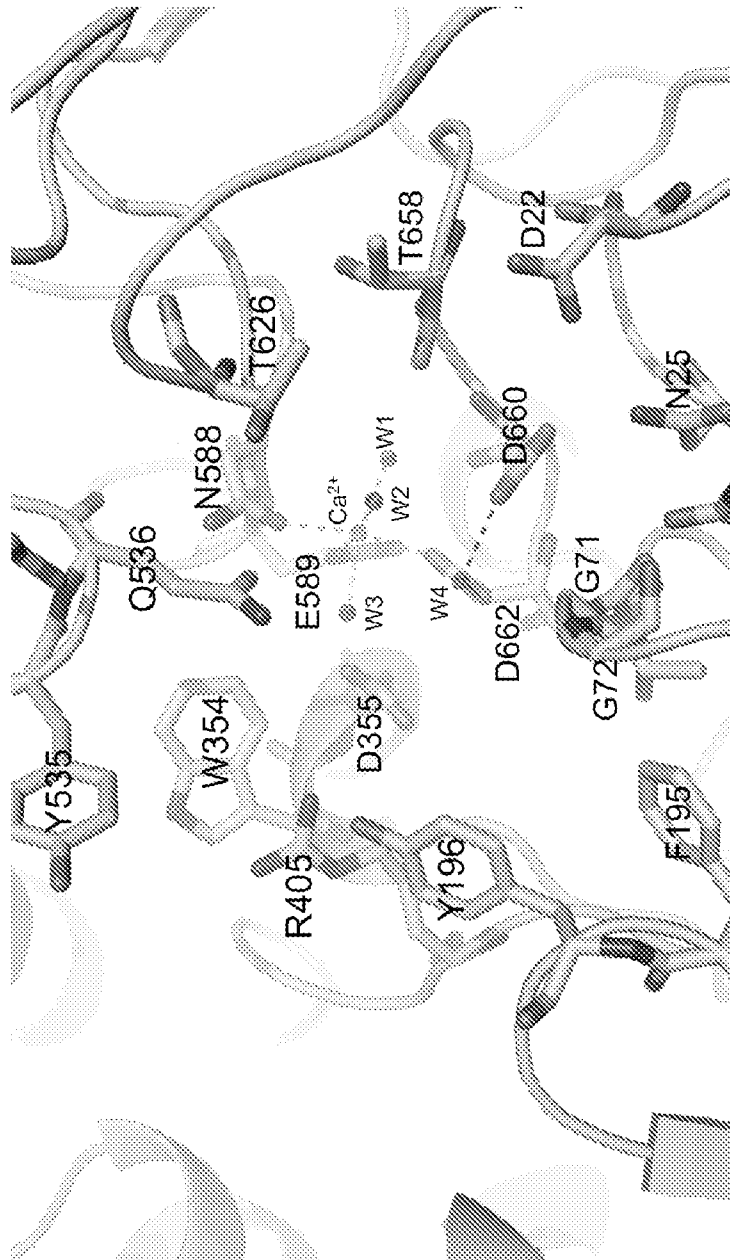


FIG. 25

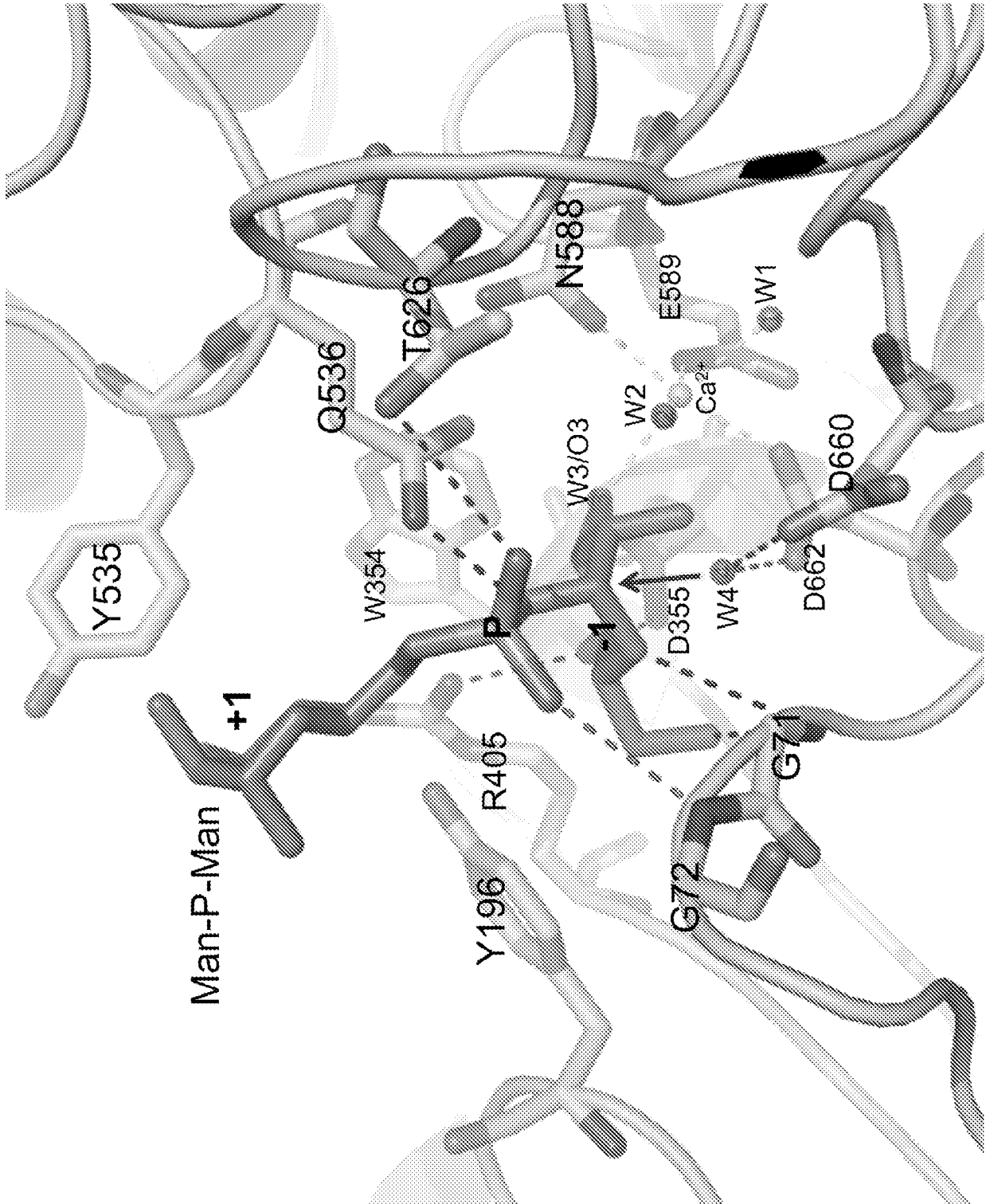


FIGURE 26A

ATGAAGCTTTCACCATCCTCTTCACAGCCTGCGCTACCTGGCCCTGGACAACGGCCTGGCCCGAACCC
CACCATGGGCTGGCTGCACTGGGAGCGATTTCATGTGTAACTGGACTGTCAGGAAGAGCCCGACTCTTGTA
TCTCTGAGAAGCTGTTTCATGGAAATGGCCGAGCTGATGGTGTCTGAGGGCTGGAAGGACGCCGGCTACGAG
TACCTGTGTATCGACGACTGTTGGATGGCCCCCAGCGAGACTCTGAGGGCCGACTCCAGGCCGACCCCA
GCGATTCCCCCACGGCATCCGACAGCTCGCCAACCTACGTGCACTCTAAGGGCCTGAAGCTGGGCATCTACG
CCGACGTGGGCAACAAGACCTGTGCCGGCTTCCCCGGCTCTTTCGGCTACTACGACATCGACGCCCGAGACC
TTCGCCGACTGGGGCGTGGACCTGCTGAAGTTCGACGGCTGTTACTGTGACTCTCTCGAGAACCTGGCCGA
CGGCTACAAGCACATGTCTCTGGCCCTGAACCGAACCGGCCGATCTATCGTGTACTCTTGTGAGTGGCCCC
TGTACATGTGGCCCTTCCAGAAGCCCAACTACACCGAGATCCGACAGTACTGTAACCACTGGCGAAACTTC
GCCGACATCGACGACTCGTGAAGTCTATCAAGTCTATTCTGGACTGGACCTCTTCAACCAGGAGCGAAT
CGTCGACGTCGCCGGACCCGGCGGATGGAACGACCCCGACATGCTGGTGTATCGGCAACTTCGGCCTGTCTT
GGAACCAGCAGGTGACCCAGATGGCCCTGTGGGCTATCATGGCTGCCCCCCTGTTTCATGTCTAACGACCTG
CGACACATCTCTCCCCAGGCCAAGGCCCTGCTCCAGGACAAGGACGTGATCGCCATCAACCAGGACCCCT
GGGCAAGCAGGGCTACCAGCTCCGACAGGGCGACAACCTTCGAGGTGTGGGAGCGACCCCTGTCTGGCCTGG
CCTGGGCCGTGGCCATGATCAACCGACAGGAGATCGGCGGACCCCGATCTTACACCATCGCCGTGGCCTCC
CTGGGAAAGGGCGTGGCCTGTAACCCCGCCTGTTTCATCACCCAGCTCCTGCCCCGTGAAGCGAAAGCTGGG
ATTCTACGAGTGGACCTCTCGACTGCGATCTCACATCAACCCACCGGCACCGTGCTGCTCCAGCTCGAGA
ACACCATGCAGATGTCTCTGAAGGACCTGCTGACGCGTGAACAAAACTCATCTCAGAAGAGGATCTGAAT
AGCGCCGTCGACCATCATCATCATCAT (SEQ ID NO:22)

FIGURE 26B

MKLSTILFTACATLALDNGLARTPTMGWLHWERFMCNLDCQEEDSCISEKLFMEMAELMVSEGWKDAGYE
YLCIDDCWMAPQRDSEGRLOADPQRFPHGIRQLANYVHSGKLGKIYADVGNKTCAGFPGSFGYYDIDAQT
FADWGVDLLKFDGCYCDSLENLADGYKHMSLALNRTGRSIVYSCEWPLYMWPFQKPNYTEIRQYCNHWRNF
ADIDDSWKSIXSILDWTSFNQERIVDVAGPGGWNDPMLVIGNFGLSWNQVVTQMALWAIMAAPLFMSNDL
RHISPQAKALLQDKDVIAINQDPLGKQGYQLRQGDNFEVWERPLSGLAWAVAMINRQEIGGPRSYTIAVAS
LGKGVACNPACFITQLLPVKKRKLGFYEWTSRLRSHINPTGTVLLQLENTMQMSLKDLLTREQKLISEEDLN
SAVDHHHHHH (SEQ ID NO:23)

FIGURE 27A

ATGAAGCTTTCCACCATCCTCTTCACAGCCTGCGCTACCCTGGCTGCCGCCAGCAGGGAGCCT
CTCGACCCGGACCCCGAGATGCCAGGCTCACCCCGGACGACCTCGAGCTGTGCCACCCAGTGTG
ACGTGCCCCCAACTCTCGATTGACTGTGCCCCGACAAGGCCATCACCCAGGAGCAGTGCAGAGG
CCCGAGGCTGTTGTTACATCCCCGTAAGCAGGGCCTGCAGGGCGCTCAGATGGGCCAGCCCTGGT
GTTTCTCCCCCCTCTTACCCCTCCTACAAGCTGGAGAACCTGTCCTCTTCGAGATGGGCTACAC
CGCCACCTGACCCGAACCACCCCACTTTTTCCCAAGGACATCCTGACCCTGCGACTGGACGTG
ATGATGGAGACCGAGAACCGACTGCACTTCACCATCAAGGACCCCGCCAACCGACGATACGAGGT
GCCCCCTGGAGACCCCCACGTGCACTCTCGAGCCCCCTCCCCCTGTACTCTGTGGAGTTCTCTGAG
GAGCCCTTCGGCGTGATCGTGCAGACAGCTGGACGGCCGAGTGTGCTGTAACACCACCGTGGCC
CCCCTGTTCTTCGCCGACCAGTTCCTGCAGCTGTCTACCTCTCTGCCCTCTCAGTACATCACCGCCT
GGCCGAGCACCTGTCCCCCTGATGCTGTCCACCTCTTGACTCGAATCACCTGTGGAACCGAGA
CCTGGCCCCACCCCGGTGCCAACCTGTACGGCTCTCACCCCTTCTACCTGGCCCTGGAGGACGGC
GGCTCTGCCACGGCGTGTCTGTGCTGAACTCTAACGCCATGGACGTGGTGTGTCAGCCCTCTCCCG
CCCTGTCTTGCGATCTACCGCGGCATCCTGGACGTGTACATCTTCTGGGCCCTGAGCCCAAGTC
TGTGGTCCAGCAGTACCTGGACGTGGTCCGATACCCCTTCATGCCCCCTACTGGGGCCTGGGCTTC
CACCTGTGTGATGGGGCTACTCTTCTACCGCCATCACCCGACAGGTGGTGGAGAACATGACCCGA
GCCACTTCCCCCTGGACGTGCAATGGAACGACCTGGACTACATGGACTCTCGACGAGACTTCACC
TTCAACAAGGACGGCTTCCGAGACTTCCCCGCCATGGTCCAGGAGCTGCACCAGGGAGGACGACG
ATACATGATGATCGTGGACCCCGCCATCTCTTCTTCGGACCCGCGGATCTTACCGACCCTACGAC
GAGGGCCTGCGACGAGGCGTGTTCATACCAACGAGACCGGCCAGCCCTGATCGGCAAGGTGTG
GCCCCGCTCTACCGCCTTCCCCGACTTCACCAACCCACCGCCCTGGCTTGGTGGGAGGACATGGT
GGCCGAGTTCACGACCAGGTGCCCTTCGACGGCATGTGGATCGACATGAACGAGCCCTCTAACTT
CATCCGAGGCTCTGAGGACGGCTGTCCCAACAACGAGCTGGAGAACCCCCCTACGTGCCCGGCGT
GGTGGGCGGAACCTGCAGGCCGCCACCATCTGTGCCTCTTCGCACCAGTTTCTGTCTACCCACTAC
AACCTGCACAACCTGTACGGACTGACCGAGGCCATTGCCTCTCACCGAGCCCTGGTGAAGGCCCGA
GGCACCCGACCCTTCGTGATCTCTCGATCTACCTTCGCCGGCCACGGCCGATACGCCGGACACTGG
ACCGGCGATGTGTGGTCTCTTGGGAGCAGCTGGCCTCTTCTGTGCCCGAGATCCTGCAGTTCAACC
TGCTGGGCGTGGCCCTGGTGGGCGCCGACGTGTGTGGCTTCTGGGCAACACCTCTGAGGAGCTGT
GTGTTGATGGACCCAGCTCGGCGCCTTCTACCCCTTCATGCGAAACCACAACCTCCCTGCTGTCTCT
GCCCCAGGAGCCCTACTCGTTCTCTGAGCCCGCTCAGCAGGCCATGCGAAAGGCTCTGACCCTGGC
ATACGCCCTGCTGCCCCACCTGTACACCCTGTTCCACCAGGCCACGTGGCTGGAGAGACCGTGGC
CCGACCCCTGTTCCCTGGAGTTCCTAAGGACTCTTCTACCTGGACCGTGGACCATCAGCTGCTGTGG
GGCGAGGCCCTCTGATCACCCCGTGTGTCAGGCCGGCAAGGCTGAGGTGACCGGCTACTTCCCT
CTGGGCACCTGGTACGACCTGCAGACCGTGCCTGTGGAGGCCCTGGGATCTCTGCCCCCTCCTCCCG
CCGCTCCCCGAGAGCCCGCCATCCACTCTGAGGGCCAGTGGGTGACCCTGCCGCTCCCCTGGACA
CCATCAACGTGCACCTGCGAGCCGGCTACATCATCCCTCTGCAGGGACCCGGCCTGACCACCACCG
AGTCTCGACAGCAGCCCATGGCCCTGGCCGTGGCTCTGACCAAGGGCGGAGAGGCCCGAGGCGAG
CTGTTCTGGGACGATGGCGAGTCTCTGGAGGTGCTGGAGCGAGGCGCTACACCCAGGTGATCTTT
CTGGCCCGAAACAACACCATCGTGAACGAGCTGGTGCAGTGACCTCTGAGGGCGCTGGTCTGCAG
CTCCAGAAGGTGACCGTCCCTGGGCGTGGCCACCGCTCCCCAGCAGGTCTGTCTAACGGCGTGCC
GTGTCTAACTCACCTACTCTCCCGACACCAAGGTGCTGGACATCTGTGTGTCTCTGCTGATGGGCG
AGCAGTTCCTGGTGTCTTGGTGTAAAC (SEQ ID NO:24)

FIGURE 27B

MKLSTILFTACATLAAAQQGASRPGPRDAQAHPGRPRAVPTQCDVPPNSRFDCAPDK
AITQEQCEARGCCYIPAKQGLQGAQMGPWCFFPPSYPSYKLENLSSSEMGYTATLTRT
TPTFFPKDILTLRLDVMMETENRLHFTIKDPANRRYEVPLETPHVHSRAPSPLYSEFSE
EPFGVIVRRQLDGRVLLNTTVAPLFFADQFLQLSTSLPSQYITGLAEHLSPLMLSTSWTR
ITLWNRDLAPTPGANLYGSHPFYLALEDGGSAGHVFLLSNAMDVVLQPSPALSWRST
GGILDVYIFLGPEPKSVVQQYLDVVGYPFMPYWGLGFHLRCRWGYSSTAITRQVVENM
TRAHFPLDVQWNDLDYMSRRDFTFNKDGFRDFPAMVQELHQGRRYMMIVDPAISS
SGPAGSYRPYDEGLRRGVFITNETGQPLIGKVVWPGSTAFPDTNPTALAWWEDMVAEF
HDQVPFDGMWIDMNEPSNFIRGSEDGCPNNELENPPYVPGVVGGLQAATICASSHQF
LSTHYNLHNLGLTEAIAISHRALVKARGTRPFVISRSTFAGHGRYAGHWTGDVWSSWE
QLASSVPEILQFNLLGVPLVGADVCGFLGNTSEELCVRWTQLGAFYPFMRNHNSLLSLP
QEPYSFSEPAQQAMRKALTLRYALLPHLYTLFHQAHVAGETVARPLFLEFPKDSSTWT
VDHQLLWGEALLITPVLQAGKAEVTGYFPLGTWYDLQTPVEALGSLPPPPAAPREPAI
HSEGQWVTLPAPLDTINVHLRAGYIPLQGPGLTTESRQQMALAVALTKGGEARGEL
FWDDGESLEVLERGAYTQVIFLARNNTIVNELVRVTSEGAGLQLQKVTVLGVATAPQQ
VLSNGVPVSNFTYSPDTKVLDCVSLLMGEQFLVSWC* (SEQ ID NO:25)

FIGURE 28

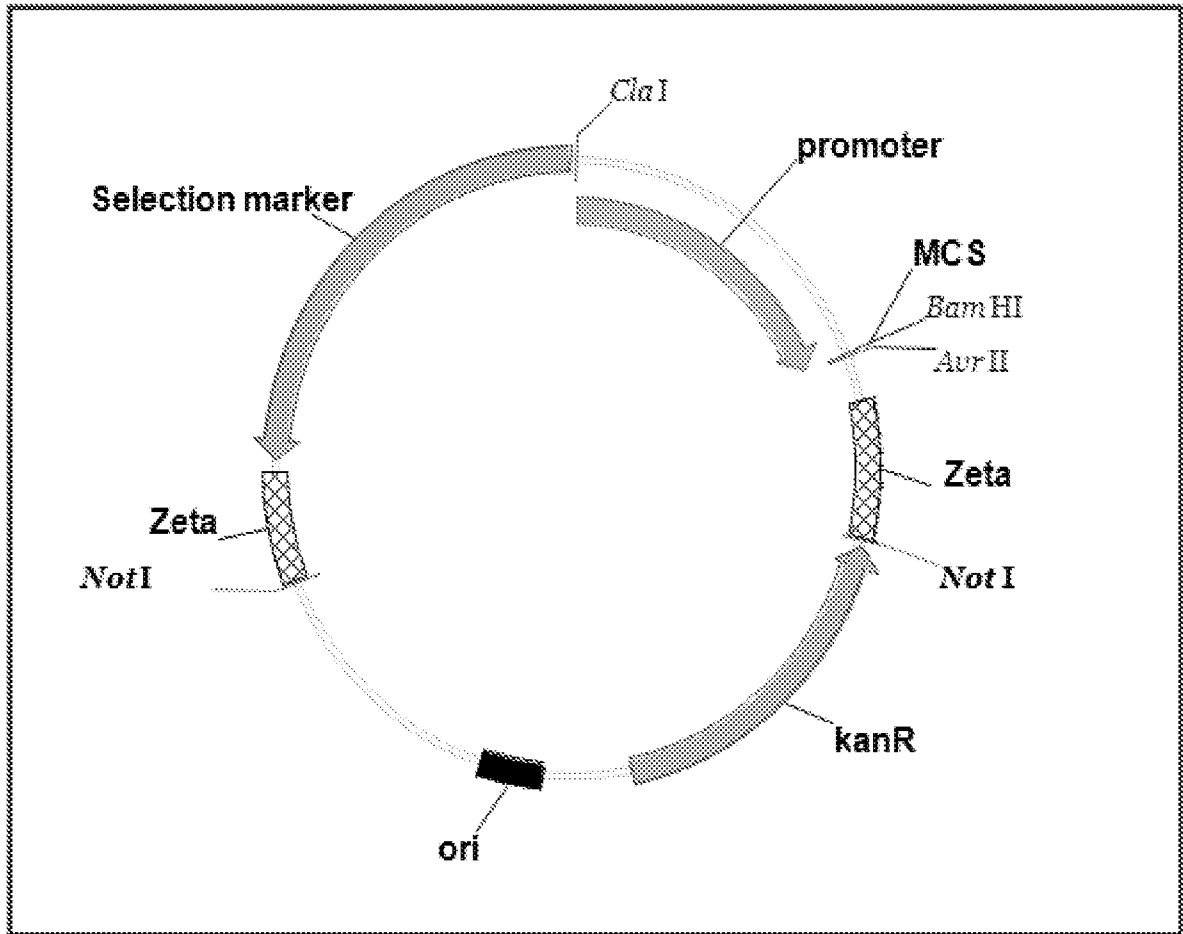


FIGURE 29

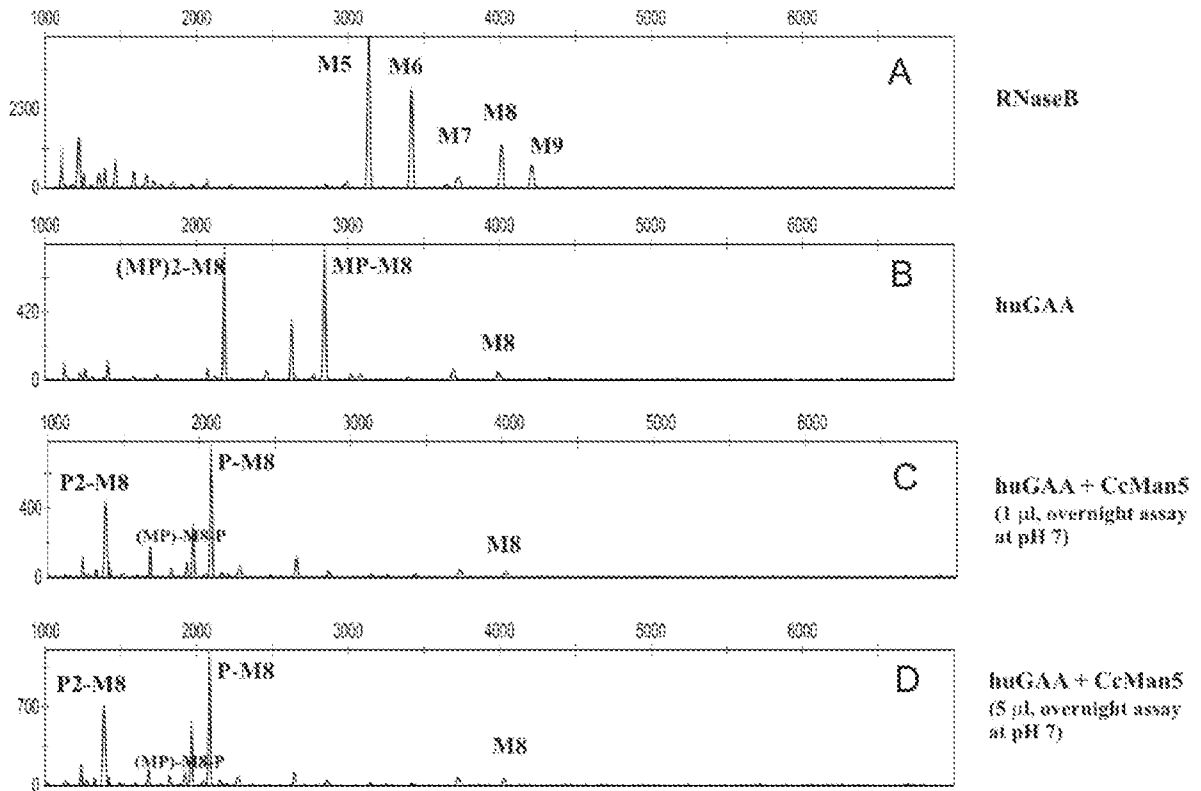
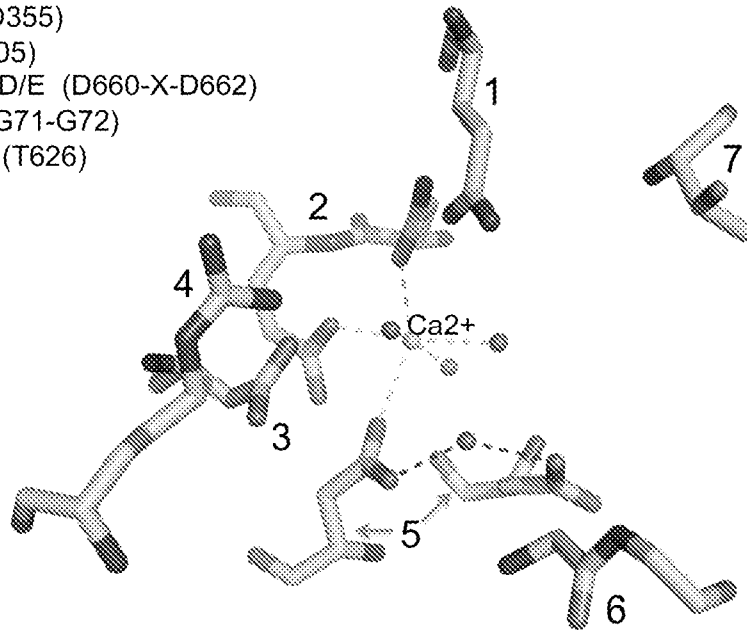


FIGURE 30

- 1: Q (Q536)
- 2: N/D-E/Q (N588-Q589)
- 3: D/E (D355)
- 4: R (R405)
- 5: D/E-X-D/E (D660-X-D662)
- 6: G-G (G71-G72)
- 7: T/S/G (T626)



Numbering for equivalent residues in SEC ID NO:50 is given in parenthesis

	10	20	30	40	50	60	70	84
CcMan5 full	(1)	1						Section 1
	(1)							84
NP_630514 Streptomyces	(1)							
ZP_02866543 Clostridium	(1)							
NP_812442 Bacteroides	(1)							
YP_003584502 Zunongwangia	(1)							
YP_003120664 Chitinophaga	(1)							
AAK22560 Caulobacter	(1)							
ACL94075 Caulobacter	(1)							
ACT03290 Paenibacillus	(1)							
ACU59240 Chitinophaga	(1)							
ACU05553 Pedobacter	(1)							
	(85)	85						
CcMan5 full	(1)							Section 2
	(1)							168
NP_630514 Streptomyces	(1)							
ZP_02866543 Clostridium	(1)							
NP_812442 Bacteroides	(1)							
YP_003584502 Zunongwangia	(1)							
YP_003120664 Chitinophaga	(1)							
AAK22560 Caulobacter	(1)							
ACL94075 Caulobacter	(1)							
ACT03290 Paenibacillus	(1)							
ACU59240 Chitinophaga	(85)							
ACU05553 Pedobacter	(1)							
	(1)							

FIG. 31

Section 5

(337) 337 CcMan5 full 350 360 370 380 390 400 410 420
 (151) 337 CcMan5 full 350 360 370 380 390 400 410 420
 (195) NP_630514 Streptomyces 350 360 370 380 390 400 410 420
 (187) ZP_02866543 Clostridium 350 360 370 380 390 400 410 420
 (127) NP_812442 Bacteroides 350 360 370 380 390 400 410 420
 (154) YP_003584502 Zunongwangia 350 360 370 380 390 400 410 420
 (141) YP_003120664 Chitinophaga 350 360 370 380 390 400 410 420
 (163) AAK22560 Caulobacter 350 360 370 380 390 400 410 420
 (163) ACL94075 Caulobacter 350 360 370 380 390 400 410 420
 (317) ACT03290 Paenibacillus 350 360 370 380 390 400 410 420
 (150) ACU59240 Chitinophaga 350 360 370 380 390 400 410 420
 (142) ACU05553 Pedobacter 350 360 370 380 390 400 410 420

Section 6

(421) 421 CcMan5 full 430 440 450 460 470 480 490 504
 (229) 421 CcMan5 full 430 440 450 460 470 480 490 504
 (274) NP_630514 Streptomyces 430 440 450 460 470 480 490 504
 (270) ZP_02866543 Clostridium 430 440 450 460 470 480 490 504
 (191) NP_812442 Bacteroides 430 440 450 460 470 480 490 504
 (214) YP_003584502 Zunongwangia 430 440 450 460 470 480 490 504
 (201) YP_003120664 Chitinophaga 430 440 450 460 470 480 490 504
 (245) AAK22560 Caulobacter 430 440 450 460 470 480 490 504
 (245) ACL94075 Caulobacter 430 440 450 460 470 480 490 504
 (395) ACT03290 Paenibacillus 430 440 450 460 470 480 490 504
 (213) ACU59240 Chitinophaga 430 440 450 460 470 480 490 504
 (205) ACU05553 Pedobacter 430 440 450 460 470 480 490 504

Section 9
 (673) 673 SADELRRGYVAGNPGA 690 QRGYQYG 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756

(485) NP_630514 Streptomyces TGTLRQGYIAGDPGT 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (478) ZP_02866543 Clostridium TSEVEKIGYVPNRVD 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (382) NP_812442 Bacteroides MKQEMERLPMRSPDQ 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (402) YP_003584502 Zunongwangia LKDAESLDYRAPHDK 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (387) YP_003120664 Chitinophaga VEKIKGLDYAHDPKA 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (459) AAK22560 Caulobacter DRSPENNRQAKSGSS 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (459) ACL94075 Caulobacter DRSPENNRQAKSGSS 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (596) ACT03290 Paenibacillus AVDADNFSISDGEISG 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (439) ACU59240 Chitinophaga TVKEVDDFERRQAVAV 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756
 (431) ACU05553 Pedobacter SVNEVHDFEKRQSVAV 700 VDAEGLTE 710 ETLRQA 720 SWPIEKLTK 730 PGAWTAADGTQVGLLTP 740 RAADGWSWQS 756

Section 10
 (757) 757 ADHAKFEAA 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840

(524) ACU05553 Pedobacter ADHAKFEAA 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 Region: Misc(534..538): LYOGT motif
 (567) NP_630514 Streptomyces VYEKFEAR 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (559) ZP_02866543 Clostridium NQDPSSTNT 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (445) NP_812442 Bacteroides AFEVMKNN 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (465) YP_003584502 Zunongwangia DVDRMQAR 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (450) YP_003120664 Chitinophaga DVDRMQAR 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (543) AAK22560 Caulobacter WDGKVHNNV 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (543) ACL94075 Caulobacter WDGKVHNNV 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (673) ACT03290 Paenibacillus AGDIQAVDRYA 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (519) ACU59240 Chitinophaga ---GQGGRDYFTENNA 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840
 (511) ACU05553 Pedobacter ---GQGGREYFTENNA 770 HMYDA 780 HMYDA 790 HMYDA 800 HMYDA 810 HMYDA 820 HMYDA 830 HMYDA 840

Section 13

(1009) 1009
 (763) APAFSMSTATDEPAEGPR-----VSAEPTTVQTDGGALDAVTLTLDG-ARLAAPAGTDLVTSGAASVVGLPDGVTAAVTV 1080 1092
 CcMan5 full 1020 1030 1040 1050 1060 1070 1080 1092

(807) EPAYSLNTDSGDDEHAPGRGTTVV SARPETVR TAADGTV DASVELRLS GRASFAARKG TSLTRTGAASVTGLPDGVTADLRV
 ZP_02866543 Clostridium (787) TAKSSDNADTSTYDDDEIAYSSAMFEESKANDGSFDQKITITLTKTKEFAGEIGEDLVATGKINITNIPEGLEASAIKTEANKV
 NP_812442 Bacteroides (654) -----
 YP_003584502 Zunongwangia (687) AWVSSIRQYL-----
 YP_003120664 Chitinophaga (672) KWIADITRQ-----
 AAK22560 Caulobacter (740) VLTVRADQPSLVDGKTSTGWR AASGQSVTFLSKAPACIAAYS SVSGPDQADPSHWTLQAYDGR AWVSVDQSNVVFDRHRHATRT
 ACL94075 Caulobacter (740) VLTVRADQPSLVDGKTSTGWR AASGQSVTFLSKAPACIAAYS SVSGPDQADPSHWTLQAYDGR AWVSVDQSNVVFDRHRHATRT
 ACT03290 Paenibacillus (895) SAAPTVDYNADMNDNFNHEQLIPEKSTWKYDDKGEAGEGWTQVDFDDSSWSSGKAMLG YDSYGKPA TTVSYGPNANNKYVTT
 ACU59240 Chitinophaga (737) QAA PPSLLNYS PAGK-----
 ACU05553 Pedobacter (729) -----

Section 14

(1093) 1093
 (839) ASPTALTVSLTGTASADARFFVHLRDAALADGVAAASLQGGQVSVRSPLRLSVASAERDALAALVDDAVLVRHGNYSVTFDRF 1160 1176
 CcMan5 full 1100 1110 1120 1130 1140 1150 1160 1176

(891) TGKRTASLRLTGTTRTDARFGITFRDRAPPHGIPASTVTGTGVSVDPLIVSAAA VHRGSLAALVDEASLVR EGNYS DGSYGIF
 ZP_02866543 Clostridium (871) EVSLNGKAKNHTLND S ISNLTIEITDGATNEPIKDSIRKTKDNV KVMFIDNQLTYSQSEFKES ESDDGAIL ETSTITLTGDTTF
 NP_812442 Bacteroides (654) -----
 YP_003584502 Zunongwangia (697) -----
 YP_003120664 Chitinophaga (681) -----
 AAK22560 Caulobacter (824) FPLAPGRYARLRWVLDGGSEASVSEVELIAGASCAAPTSGAPLL-----
 ACL94075 Caulobacter (824) FPLAPGRYARLRWVLDGGSEASVSEVELIAGASCAAPTSGAPLL-----
 ACT03290 Paenibacillus (979) YFRKTFDAKDLGDGILELDSLRDDGAI VYLNHGHEI FRTNMPTGAVNYSTFANATV GDERDKNGFIIDPSYLVEGKNVLTAEVH
 ACU59240 Chitinophaga (752) -----
 ACU05553 Pedobacter (729) -----

	2353	2360	2370	2380	2390	2400	2410	2420	Section 29
CcMan5 full (1651)	(2353)	2360	2370	2380	2390	2400	2410	2420	2436
NP_630514 Streptomyces (1987)	DLAPSI	LARTSEY	GDNRIVL	GFHYPLD	VMGGRITA	QAATVAHR	WADPEFAK	LLGQAHT	EIENVLLAR
ZP_02866543 Clostridium (1985)	NP_812442	Bacteroides (654)	NP_003584502	Zunongwangia (697)	YP_003120664	Chitinophaga (681)	AAK22560	Caulobacter (868)	ACL94075
YP_003584502 Zunongwangia (697)	ACT03290	Paenibacillus (1938)	ACU59240	Chitinophaga (752)	ACU05553	Pedobacter (729)			
NP_630514 Streptomyces (2071)	AGLSTA	QQVDRYT	QRLLTYG	FSRTGEAG	QALDAPSD	AAAALLI	TAFPDLT	AEQRAQV	LEQTATD
ZP_02866543 Clostridium (1985)	NP_812442	Bacteroides (654)	NP_003584502	Zunongwangia (697)	YP_003120664	Chitinophaga (681)	AAK22560	Caulobacter (868)	ACL94075
YP_003584502 Zunongwangia (697)	ACT03290	Paenibacillus (1938)	ACU59240	Chitinophaga (752)	ACU05553	Pedobacter (729)			
CcMan5 full (1651)	(2437)	2437	2450	2460	2470	2480	2490	2500	Section 30
CcMan5 full (1651)	(2437)	2437	2450	2460	2470	2480	2490	2500	2510
NP_630514 Streptomyces (2071)	AGLSTA	QQVDRYT	QRLLTYG	FSRTGEAG	QALDAPSD	AAAALLI	TAFPDLT	AEQRAQV	LEQTATD
ZP_02866543 Clostridium (1985)	NP_812442	Bacteroides (654)	NP_003584502	Zunongwangia (697)	YP_003120664	Chitinophaga (681)	AAK22560	Caulobacter (868)	ACL94075
YP_003584502 Zunongwangia (697)	ACT03290	Paenibacillus (1938)	ACU59240	Chitinophaga (752)	ACU05553	Pedobacter (729)			
NP_630514 Streptomyces (2071)	AGLSTA	QQVDRYT	QRLLTYG	FSRTGEAG	QALDAPSD	AAAALLI	TAFPDLT	AEQRAQV	LEQTATD
ZP_02866543 Clostridium (1985)	NP_812442	Bacteroides (654)	NP_003584502	Zunongwangia (697)	YP_003120664	Chitinophaga (681)	AAK22560	Caulobacter (868)	ACL94075
YP_003584502 Zunongwangia (697)	ACT03290	Paenibacillus (1938)	ACU59240	Chitinophaga (752)	ACU05553	Pedobacter (729)			
CcMan5 full (1651)	(2437)	2437	2450	2460	2470	2480	2490	2500	2510
CcMan5 full (1651)	(2437)	2437	2450	2460	2470	2480	2490	2500	2520
NP_630514 Streptomyces (2071)	AGLSTA	QQVDRYT	QRLLTYG	FSRTGEAG	QALDAPSD	AAAALLI	TAFPDLT	AEQRAQV	LEQTATD
ZP_02866543 Clostridium (1985)	NP_812442	Bacteroides (654)	NP_003584502	Zunongwangia (697)	YP_003120664	Chitinophaga (681)	AAK22560	Caulobacter (868)	ACL94075
YP_003584502 Zunongwangia (697)	ACT03290	Paenibacillus (1938)	ACU59240	Chitinophaga (752)	ACU05553	Pedobacter (729)			

	1	10	20	30	40	50	60	70	82
CcMan5 full	(1)								
	(1)								
NP_630514 Streptomyces	(1)								
ZP_05522540 Streptomyces	(1)								
ZP_06527366 Streptomyces	(1)								
YP_003013376 Paenibacillus	(1)								
NP_812442 Bacteroides	(1)								
ZP_04848482 Bacteroides	(1)								
ZP_03677957 Bacteroides	(1)								
YP_003584502 Zunongwangia	(1)								
ZP_01061975 Leeuwenhoekella	(1)								
ZP_07083984 Sphingobacterium	(1)								
YP_003120664 Chitinophaga	(1)								
ZP_01885202 Pedobacter	(1)								
ZP_02866543 Clostridium	(1)								
XP_367221 Magnaporthe	(1)								
ZP_07042437 Bacteroides	(1)								
ZP_05759807 Bacteroides	(1)								
ZP_05287524 Bacteroides	(1)								
ZP_06076108 Bacteroides	(1)								
YP_001302992 Parabacteroides	(1)								

FIG. 32

Section 3

246

230

220

210

200

190

180

170

165

(165)

(1) -----APEPPS-----ADYASLVDVGGTEGDFGDMPAAQENGLAKVNRPTTPG--RNN--AAQSKTSS

(37) TGGGTAVAVPVTKASPPAGERSGGTDYTKLDPFSTLGGDLGAAQPHSAXNMMITPG--RNR--NEDH

(40) TGGGTAVAVPVTKASPPAGERSGGTDYTKLDPFSTLGGDLGAAQPHSAXNMMITPG--RNR--NEDH

(12) TGGGTAVAVPVTKASPPAGERSGGTDYTKLDPFSTLGGDLGAAQPHSAXNMMITPG--RNR--NEDH

(165) IARIYELBYGVVEADQIPAEPTNLAIDYDPPNLLGNGVAAGAAAGGCGDNDP--RNR--YENKN

(1) -----MTPSVAQN---TKY---TKY---MVQAQQTSPVPGGATDRAATIGCGKPGETIP--RNG--OSQC

(4) NRSRWYALLLVVLISIMTPSVAQN---TKY---TKY---MVQAQQTSPVPGGATDRAATIGCGKPGETIP--RNG--OSQC

(1) -----MVQAQQTSPVPGGATDRAATIGCGKPGETIP--RNG--OSQC

(8) MIAAIAVALCVACQPKSQEKTSKANITDQVVFVGGGMSRSTANN--GHTNP--HNR--YAKQFD

(4) YIAVLFLLVLAFTSVAQES-----LTDYVVFVGGGMSRSTANN--GHTNP--HNR--YAKQFD

(15) FLSQFSVCLLILSSQQFVYAQRS---SLXVVFVGGGMSRSTANN--GHTNP--HNR--YAKQFD

(1) -MKKSLIYPLSLLSLTAVAQSQG---VVFVGGGMSRSTANN--GHTNP--HNR--YAKQFD

(8) NLRLLAFPLLELVNKTIIYAQQTCKD---KLVVFVGGGMSRSTANN--GHTNP--HNR--YAKQFD

(15) FTVAFSTLAVTTSRKSVAVEQEESYQYDPECDVPLFFSVVNGKSDDTYPHNTLDBA--SKLQIQ

(1) -----MAIHIFFLLSSLVSLAQAEELADYVFNITGEGGVYIYANIHLLPFTIQLPN--SMLRVPERADYTTEL

(1) -MKILHFCAAITMAAMLSGCNGGQ-----SQTANRTPVVPYIYANIHLLPFTIQLPN--SMLRVPERADYTTEL

(2) KTKLKTSMALLASAFLWVSCAGGG-----TPPSAMDPVVPYIYANIHLLPFTIQLPN--SMLRVPERADYTTEL

(1) -----MASCVYSPP-----KEPVPVYIYANIHLLPFTIQLPN--SMLRVPERADYTTEL

(2) KTKLKTSMALLASVFLWVSCAGGG-----TSPSAMDPVVPYIYANIHLLPFTIQLPN--SMLRVPERADYTTEL

- CcMan5 full
- NP_630514 Streptomyces
- ZP_05222540 Streptomyces
- ZP_06527366 Streptomyces
- YP_003013376 Paenibacillus
- NP_812442 Bacteroides
- ZP_04848482 Bacteroides
- ZP_03677957 Bacteroides
- YP_003584502 Zunongwangia
- ZP_01061975 Leeuwenhoeckiella
- ZP_07083984 Sphingobacterium
- YP_003120664 Chitinophaga
- ZP_01885202 Pedobacter
- ZP_02866543 Clostridium
- XP_367221 Magnaporthe
- ZP_07042437 Bacteroides
- ZP_05759807 Bacteroides
- ZP_05287524 Bacteroides
- ZP_06076108 Bacteroides
- YP_001302992 Parabacteroides

Section 4

	247	260	270	280	290	300	310	328
CcMan5 full	(247) THTN	GVGSGGSD	VPTS	PGTGYAHP	FSD	VS	GLGN	VAGTD - - - - - GAITGAPGTIE
	(63)	Region: Misc(69-77): glycine-rich motif						
NP_630514 Streptomyces	(117) TATN	GVGSGGSD	VPTSQQYDKR	PATSTYAHPP	DESH	SR	VLG	GLGSPS - - - - - GTID
ZP_05522540 Streptomyces	(120) TATN	GVGSGGSD	VPTSQQYDKR	PATSTYAHPP	DESH	SR	VLG	GLGSPS - - - - - GTID
ZP_06527366 Streptomyces	(92) TATN	GVGSGGSD	VPTSQQYDKR	PATSTYAHPP	DESH	SR	VLG	GLGSPS - - - - - GTID
YP_003013376 Paenibacillus	(244) SHLR	GVGSGGSD	MPETRDFTKN	VADYKQKYD	SEQA	AV	CVT	LASGIN - - - - - VQ
NP_812442 Bacteroides	(61) SVNR	GVGSGGSD	RPVAP	QELHIK	REK	FG	ST	AFTNGIK - - - - - T
ZP_04848482 Bacteroides	(79) SVNR	GVGSGGSD	RPVAP	QELHIK	REK	FG	ST	AFTNGIK - - - - - T
ZP_03677957 Bacteroides	(59) SQTR	GVGSGGSD	TPFV	TACKTKMD	ASEFH	FG	SV	LDMLK - - - - - V
YP_003584502 Zunongwangia	(87) THTH	GVGSGGSD	KPILN	DNKETEELR	VTEH	FG	EV	SEFNGID - - - - - V
ZP_01061975 Leeuwenhoekeiella	(76) THTR	GVGSGGSD	KPILN	AEVSTKLI	KTOD	FG	HV	NFENGID - - - - - V
ZP_07083984 Sphingobacterium	(91) THNR	GVGSGGSD	KPFLG	QDDQPLL	VTQ	FG	VE	IGLKNRIK - - - - - AF
YP_003120664 Chitinophaga	(73) THNR	GVGSGGSD	KPFLG	GPVKADLI	YEEQ	FG	HW	GFTNGIK - - - - - SF
ZP_01885202 Pedobacter	(84) THNR	GVGSGGSD	KPFLCA	ENDELPLV	ASET	FG	DL	AFTNGIK - - - - - GF
ZP_02866543 Clostridium	(97) SHTR	GVGSGGSD	TPTYVEYSQR	PQAQTRANN	YTD	FG	SV	ELTPKTGKDNVVKDSPEIGKIK
XP_367221 Magnaporthe	(76) SMLH	GVGSGGSD	VQMPV	LDDYLAARD	APMTEV	FG	KS	RLATGIT - - - - - V
ZP_07042437 Bacteroides	(74) LNGLPLI	GVGSGGSD	SPYQSGE	LQPIIT	YDNEHL	YS	EV	LDNDSMK - - - - - SY
ZP_05759807 Bacteroides	(74) LNGLPLI	GVGSGGSD	SPYQSGE	LRPIIT	YDNEHL	YS	EV	LDNDSMK - - - - - SY
ZP_05287524 Bacteroides	(77) LGGLPII	GVGSGGSD	CPYQSGESG	LRPVIA	SYDREKIL	YR	QV	LDNGEID - - - - - VF
ZP_06076108 Bacteroides	(56) YLADRI	GVGSGGSD	SPYQSGESG	SWIMATTGAPRINPNDYASG	DDHDFEKY	YS	SW	LLEDYDIE - - - - - VF
YP_001302992 Parabacteroides	(77) LGGLPII	GVGSGGSD	CPYQSGESG	LRPVIA	SYDREKIL	YR	QV	LDNMEED - - - - - VF

	329	330	340	350	360	370	380	390	400	410
CcMan5 full	(329) AAAAT	ETNNTSR	ETNNTSR	ETNNTSR	ETNNTSR	ETNNTSR	ETNNTSR	ETNNTSR	ETNNTSR	ETNNTSR
NP_630514 Streptomyces	TAAT	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ
ZP_05522540 Streptomyces	TAAT	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ
ZP_06527366 Streptomyces	TAAT	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ	AKAR-PQ
YP_003013376 Paenibacillus	TSSD	DTAN-TGS	DTAN-TGS	DTAN-TGS	DTAN-TGS	DTAN-TGS	DTAN-TGS	DTAN-TGS	DTAN-TGS	DTAN-TGS
NP_812442 Bacteroides	TATNAMA	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW
ZP_04848482 Bacteroides	TATNAMA	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW	RSLA-AALW
ZP_03677957 Bacteroides	TIGRTA	ATET-AGIK	ATET-AGIK	ATET-AGIK	ATET-AGIK	ATET-AGIK	ATET-AGIK	ATET-AGIK	ATET-AGIK	ATET-AGIK
YP_003584502 Zunongwangia	SVTH	NGEK--GGLF	NGEK--GGLF	NGEK--GGLF	NGEK--GGLF	NGEK--GGLF	NGEK--GGLF	NGEK--GGLF	NGEK--GGLF	NGEK--GGLF
ZP_01061975 Leeuwenhoekeiella	TVAQL	FNAN--SGLY	FNAN--SGLY	FNAN--SGLY	FNAN--SGLY	FNAN--SGLY	FNAN--SGLY	FNAN--SGLY	FNAN--SGLY	FNAN--SGLY
ZP_07083984 Sphingobacterium	AVDO	KGK--KGFS	KGK--KGFS	KGK--KGFS	KGK--KGFS	KGK--KGFS	KGK--KGFS	KGK--KGFS	KGK--KGFS	KGK--KGFS
YP_003120664 Chitinophaga	TVYK	EGK--KGF	EGK--KGF	EGK--KGF	EGK--KGF	EGK--KGF	EGK--KGF	EGK--KGF	EGK--KGF	EGK--KGF
ZP_01885202 Pedobacter	VVVK	AGK--KGF	AGK--KGF	AGK--KGF	AGK--KGF	AGK--KGF	AGK--KGF	AGK--KGF	AGK--KGF	AGK--KGF
ZP_02866543 Clostridium	TDDQ	EAGS-VN	EAGS-VN	EAGS-VN	EAGS-VN	EAGS-VN	EAGS-VN	EAGS-VN	EAGS-VN	EAGS-VN
XP_367221 Magnaporthe	AASRA	SGTPRN	SGTPRN	SGTPRN	SGTPRN	SGTPRN	SGTPRN	SGTPRN	SGTPRN	SGTPRN
ZP_07042437 Bacteroides	ALSH	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY
ZP_05759807 Bacteroides	ALSH	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY	EADKP-AY
ZP_05287524 Bacteroides	APSH	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY
ZP_06076108 Bacteroides	TVTQ	KSSE-SN	KSSE-SN	KSSE-SN	KSSE-SN	KSSE-SN	KSSE-SN	KSSE-SN	KSSE-SN	KSSE-SN
YP_001302992 Parabacteroides	APSH	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY	EKEGP-AY

Section 5

Section 7

	493	500	510	520	530	540	550	560	574
CcMan5 full	(493) AE	(500) ATGRVAID	(510) ASTATDPTG	(520) LQR	(530) FVH	(540) TSTS	(550) V	(560) AQSFT	(574) TDD
NP_630514 Streptomyces	(329) AD	(500) RT	(510) KASKKAD	(520) PDS	(530) TLT	(540) K	(550) KANGFT	(560) KANGFT	(574) TDD
ZP_05522540 Streptomyces	(332) AD	(500) RT	(510) KASKKAD	(520) PDS	(530) TLT	(540) K	(550) KANGFT	(560) KANGFT	(574) TDD
ZP_06527366 Streptomyces	(304) AD	(500) RT	(510) KASKKAD	(520) PDS	(530) TLT	(540) K	(550) KANGFT	(560) KANGFT	(574) TDD
YP_003013376 Paenibacillus	(448) AA	(500) SNV	(510) IDADE	(520) QNK	(530) R	(540) S	(550) RQASEL	(560) RQASEL	(574) TDD
ZP_812442 Bacteroides	(242) DQ	(500) KQ	(510) KGG	(520) KDRV	(530) S	(540) S	(550) ISEDFR	(560) ISEDFR	(574) TDD
ZP_04848482 Bacteroides	(260) DQ	(500) KQ	(510) KGG	(520) KDRV	(530) S	(540) S	(550) ISEDFR	(560) ISEDFR	(574) TDD
ZP_03677957 Bacteroides	(244) IA	(500) ENL	(510) QOETS	(520) DE	(530) LKTS	(540) R	(550) KSOQLP	(560) KSOQLP	(574) TDD
YP_003584502 Zunongwangia	(265) AA	(500) K	(510) R	(520) S	(530) E	(540) G	(550) OKTEGT	(560) OKTEGT	(574) TDD
ZP_01061975 Leeuwenhoeikiella	(252) AS	(500) Y	(510) T	(520) E	(530) S	(540) S	(550) ONEDYK	(560) ONEDYK	(574) TDD
ZP_07083984 Sphingobacterium	(273) QQ	(500) DIC	(510) H	(520) K	(530) S	(540) S	(550) SAKGK	(560) SAKGK	(574) TDD
YP_003120664 Chitinophaga	(250) KA	(500) DM	(510) H	(520) K	(530) S	(540) S	(550) HTNSK	(560) HTNSK	(574) TDD
ZP_01885202 Pedobacter	(264) MA	(500) DEL	(510) S	(520) D	(530) K	(540) S	(550) STOKSKEM	(560) STOKSKEM	(574) TDD
ZP_02866543 Clostridium	(324) KA	(500) DV	(510) S	(520) R	(530) E	(540) S	(550) BEANDYIH	(560) BEANDYIH	(574) TDD
XP_367221 Magnaporthe	(300) DV	(500) TD	(510) LSRVAS	(520) NDKN	(530) TTKLQHL	(540) S	(550) P	(560) P	(574) TDD
ZP_07042437 Bacteroides	(262) QI	(500) EA	(510) R	(520) I	(530) K	(540) V	(550) E	(560) E	(574) TDD
ZP_05759807 Bacteroides	(262) QI	(500) EA	(510) R	(520) I	(530) K	(540) V	(550) E	(560) E	(574) TDD
ZP_05287524 Bacteroides	(267) ND	(500) DA	(510) CKIO	(520) QGG	(530) S	(540) S	(550) EDGGRP	(560) EDGGRP	(574) TDD
ZP_06076108 Bacteroides	(254) EI	(500) NA	(510) K	(520) I	(530) E	(540) G	(550) EDTECHA	(560) EDTECHA	(574) TDD
YP_001302992 Parabacteroides	(267) ND	(500) DA	(510) CKIO	(520) QGG	(530) S	(540) S	(550) EDGGRS	(560) EDGGRS	(574) TDD

Section 8

	575	580	590	600	610	620	630	640	656
CcMan5 full	(575) KFS	Y	YALVRD	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
NP_630514 Streptomyces	(404) K	Y	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_05522540 Streptomyces	(407) K	Y	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_06527366 Streptomyces	(379) K	Y	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
YP_003013376 Paenibacillus	(524) K	Y	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
NP_812442 Bacteroides	(313) K	Y	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_04848482 Bacteroides	(331) K	Y	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_03677957 Bacteroides	(315) K	Y	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
YP_003584502 Zunongwangia	(334) Q	L	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_01061975 Leeuwenhoekella	(321) Q	L	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_07083984 Sphingobacterium	(342) Q	L	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
YP_003120664 Chitinophaga	(319) Q	L	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_01885202 Pedobacter	(333) Q	L	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_02866543 Clostridium	(399) K	L	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
XP_367221 Magnaporthe	(364) T	T	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_07042437 Bacteroides	(334) A	T	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_05759807 Bacteroides	(334) A	T	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_05287524 Bacteroides	(338) T	T	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
ZP_06076108 Bacteroides	(327) L	T	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV
YP_001302992 Parabacteroides	(338) T	T	YATYR	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV	YADAEATGTGGGLGGFVHSV

Region: Misc(404_408): VRxE motif

Section 10

	739	750	760	770	780	790	800	810	820
CcMan5 full	(739) WPIEK	TKPGAWTAADGTQV	GLL-TPRAADGSWQSA	DHAKFEAAG	DA	DA	DA	DA	DA
	(490) MATDN	VKPGAWTAADGTAV	GLL-TPRDGEGGWQGV	DYEKFEEAR	DA	DA	DA	DA	DA
NP_630514 Streptomyces	(533) MATDN	VKPGAWTAADGTAV	GLL-TPRDGEGGWQGV	DYEKFEEAR	DA	DA	DA	DA	DA
NP_05522540 Streptomyces	(536) MATDN	VKPGAWTAADGTAV	GLL-TPRDGEGGWQGV	DYEKFEEAR	DA	DA	DA	DA	DA
ZP_06527366 Streptomyces	(508) MATDN	VKPGAWTAADGTAV	GLL-TPRDGEGGWQGV	DYEKFEEAR	DA	DA	DA	DA	DA
YP_003013376 Paenibacillus	(644) S	KLNPTQVDEKGIPTG	---	FFTPNGTTVGAGDIQAVDRYA	QV	QV	QV	QV	QV
NP_812442 Bacteroides	(429) SL	EEIKK	FMNVTP	---	AFVEMKNNG	YIGSI	FMNVTP	---	AFVEMKNNG
ZP_04848482 Bacteroides	(447) SL	EEIKK	FMNVTP	---	AFVEMKNNG	YIGSI	FMNVTP	---	AFVEMKNNG
ZP_03677957 Bacteroides	(431) E	P	LO	FKD	I	G	P	---	TSDIMHGDC
YP_003584502 Zunongwangia	(449) D	E	Y	N	FKD	I	T	R	N
ZP_01061975 Leeuwenhoeikiella	(436) D	A	A	D	FKD	L	S	K	S
ZP_07083984 Sphingobacterium	(452) S	T	C	FKD	L	S	R	N	---
YP_003120664 Chitinophaga	(434) D	K	L	FQD	L	T	K	R	---
ZP_01885202 Pedobacter	(448) D	K	L	FKD	L	S	K	K	---
ZP_02866543 Clostridium	(526) N	N	T	R	AVD	S	P	F	S
XP_367221 Magnaporthe	(516) Q	S	H	N	K	A	L	G	F
ZP_07042437 Bacteroides	(494) N	N	N	N	N	N	N	N	N
ZP_05759807 Bacteroides	(494) N	N	N	N	N	N	N	N	N
ZP_05287524 Bacteroides	(499) N	N	N	N	N	N	N	N	N
ZP_06076108 Bacteroides	(483) N	N	N	N	N	N	N	N	N
YP_001302992 Parabacteroides	(499) N	N	N	N	N	N	N	N	N

Region: Misc(534_538): LYOGT motif

Section 11

	821	830	840	850	860	870	880	890	902
CcMan5 full	(821) FGEHAP	(830) -----	(840) -----	(850) DDGKAMLHSN	(860) -----	(870) -----	(880) IYTKETWNR	(890) IYTKETWNR	(902) IYTKETWNR
NP_630514 Streptomyces	(614) GEDSDA	-----	-----	DDGSTMHLHN	-----	-----	YTGETWNR	YTGETWNR	YTGETWNR
ZP_05522540 Streptomyces	(617) EDSDA	-----	-----	DDGSTMHLHN	-----	-----	YTGETWNR	YTGETWNR	YTGETWNR
ZP_06527366 Streptomyces	(589) EDSDA	-----	-----	DDGSTMHLHN	-----	-----	YTGETWNR	YTGETWNR	YTGETWNR
YP_003013376 Paenibacillus	(720) IDEY	-----	-----	MAI	-----	-----	EFTTEVVTQ	EFTTEVVTQ	EFTTEVVTQ
NP_812442 Bacteroides	(490) HHLY	-----	-----	NO	-----	-----	TEPMIHKY	TEPMIHKY	TEPMIHKY
ZP_04848482 Bacteroides	(508) HHLY	-----	-----	NO	-----	-----	TEPMIHKY	TEPMIHKY	TEPMIHKY
ZP_03677957 Bacteroides	(492) NNLFN	-----	-----	M	-----	-----	LEPTTMYG	LEPTTMYG	LEPTTMYG
YP_003584502 Zunongwangia	(511) NNHNY	-----	-----	NH	-----	-----	LDTMVQAV	LDTMVQAV	LDTMVQAV
ZP_01061975 Leeuwenhoekella	(498) EFN	-----	-----	NH	-----	-----	LDVTVQTY	LDVTVQTY	LDVTVQTY
ZP_07083984 Sphingobacterium	(514) EHYF	-----	-----	NR	-----	-----	AVDTVIQY	AVDTVIQY	AVDTVIQY
YP_003120664 Chitinophaga	(496) NDYY	-----	-----	CH	-----	-----	AVDTMVQH	AVDTMVQH	AVDTMVQH
ZP_01885202 Pedobacter	(510) NDYF	-----	-----	NK	-----	-----	ALDVTIQH	ALDVTIQH	ALDVTIQH
ZP_02866543 Clostridium	(606) MQDPDPKG	-----	-----	MQHNA	-----	-----	ETRETWNS	ETRETWNS	ETRETWNS
XP_367221 Magnaporthe	(597) PNVRPKG	-----	-----	HERFNR	-----	-----	YGKTYGVR	YGKTYGVR	YGKTYGVR
ZP_07042437 Bacteroides	(570) EPLGRSK	YEFYAQLPDHTGNVGF	-----	-----	-----	-----	KTWFRNDL	KTWFRNDL	KTWFRNDL
ZP_05759807 Bacteroides	(570) EPLGRSK	YEFYAQLPDHTGNVGF	-----	-----	-----	-----	KTWFRNDL	KTWFRNDL	KTWFRNDL
ZP_05287524 Bacteroides	(575) TPLGMSK	WQFYSTLPDHTGNVGF	-----	-----	-----	-----	NQWFRNDL	NQWFRNDL	NQWFRNDL
ZP_06076108 Bacteroides	(559) TNVPLYKKYDF	FLKQYPDMTGWIGMY	-----	-----	-----	-----	DLWYGDGP	DLWYGDGP	DLWYGDGP
YP_001302992 Parabacteroides	(575) TPLGMSK	WQFYSTLPDHTGNVGF	-----	-----	-----	-----	NQWFRNDL	NQWFRNDL	NQWFRNDL

Section 12

	903	910	920	930	940	950	960	970	984		
CcMan5 full	(903)	910	920	930	940	950	960	970	984		
	(640)	TPPLKTKVYRLDPRGMF	PIMD	DAGTMS	TMFA	AVIA	ST	TYDD	SA	TADGVSEDAFA	
NP_630514 Streptomyces	(684)	RPPVKTKAYELAPD	PIMD	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
ZP_05522540 Streptomyces	(687)	RPPVKTKAYELAPD	PIMD	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
ZP_06527366 Streptomyces	(659)	RPPVKTKAYELAPD	PIMD	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
YP_003013376 Paenibacillus	(778)	-----VYRDDPE	SPND	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
NP_812442 Bacteroides	(547)	-----KAFKNAPE	SPND	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
ZP_04848482 Bacteroides	(565)	-----KAFKNAPE	SPND	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
ZP_03677957 Bacteroides	(549)	-----KIPNTTPO	KEMD	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
YP_003584502 Zunongwangia	(569)	-----RIYQNKPK	RIND	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
ZP_01061975 Leeuwenhoekella	(556)	-----RIYQNKPK	RIND	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
ZP_07083984 Sphingobacterium	(572)	-----RIYKNESK	RIND	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
YP_003120664 Chitinophaga	(554)	-----PIYKNQ	RIND	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
ZP_01885202 Pedobacter	(568)	-----RIYKNERK	RIND	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
ZP_02866543 Clostridium	(669)	-----KMYKLSPO	ETMD	DAGTMS	TMFA	AVIA	SQFC	ST	TYDD	SA	EADGVS
XP_367221 Magnaporthe	(659)	-----	NSDAGANE	GI	WVM	GL	---	---	---	---	---
ZP_07042437 Bacteroides	(638)	-----	PGD	DC	VVF	GL	---	---	---	---	---
ZP_05759807 Bacteroides	(638)	-----	PGD	DC	VVF	GL	---	---	---	---	---
ZP_05287524 Bacteroides	(643)	-----	PGD	DC	VVF	GL	---	---	---	---	---
ZP_06076108 Bacteroides	(630)	-----	PGD	DC	VVF	GL	---	---	---	---	---
YP_001302992 Parabacteroides	(643)	-----	PGD	DC	VVF	GL	---	---	---	---	---

Section 13

	985	990	1000	1010	1020	1030	1040	1050	1066
CcMan5 full	TLDSATFGNT	DYATVVG	D	AFRGE	QPSD	TDTAPAFSMSTATDEPAEGPR			VSAEPTTVQTGDGG
NP_630514 Streptomyces	RFNNT	DAC	A	TKFD	S	EPSS	ARTEPAYSLNTDSGDGDD	EHAPGRGTTVV	SARPETVIRTAADG
ZP_05522540 Streptomyces	RFNNT	DAC	A	TKFD	S	EPSS	ARTEPAYSLNTDSGDGDD	EHAPGRGTTVV	SARPETVIRTAADG
ZP_06527366 Streptomyces	RFNNT	DAC	A	TKFD	S	EPSS	ARTEPAYSLNTDSGDGDD	EHAPGRGTTVV	SARPETVIRTAADG
YP_003013376 Paenibacillus	DFNQS	KID	A	TEFQ	S	ST	ENMS	GA	KASAAAPTVDYNADMNDNFNHEQLIPEKSTWKYDDKKGKEAG
NP_812442 Bacteroides	IHK	KADYR	HN	K	ELIYNYK				
ZP_04848482 Bacteroides	IHK	KADYR	HN	K	ELIYNYK				
ZP_03677957 Bacteroides	IHK	KADYR	HN	K	ELIYNYK				
YP_003584502 Zunongwangia	IT	GV	RS	DEI	Q		HNTDFIKSLFKK		
ZP_01061975 Leeuwenhoekella	IT	GV	RS	DEI	Q		VKEAWVSSIRQYL		
ZP_07083984 Sphingobacterium	VT	NR	NR	QSI	E		TKDKFISKVELN		
YP_003120664 Chitinophaga	AVR	QK	KL	REI	SR		NENIWITGKNIQP		
ZP_01885202 Pedobacter	YVS	QK	DR	NS	IAK		LGKWIADITRQ		
ZP_02866543 Clostridium	IT	GV	RS	DEI	Q		AGNQWVSDINAAE		
XP_367221 Magnaporthe	LV	GW	KT	WD	FA		TSPESSPPSPASEYKSGETYMGNLGGYTSNMNSGGHPSWLI		
ZP_07042437 Bacteroides	IT	GV	RS	DEI	Q		SGANAAPP	SADDK	
ZP_05759807 Bacteroides	IT	GV	RS	DEI	Q		SGANAAPP	SADDK	
ZP_05287524 Bacteroides	IT	GV	RS	DEI	Q		SGANAAPP	SADDK	
ZP_06076108 Bacteroides	IT	GV	RS	DEI	Q		SGANAAPP	SADDK	
YP_001302992 Parabacteroides	IT	GV	RS	DEI	Q		SGANAAPP	SADDK	

	1067	1080	1090	1100	1110	1120	1130	1148
CcMan5 full	(1067)	1080	1090	1100	1110	1120	1130	1148
	(795)	ALDATVTLTLDG-ARLAAAPAGTDLVTSGAASVVGLPDGVTAAVTVASPTALTVSLTGTASADARFFVHLLRDAALADGVAAAS						
NP_630514 Streptomyces	(846)	TVDASVELRLSGRASFAARKGTS	SLTRTGAASVTGLPDGV	TADLRLVTGKRTASLRLT	GTTRTDARFGITFRDR	AFPHGIPAST		
NP_05522540 Streptomyces	(849)	TVDASVELRLSGRASFAARKGTS	SLTRTGAASVTGLPDGV	TADLRLVTGKRTASLRLT	GTTRTDARFGITFRDR	AFPHGIPAST		
NP_06527366 Streptomyces	(821)	TVDASVELRLSGRASFAARKGTS	SLTRTGAASVTGLPDGV	TADLRLVTGKRTASLRLT	GTTRTDARFGITFRDR	AFPHGIPAST		
YP_003013376 Paenibacillus	(934)	EGWTQVDFDDSSWSSGKAMLGYSY	GKPAATTVSYGPNANNKYV	TYFRKTFDAKDL	DGILELDGSLIRDDGAI	VYLNHGEIF		
NP_812442 Bacteroides	(654)							
ZP_04848482 Bacteroides	(672)							
ZP_03677957 Bacteroides	(676)							
YP_003584502 Zunongwangia	(697)							
ZP_01061975 Leeuwenhoekella	(683)							
ZP_07083984 Sphingobacterium	(698)							
YP_003120664 Chitinophaga	(681)							
ZP_01885202 Pedobacter	(695)							
ZP_02866543 Clostridium	(826)	ITITLKTKEFAGEIGEDLVATGKINI	TNIPEGLEASAIKTEANKVEV	SLNGKAKNHTLNDSISNLT	IEITDGATNEPIKDSI			
XP_367221 Magnaporthe	(802)	PVAVTVP	PIVVVAFAICVTFFFIRRRRAAAQ	KALSSGSGTPESGIETL	TPTSQPVDTSKTNVQVEIVGAP	PLDSTQGANIA		
ZP_07042437 Bacteroides	(754)							
ZP_05759807 Bacteroides	(754)							
ZP_05287524 Bacteroides	(759)							
ZP_06076108 Bacteroides	(747)							
YP_001302992 Parabacteroides	(759)							

Section 14

Section 26

(2051)	2051	2060	2070	2080	2090	2100	2110	2120	2132
CcMan5 full	(1627)	VTVTATGADGTTQTVVEQVVTVPSCS							

NP_630514 Streptomyces (1676) WSARTQPAVTGDNRPQTPTVTGVTGGERVRVNWRPAGDGGFPVVGYYTVALDDGTTAHPGTTSTAVLTAAGGAKAHTATVT
 ZP_05522540 Streptomyces (1679) WSARTQPAVTGDNRPQAPTPTGVTGGERVRVNWRPAGDGGFPVVGYYTVALDDGTTAHPGTTSTAVLTAAGGAKAHTATVT
 ZP_06527366 Streptomyces (1651) WSARTQPAVTGDNRPQAPTPTGVTGGERVRVNWRPAGDGGFPVVGYYTVALDDGTTAHPGTTSTAVLTAAGGAKAHTATVT
 YP_003013376 Paenibacillus (1844) NTFAVEYAGNTTNCFTNSAVEIRLGSPTGTLVGKISTPPKAGNWTTYDVTVSGTLTQKLTGIQDVYLVLTGSAGNGETGKKYI
 NP_812442 Bacteroides (654) -----
 ZP_04848482 Bacteroides (672) -----
 ZP_03677957 Bacteroides (676) -----
 YP_003584502 Zunongwangia (697) -----
 ZP_01061975 Leeuwenhoeikiella (683) -----
 ZP_07083984 Sphingobacterium (698) -----
 YP_003120664 Chitinophaga (681) -----
 ZP_01885202 Pedobacter (695) -----
 ZP_02866543 Clostridium (1810) FIDKVTGLDSSKYTQTTWTFAPDKELTEAIAVYNDENAMQEEVNTAYSELVTAFLNLRILIPDKSLLLEDLINQANGLNGANYTK
 XP_367221 Magnaporthe (893) -----
 ZP_07042437 Bacteroides (754) -----
 ZP_05759807 Bacteroides (754) -----
 ZP_05287524 Bacteroides (759) -----
 ZP_06076108 Bacteroides (747) -----
 YP_001302992 Parabacteroides (759) -----

	2133	2140	2150	2160	2170	2180	2190	2200	2214
CcMan5 full (1651)	(2133)	2140	2150	2160	2170	2180	2190	2200	2214
NP_630514 Streptomyces (1758)									
ZP_05522540 Streptomyces (1761)									
ZP_06527366 Streptomyces (1733)									
YP_003013376 Paenibacillus (1926)									
NP_812442 Bacteroides (654)									
ZP_04848482 Bacteroides (672)									
ZP_03677957 Bacteroides (676)									
YP_003584502 Zunongwangia (697)									
ZP_01061975 Leeuwenhoekella (683)									
ZP_07083984 Sphingobacterium (698)									
YP_003120664 Chitinophaga (681)									
ZP_01885202 Pedobacter (695)									
ZP_02866543 Clostridium (1892)									
XP_367221 Magnaporthe (893)									
ZP_07042437 Bacteroides (754)									
ZP_05759807 Bacteroides (754)									
ZP_05287524 Bacteroides (759)									
ZP_06076108 Bacteroides (747)									
YP_001302992 Parabacteroides (759)									

Section 27

Section 28

(2215)	2215	2220	2230	2240	2250	2260	2270	2280	2296
CcMan5 full (1651)									

NP_630514 Streptomyces (1840) GANSEVPAGTPLGAENDRI TVSVNNAATQQQVDRAEVDASNSATVTMADGLGSRRLGPLYGEALKEGRLPKTSALFSRVNENL
 ZP_055222540 Streptomyces (1840) GANSEVPAGTPLGAENDRI TVRVNNAATQQQVDRAEVDASNSATVTMADGLGSRRLGPLYGEALKEGRLPKTSALFSRVNENL
 ZP_06527366 Streptomyces (1812) GANSEVPAGTPLGAENDRI TVRVNNAATQQQVDRAEVDASNSATVTMADGLGSRRLGPLYGEALKEGRLPKTSALFSRVNENL
 YP_003013376 Paenibacillus (1938) -----
 NP_812442 Bacteroides (654) -----
 ZP_04848482 Bacteroides (672) -----
 ZP_03677957 Bacteroides (676) -----
 YP_003584502 Zunongwangia (697) -----
 ZP_01061975 Leeuwenhoekella (683) -----
 ZP_07083984 Sphingobacterium (698) -----
 YP_003120664 Chitinophaga (681) -----
 ZP_01885202 Pedobacter (695) -----
 ZP_02866543 Clostridium (1974) AGYTIILKRKEN -----
 XP_367221 Magnaporthe (893) -----
 ZP_07042437 Bacteroides (754) -----
 ZP_05759807 Bacteroides (754) -----
 ZP_05287524 Bacteroides (759) -----
 ZP_06076108 Bacteroides (747) -----
 YP_001302992 Parabacteroides (759) -----

	2379	2400	2410	2420	2430	2440	2450	2460
CcMan5 full (1651)	(2379)	2400	2410	2420	2430	2440	2450	2460
NP_630514 Streptomyces (2004)								
ZP_05522540 Streptomyces (2004)								
ZP_06527366 Streptomyces (1976)								
YP_003013376 Paenibacillus (1938)								
NP_812442 Bacteroides (654)								
ZP_04848482 Bacteroides (672)								
ZP_03677957 Bacteroides (676)								
YP_003584502 Zunongwangia (697)								
ZP_01061975 Leeuwenhoeikiella (683)								
ZP_07083984 Sphingobacterium (698)								
YP_003120664 Chitinophaga (681)								
ZP_01885202 Pedobacter (695)								
ZP_02866543 Clostridium (1985)								
XP_367221 Magnaporthe (893)								
ZP_07042437 Bacteroides (754)								
ZP_05759807 Bacteroides (754)								
ZP_05287524 Bacteroides (759)								
ZP_06076108 Bacteroides (747)								
YP_001302992 Parabacteroides (759)								

Section 30

Section 32

2624

2610

2600

2590

2580

2570

2560

2550

2543

(2543) 2543

(1651)

CcMan5 full (1651)

NP_630514 Streptomyces (2168) VTVTTFPPDTAASAAEAVAITVGGVALDGFDPDVSTYVVDWPRNGGRIPAVGAVTAASGARVKVTSQSSSTVSSSQRGFSTRIT
 ZP_05522540 Streptomyces (2168) VTVTTFPDATAASAAEAVAITVGGVALDGFDPDVSTYVVDWPRNGGRIPAVGAVTAASGARVKVTSQSSSTVSSSQRGFSTRIT
 ZP_06527366 Streptomyces (2140) VTVTTFPDATAASAAEAVAITVGGVALDGFDPDVSTYVVDWPRNGGRIPAVGAVTAASGARVKVTSQSSSTVSSSQRGFSTRIT
 YP_003013376 Paenibacillus (1938) -----
 NP_812442 Bacteroides (654) -----
 ZP_04848482 Bacteroides (672) -----
 ZP_03677957 Bacteroides (676) -----
 YP_003584502 Zunongwangia (697) -----
 ZP_01061975 Leeuwenhoekella (683) -----
 ZP_07083984 Sphingobacterium (698) -----
 YP_003120664 Chitinophaga (681) -----
 ZP_01885202 Pedobacter (695) -----
 ZP_02866543 Clostridium (1985) -----
 XP_367221 Magnaporthe (893) -----
 ZP_07042437 Bacteroides (754) -----
 ZP_05759807 Bacteroides (754) -----
 ZP_05287524 Bacteroides (759) -----
 ZP_06076108 Bacteroides (747) -----
 YP_001302992 Parabacteroides (759) -----

FIG. 33

ATOM	5859	N	GLY	B	21	8.798	41.888	151.532	1.00	6.14	B	N
ATOM	5860	CA	GLY	B	21	7.349	41.901	151.577	1.00	6.87	B	C
ATOM	5861	C	GLY	B	21	6.718	40.662	150.976	1.00	7.36	B	C
ATOM	5862	O	GLY	B	21	7.111	39.526	151.272	1.00	7.83	B	O
ATOM	5863	N	ASP	B	22	5.749	40.896	150.115	1.00	6.97	B	N
ATOM	5864	CA	ASP	B	22	5.019	39.846	149.439	1.00	6.61	B	C
ATOM	5865	CB	ASP	B	22	3.516	40.156	149.545	1.00	6.75	B	C
ATOM	5866	CG	ASP	B	22	2.645	38.964	149.165	1.00	8.00	B	C
ATOM	5867	OD1	ASP	B	22	3.121	37.817	149.252	1.00	8.66	B	O
ATOM	5868	OD2	ASP	B	22	1.479	39.168	148.781	1.00	10.69	B	O
ATOM	5869	C	ASP	B	22	5.471	39.814	147.976	1.00	6.58	B	C
ATOM	5870	O	ASP	B	22	4.647	39.889	147.070	1.00	6.88	B	O
ATOM	5871	N	PHE	B	23	6.785	39.719	147.753	1.00	5.72	B	N
ATOM	5872	CA	PHE	B	23	7.350	39.730	146.400	1.00	5.51	B	C
ATOM	5873	CB	PHE	B	23	8.139	41.020	146.136	1.00	5.56	B	C
ATOM	5874	CG	PHE	B	23	7.315	42.266	146.256	1.00	6.28	B	C
ATOM	5875	CD1	PHE	B	23	7.127	42.875	147.497	1.00	7.33	B	C
ATOM	5876	CE1	PHE	B	23	6.344	44.032	147.621	1.00	6.32	B	C
ATOM	5877	CZ	PHE	B	23	5.755	44.584	146.490	1.00	7.38	B	C
ATOM	5878	CE2	PHE	B	23	5.941	43.979	145.235	1.00	7.39	B	C
ATOM	5879	CD2	PHE	B	23	6.713	42.824	145.132	1.00	7.45	B	C
ATOM	5880	C	PHE	B	23	8.243	38.517	146.175	1.00	5.47	B	C
ATOM	5881	O	PHE	B	23	9.345	38.640	145.612	1.00	5.14	B	O
ATOM	5882	N	GLY	B	24	7.750	37.349	146.600	1.00	4.56	B	N
ATOM	5883	CA	GLY	B	24	8.458	36.090	146.362	1.00	4.07	B	C
ATOM	5884	C	GLY	B	24	8.397	35.090	147.499	1.00	3.76	B	C
ATOM	5885	O	GLY	B	24	8.310	33.871	147.263	1.00	4.04	B	O
ATOM	5886	N	ASN	B	25	8.467	35.594	148.734	1.00	2.92	B	N
ATOM	5887	CA	ASN	B	25	8.571	34.746	149.915	1.00	2.81	B	C
ATOM	5888	CB	ASN	B	25	7.253	34.007	150.179	1.00	3.39	B	C
ATOM	5889	CG	ASN	B	25	6.227	34.911	150.842	1.00	3.67	B	C
ATOM	5890	OD1	ASN	B	25	6.478	35.461	151.917	1.00	6.16	B	O
ATOM	5891	ND2	ASN	B	25	5.090	35.098	150.199	1.00	5.91	B	N
ATOM	5892	C	ASN	B	25	9.804	33.822	149.943	1.00	2.73	B	C
ATOM	5893	O	ASN	B	25	9.747	32.691	150.423	1.00	2.49	B	O
ATOM	5894	N	ASP	B	26	10.924	34.343	149.450	1.00	3.00	B	N
ATOM	5895	CA	ASP	B	26	12.206	33.663	149.535	1.00	3.13	B	C
ATOM	5896	CB	ASP	B	26	12.975	33.829	148.235	1.00	3.67	B	C
ATOM	5897	CG	ASP	B	26	12.481	32.904	147.140	1.00	4.56	B	C
ATOM	5898	OD1	ASP	B	26	12.308	31.693	147.408	1.00	6.75	B	O
ATOM	5899	OD2	ASP	B	26	12.281	33.385	146.007	1.00	5.43	B	O
ATOM	5900	C	ASP	B	26	13.041	34.166	150.702	1.00	3.37	B	C
ATOM	5901	O	ASP	B	26	12.747	35.212	151.278	1.00	3.44	B	O
ATOM	6036	N	GLY	B	46	7.869	42.799	161.557	1.00	5.19	B	N
ATOM	6037	CA	GLY	B	46	7.346	42.212	160.323	1.00	4.86	B	C
ATOM	6038	C	GLY	B	46	8.311	41.184	159.770	1.00	5.18	B	C
ATOM	6039	O	GLY	B	46	9.481	41.143	160.175	1.00	5.13	B	O
ATOM	6040	N	ARG	B	47	7.818	40.352	158.848	1.00	5.25	B	N
ATOM	6041	CA	ARG	B	47	8.581	39.247	158.248	1.00	4.58	B	C
ATOM	6042	CB	ARG	B	47	8.650	38.046	159.210	1.00	4.70	B	C
ATOM	6043	CG	ARG	B	47	7.282	37.403	159.599	1.00	5.20	B	C
ATOM	6044	CD	ARG	B	47	7.495	36.267	160.596	1.00	6.03	B	C
ATOM	6045	NE	ARG	B	47	6.339	35.388	160.817	1.00	5.27	B	N
ATOM	6046	CZ	ARG	B	47	5.957	34.374	160.030	1.00	7.87	B	C
ATOM	6047	NH1	ARG	B	47	6.586	34.104	158.889	1.00	6.65	B	N
ATOM	6048	NH2	ARG	B	47	4.910	33.627	160.378	1.00	6.18	B	N
ATOM	6049	C	ARG	B	47	7.861	38.854	156.975	1.00	4.46	B	C
ATOM	6050	O	ARG	B	47	6.656	39.124	156.837	1.00	3.68	B	O
ATOM	6051	N	ASN	B	48	8.584	38.257	156.029	1.00	4.25	B	N
ATOM	6052	CA	ASN	B	48	7.898	37.485	154.983	1.00	4.09	B	C
ATOM	6053	CB	ASN	B	48	8.627	37.525	153.605	1.00	3.81	B	C
ATOM	6054	CG	ASN	B	48	9.824	36.572	153.502	1.00	3.63	B	C
ATOM	6055	OD1	ASN	B	48	10.191	35.864	154.455	1.00	4.51	B	O
ATOM	6056	ND2	ASN	B	48	10.441	36.556	152.325	1.00	2.00	B	N
ATOM	6057	C	ASN	B	48	7.626	36.082	155.535	1.00	4.15	B	C
ATOM	6058	O	ASN	B	48	7.928	35.813	156.710	1.00	4.08	B	O

ATOM	6059	N	ASN	B	49	7.049	35.200	154.730	1.00	3.96	B	N
ATOM	6060	CA	ASN	B	49	6.685	33.858	155.217	1.00	4.78	B	C
ATOM	6061	CB	ASN	B	49	5.892	33.072	154.155	1.00	4.58	B	C
ATOM	6062	CG	ASN	B	49	4.528	33.683	153.864	1.00	6.22	B	C
ATOM	6063	OD1	ASN	B	49	3.827	33.270	152.922	1.00	8.25	B	O
ATOM	6064	ND2	ASN	B	49	4.151	34.685	154.655	1.00	4.90	B	N
ATOM	6065	C	ASN	B	49	7.840	33.013	155.763	1.00	4.50	B	C
ATOM	6066	O	ASN	B	49	7.632	32.168	156.622	1.00	5.46	B	O
ATOM	6067	N	THR	B	50	9.054	33.243	155.273	1.00	5.02	B	N
ATOM	6068	CA	THR	B	50	10.246	32.518	155.744	1.00	4.39	B	C
ATOM	6069	CB	THR	B	50	11.413	32.651	154.748	1.00	4.34	B	C
ATOM	6070	OG1	THR	B	50	11.882	34.011	154.742	1.00	3.44	B	O
ATOM	6071	CG2	THR	B	50	10.950	32.254	153.325	1.00	4.57	B	C
ATOM	6072	C	THR	B	50	10.745	33.031	157.096	1.00	4.84	B	C
ATOM	6073	O	THR	B	50	11.450	32.306	157.804	1.00	4.54	B	O
ATOM	6074	N	GLY	B	51	10.432	34.293	157.416	1.00	4.48	B	N
ATOM	6075	CA	GLY	B	51	10.802	34.910	158.707	1.00	4.48	B	C
ATOM	6076	C	GLY	B	51	11.560	36.232	158.553	1.00	5.18	B	C
ATOM	6077	O	GLY	B	51	11.862	36.918	159.542	1.00	5.14	B	O
ATOM	6200	N	LEU	B	67	10.876	26.584	154.563	1.00	4.27	B	N
ATOM	6201	CA	LEU	B	67	10.335	26.228	153.272	1.00	4.24	B	C
ATOM	6202	CB	LEU	B	67	8.858	25.822	153.413	1.00	3.74	B	C
ATOM	6203	CG	LEU	B	67	8.558	24.462	154.063	1.00	2.90	B	C
ATOM	6204	CD1	LEU	B	67	7.076	24.167	154.019	1.00	2.92	B	C
ATOM	6205	CD2	LEU	B	67	9.361	23.328	153.400	1.00	2.00	B	C
ATOM	6206	C	LEU	B	67	10.436	27.457	152.371	1.00	4.38	B	C
ATOM	6207	O	LEU	B	67	9.926	28.517	152.720	1.00	4.30	B	O
ATOM	6208	N	ASP	B	68	11.070	27.282	151.216	1.00	4.40	B	N
ATOM	6209	CA	ASP	B	68	11.312	28.351	150.252	1.00	4.81	B	C
ATOM	6210	CB	ASP	B	68	12.398	27.937	149.254	1.00	4.70	B	C
ATOM	6211	CG	ASP	B	68	13.677	27.502	149.932	1.00	5.37	B	C
ATOM	6212	OD1	ASP	B	68	13.702	26.364	150.444	1.00	4.55	B	O
ATOM	6213	OD2	ASP	B	68	14.636	28.300	149.964	1.00	5.37	B	O
ATOM	6214	C	ASP	B	68	10.079	28.727	149.459	1.00	4.93	B	C
ATOM	6215	O	ASP	B	68	9.469	27.878	148.818	1.00	5.94	B	O
ATOM	6216	N	GLY	B	69	9.723	30.005	149.507	1.00	5.17	B	N
ATOM	6217	CA	GLY	B	69	8.769	30.598	148.558	1.00	4.56	B	C
ATOM	6218	C	GLY	B	69	7.327	30.161	148.689	1.00	4.60	B	C
ATOM	6219	O	GLY	B	69	6.573	30.236	147.713	1.00	4.30	B	O
ATOM	6220	N	VAL	B	70	6.932	29.725	149.887	1.00	3.48	B	N
ATOM	6221	CA	VAL	B	70	5.589	29.149	150.102	1.00	3.51	B	C
ATOM	6222	CB	VAL	B	70	5.627	27.927	151.063	1.00	3.54	B	C
ATOM	6223	CG1	VAL	B	70	6.419	26.772	150.442	1.00	3.34	B	C
ATOM	6224	CG2	VAL	B	70	6.209	28.314	152.422	1.00	2.91	B	C
ATOM	6225	C	VAL	B	70	4.609	30.183	150.644	1.00	3.44	B	C
ATOM	6226	O	VAL	B	70	5.033	31.257	151.119	1.00	3.41	B	O
ATOM	6227	N	GLY	B	71	3.317	29.856	150.590	1.00	3.21	B	N
ATOM	6228	CA	GLY	B	71	2.243	30.766	151.033	1.00	4.39	B	C
ATOM	6229	C	GLY	B	71	1.896	30.610	152.517	1.00	5.23	B	C
ATOM	6230	O	GLY	B	71	2.732	30.168	153.313	1.00	5.01	B	O
ATOM	6231	N	GLY	B	72	0.653	30.950	152.869	1.00	5.67	B	N
ATOM	6232	CA	GLY	B	72	0.165	30.889	154.256	1.00	5.93	B	C
ATOM	6233	C	GLY	B	72	0.949	31.862	155.107	1.00	6.07	B	C
ATOM	6234	O	GLY	B	72	1.165	33.016	154.692	1.00	6.64	B	O
ATOM	6235	N	SER	B	73	1.390	31.404	156.282	1.00	5.79	B	N
ATOM	6236	CA	SER	B	73	2.360	32.159	157.086	1.00	5.71	B	C
ATOM	6237	CB	SER	B	73	1.884	32.320	158.534	1.00	5.72	B	C
ATOM	6238	OG	SER	B	73	1.572	31.062	159.111	1.00	5.04	B	O
ATOM	6239	C	SER	B	73	3.751	31.512	157.055	1.00	5.79	B	C
ATOM	6240	O	SER	B	73	4.621	31.864	157.855	1.00	5.53	B	O
ATOM	6241	N	GLY	B	74	3.943	30.560	156.136	1.00	5.50	B	N
ATOM	6242	CA	GLY	B	74	5.232	29.886	155.962	1.00	4.98	B	C
ATOM	6243	C	GLY	B	74	5.389	28.660	156.834	1.00	4.36	B	C
ATOM	6244	O	GLY	B	74	4.702	28.512	157.834	1.00	4.36	B	O
ATOM	6245	N	GLY	B	75	6.284	27.765	156.443	1.00	4.59	B	N
ATOM	6246	CA	GLY	B	75	6.527	26.565	157.216	1.00	4.40	B	C

ATOM	6247	C	GLY	B	75	7.889	26.594	157.880	1.00	4.18	B	C
ATOM	6248	O	GLY	B	75	8.717	27.435	157.575	1.00	4.41	B	O
ATOM	6249	N	GLY	B	76	8.116	25.653	158.788	1.00	4.82	B	N
ATOM	6250	CA	GLY	B	76	9.371	25.580	159.508	1.00	4.57	B	C
ATOM	6251	C	GLY	B	76	9.475	26.759	160.434	1.00	4.29	B	C
ATOM	6252	O	GLY	B	76	8.504	27.108	161.114	1.00	4.51	B	O
ATOM	6253	N	GLY	B	77	10.647	27.390	160.433	1.00	3.80	B	N
ATOM	6254	CA	GLY	B	77	10.953	28.443	161.386	1.00	3.39	B	C
ATOM	6255	C	GLY	B	77	11.241	27.958	162.792	1.00	3.21	B	C
ATOM	6256	O	GLY	B	77	11.099	28.729	163.743	1.00	3.66	B	O
ATOM	7082	N	TYR	B	194	3.643	22.233	162.238	1.00	3.12	B	N
ATOM	7083	CA	TYR	B	194	2.613	21.951	161.250	1.00	3.30	B	C
ATOM	7084	CB	TYR	B	194	1.229	21.842	161.911	1.00	3.34	B	C
ATOM	7085	CG	TYR	B	194	0.110	21.512	160.940	1.00	3.71	B	C
ATOM	7086	CD1	TYR	B	194	-0.461	22.504	160.135	1.00	4.84	B	C
ATOM	7087	CE1	TYR	B	194	-1.490	22.204	159.240	1.00	6.19	B	C
ATOM	7088	CZ	TYR	B	194	-1.954	20.908	159.146	1.00	4.77	B	C
ATOM	7089	OH	TYR	B	194	-2.964	20.602	158.255	1.00	6.66	B	O
ATOM	7090	CE2	TYR	B	194	-1.400	19.908	159.936	1.00	5.56	B	C
ATOM	7091	CD2	TYR	B	194	-0.370	20.213	160.817	1.00	2.00	B	C
ATOM	7092	C	TYR	B	194	2.618	23.039	160.183	1.00	3.39	B	C
ATOM	7093	O	TYR	B	194	2.833	24.217	160.484	1.00	2.89	B	O
ATOM	7094	N	PHE	B	195	2.402	22.634	158.934	1.00	3.57	B	N
ATOM	7095	CA	PHE	B	195	2.176	23.583	157.852	1.00	4.28	B	C
ATOM	7096	CB	PHE	B	195	3.495	24.128	157.273	1.00	3.41	B	C
ATOM	7097	CG	PHE	B	195	3.307	25.061	156.101	1.00	2.84	B	C
ATOM	7098	CD1	PHE	B	195	2.857	26.368	156.296	1.00	2.00	B	C
ATOM	7099	CE1	PHE	B	195	2.688	27.240	155.214	1.00	2.78	B	C
ATOM	7100	CZ	PHE	B	195	2.970	26.810	153.920	1.00	2.01	B	C
ATOM	7101	CE2	PHE	B	195	3.431	25.487	153.713	1.00	3.28	B	C
ATOM	7102	CD2	PHE	B	195	3.588	24.636	154.801	1.00	2.14	B	C
ATOM	7103	C	PHE	B	195	1.340	22.977	156.739	1.00	4.75	B	C
ATOM	7104	O	PHE	B	195	1.700	21.949	156.158	1.00	4.94	B	O
ATOM	7105	N	TYR	B	196	0.234	23.655	156.460	1.00	6.22	B	N
ATOM	7106	CA	TYR	B	196	-0.598	23.421	155.296	1.00	7.19	B	C
ATOM	7107	CB	TYR	B	196	0.228	23.437	153.995	1.00	7.83	B	C
ATOM	7108	CG	TYR	B	196	-0.616	23.795	152.800	1.00	10.54	B	C
ATOM	7109	CD1	TYR	B	196	-1.022	25.116	152.596	1.00	12.24	B	C
ATOM	7110	CE1	TYR	B	196	-1.824	25.468	151.513	1.00	15.08	B	C
ATOM	7111	CZ	TYR	B	196	-2.227	24.487	150.619	1.00	16.10	B	C
ATOM	7112	OH	TYR	B	196	-3.015	24.861	149.550	1.00	19.44	B	O
ATOM	7113	CE2	TYR	B	196	-1.840	23.147	150.800	1.00	15.05	B	C
ATOM	7114	CD2	TYR	B	196	-1.038	22.812	151.890	1.00	11.35	B	C
ATOM	7115	C	TYR	B	196	-1.432	22.161	155.434	1.00	7.32	B	C
ATOM	7116	O	TYR	B	196	-2.592	22.244	155.848	1.00	8.08	B	O
ATOM	7117	N	ASN	B	197	-0.848	21.010	155.111	1.00	6.91	B	N
ATOM	7118	CA	ASN	B	197	-1.557	19.737	155.149	1.00	7.00	B	C
ATOM	7119	CB	ASN	B	197	-1.448	19.010	153.804	1.00	6.93	B	C
ATOM	7120	CG	ASN	B	197	-2.381	19.573	152.756	1.00	8.25	B	C
ATOM	7121	OD1	ASN	B	197	-3.492	19.991	153.063	1.00	6.81	B	O
ATOM	7122	ND2	ASN	B	197	-1.932	19.577	151.499	1.00	9.68	B	N
ATOM	7123	C	ASN	B	197	-1.097	18.779	156.242	1.00	6.55	B	C
ATOM	7124	O	ASN	B	197	-1.841	17.875	156.589	1.00	6.69	B	O
ATOM	7125	N	ALA	B	198	0.104	18.977	156.793	1.00	5.68	B	N
ATOM	7126	CA	ALA	B	198	0.738	17.907	157.606	1.00	5.17	B	C
ATOM	7127	CB	ALA	B	198	1.519	16.952	156.715	1.00	3.99	B	C
ATOM	7128	C	ALA	B	198	1.644	18.391	158.727	1.00	4.90	B	C
ATOM	7129	O	ALA	B	198	2.316	19.416	158.594	1.00	5.24	B	O
ATOM	8311	N	ALA	B	352	5.634	20.319	147.023	1.00	5.44	B	N
ATOM	8312	CA	ALA	B	352	4.594	21.317	146.769	1.00	6.09	B	C
ATOM	8313	CB	ALA	B	352	5.135	22.730	146.962	1.00	5.99	B	C
ATOM	8314	C	ALA	B	352	4.223	21.089	145.329	1.00	6.35	B	C
ATOM	8315	O	ALA	B	352	4.288	21.990	144.503	1.00	6.13	B	O
ATOM	8316	N	THR	B	353	3.821	19.855	145.044	1.00	7.15	B	N
ATOM	8317	CA	THR	B	353	3.818	19.352	143.690	1.00	7.57	B	C
ATOM	8318	CB	THR	B	353	3.887	17.837	143.696	1.00	7.65	B	C

ATOM	8319	OG1	THR	B	353	4.943	17.447	144.580	1.00	8.17	B	O
ATOM	8320	CG2	THR	B	353	4.184	17.295	142.296	1.00	6.93	B	C
ATOM	8321	C	THR	B	353	2.656	19.914	142.859	1.00	8.28	B	C
ATOM	8322	O	THR	B	353	2.750	19.992	141.631	1.00	8.21	B	O
ATOM	8323	N	TRP	B	354	1.581	20.325	143.531	1.00	8.91	B	N
ATOM	8324	CA	TRP	B	354	0.511	21.060	142.861	1.00	10.28	B	C
ATOM	8325	CB	TRP	B	354	-0.559	21.535	143.859	1.00	10.88	B	C
ATOM	8326	CG	TRP	B	354	-1.795	22.053	143.189	1.00	13.77	B	C
ATOM	8327	CD1	TRP	B	354	-2.904	21.331	142.861	1.00	15.75	B	C
ATOM	8328	NE1	TRP	B	354	-3.829	22.131	142.242	1.00	18.84	B	N
ATOM	8329	CE2	TRP	B	354	-3.338	23.407	142.172	1.00	19.33	B	C
ATOM	8330	CD2	TRP	B	354	-2.050	23.396	142.759	1.00	17.76	B	C
ATOM	8331	CE3	TRP	B	354	-1.317	24.588	142.806	1.00	19.35	B	C
ATOM	8332	CZ3	TRP	B	354	-1.887	25.751	142.273	1.00	22.00	B	C
ATOM	8333	CH2	TRP	B	354	-3.175	25.730	141.696	1.00	22.73	B	C
ATOM	8334	CZ2	TRP	B	354	-3.914	24.569	141.634	1.00	22.09	B	C
ATOM	8335	C	TRP	B	354	1.082	22.248	142.090	1.00	9.94	B	C
ATOM	8336	O	TRP	B	354	0.679	22.498	140.957	1.00	10.17	B	O
ATOM	8337	N	ASP	B	355	2.027	22.961	142.708	1.00	9.68	B	N
ATOM	8338	CA	ASP	B	355	2.607	24.171	142.126	1.00	9.25	B	C
ATOM	8339	CB	ASP	B	355	3.020	25.161	143.224	1.00	9.39	B	C
ATOM	8340	CG	ASP	B	355	1.920	25.410	144.250	1.00	11.18	B	C
ATOM	8341	OD1	ASP	B	355	2.026	24.884	145.380	1.00	12.17	B	O
ATOM	8342	OD2	ASP	B	355	0.957	26.138	143.927	1.00	14.96	B	O
ATOM	8343	C	ASP	B	355	3.832	23.883	141.261	1.00	8.63	B	C
ATOM	8344	O	ASP	B	355	4.010	24.505	140.198	1.00	8.49	B	O
ATOM	8345	N	ASP	B	356	4.679	22.961	141.721	1.00	7.73	B	N
ATOM	8346	CA	ASP	B	356	6.027	22.821	141.162	1.00	7.97	B	C
ATOM	8347	CB	ASP	B	356	7.084	22.835	142.282	1.00	8.30	B	C
ATOM	8348	CG	ASP	B	356	7.193	21.517	143.028	1.00	10.67	B	C
ATOM	8349	OD1	ASP	B	356	6.378	20.590	142.800	1.00	12.81	B	O
ATOM	8350	OD2	ASP	B	356	8.130	21.405	143.856	1.00	13.11	B	O
ATOM	8351	C	ASP	B	356	6.290	21.648	140.225	1.00	6.85	B	C
ATOM	8352	O	ASP	B	356	7.432	21.434	139.843	1.00	6.23	B	O
ATOM	8702	N	THR	B	403	6.656	20.211	154.931	1.00	2.24	B	N
ATOM	8703	CA	THR	B	403	6.223	19.841	153.605	1.00	2.00	B	C
ATOM	8704	CB	THR	B	403	6.095	18.282	153.520	1.00	2.05	B	C
ATOM	8705	OG1	THR	B	403	5.760	17.892	152.198	1.00	3.15	B	O
ATOM	8706	CG2	THR	B	403	5.030	17.770	154.453	1.00	2.00	B	C
ATOM	8707	C	THR	B	403	4.913	20.578	153.289	1.00	2.00	B	C
ATOM	8708	O	THR	B	403	4.316	21.193	154.178	1.00	2.00	B	O
ATOM	8709	N	VAL	B	404	4.491	20.536	152.026	1.00	2.00	B	N
ATOM	8710	CA	VAL	B	404	3.295	21.247	151.582	1.00	2.02	B	C
ATOM	8711	CB	VAL	B	404	3.633	22.358	150.571	1.00	2.00	B	C
ATOM	8712	CG1	VAL	B	404	2.406	23.239	150.320	1.00	2.00	B	C
ATOM	8713	CG2	VAL	B	404	4.800	23.187	151.064	1.00	2.00	B	C
ATOM	8714	C	VAL	B	404	2.175	20.345	151.023	1.00	2.65	B	C
ATOM	8715	O	VAL	B	404	1.183	20.118	151.702	1.00	2.92	B	O
ATOM	8716	N	ARG	B	405	2.332	19.834	149.803	1.00	3.29	B	N
ATOM	8717	CA	ARG	B	405	1.236	19.105	149.134	1.00	3.66	B	C
ATOM	8718	CB	ARG	B	405	0.170	20.101	148.633	1.00	3.90	B	C
ATOM	8719	CG	ARG	B	405	0.641	20.982	147.480	1.00	3.00	B	C
ATOM	8720	CD	ARG	B	405	-0.271	22.201	147.301	1.00	2.02	B	C
ATOM	8721	NE	ARG	B	405	-1.643	21.828	146.974	1.00	3.88	B	N
ATOM	8722	CZ	ARG	B	405	-2.594	22.700	146.644	1.00	5.53	B	C
ATOM	8723	NH1	ARG	B	405	-2.324	24.001	146.580	1.00	5.29	B	N
ATOM	8724	NH2	ARG	B	405	-3.812	22.271	146.379	1.00	6.56	B	N
ATOM	8725	C	ARG	B	405	1.717	18.221	147.981	1.00	3.69	B	C
ATOM	8726	O	ARG	B	405	2.837	18.353	147.494	1.00	3.54	B	O
ATOM	8727	N	TRP	B	406	0.861	17.315	147.540	1.00	4.26	B	N
ATOM	8728	CA	TRP	B	406	1.343	16.193	146.726	1.00	4.40	B	C
ATOM	8729	CB	TRP	B	406	1.380	14.920	147.575	1.00	4.23	B	C
ATOM	8730	CG	TRP	B	406	2.088	15.094	148.872	1.00	4.31	B	C
ATOM	8731	CD1	TRP	B	406	3.417	14.922	149.110	1.00	4.94	B	C
ATOM	8732	NE1	TRP	B	406	3.696	15.167	150.429	1.00	5.82	B	N
ATOM	8733	CE2	TRP	B	406	2.537	15.516	151.067	1.00	4.54	B	C

ATOM	8734	CD2	TRP	B	406	1.500	15.472	150.112	1.00	4.03	B	C
ATOM	8735	CE3	TRP	B	406	0.192	15.805	150.507	1.00	2.57	B	C
ATOM	8736	CZ3	TRP	B	406	-0.024	16.148	151.840	1.00	4.33	B	C
ATOM	8737	CH2	TRP	B	406	1.032	16.167	152.764	1.00	4.13	B	C
ATOM	8738	CZ2	TRP	B	406	2.316	15.859	152.397	1.00	3.80	B	C
ATOM	8739	C	TRP	B	406	0.565	15.933	145.433	1.00	4.65	B	C
ATOM	9696	N	GLY	B	533	-15.847	27.725	140.342	1.00	9.53	B	N
ATOM	9697	CA	GLY	B	533	-15.211	26.855	141.335	1.00	9.25	B	C
ATOM	9698	C	GLY	B	533	-13.705	26.691	141.187	1.00	8.71	B	C
ATOM	9699	O	GLY	B	533	-13.098	25.875	141.883	1.00	8.16	B	O
ATOM	9700	N	LEU	B	534	-13.099	27.477	140.293	1.00	8.58	B	N
ATOM	9701	CA	LEU	B	534	-11.653	27.388	140.006	1.00	8.07	B	C
ATOM	9702	CB	LEU	B	534	-11.286	28.145	138.734	1.00	7.63	B	C
ATOM	9703	CG	LEU	B	534	-11.866	27.619	137.420	1.00	6.93	B	C
ATOM	9704	CD1	LEU	B	534	-11.597	28.602	136.307	1.00	3.89	B	C
ATOM	9705	CD2	LEU	B	534	-11.303	26.245	137.079	1.00	7.25	B	C
ATOM	9706	C	LEU	B	534	-10.747	27.869	141.134	1.00	8.88	B	C
ATOM	9707	O	LEU	B	534	-11.033	28.870	141.822	1.00	7.62	B	O
ATOM	9708	N	TYR	B	535	-9.637	27.152	141.288	1.00	9.49	B	N
ATOM	9709	CA	TYR	B	535	-8.638	27.486	142.280	1.00	10.81	B	C
ATOM	9710	CB	TYR	B	535	-8.010	26.197	142.861	1.00	11.89	B	C
ATOM	9711	CG	TYR	B	535	-8.753	25.749	144.106	1.00	16.97	B	C
ATOM	9712	CD1	TYR	B	535	-8.130	25.772	145.361	1.00	20.37	B	C
ATOM	9713	CE1	TYR	B	535	-8.832	25.389	146.520	1.00	23.75	B	C
ATOM	9714	CZ	TYR	B	535	-10.184	25.009	146.424	1.00	23.62	B	C
ATOM	9715	OH	TYR	B	535	-10.865	24.649	147.562	1.00	24.62	B	O
ATOM	9716	CE2	TYR	B	535	-10.827	24.987	145.193	1.00	22.59	B	C
ATOM	9717	CD2	TYR	B	535	-10.111	25.365	144.038	1.00	20.68	B	C
ATOM	9718	C	TYR	B	535	-7.604	28.447	141.703	1.00	10.16	B	C
ATOM	9719	O	TYR	B	535	-6.865	28.094	140.801	1.00	10.24	B	O
ATOM	9720	N	GLN	B	536	-7.600	29.677	142.211	1.00	9.98	B	N
ATOM	9721	CA	GLN	B	536	-6.568	30.681	141.905	1.00	10.33	B	C
ATOM	9722	CB	GLN	B	536	-5.234	30.334	142.598	1.00	10.30	B	C
ATOM	9723	CG	GLN	B	536	-5.313	30.141	144.128	1.00	13.24	B	C
ATOM	9724	CD	GLN	B	536	-3.939	29.900	144.772	1.00	16.30	B	C
ATOM	9725	OE1	GLN	B	536	-2.909	29.886	144.095	1.00	17.97	B	O
ATOM	9726	NE2	GLN	B	536	-3.929	29.725	146.091	1.00	18.13	B	N
ATOM	9727	C	GLN	B	536	-6.337	30.936	140.402	1.00	10.03	B	C
ATOM	9728	O	GLN	B	536	-5.217	31.217	139.967	1.00	10.11	B	O
ATOM	9729	N	GLY	B	537	-7.397	30.850	139.608	1.00	10.06	B	N
ATOM	9730	CA	GLY	B	537	-7.264	31.159	138.187	1.00	8.79	B	C
ATOM	9731	C	GLY	B	537	-8.584	31.268	137.454	1.00	8.30	B	C
ATOM	9732	O	GLY	B	537	-9.659	31.075	138.037	1.00	7.71	B	O
ATOM	10119	N	ASN	B	586	1.158	35.774	134.842	1.00	4.61	B	N
ATOM	10120	CA	ASN	B	586	2.561	35.487	135.130	1.00	4.87	B	C
ATOM	10121	CB	ASN	B	586	3.446	36.671	134.695	1.00	4.94	B	C
ATOM	10122	CG	ASN	B	586	3.354	36.964	133.194	1.00	4.99	B	C
ATOM	10123	OD1	ASN	B	586	2.930	38.041	132.796	1.00	8.76	B	O
ATOM	10124	ND2	ASN	B	586	3.754	36.017	132.371	1.00	4.46	B	N
ATOM	10125	C	ASN	B	586	2.791	35.182	136.618	1.00	5.09	B	C
ATOM	10126	O	ASN	B	586	3.852	34.691	136.997	1.00	3.71	B	O
ATOM	10127	N	ALA	B	587	1.777	35.465	137.446	1.00	5.34	B	N
ATOM	10128	CA	ALA	B	587	1.917	35.442	138.923	1.00	6.16	B	C
ATOM	10129	CB	ALA	B	587	1.006	36.491	139.546	1.00	5.10	B	C
ATOM	10130	C	ALA	B	587	1.688	34.108	139.635	1.00	6.83	B	C
ATOM	10131	O	ALA	B	587	2.084	33.957	140.784	1.00	6.45	B	O
ATOM	10132	N	ASN	B	588	0.997	33.179	138.977	1.00	8.41	B	N
ATOM	10133	CA	ASN	B	588	0.684	31.857	139.540	1.00	9.70	B	C
ATOM	10134	CB	ASN	B	588	-0.790	31.765	139.954	1.00	10.13	B	C
ATOM	10135	CG	ASN	B	588	-1.026	30.752	141.077	1.00	12.26	B	C
ATOM	10136	OD1	ASN	B	588	-0.090	30.198	141.622	1.00	15.20	B	O
ATOM	10137	ND2	ASN	B	588	-2.286	30.513	141.418	1.00	15.01	B	N
ATOM	10138	C	ASN	B	588	1.027	30.786	138.509	1.00	10.25	B	C
ATOM	10139	O	ASN	B	588	1.296	31.109	137.356	1.00	10.70	B	O
ATOM	10140	N	GLU	B	589	1.014	29.523	138.913	1.00	10.86	B	N
ATOM	10141	CA	GLU	B	589	1.574	28.445	138.079	1.00	11.52	B	C

ATOM	10142	CB	GLU	B	589	2.198	27.370	138.976	1.00	11.45	B	C
ATOM	10143	CG	GLU	B	589	3.501	27.805	139.630	1.00	9.96	B	C
ATOM	10144	CD	GLU	B	589	3.311	28.386	141.028	1.00	10.68	B	C
ATOM	10145	OE1	GLU	B	589	2.146	28.629	141.434	1.00	10.43	B	O
ATOM	10146	OE2	GLU	B	589	4.335	28.606	141.722	1.00	8.00	B	O
ATOM	10147	C	GLU	B	589	0.608	27.804	137.071	1.00	11.76	B	C
ATOM	10148	O	GLU	B	589	1.027	27.008	136.207	1.00	11.79	B	O
ATOM	10149	N	ILE	B	590	-0.670	28.170	137.171	1.00	11.86	B	N
ATOM	10150	CA	ILE	B	590	-1.724	27.523	136.394	1.00	11.76	B	C
ATOM	10151	CB	ILE	B	590	-3.097	28.106	136.760	1.00	11.88	B	C
ATOM	10152	CG1	ILE	B	590	-3.531	27.471	138.085	1.00	12.53	B	C
ATOM	10153	CD1	ILE	B	590	-4.228	28.396	138.985	1.00	13.66	B	C
ATOM	10154	CG2	ILE	B	590	-4.143	27.847	135.665	1.00	12.30	B	C
ATOM	10155	C	ILE	B	590	-1.461	27.444	134.877	1.00	11.77	B	C
ATOM	10156	O	ILE	B	590	-1.563	26.362	134.286	1.00	11.66	B	O
ATOM	10444	CA	ILE	B	624	-2.166	38.601	140.777	1.00	6.95	B	C
ATOM	10445	CB	ILE	B	624	-2.394	39.529	142.009	1.00	7.43	B	C
ATOM	10446	CG1	ILE	B	624	-3.881	39.873	142.189	1.00	6.21	B	C
ATOM	10447	CD1	ILE	B	624	-4.256	40.384	143.612	1.00	6.93	B	C
ATOM	10448	CG2	ILE	B	624	-1.589	40.820	141.848	1.00	5.38	B	C
ATOM	10449	C	ILE	B	624	-2.529	37.158	141.097	1.00	8.00	B	C
ATOM	10450	O	ILE	B	624	-3.395	36.597	140.423	1.00	7.71	B	O
ATOM	10451	N	ALA	B	625	-1.876	36.566	142.107	1.00	8.28	B	N
ATOM	10452	CA	ALA	B	625	-2.036	35.146	142.429	1.00	9.17	B	C
ATOM	10453	CB	ALA	B	625	-0.672	34.532	142.890	1.00	8.73	B	C
ATOM	10454	C	ALA	B	625	-3.121	34.848	143.476	1.00	9.67	B	C
ATOM	10455	O	ALA	B	625	-3.305	33.688	143.871	1.00	9.55	B	O
ATOM	10456	N	THR	B	626	-3.813	35.889	143.941	1.00	10.19	B	N
ATOM	10457	CA	THR	B	626	-4.883	35.743	144.940	1.00	9.61	B	C
ATOM	10458	CB	THR	B	626	-4.311	35.718	146.381	1.00	10.06	B	C
ATOM	10459	OG1	THR	B	626	-5.328	35.322	147.300	1.00	9.01	B	O
ATOM	10460	CG2	THR	B	626	-3.722	37.088	146.791	1.00	10.40	B	C
ATOM	10461	C	THR	B	626	-5.890	36.882	144.760	1.00	9.89	B	C
ATOM	10462	O	THR	B	626	-5.682	37.753	143.914	1.00	9.41	B	O
ATOM	10463	N	GLY	B	627	-6.954	36.880	145.563	1.00	10.02	B	N
ATOM	10464	CA	GLY	B	627	-8.057	37.828	145.428	1.00	11.00	B	C
ATOM	10465	C	GLY	B	627	-7.684	39.294	145.557	1.00	12.06	B	C
ATOM	10466	O	GLY	B	627	-8.185	40.135	144.815	1.00	12.64	B	O
ATOM	10467	N	SER	B	628	-6.793	39.604	146.489	1.00	13.03	B	N
ATOM	10468	CA	SER	B	628	-6.410	40.988	146.753	1.00	14.00	B	C
ATOM	10469	CB	SER	B	628	-7.482	41.650	147.614	1.00	14.26	B	C
ATOM	10470	OG	SER	B	628	-7.266	43.039	147.649	1.00	15.26	B	O
ATOM	10471	C	SER	B	628	-5.047	41.111	147.452	1.00	14.28	B	C
ATOM	10472	O	SER	B	628	-4.640	40.217	148.191	1.00	14.61	B	O
ATOM	10473	N	SER	B	629	-4.362	42.229	147.219	1.00	14.43	B	N
ATOM	10474	CA	SER	B	629	-3.055	42.511	147.827	1.00	14.96	B	C
ATOM	10475	CB	SER	B	629	-1.950	42.366	146.779	1.00	14.81	B	C
ATOM	10476	OG	SER	B	629	-0.708	42.826	147.280	1.00	15.07	B	O
ATOM	10477	C	SER	B	629	-2.997	43.923	148.390	1.00	15.25	B	C
ATOM	10478	O	SER	B	629	-3.433	44.866	147.747	1.00	15.02	B	O
ATOM	10680	N	PRO	B	657	4.050	41.388	143.073	1.00	6.53	B	N
ATOM	10681	CA	PRO	B	657	3.402	41.236	144.385	1.00	6.71	B	C
ATOM	10682	CB	PRO	B	657	2.633	42.551	144.560	1.00	6.75	B	C
ATOM	10683	CG	PRO	B	657	2.497	43.125	143.179	1.00	6.81	B	C
ATOM	10684	CD	PRO	B	657	3.695	42.650	142.401	1.00	6.65	B	C
ATOM	10685	C	PRO	B	657	2.446	40.037	144.343	1.00	7.09	B	C
ATOM	10686	O	PRO	B	657	1.785	39.823	143.317	1.00	7.31	B	O
ATOM	10687	N	THR	B	658	2.382	39.270	145.438	1.00	6.82	B	N
ATOM	10688	CA	THR	B	658	1.659	37.972	145.528	1.00	7.41	B	C
ATOM	10689	CB	THR	B	658	0.160	38.025	145.092	1.00	7.22	B	C
ATOM	10690	OG1	THR	B	658	0.091	38.108	143.663	1.00	8.46	B	O
ATOM	10691	CG2	THR	B	658	-0.562	39.183	145.720	1.00	7.56	B	C
ATOM	10692	C	THR	B	658	2.281	36.761	144.802	1.00	6.95	B	C
ATOM	10693	O	THR	B	658	1.850	35.614	145.012	1.00	7.00	B	O
ATOM	10694	N	MET	B	659	3.266	37.002	143.944	1.00	6.43	B	N
ATOM	10695	CA	MET	B	659	3.892	35.925	143.184	1.00	5.95	B	C

FIG. 34

CRYST1	82.202	91.158	224.523	90.00	90.00	90.00	P 2 2 21					
SCALE1	0.012165	0.000000	0.000000			0.000000						
SCALE2	-0.000000	0.010970	0.000000			0.000000						
SCALE3	0.000000	-0.000000	0.004454			0.000000						
ATOM	2	CA	ALA	A	7	74.405	12.350	86.541	1.00	19.71	A	C
ATOM	7	CA	ASP	A	8	72.571	11.287	83.330	1.00	16.72	A	C
ATOM	15	CA	TYR	A	9	68.912	11.738	84.261	1.00	12.13	A	C
ATOM	27	CA	ALA	A	10	68.003	12.081	80.575	1.00	8.97	A	C
ATOM	32	CA	SER	A	11	68.548	8.297	80.172	1.00	8.34	A	C
ATOM	38	CA	LEU	A	12	65.749	7.661	82.703	1.00	6.43	A	C
ATOM	46	CA	VAL	A	13	63.113	9.628	80.736	1.00	5.75	A	C
ATOM	53	CA	ASP	A	14	60.886	7.538	78.450	1.00	6.27	A	C
ATOM	61	CA	VAL	A	15	59.534	10.004	75.843	1.00	4.66	A	C
ATOM	68	CA	PHE	A	16	57.177	7.311	74.498	1.00	3.87	A	C
ATOM	79	CA	VAL	A	17	55.121	7.160	77.667	1.00	3.47	A	C
ATOM	86	CA	GLY	A	18	51.656	8.454	76.810	1.00	3.67	A	C
ATOM	90	CA	THR	A	19	52.191	8.387	72.991	1.00	3.13	A	C
ATOM	97	CA	GLU	A	20	49.720	5.556	72.200	1.00	4.27	A	C
ATOM	106	CA	GLY	A	21	46.188	6.265	70.980	1.00	5.40	A	C
ATOM	110	CA	ASP	A	22	43.990	8.104	73.483	1.00	5.27	A	C
ATOM	118	CA	PHE	A	23	46.225	7.338	76.520	1.00	4.39	A	C
ATOM	129	CA	GLY	A	24	47.673	10.774	77.312	1.00	3.08	A	C
ATOM	133	CA	ASN	A	25	48.095	12.630	73.991	1.00	2.47	A	C
ATOM	141	CA	ASP	A	26	51.797	13.270	74.588	1.00	3.20	A	C
ATOM	149	CA	MET	A	27	54.547	13.334	71.981	1.00	3.36	A	C
ATOM	157	CA	PRO	A	28	57.941	11.662	71.502	1.00	3.20	A	C
ATOM	164	CA	ALA	A	29	59.066	14.935	69.784	1.00	3.06	A	C
ATOM	169	CA	ALA	A	30	62.625	16.198	70.105	1.00	3.69	A	C
ATOM	174	CA	GLN	A	31	62.423	19.283	72.321	1.00	5.14	A	C
ATOM	183	CA	ALA	A	32	64.432	21.430	74.735	1.00	4.55	A	C
ATOM	188	CA	PRO	A	33	63.094	22.317	78.204	1.00	4.64	A	C
ATOM	195	CA	ASN	A	34	60.021	24.568	77.625	1.00	4.19	A	C
ATOM	203	CA	GLY	A	35	60.994	24.689	73.959	1.00	3.44	A	C
ATOM	207	CA	LEU	A	36	59.041	26.309	71.169	1.00	3.34	A	C
ATOM	215	CA	ALA	A	37	60.477	23.938	68.541	1.00	3.52	A	C
ATOM	220	CA	LYS	A	38	58.932	20.498	68.910	1.00	3.01	A	C
ATOM	229	CA	VAL	A	39	60.209	18.174	66.213	1.00	2.00	A	C
ATOM	236	CA	ASN	A	40	57.543	15.463	66.213	1.00	2.24	A	C
ATOM	244	CA	PRO	A	41	57.420	12.468	63.910	1.00	3.24	A	C
ATOM	251	CA	ARG	A	42	53.989	12.144	62.244	1.00	4.43	A	C
ATOM	262	CA	THR	A	43	52.366	8.720	62.032	1.00	5.82	A	C
ATOM	269	CA	THR	A	44	49.941	7.692	59.271	1.00	7.54	A	C
ATOM	276	CA	PRO	A	45	47.070	6.985	58.732	1.00	7.64	A	C
ATOM	283	CA	GLY	A	46	46.363	7.283	62.482	1.00	5.22	A	C
ATOM	287	CA	ARG	A	47	47.876	9.677	65.005	1.00	4.33	A	C
ATOM	298	CA	ASN	A	48	47.297	10.971	68.487	1.00	2.87	A	C
ATOM	306	CA	ASN	A	49	46.469	14.686	68.971	1.00	3.59	A	C
ATOM	314	CA	THR	A	50	50.174	15.553	68.695	1.00	2.96	A	C
ATOM	321	CA	GLY	A	51	50.582	13.739	65.384	1.00	2.16	A	C
ATOM	325	CA	TYR	A	52	52.147	10.463	66.622	1.00	2.75	A	C
ATOM	337	CA	ASP	A	53	50.289	7.308	67.663	1.00	3.03	A	C
ATOM	345	CA	TYR	A	54	52.612	4.563	69.034	1.00	3.42	A	C
ATOM	357	CA	ALA	A	55	50.295	1.835	67.698	1.00	3.80	A	C
ATOM	362	CA	GLN	A	56	50.932	2.905	64.064	1.00	4.22	A	C
ATOM	371	CA	SER	A	57	53.533	1.527	61.658	1.00	6.71	A	C
ATOM	377	CA	LYS	A	58	54.051	4.328	59.145	1.00	6.64	A	C
ATOM	386	CA	ILE	A	59	55.576	7.777	59.431	1.00	5.57	A	C
ATOM	394	CA	SER	A	60	55.344	10.584	56.904	1.00	4.00	A	C
ATOM	400	CA	GLY	A	61	57.749	13.214	58.346	1.00	3.12	A	C
ATOM	404	CA	PHE	A	62	58.483	15.778	61.088	1.00	4.01	A	C
ATOM	415	CA	THR	A	63	56.355	18.741	62.206	1.00	3.06	A	C
ATOM	422	CA	HIS	A	64	57.952	21.581	64.185	1.00	2.46	A	C
ATOM	432	CA	THR	A	65	55.187	23.059	66.366	1.00	2.07	A	C
ATOM	439	CA	ASN	A	66	52.932	21.418	68.894	1.00	2.86	A	C
ATOM	447	CA	LEU	A	67	51.019	21.341	72.178	1.00	3.42	A	C
ATOM	455	CA	ASP	A	68	51.501	18.692	74.851	1.00	2.88	A	C

ATOM	463	CA	GLY	A	69	48.598	16.584	76.017	1.00	3.16	A	C
ATOM	467	CA	VAL	A	70	45.743	18.719	74.687	1.00	2.95	A	C
ATOM	474	CA	GLY	A	71	42.319	17.506	73.447	1.00	4.24	A	C
ATOM	478	CA	GLY	A	72	40.310	18.076	70.238	1.00	5.07	A	C
ATOM	482	CA	SER	A	73	42.497	16.981	67.319	1.00	5.30	A	C
ATOM	488	CA	GLY	A	74	45.570	18.647	68.860	1.00	4.44	A	C
ATOM	492	CA	GLY	A	75	47.211	22.012	68.196	1.00	4.03	A	C
ATOM	496	CA	GLY	A	76	50.230	23.149	66.192	1.00	3.47	A	C
ATOM	500	CA	GLY	A	77	51.645	20.417	63.910	1.00	2.77	A	C
ATOM	504	CA	ASP	A	78	53.003	23.083	61.502	1.00	3.19	A	C
ATOM	512	CA	LEU	A	79	55.816	22.729	58.975	1.00	3.46	A	C
ATOM	520	CA	LEU	A	80	55.960	19.099	57.867	1.00	3.19	A	C
ATOM	528	CA	VAL	A	81	59.330	17.935	56.541	1.00	2.38	A	C
ATOM	535	CA	VAL	A	82	59.151	14.633	54.633	1.00	2.66	A	C
ATOM	542	CA	PRO	A	83	62.241	12.791	53.307	1.00	3.10	A	C
ATOM	549	CA	THR	A	84	61.709	10.950	50.007	1.00	3.64	A	C
ATOM	556	CA	SER	A	85	63.546	9.484	47.006	1.00	3.93	A	C
ATOM	562	CA	GLY	A	86	60.474	10.171	44.894	1.00	4.58	A	C
ATOM	566	CA	SER	A	87	59.775	13.000	42.432	1.00	6.80	A	C
ATOM	572	CA	TYR	A	88	56.783	15.353	42.023	1.00	6.34	A	C
ATOM	584	CA	THR	A	89	55.158	17.278	39.133	1.00	6.35	A	C
ATOM	591	CA	ALA	A	90	52.313	18.731	41.257	1.00	5.37	A	C
ATOM	596	CA	ARG	A	91	51.505	19.377	44.932	1.00	5.13	A	C
ATOM	607	CA	PRO	A	92	52.147	16.058	46.755	1.00	5.88	A	C
ATOM	614	CA	GLY	A	93	49.327	13.739	47.757	1.00	6.88	A	C
ATOM	618	CA	THR	A	94	49.802	13.201	51.509	1.00	7.19	A	C
ATOM	625	CA	GLY	A	95	49.477	9.410	51.104	1.00	6.49	A	C
ATOM	629	CA	THR	A	96	52.770	9.459	49.173	1.00	4.50	A	C
ATOM	636	CA	TYR	A	97	54.650	10.619	52.300	1.00	3.46	A	C
ATOM	648	CA	ALA	A	98	54.035	7.311	54.100	1.00	3.39	A	C
ATOM	653	CA	HIS	A	99	57.136	5.240	55.089	1.00	4.47	A	C
ATOM	663	CA	PRO	A	100	57.203	1.916	56.975	1.00	5.44	A	C
ATOM	670	CA	PHE	A	101	58.803	2.128	60.401	1.00	5.59	A	C
ATOM	681	CA	SER	A	102	59.416	-0.191	63.342	1.00	7.19	A	C
ATOM	687	CA	HIS	A	103	59.895	0.563	67.048	1.00	6.56	A	C
ATOM	697	CA	ASP	A	104	62.859	-1.852	66.789	1.00	10.20	A	C
ATOM	705	CA	ASP	A	105	64.517	0.789	64.587	1.00	10.01	A	C
ATOM	713	CA	GLU	A	106	63.589	3.886	66.611	1.00	9.26	A	C
ATOM	722	CA	ASP	A	107	65.354	5.697	69.473	1.00	9.20	A	C
ATOM	730	CA	ALA	A	108	64.518	8.899	71.371	1.00	5.45	A	C
ATOM	735	CA	GLY	A	109	65.127	10.798	74.597	1.00	4.60	A	C
ATOM	739	CA	PRO	A	110	65.502	14.365	75.895	1.00	3.95	A	C
ATOM	746	CA	GLY	A	111	66.404	16.609	72.933	1.00	4.26	A	C
ATOM	750	CA	PHE	A	112	66.407	13.997	70.163	1.00	4.28	A	C
ATOM	761	CA	TYR	A	113	64.446	11.529	68.079	1.00	5.36	A	C
ATOM	773	CA	SER	A	114	65.729	8.957	65.540	1.00	5.75	A	C
ATOM	779	CA	VAL	A	115	64.016	6.387	63.260	1.00	4.53	A	C
ATOM	786	CA	GLY	A	116	64.781	4.199	60.259	1.00	5.51	A	C
ATOM	790	CA	LEU	A	117	62.194	4.893	57.576	1.00	5.77	A	C
ATOM	798	CA	GLY	A	118	61.496	2.539	54.687	1.00	6.00	A	C
ATOM	802	CA	ASN	A	119	62.614	4.169	51.445	1.00	5.77	A	C
ATOM	810	CA	VAL	A	120	59.965	5.458	49.014	1.00	5.26	A	C
ATOM	817	CA	ALA	A	121	60.384	6.350	45.306	1.00	6.56	A	C
ATOM	822	CA	GLY	A	122	58.455	6.925	42.071	1.00	6.12	A	C
ATOM	826	CA	THR	A	123	56.572	9.913	40.635	1.00	7.50	A	C
ATOM	833	CA	ASP	A	124	53.544	11.573	42.245	1.00	7.71	A	C
ATOM	841	CA	GLY	A	125	50.654	9.176	43.087	1.00	7.97	A	C
ATOM	845	CA	ALA	A	126	52.803	6.284	41.903	1.00	8.07	A	C
ATOM	850	CA	ILE	A	127	55.283	6.769	44.795	1.00	8.60	A	C
ATOM	858	CA	THR	A	128	55.591	3.528	46.766	1.00	9.40	A	C
ATOM	865	CA	GLY	A	129	58.010	1.444	48.876	1.00	9.09	A	C
ATOM	869	CA	ALA	A	130	61.528	1.221	47.442	1.00	8.74	A	C
ATOM	874	CA	PRO	A	131	64.650	-0.773	48.532	1.00	9.45	A	C
ATOM	881	CA	GLY	A	132	66.719	0.579	51.405	1.00	9.13	A	C
ATOM	885	CA	THR	A	133	66.380	2.820	54.435	1.00	8.85	A	C
ATOM	892	CA	ILE	A	134	66.150	6.548	55.020	1.00	7.14	A	C

ATOM	900	CA	GLU	A	135	67.868	6.967	58.400	1.00	7.03	A	C
ATOM	909	CA	ALA	A	136	66.326	10.008	60.046	1.00	5.25	A	C
ATOM	914	CA	GLU	A	137	67.736	11.833	63.043	1.00	3.46	A	C
ATOM	923	CA	VAL	A	138	66.270	15.029	64.552	1.00	2.00	A	C
ATOM	930	CA	ALA	A	139	67.283	17.116	67.586	1.00	2.00	A	C
ATOM	935	CA	ALA	A	140	66.021	20.401	69.089	1.00	3.29	A	C
ATOM	940	CA	ALA	A	141	67.192	23.587	70.725	1.00	5.41	A	C
ATOM	945	CA	THR	A	142	64.774	26.112	72.324	1.00	3.98	A	C
ATOM	952	CA	ARG	A	143	63.747	27.685	69.007	1.00	3.05	A	C
ATOM	963	CA	SER	A	144	65.230	25.253	66.475	1.00	3.15	A	C
ATOM	969	CA	GLY	A	145	64.814	21.822	64.969	1.00	3.61	A	C
ATOM	973	CA	VAL	A	146	67.745	20.138	63.259	1.00	3.66	A	C
ATOM	980	CA	HIS	A	147	67.601	17.212	60.859	1.00	4.27	A	C
ATOM	990	CA	ARG	A	148	70.193	14.697	59.628	1.00	5.28	A	C
ATOM	1001	CA	TYR	A	149	69.133	12.182	56.930	1.00	5.88	A	C
ATOM	1013	CA	ALA	A	150	70.978	9.286	55.295	1.00	6.54	A	C
ATOM	1018	CA	PHE	A	151	69.444	8.138	51.980	1.00	6.34	A	C
ATOM	1029	CA	PRO	A	152	70.329	4.881	50.196	1.00	8.11	A	C
ATOM	1036	CA	ALA	A	153	73.284	5.138	47.799	1.00	9.27	A	C
ATOM	1041	CA	GLY	A	154	72.114	6.052	44.300	1.00	9.43	A	C
ATOM	1045	CA	SER	A	155	69.016	7.860	45.553	1.00	8.94	A	C
ATOM	1051	CA	THR	A	156	68.111	11.379	44.480	1.00	5.81	A	C
ATOM	1058	CA	PRO	A	157	67.363	12.771	47.997	1.00	5.54	A	C
ATOM	1065	CA	SER	A	158	64.501	15.181	48.463	1.00	5.58	A	C
ATOM	1071	CA	LEU	A	159	62.757	16.902	51.348	1.00	4.75	A	C
ATOM	1079	CA	VAL	A	160	59.189	18.092	50.985	1.00	3.15	A	C
ATOM	1086	CA	VAL	A	161	58.408	21.099	53.170	1.00	2.91	A	C
ATOM	1093	CA	ASP	A	162	54.597	21.001	53.529	1.00	2.92	A	C
ATOM	1101	CA	LEU	A	163	53.165	24.242	54.913	1.00	4.33	A	C
ATOM	1109	CA	GLU	A	164	49.518	23.198	54.922	1.00	4.22	A	C
ATOM	1118	CA	THR	A	165	49.840	20.473	57.636	1.00	3.08	A	C
ATOM	1125	CA	ASN	A	166	48.201	21.399	60.948	1.00	3.13	A	C
ATOM	1133	CA	ASN	A	167	46.701	19.356	63.804	1.00	3.44	A	C
ATOM	1141	CA	THR	A	168	43.479	21.366	63.648	1.00	4.11	A	C
ATOM	1148	CA	SER	A	169	43.334	24.170	61.080	1.00	2.77	A	C
ATOM	1154	CA	ARG	A	170	45.644	25.892	58.648	1.00	3.20	A	C
ATOM	1165	CA	ARG	A	171	44.592	29.537	58.189	1.00	2.85	A	C
ATOM	1176	CA	SER	A	172	47.387	30.959	56.027	1.00	2.36	A	C
ATOM	1182	CA	SER	A	173	50.992	30.215	55.065	1.00	3.42	A	C
ATOM	1188	CA	SER	A	174	53.845	31.585	52.935	1.00	4.21	A	C
ATOM	1194	CA	VAL	A	175	57.213	30.528	51.618	1.00	4.87	A	C
ATOM	1201	CA	GLN	A	176	60.036	32.533	50.050	1.00	7.10	A	C
ATOM	1210	CA	VAL	A	177	63.000	30.700	48.499	1.00	8.23	A	C
ATOM	1217	CA	GLU	A	178	66.645	31.960	48.440	1.00	10.90	A	C
ATOM	1226	CA	THR	A	179	69.583	30.187	46.804	1.00	9.85	A	C
ATOM	1233	CA	ARG	A	180	72.927	31.368	48.203	1.00	10.39	A	C
ATOM	1244	CA	ALA	A	181	76.419	31.485	46.632	1.00	11.40	A	C
ATOM	1249	CA	ASP	A	182	77.367	28.014	47.948	1.00	11.35	A	C
ATOM	1257	CA	GLY	A	183	74.201	26.531	46.431	1.00	9.02	A	C
ATOM	1261	CA	THR	A	184	72.333	25.942	49.723	1.00	6.71	A	C
ATOM	1268	CA	VAL	A	185	68.670	26.993	50.031	1.00	5.35	A	C
ATOM	1275	CA	GLU	A	186	66.959	29.077	52.696	1.00	5.35	A	C
ATOM	1284	CA	LEU	A	187	63.146	29.005	52.987	1.00	3.46	A	C
ATOM	1292	CA	SER	A	188	61.169	31.481	55.086	1.00	2.87	A	C
ATOM	1298	CA	GLY	A	189	57.690	32.795	55.787	1.00	2.92	A	C
ATOM	1302	CA	GLN	A	190	54.805	32.417	58.222	1.00	2.71	A	C
ATOM	1311	CA	VAL	A	191	52.276	29.832	59.268	1.00	2.84	A	C
ATOM	1318	CA	THR	A	192	48.954	30.792	60.850	1.00	3.69	A	C
ATOM	1325	CA	GLY	A	193	46.998	28.051	62.583	1.00	3.21	A	C
ATOM	1329	CA	TYR	A	194	44.116	27.757	64.995	1.00	3.16	A	C
ATOM	1341	CA	PHE	A	195	43.413	25.739	68.120	1.00	3.26	A	C
ATOM	1352	CA	TYR	A	196	40.408	25.930	70.460	1.00	3.84	A	C
ATOM	1364	CA	ASN	A	197	39.874	29.693	71.045	1.00	4.26	A	C
ATOM	1372	CA	ALA	A	198	42.510	31.491	69.033	1.00	3.45	A	C
ATOM	1377	CA	ALA	A	199	44.567	31.753	65.914	1.00	3.47	A	C
ATOM	1382	CA	TYR	A	200	48.369	32.096	66.243	1.00	2.36	A	C

ATOM	1394	CA	THR	A	201	51.191	33.045	63.858	1.00	2.69	A	C
ATOM	1401	CA	LEU	A	202	54.816	31.843	63.819	1.00	2.11	A	C
ATOM	1409	CA	TYR	A	203	57.621	32.892	61.467	1.00	2.45	A	C
ATOM	1421	CA	TYR	A	204	60.210	30.396	60.264	1.00	2.82	A	C
ATOM	1433	CA	THR	A	205	63.573	30.086	58.500	1.00	2.58	A	C
ATOM	1440	CA	ALA	A	206	64.904	26.723	57.213	1.00	4.47	A	C
ATOM	1445	CA	ARG	A	207	68.253	26.305	55.508	1.00	5.09	A	C
ATOM	1456	CA	THR	A	208	69.773	23.209	53.912	1.00	4.88	A	C
ATOM	1463	CA	LEU	A	209	73.404	22.419	54.654	1.00	5.26	A	C
ATOM	1471	CA	GLN	A	210	73.983	20.900	51.175	1.00	6.01	A	C
ATOM	1480	CA	PRO	A	211	73.274	22.391	47.732	1.00	6.91	A	C
ATOM	1487	CA	ALA	A	212	69.689	21.962	46.560	1.00	7.18	A	C
ATOM	1492	CA	THR	A	213	67.307	23.100	43.847	1.00	5.83	A	C
ATOM	1499	CA	VAL	A	214	63.724	23.957	44.710	1.00	5.43	A	C
ATOM	1506	CA	GLN	A	215	60.145	24.166	43.470	1.00	5.10	A	C
ATOM	1515	CA	THR	A	216	57.130	25.576	45.286	1.00	4.16	A	C
ATOM	1522	CA	TRP	A	217	53.372	25.146	45.243	1.00	3.72	A	C
ATOM	1536	CA	GLY	A	218	50.356	27.288	46.105	1.00	3.16	A	C
ATOM	1540	CA	ASP	A	219	46.606	26.736	46.322	1.00	3.69	A	C
ATOM	1548	CA	ASP	A	220	46.180	25.735	42.659	1.00	3.99	A	C
ATOM	1556	CA	ASP	A	221	48.330	22.647	43.374	1.00	5.99	A	C
ATOM	1564	CA	ARG	A	222	50.800	23.609	40.598	1.00	6.95	A	C
ATOM	1575	CA	LEU	A	223	54.426	22.727	41.314	1.00	6.51	A	C
ATOM	1583	CA	VAL	A	224	56.418	25.552	39.779	1.00	7.29	A	C
ATOM	1590	CA	ASP	A	225	59.828	27.203	39.638	1.00	8.47	A	C
ATOM	1598	CA	ALA	A	226	58.537	30.421	41.281	1.00	8.13	A	C
ATOM	1603	CA	THR	A	227	60.445	31.382	44.417	1.00	7.82	A	C
ATOM	1610	CA	ALA	A	228	57.480	32.859	46.386	1.00	7.30	A	C
ATOM	1615	CA	GLN	A	229	54.054	31.642	47.513	1.00	5.21	A	C
ATOM	1624	CA	ASP	A	230	51.457	33.169	49.802	1.00	5.36	A	C
ATOM	1632	CA	GLY	A	231	48.063	31.590	50.521	1.00	3.52	A	C
ATOM	1636	CA	VAL	A	232	46.718	28.704	52.595	1.00	2.62	A	C
ATOM	1643	CA	ASP	A	233	48.105	25.470	51.111	1.00	3.49	A	C
ATOM	1651	CA	THR	A	234	51.701	26.281	50.149	1.00	3.46	A	C
ATOM	1658	CA	GLY	A	235	54.937	24.312	50.148	1.00	3.13	A	C
ATOM	1662	CA	ALA	A	236	58.416	23.735	48.763	1.00	3.15	A	C
ATOM	1667	CA	ILE	A	237	60.286	20.673	47.495	1.00	3.80	A	C
ATOM	1675	CA	LEU	A	238	64.059	20.549	47.934	1.00	4.57	A	C
ATOM	1683	CA	THR	A	239	66.059	18.244	45.649	1.00	6.16	A	C
ATOM	1690	CA	PHE	A	240	69.681	17.295	46.371	1.00	8.00	A	C
ATOM	1701	CA	ASP	A	241	72.537	15.737	44.401	1.00	10.79	A	C
ATOM	1709	CA	PRO	A	242	72.834	11.907	44.637	1.00	10.26	A	C
ATOM	1716	CA	ALA	A	243	76.443	12.489	45.768	1.00	9.54	A	C
ATOM	1721	CA	ASP	A	244	75.082	13.939	49.047	1.00	9.68	A	C
ATOM	1729	CA	ALA	A	245	72.792	10.984	49.778	1.00	9.01	A	C
ATOM	1734	CA	GLY	A	246	75.008	9.953	52.749	1.00	8.32	A	C
ATOM	1738	CA	GLU	A	247	74.244	13.241	54.603	1.00	8.02	A	C
ATOM	1747	CA	ILE	A	248	71.325	15.626	54.100	1.00	6.42	A	C
ATOM	1755	CA	GLY	A	249	71.023	18.320	56.806	1.00	5.33	A	C
ATOM	1759	CA	LEU	A	250	68.338	20.892	57.577	1.00	3.68	A	C
ATOM	1767	CA	GLN	A	251	67.982	23.542	60.272	1.00	3.91	A	C
ATOM	1776	CA	VAL	A	252	64.613	25.109	61.086	1.00	4.45	A	C
ATOM	1783	CA	THR	A	253	64.148	28.007	63.522	1.00	4.13	A	C
ATOM	1790	CA	LEU	A	254	60.850	29.555	64.744	1.00	3.15	A	C
ATOM	1798	CA	SER	A	255	60.066	33.075	65.957	1.00	3.13	A	C
ATOM	1804	CA	PRO	A	256	56.835	34.755	67.030	1.00	2.32	A	C
ATOM	1811	CA	VAL	A	257	58.462	38.034	65.945	1.00	2.47	A	C
ATOM	1818	CA	SER	A	258	59.667	37.831	62.318	1.00	3.37	A	C
ATOM	1824	CA	VAL	A	259	61.516	35.892	59.699	1.00	3.32	A	C
ATOM	1831	CA	GLU	A	260	64.595	38.024	60.217	1.00	4.54	A	C
ATOM	1840	CA	GLN	A	261	64.385	37.533	63.976	1.00	2.88	A	C
ATOM	1849	CA	ALA	A	262	64.235	33.748	63.289	1.00	3.15	A	C
ATOM	1854	CA	ARG	A	263	67.445	34.085	61.196	1.00	4.21	A	C
ATOM	1865	CA	ILE	A	264	69.210	35.928	64.012	1.00	4.33	A	C
ATOM	1873	CA	ASP	A	265	67.932	33.358	66.544	1.00	4.14	A	C
ATOM	1881	CA	GLN	A	266	69.289	30.623	64.311	1.00	6.08	A	C

ATOM	1890	CA	GLN	A	267	72.772	32.164	64.108	1.00	9.62	A	C
ATOM	1899	CA	VAL	A	268	72.748	32.711	67.907	1.00	8.96	A	C
ATOM	1906	CA	GLU	A	269	71.365	29.282	68.815	1.00	8.04	A	C
ATOM	1915	CA	LEU	A	270	73.089	27.042	66.271	1.00	8.55	A	C
ATOM	1923	CA	GLY	A	271	76.001	29.052	64.802	1.00	10.23	A	C
ATOM	1927	CA	ASP	A	272	78.321	26.600	63.009	1.00	10.69	A	C
ATOM	1935	CA	LEU	A	273	77.764	23.840	65.566	1.00	8.02	A	C
ATOM	1943	CA	SER	A	274	77.383	20.316	64.224	1.00	5.99	A	C
ATOM	1949	CA	PHE	A	275	74.219	18.210	64.582	1.00	4.80	A	C
ATOM	1960	CA	ASP	A	276	75.912	16.069	67.289	1.00	5.78	A	C
ATOM	1968	CA	ALA	A	277	76.925	19.139	69.337	1.00	5.08	A	C
ATOM	1973	CA	ILE	A	278	73.442	20.669	69.150	1.00	5.84	A	C
ATOM	1981	CA	ARG	A	279	71.924	17.315	70.206	1.00	6.83	A	C
ATOM	1992	CA	ASP	A	280	74.549	16.902	72.944	1.00	7.77	A	C
ATOM	2000	CA	ARG	A	281	74.023	20.429	74.293	1.00	7.74	A	C
ATOM	2011	CA	THR	A	282	70.270	19.953	74.599	1.00	6.38	A	C
ATOM	2018	CA	ARG	A	283	70.839	16.638	76.431	1.00	5.81	A	C
ATOM	2029	CA	ALA	A	284	73.047	18.504	78.940	1.00	5.99	A	C
ATOM	2034	CA	GLU	A	285	70.461	21.247	79.310	1.00	6.57	A	C
ATOM	2043	CA	TRP	A	286	67.904	18.490	80.139	1.00	5.67	A	C
ATOM	2057	CA	ASN	A	287	70.291	16.909	82.648	1.00	8.00	A	C
ATOM	2065	CA	ALA	A	288	70.657	20.275	84.422	1.00	8.86	A	C
ATOM	2070	CA	THR	A	289	66.862	20.740	84.464	1.00	7.09	A	C
ATOM	2077	CA	LEU	A	290	66.138	17.180	85.630	1.00	6.19	A	C
ATOM	2085	CA	GLY	A	291	69.085	17.378	88.069	1.00	6.65	A	C
ATOM	2089	CA	ARG	A	292	67.121	19.931	90.104	1.00	6.77	A	C
ATOM	2100	CA	VAL	A	293	65.530	16.844	91.707	1.00	8.09	A	C
ATOM	2107	CA	ALA	A	294	67.845	14.070	92.912	1.00	10.40	A	C
ATOM	2112	CA	ILE	A	295	66.318	10.811	94.046	1.00	13.23	A	C
ATOM	2120	CA	ASP	A	296	67.534	7.879	96.121	1.00	14.69	A	C
ATOM	2128	CA	ALA	A	297	65.001	5.043	95.685	1.00	12.88	A	C
ATOM	2133	CA	SER	A	298	65.612	1.701	97.466	1.00	11.58	A	C
ATOM	2139	CA	THR	A	299	64.657	-1.726	96.152	1.00	9.13	A	C
ATOM	2146	CA	ALA	A	300	61.955	-1.728	98.846	1.00	7.00	A	C
ATOM	2151	CA	THR	A	301	59.955	1.150	97.314	1.00	6.08	A	C
ATOM	2158	CA	ASP	A	302	61.022	0.632	93.655	1.00	6.93	A	C
ATOM	2166	CA	PRO	A	303	62.048	-3.016	93.116	1.00	7.98	A	C
ATOM	2173	CA	THR	A	304	62.107	-2.745	89.277	1.00	8.90	A	C
ATOM	2180	CA	GLY	A	305	63.438	0.816	88.893	1.00	8.26	A	C
ATOM	2184	CA	GLU	A	306	60.285	1.808	86.935	1.00	8.98	A	C
ATOM	2193	CA	LEU	A	307	59.007	4.206	89.571	1.00	6.57	A	C
ATOM	2201	CA	GLN	A	308	62.236	6.260	89.308	1.00	7.75	A	C
ATOM	2210	CA	ARG	A	309	61.928	6.228	85.486	1.00	7.83	A	C
ATOM	2221	CA	LEU	A	310	58.276	7.252	85.828	1.00	6.95	A	C
ATOM	2229	CA	PHE	A	311	59.288	10.140	88.071	1.00	5.21	A	C
ATOM	2240	CA	TYR	A	312	61.933	11.475	85.641	1.00	4.28	A	C
ATOM	2252	CA	THR	A	313	59.676	10.962	82.611	1.00	3.87	A	C
ATOM	2259	CA	HIS	A	314	56.982	13.080	84.288	1.00	4.29	A	C
ATOM	2269	CA	LEU	A	315	59.509	15.636	85.519	1.00	3.00	A	C
ATOM	2277	CA	TYR	A	316	60.512	15.912	81.827	1.00	2.15	A	C
ATOM	2289	CA	ARG	A	317	56.827	16.467	80.909	1.00	3.71	A	C
ATOM	2300	CA	MET	A	318	56.497	19.172	83.611	1.00	6.35	A	C
ATOM	2308	CA	PHE	A	319	58.967	21.419	81.712	1.00	7.83	A	C
ATOM	2319	CA	ALA	A	320	57.045	21.807	78.404	1.00	7.53	A	C
ATOM	2324	CA	MET	A	321	54.323	24.487	78.650	1.00	6.18	A	C
ATOM	2332	CA	PRO	A	322	54.063	27.434	78.638	1.00	5.02	A	C
ATOM	2339	CA	MET	A	323	56.762	27.568	75.935	1.00	4.44	A	C
ATOM	2347	CA	ASN	A	324	59.593	30.069	75.619	1.00	3.49	A	C
ATOM	2355	CA	ALA	A	325	57.989	32.716	73.341	1.00	2.49	A	C
ATOM	2360	CA	THR	A	326	61.012	35.098	73.515	1.00	2.79	A	C
ATOM	2367	CA	SER	A	327	63.597	35.861	70.819	1.00	3.68	A	C
ATOM	2373	CA	THR	A	328	67.381	35.744	71.383	1.00	4.17	A	C
ATOM	2380	CA	SER	A	329	67.191	39.550	71.284	1.00	4.95	A	C
ATOM	2386	CA	GLY	A	330	64.860	39.602	74.327	1.00	4.52	A	C
ATOM	2390	CA	THR	A	331	61.681	40.533	72.420	1.00	2.64	A	C
ATOM	2397	CA	TYR	A	332	58.212	38.956	72.097	1.00	2.99	A	C

ATOM	2409	CA	ARG	A	333	55.039	39.623	70.103	1.00	4.36	A	C
ATOM	2420	CA	GLY	A	334	52.050	40.974	71.967	1.00	4.85	A	C
ATOM	2424	CA	VAL	A	335	48.352	40.362	71.328	1.00	5.79	A	C
ATOM	2431	CA	ASP	A	336	48.362	43.923	69.904	1.00	8.91	A	C
ATOM	2439	CA	GLY	A	337	50.391	42.488	67.010	1.00	8.05	A	C
ATOM	2443	CA	ALA	A	338	53.464	44.524	67.878	1.00	7.17	A	C
ATOM	2448	CA	VAL	A	339	56.967	43.729	69.093	1.00	6.49	A	C
ATOM	2455	CA	HIS	A	340	57.792	44.405	72.780	1.00	8.62	A	C
ATOM	2465	CA	ALA	A	341	60.735	44.008	75.137	1.00	9.13	A	C
ATOM	2470	CA	ALA	A	342	60.679	41.217	77.711	1.00	11.07	A	C
ATOM	2475	CA	GLN	A	343	62.952	43.339	79.907	1.00	14.61	A	C
ATOM	2484	CA	GLY	A	344	64.997	41.352	82.439	1.00	14.36	A	C
ATOM	2488	CA	PHE	A	345	62.907	38.172	82.163	1.00	11.18	A	C
ATOM	2499	CA	THR	A	346	61.913	35.493	79.671	1.00	8.35	A	C
ATOM	2506	CA	TYR	A	347	58.429	35.786	78.204	1.00	5.52	A	C
ATOM	2518	CA	TYR	A	348	56.503	32.511	78.127	1.00	4.55	A	C
ATOM	2530	CA	ASP	A	349	53.244	31.880	76.254	1.00	3.32	A	C
ATOM	2538	CA	SER	A	350	50.780	29.020	75.499	1.00	3.95	A	C
ATOM	2544	CA	TRP	A	351	48.175	28.588	78.184	1.00	3.88	A	C
ATOM	2558	CA	ALA	A	352	45.515	26.000	79.064	1.00	3.39	A	C
ATOM	2563	CA	THR	A	353	44.865	27.385	82.495	1.00	3.82	A	C
ATOM	2570	CA	TRP	A	354	41.278	26.050	82.947	1.00	4.26	A	C
ATOM	2584	CA	ASP	A	355	42.871	22.556	82.994	1.00	4.99	A	C
ATOM	2592	CA	ASP	A	356	46.325	23.205	84.330	1.00	5.97	A	C
ATOM	2600	CA	PHE	A	357	46.209	25.899	87.012	1.00	5.02	A	C
ATOM	2611	CA	ARG	A	358	47.423	23.675	89.886	1.00	4.11	A	C
ATOM	2622	CA	LYS	A	359	50.623	22.752	87.999	1.00	2.82	A	C
ATOM	2631	CA	PHE	A	360	52.171	26.201	88.467	1.00	2.32	A	C
ATOM	2642	CA	SER	A	361	52.008	25.888	92.261	1.00	2.68	A	C
ATOM	2648	CA	VAL	A	362	54.293	22.814	91.937	1.00	3.61	A	C
ATOM	2655	CA	ILE	A	363	56.639	24.444	89.399	1.00	3.43	A	C
ATOM	2663	CA	ALA	A	364	57.066	27.142	92.096	1.00	3.80	A	C
ATOM	2668	CA	TYR	A	365	58.855	24.692	94.414	1.00	4.70	A	C
ATOM	2680	CA	ILE	A	366	60.869	22.863	91.729	1.00	6.01	A	C
ATOM	2688	CA	ASP	A	367	62.106	25.826	89.627	1.00	5.57	A	C
ATOM	2696	CA	PRO	A	368	61.194	29.124	91.362	1.00	3.96	A	C
ATOM	2703	CA	ALA	A	369	62.922	31.367	88.734	1.00	3.74	A	C
ATOM	2708	CA	LEU	A	370	61.088	29.740	85.833	1.00	3.88	A	C
ATOM	2716	CA	TYR	A	371	57.804	30.009	87.729	1.00	4.80	A	C
ATOM	2728	CA	ARG	A	372	58.430	33.689	88.384	1.00	5.45	A	C
ATOM	2739	CA	ASP	A	373	59.035	34.310	84.651	1.00	6.19	A	C
ATOM	2747	CA	MET	A	374	55.843	32.441	83.776	1.00	3.88	A	C
ATOM	2755	CA	VAL	A	375	53.730	34.523	86.171	1.00	2.79	A	C
ATOM	2762	CA	GLN	A	376	55.358	37.744	84.962	1.00	3.18	A	C
ATOM	2771	CA	SER	A	377	54.574	36.606	81.413	1.00	3.14	A	C
ATOM	2777	CA	LEU	A	378	50.953	35.908	82.336	1.00	3.14	A	C
ATOM	2785	CA	VAL	A	379	50.799	39.454	83.766	1.00	3.57	A	C
ATOM	2792	CA	TYR	A	380	52.193	40.950	80.508	1.00	4.06	A	C
ATOM	2804	CA	LEU	A	381	49.694	38.961	78.456	1.00	4.18	A	C
ATOM	2812	CA	PHE	A	382	46.697	40.475	80.286	1.00	4.57	A	C
ATOM	2823	CA	ALA	A	383	48.397	43.881	80.475	1.00	6.03	A	C
ATOM	2828	CA	ASP	A	384	48.684	43.708	76.664	1.00	7.44	A	C
ATOM	2836	CA	ALA	A	385	44.974	42.882	76.223	1.00	9.01	A	C
ATOM	2841	CA	GLU	A	386	44.199	45.935	78.362	1.00	10.91	A	C
ATOM	2850	CA	ALA	A	387	46.670	48.106	76.365	1.00	12.41	A	C
ATOM	2855	CA	THR	A	388	44.776	47.471	73.087	1.00	13.68	A	C
ATOM	2862	CA	GLY	A	389	41.944	49.514	74.624	1.00	14.39	A	C
ATOM	2866	CA	THR	A	390	39.452	47.191	72.889	1.00	15.63	A	C
ATOM	2873	CA	GLY	A	391	38.019	45.522	76.019	1.00	14.99	A	C
ATOM	2877	CA	GLY	A	392	37.896	42.282	73.993	1.00	13.79	A	C
ATOM	2881	CA	GLY	A	393	38.759	38.874	75.470	1.00	11.07	A	C
ATOM	2885	CA	LEU	A	394	42.017	37.084	74.740	1.00	7.81	A	C
ATOM	2893	CA	GLY	A	395	40.142	34.863	72.238	1.00	7.24	A	C
ATOM	2897	CA	GLY	A	396	39.767	37.812	69.847	1.00	7.80	A	C
ATOM	2901	CA	PHE	A	397	43.479	38.330	69.213	1.00	7.11	A	C
ATOM	2912	CA	VAL	A	398	46.038	36.458	67.145	1.00	5.79	A	C

ATOM	2919	CA	HIS	A	399	48.445	34.799	69.624	1.00	3.71	A	C
ATOM	2929	CA	SER	A	400	52.295	34.534	69.502	1.00	3.42	A	C
ATOM	2935	CA	VAL	A	401	52.383	30.703	69.819	1.00	2.51	A	C
ATOM	2942	CA	PRO	A	402	49.744	27.945	69.645	1.00	3.03	A	C
ATOM	2949	CA	THR	A	403	47.621	28.327	72.753	1.00	2.63	A	C
ATOM	2956	CA	VAL	A	404	44.423	27.083	74.389	1.00	2.59	A	C
ATOM	2963	CA	ARG	A	405	42.502	28.941	77.121	1.00	2.87	A	C
ATOM	2974	CA	TRP	A	406	43.027	31.341	80.033	1.00	3.31	A	C
ATOM	2988	CA	GLU	A	407	40.430	30.754	82.763	1.00	4.40	A	C
ATOM	2997	CA	ARG	A	408	41.810	30.579	86.344	1.00	3.53	A	C
ATOM	3008	CA	SER	A	409	44.708	32.938	85.445	1.00	3.56	A	C
ATOM	3014	CA	SER	A	410	43.670	35.101	88.462	1.00	4.17	A	C
ATOM	3020	CA	VAL	A	411	44.366	32.099	90.707	1.00	3.23	A	C
ATOM	3027	CA	VAL	A	412	47.816	31.532	89.197	1.00	2.11	A	C
ATOM	3034	CA	VAL	A	413	48.819	35.209	89.648	1.00	2.29	A	C
ATOM	3041	CA	ALA	A	414	47.478	35.051	93.214	1.00	2.19	A	C
ATOM	3046	CA	ASP	A	415	49.593	31.906	93.728	1.00	3.52	A	C
ATOM	3054	CA	ALA	A	416	52.816	33.913	93.216	1.00	3.54	A	C
ATOM	3059	CA	ILE	A	417	51.560	36.792	95.351	1.00	4.96	A	C
ATOM	3067	CA	ALA	A	418	50.526	34.493	98.238	1.00	7.10	A	C
ATOM	3072	CA	LYS	A	419	53.976	32.864	97.904	1.00	7.37	A	C
ATOM	3081	CA	GLY	A	420	55.694	36.259	98.483	1.00	8.91	A	C
ATOM	3085	CA	PHE	A	421	56.789	37.036	94.914	1.00	9.86	A	C
ATOM	3096	CA	ASP	A	422	56.741	40.741	94.170	1.00	13.61	A	C
ATOM	3104	CA	GLY	A	423	57.809	43.423	91.676	1.00	13.34	A	C
ATOM	3108	CA	PHE	A	424	55.491	42.086	88.966	1.00	11.76	A	C
ATOM	3119	CA	ASP	A	425	55.749	44.833	86.387	1.00	12.47	A	C
ATOM	3127	CA	ARG	A	426	52.408	45.949	84.848	1.00	10.18	A	C
ATOM	3138	CA	LEU	A	427	50.173	44.147	87.370	1.00	9.19	A	C
ATOM	3146	CA	ASP	A	428	48.058	47.323	87.662	1.00	10.66	A	C
ATOM	3154	CA	GLU	A	429	47.378	47.038	83.880	1.00	10.07	A	C
ATOM	3163	CA	ALA	A	430	46.738	43.286	83.995	1.00	8.16	A	C
ATOM	3168	CA	TYR	A	431	44.091	43.690	86.747	1.00	8.25	A	C
ATOM	3180	CA	PRO	A	432	41.138	45.159	84.775	1.00	8.05	A	C
ATOM	3187	CA	ALA	A	433	41.670	42.554	82.030	1.00	7.78	A	C
ATOM	3192	CA	LEU	A	434	41.696	39.872	84.724	1.00	7.30	A	C
ATOM	3200	CA	GLN	A	435	38.424	41.359	86.060	1.00	9.52	A	C
ATOM	3209	CA	ARG	A	436	36.760	41.034	82.610	1.00	8.83	A	C
ATOM	3220	CA	LEU	A	437	38.067	37.458	82.349	1.00	8.34	A	C
ATOM	3228	CA	VAL	A	438	36.448	36.553	85.664	1.00	9.83	A	C
ATOM	3235	CA	GLY	A	439	33.227	38.399	84.746	1.00	9.87	A	C
ATOM	3239	CA	GLN	A	440	30.557	39.769	87.076	1.00	11.48	A	C
ATOM	3248	CA	TYR	A	441	27.387	38.162	88.403	1.00	9.32	A	C
ATOM	3260	CA	SER	A	442	24.326	39.771	86.747	1.00	9.01	A	C
ATOM	3266	CA	ALA	A	443	21.930	41.944	88.804	1.00	10.00	A	C
ATOM	3271	CA	ASP	A	444	19.645	38.962	89.504	1.00	11.36	A	C
ATOM	3279	CA	GLU	A	445	22.683	36.814	90.345	1.00	10.65	A	C
ATOM	3288	CA	LEU	A	446	24.057	39.420	92.752	1.00	12.13	A	C
ATOM	3296	CA	ARG	A	447	20.688	39.484	94.519	1.00	14.84	A	C
ATOM	3307	CA	ARG	A	448	20.411	35.689	94.956	1.00	13.37	A	C
ATOM	3318	CA	GLY	A	449	24.148	34.933	95.289	1.00	11.12	A	C
ATOM	3322	CA	TYR	A	450	24.570	32.356	92.478	1.00	11.01	A	C
ATOM	3334	CA	VAL	A	451	24.024	31.514	88.794	1.00	10.44	A	C
ATOM	3341	CA	ALA	A	452	20.597	29.813	88.469	1.00	11.06	A	C
ATOM	3346	CA	GLY	A	453	20.832	26.118	87.493	1.00	10.92	A	C
ATOM	3350	CA	ASN	A	454	24.597	26.415	86.920	1.00	9.68	A	C
ATOM	3358	CA	PRO	A	455	26.517	25.184	90.002	1.00	9.01	A	C
ATOM	3365	CA	GLY	A	456	29.710	24.813	87.923	1.00	7.30	A	C
ATOM	3369	CA	ALA	A	457	29.902	28.429	86.804	1.00	5.98	A	C
ATOM	3374	CA	SER	A	458	29.216	29.530	90.428	1.00	5.00	A	C
ATOM	3380	CA	VAL	A	459	32.021	27.538	92.097	1.00	3.77	A	C
ATOM	3387	CA	GLN	A	460	34.377	28.579	89.287	1.00	3.69	A	C
ATOM	3396	CA	ARG	A	461	33.738	32.298	89.725	1.00	3.05	A	C
ATOM	3407	CA	GLY	A	462	34.143	31.742	93.464	1.00	2.45	A	C
ATOM	3411	CA	TYR	A	463	37.707	30.519	92.994	1.00	3.51	A	C
ATOM	3423	CA	ASP	A	464	38.419	33.243	90.372	1.00	3.80	A	C

ATOM	3431	CA	GLN	A	465	37.202	35.909	92.790	1.00	4.82	A	C
ATOM	3440	CA	TYR	A	466	39.300	34.526	95.621	1.00	5.23	A	C
ATOM	3452	CA	GLY	A	467	42.325	34.871	93.265	1.00	5.67	A	C
ATOM	3456	CA	LEU	A	468	41.385	38.365	92.150	1.00	6.17	A	C
ATOM	3464	CA	SER	A	469	40.991	39.304	95.828	1.00	5.97	A	C
ATOM	3470	CA	VAL	A	470	44.682	38.455	96.482	1.00	6.48	A	C
ATOM	3477	CA	ILE	A	471	45.705	40.579	93.461	1.00	6.17	A	C
ATOM	3485	CA	ALA	A	472	43.411	43.420	94.600	1.00	8.07	A	C
ATOM	3490	CA	ASP	A	473	45.060	43.606	98.080	1.00	10.08	A	C
ATOM	3498	CA	GLU	A	474	48.496	43.569	96.443	1.00	10.69	A	C
ATOM	3507	CA	LEU	A	475	47.508	46.603	94.342	1.00	11.15	A	C
ATOM	3515	CA	GLY	A	476	46.177	48.346	97.453	1.00	13.34	A	C
ATOM	3519	CA	LEU	A	477	42.538	48.006	96.344	1.00	15.39	A	C
ATOM	3527	CA	THR	A	478	41.434	47.034	99.867	1.00	17.62	A	C
ATOM	3534	CA	GLU	A	479	37.723	47.742	99.394	1.00	18.74	A	C
ATOM	3543	CA	GLU	A	480	37.595	45.763	96.110	1.00	16.33	A	C
ATOM	3552	CA	ALA	A	481	39.372	42.864	97.813	1.00	13.75	A	C
ATOM	3557	CA	GLU	A	482	36.715	42.720	100.595	1.00	14.63	A	C
ATOM	3566	CA	THR	A	483	33.842	42.673	98.101	1.00	12.95	A	C
ATOM	3573	CA	LEU	A	484	35.614	39.932	96.121	1.00	11.73	A	C
ATOM	3581	CA	ARG	A	485	36.127	37.758	99.233	1.00	11.05	A	C
ATOM	3592	CA	GLU	A	486	32.458	38.265	100.146	1.00	10.61	A	C
ATOM	3601	CA	GLN	A	487	31.331	37.147	96.664	1.00	8.60	A	C
ATOM	3610	CA	ALA	A	488	33.786	34.225	96.759	1.00	6.52	A	C
ATOM	3615	CA	SER	A	489	32.029	32.893	99.875	1.00	7.12	A	C
ATOM	3621	CA	TRP	A	490	28.661	32.726	98.010	1.00	7.33	A	C
ATOM	3635	CA	PRO	A	491	28.980	29.421	96.072	1.00	6.73	A	C
ATOM	3642	CA	ILE	A	492	29.748	27.439	99.257	1.00	8.58	A	C
ATOM	3650	CA	GLU	A	493	27.034	29.091	101.368	1.00	11.10	A	C
ATOM	3659	CA	LYS	A	494	24.283	29.217	98.744	1.00	10.38	A	C
ATOM	3668	CA	LEU	A	495	24.772	25.992	96.727	1.00	8.08	A	C
ATOM	3676	CA	THR	A	496	25.642	23.390	99.397	1.00	9.94	A	C
ATOM	3683	CA	LYS	A	497	22.269	21.696	99.874	1.00	10.32	A	C
ATOM	3692	CA	PRO	A	498	21.844	20.282	103.387	1.00	9.82	A	C
ATOM	3699	CA	GLY	A	499	21.074	16.559	103.567	1.00	10.14	A	C
ATOM	3703	CA	ALA	A	500	21.838	16.042	99.846	1.00	10.82	A	C
ATOM	3708	CA	TRP	A	501	23.386	12.728	100.863	1.00	11.71	A	C
ATOM	3722	CA	THR	A	502	22.581	10.696	104.006	1.00	12.62	A	C
ATOM	3729	CA	ALA	A	503	25.365	8.647	105.597	1.00	13.78	A	C
ATOM	3734	CA	ALA	A	504	24.951	4.994	106.737	1.00	15.02	A	C
ATOM	3739	CA	ASP	A	505	24.357	6.130	110.325	1.00	14.83	A	C
ATOM	3747	CA	GLY	A	506	21.856	8.861	109.305	1.00	12.52	A	C
ATOM	3751	CA	THR	A	507	24.298	11.810	109.343	1.00	10.12	A	C
ATOM	3758	CA	GLN	A	508	23.119	14.588	106.989	1.00	8.56	A	C
ATOM	3767	CA	VAL	A	509	25.765	15.604	104.451	1.00	6.31	A	C
ATOM	3774	CA	GLY	A	510	25.468	18.758	102.362	1.00	5.03	A	C
ATOM	3778	CA	LEU	A	511	26.594	18.750	98.722	1.00	4.48	A	C
ATOM	3786	CA	LEU	A	512	26.884	21.120	95.793	1.00	5.45	A	C
ATOM	3794	CA	THR	A	513	23.470	21.012	94.071	1.00	7.09	A	C
ATOM	3801	CA	PRO	A	514	21.941	23.109	91.250	1.00	7.88	A	C
ATOM	3808	CA	ARG	A	515	19.517	25.737	92.573	1.00	9.63	A	C
ATOM	3819	CA	ALA	A	516	16.670	27.438	90.743	1.00	11.40	A	C
ATOM	3824	CA	ALA	A	517	16.201	31.213	90.509	1.00	13.83	A	C
ATOM	3829	CA	ASP	A	518	13.518	31.125	93.261	1.00	16.58	A	C
ATOM	3837	CA	GLY	A	519	15.865	29.277	95.648	1.00	16.47	A	C
ATOM	3841	CA	SER	A	520	14.202	25.852	95.069	1.00	15.99	A	C
ATOM	3847	CA	TRP	A	521	16.785	23.089	94.870	1.00	13.14	A	C
ATOM	3861	CA	GLN	A	522	16.878	21.329	91.494	1.00	12.01	A	C
ATOM	3870	CA	SER	A	523	16.780	17.550	91.127	1.00	12.74	A	C
ATOM	3876	CA	ALA	A	524	20.170	15.972	90.267	1.00	11.18	A	C
ATOM	3881	CA	ASP	A	525	22.087	12.671	90.488	1.00	8.84	A	C
ATOM	3889	CA	HIS	A	526	25.271	13.887	92.225	1.00	6.57	A	C
ATOM	3899	CA	ALA	A	527	27.293	11.003	90.717	1.00	6.14	A	C
ATOM	3904	CA	LYS	A	528	26.151	11.481	87.106	1.00	7.50	A	C
ATOM	3913	CA	PHE	A	529	28.818	12.767	84.690	1.00	7.13	A	C
ATOM	3924	CA	GLU	A	530	27.867	16.202	83.304	1.00	7.91	A	C

ATOM	3933	CA	ALA	A	531	24.533	16.513	85.106	1.00	8.44	A	C
ATOM	3938	CA	ALA	A	532	23.397	20.032	86.170	1.00	7.77	A	C
ATOM	3943	CA	GLY	A	533	25.194	21.889	83.332	1.00	6.76	A	C
ATOM	3947	CA	LEU	A	534	28.619	20.734	84.578	1.00	5.97	A	C
ATOM	3955	CA	TYR	A	535	31.682	20.411	82.372	1.00	8.46	A	C
ATOM	3967	CA	GLN	A	536	33.627	17.123	82.410	1.00	6.34	A	C
ATOM	3976	CA	GLY	A	537	32.521	16.050	85.901	1.00	4.04	A	C
ATOM	3980	CA	THR	A	538	29.936	15.281	88.564	1.00	3.50	A	C
ATOM	3987	CA	LEU	A	539	28.500	17.232	91.498	1.00	3.71	A	C
ATOM	3995	CA	TRP	A	540	30.500	15.072	93.986	1.00	3.80	A	C
ATOM	4009	CA	GLN	A	541	33.680	16.012	92.073	1.00	3.42	A	C
ATOM	4018	CA	TYR	A	542	32.808	19.714	91.708	1.00	4.03	A	C
ATOM	4030	CA	HIS	A	543	31.606	19.795	95.352	1.00	4.44	A	C
ATOM	4040	CA	TRP	A	544	35.221	20.315	96.537	1.00	4.40	A	C
ATOM	4054	CA	TYR	A	545	36.045	23.019	93.950	1.00	4.18	A	C
ATOM	4066	CA	ASP	A	546	36.276	26.024	96.237	1.00	5.25	A	C
ATOM	4074	CA	ALA	A	547	39.546	24.793	97.671	1.00	5.51	A	C
ATOM	4079	CA	TYR	A	548	40.358	28.217	99.183	1.00	6.10	A	C
ATOM	4091	CA	ASP	A	549	37.753	27.586	101.917	1.00	6.22	A	C
ATOM	4099	CA	MET	A	550	37.554	24.017	103.177	1.00	6.61	A	C
ATOM	4107	CA	ASP	A	551	36.549	25.382	106.586	1.00	8.26	A	C
ATOM	4115	CA	ALA	A	552	33.422	27.067	105.225	1.00	7.80	A	C
ATOM	4120	CA	LEU	A	553	32.800	24.021	103.028	1.00	7.77	A	C
ATOM	4128	CA	VAL	A	554	32.983	21.675	106.066	1.00	8.48	A	C
ATOM	4135	CA	GLU	A	555	30.421	23.906	107.886	1.00	10.05	A	C
ATOM	4144	CA	ALA	A	556	28.101	24.172	104.869	1.00	8.14	A	C
ATOM	4149	CA	MET	A	557	28.262	20.355	104.586	1.00	7.75	A	C
ATOM	4157	CA	GLY	A	558	26.833	20.080	108.112	1.00	7.77	A	C
ATOM	4161	CA	GLY	A	559	29.995	19.989	110.230	1.00	8.36	A	C
ATOM	4165	CA	HIS	A	560	33.086	17.891	110.775	1.00	10.24	A	C
ATOM	4175	CA	GLU	A	561	31.496	14.448	111.036	1.00	10.61	A	C
ATOM	4184	CA	ALA	A	562	29.420	15.118	107.900	1.00	9.17	A	C
ATOM	4189	CA	ALA	A	563	32.594	16.099	105.943	1.00	7.47	A	C
ATOM	4194	CA	ARG	A	564	34.534	13.129	107.304	1.00	6.15	A	C
ATOM	4205	CA	LEU	A	565	31.813	10.683	106.236	1.00	5.27	A	C
ATOM	4213	CA	GLY	A	566	31.408	12.469	102.857	1.00	4.45	A	C
ATOM	4217	CA	MET	A	567	35.136	12.086	102.137	1.00	5.05	A	C
ATOM	4225	CA	ARG	A	568	35.000	8.412	103.020	1.00	6.55	A	C
ATOM	4236	CA	HIS	A	569	31.985	7.943	100.689	1.00	7.31	A	C
ATOM	4246	CA	MET	A	570	33.786	9.924	97.906	1.00	7.92	A	C
ATOM	4254	CA	PHE	A	571	36.388	7.150	97.840	1.00	7.04	A	C
ATOM	4265	CA	GLY	A	572	34.058	4.168	98.426	1.00	8.17	A	C
ATOM	4269	CA	GLU	A	573	35.790	3.316	101.736	1.00	10.14	A	C
ATOM	4278	CA	HIS	A	574	32.693	1.377	102.888	1.00	11.27	A	C
ATOM	4288	CA	ALA	A	575	32.914	-0.845	99.770	1.00	11.91	A	C
ATOM	4293	CA	PRO	A	576	36.615	-1.198	98.736	1.00	13.14	A	C
ATOM	4300	CA	ASP	A	577	35.923	-3.909	96.131	1.00	13.69	A	C
ATOM	4308	CA	ASP	A	578	32.994	-2.108	94.404	1.00	12.41	A	C
ATOM	4316	CA	GLY	A	579	34.247	-0.097	91.423	1.00	11.12	A	C
ATOM	4320	CA	LYS	A	580	30.905	1.748	91.259	1.00	11.21	A	C
ATOM	4329	CA	ALA	A	581	31.370	3.276	94.757	1.00	8.19	A	C
ATOM	4334	CA	MET	A	582	34.358	5.303	93.506	1.00	5.86	A	C
ATOM	4342	CA	LEU	A	583	33.022	8.889	93.198	1.00	4.23	A	C
ATOM	4350	CA	HIS	A	584	36.434	10.466	92.671	1.00	3.37	A	C
ATOM	4360	CA	SER	A	585	37.816	10.665	89.125	1.00	3.12	A	C
ATOM	4366	CA	ASN	A	586	41.508	10.091	88.359	1.00	3.90	A	C
ATOM	4374	CA	ALA	A	587	41.103	10.824	84.638	1.00	5.72	A	C
ATOM	4379	CA	ASN	A	588	40.425	14.577	84.853	1.00	7.03	A	C
ATOM	4387	CA	GLU	A	589	41.649	17.614	86.824	1.00	7.17	A	C
ATOM	4396	CA	ILE	A	590	38.295	18.647	88.338	1.00	7.24	A	C
ATOM	4404	CA	ASP	A	591	38.863	16.848	91.650	1.00	7.24	A	C
ATOM	4412	CA	LEU	A	592	42.668	16.421	91.471	1.00	6.70	A	C
ATOM	4420	CA	GLN	A	593	42.981	17.626	95.053	1.00	4.59	A	C
ATOM	4429	CA	ALA	A	594	40.356	15.206	96.428	1.00	4.71	A	C
ATOM	4434	CA	PRO	A	595	42.784	12.503	97.633	1.00	5.03	A	C
ATOM	4441	CA	TYR	A	596	44.506	15.069	99.845	1.00	6.02	A	C

ATOM	4453	CA	LEU	A	597	41.304	16.226	101.591	1.00	4.94	A	C
ATOM	4461	CA	PHE	A	598	41.417	13.474	104.213	1.00	5.47	A	C
ATOM	4472	CA	ASN	A	599	44.037	15.684	105.929	1.00	6.53	A	C
ATOM	4480	CA	TYR	A	600	41.176	18.148	106.491	1.00	7.92	A	C
ATOM	4492	CA	THR	A	601	38.801	15.556	107.938	1.00	9.25	A	C
ATOM	4499	CA	GLY	A	602	41.334	14.393	110.555	1.00	9.27	A	C
ATOM	4503	CA	GLU	A	603	42.259	11.247	108.634	1.00	8.63	A	C
ATOM	4512	CA	PRO	A	604	45.684	11.982	107.132	1.00	7.11	A	C
ATOM	4519	CA	SER	A	605	46.475	8.224	106.915	1.00	5.46	A	C
ATOM	4525	CA	LEU	A	606	43.781	7.928	104.201	1.00	4.96	A	C
ATOM	4533	CA	THR	A	607	45.390	10.729	102.175	1.00	4.57	A	C
ATOM	4540	CA	GLN	A	608	48.682	8.824	102.399	1.00	5.02	A	C
ATOM	4549	CA	LYS	A	609	47.008	5.602	101.265	1.00	5.44	A	C
ATOM	4558	CA	TRP	A	610	45.214	7.208	98.331	1.00	5.42	A	C
ATOM	4572	CA	ALA	A	611	48.211	9.266	97.205	1.00	6.31	A	C
ATOM	4577	CA	ARG	A	612	50.436	6.184	97.099	1.00	5.71	A	C
ATOM	4588	CA	ALA	A	613	47.643	4.074	95.492	1.00	4.20	A	C
ATOM	4593	CA	ILE	A	614	46.445	6.379	92.691	1.00	4.67	A	C
ATOM	4601	CA	TYR	A	615	49.973	7.212	91.557	1.00	4.85	A	C
ATOM	4613	CA	THR	A	616	51.897	3.953	92.071	1.00	4.62	A	C
ATOM	4620	CA	LYS	A	617	49.365	1.088	92.368	1.00	6.36	A	C
ATOM	4629	CA	GLU	A	618	46.132	-0.236	90.829	1.00	7.91	A	C
ATOM	4638	CA	THR	A	619	42.939	1.583	91.795	1.00	6.88	A	C
ATOM	4645	CA	TRP	A	620	39.227	1.357	91.008	1.00	7.46	A	C
ATOM	4659	CA	ASN	A	621	38.296	3.692	88.170	1.00	6.73	A	C
ATOM	4667	CA	ARG	A	622	34.663	4.574	87.576	1.00	4.87	A	C
ATOM	4678	CA	TYR	A	623	35.087	7.709	85.473	1.00	5.03	A	C
ATOM	4690	CA	ILE	A	624	36.716	8.661	82.149	1.00	5.64	A	C
ATOM	4698	CA	ALA	A	625	37.316	12.291	81.127	1.00	6.35	A	C
ATOM	4703	CA	THR	A	626	34.590	12.516	78.472	1.00	6.19	A	C
ATOM	4710	CA	GLY	A	627	31.208	10.937	77.589	1.00	5.93	A	C
ATOM	4714	CA	SER	A	628	32.592	7.840	75.913	1.00	7.00	A	C
ATOM	4720	CA	SER	A	629	35.720	6.302	74.355	1.00	9.77	A	C
ATOM	4726	CA	SER	A	630	36.292	3.632	71.688	1.00	11.33	A	C
ATOM	4732	CA	ALA	A	631	39.492	2.767	73.559	1.00	10.90	A	C
ATOM	4737	CA	VAL	A	632	37.659	1.102	76.492	1.00	8.53	A	C
ATOM	4744	CA	PRO	A	633	34.094	-0.107	77.097	1.00	7.28	A	C
ATOM	4751	CA	SER	A	634	32.472	3.099	78.345	1.00	5.04	A	C
ATOM	4757	CA	GLY	A	635	29.414	5.345	78.326	1.00	4.96	A	C
ATOM	4761	CA	GLY	A	636	27.832	8.244	80.176	1.00	6.17	A	C
ATOM	4765	CA	GLY	A	637	31.304	9.416	81.298	1.00	5.99	A	C
ATOM	4769	CA	GLU	A	638	32.099	6.071	82.923	1.00	5.01	A	C
ATOM	4778	CA	PHE	A	639	34.109	2.868	82.500	1.00	4.24	A	C
ATOM	4789	CA	THR	A	640	31.342	0.261	81.956	1.00	5.80	A	C
ATOM	4796	CA	PRO	A	641	32.109	-1.741	83.969	1.00	6.76	A	C
ATOM	4803	CA	PRO	A	642	34.393	0.103	86.417	1.00	8.12	A	C
ATOM	4810	CA	LEU	A	643	38.016	-0.971	85.969	1.00	9.82	A	C
ATOM	4818	CA	LYS	A	644	40.825	-1.805	88.435	1.00	11.06	A	C
ATOM	4827	CA	THR	A	645	43.998	-0.565	86.793	1.00	8.37	A	C
ATOM	4834	CA	LYS	A	646	47.067	1.637	87.273	1.00	7.54	A	C
ATOM	4843	CA	VAL	A	647	46.307	5.272	86.488	1.00	5.75	A	C
ATOM	4850	CA	TYR	A	648	49.922	5.925	85.527	1.00	5.56	A	C
ATOM	4862	CA	ARG	A	649	51.964	3.328	83.533	1.00	7.76	A	C
ATOM	4873	CA	LEU	A	650	55.512	3.322	82.177	1.00	6.76	A	C
ATOM	4881	CA	ASP	A	651	54.026	2.407	78.809	1.00	6.42	A	C
ATOM	4889	CA	PRO	A	652	53.043	4.126	75.570	1.00	5.92	A	C
ATOM	4896	CA	ARG	A	653	49.549	3.680	77.020	1.00	4.60	A	C
ATOM	4907	CA	GLY	A	654	50.724	6.058	79.774	1.00	4.89	A	C
ATOM	4911	CA	MET	A	655	47.517	7.356	81.356	1.00	4.93	A	C
ATOM	4919	CA	LEU	A	656	43.963	6.075	81.567	1.00	3.04	A	C
ATOM	4927	CA	PRO	A	657	42.105	6.134	78.251	1.00	3.10	A	C
ATOM	4934	CA	THR	A	658	40.754	9.734	77.676	1.00	3.53	A	C
ATOM	4941	CA	MET	A	659	43.088	11.046	80.398	1.00	3.65	A	C
ATOM	4949	CA	ASP	A	660	44.895	13.859	78.555	1.00	3.95	A	C
ATOM	4957	CA	ASN	A	661	48.031	15.161	80.276	1.00	3.15	A	C
ATOM	4965	CA	ASP	A	662	46.843	18.761	79.522	1.00	2.87	A	C

ATOM	4973	CA	ALA	A	663	50.075	20.739	79.098	1.00	3.12	A	C
ATOM	4978	CA	GLY	A	664	51.894	18.459	81.548	1.00	2.70	A	C
ATOM	4982	CA	THR	A	665	49.328	18.721	84.376	1.00	2.81	A	C
ATOM	4989	CA	MET	A	666	48.910	14.993	85.026	1.00	2.55	A	C
ATOM	4997	CA	SER	A	667	52.753	14.620	84.897	1.00	2.87	A	C
ATOM	5003	CA	THR	A	668	53.193	17.564	87.294	1.00	3.50	A	C
ATOM	5010	CA	MET	A	669	50.719	16.032	89.771	1.00	2.90	A	C
ATOM	5018	CA	PHE	A	670	52.751	12.810	89.639	1.00	2.90	A	C
ATOM	5029	CA	VAL	A	671	55.921	14.716	90.519	1.00	3.29	A	C
ATOM	5036	CA	ALA	A	672	54.049	16.446	93.385	1.00	3.04	A	C
ATOM	5041	CA	ALA	A	673	52.740	13.126	94.733	1.00	3.21	A	C
ATOM	5046	CA	ALA	A	674	56.289	11.665	94.548	1.00	4.16	A	C
ATOM	5051	CA	VAL	A	675	57.770	14.572	96.510	1.00	3.68	A	C
ATOM	5058	CA	GLY	A	676	54.729	14.329	98.773	1.00	3.81	A	C
ATOM	5062	CA	LEU	A	677	53.628	17.976	98.689	1.00	3.63	A	C
ATOM	5070	CA	PHE	A	678	50.401	18.865	96.910	1.00	4.98	A	C
ATOM	5081	CA	PRO	A	679	48.736	22.236	96.308	1.00	4.89	A	C
ATOM	5088	CA	VAL	A	680	45.131	21.802	97.449	1.00	3.54	A	C
ATOM	5095	CA	THR	A	681	44.815	25.594	97.517	1.00	3.01	A	C
ATOM	5102	CA	ALA	A	682	46.410	27.062	94.421	1.00	2.78	A	C
ATOM	5107	CA	GLY	A	683	46.558	30.790	95.067	1.00	4.10	A	C
ATOM	5111	CA	SER	A	684	47.895	30.234	98.585	1.00	5.30	A	C
ATOM	5117	CA	SER	A	685	51.390	29.528	99.913	1.00	6.30	A	C
ATOM	5123	CA	GLN	A	686	50.476	26.083	101.310	1.00	5.66	A	C
ATOM	5132	CA	PHE	A	687	51.101	22.447	100.320	1.00	5.37	A	C
ATOM	5143	CA	GLN	A	688	49.438	19.356	101.748	1.00	4.94	A	C
ATOM	5152	CA	VAL	A	689	51.517	16.390	102.936	1.00	5.79	A	C
ATOM	5159	CA	GLY	A	690	50.928	13.088	101.117	1.00	6.69	A	C
ATOM	5163	CA	SER	A	691	52.974	9.889	100.906	1.00	7.27	A	C
ATOM	5169	CA	PRO	A	692	56.528	10.626	99.669	1.00	6.85	A	C
ATOM	5176	CA	PHE	A	693	57.581	7.765	97.347	1.00	6.34	A	C
ATOM	5187	CA	PHE	A	694	61.346	7.747	97.796	1.00	6.87	A	C
ATOM	5198	CA	ASP	A	695	63.891	7.202	100.575	1.00	7.56	A	C
ATOM	5206	CA	SER	A	696	65.365	10.632	99.830	1.00	6.91	A	C
ATOM	5212	CA	THR	A	697	64.278	13.400	97.424	1.00	7.09	A	C
ATOM	5219	CA	THR	A	698	66.465	16.572	97.123	1.00	7.46	A	C
ATOM	5226	CA	ILE	A	699	65.459	19.752	95.308	1.00	6.95	A	C
ATOM	5234	CA	THR	A	700	68.553	21.914	94.670	1.00	8.40	A	C
ATOM	5241	CA	TYR	A	701	67.913	25.630	94.091	1.00	9.41	A	C
ATOM	5253	CA	ASP	A	702	69.971	28.028	91.946	1.00	11.48	A	C
ATOM	5261	CA	ASP	A	703	72.068	29.199	94.915	1.00	13.97	A	C
ATOM	5269	CA	GLY	A	704	73.018	25.617	95.847	1.00	13.70	A	C
ATOM	5273	CA	SER	A	705	70.765	25.419	98.907	1.00	12.46	A	C
ATOM	5279	CA	ALA	A	706	68.419	22.383	98.854	1.00	10.18	A	C
ATOM	5284	CA	PHE	A	707	65.052	21.131	100.137	1.00	7.55	A	C
ATOM	5295	CA	THR	A	708	65.351	17.581	101.387	1.00	7.30	A	C
ATOM	5302	CA	VAL	A	709	62.397	15.352	102.091	1.00	7.65	A	C
ATOM	5309	CA	THR	A	710	63.206	11.874	103.424	1.00	8.06	A	C
ATOM	5316	CA	ALA	A	711	60.926	8.889	103.973	1.00	8.76	A	C
ATOM	5321	CA	ASP	A	712	62.785	6.541	106.270	1.00	10.03	A	C
ATOM	5329	CA	GLY	A	713	61.790	2.931	105.687	1.00	7.18	A	C
ATOM	5333	CA	VAL	A	714	59.420	3.754	102.770	1.00	5.21	A	C
ATOM	5340	CA	SER	A	715	58.461	0.754	100.621	1.00	5.75	A	C
ATOM	5346	CA	GLU	A	716	55.621	-0.434	98.399	1.00	7.40	A	C
ATOM	5355	CA	ASP	A	717	53.911	-1.492	101.619	1.00	7.77	A	C
ATOM	5363	CA	ALA	A	718	55.257	1.143	103.999	1.00	5.28	A	C
ATOM	5368	CA	PHE	A	719	53.472	4.224	102.665	1.00	3.86	A	C
ATOM	5379	CA	TYR	A	720	51.935	5.573	105.926	1.00	5.07	A	C
ATOM	5391	CA	VAL	A	721	53.647	8.305	107.943	1.00	6.16	A	C
ATOM	5398	CA	GLN	A	722	54.378	7.196	111.544	1.00	8.68	A	C
ATOM	5407	CA	SER	A	723	56.107	10.362	112.750	1.00	9.44	A	C
ATOM	5413	CA	ALA	A	724	57.936	13.385	111.311	1.00	8.65	A	C
ATOM	5418	CA	THR	A	725	60.561	16.054	112.006	1.00	8.30	A	C
ATOM	5425	CA	LEU	A	726	61.170	19.486	110.467	1.00	8.74	A	C
ATOM	5433	CA	ASP	A	727	64.804	20.662	110.723	1.00	9.93	A	C
ATOM	5441	CA	GLY	A	728	65.330	18.182	113.597	1.00	11.60	A	C

ATOM	5445	CA	ALA	A	729	62.241	19.122	115.635	1.00	11.25	A	C
ATOM	5450	CA	THR	A	730	58.993	17.157	116.102	1.00	10.67	A	C
ATOM	5457	CA	PHE	A	731	56.579	18.068	113.320	1.00	9.05	A	C
ATOM	5468	CA	GLY	A	732	52.846	17.319	113.474	1.00	8.76	A	C
ATOM	5472	CA	ASN	A	733	51.196	19.402	110.742	1.00	8.99	A	C
ATOM	5480	CA	THR	A	734	49.869	17.843	107.528	1.00	9.35	A	C
ATOM	5487	CA	TRP	A	735	50.725	20.969	105.477	1.00	8.91	A	C
ATOM	5501	CA	VAL	A	736	53.901	22.986	104.955	1.00	8.70	A	C
ATOM	5508	CA	ASP	A	737	54.483	26.555	103.781	1.00	8.20	A	C
ATOM	5516	CA	TYR	A	738	56.186	27.338	100.476	1.00	6.61	A	C
ATOM	5528	CA	ALA	A	739	58.753	29.427	102.391	1.00	6.41	A	C
ATOM	5533	CA	THR	A	740	59.787	26.356	104.396	1.00	7.06	A	C
ATOM	5540	CA	VAL	A	741	60.397	24.354	101.242	1.00	6.62	A	C
ATOM	5547	CA	VAL	A	742	62.422	26.857	99.172	1.00	7.73	A	C
ATOM	5554	CA	GLY	A	743	64.172	27.894	102.414	1.00	9.90	A	C
ATOM	5558	CA	GLY	A	744	65.997	24.543	102.207	1.00	9.66	A	C
ATOM	5562	CA	ALA	A	745	64.348	22.686	105.078	1.00	9.46	A	C
ATOM	5567	CA	ASP	A	746	64.950	19.052	106.107	1.00	9.46	A	C
ATOM	5575	CA	LEU	A	747	61.574	17.315	106.252	1.00	7.98	A	C
ATOM	5583	CA	ALA	A	748	62.092	13.775	107.535	1.00	8.59	A	C
ATOM	5588	CA	PHE	A	749	59.312	11.203	107.659	1.00	9.99	A	C
ATOM	5599	CA	ARG	A	750	59.298	7.807	109.398	1.00	11.09	A	C
ATOM	5610	CA	MET	A	751	57.231	5.392	107.265	1.00	8.36	A	C
ATOM	5618	CA	GLY	A	752	55.426	2.175	108.214	1.00	8.95	A	C
ATOM	5622	CA	GLU	A	753	53.025	-0.439	106.937	1.00	10.36	A	C
ATOM	5631	CA	GLN	A	754	50.077	0.533	109.170	1.00	10.20	A	C
ATOM	5640	CA	PRO	A	755	48.009	3.743	109.500	1.00	9.59	A	C
ATOM	5647	CA	SER	A	756	49.021	5.965	112.430	1.00	10.96	A	C
ATOM	5653	CA	ASP	A	757	47.455	8.970	114.180	1.00	12.99	A	C
ATOM	5661	CA	TRP	A	758	50.369	11.238	113.073	1.00	11.54	A	C
ATOM	5675	CA	GLY	A	759	49.073	14.673	112.067	1.00	12.17	A	C
ATOM	5679	CA	THR	A	760	45.608	14.275	113.614	1.00	12.83	A	C
ATOM	5686	CA	ASP	A	761	46.374	17.270	115.851	1.00	13.13	A	C
ATOM	5694	CA	THR	A	762	47.486	19.473	112.939	1.00	10.45	A	C
ATOM	5701	CA	ALA	A	763	47.395	23.279	113.167	1.00	9.30	A	C
ATOM	5706	CA	PRO	A	764	44.545	24.274	110.760	1.00	8.86	A	C
ATOM	5713	CA	ALA	A	765	45.404	24.788	107.092	1.00	8.11	A	C
ATOM	5718	CA	PHE	A	766	44.495	27.881	105.084	1.00	9.09	A	C
ATOM	5729	CA	SER	A	767	40.845	28.777	104.561	1.00	9.56	A	C
ATOM	5735	CA	MET	A	768	39.792	32.266	103.433	1.00	11.12	A	C
ATOM	5743	CA	SER	A	769	37.000	32.501	106.061	1.00	14.28	A	C
ATOM	5749	CA	THR	A	770	39.217	31.636	109.046	1.00	17.50	A	C
ATOM	5756	CA	ALA	A	771	42.527	33.224	107.955	1.00	19.68	A	C
ATOM	5761	CA	ASP	B	8	33.160	32.500	141.172	1.00	18.72	B	C
ATOM	5769	CA	TYR	B	9	29.570	31.506	140.304	1.00	14.20	B	C
ATOM	5781	CA	ALA	B	10	28.422	31.895	143.925	1.00	10.68	B	C
ATOM	5786	CA	SER	B	11	29.320	35.614	143.693	1.00	10.07	B	C
ATOM	5792	CA	LEU	B	12	26.696	36.062	140.938	1.00	8.36	B	C
ATOM	5800	CA	VAL	B	13	23.923	34.801	143.242	1.00	6.94	B	C
ATOM	5807	CA	ASP	B	14	21.880	37.475	145.012	1.00	6.81	B	C
ATOM	5815	CA	VAL	B	15	20.179	35.726	147.955	1.00	4.17	B	C
ATOM	5822	CA	PHE	B	16	18.230	38.889	148.802	1.00	3.40	B	C
ATOM	5833	CA	VAL	B	17	16.289	38.686	145.547	1.00	3.53	B	C
ATOM	5840	CA	GLY	B	18	12.666	38.026	146.520	1.00	4.03	B	C
ATOM	5844	CA	THR	B	19	13.196	38.689	150.262	1.00	5.26	B	C
ATOM	5851	CA	GLU	B	20	10.979	41.797	150.516	1.00	5.53	B	C
ATOM	5860	CA	GLY	B	21	7.349	41.901	151.577	1.00	6.87	B	C
ATOM	5864	CA	ASP	B	22	5.019	39.846	149.439	1.00	6.61	B	C
ATOM	5872	CA	PHE	B	23	7.350	39.730	146.400	1.00	5.51	B	C
ATOM	5883	CA	GLY	B	24	8.458	36.090	146.362	1.00	4.07	B	C
ATOM	5887	CA	ASN	B	25	8.571	34.746	149.915	1.00	2.81	B	C
ATOM	5895	CA	ASP	B	26	12.206	33.663	149.535	1.00	3.13	B	C
ATOM	5903	CA	MET	B	27	14.936	33.714	152.197	1.00	4.16	B	C
ATOM	5911	CA	PRO	B	28	18.521	35.051	152.473	1.00	3.52	B	C
ATOM	5918	CA	ALA	B	29	19.101	32.077	154.801	1.00	2.82	B	C
ATOM	5923	CA	ALA	B	30	22.436	30.302	154.790	1.00	2.98	B	C

ATOM	5928	CA	GLN	B	31	21.896	26.852	153.161	1.00	4.61	B	C
ATOM	5937	CA	ALA	B	32	23.671	24.106	151.121	1.00	5.14	B	C
ATOM	5942	CA	PRO	B	33	22.280	22.821	147.805	1.00	6.54	B	C
ATOM	5949	CA	ASN	B	34	18.900	21.202	148.694	1.00	5.61	B	C
ATOM	5957	CA	GLY	B	35	19.876	21.616	152.349	1.00	4.62	B	C
ATOM	5961	CA	LEU	B	36	17.780	20.633	155.336	1.00	5.14	B	C
ATOM	5969	CA	ALA	B	37	19.361	23.299	157.533	1.00	3.87	B	C
ATOM	5974	CA	LYS	B	38	18.219	26.746	156.507	1.00	3.49	B	C
ATOM	5983	CA	VAL	B	39	19.767	29.259	158.889	1.00	3.32	B	C
ATOM	5990	CA	ASN	B	40	17.444	32.228	158.346	1.00	3.22	B	C
ATOM	5998	CA	PRO	B	41	17.676	35.552	160.107	1.00	3.73	B	C
ATOM	6005	CA	ARG	B	42	14.314	36.534	161.600	1.00	3.05	B	C
ATOM	6016	CA	THR	B	43	13.161	40.161	161.230	1.00	2.90	B	C
ATOM	6023	CA	THR	B	44	10.738	41.938	163.588	1.00	5.21	B	C
ATOM	6030	CA	PRO	B	45	8.030	43.161	163.977	1.00	6.45	B	C
ATOM	6037	CA	GLY	B	46	7.346	42.212	160.323	1.00	4.86	B	C
ATOM	6041	CA	ARG	B	47	8.581	39.247	158.248	1.00	4.58	B	C
ATOM	6052	CA	ASN	B	48	7.898	37.485	154.983	1.00	4.09	B	C
ATOM	6060	CA	ASN	B	49	6.685	33.858	155.217	1.00	4.78	B	C
ATOM	6068	CA	THR	B	50	10.246	32.518	155.744	1.00	4.39	B	C
ATOM	6075	CA	GLY	B	51	10.802	34.910	158.707	1.00	4.48	B	C
ATOM	6079	CA	TYR	B	52	12.692	37.754	157.012	1.00	5.15	B	C
ATOM	6091	CA	ASP	B	53	11.399	40.842	155.301	1.00	5.15	B	C
ATOM	6099	CA	TYR	B	54	14.064	43.028	153.656	1.00	5.43	B	C
ATOM	6111	CA	ALA	B	55	11.984	46.169	154.374	1.00	5.97	B	C
ATOM	6116	CA	GLN	B	56	12.446	45.769	158.155	1.00	6.73	B	C
ATOM	6125	CA	SER	B	57	15.268	47.086	160.349	1.00	7.18	B	C
ATOM	6131	CA	LYS	B	58	15.306	44.770	163.351	1.00	4.81	B	C
ATOM	6140	CA	ILE	B	59	16.408	41.138	163.680	1.00	4.41	B	C
ATOM	6148	CA	SER	B	60	15.847	38.827	166.673	1.00	4.63	B	C
ATOM	6154	CA	GLY	B	61	17.915	35.758	165.791	1.00	3.36	B	C
ATOM	6158	CA	PHE	B	62	18.275	32.810	163.426	1.00	3.48	B	C
ATOM	6169	CA	THR	B	63	15.870	29.935	162.748	1.00	3.46	B	C
ATOM	6176	CA	HIS	B	64	17.063	26.576	161.377	1.00	3.52	B	C
ATOM	6186	CA	THR	B	65	14.187	25.010	159.402	1.00	4.13	B	C
ATOM	6193	CA	ASN	B	66	12.141	26.467	156.576	1.00	4.29	B	C
ATOM	6201	CA	LEU	B	67	10.335	26.228	153.272	1.00	4.24	B	C
ATOM	6209	CA	ASP	B	68	11.312	28.351	150.252	1.00	4.81	B	C
ATOM	6217	CA	GLY	B	69	8.769	30.598	148.558	1.00	4.56	B	C
ATOM	6221	CA	VAL	B	70	5.589	29.149	150.102	1.00	3.51	B	C
ATOM	6228	CA	GLY	B	71	2.243	30.766	151.033	1.00	4.39	B	C
ATOM	6232	CA	GLY	B	72	0.165	30.889	154.256	1.00	5.93	B	C
ATOM	6236	CA	SER	B	73	2.360	32.159	157.086	1.00	5.71	B	C
ATOM	6242	CA	GLY	B	74	5.232	29.886	155.962	1.00	4.98	B	C
ATOM	6246	CA	GLY	B	75	6.527	26.565	157.216	1.00	4.40	B	C
ATOM	6250	CA	GLY	B	76	9.371	25.580	159.508	1.00	4.57	B	C
ATOM	6254	CA	GLY	B	77	10.953	28.443	161.386	1.00	3.39	B	C
ATOM	6258	CA	ASP	B	78	11.986	26.048	164.188	1.00	3.33	B	C
ATOM	6266	CA	LEU	B	79	14.888	26.547	166.612	1.00	5.32	B	C
ATOM	6274	CA	LEU	B	80	15.348	30.284	167.110	1.00	5.08	B	C
ATOM	6282	CA	VAL	B	81	18.851	31.213	168.309	1.00	4.86	B	C
ATOM	6289	CA	VAL	B	82	19.084	34.745	169.745	1.00	5.18	B	C
ATOM	6296	CA	PRO	B	83	22.375	36.414	170.798	1.00	5.18	B	C
ATOM	6303	CA	THR	B	84	22.055	38.823	173.750	1.00	4.92	B	C
ATOM	6310	CA	SER	B	85	24.059	40.445	176.538	1.00	3.51	B	C
ATOM	6316	CA	GLY	B	86	20.865	40.604	178.572	1.00	3.93	B	C
ATOM	6320	CA	SER	B	87	19.752	38.579	181.582	1.00	5.35	B	C
ATOM	6326	CA	TYR	B	88	16.498	36.652	182.221	1.00	5.05	B	C
ATOM	6338	CA	THR	B	89	14.650	35.335	185.336	1.00	5.80	B	C
ATOM	6345	CA	ALA	B	90	11.608	33.987	183.442	1.00	4.94	B	C
ATOM	6350	CA	ARG	B	91	10.687	32.820	179.933	1.00	5.95	B	C
ATOM	6361	CA	PRO	B	92	11.913	35.587	177.603	1.00	5.93	B	C
ATOM	6368	CA	GLY	B	93	9.414	38.047	176.166	1.00	5.97	B	C
ATOM	6372	CA	THR	B	94	9.767	37.813	172.371	1.00	6.48	B	C
ATOM	6379	CA	GLY	B	95	10.124	41.588	172.055	1.00	4.67	B	C
ATOM	6383	CA	THR	B	96	13.359	41.564	174.059	1.00	4.62	B	C

ATOM	6390	CA	TYR	B	97	15.098	39.627	171.228	1.00	4.32	B	C
ATOM	6402	CA	ALA	B	98	15.015	42.620	168.846	1.00	4.26	B	C
ATOM	6407	CA	HIS	B	99	18.311	44.130	167.648	1.00	4.45	B	C
ATOM	6417	CA	PRO	B	100	18.684	47.027	165.199	1.00	4.56	B	C
ATOM	6424	CA	PHE	B	101	20.304	46.145	161.878	1.00	5.62	B	C
ATOM	6435	CA	SER	B	102	21.236	47.834	158.588	1.00	6.73	B	C
ATOM	6441	CA	HIS	B	103	21.603	46.324	155.111	1.00	7.29	B	C
ATOM	6451	CA	ASP	B	104	24.879	48.336	155.037	1.00	9.91	B	C
ATOM	6459	CA	ASP	B	105	26.290	45.946	157.703	1.00	10.20	B	C
ATOM	6467	CA	GLU	B	106	24.917	42.703	156.229	1.00	9.23	B	C
ATOM	6476	CA	ASP	B	107	26.499	40.286	153.765	1.00	9.46	B	C
ATOM	6484	CA	ALA	B	108	25.242	36.970	152.370	1.00	6.84	B	C
ATOM	6489	CA	GLY	B	109	25.559	34.488	149.524	1.00	4.80	B	C
ATOM	6493	CA	PRO	B	110	25.491	30.714	148.861	1.00	4.09	B	C
ATOM	6500	CA	GLY	B	111	26.276	29.034	152.198	1.00	4.03	B	C
ATOM	6504	CA	PHE	B	112	26.451	32.010	154.507	1.00	4.63	B	C
ATOM	6515	CA	TYR	B	113	24.841	35.051	156.062	1.00	4.43	B	C
ATOM	6527	CA	SER	B	114	26.381	37.783	158.200	1.00	5.09	B	C
ATOM	6533	CA	VAL	B	115	24.962	40.865	159.954	1.00	4.89	B	C
ATOM	6540	CA	GLY	B	116	25.874	43.467	162.573	1.00	6.35	B	C
ATOM	6544	CA	LEU	B	117	23.215	43.427	165.294	1.00	6.43	B	C
ATOM	6552	CA	GLY	B	118	22.939	46.299	167.802	1.00	6.20	B	C
ATOM	6556	CA	ASN	B	119	23.960	44.928	171.238	1.00	5.86	B	C
ATOM	6564	CA	VAL	B	120	21.162	44.472	173.803	1.00	4.83	B	C
ATOM	6571	CA	ALA	B	121	21.260	44.132	177.613	1.00	4.23	B	C
ATOM	6576	CA	GLY	B	122	19.365	44.559	180.892	1.00	4.26	B	C
ATOM	6580	CA	THR	B	123	16.919	42.174	182.582	1.00	5.43	B	C
ATOM	6587	CA	ASP	B	124	13.711	40.546	181.298	1.00	5.91	B	C
ATOM	6595	CA	GLY	B	125	11.132	43.006	179.823	1.00	6.02	B	C
ATOM	6599	CA	ALA	B	126	13.463	45.973	180.415	1.00	5.90	B	C
ATOM	6604	CA	ILE	B	127	16.092	44.620	177.976	1.00	5.52	B	C
ATOM	6612	CA	THR	B	128	17.063	47.192	175.369	1.00	4.75	B	C
ATOM	6619	CA	GLY	B	129	19.991	48.703	173.407	1.00	4.79	B	C
ATOM	6623	CA	ALA	B	130	23.437	48.378	175.037	1.00	5.28	B	C
ATOM	6628	CA	PRO	B	131	26.932	49.536	173.894	1.00	5.84	B	C
ATOM	6635	CA	GLY	B	132	28.389	48.225	170.633	1.00	5.70	B	C
ATOM	6639	CA	THR	B	133	27.548	45.592	168.062	1.00	6.06	B	C
ATOM	6646	CA	ILE	B	134	27.013	41.848	168.174	1.00	7.11	B	C
ATOM	6654	CA	GLU	B	135	28.833	40.654	165.045	1.00	8.84	B	C
ATOM	6663	CA	ALA	B	136	26.820	37.651	163.802	1.00	6.10	B	C
ATOM	6668	CA	GLU	B	137	27.966	35.088	161.226	1.00	4.63	B	C
ATOM	6677	CA	VAL	B	138	26.200	31.897	160.196	1.00	3.25	B	C
ATOM	6684	CA	ALA	B	139	26.905	29.212	157.622	1.00	3.25	B	C
ATOM	6689	CA	ALA	B	140	25.244	25.933	156.657	1.00	4.37	B	C
ATOM	6694	CA	ALA	B	141	26.177	22.444	155.558	1.00	5.98	B	C
ATOM	6699	CA	THR	B	142	23.483	19.964	154.381	1.00	4.34	B	C
ATOM	6706	CA	ARG	B	143	22.140	19.123	157.865	1.00	4.38	B	C
ATOM	6717	CA	SER	B	144	23.954	21.728	159.957	1.00	4.49	B	C
ATOM	6723	CA	GLY	B	145	23.839	25.419	160.895	1.00	5.05	B	C
ATOM	6727	CA	VAL	B	146	26.994	26.868	162.414	1.00	4.62	B	C
ATOM	6734	CA	HIS	B	147	27.242	30.186	164.272	1.00	4.38	B	C
ATOM	6744	CA	ARG	B	148	30.103	32.540	165.208	1.00	4.25	B	C
ATOM	6755	CA	TYR	B	149	29.335	35.704	167.264	1.00	4.46	B	C
ATOM	6767	CA	ALA	B	150	31.522	38.561	168.553	1.00	5.60	B	C
ATOM	6772	CA	PHE	B	151	30.094	40.472	171.553	1.00	6.42	B	C
ATOM	6783	CA	PRO	B	152	31.380	43.820	172.738	1.00	8.35	B	C
ATOM	6790	CA	ALA	B	153	34.133	43.586	175.369	1.00	8.99	B	C
ATOM	6795	CA	GLY	B	154	32.667	43.385	178.865	1.00	8.42	B	C
ATOM	6799	CA	SER	B	155	29.390	41.759	177.743	1.00	8.54	B	C
ATOM	6805	CA	THR	B	156	28.171	38.643	179.484	1.00	7.06	B	C
ATOM	6812	CA	PRO	B	157	27.319	36.804	176.247	1.00	7.08	B	C
ATOM	6819	CA	SER	B	158	24.232	34.650	176.044	1.00	6.71	B	C
ATOM	6825	CA	LEU	B	159	22.370	32.582	173.502	1.00	4.87	B	C
ATOM	6833	CA	VAL	B	160	18.646	31.941	173.847	1.00	3.09	B	C
ATOM	6840	CA	VAL	B	161	17.419	28.777	172.147	1.00	3.34	B	C
ATOM	6847	CA	ASP	B	162	13.682	29.337	171.720	1.00	4.07	B	C

ATOM	6855	CA	LEU	B	163	11.876	26.065	170.870	1.00	3.79	B	C
ATOM	6863	CA	GLU	B	164	8.402	27.636	170.688	1.00	4.18	B	C
ATOM	6872	CA	THR	B	165	9.046	29.648	167.474	1.00	2.64	B	C
ATOM	6879	CA	ASN	B	166	7.417	28.351	164.312	1.00	2.42	B	C
ATOM	6887	CA	ASN	B	167	6.159	30.090	161.193	1.00	2.44	B	C
ATOM	6895	CA	THR	B	168	2.700	28.469	161.520	1.00	2.77	B	C
ATOM	6902	CA	SER	B	169	2.191	26.088	164.470	1.00	3.44	B	C
ATOM	6908	CA	ARG	B	170	4.367	24.663	167.209	1.00	3.61	B	C
ATOM	6919	CA	ARG	B	171	2.777	21.329	168.092	1.00	4.09	B	C
ATOM	6930	CA	SER	B	172	5.367	20.136	170.618	1.00	3.92	B	C
ATOM	6936	CA	SER	B	173	9.026	20.436	171.605	1.00	4.52	B	C
ATOM	6942	CA	SER	B	174	11.644	19.095	174.010	1.00	5.26	B	C
ATOM	6948	CA	VAL	B	175	15.144	19.964	175.194	1.00	7.61	B	C
ATOM	6955	CA	GLN	B	176	17.707	17.942	177.116	1.00	9.87	B	C
ATOM	6964	CA	VAL	B	177	20.827	19.575	178.479	1.00	10.75	B	C
ATOM	6971	CA	GLU	B	178	24.266	17.972	178.853	1.00	11.75	B	C
ATOM	6980	CA	THR	B	179	27.408	19.621	180.271	1.00	10.76	B	C
ATOM	6987	CA	ARG	B	180	30.637	17.869	179.161	1.00	11.07	B	C
ATOM	6998	CA	ALA	B	181	34.052	17.533	180.867	1.00	12.22	B	C
ATOM	7003	CA	ASP	B	182	35.476	20.584	179.033	1.00	11.74	B	C
ATOM	7011	CA	GLY	B	183	32.508	22.707	180.228	1.00	9.15	B	C
ATOM	7015	CA	THR	B	184	30.755	22.961	176.837	1.00	5.80	B	C
ATOM	7022	CA	VAL	B	185	27.013	22.306	176.541	1.00	5.32	B	C
ATOM	7029	CA	GLU	B	186	24.991	20.053	174.245	1.00	6.37	B	C
ATOM	7038	CA	LEU	B	187	21.225	20.551	173.801	1.00	5.44	B	C
ATOM	7046	CA	SER	B	188	19.138	17.900	172.108	1.00	4.09	B	C
ATOM	7052	CA	GLY	B	189	15.468	17.073	171.486	1.00	3.12	B	C
ATOM	7056	CA	GLN	B	190	12.682	17.282	168.938	1.00	2.37	B	C
ATOM	7065	CA	VAL	B	191	10.502	19.973	167.357	1.00	2.26	B	C
ATOM	7072	CA	THR	B	192	7.128	19.112	165.863	1.00	2.53	B	C
ATOM	7079	CA	GLY	B	193	5.491	21.715	163.683	1.00	2.57	B	C
ATOM	7083	CA	TYR	B	194	2.613	21.951	161.250	1.00	3.30	B	C
ATOM	7095	CA	PHE	B	195	2.176	23.583	157.852	1.00	4.28	B	C
ATOM	7106	CA	TYR	B	196	-0.598	23.421	155.296	1.00	7.19	B	C
ATOM	7118	CA	ASN	B	197	-1.557	19.737	155.149	1.00	7.00	B	C
ATOM	7126	CA	ALA	B	198	0.738	17.907	157.606	1.00	5.17	B	C
ATOM	7131	CA	ALA	B	199	2.648	17.834	160.876	1.00	4.90	B	C
ATOM	7136	CA	TYR	B	200	6.409	17.069	160.789	1.00	4.22	B	C
ATOM	7148	CA	THR	B	201	9.101	16.185	163.362	1.00	5.31	B	C
ATOM	7155	CA	LEU	B	202	12.799	16.977	163.328	1.00	3.48	B	C
ATOM	7163	CA	TYR	B	203	15.415	15.973	165.896	1.00	3.48	B	C
ATOM	7175	CA	TYR	B	204	18.277	18.294	166.724	1.00	4.54	B	C
ATOM	7187	CA	THR	B	205	21.584	18.490	168.504	1.00	5.64	B	C
ATOM	7194	CA	ALA	B	206	23.308	21.765	169.341	1.00	5.72	B	C
ATOM	7199	CA	ARG	B	207	26.654	22.253	170.998	1.00	5.49	B	C
ATOM	7210	CA	THR	B	208	28.601	25.232	172.226	1.00	4.80	B	C
ATOM	7217	CA	LEU	B	209	32.264	25.598	171.305	1.00	4.71	B	C
ATOM	7225	CA	GLN	B	210	33.041	27.516	174.554	1.00	6.35	B	C
ATOM	7234	CA	PRO	B	211	32.110	26.663	178.148	1.00	7.67	B	C
ATOM	7241	CA	ALA	B	212	28.543	27.611	179.143	1.00	7.25	B	C
ATOM	7246	CA	THR	B	213	26.099	27.434	182.004	1.00	7.17	B	C
ATOM	7253	CA	VAL	B	214	22.470	26.720	181.121	1.00	6.69	B	C
ATOM	7260	CA	GLN	B	215	18.911	27.268	182.283	1.00	6.38	B	C
ATOM	7269	CA	THR	B	216	15.794	25.836	180.631	1.00	4.98	B	C
ATOM	7276	CA	TRP	B	217	12.111	26.702	180.466	1.00	3.64	B	C
ATOM	7290	CA	GLY	B	218	8.884	24.866	180.016	1.00	5.63	B	C
ATOM	7294	CA	ASP	B	219	5.244	25.649	179.622	1.00	8.12	B	C
ATOM	7302	CA	ASP	B	220	4.977	27.146	183.127	1.00	12.50	B	C
ATOM	7310	CA	ASP	B	221	7.168	30.093	181.953	1.00	12.91	B	C
ATOM	7318	CA	ARG	B	222	9.634	29.500	184.787	1.00	11.75	B	C
ATOM	7329	CA	LEU	B	223	13.374	29.633	184.047	1.00	9.26	B	C
ATOM	7337	CA	VAL	B	224	15.093	26.886	186.002	1.00	9.70	B	C
ATOM	7344	CA	ASP	B	225	18.254	24.870	186.542	1.00	9.71	B	C
ATOM	7352	CA	ALA	B	226	16.446	21.645	185.479	1.00	8.94	B	C
ATOM	7357	CA	THR	B	227	18.172	19.922	182.585	1.00	8.94	B	C
ATOM	7364	CA	ALA	B	228	15.027	18.498	180.872	1.00	8.41	B	C

ATOM	7369	CA	GLN	B	229	11.887	19.991	179.343	1.00	7.11	B	C
ATOM	7378	CA	ASP	B	230	9.154	18.361	177.262	1.00	7.62	B	C
ATOM	7386	CA	GLY	B	231	5.982	20.180	176.272	1.00	5.49	B	C
ATOM	7390	CA	VAL	B	232	5.019	22.814	173.687	1.00	3.91	B	C
ATOM	7397	CA	ASP	B	233	6.717	26.124	174.666	1.00	3.59	B	C
ATOM	7405	CA	THR	B	234	10.207	25.219	175.870	1.00	3.22	B	C
ATOM	7412	CA	GLY	B	235	13.760	26.456	175.485	1.00	2.52	B	C
ATOM	7416	CA	ALA	B	236	17.165	27.054	176.932	1.00	2.01	B	C
ATOM	7421	CA	ILE	B	237	19.409	30.014	177.730	1.00	2.62	B	C
ATOM	7429	CA	LEU	B	238	23.188	29.544	177.393	1.00	3.80	B	C
ATOM	7437	CA	THR	B	239	25.439	31.941	179.352	1.00	5.81	B	C
ATOM	7444	CA	PHE	B	240	29.137	32.391	178.663	1.00	7.71	B	C
ATOM	7455	CA	ASP	B	241	32.053	34.026	180.461	1.00	10.49	B	C
ATOM	7463	CA	PRO	B	242	32.976	37.585	179.419	1.00	10.21	B	C
ATOM	7470	CA	ALA	B	243	36.482	36.270	178.652	1.00	10.56	B	C
ATOM	7475	CA	ASP	B	244	34.898	34.545	175.608	1.00	10.22	B	C
ATOM	7483	CA	ALA	B	245	32.993	37.584	174.265	1.00	9.19	B	C
ATOM	7488	CA	GLY	B	246	35.260	37.861	171.199	1.00	9.07	B	C
ATOM	7492	CA	GLU	B	247	34.117	34.471	169.883	1.00	8.40	B	C
ATOM	7501	CA	ILE	B	248	30.976	32.504	170.743	1.00	6.26	B	C
ATOM	7509	CA	GLY	B	249	30.419	29.317	168.700	1.00	5.26	B	C
ATOM	7513	CA	LEU	B	250	27.365	27.148	168.128	1.00	4.75	B	C
ATOM	7521	CA	GLN	B	251	26.729	24.080	165.940	1.00	4.54	B	C
ATOM	7530	CA	VAL	B	252	23.221	22.815	165.229	1.00	4.09	B	C
ATOM	7537	CA	THR	B	253	22.517	19.573	163.382	1.00	3.14	B	C
ATOM	7544	CA	LEU	B	254	19.085	18.325	162.254	1.00	3.66	B	C
ATOM	7552	CA	SER	B	255	17.710	14.818	161.568	1.00	3.72	B	C
ATOM	7558	CA	PRO	B	256	14.350	13.322	160.581	1.00	3.90	B	C
ATOM	7565	CA	VAL	B	257	15.675	10.122	162.247	1.00	4.49	B	C
ATOM	7572	CA	SER	B	258	16.553	10.685	165.919	1.00	5.09	B	C
ATOM	7578	CA	VAL	B	259	18.811	12.681	168.270	1.00	6.85	B	C
ATOM	7585	CA	GLU	B	260	21.309	9.841	168.225	1.00	8.56	B	C
ATOM	7594	CA	GLN	B	261	21.425	10.162	164.416	1.00	7.14	B	C
ATOM	7603	CA	ALA	B	262	21.764	13.965	164.573	1.00	6.00	B	C
ATOM	7608	CA	ARG	B	263	24.840	13.567	166.780	1.00	6.59	B	C
ATOM	7619	CA	ILE	B	264	26.315	11.008	164.363	1.00	7.36	B	C
ATOM	7627	CA	ASP	B	265	25.571	13.292	161.370	1.00	6.50	B	C
ATOM	7635	CA	GLN	B	266	27.201	16.188	163.205	1.00	7.37	B	C
ATOM	7644	CA	GLN	B	267	30.403	14.178	163.786	1.00	11.20	B	C
ATOM	7653	CA	VAL	B	268	30.481	13.075	160.124	1.00	10.88	B	C
ATOM	7660	CA	GLU	B	269	29.430	16.369	158.505	1.00	11.16	B	C
ATOM	7669	CA	LEU	B	270	31.314	18.882	160.704	1.00	10.96	B	C
ATOM	7677	CA	GLY	B	271	33.869	16.943	162.740	1.00	11.80	B	C
ATOM	7681	CA	ASP	B	272	36.639	19.186	164.002	1.00	11.87	B	C
ATOM	7689	CA	LEU	B	273	36.374	21.606	161.059	1.00	9.30	B	C
ATOM	7697	CA	SER	B	274	36.585	25.316	161.684	1.00	8.10	B	C
ATOM	7703	CA	PHE	B	275	33.692	27.729	161.001	1.00	7.57	B	C
ATOM	7714	CA	ASP	B	276	35.633	29.179	158.049	1.00	9.11	B	C
ATOM	7722	CA	ALA	B	277	36.288	25.679	156.588	1.00	8.17	B	C
ATOM	7727	CA	ILE	B	278	32.607	24.690	156.805	1.00	6.31	B	C
ATOM	7735	CA	ARG	B	279	31.510	27.935	155.180	1.00	7.50	B	C
ATOM	7746	CA	ASP	B	280	34.193	27.585	152.447	1.00	8.14	B	C
ATOM	7754	CA	ARG	B	281	33.248	23.952	151.718	1.00	7.53	B	C
ATOM	7765	CA	THR	B	282	29.590	24.914	151.207	1.00	7.03	B	C
ATOM	7772	CA	ARG	B	283	30.749	27.741	148.908	1.00	8.06	B	C
ATOM	7783	CA	ALA	B	284	32.685	25.192	146.809	1.00	7.18	B	C
ATOM	7788	CA	GLU	B	285	29.647	22.838	146.826	1.00	8.04	B	C
ATOM	7797	CA	TRP	B	286	27.513	25.684	145.409	1.00	6.44	B	C
ATOM	7811	CA	ASN	B	287	30.194	26.541	142.822	1.00	7.77	B	C
ATOM	7819	CA	ALA	B	288	30.163	22.844	141.765	1.00	8.01	B	C
ATOM	7824	CA	THR	B	289	26.340	22.924	141.481	1.00	7.57	B	C
ATOM	7831	CA	LEU	B	290	25.928	26.258	139.687	1.00	6.68	B	C
ATOM	7839	CA	GLY	B	291	28.902	25.412	137.486	1.00	8.30	B	C
ATOM	7843	CA	ARG	B	292	26.832	22.721	135.807	1.00	8.97	B	C
ATOM	7854	CA	VAL	B	293	25.594	25.636	133.725	1.00	10.54	B	C
ATOM	7861	CA	ALA	B	294	28.121	27.918	132.046	1.00	12.46	B	C

ATOM	7866	CA	ILE	B	295	26.911	31.025	130.202	1.00	15.01	B	C
ATOM	7874	CA	ASP	B	296	28.503	33.444	127.742	1.00	16.68	B	C
ATOM	7882	CA	ALA	B	297	26.492	36.686	127.506	1.00	16.02	B	C
ATOM	7887	CA	SER	B	298	27.400	39.653	125.271	1.00	15.25	B	C
ATOM	7893	CA	THR	B	299	26.635	43.276	126.121	1.00	14.42	B	C
ATOM	7900	CA	ALA	B	300	24.046	43.110	123.326	1.00	11.48	B	C
ATOM	7905	CA	THR	B	301	21.783	40.582	125.103	1.00	9.84	B	C
ATOM	7912	CA	ASP	B	302	22.704	41.528	128.706	1.00	9.77	B	C
ATOM	7920	CA	PRO	B	303	23.868	45.197	128.715	1.00	10.54	B	C
ATOM	7927	CA	THR	B	304	23.489	45.589	132.512	1.00	11.07	B	C
ATOM	7934	CA	GLY	B	305	24.761	42.154	133.524	1.00	11.11	B	C
ATOM	7938	CA	GLU	B	306	21.483	41.714	135.432	1.00	10.64	B	C
ATOM	7947	CA	LEU	B	307	20.268	38.736	133.293	1.00	10.30	B	C
ATOM	7955	CA	GLN	B	308	23.365	36.685	134.127	1.00	10.19	B	C
ATOM	7964	CA	ARG	B	309	22.914	37.468	137.835	1.00	8.46	B	C
ATOM	7975	CA	LEU	B	310	19.207	36.600	137.644	1.00	6.90	B	C
ATOM	7983	CA	PHE	B	311	20.109	33.341	135.894	1.00	5.42	B	C
ATOM	7994	CA	TYR	B	312	22.509	32.208	138.679	1.00	4.65	B	C
ATOM	8006	CA	THR	B	313	20.288	33.567	141.446	1.00	3.72	B	C
ATOM	8013	CA	HIS	B	314	17.411	31.465	140.078	1.00	3.77	B	C
ATOM	8023	CA	LEU	B	315	19.597	28.424	139.419	1.00	4.98	B	C
ATOM	8031	CA	TYR	B	316	20.467	28.753	143.144	1.00	4.55	B	C
ATOM	8043	CA	ARG	B	317	16.731	28.839	144.007	1.00	5.20	B	C
ATOM	8054	CA	MET	B	318	16.085	25.817	141.789	1.00	7.54	B	C
ATOM	8062	CA	PHE	B	319	18.175	23.558	144.063	1.00	8.02	B	C
ATOM	8073	CA	ALA	B	320	16.316	24.249	147.345	1.00	7.25	B	C
ATOM	8078	CA	MET	B	321	13.403	21.781	147.467	1.00	5.88	B	C
ATOM	8086	CA	PRO	B	322	12.863	18.952	147.903	1.00	4.25	B	C
ATOM	8093	CA	MET	B	323	15.380	18.850	150.760	1.00	3.69	B	C
ATOM	8101	CA	ASN	B	324	17.944	16.177	151.447	1.00	3.38	B	C
ATOM	8109	CA	ALA	B	325	16.107	14.123	154.077	1.00	3.72	B	C
ATOM	8114	CA	THR	B	326	18.714	11.357	154.443	1.00	3.39	B	C
ATOM	8121	CA	SER	B	327	21.071	10.805	157.383	1.00	4.75	B	C
ATOM	8127	CA	THR	B	328	24.861	10.478	156.953	1.00	5.56	B	C
ATOM	8134	CA	SER	B	329	24.175	6.783	157.673	1.00	5.03	B	C
ATOM	8140	CA	GLY	B	330	21.990	6.586	154.509	1.00	5.56	B	C
ATOM	8144	CA	THR	B	331	18.681	6.308	156.385	1.00	4.67	B	C
ATOM	8151	CA	TYR	B	332	15.380	8.194	156.221	1.00	4.08	B	C
ATOM	8163	CA	ARG	B	333	12.110	8.234	158.156	1.00	5.90	B	C
ATOM	8174	CA	GLY	B	334	8.940	7.045	156.393	1.00	7.33	B	C
ATOM	8178	CA	VAL	B	335	5.362	8.276	156.776	1.00	9.86	B	C
ATOM	8185	CA	ASP	B	336	4.715	4.745	158.108	1.00	11.83	B	C
ATOM	8193	CA	GLY	B	337	6.472	5.889	161.292	1.00	12.67	B	C
ATOM	8197	CA	ALA	B	338	9.546	3.712	160.784	1.00	10.29	B	C
ATOM	8202	CA	VAL	B	339	13.181	4.400	159.827	1.00	8.36	B	C
ATOM	8209	CA	HIS	B	340	14.277	2.783	156.523	1.00	8.22	B	C
ATOM	8219	CA	ALA	B	341	17.345	2.468	154.295	1.00	9.30	B	C
ATOM	8224	CA	ALA	B	342	17.738	4.699	151.232	1.00	10.96	B	C
ATOM	8229	CA	GLN	B	343	19.736	1.973	149.549	1.00	14.92	B	C
ATOM	8238	CA	GLY	B	344	22.061	3.243	146.798	1.00	14.18	B	C
ATOM	8242	CA	PHE	B	345	20.330	6.658	146.470	1.00	9.94	B	C
ATOM	8253	CA	THR	B	346	19.663	9.851	148.454	1.00	5.70	B	C
ATOM	8260	CA	TYR	B	347	16.132	10.341	149.769	1.00	3.08	B	C
ATOM	8272	CA	TYR	B	348	14.568	13.786	149.336	1.00	2.49	B	C
ATOM	8284	CA	ASP	B	349	11.394	15.091	150.938	1.00	2.40	B	C
ATOM	8292	CA	SER	B	350	9.337	18.340	151.066	1.00	3.13	B	C
ATOM	8298	CA	TRP	B	351	6.904	18.658	148.223	1.00	3.44	B	C
ATOM	8312	CA	ALA	B	352	4.594	21.317	146.769	1.00	6.09	B	C
ATOM	8317	CA	THR	B	353	3.818	19.352	143.690	1.00	7.57	B	C
ATOM	8324	CA	TRP	B	354	0.511	21.060	142.861	1.00	10.28	B	C
ATOM	8338	CA	ASP	B	355	2.607	24.171	142.126	1.00	9.25	B	C
ATOM	8346	CA	ASP	B	356	6.027	22.821	141.162	1.00	7.97	B	C
ATOM	8354	CA	PHE	B	357	5.476	19.675	139.069	1.00	5.45	B	C
ATOM	8365	CA	ARG	B	358	7.086	21.219	135.922	1.00	4.54	B	C
ATOM	8376	CA	LYS	B	359	10.204	22.174	137.907	1.00	3.86	B	C
ATOM	8385	CA	PHE	B	360	11.251	18.513	137.948	1.00	3.85	B	C

ATOM	8396	CA	SER	B	361	11.230	18.254	134.132	1.00	4.19	B	C
ATOM	8402	CA	VAL	B	362	13.848	21.051	134.057	1.00	4.66	B	C
ATOM	8409	CA	ILE	B	363	15.981	19.656	136.859	1.00	3.60	B	C
ATOM	8417	CA	ALA	B	364	15.986	16.487	134.724	1.00	4.12	B	C
ATOM	8422	CA	TYR	B	365	18.152	18.190	132.040	1.00	5.46	B	C
ATOM	8434	CA	ILE	B	366	20.412	20.182	134.397	1.00	6.51	B	C
ATOM	8442	CA	ASP	B	367	21.232	17.592	137.055	1.00	6.76	B	C
ATOM	8450	CA	PRO	B	368	19.936	14.178	135.858	1.00	6.15	B	C
ATOM	8457	CA	ALA	B	369	21.272	12.238	138.895	1.00	5.16	B	C
ATOM	8462	CA	LEU	B	370	19.595	14.581	141.391	1.00	5.66	B	C
ATOM	8470	CA	TYR	B	371	16.368	14.351	139.411	1.00	5.00	B	C
ATOM	8482	CA	ARG	B	372	16.537	10.538	139.507	1.00	6.13	B	C
ATOM	8493	CA	ASP	B	373	16.991	10.482	143.321	1.00	4.65	B	C
ATOM	8501	CA	MET	B	374	14.069	12.884	143.690	1.00	3.74	B	C
ATOM	8509	CA	VAL	B	375	11.706	10.681	141.648	1.00	3.94	B	C
ATOM	8516	CA	GLN	B	376	12.853	7.548	143.481	1.00	3.66	B	C
ATOM	8525	CA	SER	B	377	12.139	9.421	146.732	1.00	3.93	B	C
ATOM	8531	CA	LEU	B	378	8.564	10.288	145.559	1.00	4.51	B	C
ATOM	8539	CA	VAL	B	379	8.064	6.634	144.704	1.00	3.56	B	C
ATOM	8546	CA	TYR	B	380	9.204	5.663	148.259	1.00	4.32	B	C
ATOM	8558	CA	LEU	B	381	6.959	8.247	149.897	1.00	3.49	B	C
ATOM	8566	CA	PHE	B	382	3.838	6.774	148.265	1.00	3.82	B	C
ATOM	8577	CA	ALA	B	383	5.141	3.221	148.777	1.00	4.94	B	C
ATOM	8582	CA	ASP	B	384	5.471	3.995	152.503	1.00	6.79	B	C
ATOM	8590	CA	ALA	B	385	1.923	5.413	152.611	1.00	9.08	B	C
ATOM	8595	CA	GLU	B	386	0.765	2.141	150.976	1.00	10.89	B	C
ATOM	8604	CA	ALA	B	387	2.810	0.119	153.498	1.00	13.19	B	C
ATOM	8609	CA	THR	B	388	0.740	1.552	156.445	1.00	15.38	B	C
ATOM	8616	CA	GLY	B	389	-2.271	-0.495	155.224	1.00	15.84	B	C
ATOM	8620	CA	THR	B	390	-4.587	2.321	156.337	1.00	14.91	B	C
ATOM	8627	CA	GLY	B	391	-5.415	3.542	152.829	1.00	12.96	B	C
ATOM	8631	CA	GLY	B	392	-5.069	7.090	154.245	1.00	10.55	B	C
ATOM	8635	CA	GLY	B	393	-3.986	10.168	152.254	1.00	8.29	B	C
ATOM	8639	CA	LEU	B	394	-0.516	11.679	152.818	1.00	6.39	B	C
ATOM	8647	CA	GLY	B	395	-1.926	14.514	154.938	1.00	6.61	B	C
ATOM	8651	CA	GLY	B	396	-2.978	12.076	157.705	1.00	7.77	B	C
ATOM	8655	CA	PHE	B	397	0.603	11.030	158.585	1.00	7.05	B	C
ATOM	8666	CA	VAL	B	398	3.457	12.825	160.317	1.00	6.28	B	C
ATOM	8673	CA	HIS	B	399	6.082	13.832	157.779	1.00	4.37	B	C
ATOM	8683	CA	SER	B	400	9.868	13.714	158.080	1.00	4.62	B	C
ATOM	8689	CA	VAL	B	401	10.519	17.328	157.074	1.00	3.81	B	C
ATOM	8696	CA	PRO	B	402	8.279	20.398	156.732	1.00	2.81	B	C
ATOM	8703	CA	THR	B	403	6.223	19.841	153.605	1.00	2.00	B	C
ATOM	8710	CA	VAL	B	404	3.295	21.247	151.582	1.00	2.02	B	C
ATOM	8717	CA	ARG	B	405	1.236	19.105	149.134	1.00	3.66	B	C
ATOM	8728	CA	TRP	B	406	1.343	16.193	146.726	1.00	4.40	B	C
ATOM	8742	CA	GLU	B	407	-1.154	16.632	143.859	1.00	4.95	B	C
ATOM	8751	CA	ARG	B	408	0.354	16.056	140.358	1.00	4.63	B	C
ATOM	8762	CA	SER	B	409	2.805	13.445	141.798	1.00	4.41	B	C
ATOM	8768	CA	SER	B	410	1.577	10.955	139.158	1.00	5.65	B	C
ATOM	8774	CA	VAL	B	411	2.871	13.412	136.534	1.00	4.55	B	C
ATOM	8781	CA	VAL	B	412	6.352	13.733	138.090	1.00	3.62	B	C
ATOM	8788	CA	VAL	B	413	6.857	9.945	138.202	1.00	3.87	B	C
ATOM	8795	CA	ALA	B	414	5.438	9.658	134.653	1.00	5.13	B	C
ATOM	8800	CA	ASP	B	415	7.928	12.368	133.664	1.00	4.82	B	C
ATOM	8808	CA	ALA	B	416	10.908	10.154	134.619	1.00	4.44	B	C
ATOM	8813	CA	ILE	B	417	9.427	7.020	132.987	1.00	5.16	B	C
ATOM	8821	CA	ALA	B	418	8.712	8.909	129.732	1.00	6.23	B	C
ATOM	8826	CA	LYS	B	419	12.303	10.194	129.851	1.00	8.29	B	C
ATOM	8835	CA	GLY	B	420	13.794	6.686	129.922	1.00	10.82	B	C
ATOM	8839	CA	PHE	B	421	14.569	6.285	133.624	1.00	11.95	B	C
ATOM	8850	CA	ASP	B	422	13.949	2.885	135.136	1.00	14.48	B	C
ATOM	8858	CA	GLY	B	423	14.815	0.654	138.084	1.00	14.15	B	C
ATOM	8862	CA	PHE	B	424	12.489	2.574	140.366	1.00	12.52	B	C
ATOM	8873	CA	ASP	B	425	12.381	0.289	143.387	1.00	12.13	B	C
ATOM	8881	CA	ARG	B	426	8.874	-0.058	144.898	1.00	9.62	B	C

ATOM	8892	CA	LEU	B	427	6.950	1.477	141.964	1.00	9.74	B	C
ATOM	8900	CA	ASP	B	428	4.503	-1.436	142.300	1.00	11.74	B	C
ATOM	8908	CA	GLU	B	429	3.837	-0.485	145.942	1.00	10.85	B	C
ATOM	8917	CA	ALA	B	430	3.557	3.228	145.139	1.00	9.56	B	C
ATOM	8922	CA	TYR	B	431	0.970	2.649	142.393	1.00	9.69	B	C
ATOM	8934	CA	PRO	B	432	-2.178	2.019	144.553	1.00	8.88	B	C
ATOM	8941	CA	ALA	B	433	-1.368	5.086	146.736	1.00	7.60	B	C
ATOM	8946	CA	LEU	B	434	-0.914	7.129	143.566	1.00	7.98	B	C
ATOM	8954	CA	GLN	B	435	-4.332	5.893	142.378	1.00	9.79	B	C
ATOM	8963	CA	ARG	B	436	-5.876	6.985	145.714	1.00	8.63	B	C
ATOM	8974	CA	LEU	B	437	-4.175	10.398	145.345	1.00	8.82	B	C
ATOM	8982	CA	VAL	B	438	-5.651	10.832	141.863	1.00	10.72	B	C
ATOM	8989	CA	GLY	B	439	-9.074	9.520	142.973	1.00	11.53	B	C
ATOM	8993	CA	GLN	B	440	-11.952	7.945	141.034	1.00	13.71	B	C
ATOM	9002	CA	TYR	B	441	-14.865	9.454	139.124	1.00	12.01	B	C
ATOM	9014	CA	SER	B	442	-18.189	8.549	140.760	1.00	12.74	B	C
ATOM	9020	CA	ALA	B	443	-20.666	6.339	138.839	1.00	12.68	B	C
ATOM	9025	CA	ASP	B	444	-22.553	9.434	137.687	1.00	12.69	B	C
ATOM	9033	CA	GLU	B	445	-19.307	11.199	136.788	1.00	10.37	B	C
ATOM	9042	CA	LEU	B	446	-18.282	8.164	134.766	1.00	10.75	B	C
ATOM	9050	CA	ARG	B	447	-21.517	8.211	132.730	1.00	12.18	B	C
ATOM	9061	CA	ARG	B	448	-21.331	11.954	131.904	1.00	11.28	B	C
ATOM	9072	CA	GLY	B	449	-17.521	12.199	131.769	1.00	9.56	B	C
ATOM	9076	CA	TYR	B	450	-16.840	15.171	134.104	1.00	8.69	B	C
ATOM	9088	CA	VAL	B	451	-17.362	16.664	137.586	1.00	8.42	B	C
ATOM	9095	CA	ALA	B	452	-20.609	18.655	137.398	1.00	9.63	B	C
ATOM	9100	CA	GLY	B	453	-19.943	22.350	137.910	1.00	9.76	B	C
ATOM	9104	CA	ASN	B	454	-16.242	21.775	138.624	1.00	8.50	B	C
ATOM	9112	CA	PRO	B	455	-14.156	22.194	135.439	1.00	8.32	B	C
ATOM	9119	CA	GLY	B	456	-11.107	22.756	137.692	1.00	8.02	B	C
ATOM	9123	CA	ALA	B	457	-11.159	19.292	139.280	1.00	6.90	B	C
ATOM	9128	CA	SER	B	458	-11.963	17.688	135.889	1.00	6.05	B	C
ATOM	9134	CA	VAL	B	459	-8.939	18.989	133.896	1.00	5.70	B	C
ATOM	9141	CA	GLN	B	460	-6.681	18.365	136.882	1.00	5.79	B	C
ATOM	9150	CA	ARG	B	461	-7.804	14.732	137.121	1.00	6.42	B	C
ATOM	9161	CA	GLY	B	462	-7.221	14.406	133.370	1.00	5.35	B	C
ATOM	9165	CA	TYR	B	463	-3.539	15.278	133.771	1.00	5.04	B	C
ATOM	9177	CA	ASP	B	464	-3.270	13.035	136.851	1.00	5.57	B	C
ATOM	9185	CA	GLN	B	465	-4.804	10.198	134.841	1.00	5.78	B	C
ATOM	9194	CA	TYR	B	466	-2.483	10.769	131.919	1.00	6.49	B	C
ATOM	9206	CA	GLY	B	467	0.439	10.511	134.378	1.00	5.94	B	C
ATOM	9210	CA	LEU	B	468	-1.027	7.423	136.006	1.00	7.08	B	C
ATOM	9218	CA	SER	B	469	-1.447	5.806	132.563	1.00	7.35	B	C
ATOM	9224	CA	VAL	B	470	2.324	6.119	131.916	1.00	7.31	B	C
ATOM	9231	CA	ILE	B	471	3.054	4.452	135.299	1.00	7.55	B	C
ATOM	9239	CA	ALA	B	472	0.432	1.758	134.635	1.00	8.69	B	C
ATOM	9244	CA	ASP	B	473	2.077	0.765	131.318	1.00	10.11	B	C
ATOM	9252	CA	GLU	B	474	5.524	0.684	132.942	1.00	11.08	B	C
ATOM	9261	CA	LEU	B	475	4.081	-1.770	135.471	1.00	10.77	B	C
ATOM	9269	CA	GLY	B	476	2.557	-3.863	132.643	1.00	12.28	B	C
ATOM	9273	CA	LEU	B	477	-1.005	-2.946	133.703	1.00	14.48	B	C
ATOM	9281	CA	THR	B	478	-1.957	-2.607	130.034	1.00	17.13	B	C
ATOM	9288	CA	GLU	B	479	-5.736	-2.506	130.474	1.00	18.35	B	C
ATOM	9297	CA	GLU	B	480	-5.722	0.045	133.355	1.00	15.71	B	C
ATOM	9306	CA	ALA	B	481	-3.447	2.289	131.256	1.00	13.30	B	C
ATOM	9311	CA	GLU	B	482	-5.993	2.198	128.387	1.00	13.38	B	C
ATOM	9320	CA	THR	B	483	-8.838	3.007	130.776	1.00	11.91	B	C
ATOM	9327	CA	LEU	B	484	-6.802	5.857	132.279	1.00	9.11	B	C
ATOM	9335	CA	ARG	B	485	-6.030	7.235	128.765	1.00	8.25	B	C
ATOM	9346	CA	GLU	B	486	-9.735	7.030	127.831	1.00	8.59	B	C
ATOM	9355	CA	GLN	B	487	-10.721	8.982	130.961	1.00	7.84	B	C
ATOM	9364	CA	ALA	B	488	-7.917	11.568	130.491	1.00	6.89	B	C
ATOM	9369	CA	SER	B	489	-9.413	12.554	127.143	1.00	6.88	B	C
ATOM	9375	CA	TRP	B	490	-12.765	13.388	128.798	1.00	7.07	B	C
ATOM	9389	CA	PRO	B	491	-12.065	16.904	130.180	1.00	7.75	B	C
ATOM	9396	CA	ILE	B	492	-11.008	18.232	126.748	1.00	10.03	B	C

ATOM	9404	CA	GLU	B	493	-13.896	16.510	124.905	1.00	12.06	B	C
ATOM	9413	CA	LYS	B	494	-16.671	17.061	127.427	1.00	10.91	B	C
ATOM	9422	CA	LEU	B	495	-15.887	20.513	128.833	1.00	9.26	B	C
ATOM	9430	CA	THR	B	496	-14.827	22.544	125.800	1.00	9.84	B	C
ATOM	9437	CA	LYS	B	497	-17.972	24.524	124.878	1.00	10.36	B	C
ATOM	9446	CA	PRO	B	498	-18.062	25.503	121.171	1.00	10.16	B	C
ATOM	9453	CA	GLY	B	499	-18.441	29.251	120.587	1.00	11.73	B	C
ATOM	9457	CA	ALA	B	500	-17.533	30.238	124.169	1.00	12.89	B	C
ATOM	9462	CA	TRP	B	501	-15.579	33.120	122.622	1.00	13.32	B	C
ATOM	9476	CA	THR	B	502	-16.087	34.739	119.196	1.00	14.82	B	C
ATOM	9483	CA	ALA	B	503	-13.072	36.086	117.234	1.00	17.18	B	C
ATOM	9488	CA	ALA	B	504	-12.857	39.344	115.232	1.00	19.86	B	C
ATOM	9493	CA	ASP	B	505	-13.256	36.963	112.234	1.00	21.28	B	C
ATOM	9501	CA	GLY	B	506	-16.585	35.755	113.609	1.00	20.03	B	C
ATOM	9505	CA	THR	B	507	-14.731	32.439	114.087	1.00	18.03	B	C
ATOM	9512	CA	GLN	B	508	-16.251	30.532	117.010	1.00	15.34	B	C
ATOM	9521	CA	VAL	B	509	-13.708	29.336	119.570	1.00	11.34	B	C
ATOM	9528	CA	GLY	B	510	-14.366	26.537	122.049	1.00	9.07	B	C
ATOM	9532	CA	LEU	B	511	-13.197	27.026	125.631	1.00	7.86	B	C
ATOM	9540	CA	LEU	B	512	-13.219	25.124	128.921	1.00	7.01	B	C
ATOM	9548	CA	THR	B	513	-16.601	25.890	130.550	1.00	8.85	B	C
ATOM	9555	CA	PRO	B	514	-18.393	24.554	133.677	1.00	10.28	B	C
ATOM	9562	CA	ARG	B	515	-21.081	22.039	132.717	1.00	12.26	B	C
ATOM	9573	CA	ALA	B	516	-24.111	20.973	134.766	1.00	13.16	B	C
ATOM	9578	CA	ALA	B	517	-24.951	17.342	135.665	1.00	14.35	B	C
ATOM	9583	CA	ASP	B	518	-27.642	17.167	132.923	1.00	16.53	B	C
ATOM	9591	CA	GLY	B	519	-25.191	18.281	130.210	1.00	16.33	B	C
ATOM	9595	CA	SER	B	520	-26.240	21.945	129.971	1.00	15.96	B	C
ATOM	9601	CA	TRP	B	521	-23.559	24.648	130.008	1.00	15.21	B	C
ATOM	9615	CA	GLN	B	522	-23.432	26.929	133.060	1.00	15.39	B	C
ATOM	9624	CA	SER	B	523	-22.733	30.599	132.427	1.00	15.49	B	C
ATOM	9630	CA	ALA	B	524	-19.406	32.190	133.361	1.00	13.08	B	C
ATOM	9635	CA	ASP	B	525	-16.958	34.949	132.667	1.00	10.42	B	C
ATOM	9643	CA	HIS	B	526	-13.996	33.026	131.216	1.00	9.11	B	C
ATOM	9653	CA	ALA	B	527	-11.679	35.887	132.307	1.00	9.10	B	C
ATOM	9658	CA	LYS	B	528	-12.918	36.154	135.899	1.00	10.32	B	C
ATOM	9667	CA	PHE	B	529	-10.563	35.052	138.686	1.00	8.73	B	C
ATOM	9678	CA	GLU	B	530	-11.981	32.026	140.571	1.00	8.70	B	C
ATOM	9687	CA	ALA	B	531	-15.175	31.800	138.513	1.00	9.23	B	C
ATOM	9692	CA	ALA	B	532	-16.810	28.339	138.183	1.00	10.00	B	C
ATOM	9697	CA	GLY	B	533	-15.211	26.855	141.335	1.00	9.25	B	C
ATOM	9701	CA	LEU	B	534	-11.653	27.388	140.006	1.00	8.07	B	C
ATOM	9709	CA	TYR	B	535	-8.638	27.486	142.280	1.00	10.81	B	C
ATOM	9721	CA	GLN	B	536	-6.568	30.681	141.905	1.00	10.33	B	C
ATOM	9730	CA	GLY	B	537	-7.264	31.159	138.187	1.00	8.79	B	C
ATOM	9734	CA	THR	B	538	-9.626	31.771	135.278	1.00	7.44	B	C
ATOM	9741	CA	LEU	B	539	-11.154	29.505	132.643	1.00	8.31	B	C
ATOM	9749	CA	TRP	B	540	-8.976	31.006	129.900	1.00	6.79	B	C
ATOM	9763	CA	GLN	B	541	-6.001	30.062	132.087	1.00	5.72	B	C
ATOM	9772	CA	TYR	B	542	-7.247	26.567	133.014	1.00	6.07	B	C
ATOM	9784	CA	HIS	B	543	-8.336	25.987	129.378	1.00	6.44	B	C
ATOM	9794	CA	TRP	B	544	-4.804	24.912	128.489	1.00	5.93	B	C
ATOM	9808	CA	TYR	B	545	-4.279	22.715	131.546	1.00	4.79	B	C
ATOM	9820	CA	ASP	B	546	-4.407	19.324	129.817	1.00	4.56	B	C
ATOM	9828	CA	ALA	B	547	-0.984	19.835	128.277	1.00	5.44	B	C
ATOM	9833	CA	TYR	B	548	-0.609	16.142	127.393	1.00	6.08	B	C
ATOM	9845	CA	ASP	B	549	-3.092	16.487	124.510	1.00	5.83	B	C
ATOM	9853	CA	MET	B	550	-2.830	19.820	122.688	1.00	7.27	B	C
ATOM	9861	CA	ASP	B	551	-3.867	18.042	119.495	1.00	9.92	B	C
ATOM	9869	CA	ALA	B	552	-7.181	17.034	121.101	1.00	9.29	B	C
ATOM	9874	CA	LEU	B	553	-7.610	20.544	122.602	1.00	8.74	B	C
ATOM	9882	CA	VAL	B	554	-6.969	22.270	119.281	1.00	9.28	B	C
ATOM	9889	CA	GLU	B	555	-9.660	20.056	117.696	1.00	10.79	B	C
ATOM	9898	CA	ALA	B	556	-12.151	20.721	120.539	1.00	8.98	B	C
ATOM	9903	CA	MET	B	557	-11.629	24.480	120.201	1.00	9.27	B	C
ATOM	9911	CA	GLY	B	558	-12.831	24.096	116.592	1.00	11.18	B	C

ATOM	9915	CA	GLY	B	559	-9.648	23.449	114.651	1.00	12.57	B	C
ATOM	9919	CA	HIS	B	560	-6.240	24.925	113.907	1.00	13.76	B	C
ATOM	9929	CA	GLU	B	561	-7.665	28.397	113.047	1.00	13.40	B	C
ATOM	9938	CA	ALA	B	562	-9.852	28.561	116.156	1.00	11.93	B	C
ATOM	9943	CA	ALA	B	563	-6.829	27.744	118.342	1.00	11.17	B	C
ATOM	9948	CA	ARG	B	564	-4.548	30.202	116.523	1.00	10.14	B	C
ATOM	9959	CA	LEU	B	565	-6.933	33.107	117.193	1.00	9.78	B	C
ATOM	9967	CA	GLY	B	566	-7.517	31.781	120.725	1.00	8.93	B	C
ATOM	9971	CA	MET	B	567	-3.792	31.948	121.520	1.00	9.45	B	C
ATOM	9979	CA	ARG	B	568	-3.398	35.497	120.126	1.00	9.92	B	C
ATOM	9990	CA	HIS	B	569	-6.452	36.629	122.170	1.00	9.91	B	C
ATOM	10000	CA	MET	B	570	-4.961	34.969	125.270	1.00	10.22	B	C
ATOM	10008	CA	PHE	B	571	-1.992	37.376	124.985	1.00	9.43	B	C
ATOM	10019	CA	GLY	B	572	-3.956	40.469	123.877	1.00	11.08	B	C
ATOM	10023	CA	GLU	B	573	-2.195	40.540	120.475	1.00	13.30	B	C
ATOM	10032	CA	HIS	B	574	-5.029	42.598	118.931	1.00	14.86	B	C
ATOM	10042	CA	ALA	B	575	-4.622	45.304	121.645	1.00	15.90	B	C
ATOM	10047	CA	PRO	B	576	-0.972	45.390	122.696	1.00	17.21	B	C
ATOM	10054	CA	ASP	B	577	-1.328	48.595	124.695	1.00	18.80	B	C
ATOM	10062	CA	ASP	B	578	-4.343	47.440	126.680	1.00	16.37	B	C
ATOM	10070	CA	GLY	B	579	-3.447	45.773	129.999	1.00	14.31	B	C
ATOM	10074	CA	LYS	B	580	-6.893	44.273	130.536	1.00	14.35	B	C
ATOM	10083	CA	ALA	B	581	-6.401	42.289	127.289	1.00	11.55	B	C
ATOM	10088	CA	MET	B	582	-3.732	40.150	128.992	1.00	9.08	B	C
ATOM	10096	CA	LEU	B	583	-5.611	36.913	129.818	1.00	6.75	B	C
ATOM	10104	CA	HIS	B	584	-2.406	35.066	130.755	1.00	5.26	B	C
ATOM	10114	CA	SER	B	585	-1.183	35.314	134.331	1.00	5.08	B	C
ATOM	10120	CA	ASN	B	586	2.561	35.487	135.130	1.00	4.87	B	C
ATOM	10128	CA	ALA	B	587	1.917	35.442	138.923	1.00	6.16	B	C
ATOM	10133	CA	ASN	B	588	0.684	31.857	139.540	1.00	9.70	B	C
ATOM	10141	CA	GLU	B	589	1.574	28.445	138.079	1.00	11.52	B	C
ATOM	10150	CA	ILE	B	590	-1.724	27.523	136.394	1.00	11.76	B	C
ATOM	10158	CA	ASP	B	591	-0.787	28.589	132.835	1.00	11.65	B	C
ATOM	10166	CA	LEU	B	592	3.036	28.784	133.060	1.00	10.00	B	C
ATOM	10174	CA	GLN	B	593	3.220	26.855	129.797	1.00	7.99	B	C
ATOM	10183	CA	ALA	B	594	0.955	29.258	127.891	1.00	7.42	B	C
ATOM	10188	CA	PRO	B	595	3.697	31.434	126.264	1.00	6.47	B	C
ATOM	10195	CA	TYR	B	596	5.040	28.300	124.524	1.00	5.99	B	C
ATOM	10207	CA	LEU	B	597	1.684	27.269	123.043	1.00	5.41	B	C
ATOM	10215	CA	PHE	B	598	2.175	29.492	119.962	1.00	6.82	B	C
ATOM	10226	CA	ASN	B	599	4.570	26.728	118.788	1.00	7.99	B	C
ATOM	10234	CA	TYR	B	600	1.469	24.494	118.559	1.00	8.95	B	C
ATOM	10246	CA	THR	B	601	-0.592	27.029	116.565	1.00	9.34	B	C
ATOM	10253	CA	GLY	B	602	2.026	27.493	113.815	1.00	9.48	B	C
ATOM	10257	CA	GLU	B	603	3.299	30.812	115.139	1.00	9.51	B	C
ATOM	10266	CA	PRO	B	604	6.593	29.991	116.946	1.00	8.50	B	C
ATOM	10273	CA	SER	B	605	7.845	33.578	116.478	1.00	7.57	B	C
ATOM	10279	CA	LEU	B	606	5.226	34.638	119.060	1.00	7.18	B	C
ATOM	10287	CA	THR	B	607	6.464	32.030	121.578	1.00	5.66	B	C
ATOM	10294	CA	GLN	B	608	9.959	33.487	121.141	1.00	5.96	B	C
ATOM	10303	CA	LYS	B	609	8.701	37.059	121.593	1.00	6.51	B	C
ATOM	10312	CA	TRP	B	610	6.714	36.241	124.769	1.00	7.00	B	C
ATOM	10326	CA	ALA	B	611	9.360	33.957	126.314	1.00	7.18	B	C
ATOM	10331	CA	ARG	B	612	11.852	36.768	125.860	1.00	7.07	B	C
ATOM	10342	CA	ALA	B	613	9.502	39.547	127.021	1.00	7.16	B	C
ATOM	10347	CA	ILE	B	614	8.036	37.923	130.160	1.00	7.15	B	C
ATOM	10355	CA	TYR	B	615	11.442	37.037	131.623	1.00	8.13	B	C
ATOM	10367	CA	THR	B	616	13.573	39.966	130.409	1.00	8.81	B	C
ATOM	10374	CA	LYS	B	617	11.308	42.977	129.716	1.00	9.89	B	C
ATOM	10383	CA	GLU	B	618	8.250	44.801	130.976	1.00	10.62	B	C
ATOM	10392	CA	THR	B	619	4.925	43.183	130.129	1.00	10.85	B	C
ATOM	10399	CA	TRP	B	620	1.277	43.848	130.780	1.00	10.87	B	C
ATOM	10413	CA	ASN	B	621	0.104	42.242	133.955	1.00	8.54	B	C
ATOM	10421	CA	ARG	B	622	-3.614	41.858	134.675	1.00	6.42	B	C
ATOM	10432	CA	TYR	B	623	-3.638	39.092	137.312	1.00	6.84	B	C
ATOM	10444	CA	ILE	B	624	-2.166	38.601	140.777	1.00	6.95	B	C

ATOM	10452	CA	ALA	B	625	-2.036	35.146	142.429	1.00	9.17	B	C
ATOM	10457	CA	THR	B	626	-4.883	35.743	144.940	1.00	9.61	B	C
ATOM	10464	CA	GLY	B	627	-8.057	37.828	145.428	1.00	11.00	B	C
ATOM	10468	CA	SER	B	628	-6.410	40.988	146.753	1.00	14.00	B	C
ATOM	10474	CA	SER	B	629	-3.055	42.511	147.827	1.00	14.96	B	C
ATOM	10480	CA	SER	B	630	-2.186	45.392	150.152	1.00	15.87	B	C
ATOM	10486	CA	ALA	B	631	1.099	45.793	148.230	1.00	15.15	B	C
ATOM	10491	CA	VAL	B	632	-0.451	46.993	144.954	1.00	12.68	B	C
ATOM	10498	CA	PRO	B	633	-3.837	48.543	144.078	1.00	11.90	B	C
ATOM	10505	CA	SER	B	634	-5.790	45.403	143.232	1.00	10.24	B	C
ATOM	10511	CA	GLY	B	635	-9.047	43.424	143.655	1.00	10.87	B	C
ATOM	10515	CA	GLY	B	636	-10.965	40.434	142.243	1.00	11.37	B	C
ATOM	10519	CA	GLY	B	637	-7.618	38.717	141.514	1.00	10.74	B	C
ATOM	10523	CA	GLU	B	638	-6.354	41.552	139.331	1.00	10.06	B	C
ATOM	10532	CA	PHE	B	639	-4.034	44.542	139.288	1.00	10.96	B	C
ATOM	10543	CA	THR	B	640	-6.497	47.479	139.174	1.00	13.37	B	C
ATOM	10550	CA	PRO	B	641	-5.496	48.960	136.843	1.00	14.12	B	C
ATOM	10557	CA	PRO	B	642	-3.275	46.456	134.964	1.00	13.72	B	C
ATOM	10564	CA	LEU	B	643	0.396	47.404	135.238	1.00	14.65	B	C
ATOM	10572	CA	LYS	B	644	3.137	47.317	132.691	1.00	13.96	B	C
ATOM	10581	CA	THR	B	645	6.095	45.936	134.647	1.00	11.78	B	C
ATOM	10588	CA	LYS	B	646	8.969	43.450	134.688	1.00	11.14	B	C
ATOM	10597	CA	VAL	B	647	7.800	40.166	136.218	1.00	8.36	B	C
ATOM	10604	CA	TYR	B	648	11.303	39.198	137.406	1.00	7.74	B	C
ATOM	10616	CA	ARG	B	649	13.591	41.824	139.020	1.00	9.04	B	C
ATOM	10627	CA	LEU	B	650	17.093	41.707	140.433	1.00	7.53	B	C
ATOM	10635	CA	ASP	B	651	15.807	43.347	143.621	1.00	5.17	B	C
ATOM	10643	CA	PRO	B	652	14.545	42.350	147.055	1.00	5.40	B	C
ATOM	10650	CA	ARG	B	653	11.139	42.853	145.381	1.00	6.53	B	C
ATOM	10661	CA	GLY	B	654	12.108	40.059	142.996	1.00	6.41	B	C
ATOM	10665	CA	MET	B	655	8.705	38.795	141.823	1.00	6.72	B	C
ATOM	10673	CA	LEU	B	656	5.334	40.460	141.196	1.00	6.70	B	C
ATOM	10681	CA	PRO	B	657	3.402	41.236	144.385	1.00	6.71	B	C
ATOM	10688	CA	THR	B	658	1.659	37.972	145.528	1.00	7.41	B	C
ATOM	10695	CA	MET	B	659	3.892	35.925	143.184	1.00	5.95	B	C
ATOM	10703	CA	ASP	B	660	5.190	33.199	145.543	1.00	4.96	B	C
ATOM	10711	CA	ASN	B	661	8.255	31.368	144.118	1.00	5.08	B	C
ATOM	10719	CA	ASP	B	662	6.702	28.114	145.513	1.00	4.85	B	C
ATOM	10727	CA	ALA	B	663	9.647	25.783	146.215	1.00	5.19	B	C
ATOM	10732	CA	GLY	B	664	11.674	27.503	143.453	1.00	5.63	B	C
ATOM	10736	CA	THR	B	665	9.108	26.987	140.680	1.00	6.11	B	C
ATOM	10743	CA	MET	B	666	9.161	30.568	139.404	1.00	7.32	B	C
ATOM	10751	CA	SER	B	667	12.973	30.544	139.595	1.00	7.14	B	C
ATOM	10757	CA	THR	B	668	13.107	27.187	137.738	1.00	8.41	B	C
ATOM	10764	CA	MET	B	669	10.894	28.674	134.975	1.00	8.61	B	C
ATOM	10772	CA	PHE	B	670	13.293	31.613	134.474	1.00	5.87	B	C
ATOM	10783	CA	VAL	B	671	16.232	29.234	134.165	1.00	5.19	B	C
ATOM	10790	CA	ALA	B	672	14.275	27.232	131.532	1.00	5.77	B	C
ATOM	10795	CA	ALA	B	673	13.327	30.366	129.576	1.00	6.53	B	C
ATOM	10800	CA	ALA	B	674	17.004	31.392	129.591	1.00	6.52	B	C
ATOM	10805	CA	VAL	B	675	18.093	27.945	128.278	1.00	6.08	B	C
ATOM	10812	CA	GLY	B	676	15.236	28.143	125.757	1.00	5.59	B	C
ATOM	10816	CA	LEU	B	677	13.801	24.680	126.537	1.00	4.36	B	C
ATOM	10824	CA	PHE	B	678	10.456	24.513	128.310	1.00	4.27	B	C
ATOM	10835	CA	PRO	B	679	8.476	21.441	129.468	1.00	4.23	B	C
ATOM	10842	CA	VAL	B	680	4.935	22.169	128.193	1.00	4.71	B	C
ATOM	10849	CA	THR	B	681	4.133	18.524	128.725	1.00	4.02	B	C
ATOM	10856	CA	ALA	B	682	5.534	17.401	132.028	1.00	2.99	B	C
ATOM	10861	CA	GLY	B	683	5.162	13.611	132.013	1.00	4.05	B	C
ATOM	10865	CA	SER	B	684	6.552	13.443	128.463	1.00	5.13	B	C
ATOM	10871	CA	SER	B	685	10.143	13.369	127.150	1.00	6.66	B	C
ATOM	10877	CA	GLN	B	686	9.863	16.641	125.200	1.00	6.38	B	C
ATOM	10886	CA	PHE	B	687	10.738	20.344	125.639	1.00	5.59	B	C
ATOM	10897	CA	GLN	B	688	9.427	23.232	123.520	1.00	6.03	B	C
ATOM	10906	CA	VAL	B	689	11.793	25.796	122.043	1.00	6.55	B	C
ATOM	10913	CA	GLY	B	690	11.579	29.419	123.274	1.00	7.04	B	C

ATOM	10917	CA	SER	B	691	14.106	32.286	122.899	1.00	7.79	B	C
ATOM	10923	CA	PRO	B	692	17.484	31.426	124.521	1.00	7.84	B	C
ATOM	10930	CA	PHE	B	693	18.864	34.550	126.227	1.00	7.82	B	C
ATOM	10941	CA	PHE	B	694	22.594	34.064	125.890	1.00	7.33	B	C
ATOM	10952	CA	ASP	B	695	25.205	33.655	123.125	1.00	9.43	B	C
ATOM	10960	CA	SER	B	696	26.210	30.332	124.574	1.00	9.32	B	C
ATOM	10966	CA	THR	B	697	24.781	28.206	127.383	1.00	8.98	B	C
ATOM	10973	CA	THR	B	698	26.586	24.967	128.241	1.00	8.91	B	C
ATOM	10980	CA	ILE	B	699	25.297	22.278	130.616	1.00	9.45	B	C
ATOM	10988	CA	THR	B	700	28.086	19.908	131.647	1.00	10.12	B	C
ATOM	10995	CA	TYR	B	701	27.061	16.493	132.887	1.00	9.88	B	C
ATOM	11007	CA	ASP	B	702	28.712	14.246	135.442	1.00	11.45	B	C
ATOM	11015	CA	ASP	B	703	30.657	12.267	132.819	1.00	13.40	B	C
ATOM	11023	CA	GLY	B	704	32.161	15.461	131.345	1.00	11.98	B	C
ATOM	11027	CA	SER	B	705	29.921	15.485	128.268	1.00	10.92	B	C
ATOM	11033	CA	ALA	B	706	27.904	18.613	127.584	1.00	10.52	B	C
ATOM	11038	CA	PHE	B	707	24.798	20.036	125.938	1.00	9.26	B	C
ATOM	11049	CA	THR	B	708	25.464	23.375	124.241	1.00	8.88	B	C
ATOM	11056	CA	VAL	B	709	22.737	25.748	123.064	1.00	9.09	B	C
ATOM	11063	CA	THR	B	710	24.043	28.747	121.128	1.00	11.17	B	C
ATOM	11070	CA	ALA	B	711	22.134	31.834	120.050	1.00	12.24	B	C
ATOM	11075	CA	ASP	B	712	24.141	33.564	117.322	1.00	14.25	B	C
ATOM	11083	CA	GLY	B	713	23.722	37.334	117.271	1.00	12.57	B	C
ATOM	11087	CA	VAL	B	714	21.303	37.259	120.194	1.00	12.10	B	C
ATOM	11094	CA	SER	B	715	20.477	40.687	121.622	1.00	11.79	B	C
ATOM	11100	CA	GLU	B	716	17.633	42.447	123.438	1.00	13.43	B	C
ATOM	11109	CA	ASP	B	717	16.112	43.108	119.987	1.00	12.74	B	C
ATOM	11117	CA	ALA	B	718	17.337	39.953	118.227	1.00	11.10	B	C
ATOM	11122	CA	PHE	B	719	15.169	37.456	120.135	1.00	10.51	B	C
ATOM	11133	CA	TYR	B	720	13.602	35.772	117.045	1.00	10.60	B	C
ATOM	11145	CA	VAL	B	721	14.971	32.509	115.687	1.00	9.51	B	C
ATOM	11152	CA	GLN	B	722	15.866	32.757	111.949	1.00	9.69	B	C
ATOM	11161	CA	SER	B	723	17.260	29.236	111.377	1.00	10.67	B	C
ATOM	11167	CA	ALA	B	724	18.749	26.372	113.367	1.00	10.44	B	C
ATOM	11172	CA	THR	B	725	21.057	23.385	113.295	1.00	10.19	B	C
ATOM	11179	CA	LEU	B	726	21.285	20.299	115.497	1.00	9.79	B	C
ATOM	11187	CA	ASP	B	727	24.740	18.682	115.470	1.00	10.78	B	C
ATOM	11195	CA	GLY	B	728	25.486	20.479	112.168	1.00	9.59	B	C
ATOM	11199	CA	ALA	B	729	22.291	19.355	110.407	1.00	9.37	B	C
ATOM	11204	CA	THR	B	730	19.314	21.561	109.427	1.00	9.54	B	C
ATOM	11211	CA	PHE	B	731	16.911	21.620	112.380	1.00	9.46	B	C
ATOM	11222	CA	GLY	B	732	13.291	22.799	111.894	1.00	10.01	B	C
ATOM	11226	CA	ASN	B	733	11.327	21.345	114.825	1.00	10.28	B	C
ATOM	11234	CA	THR	B	734	9.967	23.567	117.612	1.00	10.95	B	C
ATOM	11241	CA	TRP	B	735	10.620	20.889	120.262	1.00	9.83	B	C
ATOM	11255	CA	VAL	B	736	13.510	18.634	121.295	1.00	9.57	B	C
ATOM	11262	CA	ASP	B	737	13.581	15.277	123.044	1.00	9.41	B	C
ATOM	11270	CA	TYR	B	738	15.214	14.889	126.458	1.00	7.09	B	C
ATOM	11282	CA	ALA	B	739	17.644	12.232	125.051	1.00	6.73	B	C
ATOM	11287	CA	THR	B	740	19.020	14.795	122.576	1.00	6.66	B	C
ATOM	11294	CA	VAL	B	741	19.805	17.297	125.352	1.00	6.54	B	C
ATOM	11301	CA	VAL	B	742	21.387	14.908	127.847	1.00	6.53	B	C
ATOM	11308	CA	GLY	B	743	23.160	13.136	124.956	1.00	8.60	B	C
ATOM	11312	CA	GLY	B	744	25.256	16.293	124.535	1.00	8.96	B	C
ATOM	11316	CA	ALA	B	745	23.948	17.869	121.301	1.00	8.77	B	C
ATOM	11321	CA	ASP	B	746	24.948	21.245	119.824	1.00	9.99	B	C
ATOM	11329	CA	LEU	B	747	21.727	23.156	119.256	1.00	9.20	B	C
ATOM	11337	CA	ALA	B	748	22.723	26.298	117.347	1.00	9.57	B	C
ATOM	11342	CA	PHE	B	749	20.245	29.087	116.636	1.00	9.42	B	C
ATOM	11353	CA	ARG	B	750	20.864	32.051	114.353	1.00	10.76	B	C
ATOM	11364	CA	MET	B	751	18.965	35.015	115.906	1.00	10.21	B	C
ATOM	11372	CA	GLY	B	752	17.572	38.240	114.387	1.00	12.06	B	C
ATOM	11376	CA	GLU	B	753	15.318	41.247	114.999	1.00	14.15	B	C
ATOM	11385	CA	GLN	B	754	12.309	40.094	112.922	1.00	15.06	B	C
ATOM	11394	CA	PRO	B	755	9.930	37.094	113.128	1.00	14.14	B	C
ATOM	11401	CA	SER	B	756	10.685	34.248	110.709	1.00	14.80	B	C

ATOM	11407	CA	ASP	B	757	8.804	31.176	109.465	1.00	14.96	B	C
ATOM	11415	CA	TRP	B	758	11.407	28.912	111.150	1.00	13.21	B	C
ATOM	11429	CA	GLY	B	759	9.689	25.918	112.773	1.00	13.37	B	C
ATOM	11433	CA	THR	B	760	6.357	26.434	111.014	1.00	13.94	B	C
ATOM	11440	CA	ASP	B	761	6.914	23.033	109.382	1.00	13.96	B	C
ATOM	11448	CA	THR	B	762	7.829	21.295	112.640	1.00	10.47	B	C
ATOM	11455	CA	ALA	B	763	7.173	17.563	113.052	1.00	9.77	B	C
ATOM	11460	CA	PRO	B	764	4.213	17.436	115.464	1.00	9.45	B	C
ATOM	11467	CA	ALA	B	765	4.950	17.568	119.216	1.00	7.60	B	C
ATOM	11472	CA	PHE	B	766	3.648	14.939	121.659	1.00	7.84	B	C
ATOM	11483	CA	SER	B	767	-0.100	14.561	122.179	1.00	8.05	B	C
ATOM	11489	CA	MET	B	768	-1.670	11.546	123.865	1.00	10.37	B	C
ATOM	11497	CA	SER	B	769	-4.411	11.138	121.212	1.00	13.56	B	C
ATOM	11503	CA	THR	B	770	-1.982	11.233	118.242	1.00	17.85	B	C
ATOM	11510	CA	ALA	B	771	0.792	9.094	119.762	1.00	20.44	B	C
ATOM	12382	CA	CA	C	1	41.425	16.915	81.268	1.00	30.05	C	CA
ATOM	12383	CA	CA	C	6	1.503	30.134	143.317	1.00	32.32	C	CA

END