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(54) Title: RNAi FOR THE CONTROL OF FUNGI AND OOMYCETES BY INHIBITING SACCHAROPINE DEHYDROGENASE GENE

(57) Abstract: The present invention relates to control of plant pathogens, particularly fungi or oomycetes, by inhibiting one or more biological functions, particularly by inhibiting saccharopine dehydrogenase gene(s) using RNA interference. The invention provides methods and compositions using RNA interference of plant pathogens target genes for such control. The invention is also directed to methods for making transgenic plants tolerant to said plant pathogens, and to transgenic plants and seeds generated thereof.



WO 2013/050410 A1

**RNAi for the control of fungi and oomycetes by inhibiting saccharopine dehydrogenase gene**

The present invention relates to control of plant pathogens and pests, particularly fungi or oomycetes, by inhibiting one or more biological functions, particularly by inhibiting fungi saccharopine dehydrogenase gene involved in the  $\alpha$ -amino adipate pathway for lysine biosynthesis and their oomycetes homologs using RNA interference.

The invention provides methods and compositions using RNA interference of fungi or oomycetes target genes for such control. The invention is also directed to methods for making transgenic plants tolerant to said fungi or oomycetes, and to transgenic plants and seeds generated thereof.

The technology used in the context of the present invention is RNA interference or RNAi.

The expression in an organism of a sequence homologous to a target-gene capable of inducing the formation of small double-stranded RNA (dsRNA) makes it possible, very specifically, to extinguish this gene and to observe the phenotype that results therefrom (Xiao *et al.*, 2003).

The dsRNA triggers the specific degradation of a homologous RNA only in the region of identity with the dsRNA (Zamore *et al.*, 2000; Tang *et al.*, 2003). The dsRNA is an RNA molecule which contains a double-stranded sequence, generally of at least 19 base pairs (bp) including a sense strand and an antisense strand. The dsRNA molecules are also characterized by the very large degree of complementarity between the two complementary RNA strands. The dsRNA is degraded into RNA fragments of generally 18 to 25 nucleotides (siRNA) and the cleavage sites on the target RNA are evenly spaced apart by 18 to 25 nucleotides. The small siRNAs resulting therefrom exhibit a very high degree of identity with respect to the target RNA; however, mismatches of 3 to 4 nucleotides between the siRNA and the corresponding portion of the target RNA nevertheless make it possible for the system to operate (Tang *et al.*, 2003). It has thus been suggested that these fragments of 18 to 25 nucleotides constitute RNA guides for recognition of the target (Zamore *et al.*, 2000). These small RNAs have also been detected in extracts prepared from Schneider 2 cells of *Drosophila melanogaster* which had been transfected with dsRNAs before cell lysis (Hammond *et al.*, 2000). The guiding role of the fragments of 18 to 25 nucleotides in the cleavage of the mRNAs is supported by the observation that these fragments of 19 to 25 nucleotides isolated from dsRNA are capable of being involved in the degradation of mRNA (Zamore *et al.*, 2000). Sizable homologous RNA molecules also accumulate in plant tissues which undergo the PTGS phenomenon (Post Transcriptional Gene Silencing, Hamilton and Baulcome, 1999). These small RNAs can regulate gene expression at three different levels:

- transcription (TGS for Transcriptional Gene Silencing),
- messenger RNA degradation (PTGS for Post Transcriptional Gene Silencing),
- miRNA pathway
- translation.

Regulation involving messenger RNA degradation appears to exist in all eukaryotes, whereas regulation at the transcriptional level has only been described in mammals, plants, drosophila and *C. elegans*. As regards the regulation of translation, it has been characterized in *C. elegans* and drosophila and appears also to exist in mammals (Hannon, 2002) and plants (Ruiz Ferrer and Voinnet, 2009). In the literature, reference is made to RNAi, to PTGS, to cosuppression or to quelling (reserved for fungi) when referring to this phenomenon, depending on the organisms in which it is studied.

RNAi has in particular proved that it is effective when double-stranded RNA (dsRNA) is injected into the nematode *Caenorhabditis elegans* (Fire et al. 1998; Montgomery et al., 1998; WO99/32619). Inhibition of the expression of an insect target gene was also observed when this insect is fed with bacteria expressing small double-stranded RNAs corresponding to said insect target gene (WO 01/37654).

More recently, pharmaceutical compositions comprising dsRNA substantially complementary to at least part of a gene suspected to be involved in the human papilloma virus (HPV) infection together with a pharmaceutically acceptable carrier have been disclosed to treating said HPV infection (WO2009/0247607).

The introduction of dsRNA was carried out in plants in order to induce silencing of an endogenous target gene (Hamilton et al., 1998, WO99/15682), to induce resistance to RNA viruses by means of the use of a transgene expressing a dsRNA having substantial identity with respect to the viral genes (Waterhouse et al., 1998; Pandolfini et al., 2003, WO98/36083, WO99/15682, US 5,175,102), but also to induce resistance to nematodes (Chuang and Meyerowitz, 2000, WO01/96584) or alternatively to the bacterium *Agrobacterium* (WO00/26346, Escobar et al., 2001). More recently, it has been shown that plants expressing dsRNA having substantial identity against a fungal gene essential to the growth of the fungus or to its pathogenicity may also induced resistance to this fungus (WO05/071091).

Nevertheless, since that time, only few preliminary results and no commercial examples exist on RNAi-mediated resistance or tolerance to phytopathogenic fungi where the double-stranded (dsRNA) or small interfering (siRNA) molecules are expressed in the plant, or applied as part of an external composition to the seed, the plant or to the fruit of the plant or to soil or to inert substrate wherein the plant is growing or wherein it is desired to grow.

Among others, one difficulty is to find an appropriate target gene, whose inhibition by dsRNA or siRNA induces a good level of fungi tolerance, up to a level suitable for a commercial use, without deleterious effect on the plant expressing said dsRNA or siRNA or on which a composition comprising said dsRNA or siRNA is applied.

Stärkel C. attempted in her Ph. D. thesis ["Host induced gene silencing – strategies for the improvement of resistance against *Cercospora beticola* in sugar beet (*B. vulgaris* L.) and against *Fusarium graminearum* in wheat (*T. aestivum* L.) and maize (*Z. mays* L.)", defended in June 2011] to inhibit the growth of *Fusarium graminearum* by transforming wheat with silencing constructs targeting the homoaconitase gene, an essential gene in the lysine biosynthesis pathway. Nevertheless, no transgenic wheat plants could be generated. Moreover, the attempt to delete or silence the

homoaconitase gene in *Fusarium graminearum* by transformation of the fungus with a construct designed for the inducible silencing of the homoaconitase gene were also unsuccessful.

The present inventors have surprisingly demonstrated that inhibition of fungus or oomycetes saccharopine dehydrogenase gene, which is involved in the  $\alpha$ -aminoadipate (AAA) pathway, via RNAi methodology causes cessation of infection, growth, development, reproduction and/or pathogenicity, and eventually results in the death of the organism.

This new target for the RNAi technology is particularly suitable, considering that AAA pathway is specifically found in some plant pathogens, including higher fungi, and not in plants, humans and animals.

Among the 20 common proteinogenic amino acids, L-lysine is the only one known to have a biosynthetic pathway which differs in plants and in higher fungi. In plants and bacteria, L-lysine is obtained through the diaminopimelate (DAP) pathway. In higher fungi and euglenoids, L-lysine is obtained through the  $\alpha$ -aminoadipate (AAA) pathway. Saccharopine dehydrogenase, homocitrate synthase, homoaconitase, homoisocitrate dehydrogenase,  $\alpha$ -Aminoadipate aminotransferase,  $\alpha$ -Aminoadipate reductase and saccharopine reductase are enzymes involved in the  $\alpha$ -aminoadipate pathway for the biosynthesis of L-lysine.

None of the enzymes involved in these two distinct pathways (DAP and AAA pathways) are common (Xu et al., 2006; Bhattacharjee, J. K., 1985; Bhattacharjee, J. K., 1992; Vogel, H. J., 1965). As an example, fungi saccharopine dehydrogenase is involved in the Lysine biosynthesis, when the lysine-ketoglutarate from plant, sometimes also called saccharopine dehydrogenase, is responsible for lysine catabolism (Houmard et al., 2007). For humans and animals, L-lysine is an essential amino acid which can only be obtained from protein in the diet. Additionally, enzymes involved in the fungal AAA pathway are unique to lysine synthesis (Umbargar, H. E., 1978; Bhattacharjee, J. K., 1992).

The presence of a specific  $\alpha$ -aminoadipate (AAA) pathway in higher fungi, which is not present nor in plant nor in humans or animals, leads to consider the enzymes involved in said AAA pathway as particularly attractive targets for the control of plant pathogens, particularly for fungal pathogens.

Interestingly, although lysine has been reported to be synthesized in oomycetes via the diaminopimelic pathway (Born and Blanchard, 1999), genes homologous to fungus genes of AAA pathway have been found in oomycetes, which may indicate that both pathways might be present in oomycetes. The inventors have shown that dsRNA against oomycete saccharopine dehydrogenase gene substantially reduced the growth of said oomycete, and that plants expressing dsRNA against oomycete saccharopine dehydrogenase genes could be less susceptible to said oomycete infection.

The presence of the AAA pathway for lysine biosynthesis has been demonstrated and studied for example in *Saccharomyces cerevisiae* (Broquist, H. P., 1971; Bhattacharjee, J. K., 1992), the human pathogenic fungi *Candida albicans* (Garrad, R. C. and Bhattacharjee, J. K., 1992), and the plant

pathogen *Magnaporthe grisea* (Umbargar, H. E., 1978). The saccharopine dehydrogenase enzymes involved in the AAA pathway have been intensively studied and compared in different organisms, and their technical features, including their nucleotides and aminoacids sequences, kinetics, substrate specificity, function, 3D- structure, as well as the way to purify and characterize them, are well known from the skilled man (see Xu et al., 2001, the content of which is incorporated herein by way of reference). Numerous genes from different fungi or oomycetes and their sequence data are disclosed and available in searchable public database, such as genBank.

Randall, T.A. et al. (2005) reports a list of genes of AAA pathway from the oomycete *Phytophthora infestans*, identified by their accession numbers and the sequence data thereof, incorporated herein by reference, are available in searchable public database (s) (Randall, T.A., 2005, MPMI, 18, 229-243; see in particular table 8 p 239).

In one embodiment, the present invention provides a dsRNA molecule comprising  
1) a first strand comprising a sequence substantially identical to at least 18 contiguous nucleotides of a fungus or oomycete gene and ii) a second strand comprising a sequence substantially complementary to the first strand, wherein said fungus or oomycete gene is a saccharopine dehydrogenase gene.

As used herein, "RNAi" or "RNA interference" refers to the process of sequence- specific gene silencing, mediated by double-stranded RNA (dsRNA). As used herein, "dsRNA" or "dsRNA molecule" refers to RNA that is partially or completely double stranded. Double stranded RNA is also referred to small or short interfering RNA (siRNA), short interfering nucleic acid (siNA), short interfering RNA, micro-RNA (miRNA), circular interfering RNA (ciRNA), short hairpin RNA (shRNA) and the like.

As used herein, taking into consideration the substitution of uracil for thymine when comparing RNA and DNA sequences, the term "substantially identical" or "essentially homologous" as applied to dsRNA means that the nucleotide sequence of one strand of the dsRNA is at least about 80%, at least 85% identical to 18 or more contiguous nucleotides of the target gene, more preferably at least about 90 % identical to 18 or more contiguous nucleotides of the target gene, and most preferably at least about 95%, 96%, 97%, 98% or 99% identical or absolutely identical to 18 or more contiguous nucleotides of the target gene. 18 or more nucleotides means a portion, being at least about 18, 20, 21 , 22, 23, 24, 25, 50, 100, 200, 300, 400, 500, 1000, 1500, or 2000 consecutive bases or up to the full length of the target gene.

As used herein, "complementary" polynucleotides are those that are capable of base pairing according to the standard Watson-Crick complementarity rules. Specifically, purines will base pair with pyrimidines to form a combination of guanine paired with cytosine (G:C) and adenine paired with either thymine (A:T) in the case of DNA, or adenine paired with uracil (A:U) in the case of RNA. It is understood that two polynucleotides may hybridize to each other even if they are not completely

complementary to each other, provided that each has at least one region that is substantially complementary to the other. As used herein, the term "substantially complementary" means that two nucleic acid sequences are complementary over at least 80% of their nucleotides. Preferably, the two nucleic acid sequences are complementary over at least 85%, 90%, 95%, 96%, 97%, 98%, 99% or more or all of their nucleotides. Alternatively, "substantially complementary" means that two nucleic acid sequences can hybridize under high stringency conditions. As used herein, the term "substantially identical" or "corresponding to" means that two nucleic acid sequences have at least 80% sequence identity. Preferably, the two nucleic acid sequences have at least 85%, 90%, 95%, 96%, 97%, 98%, 99% or 100% of sequence identity.

Also as used herein, the terms "nucleic acid" and "polynucleotide" refer to RNA or DNA that is linear or branched, single or double stranded, or a hybrid thereof. The term also encompasses RNA/DNA hybrids. When dsRNA is produced synthetically, less common bases, such as inosine, 5-methylcytosine, 6-methyladenine, hypoxanthine and others can also be used for antisense, dsRNA, and ribozyme pairing. For example, polynucleotides that contain C-5 propyne analogues of uridine and cytidine have been shown to bind RNA with high affinity and to be potent antisense inhibitors of gene expression. Other modifications, such as modification to the phosphodiester backbone, locked nucleic acid or the 2'-hydroxy in the ribose sugar group of the RNA can also be made.

Accordingly to the invention, the first strand and second strand may have identical sizes. Alternatively, the size of the first strand may be greater than that of the second strand. By way of example, the size of the first strand can be about 200 nucleotides greater than the size of the second strand. In another aspect of the invention, the size of second strand is greater than that of the first strand.

Accordingly to the invention, the dsRNA molecule comprises a first strand comprising a sequence substantially identical to at least 18 contiguous nucleotides of a fungus or oomycete saccharopine dehydrogenase gene.

In a particular embodiment, the invention provides a dsRNA molecule comprising  
 1) a first strand comprising a sequence substantially identical to at least 18 contiguous nucleotides of a fungus or oomycete gene and ii) a second strand comprising a sequence substantially complementary to the first strand, wherein said fungus or oomycete gene is selected from the list consisting of:

a) a polynucleotide comprising a sequence as set forth in SEQ ID N0: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43;

b) a polynucleotide encoding a polypeptide having a sequence as set forth in SEQ ID N0: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44;

c) a polynucleotide having at least 70% sequence identity, preferably at least 80%, more preferably at least 90%, even more preferably at least 95% to a polynucleotide having a sequence as set forth in SEQ ID N0: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43;

- d) a polynucleotide encoding a polypeptide having at least 70% sequence identity, preferably at least 80%, more preferably at least 90%, even more preferably at least 95%, to a polypeptide having a sequence as set forth in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44:
- 5 e) a polynucleotide hybridizing under stringent conditions to a polynucleotide having a sequence as set forth in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43; and
- f) a polynucleotide hybridizing under stringent conditions to a polynucleotide encoding a polypeptide having at least 70% sequence identity to a polypeptide having a sequence as set forth in SEQ ID NO:
- 10 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44.

In accordance with the present invention, the term "identity" is to be understood to mean the number of amino acids/nucleotides corresponding with the amino acids/nucleotides of other protein/nucleic acid, expressed as a percentage. Identity is preferably determined by comparing the Seq. ID disclosed

15 herein with other protein/nucleic acid with the help of computer programs. If sequences that are compared with one another have different lengths, the identity is to be determined in such a way that the number of amino acids, which have the shorter sequence in common with the longer sequence, determines the percentage quotient of the identity. Preferably, identity is determined by means of the computer program ClustalW, which is well known and available to the public (Thompson et al., 1994).

20 ClustalW is made publicly available on <http://www.ebi.ac.uk/tools/clustalW2/index.html>.

Preferably, Version 2.1 of the ClustalW computer program is used to determine the identity between proteins according to the invention and other proteins. In doing so, the following parameters must be set: KTUPLE=1, TOPDIAG=5, WINDOW=5, PAIRGAP=3, GAOPEN=10, GAPEXTEND=0.05,

25 GAPDIST=8, MAXDIV=40, MATRIX=GONNET, ENDGAPS(OFF), NOPGAP, NOHGAP.

Preferably, Version 2.1 of the ClustalW computer program is used to determine the identity between the nucleotide sequence of the nucleic acid molecules according to the invention, for example, and the nucleotide sequence of other nucleic acid molecules. In doing so, the following parameters must be set:

30 KTUPLE=2, TOPDIAGS=4, PAIRGAP=5, DNAMATRIX:IUB, GAOPEN=10, GAPEXT=5, MAXDIV=40, TRANSITIONS: unweighted.

In accordance with the present invention, the term "hybridizing under stringent conditions" refers to polynucleotides or nucleic acid sequences which hybridize with a reference nucleic acid sequence at a

35 level significantly greater than the background noise. The background noise may be associated with the hybridization of other DNA sequences present, in particular of other cDNAs present in a cDNA library. The level of the signal generated by the interaction between the sequence capable of selectively hybridizing and the sequences defined by the sequence IDs above according to the invention is generally 10 times, preferably 100 times, greater than that of the interaction of the other

40 DNA sequences generating the background noise. The level of interaction can be measured, for

example, by labeling the probe with radioactive elements such as  $^{32}\text{P}$ . The selective hybridization is generally obtained by using very severe conditions for the medium (for example 0.03 M NaCl and 0.03 M sodium citrate at approximately 50°C-60°C). The hybridization can of course be carried out according to the usual methods of the state of the art (in particular Sambrook et al., 2001, Molecular Cloning: A Laboratory Manual, third edition).

In a particular embodiment of the invention, the dsRNA molecule is applied on the plant pathogen, particularly fungi or oomycete, and/or on the plant or crop to be protected. The present invention therefore also relates to a composition comprising an effective and non-phytotoxic amount of a dsRNA molecule as defined herein.

dsRNA molecules accordingly to the invention may be made by classical chemical synthesis, by means of *in vitro* transcription or produced in organisms like animals cells, bacteria, yeasts, or plants by heterologous expression (Aalto A.P. et al, 2007 RNA. 13(3):422-9.)

The present invention therefore relates to a micro-organism producing a dsRNA molecule as herein defined.

The present invention also relates to a genetic construct which comprises at least one DNA sequence as well as heterologous regulatory element(s) in the 5' and optionally in the 3' positions, characterized in that the DNA sequence is able to form a dsRNA molecule as herein defined.

The present invention also relates to a cloning and/or expression vector, characterized in that it contains at least one genetic construct as herein defined.

The expression "effective and non-phytotoxic amount" means an amount of composition according to the invention which is sufficient to control or destroy the pathogen present or liable to appear on the crops and which does not entail any appreciable symptom of phytotoxicity for the said crops. Such an amount can vary within a wide range depending on the pathogen to be controlled, the type of crop, the climatic conditions and the compounds included in the composition according to the invention. This amount can be determined by systematic field trials, which are within the capabilities of a person skilled in the art.

Thus, according to the invention, there is provided a composition comprising, as an active ingredient, an effective amount of a dsRNA molecule as herein defined and an agriculturally acceptable support, carrier, filler and/or surfactant.

According to the invention, the term "support" denotes a natural or synthetic organic or inorganic compound with which the active compound of formula (I) is combined or associated to make it easier to apply, notably to the parts of the plant. This support is thus generally inert and should be agriculturally acceptable. The support can be a solid or a liquid. Examples of suitable supports include

clays, natural or synthetic silicates, silica, resins, waxes, solid fertilisers, water, alcohols, in particular butanol organic solvents, mineral and plant oils and derivatives thereof. Mixtures of such supports can also be used.

5 The composition according to the invention can also comprise additional components such as, but not limited to, surfactant, protective colloids, adhesives, thickeners, thixotropic agents, penetration agents, stabilisers, sequestering agents. More generally, the active compounds can be combined with any solid or liquid additive, which complies with the usual formulation techniques.

10 In general, the composition according to the invention can contain from 0.05 to 99% by weight of active compound, preferably 10 to 70% by weight.

Compositions according to the invention can be used in various forms such as aerosol dispenser, capsule suspension, cold fogging concentrate, dustable powder, emulsifiable concentrate, emulsion oil  
15 in water, emulsion water in oil, encapsulated granule, fine granule, flowable concentrate for seed treatment, gas (under pressure), gas generating product, granule, hot fogging concentrate, macrogranule, microgranule, oil dispersible powder, oil miscible flowable concentrate, oil miscible liquid, paste, plant rodlet, powder for dry seed treatment, seed coated with a pesticide, soluble concentrate, soluble powder, solution for seed treatment, suspension concentrate (flowable  
20 concentrate), ultra low volume (ULV) liquid, ultra low volume (ULV) suspension, water dispersible granules or tablets, water dispersible powder for slurry treatment, water soluble granules or tablets, water soluble powder for seed treatment and wettable powder. These compositions include not only compositions which are ready to be applied to the plant or seed to be treated by means of a suitable device, such as a spraying or dusting device, but also concentrated commercial compositions which  
25 must be diluted before application to the crop.

The compounds according to the invention can also be mixed with one or more phytopharmaceutical or plant growth promoting compound, such as a fungicide, herbicide, insecticide, nematocide, acaricide, molluscicide, resistance inducer, safeners, signal compounds, biologicals, pheromone active  
30 substance or other compounds with biological activity. The mixtures thus obtained have a broadened spectrum of activity. The mixtures with other fungicide compounds are particularly advantageous.

In a particular embodiment of the invention, the dsRNA is introduced or produced into the plant to be protected. After introduction or production into the plant, the dsRNA may further be processed into  
35 relatively small fragments (siRNAs) and can subsequently become distributed throughout the plant. Alternatively, the dsRNA is introduced or produced into the plant using a regulatory element or promoter that results in expression of the dsRNA in a tissue, temporal, spatial or inducible manner and may further be processed into relatively small fragments by a plant cell containing the RNAi processing machinery.

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The invention therefore relates to a genetic construct or chimeric gene which is able to produce the dsRNA of the invention inside a plant cell. Said genetic construct or chimeric gene comprises at least one DNA sequence as well as well as heterologous regulatory element(s) in the 5' and optionally in the 3' positions which are able to function in a plant, characterized in that the DNA sequence(s) is (are) able to form a dsRNA molecule as herein defined once expressed in the plant.

In a particular embodiment, the genetic construct or chimeric gene comprises:

- a promoter regulatory sequence that is functional in plant cells, operably linked to
- a DNA sequence which, when it is transcribed, generates an RNA molecule comprising at least a sense sequence and an antisense sequence which are at least partially complementary, said sense sequence comprising a sequence substantially identical to at least 18 contiguous nucleotides of a target gene (i.e in the meaning of the invention a saccharopine dehydrogenase gene), and said antisense sequence comprising a sequence substantially complementary to the sense sequence, and
- optionally a terminator regulatory sequence.

In said embodiment, the DNA sequence according to the invention may have more particularly two aspects; in the first, it comprises two nucleotide sequences, which are sense and antisense, separated by a spacer nucleotide sequence or an intron that does not exhibit any homology with the target gene. The sequence cloned in the sense and antisense orientation is that whose expression in the pathogen it is intended to inhibit. The transcription of this DNA sequence thus gives a large single-stranded RNA corresponding to the sense/spacer-intron/antisense construct. This long RNA transcript can be detected by RT-PCR. Since the sense and antisense sequences are homologous, they will pair, and the spacer or intron which separates them plays the role of a loop for folding. A dsRNA is then obtained over all the homologous regions. The dsRNA is subsequently specifically degraded by an enzymatic complex called "DICER". The degradation of the dsRNAs then forms siRNAs ("siRNA" in the figure), small double-stranded RNAs having a size of between 19 and 25 bases. These are then the siRNAs which, by pairing with the transcribed RNAs derived from the target gene will lead to their degradation via the RNA silencing machinery enzymatic machinery.

In the second aspect, the DNA sequence comprises two nucleotide sequences, which are sense and antisense, of different sizes, the loop structure corresponding to the part of the nucleotide sequence that does not exhibit any homology with the other nucleotide sequence. The nucleotide sequence cloned in the sense orientation is essentially homologous to the sequence of the target gene whose expression it is intended to inhibit. The antisense nucleotide sequence is essentially homologous to the complementary strand of the sequence of said target gene. The transcription of this DNA sequence thus gives a large single-stranded RNA corresponding to the sense/antisense construct. This long RNA transcript can be detected by RT-PCR. The homologous sense/antisense sequences are paired. A dsRNA is then obtained over all the homologous regions. The dsRNA is subsequently specifically degraded by an enzymatic complex called "DICER". The degradation of the dsRNAs then forms siRNAs ("siRNA" in the figure), small doubled-stranded RNAs having a size of between 18 and

25 bases. These are then the siRNAs which, by pairing with the target RNAs, will lead to their degradation via the RNA silencing machinery plant's enzymatic machinery.

In another particular embodiment, the genetic construct comprises:

- 5 - two promoter regulatory sequences that are functional in plant cells, wherein the first promoter regulatory sequence is operably linked to a DNA sequence which, when it is transcribed, generates an RNA molecule comprising at least a sense sequence, and the second promoter regulatory sequence is operably linked to a DNA sequence which, when it is transcribed, generates an RNA molecule comprising at least an antisense sequence partially complementary to the sense sequence, and
- 10 wherein said sense sequence comprises a sequence substantially identical to at least 18 contiguous nucleotides of the target gene, and
- optionally terminator regulatory sequence(s).

In this particular embodiment, the genetic construct may be comprised as two chimeric genes, one comprising the first promoter regulatory sequence operably linked to the first DNA sequence which, when it is transcribed, generates an RNA molecule comprising at least a sense sequence substantially identical to at least 18 contiguous nucleotides of the target gene, and optionally a terminator regulatory sequence, and the second chimeric genes comprising the second promoter regulatory sequence operably linked to the second DNA sequence which, when it is transcribed, generates an RNA molecule comprising at least an antisense sequence partially complementary to the sense sequence, and optionally a terminator regulatory sequence.

These two chimeric genes are preferably introduced into the plant cell conjointly, but not necessary, in order to favorize the hybridization of the two RNA single strands to form the dsRNA.

Alternatively, the genetic construct may be comprised as a construct comprising:

- 25 - a first promoter, operably linked to
- a double strand DNA sequence wherein one strand when it is transcribed under the control of the first promoter, generates an RNA molecule comprising at least a sense sequence substantially identical to at least 18 contiguous nucleotides of the target gene, and optionally a terminator regulatory sequence, and wherein the second strand, when it is transcribed under the control of the
- 30 second promoter, generates an RNA molecule comprising at least an antisense sequence partially complementary to the sense sequence, and optionally a terminator regulatory sequence, and
- a second promoter, in the opposite direction that the first one.

First and second promoter regulatory sequences may be different or identical, preferably different.

The invention further relates to a cloning and/or expression vector for transforming a plant, characterized in that it contains at least one chimeric gene or genetic construct as defined herein.

The present invention further relates to a transgenic plant cell containing the dsRNA molecule of the invention and as herein defined.

The present invention therefore relates to a transgenic plant cell containing the genetic construct or chimeric gene of the invention as herein defined.

5 In a particular embodiment of the invention, the transgenic plant cell is a soybean, oilseed, rice or potato plant cell.

The present invention further relates to a transgenic plant, seed or part thereof, comprising a transgenic plant cell according to the invention.

10 In a particular embodiment of the invention, the transgenic plant, seed or part thereof, is a soybean, oilseed, rice or potato plant, seed or part thereof.

15 According to the invention, the expression "chimeric gene" or "expression cassette" is intended to mean a nucleotide sequence comprising, functionally linked to one another in the direction of transcription, a regulatory promoter sequence that is functional in plants, a sequence encoding a protein or an RNA chain, and, optionally, a terminator that is functional in plant cells. The term "chimeric gene" or "expression cassette" is generally intended to mean a gene for which certain elements are not present in the native gene, but have been substituted for elements present in the native gene or have been added.

20 According to the invention, the terms "chimeric gene" or "expression cassette" may also correspond to the case where all the elements of the gene are present in the native gene, and alternatively, the term "gene" may correspond to a chimeric gene.

25 The expression "chimeric gene" or "expression cassette" may also correspond to the case where the sequence encoding a protein or a RNA chain is not directly linked to a promoter, but is part, for example, of a polycistronic construct comprising several coding sequences under the control of the same promoter. In that case, each coding sequences under the control of the promoter is designed as a "chimeric gene" or "expression cassette".

30 According to the invention, the expression "functionally linked to one another" means that said elements of the elemental chimeric gene are linked to one another in such a way that their function is coordinated and allows the expression of the coding sequence. By way of example, a promoter is functionally linked to a coding sequence when it is capable of ensuring the expression of said coding sequence. The construction of the chimeric gene according to the invention and the assembly of its various elements can be carried out using techniques well known to those skilled in the art, in particular those described in Sambrook et al. (2001, Molecular Cloning : A Laboratory Manual (third edition), Nolan C. ed., New York: Cold Spring Harbor Laboratory Press). The choice of the regulatory elements constituting the chimeric gene depends essentially on the plant and on the type of cell in which they must function, and those skilled in the art are capable of selecting regulatory elements that are functional in a given plant.

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According to the invention, the term "promoter regulatory sequence" is intended to mean any promoter regulatory sequence of a gene that is naturally expressed in plants, in particular a promoter that is expressed especially in the leaves of plants, for instance promoters referred to as constitutive of bacterial, viral or plant origin, or else promoters referred to as light-dependent, such as that of a plant ribulose-biscarboxylase/oxygenase (RuBisCO) small subunit gene, or any known suitable promoter that can be used. Among the promoters of plant origin, mention will be made of the histone promoters as described in application EP 0 507 698, or the rice actin promoter (US 5,641,876). Among the promoters of a plant virus gene, mention will be made of that of the cauliflower mosaic virus (CaMV 19S or 35S) or of the cassava vein mosaic virus (CsVMV: WO97/48819) or the circovirus promoter (AU 689 311). Use may also be made of a promoter regulatory sequence specific for particular regions or tissues of plants, and more particularly seed-specific promoters (Datla, R. *et al.*, 1997, Biotechnology Ann. Rev., 3, 269-296), especially the napin (EP 255 378), phaseolin, glutenin, helianthinin (WO 92/17580), albumin (WO 98/45460) and oleosin (WO 98/45461) promoters. An inducible promoter can also be used, it can be advantageously chosen from the promoters of phenylalanine ammonia lyase (PAL), of HMG-CoA reductase (HMG), of chitinases, of glucanases, of proteinase inhibitors (PI), of genes of the PR1 family, of nopaline synthase (nos) or of the vspB gene (US 5 670 349), the HMG2 promoter (US 5 670 349), the apple beta-galactosidase (ABG1) promoter or the apple amino cyclopropane carboxylate synthase (ACC synthase) promoter (WO 98/45445).

The term "terminator regulatory sequence" is intended to mean any sequence that is functional in plant cells or plants, also comprising polyadenylation sequences, whether they are of bacterial origin, for instance the nos or ocs terminator of *Agrobacterium tumefaciens*, of viral origin, for instance the CaMV 35S terminator, or else of plant origin, for instance a histone terminator as described in application EP 0 633 317.

The selection step for identifying the transformed cells and/or plants having integrated the construct according to the invention can be carried out by virtue of the presence of a selectable gene present in the construct according to the invention or in the plasmid used for the transformation of the cells or of the plants and comprising said construct. The selectable gene may be in the form of a chimeric gene comprising the following elements, functionally linked in the direction of transcription: a promoter regulatory sequence that is functional in plant cells, a sequence encoding a selectable marker, and a terminator regulatory sequence that is functional in plant cells.

Among the selectable markers that can be used, mention may be made of markers containing genes for resistance to antibiotics, such as, for example, that of the hygromycin phosphotransferase gene (Gritz *et al.*, 1983, Gene 25: 179-188), of the neomycin phosphotransferase II gene inducing resistance to kanamycin (Wirtz *et al.*, 1987, DNA, 6(3): 245-253), or of the aminoglycoside 3"-adenyltransferase gene, but also markers containing genes for tolerance to herbicides, such as the *bar* gene (White *et al.*, NAR 18: 1062, 1990) for tolerance to bialaphos, the EPSPS gene (US 5,188,642) for tolerance to glyphosate or else the HPPD gene (WO 96/38567) for tolerance to

isoxazoles. Mention may also be made of genes encoding readily indentifiable enzymes, such as the GUS enzyme, GFP protein or genes encoding pigments or enzymes regulating pigment production in the transformed cells. Such selectable marker genes are in particular described in patent applications WO 91/02071, WO 95/06128, WO 96/38567, and WO 97/04103.

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The present invention further relates to a method of making a transgenic plant cell or plant capable of expressing a dsRNA that inhibits a fungus or oomycete saccharopine dehydrogenase gene, wherein said method comprises the steps of transforming a plant cell with a chimeric gene or genetic construct according to the invention.

10 The method may further comprise the step of selecting the plant cell which has been transformed.

In a particular embodiment of the invention, the invention relates to a method of making a transgenic plant cell or plant capable of expressing a dsRNA that inhibits fungus or oomycete saccharopine dehydrogenase gene , wherein said method comprises the steps of transforming a plant cell with a chimeric gene or genetic construct according to the invention, and wherein said plant cell is a soybean, oilseed, rice or potato plant cell or said plant is a soybean, oilseed, rice or potato plant.

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The present invention also relates to the transformed plants or part thereof, and to plants or part thereof which are derived by cultivating and/or crossing the above regenerated plants, and to the seeds of the transformed plants.

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The present invention also relates to the end products such as the meal, oil, fiber which are obtained from the plants, part thereof, or seeds of the invention.

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To obtain the cells or plants according to the invention, those skilled in the art can use one of the numerous known methods of transformation.

One of these methods consists in bringing the cells or tissues of the host organisms to be transformed into contact with polyethylene glycol (PEG) and the vectors of the invention (Chang and Cohen, 1979, Mol. Gen. Genet. 168(1), 111-115; Mercenier and Chassy, 1988, Biochimie 70(4), 503-517). Electroporation is another method, which consists in subjecting the cells or tissues to be transformed and the vectors of the invention to an electric field (Andreason and Evans, 1988, Biotechniques 6(7), 650-660; Shigekawa and Dower, 1989, Aust. J. Biotechnol. 3(1), 56-62). Another method consists in directly injecting the vectors into the cells or the tissues by microinjection (Gordon and Ruddle, 1985, Gene 33(2), 121-136). Advantageously, the "biolistic" method may be used. It consists in bombarding cells or tissues with particles onto which the vectors of the invention are adsorbed (Bruce *et al.*, 1989, Proc. Natl. Acad. Sci. USA 86(24), 9692-9696; Klein *et al.*, 1992, Biotechnology 10(3), 286-291; US Patent No. 4,945,050). Preferably, the transformation of plant cells or tissues can be carried out using bacteria of the *Agrobacterium* genus, preferably by infection of the cells or tissues of said plants with *A. tumefaciens* (Knopf, 1979, Subcell. Biochem. 6, 143-173; Shaw *et al.*, 1983, Gene 23(3): 315-330) or *A. rhizogenes* (Bevan and Chilton, 1982, Annu. Rev. Genet. 16: 357-384; Tepfer and Casse-Delbart, 1987, Microbiol. Sci. 4(1), 24-28). Preferably, the transformation of plant cells or tissues with

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*Agrobacterium tumefaciens* is carried out according to the protocol described by Hiei *et al.*, (1994, Plant J. 6(2): 271-282). Those skilled in the art will choose the appropriate method according to the nature of the host organisms to be transformed.

- 5 The plants according to the invention contain transformed plant cells as defined above. In particular, the transformed plants can be obtained by regeneration of the transformed plant cells described above. The regeneration is obtained by any appropriate method, which depends on the nature of the species.

10 The invention also comprises parts of these plants, and the progeny of these plants. The term "part of these plants" is intended to mean any organ of these plants, whether above ground or below ground. The organs above ground are the stems, the leaves and the flowers comprising the male and female reproductive organs. The organs below ground are mainly the roots, but they may also be tubers. The term "progeny" is intended to mean mainly the seeds containing the embryos derived from the reproduction of these plants with one another. By extension, the term "progeny" applies to all the

15 seeds formed at each new generation derived from crosses between the transformed plants according to the invention. Progeny and seeds can also be obtained by vegetative multiplication of said transformed plants. The seeds according to the invention can be coated with an agrochemical composition comprising at least one active product having an activity selected from fungicidal, herbicidal, insecticidal, nematocidal, bactericidal or virucidal activities.

20 The invention further relates to a method for controlling a plant pathogen, particularly a fungus or an oomycete, comprising providing to said pathogen a dsRNA molecule according to the invention and as herein defined.

- 25 In a particular embodiment of the invention, the method relates to a method for controlling a plant pathogen, particularly a fungus or an oomycete, comprising providing to said pathogen a dsRNA according to the invention and as herein defined, or a composition comprising said dsRNA, wherein said plant pathogen is *Magnaporthe grisea*, *Phytophthora infestans*, *Sclerotinia sclerotiorum* or *Phakopsora pachyrhizi*.

30 In a particular embodiment of the invention, the method relates to a method for controlling a plant pathogen, particularly a fungus or an oomycete, comprising providing to said pathogen a dsRNA molecule according to the invention and as herein defined, or a composition comprising said dsRNA, wherein said plant is a soybean, oilseed, rice or potato plant.

35 The invention further relates to a method for controlling a plant, crop or seed pathogen, particularly a fungus or an oomycete, characterized in that an agronomically effective and substantially non-phytotoxic quantity of dsRNA molecule according to the invention or composition according to the invention is applied as seed treatment, foliar application, stem application, drench or drip application

40 (chemigation) to the seed, the plant or to the fruit of the plant or to soil or to inert substrate (e.g.

inorganic substrates like sand, rockwool, glasswool; expanded minerals like perlite, vermiculite, zeolite or expanded clay), Pumice, Pyroclastic materials or stuff, synthetic organic substrates (e.g. polyurethane) organic substrates (e.g. peat, composts, tree waste products like coir, wood fibre or chips, tree bark) or to a liquid substrate (e.g. floating hydroponic systems, Nutrient Film Technique, 5 Aeroponics) wherein the plant is growing or wherein it is desired to grow.

The invention therefore relates to a method for controlling a plant pathogen, particularly a fungus or oomycete, characterized in that an effective quantity of a dsRNA molecule according to the invention or a composition according to the invention is applied to the soil where plants grow or are capable of 10 growing, to the leaves and/or the fruit of plants or to the seeds of plants.

In a particular embodiment of the invention, the invention relates to a method for controlling a plant pathogen, particularly a fungus or oomycete, characterized in that an effective quantity of a dsRNA molecule according to the invention or a composition according to the invention is applied to the soil 15 where plants grow or are capable of growing, to the leaves and/or the fruit of plants or to the seeds of plants, wherein said plant pathogen is *Magnaporthe grisea*, *Phytophthora infestans*, *Sclerotinia sclerotinium* or *Phakopsora pachyrhizi*.

In a particular embodiment of the invention, the invention relates to a method for controlling a plant 20 pathogen, particularly a fungus or oomycete, characterized in that an effective quantity of a dsRNA molecule according to the invention or a composition according to the invention is applied to the soil where plants grow or are capable of growing, to the leaves and/or the fruit of plants or to the seeds of plants, wherein said plant is a soybean, oilseed, rice or potato plant.

25 The expression "are applied to the plants to be treated" is understood to mean, for the purposes of the present invention, that the pesticide composition which is the subject of the invention can be applied by means of various methods of treatment such as:

- spraying onto the aerial parts of the said plants a liquid comprising one of the said compositions,
- 30 • dusting, the incorporation into the soil of granules or powders, spraying, around the said plants and in the case of trees injection or daubing,
- coating or film-coating the seeds of the said plants with the aid of a plant-protection mixture comprising one of the said compositions.

The method according to the invention can either be a curing, preventing or eradicating method.

35 In this method, a composition used can be prepared beforehand by mixing the two or more active compounds according to the invention.

According to an alternative of such a method, it is also possible to apply simultaneously, successively or separately compounds (A) and (B) so as to have the conjugated (A)/(B) effects, of distinct compositions each containing one of the two or three active ingredients (A) or (B).

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The dose of active dsRNA compound usually applied in the method of treatment according to the invention is generally and advantageously

- for foliar treatments: from 0.0001 to 10,000 g/ha, preferably from 0.0001 to 1000 g/ha, more preferably from 0.001 to 300g/ha; in case of drench or drip application, the dose can even be reduced, especially while using inert substrates like rockwool or perlite;
- for seed treatment: from 0.0001 to 200 g per 100 kilogram of seed, preferably from 0.001 to 150 g per 100 kilogram of seed;
- for soil treatment: from 0.0001 to 10,000 g/ha, preferably from 0.001 to 5,000 g/ha.

When the dsRNA of the invention is mixed with another active phytopharmaceutical or plant growth promoting compound, said phytopharmaceutical or plant growth promoting compound is used in the dose usually applied.

Said phytopharmaceutical or plant growth promoting compound may be a fungicide, herbicide, insecticide, nematicide, acaricide, molluscicide, resistance inducer, safeners, or signal compounds.

The dose of phytopharmaceutical active compound usually applied in the method of treatment according to the invention is generally and advantageously from 10 to 800 g/ha, preferably from 50 to 300 g/ha for applications in foliar treatment. The dose of active substance applied is generally and advantageously from 2 to 200 g per 100 kg of seed, preferably from 3 to 150 g per 100 kg of seed in the case of seed treatment.

The doses herein indicated are given as illustrative Examples of method according to the invention. A person skilled in the art will know how to adapt the application doses, notably according to the nature of the plant or crop to be treated.

Under specific conditions, for example according to the nature of the pathogen, phytopathogenic fungus or oomycete to be treated or controlled, a lower dose can offer adequate protection. Certain climatic conditions, resistance or other factors like the nature of the pathogen or the degree of infestation, for example, of the plants with these pathogens, can require higher doses of combined active ingredients. The optimum dose usually depends on several factors, for example on the type of pathogen to be treated, on the type or level of development of the infested plant or plant material, on the density of vegetation or alternatively on the method of application.

Without it being limiting, the crop treated with the pesticide composition or combination according to the invention is, for example, grapevine, cereals, vegetables, lucerne, soybean, market garden crops, turf, wood, tree or horticultural plants.

The method of treatment according to the invention can also be useful to treat propagation material such as tubers or rhizomes, but also seeds, seedlings or seedlings pricking out and plants or plants pricking out. This method of treatment can also be useful to treat roots. The method of treatment according to the invention can also be useful to treat the over-ground parts of the plant such as trunks, stems or stalks, leaves, flowers and fruit of the concerned plant, and in general every material which is susceptible to fungal infection (e.g due to storage like hay)

The invention further relates to a method of controlling a plant pathogen, particularly a fungus or an

oomycete, comprising providing in the host plant of said plant pathogen a transformed plant cell according to the invention.

The invention further relates to a method of controlling a plant pathogen, particularly a fungus or an oomycete, comprising providing in the host plant of said plant pathogen a transformed plant cell

5 containing a dsRNA as herein defined.

The invention further relates to a method of controlling a plant pathogen, particularly a fungus or an oomycete, comprising transforming the plant with a genetic construct according to the invention.

10 The invention further relates to a method of controlling a plant pathogen, particularly a fungus or an oomycete, comprising the following steps:

i) transforming a plant cell with a chimeric gene according to the invention;  
ii) placing the cells thus transformed under conditions that allow the transcription of said construct,

15 iii) having the cells into contact with the pathogen.

In a particular embodiment of the invention, methods according to the invention are controlling a plant pathogen selected from *Magnaporthe grisea*, *Phytophthora infestans*, *Sclerotinia sclerotinium* or *Phakopsora pachyrhizi*.

20 In a particular embodiment of the invention, methods according to the invention are controlling a plant pathogen wherein the plant is a soybean, oilseed, rice or potato plant.

The invention further relates to a method for inhibiting the expression of a plant pathogen, particularly fungus or oomycete, saccharopine dehydrogenase gene, comprising the following steps:

25 i) transforming a plant cell with a chimeric gene according to the invention;  
ii) placing the cells thus transformed under conditions that allow the transcription of said construct,

iii) having the cells into contact with the pathogen.

30 In a particular embodiment of the invention, the method according to the invention is inhibiting a fungal or oomycete saccharopine dehydrogenase gene, wherein the fungus or oomycete is *Magnaporthe grisea*, *Phytophthora infestans*, *Sclerotinia sclerotinium* or *Phakopsora pachyrhizi*.

In a particular embodiment of the invention, the method according to the invention inhibites a fungal or oomycete saccharopine dehydrogenase gene, said method comprises the following steps:

35 i) transforming a plant cell with a chimeric gene according to the invention;  
ii) placing the cells thus transformed under conditions that allow the transcription of said construct,

- iii) having the cells into contact with the pathogen;  
wherein the plant is a soybean, oilseed, rice or potato plant.

According to the invention all plants and plant parts can be treated. By plants is meant all plants and plant populations such as desirable and undesirable wild plants, cultivars and plant varieties (whether or not protectable by plant variety or plant breeder's rights). Cultivars and plant varieties can be plants obtained by conventional propagation and breeding methods which can be assisted or supplemented by one or more biotechnological methods such as by use of double haploids, protoplast fusion, random and directed mutagenesis, molecular or genetic markers or by bioengineering and genetic engineering methods. By plant parts is meant all above ground and below ground parts and organs of plants such as shoot, leaf, blossom and root, whereby for example leaves, needles, stems, branches, blossoms, fruiting bodies, fruits and seed as well as roots, corms and rhizomes are listed. Crops and vegetative and generative propagating material, for example cuttings, corms, rhizomes, runners and seeds also belong to plant parts.

Among the plants that can be protected by the method according to the invention, mention may be made of major field crops like corn, soybean, cotton, *Brassica* oilseeds such as *Brassica napus* (e.g. canola), *Brassica rapa*, *B. juncea* (e.g. mustard) and *Brassica carinata*, rice, wheat, sugarbeet, sugarcane, oats, rye, barley, millet, triticale, flax, vine and various fruits and vegetables of various botanical taxa such as *Rosaceae* sp. (for instance pip fruit such as apples and pears, but also stone fruit such as apricots, cherries, almonds and peaches, berry fruits such as strawberries), *Ribesioideae* sp., *Juglandaceae* sp., *Betulaceae* sp., *Anacardiaceae* sp., *Fagaceae* sp., *Moraceae* sp., *Oleaceae* sp., *Actinidaceae* sp., *Lauraceae* sp., *Musaceae* sp. (for instance banana trees and plantings), *Rubiaceae* sp. (for instance coffee), *Theaceae* sp., *Sterculiaceae* sp., *Rutaceae* sp. (for instance lemons, oranges and grapefruit); *Solanaceae* sp. (for instance tomatoes, potatoes, peppers, eggplant), *Liliaceae* sp., *Compositae* sp. (for instance lettuce, artichoke and chicory - including root chicory, endive or common chicory), *Umbelliferae* sp. (for instance carrot, parsley, celery and celeriac), *Cucurbitaceae* sp. (for instance cucumber - including pickling cucumber, squash, watermelon, gourds and melons), *Alliaceae* sp. (for instance onions and leek), *Cruciferae* sp. (for instance white cabbage, red cabbage, broccoli, cauliflower, brussels sprouts, pak choi, kohlrabi, radish, horseradish, cress, Chinese cabbage), *Leguminosae* sp. (for instance peanuts, peas and beans - such as climbing beans and broad beans), *Chenopodiaceae* sp. (for instance mangold, spinach beet, spinach, beetroots), *Malvaceae* (for instance okra), *Asparagaceae* (for instance asparagus); horticultural and forest crops; ornamental plants; as well as genetically modified homologues of these crops.

The method of treatment according to the invention can be used in the treatment of genetically modified organisms (GMOs), e.g. plants or seeds. Genetically modified plants (or transgenic plants) are plants of which a heterologous gene has been stably integrated into genome. The expression "heterologous gene" essentially means a gene which is provided or assembled outside the plant and when introduced in the nuclear, chloroplastic or mitochondrial genome gives the transformed plant new or improved agronomic or other properties by expressing a protein or polypeptide of interest or by downregulating or silencing other gene(s) which are present in the plant (using for example, antisense technology, cosuppression technology

or RNA interference – RNAi - technology). A heterologous gene that is located in the genome is also called a transgene. A transgene that is defined by its particular location in the plant genome is called a transformation or transgenic event.

Depending on the plant species or plant cultivars, their location and growth conditions (soils, climate, vegetation period, diet), the treatment according to the invention may also result in superadditive ("synergistic") effects. Thus, for example, reduced application rates and/or a widening of the activity spectrum and/or an increase in the activity of the active compounds and compositions which can be used according to the invention, better plant growth, increased tolerance to high or low temperatures, increased tolerance to drought or to water or soil salt content, increased flowering performance, easier harvesting, accelerated maturation, higher harvest yields, bigger fruits, larger plant height, greener leaf color, earlier flowering, higher quality and/or a higher nutritional value of the harvested products, higher sugar concentration within the fruits, better storage stability and/or processability of the harvested products are possible, which exceed the effects which were actually to be expected.

At certain application rates, the active compound combinations according to the invention may also have a strengthening effect in plants. Accordingly, they are also suitable for mobilizing the defense system of the plant against attack by unwanted microorganisms. This may, if appropriate, be one of the reasons of the enhanced activity of the combinations according to the invention, for example against fungi. Plant-strengthening (resistance-inducing) substances are to be understood as meaning, in the present context, those substances or combinations of substances which are capable of stimulating the defense system of plants in such a way that, when subsequently inoculated with unwanted microorganisms, the treated plants display a substantial degree of resistance to these microorganisms. In the present case, unwanted microorganisms are to be understood as meaning phytopathogenic fungi, bacteria and viruses. Thus, the substances according to the invention can be employed for protecting plants against attack by the abovementioned pathogens within a certain period of time after the treatment. The period of time within which protection is effected generally extends from 1 to 10 days, preferably 1 to 7 days, after the treatment of the plants with the active compounds.

As already mentioned above, it is possible to treat all plants and their parts in accordance with the invention. In a preferred embodiment, wild plant species and plant cultivars, or those obtained by conventional biological breeding methods, such as crossing or protoplast fusion, and also parts thereof, are treated. In a further preferred embodiment, transgenic plants and plant cultivars obtained by genetic engineering methods, if appropriate in combination with conventional methods (Genetically Modified Organisms), and parts thereof are treated. The terms "parts" or "parts of plants" or "plant parts" have been explained above. More preferably, plants of the plant cultivars which are commercially available or are in use are treated in accordance with the invention. Plant cultivars are understood to mean plants which have new properties ("traits") and have been obtained by conventional breeding, by mutagenesis or by recombinant DNA techniques. They can be cultivars, varieties, bio- or genotypes.

The method of treatment according to the invention can be used in the treatment of genetically modified organisms (GMOs), e.g. plants or seeds. Genetically modified plants (or transgenic plants)

are plants of which a heterologous gene has been stably integrated into genome. The expression "heterologous gene" essentially means a gene which is provided or assembled outside the plant and when introduced in the nuclear, chloroplastic or mitochondrial genome gives the transformed plant new or improved agronomic or other properties by expressing a protein or polypeptide of interest or by downregulating or silencing other gene(s) which are present in the plant (using for example, antisense technology, cosuppression technology, RNA interference – RNAi – technology or microRNA – miRNA - technology). A heterologous gene that is located in the genome is also called a transgene. A transgene that is defined by its particular location in the plant genome is called a transformation or transgenic event.

- 10 Depending on the plant species or plant cultivars, their location and growth conditions (soils, climate, vegetation period, diet), the treatment according to the invention may also result in superadditive ("synergistic") effects. Thus, for example, reduced application rates and/or a widening of the activity spectrum and/or an increase in the activity of the active compounds and compositions which can be used according to the invention, better plant growth, increased tolerance to high or low temperatures, increased tolerance to drought or to water or soil salt content, increased flowering performance, easier harvesting, accelerated maturation, higher harvest yields, bigger fruits, larger plant height, greener leaf color, earlier flowering, higher quality and/or a higher nutritional value of the harvested products, higher sugar concentration within the fruits, better storage stability and/or processability of the harvested products are possible, which exceed the effects which were actually to be expected.
- 20 At certain application rates, the active compound combinations according to the invention may also have a strengthening effect in plants. Accordingly, they are also suitable for mobilizing the defense system of the plant against attack by unwanted microorganisms. This may, if appropriate, be one of the reasons of the enhanced activity of the combinations according to the invention, for example against fungi. Plant-strengthening (resistance-inducing) substances are to be understood as meaning, in the present context, those substances or combinations of substances which are capable of stimulating the defense system of plants in such a way that, when subsequently inoculated with unwanted microorganisms, the treated plants display a substantial degree of resistance to these microorganisms. In the present case, unwanted microorganisms are to be understood as meaning phytopathogenic fungi, bacteria and viruses. Thus, the substances according to the invention can be employed for protecting plants against attack by the abovementioned pathogens within a certain period of time after the treatment. The period of time within which protection is effected generally extends from 1 to 10 days, preferably 1 to 7 days, after the treatment of the plants with the active compounds.

- 35 Plants and plant cultivars which are preferably to be treated according to the invention include all plants which have genetic material which impart particularly advantageous, useful traits to these plants (whether obtained by breeding and/or biotechnological means).

- Plants and plant cultivars which are also preferably to be treated according to the invention are resistant against one or more biotic stresses, i.e. said plants show a better defense against animal and microbial pests, such as against nematodes, insects, mites, phytopathogenic fungi, bacteria, viruses and/or viroids.
- 40

Examples of nematode or insect resistant plants are described in e.g. U.S. Patent Applications 11/765,491, 11/765,494, 10/926,819, 10/782,020, 12/032,479, 10/783,417, 10/782,096, 11/657,964, 12/192,904, 11/396,808, 12/166,253, 12/166,239, 12/166,124, 12/166,209, 11/762,886, 12/364,335, 11/763,947, 12/252,453, 12/209,354, 12/491,396, 12/497,221, 12/644,632, 12/646,004, 12/701,058, 12/718,059, 12/721,595, 12/638,591.

Plants and plant cultivars which may also be treated according to the invention are those plants which are resistant to one or more abiotic stresses. Abiotic stress conditions may include, for example, drought, cold temperature exposure, heat exposure, osmotic stress, flooding, increased soil salinity, increased mineral exposure, ozone exposure, high light exposure, limited availability of nitrogen nutrients, limited availability of phosphorus nutrients, shade avoidance.

Plants and plant cultivars which may also be treated according to the invention, are those plants characterized by enhanced yield characteristics. Increased yield in said plants can be the result of, for example, improved plant physiology, growth and development, such as water use efficiency, water retention efficiency, improved nitrogen use, enhanced carbon assimilation, improved photosynthesis, increased germination efficiency and accelerated maturation. Yield can furthermore be affected by improved plant architecture (under stress and non-stress conditions), including but not limited to, early flowering, flowering control for hybrid seed production, seedling vigor, plant size, internode number and distance, root growth, seed size, fruit size, pod size, pod or ear number, seed number per pod or ear, seed mass, enhanced seed filling, reduced seed dispersal, reduced pod dehiscence and lodging resistance. Further yield traits include seed composition, such as carbohydrate content, protein content, oil content and composition, nutritional value, reduction in anti-nutritional compounds, improved processability and better storage stability.

Plants that may be treated according to the invention are hybrid plants that already express the characteristic of heterosis or hybrid vigor which results in generally higher yield, vigor, health and resistance towards biotic and abiotic stresses). Such plants are typically made by crossing an inbred male-sterile parent line (the female parent) with another inbred male-fertile parent line (the male parent). Hybrid seed is typically harvested from the male sterile plants and sold to growers. Male sterile plants can sometimes (e.g. in corn) be produced by detasseling, i.e. the mechanical removal of the male reproductive organs (or males flowers) but, more typically, male sterility is the result of genetic determinants in the plant genome. In that case, and especially when seed is the desired product to be harvested from the hybrid plants it is typically useful to ensure that male fertility in the hybrid plants is fully restored. This can be accomplished by ensuring that the male parents have appropriate fertility restorer genes which are capable of restoring the male fertility in hybrid plants that contain the genetic determinants responsible for male-sterility. Genetic determinants for male sterility may be located in the cytoplasm. Examples of cytoplasmic male sterility (CMS) were for instance described in Brassica species (WO 92/05251, WO 95/09910, WO 98/27806, WO 05/002324, WO 06/021972 and US 6,229,072). However, genetic determinants for male sterility can also be located in the nuclear genome. Male sterile plants can also be obtained by plant biotechnology methods such as genetic engineering. A particularly useful means of obtaining male-sterile plants is described in WO 89/10396 in which, for example, a ribonuclease such as barnase is selectively expressed in the

tapetum cells in the stamens. Fertility can then be restored by expression in the tapetum cells of a ribonuclease inhibitor such as barstar (e.g. WO 91/02069).

Plants or plant cultivars (obtained by plant biotechnology methods such as genetic engineering) which may be treated according to the invention are herbicide-tolerant plants, i.e. plants made tolerant to one or more given herbicides. Such plants can be obtained either by genetic transformation, or by selection of plants containing a mutation imparting such herbicide tolerance.

Herbicide-resistant plants are for example glyphosate-tolerant plants, i.e. plants made tolerant to the herbicide glyphosate or salts thereof. Plants can be made tolerant to glyphosate through different means. For example, glyphosate-tolerant plants can be obtained by transforming the plant with a gene encoding the enzyme 5-enolpyruvylshikimate-3-phosphate synthase (EPSPS). Examples of such EPSPS genes are the AroA gene (mutant CT7) of the bacterium *Salmonella typhimurium* (Science 1983, 221, 370-371), the CP4 gene of the bacterium *Agrobacterium* sp. (Curr. Topics Plant Physiol. 1992, 7, 139-145), the genes encoding a *Petunia* EPSPS (Science 1986, 233, 478-481), a Tomato EPSPS (J. Biol. Chem. 1988, 263, 4280-4289), or an Eleusine EPSPS (WO 01/66704). It can also be a mutated EPSPS as described in for example EP 0837944, WO 00/66746, WO 00/66747 or WO 02/26995. Glyphosate-tolerant plants can also be obtained by expressing a gene that encodes a glyphosate oxido-reductase enzyme as described in US 5,776,760 and US 5,463,175. Glyphosate-tolerant plants can also be obtained by expressing a gene that encodes a glyphosate acetyl transferase enzyme as described in for example WO 02/036782, WO 03/092360, WO 2005/012515 and WO 2007/024782. Glyphosate-tolerant plants can also be obtained by selecting plants containing naturally-occurring mutations of the above-mentioned genes, as described in for example WO 01/024615 or WO 03/013226. Plants expressing EPSPS genes that confer glyphosate tolerance are described in e.g. U.S. Patent Applications 11/517,991, 10/739,610, 12/139,408, 12/352,532, 11/312,866, 11/315,678, 12/421,292, 11/400,598, 11/651,752, 11/681,285, 11/605,824, 12/468,205, 11/760,570, 11/762,526, 11/769,327, 11/769,255, 11/943801 or 12/362,774. Plants comprising other genes that confer glyphosate tolerance, such as decarboxylase genes, are described in e.g. U.S. Patent Applications 11/588,811, 11/185,342, 12/364,724, 11/185,560 or 12/423,926.

Other herbicide resistant plants are for example plants that are made tolerant to herbicides inhibiting the enzyme glutamine synthase, such as bialaphos, phosphinothricin or glufosinate. Such plants can be obtained by expressing an enzyme detoxifying the herbicide or a mutant glutamine synthase enzyme that is resistant to inhibition, e.g. described in U.S. Patent Application 11/760,602. One such efficient detoxifying enzyme is an enzyme encoding a phosphinothricin acetyltransferase (such as the bar or pat protein from *Streptomyces* species). Plants expressing an exogenous phosphinothricin acetyltransferase are for example described in U.S. Patents 5,561,236; 5,648,477; 5,646,024; 5,273,894; 5,637,489; 5,276,268; 5,739,082; 5,908,810 and 7,112,665.

Further herbicide-tolerant plants are also plants that are made tolerant to the herbicides inhibiting the enzyme hydroxyphenylpyruvate dioxygenase (HPPD). HPPD is an enzyme that catalyze the reaction in which para-hydroxyphenylpyruvate (HPP) is transformed into homogentisate. Plants tolerant to HPPD-inhibitors can be transformed with a gene encoding a naturally-occurring resistant HPPD enzyme, or a gene encoding a mutated or chimeric HPPD enzyme as described in WO 96/38567, WO

99/24585, WO 99/24586, WO 09/144079, WO 02/046387, or US 6,768,044. Tolerance to HPPD-inhibitors can also be obtained by transforming plants with genes encoding certain enzymes enabling the formation of homogentisate despite the inhibition of the native HPPD enzyme by the HPPD-inhibitor. Such plants and genes are described in WO 99/34008 and WO 02/36787. Tolerance of plants to HPPD inhibitors can also be improved by transforming plants with a gene encoding an enzyme having prephenate deshydrogenase (PDH) activity in addition to a gene encoding an HPPD-tolerant enzyme, as described in WO 04/024928. Further, plants can be made more tolerant to HPPD-inhibitor herbicides by adding into their genome a gene encoding an enzyme capable of metabolizing or degrading HPPD inhibitors, such as the CYP450 enzymes shown in WO 2007/103567 and WO 2008/150473.

Still further herbicide resistant plants are plants that are made tolerant to acetolactate synthase (ALS) inhibitors. Known ALS-inhibitors include, for example, sulfonylurea, imidazolinone, triazolopyrimidines, pyrimidinyoxy(thio)benzoates, and/or sulfonylaminocarbonyltriazolinone herbicides. Different mutations in the ALS enzyme (also known as acetohydroxyacid synthase, AHAS) are known to confer tolerance to different herbicides and groups of herbicides, as described for example in Tranel and Wright (Weed Science 2002, 50, 700-712), but also, in U.S. Patents 5,605,011, 5,378,824, 5,141,870, and 5,013,659. The production of sulfonylurea-tolerant plants and imidazolinone-tolerant plants is described in U.S. Patents 5,605,011; 5,013,659; 5,141,870; 5,767,361; 5,731,180; 5,304,732; 4,761,373; 5,331,107; 5,928,937; and 5,378,824; and WO 96/33270. Other imidazolinone-tolerant plants are also described in for example WO 2004/040012, WO 2004/106529, WO 2005/020673, WO 2005/093093, WO 2006/007373, WO 2006/015376, WO 2006/024351, and WO 2006/060634. Further sulfonylurea- and imidazolinone-tolerant plants are also described in for example WO 2007/024782 and U.S. Patent Application 61/288958.

Other plants tolerant to imidazolinone and/or sulfonylurea can be obtained by induced mutagenesis, selection in cell cultures in the presence of the herbicide or mutation breeding as described for example for soybeans in US 5,084,082, for rice in WO 97/41218, for sugar beet in US 5,773,702 and WO 99/057965, for lettuce in US 5,198,599, or for sunflower in WO 01/065922.

Plants or plant cultivars (obtained by plant biotechnology methods such as genetic engineering) which may also be treated according to the invention are insect-resistant transgenic plants, i.e. plants made resistant to attack by certain target insects. Such plants can be obtained by genetic transformation, or by selection of plants containing a mutation imparting such insect resistance.

An "insect-resistant transgenic plant", as used herein, includes any plant containing at least one transgene comprising a coding sequence encoding:

- 1) an insecticidal crystal protein from *Bacillus thuringiensis* or an insecticidal portion thereof, such as the insecticidal crystal proteins listed by Crickmore et al. (1998, Microbiology and Molecular Biology Reviews, 62: 807-813), updated by Crickmore et al. (2005) at the *Bacillus thuringiensis* toxin nomenclature, online at: [http://www.lifesci.sussex.ac.uk/Home/Neil\\_Crickmore/Bt/](http://www.lifesci.sussex.ac.uk/Home/Neil_Crickmore/Bt/)), or insecticidal portions thereof, e.g., proteins of the Cry protein classes Cry1Ab, Cry1Ac, Cry1B, Cry1C, Cry1D, Cry1F, Cry2Ab, Cry3Aa, or Cry3Bb or insecticidal portions thereof (e.g. EP-

- A 1 999 141 and WO 2007/107302), or such proteins encoded by synthetic genes as e.g. described in and U.S. Patent Application 12/249,016 ; or
- 2) a crystal protein from *Bacillus thuringiensis* or a portion thereof which is insecticidal in the presence of a second other crystal protein from *Bacillus thuringiensis* or a portion thereof, such as the binary toxin made up of the Cry34 and Cry35 crystal proteins (Nat. Biotechnol. 2001, 19, 668-72; Applied Environm. Microbiol. 2006, 71, 1765-1774) or the binary toxin made up of the Cry1A or Cry1F proteins and the Cry2Aa or Cry2Ab or Cry2Ae proteins (U.S. Patent Application 12/214,022 and EP-A 2 300 618); or
  - 3) a hybrid insecticidal protein comprising parts of different insecticidal crystal proteins from *Bacillus thuringiensis*, such as a hybrid of the proteins of 1) above or a hybrid of the proteins of 2) above, e.g., the Cry1A.105 protein produced by corn event MON89034 (WO 2007/027777); or
  - 4) a protein of any one of 1) to 3) above wherein some, particularly 1 to 10, amino acids have been replaced by another amino acid to obtain a higher insecticidal activity to a target insect species, and/or to expand the range of target insect species affected, and/or because of changes introduced into the encoding DNA during cloning or transformation, such as the Cry3Bb1 protein in corn events MON863 or MON88017, or the Cry3A protein in corn event MIR604; or
  - 5) an insecticidal secreted protein from *Bacillus thuringiensis* or *Bacillus cereus*, or an insecticidal portion thereof, such as the vegetative insecticidal (VIP) proteins listed at:  
[http://www.lifesci.sussex.ac.uk/home/Neil\\_Crickmore/Bt/vip.html](http://www.lifesci.sussex.ac.uk/home/Neil_Crickmore/Bt/vip.html), e.g., proteins from the VIP3Aa protein class; or
  - 6) a secreted protein from *Bacillus thuringiensis* or *Bacillus cereus* which is insecticidal in the presence of a second secreted protein from *Bacillus thuringiensis* or *B. cereus*, such as the binary toxin made up of the VIP1A and VIP2A proteins (WO 94/21795); or
  - 7) a hybrid insecticidal protein comprising parts from different secreted proteins from *Bacillus thuringiensis* or *Bacillus cereus*, such as a hybrid of the proteins in 1) above or a hybrid of the proteins in 2) above; or
  - 8) a protein of any one of 5) to 7) above wherein some, particularly 1 to 10, amino acids have been replaced by another amino acid to obtain a higher insecticidal activity to a target insect species, and/or to expand the range of target insect species affected, and/or because of changes introduced into the encoding DNA during cloning or transformation (while still encoding an insecticidal protein), such as the VIP3Aa protein in cotton event COT102; or
  - 9) a secreted protein from *Bacillus thuringiensis* or *Bacillus cereus* which is insecticidal in the presence of a crystal protein from *Bacillus thuringiensis*, such as the binary toxin made up of VIP3 and Cry1A or Cry1F (U.S. Patent Applications 61/126083 and 61/195019), or the binary

toxin made up of the VIP3 protein and the Cry2Aa or Cry2Ab or Cry2Ae proteins (U.S. Patent Application 12/214,022 and EP-A 2 300 618).

- 10) a protein of 9) above wherein some, particularly 1 to 10, amino acids have been replaced by another amino acid to obtain a higher insecticidal activity to a target insect species, and/or to expand the range of target insect species affected, and/or because of changes introduced into the encoding DNA during cloning or transformation (while still encoding an insecticidal protein)

Of course, an insect-resistant transgenic plant, as used herein, also includes any plant comprising a combination of genes encoding the proteins of any one of the above classes 1 to 10. In one embodiment, an insect-resistant plant contains more than one transgene encoding a protein of any one of the above classes 1 to 10, to expand the range of target insect species affected when using different proteins directed at different target insect species, or to delay insect resistance development to the plants by using different proteins insecticidal to the same target insect species but having a different mode of action, such as binding to different receptor binding sites in the insect.

An "insect-resistant transgenic plant", as used herein, further includes any plant containing at least one transgene comprising a sequence producing upon expression a double-stranded RNA which upon ingestion by a plant insect pest inhibits the growth of this insect pest, as described e.g. in WO 2007/080126, WO 2006/129204, WO 2007/074405, WO 2007/080127 and WO 2007/035650.

Plants or plant cultivars (obtained by plant biotechnology methods such as genetic engineering) which may also be treated according to the invention are tolerant to abiotic stresses. Such plants can be obtained by genetic transformation, or by selection of plants containing a mutation imparting such stress resistance. Particularly useful stress tolerance plants include:

- 1) plants which contain a transgene capable of reducing the expression and/or the activity of poly(ADP-ribose) polymerase (PARP) gene in the plant cells or plants as described in WO 00/04173, WO 2006/045633, EP-A 1 807 519, or EP-A 2 018 431.
- 2) plants which contain a stress tolerance enhancing transgene capable of reducing the expression and/or the activity of the PARG encoding genes of the plants or plants cells, as described e.g. in WO 2004/090140.
- 3) plants which contain a stress tolerance enhancing transgene coding for a plant-functional enzyme of the nicotineamide adenine dinucleotide salvage synthesis pathway including nicotinamidase, nicotinate phosphoribosyltransferase, nicotinic acid mononucleotide adenylyl transferase, nicotinamide adenine dinucleotide synthetase or nicotine amide phosphorybosyltransferase as described e.g. in EP-A 1 794 306, WO 2006/133827, WO 2007/107326, EP-A 1 999 263, or WO 2007/107326.

Plants or plant cultivars (obtained by plant biotechnology methods such as genetic engineering) which may also be treated according to the invention show altered quantity, quality and/or storage-stability of the harvested product and/or altered properties of specific ingredients of the harvested product such as:

- 1) transgenic plants which synthesize a modified starch, which in its physical-chemical characteristics, in particular the amylose content or the amylose/amylopectin ratio, the degree of branching, the average chain length, the side chain distribution, the viscosity behaviour, the gelling strength, the starch grain size and/or the starch grain morphology, is changed in comparison with the synthesised starch in wild type plant cells or plants, so that this is better suited for special applications. Said transgenic plants synthesizing a modified starch are disclosed, for example, in EP-A 0 571 427, WO 95/04826, EP-A 0 719 338, WO 96/15248, WO 96/19581, WO 96/27674, WO 97/11188, WO 97/26362, WO 97/32985, WO 97/42328, WO 97/44472, WO 97/45545, WO 98/27212, WO 98/40503, WO 99/58688, WO 99/58690, WO 99/58654, WO 00/08184, WO 00/08185, WO 00/08175, WO 00/28052, WO 00/77229, WO 01/12782, WO 01/12826, WO 02/101059, WO 03/071860, WO 04/056999, WO 05/030942, WO 2005/030941, WO 2005/095632, WO 2005/095617, WO 2005/095619, WO 2005/095618, WO 2005/123927, WO 2006/018319, WO 2006/103107, WO 2006/108702, WO 2007/009823, WO 00/22140, WO 2006/063862, WO 2006/072603, WO 02/034923, WO 2008/017518, WO 2008/080630, WO 2008/080631, EP 07090007.1, WO 2008/090008, WO 01/14569, WO 02/79410, WO 03/33540, WO 2004/078983, WO 01/19975, WO 95/26407, WO 96/34968, WO 98/20145, WO 99/12950, WO 99/66050, WO 99/53072, US 6,734,341, WO 00/11192, WO 98/22604, WO 98/32326, WO 01/98509, WO 01/98509, WO 2005/002359, US 5,824,790, US 6,013,861, WO 94/04693, WO 94/09144, WO 94/11520, WO 95/35026, WO 97/20936, WO 2010/012796, WO 2010/003701,
- 2) transgenic plants which synthesize non starch carbohydrate polymers or which synthesize non starch carbohydrate polymers with altered properties in comparison to wild type plants without genetic modification. Examples are plants producing polyfructose, especially of the inulin and levan-type, as disclosed in EP-A 0 663 956, WO 96/01904, WO 96/21023, WO 98/39460, and WO 99/24593, plants producing alpha-1,4-glucans as disclosed in WO 95/31553, US 2002031826, US 6,284,479, US 5,712,107, WO 97/47806, WO 97/47807, WO 97/47808 and WO 00/14249, plants producing alpha-1,6 branched alpha-1,4-glucans, as disclosed in WO 00/73422, plants producing alternan, as disclosed in e.g. WO 00/47727, WO 00/73422, EP 06077301.7, US 5,908,975 and EP-A 0 728 213,
- 3) transgenic plants which produce hyaluronan, as for example disclosed in WO 2006/032538, WO 2007/039314, WO 2007/039315, WO 2007/039316, JP-A 2006-304779, and WO 2005/012529.
- 4) transgenic plants or hybrid plants, such as onions with characteristics such as 'high soluble solids content', 'low pungency' (LP) and/or 'long storage' (LS), as described in U.S. Patent Applications 12/020,360 and 61/054,026.

Plants or plant cultivars (that can be obtained by plant biotechnology methods such as genetic engineering) which may also be treated according to the invention are plants, such as cotton plants,

with altered fiber characteristics. Such plants can be obtained by genetic transformation, or by selection of plants contain a mutation imparting such altered fiber characteristics and include:

- a) Plants, such as cotton plants, containing an altered form of cellulose synthase genes as described in WO 98/00549.
- 5 b) Plants, such as cotton plants, containing an altered form of rsw2 or rsw3 homologous nucleic acids as described in WO 2004/053219.
- c) Plants, such as cotton plants, with increased expression of sucrose phosphate synthase as described in WO 01/17333.
- 10 d) Plants, such as cotton plants, with increased expression of sucrose synthase as described in WO 02/45485.
- e) Plants, such as cotton plants, wherein the timing of the plasmodesmatal gating at the basis of the fiber cell is altered, e.g. through downregulation of fiber-selective  $\beta$ -1,3-glucanase as described in WO 2005/017157, or as described in WO 2009/143995.
- 15 f) Plants, such as cotton plants, having fibers with altered reactivity, e.g. through the expression of N-acetylglucosaminetransferase gene including nodC and chitin synthase genes as described in WO 2006/136351.

Plants or plant cultivars (that can be obtained by plant biotechnology methods such as genetic engineering) which may also be treated according to the invention are plants, such as oilseed rape or related Brassica plants, with altered oil profile characteristics. Such plants can be obtained by genetic transformation, or by selection of plants contain a mutation imparting such altered oil profile characteristics and include:

- a) Plants, such as oilseed rape plants, producing oil having a high oleic acid content as described e.g. in US 5,969,169, US 5,840,946 or US 6,323,392 or US 6,063,947
- 25 b) Plants such as oilseed rape plants, producing oil having a low linolenic acid content as described in US 6,270,828, US 6,169,190, or US 5,965,755
- c) Plant such as oilseed rape plants, producing oil having a low level of saturated fatty acids as described e.g. in US 5,434,283 or U.S. Patent Application 12/668303

Plants or plant cultivars (that can be obtained by plant biotechnology methods such as genetic engineering) which may also be treated according to the invention are plants, such as oilseed rape or related Brassica plants, with altered seed shattering characteristics. Such plants can be obtained by genetic transformation, or by selection of plants contain a mutation imparting such altered seed shattering characteristics and include plants such as oilseed rape plants with delayed or reduced seed shattering as described in U.S. Patent Application 61/135,230, WO 2009/068313 and WO 2010/006732.

Plants or plant cultivars (that can be obtained by plant biotechnology methods such as genetic engineering) which may also be treated according to the invention are plants, such as Tobacco plants, with altered post-translational protein modification patterns, for example as described in WO 2010/121818 and WO 2010/145846.

- 5 Particularly useful transgenic plants which may be treated according to the invention are plants containing transformation events, or combination of transformation events, that are the subject of petitions for non-regulated status, in the United States of America, to the Animal and Plant Health Inspection Service (APHIS) of the United States Department of Agriculture (USDA) whether such petitions are granted or are still pending. At any time this information is readily available from APHIS
- 10 (4700 River Road, Riverdale, MD 20737, USA), for instance on its internet site (URL [http://www.aphis.usda.gov/brs/not\\_reg.html](http://www.aphis.usda.gov/brs/not_reg.html)). On the filing date of this application the petitions for nonregulated status that were pending with APHIS or granted by APHIS were those which contains the following information:
- Petition: the identification number of the petition. Technical descriptions of the transformation
  - 15 events can be found in the individual petition documents which are obtainable from APHIS, for example on the APHIS website, by reference to this petition number. These descriptions are herein incorporated by reference.
  - Extension of Petition: reference to a previous petition for which an extension is requested.
  - Institution: the name of the entity submitting the petition.
  - 20 – Regulated article: the plant species concerned.
  - Transgenic phenotype: the trait conferred to the plants by the transformation event.
  - Transformation event or line: the name of the event or events (sometimes also designated as lines or lines) for which nonregulated status is requested.
  - APHIS documents: various documents published by APHIS in relation to the Petition and which
  - 25 can be requested with APHIS.

Additional particularly useful plants containing single transformation events or combinations of transformation events are listed for example in the databases from various national or regional regulatory agencies (see for example [http://gmoinfo.jrc.it/gmp\\_browse.aspx](http://gmoinfo.jrc.it/gmp_browse.aspx) and <http://www.agbios.com/dbase.php>).

- 30 Particularly useful transgenic plants which may be treated according to the invention are plants containing transformation events, or a combination of transformation events, and that are listed for example in the databases for various national or regional regulatory agencies including Event 1143-14A (cotton, insect control, not deposited, described in WO 2006/128569); Event 1143-51B (cotton, insect control, not deposited, described in WO 2006/128570); Event 1445 (cotton, herbicide tolerance,
- 35 not deposited, described in US-A 2002-120964 or WO 02/034946); Event 17053 (rice, herbicide

tolerance, deposited as PTA-9843, described in WO 2010/117737); Event 17314 (rice, herbicide tolerance, deposited as PTA-9844, described in WO 2010/117735); Event 281-24-236 (cotton, insect control - herbicide tolerance, deposited as PTA-6233, described in WO 2005/103266 or US-A 2005-216969); Event 3006-210-23 (cotton, insect control - herbicide tolerance, deposited as PTA-6233, described in US-A 2007-143876 or WO 2005/103266); Event 3272 (corn, quality trait, deposited as PTA-9972, described in WO 2006/098952 or US-A 2006-230473); Event 40416 (corn, insect control - herbicide tolerance, deposited as ATCC PTA-11508, described in WO 2011/075593); Event 43A47 (corn, insect control - herbicide tolerance, deposited as ATCC PTA-11509, described in WO 2011/075595); Event 5307 (corn, insect control, deposited as ATCC PTA-9561, described in WO 2010/077816); Event ASR-368 (bent grass, herbicide tolerance, deposited as ATCC PTA-4816, described in US-A 2006-162007 or WO 2004/053062); Event B16 (corn, herbicide tolerance, not deposited, described in US-A 2003-126634); Event BPS-CV127-9 (soybean, herbicide tolerance, deposited as NCIMB No. 41603, described in WO 2010/080829); Event CE43-67B (cotton, insect control, deposited as DSM ACC2724, described in US-A 2009-217423 or WO2006/128573); Event CE44-69D (cotton, insect control, not deposited, described in US-A 2010-0024077); Event CE44-69D (cotton, insect control, not deposited, described in WO 2006/128571); Event CE46-02A (cotton, insect control, not deposited, described in WO 2006/128572); Event COT102 (cotton, insect control, not deposited, described in US-A 2006-130175 or WO 2004/039986); Event COT202 (cotton, insect control, not deposited, described in US-A 2007-067868 or WO 2005/054479); Event COT203 (cotton, insect control, not deposited, described in WO 2005/054480); Event DAS40278 (corn, herbicide tolerance, deposited as ATCC PTA-10244, described in WO 2011/022469); Event DAS-59122-7 (corn, insect control - herbicide tolerance, deposited as ATCC PTA 11384 , described in US-A 2006-070139); Event DAS-59132 (corn, insect control - herbicide tolerance, not deposited, described in WO 2009/100188); Event DAS68416 (soybean, herbicide tolerance, deposited as ATCC PTA-10442, described in WO 2011/066384 or WO 2011/066360); Event DP-098140-6 (corn, herbicide tolerance, deposited as ATCC PTA-8296, described in US-A 2009-137395 or WO 2008/112019); Event DP-305423-1 (soybean, quality trait, not deposited, described in US-A 2008-312082 or WO 2008/054747); Event DP-32138-1 (corn, hybridization system, deposited as ATCC PTA-9158, described in US-A 2009-0210970 or WO 2009/103049); Event DP-356043-5 (soybean, herbicide tolerance, deposited as ATCC PTA-8287, described in US-A 2010-0184079 or WO 2008/002872); Event EE-1 (brinjal, insect control, not deposited, described in WO 2007/091277); Event FI117 (corn, herbicide tolerance, deposited as ATCC 209031, described in US-A 2006-059581 or WO 98/044140); Event GA21 (corn, herbicide tolerance, deposited as ATCC 209033, described in US-A 2005-086719 or WO 98/044140); Event GG25 (corn, herbicide tolerance, deposited as ATCC 209032, described in US-A 2005-188434 or WO 98/044140); Event GHB119 (cotton, insect control - herbicide tolerance, deposited as ATCC PTA-8398, described in WO 2008/151780); Event GHB614 (cotton, herbicide tolerance, deposited as ATCC PTA-6878, described in US-A 2010-050282 or WO 2007/017186); Event GJ11 (corn, herbicide tolerance, deposited as ATCC 209030, described in US-A 2005-188434 or WO 98/044140); Event GM RZ13 (sugar beet, virus resistance , deposited as NCIMB-41601, described in WO 2010/076212); Event H7-1 (sugar beet, herbicide tolerance, deposited as NCIMB 41158 or NCIMB 41159, described

in US-A 2004-172669 or WO 2004/074492); Event JOPLIN1 (wheat, disease tolerance, not deposited, described in US-A 2008-064032); Event LL27 (soybean, herbicide tolerance, deposited as NCIMB41658, described in WO 2006/108674 or US-A 2008-320616); Event LL55 (soybean, herbicide tolerance, deposited as NCIMB 41660, described in WO 2006/108675 or US-A 2008-196127); Event

5 LLcotton25 (cotton, herbicide tolerance, deposited as ATCC PTA-3343, described in WO 03/013224 or US-A 2003-097687); Event LLRICE06 (rice, herbicide tolerance, deposited as ATCC-23352, described in US 6,468,747 or WO 00/026345); Event LLRICE601 (rice, herbicide tolerance, deposited as ATCC PTA-2600, described in US-A 2008-2289060 or WO 00/026356); Event LY038 (corn, quality trait, deposited as ATCC PTA-5623, described in US-A 2007-028322 or WO 2005/061720); Event

10 MIR162 (corn, insect control, deposited as PTA-8166, described in US-A 2009-300784 or WO 2007/142840); Event MIR604 (corn, insect control, not deposited, described in US-A 2008-167456 or WO 2005/103301); Event MON15985 (cotton, insect control, deposited as ATCC PTA-2516, described in US-A 2004-250317 or WO 02/100163); Event MON810 (corn, insect control, not deposited, described in US-A 2002-102582); Event MON863 (corn, insect control, deposited as ATCC

15 PTA-2605, described in WO 2004/011601 or US-A 2006-095986); Event MON87427 (corn, pollination control, deposited as ATCC PTA-7899, described in WO 2011/062904); Event MON87460 (corn, stress tolerance, deposited as ATCC PTA-8910, described in WO 2009/111263 or US-A 2011-0138504); Event MON87701 (soybean, insect control, deposited as ATCC PTA-8194, described in US-A 2009-130071 or WO 2009/064652); Event MON87705 (soybean, quality trait - herbicide

20 tolerance, deposited as ATCC PTA-9241, described in US-A 2010-0080887 or WO 2010/037016); Event MON87708 (soybean, herbicide tolerance, deposited as ATCC PTA9670, described in WO 2011/034704); Event MON87754 (soybean, quality trait, deposited as ATCC PTA-9385, described in WO 2010/024976); Event MON87769 (soybean, quality trait, deposited as ATCC PTA-8911, described in US-A 2011-0067141 or WO 2009/102873); Event MON88017 (corn, insect control -

25 herbicide tolerance, deposited as ATCC PTA-5582, described in US-A 2008-028482 or WO 2005/059103); Event MON88913 (cotton, herbicide tolerance, deposited as ATCC PTA-4854, described in WO 2004/072235 or US-A 2006-059590); Event MON89034 (corn, insect control, deposited as ATCC PTA-7455, described in WO 2007/140256 or US-A 2008-260932); Event

30 MON89788 (soybean, herbicide tolerance, deposited as ATCC PTA-6708, described in US-A 2006-282915 or WO 2006/130436); Event MS11 (oilseed rape, pollination control - herbicide tolerance, deposited as ATCC PTA-850 or PTA-2485, described in WO 01/031042); Event MS8 (oilseed rape, pollination control - herbicide tolerance, deposited as ATCC PTA-730, described in WO 01/041558 or US-A 2003-188347); Event NK603 (corn, herbicide tolerance, deposited as ATCC PTA-2478, described in US-A 2007-292854); Event PE-7 (rice, insect control, not deposited, described in WO

35 2008/114282); Event RF3 (oilseed rape, pollination control - herbicide tolerance, deposited as ATCC PTA-730, described in WO 01/041558 or US-A 2003-188347); Event RT73 (oilseed rape, herbicide tolerance, not deposited, described in WO 02/036831 or US-A 2008-070260); Event T227-1 (sugar beet, herbicide tolerance, not deposited, described in WO 02/44407 or US-A 2009-265817); Event T25 (corn, herbicide tolerance, not deposited, described in US-A 2001-029014 or WO 01/051654);

40 Event T304-40 (cotton, insect control - herbicide tolerance, deposited as ATCC PTA-8171, described

in US-A 2010-077501 or WO 2008/122406); Event T342-142 (cotton, insect control, not deposited, described in WO 2006/128568); Event TC1507 (corn, insect control - herbicide tolerance, not deposited, described in US-A 2005-039226 or WO 2004/099447); Event VIP1034 (corn, insect control - herbicide tolerance, deposited as ATCC PTA-3925., described in WO 03/052073), Event 32316 (corn, insect control-herbicide tolerance, deposited as PTA-11507, described in WO 2011/084632), Event 4114 (corn, insect control-herbicide tolerance, deposited as PTA-11506, described in WO 2011/084621).

The composition according to the invention can also be used against fungal diseases liable to grow on or inside timber. The term "timber" means all types of species of wood and all types of working of this wood intended for construction, for example solid wood, high-density wood, laminated wood and plywood. The method for treating timber according to the invention mainly consists in contacting one or more compounds according to the invention or a composition according to the invention; this includes for example direct application, spraying, dipping, injection or any other suitable means.

Among the diseases of plants or crops that can be controlled by the methods according to the invention, mention can be made of :

Powdery mildew diseases such as :

Blumeria diseases, caused for example by *Blumeria graminis* ;

Podosphaera diseases, caused for example by *Podosphaera leucotricha* ;

Sphaerotheca diseases, caused for example by *Sphaerotheca fuliginea* ;

Uncinula diseases, caused for example by *Uncinula necator* ;

Rust diseases such as :

Gymnosporangium diseases, caused for example by *Gymnosporangium sabinae* ;

Hemileia diseases, caused for example by *Hemileia vastatrix* ;

Phakopsora diseases, caused for example by *Phakopsora pachyrhizi* or *Phakopsora meibomia* ;

Puccinia diseases, caused for example by *Puccinia recondite*, *Puccinia graminis* or *Puccinia striiformis* ;

Uromyces diseases, caused for example by *Uromyces appendiculatus* ;

Oomycete diseases such as :

Albugo diseases caused for example by *Albugo candida* ;

Bremia diseases, caused for example by *Bremia lactucae* ;

Peronospora diseases, caused for example by *Peronospora pisi* or *P. brassicae* ;

Phytophthora diseases, caused for example by *Phytophthora infestans* ;

Plasmopara diseases, caused for example by *Plasmopara viticola* ;

Pseudoperonospora diseases, caused for example by *Pseudoperonospora humuli* or *Pseudoperonospora cubensis* ;

Pythium diseases, caused for example by *Pythium ultimum* ;

Leafspot, leaf blotch and leaf blight diseases such as :

- Alternaria diseases, caused for example by *Alternaria solani* ;  
 Cercospora diseases, caused for example by *Cercospora beticola* ;  
 Cladosporium diseases, caused for example by *Cladosporium cucumerinum* ;  
 Cochliobolus diseases, caused for example by *Cochliobolus sativus* (Conidiaform:  
 5 Drechslera, Syn: Helminthosporium) or *Cochliobolus miyabeanus* ;  
 Colletotrichum diseases, caused for example by *Colletotrichum lindemuthianum* ;  
 Cycloconium diseases, caused for example by *Cycloconium oleaginum* ;  
 Diaporthe diseases, caused for example by *Diaporthe citri* ;  
 Elsinoe diseases, caused for example by *Elsinoe fawcettii* ;  
 10 Gloeosporium diseases, caused for example by *Gloeosporium laeticolor* ;  
 Glomerella diseases, caused for example by *Glomerella cingulata* ;  
 Guignardia diseases, caused for example by *Guignardia bidwelli* ;  
 Leptosphaeria diseases, caused for example by *Leptosphaeria maculans* ; *Leptosphaeria nodorum* ;  
 Magnaporthe diseases, caused for example by *Magnaporthe grisea* ;  
 15 Mycosphaerella diseases, caused for example by *Mycosphaerella graminicola* ; *Mycosphaerella*  
*arachidicola* ; *Mycosphaerella fijiensis* ;  
 Phaeosphaeria diseases, caused for example by *Phaeosphaeria nodorum* ;  
 Pyrenophora diseases, caused for example by *Pyrenophora teres*, or *Pyrenophora tritici*  
*repentis* ;  
 20 Ramularia diseases, caused for example by *Ramularia collo-cygni* , or *Ramularia areola* ;  
 Rhynchosporium diseases, caused for example by *Rhynchosporium secalis* ;  
 Septoria diseases, caused for example by *Septoria apii* or *Septoria lycopersici* ;  
 Typhula diseases, caused for example by *Typhula incarnata* ;  
 Venturia diseases, caused for example by *Venturia inaequalis* ;  
 25 Root, Sheath and stem diseases such as :  
 Corticium diseases, caused for example by *Corticium graminearum* ;  
 Fusarium diseases, caused for example by *Fusarium oxysporum* ;  
 Gaeumannomyces diseases, caused for example by *Gaeumannomyces graminis* ;  
 Rhizoctonia diseases, caused for example by *Rhizoctonia solani* ;  
 30 Sarocladium diseases caused for example by *Sarocladium oryzae* ;  
 Sclerotium diseases caused for example by *Sclerotium oryzae* ;  
 Tapesia diseases, caused for example by *Tapesia acuformis* ;  
 Thielaviopsis diseases, caused for example by *Thielaviopsis basicola* ;  
 Ear and panicle diseases such as :  
 35 Alternaria diseases, caused for example by *Alternaria spp.* ;  
 Aspergillus diseases, caused for example by *Aspergillus flavus* ;  
 Cladosporium diseases, caused for example by *Cladosporium spp.* ;  
 Claviceps diseases, caused for example by *Claviceps purpurea* ;  
 Fusarium diseases, caused for example by *Fusarium culmorum* ;  
 40 Gibberella diseases, caused for example by *Gibberella zeae* ;

- Monographella diseases, caused for example by *Monographella nivalis* ;
- Smut and bunt diseases such as :
- Sphacelotheca diseases, caused for example by *Sphacelotheca reiliana* ;
- Tilletia diseases, caused for example by *Tilletia caries* ;
- 5 Urocystis diseases, caused for example by *Urocystis occulta* ;
- Ustilago diseases, caused for example by *Ustilago nuda* ;
- Fruit rot and mould diseases such as :
- Aspergillus diseases, caused for example by *Aspergillus flavus* ;
- Botrytis diseases, caused for example by *Botrytis cinerea* ;
- 10 Penicillium diseases, caused for example by *Penicillium expansum* ;
- Rhizopus diseases caused by example by *Rhizopus stolonifer*
- Sclerotinia diseases, caused for example by *Sclerotinia sclerotiorum* ;
- Verticillium diseases, caused for example by *Verticillium alboatrum* ;
- 15 Seed and soilborne decay, mould, wilt, rot and damping-off diseases :
- Alternaria diseases, caused for example by *Alternaria brassicicola*
- Aphanomyces diseases, caused for example by *Aphanomyces euteiches*
- Ascochyta diseases, caused for example by *Ascochyta lentis*
- Aspergillus diseases, caused for example by *Aspergillus flavus*
- 20 Cladosporium diseases, caused for example by *Cladosporium herbarum*
- Cochliobolus diseases, caused for example by *Cochliobolus sativus*  
(Conidiaform: *Drechslera*, *Bipolaris* Syn: *Helminthosporium*);
- Colletotrichum diseases, caused for example by *Colletotrichum coccodes*;
- Fusarium diseases, caused for example by *Fusarium culmorum*;
- 25 Gibberella diseases, caused for example by *Gibberella zeae*;
- Macrophomina diseases, caused for example by *Macrophomina phaseolina*
- Monographella diseases, caused for example by *Monographella nivalis*;
- Penicillium diseases, caused for example by *Penicillium expansum*
- Phoma diseases, caused for example by *Phoma lingam*
- 30 Phomopsis diseases, caused for example by *Phomopsis sojae*;
- Phytophthora diseases, caused for example by *Phytophthora cactorum*;
- Pyrenophora diseases, caused for example by *Pyrenophora graminea*
- Pyricularia diseases, caused for example by *Pyricularia oryzae*;
- Pythium diseases, caused for example by *Pythium ultimum*;
- 35 Rhizoctonia diseases, caused for example by *Rhizoctonia solani*;
- Rhizopus diseases, caused for example by *Rhizopus oryzae*
- Sclerotium diseases, caused for example by *Sclerotium rolfsii*;
- Septoria diseases, caused for example by *Septoria nodorum*;
- Typhula diseases, caused for example by *Typhula incarnata*;
- 40 Verticillium diseases, caused for example by *Verticillium dahliae* ;

Canker, broom and dieback diseases such as :

Nectria diseases, caused for example by *Nectria galligena* ;

Blight diseases such as :

Monilinia diseases, caused for example by *Monilinia laxa* ;

5 Leaf blister or leaf curl diseases such as :

Exobasidium diseases caused for example by *Exobasidium vexans*

Taphrina diseases, caused for example by *Taphrina deformans* ;

Decline diseases of wooden plants such as :

Esca diseases, caused for example by *Phaemoniella clamydospora* ;

10 Eutypa dyebark, caused for example by *Eutypa lata* ;

Ganoderma diseases caused for example by *Ganoderma boninense*;

Rigidoporus diseases caused for example by *Rigidoporus lignosus*

Diseases of Flowers and Seeds such as

15 Botrytis diseases caused for example by *Botrytis cinerea*;

Diseases of Tubers such as

Rhizoctonia diseases caused for example by *Rhizoctonia solani*;

Helminthosporium diseases caused for example by *Helminthosporium solani*;

20

Club root diseases such as

Plasmodiophora diseases, cause for example by *Plasmodiophora brassicae*.

Diseases caused by Bacterial Organisms such as

25 Xanthomonas species for example *Xanthomonas campestris* pv. *oryzae*;

Pseudomonas species for example *Pseudomonas syringae* pv. *lachrymans*;

Erwinia species for example *Erwinia amylovora*.

### 30 Legend of the figures

**Fig 1 :** Measurement of *Phytophthora infestans* growth inhibition in the presence of dsRNA targeting the saccharopine dehydrogenase messenger RNA.

**Fig 2 :** Analysis of saccharopine dehydrogenase mRNA level by qRT PCR.

### 35 Sequence listing:

- SEQ ID N1: Saccharopine dehydrogenase (LYS1) from *Aspergillus clavatus*.  
SEQ ID N2: Protein encoded by the above nucleic acid sequence.  
SEQ ID N3: Saccharopine dehydrogenase (LYS1) from *Aspergillus fumigatus*.  
SEQ ID N4: Protein encoded by the above nucleic acid sequence.
- 5 SEQ ID N5: Saccharopine dehydrogenase (LYS1) from *Botrytis cinerea*.  
SEQ ID N6: Protein encoded by the above nucleic acid sequence.  
SEQ ID N7: Saccharopine dehydrogenase (LYS1) from *Fusarium graminearum*.  
SEQ ID N8: Protein encoded by the above nucleic acid sequence.  
SEQ ID N9: Saccharopine dehydrogenase (LYS1) from *Fusarium oxysporum*.
- 10 SEQ ID N10: Protein encoded by the above nucleic acid sequence.  
SEQ ID N11: Saccharopine dehydrogenase (LYS1) from *Fusarium verticilloides*.  
SEQ ID N12: Protein encoded by the above nucleic acid sequence.  
SEQ ID N13: Saccharopine dehydrogenase (LYS1) from *Fusarium verticilloides*.  
SEQ ID N14: Protein encoded by the above nucleic acid sequence.
- 15 SEQ ID N15: Saccharopine dehydrogenase (LYS1) from *Mycosphaerella fijiensis*.  
SEQ ID N16: Polypeptide encoded by the above nucleic acid sequence.  
SEQ ID N17: Saccharopine dehydrogenase (LYS1) from *Magnaporthe grisea*.  
SEQ ID N18: Protein encoded by the above nucleic acid sequence.  
SEQ ID N19: Saccharopine dehydrogenase (LYS1) from *Monoliophthora pernicios*.
- 20 SEQ ID N20: Protein encoded by the above nucleic acid sequence.  
SEQ ID N21: Saccharopine dehydrogenase (LYS1) from *Puccinia graminis*.  
SEQ ID N22: Protein encoded by the above nucleic acid sequence.  
SEQ ID N23: Saccharopine dehydrogenase (LYS1) from *Phytophthora infestans*.  
SEQ ID N24: Protein encoded by the above nucleic acid sequence.
- 25 SEQ ID N25: Saccharopine dehydrogenase (LYS1) (from *Phytophthora ramorum*).  
SEQ ID N26: Protein encoded by the above nucleic acid sequence.  
SEQ ID N27: Saccharopine dehydrogenase (LYS1) from *Phytophthora sojae*.  
SEQ ID N28: Protein encoded by the above nucleic acid sequence.  
SEQ ID N29: Saccharopine dehydrogenase (LYS1) from *Pyrenophora tritici-repentis*.
- 30 SEQ ID N30: Protein encoded by the above nucleic acid sequence.  
SEQ ID N31: Saccharopine dehydrogenase (LYS1) from *Sclerotinia sclerotiorum*.  
SEQ ID N32: Protein encoded by the above nucleic acid sequence.  
SEQ ID N33: Saccharopine dehydrogenase (LYS1) from *Trichoderma reesei*.  
SEQ ID N34: Protein encoded by the above nucleic acid sequence.
- 35 SEQ ID N35: Saccharopine dehydrogenase (LYS1) from *Ustilago maydis*.  
SEQ ID N36: Protein encoded by the above nucleic acid sequence.  
SEQ ID N37: Saccharopine dehydrogenase (LYS1) from *Verticillium albo-atrum*.  
SEQ ID N38: Protein encoded by the above nucleic acid sequence.  
SEQ ID N39: Saccharopine dehydrogenase (LYS1) from *Mycosphaerella graminicola*.
- 40 SEQ ID N40: Protein encoded by the above nucleic acid sequence.

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**SEQ ID N°41:** Saccharopine dehydrogenase (LYS1) from *Fusarium moniliform*.

**SEQ ID N°42:** Protein encoded by the above nucleic acid sequence.

**SEQ ID N°43:** Saccharopine dehydrogenase (LYS1) from *Claviceps purpurea*.

**SEQ ID N°44:** Protein encoded by the above nucleic acid sequence.

**SEQ ID N°45:** Primer SACdh\_Pi\_T7\_F

**SEQ ID N°46:** Primer SACdh\_Pi\_T7\_R

**SEQ ID N°47:** Primer Actin forward

**SEQ ID N°48:** Primer Actin reverse

**SEQ ID N°49:** Primer  $\beta$ -Tub forward

**SEQ ID N°50:** Primer  $\beta$ -Tub reverse

**SEQ ID N°51:** Primer SACdh forward

**SEQ ID N°52:** Primer SACdh reverse

**SEQ ID N°53:** Primer pBINB33-1

**SEQ ID N°54:** Primer pBINB33-2

**SEQ ID N°55:** Primer SacdhPI R

**SEQ ID N°56:** Primer SacdhPI F

**SEQ ID N°57:** Primer LYS1 Pot 117-F

**SEQ ID N°58:** Primer LYS1 Pot 117-R

In the claims which follow and in the preceding description of the invention, except where the context requires otherwise due to express language or necessary implication, the word "comprise" or variations such as "comprises" or "comprising" is used in an inclusive sense, i.e. to specify the presence of the stated features but not to preclude the presence or addition of further features in various embodiments of the invention.

It is to be understood that, if any prior art publication is referred to herein, such reference does not constitute an admission that the publication forms a part of the common general knowledge in the art, in Australia or any other country.

The various aspects of the invention will be understood more fully by means of the experimental examples below.

All the methods or operations described below are given by way of example and correspond to a choice, made among the various methods available for achieving the same result. This choice has no effect on the quality of the result, and, consequently, any appropriate method can be used by those skilled in the art to achieve the same result. In particular, and unless otherwise specified in the examples, all the recombinant DNA techniques employed are carried out according to the standard protocols described in Sambrook and Russel (2001, *Molecular cloning: A laboratory manual*, Third edition, Cold Spring Harbor Laboratory Press, NY) in Ausubel *et al.* (1994, *Current Protocols in Molecular Biology*, Current protocols, USA, Volumes 1 and 2), and in Brown (1998, *Molecular Biology LabFax*, Second edition, Academic Press, UK). Standard materials and methods for plant molecular biology are described in Croy R.D.D. (1993, *Plant Molecular Biology LabFax*, BIOS Scientific Publications Ltd (UK) and Blackwell Scientific Publications (UK)). Standard materials and methods for PCR (Polymerase Chain Reaction) are also described in Dieffenbach and Dveksler (1995, *PCR Primer: A laboratory manual*, Cold Spring Harbor Laboratory Press, NY) and in McPherson *et al.* (2000, *PCR - Basics: From background to bench*, First edition, Springer Verlag, Germany).

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## 5 Examples

### Example 1 : *In vitro* cultivation of *Magnaporthe Grisea*

Assays were carried out using the *Magnaporthe grisea* wild-type strain P1.2 originated from the collection of the phytopathology laboratory of the CIRAD (Centre de coopération internationale en recherche agronomique pour le développement) in Montpellier. Conditions for culturing, the composition of the rice-agar medium, maintenance, and sporulation as well as protoplasts preparations are described by Silué *et al.* (1998).

### Example 2 : *Magnaporthe grisea* transfection with dsRNA targeting saccharopine dehydrogenase and measurement of growth inhibition

The *Magnaporthe grisea* saccharopine dehydrogenase (SACdh) gene sequence Lys-1 (MGG\_01359.6: 1426 bp) was obtained from the Broad Institute (<http://www.broadinstitute.org/>). A region of about 325 bp was selected for dsRNA, comprizing the nucleotides 301 through 626, was synthesized by the Geneart company and cloned into the plasmid .

For transfection, ds RNA of saccharopine dehydrogenase from *Magnaporthe grisea* was produced using the MEGAscript RNAi Kit (Ambion) according to the manufacturers' instructions. Different amounts (200µg to 2µg of ds RNA) were treated with transfection agent Lipofectamin RNAi max (Invitrogen) following manufactures' instructions. Lipofectamin-ds RNA complexes were added to  $2,5 \times 10^6$  *Magnaporthe grisea* protoplast in a microtiterplate with TB3 media (Villalba *et al.*, 2008), and growth was monitored for 5-7 days at a OD of 600 nm using a Infinite M1000 (Tecan) microplate reader..

Growth of *Magnaporthe grisea* protoplasts treated with ds RNA of saccharopine dehydrogenase comparing to untreated control.was monitored at several time points and showed a significant difference in growth.

### Example 3 : *In vitro* cultivation *Phytophthora infestans* and zoospores preparation.

*Phytophthora infestans* strain PT78 was cultivated *in vitro* in 9cm petri dishes on pea agar medium (125g/l boiled and crushed peas, 20 g/l agar agar, carbenicillin 100mg/l ) at 21°C in the dark. Every 15 days, new medium was inoculated with four 5mm cubic plugs of mycelium.

To release the zoospores, 12 ml of ice cold water were put on a 10 days old culture and the culture was placed at 4°C for 2 hours. The supernatant was then collected without dwasturbing the mycelium and filtered through a 100 µm sieve to remove hyphal fragments. The zoospores were placed on ice and counted with a haemocytometer.

### Example 4 : *Phytophthora infestans* transfection with dsRNA targeting saccharopine dehydrogenase and measurement of growth inhibition

The *Phytophthora infestans* saccharopine dehydrogenase gene sequence Lys-1 (PITG\_03530 : 3020 bp) was obtained from the *P. infestans* database of the Broad Institute. A region about 500 bp offering the best siRNA according to the BLOCKit RNAi designer software (Invitrogen) and comprising the nucleotides 2251 through 2750, was synthesized by the Geneart company and cloned into the plasmid 0920357\_SacDH\_Pi\_pMA..

Synthesis of dsRNA was carried out using the Megascript RNAi kit (Ambion) following the manufacturer's protocol and using as a template a PCR product amplified from the plasmid 0920357\_SacDH\_Pi\_pMA. The forward primer used was SACdh\_Pi\_T7\_F: 5' TAATACGACTCACTATAGGGTTGCAGGAGAGCGCAGAAAGC and the reverse primer was SACdh\_Pi\_T7\_R: TAATACGACTCACTATAGGGTCAGTTGGAGTCCGCGTGGTGT.

dsRNA were then precipitated with 100% ethanol and sodium acetate 3M, pH5.2, washed 2 times with 70 % ethanol and the pellets were resuspended in RNase free water.

The transfection mixes were prepared in a 48 well plate by adding sequentially V8 medium (5% of V8 juice (Campbell Foods Belgium), pH5) , the appropriate amount of dsRNA and 10 µl of lipofectamine RNAi max (Invitrogen) in a final volume of 200 µl. The transfection mixture was incubated during 15 min at room temperature..

Zoospores were diluted in V8 medium to a concentration of  $5 \times 10^4$  zoospores/ml. Then, 800 µL of the zoospores solution were added in each well of the plate. The final concentration of zoospores was  $4 \times 10^5$  zoospores / ml.

Three controls were added on each plate: V8 medium, V8 medium + zoospores, V8 medium + zoospores + lipofectamine. The plates were incubated at 21°C, in the dark.

The growth of the fungus was followed by measuring the absorbance at 620 nm in a plate reader (Infinite 1000, Tecan) over 8 days. The percentage of growth inhibition was calculated using the following formula:  $100 - (OD_{dsRNA} \times 100 / OD_{control \text{ lipofectamine}})$ . The growth of the fungi was reduced in the presence of dsRNA directed against saccharopine dehydrogenase in a concentration dependant manner (100nM and 200nM respectively) as shown in Figure 1

#### **Example 5 : Quantitative PCR analysis of *P. infestans* saccharopine dehydrogenase messenger RNA:**

To yield sufficient RNA for cDNA synthesis and real-time RT-PCR, several wells of the 48 wells plate were pooled for one concentration of dsRNA tested : 10 wells for 72h time point, 6 wells for 96h time point, 3 wells for 120h time point.

After 72h, 96h and 120h of treatment with the dsRNA, the mycelia were collected. The samples were centrifuged to remove the medium. The samples were frozen in liquid nitrogen and then lyophilized overnight.

Before RNA extraction, the mycelium was grinded. Total RNA was extracted using the RNeasy Plant mini kit (Qiagen) following the manufacturer's protocol. DNA contamination of the RNA samples was removed by DNase digestion (DNA free, Ambion). Integrity of the RNA was tested on the 2100 Bioanalyzer (RNA 6000 nano kit, Agilent) following the protocol supplied by the manufacturer. The cDNA were synthesized from 2 µg of total RNA by oligo dT priming using the kit Thermoscript RT-PCR

system (Invitrogen) following the manufacturer's protocol. The cDNA were precipitated with 100 %EtOH and sodium acetate 3M, pH5.2, washed 2 times with 70 % EtOH and the pellets were resuspended in 10 µL of RNase free water. The cDNA were diluted a hundred fold for the qPCR test.

- 5 Primer pairs were designed for each gene sequence by using the Primer Express 3 software (Applied Biosystems). Real time RT-PCR was performed on a 7900 Real Time PCR system (Applied Biosystems) with Power SYBR green PCR master mix (Applied Biosystems) following the manufacturer's protocol. Q-PCR was performed as follows : 95°C for 10 min, 45 cycles at 95°C for 15s and 60°C 1min, followed by a dissociation stage at 95°C for 15s, 60°C for 1min and 95°C for 15s.
- 10 The actin and  $\beta$ -tubuline genes were used as endogenous controls. The relative expression of genes was calculated with the  $2\Delta\Delta C_t$  method. The figure 2 shows a significant reduction of the level of the saccharopine dehydrogenase messenger RNA and this reduction correlates with growth inhibition.

Table 1: Sequence of qPCR primers :

Primer name	Forward primer sequence	Reverse primer sequence
actin	CGACTCTGGTGACGGTGTGT	GCGTGAGGAAGAGCGTAACC
$\beta$ -Tub	CCGCCCAGACAATTTTCGT	CCTTGCCCCAGTTGTTACCA
SACdh	TGGGTGGTTTCCAAGGTCTTC	AAAGGCACCAAGCCACTGAA

15

**Example 6 : Construction of transformation vectors containing the *Phytophthora infestans* Saccharopine Dehydrogenase gene.**

a) Preparation of the plant expression vector IR 47-71

- The plasmid pBinAR is a derivative of the binary vector plasmid pBin19 (Bevan, 1984) which was constructed as follows: A fragment of a length of 529 bp which comprised the nucleotides 6909-7437 of the 35S promoter of the cauliflower mosaic virus was isolated as EcoR \Kpn I fragment from the plasmid pDH51 (Pietrzak et al, 1986) and ligated between the EcoR I and Kpn I restriction sites of the polylinker of pUC18. In this manner, the plasmid pUC18-35S was formed. Using the restriction endonucleases Hind III and Pvu II, a fragment of a length of 192 bp which included the polyadenylation signal (3' terminus) of the Octopine Synthase gene (gene 3) of the T-DNA of the Ti plasmid pTiACH $\delta$  (Gielen et al, 1984) (nucleotides 11 749-11 939) was isolated from the plasmid pAGV40 (Herrera-Estrella et al, 1983). Following addition of Sph I linkers to the Pvu II restriction site, the fragment was ligated between the Sph I and Hind III restriction sites of pUC18-35S. This gave the plasmid pA7. Here, the entire polylinker comprising the 35S promoter and Ocs terminator was removed using EcoR I and Hind III and ligated into the appropriately cleaved vector pBin19. This gave the plant expression vector pBinAR (Hofgen and Willmitzer, 1990).
- 20
- 25
- 30

- The promoter of the patatin gene B33 from *Solanum tuberosum* (Rocha-Sosa et al., 1989) was, as Dra I fragment (nucleotides -1512 - +14), ligated into the Ssf I-cleaved vector pUC19 whose ends had been blunted using T4-DNA polymerase. This gave the plasmid pUC19-B33. From this plasmid, the B33 promoter was removed using EcoR I and Sma I and ligated into the appropriately restricted vector pBinAR. This gave the plant expression vector pBinB33. To facilitate further cloning steps, the MCS
- 35

(Multiple Cloning Site) was extended. To this end, two complementary oligonucleotides were synthesized, heated at 95°C for 5 minutes, slowly cooled to room temperature to allow good fixation (annealing) and cloned into the Sal I and Kpn I restriction sites of pBinB33. The oligonucleotides used for this purpose had the following sequence:

- 5 pBINB33-1: 5'-TCG ACA GGC CTG GAT CCT TAA TTA AAC TAG TCT CGA GGA GCT CGG TAC-3'  
pBINB33-2: 5'-CGA GCT CCT CGA GAC TAG TTT AAT TAA GGA TCC AGG CCT G-3'

The plasmid obtained was named IR 47-71.

- b) Preparation of the plant expression vectors pEPA248 and pEPA262 comprising a nucleic acid sequences for the *Phytophthora infestans* Saccharopine Dehydrogenase gene.
- 10 The saccharopine dehydrogenase sequence (PITG\_03530: 3020 bp) was obtained from the P. infestans ORF Prot V1 database. A region about 500 bp offering the best siRNA according to the BLOCKit RNAi designer software (Invitrogen), was synthesized by the Geneart company.
- A 300 bp fragment was amplified by PCR from this sequence DNA with the primers SacdhPI R (5'-agaggtaccaagcttgctagctgg-3') and SacdhPI F (5'-tatctcgagcttagacaacgccattggttac-3').
- 15 The amplified fragment was cloned into pCRII-Topo (Invitrogen) to obtain the plasmid pEPA250. According to Wesley *et al.* (2001), the sequence of interest was cloned in pHannibal vector to give plasmid pEPA241. Then the dsRNA expression cassette was sub-cloned into different binary (plant expression) vectors pART27 (Gleave AP, PMB 20, (1992), 1203-1207) and IR 47 to produce the plant
- 20 expression vectors pEPA248 and pEPA262, respectively.
- Vector pEPA248 and pEPA262 were introduced into respectively GV3101 and C58C1RIF (pGV2260) agrobacteria cells by electroporation (Rocha-Sosa et al.(1989)), in order to further transform potato plants.

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**Example 7: Construction of transformation vectors targeting the *Sclerotinia sclerotiorum* Saccharopine Dehydrogenase gene *lys1*.**

- The 351bp of a region of the *S. sclerotiorum* *Lys1* coding sequence (saccharopine dehydrogenase SS1G\_06166.1) was synthesized by the Geneart company (pEPA293), and flanked by internal (*Xba*I,
- 30 *Hind*III) and external (*Xho*I, *Kpn*I) restriction sites designed to perform a two-step cloning into the pHannibal vector (Wesley et al., 2001). The intermediate plasmid harbored two inverted copies of the *Lys1* gene fragment spaced by the pHannibal *PdK* intron and regulated by the cauliflower mosaic virus (CaMV) 35S promoter and the OCS terminator.

- The entire DNA cassette was then excised with *Not*I and inserted into the pART27 binary vector
- 35 (Gleave, 1992), giving the final plasmid pEPA307 with a plant selection cassette based on kanamycin resistance (*nptII* gene regulated by the *Nos* promoter and terminator).

The same *Not*I cassette was also inserted in a binary vector (pFCO31) with a plant selection marker based on an HPPD inhibitors resistance, to be used in Soybean transformation. The final plasmid can then transform plants with a T-DNA comprising in between the Right and Left borders, our cassette of

interest and an HPPD gene regulated by a CsVMV promoter, a chloroplast transit peptide sequence and a 3'Nos terminator.

**Example 8 : Construction of transformation vectors targeting the *Phakopsora pachirizi***

**5 Saccharopine Dehydrogenase gene *lys1*.**

The 364bp of a region of a *Phakopsora pachirizi* *Lys1* E.S.T. (saccharopine dehydrogenase PHAPC\_EH247326.1) was synthesized by the Geneart company (pCED42), and flanked by internal (*XbaI*, *HindIII*) and external (*XhoI*, *KpnI*), restriction sites designed to perform a two-step cloning into the pHannibal vector (Wesley et al., 2001). The intermediate plasmid harbored two inverted copies of the *Lys1* gene fragment spaced by the pHannibal *PdK* intron and regulated by the cauliflower mosaic virus (CaMV) 35S promoter and the OCS terminator.

The entire DNA cassette was then excised with *NotI* and inserted into the pART27 binary vector (Gleave, 1992), giving the final plasmid pCED45 with a plant selection cassette based on kanamycin resistance (*nptII* gene regulated by the *Nos* promoter and terminator).

15 The same *NotI* cassette was also inserted in a binary vector (pFCO31) with a plant selection marker based on an HPPD inhibitors resistance, to be used in Soybean transformation. The final plasmid (pCED87) can then transform plants with a T-DNA comprising inbetween the Right and Left borders, our cassette of interest and an HPPD gene regulated by a CsVMV promoter, a chloroplast transit peptide sequence and a 3'Nos terminator.

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**Example 9: Transformation of potato plants with plant expression vectors comprising nucleic acid molecules coding for hairpin saccharopine dehydrogenase construct pEPA262**

Potato plants were transformed via *Agrobacterium* using the plant expression vector pEPA262, which comprises a coding nucleic acid sequence for saccharopine dehydrogenase under the control of the promoter of the patatin gene B33 from *Solanum tuberosum* as described by Rocha-Sosa et al. (1989).

The transgenic potato plants transformed with the plasmid pEPA262, were named "537 ES".

Molecular analysis of the events of "537 ES" was performed using standart PCR methods (Sambrook et al.) to detect the presence of the nucleic acid sequence for saccharopine dehydrogenase using the following primers SacDH PI F: 5'-TATCTCGAGTCTAGACAACGCCATTGGTTAC-3' and SacDH PI R: 5'-AGAGGTACCAAGCTTGCGTAGCTGG-3'. Further selection was accomplished either by Northern blotting or by expression analysis of the the nucleic acid sequence for saccharopine dehydrogenase via RT-Q PCR leading to a selection of different events. The oligonucleotides used for this purpose had the following sequence: LYS1\_Pot 117-F: 5'-TCA ATA GAA GCG AAC GCG TAA A-3' and LYS1\_Pot 117-R: 5'-GTT CGG GAT CTG CTC GAT GT-3'

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**Example 10: *Agrobacterium*-mediated transformation of *Arabidopsis thaliana*.**

The pART27 derived plasmids was introduced into *Agrobacterium tumefaciens* strain LBA4404 (Invitrogen Electromax) by electroporation. The obtained bacterial strains were then used for the floral

dip infiltration of the *A. thaliana* Col-0 or Wassilewskija plants as described by Clough & Bent (PlantJ 1998).

**Example 11: Agrobacterium-mediated transformation of *Glycine max*.**

- 5 The pFCO31 derived plasmids were introduced into *Agrobacterium tumefaciens* strain LBA4404 (Invitrogen Electromax) by electroporation. The obtained bacterial strains were then used for Soybean transformation as described below.

- Soybean seeds are sterilized for 24h with Chlorine gas (Cl<sub>2</sub>). Seeds are then placed in Petri dishes and soaked in sterile deionized water for 20 hours prior to inoculation, in the dark, at room temperature. An overnight culture grown at 28 °C and 200rpm agitation of *Agrobacterium tumefaciens* in 200ml of YEP (5 g/L Yeast extract, 10 g/L Peptone, 5 g/L NaCl<sub>2</sub>, pH to 7.0) containing the appropriate antibiotic is centrifugated at 4000rpm, 4 °C, 15min. The pellet is resuspended in 40 to 50mL of infection medium to a final OD<sub>600nm</sub> between 0.6 and 1 and stored on ice. Soaked seeds are dissected, under sterile conditions, using a #15 scalpel blade to separate the cotyledons and remove the primary leaves attached to them. Each cotyledon is kept as explant for inoculation. About 100 explants are prepared and subsequently inoculated together, for 30 minutes in the *Agrobacterium* inoculum, with occasional agitation. Cocultivation is performed in classical Petri dishes containing 4 papers filter (Whatman® grade 1) and 4 mL of Cocultivation medium (1/10X B5 major salts, 1/10X B5 minor salts, 2.8 mg/L Ferrous, 3.8 mg/L NaEDTA, 30 g/L Sucrose, 3.9 g/L MES (pH 5.4). Filter sterilized 1X B5 vitamins, GA3 (0.25 mg/L), BAP (1.67 mg/L), Cysteine (400 mg/L), Dithiothrietol (154.2 mg/L), and 200µM acetosyringone). Explants are placed on co-cultivation plates (9 per plate), adaxial (flat) side down and sealed with a single vertical string of tape (Leucopore®) and further incubated for 5 days, at 24°C, in a 18:6 photoperiod. At the end of cocultivation, the explants are placed (6 per plate) on the Shoot Initiation Medium (1X B5 major salts, 1X B5 minor salts, 28 mg/L Ferrous, 38 mg/L NaEDTA, 30 g/L Sucrose, 0.56 g/L MES, and 8 g/L agar (pH 5.6). Filter sterilized 1X B5 vitamins, BAP (1.67 mg/L), Timentin (50 mg/L), Cefotaxime (50 mg/L), Vancomycin (50 mg/L) and Tembotrione (0.1 mg/L)), inclined at 45 °, with the cotyledonary node area imbedded in the medium and upwards. The Shoot Initiation step lasts 1 month (24°C 16/8 photoperiod). After one more month, explants with green shoots are transferred on Shoot Elongation Medium (1X MS/B5 medium amended with 1 mg/l zeatin riboside (ZR), 0.1 mg/l IAA, 0.5 mg/l GA3, 3% sucrose, 100 mg/l pyroglutamic acid, 50 mg/l asparagine, 0.56 g/L MES, pH 5.6, solidified with 0.8% agar, ticarcillin (50 mg/l), cefotaxime (50 mg/l) and vancomycin (50 mg/L)), with fresh transfer every 2 weeks. Plantlets that are more than 2cm high are transferred on Rooting medium. Plantlets are cut and placed on rooting medium (1/2 MS major salts, minor salts and vitamins B5, 15 g/L Sucrose, 1mg/L IBA 8 g/L Noble agar, pH 5.7). in an 180mL vertical plastic container.

Once the roots are well formed and the apex is strong, plants are placed into soil in the greenhouse and covered with a green plastic box for acclimatization for 5 days on a 36°C heating bed. After 10 days of acclimatization, the plants are transferred into big pots, without heating bed.

**Example 12: Asian Soybean Rust (*Phakopsora pachyrhizi*) assay.**

Soybean plants expressing dsRNA directed against *Phakopsora pachyrhizi* *Lys1* were grown in the greenhouse in 7,5 cm pots (28,5 °C, 50% humidity, 14h light). In an incubator, plants were sprayed with a conidia suspension (50ml at  $10\text{--}15 \times 10^4$  spores/ml obtained from artificially infected soybean plants serving as a source of inoculum, for one tray of dimensions 55X34X5cm containing 15 pots). Suspension includes Tween20 at 0.033%. To ensure even inoculation multidirectional spraying is necessary. Plants are then incubated for 4 days at ca. 25 °C (daytime) and ca. 20 °C (night) with very high humidity (90% to saturation). After this period plants are transferred back to normal growing conditions. Asian soybean rust development is evaluated at regular intervals to follow kinetics of disease development and severity of symptoms. All experiments with Asian soybean rust are performed in L2 safety level culture chambers or incubators according to HCB requirements.

**Example 13: *Sclerotinia sclerotiorum* assay**

Development of the Wild-type *S. sclerotiorum* isolate 1980 as well as the pac 1 mutant (Rollins, 2003) fungus was studied on a whole plant assay. *S. sclerotiorum* was stored at 4 °C on potato dextrose agar (PDA, potato 200 g/l, glucose 20 g/l, agar 18 g/l). The fungus was cultured in a Petri dish containing PDA by placing a mycelial plug in the centre and was maintained under static conditions at 21°C for 4 days. 4 weeks old *Arabidopsis* wild-type and transgenic plants were inoculated with 12-mm diameter agar-mycelium plugs excised from the actively growing margin of the fungal colony in the centre of the plant. Inoculated plants were kept in a growth chamber at 21°C with 100% relative humidity under a 12-h light photoperiod with a light intensity of  $34 \text{ mmol m}^{-2} \text{ s}^{-1}$  using fluorescent white lights and were monitored every 12 h to observe fungal development. Disease symptoms were monitored by number of infected leaves as well as lengths and widths of lesions.

The claims defining the invention are as follows:

1. A dsRNA molecule comprising i) a first strand comprising a sequence substantially identical to at least 18 contiguous nucleotides of a fungus or oomycete saccharopine dehydrogenase gene and ii) a second strand comprising a sequence substantially complementary to the first strand, wherein the fungus or oomycete gene is selected from the group consisting of:
  - a) a polynucleotide comprising a sequence as set forth in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43;
  - b) a polynucleotide encoding a polypeptide having a sequence as set forth in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44;
  - c) a polynucleotide having at least 70% sequence identity, preferably at least 80%, more preferably at least 90%, even more preferably at least 95% to a polynucleotide having a sequence as set forth in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43;
  - d) a polynucleotide encoding a polypeptide having at least 70% sequence identity, preferably at least 80%, more preferably at least 90%, even more preferably at least 95%, to a polypeptide having a sequence as set forth in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44;
  - e) a polynucleotide hybridizing under stringent conditions to a polynucleotide having a sequence as set forth in SEQ ID NO: 1, 3, 5, 7, 9, 11, 13, 15, 17, 19, 21, 23, 25, 27, 29, 31, 33, 35, 37, 39, 41, 43; and
  - f) a polynucleotide hybridizing under stringent conditions to a polynucleotide encoding a polypeptide having at least 70% sequence identity to a polypeptide having a sequence as set forth in SEQ ID NO: 2, 4, 6, 8, 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 32, 34, 36, 38, 40, 42, 44.
2. A composition comprising at least a dsRNA molecule according to claim 1.
3. A composition according to claim 2, further comprising an agriculturally acceptable support, carrier, filler and/or surfactant.
4. A composition according to claim 2 or 3 further comprising a phytopharmaceutical or plant growth promoting compound.
5. A micro-organism producing a dsRNA molecule according to claim 1.
6. A genetic construct which comprises at least one DNA sequence as well as heterologous regulatory element(s) in the 5' and optionally in the 3' positions, wherein the DNA sequence(s) is able to form a dsRNA molecule according to claim 1.
7. A cloning and/or expression vector, containing at least one genetic construct according to claim 6.

8. A transgenic plant cell which comprises a dsRNA molecule according to claim 1 or a construct according to claim 6.

9. A transgenic plant, seed or part thereof, comprising a transgenic plant cell according to claim 8.

10. A transgenic plant cell according to claim 8 or transgenic plant, seed or part thereof, according to claim 9 wherein said plant is a soybean, oilseed, rice or potato plant.

11. A method of making a transgenic plant cell capable of expressing a dsRNA that inhibits a fungus or oomycete saccharopine dehydrogenase gene, said method comprising the steps of transforming a plant cell with a genetic construct according to claim 6.

12. A method of controlling a plant pathogen, particularly a fungus or oomycete, comprising providing to said pathogen a dsRNA molecule according to claim 1, or a composition according to any one of claims 2 to 4.

13. A method according to claim 12, wherein an effective quantity of a dsRNA molecule according to claim 1 or a composition according to any one of claims 2 to 4 is applied to the soil where plants grow or are capable of growing, to the leaves and/or the fruit of plants or to the seeds of plants.

14. A method according to claim 12 comprising providing in the host plant of said plant pathogen a transformed plant cell according to claim 8.

15. A method for inhibiting the expression of a plant pathogen gene, comprising the following steps:

- i) transforming a plant cell with a genetic construct according to claim 6;
- ii) placing the cells thus transformed under conditions that allow the transcription of said construct; and
- iii) bringing the cells into contact with the plant pathogen.

16. A method according to any one of claims 11 to 15 wherein said plant pathogen is *Magnaporthe grisea*, *Phytophthora infestans*, *Sclerotinia sclerotinium* or *Phakopsora pachyrhizi*.

17. A method according to any one of claims 11 to 15 wherein said plant is soybean, oilseed, rice or potato.

Figure 1

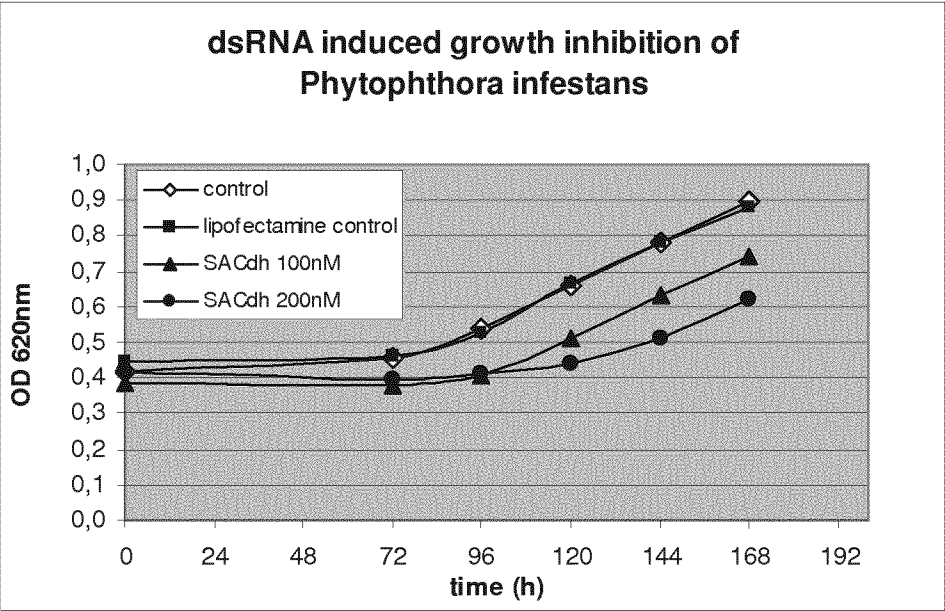
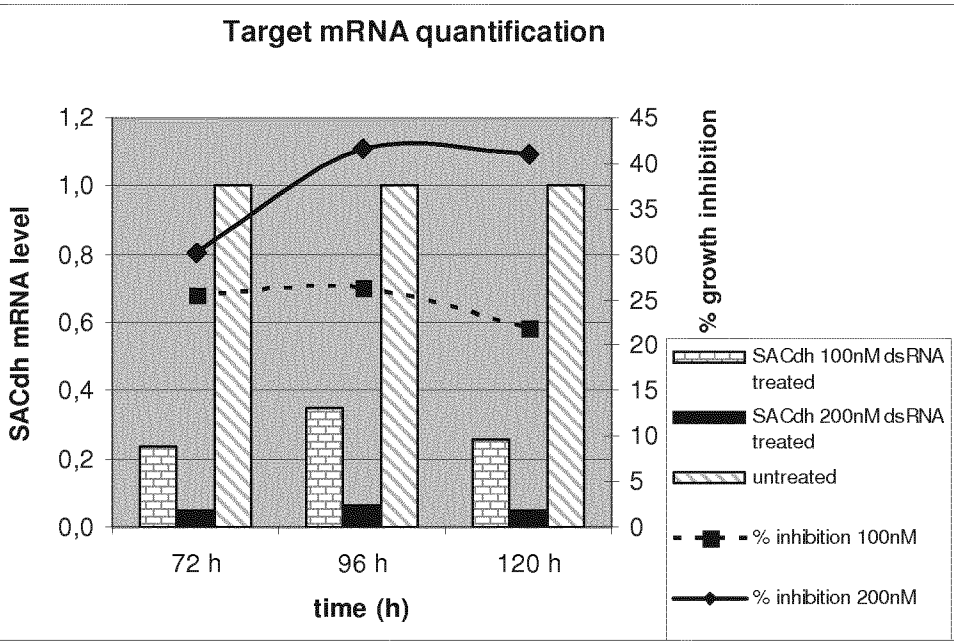


Figure 2



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Lys	Glu	Leu	Pro	Glu	Asp	Asp	Phe	Pro	Leu	Glu	His	Val	His	Ile	Ser	
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Pro	Glu	Gly	Glu	Pro	Leu	Pro	Gly	Glu	Thr	Pro	Tyr	Glu	Asn	Gln	Asp	
				165					170					175		

eof-seql.txt

ctt	ctg	atc	gcg	tcc	gtg	aag	gag	tcg	ctg	gag	ggt	ggc	aag	aag	cag	576
Leu	Leu	Ile	Ala	Ser	Val	Lys	Glu	Ser	Leu	Glu	Val	Gly	Lys	Lys	Gln	
			180					185					190			
tct	gga	aag	tcg	ccc	aag	gtg	ctt	gtc	att	gga	gct	ctt	ggc	cgt	tgc	624
Ser	Gly	Lys	Ser	Pro	Lys	Val	Leu	Val	Ile	Gly	Ala	Leu	Gly	Arg	Cys	
		195					200					205				
ggc	aag	ggt	gct	gtg	cag	ctg	gcc	aag	gat	ggt	ggc	att	ccc	gag	tct	672
Gly	Lys	Gly	Ala	Val	Gln	Leu	Ala	Lys	Asp	Val	Gly	Ile	Pro	Glu	Ser	
	210					215					220					
gat	atc	atc	cag	tgg	gat	att	gaa	gag	acc	aag	aag	ggt	gga	ccc	ttc	720
Asp	Ile	Ile	Gln	Trp	Asp	Ile	Glu	Glu	Thr	Lys	Lys	Gly	Gly	Pro	Phe	
	225				230					235					240	
cgt	gag	atc	gtt	gag	gac	gtt	gat	atc	ttc	gtc	aac	tgc	atc	tac	ctc	768
Arg	Glu	Ile	Val	Glu	Asp	Val	Asp	Ile	Phe	Val	Asn	Cys	Ile	Tyr	Leu	
				245					250					255		
tcc	gcc	aag	atc	cct	cct	ttc	gtc	aac	gcc	gag	acc	ctc	tct	act	cct	816
Ser	Ala	Lys	Ile	Pro	Pro	Phe	Val	Asn	Ala	Glu	Thr	Leu	Ser	Thr	Pro	
			260					265					270			
aac	cgc	cgc	ttg	tct	gtc	att	tgc	gac	gtg	agc	gct	gac	aca	acc	aac	864
Asn	Arg	Arg	Leu	Ser	Val	Ile	Cys	Asp	Val	Ser	Ala	Asp	Thr	Thr	Asn	
		275					280					285				
ccc	cac	aac	cct	atc	ccc	gtc	tac	tcg	atc	acc	acc	acc	ttt	gat	aag	912
Pro	His	Asn	Pro	Ile	Pro	Val	Tyr	Ser	Ile	Thr	Thr	Thr	Phe	Asp	Lys	
	290					295					300					
ccc	acg	gtg	cca	gtg	acc	ctg	tcc	gcc	ggc	gcc	cag	ggt	ccc	cct	ctg	960
Pro	Thr	Val	Pro	Val	Thr	Leu	Ser	Ala	Gly	Ala	Gln	Gly	Pro	Pro	Leu	
	305				310				315						320	
agc	gtg	atc	agc	att	gat	cat	ctc	ccc	tct	ctc	ttg	ccc	cgt	gag	agc	1008
Ser	Val	Ile	Ser	Ile	Asp	His	Leu	Pro	Ser	Leu	Leu	Pro	Arg	Glu	Ser	
				325					330					335		
tcc	gag	atg	ttc	agc	gaa	gcg	ctg	ctg	cct	agc	ctg	ctg	cag	ctc	aag	1056
Ser	Glu	Met	Phe	Ser	Glu	Ala	Leu	Leu	Pro	Ser	Leu	Leu	Gln	Leu	Lys	
			340					345					350			
gac	aga	aag	aac	gct	cgc	gtc	tgg	aag	cag	gcg	gag	gat	ctg	ttt	aac	1104
Asp	Arg	Lys	Asn	Ala	Arg	Val	Trp	Lys	Gln	Ala	Glu	Asp	Leu	Phe	Asn	
		355					360					365				
gag	aag	gtc	gct	acg	ttg	ccc	gag	tcg	atg	cgc	gca	taa				1143
Glu	Lys	Val	Ala	Thr	Leu	Pro	Glu	Ser	Met	Arg	Ala					
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Ala	Arg	Ser	Ala	Leu	Thr	Pro	Thr	Thr	Cys	Lys	Ala	Leu	Met	Asp	Ala	
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eof-seql.txt

Gly Tyr Asp Val Thr Val Glu Arg Ser Thr Gln Arg Ile Phe Asp Gly  
 35 40 45  
 Glu Asp Asn Ile Leu Ser Tyr Leu Ile Gly Ala Pro Leu Val Glu Glu  
 50 55 60  
 Gly Ser Trp Val Lys Asp Ala Pro Lys Asp Ala Tyr Val Leu Gly Leu  
 65 70 75 80  
 Lys Glu Leu Pro Glu Asp Asp Phe Pro Leu Glu His Val His Ile Ser  
 85 90 95  
 Phe Ala His Cys Tyr Lys Glu Gln Gly Gly Trp Glu Lys Val Leu Ser  
 100 105 110  
 Arg Trp Pro Arg Gly Gly Gly Thr Leu Leu Asp Leu Glu Phe Leu Thr  
 115 120 125  
 Asp Asp Val Gly Arg Arg Val Ala Ala Phe Gly Tyr Ser Ala Gly Tyr  
 130 135 140  
 Ala Gly Ser Ala Leu Ala Val Lys Asn Trp Ala Trp Gln Leu Thr His  
 145 150 155 160  
 Pro Glu Gly Glu Pro Leu Pro Gly Glu Thr Pro Tyr Glu Asn Gln Asp  
 165 170 175  
 Leu Leu Ile Ala Ser Val Lys Glu Ser Leu Glu Val Gly Lys Lys Gln  
 180 185 190  
 Ser Gly Lys Ser Pro Lys Val Leu Val Ile Gly Ala Leu Gly Arg Cys  
 195 200 205  
 Gly Lys Gly Ala Val Gln Leu Ala Lys Asp Val Gly Ile Pro Glu Ser  
 210 215 220  
 Asp Ile Ile Gln Trp Asp Ile Glu Glu Thr Lys Lys Gly Gly Pro Phe  
 225 230 235 240  
 Arg Glu Ile Val Glu Asp Val Asp Ile Phe Val Asn Cys Ile Tyr Leu  
 245 250 255  
 Ser Ala Lys Ile Pro Pro Phe Val Asn Ala Glu Thr Leu Ser Thr Pro  
 260 265 270  
 Asn Arg Arg Leu Ser Val Ile Cys Asp Val Ser Ala Asp Thr Thr Asn  
 275 280 285  
 Pro His Asn Pro Ile Pro Val Tyr Ser Ile Thr Thr Thr Phe Asp Lys  
 290 295 300

eof-seq1.txt

Pro Thr Val Pro Val Thr Leu Ser Ala Gly Ala Gln Gly Pro Pro Leu  
305 310 315 320

Ser Val Ile Ser Ile Asp His Leu Pro Ser Leu Leu Pro Arg Glu Ser  
325 330 335

Ser Glu Met Phe Ser Glu Ala Leu Leu Pro Ser Leu Leu Gln Leu Lys  
340 345 350

Asp Arg Lys Asn Ala Arg Val Trp Lys Gln Ala Glu Asp Leu Phe Asn  
355 360 365

Glu Lys Val Ala Thr Leu Pro Glu Ser Met Arg Ala  
370 375 380

<210> 3  
<211> 1125  
<212> DNA  
<213> Aspergillus fumigatus

<220>  
<221> CDS  
<222> (1)..(1125)  
<223> Lys1

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gct cgg tct gct ttg acc ccg act acc tgc aag gcc ctt atg gat gct	96
Ala Arg Ser Ala Leu Thr Pro Thr Thr Cys Lys Ala Leu Met Asp Ala	
20 25 30	
ggc tac gag gtg acc gtg gaa cgt tcc aga cag cgg att ttc gac gtt	144
Gly Tyr Glu Val Thr Val Glu Arg Ser Arg Gln Arg Ile Phe Asp Val	
35 40 45	
gta cag atc ggc gcc ccc ctc gtc gag gaa ggt tca tgg gta aag gac	192
Val Gln Ile Gly Ala Pro Leu Val Glu Glu Gly Ser Trp Val Lys Asp	
50 55 60	
gca ccc aaa gat gcc tac atc ctc ggt ctg aag gag ctt ccc gag gac	240
Ala Pro Lys Asp Ala Tyr Ile Leu Gly Leu Lys Glu Leu Pro Glu Asp	
65 70 75 80	
gat ttt ccg ctt gag cac gta cac atc tcc ttt gcg cac tgc tac aag	288
Asp Phe Pro Leu Glu His Val His Ile Ser Phe Ala His Cys Tyr Lys	
85 90 95	
caa cag gct ggc tgg gag aag gtg ctc agc cgg tgg ccc cgc gga ggc	336
Gln Gln Ala Gly Trp Glu Lys Val Leu Ser Arg Trp Pro Arg Gly Gly	
100 105 110	
ggc acc ctc ttg gac ttg gag ttc ctc aca gat gag act gga cgc cga	384
Gly Thr Leu Leu Asp Leu Glu Phe Leu Thr Asp Glu Thr Gly Arg Arg	
115 120 125	
gta gct gct ttt ggg tac tcc gcc ggt tac gca ggt tct gct ttg gcc	432
Val Ala Ala Phe Gly Tyr Ser Ala Gly Tyr Ala Gly Ser Ala Leu Ala	
130 135 140	

eof-seq1.txt

att Ile 145	aag Lys	aac Asn	tgg Trp	gcc Ala 150	tgg Trp 150	caa Gln	ttg Leu	acg Thr	cat His	cct Pro 155	gag Glu	ggc Gly	gag Glu	ccg Pro	ctt Leu 160	480
cct Pro	ggc Gly	gag Glu	act Thr	ccc Pro 165	tac Tyr	gca Ala	aac Asn	cag Gln	gat Asp 170	ctg Leu	ttg Leu	att Ile	gag Glu	tca Ser 175	gtg Val	528
aag Lys	gag Glu	tcg Ser	ttg Leu 180	gag Glu	tct Ser	ggc Gly	aag Lys	aag Lys 185	ctg Leu	tcc Ser	ggc Gly	agg Arg	ccg Pro 190	ccc Pro	aag Lys	576
gtg Val	ctt Leu	gtc Val 195	att Ile	gga Gly	gct Ala	ctt Leu	gga Gly 200	cgc Arg	tgt Cys	ggc Gly	aaa Lys	gga Gly 205	gcg Ala	gtt Val	cag Gln	624
ctg Leu 210	gcc Ala	aag Lys	gat Asp	gtc Val	ggc Gly 215	att Ile 215	cct Pro	gag Glu	tcg Ser	gat Asp 220	atc Ile 220	atc Ile	cag Gln	tgg Trp	gat Asp	672
ata Ile 225	gaa Glu	gag Glu	acc Thr	aag Lys	aag Lys 230	ggt Gly	ggc Gly	ccc Pro	ttc Phe	aga Arg 235	gag Glu	att Ile	gtc Val	gag Glu	gat Asp 240	720
gca Ala	gac Asp	att Ile	ttc Phe	atc Ile 245	aac Asn	tgc Cys	atc Ile	tac Tyr	ctc Leu 250	tcc Ser	gct Ala	aag Lys	atc Ile	cct Pro 255	cct Pro	768
ttc Phe	gtc Val	aac Asn	acc Thr 260	gaa Glu	act Thr	ctg Leu	tct Ser	tct Ser 265	cct Pro	aac Asn	cgc Arg	cgc Arg	ttg Leu 270	tcc Ser	gtc Val	816
att Ile	tgt Cys	gac Asp 275	gtg Val	agc Ser	gcc Ala	gac Asp 280	aca Thr	acc Thr	aat Asn	ccc Pro	aac Asn	aat Asn 285	cct Pro	atc Ile	cct Pro	864
gtc Val 290	tat Tyr	tcc Ser	atc Ile	aca Thr	acc Thr	act Thr 295	ttc Phe	gac Asp	aag Lys	ccc Pro	aca Thr 300	gtc Val	act Thr	gtt Val	cct Pro	912
ctt Leu 305	ccg Pro	gaa Glu	ctg Leu	gcc Ala	cag Gln 310	ggc Gly	cct Pro	cca Pro	ttg Leu	agc Ser 315	gtg Val	atc Ile	agc Ser	atc Ile	gac Asp 320	960
cac His	ctc Leu	ccc Pro	tcc Ser	ctc Leu 325	ctt Leu	cct Pro	cgt Arg	gaa Glu	agc Ser 330	tcc Ser	gag Glu	atg Met	ttc Phe	agc Ser 335	gaa Glu	1008
gcc Ala	tta Leu	ctg Leu	ccg Pro 340	agc Ser	cta Leu	ctg Leu	caa Gln 345	ctc Leu	aag Lys	gat Asp	aga Arg	aag Lys	aac Asn 350	gct Ala	cgt Arg	1056
gtc Val	tgg Trp	aag Lys 355	caa Gln	gca Ala	gag Glu	gac Asp	ttg Leu 360	ttc Phe	aac Asn	gaa Glu	aag Lys	gtt Val 365	gct Ala	acc Thr	ttg Leu	1104
ccc Pro 370	gag Glu	tcg Ser	atg Met	cgc Arg	gct Ala	taa										1125

<210> 4  
 <211> 374  
 <212> PRT  
 <213> Aspergillus fumigatus

<400> 4

eof-seq1.txt

Met Ser Ser Asn Lys Ile Trp Leu Arg Ala Glu Thr Lys Pro Ala Glu  
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Ala Arg Ser Ala Leu Thr Pro Thr Thr Cys Lys Ala Leu Met Asp Ala  
20 25 30

Gly Tyr Glu Val Thr Val Glu Arg Ser Arg Gln Arg Ile Phe Asp Val  
35 40 45

Val Gln Ile Gly Ala Pro Leu Val Glu Glu Gly Ser Trp Val Lys Asp  
50 55 60

Ala Pro Lys Asp Ala Tyr Ile Leu Gly Leu Lys Glu Leu Pro Glu Asp  
65 70 75 80

Asp Phe Pro Leu Glu His Val His Ile Ser Phe Ala His Cys Tyr Lys  
85 90 95

Gln Gln Ala Gly Trp Glu Lys Val Leu Ser Arg Trp Pro Arg Gly Gly  
100 105 110

Gly Thr Leu Leu Asp Leu Glu Phe Leu Thr Asp Glu Thr Gly Arg Arg  
115 120 125

Val Ala Ala Phe Gly Tyr Ser Ala Gly Tyr Ala Gly Ser Ala Leu Ala  
130 135 140

Ile Lys Asn Trp Ala Trp Gln Leu Thr His Pro Glu Gly Glu Pro Leu  
145 150 155 160

Pro Gly Glu Thr Pro Tyr Ala Asn Gln Asp Leu Leu Ile Glu Ser Val  
165 170 175

Lys Glu Ser Leu Glu Ser Gly Lys Lys Leu Ser Gly Arg Pro Pro Lys  
180 185 190

Val Leu Val Ile Gly Ala Leu Gly Arg Cys Gly Lys Gly Ala Val Gln  
195 200 205

Leu Ala Lys Asp Val Gly Ile Pro Glu Ser Asp Ile Ile Gln Trp Asp  
210 215 220

Ile Glu Glu Thr Lys Lys Gly Gly Pro Phe Arg Glu Ile Val Glu Asp  
225 230 235 240

Ala Asp Ile Phe Ile Asn Cys Ile Tyr Leu Ser Ala Lys Ile Pro Pro  
245 250 255

Phe Val Asn Thr Glu Thr Leu Ser Ser Pro Asn Arg Arg Leu Ser Val  
260 265 270

eof-seql.txt

Ile Cys Asp Val Ser Ala Asp Thr Thr Asn Pro Asn Asn Pro Ile Pro  
275 280 285

Val Tyr Ser Ile Thr Thr Thr Phe Asp Lys Pro Thr Val Thr Val Pro  
290 295 300

Leu Pro Glu Leu Ala Gln Gly Pro Pro Leu Ser Val Ile Ser Ile Asp  
305 310 315 320

His Leu Pro Ser Leu Leu Pro Arg Glu Ser Ser Glu Met Phe Ser Glu  
325 330 335

Ala Leu Leu Pro Ser Leu Leu Gln Leu Lys Asp Arg Lys Asn Ala Arg  
340 345 350

Val Trp Lys Gln Ala Glu Asp Leu Phe Asn Glu Lys Val Ala Thr Leu  
355 360 365

Pro Glu Ser Met Arg Ala  
370

<210> 5  
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<212> DNA  
<213> Botrytis cinerea

<220>  
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<222> (1)..(1119)  
<223> Lys1

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gag cat cga tcc gct ctt acc ccc acc acc gcc aaa gct ctc atc gat	96
Glu His Arg Ser 20 Ala Leu Thr Pro 25 Thr Ala Lys Ala 30 Leu Ile Asp	
gct gga tac aca atc aac gtt gag cgc agt cca gag cgt ata ttc gac	144
Ala Gly Tyr Thr 35 Ile Asn Val Glu Arg Ser Pro Glu Arg 45 Ile Phe Asp	
gat gaa gag ttc gag aag gtt ggt gct act ctt gtg cca gag aac aca	192
Asp Glu Glu Phe Glu Lys 55 Val Gly Ala Thr Leu Val 60 Pro Glu Asn Thr	
tgg aga caa gca cca aag gat cac att atc att ggg ttg aag gaa ctg	240
Trp Arg Gln Ala Pro Lys 70 Asp His Ile Ile 75 Gly Leu Lys Glu 80 Leu	
ccc gtc gaa gaa ttt cct ctc gag cat gtt cac gta caa ttc gca cat	288
Pro Val Glu Glu 85 Phe Pro Leu Glu His 90 Val His Val Gln Phe Ala His	
tgt tat aaa caa cag ggc ggc tgg gac act gtt cta tca cga ttc cct	336
Cys Tyr Lys 100 Gln Gln Gly Gly Trp 105 Asp Thr Val Leu Ser Arg 110 Phe Pro	

eof-seql.txt

cgt Arg	gga Gly	ggt Gly 115	gga Gly	act Thr	ctc Leu	tta Leu	gat Asp 120	ctt Leu	gag Glu	ttt Phe	ttg Leu	aca Thr 125	gac Asp	gac Asp	aga Arg	384
ggc Gly	aga Arg 130	aga Arg	gtt Val	gca Ala	gcc Ala	ttt Phe 135	gga Gly	tac Tyr	cat His	gct Ala	gga Gly 140	ttt Phe	gct Ala	ggt Gly	gca Ala	432
gca Ala 145	ttg Leu	gca Ala	ctc Leu	gaa Glu	aat Asn 150	tgg Trp	gcg Ala	tgg Trp	caa Gln	ctc Leu 155	acc Thr	cac His	cca Pro	gca Ala	tcc Ser 160	480
gag Glu	ccc Pro	ttc Phe	cca Pro	agt Ser 165	gta Val	tgc Ser	agc Ser	tac Tyr	ccc Pro 170	aac Asn	gaa Glu	gat Asp	gaa Glu	ttg Leu 175	att Ile	528
gtg Val	gat Asp	gtt Val	aag Lys 180	aag Lys	gca Ala	atc Ile	gca Ala 185	gct Ala	gga Gly	caa Gln	gag Glu	aag Lys	acc Thr 190	ggc Gly	aag Lys	576
gca Ala	cca Pro	cga Arg 195	gtt Val	ttg Leu	gtt Val	att Ile	ggt Gly 200	gca Ala	tta Leu	ggc Gly	aga Arg	tgt Cys 205	gga Gly	agt Ser	gga Gly	624
gca Ala 210	gtt Val	gac Asp	ctc Leu	tgc Cys	ttg Leu	aga Arg 215	gct Ala	ggt Gly	gtg Val	cca Pro	acc Thr 220	gaa Glu	aat Asn	gtg Val	ttg Leu	672
aag Lys 225	tgg Trp	gat Asp	atg Met	gcc Ala	gag Glu 230	acc Thr	gct Ala	aag Lys	gga Gly	ggg Gly 235	cca Pro	ttc Phe	cca Pro	gag Glu	atc Ile 240	720
gtt Val	gag Glu	agt Ser	gac Asp	atc Ile 245	ttc Phe	atc Ile	aac Asn	tgc Cys	ata Ile 250	tat Tyr	ctc Leu	atg Met	tcc Ser	aag Lys 255	att Ile	768
cca Pro	aac Asn	ttt Phe	gtc Val 260	gac Asp	atg Met	caa Gln	agc Ser	ctc Leu 265	gat Asp	acc Thr	cca Pro	aac Asn	cgc Arg 270	aaa Lys	ttg Leu	816
tca Ser	gtc Val 275	gtc Val	tgc Cys	gat Asp	gtc Val	agt Ser	gct Ala 280	gat Asp	acc Thr	acc Thr	aac Asn	ccc Pro 285	aac Asn	aac Asn	cca Pro	864
att Ile 290	cca Pro	atc Ile	tat Tyr	act Thr	gtt Val	gca Ala 295	acc Thr	aca Thr	ttt Phe	tca Ser	gag Glu 300	cca Pro	act Thr	gtt Val	cca Pro	912
gtt Val 305	gag Glu	gtc Val	aag Lys	gga Gly	gaa Glu 310	cca Pro	aga Arg	tta Leu	agt Ser	gtc Val 315	atc Ile	agt Ser	att Ile	gat Asp	cac His 320	960
ttg Leu	cca Pro	agt Ser	tta Leu	ttg Leu 325	cca Pro	aga Arg	gag Glu	gca Ala 330	agt Ser	gag Glu	gca Ala	ttt Phe	agc Ser	aag Lys 335	gac Asp	1008
tta Leu	tta Leu	cca Pro	agt Ser 340	ttg Leu	ttg Leu	tct Ser	ttg Leu	aag Lys 345	gat Asp	tgg Trp	aga Arg	aat Asn	act Thr 350	cca Pro	gtc Val	1056
tgg Trp	gcc Ala	aag Lys 355	gca Ala	gag Glu	aag Lys	tta Leu	ttc Phe 360	cag Gln	gaa Glu	aag Lys	gtt Val	gct Ala 365	act Thr	ttg Leu	ccc Pro	1104
aag Lys	aac Asn	gag Glu	gca Ala	tga												1119

eof-seql.txt

<210> 6  
 <211> 372  
 <212> PRT  
 <213> Botrytis cinerea

<400> 6

Met Ser Gly Thr Thr Leu His Leu Arg Ser Glu Leu Gly Lys Ala Leu  
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Glu His Arg Ser Ala Leu Thr Pro Thr Thr Ala Lys Ala Leu Ile Asp  
 20 25 30

Ala Gly Tyr Thr Ile Asn Val Glu Arg Ser Pro Glu Arg Ile Phe Asp  
 35 40 45

Asp Glu Glu Phe Glu Lys Val Gly Ala Thr Leu Val Pro Glu Asn Thr  
 50 55 60

Trp Arg Gln Ala Pro Lys Asp His Ile Ile Ile Gly Leu Lys Glu Leu  
 65 70 75 80

Pro Val Glu Glu Phe Pro Leu Glu His Val His Val Gln Phe Ala His  
 85 90 95

Cys Tyr Lys Gln Gln Gly Gly Trp Asp Thr Val Leu Ser Arg Phe Pro  
 100 105 110

Arg Gly Gly Gly Thr Leu Leu Asp Leu Glu Phe Leu Thr Asp Asp Arg  
 115 120 125

Gly Arg Arg Val Ala Ala Phe Gly Tyr His Ala Gly Phe Ala Gly Ala  
 130 135 140

Ala Leu Ala Leu Glu Asn Trp Ala Trp Gln Leu Thr His Pro Ala Ser  
 145 150 155 160

Glu Pro Phe Pro Ser Val Ser Ser Tyr Pro Asn Glu Asp Glu Leu Ile  
 165 170 175

Val Asp Val Lys Lys Ala Ile Ala Ala Gly Gln Glu Lys Thr Gly Lys  
 180 185 190

Ala Pro Arg Val Leu Val Ile Gly Ala Leu Gly Arg Cys Gly Ser Gly  
 195 200 205

Ala Val Asp Leu Cys Leu Arg Ala Gly Val Pro Thr Glu Asn Val Leu  
 210 215 220

Lys Trp Asp Met Ala Glu Thr Ala Lys Gly Gly Pro Phe Pro Glu Ile  
 225 230 235 240

eof-seql.txt

Val Glu Ser Asp Ile Phe Ile Asn Cys Ile Tyr Leu Met Ser Lys Ile  
245 250 255

Pro Asn Phe Val Asp Met Gln Ser Leu Asp Thr Pro Asn Arg Lys Leu  
260 265 270

Ser Val Val Cys Asp Val Ser Ala Asp Thr Thr Asn Pro Asn Asn Pro  
275 280 285

Ile Pro Ile Tyr Thr Val Ala Thr Thr Phe Ser Glu Pro Thr Val Pro  
290 295 300

Val Glu Val Lys Gly Glu Pro Arg Leu Ser Val Ile Ser Ile Asp His  
305 310 315 320

Leu Pro Ser Leu Leu Pro Arg Glu Ala Ser Glu Ala Phe Ser Lys Asp  
325 330 335

Leu Leu Pro Ser Leu Leu Ser Leu Lys Asp Trp Arg Asn Thr Pro Val  
340 345 350

Trp Ala Lys Ala Glu Lys Leu Phe Gln Glu Lys Val Ala Thr Leu Pro  
355 360 365

Lys Asn Glu Ala  
370

<210> 7  
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<212> DNA  
<213> Fusarium graminearum

<220>  
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<222> (1)..(1173)  
<223> Lys1

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gag cac cga tct ttc tct ccc tca att atc aag act ctt gtc gac gct	96
Glu His Arg Ser Phe Ser Pro Ser Ile Ile Lys Thr Leu Val Asp Ala	
20 25 30	
gga tat ccc gtc tct gtc gag cga tca tct aca gac ccc aag ttc aag	144
Gly Tyr Pro Val Ser Val Glu Arg Ser Ser Thr Asp Pro Lys Phe Lys	
35 40 45	
cgc atc ttt gaa gac tcc gag tat gag gct gct ggt gcg cgc ctc gtc	192
Arg Ile Phe Glu Asp Ser Glu Tyr Glu Ala Ala Gly Ala Arg Leu Val	
50 55 60	
aac gag ggc acc tgg ccc aac gcc gaa gcc gga act ttg att ctg ggc	240
Asn Glu Gly Thr Trp Pro Asn Ala Glu Ala Gly Thr Leu Ile Leu Gly	
65 70 75 80	

eof-seq1.txt

ctc Leu	aag Lys	gag Glu	att Ile	ccc Pro 85	gag Glu	gaa Glu	gac Asp	ttt Phe	cct Pro 90	ctc Leu	aag Lys	aac Asn	gac Asp	cac His 95	att Ile	288
tca Ser	ttt Phe	gcc Ala	cat His 100	tgt Cys	tac Tyr	aag Lys	aac Asn	caa Gln 105	ggt Gly	gga Gly	tgg Trp	gaa Glu	aag Lys 110	gtc Val	ctc Leu	336
ggc Gly	cgc Arg	ttc Phe 115	cct Pro	caa Gln	gga Gly	agc Ser	agc Ser 120	gtt Val	cta Leu	tac Tyr	gat Asp	ttg Leu 125	gag Glu	ttc Phe	ctt Leu	384
gtc Val	gat Asp 130	gag Glu	caa Gln	gga Gly	cgc Arg	cga Arg 135	gtt Val	tct Ser	gct Ala	ttc Phe	ggt Gly 140	ttc Phe	cac His	gct Ala	gga Gly	432
ttc Phe 145	gct Ala	ggc Gly	gct Ala	gcc Ala	ctc Leu 150	ggt Gly	atc Ile	aag Lys	act Thr	ctt Leu 155	gcc Ala	cac His	cag Gln	ctt Leu	cag Gln 160	480
gac Asp	ccc Pro	tcc Ser	tcc Ser	aag Lys 165	ctc Leu	ccc Pro	tca Ser	gtg Val	gag Glu 170	aca Thr	ttc Phe	acc Thr	gat Asp	ggt Gly 175	cgt Arg	528
gga Gly	tac Tyr	tac Tyr	cta Leu 180	aac Asn	gag Glu	gag Glu	gag Glu	ctc Leu 185	gtt Val	aac Asn	cag Gln	atc Ile	cga Arg 190	gaa Glu	gac Asp	576
ctt Leu	gcc Ala	aag Lys 195	gcc Ala	gag Glu	aag Lys	tct Ser	ctt Leu 200	gga Gly	cgc Arg	aag Lys	ccc Pro	act Thr 205	gct Ala	ctc Leu	gtc Val	624
ctc Leu	ggt Gly 210	gct Ala	ctt Leu	gga Gly	cga Arg	tgt Cys 215	ggt Gly	aag Lys	ggt Gly	gcc Ala	gtt Val 220	gac Asp	ctg Leu	ttc Phe	ctc Leu	672
aag Lys 225	gct Ala	ggc Gly	atg Met	ccc Pro	gac Asp 230	gag Glu	aac Asn	atc Ile	acc Thr	cgc Arg 235	tgg Trp	gac Asp	ttg Leu	aac Asn	gag Glu 240	720
acc Thr	aag Lys	gac Asp	cga Arg	gat Asp 245	ggt Gly	cct Pro	tat Tyr	gag Glu	gag Glu 250	att Ile	gcc Ala	cag Gln	gcc Ala	gat Asp 255	gtc Val	768
ttc Phe	ctt Leu	aac Asn	gcc Ala 260	atc Ile	tac Tyr	ctt Leu	tcc Ser	aag Lys 265	ccc Pro	atc Ile	cct Pro	cct Pro	ttc Phe 270	atc Ile	aac Asn	816
gag Glu	gag Glu	ctt Leu 275	ctc Leu	gcc Ala	aag Lys	tct Ser	ggc Gly 280	cgc Arg	aac Asn	ttg Leu	gct Ala	gtt Val 285	gtt Val	atc Ile	gat Asp	864
gtc Val	tcc Ser 290	tgc Cys	gac Asp	acc Thr	acc Thr	aac Asn 295	cct Pro	cac His	aac Asn	ccc Pro	atc Ile 300	ccc Pro	atc Ile	tac Tyr	tct Ser	912
atc Ile 305	aac Asn	acc Thr	aca Thr	ttc Phe	gag Glu 310	gag Glu	ccc Pro	acc Thr	gtc Val	ccc Pro 315	gtc Val	gag Glu	atc Ile	aag Lys	aac Asn 320	960
gac Asp	cag Gln	aac Asn	tct Ser	ctc Leu 325	cct Pro	cta Leu	tca Ser	gtt Val	att Ile 330	agc Ser	atc Ile	gac Asp	cat His	ctc Leu 335	ccc Pro	1008
tca Ser	atg Met	ctg Leu	ccc Pro 340	cgt Arg	gag Glu	gct Ala	agt Ser	gag Glu 345	gcc Ala	ttt Phe	agc Ser	gag Glu	ggc Gly 350	ctc Leu	aag Lys	1056

eof-seq1.txt

gag tct ctc ctt aca ctc aag gac cgc aag act tcg cga gtg tgg gcc 1104  
Glu Ser Leu Leu Thr Leu Lys Asp Arg Lys Thr Ser Arg Val Trp Ala  
355 360 365

gac gcc gag aag ctc ttc aac gac aag gtt gct ctg ctt ccc gag tcc 1152  
Asp Ala Glu Lys Leu Phe Asn Glu Lys Val Ala Leu Leu Pro Glu Ser  
370 375 380

ctg cga acc aag aga gtt taa 1173  
Leu Arg Thr Lys Arg Val  
385 390

<210> 8  
<211> 390  
<212> PRT  
<213> Fusarium graminearum

<400> 8

Met Ser Gln Tyr Pro His Ile Leu Leu Arg Ala Glu Glu Lys Pro Leu  
1 5 10 15

Glu His Arg Ser Phe Ser Pro Ser Ile Ile Lys Thr Leu Val Asp Ala  
20 25 30

Gly Tyr Pro Val Ser Val Glu Arg Ser Ser Thr Asp Pro Lys Phe Lys  
35 40 45

Arg Ile Phe Glu Asp Ser Glu Tyr Glu Ala Ala Gly Ala Arg Leu Val  
50 55 60

Asn Glu Gly Thr Trp Pro Asn Ala Glu Ala Gly Thr Leu Ile Leu Gly  
65 70 75 80

Leu Lys Glu Ile Pro Glu Glu Asp Phe Pro Leu Lys Asn Asp His Ile  
85 90 95

Ser Phe Ala His Cys Tyr Lys Asn Gln Gly Gly Trp Glu Lys Val Leu  
100 105 110

Gly Arg Phe Pro Gln Gly Ser Ser Val Leu Tyr Asp Leu Glu Phe Leu  
115 120 125

Val Asp Glu Gln Gly Arg Arg Val Ser Ala Phe Gly Phe His Ala Gly  
130 135 140

Phe Ala Gly Ala Ala Leu Gly Ile Lys Thr Leu Ala His Gln Leu Gln  
145 150 155 160

Asp Pro Ser Ser Lys Leu Pro Ser Val Glu Thr Phe Thr Asp Gly Arg  
165 170 175

Gly Tyr Tyr Leu Asn Glu Glu Glu Leu Val Asn Gln Ile Arg Glu Asp  
180 185 190

eof-seq1.txt

Leu Ala Lys Ala Glu Lys Ser Leu Gly Arg Lys Pro Thr Ala Leu Val  
195 200 205

Leu Gly Ala Leu Gly Arg Cys Gly Lys Gly Ala Val Asp Leu Phe Leu  
210 215 220

Lys Ala Gly Met Pro Asp Glu Asn Ile Thr Arg Trp Asp Leu Asn Glu  
225 230 235 240

Thr Lys Asp Arg Asp Gly Pro Tyr Glu Glu Ile Ala Gln Ala Asp Val  
245 250 255

Phe Leu Asn Ala Ile Tyr Leu Ser Lys Pro Ile Pro Pro Phe Ile Asn  
260 265 270

Glu Glu Leu Leu Ala Lys Ser Gly Arg Asn Leu Ala Val Val Ile Asp  
275 280 285

Val Ser Cys Asp Thr Thr Asn Pro His Asn Pro Ile Pro Ile Tyr Ser  
290 295 300

Ile Asn Thr Thr Phe Glu Glu Pro Thr Val Pro Val Glu Ile Lys Asn  
305 310 315 320

Asp Gln Asn Ser Leu Pro Leu Ser Val Ile Ser Ile Asp His Leu Pro  
325 330 335

Ser Met Leu Pro Arg Glu Ala Ser Glu Ala Phe Ser Glu Gly Leu Lys  
340 345 350

Glu Ser Leu Leu Thr Leu Lys Asp Arg Lys Thr Ser Arg Val Trp Ala  
355 360 365

Asp Ala Glu Lys Leu Phe Asn Glu Lys Val Ala Leu Leu Pro Glu Ser  
370 375 380

Leu Arg Thr Lys Arg Val  
385 390

<210> 9  
<211> 591  
<212> DNA  
<213> Fusarium oxysporum

<220>  
<221> CDS  
<222> (1).. (591)  
<223> Lys1

<400> 9  
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Met Ser Glu Tyr Pro His Ile Leu Leu Arg Ala Glu Glu Lys Pro Leu  
1 5 10 15

eof-seq1.txt

gag	cac	cga	tct	ttc	tcc	ccc	gcc	gtt	atc	aag	aca	ctc	gtc	gat	gct	96
Glu	His	Arg	Ser	Phe	Ser	Pro	Ala	Val	Ile	Lys	Thr	Leu	Val	Asp	Ala	
			20					25					30			
gga	tac	ccc	att	tcc	gtc	gag	cga	tcg	tcc	aca	gac	ccc	aag	ttt	aag	144
Gly	Tyr	Pro	Ile	Ser	Val	Glu	Arg	Ser	Ser	Thr	Asp	Pro	Lys	Phe	Lys	
		35					40					45				
cgt	atc	ttt	gag	gac	tca	gag	tat	gag	gct	gct	ggc	gct	cgt	ctt	gtc	192
Arg	Ile	Phe	Glu	Asp	Ser	Glu	Tyr	Glu	Ala	Ala	Gly	Ala	Arg	Leu	Val	
	50					55					60					
gat	acg	ggt	gtc	tgg	ccc	aac	gct	gag	cct	ggg	aca	atc	atc	ctg	ggg	240
Asp	Thr	Gly	Val	Trp	Pro	Asn	Ala	Glu	Pro	Gly	Thr	Ile	Ile	Leu	Gly	
65					70				75					80		
ctg	aag	gag	ata	ccc	gag	gag	gac	ttc	ccc	ctc	aag	aat	gac	cac	atc	288
Leu	Lys	Glu	Ile	Pro	Glu	Glu	Asp	Phe	Pro	Leu	Lys	Asn	Asp	His	Ile	
				85					90					95		
aca	ttc	gct	cac	tgc	tac	aag	aat	caa	ggg	ggc	tgg	gag	aag	gta	ctc	336
Thr	Phe	Ala	His	Cys	Tyr	Lys	Asn	Gln	Gly	Gly	Trp	Glu	Lys	Val	Leu	
			100					105					110			
ggt	cgc	tgg	gca	caa	ggc	ggc	tca	gtt	ttg	tat	gac	ttg	gag	ttc	ttg	384
Gly	Arg	Trp	Ala	Gln	Gly	Gly	Ser	Val	Leu	Tyr	Asp	Leu	Glu	Phe	Leu	
		115					120					125				
cac	gat	tct	gaa	ggt	cgt	cgg	gtt	tcg	gcc	ttc	ggt	ttc	cac	gct	ggt	432
His	Asp	Ser	Glu	Gly	Arg	Arg	Val	Ser	Ala	Phe	Gly	Phe	His	Ala	Gly	
	130					135					140					
ttt	gca	gga	gct	gcg	ctc	gga	att	aag	acg	ctt	gct	cac	cag	ctg	cag	480
Phe	Ala	Gly	Ala	Ala	Leu	Gly	Ile	Lys	Thr	Leu	Ala	His	Gln	Leu	Gln	
145					150					155				160		
gac	tcg	tct	tct	aag	ctc	cct	tca	gtc	gag	aca	ttc	acc	gat	ggc	cgc	528
Asp	Ser	Ser	Ser	Lys	Leu	Pro	Ser	Val	Glu	Thr	Phe	Thr	Asp	Gly	Arg	
				165					170					175		
gga	tac	tac	ctg	aac	gag	gat	gag	ctc	gtc	aac	cca	gat	tcg	cga	gga	576
Gly	Tyr	Tyr	Leu	Asn	Glu	Asp	Glu	Leu	Val	Asn	Pro	Asp	Ser	Arg	Gly	
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tct	cac	caa	ggc	tga												591
Ser	His	Gln	Gly													
		195														

<210> 10  
 <211> 196  
 <212> PRT  
 <213> Fusarium oxysporum

<400> 10

Met	Ser	Glu	Tyr	Pro	His	Ile	Leu	Leu	Arg	Ala	Glu	Glu	Lys	Pro	Leu	
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Glu	His	Arg	Ser	Phe	Ser	Pro	Ala	Val	Ile	Lys	Thr	Leu	Val	Asp	Ala	
			20					25					30			
Gly	Tyr	Pro	Ile	Ser	Val	Glu	Arg	Ser	Ser	Thr	Asp	Pro	Lys	Phe	Lys	
		35					40					45				

eof-seq1.txt

Arg Ile Phe Glu Asp Ser Glu Tyr Glu Ala Ala Gly Ala Arg Leu Val  
50 55 60

Asp Thr Gly Val Trp Pro Asn Ala Glu Pro Gly Thr Ile Ile Leu Gly  
65 70 75 80

Leu Lys Glu Ile Pro Glu Glu Asp Phe Pro Leu Lys Asn Asp His Ile  
85 90 95

Thr Phe Ala His Cys Tyr Lys Asn Gln Gly Gly Trp Glu Lys Val Leu  
100 105 110

Gly Arg Trp Ala Gln Gly Gly Ser Val Leu Tyr Asp Leu Glu Phe Leu  
115 120 125

His Asp Ser Glu Gly Arg Arg Val Ser Ala Phe Gly Phe His Ala Gly  
130 135 140

Phe Ala Gly Ala Ala Leu Gly Ile Lys Thr Leu Ala His Gln Leu Gln  
145 150 155 160

Asp Ser Ser Ser Lys Leu Pro Ser Val Glu Thr Phe Thr Asp Gly Arg  
165 170 175

Gly Tyr Tyr Leu Asn Glu Asp Glu Leu Val Asn Pro Asp Ser Arg Gly  
180 185 190

Ser His Gln Gly  
195

<210> 11  
<211> 1104  
<212> DNA  
<213> Fusarium verticillioides

<220>  
<221> CDS  
<222> (1)..(1104)  
<223> Lys1

<400> 11  
atg tcg cct gtc gtc att cat ctc cgg acc gag aca aag cct cta gag 48  
Met Ser Pro Val Val Ile His Leu Arg Thr Glu Thr Lys Pro Leu Glu  
1 5 10 15

cgt cgc tct cca ttg tca cct gcg act gct aag gct ttg ctc ggc gct 96  
Arg Arg Ser Pro Leu Ser Pro Ala Thr Ala Lys Ala Leu Leu Gly Ala  
20 25 30

ggc tac aca gtt cga gtt gag gaa tcc ccg gac cgc att tac aag att 144  
Gly Tyr Thr Val Arg Val Glu Glu Ser Pro Asp Arg Ile Tyr Lys Ile  
35 40 45

gac gag ttc aga gat gtt ggg gct gaa atc gta ccc gct ggc tcg tgg 192  
Asp Glu Phe Arg Asp Val Gly Ala Glu Ile Val Pro Ala Gly Ser Trp  
50 55 60

eof-seql.txt

gtg Val 65	aat Asn	gca Ala	ccc Pro	aag Lys	gag Glu 70	gat Asp	att Ile	atc Ile	ctg Leu	gga Gly 75	ttg Leu	aag Lys	gag Glu	att Ile	gag Glu 80	240
gcg Ala	aat Asn	ggt Gly	aca Thr	ccg Pro 85	tta Leu	ctt Leu	cac His	acc Thr	tac Tyr 90	att Ile	cac His	ttt Phe	gct Ala	cat His 95	gta Val	288
ttc Phe	aag Lys	aag Lys	caa Gln 100	agc Ser	ggc Gly	tgg Trp	gct Ala	act Thr 105	gag Glu	ctg Leu	tcc Ser	cgc Arg	ttc Phe 110	gca Ala	aac Asn	336
gcg Ala	ggt Gly	ggc Gly 115	tta Leu	cta Leu	tat Tyr	gac Asp	ttg Leu 120	gaa Glu	ttt Phe	ctt Leu	acg Thr	gac Asp 125	caa Gln	gat Asp	gga Gly	384
cgc Arg	cgg Arg 130	gta Val	gct Ala	gca Ala	ttc Phe	gga Gly 135	tat Tyr	tgg Trp	gcg Ala	gga Gly	tat Tyr 140	gcc Ala	ggg Gly	acc Thr	gct Ala	432
ctg Leu 145	gca Ala	ctc Leu	ctg Leu	tct Ser	tgg Trp 150	gct Ala	cac His	cag Gln	ctg Leu	ctc Leu 155	aac Asn	cct Pro	ggc Gly	gta Val	ccc Pro 160	480
caa Gln	gga Gly	ccg Pro	gtt Val	ccc Pro 165	gtc Val	ttc Phe	gac Asp	tct Ser	gcc Ala 170	tct Ser	gct Ala	ttg Leu	act Thr	gaa Glu 175	ctt Leu	528
gtc Val	aag Lys	ggt Gly	aaa Lys 180	gtc Val	gac Asp	gct Ala	gca Ala	cgc Arg 185	tcc Ser	gcc Ala	aac Asn	cac His	ggc Gly 190	gcg Ala	ctc Leu	576
cct Pro	cga Arg	ctg Leu 195	atc Ile	gta Val	atc Ile	ggt Gly	gcc Ala 200	ctg Leu	ggc Gly	cgc Arg	tgc Cys	ggc Gly 205	aaa Lys	ggt Gly	gcc Ala	624
atc Ile 210	gca Ala	gca Ala	gct Ala	gag Glu	gcc Ala	att Ile 215	ggt Gly	gtt Val	agt Ser	gac Asp	att Ile 220	ctg Leu	aaa Lys	tgg Trp	gac Asp	672
att Ile 225	gct Ala	gag Glu	acg Thr	agc Ser	aag Lys 230	ggt Gly	ggc Gly	cca Pro	ttt Phe	cct Pro 235	gaa Glu	gtc Val	gcc Ala	tgc Ser	tct Ser 240	720
gat Asp	att Ile	ttt Phe	gtg Val	aac Asn 245	tgc Cys	gtc Val	tac Tyr	cta Leu	ggt Gly 250	tcc Ser	aac Asn	aag Lys	att Ile	cca Pro 255	ccc Pro	768
ttc Phe	aca Thr	act Thr	ttt Phe 260	gaa Glu	gca Ala	ctc Leu	tca Ser	gga Gly 265	ccc Pro	ggc Gly	aga Arg	cga Arg	ctc Leu 270	cgg Arg	gtc Val	816
att Ile	tgc Cys	gac Asp 275	gtc Val	agt Ser	tgc Cys	gac Asp	ccc Pro 280	aac Asn	agc Ser	gag Glu	aat Asn	aac Asn 285	cct Pro	gtt Val	ccc Pro	864
gtc Val	tac Tyr 290	tcc Ser	agc Ser	tac Tyr	agt Ser	tgc Ser 295	ttc Phe	gaa Glu	aac Asn	ccg Pro	act Thr 300	gtc Val	cct Pro	gct Ala	tct Ser	912
gag Glu 305	cat His	att Ile	gat Asp	ggt Gly	cct Pro 310	gaa Glu	ctg Leu	cgc Arg	atc Ile	att Ile 315	gct Ala	atc Ile	gat Asp	cat His	ctt Leu 320	960
cct Pro	acc Thr	atg Met	gtt Val	gca Ala 325	cgc Arg	gag Glu	tgc Ser	agt Ser	gac Asp 330	gag Glu	tat Tyr	tcc Ser	tca Ser	cta Leu 335	ctt Leu	1008

eof-seql.txt

ctg cca agt ttg ttg act ctg gac cgc cgg gat act gag ggg gtt tgg 1056  
Leu Pro Ser Leu Leu Thr Leu Asp Arg Arg Asp Thr Glu Gly Val Trp  
340 345 350

cag cga gca gaa cgg atc ttt cgc gag aag gtt gca gag ctg ccc tag 1104  
Gln Arg Ala Glu Arg Ile Phe Arg Glu Lys Val Ala Glu Leu Pro  
355 360 365

<210> 12  
<211> 367  
<212> PRT  
<213> Fusarium verticillioides  
<400> 12

Met Ser Pro Val Val Ile His Leu Arg Thr Glu Thr Lys Pro Leu Glu  
1 5 10 15

Arg Arg Ser Pro Leu Ser Pro Ala Thr Ala Lys Ala Leu Leu Gly Ala  
20 25 30

Gly Tyr Thr Val Arg Val Glu Glu Ser Pro Asp Arg Ile Tyr Lys Ile  
35 40 45

Asp Glu Phe Arg Asp Val Gly Ala Glu Ile Val Pro Ala Gly Ser Trp  
50 55 60

Val Asn Ala Pro Lys Glu Asp Ile Ile Leu Gly Leu Lys Glu Ile Glu  
65 70 75 80

Ala Asn Gly Thr Pro Leu Leu His Thr Tyr Ile His Phe Ala His Val  
85 90 95

Phe Lys Lys Gln Ser Gly Trp Ala Thr Glu Leu Ser Arg Phe Ala Asn  
100 105 110

Ala Gly Gly Leu Leu Tyr Asp Leu Glu Phe Leu Thr Asp Gln Asp Gly  
115 120 125

Arg Arg Val Ala Ala Phe Gly Tyr Trp Ala Gly Tyr Ala Gly Thr Ala  
130 135 140

Leu Ala Leu Leu Ser Trp Ala His Gln Leu Leu Asn Pro Gly Val Pro  
145 150 155 160

Gln Gly Pro Val Pro Val Phe Asp Ser Ala Ser Ala Leu Thr Glu Leu  
165 170 175

Val Lys Gly Lys Val Asp Ala Ala Arg Ser Ala Asn His Gly Ala Leu  
180 185 190

Pro Arg Leu Ile Val Ile Gly Ala Leu Gly Arg Cys Gly Lys Gly Ala  
195 200 205

eof-seq1.txt

Ile Ala Ala Ala Glu Ala Ile Gly Val Ser Asp Ile Leu Lys Trp Asp  
210 215 220

Ile Ala Glu Thr Ser Lys Gly Gly Pro Phe Pro Glu Val Ala Ser Ser  
225 230 235 240

Asp Ile Phe Val Asn Cys Val Tyr Leu Gly Ser Asn Lys Ile Pro Pro  
245 250 255

Phe Thr Thr Phe Glu Ala Leu Ser Gly Pro Gly Arg Arg Leu Arg Val  
260 265 270

Ile Cys Asp Val Ser Cys Asp Pro Asn Ser Glu Asn Asn Pro Val Pro  
275 280 285

Val Tyr Ser Ser Tyr Ser Ser Phe Glu Asn Pro Thr Val Pro Ala Ser  
290 295 300

Glu His Ile Asp Gly Pro Glu Leu Arg Ile Ile Ala Ile Asp His Leu  
305 310 315 320

Pro Thr Met Val Ala Arg Glu Ser Ser Asp Glu Tyr Ser Ser Leu Leu  
325 330 335

Leu Pro Ser Leu Leu Thr Leu Asp Arg Arg Asp Thr Glu Gly Val Trp  
340 345 350

Gln Arg Ala Glu Arg Ile Phe Arg Glu Lys Val Ala Glu Leu Pro  
355 360 365

<210> 13  
<211> 1173  
<212> DNA  
<213> Fusarium verticillioides

<220>  
<221> CDS  
<222> (1)..(1173)  
<223> Lys1

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gga tac ccc atc tcc gtc gag cgg tca tcc acc gat ccc aaa ttt agg Gly Tyr Pro Ile Ser Val Glu Arg Ser Ser Thr Asp Pro Lys Phe Arg 35 40 45	144
cgt atc ttt gag gac tct gaa tat gaa gct gct ggt gct cgt ctt gtc Arg Ile Phe Glu Asp Ser Glu Tyr Glu Ala Ala Gly Ala Arg Leu Val 50 55 60	192

eof-seq1.txt

gac Asp 65	acg Thr	ggt Gly	gtc Val	tgg Trp 70	ccc Pro	aac Asn	gct Ala	gag Glu	cct Pro	ggg Gly 75	aca Thr	atc Ile	ata Ile	ctc Leu	gga Gly 80	240
ttg Leu	aag Lys	gag Glu	att Ile 85	ccc Pro	gag Glu	gag Glu	gat Asp	ttc Phe 90	cct Pro	ctg Leu	aag Lys	aat Asn	gat Asp	cac His 95	atc Ile	288
aca Thr	ttt Phe	gca Ala 100	cac His	tgc Cys	tac Tyr	aaa Lys	aac Asn	cag Gln 105	gga Gly	ggc Gly	tgg Trp	gag Glu	aag Lys 110	gtt Val	ctc Leu	336
ggt Gly	cgt Arg	tgg Trp 115	tct Ser	cgc Arg	ggc Gly	ggg Gly	tct Ser 120	act Thr	ctg Leu	tat Tyr	gac Asp	ttg Leu 125	gag Glu	ttt Phe	ttg Leu	384
cat His 130	gac Asp	gct Ala	gaa Glu	ggc Gly	cga Arg	cgt Arg 135	gtt Val	tct Ser	gcg Ala	ttt Phe	ggt Gly 140	ttc Phe	cac His	gca Ala	ggc Gly	432
ttc Phe 145	gcc Ala	ggg Gly	gct Ala	gcg Ala	ctt Leu 150	gga Gly	ata Ile	aag Lys	aca Thr	ctt Leu 155	tcc Ser	cac His	cag Gln	ctg Leu	cag Gln 160	480
gac Asp	ccg Pro	tca Ser	tct Ser	aag Lys 165	ctt Leu	ccg Pro	tct Ser	gtc Val	gag Glu 170	aca Thr	ttc Phe	acc Thr	gat Asp	ggc Gly 175	cgc Arg	528
gga Gly	tat Tyr	tac Tyr	ctg Leu 180	aac Asn	gaa Glu	gat Asp	gag Glu	ctc Leu 185	gtc Val	aac Asn	cag Gln	att Ile	cgc Arg 190	gag Glu	gat Asp	576
ctc Leu	gcc Ala	aag Lys 195	gct Ala	gag Glu	aag Lys	gct Ala	ctc Leu 200	gga Gly	cgc Arg	aag Lys	ccc Pro	act Thr 205	gct Ala	ctc Leu	gtc Val	624
ctt Leu 210	ggt Gly	gct Ala	ctt Leu	gga Gly	cga Arg	tgt Cys 215	ggc Gly	aag Lys	ggt Gly	gct Ala	gtg Val 220	gac Asp	ctt Leu	ttc Phe	ctc Leu	672
aag Lys 225	gcc Ala	ggc Gly	atg Met	cct Pro	gat Asp 230	gag Glu	aac Asn	atc Ile	acc Thr	cgt Arg 235	tgg Trp	gac Asp	ttg Leu	aac Asn	gaa Glu 240	720
acc Thr	aag Lys	gac Asp	cgt Arg	gat Asp 245	gga Gly	cct Pro	tac Tyr	gag Glu	gaa Glu 250	att Ile	gcg Ala	aag Lys	gct Ala	gat Asp 255	gtc Val	768
ttc Phe	ctt Leu	aac Asn 260	gcc Ala	atc Ile	tac Tyr	ctc Leu	tcc Ser	aag Lys 265	ccc Pro	atc Ile	ccc Pro	cct Pro	ttc Phe 270	atc Ile	aac Asn	816
caa Gln	gaa Glu 275	ctc Leu	ctc Leu	gcc Ala	aag Lys	aag Lys	ggt Gly 280	cgc Arg	aac Asn	ctc Leu	gct Ala	gta Val 285	gtc Val	atc Ile	gac Asp	864
gtt Val 290	tct Ser	tgt Cys	gat Asp	acc Thr	aca Thr	aac Asn 295	ccc Pro	cac His	aac Asn	ccc Pro	atc Ile 300	ccc Pro	atc Ile	tac Tyr	tcc Ser	912
atc Ile 305	aac Asn	acc Thr	acc Thr	ttc Phe	gag Glu 310	gac Asp	ccc Pro	acc Thr	gtc Val	ccc Pro 315	gtc Val	gag Glu	atc Ile	aag Lys	gac Asp 320	960
gac Asp	cag Gln	aac Asn	aac Asn	ctc Leu 325	ccc Pro	ctt Leu	tcc Ser	gtc Val	atc Ile 330	agc Ser	atc Ile	gac Asp	cac His	ctt Leu 335	ccc Pro	1008

eof-seq1.txt

tcc atg ctt ccc cgc gag gct agt gag gcc ttt agt gag ggt ctc aag 1056  
Ser Met Leu Pro Arg Glu Ala Ser Glu Ala Phe Ser Glu Gly Leu Lys  
340 345 350

gag tct ctg ctt aca ctc aag gat cgc aaa act tcg cgg gtg tgg gct 1104  
Glu Ser Leu Leu Thr Leu Lys Asp Arg Lys Thr Ser Arg Val Trp Ala  
355 360 365

gat gct gag aag ctc ttc cat gag aag gtt gct acg ttg ccg gag gag 1152  
Asp Ala Glu Lys Leu Phe His Glu Lys Val Ala Thr Leu Pro Glu Glu  
370 375 380

ttg aga acc aag aac gtt taa 1173  
Leu Arg Thr Lys Asn Val  
385 390

<210> 14  
<211> 390  
<212> PRT  
<213> Fusarium verticillioides

<400> 14

Met Ser Glu Tyr Pro His Ile Leu Leu Arg Ala Glu Glu Lys Pro Leu  
1 5 10 15

Glu His Arg Ser Phe Ser Pro Ala Val Ile Lys Thr Leu Val Asp Ala  
20 25 30

Gly Tyr Pro Ile Ser Val Glu Arg Ser Ser Thr Asp Pro Lys Phe Arg  
35 40 45

Arg Ile Phe Glu Asp Ser Glu Tyr Glu Ala Ala Gly Ala Arg Leu Val  
50 55 60

Asp Thr Gly Val Trp Pro Asn Ala Glu Pro Gly Thr Ile Ile Leu Gly  
65 70 75 80

Leu Lys Glu Ile Pro Glu Glu Asp Phe Pro Leu Lys Asn Asp His Ile  
85 90 95

Thr Phe Ala His Cys Tyr Lys Asn Gln Gly Gly Trp Glu Lys Val Leu  
100 105 110

Gly Arg Trp Ser Arg Gly Gly Ser Thr Leu Tyr Asp Leu Glu Phe Leu  
115 120 125

His Asp Ala Glu Gly Arg Arg Val Ser Ala Phe Gly Phe His Ala Gly  
130 135 140

Phe Ala Gly Ala Ala Leu Gly Ile Lys Thr Leu Ser His Gln Leu Gln  
145 150 155 160

Asp Pro Ser Ser Lys Leu Pro Ser Val Glu Thr Phe Thr Asp Gly Arg  
165 170 175

eof-seql.txt

Gly Tyr Tyr Leu Asn Glu Asp Glu Leu Val Asn Gln Ile Arg Glu Asp  
180 185 190

Leu Ala Lys Ala Glu Lys Ala Leu Gly Arg Lys Pro Thr Ala Leu Val  
195 200 205

Leu Gly Ala Leu Gly Arg Cys Gly Lys Gly Ala Val Asp Leu Phe Leu  
210 215 220

Lys Ala Gly Met Pro Asp Glu Asn Ile Thr Arg Trp Asp Leu Asn Glu  
225 230 235 240

Thr Lys Asp Arg Asp Gly Pro Tyr Glu Glu Ile Ala Lys Ala Asp Val  
245 250 255

Phe Leu Asn Ala Ile Tyr Leu Ser Lys Pro Ile Pro Pro Phe Ile Asn  
260 265 270

Gln Glu Leu Leu Ala Lys Lys Gly Arg Asn Leu Ala Val Val Ile Asp  
275 280 285

Val Ser Cys Asp Thr Thr Asn Pro His Asn Pro Ile Pro Ile Tyr Ser  
290 295 300

Ile Asn Thr Thr Phe Glu Asp Pro Thr Val Pro Val Glu Ile Lys Asp  
305 310 315 320

Asp Gln Asn Asn Leu Pro Leu Ser Val Ile Ser Ile Asp His Leu Pro  
325 330 335

Ser Met Leu Pro Arg Glu Ala Ser Glu Ala Phe Ser Glu Gly Leu Lys  
340 345 350

Glu Ser Leu Leu Thr Leu Lys Asp Arg Lys Thr Ser Arg Val Trp Ala  
355 360 365

Asp Ala Glu Lys Leu Phe His Glu Lys Val Ala Thr Leu Pro Glu Glu  
370 375 380

Leu Arg Thr Lys Asn Val  
385 390

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<211> 1223  
<212> DNA  
<213> Mycosphaerella fijiensis

<220>  
<221> CDS  
<222> (3).. (1223)  
<223> Lys1

eol f-seql . txt

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  Arg Ser Thr Leu Lys Lys Asp Ser Tyr Ser Ala Ser Thr Ile Ile
    1          5          10          15

gta caa gat cag tgt ctc aca atg tgc cct aca gtc ctt cat ttc aga      95
Val Gln Asp Gln Cys Leu Thr Met Ser Pro Thr Val Leu His Phe Arg
          20          25          30

gca gag acc aag cca tta gag cat cgc tgc gca gtc aca ccg aca atc      143
Ala Glu Thr Lys Pro Leu Glu His Arg Ser Ala Val Thr Pro Thr Ile
          35          40          45

gca aag aaa ctt gtc gaa gct ggc tat gaa gtc cac gtc gaa cga tgc      191
Ala Lys Lys Leu Val Glu Ala Gly Tyr Glu Val His Val Glu Arg Ser
          50          55          60

cag ctc agc ata ttc gca gac tcc gag tat gaa ggc act ggt gct aag      239
Gln Leu Ser Ile Phe Ala Asp Ser Glu Tyr Glu Gly Thr Gly Ala Lys
          65          70          75

cta gtg cct act gga tgc tgg aca gag gca ccg aag gag cac att gtc      287
Leu Val Pro Thr Gly Ser Trp Thr Glu Ala Pro Lys Glu His Ile Val
          80          85          90          95

att ggc ttg aag gaa ctc ccc gag gag gac ttt cct ttg aag cat gtt      335
Ile Gly Leu Lys Glu Leu Pro Glu Glu Asp Phe Pro Leu Lys His Val
          100          105          110

cat gtg caa ttt gct cac tgc tac aaa gga caa ggt ggc tgg gac aag      383
His Val Gln Phe Ala His Cys Tyr Lys Gly Gln Gly Gly Trp Asp Lys
          115          120          125

gtt ctc agt aga ttt cca aac ggc aga ggc aca tta ctt gac ctt gaa      431
Val Leu Ser Arg Phe Pro Asn Gly Arg Gly Thr Leu Leu Asp Leu Glu
          130          135          140

ttc ttg gaa gat gag caa gga cga aga gtc gct gca ttt ggc tac cat      479
Phe Leu Glu Asp Glu Gln Gly Arg Arg Val Ala Ala Phe Gly Tyr His
          145          150          155

gcc gga ttt gct ggt gcc gca ctt gct ctc atg gcc tgg tct cac cag      527
Ala Gly Phe Ala Gly Ala Ala Leu Ala Leu Met Ala Trp Ser His Gln
          160          165          170          175

ctc gtg cac ggc aag gac agc cct ctt ccg gga gtg aca cca tac gaa      575
Leu Val His Gly Lys Asp Ser Pro Leu Pro Gly Val Thr Pro Tyr Glu
          180          185          190

aac gaa ggt ctc ctt att tcc gat gtc aag aag gcc atc gaa gct ggc      623
Asn Glu Gly Leu Leu Ile Ser Asp Val Lys Lys Ala Ile Glu Ala Gly
          195          200          205

aag gcc aag ggc ggt tgc ctc cca aga gtg ctc gtt att ggc gga ctc      671
Lys Ala Lys Gly Gly Cys Leu Pro Arg Val Leu Val Ile Gly Gly Leu
          210          215          220

ggc aga tgc ggt cgt ggt gcc gtt gac ctc tgt gtc aag gct ggc gtg      719
Gly Arg Cys Gly Arg Gly Ala Val Asp Leu Cys Val Lys Ala Gly Val
          225          230          235

gaa gac att cta aag tgg gat ctt cca gaa aca agt gcc aaa cca gga      767
Glu Asp Ile Leu Lys Trp Asp Leu Pro Glu Thr Ser Ala Lys Pro Gly
          240          245          250          255

cca tac cag gag atc atc gag tct gac gtg ttt gtc aac tgc atc tac      815
Pro Tyr Gln Glu Ile Ile Glu Ser Asp Val Phe Val Asn Cys Ile Tyr
          260          265          270

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eof-seql . txt

ttg tcg gcc aag atc ccg ccc ttc atc gac cag gca tct ctc gca tcg	863
Leu Ser Ala Lys Ile Pro Pro Phe Ile Asp Gl n Ala Ser Leu Ala Ser	
	275
	280
	285
cca aac cgc aaa ctg tct gtg gtc tgt gat gta tcc tgc gat acc acg	911
Pro Asn Arg Lys Leu Ser Val Val Cys Asp Val Ser Cys Asp Thr Thr	
	290
	295
	300
aac cca aac aat cct atc cca atc tac gac atc aac acg act ttc gac	959
Asn Pro Asn Asn Pro Ile Pro Ile Tyr Asp Ile Asn Thr Thr Phe Asp	
	305
	310
	315
aag cca acg gtg cct gtc aag ctc tcc gct gag gcc aat gat ctt ccg	1007
Lys Pro Thr Val Pro Val Lys Leu Ser Ala Gl u Ala Asn Asp Leu Pro	
	320
	325
	330
	335
ttg agc gta atc agt atc gac cat ctc cca tcc tta ctg ccg cga gaa	1055
Leu Ser Val Ile Ser Ile Asp His Leu Pro Ser Leu Leu Pro Arg Gl u	
	340
	345
	350
gct tcc gaa gct ttc agc tcc gcg ctg ctg ccc agc ttg ctt caa ttg	1103
Ala Ser Gl u Ala Phe Ser Ser Ala Leu Leu Pro Ser Leu Leu Gl n Leu	
	355
	360
	365
aac gac tgg aag aac gca cgt gtt tgg caa caa gcc gag aag ctg ttc	1151
Asn Asp Trp Lys Asn Ala Arg Val Trp Gl n Gl n Ala Gl u Lys Leu Phe	
	370
	375
	380
aag gac aaa tgc gcc act ctc cca gaa ggc gca atc gat agc cat gcc	1199
Lys Asp Lys Cys Ala Thr Leu Pro Gl u Gly Ala Ile Asp Ser His Ala	
	385
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	395
gag ctt ctt gct tcg caa tct tga	1223
Gl u Leu Leu Ala Ser Gl n Ser	
	400
	405

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Gl n Asp Gl n Cys Leu Thr Met Ser Pro Thr Val Leu His Phe Arg Ala	
	20 25 30
Gl u Thr Lys Pro Leu Gl u His Arg Ser Ala Val Thr Pro Thr Ile Ala	
	35 40 45
Lys Lys Leu Val Gl u Ala Gly Tyr Gl u Val His Val Gl u Arg Ser Gl n	
	50 55 60
Leu Ser Ile Phe Ala Asp Ser Gl u Tyr Gl u Gly Thr Gly Ala Lys Leu	
	65 70 75 80
Val Pro Thr Gly Ser Trp Thr Gl u Ala Pro Lys Gl u His Ile Val Ile	
	85 90 95

eof-seq1.txt

Gly Leu Lys Glu Leu Pro Glu Glu Asp Phe Pro Leu Lys His Val His  
 100 105 110  
 Val Gln Phe Ala His Cys Tyr Lys Gly Gln Gly Gly Trp Asp Lys Val  
 115 120 125  
 Leu Ser Arg Phe Pro Asn Gly Arg Gly Thr Leu Leu Asp Leu Glu Phe  
 130 135 140  
 Leu Glu Asp Glu Gln Gly Arg Arg Val Ala Ala Phe Gly Tyr His Ala  
 145 150 155 160  
 Gly Phe Ala Gly Ala Ala Leu Ala Leu Met Ala Trp Ser His Gln Leu  
 165 170 175  
 Val His Gly Lys Asp Ser Pro Leu Pro Gly Val Thr Pro Tyr Glu Asn  
 180 185 190  
 Glu Gly Leu Leu Ile Ser Asp Val Lys Lys Ala Ile Glu Ala Gly Lys  
 195 200 205  
 Ala Lys Gly Gly Cys Leu Pro Arg Val Leu Val Ile Gly Gly Leu Gly  
 210 215 220  
 Arg Cys Gly Arg Gly Ala Val Asp Leu Cys Val Lys Ala Gly Val Glu  
 225 230 235 240  
 Asp Ile Leu Lys Trp Asp Leu Pro Glu Thr Ser Ala Lys Pro Gly Pro  
 245 250 255  
 Tyr Gln Glu Ile Ile Glu Ser Asp Val Phe Val Asn Cys Ile Tyr Leu  
 260 265 270  
 Ser Ala Lys Ile Pro Pro Phe Ile Asp Gln Ala Ser Leu Ala Ser Pro  
 275 280 285  
 Asn Arg Lys Leu Ser Val Val Cys Asp Val Ser Cys Asp Thr Thr Asn  
 290 295 300  
 Pro Asn Asn Pro Ile Pro Ile Tyr Asp Ile Asn Thr Thr Phe Asp Lys  
 305 310 315 320  
 Pro Thr Val Pro Val Lys Leu Ser Ala Glu Ala Asn Asp Leu Pro Leu  
 325 330 335  
 Ser Val Ile Ser Ile Asp His Leu Pro Ser Leu Leu Pro Arg Glu Ala  
 340 345 350  
 Ser Glu Ala Phe Ser Ser Ala Leu Leu Pro Ser Leu Leu Gln Leu Asn  
 355 360 365

eof-seq1.txt

Asp Trp Lys Asn Ala Arg Val Trp Gln Gln Ala Glu Lys Leu Phe Lys  
370 375 380

Asp Lys Cys Ala Thr Leu Pro Glu Gly Ala Ile Asp Ser His Ala Glu  
385 390 395 400

Leu Leu Ala Ser Gln Ser  
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<213> Magnaporthe grisea

<220>  
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tac acc gtc aat gtc gag cgc agt ccc gtc cgc atc ttc gac gac gcc Tyr Thr Val Asn Val Glu Arg Ser Pro Val Arg Ile Phe Asp Asp Ala 35 40 45	144
gag ttc gag gct gtt ggc gca acc ctc gtc ccc gag ggc agc tgg gag Glu Phe Glu Ala Val Gly Ala Thr Leu Val Pro Glu Gly Ser Trp Glu 50 55 60	192
cag att ccc aag gac cac atc gtc att gga ctg aag gag ttg gag gag Gln Ile Pro Lys Asp His Ile Val Ile Gly Leu Lys Glu Leu Glu Glu 65 70 75 80	240
aag gac ttc cct ttg aag cac aca cac atc cag ttc gcc cac tgc tac Lys Asp Phe Pro Leu Lys His Thr His Ile Gln Phe Ala His Cys Tyr 85 90 95	288
aag aac caa gga ggc tgg gac aag gtc ctc cgg agg tat cac gat ggc Lys Asn Gln Gly Gly Trp Asp Lys Val Leu Arg Arg Tyr His Asp Gly 100 105 110	336
gag gga atg ctc ctc gac att gag ttc ctc gag aag gac ggc cgt cgt Glu Gly Met Leu Leu Asp Ile Glu Phe Leu Glu Lys Asp Gly Arg Arg 115 120 125	384
gtt gct gct ttc ggc tac tgg gcc ggc ttt gcc ggt gcc gcg ctg gcc Val Ala Ala Phe Gly Tyr Trp Ala Gly Phe Ala Glu Ala Ala Leu Ala 130 135 140	432
ctt cag aac tgg gcc tgg cag ctg acc aac agc ggc gag ccc ctg ccg Leu Gln Asn Trp Ala Trp Gln Leu Thr Asn Ser Gly Glu Pro Leu Pro 145 150 155 160	480
agc gtc gag agc cgc cct aac gag gct gcc ttg gtc ggc gac atc aag Ser Val Glu Ser Arg Pro Asn Glu Ala Ala Leu Val Gly Asp Ile Lys 165 170 175	528

eof-seq1.txt

gag Glu	gct Ala	ctt Leu	gct Ala 180	ggg Gly	ggc Gly	aag Lys	gaa Glu	aag Lys 185	gcc Ala	ggc Gly	agg Arg	ctg Leu	ccc Pro 190	cgc Arg	gtc Val	576
atc Ile	gtc Val	atc Ile 195	ggt Gly	gcc Ala	ctg Leu	ggc Gly	cgc Arg 200	tgc Cys	ggt Gly	cgt Arg	ggt Gly	gcc Ala 205	gtc Val	gac Asp	atg Met	624
tgc Cys	aag Lys 210	aag Lys	gcc Ala	ggc Gly	ata Ile	ccg Pro 215	gat Asp	gag Glu	aac Asn	atc Ile	ctc Leu 220	aag Lys	tgg Trp	gat Asp	atg Met	672
gag Glu 225	gag Glu	acg Thr	gcc Ala	ccc Pro	ggt Gly 230	gga Gly	ccg Pro	ttc Phe	aag Lys	gag Glu 235	att Ile	gtc Val	gag Glu	agt Ser	gac Asp 240	720
atc Ile	ttt Phe	gtc Val	aac Asn	tgc Cys 245	atc Ile	tac Tyr	ctg Leu	aac Asn	aag Lys 250	cag Gln	atc Ile	ccg Pro	ccg Pro	ttt Phe 255	gtg Val	768
acc Thr	atg Met	gag Glu	tgc Ser 260	ctc Leu	aac Asn	acg Thr	cca Pro	gag Glu 265	cgc Arg	aag Lys	ctc Leu	tcc Ser	gtc Val 270	atc Ile	aac Asn	816
gac Asp	tgc Cys	tgc Ser 275	gct Ala	gac Asp	acg Thr	acc Thr	aac Asn 280	ccc Pro	ttt Phe	aac Asn	cct Pro	gtt Val 285	ccc Pro	gtg Val	tac Tyr	864
act Thr	gtg Val 290	gcc Ala	acg Thr	act Thr	ttc Phe	gac Asp 295	aag Lys	ccc Pro	act Thr	gtg Val	ccg Pro 300	gtc Val	gat Asp	ggc Gly	ttg Leu	912
agc Ser 305	cat His	ggc Gly	ccg Pro	ccc Pro	ctg Leu 310	agc Ser	gtt Val	atc Ile	agc Ser	atc Ile 315	gac Asp	cac His	ctg Leu	ccg Pro	agt Ser 320	960
ctg Leu	ctc Leu	ccg Pro	cgg Arg	gag Glu 325	gct Ala	tgc Ser	gag Glu	tgc Ser	ttc Phe 330	agc Ser	aac Asn	gac Asp	ttg Leu 335	ttg Leu	cca Pro	1008
tat Tyr	ctc Leu	ctg Leu	aag Lys 340	ctg Leu	aag Lys	gac Asp	tgg Trp	aag Lys 345	agt Ser	gac Asp	ccg Pro	gtg Val	tgg Trp 350	gct Ala	ggc Gly	1056
gcc Ala	gag Glu	aag Lys 355	ctg Leu	ttc Phe	cac His	gag Glu	aag Lys 360	gtc Val	aag Lys	acc Thr	ctg Leu	ccg Pro 365	tag			1098

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<400> 18

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Arg Ser Ala Leu Thr Pro Thr Thr Thr Lys Ala Leu Ile Glu Ala Gly  
 20 25 30

Tyr Thr Val Asn Val Glu Arg Ser Pro Val Arg Ile Phe Asp Asp Ala  
 35 40 45

eof-seql.txt

Gl u Phe Gl u Al a Val Gly Al a Thr Leu Val Pro Gl u Gly Ser Trp Gl u  
 50 55 60  
 Gl n Ile Pro Lys Asp Hi s Ile Val Ile Gly Leu Lys Gl u Leu Gl u Gl u  
 65 70 75 80  
 Lys Asp Phe Pro Leu Lys Hi s Thr Hi s Ile Gl n Phe Al a Hi s Cys Tyr  
 85 90 95  
 Lys Asn Gl n Gly Gly Trp Asp Lys Val Leu Arg Arg Tyr Hi s Asp Gly  
 100 105 110  
 Gl u Gly Met Leu Leu Asp Ile Gl u Phe Leu Gl u Lys Asp Gly Arg Arg  
 115 120 125  
 Val Al a Al a Phe Gly Tyr Trp Al a Gly Phe Al a Gly Al a Al a Leu Al a  
 130 135 140  
 Leu Gl n Asn Trp Al a Trp Gl n Leu Thr Asn Ser Gly Gl u Pro Leu Pro  
 145 150 155 160  
 Ser Val Gl u Ser Arg Pro Asn Gl u Al a Al a Leu Val Gly Asp Ile Lys  
 165 170 175  
 Gl u Al a Leu Al a Gly Gly Lys Gl u Lys Al a Gly Arg Leu Pro Arg Val  
 180 185 190  
 Ile Val Ile Gly Al a Leu Gly Arg Cys Gly Arg Gly Al a Val Asp Met  
 195 200 205  
 Cys Lys Lys Al a Gly Ile Pro Asp Gl u Asn Ile Leu Lys Trp Asp Met  
 210 215 220  
 Gl u Gl u Thr Al a Pro Gly Gly Pro Phe Lys Gl u Ile Val Gl u Ser Asp  
 225 230 235 240  
 Ile Phe Val Asn Cys Ile Tyr Leu Asn Lys Gl n Ile Pro Pro Phe Val  
 245 250 255  
 Thr Met Gl u Ser Leu Asn Thr Pro Gl u Arg Lys Leu Ser Val Ile Asn  
 260 265 270  
 Asp Cys Ser Al a Asp Thr Thr Asn Pro Phe Asn Pro Val Pro Val Tyr  
 275 280 285  
 Thr Val Al a Thr Thr Phe Asp Lys Pro Thr Val Pro Val Asp Gly Leu  
 290 295 300  
 Ser Hi s Gly Pro Pro Leu Ser Val Ile Ser Ile Asp Hi s Leu Pro Ser  
 305 310 315 320

eof-seql.txt

Leu Leu Pro Arg Glu Ala Ser Glu Ser Phe Ser Asn Asp Leu Leu Pro  
325 330 335

Tyr Leu Leu Lys Leu Lys Asp Trp Lys Ser Asp Pro Val Trp Ala Gly  
340 345 350

Ala Glu Lys Leu Phe His Glu Lys Val Lys Thr Leu Pro  
355 360 365

<210> 19  
<211> 453  
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<213> Monilophthora pernicios

<220>  
<221> CDS  
<222> (1).. (453)  
<223> Lys1

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1 5 10 15  
cgg gaa gac cct act cga att tgg gaa cga cgg gct cct cta acc ccc 96  
Arg Glu Asp Pro Thr Arg Ile Trp Glu Arg Arg Ala Pro Leu Thr Pro  
20 25 30  
gac tcc gtt tac gag ctt gtc aag gac aag gcg gta caa gtt cat gtc 144  
Asp Ser Val Tyr Glu Leu Val Lys Asp Lys Ala Val Glu Val His Val  
35 40 45  
gaa ggc tgc gac agg agg ata ttc aag gac gag gaa tac ata aag gcc 192  
Glu Gly Cys Asp Arg Arg Ile Phe Lys Asp Glu Glu Tyr Ile Lys Ala  
50 55 60  
ggt gca aca atc cga ccc aac ctg aac gat gca cat gtt gta atg gcc 240  
Gly Ala Thr Ile Arg Pro Asn Leu Asn Asp Ala His Val Val Met Gly  
65 70 75 80  
att aaa gag ccc cca ttg gac aga tta ttg ctt gat cct ctt cct cta 288  
Ile Lys Glu Pro Pro Leu Asp Arg Leu Leu Leu Asp Pro Leu Pro Leu  
85 90 95  
tcc aac acc acg tcg aaa cat gag cga aca tac atg aag ttt tct cat 336  
Ser Asn Thr Thr Ser Lys His Glu Arg Thr Tyr Met Lys Phe Ser His  
100 105 110  
act tgg aaa ggt caa gcc tac aat atg cca tta ctt tct gca ttc tta 384  
Thr Trp Lys Gly Glu Ala Tyr Asn Met Pro Leu Leu Ser Ala Phe Leu  
115 120 125  
aac att cac cca ttc cat ggc gct tat aat gac cca ctg gcc cac act 432  
Asn Ile His Pro Phe His Gly Ala Tyr Asn Asp Pro Leu Ala His Thr  
130 135 140  
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Asn Trp Ile Met Asn Ser  
145 150

<210> 20  
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eof-seql.txt

<212> PRT  
 <213> Moniliophthora perniciosa  
 <400> 20

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Met Lys Phe Thr Leu Pro Arg Leu Arg Pro Leu Thr Ile Gly Ile Arg
1      5      10      15

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      20      25      30

Asp Ser Val Tyr Glu Leu Val Lys Asp Lys Ala Val Gln Val His Val
      35      40      45

Glu Gly Cys Asp Arg Arg Ile Phe Lys Asp Glu Glu Tyr Ile Lys Ala
      50      55      60

Gly Ala Thr Ile Arg Pro Asn Leu Asn Asp Ala His Val Val Met Gly
65      70      75      80

Ile Lys Glu Pro Pro Leu Asp Arg Leu Leu Leu Asp Pro Leu Pro Leu
      85      90      95

Ser Asn Thr Thr Ser Lys His Glu Arg Thr Tyr Met Lys Phe Ser His
      100      105      110

Thr Trp Lys Gly Gln Ala Tyr Asn Met Pro Leu Leu Ser Ala Phe Leu
      115      120      125

Asn Ile His Pro Phe His Gly Ala Tyr Asn Asp Pro Leu Ala His Thr
      130      135      140

Asn Trp Ile Met Asn Ser
145      150
  
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<210> 21  
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 <212> DNA  
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<220>  
 <221> CDS  
 <222> (1)..(1116)  
 <223> Lys1

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Met Ser Gly Asp Gln Pro Leu Leu Trp Leu Arg Cys Glu Thr Lys Pro
1      5      10      15

ttc gag cat cgc tca gca tta act ccc ttg acc gcc aaa aaa ctc atc      96
Phe Glu His Arg Ser Ala Leu Thr Pro Leu Thr Ala Lys Lys Leu Ile
      20      25      30

gat gct gga ttc aag ctg gta gta gaa cgg gat cct caa cgg ttc ttc      144
Asp Ala Gly Phe Lys Leu Val Val Glu Arg Asp Pro Gln Arg Phe Phe
      35      40      45
  
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eof-seq1.txt

gct Ala	gat Asp 50	gac Asp	gaa Glu	ttt Phe	gca Ala	aaa Lys 55	gtc Val	gga Gly	tgt Cys	gaa Glu 60	ctg Val	gtc Glu	gaa His	cac Asn	192	
tcg Ser 65	tgg Trp	ccc Pro	aag Lys	gct Ala 70	ccc Pro	tcc Ser	aat Asn	gca Ala	att Ile 75	atc Ile	atc Ile	gga Gly	ttg Leu	aag Lys	gag Glu 80	240
ttg Leu	cct Pro	ccg Pro	aat Asn	gat Asp 85	gat Asp	tcg Ser	ccc Pro	ctc Leu	atc Ile 90	cat His	acc Thr	cat His	gtc Val	atg Met 95	ttc Phe	288
ggc Gly	cat His	tgt Cys	tac Tyr 100	aag Lys	caa Gln	caa Gln	gca Ala	ggc Gly 105	tac Tyr	cag Gln	gac Asp	atc Ile	cta Leu 110	agt Ser	cga Arg	336
ttc Phe	aaa Lys	cga Arg 115	ggc Gly	ggg Gly	gga Gly	act Thr	ttg Leu 120	ctc Leu	gac Asp	atg Met	gaa Glu	ttc Phe 125	ttg Leu	cag Gln	gac Asp	384
gag Glu	cac His 130	acc Thr	aaa Lys	cgc Arg	aga Arg	gtg Val 135	gct Ala	gcc Ala	ttt Phe	ggc Gly	ttc Phe 140	cac His	gcc Ala	gga Gly	ttc Phe	432
aat Asn 145	ggc Gly	tct Ser	gct Ala	gtc Val	ggt Gly 150	cta Leu	ctt Leu	gcg Ala	ctt Leu	ggg Gly 155	tcg Ser	atg Met	tta Leu	tcc Ser	ggg Gly 160	480
gag Glu	gga Gly	agt Ser	ttg Leu	aaa Lys 165	gga Gly	ttg Leu	aag Lys	cct Pro	ttc Phe 170	aaa Lys	gat Asp	gaa Glu	gac Asp	gaa Glu 175	ttg Leu	528
att Ile	act Thr	cga Arg	ggg Gly 180	aag Lys	aaa Lys	gaa Glu	ttc Phe	gat Asp 185	cga Arg	gtg Val	gtc Val	gcc Ala	aag Lys 190	cta Leu	ggt Gly	576
cga Arg	cat His	cct Pro 195	aag Lys	gcc Ala	cta Leu	gtg Val	att Ile 200	ggt Gly	tca Ser	ttg Leu	gga Gly	cgc Arg 205	tgt Cys	ggc Gly	tcc Ser	624
gga Gly 210	gct Ala	gtt Val	aca Thr	ttt Phe	ttc Phe	aag Lys 215	aaa Lys	att Ile	gga Gly	ttg Leu	aac Asn 220	aag Lys	gat Asp	gac Asp	gtc Val	672
gtt Val 225	gaa Glu	tgg Trp	gac Asp	atg Met	gca Ala 230	gag Glu	act Thr	gcc Ala	aaa Lys	ggt Gly 235	ggc Gly	cca Pro	ttc Phe	caa Gln	gaa Glu 240	720
att Ile	ctt Leu	gaa Glu	gcc Ala	gat Asp 245	atc Ile	ttc Phe	atc Ile	aat Asn	tgc Cys 250	atc Ile	tac Tyr	ttg Leu	agt Ser	agc Ser 255	aaa Lys	768
atc Ile	ccc Pro	agc Ser	ttt Phe 260	gtc Val	acc Thr	cgg Arg	gag Glu	aca Thr 265	ata Ile	gcc Ala	gca Ala	gca Ala	gga Gly 270	gat Asp	tct Ser	816
cgc Arg	cag Gln 275	ctg Leu	agg Arg	gtt Val	gta Val	gtc Val	gat Asp 280	gtg Val	tcc Ser	tgt Cys	gat Asp	acc Thr 285	acc Thr	aat Asn	ccg Pro	864
aac Asn 290	aat Asn	cct Pro	att Ile	ccg Pro	ata Ile	tat Tyr 295	gac Asp	gtc Val	aac Asn	act Thr	aca Thr 300	ttc Phe	gat Asp	tct Ser	cca Pro	912
acc Thr 305	gtg Val	cct Pro	gtc Val	cag Gln	cta Leu 310	gat Asp	gct Ala	gga Gly	ctg Leu	cca Pro 315	agc Ser	tta Leu	gaa Glu	gtt Val	tgc Cys 320	960

eof-seq1.txt

tcg	ata	gat	cat	ctg	cca	aca	ctt	ctg	ccc	aga	gaa	gcc	tcc	gaa	caa		1008
Ser	Ile	Asp	His	Leu	Pro	Thr	Leu	Leu	Pro	Arg	Glu	Ala	Ser	Glu	Gln		
				325					330					335			

ttt	tcg	aac	gat	ctc	ctt	cct	acg	ctt	ttg	caa	tta	aaa	acg	ttg	gat		1056
Phe	Ser	Asn	Asp	Leu	Leu	Pro	Thr	Leu	Leu	Gln	Leu	Lys	Thr	Leu	Asp		
			340					345					350				

caa	tct	aaa	gtt	tgg	act	gaa	gca	agg	gat	ttg	ttt	cac	aag	atg	gtc		1104
Gln	Ser	Lys	Val	Trp	Thr	Glu	Ala	Arg	Asp	Leu	Phe	His	Lys	Met	Val		
		355					360					365					

aat	tcc	atc	tag														1116
Asn	Ser	Ile															
		370															

<210> 22  
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 <212> PRT  
 <213> Puccini a graminis

<400> 22

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Phe	Glu	His	Arg	Ser	Ala	Leu	Thr	Pro	Leu	Thr	Ala	Lys	Lys	Leu	Ile	
			20					25					30			

Asp	Ala	Gly	Phe	Lys	Leu	Val	Val	Glu	Arg	Asp	Pro	Gln	Arg	Phe	Phe	
		35					40					45				

Ala	Asp	Asp	Glu	Phe	Ala	Lys	Val	Gly	Cys	Glu	Leu	Val	Glu	His	Asn	
	50					55					60					

Ser	Trp	Pro	Lys	Ala	Pro	Ser	Asn	Ala	Ile	Ile	Ile	Gly	Leu	Lys	Glu	
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Leu	Pro	Pro	Asn	Asp	Asp	Ser	Pro	Leu	Ile	His	Thr	His	Val	Met	Phe	
			85					90						95		

Gly	His	Cys	Tyr	Lys	Gln	Gln	Ala	Gly	Tyr	Gln	Asp	Ile	Leu	Ser	Arg	
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Phe	Lys	Arg	Gly	Gly	Gly	Thr	Leu	Leu	Asp	Met	Glu	Phe	Leu	Gln	Asp	
		115					120					125				

Glu	His	Thr	Lys	Arg	Arg	Val	Ala	Ala	Phe	Gly	Phe	His	Ala	Gly	Phe	
	130					135					140					

Asn	Gly	Ser	Ala	Val	Gly	Leu	Leu	Ala	Leu	Gly	Ser	Met	Leu	Ser	Gly	
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Glu	Gly	Ser	Leu	Lys	Gly	Leu	Lys	Pro	Phe	Lys	Asp	Glu	Asp	Glu	Leu	
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eof-seq1.txt

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Arg His Pro Lys Ala Leu Val Ile Gly Ser Leu Gly Arg Cys Gly Ser  
195 200 205

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210 215 220

Val Glu Trp Asp Met Ala Glu Thr Ala Lys Gly Gly Pro Phe Gln Glu  
225 230 235 240

Ile Leu Glu Ala Asp Ile Phe Ile Asn Cys Ile Tyr Leu Ser Ser Lys  
245 250 255

Ile Pro Ser Phe Val Thr Arg Glu Thr Ile Ala Ala Ala Gly Asp Ser  
260 265 270

Arg Gln Leu Arg Val Val Val Asp Val Ser Cys Asp Thr Thr Asn Pro  
275 280 285

Asn Asn Pro Ile Pro Ile Tyr Asp Val Asn Thr Thr Phe Asp Ser Pro  
290 295 300

Thr Val Pro Val Gln Leu Asp Ala Gly Leu Pro Ser Leu Glu Val Cys  
305 310 315 320

Ser Ile Asp His Leu Pro Thr Leu Leu Pro Arg Glu Ala Ser Glu Gln  
325 330 335

Phe Ser Asn Asp Leu Leu Pro Thr Leu Leu Gln Leu Lys Thr Leu Asp  
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Asn Ser Ile  
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eof-seq1.txt

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cgt Arg	ggc Gly	gtg Val 35	cag Gln	gtg Val	ctg Leu	gtt Val	cag Gln 40	ccg Pro	tcc Ser	acg Thr	gca Ala	cgc Arg 45	gtc Val	ttc Phe	tca Ser	144
gac Asp	gac Asp 50	cag Gln	tat Tyr	gtt Val	cgc Arg	gcc Ala 55	gga Gly	gcc Ala	aca Thr	ctg Leu	tcc Ser 60	gag Glu	aat Asn	ttg Leu	gcg Ala	192
ccg Pro 65	gcg Ala	aat Asn	gtc Val	att Ile	gtg Val 70	ggg Gly	gtc Val	aag Lys	cag Gln	gtg Val 75	ccc Pro	gag Glu	ccg Pro	gca Ala	ctg Leu 80	240
ctg Leu	gcg Ala	gac Asp	aag Lys	acg Thr 85	tac Tyr	ctc Leu	ttc Phe	ttc Phe	agt Ser 90	cat His	acc Thr	atc Ile	aaa Lys	gcg Ala 95	cag Gln	288
ccc Pro	gag Glu	aac Asn	atg Met 100	gcg Ala	ctg Leu	ctg Leu	gac Asp	gcc Ala 105	gtg Val	cta Leu	cag Gln	cgt Arg	cgc Arg 110	gtc Val	acg Thr	336
cta Leu	atc Ile	gac Asp 115	tac Tyr	gaa Glu	tgt Cys	atc Ile	aca Thr 120	gag Glu	gag Glu	agt Ser	ggt Gly	aag Lys 125	cgt Arg	ctc Leu	att Ile	384
gcc Ala	ttc Phe 130	ggc Gly	gga Gly	aat Asn	gcc Ala	ggt Gly 135	cgt Arg	gca Ala	gga Gly	atg Met	att Ile 140	gcg Ala	ggc Gly	ttc Phe	cga Arg	432
ggt Gly 145	ctc Leu	gga Gly	gaa Glu	cgt Arg	ctc Leu 150	atc Ile	aac Asn	atg Met	gga Gly	atc Ile 155	tcc Ser	tgc Ser	ccc Pro	ttc Phe	gtc Val 160	480
aac Asn	atc Ile	gcg Ala	tcc Ser	tcc Ser 165	tac Tyr	atg Met	tac Tyr	gca Ala	gac Asp 170	tta Leu	gag Glu	cac His	gct Ala	aaa Lys 175	gac Asp	528
gca Ala	gtg Val	gaa Glu	gct Ala 180	gca Ala	ggt Gly	aga Arg	agg Arg	att Ile 185	cga Arg	aca Thr	gat Asp	ggt Gly	ctg Leu 190	ccg Pro	ggt Gly	576
gac Asp	ttg Leu	gcg Ala 195	cca Pro	atg Met	aca Thr	ttc Phe	gcc Ala 200	ttt Phe	act Thr	ggc Gly	aat Asn	ggc Gly 205	aat Asn	gtg Val	tct Ser	624
aaa Lys	gga Gly 210	gct Ala	cag Gln	gag Glu	atc Ile	ttc Phe 215	aag Lys	ctc Leu	atg Met	ccg Pro	cac His 220	gag Glu	atg Met	gtg Val	cat His	672
cct Pro 225	tgc Ser	gaa Glu	cta Leu	cca Pro	aag Lys 230	ctg Leu	cca Pro	aaa Lys	aac Asn	aat Asn 235	cgt Arg	att Ile	ctc Leu	tac Tyr	ggt Gly 240	720
aca Thr	gtc Val	atc Ile	gac Asp	gat Asp 245	ccg Pro	gcc Ala	tac Tyr	ttt Phe	gtt Val 250	aag Lys	cct Pro	cag Gln	ccg Pro	ggt Gly 255	ttc Phe	768
acg Thr	ggt Gly	gag Glu	act Thr 260	acg Thr	aga Arg	gct Ala	cac His	tac Tyr 265	tac Tyr	cag Gln	aat Asn	cca Pro	cac His 270	caa Gln	tac Tyr	816
gaa Glu	gct Ala	gct Ala 275	ttc Phe	cat His	gaa Glu	aaa Lys	gtg Val 280	ttg Leu	cca Pro	tac Tyr	acc Thr	tca Ser 285	atg Met	ctc Leu	gtc Val	864

eof-seq1.txt

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cag Gln 305	att Ile	cgt Arg	gag Glu	ctt Leu	cgc Arg 310	aag Lys	tct Ser	ggg Gly	aac Asn	cac His 315	aaa Lys	ttg Leu	ctc Leu	ggg Gly	atc Ile 320	960
gct Ala	gat Asp	atc Ile	tcg Ser	tgc Cys 325	gat Asp	atc Ile	ggc Gly	gga Gly	agt Ser 330	atc Ile	gag Glu	ttt Phe	ctg Leu	gag Glu 335	cgt Arg	1008
gct Ala	act Thr	gaa Glu	att Ile 340	gag Glu	cgt Arg	ccg Pro	ttc Phe	gct Ala 345	ctg Leu	tac Tyr	gac Asp	gtt Val	gtt Val 350	gaa Glu	gat Asp	1056
aaa Lys	atg Met	cgc Arg 355	gaa Glu	gat Asp	gga Gly	gac Asp	agc Ser 360	cga Arg	ggc Gly	ctc Leu	gaa Glu	gga Gly 365	gac Asp	gac Asp	ggc Gly	1104
att Ile 370	atg Met	atg Met	atg Met	ggg Gly	gtc Val	gat Asp 375	atc Ile	tta Leu	ccg Pro	agt Ser	gaa Glu 380	ctg Leu	gca Ala	cgc Arg	gag Glu	1152
tcg Ser 385	agt Ser	cag Gln	cag Gln	ttc Phe	ggg Gly 390	gac Asp	cgc Arg	ttg Leu	gtc Val	gga Gly 395	tac Tyr	gtc Val	acg Thr	gcg Ala	ctg Leu 400	1200
tcg Ser	agt Ser	gct Ala	tct Ser	tcg Ser 405	tcc Ser	aac Asn	gtt Val	cct Pro	cta Leu 410	cat His	gaa Glu	cag Gln	aaa Lys	gag Glu 415	ctg Leu	1248
cct Pro	gct Ala	gag Glu	cta Leu 420	cgt Arg	gga Gly	gct Ala	tgt Cys	atc Ile 425	gcc Ala	agc Ser	aat Asn	ggg Gly	gtg Val 430	ctt Leu	gca Ala	1296
cct Pro	aaa Lys	tac Tyr 435	gag Glu	tat Tyr	atc Ile	cac His	cgt Arg 440	atg Met	cga Arg	gcc Ala	gag Glu	cgc Arg 445	gag Glu	cgc Arg	agc Ser	1344
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tgc Cys 465	gtt Val	cta Leu	ctg Leu	gag Glu	ggg Gly 470	cac His	ctg Leu	ttt Phe	gat Asp 475	acc Thr	gga Gly	ctt Leu	att Ile	aac Asn	cag Gln 480	1440
gta Val	ctc Leu	aac Asn	ctc Leu	att Ile 485	gaa Glu	gac Asp	cat His	gat Asp	ggg Gly 490	ggg Gly	ttc Phe	cac His	ctt Leu	cta Leu 495	gac Asp	1488
tgc Cys	gaa Glu	gta Val	cgc Arg 500	cca Pro	aat Asn	gtt Val	gga Gly	gtc Val 505	ggg Gly	gac Asp	agt Ser	ggg Gly	gac Asp 510	agc Ser	acc Thr	1536
agt Ser	tcg Ser	aac Asn 515	gcc Ala	atc Ile	gtg Val	cag Gln	att Ile 520	agt Ser	atg Met	agt Ser	gac Asp	cgt Arg 525	gaa Glu	gcg Ala	ctg Leu	1584
gat Asp	gct Ala 530	atc Ile	atc Ile	acc Thr	aag Lys	atc Ile 535	cgc Arg	tcc Ser	tta Leu	gct Ala	gag Glu 540	ctt Leu	act Thr	tct Ser	ggg Gly	1632
gcg Ala 545	aag Lys	gct Ala	acg Thr	gtc Val	aca Thr 550	gaa Glu	cta Leu	ccc Pro	gat Asp	ctg Leu 555	tgc Cys	ggc Gly	aca Thr	gac Asp	tac Tyr 560	1680

eol f-seql . txt

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gct Ala	gaa Glu	gta Val	tcg Ser 580	gtc Val	tcc Ser	agc Ser	ccg Pro	aag Lys 585	aag Lys	cgg Arg	aag Lys	gtt Val	gtg Val 590	tgt Cys	ttt Phe	1776
ggc Gly	gct Ala	gga Gly 595	ttg Leu	gtg Val	gcg Ala	tca Ser	cca Pro 600	ctt Leu	gtg Val	gag Glu	tat Tyr	cta Leu 605	tca Ser	aga Arg	gag Glu	1824
caa Gln 610	gga Gly	aac Asn	gaa Glu	gtt Val	cat His	gtg Val 615	gtg Val	tcg Ser	ggc Gly	att Ile 620	gag Glu	agg Arg	gaa Glu	gta Val	aag Lys	1872
gag Glu 625	atg Met	atg Met	cgc Arg	aaa Lys	atc Ile 630	tct Ser	cgt Arg	cga Arg	aac Asn	atc Ile 635	aag Lys	cct Pro	cac His	gta Val	gtg Val 640	1920
aac Asn	gtc Val	gct Ala	gaa Glu 645	gat Asp	gct Ala	gct Ala	gga Gly	gtc Val	gac Asp 650	aaa Lys	ctt Leu	tgc Cys	gct Ala	gaa Glu 655	gcc Ala	1968
gat Asp	tgc Cys	gtt Val	gtg Val 660	tcg Ser	cta Leu	ctg Leu	cca Pro	gcg Ala 665	acc Thr	atg Met	cac His	acg Thr 670	aca Thr	att Ile	gct Ala	2016
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tca Ser 690	cca Pro	gaa Glu	atg Met	aag Lys	gag Glu 695	ctg Leu	gac Asp	gct Ala	agg Arg	gca Ala	aag Lys 700	aag Lys	gca Ala	ggc Gly	att Ile	2112
ccg Pro 705	atc Ile	ctc Leu	tgt Cys	gag Glu 710	att Ile	ggc Gly	ctc Leu	gac Asp	cct Pro	ggc Gly 715	atg Met	gat Asp	cac His	atg Met	agt Ser 720	2160
gcc Ala	atg Met	aaa Lys	gtc Val 725	att Ile	gat Asp	gaa Glu	gtg Val	aag Lys	gcc Ala 730	ctc Leu	tcc Ser	ggg Gly	aag Lys	gtc Val 735	atg Met	2208
acc Thr	ttc Phe	tcg Ser	tct Ser 740	gtc Val	tgt Cys	ggc Gly	ggc Gly	ttg Leu 745	cca Pro	gct Ala	cct Pro	gaa Glu 750	gca Ala	gcg Ala	gac Asp	2256
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gcg Ala 770	gca Ala	ctg Leu	aat Asn	gca Ala	gcc Ala	cag Gln 775	tac Tyr	cgc Arg	aaa Lys	gac Asp	ggc Gly 780	aaa Lys	gtc Val	atc Ile	aac Asn	2352
gtg Val 785	gca Ala	ggt Gly	gaa Glu	gac Asp 790	tta Leu	ctc Leu	aat Asn	aga Arg	agc Ser	gaa Glu 795	cgc Arg	gta Val	aac Asn	ttc Phe	ttg Leu 800	2400
ccg Pro	gcg Ala	ttc Phe	aac Asn 805	atc Ile	gag Glu	cag Gln	atc Ile	ccg Pro	aac Asn 810	cgt Arg	aat Asn	tcg Ser	ctt Leu	cct Pro 815	tac Tyr	2448
ggc Gly	gat Asp	atc Ile	tat Tyr 820	ggc Gly	atc Ile	cca Pro	gag Glu	gca Ala 825	cac His	tcg Ser	ctg Leu	tat Tyr	cgt Arg 830	gga Gly	act Thr	2496

eof-seql.txt

ctg cga tac ggt gga tgc tgt caa att ctg tac cag cta cgc aag ctt Leu Arg Tyr Gly Gly Cys Cys Gln Ile Leu Tyr Gln Leu Arg Lys Leu 835 840 845	2544
ggc ctc ttt gac atg gac cca tcc aaa ccc att cca gcg act tgg ccg Gly Leu Phe Asp Met Asp Pro Ser Lys Pro Ile Pro Ala Thr Trp Pro 850 855 860	2592
gat ctt ctg acg cag ttg ggt ggt ttc caa ggt ctt cgc gaa gac gcc Asp Leu Leu Thr Gln Leu Gly Gly Phe Gln Gly Leu Arg Glu Asp Ala 865 870 875 880	2640
cat ggg ttc ctt cag tgg ctt ggt gcc ttt gat aaa tca aat cct gtg His Gly Phe Leu Gln Trp Leu Gly Ala Phe Asp Lys Ser Asn Pro Val 885 890 895	2688
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aag ttg tcg tac caa cca gga gaa cgc gac atg gcc atc atg cat cat Lys Leu Ser Tyr Gln Pro Gly Glu Arg Asp Met Ala Ile Met His His 915 920 925	2784
gag ttc ggc atc gag tac gag gat ggc aag aag gaa aag cgt acg tca Glu Phe Gly Ile Glu Tyr Glu Asp Gly Lys Lys Glu Lys Arg Thr Ser 930 935 940	2832
aca ttc gtg ggc tac ggg tcc gag aag ggc gac acg atc atg gct aaa Thr Phe Val Gly Tyr Gly Ser Glu Lys Gly Asp Thr Ile Met Ala Lys 945 950 955 960	2880
acg gtt gga tta agt gct gct atc ggc gtc cag ttg atc ttg cag gac Thr Val Gly Leu Ser Ala Ala Ile Gly Val Gln Leu Ile Leu Gln Asp 965 970 975	2928
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Arg Gly Val Gln Val Leu Val Gln Pro Ser Thr Ala Arg Val Phe Ser 35 40 45

eof-seql.txt

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 Leu Ala Asp Lys 85 Thr Tyr Leu Phe Phe Ser 90 His Thr Ile Lys Ala 95 Gln  
 Pro Glu Asn Met 100 Ala Leu Leu Asp Ala 105 Val Leu Gln Arg Arg 110 Val Thr  
 Leu Ile Asp 115 Tyr Glu Cys Ile Thr 120 Glu Glu Ser Gly Lys 125 Arg Leu Ile  
 Ala Phe 130 Gly Gly Asn Ala Gly 135 Arg Ala Gly Met Ile Ala Gly Phe Arg  
 Gly 145 Leu Gly Glu Arg Leu 150 Ile Asn Met Gly Ile 155 Ser Ser Pro Phe Val 160  
 Asn Ile Ala Ser 165 Ser Tyr Met Tyr Ala Asp 170 Leu Glu His Ala Lys 175 Asp  
 Ala Val Glu Ala 180 Ala Gly Arg Arg Ile Arg Thr Asp Gly Leu Pro Gly  
 Asp Leu Ala 195 Pro Met Thr Phe Ala 200 Phe Thr Gly Asn Gly 205 Asn Val Ser  
 Lys Gly 210 Ala Gln Glu Ile Phe 215 Lys Leu Met Pro His 220 Glu Met Val His  
 Pro Ser Glu Leu Pro Lys 230 Leu Pro Lys Asn Asn 235 Arg Ile Leu Tyr Gly 240  
 Thr Val Ile Asp Asp 245 Pro Ala Tyr Phe Val 250 Lys Pro Gln Pro Gly 255 Phe  
 Thr Gly Glu Thr 260 Thr Arg Ala His Tyr 265 Tyr Gln Asn Pro His 270 Gln Tyr  
 Glu Ala Ala 275 Phe His Glu Lys Val 280 Leu Pro Tyr Thr Ser 285 Met Leu Val  
 Asn Cys 290 Met Tyr Trp Asp Asp 295 Arg Phe Pro Arg Leu Ala Thr Arg Glu  
 Gln Ile Arg Glu Leu Arg 310 Lys Ser Gly Asn His 315 Lys Leu Leu Gly Ile 320

eof-seq1.txt

Ala Asp Ile Ser Cys Asp Ile Gly Gly Ser Ile Glu Phe Leu Glu Arg  
325 330 335

Ala Thr Glu Ile Glu Arg Pro Phe Ala Leu Tyr Asp Val Val Glu Asp  
340 345 350

Lys Met Arg Glu Asp Gly Asp Ser Arg Gly Leu Glu Gly Asp Asp Gly  
355 360 365

Ile Met Met Met Gly Val Asp Ile Leu Pro Ser Glu Leu Ala Arg Glu  
370 375 380

Ser Ser Gln Gln Phe Gly Asp Arg Leu Val Gly Tyr Val Thr Ala Leu  
385 390 395 400

Ser Ser Ala Ser Ser Ser Asn Val Pro Leu His Glu Gln Lys Glu Leu  
405 410 415

Pro Ala Glu Leu Arg Gly Ala Cys Ile Ala Ser Asn Gly Val Leu Ala  
420 425 430

Pro Lys Tyr Glu Tyr Ile His Arg Met Arg Ala Glu Arg Glu Arg Ser  
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Lys Gln Tyr Lys Phe Leu Asp Ala Gln Gln Gln Val Ala Gly Ser Thr  
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Cys Val Leu Leu Glu Gly His Leu Phe Asp Thr Gly Leu Ile Asn Gln  
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Val Leu Asn Leu Ile Glu Asp His Asp Gly Gly Phe His Leu Leu Asp  
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Cys Glu Val Arg Pro Asn Val Gly Val Gly Asp Ser Gly Asp Ser Thr  
500 505 510

Ser Ser Asn Ala Ile Val Gln Ile Ser Met Ser Asp Arg Glu Ala Leu  
515 520 525

Asp Ala Ile Ile Thr Lys Ile Arg Ser Leu Ala Glu Leu Thr Ser Gly  
530 535 540

Ala Lys Ala Thr Val Thr Glu Leu Pro Asp Leu Cys Gly Thr Asp Tyr  
545 550 555 560

Ser Lys Thr Arg Gly Val Val Arg Lys Asp Ala Ala Ala Asn Thr Met  
565 570 575

Ala Glu Val Ser Val Ser Ser Pro Lys Lys Arg Lys Val Val Cys Phe  
580 585 590

eof-seql.txt

Gly Ala Gly Leu Val Ala Ser Pro Leu Val Glu Tyr Leu Ser Arg Glu  
 595 600 605  
 Gln Gly Asn Glu Val His Val Val Ser Gly Ile Glu Arg Glu Val Lys  
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 625 630 635 640  
 Asn Val Ala Glu Asp Ala Ala Gly Val Asp Lys Leu Cys Ala Glu Ala  
 645 650 655  
 Asp Cys Val Val Ser Leu Leu Pro Ala Thr Met His Thr Thr Ile Ala  
 660 665 670  
 Gln Arg Cys Ile Gln His Ala Thr Pro Leu Val Thr Ala Ser Tyr Val  
 675 680 685  
 Ser Pro Glu Met Lys Glu Leu Asp Ala Arg Ala Lys Lys Ala Gly Ile  
 690 695 700  
 Pro Ile Leu Cys Glu Ile Gly Leu Asp Pro Gly Met Asp His Met Ser  
 705 710 715 720  
 Ala Met Lys Val Ile Asp Glu Val Lys Ala Leu Ser Gly Lys Val Met  
 725 730 735  
 Thr Phe Ser Ser Val Cys Gly Gly Leu Pro Ala Pro Glu Ala Ala Asp  
 740 745 750  
 Asn Ala Ile Gly Tyr Lys Phe Ser Trp Ser Pro Arg Gly Val Leu Thr  
 755 760 765  
 Ala Ala Leu Asn Ala Ala Gln Tyr Arg Lys Asp Gly Lys Val Ile Asn  
 770 775 780  
 Val Ala Gly Glu Asp Leu Leu Asn Arg Ser Glu Arg Val Asn Phe Leu  
 785 790 795 800  
 Pro Ala Phe Asn Ile Glu Gln Ile Pro Asn Arg Asn Ser Leu Pro Tyr  
 805 810 815  
 Gly Asp Ile Tyr Gly Ile Pro Glu Ala His Ser Leu Tyr Arg Gly Thr  
 820 825 830  
 Leu Arg Tyr Gly Gly Cys Cys Gln Ile Leu Tyr Gln Leu Arg Lys Leu  
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 Gly Leu Phe Asp Met Asp Pro Ser Lys Pro Ile Pro Ala Thr Trp Pro  
 850 855 860

eof-seq1.txt

Asp Leu Leu Thr Gln Leu Gly Gly Phe Gln Gly Leu Arg Glu Asp Ala  
865 870 875 880

His Gly Phe Leu Gln Trp Leu Gly Ala Phe Asp Lys Ser Asn Pro Val  
885 890 895

Val Lys Ala Pro Ser Ile Leu Asp Ala Phe Cys Ala Leu Leu Gln Asp  
900 905 910

Lys Leu Ser Tyr Gln Pro Gly Glu Arg Asp Met Ala Ile Met His His  
915 920 925

Glu Phe Gly Ile Glu Tyr Glu Asp Gly Lys Lys Glu Lys Arg Thr Ser  
930 935 940

Thr Phe Val Gly Tyr Gly Ser Glu Lys Gly Asp Thr Ile Met Ala Lys  
945 950 955 960

Thr Val Gly Leu Ser Ala Ala Ile Gly Val Gln Leu Ile Leu Gln Asp  
965 970 975

Ala Val Gln Gly Arg Gly Val Leu Thr Pro Thr Thr Pro Asp Ile Tyr  
980 985 990

Gly Pro Ala Leu Ala Arg Leu Glu Val Glu Gly Val Arg Phe Ile Glu  
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Leu Thr Pro Ala His Val Arg Glu Leu Val Gln Arg Gly Val Gln Val	
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ttg gtg cag ccg tct acg gcg cgc gtc ttc tca gac gag cag tac gtg	144
Leu Val Gln Pro Ser Thr Ala Arg Val Phe Ser Asp Glu Gln Tyr Val	
35 40 45	
cga gcg ggc gcc aag ctg gcc gag gac ctg gcg ccc gcc aat atc atc	192
Arg Ala Gly Ala Lys Leu Ala Glu Asp Leu Ala Pro Ala Asn Ile Ile	
50 55 60	

eof-seq1.txt

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tac Tyr	ctc Leu	ttc Phe	ttc Phe	agc Ser 85	cat His	acc Thr	atc Ile	aag Lys	gcg Ala 90	cag Gln	cct Pro	gag Glu	aac Asn	atg Met 95	gcg Ala	288
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tgc Cys	atc Ile	aca Thr 115	gag Glu	gag Glu	agc Ser	ggc Gly	aag Lys 120	cg Arg	ctc Leu	atc Ile	gcc Ala	ttc Phe 125	gga Gly	ggc Gly	aac Asn	384
gcg Ala 130	ggg Gly	cga Arg	gca Ala	gga Gly	atg Met	atc Ile 135	gcg Ala	ggc Gly	ttc Phe	cgg Arg	ggg Gly 140	ctt Leu	ggg Gly	gaa Glu	cg Arg	432
ctc Leu 145	atc Ile	aac Asn	atg Met	ggc Gly	atc Ile 150	tcc Ser	tcg Ser	ccc Pro	ttt Phe	gtg Val 155	aac Asn	gta Val	gcg Ala	tcg Ser	gcc Ala 160	480
tac Tyr	atg Met	tac Tyr	tcg Ser	gac Asp 165	ttg Leu	gag Glu	cac His	gca Ala	aaa Lys 170	gac Asp	gcc Ala	gta Val	gaa Glu	gct Ala 175	gcg Ala	528
ggc Gly	agg Arg	agg Arg	att Ile 180	cgg Arg	tcc Ser	gat Asp	gga Gly	ttg Leu 185	cca Pro	ggc Gly	gag Glu	ctg Leu	gcg Ala 190	ccc Pro	atg Met	576
gcc Ala	ttc Phe	gcc Ala 195	ttc Phe	acg Thr	ggc Gly	aat Asn	ggc Gly 200	aac Asn	gtg Val	tcc Ser	aaa Lys	ggg Gly 205	gcg Ala	cag Gln	gag Glu	624
atc Ile 210	ttc Phe	aag Lys	ctc Leu	atg Met	cca Pro	cat His 215	gaa Glu	atg Met	gtg Val	cat His	ccg Pro 220	tca Ser	gag Glu	ctg Leu	cca Pro	672
aac Asn 225	ttg Leu	ccg Pro	aag Lys	aac Asn	aac Asn 230	cac His	atc Ile	ctg Leu	tac Tyr	ggc Gly 235	acc Thr	gtg Val	gtt Val	gat Asp	gac Asp 240	720
ccg Pro	gcc Ala	ttc Phe	ttt Phe	gtg Val 245	aag Lys	ccg Pro	cag Gln	gct Ala	ggc Gly 250	aac Asn	agc Ser	ggc Gly	acg Thr	gcc Ala 255	tcg Ser	768
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gag Glu	aag Lys	gtg Val 275	ctg Leu	ccg Pro	tac Tyr	acg Thr	tcc Ser 280	atg Met	ttg Leu	gtg Val	aac Asn	tgc Cys 285	atg Met	tac Tyr	tgg Trp	864
gac Asp 290	gac Asp	cgt Arg	ttc Phe	ccg Pro	cga Arg	ttg Leu 295	gtt Val	acg Thr	cg Arg	gaa Glu	cag Gln 300	atc Ile	cgt Arg	gag Glu	ctt Leu	912
cga Arg 305	ggc Gly	tct Ser	gga Gly	aac Asn	caa Gln 310	aag Lys	ctg Leu	ctg Leu	ggc Gly	att Ile 315	gcc Ala	gat Asp	att Ile	tcg Ser	tgc Cys 320	960
gat Asp	att Ile	ggc Gly	ggc Gly	agt Ser 325	gtt Val	gaa Glu	ttt Phe	ttg Leu	gag Glu 330	cg Arg	gcg Ala	acg Thr	gaa Glu	att Ile 335	gaa Glu	1008

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Arg	Pro	Phe	Ala	Leu	Tyr	Asp	Val	Ala	Glu	Asp	Lys	Met	Arg	Glu	Asp	
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ggc	gat	agt	cga	ggg	ctc	gaa	ggg	gat	aac	ggc	atc	atg	atg	atg	ggg	1104
Gly	Asp	Ser	Arg	Gly	Leu	Glu	Gly	Asp	Asn	Gly	Ile	Met	Met	Met	Gly	
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gtc	gat	ata	ttg	cca	agc	gaa	ctt	cca	cgc	gag	tct	agt	caa	cag	ttc	1152
Val	Asp	Ile	Leu	Pro	Ser	Glu	Leu	Pro	Arg	Glu	Ser	Ser	Gln	Gln	Phe	
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Gly	Asp	Arg	Leu	Val	Ala	Tyr	Val	Ala	Ala	Leu	Ser	Asn	Ala	Ser	Ser	
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tcc	agt	gtc	cca	ctt	cat	gag	caa	aag	gag	cta	cct	gct	gag	ctc	cgt	1248
Ser	Ser	Val	Pro	Leu	His	Glu	Gln	Lys	Glu	Leu	Pro	Ala	Glu	Leu	Arg	
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ggc	gcc	tgc	atc	gca	agc	aaa	gga	gta	ctc	gcc	cca	aaa	tac	gag	tac	1296
Gly	Ala	Cys	Ile	Ala	Ser	Lys	Gly	Val	Leu	Ala	Pro	Lys	Tyr	Glu	Tyr	
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Ile	His	Arg	Met	Arg	Ala	Glu	Arg	Glu	Arg	Ser	Lys	Gln	Tyr	Lys	Phe	
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Leu	Asp	Ala	Gln	Gln	Glu	Val	Ala	Gly	Ser	Thr	Cys	Ile	Leu	Leu	Glu	
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ggt	cac	ctc	ttc	gat	aca	gga	ctt	atc	aac	caa	gtg	ctc	aac	ctc	att	1440
Gly	His	Leu	Phe	Asp	Thr	Gly	Leu	Ile	Asn	Gln	Val	Leu	Asn	Leu	Ile	
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Glu	Asp	His	Asp	Gly	Gly	Phe	His	Leu	Val	Asp	Cys	Glu	Val	Arg	Pro	
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aac	gtt	gct	gtc	agc	gac	agt	ggc	gac	tgt	acc	ata	tcg	aat	gct	atc	1536
Asn	Val	Ala	Val	Ser	Asp	Ser	Gly	Asp	Cys	Thr	Ile	Ser	Asn	Ala	Ile	
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Val	Gln	Ile	Ser	Met	Asn	Asp	Arg	Ala	Ala	Leu	Asp	Glu	Ile	Ile	Ser	
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Lys	Val	Arg	Ser	Leu	Ala	Asp	Leu	Thr	Ser	Gly	Ala	Lys	Ala	Thr	Val	
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act	gag	ctt	ccc	gac	ttg	tgc	ggt	aca	aac	tac	tcg	aag	aca	cgt	ggt	1680
Thr	Glu	Leu	Pro	Asp	Leu	Cys	Gly	Thr	Asn	Tyr	Ser	Lys	Thr	Arg	Gly	
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gct	gta	gta	cgg	aag	gac	gcg	gct	gcc	aat	acg	act	acc	gac	gtg	tcg	1728
Ala	Val	Val	Arg	Lys	Asp	Ala	Ala	Ala	Asn	Thr	Thr	Thr	Asp	Val	Ser	
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gtc	tca	agc	cca	aag	agg	cga	aag	att	gta	tgc	ttt	ggc	gcc	ggg	ttg	1776
Val	Ser	Ser	Pro	Lys	Arg	Arg	Lys	Ile	Val	Cys	Phe	Gly	Ala	Gly	Leu	
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gtg	gca	tcg	ccc	ctt	gtg	gag	tat	ctg	tcg	cgt	gaa	caa	gtc	aac	gaa	1824
Val	Ala	Ser	Pro	Leu	Val	Glu	Tyr	Leu	Ser	Arg	Glu	Gln	Val	Asn	Glu	
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gtt Val	cat His 610	gtg Val	gtg Val	tct Ser	ggc Gly	ctc Leu 615	gag Glu	ggg Gly	gag Glu	gta Val	aag Lys 620	gga Gly	att Ile	atg Met	cgc Arg	1872
aag Lys 625	atc Ile	tct Ser	cgc Arg	cga Arg	aac Asn 630	atc Ile	aag Lys	cct Pro	cat His	gtt Val 635	gta Val	aac Asn	gta Val	gct Ala	gaa Glu 640	1920
gac Asp	act Thr	gcc Ala	gga Gly	gtt Val 645	gac Asp	aag Lys	ctt Leu	tgt Cys	gca Ala 650	gaa Glu	gct Ala	gat Asp	tgt Cys	gtc Val 655	gtg Val	1968
tca Ser	ctt Leu	tig Leu	cca Pro 660	gcg Ala	acc Thr	atg Met	cac His	aca Thr 665	acg Thr	att Ile	gcc Ala	gag Glu	cgc Arg 670	tgt Cys	att Ile	2016
cag Gln	cac His	gcg Ala 675	aca Thr	cca Pro	ctt Leu	gtg Val	act Thr 680	gca Ala	agt Ser	tat Tyr	gtg Val	tct Ser 685	ccg Pro	gag Glu	atg Met	2064
aaa Lys 690	gag Glu	ctg Leu	gac Asp	tcc Ser	aag Lys	gca Ala 695	aag Lys	caa Gln	gct Ala	ggc Gly	atc Ile 700	cct Pro	atc Ile	ctc Leu	tgt Cys	2112
gaa Glu 705	att Ile	ggg Gly	ctt Leu	gac Asp	cct Pro 710	ggc Gly	atg Met	gat Asp	cac His	atg Met 715	agc Ser	gcg Ala	atg Met	aaa Lys	gtt Val 720	2160
att Ile	gat Asp	gag Glu	gtg Val	aag Lys 725	gcg Ala	cac His	tct Ser	ggg Gly	aag Lys 730	atc Ile	aag Lys	tct Ser	ttc Phe	tcg Ser 735	tct Ser	2208
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gca Ala 770	gct Ala	caa Gln	tac Tyr	cgc Arg	aaa Lys	gac Asp 775	gac Asp	aaa Lys	gtc Val	att Ile 780	aat Asn	gtg Val	gcc Ala	ggg Gly	gaa Glu	2352
gac Asp 785	ttg Leu	cta Leu	aac Asn	agc Ser	agt Ser 790	gag Glu	cgc Arg	gtg Val	aac Asn	ttc Phe 795	ctg Leu	cca Pro	gcg Ala	ttc Phe	aac Asn 800	2400
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ggc Gly	tgc Cys	tgt Cys 835	caa Gln	att Ile	ttg Leu	tac Tyr	caa Gln 840	cta Leu	cgt Arg	aag Lys	ctc Leu	ggc Gly 845	ctc Leu	ttc Phe	gac Asp	2544
atg Met 850	gac Asp	cca Pro	tcc Ser	aag Lys	ccc Pro	att Ile 855	cca Pro	gct Ala	act Thr	tgg Trp	cca Pro 860	gac Asp	ctt Leu	ctc Leu	act Thr	2592
caa Gln 865	tta Leu	ggg Gly	gga Gly	cac His	caa Gln 870	gac Asp	ctt Leu	cgt Arg	gaa Glu 875	gat Asp	gcc Ala	agc Ser	ggg Gly	ttc Phe	ctt Leu 880	2640

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Gln Trp Leu Gly Ala Phe Asp Arg Thr Thr Pro Ile Val Lys Ala Pro	
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tcc gtt ctc gac gct ttc tgt aca ttg ctg cag gac aag tta tcg tac	2736
Ser Val Leu Asp Ala Phe Cys Thr Leu Leu Gln Asp Lys Leu Ser Tyr	
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cag ccg ggt gaa cgc gac atg gcc atc atg cac cat gaa ttc ggc att	2784
Gln Pro Gly Glu Arg Asp Met Ala Ile Met His His Glu Phe Gly Ile	
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gag tac gaa gat ggc aaa agg gaa aaa cgc acg tca acg ttc gtg ggc	2832
Glu Tyr Glu Asp Gly Lys Arg Glu Lys Arg Thr Ser Thr Phe Val Gly	
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	935
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tat ggc tcc gag aag ggc gac aca atc atg gca aaa act gtc ggc ttg	2880
Tyr Gly Ser Glu Lys Gly Asp Thr Ile Met Ala Lys Thr Val Gly Leu	
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agt gca gct att ggc gtg cag ctg att ttg cag gac gcc gtc cag ggc	2928
Ser Ala Ala Ile Gly Val Gln Leu Ile Leu Gln Asp Ala Val Gln Gly	
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	970
	975
cga ggt gtg cta acc ccg aca acc cct gac atc tac ggc cca gcc ctc	2976
Arg Gly Val Leu Thr Pro Thr Thr Pro Asp Ile Tyr Gly Pro Ala Leu	
	980
	985
	990
gcc cgt ctc gaa gtc gaa ggt gtg cgc ttc atc gag aag	3015
Ala Arg Leu Glu Val Glu Gly Val Arg Phe Ile Glu Lys	
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35 40 45	
Arg Ala Gly Ala Lys Leu Ala Glu Asp Leu Ala Pro Ala Asn Ile Ile	
50 55 60	
Val Gly Val Lys Gln Val Pro Glu Pro Ala Leu Leu Ala Asp Lys Thr	
65 70 75 80	
Tyr Leu Phe Phe Ser His Thr Ile Lys Ala Gln Pro Glu Asn Met Ala	
85 90 95	
Leu Leu Asp Ala Val Leu Gln Arg Arg Val Thr Leu Ile Asp Tyr Glu	
100 105 110	

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Cys Ile Thr Glu Glu Ser Gly Lys Arg Leu Ile Ala Phe Gly Gly Asn  
 115 120 125  
 Ala Gly Arg Ala Gly Met Ile Ala Gly Phe Arg Gly Leu Gly Glu Arg  
 130 135 140  
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 145 150 155 160  
 Tyr Met Tyr Ser Asp Leu Glu His Ala Lys Asp Ala Val Glu Ala Ala  
 165 170 175  
 Gly Arg Arg Ile Arg Ser Asp Gly Leu Pro Gly Glu Leu Ala Pro Met  
 180 185 190  
 Ala Phe Ala Phe Thr Gly Asn Gly Asn Val Ser Lys Gly Ala Gln Glu  
 195 200 205  
 Ile Phe Lys Leu Met Pro His Glu Met Val His Pro Ser Glu Leu Pro  
 210 215 220  
 Asn Leu Pro Lys Asn Asn His Ile Leu Tyr Gly Thr Val Val Asp Asp  
 225 230 235 240  
 Pro Ala Phe Phe Val Lys Pro Gln Ala Gly Asn Ser Gly Thr Ala Ser  
 245 250 255  
 Arg Ala His Tyr Tyr Gln Asn Pro His Gln Tyr Asp Pro Ala Phe His  
 260 265 270  
 Glu Lys Val Leu Pro Tyr Thr Ser Met Leu Val Asn Cys Met Tyr Trp  
 275 280 285  
 Asp Asp Arg Phe Pro Arg Leu Val Thr Arg Glu Gln Ile Arg Glu Leu  
 290 295 300  
 Arg Gly Ser Gly Asn Gln Lys Leu Leu Gly Ile Ala Asp Ile Ser Cys  
 305 310 315 320  
 Asp Ile Gly Gly Ser Val Glu Phe Leu Glu Arg Ala Thr Glu Ile Glu  
 325 330 335  
 Arg Pro Phe Ala Leu Tyr Asp Val Ala Glu Asp Lys Met Arg Glu Asp  
 340 345 350  
 Gly Asp Ser Arg Gly Leu Glu Gly Asp Asn Gly Ile Met Met Met Gly  
 355 360 365  
 Val Asp Ile Leu Pro Ser Glu Leu Pro Arg Glu Ser Ser Gln Gln Phe  
 370 375 380

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Gly Asp Arg Leu Val Ala Tyr Val Ala Ala Leu Ser Asn Ala Ser Ser  
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 Ser Ser Val Pro Leu His Glu Gln Lys Glu Leu Pro Ala Glu Leu Arg  
 405 410 415  
 Gly Ala Cys Ile Ala Ser Lys Gly Val Leu Ala Pro Lys Tyr Glu Tyr  
 420 425 430  
 Ile His Arg Met Arg Ala Glu Arg Glu Arg Ser Lys Gln Tyr Lys Phe  
 435 440 445  
 Leu Asp Ala Gln Gln Glu Val Ala Gly Ser Thr Cys Ile Leu Leu Glu  
 450 455 460  
 Gly His Leu Phe Asp Thr Gly Leu Ile Asn Gln Val Leu Asn Leu Ile  
 465 470 475 480  
 Glu Asp His Asp Gly Gly Phe His Leu Val Asp Cys Glu Val Arg Pro  
 485 490 495  
 Asn Val Ala Val Ser Asp Ser Gly Asp Cys Thr Ile Ser Asn Ala Ile  
 500 505 510  
 Val Gln Ile Ser Met Asn Asp Arg Ala Ala Leu Asp Glu Ile Ile Ser  
 515 520 525  
 Lys Val Arg Ser Leu Ala Asp Leu Thr Ser Gly Ala Lys Ala Thr Val  
 530 535 540  
 Thr Glu Leu Pro Asp Leu Cys Gly Thr Asn Tyr Ser Lys Thr Arg Gly  
 545 550 555 560  
 Ala Val Val Arg Lys Asp Ala Ala Ala Asn Thr Thr Thr Asp Val Ser  
 565 570 575  
 Val Ser Ser Pro Lys Arg Arg Lys Ile Val Cys Phe Gly Ala Gly Leu  
 580 585 590  
 Val Ala Ser Pro Leu Val Glu Tyr Leu Ser Arg Glu Gln Val Asn Glu  
 595 600 605  
 Val His Val Val Ser Gly Leu Glu Gly Glu Val Lys Gly Ile Met Arg  
 610 615 620  
 Lys Ile Ser Arg Arg Asn Ile Lys Pro His Val Val Asn Val Ala Glu  
 625 630 635 640  
 Asp Thr Ala Gly Val Asp Lys Leu Cys Ala Glu Ala Asp Cys Val Val  
 645 650 655

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Ser Leu Leu Pro Ala Thr Met His Thr Thr Ile Ala Glu Arg Cys Ile  
660 665 670

Gln His Ala Thr Pro Leu Val Thr Ala Ser Tyr Val Ser Pro Glu Met  
675 680 685

Lys Glu Leu Asp Ser Lys Ala Lys Gln Ala Gly Ile Pro Ile Leu Cys  
690 695 700

Glu Ile Gly Leu Asp Pro Gly Met Asp His Met Ser Ala Met Lys Val  
705 710 715 720

Ile Asp Glu Val Lys Ala His Ser Gly Lys Ile Lys Ser Phe Ser Ser  
725 730 735

Val Cys Gly Gly Leu Pro Ala Pro Glu Ala Ala Asp Asn Ala Ile Gly  
740 745 750

Tyr Lys Phe Ser Trp Ser Pro Arg Gly Val Leu Thr Ala Ala Leu Asn  
755 760 765

Ala Ala Gln Tyr Arg Lys Asp Asp Lys Val Ile Asn Val Ala Gly Glu  
770 775 780

Asp Leu Leu Asn Ser Ser Glu Arg Val Asn Phe Leu Pro Ala Phe Asn  
785 790 795 800

Ile Glu Gln Ile Pro Asn Arg Asn Ser Leu Pro Tyr Gly Asp Ile Tyr  
805 810 815

Gly Ile Pro Glu Ala His Ser Leu Tyr Arg Gly Thr Leu Arg Tyr Gly  
820 825 830

Gly Cys Cys Gln Ile Leu Tyr Gln Leu Arg Lys Leu Gly Leu Phe Asp  
835 840 845

Met Asp Pro Ser Lys Pro Ile Pro Ala Thr Trp Pro Asp Leu Leu Thr  
850 855 860

Gln Leu Gly Gly His Gln Asp Leu Arg Glu Asp Ala Ser Gly Phe Leu  
865 870 875 880

Gln Trp Leu Gly Ala Phe Asp Arg Thr Thr Pro Ile Val Lys Ala Pro  
885 890 895

Ser Val Leu Asp Ala Phe Cys Thr Leu Leu Gln Asp Lys Leu Ser Tyr  
900 905 910

Gln Pro Gly Glu Arg Asp Met Ala Ile Met His His Glu Phe Gly Ile  
915 920 925

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Glu Tyr Glu Asp Gly Lys Arg Glu Lys Arg Thr Ser Thr Phe Val Gly  
930 935 940

Tyr Gly Ser Glu Lys Gly Asp Thr Ile Met Ala Lys Thr Val Gly Leu  
945 950 955 960

Ser Ala Ala Ile Gly Val Gln Leu Ile Leu Gln Asp Ala Val Gln Gly  
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Arg Gly Val Leu Thr Pro Thr Thr Pro Asp Ile Tyr Gly Pro Ala Leu  
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Glu Arg Arg Ala Pro Leu Thr Pro Ala His Val Arg Glu Leu Val Gln	
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Arg Gly Ile Gln Val Leu Val Gln Pro Ser Thr Ala Arg Val Phe Ser	
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Asp Glu Gln Tyr Val Arg Ala Gly Ala Thr Leu Ser Glu Asp Leu Ala	
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ccc gcc aac gtc atc gtg ggc gtc aag cag gtg ccc gag ccg gcg ctg	240
Pro Ala Asn Val Ile Val Gly Val Lys Gln Val Pro Glu Pro Ala Leu	
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Leu Ala Asp Lys Thr Tyr Leu Phe Phe Ser His Thr Ile Lys Ala Gln	
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Pro Glu Asn Met Ala Leu Leu Asp Ala Val Leu Gln Arg Arg Ile Thr	
100 105 110	
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Leu Val Asp Tyr Glu Cys Ile Thr Glu Glu Ser Gly Lys Arg Leu Ile	
115 120 125	
gcc ttc ggg ggc aac gcc ggc cgc gcc ggt atg atc gcg ggc ttc cgg	432
Ala Phe Gly Gly Asn Ala Gly Arg Ala Gly Met Ile Ala Gly Phe Arg	
130 135 140	

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aac Asn	gtc Val	gcg Ala	tcc Ser	gcc Ala 165	tac Tyr	atg Met	tac Tyr	tcg Ser	gac Asp 170	ttg Leu	gag Glu	cac His	gcc Ala	aag Lys 175	gac Asp	528
gcg Ala	gtg Val	gag Glu	gcc Ala 180	gcg Ala	ggc Gly	aag Lys	agg Arg	atc Ile 185	cgc Arg	tcg Ser	gac Asp	gga Gly	ctg Leu 190	ccg Pro	agc Ser	576
gag Glu	ctg Leu	gtg Val 195	ccc Pro	atg Met	acg Thr	ttc Phe	gca Ala 200	ttc Phe	acg Thr	ggc Gly	aac Asn	ggc Gly 205	aat Asn	gtg Val	tca Ser	624
aag Lys	ggc Gly 210	gcg Ala	cag Gln	gag Glu	atc Ile	ttc Phe 215	aag Lys	ctt Leu	atg Met	ccc Pro	cac His 220	gag Glu	atg Met	gtg Val	cat His	672
ccg Pro 225	tcg Ser	gag Glu	ctg Leu	ccc Pro	aag Lys 230	cta Leu	ccg Pro	aag Lys	aac Asn	aac Asn 235	cgt Arg	att Ile	ctg Leu	tac Tyr	ggc Gly 240	720
aca Thr	gtg Val	atc Ile	gac Asp	aac Asn 245	ccg Pro	gac Asp	tac Tyr	ttc Phe	gtc Val 250	aag Lys	ccc Pro	cag Gln	ccg Pro	gga Gly 255	ttc Phe	768
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aac Asn	tgc Cys 290	atg Met	tac Tyr	tgg Trp	gac Asp	gac Asp 295	cgc Arg	ttc Phe	ccg Pro	cgg Arg	cta Leu 300	gtc Val	acg Thr	cga Arg	gag Glu	912
cag Gln 305	atc Ile	cga Arg	gag Glu	ctt Leu	cga Arg 310	ggc Gly	tct Ser	ggg Gly	aac Asn	cat His 315	aag Lys	ctg Leu	ctg Leu	ggg Gly	atc Ile 320	960
gcg Ala	gac Asp	att Ile	tcg Ser	tgc Cys 325	gat Asp	att Ile	ggc Gly	ggc Gly	agc Ser 330	gtg Val	gag Glu	ttc Phe	ttg Leu	gag Glu 335	cgc Arg	1008
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att Ile 370	atg Met	atg Met	atg Met	ggc Gly	gtc Val	gat Asp 375	atc Ile	ctg Leu	ccg Pro	agc Ser	gag Glu 380	ttg Leu	gcg Ala	cgc Arg	gag Glu	1152
tca Ser 385	agc Ser	cag Gln	cag Gln	ttt Phe	ggt Gly 390	gac Asp	cgc Arg	ttg Leu	gtg Val	gga Gly 395	tac Tyr	gtc Val	acg Thr	gcg Ala	cta Leu 400	1200
tcg Ser	agt Ser	gtc Val	act Thr	tca Ser 405	tcg Ser	aat Asn	gta Val	cct Pro	ctg Leu 410	cac His	gag Glu	caa Gln	aag Lys	gag Glu 415	ctg Leu	1248

eof-seql.txt

ccg gct gaa ctg cgt ggc gct tgt att gcc agc aaa ggt gtt ttg gcg Pro Ala Glu Leu Arg Gly Ala Cys Ile Ala Ser Lys Gly Val Leu Ala	1296
ccc agg tac gag tac att cat cgg atg cgt gcc gaa cgt gag cgc agc Pro Arg Tyr Glu Tyr Ile His Arg Met Arg Ala Glu Arg Glu Arg Ser	1344
aag cag ttc aag ttc ctg gat gct cag cag gag gtt gca ggc agc acg Lys Gln Phe Lys Phe Leu Asp Ala Gln Gln Glu Val Ala Gly Ser Thr	1392
tgc ctg tta ctg gaa gga cac ctc ttt gat aca ggc ctc atc aac cag Cys Leu Leu Leu Glu Gly His Leu Phe Asp Thr Gly Leu Ile Asn Gln	1440
gta ctg aat ctc atc gag gac cac gat gga ggc ttc cac ctc gtg gac Val Leu Asn Leu Ile Glu Asp His Asp Gly Gly Phe His Leu Val Asp	1488
tgc gag gtc cga ccg aac att gca gcc gga gac agc ggc ttc agc act Cys Glu Val Arg Pro Asn Ile Ala Ala Gly Asp Ser Gly Phe Ser Thr	1536
gtg tcg aat gct ata gtc cag gtt agc atg agt gac cgt gct gct ttg Val Ser Asn Ala Ile Val Gln Val Ser Met Ser Asp Arg Ala Ala Leu	1584
gac gac att att gcg aag atc cgc tcc ctt gcc gat ctt acg tct ggt Asp Asp Ile Ile Ala Lys Ile Arg Ser Leu Ala Asp Leu Thr Ser Gly	1632
gcc aag gcc atc gtc acg gag ctt ccc gac ttg tgc ggc acg aat tat Ala Lys Ala Ile Val Thr Glu Leu Pro Asp Leu Cys Gly Thr Asn Tyr	1680
tcg aag act cgt ggt gcc gtg gtg cgc aag gat gct gct gct aat acg Ser Lys Thr Arg Gly Ala Val Val Arg Lys Asp Ala Ala Ala Asn Thr	1728
gcg gcg gac gtg tca gtc tcg agc ccg aag aag cga cag atc gtg tgt Ala Ala Asp Val Ser Val Ser Pro Lys Lys Arg Gln Ile Val Cys	1776
ttt ggc gct gga tta gtg gca tcc ccg ctt gtt gag tac ctg tct cgc Phe Gly Ala Gly Leu Val Ala Ser Pro Leu Val Glu Tyr Leu Ser Arg	1824
gag caa ggc aat gaa gtg cac gtg gtg tcc ggt atc gag ggt gaa gtg Glu Gln Gly Asn Glu Val His Val Val Ser Gly Ile Glu Gly Glu Val	1872
aag ggg gtg atg cgc aaa atc tct cgc cgt aac att aag cct cac gtg Lys Gly Val Met Arg Lys Ile Ser Arg Arg Asn Ile Lys Pro His Val	1920
gtg aac gta gct gag gac ggt gct ggc gtt gac aag ctc tgt gca gaa Val Asn Val Ala Glu Asp Gly Ala Gly Val Asp Lys Leu Cys Ala Glu	1968
gcc gat tgt gtt gtg tca cta tta cca gct aca atg cac aca acc atc Ala Asp Cys Val Val Ser Leu Leu Pro Ala Thr Met His Thr Thr Ile	2016
gcc cag cgt tgc atc caa cat gga acg cct ctc gtg act gca agc tac Ala Gln Arg Cys Ile Gln His Gly Thr Pro Leu Val Thr Ala Ser Tyr	2064

eof-seql.txt

gtg Val	tct Ser 690	ccg Pro	gag Glu	atg Met	aaa Lys	gag Glu 695	ttg Leu	gat Asp	gcc Ala	aaa Lys	gcc Ala 700	aag Lys	aaa Lys	gcc Ala	ggc Gly	2112
att Ile 705	cca Pro	ata Ile	ctg Leu	tgt Cys	gaa Glu 710	att Ile	ggc Gly	ctt Leu	gat Asp	cct Pro 715	ggc Gly	atg Met	gat Asp	cac His	atg Met 720	2160
agc Ser	gct Ala	atg Met	aag Lys	gtc Val 725	atc Ile	gac Asp	gaa Glu	gtg Val	aag Lys 730	gct Ala	cac His	tcc Ser	gga Gly	aag Lys 735	atc Ile	2208
ctg Leu	tca Ser	ttt Phe	tca Ser 740	tcc Ser	gtg Val	tgt Cys	ggg Gly	gga Gly 745	ctt Leu	cca Pro	gct Ala	cct Pro	gaa Glu 750	gct Ala	gct Ala	2256
gac Asp	aac Asn	gcc Ala 755	att Ile	ggg Gly	tac Tyr	aag Lys	ttc Phe 760	agc Ser	tgg Trp	agt Ser	ccg Pro	cgc Arg 765	ggc Gly	gtg Val	ctt Leu	2304
act Thr 770	gca Ala	gca Ala	ctg Leu	aac Asn	gcg Ala	gcg Ala 775	caa Gln	tac Tyr	cgc Arg	aaa Lys	gac Asp 780	ggc Gly	aag Lys	atc Ile	gtc Val	2352
aac Asn 785	gtc Val	gca Ala	ggc Gly	gaa Glu	gac Asp 790	ttg Leu	ctt Leu	aac Asn	agc Ser	agc Ser 795	gaa Glu	cct Pro	gtg Val	aac Asn	ttc Phe 800	2400
ttg Leu	cct Pro	gcg Ala	ttt Phe	aac Asn 805	atc Ile	gag Glu	cag Gln	atc Ile	ccg Pro 810	aac Asn	cgt Arg	gac Asp	tca Ser	ctg Leu 815	ccg Pro	2448
tac Tyr	ggc Gly	gag Glu	atc Ile 820	tac Tyr	gtc Val	att Ile	cct Pro	gag Glu 825	gcg Ala	cac His	tgc Ser	ctg Leu	tat Tyr 830	cgt Arg	gga Gly	2496
aca Thr	ctg Leu	cgc Arg 835	tac Tyr	gga Gly	ggg Gly	tgt Cys	tgc Cys 840	cga Arg	atc Ile	ctg Leu	tac Tyr	cag Gln 845	ctg Leu	cgc Arg	aag Lys	2544
ctt Leu 850	ggg Gly	ctc Leu	ttt Phe	gac Asp	atg Met	gac Asp 855	ccg Pro	tcc Ser	aag Lys	ccc Pro	att Ile 860	cca Pro	gct Ala	acg Thr	tgg Trp	2592
cca Pro 865	gac Asp	ctt Leu	atc Ile	tcc Ser	cag Gln 870	ctt Leu	ggg Gly	gga Gly	cac His	caa Gln 875	ggc Gly	ctt Leu	cgc Arg	gaa Glu	gac Asp 880	2640
gcc Ala	aat Asn	gga Gly	ttc Phe	ctt Leu 885	caa Gln	tgg Trp	ctc Leu	ggg Gly	gcc Ala 890	ttc Phe	gat Asp	cac His	agc Ser	act Thr 895	ccc Pro	2688
gta Val	gtg Val	cga Arg	gcc Ala 900	cca Pro	tcc Ser	atc Ile	ctc Leu	gac Asp 905	gca Ala	ttc Phe	tgc Cys	gca Ala	ctg Leu 910	ttg Leu	cag Gln	2736
gac Asp	aag Lys	tta Leu 915	tct Ser	tac Tyr	caa Gln	cct Pro	gga Gly 920	gaa Glu	cgt Arg	gat Asp	atg Met	gcc Ala 925	atc Ile	atg Met	cac His	2784
cat His 930	gaa Glu	ttc Phe	ggc Gly	atc Ile	gag Glu	tac Tyr 935	gaa Glu	gac Asp	ggc Gly	aag Lys	aag Lys 940	gaa Glu	aag Lys	cgc Arg	acg Thr	2832
tca Ser 945	acg Thr	ttc Phe	gtg Val	ggg Gly	tac Tyr 950	ggc Gly	tcc Ser	gat Asp	aag Lys	ggc Gly 955	gac Asp	acc Thr	atc Ile	atg Met	gct Ala 960	2880

eof-seql.txt

aaa acc gtt ggg ttg agt gca gcg att ggt gtg cag ttg atc ttg cag 2928  
Lys Thr Val Gly Leu Ser Ala Ala Ile Gly Val Gln Leu Ile Leu Gln 965 970 975

gac gca gtc caa ggc cga ggc gtg ctt aca ccg acg acc cct gac atc 2976  
Asp Ala Val Gln Gly Arg Gly Val Leu Thr Pro Thr Thr Pro Asp Ile 980 985 990

tac ggg cca gcc ctc gcg cga ctt gag gtt gaa ggt gtt cgc ttc atc 3024  
Tyr Gly Pro Ala Leu Ala Arg Leu Glu Val Glu Gly Val Arg Phe Ile 995 1000 1005

gaa aag acg ttc ccg cag cct taa 3048  
Glu Lys Thr Phe Pro Gln Pro 1010 1015

<210> 28  
<211> 1015  
<212> PRT  
<213> Phytophthora sojae

<400> 28

Met Thr Gly Lys Cys Val Gly Ile Val Arg Glu Val Tyr Asn Lys Trp  
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Glu Arg Arg Ala Pro Leu Thr Pro Ala His Val Arg Glu Leu Val Gln  
20 25 30

Arg Gly Ile Gln Val Leu Val Gln Pro Ser Thr Ala Arg Val Phe Ser  
35 40 45

Asp Glu Gln Tyr Val Arg Ala Gly Ala Thr Leu Ser Glu Asp Leu Ala  
50 55 60

Pro Ala Asn Val Ile Val Gly Val Lys Gln Val Pro Glu Pro Ala Leu  
65 70 75 80

Leu Ala Asp Lys Thr Tyr Leu Phe Phe Ser His Thr Ile Lys Ala Gln  
85 90 95

Pro Glu Asn Met Ala Leu Leu Asp Ala Val Leu Gln Arg Arg Ile Thr  
100 105 110

Leu Val Asp Tyr Glu Cys Ile Thr Glu Glu Ser Gly Lys Arg Leu Ile  
115 120 125

Ala Phe Gly Gly Asn Ala Gly Arg Ala Gly Met Ile Ala Gly Phe Arg  
130 135 140

Gly Leu Gly Glu Arg Leu Ile Asn Met Gly Val Ser Ser Pro Phe Val  
145 150 155 160

Asn Val Ala Ser Ala Tyr Met Tyr Ser Asp Leu Glu His Ala Lys Asp  
165 170 175

eof-seql.txt

Ala Val Glu Ala Ala Gly Lys Arg Ile Arg Ser Asp Gly Leu Pro Ser  
180 185 190

Glu Leu Val Pro Met Thr Phe Ala Phe Thr Gly Asn Gly Asn Val Ser  
195 200 205

Lys Gly Ala Gln Glu Ile Phe Lys Leu Met Pro His Glu Met Val His  
210 215 220

Pro Ser Glu Leu Pro Lys Leu Pro Lys Asn Asn Arg Ile Leu Tyr Gly  
225 230 235 240

Thr Val Ile Asp Asn Pro Asp Tyr Phe Val Lys Pro Gln Pro Gly Phe  
245 250 255

Ser Gly Thr Pro Ser Arg Ala His Tyr Tyr Gln Asn Pro His Gln Tyr  
260 265 270

Glu Pro Ala Phe His Glu Lys Val Leu Pro Tyr Thr Ser Met Leu Val  
275 280 285

Asn Cys Met Tyr Trp Asp Asp Arg Phe Pro Arg Leu Val Thr Arg Glu  
290 295 300

Gln Ile Arg Glu Leu Arg Gly Ser Gly Asn His Lys Leu Leu Gly Ile  
305 310 315 320

Ala Asp Ile Ser Cys Asp Ile Gly Gly Ser Val Glu Phe Leu Glu Arg  
325 330 335

Ala Thr Glu Ile Glu Arg Pro Phe Ala Leu Tyr Asp Val Ala Glu Asp  
340 345 350

Lys Met Arg Glu Asp Gly Asp Ser Arg Gly Leu Glu Gly Asp Asp Gly  
355 360 365

Ile Met Met Met Gly Val Asp Ile Leu Pro Ser Glu Leu Ala Arg Glu  
370 375 380

Ser Ser Gln Gln Phe Gly Asp Arg Leu Val Gly Tyr Val Thr Ala Leu  
385 390 395 400

Ser Ser Val Thr Ser Ser Asn Val Pro Leu His Glu Gln Lys Glu Leu  
405 410 415

Pro Ala Glu Leu Arg Gly Ala Cys Ile Ala Ser Lys Gly Val Leu Ala  
420 425 430

Pro Arg Tyr Glu Tyr Ile His Arg Met Arg Ala Glu Arg Glu Arg Ser  
435 440 445

eof-seql.txt

Lys Gln Phe Lys Phe Leu Asp Ala Gln Gln Glu Val Ala Gly Ser Thr  
 450 455 460  
 Cys Leu Leu Leu Glu Gly His Leu Phe Asp Thr Gly Leu Ile Asn Gln  
 465 470 475 480  
 Val Leu Asn Leu Ile Glu Asp His Asp Gly Gly Phe His Leu Val Asp  
 485 490 495  
 Cys Glu Val Arg Pro Asn Ile Ala Ala Gly Asp Ser Gly Phe Ser Thr  
 500 505 510  
 Val Ser Asn Ala Ile Val Gln Val Ser Met Ser Asp Arg Ala Ala Leu  
 515 520 525  
 Asp Asp Ile Ile Ala Lys Ile Arg Ser Leu Ala Asp Leu Thr Ser Gly  
 530 535 540  
 Ala Lys Ala Ile Val Thr Glu Leu Pro Asp Leu Cys Gly Thr Asn Tyr  
 545 550 555 560  
 Ser Lys Thr Arg Gly Ala Val Val Arg Lys Asp Ala Ala Ala Asn Thr  
 565 570 575  
 Ala Ala Asp Val Ser Val Ser Ser Pro Lys Lys Arg Gln Ile Val Cys  
 580 585 590  
 Phe Gly Ala Gly Leu Val Ala Ser Pro Leu Val Glu Tyr Leu Ser Arg  
 595 600 605  
 Glu Gln Gly Asn Glu Val His Val Val Ser Gly Ile Glu Gly Glu Val  
 610 615 620  
 Lys Gly Val Met Arg Lys Ile Ser Arg Arg Asn Ile Lys Pro His Val  
 625 630 635 640  
 Val Asn Val Ala Glu Asp Gly Ala Gly Val Asp Lys Leu Cys Ala Glu  
 645 650 655  
 Ala Asp Cys Val Val Ser Leu Leu Pro Ala Thr Met His Thr Thr Ile  
 660 665 670  
 Ala Gln Arg Cys Ile Gln His Gly Thr Pro Leu Val Thr Ala Ser Tyr  
 675 680 685  
 Val Ser Pro Glu Met Lys Glu Leu Asp Ala Lys Ala Lys Lys Ala Gly  
 690 695 700  
 Ile Pro Ile Leu Cys Glu Ile Gly Leu Asp Pro Gly Met Asp His Met  
 705 710 715 720

eof-seq1.txt

Ser Ala Met Lys Val Ile Asp Glu Val Lys Ala His Ser Gly Lys Ile  
725 730 735

Leu Ser Phe Ser Ser Val Cys Gly Gly Leu Pro Ala Pro Glu Ala Ala  
740 745 750

Asp Asn Ala Ile Gly Tyr Lys Phe Ser Trp Ser Pro Arg Gly Val Leu  
755 760 765

Thr Ala Ala Leu Asn Ala Ala Gln Tyr Arg Lys Asp Gly Lys Ile Val  
770 775 780

Asn Val Ala Gly Glu Asp Leu Leu Asn Ser Ser Glu Pro Val Asn Phe  
785 790 795 800

Leu Pro Ala Phe Asn Ile Glu Gln Ile Pro Asn Arg Asp Ser Leu Pro  
805 810 815

Tyr Gly Glu Ile Tyr Val Ile Pro Glu Ala His Ser Leu Tyr Arg Gly  
820 825 830

Thr Leu Arg Tyr Gly Gly Cys Cys Arg Ile Leu Tyr Gln Leu Arg Lys  
835 840 845

Leu Gly Leu Phe Asp Met Asp Pro Ser Lys Pro Ile Pro Ala Thr Trp  
850 855 860

Pro Asp Leu Ile Ser Gln Leu Gly Gly His Gln Gly Leu Arg Glu Asp  
865 870 875 880

Ala Asn Gly Phe Leu Gln Trp Leu Gly Ala Phe Asp His Ser Thr Pro  
885 890 895

Val Val Arg Ala Pro Ser Ile Leu Asp Ala Phe Cys Ala Leu Leu Gln  
900 905 910

Asp Lys Leu Ser Tyr Gln Pro Gly Glu Arg Asp Met Ala Ile Met His  
915 920 925

His Glu Phe Gly Ile Glu Tyr Glu Asp Gly Lys Lys Glu Lys Arg Thr  
930 935 940

Ser Thr Phe Val Gly Tyr Gly Ser Asp Lys Gly Asp Thr Ile Met Ala  
945 950 955 960

Lys Thr Val Gly Leu Ser Ala Ala Ile Gly Val Gln Leu Ile Leu Gln  
965 970 975

Asp Ala Val Gln Gly Arg Gly Val Leu Thr Pro Thr Thr Pro Asp Ile  
980 985 990

eof-seq1.txt

Tyr Gly Pro Ala Leu Ala Arg Leu Glu Val Glu Gly Val Arg Phe Ile  
995 1000 1005

Glu Lys Thr Phe Pro Gln Pro  
1010 1015

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<211> 1164  
<212> DNA  
<213> Pyrenophora tritici-repentis

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<222> (1)..(1164)  
<223> Lys1

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cac cgc tcg tgt ctc aca ccc aca acc gca aag aag ctg ctc gat gct 96  
His Arg Ser Cys Leu Thr Pro Thr Thr Ala Lys Lys Leu Leu Asp Ala  
20 25 30  
ggc tac cct gtc ctc gtc gag cgc tcg ccc aag gac ccc aac tac gcc 144  
Gly Tyr Pro Val Leu Val Glu Arg Ser Pro Lys Asp Pro Asn Tyr Ala  
35 40 45  
cgt atc ttc aaa gac gac gag ttc gag gag gta ggc gcg acg ctc atc 192  
Arg Ile Phe Lys Asp Asp Glu Phe Glu Glu Val Gly Ala Thr Leu Ile  
50 55 60  
gag gaa ggc gca tac aag acg gcc ccc aag gac cgc atc atc atc ggc 240  
Glu Glu Gly Ala Tyr Lys Thr Ala Pro Lys Asp Arg Ile Ile Ile Gly  
65 70 75 80  
cta aag gag ctg cca gag gac aag ttc ccg ctc gag cac aca ttc gta 288  
Leu Lys Glu Leu Pro Glu Asp Lys Phe Pro Leu Glu His Thr Phe Val  
85 90 95  
cac ttt gca cat tgc tac aag cag caa ggc gga tgg gaa aac gta ttg 336  
His Phe Ala His Cys Tyr Lys Gln Gln Gly Gly Trp Glu Asn Val Leu  
100 105 110  
gcg agg ttc ccc cgt ggc ggt ggc aca ttg tat gac ctc gag ttc ctg 384  
Ala Arg Phe Pro Arg Gly Gly Gly Thr Leu Tyr Asp Leu Glu Phe Leu  
115 120 125  
caa gat gag tca ggc agg cgc gtt gct gcc ttt ggc tac cac gcc ggt 432  
Gln Asp Glu Ser Gly Arg Arg Val Ala Ala Phe Gly Tyr His Ala Gly  
130 135 140  
ttc gtc ggc gct gct ctt gcc atc aag aca tgg gct tgg caa ctc acc 480  
Phe Val Gly Ala Ala Leu Ala Ile Lys Thr Trp Ala Trp Gln Leu Thr  
145 150 155 160  
cat ccc aac ggc gag ccc ctt cct ggt cta gag acc ttc act gag gga 528  
His Pro Asn Gly Glu Pro Leu Pro Gly Leu Glu Thr Phe Thr Glu Gly  
165 170 175  
cgc ggc tac tac aac aac gag agc gag ttg att acc caa ctc aag gaa 576  
Arg Gly Tyr Tyr Asn Asn Glu Ser Glu Leu Ile Thr Gln Leu Lys Glu  
180 185 190

eof-seql.txt

gat Asp	gtc Val	gca Ala 195	gcc Ala	ggc Gly	gaa Glu	aag Lys	atc Ile 200	gca Ala	gga Gly	cac His	aag Lys	ccc Pro 205	agc Ser	agt Ser	ctg Leu	624
gtg Val 210	cta Leu	ggt Gly	gct Ala	ctc Leu	ggc Gly	cgc Arg 215	tgc Cys	ggc Gly	tct Ser	ggt Gly	gcc Ala 220	gtc Val	gac Asp	ctc Leu	ctt Leu	672
gag Glu 225	aag Lys	att Ile	ggc Gly	tgc Cys	cct Pro 230	gag Glu	atc Ile	aag Lys	aag Lys	tgg Trp 235	gat Asp	ctt Leu	gcc Ala	gag Glu	acc Thr 240	720
aag Lys	gag Glu	cgc Arg	gac Asp	ggc Gly 245	cca Pro	tac Tyr	ccc Pro	gaa Glu	att Ile 250	gtc Val	gaa Glu	tct Ser	gac Asp	atc Ile 255	ttc Phe	768
gtc Val	aac Asn	tgc Cys	atc Ile 260	tac Tyr	ctt Leu	tcc Ser	aag Lys	ccc Pro 265	atc Ile	ccg Pro	cct Pro	ttt Phe	gtc Val 270	aac Asn	aag Lys	816
gag Glu	agt Ser	ctc Leu 275	aag Lys	tgc Ser	cca Pro	aac Asn	cgc Arg 280	aga Arg	ctc Leu	agc Ser	gtc Val	gtt Val 285	tgc Cys	gat Asp	gtc Val	864
agc Ser	tgt Cys 290	gac Asp	act Thr	act Thr	aac Asn	cct Pro 295	cat His	aac Asn	ccc Pro	att Ile	ccc Pro 300	atc Ile	tac Tyr	gat Asp	atc Ile	912
aac Asn 305	acc Thr	acc Thr	ttt Phe	gac Asp	aag Lys 310	ccc Pro	aca Thr	gtt Val	gag Glu	gtt Val 315	tct Ser	gtc Val	gag Glu	ggt Gly	gac Asp 320	960
ggt Gly	cct Pro	aga Arg	cta Leu	tct Ser 325	gtc Val	atc Ile	tcc Ser	att Ile	gac Asp 330	cac His	ctt Leu	ccc Pro	tct Ser	gcg Ala 335	ttg Leu	1008
cct Pro	cgt Arg	gag Glu	tct Ser 340	tca Ser	gaa Glu	gcc Ala	ttt Phe	agc Ser 345	aac Asn	gct Ala	ctg Leu	ctg Leu	cct Pro 350	agc Ser	ttg Leu	1056
atg Met	gct Ala	ctg Leu 355	aag Lys	gac Asp	cgc Arg	gcg Ala	aca Thr 360	acg Thr	cct Pro	gtg Val	tgg Trp	cag Gln 365	ggt Gly	gct Ala	gag Glu	1104
aag Lys	ctc Leu 370	ttc Phe	caa Gln	gag Glu	aag Lys	gtt Val 375	cag Gln	acg Thr	ctg Leu	ccg Pro	ggt Gly 380	ggt Gly	gtc Val	ccc Pro	gca Ala	1152
aag Lys 385	gag Glu	gta Val	tag													1164

<210> 30  
 <211> 387  
 <212> PRT  
 <213> Pyrenophora tritici-repentis

<400> 30

Met 1	Ala	Phe	Pro	Thr 5	Leu	His	Ala	Arg	Ala 10	Glu	Ala	Lys	Pro	Leu 15	Glu	
His	Arg	Ser	Cys 20	Leu	Thr	Pro	Thr 25	Thr	Ala	Lys	Lys	Leu 30	Leu	Asp	Ala	

eof-seql.txt

Gly Tyr Pro Val Leu Val Glu Arg Ser Pro Lys Asp Pro Asn Tyr Ala  
 35 40 45  
 Arg Ile Phe Lys Asp Asp Glu Phe Glu Glu Val Gly Ala Thr Leu Ile  
 50 55 60  
 Glu Glu Gly Ala Tyr Lys Thr Ala Pro Lys Asp Arg Ile Ile Ile Gly  
 65 70 75 80  
 Leu Lys Glu Leu Pro Glu Asp Lys Phe Pro Leu Glu His Thr Phe Val  
 85 90 95  
 His Phe Ala His Cys Tyr Lys Gln Gln Gly Gly Trp Glu Asn Val Leu  
 100 105 110  
 Ala Arg Phe Pro Arg Gly Gly Gly Thr Leu Tyr Asp Leu Glu Phe Leu  
 115 120 125  
 Gln Asp Glu Ser Gly Arg Arg Val Ala Ala Phe Gly Tyr His Ala Gly  
 130 135 140  
 Phe Val Gly Ala Ala Leu Ala Ile Lys Thr Trp Ala Trp Gln Leu Thr  
 145 150 155 160  
 His Pro Asn Gly Glu Pro Leu Pro Gly Leu Glu Thr Phe Thr Glu Gly  
 165 170 175  
 Arg Gly Tyr Tyr Asn Asn Glu Ser Glu Leu Ile Thr Gln Leu Lys Glu  
 180 185 190  
 Asp Val Ala Ala Gly Glu Lys Ile Ala Gly His Lys Pro Ser Ser Leu  
 195 200 205  
 Val Leu Gly Ala Leu Gly Arg Cys Gly Ser Gly Ala Val Asp Leu Leu  
 210 215 220  
 Glu Lys Ile Gly Cys Pro Glu Ile Lys Lys Trp Asp Leu Ala Glu Thr  
 225 230 235 240  
 Lys Glu Arg Asp Gly Pro Tyr Pro Glu Ile Val Glu Ser Asp Ile Phe  
 245 250 255  
 Val Asn Cys Ile Tyr Leu Ser Lys Pro Ile Pro Pro Phe Val Asn Lys  
 260 265 270  
 Glu Ser Leu Lys Ser Pro Asn Arg Arg Leu Ser Val Val Cys Asp Val  
 275 280 285  
 Ser Cys Asp Thr Thr Asn Pro His Asn Pro Ile Pro Ile Tyr Asp Ile  
 290 295 300

eof-seq1.txt

Asn Thr Thr Phe Asp Lys Pro Thr Val Glu Val Ser Val Glu Gly Asp  
305 310 315 320

Gly Pro Arg Leu Ser Val Ile Ser Ile Asp His Leu Pro Ser Ala Leu  
325 330 335

Pro Arg Glu Ser Ser Glu Ala Phe Ser Asn Ala Leu Leu Pro Ser Leu  
340 345 350

Met Ala Leu Lys Asp Arg Ala Thr Thr Pro Val Trp Gln Gly Ala Glu  
355 360 365

Lys Leu Phe Gln Glu Lys Val Gln Thr Leu Pro Gly Gly Val Pro Ala  
370 375 380

Lys Glu Val  
385

<210> 31  
<211> 1119  
<212> DNA  
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Glu His Arg Ser Ala Leu Thr Pro Thr Ala Lys Ala Leu Ile Asp	
20 25 30	
gct gga tac aca att aat gtc gag cgc agc cca gaa cgt ata ttc gac	144
Ala Gly Tyr Thr Ile Asn Val Glu Arg Ser Pro Glu Arg Ile Phe Asp	
35 40 45	
gat gag gag ttc gag aag gtt gga gct act ctc gtg cca gaa gat aca	192
Asp Glu Glu Phe Glu Lys Val Gly Ala Thr Leu Val Pro Glu Asp Thr	
50 55 60	
tgg aga caa gca cca aag gat cat att att att ggt ttg aag gaa cta	240
Trp Arg Gln Ala Pro Lys Asp His Ile Ile Ile Gly Leu Lys Glu Leu	
65 70 75 80	
cct gtc gaa gac ttt ccc ctc gag cat gtc cac gta caa ttc gca cac	288
Pro Val Glu Asp Phe Pro Leu Glu His Val His Val Gln Phe Ala His	
85 90 95	
tgt tac aaa caa caa ggt ggc tgg gac acc gtt cta tca cga ttt cct	336
Cys Tyr Lys Gln Gln Gly Gly Trp Asp Thr Val Leu Ser Arg Phe Pro	
100 105 110	
cgc ggc ggc gga act ctc tta gat ctc gaa ttt ttg aca gac gat aaa	384
Arg Gly Gly Gly Thr Leu Leu Asp Leu Glu Phe Leu Thr Asp Asp Lys	
115 120 125	

eof-seql.txt

ggc Gly	aga Arg 130	aga Arg	gtt Val	gca Ala	gct Ala	ttc Phe 135	gga Gly	tac Tyr	cac His	gct Ala	gga Gly 140	ttt Phe	gct Ala	ggt Gly	gca Ala	432
gca Ala 145	ttg Leu	gca Ala	ctc Leu	gaa Glu	act Thr 150	tgg Trp	gca Ala	tgg Trp	caa Gln	ctc Leu 155	act Thr	cat His	tca Ser	gca Ala	tca Ser 160	480
gag Glu	cca Pro	ttc Phe	cca Pro	agc Ser 165	gta Val	tca Ser	agc Ser	tat Tyr	cct Pro 170	aac Asn	gaa Glu	gat Asp	gag Glu	ttg Leu 175	att Ile	528
gcc Ala	gac Asp	gta Val	aag Lys 180	aaa Lys	gca Ala	atc Ile	gca Ala	gct Ala 185	gga Gly	caa Gln	gaa Glu	aag Lys	aca Thr 190	ggc Gly	aag Lys	576
gca Ala	cca Pro	cga Arg 195	gtt Val	ttg Leu	gtt Val	atc Ile	gga Gly 200	gca Ala	ttg Leu	ggc Gly	aga Arg	tgt Cys 205	gga Gly	agt Ser	gga Gly	624
gca Ala 210	gtt Val	gat Asp	ctc Leu	tgc Cys	ttg Leu	aga Arg 215	gcc Ala	ggt Gly	gtg Val	cca Pro	acc Thr 220	gag Glu	aat Asn	gtc Val	ttg Leu	672
aag Lys 225	tgg Trp	gat Asp	atg Met	gca Ala	gag Glu 230	act Thr	gcc Ala	aag Lys	gga Gly	gga Gly 235	cca Pro	ttt Phe	cca Pro	gag Glu	att Ile 240	720
gtc Val	gag Glu	agt Ser	gac Asp	atc Ile 245	ttc Phe	att Ile	aac Asn	tgc Cys	ata Ile 250	tac Tyr	ctc Leu	atg Met	tcc Ser	aag Lys 255	att Ile	768
ccg Pro	aac Asn	ttt Phe	gtc Val 260	gac Asp	atg Met	caa Gln	agc Ser	ctc Leu 265	gat Asp	acc Thr	cca Pro	aac Asn	cgc Arg 270	aaa Lys	ttg Leu	816
tca Ser	gtc Val	gtc Val 275	tgc Cys	gac Asp	gtc Val	agt Ser	gct Ala 280	gat Asp	acc Thr	aca Thr	aat Asn	ccc Pro 285	aac Asn	aat Asn	cca Pro	864
att Ile 290	cca Pro	atc Ile	tat Tyr	acc Thr	gtt Val	gcg Ala 295	acc Thr	aca Thr	ttt Phe	tca Ser	gag Glu 300	cca Pro	act Thr	gtc Val	cca Pro	912
gtc Val 305	gag Glu	gtc Val	aag Lys	gga Gly	gac Asp 310	ccc Pro	aga Arg	cta Leu	agc Ser	gtc Val 315	atc Ile	agc Ser	att Ile	gat Asp	cac His 320	960
tta Leu	cca Pro	agt Ser	cta Leu	ttg Leu 325	cca Pro	aga Arg	gaa Glu	gca Ala	agc Ser 330	gag Glu	gca Ala	ttc Phe	agc Ser	aag Lys 335	gac Asp	1008
ttg Leu	cta Leu	cca Pro	agt Ser 340	ttg Leu	ttg Leu	aca Thr	tta Leu	aag Lys 345	gac Asp	tgg Trp	aga Arg	agt Ser	act Thr 350	cca Pro	gtc Val	1056
tgg Trp	gcg Ala	aag Lys 355	gcc Ala	gag Glu	aag Lys	ttg Leu	ttt Phe 360	cag Gln	gag Glu	aag Lys	gtt Val	gcc Ala 365	aca Thr	ttg Leu	cct Pro	1104
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<210> 32  
<211> 372

&lt;212&gt; PRT

&lt;213&gt; Sclerotinia sclerotiorum

&lt;400&gt; 32

Met Ser Gly Thr Thr Leu His Leu Arg Ser Glu Leu Gly Lys Ala Leu  
1 5 10 15

Glu His Arg Ser Ala Leu Thr Pro Thr Thr Ala Lys Ala Leu Ile Asp  
20 25 30

Ala Gly Tyr Thr Ile Asn Val Glu Arg Ser Pro Glu Arg Ile Phe Asp  
35 40 45

Asp Glu Glu Phe Glu Lys Val Gly Ala Thr Leu Val Pro Glu Asp Thr  
50 55 60

Trp Arg Gln Ala Pro Lys Asp His Ile Ile Ile Gly Leu Lys Glu Leu  
65 70 75 80

Pro Val Glu Asp Phe Pro Leu Glu His Val His Val Gln Phe Ala His  
85 90 95

Cys Tyr Lys Gln Gln Gly Gly Trp Asp Thr Val Leu Ser Arg Phe Pro  
100 105 110

Arg Gly Gly Gly Thr Leu Leu Asp Leu Glu Phe Leu Thr Asp Asp Lys  
115 120 125

Gly Arg Arg Val Ala Ala Phe Gly Tyr His Ala Gly Phe Ala Gly Ala  
130 135 140

Ala Leu Ala Leu Glu Thr Trp Ala Trp Gln Leu Thr His Ser Ala Ser  
145 150 155 160

Glu Pro Phe Pro Ser Val Ser Ser Tyr Pro Asn Glu Asp Glu Leu Ile  
165 170 175

Ala Asp Val Lys Lys Ala Ile Ala Ala Gly Gln Glu Lys Thr Gly Lys  
180 185 190

Ala Pro Arg Val Leu Val Ile Gly Ala Leu Gly Arg Cys Gly Ser Gly  
195 200 205

Ala Val Asp Leu Cys Leu Arg Ala Gly Val Pro Thr Glu Asn Val Leu  
210 215 220

Lys Trp Asp Met Ala Glu Thr Ala Lys Gly Gly Pro Phe Pro Glu Ile  
225 230 235 240

Val Glu Ser Asp Ile Phe Ile Asn Cys Ile Tyr Leu Met Ser Lys Ile  
245 250 255

eof-seql.txt

Pro Asn Phe Val Asp Met Gln Ser Leu Asp Thr Pro Asn Arg Lys Leu  
260 265 270

Ser Val Val Cys Asp Val Ser Ala Asp Thr Thr Asn Pro Asn Asn Pro  
275 280 285

Ile Pro Ile Tyr Thr Val Ala Thr Thr Phe Ser Glu Pro Thr Val Pro  
290 295 300

Val Glu Val Lys Gly Asp Pro Arg Leu Ser Val Ile Ser Ile Asp His  
305 310 315 320

Leu Pro Ser Leu Leu Pro Arg Glu Ala Ser Glu Ala Phe Ser Lys Asp  
325 330 335

Leu Leu Pro Ser Leu Leu Thr Leu Lys Asp Trp Arg Ser Thr Pro Val  
340 345 350

Trp Ala Lys Ala Glu Lys Leu Phe Gln Glu Lys Val Ala Thr Leu Pro  
355 360 365

Lys Lys Glu Ala  
370

<210> 33  
<211> 1170  
<212> DNA  
<213> Trichoderma reesei

<220>  
<221> CDS  
<222> (1)..(1170)  
<223> Lys1

<400> 33  
gtt caa cta ttt tcc ccc tca tta tca tac cac aaa aaa caa caa caa 48  
Val Gln Leu Phe Ser Pro Ser Leu Ser Tyr His Lys Lys Gln Gln Gln  
1 5 10 15

tca att cgt gcc aaa atg cct acc gtc atc cat ctg aga gcc gat acc 96  
Ser Ile Arg Ala Lys Met Pro Thr Val Ile His Leu Arg Ala Asp Thr  
20 25 30

aag ccg ttt gag cgt cgc tcg ccg ctg tcc ccc cca acc gcc aag gcc 144  
Lys Pro Phe Glu Arg Arg Ser Pro Leu Ser Pro Pro Thr Ala Lys Ala  
35 40 45

ctc ctc gac gct ggc tac gtc gtc cgc gtt gag cga tct tca gag cgc 192  
Leu Leu Asp Ala Gly Tyr Val Val Arg Val Glu Arg Ser Ser Glu Arg  
50 55 60

atc tac aag gac gaa gag ttc gag gcc gtt ggc gcc gaa ttg gtc cct 240  
Ile Tyr Lys Asp Glu Glu Phe Glu Ala Val Gly Ala Glu Leu Val Pro  
65 70 75 80

gcc gga tca tgg atc aag gct ccc aag gag gac atc atc ctg ggc ctc 288  
Ala Gly Ser Trp Ile Lys Ala Pro Lys Glu Asp Ile Ile Leu Gly Leu  
85 90 95

eof-seq1.txt

aag Lys	gag Glu	ctg Leu	ccc Pro 100	gac Asp	gac Asp	gac Asp	att Ile	gac Asp 105	ctg Leu	cct Pro	cac His	aca Thr	tac Tyr 110	atc Ile	cac His	336
ttc Phe	cag Gln 115	cac His	atc Ile	ttc Phe	aag Lys	aag Lys	cag Gln 120	ctc Leu	ggc Gly	tgg Trp	gcg Ala	ccc Pro 125	tcc Ser	ctg Leu	aag Lys	384
cgc Arg 130	ttt Phe	gct Ala	cgc Arg	gcc Ala	ggc Gly	ggc Gly 135	acg Thr	ctc Leu	tac Tyr	gac Asp	ctg Leu 140	gaa Glu	ttc Phe	ctg Leu	act Thr	432
gag Glu 145	gag Glu	aac Asn	ggc Gly	cgc Arg	aga Arg 150	atc Ile	gcg Ala	gcc Ala	ttt Phe	ggc Gly 155	tac Tyr	ttt Phe	gct Ala	gga Gly	tat Tyr 160	480
gcc Ala	ggc Gly	gcc Ala	gcc Ala	att Ile 165	gcc Ala	ttc Phe	atc Ile	tgc Ser	tgg Trp 170	gct Ala	cac His	cag Gln	atc Ile	ctc Leu 175	aac Asn	528
ccc Pro	ggc Gly	gtc Val	ccc Pro 180	cag Gln	ccc Pro	ccg Pro	gtg Val	cct Pro 185	ctg Leu	ttc Phe	gac Asp	agc Ser	gcg Ala 190	cct Pro	gct Ala	576
ctg Leu	gtc Val	gcg Ala 195	cat His	gtc Val	aag Lys	gct Ala	gcg Ala 200	ctc Leu	gag Glu	ccc Pro	gcg Ala	att Ile 205	cgt Arg	gcc Ala	aac Asn	624
aac Asn 210	ggc Gly	caa Gln	ctt Leu	ccc Pro	cgc Arg	gtc Val 215	att Ile	gtg Val	att Ile	ggc Gly	gct Ala 220	ctg Leu	gga Gly	cga Arg	tgc Cys	672
ggc Gly 225	aag Lys	gga Gly	gcc Ala	gtg Val	gac Asp 230	ttt Phe	tgc Cys	cgc Arg	gag Glu	gtt Val 235	ggt Gly	ctt Leu	cct Pro	gag Glu	gac Asp 240	720
tcc Ser	atc Ile	ctc Leu	aag Lys	tgg Trp 245	gat Asp	atg Met	cag Gln	gag Glu	act Thr 250	gcc Ala	aag Lys	gga Gly	ggg Gly	cct Pro 255	ttc Phe	768
gag Glu	gaa Glu	atc Ile	acc Thr 260	act Thr	tcc Ser	gac Asp	atc Ile	ttc Phe 265	atc Ile	aac Asn	tgc Cys	gtc Val	tac Tyr 270	ctt Leu	ggc Gly	816
cct Pro	act Thr	ccc Pro 275	act Thr	cct Pro	ccc Pro	ttt Phe	gtc Val 280	aca Thr	ttc Phe	gaa Glu	tgc Ser	ctt Leu 285	gcg Ala	acc Thr	cca Pro	864
gag Glu 290	aga Arg	cga Arg	ctc Leu	cgc Arg	gtc Val	att Ile 295	gcc Ala	gat Asp	atc Ile	tgc Ser	tgt Cys 300	gat Asp	ccc Pro	aac Asn	agc Ser	912
gag Glu 305	aac Asn	aac Asn	ccg Pro	atc Ile	ccc Pro 310	ttg Leu	tac Tyr	tcc Ser	acc Thr	tgg Trp 315	tcc Ser	agc Ser	ttt Phe	gac Asp	aag Lys 320	960
ccc Pro	acg Thr	atc Ile	ccg Pro	aca Thr 325	tcc Ser	aag Lys	ccc Pro	gtt Val	gac Asp 330	ggc Gly	cct Pro	gag Glu	ctg Leu	aga Arg 335	atc Ile	1008
att Ile	gcc Ala	atc Ile	gac Asp 340	cac His	ctc Leu	ccc Pro	acc Thr	ttg Leu 345	att Ile	gcc Ala	cgc Arg	gag Glu	tcc Ser 350	agc Ser	gat Asp	1056
gag Glu	tac Tyr	tct Ser 355	ggc Gly	ctg Leu	ctg Leu	ctg Leu	ccg Pro 360	gcc Ala	ctg Leu	ctc Leu	act Thr	ctg Leu 365	gac Asp	aag Lys	cgg Arg	1104

eol f-seql . txt

gat act gag ggt gtc tgg acg aga gcg gag aag aca tac aag gac aga 1152  
Asp Thr Glu Gly Val Trp Thr Arg Ala Glu Lys Thr Tyr Lys Asp Arg  
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gtg gct gag ctg cct tag 1170  
Val Ala Glu Leu Pro  
385

<210> 34  
<211> 389  
<212> PRT  
<213> Trichoderma reesei  
<400> 34

Val Gln Leu Phe Ser Pro Ser Leu Ser Tyr His Lys Lys Gln Gln Gln  
1 5 10 15

Ser Ile Arg Ala Lys Met Pro Thr Val Ile His Leu Arg Ala Asp Thr  
20 25 30

Lys Pro Phe Glu Arg Arg Ser Pro Leu Ser Pro Pro Thr Ala Lys Ala  
35 40 45

Leu Leu Asp Ala Gly Tyr Val Val Arg Val Glu Arg Ser Ser Glu Arg  
50 55 60

Ile Tyr Lys Asp Glu Glu Phe Glu Ala Val Gly Ala Glu Leu Val Pro  
65 70 75 80

Ala Gly Ser Trp Ile Lys Ala Pro Lys Glu Asp Ile Ile Leu Gly Leu  
85 90 95

Lys Glu Leu Pro Asp Asp Asp Ile Asp Leu Pro His Thr Tyr Ile His  
100 105 110

Phe Gln His Ile Phe Lys Lys Gln Leu Gly Trp Ala Pro Ser Leu Lys  
115 120 125

Arg Phe Ala Arg Ala Gly Gly Thr Leu Tyr Asp Leu Glu Phe Leu Thr  
130 135 140

Glu Glu Asn Gly Arg Arg Ile Ala Ala Phe Gly Tyr Phe Ala Gly Tyr  
145 150 155 160

Ala Gly Ala Ala Ile Ala Phe Ile Ser Trp Ala His Gln Ile Leu Asn  
165 170 175

Pro Gly Val Pro Gln Pro Pro Val Pro Leu Phe Asp Ser Ala Pro Ala  
180 185 190

Leu Val Ala His Val Lys Ala Ala Leu Glu Pro Ala Ile Arg Ala Asn  
195 200 205

eof-seql.txt

Asn Gly Gln Leu Pro Arg Val Ile Val Ile Gly Ala Leu Gly Arg Cys  
210 215 220

Gly Lys Gly Ala Val Asp Phe Cys Arg Glu Val Gly Leu Pro Glu Asp  
225 230 235 240

Ser Ile Leu Lys Trp Asp Met Gln Glu Thr Ala Lys Gly Gly Pro Phe  
245 250 255

Glu Glu Ile Thr Thr Ser Asp Ile Phe Ile Asn Cys Val Tyr Leu Gly  
260 265 270

Pro Thr Pro Thr Pro Pro Phe Val Thr Phe Glu Ser Leu Ala Thr Pro  
275 280 285

Glu Arg Arg Leu Arg Val Ile Ala Asp Ile Ser Cys Asp Pro Asn Ser  
290 295 300

Glu Asn Asn Pro Ile Pro Leu Tyr Ser Thr Trp Ser Ser Phe Asp Lys  
305 310 315 320

Pro Thr Ile Pro Thr Ser Lys Pro Val Asp Gly Pro Glu Leu Arg Ile  
325 330 335

Ile Ala Ile Asp His Leu Pro Thr Leu Ile Ala Arg Glu Ser Ser Asp  
340 345 350

Glu Tyr Ser Gly Leu Leu Leu Pro Ala Leu Leu Thr Leu Asp Lys Arg  
355 360 365

Asp Thr Glu Gly Val Trp Thr Arg Ala Glu Lys Thr Tyr Lys Asp Arg  
370 375 380

Val Ala Glu Leu Pro  
385

<210> 35  
<211> 1173  
<212> DNA  
<213> Ustilago maydis

<220>  
<221> CDS  
<222> (1)..(1173)  
<223> Lys1

<400> 35  
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Met Ser Thr Ser Arg Gln Pro Leu Tyr Leu Arg Cys Glu Met Lys Pro  
1 5 10 15

gcc gag cac cgt gcc gcg ctc acc cca acc acc gcc aag gca ctc att 96  
Ala Glu His Arg Ala Ala Leu Thr Pro Thr Thr Ala Lys Ala Leu Ile  
20 25 30

eof-seql.txt

gac Asp	gct Ala	ggg Gly 35	ttc Phe	gac Asp	atc Ile	acg Thr	gtc Val 40	gag Glu	tcc Ser	gac Asp	cct Pro	caa Gln 45	cgt Arg	atc Ile	ttt Phe	144
gac Asp	gac Asp 50	aaa Lys	gag Glu	tac Tyr	acc Thr	gag Glu 55	gta Val	ggg Gly	tgc Cys	aag Lys	ctt Leu 60	gcc Ala	cct Pro	cac His	aac Asn	192
acg Thr 65	ttt Phe	cac His	tcc Ser	ctg Leu	ccc Pro 70	gcc Ala	gac Asp	atc Ile	ccc Pro	atc Ile 75	atc Ile	ggg Gly	ctc Leu	aag Lys	gag Glu 80	240
ctt Leu	gag Glu	gag Glu	cct Pro	ggg Gly 85	ccg Pro	gat Asp	ctg Leu	cct Pro	cac His 90	acc Thr	cac His	att Ile	cag Gln	ttt Phe 95	gct Ala	288
cac His	tgc Cys	tac Tyr	aag Lys 100	aag Lys	cag Gln	gcc Ala	ggg Gly	tgg Trp 105	gcg Ala	gat Asp	gtt Val	ctg Leu	ggg Gly 110	cga Arg	ttc Phe	336
aag Lys	cgc Arg	gga Gly 115	ggc Gly	gga Gly	aag Lys	ctc Leu	tac Tyr 120	gat Asp	ttg Leu	gag Glu	ttt Phe	ttg Leu 125	gag Glu	gac Asp	aag Lys	384
aac Asn	ggc Gly 130	cgt Arg	cgt Arg	gta Val	gcc Ala	gcg Ala 135	ttt Phe	ggg Gly	tgg Trp	cac His	gcc Ala 140	ggc Gly	ttt Phe	gca Ala	ggc Gly	432
gct Ala 145	gca Ala	ctt Leu	ggg Gly	ctg Leu	ttg Leu 150	gct Ala	ttg Leu	gct Ala	gaa Glu	cag Gln 155	gtg Val	caa Gln	ggg Gly	gaa Glu	gac Asp 160	480
caa Gln	agg Arg	ctg Leu	ggg Gly	gct Ala 165	caa Gln	aag Lys	gct Ala	tac Tyr	ccc Pro 170	aac Asn	gag Glu	cag Gln	gcg Ala	ctc Leu 175	atc Ile	528
gca Ala	cac His	gcc Ala	aag Lys 180	cag Gln	cag Gln	att Ile	gag Glu	ttc Phe 185	atc Ile	aag Lys	aaa Lys	tcg Ser	cgc Arg 190	tcg Ser	gac Asp	576
ggc Gly	aaa Lys	gtg Val 195	aag Lys	gcg Ala	ctc Leu	gtc Val	gtc Val 200	gga Gly	gct Ala	ttg Leu	gga Gly	cga Arg 205	tgt Cys	ggg Gly	cgt Arg	624
gga Gly	gcc Ala 210	att Ile	gac Asp	ttt Phe	ttc Phe	gaa Glu 215	aag Lys	gcc Ala	ggc Gly	gtc Val	gct Ala 220	tct Ser	gag Glu	gac Asp	att Ile	672
gtc Val 225	cgc Arg	tgg Trp	gac Asp	atc Ile	cag Gln 230	gag Glu	acc Thr	tca Ser	gcc Ala	aag Lys 235	cac His	ggg Gly	ccc Pro	tac Tyr	caa Gln 240	720
gag Glu	ctg Leu	ctc Leu	gac Asp	gta Val 245	gac Asp	atc Ile	ttt Phe	gtc Val	aac Asn 250	tgc Cys	atc Ile	tac Tyr	ctc Leu	acc Thr 255	tct Ser	768
aaa Lys	atc Ile	ccg Pro	cct Pro 260	ttc Phe	ctc Leu	gac Asp	caa Gln	cct Pro 265	acg Thr	atc Ile	caa Gln	gcc Ala	gct Ala 270	ggg Gly	ccc Pro	816
tcg Ser	cgt Arg 275	cgt Arg	ctc Leu	ggc Gly	gtc Val	gtc Val	gtc Val 280	gac Asp	gta Val	tcg Ser	tgc Cys	gac Asp 285	act Thr	acc Thr	aac Asn	864
ccc Pro	aac Asn 290	aac Asn	cct Pro	ctc Leu	ccc Pro	atc Ile 295	tac Tyr	gac Asp	atc Ile	aac Asn	acc Thr 300	acc Thr	ttt Phe	gac Asp	aaa Lys	912

eof-seq1.txt

cct acc gtc gac gtc aac acg ggc aaa ggt aac ccg agc ttg acc gtc 960  
Pro Thr Val Asp Val Asn Thr Gly Lys Gly Asn Pro Ser Leu Thr Val  
305 310 315 320

atc tcg atc gac cat ctg ccc act ctg ctg cct agg gag agc tcc gag 1008  
Ile Ser Ile Asp His Leu Pro Thr Leu Leu Pro Arg Glu Ser Ser Glu  
325 330 335

ggt ttc agc aac gac ctg ttg ccc agt ctg ttg cag ttg ccc tac gtg 1056  
Gly Phe Ser Asn Asp Leu Leu Pro Ser Leu Leu Glu Leu Pro Tyr Val  
340 345 350

ctc ggt aag gac acc acc aag ctc gac acg ttg gac gaa gga aag gga 1104  
Leu Gly Lys Asp Thr Thr Lys Leu Asp Thr Leu Asp Glu Gly Lys Glu  
355 360 365

gcc gtc tgg cag aga gcc gaa aag ctc ttc cat aag cac ttg gca gag 1152  
Ala Val Trp Glu Arg Ala Glu Lys Leu Phe His Lys His Leu Ala Glu  
370 375 380

gct gag cag cac aca gct tga 1173  
Ala Glu Glu His Thr Ala  
385 390

<210> 36  
<211> 390  
<212> PRT  
<213> Ustilago maydis  
<400> 36

Met Ser Thr Ser Arg Glu Pro Leu Tyr Leu Arg Cys Glu Met Lys Pro  
1 5 10 15

Ala Glu His Arg Ala Ala Leu Thr Pro Thr Thr Ala Lys Ala Leu Ile  
20 25 30

Asp Ala Gly Phe Asp Ile Thr Val Glu Ser Asp Pro Glu Arg Ile Phe  
35 40 45

Asp Asp Lys Glu Tyr Thr Glu Val Gly Cys Lys Leu Ala Pro His Asn  
50 55 60

Thr Phe His Ser Leu Pro Ala Asp Ile Pro Ile Ile Gly Leu Lys Glu  
65 70 75 80

Leu Glu Glu Pro Gly Pro Asp Leu Pro His Thr His Ile Glu Phe Ala  
85 90 95

His Cys Tyr Lys Lys Glu Ala Gly Trp Ala Asp Val Leu Gly Arg Phe  
100 105 110

Lys Arg Gly Gly Gly Lys Leu Tyr Asp Leu Glu Phe Leu Glu Asp Lys  
115 120 125

Asn Gly Arg Arg Val Ala Ala Phe Gly Trp His Ala Gly Phe Ala Gly  
130 135 140

eof-seq1.txt

Ala Ala Leu Gly Leu Leu Ala Leu Ala Glu Gln Val Gln Gly Glu Asp  
145 150 155 160

Gln Arg Leu Gly Ala Gln Lys Ala Tyr Pro Asn Glu Gln Ala Leu Ile  
165 170 175

Ala His Ala Lys Gln Gln Ile Glu Phe Ile Lys Lys Ser Arg Ser Asp  
180 185 190

Gly Lys Val Lys Ala Leu Val Val Gly Ala Leu Gly Arg Cys Gly Arg  
195 200 205

Gly Ala Ile Asp Phe Phe Glu Lys Ala Gly Val Ala Ser Glu Asp Ile  
210 215 220

Val Arg Trp Asp Ile Gln Glu Thr Ser Ala Lys His Gly Pro Tyr Gln  
225 230 235 240

Glu Leu Leu Asp Val Asp Ile Phe Val Asn Cys Ile Tyr Leu Thr Ser  
245 250 255

Lys Ile Pro Pro Phe Leu Asp Gln Pro Thr Ile Gln Ala Ala Gly Pro  
260 265 270

Ser Arg Arg Leu Gly Val Val Val Asp Val Ser Cys Asp Thr Thr Asn  
275 280 285

Pro Asn Asn Pro Leu Pro Ile Tyr Asp Ile Asn Thr Thr Phe Asp Lys  
290 295 300

Pro Thr Val Asp Val Asn Thr Gly Lys Gly Asn Pro Ser Leu Thr Val  
305 310 315 320

Ile Ser Ile Asp His Leu Pro Thr Leu Leu Pro Arg Glu Ser Ser Glu  
325 330 335

Gly Phe Ser Asn Asp Leu Leu Pro Ser Leu Leu Gln Leu Pro Tyr Val  
340 345 350

Leu Gly Lys Asp Thr Thr Lys Leu Asp Thr Leu Asp Glu Gly Lys Gly  
355 360 365

Ala Val Trp Gln Arg Ala Glu Lys Leu Phe His Lys His Leu Ala Glu  
370 375 380

Ala Glu Gln His Thr Ala  
385 390

<210> 37  
<211> 1137  
<212> DNA

&lt;213&gt; Verticillium albo-atrum

&lt;220&gt;

&lt;221&gt; CDS

&lt;222&gt; (1).. (1137)

&lt;223&gt; Lys1

&lt;400&gt; 37

atg	tct	ggc	atc	acc	cta	cac	ctc	cgg	tcc	gag	acc	aag	ccg	ctg	gag	48
Met	Ser	Gly	Ile	Thr	Leu	His	Leu	Arg	Ser	Glu	Thr	Lys	Pro	Leu	Glu	
1				5					10					15		

cac	cgc	agt	gct	ctg	acc	ccc	ttc	acg	gca	tcc	gag	ctg	atc	aag	gct	96
His	Arg	Ser	Ala	Leu	Thr	Pro	Phe	Thr	Ala	Ser	Glu	Leu	Ile	Lys	Ala	
			20					25					30			

ggc	tac	acc	ctc	aat	gtc	gag	cgc	agc	cct	gtt	cgc	atc	ttc	gac	gat	144
Gly	Tyr	Thr	Leu	Asn	Val	Glu	Arg	Ser	Pro	Val	Arg	Ile	Phe	Asp	Asp	
		35					40					45				

gcc	gag	ttc	gag	aag	att	ggc	gcc	acc	ctc	gtc	ccc	gag	ggg	agc	tgg	192
Ala	Glu	Phe	Glu	Lys	Ile	Gly	Ala	Thr	Leu	Val	Pro	Glu	Gly	Ser	Trp	
	50					55					60					

acc	gaa	gcc	ccc	caa	gac	cac	atc	att	gtt	ggc	ctc	aag	gag	ctt	ctt	240
Thr	Glu	Ala	Pro	Gln	Asp	His	Ile	Ile	Val	Gly	Leu	Lys	Glu	Leu	Leu	
65					70					75					80	

gag	gag	gat	ttc	cct	ctg	aag	cac	gtt	cac	gtc	cag	ttt	gcg	cac	tgc	288
Glu	Glu	Asp	Phe	Pro	Leu	Lys	His	Val	His	Val	Gln	Phe	Ala	His	Cys	
				85				90						95		

tac	aag	cag	cag	ggc	ggg	tgg	gag	aac	gtc	ctg	gcc	cgc	ttc	cct	cgt	336
Tyr	Lys	Gln	Gln	Gly	Gly	Trp	Glu	Asn	Val	Leu	Ala	Arg	Phe	Pro	Arg	
			100					105					110			

ggg	ggc	ggc	acc	ctc	tac	gac	ctc	gaa	ttc	ctc	gag	aag	gag	gtc	gcc	384
Gly	Gly	Gly	Thr	Leu	Tyr	Asp	Leu	Glu	Phe	Leu	Glu	Lys	Glu	Val	Ala	
		115					120					125				

ccc	ggc	cgc	ttt	gcc	cga	gtc	gcc	gcc	ttt	ggc	tgg	agc	gcc	ggg	ttc	432
Pro	Gly	Arg	Phe	Ala	Arg	Val	Ala	Ala	Phe	Gly	Trp	Ser	Ala	Gly	Phe	
	130					135					140					

tct	ggg	gct	gct	ctg	gct	ctc	cag	aac	tgg	gcc	tgg	cag	ctc	aac	aac	480
Ser	Gly	Ala	Ala	Leu	Ala	Leu	Gln	Asn	Trp	Ala	Trp	Gln	Leu	Asn	Asn	
145					150					155					160	

ccc	ggc	aag	ccc	ctg	cct	tgc	gtc	gag	agc	tat	ccc	aac	gag	gac	gag	528
Pro	Gly	Lys	Pro	Leu	Pro	Ser	Val	Glu	Ser	Tyr	Pro	Asn	Glu	Asp	Glu	
				165					170					175		

ctc	atc	act	gct	gtc	aag	aag	agc	att	gcc	gag	ggc	aag	gag	aag	gcc	576
Leu	Ile	Thr	Ala	Val	Lys	Lys	Ser	Ile	Ala	Glu	Gly	Lys	Glu	Lys	Ala	
			180					185					190			

ggc	aag	cta	ccc	cag	gtc	ctc	gtg	atc	ggc	gcc	ctc	ggc	cgc	tgc	ggc	624
Gly	Lys	Leu	Pro	Gln	Val	Leu	Val	Ile	Gly	Ala	Leu	Gly	Arg	Cys	Gly	
		195					200					205				

agc	ggg	gcc	gtc	gag	ctg	tgc	cgc	cgc	gtc	ggc	ctg	ccc	gag	gag	cag	672
Ser	Gly	Ala	Val	Glu	Leu	Cys	Arg	Arg	Val	Gly	Leu	Pro	Glu	Glu	Gln	
	210					215					220					

atc	ctc	aag	tgg	gat	ctc	gag	gag	acg	aag	aag	ggc	ggc	ccc	ttc	acc	720
Ile	Leu	Lys	Trp	Asp	Leu	Glu	Glu	Thr	Lys	Lys	Gly	Gly	Pro	Phe	Thr	
225					230					235					240	

eof-seq1.txt

gag att gtt gag agc gac atc ttt gtc aac tgc atc tac ctc aac agc	768
Glu Ile Val Glu Ser 245 Asp Ile Phe Val Asn 250 Cys Ile Tyr Leu Asn 255 Ser	
aag att ccc aac ttt gtc gac ttt gag tct ctc aag tcg ccc aag agg	816
Lys Ile Pro Asn 260 Phe Val Asp Phe Glu 265 Ser Leu Lys Ser Pro 270 Lys Arg	
cag ctg tcc gtc gtc tgc gac gtc tct gct gac acc acc aac ccc aac	864
Gln Leu Ser 275 Val Val Cys Asp Val 280 Ser Ala Asp Thr 285 Asn Pro Asn	
aac cct gtc ccc atc tac acc gtc gcc aca acc ttc gac aag ccc aca	912
Asn Pro 290 Val Pro Ile Tyr Thr 295 Val Ala Thr Thr Phe 300 Asp Lys Pro Thr	
gtg ccc gtc gag gga ctc gag aac ccg ccc ctg agc gtc atc agc att	960
Val Pro Val Glu Gly 310 Leu Glu Asn Pro Pro Leu 315 Ser Val Ile Ser Ile 320	
gat cac ctt ccc agc ctc ctc ccc cgt gag gcc tcc gag acg tac agc	1008
Asp His Leu Pro Ser 325 Leu Leu Pro Arg Glu 330 Ala Ser Glu Thr Tyr 335 Ser	
aag gac ctg ctg ccg tac ctg ctc acg ctg aag gac cgc aag acg gac	1056
Lys Asp Leu Leu 340 Pro Tyr Leu Leu Thr 345 Leu Lys Asp Arg Lys 350 Thr Asp	
ccc gtc tgg acg agg gct gag aag ctc ttc aac gac aag gtc gct acg	1104
Pro Val Trp 355 Thr Arg Ala Glu Lys 360 Leu Phe Asn Asp Lys 365 Val Ala Thr	
ctg ccg gcc gac ctg cag aag cct gct cag tga	1137
Leu Pro Ala Asp Leu Gln Lys 375 Pro Ala Gln	

<210> 38

<211> 378

<212> PRT

<213> Verticillium albo-atrum

<400> 38

Met Ser Gly Ile Thr Leu His Leu Arg Ser Glu Thr Lys Pro Leu Glu	
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His Arg Ser Ala Leu Thr Pro Phe Thr Ala Ser Glu Leu Ile Lys Ala	
20 25 30	
Gly Tyr Thr Leu Asn Val Glu Arg Ser Pro Val Arg Ile Phe Asp Asp	
35 40 45	
Ala Glu Phe Glu Lys Ile Gly Ala Thr Leu Val Pro Glu Gly Ser Trp	
50 55 60	
Thr Glu Ala Pro Gln Asp His Ile Ile Val Gly Leu Lys Glu Leu Leu	
65 70 75 80	
Glu Glu Asp Phe Pro Leu Lys His Val His Val Gln Phe Ala His Cys	
85 90 95	

eof-seql.txt

Tyr Lys Gln Gln Gly Gly Trp Glu Asn Val Leu Ala Arg Phe Pro Arg  
 100 105 110  
 Gly Gly Gly Thr Leu Tyr Asp Leu Glu Phe Leu Glu Lys Glu Val Ala  
 115 120 125  
 Pro Gly Arg Phe Ala Arg Val Ala Ala Phe Gly Trp Ser Ala Gly Phe  
 130 135 140  
 Ser Gly Ala Ala Leu Ala Leu Gln Asn Trp Ala Trp Gln Leu Asn Asn  
 145 150 155 160  
 Pro Gly Lys Pro Leu Pro Ser Val Glu Ser Tyr Pro Asn Glu Asp Glu  
 165 170 175  
 Leu Ile Thr Ala Val Lys Lys Ser Ile Ala Glu Gly Lys Glu Lys Ala  
 180 185 190  
 Gly Lys Leu Pro Gln Val Leu Val Ile Gly Ala Leu Gly Arg Cys Gly  
 195 200 205  
 Ser Gly Ala Val Glu Leu Cys Arg Arg Val Gly Leu Pro Glu Glu Gln  
 210 215 220  
 Ile Leu Lys Trp Asp Leu Glu Glu Thr Lys Lys Gly Gly Pro Phe Thr  
 225 230 235 240  
 Glu Ile Val Glu Ser Asp Ile Phe Val Asn Cys Ile Tyr Leu Asn Ser  
 245 250 255  
 Lys Ile Pro Asn Phe Val Asp Phe Glu Ser Leu Lys Ser Pro Lys Arg  
 260 265 270  
 Gln Leu Ser Val Val Cys Asp Val Ser Ala Asp Thr Thr Asn Pro Asn  
 275 280 285  
 Asn Pro Val Pro Ile Tyr Thr Val Ala Thr Thr Phe Asp Lys Pro Thr  
 290 295 300  
 Val Pro Val Glu Gly Leu Glu Asn Pro Pro Leu Ser Val Ile Ser Ile  
 305 310 315 320  
 Asp His Leu Pro Ser Leu Leu Pro Arg Glu Ala Ser Glu Thr Tyr Ser  
 325 330 335  
 Lys Asp Leu Leu Pro Tyr Leu Leu Thr Leu Lys Asp Arg Lys Thr Asp  
 340 345 350  
 Pro Val Trp Thr Arg Ala Glu Lys Leu Phe Asn Asp Lys Val Ala Thr  
 355 360 365

eol f-seql . txt

Leu Pro Al a Asp Leu Gl n Lys Pro Al a Gl n  
370 375

<210> 39  
<211> 1125  
<212> DNA  
<213> Mycosphaerel l a grami ni col a

<220>  
<221> CDS  
<222> (1).. (1125)  
<223> Lys1

<400> 39	atg tct tct tca cca cta act ttg cac atc cgc gcg gaa acc aag ccc	48
Met Ser Ser Ser Pro Leu Thr Leu His Ile Arg Ala Glu Thr Lys Pro	1 5 10 15	
ctc gaa cat cgc acc gct gtt ccg ccc aag gtt gcg agg aaa ctt gtc	96	
Leu Glu His Arg Thr Ala Val Pro Pro Lys Val Ala Arg Lys Leu Val	20 25 30	
gag gcg ggt tac gtg gtc aac gtc gag cgg agt cca ttg agc atc ttc	144	
Gl u Ala Gly Tyr Val Val Asn Val Gl u Arg Ser Pro Leu Ser Ile Phe	35 40 45	
ccg gac aac gag tac gaa gga aca gga gct acg ctt gtg ccg act ggt	192	
Pro Asp Asn Glu Tyr Glu Gly Thr Gly Ala Thr Leu Val Pro Thr Gly	50 55 60	
tct tgg acg gaa gca ccc aag gac cat atc gtt gtg gga ttg aag gag	240	
Ser Trp Thr Glu Ala Pro Lys Asp His Ile Val Val Gly Leu Lys Gl u	65 70 75 80	
ttg ccg gag gag gat ttc gcg ttg gtg cat acg cat gtt cag ttc gca	288	
Leu Pro Glu Glu Asp Phe Ala Leu Val His Thr His Val Gl n Phe Ala	85 90 95	
cat tgc tat aag aat cag ggc gga tgg gag aaa gtc ttg agt cga ttc	336	
His Cys Tyr Lys Asn Gl n Gly Gly Trp Glu Lys Val Leu Ser Arg Phe	100 105 110	
ccg aga ggt gga ggg acg ctg ctt gac ttg gag ttc ctc gaa gat gag	384	
Pro Arg Gly Gly Gly Thr Leu Leu Asp Leu Glu Phe Leu Gl u Asp Gl u	115 120 125	
caa gga cga cga gtt gcg gct ttc ggc tac cac gcc gga ttc gca gga	432	
Gl n Gly Arg Arg Val Ala Ala Phe Gly Tyr His Ala Gly Phe Ala Gly	130 135 140	
gca gct ctc tcc ctc atc aca tgg gcc tgg caa ctc gag cac ggc acc	480	
Ala Ala Leu Ser Leu Ile Thr Trp Ala Trp Gl n Leu Gl u His Gly Thr	145 150 155 160	
tcc aag ccc gtg cct gga gtc acc gcc tac gag aac gaa aca ctc ctc	528	
Ser Lys Pro Val Pro Gly Val Thr Ala Tyr Glu Asn Gl u Thr Leu Leu	165 170 175	
gtc aac gac gtg aag aaa gcc gtc gag aag ggc aag tgc atc gcc ggc	576	
Val Asn Asp Val Lys Lys Ala Val Gl u Lys Gly Lys Ser Ile Ala Gly	180 185 190	
cac ctt ccg cga gtt ctg gtc atc ggc gct ctc gga cgt tgt ggc cgt	624	
His Leu Pro Arg Val Leu Val Ile Gly Ala Leu Gl y Arg Cys Gly Arg	195 200 205	

eof-seql . txt

gga gcc gtc gac ctc tgc gtc aaa gct ggt ctg caa gac atc ctc aaa	672
Gly Ala Val Asp Leu Cys Val Lys Ala Gly Leu Gln Asp Ile Leu Lys	
210 215 220	
tg gac ctg caa gag acc aaa gcc aaa cca ggc ccc tac caa gaa atc	720
Trp Asp Leu Gln Glu Thr Lys Ala Lys Pro Gly Pro Tyr Gln Glu Ile	
225 230 235 240	
atc gag tcc gac gtc ttt gta aac tgc atc tac ctg tcc gcc aaa atc	768
Ile Glu Ser Asp Val Phe Val Asn Cys Ile Tyr Leu Ser Ala Lys Ile	
245 250 255	
cct cca ttc atc gac gca cct tcc ctg gcc tcc ccg acc cgc aaa ctg	816
Pro Pro Phe Ile Asp Ala Pro Ser Leu Ala Ser Pro Thr Arg Lys Leu	
260 265 270	
agc gtc gtc tgt gac gtc tcc tgc gac acc acg aat cca cac aat ccg	864
Ser Val Val Cys Asp Val Ser Cys Asp Thr Thr Asn Pro His Asn Pro	
275 280 285	
att ccg att tac tcc atc aat acg acg ttt gac aag ccc acc gtg cct	912
Ile Pro Ile Tyr Ser Ile Asn Thr Thr Phe Asp Lys Pro Thr Val Pro	
290 295 300	
gtg gaa ctg tcc tca gag gcg aac gat gtg ccg ttg agt gtg att agc	960
Val Glu Leu Ser Ser Glu Ala Asn Asp Val Pro Leu Ser Val Ile Ser	
305 310 315 320	
atc gac cac ttg ccg agt ttg ttg ccg aga gag gcg agc gag gca ttc	1008
Ile Asp His Leu Pro Ser Leu Leu Pro Arg Glu Ala Ser Glu Ala Phe	
325 330 335	
agc gag gca ttg ttg ccg agt ttg ttg gag ttg aag gag agg aag acg	1056
Ser Glu Ala Leu Leu Pro Ser Leu Leu Glu Leu Lys Glu Arg Lys Thr	
340 345 350	
gcg ccg gtg tgg agg cag gcg gag aag ttg ttc gag gac aag gtg gct	1104
Ala Arg Val Trp Arg Gln Ala Glu Lys Leu Phe Glu Asp Lys Val Ala	
355 360 365	
agt ctg ccg aag ggt tgc tac	1125
Ser Leu Pro Lys Gly Ser Tyr	
370 375	

<210> 40  
 <211> 375  
 <212> PRT  
 <213> Mycosphaerella graminicola

<400> 40

Met Ser Ser Ser Pro Leu Thr Leu His Ile Arg Ala Glu Thr Lys Pro
1 5 10 15
Leu Glu His Arg Thr Ala Val Pro Pro Lys Val Ala Arg Lys Leu Val
20 25 30
Glu Ala Gly Tyr Val Val Asn Val Glu Arg Ser Pro Leu Ser Ile Phe
35 40 45
Pro Asp Asn Glu Tyr Glu Gly Thr Gly Ala Thr Leu Val Pro Thr Gly
50 55 60

eof-seql.txt

Ser Trp Thr Glu Ala Pro Lys Asp His Ile Val Val Gly Leu Lys Glu  
 65 70 75 80  
 Leu Pro Glu Glu Asp Phe Ala Leu Val His Thr His Val Gln Phe Ala  
 85 90 95  
 His Cys Tyr Lys Asn Gln Gly Gly Trp Glu Lys Val Leu Ser Arg Phe  
 100 105 110  
 Pro Arg Gly Gly Gly Thr Leu Leu Asp Leu Glu Phe Leu Glu Asp Glu  
 115 120 125  
 Gln Gly Arg Arg Val Ala Ala Phe Gly Tyr His Ala Gly Phe Ala Gly  
 130 135 140  
 Ala Ala Leu Ser Leu Ile Thr Trp Ala Trp Gln Leu Glu His Gly Thr  
 145 150 155 160  
 Ser Lys Pro Val Pro Gly Val Thr Ala Tyr Glu Asn Glu Thr Leu Leu  
 165 170 175  
 Val Asn Asp Val Lys Lys Ala Val Glu Lys Gly Lys Ser Ile Ala Gly  
 180 185 190  
 His Leu Pro Arg Val Leu Val Ile Gly Ala Leu Gly Arg Cys Gly Arg  
 195 200 205  
 Gly Ala Val Asp Leu Cys Val Lys Ala Gly Leu Gln Asp Ile Leu Lys  
 210 215 220  
 Trp Asp Leu Gln Glu Thr Lys Ala Lys Pro Gly Pro Tyr Gln Glu Ile  
 225 230 235 240  
 Ile Glu Ser Asp Val Phe Val Asn Cys Ile Tyr Leu Ser Ala Lys Ile  
 245 250 255  
 Pro Pro Phe Ile Asp Ala Pro Ser Leu Ala Ser Pro Thr Arg Lys Leu  
 260 265 270  
 Ser Val Val Cys Asp Val Ser Cys Asp Thr Thr Asn Pro His Asn Pro  
 275 280 285  
 Ile Pro Ile Tyr Ser Ile Asn Thr Thr Phe Asp Lys Pro Thr Val Pro  
 290 295 300  
 Val Glu Leu Ser Ser Glu Ala Asn Asp Val Pro Leu Ser Val Ile Ser  
 305 310 315 320  
 Ile Asp His Leu Pro Ser Leu Leu Pro Arg Glu Ala Ser Glu Ala Phe  
 325 330 335

eof-seql.txt

Ser Glu Ala Leu Leu Pro Ser Leu Leu Glu Leu Lys Glu Arg Lys Thr  
340 345 350

Ala Arg Val Trp Arg Gln Ala Glu Lys Leu Phe Glu Asp Lys Val Ala  
355 360 365

Ser Leu Pro Lys Gly Ser Tyr  
370 375

<210> 41  
<211> 1170  
<212> DNA  
<213> Fusarium moniliforme

<220>  
<221> CDS  
<222> (1).. (1170)  
<223> Lys1

<400> 41	atg tct gac tat ccc cac att ctc ctt cgc gct gag gag aag cct ctc	48
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	gag cac cga tct ttc tcc ccc gca att atc aag aca ctc gtt gat gct	96
Glu His Arg Ser Phe Ser Pro Ala Ile Ile Lys Thr Leu Val Asp Ala	20 25 30	
	gga tac ccc att tcc gtc gag cga tcg tct acc gac ccc aag ttc aag	144
Gly Tyr Pro Ile Ser Val Glu Arg Ser Ser Thr Asp Pro Lys Phe Lys	35 40 45	
	cgt atc ttt gag gac tca gaa tat gag gct gct ggt gct cgt ctt gtc	192
Arg Ile Phe Glu Asp Ser Glu Tyr Glu Ala Ala Gly Ala Arg Leu Val	50 55 60	
	gat acg ggt gtc tgg ccc aac gct gag cct gga aca atc att ctc ggc	240
Asp Thr Gly Val Trp Pro Asn Ala Glu Pro Gly Thr Ile Ile Leu Gly	65 70 75 80	
	cta aaa gag cta cct tct gag gac ttc ccg ctc aag aat gac cac att	288
Leu Lys Glu Leu Pro Ser Glu Asp Phe Pro Leu Lys Asn Asp His Ile	85 90 95	
	aca ttt gca cat tgt tac aag aac caa ggc ggg tgg gag cag gtc ctc	336
Thr Phe Ala His Cys Tyr Lys Asn Gln Gly Gly Trp Glu Gln Val Leu	100 105 110	
	ggt cgc tgg gca cgc ggt ggc agc cgg ctt tac gac ctc gag ttt ctt	384
Gly Arg Trp Ala Arg Gly Gly Ser Arg Leu Tyr Asp Leu Glu Phe Leu	115 120 125	
	gtg gac gaa caa ggt cgg cgt gtt tct gca ttt ggg tac cac gcg ggc	432
Val Asp Glu Gln Gly Arg Arg Val Ser Ala Phe Gly Tyr His Ala Gly	130 135 140	
	ttc gcc ggt gct gcg ctc gga atc aag aca ctc gct cac cag ctg cag	480
Phe Ala Gly Ala Ala Leu Gly Ile Lys Thr Leu Ala His Gln Leu Gln	145 150 155 160	
	ggc tcg tcc tct aaa ctt cct tct gtc gag aca ttc act gat ggc cgc	528
Gly Ser Ser Ser Lys Leu Pro Ser Val Glu Thr Phe Thr Asp Gly Arg	165 170 175	

eof-seq1.txt

gga	tat	tac	tta	aac	gaa	gat	gag	ctc	gtc	aac	cag	att	cgt	gag	gat	576
Gly	Tyr	Tyr	Leu	Asn	Glu	Asp	Glu	Leu	Val	Asn	Gln	Ile	Arg	Glu	Asp	
			180					185					190			
ctc	gct	aag	gct	gaa	aag	gct	ctc	gga	cgt	aag	ccc	act	gct	ctc	gtc	624
Leu	Ala	Lys	Ala	Glu	Lys	Ala	Leu	Gly	Arg	Lys	Pro	Thr	Ala	Leu	Val	
		195					200					205				
ctt	ggt	gct	ctt	gga	cga	tgt	ggt	aag	ggt	gcc	gtg	gac	ctt	ttc	ctg	672
Leu	Gly	Ala	Leu	Gly	Arg	Cys	Gly	Lys	Gly	Ala	Val	Asp	Leu	Phe	Leu	
	210					215					220					
aag	gcc	ggc	atg	cct	gat	gac	aac	att	acc	cgc	tgg	gac	ttg	aac	gag	720
Lys	Ala	Gly	Met	Pro	Asp	Asp	Asn	Ile	Thr	Arg	Trp	Asp	Leu	Asn	Glu	
225					230					235					240	
act	aag	gac	cgt	gat	ggc	cct	tac	gag	gag	atc	gcc	aag	gct	gat	gtc	768
Thr	Lys	Asp	Arg	Asp	Gly	Pro	Tyr	Glu	Glu	Ile	Ala	Lys	Ala	Asp	Val	
				245					250					255		
ttc	ctc	aac	gcc	atc	tac	ctc	tcc	aag	ccc	att	ccc	cct	ttc	atc	aac	816
Phe	Leu	Asn	Ala	Ile	Tyr	Leu	Ser	Lys	Pro	Ile	Pro	Pro	Phe	Ile	Asn	
			260					265					270			
caa	gaa	ctc	ctt	gcc	aag	caa	ggt	cgc	aac	ctc	gct	gtt	gtc	atc	gac	864
Gln	Glu	Leu	Leu	Ala	Lys	Gln	Gly	Arg	Asn	Leu	Ala	Val	Val	Ile	Asp	
		275					280					285				
gtt	tct	tgt	gac	acc	aca	aac	cct	cac	aac	cct	atc	ccc	atc	tac	tcc	912
Val	Ser	Cys	Asp	Thr	Thr	Asn	Pro	His	Asn	Pro	Ile	Pro	Ile	Tyr	Ser	
	290					295					300					
atc	aac	acc	acc	ttt	gag	gac	cca	acc	gtc	ccc	gtt	gag	atc	aag	gac	960
Ile	Asn	Thr	Thr	Phe	Glu	Asp	Pro	Thr	Val	Pro	Val	Glu	Ile	Lys	Asp	
305					310					315					320	
gat	cag	aac	aac	ctc	ccc	cta	tcc	gtc	atc	agc	att	gat	cac	ctc	cct	1008
Asp	Gln	Asn	Asn	Leu	Pro	Leu	Ser	Val	Ile	Ser	Ile	Asp	His	Leu	Pro	
				325					330					335		
tcc	atg	ctt	ccc	cgc	gag	gct	agt	gag	gcc	ttt	agt	gag	ggg	ctc	aag	1056
Ser	Met	Leu	Pro	Arg	Glu	Ala	Ser	Glu	Ala	Phe	Ser	Glu	Gly	Leu	Lys	
			340					345					350			
gag	tct	ctg	ctc	aca	ctc	aag	gat	cgc	gag	act	tcg	cgg	gtg	tgg	act	1104
Glu	Ser	Leu	Leu	Thr	Leu	Lys	Asp	Arg	Glu	Thr	Ser	Arg	Val	Trp	Thr	
		355					360					365				
gat	gct	gag	aag	ctc	ttc	cat	gag	aag	gtt	gcc	ctg	ttg	ccc	gag	gag	1152
Asp	Ala	Glu	Lys	Leu	Phe	His	Glu	Lys	Val	Ala	Leu	Leu	Pro	Glu	Glu	
	370					375					380					
ttg	aga	acc	aag	agt	gtt											1170
Leu	Arg	Thr	Lys	Ser	Val											
385					390											

<210> 42  
 <211> 390  
 <212> PRT  
 <213> Fusarium moniliforme  
 <400> 42

Met Ser Asp Tyr Pro His Ile Leu Leu Arg Ala Glu Glu Lys Pro Leu  
 1 5 10 15

eof-seql.txt

Glu His Arg Ser Phe Ser Pro Ala Ile Ile Lys Thr Leu Val Asp Ala  
 20 30  
 Gly Tyr Pro Ile Ser Val Glu Arg Ser Ser Thr Asp Pro Lys Phe Lys  
 35 40 45  
 Arg Ile Phe Glu Asp Ser Glu Tyr Glu Ala Ala Gly Ala Arg Leu Val  
 50 55 60  
 Asp Thr Gly Val Trp Pro Asn Ala Glu Pro Gly Thr Ile Ile Leu Gly  
 65 70 75 80  
 Leu Lys Glu Leu Pro Ser Glu Asp Phe Pro Leu Lys Asn Asp His Ile  
 85 90 95  
 Thr Phe Ala His Cys Tyr Lys Asn Gln Gly Gly Trp Glu Gln Val Leu  
 100 105 110  
 Gly Arg Trp Ala Arg Gly Gly Ser Arg Leu Tyr Asp Leu Glu Phe Leu  
 115 120 125  
 Val Asp Glu Gln Gly Arg Arg Val Ser Ala Phe Gly Tyr His Ala Gly  
 130 135 140  
 Phe Ala Gly Ala Ala Leu Gly Ile Lys Thr Leu Ala His Gln Leu Gln  
 145 150 155 160  
 Gly Ser Ser Ser Lys Leu Pro Ser Val Glu Thr Phe Thr Asp Gly Arg  
 165 170 175  
 Gly Tyr Tyr Leu Asn Glu Asp Glu Leu Val Asn Gln Ile Arg Glu Asp  
 180 185 190  
 Leu Ala Lys Ala Glu Lys Ala Leu Gly Arg Lys Pro Thr Ala Leu Val  
 195 200 205  
 Leu Gly Ala Leu Gly Arg Cys Gly Lys Gly Ala Val Asp Leu Phe Leu  
 210 215 220  
 Lys Ala Gly Met Pro Asp Asp Asn Ile Thr Arg Trp Asp Leu Asn Glu  
 225 230 235 240  
 Thr Lys Asp Arg Asp Gly Pro Tyr Glu Glu Ile Ala Lys Ala Asp Val  
 245 250 255  
 Phe Leu Asn Ala Ile Tyr Leu Ser Lys Pro Ile Pro Pro Phe Ile Asn  
 260 265 270  
 Gln Glu Leu Leu Ala Lys Gln Gly Arg Asn Leu Ala Val Val Ile Asp  
 275 280 285

eof-seq1.txt

Val Ser Cys Asp Thr Thr Asn Pro His Asn Pro Ile Pro Ile Tyr Ser  
 290 295 300

Ile Asn Thr Thr Phe Glu Asp Pro Thr Val Pro Val Glu Ile Lys Asp  
 305 310 315 320

Asp Gln Asn Asn Leu Pro Leu Ser Val Ile Ser Ile Asp His Leu Pro  
 325 330 335

Ser Met Leu Pro Arg Glu Ala Ser Glu Ala Phe Ser Glu Gly Leu Lys  
 340 345 350

Glu Ser Leu Leu Thr Leu Lys Asp Arg Glu Thr Ser Arg Val Trp Thr  
 355 360 365

Asp Ala Glu Lys Leu Phe His Glu Lys Val Ala Leu Leu Pro Glu Glu  
 370 375 380

Leu Arg Thr Lys Ser Val  
 385 390

<210> 43  
 <211> 856  
 <212> DNA  
 <213> Claviceps purpurea

<220>  
 <221> CDS  
 <222> (3)..(854)  
 <223> Lys1

<400> 43  
 ct ggc tac aaa gtt tcc gtg gag cgc tca cca tcg gac ccc gac ttt 47  
 Gly Tyr Lys Val Ser Val Glu Arg Ser Pro Ser Asp Pro Asp Phe  
 1 5 10 15

gtc cgc atc ttc caa gac tct gaa tat gag gcg gct gga gct tct ctg 95  
 Val Arg Ile Phe Gln Asp Ser Glu Tyr Glu Ala Ala Gly Ala Ser Leu  
 20 25 30

gtt ccc act ggc gtg tgg ccc aag gct gcg cca aac act ctg atc gtt 143  
 Val Pro Thr Gly Val Trp Pro Lys Ala Ala Pro Asn Thr Leu Ile Val  
 35 40 45

ggt ctc aag gag atc ccc gaa gag ggc ttt tct ctc acc aat gac cac 191  
 Gly Leu Lys Glu Ile Pro Glu Glu Gly Phe Ser Leu Thr Asn Asp His  
 50 55 60

ctg acc ttt gcc cat tgc tac aag aag caa gct ggc tgg aca cag gtt 239  
 Leu Thr Phe Ala His Cys Tyr Lys Lys Gln Ala Gly Trp Thr Gln Val  
 65 70 75

ctc gga cgc ttt gtc gca ggc aaa tcg acc ctc tac gac ctg gag ttt 287  
 Leu Gly Arg Phe Val Ala Gly Lys Ser Thr Leu Tyr Asp Leu Glu Phe  
 80 85 90 95

ctg gtc gat agc act gga aga cgc atc tct gct ttc gga ttc cac gct 335  
 Leu Val Asp Ser Thr Gly Arg Arg Ile Ser Ala Phe Gly Phe His Ala  
 100 105 110

eof-seql.txt

ggc	ttc	acg	ggc	gct	gct	ttg	gga	gtg	aag	aac	tgg	gct	tgg	caa	ctg	383
Gly	Phe	Thr	Gly	Ala	Ala	Leu	Gly	Val	Lys	Asn	Trp	Ala	Trp	Gln	Leu	
			115					120					125			
gcc	aat	cca	gcc	aag	aag	ctt	ccc	gcc	gtg	tcc	aca	ttc	act	gac	ggc	431
Ala	Asn	Pro	Ala	Lys	Lys	Leu	Pro	Ala	Val	Ser	Thr	Phe	Thr	Asp	Gly	
		130					135					140				
aag	gga	tac	tat	gtg	aac	gaa	acc	gag	ttg	gtg	gag	caa	atc	cgc	aag	479
Lys	Gly	Tyr	Tyr	Val	Asn	Glu	Thr	Glu	Leu	Val	Glu	Gln	Ile	Arg	Lys	
	145					150					155					
gat	ctc	gag	gca	gga	gag	aag	att	ctt	ggc	cgt	aag	cca	aca	gcc	ttt	527
Asp	Leu	Glu	Ala	Gly	Glu	Lys	Ile	Leu	Gly	Arg	Lys	Pro	Thr	Ala	Phe	
					165					170					175	
gtc	ctg	gga	gct	ttg	ggc	cgc	tgc	ggc	agg	ggc	gct	tgc	gat	ttg	ttc	575
Val	Leu	Gly	Ala	Leu	Gly	Arg	Cys	Gly	Arg	Gly	Ala	Cys	Asp	Leu	Phe	
				180				185						190		
ctc	aag	gca	ggc	ttg	ccc	gaa	gag	aac	atc	acc	cga	tgg	gat	ttg	gcc	623
Leu	Lys	Ala	Gly	Leu	Pro	Glu	Glu	Asn	Ile	Thr	Arg	Trp	Asp	Leu	Ala	
			195					200					205			
gaa	acc	cgc	gac	cgc	caa	ggc	ccg	tac	gag	gag	att	gct	cag	cac	gat	671
Glu	Thr	Arg	Asp	Arg	Gln	Gly	Pro	Tyr	Glu	Glu	Ile	Ala	Gln	His	Asp	
		210					215					220				
atc	ttc	ctg	aac	gct	atc	tat	ctc	tcc	gag	ccc	atc	cca	cct	ttt	gtc	719
Ile	Phe	Leu	Asn	Ala	Ile	Tyr	Leu	Ser	Glu	Pro	Ile	Pro	Pro	Phe	Val	
	225					230					235					
aac	aac	gag	ctt	ctt	tct	aag	cct	ggc	cgc	aaa	ttg	agt	ggt	gtc	ata	767
Asn	Asn	Glu	Leu	Leu	Ser	Lys	Pro	Gly	Arg	Lys	Leu	Ser	Val	Val	Ile	
					245					250					255	
gac	gtc	tcc	tgc	gac	acc	acc	aac	ccc	cac	aac	ccc	atc	ccc	atc	tac	815
Asp	Val	Ser	Cys	Asp	Thr	Thr	Asn	Pro	His	Asn	Pro	Ile	Pro	Ile	Tyr	
				260					265					270		
agc	atc	aac	acc	acc	ttc	gac	agc	ccc	acc	gtt	gcc	gtc	aa			856
Ser	Ile	Asn	Thr	Thr	Phe	Asp	Ser	Pro	Thr	Val	Ala	Val				
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Arg	Ile	Phe	Gln	Asp	Ser	Glu	Tyr	Glu	Ala	Ala	Gly	Ala	Ser	Leu	Val
			20					25					30		
Pro	Thr	Gly	Val	Trp	Pro	Lys	Ala	Ala	Pro	Asn	Thr	Leu	Ile	Val	Gly
		35					40					45			
Leu	Lys	Glu	Ile	Pro	Glu	Glu	Gly	Phe	Ser	Leu	Thr	Asn	Asp	His	Leu
	50					55					60				

eof-seql.txt

Thr Phe Ala His Cys Tyr Lys Lys Gln Ala Gly Trp Thr Gln Val Leu  
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Gly Arg Phe Val Ala Gly Lys Ser Thr Leu Tyr Asp Leu Glu Phe Leu  
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Val Asp Ser Thr Gly Arg Arg Ile Ser Ala Phe Gly Phe His Ala Gly  
100 105 110

Phe Thr Gly Ala Ala Leu Gly Val Lys Asn Trp Ala Trp Gln Leu Ala  
115 120 125

Asn Pro Ala Lys Lys Leu Pro Ala Val Ser Thr Phe Thr Asp Gly Lys  
130 135 140

Gly Tyr Tyr Val Asn Glu Thr Glu Leu Val Glu Gln Ile Arg Lys Asp  
145 150 155 160

Leu Glu Ala Gly Glu Lys Ile Leu Gly Arg Lys Pro Thr Ala Phe Val  
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Leu Gly Ala Leu Gly Arg Cys Gly Arg Gly Ala Cys Asp Leu Phe Leu  
180 185 190

Lys Ala Gly Leu Pro Glu Glu Asn Ile Thr Arg Trp Asp Leu Ala Glu  
195 200 205

Thr Arg Asp Arg Gln Gly Pro Tyr Glu Glu Ile Ala Gln His Asp Ile  
210 215 220

Phe Leu Asn Ala Ile Tyr Leu Ser Glu Pro Ile Pro Pro Phe Val Asn  
225 230 235 240

Asn Glu Leu Leu Ser Lys Pro Gly Arg Lys Leu Ser Val Val Ile Asp  
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