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(71) Applicant: **EVOLVA SA** [CH/CH]; Duggingerstrasse 23, 4153 Reinach (CH).

(72) Inventors: **OLSSON, Kim**; Finsensvej 46B, 1tv., 2000 Frederiksberg (DK). **SIMON, Ernesto**; Svaetegade 7, 2nd Floor, 1118 Copenhagen (DK). **DALGAARD MIKKELSEN, Michael**; 1 Bregnevej, 3500 Vaerløse (DK). **CARLSEN, Simon**; Duggingerstrasse 23, CH-4153 Reinach (CH). **DOUCHIN, Veronique**; Aurikelvej 2-4th, 2000 Frederiksberg (DK). **LIM, Swee Chuang**; Amalieparken 87, 4-4, 2665 Vallensbaek Strand (DK). **DURING, Louis**; Classensgade 23, 4th, 2100 Copenhagen (DK).

(74) Agent: **SMAGGASGALE, Gillian**; 55 Drury Lane, London WC2B 5SQ (GB).

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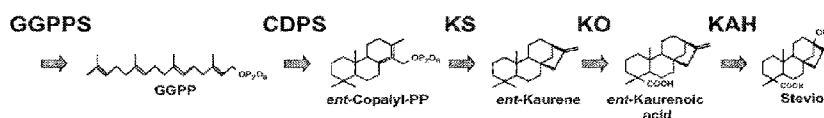
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(54) Title: PRODUCTION OF STEVIOL GLYCOSIDES IN RECOMBINANT HOSTS

Figure 1



(57) Abstract: The invention relates to recombinant microorganisms and methods for producing steviol glycosides, glycosylated ent-kaurenol, and glycosylated ent-kaurenoic acid.



## PRODUCTION OF STEVIOL GLYCOSIDES IN RECOMBINANT HOSTS

### BACKGROUND OF THE INVENTION

#### Field of the Invention

**[0001]** This disclosure relates to recombinant production of steviol glycosides and steviol glycoside precursors in recombinant hosts. In particular, this disclosure relates to production of steviol glycosides comprising steviol-13-O-glucoside (13-SMG), steviol-19-O-glucoside (19-SMG), steviol-1,2-bioside, steviol-1,3-bioside, 1,2-stevioside, 1,3-stevioside, rubusoside (Rubu), rebaudioside A (RebA), rebaudioside B (RebB), rebaudioside D (RebD), rebaudioside E (RebE), rebaudioside M (RebM), rebaudioside Q (RebQ), rebaudioside I (RebI), di-glycosylated steviol, tri-glycosylated steviol, tetra-glycosylated steviol, penta-glycosylated steviol, hexa-glycosylated steviol, hepta-glycosylated steviol, glycosylated ent-kaurenol, glycosylated ent-kaurenoic acid, and/or isomers thereof in recombinant hosts.

#### Description of Related Art

**[0002]** Sweeteners are well known as ingredients used most commonly in the food, beverage, or confectionary industries. The sweetener can either be incorporated into a final food product during production or for stand-alone use, when appropriately diluted, as a tabletop sweetener or an at-home replacement for sugars in baking. Sweeteners include natural sweeteners such as sucrose, high fructose corn syrup, molasses, maple syrup, and honey and artificial sweeteners such as aspartame, saccharine, and sucralose. Stevia extract is a natural sweetener that can be isolated and extracted from a perennial shrub, *Stevia rebaudiana*. Stevia is commonly grown in South America and Asia for commercial production of stevia extract. Stevia extract, purified to various degrees, is used commercially as a high intensity sweetener in foods and in blends or alone as a tabletop sweetener.

**[0003]** Chemical structures for several steviol glycosides are shown in Figure 1, including the diterpene steviol and various steviol glycosides. Extracts of the Stevia plant generally comprise steviol glycosides that contribute to the sweet flavor, although the amount of each steviol glycoside often varies, *inter alia*, among different production batches.

**[0004]** As recovery and purification of steviol glycosides from the Stevia plant have proven to be labor intensive and inefficient, there remains a need for a recombinant production system

that can accumulate high yields of desired steviol glycosides, such as RebD and RebM. There also remains a need for improved production of steviol glycosides in recombinant hosts for commercial uses. As well, there remains a need for identifying enzymes selective towards particular substrates to produce one or more specific steviol glycosides. In some aspects, there remains a need to increase the catalytic capability of enzymes with 19-O glycosylation activity in order to produce higher yields of steviol glycosides.

### SUMMARY OF THE INVENTION

**[0005]** It is against the above background that the present invention provides certain advantages and advancements over the prior art.

**[0006]** Although this invention as disclosed herein is not limited to specific advantages or functionalities, the invention provides a recombinant host cell, comprising at least one recombinant gene that is:

(a) a gene encoding a UGT91D2e polypeptide having at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO:11;

(b) a gene encoding a chimeric polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:17 or SEQ ID NO:18;

(c) a gene encoding a UGT85C2 polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7; and/or

(d) a gene encoding a UGT76G1 polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;

wherein the recombinant host cell is capable of producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or a glycosylated ent-kaurenoic acid compound in a cell culture broth.

**[0007]** In one aspect of the recombinant host cell disclosed herein, the UGT91D2e polypeptide comprises a UGT91D2e polypeptide having at least one amino acid substitution at residues 93, 99, 114, 144, 148, 152, 195, 196, 199, 211, 213, 221, 286, 384, 426, 438, or 466 of SEQ ID NO:11.

**[0008]** In one aspect of the recombinant host cell disclosed herein, the UGT85C2 polypeptide comprises a UGT85C2 polypeptide having at least one amino acid substitution at residues 21, 48, 49, 84, 86, 87, 91, 92, 95, 122, 334, or 334 of SEQ ID NO:7.

**[0009]** In one aspect of the recombinant host cell disclosed herein, the UGT76G1 polypeptide comprises a UGT76G1 polypeptide having at least one amino acid substitution at residues 23, 26, 55, 146, 257, 283, and 337 of SEQ ID NO:9.

**[0010]** In one aspect of the recombinant host cell disclosed herein, the UGT91D2e polypeptide comprises one or more of the UGT91D2e polypeptide variants comprising: P93V, S99I, S114F, T144K, T144L, T144M, A148K, M152T, L195G, L195C, L195S, L195N, L195V, V196P, K199C, L211H, L211M, L211I, L211C, L211T, L213E, S221I, V286C, V286N, V286S, G384W, G384K, G384Y, E426G, E438H, 3438M or A466V of SEQ ID NO:11.

**[0011]** In one aspect of the recombinant host cell disclosed herein, the UGT85C2 polypeptide comprises one or more of the UGT85C2 polypeptide variants comprising: Q21L, Q21T, Q21V, F48S, F48H, F48Y, F48R, F48Q, F48W, F48T, I49V, S84G, S84A, S84T, S84C, S84P, S84N, S84V, P86R, P86G, I87H, I87P, I87M, I87Y, L91K, L91R, L91T, L92F, L92I, L92M, I95K, F122S, L334S or L334M of SEQ ID NO:7.

**[0012]** In one aspect of the recombinant host cell disclosed herein, the UGT76G1 polypeptide comprises one or more of the UGT76G1 polypeptide variants comprising: Q23H, I26W, T146G, H155L, L257G, S253W, T284G, S283N, K337P or T55K of SEQ ID NO:9.

**[0013]** In one aspect the recombinant host cell disclosed herein further comprises at least one recombinant gene that is:

- (a) a gene encoding a geranylgeranyl diphosphate synthase (GGPPS) polypeptide;
- (b) a gene encoding an ent-copalyl diphosphate synthase (CDPS) polypeptide;
- (c) a gene encoding an ent-kaurene synthase (KS) polypeptide;
- (d) a gene encoding an ent-kaurene oxidase (KO) polypeptide;
- (e) a gene encoding a cytochrome P450 reductase (CPR) polypeptide; and
- (f) a gene encoding an ent-kaurenoic acid hydroxylase (KAH) polypeptide;
- (g) a gene encoding a UGT74G1 polypeptide; and/or
- (h) a gene encoding an EUGT11 polypeptide;

wherein the recombinant host cell capable of producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or a glycosylated ent-kaurenoic acid compound in a cell culture broth.

**[0014]** In one aspect of the recombinant host cell disclosed herein,

(a) the GGPPS polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, or SEQ ID NO:116;

(b) the CDPS polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, or SEQ ID NO:42;

(c) the KS polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, or SEQ ID NO:52;

(d) the KO polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:117, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:74, or SEQ ID NO:76;

(e) the CPR polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:82, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:88, SEQ ID NO:90, SEQ ID NO:92;

(f) the KAH polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:94, SEQ ID NO:97, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:106, SEQ ID NO:108, SEQ ID NO:110, SEQ ID NO:112, or SEQ ID NO:114;

(g) the UGT74G1 polypeptide comprises a polypeptide having at least 55% identity to an amino acid sequence set forth in SEQ ID NO:4;

(h) the EUGT11 polypeptide comprises a polypeptide having at least 65% identity to an amino acid sequence set forth in SEQ ID NO:16.

**[0015]** In one aspect of the recombinant host cell disclosed herein, the cell culture broth comprises:

(a) the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound produced by the recombinant host cell,

(b) glucose, fructose and/or sucrose; and/or

(c) supplemental nutrients comprising trace metals, vitamins, salts, yeast nitrogen base (YNB), and/or amino acids.

**[0016]** In one aspect of the recombinant host cell disclosed herein, the recombinant host comprises a plant cell, a mammalian cell, an insect cell, a fungal cell, an algal cell, or a bacterial cell.

**[0017]** In one aspect of the recombinant host cell disclosed herein, the bacterial cell comprises *Escherichia* cells, *Lactobacillus* cells, *Lactococcus* cells, *Cornibacterium* cells, *Acetobacter* cells, *Acinetobacter* cells, or *Pseudomonas* cells.

**[0018]** In one aspect of the recombinant host cell disclosed herein, the fungal cell comprises a yeast cell.

**[0019]** In one aspect of the recombinant host cell disclosed herein, the yeast cell is a cell from *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Yarrowia lipolytica*, *Candida glabrata*, *Ashbya gossypii*, *Cyberlindnera jadinii*, *Pichia pastoris*, *Kluyveromyces lactis*, *Hansenula polymorpha*, *Candida boidinii*, *Arxula adenivorans*, *Xanthophyllomyces dendrorhous*, or *Candida albicans* species.

**[0020]** In one aspect of the recombinant host cell disclosed herein, the yeast cell is a *Saccharomycete*.

**[0021]** In one aspect of the recombinant host cell disclosed herein, the yeast cell is a cell from the *Saccharomyces cerevisiae* species.

**[0022]** The invention also provides a method of producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or glycosylated ent-kaurenoic acid compound in a cell culture broth, comprising growing the recombinant host cell disclosed herein in a culture medium, under conditions in which one or more of the genes are expressed;

wherein at least one of the genes is a recombinant gene;

wherein the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound is produced by the recombinant host cell.

**[0023]** In one aspect of the methods disclosed herein, one or more of the genes is constitutively expressed and/or expression of one or more of the genes is induced.

**[0024]** The invention also provides a method for producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound comprising whole-cell bioconversion of plant-derived components or synthetic steviol or steviol glycosides using one or more of:

- (a) a UGT91D2e polypeptide having at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO:11;
- (b) a chimeric polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:17 or SEQ ID NO:18;
- (c) a UGT85C2 polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7; and/or
- (d) a UGT76G1 polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;

wherein at least one of the polypeptides is a recombinant polypeptide.

**[0025]** In one aspect of the methods disclosed herein, the whole cell is the recombinant host cell disclosed herein.

**[0026]** In one aspect of the methods disclosed herein, the recombinant host cell is grown in a fermentor at a temperature for a period of time, wherein the temperature and period of time facilitate the production of the steviol glycoside, glycosylated ent-kaurenol compound, and/or glycosylated ent-kaurenoic acid compound.

**[0027]** The invention also provides an *in vitro* method for producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound, comprising adding one or more of:

- (a) a UGT91D2e polypeptide having at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO:11;
- (b) a chimeric polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:17 or SEQ ID NO:18;
- (c) a UGT85C2 polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7; and/or
- (d) a UGT76G1 polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9,

and plant-derived components or synthetic steviol or steviol glycosides to a reaction mixture;

wherein at least one of the polypeptides is a recombinant polypeptide; and

- (b) synthesizing steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound in the reaction mixture.

**[0028]** In one aspect, methods disclosed herein further comprise isolating the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound, alone or in combination from the cell culture broth.

**[0029]** In one aspect of the methods disclosed herein, the isolating step comprises:

- (a) providing the cell culture broth comprising the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or in combination;
- (b) separating a liquid phase of the cell culture broth from a solid phase of the cell culture broth to obtain a supernatant comprising the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or in combination;
- (c) providing one or more adsorbent resins, comprising providing the adsorbent resins in a packed column; and
- (d) contacting the supernatant of step (b) with the one or more adsorbent resins in order to obtain at least a portion of the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or in combination thereby isolating the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or in combination.

**[0030]** In one aspect, methods disclosed herein further comprise recovering the the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or a composition comprising the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound.

**[0031]** In one aspect of the methods disclosed herein, the recovered composition is enriched for the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound relative to a steviol glycoside composition of Stevia plant and has a reduced level of non-steviol glycoside Stevia plant-derived components relative to a plant-derived stevia extract.

**[0032]** In one aspect of the methods disclosed herein, the cell culture broth comprises:

- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or glycosylated ent-kaurenoic acid compounds produced by the recombinant host cell disclosed herein,
- (b) glucose, fructose, and/or sucrose; and/or
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids.

**[0033]** In one aspect of the methods disclosed herein, the reaction mixture comprising:

- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or a glycosylated ent-kaurenoic acid compounds produced in the reaction mixture;
- (b) a UGT polypeptide;
- (c) UDP-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and/or
- (d) reaction buffer and/or salts.

**[0034]** In one aspect of the methods disclosed herein, the recombinant host cell comprises a plant cell, a mammalian cell, an insect cell, a fungal cell, an algal cell, or a bacterial cell.

**[0035]** In one aspect of the methods disclosed herein, the bacterial cell comprises *Escherichia* cells, *Lactobacillus* cells, *Lactococcus* cells, *Cornibacterium* cells, *Acetobacter* cells, *Acinetobacter* cells, or *Pseudomonas* cells.

**[0036]** In one aspect of the methods disclosed herein, the fungal cell comprises a yeast cell.

**[0037]** In one aspect of the methods disclosed herein, the yeast cell is a cell from *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Yarrowia lipolytica*, *Candida glabrata*, *Ashbya gossypii*, *Cyberlindnera jadinii*, *Pichia pastoris*, *Kluyveromyces lactis*, *Hansenula polymorpha*, *Candida boidinii*, *Arxula adenivorans*, *Xanthophyllomyces dendrorhous*, or *Candida albicans* species.

**[0038]** In one aspect of the methods disclosed herein, the yeast cell is a *Saccharomycete*.

**[0039]** In one aspect of the methods disclosed herein, the yeast cell is a cell from the *Saccharomyces cerevisiae* species.

**[0040]** In one aspect of the recombinant hosts and methods disclosed herein,

- (a) the steviol glycoside comprises 13-SMG, 19-SMG, Steviol-1,2-bioside, Steviol-1,3-bioside, 1,2-stevioside, 1,3-stevioside, rubusoside, RebA, RebB, RebD,

RebE, RebM, di-glycosylated tri-glycosylated steviol, tetra-glycosylated steviol, penta-glycosylated steviol, hexa-glycosylated steviol, hepta-glycosylated steviol, and/or isomers thereof;

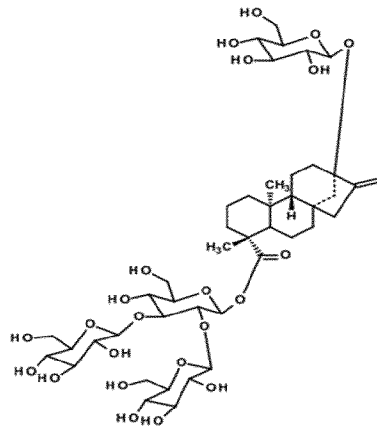
- (b) the glycosylated ent-kaurenol compound comprises di-glycosylated ent-kaurenol, tri-glycosylated ent-kaurenol, and/or isomers thereof; and/or
- (c) the glycosylated ent-kaurenoic acid compound comprises di-glycosylated ent-kaurenoic acid, tri-glycosylated ent-kaurenoic acid, and/or isomers thereof.

**[0041]** In one aspect of the recombinant hosts and methods disclosed herein,

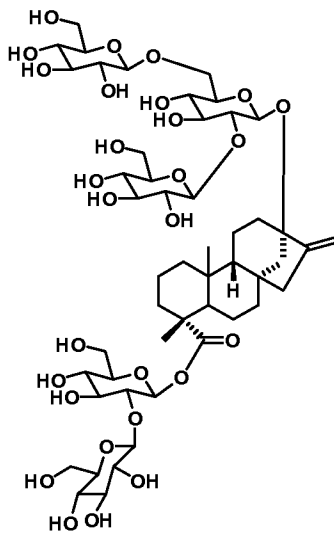
- (a) the di-glycosylated steviol comprises compound 2.23 of Table 1;
- (b) the tri-glycosylated steviol comprises compound 3.1 and/or compound 3.34 of Table 1;
- (c) the tetra-glycosylated steviol comprises compound 4.26 and/or compound 4.33 of Table 1;
- (d) the penta-glycosylated steviol comprises compound 5.22, compound 5.24, and/or compound 5.25 of Table 1;
- (e) the hexa-glycosylated steviol comprises compound 6.1 and/or compound 6.23 of Table 1;
- (f) the hepta-glycosylated steviol comprises compound 7.2, compound 7.5, and/or compound 7.13 of Table 1;
- (g) the glycosylated ent-kaurenoic acid compound comprises compound KA3.1, compound KA3.2, and/or compound KA2.7 of Table 1; and/or
- (h) the glycosylated ent-kaurenol compound comprises compound KL2.8 and/or compound KL3.1 co-eluted with compound KL3.6 of Table 1.

**[0042]** In one aspect of the recombinant hosts and methods disclosed herein,

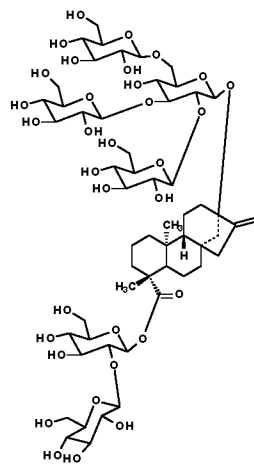
- (a) compound 4.26 has the structure:



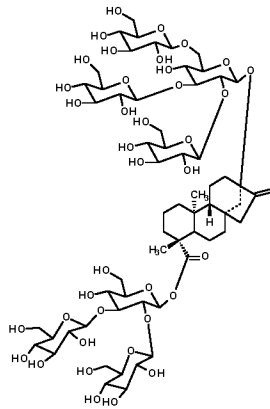
(b) compound 5.22 has the structure:



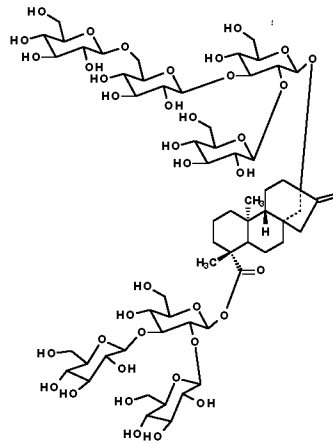
(c) compound 6.1 has the structure:



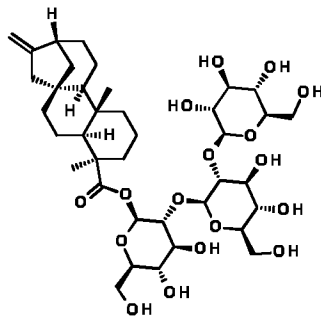
(d) compound 7.2 has the structure:



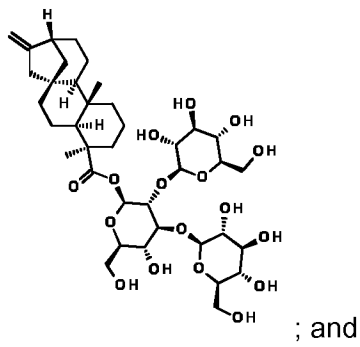
(e) compound 7.5 has the structure:



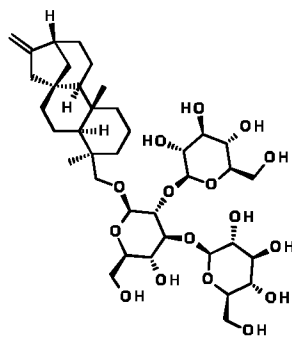
(f) compound KA3.1 has the structure:



(g) compound KA3.2 has the structure:

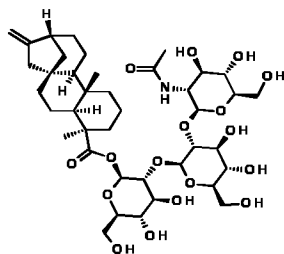


(h) compound KL3.1 has the structure:

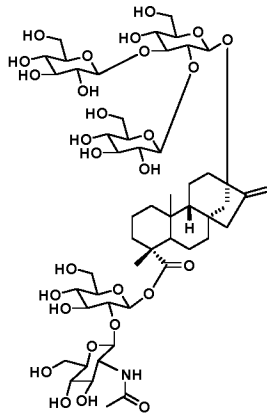


**[0043]** In one aspect of the recombinant hosts and methods disclosed herein,

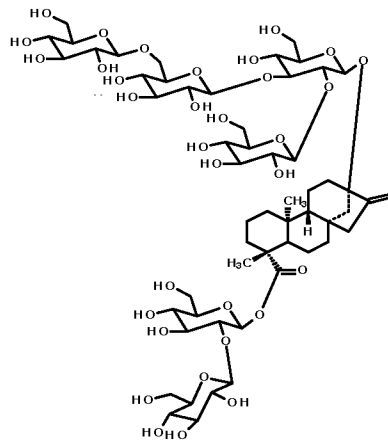
(a) the tri-glycosylated ent-kaurenoic acid comprises a compound having the structure:



(b) the penta-glycosylated steviol comprises a compound having the structure:

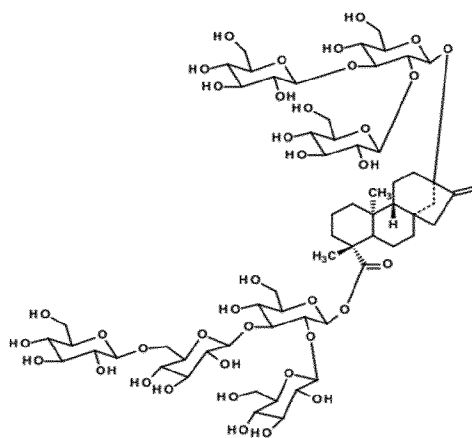


(c) the hexa-glycosylated steviol comprises a compound having the structure:



; and

(d) the hepta-glycosylated steviol comprises a compound having the structure:



**[0044]** The invention also provides a steviol glycoside composition produced by the recombinant host cell disclosed herein or the method disclosed herein, wherein the composition has a steviol glycoside composition enriched for RebD, RebM, or isomers thereof relative to a steviol glycoside composition of Stevia plant and has a reduced level of non-steviol glycoside Stevia plant-derived components relative to a plant-derived stevia extract.

**[0045]** The invention also provides a cell culture broth comprising:

- (a) the recombinant host cell disclosed herein; and
- (b) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or glycosylated ent-kaurenoic acid compounds produced by the recombinant host cell;

wherein one or more steviol glycosides is present at a concentration of at least 1 mg/liter of the culture broth.

**[0046]** The invention also provides a cell culture broth comprising:

- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or glycosylated ent-kaurenoic acid compounds produced by the recombinant host cell disclosed herein,
- (b) glucose, fructose, sucrose, xylose, ethanol, and/or glycerol; and/or
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids.

**[0047]** The invention also provides a cell lysate comprising:

- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or glycosylated ent-kaurenoic acid compounds produced by the recombinant host cell disclosed herein,
- (b) glucose, fructose, sucrose, xylose, ethanol, glycerol, uridine diphosphate (UDP)-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and/or
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids.

**[0048]** The invention also provides a reaction mixture comprising:

- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or a glycosylated ent-kaurenoic acid compounds produced in the reaction mixture;

- (b) a UGT polypeptide;
- (c) glucose, fructose, sucrose, xylose, ethanol, glycerol, uridine diphosphate (UDP)-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and/or
- (d) reaction buffer and/or salts.

**[0049]** These and other features and advantages of the present invention will be more fully understood from the following detailed description taken together with the accompanying claims. It is noted that the scope of the claims is defined by the recitations therein and not by the specific discussion of features and advantages set forth in the present description.

### BRIEF DESCRIPTION OF THE DRAWINGS

**[0050]** The following detailed description of the embodiments of the present invention can be best understood when read in conjunction with the following drawings, where like structure is indicated with like reference numerals and in which:

**[0051]** Figure 1 shows a schematic of the engineered biosynthetic pathway for producing steviol in yeast from geranylgeranyl diphosphate using geranylgeranyl diphosphate synthase (GGPPS), ent-copalyl diphosphate synthase (CDPS), ent-kaurene synthase (KS), ent-kaurene oxidase (KO), and ent-kaurenoic acid hydroxylase (KAH) polypeptides.

**[0052]** Figure 2 shows representative steviol glycoside glycosylation reactions catalyzed by suitable uridine 5'-diphospho (UDP) glycosyl transferases (UGT) enzymes and chemical structures for several steviol glycoside compounds.

**[0053]** Figure 3 shows the steviol synthetic intermediate, ent-kaurenol, and its bioconversion product, ent-kaurenoic acid, for the steviol pathway step catalyzed by a KO, along with potential glycosylation by-products (mono-, di-, and/or tri-glycosylated ent-kaurenol and mono-, di-, or tri-glycosylated ent-kaurenoic acid).

**[0054]** Figure 4A shows accumulation of ent-kaurenoic acid+2Glc (#7), ent-kaurenoic acid+3Glc (isomer 1), and ent-kaurenoic acid+3Glc (isomer 2) by a steviol glycoside-producing *S. cerevisiae* strain deleted of UGT85C2 (SEQ ID NO:7). Figure 4B shows accumulation of 19-SMG by a steviol glycoside-producing *S. cerevisiae* strain deleted of UGT85C2 (SEQ ID NO:7). Figure 4C shows accumulation of steviol, steviol+2Glc (#23), and steviol+3Glc (#34) by a steviol glycoside-producing *S. cerevisiae* strain deleted of UGT85C2 (SEQ ID NO:7). See Example 6.

**[0055]** Figure 5 shows conversion of steviol to rubusoside by bacterial lysates comprising UGT85C2 variants. Bacterial lysates were incubated with steviol for 24 h. See Example 7.

**[0056]** Figure 6A shows production of RebM, RebD, RebA, RebB, 13-SMG, and rubusoside in a steviol glycoside-producing strain expressing UGT76G1 H155L (gray bars), compared to the control steviol glycoside-producing strain expressing wild-type UGT76G1 (black bars). Figure 6B shows production of 1,2-bioside, rubusoside (Rubu), RebG, and RebE in a steviol glycoside-producing strain expressing UGT76G1 H155L (gray bars), compared to a control strain expressing wild-type UGT76G1 (black bars). Figure 6C shows production of quantifiable steviol glycosides (13-SMG + 1,2-bioside + Rubu + RebG + RebB + RebA + RebE + RebD + RebM) and RebD plus RebM titers in a steviol glycoside-producing strain expressing UGT76G1 H155L (gray bars), compared to a control strain expressing wild-type UGT76G1 (black bars). Figure 6D shows production of a tri-glycosylated steviol molecule (steviol+3Glc (#1)), a tetra-glycosylated steviol molecule (steviol+4Glc (#26)), three penta-glycosylated steviol molecules (steviol+5Glc (#22), steviol+5Glc (#24), and steviol+5Glc (#25)), two hexa-glycosylated steviol molecules (steviol+6Glc (isomer 1) and steviol+6Glc (#23)), and two hepta-glycosylated steviol molecules (steviol+7Glc (isomer 2) and steviol+7Glc (#13)) in a steviol glycoside-producing strain expressing UGT76G1 H155L (gray bars), compared to a control strain expressing wild-type UGT76G1 (black bars). See Example 9.

**[0057]** Figure 7A shows NMR-elucidated structures of tri-glycosylated ent-kaurenoic acid (Ent-Kaurenoic Acid+3Glc (isomers 1 and 2)), ent-kaurenoic acid+2Glc+1GlcNAc, and tri-glycosylated ent-kaurenol (ent-kaurenol+3Glc (isomer 1)). Figure 7B shows NMR-elucidated structures of steviol+6Glc (isomer 1) and steviol+7Glc (isomer 2). Figure 7C shows NMR-elucidated structures of steviol+6Glc (isomer 4) and steviol+7Glc (isomer 5). Figure 7D shows NMR-elucidated structures of steviol+4Glc+1GlcNAc (#11) and steviol+4Glc (#26). Figure 7E shows NMR-elucidated structures of steviol+5Glc (#22) and steviol+7Glc (#14). See Examples 6, 8, and 9.

**[0058]** Figures 8A, 8B, and 8C show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for ent-kaurenoic acid+3Glc (isomer 1). Figures 8D, 8E, and 8F show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for ent-kaurenoic acid+3Glc (isomer 2). Figures 8G, 8H, and 8I show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for ent-kaurenoic acid+2Glc+1GlcNAc. Figures 8J, 8K, and 8L show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for ent-kaurenol+3Glc (isomer 1). Figures 8M, 8N, 8O, and 8P show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for steviol+6Glc

(isomer 1). Figures 8Q, 8R, 8S, and 8T show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for steviol+7Glc (isomer 2). Figures 8U, 8V, 8W, and 8X show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for steviol+6Glc (isomer 4). Figures 8Y, 8Z, 8AA, and 8AB show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for steviol+7Glc (isomer 5). Figures 8AC, 8AD, 8AE, and 8AF show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for steviol+4Glc+1GlcNAc (#11). Figures 8AG, 8AH, 8AI, and 8AJ show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for steviol+4Glc (#26). Figures 8AK, 8AL, 8AM, and 8AN show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for steviol+5Glc (#22). Figures 8AO, 8AP, 8AQ, and 8AR show a  $^1\text{H}$  NMR spectrum and  $^1\text{H}$  and  $^{13}\text{C}$  NMR chemical shifts (in ppm) for steviol+7Glc (#14). See Examples 6, 8, and 9.

**[0059]** Figure 9A shows accumulation of ent-kaurenoic acid+2Glc (#7), ent-kaurenoic acid+3Glc (isomer 1), and ent-kaurenoic acid+3Glc (isomer 2) in *S. cerevisiae* expressing UGT76G1 variants. Figure 9B shows accumulation of ent-kaurenol+2Glc (#8) and ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6) in *S. cerevisiae* expressing UGT76G1 variants. See Example 8.

**[0060]** Figure 10A shows accumulation of 1,2-stevioside, RebG, steviol+3Glc (#1), steviol+4Glc (#26), steviol+5Glc (#22), steviol+5Glc (#24), steviol+5Glc (#25), steviol+6Glc (isomer 1), and steviol+6Glc (#23) in *S. cerevisiae* expressing RebD-producing UGT76G1 variants. Figure 10B shows accumulation of 1,2-stevioside, RebG, steviol+3Glc (#1), steviol+4Glc (#26), steviol+5Glc (#22), steviol+5Glc (#24), steviol+5Glc (#25), steviol+6Glc (isomer 1), and steviol+6Glc (#23) in *S. cerevisiae* expressing RebM-producing UGT76G1 variants. Figure 10C shows accumulation of 13-SMG, 1,2-bioside, rubusoside, RebA, RebB, RebD, RebE, and RebM in *S. cerevisiae* expressing UGT76G1 variants. See Example 8.

**[0061]** Figure 11A shows accumulation of ent-kaurenoic acid+2Glc (#7), ent-kaurenoic acid+3Glc (isomer 1), ent-kaurenoic acid+3Glc (isomer 2), ent-kaurenol+2Glc (#8), and ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6) in an *S. cerevisiae* steviol glycoside production strain (control strain comprised three copies of wild-type UGT76G1 (SEQ ID NO:9); variant strains comprised two copies of wild-type UGT76G1 and one copy of a UGT76G1 variant). Figure 11B shows total levels of glycosylated ent-kaurenoic acid (ent-kaurenoic acid+2Glc (#7) + ent-kaurenoic acid+3Glc (isomer 1) + ent-kaurenoic acid+3Glc (isomer 2)) in an *S. cerevisiae* steviol glycoside production strain expressing UGT76G1 variants. Figure 11C shows total levels of glycosylated ent-kaurenol (ent-kaurenol+3Glc (isomer 1) co-

eluted with ent-kaurenol+3Glc (#6) and ent-kaurenol+2Glc (#8) in an *S. cerevisiae* steviol glycoside production strain expressing UGT76G1 variants. Figure 11D shows accumulation of 1,2-bioside, 1,2-stevioside, steviol+3Glc (#1), steviol+4Glc (#26), steviol+5Glc (#22), steviol+5Glc (#24), steviol+5Glc (#25), steviol+6Glc (isomer 1), steviol+6Glc (#23), steviol+7Glc (isomer 2), and steviol+7Glc (isomer 5) in an *S. cerevisiae* steviol glycoside production strain expressing UGT76G1 variants. Figure 11E shows accumulation of 13-SMG, 1,2-bioside, rubusoside, RebG, RebA, RebB, RebD, RebE, and RebM in an *S. cerevisiae* steviol glycoside production strain expressing UGT76G1 variants. See Example 8.

**[0062]** Skilled artisans will appreciate that elements in the Figures are illustrated for simplicity and clarity and have not necessarily been drawn to scale. For example, the dimensions of some of the elements in the Figures can be exaggerated relative to other elements to help improve understanding of the embodiment(s) of the present invention.

#### DETAILED DESCRIPTION OF THE INVENTION

**[0063]** Before describing the present invention in detail, a number of terms will be defined. As used herein, the singular forms “a,” “an,” and “the” include plural referents unless the context clearly dictates otherwise. For example, reference to a “nucleic acid” means one or more nucleic acids.

**[0064]** It is noted that terms like “preferably,” “commonly,” and “typically” are not utilized herein to limit the scope of the claimed invention or to imply that certain features are critical, essential, or even important to the structure or function of the claimed invention. Rather, these terms are merely intended to highlight alternative or additional features that can or cannot be utilized in a particular embodiment of the present invention.

**[0065]** For the purposes of describing and defining the present invention it is noted that the term “substantially” is utilized herein to represent the inherent degree of uncertainty that can be attributed to any quantitative comparison, value, measurement, or other representation. The term “substantially” is also utilized herein to represent the degree by which a quantitative representation can vary from a stated reference without resulting in a change in the basic function of the subject matter at issue.

**[0066]** Methods well known to those skilled in the art can be used to construct genetic expression constructs and recombinant cells according to this invention. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, *in vivo* recombination

techniques, and polymerase chain reaction (PCR) techniques. See, for example, techniques as described in Green & Sambrook, 2012, MOLECULAR CLONING: A LABORATORY MANUAL, Fourth Edition, Cold Spring Harbor Laboratory, New York; Ausubel *et al.*, 1989, CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, Greene Publishing Associates and Wiley Interscience, New York, and PCR Protocols: A Guide to Methods and Applications (Innis *et al.*, 1990, Academic Press, San Diego, CA).

**[0067]** As used herein, the terms “polynucleotide,” “nucleotide,” “oligonucleotide,” and “nucleic acid” can be used interchangeably to refer to nucleic acid comprising DNA, RNA, derivatives thereof, or combinations thereof, in either single-stranded or double-stranded embodiments depending on context as understood by the skilled worker.

**[0068]** As used herein, the terms “microorganism,” “microorganism host,” “microorganism host cell,” “recombinant host,” and “recombinant host cell” can be used interchangeably. As used herein, the term “recombinant host” is intended to refer to a host, the genome of which has been augmented by at least one DNA sequence. Such DNA sequences include but are not limited to genes that are not naturally present, DNA sequences that are not normally transcribed into RNA or translated into a protein (“expressed”), and other genes or DNA sequences which one desires to introduce into a host. It will be appreciated that typically the genome of a recombinant host described herein is augmented through stable introduction of one or more recombinant genes. Generally, introduced DNA is not originally resident in the host that is the recipient of the DNA, but it is within the scope of this disclosure to isolate a DNA segment from a given host, and to subsequently introduce one or more additional copies of that DNA into the same host, *e.g.*, to enhance production of the product of a gene or alter the expression pattern of a gene. In some instances, the introduced DNA will modify or even replace an endogenous gene or DNA sequence by, *e.g.*, homologous recombination or site-directed mutagenesis. Suitable recombinant hosts include microorganisms.

**[0069]** As used herein, the term “recombinant gene” refers to a gene or DNA sequence that is introduced into a recipient host, regardless of whether the same or a similar gene or DNA sequence may already be present in such a host. “Introduced,” or “augmented” in this context, is known in the art to mean introduced or augmented by the hand of man. Thus, a recombinant gene can be a DNA sequence from another species or can be a DNA sequence that originated from or is present in the same species but has been incorporated into a host by recombinant methods to form a recombinant host. It will be appreciated that a recombinant gene that is introduced into a host can be identical to a DNA sequence that is normally present in the host

being transformed, and is introduced to provide one or more additional copies of the DNA to thereby permit overexpression or modified expression of the gene product of that DNA. In some aspects, said recombinant genes are encoded by cDNA. In other embodiments, recombinant genes are synthetic and/or codon-optimized for expression in *S. cerevisiae*.

**[0070]** As used herein, the term “engineered biosynthetic pathway” refers to a biosynthetic pathway that occurs in a recombinant host, as described herein. In some aspects, one or more steps of the biosynthetic pathway do not naturally occur in an unmodified host. In some embodiments, a heterologous version of a gene is introduced into a host that comprises an endogenous version of the gene.

**[0071]** As used herein, the term “endogenous” gene refers to a gene that originates from and is produced or synthesized within a particular organism, tissue, or cell. In some embodiments, the endogenous gene is a yeast gene. In some embodiments, the gene is endogenous to *S. cerevisiae*, including, but not limited to *S. cerevisiae* strain S288C. In some embodiments, an endogenous yeast gene is overexpressed. As used herein, the term “overexpress” is used to refer to the expression of a gene in an organism at levels higher than the level of gene expression in a wild type organism. See, e.g., Prelich, 2012, *Genetics* 190:841-54. In some embodiments, an endogenous yeast gene, for example ADH, is deleted. See, e.g., Giaever & Nislow, 2014, *Genetics* 197(2):451-65. As used herein, the terms “deletion,” “deleted,” “knockout,” and “knocked out” can be used interchangeably to refer to an endogenous gene that has been manipulated to no longer be expressed in an organism, including, but not limited to, *S. cerevisiae*.

**[0072]** As used herein, the terms “heterologous sequence” and “heterologous coding sequence” are used to describe a sequence derived from a species other than the recombinant host. In some embodiments, the recombinant host is an *S. cerevisiae* cell, and a heterologous sequence is derived from an organism other than *S. cerevisiae*. A heterologous coding sequence, for example, can be from a prokaryotic microorganism, a eukaryotic microorganism, a plant, an animal, an insect, or a fungus different than the recombinant host expressing the heterologous sequence. In some embodiments, a coding sequence is a sequence that is native to the host.

**[0073]** A “selectable marker” can be one of any number of genes that complement host cell auxotrophy, provide antibiotic resistance, or result in a color change. Linearized DNA fragments of the gene replacement vector then are introduced into the cells using methods well known in the art (see below). Integration of the linear fragments into the genome and the disruption of the

gene can be determined based on the selection marker and can be verified by, for example, PCR or Southern blot analysis. Subsequent to its use in selection, a selectable marker can be removed from the genome of the host cell by, e.g., Cre-LoxP systems (see, e.g., Gossen *et al.*, 2002, *Ann. Rev. Genetics* 36:153-173 and U.S. 2006/0014264). Alternatively, a gene replacement vector can be constructed in such a way as to include a portion of the gene to be disrupted, where the portion is devoid of any endogenous gene promoter sequence and encodes none, or an inactive fragment of, the coding sequence of the gene.

**[0074]** As used herein, the terms “variant” and “mutant” are used to describe a protein sequence that has been modified at one or more amino acids, compared to the wild-type sequence of a particular protein.

**[0075]** As used herein, the term “inactive fragment” is a fragment of the gene that encodes a protein having, e.g., less than about 10% (e.g., less than about 9%, less than about 8%, less than about 7%, less than about 6%, less than about 5%, less than about 4%, less than about 3%, less than about 2%, less than about 1%, or 0%) of the activity of the protein produced from the full-length coding sequence of the gene. Such a portion of a gene is inserted in a vector in such a way that no known promoter sequence is operably linked to the gene sequence, but that a stop codon and a transcription termination sequence are operably linked to the portion of the gene sequence. This vector can be subsequently linearized in the portion of the gene sequence and transformed into a cell. By way of single homologous recombination, this linearized vector is then integrated in the endogenous counterpart of the gene with inactivation thereof.

**[0076]** As used herein, the term “steviol glycoside” refers to rebaudioside A (RebA) (CAS # 58543-16-1), rebaudioside B (RebB) (CAS # 58543-17-2), rebaudioside C (RebC) (CAS # 63550-99-2), rebaudioside D (RebD) (CAS # 63279-13-0), rebaudioside E (RebE) (CAS # 63279-14-1), rebaudioside F (RebF) (CAS # 438045-89-7), rebaudioside M (RebM) (CAS # 1220616-44-3), rubusoside (CAS # 63849-39-4), dulcoside A (CAS # 64432-06-0), rebaudioside I (RebI) (MassBank Record: FU000332), rebaudioside Q (RebQ), 1,2-stevioside (CAS # 57817-89-7), 1,3-stevioside (RebG), 1,2-bioside (MassBank Record: FU000299), 1,3-bioside, steviol-13-O-glucoside (13-SMG), steviol-19-O-glucoside (19-SMG), a di-glycosylated steviol, a tri-glycosylated steviol, a tetra-glycosylated steviol, a penta-glycosylated steviol, a hexa-glycosylated steviol, a hepta-glycosylated steviol, and/or isomers thereof. See Figure 2; see also, Steviol Glycosides Chemical and Technical Assessment 69th JECFA, 2007, prepared by Harriet Wallin, Food Agric. Org. See Figure 2, Figure 7, Figure 8, and Table 1; see also, Steviol Glycosides Chemical and Technical Assessment 69th JECFA, 2007, prepared by Harriet Wallin,

Food Agric. Org. Glycosylated steviol compounds can comprise one or more glucose, N-acetylglucosamine (GlcNAc), rhamnose, and/or xylose moieties. Non-limiting examples of steviol glycosides that can be produced by methods described herein are shown in Table 1, Figure 7, and Figure 8.

**[0077]** As used herein, the term “glycosylated ent-kaurenol compound” refers to di-glycosylated ent-kaurenol or tri-glycosylated ent-kaurenol. As used herein, the term “glycosylated ent-kaurenoic acid compound” refers to di-glycosylated ent-kaurenoic acid or tri-glycosylated ent-kaurenoic acid. See Figure 7, Figure 8, and Table 1. Glycosylated ent-kaurenol compounds and glycosylated ent-kaurenoic acid compounds can comprise one or more glucose, GlcNAc, rhamnose, and/or xylose moieties. Non-limiting examples of glycosylated ent-kaurenol compounds and glycosylated ent-kaurenoic acid compounds that can be produced by methods described herein are shown in Table 1, Figure 7, and Figure 8.

**[0078]** As used herein, the terms “steviol glycoside precursor” and “steviol glycoside precursor compound” are used to refer to intermediate compounds in the steviol glycoside biosynthetic pathway. Steviol glycoside precursors include, but are not limited to, geranylgeranyl diphosphate (GGPP), ent-copalyl-diphosphate, ent-kaurene, ent-kaurenol, ent-kaurenal, ent-kaurenoic acid, and steviol. See Figure 1. In some embodiments, steviol glycoside precursors are themselves steviol glycoside compounds. For example, 19-SMG, rubusoside, stevioside, and RebE are steviol glycoside precursors of RebM. See Figure 2. Steviol glycosides and/or steviol glycoside precursors can be produced *in vivo* (*i.e.*, in a recombinant host), *in vitro* (*i.e.*, enzymatically), or by whole cell bioconversion. As used herein, the terms “produce” and “accumulate” can be used interchangeably to describe synthesis of steviol glycosides and steviol glycoside precursors *in vivo*, *in vitro*, or by whole cell bioconversion.

**[0079]** As used herein, the term “cell culture broth” can be used to refer to a liquid that can support or has supported growth of a host cell, including, but not limited to, a yeast host cell. The components of a cell culture broth can include, for example, a steviol glycoside, a glycosylated ent-kaurenol compound, and/or a glycosylated ent-kaurenoic acid compound produced by the host cell, glucose, fructose, sucrose, trace metals, vitamins, salts, yeast nitrogen base (YNB), and/or amino acids.

**[0080]** As used herein, the term “cell lysate” can be used to refer to a fluid comprising the components of a lysed cell, *i.e.*, a cell whose membrane has been disrupted chemically or mechanically. A cell lysate can further comprise a steviol glycoside, a glycosylated ent-kaurenol

compound, and/or a glycosylated ent-kaurenoic acid compound produced by the host cell, glucose, fructose, sucrose, xylose, rhamnose, uridine diphosphate (UDP)-glucose, UDP-rhamnose, UDP-xylose, GlcNAc, trace metals, vitamins, salts, YNB, and/or amino acids. In some aspects, a cell lysate is a yeast cell lysate, such as an *S. cerevisiae* cell lysate, or a bacterial cell lysate, such as an *E. coli* cell lysate.

**[0081]** As used herein, the term “reaction mixture” refers to a solution for conducting an *in vitro* reaction. The components of a reaction mixture can include, but are not limited to, a steviol glycoside, a glycosylated ent-kaurenol compound, a glycosylated ent-kaurenoic acid compound, a polypeptide such as a UGT polypeptide, UDP-glucose, UDP-rhamnose, UDP-xylose, GlcNAc, a buffer, and/or salts.

**[0082]** Recombinant steviol glycoside-producing *Saccharomyces cerevisiae* (*S. cerevisiae*) strains are described in WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328. Methods of producing steviol glycosides in recombinant hosts, by whole cell bio-conversion, and *in vitro* are also described in WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328.

**[0083]** In some embodiments, steviol glycosides and/or steviol glycoside precursors are produced *in vivo* through expression of one or more enzymes involved in the steviol glycoside biosynthetic pathway in a recombinant host. For example, a steviol-producing recombinant host expressing one or more of a gene encoding a GGPPS polypeptide, a gene encoding a CDPS polypeptide, a gene encoding a KS polypeptide, a gene encoding a KO polypeptide, a gene encoding a KAH polypeptide, a gene encoding a CPR polypeptide, and a gene encoding a UGT polypeptide can produce a steviol glycoside and/or steviol glycoside precursors *in vivo*. See, e.g., Figures 1 and 2. The skilled worker will appreciate that one or more of these genes can be endogenous to the host provided that at least one (and in some embodiments, all) of these genes is a recombinant gene introduced into the recombinant host.

**[0084]** A recombinant host described herein can comprise a gene encoding a polypeptide capable of synthesizing geranylgeranyl pyrophosphate (GGPP) from farnesyl diphosphate (FPP) and isopentenyl diphosphate (IPP), a gene encoding a polypeptide capable of synthesizing ent-copalyl diphosphate from GGPP; a gene encoding a polypeptide capable of synthesizing ent-kaurene from ent-copalyl pyrophosphate, a gene encoding a polypeptide capable of synthesizing ent-kaurenoic acid from ent-kaurene, a gene encoding a polypeptide capable of synthesizing steviol from ent-kaurenoic acid; and/or a gene encoding a polypeptide capable of converting NADPH to NADP<sup>+</sup>. A GGPPS polypeptide can synthesize GGPP from

FPP and IPP. A CDPS polypeptide can synthesize ent-copalyl diphosphate from GGPP. A KS polypeptide can synthesize ent-kaurene from ent-copalyl pyrophosphate. A KO polypeptide can synthesize ent-kaurenoic acid from ent-kaurene. A KAH polypeptide can synthesize steviol from ent-kaurenoic acid. A CPR polypeptide can convert NADPH to NADP+.

**[0085]** In another example, a recombinant host expressing a gene encoding a GGPPS polypeptide, a gene encoding a CDPS polypeptide, a gene encoding a KS polypeptide, a gene encoding a KO polypeptide, a gene encoding a KAH polypeptide, and a gene encoding a CPR polypeptide can produce steviol *in vivo*. See, e.g., Figure 1. The skilled worker will appreciate that one or more of these genes can be endogenous to the host provided that at least one (and in some embodiments, all) of these genes is a recombinant gene introduced into the recombinant host.

**[0086]** In another example, a recombinant host expressing a gene encoding a GGPPS polypeptide, a gene encoding a CDPS polypeptide, a gene encoding a KS polypeptide, a gene encoding a KO polypeptide, a gene encoding a KAH polypeptide, a gene encoding a CPR polypeptide, and one or more of a gene encoding a UGT polypeptide can produce a steviol glycoside *in vivo*. See, e.g., Figures 1 and 2. The skilled worker will appreciate that one or more of these genes can be endogenous to the host provided that at least one (and in some embodiments, all) of these genes is a recombinant gene introduced into the recombinant host.

**[0087]** In some aspects, the GGPPS polypeptide comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:20 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:19), SEQ ID NO:22 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:21), SEQ ID NO:24 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:23), SEQ ID NO:26 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:25), SEQ ID NO:28 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:27), SEQ ID NO:30 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:29), SEQ ID NO:32 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:31), or SEQ ID NO:116 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:115).

**[0088]** In some aspects, the CDPS polypeptide comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:34 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:33), SEQ ID NO:36 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:35), SEQ ID NO:38 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:37), SEQ ID NO:40 (which can be encoded by the nucleotide sequence

set forth in SEQ ID NO:39), or SEQ ID NO:42 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:41). In some embodiments, the CDPS polypeptide lacks a chloroplast transit peptide.

**[0089]** In some aspects, the KS polypeptide comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:44 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:43), SEQ ID NO:46 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:45), SEQ ID NO:48 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:47), SEQ ID NO:50 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:49), or SEQ ID NO:52 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:51).

**[0090]** In some embodiments, a recombinant host comprises a gene encoding a CDPS-KS polypeptide. In some aspects, the CDPS-KS polypeptide comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:54 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:53), SEQ ID NO:56 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:55), or SEQ ID NO:58 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:57).

**[0091]** In some aspects, the KO polypeptide comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:60 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:59), SEQ ID NO:62 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:61), SEQ ID NO:117 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:63 or SEQ ID NO:64), SEQ ID NO:66 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:65), SEQ ID NO:68 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:67), SEQ ID NO:70 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:69), SEQ ID NO:72 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:71), SEQ ID NO:74 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:73), or SEQ ID NO:76 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:75).

**[0092]** In some aspects, the CPR polypeptide comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:78 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:77), SEQ ID NO:80 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:79), SEQ ID NO:82 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:81), SEQ ID NO:84 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:83), SEQ ID NO:86 (which can be encoded by the nucleotide sequence

set forth in SEQ ID NO:85), SEQ ID NO:88 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:87), SEQ ID NO:90 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:89), or SEQ ID NO:92 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:91).

**[0093]** In some aspects, the KAH polypeptide comprises a polypeptide having an amino acid sequence set forth in SEQ ID NO:94 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:93), SEQ ID NO:97 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:95 or SEQ ID NO:96), SEQ ID NO:100 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:98 or SEQ ID NO:99), SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:106 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:105), SEQ ID NO:108 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:107), SEQ ID NO:110 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:109), SEQ ID NO:112 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:111), or SEQ ID NO:114 (which can be encoded by the nucleotide sequence set forth in SEQ ID NO:113).

**[0094]** In some embodiments, a recombinant host comprises a nucleic acid encoding a UGT85C2 polypeptide (SEQ ID NO:7), a nucleic acid encoding a UGT76G1 polypeptide (SEQ ID NO:9), a nucleic acid encoding a UGT74G1 polypeptide (SEQ ID NO:4), a nucleic acid encoding a UGT91D2 polypeptide, and/or a nucleic acid encoding a EUGT11 polypeptide (SEQ ID NO:16). In some aspects, the UGT91D2 polypeptide can be a UGT91D2e polypeptide (SEQ ID NO:11) or a UGT91D2e-b polypeptide (SEQ ID NO:13). In some aspects, the UGT85C2 polypeptide can be encoded by the nucleotide sequence set forth in SEQ ID NO:5 or SEQ ID NO:6, the UGT76G1 polypeptide can be encoded by the nucleotide sequence set forth in SEQ ID NO:8, the UGT74G1 polypeptide can be encoded by the nucleotide sequence set forth in SEQ ID NO:3, the UGT91D2e polypeptide can be encoded by the nucleotide sequence set forth in SEQ ID NO:10, the UGT91D2e-b polypeptide can be encoded by the nucleotide sequence set forth in SEQ ID NO:12, and the EUGT11 polypeptide can be encoded by the nucleotide sequence set forth in SEQ ID NO:14 or SEQ ID NO:15. The skilled worker will appreciate that expression of these genes may be necessary to produce a particular steviol glycoside but that one or more of these genes can be endogenous to the host provided that at least one (and in some embodiments, all) of these genes is a recombinant gene introduced into the recombinant host. In a particular embodiment, a steviol-producing recombinant microorganism comprises exogenous nucleic acids encoding UGT85C2, UGT76G1, or UGT91D2 polypeptides.

**[0095]** In another particular embodiment, a steviol-producing recombinant microorganism comprises exogenous nucleic acids encoding UGT85C2, UGT76G1, UGT74G1, and UGT91D2 polypeptides. In yet another particular embodiment, a steviol-producing recombinant microorganism comprises exogenous nucleic acids encoding UGT85C2, UGT76G1, UGT74G1, and EUGT11 polypeptides. In yet another particular embodiment, a steviol-producing recombinant microorganism comprises exogenous nucleic acids encoding UGT85C2, UGT76G1, UGT74G1, UGT91D2 (including *inter alia* UGT91D2e, UGT91D2m, UGT91D2e-b, and functional homologs thereof), and EUGT11 polypeptides. In yet another particular embodiment, a steviol-producing recombinant microorganism comprises exogenous nucleic acids encoding UGT85C2, UGT76G1, UGT74G1, UGT91D2, and/or EUGT11 polypeptides. In yet another particular embodiment, a steviol-producing recombinant microorganism comprises exogenous nucleic acids encoding UGT85C2, UGT76G1, UGT74G1, UGT91D2, and/or EUGT11 polypeptides.

**[0096]** In some embodiments, a recombinant host comprises: (a) a gene encoding a polypeptide capable of beta 1,2 glucosylation of the C2' of the 19-O glucose of a steviol glycoside; (b) a gene encoding a polypeptide capable of beta 1,2 glucosylation of the C2' of the 13-O-glucose of a steviol glycoside; (c) a gene encoding a polypeptide capable of beta 1,3 glucosylation of the C3' of the 19-O-glucose of a steviol glycoside; (d) a gene encoding a polypeptide capable of beta 1,3 glucosylation of the C3' of the 13-O-glucose of a steviol glycoside; (e) a gene encoding a polypeptide capable of beta 1,6 glucosylation of the C6' of the 13-O-glucose of a steviol glycoside; (f) a gene encoding a polypeptide capable of beta 1,6 glucosylation of the C6' of the 1,3-glucose of a 13-O diglucoside moiety of a steviol glycoside; (g) a gene encoding a polypeptide capable of glucosylation of the 13-OH of steviol or a steviol glycoside; (h) a gene encoding a polypeptide capable of glucosylation of the C-19 carboxyl of steviol or a steviol glycoside; (i) a gene encoding a polypeptide capable of beta 1,2 rhamnosylation of the C2' of the 13-O-glucose of a steviol glycoside; (j) a gene encoding a polypeptide capable of beta 1,2 xylosylation of the C2' of the 13-O-glucose of a steviol glycoside; (k) a gene encoding a polypeptide capable of beta 1,2 GlcNAc transfer to the C2' of the 19-O glucose of a steviol glycoside; (l) a gene encoding a polypeptide capable of beta 1,3 GlcNAc transfer to the C2' of the 19-O glucose of a steviol glycoside; (m) a gene encoding a polypeptide capable of beta 1,3 GlcNAc transfer to the C2' of the 13-O-glucose of a steviol glycoside; (n) a gene encoding a polypeptide capable of GlcNAc transfer to the C-19 carboxyl of steviol or a steviol glycoside; (o) a gene encoding a polypeptide capable of glucosylation of the C-19 carboxyl of kaurenoic acid or kaurenol; (p) a gene encoding a polypeptide capable of

beta 1,2 glucosylation of the C2' of the 19-O glucose of a kaurenoic acid glycoside or kaurenol glycoside; (p) a gene encoding a polypeptide capable of a beta 1,2 glucosylation of a beta 1,2 diglucoside of kaurenoic acid; (q) a gene encoding a polypeptide capable of beta 1,2 GlcNAc transfer of a beta 1,2 diglucoside of kaurenoic acid; (r) a gene encoding a polypeptide capable of beta 1,3 glucosylation of the C3' of the 19-O-glucose of a kaurenoic acid glycoside or kaurenol glycoside; and/or (s) a gene encoding a polypeptide capable of beta 1,6 glucosylation of the C6' of the 1,3-glucose of a 19-O diglucoside moiety of a steviol glycoside.

**[0097]** In some aspects, EUGT11 (SEQ ID NO:14/SEQ ID NO:15, SEQ ID NO:16), UGT91D2e (SEQ ID NO:10, SEQ ID NO:11), UGT91D2e-b (SEQ ID NO:12, SEQ ID NO:13), a variant thereof, or a chimeric protein thereof catalyzes beta 1,2 glucosylation of the C2' of the 19-O glucose of a steviol glycoside. Exemplary UGT91D2e variant sequences are set forth in SEQ ID NOs:1, 2, 118-121, 123, and 191-214. In some aspects, UGT91D2e (SEQ ID NO:10, SEQ ID NO:11), UGT91D2e-b (SEQ ID NO:12, SEQ ID NO:13), a variant thereof, or a chimeric protein thereof catalyzes beta 1,2 glucosylation of the C2' of the 13-O-glucose of a steviol glycoside. Exemplary UGT91D2e variant sequences are set forth in SEQ ID NOs:1, 2, 118-121, 123, and 191-214. Exemplary UGT91D2e-EUGT11 chimeric protein sequences are set forth in SEQ ID NO:17 and SEQ ID NO:18. In some aspects, UGT76G1 (SEQ ID NO:8, SEQ ID NO:9), a variant thereof, or a chimeric protein thereof catalyzes beta 1,3 glucosylation of the C3' of the 19-O-glucose of a steviol glycoside and/or beta 1,3 glucosylation of the C3' of the 13-O-glucose of a steviol glycoside. Exemplary UGT76G1 variant sequences are set forth in SEQ ID NOs:181-190 and 217-220. In some aspects, UGT85C2 (SEQ ID NO:5/SEQ ID NO:6, SEQ ID NO:7), a variant thereof, or a chimeric protein thereof catalyzes glucosylation of the 13-OH of steviol or a steviol glycoside. Exemplary UGT85C2 variant sequences are set forth in SEQ ID NOs:127 and 147-180. In some aspects, UGT74G1 (SEQ ID NO:3, SEQ ID NO:4), a variant thereof, or a chimeric protein thereof catalyzes glucosylation of the C-19 carboxyl of steviol or a steviol glycoside. In some aspects, EUGT11 (SEQ ID NO:14/SEQ ID NO:15, SEQ ID NO:16), UGT91D2e (SEQ ID NO:10, SEQ ID NO:11), UGT74G1 (SEQ ID NO:3, SEQ ID NO:4), and/or UGT76G1 (SEQ ID NO:8, SEQ ID NO:9) can accept uridine diphosphate N-acetylglucosamine (UDP-Glc-NAc) as a substrate. In some aspects, UGT74G1 glycosylates ent-kaurenol and ent-kaurenoic acid; UGT76G1 and UGT91D2e subsequently add additional glucose or GlcNAc moieties by either a 1,3- or 1,2-linkage to form tri-glycosylated compounds. See Figures 3, 7 and 8.

**[0098]** In some embodiments, steviol glycosides and/or steviol glycoside precursors are produced through contact of a steviol glycoside precursor with one or more enzymes involved in the steviol glycoside pathway *in vitro*. For example, contacting steviol with a UGT polypeptide can result in production of a steviol glycoside *in vitro*. In some embodiments, a steviol glycoside precursor is produced through contact of an upstream steviol glycoside precursor with one or more enzymes involved in the steviol glycoside pathway *in vitro*. For example, contacting entkaurenoic acid with a KAH enzyme can result in production of steviol *in vitro*.

**[0099]** In some embodiments, a steviol glycoside or steviol glycoside precursor is produced by whole cell bioconversion. For whole cell bioconversion to occur, a host cell expressing one or more enzymes involved in the steviol glycoside pathway takes up and modifies a steviol glycoside precursor in the cell; following modification *in vivo*, a steviol glycoside remains in the cell and/or is excreted into the culture medium. For example, a host cell expressing a gene encoding a UGT polypeptide can take up steviol and glycosylate steviol in the cell; following glycosylation *in vivo*, a steviol glycoside can be excreted into the culture medium. In some embodiments, the cell is permeabilized to take up a substrate to be modified or to excrete a modified product.

**[00100]** In some embodiments, steviol, one or more steviol glycoside precursors, and/or one or more steviol glycosides are produced by co-culturing of two or more hosts. In some embodiments, one or more hosts, each expressing one or more enzymes involved in the steviol glycoside pathway, produce steviol, one or more steviol glycoside precursors, and/or one or more steviol glycosides. For example, a host comprising a GGPPS, a CDPS, a KO, a KS, a KAH, and/or a CPR and a host comprising one or more UGTs produce one or more steviol glycosides.

**[00101]** In some embodiments, polypeptides suitable for producing steviol glycosides, such as 1,2-stevioside and RebD, *in vitro*, in a recombinant host, or by whole cell bioconversion include functional homologs of UGT91D2e (SEQ ID NO:10, SEQ ID NO:11), including UGT91D2e-b (SEQ ID NO:12, SEQ ID NO:13); UGT91D2e V286C (SEQ ID NO:1); UGT91D2e G384W (SEQ ID NO:2); UGT91D2e L211M (SEQ ID NO:118); UGT91D2e L195G (SEQ ID NO:119); UGT91D2e V196P (SEQ ID NO:120); UGT91D2e L211H (SEQ ID NO:121); UGT91D2e L213E (SEQ ID NO:191); UGT91D2e S221Y (SEQ ID NO:192); UGT91D2e E438H (SEQ ID NO:193); UGT91D2e M152T (SEQ ID NO:194); UGT91D2e L211C (SEQ ID NO:195); UGT91D2e L195S (SEQ ID NO:196); UGT91D2e L195V (SEQ ID NO:197); UGT91D2e V286S (SEQ ID NO:198); UGT91D2e S221S (SEQ ID NO:199); UGT91D2e P93V M152G (SEQ ID

NO:200); UGT91D2e S99I (SEQ ID NO:201); UGT91D2e T144K P201P (SEQ ID NO:202); UGT91D2e T144L (SEQ ID NO:203); UGT91D2e T144M (SEQ ID NO:204); UGT91D2e A148K L211I (SEQ ID NO:205); UGT91D2e L195N (SEQ ID NO:206); UGT91D2e K199C (SEQ ID NO:207); UGT91D2e L211M E426G A466V (SEQ ID NO:208); UGT91D2e L211T I303I (SEQ ID NO:209); UGT91D2e V286N (SEQ ID NO:210); UGT91D2e S114F V286S (SEQ ID NO:211); UGT91D2e G384K (SEQ ID NO:212); UGT91D2e G384Y (SEQ ID NO:213); UGT91D2e E438M (SEQ ID NO:214); and UGT91D2e L195C (SEQ ID NO:123). See Example 3.

**[00102]** In some embodiments, a useful UGT91D2 homolog can have one or more amino acid substitutions at residues 93, 99, 114, 144, 148, 152, 195, 196, 199, 211, 213, 221, 286, and 384. See Table 2. Non-limiting examples of useful UGT91D2e homologs include polypeptides having substitutions (with respect to SEQ ID NO:11) at residue 93 (*e.g.*, a valine at residue 93); 99 (*e.g.*, an isoleucine at residue 99), 114 (*e.g.*, a phenylalanine at residue 114); 144 (*e.g.*, a lysine, leucine, or methionine at residue 144); 148 (*e.g.*, a lysine at residue 148); 152 (*e.g.*, a threonine at residue 152); 195 (*e.g.*, a glycine, cysteine, serine, arginine, or valine at residue 195); 196 (*e.g.*, a proline at residue 196); 199 (*e.g.*, a cysteine at residue 199); 211 (*e.g.*, a methionine, histidine, threonine, cysteine, or isoleucine at residue 211); 213 (*e.g.*, a glutamic acid at 213); 221 (*e.g.*, an isoleucine at residue 221); 286 (*e.g.*, an alanine, cysteine, asparagine, or serine at residue 286); 384 (*e.g.*, a tryptophan, lysine, or tyrosine at residue 384); 426 (*e.g.*, a glycine at residue 426); 438 (*e.g.*, a histidine or methionine at residue 438); or 466 (*e.g.*, a valine at residue 466). See Example 3.

**[00103]** In some embodiments, UGT91D2e variants comprise silent mutations. For example, in some embodiments, UGT91D2e variants comprise silent mutations at residues not limited to residue 130, residue 201, or residue 221. See Example 3.

**[00104]** In some embodiments, UGT91D2e variants not limited to UGT91D2e V286C (SEQ ID NO:1), UGT91D2e G384W (SEQ ID NO:2), UGT91D2e L195V (SEQ ID NO:197), UGT91D2e V286S (SEQ ID NO:198), UGT91D2e T144K P201P (SEQ ID NO:202), UGT91D2e L211T I130I (SEQ ID NO:184), UGT91D2e S11F V286S (SEQ ID NO:211), and UGT91D2e E438M (SEQ ID NO:214) are selective towards rubusoside, with preferential accumulation of 1,2-stevioside. In some embodiments, UGT91D2e variants not limited to UGTD1D2e P93V M152G (SEQ ID NO:200), UGT91D2e S99I (SEQ ID NO:201), UGT91D2e T144L (SEQ ID NO:203), UGT91D2e A148K L221I (SEQ ID NO:205), and UGT91D2e G384K (SEQ ID NO:212) are selective towards RebA, with preferential accumulation of RebD. In some embodiments, UGT91D2e variants not limited to a UGT91D2e variant with a mutation at residue 211 (*e.g.*, UGT91D2e L211M of SEQ ID NO:118) catalyze conversion of rubusoside to 1,2-stevioside and

conversion of RebA to RebD, with preferential accumulation of 1,2-stevioside. See Example 3 and Tables 2 and 3.

**[00105]** In some embodiments, polypeptides suitable for producing steviol glycosides, such as RebA, RebD, rubusoside, and/or 1,2-stevioside in a recombinant host include UGT91D2e-b-EUGT11 chimeric enzymes, such as Chim\_3 (SEQ ID NO:17) or Chim\_7 (SEQ ID NO:18). See Example 4 and Table 5.

**[00106]** In some embodiments, Chim\_7 (SEQ ID NO:18) more efficiently converts rubusoside to 1,2-stevioside, compared to EUGT11 and UGT91D2e. In some embodiments, Chim\_7 (SEQ ID NO:18) fully consumes a supplied amount of rubusoside. In some embodiments, Chim\_7 (SEQ ID NO:18) demonstrates 1.75-fold higher activity towards RebA than UGT91D2e-b (SEQ ID NO:12, SEQ ID NO:13). In some embodiments, Chim\_3 (SEQ ID NO:17) selectively converts rubusoside to 1,2-stevioside. See Example 4 and Table 5.

**[00107]** In some embodiments, UGT91D2e-b-EUGT11 chimeric enzymes such as Chim\_2 (SEQ ID NO:122); Chim\_4 (SEQ ID NO:124); Chim\_5 (SEQ ID NO:125); Chim\_6 (SEQ ID NO:126); Chim\_7 (SEQ ID NO:18); Chim\_8 (SEQ ID NO:128); Chim\_9 (SEQ ID NO:129); Chim\_10 (SEQ ID NO:130); Chim\_11 (SEQ ID NO:131); Chim\_12 (SEQ ID NO:132); Chim\_13 (SEQ ID NO:133); Chim\_14 (SEQ ID NO:134) are used to produce steviol glycosides and/or steviol glycoside precursors.

**[00108]** In some embodiments, a useful UGT85C2 homolog can have one or more amino acid substitutions at residues 21, 48, 49, 84, 86, 87, 91, 92, 95, 122, 304, and 334. See Table 7. Non-limiting examples of useful UGT85C2 homologs include polypeptides having substitutions (with respect to SEQ ID NO:7) at residue 21 (*e.g.*, a lysine, threonine, or valine at residue 21), 48 (*e.g.*, a serine, histidine, tyrosine, arginine, glutamine, or tryptophan at residue 48), 49 (*e.g.*, a valine at residue 49), 84 (*e.g.*, a glycine, alanine, threonine, cysteine, proline, valine, or asparagine at residue 84), 86 (*e.g.*, an arginine or glycine at residue 86); 87 (*e.g.*, an histidine, proline, methionine or tyrosine at residue 87); 91 (*e.g.*, an lysine, arginine, or threonine at residue 91); 92 (*e.g.*, an phenylalanine, isoleucine, methionine, or lysine at residue 92); 122 (*e.g.*, an serine at residue 122); 304 (*e.g.*, a serine at residue 304); and 334 (*e.g.*, an serine or methionine at residue 334). See SEQ ID NOs:127 and 147-180, Table 7A for UGT85C2 variants analyzed that preferentially catalyze conversion of 19-SMG over conversion of steviol, Table 7B for UGT85C2 variants that preferentially catalyze conversion of steviol over conversion of 19-SMG, and Table 7C for additional UGT85C2 variants that catalyze conversion of 19-SMG and steviol. Also see Example 5.

**[00109]** In some embodiments, a steviol glycoside-producing *S. cerevisiae* strain comprising a recombinant gene encoding a *Synechococcus sp.* GGPPS polypeptide (SEQ ID NO:19, SEQ ID NO:20), a recombinant gene encoding a truncated *Z. mays* CDPS polypeptide (SEQ ID NO:39, SEQ ID NO:40), a recombinant gene encoding an *A. thaliana* KS polypeptide (SEQ ID NO:51, SEQ ID NO:52), a recombinant gene encoding a recombinant *S. rebaudiana* KO polypeptide (SEQ ID NO:59, SEQ ID NO:60), a recombinant gene encoding an *A. thaliana* ATR2 polypeptide (SEQ ID NO:91, SEQ ID NO:92), a recombinant gene encoding an *O. sativa* EUGT11 polypeptide (SEQ ID NO:14/SEQ ID NO:15, SEQ ID NO:16), a recombinant gene encoding an SrKAHe1 polypeptide (SEQ ID NO:93, SEQ ID NO:94), a recombinant gene encoding an *S. rebaudiana* CPR8 polypeptide (SEQ ID NO:85, SEQ ID NO:86), a recombinant gene encoding an *S. rebaudiana* UGT74G1 polypeptide (SEQ ID NO:3, SEQ ID NO:4), a recombinant gene encoding an *S. rebaudiana* UGT76G1 polypeptide (SEQ ID NO:8, SEQ ID NO:9), a recombinant gene encoding an *S. rebaudiana* UGT91D2e polypeptide (SEQ ID NO:10, SEQ ID NO:11), a recombinant KO gene encoded by the nucleotide sequence set forth in SEQ ID NO:67 (corresponding to the amino acid sequence set forth in SEQ ID NO:117), and a recombinant CPR1 gene encoding (SEQ ID NO:77, SEQ ID NO:78) accumulates ent-kaurenoic acid+2Glc (#7), ent-kaurenoic acid+3Glc (isomer 1), ent-kaurenoic acid+3Glc (isomer 2), 19-SMG, steviol, steviol+2Glc (#23), and steviol+3Glc (#34) but does not accumulate ent-kaurenol glycosides. See Example 6 and Figures 4A-4C.

**[00110]** In some embodiments, the S84V F48S, F48H, F48Y, F48R, F48Q, F48T, F48S, I49V, P86R, P86G, and F122S variants of UGT85C2 are selective towards 19-SMG, compared to steviol (Table 7A). In some embodiments, the S84T, I87M I87P, I87Y, L91K, L91R, L91T, L92M, and I95K variants of UGT85C2 are selective towards steviol, compared to 19-SMG (Table 7B). In some embodiments, expression of UGT85C2 T304S (SEQ ID NO:127) in a steviol glycoside-producing host increases accumulation of steviol glycosides, compared to a steviol glycoside-producing host not expressing UGT85C2 T304S (SEQ ID NO:127). See Example 5.

**[00111]** In some embodiments, cell lysates comprising UGT85C2 or a UGT85C2 variant show a preference for either steviol or 19-SMG for a substrate. In some aspects, using steviol as a substrate, the F48H, F48Y, F48T, I49V, S84A, and L92F UGT85C2 variants exhibit high activity during incubation periods of under 40 min, and the F48H, F48Y, F48T, and I49V UGT85C2 variants exhibit high activity during incubation periods of over 40 min (Table 8A). Using 19-SMG as a substrate, the F48H, F48Y, F48T, I49V, and S84A UGT85C2 variants

exhibit high activity during incubation periods of under 40 min, and the F48H, I49V, S84A, S84V, L91K, and L92F UGT85C2 variants, as well as the wild-type UGT85C2, exhibit high activity during incubation periods of over 40 min (Table 8B). In some aspects, the L91K, L91R, and L92F UGT85C2 variants exhibit a high 13-SMG/rubusoside ratio, whereas the F48Y, F48T, P86G UGT85C2 variants exhibit a low 13-SMG/rubusoside ratio. See Example 7.

**[00112]** In some embodiments, a useful UGT76G1 homolog can have one or more amino acid substitutions at residues 23, 26, 55, 146, 257, 283, and 337. See Example 4. Non-limiting examples of useful UGT76G1 homologs include polypeptides having substitutions (with respect to SEQ ID NO:9) at residue 21 (e.g., a lysine, threonine or valine at residue 21), residue 23 (e.g., a histidine at residue 23); residue 26 (e.g., a tryptophan at residue 26); residue 55 (e.g., a lysine at residue 55); residue 146 (e.g., a glycine at residue 146); residue 257 (e.g., a glycine at residue 257); residue 283 (e.g., a asparagine at residue 283); and residue 337 (e.g., a proline at residue 337). See SEQ ID NOs: 181-190. See Table 9 and Examples 8 and 9.

**[00113]** In some embodiments, expression of UGT76G1 variants that increase accumulation of RebD or RebM in steviol glycoside-producing *S. cerevisiae* strains (see WO 2014/122227, which has been incorporated by reference in its entirety) alter accumulation of 13-SMG, 1,2-bioside, rubusoside, RebA, RebB, RebD, RebE, RebM, RebG (1,3-stevioside), steviol+3Glc (#1), steviol+4Glc (#26), steviol+5Glc (#22), steviol+5Glc (#24), steviol+5Glc (#25), steviol+6Glc (isomer 1), and steviol+6Glc (#23), compared to expression of wild-type UGT76G1 (SEQ ID NO:9) in steviol glycoside-producing *S. cerevisiae* strains. See Figures 6, 10, 11D, and 11E and Examples 8 and 9.

**[00114]** In some embodiments, expression of UGT variants that increase RebD levels in *S. cerevisiae* also results in increased accumulation of steviol+5Glc (#22), 1,2-stevioside, steviol+6Glc (isomer 1), and steviol+3Glc (#1) but decreased accumulation of steviol+4Glc (#26), steviol+5Glc (#24), and RebG (1,3-stevioside). In some embodiments, expression of UGT76G1 H155L (SEQ ID NO:184) results in increased accumulation of steviol+5Glc (#25) but decreased accumulation of 1,2-stevioside, steviol+3Glc (#1), steviol+4Glc (#26), steviol+5Glc (#22), steviol+6Glc (isomer 1), and steviol+6Glc (#23). In some embodiments, expression of UGT76G1 S253W (SEQ ID NO:186) results in decreased accumulation of 1,2-stevioside and steviol+6Glc (isomer 1). In some embodiments, expression of UGT76G1 284G results in increased accumulation of 1,2-stevioside and steviol+6Glc (isomer 1) but decreased accumulation of RebG, steviol+4Glc (#26), steviol+5Glc (#25), and steviol+6Glc (#23). See Figure 10 and Example 8.

**[00115]** In some embodiments, expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 H155L (SEQ ID NO:184), UGT76G1 L257G (SEQ ID NO:185), and UGT76G1 S283N (SEQ ID NO:188) decrease accumulation of steviol+4Glc (#26). In some embodiments, expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188), all of which increase production of RebD, decrease accumulation of steviol+5Glc (#25), compared to a control strain expressing wild-type UGT76G1. In some embodiments, expression of UGT76G1 H155L (SEQ ID NO:184), which increases RebM production, increases accumulation of steviol+5Glc (#25). See Figure 11D and Example 8.

**[00116]** In some embodiments, expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) increases accumulation of steviol+6Glc (#23), compared to a control strain expressing wild-type UGT76G1. In some embodiments, expression of UGT76G1 H155L (SEQ ID NO:184) decreases accumulation of steviol+6Glc (#23). In some embodiments, expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) increases accumulation of steviol+7Glc (isomer 2), compared to a control strain expressing wild-type UGT76G1. In some embodiments, expression of UGT76G1 H155L (SEQ ID NO:184) decreases accumulation of steviol+7Glc (isomer 2). In some embodiments, expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) increases accumulation of steviol+7Glc (isomer 5). See Figure 11D and Example 8.

**[00117]** In some embodiments, a host expressing a gene encoding a UGT variant or UGT chimeric polypeptide produces an increased level of glycosylated ent-kaurenoic acid and/or ent-kaurenol relative to a host not expressing a gene encoding a UGT variant or UGT chimeric polypeptide. In some embodiments, the UGT variant or UGT chimeric polypeptide comprises a UGT91D2e variant, a gene encoding a UGT91D2e-b-EUGT11 chimeric polypeptide, a gene encoding a UGT85C2 variant, and/or a gene encoding a UGT76G1 variant.

**[00118]** In some embodiments, a host expressing a gene encoding a UGT variant or UGT chimeric polypeptide produces a decreased level of glycosylated ent-kaurenoic acid and/or ent-kaurenol relative to a host not expressing a gene encoding a UGT variant or UGT chimeric

polypeptide. In some embodiments, the UGT variant or UGT chimeric polypeptide comprises a UGT91D2e variant, a gene encoding a UGT91D2e-b-EUGT11 chimeric polypeptide, a gene encoding a UGT85C2 variant, and/or a gene encoding a UGT76G1 variant.

**[00119]** In some embodiments, levels of ent-kaurenoic acid+2Glc (#7), ent-kaurenoic acid+3Glc (isomer 1), ent-kaurenoic acid+3Glc (isomer 2), ent-kaurenol+2Glc (#8), and ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6) are altered in steviol glycoside-producing *S. cerevisiae* strains expressing wild-type UGT76G1 (SEQ ID NO:9), compared to *S. cerevisiae* strains expressing UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 H155L (SEQ ID NO:184), UGT76G1 L257G (SEQ ID NO:185), UGT76G1 S253W (SEQ ID NO:186), UGT76G1 T284G (SEQ ID NO:187), UGT76G1 S283N (SEQ ID NO:188), UGT76G1 K337P (SEQ ID NO:189), or UGT76G1 T55K (SEQ ID NO:190). See Figure 9, Figures 11A-11C, and Example 8.

**[00120]** In some embodiments, *S. cerevisiae* strains expressing UGT76G1 variants that increase RebD levels also increase accumulation of ent-kaurenoic acid+2Glc (#7) and ent-kaurenoic acid+2Glc (isomer 1) but decrease accumulation of ent-kaurenoic acid+3Glc (isomer 2), compared to an *S. cerevisiae* strain expressing wild-type UGT76G1. In some embodiments, UGT76G1 variants that increase RebD levels also increase accumulation of ent-kaurenol+2Glc (#8) but decrease accumulation of ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6). In some embodiments, expression of UGT76G1 H155L (SEQ ID NO:184), a variant that increases levels of RebM, decreases accumulation of ent-kaurenoic acid+2Glc (#7) and ent-kaurenoic acid+3Glc (isomer 1). See Figure 9 and Example 8.

**[00121]** In some embodiments, total levels of glycosylated ent-kaurenoic acid (ent-kaurenoic acid+2Glc (#7) + ent-kaurenoic acid+3Glc (isomer 1) + ent-kaurenoic acid+3Glc (isomer 2)) are increased in steviol glycoside-producing *S. cerevisiae* strains expressing UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), and UGT L257G (SEQ ID NO:185). In some embodiments, total levels of glycosylated ent-kaurenol (ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6) and ent-kaurenol+2Glc (#8) are altered for in steviol glycoside-producing *S. cerevisiae* strains expressing UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), and UGT76G1 T146G (SEQ ID NO:183). See Figures 11B and 11C and Example 8.

**[00122]** In some embodiments, UGT variants not limited to variants of UGT76G1, UGT85C2, and/or UGT91D2e alter ratios of steviol glycosides produced to GlcNAc compounds and isomers thereof produced *in vitro*, *in vivo* in a host, and/or by whole cell bioconversion.

Exemplary GlcNAc structures include ent-kaurenoic acid+2Glc+1GlcNAc and steviol+4Glc+1GlcNAc (#11). See, e.g., Figures 7A, 7D, 8G-8I, and 8AC-8AF and Examples 6, 8, and 9.

**[00123]** In some embodiments, a steviol glycoside or steviol glycoside precursor composition produced *in vivo*, *in vitro*, or by whole cell bioconversion comprises fewer contaminants or less of any particular contaminant than a stevia extract from, *inter alia*, a stevia plant. Contaminants can include plant-derived compounds that contribute to off-flavors. Potential contaminants include pigments, lipids, proteins, phenolics, saccharides, spathulenol and other sesquiterpenes, labdane diterpenes, monoterpenes, decanoic acid, 8,11,14-eicosatrienoic acid, 2-methyloctadecane, pentacosane, octacosane, tetracosane, octadecanol, stigmaterol,  $\beta$ -sitosterol,  $\alpha$ -amyrin,  $\beta$ -amyrin, lupeol,  $\beta$ -amyrin acetate, pentacyclic triterpenes, centaureidin, quercetin, epi- $\alpha$ -cadinol, carophyllenes and derivatives, beta-pinene, beta-sitosterol, and gibberellins.

**[00124]** As used herein, the terms “detectable amount,” “detectable concentration,” “measurable amount,” and “measurable concentration” refer to a level of steviol glycosides measured in area-under-curve (AUC),  $\mu\text{M}/\text{OD}_{600}$ , mg/L,  $\mu\text{M}$ , or mM. Steviol glycoside production (*i.e.*, total, supernatant, and/or intracellular steviol glycoside levels) can be detected and/or analyzed by techniques generally available to one skilled in the art, for example, but not limited to, liquid chromatography-mass spectrometry (LC-MS), thin layer chromatography (TLC), high-performance liquid chromatography (HPLC), ultraviolet-visible spectroscopy/spectrophotometry (UV-Vis), mass spectrometry (MS), and nuclear magnetic resonance spectroscopy (NMR).

**[00125]** As used herein, the term “undetectable concentration” refers to a level of a compound that is too low to be measured and/or analyzed by techniques such as TLC, HPLC, UV-Vis, MS, or NMR. In some embodiments, a compound of an “undetectable concentration” is not present in a steviol glycoside or steviol glycoside precursor composition.

**[00126]** As used herein, the terms “or” and “and/or” is utilized to describe multiple components in combination or exclusive of one another. For example, “x, y, and/or z” can refer to “x” alone, “y” alone, “z” alone, “x, y, and z,” “(x and y) or z,” “x or (y and z),” or “x or y or z.” In some embodiments, “and/or” is used to refer to the exogenous nucleic acids that a recombinant cell comprises, wherein a recombinant cell comprises one or more exogenous nucleic acids selected from a group. In some embodiments, “and/or” is used to refer to production of steviol glycosides and/or steviol glycoside precursors. In some embodiments, “and/or” is used to refer

to production of steviol glycosides, wherein one or more steviol glycosides are produced. In some embodiments, “and/or” is used to refer to production of steviol glycosides, wherein one or more steviol glycosides are produced through one or more of the following steps: culturing a recombinant microorganism, synthesizing one or more steviol glycosides in a recombinant microorganism, and/or isolating one or more steviol glycosides.

### **Functional Homologs**

**[00127]** Functional homologs of the polypeptides described above are also suitable for use in producing steviol glycosides in a recombinant host. A functional homolog is a polypeptide that has sequence similarity to a reference polypeptide, and that carries out one or more of the biochemical or physiological function(s) of the reference polypeptide. A functional homolog and the reference polypeptide can be a natural occurring polypeptide, and the sequence similarity can be due to convergent or divergent evolutionary events. As such, functional homologs are sometimes designated in the literature as homologs, or orthologs, or paralogs. Variants of a naturally occurring functional homolog, such as polypeptides encoded by mutants of a wild type coding sequence, can themselves be functional homologs. Functional homologs can also be created via site-directed mutagenesis of the coding sequence for a polypeptide, or by combining domains from the coding sequences for different naturally-occurring polypeptides (“domain swapping”). Techniques for modifying genes encoding functional polypeptides described herein are known and include, *inter alia*, directed evolution techniques, site-directed mutagenesis techniques and random mutagenesis techniques, and can be useful to increase specific activity of a polypeptide, alter substrate specificity, alter expression levels, alter subcellular location, or modify polypeptide-polypeptide interactions in a desired manner. Such modified polypeptides are considered functional homologs. The term “functional homolog” is sometimes applied to the nucleic acid that encodes a functionally homologous polypeptide.

**[00128]** Functional homologs can be identified by analysis of nucleotide and polypeptide sequence alignments. For example, performing a query on a database of nucleotide or polypeptide sequences can identify homologs of steviol glycoside biosynthesis polypeptides. Sequence analysis can involve BLAST, Reciprocal BLAST, or PSI-BLAST analysis of non-redundant databases using a UGT amino acid sequence as the reference sequence. Amino acid sequence is, in some instances, deduced from the nucleotide sequence. Those polypeptides in the database that have greater than 40% sequence identity are candidates for further evaluation for suitability as a steviol glycoside biosynthesis polypeptide. Amino acid

sequence similarity allows for conservative amino acid substitutions, such as substitution of one hydrophobic residue for another or substitution of one polar residue for another. If desired, manual inspection of such candidates can be carried out in order to narrow the number of candidates to be further evaluated. Manual inspection can be performed by selecting those candidates that appear to have domains present in steviol glycoside biosynthesis polypeptides, e.g., conserved functional domains. In some embodiments, nucleic acids and polypeptides are identified from transcriptome data based on expression levels rather than by using BLAST analysis.

**[00129]** Conserved regions can be identified by locating a region within the primary amino acid sequence of a steviol glycoside biosynthesis polypeptide that is a repeated sequence, forms some secondary structure (e.g., helices and beta sheets), establishes positively or negatively charged domains, or represents a protein motif or domain. See, e.g., the Pfam web site describing consensus sequences for a variety of protein motifs and domains on the World Wide Web at [sanger.ac.uk/Software/Pfam/](http://sanger.ac.uk/Software/Pfam/) and [pfam.janelia.org/](http://pfam.janelia.org/). The information included at the Pfam database is described in Sonnhammer *et al.*, *Nucl. Acids Res.*, 26:320-322 (1998); Sonnhammer *et al.*, *Proteins*, 28:405-420 (1997); and Bateman *et al.*, *Nucl. Acids Res.*, 27:260-262 (1999). Conserved regions also can be determined by aligning sequences of the same or related polypeptides from closely related species. Closely related species preferably are from the same family. In some embodiments, alignment of sequences from two different species is adequate to identify such homologs.

**[00130]** Typically, polypeptides that exhibit at least about 40% amino acid sequence identity are useful to identify conserved regions. Conserved regions of related polypeptides exhibit at least 45% amino acid sequence identity (e.g., at least 50%, at least 60%, at least 70%, at least 80%, or at least 90% amino acid sequence identity). In some embodiments, a conserved region exhibits at least 92%, 94%, 96%, 98%, or 99% amino acid sequence identity.

**[00131]** For example, polypeptides suitable for producing steviol in a recombinant host include functional homologs of UGTs.

**[00132]** Methods to modify the substrate specificity of, for example, a UGT, are known to those skilled in the art, and include without limitation site-directed/rational mutagenesis approaches, random directed evolution approaches and combinations in which random mutagenesis/saturation techniques are performed near the active site of the enzyme. For example see Osmani *et al.*, 2009, *Phytochemistry* 70: 325–347.

**[00133]** A candidate sequence typically has a length that is from 80% to 200% of the length of the reference sequence, *e.g.*, 82, 85, 87, 89, 90, 93, 95, 97, 99, 100, 105, 110, 115, 120, 130, 140, 150, 160, 170, 180, 190, or 200% of the length of the reference sequence. A functional homolog polypeptide typically has a length that is from 95% to 105% of the length of the reference sequence, *e.g.*, 90, 93, 95, 97, 99, 100, 105, 110, 115, or 120% of the length of the reference sequence, or any range between. A % identity for any candidate nucleic acid or polypeptide relative to a reference nucleic acid or polypeptide can be determined as follows. A reference sequence (*e.g.*, a nucleic acid sequence or an amino acid sequence described herein) is aligned to one or more candidate sequences using the computer program Clustal Omega (version 1.2.1, default parameters), which allows alignments of nucleic acid or polypeptide sequences to be carried out across their entire length (global alignment). Chenna *et al.*, 2003, *Nucleic Acids Res.* 31(13):3497-500.

**[00134]** Clustal Omega calculates the best match between a reference and one or more candidate sequences, and aligns them so that identities, similarities and differences can be determined. Gaps of one or more residues can be inserted into a reference sequence, a candidate sequence, or both, to maximize sequence alignments. For fast pairwise alignment of nucleic acid sequences, the following default parameters are used: word size: 2; window size: 4; scoring method: %age; number of top diagonals: 4; and gap penalty: 5. For multiple alignment of nucleic acid sequences, the following parameters are used: gap opening penalty: 10.0; gap extension penalty: 5.0; and weight transitions: yes. For fast pairwise alignment of protein sequences, the following parameters are used: word size: 1; window size: 5; scoring method: %age; number of top diagonals: 5; gap penalty: 3. For multiple alignment of protein sequences, the following parameters are used: weight matrix: blosum; gap opening penalty: 10.0; gap extension penalty: 0.05; hydrophilic gaps: on; hydrophilic residues: Gly, Pro, Ser, Asn, Asp, Gln, Glu, Arg, and Lys; residue-specific gap penalties: on. The Clustal Omega output is a sequence alignment that reflects the relationship between sequences. Clustal Omega can be run, for example, at the Baylor College of Medicine Search Launcher site on the World Wide Web ([searchlauncher.bcm.tmc.edu/multi-align/multi-align.html](http://searchlauncher.bcm.tmc.edu/multi-align/multi-align.html)) and at the European Bioinformatics Institute site at <http://www.ebi.ac.uk/Tools/msa/clustalo/>.

**[00135]** To determine a % identity of a candidate nucleic acid or amino acid sequence to a reference sequence, the sequences are aligned using Clustal Omega, the number of identical matches in the alignment is divided by the length of the reference sequence, and the result is multiplied by 100. It is noted that the % identity value can be rounded to the nearest tenth. For

example, 78.11, 78.12, 78.13, and 78.14 are rounded down to 78.1, while 78.15, 78.16, 78.17, 78.18, and 78.19 are rounded up to 78.2.

**[00136]** It will be appreciated that functional UGT proteins can include additional amino acids that are not involved in the enzymatic activities carried out by the enzymes. In some embodiments, UGT proteins are fusion proteins. The terms “chimera,” “fusion polypeptide,” “fusion protein,” “fusion enzyme,” “fusion construct,” “chimeric protein,” “chimeric polypeptide,” “chimeric construct,” and “chimeric enzyme” can be used interchangeably herein to refer to proteins engineered through the joining of two or more genes that code for different proteins. In some embodiments, a nucleic acid sequence encoding a UGT polypeptide can include a tag sequence that encodes a “tag” designed to facilitate subsequent manipulation (e.g., to facilitate purification or detection), secretion, or localization of the encoded polypeptide. Tag sequences can be inserted in the nucleic acid sequence encoding the polypeptide such that the encoded tag is located at either the carboxyl or amino terminus of the polypeptide. Non-limiting examples of encoded tags include green fluorescent protein (GFP), human influenza hemagglutinin (HA), glutathione S transferase (GST), polyhistidine-tag (HIS tag), and Flag™ tag (Kodak, New Haven, CT). Other examples of tags include a chloroplast transit peptide, a mitochondrial transit peptide, an amyloplast peptide, signal peptide, or a secretion tag.

**[00137]** In some embodiments, a fusion protein is a protein altered by domain swapping. As used herein, the term “domain swapping” is used to describe the process of replacing a domain of a first protein with a domain of a second protein. In some embodiments, the domain of the first protein and the domain of the second protein are functionally identical or functionally similar. In some embodiments, the structure and/or sequence of the domain of the second protein differs from the structure and/or sequence of the domain of the first protein. In some embodiments, a UGT polypeptide is altered by domain swapping.

### **Steviol and Steviol Glycoside Biosynthesis Nucleic Acids**

**[00138]** A recombinant gene encoding a polypeptide described herein comprises the coding sequence for that polypeptide, operably linked in sense orientation to one or more regulatory regions suitable for expressing the polypeptide. Because many microorganisms are capable of expressing multiple gene products from a polycistronic mRNA, multiple polypeptides can be expressed under the control of a single regulatory region for those microorganisms, if desired. A coding sequence and a regulatory region are considered to be operably linked when the

regulatory region and coding sequence are positioned so that the regulatory region is effective for regulating transcription or translation of the sequence. Typically, the translation initiation site of the translational reading frame of the coding sequence is positioned between one and about fifty nucleotides downstream of the regulatory region for a monocistronic gene.

**[00139]** In many cases, the coding sequence for a polypeptide described herein is identified in a species other than the recombinant host, *i.e.*, is a heterologous nucleic acid. Thus, if the recombinant host is a microorganism, the coding sequence can be from other prokaryotic or eukaryotic microorganisms, from plants or from animals. In some case, however, the coding sequence is a sequence that is native to the host and is being reintroduced into that organism. A native sequence can often be distinguished from the naturally occurring sequence by the presence of non-natural sequences linked to the exogenous nucleic acid, *e.g.*, non-native regulatory sequences flanking a native sequence in a recombinant nucleic acid construct. In addition, stably transformed exogenous nucleic acids typically are integrated at positions other than the position where the native sequence is found. "Regulatory region" refers to a nucleic acid having nucleotide sequences that influence transcription or translation initiation and rate, and stability and/or mobility of a transcription or translation product. Regulatory regions include, without limitation, promoter sequences, enhancer sequences, response elements, protein recognition sites, inducible elements, protein binding sequences, 5' and 3' untranslated regions (UTRs), transcriptional start sites, termination sequences, polyadenylation sequences, introns, and combinations thereof. A regulatory region typically comprises at least a core (basal) promoter. A regulatory region also may include at least one control element, such as an enhancer sequence, an upstream element or an upstream activation region (UAR). A regulatory region is operably linked to a coding sequence by positioning the regulatory region and the coding sequence so that the regulatory region is effective for regulating transcription or translation of the sequence. For example, to operably link a coding sequence and a promoter sequence, the translation initiation site of the translational reading frame of the coding sequence is typically positioned between one and about fifty nucleotides downstream of the promoter. A regulatory region can, however, be positioned as much as about 5,000 nucleotides upstream of the translation initiation site, or about 2,000 nucleotides upstream of the transcription start site.

**[00140]** The choice of regulatory regions to be included depends upon several factors, including, but not limited to, efficiency, selectability, inducibility, desired expression level, and preferential expression during certain culture stages. It is a routine matter for one of skill in the art to modulate the expression of a coding sequence by appropriately selecting and positioning

regulatory regions relative to the coding sequence. It will be understood that more than one regulatory region may be present, *e.g.*, introns, enhancers, upstream activation regions, transcription terminators, and inducible elements.

**[00141]** One or more genes can be combined in a recombinant nucleic acid construct in “modules” useful for a discrete aspect of steviol and/or steviol glycoside production. Combining a plurality of genes in a module, particularly a polycistronic module, facilitates the use of the module in a variety of species. For example, a steviol biosynthesis gene cluster, or a UGT gene cluster, can be combined in a polycistronic module such that, after insertion of a suitable regulatory region, the module can be introduced into a wide variety of species. As another example, a UGT gene cluster can be combined such that each UGT coding sequence is operably linked to a separate regulatory region, to form a UGT module. Such a module can be used in those species for which monocistronic expression is necessary or desirable. In addition to genes useful for steviol or steviol glycoside production, a recombinant construct typically also contains an origin of replication, and one or more selectable markers for maintenance of the construct in appropriate species.

**[00142]** It will be appreciated that because of the degeneracy of the genetic code, a number of nucleic acids can encode a particular polypeptide; *i.e.*, for many amino acids, there is more than one nucleotide triplet that serves as the codon for the amino acid. Thus, codons in the coding sequence for a given polypeptide can be modified such that optimal expression in a particular host is obtained, using appropriate codon bias tables for that host (*e.g.*, microorganism). As isolated nucleic acids, these modified sequences can exist as purified molecules and can be incorporated into a vector or a virus for use in constructing modules for recombinant nucleic acid constructs.

**[00143]** In some cases, it is desirable to inhibit one or more functions of an endogenous polypeptide in order to divert metabolic intermediates towards steviol or steviol glycoside biosynthesis. For example, it may be desirable to downregulate synthesis of sterols in a yeast strain in order to further increase steviol or steviol glycoside production, *e.g.*, by downregulating squalene epoxidase. As another example, it may be desirable to inhibit degradative functions of certain endogenous gene products, *e.g.*, glycohydrolases that remove glucose moieties from secondary metabolites or phosphatases as discussed herein. In such cases, a nucleic acid that overexpresses the polypeptide or gene product may be included in a recombinant construct that is transformed into the strain. Alternatively, mutagenesis can be used to generate mutants in genes for which it is desired to increase or enhance function.

### Host Microorganisms

**[00144]** Recombinant hosts can be used to express polypeptides for the producing steviol glycosides. A number of prokaryotes and eukaryotes are suitable for use in constructing the recombinant microorganisms described herein, e.g., gram-negative bacteria, fungi (i.e., yeast), mammalian, insect, plant, and algae cells. A species and strain selected for use as a steviol glycoside production strain is first analyzed to determine which production genes are endogenous to the strain and which genes are not present. Genes for which an endogenous counterpart is not present in the strain are advantageously assembled in one or more recombinant constructs, which are then transformed into the strain in order to supply the missing function(s).

**[00145]** Typically, the recombinant microorganism is grown in a fermenter at a temperature(s) for a period of time, wherein the temperature and period of time facilitate the production of a steviol glycoside. The constructed and genetically engineered microorganisms provided by the invention can be cultivated using conventional fermentation processes, including, *inter alia*, chemostat, batch, fed-batch cultivations, semi-continuous fermentations such as draw and fill, continuous perfusion fermentation, and continuous perfusion cell culture. Depending on the particular microorganism used in the method, other recombinant genes such as isopentenyl biosynthesis genes and terpene synthase and cyclase genes may also be present and expressed. Levels of substrates and intermediates, e.g., isopentenyl diphosphate, dimethylallyl diphosphate, GGPP, ent-kaurene and ent-kaurenoic acid, can be determined by extracting samples from culture media for analysis according to published methods.

**[00146]** Carbon sources of use in the instant method include any molecule that can be metabolized by the recombinant host cell to facilitate growth and/or production of the steviol glycosides. Examples of suitable carbon sources include, but are not limited to, sucrose (e.g., as found in molasses), fructose, xylose, ethanol, glycerol, glucose, cellulose, starch, cellobiose or other glucose-comprising polymer. In embodiments employing yeast as a host, for example, carbon sources such as sucrose, fructose, xylose, ethanol, glycerol, and glucose are suitable. The carbon source can be provided to the host organism throughout the cultivation period or alternatively, the organism can be grown for a period of time in the presence of another energy source, e.g., protein, and then provided with a source of carbon only during the fed-batch phase.

**[00147]** After the recombinant microorganism has been grown in culture for the period of time, wherein the temperature and period of time facilitate the production of a steviol glycoside, steviol and/or one or more steviol glycosides can then be recovered from the culture using various techniques known in the art. In some embodiments, a permeabilizing agent can be added to aid the feedstock entering into the host and product getting out. For example, a crude lysate of the cultured microorganism can be centrifuged to obtain a supernatant. The resulting supernatant can then be applied to a chromatography column, e.g., a C-18 column, and washed with water to remove hydrophilic compounds, followed by elution of the compound(s) of interest with a solvent such as methanol. The compound(s) can then be further purified by preparative HPLC. See also, WO 2009/140394.

**[00148]** It will be appreciated that the various genes and modules discussed herein can be present in two or more recombinant hosts rather than a single host. When a plurality of recombinant hosts is used, they can be grown in a mixed culture to accumulate steviol and/or steviol glycosides.

**[00149]** Alternatively, the two or more hosts each can be grown in a separate culture medium and the product of the first culture medium, e.g., steviol, can be introduced into second culture medium to be converted into a subsequent intermediate, or into an end product such as, for example, RebA. The product produced by the second, or final host is then recovered. It will also be appreciated that in some embodiments, a recombinant host is grown using nutrient sources other than a culture medium and utilizing a system other than a fermenter.

**[00150]** Exemplary prokaryotic and eukaryotic species are described in more detail below. However, it will be appreciated that other species can be suitable. For example, suitable species can be in a genus such as *Agaricus*, *Aspergillus*, *Bacillus*, *Candida*, *Corynebacterium*, *Eremothecium*, *Escherichia*, *Fusarium/Gibberella*, *Kluyveromyces*, *Laetiporus*, *Lentinus*, *Phaffia*, *Phanerochaete*, *Pichia*, *Physcomitrella*, *Rhodoturula*, *Saccharomyces*, *Schizosaccharomyces*, *Sphaceloma*, *Xanthophyllomyces* or *Yarrowia*. Exemplary species from such genera include *Lentinus tigrinus*, *Laetiporus sulphureus*, *Phanerochaete chrysosporium*, *Pichia pastoris*, *Cyberlindnera jadinii*, *Physcomitrella patens*, *Rhodoturula glutinis*, *Rhodoturula mucilaginoso*, *Phaffia rhodozyma*, *Xanthophyllomyces dendrorhous*, *Fusarium fujikuroi/Gibberella fujikuroi*, *Candida utilis*, *Candida glabrata*, *Candida albicans*, and *Yarrowia lipolytica*.

**[00151]** In some embodiments, a microorganism can be a prokaryote such as *Escherichia* bacteria cells, for example, *Escherichia coli* cells; *Lactobacillus* bacteria cells; *Lactococcus*

bacteria cells; *Cornibacterium* bacteria cells; *Acetobacter* bacteria cells; *Acinetobacter* bacteria cells; or *Pseudomonas* bacterial cells.

**[00152]** In some embodiments, a microorganism can be an Ascomycete such as *Gibberella fujikuroi*, *Kluyveromyces lactis*, *Schizosaccharomyces pombe*, *Aspergillus niger*, *Yarrowia lipolytica*, *Ashbya gossypii*, or *S. cerevisiae*.

**[00153]** In some embodiments, a microorganism can be an algal cell such as *Blakeslea trispora*, *Dunaliella salina*, *Haematococcus pluvialis*, *Chlorella sp.*, *Undaria pinnatifida*, *Sargassum*, *Laminaria japonica*, *Scenedesmus almeriensis* species.

**[00154]** In some embodiments, a microorganism can be a cyanobacterial cell such as *Blakeslea trispora*, *Dunaliella salina*, *Haematococcus pluvialis*, *Chlorella sp.*, *Undaria pinnatifida*, *Sargassum*, *Laminaria japonica*, *Scenedesmus almeriensis*.

*Saccharomyces spp.*

**[00155]** *Saccharomyces* is a widely used chassis organism in synthetic biology, and can be used as the recombinant microorganism platform. For example, there are libraries of mutants, plasmids, detailed computer models of metabolism and other information available for *S. cerevisiae*, allowing for rational design of various modules to enhance product yield. Methods are known for making recombinant microorganisms.

*Aspergillus spp.*

**[00156]** *Aspergillus* species such as *A. oryzae*, *A. niger* and *A. sojae* are widely used microorganisms in food production and can also be used as the recombinant microorganism platform. Nucleotide sequences are available for genomes of *A. nidulans*, *A. fumigatus*, *A. oryzae*, *A. clavatus*, *A. flavus*, *A. niger*, and *A. terreus*, allowing rational design and modification of endogenous pathways to enhance flux and increase product yield. Metabolic models have been developed for *Aspergillus*, as well as transcriptomic studies and proteomics studies. *A. niger* is cultured for the industrial production of a number of food ingredients such as citric acid and gluconic acid, and thus species such as *A. niger* are generally suitable for producing steviol glycosides.

*E. coli*

**[00157]** *E. coli*, another widely used platform organism in synthetic biology, can also be used as the recombinant microorganism platform. Similar to *Saccharomyces*, there are libraries of mutants, plasmids, detailed computer models of metabolism and other information available for

*E. coli*, allowing for rational design of various modules to enhance product yield. Methods similar to those described above for *Saccharomyces* can be used to make recombinant *E. coli* microorganisms.

*Agaricus, Gibberella, and Phanerochaete spp.*

**[00158]** *Agaricus, Gibberella, and Phanerochaete* spp. can be useful because they are known to produce large amounts of isoprenoids in culture. Thus, the terpene precursors for producing large amounts of steviol glycosides are already produced by endogenous genes. Thus, modules comprising recombinant genes for steviol glycoside biosynthesis polypeptides can be introduced into species from such genera without the necessity of introducing mevalonate or MEP pathway genes.

*Arxula adenivorans (Blastobotrys adenivorans)*

**[00159]** *Arxula adenivorans* is dimorphic yeast (it grows as budding yeast like the baker's yeast up to a temperature of 42°C, above this threshold it grows in a filamentous form) with unusual biochemical characteristics. It can grow on a wide range of substrates and can assimilate nitrate. It has successfully been applied to the generation of strains that can produce natural plastics or the development of a biosensor for estrogens in environmental samples.

*Yarrowia lipolytica*

**[00160]** *Yarrowia lipolytica* is dimorphic yeast (see *Arxula adenivorans*) and belongs to the family Hemiascomycetes. The entire genome of *Yarrowia lipolytica* is known. *Yarrowia* species is aerobic and considered to be non-pathogenic. *Yarrowia* is efficient in using hydrophobic substrates (e.g. alkanes, fatty acids, oils) and can grow on sugars. It has a high potential for industrial applications and is an oleaginous microorganism. *Yarrowia lipolytica* can accumulate lipid content to approximately 40% of its dry cell weight and is a model organism for lipid accumulation and remobilization. See e.g., Nicaud, 2012, *Yeast* 29(10):409-18; Beopoulos et al., 2009, *Biochimie* 91(6):692-6; Bankar et al., 2009, *Appl Microbiol Biotechnol.* 84(5):847-65.

*Rhodotorula sp.*

**[00161]** *Rhodotorula* is unicellular, pigmented yeast. The oleaginous red yeast, *Rhodotorula glutinis*, has been shown to produce lipids and carotenoids from crude glycerol (Saenge et al., 2011, *Process Biochemistry* 46(1):210-8). *Rhodotorula toruloides* strains have been shown to

be an efficient fed-batch fermentation system for improved biomass and lipid productivity (Li *et al.*, 2007, *Enzyme and Microbial Technology* 41:312-7).

*Rhodosporidium toruloides*

**[00162]** *Rhodosporidium toruloides* is oleaginous yeast and useful for engineering lipid-production pathways (See e.g. Zhu *et al.*, 2013, *Nature Commun.* 3:1112; Ageitos *et al.*, 2011, *Applied Microbiology and Biotechnology* 90(4):1219-27).

*Candida boidinii*

**[00163]** *Candida boidinii* is methylotrophic yeast (it can grow on methanol). Like other methylotrophic species such as *Hansenula polymorpha* and *Pichia pastoris*, it provides an excellent platform for producing heterologous proteins. Yields in a multigram range of a secreted foreign protein have been reported. A computational method, IPRO, recently predicted mutations that experimentally switched the cofactor specificity of *Candida boidinii* xylose reductase from NADPH to NADH. See, e.g., Mattanovich *et al.*, 2012, *Methods Mol Biol.* 824:329-58; Khoury *et al.*, 2009, *Protein Sci.* 18(10):2125-38.

*Hansenula polymorpha (Pichia angusta)*

**[00164]** *Hansenula polymorpha* is methylotrophic yeast (see *Candida boidinii*). It can furthermore grow on a wide range of other substrates; it is thermo-tolerant and can assimilate nitrate (see also *Kluyveromyces lactis*). It has been applied to producing hepatitis B vaccines, insulin and interferon alpha-2a for the treatment of hepatitis C, furthermore to a range of technical enzymes. See, e.g., Xu *et al.*, 2014, *Virol Sin.* 29(6):403-9.

*Kluyveromyces lactis*

**[00165]** *Kluyveromyces lactis* is yeast regularly applied to the production of kefir. It can grow on several sugars, most importantly on lactose which is present in milk and whey. It has successfully been applied among others for producing chymosin (an enzyme that is usually present in the stomach of calves) for producing cheese. Production takes place in fermenters on a 40,000 L scale. See, e.g., van Ooyen *et al.*, 2006, *FEMS Yeast Res.* 6(3):381-92.

*Pichia pastoris*

**[00166]** *Pichia pastoris* is methylotrophic yeast (see *Candida boidinii* and *Hansenula polymorpha*). It provides an efficient platform for producing foreign proteins. Platform elements

are available as a kit and it is worldwide used in academia for producing proteins. Strains have been engineered that can produce complex human N-glycan (yeast glycans are similar but not identical to those found in humans). See, e.g., Piirainen *et al.*, 2014, *N Biotechnol.* 31(6):532-7.

*Physcomitrella spp.*

**[00167]** *Physcomitrella mosses*, when grown in suspension culture, have characteristics similar to yeast or other fungal cultures. This genera can be used for producing plant secondary metabolites, which can be difficult to produce in other types of cells.

### **Steviol Glycoside Compositions**

**[00168]** Steviol glycosides do not necessarily have equivalent performance in different food systems. It is therefore desirable to have the ability to direct the synthesis to steviol glycoside compositions of choice. Recombinant hosts described herein can produce compositions that are selectively enriched for specific steviol glycosides (e.g., RebD or RebM) and have a consistent taste profile. As used herein, the term “enriched” is used to describe a steviol glycoside composition with an increased proportion of a particular steviol glycoside, compared to a steviol glycoside composition (extract) from a stevia plant. Thus, the recombinant hosts described herein can facilitate the production of compositions that are tailored to meet the sweetening profile desired for a given food product and that have a proportion of each steviol glycoside that is consistent from batch to batch. In some embodiments, hosts described herein do not produce or produce a reduced amount of undesired plant by-products found in *Stevia* extracts. Thus, steviol glycoside compositions produced by the recombinant hosts described herein are distinguishable from compositions derived from *Stevia* plants.

**[00169]** It will be appreciated that the amount of an individual steviol glycoside (e.g., RebA, RebB, RebD, or RebM) produced by the recombinant host cell disclosed herein can accumulate in the cell culture broth from about 1 to about 7,000 mg/L, e.g., about 1 to about 10 mg/L, about 3 to about 10 mg/L, about 5 to about 20 mg/L, about 10 to about 50 mg/L, about 10 to about 100 mg/L, about 25 to about 500 mg/L, about 100 to about 1,500 mg/L, or about 200 to about 1,000 mg/L, at least about 1,000 mg/L, at least about 1,200 mg/L, at least about at least 1,400 mg/L, at least about 1,600 mg/L, at least about 1,800 mg/L, at least about 2,800 mg/L, or at least about 7,000 mg/L. In some aspects, the amount of an individual steviol glycoside produced by the recombinant host cell disclosed herein can exceed 7,000 mg/L in the cell culture broth.

**[00170]** It will be appreciated that the amount of a combination of steviol glycosides (*e.g.*, RebA, RebB, RebD, or RebM) produced by the recombinant host cell disclosed herein can accumulate in the cell culture broth from about 1 mg/L to about 7,000 mg/L, *e.g.*, about 200 to about 1,500, at least about 2,000 mg/L, at least about 3,000 mg/L, at least about 4,000 mg/L, at least about 5,000 mg/L, at least about 6,000 mg/L, or at least about 7,000 mg/L. In some aspects, the amount of a combination of steviol glycosides produced by the recombinant host cell disclosed herein can exceed 7,000 mg/L. In general, longer culture times will lead to greater amounts of product. Thus, the recombinant microorganism can be cultured for from 1 day to 7 days, from 1 day to 5 days, from 3 days to 5 days, about 3 days, about 4 days, or about 5 days.

**[00171]** It will be appreciated that the various genes and modules discussed herein can be present in two or more recombinant microorganisms rather than a single microorganism. When a plurality of recombinant microorganisms is used, they can be grown in a mixed culture to produce steviol and/or steviol glycosides. For example, a first microorganism can comprise one or more biosynthesis genes for producing a steviol glycoside precursor, while a second microorganism comprises steviol glycoside biosynthesis genes. The product produced by the second, or final microorganism is then recovered. It will also be appreciated that in some embodiments, a recombinant microorganism is grown using nutrient sources other than a culture medium and utilizing a system other than a fermenter.

**[00172]** Alternatively, the two or more microorganisms each can be grown in a separate culture medium and the product of the first culture medium, *e.g.*, steviol, can be introduced into second culture medium to be converted into a subsequent intermediate, or into an end product such as RebA. The product produced by the second, or final microorganism is then recovered. It will also be appreciated that in some embodiments, a recombinant microorganism is grown using nutrient sources other than a culture medium and utilizing a system other than a fermenter.

**[00173]** Steviol glycosides and compositions obtained by the methods disclosed herein can be used to make food products, dietary supplements and sweetener compositions. See, *e.g.*, WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328.

**[00174]** For example, substantially pure steviol or steviol glycoside such as RebM or RebD can be included in food products such as ice cream, carbonated beverages, fruit juices, yogurts, baked goods, chewing gums, hard and soft candies, and sauces. Substantially pure steviol or

steviol glycoside can also be included in non-food products such as pharmaceutical products, medicinal products, dietary supplements and nutritional supplements. Substantially pure steviol or steviol glycosides may also be included in animal feed products for both the agriculture industry and the companion animal industry. Alternatively, a mixture of steviol and/or steviol glycosides can be made by culturing recombinant microorganisms separately, each producing a specific steviol or steviol glycoside, recovering the steviol or steviol glycoside in substantially pure form from each microorganism and then combining the compounds to obtain a mixture comprising each compound in the desired proportion. The recombinant microorganisms described herein permit more precise and consistent mixtures to be obtained compared to current *Stevia* products.

**[00175]** In another alternative, a substantially pure steviol or steviol glycoside can be incorporated into a food product along with other sweeteners, *e.g.* saccharin, dextrose, sucrose, fructose, erythritol, aspartame, sucralose, monatin, or acesulfame potassium. The weight ratio of steviol or steviol glycoside relative to other sweeteners can be varied as desired to achieve a satisfactory taste in the final food product. See, *e.g.*, U.S. 2007/0128311. In some embodiments, the steviol or steviol glycoside may be provided with a flavor (*e.g.*, citrus) as a flavor modulator.

**[00176]** Compositions produced by a recombinant microorganism described herein can be incorporated into food products. For example, a steviol glycoside composition produced by a recombinant microorganism can be incorporated into a food product in an amount ranging from about 20 mg steviol glycoside/kg food product to about 1800 mg steviol glycoside/kg food product on a dry weight basis, depending on the type of steviol glycoside and food product. For example, a steviol glycoside composition produced by a recombinant microorganism can be incorporated into a dessert, cold confectionary (*e.g.*, ice cream), dairy product (*e.g.*, yogurt), or beverage (*e.g.*, a carbonated beverage) such that the food product has a maximum of 500 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism can be incorporated into a baked good (*e.g.*, a biscuit) such that the food product has a maximum of 300 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism can be incorporated into a sauce (*e.g.*, chocolate syrup) or vegetable product (*e.g.*, pickles) such that the food product has a maximum of 1000 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism can be incorporated into bread such that the food product has a maximum of 160 mg steviol glycoside/kg food on a dry

weight basis. A steviol glycoside composition produced by a recombinant microorganism, plant, or plant cell can be incorporated into a hard or soft candy such that the food product has a maximum of 1600 mg steviol glycoside/kg food on a dry weight basis. A steviol glycoside composition produced by a recombinant microorganism, plant, or plant cell can be incorporated into a processed fruit product (e.g., fruit juices, fruit filling, jams, and jellies) such that the food product has a maximum of 1000 mg steviol glycoside/kg food on a dry weight basis. In some embodiments, a steviol glycoside composition produced herein is a component of a pharmaceutical composition. See, e.g., Steviol Glycosides Chemical and Technical Assessment 69th JECFA, 2007, prepared by Harriet Wallin, Food Agric. Org.; EFSA Panel on Food Additives and Nutrient Sources added to Food (ANS), "Scientific Opinion on the safety of steviol glycosides for the proposed uses as a food additive," 2010, *EFSA Journal* 8(4):1537; U.S. Food and Drug Administration GRAS Notice 323; U.S Food and Drug Administration GRAS Notice Notice 329; WO 2011/037959; WO 2010/146463; WO 2011/046423; and WO 2011/056834.

**[00177]** For example, such a steviol glycoside composition can have from 90-99 weight % RebA and an undetectable amount of stevia plant-derived contaminants, and be incorporated into a food product at from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000 mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis.

**[00178]** Such a steviol glycoside composition can be a RebB-enriched composition having greater than 3 weight % RebB and be incorporated into the food product such that the amount of RebB in the product is from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000 mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis. Typically, the RebB-enriched composition has an undetectable amount of stevia plant-derived contaminants.

**[00179]** Such a steviol glycoside composition can be a RebD-enriched composition having greater than 3 weight % RebD and be incorporated into the food product such that the amount of RebD in the product is from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000 mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis. Typically, the RebD-enriched composition has an undetectable amount of stevia plant-derived contaminants.

**[00180]** Such a steviol glycoside composition can be a RebE-enriched composition having greater than 3 weight % RebE and be incorporated into the food product such that the amount of RebE in the product is from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000

mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis. Typically, the RebE-enriched composition has an undetectable amount of stevia plant-derived contaminants.

**[00181]** Such a steviol glycoside composition can be a RebM-enriched composition having greater than 3 weight % RebM and be incorporated into the food product such that the amount of RebM in the product is from 25-1600 mg/kg, e.g., 100-500 mg/kg, 25-100 mg/kg, 250-1000 mg/kg, 50-500 mg/kg or 500-1000 mg/kg on a dry weight basis. Typically, the RebM-enriched composition has an undetectable amount of stevia plant-derived contaminants.

**[00182]** In some embodiments, a substantially pure steviol or steviol glycoside is incorporated into a tabletop sweetener or "cup-for-cup" product. Such products typically are diluted to the appropriate sweetness level with one or more bulking agents, e.g., maltodextrins, known to those skilled in the art. Steviol glycoside compositions enriched for RebA, RebB, RebD, RebE, or RebM, can be package in a sachet, for example, at from 10,000 to 30,000 mg steviol glycoside/kg product on a dry weight basis, for tabletop use. In some embodiments, a steviol glycoside produced *in vitro*, *in vivo*, or by whole cell bioconversion

**[00183]** The invention will be further described in the following examples, which do not limit the scope of the invention described in the claims.

## EXAMPLES

**[00184]** The Examples that follow are illustrative of specific embodiments of the invention, and various uses thereof. They are set forth for explanatory purposes only, and are not to be taken as limiting the invention.

### Example 1: LC-MS Analytical Procedures

**[00185]** LC-MS analyses for Examples 3 and 4 were performed using an Agilent 1200 Series HPLC system (Agilent Technologies) fitted with a Phenomenex<sup>®</sup> Kinetex C18 column (150 x 2.1 mm, 2.6  $\mu$ m particles, 100 Å pore size) connected to a TSQ Quantum Access (ThermoFisher Scientific) triple quadrupole mass spectrometer with a heated electrospray ion (HESI) source. Elution was carried out using a mobile phase of eluent B (MeCN with 0.1% Formic acid) and eluent A (water with 0.1% Formic acid) by increasing the gradient from 10-40% B from min 0.0 to 1.0, increasing 40-50% B in min 1.0 to 6.5, and increasing 50-100% B from min 6.5 to 7.0.

The flow rate was 0.4 mL/min, and the column temperature was 30°C. 1,2-stevioside and RebD were detected using SIM (Single Ion Monitoring) in positive mode.

**[00186]** LC-MS analyses for Examples 8 and 9 were performed on Waters ACQUITY UPLC® (Waters Corporation) with a Waters ACQUITY UPLC® BEH C18 column (2.1 x 50 mm, 1.7 µm particles, 130 Å pore size) equipped with a pre-column (2.1 x 5 mm, 1.7 µm particles, 130 Å pore size) coupled to a Waters ACQUITY TQD triple quadrupole mass spectrometer with electrospray ionization (ESI) operated in negative ionization mode. Compound separation was achieved using a gradient of the two mobile phases: A (water with 0.1% formic acid) and B (MeCN with 0.1% formic acid) by increasing from 20% to 50 % B between 0.3 to 2.0 min, increasing to 100% B at 2.01 min, holding 100% B for 0.6 min, and re-equilibrating for 0.6 min. The flow rate was 0.6 mL/min, and the column temperature was set at 55°C. Steviol glycosides were monitored using SIM (Single Ion Monitoring) and quantified by comparing against authentic standards. See Table 1 for m/z trace and retention time values of steviol glycosides detected.

**Table 1: LC-MS Analytical Data for Steviol and Steviol Glycosides**

Compound	MS Trace	RT (min)	Figure(s)	Table(s)
steviol+5Glc (#22) [also referred to as compound 5.22]	1127.48	0.85	6D, 7E, 8AK-8AN, 10A, 10B, 11D	9C, 9F, 9I
steviol+6Glc (isomer 1) [also referred to as compound 6.1]	1289.53	0.87	6D, 7B, 8M- 8P, 10A, 10B, 11D	9C, 9F, 9I
steviol+7Glc (isomer 2) [also referred to as compound 7.2]	1451.581	0.94	6D, 7B, 8Q- 8T, 11D	9C, 9F, 9I
steviol+6Glc (#23) [also referred to as compound 6.23]	1289.53	0.97	6D, 10A, 10B, 11D	9F, 9I
RebE	965.42	1.06	6B, 6C, 10C, 11E	9A, 9D, 9G
RebD	1127.48	1.08	6A, 6C, 10C, 11E	2, 3, 5, 9A, 9D, 9G
RebM	1289.53	1.15	6A, 6C, 10C, 11E	9A, 9D, 9G
steviol+7Glc (isomer 5) [also referred to as compound 7.5]	1451.581	1.09	7C, 8Y-8AB, 11D	9F, 9I
steviol+7Glc (#13)	1451.581	0.94	6D	

Compound	MS Trace	RT (min)	Figure(s)	Table(s)
[also referred to as compound 7.13]				
steviol+4Glc (#26) [also referred to as compound 4.26]	965.42	1.21	6D, 7D, 8AG-8AJ, 10A, 10B, 11D	9C, 9F, 9H
steviol+4Glc (#33) [also referred to as compound 4.33]	965.42	1.49		9C, 9I
steviol+5Glc (#24) [also referred to as compound 5.24]	1127.48	1.18	6D, 10A, 10B, 11D	9F, 9I
steviol+4Glc (#25) [also referred to as compound 5.25]	1127.48	1.40	6D, 10A, 10B, 11D	5, 9C, 9F, 9I
RebA	965.42	1.43	6A, 6C, 10C, 11E	9A, 9D, 9G
RebI	1127.48	1.4		9H
1,2-stevioside	803.37	1.43	10B, 11D	2, 3, 5, 9B, 9E, 9H
steviol+3Glc (#1) [also referred to as compound 3.1]	803.37	1.52	6D, 10A, 10B, 11D	9B, 9E
steviol+2Glc (#23) [also referred to as compound 2.23]	641.32	1.57	4C	
steviol+3Glc (#34) [also referred to as compound 3.34]	803.37		4C	9C, 9E
RebQ	965.42	1.59		
1,3-stevioside (RebG)	803.37	1.60	6B-6D, 10B, 11E	9D, 9G
rubusoside	641.32	1.67	5, 6B, 6C, 10C, 11E	5, 8B, 8C, 9D, 9G
RebB	803.37	1.76	6A, 6C, 10C, 11E	9A, 9D, 9G
1,2-bioside	641.32	1.80	6B-D, 10C, 11D, 11E	9A, 9D, 9G
1,3-bioside	641.32	1.95		9E
13-SMG	479.26	2.04	4B, 6A, 6C, 10C, 11E	8A, 8B, 8C, 9A, 9D, 9G
19-SMG	525.27	1.98	4B	7A, 7B, 7C, 8B, 8C, 9E, 9H

Compound	MS Trace	RT (min)	Figure(s)	Table(s)
ent-kaurenoic acid+3Glc (isomer 1) [also referred to as compound KA3.1]	787.37	2.16	4A, 7A, 8A-8C, 9A, 11A, 11B	9B, 9E, 9H
ent-kaurenoic acid+3Glc (isomer 2) [also referred to as compound KA3.2]	787.37	2.28	4A, 7A, 8D-8F, 9A, 11A, 11B	9B, 9E, 9H
ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6) [also referred to as compounds KL3.1 and KL3.6]	773.4	2.36	4A, 7A, 8J-8L, 9B, 11A, 11C	
ent-kaurenoic acid+2Glc (#7) [also referred to as compound KA2.7]	625.32	2.35	4A, 9A, 11A, 11B	9B, 9D, 9H
ent-kaurenol+2Glc (#8) [also referred to as compound KL2.8]	611.34	2.38	9B, 7B, 11A, 11C	9B, 9E
Steviol	317.21	2.39	4C	7A, 7B, 7C, 8A, 8B, 8C, 9F

**[00187]** Steviol glycosides, including GlcNAc-derivatives, glycosylated ent-kaurenol, and/or glycosylated ent-kaurenoic acid can be isolated using a method described herein. For example, following fermentation, a culture broth can be centrifuged for 30 min at 7000 rpm at 4°C to remove cells, or cells can be removed by filtration. The cell-free lysate can be obtained, for example, by mechanical disruption or enzymatic disruption of the host cells and additional centrifugation to remove cell debris. Mechanical disruption of the dried broth materials can also be performed, such as by sonication. The dissolved or suspended broth materials can be filtered using a micron or sub-micron prior to further purification, such as by preparative chromatography. The fermentation media or cell-free lysate can optionally be treated to remove low molecular weight compounds such as salt; and can optionally be dried prior to purification and re-dissolved in a mixture of water and solvent. The supernatant or cell-free lysate can be purified as follows: a column can be filled with, for example, HP20 Diaion® resin (Supelco) or other suitable non-polar adsorbent or reverse phase chromatography resin, and an aliquot of supernatant or cell-free lysate can be loaded on to the column and washed with water to remove the hydrophilic components. The steviol glycoside product can be eluted by stepwise incremental increases in the solvent concentration in water or a gradient from, e. g., 0% → 100% methanol). The levels of steviol glycosides, glycosylated ent-kaurenol, and/or glycosylated ent-kaurenoic acid in each fraction, including the flow-through, can then be analyzed by LC-MS. Fractions can then be combined and reduced in volume using a vacuum

evaporator. Additional purification steps can be utilized, if desired, such as additional chromatography steps and crystallization.

### **Example 2: Strain Engineering and Fermentation**

**[00188]** Steviol glycoside-producing *S. cerevisiae* strains were constructed as described in WO 2011/153378, WO 2013/022989, WO 2014/122227, and WO 2014/122328, each of which is incorporated by reference in their entirety. For example, a yeast strain comprising one or more copies of a recombinant gene encoding a *Synechococcus sp.* GGPPS polypeptide (SEQ ID NO:19, SEQ ID NO:20), a recombinant gene encoding a truncated *Z. mays* CDPS polypeptide (SEQ ID NO:39, SEQ ID NO:40), a recombinant gene encoding an *A. thaliana* KS polypeptide (SEQ ID NO:51, SEQ ID NO:52), a recombinant gene encoding a recombinant *S. rebaudiana* KO polypeptide (SEQ ID NO:59, SEQ ID NO:60), a recombinant gene encoding an *A. thaliana* ATR2 polypeptide (SEQ ID NO:91, SEQ ID NO:92), a recombinant gene encoding an *O. sativa* EUGT11 polypeptide (SEQ ID NO:14/SEQ ID NO:15, SEQ ID NO:16), a recombinant gene encoding an SrKAHe1 polypeptide (SEQ ID NO:93, SEQ ID NO:94), a recombinant gene encoding an *S. rebaudiana* CPR8 polypeptide (SEQ ID NO:85, SEQ ID NO:86), a recombinant gene encoding an *S. rebaudiana* UGT85C2 polypeptide (SEQ ID NO:5/SEQ ID NO:6, SEQ ID NO:7) or a UGT85C2 variant (or functional homolog) of SEQ ID NO:7, a recombinant gene encoding an *S. rebaudiana* UGT74G1 polypeptide (SEQ ID NO:3, SEQ ID NO:4) or a UGT74G1 variant (or functional homolog) of SEQ ID NO:4, a recombinant gene encoding an *S. rebaudiana* UGT76G1 polypeptide (SEQ ID NO:8, SEQ ID NO:9) or a UGT76G1 variant (or functional homolog) of SEQ ID NO:9, and a recombinant gene encoding an *S. rebaudiana* UGT91D2e polypeptide (SEQ ID NO:10, SEQ ID NO:11) or a UGT91D2e variant (or functional homolog) of SEQ ID NO:11 such as a UGT91D2e-b (SEQ ID NO:12, SEQ ID NO:13) polypeptide produced steviol glycosides.

### **Example 3: Modulation of Substrate-Specificity of UGT91D2e**

**[00189]** UGT91D1 (GenBank Accession No. AY345980) is highly expressed in the Stevia plant and thought to be a functional UGT. However, its substrate is not a steviol glycoside. This suggests that UGT91D1 has a different substrate than UGT91D2e, which may be defined by the 22 amino acids with which it differs from UGT91D2e. A UGT91D2e site saturation library (SSL) screen of the 22 amino acids differing from UGT91D1 was prepared using Geneart® (Life Technologies) and degenerate NNK-primers.

**[00190]** UGT91D2 SSL clones were expressed in *E. coli* XJb (DE3) Autolysis™ cells (Zymo Research). Colonies were grown overnight in 96 deep-well plates at 37°C with 1 mL NZCYM (pH 7.0) comprising 15 g Tryptone, 7.5 g NaCl, 7.5 g yeast extract, 1.5 g casamino acids, 3 g MgSO<sub>4</sub> and fortified with 100 mg/L ampicillin and 33 mg/L chloramphenicol. 150 µL overnight cultures were transferred to 24 deep-well plates comprising 3 mL NZCYM with ampicillin, 0.1 mM isopropyl-β-D-1-thiogalactopyranoside (IPTG), 3 mM L-arabinose, and 2% (v/v) ethanol and incubated 20 h at 20°C. Cells were pelleted and lysed in 100 µL lysis buffer (10 mM Tris-HCl pH 8.0, 5 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>, 3 tablets/100 mL Complete mini protease inhibitor cocktail (Roche)) by a single freeze-thaw cycle and 50 µL DNase mix (1 µL 1.4 mg/mL deoxyribonuclease (Calbiochem), 1.2 µL 500 mM MgCl<sub>2</sub>, and 47.8 µL of 4x PBS buffer). Plates were shaken at 500 rpm for 5 min at 25°C to allow degradation of genomic DNA. Plates were then spun down at 4000 rpm for 30 min at 4°C. See WO 2013/022989, which is incorporated by reference in its entirety.

**[00191]** Activity of UGT91D2e variants was tested *in vitro* to assess the specificity of the UGT91D2e variants towards the substrates, rubusoside and RebA. 6 µL of the lysates were diluted with 24 µL of reaction mixture (final concentration: 100 mM Tris-HCl (pH 8.0), 5 mM MgCl<sub>2</sub>, 1 mM KCl, 300 µM uridine diphosphate glucose (UDPG), and 100 µM rubusoside or RebA). The reaction mixture was incubated at 30°C for 24 h, and 1,2-stevioside and RebD production was measured by LC-MS. Results are shown in Table 2.

**Table 2. Activity of UGT91D2e-b and UGT91D2e variants on rubusoside and RebA, producing 1,2-stevioside and RebD, respectively.**

	1,2-stevioside (µM)	RebD (µM)	1,2-stevioside/RebD
UGT91D2e-b (SEQ ID NO:13)	264.9	2.7	98.1
UGT91D2e V286C (SEQ ID NO:1)	59.3	0.0	N/A (No activity on RebA)
UGT91D2e G384W (SEQ ID NO:2)	205.6	0.0	N/A (No activity on RebA)
UGT91D2e L211M (SEQ ID NO:118)	129.7	3.7	35.1
UGT91D2e L195G (SEQ ID NO:119)	178.4	0.9	198.2
UGT91D2e V196P (SEQ ID NO:120)	162.1	2.4	67.5
UGT91D2e L211H (SEQ ID NO:121)	123.5	5.1	24.2

**[00192]** As shown in Table 2, rubusoside and RebA were substrates of UGT91D2e-b (SEQ ID NO:13), UGT91D2e L211M (SEQ ID NO:118), UGT91D2e L195G (SEQ ID NO:119), UGT91D2e V196P (SEQ ID NO:120), and UGT91D2e L211H (SEQ ID NO:121), as 1,2-stevioside and RebD were produced upon contact of the enzymes with either rubusoside or RebA. However, the ratio of 1,2-stevioside/RebD produced by UGT91D2e-b (SEQ ID NO:13), UGT91D2e L211M (SEQ ID NO:118), UGT91D2e L195G (SEQ ID NO:119), UGT91D2e V196P (SEQ ID NO:120), and UGT91D2e L211H (SEQ ID NO:121) fluctuated from 24.2 to 198.2, indicating that the enzymes were not equally selective towards either substrate. The UGT91D2e V286C and UGT91D2e G384W variants were selective towards rubusoside; no RebD was produced upon contact of either variant with RebA.

**[00193]** Additional variants of UGT91D2e were found to demonstrate substrate specificity towards rubusoside or RebA using the above-described assay. See Table 3. The variants of SEQ ID NO:200 (P93V M152G), SEQ ID NO:201 (S99I), SEQ ID NO:203 (T144L), SEQ ID NO:205 (A148K L221I), SEQ ID NO:212 (G384K) were selective towards RebA. The UGT91D2e variants of SEQ ID NO:197 (L195V), SEQ ID NO:198 (V286S), SEQ ID NO:202 (T144K P201P (*silent*)), SEQ ID NO:209 (L211T I130I (*silent*)), SEQ ID NO:211 (S114F V286S), SEQ ID NO:214 (E438M) were selective towards rubusoside.

**Table 3. Activity of UGT91D2e variants on rubusoside and RebA, producing 1,2-stevioside and RebD, respectively.**

Variant	1,2-stevioside ( $\mu\text{M}$ )	RebD ( $\mu\text{M}$ )	1,2-stevioside/RebD
UGT91D2e L213E (SEQ ID NO:191)	13.6	1.1	12.4
UGT91D2e S221Y (SEQ ID NO:192)	13.1	27.1	0.5
UGT91D2e E438H (SEQ ID NO:193)	5.1	1.4	3.6
UGT91D2e M152T (SEQ ID NO:194)	16.8	1.5	11.2
UGT91D2e L211C (SEQ ID NO:195)	7.3	1.6	15.8
UGT91D2e L195S (SEQ ID NO:196)	16.4	1.4	11.7
UGT91D2e L195V (SEQ ID NO:197)	35.9	0.0	N/A (No activity on RebA)
UGT91D2e V286S	14.2	0.0	N/A (No activity on

Variant	1,2-stevioside ( $\mu\text{M}$ )	RebD ( $\mu\text{M}$ )	1,2-stevioside/RebD
(SEQ ID NO:198)			RebA)
UGT91D2e S221S ( <i>silent</i> ) (SEQ ID NO:199)	16.2	1.7	9.5
UGT91D2e P93V M152G (SEQ ID NO:200)	0.2	2.5	0.1
UGT91D2e S99I (SEQ ID NO:201)	0.2	2.6	0.1
UGT91D2e T144K P201P ( <i>silent</i> ) (SEQ ID NO:202)	1.6	0.0	N/A (No activity on RebA)
UGT91D2e T144L (SEQ ID NO:203)	0.0	2.6	0.0 (No activity on rubusoside)
UGT91D2e T144M (SEQ ID NO:204)	1.3	1.6	0.8
UGT91D2e A148K L211I (SEQ ID NO:205)	0.2	2.7	0.1
UGT91D2e L195N (SEQ ID NO:206)	5.1	1.0	5.1
UGT91D2e K199C (SEQ ID NO:207)	2.6	1.3	2.0
UGT91D2e L211M E426G A466V (SEQ ID NO:208)	79.1	1.1	71.9
UGT91D2e L211T I303I ( <i>silent</i> ) (SEQ ID NO:209)	2.7	0.0	N/A (No activity on RebA)
UGT91D2e V286N (SEQ ID NO:210)	3.0	0.0	N/A (No activity on RebA)
UGT91D2e S114F V286S (SEQ ID NO:211)	5.9	0.0	N/A (No activity on RebA)
UGT91D2e G384K (SEQ ID NO:212)	0.0	2.2	0.0 (No activity on rubusoside)
UGT91D2e G384Y (SEQ ID NO:213)	2.9	1.9	1.5
UGT91D2e E438M (SEQ ID NO:214)	4.7	0.0	N/A (No activity on RebA)
UGT91D2e L195C (SEQ ID NO:123)	3.2	1.3	2.5

**Example 4: Evaluation of UGT91D2e-b-EUGT11 Chimeric Enzymes**

**[00194]** UGT91D2e-b-EUGT11 chimeric enzymes were tested *in vitro* to access activity on the substrates, rubusoside and RebA. UGT91D2e-b-EUGT11 chimeras were created by polymerase chain reaction (PCR)-amplification and overlap extension PCR using the primers in Table 4.

**Table 4. Primers Used to Create UGT91D2e-b-EUGT11 Chimeric Enzymes.**

Description	Sequence	SEQ ID
Vector (forward)	GGCAAGCCACGTTTGGTG	SEQ ID NO:135
Vector (reverse)	GGAGCTGCATGTGTCAGAGG	SEQ ID NO:136
EUGT11 fragment 1 / UGT91D2e-b fragment 2 (forward)	CGATGTATTTCACTACTGGTTGCC ATCCATCGCGGCT	SEQ ID NO:137
EUGT11 / UGT91D2e-b fragment 2 (reverse)	AGCCGCGATGGATGGCAACCAGT GATGAAATACATCG	SEQ ID NO:138
UGT91D2e-b fragment 1 / EUGT11 fragment 2 (forward)	TTATGATTATACTCACTACTGGGC TGCTGCAGCCGCATTG	SEQ ID NO:139
UGT91D2e-b fragment 1 / EUGT11 fragment 2 (reverse)	AGCCGCGATGGATGGCAACCAGT GATGAAATACATCG	SEQ ID NO:140
EUGT11 fragment 2 / UGT91D2e-b fragment 3 (forward)	CAAACCTATTACTTTTCCTTGGTTT ACTGCCACCGGAAATAC	SEQ ID NO:141
EUGT11 fragment 2 / UGT91D2e-b fragment 3 (reverse)	GTTTTCCGGTGGCAGTAAACCA AGGAAAGTAATAGGTTTG	SEQ ID NO:142
UGT91D2e-b fragment 2 / EUGT11 fragment 3 (forward)	CCGGTGGTTCCGGTGGGACTAAT GCCTCCATTACATGA	SEQ ID NO:143
UGT91D2e-b fragment 2 / EUGT11 fragment 3 (reverse)	TCATGTAATGGAGGCATTAGTCCC ACCGGAACCACCGG	SEQ ID NO:144
EUGT11 fragment 3 / UGT91D2e-b fragment 4 (forward)	GAACGCAGGTCTGCAGGTTCCAA GAAATGAGGAAGATGG	SEQ ID NO:145
EUGT11 fragment 3 / UGT91D2e-b fragment 4 (reverse)	CCATCTTCCTCATTTCTTGAACC TGCAGACCTGCGTTC	SEQ ID NO:146

**[00195]** UGT91D2e-b-EUGT11 chimeric enzymes were expressed in *E. coli* XJb(DE3) Autolysis™ cells (Zymo Research). Colonies were grown in 50 mL NZCYM (pH 7.0) with ampicillin and chloramphenicol and re-inoculated into 500 mL NZCYM with IPTG, L-arabinose, and ethanol. Cell lysate preparations were done in 15 mL lysis buffer followed by 150  $\mu$ L DNase and 200  $\mu$ L 500 mM MgCl<sub>2</sub>. GST-tag affinity purification of the chimeras was performed by adding 1/3 volume of 4x PBS buffer (560 mM NaCl, 10.8 mM KCl, 40 mM Na<sub>2</sub>HPO<sub>4</sub>, 7.2 mM KH<sub>2</sub>PO<sub>4</sub> (pH 7.3)) to the lysate supernatant, followed by incubation (2 h, 4°C) with Glutathione Sepharose 4B (GE Healthcare) and loading onto Poly-Prep® Chromatography Columns (Bio-Rad). The beads were washed twice with 1X PBS buffer and eluted with 50 mM Tris-HCl (pH 8.0) and 10 mM reduced glutathione. Eluted protein was stabilized by addition of glycerol to a final concentration of 50%. SDS-PAGE was performed using NuPAGE® 4-12 % Bis-Tris 1.0 mm precast gels (Invitrogen), NuPAGE MOPS (Invitrogen) running buffer and SimplyBlue SafeStain (Invitrogen). The amounts of chimeras produced were determined from the relative staining intensity of the gel images using ImageJ software.

**[00196]** Chimeras were screened by adding 20  $\mu$ L purified UGT91D2e-b, EUGT11, or UGT91D2e-b-EUGT11 chimeric enzymes (0.02 mg/mL) to a total volume of 80  $\mu$ L reaction mixture comprising 100 mM Tris-HCl (pH 8.0), 5 mM MgCl<sub>2</sub>, 1 mM KCl, 300  $\mu$ M uridine diphosphate glucose (UDPG), and 100  $\mu$ M rubusoside or RebA. The reactions were incubated at 30°C for 24 h, and levels of RebA, RebD, rubusoside, and 1,2-stevioside were measured by LC-MS. Not all of the chimeras purified were active in the above described assay (see Table 5 for enzymes having activity on rubusoside and/or RebA).

**Table 5. EUGT11, UGT91D2e-b, and EUGT11-UGT91D2e-b chimeric enzyme activity on RebA and rubusoside.**

	RebA ( $\mu$ M)	RebD ( $\mu$ M)	rubusoside ( $\mu$ M)	1,2-stevioside (AUC)
EUGT11 (SEQ ID NO:16)	32.230	101.300	34.899	1188497
UGT91D2e-b (SEQ ID NO:13)	97.314	6.580	41.157	2660570
Chim_3 (SEQ ID NO:17)	109.764	NF	138.911	11435
Chim_7 (SEQ ID NO:18)	88.502	11.510	NF	3693895

\*NF=Not Found

**[00197]** As shown in Table 5, Chim\_7 (SEQ ID NO:18) more efficiently converted rubusoside to 1,2-stevioside, compared to EUGT11 and UGT91D2e. Chim\_7 (SEQ ID NO:18) fully consumed the supplied amount of rubusoside, unlike EUGT11 or UGT91D2e. When incubating EUGT11 with rubusoside, the C19-position of rubusoside was 1,2-glycosylated, and RebE and 1,2-stevioside were also produced (Table 5). Additionally, Chim\_7 (SEQ ID NO:18) demonstrated 1.75-fold higher activity towards RebA than UGT91D2e-b. Chim\_3 (SEQ ID NO:17) selectively converted rubusoside to 1,2-stevioside; no RebA was converted to RebD by Chim\_3 (SEQ ID NO:17) (Table 5).

#### Example 5: Evaluation of UGT85C2 Variants

**[00198]** Three homology models of UGT85C2 were generated with the ORCHESTRA module in Sybyl-X 2.0 (Certara) using a combination of the three PDB templates (Model 1: 2PQ6, 2VCE, 2C1X; Model 2: 2PQ6; Model 3: 2PQ6, 2C1X) and using standard settings and sequences for UGT85H2, UGT72B1, and VvGT1 (see PDB2PQ6, PDB2VCE, and PCB2C1X). Model geometry and quality were checked with the molprobit and ProQ webserver (see Chen *et al.*, Acta Crystallographica. Section D, Biological Crystallography 66(Pt 1):12-21 (2010), Davis *et al.*, Nucleic Acids Research 35:W375-83 (2007), Wallner & Elofsson, Protein Science : A Publication of the Protein Society 12(5):1073-86 (2003). The fluorinated UDPG sugar donor analog, UDP-2FGlc, from PDB:2VCE was imported into the UDPG binding site of UGT85C2 prior to the acceptors steviol, 13-SMG, 19-SMG, or rubusoside. Steviol and steviol glycosides were prepared using the Sybyl-X small molecule builder and docked into the active site of the enzyme with the Surflex Dock suite using standard GeomX settings. The sites for the site saturation library (SSL) were determined by selecting all the residues within 3 Å of the ligands in the docking analysis that were not 100% conserved in the PDB-templates. See Table 6.

**Table 6. SSL residues for UGT85C2 Docking Analysis.**

	UGT85C2 Model #1	UGT85C2 Model #2	UGT85C2 Model #3	Conserved
<b>Phe18</b>	x	x	x	
<b>Pro19</b>	x	x	x	C
<b>Ala20</b>	x	x	x	
<b>Gln21</b>	x	x	x	

	UGT85C2 Model #1	UGT85C2 Model #2	UGT85C2 Model #3	Conserved
Ser22	x	x	x	
His23	x	x	x	C
Lys25		x	x	
Phe48		x	x	
Ile49			x	
Gln52			x	
Glu82		x		
Ala83		x		
Ser84		x		
Pro86			x	
Ile87			x	
Arg88	x		x	
Leu91	x		x	
Leu92	x			
Ile95	x			
Phe122	x			
Thr143	x	x		
Leu144	x	x	x	
Asp198		x		
Val207		x		
Phe210	x			
Thr211	x			
Asn300	x			
Phe301	x			C
Gly302	x		x	C
Ser303	x		x	
Thr304	x	x	x	
Thr305	x	x	x	
Val306			x	
Leu334			x	
Trp359			x	C
Gln362	x			C
His377	x	x		C
Gly379		x	x	C
Trp380	x	x	x	C
Gly381		x	x	
Ser382	x	x	x	C
Tyr398		x	x	

	UGT85C2 Model #1	UGT85C2 Model #2	UGT85C2 Model #3	Conserved
<b>Trp400</b>	x	x	x	
<b>Asp401</b>	x	x	x	
<b>Gln402</b>	x	x		C

x: Residue within 3 Å of steviol, 19-SMG, and UDPG in the docking analysis

C: Conserved residue

**[00199]** SSL clones were generated for the 34 non-conserved amino acids in Table 6 predicted to be within 3 Å of the ligands residues. A modified version of the whole plasmid amplification method (Zheng *et al.* Nucleic Acids Research 32(14):e115 (2004)) was used with overlapping NNK-primers and Phusion polymerase. 10 µL PCR reaction was treated with 10 U DpnI (New England Biolabs) at 37°C for 1 h, heat inactivated at 65°C for 20 min, and transformed into *E. coli* DH5α cells. Colonies were selected on Luria Broth (LB) + kanamycin agar plates and grown in 4 mL LB fortified with kanamycin. Plasmids were purified using the GeneJET™ miniprep kit (Thermo Fisher Scientific) and sequenced.

**[00200]** The sequence-verified site saturation library (SSL) clones were transformed into *E. coli* XJb(DE3) Autolysis™ cells (Zymo Research) and selected on LB + kanamycin agar plates. Single colonies were inoculated into 1 mL NZCYM fortified with 30 mg/L kanamycin and incubated overnight at 37°C and 200 rpm orbital shaking. 50 µL of the overnight culture were transferred into 1 mL of fresh NZCYM fortified with 30 mg/L kanamycin, 3 mM arabinose, and 0.1 mM IPTG and incubated overnight at 20°C and 200 rpm orbital shaking. The cells were spun down at 3220 g / 10 min at 4°C and resuspended in 50 µL GT-buffer (10 mM Tris-HCl (pH 7.5), 5 mM MgCl<sub>2</sub>, 1 mM CaCl<sub>2</sub>) comprising complete Mini EDTA free protease inhibitor cocktail (1 tablet / 25 mL GT-buffer; Roche Diagnostics). Pellets were resuspended by orbital shaking at 200 rpm / 5 min at 4°C. Cells were incubated at -80°C for minimum 15 min before initiation of lysing step.

**[00201]** The cells were lysed by heating the samples to 25°C and adding 25 µL DNase I mix comprising of 2.39 mL 4x His binding buffer (80 mM Tris-HCl (pH 7.5), 500 mM NaCl, 10 mM Imidazole) with 50 µL 1.4 mg/mL DNase I bovine pancreas (Calbiochem) and 60 µL MgCl<sub>2</sub> (500 mM). The lysates were filtered through a 1.2 µm 96-well filterplate (EMD Millipore) and transferred to another 1.2 µm filterplate comprising 50 µL His-select beads (Sigma-Aldrich) prewashed twice with 1X binding buffer. The lysates and beads were then incubated for 2 h at 4°C with 500 rpm orbital shaking. The plates were spun down at 450 g / 2 min. Total protein

concentration in the flow-through was measured using the Bradford assay reagent (Sigma-Aldrich), the samples were washed twice by centrifuging the samples, removing supernatants and adding 50  $\mu$ L 1X His binding buffer. Elution buffer (20 mM Tris-HCl (pH 7.5), 500 mM NaCl, 250 mM imidazole) was added to the beads and incubated for 5 min at 4 °C at 500 rpm orbital shaking and the proteins eluted into a 96 well PCR plate (FrameStar 96, 4titude). The purifications were evaluated by running samples of the flow-through, washing steps and eluate on NuPAGE® SDS-PAGE gel system with 4-12% Bis-Tris precast gels (Invitrogen).

**[00202]** Activity of the purified UGT85C2 variants was measured. 2.0  $\mu$ g/mL UGT85C2 variant was incubated for 20 min at 37°C with reaction buffer (100 mM Tris-HCl (pH 8.0), 1 mM KCl, Calf Intestinal Alkaline Phosphatase (New England Biolabs), 120  $\mu$ M UDPG, and either 40  $\mu$ M steviol or 40  $\mu$ M 19-SMG). In this assay, the glucose on UDPG was transferred to steviol or 19-SMG; the products were UDP and either 13-SMG or rubusoside. The phosphates on UDP were then released by a phosphatase, and the amount of phosphate released was measured at Abs<sub>600</sub> using the Malachite green protocol (Baykov *et al.*, Analytical Biochemistry 171(2):266-70). Values were normalized by total protein released measured by using Bradford reagent (Sigma-Aldrich).

**[00203]** Candidates were selected as having activity of one standard deviation or higher than wild-type activity or having less than 50% activity on one substrate while maintaining wild-type activity on the other (*e.g.*, exhibiting substrate-specificity). The Abs<sub>600</sub> ratios of a steviol sample to a 19-SMG sample for wild-type UGT85C2 (SEQ ID NO:7) averaged 0.94, indicating that the wild-type UGT85C2 catalyzes conversion of steviol and 19-SMG with little or no preference of substrate. Table 7A shows the UGT85C2 variants analyzed that preferentially catalyzed conversion of 19-SMG over conversion of steviol, Table 7B shows the UGT85C2 variants analyzed that preferentially catalyzed conversion of steviol over conversion of 19-SMG, and Table 7C shows the UGT85C2 variants analyzed that catalyzed conversion of 19-SMG and steviol with little preference for either substrate. Particular clones generated by the site saturation library (SSL) screen were selected more than once, corresponding to more than one entry in Tables 7A-C.

**Table 7A. UGT85C2 SSL screen candidates that were selective towards 19-SMG as a substrate.**

Steviol (Abs <sub>600</sub> )	19-SMG (Abs <sub>600</sub> )	Steviol/19- SMG Abs <sub>600</sub> Ratio	Sum (Abs <sub>600</sub> )	Mutation	UGT85C2 Variant SEQ ID
0.105	0.165	0.636	0.27	F48S	SEQ ID NO:150
0.099	0.136	0.728	0.235	F48H	SEQ ID NO:151
0.089	0.142	0.627	0.231	F48Y	SEQ ID NO:152
0.080	0.117	0.684	0.197	F48R	SEQ ID NO:153
0.068	0.126	0.540	0.194	F48Q	SEQ ID NO:154
0.068	0.112	0.607	0.18	F48T	SEQ ID NO:156
0.065	0.114	0.570	0.179	F48S	SEQ ID NO:150
0.094	0.141	0.667	0.235	I49V	SEQ ID NO:157
0.078	0.111	0.703	0.189	I49V	SEQ ID NO:157
0.116	0.238	0.487	0.354	S84V	SEQ ID NO:164
-0.020	0.153	19-SMG	0.133	S84V	SEQ ID NO:164
0.096	0.230	0.417	0.326	P86R	SEQ ID NO:165
0.083	0.196	0.423	0.279	P86R	SEQ ID NO:165
0.065	0.17	0.382	0.235	P86R	SEQ ID NO:165
0.042	0.18	0.233	0.222	P86G	SEQ ID NO:166
-0.003	0.169	19-SMG	0.166	P86R	SEQ ID NO:165

**Table 7B. UGT85C2 SSL screen candidates that were selective towards steviol as a substrate.**

Steviol (Abs <sub>600</sub> )	19-SMG (Abs <sub>600</sub> )	Steviol/19- SMG Ratio	Sum (Abs <sub>600</sub> )	Mutation	UGT85C2 Variant SEQ ID
0.382	-0.081	Steviol	0.301	S84T	SEQ ID NO:160
0.242	-0.083	Steviol	0.159	S84T	SEQ ID NO:160
0.521	-0.033	Steviol	0.488	I87M	SEQ ID NO:169
0.261	0.190	1.374	0.451	I87Y	SEQ ID NO:170
0.372	0.159	2.340	0.531	L91K	SEQ ID NO:171
0.369	0.134	2.754	0.503	L91K	SEQ ID NO:171
0.228	0.104	2.192	0.332	L91R	SEQ ID NO:172
0.202	0.079	2.557	0.281	L91R	SEQ ID NO:172
0.147	0.041	3.585	0.188	L91T	SEQ ID NO:173
0.606	0.266	2.278	0.872	I95K	SEQ ID NO:177

**Table 7C. UGT85C2 SSL screen candidates that were not substrate selective towards steviol or 19-SMG.**

Steviol (Abs <sub>600</sub> )	19-SMG (Abs <sub>600</sub> )	Steviol/19- SMG Ratio	Sum (Abs <sub>600</sub> )	Mutation	UGT85C2 Variant SEQ ID
0.229	0.268	0.854	0.497	Q21L	SEQ ID NO:147
0.231	0.261	0.885	0.492	Q21T	SEQ ID NO:148
0.214	0.252	0.849	0.466	Q21V	SEQ ID NO:149
0.083	0.098	0.847	0.181	F48W	SEQ ID NO:155
0.359	0.332	1.081	0.691	S84G	SEQ ID NO:158
0.306	0.331	0.924	0.637	S84A	SEQ ID NO:159
0.296	0.292	1.014	0.588	S84C	SEQ ID NO:161
0.250	0.299	0.836	0.549	S84P	SEQ ID NO:162
0.250	0.256	0.977	0.506	S84A	SEQ ID NO:159
0.219	0.262	0.836	0.481	S84N	SEQ ID NO:163
0.355	0.306	1.160	0.661	I87H	SEQ ID NO:167
0.326	0.274	1.190	0.600	I87P	SEQ ID NO:168
0.308	0.282	1.092	0.590	I87M	SEQ ID NO:169
0.279	0.216	1.292	0.495	I87Y	SEQ ID NO:170
0.474	0.426	1.113	0.900	L92F	SEQ ID NO:174
0.387	0.331	1.169	0.718	L92I	SEQ ID NO:175
0.342	0.260	1.315	0.602	L92M	SEQ ID NO:176
0.39	0.598	0.652	0.988	F122S	SEQ ID NO:178
0.297	0.248	1.198	0.545	L334S	SEQ ID NO:179
0.27	0.233	1.159	0.503	L334M	SEQ ID NO:180

**[00204]** The purified S84V and P86R variants of UGT85C2 were selective towards 19-SMG; UGT85C2 S84V and UGT85C2 P86R did not demonstrate activity on steviol (Table 7A). The purified F48S, F48H, F48Y, F48R, F48Q, F48T, F48S, I49V, P86R, P86G, and F122S UGT85C2 variants also showed selectivity towards 19-SMG (Table 7A). However, the purified S84T and I87M variants of UGT85C2 were selective towards steviol; UGT85C2 S84T and UGT85C2 I87M did not demonstrate activity on 19-SMG (Table 7B). The purified I87P, I87Y, L91K, L91R, L91T, L92M, and I95K UGT85C2 variants also showed selectivity towards steviol (Table 7B).

**Example 6: Characterization of Steviol Glycoside-Producing Yeast Strain Deleted of UGT85C2**

**[00205]** A modified version of the steviol glycoside-producing *S. cerevisiae* strain described in Example 2, a recombinant KO gene encoded by the nucleotide sequence set forth in SEQ ID NO:67 (corresponding to the amino acid sequence set forth in SEQ ID NO:117) and a recombinant CPR1 gene encoding (SEQ ID NO:77, SEQ ID NO:78) was deleted for *S. rebaudiana* UGT85C2 polypeptide (SEQ ID NO:5/SEQ ID NO:6, SEQ ID NO:7). Sixteen independent clones were grown in Synthetic Complete (SC) medium at 30°C for 5 days with shaking (400 rpm for deep wells) prior to harvest. Culture samples (without cell removal) were heated in the presence of DMSO for detection of total glycoside levels with LC-MS.

**[00206]** As shown in Figure 4A, culture samples of cells deleted of UGT85C2 did not accumulate ent-kaurenol glycosides (ent-kaurenol+3Glc (isomer 1), ent-kaurenol+3Glc (#6), or ent-kaurenol\_2Glc (#8), as compared to the control strain (not deleted for UGT85C2). This result suggests that UGT85C2 is responsible for the 19-O-glucosylation of ent-kaurenol. Also as shown in Figure 4A, culture samples of cells deleted of UGT85C2 did accumulate ent-kaurenoic acid glycosides (ent-kaurenoic acid+2Glc (#7), ent-kaurenoic acid+3Glc (isomer 1), and ent-kaurenoic acid+3Glc (isomer 2)). Whereas control samples accumulated 13-SMG, culture samples of cells deleted of UGT85C2 accumulated 19-SMG, steviol, steviol+2Glc (#23), and steviol+3Glc (#34). See Figures 4B and 4C. Steviol+2Glc (#23) and steviol+3Glc (#34) likely have two or three glucose moieties, respectively, attached on the 19 position of the steviol backbone.

**[00207]** Structures of isolated tri-glycosylated ent-kaurenoic acid, elucidated by NMR, are shown in Figure 7A, along with a structure of tri-glycosylated ent-kaurenol. These structures were solved by means of standard homo- and heteronuclear multipulse NMR experiments, <sup>1</sup>H,<sup>1</sup>H-COSY, <sup>1</sup>H,<sup>1</sup>H-ROESY, <sup>1</sup>H,<sup>13</sup>C-HSQC, and <sup>1</sup>H,<sup>13</sup>C-HMBC. Compounds were dissolved in 60 μL DMSO-d6 and measured at 25°C. Spectra of these compounds were acquired on an 800 MHz Bruker Avance instrument (800 MHz for <sup>1</sup>H, 201 MHz for <sup>13</sup>C) equipped with a cryogenic probe (5 mm CPTCI 1H-13C/15N/D Z-GRD Z44909/0010). In addition, <sup>1</sup>H-NMR spectra were obtained for 3 molecules detected by LC-MS that were concordant with a general ent-kaurenoic acid+2Glc, ent-kaurenol+3Glc (isomer 2), and ent-kaurenol+Glc+GlcNAc structures. See Figures 8A-8L for <sup>1</sup>H NMR spectra and <sup>1</sup>H and <sup>13</sup>C NMR chemical shifts for these compounds.

**[00208]** UGT85C2 variants were subsequently cloned into USER vectors (for integration at ChrXII-1) using a forward primer (SEQ ID NO:215) and a reverse primer (SEQ ID NO:216) and the PGK1 promoter. The UGT85C2 variants were then integrated into the steviol glycoside-

producing strain deleted of UGT85C2. Transformants were re-streaked from transformation plates. Pre-cultures were set up from re-streaked plates in 500  $\mu$ L synthetic complete-URA (SC-URA) media in a 96 deep well plate (DWP) and grown at 30°C and 300 rpm overnight. Cultures were set up by transferring 50  $\mu$ L of the pre-cultures to a 96 well DWP comprising 500  $\mu$ L SC-URA media.

**[00209]** After 1 day of incubation, cultures were set up from pre-cultures (50  $\mu$ L in 500  $\mu$ L SC-URA) and grown in Duetz system for 5 days (same conditions as for pre-cultures). The OD<sub>600</sub> was measured on plate reader in a 1:10 dilution, and samples were harvested by transferring 50  $\mu$ L sample to 50  $\mu$ L 100% DMSO. The mixtures were heated to 80°C for 10 min and subsequently spun down (4000 rcf, 4°C, 10 min). 15  $\mu$ L of each supernatant were mixed with 105  $\mu$ L 50% DMSO (total dilution of 1:16), and the samples were analyzed by LC-MS.

#### **Example 7: Assessment of UGT85C2 Variant Activity in Cell Lysates**

**[00210]** Purified variant UGT85C2 DNA from Example 6 was individually transformed into XJB autolysis z-competent cells. Pre-cultures of three colonies from each transformation plate were inoculated into 600  $\mu$ L LB comprising kanamycin (600 mg/L) and incubated overnight at 200 rpm and 37°C in a 96 well DWP. Protein production and cell wall degradation were induced by transferring 50  $\mu$ L of the pre-cultures to a new 96 well DWP comprising 1mL/well of NZCYM broth comprising kanamycin (600 mg/L) + 3 mL/L 1M Arabinose and 100  $\mu$ L/L 1M IPTG. Cultures were incubated at 20°C, 200 rpm for approximately 20 h before pelleting the cells (4000 rcf, 5 min, 4°C) and removing the supernatant. To each well, 50  $\mu$ L GT buffer with protease inhibitor (cOmplete™, Mini, EDTA-free Protease Inhibitor Cocktail Tablets, 11836170001 Roche) was added. Pellets were resuspended by shaking at 200 rpm for 5 min at 4°C. A 75  $\mu$ L aliquot of each sample was transferred to a PCR plate and frozen at -80°C. Pellets were thawed at room temperature, and 25  $\mu$ L/well DNase mix (2,39 mL 4x binding buffer + 50  $\mu$ L DNase I (1.4 mg/mL) + 60  $\mu$ L MgCl<sub>2</sub> (1 M) per plate) were added when samples were nearly thawed. The plate was incubated at room temperature for 5 min with gentle shaking and subsequently centrifuged at 4000 rcf for 5 min. Each supernatant was transferred to a fresh PCR plate for activity measurements.

**[00211]** Each supernatant was incubated in an assay reaction mix comprising a final concentration of 100 mM Tris (pH 8.0), 4 mM MgCl<sub>2</sub>, 1 mM KCl, 300  $\mu$ M UDP-Glucose, and 100  $\mu$ M substrate. The substrates were either steviol or 19-SMG. A purified wild-type UGT85C2

enzyme and a UGT85C2 bacterial lysate were used as positive controls. Reactions were incubated at 30°C (on a plate shaker), and the reactions were stopped after 20 min, 40 min, and 19 h by mixing 20 µL sample with 20 µL 100% DMSO. The samples were further diluted by adding 60 µL 50% DMSO and subsequently analyzed by LC-MS. AUC values corresponding to measured 13-SMG, 19-SMG, rubusoside, and steviol levels are shown in Tables 8A-C.

**Table 8A. Measured 13-SMG and steviol AUC values in UGT85C2 variant activity assay using steviol as a substrate.**

UGT85C2 Variant	13-SMG			Steviol		
	20 min	40 min	19 h	20 min	40 min	19 h
F48S (SEQ ID NO:150)	38195	55395	76045	21355	9955	
F48H (SEQ ID NO:151)	49840	64105	79000	17670	4035	
F48Y (SEQ ID NO:152)	36980	53005	83100	26675	16135	
F48R (SEQ ID NO:153)	37990	55510	71810	25540	11075	
F48Q (SEQ ID NO:154)	33660	46010	72550	30565	16135	
F48W (SEQ ID NO:155)	37580	56220	76490	25280	8615	
F48T (SEQ ID NO:156)	40505	57280	78080	20405	10340	
I49V (SEQ ID NO:157)	48345	60720	75420	17545	4305	
S84G (SEQ ID NO:158)	33960	50770	76070	29500	15870	
S84A (SEQ ID NO:159)	43135	62000	75715	21445	5190	
S84C (SEQ ID NO:161)	25780	39330	71060	34285	22700	
S84V (SEQ ID NO:164)	27045	43200	74505	32100	17715	
P86R (SEQ ID NO:165)	23240	34440	71955	33670	25395	
P86G (SEQ ID NO:166)	28000	43525	74300	27640	14380	
I87H (SEQ ID NO:167)	7290	10465	43495	51340	41690	21865
I87P (SEQ ID NO:168)	32165	48565	76700	29475	13945	
I87Y (SEQ ID NO:170)	36905	47250	71390	31220	14065	
L91K (SEQ ID NO:171)	25810	37830	72435	29455	19015	2770
L91R (SEQ ID NO:172)	27560	40235	75830	34275	22140	2470
L92F (SEQ ID NO:174)	49205	62540	72385	15635	3570	

**Table 8B. Measured 13-SMG, 19-SMG, and rubusoside AUC values in UGT85C2 variant activity assay using 19-SMG as a substrate.**

	19-SMG	rubusoside

<b>UGT85C2 Variant</b>	20 min	40 min	19 h	20 min	40 min	19 h
F48S (SEQ ID NO:150)	171625	147690	3720	18935	30650	92800
F48H (SEQ ID NO:151)	165365	129495	1830	24415	40520	99660
F48Y (SEQ ID NO:152)	161680	128705	2815	23130	39385	97180
F48R (SEQ ID NO:153)	166035	142095	6120	17335	30075	93750
F48Q (SEQ ID NO:154)	169560	145130	3235	16570	28495	81190
F48W (SEQ ID NO:155)	168175	147640	3920	16040	28030	95530
F48T (SEQ ID NO:156)	166190	134425	2960	22445	37520	96620
I49V (SEQ ID NO:157)	170460	133705	1935	20340	35300	97440
S84G (SEQ ID NO:158)	175515	147045	3165	14645	24745	91945
S84A (SEQ ID NO:159)	163565	131735	1790	19805	31845	90090
S84C (SEQ ID NO:161)	183175	159805	44230	11040	17040	77130
S84V (SEQ ID NO:164)	183415	168240	6600	11975	20075	98555
P86R (SEQ ID NO:165)	186925	154290	12670	12075	20350	85755
P86G (SEQ ID NO:166)	175265	146080	5720	17660	29815	93195
I87H (SEQ ID NO:167)	197170	191250	149025	3045	5300	27610
I87P (SEQ ID NO:168)	167935	143945	8795	16675	28290	96865
I87Y (SEQ ID NO:170)	176815	142820	4750	16635	26615	93205
L91K (SEQ ID NO:171)	188110	182210	177120	5350	8545	20345
L91R (SEQ ID NO:172)	188750	180040	149165	7535	12140	29160
L92F (SEQ ID NO:174)	187295	155170	2695	11335	22340	98920

**Table 8C. Measured 13-SMG, 19-SMG, rubusoside, and steviol AUC values in control UGT85C2 assays.**

	<b>13-SMG</b>			<b>19-SMG</b>			<b>rubusoside</b>			<b>Steviol</b>		
	20 min	40 min	19 h	20 min	40 min	19 h	20 min	40 min	19 h	20 min	40 min	19 h
Substrate: Steviol WT UGT85C2 (SEQ ID NO:7)	60635	67575	73750						490			
Substrate: 19-SMG WT UGT85C2 (SEQ ID NO:7)				53380	4635	1775	85560	108620	100300			
Substrate: Steviol No UGT85C2										53745	46585	54250
Substrate: 19-SMG No UGT85C2				224605	206230	199490						

**[00212]** Accumulation of 19-SMG and rubusoside was not observed in UGT85C2 variant activity assays using steviol as a substrate. Using steviol as the substrate, the F48H, F48Y,

F48T, I49V, S84A, and L92F UGT85C2 variants demonstrated high activity during incubation periods of under 40 min, and the F48H, F48Y, F48T, and I49V UGT85C2 variants demonstrated high activity during incubation periods of over 40 min (Table 8A). Using 19-SMG as the substrate, the F48H, F48Y, F48T, I49V, and S84A UGT85C2 variants demonstrated high activity during incubation periods of under 40 min, and the F48H, I49V, S84A, S84V, L91K, and L92F UGT85C2 variants, as well as the wild-type UGT85C2, demonstrated high activity during incubation periods of over 40 min (Table 8B). Slow conversion of steviol and 19-SMG was observed for UGT85C2 I87H (Tables 8A and 8B).

**[00213]** 13-SMG/rubusoside ratios were calculated for the UGT85C2 variants. A high 13-SMG/rubusoside ratio indicates preference of a UGT85C2 variant for steviol, whereas a low 13-SMG/rubusoside ratio indicates preference of a UGT85C2 variant for 19-SMG. The L91K, L91R, and L92F UGT85C2 variants demonstrated a high 13-SMG/rubusoside ratio, whereas the F48Y, F48T, P86G UGT85C2 variants demonstrated a low 13-SMG/rubusoside ratio.

**[00214]** The UGT85C2 variants were found to convert steviol to rubusoside after 24 h. Rubusoside levels (in AUC) are shown in Figure 5. Mutations in the amino acid 48 and 49 positions produced increased levels of rubusoside, as compared to the control. The variants with mutations in amino acids at position 86, 91 and 92 seem to produce lower levels of rubusoside.

### **Example 8: Evaluation of UGT76G1 Variants**

**[00215]** UGT76G1 variants were tested in a modified version of a steviol glycoside-producing *S. cerevisiae* strain as described in Example 2 to determine the effects on steviol glycosides, tri-glycosylated ent-kaurenol, and tri-glycosylated ent-kaurenoic acid levels. The background strain was described in Example 9 of WO 2014/122227, wherein both copies of UGT76G1 were deleted by homologous recombination using selective markers. The strain comprised a reintegrated wild-type UGT76G1 (WT control) or variants of UGT76G1 at the chromosome level.

**[00216]** Expression of UGT76G1 H155L (SEQ ID NO:184) increased the ratio of RebM/RebD produced, as compared to wild-type UGT76G1. Expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) in the strain all resulted in increased accumulation of ent-kaurenoic acid+2Glc (#7), 1,2-bioside, 1,2-stevioside, RebE, RebD, steviol+5Glc (#22), and steviol+6Glc (isomer 1), increased the ratio of RebD/RebM produced, and decreased

accumulation of RebB and RebA, as compared to wild-type UGT76G1. See Tables 9A-9C. Specifically, expression of UGT76G1 T146G (SEQ ID NO:183), resulted in increased accumulation of ent-kaurenoic acid+3Glc (isomer 1), steviol+3Glc (#1), and Stev3Glc (#34), as compared to wild-type UGT76G1. Expression of UGT76G1 L257G (SEQ ID NO:185) increased the amount of steviol+7Glc (isomer 2), as compared to wild-type UGT76G1. Expression of UGT76G1 S283N (SEQ ID NO:188) increased the amount of steviol+3Glc (#1) and Stev3Glc (#34), as compared to wild-type UGT76G1. See Tables 9A-9C.

**Table 9A. Accumulation of steviol glycosides (in  $\mu\text{M}$ ) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	13-SMG	1,2-bioside	RebB	RebA	RebE	RebD	RebM
<b>Wild-type</b> (SEQ ID NO:9)	13.5 $\pm$ 3.8	N/A	1.5 $\pm$ 0.4	4.7 $\pm$ 1.9	N/A	5.2 $\pm$ 2.5	29.3 $\pm$ 15.5
<b>H155L</b> (SEQ ID NO:184)	13.9 $\pm$ 2.4	N/A	1.8 $\pm$ 0.2	6.5 $\pm$ 1.5	N/A	2.1 $\pm$ 0.3	38.8 $\pm$ 12.6
<b>Q23H</b> (SEQ ID NO:181)	13.4 $\pm$ 2.2	1.8 $\pm$ 0.4	0.9 $\pm$ 0.1	1.3 $\pm$ 0.2	4.6 $\pm$ 0.6	17.7 $\pm$ 6.4	1.9 $\pm$ 0.7
<b>T146G</b> (SEQ ID NO:183)	13.9 $\pm$ 2.7	2.0 $\pm$ 0.4	0.6 $\pm$ 0.3	0.7 $\pm$ 0.5	7.4 $\pm$ 1.9	14.1 $\pm$ 3.5	1.1 $\pm$ 0.2
<b>L257G</b> (SEQ ID NO:185)	13.6 $\pm$ 0.9	1.2 $\pm$ 0.1	0.9 $\pm$ 0.2	2.3 $\pm$ 0.3	2.8 $\pm$ 0.4	32.0 $\pm$ 6.1	7.0 $\pm$ 1.5
<b>S283N</b> (SEQ ID NO:188)	13.5 $\pm$ 1.4	2.1 $\pm$ 0.4	0.5 $\pm$ 0.1	0.3 $\pm$ 0.5	7.9 $\pm$ 1.0	14.4 $\pm$ 3.9	0.9 $\pm$ 0.4
<b>Q23H+H155L</b> (SEQ ID NO:217)	12.4 $\pm$ 1.1	1.4 $\pm$ 0.3	0.8 $\pm$ 0.1	1.9 $\pm$ 0.5	4.0 $\pm$ 0.4	22.4 $\pm$ 5.9	8.4 $\pm$ 3.4
<b>T146G+H155L</b> (SEQ ID NO:218)	13.8 $\pm$ 1.3	1.4 $\pm$ 0.2	0.8 $\pm$ 0.1	2.2 $\pm$ 0.1	3.4 $\pm$ 0.4	26.5 $\pm$ 2.5	9.5 $\pm$ 1.9
<b>L257G+H155L</b> (SEQ ID NO:219)	14.1 $\pm$ 1.3	0.9 $\pm$ 0.4	1.0 $\pm$ 0.1	3.1 $\pm$ 0.5	1.8 $\pm$ 0.5	23.8 $\pm$ 5.2	15.9 $\pm$ 1.5
<b>S283N+H155L</b> (SEQ ID NO:220)	13.4 $\pm$ 2.6	2.3 $\pm$ 0.5	0.5 $\pm$ 0.3	0.3 $\pm$ 0.5	7.2 $\pm$ 1.8	10.1 $\pm$ 4.3	1.2 $\pm$ 0.6

**Table 9B. Accumulation of steviol glycosides, glycosylated ent-kaurenoic acid, or glycosylated kaurenol (in AUC) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	KA+2Glc (#7)	KA+3Glc (isomer 1)	KA+3Glc (isomer 2)	KL+2Glc (#8)	KL+3Glc (isomer 1 and isomer 2)	1,2-stevioside	steviol+3Glc (#1)
<b>Wild-type</b> (SEQ ID NO:9)	N/A	N/A	859 ± 1089	N/A	N/A	887 ± 668	N/A
<b>H155L</b> (SEQ ID NO:184)	N/A	N/A	1862 ± 1825	N/A	550 ± 1035	874 ± 754	N/A
<b>Q23H</b> (SEQ ID NO:181)	3118 ± 1068	592 ± 1165	N/A	N/A	N/A	6716 ± 966	466 ± 500
<b>T146G</b> (SEQ ID NO:183)	3109 ± 1441	1355 ± 951	N/A	N/A	N/A	8313 ± 1498	1243 ± 601
<b>L257G</b> (SEQ ID NO:185)	2562 ± 1267	1062 ± 1199	N/A	N/A	N/A	5716 ± 837	N/A
<b>S283N</b> (SEQ ID NO:188)	3872 ± 1086	1200 ± 1929	N/A	N/A	N/A	8572 ± 1325	1162 ± 644
<b>Q23H+H155L</b> (SEQ ID NO:217)	2690 ± 423	N/A	236 ± 668	N/A	N/A	6690 ± 734	110 ± 311
<b>T146G+H155L</b> (SEQ ID NO:218)	2416 ± 555	N/A	N/A	N/A	N/A	6172 ± 524	208 ± 385
<b>L257G+H155L</b> (SEQ ID NO:219)	1634 ± 1227	212 ± 600	1524 ± 1318	N/A	222 ± 628	5458 ± 1068	N/A
<b>S283N+H155L</b> (SEQ ID NO:220)	3886 ± 750	496 ± 929	N/A	408 ± 1154	N/A	8036 ± 1601	1118 ± 614

–KA:ent-kaurenoic acid

–KL:ent-kaurenol

**Table 9C. Accumulation of steviol glycosides (in AUC) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	steviol +3 Glc (#34)	steviol+4 Glc (#26)	steviol+4 Glc (#33)	steviol+5 Glc (#22)	steviol+5 Glc (#25)	steviol +6 Glc (isomer 1)	steviol+7 Glc (isomer 2)
<b>Wild-type</b> (SEQ ID NO:9)	N/A	2443 ± 1164	N/A	N/A	N/A	N/A	N/A
<b>H155L</b> (SEQ ID NO:184)	N/A	1020 ± 731	N/A	N/A	938 ± 1039	N/A	N/A
<b>Q23H</b> (SEQ ID NO:181)	472 ± 507	818 ± 726	N/A	19804 ± 4600	N/A	7350 ± 4013	N/A

NO:181)							
<b>T146G</b> (SEQ ID NO:183)	1262 ± 605	1509 ± 376	114 ± 302	38469 ± 8953	N/A	7365 ± 3483	N/A
<b>L257G</b> (SEQ ID NO:185)	104 ± 294	1038 ± 459	N/A	11638± 2268	N/A	10722 ± 1871	3870 ± 2463
<b>S283N</b> (SEQ ID NO:188)	1168 ± 655	1572 ± 625	104 ± 294	44460 ± 11455	N/A	12174 ± 5214	N/A
<b>Q23H+H15</b> <b>5L</b> (SEQ ID NO:217)	122 ± 345	964 ± 459	N/A	16600 ± 3617	N/A	4404 ± 2744	5230 ± 3262
<b>T146G+H1</b> <b>55L</b> (SEQ ID NO:218)	212 ± 383	1114 ± 192	N/A	14362 ± 1802	N/A	2498 ± 2743	4840 ± 2053
<b>L257G+H1</b> <b>55L</b> (SEQ ID NO:219)	N/A	782 ± 725	N/A	6354 ± 4578	N/A	2408 ± 2584	5780 ± 977
<b>S283N+H1</b> <b>55L</b> (SEQ ID NO:220)	1186 ± 673	1020 ± 739	N/A	38410 ± 17463	N/A	3864 ± 3520	N/A

**[00217]** The double UGT76G1 variants were also tested. The double variants were: UGT76G1 Q23H H155L (SEQ ID NO:217), UGT76G1 T146G H155L (SEQ ID NO:218), UGT76G1 L257G H155L (SEQ ID NO:219), and UGT76G1 S283N H155L (SEQ ID NO:220). Double variants UGT76G1 Q23H H155L (SEQ ID NO:217), UGT76G1 T146G H155L (SEQ ID NO:218), and UGT76G1 L257G H155L (SEQ ID NO:219) resulted in increased RebM accumulation, as compared to the three single variants UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 T146G (SEQ ID NO:183), and UGT76G1 L257G (SEQ ID NO:185). See Tables 9A-9C. Specifically, expression of UGT76G1 Q23H H155L (SEQ ID NO:217) increased the amount of RebM and steviol+7Glc (isomer 2), compared to the UGT76G1 Q23H (SEQ ID NO:181) variant. Expression of UGT76G1 T146G H155L (SEQ ID NO:218) increased accumulation of RebA, RebD, RebM, and steviol+7Glc (isomer 2) and decreased accumulation of ent-kaurenoic acid+3Glc (isomer1), 1,2-bioside, 1,2-stevioside, steviol+3Glc (#1), Stev3Glc (#34), RebE, and steviol+5Glc (#22), as compared to the UGT76G1 T146G (SEQ ID NO:183) variant. Expression of UGT76G1 L257G H155L (SEQ ID NO:219) increased accumulation of ent-kaurenoic acid+3Glc (isomer 2), RebA, and RebM and decreased accumulation of RebE and steviol+6Glc (isomer 1), as compared to the UGT76G1 L257G (SEQ ID NO:185) variant. See Tables 9A-9C. Thus, synergistic effects were observed for UGT76G1 double variants.

**[00218]** UGT76G1 variants were also analyzed in a modified version of the strain described above, which comprised a higher copy number of UGT91D2e (SEQ ID NO:10, SEQ ID NO:11), UGT74G1 (SEQ ID NO:3, SEQ ID NO:4), and ATR2 (SEQ ID NO:91, SEQ ID NO:92). Steviol glycoside-producing *S. cerevisiae* strains expressing UGT76G1 variants that resulted in increased RebD levels, including UGT76G1 Q23H, UGT76G T146G, and S283N, also increased accumulation of ent-kaurenoic acid+2Glc (#7) and ent-kaurenoic acid+2Glc (isomer 1) but decreased accumulation of ent-kaurenoic acid+3Glc (isomer 2), compared to steviol glycoside-producing *S. cerevisiae* strains expressing wild-type UGT76G1. See Figure 9A. UGT76G1 variants that increased RebD levels also increased accumulation of ent-kaurenol+2Glc (#8) but decreased accumulation of ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6) (Figure 9B).

**[00219]** Expression of the UGT76G1 H155L variant (SEQ ID NO:184), a variant that increased levels of RebM, resulted in decreased accumulation of ent-kaurenoic acid+2Glc (#7) and ent-kaurenoic acid+3Glc (isomer 1) (Figure 9A). Levels of ent-kaurenol glycosides were not significantly altered upon expression of UGT76G1 variants that increased levels of RebM, compared to strains expressing wild-type UGT76G1 (Figure 9B).

**[00220]** Levels of 13-SMG, 1,2-bioside, rubusoside, RebA, RebB, RebD, RebE, RebM, RebG (1,3-stevioside), steviol+3Glc (#1), steviol+4Glc (#26), steviol+5Glc (#22), steviol+5Glc (#24), steviol+5Glc (#25), steviol+6Glc (isomer 1), and steviol+6Glc (#23) produced in the steviol glycoside-producing strain are shown in Figures 10A-10C. Expression of UGT variants that resulted in increased RebD levels also increased accumulation of steviol+5Glc (#22), 1,2-stevioside, steviol+6Glc (isomer 1), and Stevio+3Glc (#1) but decreased accumulation of steviol+4Glc (#26), steviol+5Glc (#24), and RebG (1,3-stevioside) (Figure 10A). Expression of UGT76G1 H155L (SEQ ID NO:184) resulted in increased accumulation of steviol+5Glc (#25) but decreased accumulation of 1,2-stevioside, steviol+3Glc (#1), steviol+4Glc (#26), steviol+5Glc (#22), steviol+6Glc (isomer 1), and steviol+6Glc (#23) (Figure 10B). Expression of UGT76G1 S253W (SEQ ID NO:186) resulted in decreased accumulation of 1,2-stevioside and steviol+6Glc (isomer 1) (Figure 10B). Expression of UGT76G1 284G resulted in increased accumulation of 1,2-stevioside and steviol+6Glc (isomer 1) but decreased accumulation of RebG, steviol+4Glc (#26), steviol+5Glc (#25), and steviol+6Glc (#23) (Figure 10B). Figure 10C shows accumulation of 13-SMG, 1,2-bioside, rubusoside, RebA, RebB, RebD, RebE, and RebM in *S. cerevisiae* expressing wild-type UGT76G1 (SEQ ID NO:9) or a UGT76G1 variant that increases accumulation of RebD or RebM.

**[00221]** The steviol glycoside-producing strain comprising a higher copy number of UGT91D2e (SEQ ID NO:10, SEQ ID NO:11), UGT74G1 (SEQ ID NO:3, SEQ ID NO:4), and ATR2 (SEQ ID NO:91, SEQ ID NO:92) was further tested in a separate experiment. As shown in Tables 9D-9F, expression of UGT76G1 H155L (SEQ ID NO:184) resulted in increased accumulation of steviol+5Glc (#25), increased the ratio of RebM/RebD produced, and decreased accumulation of 1,2-bioside, steviol+3Glc (#1), RebE, steviol+6Glc (isomer 1), and steviol+6Glc (#23), as compared to wild-type UGT76G1. Expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) increased accumulation of 1,2-bioside, 1,2-stevioside, steviol+3Glc (#1), Stev+3Glc (#34), RebE, and steviol+5Glc (#22), increased the ratio of RebD/RebM produced, and decreased accumulation of RebG, RebA, steviol+5Glc (#25), steviol+7Glc (isomer 2), and steviol+7Glc (isomer 5). Specifically, expression of UGT76G1 Q23H (SEQ ID NO:181) resulted in increased accumulation of rubusoside, steviol+6Glc (isomer 1) and decreased accumulation of RebB and steviol+5Glc (#24). Expression of UGT76G1 T146G (SEQ ID NO:183) resulted in increased accumulation of rubusoside and decreased accumulation of RebB, steviol+5Glc (#24) and steviol+6Glc (#23). Expression of UGT76G1 L257G (SEQ ID NO:185) resulted in increased accumulation of steviol+6Glc (isomer 1). Expression of UGT76G1 S283N (SEQ ID NO:188) resulted in increased accumulation of rubusoside and decreased accumulation of RebB, steviol+5Glc (#24) and steviol+6Glc (#23). See Tables 9D-F.

**Table 9D. Accumulation of steviol glycosides (in  $\mu\text{M}$ ) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	13-SMG	1,2-bioside	Rubu	RebG	RebB	RebA	RebE	RebD	RebM
<b>Wild-type</b> (SEQ ID NO:9)	37.6 $\pm$ 8.8	1.3 $\pm$ 0.5	1.2 $\pm$ 0.2	0.2 $\pm$ 0.2	8.4 $\pm$ 2.3	32.5 $\pm$ 7.5	0.4 $\pm$ 0.1	30.4 $\pm$ 12.5	43.0 $\pm$ 9.6
<b>H155L</b> (SEQ ID NO:184)	35.3 $\pm$ 7.0	0.4 $\pm$ 0.1	1.3 $\pm$ 0.1	0.2 $\pm$ 0.2	8.9 $\pm$ 2.1	35.2 $\pm$ 9.3	0.1 $\pm$ 0.1	5.7 $\pm$ 1.8	64.5 $\pm$ 7.1
<b>Q23H</b> (SEQ ID NO:181)	40.8 $\pm$ 6.9	11.1 $\pm$ 1.5	2.4 $\pm$ 0.4	N/A	4.3 $\pm$ 1.3	7.2 $\pm$ 2.0	11.8 $\pm$ 4.5	35.1 $\pm$ 6.5	1.0 $\pm$ 0.4
<b>T146G</b> (SEQ ID NO:183)	41.4 $\pm$ 6.9	16.1 $\pm$ 1.4	3.1 $\pm$ 0.4	N/A	1.5 $\pm$ 0.5	2.4 $\pm$ 1.1	19.2 $\pm$ 3.2	15.0 $\pm$ 5.3	0.2 $\pm$ 0.2
<b>L257G</b> (SEQ ID NO:185)	32.4 $\pm$ 6.2	6.9 $\pm$ 1.0	1.8 $\pm$ 0.5	N/A	5.2 $\pm$ 1.8	12.1 $\pm$ 4.8	4.7 $\pm$ 1.6	41.7 $\pm$ 10.4	2.3 $\pm$ 0.9

<b>S283N</b> (SEQ ID NO:188)	39.8 ± 7.2	15.1 ± 2.8	2.6 ± 0.4	N/A	1.5 ± 0.5	2.9 ± 1.2	16.2 ± 4.8	19.2 ± 6.9	0.3 ± 0.1
<b>Q23H+H155L</b> (SEQ ID NO:217)	39.4 ± 4.5	9.0 ± 1.3	2.1 ± 0.2	N/A	4.7 ± 0.9	8.3 ± 2.6	8.8 ± 1.6	34.1 ± 4.5	3.0 ± 1.2
<b>T146G+H155L</b> (SEQ ID NO:218)	33.0 ± 8.0	8.5 ± 2.0	1.9 ± 0.7	N/A	3.8 ± 1.0	9.2 ± 2.9	6.6 ± 1.7	36.5 ± 4.7	3.1 ± 0.9
<b>L257G+H155L</b> (SEQ ID NO:219)	44.4 ± 6.6	4.9 ± 0.9	1.5 ± 0.3	N/A	8.2 ± 1.2	19.2 ± 4.0	3.4 ± 1.0	47.8 ± 4.5	12.3 ± 3.3
<b>S283N+H155L</b> (SEQ ID NO:220)	42.9 ± 6.6	14.5 ± 1.1	2.8 ± 0.2	N/A	2.1 ± 0.7	2.7 ± 0.9	16.7 ± 1.9	17.2 ± 3.7	0.7 ± 0.3

**Table 9E. Accumulation of steviol glycosides, glycosylated ent-kaurenoic acid, or glycosylated kaurenol (in AUC) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	<b>KA+2 Glc (#7)</b>	<b>KA+3 Glc (isomer 1)</b>	<b>KA+3 Glc (isomer 2)</b>	<b>KL+2 Glc (#8)</b>	<b>KL+3 Glc (isomer 1 and isomer 2)</b>	<b>19-SMG</b>	<b>1,3-bioside</b>	<b>1,2-stevio side</b>	<b>stevio I+3Glc (#1)</b>	<b>stevio I+3Glc (#34)</b>
<b>Wild-type</b> (SEQ ID NO:9)	14444 ± 5537	2472 ± 1360	47650 ± 20783	8102 ± 4937	12328 ± 20872	2174 ± 1054	274 ± 775	23410 ± 10331	2226 ± 1961	1512 ± 2135
<b>H155L</b> (SEQ ID NO:184)	1096 ± 1570	N/A	48264 ± 17847	1770 ± 1118	10590 ± 33369	2072 ± 940	N/A	13466 ± 2764	N/A	N/A
<b>Q23H</b> (SEQ ID NO:181)	14033 ± 26599	10386 ± 2233	2914 ± 2162	18346 ± 22523	53058 ± 11295	2364 ± 520	N/A	19950 ± 50824	21436 ± 6924	21436 ± 6924
<b>T146G</b> (SEQ ID NO:183)	15824 ± 18966	7339 ± 2016	N/A	26653 ± 21693	21515 ± 3812	1961 ± 1049	N/A	23720 ± 38885	27438 ± 6704	27438 ± 6704
<b>L257G</b> (SEQ ID NO:185)	11115 ± 39204	9732 ± 3604	7486 ± 3428	10014 ± 34855	67696 ± 22294	2010 ± 480	N/A	12374 ± 31888	13040 ± 2074	13070 ± 2086
<b>S283N</b> (SEQ ID NO:188)	14905 ± 55275	8722 ± 3756	N/A	22283 ± 63472	19864 ± 6586	1980 ± 875	N/A	20512 ± 58796	28660 ± 10712	28660 ± 10712
<b>Q23H+H155L</b> (SEQ ID NO:217)	10793 ± 18511	9230 ± 944	15348 ± 3586	86190 ± 13792	84080 ± 7629	2712 ± 674	N/A	16226 ± 12368	19104 ± 3180	19148 ± 3184
<b>T146G+H155L</b>	10414 ± 6	9346 ± 1964	13674 ± 4859	98980 ±	81762 ±	2034 ± 768	N/A	13851 ± 0	18846 ± 4723	18900

(SEQ ID NO:218)	17815			30306	19834			32208		± 4624
<b>L257G+H 155L</b> (SEQ ID NO:219)	68986 ± 17561	7974 ± 1665	34450 ± 6021	34730 ± 9050	99436 ± 7792	2800 ± 1291	N/A	118750 ± 15972	10356 ± 1814	10376 ± 1838
<b>S283N+H 155L</b> (SEQ ID NO:220)	14670 ± 15045	8168 ± 1243	1706 ± 1880	19180 ± 25165	31296 ± 6636	2694 ± 574	N/A	200156 ± 11694	25406 ± 6048	25406 ± 6048

–KA:ent-kaurenoic acid

–KL:ent-kaurenol

**Table 9F. Accumulation of steviol glycosides (in AUC) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	steviol+ 4Glc (#26)	steviol+ 5Glc (#22)	steviol+ 5Glc (#24)	steviol+ 5Glc (#25)	steviol+ 6Glc (isomer 1)	steviol+ 6Glc (#23)	steviol+ 7Glc (isomer 2)	steviol+ 7Glc (isomer 5)	Steviol
<b>Wild-type</b> (SEQ ID NO:9)	38936 ± 21188	3288 ± 3892	2194 ± 2020	9068 ± 3994	12294 ± 10105	5838 ± 2979	13784 ± 4806	7630 ± 3054	N/A
<b>H155L</b> (SEQ ID NO:184)	20000 ± 4629	178 ± 503	1530 ± 2310	29526 ± 15999	122 ± 345	2000 ± 830	6494 ± 2530	10782 ± 2519	N/A
<b>Q23H</b> (SEQ ID NO:181)	26366 ± 7357	161044 ± 57250	N/A	N/A	26590 ± 3671	3108 ± 1514	2964 ± 1547	918 ± 1268	N/A
<b>T146G</b> (SEQ ID NO:183)	25070 ± 6192	224315 ± 53331	N/A	N/A	10320 ± 3647	304 ± 804	322 ± 853	286 ± 756	N/A
<b>L257G</b> (SEQ ID NO:185)	17638 ± 5814	81252 ± 31941	258 ± 730	N/A	31616 ± 5164	5088 ± 1171	5154 ± 1398	1590 ± 1335	1246 ± 3524
<b>S283N</b> (SEQ ID NO:188)	24980 ± 8098	219964 ± 61935	N/A	N/A	19666 ± 5418	846 ± 1170	264 ± 747	296 ± 837	N/A
<b>Q23H+H 155L</b> (SEQ ID NO:217)	23100 ± 2234	142460 ± 24407	N/A	N/A	15108 ± 1958	3582 ± 819	5996 ± 1705	596 ± 1121	N/A
<b>T146G+H 155L</b> (SEQ ID NO:218)	19064 ± 3666	120990 ± 34224	N/A	N/A	13048 ± 2270	4288 ± 889	4640 ± 1866	1306 ± 1449	N/A

<b>L257G+ H155L</b> (SEQ ID NO:219)	17126 ± 2237	56416 ± 15937	928 ± 1293	N/A	17756 ± 2361	5856 ± 960	15114 ± 1900	2230 ± 985	N/A
<b>S283N+ H155L</b> (SEQ ID NO:220)	23536 ± 2818	213846 ± 31505	N/A	N/A	11222 ± 2649	1162 ± 1288	1042 ± 1117	N/A	N/A

**[00222]** Expression of UGT76G1 Q23H H155L (SEQ ID NO:217) increased accumulation of ent-kaurenoic acid+3Glc (isomer 2) and ent-kaurenol+3Glc (isomer 1) and decreased accumulation of ent-kaurenol+2Glc (#8) and steviol+6Glc (isomer 1), as compared to UGT76G1 Q23H (SEQ ID NO:181). UGT76G1 T146G H155L (SEQ ID NO:218) increased accumulation of ent-kaurenoic acid+3Glc (isomer 2), ent-kaurenol+3Glc (isomer 1), RebB, RebA, RebD, steviol+6Glc (#23), and steviol+7Glc (isomer 2) and decreased accumulation of ent-kaurenoic acid+2Glc (#7), ent-kaurenol+2Glc (#8), 1,2-bioside, rubusoside, 1,2-stevioside, RebE, steviol+5Glc (#22), as compared to UGT76G1 T146G (SEQ ID NO:183). Expression of UGT76G1 L257G H155L (SEQ ID NO:219) increased accumulation of ent-kaurenoic acid+3Glc (isomer 2), ent-kaurenol+3Glc (isomer 1), and steviol+7Glc (isomer 2) and decreased accumulation of ent-kaurenol+2Glc (#8), 1,2-bioside, and steviol+6Glc (isomer 1), as compared to UGT76G1 L257G (SEQ ID NO:185). As well, UGT76G1 L257G H155L (SEQ ID NO:219) increased accumulation of RebD, as compared to wild-type UGT76G1. Expression of UGT76G1 S283N H155L (SEQ ID NO:220) decreased accumulation of steviol+6Glc (isomer 1), as compared to UGT76G1 S283N (SEQ ID NO:188). See Tables 9D-F.

**[00223]** UGT76G1 variants were also expressed in a steviol glycoside-producing strain comprising an extra copy of CPR1 (SEQ ID NO:77, SEQ ID NO:78), an extra copy of SrKAHe1 (SEQ ID NO:93, SEQ ID NO:94), and an extra copy of a UGT76G1 (SEQ ID NO:8, SEQ ID NO:9) or a UGT76G1 variant. Accumulation of steviol glycosides, tri-glycosylated ent-kaurenol, and tri-glycosylated ent-kaurenoic acid levels were measured. See Figure 11.

**[00224]** UGT76G1 variants that increased accumulation of RebD or RebM were also expressed in a steviol glycoside production *S. cerevisiae* strain comprising an extra copy of CPR1 (SEQ ID NO:77, SEQ ID NO:78) and an extra copy of SrKAHe1 (SEQ ID NO:93, SEQ ID NO:94). The control steviol glycoside production strain comprised three copies of wild-type UGT76G1 (SEQ ID NO:9), and the variant-comprising strains comprised two copies of wild-type UGT76G1 (SEQ ID NO:9) and one copy of a UGT76G1 variant. Figure 11A shows levels of

ent-kaurenoic acid+2Glc (#7), ent-kaurenoic acid+3Glc (isomer 1), ent-kaurenoic acid+3Glc (isomer 2), ent-kaurenol+2Glc (#8), and ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6) in production strains expressing wild-type UGT76G1 (SEQ ID NO:9), UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 H155L (SEQ ID NO:184), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188). Total levels of glycosylated ent-kaurenoic acid (ent-kaurenoic acid+2Glc (#7) + ent-kaurenoic acid+3Glc (isomer 1) + ent-kaurenoic acid+3Glc (isomer 2)) were most significantly increased in production strains expressing UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), and UGT76G1 L257G (SEQ ID NO:185) (Figure 11B), and total levels of glycosylated ent-kaurenol (ent-kaurenol+3Glc (isomer 1) co-eluted with ent-kaurenol+3Glc (#6) and ent-kaurenol+2Glc (#8) were most significantly affected for production strains expressing UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), and UGT76G1 T146G (SEQ ID NO:183) (Figure 11C).

**[00225]** Figure 11D and 11E show accumulation of 1,2-bioside, 1,2-stevioside, steviol+3Glc (#1), steviol+4Glc (#26), steviol+5Glc (#22), steviol+5Glc (#24), steviol+5Glc (#25), steviol+6Glc (isomer 1), steviol+6Glc (#23), steviol+7Glc (isomer 2), steviol+7Glc (isomer 5), 13-SMG, rubusoside, RebG (1,3-stevioside), RebA, RebB, RebD, RebE, and RebM in production strains expressing wild-type UGT76G1 (SEQ ID NO:9), UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 H155L (SEQ ID NO:184), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188).

**[00226]** All UGT76G1 variants tested in Figure 11D showed decreased accumulation of steviol+4Glc (#26). Expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188), all of which increased production of RebD, resulted in decreased accumulation of steviol+5Glc (#25), compared to a control strain expressing wild-type UGT76G1 (Figure 11D). However, expression of the UGT76G1 H155L (SEQ ID NO:184) variant, which increased RebM production, resulted in increased accumulation of steviol+5Glc (#25) (Figure 11D).

**[00227]** Expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) resulted in increased accumulation of steviol+6Glc (#23), compared to a control strain expressing wild-type UGT76G1, whereas expression of the UGT76G1 H155L (SEQ ID NO:184) variant resulted in decreased accumulation of steviol+6Glc

(#23) (Figure 11D). Expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) resulted in increased accumulation of steviol+7Glc (isomer 2), compared to a control strain expressing wild-type UGT76G1, whereas expression of the UGT76G1 H155L (SEQ ID NO:184) variant resulted in decreased accumulation of steviol+7Glc (isomer 2) (Figure 11D). Expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 I26W (SEQ ID NO:182), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) resulted in increased accumulation of steviol+7Glc (isomer 5) (Figure 11D).

**[00228]** The steviol glycoside-producing strain comprising a higher copy number of CPR1 (SEQ ID NO:77, SEQ ID NO:78) and SrKAHe1 (SEQ ID NO:93, SEQ ID NO:94) was further tested in a separate experiment. As shown in Tables 9G-9I, expression of UGT76G1 H155L (SEQ ID NO:184) reduced the levels of ent-kaurenoic acid+3Glc (isomer 1), RebD, steviol+6Glc (#23), steviol+7Glc (isomer 2), as compared to wild-type UGT76G1. Expression of UGT76G1 Q23H (SEQ ID NO:181), UGT76G1 T146G (SEQ ID NO:183), UGT76G1 L257G (SEQ ID NO:185), or UGT76G1 S283N (SEQ ID NO:188) each reduced accumulation of steviol+4Glc (#26) and steviol+5Glc (#24), as compared to wild-type UGT76G1. Specifically, expression UGT76G1 T146G (SEQ ID NO:183) increased the amount of ent-kaurenoic acid+2Glc (#7), ent-kaurenoic acid+3Glc (isomer 1), RebD, steviol+6Glc (#23), and steviol+7Glc (isomer 2) and reduced the amount of RebG, steviol+5Glc #25, as compared to wild-type UGT76G1. Expression of UGT76G1 L257G (SEQ ID NO:185) increased accumulation of ent-kaurenoic acid+3Glc (isomer 1) and reduced accumulation of ent-kaurenoic acid+3Glc (isomer 2) and steviol+5Glc (#25), as compared to wild-type UGT76G1. Expression of UGT76G1 S283N (SEQ ID NO:188) increased accumulation of ent-kaurenoic acid+3Glc (isomer 1), RebD, steviol+6Glc (isomer 1), and steviol+7Glc (isomer 2) and reduced accumulation of RebG and steviol+5Glc (#25), as compared to wild-type UGT76G1. Expression of UGT76G1 L257G H155L reduced accumulation of ent-kaurenoic acid+3Glc (isomer 1), as compared to the single variant UGT76G1 L257G. Expression of the double variant UGT76G1 Q23H H155L reduced accumulation of steviol+5Glc (#25), as compared to wild-type UGT76G1. Expression of the double variant UGT76G1 S283N H155L reduced accumulation of ent-kaurenoic acid+3Glc (isomer 2), as compared to wild-type UGT76G1. See Tables 9G-9I.

**Table 9G. Accumulation of steviol glycosides (in  $\mu\text{M}$ ) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	13-SMG	1,2-bioside	Rubu	RebG	RebB	RebA	RebE	RebD	RebM
<b>Wild-type</b> (SEQ ID NO:9)	66.9 $\pm$ 4.7	0.4 $\pm$ 0.1	1.2 $\pm$ 0.2	0.7 $\pm$ 0.3	5.6 $\pm$ 0.4	30.3 $\pm$ 2.4	0.5 $\pm$ 0.4	31.0 $\pm$ 6.7	199.3 $\pm$ 14.2
<b>H155L</b> (SEQ ID NO:184)	63.1 $\pm$ 4.6	0.3 $\pm$ 0.1	1.3 $\pm$ 0.3	0.9 $\pm$ 0.3	5.5 $\pm$ 0.5	29.6 $\pm$ 1.9	0.1 $\pm$ 0.2	12.0 $\pm$ 10.8	210.0 $\pm$ 19.3
<b>Q23H</b> (SEQ ID NO:181)	62.2 $\pm$ 13.9	0.4 $\pm$ 0.1	0.8 $\pm$ 0.3	0.2 $\pm$ 0.3	5.2 $\pm$ 0.9	27.7 $\pm$ 3.3	0.6 $\pm$ 0.2	42.0 $\pm$ 9.8	179.2 $\pm$ 19.6
<b>T146G</b> (SEQ ID NO:183)	64.8 $\pm$ 5.2	0.5 $\pm$ 0.2	1.0 $\pm$ 0.1	0.1 $\pm$ 0.2	5.3 $\pm$ 0.8	27.9 $\pm$ 3.1	0.8 $\pm$ 0.1	46.2 $\pm$ 6.7	180.4 $\pm$ 24.2
<b>L257G</b> (SEQ ID NO:185)	68.7 $\pm$ 9.2	0.4 $\pm$ 0.1	0.6 $\pm$ 0.4	0.2 $\pm$ 0.3	5.5 $\pm$ 0.6	29.6 $\pm$ 3.4	0.6 $\pm$ 0.4	45.6 $\pm$ 9.3	187.3 $\pm$ 14.7
<b>S283N</b> (SEQ ID NO:188)	67.4 $\pm$ 13.3	0.4 $\pm$ 0.1	0.7 $\pm$ 0.5	0.1 $\pm$ 0.2	5.7 $\pm$ 0.7	32.0 $\pm$ 4.2	0.8 $\pm$ 0.4	52.7 $\pm$ 7.4	189.2 $\pm$ 14.1
<b>Q23H+H155L</b> (SEQ ID NO:217)	65.2 $\pm$ 4.3	0.3 $\pm$ 0.0	0.8 $\pm$ 0.4	0.3 $\pm$ 0.3	5.3 $\pm$ 0.3	27.1 $\pm$ 2.8	0.7 $\pm$ 0.3	37.5 $\pm$ 5.4	187.5 $\pm$ 10.8
<b>T146G+H155L</b> (SEQ ID NO:218)	64.3 $\pm$ 9.8	0.5 $\pm$ 0.1	0.8 $\pm$ 0.3	0.1 $\pm$ 0.2	5.4 $\pm$ 0.6	27.3 $\pm$ 4.3	0.7 $\pm$ 0.4	40.0 $\pm$ 8.7	171.2 $\pm$ 29.8
<b>L257G+H155L</b> (SEQ ID NO:219)	58.5 $\pm$ 15.9	0.3 $\pm$ 0.1	0.5 $\pm$ 0.5	0.3 $\pm$ 0.3	5.2 $\pm$ 1.5	25.1 $\pm$ 7.9	0.7 $\pm$ 0.3	30.4 $\pm$ 13.3	167.6 $\pm$ 33.6
<b>S283N+H155L</b> (SEQ ID NO:220)	61.2 $\pm$ 11.8	0.4 $\pm$ 0.1	0.6 $\pm$ 0.5	0.0 $\pm$ 0.0	5.2 $\pm$ 1.0	25.0 $\pm$ 5.5	0.6 $\pm$ 0.5	37.5 $\pm$ 12.0	152.5 $\pm$ 35.2

**Table 9H. Accumulation of steviol glycosides, glycosylated ent-kaurenoic acid, or glycosylated kaurenol (in AUC) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	KA+2Glc (#7)	KA+3Glc (isomer 1)	KA+3Glc (isomer 2)	KL+3Glc (isomer 1 and isomer 2)	19-SMG	1,2-stevioside	RebI	steviol+4Glc (#26)
<b>Wild-type</b> (SEQ ID NO:9)	2422 $\pm$ 419	1962 $\pm$ 383	40290 $\pm$ 3139	11500 $\pm$ 1169	422 $\pm$ 270	4712 $\pm$ 656	N/A	11194 $\pm$ 2466
<b>H155L</b> (SEQ	2894 $\pm$	418 $\pm$	40350 $\pm$	10326 $\pm$	376 $\pm$	4466 $\pm$	512 $\pm$	9086 $\pm$

ID NO:184)	401	841	2392	759	316	359	992	1374
<b>Q23H</b> (SEQ ID NO:181)	3340 ± 1018	3044 ± 747	41140 ± 5158	11404 ± 1306	476 ± 317	4452 ± 595	N/A	6550 ± 771
<b>T146G</b> (SEQ ID NO:183)	3362 ± 509	2934 ± 399	40636 ± 5193	10880 ± 872	400 ± 350	4600 ± 511	N/A	6996 ± 695
<b>L257G</b> (SEQ ID NO:185)	2816 ± 240	2712 ± 264	34402 ± 2377	10820 ± 708	254 ± 272	4770 ± 642	N/A	5884 ± 674
<b>S283N</b> (SEQ ID NO:188)	3114 ± 585	2914 ± 346	35830 ± 2929	11430 ± 641	188 ± 348	4986 ± 562	N/A	5734 ± 442
<b>Q23H+H155 L</b> (SEQ ID NO:217)	2622 ± 286	2250 ± 408	37176 ± 3860	10376 ± 1049	264 ± 283	4404 ± 416	N/A	6036 ± 906
<b>T146G+H155 L</b> (SEQ ID NO:218)	2884 ± 354	2424 ± 324	34100 ± 5312	10026 ± 1326	248 ± 347	4438 ± 1060	N/A	5836 ± 10777
<b>L257G+H155 L</b> (SEQ ID NO:219)	2364 ± 691	1798 ± 368	32044 ± 5509	9472 ± 1812	256 ± 363	3690 ± 1217	N/A	5254 ± 1189
<b>S283N+H155 L</b> (SEQ ID NO:220)	3162 ± 1250	2656 ± 980	31504 ± 4414	9386 ± 1425	384 ± 331	4014 ± 925	N/A	5638 ± 1696

–KA:ent-kaurenoic acid

–KL:ent-kaurenol

**Table 9I. Accumulation of steviol glycosides (in AUC) in a host comprising wild-type UGT76G1 or a UGT76G1 variant.**

	steviol+ 4Glc (#33)	steviol+ 5Glc (#22)	steviol+ 5Glc (#24)	steviol+ 5Glc (#25)	steviol+ 6Glc (isomer 1)	steviol+ 6Glc (#23)	steviol+ 7Glc (isomer 2)	steviol+ 7Glc (isomer 5)
<b>Wild-type</b> (SEQ ID NO:9)	N/A	N/A	7416 ± 1103	5230 ± 789	1572 ± 1044	3622 ± 590	7078 ± 912	4474 ± 2521
<b>H155L</b> (SEQ ID NO:184)	122 ± 345	N/A	7452 ± 2166	9450 ± 4068	320 ± 905	1868 ± 825	3894 ± 1243	4760 ± 1318
<b>Q23H</b> (SEQ ID NO:181)	N/A	108 ± 305	4382 ± 1490	3412 ± 1176	2792 ± 1053	4520 ± 985	9388 ± 1677	4158 ± 1528
<b>T146G</b> (SEQ ID NO:183)	N/A	114 ± 322	3598 ± 1630	2996 ± 745	3356 ± 1047	5438 ± 636	10406 ± 910	3700 ± 1726
<b>L257G</b>	N/A	N/A	4336 ±	3484 ±	2860 ±	4158 ±	9348 ±	4420 ±

(SEQ ID NO:185)			1158	754	842	1149	1429	1036
<b>S283N</b> (SEQ ID NO:188)	N/A	N/A	4834 ± 1338	3358 ± 546	3566 ± 784	4350 ± 909	9796 ± 1619	3924 ± 1203
<b>Q23H+H155L</b> (SEQ ID NO:217)	N/A	N/A	4468 ± 1172	3668 ± 679	1932 ± 380	3798 ± 619	8764 ± 1384	3528 ± 2244
<b>T146G+H155L</b> (SEQ ID NO:218)	N/A	N/A	3682 ± 1715	3008 ± 775	2176 ± 698	4022 ± 898	8712 ± 879	3284 ± 1803
<b>L257G+H155L</b> (SEQ ID NO:219)	N/A	N/A	3566 ± 1693	2974 ± 781	956 ± 1073	2988 ± 772	7046 ± 1660	3072 ± 1631
<b>S283N+H155L</b> (SEQ ID NO:220)	N/A	N/A	2670 ± 1807	2554 ± 444	2430 ± 1647	3874 ± 1837	9450 ± 3268	2758 ± 1204

#### Example 9: Further characterization of UGT76G1 H155L Variant

**[00229]** UGT76G1 H155L (SEQ ID NO:184) was expressed in the steviol glycoside-producing *S. cerevisiae* strain described in Examples 2 and 8. As shown in Figure 6A, the strain expressing UGT76G1 H155L (gray bars) produced higher levels of RebM, RebA, RebB, 13-SMG, and rubusoside, compared to the control strain expressing wild-type UGT76G1 (black bars). The steviol glycoside-producing strain expressing UGT76G1 H155L produced higher titers of RebM than RebD (Figure 6A).

**[00230]** The strain expressing UGT76G1 H155L (SEQ ID NO:184) produced greater total levels of steviol glycosides (13-SMG + 1,2-bioside + rubusoside + RebG + RebB + RebA + RebE + RebD + RebM) and RebD + RebM (gray bars), compared to the control strain expressing wild-type UGT76G1 (black bars) (Figure 6B). Thus, the steviol glycoside-producing strain expressing UGT76G1 H155L (gray bars) demonstrated a 20% increase in steviol glycoside production and a 10% increase in RebD and RebM titers, compared to the control strain expressing wild-type UGT76G1 (black bars) (Figure 6C).

**[00231]** The strain expressing UGT76G1 H155L (gray bars) also produced lesser amounts of a 1,2-bioside, 1,2-stevioside, a tri-glycosylated steviol molecule (steviol+3Glc (#1)), a penta-

glycosylated steviol molecule (steviol+5Glc (#22), two hexa-glycosylated steviol molecules (steviol+6Glc (isomer 1 and #23)), and a hepta-glycosylated steviol molecule (steviol+7Glc (isomer 2)) but increased amounts of a tetra-glycosylated molecule (steviol+4Glc (#26)) and two penta-glycosylated steviol molecules (Steviol+5Glc (#24 and #25)), compared to the control strain expressing wild-type UGT76G1 (black bars) (Figure 6D). See Figures 1, 7, and 8 for structures of particular steviol glycosides detected.

**[00232]** Having described the invention in detail and by reference to specific embodiments thereof, it will be apparent that modifications and variations are possible without departing from the scope of the invention defined in the appended claims. More specifically, although some aspects of the present invention are identified herein as particularly advantageous, it is contemplated that the present invention is not necessarily limited to these particular aspects of the invention.

**Table 10. Sequences disclosed herein.**

**SEQ ID NO:1**

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIIY	120
DYTHYWLPSI	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSECLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

**SEQ ID NO:2**

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIIY	120
DYTHYWLPSI	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFWDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

**SEQ ID NO:3**

atggcagagc	aacaaaagat	caaaaagtca	cctcacgtct	tacttattcc	atttctctg	60
caaggacata	tcaaccatt	catacaattt	gggaaaagat	tgattagtaa	gggtgtaaag	120
acaacactgg	taaccactat	ccacactttg	aattctactc	tgaaccactc	aaatactact	180
actacaagta	tagaaattca	agctatatca	gacggatgcg	atgaggggtg	ctttatgtct	240
gccggtgaat	cttacttga	aacattcaag	caagtgggat	ccaagtctct	ggccgatcta	300
atcaaaaagt	tacagagtga	aggcaccaca	attgacgcca	taatctacga	ttctatgaca	360
gagtggtgtt	tagacgttgc	tatcgaaatt	ggtattgatg	gaggttcctt	tttcacacaa	420
gcatgtgttg	tgaattctct	atactaccat	gtgcataaag	ggttaatctc	tttaccattg	480

ggtgaaactg	tttcagttcc	aggttttcca	gtgttacaac	gttgggaaac	cccattgatc	540
ttacaaaatc	atgaacaaat	acaatcacct	tggtcccaga	tgttgtttgg	tcaattcgct	600
aacatcgatc	aagcaagatg	ggtcctttact	aattcattct	ataagttaga	ggaagaggta	660
attgaatgga	ctaggaagat	ctggaatttg	aaagtcattg	gtccaacatt	gccatcaatg	720
tatttgga	aaagacttga	tgatgataaa	gataatggtt	tcaatttgta	caaggetaat	780
catcacgaat	gtatgaattg	gctggatgac	aaaccaaagg	aatcagttgt	atatgttgct	840
ttcggctctc	ttgttaaaca	tggtccagaa	caagttgagg	agattacaag	agcacttata	900
gactctgacg	taaacttttt	gtgggtcatt	aagcacaag	aggaggggaa	actgccagaa	960
aacctttctg	aagtgataaa	gaccggaaaa	ggtctaactg	ttgcttgggtg	taaacaattg	1020
gatgttttag	ctcatgaatc	tgtaggtgtg	tttgtaacac	attgcggtt	caactctaca	1080
ctagaagcca	tttccttagg	cgtacctgtc	gttgcaatgc	ctcagttctc	cgatcagaca	1140
accaacgcta	aacttttggg	cgaaatacta	ggggtgggtg	tcagagttaa	agcagacgag	1200
aatggtatcg	tcagaagagg	gaacctagct	tcatgtatca	aaatgatcat	ggaagaggaa	1260
agaggagtta	tcataaggaa	aaacgcagtt	aagtggaagg	atcttgcaaa	ggttgccgtc	1320
catgaaggcg	gctcttcaga	taatgatatt	gttgaatttg	tgtccgaact	aatcaaagcc	1380
taa						1383

SEQ ID NO:4

MAEQQKIKKS	PHVLLIPFPL	QGHINPFIQF	GKRLISKGVK	TTLVTTIHTL	NSTLNHSNTT	60
TTSIEIQAIS	DGCDEGGFMS	AGESYLETFK	QVGSKSLADL	IKKLQSEGTT	IDAIYDSMT	120
EWVLDVAIEF	GIDGGSFFTQ	ACVVNSLYYH	VHKGLISLPL	GETVSVPGFP	VLQRWETPLI	180
LQNHQIQSP	WSQMLFGQFA	NIDQARWVFT	NSFYKLEEEV	IEWTRKIWNL	KVIGPTLPSM	240
YLDKRLDDDK	DNGFNLYKAN	HHECMNWLDD	KPKESVYVVA	FGSLVKHGPE	QVEEITRALI	300
DSDVNFLWVI	KHKEEGKLP	NLSEVIKTGK	GLIVAWCKQL	DVLAHESVGC	FVTHCGFNST	360
LEAISLGPV	VAMPQFSDQT	TNAKLLDEIL	GVGVRVKADE	NGIVRRGNLA	SCIKMIMEEE	420
RGVIIRKNAV	KWKDLAKVAV	HEGGSSDNDI	VEFVSELIKA			460

SEQ ID NO:5

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caaagccaca	ttaaagccat	gctcaaaacta	gcacaacttc	tccaccacaa	aggactccag	120
ataaccttcg	tcaacaccga	cttcatccac	aaccagtttc	ttgaatcatc	gggccacatc	180
tgtctagacg	gtgcaccggg	tttccggttc	gaaaccattc	cggatggtgt	ttctcacagt	240
ccggaagcga	gcatcccaat	cagagaatca	ctcttgagat	ccattgaaac	caacttcttg	300
gatcgtttca	ttgatcttgt	aaccaaactt	ccggatcctc	cgacttgat	tatctcagat	360
gggttcttgt	cggttttcac	aattgacgct	gcaaaaaagc	ttggaattcc	ggtcatgatg	420
tattggacac	ttgtcgctg	tgggttcatg	ggtttttacc	atattcattc	tctcattgag	480
aaaggatttg	caccacttaa	agatgcaagt	tacttgacaa	atgggtattt	ggacaccgtc	540
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cccgaagaga	aaaagcaaac	tggaattacg	agtctccatg	gatacagttt	agtaaaagaa	840
gaaccagagt	gtttccagtg	gcttcagctc	aaagaaccaa	attccgtcgt	ttatgtaaat	900
tttggaaagta	ctacagtaat	gtctttagaa	gacatgacgg	aatttggttg	gggacttgct	960
aatagcaacc	attatttcct	ttggatcatc	cgatcaaact	tggtgatagg	ggaaaatgca	1020
gttttgcccc	ctgaacttga	ggaacatata	aagaaaagag	gctttattgc	tagctggtgt	1080
tcacaagaaa	aggtcttgaa	gcacccttcg	gttggagggg	tcttgactca	ttgtgggtgg	1140
ggatcgacca	tcgagagctt	gtctgctggg	gtgccaatga	tatgctggcc	ttattcgtgg	1200
gaccagctga	ccaactgtag	gtatatatgc	aaagaatggg	aggttgggct	cgagatggga	1260
accaaagtga	aacgagatga	agtcaagagg	cttgtaacaag	agttgatggg	agaaggaggt	1320
cacaaaatga	ggaacaaggc	taaagattgg	aaagaaaagg	ctcgcattgc	aatagctcct	1380
aacggttcat	cttctttgaa	catagacaaa	atggtcaagg	aaatcaccgt	gctagcaaga	1440
aactagttac	aaagttggtt	cacattgtgc	tttctattta	agatgtaact	ttgttctaata	1500
ttaatattgt	ctagatgtat	tgaaccataa	gtttagttgg	tctcaggaat	tgatttttaa	1560
tgaataaatg	gtcattaggg	gtgagt				1586

SEQ ID NO:6

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ataactttcg	tgaataccga	cttcatccat	aatcaatttc	tggaatctag	tggccctcat	180
tgtttggaag	gagccccagg	gtttagattc	gaaacaattc	ctgacgggtg	ttcacattcc	240
ccagaggcct	ccatcccaat	aagagagagt	ttactgaggt	caatagaaac	caactttttg	300
gatcgtttca	ttgacttggg	cacaaaactt	ccagaccac	caacttgcac	aatctctgat	360
ggctttctgt	cagtgtttac	tatcgacgct	gccaaaaagt	tgggtatccc	agttatgatg	420
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acagacctta	atgataaagt	attgatgttt	actacagaag	ctccacaaag	atctcataag	660
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tctctaagat	acaatcatal	ctacactatt	ggtccattac	aattacttct	agatcaaatt	780
cctgaagaga	aaaagcaaac	tggtattaca	tccttacacg	gctactcttt	agtgaagag	840
gaaccagaat	gttttcaatg	gctacaaagt	aaagagccta	attctgtggg	ctacgtcaac	900
ttcgggaagta	caacagtcac	gtccttggaa	gatatgactg	aatttgggtg	gggecttget	960
aattcaaadc	attactttct	atggattatc	aggccaatt	tggtaatagg	ggaaaacgcc	1020
gtattacctc	cagaattgga	ggaacacatc	aaaaagagag	gtttcattgc	ttcctgggtg	1080
tctcagaaa	aggtattgaa	acatccttct	gttggtgggt	tccttactca	ttgcggttgg	1140
ggctctacaa	tcgaatcact	aagtgcagga	gttccaatga	tttgttggcc	atattcatgg	1200
gaccaactta	caaattgtag	gtatatctgt	aaagagtggg	aagttggatt	agaaatggga	1260
acaaaggtta	aacgtgatga	agtgaaaaga	ttggttcagg	agttgatggg	ggaaggtggc	1320
cacaagatga	gaaacaaggc	caaagattgg	aaggaaaaag	ccagaattgc	tattgetcct	1380
aacgggtcat	cctctctaaa	cattgataag	atggtcaaag	agattacagt	cttagccaga	1440
aactaa						1446

SEQ ID NO:7

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSFS	PEASIPRES	LLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLME	TTEAPQRSBK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEHI	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:8

atggaaaaca	agaccgaaac	aacagttaga	cgtaggcgta	gaatcattct	gtttccagta	60
ccttttcaag	ggcacatcaa	tccaatacta	caactagcca	acgttttcta	ctctaaaggt	120
ttttctatta	caatctttca	caccaatttc	aacaaaccaa	aaacatccaa	ttaccacat	180
ttcacattca	gattcatact	tgataatgat	ccacaagatg	aacgtatttc	aaacttacct	240
accacgggac	ctttagctgg	aatgagaatt	ccaatcatca	atgaacatgg	tgccgatgag	300
cttagaagag	aattagagtt	acttatggtg	gcatecgaag	aggacgagga	agtctcttgt	360
ctgattactg	acgctctatg	gtactttgcc	caatctgtgg	ctgatagttt	gaatttgagg	420
agattggtac	taatgacatc	cagtctgttt	aaactttcacg	ctcatgttag	tttaccacaa	480
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SEQ ID NO:9

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FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFWLWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGYYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:10

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SEQ ID NO:11

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DYTHYWLPSI	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDRKVEK	EYVSQFVDYL	EKNARAVAI	HES	473

SEQ ID NO:12

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SEQ ID NO:13

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DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	MVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEALVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAI	HES	473

SEQ ID NO:14

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SEQ ID NO:15

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SEQ ID NO:16

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CADWVIVDVF	HHWAAAAALE	HKVPCAMMLL	GSAHMIASIA	DRRLERAETE	SPAAAGQGRP	180
AAAPTFEVAR	MKLIRTKGSS	GMSLAERFSL	TLRSSSLVVG	RSCVEFEPET	VPLLSTLRGK	240
PITFLGLMPP	LHEGRREDGE	DATVRWLDAQ	PAKSVVYVAL	GSEVPLGVEK	VHELALGLEL	300
AGTRFLWALR	KPTGVSDADL	LPAGFEERTR	GRGVVATRWV	PQMSILAHAA	VGAFALTHCGW	360
NSTIEGLMFG	HPLIMLPIFG	DQGPNARLIE	AKNAGLQVAR	NDGDGSFDRE	GVA AIRAVA	420
VEEESKVFQ	AKAKKLQEI	ADMACHERYI	DGFIQQLRSY	KD		462

SEQ ID NO:17

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CADWVIVDVF	HHWAAAAALE	HKVPCAMMLL	GSAHMIASIA	DRRLERAETE	SPAAAGQGRP	180
AAAPTFEVAR	MKLIRTKGSS	GMSLAERFSL	TLRSSSLVVG	RSCVEFEPET	VPLLSTLRGK	240
PITFLGLMPP	EIPGDEKDET	WVSIKKWLDG	KQKGSVVYVA	LGSEALVSQT	EVVELALGLE	300
LSGLPFVWAY	RKPKGPAKSD	SVELPDGFVE	RTRDRGLVWT	SWAPQLRILS	HESVCGFLTH	360
CGSGSIVEGL	MFGHPLIMLP	IFGDQPLNAR	LLEDKQVGIE	IARNDDGDSF	DREGVAAAIR	420
AVAVEEESK	VFQAKAKKLQ	EIVADMACHE	RYIDGFIQQL	RSYKD		465

SEQ ID NO:18

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FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	MVLKGSDCLL	SKCYHEFGTQ	WLPLETLHQ	240
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WNSTIEGLMF	GHPLIMLPIF	GDQGPNARLI	EAKNAGLQVP	RNEEDGCLTK	ESVARSLRSV	420
VVEKEGEIYK	ANARELSKIY	NDTKVEKEYV	SQFVDYLEKN	ARAVAIHDES		470

SEQ ID NO:19

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SEQ ID NO:20

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DALLSLSFEH	IATATKGVSK	DRIVRAIGEL	ARSVGSEGLV	AGQVVDILSE	GADVGLDHLE	240
YIHIHKTAML	LESSVIGAI	MGGGSDQQIE	KLRKFARSIG	LLFQVVDIL	DVTKSTEELG	300
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N						361

SEQ ID NO:21

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ggcaagtact	cactaacact	tattcatgcc	ctccaaactg	attcatccga	tctactgacc	960
aacatccttt	caatgagaag	agtgcaggga	aagttaacgg	cacaaaagag	atgttggttc	1020
tggaatga						1029

SEQ ID NO:22

MAEQQISNLL	SMFDASHASQ	KLEITVQMM	TYHYRETPD	SSSSEGGSL	RYDERRVSLP	60
LSHNAASPDI	VSQLCFSTAM	SSELNHRWKS	QRLKVDADPY	NYILTLPSKG	IRGAFIDSLN	120
VWLEVPEDET	SVIKEVIGML	HNSSLIIDDF	QDNSPLRRGK	PSTHTVFGPA	QAINATYVI	180
VKAIEKIQDI	VGHADALADVT	GTITTFQGG	AMDLWWTANA	IVPSIQEYLL	MVNDKTGALF	240
RLSLELLALN	SEASISDSAL	ESLSSAVSLL	GOYFQIRDDY	MNLIDNKYTD	QKGFCELDDE	300
GKYSLTLIHA	LQTDSSDLLT	NILSMRRVQG	KLTAQKRCWF	WK		342

SEQ ID NO:23

atggaaaaga	ctaaggagaa	agcagaacgt	atcttgctgg	agccatacag	atacttatta	60
caactaccag	gaaagcaagt	ccgttctaaa	ctatcacaag	cgttcaatca	ctggttaaaa	120
gttcttgaag	ataagttaca	aatcattatt	gaagtcacag	aaatgctaca	caatgcttct	180
ttactgatcg	atgatataga	ggattcttcc	aaactgagaa	gagggtttcc	tgtegcctcat	240
tccatatacg	gggtaccaag	tgtaatcaac	tcagctaatt	acgtctactt	cttgggattg	300
gaaaaagtat	tgacattaga	tcattccagac	gctgtaaagc	tattcaccag	acaacttctt	360
gaattgcatc	aaggtcaagg	tttgatatac	tattggagag	acacttatac	ttgccaaca	420
gaagaggagt	acaaagcaat	ggttctacaa	aagactggcg	gtttgttcgg	acttgccggt	480
ggtctgatgc	aacttttctc	tgattacaag	gaggacttaa	agcctctggt	ggataccttg	540
ggcttgtttt	tccagattag	agatgactac	gctaacttac	attcaaagga	atattcagaa	600
aacaaatcat	tctgtgaaga	tttgactgaa	gggaagttaa	gttttccaac	aatccacgcc	660
atgttggtcaa	gaccagaatc	tactcaagtg	caaaacattc	tgcgtcagag	aacagagaat	720
attgacatca	aaaagtattg	tgttcagtac	ttggaagatg	ttggttcttt	tgcttacaca	780
agacatacac	ttagagaatt	agaggcaaaa	gcatacaagc	aaatagaagc	ctgtggaggc	840
aatccttctc	tagtggcatt	ggttaaacaat	ttgtccaaaa	tgttcaccga	ggaaaacaag	900
taa						903

SEQ ID NO:24

MEKTKEKAER	ILLEPYRYLL	QLPGKQVRSK	LSQAFNHWLK	VPEDKLQIII	EVTEMLHNAS	60
LLIDDIEDSS	KLRGFPVAH	SIYGVPSVIN	SANYVYFLGL	EKVLTLDHPD	AVKLFTRQLL	120
ELHQGQGLDI	YWRDITYCPT	EEEYKAMVLQ	KTGGLFGLAV	GLMQLFSDYK	EDLKPLDRTL	180
GLFFQIRDDY	ANLHSKEYSE	NKSFCEDLTE	GKFSFPTIHA	IWSRPESTQV	QNILRQRTE	240
IDIKKYCVQY	LEDVGSFAYT	RHTLRELEAK	AYKQIEACGG	NPSLVALVKH	LSKMFTEENK	300

SEQ ID NO:25

atggcaagat	tctattttct	taacgcacta	ttgatggtta	tctcattaca	atcaactaca	60
gccttcactc	cagctaaaact	tgcttatcca	acaacaacaa	cagctctaaa	tgtegcctcc	120
gccgaaactt	ctttcagctc	agatgaatac	ttggcctcta	agataggacc	tatagagtct	180
gccttgggaag	catcagtcaa	atccagaatt	ccacagaccg	ataagatctg	cgaatctatg	240
gcctactctt	tgatggcagg	aggcaagaga	attagaccag	tggtgtgat	cgctgcatgt	300
gagatgttcg	gtggatccca	agatgtcgct	atgcctactg	ctgtggcatt	agaaatgata	360
cacacaatgt	cttgattcca	tgatgatttg	ccatccatgg	ataacgatga	cttgagaaga	420
ggtaaaccba	caaaccatgt	cgttttcggc	gaagatgtag	ctattcttgc	aggtgactct	480
ttattgtcaa	cttccttcga	gcacgtcgct	agagaaacaa	aaggagtgtc	agcagaaaag	540
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caagttatgg	acttagaatg	tgaagctaaa	ccaggtacca	cattagacga	cttgaaatgg	660
attcatatcc	ataaaaaccgc	tacattgtta	caagttgctg	tagcttctgg	tgcaattcta	720
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gcctttcaag	ttgccgacga	tatccttgat	gtaaccgctt	catcagaaga	tttgggtaaa	840
actgcaggca	aagatgaagc	tactgataag	acaacttacc	caaagttatt	aggattagaa	900
gagagtaagg	catacgcaag	acaactaatc	gatgaagcca	aggaaagttt	ggctcctttt	960
ggagatagag	ctgccccttt	attggccatt	gcagatttca	ttattgatag	aaagaattga	1020

SEQ ID NO:26

MARFYFLNAL	LMVISLQSTT	AFTPAKLAYP	TTTTALNVAS	AETSFSLDEY	LASKIGPIES	60
ALEASVKSRI	PQTDKICESM	AYSLMAGGKR	IRPVLCIAAC	EMFGGSQDVA	MPTAVALEMI	120
HTMSLIHDDL	PSMDNDDLRR	GKPTNHVVFV	EDVAILAGDS	LLSTSFHVA	RETKGVSAEK	180
IVDVIARLGL	SVGAEGLAGG	QVMDLECEAK	PGTTLDDLKW	IHIHKTATLL	QVAVASGAVL	240
GGATPEEVAA	CELFAMNIGL	AFQVADDILD	VTASSEDLGK	TAGKDEATDK	TTYPKLLGLE	300
ESKAYARQLI	DEAKESLAPF	GDRAAPLLAI	ADFIIDRKN			339

SEQ ID NO:27

atgcacttag	caccacgtag	agtccttaga	ggtagaagat	caccacctga	cagagttcct	60
gaaagacaag	gtgccttggg	tagaagacgt	ggagctggct	ctactggctg	tgcccgtgct	120
gctgctgggtg	ttcaccgtag	aagaggagga	ggcaggctg	atccatcagc	tgctgtgcat	180
agaggctggc	aagccgggtg	tggcaccggt	ttgctgatg	aggtgggtgc	taccgagcc	240
gccttagaaa	tgttcatgc	ttttgcttta	atccatgatg	atatcatgga	tgatagtga	300

actagaagag	gctcccaac	tgttcacaga	gccctagctg	atcgtttagg	cgctgctctg	360
gaccagatc	aggccggtca	actaggagtt	tctactgcta	tcttggttg	agatctggct	420
ttgacatggt	ccgatgaatt	gttatacgtt	ccattgactc	cacatagact	ggcagcagta	480
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gcagcttaca	caatggaacg	tccactgcac	attggtgcag	ccctggctgg	ggcaagacca	660
gaactattag	cagggctttc	agcatagccc	ttgccagctg	gagaagcctt	ccaattggca	720
gatgacctgc	taggcgtctt	cggtgatoca	agacgtacag	ggaaacctga	cctagatgat	780
cttagaggty	gaaagcatac	tgtcttagtc	gccttgccaa	gagaacatgc	cactccagaa	840
cagagacaca	cattggatac	attattgggt	acaccaggtc	ttgatagaca	aggecgttca	900
agactaagat	gcgtattggt	agcaactggt	gcaagagccg	aagccgaaag	acttattaca	960
gagagaagag	atcaagcatt	aactgcattg	aacgcattaa	cactgccacc	tcctttagct	1020
gaggcattag	caagattgac	attagggctc	acagctcatc	ctgcctaa		1068

SEQ ID NO:28

MHLAPRRVPR	GRRSPPDRVP	ERQALGRRR	GAGSTGCARA	AAGVHRRRG	GEADPSAAVH	60
RGWQAGGGTG	LPDEVVSTAA	ALEMFHAFAL	IHDDIMDDSA	TRRSPTVHR	ALADRLGAAL	120
DPDQAGQLGV	STAILVGDLA	LTWSEDELLYA	PLTPHRLAAV	LPLVTAMRAE	TVHGQYLDIT	180
SARRPGTDT	LALRIARYKT	AAYTMERPLH	IGAALAGARP	ELLAGLSAYA	LPAGEAFQLA	240
DDLLGVFGDP	RRTGKPDLLD	LRGGKHTVLV	ALAREHATPE	QRHTLDTLLG	TPGLDRQGAS	300
RLRCVLVATG	ARAEAEERLIT	ERRDQALTAL	NALTLPPPLA	EALARLTLGS	TAHPA	355

SEQ ID NO:29

atgtcatatt	tcgataacta	cttcaatgag	atagttaatt	ccgtgaacga	catcattaag	60
tcttacatct	ctggcgacgt	accaaaacta	tacgaagcct	cctaccattt	gtttacatca	120
ggagaaaga	gactaaagacc	attgatcctt	acaatttctt	ctgatctttt	cggtgacag	180
agagaaagag	catactatgc	tgccgcagca	atcgaagttt	tgcacacatt	cactttggtt	240
cacgatgata	tcatggatca	agataacatt	cgtagaggtc	ttcctactgt	acatgtcaag	300
tatggcctac	ctttggccat	tttagctggt	gacttattgc	atgcaaaagc	ctttcaattg	360
ttgactcagg	cattgagagg	tctaccatct	gaaactatca	tcaaggcgtt	tgatattctt	420
acaagatcta	tattatcat	atcagaaggt	caagctgtcg	atatggaatt	cgaagataga	480
attgatataca	aggaacaaga	gtatttggt	atgatattct	gtaaaaccgc	tgcttattc	540
tcagcttctt	cttccattgg	ggcgttgata	gctggagcta	atgataacga	tgtgagatta	600
atgtccgatt	tcggtacaaa	tcttgggatc	gcatttcaaa	ttgtagatga	tatacttgg	660
ttaacagctg	atgaaaaaga	gctaggaaaa	cctgttttca	gtgatatacag	agaaggtaaa	720
aagaccatat	tagtcattaa	gactttagaa	ttgtgtaagg	aagacgagaa	aaagattgtg	780
ttaaaagcgc	taggcaacaa	gtcagcatca	aaggaagagt	tgatgagttc	tgctgacata	840
atcaaaaagt	actcattgga	ttacgcctac	aacttagctg	agaaatacta	caaaaacgcc	900
atcgattctc	taaatcaagt	ttcaagtaaa	agtgatattc	caggaaggc	attgaaatat	960
cttgctgaat	tcaccatcag	aagacgtaag	taa			993

SEQ ID NO:30

MSYFDNYFNE	IVNSVNDI IK	SYISGDV PKL	YEASYHLF TS	GGKRLRPL IL	TISSDLFGGQ	60
RERAYYAGAA	IEVLHTFTLV	HDDIMDQDNI	RRGLPTVHV K	YGLPLAILAG	DLLHAKAFQL	120
LTQALRGLPS	ETI IKAFDIF	TRSII I ISEG	QAVDMEFEDR	IDIKEQEYLD	MISRKTAALF	180
SASSSIGALI	AGANDNDVRL	MSDFGTNLGI	AFQIVDDILG	LTADKEKELGK	PVFS DIREGK	240
KTILVIK TLE	LCKEDEKKIV	LKALGNKSAS	KEELMSSADI	IKKYSLDYAY	NLAEKYYKNA	300
IDSLNQVSSK	SDIPGKALKY	LA EFTIRRRK				330

SEQ ID NO:31

atggtcgcac	aaactttcaa	cctggataacc	tacttatccc	aaagacaaca	acaagttgaa	60
gaggccctaa	gtgctgctct	tgtgccagct	tatcctgaga	gaatatacga	agctatgaga	120
tactcctec	tggcaggtg	caaaagatta	agacctatct	tatgtttagc	tgcttgcgaa	180
ttggcaggtg	gttctgttga	acaagccatg	ccaactgcgt	gtgcacttga	aatgatccat	240
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aagccaacta	atcacaaggt	gttcggggaa	gatatagcca	tcttagcggg	tgatgcgctt	360
ttagcttacg	cttttgaaca	tattgcttct	caaacaagag	gagtaccacc	tcaattggty	420

ctacaagtta	ttgctagaat	cggacacgcc	gttgctgcaa	caggcctcgt	tggaggccaa	480
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tcacataaga	ctggagcctt	gctggaagca	tcagttgtct	caggcgggat	tctcgcaggg	600
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ggtaaagacc	aggcagccgc	aaaggcaact	tatccaagtc	tattgggttt	agaagcctct	780
agacagaaag	cggaaagagt	gattcaatct	gctaaggaag	ccttaagacc	ttacggttca	840
caagcagagc	cactcctagc	gctggcagac	ttcatcacac	gtcgtcagca	ttaa	894

SEQ ID NO:32

MVAQTFNLDT	YLSQRQQQVE	EALSAALVPA	YPERIYEAMR	YSLLAGGKRL	RPILCLAACE	60
LAGGSVEQAM	PTACALEMIH	TMSLIHDDL	AMDNDDFRRG	KPTNHKVFGE	DIAILAGDAL	120
LAYAFEHIAS	QTRGVPPQLV	LQVIARIGHA	VAATGLVGGQ	VVDLESEGKA	ISLETLEYIH	180
SHKTGALLEA	SVVSGGILAG	ADEELLARLS	HYARDIGLAF	QIVDDILDVT	ATSEQLGKTA	240
GKDQAAAKAT	YPSLLGLEAS	RQKAEELIQS	AKEALRPYGS	QAEPLLALAD	FITRRQH	297

SEQ ID NO:33

atgaaaaccg	ggtttatctc	accagcaaca	gtatttcate	acagaatctc	accagcgacc	60
actttcagac	atcacttate	acctgctact	acaaactcta	caggcattgt	cgccctaaga	120
gacatcaact	tcagatgtaa	agcagtttct	aaagagtact	ctgatctggt	gcagaaagat	180
gaggcttctt	tcacaaaatg	ggacgatgac	aaggtgaaag	atcatcttga	taccaacaaa	240
aacttatacc	caaatgatga	gattaaggaa	tttgttgaat	cagtaaaggc	tatgttcggt	300
agtatgaatg	acggggagat	aaacgtctct	gcatacgata	ctgcatgggt	tgctttgggt	360
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ccagtcgatt	tgtttgaaca	tatttggggt	gttgatagac	tgacagagatt	ggggattgcc	1020
agatacttca	aatcagagat	aaaagattgt	gtagagtata	tcaataagta	ctggaccaa	1080
aatggaatth	gttgggctag	aaatactcac	gttcaagata	tcgatgatac	agccatggga	1140
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aaagatggta	aattcgtttg	ctttgcaggg	caatcaacac	aagccgtgac	aggaatggtt	1260
aacgtttaca	gagcctctca	aatggtgttc	ccaggggaga	gaattttgga	agatgccaaa	1320
aagttctctt	acaattactt	aaaggaaaag	caaagtacca	acgaattgct	ggataaatgg	1380
ataatcgcta	aagatctacc	tggtgaagtt	ggttatgctc	tgatataccc	atggtatgct	1440
tccttaccaa	gattggaaac	tcgttattac	cttgaacaat	acggcgggtga	agatgatgct	1500
tgatagggca	agacattata	cagaatgggt	tacgtgtcca	ataacacata	tctagaaatg	1560
gcaaagctgg	attacaataa	ctatgttgca	gtccttcaat	tagaatggtg	cacaatacaa	1620
caatggtacg	tcgatattgg	tatagagaag	ttcgaatctg	acaacatcaa	gtcagtcctg	1680

SEQ ID NO:34

MKTGFISPAT	VFHHRISPAT	TFRHHLSPAT	TNSTGIVALR	DINFRCKAVS	KEYSDLLQKD	60
EASF TKWDDD	KVKDHLDTNK	NLYPNDEIKE	FVESVKAMFG	SMNDGEINVS	AYDTAWVALV	120
QVDVSGSPQ	FPSSLEWIAN	NQLSDGSWGD	HLLFSAHDRI	INTLACVIAL	TSWNVHPSKC	180
EKGLNFLREN	ICKLEDENAE	HMPIGFEVTF	PSLIDIAKKL	NIEVPEDTPA	LKEIYARRDI	240
KLTKIPMEVL	HKVPTLLHS	LEGMPDLEWE	KLLKLQCKDG	SFLFSPSSTA	FALMQTKDEK	300
CLOYLTNIIVT	KFNGGVPNVY	PVDLFEHIWV	VDRLQRLGIA	RYFKSEIKDC	VEYINKYWTK	360
NGICWARNTN	VQDIDDTAMG	FRVLRAHGYD	VTPDVFRQFE	KDGKFCVCFAG	QSTQAVTGMF	420
NLYRASQMLF	PERILEDAK	KFSYNYLKEK	QSTNELLDKW	IIAKDLPGEV	GYALDIPWYA	480
SLPRLETRY	LEQYGGEDDV	WIGKTLYRMG	YVSNNTYLEM	AKLDYNNYVA	VLQLEWYTIQ	540
QWYVDIGIEK	FESDNIKSVL	VSYYLAAASI	FEPERSKERI	AWAKTTILVD	KITSIFDSSQ	600

SSKEDITAFI	DKFRNKSSSK	KHSINGEPWH	EVMVALKKTLL	HGFALDALMT	HSQDIHPQLH	660
QAWEMWLTKL	QDGDVDTAEL	MVQMINMTAG	RWVSKELLTH	PQYQRLSTVT	NSVCHDITKL	720
HNFKENSTTV	DSKVQELVQL	VFSDTPPDDL	QDMKQTFLLTV	MKTFYYKAWC	DPNTINDHIS	780
KVFEIVI						787

SEQ ID NO:35

atgcctgatg	cacacgatgc	tccacctcca	caaataagac	agagaacact	agtagatgag	60
gctacccaac	tgctaactga	gtccgcagaa	gatgcatggg	gtgaagtcag	tgtgtcagaa	120
tacgaaacag	caaggctagt	tgccccatgct	acatggtttag	gtggacacgc	cacaagagtg	180
gccttccttc	tggagagaca	acacgaagac	gggtcatggg	gtccaccagg	tggatatagg	240
ttagtccta	cattatctgc	tgttcacgca	ttattgacat	gtcttgcttc	tcctgctcag	300
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agaagattgg	ggacatctga	ctccccacct	gatactatag	cagttgagct	ggttatcca	420
tctttgctag	agggcattca	acacttactg	gaccctgctc	atcctcatag	tagaccagcc	480
ttctctcaac	atagaggctc	tcttgtttgt	cctgggtggac	tagatgggag	aactctagga	540
gctttgagat	cacacgccgc	agcaggtaga	ccagtaccag	gaaaagtctg	gcacgcttcc	600
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gggtgctctg	ctgctgccac	agcaacatgg	ctaaccaggg	ttgcaccatc	tcaacagtca	720
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gggtgctcctg	ctggagcagg	attgcctcca	gatgctgatg	atacagccgc	tgtgttgcct	960
gcattggcaa	cacatgggag	aggtagaaga	ccagaagtac	tgatggatta	caggactgac	1020
gggtatttcc	aatgctttat	tggggaaagg	actccatcaa	tttcaacaaa	cgtcacgta	1080
ttggaacat	tagggcatca	tgtggcccaa	catccacaag	atagagccag	atacggatca	1140
gccatggata	ccgcatcagc	ttggctgctg	gcagctcaaa	agcaagatgg	ctcttggtta	1200
gataaatggc	atgcctcacc	atactacgct	actgtttgtt	gcacacaagc	cctagccgct	1260
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ttgtatactc	cagtaagagt	agtcagagct	gccagagctg	ctgctctgta	cactaccaga	1560
gatctattgt	taccaccatt	gtaa				1584

SEQ ID NO:36

MPDAHDAAPP	QIRQRTLVDL	ATQLLTESAE	DAWGEVSVSE	YETARLVAHA	TWLGGHATRV	60
AFLLERQHED	GSWGPPGGYR	LVPTLSAVHA	LLTCLASPAQ	DHGVPDRLL	RAVDAGLTAL	120
RRLGTSDSPP	DTIAVELVIP	SLLEGIQHLL	DPAHPHSRPA	FSQHRGSLVC	PGGLDGRTLG	180
ALRSHAAAGT	PVPGKVWHAS	ETLGLSTEEA	SHLQPAQGI	GGSAATATW	LTRVAPSQQS	240
DSARRYLEEL	QHRYSGPVPS	ITPITYFERA	WLLNNFAAAG	VPCEAPAALL	DSLEAALTPQ	300
GAPAGAGLPP	DADDTAAVLL	ALATHGRGRR	PEVLMYDRTD	GYFQCFIGER	TPSISTNAHV	360
LETLGHVVAQ	HPQDRARYGS	AMDTASAWLL	AAQKQDGSWL	DKWHASPYYA	TVCTQALAA	420
HASPATAPAR	QRAVRWVLAT	QRSDDGGWGLW	HSTVEETAYA	LQILAPPSGG	GNIPIVQQALT	480
RGRARLCGAL	PLTPLWHDKD	LYTPVRVVRA	ARAAALYTTR	DLLLPL		527

SEQ ID NO:37

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SEQ ID NO:38

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AELILPQFCG	EAAWLLGGVA	FPRHPALLPL	RQACLVKLGA	VAMLPSGHPL	LHSWEAWGTS	180
PTTACPDDDG	SIGISPAATA	AWRAQAVTRG	STPQVGRADA	YLQMASRATR	SGIEGVFPNV	240
WPINVFPCW	SLYTLHLAGL	FAHPALAEAV	RVIVAQLEAR	LGVHGLGPAL	HFAADADDTA	300
VALCVLHLAG	RDPAVDALRH	FEIGELFVTF	PGERNASVST	NIHALHALRL	LGKPAAGASA	360
YVEANRNPHG	LWDNEKWHVS	WLYPTAHAVA	ALAQGKQWR	DERALAALLQ	AQRDDGGWGA	420
GRGSTFEETA	YALFALHVMD	GSEEATGRRR	IAQVVARALE	WMLARHAAHG	LPQTPWIGK	480
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SEQ ID NO:39

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SEQ ID NO:40

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GRGLSFLGRN	MWKLATEDEE	SMPIGFELAF	PSLIELAKSL	GVHDFPYDHQ	ALQGIYSSRE	240
IKMKRIPKEV	MHTVPTSILH	SLEGMPGLDW	AKLLKLQSSD	GSFLFSPAAT	AYALMNTGDD	300
RCFSYIDRTV	KKFNGGVPNV	YPVDLFEHIW	AVDRLERLGI	SRYFQKEIEQ	CMDYVNRHWT	360
EDGICWARNS	DVKEVDDTAM	AFRLRLRHGY	SVSPDVFKNF	EKDGEFFAFV	GQSNQAVTGM	420
YNLNRASQIS	FPGEDVLHRA	GAFSYEFLRR	KEAEGALRDK	WIISKDLPGE	VVYTLDFPWY	480
GNLPRVEARD	YLEQYGGGDD	VWIGKTLYRM	PLVNNDVYLE	LARMDFNHCQ	ALHQLEWQGL	540
KRWYTENRLM	DFGVAQEDAL	RAYFLAAASV	YEPCAAERL	AWARAAILAN	AVSTHLRNSP	600
SFRERLEHSL	RCRPSEETDG	SWFNSSSGSD	AVLVKAVLRL	TDSLAREAQP	IHGDPEDI I	660
HKLLRSAAE	WVREKADAAD	SVCNGSSAVE	QEGSRMVHDK	QTCLLLARMI	EISAGRAAGE	720
AASEDGDRRI	IQLTGSICDS	LKQKMLVSQD	PEKNEEMMSH	VDDELKLRIR	EFVQYLLRLG	780
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SEQ ID NO:41

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SEQ ID NO:42

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PHQCNGKITF	FRENIGKLED	ENDEHMPIGF	EVAFPSLLEI	ARGINIDVPY	DSPVLKDIYA	240
KKELKLTRIP	KEIMHKIPTT	LLHSLEGMRD	LDWEKLLKLO	SQDGSFLFSP	SSTAFAFMQT	300
RDSNCLEYLR	NAVKRENGGV	PNVFPVDLFE	HIWIVDRLQR	LGISRYFEEE	IKECLDYVHR	360
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DIFQKWYEEN	RLSEWGVRRS	ELLECYYLAA	ATIFESERSH	ERMVWAKSSV	LVKAISSEFG	600
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FMSHGRDVNN	LLYLSWGDWM	EKWKLYGDEG	EGELMVKMII	LMKNNDLTNF	FTHTHFVRLA	720
EIINRICLPR	QYLKARRNDE	KEKTIKSMEK	EMGKMVELAL	SESDFTRDVS	ITFLDVAKAF	780
YFALCGDHL	QTHISKVLFQ	KV				802

SEQ ID NO:43

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SEQ ID NO:44

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LSKQTDfSLM	LHKRELEQKR	CHSNEMDGYL	AYISEGLGNL	YDWNMVKKYQ	MKNGSVFNSP	240
SATAAAFINH	QNPGLNYLN	SLLDKFGNAV	PTVYPHDLFI	RLSMVDTIER	LGISHHFRVE	300
IKNVLDETYR	CWVERDEQIF	MDVVTCALAF	RLLRINGYEV	SPDPLAEITN	ELALKDEYAA	360
LETYHASHIL	YQEDLSSGKQ	ILKSADFLKE	IISTDSNRLS	KLIHKEVENA	LKFPINTGLE	420
RINTRRNIQL	YNVDNTRILK	TTYHSSNISN	TDYLRLAVED	FYTCQSIYRE	ELKGLERWV	480
ENKLDQLKFA	RQKTAYCYFS	VAATLSSPEL	SDARISWAKN	GILTTVVDDF	FDIGGTIDEL	540
TNLIQCVEKW	NVDVDKDCCS	EHVRILFLAL	KDAICWIGDE	AFKWQARDVT	SHVIQTWLEL	600
MNSMLREAIW	TRDAYVPTLN	EYMENAYVSF	ALGPVVKPAI	YFVGPKLSEE	IVESSEYHNL	660
FKLMSTQGR	LNDIHSFKRE	FKEGKLNVA	LHLSNGESGK	VEEEVVEEMM	MMIKNKRKEL	720
MKLIFEENG	IVPRACKDAF	WNMCHVLNFF	YANDDGFTGN	TILDTVKDII	YNPLVLVNE	780
EEQR						784

SEQ ID NO:45

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ataatcgata	ctactaagga	gagaatccaa	aagctattca	aaaatgthtga	aatctcagta	180
tcactctatg	acaccgcatg	ggtthtgaatg	gtgccatcac	ctaattcccc	aaaaagtcca	240
tgthttccag	agtgcttgaa	ttggttaatc	aataatcagt	taaacgatgg	ttctthtgggt	300
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tgccattcta	acgaaattga	cgggtactta	gcataatct	cagaaggtht	gggthaattg	660
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tctgcaactg	ccgcagcatt	cattaatcat	caaaacctg	ggtgthtctaa	ctactthtgaac	780
tcactattag	ataagthtgg	aaatgcagtht	ccaacagtht	atcctthtgg	ctthtgcac	840
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aaattgtht	acaaggaagtht	agaaaacgca	ctaaagtht	ctattaacac	tggtthtgg	1260

agaatcaata	ctaggagaaa	cattcagctg	tacaacgtag	ataatacaag	gattccttaag	1320
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caaaacaagt	tggatcaact	gaagtttgct	agacagaaga	cagcatactg	ttattttctct	1500
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acaatattgg	atacagttaa	agatatcatc	tacaaccac	ttgttttggt	caatgagaac	2340
gaggaacaaa	gataa					2355

SEQ ID NO:46

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SSYDTAWVAM	VSPNSPKSP	CFPECLNWL	NNQLNDGSWG	LVNHTHNHNH	PLLKDSLSST	120
LACIVALKRW	NVGEDQINKG	LSFIESNLAS	ATDKSQPSPI	GFDIIFPGLL	EYAKNLDINL	180
LSKQTDfSLM	LHKRELEQKR	CHSNEIDGYL	AYISEGLGNL	YDWNMVKKYQ	MKNGSVFNSP	240
SATAAAFINH	QNPGLNYLN	SLLDKFGNAV	PTVYPLDLYI	RLSMVDTIER	LGISHHFRVE	300
IKNVLDETYR	CWVERDEQIF	MDVVTCALAF	RLLRIHGYKV	SPDQLAEITN	ELAFKDEYAA	360
LETYHASQIL	YQEDLSSGKQ	ILKSADFLKG	ILSTDSNRLS	KLIHKEVENA	LKFPINTGLE	420
RINTRRNIQL	YNVDNTRILK	TTYHSSNISN	TYYLRLAVED	FYTCQSIYRE	ELKGLERWV	480
QNKLDQLKFA	RQKTAYCYFS	VAATLSSPEL	SDARISWAKN	GILTTVVDDF	FDIGGTIDEL	540
TNLIQCVEKW	NVDVKDCCS	EHVRILFLAL	KDAICWIGDE	AFKWQARDVT	SHVIQWLEL	600
MNSMLREAIW	TRDAYVPTLN	EYMENAYVSF	ALGPIVKPAI	YFVGPKLSEE	IVESSEYHNL	660
FKLMSTQGR	LNDIHSFKRE	FKEGKLNVA	LHLSNGESGK	VEEEVVEEMM	MMIKNKRKEL	720
MKLIFEENG	IVPRACKDAF	WNMCHVLNFF	YANDDGFTGN	TILDTVKDII	YNPLVLVNN	780
EEQR						784

SEQ ID NO:47

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cctacccaaa	gatctacttc	ttcctctact	actagaccag	ctgccgaagt	gtcatcaggt	180
aagagtaaac	aacatgatca	ggaagctagt	gaagcgacta	tcagacaaca	attacaactt	240
gtggatgtcc	tggagaatat	gggaatatcc	agacattttg	ctgcagagat	aaagtgcata	300
ctagacagaa	cttacagatc	ttggttacia	agacacgagg	aatcatgctg	ggacactatg	360
acatgtgcta	tggcttttag	aatcctaaga	ttgaacggat	acaacgtttc	atcagatgaa	420
ctataccacg	ttgtagaggc	atctggtctg	cataattcct	tgggtgggta	tcttaacgat	480
accagaacac	tacttgaatt	acacaaggct	tcaacagtta	gtatctctga	ggatgaatct	540
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ggcgactga	gaaagccttc	tttattcaaa	gagggtgaac	atgactgga	tggacctttt	660
tacaccacac	ttgatagact	tcatcatagg	tggaaatattg	aaaacttcaa	cattattgag	720
caacacatgt	tggagactcc	atacttatct	aaccagcata	catcaaggga	tatcctagca	780
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tctagtcaag	tcgaaagagc	aaaagaggta	gacgctgtca	taaatgagcc	actgaagttg	1740
caaggttctc	atacactggt	atctgatggt	taa			1773

SEQ ID NO:48

MAMPVKLTPA	SLSLKAVCCR	FSSGGHALRF	GSSSLPCWRRT	PTQRSTSSST	TRPAAEVSSG	60
KSKQHDQEAS	EATIRQQQLQ	VDVLENMGIS	RHFAAEIKCI	LDRTYRSWLQ	RHEEIMLDTM	120
TCAMAFRILR	LNGYNVSSDE	LYHVVEASGL	HNSLGGYLND	TRTLLELHKA	STVSISEDES	180
ILDSIGSRSR	TLLREQLESG	GALRKPSLFF	EVEHALDGP	YTTLDRLHHR	WNIENFNIE	240
QHMLETPYLS	NQHTSRDILA	LSIRDFSSSQ	FTYQQELQHL	ESWVKECRLD	QLQFARQKLA	300
YFYLSAAGTM	FSEPLSDART	LWAKNGVLT	IVDDFFDVAG	SKEELENLVM	LVEMWDEHHK	360
VEFYSEQVEI	IFSSYDSVN	QLGEKASLVQ	DRSITKHLVE	IWL DLLKSMM	TEVEWRLSKY	420
VPTEKEYMIN	ASLIFGLGPI	VLPALYFVGP	KISESIVKDP	EYDELFLKMS	TCGRLLNDVQ	480
TFEREYNEGK	LNSVSLLVHL	GGPMSISDAK	RKLQKPIDTC	RRDLLSLVLR	EESVVRPCK	540
ELFWKMCKVC	YFFYSTTDGF	SSQVERAKEV	DAVINEPLKL	QGSHTLVSDV		590

SEQ ID NO:49

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SEQ ID NO:50

MSCIRPWFCP	SSISATLTDP	ASKLVTGEFK	TTSLNFHGTK	ERIKKMFDKI	ELSVSSYDTA	60
WVAMVPSPDC	PETPCFPECT	KWILENQLGD	GSWSLPHGNP	LLVKDALSSST	LACILALKRW	120
GIGEEQINKG	LRFIELNSAS	VDNEQHKPI	GFDIIFPGMI	EYAKDLDLNL	PLKPTDINSM	180
LHRALELTS	GGGNLEGRR	AYLAYVSEGI	GKLQDWEMAM	KYQRKNGSLF	NSPSTTAAAF	240
IHIQDAECLH	YIRSLLOKFG	NAVPTIYPLD	IYARLSMVDA	LERLGIDRFH	RKERKFVLDE	300
TYRFLWQGEE	EIFSDNATCA	LAFRILRLNG	YDVSLEDHFS	NSLGGYLKDS	GAALELYRAL	360
QLSYPDESLL	EKQNSRYSYF	LKQGLSNVSL	CGDRLRKNI I	GEVHDALNFP	DHANLQRLAI	420
RRRIKHYATD	DTRILKTSYR	CSTIGNQDFL	KLAVEDFNIC	QSIQREEFKH	IERWVVERRL	480
DKLKFARQKE	AYCYFSAAAT	LFAPELSDAR	MSWAKNGVLT	TVVDDFFDVG	GSEELVNLI	540
ELIERWDVNG	SADFCSEEVE	IIYSAIHSTI	SEIGDKSFGV	QGRDVKSHVI	KIWL DLLKSM	600
LTEAQWSSNK	SVPTLDEYMT	TAHVSFALGP	IVLPALYFVG	PKLSEEVAGH	PELLNLYKVM	660
STCGRLLNDW	RSFKRESEEG	KLNAISLYMI	HSGGASTEVE	TIEHFKGLID	SQRRQLQLV	720
LQEKDSI IPR	PCKDLFWNMI	KLLHTFYMKD	DGFTSNEMRN	VVKAIINEPI	SLDEL	775

SEQ ID NO:51

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aaagaatctt	taacttga					2358

SEQ ID NO:52

MSINLRSSGC	SSPISATLER	GLDSEVQTRA	NNVSFEQTK	KIRKMLEKVE	LSVSAYDTSW	60
VAMVPSRSSQ	NAPLFPQCVK	WLLDNQHEDG	SWGLDNHDHQ	SLKKDVLSS	LASILALKKW	120
GIGERQINKG	LQFIELNSAL	VTDETIQKPT	GFDIIFPGMI	KYARDLNLI	PLGSEVVDDM	180
IRKRDLKLC	DSEKFSKGRE	AYLAYVLEGT	RNLKDWDLIV	KYQRKNGSLF	DSPATTAAAF	240
TQFGNDGCLR	YLCSLLQKFE	AAVPSVYPFD	QYARLSIIVT	LES LGIDRDF	KTEIKSILDE	300
TYRYWLRGDE	EICLDLATCA	LAFRLLLAHG	YDVSYDPLKP	FAEESGFSDT	LEGYVKNFTS	360
VLELFKAAQS	YPHESALKKQ	CCWTKQYLEM	ELSSWVKTSV	RDKYLKKEVE	DALAFPSYAS	420
LERSDHRRI	LNGSAVENTR	VTKTSYRLHN	ICTSDILKLA	VDDFNFCQSI	HREEMERLDR	480
WIVENRLQEL	KFARQKLAYC	YFSGAATLFS	PELSDARISW	AKGGVLTTVV	DDFFDVGGSK	540
EELENLIHLV	EKWDLNGVPE	YSSEHVEIIF	SVLRDTILET	GDKAFTYQGR	NVTHHIVKIW	600
LDLLKSMLE	AEWSSDKSTP	SLEDYMENAY	ISFALGPIVL	PATYLDIGPPL	PEKTVDSHQY	660
NQLYKLVSTM	GRLNDIQGF	KRESAEGKLN	AVSLHMKHER	DNRSKEVIEE	SMKGLAERKR	720
EELHKLVL	KGSVVPRECK	EAFKMSKVL	NLFYRKDDGF	TSNDLMSLVK	SVIYEPVSLQ	780
KESLT						858

SEQ ID NO:53

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SEQ ID NO:54

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RPEYLYGKQP	MTALHSLEAF	IGKIDFDKVR	HHRTHGSMMG	SPSSTAAYLM	HASQWDGDSE	240
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SFEKEGGAIG	YAPGFQADVD	DTAKTISTLA	VLGRDATPRQ	MIKVFETHANTH	FRTYPERDP	360
SLTANCNALS	ALLHQPDAAAM	YGSQIQKITK	FVCDYWWKSD	GKIKDKWNTC	YLYPSVLLVE	420
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RSPEYDLVFS	ALSTFTKHVL	QHPSIQSASV	WDRKLLARE	KAYLLAHIQQ	AEDSTPLSEL	780
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SEQ ID NO:55

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SEQ ID NO:56

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IIDNQLPDGD	WGEPSLFLGY	DRVCNTLACV	IALKTWGVGA	QNVERGIQFL	QSNIYKMEED	240
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SEQ ID NO:57

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SEQ ID NO:58

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LERMHGEKLG	HFDLEQVYVK	PSSLLHSLEA	FLGKLDLDFRL	SHHLYHGSMM	ASPSSTAAYL	240
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IESSFFVPLL	QAQRVEIYPR	DNIKVDEDKY	LSIIPFTWVG	CNNRSRTFAS	NRWLYDMMYL	660
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SEQ ID NO:59

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ctgaaaatca	ctatgaatag	agacgaaatc	tttcaagtcc	ttgttggtga	tccaatgatg	720
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aagttcgaaa	atactattca	acaaatgtac	atcagaagag	aagctgttat	gaaatcttta	840
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cttttatctg	aagctcaaac	tttaaccgat	cagcaactat	tgatgtcctt	gtgggaacca	960
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ggttccttgc	aagccctttt	aactgcatct	attgggattg	ggagaatggg	tcaagagttc	1440
gaatggaaac	tgaaggat	gactcaagag	gaagtgaaca	cgataggcct	aactacaaa	1500
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SEQ ID NO:60

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KALKVLTADK	TMVAMSDYDD	YHKTVKRHIL	TAVLGPNAQK	KHRIHRDIMM	DNISTQLHEF	180
VKNNPEQEEV	DLRKIFQSEL	FGLAMRQALG	KDVESLYVED	LKITMNRDEI	FQVLVDPMM	240
GAIDVDWRDF	FPYLKWPVKN	KFENTIQQMY	IRREAVMKSL	IKEHKKRIAS	GEKLNSYIDY	300
LLSEAQTLTD	QQLLMSLWEP	IIESSDITMV	TTEWAMYELA	KNPKLQDRLY	RDIKSVCGSE	360
KITEEHLSQL	PYITAFHET	LRRHSPVPII	PLRHVHEDTV	LGGYHVPAGT	ELAVNIYGCN	420
MDKNVWENPE	EWNPERFMKE	NETIDFQKTM	AFGGGKRVCA	GSLQALLTAS	IGIGRMVQEF	480
EWKLDMTQE	EVNTIGLTTQ	MLRPLRAIIK	PRI			513

SEQ ID NO:61

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tcctacgctt	ccccatctca	tcatttcaat	catttgccac	cagtacctga	agttccaggt	180
gttccagttt	tgggtaattt	gttgcaattg	aaagaaaaaa	agccttacat	gaccttcacc	240
aagtgggctg	aaatgtatgg	tccaatctac	tctattagaa	ctgggtctac	ttccatgggt	300
gttgtctctt	ctaacgaaat	cgccaaagaa	gttgttgta	ccagattccc	atctatctct	360
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ccgcgg						1566

SEQ ID NO:62

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FEKNPNQEVN	LKIFQSQLE	GLAMKQALGK	DVESIYVKDL	ETTMKREEIF	EVLVVDPMMG	240
AIEVDWRDFF	PYLKWPKNKS	FENI IHRMYT	RREAVMKALI	QEHKKRIASG	ENLNSYIDYL	300
LSEAQTLTDK	QLLMSLWEP I	IESSDTTMVT	TEWAMYELAK	NPNMQDRLYE	EIQSVCGSEK	360
ITEENLSQLP	YLYAVFQETL	RKHCPVPIMP	LRYVHENTVL	GGYHVPAGTE	VAINIYGCNM	420
DKKVWENPEE	WNPERFLSEK	ESMDLYKTMA	FGGKRVKAG	SLQAMVISCI	GIGRLVQDFE	480
WKLKDDAEED	VNTLGLTTQK	LHPLLLALINP	RK			512

SEQ ID NO:63

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cagaagaagc	gaattgcttc	aggagaggaa	atcaactggt	atctcgactt	cttgcttaag	900
gaagggaaga	cactgacaat	ggaccaaata	agtatgttgc	tttgggagac	ggttattgaa	960
acagcagata	ctacaatggt	aacgacagaa	tgggctatgt	atgaagttgc	taaagactca	1020
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cctatggatt	tgtacaagac	catggctttt	ggggctggaa	agaggggatg	tgctggttct	1380
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aagctgagag	atggagaaga	agaaaatgta	gatactgttg	ggctcaccac	tcacaaacgc	1500
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SEQ ID NO:64

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SEQ ID NO:65

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SEQ ID NO:66

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SKALELLTSN	KSMVATSDYN	EFHKMVKKYI	LAELLGANAQ	KRHRIHRDTL	IENVLNKLHA	180
HTKNSPLQAV	NFRKIFESEL	FGLAMQALG	YDVDSLVEE	LGTTLSREEI	YNVLVSDMLK	240
GAIEVDWRDF	FPYLKWIPNK	SFEMKIQRLA	SRRQAVMNSI	VKEQKKSIA	GKGENCYLNY	300
LLSEAKTLTE	KQISILAWET	I IETADTTVV	TTEWAMYELA	KNPKQDRLY	NEIQNVCSTD	360
KITEEHLTKL	PYLSAVFHE	LRKYSPLV	PLRYAHEDTQ	LGGYVVPAGT	EIAVNIYGCN	420
MDKNQWETPE	EWKPERFLDE	KYDPMDMYKT	MSFGSGKRV	AGSLQASLIA	CTSIGRLVQE	480
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SEQ ID NO:67

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SEQ ID NO:68

MASMISLLLG	FVVSFLFIF	FLKLLFFFS	RHKMSEVSR	PSVPVPGFPL	IGNLLQLKEK	60
KPHKTFTKWS	ELYGPIYSIK	MGSSSLIVLN	SIETAKEAMV	SRFSSISTRK	LSNALTVLTC	120
NKSMVATSDY	DDFHKFVKRC	LLNGLLGANA	QERKRHYRDA	LIENVTSKLN	AHTRNHPQEP	180
VNFRAIFEHE	LFGVALKQAF	GKDVESIYVK	ELGVTLSRDE	IFKVLVHDM	EGAIDVDWRD	240
FFPYLKWIPN	NSFEARIQOK	HKRRLAVMNA	LIQDRLNQND	SESDDDCYLN	FLMSEAKTLT	300
MEQIAILVWE	TIIETADTTL	VTEWAMYEL	AKHQSVQDRL	FKEIQSVC GG	EKIKEEQLPR	360
LPYVNGVFHE	TLRKYSAPL	VPIRYAHEDT	QIGGYHIPAG	SEIAINIYGC	NMDKKRWERP	420
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SEQ ID NO:69

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ttgactgaag	aaaacttgtc	caagttgcc	tacttgaact	ctgttttcca	cgaaaccttg	1140
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ggtggttacc	atattccagc	tggttctcaa	attgccatta	acatctacgg	ttgcaacatg	1260
aacaaaaagc	aatgggaaaa	tctgaagaa	tggaagccag	aaagattctt	ggacgaaaag	1320
tatgacttga	tggacttgc	taagactatg	gcttttggtg	gtggtaaaag	agtttgtgct	1380
ggtgctttac	aagcaatggt	gattgcttgc	acttccatcg	gtagattcgt	tcaagaattt	1440
gaatggaagt	tgatgggtgg	tgaagaagaa	aacgttgata	ctgttgcttt	gacctcccaa	1500
aaattgcatc	caatgcaagc	cattattaag	gccagagaat	gactcgagcc	gcgg	1554

SEQ ID NO:70

MDDMGIEAVP	FATAVVLGGI	SLVVLIFIRR	FVSNRKRSVE	GLPPVPDIPG	LPLIGNLLQL	60
KEKKPHKTFA	RWAETYGPIF	SIRTGASTMI	VLNSSEVAKE	AMVTRFSSIS	TRKLSNALKI	120
LTFDKCMVAT	SDYNDFHKMV	KGFILRNVLG	APAQKRHRCH	RDTLIENISK	YLHAHVKTSP	180
LEPVVLKKIF	ESEIFGLALK	QALGKDIESI	YVEELGTTLS	REEIFAVLVV	DPMAGAIEVD	240
WRDFFPYLSW	IPNKS MEMKI	QRMDFRRGAL	MKALIGEQQK	RIGSGEEKNS	YIDFLLSEAT	300
TLTEKQIAML	IWETIIEISD	TTLVTSEWAM	YELAKDPNRQ	EILYREIHKV	CGSNKLT EEN	360
LSKLPYLNSV	FHETLRKYSP	APMVPVRYAH	EDTQLGGYHI	PAGSQIAINI	YGCNMNKKQW	420
ENPEEWKPER	FLDEKYDLMD	LHKTMAFGGG	KRVCAGALQA	MLIACTSIGR	FVQEF EWKLM	480
GGEEENVDTV	ALTSQKLHPM	QAI IKARE				508

SEQ ID NO:71

aagcttaaaa	tgagtaagtc	taatagtatg	aattctacat	cacacgaaac	cctttttcaa	60
caattggtct	tgggtttgga	ccgtatgcc	ttgatggatg	ttcactgggt	gatctacggt	120
gctttcggcg	catggttatg	ttcttatgtg	atacatgttt	tatcatcttc	ctctacagta	180
aaagtgccag	ttgttgata	caggtctgta	ttcgaacct	catggttgct	tagacttaga	240
ttcgtctggg	aaggtggctc	tatcatagg	caaggttaca	ataagttta	agactctatt	300
ttccaagtta	ggaaattggg	aactgatatt	gtcattatac	cacctaact	tattgatgaa	360
gtgagaaaa	tgtcacagga	caagactaga	tcagttgaac	ctttcattaa	tgattttgca	420
ggtcaataca	caagaggcat	ggttttcttg	caatctgact	tacaaaaccg	tggtatacaa	480
caaagactaa	ctccaaaatt	ggtttccttg	accaaggtca	tgaaggaaga	gttggattat	540
gctttaacaa	aagagatgcc	tgatatgaaa	aatgacgaat	gggtagaagt	agatatcagt	600
agtataatgg	tgagattgat	ttccaggatc	tccgccagag	tctttctagg	gcctgaacac	660
tgctgtaacc	aggaatggtt	gactactaca	gcagaatatt	cagaatcact	tttcattaca	720
gggtttatct	taagagttgt	acctcatatc	ttaagaccat	tcatcgcccc	tctattacct	780
tcatacagga	ctctacttag	aaacgtttca	agtggtagaa	gagtcacg	tgacatcata	840
agatctcagc	aaggggatgg	taacgaagat	atactttcct	ggatgagaga	tgctgccaca	900
ggagaggaaa	agcaaatcga	taacattgct	cagagaatgt	taattctttc	tttagcatca	960
atccacacta	ctgcgatgac	catgacacat	gccatgtacg	atctatgtgc	ttgcctgag	1020
tacattgaac	cattaagaga	tgaagttaaa	tctgttggtg	gggcttctgg	ctgggacaag	1080
acagcgtaa	acagatttca	taagttggac	tccttctctaa	aagagtcaca	aagattcaac	1140
ccagatattct	tattgacatt	caatagaatc	taccatcaat	ctatgacctt	atcagatggc	1200
actaacattc	catctggaac	acgtattgct	gttccatcac	acgcaatggt	gcaagattct	1260
gcacatgtcc	caggtccaac	cccacctact	gaatttgatg	gattcagata	tagtaagata	1320
cgttctgata	gtaactacgc	acaaaagtac	ctattctcca	tgaccgattc	ttcaaacatg	1380
gctttcggat	acggcaagta	tgcttggtcca	ggtagatttt	acgcgtctaa	tgagatgaaa	1440
ctaacattag	ccattttggt	gctacaattt	gagttcaaac	taccagatgg	taaaggtcgt	1500
cctagaaata	tcactatcga	ttctgatatg	attccagacc	caagagctag	actttgcgct	1560
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SEQ ID NO:72

MSKSNSMNST	SHETLFQQLV	LGLDRMPLMD	VHWLIYVAFG	AWLCSYVIHV	LSSSSTVKVP	60
VVGYRSVFEP	TWLLRLRFVW	EGGSIIGQGY	NKFKDSIFQV	RKLGTDIVII	PPNYIDEVRK	120
LSQDKTRSVE	PFINDFAGQY	TRGMVFLQSD	LQNRVIQQRL	TPKLVSLTKV	MKEELDYALT	180
KEMPD MNDE	WVEVDISSIM	VRLISRISAR	VFLGPEHCRN	QEWLTTTAEY	SESLFITGFI	240
LRVPHILRP	FIAPLLPSYR	TLLRNVSSGR	RVIGDIIRSQ	QGDGNEDILS	WMRDAATGEE	300

KQIDNIAQRM	LILSLASIHT	TAMTMTHAMY	DLCACPEYIE	PLRDEVKSVV	GASGWDKTAL	360
NRFHKLDSFL	KESQRFNPFV	LLTFNRIYHQ	SMTLSDGTNI	PSGTRIAVPS	HAMLQDSAHV	420
PGPTPTEFD	GFRYSKIRSD	SNYAQKYLFS	MTDSSNMAFG	YGKYACPGRF	YASNEMKLTL	480
AILLQFEFK	LPDGKGRPRN	ITIDSDMIPD	PRARLCVRKR	SLRDE		525

SEQ ID NO:73

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caagagggat	atgatggcta	cagaggatct	acattcaaaa	tcgcgatggt	agaccgttgg	240
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ttaaacttta	tgacggatt	aggagcattc	gtccaaacta	agtacacctt	aggtgaagct	360
attcataacg	atccatacca	tgtcgatata	ataagagaaa	aactaacaag	aggccttcca	420
gccgtgcttc	ctgatgtcat	tgaagagttg	acacttgccg	ttagacagta	cattccaaca	480
gaaggtgatg	aatgggtgct	cgtaaactgt	tcaaaggccg	caagagatat	tgttgctaga	540
gcttctaata	gagtctttgt	aggtttgctt	gcttgacaga	accaaggtta	cttagatttg	600
gcaatagact	ttacattgtc	tgttgtaacg	gatagagcca	tcatcaatat	gtttccagaa	660
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gttccttttg	ttgtctccatt	ggtggaggaa	agacgtagac	ttatggaaga	gtacggtgaa	780
gactggtctg	aaaaacctaa	tgatatgta	cagtgataaa	tggatgaagc	tgcatccaga	840
gatagttcag	tgaaggcaat	cgcagagaga	ttgttaatgg	tgaacttcgc	ggctattcat	900
acctcatcaa	acactatcac	tcatgctttg	taccaccttg	ccgaaatgcc	tgaaactttg	960
caaccactta	gagaagagat	cgaaccatta	gtcaaagagg	agggctggac	caaggctgct	1020
atgggaaaaa	tgtggtggtt	agattcattt	ctaagagaat	ctcaaagata	caatggcatt	1080
aacatcgtat	ctttaactag	aatggctgac	aaagatatta	cattgagtga	tggcacattt	1140
ttgccaaaag	gtactctagt	ggccgttcca	gcgtattcta	ctcatagaga	tgatgctgct	1200
tacgctgatg	ccttagtatt	cgatcctttc	agattctcac	gtatgagagc	gagagaaggt	1260
gaaggtacaa	agcaccagtt	cgtaataact	tcagtgcagt	acgttccatt	tggtcacgga	1320
aagcatgctt	gtccaggaag	attcttcgcc	gcaaacgaat	tgaaagcaat	gttggettac	1380
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tggggtccaa	cagttttgcc	tgcaccagca	ggccaagtat	tgttcagaaa	gagacaagtt	1500
agtctataac	cgcgg					1515

SEQ ID NO:74

MEDPTVLYAC	LAIAVATFVV	RWYRDPLRSI	PTVGGSDLPI	LSYIGALRWT	RRGREILQEG	60
YDGYRGSTFK	IAMLDRWIVI	ANGPKLADEV	RRRPDEELNF	MDGLGAFVQT	KYTLGEAIHN	120
DPYHVDIIRE	KLTRGLPAVL	PDVIEELTLA	VRQYIPTEGD	EWVSVNCSKA	ARDIVARASN	180
RVFVGLPACR	NQGYLDLAID	FTLSVVKDRA	IINMFPELLK	PIVGRVVGNA	TRNVRRAVPF	240
VAPLVEERRR	LMEEYGEDWS	EKPNMQLQWI	MDEAASRDSS	VKAI AERLLM	VNFAAIHTSS	300
NTITHALYHL	AEMPETLQPL	REEIEPLVKE	EGWTKAAMGK	MWWLDSFLRE	SQRYNGINIV	360
SLTRMADKDI	TLSDGTFLPK	GTLVAVPAYS	THRDDAVYAD	ALVFDPPFRFS	RMRAREGEGT	420
KHQFVNVSVE	YVPFGHGKHA	CPGRFFAANE	LKAMLAYIVL	NYDVKLPGDG	KRPLNMYWGP	480
TVLPAPAGQV	LFRKRQVSL					499

SEQ ID NO:75

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ttgccaaagt	ttccagtagt	gcctggtttt	ccagttattg	ggaatttggt	gcaactaaag	180
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gccttcggta	aagacgtaga	atccatatac	gtcaaggagt	taggcgtaac	attatcaaaa	660
gatgaaatct	ttaaggtgct	tgtacatgat	atgatggagg	gtgcaattga	tgtagattgg	720

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aatgggtctg	aatcagatga	tgattgttac	cttaacttct	taatgtctga	ggctaaaaca	900
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accttagtca	caactgaatg	ggccatatac	gagctagcca	aacatccatc	tgtgcaagat	1020
aggttgtgta	aggagatcca	gaacgtgtgt	ggtggagaga	aattcaagga	agagcagttg	1080
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atggcaatca	tcaatcctag	aagatcctaa				1530

SEQ ID NO:76

MAFFSMISIL	LGFVISSFIF	IFFFKLLSF	SRKNMSEVST	LPSVPVVPGF	PVIGNLLQLK	60
EKKPHKTFTR	WSEIYGPIYS	IKMGSSSLIV	LNSTETAKEA	MVTRFSSIST	RKLSNALTVL	120
TCDKSMVATS	DYDDFHLVK	RCLLNGLLGA	NAQKRKRHYR	DALIENVSSK	LHAHARDHPQ	180
EPVNFRAIFE	HELFGVALKQ	AFGKDVESY	VKELGVTLK	DEIFKVLVHD	MMEGAIDVDW	240
RDFFPYLKWI	PNKSFEARIQ	QKHRRRLAVM	NALIQDRLKQ	NGSESDDDCY	LNFLMSEAKT	300
LTKEQIAILV	WETIETADT	TLVTTEWAIY	ELAKHPSVQD	RLCKEIQNVC	GGEKFKEEQL	360
SQVPYLNQVF	HETLRKYSPA	PLVPIRYAHE	DTQIGGYHVP	AGSEIAINIY	GCMNDKKRWE	420
RPEDWWPERF	LDDGKYETSD	LHKTMAFGAG	KRVCAGALQA	SLMAGIAIGR	LVQEFEWKLR	480
DGEEENVDTY	GLTSQKLYPL	MAIINPRRS				509

SEQ ID NO:77

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aagatgctag	ttgaaaatag	agaattgttg	acactgttca	caacttcctt	cgcagttcct	180
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ccagttccac	aagttatcgt	tgtaaagaag	aaagagaagg	agtcagaggt	tgatgacggg	300
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ggtcatgttg	ttcatgatgc	acagcatcct	tcaagatcta	atgtggcttt	caaaaaggaa	960
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gttcaggaac	aaggagctct	ggattctctc	aaggctgaat	tgtacgtcaa	aaacttacag	2100
atgtctggaa	gatacttaag	agatgthttg	taa			2133

SEQ ID NO:78

MQSDSVKVSP	FDLVSAAMNG	KAMEKLNASE	SEDPTTLPAL	KMLVENRELL	TLFTTSFAVL	60
IGCLVFLMWR	RSSSKKLVQD	PVPQVIVVKK	KEKESEVDDG	KKKVSIFYGT	QTGTAEGFAK	120
ALVEEAKVRY	EKTSFKVIDL	DDYAADDDEY	EEKLKESLA	FFFLATYGDG	EPTDNAANFY	180
KWFTEGDDKG	EWLKKLQYGV	FGLGNRQYEH	FNKIAIVVDD	KLTEMGAKRL	VPVGLGDDQ	240
CIEDDFTAWK	ELVWPELDQL	LRDEDDTSVT	TPYTAADVLEY	RVVYHDKPAD	SYAEDQHTN	300
GHVVHDAQHP	SRSNVAFKKE	LHTSQSDRSC	THLEFDISHT	GLSYETGDHV	GVYSENLSEV	360
VDEALKLLGL	SPDYFSVHA	DKEDGTPIGG	ASLPPFPFPC	TLRDALTRYA	DVLSSPKKVA	420
LLALAAHASD	PSEADRLKFL	ASPAGKDEYA	QWIVANQRSL	LEVMSQSFPSA	KPPLGVFFAA	480
VAPRLQPRYY	SISSSPKMSP	NRIHVTCALV	YETTPAGRIH	RGLCSTWMKN	AVPLTESPDC	540
SQASIFVRTS	NFRLPVDPKV	PVIMIGPAGT	LAPFRGFLQE	RLALKESGTE	LGSSIFFFGC	600
RNRKVDFIYE	DELNFVETG	ALSELIVAFS	REGTAKEYVQ	HKMSQKASDI	WKLLSEGAYL	660
YVCGDAKGMA	KDVHRTLHTI	VQEQGLDSS	KAELYVKNLQ	MSGRYLRDVM		710

SEQ ID NO:79

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gaatthggth	catccatthtt	gthttthccggt	thcgaaaca	gaaagatthg	thtacctctac	1800
gaagatgaa	tgaacaactt	cgthgaaacc	gthgctthgt	ccgaatthgth	tattgctthtt	1860
thtagagaag	gthctaccaa	agaatacgtc	caacataaga	tgthgthgaaa	gctthctgth	1920
actthgaa	tgatthctga	agthgctth	thgtactgthtt	gthgthgatgc	thaaagthgth	1980
gctaaagthg	thtcatgaa	ctthcatacc	atcatgcaag	aacaagtht	thtggatthct	2040
thccaagctg	aatccatgth	caagaactthg	caaatgaa	gthagatactt	aagatgtht	2100

tggtaa

2106

SEQ ID NO:80

MKVSPFEFMS	AIKGRMDPS	NSSFESTGEV	ASVIFENREL	VAILTTSIAV	MIGCFVVLMMW	60
RRAGSRKVKV	VELPKPLIVH	EPEPEVEDGK	KKVSIFFGTQ	TGTAEGFAKA	LADEAKARYE	120
KATFRVVDLD	DYAADDQYE	EKLKNESFAV	FLLATYGDGE	PTDNAARFYK	WFAEGKERGE	180
WLQNLHYAVF	GLGNRQYEHF	NKIAKVADEL	LEAQGGNRLV	KVGLGDDDDQC	IEDDFSARE	240
SLWPELDMLL	RDEDDATTVT	TPYTAAVLEY	RVVFHDSADV	AAEDKSWINA	NGHAVHDAQH	300
PFRSNVVRK	ELHTSASDRS	CSHLEFNISG	SALNYETGDH	VGVCENLTE	TVDEALNLLG	360
LSPETYFSIY	TDNEDGTPLG	GSSLPPFPFS	CTLRTALTRY	ADLLNSPKKS	ALLALAAHAS	420
NPVEADRLRY	LASPAGKDEY	AQSVIGSQKS	LLEVMAEFPS	AKPPLGVFFA	AVAPRLQPRF	480
YSISSSPRMA	PSRIHVTAL	VYDKMPTGRI	HKGVCSTWMK	NSVPMESHE	CSWAPIFVRQ	540
SNFKLPAESK	VPIIMVPGT	GLAPFRGFLQ	ERLALKESGV	ELGPSILFFG	CRNRRMDYIY	600
EDELNNFVET	GALSELVIAF	SREGPTKEYV	QHKMAEKASD	IWNLISEGAY	LYVCGDAKGM	660
AKDVHRTLHT	IMQEQGLDS	SKAESMVKNL	QMNGRYLRDV	W		701

SEQ ID NO:81

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gcatacttta	ctaagggtaa	attgtgggggt	gttaccaagg	atccatacgc	taacggattc	120
gctgcagggtg	gtgcttccaa	gcctggcaga	actagaaaca	tcgtcgaagc	tatggaggaa	180
tcaggtaaaa	actgtgttgt	tttctacggc	agtcaaacag	gtacagcgga	ggattacgca	240
tcaagacttg	caaaggaagg	aaagtccaga	ttcggtttga	acactatgat	cgccgatcta	300
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gcagaatcat	acgaactttt	ctcagctaag	gatagaaatt	gtctgcatat	ggaaattgat	900
atctctggta	gtaactctaa	gtatgaaaca	ggcgaccata	tcgcatctg	gcctaccaac	960
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tacgatgcta	tattgagata	ccatctggaa	atatgcgctc	cagtttctag	acagtttgtc	1140
tcaactttag	cagcattcgc	ccctaattgat	gatatcaaag	ctgagatgaa	ccgtttggga	1200
tcagacaaag	attactcca	cgaaaagaca	ggaccacatt	actacaatat	cgctagattt	1260
ttggcctcag	tctctaaaag	tgaaaaatgg	acaaagatac	cattttctgc	tttcatagaa	1320
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ggcgacaaat	tcgaaatgat	tacagctttt	tcaagagaag	gatctaaaaa	ggtttatggt	1860
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agatcagcaa	atcaatacca	agtgtgttct	gatttcgtaa	ctttacactg	taaagagaca	2100
acatacgcga	attcagaatt	gcaagaggat	gtctggagtt	aa		2142

SEQ ID NO:82

MAELDTLDIV	VLGVIFLGTV	AYFTKGLWG	VTKDPYANGF	AAGGASKPGR	TRNIVEAMEE	60
SGKNCVVFY	SQTGTAEDYA	SRLAKEGKSR	FGLNTMIADL	EDYDFDNLDT	VPSDNIVMVF	120

LATYGEGET	DNAVDFYEFI	TGEDASFNEG	NDPPLGNLNY	VAFGLGNNTY	EHYNSMVRNV	180
NKALEKLGAAH	RIGEAGEGDD	GAGTMEEDFL	AWKDPMWEAL	AKKMGLEERE	AVYEPIFAIN	240
ERDDLTPKAN	EVYLGEPNKL	HLEGTAKGPF	NSHNPYIAPI	AESYELFSAK	DRNCLHMEID	300
ISGSNLKYET	GDHIAIWPTN	PGEEVNKFLD	ILDLSGKQHS	VVTVKALEPT	AKVFPNPTT	360
YDAILRYHLE	ICAPVSRQFV	STLAAFAPND	DIKAEMNRLG	SDKDYFHEKT	GPHYNYIARF	420
LASVSKGEKW	TKIPFSAFIE	GLTKLQPRYY	SISSSSLVQP	KKISITAVVE	SQQIPGRDDP	480
FRGVATNYLF	ALKQKQNGDP	NPAPFGQSYE	LTGPRNKYDG	IHVPVHVRHS	NFKLPSDPGK	540
PIIMIGPGTG	VAPFRGFVQE	RAKQARDGVE	VGKTLFFGFC	RKSTEDFMYQ	KEWQEYKEAL	600
GDKFEMITAF	SREGSKKVYV	QHRLKERSKE	VSDLQSQKAY	FYVCGDAAHM	AREVNTVLAQ	660
IIAEGRGVSE	AKGEEIVKNM	RSANQYQVCS	DFVTLHCKET	TYANSELQED	VWS	713

SEQ ID NO:83

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gcgatgatgt	tcgaaattcg	tgatctgttg	ctgattttga	ctacgtcagt	tgctgttttg	180
gtcggatggt	tcgttgtttt	ggtgtggaag	agatcgctcg	ggaagaagtc	cggcaaggaa	240
ttggagccgc	cgaagatcgt	tgtgccgaag	aggcggctgg	agcaggaggt	tgatgatggt	300
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cgcgtcgtat	ttcatgacaa	acccgatgcg	ttttctgatg	atcactactca	aaccaatggt	900
catgctgttc	atgatgctca	acatccatgc	agatccaatg	tggctgttaa	aaaagagctt	960
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gaagaagctg	ggaaattggt	aggattatca	acagataact	atttctcgtt	acatattgat	1140
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aaccgcaaag	tggattacat	atatgagaat	gaactcaaca	actttgttga	aatgggtgcg	1860
ctttctgagc	ttgatgttgc	tttctcccgc	gatggcccga	cgaaagaata	cgtgcaacat	1920
aaaatgacct	aaaaggcttc	tgaaatatgg	aatatgcttt	ctgagggagc	atatttatat	1980
gtatgtggtg	atgctaaaag	catggctaaa	gatgtacacc	gtacacttca	caccattgtg	2040
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tcaggaagat	acctccgtga	tgtttggtaa				2130

SEQ ID NO:84

MQSESVEAST	IDLMTAVLKD	TVIDTANASD	NGDSKMPPAL	AMMFEIRDLL	LILTTSVAVL	60
VGCFVVLVWK	RSSGKKSKE	LEPPKIVVVK	RRLEQEVDDG	KKKVTIFFGT	QTGTAEGFAK	120
ALFEEAKARY	EKAFAKVIDL	DDYAADLDEY	AEKLLKETYA	FFFLATYGDG	EPTDNAAKFY	180
KWFTEGDEK	VWLQKLQYGV	FGLGNRQYEH	FNKIGIVDD	GLTEQGAKRI	VPVGLDDDDQ	240
SIEDDFSAAK	ELVWPELDDL	LRDEDDKAAA	TPYTAAIPEY	RVVFHDKPDA	FSDDHTQNG	300
HAVHDAQHPC	RSNVAVKKEL	HTPESDRSCT	HLEFDISHTG	LSYETGDHVG	VCENLIEVV	360
EEAGKLLGLS	TDTYFSLHID	NEDGSPLGGP	SLQPPFPCT	LRKALTNYAD	LLSSPKKSTL	420

LALAAHASDP	TEADRLRFLA	SREGKDEYAE	WVVANQRSLL	EVMEAFPSAR	PPLGVFFAAV	480
APRLQPRYS	ISSSPKMEPN	RIHVTCALVY	EKTPAGRIHK	GICSTWMKNA	VPLTESQDCS	540
WAPIFVRTSN	FRLPIDPKVP	VIMIGPGTGL	APFRGFLQER	LALKESGTEL	GSSILFFGCR	600
NRKVDYIYEN	ELNNFVENGA	LSELDVAFSR	DGPTKEYVQH	KMTQKASEIW	NMLSEGAYLY	660
VCGDAKGMMAK	DVHRTLHTIV	QEQGLDSSK	AELYVKNLQM	SGRYLRD VW		709

SEQ ID NO:85

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agatactcc	gtgacgtttg	gtaa				2124

SEQ ID NO:86

MQSNSVKISP	LDLVTALFSG	KVLDTSNASE	SGESAMLPTI	AMIMENRELL	MILTTSVAVL	60
IGCVVVLVWR	RSSTKKSAL	PPVIVVPKRV	QEEVDDGKK	KVTVFFGTQT	GTAEGFAKAL	120
VEEAKARYEK	AVFKVIDLDD	YAADDDEYEE	KLKKEFLAFF	FLATYGDGEP	TDNAARFYKW	180
FTEGDAKGEW	LNKLQYGVFG	LGNRQYEHFN	KIAKVDDGL	VEQGAKRLVP	VGLGDDQCI	240
EDDFTAWKEL	VWPELDQLLR	DEDDTTVATP	YTAAVAERYV	VFHEKPDALS	EDYSYTNNGHA	300
VHDAQHPCRS	NVAVKKELHS	PESDRSCTHL	EFDISNTGLS	YETGDHVG VY	CENLSEVVND	360
AERLVGLPPD	TYSSIHTDSE	DGSPLGGASL	PPFPFPCTLR	KALTCYADVL	SSPKKSALLA	420
LAAHATDPSE	ADRLKFLASP	AGKDEYSQWI	VASQRSLLLEV	MEAFPSAKPS	LGVFFASVAP	480
RLQPRYSIS	SSPKMAPDRI	HVTCALVYEK	TPAGRIHKGV	CSTWMKNAV P	MTESQDCS W A	540
PIYVRTSNFR	LSPDPKVPVI	MIGPGTGLAP	FRGFLQERLA	LKEAGTDLGL	SILFFGCRNR	600
KVDFIYENEL	NNFVETGALS	ELIVAFSREG	PTKEYVQHKM	SEKASDIWNL	LSEGAYLYVC	660
GDAKGMMAKDV	HRTLHTIVQE	QGLDSSKAE	LYVKNLQMSG	RYLRD VW		709

SEQ ID NO:87

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gacggtagat	acttgagaga	tgtttgggtga				2070

SEQ ID NO:88

MSSNSDLVRR	LESVLGVSFG	GSVTDSVVVI	ATTSIALVIG	VLVLLWRRSS	DRSREVKQLA	60
VPKPVITVEE	EDEFEVASGK	TRVSI FYGTQ	TGTAEGFAKA	LAEEIKARYE	KA AVKVIDLD	120
DYTAEDDKYG	EKLKKE TMAF	FMLATYGDGE	PTDNAARFYK	WFTEG TDRGV	WLEHLRYGVF	180
GLGNRQYEHF	NKI AKVVDDL	LVEQGA KRLV	TVGLGD DDDQC	IEDDFSAWKE	ALWPELDQLL	240
QDDTNTVSTP	YTAVIPEYRV	VIHDP SVTSY	EDPYSNMANG	NASYDIH HPC	RANVAVQKEL	300
HKPESDRSCI	HLEFDIFATG	LYETGDHVG	VYADNCDDTV	EEAAKLLGQP	LDLLFSIHTD	360
NNDG TSLGSS	LPPFPFGPCT	LRTALARYAD	LLNPPKKAAL	I ALAAHADEP	SEAERLKFLS	420
SPQ GKDEYSK	WVVGSRSLV	EVMAEFPSAK	PPLGVFFAAV	VPRLQPRYYS	I SSSPRFAPH	480
RVHVTALVY	GPTPTGRIHR	GVCSFWMKNV	VPLEKSQNCS	WAPIFIRQSN	FKLPADHSVP	540
IVMVGPGTGL	APFRGFLQER	LALKEEGAQV	GPALLFFGCR	NRQMDFIYEV	ELNNFVEQGA	600
LSELIVAFSR	EGPSKEYVQH	KMVEKAA YMW	NLISQGGYFY	VCGDAKGMAR	DVHRTLHTIV	660
QQEEKVDSTK	AESIVKKLQM	DGRYLRD VW				689

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atgacttctg	cactttatgc	ctccgatctt	ttcaaacaaat	tgaaaagtat	catgggaacg	60
gattctttgt	ccgatgatgt	tgtattagtt	atgtctacaa	cttctctggc	actgggtgct	120
ggtttcgttg	tcttattgtg	gaaaaagacc	accgacagatc	gttccggcga	gctaagcca	180
cta atgatcc	cta agtctct	gatggcgaaa	gatgaggatg	atgacttaga	tctaggttct	240

ggaaaaacga	gagtctctat	cttcttcggc	acacaaaaccg	gaacagccga	aggattcgc	300
aaagcacttt	cagaagagat	caaagcaaga	tacgaaaagg	cggtctgtaa	agtaatcgat	360
ttggatgatt	acgctgccga	tgatgaccaa	tatgaggaaa	agttgaaaa	ggaaacattg	420
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gaagagttat	gcaaaaagg	tgcaagaga	ttgattgaag	tcggtttagg	agatgatgat	660
caatctatcg	aggatgactt	taatgcatgg	aaggaatctt	tgtggtctga	attagataag	720
ttacttaagg	acgaagatga	taaatccggt	gccactccat	acacagccgt	cattccagaa	780
tatagagtag	ttactcatga	tccaagattc	acaacacaga	aatcaatgga	aagtaatgtg	840
gctaattggt	atactaccat	cgatattcat	catccatgta	gagtagacgt	tgcaagttca	900
aaggaattgc	acactcatga	atcagacaga	tcttgcatat	atcttgaatt	tgatataatc	960
cgtactggta	tcacttacga	aacaggtgat	cacgtgggtg	tctacgctga	aaacatggt	1020
gaaattgtag	aggaagctgg	aaagttggtg	ggccatagtt	tagatcttgt	tttctcaatt	1080
catgccgata	aagaggatgg	ctcaccacta	gaaagtgcag	tgctccacc	atttccagga	1140
ccatgcacc	taggtaccgg	tttagctcgt	tacgcggatc	tgtaaatacc	tccacgtaa	1200
tcagctctag	tgcccttggc	tgcgtagcgc	acagaacctt	ctgaggcaga	aaaactgaa	1260
catctaact	caccagatgg	taaggatgaa	tactcacaat	ggatagtagc	tagtcaactg	1320
tcttactag	aagttagggc	tgctttccca	tcctgtaaac	ctcctttggg	tgttttcttc	1380
gccgcaatag	cgcttagact	gcaaccaaga	tactattcaa	tttcatcctc	acctagactg	1440
gcaccatcaa	gagttcatgt	cacatccgct	ttagtgtacg	gtccaactcc	tactggtaga	1500
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tctactccta	ttgtcatggt	cggtcctggt	acaggtcctg	ctccattcag	aggtttctta	1680
caagagagaa	tgcccttaaa	ggaggatggt	gaagagttgg	gatcttcttt	gttgtttttc	1740
ggctgtagaa	acagacaaat	ggatttcate	tacgaagatg	aactgaataa	ctttgtagat	1800
caaggagtta	tttcagagtt	gataatggct	ttttctagag	aaggtgctca	gaaggagtac	1860
gtccaacaca	aatgatgga	aaaggccgca	caagtttggg	acttaataca	agaggaaggc	1920
tatctatatg	tctgtggtga	tgcaaagggt	atggcaagag	atgttcacag	aacacttcat	1980
actatagtcc	aggaacagga	aggcgtagt	tcttctgaag	cggaagcaat	tgtgaaaaag	2040
ttacaaacag	aggaagata	cttgagagat	gtgtggttaa			2079

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MTSALYASDL	FKQLKSI	MTGTDLSL	DDVVLV	IATTS	LALVA	GFVLL	LWKKT	TADR	SGELKP	60	
LMIPKSLMAK	DEDDDL	DLGSGK	TRVSI	FFGTQT	GTAEGFA	KALSEE	EIKAR	YEKA	AVKVID	120	
LDDYAADDQ	YEEKL	KKETL	AFFCV	ATYGD	GEPTD	NAARF	YKWF	TEENER	DIKLQ	QLAYG	180
VFALGNRQYE	HFNKI	GIVLD	EELCK	KGAKR	LIEVGL	GDDDD	QSIED	DFNAW	KESLW	SELDK	240
LLKDEDDKSV	ATPYT	AVIPE	YRVVT	HDPFR	TTQKS	MESNV	ANGNT	TIDIH	HPCR	VDVAVQ	300
KELHTHESDR	SCIHL	EFDIS	RTGIT	YETGD	HVGVA	AENHV	EIVVE	EAGKLL	GHSLD	LVFSI	360
HADKEDGSPL	ESAVP	PPFP	PCTLG	TGLAR	YADLL	NPPRK	SALVA	LAAYA	TEPSE	AEKLL	420
HLTSPDGKDE	YSQW	IVASQR	SLLEV	MAAFP	SAKP	PLGVFF	AAIAP	RLOPR	YYSI	SSSRL	480
APSRVHVTSA	LVYG	PPTGR	IHKGV	CSTWM	KNAV	PAEKSH	ECSG	APIFIR	ASNFK	LPSNP	540
STPIVMVGP	TGLAP	FRGFL	QERM	ALKEDG	EELGS	SLFF	GCRNR	QMDFI	YEDEL	NNFVD	600
QGVISELIMA	FSREG	AQKEY	VQHKM	MEKAA	QVWDL	IKEEG	YLYVC	GDAKG	MARDV	HRTLH	660
TIVQEQEGVS	SSEAE	AIVKK	LQTE	GRYLRD	VW						692

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ggtgaaccag	ttatcgtctc	cgaccagca	aatgcctctg	cttatgaatc	agttgctgca	120
gaattgtctt	caatgttgat	cgaaaacaga	caattcgcca	tgatcgtaac	tacatcaatc	180
gctgttttga	tcggttgat	tgatcattg	gtatggagaa	gatccggtag	tggtaatctt	240
aaaagagtcg	aacctttgaa	accattagta	attaagccaa	gagaagaaga	aatagatgac	300
ggtagaaaga	aagttacaat	atttttcggg	acccaaactg	gtacagctga	aggttttgca	360
aaagccttag	gtgaagaagc	taaggcaaga	tacgaaaaga	ctagattcaa	gatagtcgat	420
ttgattgact	atgccgctga	tgacgatgaa	tacgaagaaa	agttgaagaa	agaagatggt	480
gcatttttct	tttgccaac	ctatggtgac	gggtgaacaa	ctgacaatgc	agccagattc	540
tacaaatggt	ttacagaggg	taatgatcgt	gggtgaatggt	tgaaaaactt	aaagtacggg	600

gttttcgggt	tgggtaacag	acaatacgaa	catttcaaca	aagttgcaa	ggttgtcgac	660
gatattttgg	tcgaacaagg	tgctcaaaga	ttagtccaag	taggtttggg	tgacgatgac	720
caatgtatag	aagatgactt	tactgcctgg	agagaagcct	tgtggcctga	attagacaca	780
atcttgagag	aagaaggtga	caccgccggt	gctaccccat	atactgctgc	agtattagaa	840
tacagagttt	ccatccatga	tagtgaagac	gcaaagttta	atgatatcac	tttggccaat	900
ggtaacgggt	atacagtttt	cgatgcacaa	cacccttaca	aagctaacgt	tgacgtcaag	960
agagaattac	atacaccaga	atccgacaga	agttgtatac	acttgggaatt	tgatatacgt	1020
ggttccgggt	taaccatgaa	gttgggtgac	catgtagggt	ttttatgcga	caatttgtct	1080
gaaactggtg	atgaagcatt	gagattggtg	gatatgtccc	ctgacactta	ttttagtttg	1140
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gccttggttg	ctttagccgc	tcatgctagt	gacccactg	aagcagaaag	attgaaacac	1320
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ggtgtagcac	ctagattgca	accaagattc	tactcaatca	gttcttcacc	taagatcgct	1500
gaaactagaa	ttcatgttac	atgtgcatta	gtctacgaaa	agatgccaac	cggtagaatt	1560
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aaggttccaa	taatcatgat	aggtcctggt	acaggtttag	ccccattcag	aggtttcttg	1740
caagaaagat	tggctttagt	tgaatctggg	gtcgaattag	gtccttcagt	tttgttcttt	1800
ggtttagtaa	acagaagaat	ggatttcatc	tatgaagaag	aattgcaaag	attcgtcgaa	1860
tctggtgcat	tggccgaatt	atctgtagct	ttttcaagag	aaggtccaac	taaggaatac	1920
gttcaacata	agatgatgga	taaggcatcc	gacatatgga	acatgatcag	tcaaggtgct	1980
tattttagac	tttgccgtga	cgcaaagggg	atggccagag	atgtccatag	atctttgcac	2040
acaattgctc	aagaacaagg	ttccatggat	agtaccaaaag	ctgaagggtt	cgtaaagaac	2100
ttacaaactt	ccgtagata	cttgagagat	gtctggtga			2139

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MSSSSSSSTS	MIDLMAAIK	GEPVIVSDPA	NASAYESVAA	ELSSMLIENR	QFAMIVTTSI	60
AVLIGCIVML	VWRRSGSGNS	KRVEPLKPLV	IKPREEEIDD	GRKKVTIFFG	TQTGTAEGFA	120
KALGEEAKAR	YEKTRFKIVD	LDDYAADDDE	YEEKLKKEDV	AFFFLATYGD	GEPTDNAARF	180
YKWFTEGNDR	GEWLKLNLYG	VFGLGNRQYE	HFNKVAKVVD	DILVEQGAQR	LVQVGLGDDD	240
QCIEDDFTAW	REALWPELDT	ILREEGDTAV	ATPYTAAVLE	YRVSIHDSER	AKFNDITLAN	300
GNGYTVFDAQ	HPYKANVAVK	RELHTPESDR	SCIHLEFDIA	GSGLTMKLGD	HVGVLCNLS	360
ETVDEALRLL	DMSPDYFSL	HAEKEDGTPI	SSSLPPFPFP	CNLRTALTRY	ACLLSSPKKS	420
ALVALAAHAS	DPTEAERLKH	LASPAGKDEY	SKWVVESQRS	LLEVMAEFPS	AKPPLGVFFA	480
GVAPRLQPRF	YSISSSPKIA	ETRIHVTCAL	VYEKMPGTGR	HKGVCSTWMK	NAVPEKSEK	540
LFLGRPIFVR	QSNFKLPSDS	KVPIIMIGPG	TGLAPFRGFL	QERLALVESG	VELGPSVLFF	600
GCRNRRMDFI	YEEELQRFVE	SGALAELSVA	FSREGPTKEY	VQHKMMDKAS	DIWNMISQGA	660
YLYVCGDAKG	MARDVHRSLSH	TIAQEQGSMD	STKAEGFVKN	LQTSGRYLRD	VW	712

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actcaactta	gaaggaagag	cgctaatacta	ccaccaaccg	tgtttccatc	aataccaatc	120
attggacact	tatacttact	caaaaagcct	ctttatagaa	cttttagcaaa	aattgccgct	180
aagtacggac	caatactgca	attacaactc	ggctacagac	gtgttctggt	gatttctctca	240
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tggcgtaatc	taaggagagt	agcttctatc	gaaatcctat	cagttcatag	gttgaacgaa	420
tttcatgata	tcagagtgga	tgagaacaga	ttgttaatta	gaaaacttag	aagttcatct	480
tctcctgtta	ctcttataac	agtcttttat	gctctaacat	tgaacgtcat	tatgagaatg	540
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aaagtaggca	aaggtagaaa	aacgatgac	gaactcttat	tatctttgca	agagtacgaa	840
cctgagtact	atacagatgc	tatgataaga	tcttttgtcc	taggtctgct	ggctgcagg	900

agtgatactt	cagcggggcac	tatggaatgg	gccatgagct	tactgggtcaa	tcacccacat	960
gtattgaaga	aagctcaagc	tgaaatcgat	agagttatcg	gtaataacag	attgattgac	1020
gagtcagaca	ttggaaatat	cccttacatc	gggtgtatta	tcaatgaaac	tctaagactc	1080
tatccagcag	ggccattggt	gttcccacat	gaaagttctg	ccgactgcgt	tatttccggg	1140
tacaatatac	ctagaggtac	aatgttaatc	gtaaaccaat	gggcgattca	tcacgatcct	1200
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gttccattag	ttgccaaaatg	taagccacgt	tccgaaatga	ctaattctcct	atccgaactt	1500
taa						1503

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MEASYLYISI	LLLLASYLFT	TQLRRKSANL	PPTVFPSIPI	IGHLYLLKKP	LYRTLAKIAA	60
KYGPILQLQL	GYRRVLVISS	PSAAEECFN	NDVIFANRPK	TLFGKIVGGT	SLGSLSYGDQ	120
WRNLRRVASI	EILSVHRLNE	FHDIRVDENR	LLIRKLRSSS	SPVTLITVFI	ALTINVIMRM	180
ISGKRYFDSG	DRELEEEGKR	FREILDETLI	LAGASNVDY	LPILNWLGVK	SLEKLIALQ	240
KKRDDFFQGL	IEQVRKSRGA	KVGKGRKMTI	ELLLSLQESE	PEYYTDAMIR	SFVLGLLAAG	300
SDTSAGTMEW	AMSLLVNHPH	VLKKAQAEID	RVIGNNRLID	ESDIGNIPYI	GCIINETLRL	360
YPAGPLLFPH	ESSADCVISG	YNI PRGTMLI	VNQWAIHHD	KVWDDPETFK	PERFQGLEGT	420
RDGFKLMPFG	SGRRGCPGEG	LAIRLLGMTL	GSVIQCFDWE	RVGDEMVDMT	EGLGVTLPKA	480
VPLVAKCKPR	SEMTNLLSEL					500

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ccaacattcc	attcggagag	gctaaagcgt	atgttacctt	catttcacca	aagttgtaat	540
gagatgggtca	aggaatggga	gagcttgggtg	tcaaaagagg	gttcatcatg	tgagttggat	600
gtctggcctt	ttcttgaaaa	tatgtcggca	gatgtgatct	cgagaacagc	atttggact	660
agctacaaaa	aaggacagaa	aatctttgaa	ctcttgagag	agcaagtaat	atatgtaacg	720
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tcaaacttga	aggacattcg	ggaacatggg	aaaaacaaca	aaaatgttgg	gatgagtatt	960
gaagatgtaa	ttcaggagtg	taagctgttt	tactttgtctg	ggcaagaaac	cacttcagtg	1020
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caagaggttt	tgcaagtctt	tggaagcagc	aagccagatt	ttgatggtct	agctcacctt	1140
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gcaaagttgg	ccttagcatt	gatcttgcga	cacttcacct	ttgagctttc	tccatctcat	1500
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catcgacggt	ag					1572

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atggaagtca	ctgtcgcctc	ttctgtcgtt	ttatccttag	tcttcatttc	cattgtcgtc	60
agatgggctt	ggccggttgt	caactgggtt	tggttcaaac	caaagaagtt	ggaaagattc	120
ttgagagagc	aaggtttgaa	gggtaattct	tatagattct	tgtacggtga	catgaaggaa	180
aattctattt	tgttgaagca	agccagatoc	aaaccaatga	acttgtctac	ctctcatgat	240

attgctccac	aagtactcc	attcgctgat	caaactgtta	aagcctacgg	taagaactct	300
ttcaattggg	ttggtccaat	tcctagagtt	aacatcatga	accagaaga	tttgaaggat	360
gtcttgacca	agaacgttga	cttcgttaag	ccaatttcca	accattgat	taaattgttg	420
gctactggta	ttgccattta	cgaaggtgaa	aagtggacta	agcatagaag	aatcatcaac	480
cctaccttcc	actctgaaag	attgaagaga	atgtttaccat	ctttccatca	atcctgtaat	540
gaaatgggta	aggaatggga	atccttggtt	tctaaagaag	gttcttcttg	cgaattggat	600
gtttggccat	tcttgaaaa	tatgtctgct	gatgtcattt	ccagaaccgc	tttcggtagc	660
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cgtatgaacg	agatcaacga	agaaattaaa	ggtttgatca	gaggtagtat	tatcgacaga	840
gaacaaatta	ttaaagctgg	tgaagaaacc	aacgatgatt	tgttgggtgc	tttgatggag	900
tccaacttga	aggatattag	agaacatggt	aagaacaaca	agaatggttg	tatgtctatt	960
gaagatgta	ttcaagaatg	taagttatct	tacttcgctg	gtcaagagac	cacttctggt	1020
ttgtagcct	ggactatggt	cttgtagggt	caaaaccaa	attggcaaga	tagagctaga	1080
caagaagttt	tgcaagtctt	cggttcttcc	aagccagact	ttgatgggtt	ggcccacttg	1140
aagggtgta	ctatgatttt	gttagaagtt	ttgagattgt	accaccagt	cattgagtta	1200
atcagaacca	ttcataaaaa	gactcaattg	ggtaaattat	ctttgccaga	aggtggtgaa	1260
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caatttaatc	cagaaagatt	ttccgaaggt	gtttccaagg	ctaccaaaaa	ccgtttgctc	1380
ttcttcccat	ttggtgctgg	tccacgtatt	tgtatcggtc	aaaacttttc	catgatggaa	1440
gccaagttgg	ctttggcttt	aatcttgcaa	cacttcactt	tcaaatgttc	tccatcccat	1500
gcccacgctc	cttctcatag	aatcacttta	caaccacaat	acggtgtcag	aatcatctta	1560
cacagaagat	aa					1572

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MEVTVAASSVA	LSLVFISIVV	RWAWSVVNWV	WFKPKKLERF	LREQGLKGN	YRFLYGDMKE	60
NSILLKQARS	KPMNLSTSHD	IAPQVTPFVD	QTVKAYGKNS	FNWVGPIPRV	NIMNPEDLKD	120
VLTKNVDFVK	PISNPLIKLL	ATGIAIYEGE	KWTKHRIIN	PTFHSERLKR	MLPSFHQSCN	180
EMVKEWESLV	SKEGSSCELD	VWPFLENMSA	DVISRTAFGT	SYKKGQKIFE	LLREQVIYVT	240
KGFQSFYIPG	WRFLPTKMNK	RMNEINEEIK	GLIRGIIIDR	EQIIKAGEET	NDDLLGALME	300
SNLKDIREHG	KNNKNVGMIS	EDVIOECKLF	YFAGQETTSV	LLAWTMVLLG	QNQNWDQDRAR	360
QEVLQVFGSS	KPDFDGLAHL	KVVTMILLEV	LRLYPPVIEL	IRTIHKKTQL	GKLSLPEGVE	420
VRLPTLLIHH	DKELWGDDAN	QFNPERFSEG	VSKATKNRSL	FFPFGAGPRI	CIGQNFMSME	480
AKLALALILQ	HFTFELSPSH	AHAPSHRITL	QPQYGVRIIL	HRR		523

SEQ ID NO:98

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ttgagggagc	aaggccttac	aggcaattct	tacaggcttt	tgtttgagag	caccaaggat	180
ctctcgaaga	tgctggaaca	aacacaatcc	aaaccatca	aactctccac	ctcccatgat	240
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gtgacctatga	tttacttga	agttcttcga	ttatacccat	cagtcggtgc	gcttctcga	1200
accactcaca	agaaaacaca	gcttgaaaaa	ttatcattac	cagctggagt	ggaagtctcc	1260

ttgccatac	tgcttgttca	ccatgacaaa	gagttgtggg	gtgaggatgc	aaatgagttc	1320
aagccagaga	ggttttcaga	gggagtttca	aaggcaacaa	agaacaaatt	tacatactta	1380
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cgttga						1566

SEQ ID NO:99

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gctttcaaca	gacatgatga	tttccataag	accgtcaaga	acccaattat	gaagtctcca	420
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gctttgagat	ctgtttacat	tccaggttgg	agattcttgc	caactaagca	aaacaaaaag	780
accaaagaaa	tccacaacga	aatcaagggg	ttgttgaagg	gtatcatcaa	caagagagaa	840
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gctccatctg	ctgttattac	attgcaacca	caatttgggtg	cccatatcat	cttgcataag	1560
agataac						1567

SEQ ID NO:100

MEASRASCVA	LCVVVWSIVI	TLAWRVLNWV	WLRPKKLERC	LREQGLTGNS	YRLLFGDTKD	60
LSKMLEQTQS	KPIKLTSHD	IAPRVTPFFH	RTVNSNGKNS	FVWMGPIPRV	HIMNPEDLKD	120
AFNRHDDFHK	TVKNPIMKSP	PPGIVGIEGE	QWAKHRKIIN	PAFHLEKLKG	MVPIFYQSCS	180
EMINKWESLV	SKESCELDV	WPYLENFTSD	VISRAAFGSS	YEGRKIFQL	LREEAKVYSV	240
ALRSVIYPGW	RFLPTKQNKK	TKEIHNEIKG	LLKGIINKRE	EAMKAGEATK	DDLLGILMES	300
NFREIQEHGN	NKNAGMSIED	VIGECKLFYF	AGQETTSVLL	VWTMILLSQN	QDWQARAREE	360
VLKVFGSNIP	TYEELSHLKV	VTMILLEVLR	LYPSVVALPR	TTHKKTQLGK	LSPAGVEVS	420
LPILLVHHDK	ELWGEDANEF	KPERFSEGVS	KATKNKFTYL	PFGGGPRICI	GQNFAMVEAK	480
LALALILQHF	AFELSPSYAH	APSAVITLQP	QFGAHIILHK	R		521

SEQ ID NO:101

ASWVAVLSVV	WVSMVIAWAW	RVLNWWVLRP	KKLEKCLREQ	GLAGNSYRLL	FGDTKDLSKM	60
LEQTQSKPIK	LSTSHDIAPH	VTPFFHQTVN	SYGKNSFVWM	GPIPRVHIMN	PEDLKDTFNR	120
HDDFHKVVKN	PIMKSLPQGI	VGIEGEQWAK	HRKIINPAFH	LEKLKGMVPI	FYRSCSEMIN	180
KWESLVSKES	SCELDVWPYL	ENFTSDVISR	AAFSSYEEG	RKIFQLLREE	AKIYTVAMRS	240
VYIPGWRFPL	TKQNKKAKEI	HNEIKGLLKG	IINKREEAMK	AGEATKDDLL	GILMESNFRE	300
IQEHGNNKNA	GMSIEDVIGE	CKLFYFAGQE	TTSVLLVWTM	VLLSQNDWQ	ARAREEVLOV	360
FGSNIPTYEE	LSQLKVVTMI	LLEVLRLYPS	VVALPRTHK	KTQLGKLSLP	AGVEVSLPIL	420
LVHHDKELWG	EDANEFKPER	FSEGVSKATK	NQFTYFPFGG	GPRICIGQNF	AMMEAKLALS	480
LILRHFALEL	SPLYAHAPSV	TITLQPQYGA	HIILHKR			517

SEQ ID NO:102

MEASRPSCVA	LSVVLVSVI	AWAWRVLNWV	WLRPNKLERC	LREQGLTGNS	YRLLFGDTKE	60
ISMVVEQAQS	KPIKLSSTHD	IAPRVIPFSH	QIVYTYGRNS	FVWMGPTPRV	TIMNPEDLKD	120
AFNKSDDEFQR	AINSPIVKS I	SQGLSSLEGE	KWAKHRKI IN	PAFHLEKLGK	MLPTFYQSCS	180
EMINKWESLV	FKEGSREMDV	WPYLENLTS D	VISRAAFGSS	YEEGRKIFQL	LREEAKFYTI	240
AARSVYIPGW	RFLPTKQNK R	MKEIHKEVRG	LLKGIINKRE	DAIKAGEAAK	GNULLGILMES	300
NFREIQEHGN	NKNAGMSIED	VIGECKLFYF	AGQETTSVLL	VWTLVLLSQ N	QDWQARAREE	360
VLQVFGTNIP	TYDQLSHLKV	VTMILLEVL R	LYPAVVELPR	TTYKKTQLGK	FLLPAGVEVS	420
LHIMLAHHDK	ELWGEDAKEF	KPERFSEGVS	KATKNQFTYF	PFGAGPRICI	GQNFAMLEAK	480
LALSLLQHF	TFELSPSYAH	APSVTITLHP	QFGAHFILHK	R		521

SEQ ID NO:103

CVALSVVLS	IIVAWAVRVL	NWVWLRPNKL	ERCLREQGLT	GNSYRLLFGD	TKEISMMVEQ	60
AQSKPIKLST	THDIAPRVIP	FSHQIVYTYG	RNSFVWMGPT	PRVTIMNPED	LKD AFNKSD E	120
FQRAISNPIV	KSISQGLSSL	EGEKWAKHRK	I INPAFHLEK	LKGMLPTFYQ	SCSEMINKWE	180
SLVFKEGSRE	MDVWPYLENL	TSDVISRAAF	GSSYEEGRKI	FQLLREEAKF	YTIAARSVYI	240
PGWRFLPTKQ	NKRMEIHKE	VRGLLKG I IN	KREDAIKAGE	AAKGNLLGIL	MESNFREIQE	300
HGNKNAGMS	IEDVIGECKL	FYFAGQETTS	VLLVWTLVLL	SQNQDWQARA	REEVLQVFGT	360
NIPTYDQLSH	LKVVTMILLE	VLRLYPVAVE	LPRTTYKKTQ	LGKFLLPAGV	EVSLHIMLAH	420
HDKELWGEDA	KEFKPERFSE	GVSKATKNQF	TYFPFGAGPR	ICIGQNFAML	EAKLALSLLI	480
QHFTFELSPS	YAHAPSVTIT	LHPQFGAHFI	LHKR			514

SEQ ID NO:104

MGPIPRVHIM	NPEDLKDTFN	RHDDFHKVVK	NPIMKSLPQG	IVGIEGDQWA	KHRKIINPAF	60
HLEKLGKGMVP	IFYQSCSEMI	NIWKSLSVKE	SCELDVWPY	LENFTSDVIS	RAAFGSSYEE	120
GRKIFQLLRE	EAKVYTVAVR	SVYIPGWRFL	PTKQNKKTKE	IHNEIKGLLK	GIINKREEAM	180
KAGEATKDDL	LGILMESNFR	EIQEHGNKN	AGMSIEDVIG	ECKLFYFAGQ	ETTSVLLVWT	240
MVLLSQNQDW	QARAREEVLO	VFGSNIPTYE	ELSHLKVVTM	ILLEVLRLYP	SVVALPRTH	300
KKTQLGKLSL	PAGVEVSLPI	LLVHHDKELW	GEDANEFKPE	RFSEGVSKAT	KNQFTYFPFG	360
GGPRICIQON	FAMMEAKLAL	SLILQHFTFE	LSPQYSHAPS	VTITLQPQYG	AHLILHKR	418

SEQ ID NO:105

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tcaagtggtc	tacctattat	cttagcactt	gcctctttag	cagacagatg	tggtcctatt	240
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caagctaacg	atgacaaaaa	ggaggatatg	gatttcatgg	atatcatgat	ctccatgaca	900
gaagcaaat	caccacttga	aggatacggc	actgatacta	ttatcaagac	cacatgatg	960
actttgattg	tttcaggagt	tgatacaacc	tcaatcgtac	ttacttgggc	cttatcactt	1020
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aaaggaagac	aagtcaacga	gtctgatctt	gttaacttga	tatacttggg	agcagtgcct	1140
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cgtgttaaat	ggtcctaa					1578

SEQ ID NO:106

MGLFPLEDSY	ALVFEGLAIT	LALYYLLSFI	YKTSKKTCTP	PKASGEHPIT	GHLNLLSGSS	60
GLPHLALASL	ADRCGPIFTI	RLGIRRVLVV	SNWEIAKEIF	TTHDLIVSNR	PKYLAAKILG	120
FNYVSFSFAP	YGPYWVGIRK	IIATKLMSSS	RLQKLQFVRV	FELENSMKSI	RESWKEKKDE	180
EGKVLVEMKK	WFWELNMNIV	LRTVAGKQYT	GTVDDADAKR	ISELFREWFH	YTGRFVVGDA	240
FPFLGWLDLG	GYKTMELVA	SRLDSMVSKW	LDEHRKKQAN	DDKKEDMDFM	DIMISMTEAN	300
SPLEGYGTDI	IIKTTCMTLI	VSGVDTTSIV	LTWALSLLL	NRDTLKKAE	ELDMCVGKGR	360
QVNESDLVNL	IYLEAVLKEA	LRLYPAAFLG	GPRAFLEDCT	VAGYRIPKGT	CLLINMWKLH	420
RDPNIWSDPC	EFKPERFLTP	NQKDVVDVIGM	DFELIPFGAG	RRYCPGTRLA	LQMLHIVLAT	480
LLQNFEMSTP	NDAFVDMTAS	VGMTNAKASP	LEVLLSPRVK	WS		522

SEQ ID NO:107

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agaatgtgtt	taggcaaaga	gtttgccagg	ttagaagtgt	tagcatttct	ccacaacatt	1320
gttaccact	ttaagtggga	tcttctaate	cctgatgaga	agatcgaata	tgatccaatg	1380
gctactccag	ctaagggtct	gccaaattaga	cttcatccac	accaagtcta	a	1431

SEQ ID NO:108

MIQVLTPIIL	FLIFFVFWKV	YKHQKTKINL	PPGSFGWPFL	GETLALLRAG	WDSEPERFVR	60
ERIKKHGSP	VFKTSLFGDR	FAVLCGPAGN	KFLFCNENKL	VASWWPVPVR	KLFGKSLITI	120
RGDEAKWMRK	MLLSYLGPD	FATHYAVTMD	VVTRRHIDVH	WRGKEEVNMF	QTVKLYAFEL	180
ACRLFMNLDD	PNHIAKLGSL	FNIFLKGIE	LPIDVPGTRF	YSSKKAIAI	RIELKKLIKA	240
RKLELKEGKA	SSSQDLLSHL	LTSPDENGFM	LTEEEIVDNI	LLLLFAGHDT	SALSITLLMK	300
TLGEHSDVYD	KVLKEQLEIS	KTKEAWESLK	WEDIQMKYS	WSVICEVMRL	NPPVIGTYRE	360
ALVDIDYAGY	TIPKGWKLHW	SAVSTORDEA	NFEDVTRFDP	SRFEGAGPTP	FTFVPGGGP	420
RMCLGKEFAR	LEVLAFLHNI	VTNFKWDLII	PDEKIEYDPM	ATPAKGLPIR	LHPHQV	476

SEQ ID NO:109

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gaaatgcaac	gtatccaatc	cgaagctaaa	cactgctctg	gcgataacat	tatctcacat	240
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agtacagctg	tctcagtgtc	atgggtgttg	atggttactgg	ccctaaacct	atcatggcaa	1080
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gtaattagag	tggtttaa					1578

SEQ ID NO:110

MESLVVHTVN	AIWCIVIVGI	FSVGYHVVYGR	AVVEQWRMRR	SLKLQGVKGP	PPSIFNGNVS	60
EMQRIQSEAK	HCSGDNIIISH	DYSSSLFPHF	DHWRKQYGRI	YTYSTGLKQH	LYINHPEMVK	120
ELSQTNTLNL	GRITHITKRL	NPILNGIIT	SNGPHWAHQ	RIIAYEFTHD	KIKGMVGLMV	180
ESAMPMLNKW	EEMVKRGGEM	GCDIRVDEDL	KDVSADVIK	ACFGSSFSKG	KAIFSMIRDL	240
LTAITKRSVL	FRFNGFTDMV	FGSKKHGDVD	IDALEMELES	SIWETVKERE	IECKDTHKKD	300
LMQLILEGAM	RSCDGNLWDK	SAYRRFVVVDN	CKSIYFAGHD	STAVSVSWCL	MLLALNPSWQ	360
VKIRDEILSS	CKNGIPDAES	IPNLKVTVMV	IQETMRLYPP	APIVGREASK	DIRLGLDLPV	420
KGVCIWTLIP	ALHRDPEIWG	PDANDFKPER	FSEGISKACK	YPQSYIPFGL	GPRTCVGKNF	480
GMMEVKVLVS	LIVSKFSFTL	SPTYQHSPSH	KLLVEPQHG	VIRVV		525

SEQ ID NO:111

atgtacttcc	tactacaata	cctcaacatc	acaaccgttg	gtgtctttgc	cacattgttt	60
ctctcttatt	gtttacttct	ctggagaagt	agagcgggta	acaaaaagat	tgcccagaa	120
gctgccgctg	catggcctat	tatcgccac	ctccacttac	ttgcaggtgg	atcccatcaa	180
ctaccacata	ttacattggg	taacatggca	gataagtacg	gtcctgtatt	cacaatcaga	240
ataggcttgc	atagagctgt	agttgtctca	tottgggaaa	tggcaaagga	atgttcaaca	300
gctaagtatc	aagtgtcttc	ttcaagacct	gaactattag	cttctaagtt	gttgggttat	360
aactacgcca	tgtttggttt	ttcaccatac	ggttcatact	ggagagaaat	gagaaagatc	420
atctctctcg	aattactatc	taattccaga	ttggaactat	tgaaagatgt	tagagcctca	480
gaagttgtca	catctattaa	ggaactatac	aaattgtggg	cggaaaagaa	gaatgagtca	540
ggattggttt	ctgtcgagat	gaaacaatgg	ttcggagatt	tgactttaaa	cgtgatcttg	600
agaatgggtg	ctggtaaaaag	atacttctcc	gcgagtgcag	cttcagaaaa	caaacaggcc	660
cagcgttgta	gaagagtctt	cagagaatcc	ttccatctct	ccggcttggt	tgtggttgct	720
gatgctatac	cttttcttgg	atggctcgat	tggggaagac	acgagaagac	cttgaaaaag	780
accgccatag	aatggattc	catcgcccag	gagtggtctg	aggaacatag	acgtagaaaa	840
gattctggag	atgataattc	tacccaagat	ttcatggacg	ttatgcaatc	tgtgctagat	900
ggcaaaaatc	taggcggata	cgatgctgat	acgattaaca	aggctacatg	cttaactctt	960
atatacagg	gcagtgatac	tactgtagtt	tctttgacat	gggctcttag	tcttgtgtta	1020
aacaatagag	atactttgaa	aaaggcacag	gaagagttag	acatccaagt	cggtaaggaa	1080
agattgggta	acgagcaaga	catcagtaag	ttagtttact	tgcaagcaat	agtaaaagag	1140
acactcagac	tttatccacc	aggtcctttg	ggtgggttga	gacaattcac	tgaagattgt	1200
acactaggtg	gctatcacgt	ttcaaaaagga	actagattaa	tcatgaactt	atccaagatt	1260
caaaaagatc	cacgtatttg	gtctgatctc	actgaattcc	aaccagagag	attccttacg	1320

actcataaag	atgtcgatcc	acgtggtaaa	cactttgaat	tcattccatt	cgggtgcagga	1380
agacgtgcat	gtcctgggat	cacattcgga	ttacaagtac	tacatctaac	attggcatct	1440
ttcttgcgat	cgtttgaatt	ttcaacacca	tcaaatgagc	aggttaacat	gagagaatca	1500
ttaggtctta	cgaatatgaa	atctacccca	ttagaagttt	tgattttctcc	aagactatcc	1560
cttaattgct	tcaaccttat	gaaaatttga				1590

SEQ ID NO:112

MYFLLQYLNI	TTVGVFATLF	LSYCLLLWRS	RAGNKKIAPE	AAAAWPIIGH	LHLLAGGSHQ	60
LPHITLGNMA	DKYGPVFTIR	IGLHRAVVVS	SWEMAKECST	ANDQVSSSRP	ELLASKLLGY	120
NYAMFGFSY	GSYWREMRKI	ISLELLSNSR	LELLKDVRAS	EVVTSIKELY	KLWAEKKNES	180
GLVSVEMKQW	FGDLTLNVIL	RMVAGKRYFS	ASDASENKQA	QRCRRVFREF	FHLSGLFVVA	240
DAIPFLGWLD	WGRHEKTLKK	TAIEMDSIAQ	EWLEEHRRRK	DSGDDNSTQD	FMDVMQSVLD	300
GKNLGGYDAD	TINKATCLTL	ISGGSDDTVV	SLTWALSLVL	NNRDTLKKAQ	EELDIQVGKE	360
RLVNEQDISK	LVYLQAIVKE	TLRLYPPGPL	GGLRQFTEDC	TLGGYHVSKE	TRLIMNLSKI	420
QKDPRIWSDP	TEFQPERFLT	THKDVDPRGK	HFEFIPFGAG	RRACPGITFG	LQVLHLLTAS	480
FLHAFEFSTP	SNEQVNMRES	LGLTNMKSTP	LEVLISPRLS	SCSLYN		526

SEQ ID NO:113

atggaaccta	acttttactt	gtcattacta	ttggtgttcg	tgaccttcat	ttctttaagt	60
ctgtttttca	tcttttacia	acaaaagtcc	ccattgaatt	tgccaccagg	gaaaatgggt	120
taccctatca	taggtgaaag	tttagaattc	ctatccacag	gctggaaggg	acatcctgaa	180
aagttcatat	ttgatagaat	gcgtaagtac	agtagtgagt	tattcaagac	ttctattgta	240
ggcgaatcca	cagttgtttg	ctgtggggca	gctagtaaca	aattcctatt	ctctaacgaa	300
aacaaactgg	taactgcctg	gtggccagat	tctgttaaca	aaatcttccc	aacaacttca	360
ctggattcta	atltgaaagga	ggaatctata	aagatgagaa	agttgctgcc	acagttcttc	420
aaaccagaag	cacttcaaag	atacgtcggc	gttatggatg	taatcgaca	aagacatttt	480
gtcactcact	gggacaacaa	aaatgagatc	acagtttata	cacttgctaa	aagatacact	540
ttcttgcttg	cgtgtagact	gttcatgtct	gttgaggatg	aaaatcatgt	ggcgaaattc	600
tcagacccat	tccaactaat	cgctgcagge	atcatttcac	ttcctatcga	tcttctctgg	660
actccattca	acaaggccat	aaaggcttca	aatttcatta	gaaaagagct	gataaagatt	720
atcaaacaaa	gacgtgttga	tctggcagag	ggtacagcat	ctccaacca	ggataltctg	780
tcacatatgc	tattaacatc	tgatgaaaac	ggtaaatcta	tgaacgagtt	gaacattgcc	840
gacaagattc	ttggactatt	gataggaggc	cacgatacag	cttcagtagc	ttgcacattt	900
ctagtgaagt	acttaggaga	attaccacat	atctacgata	aagtctacca	agagcaaatg	960
gaaattgccca	agtccaaacc	tgctggggaa	ttggtgaatt	gggatgactt	gaaaaagatg	1020
aagtattcat	ggaatgtggc	atgtgaggta	atgagattgt	caccaccttt	acaagggtgg	1080
tttagagagg	ctataactga	ctttatgttt	aacggtttct	ctattccaaa	aggggtggaag	1140
ttatactggg	ccgccaactc	tacacacaaa	aatgcagaat	gtttcccaat	gcctgagaaa	1200
ttcgatccta	ccagatttga	aggtaatggg	ccagcgcctt	atacatttgt	accattcggg	1260
ggaggcccta	gaatgtgtcc	tggaaaggaa	tacgctagat	tagaaatctt	ggttttcatg	1320
cataatctgg	tcaaactgtt	taagtgggaa	aaggttattc	cagacgaaaa	gattattgtc	1380
gatccattcc	caatcccagc	taaagatctt	ccaatccggt	tgtatcctca	caaagcttaa	1440

SEQ ID NO:114

MEPNFYLSLL	LLFVTFISLS	LFFIFYKQKS	PLNLPPGKMG	YPIIGESLEF	LSTGWKGHPE	60
KFIFDRMRKY	SSELFKTSIV	GESTVVCCGA	ASNKFLFSNE	NKLVTAWWPD	SVNKIFPTTS	120
LDSNLKEESI	KMRKLLPQFF	KPEALQRYVG	VMDVIAQRHF	VTHWDNKNEI	TVYPLAKRYT	180
FLLACRLFMS	VEDENHVAKF	SDPFQLIAAG	IISLPIDLPG	TPFNKAIKAS	NFIRKELIKI	240
IKQRRVDLAE	GTASPTQDIL	SHMLLTSDEN	GKSMNELNIA	DKILGLLIGG	HDTASVACTF	300
LVKYLGELPH	IYDKVYQEQM	EIAKSKPAGE	LLNWDDLKMM	KYSWNVACEV	MRLSPPLQGG	360
FREAITDFMF	NGFSIPKGWK	LYWSANSTHK	NAECFPMPEK	FDPTRFEGNG	PAPYTFVPFG	420
GGPRMCPGKE	YARLEILVFM	HNLVKRFKWE	KVIPDEKIIIV	DPFPIPAKDL	PIRLYPHKA	479

SEQ ID NO:115

atggcctctg	ttactttggg	ttcctggatc	gtcgtccacc	accataacca	tcaccatcca	60
tcactatctc	taactaaatc	tcgttcaaga	tcctgtccta	ttacactaac	caaccaatc	120
tcttttcggt	caaagagaac	agtttctctc	agtagttcta	tcgtgtcctc	tagtgcctc	180

actaaggaag	acaatctgag	acagtctgaa	ccttcttct	ttgatttcat	gtcatatatac	240
attactaagg	cagaactagt	gaataaggct	cttgattcag	cagttccatt	aagagagcca	300
ttgaaaatcc	atgaagcaat	gagatactct	cttctagctg	gcggaagag	agtcagacct	360
gtactctgca	tagcagcgtg	cgaattagtt	ggtggcgagg	aatcaaccgc	tatgectgcc	420
gcttgtgctg	tagaaatgat	tcatacaatg	tcactgatac	acgatgattt	gccatgtatg	480
gataacgatg	atctgagaag	gggtaagcca	actaaccata	aggttttcgg	cgaagatggt	540
gccgtcttag	ctggtgatgc	tttgttatct	ttegcgttcg	aacatttggc	atccgcaaca	600
tcaagtgatg	ttgtgtcacc	agtaagagta	gtagagcag	ttggagaact	ggctaaagct	660
attggaactg	agggtttagt	tgcagggtcaa	gtcgtcgata	tctcttccga	aggctctgat	720
ttgaatgatg	taggtcttga	acatctcgaa	ttcatccatc	ttcacaagac	agctgcactt	780
ttagaagcca	gtgctgttct	cggcgcaatt	gtaggaggag	ggagtgatga	cgaaattgag	840
agattgagga	agtttgctag	atgtatagga	ttactgttcc	aagtagtaga	cgatatacta	900
gatgtgacaa	agtcttccaa	agagttggga	aaaacagctg	gtaaagattt	gattgccgac	960
aaattgacct	accctaagat	tatggggcta	gaaaaatcaa	gagaatttgc	cgagaaactc	1020
aatagagagg	cgcgtgatca	actggtgggt	ttcgattctg	ataaagttgc	accactctta	1080
gccttagcca	actacatcgc	ttacagacaa	aactaa			1116

SEQ ID NO:116

MASVTLGSKI	VVHHHHHHHP	SSILTKRSR	SCPITLTKPI	SFRSKRTVSS	SSSIVSSSVV	60
TKEDNLRQSE	PSSFDMSYI	ITKAELVNKA	LDSAVPLREP	LKIHEAMRYS	LLAGGKRVRP	120
VLCIAACELV	GGEESTAMPA	ACAVEMIHTM	SLIHDDLPCM	DNDDLRRGKP	TNHKVFGEDEV	180
AVLAGDALLS	FAFEHLASAT	SSDVVSPVRV	VRAVELAKA	IGTEGLVAGQ	VVDISSEGLD	240
LNDVBLEHLE	FIHLHKTAAL	LEASAVLGAI	VGGGSDDEIE	RLRKFCARCIG	LLFQVDDIL	300
DVTKSSKELG	KTAGKDLIAD	KLTYPKIMGL	EKSREFAEKL	NREARDQLLG	FDSKQVAPLL	360
ALANYIAYRQ	N					371

SEQ ID NO:117

MATLLEHFQA	MPFAIPIALA	ALSWLFLFYI	KVSFFSNKSA	QAKLPPVPVV	PGLPVIIGNLL	60
QLKEKPYQT	FTRWAEYGP	IYSIRTGAST	MVVLNTTQVA	KEAMVTRYLS	ISTRKLSNAL	120
KILTADKCMV	AISDYNDFHK	MIKRYILSNV	LGPSAQKRHR	SNRDTLRANV	CSRLHSQVKN	180
SPREAVNFRR	VFEWELFGIA	LKQAFGKDIE	KPIYVEELGT	TLRDEIFKV	LVLDMEGAI	240
EVDWRDFFPY	LRWIPNTRME	TKIQRLYFRR	KAVMTALINE	QKKRIASGEE	INCYIDFLK	300
EGKTLTMDQI	SMLLWETVIE	TADTTMVTTE	WAMYEVAKDS	KRQDRLYQEI	QKVCGSEMVT	360
EEYLSQLPYL	NAVFHETLRK	HSPAALVPLR	YAHEDTQLGG	YYIPAGTEIA	INIYGCNMDK	420
HQWESPEEWK	PERFLDPKFD	PMDLYKTMAF	GAGKRVCAGS	LQAMLIACPT	IGRLVQEFEW	480
KLRDGEENV	DTVGLTTHKR	YPMHAILKPR	S			511

SEQ ID NO:118

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVYKA	PGISDGYRMG	MVLKGSDDL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKEVSARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAI	HES	473

SEQ ID NO:119

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARGVPYKA	PGISDGYRMG	LVLKGSDDL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKEVSARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAI	HES	473

SEQ ID NO:120

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPST	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLPPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:121

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPST	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVYKA	PGISDGYRMG	HVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:122

MDSGYSSSYA	AAAGMHVVIC	PWLAFGHLLP	CLDLAQRLAS	RGHRVSFVST	PRNISRLPPV	60
RPALAPLVAF	VALPLPRVEG	LPDGAESTND	VPHDRPDMVE	LHRRAFDGLA	APFSEFLGTA	120
CADWVIVDVF	HHWAAAAALE	HKVPCAMMLL	GSAHMIAASIA	DRRLERAETE	SPAAAGQGRP	180
AAAPTFEVAR	MKLIRTKGSS	GMSLAERFSL	TLSRSSLVVG	RSCVEFEPET	VPLLSTLRGK	240
PITFLGLMPP	LHEGREDGE	DATVRWLDAQ	PAKSVVYVAL	GSEVPLGVEK	VHELALGLEL	300
AGTRFLWALR	KPTGVSDADL	LPAGFEERTR	GRGVVATRWW	PQMSILAHAA	VGAFTHCGW	360
NSTIEGLMFG	HPLIMLPFIG	DQGPNARLIE	AKNAGLQVPR	NEEDGCLTKE	SVARSLRSVV	420
VEKEGEIYKA	NARELSKIYN	DTKVEKEYVS	QFVDYLEKNA	RAVAIDHES		469

SEQ ID NO:123

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPST	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARCVPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:124

MDSGYSSSYA	AAAGMHVVIC	PWLAFGHLLP	CLDLAQRLAS	RGHRVSFVST	PRNISRLPPV	60
RPALAPLVAF	VALPLPRVEG	LPDGAESTND	VPHDRPDMVE	LHRRAFDGLA	APFSEFLGTA	120
CADWVIVDVF	HHWLPSIAAS	LGISRAHFSV	TTPWAIAYMG	PSADAMINGS	DGRITVEDLT	180
TPPKWFPFPT	KVCWRKHDLA	RLVPYKAPGI	SDGYRMGMVL	KGSDCLLSKC	YHEFGTQWLP	240
LLETLHQVPV	VPVGLMPPLH	EGRREDGEDA	TVRWLDAQPA	KSVVYVALGS	EVPLGVEKVH	300
ELALGLELAG	TRFLWALRKP	TGVSDADLLP	AGFEERTRGR	GVVATRWPVQ	MSILAHAAVG	360
AFLTHCGWNS	TIEGLMFGHP	LIMLPFIGDQ	GPNARLIEAK	NAGLQVARND	GDGSFDREGV	420
AAAIRAVAVE	EESKVFQAK	AKKLOEIVAD	MACHERYIDG	FIQQLRSYKD		470

SEQ ID NO:125

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWAAAA	ALEHKVPCAM	MLLGSAHMIA	SIADRRLERA	ETESPAAAGQ	GRPAAAPTFE	180
VARMKLIRTK	GSSGMSLAER	FSLTLSRSSL	VVGRSCVEFE	PETVPLLSTL	RGKPITFLGL	240
MPPLHEGRRE	DGEDATVRWL	DAQPAKSVVY	VALGSEVPLG	VEKVHELALG	LELAGTRFLW	300
ALRKPTGVSD	ADLLPAGFEE	RTRGRGVVAT	RWVPQMSILA	HAAVGAFLTH	CGWNSTIEGL	360

MFGHPLIMLP IFGDQGP NAR LIEAKNAGLQ VARNDGDGSF DREGVAAAIR AVAVEEESSK 420  
 VFQAKAKKLQ EIVADMACHE RYIDGFIQQL RSYKD 455

SEQ ID NO:126

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLK KASD GLQPEVTRFL EQHSPDWI IY 120  
 DYTHYWLPSI AASLGISRAH FSVTPWAIA YMGPSADAMI NGSDGR TTVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVPYKA FGISDGYRMG MVLKGS DCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEALVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIARN DGDGS FDREGVAAAI 420  
 RAVAVEEESS KVFQAKAKKL QEIVADMACH ERYIDGFIQQL LRSYKD 466

SEQ ID NO:127

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIP IRES LLRSIETNFL DRFIDLVT KL PDPPTCIISD 120  
 GFSLVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVLMI TTEAPQRS HK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVVYVN 300  
 FGSSTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGF IASWC 360  
 SQEKVLKHP S VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITV LAR 480  
 N 481

SEQ ID NO:128

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLK KASD GLQPEVTRFL EQHSPDWI IY 120  
 DYTHYWAAAA ALEHKVPCAM MLLGSAHMIA SIADRR LERA ETESPAAAGQ GRPAAAPTFE 180  
 VARMKLIRTK GSSGMSLAER FSLT LSRSSL VVGRSCVEFE PETVPLLSTL RGKPI TFLGL 240  
 LPPEIPGDEK DETWVSIKKW LDGKQKGSVV YVALGSEALV SQTEVVELAL GLELSGLPFV 300  
 WAYRKPKGPA KSDSVELPDG FVERTDRGL VWTSWAPQLR ILSHESVCGF LTHCGSGSIV 360  
 EGLMFGHPLI MLP IFGDQPL NARLLEDKQV GIEIPRNEED GCLTKESVAR SLRSVVVEKE 420  
 GEIYKANARE LSKIYNDTKV EKEYVSQFVD YLEKNARAVA IDHES 465

SEQ ID NO:129

MDSGYSSSYA AAAGMHVVIC PWLAFGHLLP CLDLAQRLAS RGHRSV FVST PRNISRLPPV 60  
 RPALAPL VAF VALPLPRVEG LPDGAESTND VPHDRPDMVE LHRRAFDGLA APFSEFLGTA 120  
 CADWVIVDVF HHWLPSIAAS LGISRAHFSV TTPWAIAYMG PSADAMINGS DGRTTVEDLT 180  
 TPPKWFPPPT KVCWRKHDLA RLVYPYKAPGI SDGYRMGMVL KGS DCLLSK YHEFGTQWLP 240  
 LLET LHQVPV VPVGLLPEI PGDEKDETVV SIKKWLDGKQ KGSVVYVALG SEALVSQTEV 300  
 VELALGLELS GLPFVWAYRK PKGPAKSDSV ELPDGFVERT RDRGLVWTSW APQLRILSHE 360  
 SVCGFLTHCG SGSIVEGLMF GHPLIMLP I FGDQPLNARLL EDKQVGIEIP RNEEDGCLTK 420  
 ESVARSLRSV VVEKEGEIYK ANARELSKIY NDTKVEKEYV SQFVDYLEKN ARAVAIDHES 480

SEQ ID NO:130

MDSGYSSSYA AAAGMHVVIC PWLAFGHLLP CLDLAQRLAS RGHRSV FVST PRNISRLPPV 60  
 RPALAPL VAF VALPLPRVEG LPDGAESTND VPHDRPDMVE LHRRAFDGLA APFSEFLGTA 120  
 CADWVIVDVF HHWAAAAALE HKVPCAMMLL GSAHMIASIA DRRLERAE TE SPAAAGQGRP 180  
 AAAPTFEVAR MKLIRTKGSS GMSLAERFSL T LSRSSLVVG RSCVEFEPET VPLLSTLRGK 240  
 PITFLGLLPP EIPGDEKDET WWSIKKWLDG KQKGSVVYVA LGSEALVSQT EVVELALGLE 300  
 LSGLPFVWAY RKPKGPAKSD SVELPDGFVE RTRDRGLVWT SWAPQLRILS HESVCGFLTH 360  
 CGSGSIVEGL MFGHPLIMLP IFGDQPLNAR LLEDKQVGIE IPRNEEDGCL TKESVARSLR 420  
 SVVVEKEGEI YKANARELSK IYNDTKVEKE YVSQFVDYLE KNARAVAI DH ES 472

SEQ ID NO:131

MDSGYSSSYA AAAGMHVVIC PWLAFGHLLP CLDLAQRLAS RGHRSV FVST PRNISRLPPV 60  
 RPALAPL VAF VALPLPRVEG LPDGAESTND VPHDRPDMVE LHRRAFDGLA APFSEFLGTA 120

CADWVIVDVF	HHWLPSIAAS	LGISRAHFSV	TTPWAIAYMG	PSADAMINGS	DGRTTVEDLT	180
TPPKWFPFPT	KVCWRKHDLA	RLVPYKAPGI	SDGYRMGMVL	KGSDCLLSKC	YHEFGTQWLP	240
LLETLHQVPV	VPVGLLPPEI	PGDEKDETVV	SIKKWLDGKQ	KGSVVYVALG	SEALVSQTEV	300
VELALGLELS	GLPFVWAYRK	PKGPAKSDSV	ELPDGFVERT	RDRGLVWTSW	APQLRILSHE	360
SVCGFLTHCG	SGSIVEGLMF	GHPLIMLPIF	GDQPLNARLL	EDKQVGIEIA	RNDGDGSFDR	420
EGVAAAIRAV	AVEEESKVF	QAKAKKLOEI	VADMACHERY	IDGFIQQLRS	YKD	473

SEQ ID NO:132

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIIY	120
DYTHYWLPST	AASLGISRAH	FSVTPWAIAT	YMGPSADAMI	NGSDGRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	MVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLMP	PLHEGRREDG	EDATVRWLDA	QPAKSVVYVA	LGSEVPLGVE	KVHELALGLE	300
LAGTRFLWAL	RKPTGVSDAD	LLPAGFEERT	RGRGVVATRW	VPQMSILAHA	AVGAFSLTHCG	360
WNSTIEGLMF	GHPLIMLPIF	GDQGPANARLI	EAKNAGLQVA	RNDGDGSFDR	EGVAAAIRAV	420
AVEEESKVF	QAKAKKLOEI	VADMACHERY	IDGFIQQLRS	YKD		463

SEQ ID NO:133

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIIY	120
DYTHYWAAAA	ALEHKVPCAM	MLLGSAHMIA	SIADRRLERA	ETESPAAAGQ	GRPAAAPTFE	180
VARMKLIRTK	GSSGMSLAER	FSLTSLRSSL	VVGRSCVEFE	PETVPLLSTL	RGKPITFLGL	240
MPPLHEGRRE	DGEDATVRWL	DAQPAKSVVY	VALGSEVPLG	VEKVHELALG	LELAGTRFLW	300
ALRKPTGVSD	ADLLPAGFEE	RTRGRGVVAT	RWVPQMSILA	HAAVGAFSLH	CGWNSTIEGL	360
MFGHPLIMLP	IFGDQGPANAR	LIEAKNAGLQ	VPRNEEDGCL	TKESVARSLR	SVVVEKEGEI	420
YKANARELSK	IYNDTKVEKE	YVSQFVDYLE	KNARAVAIIDH	ES		462

SEQ ID NO:134

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIIY	120
DYTHYWAAAA	ALEHKVPCAM	MLLGSAHMIA	SIADRRLERA	ETESPAAAGQ	GRPAAAPTFE	180
VARMKLIRTK	GSSGMSLAER	FSLTSLRSSL	VVGRSCVEFE	PETVPLLSTL	RGKPITFLGL	240
LPPEIPGDEK	DETVWSIKKW	LDGKQKGSVV	YVALGSEALV	SQTEVVELAL	GLELSGLPFV	300
WAYRKPKGPA	KSDSVELPDG	FVERTDRGL	VWTSWAPQLR	ILSHESVCGF	LTHCGSGSIV	360
EGLMFGHPLI	MLPIFGDQPL	NARLLEDKQV	GIEIARNDDG	GSFDREGVAA	AIRAVAVEEE	420
SSKVFQAKAK	KLQEIADMA	CHERYIDGFI	QQLRSYKD			458

SEQ ID NO:135

ggcaagccac gtttggtg 18

SEQ ID NO:136

ggagctgcat gtgtcagagg 20

SEQ ID NO:137

cgatgtattt catcactggt tgccatccat cgcggct 37

SEQ ID NO:138

agccgcgatg gatggcaacc agtgatgaaa tacatcg 37

SEQ ID NO:139

ttatgattat actcactact gggctgctgc agccgcattg 40

SEQ ID NO:140

agccgcgatg gatggcaacc agtgatgaaa tacatcg 37

SEQ ID NO:141

caaacctatt actttccttg gtttactgcc accggaata c 41

**SEQ ID NO:142**

gtatttcggg tggcagtaaa ccaaggaaag taataggttt g 41

**SEQ ID NO:143**

ccggtgggtc cggtaggact aatgcctcca ttacatga 38

**SEQ ID NO:144**

tcatgtaatg gaggcattag tcccaccgga accaccgg 38

**SEQ ID NO:145**

gaacgcaggt ctgcaggttc caagaaatga ggaagatgg 39

**SEQ ID NO:146**

ccatcttcct catttcttg aacctgcaga cctgcgttc 39

**SEQ ID NO:147**

MDAMATTEKK PHVIFIPFPA LSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPRES LLRSIETNFL DRFIDLVTCL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVLME TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFASWC 360  
 SQEKVLKHPV VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

**SEQ ID NO:148**

MDAMATTEKK PHVIFIPFPA TSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPRES LLRSIETNFL DRFIDLVTCL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVLME TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFASWC 360  
 SQEKVLKHPV VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

**SEQ ID NO:149**

MDAMATTEKK PHVIFIPFPA VSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPRES LLRSIETNFL DRFIDLVTCL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVLME TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFASWC 360  
 SQEKVLKHPV VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

**SEQ ID NO:150**

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDSIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPRES LLRSIETNFL DRFIDLVTCL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVLME TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300

FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH	KKRGFIASWC	360
SQEKVLKHP	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:151

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDHIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPPIRES	LLRSIETNFL	DRFIDLVTCL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLME	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH	KKRGFIASWC	360
SQEKVLKHP	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:152

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDYIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPPIRES	LLRSIETNFL	DRFIDLVTCL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLME	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH	KKRGFIASWC	360
SQEKVLKHP	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:153

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDRIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPPIRES	LLRSIETNFL	DRFIDLVTCL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLME	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH	KKRGFIASWC	360
SQEKVLKHP	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:154

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDQIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPPIRES	LLRSIETNFL	DRFIDLVTCL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLME	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH	KKRGFIASWC	360
SQEKVLKHP	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:155

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDWIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPPIRES	LLRSIETNFL	DRFIDLVTCL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLME	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH	KKRGFIASWC	360
SQEKVLKHP	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420

TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

**SEQ ID NO:156**

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDTIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFASWC 360  
 SQEKVLKHP S VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

**SEQ ID NO:157**

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFVH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFASWC 360  
 SQEKVLKHP S VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

**SEQ ID NO:158**

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEAGIPIRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFASWC 360  
 SQEKVLKHP S VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

**SEQ ID NO:159**

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEAAIPIRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFASWC 360  
 SQEKVLKHP S VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

**SEQ ID NO:160**

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEATIPIRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFASWC 360  
 SQEKVLKHP S VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

SEQ ID NO:161

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEACIPIRES	LLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLMF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEHI	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:162

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEAPIPIRES	LLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLMF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEHI	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:163

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEANIPIRES	LLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLMF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEHI	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:164

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEAVIPIRES	LLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLMF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEHI	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:165

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIRIRES	LLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVLMF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVYN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEHI	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLAR	480
N						481

SEQ ID NO:166

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIGIRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFIASWC 360  
 SQEKVLKHPS VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

SEQ ID NO:167

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPHRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFIASWC 360  
 SQEKVLKHPS VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

SEQ ID NO:168

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPPRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFIASWC 360  
 SQEKVLKHPS VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

SEQ ID NO:169

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPMRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFIASWC 360  
 SQEKVLKHPS VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

SEQ ID NO:170

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60  
 CLDGAPGFRF ETIPDGVSHS PEASIPYRES LLRSIETNFL DRFIDLVTKL PDPPTCIISD 120  
 GFLSVFTIDA AKKLGIPVMM YWTLAACGFM GFYHIHSLIE KGFAPLKDAS YLTNGYLDTV 180  
 IDWVPGMEGI RLKDFPLDWS TDLNDKVL MF TTEAPQRSHK VSHHIFHTFD ELEPSIIKTL 240  
 SLRYNHIYTI GPLQLLLDQI PEEKKQTGIT SLHGYSLVKE EPECFQWLQS KEPNSVYVYN 300  
 FGSTTVMSLE DMTEFGWGLA NSNHYFLWII RSNLVIGENA VLPPELEEHI KKRGFIASWC 360  
 SQEKVLKHPS VGGFLTHCGW GSTIESLSAG VPMICWPYSW DQLTNCRYIC KEWEVGLEMG 420  
 TKVKRDEVKR LVQELMGEGG HKMRNKAKDW KEKARIAIAP NGSSSLNIDK MVKEITVLAR 480  
 N 481

SEQ ID NO:171

MDAMATTEKK PHVIFIPFPA QSHIKAMLKL AQLLHHKGLQ ITFVNTDFIH NQFLESSGPH 60

CLDGAPGFRF	ETIPDGVSHS	PEASIPARES	KLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITV LAR	480
N						481

SEQ ID NO:172

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPARES	RLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITV LAR	480
N						481

SEQ ID NO:173

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPARES	TLRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITV LAR	480
N						481

SEQ ID NO:174

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPARES	LFRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITV LAR	480
N						481

SEQ ID NO:175

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPARES	LIRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITV LAR	480
N						481

SEQ ID NO:176

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIPARES	LMRSIETNFL	DRFIDLVTKL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180

IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLR	480
N						481

SEQ ID NO:177

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIP IRES	LLRSKETNFL	DRFIDLVT KL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLR	480
N						481

SEQ ID NO:178

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIP IRES	LLRSIETNFL	DRFIDLVT KL	PDPPTCIISD	120
GSLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNLVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLR	480
N						481

SEQ ID NO:179

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIP IRES	LLRSIETNFL	DRFIDLVT KL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNSVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLR	480
N						481

SEQ ID NO:180

MDAMATTEKK	PHVIFIPFPA	QSHIKAMLKL	AQLLHHKGLQ	ITFVNTDFIH	NQFLESSGPH	60
CLDGAPGFRF	ETIPDGVSHS	PEASIP IRES	LLRSIETNFL	DRFIDLVT KL	PDPPTCIISD	120
GFLSVFTIDA	AKKLGIPVMM	YWTLAACGFM	GFYHIHSLIE	KGFAPLKDAS	YLTNGYLDTV	180
IDWVPGMEGI	RLKDFPLDWS	TDLNDKVL MF	TTEAPQRSHK	VSHHIFHTFD	ELEPSIIKTL	240
SLRYNHIYTI	GPLQLLLDQI	PEEKKQTGIT	SLHGYSLVKE	EPECFQWLQS	KEPNSVYVN	300
FGSTTVMSLE	DMTEFGWGLA	NSNHYFLWII	RSNMVIGENA	VLPPELEEH I	KKRGFIASWC	360
SQEKVLKHPS	VGGFLTHCGW	GSTIESLSAG	VPMICWPYSW	DQLTNCRYIC	KEWEVGLEMG	420
TKVKRDEVKR	LVQELMGEGG	HKMRNKAKDW	KEKARIAIAP	NGSSSLNIDK	MVKEITVLR	480
N						481

SEQ ID NO:181

MENKTETTVR	RRRIILFPV	PFHGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGFLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFAHAVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300

DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

**SEQ ID NO:182**

MENKTETTVR	RRRRIILFPV	PFQGHWPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

**SEQ ID NO:183**

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMGSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

**SEQ ID NO:184**

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHALVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

**SEQ ID NO:185**

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSGLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

**SEQ ID NO:186**

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TAWSSSLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

**SEQ ID NO:187**

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120

LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFWLDQ	QPPSSVLYVS	FGSGSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:188

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGFLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFWLDQ	QPPSSVLYVS	FGNTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:189

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGFLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVPWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:190

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKKSNYPH	60
FTFRFILDND	PQDERISNLP	THGFLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHAHVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMVDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:191

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	LVEKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVSCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAI	HES	473

SEQ ID NO:192

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTTPWAIA	YMGPSADAMI	NGSDGRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	LVLKGSDCLL	YKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVSCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAI	HES	473

SEQ ID NO:193

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARHLS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:194

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YTGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:195

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVYKA	PGISDGYRMG	CVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:196

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:197

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:198

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRITVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHEVCGFLT	360

HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:199**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKASD GLQPEVTRFL EQHSPDWIYY 120  
 DYTHYWLPSI AASLGISRAH FSVTPWAIA YMGPSADAMI NGSDGRITVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEVLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:200**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIVYLKASD GLQPEVTRFL EQHSPDWIYY 120  
 DYTHYWLPSI AASLGISRAH FSVTPWAIA YGSPADAMI NGSDGRITVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEVLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:201**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKASD GLQPEVTRFL EQHSPDWIYY 120  
 DYTHYWLPSI AASLGISRAH FSVTPWAIA YMGPSADAMI NGSDGRITVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEVLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:202**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKASD GLQPEVTRFL EQHSPDWIYY 120  
 DYTHYWLPSI AASLGISRAH FSVKTPWAIA YMGPSADAMI NGSDGRITVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEVLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:203**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKASD GLQPEVTRFL EQHSPDWIYY 120  
 DYTHYWLPSI AASLGISRAH FSVLTPWAIA YMGPSADAMI NGSDGRITVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEVLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:204**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKASD GLQPEVTRFL EQHSPDWIYY 120  
 DYTHYWLPSI AASLGISRAH FSVMPWAIA YMGPSADAMI NGSDGRITVE DLTPPKWFP 180

FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:205

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPPWKIA	YMGPSADAMI	NGSDGRRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	IVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:206

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARNVPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:207

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	LVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:208

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	MVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVGEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARVVAID	HES	473

SEQ ID NO:209

MATSDSIVDD	RKQLHVATFP	WLAFGHILPY	LQLSKLIAEK	GHKVSFLSTT	RNIQRLSSHI	60
SPLINVVQLT	LPRVQELPED	AEATTDVHPE	DIPYLKKASD	GLQPEVTRFL	EQHSPDWIYY	120
DYTHYWLPSI	AASLGISRAH	FSVTPWAIA	YMGPSADAMI	NGSDGRRTTVE	DLTTPPKWFP	180
FPTKVCWRKH	DLARLVPYKA	PGISDGYRMG	TVLKGSDCLL	SKCYHEFGTQ	WLPLLETLHQ	240
VPVVPVGLLP	PEIPGDEKDE	TWVSIKKWLD	GKQKGSVVYV	ALGSEVLVSQ	TEVVELALGL	300
ELSGLPFVWA	YRKPKGPAKS	DSVELPDGFV	ERTRDRGLVW	TSWAPQLRIL	SHESVCGFLT	360
HCGSGSIVEG	LMFGHPLIML	PIFGDQPLNA	RLLEDKQVGI	EIPRNEEDGC	LTKESVARSL	420
RSVVVEKEGE	IYKANARELS	KIYNDTKVEK	EYVSQFVDYL	EKNARAVAID	HES	473

SEQ ID NO:210

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKKASD GLQPEVTRFL EQHSPDWIIY 120  
 DYTHYWLPSI AASLGISRAH FSVTTPWAIA YMGPSADAMI NGSDGRTTVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVPYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSENLSVQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:211**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKKASD GLQPEVTRFL EQHSPDWIIY 120  
 DYTHYWLPSI AASLGISRAH FSVTTPWAIA YMGPSADAMI NGSDGRTTVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVPYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSESLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:212**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKKASD GLQPEVTRFL EQHSPDWIIY 120  
 DYTHYWLPSI AASLGISRAH FSVTTPWAIA YMGPSADAMI NGSDGRTTVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVPYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEVLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFKDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:213**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKKASD GLQPEVTRFL EQHSPDWIIY 120  
 DYTHYWLPSI AASLGISRAH FSVTTPWAIA YMGPSADAMI NGSDGRTTVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVPYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEVLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFYDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARELS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:214**

MATSDSIVDD RKQLHVATFP WLAFGHILPY LQLSKLIAEK GHKVSFLSTT RNIQRLSSHI 60  
 SPLINVVQLT LPRVQELPED AEATTDVHPE DIPYLKKASD GLQPEVTRFL EQHSPDWIIY 120  
 DYTHYWLPSI AASLGISRAH FSVTTPWAIA YMGPSADAMI NGSDGRTTVE DLTPPKWFP 180  
 FPTKVCWRKH DLARLVPYKA PGISDGYRMG LVLKGSDCLL SKCYHEFGTQ WLPLLETLHQ 240  
 VPVVPVGLLP PEIPGDEKDE TWVSIKKWLD GKQKGSVVYV ALGSEVLVSQ TEVVELALGL 300  
 ELSGLPFVWA YRKPKGPAKS DSVELPDGFV ERTRDRGLVW TSWAPQLRIL SHESVCGFLT 360  
 HCGSGSIVEG LMFHGHLIML PIFGDQPLNA RLLEDKQVGI EIPRNEEDGC LTKESVARSL 420  
 RSVVVEKEGE IYKANARMLS KIYNDTKVEK EYVSQFVDYL EKNARAVAID HES 473

**SEQ ID NO:215**

ATCAACGGGUA AAAATGGATGCTATGGCTACCACCG

**SEQ ID NO:216**

CGTGCGAUTCAGTTTCTGGCCAAAACGGTGATT

**SEQ ID NO:217**

MENKTETTVR	RRRRIILFPV	PFHGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHALVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMDDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:218

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMGSSLF	NFHALVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMDDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:219

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHALVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSGLDH	DRTVFQWLDQ	QPPSSVLYVS	FGSTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMDDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

SEQ ID NO:220

MENKTETTVR	RRRRIILFPV	PFQGHINPIL	QLANVLYSKG	FSITIFHTNF	NKPKTSNYPH	60
FTFRFILDND	PQDERISNLP	THGPLAGMRI	PIINEHGADE	LRRELELLML	ASEEDEEVSC	120
LITDALWYFA	QSVADSLNLR	RLVLMTSSLF	NFHALVSLPQ	FDELGYLDPD	DKTRLEEQAS	180
GFPMLKVKDI	KSAYSNWQIL	KEILGKMIKQ	TKASSGVIWN	SFKELEESEL	ETVIREIPAP	240
SFLIPLPKHL	TASSSSLLDH	DRTVFQWLDQ	QPPSSVLYVS	FGNTSEVDEK	DFLEIARGLV	300
DSKQSFLWV	RPGFVKGSTW	VEPLPDGFLG	ERGRIVKWVP	QQEVLAHGAI	GAFWTHSGWN	360
STLESVCEGV	PMIFSDFGLD	QPLNARYMSD	VLKVGVYLEN	GWERGEIANA	IRRVMDDEEG	420
EYIRQNARVL	KQKADVSLMK	GGSSYESLES	LVSYISSL			458

**WHAT IS CLAIMED IS:**

1. A recombinant host cell, comprising at least one recombinant gene that is:
  - (a) a gene encoding a UGT91D2e polypeptide having at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO:11;
  - (b) a gene encoding a chimeric polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:17 or SEQ ID NO:18;
  - (c) a gene encoding a UGT85C2 polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7; and/or
  - (d) a gene encoding a UGT76G1 polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;wherein the recombinant host cell is capable of producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or a glycosylated ent-kaurenoic acid compound in a cell culture broth.
2. The recombinant host cell of claim 1, wherein the UGT91D2e polypeptide comprises a UGT91D2e polypeptide having at least one amino acid substitution at residues 93, 99, 114, 144, 148, 152, 195, 196, 199, 211, 213, 221, 286, 384, 426, 438, or 466 of SEQ ID NO:11.
3. The recombinant host cell of claim 1, wherein the UGT85C2 polypeptide comprises a UGT85C2 polypeptide having at least one amino acid substitution at residues 21, 48, 49, 84, 86, 87, 91, 92, 95, 122, 334, or 334 of SEQ ID NO:7.
4. The recombinant host cell of claim 1, wherein the UGT76G1 polypeptide comprises a UGT76G1 polypeptide having at least one amino acid substitution at residues 23, 26, 55, 146, 257, 283, and 337 of SEQ ID NO:9.
5. The recombinant host cell of claim 1, wherein the UGT91D2e polypeptide comprises one or more of the UGT91D2e polypeptide variants comprising: P93V, S99I, S114F, T144K, T144L, T144M, A148K, M152T, L195G, L195C, L195S, L195N, L195V, V196P, K199C, L211H, L211M, L211I, L211C, L211T, L213E, S221I, V286C, V286N, V286S, G384W, G384K, G384Y, E426G, E438H, 3438M or A466V of SEQ ID NO:11.

6. The recombinant host cell of claim 1, wherein the UGT85C2 polypeptide comprises one or more of the UGT85C2 polypeptide variants comprising: Q21L, Q21T, Q21V, F48S, F48H, F48Y, F48R, F48Q, F48W, F48T, I49V, S84G, S84A, S84T, S84C, S84P, S84N, S84V, P86R, P86G, I87H, I87P, I87M, I87Y, L91K, L91R, L91T, L92F, L92I, L92M, I95K, F122S, L334S or L334M of SEQ ID NO:7.
7. The recombinant host cell of claim 1, wherein the UGT76G1 polypeptide comprises one or more of the UGT76G1 polypeptide variants comprising: Q23H, I26W, T146G, H155L, L257G, S253W, T284G, S283N, K337P or T55K of SEQ ID NO:9.
8. The recombinant host cell of any one of claims 1-7, further comprising at least one recombinant gene that is:
  - (a) a gene encoding a geranylgeranyl diphosphate synthase (GGPPS) polypeptide;
  - (b) a gene encoding an ent-copalyl diphosphate synthase (CDPS) polypeptide;
  - (c) a gene encoding an ent-kaurene synthase (KS) polypeptide;
  - (d) a gene encoding an ent-kaurene oxidase (KO) polypeptide;
  - (e) a gene encoding a cytochrome P450 reductase (CPR) polypeptide; and
  - (f) a gene encoding an ent-kaurenoic acid hydroxylase (KAH) polypeptide;
  - (g) a gene encoding a UGT74G1 polypeptide; and/or
  - (h) a gene encoding an EUGT11 polypeptide;wherein the recombinant host cell capable of producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or a glycosylated ent-kaurenoic acid compound in a cell culture broth.
9. The recombinant host cell of claim 8, wherein:
  - (a) the GGPPS polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:20, SEQ ID NO:22, SEQ ID NO:24, SEQ ID NO:26, SEQ ID NO:28, SEQ ID NO:30, SEQ ID NO:32, or SEQ ID NO:116;

- (b) the CDPS polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:34, SEQ ID NO:36, SEQ ID NO:38, SEQ ID NO:40, or SEQ ID NO:42;
  - (c) the KS polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:44, SEQ ID NO:46, SEQ ID NO:48, SEQ ID NO:50, or SEQ ID NO:52;
  - (d) the KO polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:60, SEQ ID NO:62, SEQ ID NO:117, SEQ ID NO:66, SEQ ID NO:68, SEQ ID NO:70, SEQ ID NO:72, SEQ ID NO:74, or SEQ ID NO:76;
  - (e) the CPR polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:78, SEQ ID NO:80, SEQ ID NO:82, SEQ ID NO:84, SEQ ID NO:86, SEQ ID NO:88, SEQ ID NO:90, SEQ ID NO:92;
  - (f) the KAH polypeptide comprises a polypeptide having at least 70% identity to an amino acid sequence set forth in SEQ ID NO:94, SEQ ID NO:97, SEQ ID NO:100, SEQ ID NO:101, SEQ ID NO:102, SEQ ID NO:103, SEQ ID NO:104, SEQ ID NO:106, SEQ ID NO:108, SEQ ID NO:110, SEQ ID NO:112, or SEQ ID NO:114;
  - (g) the UGT74G1 polypeptide comprises a polypeptide having at least 55% identity to an amino acid sequence set forth in SEQ ID NO:4;
  - (h) the EUGT11 polypeptide comprises a polypeptide having at least 65% identity to an amino acid sequence set forth in SEQ ID NO:16.
10. The recombinant host cell of any one of claims 1-9, wherein the cell culture broth comprises:
- (a) the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound produced by the recombinant host cell,
  - (b) glucose, fructose and/or sucrose; and/or
  - (c) supplemental nutrients comprising trace metals, vitamins, salts, yeast nitrogen base (YNB), and/or amino acids.

11. The recombinant host cell of any one of claims 1-10, wherein the recombinant host comprises a plant cell, a mammalian cell, an insect cell, a fungal cell, an algal cell, or a bacterial cell.
12. The recombinant host cell of claim 11, wherein the bacterial cell comprises *Escherichia* cells, *Lactobacillus* cells, *Lactococcus* cells, *Cornibacterium* cells, *Acetobacter* cells, *Acinetobacter* cells, or *Pseudomonas* cells.
13. The recombinant host cell of claim 11, wherein the fungal cell comprises a yeast cell.
14. The recombinant host cell of claim 13, wherein the yeast cell is a cell from *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Yarrowia lipolytica*, *Candida glabrata*, *Ashbya gossypii*, *Cyberlindnera jadinii*, *Pichia pastoris*, *Kluyveromyces lactis*, *Hansenula polymorpha*, *Candida boidinii*, *Arxula adenivorans*, *Xanthophyllomyces dendrorhous*, or *Candida albicans* species.
15. The recombinant host cell of claim 14, wherein the yeast cell is a *Saccharomycete*.
16. The recombinant host cell of claim 15, wherein the yeast cell is a cell from the *Saccharomyces cerevisiae* species.
17. A method of producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or glycosylated ent-kaurenoic acid compound in a cell culture broth, comprising growing the recombinant host cell of any one of claims 1-16 in a culture medium, under conditions in which one or more of the genes are expressed;
  - wherein at least one of the genes is a recombinant gene;
  - wherein the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound is produced by the recombinant host cell.
18. The method of claim 17, wherein one or more of the genes is constitutively expressed and/or expression of one or more of the genes is induced.

19. A method for producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound comprising whole-cell bioconversion of plant-derived components or synthetic steviol or steviol glycosides using one or more of:
- (a) a UGT91D2e polypeptide having at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO:11;
  - (b) a chimeric polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:17 or SEQ ID NO:18;
  - (c) a UGT85C2 polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7; and/or
  - (d) a UGT76G1 polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9;
- wherein at least one of the polypeptides is a recombinant polypeptide.
20. The method of claim 19, wherein the whole cell is the recombinant host cell of any one of claims 1-16.
21. The method of any one of claims 17-20, wherein the recombinant host cell is grown in a fermentor at a temperature for a period of time, wherein the temperature and period of time facilitate the production of the steviol glycoside, glycosylated ent-kaurenol compound, and/or glycosylated ent-kaurenoic acid compound.
22. An *in vitro* method for producing a steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound, comprising adding one or more of:
- (a) a UGT91D2e polypeptide having at least 90% sequence identity to the amino acid sequence set forth in SEQ ID NO:11;
  - (b) a chimeric polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO:17 or SEQ ID NO:18;
  - (c) a UGT85C2 polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID NO:7; and/or
  - (d) a UGT76G1 polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID NO:9,
- and plant-derived components or synthetic steviol or steviol glycosides to a reaction mixture;

- wherein at least one of the polypeptides is a recombinant polypeptide; and synthesizing the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound in the reaction mixture.
23. The method of any one of claims 17-22, that further comprises isolating the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound, alone or in combination from the cell culture broth.
24. The method of claim 23, wherein the isolating step comprises:
- (a) providing the cell culture broth comprising the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or in combination;
  - (b) separating a liquid phase of the cell culture broth from a solid phase of the cell culture broth to obtain a supernatant comprising the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or in combination;
  - (c) providing one or more adsorbent resins, comprising providing the adsorbent resins in a packed column; and
  - (d) contacting the supernatant of step (b) with the one or more adsorbent resins in order to obtain at least a portion of the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or in combination thereby isolating the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or in combination.
25. The method of any one of claims 17-22, that further comprises recovering the the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound alone or a composition comprising the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound.
26. The method of claim 25, wherein the recovered composition is enriched for the steviol glycoside, glycosylated ent-kaurenol compound, and/or the glycosylated ent-kaurenoic acid compound relative to a steviol glycoside composition of Stevia plant and has a

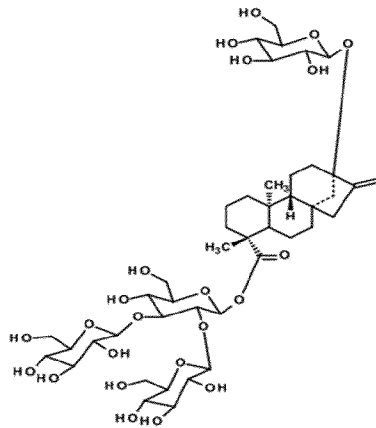
- reduced level of non-steviol glycoside Stevia plant-derived components relative to a plant-derived stevia extract.
27. The method of any one of claims 17-22, wherein the cell culture broth comprises:
- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or glycosylated ent-kaurenoic acid compounds produced by the recombinant host cell of any one of claims 1-12,
  - (b) glucose, fructose, and/or sucrose; and/or
  - (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids.
28. The method of claim 22, wherein the reaction mixture comprising:
- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or a glycosylated ent-kaurenoic acid compounds produced in the reaction mixture;
  - (b) a UGT polypeptide;
  - (c) UDP-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetylglucosamine; and/or
  - (d) reaction buffer and/or salts.
29. The method of any one of claims 17-28, wherein the recombinant host cell comprises a plant cell, a mammalian cell, an insect cell, a fungal cell, an algal cell, or a bacterial cell.
30. The method of claim 29, wherein the bacterial cell comprises *Escherichia* cells, *Lactobacillus* cells, *Lactococcus* cells, *Cornibacterium* cells, *Acetobacter* cells, *Acinetobacter* cells, or *Pseudomonas* cells.
31. The method of claim 29, wherein the fungal cell comprises a yeast cell.
32. The method of claim 31, wherein the yeast cell is a cell from *Saccharomyces cerevisiae*, *Schizosaccharomyces pombe*, *Yarrowia lipolytica*, *Candida glabrata*, *Ashbya gossypii*, *Cyberlindnera jadinii*, *Pichia pastoris*, *Kluyveromyces lactis*, *Hansenula polymorpha*, *Candida boidinii*, *Arxula adenivorans*, *Xanthophyllomyces dendrorhous*, or *Candida albicans* species.

33. The method of claim 31, wherein the yeast cell is a *Saccharomycete*.
34. The method of claim 33, wherein the yeast cell is a cell from the *Saccharomyces cerevisiae* species.
35. The recombinant host cell of any one of claims 1-16 or the method of any one of claims 17-34, wherein:
- (a) the steviol glycoside comprises 13-SMG, 19-SMG, Steviol-1,2-bioside, Steviol-1,3-bioside, 1,2-stevioside, 1,3-stevioside, rubusoside, RebA, RebB, RebD, RebE, RebM, di-glycosylated tri-glycosylated steviol, tetra-glycosylated steviol, penta-glycosylated steviol, hexa-glycosylated steviol, hepta-glycosylated steviol, and/or isomers thereof;
  - (b) the glycosylated ent-kaurenol compound comprises di-glycosylated ent-kaurenol, tri-glycosylated ent-kaurenol, and/or isomers thereof; and/or
  - (c) the glycosylated ent-kaurenoic acid compound comprises di-glycosylated ent-kaurenoic acid, tri-glycosylated ent-kaurenoic acid, and/or isomers thereof.
36. The recombinant host cell or the method of claim 35, wherein:
- (a) the di-glycosylated steviol comprises compound 2.23 of Table 1;
  - (b) the tri-glycosylated steviol comprises compound 3.1 and/or compound 3.34 of Table 1;
  - (c) the tetra-glycosylated steviol comprises compound 4.26 and/or compound 4.33 of Table 1;
  - (d) the penta-glycosylated steviol comprises compound 5.22, compound 5.24, and/or compound 5.25 of Table 1;
  - (e) the hexa-glycosylated steviol comprises compound 6.1 and/or compound 6.23 of Table 1;
  - (f) the hepta-glycosylated steviol comprises compound 7.2, compound 7.5, and/or compound 7.13 of Table 1;
  - (g) the glycosylated ent-kaurenoic acid compound comprises compound KA3.1, compound KA3.2, and/or compound KA2.7 of Table 1; and/or

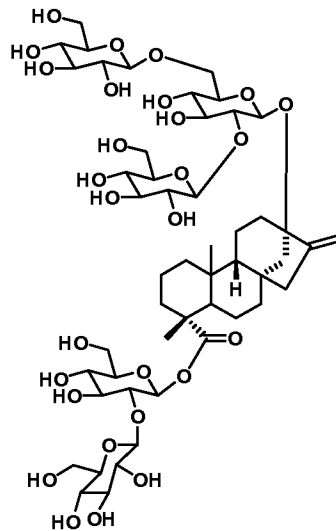
- (h) the glycosylated ent-kaurenol compound comprises compound KL2.8 and/or compound KL3.1 co-eluted with compound KL3.6 of Table 1.

37. The recombinant host cell or method of claim 36, wherein:

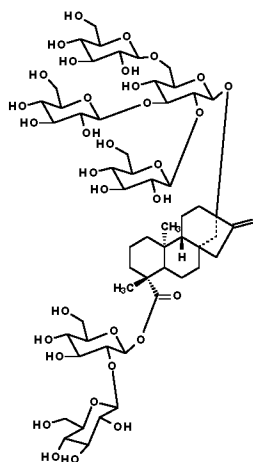
- (a) compound 4.26 has the structure:



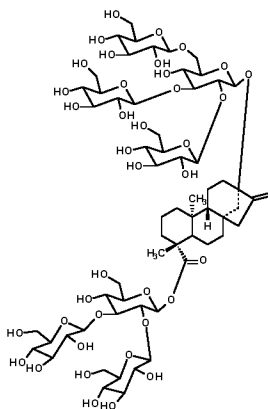
- (b) compound 5.22 has the structure:



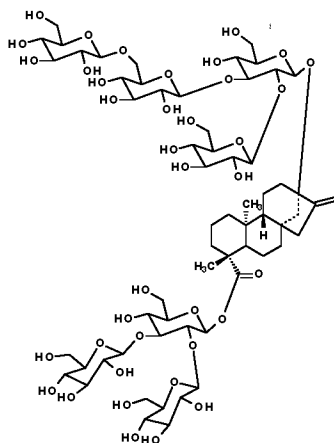
- (c) compound 6.1 has the structure:



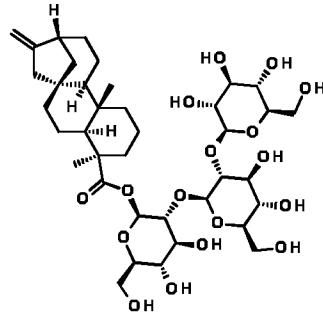
(d) compound 7.2 has the structure:



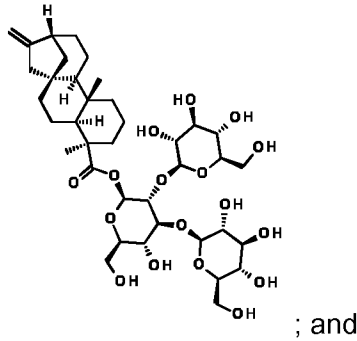
(e) compound 7.5 has the structure:



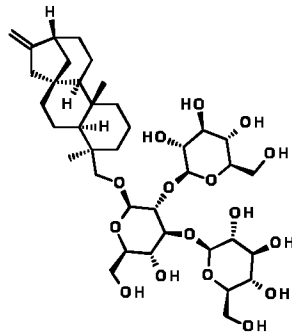
(f) compound KA3.1 has the structure:



(g) compound KA3.2 has the structure:



(h) compound KL3.1 has the structure:



38. A steviol glycoside composition produced by the recombinant host cell of any one of claims 1-16 or the method of any one of claims 17-34, wherein the composition has a steviol glycoside composition enriched for RebD, RebM, or isomers thereof relative to a steviol glycoside composition of Stevia plant and has a reduced level of non-steviol glycoside Stevia plant-derived components relative to a plant-derived stevia extract.

39. A cell culture broth comprising:

(a) the recombinant host cell of any one of claims 1-16; and

- (b) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or glycosylated ent-kaurenoic acid compounds produced by the recombinant host cell;

wherein one or more steviol glycosides is present at a concentration of at least 1 mg/liter of the culture broth.

40. A cell culture broth comprising:

- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or glycosylated ent-kaurenoic acid compounds produced by the recombinant host cell of any one of claims 1-16,
- (b) glucose, fructose, sucrose, xylose, ethanol, and/or glycerol; and/or
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids.

41. A cell lysate comprising:

- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or glycosylated ent-kaurenoic acid compounds produced by the recombinant host cell of any one of claims 1-16,
- (b) glucose, fructose, sucrose, xylose, ethanol, glycerol, uridine diphosphate (UDP)-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and/or
- (c) supplemental nutrients comprising trace metals, vitamins, salts, YNB, and/or amino acids.

42. A reaction mixture comprising:

- (a) one or more steviol glycosides, glycosylated ent-kaurenol compounds, and/or a glycosylated ent-kaurenoic acid compounds produced in the reaction mixture;
- (b) a UGT polypeptide;
- (c) glucose, fructose, sucrose, xylose, ethanol, glycerol, uridine diphosphate (UDP)-glucose, UDP-rhamnose, UDP-xylose, and/or N-acetyl-glucosamine; and/or
- (d) reaction buffer and/or salts.

Figure 1

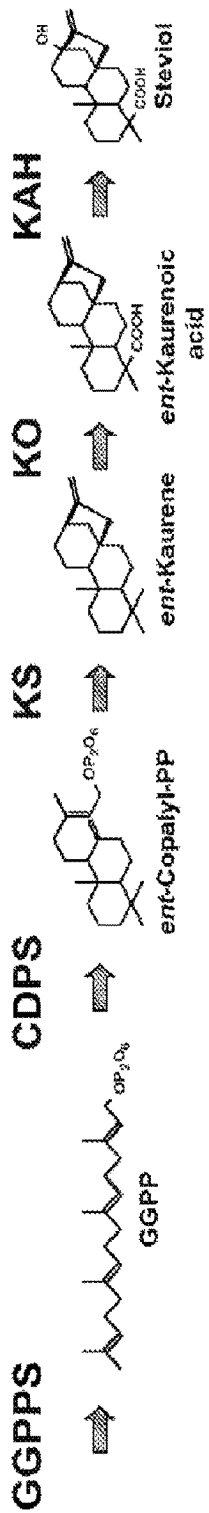


Figure 2

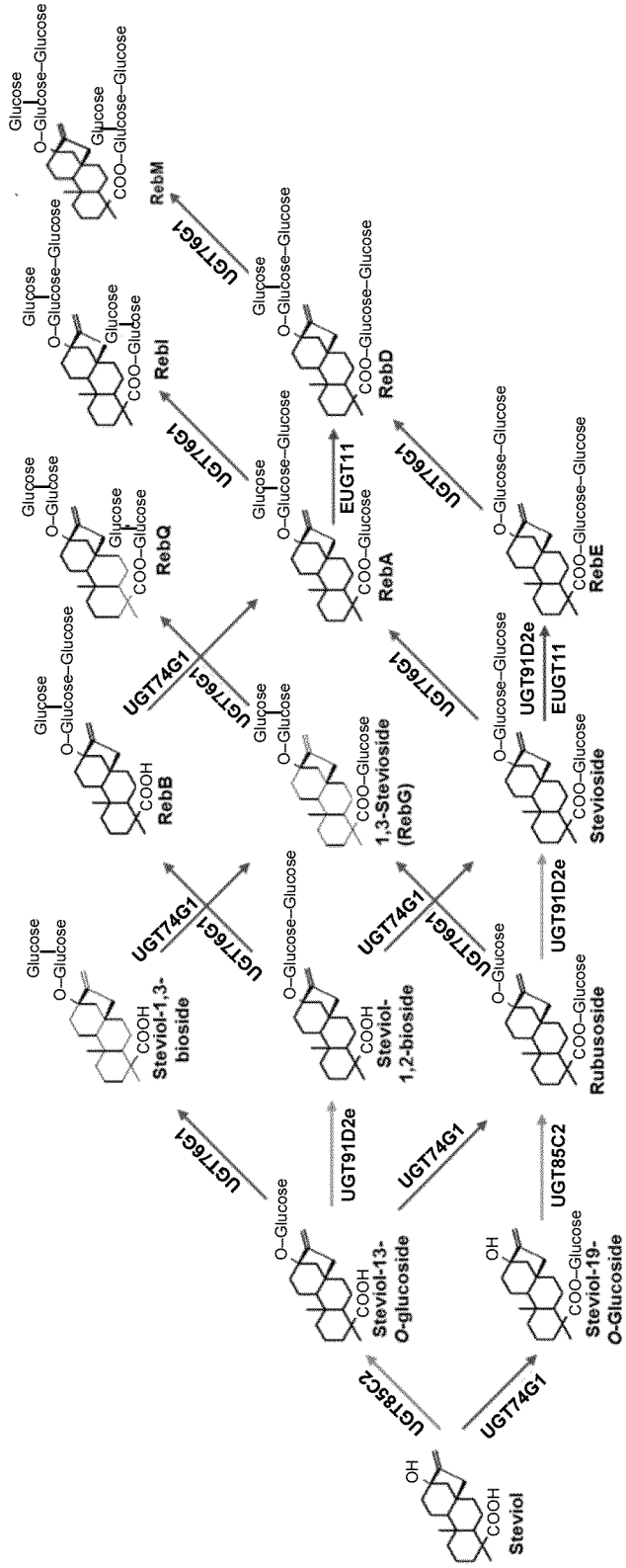


Figure 3

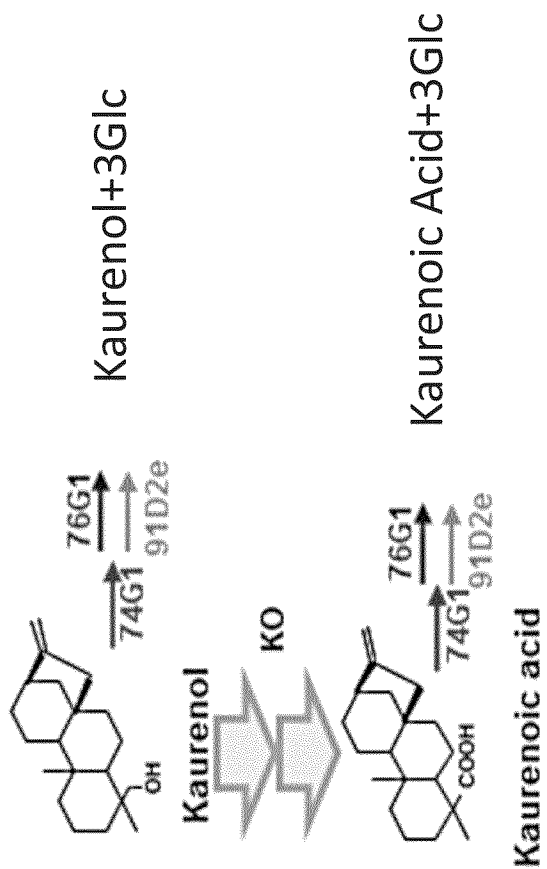


Figure 4A

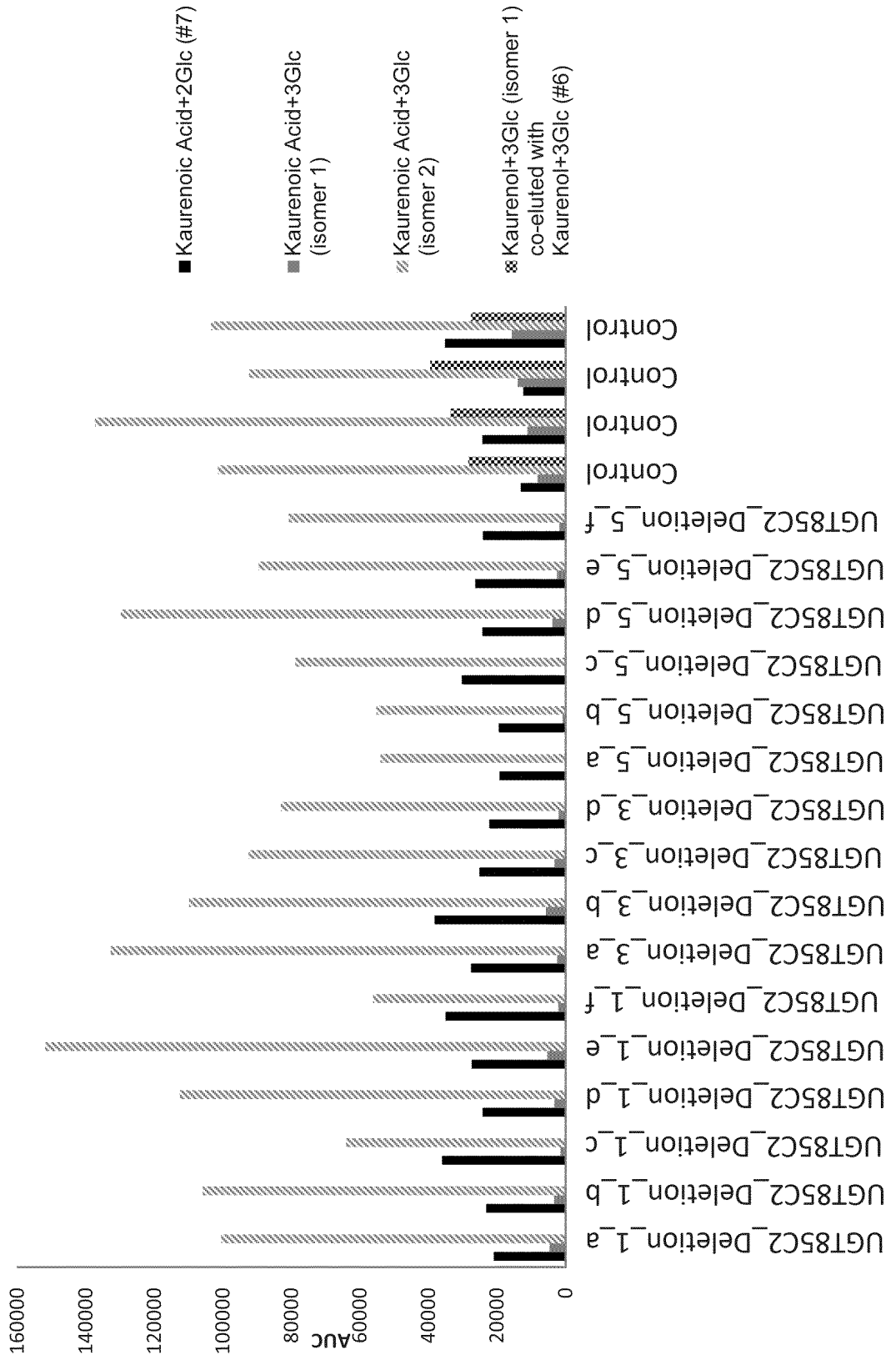


Figure 4B

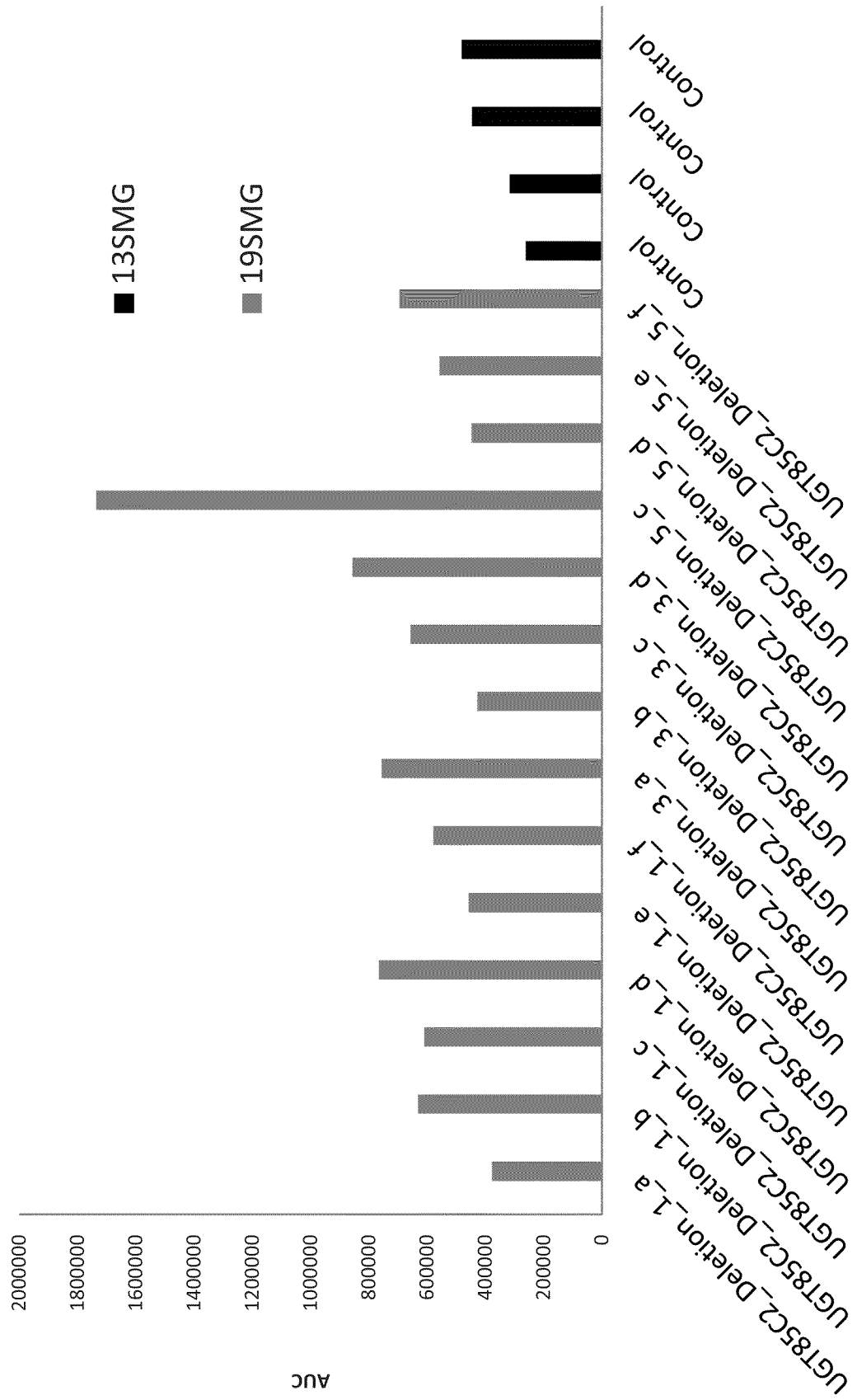


Figure 4C

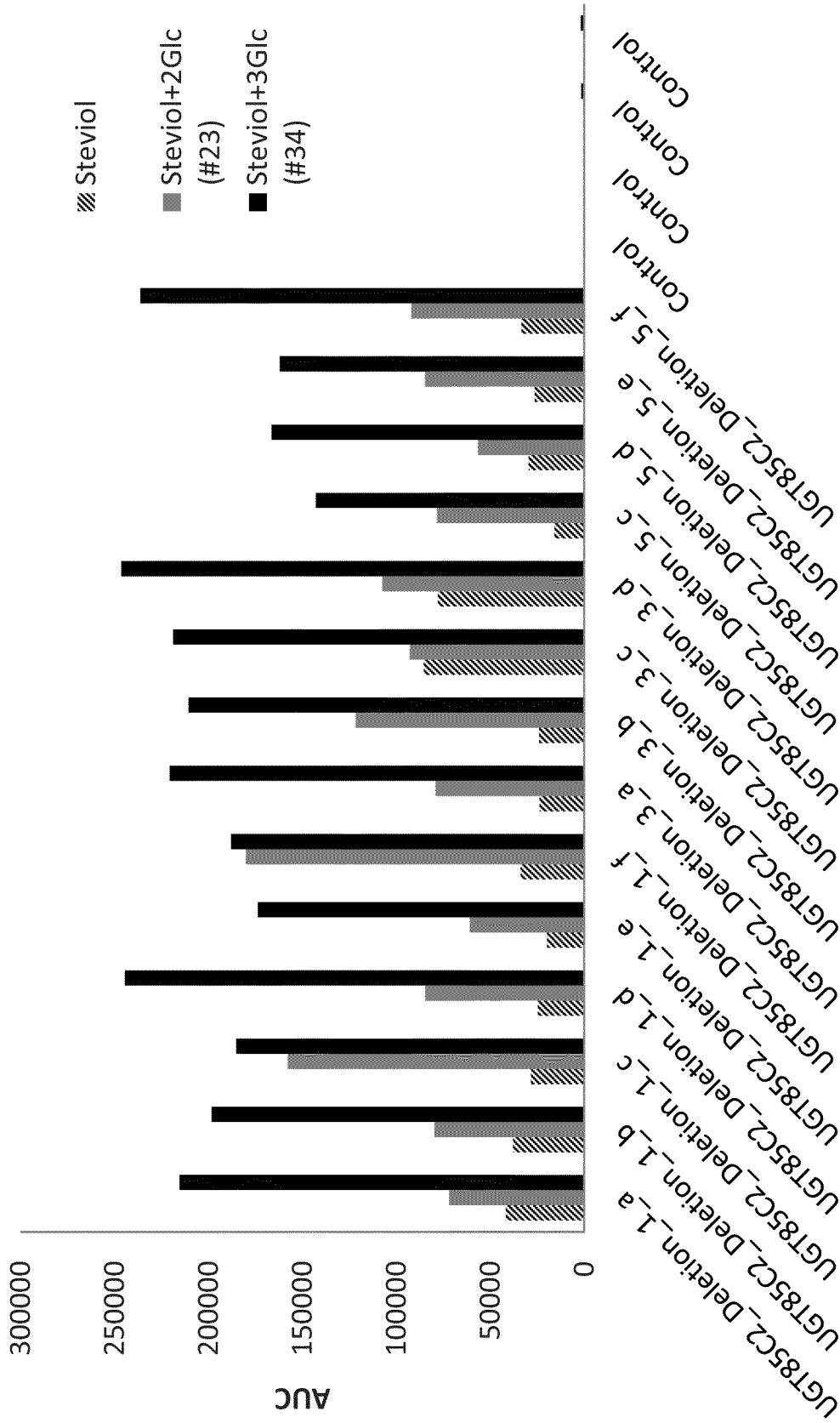


Figure 5

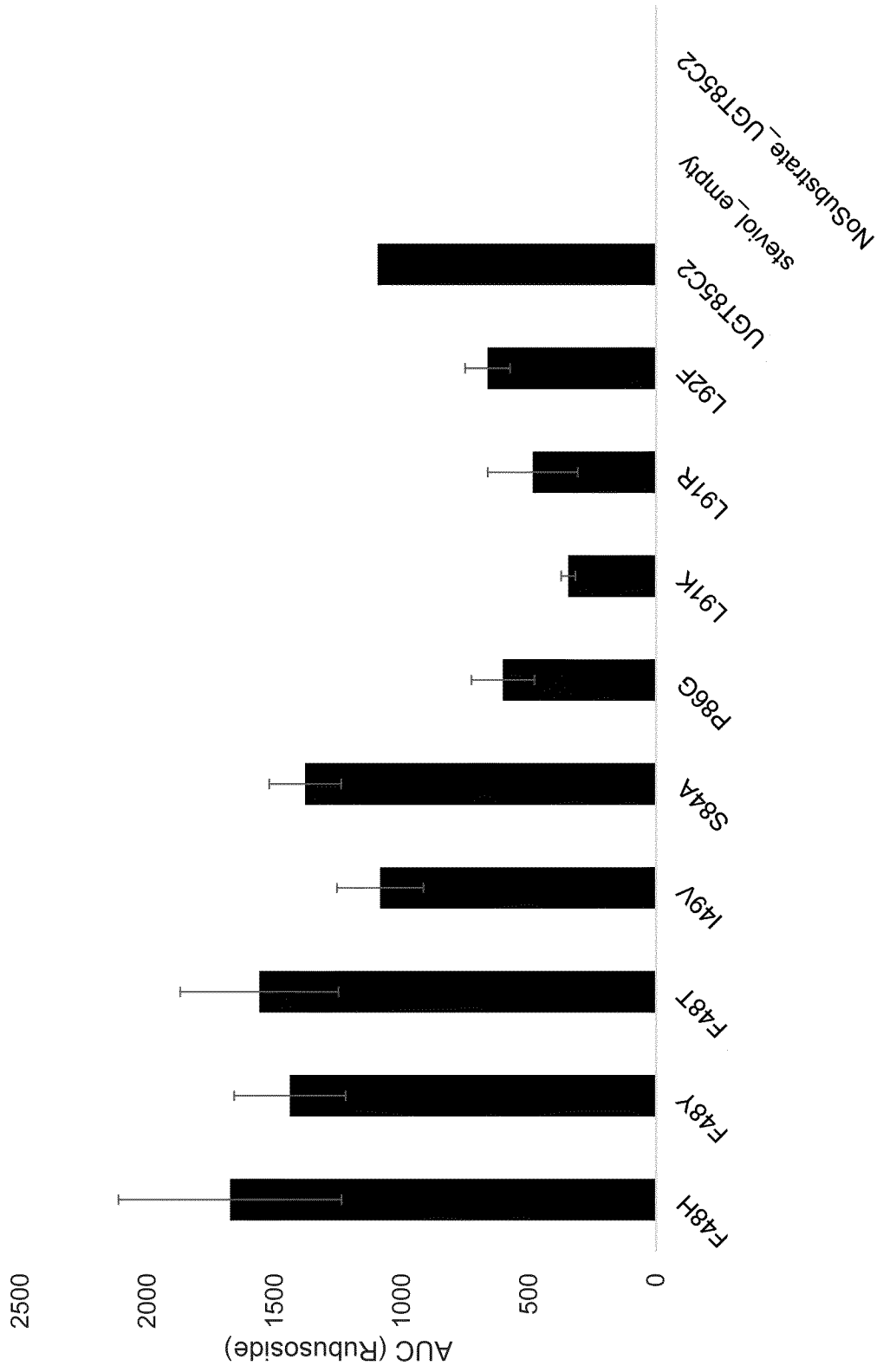


Figure 6A

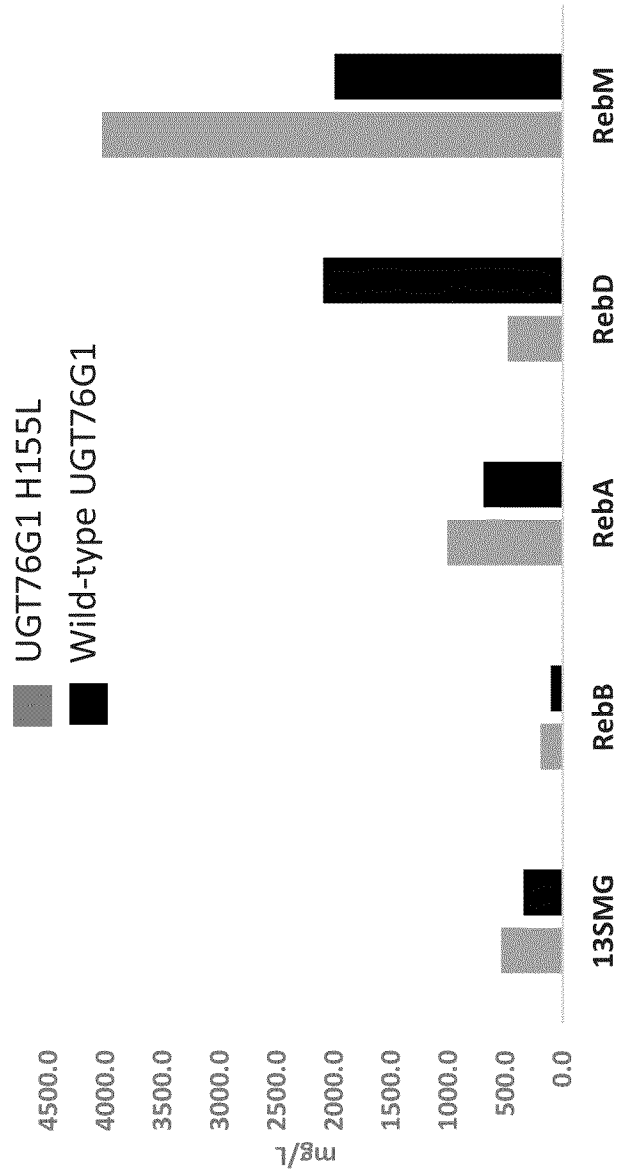


Figure 6B

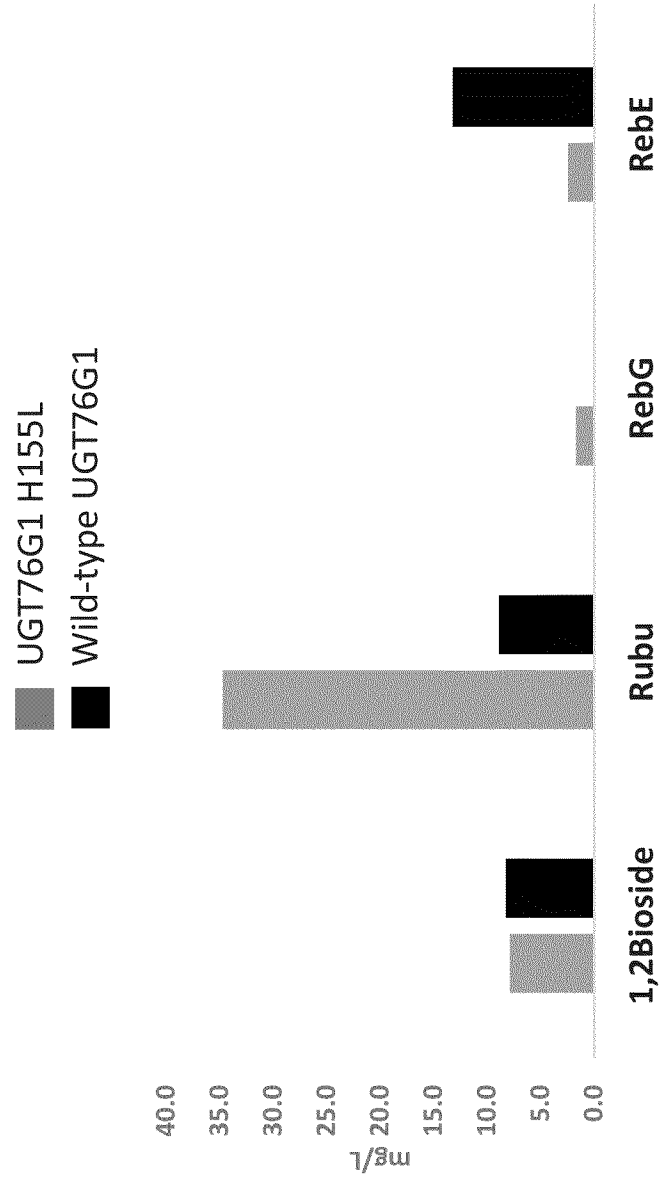


Figure 6C

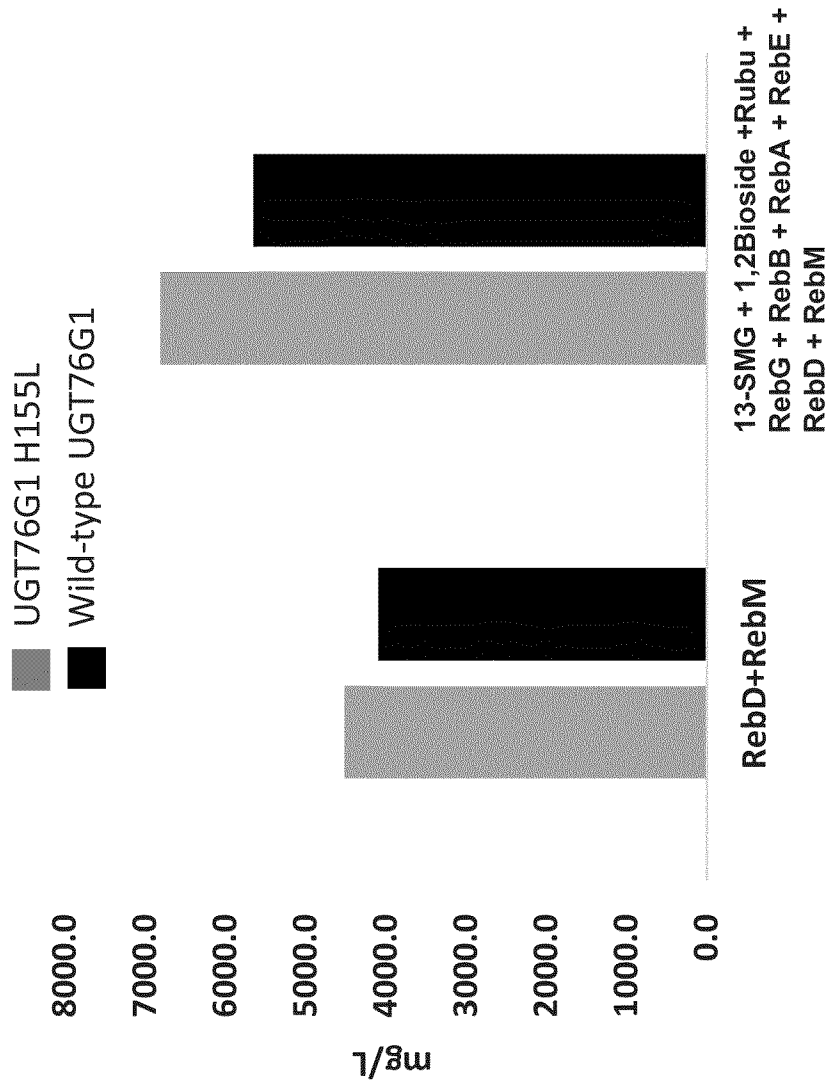


Figure 6D

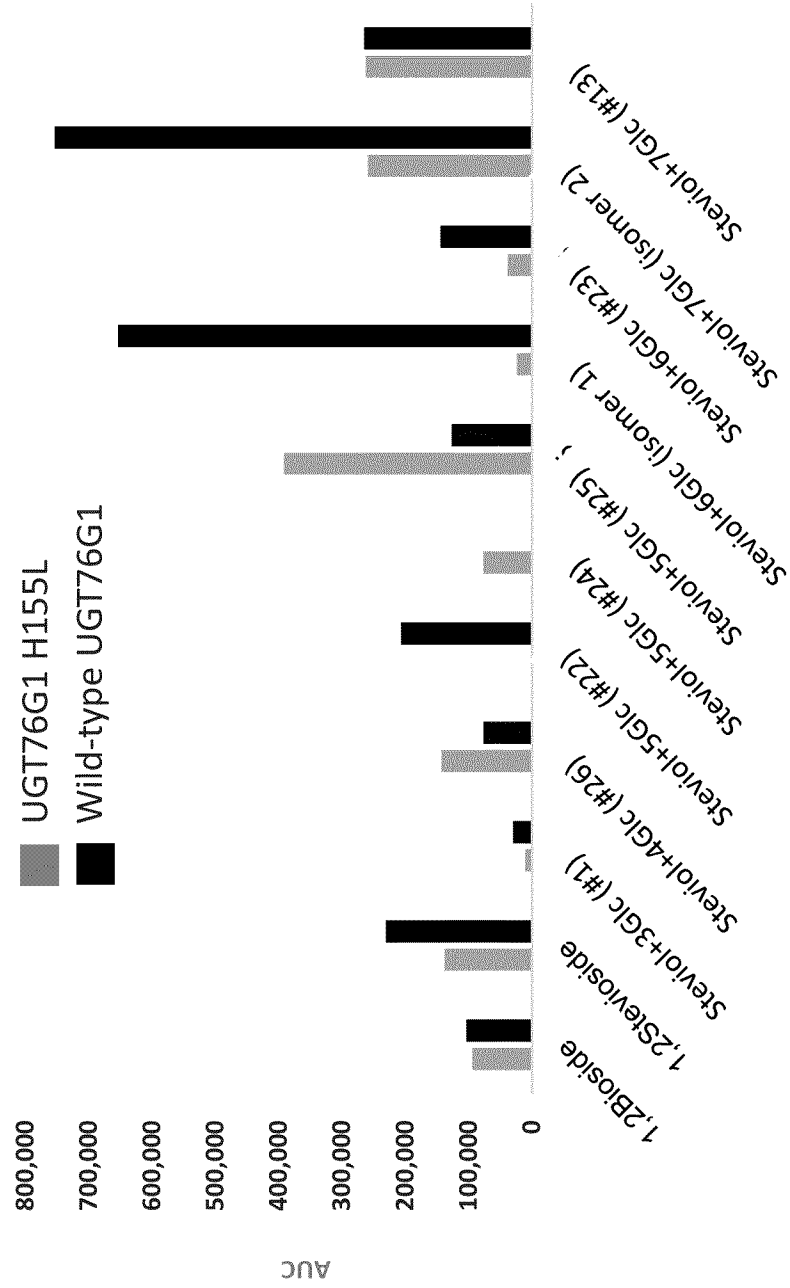
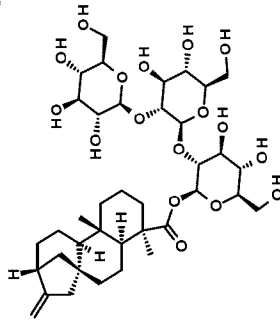


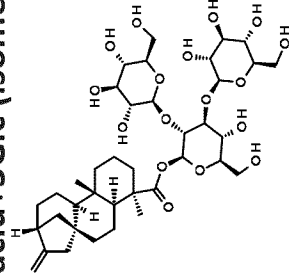
Figure 7A

**Kaurenoic acid+3Glc (isomer 1)**



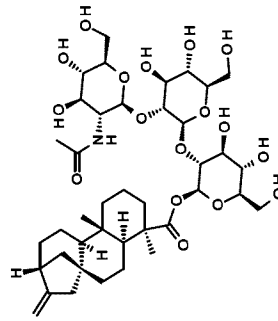
Formula Weight : 788.87(3)  
 Exact Mass : 788.38305050(3)  
 Formula : C<sub>38</sub>H<sub>60</sub>O<sub>17</sub>

**Kaurenoic acid+3Glc (isomer 2)**



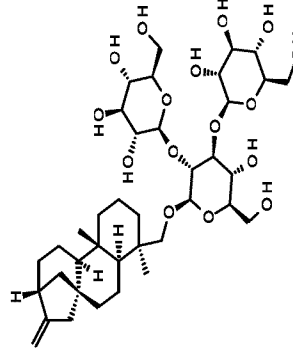
Formula Weight : 788.87(3)  
 Exact Mass : 788.38305050(3)  
 Formula : C<sub>38</sub>H<sub>60</sub>O<sub>17</sub>

**Kaurenoic acid+2Glc+1GlcNAc**



Formula Weight : 829.92(3)  
 Exact Mass : 829.40959960(4)  
 Formula : C<sub>40</sub>H<sub>63</sub>NO<sub>17</sub>

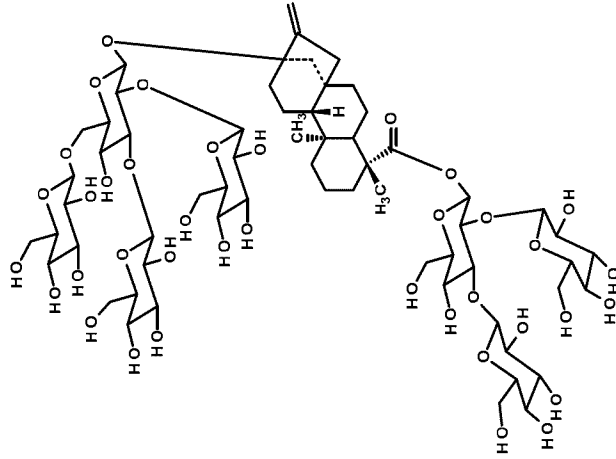
**Kaurenol+3Glc (isomer 1)**



Formula Weight : 774.89(3)  
 Exact Mass : 774.40378594(3)  
 Formula : C<sub>38</sub>H<sub>62</sub>O<sub>16</sub>

Figure 7B

Steviol+7Glc (isomer 2)



Steviol+6Glc (isomer 1)

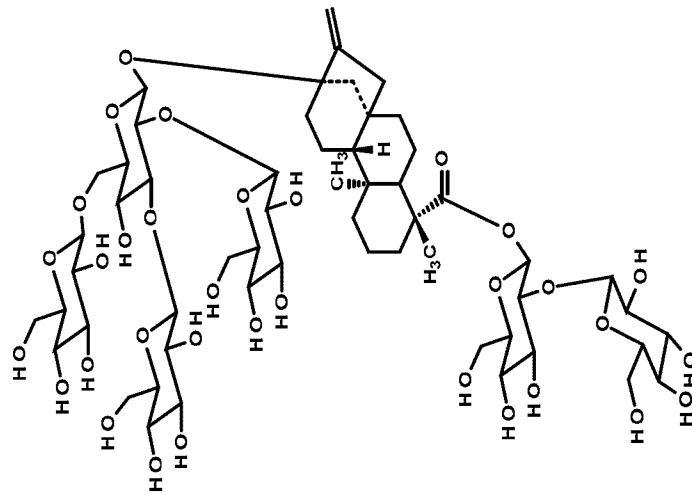
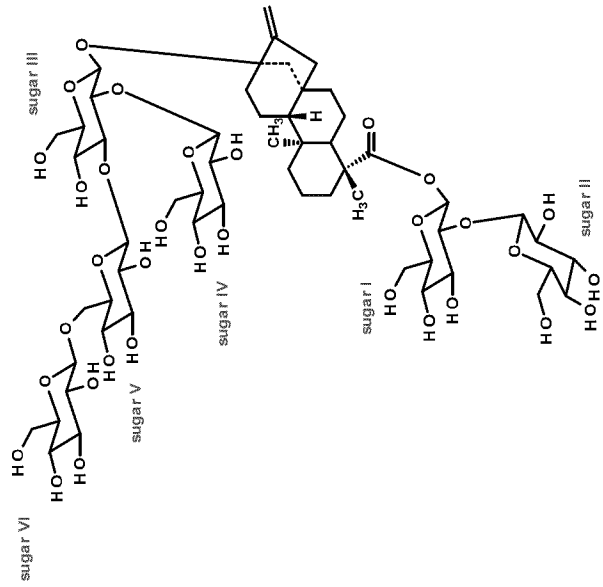


Figure 7C

**Steviol+6Glc (isomer 4)**



**Steviol+7Glc (isomer 5)**

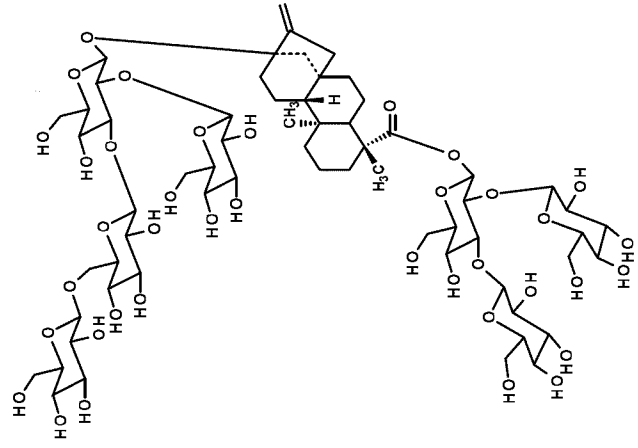
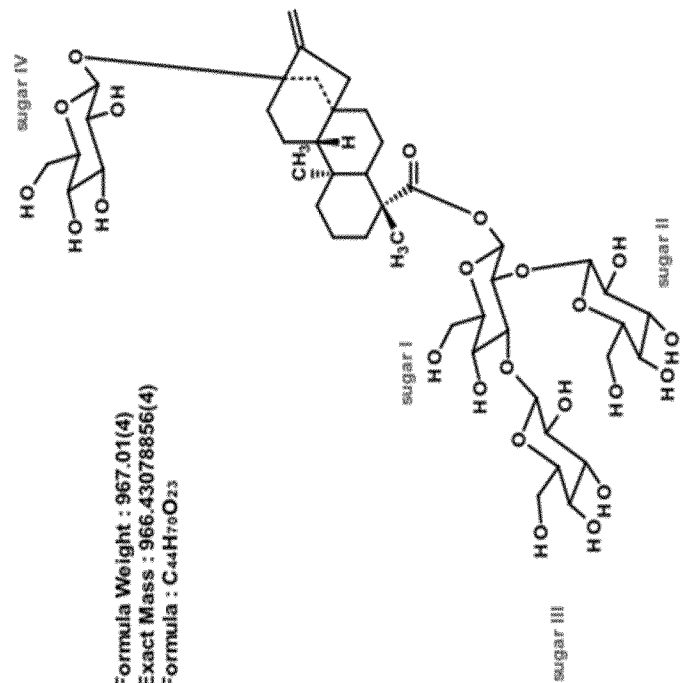


Figure 7D

**Steviol+4Glc (#26)**



**Steviol+4Glc+1GlcNAc (#11)**

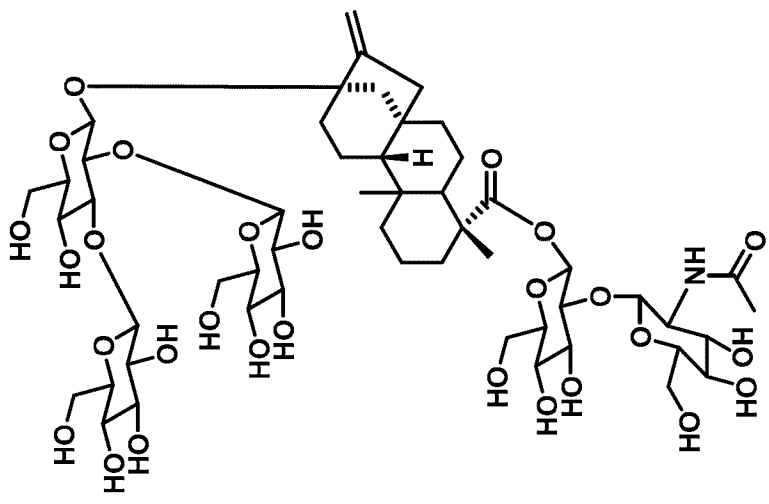


Figure 7E

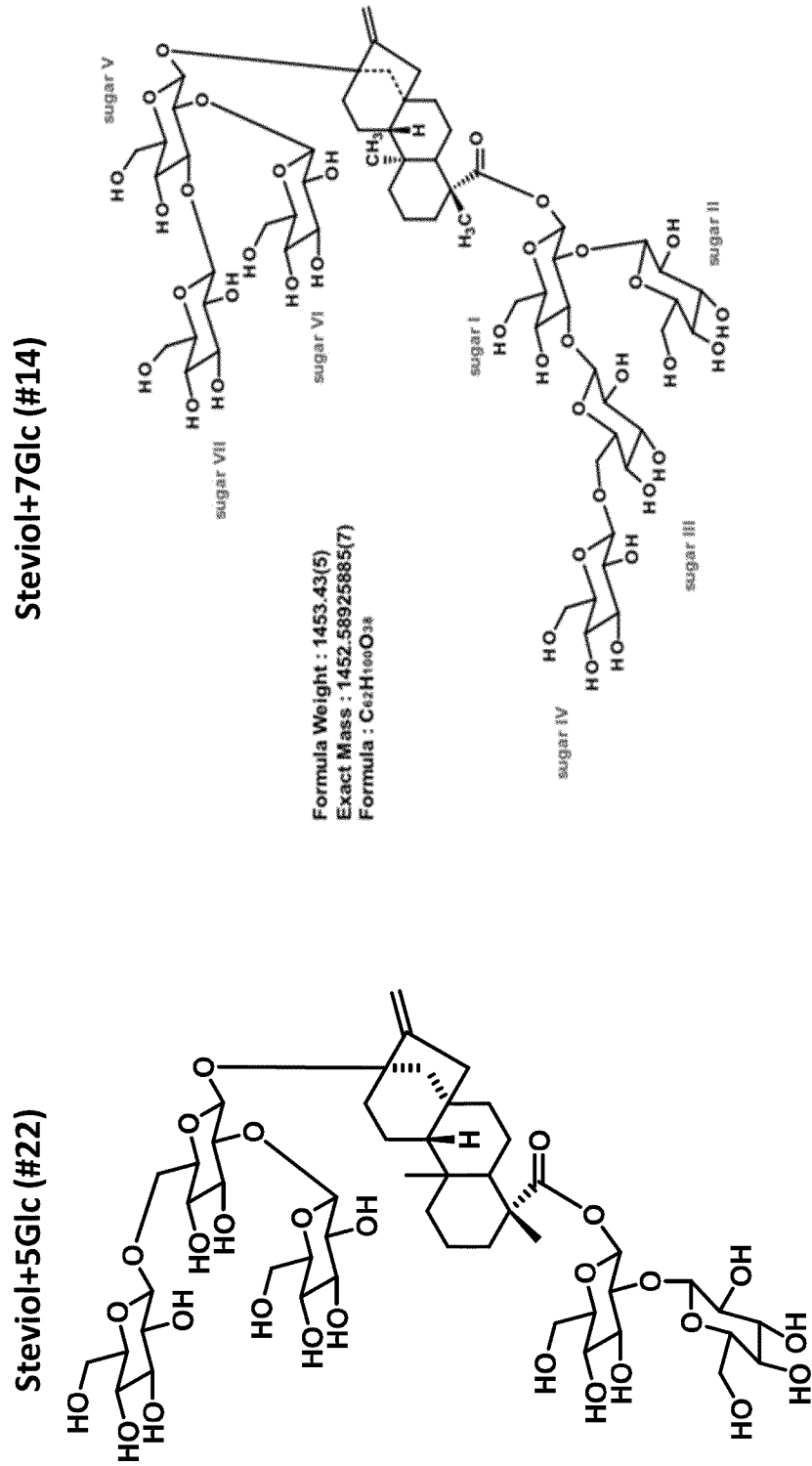


Figure 8A

Kaurenoic Acid+3Glc (isomer 1)

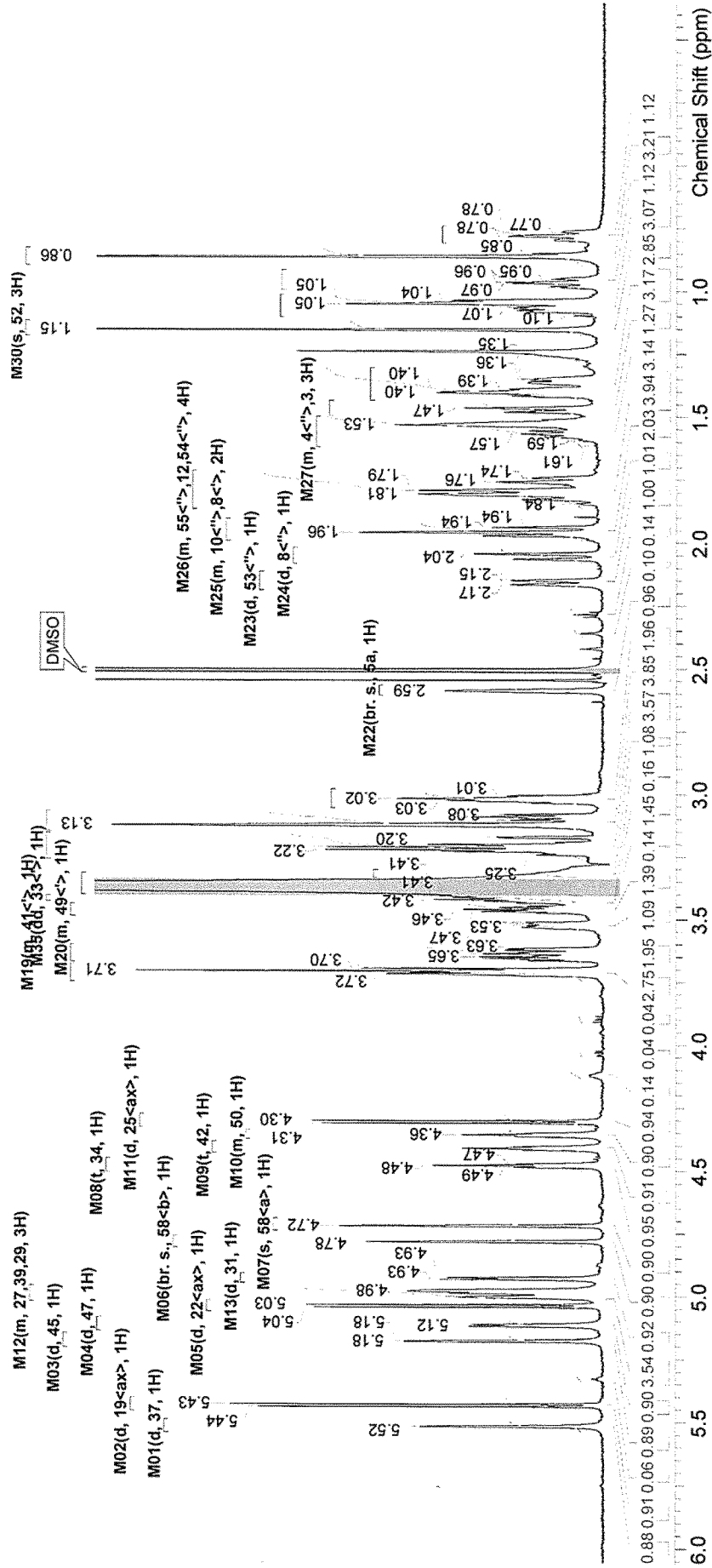


Figure 8B

Atom1	Shift1 (ppm)	H's	Type	J (Hz)	Multiplet1	(ppm)
55<>	0.78	1	m	-	M34	[0.74 .. 0.81]
57	0.86	3	m	-	M33	[0.83 .. 0.90]
53<>	0.97	1	td	13.33, 4.16	M32	[0.92 .. 1.00]
13a, 2a, 10<>	1.07	3	m	-	M31	[1.02 .. 1.10]
52	1.15	3	s	-	M30	[1.12 .. 1.18]
11<>, 4<>, 54<>	1.38	3	m	-	M29	[1.30 .. 1.44]
11<>	1.47	1	m	-	M28	[1.44 .. 1.50]
4<>, 3	1.56	3	m	-	M27	[1.50 .. 1.62]
55<>, 12, 54<>	1.79	4	m	-	M26	[1.72 .. 1.87]
10<>, 8<>	1.95	2	m	-	M25	[1.91 .. 1.99]
8<>	2.06	1	d	17.12	M24	[2.02 .. 2.08]
53<>	2.16	1	d	12.72	M23	[2.12 .. 2.19]
5a	2.59	1	br. s.	-	M22	[2.57 .. 2.61]
38<ax>, 26<ax>	3.02	2	m	-	M14	[2.98 .. 3.06]
30<ax>, 40<ax>, 28<ax>, 32<ax>	3.10	4	m	-	M15	[3.07 .. 3.15]
46<ax>, 23<ax>, 48<ax>	3.22	3	m	-	M16	[3.16 .. 3.26]
36<ax>	3.32	1	m	-	M21	[3.30 .. 3.33]
33<>	3.42	1	dd	11.00, 5.14	M35	[3.41 .. 3.43]
49<>	3.46	1	m	-	M20	[3.44 .. 3.49]
41<>	3.52	1	m	-	M19	[3.50 .. 3.55]
44<ax>, 49<>	3.64	2	m	-	M18	[3.60 .. 3.67]
41<>, 33<>, 20<ax>	3.71	3	m	-	M17	[3.68 .. 3.75]
25<ax>	4.31	1	d	7.82	M11	[4.28 .. 4.33]
50	4.35	1	m	-	M10	[4.34 .. 4.37]
42	4.41	1	t	5.38	M09	[4.39 .. 4.43]
34	4.48	1	t	5.62	M08	[4.45 .. 4.51]
58<a>	4.72	1	s	-	M07	[4.69 .. 4.74]
58<b>	4.78	1	br. s.	-	M06	[4.76 .. 4.81]
31	4.93	1	d	5.38	M13	[4.91 .. 4.95]
27, 39, 29	4.99	3	m	-	M12	[4.96 .. 5.02]
22<ax>	5.04	1	d	7.83	M05	[5.02 .. 5.06]
47	5.12	1	d	5.38	M04	[5.09 .. 5.14]
45	5.18	1	d	2.45	M03	[5.15 .. 5.21]
19<ax>	5.43	1	d	7.83	M02	[5.40 .. 5.46]
37	5.52	1	d	2.45	M01	[5.49 .. 5.54]

### Kaurenoic Acid+3Glc (isomer 1)

<sup>1</sup>H NMR (800 MHz, DMSO *d*<sub>6</sub>) δ ppm 0.74 - 0.81 (m, 1 H) 0.83 - 0.90 (m, 3 H) 0.97 (td, *J*=13.33, 4.16 Hz, 1 H) 1.02 - 1.10 (m, 3 H) 1.15 (s, 3 H) 1.30 - 1.44 (m, 3 H) 1.44 - 1.50 (m, 1 H) 1.50 - 1.62 (m, 3 H) 1.72 - 1.87 (m, 4 H) 1.91 - 1.99 (m, 2 H) 2.06 (d, *J*=17.12 Hz, 1 H) 2.16 (d, *J*=12.72 Hz, 1 H) 2.59 (br. s., 1 H) 2.98 - 3.06 (m, 2 H) 3.07 - 3.15 (m, 4 H) 3.16 - 3.26 (m, 3 H) 3.30 - 3.33 (m, 1 H) 3.42 (dd, *J*=11.00, 5.14 Hz, 1 H) 3.44 - 3.49 (m, 1 H) 3.50 - 3.55 (m, 1 H) 3.60 - 3.67 (m, 2 H) 3.68 - 3.75 (m, 3 H) 4.31 (d, *J*=7.82 Hz, 1 H) 4.34 - 4.37 (m, 1 H) 4.41 (t, *J*=5.38 Hz, 1 H) 4.48 (t, *J*=5.62 Hz, 1 H) 4.72 (s, 1 H) 4.78 (br. s., 1 H) 4.93 (d, *J*=5.38 Hz, 1 H) 4.96 - 5.02 (m, 3 H) 5.04 (d, *J*=7.83 Hz, 1 H) 5.12 (d, *J*=5.38 Hz, 1 H) 5.18 (d, *J*=2.45 Hz, 1 H) 5.43 (d, *J*=7.83 Hz, 1 H) 5.52 (d, *J*=2.45 Hz, 1 H)

<sup>13</sup>C NMR (201.21 MHz, DMSO *d*<sub>6</sub>) δ ppm 175.4 (1C), 155.3 (1C), 104.6 (1C), 103.6 (1C), 99.8 (1C), 92.4 (1C), 83.4 (1C), 77.6 (1C), 77.5 (1C), 77.2 (1C), 76.9 (1C), 76.2 (1C), 76.1 (1C), 75.8 (1C), 75.2 (1C), 70.8 (1C), 70.0 (1C), 69.8 (1C), 61.5 (1C), 61.4 (1C), 60.6 (1C), 56.7 (1C), 54.6 (1C), 48.7 (1C), 43.9 (1C), 43.4 (1C), 43.3 (1C), 41.2 (1C), 40.4 (1C), 40.0 (1C), 39.4 (1C), 37.8 (1C), 32.9 (1C), 28.5 (1C), 21.6 (1C), 19.1 (1C), 18.5 (1C), 16.1 (1C)

Figure 8C  
Kaurenoic Acid+3Glc (isomer 1)

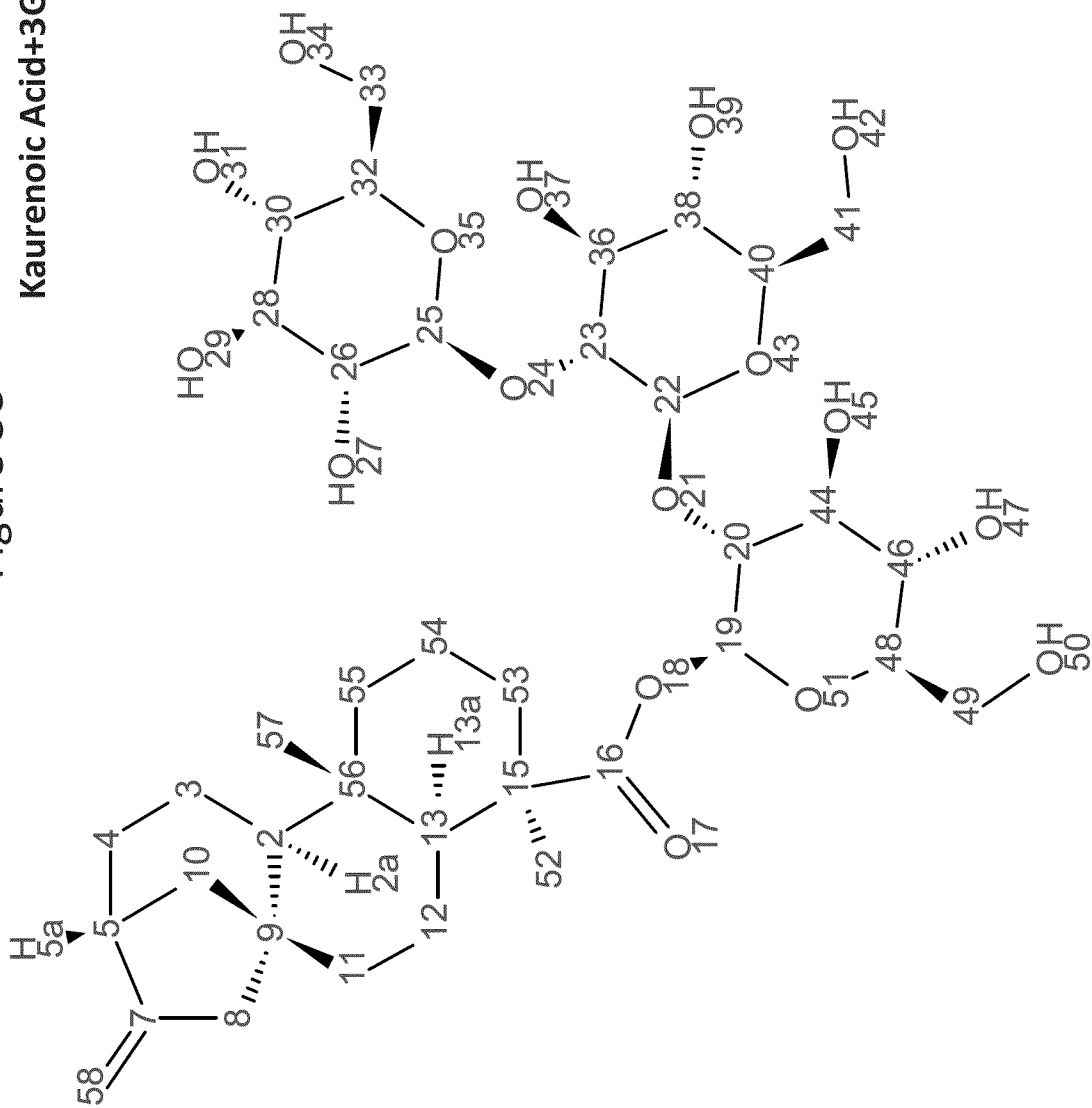




Figure 8E

Atom1	Shift1 (ppm)	H's	Type	J (Hz)	Multiplet1	(ppm)
46<->	0.77	1	m	-	M40	[0.73 .. 0.80]
61	0.83	3	s	-	M07	[0.81 .. 0.85]
44<->	0.92	1	td	13.33, 4.16	M39	[0.89 .. 0.96]
2a	1.03	1	m	-	M38	[1.00 .. 1.04]
57<->, 48a	1.07	2	m	-	M37	[1.04 .. 1.10]
43	1.17	3	s	-	M06	[1.13 .. 1.21]
45<->	1.36	1	m	-	M36	[1.33 .. 1.38]
51<->, 58<->	1.40	2	m	-	M35	
58<->	1.49	1	d	12.72	M34	
50, 51<->	1.54	3	m	-	M33	
46<->, 45<->, 59	1.78	4	m	-	M32	
57<->	1.90	1	m	-	M05	
55<->	1.95	1	m	-	M04	
55<->	2.06	1	m	-	M03	
44<->	2.31	1	m	-	M02	
52a	2.58	1	br. s.	-	M01	
19<ax>	2.86	1	m	-	M31	
15<ax>	2.92	1	m	-	M30	
31<ax>, 24<ax>, 29<ax>	3.07	2	m	-	M29	
17<ax>, 12<ax>, 21<ax>	3.16	4	m	-	M28	
33<ax>	3.21	1	m	-	M27	
39<ax>, 37<ax>	3.32	2	m	-	M26	
34<->, 13<->	3.41	2	m	-	M25	
40<->	3.50	1	m	-	M24	
40<->	3.64	1	dd	10.51, 4.16	M23	
34<->, 13<->, 21<ax>	3.73	3	d	7.83	M22	
8<ax>	3.78	1	s	-	M21	
25	4.12	1	m	-	M41	
14	4.34	1	t	5.14	M20	
23<ax>	4.48	1	d	7.83	M08	
41	4.56	1	t	5.62	M19	
35	4.66	1	t	4.89	M18	
20	4.69	1	d	2.45	M17	
60<a>	4.72	1	s	-	M16	
60<b>	4.79	1	br. s.	-	M15	
10<ax>	4.86	1	d	7.83	M09	[4.84 .. 4.88]
38, 30	4.95	2	br. s.	-	M14	[4.93 .. 4.99]
18	5.04	1	d	4.89	M13	[5.02 .. 5.06]
32	5.11	1	br. s.	-	M12	[5.09 .. 5.13]
16	5.16	1	d	6.36	M11	[5.14 .. 5.18]
7<ax>	5.49	2	d	8.31	M10	[5.46 .. 5.53]

## Kaurenoic Acid+3Glc (isomer 2)

<sup>1</sup>H NMR (800 MHz, DMSO d<sub>6</sub>) δ ppm 0.73 - 0.80 (m, 1 H) 0.83 (s, 3 H) 0.92 (td, *J*=13.33, 4.16 Hz, 1 H) 1.00 - 1.04 (m, 1 H) 1.04 - 1.10 (m, 2 H) 1.17 (s, 3 H) 1.33 - 1.38 (m, 1 H) 1.38 - 1.44 (m, 2 H) 1.49 (d, *J*=12.72 Hz, 1 H) 1.51 - 1.59 (m, 3 H) 1.75 - 1.83 (m, 4 H) 1.88 - 1.93 (m, 1 H) 1.93 - 1.98 (m, 1 H) 2.03 - 2.09 (m, 1 H) 2.27 - 2.34 (m, 1 H) 2.58 (br. s., 1 H) 2.83 - 2.88 (m, 1 H) 2.89 - 2.95 (m, 1 H) 3.04 - 3.10 (m, 2 H) 3.12 - 3.19 (m, 4 H) 3.19 - 3.23 (m, 1 H) 3.30 - 3.35 (m, 2 H) 3.39 - 3.44 (m, 2 H) 3.4 - 3.53 (m, 1 H) 3.64 (dd, *J*=10.51, 4.16 Hz, 1 H) 3.73 (d, *J*=7.83 Hz, 3 H) 3.78 (s, 1 H) 4.09 - 4.14 (m, 1 H) 4.34 (t, *J*=5.14 Hz, 1 H) 4.48 (d, *J*=7.83 Hz, 1 H) 4.56 (t, *J*=5.62 Hz, 1 H) 4.66 (t, *J*=4.89 Hz, 1 H) 4.69 (d, *J*=2.45 Hz, 1 H) 4.72 (s, 1 H) 4.79 (br. s., 1 H) 4.86 (d, *J*=7.83 Hz, 1 H) 4.95 (br. s., 2 H) 5.04 (d, *J*=4.89 Hz, 1 H) 5.11 (br. s., 1 H) 5.16 (d, *J*=6.36 Hz, 1 H) 5.49 (d, *J*=8.31 Hz, 2 H)

<sup>13</sup>C NMR (201.21 MHz, DMSO d<sub>6</sub>) δ ppm 175.1 (1C), 155.4 (1C), 103.7 (1C), 103.2 (1C), 101.5 (1C), 92.1 (1C), 86.8 (1C), 77.9 (1C), 77.3 (1C), 77.1 (1C), 77.0 (1C), 76.8 (1C), 76.7 (1C), 74.4 (1C), 73.9 (1C), 71.2 (1C), 70.2 (1C), 68.2 (1C), 61.8 (1C), 61.1 (1C), 60.6 (1C), 56.7 (1C), 54.6 (1C), 48.8 (1C), 44.0 (1C), 43.6 (1C), 43.4 (1C), 41.1 (1C), 40.0 (1C), 39.5 (1C), 39.4 (1C), 37.1 (1C), 32.8 (1C), 28.4 (1C), 21.6 (1C), 19.3 (1C), 18.2 (1C), 16.2 (1C)

Figure 8F

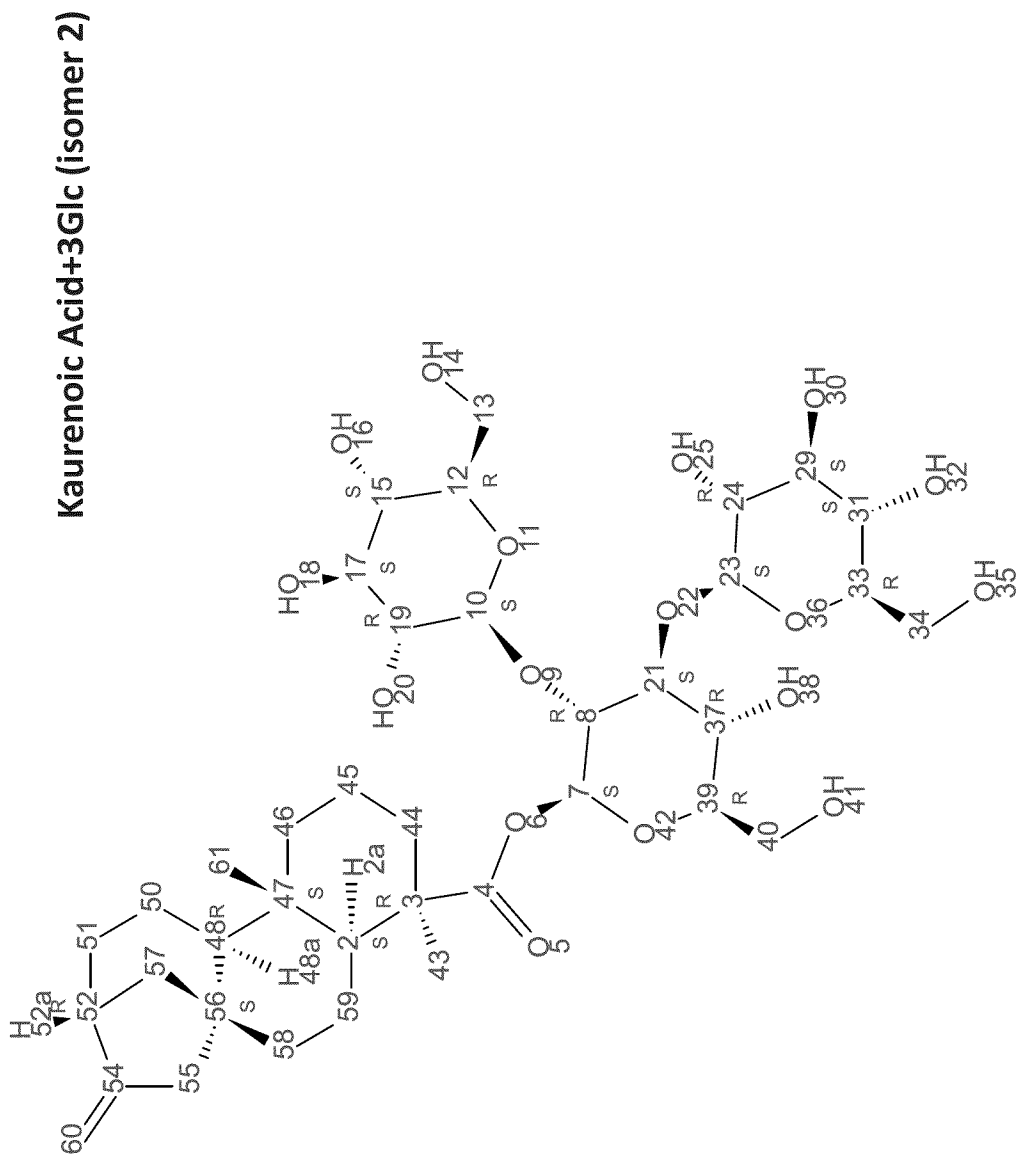




Figure 8H

## Kaurenoic Acid+2Glc+1GlcNAC

Atom1	Shift1 (ppm)	H's	Type	J (Hz)	Multiplet1	(ppm)
55<->	0.77	1	br. s.	-	M12	[0.75 .. 0.80]
57	0.87	3	s	-	M14	[0.86 .. 0.88]
53<->	0.96	1	m	-	M13	[0.93 .. 1.00]
2a, 13a, 10<->	1.05	3	m	-	M15	[1.01 .. 1.08]
52	1.15	3	s	-	M16	[1.13 .. 1.17]
54<->, 4<->, 11<->	1.38	3	m	-	M17	[1.33 .. 1.43]
11<->	1.47	1	d	13.20	M18	[1.45 .. 1.49]
3<->	1.53	1	d	10.27	M19	[1.50 .. 1.55]
3<->, 4<->	1.57	2	d	11.74	M20	[1.55 .. 1.62]
12<->	1.76	1	br. s.	-	M21	[1.72 .. 1.77]
12<->, 55<->, 54<->, 3	1.82	6	m	-	M37	[1.77 .. 1.87]
10<->, 8<->	1.95	2	d	14.18	M22	[1.93 .. 2.00]
8<->	2.05	1	d	17.12	M23	[2.03 .. 2.08]
53<->	2.18	1	d	12.23	M24	[2.14 .. 2.20]
5a	2.57	1	br. s.	-	M25	[2.56 .. 2.59]
38<ax>	3.00	1	br. s.	-	M28	[2.98 .. 3.03]
48<ax>, 23<ax>	3.08	2	m	-	M29	[3.04 .. 3.12]
32<ax>, 30	3.15	2	m	-	M30	[3.13 .. 3.19]
40, 46, 36, 28	3.24	4	m	-	M31	[3.20 .. 3.31]
41<->, 26	3.41	2	m	-	M36	[3.38 .. 3.43]
49<->	3.46	1	m	-	M35	[3.43 .. 3.49]
33<->	3.56	1	m	-	M34	[3.53 .. 3.60]
33<->, 44<ax>	3.65	2	m	-	M33	[3.60 .. 3.67]
41<->, 49<->, 20<ax>	3.72	3	m	-	M32	[3.67 .. 3.77]
50	4.33	1	t	5.38	M03	[4.31 .. 4.35]
37, 34, 42	4.47	3	m	-	M04	[4.43 .. 4.49]
25<ax>	4.53	1	d	8.31	M05	[4.51 .. 4.54]
58<a>	4.72	1	br. s.	-	M06	[4.71 .. 4.73]
58<b>	4.78	1	br. s.	-	M07	[4.78 .. 4.79]
29	4.89	1	d	5.38	M08	[4.86 .. 4.90]
22<ax>	4.95	1	d	7.83	M09	[4.93 .. 4.97]
31, 39, 45	5.01	3	m	-	M10	[4.97 .. 5.05]
47	5.08	1	d	5.38	M11	[5.05 .. 5.10]
19<ax>	5.46	1	d	7.82	M02	[5.43 .. 5.49]
27	7.82	1	m	-	M01	[7.79 .. 7.84]

<sup>1</sup>H NMR (800 MHz, DMSO *d*<sub>6</sub>) δ ppm 0.77 (br. s., 1 H) 0.87 (s, 3 H) 0.93 - 1.00 (m, 1 H) 1.01 - 1.08 (m, 3 H) 1.15 (s, 3 H) 1.33 - 1.43 (m, 3 H) 1.47 (d, *J*=13.20 Hz, 1 H) 1.53 (d, *J*=10.27 Hz, 1 H) 1.57 (d, *J*=11.74 Hz, 2 H) 1.76 (br. s., 1 H) 1.77 - 1.87 (m, 6 H) 1.95 (d, *J*=14.18 Hz, 2 H) 2.05 (d, *J*=17.12 Hz, 1 H) 2.18 (d, *J*=12.23 Hz, 1 H) 2.57 (br. s., 1 H) 3.00 (br. s., 1 H) 3.04 - 3.12 (m, 2 H) 3.13 - 3.19 (m, 2 H) 3.20 - 3.31 (m, 4 H) 3.38 - 3.43 (m, 2 H) 3.43 - 3.49 (m, 1 H) 3.53 - 3.60 (m, 1 H) 3.60 - 3.67 (m, 2 H) 3.67 - 3.77 (m, 3 H) 4.33 (t, *J*=5.38 Hz, 1 H) 4.43 - 4.49 (m, 3 H) 4.53 (d, *J*=8.31 Hz, 1 H) 4.72 (br. s., 1 H) 4.78 (br. s., 1 H) 4.89 (d, *J*=5.38 Hz, 1 H) 4.95 (d, *J*=7.83 Hz, 1 H) 4.97 - 5.05 (m, 3 H) 5.08 (d, *J*=5.38 Hz, 1 H) 5.46 (d, *J*=7.82 Hz, 1 H) 7.79 - 7.84 (m, 1 H)

<sup>13</sup>C NMR (201.21 MHz, DMSO *d*<sub>6</sub>) δ ppm 175.6 (1C), 171.6 (1C), 155.5 (1C), 103.6 (1C), 101.9 (1C), 99.9 (1C), 92.5 (1C), 82.1 (1C), 77.6 (1C), 77.3 (1C), 77.0 (1C), 77.0 (1C), 76.1 (1C), 75.9 (1C), 74.0 (1C), 70.5 (1C), 70.3 (1C), 69.9 (1C), 61.8 (1C), 61.0 (1C), 60.9 (1C), 57.2 (1C), 56.7 (1C), 54.6 (1C), 48.8 (1C), 44.1 (1C), 43.8 (1C), 43.4 (1C), 41.3 (1C), 40.4 (1C), 39.9 (1C), 39.5 (1C), 37.6 (1C), 32.9 (1C), 28.5 (1C), 23.4 (1C), 21.6 (1C), 19.2 (1C), 18.4 (1C), 16.2 (1C)

Kaurenoic Acid+2Glc+1GlcNAc

Figure 81

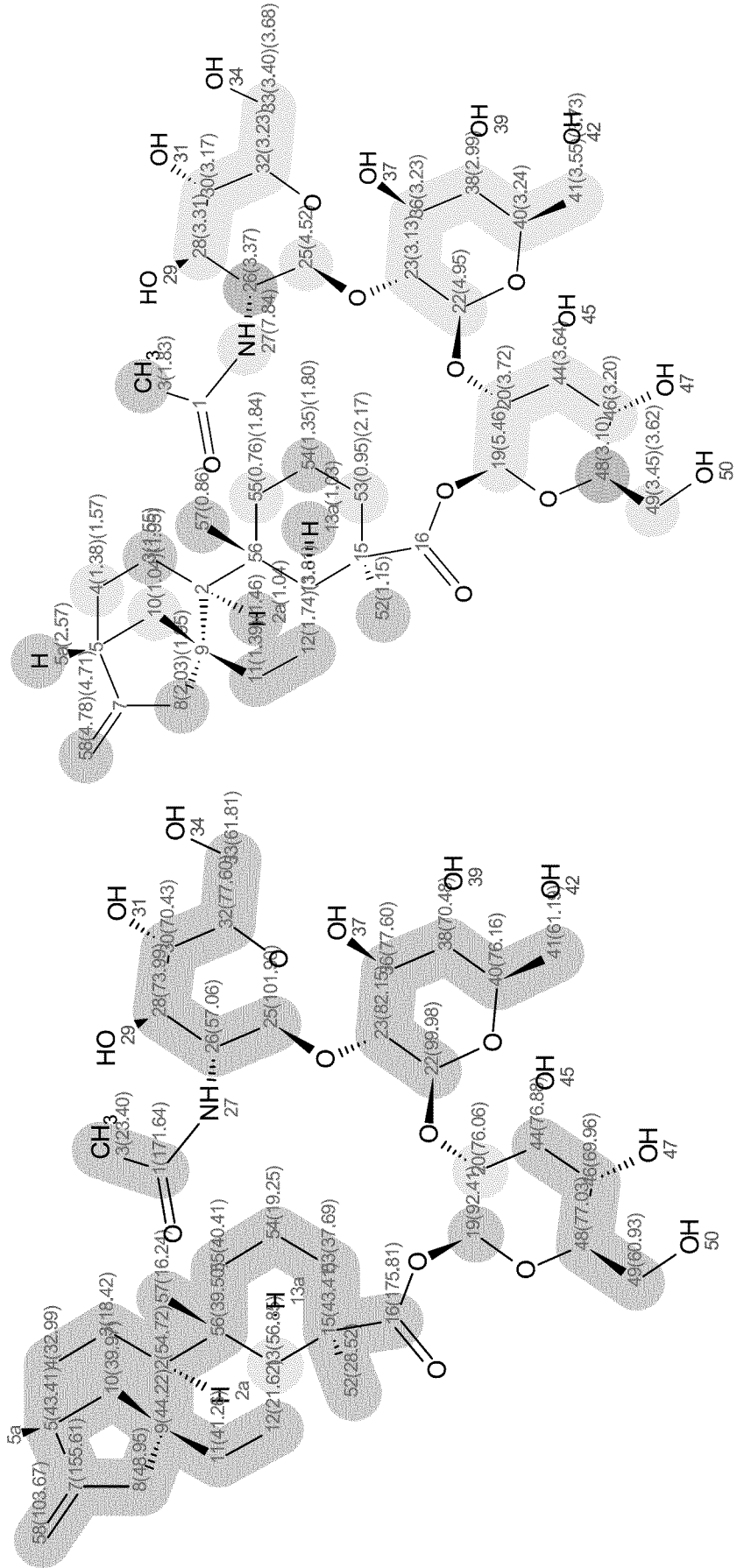


Figure 8J

Kaurenol+3Glc (isomer 1)

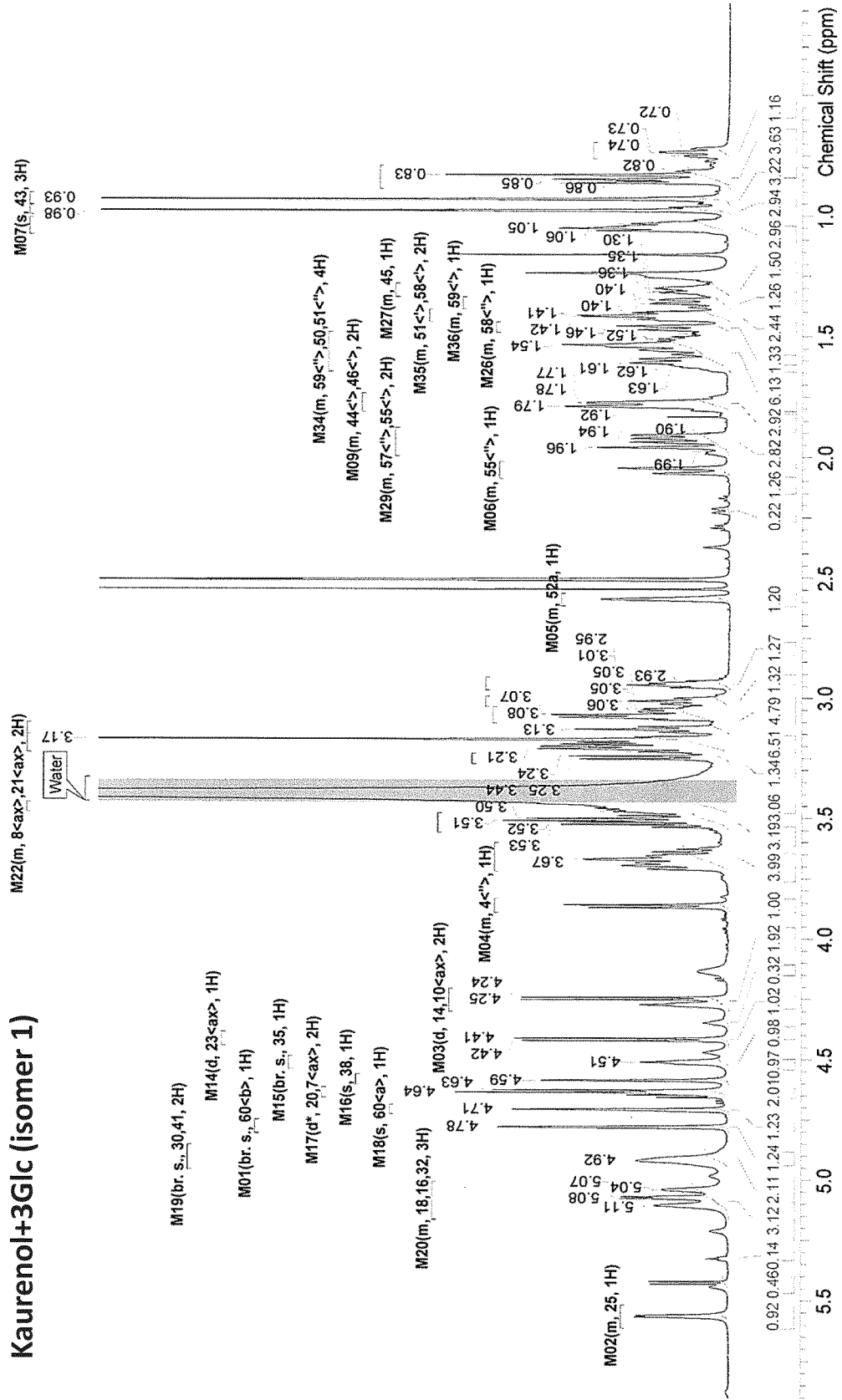
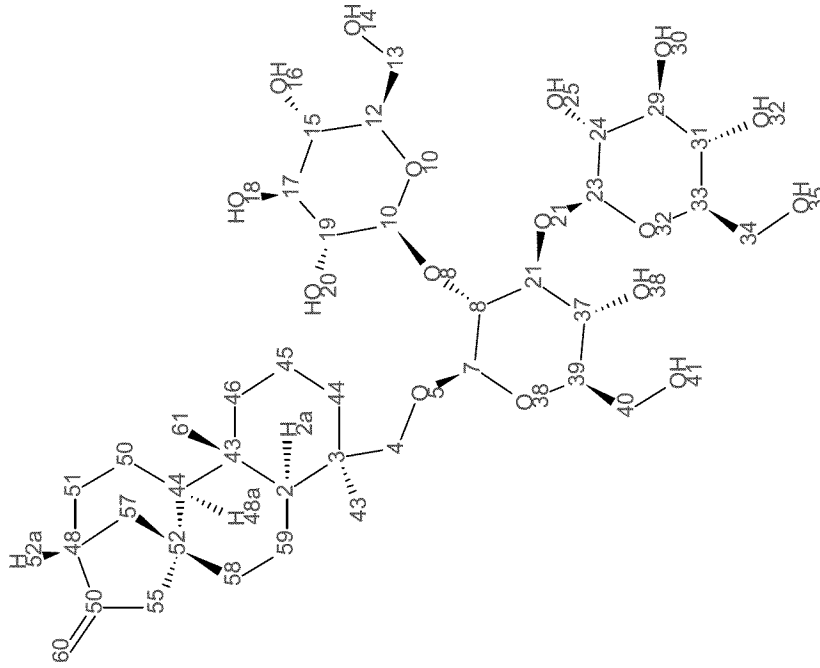


Figure 8K

Kaurenol+3Glc (isomer 1)

Atom1	Shift1 (ppm)	H's	Type	J (Hz)	Multiplet1	(ppm)
46<sup>1</sup>	0.74	1	m	-	M32	[0.70 .. 0.76]
44<sup>1</sup>, 2a	0.84	2	m	-	M25	[0.79 .. 0.89]
61	0.93	3	s	-	M08	[0.91 .. 0.96]
43	0.98	3	s	-	M07	[0.96 .. 1.00]
48a, 57<sup>1</sup>	1.04	2	m	-	M28	[1.01 .. 1.08]
45	1.32	1	m	-	M27	[1.28 .. 1.34]
59<sup>1</sup>	1.36	1	m	-	M36	[1.34 .. 1.39]
51<sup>1</sup>, 58<sup>1</sup>	1.41	2	m	-	M35	[1.39 .. 1.44]
58<sup>1</sup>	1.46	1	m	-	M26	[1.44 .. 1.48]
59<sup>1</sup>, 50, 51<sup>1</sup>	1.56	4	m	-	M34	[1.48 .. 1.65]
44<sup>1</sup>, 46<sup>1</sup>	1.78	2	m	-	M09	[1.74 .. 1.82]
57<sup>1</sup>, 55<sup>1</sup>	1.94	2	m	-	M29	[1.88 .. 2.00]
55<sup>1</sup>	2.06	1	m	-	M06	[2.02 .. 2.09]
52a	2.59	1	m	-	M05	[2.57 .. 2.62]
19<sup>1</sup>	2.94	1	m	-	M24	[2.91 .. 2.97]
37<sup>1</sup>	3.02	1	m	-	M23	[2.99 .. 3.04]
31<sup>1</sup>, 24<sup>1</sup>	3.07	2	m	-	M30	[3.04 .. 3.11]
15<sup>1</sup>	3.13	1	t	9.05	M11	[3.11 .. 3.15]
17<sup>1</sup>, 29<sup>1</sup>, 12<sup>1</sup>, 33<sup>1</sup>, 39<sup>1</sup>	3.19	5	m	-	M31	[3.15 .. 3.23]
4<sup>1</sup>	3.24	1	m	-	M33	[3.23 .. 3.28]
8<sup>1</sup>, 21<sup>1</sup>	3.46	2	m	-	M22	[3.44 .. 3.48]
13<sup>1</sup>, 34<sup>1</sup>, 40<sup>1</sup>	3.51	3	dd	11.74, 7.82	M21	[3.48 .. 3.56]
13<sup>1</sup>, 34<sup>1</sup>, 40<sup>1</sup>	3.67	3	br. s.	-	M12	[3.60 .. 3.72]
4<sup>1</sup>	3.86	1	m	-	M04	[3.83 .. 3.89]
14, 10<sup>1</sup>	4.25	2	d	7.34	M03	[4.21 .. 4.30]
23<sup>1</sup>	4.42	1	d	7.83	M14	[4.39 .. 4.45]
35	4.51	1	br. s.	-	M15	[4.49 .. 4.54]
38	4.59	1	s	-	M16	[4.56 .. 4.61]
20, 7<sup>1</sup>	4.63	2	d	8.31	M17	[4.62 .. 4.66]
60<sup>1</sup>	4.71	1	s	-	M18	[4.69 .. 4.73]
60<sup>1</sup>	4.78	1	br. s.	-	M01	[4.75 .. 4.81]
30, 41	4.92	2	br. s.	-	M19	[4.86 .. 4.96]
18, 16, 32	5.07	3	m	-	M20	[5.01 .. 5.17]
25	5.57	1	m	-	M02	[5.52 .. 5.62]



## Figure 8L

## Kaurenol+3Glc (isomer 1)

<sup>1</sup>H NMR (800 MHz, DMSO *d*<sub>6</sub>) δ ppm 0.70 - 0.76 (m, 1 H) 0.79 - 0.89 (m, 2 H) 0.93 (s, 3 H) 0.98 (s, 3 H) 1.01 - 1.08 (m, 2 H) 1.28 - 1.34 (m, 1 H) 1.34 - 1.39 (m, 1 H) 1.39 - 1.44 (m, 2 H) 1.44 - 1.48 (m, 1 H) 1.48 - 1.65 (m, 4 H) 1.74 - 1.82 (m, 2 H) 1.88 - 2.00 (m, 2 H) 2.02 - 2.09 (m, 1 H) 2.57 - 2.62 (m, 1 H) 2.91 - 2.97 (m, 1 H) 2.99 - 3.04 (m, 1 H) 3.04 - 3.11 (m, 2 H) 3.13 (t, *J*=9.05 Hz, 1 H) 3.15 - 3.23 (m, 5 H) 3.23 - 3.28 (m, 1 H) 3.44 - 3.48 (m, 2 H) 3.51 (dd, *J*=11.74, 7.82 Hz, 3 H) 3.67 (br. s., 3 H) 3.83 - 3.89 (m, 1 H) 4.25 (d, *J*=7.34 Hz, 2 H) 4.42 (d, *J*=7.83 Hz, 1 H) 4.51 (br. s., 1 H) 4.59 (s, 1 H) 4.63 (d, *J*=8.31 Hz, 2 H) 4.71 (s, 1 H) 4.78 (br. s., 1 H) 4.92 (br. s., 2 H) 5.01 - 5.17 (m, 3 H) 5.52 - 5.62 (m, 1 H)

<sup>13</sup>C NMR (201.21 MHz, DMSO *d*<sub>6</sub>) δ ppm 155.8 (1C), 103.6 (1C), 103.4 (1C), 102.3 (1C), 102.1 (1C), 86.7 (1C), 78.6 (1C), 77.1 (1C), 77.0 (1C), 76.5 (1C), 76.4 (1C), 76.2 (1C), 74.7 (1C), 73.8 (1C), 72.2 (1C), 70.5 (1C), 70.2 (1C), 68.8 (1C), 61.5 (1C), 61.3 (1C), 61.1 (1C), 56.7 (1C), 55.9 (1C), 48.9 (1C), 44.1 (1C), 43.6 (1C), 41.4 (1C), 40.4 (1C), 39.4 (1C), 39.1 (1C), 37.7 (1C), 36.4 (1C), 33.1 (1C), 28.0 (1C), 20.4 (1C), 18.4 (1C), 18.2 (1C), 18.1 (1C)

Figure 8M

Steviol+6Glc (isomer 1)

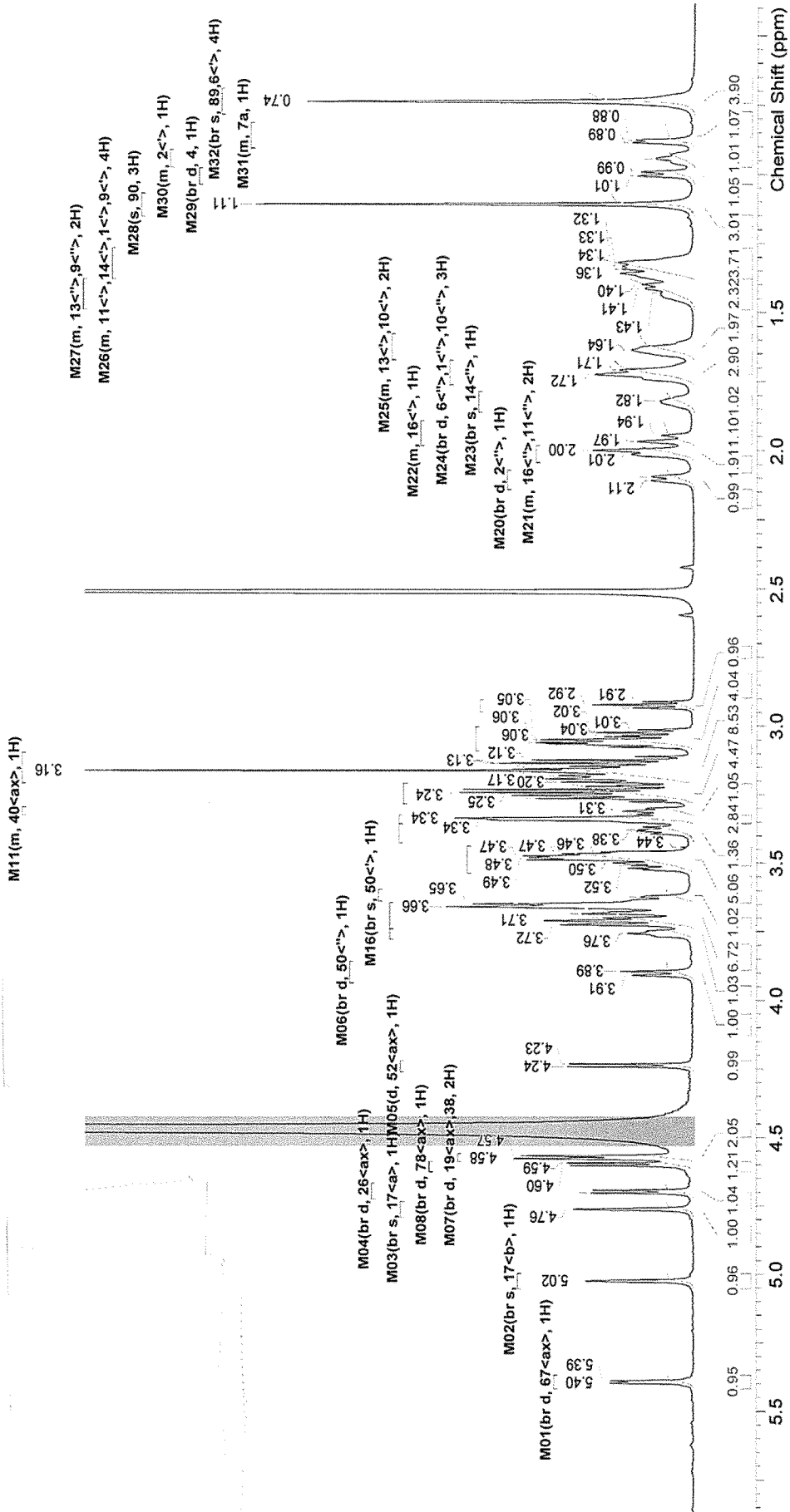
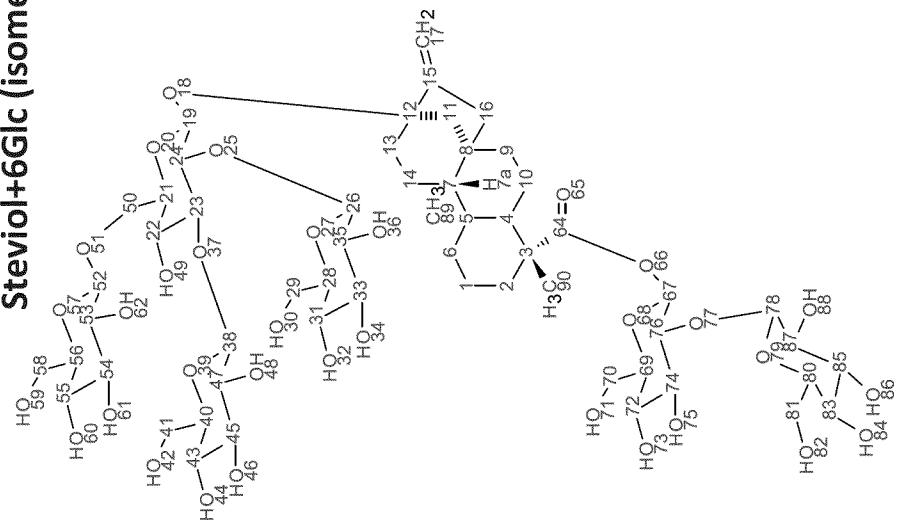


Figure 8N

Steviol+6Glc (isomer 1)

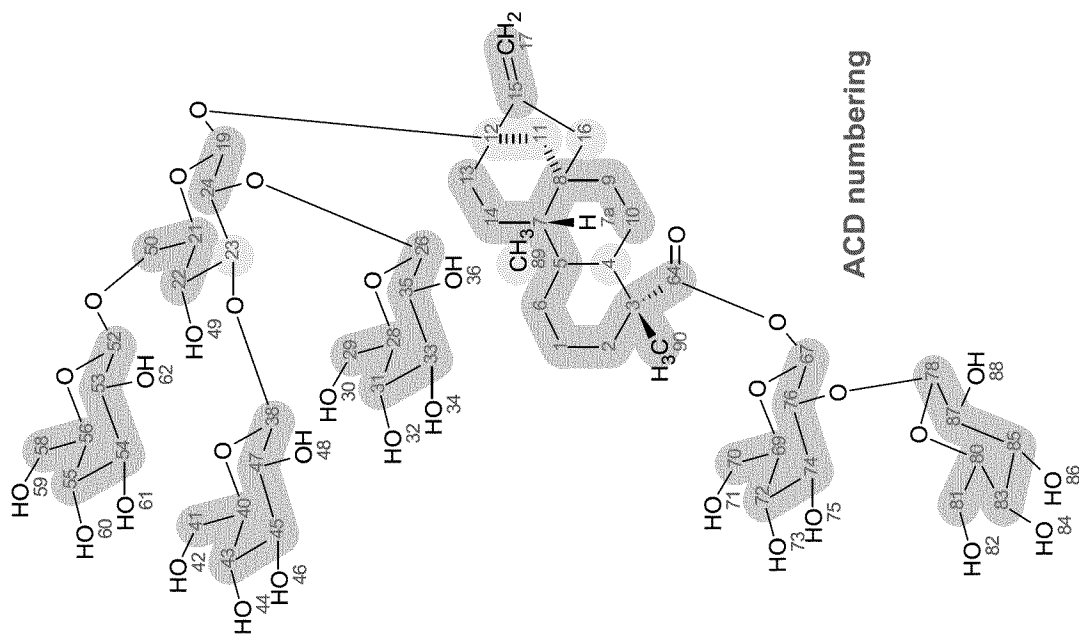


No.	Atom1	Shift1 (ppm)	H's	Type	J (Hz)	Multiplet1	(ppm)
1	89, 6<->	0.74	4	br s	-	M32	[0.69 .. 0.78]
2	7a	0.88	1	m	-	M31	[0.81 .. 0.91]
3	2<->	0.94	1	m	-	M30	[0.91 .. 0.98]
4	4	1.00	1	br d	12.72	M29	[0.98 .. 1.04]
5	90	1.11	3	s	-	M28	[1.07 .. 1.15]
6	11<->, 14<->, 1<->, 9<->	1.34	4	m	-	M26	[1.27 .. 1.38]
7	13<->, 9<->	1.42	2	m	-	M27	[1.38 .. 1.49]
8	13<->, 10<->	1.64	2	m	-	M25	[1.58 .. 1.67]
9	6<->, 1<->, 10<->	1.72	3	br d	12.23	M24	[1.67 .. 1.76]
10	14<->	1.82	1	br s	-	M23	[1.79 .. 1.86]
11	16<->	1.96	1	m	-	M22	[1.89 .. 1.98]
12	16<->, 11<->	2.00	2	m	-	M21	[1.98 .. 2.04]
13	2<->	2.10	1	br d	11.49	M20	[2.07 .. 2.14]
14	31<ax>	2.92	1	m	-	M15	[2.90 .. 2.95]
15	85<ax>, 87<ax>, 35<ax>, 53<ax>	3.05	4	m	-	M14	[3.00 .. 3.09]
16	80<ax>, 83<ax>, 43<ax>, 55<ax>, 56<ax>, 28<ax>, 47<ax>, 33<ax>	3.16	8	m	-	M13	[3.10 .. 3.21]
17	45<ax>, 54<ax>, 74<ax>, 72<ax>	3.25	4	br dd	17.24, 8.93	M19	[3.21 .. 3.28]
18	40<ax>	3.31	1	m	-	M11	[3.30 .. 3.32]
19	69<ax>, 21<ax>, 22<ax>	3.34	3	br d	4.65	M18	[3.32 .. 3.35]
20	29<->	3.38	1	m	-	M12	[3.36 .. 3.43]
21	81<->, 41<->, 58<->, 70<->, 24<ax>	3.48	5	m	-	M10	[3.44 .. 3.54]
22	50<->	3.62	1	br s	-	M16	[3.59 .. 3.64]
23	81<->, 41<->, 58<->, 76<ax>, 70<->, 29<->	3.68	6	m	-	M09	[3.64 .. 3.74]
24	23<ax>	3.75	1	br d	8.56	M17	[3.74 .. 3.78]
25	50<->	3.90	1	br d	11.25	M06	[3.86 .. 3.94]
26	52<ax>	4.24	1	d	7.82	M05	[4.22 .. 4.26]
27	19<ax>, 38	4.57	2	br d	7.83	M07	[4.56 .. 4.59]
28	78<ax>	4.60	1	br d	7.58	M08	[4.59 .. 4.63]
29	26<ax>	4.70	1	br d	7.82	M04	[4.67 .. 4.72]
30	17<a>	4.76	1	br s	-	M03	[4.73 .. 4.79]
31	17<b>	5.02	1	br s	-	M02	[5.00 .. 5.05]
32	67<ax>	5.39	1	br d	7.34	M01	[5.37 .. 5.42]

<sup>1</sup>H NMR (800 MHz, DMSO-d<sub>6</sub>) δ ppm 0.74 (br s, 4 H) 0.81 - 0.91 (m, 1 H) 0.91 - 0.98 (m, 1 H) 1.00 (br d, J=12.72 Hz, 1 H) 1.11 (s, 3 H) 1.27 - 1.38 (m, 4 H) 1.38 - 1.49 (m, 2 H) 1.58 - 1.67 (m, 2 H) 1.72 (br d, J=12.23 Hz, 3 H) 1.82 (br s, 1 H) 1.89 - 1.98 (m, 1 H) 1.98 - 2.04 (m, 2 H) 2.10 (br d, J=11.49 Hz, 1 H) 2.90 - 2.95 (m, 1 H) 3.00 - 3.09 (m, 4 H) 3.10 - 3.21 (m, 8 H) 3.25 (br dd, J=17.24, 8.93 Hz, 4 H) 3.30 - 3.32 (m, 1 H) 3.34 (br d, J=4.65 Hz, 3 H) 3.36 - 3.43 (m, 1 H) 3.44 - 3.54 (m, 5 H) 3.62 (br s, 1 H) 3.64 - 3.74 (m, 6 H) 3.75 (br d, J=8.56 Hz, 1 H) 3.90 (br d, J=11.25 Hz, 1 H) 4.24 (d, J=7.82 Hz, 1 H) 4.57 (br d, J=7.83 Hz, 2 H) 4.60 (br d, J=7.58 Hz, 1 H) 4.70 (br d, J=7.82 Hz, 1 H) 4.76 (br s, 1 H) 5.02 (br s, 1 H) 5.39 (br d, J=7.34 Hz, 1 H)

Figure 80

Steviol+6Glc (isomer 1)



F2 Atom	F2 (ppm)	F2 Atom	F2 (ppm)	F1 Atom	F1 (ppm)	F1 Atom	F1 (ppm)
1<->	1.71	31<-ax>	2.91	1	19.9	31	71.5
1<->	1.31	33<-ax>	3.18			33	77.2
2<->	2.09	35<-ax>	3.01	2	37.7	35	75.4
2<->	0.93	38<-ax>	4.56			38	103.3
4	0.99	40<-ax>	3.33	3	44.8	40	77.8
		41<->	3.69	4	57.4	41	61.9
		41<->	3.46	5	40.3		
6<->	1.71	43<-ax>	3.24	6	40.8	43	70.3
6<->	0.73	45<-ax>	3.23	7	54	45	76.9
7a	0.87	47<-ax>	3.12			47	74.4
		50<->	3.89	8	42.2	50	69.3
		50<->	3.62	9	42.1		
9<->	1.39	52<-ax>	4.23			52	103.9
9<->	1.31	53<-ax>	3.04	10	22.7	53	74.2
10<->	1.72	54<-ax>	3.18			54	77.2
10<->	1.62	55<-ax>	3.33	11	44.4	55	69.1
11<->	2	56<-ax>	3.3			56	77.2
11<->	1.34	58<->	3.68	12	88.4	58	62
		58<->	3.46	13	38.3		
13<->	1.81					64	178
13<->	1.36	67<-ax>	5.38	14	20.6	67	93.6
14<->	1.62	69<-ax>	3.23			69	76.9
14<->	1.43	70<->	3.64	15	153.2	70	61.6
		70<->	3.5	16	47.4		
16<->	1.99	72<-ax>	3.15			72	70.8
16<->	1.94	74<-ax>	3.65	17	105.8	74	77.2
17<-ax>	4.75	76	3.65			76	78.4
17<-b>	5.02	78<-ax>	4.59	19	96.4	78	103.3
19<-ax>	4.57	80<-ax>	3.33	21	77.6	80	75.6
21<-ax>	3.15	81<->	3.66	22	70.8	81	62.7
22<-ax>	3.12	81<->	3.37	23	86.5		
23<-ax>	3.75	83<-ax>	3.06	24	79.6	83	71.3
24<-ax>	3.48	85<-ax>	3.18	26	103	85	77.2
26<-ax>	4.69	87<-ax>	3.05	28	77.2	87	75.1
28<-ax>	3.3	89	0.73	29	62.1	89	17.1
29<->	3.71	90	1.1			90	29.2
29<->	3.69						

<sup>13</sup>C NMR (201 MHz, DMSO-d<sub>6</sub>) δ ppm 17.1, 19.9, 20.6, 22.7, 29.2, 37.7, 38.3, 40.3, 40.8, 42.1, 42.2, 44.4, 44.8, 47.4, 54.0, 57.4, 61.6, 61.9, 62.0, 62.1, 62.7, 69.1, 69.3, 70.3, 70.8, 70.8, 71.3, 71.5, 74.2, 74.4, 75.1, 75.4, 75.6, 76.9, 77.2, 77.2, 77.2, 77.2, 77.2, 77.2, 77.2, 77.6, 77.8, 78.4, 79.6, 86.5, 88.4, 93.6, 96.4, 103.0, 103.3, 103.3, 103.3, 103.3, 105.8, 153.2, 178.0

Figure 8P

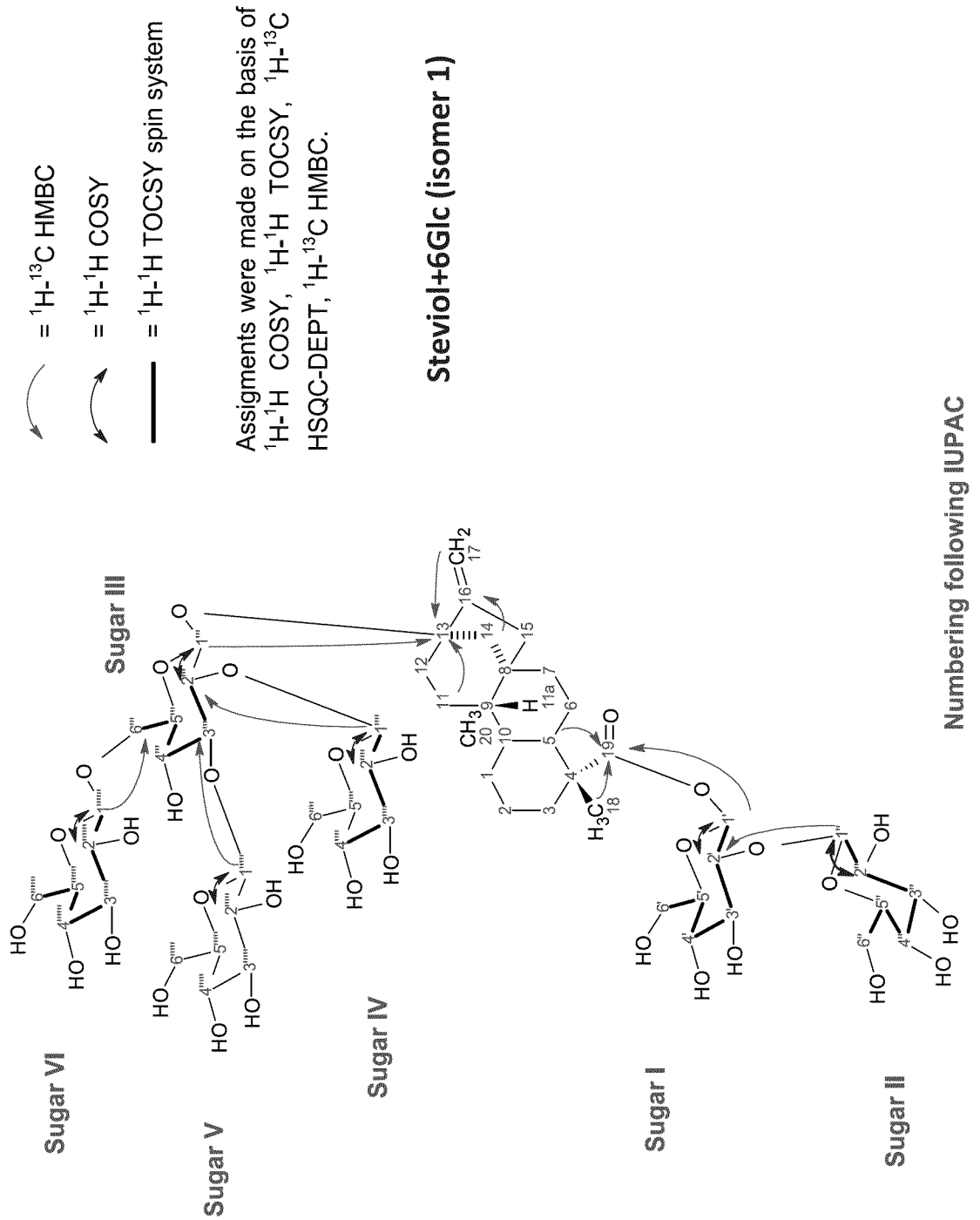
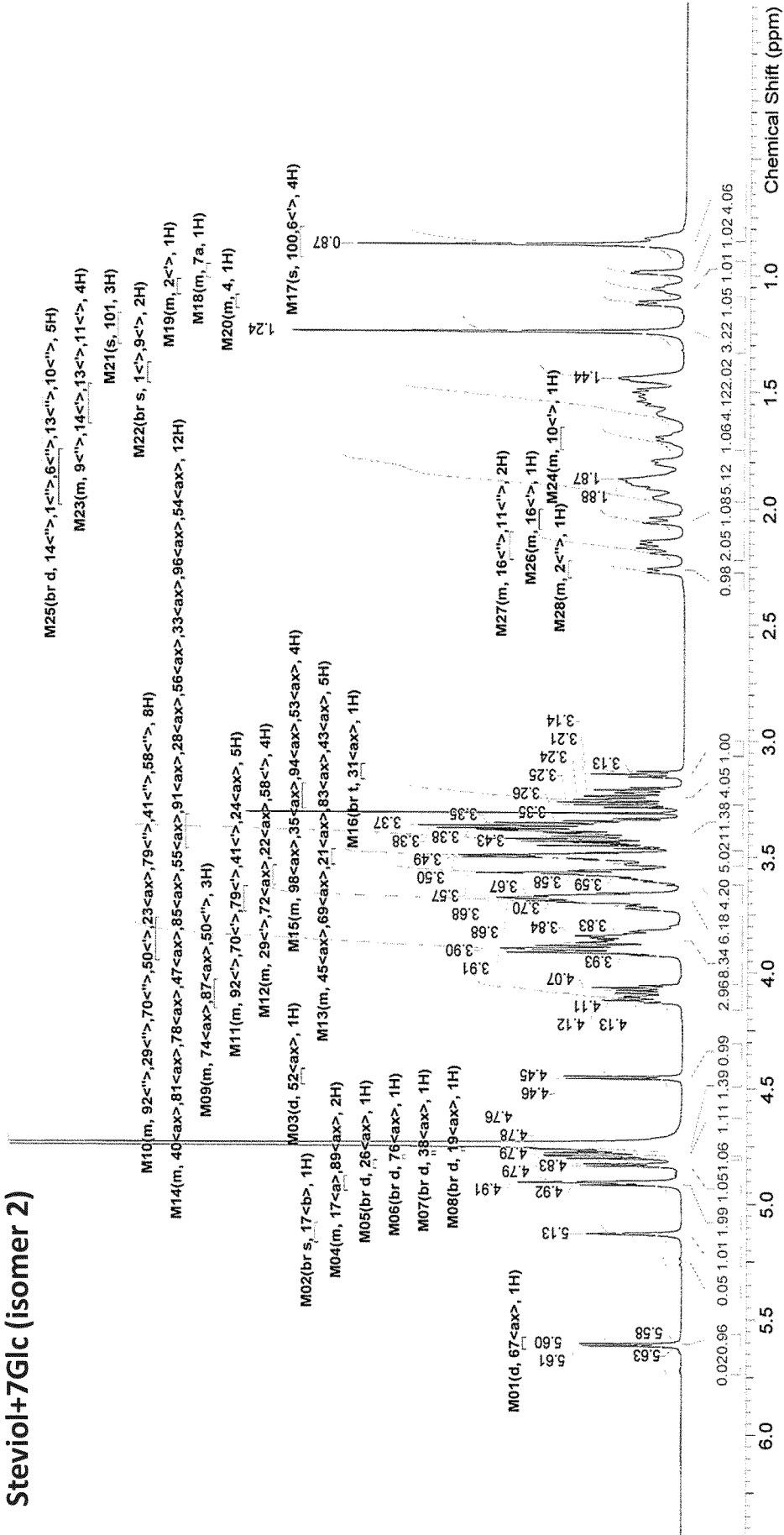


Figure 8Q

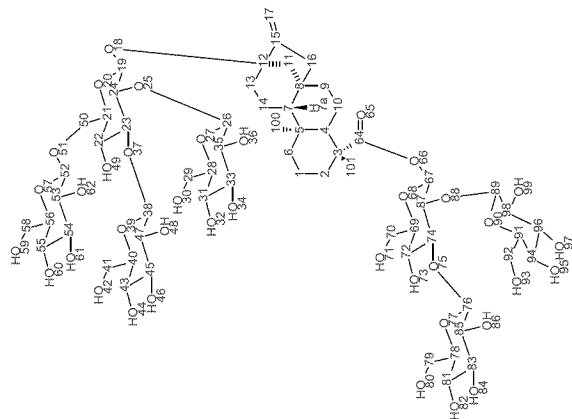
Steviol+7Glc (isomer 2)



## Figure 8R

## Steviol+7Glc (isomer 2)

No.	Atom1	Shift1 (ppm)	H's	Type	J (Hz)	Multiplet1	(ppm)
1	100, 6<->	0.87	4	s	-	M17	[0.80 .. 0.93]
2	7a	0.99	1	m	-	M18	[0.96 .. 1.02]
3	2<->	1.06	1	m	-	M19	[1.02 .. 1.09]
4	4	1.12	1	m	-	M20	[1.09 .. 1.15]
5	101	1.24	3	s	-	M21	[1.17 .. 1.30]
6	1<->, 9<->	1.44	2	br s	-	M22	[1.38 .. 1.47]
7	9<->, 14<->, 13<->, 11<->	1.56	4	m	-	M23	[1.47 .. 1.65]
8	10<->	1.70	1	m	-	M24	[1.65 .. 1.75]
9	14<->, 1<->, 6<->, 13<->, 10<->	1.88	5	br d	10.51	M25	[1.76 .. 1.99]
10	16<->	2.05	1	m	-	M26	[2.00 .. 2.09]
11	16<->, 11<->	2.16	2	m	-	M27	[2.10 .. 2.22]
12	2<->	2.26	1	m	-	M28	[2.23 .. 2.30]
13	31<-ax>	3.14	1	br t	9.41	M16	[3.10 .. 3.17]
14	98<-ax>, 35<-ax>, 94<-ax>, 53<-ax>	3.24	4	m	-	M15	[3.19 .. 3.30]
15	40<-ax>, 81<-ax>, 78<-ax>, 47<-ax>, 85<-ax>, 55<-ax>, 91<-ax>, 28<-ax>, 56<-ax>, 33<-ax>, 96<-ax>, 54<-	3.40	12	m	-	M14	[3.32 .. 3.47]
16	45<-ax>, 69<-ax>, 21<-ax>, 83<-ax>, 43<-ax>	3.50	5	m	-	M13	[3.47 .. 3.54]
17	29<->, 72<-ax>, 22<-ax>, 58<->	3.58	4	m	-	M12	[3.54 .. 3.63]
18	92<->, 70<->, 79<->, 41<->, 24<-ax>	3.69	5	m	-	M11	[3.63 .. 3.74]
19	92<->, 29<->, 70<->, 50<->, 23<-ax>, 79<->, 41<->, 58<->	3.86	8	m	-	M10	[3.77 .. 3.95]
20	74<-ax>, 87<-ax>, 50<->	4.09	3	m	-	M09	[4.04 .. 4.16]
21	52<-ax>	4.45	1	d	7.82	M03	[4.42 .. 4.49]
22	19<-ax>	4.77	1	br d	8.07	M08	[4.76 .. 4.77]
23	38<-ax>	4.78	1	br d	8.07	M07	[4.78 .. 4.79]
24	76<-ax>	4.80	1	br d	8.07	M06	[4.79 .. 4.81]
25	26<-ax>	4.83	1	br d	7.83	M05	[4.81 .. 4.85]
26	17<-a>, 89<-ax>	4.91	2	m	-	M04	[4.88 .. 4.95]
27	17<-b>	5.13	1	br s	-	M02	[5.09 .. 5.17]
28	67<-ax>	5.61	1	d	7.83	M01	[5.58 .. 5.63]



<sup>1</sup>H NMR (800 MHz, DEUTERIUM OXIDE) δ ppm 0.87 (s, 4 H) 0.96 - 1.02 (m, 1 H) 1.02 - 1.09 (m, 1 H) 1.09 - 1.15 (m, 1 H) 1.24 (s, 3 H) 1.44 (br s, 2 H) 1.47 - 1.65 (m, 4 H) 1.65 - 1.75 (m, 1 H) 1.88 (br d, *J*=10.51 Hz, 5 H) 2.00 - 2.09 (m, 1 H) 2.10 - 2.22 (m, 2 H) 2.23 - 2.30 (m, 1 H) 3.14 (br t, *J*=9.41 Hz, 1 H) 3.19 - 3.30 (m, 4 H) 3.32 - 3.47 (m, 12 H) 3.47 - 3.54 (m, 5 H) 3.54 - 3.63 (m, 4 H) 3.63 - 3.74 (m, 5 H) 3.77 - 3.95 (m, 8 H) 4.04 - 4.16 (m, 3 H) 4.45 (d, *J*=7.82 Hz, 1 H) 4.77 (br d, *J*=8.07 Hz, 1 H) 4.78 (br d, *J*=8.07 Hz, 1 H) 4.80 (br d, *J*=8.07 Hz, 1 H) 4.83 (br d, *J*=8.07 Hz, 1 H) 4.88 - 4.95 (m, 2 H) 5.13 (br s, 1 H) 5.61 (d, *J*=7.83 Hz, 1 H)



Figure 8T

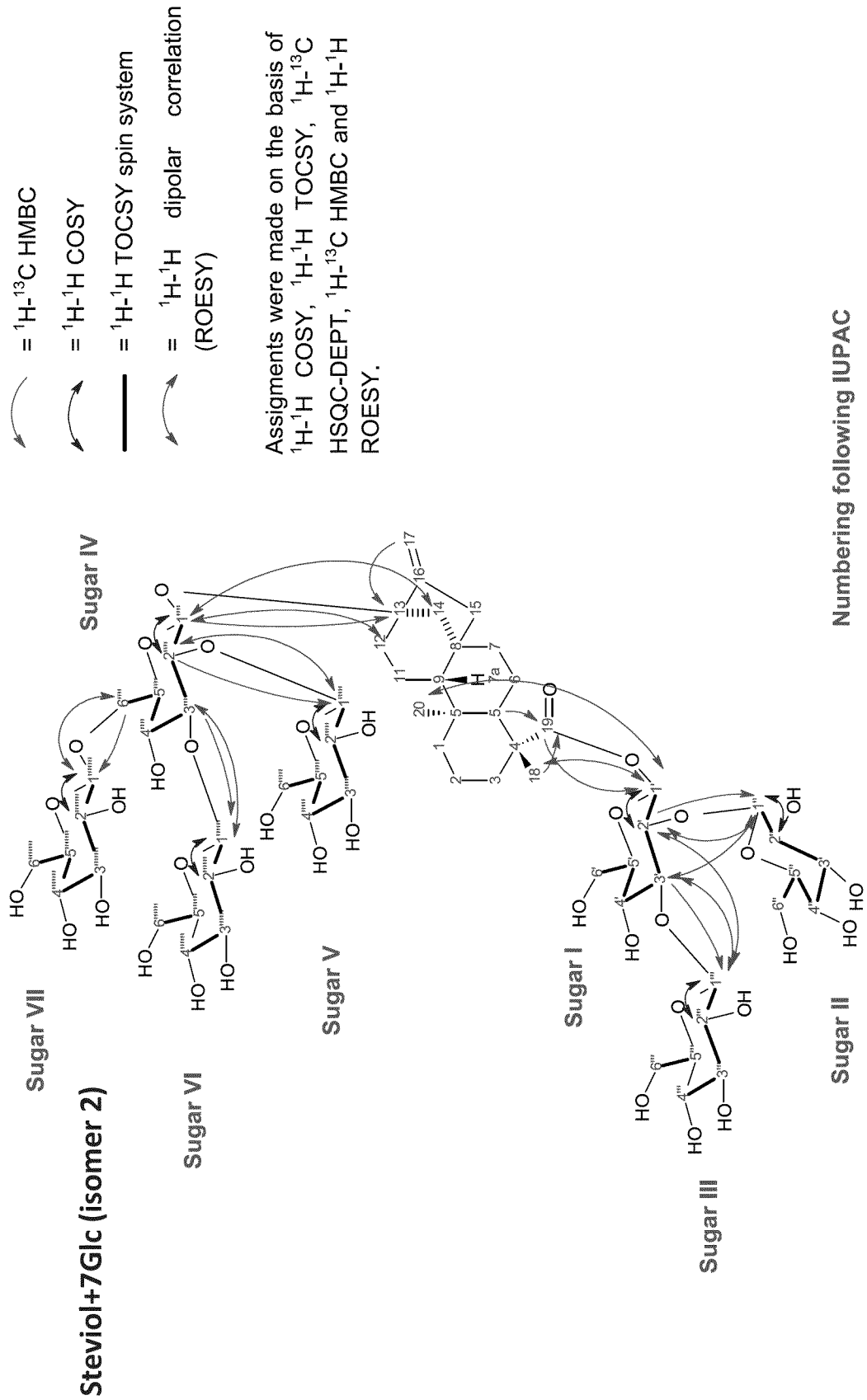


Figure 8U

Steviol+6Glc (isomer 4)

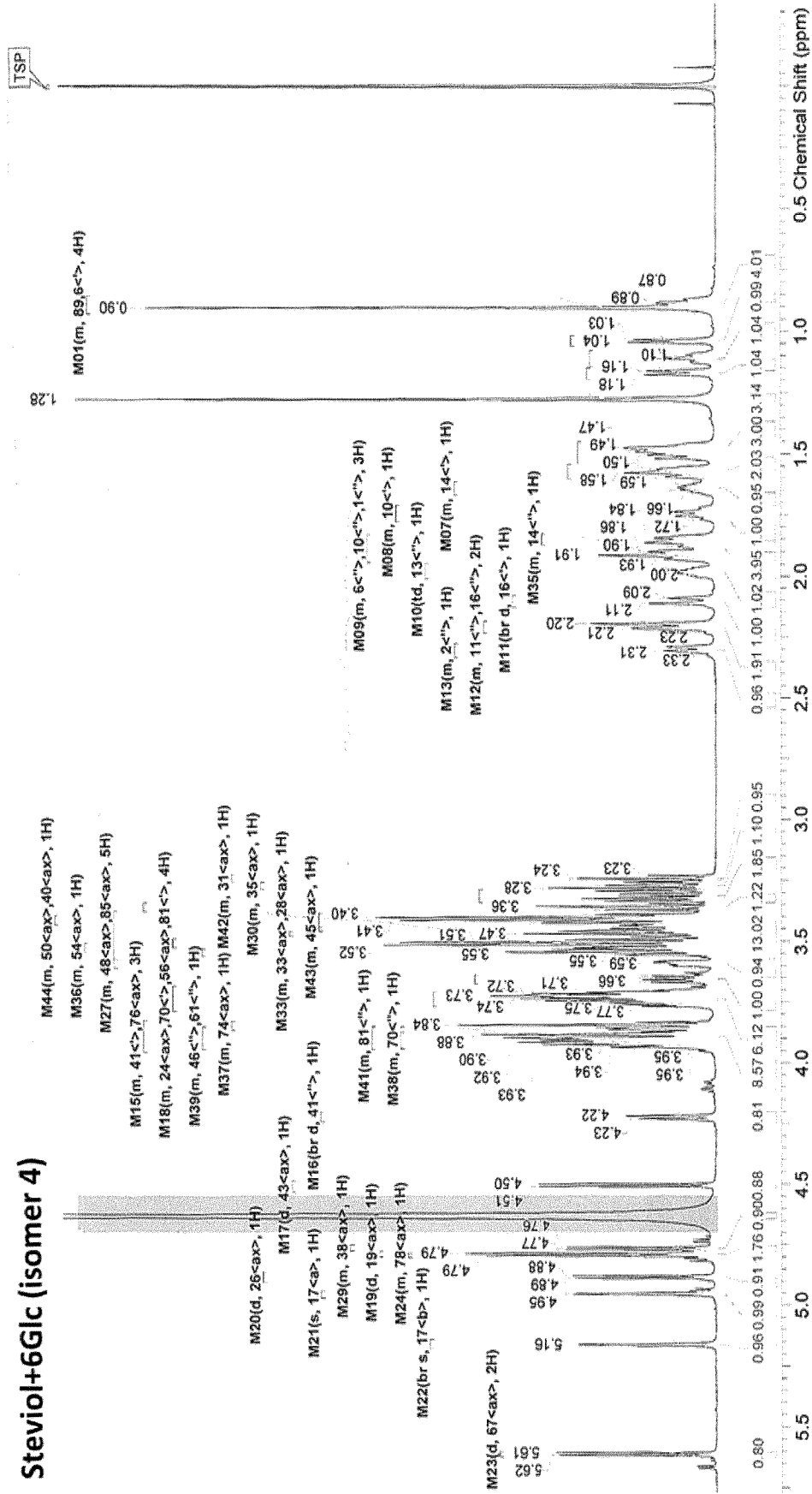


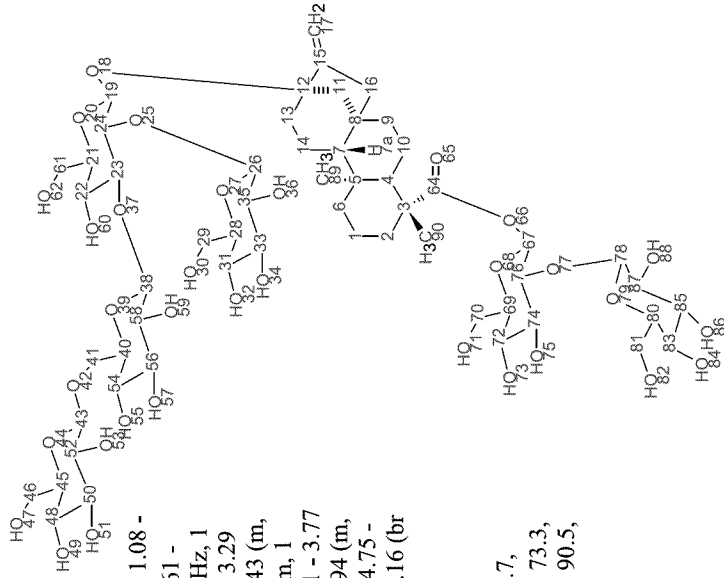
Figure 8V

Multiplet1	Shift1 (ppm)	Atom1	H's	Type	J (Hz)	(ppm)
M01	0.90	89, 6<->	4	m	-	[0.85 .. 0.93]
M02	1.04	7a	1	br d	8.07	[1.02 .. 1.06]
M03	1.11	2<->	1	m	-	[1.08 .. 1.14]
M04	1.17	4	1	br d	12.47	[1.15 .. 1.20]
M05	1.28	90	3	s	-	[1.25 .. 1.30]
M06	1.49	11<->, 9<->, 1<->	3	m	-	[1.45 .. 1.54]
M051	1.57	13<->, 9<->	2	m	-	[1.54 .. 1.61]
M07	1.64	14<->	1	m	-	[1.61 .. 1.67]
M08	1.74	10<->	1	m	-	[1.71 .. 1.78]
M35	1.85	14<->	1	m	-	[1.83 .. 1.87]
M09	1.89	6<->, 10<->, 1<->	3	m	-	[1.83 .. 1.94]
M10	1.98	13<->	1	td	12.10, 6.11	[1.95 .. 2.01]
M11	2.10	16<->	1	br d	17.12	[2.08 .. 2.13]
M12	2.20	11<->, 16<->	2	m	-	[2.18 .. 2.24]
M13	2.30	2<->	1	m	-	[2.28 .. 2.33]
M42	3.24	31<-ax>	1	m	-	[3.23 .. 3.26]
M30	3.27	35<-ax>	1	m	-	[3.26 .. 3.29]
M14	3.32	87<-ax>, 52<-ax>	2	m	-	[3.29 .. 3.34]
M28	3.36	83<-ax>	1	m	-	[3.34 .. 3.38]
M44	3.41	50<-ax>, 40<-ax>	2	m	-	[3.39 .. 3.43]
M32	3.42	22<-ax>, 58<-ax>	3	m	-	[3.38 .. 3.46]
M43	3.45	45<-ax>	1	m	-	[3.44 .. 3.46]
M33	3.47	33<-ax>, 28<-ax>	2	m	-	[3.46 .. 3.48]
M27	3.48	48<-ax>, 85<-ax>	3	m	-	[3.38 .. 3.57]
M31	3.51	72<-ax>	1	m	-	[3.49 .. 3.53]
M36	3.53	54<-ax>	1	m	-	[3.51 .. 3.55]
M34	3.55	80<-ax>	1	m	-	[3.53 .. 3.56]
M26	3.59	69<-ax>	1	ddd	9.72, 5.07, 2.32	[3.57 .. 3.61]
M25	3.66	29<->	1	dd	12.23, 7.09	[3.64 .. 3.68]
M18	3.73	24<-ax>, 70<->, 56<-ax>, 81<->	4	m	-	[3.68 .. 3.79]
M47	3.74	46<->	1	m	-	[3.71 .. 3.77]
M49	3.76	61<->	1	m	-	[3.74 .. 3.78]
M37	3.85	74<-ax>	1	m	-	[3.83 .. 3.87]
M15	3.89	41<->, 76<-ax>	2	m	-	[3.82 .. 3.96]
M40	3.90	23<-ax>, 29<->	2	m	-	[3.85 .. 3.94]
M39	3.90	46<->, 61<->	2	m	-	[3.85 .. 3.94]
M41	3.90	81<->	1	m	-	[3.85 .. 3.94]
M38	3.91	70<->	1	m	-	[3.87 .. 3.94]
M16	4.22	41<->	1	br d	9.54	[4.20 .. 4.24]
M17	4.50	43<-ax>	1	d	7.83	[4.49 .. 4.53]
M29	4.77	38<-ax>	1	m	-	[4.75 .. 4.78]
M19	4.79	19<-ax>	1	d	7.83	[4.78 .. 4.80]
M24	4.79	78<-ax>	1	m	-	[4.78 .. 4.80]
M20	4.88	26<-ax>	1	d	8.07	[4.86 .. 4.90]
M21	4.95	17<-ax>	1	s	-	[4.94 .. 4.97]
M22	5.16	17<-b>	1	br s	-	[5.14 .. 5.18]
M23	5.61	67<-ax>	1	d	7.82	[5.60 .. 5.62]

Steviol+6Glc (isomer 4)

Figure 8W

## Steviol+6Glc (isomer 4)

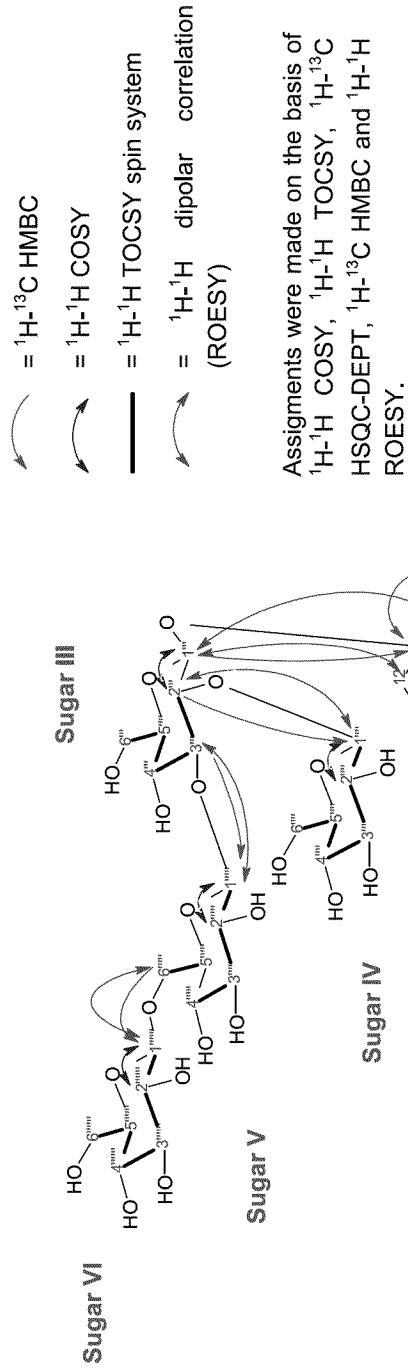


<sup>1</sup>H NMR (800 MHz, *DEUTERIUM OXIDE*)  $\delta$  ppm 0.85 - 0.93 (m, 4 H) 1.04 (br d,  $J=8.07$  Hz, 1 H) 1.08 - 1.14 (m, 1 H) 1.17 (br d,  $J=12.47$  Hz, 1 H) 1.28 (s, 3 H) 1.45 - 1.54 (m, 3 H) 1.54 - 1.61 (m, 2 H) 1.61 - 1.67 (m, 1 H) 1.71 - 1.78 (m, 1 H) 1.83 - 1.87 (m, 1 H) 1.83 - 1.94 (m, 3 H) 1.98 (td,  $J=12.10, 6.11$  Hz, 1 H) 2.10 (br d,  $J=17.12$  Hz, 1 H) 2.18 - 2.24 (m, 2 H) 2.28 - 2.33 (m, 1 H) 3.23 - 3.26 (m, 1 H) 3.26 - 3.29 (m, 1 H) 3.29 - 3.34 (m, 2 H) 3.34 - 3.38 (m, 1 H) 3.38 - 3.57 (m, 3 H) 3.38 - 3.46 (m, 3 H) 3.39 - 3.43 (m, 2 H) 3.44 - 3.46 (m, 1 H) 3.46 - 3.48 (m, 2 H) 3.49 - 3.53 (m, 1 H) 3.51 - 3.55 (m, 1 H) 3.53 - 3.56 (m, 1 H) 3.59 (ddd,  $J=9.72, 5.07, 2.32$  Hz, 1 H) 3.66 (dd,  $J=12.23, 7.09$  Hz, 1 H) 3.68 - 3.79 (m, 4 H) 3.71 - 3.77 (m, 1 H) 3.74 - 3.78 (m, 1 H) 3.82 - 3.96 (m, 2 H) 3.83 - 3.87 (m, 1 H) 3.85 - 3.94 (m, 2 H) 3.85 - 3.94 (m, 2 H) 3.85 - 3.94 (m, 1 H) 3.87 - 3.94 (m, 1 H) 4.22 (br d,  $J=9.54$  Hz, 1 H) 4.50 (d,  $J=7.83$  Hz, 1 H) 4.75 - 4.78 (m, 1 H) 4.79 (d,  $J=7.83$  Hz, 1 H) 4.78 - 4.80 (m, 1 H) 4.88 (d,  $J=8.07$  Hz, 1 H) 4.95 (s, 1 H) 5.16 (br s, 1 H) 5.61 (d,  $J=7.82$  Hz, 1 H)

<sup>13</sup>C NMR (201 MHz, *DEUTERIUM OXIDE*)  $\delta$  ppm 18.8, 21.8, 22.6, 24.2, 31.1, 39.6, 39.6, 42.0, 42.7, 43.7, 44.1, 46.8, 46.8, 49.5, 55.8, 59.5, 63.3, 63.3, 63.7, 64.3, 64.3, 71.3, 71.5, 72.2, 72.2, 72.3, 72.3, 72.5, 72.8, 73.3, 75.9, 76.1, 77.0, 77.5, 78.0, 78.4, 78.4, 78.5, 78.6, 78.7, 78.7, 78.8, 79.0, 79.0, 79.3, 80.8, 81.3, 89.0, 90.5, 95.4, 98.4, 104.8, 105.2, 105.3, 105.3, 107.1, 155.9, 181.1

Figure 8X

Steviol+6Glc (isomer 4)



Numbering following IUPAC

Figure 8Y

Steviol+7Glc (isomer 5)

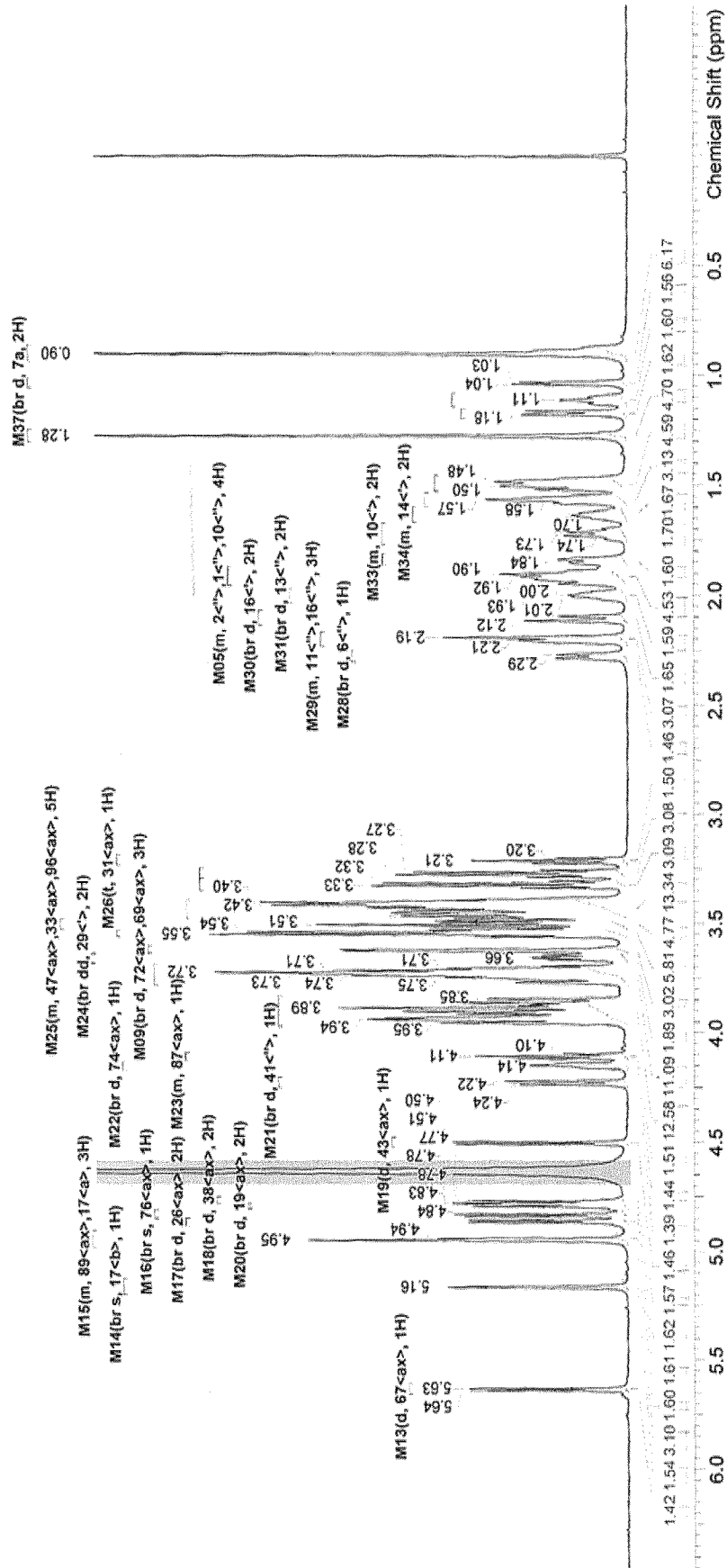
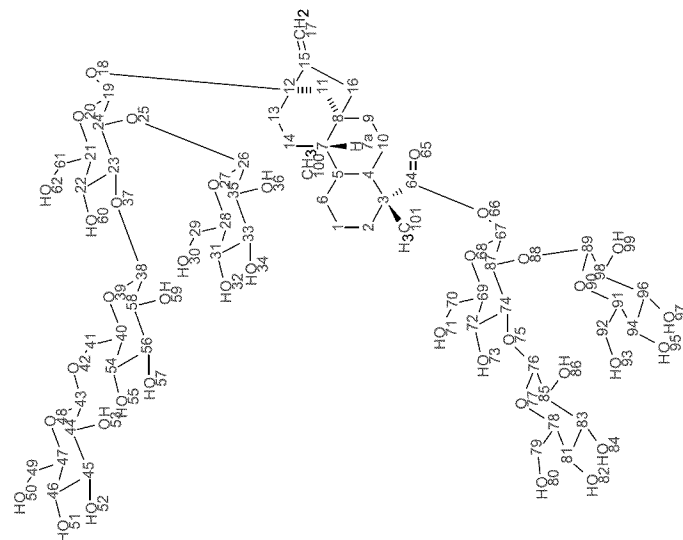


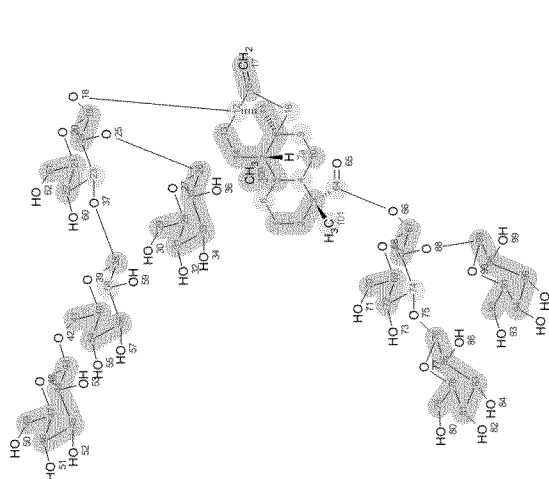
Figure 8Z

Steviol+7Glc (isomer 5)



Multiplet1	Shift1 (ppm)	Atom1	H's	Type	J (Hz)	(ppm)
M01	0.90	100, 6<->	4	s	-	[0.86 .. 0.94]
M37	1.04	7a	1	brd	7.83	[1.01 .. 1.06]
M36	1.11	2<->	1	m	-	[1.08 .. 1.14]
M35	1.17	4	1	brd	12.23	[1.15 .. 1.20]
M02	1.28	101	3	s	-	[1.25 .. 1.31]
M03	1.50	11<->, 1<->, 9<->	3	m	-	[1.46 .. 1.53]
M04	1.57	13<->, 9<->	2	m	-	[1.54 .. 1.60]
M34	1.63	14<->	1	m	-	[1.60 .. 1.67]
M33	1.72	10<->	1	m	-	[1.67 .. 1.77]
M32	1.84	14<->	1	brd	9.29	[1.81 .. 1.86]
M05	1.92	10<->, 1<->, 2<->	3	m	-	[1.87 .. 1.96]
M31	2.00	13<->	1	brd	6.36	[1.97 .. 2.04]
M30	2.11	16<->	1	brd	16.14	[2.07 .. 2.14]
M29	2.20	16<->, 11<->	2	m	-	[2.17 .. 2.23]
M28	2.28	6<->	1	brd	12.72	[2.25 .. 2.30]
M26	3.21	31<-ax>	1	t	9.29	[3.19 .. 3.23]
M06	3.27	98<-ax>, 35<-ax>	2	q	8.31	[3.25 .. 3.30]
M27	3.32	44<-ax>, 94<-ax>	2	m	-	[3.30 .. 3.35]
M07	3.43	46<-ax>, 21<-ax>, 23<-ax>, 58<-ax>, 28<-ax>, 91<-ax>, 78<-ax>, 83<-ax>, 85<-ax>	9	m	-	[3.39 .. 3.48]
M25	3.50	96<-ax>, 33<-ax>, 47<-ax>	3	m	-	[3.48 .. 3.53]
M08	3.54	56<-ax>, 81<-ax>, 54<-ax>, 45<-ax>	4	m	-	[3.53 .. 3.56]
M09	3.62	69<-ax>, 72<-ax>	2	brd	4.40	[3.60 .. 3.64]
M24	3.66	29<->	1	brdd	12.23, 7.34	[3.64 .. 3.68]
M10	3.73	70<->, 40<-ax>, 24<-ax>, 61<->, 79<->, 92<->, 49<->	7	m	-	[3.69 .. 3.78]
M11	3.90	41<->, 49<->, 79<->, 22<-ax>, 70<->, 92<->, 61<->, 29<->	8	m	-	[3.83 .. 3.97]
M23	4.11	87<-ax>	1	m	-	[4.09 .. 4.13]
M22	4.15	74<-ax>	1	brd	3.91	[4.13 .. 4.17]
M21	4.23	41<->	1	brd	10.76	[4.20 .. 4.25]
M19	4.50	43<-ax>	1	d	7.83	[4.48 .. 4.52]
M18	4.77	38<-ax>	1	brd	7.83	[4.75 .. 4.78]
M20	4.79	19<-ax>	1	brd	8.31	[4.78 .. 4.80]
M16	4.81	76<-ax>	1	brs	-	[4.81 .. 4.85]
M17	4.86	26<-ax>	1	brd	7.82	[4.85 .. 4.88]
M15	4.95	17<-a>, 89<-ax>	2	m	-	[4.90 .. 4.98]
M14	5.16	17<-b>	1	brs	-	[5.13 .. 5.20]
M13	5.63	67<-ax>	1	d	7.34	[5.61 .. 5.66]

Figure 8AA



<sup>1</sup>H NMR (800 MHz, DEUTERIUM OXIDE) δ ppm 0.90 (s, 4 H) 1.04 (br d, J=7.83 Hz, 1 H) 1.08 - 1.14 (m, 1 H) 1.17 (br d, J=12.23 Hz, 1 H) 1.28 (s, 3 H) 1.46 - 1.53 (m, 3 H) 1.54 - 1.60 (m, 2 H) 1.60 - 1.67 (m, 1 H) 1.67 - 1.77 (m, 1 H) 1.84 (br d, J=9.29 Hz, 1 H) 1.87 - 1.96 (m, 3 H) 2.00 (br d, J=6.36 Hz, 1 H) 2.11 (br d, J=16.14 Hz, 1 H) 2.17 - 2.23 (m, 2 H) 2.28 (br d, J=12.72 Hz, 1 H) 3.21 (t, J=9.29 Hz, 1 H) 3.27 (q, J=8.31 Hz, 2 H) 3.30 - 3.35 (m, 2 H) 3.39 - 3.48 (m, 9 H) 3.48 - 3.53 (m, 3 H) 3.53 - 3.56 (m, 4 H) 3.62 (br d, J=4.40 Hz, 2 H) 3.66 (br dd, J=12.23, 7.34 Hz, 1 H) 3.69 - 3.78 (m, 7 H) 3.83 - 3.97 (m, 8 H) 4.09 - 4.13 (m, 1 H) 4.15 (br d, J=3.91 Hz, 1 H) 4.23 (br d, J=10.76 Hz, 1 H) 4.50 (d, J=7.83 Hz, 1 H) 4.77 (br d, J=7.83 Hz, 1 H) 4.79 (br d, J=8.31 Hz, 1 H) 4.81 (br s, 1 H) 4.86 (br d, J=7.82 Hz, 1 H) 4.90 - 4.98 (m, 2 H) 5.16 (br s, 1 H) 5.63 (d, J=7.34 Hz, 1 H)

<sup>13</sup>C NMR (201 MHz, DEUTERIUM OXIDE) δ ppm 18.8, 21.7, 22.7, 24.5, 31.0, 39.7, 39.9, 41.9, 42.7, 43.9, 44.1, 46.7, 46.8, 49.4, 55.9, 59.5, 63.5, 63.6, 63.7, 64.5, 64.5, 71.0, 71.5, 71.6, 72.4, 72.5, 72.5, 73.4, 73.4, 76.0, 76.3, 76.8, 77.0, 77.6, 78.0, 78.6, 78.6, 78.8, 78.8, 78.8, 78.8, 78.8, 79.1, 79.1, 79.1, 81.4, 81.4, 87.5, 89.2, 90.7, 95.6, 98.3, 104.5, 104.9, 105.0, 105.2, 105.4, 107.1, 155.7, 181.2

Steviol+7Glc (isomer 5)

F2 Atom	F2 (ppm)	F1 Atom	F1 (ppm)
1<'>	1.9	1	21.7
1<>	1.49		
2<'>	2.28	2	39.7
2<>	1.11		
4	1.17	3	46.8
		4	59.5
		5	41.9
6<'>	1.91	6	42.7
6<>	0.89		
7a	1.04	7	55.9
		8	44.1
9<'>	1.57	9	43.9
9<>	1.48		
10<'>	1.94	10	24.5
10<>	1.72		
11<'>	2.2	11	46.7
11<>	1.51		
		12	90.7
13<'>	2	13	39.9
13<>	1.56		
14<'>	1.84	14	22.7
14<>	1.63		
		15	155.7
16<'>	2.2	16	49.4
16<>	2.11		
17<a>	4.95	17	107.1
17<b>	5.16		
19<a>	4.79	19	98.3
21<a>	3.72	21	77.6

22<ax>	3.54	22	71.5
23<ax>	3.93	23	89.2
24<ax>	3.71	24	81.4
26<ax>	4.87	26	105
28<ax>	3.47	28	78.8
29<'>	3.9	29	64.5
29<>	3.65		
31<ax>	3.21	31	73.4
33<ax>	3.39	33	78
35<ax>	3.27	35	77
38<ax>	4.77	38	105.2
40<ax>	3.47	40	78.8
41<'>	4.23	41	71.6
41<>	3.87		
43<ax>	4.5	43	105.4
44<ax>	3.33	44	76
45<ax>	3.55	45	78.8
46<ax>	3.41	46	72.5
47<ax>	3.5	47	78.6
49<'>	3.92	49	63.6
49<>	3.95		
54<ax>	3.51	54	72.4
56<ax>	3.5	56	78.6
58<ax>	3.42	58	79.1
61<'>	3.73	61	63.7
61<'>	3.85		
64		64	181.2
67<ax>	5.63	67	95.6
69<ax>	3.62	69	79.1
70<'>	3.74	70	63.6
70<>	3.73		

Figure 8AB

Steviol+7Glc (isomer 5)

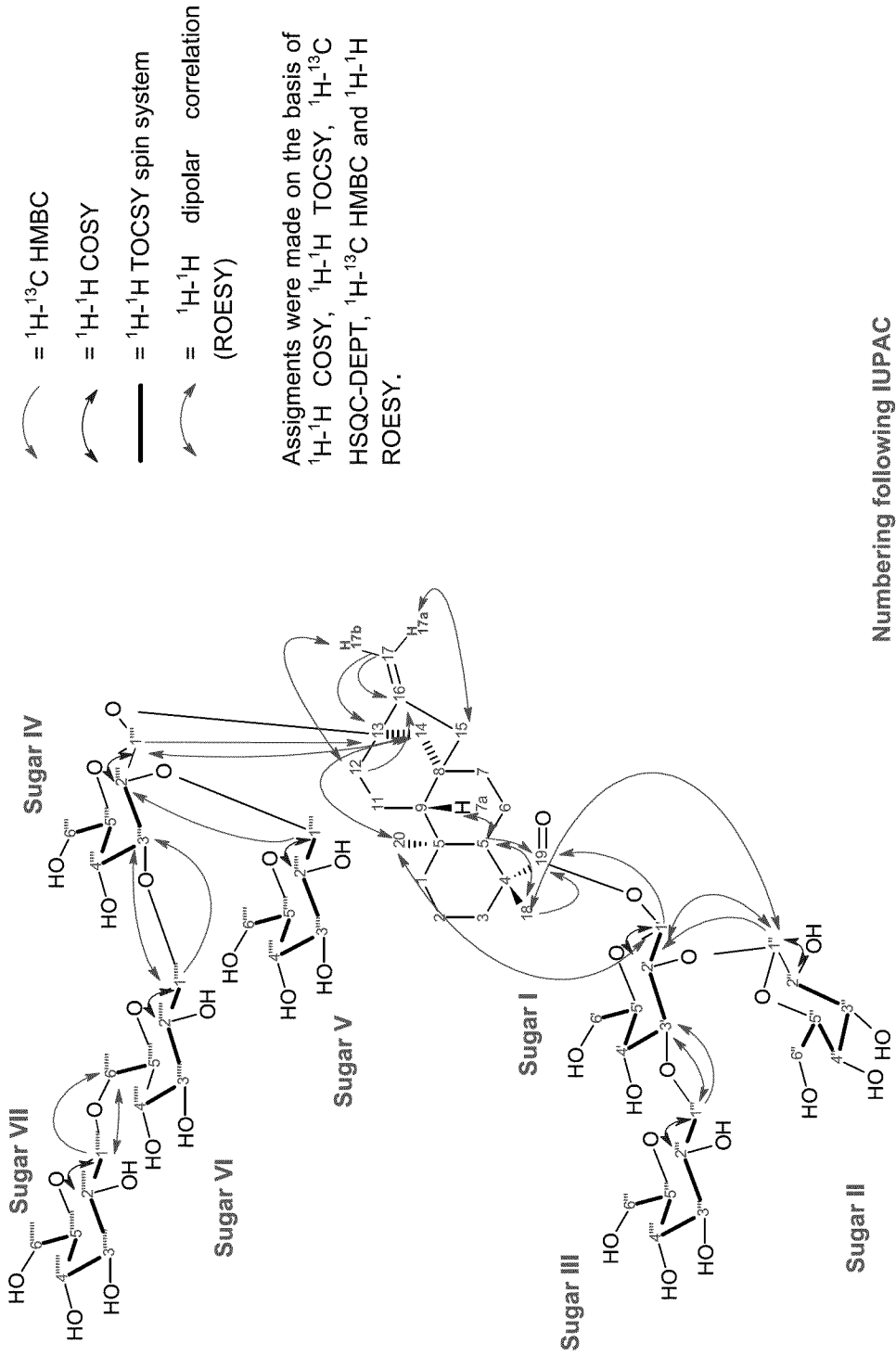


Figure 8AC

Steviol+4Glc+1GlcNAc (#11)

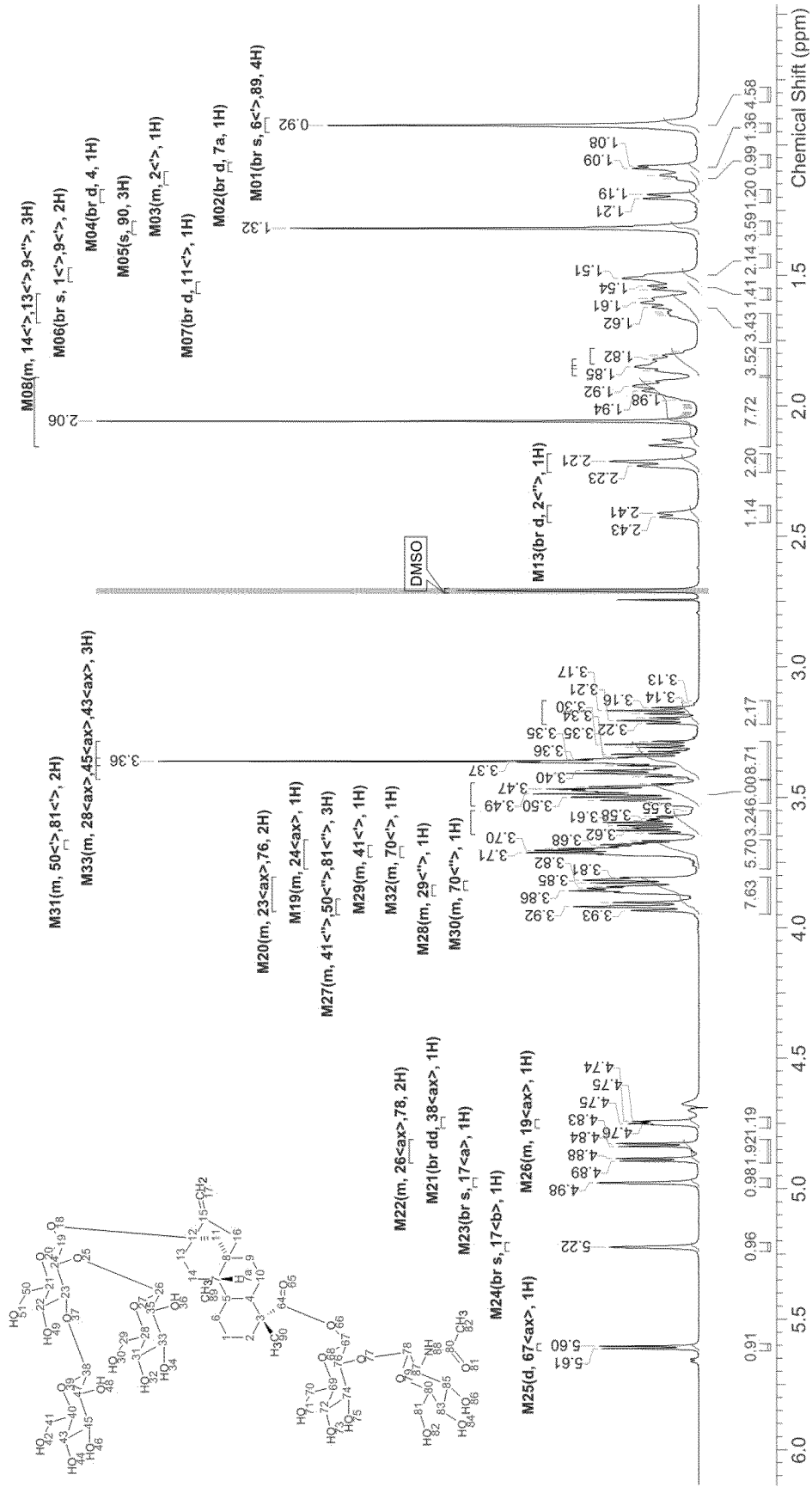
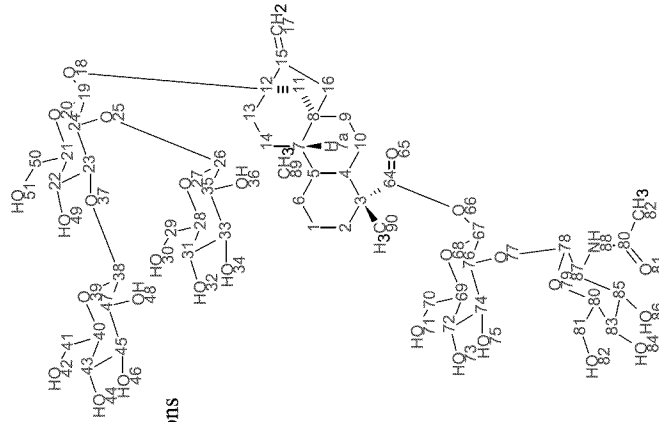


Figure 8AD

Steviol+4Glc+1GlcNAc (#11)

Multiplet1	Shift1 (ppm)	Atom1	H's	Type	J (Hz)	(ppm)
M25	5.61	67<ax>	1	d	7.43	[5.60..5.62]
M24	5.22	17<b>	1	br s	-	[5.21..5.24]
M23	4.98	17<a>	1	br s	-	[4.96..4.99]
M22	4.86	26<ax>, 78	2	m	-	[4.81..4.90]
M26	4.75	19<ax>	1	m	-	[4.73..4.77]
M21	4.75	38<ax>	1	br dd	7.63, 4.11	[4.73..4.77]
M27	3.92	41<*>, 50<*>, 81<*>	3	m	-	[3.89..3.95]
M20	3.86	23<ax>, 76	2	m	-	[3.80..3.94]
M28	3.86	29<*>	1	m	-	[3.84..3.88]
M30	3.84	70<*>	1	m	-	[3.82..3.86]
M32	3.71	70<*>	1	m	-	[3.69..3.73]
M29	3.71	41<*>	1	m	-	[3.68..3.73]
M19	3.70	24<ax>	1	m	-	[3.66..3.77]
M31	3.68	50<*>, 81<*>	2	m	-	[3.67..3.70]
M18	3.61	29<*>, 74, 87<ax>	3	m	-	[3.55..3.64]
M17	3.48	21<ax>, 40<ax>, 85<ax>, 72<ax>, 69<ax>, 22	6	m	-	[3.45..3.53]
M34	3.40	80<ax>, 33<ax>	2	m	-	[3.38..3.42]
M33	3.36	28<ax>, 45<ax>, 43<ax>	3	m	-	[3.35..3.38]
M16	3.36	83<ax>, 47<ax>	2	m	-	[3.29..3.43]
M15	3.19	31<ax>, 35<ax>	2	m	-	[3.13..3.22]
M13	2.42	2<*>	1	br d	12.52	[2.38..2.45]
M12	2.22	11<*>, 16<*>	2	br d	13.69	[2.18..2.25]
M11	2.03	10<*>, 13<*>, 16<*>, 6<*>, 82	7	m	-	[1.89..2.16]
M10	1.86	14<*>	1	br d	10.95	[1.85..1.89]
M35	1.84	1<*>	1	m	-	[1.82..1.86]
M09	1.81	10<*>	1	m	-	[1.78..1.84]
M08	1.62	14<*>, 13<*>, 9<*>	3	m	-	[1.57..1.68]
M07	1.55	11<*>	1	br d	10.95	[1.53..1.57]
M06	1.51	1<*>, 9<*>	2	br s	-	[1.47..1.53]
M05	1.32	90	3	s	-	[1.29..1.34]
M04	1.20	4	1	br d	12.13	[1.17..1.22]
M03	1.12	2<*>	1	m	-	[1.10..1.15]
M02	1.09	7a	1	br d	7.82	[1.07..1.10]
M01	0.92	6<*>, 89	4	br s	-	[0.90..0.95]



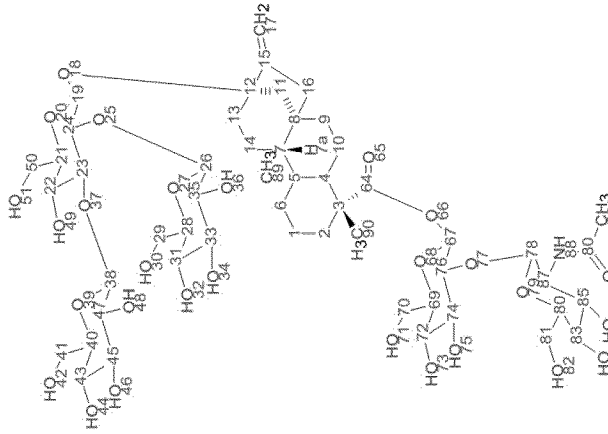
Homonuclear dipolar correlations

between:

- H-38 and H-23
- H-26 and H-24
- H-78 and H-76
- H-67 and Me-90, Me-89
- H-19 and H-11', H-13", H-16"
- H-17a and H-16', H-16"
- H-17b and H-13'
- H-4 and H-7a, Me-90
- Me-89 and H-11"
- H-7a and H-16'

<sup>1</sup>H NMR (800 MHz, *Solvent*) δ ppm 0.92 (br s, 4 H) 1.09 (br d, =7.82 Hz, 1 H) 1.10 - 1.15 (m, 1 H) 1.20 (br d, =12.13 Hz, 1 H) 1.32 (s, 3 H) 1.51 (br s, 2 H) 1.55 (br d, =10.95 Hz, 1 H) 1.57 - 1.68 (m, 3 H) 1.78 - 1.84 (m, 1 H) 1.82 - 1.86 (m, 1 H) 1.86 (br d, =10.95 Hz, 1 H) 1.89 - 2.16 (m, 7 H) 2.22 (br d, =13.69 Hz, 2 H) 2.42 (br d, =12.52 Hz, 1 H) 3.13 - 3.22 (m, 2 H) 3.29 - 3.43 (m, 2 H) 3.35 - 3.38 (m, 3 H) 3.38 - 3.42 (m, 2 H) 3.45 - 3.53 (m, 6 H) 3.55 - 3.64 (m, 3 H) 3.66 - 3.77 (m, 1 H) 3.67 - 3.70 (m, 2 H) 3.68 - 3.73 (m, 1 H) 3.69 - 3.73 (m, 1 H) 3.80 - 3.94 (m, 2 H) 3.82 - 3.86 (m, 1 H) 3.84 - 3.88 (m, 1 H) 3.89 - 3.95 (m, 3 H) 4.75 (br dd, =7.63, 4.11 Hz, 1 H) 4.73 - 4.77 (m, 1 H) 4.81 - 4.90 (m, 2 H) 4.98 (br s, 1 H) 5.22 (br s, 1 H) 5.61 (d, =7.43 Hz, 1 H)

Figure 8AE



Heteronuclear scalar correlations between:

- C=O(64) and H-67, Me-90, H-4
- C-12 and H-19, H-17a/b
- H-26 and C-24 (C-26 and H-24)
- H-38 and C-23 (C-38 and H-23)
- H-78 and C-76 (C-78 and H-76)
- C=O(80) and H-87, Me-82

<sup>13</sup>C NMR (201 MHz, Solvent)  $\delta$  ppm 178.1 (1C), 105.7 (1C), 103.4 (1C), 103.0 (1C), 101.7 (1C), 96.7 (1C), 93.2 (1C), 88.4 (1C), 86.7 (1C), 79.5 (1C), 78.4 (1C), 77.7 (1C), 77.7 (1C), 77.6 (1C), 77.2 (1C), 77.2 (1C), 77.2 (1C), 76.6 (1C), 75.4 (1C), 75.2 (1C), 74.6 (1C), 71.6 (1C), 71.6 (1C), 70.8 (1C), 70.5 (1C), 70.5 (1C), 69.3 (1C), 62.6 (1C), 62.6 (1C), 62.4 (1C), 61.8 (1C), 61.8 (1C), 61.7 (1C), 57.6 (1C), 57.1 (1C), 53.9 (1C), 47.9 (1C), 45.1 (1C), 45.0 (1C), 42.5 (1C), 42.0 (1C), 41.1 (1C), 40.1 (1C), 38.1 (1C), 37.6 (1C), 29.7 (1C), 23.7 (1C), 22.4 (1C), 21.0 (1C), 20.2 (1C), 17.3 (1C),

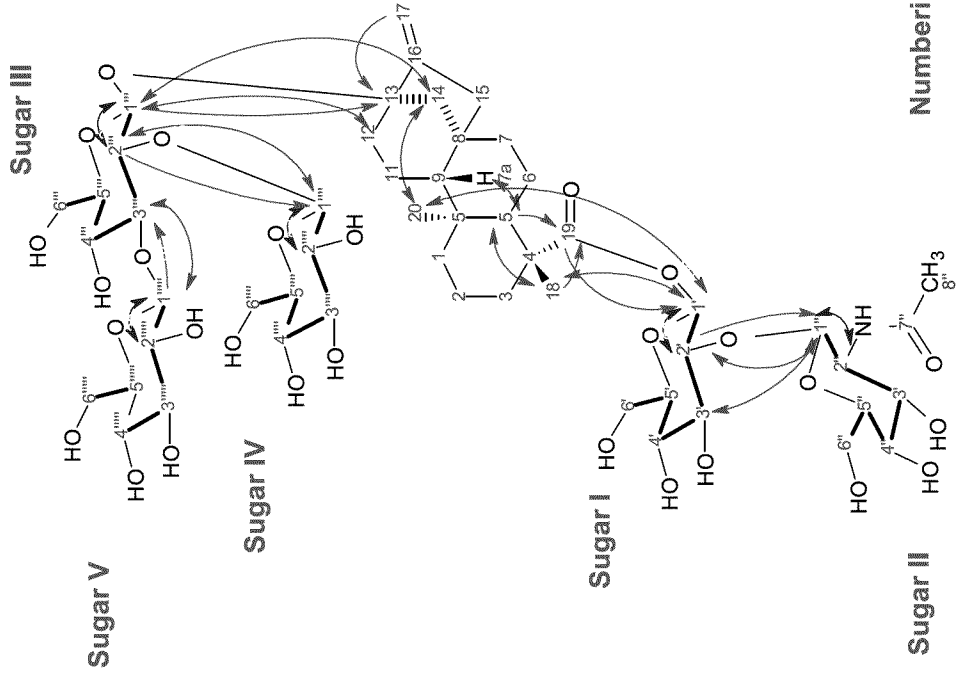
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24<ax>	3.72	24	79.5
26<ax>	4.89	26	103
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29<''>	3.86	29	62.6
29<'>	3.58	29	62.6
31<ax>	3.17	31	71.6
33<ax>	3.41	33	77.2
35<ax>	3.2	35	75.4
38<ax>	4.75	38	103.4
40<ax>	3.47	40	77.2
41<''>	3.93	41	62.4
41<'>	3.71	41	62.4
43<ax>	3.37	43	70.8
45<ax>	3.36	45	76.6
47<ax>	3.33	47	74.6
50<''>	3.92	50	61.8
50<'>	3.69		
67<ax>	5.61	64	178.1
69<ax>	3.52	67	93.2
70<''>	3.84	69	77.7
70<'>	3.71	70	61.7
72<ax>	3.46	72	70.5
74	3.63	74	77.6
76	3.82	76	78.4
78	4.83	78	101.7
80<ax>	3.41	80	77.2
81<''>	3.92	81	61.8
81<'>	3.69		
82	2.06	82	23.7

Steviol+4Glc+1GlcNAc (#11)

F2Atom	$\delta$ (1H) (ppm)	F1Atom	$\delta$ (13C) (ppm)
1<'>	1.84	1	20.2
1<'>	1.51		
2<'>	2.42	2	37.6
2<'>	1.12		
4	1.2	3	45.1
		4	57.6
		5	40.1
6<'>	1.91	6	41.1
6<'>	0.92		
7a	1.08	7	53.9
		8	42.5
9<'>	1.62	9	42
9<'>	1.51		
10<'>	1.93	10	22.4
10<'>	1.81		
11<'>	2.22	11	45
11<'>	1.55		
		12	88.4
13<'>	1.96	13	38.1
13<'>	1.59		
14<'>	1.86	14	21
14<'>	1.65		
16<'>	2.22	16	47.9
16<'>	2.14		
17<a>	4.98	17	105.7
17<b>	5.22		
19<ax>	4.75	19	96.7
21<ax>	3.5	21	75.2

Figure 8AF

**Steviol+4Glc+1GlcNAc (#11)**



Numbering following IUPAC

- =  $^1\text{H}-^{13}\text{C}$  HMBC
- =  $^1\text{H}-^1\text{H}$  COSY
- =  $^1\text{H}-^1\text{H}$  TOCSY spin system
- =  $^1\text{H}-^1\text{H}$  dipolar correlation (ROESY)

Assignments were made on the basis of  $^1\text{H}-^1\text{H}$  COSY,  $^1\text{H}-^1\text{H}$  TOCSY,  $^1\text{H}-^{13}\text{C}$  HSQC-DEPT,  $^1\text{H}-^{13}\text{C}$  HMBC and  $^1\text{H}-^1\text{H}$  ROESY.

Figure 8AG

**Steviol+4Glc (#26)**

Homonuclear dipolar correlations between:

- H-2 and Me-53, Me-54
- H-11 and H-3, H-18
- H-18 and H-4
- H-68 and H-28, Me-53, H-38'
- H-28 and H-38', Me-53
- Me-54 and H-32''

Heteronuclear scalar correlations between:

- C=O(-8) and H-2, Me-53, H-28
- H-11 and C-3 (H-3 and C-11)
- H-18 and C-4 (H-4 and C18)
- H-56 and C-33

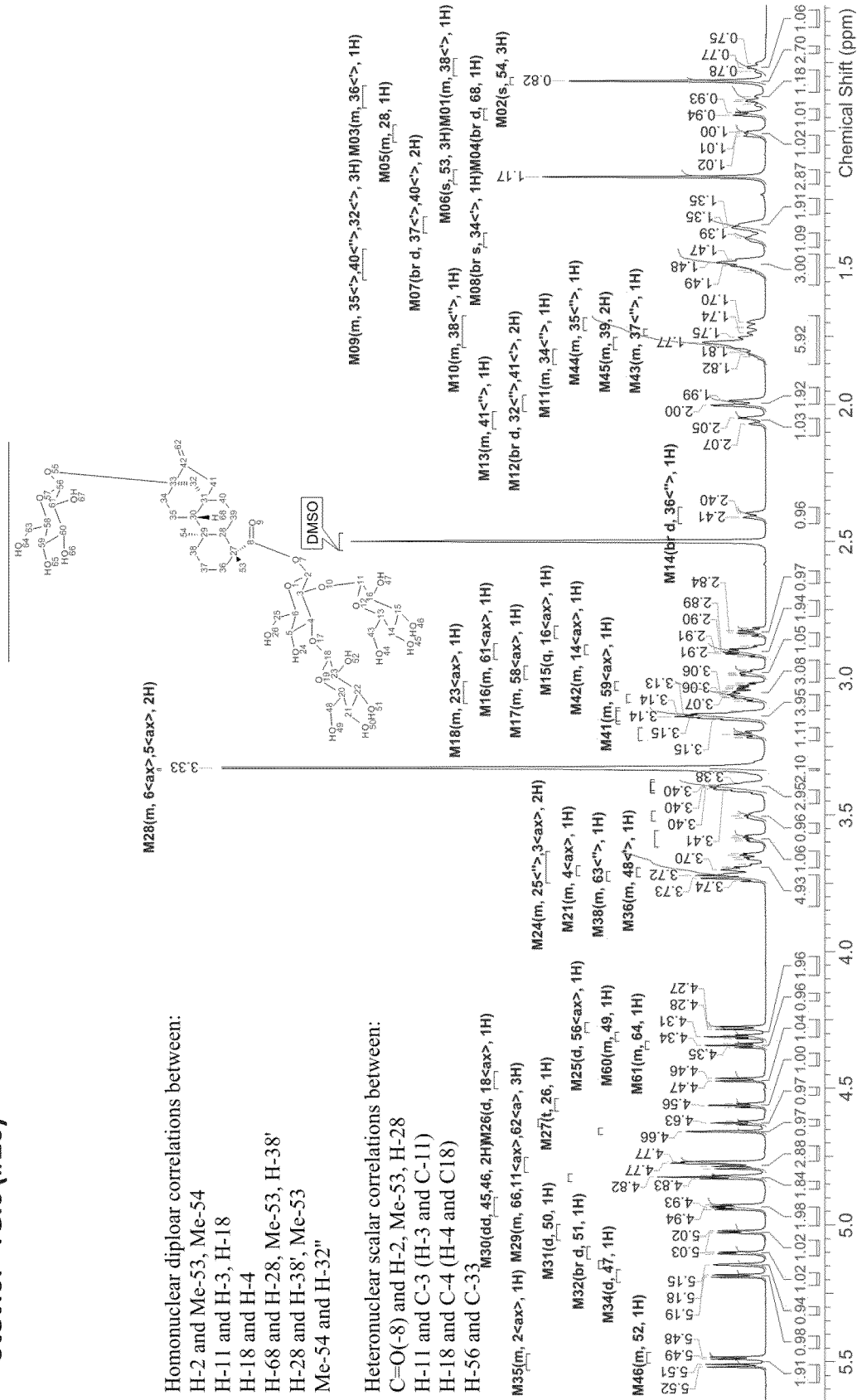
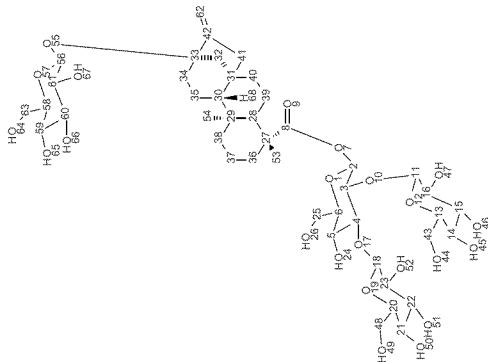


Figure 8AH

Steviol+4Glc (#26)



<sup>1</sup>H NMR (800 MHz, DMSO-d<sub>6</sub>) δ ppm 0.73 - 0.80 (m, 1 H) 0.82 (s, 3 H) 0.83 - 0.91 (m, 1 H) 0.94 (br d, J=8.07 Hz, 1 H) 0.98 - 1.04 (m, 1 H) 1.17 (s, 3 H) 1.35 (br d, J=4.40 Hz, 2 H) 1.39 (br s, 1 H) 1.43 - 1.54 (m, 3 H) 1.68 - 1.79 (m, 1 H) 1.68 - 1.73 (m, 1 H) 1.72 - 1.75 (m, 1 H) 1.75 - 1.80 (m, 2 H) 1.80 - 1.85 (m, 1 H) 1.99 (br d, J=13.45 Hz, 2 H) 2.03 - 2.09 (m, 1 H) 2.40 (br d, J=12.47 Hz, 1 H) 2.83 (q, J=7.99 Hz, 1 H) 2.87 - 2.93 (m, 1 H) 2.88 - 2.91 (m, 1 H) 2.96 - 3.00 (m, 1 H) 3.01 - 3.09 (m, 1 H) 3.01 - 3.05 (m, 1 H) 3.06 - 3.09 (m, 1 H) 3.11 - 3.17 (m, 3 H) 3.12 - 3.16 (m, 1 H) 3.18 - 3.23 (m, 1 H) 3.33 - 3.34 (m, 2 H) 3.37 - 3.41 (m, 1 H) 3.38 - 3.42 (m, 1 H) 3.38 - 3.42 (m, 1 H) 3.49 - 3.52 (m, 1 H) 3.59 (br dd, J=9.90, 5.50 Hz, 1 H) 3.63 - 3.75 (m, 2 H) 3.69 - 3.72 (m, 1 H) 3.69 - 3.73 (m, 1 H) 3.71 - 3.74 (m, 1 H) 4.28 (d, J=7.82 Hz, 1 H) 4.30 - 4.33 (m, 1 H) 4.33 - 4.36 (m, 1 H) 4.47 (d, J=7.83 Hz, 1 H) 4.56 (t, J=5.50 Hz, 1 H) 4.61 - 4.64 (m, 1 H) 4.65 - 4.67 (m, 1 H) 4.75 - 4.81 (m, 3 H) 4.81 - 4.84 (m, 2 H) 4.93 (dd, J=8.80, 4.89 Hz, 2 H) 5.03 (d, J=5.38 Hz, 1 H) 5.10 (br d, J=4.89 Hz, 1 H) 5.15 (br s, 1 H) 5.19 (d, J=6.60 Hz, 1 H) 5.47 - 5.50 (m, 1 H) 5.47 - 5.53 (m, 1 H)

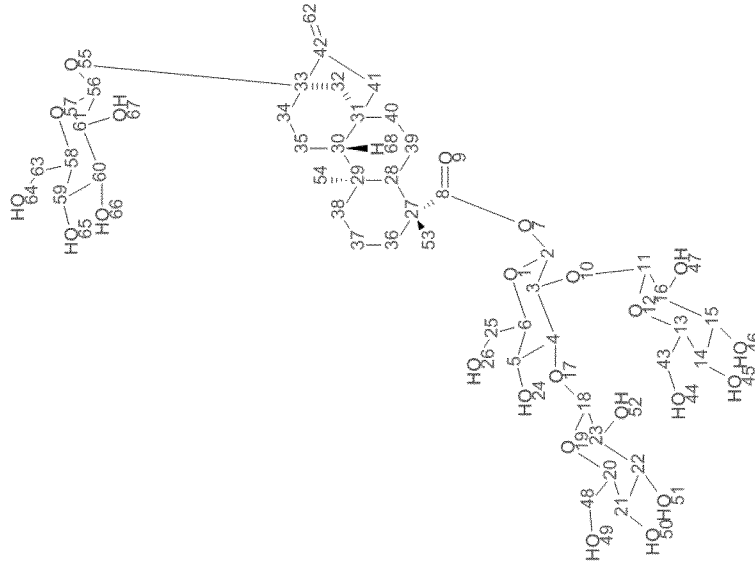
Multiplet1	Shift1 (ppm)	Atom1	H's	Type	J (Hz)	(ppm)
M35	5.50	2<ax>	1	m	-	[5.47 .. 5.53]
M46	5.49	52	1	m	-	[5.47 .. 5.50]
M34	5.19	47	1	d	6.60	[5.16 .. 5.21]
M33	5.15	62<b>	1	br s	-	[5.13 .. 5.16]
M32	5.10	51	1	br d	4.89	[5.08 .. 5.13]
M31	5.03	50	1	d	5.38	[5.00 .. 5.06]
M30	4.93	45, 46	2	dd	8.80, 4.89	[4.90 .. 4.97]
M58	4.83	65, 67	2	m	-	[4.81 .. 4.84]
M29	4.78	66, 11<ax>, 62<a>	3	m	-	[4.75 .. 4.81]
M59	4.66	24	1	m	-	[4.65 .. 4.67]
M62	4.63	44	1	m	-	[4.61 .. 4.64]
M27	4.56	26	1	t	5.50	[4.54 .. 4.58]
M26	4.47	18<ax>	1	d	7.83	[4.44 .. 4.50]
M61	4.34	64	1	m	-	[4.33 .. 4.36]
M60	4.31	49	1	m	-	[4.30 .. 4.33]
M25	4.28	56<ax>	1	d	7.82	[4.26 .. 4.30]
M38	3.73	63<*>	1	m	-	[3.71 .. 3.74]
M36	3.71	48<*>	1	m	-	[3.69 .. 3.73]
M21	3.71	4<ax>	1	m	-	[3.69 .. 3.72]
M24	3.70	25<*>, 3<ax>	2	m	-	[3.63 .. 3.75]
M23	3.59	43<*>	1	br dd	9.90, 5.50	[3.56 .. 3.62]
M22	3.50	25<*>	1	m	-	[3.49 .. 3.52]
M47	3.40	43<*>	1	m	-	[3.38 .. 3.42]
M37	3.40	48<*>	1	m	-	[3.38 .. 3.42]
M39	3.39	63<*>	1	m	-	[3.37 .. 3.41]
M28	3.33	6<ax>, 5<ax>	2	m	-	[3.33 .. 3.34]
M20	3.21	20<ax>	1	m	-	[3.18 .. 3.23]
M19	3.14	60<ax>, 15<ax>, 22<ax>	3	m	-	[3.11 .. 3.17]
M48	3.14	13<ax>	1	m	-	[3.12 .. 3.16]
M40	3.07	21<ax>	1	m	-	[3.06 .. 3.09]
M18	3.05	23<ax>	1	m	-	[3.01 .. 3.09]
M41	3.03	59<ax>	1	m	-	[3.01 .. 3.05]
M17	2.98	58<ax>	1	m	-	[2.96 .. 3.00]
M16	2.90	61<ax>	1	m	-	[2.87 .. 2.93]
M42	2.90	14<ax>	1	m	-	[2.88 .. 2.91]
M15	2.83	16<ax>	1	q	7.99	[2.81 .. 2.85]
M14	2.40	36<*>	1	br d	12.47	[2.38 .. 2.43]
M13	2.06	41<*>	1	m	-	[2.03 .. 2.09]
M12	1.99	32<*>, 41<*>	2	br d	13.45	[1.96 .. 2.03]
M11	1.82	34<*>	1	m	-	[1.80 .. 1.85]
M45	1.78	39	2	m	-	[1.75 .. 1.80]
M10	1.75	38<*>	1	m	-	[1.68 .. 1.79]
M43	1.74	37<*>	1	m	-	[1.72 .. 1.75]
M44	1.71	35<*>	1	m	-	[1.68 .. 1.73]
M09	1.49	35<*>, 40<*>, 32<*>	3	m	-	[1.43 .. 1.54]
M08	1.39	34<*>	1	br s	-	[1.37 .. 1.42]
M07	1.35	37<*>, 40<*>	2	br d	4.40	[1.31 .. 1.37]
M06	1.17	53	3	s	-	[1.14 .. 1.19]
M05	1.01	28	1	m	-	[0.98 .. 1.04]
M04	0.94	68	1	br d	8.07	[0.92 .. 0.96]
M03	0.89	36<*>	1	m	-	[0.83 .. 0.91]
M02	0.82	54	3	s	-	[0.80 .. 0.83]
M01	0.77	38<*>	1	m	-	[0.73 .. 0.80]

Figure 8AI

**Steviol+4Glc (#26)**

F2 Atom	$\delta(1H)$ (ppm)	F1 Atom	$\delta(13C)$ (ppm)
2<ax>	5.52	2	91.9
3<ax>	3.73	3	75.5
4<ax>	3.69	4	87
5<ax>	3.34	5	68.3
6<ax>	3.33	6	77.8
		8	174.9
11<ax>	4.79	11	101.8
13<ax>	3.14	13	77.3
14<ax>	2.89	14	71.4
15<ax>	3.14	15	77
16<ax>	2.83	16	74.3
18<ax>	4.47	18	103.3
20<ax>	3.21	20	77.3
21<ax>	3.07	21	70.3
22<ax>	3.14	22	77
23<ax>	3.06	23	74
25<''>	3.65	25	60.7
25<<>	3.5		
27		27	43.6
28	1.01	28	56.7
29		29	39.1
31		31	41.6
32<'>	2	32	43.6
32<<>	1.48		
33		33	85.8
34<'>	1.82	34	37.9
34<<>	1.39		
35<'>	1.7	35	20.1





35<'>	1.5	36	36.6
36<'>	2.4	37	19.5
36<<>	0.89		
37<'>	1.74	38	40.2
37<<>	1.35		
38<'>	1.78		
38<<>	0.77		
39	1.78	39	21.4
40<'>	1.49	40	41.3
40<<>	1.35		
41<'>	2.06	41	47.8
41<<>	2		
42		42	152.6
43<'>	3.59	43	61.4
43<<>	3.39		
48<'>	3.4	48	61.3
48<<>	3.71		
53	1.17	53	28.5
54	0.82	54	16.4
56<ax>	4.28	56	98.1
58<ax>	2.98	58	76.9
59<ax>	3.03	59	70.5
60<ax>	3.14	60	77
61<ax>	2.91	61	74
62<a>	4.77	62	104.8
62<b>	5.15		
63<'>	3.73	63	62
63<<>	3.39		
68	0.94	30	53.4



<sup>13</sup>C NMR (201 MHz, DMSO d<sub>6</sub>)  $\delta$  ppm 174.9 (1C), 152.6 (1C), 104.8 (1C), 103.3 (1C), 101.8 (1C), 98.1 (1C), 91.9 (1C), 87.0 (1C), 85.8 (1C), 77.8 (1C), 77.3 (2C), 77.0 (3C), 76.9 (1C), 75.5 (1C), 74.3 (1C), 74.0 (2C), 71.4 (1C), 70.5 (1C), 70.3 (1C), 68.3 (1C), 62.0 (1C), 61.4 (1C), 61.3 (1C), 60.7 (1C), 56.7 (1C), 53.4 (1C), 47.8 (1C), 43.6 (2C), 41.6 (1C), 41.3 (1C), 40.2 (1C), 39.1 (1C), 37.9 (1C), 36.6 (1C), 28.5 (1C), 21.4 (1C), 20.1 (1C), 19.5 (1C), 16.4 (1C)

Figure 8AJ

Steviol+4Glc (#26)

-  = <sup>1</sup>H-<sup>13</sup>C HMBC
-  = <sup>1</sup>H-<sup>1</sup>H COSY
-  = <sup>1</sup>H-<sup>1</sup>H TOCSY spin system
-  = <sup>1</sup>H-<sup>1</sup>H dipolar correlation (ROESY)

Assignments were made on the basis of <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>1</sup>H TOCSY, <sup>1</sup>H-<sup>13</sup>C HSQC-DEPT, <sup>1</sup>H-<sup>13</sup>C HMBC and <sup>1</sup>H-<sup>1</sup>H ROESY.

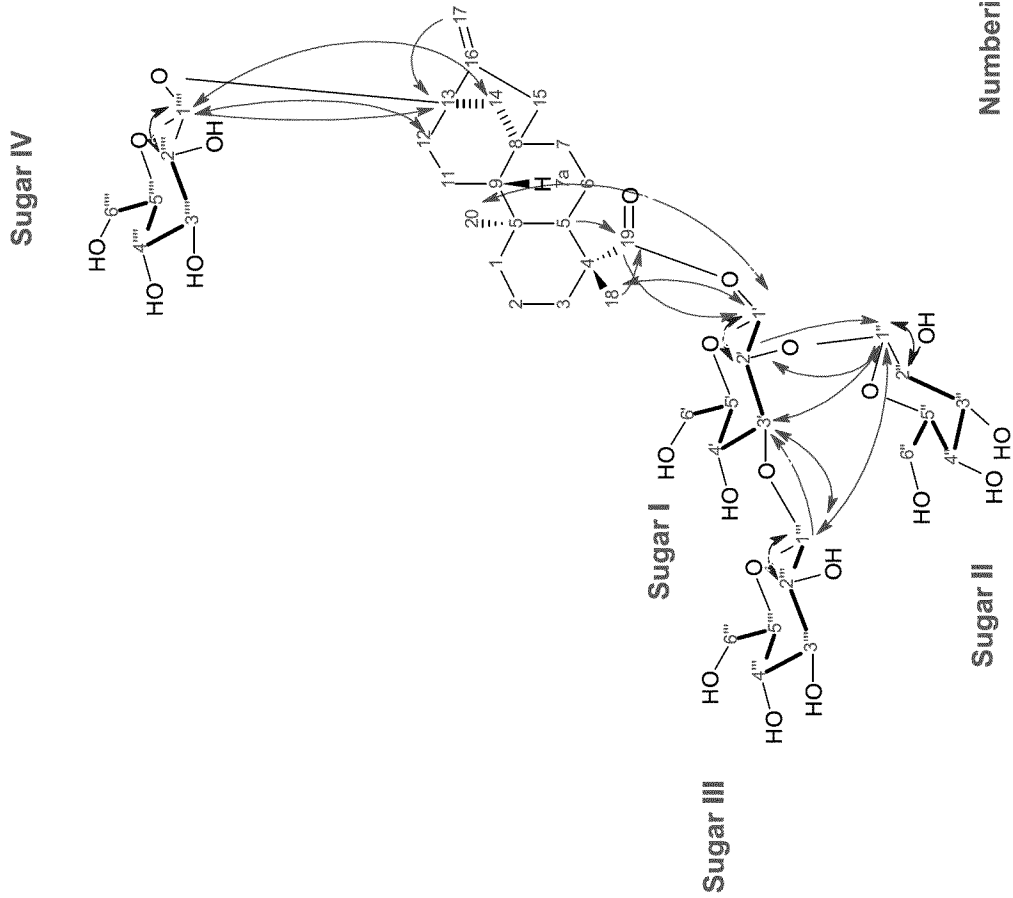


Figure 8AK

Steviol+5Glc (#22)

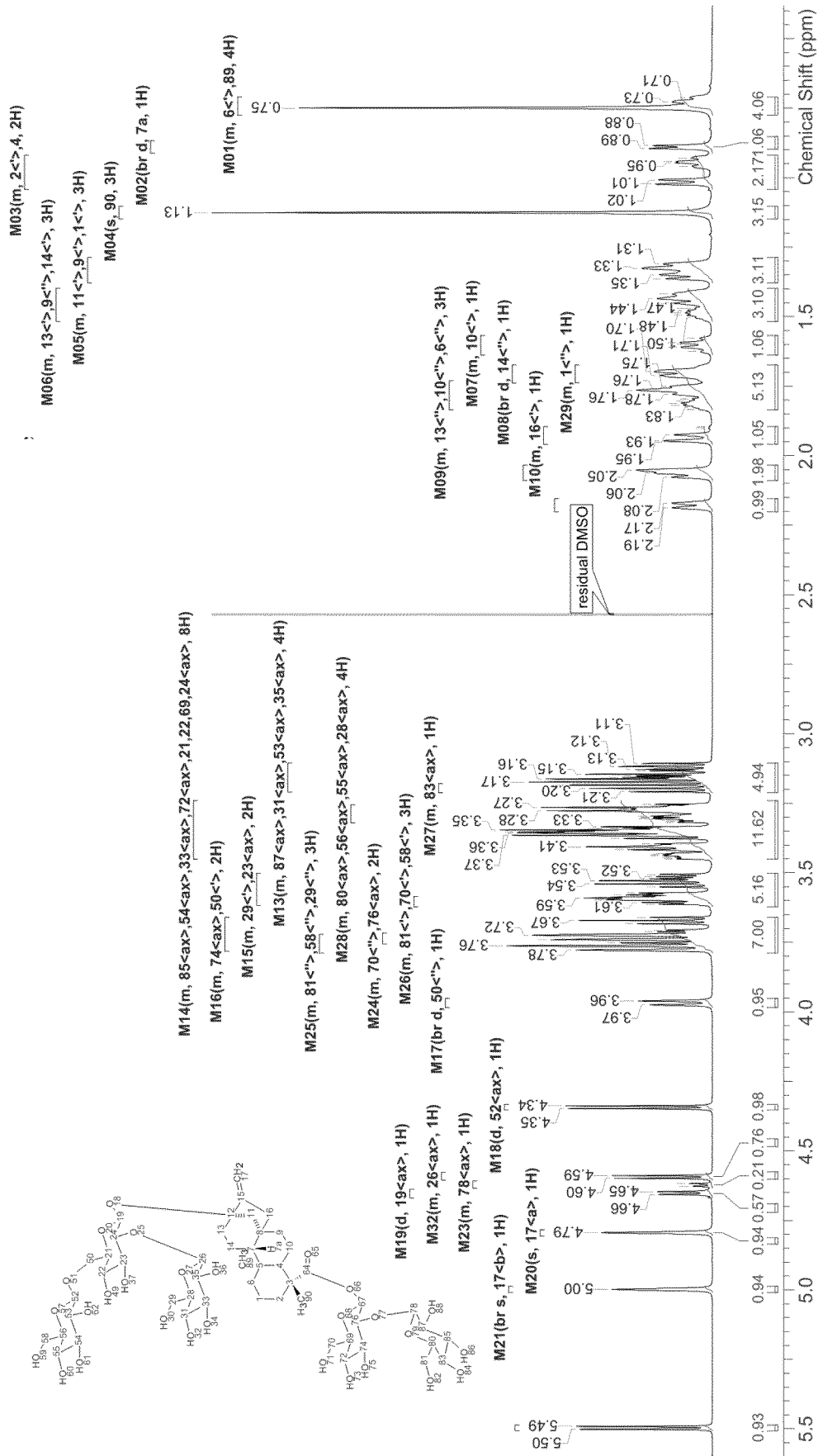
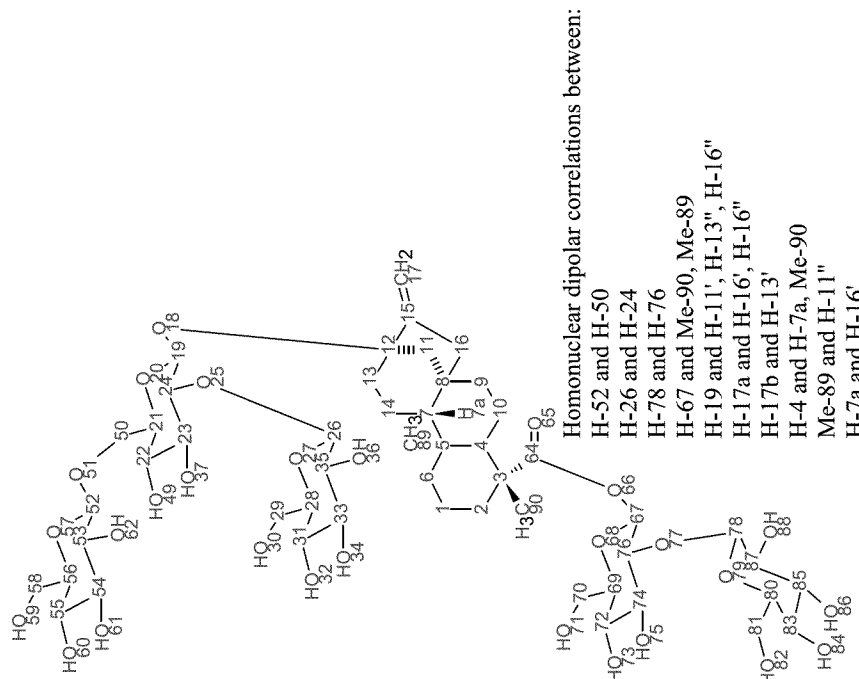


Figure 8AL

Steviol+5Glc (#22)

Multiplet1	Shift1 (ppm)	Atom1	H's	Type	J (Hz)	(ppm)
M22	5.50	67<ax>	1	d	7.82	[5.49 .. 5.51]
M21	5.00	17<b>	1	br s	-	[4.99 .. 5.01]
M20	4.79	17<a>	1	s	-	[4.78 .. 4.81]
M19	4.65	19<ax>	1	d	7.82	[4.64 .. 4.67]
M23	4.62	78<ax>	1	m	-	[4.61 .. 4.64]
M32	4.59	26<ax>	1	m	-	[4.58 .. 4.61]
M18	4.34	52<ax>	1	d	8.07	[4.33 .. 4.35]
M17	3.97	50<f>	1	br d	10.51	[3.95 .. 3.98]
M25	3.76	81<f>, 58<f>, 29<f>	3	m	-	[3.72 .. 3.79]
M24	3.74	70<f>, 76<ax>	2	m	-	[3.72 .. 3.76]
M16	3.73	74<ax>, 50<f>	2	m	-	[3.66 .. 3.78]
M26	3.60	81<f>, 70<f>, 58<f>	3	m	-	[3.58 .. 3.62]
M15	3.57	29<f>, 23<ax>	2	m	-	[3.50 .. 3.62]
M14	3.34	85<ax>, 54<ax>, 33<ax>, 72<ax>, 21, 22, 69, 24<ax>	8	m	-	[3.24 .. 3.45]
M28	3.29	80<ax>, 56<ax>, 55<ax>, 28<ax>	4	m	-	[3.26 .. 3.32]
M27	3.20	83<ax>	1	m	-	[3.18 .. 3.21]
M23	3.16	87<ax>, 31<ax>, 53<ax>, 35<ax>	4	m	-	[3.11 .. 3.21]
M12	2.18	2<f>	1	br d	13.20	[2.16 .. 2.20]
M11	2.06	11<f>, 16<f>	2	m	-	[2.03 .. 2.09]
M10	1.93	16<f>	1	m	-	[1.90 .. 1.96]
M09	1.78	13<f>, 10<f>, 6<f>	3	m	-	[1.73 .. 1.84]
M29	1.71	1<f>	1	m	-	[1.67 .. 1.74]
M08	1.70	14<f>	1	br d	10.51	[1.67 .. 1.74]
M07	1.60	10<f>	1	m	-	[1.57 .. 1.64]
M06	1.46	13<f>, 9<f>, 14<f>	3	m	-	[1.40 .. 1.52]
M05	1.34	11<f>, 9<f>, 1<f>	3	m	-	[1.29 .. 1.38]
M04	1.13	90	3	s	-	[1.10 .. 1.15]
M03	0.98	2<f>, 4	2	m	-	[0.92 .. 1.04]
M02	0.89	7a	1	br d	7.83	[0.87 .. 0.91]
M01	0.75	6<f>, 89	4	m	-	[0.71 .. 0.77]



Homonuclear dipolar correlations between:  
 H-52 and H-50  
 H-26 and H-24  
 H-78 and H-76  
 H-67 and Me-90, Me-89  
 H-19 and H-11', H-13", H-16"  
 H-17a and H-16', H-16"  
 H-17b and H-13'  
 H-4 and H-7a, Me-90  
 Me-89 and H-11"  
 H-7a and H-16'

<sup>1</sup>H NMR (800 MHz, DEUTERIUM OXIDE) δ ppm 0.71 - 0.77 (m, 4 H) 0.89 (br d, J=7.83 Hz, 1 H) 0.92 - 1.04 (m, 2 H) 1.13 (s, 3 H) 1.29 - 1.38 (m, 3 H) 1.40 - 1.52 (m, 3 H) 1.57 - 1.64 (m, 1 H) 1.70 (br d, J=10.51 Hz, 1 H) 1.67 - 1.74 (m, 1 H) 1.73 - 1.84 (m, 3 H) 1.90 - 1.96 (m, 1 H) 2.03 - 2.09 (m, 2 H) 2.18 (br d, J=13.20 Hz, 1 H) 3.11 - 3.21 (m, 4 H) 3.18 - 3.21 (m, 1 H) 3.24 - 3.45 (m, 8 H) 3.26 - 3.32 (m, 4 H) 3.50 - 3.62 (m, 2 H) 3.58 - 3.62 (m, 3 H) 3.66 - 3.78 (m, 2 H) 3.72 - 3.76 (m, 2 H) 3.72 - 3.79 (m, 3 H) 3.97 (br d, J=10.51 Hz, 1 H) 4.34 (d, J=8.07 Hz, 1 H) 4.58 - 4.61 (m, 1 H) 4.61 - 4.64 (m, 1 H) 4.65 (d, J=7.82 Hz, 1 H) 4.79 (s, 1 H) 5.00 (br s, 1 H) 5.50 (br d, J=7.82 Hz, 1 H)

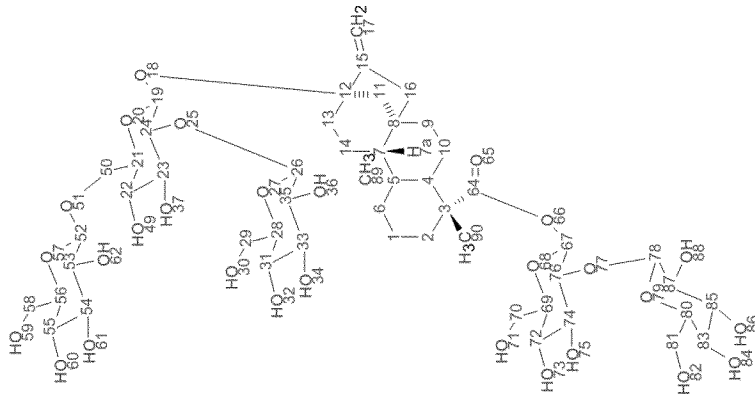


Figure 8AM

**Steviol+5Glc (#22)**

F2 Atom	$\delta(1H)$ (ppm)	F1 Atom	$\delta(13C)$ (ppm)	J2	22	69
1<>	1.71	1	18.9	23<ax>	23	76
1<>	1.32			24<ax>	24	74.5
2<>	2.18	2	36.6	26<ax>	26	102.7
2<>	0.95			28<ax>	28	76.1
		3	44	29<'>	29	61
4	1.02	4	56.6	29<>	29	61
		5	39	31<ax>	31	69.9
6<'>	1.76	6	39.8	33<ax>	33	75.3
6<>	0.73			35<ax>	35	74.1
7a	0.89	7	52.9	50<'>	50	68.2
9<'>	1.44	9	40.7	50<>		
9<>	1.33			52<ax>	52	102.5
10<'>	1.77	10	21.2	53<ax>	53	72.9
10<>	1.6			54<ax>	54	75.3
11<'>	2.06	11	44	55<ax>	55	69.4
11<>	1.36			56<ax>	56	75.8
		12	87.4	58<'>	58	60.7
13<'>	1.81	13	36.6	58<>		
13<>	1.42			64	64	178
14<'>	1.71	14	19.9	67<ax>	67	92.2
14<>	1.49			69	69	76.4
		15	153.1	70<'>	70	60.4
16<'>	2.07	16	46.8	70<>		
16<>	1.93			72<ax>	72	69
17<a>	4.79	17	104.3	74<ax>	74	75.8
17<b>	5			76<ax>	76	78
19<ax>	4.65	19	95.5	78<ax>	78	102.4
21	3.41	21	74.5	80<ax>	80	76.1
				81<'>	81	60.7
				81<>		
				83<ax>	83	69.7

Heteronuclear scalar correlations between:

- C=O(64) and H-67, Me-90, H-4
- C-12 and H-19, H-17a/b
- H-26 and C-24 (C-26 and H-24)
- H-52 and C-50 (C-52 and H-50)
- H-78 and C-76 (C-78 and H-76)

$^{13}C$  NMR (201 MHz, DEUTERIUM OXIDE)  $\delta$  ppm 178.0 (1C), 153.1 (1C), 104.3 (1C), 102.7 (1C), 102.5 (1C), 102.4 (1C), 95.5 (1C), 92.2 (1C), 87.4 (1C), 78.0 (1C), 76.4 (1C), 76.1 (2C), 76.0 (1C), 75.8 (2C), 75.3 (3C), 74.5 (2C), 74.1 (1C), 73.8 (1C), 72.9 (1C), 69.9 (1C), 69.7 (1C), 69.4 (1C), 69.0 (2C), 68.2 (1C), 61.0 (2C), 60.7 (2C), 60.4 (1C), 56.6 (1C), 52.9 (1C), 46.8 (1C), 44.0 (2C), 40.7 (1C), 39.8 (1C), 39.0 (1C), 36.6 (1C), 36.6 (1C), 28.4 (1C), 21.2 (1C), 19.9 (1C), 18.9 (1C), 15.8 (1C)

Figure 8AN

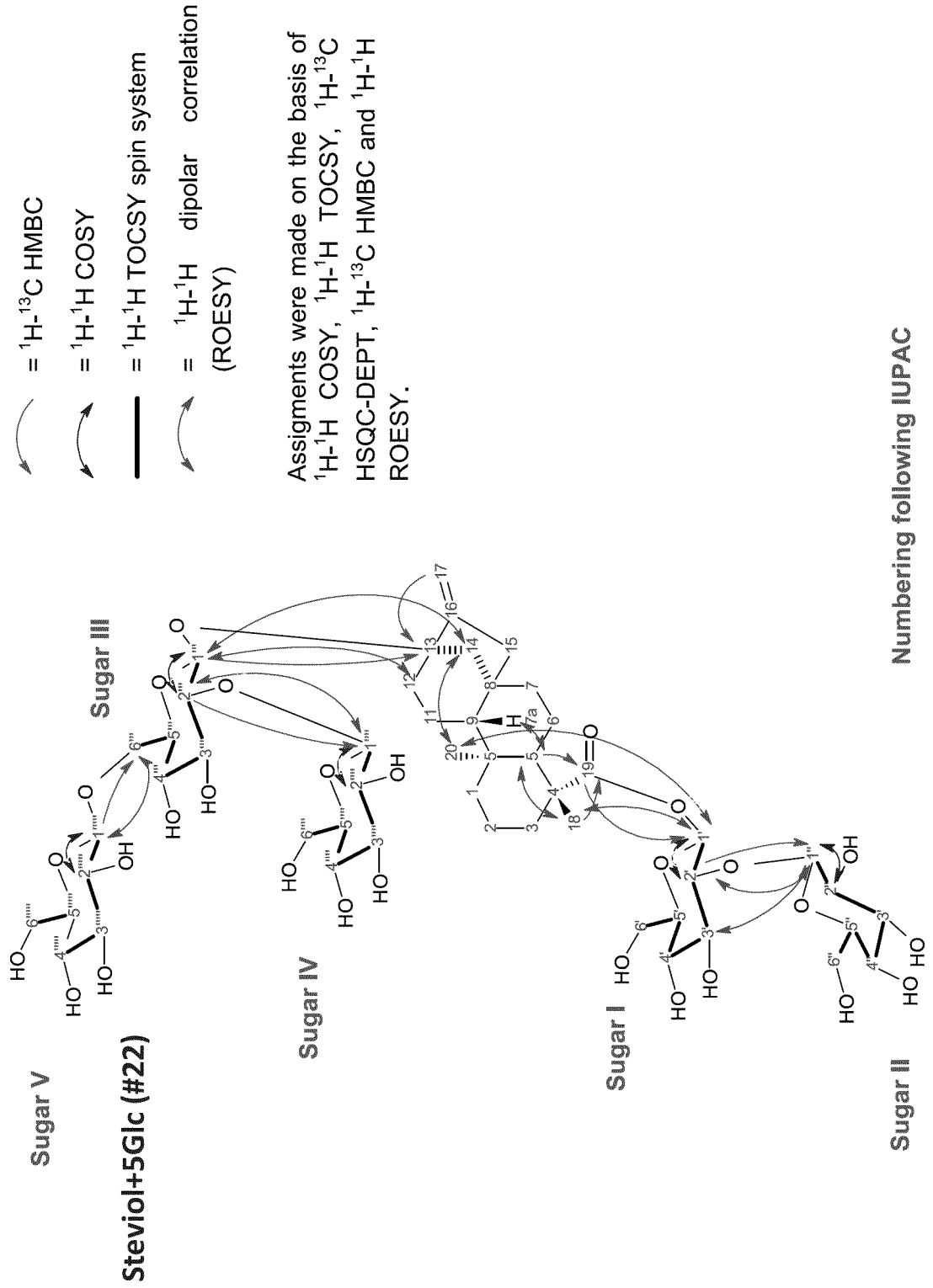


Figure 8AO

Steviol+7Glc (#14)

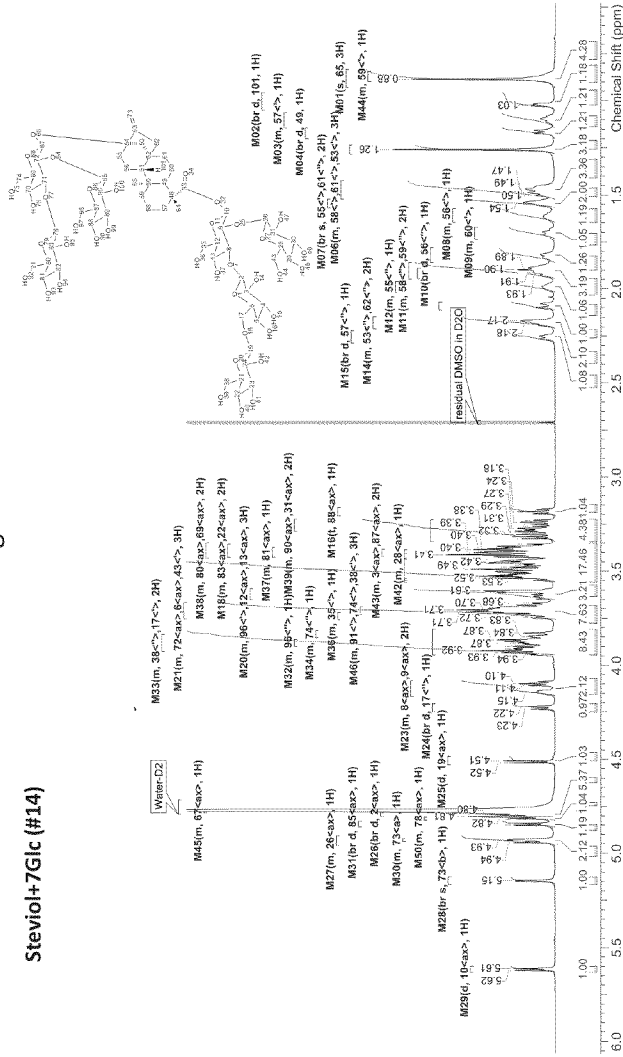


Figure 8AP

Steviol+7Glc (#14)

Multiplet	Shift1 (ppm)	Atom1	H's	Type	J (Hz)	(ppm)
M29	5.62	10<ax>	1	d	7.83	[5.60 .. 5.63]
M28	5.15	73<b>	1	br s	-	[5.13 .. 5.17]
M27	4.93	26<ax>	1	m	-	[4.91 .. 4.96]
M30	4.93	73<a>	1	m	-	[4.92 .. 4.94]
M31	4.85	85<ax>	1	br d	7.83	[4.83 .. 4.86]
M50	4.82	78<ax>	1	m	-	[4.81 .. 4.82]
M26	4.80	2<ax>	1	br d	8.56	[4.78 .. 4.81]
M45	4.78	67<ax>	1	m	-	[4.78 .. 4.79]
M25	4.51	19<ax>	1	d	8.07	[4.49 .. 4.53]
M24	4.23	17<e>	1	br d	10.03	[4.20 .. 4.25]
M23	4.12	8<ax>, 9<ax>	2	m	-	[4.08 .. 4.17]
M22	3.89	71<ax>, 43<e>	2	m	-	[3.81 .. 3.95]
M35	3.88	91<e>, 35<e>	2	m	-	[3.84 .. 3.93]
M33	3.88	38<e>, 17<e>	2	m	-	[3.84 .. 3.93]
M32	3.88	96<e>	1	m	-	[3.86 .. 3.90]
M34	3.83	74<e>	1	m	-	[3.82 .. 3.85]
M36	3.74	35<e>	1	m	-	[3.72 .. 3.76]
M46	3.71	91<e>, 74<e>, 38<e>	3	m	-	[3.69 .. 3.73]
M21	3.71	72<ax>, 6<ax>, 43<e>	3	m	-	[3.67 .. 3.75]
M20	3.62	96<e>, 12<ax>, 13<ax>	3	m	-	[3.59 .. 3.65]
M19	3.49	5<ax>	1	m	-	[3.44 .. 3.54]
M40	3.47	82<ax>, 70<ax>, 23<ax>	3	m	-	[3.39 .. 3.56]
M41	3.47	89<ax>, 21<ax>, 4<ax>, 30<ax>	4	m	-	[3.39 .. 3.55]
M42	3.42	28<ax>	1	m	-	[3.41 .. 3.43]
M43	3.40	3<ax>, 87<ax>	2	m	-	[3.38 .. 3.42]
M37	3.40	81<ax>	1	m	-	[3.37 .. 3.43]
M18	3.39	83<ax>, 22<ax>	2	m	-	[3.36 .. 3.43]
M38	3.39	80<ax>, 69<ax>	2	m	-	[3.35 .. 3.43]
M17	3.28	24<ax>, 29<ax>	2	m	-	[3.23 .. 3.34]
M39	3.26	90<ax>, 31<ax>	2	m	-	[3.22 .. 3.29]
M16	3.18	88<ax>	1	t	9.41	[3.16 .. 3.21]
M15	2.26	57<e>	1	br d	12.23	[2.22 .. 2.29]
M14	2.18	53<e>, 62<e>	2	m	-	[2.15 .. 2.21]
M13	2.09	62<e>	1	br d	17.12	[2.07 .. 2.11]
M12	1.99	55<e>	1	m	-	[1.96 .. 2.02]
M47	1.92	60<e>	1	m	-	[1.89 .. 1.95]
M11	1.90	58<e>, 59<e>	2	m	-	[1.86 .. 1.95]
M10	1.82	56<e>	1	br d	9.78	[1.76 .. 1.86]
M09	1.70	60<e>	1	m	-	[1.67 .. 1.73]
M08	1.61	56<e>	1	m	-	[1.57 .. 1.65]
M07	1.54	55<e>, 61<e>	2	br s	-	[1.52 .. 1.57]
M06	1.48	58<e>, 61<e>, 63<e>	3	m	-	[1.42 .. 1.52]
M05	1.26	64	3	s	-	[1.24 .. 1.28]
M04	1.16	49	1	br d	12.72	[1.14 .. 1.20]
M03	1.10	57<e>	1	m	-	[1.06 .. 1.14]
M02	1.02	101	1	br d	7.58	[0.96 .. 1.05]
M01	0.88	65	3	s	-	[0.83 .. 0.93]
M44	0.87	59<e>	1	m	-	[0.84 .. 0.90]

Multiplet	Shift1 (ppm)	Atom1	H's	Type	J (Hz)	(ppm)
M29	5.62	10<ax>	1	d	7.83	[5.60 .. 5.63]
M28	5.15	73<b>	1	br s	-	[5.13 .. 5.17]
M27	4.93	26<ax>	1	m	-	[4.91 .. 4.96]
M30	4.93	73<a>	1	m	-	[4.92 .. 4.94]
M31	4.85	85<ax>	1	br d	7.83	[4.83 .. 4.86]
M50	4.82	78<ax>	1	m	-	[4.81 .. 4.82]
M26	4.80	2<ax>	1	br d	8.56	[4.78 .. 4.81]
M45	4.78	67<ax>	1	m	-	[4.78 .. 4.79]
M25	4.51	19<ax>	1	d	8.07	[4.49 .. 4.53]
M24	4.23	17<e>	1	br d	10.03	[4.20 .. 4.25]
M23	4.12	8<ax>, 9<ax>	2	m	-	[4.08 .. 4.17]
M22	3.89	71<ax>, 43<e>	2	m	-	[3.81 .. 3.95]
M35	3.88	91<e>, 35<e>	2	m	-	[3.84 .. 3.93]
M33	3.88	38<e>, 17<e>	2	m	-	[3.84 .. 3.93]
M32	3.88	96<e>	1	m	-	[3.86 .. 3.90]
M34	3.83	74<e>	1	m	-	[3.82 .. 3.85]
M36	3.74	35<e>	1	m	-	[3.72 .. 3.76]
M46	3.71	91<e>, 74<e>, 38<e>	3	m	-	[3.69 .. 3.73]
M21	3.71	72<ax>, 6<ax>, 43<e>	3	m	-	[3.67 .. 3.75]
M20	3.62	96<e>, 12<ax>, 13<ax>	3	m	-	[3.59 .. 3.65]
M19	3.49	5<ax>	1	m	-	[3.44 .. 3.54]
M40	3.47	82<ax>, 70<ax>, 23<ax>	3	m	-	[3.39 .. 3.56]

Multiplet	Shift1 (ppm)	Atom1	H's	Type	J (Hz)	(ppm)
M41	3.47	89<ax>, 21<ax>, 4<ax>, 30<ax>	4	m	-	[3.39 .. 3.55]
M42	3.42	28<ax>	1	m	-	[3.41 .. 3.43]
M43	3.40	3<ax>, 87<ax>	2	m	-	[3.38 .. 3.42]
M37	3.40	81<ax>	1	m	-	[3.37 .. 3.43]
M18	3.39	83<ax>, 22<ax>	2	m	-	[3.36 .. 3.43]
M38	3.39	80<ax>, 69<ax>	2	m	-	[3.35 .. 3.43]
M17	3.28	24<ax>, 29<ax>	2	m	-	[3.23 .. 3.34]
M39	3.26	90<ax>, 31<ax>	2	m	-	[3.22 .. 3.29]
M16	3.18	88<ax>	1	t	9.41	[3.16 .. 3.21]
M15	2.26	57<e>	1	br d	12.23	[2.22 .. 2.29]
M14	2.18	53<e>, 62<e>	2	m	-	[2.15 .. 2.21]
M13	2.09	62<e>	1	br d	17.12	[2.07 .. 2.11]
M12	1.99	55<e>	1	m	-	[1.96 .. 2.02]
M47	1.92	60<e>	1	m	-	[1.89 .. 1.95]
M11	1.90	58<e>, 59<e>	2	m	-	[1.86 .. 1.95]
M10	1.82	56<e>	1	br d	9.78	[1.76 .. 1.86]
M09	1.70	60<e>	1	m	-	[1.67 .. 1.73]
M08	1.61	56<e>	1	m	-	[1.57 .. 1.65]
M07	1.54	55<e>, 61<e>	2	br s	-	[1.52 .. 1.57]
M06	1.48	58<e>, 61<e>, 63<e>	3	m	-	[1.42 .. 1.52]
M05	1.26	64	3	s	-	[1.24 .. 1.28]
M04	1.16	49	1	br d	12.72	[1.14 .. 1.20]
M03	1.10	57<e>	1	m	-	[1.06 .. 1.14]
M02	1.02	101	1	br d	7.58	[0.96 .. 1.05]
M01	0.88	65	3	s	-	[0.83 .. 0.93]
M44	0.87	59<e>	1	m	-	[0.84 .. 0.90]

Homonuclear dipolar correlations between:  
H-26 and H-9  
H-2 and H-8  
H-19 and H-17  
H-85 and H-72  
H-78 and H-71  
H-10 and Me-65, Me-64  
H-67 and H-53', H-55"  
H-101 and H-49  
H-49 and H-59', H-101, Me-64  
Me-65 and H-60', H-53", H-53' (indirect)  
Me-64 and H-60"

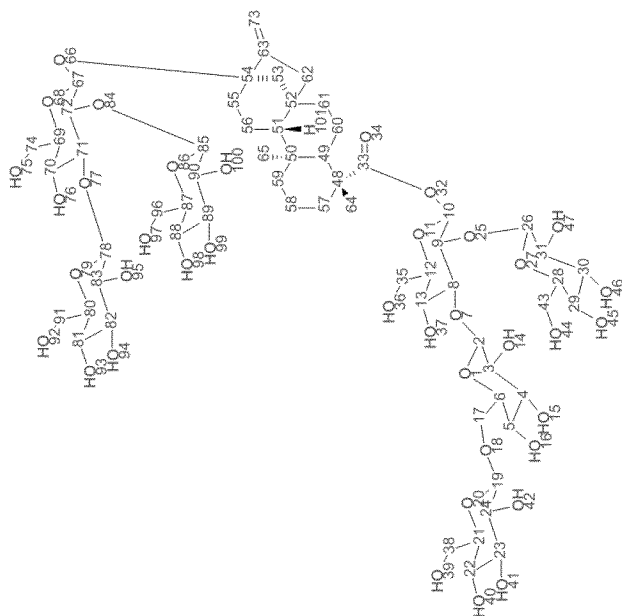
<sup>1</sup>H NMR (800 MHz, DEUTERIUM OXIDE)  $\delta$   
ppm 0.88 (s, 3 H) 0.84 - 0.90 (m, 1 H) 1.02 (br d, J=7.58 Hz, 1 H) 1.06 - 1.14 (m, 1 H) 1.16 (br d, J=12.72 Hz, 1 H) 1.26 (s, 3 H) 1.42 - 1.52 (m, 3 H) 1.54 (br s, 2 H) 1.57 - 1.65 (m, 1 H) 1.67 - 1.73 (m, 1 H) 1.82 (br d, J=9.78 Hz, 1 H) 1.86 - 1.95 (m, 2 H) 1.89 - 1.95 (m, 1 H) 1.96 - 2.02 (m, 1 H) 2.09 (br d, J=17.12 Hz, 1 H) 2.15 - 2.21 (m, 2 H) 2.26 (br d, J=12.23 Hz, 1 H) 3.18 (t, J=9.41 Hz, 1 H) 3.22 - 3.29 (m, 2 H) 3.23 - 3.34 (m, 2 H) 3.35 - 3.43 (m, 2 H) 3.36 - 3.43 (m, 2 H) 3.37 - 3.43 (m, 1 H) 3.38 - 3.42 (m, 2 H) 3.39 - 3.56 (m, 3 H) 3.39 - 3.55 (m, 4 H) 3.41 - 3.43 (m, 1 H) 3.44 - 3.54 (m, 1 H) 3.59 - 3.65 (m, 3 H) 3.67 - 3.75 (m, 3 H) 3.69 - 3.73 (m, 3 H) 3.72 - 3.76 (m, 1 H) 3.81 - 3.95 (m, 2 H) 3.82 - 3.85 (m, 1 H) 3.84 - 3.93 (m, 2 H) 3.84 - 3.93 (m, 2 H) 3.86 - 3.90 (m, 1 H) 4.08 - 4.17 (m, 2 H) 4.23 (br d, J=10.03 Hz, 1 H) 4.51 (d, J=8.07 Hz, 1 H) 4.78 - 4.79 (m, 1 H) 4.80 (br d, J=8.56 Hz, 1 H) 4.81 - 4.82 (m, 1 H) 4.85 (br d, J=7.83 Hz, 1 H) 4.91 - 4.96 (m, 1 H) 4.92 - 4.94 (m, 1 H) 5.15 (br s, 1 H) 5.62 (d, J=7.83 Hz, 1 H)

Figure 8AQ

**Steviol+7Glc (#14)**

F2 Atom	$\delta$ (1H) (ppm)	F1 Atom	$\delta$ (13C) (ppm)
2<ax>	4.8	2	102.9
3<ax>	3.4	3	77.1
4<ax>	3.52	4	76.7
5<ax>	3.49	5	70.4
6<ax>	3.71	6	75.6
8<ax>	4.14	8	86.3
9<ax>	4.1	9	76.6
10<ax>	5.62	10	93.4
12<ax>	3.61	12	76.9
13<ax>	3.61	13	69
17<'>	4.23	17	69.5
17<'>	3.86		
19<ax>	4.51	19	103.4
21<ax>	3.45	21	76.7
22<ax>	3.39	22	70.3
23<ax>	3.49	23	76.4
24<ax>	3.32	24	73.9
26<ax>	4.94	26	102.4
28<ax>	3.41	28	76.9
29<ax>	3.29	29	71.3
30<ax>	3.45	30	76.7
31<ax>	3.24	31	74.7
35<'>	3.87	33	179.3
35<'>	3.74	35	61.3
38<'>	3.91		
38<'>	3.71	38	61.4
43<'>	3.93	43	62.4
43<'>	3.7		

49	1.16	48	44.7
		49	57.4
		50	39.8
53<'>	2.17	52	41.9
53<'>	1.49	53	44.6
		54	88.8
55<'>	1.99	55	37.8
55<'>	1.54		
56<'>	1.82	56	20.5
56<'>	1.61		
57<'>	2.26	57	37.5
57<'>	1.1		
58<'>	1.88	58	19.7
58<'>	1.46		
59<'>	1.89	59	40.5
59<'>	0.87		
60<'>	1.92	60	22.5
60<'>	1.7		
61<'>	1.55	61	41.8
61<'>	1.47		
62<'>	2.18	62	47.2
62<'>	2.09		
		63	153.5
64	1.26	64	28.8
65	0.88	65	16.8
67<ax>	4.78	67	96.1
69<ax>	3.37	69	74.3
70<ax>	3.5	70	69.2
71<ax>	3.93	71	86
72<ax>	3.68	72	79.5



Heteronuclear scalar correlations between:

C=O(33) and H-10

C-54 and H-67

H-26 and C-9 (C-26 and H-9)

H-2 and C-8 (C-2 and H-8)





H-19 and C-17 (C-19 and H-17)

H-85 and C-72 (C-85 and H-72)

H-78 and C-71 (C-78 and H-71)

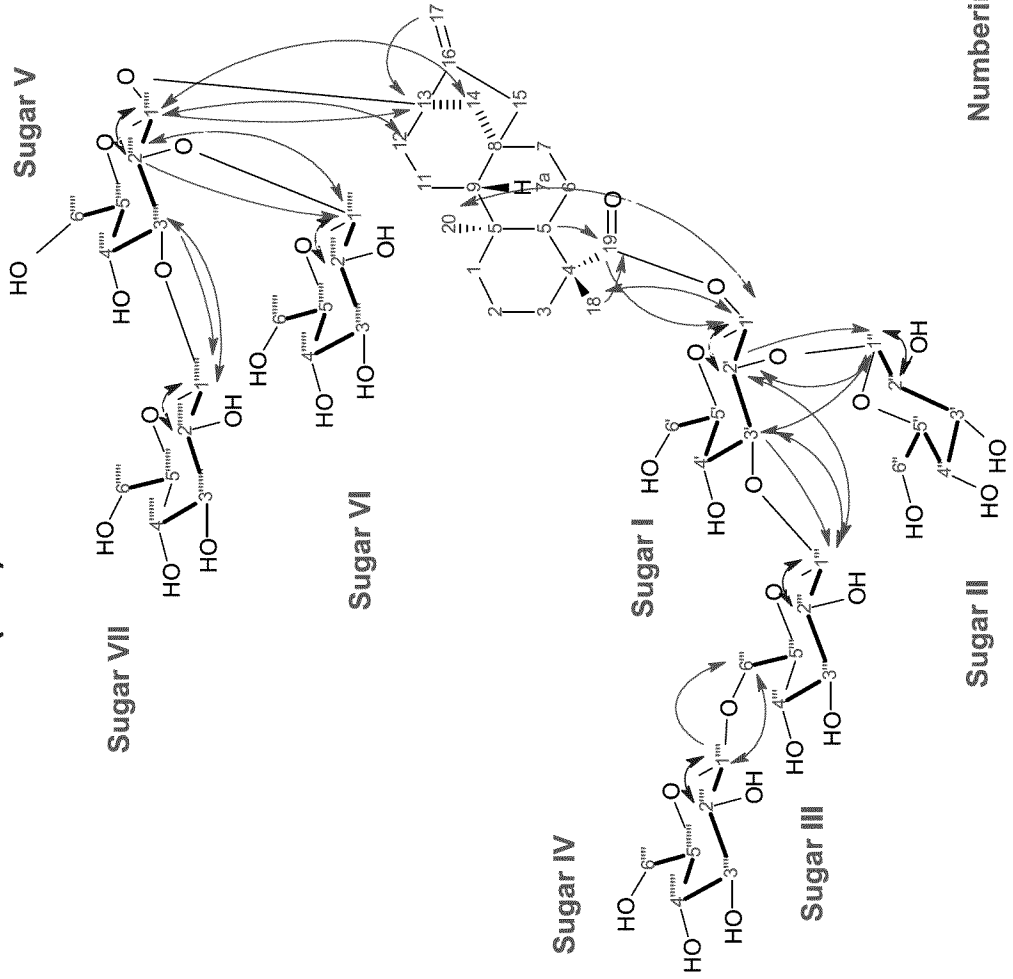
<sup>13</sup>C NMR (201 MHz, DEUTERIUM OXIDE)  $\delta$  ppm 179.3 (1C), 153.5 (1C), 105.1 (1C), 103.4 (1C), 103.0 (1C), 102.9 (2C), 102.4 (1C), 96.1 (1C), 93.4 (1C), 88.8 (1C), 86.3 (1C), 86.0 (1C), 79.5 (1C), 77.1 (2C), 76.9 (2C), 76.7 (4C), 76.6 (1C), 76.4 (2C), 76.0 (1C), 75.6 (1C), 75.0 (1C), 74.7 (1C), 74.3 (1C), 74.1 (1C), 73.9 (1C), 71.4 (1C), 71.3 (1C), 70.4 (1C), 70.3 (2C), 69.5 (1C), 69.2 (1C), 69.0 (1C), 62.4 (1C), 62.3 (1C), 61.4 (2C), 61.3 (2C), 57.4 (1C), 53.8 (1C), 47.2 (1C), 44.7 (1C), 44.6 (1C), 41.9 (1C), 41.8 (1C), 40.5 (1C), 39.8 (1C), 37.8 (1C), 37.5 (1C), 28.8 (1C), 22.5 (1C), 20.5 (1C), 19.7 (1C), 16.8 (1C)

Figure 8AR

-  =  $^1\text{H}-^{13}\text{C}$  HMBC
-  =  $^1\text{H}-^1\text{H}$  COSY
-  =  $^1\text{H}-^1\text{H}$  TOCSY spin system
-  =  $^1\text{H}-^1\text{H}$  dipolar correlation (ROESY)

Assignments were made on the basis of  $^1\text{H}-^1\text{H}$  COSY,  $^1\text{H}-^1\text{H}$  TOCSY,  $^1\text{H}-^{13}\text{C}$  HSQC-DEPT,  $^1\text{H}-^{13}\text{C}$  HMBC and  $^1\text{H}-^1\text{H}$  ROESY.

**Steviol+7Glc (#14)**



Numbering following IUPAC

Figure 9A

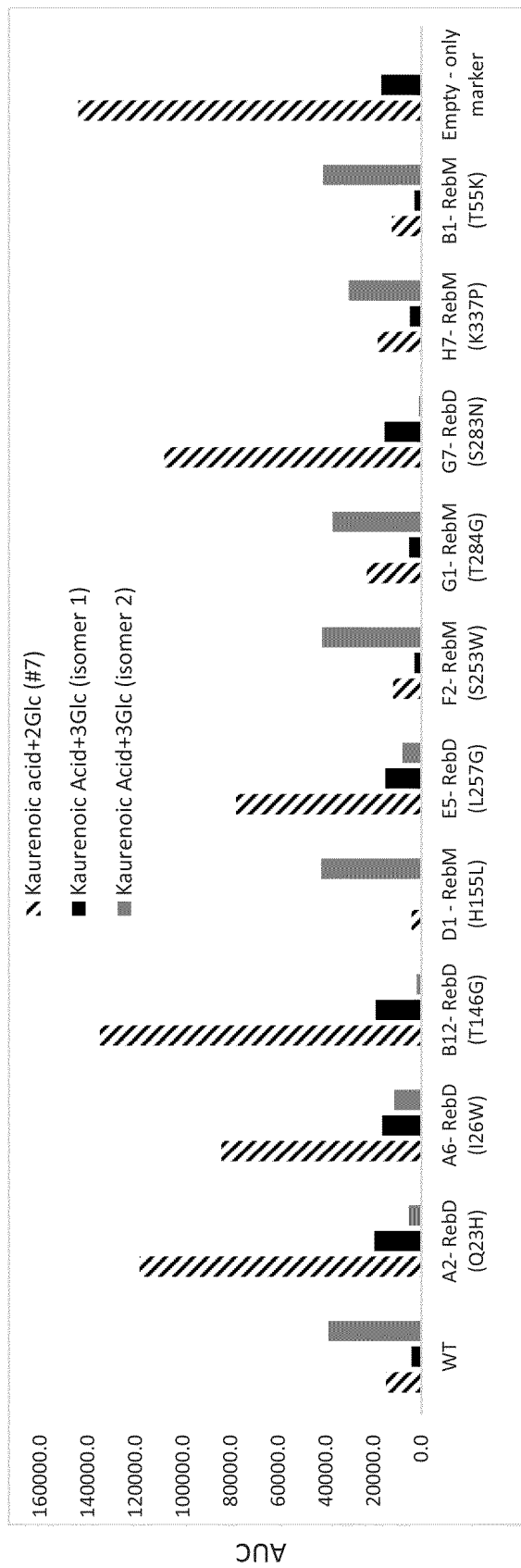


Figure 9B

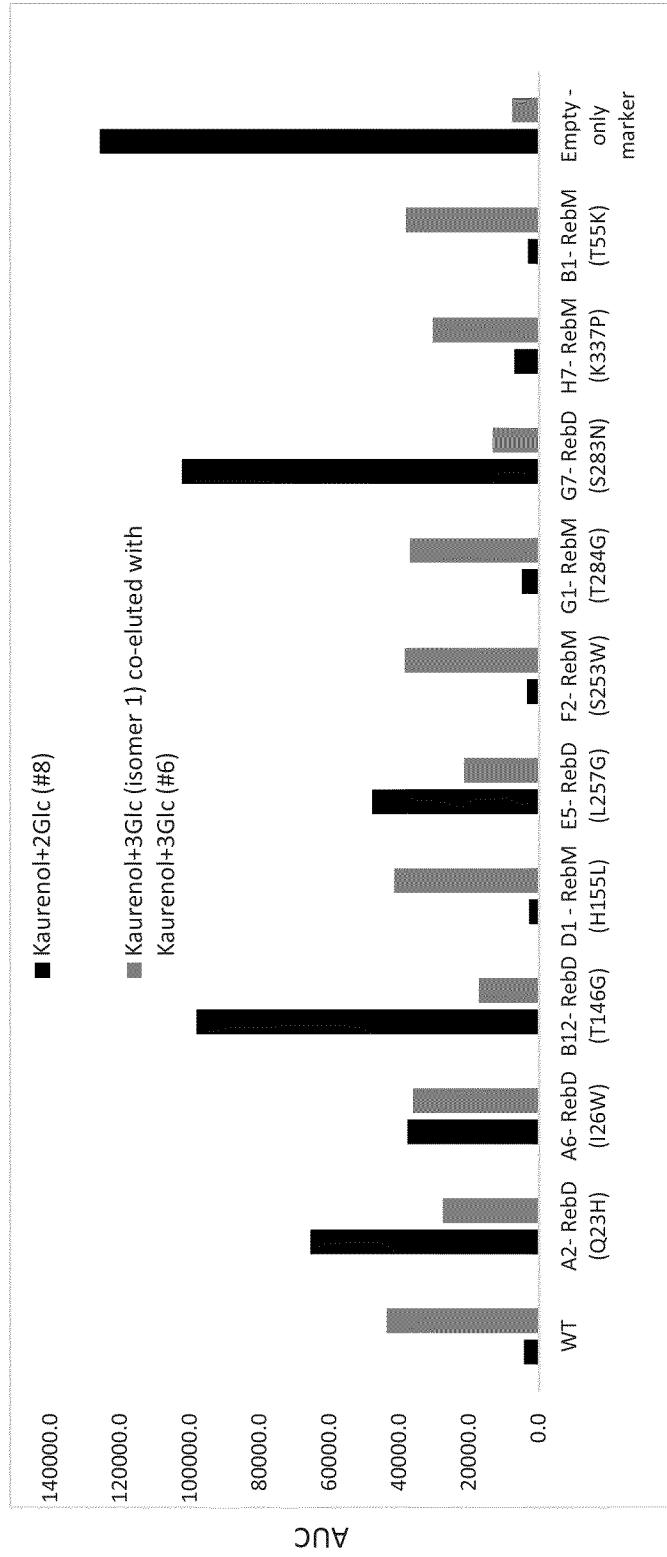


Figure 10A

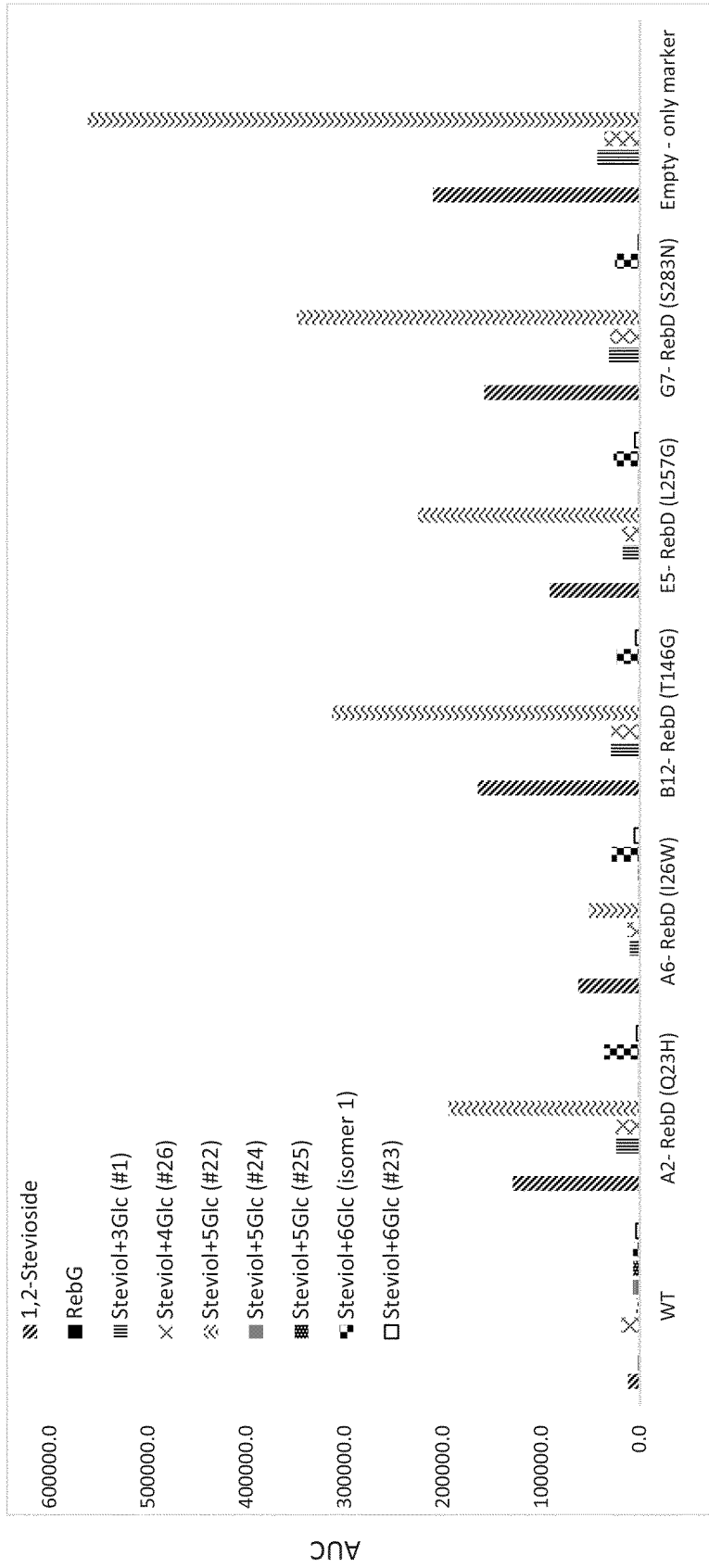


Figure 10B

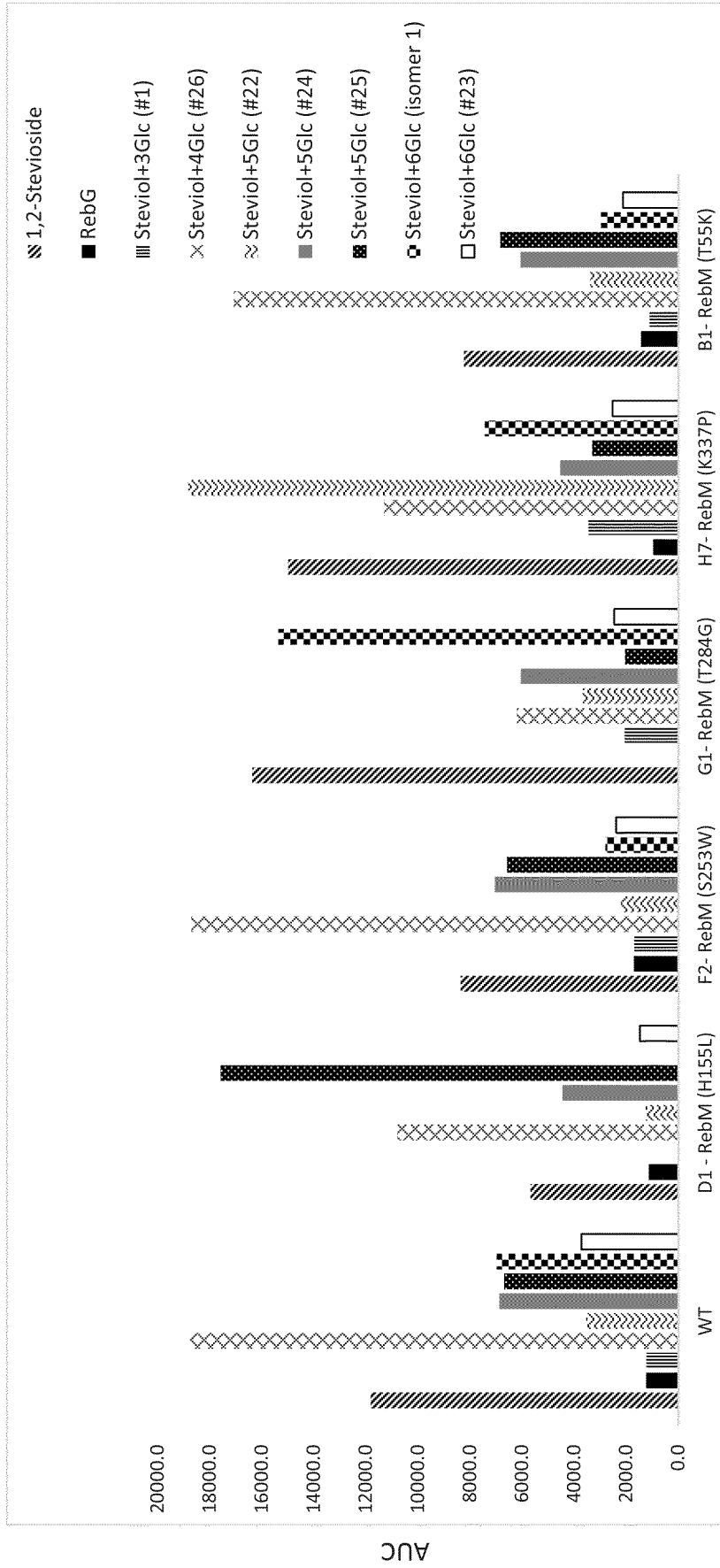


Figure 10C

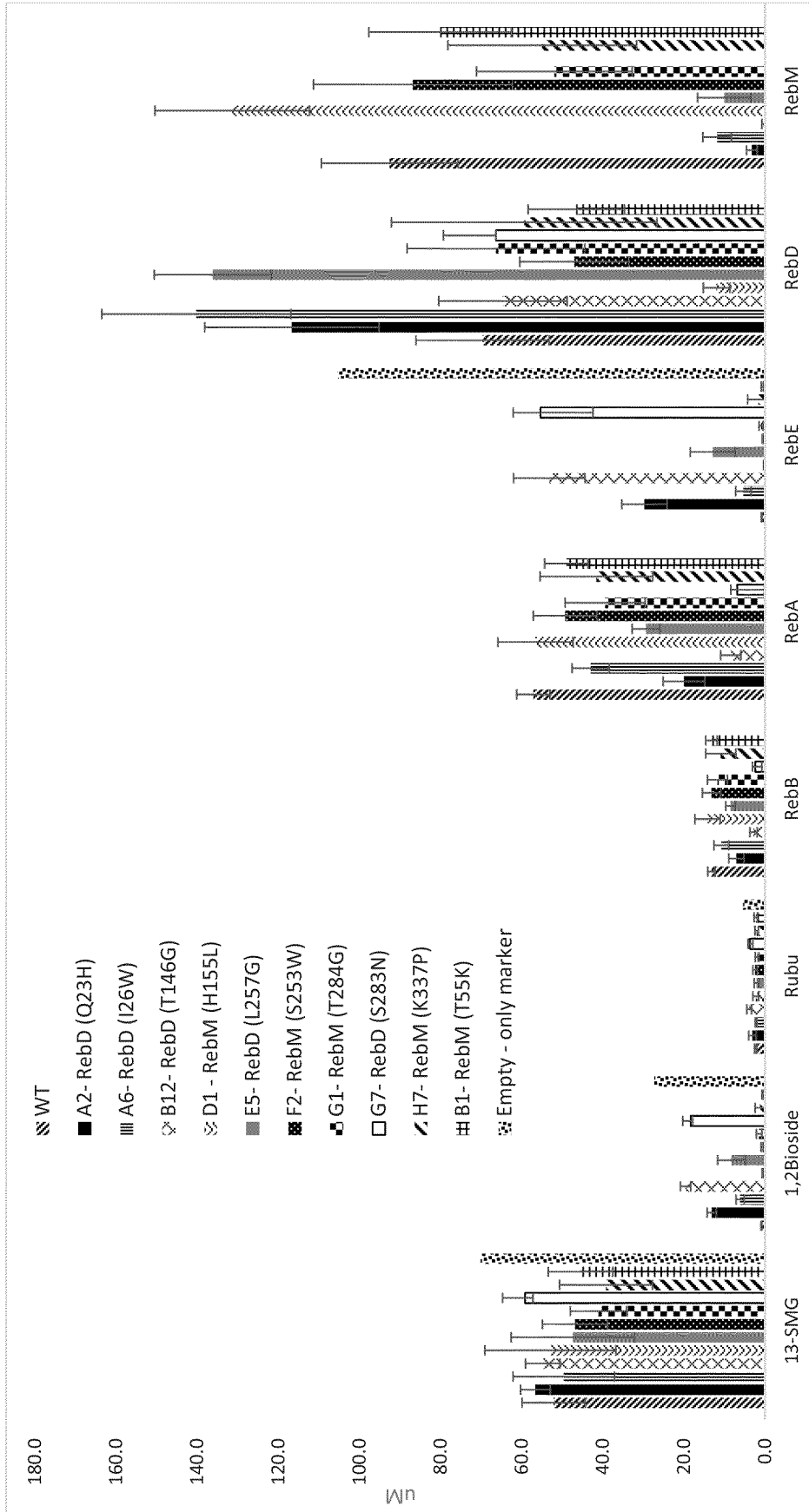


Figure 11A

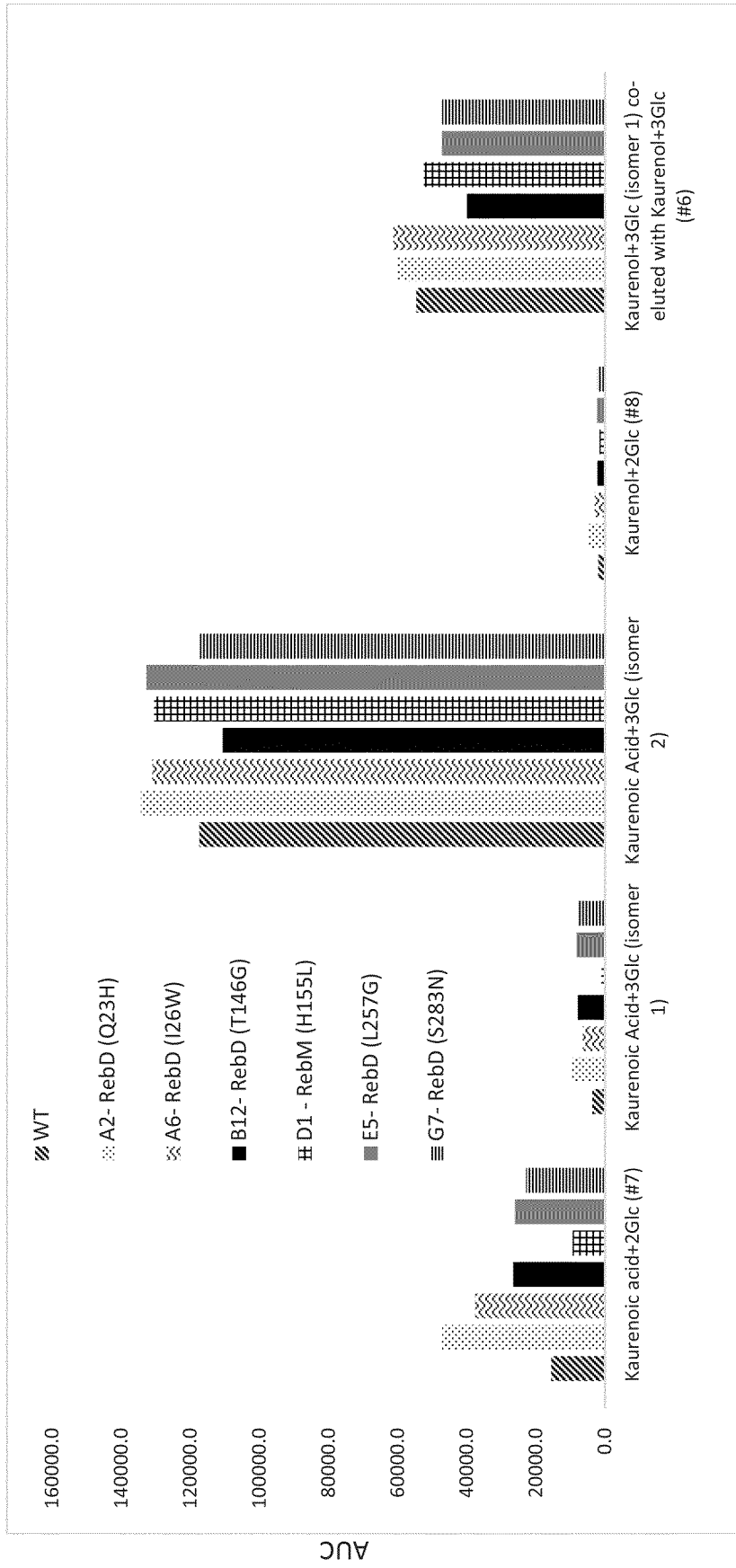


Figure 11B

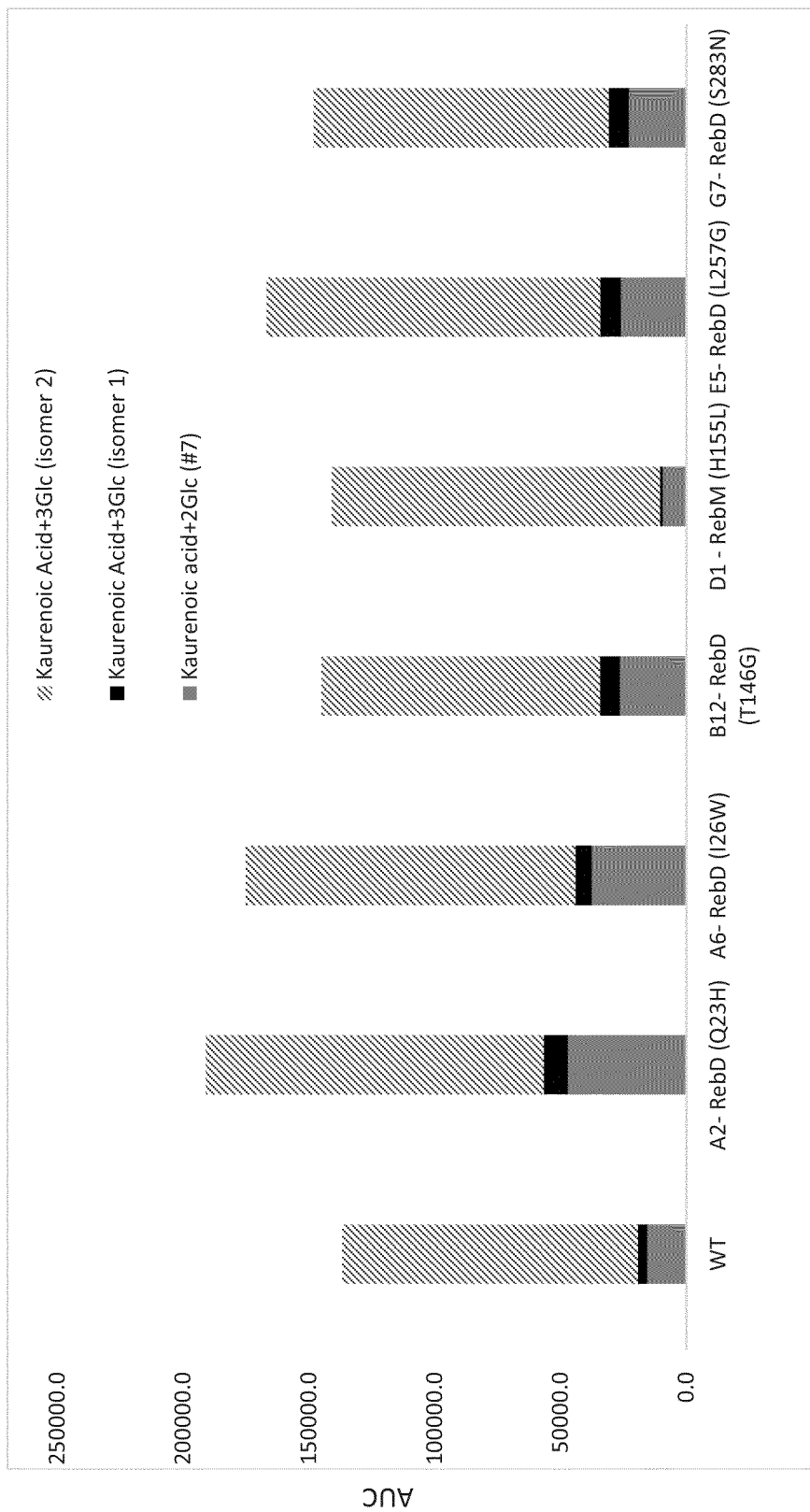


Figure 11C

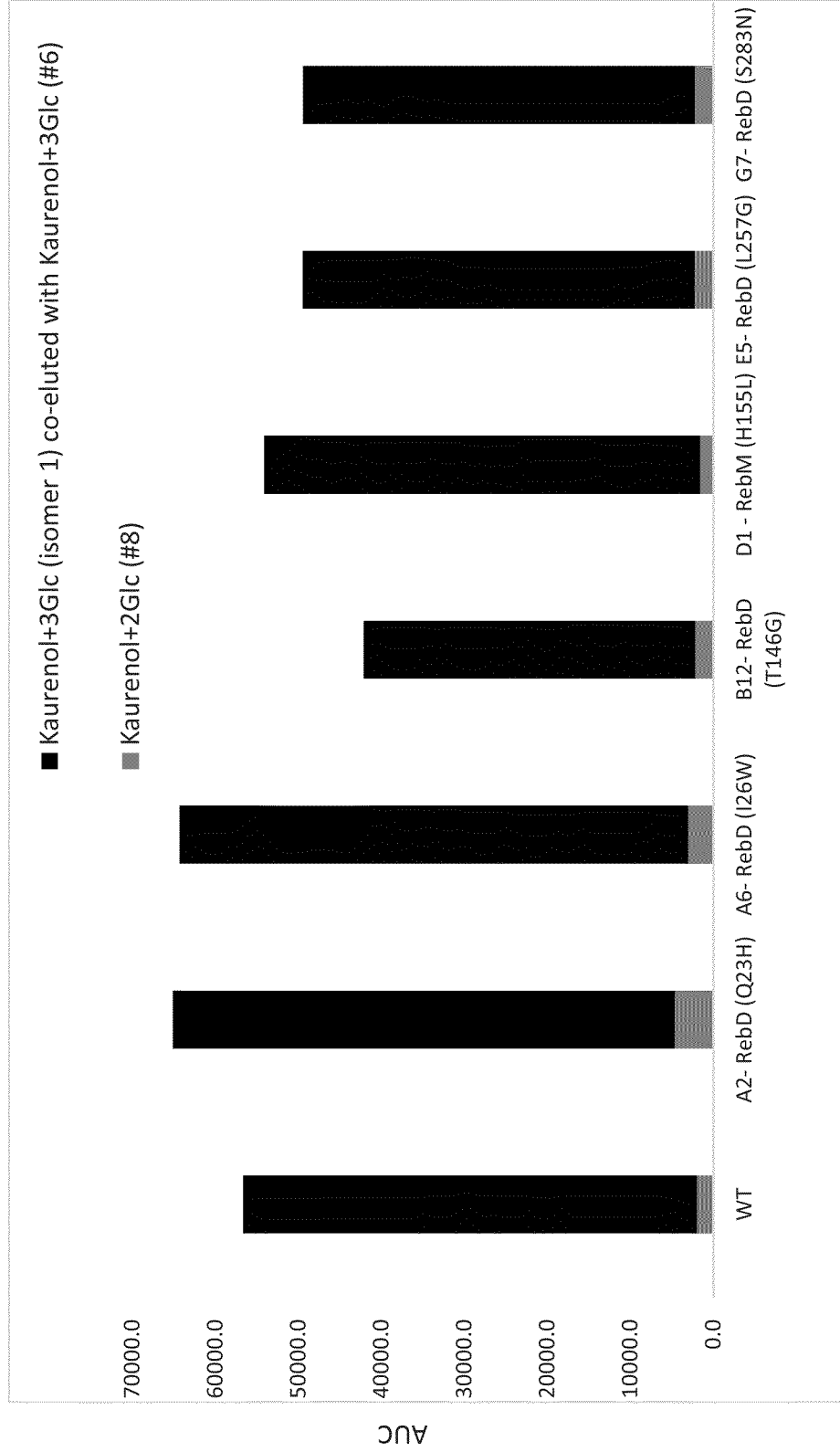


Figure 11D

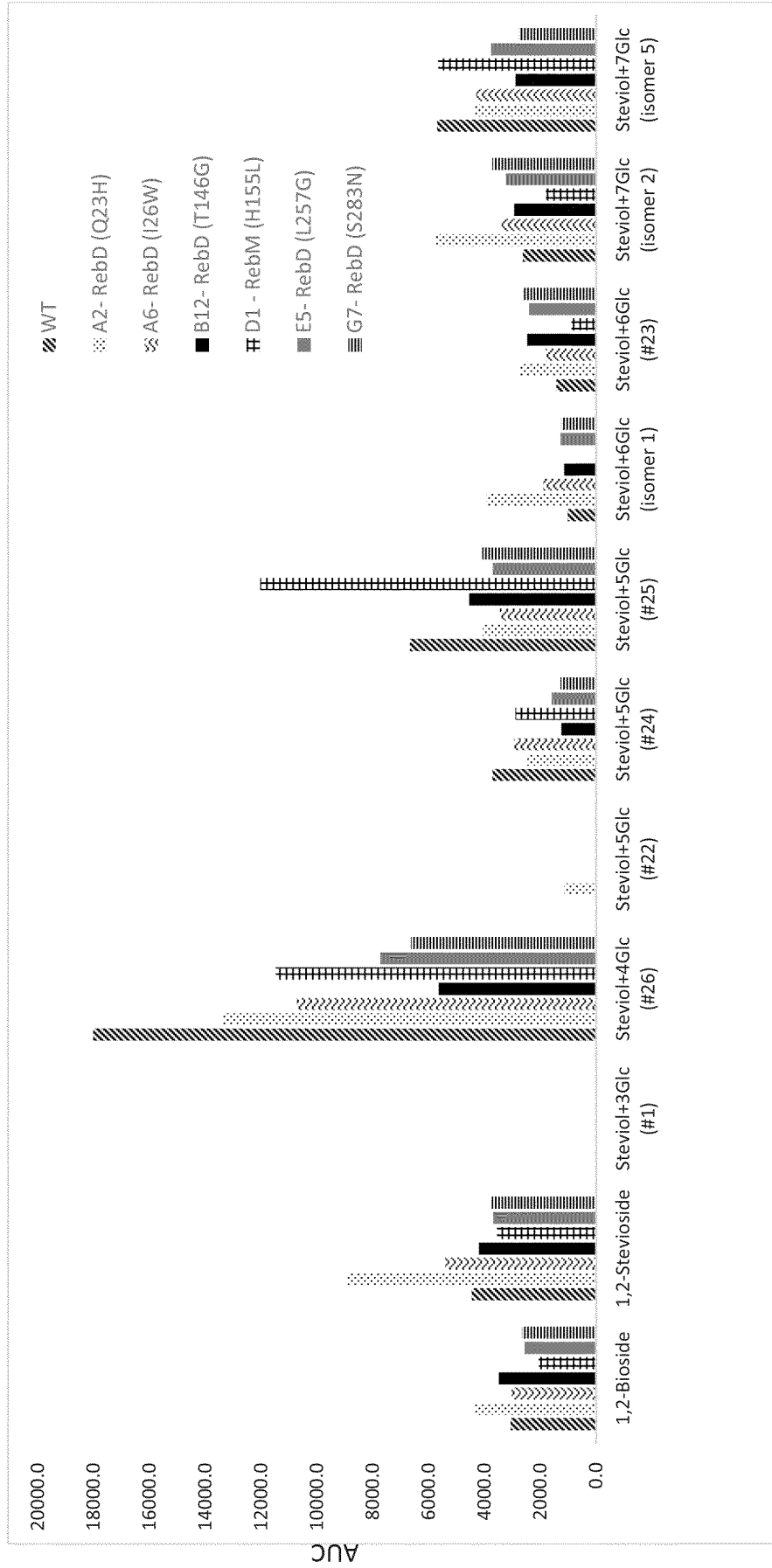
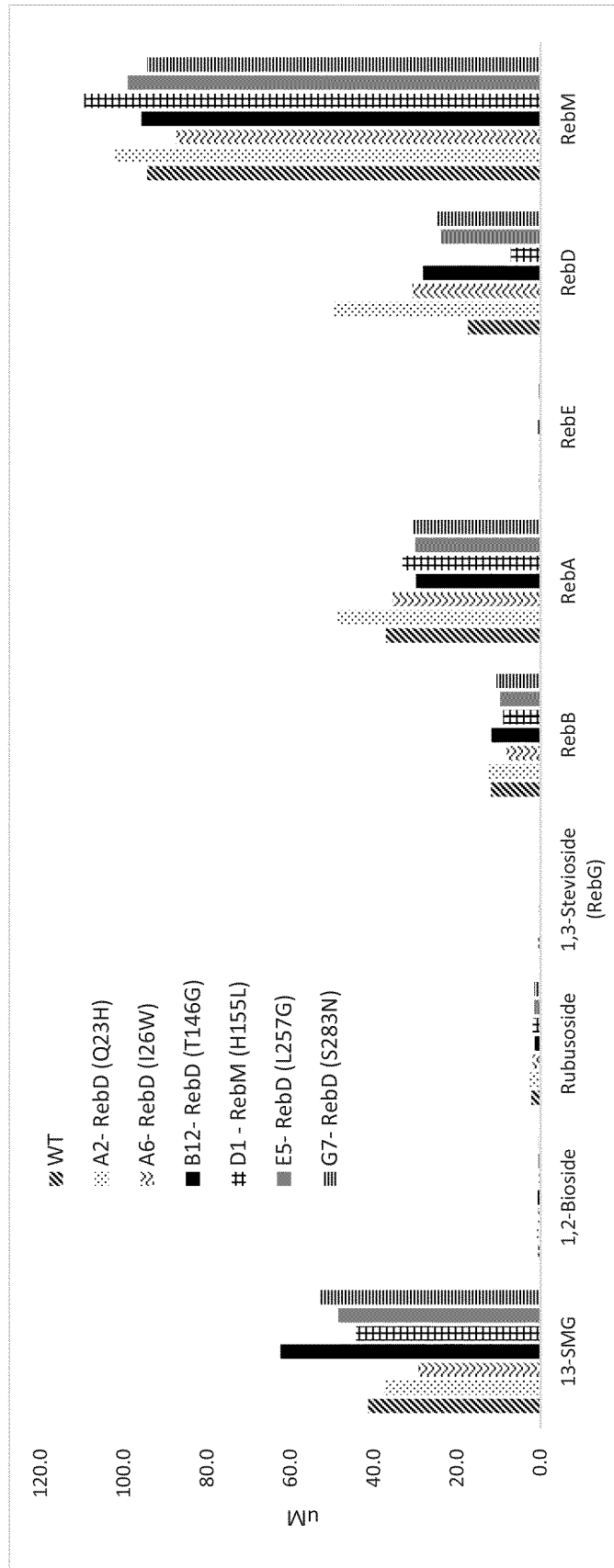


Figure 11E



**INTERNATIONAL SEARCH REPORT**

International application No  
PCT/EP2016/052007

**A. CLASSIFICATION OF SUBJECT MATTER**  
 INV. C12N9/10 C12P15/00 C12P19/56 C12N1/18  
 ADD.  
 According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**  
 Minimum documentation searched (classification system followed by classification symbols)  
 C12P C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)  
 EPO-Internal

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2014/122227 A2 (EVOLVA SA [CH]) 14 August 2014 (2014-08-14) the whole document	1-41
X	WO 2013/022989 A2 (EVOLVA SA [CH]; HOUGHTON-LARSEN JENS [DK]; HICKS PAULA M [US]; NAESBY) 14 February 2013 (2013-02-14) the whole document	1-5,8-41
X	WO 2014/086890 A1 (EVOLVA SA [CH]) 12 June 2014 (2014-06-12) the whole document	1,2,5, 8-41
X	WO 2014/122328 A1 (EVOLVA SA [CH]) 14 August 2014 (2014-08-14) the whole document	1-3,5,6, 8-21, 23-41

Further documents are listed in the continuation of Box C.

See patent family annex.

\* Special categories of cited documents :

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier application or patent but published on or after the international filing date
- "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "&" document member of the same patent family

Date of the actual completion of the international search  
 22 June 2016

Date of mailing of the international search report  
 04/07/2016

Name and mailing address of the ISA/  
 European Patent Office, P.B. 5818 Patentlaan 2  
 NL - 2280 HV Rijswijk  
 Tel. (+31-70) 340-2040,  
 Fax: (+31-70) 340-3016

Authorized officer  
 Sonnerat, Isabelle

# INTERNATIONAL SEARCH REPORT

International application No.  
PCT/EP2016/052007

## Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1.  Claims Nos.:  
because they relate to subject matter not required to be searched by this Authority, namely:
  
2.  Claims Nos.:  
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
  
3.  Claims Nos.:  
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

## Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1.  As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
  
2.  As all searchable claims could be searched without effort justifying an additional fees, this Authority did not invite payment of additional fees.
  
3.  As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:  
  
1-41
  
4.  No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

### Remark on Protest

- The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- No protest accompanied the payment of additional search fees.

**FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210**

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. claims: 2, 5(completely); 1, 8-41(partially)

Recombinant host cell comprising a gene encoding a UGT91D2e polypeptide having at least 90% sequence identity to the amino acid sequence set forth in SEQ ID No. 11, wherein the host cell is capable of producing a steviol glycoside, glycosylated ent-kaurenol compound and/or a glycosylated ent-kaurenoic acid compound in a cell culture broth; subject-matter related thereto.

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2. claims: 1, 8-41(all partially)

Recombinant host cell comprising a gene encoding a chimeric polypeptide having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID No. 17 or SEQ ID No. 18, wherein the host cell is capable of producing a steviol glycoside, glycosylated ent-kaurenol compound and/or a glycosylated ent-kaurenoic acid compound in a cell culture broth; subject-matter related thereto.

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3. claims: 3, 6(completely); 1, 8-41(partially)

Recombinant host cell comprising a gene encoding a UGT85C2 polypeptide having at least 55% sequence identity to the amino acid sequence set forth in SEQ ID No. 7, wherein the host cell is capable of producing a steviol glycoside, glycosylated ent-kaurenol compound and/or a glycosylated ent-kaurenoic acid compound in a cell culture broth; subject-matter related thereto.

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4. claims: 4, 7(completely); 1, 8-41(partially)

Recombinant host cell comprising a gene encoding a UGT76G1 polypeptide having at least 50% sequence identity to the amino acid sequence set forth in SEQ ID No. 9, wherein the host cell is capable of producing a steviol glycoside, glycosylated ent-kaurenol compound and/or a glycosylated ent-kaurenoic acid compound in a cell culture broth; subject-matter related thereto.

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5. claim: 42

Reaction mixture comprising (a) a steviol glycoside, glycosylated ent-kaurenol compound and/or a glycosylated ent-kaurenoic acid compound, (b) a UGT polypeptide, (c) glucose, fructose, sucrose, xylose, ethanol, glycerol, UDP-glucose, UDP-rhamnose, UDP-xylose and/or GlcNAc and/or (d) reaction buffer and/or salts.

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# INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No

PCT/EP2016/052007

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