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- (71) Applicant: **PIONEER HI-BRED INTERNATIONAL, INC.** [US/US]; 7100 N.W. 62nd Avenue, Johnston, Iowa 50131-1014 (US).
- (72) Inventors; and
- (71) Applicants : **CHO, Myeong-Je** [US/US]; c/o Pioneer Hi-Bred International, Inc., 7250 N.W. 62nd Avenue, Johnston, Iowa 50131-0552 (US). **ELLIS, Samuel R.** [US/US]; c/o Pioneer Hi-Bred International, Inc., 7250 N.W. 62nd

Avenue, Johnston, Iowa 50131-0552 (US). **GOR-DON-KAMM, William J.** [US/US]; c/o Pioneer Hi-Bred International, Inc., 7250 N.W. 62nd Avenue, Johnston, Iowa 50131-0552 (US). **ZHAO, Zuo-Yu** [US/US]; c/o Pioneer Hi-Bred International, Inc., 7250 N.W. 62nd Avenue, Johnston, Iowa 50131-0552 (US).

(74) Agent: **LAPPEGARD, Kathryn K.**; Pioneer Hi-Bred International, Inc., 7250 N.W. 62nd Avenue, Johnston, Iowa 50131-0552 (US).

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- (54) Title: METHODS AND COMPOSITIONS FOR PRODUCING AND SELECTING TRANSGENIC PLANTS

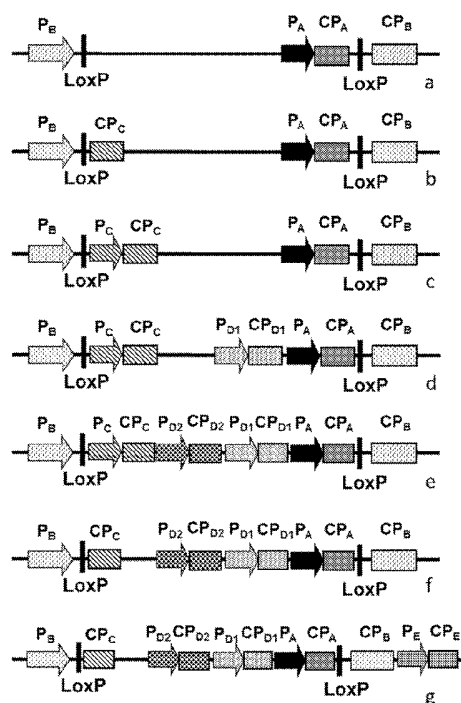


FIG. 9

(57) Abstract: Compositions and methods are provided for the production and selection of transgenic plants and plant parts, for increasing the transformation frequency of a plant or plant part, and for regulating the expression of a transgene, such as a herbicide tolerance polynucleotide. The methods and compositions allow for the delay in the expression of herbicide tolerance polynucleotides until a point in development during which herbicide selection is more efficient. Compositions comprise polynucleotide constructs comprising an excision cassette that separates a transgene, such as a herbicide tolerance polynucleotide, from its promoter and host cells comprising the same. The excision cassette comprises a polynucleotide encoding a site-specific recombinase operably linked to an inducible promoter and expression of the recombinase leads to excision of the excision cassette and expression of the transgene.



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METHODS AND COMPOSITIONS FOR PRODUCING AND SELECTING TRANSGENIC PLANTS

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5 The official copy of the sequence listing is submitted electronically via EFS-Web as an ASCII formatted sequence listing, created on March 12, 2013 and having a size of 308 kilobytes and is filed concurrently with the specification. The sequence listing contained in this ASCII formatted document is part of the specification and is herein incorporated by reference in its entirety.

FIELD OF THE INVENTION

10 The present invention relates to the genetic modification of plants. More particularly, the compositions and methods are directed to the production and selection of transgenic plants.

BACKGROUND OF THE INVENTION

15 Current genetic engineering technology allows for the production of transgenic plants with desired traits. In some instances, it is desirable to delay expression of a transgene until a certain developmental stage is reached or environmental condition is encountered. Such transgenes can confer a desired trait or can serve as a selectable
20 marker to aid in the identification of transgenic plants that have been successfully engineered with a polynucleotide of interest.

25 For example, herbicide tolerance polynucleotides, which encode polypeptides that confer tolerance to specific herbicides, can be introduced into a plant to generate a herbicide tolerant plant and/or to serve as a selectable marker for the introduction of another polynucleotide of interest. Direct selection with herbicides, such as glyphosate and sulfonylureas, during early stages of transgenic plant production (i.e., tissue proliferation) has been relatively inefficient when transforming maize and sugarcane

(Experimental Example 1 and unpublished data). Larger clusters of maize cells may be less sensitive to herbicides such as glyphosate and some nontransgenic calli may still grow in the presence of the herbicide (Wang *et al.* (2009) *Handbook of Maize: Genetics and Genomics*, J.L. Bennetzen and S. Hake, eds., pp. 609-639). As observed in wheat,
5 however, selection at the stage of regeneration was more effective and escapes were rarely regenerated (Zhou *et al.* (1995) *Plant Cell Rep* 15:159-163; Hu *et al.* (2003) *Plant Cell Rep* 21:1010-1019).

Thus, methods and compositions are needed that allow for the delayed expression of transgenes to reduce the potential for negative effects on transformed tissues,
10 particularly during development. Such methods and compositions would be especially useful for delaying the expression of herbicide tolerance polynucleotides until a stage at which herbicide selection is more efficient.

BRIEF SUMMARY OF THE INVENTION

15 Compositions and methods are provided for the production and selection of transgenic plants and plant parts, for increasing the transformation frequency of a plant or plant part, and for regulating the expression of a transgene, such as a herbicide tolerance polynucleotide. The methods and compositions allow for the delay of the expression of a transgene (e.g., herbicide tolerance polynucleotide) by the presence and subsequent
20 excision of an excision cassette that separates the transgene (e.g., herbicide tolerance polynucleotide) from a promoter that drives its expression. Excision of the excision cassette is mediated by a site-specific recombinase, the expression of which is regulated by an inducible promoter, which results in the operable linkage of the transgene (e.g., herbicide tolerance polynucleotide) and its promoter and subsequent expression of the
25 transgene (e.g., herbicide tolerance polynucleotide). These methods and compositions are useful for delaying the expression of transgenes that might otherwise negatively affect the development or growth of a transformed tissue or plant.

The herbicide tolerance polynucleotide can serve as a means for imparting herbicide tolerance to a plant or plant part and/or can function as a selectable marker,
30 aiding in the identification of a transgenic plant or plant part comprising another

polynucleotide of interest or lacking a polynucleotide of interest that has been excised from the excision cassette. In some of these embodiments, the excision of the excision cassette and expression of the herbicide tolerance polynucleotide is delayed until after the tissue proliferation stage of transgenic plant production to allow for more efficient herbicide selection.

In some embodiments, the inducible promoter regulating the expression of the recombinase, excision of the excision cassette, and expression of the herbicide tolerance polynucleotide is one that is induced by stress (e.g., cold temperatures, desiccation) or by a chemical (e.g., antibiotic, herbicide).

Compositions include polynucleotide constructs comprising a promoter that is active in a plant, a herbicide tolerance polynucleotide, and an excision cassette, wherein the excision cassette comprises an inducible promoter operably linked to a site-specific recombinase-encoding polynucleotide, and wherein excision of the excision cassette allows for the operable linkage of the promoter and the herbicide tolerance polynucleotide. Host cells, such as plant cells, and plants and plant parts comprising the polynucleotide constructs are further provided.

The following embodiments are encompassed by the present invention.

1. A polynucleotide construct comprising:

- a) an excision cassette comprising an expression cassette A (EC_A) comprising:
 - i) a promoter A (P_A), wherein said P_A is an inducible promoter; and
 - ii) a coding polynucleotide A (CP_A) encoding a site-specific recombinase;

wherein said P_A is operably linked to said CP_A ; and

wherein said excision cassette is flanked by a first and a second recombination site, wherein said first and said second recombination sites are recombinogenic with respect to one another and are directly repeated, and wherein said site-specific recombinase can recognize and implement recombination at said first and said second recombination sites; thereby excising said excision cassette;

- b) a coding polynucleotide B (CP_B) encoding a herbicide tolerance polypeptide; and
- c) a promoter B (P_B), wherein said P_B is operably linked to said CP_B after excision of said excision cassette;
- 5 wherein said P_A and P_B are active in a plant cell.

2. The polynucleotide construct of embodiment 1, wherein said inducible promoter is selected from the group consisting of a stress-inducible promoter and a chemical-inducible promoter.

- 10 3. The polynucleotide construct of embodiment 2, wherein said chemical-inducible promoter comprises a promoter comprising a tet operator.

4. The polynucleotide construct of embodiment 3, wherein said polynucleotide construct further comprises a coding polynucleotide F (CP_F) encoding a sulfonylurea-responsive transcriptional repressor protein, wherein said CP_F is operably
15 linked to a promoter active in a plant cell.

5. The polynucleotide construct of embodiment 2, wherein said stress-inducible promoter can be induced in response to cold, drought, high salinity, desiccation, or a combination thereof.

6. The polynucleotide construct of embodiment 2 or 5, wherein said stress-
20 inducible promoter is a maize *rab17* promoter or an active variant or fragment thereof.

7. The polynucleotide construct of any one of embodiments 2, 5 and 6, wherein said stress-inducible promoter has a nucleotide sequence selected from the group consisting of:

- a) the nucleotide sequence having the sequence set forth in SEQ ID
25 NO: 18;
- b) a nucleotide sequence having at least 70% sequence identity to the sequence set forth in SEQ ID NO: 18;

- c) a nucleotide sequence comprising at least 50 contiguous nucleotides of the sequence set forth in SEQ ID NO: 18;
- d) the nucleotide sequence set forth in nucleotides 291-430 of SEQ ID NO: 18; and
- 5 e) a nucleotide sequence having at least 70% sequence identity to the sequence set forth in nucleotides 291-430 of SEQ ID NO: 18.

8. The polynucleotide construct of embodiment 6 or 7, wherein said EC_A further comprises an attachment B (attB) site between said stress-inducible promoter and
10 said CP_A.

9. The polynucleotide construct of embodiment 8, wherein said attB site has a nucleotide sequence selected from the group consisting of:

- 15 a) a nucleotide sequence having at least 70% sequence identity to the sequence set forth in SEQ ID NO: 20; and
- b) the nucleotide sequence set forth in SEQ ID NO: 20.

10. The polynucleotide construct of any one of embodiments 1-9, wherein said site-specific recombinase is selected from the group consisting of FLP, Cre, S-CRE, V-CRE, Dre, SSV1, lambda Int, phi C31 Int, HK022, R, Gin, Tn1721, CinH, ParA, Tn5053, Bxb1, TP907-1, and U153.

11. The polynucleotide construct of any one of embodiments 1-10, wherein said CP_A has the nucleotide sequence selected from the group consisting of:

- 25 a) the nucleotide sequence set forth in SEQ ID NO: 33 or 35;
- b) a nucleotide sequence having at least 70% sequence identity to SEQ ID NO: 33 or 35;
- c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 34 or 36; and

d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 70% sequence identity to SEQ ID NO: 34 or 36.

12. The polynucleotide construct of any one of embodiments 1-11, wherein P_B is a constitutive promoter.

13. The polynucleotide construct of embodiment 12, wherein said P_B is selected from the group consisting of a ubiquitin promoter, an oleosin promoter, an actin promoter, and a *Mirabilis* mosaic virus (MMV) promoter.

14. The polynucleotide construct of any one of embodiments 1-13, wherein said excision cassette further comprises a coding polynucleotide C (CP_C) encoding a selectable marker, wherein said CP_C is operably linked to a promoter active in a plant cell.

15. The polynucleotide construct of embodiment 14, wherein said CP_C is operably linked to P_B before excision of the excision cassette.

16. The polynucleotide construct of embodiment 14, wherein said excision cassette further comprises a promoter C (P_C), wherein P_C is operably linked to said CP_C.

17. The polynucleotide construct of embodiment 16, wherein said P_C is a constitutive promoter.

18. The polynucleotide construct of embodiment 17, wherein said P_C is selected from the group consisting of an ubiquitin promoter, an oleosin promoter, an actin promoter, and a *Mirabilis* mosaic virus (MMV) promoter.

19. The polynucleotide construct of any one of embodiments 14-18, wherein said selectable marker is selected from the group consisting of a fluorescent protein, an

antibiotic resistance polypeptide, a herbicide tolerance polypeptide, and a metabolic enzyme.

20. The polynucleotide construct of embodiment 19, wherein said fluorescent
5 protein is selected from the group consisting of a yellow fluorescent protein, a red fluorescent protein, a cyan fluorescent protein, and a green fluorescent protein.

21. The polynucleotide construct of embodiment 19, wherein said fluorescent
protein comprises a *Discosoma* red fluorescent protein.

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22. The polynucleotide construct of embodiment 19, wherein said antibiotic
resistance polypeptide comprises a neomycin phosphotransferase II.

23. The polynucleotide construct of embodiment 19, wherein said herbicide
15 tolerance polypeptide encoded by CP_C comprises a phosphinothricin acetyl transferase.

24. The polynucleotide construct of embodiment 19, wherein said metabolic
enzyme comprises a phosphomannose isomerase.

20 25. The polynucleotide construct of any one of embodiments 14-24, wherein
said excision cassette comprises more than one polynucleotide encoding a distinct
selectable marker, wherein said polynucleotide encoding a selectable marker is operably
linked to a promoter active in a plant cell.

25 26. The polynucleotide construct of embodiment 25, wherein said excision
cassette comprises at least a first and a second polynucleotide encoding a selectable
marker, wherein said first polynucleotide encodes a yellow fluorescent protein, and
wherein said second polynucleotide encodes a phosphinothricin acetyl transferase or a
neomycin phosphotransferase II.

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27. The polynucleotide construct of any one of embodiments 1-26, wherein said herbicide tolerance polypeptide encoded by CP_B confers tolerance to a herbicide selected from the group consisting of glyphosate, an ALS inhibitor, an acetyl Co-A carboxylase inhibitor, a synthetic auxin, a protoporphyrinogen oxidase (PPO) inhibitor herbicide, a pigment synthesis inhibitor herbicide, a phosphinothricin acetyltransferase, a phytoene desaturase inhibitor, a glutamine synthase inhibitor, a hydroxyphenylpyruvate dioxygenase inhibitor, and a protoporphyrinogen oxidase inhibitor.

28. The polynucleotide construct of embodiment 27, wherein said ALS inhibitor is selected from the group consisting of a sulfonylurea, a triazolopyrimidine, a pyrimidinyloxy(thio)benzoate, an imidazolinone, and a sulfonylaminocarbonyltriazolinone.

29. The polynucleotide construct of any one of embodiments 1-28, wherein said herbicide tolerance polypeptide encoded by CP_B comprises a glyphosate-N-acetyltransferase (GLYAT) polypeptide or an ALS inhibitor-tolerance polypeptide.

30. The polynucleotide construct of embodiment 29, wherein said polynucleotide encoding said GLYAT polypeptide has a nucleotide sequence selected from the group consisting of:

- a) the nucleotide sequence set forth in SEQ ID NO: 47 or 49;
- b) a nucleotide sequence having at least 95% sequence identity to SEQ ID NO: 47 or 49;
- c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 48 or 50; and
- d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 95% sequence identity to SEQ ID NO: 48 or 50.

31. The polynucleotide construct of embodiment 29, wherein said ALS inhibitor-tolerance polypeptide comprises the highly resistant ALS (HRA) mutation of acetolactate synthase.

5 32. The polynucleotide constructs of any one of embodiments 1-31, wherein said polynucleotide construct comprises more than one polynucleotide encoding a distinct herbicide tolerance polypeptide, wherein the polynucleotide encoding a herbicide tolerance polypeptide is operably linked to a promoter active in a plant cell.

10 33. The polynucleotide construct of embodiment 32, wherein said polynucleotide construct comprises at least a first and a second polynucleotide encoding a herbicide tolerance polypeptide, wherein said first polynucleotide encodes an ALS inhibitor-tolerance polypeptide and wherein said second polynucleotide encodes a GLYAT polypeptide.

15 34. The polynucleotide construct of any one of embodiments 1-33, wherein said excision cassette further comprises a coding polynucleotide D (CP_D) encoding a cell proliferation factor, wherein said CP_D is operably linked to a promoter active in a plant cell.

20 35. The polynucleotide construct of embodiment 34, wherein said cell proliferation factor is selected from the group consisting of a Lec1 polypeptide, a Kn1 polypeptide, a WUSCHEL polypeptide, a Zwillie polypeptide, a babyboom polypeptide, an Aintegumenta polypeptide (ANT), a FUS3 polypeptide, a Kn1 polypeptide, a STM
25 polypeptide, an OSH1 polypeptide, and a SbH1 polypeptide.

36. The polynucleotide construct of embodiment 35, wherein said cell proliferation factor is selected from the group consisting of a WUSCHEL polypeptide and a babyboom polypeptide.

37. The polynucleotide construct of any one of embodiments 34-36, wherein said babyboom polypeptide comprises at least two AP2 domains and at least one of the following amino acid sequences:

5 a) the amino acid sequence set forth in SEQ ID NO: 67 or an amino acid sequence that differs from the amino acid sequence set forth in SEQ ID NO: 67 by one amino acid; and

b) the amino acid sequence set forth in SEQ ID NO: 68 or an amino acid sequence that differs from the amino acid sequence set forth in SEQ ID NO: 68 by one amino acid.

10

38. The polynucleotide construct of any one of embodiments 34-36, wherein said CP_D has a nucleotide sequence selected from the group consisting of:

a) the nucleotide sequence set forth in SEQ ID NO: 55, 57, 58, 60, 74, 76, 78, 80, 82, 84, 86, 87, 88, 90, 92, 94, 96, 98, 99, or 101;

15 b) a nucleotide sequence having at least 70% sequence identity to SEQ ID NO: 55, 57, 58, 60, 74, 76, 78, 80, 82, 84, 86, 87, 88, 90, 92, 94, 96, 98, 99, or 101;

c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 56, 59, 75, 77, 79, 81, 83, 85, 89, 91, 93, 95, 97, 100, or 102; and

20

d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO: 56, 59, 75, 77, 79, 81, 83, 85, 89, 91, 93, 95, 97, 100, or 102.

25 39. The polynucleotide construct of any one of embodiments 34-38, wherein said excision cassette further comprises a promoter D (P_D) operably linked to said CP_D.

40. The polynucleotide construct of embodiment 39, wherein said P_D is a constitutive promoter.

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41. The polynucleotide construct of embodiment 40, wherein said P_D is a ubiquitin promoter or an oleosin promoter.

42. The polynucleotide construct of any one of embodiments 36-41, wherein
5 said excision cassette comprises more than one coding polynucleotide D (CP_D) encoding a distinct cell proliferation factor, wherein the CP_D is operably linked to a promoter active in a plant cell.

43. The polynucleotide construct of embodiment 42, wherein said excision
cassette comprises at least a first coding polynucleotide D (CP_{D1}) encoding a babyboom
10 polypeptide and a second coding polynucleotide D (CP_{D2}) encoding a WUSCHEL polypeptide.

44. The polynucleotide construct of any one of embodiments 35, 36, 42, and
43, wherein said polynucleotide encoding a WUSCHEL polypeptide has a nucleotide
15 sequence selected from the group consisting of:

a) the nucleotide sequence set forth in SEQ ID NO: 103, 105, 107, or
109; and

b) a nucleotide sequence having at least 70% sequence identity to
SEQ ID NO: 103, 105, 107, or 109;

20 c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 104, 106, 108, or 110; and

d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 70% sequence identity to SEQ ID NO: 104, 106, 108, or
110.

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45. The polynucleotide construct of any one of embodiments 35, 36, 42, 43, and 44, wherein said polynucleotide encoding a WUSCHEL polypeptide is operably linked to a maize In2-2 promoter or a nopaline synthase promoter.

46. The polynucleotide construct of any one of embodiments 1-45, wherein said polynucleotide construct further comprises a coding polynucleotide E (CP_E) encoding a polypeptide of interest, wherein said CP_E is operably linked to a promoter active in a plant cell.

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47. The polynucleotide construct of embodiment 46, wherein said excision cassette comprises said CP_E.

48. The polynucleotide construct of embodiment 46, wherein said CP_E is outside of the excision cassette.

10

49. The polynucleotide construct of any one of embodiments 46-48, wherein said polynucleotide construct further comprises a promoter E (P_E) operably linked to said CP_E.

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50. The polynucleotide construct of embodiment 1, wherein said polynucleotide construct comprises:

- a) a first ubiquitin promoter;
- b) an excision cassette flanked by loxP recombination sites that are are recombinogenic with respect to one another and are directly repeated, wherein said excision cassette comprises:
 - i) a polynucleotide encoding a phosphinothricin acetyl transferase (PAT) or a neomycin phosphotransferase II (NPTII);
 - ii) a second ubiquitin promoter;
 - iii) a polynucleotide encoding a yellow fluorescent protein;
 - iv) a promoter comprising a maize *rab17* promoter and an attachment B (attB) site;
 - v) a polynucleotide encoding a CRE recombinase;
 - vi) a nopaline synthase promoter;
 - vii) a polynucleotide encoding a maize Wuschel 2 polypeptide;

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- viii) a third ubiquitin promoter; and
- ix) a babyboom polynucleotide; and
- c) a GLYAT polynucleotide;

wherein said first ubiquitin promoter is operably linked to said polynucleotide
5 encoding said PAT or NPTII and wherein said first ubiquitin promoter is operably linked
to said GLYAT polynucleotide upon excision of said excision cassette;

wherein said second ubiquitin promoter is operably linked to said polynucleotide
encoding said yellow fluorescent protein;

wherein said promoter comprising said maize *rab17* promoter and said attB site is
10 operably linked to said polynucleotide encoding said CRE recombinase;

wherein said nopaline synthase promoter is operably linked to said polynucleotide
encoding said maize Wuschel 2 polypeptide;

and wherein said third ubiquitin promoter is operably linked to said babyboom
polynucleotide.

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51. The polynucleotide construct of embodiment 1, wherein said
polynucleotide construct comprises:

- a) a ubiquitin promoter;
- b) an excision cassette flanked by loxP recombination sites that are

20 are recombinogenic with respect to one another and are directly repeated, wherein said
excision cassette comprises:

- i) a polynucleotide encoding a *Discosoma* red fluorescent
protein;

- ii) a promoter comprising a maize *rab17* promoter and an
25 attachment B (attB) site; and

- iii) a polynucleotide encoding a CRE recombinase; and

- c) a GLYAT polynucleotide;

wherein said ubiquitin promoter is operably linked to said polynucleotide
encoding said *Discosoma* red fluorescent protein and wherein said ubiquitin promoter is

operably linked to said GLYAT polynucleotide upon excision of said excision cassette;
and

wherein said promoter comprising said maize *rab17* promoter and said attB site is
operably linked to said polynucleotide encoding said CRE recombinase.

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52. The polynucleotide construct of embodiment 1, wherein said
polynucleotide construct comprises:

- a) a ubiquitin promoter;
- b) an excision cassette flanked by loxP recombination sites that are
10 are recombinogenic with respect to one another and are directly repeated, wherein said
excision cassette comprises:
 - i) an actin promoter;
 - ii) a polynucleotide encoding a *Discosoma* red fluorescent
protein;
 - 15 iii) a promoter comprising a maize *rab17* promoter and an
attachment B (attB) site; and
 - iv) a polynucleotide encoding a CRE recombinase; and
- c) a GLYAT polynucleotide;

wherein said ubiquitin promoter is operably linked to said GLYAT polynucleotide
20 upon excision of said excision cassette;

wherein said actin promoter is operably linked to said polynucleotide encoding
said *Discosoma* red fluorescent protein; and

wherein said promoter comprising said maize *rab17* promoter and said attB site is
operably linked to said polynucleotide encoding said CRE recombinase.

25

53. A host cell comprising the polynucleotide construct of any one of
embodiments 1-52.

54. A plant cell comprising the polynucleotide construct of any one of
30 embodiments 1-52.

55. A plant or plant part comprising said plant cell of embodiment 54.

56. The plant or plant part of embodiment 55, wherein said plant or plant part
5 is a dicot.

57. The plant or plant part of embodiment 55, wherein said plant or plant part
is a monocot.

10 58. The plant or plant part of embodiment 57, wherein said monocot is
selected from the group consisting of maize, rice, sorghum, barley, wheat, millet, oat, rye,
triticale, sugarcane, switchgrass, and turf/forage grass.

59. The plant or plant part of any one of embodiments 55-58, wherein said
15 plant or plant part is recalcitrant.

60. The plant or plant part of embodiment 59, wherein said plant or plant part
is a sugarcane cultivar selected from the group consisting of CP96-1252, CP01-1372,
CPCL97-2730, HoCP85-845, CP89-2143, and KQ228.

20 61. The plant or plant part of any one of embodiments 55-60, wherein said
plant part is a seed.

62. A method for producing a transgenic plant or plant part, said method
25 comprising introducing said polynucleotide construct of any one of embodiments 1-52
into a plant or plant part.

63. A method for regulating the expression of a herbicide tolerance
polynucleotide, wherein said method comprises:

- a) providing the host cell of embodiment 53, the plant cell of embodiment 54, or the plant or plant part of any one of embodiments 55-61; and,
- b) inducing the expression of said site-specific recombinase, thereby excising said excision cassette from said polynucleotide construct and expressing said herbicide tolerance polynucleotide.

64. A method for selecting a herbicide tolerant plant cell, said method comprising the steps of:

- A) providing a population of plant cells, wherein at least one plant cell in the population comprises a polynucleotide construct comprising:

- a) an excision cassette comprising an expression cassette A (EC_A) comprising:
- i) a promoter A (P_A), wherein said P_A is an inducible promoter; and
- ii) a coding polynucleotide A (CP_A) encoding a site-specific recombinase;

wherein said P_A is operably linked to said CP_A;

- b) a coding polynucleotide B (CP_B) encoding a herbicide tolerance polypeptide; and
- c) a promoter B (P_B), wherein said P_B is operably linked to said CP_B after excision of said excision cassette;

wherein said P_A and P_B are active in a plant cell; and

- wherein said excision cassette is flanked by a first and a second recombination site, wherein said first and said second recombination sites are recombinogenic with respect to one another and are directly repeated, and wherein said site-specific recombinase can recognize and implement recombination at said first and said second recombination sites; thereby excising said excision cassette;

- B) inducing the expression of said site-specific recombinase; and

C) contacting said population of plant cells with a herbicide to which said herbicide tolerance polypeptide confers tolerance, thereby selecting for a plant cell having tolerance to said herbicide.

5 65. The method of embodiment 64, wherein said provided population of plant cells is cultured into a population of plant tissues or plants prior to, during, or after said step B), and wherein said step C) comprises contacting said population of plant tissues or plants with said herbicide.

10 66. The method of embodiment 65, wherein said step C) occurs during or after regeneration of said provided population of plant cells into a population of plants.

 67. The method of embodiment 64, wherein said provided population of plant cells is a population of immature or mature seeds, wherein at least one immature or
15 mature seed within said population of immature or mature seeds comprises said polynucleotide construct.

 68. The method of embodiment 67, wherein said provided population of seeds is planted prior to, during, or after said step B) to produce a population of plants, and
20 wherein said step C) comprises contacting said population of plants with said herbicide.

 69. The method of embodiment 75, wherein said provided population of plant cells is a population of plant tissues, wherein at least one plant tissue within said population of plant tissues comprises said polynucleotide construct.

25 70. The method of embodiment 69, wherein said provided population of plant tissues is cultured into a population of plants prior to, during, or after said step B), and wherein said step C) comprises contacting said population of plants with said herbicide.

71. The method of embodiment 64, wherein said provided population of plant cells is a population of plants, wherein at least one plant within said population of plants comprises said polynucleotide construct.

5 72. The method of any one of embodiments 64-71, wherein said method further comprises introducing said polynucleotide construct into said at least one plant cell before step A).

73. The method of any one of embodiments 64-72, wherein said inducible
10 promoter P_A is selected from the group consisting of a stress-inducible promoter and a chemical-inducible promoter.

74. The method of embodiment 73, wherein said chemical-inducible promoter comprises a promoter comprising a tet operator.

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75. The method of embodiment 74, wherein said polynucleotide construct or said at least one plant cell further comprises a coding polynucleotide F (CP_F) encoding a sulfonylurea-responsive transcriptional repressor protein, wherein said CP_F is operably linked to a promoter active in a plant cell, and wherein said inducing comprises
20 contacting said population of plant cells with a sulfonylurea compound.

76. The method of embodiment 73, wherein said stress-inducible promoter is induced in response to cold, drought, desiccation, high salinity, or a combination thereof.

77. The method of embodiment 73 or 76, wherein said stress-inducible promoter comprises a drought-inducible promoter, and wherein said inducing comprises
25 desiccating said population of plant cells.

78. The method of embodiment 77, wherein said desiccating occurs during the maturation of an immature seed.

79. The method of embodiment 73, wherein said stress-inducible promoter is a maize *rab17* promoter or an active variant or fragment thereof.

80. The method of embodiment 73, wherein said stress-inducible promoter has a nucleotide sequence selected from the group consisting of:

- 5 a) the nucleotide sequence having the sequence set forth in SEQ ID NO: 18;
- b) a nucleotide sequence having at least 70% sequence identity to the sequence set forth in SEQ ID NO: 18;
- c) a nucleotide sequence comprising at least 50 contiguous
10 nucleotides of the sequence set forth in SEQ ID NO: 18;
- d) the nucleotide sequence set forth in nucleotides 291-430 of SEQ ID NO: 18; and
- e) a nucleotide sequence having at least 70% sequence identity to the sequence set forth in nucleotides 291-430 of SEQ ID NO: 18.

15

81. The method of embodiment 79 or 80, wherein said EC_A further comprises an attachment B (attB) site between said stress-inducible promoter and said CP_A.

82. The method of embodiment 81, wherein said attB site has a nucleotide
20 sequence selected from the group consisting of:

- a) a nucleotide sequence having at least 70% sequence identity to the sequence set forth in SEQ ID NO: 20; and
- b) the nucleotide sequence set forth in SEQ ID NO: 20.

25 83. The method of any one of embodiments 64-82, wherein said site-specific recombinase is selected from the group consisting of FLP, Cre, S-CRE, V-CRE, Dre, SSV1, lambda Int, phi C31 Int, HK022, R, Gin, Tn1721, CinH, ParA, Tn5053, Bxb1, TP907-1, and U153.

84. The method of any one of embodiments 64-83, wherein said CP_A has the nucleotide sequence selected from the group consisting of:

- a) the nucleotide sequence set forth in SEQ ID NO: 33 or 35;
- b) a nucleotide sequence having at least 70% sequence identity to
5 SEQ ID NO: 33 or 35;
- c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 34 or 36; and
- d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 70% sequence identity to SEQ ID NO: 34 or 36.

10

85. The method of any one of embodiments 64-84, wherein P_B is a constitutive promoter.

86. The method of embodiment 85, wherein said P_B is selected from the group
15 consisting of a ubiquitin promoter, an oleosin promoter, an actin promoter, and a *Mirabilis* mosaic virus promoter.

87. The method of any one of embodiments 64-86, wherein said excision cassette further comprises a coding polynucleotide C (CP_C), wherein said CP_C encodes a
20 selectable marker, wherein said CP_C is operably linked to a promoter active in a plant cell, and wherein said method further comprises a selection step prior to step B), wherein those plant cells within said population of plant cells that comprise said selectable marker are identified and wherein these selected plant cells comprise the population of plant cells that are induced in step B).

25

88. The method of embodiment 87, wherein said CP_C is operably linked to P_B.

89. The method of embodiment 87, wherein said excision cassette further comprises a promoter C (P_C), wherein P_C is operably linked to said CP_C.

30

90. The method of embodiment 89, wherein P_C is a constitutive promoter.

91. The method of embodiment 90, wherein said P_C is selected from the group consisting of a ubiquitin promoter, an oleosin promoter, an actin promoter, and a
5 *Mirabilis* mosaic virus promoter.

92. The method of any one of embodiments 87-91, wherein said selectable marker is selected from the group consisting of a fluorescent protein, an antibiotic resistance polypeptide, a herbicide tolerance polypeptide, and a metabolic enzyme.

10

93. The method of embodiment 92, wherein said fluorescent protein is selected from the group consisting of a yellow fluorescent protein, a red fluorescent protein, a cyan fluorescent protein, and a green fluorescent protein.

15 94. The method of embodiment 92, wherein said fluorescent protein comprises a *Discosoma* red fluorescent protein.

95. The method of embodiment 92, wherein said antibiotic resistance polypeptide comprises a neomycin phosphotransferase II.

20

96. The method of embodiment 92, wherein said herbicide tolerance polypeptide encoded by CP_C comprises a phosphinothricin acetyl transferase.

97. The method of embodiment 92, wherein said metabolic enzyme comprises
25 a phosphomannose isomerase.

98. The method of any one of embodiments 87-97, wherein said excision cassette comprises more than one polynucleotide encoding a distinct selectable marker, wherein said polynucleotide encoding a selectable marker is operably linked to a
30 promoter active in a plant cell.

99. The method of embodiment 98, wherein said excision cassette comprises at least a first and a second polynucleotide encoding a selectable marker, wherein said first polynucleotide encodes a yellow fluorescent protein, and wherein said second
5 polynucleotide encodes a phosphinothricin acetyl transferase or a neomycin phosphotransferase II.

100. The method of any one of embodiments 64-99, wherein said herbicide tolerance polypeptide encoded by CP_B confers tolerance to a herbicide selected from the
10 group consisting of glyphosate, an ALS inhibitor, an acetyl Co-A carboxylase inhibitor, a synthetic auxin, a protoporphyrinogen oxidase (PPO) inhibitor herbicide, a pigment synthesis inhibitor herbicide, a phosphinothricin acetyltransferase, a phytoene desaturase inhibitor, a glutamine synthase inhibitor, a hydroxyphenylpyruvatedioxygenase inhibitor, and a protoporphyrinogen oxidase inhibitor.

101. The method of embodiment 100, wherein said ALS inhibitor is selected from the group consisting of a sulfonylurea, a triazolopyrimidine, a pyrimidinyloxy(thio)benzoate, an imidazolinone, and a
15 sulfonylaminocarbonyltriazolinone.

102. The method of any one of embodiments 64-101, wherein said herbicide tolerance polypeptide encoded by CP_B comprises a glyphosate-N-acetyltransferase (GLYAT) polypeptide or an ALS inhibitor-tolerance polypeptide.

103. The method of embodiment 102, wherein said polynucleotide encoding said GLYAT polypeptide has a nucleotide sequence selected from the group consisting of:

- a) the nucleotide sequence set forth in SEQ ID NO: 47 or 49;
- b) a nucleotide sequence having at least 95% sequence identity to

30 SEQ ID NO: 47 or 49;

c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 48 or 50; and

d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 95% sequence identity to SEQ ID NO: 48 or 50.

5

104. The method of embodiment 102, wherein said ALS inhibitor-tolerance polypeptide comprises the highly resistant ALS (HRA) mutation of acetolactate synthase.

105. The method of any one of embodiments 64-104, wherein said
10 polynucleotide construct comprises more than one polynucleotide encoding a distinct herbicide tolerance polypeptide, wherein said polynucleotide encoding a herbicide tolerance polypeptide is operably linked to a promoter active in a plant cell.

106. The method of embodiment 105, wherein said polynucleotide construct
15 comprises at least a first and a second polynucleotide encoding a herbicide tolerance polypeptide, wherein said first polynucleotide encodes an ALS inhibitor-tolerance polypeptide, and wherein said second polynucleotide encodes a GLYAT polypeptide.

107. The method of any one of embodiments 64-106, wherein said excision
20 cassette further comprises a coding polynucleotide D (CP_D), wherein said CP_D encodes a cell proliferation factor, and wherein said CP_D is operably linked to a promoter active in a plant cell.

108. The method of embodiment 107, wherein said cell proliferation factor is
25 selected from the group consisting of a Lec1 polypeptide, a Kn1 polypeptide, a WUSCHEL polypeptide, a Zwiller polypeptide, a babyboom polypeptide, an Aintegumenta polypeptide (ANT), a FUS3 polypeptide, a Kn1 polypeptide, a STM polypeptide, an OSH1 polypeptide, and a SbH1 polypeptide.

109. The method of embodiment 108, wherein said cell proliferation factor is selected from the group consisting of a WUSCHEL polypeptide and a babyboom polypeptide.

5 110. The method of any one of embodiments 107-109, wherein said babyboom polypeptide comprises at least two AP2 domains and at least one of the following amino acid sequences:

a) the amino acid sequence set forth in SEQ ID NO: 67 or an amino acid sequence that differs from the amino acid sequence set forth in SEQ ID NO: 67 by
10 one amino acid; and

b) the amino acid sequence set forth in SEQ ID NO: 68 or an amino acid sequence that differs from the amino acid sequence set forth in SEQ ID NO: 68 by one amino acid.

15 111. The method of any one of embodiments 107-109, wherein said CP_D has a nucleotide sequence selected from the group consisting of:

a) the nucleotide sequence set forth in SEQ ID NO: 55, 57, 58, 60, 74, 76, 78, 80, 82, 84, 86, 87, 88, 90, 92, 94, 96, 98, 99, or 101;

b) a nucleotide sequence having at least 70% sequence identity to
20 SEQ ID NO: 55, 57, 58, 60, 74, 76, 78, 80, 82, 84, 86, 87, 88, 90, 92, 94, 96, 98, 99, or 101;

c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 56, 59, 75, 77, 79, 81, 83, 85, 89, 91, 93, 95, 97, 100, or 102; and

25 d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO: 56, 59, 75, 77, 79, 81, 83, 85, 89, 91, 93, 95, 97, 100, or 102.

112. The method of any one of embodiments 107-111, wherein said excision cassette further comprises a promoter D (P_D), wherein said P_D is operably linked to said CP_D .

5 113. The method of embodiment 112, wherein said P_D is a constitutive promoter.

114. The method of embodiment 112 or 113, wherein said P_D is an ubiquitin promoter or an oleosin promoter.

10

115. The method of any one of embodiments 107-114, wherein said excision cassette comprises more than one polynucleotide encoding a distinct cell proliferation factor, wherein the polynucleotide encoding a cell proliferation factor is operably linked to a promoter active in a plant cell.

15

116. The method of embodiment 115, wherein said excision cassette comprises at least a first coding polynucleotide D (CP_{D1}) encoding a babyboom polypeptide and a second coding polynucleotide D (CP_{D2}) encoding a WUSCHEL polypeptide.

20 117. The method of any one of embodiments 108, 109, and 116, wherein said polynucleotide encoding a WUSCHEL polypeptide has a nucleotide sequence selected from the group consisting of:

a) the nucleotide sequence set forth in SEQ ID NO: 103, 105, 107, or 109; and

25 b) a nucleotide sequence having at least 70% sequence identity to SEQ ID NO: 103, 105, 107, or 109;

c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 104, 106, 108, or 110; and

d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 70% sequence identity to SEQ ID NO: 104, 106, 108, or 110.

5 118. The method of any one of embodiments 108, 109, 116, and 117, wherein said polynucleotide encoding a WUSCHEL polypeptide is operably linked to a maize In2-2 promoter or a nopaline synthase promoter.

10 119. The method of any one of embodiments 64-118, wherein said polynucleotide construct further comprises a coding polynucleotide E (CP_E) encoding a polypeptide of interest, wherein the CP_E is operably linked to a promoter active in a plant cell.

15 120. The method of embodiment 119, wherein said excision cassette comprises said CP_E, and wherein said selected herbicide tolerant plant cell lacks said CP_E.

20 121. The method of embodiment 119, wherein said CP_E is outside of the excision cassette, and wherein said selected herbicide tolerant plant cell comprises said CP_E.

122. The method of any one of embodiments 119-121, wherein said polynucleotide construct further comprises a promoter E (P_E) operably linked to said CP_E.

25 123. The method of embodiment 64, wherein said polynucleotide construct comprises:

- a) a first ubiquitin promoter;
- b) an excision cassette flanked by loxP recombination sites that are recombinogenic with respect to one another and are directly repeated, wherein said excision cassette comprises:

- i) a polynucleotide encoding a phosphinothricin acetyl transferase (PAT) or a neomycin phosphotransferase II (NPTII);
- ii) a second ubiquitin promoter;
- iii) a polynucleotide encoding a yellow fluorescent protein;
- 5 iv) a promoter comprising a maize *rab17* promoter and an attachment B (attB) site;
- v) a polynucleotide encoding a CRE recombinase;
- vi) a nopaline synthase promoter;
- vii) a polynucleotide encoding a maize Wuschel 2 polypeptide;
- 10 viii) a third ubiquitin promoter; and
- ix) a babyboom polynucleotide; and
- c) a GLYAT polynucleotide;

wherein said first ubiquitin promoter is operably linked to said polynucleotide encoding said PAT or NPTII and wherein said first ubiquitin promoter is operably linked to said GLYAT polynucleotide upon excision of said excision cassette;

wherein said second ubiquitin promoter is operably linked to said polynucleotide encoding said yellow fluorescent protein;

wherein said promoter comprising said maize *rab17* promoter and said attB site is operably linked to said polynucleotide encoding said CRE recombinase;

wherein said nopaline synthase promoter is operably linked to said polynucleotide encoding said maize Wuschel 2 polypeptide;

and wherein said third ubiquitin promoter is operably linked to said babyboom polynucleotide.

124. The method of embodiment 64, wherein said polynucleotide construct comprises:

- a) a ubiquitin promoter;
- b) an excision cassette flanked by loxP recombination sites that are recombinationogenic with respect to one another and are directly repeated, wherein said excision cassette comprises:

i) a polynucleotide encoding a *Discosoma* red fluorescent protein;

ii) a promoter comprising a maize *rab17* promoter and an attachment B (attB) site; and

5 iii) a polynucleotide encoding a CRE recombinase; and

c) a GLYAT polynucleotide;

wherein said ubiquitin promoter is operably linked to said polynucleotide encoding said *Discosoma* red fluorescent protein and wherein said ubiquitin promoter is operably linked to said GLYAT polynucleotide upon excision of said excision cassette;

10 and

wherein said promoter comprising said maize *rab17* promoter and said attB site is operably linked to said polynucleotide encoding said CRE recombinase.

125. The method of embodiment 64, wherein said polynucleotide construct
15 comprises:

a) a ubiquitin promoter;

b) an excision cassette flanked by loxP recombination sites that are are recombinogenic with respect to one another and are directly repeated, wherein said excision cassette comprises:

20 i) an actin promoter;

ii) a polynucleotide encoding a *Discosoma* red fluorescent protein;

iii) a promoter comprising a maize *rab17* promoter and an attachment B (attB) site; and

25 iv) a polynucleotide encoding a CRE recombinase; and

c) a GLYAT polynucleotide;

wherein said ubiquitin promoter is operably linked to said GLYAT polynucleotide upon excision of said excision cassette;

wherein said actin promoter is operably linked to said polynucleotide encoding
30 said *Discosoma* red fluorescent protein; and

wherein said promoter comprising said maize *rab17* promoter and said attB site is operably linked to said polynucleotide encoding said CRE recombinase.

126. The method of any one of embodiments 64-125, wherein said plant cells
5 are dicotyledonous.

127. The method of any one of embodiments 64-125, wherein said plant cells are monocotyledonous.

10 128. The method of embodiment 127, wherein said monocotyledonous plant cell is selected from the group consisting of maize, rice, sorghum, barley, wheat, millet, oat, rye, triticale, sugarcane, switchgrass, and turf/forage grass.

129. The method of any one of embodiments 64-128, wherein said plant cells
15 are recalcitrant.

130. The method of embodiment 129, wherein said recalcitrant plant cells are cells of a sugarcane cultivar selected from the group consisting of CP96-1252, CP01-1372, CPCL97-2730, HoCP85-845, CP89-2143, and KQ228.
20

131. A method for increasing the transformation frequency of a plant tissue, the method comprising the steps of:

- a) providing a population of plant cells, wherein at least one plant cell in the population comprises the polynucleotide construct of any one of claims 1-52;
- 25 b) culturing the population of plant cells in the absence of a herbicide to which the herbicide tolerance polypeptide confers herbicide resistance for a period of time sufficient for the population of plant cells to proliferate;
- c) inducing the expression of the site-specific recombinase, thereby excising the excision cassette;

d) contacting the population of plant cells from c) with the herbicide to which the herbicide tolerance polypeptide confers tolerance; and

e) selecting for a plant cell having tolerance to the herbicide, wherein the transformation frequency is increased compared to a comparable plant cell not comprising the excision cassette and selected directly by herbicide selection.

132. The method of embodiment 131, wherein the inducing comprises desiccating the population of plant cells.

133. The method of embodiment 131 or 132, wherein the population of plant cells is cultured in the absence of the herbicide to which the herbicide tolerance polypeptide confers herbicide resistance for about 1 hour to about 6 weeks prior to excision.

BRIEF DESCRIPTION OF THE FIGURES

Figure 1 provides a depiction of vector PHP35648. The vector comprises a coding sequence for the cyan fluorescent protein (CFP), the expression of which is regulated by the ubiquitin promoter (Ubi Pro; comprising the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBIZM INTRON1; SEQ ID NO: 113)). The PHP35648 vector comprises the maize *rab17* promoter with an attachment B site (Rab17 Pro) that drives the expression of the CRE site-specific recombinase. The vector further comprises expression cassettes for the maize Wuschel 2 (WUS2) protein (the expression of which is regulated by the nopaline synthase (Nos) promoter), the maize babyboom (BBM) protein and the maize optimized phosphinothricin acetyl transferase (moPAT) (both of which are regulated by the ubiquitin promoter; comprising the maize ubiquitin promoter (Ubi Pro; comprising the UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBIZM INTRON1; SEQ ID NO: 113)). The yellow fluorescent protein (YFP) is expressed when a fragment of the vector that is

flanked by LoxP recombination sites (the excision cassette) is excised by the CRE recombinase.

Figure 2 provides a depiction of vector PHP54561. The vector comprises a coding sequence for moPAT or neomycin phosphotransferase II (nptII), the expression of which is regulated by the ubiquitin promoter (Ubi Pro; comprising the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBIZM INTRON1; SEQ ID NO: 113)). An ubiquitin promoter (Ubi Pro) also regulates the expression of yellow fluorescent protein (YFP) and the maize BBM protein. The PHP54561 vector further comprises the maize *rab17* promoter with an attachment B site (Rab17 Pro) that drives the expression of the CRE recombinase and an expression cassette for WUS2 under the regulation of the Nos promoter. The ubiquitin promoter (Ubi Pro) regulates the expression of the glyphosate-N-acetyltransferase (GLYAT) gene when an excision cassette flanked by LoxP sites is excised by the CRE recombinase.

Figure 3 provides an image of glyphosate selection on tissue proliferation/regeneration medium of tissues of sugarcane cultivars CP01-1372 (top) and CP88-1762 (bottom) that had been transformed with the PHP54561 vector and desiccated.

Figure 4 provides images of glyphosate selection on regeneration/rooting medium of sugarcane cultivars CP01-1372 (left) and CP88-1762 (right) that had been transformed with the PHP54561 vector and desiccated.

Figure 5 provides images of a second round of glyphosate selection on rooting medium containing 30 μ M glyphosate of sugarcane that had been transformed with the PHP54561 vector and desiccated.

Figure 6 provides a depiction of vector PHP54353. The vector comprises a coding sequence for the red fluorescent protein from *Discosoma* (dsRED), the expression of which is regulated by the ubiquitin promoter (Ubi Pro; comprising the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBIZM INTRON1; SEQ ID NO: 113)). The PHP54353 vector comprises the maize *rab17* promoter with an attachment B site (Rab17

Pro) that drives the expression of the CRE site-specific recombinase. The ubiquitin promoter (Ubi Pro) regulates the expression of the glyphosate-N-acetyltransferase (GLYAT) gene when an excision cassette flanked by LoxP sites is excised by the CRE recombinase.

5 Figure 7 provides a depiction of another polynucleotide construct embodiment. The vector comprises a coding sequence for the red fluorescent protein from *Discosoma* (dsRED), the expression of which is regulated by the actin promoter (Actin Pro). The vector further comprises the maize *rab17* promoter with an attachment B site (Rab17 Pro) that drives the expression of the CRE site-specific recombinase. The ubiquitin promoter
10 (Ubi Pro; comprising the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBIZM INTRON1; SEQ ID NO: 113) regulates the expression of the glyphosate-N-acetyltransferase (GLYAT) gene when an excision cassette flanked by LoxP sites is excised by the CRE recombinase.

15 Figure 8 provides a depiction of vector PHP55062. The vector comprises a coding sequence for the red fluorescent protein from *Discosoma* (dsRED), the expression of which is regulated by the enhanced *Mirabilis* mosaic virus (dMMV) promoter. The vector further comprises the maize *rab17* promoter with an attachment B site (Rab17 Pro) that drives the expression of the CRE site-specific recombinase. A separate dMMV
20 promoter regulates the expression of a hygromycin phosphotransferase (Hyg (hpt)) gene and also regulates the expression of the glyphosate-N-acetyltransferase (GLYAT) gene when an excision cassette flanked by LoxP sites is excised by the CRE recombinase.

Figure 9 provides depictions of various embodiments of the presently disclosed polynucleotide constructs. The constructs all comprise an excision cassette (flanked by
25 LoxP sites) comprising a polynucleotide encoding a site-specific recombinase (CP_A), the expression of which is regulated by an inducible promoter A (P_A). Upon activation of P_A and excision of the excision cassette, promoter B (P_B) is operably linked to the polynucleotide encoding a herbicide tolerance polypeptide (CP_B) and the herbicide tolerance polypeptide is produced. The excision cassette of the constructs of Figures 9b-
30 9g further comprise a polynucleotide encoding a selectable marker (CP_C) in the excision

cassette that is either operably linked to P_B or to another promoter (P_C). The excision cassettes of the constructs of Figures 9d-9g further comprises at least one polynucleotide encoding a cell proliferation factor (CP_{D1} and CP_{D2}), each of which are operably linked to a promoter (P_{D1} or P_{D2} , respectively). The polynucleotide construct of Figure 9g further
5 comprises (outside of the excision cassette) a polynucleotide encoding a polypeptide of interest (CP_E) that is operably linked to a promoter E (P_E).

DETAILED DESCRIPTION OF THE INVENTION

Compositions and methods are provided for regulating the expression of a
10 transgene, such as a herbicide tolerance polynucleotide, for producing and selecting transgenic plants and plant parts, and for increasing the transformation frequency of a plant or plant part. Compositions include polynucleotide constructs comprising an excision cassette, a transgene (e.g., herbicide tolerance polynucleotide) and a promoter that becomes operably linked to the transgene (e.g., herbicide tolerance polynucleotide)
15 upon excision of the excision cassette from the polynucleotide construct. The excision cassette comprises an inducible promoter operably linked to a polynucleotide that encodes a site-specific recombinase and the excision cassette is flanked by a first and a second recombination site, wherein the first and second recombination sites are recombinogenic with respect to one another and are directly repeated, and wherein the
20 site-specific recombinase can recognize and implement recombination at the first and second recombination sites, thereby excising the excision cassette and allowing for the operable linkage of the transgene (e.g., herbicide tolerance polynucleotide) with its promoter. In some embodiments, the polynucleotide construct further comprises a polynucleotide of interest, either within or outside of the excision cassette. In certain
25 embodiments, the excision cassette further comprises at least one coding polynucleotide for a cell proliferation factor, such as a babyboom polypeptide or a Wuschel polypeptide.

In some embodiments, the polynucleotide construct further comprises at least one selectable marker. In some embodiments, the selectable marker is selected from the group consisting of a fluorescent protein, an antibiotic resistance polypeptide, a herbicide
30 tolerance polypeptide, and a metabolic enzyme. In some embodiments, the plant or plant

part is recalcitrant to transformation. In some embodiments, the plant or plant part is a monocotyledonous. In some embodiments the plant or plant part is maize, rice, wheat, barley, sorghum, oats, rye, triticale and sugarcane.

It is intended that the excision cassette is not limited by the number and or order of the coding polynucleotides within the excision cassette. It is envisioned that the excision cassette can be constructed with any number of coding polynucleotides in any order. It is also intended that the polynucleotide construct may also include, beyond the promoter and polynucleotide encoding the herbicide tolerance polypeptide flanking the recombination sites, one or more polynucleotide encoding polypeptide(s) of interest.

The use of the term "polynucleotide" is not intended to limit compositions to polynucleotides comprising DNA. Polynucleotides can comprise ribonucleotides and combinations of ribonucleotides and deoxyribonucleotides. Such deoxyribonucleotides and ribonucleotides include both naturally occurring molecules and synthetic analogues. The polynucleotides also encompass all forms of sequences including, but not limited to, single-, double-, or multi-stranded forms, hairpins, stem-and-loop structures, circular plasmids, and the like.

An "isolated" or "purified" polynucleotide or protein, or biologically active portion thereof, is substantially or essentially free from components that normally accompany or interact with the polynucleotide or protein as found in its naturally occurring environment. Thus, an isolated or purified polynucleotide or protein is substantially free of other cellular material, or culture medium when produced by recombinant techniques, or substantially free of chemical precursors or other chemicals when chemically synthesized. Optimally, an "isolated" polynucleotide is free of sequences (optimally protein encoding sequences) that naturally flank the polynucleotide (*i.e.*, sequences located at the 5' and 3' ends of the polynucleotide) in the genomic DNA of the organism from which the polynucleotide is derived. For example, in various embodiments, the isolated polynucleotide can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb, or 0.1 kb of nucleotide sequence that naturally flank the polynucleotide in genomic DNA of the cell from which the polynucleotide is derived. A protein that is substantially free of cellular material includes preparations of protein having less than

about 30%, 20%, 10%, 5%, or 1% (by dry weight) of contaminating protein. When the protein or biologically active portion thereof is recombinantly produced, optimally culture medium represents less than about 30%, 20%, 10%, 5%, or 1% (by dry weight) of chemical precursors or non-protein-of-interest chemicals.

5 As used herein, a “polynucleotide construct” refers to a polynucleotide molecule comprised of various types of nucleotide sequences having different functions and/or activities. For example, a polynucleotide construct may comprise one or more of any of the following: expression cassettes, coding polynucleotides, regulatory sequences (e.g., enhancers, promoters, termination sequences), origins of replication, restriction sites,
10 recombination sites, and excision cassettes.

 The presently disclosed polynucleotide constructs can comprise one or more expression cassettes, wherein a coding polynucleotide is operably linked to a regulatory sequence.

 As used herein, a “coding polynucleotide” refers to a polynucleotide that encodes
15 a polypeptide and therefore comprises the requisite information to direct translation of the nucleotide sequence into a specified polypeptide. Alternatively, a “coding polynucleotide” can refer to a polynucleotide that encodes a silencing polynucleotide that reduces the expression of target genes. Non-limiting examples of a silencing polynucleotide include a small interfering RNA, micro RNA, antisense RNA, a hairpin
20 structure, and the like.

 As used herein, an “expression cassette” refers to a polynucleotide that comprises at least one coding polynucleotide operably linked to regulatory sequences sufficient for the expression of the coding polynucleotide. “Operably linked” is intended to mean a functional linkage between two or more elements. For example, an operable linkage
25 between a coding polynucleotide and a regulatory sequence (i.e., a promoter) is a functional link that allows for expression of the coding polynucleotide. Operably linked elements may be contiguous or non-contiguous. When used to refer to the joining of two protein coding regions, by operably linked is intended that the coding regions are in the same reading frame.

An expression cassette will include in the 5'-3' direction of transcription, a transcriptional and translational initiation region (i.e., a promoter), a coding polynucleotide, and a transcriptional and translational termination region (i.e., termination region) functional in plants. The regulatory regions (i.e., promoters, transcriptional regulatory regions, and translational termination regions) and/or the coding polynucleotide may be native/analogous to a host cell comprising the presently disclosed polynucleotide constructs or to each other. Alternatively, the regulatory regions and/or the coding polynucleotide may be heterologous to the host cell or to each other. As used herein, "heterologous" in reference to a sequence is a sequence that originates from a foreign species, or, if from the same species, is substantially modified from its native form in composition and/or genomic locus by deliberate human intervention. A heterologous polynucleotide is also referred to herein as a "transgene". For example, a promoter operably linked to a heterologous polynucleotide is from a species different from the species from which the polynucleotide was derived, or, if from the same/analogous species, one or both are substantially modified from their original form and/or genomic locus, or the promoter is not the native promoter for the operably linked polynucleotide. While it may be optimal to express the sequences using heterologous promoters, the native promoter sequences may be used.

The termination region may be native with the transcriptional initiation region, may be native with the operably linked coding polynucleotide, may be native with the host cell, or may be derived from another source (i.e., foreign or heterologous) to the promoter, the coding polynucleotide, the host cell, or any combination thereof. Convenient termination regions are available from the potato proteinase inhibitor (PinII) gene or the Ti-plasmid of *A. tumefaciens*, such as the octopine synthase and nopaline synthase termination regions. See also Guerineau *et al.* (1991) *Mol. Gen. Genet.* 262:141-144; Proudfoot (1991) *Cell* 64:671-674; Sanfacon *et al.* (1991) *Genes Dev.* 5:141-149; Mogen *et al.* (1990) *Plant Cell* 2:1261-1272; Munroe *et al.* (1990) *Gene* 91:151-158; Ballas *et al.* (1989) *Nucleic Acids Res.* 17:7891-7903; and Joshi *et al.* (1987) *Nucleic Acid Res.* 15:9627-9639. In some embodiments, the termination sequence that is operably linked to at least one of the site-specific recombinase-encoding polynucleotide,

the selectable marker-encoding polynucleotide, the cell proliferation marker-encoding polynucleotide, the herbicide tolerance polynucleotide, and the polynucleotide of interest is the termination region from the pinII gene. In some of these embodiments, the termination region has the sequence set forth in SEQ ID NO: 1 or an active variant or
5 fragment thereof that is capable of terminating transcription and/or translation in a plant cell.

The expression cassettes may additionally contain 5' leader sequences. Such leader sequences can act to enhance translation. Translation leaders are known in the art and include: picornavirus leaders, for example, EMCV leader (encephalomyocarditis 5' noncoding region) (Elroy-Stein *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86:6126-6130);
10 potyvirus leaders, for example, TEV leader (tobacco etch virus) (Gallie *et al.* (1995) *Gene* 165(2):233-238), MDMV leader (maize dwarf mosaic virus) (*Virology* 154:9-20), and human immunoglobulin heavy-chain binding protein (BiP) (Macejak *et al.* (1991) *Nature* 353:90-94); untranslated leader from the coat protein mRNA of alfalfa mosaic virus
15 (AMV RNA 4) (Jobling *et al.* (1987) *Nature* 325:622-625); tobacco mosaic virus leader (TMV) (Gallie *et al.* (1989) in *Molecular Biology of RNA*, ed. Cech (Liss, New York), pp. 237-256); and maize chlorotic mottle virus leader (MCMV) (Lommel *et al.* (1991) *Virology* 81:382-385). See also, Della-Cioppa *et al.* (1987) *Plant Physiol.* 84:965-968.

For example, in some of the embodiments, wherein the herbicide tolerance
20 polynucleotide is a GLYAT polynucleotide, the cauliflower mosaic virus (CaMV) 35S enhancer region or tobacco mosaic virus (TMV) omega 5' UTR translational enhancer element is included upstream of a promoter that is operably linked (when the excision cassette is excised) to the GLYAT polynucleotide to enhance transcription (see, for example, U.S. Patent Nos. 7,928,296 and 7,622,641, each of which is herein incorporated
25 by reference in its entirety).

In preparing the expression cassette or polynucleotide construct, the various DNA fragments may be manipulated, so as to provide for the DNA sequences in the proper orientation and, as appropriate, in the proper reading frame. Toward this end, adapters or linkers may be employed to join the DNA fragments or other manipulations may be
30 involved to provide for convenient restriction sites, removal of superfluous DNA,

removal of restriction sites, or the like. For this purpose, *in vitro* mutagenesis, primer repair, restriction, annealing, resubstitutions, e.g., transitions and transversions, may be involved.

Expression cassettes comprise a promoter operably linked to a coding
5 polynucleotide. As used herein, the term "promoter" includes reference to a region of DNA involved in the recognition and binding of RNA polymerase and other proteins to initiate transcription of a coding sequence. Promoters may be naturally occurring promoters, a variant or fragment thereof, or synthetically derived. The term "promoter" refers to the minimal sequences necessary to direct transcription (minimal promoter) as
10 well as sequences comprising the minimal promoter and any number of additional elements, such as operator sequences, enhances, modulators, restriction sites, recombination sites, sequences located in between the minimal promoter and the coding sequence, and sequences of the 5'-untranslated region (5'-UTR), which is the region of a transcript that is transcribed, but is not translated into a polypeptide, which may or may
15 not influence transcription levels in a desired manner. A "plant promoter" refers to a promoter isolated from a plant or a promoter derived therefrom or a heterologous promoter that functions in a plant.

Although according to the invention, the promoter that drives the expression of the site-specific recombinase is an inducible promoter, various types of promoters can be
20 used for the regulation of the expression of the remaining coding polynucleotides in the presently disclosed polynucleotide constructs. The promoter may be selected based on the desired outcome or expression pattern (for a review of plant promoters, see Potenza *et al.* (2004) *In Vitro Cell Dev Biol* 40:1-22).

Constitutive promoters include, for example, the core promoter of the Rsyn7
25 promoter and other constitutive promoters disclosed in WO 99/43838 and U.S. Patent No. 6,072,050; the core CaMV 35S promoter (Odell *et al.* (1985) *Nature* 313:810-812); rice actin (McElroy *et al.* (1990) *Plant Cell* 2:163-171); ubiquitin (Christensen *et al.* (1989) *Plant Mol. Biol.* 12:619-632 and Christensen *et al.* (1992) *Plant Mol. Biol.* 18:675-689); pEMU (Last *et al.* (1991) *Theor. Appl. Genet.* 81:581-588); MAS (Velten *et al.* (1984) *EMBO J.* 3:2723-2730); ALS promoter (U.S. Patent No. 5,659,026), the
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Agrobacterium nopaline synthase (NOS) promoter (Bevan *et al.* (1983) *Nucl. Acids Res.* 11:369-385); *Mirabilis* mosaic virus (MMV) promoter (Dey & Maiti (1999) *Plant Mol Biol* 40:771-782; Dey & Maiti (1999) *Transgenics* 3:61-70); histone 2B (H2B) (International Application Publication No. WO 99/43797); banana streak virus (BSV) promoter (Remans *et al.* (2005) *Virus Research* 108:177-186); chloris striate mosaic virus (CSMV) promoter (Zhan *et al.* (1993) *Virology* 193:498-502); Cassava vein mosaic virus (CSVMV) promoter (Verdaguer *et al.* (1998) *Plant Mol Biol* 37:1055-1067); figwort mosaic virus (FMV) promoter (U.S. Patent No. 6,018,100), rice alpha-tubulin (OsTUBA1) promoter (Jeon *et al.* (2000) *Plant Physiol* 123:1005-1014); rice cytochrome C (OsCC1) promoter (Jang *et al.* (2002) *Plant Physiol* 129:1473-1481); maize alcohol dehydrogenase1 (ZmADH1) promoter (Kyoizuka *et al.* (1990) *Maydica* 35:353-357; an oleosin promoter (*e.g.*, SEQ ID NO: 2 or a variant or fragment thereof) and the like; each of which is herein incorporated by reference in its entirety. Other constitutive promoters are described in, for example, U.S. Patent Nos. 5,608,149; 5,608,144; 5,604,121; 5,569,597; 5,466,785; 5,399,680; 5,268,463; 5,608,142; and 6,177,611; each of which is herein incorporated by reference in its entirety.

In some embodiments, an inducible promoter can be used, such as from a pathogen-inducible promoter. Such promoters include those from pathogenesis-related proteins (PR proteins), which are induced following infection by a pathogen; *e.g.*, PR proteins, SAR proteins, beta-1,3-glucanase, chitinase, etc. See, for example, Redolfi *et al.* (1983) *Neth. J. Plant Pathol.* 89:245-254; Uknes *et al.* (1992) *Plant Cell* 4:645-656; and Van Loon (1985) *Plant Mol. Virol.* 4:111-116. See also WO 99/43819, herein incorporated by reference. Promoters that are expressed locally at or near the site of pathogen infection include, for example, Marineau *et al.* (1987) *Plant Mol. Biol.* 9:335-342; Matton *et al.* (1989) *Mol Plant-Microbe Interact* 2:325-331; Somsisch *et al.* (1986) *Proc. Natl. Acad. Sci. USA* 83:2427-2430; Somsisch *et al.* (1988) *Mol. Gen. Genet.* 2:93-98; and Yang (1996) *Proc. Natl. Acad. Sci. USA* 93:14972-14977. See also, Chen *et al.* (1996) *Plant J.* 10:955-966; Zhang *et al.* (1994) *Proc. Natl. Acad. Sci. USA* 91:2507-2511; Warner *et al.* (1993) *Plant J.* 3:191-201; Siebertz *et al.* (1989) *Plant Cell* 1:961-968; U.S. Patent No. 5,750,386 (nematode-inducible); and the references cited therein.

Additional promoters include the inducible promoter for the maize PRms gene, whose expression is induced by the pathogen *Fusarium moniliforme* (see, for example, Cordero *et al.* (1992) *Physiol. Mol. Plant Path.* 41:189-200). Wound-inducible promoters include potato proteinase inhibitor (pin II) gene (Ryan (1990) *Ann. Rev. Phytopath.* 28:425-449; Duan *et al.* (1996) *Nat Biotechnol* 14:494-498); *wun1* and *wun2*, U.S. Patent No. 5,428,148; *win1* and *win2* (Stanford *et al.* (1989) *Mol. Gen. Genet.* 215:200-208); systemin (McGurl *et al.* (1992) *Science* 225:1570-1573); WIP1 (Rohmeier *et al.* (1993) *Plant Mol. Biol.* 22:783-792; Eckelkamp *et al.* (1993) *FEBS Lett* 323:73-76); MPI gene (Corderok *et al.* (1994) *Plant J.* 6:141-150); and the like, herein incorporated by reference.

Other inducible promoters useful for regulating the expression of any of the coding sequences of the presently disclosed polynucleotide constructs include stress-inducible promoters, such as those described elsewhere herein.

Chemical-regulated promoters can be used to modulate the expression of a gene in a plant through the application of an exogenous chemical regulator. The promoter may be a chemical-inducible promoter, where application of the chemical induces gene expression, or a chemical-repressible promoter, where application of the chemical represses gene expression. Chemical-inducible promoters are known in the art and include, but are not limited to, the maize In2-2 promoter, which is activated by benzenesulfonamide herbicide safeners (De Veylder *et al.* (1997) *Plant Cell Physiol.* 38:568-77), the maize GST promoter (GST-II-27, WO 93/01294), which is activated by hydrophobic electrophilic compounds that are used as pre-emergent herbicides, the PR-1 promoter (Cao *et al.* (2006) *Plant Cell Reports* 6:554-60), which is activated by BTH or benzo(1,2,3)thiadiazole-7-carbothioic acid s-methyl ester, the tobacco PR-1a promoter (Ono *et al.* (2004) *Biosci. Biotechnol. Biochem.* 68:803-7), which is activated by salicylic acid, the copper inducible ACE1 promoter (Mett *et al.* (1993) *PNAS* 90:4567-4571), the ethanol-inducible promoter AlcA (Caddick *et al.* (1988) *Nature Biotechnol* 16:177-80), an estradiol-inducible promoter (Bruce *et al.* (2000) *Plant Cell* 12:65-79), the XVE estradiol-inducible promoter (Zao *et al.* (2000) *Plant J* 24:265-273), the VGE methoxyfenozide inducible promoter (Padidam *et al.* (2003) *Transgenic Res* 12:101-109),

and the TGV dexamethasone-inducible promoter (Bohner *et al.* (1999) *Plant J* 19:87-95). Other chemical-regulated promoters of interest include steroid-responsive promoters (see, for example, the glucocorticoid-inducible promoter in Schena *et al.* (1991) *Proc. Natl. Acad. Sci. USA* 88:10421-10425 and McNellis *et al.* (1998) *Plant J.* 14(2):247-257) and
5 tetracycline-inducible and tetracycline-repressible promoters (see, for example, Gatz *et al.* (1991) *Mol. Gen. Genet.* 227:229-237; Gatz *et al.* (1992) *Plant J* 2:397-404; and U.S. Patent Nos. 5,814,618 and 5,789,156), herein incorporated by reference.

One particular chemical-inducible promoter that is described in more detail elsewhere herein and that can be used in the presently disclosed compositions and
10 methods, particularly to regulate the expression of the site-specific recombinase, is a promoter responsive to sulfonylurea, wherein the promoter comprises operator sequences capable of binding to a sulfonylurea-responsive transcriptional repressor (SuR) protein, such as those described in U.S. Application Publication Nos. 2010/0105141 and 2011/0287936, each of which is herein incorporated by reference in its entirety.

15 Tissue-preferred promoters can be utilized to target enhanced expression of a coding polynucleotide within a particular plant tissue. Tissue-preferred promoters include Kawamata *et al.* (1997) *Plant Cell Physiol.* 38(7):792-803; Hansen *et al.* (1997) *Mol. Gen. Genet.* 254(3):337-343; Russell *et al.* (1997) *Transgenic Res.* 6(2):157-168; Rinehart *et al.* (1996) *Plant Physiol.* 112(3):1331-1341; Van Camp *et al.* (1996) *Plant*
20 *Physiol.* 112(2):525-535; Canevascini *et al.* (1996) *Plant Physiol.* 112(2):513-524; Lam (1994) *Results Probl. Cell Differ.* 20:181-196; and Guevara-Garcia *et al.* (1993) *Plant J.* 4(3):495-505.

Leaf-preferred promoters are known in the art. See, for example, Yamamoto *et al.* (1997) *Plant J.* 12:255-265; Kwon *et al.* (1994) *Plant Physiol.* 105:357-67; Yamamoto *et al.* (1994) *Plant Cell Physiol.* 35:773-778; Gotor *et al.* (1993) *Plant J.* 3:509-18; Orozco
25 *et al.* (1993) *Plant Mol. Biol.* 23:1129-1138; and Matsuoka *et al.* (1993) *Proc. Natl. Acad. Sci. USA* 90:9586-9590. In addition, promoter of cab and rubisco can also be used. See, for example, Simpson *et al.* (1958) *EMBO J* 4:2723-2729 and Timko *et al.* (1988) *Nature* 318:57-58.

Root-preferred promoters are known and can be selected from the many available. See, for example, Hire *et al.* (1992) *Plant Mol. Biol.* 20:207-218 (soybean root-specific glutamine synthase gene); Keller and Baumgartner (1991) *Plant Cell* 3:1051-1061 (root-specific control element in the GRP 1.8 gene of French bean); Sanger *et al.* (1990) *Plant Mol. Biol.* 14:433-443 (root-specific promoter of the mannopine synthase (MAS) gene of *Agrobacterium tumefaciens*); and Miao *et al.* (1991) *Plant Cell* 3:11-22 (full-length cDNA clone encoding cytosolic glutamine synthase (GS), which is expressed in roots and root nodules of soybean). See also Bogusz *et al.* (1990) *Plant Cell* 2:633-641, where two root-specific promoters isolated from hemoglobin genes from the nitrogen-fixing nonlegume *Parasponia andersonii* and the related non-nitrogen-fixing nonlegume *Trema tomentosa* are described. Leach and Aoyagi (1991) describe their analysis of the promoters of the highly expressed rolC and rolD root-inducing genes of *Agrobacterium rhizogenes* (see *Plant Sci* (Limerick) 79:69-76). Teeri *et al.* (1989) used gene fusion to lacZ to show that the *Agrobacterium* T-DNA gene encoding octopine synthase is especially active in the epidermis of the root tip and that the TR2' gene is root specific in the intact plant and stimulated by wounding in leaf tissue (see *EMBO J.* 8:343-350). The TR1' gene, fused to *nptII* (neomycin phosphotransferase II) showed similar characteristics. Additional root-preferred promoters include the VfENOD-GRP3 gene promoter (Kuster *et al.* (1995) *Plant Mol. Biol.* 29:759-772); and rolB promoter (Capana *et al.* (1994) *Plant Mol. Biol.* 25:681-691. See also U.S. Patent Nos. 5,837,876; 5,750,386; 5,633,363; 5,459,252; 5,401,836; 5,110,732; and 5,023,179. Another root-preferred promoter includes the promoter of the phaseolin gene (Murai *et al.* (1983) *Science* 23:476-482 and Sengopta-Gopalen *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 82:3320-3324.

Seed-preferred promoters include both those promoters active during seed development as well as promoters active during seed germination. See Thompson *et al.* (1989) *BioEssays* 10:108, herein incorporated by reference. Such seed-preferred promoters include, but are not limited to, Cim1 (cytokinin-induced message); cZ19B1 (maize 19 kDa zein); and milps (myo-inositol-1-phosphate synthase); (see WO 00/11177 and U.S. Patent No. 6,225,529; herein incorporated by reference). For dicots, seed-

preferred promoters include, but are not limited to, bean β -phaseolin, napin, β -conglycinin, soybean lectin, cruciferin, and the like. For monocots, seed-preferred promoters include, but are not limited to, maize 15 kDa zein, 22 kDa zein, 27 kDa gamma zein, waxy, shrunken 1, shrunken 2, globulin 1, oleosin, nuc1, etc. See also WO 00/12733, where seed-preferred promoters from *end1* and *end2* genes are disclosed; herein incorporated by reference.

Where low-level expression is desired, weak promoters will be used. Generally, by "weak promoter" is intended a promoter that drives expression of a coding sequence at a low level. By low level is intended at levels of about 1/1000 transcripts to about 1/100,000 transcripts to about 1/500,000 transcripts. Alternatively, it is recognized that weak promoters also encompasses promoters that are expressed in only a few cells and not in others to give a total low level of expression. Where a promoter is expressed at unacceptably high levels, portions of the promoter sequence can be deleted or modified to decrease expression levels. Such weak constitutive promoters include, for example, the core promoter of the Rsyn7 promoter (WO 99/43838 and U.S. Patent No. 6,072,050), the core 35S CaMV promoter, and the like.

In some embodiments, at least one of the following promoters is a constitutive promoter: the promoter regulating the expression of the herbicide tolerance polypeptide, the promoter operably linked to the cell proliferation marker, and the promoter driving the expression of the selectable marker present within the excision cassette. In particular embodiments, the selectable marker present within the excision cassette of the presently disclosed polynucleotide constructs is operably linked to a constitutive promoter such that the selectable marker is constitutively expressed until excision of the excision cassette, and the same constitutive promoter then regulates the expression of the herbicide tolerance polypeptide upon excision of the cassette. In some of these embodiments, the constitutive promoter is the maize ubiquitin promoter (Christensen *et al.* (1989) *Plant Mol. Biol.* 12:619-632 and Christensen *et al.* (1992) *Plant Mol. Biol.* 18:675-689), which in some embodiments comprises the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBIZM INTRON1; SEQ ID NO: 113). In other embodiments, the constitutive

promoter regulating the expression of the selectable marker present within the excision cassette is the enhanced *Mirabilis* mosaic virus (MMV) promoter (Dey & Maiti (1999) Plant Mol Biol 40:771-782; Dey & Maiti (1999) Transgenics 3:61-70). In some embodiments, the polynucleotide encoding a cell proliferation factor (e.g., babyboom polypeptide) is operably linked to a maize ubiquitin promoter (which in some embodiments comprises the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBI1ZM INTRON1; SEQ ID NO: 113) or a maize oleosin promoter (e.g., SEQ ID NO: 2 or a variant or fragment thereof).

According to the invention, the promoter that regulates the expression of the site-specific recombinase is an inducible promoter. In some embodiments, the inducible promoter that is operably linked to the site-specific recombinase-encoding polynucleotide comprises a stress-inducible promoter. As used herein, a “stress-inducible promoter” refers to a promoter that initiates transcription when the host cell (e.g., plant cell) or host (e.g., plant or plant part) undergoes stress, including abiotic stress. Non-limiting examples of conditions that can activate stress-inducible promoters include drought, salinity, flood, and suboptimal temperature. Some stress-inducible promoters are only activated by a particular stress (e.g., drought), whereas other stress-inducible promoters can be activated by any type of stress, particularly any type of abiotic stress.

Stress-inducible promoters include those that become activated in response to drought and high salinity (drought-inducible promoters) and cold temperatures (cold-inducible promoters). Some promoters are both drought-inducible and cold-inducible. Many stress-inducible promoters are also activated by abscisic acid (ABA), a phytohormone that is often expressed by plants in response to drought and high-salinity stress. Regulatory pathways by which stress-inducible promoters can become activated include those that are ABA-dependent as well as those that are ABA-independent. Thus, some stress-inducible promoters comprise an ABA-responsive element (ABRE) and respond to ABA. Some of those stress-inducible promoters that are responsive to drought, high salinity, and/or cold temperatures comprise a dehydration-responsive (DRE)/C-repeat (CRT) element. The C-repeat binding factor (CBF)/DREB1

transcription factor, the expression of which is induced by cold stress, and the DREB2 transcription factor, which is induced by dehydration, bind to DRE/CRT elements. In some embodiments, stress-inducible promoters comprise any one of the following *cis*-acting stress-responsive elements: ABRE, CE1, CE3, MYB recognition site (MYBR), MYC recognition site (MYCR), DRE, CRT, low-temperature-responsive element (LTRE), NAC recognition site (NACR), zinc-finger homeodomain recognition site (ZFHDR) and an inducer of CBF expression (ICE) recognition site. Table 1 provides the sequences of these *cis*-acting stress-responsive elements. See Yamaguchi-Shinozaki and Shinozaki (2005) *Trends Plant Sci* 10:1360-1385 and Shinozaki *et al.* (2003) *Curr Opin Plant Biol* 6:410-417, each of which is incorporated by reference in its entirety, for reviews of stress-inducible promoters and the regulatory pathways controlling the same.

Table 1. *cis*-Acting regulatory elements in stress-inducible gene expression.*

<i>cis</i> element	Sequence (SEQ ID NO:)	Type of transcription factors that bind to <i>cis</i> elements	Gene	Stress condition
ABRE	PyACGTGGC (3)	bZIP	Em, RAB16	Water deficit, ABA
CE1	TGCCACCGG (4)	ERF/AP2	HVA1	ABA
CE3	ACGCGTGCCTC (5)	Not known	HVA22	ABA
ABRE	ACGTGTC (6)	bZIP	Osem	ABA
ABRE	ACGTGGC (7), ACGTGTC (8)	bZIP	RD29B	Water deficit, ABA
MYBR	TGGTTAG (9)	MYB	RD22	Water deficit, ABA
MYCR	CACATG (10)	bHLH	RD22	Water deficit, ABA
DRE	TACCGACAT (11)	ERF/AP2	RD29A	Water deficit, cold
CRT	GGCCGACAT (12)	ERF/AP2	Cor15A	Cold
LTRE	GGCCGACGT (13)	ERF/AP2	BN115	Cold
NACR	ACACGCATGT (14)	NAC	ERD1	Water deficit
ZFHDR	Not yet reported	ZFHD	ERD1	Water deficit
ICEr1	GGACACATGTCAGA	Not known	CBF2/	Cold

<i>cis</i> element	Sequence (SEQ ID NO:)	Type of transcription factors that bind to <i>cis</i> elements	Gene	Stress condition
	(15)		DREB1C	
ICEr2	ACTCCG (16)	Not known	CBF2/ DREB1C	Cold

*Adopted from Yamaguchi-Shinozaki and Shinozaki (2005) *Trends Plant Sci* 10:1360-1385

In some embodiments, the inducible promoter that is operably linked to the
5 polynucleotide encoding a site-specific recombinase is a cold-inducible promoter. As
used herein, a “cold-inducible promoter” is a promoter that is activated at temperatures
that are below optimal temperatures for plant growth. In some embodiments, the cold-
inducible promoter is one that is induced in response to temperatures less than about
20°C, less than about 19°C, less than about 18°C, less than about 17°C, less than about
10 16°C, less than about 15°C, less than about 14°C, less than about 13°C, less than about
12°C, less than about 11°C, less than about 10°C, less than about 9°C, less than about
8°C, less than about 7°C, less than about 6°C, less than about 5°C, less than about 4°C,
less than about 3°C, less than about 2°C, less than about 1°C, or less than about 0°C.

Cold-inducible promoters may be activated by exposing a plant or plant part to
15 cold temperatures for a period of about 12 hours, about 1 day, about 2 days, about 3 days,
about 4 days, about 5 days, about 6 days, about 1 week, about 2 weeks, about 3 weeks,
about 4 weeks, about 5 weeks, about 6 weeks, about 8 weeks, about 9 weeks, about 10
weeks, about 3 months, or more. The temperature required or the necessary amount of
time the plant or plant part is exposed to the cold temperatures will vary based on, for
20 example, the promoter, the plant species, the type of explant, and the size of the plant
tissue, and can be determined by one of skill in the art.

Cold-inducible promoters can comprise a C-repeat (CRT) and/or a low-
temperature-responsive element (LTRE), both of which contain an A/GCCGAC motif
that forms the core of the DRE sequence, as well. Non-limiting examples of cold-

inducible promoters include the maize *rab17* promoter (Vilardell *et al.* (1990) *Plant Mol Biol* 14:423-432), the *RD29A* promoter (Uno *et al.* (2000) *PNAS* 97:11632-11637), the *Cor15A* promoter (Baker *et al.* (1994) *Plant Mol Biol* 24:701-713), the *BN115* promoter (Jiang *et al.* (1996) *Plant Mol Biol* 30:679-684), and the *CBF2/DREB1C* promoter (Zarka
5 *et al.* (2003) *Plant Physiol* 133:910-918); each of which is herein incorporated by reference in its entirety.

In some embodiments, the inducible promoter that regulates the expression of the site-specific recombinase is a vernalization promoter, which is a promoter that responds to cold exposure to trigger flowering in plants. Vernalization promoters generally require
10 exposure to cold temperatures for an extended period of time (e.g., at least 2 weeks) for activation. In certain embodiments, activation of a vernalization promoter requires exposure to temperatures less than about 20°C, less than about 19°C, less than about 18°C, less than about 17°C, less than about 16°C, less than about 15°C, less than about 14°C, less than about 13°C, less than about 12°C, less than about 11°C, less than about
15 10°C, less than about 9°C, less than about 8°C, less than about 7°C, less than about 6°C, less than about 5°C, less than about 4°C, less than about 3°C, less than about 2°C, less than about 1°C, or less than about 0°C for at least 2 weeks, at least 3 weeks, at least 4 weeks, at least 5 weeks, at least 6 weeks, at least 7 weeks, at least 8 weeks, at least 9 weeks, at least 10 weeks, at least 11 weeks, at least 12 weeks, at least 13 weeks, at least
20 14 weeks, at least 15 weeks, at least 16 weeks, or more. In certain embodiments, activation of a vernalization promoter requires exposure to a temperature of about 4°C for about 2 weeks.

In some embodiments, the vernalization promoter comprises a putative MADS-box protein binding site, referred to herein as CarG-box, the sequence of which is set
25 forth in SEQ ID NO: 114. A non-limiting example of a vernalization promoter is the *Triticum monococcum* VRN1/AP1 promoter set forth in SEQ ID NO: 115 and described in Yan *et al.* (2003) *Proc Natl Acad Sci USA* 100:6263-6268 and U.S. Application Publication No. 2004/0203141, each of which is herein incorporated by reference in its entirety.

In some of those embodiments wherein the inducible promoter that regulates the expression of the site-specific recombinase is a vernalization promoter, the host cell of the polynucleotide construct is a *Brassica* sp., winter wheat, barley, oat, or rye.

In other embodiments, the inducible promoter that regulates the expression of the site-specific recombinase is a drought-inducible promoter. As used herein, a “drought-inducible promoter” or “desiccation-inducible promoter” refers to a promoter that initiates transcription in response to drought conditions, high salinity, and/or dessication of a plant or plant part. Drought-inducible promoters can drive expression in a number of different plant tissues including, but not limited to, root tissue (e.g., root endodermis, root epidermis, or root vascular tissues) and leaf tissue (e.g. epidermis, mesophyll or leaf vascular tissue).

In some embodiments, the drought-inducible promoter comprises a DRE or an early responsive to dehydration 1 (ERD1) cis-acting element (Yamaguchi-Shinozaki and Shinozaki (2004) *Trends Plant Sci* 10:1360-1385; and Shinozaki *et al.* (2003) *Curr Opin Plant Biol* 6:410-417).

The drought-inducible promoter is activated when the plant or plant part comprising the same is desiccated. As used herein, the term “desiccate” refers to a process by which the water content of a plant or plant part is reduced, and can include reference to the natural desiccation process that occurs during the maturation of seeds. Thus, in some embodiments, the drought-inducible promoter is activated in a plant cell comprising the presently disclosed polynucleotide constructs and excision of the excision cassette occurs during the maturation of a seed comprising the plant cell.

A desiccated plant or plant part can comprise about 90%, 85%, 80%, 75%, 70%, 65%, 60%, 55%, 50%, 45%, 40%, 35%, 30%, 25%, 20%, 15%, 10%, 5%, 1%, 0.1% or less water than a plant or plant part that has not been dried. The amount of desiccation necessary to activate a drought-inducible promoter or the amount of time needed to desiccate a plant or plant part will vary based on, for example, the promoter, the plant species, the explant type, and the size of the plant tissue.

In some embodiments, a plant or plant part is desiccated and the drought-inducible promoter is activated by exposing the plant or plant part comprising the

drought-inducible promoter to drought conditions. As used herein, “drought” or “drought conditions” can be defined as the set of environmental conditions under which a plant or plant part will begin to suffer the effects of water deprivation, such as decreased stomatal conductance and photosynthesis, decreased growth rate, loss of turgor (wilting), or ovule abortion. For these reasons, plants experiencing drought stress typically exhibit a significant reduction in biomass and yield. Water deprivation may be caused by lack of rainfall or limited irrigation. Alternatively, water deficit may also be caused by high temperatures, low humidity, saline soils, freezing temperatures or water-logged soils that damage roots and limit water uptake to the shoot. Since plant species vary in their capacity to tolerate water deficit, the precise environmental conditions that cause drought stress cannot be generalized.

The drought-inducible promoter may be activated by exposing a plant or plant part to drought conditions for a period of about 1 day, about 2 days, about 3 days, about 4 days, about 5 days, about 6 days, about 1 week, about 2 weeks, about 3 weeks, or more.

In some embodiments, the plant or plant part is desiccated and the drought-inducible promoter activated by incubating the plant or plant part in the absence of liquid medium and optionally on dry filter paper. In some embodiments, the plant or plant part is desiccated by incubating the plant or plant part in a sealed container with a saturated salt solution (e.g., $(\text{NH}_4)_2\text{SO}_4$). In some embodiments, the plant or plant part is incubated in the absence of liquid medium, and optionally, on dry filter paper, and in some embodiments, in a sealed container with a saturated salt solution for about 1 day, about 1.5 days, about 2 days, about 2.5 days, about 3 days, about 3.5 days, about 4 days, about 4.5 days, about 5 days, about 5.5 days, about 6 days, about 6.5 days, about 7 days, about 7.5 days, about 8 days, about 8.5 days, about 9 days, about 9.5 days, about 10 days, or more in order to induce the expression of the drought-inducible promoter.

Non-limiting examples of drought-inducible promoters include the promoters of maize *rab17* (Vilardell *et al.* (1990) *Plant Mol Biol* 14:423-432); *Oryza sativa* *Em* (Guiltinan *et al.* (1990) *Science* 250:267-271); *Rab16* (Mundy *et al.* (1990) *PNAS* 87:406-410); *HVA1* (Hobo *et al.* (1999) *Plant J* 19:679-689); *HVA22* (Su *et al.* (1998) *Plant Physiol* 117:913-922); *RD29B* and *RD29A* (Uno *et al.* (2000) *PNAS* 97:11632-11637);

RD22 (Abe *et al.* (1997) *Plant Cell* 9:1859-1868); *Cor15A* (Baker *et al.* (1994) *Plant Mol Biol* 24:701-713); *BN115* (Jiang *et al.* (1996) *Plant Mol Biol* 30:679-684); *ERD1* (Tran *et al.* (2004) *Plant Cell* 16:2481-2498); *Oryza sativa LEA3* (Xiao *et al.* (2007) *Theor Appl Genet* 115:35-46); *Oryza sativa rab16Bj* (Xiao and Xue (2001) *Plant Cell Rep* 20:667-73); *Brassica LEA3-1* (U.S. Application Publication No. US 2008/0244793); *LEA D7*, *LEA D11*, *LEA D19*, *LEA d34*, and *LEA D113* (Baker *et al.* (1988) *Plant Mol Biol* 11:277-291); *Oryza sativa RAB16* and *Sorghum bicolor DHN2* (Buchanan *et al.* (2004) *Genetics* 168:1639-1654); *Oryza sativa ASR1* (Kuriakose *et al.* (2009) *African J Biotech* 8:4765-73); *Oryza sativa NAC6* (Nakashima *et al.* (2007) *Plant J* 51:617-630); *Oryza sativa SALT* (Garcia *et al.* (1998) *Planta* 207:172-180); *Oryza sativa LIP9* (Aguan *et al.* (1993) *Mol Gen Genet* 240:1-8); *Oryza sativa WSI724* (Takahashi *et al.* (1994) *Plant Mol Biol* 26:339-352); *Oryza sativa WSI18* (Oh *et al.* (2005) *Plant Physiol* 138:341-351); *AREB1*, *AREB2*, and *ABF3* (Yoshida *et al.* (2010) *Plant J* 61:672-685); *Oryza sativa DIP1*, *UGE1*, *RIG1B*, and *RAB21* promoters (Yi *et al.* (2010) *Planta* 232:743-754); cotton *D113* (Luo *et al.* (2008) *Plant Cell Rep* 27:707-717); the *dehydrin* promoter; the *ASI* promoter; the *WGA* promoter; the *P511* promoter; and the *HS70* promoter; the *dehydrin* (DHN) promoter (Robertson *et al.* (1995) *Physiol Plant* 94:470-478); the alpha-amylase/subtilisin inhibitor (ASI) promoter (Furtado *et al.* (2003) *Plant Mol Biol* 52:787-799); the *WGA* promoter; and the *HS70* promoter; each of which is herein incorporated by reference in its entirety.

In some embodiments, the inducible promoter that drives the expression of a site-specific recombinase and subsequent excision of the excision cassette is a *Rab17* promoter, such as the maize *rab17* promoter or an active variant or fragment thereof. The maize *rab17* (responsive to abscisic acid) gene (GenBank Accession No. X15994; Vilardell *et al.* (1990) *Plant Mol Biol* 14:423-432; Vilardell *et al.* (1991) *Plant Mol Biol* 17:985-993; each of which is herein incorporated in its entirety) is expressed in late embryos, but its expression can be induced by exposure to abscisic acid, cold temperatures, or water stress. The sequence of the maize *rab17* promoter corresponds to nucleotides 1-558 of GenBank Accession No. X15994, which was disclosed in Vilardell *et al.* (1990) *Plant Mol Biol* 14:423-432 and is set forth in SEQ ID NO: 17. An

alternative maize *rab17* promoter was disclosed in U.S. Patent Nos. 7,253,000 and 7,491,813, each of which is herein incorporated by reference in its entirety, and is set forth in SEQ ID NO: 18. The *rab17* promoter contains four abscisic acid responsive elements (ABRE) (Busk *et al.* (1997) *Plant J* 11:1285-1295, which is herein incorporated by reference in its entirety). The ABRE elements in the maize *rab17* promoter can be found at nucleotides 304-309, 348-353, 363-368, 369-374, 414-419, and 427-432 of SEQ ID NO: 18. The *rab17* promoter also contains drought-responsive elements (DRE), of which the core sequence is identical to the DRE (drought-responsive) and CRT (cold-response elements) elements in *Arabidopsis*. The drought-responsive elements of the maize *rab17* promoter are found at nucleotides 233-238, 299-304, and 322-327 of SEQ ID NO: 18. The CAAT and TATAA box can be found from nucleotides 395 to 398 and 479 to 483 of SEQ ID NO: 18, respectively. In those embodiments wherein the inducible promoter that regulates the expression of the site-specific recombinase is a *rab17* promoter, the expression of the recombinase can be induced by desiccating a host cell (e.g., plant cell) or host (e.g., plant or plant part) or exposing the host cell or host to drought conditions, cold temperatures, or abscisic acid.

In some embodiments, the stress-inducible promoter of the presently disclosed polynucleotide constructs has the sequence set forth in SEQ ID NO: 18 or an active variant or fragment thereof. In other embodiments, the stress-inducible promoter of the presently disclosed polynucleotide constructs has the sequence set forth in SEQ ID NO: 17 or 19 or an active variant or fragment thereof.

In some embodiments of the methods and compositions, the polynucleotide constructs comprise active variants or fragments of the maize *rab17* promoter. An active variant or fragment of a maize *rab17* promoter (e.g., SEQ ID NO: 17, 18, 19) is a polynucleotide variant or fragment that retains the ability to initiate transcription in response to drought conditions, desiccation, cold, and/or ABA. In some of these embodiments, the promoter comprises at least one DRE element. In some embodiments, an active fragment of a maize *rab17* promoter may comprise at least about 50, 100, 150, 200, 250, 300, 350, 400, 450, or 500 contiguous nucleotides of SEQ ID NO: 17, 18, or 19, or may have at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%,

90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity to SEQ ID NO: 17, 18, or 19. In particular embodiments, the promoter of the compositions and methods comprises from about -219 to about -102 of the maize *rab17* promoter (corresponding to nucleotides 291 to 408 of SEQ ID NO: 18). In other embodiments, the
5 active maize *rab17* promoter fragment comprises from about -219 to about -80 of the maize *rab17* promoter (nucleotides 291 to 430 of SEQ ID NO: 18), which comprises most of the DRE and ABRE elements.

In some embodiments, the expression of the site-specific recombinase is regulated by a promoter comprising a maize *rab17* promoter or a fragment or variant thereof, and
10 an attachment site, such as an attachment B (attB) site as described in U.S. Application Publication No. 2011/0167516 (which is herein incorporated by reference in its entirety), and in some of these embodiments, the attB site modifies the activity of the maize *rab17* promoter.

As used herein, a “modulator” refers to a polynucleotide that when present
15 between a promoter and a coding sequence, serves to increase or decrease the activity of the promoter. Non-limiting examples of modulators include recombination sites, operators, and insulators.

Attachment sites are site-specific recombination sites found in viral and bacterial genomes that facilitate the integration or excision of the viral genome into and out of its
20 host genome. Non-limiting examples of a viral and bacterial host system that utilize attachment sites is the lambda bacteriophage and *E. coli* system (Weisberg and Landy (1983) In Lambda II, eds. Hendrix *et al.* (Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.) pp.211-250). The modulator of the maize *rab17* promoter can be an *E.coli* attachment site B (attB) site. The attB site can be a naturally occurring *E. coli* attB site or
25 an active variant or fragment thereof or a synthetically derived sequence. Synthetically derived attB sites and active variants and fragments of naturally occurring attB sites are those that are capable of recombining with a bacteriophage lambda attachment P site, a process that is catalyzed by the bacteriophage lambda Integrase (Int) and the *E. coli* Integration Host Factor (IHF) proteins (Landy (1989) *Ann Rev Biochem* 58: 913-949,
30 which is herein incorporated by reference in its entirety). AttB sites typically have a

length of about 25 nucleotides, with a core 15-base pair sequence that is involved in the actual crossover event. Alternatively, active variants and fragments of naturally occurring attB sites are those that are capable of modulating the activity of a promoter. Non-limiting examples of attB sites that can be used include attB1 (SEQ ID NO: 20), attB2 (SEQ ID NO: 21), attB3 (SEQ ID NO: 22), and attB4 (SEQ ID NO: 23), and variants or fragments thereof. In some embodiments, the modulator is an active variant or fragment of an attB site that is capable of modulating (i.e., increasing, decreasing) the activity of a promoter, but is not capable of recombination with an attachment P site. Non-limiting examples of such active variants of an attB site include those having the sequence set forth in SEQ ID NO: 24, 25, or 26.

In some embodiments, the distance of the modulator (e.g., attB site) from the promoter impacts the ability of the modulator to modify the activity of the promoter. The modulator may be contiguous with the promoter and/or the coding polynucleotide. In other embodiments, a linker sequence separates the promoter sequence and the modulator (e.g., attB site). As used herein, a “linker sequence” is a nucleotide sequence that functions to link one functional sequence with another without otherwise contributing to the expression or translation of a coding polynucleotide. Accordingly, the actual sequence of the linker sequence can vary. The linker sequence can comprise plasmid sequences, restriction sites, and/or regions of the 5'-untranslated region (5'-UTR) of the gene from which the promoter is derived. The linker sequence separating the promoter and the modulator (e.g., attB site) can have a length of about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 150, 200, 250, 300, 400, 500, 1000 nucleotides or greater. In certain embodiments, a linker sequence of about 133 nucleotides separates the maize *rab17* promoter and the modulator (e.g., attB site). In some embodiments, the linker sequence comprises a fragment of the *rab17* 5'-UTR. The fragment of the 5'-UTR can be about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100 nucleotides, or greater, in length. In certain embodiments, the promoter comprises a linker sequence separating the maize *rab17* promoter and the modulator (e.g., attB site) that comprises 95 nucleotides of the maize *rab17* 5'-UTR. In some of these embodiments, the 95 nucleotide

sequence has the sequence set forth in SEQ ID NO: 27. In certain embodiments, the linker sequence between the maize *rab17* promoter and modulator (e.g., attB site) has the sequence set forth in SEQ ID NO: 28 or a variant or fragment thereof.

In some embodiments, the promoter comprises a linker sequence separating the modulator (e.g., attB site) and the site-specific recombinase-coding polynucleotide. The length and sequence of this linker may also vary and can be about 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 25, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 150, 200, 250, 300, 400, 500, 1000 nucleotides or greater in length. In certain embodiments, a linker sequence of about 61 nucleotides separates the modulator (e.g., attB site) and the recombinase-encoding polynucleotide. In certain embodiments, the linker sequence between the modulator (e.g., attB site) and the coding polynucleotide has the sequence set forth in SEQ ID NO: 29 or a variant or fragment thereof. In other embodiments, a linker sequence of about 25 nucleotides separates the modulator (e.g., attB site) and the coding polynucleotide. In certain embodiments, the linker sequence between the modulator (e.g., attB site) and the coding polynucleotide has the sequence set forth in SEQ ID NO: 30.

In certain embodiments, the stress-inducible promoter that regulates the expression of the site-specific recombinase has the sequence set forth in SEQ ID NO: 31 or a variant or fragment thereof.

In other embodiments of the presently disclosed compositions and methods, the inducible promoter that regulates the expression of the site-specific recombinase is a chemical-inducible promoter. In some of these embodiments, the chemical-inducible promoter is a sulfonylurea (SU)-inducible promoter that has at least one operator sequence capable of binding to a sulfonylurea-responsive transcriptional repressor (SuR) protein, such as those disclosed in U.S. Application Publication Nos. 2010/0105141 and 2011/0287936.

As used herein, a “sulfonylurea-responsive transcriptional repressor” or “SuR” refers to a transcriptional repressor protein whose binding to an operator sequence is controlled by a ligand comprising a sulfonylurea compound. The SuR proteins useful in the presently disclosed methods and compositions include those that bind specifically to an operator sequence in the absence of a sulfonylurea ligand.

In some embodiments, the SuR protein is one that specifically binds to a tetracycline operator, wherein the specific binding is regulated by a sulfonylurea compound. Thus, in some embodiments, the sulfonylurea-inducible promoter comprises at least one tetracycline (tet) operator sequence. Tetracycline operator sequences are known in the art and include the tet operator sequence set forth in SEQ ID NO: 32. The tet operator sequence can be located within 0-30 nucleotides 5' or 3' of the TATA box of the chemical-regulated promoter, including, for example, within 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1, or 0 nt of the TATA box. In other instances, the tet operator sequence may partially overlap with the TATA box sequence. In one non-limiting example, the tet operator sequence is SEQ ID NO: 32 or an active variant or fragment thereof.

Useful tet operator containing promoters include, for example, those known in the art (see, e.g., Matzke *et al.* (2003) *Plant Mol Biol Rep* 21:9-19; Padidam (2003) *Curr Op Plant Biol* 6:169-177; Gatz & Quail (1988) *PNAS* 85:1394-1397; Ulmasov *et al.* (1997) *Plant Mol Biol* 35:417-424; Weinmann *et al.* (1994) *Plant J* 5:559-569; each of which is herein incorporated by reference in its entirety). One or more tet operator sequences can be added to a promoter in order to produce a sulfonylurea-inducible promoter. See, for example, Weinmann *et al.* (1994) *Plant J* 5:559-569; Love *et al.* (2000) *Plant J* 21:579-588. In addition, the widely tested tetracycline regulated expression system for plants using the CaMV 35S promoter (Gatz *et al.* (1992) *Plant J* 2:397-404; which is herein incorporated by reference in its entirety) having three tet operators introduced near the TATA box (3XOpT 35S) can be used as the sulfonylurea-inducible promoter.

Thus, a SU-inducible promoter comprising at least one, two, three or more operators capable of binding a SuR (including a tet operator, such as that set forth in SEQ ID NO:32 or an active variant or fragment thereof) can be used to regulate the expression of the site-specific recombinase. Any promoter can be combined with an operator capable of binding a SuR to generate a SU-inducible promoter. In specific embodiments, the promoter is active in plant cells. The promoter can be a constitutive promoter or a non-constitutive promoter. Non-constitutive promoters include tissue-preferred

promoter, such as a promoter that is primarily expressed in roots, leaves, stems, flowers, silks, anthers, pollen, meristem, seed, endosperm, or embryos.

In particular embodiments, the promoter is a plant actin promoter, a banana streak virus promoter (BSV), an MMV promoter, an enhanced MMV promoter (dMMV), a
5 plant P450 promoter, or an elongation factor 1a (EF1A) promoter (U.S. Application Publication No. 20080313776, which is herein incorporated by reference in its entirety).

In those embodiments wherein the inducible promoter that is operably linked to the polynucleotide encoding the site-specific recombinase is a SU-inducible promoter, the host cell further comprises a sulfonylurea-responsive transcriptional repressor (SuR) or
10 the polynucleotide construct comprises a polynucleotide encoding a SuR. Non-limiting examples of SuR polynucleotide and polypeptide sequences include those disclosed in U.S. Application Publication No. 2011/0287936, such as the polypeptide sequences set forth in SEQ ID NOs: 3-419 and the polynucleotide sequences set forth in SEQ ID NOs: 420-836 of U.S. Application Publication No. 2011/0287936, which is herein incorporated
15 by reference in its entirety. Additional non-limiting examples of SuR polynucleotide and polypeptide sequences include those disclosed in U.S. Application Publication No. 2010/0105141, such as the polypeptide sequences set forth in SEQ ID NO: 3-401, 1206-1213, 1228-1233, and 1240-1243 and the polynucleotide sequences set forth in SEQ ID NO: 434-832, 1214-1221, 1222-1227, 1234-1239, and 1244-1247 of U.S. Application
20 Publication No. 2010/0105141, which is herein incorporated by reference in its entirety.

In those embodiments wherein the presently disclosed polynucleotide constructs further comprise a polynucleotide encoding a SuR, the SuR-encoding polynucleotide is operably linked to a promoter that is active in a plant. The promoter may be a constitutive or a non-constitutive promoter, including a tissue-preferred promoter.

25 In particular embodiments, the promoter that is operably linked to the SuR-encoding polynucleotide comprises operator sequences that are capable of binding to SuR, which allows for autoregulation of the repressor and enhanced induction of the SU-inducible promoter and expression of the site-specific recombinase. See, for example, U.S. Application Publication No. 2011/0287936.

In particular embodiments, the SuR-encoding polynucleotide and optionally, the promoter operably linked thereto, is present within the excision cassette of the presently disclosed polynucleotide constructs, such that the polynucleotide is excised upon induction of the SU-inducible promoter and expression of the site-specific recombinase.

5 A variety of SU compounds can be used to bind to the SuR and induce the SU-inducible promoter. Sulfonylurea molecules comprise a sulfonylurea moiety ($-S(O)_2NHC(O)NH(R)-$). In sulfonylurea herbicides, the sulfonyl end of the sulfonylurea moiety is connected either directly or by way of an oxygen atom or an optionally substituted amino or methylene group to a typically substituted cyclic or acyclic group.

10 At the opposite end of the sulfonylurea bridge, the amino group, which may have a substituent such as methyl (R being CH_3) instead of hydrogen, is connected to a heterocyclic group, typically a symmetric pyrimidine or triazine ring, having one or two substituents such as methyl, ethyl, trifluoromethyl, methoxy, ethoxy, methylamino, dimethylamino, ethylamino and the halogens. Sulfonylurea herbicides can be in the form of the free acid or a salt. In the free acid form, the sulfonamide nitrogen on the bridge is not deprotonated (*i.e.*, $-S(O)_2NHC(O)NH(R)$), while in the salt form, the sulfonamide nitrogen atom on the bridge is deprotonated, and a cation is present, typically of an alkali metal or alkaline earth metal, most commonly sodium or potassium. Sulfonylurea compounds include, for example, compound classes such as pyrimidinylsulfonylurea
15 compounds, triazinylsulfonylurea compounds, thiadiazolylurea compounds, and pharmaceuticals such as antidiabetic drugs, as well as salts and other derivatives thereof. Examples of pyrimidinylsulfonylurea compounds include amidosulfuron, azimsulfuron, bensulfuron, bensulfuron-methyl, chlorimuron, chlorimuron-ethyl, cyclosulfamuron, ethoxysulfuron, flazasulfuron, flucetosulfuron, flupyrsulfuron, flupyrsulfuron-methyl, foramsulfuron, halosulfuron, halosulfuron-methyl, imazosulfuron, mesosulfuron, mesosulfuron-methyl, nicosulfuron, orthosulfamuron, oxasulfuron, primisulfuron, primisulfuron-methyl, pyrazosulfuron, pyrazosulfuron-ethyl, rimsulfuron, sulfometuron, sulfometuron-methyl, sulfosulfuron, trifloxysulfuron and salts and derivatives thereof. Examples of triazinylsulfonylurea compounds include chlorsulfuron, cinosulfuron,
20 ethametsulfuron, ethametsulfuron-methyl, iodosulfuron, iodosulfuron-methyl,

metsulfuron, metsulfuron-methyl, prosulfuron, thifensulfuron, thifensulfuron-methyl, triasulfuron, tribenuron, tribenuron-methyl, triflusulfuron, triflusulfuron-methyl, tritosulfuron and salts and derivatives thereof. Examples of thiadiazolylurea compounds include buthiuron, ethidimuron, tebuthiuron, thiazafluron, thidiazuron, 5 pyrimidinylsulfonylurea compound (e.g., amidosulfuron, azimsulfuron, bensulfuron, chlorimuron, cyclosulfamuron, ethoxysulfuron, flazasulfuron, flucetosulfuron, flupyrsulfuron, foramsulfuron, halosulfuron, imazosulfuron, mesosulfuron, nicosulfuron, orthosulfamuron, oxasulfuron, primisulfuron, pyrazosulfuron, rimsulfuron, sulfometuron, sulfosulfuron and trifloxysulfuron); a triazinylsulfonylurea compound (e.g., 10 chlorsulfuron, cinosulfuron, ethametsulfuron, iodosulfuron, metsulfuron, prosulfuron, thifensulfuron, triasulfuron, tribenuron, triflusulfuron and tritosulfuron); or a thiadazolylurea compound (e.g., cloransulam, diclosulam, florasulam, flumetsulam, metosulam, and penoxsulam) and salts and derivatives thereof. Examples of antidiabetic drugs include acetohexamide, chlorpropamide, tolbutamide, tolazamide, glipizide, 15 gliclazide, glibenclamide (glyburide), gliquidone, glimepiride and salts and derivatives thereof. In some systems, the SuR polypeptides specifically bind to more than one sulfonylurea compound, so one can chose which SU ligand to apply to the plant.

In some examples, the sulfonylurea compound is selected from the group consisting of chlorsulfuron, ethametsulfuron-methyl, metsulfuron-methyl, thifensulfuron- 20 methyl, sulfometuron-methyl, tribenuron-methyl, chlorimuron-ethyl, nicosulfuron, and rimsulfuron.

In other embodiments, the sulfonylurea compound comprises a pyrimidinylsulfonylurea, a triazinylsulfonylurea, a thiadazolylurea, a chlorosulfuron, an ethametsulfuron, a thifensulfuron, a metsulfuron, a sulfometuron, a tribenuron, a 25 chlorimuron, a nicosulfuron, or a rimsulfuron compound.

In some embodiments, it may be necessary for a plant or plant part that is contacted with a SU in order to induce the SU-inducible promoter to have tolerance to the SU. A host (e.g., a plant or plant part) may be naturally tolerant to the SU ligand, or the host (e.g., the plant or plant part) may be tolerant to the SU ligand as a result of human 30 intervention such as, for example, by the use of a recombinant construct, plant breeding

or genetic engineering. Thus, the host (e.g., the plant or plant part) employed in the various methods disclosed herein can comprise a native or a heterologous sequence that confers tolerance to the sulfonylurea compound.

5 In some of these embodiments, the presently disclosed polynucleotide constructs can comprise a polynucleotide encoding a sulfonylurea-tolerance polypeptide, which is a polypeptide that when expressed in a host (e.g., plant or plant part) confers tolerance to at least one sulfonylurea. In some of these embodiments, the polynucleotide encoding the SU-tolerance polypeptide is comprised within the excision cassette.

10 In other embodiments, the herbicide tolerance polypeptide that is expressed upon excision of the excision cassette is a SU-tolerance polypeptide, such that the plant or plant part does not have tolerance to SU prior to the addition of SU to the plant or plant part, but upon the addition of SU, the excision cassette is excised and the SU-tolerance polypeptide is subsequently expressed, which allows for protection of the plant or plant part from damage due to the SU.

15 Sulfonylurea herbicides inhibit growth of higher plants by blocking acetolactate synthase (ALS), also known as, acetohydroxy acid synthase (AHAS). Thus, in some embodiments, the SU-tolerance polypeptide is an ALS inhibitor-tolerance polypeptide, as described elsewhere herein.

20 When the inducible promoter of the presently disclosed polynucleotide constructs is activated, a site-specific recombinase is expressed, which catalyzes the excision of the excision cassette comprised within the polynucleotide construct. As used herein, an “excision cassette” refers to a polynucleotide that is flanked by recombination sites that are recombinogenic with one another and directly repeated, such that when acted upon by a site-specific recombinase that recognizes the recombination sites, the nucleotide
25 sequence within the recombination sites is excised from the remaining polynucleotide. The excision cassette of the presently disclosed polynucleotide constructs comprise a first expression cassette comprising a site-specific recombinase-encoding polynucleotide operably linked to an inducible promoter and optionally, at least one of a polynucleotide encoding a selectable marker, a polynucleotide encoding a cell proliferation factor, a

polynucleotide encoding a herbicide tolerance polypeptide, and a polynucleotide of interest.

A site-specific recombinase, also referred to herein as a recombinase, is a polypeptide that catalyzes conservative site-specific recombination between its compatible recombination sites, and includes native polypeptides as well as derivatives, variants and/or fragments that retain activity, and native polynucleotides, derivatives, variants, and/or fragments that encode a recombinase that retains activity. The recombinase used in the methods and compositions can be a native recombinase or a biologically active fragment or variant of the recombinase. For reviews of site-specific recombinases and their recognition sites, see Sauer (1994) *Curr Op Biotechnol* 5:521-527; and Sadowski (1993) *FASEB* 7:760-767, each of which is herein incorporated by reference in its entirety.

Any recombinase system can be used in the presently disclosed methods and compositions. Non-limiting examples of site-specific recombinases include FLP, Cre, S-CRE, V-CRE, Dre, SSV1, lambda Int, phi C31 Int, HK022, R, Gin, Tn1721, CinH, ParA, Tn5053, Bxb1, TP907-1, U153, and other site-specific recombinases known in the art, including those described in Thomson and Ow (2006) *Genesis* 44:465-476, which is herein incorporated by reference in its entirety. Examples of site-specific recombination systems used in plants can be found in U.S. Patent Nos. 5,929,301, 6,175,056, 6,331,661; and International Application Publication Nos. WO 99/25821, WO 99/25855, WO 99/25841, and WO 99/25840, the contents of each are herein incorporated by reference.

In some embodiments, the recombinase is a member of the Integrase or Resolvase families, including biologically active variants and fragments thereof. The Integrase family of recombinases has over one hundred members and includes, for example, FLP, Cre, lambda integrase, and R. For other members of the Integrase family, see, for example, Esposito *et al.* (1997) *Nucleic Acids Res* 25:3605-3614; and Abremski *et al.* (1992) *Protein Eng* 5:87-91; each of which are herein incorporated by reference in its entirety. Other recombination systems include, for example, the *Streptomyces* bacteriophage phi C31 (Kuhstoss *et al.* (1991) *J Mol Biol* 20:897-908); the SSV1 site-specific recombination system from *Sulfolobus shibatae* (Maskhelishvili *et al.* (1993) *Mol*

Gen Genet 237:334-342); and a retroviral integrase-based integration system (Tanaka *et al.* (1998) *Gene* 17:67-76). In some embodiments, the recombinase does not require cofactors or a supercoiled substrate. Such recombinases include Cre, FLP, or active variants or fragments thereof.

5 The FLP recombinase is a protein that catalyzes a site-specific reaction that is involved in amplifying the copy number of the two-micron plasmid of *S. cerevisiae* during DNA replication. FLP recombinase catalyzes site-specific recombination between two FRT sites. The FLP protein has been cloned and expressed (Cox (1993) *Proc Natl Acad Sci USA* 80:4223-4227, which is herein incorporated by reference in its entirety).

10 The FLP recombinase for use in the methods and compositions may be derived from the genus *Saccharomyces*. In some embodiments, a recombinase polynucleotide modified to comprise more plant-preferred codons is used. A recombinant FLP enzyme encoded by a nucleotide sequence comprising maize preferred codons (FLPm) that catalyzes site-specific recombination events is known (the polynucleotide and polypeptide sequence of
15 which is set forth in SEQ ID NO: 33 and 34, respectively; see, *e.g.*, US Patent 5,929,301, which is herein incorporated by reference in its entirety). Additional functional variants and fragments of FLP are known (Buchholz *et al.* (1998) *Nat Biotechnol* 16:657-662; Hartung *et al.* (1998) *J Biol Chem* 273:22884-22891; Saxena *et al.* (1997) *Biochim Biophys Acta* 1340:187-204; Hartley *et al.* (1980) *Nature* 286:860-864; Voziyanov *et al.*
20 (2002) *Nucleic Acids Res* 30:1656-1663; Zhu & Sadowski (1995) *J Biol Chem* 270:23044-23054; and U.S. Patent No. 7,238,854, each of which is herein incorporated by reference in its entirety).

 The bacteriophage recombinase Cre catalyzes site-specific recombination between two *lox* sites. The Cre recombinase is known (Guo *et al.* (1997) *Nature* 389:40-46;
25 Abremski *et al.* (1984) *J Biol Chem* 259:1509-1514; Chen *et al.* (1996) *Somat Cell Mol Genet* 22:477-488; Shaikh *et al.* (1977) *J Biol Chem* 272:5695-5702; and, Buchholz *et al.* (1998) *Nat Biotechnol* 16:657-662, each of which is herein incorporated by reference in its entirety). Cre polynucleotide sequences may also be synthesized using plant-preferred codons, for example such sequences (moCre; the polynucleotide and polypeptide
30 sequence of which is set forth in SEQ ID NO: 35 and 36, respectively) are described, for

example, in International Application Publication No. WO 99/25840, which is herein incorporated by reference in its entirety. Variants of the Cre recombinase are known (see, for example U.S. Patent No. 6,890,726; Rufer & Sauer (2002) *Nucleic Acids Res* 30:2764-2772; Wierzbicki *et al.* (1987) *J Mol Biol* 195:785-794; Petyuk *et al.* (2004) *J Biol Chem* 279:37040-37048; Hartung & Kisters-Woike (1998) *J Biol Chem* 273:22884-22891; Santoro & Schultz (2002) *Proc Natl Acad Sci USA* 99:4185-4190; Koresawa *et al.* (2000) *J Biochem* (Tokyo) 127:367-372; and Vergunst *et al.* (2000) *Science* 290:979-982, each of which are herein incorporated by reference in its entirety).

In some embodiments, the recombinase is a S-CRE, V-CRE recombinase (Suzuki & Nakayama (2011) *Nucl Acid Res* 39(8):e49) or Dre recombinase (Sauer & McDermott (2004) *Nucl Acid Res* 32(20):6086-6095), each of which is herein incorporated by reference in its entirety.

In some embodiments, the recombinase is a chimeric recombinase, which is a recombinant fusion protein that is capable of catalyzing site-specific recombination between recombination sites that originate from different recombination systems. For example, if the set of recombination sites comprises a FRT site and a LoxP site, a chimeric FLP/Cre recombinase or active variant or fragment thereof can be used, or both recombinases may be separately provided. Methods for the production and use of such chimeric recombinases or active variants or fragments thereof are described, for example, in International Application Publication No. WO 99/25840; and Shaikh & Sadowski (2000) *J Mol Biol* 302:27-48, each of which are herein incorporated by reference in its entirety.

In other embodiments, a variant recombinase is used. Methods for modifying the kinetics, cofactor interaction and requirements, expression, optimal conditions, and/or recognition site specificity, and screening for activity of recombinases and variants are known, see for example Miller *et al.* (1980) *Cell* 20:721-9; Lange-Gustafson and Nash (1984) *J Biol Chem* 259:12724-32; Christ *et al.* (1998) *J Mol Biol* 288:825-36; Lorbach *et al.* (2000) *J Mol Biol* 296:1175-81; Vergunst *et al.* (2000) *Science* 290:979-82; Dorgai *et al.* (1995) *J Mol Biol* 252:178-88; Dorgai *et al.* (1998) *J Mol Biol* 277:1059-70; Yagu *et al.* (1995) *J Mol Biol* 252:163-7; Scilimente *et al.* (2001) *Nucleic Acids Res* 29:5044-51;

Santoro and Schultze (2002) *Proc Natl Acad Sci USA* 99:4185-90; Buchholz and Stewart (2001) *Nat Biotechnol* 19:1047-52; Voziyanov *et al.* (2002) *Nucleic Acids Res* 30:1656-63; Voziyanov *et al.* (2003) *J Mol Biol* 326:65-76; Klippel *et al.* (1988) *EMBO J* 7:3983-9; Arnold *et al.* (1999) *EMBO J* 18:1407-14; and International Application Publication
5 Nos. WO 03/08045, WO 99/25840, and WO 99/25841; each of which is herein incorporated by reference in its entirety.

By "recombination site" is intended a polynucleotide (native or synthetic/artificial) that is recognized by the recombinase enzyme of interest. As outlined above, many recombination systems are known in the art and one of skill will recognize the
10 appropriate recombination site to be used with the recombinase of interest.

Non-limiting examples of recombination sites include FRT sites including, for example, the native FRT site (FRT1, SEQ ID NO:37), and various functional variants of FRT, including but not limited to, FRT5 (SEQ ID NO:38), FRT6 (SEQ ID NO:39), FRT7 (SEQ ID NO:40), FRT12 (SEQ ID NO: 41), and FRT87 (SEQ
15 ID NO:42). See, for example, International Application Publication Nos. WO 03/054189, WO 02/00900, and WO 01/23545; and Schlake *et al.* (1994) *Biochemistry* 33:12745-12751, each of which is herein incorporated by reference. Recombination sites from the Cre/Lox site-specific recombination system can be used. Such recombination sites include, for example, native LOX sites and
20 various functional variants of LOX.

In some embodiments, the recombination site is a functional variant of a FRT site or functional variant of a LOX site, any combination thereof, or any other combination of recombinogenic or non-recombinogenic recombination sites known. Functional variants include chimeric recombination sites, such as an FRT site fused to a
25 LOX site (see, for example, Luo *et al.* (2007) *Plant Biotech J* 5:263-274, which is herein incorporated by reference in its entirety). Functional variants also include minimal sites (FRT and/or LOX alone or in combination). The minimal native FRT recombination site (SEQ ID NO: 37) has been characterized and comprises a series of domains comprising a pair of 11 base pair symmetry elements, which are the FLP binding sites; the 8 base pair
30 core, or spacer, region; and the polypyrimidine tracts. In some embodiments, at least one

modified FRT recombination site is used. Modified or variant FRT recombination sites are sites having mutations such as alterations, additions, or deletions in the sequence. The modifications include sequence modification at any position, including but not limited to, a modification in at least one of the 8 base pair spacer domain, a symmetry element, and/or a polypyrimidine tract. FRT variants include minimal sites (see, *e.g.*,
5 Broach *et al.* (1982) *Cell* 29:227-234; Senecoff *et al.* (1985) *Proc Natl Acad Sci USA* 82:7270-7274; Gronostajski & Sadowski (1985) *J Biol Chem* 260:12320-12327; Senecoff *et al.* (1988) *J Mol Biol* 201:405-421; and International Application Publication No. WO99/25821), and sequence variants (see, for example, Schlake & Bode (1994)
10 *Biochemistry* 33:12746-12751; Seibler & Bode (1997) *Biochemistry* 36:1740-1747; Umlauf & Cox (1988) *EMBO J* 7:1845-1852; Senecoff *et al.* (1988) *J Mol Biol* 201:405-421; Voziyanov *et al.* (2002) *Nucleic Acids Res* 30:7; International Application Publication Nos. WO 07/011733, WO 99/25854, WO 99/25840, WO 99/25855, WO 99/25853 and WO 99/25821; and U.S. Patent Nos. 7,060,499 and 7,476,539; each of
15 which are herein incorporated by reference in its entirety).

An analysis of the recombination activity of variant LOX sites is presented in Lee *et al.* (1998) *Gene* 216:55-65 and in U.S. Patent No. 6,465,254. Also, see for example, Huang *et al.* (1991) *Nucleic Acids Res* 19:443-448; Sadowski (1995) In *Progress in Nucleic Acid Research and Molecular Biology*
20 Vol. 51, pp. 53-91; U.S. Patent No. 6,465,254; Cox (1989) In *Mobile DNA*, Berg and Howe (eds) American Society of Microbiology, Washington D.C., pp. 116-670; Dixon *et al.* (1995) *Mol Microbiol* 18:449-458; Buchholz *et al.* (1996) *Nucleic Acids Res* 24:3118-3119; Kilby *et al.* (1993) *Trends Genet* 9:413-421; Rossant & Geagy (1995) *Nat Med* 1:592-594; Albert *et al.* (1995) *Plant J* 7:649-
25 659; Bayley *et al.* (1992) *Plant Mol Biol* 18:353-361; Odell *et al.* (1990) *Mol Gen Genet* 223:369-378; Dale & Ow (1991) *Proc Natl Acad Sci USA* 88:10558-10562; Qui *et al.* (1994) *Proc Natl Acad Sci USA* 91:1706-1710; Stuurman *et al.* (1996) *Plant Mol Biol* 32:901-913; Dale *et al.* (1990) *Gene* 91:79-85; and International Application Publication No. WO 01/111058; each of which is herein incorporated
30 by reference in its entirety.

Naturally occurring recombination sites or biologically active variants thereof are of use. Methods to determine if a modified recombination site is recombinogenic are known (see, for example, International Application Publication No. WO 07/011733, which is herein incorporated by reference in its entirety). Variant recognition sites are known, see for example, Hoess *et al.* (1986) *Nucleic Acids Res* 14:2287-300; Albert *et al.* (1995) *Plant J* 7:649-59; Thomson *et al.* (2003) *Genesis* 36:162-7; Huang *et al.* (1991) *Nucleic Acids Res* 19:443-8; Siebler and Bode (1997) *Biochemistry* 36:1740-7; Schlake and Bode (1994) *Biochemistry* 33:12746-51; Thygarajan *et al.* (2001) *Mol Cell Biol* 21:3926-34; Umlauf and Cox (1988) *EMBO J* 7:1845-52; Lee and Saito (1998) *Gene* 216:55-65; International Application Publication Nos. WO 01/23545, WO 99/25851, WO 01/11058, WO 01/07572; and U.S. Patent No. 5,888,732; each of which is herein incorporated by reference in its entirety.

The recombination sites employed in the methods and compositions can be identical or dissimilar sequences, so long as the sites are recombinogenic with respect to one another.

By "recombinogenic" is intended that the set of recombination sites (*i.e.*, dissimilar or corresponding) are capable of recombining with one another. Alternatively, by "non-recombinogenic" is intended the set of recombination sites, in the presence of the appropriate recombinase, will not recombine with one another or recombination between the sites is minimal. Accordingly, it is recognized that any suitable set of recombinogenic recombination sites may be utilized, including a FRT site or functional variant thereof, a LOX site or functional variant thereof, any combination thereof, or any other combination of recombination sites known in the art.

In some embodiments, the recombination sites are asymmetric, and the orientation of any two sites relative to each other will determine the recombination reaction product. Directly repeated recombination sites are those recombination sites in a set of recombinogenic recombination sites that are arranged in the same orientation, such that recombination between these sites results in excision, rather than inversion, of the intervening DNA sequence. Inverted recombination sites are those recombination sites in a set of recombinogenic recombination sites that are arranged in the opposite orientation,

so that recombination between these sites results in inversion, rather than excision, of the intervening DNA sequence. The presently disclosed polynucleotide constructs comprise recombination sites that are recombinogenic with one another and directly repeated so as to result in excision of the excision cassette.

5 The presently disclosed compositions and methods utilize at least one polynucleotide that confers herbicide tolerance. Tolerance to specific herbicides can be conferred by engineering genes into plants which encode appropriate herbicide metabolizing enzymes and/or insensitive herbicide targets. Such polypeptides are referred to as “herbicide tolerance polypeptides”. In some embodiments these enzymes,
10 and the nucleic acids that encode them, originate from a plant. In other embodiments, they are derived from other organisms, such as microbes. *See, e.g.,* Padgett *et al.* (1996) “New weed control opportunities: Development of soybeans with a Roundup Ready[®] gene” and Vasil (1996) “Phosphinothricin-resistant crops,” both in *Herbicide-Resistant Crops*, ed. Duke (CRC Press, Boca Raton, Florida) pp. 54-84 and pp. 85-91.

15 An “herbicide” is a chemical that causes temporary or permanent injury to a plant. Non-limiting examples of herbicides that can be employed in the various methods and compositions of the invention are discussed in further detail elsewhere herein. A herbicide may be incorporated into the plant or plant part, or it may act on the plant or plant part without being incorporated into the plant or plant part. An “active ingredient”
20 is the chemical in a herbicide formulation primarily responsible for its phytotoxicity and which is identified as the active ingredient on the product label. Product label information is available from the U.S. Environmental Protection Agency and is updated online at the url oaspub.epa.gov/pestlabl/ppls.own; product label information is also available online at the url www.cdms.net.

25 “Herbicide-tolerant” or “tolerant” in the context of herbicide or other chemical treatment as used herein means that a plant or plant part treated with a particular herbicide or class or subclass of herbicide or other chemical or class or subclass of other chemical will show no significant damage or less damage following that treatment in comparison to an appropriate control plant or plant part. A plant or plant part may be naturally
30 tolerant to a particular herbicide or chemical, or a plant or plant part may be herbicide-

tolerant as a result of human intervention such as, for example, breeding or genetic engineering. An “herbicide-tolerance polypeptide” is a polypeptide that confers herbicide tolerance on a plant or other organism expressing it (*i.e.*, that makes a plant or other organism herbicide-tolerant), and an “herbicide-tolerance polynucleotide” is a polynucleotide that encodes a herbicide-tolerance polypeptide. For example, a sulfonyleurea-tolerance polypeptide is one that confers tolerance to sulfonyleurea herbicides on a plant or other organism that expresses it, an imidazolinone-tolerance polypeptide is one that confers tolerance to imidazolinone herbicides on a plant or other organism that expresses it; and a glyphosate-tolerance polypeptide is one that confers tolerance to glyphosate on a plant or other organism that expresses it.

Thus, a plant or plant part is tolerant to a herbicide or other chemical if it shows damage in comparison to an appropriate control plant or plant part that is less than the damage exhibited by the control plant or plant part by at least 5%, 10%, 15%, 20%, 25%, 30%, 35%, 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 90%, 100%, 150%, 200%, 250%, 300%, 400%, 500%, 600%, 700%, 800%, 900%, or 1000% or more. In this manner, a plant or plant part that is tolerant to a herbicide or other chemical shows “improved tolerance” in comparison to an appropriate control plant or plant part.

Damage resulting from herbicide or other chemical treatment is assessed by evaluating any parameter of plant growth or well-being deemed suitable by one of skill in the art.

Damage can be assessed by visual inspection and/or by statistical analysis of suitable parameters of individual plants or plant parts or of a group of plants or plant parts. Thus, damage may be assessed by evaluating, for example, parameters such as plant height, plant weight, leaf color, leaf length, flowering, fertility, silking, yield, seed production, and the like. Damage may also be assessed by evaluating the time elapsed to a particular stage of development (*e.g.*, silking, flowering, or pollen shed) or the time elapsed until a plant has recovered from treatment with a particular chemical and/or herbicide.

In making such assessments, particular values may be assigned to particular degrees of damage so that statistical analysis or quantitative comparisons may be made. The use of ranges of values to describe particular degrees of damage is known in the art, and any suitable range or scale may be used. For example, herbicide injury scores (also

called tolerance scores) can be assigned as set forth in Table 2. In this scale, a rating of 9 indicates that a herbicide treatment had no effect on a crop, *i.e.*, that no crop reduction or injury was observed following the herbicide treatment. Thus, in this scale, a rating of 9 indicates that the crop exhibited no damage from the herbicide and therefore that the crop is tolerant to the herbicide. As indicated above, herbicide tolerance is also indicated by other ratings in this scale where an appropriate control plant exhibits a lower score on the scale, or where a group of appropriate control plants exhibits a statistically lower score in response to a herbicide treatment than a group of subject plants.

Table 2. Herbicide injury scale (1 to 9 scale scoring system).

Rating	Main categories	Detailed description
9	No Effect	No crop reduction or injury
8	Slight Effect	Slight crop discoloration or stunting
7		Some crop discoloration, stunting, or stunt loss
6		Crop injury more pronounced, but not lasting
5	Moderate Effect	Moderate injury, crop usually recovers
4		Crop injury more lasting, recovery doubtful
3		Lasting crop injury, no recovery

A herbicide does not “significantly damage” a plant or plant part when it either has no effect on a plant or plant part or when it has some effect on a plant or plant part from which the plant later recovers, or when it has an effect which is detrimental but which is offset, for example, by the impact of the particular herbicide on weeds. Thus, for example, a plant or plant part is not “significantly damaged by” a herbicide or other treatment if it exhibits less than 50%, 40%, 30%, 25%, 20%, 15%, 10%, 9%, 8%, 7%, 6%, 5%, 4%, 3%, 2%, or 1% decrease in at least one suitable parameter that is indicative of plant health and/or productivity in comparison to an appropriate control plant or plant part (*e.g.*, an untreated plant or plant part). Suitable parameters that are indicative of plant health and/or productivity include, for example, plant height, plant weight, leaf length, time elapsed to a particular stage of development, flowering, yield, seed production, and the like. The evaluation of a parameter can be by visual inspection and/or by statistical analysis of any suitable parameter. Comparison may be made by

visual inspection and/or by statistical analysis. Accordingly, a plant or plant part is not “significantly damaged by” a herbicide or other treatment if it exhibits a decrease in at least one parameter but that decrease is temporary in nature and the plant or plant part recovers fully within 1 week, 2 weeks, 3 weeks, 4 weeks, or 6 weeks.

5 Conversely, a plant or plant part is significantly damaged by a herbicide or other treatment if it exhibits more than a 50%, 60%, 70%, 80%, 90%, 100%, 110%, 120%, 150%, 170% decrease in at least one suitable parameter that is indicative of plant health and/or productivity in comparison to an appropriate control plant or plant part. Thus, a plant or plant part is significantly damaged if it exhibits a decrease in at least one
10 parameter and the plant or plant part does not recover fully within 1 week, 2 weeks, 3 weeks, 4 weeks, or 6 weeks.

Damage resulting from a herbicide or other chemical treatment of a plant or plant part can be assessed by visual inspection by one of skill in the art and can be evaluated by statistical analysis of suitable parameters. The plant or plant part being evaluated is
15 referred to as the “test plant” or “test plant part.” Typically, an appropriate control plant or plant part is one that expresses the same herbicide-tolerance polypeptide(s) as the plant or plant part being evaluated for herbicide tolerance (*i.e.*, the “test plant”) but that has not been treated with herbicide. In some circumstances, the control plant or plant part is one that has been subjected to the same herbicide treatment as the plant or plant part being
20 evaluated (*i.e.*, the test plant or plant part) but that does not express the enzyme intended to provide tolerance to the herbicide of interest in the test plant or plant part. One of skill in the art will be able to design, perform, and evaluate a suitable controlled experiment to assess the herbicide tolerance of a plant or plant part of interest, including the selection of appropriate test plants or plant part, control plants or plant part, and treatments.

25 Damage caused by a herbicide or other chemical can be assessed at various times after a plant or plant part has been contacted with a herbicide, although in some embodiments, assessment of the plant or plant part for herbicide tolerance occurs during or after rooting/regeneration of the plant or plant part. Often, damage is assessed at about the time that the control plant or plant part exhibits maximum damage. Sometimes,
30 damage is assessed after a period of time in which a control plant or plant part that was

not treated with herbicide has measurably grown and/or developed in comparison to the size or stage at which the treatment was administered. Damage can be assessed at various times, for example, at 12 hours or at 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14 days, or three weeks, four weeks, or longer after the test plant or plant part was treated with herbicide. Any time of assessment is suitable as long as it permits detection of a difference in response to a treatment of test and control plants or plant parts.

Thus, as used herein, a “test plant” or “test plant part” is one which has been transformed with the presently disclosed polynucleotide constructs or is a plant or plant part which is descended from a plant or plant part so altered and which comprises the herbicide tolerance polynucleotide.

A “control” or “control plant” or “control plant part” provides a reference point for measuring changes in phenotype of the subject plant or plant part, and may be any suitable plant or plant part. A control plant or plant part may comprise, for example: (a) a wild-type plant or plant part, *i.e.*, an untransformed plant of the same genotype as the test plant or plant part prior to transformation; (b) a plant or plant part of the same genotype as the starting material but which has been transformed with a null construct (*i.e.*, with a construct which has no known effect on the trait of interest, such as a construct comprising a marker gene); (c) a plant or plant part which is a non-transformed segregant among progeny of a subject plant or plant part; (d) a plant or plant part which is genetically identical to the subject plant or plant part but which is not exposed to the same treatment (*e.g.*, herbicide treatment) as the subject plant or plant part; (e) the subject plant or plant part itself, under conditions in which the herbicide tolerance polynucleotide is not expressed; or (f) the subject plant or plant part itself, under conditions in which it has not been exposed to a particular treatment such as, for example, a herbicide or combination of herbicides and/or other chemicals. In some instances, an appropriate control maize plant or plant part comprises a NK603 event (Nielson *et al.* (2004) *European Food Research and Technology* 219:421-427 and Ridley *et al.* (2002) *Journal of Agriculture and Food Chemistry* 50: 7235-7243), an elite stiff stalk inbred plant, a P3162 plant (Pioneer Hi-Bred International), a 39T66 plant (Pioneer Hi-Bred International), or a 34M91 plant

(Pioneer Hi-Bred International). In some instances, an appropriate control soybean plant or plant part is a “Jack” soybean plant (Illinois Foundation Seed, Champaign, Illinois).

The herbicide tolerance polypeptides used in the presently disclosed compositions and methods can confer tolerance to any respective herbicide. In some embodiments, the herbicide tolerance polypeptide confers tolerance to a herbicide selected from the group consisting of glyphosate, an ALS inhibitor (e.g., a sulfonyleurea), an acetyl Co-A carboxylase inhibitor, a synthetic auxin, a protoporphyrinogen oxidase (PPO) inhibitor herbicide, a pigment synthesis inhibitor herbicide, a phosphinothricin acetyltransferase or a phytoene desaturase inhibitor, a glutamine synthase inhibitor, a hydroxyphenylpyruvate dioxygenase inhibitor, and a protoporphyrinogen oxidase inhibitor.

One herbicide which has been studied extensively is N-phosphonomethylglycine, commonly referred to as glyphosate. Glyphosate is a broad spectrum herbicide that kills both broadleaf and grass-type plants due to inhibition of the enzyme 5-enolpyruvylshikimate-3-phosphate synthase (also referred to as “EPSP synthase” or “EPSPS”), an enzyme which is part of the biosynthetic pathway for the production of aromatic amino acids, hormones, and vitamins. Glyphosate-resistant transgenic plants have been produced which exhibit a commercially viable level of glyphosate resistance due to the introduction of a modified *Agrobacterium* CP4 EPSPS. This modified enzyme is targeted to the chloroplast where, even in the presence of glyphosate, it continues to synthesize EPSP from phosphoenolpyruvic acid (“PEP”) and shikimate-3-phosphate. CP4 glyphosate-resistant soybean transgenic plants are presently in commercial use (e.g., as sold by Monsanto under the name “Roundup Ready[®]”).

In some embodiments, the presently disclosed methods and compositions utilize a polynucleotide that encodes a herbicide tolerance polypeptide that confers tolerance to glyphosate. Various sequences which confer tolerance to glyphosate can be employed in the presently disclosed methods and compositions. In some embodiments, the herbicide tolerance polypeptide that confers resistance to glyphosate has glyphosate transferase activity. As used herein, a “glyphosate transferase” polypeptide has the ability to transfer the acetyl group from acetyl CoA to the N of glyphosate, transfer the propionyl group of

propionyl CoA to the N of glyphosate, or to catalyze the acetylation of glyphosate analogs and/or glyphosate metabolites, *e.g.*, aminomethylphosphonic acid. Methods to assay for this activity are disclosed, for example, in U.S. Publication No. 2003/0083480, U.S. Publication No. 2004/0082770, and U.S. Patent No. 7,405,074, WO2005/012515,
5 WO2002/36782 and WO2003/092360. In one embodiment, the transferase polypeptide comprises a glyphosate-N-acetyltransferase “GLYAT” polypeptide.

As used herein, a GLYAT polypeptide or enzyme comprises a polypeptide which has glyphosate-N-acetyltransferase activity (“GLYAT” activity), *i.e.*, the ability to catalyze the acetylation of glyphosate. In specific embodiments, a polypeptide having
10 glyphosate-N-acetyltransferase activity can transfer the acetyl group from acetyl CoA to the N of glyphosate. In addition, some GLYAT polypeptides transfer the propionyl group of propionyl CoA to the N of glyphosate. Some GLYAT polypeptides are also capable of catalyzing the acetylation of glyphosate analogs and/or glyphosate metabolites, *e.g.*, aminomethylphosphonic acid. GLYAT polypeptides are characterized by their structural
15 similarity to one another, *e.g.*, in terms of sequence similarity when the GLYAT polypeptides are aligned with one another. Exemplary GLYAT polypeptides and the polynucleotides encoding them are known in the art and particularly disclosed, for example, in U.S. App. Publ. No. 2003/0083480, and U.S. Patent Nos. 7,462,481, 7,531,339, 7,622,641, and 7,405,074, each of which is herein incorporated by reference in
20 its entirety. In some embodiments, GLYAT polypeptides used in the presently disclosed methods and compositions comprise the amino acid sequence set forth in: SEQ ID NO: 43, 44, 45, 46, 48, or 50. In some embodiments, the GLYAT polynucleotide that encodes the GLYAT polypeptide that is used in the presently disclosed methods and compositions are set forth in SEQ ID NO: 47 or 49. As discussed in further detail elsewhere herein,
25 the use of fragments and variants of GLYAT polynucleotides and other known herbicide-tolerance polynucleotides and polypeptides encoded thereby is also encompassed by the present invention.

Active variants of SEQ ID NOS: 43, 44, 45, 46, 48, or 50 which retain glyphosate N-acetyltransferase activity include sequences which generate a similarity score of at least
30 430 using the BLOSUM62 matrix, a gap existence penalty of 11, and a gap extension

penalty of 1 when optimally aligned with any one of SEQ ID NO. Some aspects of the invention pertain to GAT polypeptides comprising an amino acid sequence that can be optimally aligned with an amino acid sequence selected from the group consisting of SEQ ID NOS: 43, 44, 45, 46, 48, and 50 to generate a similarity score of at least 440, 445, 450, 455, 460, 465, 470, 475, 480, 485, 490, 495, 500, 505, 510, 515, 520, 525, 530, 535, 540, 545, 550, 555, 560, 565, 570, 575, 580, 585, 590, 595, 600, 605, 610, 615, 620, 625, 630, 635, 640, 645, 650, 655, 660, 665, 670, 675, 680, 685, 690, 695, 700, 705, 710, 715, 720, 725, 730, 735, 740, 745, 750, 755, or 760 using the BLOSUM62 matrix, a gap existence penalty of 11, and a gap extension penalty of 1. Two sequences are “optimally aligned” when they are aligned for similarity scoring using a defined amino acid substitution matrix (e.g., BLOSUM62), gap existence penalty and gap extension penalty so as to arrive at the highest score possible for that pair of sequences.

Plants expressing GLYAT that have been treated with glyphosate contain the glyphosate metabolite N-acetylglyphosate (“NAG”). The presence of N-acetylglyphosate can serve as a diagnostic marker for the presence of an active GLYAT gene in a plant and can be evaluated by methods known in the art, for example, by mass spectrometry or by immunoassay. Generally, the level of NAG in a plant containing a GLYAT gene that has been treated with glyphosate is correlated with the activity of the GLYAT gene and the amount of glyphosate with which the plant has been treated.

Polynucleotides that encode glyphosate tolerance polypeptides that can be used in the presently disclosed methods and compositions include those that encode a glyphosate oxido-reductase enzyme as described more fully in U.S. Patent Nos. 5,776,760 and 5,463,175, which are incorporated herein by reference in their entireties for all purposes.

Other herbicides commonly used for commercial crop production include glufosinate (phosphinothricin) and acetolactate synthase (ALS) chemistry such as the sulfonylurea herbicides. Glufosinate is a broad spectrum herbicide which acts on the chloroplast glutamate synthase enzyme. Glufosinate-tolerant transgenic plants have been produced which carry the *bar* gene from *Streptomyces hygroscopicus*. The enzyme encoded by the *bar* gene has N-acetylation activity and modifies and detoxifies glufosinate. Glufosinate-tolerant plants are presently in commercial use (e.g., as sold by

Bayer under the name “Liberty Link[®]”). As described elsewhere herein, sulfonylurea herbicides inhibit growth of higher plants by blocking acetolactate synthase (ALS). Plants containing particular mutations in ALS are tolerant to the ALS herbicides including sulfonylureas.

5 In some embodiments, the herbicide tolerance polypeptide that is utilized in the presently disclosed methods and compositions is an ALS inhibitor-tolerance polypeptide. As used herein, an “ALS inhibitor-tolerance polypeptide” comprises any polypeptide which when expressed in a plant confers tolerance to at least one ALS inhibitor. A variety of ALS inhibitors are known and include, for example, sulfonylurea,
10 imidazolinone, triazolopyrimidines, pyrimidinyl(thio)benzoates, and/or sulfonylaminocarbonyltriazolinone herbicides. Additional ALS inhibitors are known and are disclosed elsewhere herein. It is known in the art that ALS mutations fall into different classes with regard to tolerance to sulfonylureas, imidazolinones, triazolopyrimidines, and pyrimidinyl(thio)benzoates, including mutations having the
15 following characteristics: (1) broad tolerance to all four of these groups; (2) tolerance to imidazolinones and pyrimidinyl(thio)benzoates; (3) tolerance to sulfonylureas and triazolopyrimidines; and (4) tolerance to sulfonylureas and imidazolinones.

 Various ALS inhibitor-tolerance polypeptides can be employed. In some embodiments, the ALS inhibitor-tolerance polynucleotides contain at least one nucleotide
20 mutation resulting in one amino acid change in the ALS polypeptide. In specific embodiments, the change occurs in one of seven substantially conserved regions of acetolactate synthase. See, for example, Hattori *et al.* (1995) *Molecular Genetics and Genomes* 246:419-425; Lee *et al.* (1998) *EMBO Journal* 7:1241-1248; Mazur *et al.* (1989) *Ann. Rev. Plant Phys.* 40:441-470; and U.S. Patent No. 5,605,011, each of which
25 is incorporated by reference in their entirety. The ALS inhibitor-tolerance polypeptide can be encoded by, for example, the SuRA or SuRB locus of ALS. In specific embodiments, the ALS inhibitor-tolerance polypeptide comprises the C3 ALS mutant, the HRA ALS mutant, the S4 mutant or the S4/HRA mutant or any combination thereof. Different mutations in ALS are known to confer tolerance to different herbicides and
30 groups (and/or subgroups) of herbicides; see, e.g., Tranel and Wright (2002) *Weed*

Science 50:700-712. See also, U.S. Patent No. 5,605,011, 5,378,824, 5,141,870, 5,013,659, and 7,622,641, each of which is herein incorporated by reference in their entirety. See also, SEQ ID NO:51 comprising a soybean HRA sequence; SEQ ID NO:52 comprising a maize HRA sequence; and SEQ ID NO:53 comprising an *Arabidopsis* HRA
5 sequence. The HRA mutation in ALS finds particular use in one embodiment of the invention. The mutation results in the production of an acetolactate synthase polypeptide which is resistant to at least one ALS inhibitor chemistry in comparison to the wild-type protein. For example, a plant expressing an ALS inhibitor-tolerant polypeptide may be tolerant of a dose of sulfonylurea, imidazolinone, triazolopyrimidines,
10 pyrimidinyloxy(thio)benzoates, and/or sulfonylaminocarbonyltriazolinone herbicide that is at least 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 50, 70, 80, 100, 125, 150, 200, 500, or 1000 times higher than a dose of the herbicide that would cause damage to an appropriate control plant. In some embodiments, an ALS inhibitor-tolerant polypeptide comprises a number of mutations.

15 In some embodiments, the ALS inhibitor-tolerance polypeptide confers tolerance to sulfonylurea and imidazolinone herbicides. Sulfonylurea and imidazolinone herbicides inhibit growth of higher plants by blocking acetolactate synthase (ALS), also known as, acetohydroxy acid synthase (AHAS). For example, plants containing particular mutations in ALS (*e.g.*, the S4 and/or HRA mutations) are tolerant to sulfonylurea herbicides. The
20 production of sulfonylurea-tolerant plants and imidazolinone-tolerant plants is described more fully in U.S. Patent Nos. 5,605,011; 5,013,659; 5,141,870; 5,767,361; 5,731,180; 5,304,732; 4,761,373; 5,331,107; 5,928,937; and 5,378,824; and international publication WO 96/33270, which are incorporated herein by reference in their entireties for all purposes. In specific embodiments, the ALS inhibitor-tolerance polypeptide comprises a
25 sulfonamide-tolerant acetolactate synthase (otherwise known as a sulfonamide-tolerant acetohydroxy acid synthase) or an imidazolinone-tolerant acetolactate synthase (otherwise known as an imidazolinone-tolerant acetohydroxy acid synthase).

Often, a herbicide-tolerance polynucleotide that confers tolerance to a particular herbicide or other chemical or a plant expressing it will also confer tolerance to other

herbicides or chemicals in the same class or subclass, for example, a class or subclass set forth in Table 3.

Table 3: Abbreviated version of HRAC Herbicide Classification

I. ALS Inhibitors (WSSA Group 2)	2. Pyriftalid
A. Sulfonyleureas	3. Pyribenzoxim
1. Azimsulfuron	4. Pyriothiac
2. Chlorimuron-ethyl	5. Pyriminobac-methyl
3. Metsulfuron-methyl	E. Imidazolinones
4. Nicosulfuron	1. Imazapyr
5. Rimsulfuron	2. Imazethapyr
6. Sulfometuron-methyl	3. Imazaquin
7. Thifensulfuron-methyl	4. Imazapic
8. Tribenuron-methyl	5. Imazamethabenz-methyl
9. Amidosulfuron	6. Imazamox
10. Bensulfuron-methyl	II. Other Herbicides--Active Ingredients/ Additional Modes of Action
11. Chlorsulfuron	A. Inhibitors of Acetyl CoA carboxylase (ACCase) (WSSA Group 1)
12. Cinosulfuron	1. Aryloxyphenoxypropionates ('FOPs')
13. Cyclosulfamuron	a. Quizalofop-P-ethyl
14. Ethametsulfuron-methyl	b. Diclofop-methyl
15. Ethoxysulfuron	c. Clodinafop-propargyl
16. Flazasulfuron	d. Fenoxaprop-P-ethyl
17. Flupyralsulfuron-methyl	e. Fluazifop-P-butyl
18. Foramsulfuron	f. Propaquizafop
19. Imazosulfuron	g. Haloxyfop-P-methyl
20. Iodosulfuron-methyl	h. Cyhalofop-butyl
21. Mesosulfuron-methyl	i. Quizalofop-P-ethyl
22. Oxasulfuron	2. Cyclohexanediones ('DIMs')
23. Primisulfuron-methyl	a. Alloxydim
24. Prosulfuron	b. Butoxydim
25. Pyrazosulfuron-ethyl	c. Clethodim
26. Sulfosulfuron	d. Cycloxydim
27. Triasulfuron	e. Sethoxydim
28. Trifloxysulfuron	f. Tepraloxydim
29. Triflusaluron-methyl	g. Tralkoxydim
30. Tritosulfuron	B. Inhibitors of Photosystem II—HRAC Group C1/ WSSA Group 5
31. Halosulfuron-methyl	1. Triazines
32. Flucetosulfuron	a. Ametryne
B. Sulfonyleureas	b. Atrazine
1. Flucarbazone	c. Cyanazine
2. Procarbazone	d. Desmetryne
C. Triazolopyrimidines	e. Dimethametryne
1. Cloransulam-methyl	f. Prometon
2. Flumetsulam	g. Prometryne
3. Diclosulam	h. Propazine
4. Florasulam	i. Simazine
5. Metosulam	j. Simetryne
6. Penoxsulam	k. Terbumeton
7. Pyroxsulam	
D. Pyrimidinyloxy(thio)benzoates	
1. Bispyribac	

l. Terbutylazine
m. Terbutryne
n. Trietazine
2. Triazinones
a. Hexazinone
b. Metribuzin
c. Metamitron
3. Triazolinone
a. Amicarbazone
4. Uracils
a. Bromacil
b. Lenacil
c. Terbacil
5. Pyridazinones
a. Pyrazon
6. Phenyl carbamates
a. Desmedipham
b. Phenmedipham
C. Inhibitors of Photosystem II--HRAC Group C2/WSSA Group 7
1. Ureas
a. Fluometuron
b. Linuron
c. Chlorobromuron
d. Chlorotoluron
e. Chloroxuron
f. Dimefuron
g. Diuron
h. Ethidimuron
i. Fenuron
j. Isoproturon
k. Isouron
l. Methabenzthiazuron
m. Metobromuron
n. Metoxuron
o. Monolinuron
p. Neburon
q. Siduron
r. Tebuthiuron
2. Amides
a. Propanil
b. Pentanochlor
D. Inhibitors of Photosystem II--HRAC Group C3/ WSSA Group 6
1. Nitriles
a. Bromofenoxim
b. Bromoxynil
c. Ioxynil
2. Benzothiadiazinone (Bentazon)
a. Bentazon
3. Phenylpyridazines
a. Pyridate

b. Pyridafol
E. Photosystem-I-electron diversion (Bipyridyliums) (WSSA Group 22)
1. Diquat
2. Paraquat
F. Inhibitors of PPO (protoporphyrinogen oxidase) (WSSA Group 14)
1. Diphenylethers
a. Acifluorfen-Na
b. Bifenox
c. Chlormethoxyfen
d. Fluoroglycofen-ethyl
e. Fomesafen
f. Halosafen
g. Lactofen
h. Oxyfluorfen
2. Phenylpyrazoles
a. Fluzolate
b. Pyraflufen-ethyl
3. N-phenylphthalimides
a. Cinidon-ethyl
b. Flumioxazin
c. Flumiclorac-pentyl
4. Thiadiazoles
a. Fluthiacet-methyl
b. Thidiazimin
5. Oxadiazoles
a. Oxadiazon
b. Oxadiargyl
6. Triazolinones
a. Carfentrazone-ethyl
b. Sulfentrazone
7. Oxazolidinediones
a. Pentoxazone
8. Pyrimidindiones
a. Benzfendizone
b. Butafenicil
9. Others
a. Pyrazogyl
b. Profluzol
G. Bleaching: Inhibition of carotenoid biosynthesis at the phytoene desaturase step (PDS) (WSSA Group 12)
1. Pyridazinones
a. Norflurazon
2. Pyridinecarboxamides
a. Diflufenican
b. Picolinafen
3. Others
a. Beflubutamid
b. Fluridone
c. Flurochloridone
d. Flurtamone

H. Bleaching: Inhibition of 4-hydroxyphenyl-pyruvate-dioxygenase (4-HPPD) (WSSA Group 28)
1. Triketones
a. Mesotrione
b. Sulcotrione
2. Isoxazoles
a. Isoxachlortole
b. Isoxaflutole
3. Pyrazoles
a. Benzofenap
b. Pyrazoxyfen
c. Pyrazolynate
4. Others
a. Benzobicyclon
I. Bleaching: Inhibition of carotenoid biosynthesis (unknown target) (WSSA Group 11 and 13)
1. Triazoles (WSSA Group 11)
a. Amitrole
2. Isoxazolidinones (WSSA Group 13)
a. Clomazone
3. Ureas
a. Fluometuron
3. Diphenylether
a. Aclonifen
J. Inhibition of EPSP Synthase
1. Glycines (WSSA Group 9)
a. Glyphosate
b. Sulfosate
K. Inhibition of glutamine synthetase
1. Phosphinic Acids
a. Glufosinate-ammonium
b. Bialaphos
L. Inhibition of DHP (dihydropteroate) synthase (WSSA Group 18)
1 Carbamates
a. Asulam
M. Microtubule Assembly Inhibition (WSSA Group 3)
1. Dinitroanilines
a. Benfluralin
b. Butralin
c. Dinitramine
d. Ethalfluralin
e. Oryzalin
f. Pendimethalin
g. Trifluralin
2. Phosphoroamidates
a. Amiprophos-methyl
b. Butamiphos
3. Pyridines
a. Dithiopyr

b. Thiazopyr
4. Benzamides
a. Pronamide
b. Tebutam
5. Benzenedicarboxylic acids
a. Chlorthal-dimethyl
N. Inhibition of mitosis/microtubule organization WSSA Group 23)
1. Carbamates
a. Chlorpropham
b. Propham
c. Carbetamide
O. Inhibition of cell division (Inhibition of very long chain fatty acids as proposed mechanism; WSSA Group 15)
1. Chloroacetamides
a. Acetochlor
b. Alachlor
c. Butachlor
d. Dimethachlor
e. Dimethanamid
f. Metazachlor
g. Metolachlor
h. Pethoxamid
i. Pretilachlor
j. Propachlor
k. Propisochlor
l. Thenylchlor
2. Acetamides
a. Diphenamid
b. Napropamide
c. Naproanilide
3. Oxyacetamides
a. Flufenacet
b. Mefenacet
4. Tetrazolinones
a. Fentrazamide
5. Others
a. Anilofos
b. Cafenstrole
c. Indanofan
d. Piperophos
P. Inhibition of cell wall (cellulose) synthesis
1. Nitriles (WSSA Group 20)
a. Dichlobenil
b. Chlorthiamid
2. Benzamides (isoxaben (WSSA Group 21))
a. Isoxaben
3. Triazolocarboxamides (flupoxam)
a. Flupoxam
Q. Uncoupling (membrane disruption):

(WSSA Group 24)
1. Dinitrophenols
a. DNOC
b. Dinoseb
c. Dinoterb
R. Inhibition of Lipid Synthesis by other than ACC inhibition
1. Thiocarbamates (WSSA Group 8)
a. Butylate
b. Cycloate
c. Dimepiperate
d. EPTC
e. Esprocarb
f. Molinate
g. Orbencarb
h. Pebulate
i. Prosulfocarb
j. Benthiocarb
k. Tiocarbazil
l. Triallate
m. Vernolate
2. Phosphorodithioates
a. Bensulide
3. Benzofurans
a. Benfuresate
b. Ethofumesate
4. Halogenated alkanolic acids (WSSA Group 26)
a. TCA
b. Dalapon
c. Flupropanate
S. Synthetic auxins (IAA-like) (WSSA Group 4)
1. Phenoxy-carboxylic acids
a. Clomeprop
b. 2,4-D
c. Mecoprop
2. Benzoic acids
a. Dicamba
b. Chloramben

c. TBA
3. Pyridine carboxylic acids
a. Clopyralid
b. Fluroxypyr
c. Picloram
d. Tricyclopyr
4. Quinoline carboxylic acids
a. Quinclorac
b. Quinmerac
5. Others (benazolin-ethyl)
a. Benazolin-ethyl
T. Inhibition of Auxin Transport
1. Phthalamates; semicarbazones (WSSA Group 19)
a. Naptalam
b. Diflufenzopyr-Na
U. Other Mechanism of Action
1. Arylamino-propionic acids
a. Flamprop-M-methyl /-isopropyl
2. Pyrazolium
a. Difenzoquat
3. Organoarsenicals
a. DSMA
b. MSMA
4. Others
a. Bromobutide
b. Cinmethylin
c. Cumyluron
d. Dazomet
e. Daimuron-methyl
f. Dimuron
g. Etobenzanid
h. Fosamine
i. Metam
j. Oxaziclomefone
k. Oleic acid
l. Pelargonic acid
m. Pyributicarb

The presently disclosed methods and compositions can utilize multiple herbicide tolerance polynucleotides. That is, the presently disclosed polynucleotide constructs can comprise more than one coding polynucleotide for a herbicide tolerance polypeptide. In some embodiments, the polynucleotide construct comprises more than one polynucleotide that encodes the same type of herbicide tolerance polypeptide (i.e., more than one GLYAT). In other embodiments, the polynucleotide constructs comprise more than one

herbicide-tolerance coding polynucleotide, wherein each of the coding polynucleotides encodes for a distinct type of herbicide tolerance polypeptide (of a different class or subclass). In some embodiments, the polynucleotide construct comprises at least a first and a second polynucleotide encoding a herbicide tolerance polypeptide, wherein the first
5 and the second polynucleotide encodes a first and a second herbicide tolerance polypeptide that confer tolerance to a first and a second herbicide, wherein the first and second herbicide have different mechanisms of action.

In some of those embodiments wherein the presently disclosed polynucleotide constructs comprise at least two herbicide tolerance polynucleotides, at least two
10 herbicide tolerance polynucleotides are located outside of the excision cassette. In other embodiments, the polynucleotide construct comprises a herbicide tolerance polynucleotide outside of the excision cassette that becomes operably linked to its promoter upon excision of the excision cassette and a second herbicide tolerance polypeptide within the excision cassette.

15 In some embodiments, the presently disclosed methods and compositions utilize polynucleotides that confer tolerance to glyphosate and at least one ALS inhibitor herbicide. In other embodiments, the presently disclosed methods and compositions utilize polynucleotides that confer tolerance to glyphosate and at least one ALS inhibitor herbicide, as well as, tolerance to at least one additional herbicide.

20 In addition to glyphosate and ALS inhibitors, the presently disclosed polynucleotide constructs can comprise polynucleotides that encode herbicide tolerance polypeptides that confer tolerance to other types of herbicides. Such additional herbicides, include but are not limited to, an acetyl Co-A carboxylase inhibitor such as quizalofop-P-ethyl, a synthetic auxin such as quinclorac, a protoporphyrinogen oxidase (PPO) inhibitor herbicide (such as sulfentrazone), a pigment synthesis inhibitor herbicide
25 such as a hydroxyphenylpyruvate dioxygenase inhibitor (*e.g.*, mesotrione or sulcotrione), a phosphinothricin acetyltransferase or a phytoene desaturase inhibitor like diflufenican or pigment synthesis inhibitor.

In some embodiments, the presently disclosed polynucleotide constructs comprise
30 polynucleotides encoding polypeptides conferring tolerance to herbicides which inhibit

the enzyme glutamine synthase, such as phosphinothricin or glufosinate (*e.g.*, the *bar* gene or *pat* gene). Glutamine synthetase (GS) appears to be an essential enzyme necessary for the development and life of most plant cells, and inhibitors of GS are toxic to plant cells. Glufosinate herbicides have been developed based on the toxic effect due to the inhibition of GS in plants. These herbicides are non-selective; that is, they inhibit growth of all the different species of plants present. The development of plants containing an exogenous phosphinothricin acetyltransferase is described in U.S. Patent Nos. 5,969,213; 5,489,520; 5,550,318; 5,874,265; 5,919,675; 5,561,236; 5,648,477; 5,646,024; 6,177,616; and 5,879,903, which are incorporated herein by reference in their entireties for all purposes. Mutated phosphinothricin acetyltransferase having this activity are also disclosed. In certain embodiments a maize-optimized PAT gene is used. In some of these embodiments, the maize-optimized PAT gene has the sequence set forth in SEQ ID NO: 54. In some embodiments, the PAT gene is used as a selectable marker as described elsewhere herein and is present within the excision cassette.

In still other embodiments, the presently disclosed polynucleotide constructs comprise polynucleotides encoding polypeptides conferring tolerance to herbicides which inhibit protox (protoporphyrinogen oxidase). Protox is necessary for the production of chlorophyll, which is necessary for all plant survival. The protox enzyme serves as the target for a variety of herbicidal compounds. These herbicides also inhibit growth of all the different species of plants present. The development of plants containing altered protox activity which are resistant to these herbicides are described in U.S. Patent Nos. 6,288,306; 6,282,837; and 5,767,373; and international publication WO 01/12825, which are incorporated herein by reference in their entireties for all purposes.

In still other embodiments, the presently disclosed polynucleotide constructs may comprise polynucleotides encoding polypeptides involving other modes of herbicide resistance. For example, hydroxyphenylpyruvate dioxygenases are enzymes that catalyze the reaction in which para-hydroxyphenylpyruvate (HPP) is transformed into homogentisate. Molecules which inhibit this enzyme and which bind to the enzyme in order to inhibit transformation of the HPP into homogentisate are useful as herbicides. Plants more resistant to certain herbicides are described in U.S. Patent Nos. 6,245,968;

6,268,549; and 6,069,115; and international publication WO 99/23886, which are incorporated herein by reference in their entireties for all purposes. Mutated hydroxyphenylpyruvate dioxygenase having this activity are also disclosed.

In some embodiments, the methods and compositions can further comprise at least one cell proliferation factor. Expression of a cell proliferation factor, such as babyboom can enhance the transformation frequency of otherwise recalcitrant plants or plant parts. A polynucleotide encoding a cell proliferation factor can be co-transformed into a plant or plant part with the presently disclosed polynucleotide constructs. In other embodiments, the presently disclosed polynucleotide constructs comprise at least one polynucleotide encoding a cell proliferation factor. In some of these embodiments, the at least one polynucleotide encoding a cell proliferation factor is located within the excision cassette of the polynucleotide construct, such that the polynucleotide is excised when the site-specific recombinase is expressed.

As used herein, a “cell proliferation factor” is a polypeptide or a polynucleotide capable of stimulating growth of a cell or tissue, including but not limited to promoting progression through the cell cycle, inhibiting cell death, such as apoptosis, stimulating cell division, and/or stimulating embryogenesis. The polynucleotides can fall into several categories, including but not limited to, cell cycle stimulatory polynucleotides, developmental polynucleotides, anti-apoptosis polynucleotides, hormone polynucleotides, or silencing constructs targeted against cell cycle repressors or pro-apoptotic factors. The following are provided as non-limiting examples of each category and are not considered a complete list of useful polynucleotides for each category: 1) cell cycle stimulatory polynucleotides including plant viral replicase genes such as RepA, cyclins, E2F, *prolifera*, *cdc2* and *cdc25*; 2) developmental polynucleotides such as *Lec1*, *Kn1* family, *WUSCHEL*, *Zwille*, *BBM*, *Aintegumenta (ANT)*, *FUS3*, and members of the *Knotted* family, such as *Kn1*, *STM*, *OSH1*, and *SbH1*; 3) anti-apoptosis polynucleotides such as *CED9*, *Bcl2*, *Bcl-X(L)*, *Bcl-W*, *A1*, *McL-1*, *Mac1*, *Boo*, and *Bax-inhibitors*; 4) hormone polynucleotides such as *IPT*, *TZS*, and *CKI-1*; and 5) silencing constructs targeted against cell cycle repressors, such as *Rb*, *CK1*, *prohibitin*, and *wee1*, or stimulators of apoptosis such as *APAF-1*, *bad*, *bax*, *CED-4*, and *caspase-3*, and repressors of plant developmental

transitions, such as Pickle and WD polycomb genes including FIE and Medea. The polynucleotides can be silenced by any known method such as antisense, RNA interference, cosuppression, chimerplasty, or transposon insertion.

The polynucleotide encoding the cell proliferation factor may be native to the cell or heterologous. Any of a number of cell proliferation factors can be used. In certain embodiments, those cell proliferation factors that are capable of stimulating embryogenesis are used to enhance transformation efficiency. Such cell proliferation factors are referred to herein as embryogenesis-stimulating polypeptides and they include, but are not limited to, babyboom polypeptides.

In some embodiments, the cell proliferation factor is a member of the AP2/ERF family of proteins. The AP2/ERF family of proteins is a plant-specific class of putative transcription factors that regulate a wide variety of developmental processes and are characterized by the presence of an AP2 DNA binding domain that is predicted to form an amphipathic alpha helix that binds DNA (PFAM Accession PF00847). The AP2/ERF proteins have been subdivided into distinct subfamilies based on the presence of conserved domains. Initially, the family was divided into two subfamilies based on the number of DNA binding domains, with the ERF subfamily having one DNA binding domain, and the AP2 subfamily having 2 DNA binding domains. As more sequences were identified, the family was subsequently subdivided into five subfamilies: AP2, DREB, ERF, RAV, and others. (Sakuma *et al.* (2002) *Biochem Biophys Res Comm* 290:998-1009).

Members of the APETALA2 (AP2) family of proteins function in a variety of biological events, including but not limited to, development, plant regeneration, cell division, embryogenesis, and cell proliferation (see, *e.g.*, Riechmann and Meyerowitz (1998) *Biol Chem* 379:633-646; Saleh and Pagés (2003) *Genetika* 35:37-50 and Database of Arabidopsis Transcription Factors at daft.cbi.pku.edu.cn). The AP2 family includes, but is not limited to, AP2, ANT, Glossy15, AtBBM, BnBBM, and maize ODP2/BBM.

U.S. Application Publication No. 2011/0167516, which is herein incorporated by reference in its entirety, describes an analysis of fifty sequences with homology to a maize BBM sequence (also referred to as maize ODP2 or ZmODP2, the polynucleotide and

amino acid sequence of the maize BBM is set forth in SEQ ID NO: 55 and 56, respectively; the polynucleotide and amino acid sequence of another ZmBBM is set forth in SEQ ID NO: 58 and 59, respectively). The analysis identified three motifs (motifs 4-6; set forth in SEQ ID NOs: 61-63), along with the AP2 domains (motifs 2 and 3; SEQ ID NOs: 64 and 65) and linker sequence that bridges the AP2 domains (motif 1; SEQ ID NO: 66), that are found in all of the BBM homologues. Thus, motifs 1-6 distinguish these BBM homologues from other AP2-domain containing proteins (*e.g.*, WRI, AP2, and RAP2.7) and these BBM homologues comprise a subgroup of AP2 family of proteins referred to herein as the BBM/PLT subgroup. In some embodiments, the cell proliferation factor that is used in the methods and compositions is a member of the BBM/PLT group of AP2 domain-containing polypeptides. In these embodiments, the cell proliferation factor comprises two AP2 domains and motifs 4-6 (SEQ ID NOs: 61-63) or a fragment or variant thereof. In some of these embodiments, the AP2 domains have the sequence set forth in SEQ ID NOs: 64 and 65 or a fragment or variant thereof, and in particular embodiments, further comprises the linker sequence of SEQ ID NO: 66 or a fragment or variant thereof. In other embodiments, the cell proliferation factor comprises at least one of motifs 4-6 or a fragment or variant thereof, along with two AP2 domains, which in some embodiments have the sequence set forth in SEQ ID NO: 64 and/or 65 or a fragment or variant thereof, and in particular embodiments have the linker sequence of SEQ ID NO: 66 or a fragment or variant thereof. Based on the phylogenetic analysis, the subgroup of BBM/PLT polypeptides can be subdivided into the BBM, AIL6/7, PLT1/2, AIL1, PLT3, and ANT groups of polypeptides.

In some embodiments, the cell proliferation factor is a babyboom (BBM) polypeptide, which is a member of the AP2 family of transcription factors. The BBM protein from *Arabidopsis* (AtBBM) is preferentially expressed in the developing embryo and seeds and has been shown to play a central role in regulating embryo-specific pathways. Overexpression of AtBBM has been shown to induce spontaneous formation of somatic embryos and cotyledon-like structures on seedlings. See, Boutilier *et al.* (2002) *The Plant Cell* 14:1737-1749. The maize BBM protein also induces embryogenesis and promotes transformation (See, U.S. Patent No. 7,579,529, which is herein incorporated by

reference in its entirety). Thus, BBM polypeptides stimulate proliferation, induce embryogenesis, enhance the regenerative capacity of a plant, enhance transformation, and as demonstrated herein, enhance rates of targeted polynucleotide modification.

In some embodiments, the babyboom polypeptide comprises two AP2 domains
 5 and at least one of motifs 7 and 10 (set forth in SEQ ID NO: 67 and 68, respectively) or a variant or fragment thereof. In certain embodiments, the AP2 domains are motifs 2 and 3 (SEQ ID NOs: 64 and 65, respectively) or a fragment or variant thereof, and in particular embodiments, the babyboom polypeptide further comprises a linker sequence between AP2 domain 1 and 2 having motif 1 (SEQ ID NO: 66) or a fragment or variant thereof. In
 10 particular embodiments, the BBM polypeptide further comprises motifs 4-6 (SEQ ID NOs 61-63) or a fragment or variant thereof. The BBM polypeptide can further comprise motifs 8 and 9 (SEQ ID NOs: 69 and 70, respectively) or a fragment or variant thereof, and in some embodiments, motif 10 (SEQ ID NO: 68) or a variant or fragment thereof. In some of these embodiments, the BBM polypeptide also comprises at least one of motif
 15 14 (set forth in SEQ ID NO: 71), motif 15 (set forth in SEQ ID NO: 72), and motif 19 (set forth in SEQ ID NO: 73), or variants or fragments thereof. The variant of a particular amino acid motif can be an amino acid sequence having at least about 40%, 50%, 60%, 70%, 75%, 80%, 85%, 90%, 95%, 96%, 97%, 98%, 99%, or greater sequence identity with the motif disclosed herein. Alternatively, variants of a particular amino acid motif
 20 can be an amino acid sequence that differs from the amino acid motif by 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 amino acids.

Non-limiting examples of babyboom polynucleotides and polypeptides that can be used in the methods and compositions include the *Arabidopsis thaliana* AtBBM (SEQ ID NOs: 74 and 75), *Brassica napus* BnBBM1 (SEQ ID NOs: 76 and 77), *Brassica napus*
 25 BnBBM2 (SEQ ID NOs: 78 and 79), *Medicago truncatula* MtBBM (SEQ ID NOs: 80 and 81), *Glycine max* GmBBM (SEQ ID NOs: 82 and 83), *Vitis vinifera* VvBBM (SEQ ID NOs: 84 and 85), *Zea mays* ZmBBM (SEQ ID NOs: 55 and 56 and genomic sequence set forth in SEQ ID NO: 57; or SEQ ID NOs: 58 and 59 and genomic sequence set forth in SEQ ID NO: 60) and ZmBBM2 (SEQ ID NOs: 101 and 102), *Oryza sativa* OsBBM
 30 (polynucleotide sequences set forth in SEQ ID NOs: 86 and 87; amino acid sequence set

forth in SEQ ID NO: 89; and genomic sequence set forth in SEQ ID NO: 88), OsBBM1 (SEQ ID NOs: 90 and 91), OsBBM2 (SEQ ID NOs: 92 and 93), and OsBBM3 (SEQ ID NOs: 94 and 95), *Sorghum bicolor* SbBBM (SEQ ID NOs: 96 and 97 and genomic sequence set forth in SEQ ID NO: 98) and SbBBM2 (SEQ ID NOs: 99 and 100) or active fragments or variants thereof. In particular embodiments, the cell proliferation factor is a maize BBM polypeptide (SEQ ID NO: 56, 59, or 102) or a variant or fragment thereof, or is encoded by a maize BBM polynucleotide (SEQ ID NO: 55, 57, 121, 116, or 101) or a variant or fragment thereof.

Thus, in some embodiments, a polynucleotide encoding a cell proliferation factor has a nucleotide sequence having at least 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity to the nucleotide sequence set forth in SEQ ID NO: 82, 96, 84, 80, 55, 101, 86, 90, 92, 94, 74, 76, 78, 99, 57, 60, 88, 87, 58, or 98 or the cell proliferation factor has an amino acid sequence having at least 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity to the amino acid sequence set forth in SEQ ID NO: 83, 97, 85, 81, 56, 102, 89, 91, 93, 95, 75, 77, 79, 59, or 100. In some of these embodiments, the cell proliferation factor has at least one of motifs 7 and 10 (SEQ ID NO: 67 and 68, respectively) or a variant or fragment thereof at the corresponding amino acid residue positions in the babyboom polypeptide. In other embodiments, the cell proliferation factor further comprises at least one of motif 14 (set forth in SEQ ID NO: 71), motif 15 (set forth in SEQ ID NO: 72), and motif 19 (set forth in SEQ ID NO: 73) or a variant or fragment thereof at the corresponding amino acid residue positions in the babyboom polypeptide.

In other embodiments, other cell proliferation factors, such as, Lec1, Kn1 family, WUSCHEL (*e.g.*, WUS1, the polynucleotide and amino acid sequence of which is set forth in SEQ ID NO: 103 and 104; WUS2, the polynucleotide and amino acid sequence of which is set forth in SEQ ID NO: 105 and 106; WUS2 alt, the polynucleotide and amino acid sequence of which is set forth in SEQ ID NO: 107 and 108; WUS3, the polynucleotide and amino acid sequence of which is set forth in SEQ ID NO: 109 and 110), Zwillie, and Aintegumeta (ANT), may be used alone, or in combination with a

babyboom polypeptide or other cell proliferation factor. See, for example, U.S. Application Publication No. 2003/0135889, International Application Publication No. WO 03/001902, and U.S. Patent No. 6,512,165, each of which is herein incorporated by reference.

5 In some embodiments, the polynucleotide construct comprises a polynucleotide encoding a Wuschel polypeptide (see International Application Publication No. WO 01/23575 and U.S. Patent No. 7,256,322, each of which are herein incorporated by reference in its entirety). In certain embodiments, the polynucleotide encoding the Wuschel polypeptide has the sequence set forth in SEQ ID NO: 103, 105, 107, or 109
10 (WUS1, WUS2, WUS2 alt, or WUS3, respectively) or an active variant or fragment thereof. In particular embodiments, the Wuschel polypeptide has the sequence set forth in SEQ ID NO: 104, 106, 108, or 110 (WUS1, WUS2, WUS2 alt, or WUS3, respectively) or an active variant or fragment thereof. In some of these embodiments, the polynucleotide encoding a Wuschel polypeptide is operably linked to a promoter active in
15 the plant, including but not limited to the maize In2-2 promoter or a nopaline synthase promoter.

 When multiple cell proliferation factors are used, or when a babyboom polypeptide is used along with any of the abovementioned polypeptides, the polynucleotides encoding each of the factors can be present on the same expression
20 cassette or on separate expression cassettes. When two or more factors are coded for by separate expression cassettes, the expression cassettes can be provided to the plant simultaneously or sequentially. In some embodiments, the polynucleotide construct comprises a polynucleotide encoding a babyboom polypeptide and a polynucleotide encoding a Wuschel polypeptide within the excision cassette such that the cell
25 proliferation factors enhance the transformation frequency of the polynucleotide construct, but are subsequently excised upon desiccation of the transformed plant cell/tissue.

 In some embodiments, herbicide tolerance polynucleotides can serve as a selectable marker for the identification of plants or plant parts that further comprise a
30 polynucleotide of interest. Thus, in certain embodiments, the presently disclosed

polynucleotide constructs can further comprise a polynucleotide of interest. In some embodiments, the polynucleotide of interest is operably linked to a promoter that is active in a plant cell. The promoter that is operably linked to the polynucleotide of interest can be a constitutive promoter, an inducible promoter, or a tissue-preferred promoter.

5 In certain embodiments, the polynucleotide of interest, and optionally the operably linked promoter, are located outside of the excision cassette on the polynucleotide construct. In other embodiments, the polynucleotide of interest and optionally its operably linked promoter are located within the excision cassette and the herbicide tolerance polynucleotide serves as a selectable marker to identify those plants or plant
10 parts from which the polynucleotide of interest has been excised.

 The polynucleotide of interest may impart various changes in the organism, particularly plants, including, but not limited to, modification of the fatty acid composition in the plant, altering the amino acid content of the plant, altering pathogen resistance, and the like. These results can be achieved by providing expression of
15 heterologous products, increased expression of endogenous products in plants, or suppressed expression of endogenous products in plants.

 General categories of polynucleotides of interest include, for example, those genes involved in information, such as zinc fingers, those involved in communication, such as kinases, those involved in biosynthetic pathways, and those involved in housekeeping,
20 such as heat shock proteins. More specific categories of transgenes, for example, include sequences encoding important traits for agronomics, insect resistance, disease resistance, sterility, grain characteristics, oil, starch, carbohydrate, phytate, protein, nutrient, metabolism, digestability, kernel size, sucrose loading, and commercial products.

 Traits such as oil, starch, and protein content can be genetically altered in addition
25 to using traditional breeding methods. Modifications include increasing content of oleic acid, saturated and unsaturated oils, increasing levels of lysine and sulfur, providing essential amino acids, and also modification of starch. Protein modifications to alter amino acid levels are described in U.S. Patent Nos. 5,703,049, 5,885,801, 5,885,802, and 5,990,389 and WO 98/20122, herein incorporated by reference.

Insect resistance genes may encode resistance to pests such as rootworm, cutworm, European Corn Borer, and the like. Such genes include, for example, *Bacillus thuringiensis* toxic protein genes (U.S. Patent Nos. 5,366,892; 5,747,450; 5,737,514; 5,723,756; 5,593,881; and Geiser *et al.* (1986) *Gene* 48:109); lectins (Van Damme *et al.* (1994) *Plant Mol. Biol.* 24:825); and the like.

Genes encoding disease resistance traits include detoxification genes, such as against fumonisin (U.S. Patent No. 5,792,931); avirulence (avr) and disease resistance (R) genes (Jones *et al.* (1994) *Science* 266:789; Martin *et al.* (1993) *Science* 262:1432; and Mindrinos *et al.* (1994) *Cell* 78:1089); and the like.

10 Sterility genes can also be encoded in an expression cassette and provide an alternative to physical detasseling. Examples of genes used in such ways include male tissue-preferred genes and genes with male sterility phenotypes such as QM, described in U.S. Patent No. 5,583,210. Other genes include kinases and those encoding compounds toxic to either male or female gametophytic development.

15 Commercial traits can also be encoded on a gene or genes that could, for example increase starch for ethanol production, or provide expression of proteins.

Although the herbicide tolerance polynucleotide can serve as a selectable marker to aid in the identification of transgenic plants that comprise a polynucleotide of interest or lack a polynucleotide of interest, an additional selectable marker may be present in the
20 excision cassette of the presently disclosed polynucleotide constructs that aids in the selection of transgenic plants or plant parts at an earlier point in development when most herbicide selection systems are less efficient. In general, the selectable marker that is present within the excision cassette is one that allows for efficient selection in early stages of plant development and production (e.g., during the tissue proliferation stage of
25 transgenic plant production). For example, the expression of a fluorescent protein can be used to select plants or plant parts that comprise a presently disclosed polynucleotide construct during or prior to tissue proliferation. Proliferating the tissue to a certain mass is generally necessary before regeneration of the tissue into a plant. The expression of the site-specific recombinase is then induced before herbicide selection, which in general,
30 occurs during or after the regeneration of the provided cells or tissues into plants.

“Regenerating” or “regeneration” of a plant cell is the process of growing a plant from the plant cell (e.g., plant protoplast, callus or explant).

Marker genes that can be present within the excision cassette include polynucleotides encoding products that provide resistance against otherwise toxic compounds (e.g. antibiotic resistance) such as those encoding neomycin phosphotransferase II (NEO or nptII) and hygromycin phosphotransferase (HPT), as well as genes conferring resistance to herbicidal compounds, such as glufosinate ammonium, bromoxynil, imidazolinones, and 2,4-dichlorophenoxyacetate (2,4-D), including but not limited to, the selectable marker gene phosphinothricin acetyl transferase (PAT) (Wohlleben *et al.* (1988) *Gene* 70:25-37), which confers resistance to the herbicide Bialaphos. In certain embodiments, the selectable marker that is present within the excision cassette is not a herbicide tolerance polynucleotide.

As used herein, “antibiotic resistance polypeptide” refers to a polypeptide that confers resistance or tolerance to an antibiotic compound to a host cell comprising or secreting the polypeptide.

Additional selectable marker-encoding polynucleotides include those that encode products that can be readily identified, including but not limited to phenotypic markers such as β -galactosidase, and visual markers, such as fluorescent proteins. As used herein, a “fluorescent protein” or “fluorescent polypeptide” refers to a polypeptide that is capable of absorbing radiation (e.g., light at a wavelength in the visible spectrum) at one wavelength and emitting radiation as light at a different wavelength. Non-limiting examples of fluorescent protein include green fluorescent protein (GFP) (Su *et al.* (2004) *Biotechnol Bioeng* 85:610-9 and Fetter *et al.* (2004) *Plant Cell* 16:215-28), cyan fluorescent protein (CYP) (Bolte *et al.* (2004) *J. Cell Science* 117:943-54 and Kato *et al.* (2002) *Plant Physiol* 129:913-42), red fluorescent protein, and yellow fluorescent protein (PhiYFP™ from Evrogen, see, Bolte *et al.* (2004) *J. Cell Science* 117:943-54). For additional selectable markers, see generally, Yarranton (1992) *Curr. Opin. Biotech.* 3:506-511; Christopherson *et al.* (1992) *Proc. Natl. Acad. Sci. USA* 89:6314-6318; Yao *et al.* (1992) *Cell* 71:63-72; Reznikoff (1992) *Mol. Microbiol.* 6:2419-2422; Barkley *et al.* (1980) in *The Operon*, pp. 177-220; Hu *et al.* (1987) *Cell* 48:555-566; Brown *et al.* (1987) *Cell*

49:603-612; Figge *et al.* (1988) *Cell* 52:713-722; Deuschle *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86:5400-5404; Fuerst *et al.* (1989) *Proc. Natl. Acad. Sci. USA* 86:2549-2553; Deuschle *et al.* (1990) *Science* 248:480-483; Gossen (1993) Ph.D. Thesis, University of Heidelberg; Reines *et al.* (1993) *Proc. Natl. Acad. Sci. USA* 90:1917-1921; Labow *et al.* (1990) *Mol. Cell. Biol.* 10:3343-3356; Zambretti *et al.* (1992) *Proc. Natl. Acad. Sci. USA* 89:3952-3956; Baim *et al.* (1991) *Proc. Natl. Acad. Sci. USA* 88:5072-5076; Wyborski *et al.* (1991) *Nucleic Acids Res.* 19:4647-4653; Hillenand-Wissman (1989) *Topics Mol. Struc. Biol.* 10:143-162; Degenkolb *et al.* (1991) *Antimicrob. Agents Chemother.* 35:1591-1595; Kleinschmidt *et al.* (1988) *Biochemistry* 27:1094-1104; Bonin (1993) Ph.D. Thesis, University of Heidelberg; Gossen *et al.* (1992) *Proc. Natl. Acad. Sci. USA* 89:5547-5551; Oliva *et al.* (1992) *Antimicrob. Agents Chemother.* 36:913-919; Hlavka *et al.* (1985) *Handbook of Experimental Pharmacology*, Vol. 78 (Springer-Verlag, Berlin); Gill *et al.* (1988) *Nature* 334:721-724. Such disclosures are herein incorporated by reference.

The presently provided methods and compositions can also utilize metabolic enzymes as selectable markers. The term “metabolic enzymes” as it relates to selectable markers refer to enzymes that confer a selectable metabolic advantage to cells. Cells expressing the metabolic enzyme are then positively selected for the ability to metabolize and utilize a particular chemical compound that cannot otherwise be metabolized or utilized by other cells not comprising the enzyme. Non-limiting examples of metabolic enzymes for use as selectable markers include D-amino oxidase (encoded by the *doaI* gene), which catalyzes the oxidative deamination of various D-amino acids (see, for example, Erikson *et al.* (2004) *Nature Biotechnology* 22:455-458, which is herein incorporated by reference in its entirety); cyanamide hydratase (encoded by the *cah* gene), which converts cyanamide into urea as a fertilizer source (see, for example, U.S. Patent No. 6,268,547, which is herein incorporated by reference in its entirety); and phosphomannose isomerase (encoded by the *pmi* gene), which catalyzes the reversible inter-conversion of mannose-6-phosphate and fructose-6-phosphate, allowing plant cells to utilize mannose as a carbon source (see, for example, Joersbo *et al.* (1998) *Molecular Breeding* 4:11-117, which is herein incorporated by reference in its entirety).

In some embodiments, the excision cassette comprises more than one selectable marker-coding polynucleotide. In some of these embodiments, the excision cassette comprises both a visual marker and an antibiotic resistance or herbicidal resistance selectable marker. In some of these embodiments, the excision cassette comprises a
5 maize optimized PAT-coding polynucleotide (such as the sequence set forth in SEQ ID NO: 54) or a polynucleotide encoding neomycin phosphotransferase II (NEO or nptII), and a polynucleotide encoding a fluorescent protein, such as yellow fluorescent protein.

The selectable marker-encoding polynucleotide within the excision cassette is operably linked to a promoter that is active in a plant cell. This promoter can be present
10 within or outside of the excision cassette. In some of the embodiments wherein the promoter that is operably linked to the selectable marker-encoding polynucleotide is outside of the excision cassette, this same promoter will become operably linked to the herbicide tolerance polynucleotide after excision of the excision cassette.

In certain embodiments, the promoter that is operably linked to the selectable
15 marker-encoding polynucleotide present within the excision cassette is a constitutive promoter such that the selectable marker will be constitutively expressed in the plant or plant part until excision of the excision cassette. In some of these embodiments, the constitutive promoter is a maize ubiquitin promoter, which in some embodiments comprises the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin
20 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBI1ZM INTRON1; SEQ ID NO: 113).

During the selection of the plant or plant part that expresses the selectable marker that is found within the excision cassette, the plant or plant part can be cultured in the presence of a selection agent. As used herein, a “selection agent” refers to a compound
25 that when contacted with a plant or plant part allows for the identification of a plant or plant part expressing a selectable marker, either positively or negatively. For example, a selection agent for an antibiotic resistance polynucleotide is the antibiotic to which the polynucleotide confers resistance. As a further non-limiting example, a selection agent for a metabolizing enzyme selectable marker is the compound that can only be
30 metabolized and utilized by the cell that expresses the selectable marker.

In particular embodiments wherein the polynucleotide construct is designed for transformation of maize, the polynucleotide construct comprises, outside of the excision cassette, the expression cassettes for a GLYAT polypeptide and an ALS-inhibitor tolerance polypeptide as present in the T-DNA region of plasmid PHP24279 described in U.S. Patent No. 7,928,296, which is herein incorporated by reference in its entirety. In these embodiments, the polynucleotide construct comprises the *glyat4621* gene that was derived from the soil bacterium *Bacillus licheniformis* and was synthesized by a gene shuffling process to optimize the acetyltransferase activity of the GLYAT4621 enzyme (Castle *et al.* (2004) *Science* 304:1151-1154). The polynucleotide construct further comprises a ZM-HRA expression cassette comprising a modified maize acetolactate synthase gene, *zm-hra* (*Zea mays*-highly resistant allele), encoding the ZM-HRA protein, which confers tolerance to a range of ALS-inhibiting herbicides, such as sulfonylureas. In these embodiments, the *glyat4621* gene cassette and the *zm-hra* gene cassette are in reverse orientation. The expression of the *glyat4621* gene is controlled by the ubiquitin regulatory region from maize (*ubiZM1* promoter (SEQ ID NO: 111), 5'UTR (SEQ ID NO: 112), and intron (SEQ ID NO: 112) (Christensen *et al.* (1992)) and the *pinII* terminator (An *et al.* (1989) *Plant Cell* 1:115-122). The expression of the *zm-hra* gene is controlled by the native maize acetolactate synthase promoter (*zm-als* promoter) (Fang *et al.* (2000)). The terminator for the *zm-hra* gene is the 3' terminator sequence from the proteinase inhibitor II gene of *Solanum tuberosum* (*pinII* terminator). Upstream of both cassettes are three copies of the enhancer region from the cauliflower mosaic virus (CaMV 35S enhancer, US Application No. 11/508,045, herein incorporated by reference) providing expression enhancement to both cassettes.

In certain embodiments wherein the polynucleotide construct is designed for transformation of soybean (*Glycine max*), the polynucleotide construct comprises, outside of the excision cassette, the expression cassettes for a GLYAT polypeptide and an ALS-inhibitor tolerance polypeptide as present in the *Not I-Asc I* fragment of plasmid PHP20163 described in U.S. Patent No. 7,622,641, which is herein incorporated by reference in its entirety. In these embodiments, the polynucleotide construct comprises the glyphosate acetyltransferase (*glyat*) gene derived from *Bacillus licheniformis* and a

modified version of the soybean acetolactate synthase gene (*zm-hra*). The *glyat* gene was functionally improved by a gene shuffling process to optimize the kinetics of glyphosate acetyltransferase (GLYAT) activity for acetylating the herbicide glyphosate. The *glyat* gene is under the control of the SCP1 promoter and Tobacco Mosaic Virus (TMV) omega
5 5' UTR translational enhancer element and the proteinase inhibitor II (*pinII*) terminator from *Solanum tuberosum*. The *zm-hra* gene is under the control of the S-adenosyl-L-methionine synthetase (SAMS) promoter and the acetolactate synthase (*gm-als*) terminator, both from soybean.

In other embodiments wherein the polynucleotide construct is designed for
10 transformation of *Brassica*, the polynucleotide construct comprises the expression cassette for a GLYAT polypeptide as present in the plasmid PHP28181 described in U.S. Appl. Publ. No. 2012/0131692, which is herein incorporated by reference in its entirety. In these embodiments, the polynucleotide construct comprises the *glyat4621* gene, which was derived from the soil bacterium *Bacillus licheniformis* and was synthesized by a gene
15 shuffling process to optimize the acetyltransferase activity of the GLYAT4621 enzyme (Castle, *et al.*, (2004) *Science* 304:1151-1154). The expression of the *glyat4621* gene is controlled by the UBQ10 regulatory region from *Arabidopsis* and the *pinII* terminator. In some of these embodiments, the polynucleotide construct further comprises an expression cassette for an ALS inhibitor tolerance polypeptide.

20 The presently disclosed compositions and methods can utilize fragments or variants of known polynucleotide or polypeptide sequences. By "fragment" is intended a portion of the polynucleotide or a portion of an amino acid sequence and hence protein encoded thereby. Fragments of a polynucleotide may retain the biological activity of the native polynucleotide and, for example, have promoter activity (promoter fragments), or
25 are capable of stimulating proliferation, inducing embryogenesis, modifying the regenerative capacity of a plant (cell proliferation factor fragments), are capable of conferring herbicide tolerance (herbicide tolerance polypeptide fragments) or catalyzing site-specific recombination (site-specific recombinase fragments). In those embodiments wherein the polynucleotide encodes a polypeptide, fragments of the polynucleotide may
30 encode protein fragments that retain the biological activity of the native protein.

Alternatively, fragments of a polynucleotide that are useful as hybridization probes generally do not retain biological activity or encode fragment proteins that retain biological activity. Thus, fragments of a nucleotide sequence may range from at least about 20, 50, 100, 150, 200, 250, 300, 400, 500 nucleotides, or greater.

5 A fragment of a polynucleotide that encodes a biologically active portion of a cell proliferation factor, for example, will encode at least 15, 25, 30, 50, 100, 150, 200, 250, 300, 400, 500 contiguous amino acids, or up to the total number of amino acids present in the full-length cell proliferation factor. Fragments of a coding polynucleotide that are useful as hybridization probes or PCR primers generally need not encode a biologically
10 active portion of a polypeptide.

 "Variants" is intended to mean substantially similar sequences. For polynucleotides, a variant comprises a polynucleotide having deletions at the 5' and/or 3' end; deletion and/or addition of one or more nucleotides at one or more internal sites in the native polynucleotide; and/or substitution of one or more nucleotides at one or more
15 sites in the native polynucleotide. As used herein, a "native" polynucleotide or polypeptide comprises a naturally occurring nucleotide sequence or amino acid sequence, respectively. For polynucleotides encoding polypeptides conservative variants include those sequences that, because of the degeneracy of the genetic code, encode the amino acid sequence the polypeptide (*e.g.*, cell proliferation factor). Naturally occurring
20 variants such as these can be identified with the use of well-known molecular biology techniques, such as, for example, with polymerase chain reaction (PCR) and hybridization techniques. Variant polynucleotides also include synthetically derived polynucleotides, such as those generated, for example, by using site-directed mutagenesis. Generally, variants of a particular will have at least about 40%, 45%, 50%, 55%, 60%,
25 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity to that particular polynucleotide as determined by sequence alignment programs and parameters.

 Variants of a particular polynucleotide that encodes a polypeptide can also be evaluated by comparison of the percent sequence identity between the polypeptide
30 encoded by a variant polynucleotide and the polypeptide encoded by the particular

polynucleotide. Percent sequence identity between any two polypeptides can be calculated using sequence alignment programs and parameters. Where any given pair of polynucleotides is evaluated by comparison of the percent sequence identity shared by the two polypeptides they encode, the percent sequence identity between the two encoded
5 polypeptides is at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity.

"Variant" protein is intended to mean a protein derived from the native protein by deletion of one or more amino acids at the N-terminal and/or C-terminal end of the native protein; deletion and/or addition of one or more amino acids at one or more internal sites
10 in the native protein; and/or substitution of one or more amino acids at one or more sites in the native protein. Variant proteins retain the desired biological activity of the native protein. For example, variant cell proliferation factors stimulate proliferation and variant babyboom polypeptides are capable of stimulating proliferation, inducing embryogenesis, modifying the regenerative capacity of a plant, increasing the transformation efficiency in
15 a plant, increasing or maintaining the yield in a plant under abiotic stress, producing asexually derived embryos in a plant, and/or enhancing rates of targeted polynucleotide modification. Such variants may result from, for example, genetic polymorphism or from human manipulation. Biologically active variants of a native protein will have at least about 40%, 45%, 50%, 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, 91%, 92%, 93%,
20 94%, 95%, 96%, 97%, 98%, 99% or more sequence identity to the amino acid sequence for the native protein as determined by sequence alignment programs and parameters. A biologically active variant of a native protein may differ from that protein by as few as 1-15 amino acid residues, as few as 1-10, such as 6-10, as few as 5, as few as 4, 3, 2, or even 1 amino acid residue.

25 Where appropriate, the coding polynucleotides may be optimized for increased expression in the transformed plant. That is, the coding polynucleotides can be synthesized using plant-preferred codons for improved expression. See, for example, Campbell and Gowri (1990) *Plant Physiol.* 92:1-11 for a discussion of host-preferred codon usage. Methods are available in the art for synthesizing plant-preferred genes.

See, for example, U.S. Patent Nos. 5,380,831, and 5,436,391, and Murray *et al.* (1989) *Nucleic Acids Res.* 17:477-498, herein incorporated by reference.

Additional sequence modifications are known to enhance gene expression in a cellular host. These include elimination of sequences encoding spurious polyadenylation signals, exon-intron splice site signals, transposon-like repeats, and other such well-
5 characterized sequences that may be deleterious to gene expression. The G-C content of the sequence may be adjusted to levels average for a given cellular host, as calculated by reference to known genes expressed in the host cell. When possible, the sequence is modified to avoid predicted hairpin secondary mRNA structures.

10 The following terms are used to describe the sequence relationships between two or more polynucleotides or polypeptides: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", and, (d) "percentage of sequence identity."

(a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison. A reference sequence may be a subset or the entirety of a
15 specified sequence; for example, as a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.

(b) As used herein, "comparison window" makes reference to a contiguous and specified segment of a polynucleotide sequence, wherein the polynucleotide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) compared to
20 the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two polynucleotides. Generally, the comparison window is at least 20 contiguous nucleotides in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide sequence a gap penalty is typically introduced and
25 is subtracted from the number of matches.

Methods of alignment of sequences for comparison are well known in the art. Thus, the determination of percent sequence identity between any two sequences can be accomplished using a mathematical algorithm. Non-limiting examples of such mathematical algorithms are the algorithm of Myers and Miller (1988) *CABIOS* 4:11-17;
30 the local alignment algorithm of Smith *et al.* (1981) *Adv. Appl. Math.* 2:482; the global

alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453; the search-for-local alignment method of Pearson and Lipman (1988) *Proc. Natl. Acad. Sci.* 85:2444-2448; the algorithm of Karlin and Altschul (1990) *Proc. Natl. Acad. Sci. USA* 87:2264, modified as in Karlin and Altschul (1993) *Proc. Natl. Acad. Sci. USA* 90:5873-5877.

Computer implementations of these mathematical algorithms can be utilized for comparison of sequences to determine sequence identity. Such implementations include, but are not limited to: CLUSTAL in the PC/Gene program (available from Intelligenetics, Mountain View, California); the ALIGN program (Version 2.0) and GAP, BESTFIT, BLAST, FASTA, and TFASTA in the GCG Wisconsin Genetics Software Package, Version 10 (available from Accelrys Inc., 9685 Scranton Road, San Diego, California, USA). Alignments using these programs can be performed using the default parameters. The CLUSTAL program is well described by Higgins *et al.* (1988) *Gene* 73:237-244 (1988); Higgins *et al.* (1989) *CABIOS* 5:151-153; Corpet *et al.* (1988) *Nucleic Acids Res.* 16:10881-90; Huang *et al.* (1992) *CABIOS* 8:155-65; and Pearson *et al.* (1994) *Meth. Mol. Biol.* 24:307-331. The ALIGN program is based on the algorithm of Myers and Miller (1988) *supra*. A PAM120 weight residue table, a gap length penalty of 12, and a gap penalty of 4 can be used with the ALIGN program when comparing amino acid sequences. The BLAST programs of Altschul *et al.* (1990) *J. Mol. Biol.* 215:403 are based on the algorithm of Karlin and Altschul (1990) *supra*. BLAST nucleotide searches can be performed with the BLASTN program, score = 100, wordlength = 12, to obtain nucleotide sequences homologous to a nucleotide sequence encoding a protein of the invention. BLAST protein searches can be performed with the BLASTX program, score = 50, wordlength = 3, to obtain amino acid sequences homologous to a protein or polypeptide of the invention. To obtain gapped alignments for comparison purposes, Gapped BLAST (in BLAST 2.0) can be utilized as described in Altschul *et al.* (1997) *Nucleic Acids Res.* 25:3389. Alternatively, PSI-BLAST (in BLAST 2.0) can be used to perform an iterated search that detects distant relationships between molecules. See Altschul *et al.* (1997) *supra*. When utilizing BLAST, Gapped BLAST, PSI-BLAST, the default parameters of the respective programs (*e.g.*, BLASTN for

nucleotide sequences, BLASTX for proteins) can be used. See www.ncbi.nlm.nih.gov. Alignment may also be performed manually by inspection.

Unless otherwise stated, sequence identity/similarity values provided herein refer to the value obtained using GAP Version 10 using the following parameters: % identity and % similarity for a nucleotide sequence using GAP Weight of 50 and Length Weight of 3, and the nws gapdna.cmp scoring matrix; % identity and % similarity for an amino acid sequence using GAP Weight of 8 and Length Weight of 2, and the BLOSUM62 scoring matrix; or any equivalent program thereof. By "equivalent program" is intended any sequence comparison program that, for any two sequences in question, generates an alignment having identical nucleotide or amino acid residue matches and an identical percent sequence identity when compared to the corresponding alignment generated by GAP Version 10.

GAP uses the algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443-453, to find the alignment of two complete sequences that maximizes the number of matches and minimizes the number of gaps. GAP considers all possible alignments and gap positions and creates the alignment with the largest number of matched bases and the fewest gaps. It allows for the provision of a gap creation penalty and a gap extension penalty in units of matched bases. GAP must make a profit of gap creation penalty number of matches for each gap it inserts. If a gap extension penalty greater than zero is chosen, GAP must, in addition, make a profit for each gap inserted of the length of the gap times the gap extension penalty. Default gap creation penalty values and gap extension penalty values in Version 10 of the GCG Wisconsin Genetics Software Package for protein sequences are 8 and 2, respectively. For nucleotide sequences the default gap creation penalty is 50 while the default gap extension penalty is 3. The gap creation and gap extension penalties can be expressed as an integer selected from the group of integers consisting of from 0 to 200. Thus, for example, the gap creation and gap extension penalties can be 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 25, 30, 35, 40, 45, 50, 55, 60, 65 or greater.

GAP presents one member of the family of best alignments. There may be many members of this family, but no other member has a better quality. GAP displays four

figures of merit for alignments: Quality, Ratio, Identity, and Similarity. The Quality is the metric maximized in order to align the sequences. Ratio is the quality divided by the number of bases in the shorter segment. Percent Identity is the percent of the symbols that actually match. Percent Similarity is the percent of the symbols that are similar.

5 Symbols that are across from gaps are ignored. A similarity is scored when the scoring matrix value for a pair of symbols is greater than or equal to 0.50, the similarity threshold. The scoring matrix used in Version 10 of the GCG Wisconsin Genetics Software Package is BLOSUM62 (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

(c) As used herein, "sequence identity" or "identity" in the context of two
10 polynucleotides or polypeptide sequences makes reference to the residues in the two sequences that are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other
15 amino acid residues with similar chemical properties (*e.g.*, charge or hydrophobicity) and therefore do not change the functional properties of the molecule. When sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences that differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means
20 for making this adjustment are well known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The
25 scoring of conservative substitutions is calculated, *e.g.*, as implemented in the program PC/GENE (Intelligenetics, Mountain View, California).

(d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may
30 comprise additions or deletions (*i.e.*, gaps) as compared to the reference sequence (which

does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total
5 number of positions in the window of comparison, and multiplying the result by 100 to yield the percentage of sequence identity.

In hybridization techniques, all or part of a known polynucleotide is used as a probe that selectively hybridizes to other corresponding polynucleotides present in a population of cloned genomic DNA fragments or cDNA fragments (*i.e.*, genomic or
10 cDNA libraries) from a chosen organism. The hybridization probes may be genomic DNA fragments, cDNA fragments, RNA fragments, or other oligonucleotides, and may be labeled with a detectable group such as ^{32}P , or any other detectable marker. Thus, for example, probes for hybridization can be made by labeling synthetic oligonucleotides based on the babyboom polynucleotide. Methods for preparation of probes for
15 hybridization and for construction of cDNA and genomic libraries are generally known in the art and are disclosed in Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York).

For example, the entire coding polynucleotide, or one or more portions thereof, may be used as a probe capable of specifically hybridizing to a corresponding coding
20 polynucleotide and messenger RNAs. To achieve specific hybridization under a variety of conditions, such probes include sequences that are unique among the particular family of coding polynucleotide sequences and are optimally at least about 10 nucleotides in length, and most optimally at least about 20 nucleotides in length. Such probes may be used to amplify corresponding coding polynucleotides from a chosen plant by PCR. This
25 technique may be used to isolate additional coding sequences from a desired plant or as a diagnostic assay to determine the presence of coding sequences in a plant. Hybridization techniques include hybridization screening of plated DNA libraries (either plaques or colonies; see, for example, Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York).

Hybridization of such sequences may be carried out under stringent conditions. By "stringent conditions" or "stringent hybridization conditions" is intended conditions under which a probe will hybridize to its target sequence to a detectably greater degree than to other sequences (*e.g.*, at least 2-fold over background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences that are 100% complementary to the probe can be identified (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous probing). Generally, a probe is less than about 1000 nucleotides in length, optimally less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (*e.g.*, 10 to 50 nucleotides) and at least about 60°C for long probes (*e.g.*, greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1.0 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C. Optionally, wash buffers may comprise about 0.1% to about 1% SDS. Duration of hybridization is generally less than about 24 hours, usually about 4 to about 12 hours. The duration of the wash time will be at least a length of time sufficient to reach equilibrium.

Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_m can be approximated from the equation of Meinkoth and Wahl (1984) *Anal. Biochem.* 138:267-284: $T_m = 81.5^{\circ}\text{C} + 16.6 (\log M) + 0.41 (\%GC) - 0.61$

(% form) - 500/L; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The T_m is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe. T_m is reduced by about 1°C for each 1% of mismatching; thus, T_m , hybridization, and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with $\geq 90\%$ identity are sought, the T_m can be decreased 10°C. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4°C lower than the thermal melting point (T_m); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10°C lower than the thermal melting point (T_m); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20°C lower than the thermal melting point (T_m). Using the equation, hybridization and wash compositions, and desired T_m , those of ordinary skill will understand that variations in the stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T_m of less than 45°C (aqueous solution) or 32°C (formamide solution), it is optimal to increase the SSC concentration so that a higher temperature can be used. An extensive guide to the hybridization of nucleic acids is found in Tijssen (1993) *Laboratory Techniques in Biochemistry and Molecular Biology—Hybridization with Nucleic Acid Probes*, Part I, Chapter 2 (Elsevier, New York); and Ausubel *et al.*, eds. (1995) *Current Protocols in Molecular Biology*, Chapter 2 (Greene Publishing and Wiley-Interscience, New York). See Sambrook *et al.* (1989) *Molecular Cloning: A Laboratory Manual* (2d ed., Cold Spring Harbor Laboratory Press, Plainview, New York).

The presently disclosed polynucleotide constructs can be introduced into a host cell. By “host cell” is meant a cell, which comprises a heterologous nucleic acid sequence. Host cells may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, insect, amphibian, or mammalian cells. In some examples, host cells are

monocotyledonous or dicotyledonous plant cells. In particular embodiments, the monocotyledonous host cell is a sugarcane host cell.

An intermediate host cell may be used, for example, to increase the copy number of the cloning vector and/or to mediate transformation of a different host cell. With an
5 increased copy number, the vector containing the nucleic acid of interest can be isolated in significant quantities for introduction into the desired plant cells. In one embodiment, plant promoters that do not cause expression of the polypeptide in bacteria are employed.

Prokaryotes most frequently are represented by various strains of *E. coli*; however, other microbial strains may also be used. Commonly used prokaryotic control
10 sequences which are defined herein to include promoters for transcription initiation, optionally with an operator, along with ribosome binding sequences, include such commonly used promoters as the beta lactamase (penicillinase) and lactose (lac) promoter systems (Chang *et al.* (1977) *Nature* 198:1056), the tryptophan (trp) promoter system (Goeddel *et al.* (1980) *Nucleic Acids Res.* 8:4057) and the lambda derived P_L promoter
15 and N-gene ribosome binding site (Shimatake *et al.* (1981) *Nature* 292:128). The inclusion of selection markers in DNA vectors transfected in *E. coli* is also useful. Examples of such markers include genes specifying resistance to ampicillin, tetracycline, or chloramphenicol.

The vector is selected to allow introduction into the appropriate host cell.
20 Bacterial vectors are typically of plasmid or phage origin. Appropriate bacterial cells are infected with phage vector particles or transfected with naked phage vector DNA. If a plasmid vector is used, the bacterial cells are transfected with the plasmid vector DNA. Expression systems for expressing a protein are available using *Bacillus sp.* and *Salmonella* (Palva *et al.* (1983) *Gene* 22:229-235); Mosbach *et al.* (1983) *Nature*
25 302:543-545).

Methods are provided for regulating the expression of a herbicide tolerance polynucleotide, wherein a host cell is provided that comprises a presently disclosed polynucleotide construct and the expression of the site-specific recombinase is induced, thereby excising the excision cassette and allowing for the operable linkage of the

herbicide tolerance polynucleotide and its promoter and the expression of the herbicide tolerance polynucleotide.

Such methods allow for the delay of the expression of a herbicide tolerance polynucleotide until a point in development at which herbicide selection is more effective.

Thus, methods are further provided for selecting a herbicide tolerant plant cell, wherein a population of plant cells are provided, wherein at least one plant cell within the population comprises a presently disclosed polynucleotide construct, inducing the expression of the recombinase, and contacting the population of cells with a herbicide to which the herbicide tolerant polypeptide confers tolerance in order to select for the herbicide tolerant plant cell.

As used herein, the term “population of plant cells” may refer to any one of the following: a grouping of individual plant cells; a grouping of plant cells present within a single tissue, plant or plant part; a population of plants; a population of plant tissues either from the same plant or different plants; a population of seeds either from the same plant or different plants; or a population of plant parts either from the same plant or different plants. The provided population of plant cells, plant tissues, plants, or plant parts may be contacted with the herbicide. Alternatively, the provided population of plant cells may be cultured into a population of plant tissues or a population of plants, which is then exposed to the herbicide. Likewise, a provided population of plant seeds may be planted to produce a population of plants, which is then exposed to the herbicide.

In some embodiments, the provided population of plant cells is cultured into a population of plant tissues or plants prior to, during, or after the induction step, and the population of plant tissues or plants is then contacted with the herbicide. In some of these embodiments, the population of plant tissues is contacted with the herbicide during the regeneration of the tissues into plants or the population of plants that were regenerated from the population of plant tissues is contacted with the herbicide.

In certain embodiments, the provided population of plant cells is a population of immature or mature seeds. In some of these embodiments, the provided population of seeds is planted prior to, during, or after the induction step to produce a population of

plants, and the population of plants are contacted with the herbicide. In those
embodiments wherein the provided population of plant cells is a population of immature
seeds and the inducible promoter that regulates the expression of the site-specific
recombinase is a drought-inducible promoter, the drought-inducible promoter is activated
5 in response to the natural desiccation that occurs during the maturation of the immature
seed into a mature seed.

In other embodiments, the provided population of plant cells is a population of
plant tissues and these plant tissues are cultured into a population of plants prior to,
during, or after the induction step and the population of plants are then contacted with the
10 herbicide.

In yet other embodiments, the provided population of plant cells is a population of
plants.

In some embodiments, the provision of a plant or plant part comprising a
presently disclosed polynucleotide construct comprises introducing the polynucleotide
15 construct into the plant or plant part.

"Introducing" is intended to mean presenting to the organism, such as a plant, or
the cell the polynucleotide or polypeptide in such a manner that the sequence gains access
to the interior of a cell of the organism or to the cell itself. The methods and
compositions do not depend on a particular method for introducing a sequence into an
20 organism or cell, only that the polynucleotide or polypeptide gains access to the interior
of at least one cell of the organism. Methods for introducing polynucleotides or
polypeptides into plants or plant parts are known in the art including, but not limited to,
stable transformation methods, transient transformation methods, and virus-mediated
methods.

25 "Stable transformation" is intended to mean that the nucleotide construct
introduced into a plant integrates into a genome of the plant and is capable of being
inherited by the progeny thereof. "Transient transformation" is intended to mean that a
polynucleotide is introduced into the plant and does not integrate into a genome of the
plant or a polypeptide is introduced into a plant.

Protocols for introducing polypeptides or polynucleotide sequences into plants may vary depending on the type of plant or plant cell, *i.e.*, monocot or dicot, targeted for transformation. Suitable methods of introducing polypeptides and polynucleotides into plant cells include microinjection (Crossway *et al.* (1986) *Biotechniques* 4:320-334),
 5 electroporation (Riggs *et al.* (1986) *Proc. Natl. Acad. Sci. USA* 83:5602-5606, *Agrobacterium*-mediated transformation (U.S. Patent No. 5,563,055 and U.S. Patent No. 5,981,840), direct gene transfer (Paszowski *et al.* (1984) *EMBO J.* 3:2717-2722), and ballistic particle acceleration (see, for example, U.S. Patent Nos. 4,945,050; U.S. Patent No. 5,879,918; U.S. Patent No. 5,886,244; and, 5,932,782; Tomes *et al.* (1995) in *Plant*
 10 *Cell, Tissue, and Organ Culture: Fundamental Methods*, ed. Gamborg and Phillips (Springer-Verlag, Berlin); McCabe *et al.* (1988) *Biotechnology* 6:923-926); and Lec1 transformation (WO 00/28058). Also see Weissinger *et al.* (1988) *Ann. Rev. Genet.* 22:421-477; Sanford *et al.* (1987) *Particulate Science and Technology* 5:27-37 (onion); Christou *et al.* (1988) *Plant Physiol.* 87:671-674 (soybean); McCabe *et al.* (1988)
 15 *Bio/Technology* 6:923-926 (soybean); Finer and McMullen (1991) *In Vitro Cell Dev. Biol.* 27P:175-182 (soybean); Singh *et al.* (1998) *Theor. Appl. Genet.* 96:319-324 (soybean); Datta *et al.* (1990) *Biotechnology* 8:736-740 (rice); Klein *et al.* (1988) *Proc. Natl. Acad. Sci. USA* 85:4305-4309 (maize); Klein *et al.* (1988) *Biotechnology* 6:559-563 (maize); U.S. Patent Nos. 5,240,855; 5,322,783; and, 5,324,646; Klein *et al.* (1988) *Plant*
 20 *Physiol.* 91:440-444 (maize); Fromm *et al.* (1990) *Biotechnology* 8:833-839 (maize); Hooykaas-Van Slogteren *et al.* (1984) *Nature* 311:763-764; U.S. Patent No. 5,736,369 (cereals); Bytebier *et al.* (1987) *Proc. Natl. Acad. Sci. USA* 84:5345-5349 (Liliaceae); De Wet *et al.* (1985) in *The Experimental Manipulation of Ovule Tissues*, ed. Chapman *et al.* (Longman, New York), pp. 197-209 (pollen); Kaeppler *et al.* (1990) *Plant Cell Rep*
 25 9:415-418 and Kaeppler *et al.* (1992) *Theor. Appl. Genet.* 84:560-566 (whisker-mediated transformation); D'Halluin *et al.* (1992) *Plant Cell* 4:1495-1505 (electroporation); Li *et al.* (1993) *Plant Cell Rep* 12:250-255 and Christou and Ford (1995) *Annals of Botany* 75:407-413 (rice); Osjoda *et al.* (1996) *Nat Biotechnol* 14:745-750 (maize via *Agrobacterium tumefaciens*); all of which are herein incorporated by reference.

In specific embodiments, the polynucleotide constructs can be provided to a plant using a variety of transient transformation methods. Such transient transformation methods include, but are not limited to, the introduction of the polynucleotide construct directly into the plant. Such methods include, for example, microinjection or particle bombardment. See, for example, Crossway *et al.* (1986) *Mol Gen. Genet.* 202:179-185; Nomura *et al.* (1986) *Plant Sci.* 44:53-58; Hepler *et al.* (1994) *Proc. Natl. Acad. Sci.* 91:2176-2180 and Hush *et al.* (1994) *J Cell Sci* 107:775-784, all of which are herein incorporated by reference. Alternatively, the polynucleotide construct can be transiently transformed into the plant using techniques known in the art. Such techniques include viral vector system and the precipitation of the polynucleotide in a manner that precludes subsequent release of the DNA. Thus, the transcription from the particle-bound DNA can occur, but the frequency with which it is released to become integrated into the genome is greatly reduced. Such methods include the use of particles coated with polyethylimine (PEI; Sigma #P3143).

In other embodiments, the polynucleotide construct may be introduced into plants or plant parts by contacting plants or plant parts with a virus or viral nucleic acids. Generally, such methods involve incorporating a nucleotide construct within a viral DNA or RNA molecule. It is recognized that the proteins encoded by the various coding polynucleotides of the polynucleotide construct may be initially synthesized as part of a viral polyprotein, which later may be processed by proteolysis *in vivo* or *in vitro* to produce the desired recombinant protein. Further, it is recognized that promoters also encompass promoters utilized for transcription by viral RNA polymerases. Methods for introducing polynucleotides into plants and expressing a protein encoded therein, involving viral DNA or RNA molecules, are known in the art. See, for example, U.S. Patent Nos. 5,889,191, 5,889,190, 5,866,785, 5,589,367, 5,316,931, and Porta *et al.* (1996) *Molecular Biotechnology* 5:209-221; herein incorporated by reference.

Other methods of introducing polynucleotides into a plant or plant part can be used, including plastid transformation methods, and the methods for introducing polynucleotides into tissues from seedlings or mature seeds.

Methods are known in the art for the targeted insertion of a polynucleotide at a specific location in the plant genome. In one embodiment, the insertion of the polynucleotide at a desired genomic location is achieved using a site-specific recombination system. See, for example, WO99/25821, WO99/25854, WO99/25840, 5 WO99/25855, and WO99/25853, all of which are herein incorporated by reference. Briefly, the polynucleotide can be contained in a transfer cassette flanked by two non-recombinogenic recombination sites. The transfer cassette is introduced into a plant or plant part having stably incorporated into its genome a target site which is flanked by two non-recombinogenic recombination sites that correspond to the sites of the transfer 10 cassette. An appropriate recombinase is provided and the transfer cassette is integrated at the target site. The polynucleotide construct is thereby integrated at a specific chromosomal position in the plant genome.

The cells that have been transformed may be grown into plants in accordance with conventional ways. See, for example, McCormick *et al.* (1986) *Plant Cell Rep* 5:81-84. 15 These plants may then be grown, and either pollinated with the same transformed strain or different strains, and the resulting hybrid having constitutive expression of the desired phenotypic characteristic identified. Two or more generations may be grown to ensure that expression of the desired phenotypic characteristic is stably maintained and inherited and then seeds harvested to ensure expression of the desired phenotypic characteristic has 20 been achieved. In this manner, transformed seed (also referred to as “transgenic seed”) having a nucleotide construct, for example, an expression cassette, stably incorporated into their genome is provided. Thus, compositions of the invention include plant cells, plant tissues, plant parts, and plants comprising the presently disclosed polynucleotide constructs. Likewise, the methods of the invention can be performed in plant cells, plant 25 tissues, plant parts, and plants.

In certain embodiments the presently disclosed polynucleotide constructs can be stacked with any combination of polynucleotide sequences of interest in order to create plants with a desired trait. A trait, as used herein, refers to the phenotype derived from a particular sequence or groups of sequences. Plants that have various stacked 30 combinations of traits can be created by any method including, but not limited to, cross-

breeding plants by any conventional or TopCross methodology, or genetic transformation. If the sequences are stacked by genetically transforming the plants, the polynucleotide sequences of interest can be combined at any time and in any order. For example, a transgenic plant comprising one or more desired traits can be used as the target to
5 introduce further traits by subsequent transformation. The traits can be introduced simultaneously in a co-transformation protocol with the polynucleotides of interest provided by any combination of transformation cassettes. For example, if two sequences will be introduced, the two sequences can be contained in separate transformation cassettes (trans) or contained on the same transformation cassette (cis). Expression of the
10 sequences can be driven by the same promoter or by different promoters. In certain cases, it may be desirable to introduce a transformation cassette that will suppress the expression of a polynucleotide of interest. This may be combined with any combination of other suppression cassettes or overexpression cassettes to generate the desired combination of traits in the plant. It is further recognized that polynucleotide sequences
15 can be stacked at a desired genomic location using a site-specific recombination system. See, for example, WO99/25821, WO99/25854, WO99/25840, WO99/25855, and WO99/25853, all of which are herein incorporated by reference.

Any plant species can be transformed, including, but not limited to, monocots and dicots. Examples of plant species of interest include, but are not limited to, corn (*Zea mays*), *Brassica* sp. (e.g., *B. napus*, *B. rapa*, *B. juncea*), particularly those *Brassica* species
20 useful as sources of seed oil, alfalfa (*Medicago sativa*), rice (*Oryza sativa*), rye (*Secale cereale*), sorghum (*Sorghum bicolor*, *Sorghum vulgare*), millet (e.g., pearl millet (*Pennisetum glaucum*), proso millet (*Panicum miliaceum*), foxtail millet (*Setaria italica*), finger millet (*Eleusine coracana*)), sunflower (*Helianthus annuus*), safflower (*Carthamus tinctorius*), wheat (*Triticum* spp.), soybean (*Glycine max*), tobacco (*Nicotiana tabacum*),
25 potato (*Solanum tuberosum*), peanuts (*Arachis hypogaea*), cotton (*Gossypium barbadense*, *Gossypium hirsutum*), sweet potato (*Ipomoea batatas*), cassava (*Manihot esculenta*), coffee (*Coffea* spp.), coconut (*Cocos nucifera*), pineapple (*Ananas comosus*), citrus trees (*Citrus* spp.), cocoa (*Theobroma cacao*), tea (*Camellia sinensis*), banana (*Musa* spp.), avocado
30 (*Persea americana*), fig (*Ficus casica*), guava (*Psidium guajava*), mango (*Mangifera*

indica), olive (*Olea europaea*), papaya (*Carica papaya*), cashew (*Anacardium occidentale*), macadamia (*Macadamia integrifolia*), almond (*Prunus amygdalus*), sugar beets (*Beta vulgaris*), sugarcane (*Saccharum* spp.), oats (*Avena*), barley (*Hordeum*), *Arabidopsis*, switchgrass, vegetables, ornamentals, grasses, and conifers.

5 Vegetables include tomatoes (*Lycopersicon esculentum*), lettuce (e.g., *Lactuca sativa*), green beans (*Phaseolus vulgaris*), lima beans (*Phaseolus limensis*), peas (*Lathyrus* spp.), and members of the genus *Cucumis* such as cucumber (*C. sativus*), cantaloupe (*C. cantalupensis*), and musk melon (*C. melo*). Ornamentals include azalea (*Rhododendron* spp.), hydrangea (*Macrophylla hydrangea*), hibiscus (*Hibiscus rosasanensis*), roses (*Rosa* spp.), tulips (*Tulipa* spp.), daffodils (*Narcissus* spp.), petunias (*Petunia hybrida*), carnation
10 (*Dianthus caryophyllus*), poinsettia (*Euphorbia pulcherrima*), and chrysanthemum.

 Conifers that may be employed in practicing the present invention include, for example, pines such as loblolly pine (*Pinus taeda*), slash pine (*Pinus elliotii*), ponderosa pine (*Pinus ponderosa*), lodgepole pine (*Pinus contorta*), and Monterey pine (*Pinus*
15 *radiata*); Douglas-fir (*Pseudotsuga menziesii*); Western hemlock (*Tsuga canadensis*); Sitka spruce (*Picea glauca*); redwood (*Sequoia sempervirens*); true firs such as silver fir (*Abies amabilis*) and balsam fir (*Abies balsamea*); and cedars such as Western red cedar (*Thuja plicata*) and Alaska yellow-cedar (*Chamaecyparis nootkatensis*). In specific embodiments, plants of the present invention are crop plants (for example, corn, alfalfa, sunflower,
20 *Brassica*, soybean, cotton, safflower, peanut, sorghum, wheat, millet, tobacco, etc.). sugarcane (*Saccharum* spp.). In other embodiments, the plants are maize, rice, sorghum, barley, wheat, millet, oats, sugarcane, turfgrass, or switch grass. In specific embodiments, the plant is sugarcane.

 Other plants of interest include grain plants that provide seeds of interest, oil-seed
25 plants, and leguminous plants. Seeds of interest include grain seeds, such as corn, wheat, barley, rice, sorghum, rye, etc. Oil-seed plants include cotton, soybean, safflower, sunflower, *Brassica*, maize, alfalfa, palm, coconut, etc. Leguminous plants include beans and peas. Beans include guar, locust bean, fenugreek, soybean, garden beans, cowpea, mungbean, lima bean, fava bean, lentils, chickpea, etc.

In certain embodiments, the plant or plant part is a winter wheat plant or plant part. As used herein, “winter wheat” refers to wheat plants or plant parts that require an extended period of low temperatures to be able to flower. Non-limiting examples of winter wheat include *Triticum aestivum* and *Triticum monococcum*.

5 As used herein, the term “plant part” refers to plant cells, plant protoplasts, plant cell tissue cultures from which plants can be regenerated, plant calli, plant clumps, and plant cells that are intact in plants or parts of plants such as embryos, pollen, ovules, seeds, leaves, flowers, branches, fruit, kernels, ears, cobs, husks, stalks, roots, root tips, anthers, and the like, as well as the parts themselves. Grain is intended to mean the
10 mature seed produced by commercial growers for purposes other than growing or reproducing the species. Progeny, variants, and mutants of the regenerated plants are also included within the scope of the invention, provided that these parts comprise the introduced polynucleotides.

 Methods are also provided for increasing transformation frequency, wherein a
15 host cell is provided that comprises a presently disclosed polynucleotide construct comprising an excision cassette separating a polynucleotide encoding a herbicide tolerance polypeptide from its promoter, wherein the excision cassette comprises a polynucleotide encoding a site-specific recombinase that when expressed can excise the excision cassette. The population of plant cells comprising the polynucleotide construct
20 is cultured in the absence of a herbicide to which the herbicide tolerance polypeptide confers herbicide resistance for a period of time sufficient for the population of plant cells to proliferate, followed by the induction of the expression of the site-specific recombinase, thereby excising the excision cassette and allowing for the operable linkage of the herbicide tolerance polynucleotide and its promoter and the expression of the
25 herbicide tolerance polynucleotide allowing for the direct herbicide selection, thereby the transformation frequency is increased compared to a comparable plant cell not comprising the excision cassette and selected directly by herbicide selection. In some embodiments, the herbicide is glyphosate. In some embodiments, the induction comprises desiccating the population of plant cells. In some embodiments the induction
30 comprises cold treatment.

By “period of time sufficient for the population cells to proliferate” is intended to mean that the population of cells has proliferated to a size and quality to produce transgenic events at an optimal level. The time period sufficient for the cells to proliferate may vary depending on the plant species, cultivar, explant and proliferation medium. In some embodiments, the population of plant cells is cultured in the absence of the herbicide to which the herbicide tolerance polypeptide confers herbicide resistance for about 1 hour to about 12 weeks, about 1 day to about 12 weeks, about 1 week to about 12 weeks, or about 1 week to 6 weeks, including but not limited to about 1 hour, 2, hours, 3 hours, 4 hours, 5 hours, 6 hours, 7 hours, 8 hours, 9 hours, 10 hours, 11 hours, 12 hours, 13 hours, 14 hours, 15 hours, 16 hours, 17 hours, 18 hours, 19 hours, 20 hours, 21 hours, 22 hours, 23 hours, 1 day, 2 days, 3 days, 4 days, 5 days, 6 days, 1 week, 2 weeks, 3 weeks, 4 weeks, 5 weeks, 6 weeks, 7 weeks, 8 weeks, 9 weeks, 10 weeks, 11 weeks, and 12 weeks. In other embodiments, the population of plant cells is cultured in the absence of the herbicide to which the herbicide tolerance polypeptide confers herbicide resistance for about 1 day to about 6 weeks, about 1 day to about 2 weeks, about 1 day to about 4 weeks, about 2 days to about 6 weeks, about 4 days to about 6 weeks, about 1 week to about 6 weeks, about 2 weeks to about 6 weeks, about 2 weeks to about 4 weeks, or about 2 weeks to about 3 weeks prior to excision.

“Transformation frequency” refers to the percentage of plant cells that are successfully transformed with a heterologous nucleic acid after performance of a transformation protocol on the cells to introduce the nucleic acid. In some embodiments, transformation further includes a selection protocol to select for those cells that are expressing one or more proteins encoded by a heterologous nucleic acid of interest. In some embodiments, transformation makes use of a “vector,” which is a nucleic acid molecule designed for transformation into a host cell.

An increased “transformation efficiency,” as used herein, refers to any improvement, such as an increase in transformation frequency, increased quality events frequency, labor saving, and/or decrease in ergonomic impact that impact overall efficiency of the transformation process by reducing the amount of resources required.

In general, upon use of the methods taught herein, transformation frequency is increased by at least about 3%, 5%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or 100% or greater, or even 1-, 2-, 3-, 4-, 5-, 6-, 7-, 8-, 9-, 10-fold or more, than the transformation frequency relative to a control. The “control” provides a reference point for measuring changes in phenotype of the subject plant or plant cell, e.g., transformation frequency/efficiency, callus quality or transformation process time. The control may include, for example, plant cells transformed with a corresponding nucleic acid without the excision cassette.

In certain embodiments, the plant or plant part useful in the presently disclosed methods and compositions is recalcitrant. As used herein, a “recalcitrant plant” or “recalcitrant plant part” is a plant or plant part in which the average transformation frequency using typical transformation methods is relatively low, and typically less than about 1%, 2%, 3%, 4%, 5%, 6%, 7%, 8%, 9%, 10%, 11%, 12%, 13%, 14%, 15%, 16%, 17%, 18%, 19%, 20%, 25%, or 30%. The transformation of species, varieties or cultivars recalcitrant to transformation is time consuming, laborious, and inefficient compared to the transformation of non-recalcitrant varieties, with respect to one or more methods of transformation (e.g., *Agrobacterium*-mediated transformation). Non-limiting examples of species recalcitrant to *Agrobacterium*-mediated transformation include, but are not limited to, species of *Lolium* (rye grass), elite varieties of maize, cultivars of sugarcane, species of rice (especially Indica), and various turf grass species. In some embodiments, the recalcitrant plant or plant part is unable to be transformed in the absence of a cell proliferation factor. In certain embodiments, the recalcitrant plant or plant part is an elite maize inbred or a cell or tissue thereof. In other embodiments, the recalcitrant plant or plant part is the sugarcane cultivar CP96-1252, CP01-1372, CPCL97-2730, HoCP85-845, or CP89-2143 or a cell or tissue thereof.

In some embodiments of the present methods the recalcitrant plant part is an explant from a model or recalcitrant inbred or cultivar. In some embodiments of the present methods and compositions, the explant is from a recalcitrant inbred having a type I callus genotype. In some embodiments of the present methods and compositions, the explant is from a recalcitrant maize inbred having a type I callus genotype. Callus in

grasses can be classified as type I or type II, based upon color, texture, regeneration system, and the amount of time required for callus initiation. The morphology of callus has been reported and described in the agronomically important monocot crops such as maize (Armstrong et al. (1985) *Planta* 164:207-214; Assam (2001) *Arab J Biotechnol* 4:247-256; Frame et al. (2000) *In Vitro Cell Dev Biol-Plant* 36:21-29; Lu et al. (1982) *L. Theor Appl Genet* 62:109-112; McCain et al. (1988) *Bot Gazette* 149:16-20; Songstad et al. (1992) *Am J Bot* 79:761-764; Welter et al. (1995) *Plant Cell Rep* 14:725-729; each of which is herein incorporated by reference in its entirety), rice (Chen et al. (1985) *Plant Cell Tissue Organ Cult* 4:51-51; Nakamura et al. (1989) *Japan J Crop Sci* 58:395-403; Rueb et al. (1994) *Plant Cell Tissue Organ Cult* 36:259-264; each of which is herein incorporated by reference in its entirety), sorghum (Jeoung et al. (2002) *Hereditas* 137:20-28; which is herein incorporated by reference in its entirety), sugarcane (Guiderdoni et al. (1988) *Plant Cell Tissue Organ Cult* 14:71-88; which is herein incorporated by reference in its entirety), wheat (Redway et al. (1990) *Theor Appl Genet* 79:609-617; which is herein incorporated by reference in its entirety), and various nonfood grasses. Type I callus is the typical and most prevalent callus formed in monocot species. It is characterized by compact form, slow-growth, white to light yellow in color, and highly organized. This callus is composed almost entirely of cytoplasmic meristematic cells that lack large vacuoles. In maize, type I callus can only be maintained for a few months and cannot be used in suspension cultures; whereas, type II callus can be maintained in culture for extended periods of time and is able to form cell suspensions. Type II callus derived from maize has been described as soft, friable, rapidly growing and exceedingly regenerative but is typically formed at lower frequencies than type I callus. Embryogenic suspension cells can be initiated from type II callus, which few maize lines can form. Although the ability to form type II callus can be backcrossed into agronomically important maize lines, in practice this is time consuming and difficult. Moreover, even for those lines that can form type II callus, the method requires a great deal of time and labor and is, therefore, impractical. Normally, recalcitrant inbred or cultivar genotypes that produce type I callus have low transformation frequencies. Typically with maize type I inbreds large numbers of embryos or other explants must be

screened to identify sufficient quantities of events, which is expensive and labor intensive.

It is to be noted that the term “a” or “an” entity refers to one or more of that entity; for example, “a polynucleotide” is understood to represent one or more polynucleotides.

5 As such, the terms “a” (or “an”), “one or more,” and “at least one” can be used interchangeably herein.

Throughout this specification and the claims, the words “comprise,” “comprises,” and “comprising” are used in a non-exclusive sense, except where the context requires otherwise.

10 As used herein, the term “about,” when referring to a value is meant to encompass variations of, in some embodiments $\pm 50\%$, in some embodiments $\pm 20\%$, in some embodiments $\pm 10\%$, in some embodiments $\pm 5\%$, in some embodiments $\pm 1\%$, in some embodiments $\pm 0.5\%$, and in some embodiments $\pm 0.1\%$ from the specified amount, as such variations are appropriate to perform the disclosed methods or employ the disclosed
15 compositions.

Further, when an amount, concentration, or other value or parameter is given as either a range, preferred range, or a list of upper preferable values and lower preferable values, this is to be understood as specifically disclosing all ranges formed from any pair of any upper range limit or preferred value and any lower range limit or preferred value,
20 regardless of whether ranges are separately disclosed. Where a range of numerical values is recited herein, unless otherwise stated, the range is intended to include the endpoints thereof, and all integers and fractions within the range. It is not intended that the scope of the presently disclosed subject matter be limited to the specific values recited when defining a range.

25 The following examples are offered by way of illustration and not by way of limitation.

30

EXPERIMENTAL

Example 1. Glyphosate Selection of Transformed Maize Inbred PHR03

Immature embryos from maize inbred PHR03 were harvested 9-13 days post-pollination with embryo sizes ranging from 0.8 – 2.5 mm length and were co-cultivated with *Agrobacterium* strain LBA4404 containing the vector PHP29204 or *Agrobacterium* strain LBA4404 containing the vector PHP32269 on PHI-T medium for 2-4 days in dark conditions. PHP29204: Ubi:DsRed + Ubi:GAT4602. PHP32269: Ubi:PMI + Ubi:MOPAT::YFP. Ubi refers to the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBI1ZM INTRON1; SEQ ID NO: 113). The tissues were then transferred to DBC3 medium without selection for one week, and then to DBC3 medium with 0.25 mM or 0.5 mM glyphosate for 3 weeks, and then DBC3 medium with 0.5 mM glyphosate for another 3-4 weeks. The embryos were then transferred to PHI-RF maturation medium with 0.1 mM glyphosate for 2-3 weeks until shoots formed, at which point, the shoots were transferred to MSB medium in Phytatrays containing 100 mg/L cefotaxime for rooting. Plants with good roots were transferred to soil for further growth and a glyphosate spray test. For PMI selection using PHP32269, DBC3 medium containing 12.5 g/L mannose and 5g/L maltose was used for selection. PHI-RF maturation medium without any selective agent or sugar modifications was used for regeneration.

PHI-T medium contains 0.1 μ M copper in MS salts 4.3 mg/L, Nicotinic acid 0.5 mg/L, Pyridoxine HCl 0.5 mg/L, Thiamine HCl 1 mg/L, Myo-inositol 100 mg/L, 2,4-D 2 mg/L, Sucrose 20 g/L, Glucose 10 g/L, L-proline 700 mg/L, MES 0.5 g/L, Acetosyringone 100 μ M, Ascorbic acid 10 mg/L and Agar 8.0 g/L.

PHI-RF is 4.3 g/L MS salts (GIBCO BRL 11117-074), 0.5 mg/L nicotinic acid, 0.1 mg/L thiamine HCl, 0.5 mg/L pyridoxine HCl, 2.0 mg/L glycine, 0.1 g/L myo-inositol, 0.49 μ M cupric sulfate, 0.5 mg/L zeatin (Sigma Z-0164), 1 mg/L IAA, 26.4 μ g/L ABA, thidiazuron 0.1 mg/L, 60 g/L sucrose, 100 mg/L cefotaxime, 8 g/L agar, pH 5.6.

Table 4. Transformation frequency of maize inbred PHR03 with PHP29204 or PHP32269.

Vector	No. of embryos	No. of T ₀ events	% Transformation	No. single copy events	% Single Copy Events
PHP29204	300	21	7	13	61.9
PHP32269	90	36	40	16	44.4

The transformation frequency with PHP29204 with glyphosate selection was only 7% in the maize inbred PHR03. Overall, glyphosate selection did not provide for a clean selection, a lot of non-transformed tissues were growing, and the morphology of both transformed and non-transformed tissues was irregular.

Example 2. *Agrobacterium*-mediated Sugarcane Transformation Using a Standard Test

Vector Without Developmental Genes

Media for Plant Transformation:

Liquid DBC3(M5G) contains MS salts (4.3 g/L) plus maltose (30 g/L); glucose (5 g/L); thiamine-HCl (1 mg/mL); myo-inositol(0.25 g/L); N-Z-amine-A (casein hydrolysate) (1 g/L); proline (0.69 g/L); CuSO₄ (4.9 μM); 2,4-D (1.0 mg/L); BAP (0.5 mg/L); Adjust volume to 1 L with ddH₂O; pH 5.8 — Adjust pH with 1 M KOH; autoclave.

DBC3 contains MS salts (4.3 g/L) plus maltose (30 g/L); thiamine-HCl (1 mg/mL); myo-inositol (0.25 g/L); N-Z-amine-A (casein hydrolysate) (1 g/L); proline (0.69 g/L); CuSO₄ (4.9 μM); 2,4-D (1.0 mg/L); BAP (0.5 mg/L); Adjust volume to 1 L with ddH₂O; pH 5.8 — Adjust pH with 1 M KOH; Phytigel (3.5 g/L); autoclave.

DBC6 contains MS salts (4.3 g/L) plus maltose (30 g/L); thiamine-HCl (1 mg/mL); myo-inositol(0.25 g/L); N-Z-amine-A (casein hydrolysate) (1 g/L); proline (0.69 g/L); CuSO₄ (4.9 μM); 2,4-D (0.5 mg/L); BAP (2.0 mg/L); Adjust volume to 1 L with ddH₂O; pH 5.8 — Adjust pH with 1 M KOH; Phytigel (3.5 g/L); autoclave.

MSB contains MS salts and vitamins (4.43 g/L) plus sucrose (20 g/L); myo-inositol (1.0 g/L); indole-3-butyric acid (IBA, 0.5 mg/L); Adjust volume to 1 L with ddH₂O; pH 5.8 — Adjust pH with 1 M KOH; Phytigel (3.5 g/L); autoclave.

Preparation of Agrobacterium Suspension:

Agrobacterium tumefaciens harboring a binary vector from a -80° frozen aliquot was streaked out onto solid PHI-L or LB medium containing an appropriate antibiotic and
5 cultured at 28°C in the dark for 2-3 days. A single colony or multiple colonies were picked from the master plate and streaked onto a plate containing PHI-M medium and incubated at 28°C in the dark for 1-2 days. *Agrobacterium* cells were collected from the solid medium using 5 mL 10 mM MgSO₄ medium (*Agrobacterium* infection medium) plus 100 µM acetosyringone. One mL of the suspension was transferred to a
10 spectrophotometer tube and the OD_{500nm} of the suspension was adjusted to 0.35-0.40 at 550 nm using the same medium.

Agrobacterium Infection and Co-cultivation:

Good quality callus tissues induced from *in vitro*-cultured plantlets were collected
15 in an empty Petri dish and exposed to air in the hood for about 30 minutes. Tissue that is younger than 2 months old is considered ideal for transformation. One mL *Agrobacterium* suspension was added to the Petri dish, the tissues were broken or chopped into small pieces, and an additional 1-3 mL *Agrobacterium* (AGL1) suspension was then added to cover all the tissues. The Petri dish was placed into a transparent
20 polycarbonate desiccator container, and the container was covered and connected to an in-house vacuum system for 20 minutes. After infection, the *Agrobacterium* suspension was drawn off from the Petri dish and the tissues were transferred onto 2 layers of VWR 415 filter paper (7.5 cm diameter) of a new Petri dish and 0.7-2.0 mL liquid DBC3 (M5G) medium plus 100 µM acetosyringone was added for cocultivation depending on the
25 amount of tissue collected. The top layer of filter paper containing the infected tissues was transferred to a fresh layer of filter paper of another new Petri dish. The infected tissues were incubated at 21°C in the dark for 3 days.

Selection and Plant Regeneration:

30 Callus tissues were transferred to first round selection DBC3 containing antibiotics (timentin and cefotaxime) and 3 mg/L bialaphos (Meiji Seika, Tokyo, Japan).

Tissues were transferred to 2nd round selection DBC6 containing antibiotics and 3-5 mg/L bialaphos and subcultured for 3 weeks at 26-28°C in dark or dim light conditions. At the 3rd round selection on DBC6 medium containing antibiotics and bialaphos, tissues were broken into smaller pieces and exposed to bright light conditions ($30-150 \mu\text{mol m}^{-2} \text{sec}^{-1}$) for 2-3 weeks. Shoot-elongated tissues were broken into small pieces and transferred to MSB regeneration/rooting medium containing antibiotics and 3 mg/L bialaphos. Single plantlets were separated and transferred to soil.

Table 5 shows the results of transformation experiments using 7 U.S. sugarcane cultivars. CP89-2376 and CP88-1762 had >100% transformation frequency at the T₀ plant level using a standard vector containing DsRED and PAT (or moPAT) while the remaining 5 cultivars, CP96-1252, CP01-1372, CPCL97-2730, HoCP85-845 and CP89-2143, were recalcitrant in transformation.

Table 5. Transformation Frequencies at T₀ Plant Level in 7 U.S. Sugarcane Cultivars Using a Standard Test Vector.

CP96-1252	CP01-1372	CP89-2376	CPCL97-2730	HoCP85-845	CP89-2143	CP88-1762
n.t.*	n.t.	75.0% (6/8)	n.t.	n.t.	n.t.	n.t.
0% (0/8)	0% (0/8)	100.0% (8/8)	0% (0/8)	n.t.	n.t.	n.t.
n.t.	n.t.	87.5% (7/8)	n.t.	n.t.	n.t.	n.t.
n.t.	n.t.	150.0% (12/8)	n.t.	0% (0/8)	n.t.	n.t.
n.t.	n.t.	n.t.	n.t.	n.t.	0% (0/8)	62.5% (5/8)
n.t.	n.t.	100.0% (8/8)	n.t.	n.t.	0% (0/8)	137.5% (11/8)
n.t.	n.t.	187.5% (15/8)	n.t.	n.t.	n.t.	137.5% (11/8)

Transformation Frequency = (# transgenic events / # explants infected with *Agrobacterium*) x 100%

*n.t.: not tested

5 *Confirmation of Transgenic Events:*

The putative stable callus/green tissues/regenerating plants were identified based on the visible RFP marker gene expression. All of these putative transgenic callus tissues were transferred to medium for plant regeneration under standard regeneration conditions. The final confirmation of stable transformation frequency was determined based on
10 molecular analysis such as PCR and Southern blot hybridization.

Example 3. Sugarcane Transformation Using a Developmental Gene (DevGene) Vector PHP35648 and Excision Test

A DevGene binary vector (PHP35648, Figure 1) with the BBM/WUS gene
15 cassette was initially compared with a standard vector containing PAT or moPAT plus DsRED without the BBM/WUS gene cassette for transformation frequency using two *Agrobacterium* strains, AGL1 and LBA4404, in cultivar CP89-2376 and the recalcitrant cultivar CP01-1372 (Table 6). The DevGene binary vector contains
Ubi::LoxP::CFP+Rab17Pro-attb1::Cre+Nos::ZmWUS2+Ubi::ZmBBM-
20 LoxP::YFP+Ubi::MOPAT (Figure 1); each gene cassette has a 3' terminator. The Lox cassette containing CFP::Cre::WUS::BBM can be excised by Cre recombinase controlled by the Rab17 promoter. The PHP35648 vector was designed to demonstrate the excision efficiency of the excision cassette using visual markers. The PHP35648 excision cassette comprises the cyan fluorescent protein (CFP) controlled by the ubiquitin promoter
25 (comprising the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBIZM INTRON1; SEQ ID NO: 113)), which is located outside of the loxP site flanking the excision cassette (see Figure 1). Transformants comprising the excision cassette can be visually identified by the presence of the cyan fluorescent protein (CFP). When the
30 excision cassette is excised, the yellow fluorescent protein (YFP) is expressed under the regulation of the ubiquitin promoter. Transformants lacking the excision cassette can be visually identified by the presence of the yellow fluorescent protein (YFP). The ratio of

cyan fluorescent protein (CFP) to yellow fluorescent protein (YFP) can be used to demonstrate the excision efficiency. In PHP35648, the ubiquitin promoter controlling the expression of the moPAT gene product was included outside of the excision cassette as a positive selection to reduce the number of escapes.

5 Callus tissues of all 5 sugarcane cultivars were induced and maintained on DBC3 medium. Tissues were infected with *Agrobacterium* containing the DevGene binary vector PHP35648 in liquid 10 mM MgSO₄ plus 100 µM acetosyringone and then co-cultivated with liquid DBC3 (M5G) medium plus 100 µM acetosyringone on filter paper in Petri dishes at 21°C in the dark. Three days after co-cultivation, the tissues were
10 transferred to DBC3 containing 100 mg/L cefotaxime and 150 mg/L timentin for AGL1 and DBC3 containing 100 mg/L carbenicillin for LBA4404, and incubated at 26°C ($\pm 1^\circ\text{C}$) in the dark or dim light for 3-7 days. Afterwards, the tissues were transferred to the same media as the previous step plus 3 or 5 mg/L bialaphos. After 2 to 3 weeks, the tissues were transferred to 2nd round selection DBC6 containing antibiotics and 3-5 mg/L
15 bialaphos. After two months from the initiation of the experiment, transformation frequency was calculated by the number of tissues showing CFP-expressing sectors divided by the number of explants infected by *Agrobacterium*. AGL1 was more efficient in transformation than LBA4404 in both CP89-2376 and CP01-1372 (Table 6, rows 1 and 2). There was also a genotype difference in transformation frequency; the CP89-2376
20 cultivar had a much higher transformation frequency than the recalcitrant cultivar CP01-1372 using either of the *Agrobacterium* strains.

AGL1 containing the DevGene binary vector PHP35648 was also used to test sugarcane germplasm screening in another set of four experiments (Table 6, rows 3-6) using 5 different cultivars (CP96-1252, CP01-1372, CP89-2376, CPCL97-2730 and
25 HoCP85-845). Callus tissues of all 5 cultivars tested were induced and maintained on DBC3 medium and tissues were infected with AGL1 containing the developmental gene binary vector PHP35648. The use of developmental genes dramatically increased transformation frequency in all 5 cultivars tested. Transformation frequencies in the most amenable cultivar, CP89-2376, using a standard binary vector averaged 116.7% (56/48)
30 (Table 6). In contrast, an average transformation frequency in CP89-2376 from the 5

experiments using the DevGene binary vector PHP35648 was >2,512.5% (>1,005 events/40 tissues infected) (see Table 6, rows 2-6). An increase in transformation frequency was also observed in the recalcitrant cultivars CP96-1252, CP01-1372, CPCL97-2730 and HoCP85-845; with transformation frequencies ranging from 62.5% to 1250.0% using AGL1 while no transgenic events were obtained using the standard vector without the BBM/WUS gene cassette from these cultivars (Table 6, row 7).

Table 6. Transformation Frequency in Sugarcane Using a BBM/WUS Developmental Gene Vector PHP35648.

<i>Agrobacterium</i>		Sugarcane Cultivar				
Strain	Binary Vector	CP96-1252	CP01-1372	CP89-2376	CPCL97-2730	HoCP85-845
AGL1	DG ^a	n.t. ^c	37.5% (3/8)	n.t.	n.t.	n.t.
LBA4404	DG	n.t.	0% (0/8)	n.t.	n.t.	n.t.
AGL1	DG	n.t.	>1,250.0% (>100/8)	>6,250.0% (>500/8)	n.t.	n.t.
LBA4404	DG	n.t.	12.5% (1/8)	>1,500% (>120/8)	n.t.	n.t.
AGL1	DG	n.t.	n.t.	687.5% (>55/8)	n.t.	n.t.
AGL1	DG	n.t.	n.t.	>2,500% (>200/8)	175.0% (14/8)	n.t.
AGL1	DG	150.0% (12/8)	62.5% (5/8)	>625.0% (>50/8)	62.5% (6/8)	n.t.
AGL1	DG	n.t.	n.t.	>2,500% (>200/8)	n.t.	187.5% (15/8)
AGL1	Std^b	0% (0/8)	0% (0/8)	116.7% (56/48)	0% (0/8)	0% (0/8)

Each transformation treatment had 8 pieces of callus tissues 0.4-0.5 cm in size.

5 DG^a: developmental gene vector with BBM/WUS gene cassette

Std^b: standard vector without BBM/WUS gene cassette

n.t.^c: not tested

Excision of the LoxP cassette by dessication monitored by visual markers

Transgenic callus tissues were desiccated on dry filter papers for one day to induce excision of the Lox cassette containing CFP::Cre::WUS::BBM by Cre recombinase driven by the Rab17 promoter (Figure 1). Excision was monitored by observing YFP expression on desiccated transgenic callus events by the presence of the UBI:loxP:YFP junction formed as a result of excision (Figure 1). Cre excision occurred on 83 of 87 transgenic events (95.4%) (Table 7). Plants from some transgenic events after excision were regenerated on MSB plus 1-3 mg/L bialaphos and antibiotics.

Table 7. Excision Efficiency of the BBM/WUS Gene Cassette in Transgenic Sugarcane Events by Desiccation.

Sugarcane Cultivar	<i>Agrobacterium</i> Strain	Binary Vector	Excision Efficiency (%)
CP89-2376	AGL1	DG ^a	93% (40/43)
CP89-2376	LBA4404	DG	100% (25/25)
CP01-1372	AGL1	DG	100% (13/13)
CP01-1372	LBA4404	DG	0% (0/1)
CP89-2376	AGL1	DG	100% (5/5)
Average			95.4% (83/87)

DG^a: developmental gene (DevGene) vector PHP35648 with BBM/WUS gene cassette

Example 4. Sugarcane Excision Induction and Plant Regeneration from Transformed Callus/Green Tissue Events Generated Using a Developmental Gene (DevGene) Vector PHP54561

Generation of Transgenic Events:

A new DevGene binary vector PHP54561 with the BBM/WUS gene cassette was designed as described in Figure 2. The DevGene binary vector PHP54561 contains Ubi::LoxP-moPAT+Ubi:YFP+Rab17Pro-attb1:Cre+Nos:ZmWUS2+Ubi:ZmBBM-LoxP::GLYAT (Figure 2); each gene cassette has a 3' terminator. The Lox cassette

containing moPAT+Ubi:YFP+Rab17Pro-attb1:Cre+Nos:ZmWUS2+Ubi:ZmBBM can be excised by Cre recombinase controlled by the Rab17 promoter. The PHP54561 excision cassette was designed to test the excision efficiency directly by glyphosate tolerance (see Figure 2). The yellow florescent protein (YFP) was included in the PHP54561 excision cassette as a visual marker and moPAT as a selection marker prior to excision (see Figure 2). Ubi refers to the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron 1 (UBIZM INTRON1; SEQ ID NO: 113).

Callus tissues of two U.S. sugarcane cultivars, CP88-1762, CP01-1372 and 1 Australian cultivar, KQ228, were induced and maintained on DBC3 or DBC6 medium. Tissues were infected with *Agrobacterium* containing the DevGene binary vector PHP54561 in liquid 10 mM MgSO₄ plus 100 µM acetosyringone and then co-cultivated with liquid DBC3 (M5G) medium plus 100 µM acetosyringone on the filter paper in Petri dishes at 21°C in the dark. Three days after co-cultivation, the tissues of CP88-1762/CP01-1372 and KQ228 were transferred to DBC3 and DBC6 containing 100 mg/L cefotaxime and 150 mg/L timentin, respectively, and incubated at 26°C (±1°C) in the dark or dim light for 3-7 days. Afterwards, the tissues were transferred to the same media as the previous step plus 3 or 5 mg/L bialaphos. After 2 to 3 weeks, the tissues were transferred to 2nd round selection DBC6 containing antibiotics and 3-5 mg/L bialaphos. YFP-expressing sectors were transferred to the same medium for proliferation. After two months from the initiation of the experiment, transformation frequency was calculated by the number of tissues showing YFP-expressing sectors divided by the number of explants infected by *Agrobacterium*. Table 8 demonstrated transformation frequency at the T₀ tissue level in 3 sugarcane cultivars. CP88-1762, an amenable cultivar had 405% transformation. Two recalcitrant cultivars, CP01-1372 and KQ228 also had high transformation frequencies, 885% and 130%, respectively.

Table 8. Transformation Frequencies at the T₀ Tissue Level in Sugarcane with Bialaphos Selection before Excision.

Cultivar	Txn Frequency (%)
CP01-1372*	270% (27/10)
CP01-1372*	1500% (150/10)
Total	885% (177/20)
CP88-1762	400% (40/10)
CP88-1762	410% (41/10)
Total	405% (81/20)
KQ228*	10% (1/10)
KQ228*	250% (25/10)
Total	130% (26/20)

*CP01-1372 and KQ228 are recalcitrant commercial cultivars.

5 *Excision of LoxP Cassette by Desiccation and Plant Regeneration with Glyphosate Selection:*

Transgenic tissues (0.3-0.5 mm in diameter) were transferred to an empty 60 mm x 25 mm Petri dish containing a piece of sterilized glass filter paper (VWR Glass Microfibre filter, 691). The Petri dish was covered with a lid and placed in a container with a tight-seal cover. A Petri dish (or beaker) with ~20 mL of sterilized water with the lid open was placed in the container. The container was kept in a dark culture room for 1-2.5 days at 28°C; the desiccation period was dependent on the degree or size of tissues. After 1-2.5 days of desiccation treatment, the desiccated tissues were transferred to DBC6 proliferation medium with antibiotics and 100 µM glyphosate. The plates were kept in

dim ($10\text{-}50\ \mu\text{mol m}^{-2}\text{ sec}^{-1}$) to moderately bright light at $26\text{-}28^{\circ}\text{C}$ for 2-3 weeks (Figure 3). If necessary, tissues were subcultured for another round on the same medium for another 2-3 weeks to get small green shoots; the plates was kept in a higher intensity of light at $26\text{-}28^{\circ}\text{C}$. Tissues with shoots were picked up and placed onto MSB

5 regeneration/rooting medium containing antibiotics and $20\text{-}30\ \mu\text{M}$ glyphosate in A175 Agar (PhytoTechnology Lab) as a gelling agent. Tissues were cultured under bright light conditions ($50\text{-}200\ \mu\text{mol m}^{-2}\text{ sec}^{-1}$) for 3-4 weeks at $26\text{-}28^{\circ}\text{C}$. When shoots were strong enough, single plantlets were separated and transferred to soil. In general, plants with complete excision exhibited a normal phenotype with greener and faster growth, while
10 plantlets from tissues without excision of the developmental genes or having incomplete excision usually showed a stunted phenotype or bleached shoots, indicating susceptibility to glyphosate (Figures 4 and 5). Plants with a normal phenotype were transferred to soil for further growth, glyphosate spray test and molecular assay.

Table 9 shows LoxP cassette excision efficiency in transgenic events of 3
15 sugarcane cultivars, CP88-1762, CP01-1372 and KQ228, based on glyphosate resistance of the events. Excision efficiencies ranged from 32% to 68% in these 3 cultivars.

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Table 9. Excision Efficiency with Glyphosate Selection of Transgenic Sugarcane Events by Desiccation.

Cultivar	Transformation Frequency*	# of events desiccated	# of events with green elongated shoots on glyphosate	Excision Efficiency (# of events excised/# of events desiccated)
CP01-1372	270% (27/10)	12	8	66.7% (8/12)
CP01-1372	1500% (150/10)	41	28	68.3% (28/41)
Total	885% (177/20)	53	36	67.9% (36/53)
CP88-1762	400% (40/10)	15	6	40.0% (6/15)
CP88-1762	410% (41/10)	38	20	52.6% (20/38)
Total	405% (81/20)	53	26	49.1% (26/53)
KQ228	10% (1/10)	1	0	0% (0/1)
KQ228	250% (25/10)	21	7	33.3% (7/21)
Total	130% (26/20)	22	7	31.8% (7/22)

*bialaphos selection before excision

5 *Glyphosate Resistance Confirmation by Glyphosate Spray Test:*

T₀ plantlets were then moved to soil and spray tested with 4X glyphosate to confirm excision/ glyphosate resistance. All 72 independent T₀ events from 3 sugarcane cultivars (Table 9) showed strong glyphosate resistance while plants of 3 nontransgenic cultivars were completely killed by glyphosate spray. The final confirmation of stable

transformation frequency is determined based on molecular analysis such as PCR and Southern blot hybridization.

Example 5. Corn Excision Induction and Plant Regeneration from Desiccated T₁

5 Immature Embryos

Corn Transformation:

A corn elite inbred, PHR03 was transformed with *Agrobacterium* strain AGL1 containing the excision vector PHP54353. The PHP54353 vector contains Ubi::LoxP-Ds
10 RED+Rab17-attB::CRE-LoxP::GLYAT (Figure 6). The Lox cassette containing Ds RED+Rab17-attB::CRE can be excised by Cre recombinase controlled by the Rab17 promoter. The PHP54353 excision cassette was designed to demonstrate direct glyphosate selection. Ubi refers to the maize ubiquitin promoter (UBI1ZM PRO; SEQ ID NO: 111), the ubiquitin 5' UTR (UBI1ZM 5UTR; SEQ ID NO: 112), and ubiquitin intron
15 1 (UBIZM INTRON1; SEQ ID NO: 113).

Immature embryos from maize inbred PHR03 were harvested 9-13 days post-pollination with embryo sizes ranging from 0.8 – 2.5 mm length and were co-cultivated with *Agrobacterium* strain AGL1 containing the excision vector PHP54353 on PHI-T medium for 3 days in dark conditions. These embryos were then transferred to DBC3
20 medium containing 100 mg/L cefotaxime in dim light conditions. RFP-expressing sectors were picked up and proliferated on the same medium. When the tissue proliferation period for each transgenic event was sufficient, tissues were moved to PHI-RF maturation medium. Regenerating shoots were transferred to MSB medium in Phytatrays containing 100 mg/L cefotaxime for rooting. Plants with good roots were
25 transferred to soil for further growth, glyphosate spray test and molecular assay.

PHI-T medium contains 0.1 µM copper in MS salts 4.3 mg/L, Nicotinic acid 0.5 mg/L, Pyridoxine HCl 0.5 mg/L, Thiamine HCl 1 mg/L, Myo-inositol 100 mg/L, 2,4-D 2 mg/L, Sucrose 20 g/L, Glucose 10 g/L, L-proline 700 mg/L, MES 0.5 g/L, Acetosyringone 100
30 µM, Ascorbic acid 10 mg/L and Agar 8.0 g/L.

PHI-RF is 4.3 g/L MS salts (GIBCO BRL 11117-074), 0.5 mg/L nicotinic acid, 0.1 mg/L thiamine HCl, 0.5 mg/L pyridoxine HCl, 2.0 mg/L glycine, 0.1 g/L myo-inositol, 0.49 μ M cupric sulfate, 0.5 mg/L zeatin (Sigma Z-0164), 1 mg/L IAA, 26.4 μ g/L ABA, thidiazuron 0.1 mg/L, 60 g/L sucrose, 100 mg/L cefotaxime, 8 g/L agar, pH 5.6.

5

Immature Embryo Isolation, Desiccation, Selection and Regeneration:

Sterilized immature embryos with 2.0-3.5 mm were placed scutellum side down on sterile fiber glass filter paper in a Petri dish. 300 μ L of DBC6 liquid medium with 100 mg/L cefotaxime was added to the filter paper to prevent over drying. Plates were wrapped with Parafilm and checked for expression of DsRed before desiccation in order to compare expression after desiccation. Plates were moved into a sterile laminar hood unwrapped and let stand for 2-4 days until the embryos appeared darker and shrunken, and were desiccated. Embryos were then placed scutellum side down onto MSA regeneration medium containing 100 mg/L cefotaxime with 10-50 μ M glyphosate for selection. Five to 10 days later, DsRed expression is checked in the emerging shoots.

15

Example 6. Natural Desiccation and Excision in Transgenic Mature Corn Seed

Immature embryos of maize inbred PHR03 were transformed with the excision vector AGL1/PHP54353, the expression of DsRed was visually confirmed, and T_0 plantlets were regenerated as described in Example 5. Before moving the T_0 plantlets to soil, the expression of DsRed was again visually confirmed.

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Glyphosate resistance confirmation

To confirm that the natural desiccation process that occurs during seed maturation would in fact allow for the excision of DsRed and resistance to glyphosate, seeds collected from T_0 plants crossed with wild-type PHR03 pollen were germinated in soil. By planting seeds straight to soil without any treatments, excision would be a result of natural processes.

25

Three random events were chosen to be tested by this method. Five mature T_1 seeds each from the following events, PHP54353 T_0 event numbers 6, 7, and 10 were placed in small pots with Metro Mix soil (Sun Gro Horticulture, McFarland, CA) with

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fertilizer and placed in the greenhouse. After plants had germinated and grown to about 12-18 cm (10-12 days after planting), the plants were then sprayed with glyphosate + surfactant at 2X or 4X concentration (1X is equivalent to what is used in the field). Before spraying, all pots were evenly spaced and positioned to ensure that they would receive an even distribution of glyphosate. The distance between the sprayer nozzle and the apical meristem of the plants was approximately 18 inches. Within 10-12 days, it was visibly evident which plants were not affected by the herbicide and which plants had been severely damaged.

The results of the spray test are presented in Table 10. From visible spray test results, all wild-type PHR03 plants had been severely damaged, as predicted. It was also clear that 2 out of 4 plants from event number 6 had no signs of damage and continued to grow at a normal rate having not lost any leaf tissue. However, all 5 plants from event number 7 did show damage equivalent to that of the wild-type PHR03 plants, which was not expected. All 4 plants from event number 10 also showed damage equivalent to that of the wild-type PHR03 plants. When the T₀ plants were analyzed for the presence of the DsRED and GLYAT genes, it was discovered that event number 10 did not have the DsRED gene and although the T₀ plant had the GLYAT gene, presumably GLYAT was not expressed because it was not operably linked to a promoter (see Table 10). In event number 13, 3 out of 5 plants showed damage and 2 out of 5 plants were tolerant.

Table 10. Glyphosate Spray Test on Plants Germinated from T₁ Mature Corn Seed

Lab event #	DS-RED2INT QPCR of T ₀	GLYAT QPCR of T ₀	Glyphosate Spray Test
6	+	+	2/4 plants damaged; 2/4 plants tolerant
7	+	+	5/5 plants damaged
10	-	+	4/4 plants damaged
13	+	+	3/5 plants damaged; 2/5 plants tolerant
Wild-type	-	-	4/4 plants damaged

Example 7. Tobacco Excision Induction and Plant Regeneration from Transformed Tissue Events

Tobacco Transformation

5 Young leaves are harvested from *in vitro*-cultured tobacco plants and cut into 0.5-1 cm size as an *Agrobacterium* infection target. AGL1/PHP55062 (a standard excision vector, Figure 8) is used for transformation. Transgenic tobacco (cv. Petite havana) plants are generated following the leaf disc method described by Horsch et al. (1985) *Science* 227:1229-1231, which is herein incorporated by reference in its entirety, and
10 50 mg/L hygromycin B was used for selection.

Excision of LoxP Cassette by Desiccation and Plant Regeneration with Glyphosate Selection

 Tobacco desiccation experiments are conducted to induce excision from
15 transformed tissue events and transformed plants are regenerated. Once tissue from each event having visual marker expression has reached a sufficient size when grown on selection medium with hygromycin, desiccation experiments can be conducted. Tissues (0.3-0.5mm in diameter) are sliced and transferred to an empty 60 mm x 25 mm Petri dish containing a piece of sterilized glass filter paper (VWR Glass Microfibre filter, 691). The
20 Petri dish is covered and placed in a container with a tight-seal cover. An open Petri dish with 15 mL of sterilized water is placed in the container. The container is placed in a dark culture room at 28°C. After 2-3 days of desiccation treatment, the tissues are either directly transferred to regeneration medium or selection medium with antibiotics and 20-50 uM glyphosate using Phytigel as a gelling agent for 2-3 weeks with sealed plates for
25 proliferation and regeneration. The tissues are transferred to regeneration medium with antibiotics and 20-50 uM glyphosate for another 2-4 weeks to generate shoots. Plates are placed in higher intensity light at 26-28°C. When shoots are strong enough, single plantlets are separated and transferred to soil. Leaf samples are collected for qPCR analysis.

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Example 8. Tobacco Excision Induction and Plant Regeneration from Desiccated T₁ Immature Seeds

T₁ immature seeds from transgenic tobacco plants are isolated, sterilized with 15% Clorox + 2 drops of Tween 20 and rinsed with autoclaved water 3 times. Sterilized
5 immature seeds are placed on sterile fiber glass filter paper in a Petri dish. The Petri dish is covered and moved into a sterile laminar hood unwrapped and incubated for 1-2 days until the seeds are desiccated. Desiccated immature seeds are then placed onto regeneration medium containing 100 mg/L cefotaxime and with 20-50 µM glyphosate for selection. One to 2 weeks later, DsRed expression is checked in the emerging shoots.

- 10 Immature seeds that have been properly desiccated have very weak or no DsRed expression as the gene is excised via the LoxP sites. Both transgenic and nontransgenic seeds without desiccation treatment will germinate well on glyphosate-free medium while germination will be completely inhibited for both of them on 20-50 µM glyphosate. Immature seeds that successfully underwent gene excision by desiccation will have
15 glyphosate resistance and regenerate on medium containing 20-50 µM glyphosate.

Healthy plantlets are transferred to regeneration medium in Phytatrays containing 100 mg/L cefotaxime and 20-50 µM glyphosate for further selection and growth.

Example 9. Natural Desiccation and Excision in Transgenic Mature Tobacco Seeds

- 20 *Mature seed sterilization, Selection/ Regeneration:*

T₁ mature tobacco seed transformed with AGL1/ PHP55062 are sterilized with 20% Clorox + 2 drops Tween 20 and rinsed with autoclaved water 3 times. Sterilized seeds are then transferred to regeneration medium containing 100 mg/L cefotaxime with 20-50 µM glyphosate for selection. After 5-10 days, DsRed expression is checked in the
25 emerging shoots. Seeds that have been excised will no longer have DsRed expression as the gene is cleaved via the Lox P sites. Those seeds that are successfully excised of DsRed will have glyphosate resistance and regenerate on medium containing glyphosate. Once seeds have healthy shoot and root formation, the plantlets are moved to soil or another regeneration medium containing 100 mg/L cefotaxime in Phytatrays with 20 or
30 50 µM glyphosate for further selection and growth.

Sowing dry tobacco T₁ seeds straight to soil and glyphosate resistance confirmation:

To confirm that the natural desiccation process that occurs during seed maturation would in fact allow for the excision of DsRed and resistance to glyphosate, seeds
5 collected from T₀ tobacco plants are germinated in soil. By planting seeds straight to soil without any treatments, excision would truly be a result of natural processes. After plants have germinated and grown to about 10-15 cm, the plants are sprayed with glyphosate + surfactant at 2X or 4X concentration (1X is equivalent to what is used in the field). Within 10-12 days, it is visibly evident which plants are not affected by the herbicide and
10 which plants are severely damaged.

Example 10. Soybean Excision Induction and Plant Regeneration from Transformed Tissue Events

Soybean Transformation:

15 Soybean (cv. Jack) mature seeds are sterilized and sliced into half longitudinally and half-seeds are used as an *Agrobacterium* infection target. *Agrobacterium* strain AGL1 containing the PHP55062 vector (a standard excision vector, Figure 8) is used for transformation. Alternatively, soybean embryogenic suspension cultures are transformed with the PHP55062 vector via *Agrobacterium*-mediated transformation as described
20 herein or by the method of particle gun bombardment (Klein *et al.* (1987) *Nature*, 327:70, which is herein incorporated by reference in its entirety).

Transgenic soybean plants are generated following the method described in U.S. Patent No. 7,473,822, which is herein incorporated by reference in its entirety, and 5 to 30 mg/L hygromycin B is used for selection.

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Excision of LoxP Cassette by Desiccation and Plant Regeneration with Glyphosate Selection:

Soybean desiccation experiments are conducted to induce excision from transformed tissue events and transformed plants are regenerated. Once tissue from each
30 event having visual marker expression has reached a sufficient size when grown on

selection medium with hygromycin, desiccation experiments can be conducted. Tissues (0.3-0.5mm in diameter) are sliced and transferred to an empty 60 mm x 25 mm Petri dish containing a piece of sterilized glass filter paper (VWR Glass Microfibre filter, 691). The Petri dish is covered and placed in a container with a tight-seal cover. An open Petri dish with 15 mL of sterilized water is placed in the container. The container is placed in a dark culture room at 28°C. After 2-3 days of desiccation treatment, the tissues are either directly transferred to regeneration medium with antibiotics and 20-50 µM glyphosate using Phytigel as a gelling agent for 2-3 weeks with sealed plates for proliferation and regeneration. The tissues are transferred to regeneration medium with antibiotics and 20-50 µM glyphosate for another 2-4 weeks to generate shoots. Plates are placed in higher intensity light at 26-28°C. When shoots are strong enough, single plantlets are separated and transferred to soil. Leaf samples were collected for qPCR analysis.

Example 11. Soybean Excision Induction and Plant Regeneration from Desiccated T₁

Immature Seeds

T₁ immature pods from transgenic soybean plants are harvested, sterilized with 15% Clorox + 2 drops of Tween 20 and rinsed with autoclaved water 3 times. Immature seeds are isolated from sterilized pods and placed on sterile fiber glass filter paper in a Petri dish. The Petri dish is covered and moved into a sterile laminar hood unwrapped and incubated for 1-2 days until the seeds are desiccated. Desiccated immature seeds are then placed onto regeneration medium containing 100 mg/L cefotaxime and with 20-50 µM glyphosate for selection. One to 2 weeks later, DsRed expression is checked in the emerging shoots. Immature seeds that have been properly desiccated will have very weak or no DsRed expression as the gene is excised via the LoxP sites. Both transgenic and nontransgenic seeds without desiccation treatment will germinate well on glyphosate-free medium while germination will be completely inhibited for both of them on 20-50 µM glyphosate. Immature seeds that successfully underwent gene excision by desiccation will have glyphosate resistance and regenerate on medium containing 20-50 µM glyphosate.

Healthy plantlets are transferred to regeneration medium in Phytatrays containing 100 mg/L cefotaxime and 20-50 μ M glyphosate for further selection and growth.

Example 12. Natural Desiccation and Excision of Transgenic Mature Soybean Seeds

5 *Mature seed sterilization, Selection/ Regeneration:*

T_1 mature soybean seed transformed with AGL1/PHP55062 are sterilized with 20% Clorox + 2 drops Tween 20 and rinsed with autoclaved water 3 times. Sterilized seeds are then transferred to regeneration medium containing 100 mg/L cefotaxime with 20-50 μ M glyphosate for selection. After 5-10 days, DsRed expression is checked in the
10 emerging shoots. Seeds that have been excised will no longer have DsRed expression as the gene is cleaved via the Lox P sites. Those seeds that are successfully excised of DsRed will have glyphosate resistance and regenerate on medium containing glyphosate. Once seeds have healthy shoot and root formation, the plantlets are moved to soil or another regeneration medium containing 100 mg/L cefotaxime in Phytatrays with 20 or
15 50 μ M glyphosate for further selection and growth.

Sowing dry soybean T_1 seeds straight to soil and glyphosate resistance confirmation:

To confirm that the natural desiccation process that occurs during seed maturation would in fact allow for the excision of DsRed and resistance to glyphosate, seeds
20 collected from T_0 soybean plants are germinated in soil. By planting seeds straight to soil without any treatments, excision would be a result of truly natural processes. After plants have germinated and grown to about 10-15 cm, the plants are sprayed with glyphosate + surfactant at 2X or 4X concentration (1X is equivalent to what is used in the field). Within 10 days, it is visibly evident which plants are not affected by the herbicide and
25 which plants are severely damaged.

Example 13. *Agrobacterium*-mediated Transformation of Wheat using Immature Embryos (IEs) with Standard and Sand treatments

Preparation of Agrobacterium Suspension:

Agrobacterium tumefaciens harboring vector of interest was streaked from a -80° frozen aliquot onto solid LB medium containing selection (kanamycin or spectinomycin). The *Agrobacterium* was cultured on the LB plate at 21°C in the dark for 2-3 days. A single colony was selected from the master plate and was streaked onto an 810D medium plate containing selection and it was incubated at 28°C in the dark overnight. A sterile spatula was used to collect *Agrobacterium* cells from the solid medium and cells were suspended in ~ 5mL wheat infection medium (WI4) with 400 uM acetosyringone (As) (Table 1). The OD of the suspension was adjusted to 0.1 at 600 nm using the same medium.

Wheat Immature Embryo Transformation:

Material preparation, sterilization and sand treatment

4-5 spikes were collected, containing immature seeds with 1.5-2.5mm embryos. Immature seeds/wheat grains were then isolated from the spike by pulling downwards on the awn and removing both sets of bracts (the lemma and palea). Wheat grains were surface-sterilized for 15 min in 20% (v/v) bleach (5.25% sodium hypochlorite) plus 1 drop of Tween 20, and then they were washed in sterile water 2-3 times. Immature embryos (IEs) were isolated from the wheat grains and were placed in 1.5 ml of the WI4 medium into 2 mL micro-centrifuge tubes. Immature embryos were isolated and placed in 1 mL of WI4 medium with 0.25 mL of autoclaved sand. The 2 mL microcentrifuge tubes containing the immature embryos were centrifuged at 6k for 30 seconds, vortexed at 4.5, 5 or 6 for 10 seconds, and then centrifuged at 6k for 30 seconds. Embryos were let stood for 20 minutes.

Embryo treatments with sand and Infection

WI4 medium was drawn off, and 1.0 ml of *Agrobacterium* suspension was added to the 2 mL microcentrifuge tubes containing the immature embryos. Embryos were let to stand for 20 minutes. The suspension of *Agrobacterium* and immature embryos was poured onto wheat co-cultivation medium, WC21 (Table 2). Any embryos left in the tube were transferred to the

plate using a sterile spatula. The immature embryos were placed embryo axis side down on the media, and it was ensured that the embryos were immersed in the solution. The plate was sealed with Parafilm tape and incubated in the dark at 25°C for 3 days of co-cultivation.

5 Media Scheme and selection

After 3 days of co-cultivation immature embryos were transferred embryo axis side down to DBC4 green tissue (GT) induction medium containing 100 mg/L cefotaxime (PhytoTechnology Lab., Shawnee Mission, KS) (Table 3). All embryos were then incubated at 26-28°C in dim light for two weeks, then were transferred to DBC6 tissue (GT) induction medium containing 100 mg/L cefotaxime for another two weeks (Table 4). Regenerable sectors appear 3-4 weeks after transformation and will be ready for regeneration after being isolated. Regenerable sectors were cut from the non-transformed tissues and placed on regeneration media MSA with 100 mg/L cefotaxime (Table 5). Sectors on MSA medium should be placed in bright light for 1.5- 2 weeks or until roots and elongated shoots have formed. After sectors have developed into small plantlets they were transferred to Phyta trays until plantlets are ready to be transferred to soil. During each transfer plantlets were checked for marker gene expression and any non-expressing or chimeric tissues were removed.

Table 11. Liquid Wheat Infection Medium

WI4	
DI water	1000mL
MS salt + Vitamins	4.43 g
Maltose	30 g
Glucose	10 g
MES	1.95 g
2,4-D (0.5 mg/L)	1 ml
Picloram (10 mg/ml)	200 µl
BAP (1 mg/L)	0.5 ml
Adjust PH to 5.8 with KOH	

Table 12. Wheat Co-cultivation Medium

WC21	
DI water	1000mL
MS salt + Vitamins	4.43 g
Maltose	30 g
MES	1.95 g
2,4-D (0.5 mg/L)	1 ml
Picloram (10 mg/ml)	200 µl
BAP (1 mg/L)	0.5 ml
50X CuSO4 (0.1 M)	49 µl
Adjust PH to 5.8 with KOH	
Add 3.5 g/L of Phytigel	

Post sterilization	
Acetosyringone (1 M)	400 μ l

Post sterilization	
Acetosyringone (1 M)	400 μ l

Table 13. DBC 4 medium

DBC4	
	1000m
dd H2O	L
MS salt	4.3 g
Maltose	30 g
Myo-inositol	0.25 g
N-Z-Amine-A	1 g
Proline	0.69 g
Thiamine-HCl (0.1 mg/mL)	10 mL
50X CuSO ₄ (0.1 M)	49 μ L
2,4-D (0.5mg/mL)	2 mL
BAP	1 mL
Adjust PH to 5.8 with KOH Add 3.5 g/L of Phytigel Post sterilization	
Cefotaxime (100 mg/ml)	1 ml

Table 14. DBC 6 medium

DBC6	
	1000m
dd H2O	L
MS salt	4.3 g
Maltose	30 g
Myo-inositol	0.25 g
N-Z-Amine-A	1 g
Proline	0.69 g
Thiamine-HCl (0.1 mg/mL)	10 mL
50X CuSO ₄ (0.1 M)	49 μ L
2,4-D (0.5 mg/mL)	1 mL
BAP	2 mL
Adjust PH to 5.8 with KOH Add 3.5 g/L of Phytigel Post sterilization	
Cefotaxime (100 mg/ml)	1 ml

Table 15. Regeneration MSA medium

MSA	
dd H2O	1000mL
MS salt + Vitamins(M519)	4.43 g
Sucrose	20g
Myo- Inositol	1 g
Adjust PH to 5.8 with KOH Add 3.5 g/L of Phytigel	
Post sterilization	
Cefotaxime (100 mg/ml)	1 ml

Wheat *Agrobacterium*-mediated transformation using immature embryos were
 5 conducted with standard treatments and sand treatments to compare the transformation
 frequencies at T0 plant level.

Table 16 shows the transformation frequencies at T0 plant level (T0) for
 transformation experiments with standard and sand treatments using Standard vector for
 Pioneer elite spring wheat variety SBC0456D; the binary vectors are difficult constructs for
 10 transformation because the visual marker is driven by weak promoter for selection. All
 experiments were performed with 4.5-6 vortex speed for both standard and sand treatments.
 Data showed that T0 frequencies ranged from 0% to 1.2% for standard treatments. For sand
 treatments, T0 frequencies ranged from 5.9% to 6.8%. Results indicated that experiments
 conducted with sand treatments had higher transformation frequencies comparing to standard
 15 treatments.

Table 16. *Agrobacterium*-mediated transformation of immature embryos using standard vector with standard and sand treatments

Treatments	Standard Vortex at 4.5	0.25mL sand Vortex at 4.5	Standard Vortex at 5	0.25mL sand Vortex at 5	Standard Vortex at 6	0.25mL sand Vortex at 6
Transformation Frequency (T0)	0% (0/52)	5.9% (3/51)	0% (0/46)	18.6% (8/43)	0% (0/48)	13.3% (6/45)
			0% (0/54)	3.7% (2/54)	0% (0/66)	1.4% (1/72)
			2.8% (2/71)	1.5% (1/65)		
Average	0% (0/52)	5.9% (3/51)	1.2% (2/171)	6.8% (11/162)	0% (0/114)	6.0% (7/117)

5 All publications and patent applications mentioned in the specification are indicative of the level of those skilled in the art to which this invention pertains. All publications and patent applications are herein incorporated by reference to the same extent as if each individual publication or patent application was specifically and individually indicated to be incorporated by reference.

10 Many modifications and other embodiments of the inventions set forth herein will come to mind to one skilled in the art to which these inventions pertain having the benefit of the teachings presented in the foregoing descriptions and the associated drawings. Therefore, it is to be understood that the inventions are not to be limited to the specific embodiments disclosed and that modifications and other embodiments are intended to be included within the scope of the appended claims. Although specific terms are employed
15 herein, they are used in a generic and descriptive sense only and not for purposes of limitation.

THAT WHICH IS CLAIMED:

1. A polynucleotide construct comprising:
 - a) an excision cassette, comprising an expression cassette A (EC_A)
5 comprising:
 - i) a coding polynucleotide A (CP_A) encoding a site specific recombinase; and
 - ii) an inducible promoter A (P_A) operably linked to the CP_A;
 - b) a first and a second recombination site flanking the excision
10 cassette;
 - c) a coding polynucleotide B (CP_B) encoding a herbicide tolerance polypeptide; and
 - d) a promoter B (P_B), wherein the P_B is operably linked to the CP_B after excision of the excision cassette.
15
2. The polynucleotide construct of claim 1, wherein the inducible promoter P_A is selected from the group consisting of a stress-inducible promoter and a chemical-inducible promoter.
- 20 3. The polynucleotide construct of claim 2, wherein said chemical-inducible promoter comprises a promoter comprising a tet operator.
4. The polynucleotide construct of claim 3, wherein said polynucleotide construct further comprises a coding polynucleotide F (CP_F) encoding a sulfonylurea-
25 responsive transcriptional repressor protein, wherein said CP_F is operably linked to a promoter active in a plant cell.
5. The polynucleotide construct of claim 2, wherein the stress-inducible promoter can be induced in response to cold, drought, high salinity, desiccation, or a
30 combination thereof.

6. The polynucleotide construct of claim 2, wherein the stress-inducible promoter comprises a nucleotide sequence selected from the group consisting of:

- 5 NO: 18;
- a) the nucleotide sequence having the sequence set forth in SEQ ID NO: 18;
 - b) a nucleotide sequence having at least 70% sequence identity to the sequence set forth in SEQ ID NO: 18;
 - c) a nucleotide sequence comprising at least 50 contiguous nucleotides of the sequence set forth in SEQ ID NO: 18;
 - 10 d) the nucleotide sequence set forth in nucleotides 291-430 of SEQ ID NO: 18; and
 - e) a nucleotide sequence having at least 70% sequence identity to the sequence set forth in nucleotides 291-430 of SEQ ID NO: 18.

15 7. The polynucleotide construct of claim 1, wherein the P_B is a constitutive promoter.

8. The polynucleotide construct of claim 7, wherein the P_B is selected from the group consisting of a ubiquitin promoter, an oleosin promoter, an actin promoter, and
20 a *Mirabilis* mosaic virus (MMV) promoter.

9. The polynucleotide construct of claim 1, wherein the excision cassette further comprises a coding polynucleotide C (CP_C) encoding a selectable marker, wherein the CP_C is operably linked to a promoter active in a plant cell.

25

10. The polynucleotide construct of claim 9, wherein the CP_C is operably linked to P_B prior to excision of the excision cassette.

11. The polynucleotide construct of claim 9, wherein the excision cassette
30 further comprises a promoter C (P_C) operably linked to the CP_C .

12. The polynucleotide construct of claim 11, wherein the P_C is a constitutive promoter.

5 13. The polynucleotide construct of claim 9, wherein the selectable marker is selected from the group consisting of a fluorescent protein, an antibiotic resistance polypeptide, a herbicide tolerance polypeptide, and a metabolic enzyme.

10 14. The polynucleotide construct of claim 1, wherein the herbicide tolerance polypeptide encoded by CP_B comprises a glyphosate-N-acetyltransferase (GLYAT) polypeptide or an ALS inhibitor-tolerance polypeptide.

15 15. The polynucleotide construct of claim 14, wherein said ALS inhibitor-tolerance polypeptide comprises the highly resistant ALS (HRA) mutation of acetolactate synthase.

20 16. The polynucleotide construct of claim 1, wherein the excision cassette further comprises a coding polynucleotide D (CP_D) encoding a cell proliferation factor operably linked to a promoter active in a plant cell.

17. The polynucleotide construct of claim 16, wherein the cell proliferation factor is selected from a WUSCHEL polypeptide and a babyboom polypeptide.

25 18. The polynucleotide construct of claim 17, wherein the babyboom polypeptide comprises at least two AP2 domains and at least one of the following amino acid sequences:

a) the amino acid sequence set forth in SEQ ID NO: 67 or an amino acid sequence that differs from the amino acid sequence set forth in SEQ ID NO: 67 by one amino acid; and

b) the amino acid sequence set forth in SEQ ID NO: 68 or an amino acid sequence that differs from the amino acid sequence set forth in SEQ ID NO: 68 by one amino acid.

5 19. The polynucleotide construct of claim 17, wherein the CP_D has a nucleotide sequence selected from the group consisting of:

a) the nucleotide sequence set forth in SEQ ID NO: 55, 57, 58, 60, 74, 76, 78, 80, 82, 84, 86, 87, 88, 90, 92, 94, 96, 98, 99, or 101;

b) a nucleotide sequence having at least 70% sequence identity to
10 SEQ ID NO: 55, 57, 58, 60, 74, 76, 78, 80, 82, 84, 86, 87, 88, 90, 92, 94, 96, 98, 99, or 101;

c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 56, 59, 75, 77, 79, 81, 83, 85, 89, 91, 93, 95, 97, 100, or 102; and

15 d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 70% sequence identity to the amino acid sequence set forth in SEQ ID NO: 56, 59, 75, 77, 79, 81, 83, 85, 89, 91, 93, 95, 97, 100, or 102.

20 20. The polynucleotide construct of claim 17, wherein the polynucleotide encoding a WUSCHEL polypeptide has a nucleotide sequence selected from the group consisting of:

a) the nucleotide sequence set forth in SEQ ID NO: 103, 105, 107, or 109; and

b) a nucleotide sequence having at least 70% sequence identity to
25 SEQ ID NO: 103, 105, 107, or 109;

c) a nucleotide sequence encoding a polypeptide having the amino acid sequence set forth in SEQ ID NO: 104, 106, 108, or 110; and

d) a nucleotide sequence encoding a polypeptide having an amino acid sequence having at least 70% sequence identity to SEQ ID NO: 104, 106, 108, or
30 110.

21. The polynucleotide construct of claim 20, wherein the polynucleotide encoding a WUSCHEL polypeptide is operably linked to a maize In2-2 promoter or a nopaline synthase promoter.

5

22. The polynucleotide construct of claim 16, wherein the excision cassette further comprises a promoter D (P_D) operably linked to the CP_D .

23. The polynucleotide construct of claim 22, wherein the P_D is a constitutive promoter.

10

24. The polynucleotide construct of claim 23, wherein the P_D is a ubiquitin promoter or an oleosin promoter.

25. The polynucleotide construct of claim 16, wherein the excision cassette comprises at least a first coding polynucleotide D (CP_{D1}) encoding a babyboom polypeptide and a second coding polynucleotide D (CP_{D2}) encoding a WUSCHEL polypeptide.

15

26. The polynucleotide construct of claim 1, wherein the polynucleotide construct further comprises a coding polynucleotide E (CP_E) encoding a polypeptide of interest, wherein the CP_E is operably linked to a promoter active in a plant cell.

20

27. The polynucleotide construct of claim 26, wherein the CP_E is outside of the first and a second recombination sites flanking the excision cassette.

25

28. A host cell comprising the polynucleotide construct of claim 1.

29. A plant cell comprising the polynucleotide construct of claim 1.

30

30. A plant or plant part comprising the plant cell of claim 29.

31. The plant or plant part of claim 30, wherein the plant or plant part is a dicot.

5

32. The plant or plant part of claim 30, wherein the plant or plant part is a monocot.

33. The plant or plant part of claim 32, wherein the monocot is selected from the group consisting of maize, rice, sorghum, barley, millet, oat, rye, triticale, sugarcane, switch grass, and turf/forage grass.

10

34. The plant or plant part of claim 30, wherein the plant or plant part is recalcitrant to transformation.

15

35. The plant or plant part of claim 30, wherein the plant part is a seed.

36. A method for producing a transgenic plant or plant part, said method comprising introducing the polynucleotide construct of claim 1 into a plant or plant part.

20

37. A method for regulating the expression of a herbicide tolerance polynucleotide, wherein the method comprises:

a) providing the host cell of claim 28; and,
b) inducing the expression of the site-specific recombinase, thereby excising the excision cassette from the polynucleotide construct and expressing the herbicide tolerance polynucleotide.

25

38. A method for selecting a herbicide tolerant plant cell, the method comprising the steps of:

A) providing a population of plant cells, wherein at least one plant cell in the population comprises the polynucleotide construct of claim 1;

B) inducing the expression of the site-specific recombinase; and

C) contacting the population of plant cells with a herbicide to which the herbicide tolerance polypeptide confers tolerance, thereby selecting for a plant cell having tolerance to the herbicide.

39. The method of claim 38, wherein the method further comprises introducing the polynucleotide construct into the at least one plant cell before step A).

40. The method of claim 38, wherein the inducible promoter A (P_A) is induced in response to cold, drought, desiccation, high salinity or a combination thereof.

41. The method of claim 38, wherein the inducing comprises desiccating the population of plant cells.

42. The method of claim 41, wherein the desiccating occurs during the maturation of an immature seed.

43. The method of claim 38, wherein the excision cassette further comprises a coding polynucleotide C (CP_C), wherein the CP_C encodes a selectable marker operably linked to a promoter, and wherein the method further comprises a selection step prior to step B), wherein those plant cells within the population of plant cells that comprise the selectable marker are identified and wherein these selected plant cells comprise the population of plant cells that are induced in step B).

44. A method for increasing the transformation efficiency of a plant tissue, the method comprising the steps of:

a) providing a population of plant cells, wherein at least one plant cell in the population comprises the polynucleotide construct of claim 1;

b) culturing the population of plant cells in the absence of a herbicide to which the herbicide tolerance polypeptide confers herbicide resistance for a period of time sufficient for the population of plant cells to proliferate;

5 c) inducing the expression of the site-specific recombinase, thereby excising the excision cassette;

d) contacting the population of plant cells from c) with the herbicide to which the herbicide tolerance polypeptide confers tolerance; and

e) selecting for a plant cell having tolerance to the herbicide, wherein the transformation frequency is increased compared to a comparable plant cell not
10 comprising the excision cassette and selected directly by herbicide selection.

45. The method of claim 44, wherein the inducing comprises desiccating the population of plant cells.

15 46. The method of claim 44, wherein the population of plant cells is cultured in the absence of the herbicide to which the herbicide tolerance polypeptide confers herbicide resistance for about 1 hour to about 6 weeks prior to excision.

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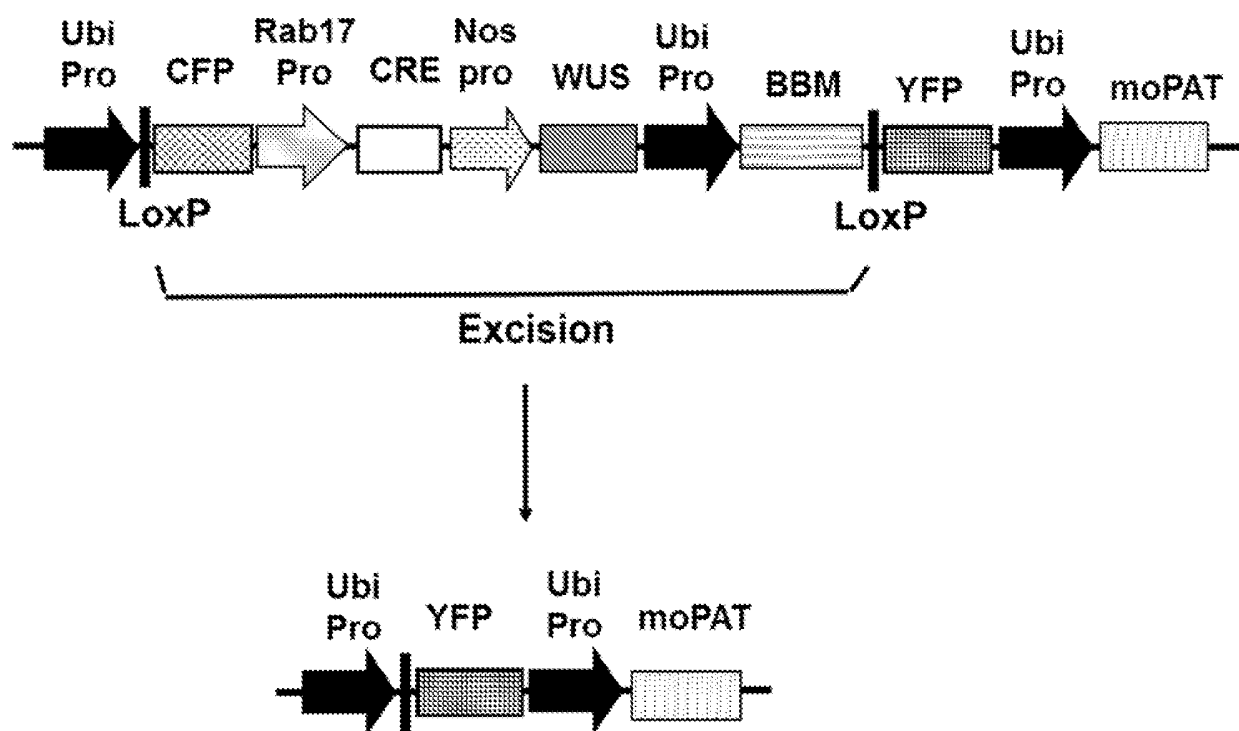


FIG. 1

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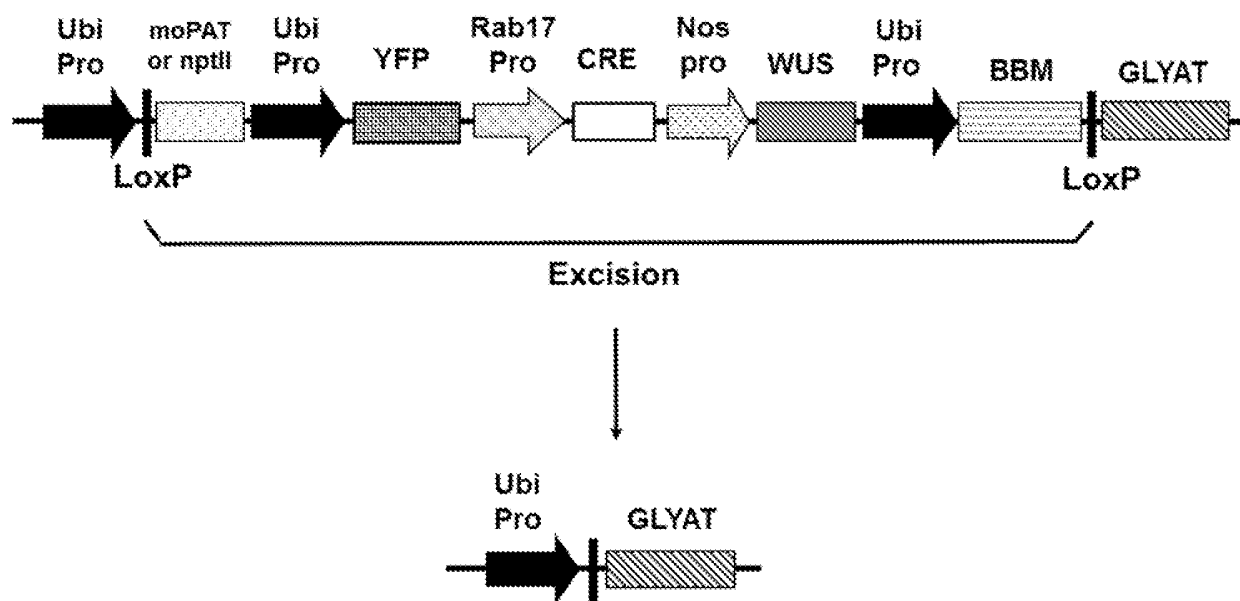


FIG. 2

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FIG. 3

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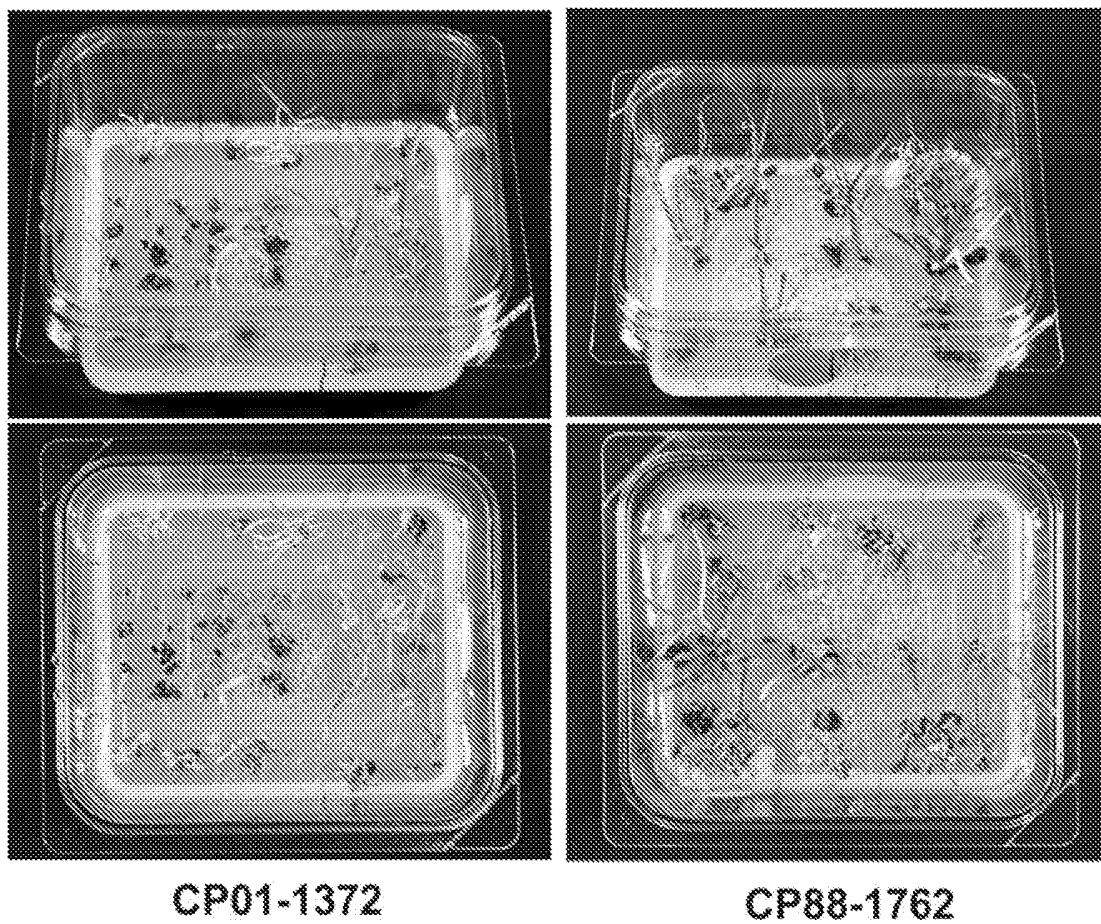


FIG. 4

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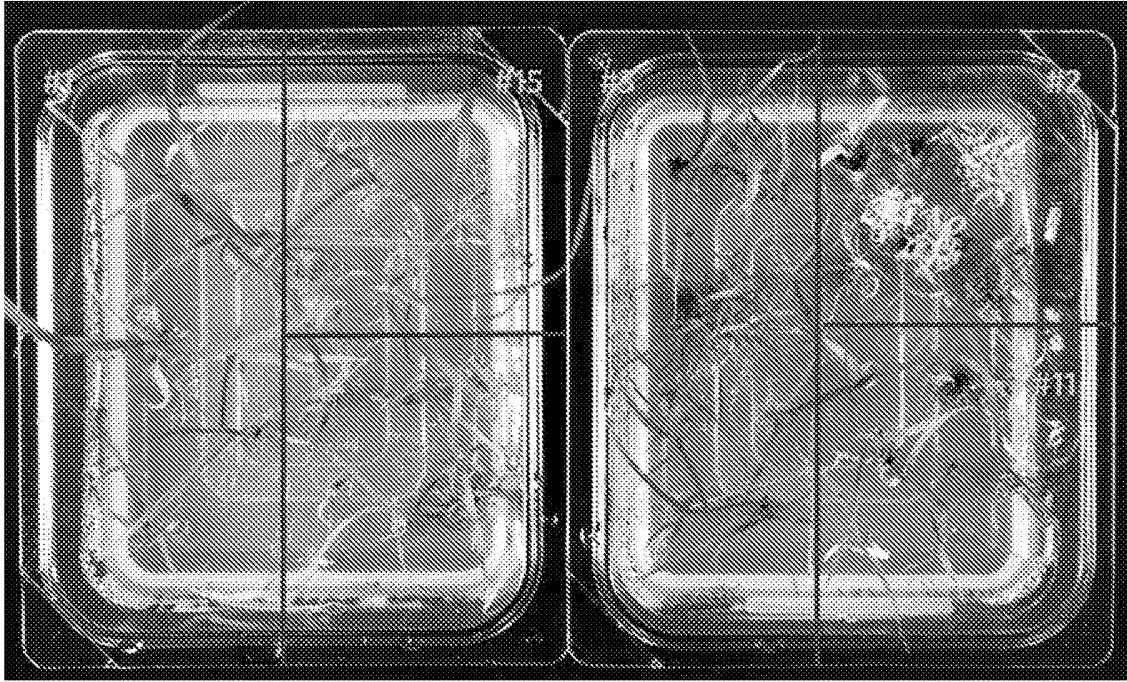


FIG. 5

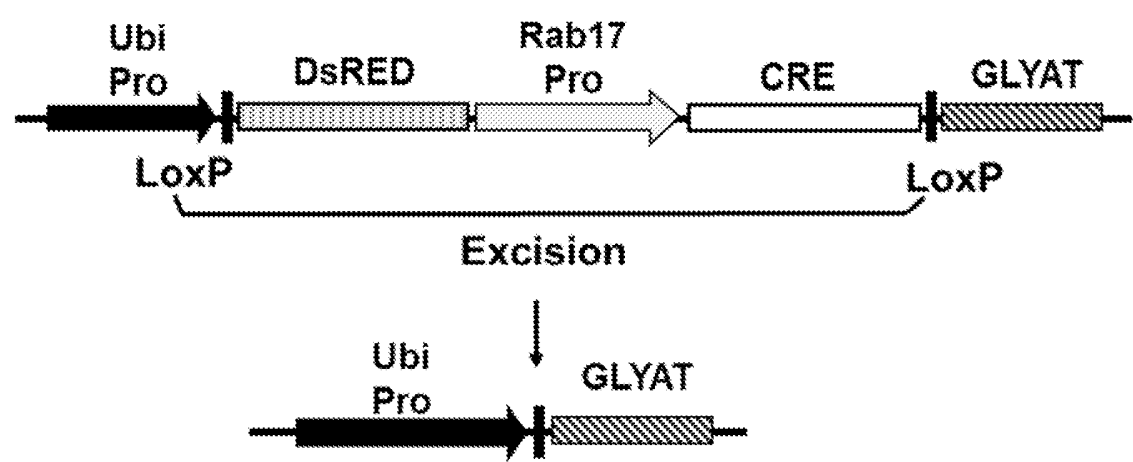


FIG. 6

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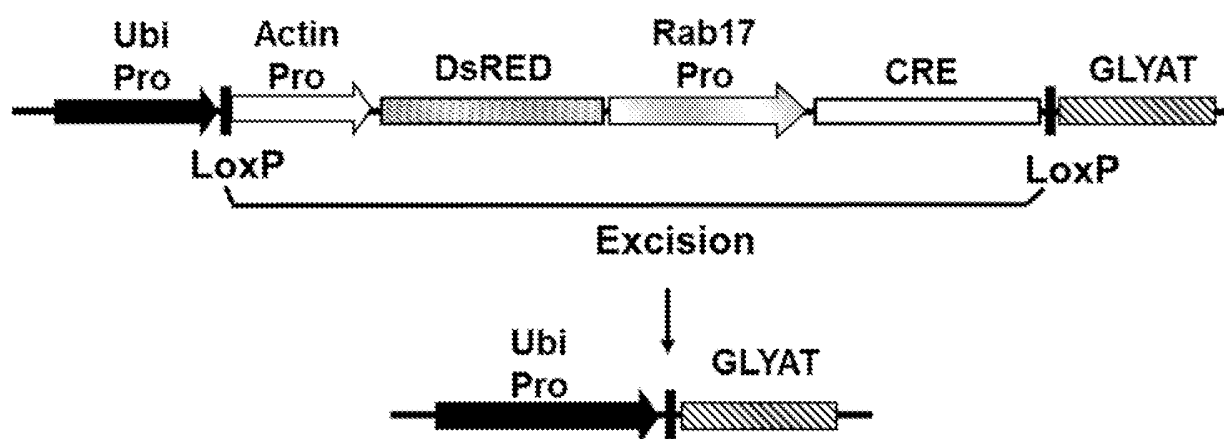


FIG. 7

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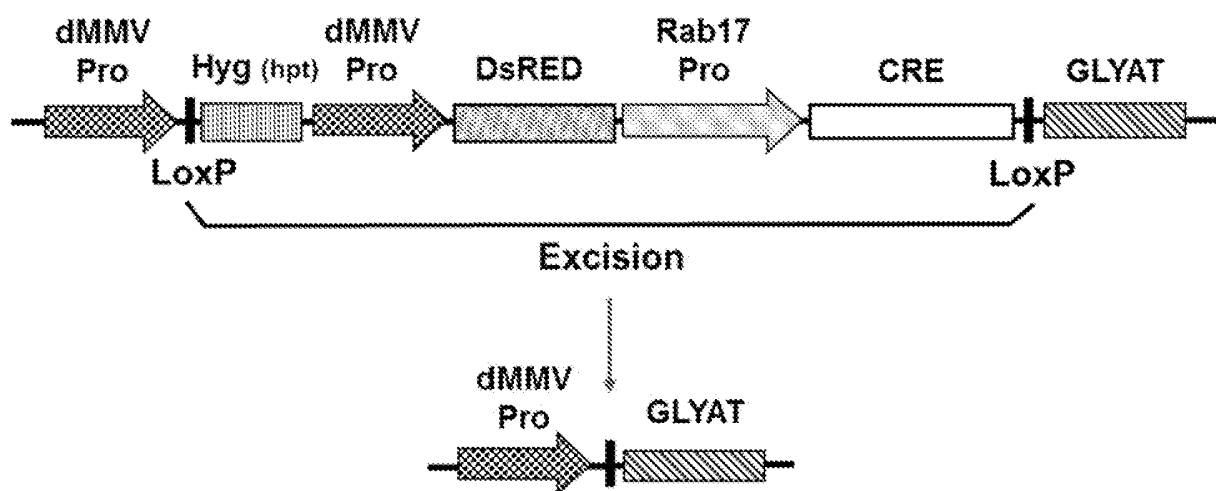


FIG. 8

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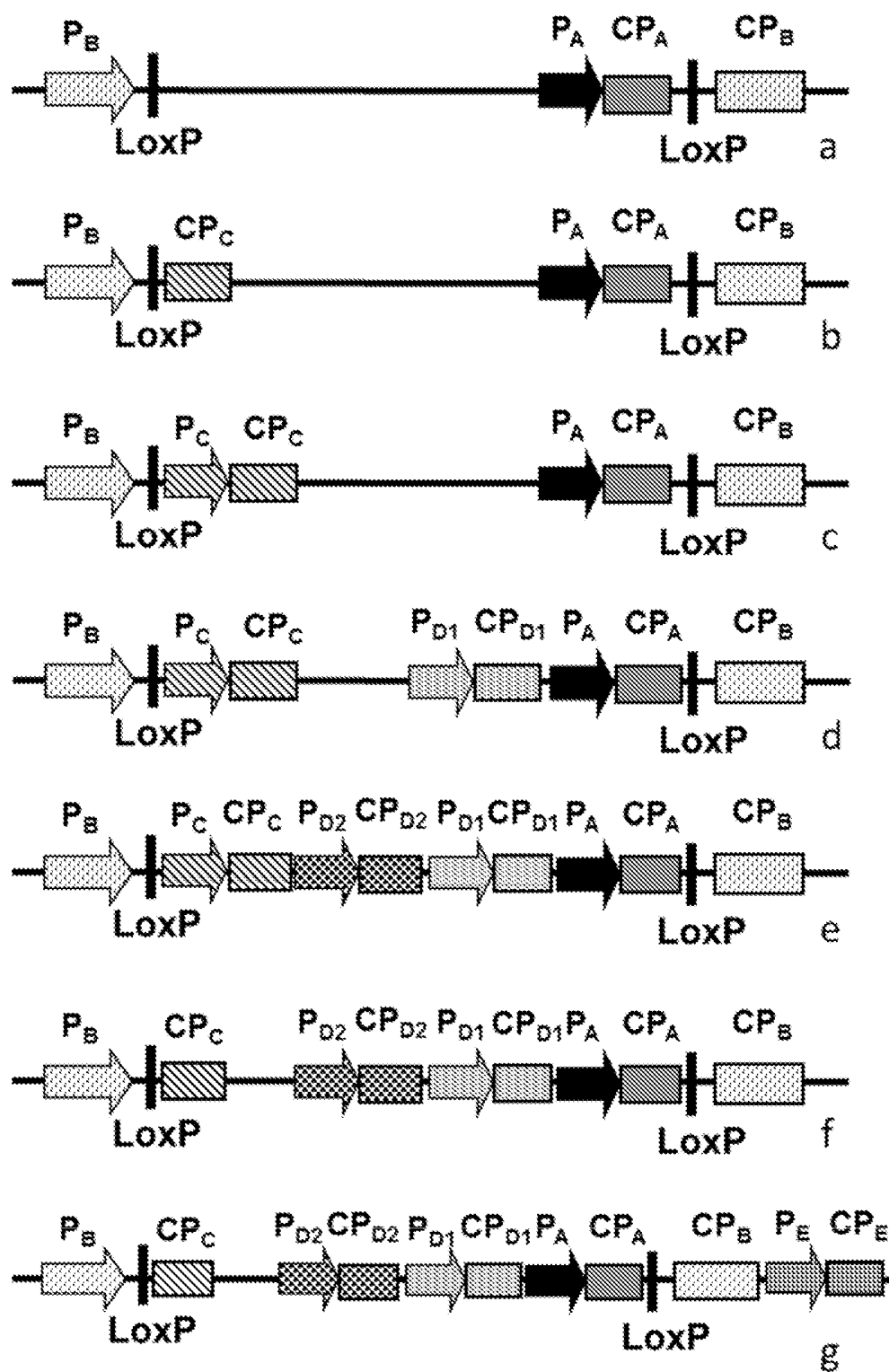


FIG. 9

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 Zhao, Zuo-Yu

<120> Methods and Compositions for Producing
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cct gcat t ga at aat ggat g agcaccggg a aaat ccgcgt acccaact t t cgagaagaac 300
cgagacgt gg cgggcccgggc caccgacgca cggcaccagc gact gcacac gt cccgccgg 360
cgt acgt gt a cgt gct gt t c cct cact ggc cgcccaat cc act cat gcat gcccacgt ac 420
acccct gccg t ggcgcgcc agat cct aat cct t t cgccg t t ct gact t ct gct gcct a 480
t aaat ggcgg cat cgaccgt cacct gct t c accaccggcg agccacat cg agaacacgat 540
cgagcacaca agcacgaaga ct cgt t t agg agaaaccaca aaccaccaag ccgt gcaagc 600
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aggct 665

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<210> 32
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 <213> Artificial Sequence

<220>
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<220>
 <223> tet operator

<400> 32
 act ct at cag t gat agagt 19

<210> 33
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Met Pro Gln Phe Asp Ile Leu Oys Lys Thr Pro Pro Lys Val Leu Val
1 5 10 15

agg cag ttc gtg gag agg ttc gag agg ccc tcc ggc gag aag atc gcc 96
Arg Gln Phe Val Glu Arg Phe Glu Arg Pro Ser Gly Glu Lys Ile Ala
20 25 30

ctc tgc gcc gcc gag ct c acc tac ct c tgc tgg atg at c acc cac aac 144
Leu Oys Ala Ala Glu Leu Thr Tyr Leu Oys Trp Met Ile Thr His Asn
35 40 45

ggc acc gcc att aag agg gcc acc ttc atg tca tac aac acc atc atc 192
Gly Thr Ala Ile Lys Arg Ala Thr Phe Met Ser Tyr Asn Thr Ile Ile
50 55 60

tcc aac tcc ct c tcc ttc gac atc gtg aac aag tcc ct c cag ttc aaa 240
Ser Asn Ser Leu Ser Phe Asp Ile Val Asn Lys Ser Leu Gln Phe Lys
65 70 75 80

tac aag acc cag aag gcc acc atc ct c gag gcc tcc ct c aag aag ct c 288
Tyr Lys Thr Gln Lys Ala Thr Ile Leu Glu Ala Ser Leu Lys Lys Leu
85 90 95

atc ccc gcc tgg gag ttc acc atc atc ccc tac tac ggc cag aag cac 336
Ile Pro Ala Trp Glu Phe Thr Ile Ile Pro Tyr Tyr Gly Gln Lys His
100 105 110

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5312WOPCT_SEQ_LI STI NG TXT

cag G n	tcc Ser	gac Asp 115	atc Ile	acc Thr	gac Asp	atc Ile	gtg Val 120	tca Ser	tcc Ser	ctc Leu	cag G n	ctt Leu 125	cag G n	ttc Phe	gag G u	384
tcc Ser	tcc Ser 130	gag G u	gag G u	gct Ala	gac Asp	aag Lys 135	ggc Gly	aac Asn	tcc Ser	cac His	tcc Ser 140	aag Lys	aag Lys	atg Met	ctg Leu	432
aag Lys 145	gcc Ala	ctc Leu	ctc Leu	tcc Ser	gag G u 150	ggc Gly	gag G u	tcc Ser	atc Ile	tgg Trp 155	gag G u	atc Ile	acc Thr	gag G u	aag Lys 160	480
atc Ile	ctc Leu	aac Asn	tcc Ser	ttc Phe 165	gag G u	tac Tyr	acc Thr	tcc Ser	agg Arg 170	ttc Phe	act Thr	aag Lys	acc Thr	aag Lys 175	acc Thr	528
ctc Leu	tac Tyr	cag G n 180	ttc Phe	ctc Leu	ttc Phe	ctc Leu	gcc Ala 185	acc Thr	ttc Phe	atc Ile	aac Asn	tgc Cys	ggc Gly 190	agg Arg	ttc Phe	576
tca Ser	gac Asp	atc Ile 195	aag Lys	aac Asn	gtg Val	gac Asp	ccc Pro 200	aag Lys	tcc Ser	ttc Phe	aag Lys	ctc Leu 205	gtg Val	cag G n	aac Asn	624
aag Lys 210	tac Tyr	ctc Leu	ggc Gly	gtg Val	atc Ile 215	atc Ile	cag G n	tgc Cys	ctc Leu	gtg Val	acc Thr 220	gag G u	acc Thr	aag Lys	acc Thr	672
tcc Ser 225	gtg Val	tcc Ser	agg Arg	cac His 230	atc Ile	tac Tyr	ttc Phe	ttc Phe	tcc Ser	gct Ala 235	cgc Arg	ggc Gly	agg Arg	atc Ile	gac Asp 240	720
ccc Pro	ctc Leu	gtg Val	tac Tyr	ctc Leu 245	gac Asp	gag G u	ttc Phe	ctc Leu	agg Arg 250	aac Asn	tca Ser	gag G u	ccc Pro	gtg Val 255	ctc Leu	768
aag Lys	agg Arg	gtg Val	aac Asn 260	agg Arg	acc Thr	ggc Gly	aac Asn	tcc Ser 265	tcc Ser	tcc Ser	aac Asn	aag Lys	cag G n 270	gag G u	tac Tyr	816
cag G n	ctc Leu	ctc Leu 275	aag Lys	gac Asp	aac Asn	ctc Leu	gtg Val 280	agg Arg	tcc Ser	tac Tyr	aac Asn	aag Lys 285	gcc Ala	ctc Leu	aag Lys	864
aag Lys 290	aac Asn	gcc Ala	ccc Pro	tac Tyr	tcc Ser	atc Ile 295	ttc Phe	gcc Ala	atc Ile	aag Lys	aac Asn 300	ggc Gly	ccc Pro	aag Lys	tcc Ser	912
cac His 305	atc Ile	ggt Gly	agg Arg	cac His 310	ctc Leu	atg Met	acc Thr	tcc Ser	ttc Phe	ctc Leu 315	tca Ser	atg Met	aag Lys	ggc Gly	ctc Leu 320	960
acc Thr	gag G u	ctc Leu	acc Thr	aac Asn 325	gtg Val	gtg Val	ggc Gly	aac Asn	tgg Trp 330	tcc Ser	gac Asp	aag Lys	agg Arg	gcc Ala 335	tcc Ser	1008
gcc Ala	gtg Val	gcc Ala	agg Arg 340	acc Thr	acc Thr	tac Tyr	acc Thr	cac His 345	cag G n	atc Ile	acc Thr	gcc Ala 350	atc Ile	ccc Pro	gac Asp	1056
cac His	tac Tyr	ttc Phe 355	gcc Ala	ctc Leu	gtg Val	tca Ser	agg Arg 360	tac Tyr	tac Tyr	gcc Ala	tac Tyr	gac Asp 365	ccc Pro	atc Ile	tcc Ser	1104
aag Lys 370	gag G u	atg Met	atc Ile	gcc Ala	ctc Leu	aag Lys 375	gac Asp	gag G u	act Thr	aac Asn	ccc Pro 380	atc Ile	gag G u	gag G u	tgg Trp	1152

5312WOPCT_SEQ_LI STI NG TXT

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G n	H i s	I l e	G u	G n	L e u	L y s	G l y	S e r	A l a	G u	G l y	S e r	I l e	A r g	T y r	
385					390					395					400	
ccc	gcc	tgg	aac	ggc	atc	atc	tcc	cag	gag	gtg	ctc	gac	tac	ctc	tcc	1248
Pro	Ala	Trp	Asn	Gly	Ile	Ile	Ser	Gln	Glu	Val	Leu	Asp	Tyr	Leu	Ser	
				405					410					415		
tcc	tac	atc	aac	agg	agg	atc	tga									1272
Ser	Tyr	Ile	Asn	Arg	Arg	Ile										
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			20					25					30			
Leu	Cys	Ala	Ala	G u	Leu	Thr	Tyr	Leu	Cys	Trp	Met	Ile	Thr	H i s	Asn	
		35					40					45				
G y	Thr	Ala	Ile	Lys	Arg	Ala	Thr	Phe	Met	Ser	Tyr	Asn	Thr	Ile	Ile	
	50					55					60					
Ser	Asn	Ser	Leu	Ser	Phe	Asp	Ile	Val	Asn	Lys	Ser	Leu	G n	Phe	Lys	
65					70				75					80		
Tyr	Lys	Thr	G n	Lys	Ala	Thr	Ile	Leu	G u	Ala	Ser	Leu	Lys	Lys	Leu	
			85					90						95		
Ile	Pro	Ala	Trp	G u	Phe	Thr	Ile	Ile	Pro	Tyr	Tyr	G y	G n	Lys	H i s	
			100					105					110			
G n	Ser	Asp	Ile	Thr	Asp	Ile	Val	Ser	Ser	Leu	G n	Leu	G n	Phe	G u	
		115					120					125				
Ser	Ser	G u	G u	Ala	Asp	Lys	G y	Asn	Ser	H i s	Ser	Lys	Lys	Met	Leu	
	130					135					140					
Lys	Ala	Leu	Leu	Ser	G u	G y	G u	Ser	Ile	Trp	G u	Ile	Thr	G u	Lys	
145					150					155					160	
Ile	Leu	Asn	Ser	Phe	G u	Tyr	Thr	Ser	Arg	Phe	Thr	Lys	Thr	Lys	Thr	
				165					170					175		
Leu	Tyr	G n	Phe	Leu	Phe	Leu	Ala	Thr	Phe	Ile	Asn	Cys	G y	Arg	Phe	
			180					185					190			
Ser	Asp	Ile	Lys	Asn	Val	Asp	Pro	Lys	Ser	Phe	Lys	Leu	Val	G n	Asn	
	195						200					205				
Lys	Tyr	Leu	G y	Val	Ile	Ile	G n	Cys	Leu	Val	Thr	G u	Thr	Lys	Thr	
	210					215										
Ser	Val	Ser	Arg	H i s	Ile	Tyr	Phe	Phe	Ser	Ala	Arg	G y	Arg	Ile	Asp	
225				230						235					240	
Pro	Leu	Val	Tyr	Leu	Asp	G u	Phe	Leu	Arg	Asn	Ser	G u	Pro	Val	Leu	
				245					250					255		
Lys	Arg	Val	Asn	Arg	Thr	G y	Asn	Ser	Ser	Asn	Lys	G n	G u	Tyr		
			260					265						270		
G n	Leu	Leu	Lys	Asp	Asn	Leu	Val	Arg	Ser	Tyr	Asn	Lys	Ala	Leu	Lys	
	275					280						285				
Lys	Asn	Ala	Pro	Tyr	Ser	Ile	Phe	Ala	Ile	Lys	Asn	G y	Pro	Lys	Ser	
	290					295					300					
H i s	Ile	G y	Arg	H i s	Leu	Met	Thr	Ser	Phe	Leu	Ser	Met	Lys	G y	Leu	
305					310					315					320	
Thr	G u	Leu	Thr	Asn	Val	Val	G y	Asn	Trp	Ser	Asp	Lys	Arg	Ala	Ser	
				325					330					335		
Ala	Val	Ala	Arg	Thr	Thr	Tyr	Thr	H i s	G n	Ile	Thr	Ala	Ile	Pro	Asp	

5312 WOPCT_SEQ_LI STI NG TXT

340
 His Tyr Phe Ala Leu Val Ser Arg Tyr Tyr Ala Tyr Asp Pro Ile Ser
 355
 Lys Glu Met Ile Ala Leu Lys Asp Glu Thr Asn Pro Ile Glu Glu Trp
 370
 Gln His Ile Glu Gln Leu Lys Gly Ser Ala Glu Gly Ser Ile Arg Tyr
 385
 Pro Ala Trp Asn Gly Ile Ile Ser Gln Glu Val Leu Asp Tyr Leu Ser
 400
 Ser Tyr Ile Asn Arg Arg Ile
 420

<210> 35
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<220>
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<220>
 <221> CDS
 <222> (1) . . . (1032)

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 1 5 10 15
 gac gcg acg tcc gat gaa gt c agg aag aac ct c at g gac at g t t c cgc 96
 Asp Ala Thr Ser Asp Glu Val Arg Lys 25 Asn Leu Met Asp Met Phe Arg
 20 25 30
 gac agg caa gcg t t c agc gag cac acc t gg aag at g ct g ct c t c c gt c 144
 Asp Arg Gln Ala Phe Ser Glu His Thr Trp Lys Met Leu Leu Ser Val
 35 40 45
 t gc cgc t cc t gg gct gca t gg t gc aag ct g aac aac agg aag t gg t t c 192
 Cys Arg Ser Trp Ala Ala Trp 55 Cys Lys Leu Asn Asn 60 Arg Lys Trp Phe
 50 55 60
 ccc gct gag ccc gag gac gt g agg gat t ac ct t ct g t ac ct g caa gct 240
 Pro Ala Glu Pro Glu Asp Val Arg Asp Tyr Leu Leu Tyr Leu Gln Ala 80
 65 70 75 80
 cgc ggg ct g gca gt g aag acc at c cag caa cac ct t gga caa ct g aac 288
 Arg Gly Leu Ala Val 85 Lys Thr Ile Gln Gln His Leu Gly Gln Leu Asn
 90 95
 at g ct t cac agg cgc t cc ggc ct c ccg cgc ccc agc gac t cg aac gcc 336
 Met Leu His Arg Arg Ser Gly Leu Pro Arg Pro Ser Asp Ser Asn Ala
 100 105 110
 gt g agc ct c gt c at g cgc cgc at c agg aag gaa aac gt c gat gcc ggc 384
 Val Ser Leu Val Met Arg Arg Ile Arg Lys Glu Asn Val Asp Ala Gly
 115 120 125
 gaa agg gca aag cag gcc ct c gcg t t c gag agg acc gat t t c gac cag 432
 Glu Arg Ala Lys Gln Ala Leu Ala Phe Glu Arg Thr 140 Asp Phe Asp Gln
 130 135 140
 gt c cgc agc ct g at g gag aac agc gac agg t gc cag gac att agg aac 480
 Val Arg Ser Leu Met Glu Asn Ser Asp Arg Cys Gln Asp Ile Arg Asn
 145 150 155 160

5312WOPCT_SEQ_LI STI NG TXT

ct g	gcg	t t c	ct c	gga	att	gca	t ac	aac	acg	ct c	ct c	agg	at c	gcg	gaa	528
Leu	Al a	Phe	Leu	G y	I l e	Al a	Tyr	Asn	Thr	Leu	Leu	Arg	I l e	Al a	G u	
				165					170					175		
att	gcc	cgc	att	cgc	gt g	aag	gac	att	agc	cgc	acc	gac	ggc	ggc	agg	576
I l e	Al a	Arg	I l e	Arg	Val	Lys	Asp	I l e	Ser	Arg	Thr	Asp	G y	G y	Arg	
			180					185					190			
at g	ct t	at c	cac	att	ggc	agg	acc	aag	acg	ct c	gt t	t cc	acc	gca	ggc	624
Met	Leu	I l e	His	I l e	G y	Arg	Thr	Lys	Thr	Leu	Val	Ser	Thr	Al a	G y	
		195					200					205				
gt c	gaa	aag	gcc	ct c	agc	ct c	gga	gt g	acc	aag	ct c	gt c	gaa	cgc	t gg	672
Val	G u	Lys	Al a	Leu	Ser	Leu	G y	Val	Thr	Lys	Leu	Val	G u	Arg	Trp	
	210					215					220					
at c	t cc	gt g	t cc	ggc	gt c	gcg	gac	gac	cca	aac	aac	t ac	ct c	t t c	t gc	720
I l e	Ser	Val	Ser	G y	Val	Al a	Asp	Asp	Pro	Asn	Asn	Tyr	Leu	Phe	Cys	
225					230					235					240	
cgc	gt c	cgc	aag	aac	ggg	gt g	gct	gcc	cct	agc	gcc	acc	agc	caa	ct c	768
Arg	Val	Arg	Lys	Asn	G y	Val	Al a	Al a	Pro	Ser	Al a	Thr	Ser	G n	Leu	
				245					250					255		
agc	acg	agg	gcc	t t g	gaa	ggt	att	t t c	gag	gcc	acc	cac	cgc	ct g	at c	816
Ser	Thr	Arg	Al a	Leu	G u	G y	I l e	Phe	G u	Al a	Thr	His	Arg	Leu	I l e	
			260					265					270			
t ac	ggc	gcg	aag	gat	gac	agc	ggt	caa	cgc	t ac	ct c	gca	t gg	t cc	ggg	864
Tyr	G y	Al a	Lys	Asp	Asp	Ser	G y	G n	Arg	Tyr	Leu	Al a	Trp	Ser	G y	
		275					280					285				
cac	t cc	gcc	cgc	gt t	gga	gct	gct	agg	gac	at g	gcc	cgc	gcc	ggt	gt t	912
His	Ser	Al a	Arg	Val	G y	Al a	Al a	Arg	Asp	Met	Al a	Arg	Al a	G y	Val	
	290					295					300					
t cc	at c	ccc	gaa	at c	at g	cag	gcg	ggt	gga	t gg	acg	aac	gt g	aac	att	960
Ser	I l e	Pro	G u	I l e	Met	G n	Al a	G y	G y	Trp	Thr	Asn	Val	Asn	I l e	
305					310					315					320	
gt c	at g	aac	t ac	att	cgc	aac	ct t	gac	agc	gag	acg	ggc	gca	at g	gt t	1008
Val	Met	Asn	Tyr	I l e	Arg	Asn	Leu	Asp	Ser	G u	Thr	G y	Al a	Met	Val	
				325					330					335		
cgc	ct c	ct g	gaa	gat	ggt	gac	t ga									1032
Arg	Leu	Leu	G u	Asp	G y	Asp										
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<220>
 <223> mai ze- opt i mi zed Cre

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 1 5 10 15
 Asp Al a Thr Ser Asp G u Val Arg Lys Asn Leu Met Asp Met Phe Arg
 20 25 30
 Asp Arg Gln Al a Phe Ser G u His Thr Trp Lys Met Leu Leu Ser Val
 35 40 45

5312WOPCT_SEQ LISTING.TXT

Cys	Arg	Ser	Trp	Ala	Ala	Trp	Cys	Lys	Leu	Asn	Asn	Arg	Lys	Trp	Phe
50						55					60				
Pro	Ala	Glu	Pro	Glu	Asp	Val	Arg	Asp	Tyr	Leu	Leu	Tyr	Leu	Gln	Ala
65					70					75					80
Arg	Gly	Leu	Ala	Val	Lys	Thr	Ile	Gln	Gln	His	Leu	Gly	Gln	Leu	Asn
				85					90					95	
Met	Leu	His	Arg	Arg	Ser	Gly	Leu	Pro	Arg	Pro	Ser	Asp	Ser	Asn	Ala
			100					105					110		
Val	Ser	Leu	Val	Met	Arg	Arg	Ile	Arg	Lys	Glu	Asn	Val	Asp	Ala	Gly
		115					120					125			
Glu	Arg	Ala	Lys	Gln	Ala	Leu	Ala	Phe	Glu	Arg	Thr	Asp	Phe	Asp	Gln
		130				135					140				
Val	Arg	Ser	Leu	Met	Glu	Asn	Ser	Asp	Arg	Cys	Gln	Asp	Ile	Arg	Asn
145					150					155					160
Leu	Ala	Phe	Leu	Gly	Ile	Ala	Tyr	Asn	Thr	Leu	Leu	Arg	Ile	Ala	Glu
			165						170					175	
Ile	Ala	Arg	Ile	Arg	Val	Lys	Asp	Ile	Ser	Arg	Thr	Asp	Gly	Gly	Arg
		180						185					190		
Met	Leu	Ile	His	Ile	Gly	Arg	Thr	Lys	Thr	Leu	Val	Ser	Thr	Ala	Gly
		195					200					205			
Val	Glu	Lys	Ala	Leu	Ser	Leu	Gly	Val	Thr	Lys	Leu	Val	Glu	Arg	Trp
		210				215					220				
Ile	Ser	Val	Ser	Gly	Val	Ala	Asp	Asp	Pro	Asn	Asn	Tyr	Leu	Phe	Cys
225					230					235					240
Arg	Val	Arg	Lys	Asn	Gly	Val	Ala	Ala	Pro	Ser	Ala	Thr	Ser	Gln	Leu
			245						250					255	
Ser	Thr	Arg	Ala	Leu	Glu	Gly	Ile	Phe	Glu	Ala	Thr	His	Arg	Leu	Ile
			260					265					270		
Tyr	Gly	Ala	Lys	Asp	Asp	Ser	Gly	Gln	Arg	Tyr	Leu	Ala	Trp	Ser	Gly
		275					280					285			
His	Ser	Ala	Arg	Val	Gly	Ala	Ala	Arg	Asp	Met	Ala	Arg	Ala	Gly	Val
		290				295					300				
Ser	Ile	Pro	Glu	Ile	Met	Gln	Ala	Gly	Gly	Trp	Thr	Asn	Val	Asn	Ile
305					310					315					320
Val	Met	Asn	Tyr	Ile	Arg	Asn	Leu	Asp	Ser	Glu	Thr	Gly	Ala	Met	Val
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Arg	Leu	Leu	Glu	Asp	Gly	Asp									
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<220>
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<400> 38
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<210> 39
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 <212> DNA
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 <220>
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 <220>
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 <211> 146
 <212> PRT
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 <220>
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 <220>
 <223> 13_6D10 Synthetic protein sequence

 <400> 43

5312WOPCT_SEQ LISTING.TXT

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Thr 31 Asp 32 Ser 33 Leu 34 Gly 35 Gly 36 Thr 37 Phe 38 His 39 Leu 40 Gly 41 Gly 42 Tyr 43 Tyr 44 Arg 45 Gly
Lys 46 Leu 47 Ile 48 Ser 49 Ile 50 Ala 51 Ser 52 Phe 53 Asn 54 Gln 55 Ala 56 Glu 57 His 58 Pro 59 Glu 60 Leu
Glu 61 Gly 62 Gln 63 Lys 64 Gln 65 Tyr 66 Gln 67 Leu 68 Arg 69 Gly 70 Met 71 Ala 72 Thr 73 Leu 74 Glu 75 Gly
Tyr 76 Arg 77 Glu 78 Gln 79 Lys 80 Ala 81 Gly 82 Ser 83 Thr 84 Leu 85 Ile 86 Arg 87 His 88 Ala 89 Glu 90 Glu
Leu 91 Leu 92 Arg 93 Lys 94 Lys 95 Gly 96 Ala 97 Asp 98 Leu 99 Leu 100 Trp 101 Cys 102 Asn 103 Ala 104 Arg 105 Thr
Ser 106 Ala 107 Ser 108 Gly 109 Tyr 110 Tyr 111 Lys 112 Lys 113 Leu 114 Gly 115 Phe 116 Ser 117 Glu 118 Gln 119 Gly 120 Glu
Val 121 Tyr 122 Asp 123 Thr 124 Pro 125 Pro 126 Val 127 Gly 128 Pro 129 His 130 Ile 131 Leu 132 Met 133 Tyr 134 Lys 135 Lys
Leu 136 Thr
145

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<210> 44
 <211> 146
 <212> PRT
 <213> Artificial Sequence

<220>
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<220>
 <223> 10_4H4 Synthetic protein sequence

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His 16 Lys 17 Ile 18 Leu 19 Arg 20 Pro 21 Asn 22 Gln 23 Pro 24 Leu 25 Glu 26 Val 27 Cys 28 Met 29 Tyr 30 Glu
Thr 31 Asp 32 Leu 33 Leu 34 Arg 35 Gly 36 Ala 37 Phe 38 His 39 Leu 40 Gly 41 Gly 42 Phe 43 Tyr 44 Arg 45 Gly
Lys 46 Leu 47 Ile 48 Ser 49 Ile 50 Ala 51 Ser 52 Phe 53 His 54 Gln 55 Ala 56 Glu 57 His 58 Ser 59 Glu 60 Leu
Gln 61 Gly 62 Gln 63 Lys 64 Gln 65 Tyr 66 Gln 67 Leu 68 Arg 69 Gly 70 Met 71 Ala 72 Thr 73 Leu 74 Glu 75 Gly
Tyr 76 Arg 77 Glu 78 Gln 79 Lys 80 Ala 81 Gly 82 Ser 83 Ser 84 Leu 85 Ile 86 Lys 87 His 88 Ala 89 Glu 90 Glu
Ile 91 Leu 92 Arg 93 Lys 94 Arg 95 Gly 96 Ala 97 Asp 98 Leu 99 Leu 100 Trp 101 Cys 102 Asn 103 Ala 104 Arg 105 Thr
Ser 106 Ala 107 Ser 108 Gly 109 Tyr 110 Tyr 111 Lys 112 Lys 113 Leu 114 Gly 115 Phe 116 Ser 117 Glu 118 Gln 119 Gly 120 Glu
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Ile 136 Thr
145

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<210> 45
 <211> 146
 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Synthesized

<220>
 <223> 0_5D3 Synthetic protein sequence

<400> 45

5312WOPCT_SEQ LISTING.TXT

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Met  Leu  Glu  Val  Lys  Pro  Ile  Asn  Ala  Glu  Asp  Thr  Tyr  Glu  Leu  Arg
 1      5      10      15
His  Arg  Ile  Leu  Arg  Pro  Asn  Gln  Pro  Ile  Glu  Ala  Cys  Met  Tyr  Glu
      20      25      30
Ser  Asp  Leu  Leu  Arg  Gly  Ala  Phe  His  Leu  Gly  Gly  Tyr  Tyr  Arg  Gly
      35      40      45
Lys  Leu  Ile  Ser  Ile  Ala  Ser  Phe  His  Gln  Ala  Glu  His  Ser  Glu  Leu
      50      55      60
Gln  Gly  Gln  Lys  Gln  Tyr  Gln  Leu  Arg  Gly  Met  Ala  Thr  Leu  Glu  Gly
65      70      75      80
Tyr  Arg  Glu  Gln  Lys  Ala  Gly  Ser  Ser  Leu  Ile  Lys  His  Ala  Glu  Glu
      85      90      95
Ile  Leu  Arg  Lys  Arg  Gly  Ala  Asp  Leu  Leu  Trp  Cys  Asn  Ala  Arg  Thr
      100      105      110
Ser  Ala  Ser  Gly  Tyr  Tyr  Lys  Lys  Leu  Gly  Phe  Ser  Glu  Gln  Gly  Glu
      115      120      125
Ile  Phe  Glu  Thr  Pro  Pro  Val  Gly  Pro  His  Ile  Leu  Met  Tyr  Lys  Arg
      130      135      140
Ile  Thr
145

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<210> 46
 <211> 146
 <212> PRT
 <213> Artificial Sequence

<220>
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<220>
 <223> R12G2 Synthetic protein sequence

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<400> 46
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His  Arg  Val  Leu  Arg  Pro  Asn  Gln  Pro  Ile  Glu  Ala  Cys  Met  Phe  Glu
      20      25      30
Ser  Asp  Leu  Thr  Arg  Ser  Ala  Phe  His  Leu  Gly  Gly  Phe  Tyr  Gly  Gly
      35      40      45
Lys  Leu  Ile  Ser  Val  Ala  Ser  Phe  His  Gln  Ala  Glu  His  Thr  Glu  Leu
      50      55      60
Gln  Gly  Lys  Lys  Gln  Tyr  Gln  Leu  Arg  Gly  Val  Ala  Thr  Leu  Glu  Gly
65      70      75      80
Tyr  Arg  Glu  Gln  Lys  Ala  Gly  Ser  Ser  Leu  Val  Lys  His  Ala  Glu  Glu
      85      90      95
Ile  Leu  Arg  Lys  Arg  Gly  Ala  Asp  Met  Ile  Trp  Cys  Asn  Ala  Arg  Thr
      100      105      110
Ser  Ala  Ser  Gly  Tyr  Tyr  Arg  Lys  Leu  Gly  Phe  Ser  Glu  Gln  Gly  Glu
      115      120      125
Val  Phe  Asp  Thr  Pro  Pro  Val  Gly  Pro  His  Ile  Leu  Met  Tyr  Lys  Arg
      130      135      140
Ile  Thr
145

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<210> 47
 <211> 442
 <212> DNA
 <213> Artificial Sequence

<220>
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<220>
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<220>

5312WOPCT_SEQ_LI STI NG TXT

<221> CDS

<222> (2)...(442)

<400> 47

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cat aga ata ctg aga cca aac cag ccg ata gaa gcg tgt atg ttt gaa 97
His Arg Ile Leu Arg Pro Asn Gln Pro Ile Gu Ala Cys Met Phe Gu
          20           25           30

agc gat tta ctt cgt ggt gca ttt cac tta ggc ggc ttt tac agg ggc 145
Ser Asp Leu Leu Arg Gly Ala Phe His Leu Gly Gly Phe Tyr Arg Gly
          35           40           45

aaa ctg att tcc ata gct tca ttc cac cag gcc gag cac tcg gaa ctg 193
Lys Leu Ile Ser Ile Ala Ser Phe His Gln Ala Gu His Ser Gu Leu
          50           55           60

caa ggc cag aaa cag tac cag ctg cga ggt atg gct acc ttg gaa ggt 241
Gln Gly Gln Lys Gln Tyr Gln Leu Arg Gly Met Ala Thr Leu Gu Gly
          65           70           75

tat cgt gag cag aaa gcg gga tca act cta gtt aaa cac gct gaa gaa 289
Tyr Arg Gu Gln Lys Ala Gly Ser Thr Leu Val Lys His Ala Gu Gu
          85           90           95

atc ctt cgt aag agg ggg gcg gac atg ctt tgg tgt aat gcg agg aca 337
Ile Leu Arg Lys Arg Gly Ala Asp Met Leu Trp Cys Asn Ala Arg Thr
          100          105          110

tcc gcc tca ggc tac tac aaa aag tta ggc ttc agc gag cag gga gag 385
Ser Ala Ser Gly Tyr Tyr Lys Lys Leu Gly Phe Ser Gu Gln Gly Gu
          115          120          125

ata ttt gac acg ccg cca gta gga cct cac atc ctg atg tat aaa agg 433
Ile Phe Asp Thr Pro Pro Val Gly Pro His Ile Leu Met Tyr Lys Arg
          130          135          140

atc aca taa
Ile Thr
145

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<210> 48

<211> 146

<212> PRT

<213> Artificial Sequence

<220>

<223> Synthesized

<220>

<223> optimized GAT sequence (GAT4601)

<400> 48

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Met Ile Gu Val Lys Pro Ile Asn Ala Gu Asp Thr Tyr Gu Leu Arg
  1           5           10           15
His Arg Ile Leu Arg Pro Asn Gln Pro Ile Gu Ala Cys Met Phe Gu
          20           25           30
Ser Asp Leu Leu Arg Gly Ala Phe His Leu Gly Gly Phe Tyr Arg Gly
          35           40           45
Lys Leu Ile Ser Ile Ala Ser Phe His Gln Ala Gu His Ser Gu Leu
          50           55           60
Gln Gly Gln Lys Gln Tyr Gln Leu Arg Gly Met Ala Thr Leu Gu Gly
          65           70           75           80
Tyr Arg Gu Gln Lys Ala Gly Ser Thr Leu Val Lys His Ala Gu Gu

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5312WOPCT_SEQ_LI STI NG TXT

I l e L e u A r g L y s A r g G l y A l a A s p M e t L e u T r p C y s A s n A l a A r g T h r
 S e r A l a S e r G l y T y r T y r L y s L y s L e u G l y P h e S e r G l u G n G l y G l u
 I l e P h e A s p T h r P r o P r o V a l G l y P r o H i s I l e L e u M e t T y r L y s A r g
 I l e T h r
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<210> 49
 <211> 441
 <212> DNA
 <213> Artificial Sequence

<220>
 <223> Synthesized

<220>
 <223> optimized GAT sequence (GAT4602)

<220>
 <221> CDS
 <222> (1)...(441)

<400> 49
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 M e t I l e G l u V a l L y s P r o I l e A s n A l a G l u A s p T h r T y r G l u L e u A r g
 1 5 10 15
 c a t a g a a t a c t c a g a c c a a a c a g c c g a t a g a a g c g t g t a t g t t t g a a 96
 H i s A r g I l e L e u A r g P r o A s n G l n P r o I l e G l u A l a C y s M e t P h e G l u
 20 25 30
 a g c g a t t t a c t t c g t g g t g c a t t t c a c t t a g g c g g c t a t t a c g g g g g c 144
 S e r A s p L e u L e u A r g G l y A l a P h e H i s L e u G l y G l y T y r T y r G l y G l y
 35 40 45
 a a a c t g a t t t c c a t a g c t t c a t t c c a c c a g g c c g a g c a c t c a g a a c t c 192
 L y s L e u I l e S e r I l e A l a S e r P h e H i s G l n A l a G l u H i s S e r G l u L e u
 50 55 60
 c a a g g c c a g a a a c a g t a c c a g c t c c g a g g t a t g g c t a c c t t g g a a g g t 240
 G n G l y G n L y s G n T y r G n L e u A r g G l y M e t A l a T h r L e u G l u G l y
 65 70 75 80
 t a t c g t g a g c a g a a g g c g g a t c g a g t c t a a t t a a a c a c g c t g a a g a a 288
 T y r A r g G l u G n L y s A l a G l y S e r S e r L e u I l e L y s H i s A l a G l u G l u
 85 90 95
 a t t c t t c g t a a g a g g g g g c g a c t t g c t t t g g t g t a a t g c g c g g a c a 336
 I l e L e u A r g L y s A r g G l y A l a A s p L e u L e u T r p C y s A s n A l a A r g T h r
 100 105 110
 t c c g c c t c a g g c t a c t a c a a a a a g t t a g g c t t c a g c g a g c a g g g a g a g 384
 S e r A l a S e r G l y T y r T y r L y s L y s L e u G l y P h e S e r G l u G n G l y G l u
 115 120 125
 g t a t t c g a c a c g c c g c c a g t a g g a c c t c a c a t c c t g a t g t a t a a a a g g 432
 V a l P h e A s p T h r P r o P r o V a l G l y P r o H i s I l e L e u M e t T y r L y s A r g
 130 135 140
 a t c a c a t a a 441
 I l e T h r
 145

5312WOPCT_SEQ_LI STI NG. TXT

<210> 50
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 <212> PRT
 <213> Artificial Sequence

<220>
 <223> Synthesized

<220>
 <223> optimized GAT sequence (GAT4602)

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 His Arg Ile Leu Arg Pro Asn Gln Pro Ile Glu Ala Cys Met Phe Glu
 20 25 30
 Ser Asp Leu Leu Arg Gly Ala Phe His Leu Gly Gly Tyr Tyr Gly Gly
 35 40 45
 Lys Leu Ile Ser Ile Ala Ser Phe His Gln Ala Glu His Ser Glu Leu
 50 55 60
 Gln Gly Gln Lys Gln Tyr Gln Leu Arg Gly Met Ala Thr Leu Glu Gly
 65 70 75 80
 Tyr Arg Glu Gln Lys Ala Gly Ser Ser Leu Ile Lys His Ala Glu Glu
 85 90 95
 Ile Leu Arg Lys Arg Gly Ala Asp Leu Leu Trp Cys Asn Ala Arg Thr
 100 105 110
 Ser Ala Ser Gly Tyr Tyr Lys Lys Leu Gly Phe Ser Glu Gln Gly Glu
 115 120 125
 Val Phe Asp Thr Pro Pro Val Gly Pro His Ile Leu Met Tyr Lys Arg
 130 135 140
 Ile Thr
 145

<210> 51
 <211> 1968
 <212> DNA
 <213> Glycine max

<220>
 <221> misc_feature
 <222> (1)... (1968)
 <223> HRA sequence

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 ct caccaaac ccaaccacgc t ct caaaat c aaat gt t cca t ct ccaaacc cccacggcg 180
 gcgccct t ca ccaaggaagc gccgaccacg gagccct t cg t gt cacggt t cgcct ccggc 240
 gaacct cgca agggcgcgga cat cct t gt g gaggcgt gg agaggcagg cgt gacgacg 300
 gt gt t cgcg t acccgggcg t gcgt cgat g gagat ccacc aggcgt cac gcgt ccgcc 360
 gccat ccgca acgt gct ccc gccccacgag cagggcggcg t ct t cgccgc cgaaggct ac 420
 gcgcgt t cct ccggcct ccc cggcgt ct gc at t gccacct ccggccccgg cgccaccaac 480
 ct cgt gagcg gcc cgccga cgct t t aat g gacagcgt cc cagt cgt cgc cat caccggc 540
 cagg t cgccc gccggat gat cggcaccgac gcct t ccaag aaaccccgat cgt ggaggt g 600
 agcagat cca t cacgaagca caact acct c at cct cgacg t cgacgacat ccccgcgct c 660
 gt cgccgagg ct t t ct t cgt cgccacct cc ggccgccccg gt ccggt cct cat cgacat t 720
 cccaaagagc t t cagcagca act cgccgt g cct aat t ggg acgagcccg t aacct cccc 780
 ggt t acct cg ccagct gcc caggcccccc gccgaggccc aat t ggaaca cat t gt caga 840
 ct cat cat gg agggccaaaa gccgct t ct c t acgt cggcg gt ggcagt t t gaat t ccagt 900
 gct gaat t ga ggcgct t t gt t gaact cact ggt at t cccg t t gct agcac t t t aat ggg 960
 ct t ggaact t t t cct at t gg t gat gaat at t cct t caga t gct ggt at gcat ggt act 1020
 gt t t at gct a act at gct gt t gacaat agt gat t t gt t gc t t gcct t t gg ggt aaggt t t 1080
 gat gaccgt g t t act gggaa gct t gaggct t t t gct agt a gggct aagat t gt t cacat t 1140
 gat at t gat t ct gccgagat t gggaagaac aagcaggcgc acgt gt cgt t t gcgcggat 1200
 t t gaagt t gg cct t gaagg aat t aat at g at t t t ggagg agaaaggagt ggaggg t aag 1260
 t t t gat ct t g gagg t ggag agaagagat t aat gt gcaga aacacaagt t t ccat t ggg 1320
 t acaagacat t ccaggacgc gat t t ct ccg cagcat gct a t cgaggt t ct t gat gagi t g 1380

5312WOPCT_SEQ_LI STI NG.TXT

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act aat ggag at gct at t gt t agt act ggg gt t gggcagc at caaat gt g ggct ggcgag 1440
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t t t ggat t gc ct gcggt at t ggt gct gct gt t gct aacc ct ggggt gt t gt ggt t gac 1560
at t gat gggg at ggt agt t t cat cat gaac gt t caggagt t ggccact at aagagt ggag 1620
aat ct ccag t t aagat at t gt t gt t gaac aat cagcat t t ggt at ggt ggt t cagt t g 1680
gaggat aggt t ct acaagt c caat agagct cacacct at c t t ggagat cc gt ct agcgag 1740
agcgagat at t cccaaacat gct caagt t t gct gat gct t gt gggat acc ggcagcgcg 1800
gt gacgaaga aggaagagct t agagcggca at t cagagaa t gt t ggacac ccct ggcccc 1860
t acct t ct t g at gt cat t gt gccccat cag gagcat gt gt t gccgat gat t cccagt aat 1920
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<210> 52
 <211> 1917
 <212> DNA
 <213> Zea mays

<220>
 <221> mi sc_f eat ur e
 <222> (1) . . . (1917)
 <223> HRA sequence

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gt ct cgagca t ct t ct t t at cgct ggcggt act t cgt t ct t ct t t gt cac acggaccgt 180
ggaat gt t ga accct t t ggc gat cgt cag aaat ct ggat at at ct cact t t cat t ct ct 240
gggt t t ccca agt at gt gt g cgct ct gt t g gcct t at aga acct gt cct c caact gcacc 300
accat ccca ggt gct ggt t gt t t agcaca aagacct t ca ct gggaggt t ct caat t cgg 360
at cat agct a gct cct gaac gt t cat gaga aagct accat ct ccat cgat gt caacaaca 420
gt gacacct g ggt t t gccac agaagcacca gcagcagccg gcaaaccaaa t cccat agcc 480
ccaagaccag ct gaagacaa ccact gcct t ggccgct t gt aagt gt agt a ct gt gccgcc 540
cacat ct ggt gct gcccaac acct gt gccg at gat gccct at gat gccct t t cgt cagct cat ca 600
agaacct gaa t agcat at t g t ggct ggat c t cct cat t ag at gt t t t at a cccaaggggg 660
aat t cct ct t ct gct gat c caact cat cg t t ccat gagc caaagt caaa gct ct t ct t t 720
gat gt gct t c ct t caagaag agcat t cat g ccct gcaaag caagct t aac at ct gcacag 780
at ggacacat gt ggct gct t gt t ct t gccca at ct cagccg gat caat at c aacgt gcaca 840
at ct t agccc t gct t gcaaa agcct caat c t t cct gt ca cgcat cat c aaaccgcaca 900
ccaagt gcaa gcaacagat c ggcct t at cc act gcat aat t t gcat acac cgt cccat gc 960
at acct agca t ggcgagaga cagt ggggt cg t cgct gggga agt t gccgag gcccat aaga 1020
gt agt t gt ga ccgggat t cc agt cagct cc acaaagcgt c gcaact cct c accagat gct 1080
gcgcgagccac cgcccacat a aagaacaggg cgccgcgat t caccaacaag acgcagcacc 1140
t gct caagca act cagt cgc aggggggt t g ggaaggcgcg caat gt accc aggcagact c 1200
at ggggt t gt cccagacagg caccgccat c t gct gct gga t gt cct t ggg gat gt cgaca 1260
agcacccggcc ct ggt cgacc agaggaggcg aggaagaaag cct cct gcac gacgcggggg 1320
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at gggcgt ct cct ggaaggc gt cgggt gcca at cat gcgt c t cggcgagcg t cccgt gat g 1440
gcgaccat gg ggacggaat c gagcagcgcg t cggcgagcg cggagact ag gt t ggt ggcg 1500
ccggggccgg aggt ggcgat gcagacgccg acgcggcccg aggagcgcg gt agcggag 1560
gcggaaggg cct cccct t g ct cgt ggcg aagaggt ggt t ggcgat gac gggggagcgg 1620
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ccgcagcgct cgagggact c gacgaggat g t cagcaccct t gcggggct c ggt gggggcc 1740
cacggccgg gcggggt ggc cgggggagcc at cggcat gg cgggt gacgc cgt gagcac 1800
ct gat gggcg cggcgagggc gcggcggggt g gccaggaggt gcgcccggcg cct cgcct t g 1860
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<210> 53
 <211> 2139
 <212> DNA
 <213> Arabidopsis

<220>
 <221> mi sc_f eat ur e
 <222> (1) . . . (2139)
 <223> HRA sequence

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<400> 53
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5312WOPCT_SEQ_LI STI NG.TXT

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ccgggat cca t ggcgggcgcc aacaacaaca acaacaacat ct t ct t cgat ct cct t ct cc 180
accaaaccat ct cct t cct c ct ccaaat ca ccat t accaa t ct ccagat t ct ccct ccca 240
t t ct ccct aa accccaacaa at cat cct cc t cct cccgcc gccgcggt at caaat ccagc 300
t ct ccct cct ccat ct ccgc cgt gct caac acaaccacca at gt cacaac cact ccct ct 360
ccaaccaaac ct accaaacc cgaacat t c at ct cccgat t cgct ccaga t caaccccgcc 420
aaaggcgct g at at cct cgt cgaagct t t a gaacgt caag gcgt agaaac cgt at t cgct 480
t accct ggag gt gcat caat ggagat t cac caagcct t aa cccgct ct t c ct caat ccgt 540
aacgt cct t c ct cgt cacga acaaggaggt gt at t cgcag cagaaggat a cgct cgat cc 600
t caggt aaac caggt at ct g t at agccact t caggt cccg gagct acaaa t ct cgt t agc 660
ggat t agccg at gcgt t gt t agat agt gt t cct ct t gt ag caat cacagg acaagt cgct 720
cgt cgt at ga t t ggt acaga t gcgt t t caa gagact ccga t t gt t gaggt aacgcgt t cg 780
at t acgaagc at aact at ct t gt gat ggat gt t gaagat a t ccct aggat t at t gaggaa 840
gct t t ct t t t t agct act t c t ggt agacct ggacct gt t t t ggt t gat gt t cct aaagat 900
at t caacaac agct t gcgat t cct aat t gg gaacaggct a t gagat t acc t ggt t at at g 960
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gagt ct aaga agcct gt gt t gt at gt t ggt ggt ggt t gt t t gaat t ct ag cgat gaat t g 1080
ggg aggt t t g t t gagct t ac ggaggat ccct gt t gcgagt a cgt t gat ggg gct gggat ct 1140
t at cct t gt g at gat gagt t gt cgt t acat at gct t ggaa t gcat gggac t gt gt at gca 1200
aat t acgct g t ggagcat ag t gat t t gt t g t t ggcgt t t g ggg aaggt t t gat gat cgt 1260
gt cacgggt a agct t gaggc t t t t gct agt agggct aaga t t gt t cat at t gat at t gac 1320
t cggct gaga t t gggaagaa t aagact cct cat gt gt ct g t gt gt ggt ga t gt t aagct g 1380
gct t t gcaag ggat gaat at gat t ct t gag agccgagcgg aggagct t aa gct t gat t t t 1440
ggagt t t gga ggaat gagt t gaacgt acag aaacagaagt t t ccgt t gag ct t t aagacg 1500
t t t ggggaag ct at t cct cc acagt at gcg at t aaggt cc t t gat gagt t gact gat gga 1560
aaagccat aa t aagt act gg t gt cgggcaa cat caaat gt gggcggcgca gt t ct acaat 1620
t acaagaac caaggcagt g gct at cat ca ggaggcct t g gagct at ggg at t t ggact t 1680
cct gct gcga t t ggagcgt c t gt t gct aac cct gat gcga t agt t gt gga t at t gacgga 1740
gat ggaagct t t at aat gaa t gt gcaagag ct agccact a t t cgt gt aga gaat ct t cca 1800
gt gaaggf ac t t t t at t aaa caaccagcat ct t ggcat gg t t at gcaat t ggaagat cgg 1860
t t ct acaaag ct aaccgagc t cacacat t t ct cggggat c cggct cagga ggacgagat a 1920
t t cccgaaca t gt t gct gt t t gcagcagct t gcgggat t c cagcggcgag ggt gacaaag 1980
aaagcagat c t ccgagaagc t at t cagaca at gct ggt a caccaggacc t t acct gt t g 2040
gat gt gat t t gt ccgcacca agaacat gt g t t gccgat ga t cccgagt gg t ggcact t t c 2100
aacgat gt ca t aacggaagg agat ggccgg at t aaat ac 2139

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<210> 54
<211> 552
<212> DNA
<213> Artificial Sequence

<220>
<223> Synthesized

<220>
<223> maize optimized PAT sequence

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cagaccccgcc aggagt ggat cgacgacct g gaggcct cc aggaccgt a cccgt ggct c 180
gt ggccgagg t ggagggcggt ggt ggccggc at cgct acg cccggcccgt g gaaggcccgcc 240
aacgccf acg act ggaccgt ggagt ccacc gt gt acgt gt cccaccgcca ccagcgccf c 300
ggcct cggct ccacct ct a caccacct c ct caagagca t ggaggcca gggct t caag 360
t ccgt ggt gg ccgt gat cgg cct cccgaac gaccgct ccg t gcgct cca cgaggccct c 420
ggct acaccg cccgcggcac cct ccgcgcc gccggct aca agcacggcgg ct ggcacgac 480
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ct g Leu	ccg Pro	ccc Pro	t cc Ser 20	cag Gln	acg Thr	acg Thr	gac Asp	t cc Ser 25	acg Thr	ct c Leu	at c Ile	t cg Ser	gcc Ala 30	gcc Ala	acc Thr	96
gcc Ala	gac Asp	cat His 35	gt c Val	t cc Ser	ggc Gly	gat Asp	gt c Val 40	t gc Cys	t t c Phe	aac Asn	at c Ile	ccc Pro 45	caa Gln	gat Asp	t gg Trp	144
agc Ser	at g Met 50	agg Arg	gga Gly	t ca Ser	gag Glu	ct t Leu 55	t cg Ser	gcg Ala	ct c Leu	gt c Val	gcg Ala 60	gag Glu	ccg Pro	aag Lys	ct g Leu	192
gag Glu 65	gac Asp	t t c Phe	ct c Leu	ggc Gly	ggc Gly 70	at c Ile	t cc Ser	t t c Phe	t cc Ser	gag Glu 75	cag Gln	cat His	cac His	aag Lys	t cc Ser 80	240
aac Asn	t gc Cys	aac Asn	t t g Leu	at a Ile 85	ccc Pro	agc Ser	act Thr	agc Ser	agc Ser 90	aca Thr	gt t Val	t gc Cys	t ac Tyr	gcg Ala 95	agc Ser	288
t ca Ser	gct Ala	gct Ala	agc Ser 100	acc Thr	ggc Gly	t ac Tyr	cat His 105	cac His	cag Gln	ct g Leu	t ac Tyr	cag Gln	ccc Pro 110	acc Thr	agc Ser	336
t cc Ser	gcg Ala	ct c Leu 115	cac His	t t c Phe	gcg Ala	gac Asp	t cc Ser 120	gt c Val	at g Met	gt g Val	gcc Ala	t cc Ser 125	t cg Ser	gcc Ala	ggt Gly	384
gt c Val	cac His 130	gac Asp	ggc Gly	ggt Gly	t cc Ser	at g Met 135	ct c Leu	agc Ser	gcg Ala	gcc Ala	gcc Ala 140	gct Ala	aac Asn	ggt Gly	gt c Val	432
gct Ala 145	ggc Gly	gct Ala	gcc Ala	agt Ser	gcc Ala 150	aac Asn	ggc Gly	ggc Gly	ggc Gly	at c Ile 155	ggg Gly	ct g Leu	t cc Ser	at g Met	at c Ile 160	480
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gcg Ala	gct Ala	gag Glu	ggc Gly 180	gcg Ala	cag Gln	ggg Gly	ct c Leu	t ct Ser 185	t t g Leu	t cc Ser	at g Met	aac Asn	at g Met 190	gcg Ala	ggg Gly	576
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gac Asp	aac Asn	acg Thr	gca Ala 260	agg Arg	aag Lys	acg Thr	gt g Val	gac Asp 265	acg Thr	t t c Phe	ggg Gly	cag Gln	cgc Arg 270	acg Thr	t cg Ser	816

5312WOPCT_SEQ_LI STI NG TXT

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Phe	Ser	Arg	Gly	Ala	Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	His	Gln	
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gac	atc	gcg	gcg	atc	aag	ttc	cgc	ggc	ctc	aac	gcc	gtc	acc	aac	ttc	1296
Asp	Ile	Ala	Ala	Ile	Lys	Phe	Arg	Gly	Leu	Asn	Ala	Val	Thr	Asn	Phe	
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Asp	Met	Ser	Arg	Tyr	Asp	Val	Lys	Ser	Ile	Leu	Asp	Ser	Ser	Ala	Leu	
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ccc	atc	ggc	agc	gcc	gcc	aag	cgt	ctc	aag	gag	gcc	gag	gcc	gca	gcg	1392
Pro	Ile	Gly	Ser	Ala	Ala	Lys	Arg	Leu	Lys	Glu	Ala	Glu	Ala	Ala	Ala	
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Ile	Ala	Ser	Gln	Leu	Gly	Asp	Gly	Gly	Ala	Leu	Ala	Ala	Ala	Tyr	Gly	
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gcg	cac	tac	cac	ggc	gcc	gcc	tgg	ccg	acc	atc	gcg	ttc	cag	ccg	ggc	1536
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gcc	gcc	acc	aca	ggc	ctg	tac	cac	ccg	tac	gcg	cag	cag	cca	atg	cgc	1584
Ala	Ala	Thr	Thr	Gly	Leu	Tyr	His	Pro	Tyr	Ala	Gln	Gln	Pro	Met	Arg	
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ggc	ggc	ggg	tgg	tgc	aag	cag	gag	cag	gac	cac	gcg	gtg	atc	gcg	gcc	1632
Gly	Gly	Gly	Trp	Cys	Lys	Gln	Glu	Gln	Asp	His	Ala	Val	Ile	Ala	Ala	
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5312WOPCT_SEQ_LI STI NG TXT

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gcg cac gac ttt ttc t c g gca ggg cag cag gcc gcc gcc gca gct gcg Al a Hi s Asp Phe Phe Ser Al a Gl y Gl n Gl n Al a Al a Al a Al a Al a Al a 565 570 575	1728
at g cac ggc ct g gct agc at c gac agt gcg t c g ct c gag cac agc acc Met Hi s Gl y Leu Al a Ser Ile Asp Ser Al a Ser Leu Gl u Hi s Ser Thr 580 585 590	1776
ggc t cc aac t cc gt c gt c t ac aac ggc ggg gt c ggc gat agc aac ggc Gl y Ser Asn Ser Val Val Tyr Asn Gl y Gl y Val Gl y Asp Ser Asn Gl y 595 600 605	1824
gcc agc gcc gt t ggc agc ggc ggt ggc t ac at g at g ccg at g agc gct Al a Ser Al a Val Gl y Ser Gl y Gl y Tyr Met Met Pro Met Ser Al a 610 615 620	1872
gcc gga gca acc act aca t c g gca at g gt g agc cac gag cag at g cat Al a Gl y Al a Thr Thr Thr Ser Al a Met Val Ser Hi s Gl u Gl n Met Hi s 625 630 635 640	1920
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Gl u	Asp	Phe	Leu	Gl y	Gl y	Ile	Ser	Phe	Ser	Gl u	Gl n	Hi s	Hi s	Lys	Ser
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5312WOPCT_SEQ_LI STI NG TXT

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Thr	Thr	G n	G y	Al a	Al a	G y	Met	185	Pro	Leu	Leu	Al a	G y	190	G u	Arg	Al a	
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Val	Thr	Al a	Pro	Lys	G u	Asp	Ser	G y	G y	Ser	G y	Val	Al a	G y	Al a	240	Al a	
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Asp	Asn	Thr	Al a	Arg	Lys	Thr	Val	Asp	250	Thr	Phe	G y	G n	Arg	270	Thr	Ser	
I l e	Tyr	Arg	G y	Val	Thr	Arg	H i s	Arg	265	Tr p	Thr	G y	Arg	Tyr	G u	Al a		
H i s	Leu	Tr p	Asp	Asn	Ser	Cys	Arg	Arg	280	G u	G y	G n	Thr	Arg	Lys	G y		
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Asn	Phe	Pro	Val	Ser	Asn	Tyr	G u	Lys	330	G u	Leu	G u	Asp	Met	Lys	H i s		
Met	Thr	Arg	G n	G u	Phe	Val	Al a	Ser	345	Leu	Arg	Arg	Lys	Ser	Ser	G y		
Phe	Ser	Arg	G y	Al a	Ser	I l e	Tyr	Arg	360	G y	Val	Thr	Arg	H i s	H i s	G n		
H i s	G y	Arg	Tr p	G n	Al a	Arg	I l e	G y	375	Arg	Val	Al a	G y	Asn	Lys	Asp	400	
Leu	Tyr	Leu	G y	Thr	Phe	Ser	Thr	G n	390	G u	G u	Al a	Al a	G u	Al a	Tyr		
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I l e	Al a	Ser	G n	Leu	G y	Asp	G y	G y	470	Al a	Leu	Al a	Al a	Al a	Tyr	G y		
Al a	H i s	Tyr	H i s	G y	Al a	Al a	Tr p	Pro	485	Thr	I l e	Al a	Phe	G n	Pro	G y		
Al a	Al a	Thr	G y	Leu	Tyr	H i s	Pro	Tyr	500	Al a	G n	G n	Pro	Met	Arg			
G y	G y	G y	Tr p	Cys	Lys	G n	G u	G n	515	Asp	H i s	Al a	Val	I l e	Al a	Al a		
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545	Al a	H i s	Asp	Phe	Phe	Ser	Al a	G y	535	G n	G n	Al a	Al a	Al a	Al a	Al a		
Met	H i s	G y	Leu	Al a	Ser	I l e	Asp	Ser	550	Al a	Ser	Leu	G u	H i s	Ser	Thr		
G y	Ser	Asn	Ser	Val	Val	Tyr	Asn	G y	565	G y	G y	Val	G y	Asp	Ser	Asn	G y	
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Al a	G y	Al a	Thr	Thr	Thr	Ser	Al a	Met	595	Val	Ser	H i s	G u	G n	Met	H i s		
625	Al a	Arg	Al a	Tyr	Asp	G u	Al a	Lys	610	G n	Al a	G n	Met	G y	Tyr	G u		
Ser	Tyr	Leu	Val	Asn	Al a	G u	Asn	Asn	630	G y	G y	G y	Arg	Met	Ser	Al a		
Tr p	G y	Thr	Val	Val	Ser	Al a	Al a	Al a	645	Al a	Al a	Al a	Al a	Ser	Ser	Asn		

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 Val Trp Asn Asp Thr
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5312WOPCT_SEQ_LI STI NG TXT

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cac Hi s	at g Mét	aca Thr	1104
ggt G y 370	ttc Phe	tcc Ser	1152
caa G n 385	cat Hi s	gga G y	1200
gat Asp	ctt Leu	tac Tyr	1248
tac Tyr	gac Asp	atc Ile	1296
ttc Phe	gac Asp	atg Mét	1344
ctc Leu	ccc Pro	atc Ile	1392
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ggc G y	gcc Al a	agg Al a	1584
cgc Arg	ggc G y	ggg G y	1632
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5312WOPCT_SEQ_LI STI NG TXT

565

570

575

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 Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu Gln His His Lys Ala
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 Ser Gly Ala Ser Thr Gly Tyr His His Gln Leu Tyr His Gln Pro Thr
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 Ser Ser Ala Leu His Phe Ala Asp Ser Val Met Val Ala Ser Ser Ala
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 Gly Val His Asp Gly Gly Ala Met Leu Ser Ala Ala Ala Ala Asn Gly
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 Val Ala Gly Ala Ala Ser Ala Asn Gly Gly Ile Gly Leu Ser Met
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 Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Met Gln Pro Arg Val
 165 170 175
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<210> 60
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<220>
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<220>
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<222> 3
<223> Xaa=Leu or Val

<220>
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<223> Xaa=Glu or Ala

<220>
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<210> 62
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<220>
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<220>
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<220>
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 <223> Xaa=Gln or Glu

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Phe	Ser	Thr	Xaa	G u	G u	Al a	Al a	G u	Al a	Tyr	Asp	Xaa	Al a	Al a
		35					40					45		
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<222> 26

<223> Xaa=Arg or Lys

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<222> 33

<223> Xaa=Val or G l y

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<223> Xaa=Tyr or Arg

<220>

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<220>

<221> VARI ANT

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<223> Xaa=G u or Asp

<220>

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<222> (58) . . . (58)

<223> Xaa=Pr o or Thr

<220>

<221> VARI ANT

<222> (61) . . . (61)

<223> Xaa=Thr or His

<220>

<221> VARI ANT

<222> (62) . . . (62)

<223> Xaa=Thr or Ile

<220>

<221> VARI ANT

<222> (66) . . . (66)

<223> Xaa=Ile, Val , or Leu

<400> 65

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			20					25					30		
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<210> 66

<211> 31

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<222> 10

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<220>

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<222> 16

<223> Xaa=Phe or Tyr

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<221> VARI ANT

<222> 17

<223> Xaa=Val or Ile

<220>

<221> VARI ANT

<222> 19

<223> Xaa=Ser or His

<400> 66

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<210> 67

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<223> Xaa=Thr or Asn

<400> 67

Xaa Leu Ser Met Ile Lys Xaa Trp Leu Arg
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<210> 68

<211> 7

<212> PRT

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<223> Synthesized

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<223> BBM consensus sequence motif 10

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<223> Xaa=Gln or Pro

<400> 68

Trp Cys Lys Xaa Glu Gln Asp
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<210> 69

<211> 8

<212> PRT

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<223> Xaa=any amino acid

<400> 69

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<210> 70

<211> 5

<212> PRT

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<400> 70

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<210> 71

<211> 7

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<220>

<223> BBM consensus sequence motif 14

<400> 71

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<210> 72

<211> 11

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<223> Xaa=Ser or Thr

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<210> 73

<211> 7

<212> PRT

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5312WOPCT_SEQ_LI STI NG TXT

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<222> (1)...(1755)

<400> 74

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G n Asn His His 20 Arg Thr Asp Val Asp 25 Ser Ser Thr Thr Arg 30 Thr Ala	
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Val Asp Val Ala Gly Gly Tyr Cys 40 Phe Asp Leu Ala Ala 45 Pro Ser Asp	
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G u Ser 50 Ser Ala Val G n Thr 55 Ser Phe Leu Ser Pro 60 Phe Gly Val Thr	
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Leu G u Ala Phe Thr Arg 70 Asp Asn Asn Ser His 75 Ser Arg Asp Trp Asp 80	
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I le Asn Gly Gly Ala Cys Asn Thr Leu Thr 90 Asn Asn G u G n Asn 95 Gly	
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Pro Lys Leu G u Asn Phe Leu Gly Arg 105 Thr Thr Thr Ile Tyr Asn Thr	
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Asn G u Thr 115 Val Val Asp Gly Asn 120 Gly Asp Cys Gly Gly Asp Gly	
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G y Gly Gly Gly Ser Leu Gly Leu Ser Met Ile Lys 140 Thr Trp Leu Ser	
aat cat tgc gtt gct aat gct aat cat caa gac aat ggt aac ggt gca	480
Asn His Ser Val Ala Asn Ala Asn His G n Asp 155 Asn Gly Asn Gly Ala 160	
cga ggc ttg tcc ctg tct atg aat tca tct act agt gat agc aac aac	528
Arg Gly Leu Ser Leu Ser Met Asn Ser Ser Thr Ser Asp Ser Asn Asn	
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act Thr	aac Asn	t tc Phe 275	ccc Pro	t tg Leu	agt Ser	gaa Glu	t at Tyr 280	gag Glu	aaa Lys	gag Glu	gt a Val	gaa Glu 285	gag Glu	at g Met	aag Lys	864		
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gac Asp	ct c Leu	t ac Tyr	t tg Leu 340	gga Gly	act Thr	t tc Phe	ggc Gly	aca Thr 345	cag Gln	gaa Glu	gag Glu	gct Ala	gct Ala 350	gag Glu	gct Ala	1056		
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							caa Gln 505
							gga Gly
							t tc Phe
							gca Ala
							at c Ile
							cct Pro 510
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gca Ala	aga Arg 530	aat Asn	cat His	t at Tyr	t ac Tyr	t at Tyr 535	gct Ala
							cag Gln
							cat His
							cag Gln
							caa Gln 540
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							cag Gln
							at t Ile
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cag Gln 545	cag Gln	t cg Ser	ccg Pro	gga Gly	gga Gly 550	gat Asp	t tt Phe
							ccg Pro
							gt g Val
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							t cg Ser
							aat Asn
							aac Asn
							cat His 560
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Val	Glu	Thr 195	Thr	Pro	Lys	Lys	Thr 200	Ile	Glu	Ser	Phe	Gly 205	Gln	Arg	Thr
Ser	Ile 210	Tyr	Arg	Gly	Val	Thr 215	Arg	His	Arg	Trp	Thr 220	Gly	Arg	Tyr	Glu
Ala 225	His	Leu	Trp	Asp	Asn 230	Ser	Cys	Lys	Arg	Glu 235	Gly	Gln	Thr	Arg	Lys 240
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Glu 465	Glu	Gln	Gln	His	Phe 470	Leu	Arg	Asn	Ser	Pro 475	Ser	His	Met	Thr	Asn 480
Val	Asp	His	His	Ser 485	Ser	Thr	Ser	Asp	Asp 490	Ser	Val	Thr	Val	Cys 495	Gly
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145					150					155						
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G y	Leu	Ser	Leu	Ser	Met	Asn	Ser	Ser	Thr	Ser	Cys	Asp	Asn	Asn	Asn	175
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Asp	Ser	Asn	Asn	Asn	Val	Val	Al a	G n	G y	Lys	Thr	I l e	Asp	Asp	Ser	190
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Al a	His	Leu	Trp	Asp	Asn	Ser	Cys	Lys	Arg	G u	G y	G n	Thr	Arg	Lys	240
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Leu	Pro	I l e	G l y	Ser	A l a	A l a	Lys	Arg	Leu	Lys	G u	A l a	Asn	Arg	Pro	
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Asn	H i s	Tyr	Tyr	Phe	A l a	G n	G n	G n	G n	Thr	G n	G n	Ser	Pro	G y	
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gga	gat	t t t	ccc	g c g	gca	a t g	acg	aat	aat	g t t	g g c	t c t	aat	a t g	t a t	1680
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			20					25					30		
Val	Ala	Gly	Glu	Tyr	Cys	Tyr	Asp	Pro	Thr	Ala	Ala	Ser	Asp	Glu	Ser
		35					40					45			
Ser	Ala	Ile	Gln	Thr	Ser	Phe	Pro	Ser	Pro	Phe	Gly	Val	Val	Val	Asp
	50					55					60				
Ala	Phe	Thr	Arg	Asp	Asn	Asn	Ser	His	Ser	Arg	Asp	Trp	Asp	Ile	Asn
65					70					75					80
Gly	Cys	Ala	Cys	Asn	Asn	Ile	His	Asn	Asp	Glu	Gln	Asp	Gly	Pro	Lys
			85						90					95	
Leu	Glu	Asn	Phe	Leu	Gly	Arg	Thr	Thr	Thr	Ile	Tyr	Asn	Thr	Asn	Glu
			100					105					110		
Asn	Val	Gly	Asp	Gly	Ser	Gly	Ser	Gly	Cys	Tyr	Gly	Gly	Gly	Asp	Gly
		115					120					125			
Gly	Gly	Gly	Ser	Leu	Gly	Leu	Ser	Met	Ile	Lys	Thr	Trp	Leu	Arg	Asn
	130					135					140				
Gln	Pro	Val	Asp	Asn	Val	Asp	Asn	Gln	Glu	Asn	Gly	Asn	Ala	Ala	Lys
145					150					155					160
Gly	Leu	Ser	Leu	Ser	Met	Asn	Ser	Ser	Thr	Ser	Cys	Asp	Asn	Asn	Asn
				165					170					175	
Asp	Ser	Asn	Asn	Asn	Val	Val	Ala	Gln	Gly	Lys	Thr	Ile	Asp	Asp	Ser
			180					185					190		
Val	Glu	Ala	Thr	Pro	Lys	Lys	Thr	Ile	Glu	Ser	Phe	Gly	Gln	Arg	Thr
		195					200					205			
Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	Arg	Trp	Thr	Gly	Arg	Tyr	Glu
	210					215					220				
Ala	His	Leu	Trp	Asp	Asn	Ser	Cys	Lys	Arg	Glu	Gly	Gln	Thr	Arg	Lys
225					230					235					240
Gly	Arg	Gln	Val	Tyr	Leu	Gly	Gly	Tyr	Asp	Lys	Glu	Glu	Lys	Ala	Ala
			245						250					255	
Arg	Ala	Tyr	Asp	Leu	Ala	Ala	Leu	Lys	Tyr	Trp	Gly	Thr	Thr	Thr	Thr
			260					265					270		
Thr	Asn	Phe	Pro	Met	Ser	Glu	Tyr	Glu	Lys	Glu	Val	Glu	Glu	Met	Lys
		275					280					285			
His	Met	Thr	Arg	Gln	Glu	Tyr	Val	Ala	Ser	Leu	Arg	Arg	Lys	Ser	Ser
	290					295					300				
Gly	Phe	Ser	Arg	Gly	Ala	Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	His
305					310					315					320
Gln	His	Gly	Arg	Trp	Gln	Ala	Arg	Ile	Gly	Arg	Val	Ala	Gly	Asn	Lys
			325						330					335	
Asp	Leu	Tyr	Leu	Gly	Thr	Phe	Gly	Thr	Gln	Glu	Glu	Ala	Ala	Glu	Ala
			340					345					350		
Tyr	Asp	Ile	Ala	Ala	Ile	Lys	Phe	Arg	Gly	Leu	Thr	Ala	Val	Thr	Asn
		355					360					365			
Phe	Asp	Met	Asn	Arg	Tyr	Asn	Val	Lys	Ala	Ile	Leu	Glu	Ser	Pro	Ser
	370					375					380				
Leu	Pro	Ile	Gly	Ser	Ala	Ala	Lys	Arg	Leu	Lys	Glu	Ala	Asn	Arg	Pro
385					390					395					400
Val	Pro	Ser	Met	Met	Met	Ile	Ser	Asn	Asn	Val	Ser	Glu	Ser	Glu	Asn
			405						410					415	
Ser	Ala	Ser	Gly	Trp	Gln	Asn	Ala	Ala	Val	Gln	His	His	Gln	Gly	Val
			420					425					430		
Asp	Leu	Ser	Leu	Leu	His	Gln	His	Gln	Glu	Arg	Tyr	Asn	Gly	Tyr	Tyr
		435					440					445			
Tyr	Asn	Gly	Gly	Asn	Leu	Ser	Ser	Glu	Ser	Ala	Arg	Ala	Cys	Phe	Lys
	450					455					460				
Gln	Glu	Asp	Asp	Gln	His	His	Phe	Leu	Ser	Asn	Thr	Gln	Ser	Leu	Met
465					470					475					480
Thr	Asn	Ile	Asp	His	Gln	Ser	Ser	Val	Ser	Asp	Asp	Ser	Val	Thr	Val
			485						490					495	

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Cys	Gly	Asn	Val	Val	Gly	Tyr	Gly	Gly	Tyr	Gln	Gly	Phe	Ala	Ala	Pro
			500					505					510		
Val	Asn	Cys	Asp	Ala	Tyr	Ala	Ala	Ser	Glu	Phe	Asp	Tyr	Asn	Ala	Arg
		515					520					525			
Asn	His	Tyr	Tyr	Phe	Ala	Gln	Gln	Gln	Thr	Gln	Gln	Ser	Pro	Gly	
	530					535					540				
Gly	Asp	Phe	Pro	Ala	Ala	Met	Thr	Asn	Asn	Val	Gly	Ser	Asn	Met	Tyr
545					550					555					560
Tyr	His	Gly	Glu	Gly	Gly	Gly	Glu	Val	Ala	Pro	Thr	Phe	Thr	Val	Trp
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Asn	Asp	Asn													

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 <212> DNA
 <213> Brassica napus

<220>
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cac cat cgt aag gac gtc tgc tct tcc acc acc aca acc gcc gta gat	96
His His Arg Lys Asp Val Cys Ser Ser Thr Thr Thr Thr Ala Val Asp	
20 25 30	
gtc gcc gga gag tac tgt tac gat ccg acc gct gcc tcc gat gag tct	144
Val Ala Gly Glu Tyr Cys Tyr Asp Pro Thr Ala Ala Ser Asp Glu Ser	
35 40 45	
tca gcc atc caa aca tgc ttt cct tct ccc ttt ggt gtc gtc ctg gat	192
Ser Ala Ile Gln Thr Ser Phe Pro Ser Pro Phe Gly Val Val Leu Asp	
50 55 60	
gct ttc acc aga gac aac aat agt cac tcc cga gat tgg gac atc aat	240
Ala Phe Thr Arg Asp Asn Asn Ser His Ser Arg Asp Trp Asp Ile Asn	
65 70 75 80	
ggt agt gca tgt aat aac atc cac aat gat gag caa gat gga cca aaa	288
Gly Ser Ala Cys Asn Asn Ile His Asn Asp Glu Gln Asp Gly Pro Lys	
85 90 95	
ctt gag aat ttc ctt ggc cgc acc acc acg att tac aac acc aac gaa	336
Leu Glu Asn Phe Leu Gly Arg Thr Thr Thr Ile Tyr Asn Thr Asn Glu	
100 105 110	
aac gtt gga gat atc gat gga agt ggg tgt tat gga gga gga gac ggt	384
Asn Val Gly Asp Ile Asp Gly Ser Gly Cys Tyr Gly Gly Gly Asp Gly	
115 120 125	
ggt ggt ggc tca cta gga ctt tgc atg ata aag aca tgg ctg aga aat	432
Gly Gly Gly Ser Leu Gly Leu Ser Met Ile Lys Thr Trp Leu Arg Asn	
130 135 140	
caa ccc gtg gat aat gtt gat aat caa gaa aat ggc aat ggt gca aaa	480
Gln Pro Val Asp Asn Val Asp Asn Gln Glu Asn Gly Asn Gly Ala Lys	
145 150 155 160	
ggc ctg tcc ctg tca atg aac tca tct act tct tgt gat aac aac aac	528
Gly Leu Ser Leu Ser Met Asn Ser Ser Thr Ser Cys Asp Asn Asn Asn	
165 170 175	

5312WOPCT_SEQ_LI STING.TXT

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			180					185					190			
gt t	gaa	gct	aca	ccg	aag	aaa	act	at t	gag	agt	ttt	gga	cag	agg	acg	624
Val	Glu	Ala	Thr	Pro	Lys	Lys	Thr	Ile	Glu	Ser	Phe	Gly	Gln	Arg	Thr	
		195					200					205				
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Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	Arg	Trp	Thr	Gly	Arg	Tyr	Glu	
	210					215					220					
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Ala	His	Leu	Trp	Asp	Asn	Ser	Cys	Lys	Arg	Glu	Gly	Gln	Thr	Arg	Lys	
225					230					235					240	
gga	aga	caa	gtt	tat	t t g	gga	ggc	t at	gac	aaa	gaa	gaa	aaa	gca	gct	768
Gly	Arg	Gln	Val	Tyr	Leu	Gly	Gly	Tyr	Asp	Lys	Glu	Glu	Lys	Ala	Ala	
			245						250					255		
agg	gct	t at	gat	tta	gcc	gca	ctc	aag	t at	t gg	gga	acc	acc	act	act	816
Arg	Ala	Tyr	Asp	Leu	Ala	Ala	Leu	Lys	Tyr	Trp	Gly	Thr	Thr	Thr	Thr	
			260					265					270			
act	aac	t t c	ccc	at g	agc	gaa	t at	gag	aaa	gag	at a	gaa	gag	at g	aag	864
Thr	Asn	Phe	Pro	Met	Ser	Glu	Tyr	Glu	Lys	Glu	Ile	Glu	Glu	Met	Lys	
		275					280					285				
cac	at g	aca	agg	caa	gag	t at	gt t	gcc	tca	ctt	cgc	agg	aaa	agt	agt	912
His	Met	Thr	Arg	Gln	Glu	Tyr	Val	Ala	Ser	Leu	Arg	Arg	Lys	Ser	Ser	
	290					295					300					
ggc	t t c	t ct	cgt	ggc	gca	t cg	att	t at	cgt	gga	gt a	aca	aga	cat	cac	960
Gly	Phe	Ser	Arg	Gly	Ala	Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	His	
305					310					315					320	
caa	cat	gga	aga	tgg	caa	gct	agg	at a	gga	aga	gt c	gcc	ggc	aac	aaa	1008
Gln	His	Gly	Arg	Trp	Gln	Ala	Arg	Ile	Gly	Arg	Val	Ala	Gly	Asn	Lys	
				325					330					335		
gac	ctc	t ac	t t g	gga	act	t t t	ggc	aca	caa	gaa	gaa	gct	gca	gag	gca	1056
Asp	Leu	Tyr	Leu	Gly	Thr	Phe	Gly	Thr	Gln	Glu	Glu	Ala	Ala	Glu	Ala	
			340					345					350			
t ac	gac	att	gcg	gcc	atc	aaa	t t c	aga	gga	tta	acc	gca	gtg	act	aac	1104
Tyr	Asp	Ile	Ala	Ala	Ile	Lys	Phe	Arg	Gly	Leu	Thr	Ala	Val	Thr	Asn	
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t t c	gac	at g	aac	aga	t ac	aac	gt t	aaa	gca	atc	ctc	gaa	agc	cct	agt	1152
Phe	Asp	Met	Asn	Arg	Tyr	Asn	Val	Lys	Ala	Ile	Leu	Glu	Ser	Pro	Ser	
	370					375					380					
ctt	cct	att	ggc	agc	gcc	gca	aaa	cgt	ctc	aag	gag	gct	aac	cgt	ccg	1200
Leu	Pro	Ile	Gly	Ser	Ala	Ala	Lys	Arg	Leu	Lys	Glu	Ala	Asn	Arg	Pro	
385					390					395					400	
gtt	cca	agt	atg	atg	atg	atc	agt	aat	aac	gtt	tca	gag	agt	gag	aat	1248
Val	Pro	Ser	Met	Met	Met	Ile	Ser	Asn	Asn	Val	Ser	Glu	Ser	Glu	Asn	
				405					410					415		
aat	gct	agc	ggc	tgg	caa	aac	gct	gcg	gtt	cag	cat	cat	cag	gga	gt a	1296
Asn	Ala	Ser	Gly	Trp	Gln	Asn	Ala	Ala	Val	Gln	His	His	Gln	Gly	Val	
			420					425					430			
gat	t t g	agc	tta	t t g	cag	caa	cat	caa	gag	agg	t ac	aat	ggc	t at	t at	1344
Asp	Leu	Ser	Leu	Leu	Gln	Gln	His	Gln	Glu	Arg	Tyr	Asn	Gly	Tyr	Tyr	
		435					440					445				

5312WOPCT_SEQ_LI STI NG TXT

t ac	aat	gga	gga	aac	t t g	t c t	t c g	gag	agt	gct	agg	gct	tgt	t t c	aaa	1392
Tyr	Asn	Gly	Gly	Asn	Leu	Ser	Ser	Glu	Ser	Ala	Arg	Ala	Cys	Phe	Lys	
	450					455					460					
caa	gag	gat	gat	caa	cac	cat	t t c	t t g	agc	aac	acg	cag	agc	ct c	at g	1440
G n	G u	Asp	Asp	G n	H i s	H i s	Phe	Leu	Ser	Asn	Thr	G n	Ser	Leu	Met	
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act	aat	at c	gat	cat	caa	agt	t c t	g t t	t c a	gat	gat	t c g	g t t	act	g t t	1488
Thr	Asn	I l e	Asp	H i s	G n	Ser	Ser	Val	Ser	Asp	Asp	Ser	Val	Thr	Val	
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tgt	gga	aat	g t t	g t t	ggt	t at	ggt	ggt	t at	caa	gga	t t t	gca	gcc	ccg	1536
Cys	Gly	Asn	Val	Val	Gly	Tyr	Gly	Gly	Tyr	G n	Gly	Phe	Ala	Ala	Pro	
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g t t	aac	tgc	gat	gcc	t ac	gct	gct	agt	gag	t t t	gac	t at	aac	gca	aga	1584
Val	Asn	Cys	Asp	Ala	Tyr	Ala	Ala	Ser	Glu	Phe	Asp	Tyr	Asn	Ala	Arg	
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aac	cat	t at	t ac	t t t	gct	cag	cag	cag	cag	acc	cag	cat	t c g	cca	gga	1632
Asn	H i s	Tyr	Tyr	Phe	Ala	G n	G n	G n	G n	Thr	G n	H i s	Ser	Pro	Gly	
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gga	gat	t t t	ccc	gcg	gca	at g	acg	aat	aat	g t t	ggc	t c t	aat	at g	t at	1680
Gly	Asp	Phe	Pro	Ala	Ala	Met	Thr	Asn	Asn	Val	Gly	Ser	Asn	Met	Tyr	
545					550					555					560	
t ac	cat	ggg	gaa	ggt	ggt	gga	gaa	g t t	gct	cca	aca	t t t	aca	g t t	t gg	1728
Tyr	H i s	Gly	Glu	Gly	Gly	Gly	Glu	Val	Ala	Pro	Thr	Phe	Thr	Val	Trp	
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Asn	Asp	Asn														

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<211> 579

<212> PRT

<213> Brassica napus

<400> 79

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			20					25					30		
Val	Ala	Gly	Glu	Tyr	Cys	Tyr	Asp	Pro	Thr	Ala	Ala	Ser	Asp	Glu	Ser
		35					40					45			
Ser	Ala	I l e	G n	Thr	Ser	Phe	Pro	Ser	Pro	Phe	Gly	Val	Val	Leu	Asp
	50					55				60					
Ala	Phe	Thr	Arg	Asp	Asn	Ser	H i s	Ser	Arg	Asp	Trp	Asp	I l e	Asn	
65					70				75					80	
Gly	Ser	Ala	Cys	Asn	Asn	I l e	H i s	Asn	Asp	Glu	G n	Asp	Gly	Pro	Lys
			85					90					95		
Leu	Glu	Asn	Phe	Leu	Gly	Arg	Thr	Thr	Thr	I l e	Tyr	Asn	Thr	Asn	Glu
			100					105					110		
Asn	Val	Gly	Asp	I l e	Asp	Gly	Ser	Gly	Cys	Tyr	Gly	Gly	Gly	Asp	Gly
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Gly	Gly	Gly	Ser	Leu	Gly	Leu	Ser	Met	I l e	Lys	Thr	Trp	Leu	Arg	Asn
		130				135					140				
G n	Pro	Val	Asp	Asn	Val	Asp	Asn	G n	Glu	Asn	Gly	Asn	Gly	Ala	Lys
145					150				155					160	
Gly	Leu	Ser	Leu	Ser	Met	Asn	Ser	Ser	Thr	Ser	Cys	Asp	Asn	Asn	Asn
				165					170					175	
Tyr	Ser	Ser	Asn	Asn	Leu	Val	Ala	G n	Gly	Lys	Thr	I l e	Asp	Asp	Ser
			180					185					190		
Val	Glu	Ala	Thr	Pro	Lys	Lys	Thr	I l e	Glu	Ser	Phe	Gly	G n	Arg	Thr

5312WOPCT_SEQ_LI STI NG TXT

Ser Ile Tyr Arg Gly Val Thr Arg His Arg Trp Thr Gly Arg Tyr Gu
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 Ala His Leu Trp Asp Asn Ser Cys Lys Arg Gu Gn Thr Arg Lys
 210 215 220
 225 Gly Arg Gn Val Tyr Leu Gly Gly Tyr Asp Lys Gu Gu Lys Ala Ala
 230 235 240
 Arg Ala Tyr Asp Leu Ala Ala Leu Lys Tyr Trp Gly Thr Thr Thr Thr
 245 250 255
 Thr Asn Phe Pro Met Ser Gu Tyr Gu Lys Gu Ile Gu Gu Met Lys
 260 265 270
 His Met Thr Arg Gn Gu Tyr Val Ala Ser Leu Arg Arg Lys Ser Ser
 275 280 285
 Gly Phe Ser Arg Gly Ala Ser Ile Tyr Arg Gly Val Thr Arg His His
 290 295 300 305
 Gn His Gly Arg Trp Gn Ala Arg Ile Gly Arg Val Ala Gly Asn Lys
 310 315 320 325
 Asp Leu Tyr Leu Gly Thr Phe Gly Thr Gn Gu Gu Ala Ala Gu Ala
 330 335 340 345
 Tyr Asp Ile Ala Ala Ile Lys Phe Arg Gly Leu Thr Ala Val Thr Asn
 350 355 360 365
 Phe Asp Met Asn Arg Tyr Asn Val Lys Ala Ile Leu Gu Ser Pro Ser
 370 375 380 385
 Leu Pro Ile Gly Ser Ala Ala Lys Arg Leu Lys Gu Ala Asn Arg Pro
 390 395 400 405
 Val Pro Ser Met Met Ile Ser Asn Asn Val Ser Gu Ser Gu Asn
 410 415 420 425
 Asn Ala Ser Gly Trp Gn Asn Ala Ala Val Gn His His Gn Gly Val
 430 435 440 445
 Asp Leu Ser Leu Leu Gn Gn His Gn Gu Arg Tyr Asn Gly Tyr Tyr
 450 455 460 465
 Tyr Asn Gly Gly Asn Leu Ser Ser Gu Ser Ala Arg Ala Cys Phe Lys
 470 475 480 485
 Gn Gu Asp Asp Gn His His Phe Leu Ser Asn Thr Gn Ser Leu Met
 490 495 500 505
 Thr Asn Ile Asp His Gn Ser Ser Val Ser Asp Asp Ser Val Thr Val
 510 515 520 525
 Cys Gly Asn Val Val Gly Tyr Gly Gly Tyr Gn Gly Phe Ala Ala Pro
 530 535 540 545
 Val Asn Cys Asp Ala Tyr Ala Ala Ser Gu Phe Asp Tyr Asn Ala Arg
 550 555 560 565
 Asn His Tyr Tyr Phe Ala Gn Gn Gn Thr Gn His Ser Pro Gly
 570 575
 Gly Asp Phe Pro Ala Ala Met Thr Asn Asn Val Gly Ser Asn Met Tyr
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<220>
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 1 5 10 15

cat cca tca aca caa gat caa acg gtg gct tcc cgt ttt ggg ttc aac 96
 His Pro Ser Thr Gn Asp Gn Thr Val Ala Ser Arg Phe Gly Phe Asn
 20 25 30

5312WOPCT_SEQ_LI STI NG TXT

cct Pro	aat Asn	gaa G u 35	atc I le	tca Ser	ggc G y	tct Ser	gat Asp 40	gtt Val	caa G n	gga G y	gat Asp	cac H s 45	tgc Cys	tat Tyr	gat Asp	144
ctc Leu	tct Ser 50	tct Ser	cac H s	aca Thr	act Thr	cct Pro 55	cat H s	cat H s	tca Ser	ctc Leu	aac Asn 60	ctt Leu	tct Ser	cat H s	cct Pro	192
ttt Phe 65	tcc Ser	att I le	tat Tyr	gaa G u	gct Al a 70	ttc Phe	cac H s	aca Thr	aat Asn	aac Asn 75	aac Asn	att I le	cac H s	acc Thr	act Thr 80	240
caa G n	gat Asp	tgg Trp	aag Lys	gag G u 85	aac Asn	tac Tyr	aac Asn	aac Asn	caa G n 90	aac Asn	cta Leu	cta Leu	ttg Leu	gga G y 95	aca Thr	288
tca Ser	tgc Cys	atg Met	aac Asn 100	caa G n	aat Asn	gtg Val	aac Asn 105	aac Asn	aac Asn	aac Asn	caa G n	caa G n	gca Al a 110	caa G n	cca Pro	336
aag Lys	cta Leu	gaa G u 115	aac Asn	ttc Phe	ctc Leu	ggt G y 120	gga G y	cac H s	tct Ser	ttc Phe	acc Thr	gac Asp 125	cat H s	caa G n	gaa G u	384
tac Tyr	ggt G y 130	ggt G y	agc Ser	aac Asn	tca Ser	tac Tyr 135	tct Ser	tca Ser	tta Leu	cac H s	ctc Leu 140	cca Pro	cct Pro	cat H s	cag G n	432
ccg Pro 145	gaa G u	gca Al a	tcc Ser	tgt Cys	ggc G y 150	ggt G y	ggt G y	gat Asp	ggt G y	agt Ser 155	aca Thr	agt Ser	aac Asn	aat Asn	aac Asn 160	480
tca Ser	ata I le	ggt G y	tta Leu	tct Ser 165	atg Met	ata I le	aaa Lys	aca Thr	tgg Trp 170	ctc Leu	aga Arg	aac Asn	caa G n	cca Pro 175	cca Pro	528
cca Pro	cca Pro	gaa G u 180	aac Asn	aac Asn	aac Asn	aat Asn	aac Asn 185	aac Asn	aat Asn	gaa G u	agt Ser	ggt G y	gca Al a 190	cgt Arg	gtg Val	576
cag G n	aca Thr	cta Leu 195	tca Ser	ctt Leu	tct Ser	atg Met	agt Ser 200	act Thr	ggc G y	tca Ser	cag G n	tca Ser 205	agt Ser	tca Ser	tct Ser	624
gtg Val 210	cct Pro	ctt Leu	ctc Leu	aat Asn	gca Al a 215	aat Asn	gtg Val	atg Met	agt Ser	ggt G y	gag G u 220	att I le	tcc Ser	tca Ser	tcg Ser	672
gaa G u 225	aac Asn	aaa Lys	caa G n	cca Pro	ccc Pro 230	aca Thr	act Thr	gca Al a	gtt Val	gta Val 235	ctt Leu	gat Asp	agc Ser	aac Asn	caa G n 240	720
aca Thr	agt Ser	gtc Val	gtt Val	gaa G u 245	agt Ser	gct Al a	gtg Val	cct Pro	aga Arg 250	aaa Lys	tcc Ser	gtt Val	gat Asp	aca Thr 255	ttt Phe	768
gga G y	caa G n	aga Arg	act Thr 260	tcc Ser	att I le	tac Tyr	cgt Arg	ggt G y 265	gta Val	aca Thr	agg Arg	cat H s	aga Arg 270	tgg Trp	aca Thr	816
ggg G y	aga Arg	tat Tyr 275	gaa G u	gct Al a	cac H s	ctt Leu	tgg Trp 280	gat Asp	aat Asn	agt Ser	tgt Cys	aga Arg 285	aga Arg	gag G u	ggg G y	864
cag G n 290	act Thr	cgc Arg	aaa Lys	gga G y	agg Arg	caa G n 295	gtt Val	tac Tyr	tta Leu	gga G y	ggt G y 300	tat Tyr	gac Asp	aaa Lys	gaa G u	912

5312WOPCT_SEQ_LI STI NG TXT

gaa G u 305	aaa Lys	gca Al a	gct Al a	aga Arg	gcc Al a 310	t at Tyr	gat Asp	t t g Leu	gca Al a 315	gca Al a	ct a Leu	aaa Lys	t at Tyr	t gg Tr p	gga G y 320	960
aca Thr	act Thr	act Thr	aca Thr 325	aca Thr	aat Asn	t t t Phe	cca Pro	att I le	agc Ser 330	cat Hi s	t at Tyr	gaa G u	aaa Lys	gaa G u 335	gt g Val	1008
gaa G u	gaa G u	at g Met	aag Lys 340	cat Hi s	at g Met	aca Thr	agg Arg	caa G n 345	gag G u	t ac Tyr	gt t Val	gcg Al a	t ca Ser 350	t t g Leu	aga Arg	1056
agg Arg	aaa Lys	agt Ser 355	agt Ser	ggg G y	t t t Phe	t ca Ser	cga Arg 360	ggg G y	gca Al a	t cc Ser	att I le	t ac Tyr 365	cga Arg	gga G y	gt a Val	1104
aca Thr 370	aga Arg	cat Hi s	cat Hi s	caa G n	cat Hi s	ggg G y 375	aga Arg	t gg Tr p	caa G n	gct Al a	agg Arg 380	att I le	gga G y	aga Arg	gt t Val	1152
gca Al a 385	ggc G y	aac Asn	aaa Lys	gat Asp	ct c Leu 390	t ac Tyr	ct a Leu	gga G y	act Thr	t t c Phe 395	agc Ser	act Thr	caa G n	gaa G u 400	gag G u	1200
gca Al a	gca Al a	gag G u	gca Al a	t at Tyr 405	gat Asp	gt g Val	gca Al a	gca Al a	at a I le 410	aaa Lys	t t c Phe	aga Arg	gga G y	ct g Leu 415	agt Ser	1248
gca Al a	gt t Val	aca Thr	aac Asn 420	t t t Phe	gac Asp	at g Met	agc Ser 425	aga Arg	t at Tyr	gat Asp	gt c Val	aaa Lys	acc Thr 430	at a I le	ct t Leu	1296
gag G u	agc Ser	agc Ser 435	aca Thr	t t a Leu	cca Pro	att I le	ggg G y 440	ggg G y	gct Al a	gca Al a	aag Lys	cgt Arg 445	t t a Leu	aaa Lys	gac Asp	1344
at g Met 450	gag G u	caa G n	gt t Val	gaa G u	t t g Leu	aat Asn 455	cat Hi s	gt g Val	aat Asn	gt t Val	gat Asp 460	att I le	agc Ser	cat Hi s	aga Arg	1392
act Thr 465	gaa G u	caa G n	gat Asp	cat Hi s	agc Ser 470	at c I le	at c I le	aac Asn	aac Asn	act Thr 475	t cc Ser	cat Hi s	t t a Leu	aca Thr	gaa G u 480	1440
caa G n	gcc Al a	at c I le	t at Tyr	gca Al a 485	gca Al a	aca Thr	aat Asn	gca Al a	t ct Ser 490	aat Asn	t gg Tr p	cat Hi s	gca Al a	ct t Leu 495	t ca Ser	1488
t t c Phe	caa G n	cat Hi s	caa G n 500	caa G n	cca Pro	cat Hi s	cat Hi s	cat Hi s 505	t ac Tyr	aat Asn	gcc Al a	aac Asn	aac Asn 510	at g Met	cag G n	1536
t t a Leu	cag G n	aat Asn 515	t at Tyr	cct Pro	t at Tyr	gga G y 520	act Thr	caa G n	act Thr	caa G n	aag Lys	ct t Leu 525	t gg Tr p	t gc Cys	aaa Lys	1584
caa G n 530	gaa G u	caa G n	gat Asp	t ct Ser	gat Asp	gat Asp 535	cat Hi s	agt Ser	act Thr	t at Tyr	act Thr 540	act Thr	gct Al a	act Thr	gat Asp	1632
att I le 545	cat Hi s	caa G n	ct a Leu	cag G n	t t a Leu 550	ggg G y	aat Asn	aat Asn	aat Asn	aac Asn 555	aat Asn	act Thr	cac Hi s	aat Asn	t t c Phe 560	1680
t t t Phe	ggg G y	t t a Leu	caa G n	aat Asn 565	at c I le	at g Met	agt Ser	at g Met	gat Asp 570	t ct Ser	gct Al a	t cc Ser	at g Met	gat Asp 575	aat Asn	1728

5312WOPCT_SEQ_LI STI NG TXT

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Ser	Ser	G y	Ser	Asn	Ser	Val	Val	Tyr	G y	G y	G y	Asp	His	G y	G y	
			580					585					590			
t at	gga	gga	aat	ggt	gga	t at	at g	att	cca	at g	gct	att	gca	aat	gat	1824
Tyr	G y	G y	Asn	G y	G y	Tyr	Met	I le	Pro	Met	Al a	I le	Al a	Asn	Asp	
		595					600					605				
ggt	aac	caa	aat	cca	aga	agc	aac	aac	aat	tt t	ggt	gag	agt	gag	att	1872
G y	Asn	G n	Asn	Pro	Arg	Ser	Asn	Asn	Asn	Phe	G y	G u	Ser	G u	I le	
	610					615					620					
aaa	gga	tt t	ggt	t at	gaa	aat	gt t	tt t	ggg	act	act	act	gat	cct	t at	1920
Lys	G y	Phe	G y	Tyr	G u	Asn	Val	Phe	G y	Thr	Thr	Thr	Asp	Pro	Tyr	
625					630					635					640	
cat	gca	cag	gca	gca	agg	aac	tt g	t ac	t at	cag	cca	caa	caa	tt a	t ct	1968
His	Al a	G n	Al a	Al a	Arg	Asn	Leu	Tyr	Tyr	G n	Pro	G n	G n	Leu	Ser	
				645					650					655		
gt t	gat	caa	gga	t ca	aat	t gg	gt t	cca	act	gct	att	cca	aca	ct t	gct	2016
Val	Asp	G n	G y	Ser	Asn	Trp	Val	Pro	Thr	Al a	I le	Pro	Thr	Leu	Al a	
			660					665					670			
cca	agg	act	acc	aat	gt c	t ct	ct a	t gt	cct	cct	tt c	act	tt g	tt g	cat	2064
Pro	Arg	Thr	Thr	Asn	Val	Ser	Leu	Cys	Pro	Pro	Phe	Thr	Leu	Leu	His	
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gaa	t ag															2070
G u																

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<400> 81

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His	Pro	Ser	Thr	G n	Asp	G n	Thr	Val	Al a	Ser	Arg	Phe	G y	Phe	Asn
			20					25					30		
Pro	Asn	G u	I le	Ser	G y	Ser	Asp	Val	G n	G y	Asp	His	Cys	Tyr	Asp
		35					40					45			
Leu	Ser	Ser	His	Thr	Thr	Pro	His	His	Ser	Leu	Asn	Leu	Ser	His	Pro
		50				55					60				
Phe	Ser	I le	Tyr	G u	Al a	Phe	His	Thr	Asn	Asn	Asn	I le	His	Thr	Thr
65					70					75					80
G n	Asp	Trp	Lys	G u	Asn	Tyr	Asn	Asn	G n	Asn	Leu	Leu	Leu	G y	Thr
			85					90					95		
Ser	Cys	Met	Asn	G n	Asn	Val	Asn	Asn	Asn	G n	G n	Al a	G n	Pro	
			100					105				110			
Lys	Leu	G u	Asn	Phe	Leu	G y	G y	His	Ser	Phe	Thr	Asp	His	G n	G u
		115					120					125			
Tyr	G y	G y	Ser	Asn	Ser	Tyr	Ser	Ser	Leu	His	Leu	Pro	Pro	His	G n
		130				135					140				
Pro	G u	Al a	Ser	Cys	G y	G y	G y	Asp	G y	Ser	Thr	Ser	Asn	Asn	Asn
145					150					155					160
Ser	I le	G y	Leu	Ser	Met	I le	Lys	Thr	Trp	Leu	Arg	Asn	G n	Pro	Pro
			165					170					175		
Pro	Pro	G u	Asn	Asn	Asn	Asn	Asn	Asn	G u	Ser	G y	Al a	Arg	Val	
			180					185				190			
G n	Thr	Leu	Ser	Leu	Ser	Met	Ser	Thr	G y	Ser	G n	Ser	Ser	Ser	Ser
		195				200					205				
Val	Pro	Leu	Leu	Asn	Al a	Asn	Val	Met	Ser	G y	G u	I le	Ser	Ser	Ser
		210				215					220				

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Glu	Asn	Lys	Gln	Pro	Pro	Thr	Thr	Ala	Val	Val	Leu	Asp	Ser	Asn	Gln
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Thr	Ser	Val	Val	Glu	Ser	Ala	Val	Pro	Arg	Lys	Ser	Val	Asp	Thr	Phe
				245					250					255	
Gly	Gln	Arg	Thr	Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	Arg	Trp	Thr
			260					265					270		
Gly	Arg	Tyr	Glu	Ala	His	Leu	Trp	Asp	Asn	Ser	Cys	Arg	Arg	Glu	Gly
		275					280					285			
Gln	Thr	Arg	Lys	Gly	Arg	Gln	Val	Tyr	Leu	Gly	Gly	Tyr	Asp	Lys	Glu
	290					295					300				
Glu	Lys	Ala	Ala	Arg	Ala	Tyr	Asp	Leu	Ala	Ala	Leu	Lys	Tyr	Trp	Gly
305					310				315					320	
Thr	Thr	Thr	Thr	Thr	Asn	Phe	Pro	Ile	Ser	His	Tyr	Glu	Lys	Glu	Val
				325					330					335	
Glu	Glu	Met	Lys	His	Met	Thr	Arg	Gln	Glu	Tyr	Val	Ala	Ser	Leu	Arg
			340					345					350		
Arg	Lys	Ser	Ser	Gly	Phe	Ser	Arg	Gly	Ala	Ser	Ile	Tyr	Arg	Gly	Val
		355					360					365			
Thr	Arg	His	His	Gln	His	Gly	Arg	Trp	Gln	Ala	Arg	Ile	Gly	Arg	Val
	370					375					380				
Ala	Gly	Asn	Lys	Asp	Leu	Tyr	Leu	Gly	Thr	Phe	Ser	Thr	Gln	Glu	Glu
385					390					395				400	
Ala	Ala	Glu	Ala	Tyr	Asp	Val	Ala	Ala	Ile	Lys	Phe	Arg	Gly	Leu	Ser
				405					410					415	
Ala	Val	Thr	Asn	Phe	Asp	Met	Ser	Arg	Tyr	Asp	Val	Lys	Thr	Ile	Leu
			420					425					430		
Glu	Ser	Ser	Thr	Leu	Pro	Ile	Gly	Gly	Ala	Ala	Lys	Arg	Leu	Lys	Asp
		435					440					445			
Met	Glu	Gln	Val	Glu	Leu	Asn	His	Val	Asn	Val	Asp	Ile	Ser	His	Arg
	450					455					460				
Thr	Glu	Gln	Asp	His	Ser	Ile	Ile	Asn	Asn	Thr	Ser	His	Leu	Thr	Glu
465					470					475				480	
Gln	Ala	Ile	Tyr	Ala	Ala	Thr	Asn	Ala	Ser	Asn	Trp	His	Ala	Leu	Ser
				485					490					495	
Phe	Gln	His	Gln	Gln	Pro	His	His	His	Tyr	Asn	Ala	Asn	Asn	Met	Gln
			500					505					510		
Leu	Gln	Asn	Tyr	Pro	Tyr	Gly	Thr	Gln	Thr	Gln	Lys	Leu	Trp	Cys	Lys
		515					520					525			
Gln	Glu	Gln	Asp	Ser	Asp	Asp	His	Ser	Thr	Tyr	Thr	Thr	Ala	Thr	Asp
	530					535					540				
Ile	His	Gln	Leu	Gln	Leu	Gly	Asn	Asn	Asn	Asn	Asn	Thr	His	Asn	Phe
545					550					555				560	
Phe	Gly	Leu	Gln	Asn	Ile	Met	Ser	Met	Asp	Ser	Ala	Ser	Met	Asp	Asn
				565					570					575	
Ser	Ser	Gly	Ser	Asn	Ser	Val	Val	Tyr	Gly	Gly	Gly	Asp	His	Gly	Gly
		580						585					590		
Tyr	Gly	Gly	Asn	Gly	Gly	Tyr	Met	Ile	Pro	Met	Ala	Ile	Ala	Asn	Asp
		595					600					605			
Gly	Asn	Gln	Asn	Pro	Arg	Ser	Asn	Asn	Asn	Phe	Gly	Glu	Ser	Glu	Ile
	610					615					620				
Lys	Gly	Phe	Gly	Tyr	Glu	Asn	Val	Phe	Gly	Thr	Thr	Thr	Asp	Pro	Tyr
625					630					635				640	
His	Ala	Gln	Ala	Ala	Arg	Asn	Leu	Tyr	Tyr	Gln	Pro	Gln	Gln	Leu	Ser
				645					650					655	
Val	Asp	Gln	Gly	Ser	Asn	Trp	Val	Pro	Thr	Ala	Ile	Pro	Thr	Leu	Ala
			660					665					670		
Pro	Arg	Thr	Thr	Asn	Val	Ser	Leu	Cys	Pro	Pro	Phe	Thr	Leu	Leu	His
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5312WOPCT_SEQ_LI STI NG TXT

<221> CDS

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<400> 82

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1 5 10 15	
cct tct agt caa gat cac tct caa acg gca cct tct cgt ttt tgc ttc	96
Pro Ser Ser Gln Asp His Ser Gln Thr Ala Pro Ser Arg Phe Cys Phe	
20 25 30	
aac cct gat gga atc tca agc act gat gta gca gga gac tgc ttt gat	144
Asn Pro Asp Gly Ile Ser Ser Thr Asp Val Ala Gly Asp Cys Phe Asp	
35 40 45	
ctc act tct gac tca act cct cat tta ctg aac ctt ccc tct tac ggc	192
Leu Thr Ser Asp Ser Thr Pro His Leu Leu Asn Leu Pro Ser Tyr Gly	
50 55 60	
ata tac gaa gct ttt cat agg agc aac aat att cac acc act caa gat	240
Ile Tyr Gu Ala Phe His Arg Ser Asn Asn Ile His Thr Thr Gln Asp	
65 70 75 80	
tgg aag gag aac tac aac agc caa aac ttg cta ttg gga act tca tgc	288
Trp Lys Gu Asn Tyr Asn Ser Gln Asn Leu Leu Gly Thr Ser Cys	
85 90 95	
agc aac caa aac atg aac cac aac cat cag caa caa caa caa caa cag	336
Ser Asn Gln Asn Met Asn His Asn His Gln Gln Gln Gln Gln Gln	
100 105 110	
cca aag ctt gaa aac ttc ctg ggt gga cac tca ttt ggt gaa cat gag	384
Pro Lys Leu Gu Asn Phe Leu Gly Gly His Ser Phe Gly Gu His Gu	
115 120 125	
caa ccc tac ggt ggt aac tca gcc tct aca gaa tac atg ttc ccg gct	432
Gln Pro Tyr Gly Gly Asn Ser Ala Ser Thr Gu Tyr Met Phe Pro Ala	
130 135 140	
cag ccg gta ttg gcc ggt ggc ggc ggc ggt ggt agc aat agc agc aac	480
Gln Pro Val Leu Ala Gly Gly Gly Gly Gly Gly Ser Asn Ser Ser Asn	
145 150 155 160	
aca agc aac agt agc tcc ata ggg tta tcc atg ata aag aca tgg ttg	528
Thr Ser Asn Ser Ser Ile Gly Leu Ser Met Ile Lys Thr Trp Leu	
165 170 175	
agg aac caa cca cca cac tca gaa aac aac aat aac aac aac aat gaa	576
Arg Asn Gln Pro Pro His Ser Gu Asn Asn Asn Asn Asn Asn Asn Gu	
180 185 190	
agt ggt ggc aat agt aga agc agt gtg cag cag act cta tca ctt tcc	624
Ser Gly Gly Asn Ser Arg Ser Ser Val Gln Gln Thr Leu Ser Leu Ser	
195 200 205	
atg agt act ggt tca caa tca agc aca tca cta ccc ctt ctg act gct	672
Met Ser Thr Gly Ser Gln Ser Ser Thr Ser Leu Pro Leu Leu Thr Ala	
210 215 220	
agt gtg gat aat gga gag agt tct tct gat aac aaa caa cca cat acc	720
Ser Val Asp Asn Gly Gu Ser Ser Ser Asp Asn Lys Gln Pro His Thr	
225 230 235 240	
acg gct gca ctt gat aca acc caa acc gga gcc att gaa act gca ccc	768
Thr Ala Ala Leu Asp Thr Thr Gln Thr Gly Ala Ile Gu Thr Ala Pro	
245 250 255	

5312WOPCT_SEQ_LI STING.TXT

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gta Val	aca Thr	agg Arg 275	cat His	agg Arg	tgg Trp	acg Thr	ggg Gly 280	agg Arg	tat Tyr	gag Glu	gct Ala	cac His 285	ctg Leu	tgg Trp	gat Asp	864
aat Asn 290	agt Ser	tgt Cys	aga Arg	aga Arg	gag Glu	gga Gly 295	caa Gln	act Thr	cgc Arg	aaa Lys	gga Gly 300	agg Arg	caa Gln	gtt Val	tac Tyr	912
ttg Leu 305	gga Gly	ggt Gly	tat Tyr	gac Asp	aaa Lys 310	gaa Glu	gaa Glu	aag Lys	gca Ala	gct Ala 315	aga Arg	gcc Ala	tac Tyr	gat Asp	ttg Leu 320	960
gca Ala	gca Ala	cta Leu	aaa Lys	tac Tyr 325	tgg Trp	gga Gly	aca Thr	act Thr	acg Thr 330	aca Thr	aca Thr	aat Asn	ttt Phe	cca Pro 335	att Ile	1008
agc Ser	cac His	tat Tyr	gag Glu 340	aaa Lys	gag Glu	ttg Leu	gaa Glu	gaa Glu 345	atg Met	aag Lys	cac His	atg Met	act Thr 350	agg Arg	caa Gln	1056
gag Glu	tac Tyr	gtt Val 355	gcg Ala	tca Ser	ttg Leu	aga Arg	agg Arg 360	aag Lys	agt Ser	agt Ser	ggg Gly	ttt Phe 365	tct Ser	cgc Arg	ggg Gly	1104
gca Ala	tcc Ser 370	att Ile	tat Tyr	cga Arg	ggt Gly	gtg Val 375	acg Thr	aga Arg	cac His	cat His	caa Gln 380	cat His	gga Gly	aga Arg	tgg Trp	1152
caa Gln 385	gcg Ala	agg Arg	att Ile	gga Gly	aga Arg	gtt Val	gct Ala	ggc Gly	aac Asn	aag Lys 395	gat Asp	ctc Leu	tac Tyr	ttg Leu	gga Gly 400	1200
act Thr	ttc Phe	agc Ser	acc Thr	caa Gln 405	gag Glu	gag Glu	gca Ala	gca Ala	gaa Glu 410	gca Ala	tat Tyr	gat Asp	gta Val	gca Ala 415	gca Ala	1248
atc Ile	aaa Lys	ttc Phe	aga Arg 420	gga Gly	cta Leu	agt Ser	gct Ala 425	gtt Val	aca Thr	aac Asn	ttt Phe	gac Asp 430	atg Met	agc Ser	aga Arg	1296
tat Tyr	gac Asp	gtg Val 435	aaa Lys	agc Ser	ata Ile	ctt Leu	gag Glu 440	agc Ser	acc Thr	act Thr	ttg Leu	cca Pro 445	att Ile	ggt Gly	ggt Gly	1344
gct Ala 450	gca Ala	aag Lys	cgt Arg	ttg Leu	aag Lys	gat Asp 455	atg Met	gag Glu	cag Gln	gtg Val	gaa Glu 460	ctg Leu	agg Arg	gtg Val	gag Glu	1392
aat Asn 465	gtt Val	cat His	aga Arg	gca Ala	gat Asp 470	caa Gln	gaa Glu	gat Asp	cat His	agt Ser 475	agc Ser	atc Ile	atg Met	aac Asn	tct Ser 480	1440
cac His	tta Leu	act Thr	caa Gln 485	gga Gly	atc Ile	att Ile	aac Asn	aac Asn	tat Tyr 490	gca Ala	gca Ala	gga Gly	gga Gly	aca Thr 495	aca Thr	1488
gcg Ala	act Thr	cat His	cat His 500	cat His	aac Asn	tgg Trp	cac His	aat Asn 505	gct Ala	ctt Leu	gca Ala	ttc Phe	cac His 510	caa Gln	cct Pro	1536
caa Gln	cct Pro	tgc Cys 515	acc Thr	acc Thr	ata Ile	cac His	tac Tyr 520	cct Pro	tat Tyr	gga Gly	caa Gln	aga Arg 525	att Ile	aat Asn	tgg Trp	1584

5312WOPCT_SEQ_LI STI NG TXT

tgc Cys	aag Lys	caa Gln	gaa Glu	caa Gln	gac Asp	aac Asn	tct Ser	gat Asp	gcc Ala	tct Ser	cac His	tct Ser	ttg Leu	tct Ser	tat Tyr	1632
530 535 540																
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545 550 555 560																
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565 570 575																
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580 585 590																
gga Gly	ggg Gly	ggg Gly	ggg Gly	ggc Gly	tat Tyr	aat Asn	gtg Val	att Ile	cct Pro	atg Met	ggg Gly	act Thr	act Thr	act Thr	act Thr	1824
595 600 605																
gtt Val	gtt Val	gca Ala	aat Asn	gat Asp	ggg Gly	gat Asp	caa Gln	aat Asn	cca Pro	aga Arg	agc Ser	aat Asn	cat His	ggg Gly	ttt Phe	1872
610 615 620																
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625 630 635 640																
aca Thr	act Thr	gat Asp	cct Pro	tat Tyr	cat His	gca Ala	cat His	gca Ala	agg Arg	aac Asn	ttg Leu	tat Tyr	tat Tyr	ctt Leu	act Thr	1968
645 650 655																
caa Gln	cag Gln	caa Gln	cca Pro	tct Ser	tct Ser	gtt Val	gat Asp	gca Ala	gtg Val	aag Lys	gct Ala	agt Ser	gca Ala	tat Tyr	gat Asp	2016
660 665 670																
caa Gln	gga Gly	tct Ser	gca Ala	tgc Cys	aat Asn	act Thr	tgg Trp	gtt Val	cca Pro	act Thr	gct Ala	att Ile	cca Pro	act Thr	cat His	2064
675 680 685																
gca Ala	cca Pro	agg Arg	tct Ser	agt Ser	act Thr	agt Ser	atg Met	gct Ala	ctc Leu	tgc Cys	cat His	ggg Gly	gct Ala	acg Thr	ccc Pro	2112
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705 710																

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 <212> PRT
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<400> 83
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 20 25 30
 Asn Pro Asp Gly Ile Ser Ser Thr Asp Val Ala Gly Asp Cys Phe Asp
 35 40 45
 Leu Thr Ser Asp Ser Thr Pro His Leu Leu Asn Leu Pro Ser Tyr Gly
 50 55 60
 Ile Tyr Glu Ala Phe His Arg Ser Asn Asn Ile His Thr Thr Gln Asp
 65 70 75 80
 Trp Lys Glu Asn Tyr Asn Ser Gln Asn Leu Leu Leu Gly Thr Ser Cys
 85 90 95
 Ser Asn Gln Asn Met Asn His Asn His Gln Gln Gln Gln Gln Gln

5312 WOPCT_SEQ_LI STI NG TXT

Pro	Lys	Leu	100	Glu	Asn	Phe	Leu	Gly	105	Gly	His	Ser	Phe	Gly	110	Glu	His	Glu
Gln	Pro	115	Tyr	Gly	Gly	Asn	Ser	120	Ala	Ser	Thr	Glu	Tyr	125	Met	Phe	Pro	Ala
Gln	Pro	130	Val	Leu	Ala	Gly	Gly	135	Gly	Gly	Gly	Gly	Ser	140	Asn	Ser	Ser	Asn
145	Thr	Ser	Asn	Ser	Ser	150	Ser	155	Ile	Gly	Leu	Ser	160	Met	Ile	Lys	Thr	Trp
Arg	Asn	Gln	Pro	165	His	Ser	Glu	170	Asn	Asn	Asn	Asn	175	Asn	Asn	Asn	Asn	Glu
Ser	Gly	Gly	Asn	Ser	Arg	Ser	Ser	185	Val	Gln	Gln	Thr	190	Leu	Ser	Leu	Ser	
Met	Ser	195	Thr	Gly	Ser	Gln	Ser	200	Ser	Thr	Ser	Leu	205	Pro	Leu	Leu	Thr	Ala
Ser	210	Val	Asp	Asn	Gly	Glu	Ser	215	Ser	Ser	Ser	Asp	220	Asn	Lys	Gln	Pro	His
225	Thr	Ala	Ala	Leu	Asp	Thr	Thr	230	Gln	Thr	Gly	Ala	235	Ile	Glu	Thr	Ala	Pro
Arg	Lys	Ser	245	Ile	Asp	Thr	Phe	Gly	Gln	Arg	Thr	Ser	250	Ile	Tyr	Arg	Gly	
Val	Thr	Arg	260	His	Arg	Trp	Thr	Gly	265	Arg	Tyr	Glu	270	Ala	His	Leu	Trp	Asp
Asn	Ser	Cys	275	Arg	Arg	Glu	Gly	280	Gln	Thr	Arg	Lys	285	Gly	Arg	Gln	Val	Tyr
Leu	305	Gly	Gly	Tyr	Asp	Lys	310	Glu	Glu	Lys	Ala	Ala	315	Arg	Ala	Tyr	Asp	Leu
Ala	Ala	Leu	Lys	Tyr	Trp	Gly	Thr	Thr	Thr	Thr	Thr	Thr	330	Thr	Asn	Phe	Pro	Ile
Ser	His	Tyr	340	Glu	Lys	Glu	Leu	Glu	345	Met	Lys	His	350	Met	Thr	Arg	Gln	
Glu	Tyr	Val	355	Ala	Ser	Leu	Arg	Arg	360	Lys	Ser	Ser	365	Gly	Phe	Ser	Arg	Gly
Ala	Ser	Ile	370	Tyr	Arg	Gly	Val	Thr	375	Arg	His	His	380	Gln	His	Gly	Arg	Trp
Gln	385	Ala	Arg	Ile	Gly	Arg	Val	Ala	390	Gly	Asn	Lys	395	Asp	Leu	Tyr	Leu	Gly
Thr	Phe	Ser	Thr	Gln	Glu	Glu	Ala	Ala	405	Glu	Ala	Tyr	410	Asp	Val	Ala	Ala	
Ile	Lys	Phe	420	Arg	Gly	Leu	Ser	Ala	425	Val	Thr	Asn	430	Phe	Asp	Met	Ser	Arg
Tyr	Asp	Val	435	Lys	Ser	Ile	Leu	Glu	440	Ser	Thr	Thr	445	Leu	Pro	Ile	Gly	Gly
Ala	Ala	Lys	450	Arg	Leu	Lys	Asp	Met	455	Glu	Gln	Val	460	Glu	Leu	Arg	Val	Glu
Asn	465	Val	His	Arg	Ala	Asp	Gln	Glu	470	Asp	His	Ser	475	Ser	Ile	Met	Asn	Ser
His	Leu	Thr	Gln	Gly	Ile	Ile	Asn	Asn	485	Tyr	Ala	Ala	490	Gly	Gly	Thr	495	Thr
Ala	Thr	His	500	His	His	Asn	Trp	His	505	Asn	Ala	Leu	510	Ala	Phe	His	Gln	Pro
Gln	Pro	Cys	515	Thr	Thr	Ile	His	Tyr	520	Pro	Tyr	Gly	525	Gln	Arg	Ile	Asn	Trp
Cys	Lys	Gln	Glu	Gln	Asp	Asn	Ser	Asp	535	Ala	Ser	His	540	Ser	Leu	Ser	Tyr	
Ser	545	Asp	Ile	His	Gln	Leu	Gln	Leu	550	Gly	Asn	Asn	555	Gly	Thr	His	Asn	Phe
Phe	His	Thr	Asn	Ser	Gly	Leu	His	Pro	565	Met	Leu	Ser	570	Met	Asp	Ser	575	Ala
Ser	Ile	Asp	Asn	Ser	Ser	Ser	Ser	Asn	580	Ser	Val	Val	585	Tyr	Asp	Gly	Tyr	
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Val	Val	Ala	Asn	Asp	Gly	Asp	Gln	Asn	615	Pro	Arg	Ser	620	Asn	His	Gly	Phe	
Gly	625	Asp	Asn	Glu	Ile	Lys	Ala	Leu	630	Gly	Tyr	Glu	635	Ser	Val	Tyr	Gly	Ser
Thr	Thr	Asp	Pro	Tyr	His	Ala	His	Ala	640	Arg	Asn	Leu	645	Tyr	Tyr	Leu	Thr	

5312WOPCT_SEQ_LI STI NG TXT

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 G n G y Ser Al a Cys Asn Thr Tr p Val Pro Thr Al a I l e Pro Thr Hi s
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 Leu Pro Pro G n Pro G u Asn Hi s Ser G n Asn Ser Val Ser Arg Leu
 20 25 30
 ggt ttc aac tct gat gaa atc tct ggg act gat gtg tca ggt gag tgt 144
 Gly Phe Asn Ser Asp G u I l e Ser Gly Thr Asp Val Ser Gly G u Cys
 35 40 45
 ttt gat ct c act tca gat tcc act gct ccc tct ct c aac ct c cct ccc 192
 Phe Asp Leu Thr Ser Asp Ser Thr Al a Pro Ser Leu Asn Leu Pro Pro
 50 55 60
 cct ttt ggg ata ctt gaa gca ttc aac agg aat aat cag ccc caa gat 240
 Pro Phe Gly I l e Leu G u Al a Phe Asn Arg Asn Asn G n Pro G n Asp
 65 70 75 80
 act aac tac aaa acc acc act tct gag ct c tcc at g ct c at g ggt agt 288
 Thr Asn Tyr Lys Thr Thr Ser G u Leu Ser Met Leu Met Gly Ser
 85 90 95
 tca tgc agt agt cat cat aac ct c gaa aac caa gaa ccc aaa ctt gaa 336
 Ser Cys Ser Ser Hi s Hi s Asn Leu G u Asn G n G u Pro Lys Leu G u
 100 105 110
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 115 120 125
 ggg tac tac att tcc att ggt tta tcc at g at c aag aca tgg ctg cgg 432
 Gly Tyr Tyr I l e Ser I l e Gly Leu Ser Met I l e Lys Thr Trp Leu Arg
 130 135 140
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 Asn G n Pro Al a Pro Thr Hi s G n Asp Asn Asn Lys Ser Thr Asp Thr
 145 150 155 160
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 Gly Pro Val Gly Gly Al a Al a Al a Gly Asn Leu Pro Asn Al a G n Thr
 165 170 175
 tta tgc ttg tcc at g agc acc ggc tgc cac cag acc ggt gcc att gaa 576
 Leu Ser Leu Ser Met Ser Thr Gly Ser Hi s G n Thr Gly Al a I l e G u
 180 185 190

5312WOPCT_SEQ_LI STI NG TXT

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t ac Tyr	cgt Arg 210	ggg Gly	gt a Val	aca Thr	agg Arg	cat His 215	aga Arg	t gg Trp	acg Thr	ggg Gly	aga Arg 220	t at Tyr	gag Glu	gct Ala	cat His	672
ct a Leu 225	t gg Trp	gac Asp	aac Asn	agt Ser	t gc Cys 230	aga Arg	aga Arg	gaa Glu	gga Gly	caa Gln 235	act Thr	cga Arg	aag Lys	gga Gly	agg Arg 240	720
caa Gln	gt t Val	t at Tyr	t t a Leu	ggg Gly 245	ggg Gly	t at Tyr	gac Asp	aaa Lys	gaa Glu 250	gaa Glu	aag Lys	gca Ala	gct Ala	agg Arg 255	gct Ala	768
t ac Tyr	gat Asp	t t a Leu	gca Ala 260	gca Ala	ct g Leu	aag Lys	t at Tyr	t gg Trp 265	ggg Gly	acc Thr	acc Thr	acc Thr	aca Thr 270	aca Thr	aat Asn	816
t t c Phe	cct Pro	att Ile 275	agc Ser	aac Asn	t at Tyr	gaa Glu	aaa Lys 280	gag Glu	at a Ile	gag Glu	gag Glu	at g Met 285	aag Lys	cac His	at g Met	864
aca Thr	agg Arg 290	cag Gln	gag Glu	t ac Tyr	gt a Val	gca Ala 295	t ct Ser	ct g Leu	cga Arg	agg Arg	aag Lys 300	agt Ser	agc Ser	ggg Gly	ttt Phe	912
t ct Ser 305	cgt Arg	gga Gly	gca Ala	t cc Ser	at a Ile 310	t at Tyr	aga Arg	gga Gly	gt g Val	acc Thr 315	aga Arg	cac His	cat His	cag Gln	cat His 320	960
ggg Gly	aga Arg	t gg Trp	cag Gln	gca Ala 325	agg Arg	att Ile	gga Gly	aga Arg	gt c Val 330	gca Ala	ggc Gly	aac Asn	aaa Lys	gat Asp 335	ct t Leu	1008
t ac Tyr	t t g Leu	gga Gly	act Thr 340	t t c Phe	agc Ser	acc Thr	caa Gln	gag Glu 345	gaa Glu	gca Ala	gca Ala	gag Glu	gcc Ala 350	t at Tyr	gac Asp	1056
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gat Asp	gga Gly	at c Ile	aac Asn 420	aac Asn	t at Tyr	gga Gly	gca Ala	cac His 425	cac His	cat His	ggc Gly	t gg Trp	cct Pro 430	act Thr	gt t Val	1296
gca Ala	t t c Phe 435	caa Gln	caa Gln	gct Ala	cag Gln	cca Pro	ttt Phe 440	agc Ser	at g Met	cac His	t ac Tyr	cct Pro 445	t at Tyr	ggc Gly	cat His	1344
cag Gln 450	cag Gln	agg Arg	gct Ala	gt t Val	t gg Trp	t gt Cys 455	aag Lys	caa Gln	gag Glu	caa Gln	gac Asp 460	cct Pro	gat Asp	ggc Gly	aca Thr	1392

5312WOPCT_SEQ_LI STI NG TXT

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ttc Phe	ttc Phe	cag Gln	cct Pro	aat Asn 485	gtt Val	ctg Leu	cac His	aac Asn	ctc Leu 490	atg Met	agc Ser	atg Met	gac Asp	tct Ser 495	tct Ser	1488
tca Ser	atg Met	gac Asp	cat His 500	agc Ser	tca Ser	ggc Gly	tcc Ser	aat Asn 505	tca Ser	gtc Val	atc Ile	tat Tyr	agc Ser 510	ggg Gly	ggg Gly	1536
gga Gly	gcc Ala	gct Ala 515	gat Asp	ggc Gly	agc Ser	gct Ala	gca Ala 520	act Thr	ggc Gly	ggc Gly	agt Ser	ggc Gly 525	agt Ser	ggg Gly	agc Ser	1584
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agc Ser 545	acc Thr	gtc Val	atc Ile	gct Ala	cat His 550	gaa Glu	ggc Gly	ggc Gly	cat His	ggc Gly 555	cag Gln	gga Gly	aat Asn	ggg Gly	ggc Gly 560	1680
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tcg Ser	aca Thr	gat Asp	cct Pro 580	tac Tyr	cat His	gct Ala	agg Arg	agc Ser 585	ttg Leu	tac Tyr	tat Tyr	ctt Leu	tca Ser 590	cag Gln	caa Gln	1776
tca Ser	tct Ser	gca Ala 595	ggc Gly	atg Met	gtg Val	aag Lys	ggc Gly 600	agt Ser	agt Ser	gca Ala	tat Tyr	gat Asp 605	cag Gln	ggg Gly	tca Ser	1824
ggg Gly 610	tgt Cys	aac Asn	aac Asn	tgg Trp	gtt Val	cca Pro 615	act Thr	gca Ala	gtt Val	cca Pro	acc Thr 620	cta Leu	gct Ala	cca Pro	agg Arg	1872
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<211> 643

<212> PRT

<213> Vitis vinifera

<400> 85

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Gly	Phe	Asn 35	Ser	Asp	Glu	Ile	Ser 40	Gly	Thr	Asp	Val	Ser 45	Gly	Glu	Cys
Phe	Asp 50	Leu	Thr	Ser	Asp	Ser 55	Thr	Ala	Pro	Ser	Leu 60	Asn	Leu	Pro	Pro
Pro 65	Phe	Gly	Ile	Leu	Glu 70	Ala	Phe	Asn	Arg	Asn 75	Asn	Gln	Pro	Gln	Asp 80
Thr	Asn	Tyr	Lys	Thr 85	Thr	Thr	Ser	Glu 90	Leu	Ser	Met	Leu	Met	Gly 95	Ser

5312WOPCT_SEQ LISTING.TXT

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Gly	Tyr	Tyr	Ile	Ser	Ile	Gly	Leu	Ser	Met	Ile	Lys	Thr	Trp	Leu	Arg
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Leu	Ser	Leu	Ser	Met	Ser	Thr	Gly	Ser	His	Gln	Thr	Gly	Ala	Ile	Glu
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Thr	Val	Pro	Arg	Lys	Ser	Ile	Asp	Thr	Phe	Gly	Gln	Arg	Thr	Ser	Ile
		195					200					205			
Tyr	Arg	Gly	Val	Thr	Arg	His	Arg	Trp	Thr	Gly	Arg	Tyr	Glu	Ala	His
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Leu	Trp	Asp	Asn	Ser	Cys	Arg	Arg	Glu	Gly	Gln	Thr	Arg	Lys	Gly	Arg
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Gln	Val	Tyr	Leu	Gly	Gly	Tyr	Asp	Lys	Glu	Glu	Lys	Ala	Ala	Arg	Ala
			245						250					255	
Tyr	Asp	Leu	Ala	Ala	Leu	Lys	Tyr	Trp	Gly	Thr	Thr	Thr	Thr	Thr	Asn
			260					265					270		
Phe	Pro	Ile	Ser	Asn	Tyr	Glu	Lys	Glu	Ile	Glu	Glu	Met	Lys	His	Met
		275					280					285			
Thr	Arg	Gln	Glu	Tyr	Val	Ala	Ser	Leu	Arg	Arg	Lys	Ser	Ser	Gly	Phe
	290					295					300				
Ser	Arg	Gly	Ala	Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	His	Gln	His
305					310					315				320	
Gly	Arg	Trp	Gln	Ala	Arg	Ile	Gly	Arg	Val	Ala	Gly	Asn	Lys	Asp	Leu
			325						330					335	
Tyr	Leu	Gly	Thr	Phe	Ser	Thr	Gln	Glu	Ala	Ala	Glu	Ala	Tyr	Asp	
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Ile	Ala	Ala	Ile	Lys	Phe	Arg	Gly	Leu	Asn	Ala	Val	Thr	Asn	Phe	Asp
		355					360					365			
Met	Ser	Arg	Tyr	Asp	Val	Asn	Ser	Ile	Leu	Glu	Ser	Ser	Thr	Leu	Pro
	370					375					380				
Ile	Gly	Gly	Ala	Ala	Lys	Arg	Leu	Lys	Asp	Ala	Glu	Gln	Ala	Glu	Met
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Thr	Ile	Asp	Gly	Gln	Arg	Thr	Asp	Asp	Glu	Met	Ser	Ser	Gln	Leu	Thr
			405						410					415	
Asp	Gly	Ile	Asn	Asn	Tyr	Gly	Ala	His	His	His	Gly	Trp	Pro	Thr	Val
			420					425					430		
Ala	Phe	Gln	Gln	Ala	Gln	Pro	Phe	Ser	Met	His	Tyr	Pro	Tyr	Gly	His
		435					440					445			
Gln	Gln	Arg	Ala	Val	Trp	Cys	Lys	Gln	Glu	Gln	Asp	Pro	Asp	Gly	Thr
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His	Asn	Phe	Gln	Asp	Leu	His	Gln	Leu	Gln	Leu	Gly	Asn	Thr	His	Asn
465					470					475				480	
Phe	Phe	Gln	Pro	Asn	Val	Leu	His	Asn	Leu	Met	Ser	Met	Asp	Ser	Ser
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Ser	Met	Asp	His	Ser	Ser	Gly	Ser	Asn	Ser	Val	Ile	Tyr	Ser	Gly	Gly
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Phe	Gln	Gly	Val	Gly	Tyr	Gly	Asn	Asn	Ile	Gly	Phe	Val	Met	Pro	Ile
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Ser	Thr	Val	Ile	Ala	His	Glu	Gly	Gly	His	Gly	Gln	Gly	Asn	Gly	Gly
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Phe	Gly	Asp	Ser	Glu	Val	Lys	Ala	Ile	Gly	Tyr	Asp	Asn	Met	Phe	Gly
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Ser	Thr	Asp	Pro	Tyr	His	Ala	Arg	Ser	Leu	Tyr	Tyr	Leu	Ser	Gln	Gln
			580					585					590		
Ser	Ser	Ala	Gly	Met	Val	Lys	Gly	Ser	Ser	Ala	Tyr	Asp	Gln	Gly	Ser
		595					600						605		
Gly	Cys	Asn	Asn	Trp	Val	Pro	Thr	Ala	Val	Pro	Thr	Leu	Ala	Pro	Arg
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 Gln Leu Pro Pro Ser Gln Thr Asn Ser Thr Leu Ile Ser Ala Ala Ala
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 Ile Pro Gln Asp Trp Ser Met Arg Gly Ser Gln Leu Ser Ala Leu Val
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 gcc gag ccg aag ct g gag gac ttc ct c ggc ggc at c tcc ttc t cg gag 240
 Ala Glu Pro Lys Leu Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu
 65 70 75
 cag cag cat cat cac ggc ggc aag ggc ggc gt g at c ccg agc agc gcc 288
 Gln Gln His His His Gly Gly Lys Gly Val Ile Pro Ser Ser Ala
 85 90 95
 gcc gct tgc t ac gcg agc tcc ggc agc agc gt c ggc t ac ct g t ac cct 336
 Ala Ala Cys Tyr Ala Ser Ser Gly Ser Ser Val Gly Tyr Leu Tyr Pro
 100 105 110
 cct cca agc t ca tcc t cg ct c cag ttc gcc gac tcc gt c at g gt g gcc 384
 Pro Pro Ser Ser Ser Ser Leu Gln Phe Ala Asp Ser Val Met Val Ala
 115 120 125
 acc tcc t cg ccc gt c gt c gcc cac gac ggc gt c agc ggc ggc ggc at g 432
 Thr Ser Ser Pro Val Val Ala His Asp Gly Val Ser Gly Gly Gly Met
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 Val Ser Ala Ala Ala Ala Ala Ala Ala Ser Gly Asn Gly Gly Ile Gly
 145 150 155 160
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 Leu Ser Met Ile Lys Asn Trp Leu Arg Ser Gln Pro Ala Pro Gln Pro
 165 170 175
 gcg cag gcg ct g t ct ct g tcc at g aac at g gcg ggc acg acg acg gcg 576
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 cag ggc ggc ggc gcc at g gcg ct c ct c gcc ggc gca ggc gag cga ggc 624
 Gln Gly Gly Gly Ala Met Ala Leu Leu Ala Gly Ala Gly Gu Arg Gly
 195 200 205
 cgg acg acg ccc gcg t ca gag agc ct g tcc acg t cg gcg cac gga gcg 672
 Arg Thr Thr Pro Ala Ser Glu Ser Leu Ser Thr Ser Ala His Gly Ala

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agc Ser	ggc Gly	agc Ser	gcc Ala	ggc Gly 245	gcc Ala	gtg Val	gtt Val	gcc Ala	gtc Val 250	ggc Gly	tcg Ser	gag Glu	tca Ser	ggc Gly 255	ggc Gly	768
agc Ser	ggc Gly	gcc Ala	gtg Val 260	gtg Val	gag Glu	gcc Ala	ggc Gly	gcg Ala 265	gcg Ala	gcg Ala	gcg Ala	gcg Ala 270	agg Arg	aag Lys		816
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agg Arg	cat His 290	aga Arg	tgg Trp	aca Thr	ggg Gly	agg Arg 295	tat Tyr	gag Glu	gct Ala	cat His 300	ctt Leu	tgg Trp	gac Asp	aac Asn	agc Ser	912
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gtc Val	aag Lys 450	agc Ser	atc Ile	ctc Leu	gac Asp	agc Ser 455	gct Ala	gcc Ala	ctc Leu	ccc Pro	gtc Val 460	ggc Gly	acc Thr	gcc Ala	gcc Ala	1392
aag Lys 465	cgc Arg	ctc Leu	aag Lys	gac Asp	gcc Ala 470	gag Glu	gcc Ala	gcc Ala	gcc Ala	gcc Ala 475	tac Tyr	gac Asp	gtc Val	ggc Gly	cgc Arg 480	1440
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5312WOPCT_SEQ_LI STI NG TXT

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gcc	gcc	gcc	gcc	gcg	cat	gac	t t c	t t c	t cg	cag	gcg	at g	cag	cag	cag	1728			
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Tyr	Ile	Met	Ala	Pro	Met	Ser	Ala	Val	Ser	Ala	Thr	Ala	Thr	Ala	Val				
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<211> 2088

<212> DNA

<213> Oryza sativa

<220>

<221> CDS

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<400> 87

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5312WOPCT_SEQ_LI STI NG TXT

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Thr	Thr	Thr	Thr	Al a	G y	Asp	Ser	Ser	Thr	G y	Asp	Val	Cys	Phe	Asn	
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caa	cag	cat	cat	cac	ggc	gga	aag	ggc	ggt	gtt	atc	cca	agc	tct	gct	288
G n	G n	His	His	His	G y	G y	Lys	G y	G y	Val	I le	Pro	Ser	Ser	Al a	
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Al a	Al a	Cys	Tyr	Al a	Ser	Ser	G y	Ser	Ser	Val	G y	Tyr	Leu	Tyr	Pro	
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ccg	cct	tca	tcc	tcg	tca	ctt	cag	ttt	gca	gac	agc	gtg	atg	gtc	gca	384
Pro	Pro	Ser	Ser	Ser	Ser	Leu	G n	Phe	Al a	Asp	Ser	Val	Met	Val	Al a	
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Thr	Ser	Ser	Pro	Val	Val	Al a	His	Asp	G y	Val	Ser	G y	G y	G y	Met	
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Leu	Ser	Met	I le	Lys	Asn	Trp	Leu	Arg	Ser	G n	Pro	Al a	Pro	G n	Pro	
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gcg	caa	gca	ctc	agc	ctg	tcg	atg	aac	atg	gct	ggt	act	act	acc	gct	576
Al a	G n	Al a	Leu	Ser	Leu	Ser	Met	Asn	Met	Al a	G y	Thr	Thr	Thr	Al a	
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caa	ggt	gga	ggc	gca	atg	gca	ctt	ctc	gca	ggc	gct	ggc	gaa	aga	gga	624
G n	G y	G y	G y	Al a	Met	Al a	Leu	Leu	Al a	G y	Al a	G y	G u	Arg	G y	
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Ser	G y	Ser	Al a	G y	Al a	Val	Val	Al a	Val	G y	Ser	G u	Ser	G y	G y	
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Ser	G y	Al a	Val	Val	G u	Al a	G y	Al a	Al a	Al a	Al a	Al a	Al a	Arg	Lys	
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agc	gtt	gat	act	ttc	ggc	caa	aga	acg	agc	atc	tac	aga	ggc	gtt	act	864
Ser	Val	Asp	Thr	Phe	G y	G n	Arg	Thr	Ser	I le	Tyr	Arg	G y	Val	Thr	
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5312WOPCT_SEQ_LI STI NG TXT

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tgt Cys 305	cgc Arg	cgc Arg	gag Glu	ggc Gly 310	caa Gln 310	act Thr	agg Arg	aag Lys	gga Gly 315	aga Arg 315	cag Gln	gtc Val	tat Tyr	cta Leu	gga Gly 320	960
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5312WOPCT_SEQ_LI STI NG TXT

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 His Gly Leu Gly Ser Ile Asp Asn Ala Ser Leu Gu His Ser Thr Gly
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 Ser Asn Ser Val Val Tyr Asn Gly Asp Asn Gly Gly Gly Gly Gly Gly
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<212> DNA

<213> Oryza sativa

<400> 88

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5312WOPCT_SEQ_LI STI NG.TXT

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gat gagcgcc act t aagcca cggccacgcg ggt ggcgagc agccacgat c acggcggcga 3720
cggcggggaag caggt gcaga t ggggt acga cagct acct c gt cggcgcag acgcct acgg 3780
cggcgggcggc gccgggagga t gccat cct g ggcat gacg ccggcgt cgg cgccggccgc 3840
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cgacacat aa aaaaaaaact aggt t agcca gct t aat t ag caggggt aaac cact gacaca 3960
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gt gat ggct t gt gaaat t ga acct ggt gt t ct t gccat ga t t t t t t t t c acaagct gcc 4140
at t t t ggggt t caggt t cag aaggat cct g at t at t at t a accagccat a t at at at aga 4200
aggggt agaaa t ggaggt at c ct gct t gt aa at t ggggcaa t ggt agct ag agt t gat gca 4260
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 <212> PRT
 <213> Oryza sativa

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 Thr Thr Thr Thr Ala Gly Asp Ser Ser Thr Gly Asp Val Cys Phe Asn
 35 40 45
 Ile Pro Gln Asp Trp Ser Met Arg Gly Ser Glu Leu Ser Ala Leu Val
 50 55 60
 Ala Glu Pro Lys Leu Glu Asp Phe Leu Gly Gly Ile Ser Phe Ser Glu
 65 70 75 80
 Gln Gln His His His Gly Gly Lys Gly Gly Val Ile Pro Ser Ser Ala
 85 90 95

5312 WOPCT_SEQ LISTING.TXT

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Thr	Ser	Ser	Pro	Val	Val	Ala	His	Asp	Gly	Val	Ser	Gly	Gly	Gly	Met
Val	Ser	Ala	Ala	Ala	Ala	Ala	Ala	Ala	Ser	Gly	Asn	Gly	Gly	Ile	Gly
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Leu	Ser	Met	Ile	Lys	Asn	Trp	Leu	Arg	Ser	Gln	Pro	Ala	Pro	Gln	Pro
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Ala	Gln	Ala	Leu	Ser	Leu	Ser	Met	Asn	Met	Ala	Gly	Thr	Thr	Thr	Ala
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Gln	Gly	Gly	Gly	Ala	Met	Ala	Leu	Leu	Ala	Gly	Ala	Gly	Glu	Arg	Gly
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Arg	Thr	Thr	Pro	Ala	Ser	Glu	Ser	Leu	Ser	Thr	Ser	Ala	His	Gly	Ala
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Thr	Thr	Ala	Thr	Met	Ala	Gly	Gly	Arg	Lys	Glu	Ile	Asn	Glu	Glu	Gly
225					230					235					240
Ser	Gly	Ser	Ala	Gly	Ala	Val	Val	Ala	Val	Gly	Ser	Glu	Ser	Gly	Gly
				245					250					255	
Ser	Gly	Ala	Val	Val	Glu	Ala	Gly	Ala	Ala	Ala	Ala	Ala	Ala	Arg	Lys
			260					265					270		
Ser	Val	Asp	Thr	Phe	Gly	Gln	Arg	Thr	Ser	Ile	Tyr	Arg	Gly	Val	Thr
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Cys	Arg	Arg	Glu	Gly	Gln	Thr	Arg	Lys	Gly	Arg	Gln	Val	Tyr	Leu	Gly
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Gly	Tyr	Asp	Lys	Glu	Glu	Lys	Ala	Ala	Arg	Ala	Tyr	Asp	Leu	Ala	Ala
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Leu	Lys	Tyr	Trp	Gly	Pro	Thr	Thr	Thr	Asn	Phe	Pro	Val	Asn	Asn	
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Val	Ala	Ser	Leu	Arg	Arg	Lys	Ser	Ser	Gly	Phe	Ser	Arg	Gly	Ala	Ser
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Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	His	Gln	His	Gly	Arg	Trp	Gln	Ala
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Arg	Ile	Gly	Arg	Val	Ala	Gly	Asn	Lys	Asp	Leu	Tyr	Leu	Gly	Thr	Phe
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Ser	Thr	Gln	Glu	Glu	Ala	Ala	Glu	Ala	Tyr	Asp	Ile	Ala	Ala	Ile	Lys
			420					425					430		
Phe	Arg	Gly	Leu	Asn	Ala	Val	Thr	Asn	Phe	Asp	Met	Ser	Arg	Tyr	Asp
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Val	Lys	Ser	Ile	Leu	Asp	Ser	Ala	Ala	Leu	Pro	Val	Gly	Thr	Ala	Ala
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Lys	Arg	Leu	Lys	Asp	Ala	Glu	Ala	Ala	Ala	Ala	Tyr	Asp	Val	Gly	Arg
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Ile	Ala	Ser	His	Leu	Gly	Gly	Asp	Gly	Ala	Tyr	Ala	Ala	His	Tyr	Gly
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His	His	His	His	Ser	Ala	Ala	Ala	Ala	Trp	Pro	Thr	Ile	Ala	Phe	Gln
			500					505					510		
Ala	Ala	Ala	Ala	Pro	Pro	Pro	His	Ala	Ala	Gly	Leu	Tyr	His	Pro	Tyr
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Ala	Gln	Pro	Leu	Arg	Gly	Trp	Cys	Lys	Gln	Glu	Gln	Asp	His	Ala	Val
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Ile	Ala	Ala	Ala	His	Ser	Leu	Gln	Asp	Leu	His	His	Leu	Asn	Leu	Gly
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Ala	Ala	Ala	Ala	Ala	His	Asp	Phe	Phe	Ser	Gln	Ala	Met	Gln	Gln	Gln
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His	Gly	Leu	Gly	Ser	Ile	Asp	Asn	Ala	Ser	Leu	Glu	His	Ser	Thr	Gly
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Ser	Asn	Ser	Val	Val	Tyr	Asn	Gly	Asp	Asn	Gly	Gly	Gly	Gly	Gly	Gly
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Tyr	Ile	Met	Ala	Pro	Met	Ser	Ala	Val	Ser	Ala	Thr	Ala	Thr	Ala	Val
		610				615					620				
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5312 WOPCT_SEQ LISTING.TXT

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 Ala Gly Arg Met Pro Ser Trp Ala Met Thr Pro Ala Ser Ala Pro Ala
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 <213> Oryza sativa

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 Gly Ala Gly Ala Asp Pro Val Leu Pro His Pro Pro Leu Gln Glu Trp
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 ggg agc gct tat gag ggc ggc ggc acg gtg gcg gcc gcc ggc ggg gag 144
 Gly Ser Ala Tyr Glu Gly Gly Gly Thr Val Ala Ala Ala Gly Gly Glu
 35 40 45
 gag acg gcg gcg ccg aag ctg gag gac ttc ctg ggc atg cag gtg cag 192
 Glu Thr Ala Ala Pro Lys Leu Glu Asp Phe Leu Gly Met Gln Val Gln
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 cag gag acg gcc gcc gcg gcg gcg ggg cac gcg cgt gga ggc agc tcg 240
 Gln Glu Thr Ala Ala Ala Ala Glu His Gly Arg Gly Gly Ser Ser
 65 70 75 80
 tcg gtg gtt ggg ctg tcc atg atc aag aac tgg cta cgc agc cag ccg 288
 Ser Val Val Gly Leu Ser Met Ile Lys Asn Trp Leu Arg Ser Gln Pro
 85 90 95
 ccg ccc gcg gtg gtt ggg gga gaa gac gct atg atg gcg ctg gcg gtg 336
 Pro Pro Ala Val Val Gly Gly Glu Asp Ala Met Met Ala Leu Ala Val
 100 105 110
 tcg acg tcg gcg tcg ccg ccg gtg gac gcg acg gtg ccg gcc tgc att 384
 Ser Thr Ser Ala Ser Pro Pro Val Asp Ala Thr Val Pro Ala Cys Ile
 115 120 125
 tcg ccg gat ggg atg ggg tcg aag gcg gcc gac ggc ggc ggc gcg gcc 432
 Ser Pro Asp Gly Met Gly Ser Lys Ala Ala Asp Gly Gly Gly Ala Ala
 130 135 140
 gag gcg gcg gcg gcg gcg gcg gcg cag agg atg aag gcg gcc atg gac 480
 Glu Ala Ala Ala Ala Ala Ala Ala Gln Arg Met Lys Ala Ala Met Asp
 145 150 155 160
 acg ttc ggg cag cgg acg tcc atc tac ccg ggt gtg acc aag cac agg 528
 Thr Phe Gly Gln Arg Thr Ser Ile Tyr Arg Gly Val Thr Lys His Arg
 165 170 175
 tgg aca gga agg tat gaa gcc cat ctg tgg gat aac agc tgc aga aga 576
 Trp Thr Gly Arg Tyr Glu Ala His Leu Trp Asp Asn Ser Cys Arg Arg
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5312WOPCT_SEQ_LI STI NG TXT

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195	200	205	
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225	230	235	240
gag ttg gat gaa atg aag cac atg aat agg cag gaa ttt gtt gca tcc G u Leu Asp G u Met Lys Hi s Met Asn Arg G n G u Phe Val Al a Ser	768		
245	250	255	
ctt aga aga aaa agc agt gga ttt tca cgt ggt gct tcc ata tat cgt Leu Arg Arg Lys Ser Ser G y Phe Ser Arg G y Al a Ser Ile Tyr Arg	816		
260	265	270	
ggt gtt aca aga cac cat cag cat gga agg tgg caa gca agg ata gga G y Val Thr Arg Hi s Hi s G n Hi s G y Arg Trp G n Al a Arg Ile G y	864		
275	280	285	
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290	295	300	
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305	310	315	320
ctc aat gct gtg aca aac ttt gac atg agc cgg tac gat gtc aag agc Leu Asn Al a Val Thr Asn Phe Asp Met Ser Arg Tyr Asp Val Lys Ser	1008		
325	330	335	
atc att gaa agc agc aat ct ccca att ggt act gga acc acc cgg cga Ile Ile G u Ser Ser Asn Leu Pro Ile G y Thr G y Thr Thr Arg Arg	1056		
340	345	350	
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370	375	380	
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385	390	395	400
agc atg cag ccg atc ccc tcg cag tac gcc aac ggc cag ccc agg gca Ser Met G n Pro Ile Pro Ser G n Tyr Al a Asn G y G n Pro Arg Al a	1248		
405	410	415	
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420	425	430	
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435	440	445	
ttc cag caa tct gat gtt cca gac gtc aca ggt ttc gtt gat gcg cct Phe G n G n Ser Asp Val Pro Asp Val Thr G y Phe Val Asp Al a Pro	1392		
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5312WOPCT_SEQ_LI STI NG TXT

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t t t Phe	cat His	ggg Gly	ct c Leu	ccg Pro 485	ggg Gly	gga Gly	at c Ile	agc Ser	t at Tyr 490	gct Ala	at g Met	ccg Pro	gt t Val	gcg Ala 495	aca Thr	1488
gcg Ala	gt g Val	gac Asp	caa Gln 500	ggg Gly	cag Gln	ggc Gly	at c Ile	cat His 505	ggc Gly	t at Tyr	gga Gly	gaa Glu	gat Asp 510	ggg Gly	gt g Val	1536
gca Ala	ggc Gly	att Ile 515	gac Asp	acc Thr	aca Thr	cat His	gac Asp 520	ct g Leu	t at Tyr	ggc Gly	agc Ser	cgt Arg 525	aat Asn	gt g Val	t ac Tyr	1584
t ac Tyr	ct t Leu 530	t c c Ser	gag Glu	ggg Gly	t c g Ser	ct t Leu 535	ct t Leu	gcc Ala	gat Asp	gt c Val	gaa Glu 540	aaa Lys	gaa Glu	ggc Gly	gac Asp	1632
t at Tyr 545	ggc Gly	caa Gln	t ct Ser	gt g Val	ggg Gly 550	ggc Gly	aac Asn	agc Ser	t gg Trp	gt t Val 555	t t g Leu	ccg Pro	aca Thr	ccg Pro	t ag	1680

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 <212> PRT
 <213> Oryza sativa

<400> 91

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Gly	Ser	Ala 35	Tyr	Glu	Gly	Gly	Gly 40	Thr	Val	Ala	Ala	Ala 45	Gly	Gly	Glu
Glu	Thr 50	Ala	Ala	Pro	Lys	Leu 55	Glu	Asp	Phe	Leu	Gly 60	Met	Gln	Val	Gln
Gln 65	Glu	Thr	Ala	Ala 70	Ala	Ala	Ala	Gly	His 75	Gly	Arg	Gly	Gly	Ser	Ser 80
Ser	Val	Val	Gly	Leu 85	Ser	Met	Ile	Lys	Asn 90	Trp	Leu	Arg	Ser	Gln 95	Pro
Pro	Pro	Ala 100	Val	Val	Gly	Gly	Glu	Asp 105	Ala	Met	Met	Ala	Leu 110	Ala	Val
Ser	Thr	Ser 115	Ala	Ser	Pro	Pro	Val	Asp 120	Ala	Thr	Val	Pro 125	Ala	Cys	Ile
Ser	Pro 130	Asp	Gly	Met	Gly	Ser 135	Lys	Ala	Ala	Asp	Gly 140	Gly	Gly	Ala	Ala
Glu 145	Ala	Ala	Ala	Ala 150	Ala	Ala	Ala	Gln	Arg	Met 155	Lys	Ala	Ala	Met	Asp 160
Thr	Phe	Gly	Gln	Arg 165	Thr	Ser	Ile	Tyr	Arg 170	Gly	Val	Thr	Lys	His 175	Arg
Trp	Thr	Gly 180	Arg	Tyr	Glu	Ala	His 185	Leu	Trp	Asp	Asn	Ser	Cys 190	Arg	Arg
Glu	Gly	Gln 195	Thr	Arg	Lys	Gly	Arg 200	Gln	Val	Tyr	Leu	Gly 205	Gly	Tyr	Asp
Lys	Glu 210	Glu	Lys	Ala	Ala	Arg 215	Ala	Tyr	Asp	Leu	Ala 220	Ala	Leu	Lys	Tyr
Trp 225	Gly	Thr	Thr	Thr	Thr	Thr	Asn	Phe	Pro	Val 235	Ser	Asn	Tyr	Glu 240	Lys
Glu	Leu	Asp	Glu	Met 245	Lys	His	Met	Asn	Arg 250	Gln	Glu	Phe	Val	Ala 255	Ser
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Gly	Val	Thr 275	Arg	His	His	Gln	His 280	Gly	Arg	Trp	Gln	Ala 285	Arg	Ile	Gly

Arg	Val 290	Ala	Gly	Asn	Lys	Asp 295	Leu	Tyr	Leu	Gly	Thr 300	Phe	Gly	Thr	Gln
Glu 305	Glu	Ala	Ala	Glu	Ala 310	Tyr	Asp	Ile	Ala	Ala 315	Ile	Lys	Phe	Arg	Gly 320
Leu	Asn	Ala	Val	Thr 325	Asn	Phe	Asp	Met	Ser 330	Arg	Tyr	Asp	Val	Lys 335	Ser
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Leu	Lys	Asp 355	Ser	Ser	Asp	His	Thr 360	Asp	Asn	Val	Met	Asp 365	Ile	Asn	Val
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Gly 385	Asn	Tyr	Gly	Ser	Gln 390	His	Tyr	Gly	Tyr	Asn 395	Gly	Trp	Ser	Pro	Ile 400
Ser	Met	Gln	Pro	Ile 405	Pro	Ser	Gln	Tyr	Ala 410	Asn	Gly	Gln	Pro	Arg 415	Ala
Trp	Leu	Lys	Gln 420	Glu	Gln	Asp	Ser	Ser 425	Val	Val	Thr	Ala	Ala 430	Gln	Asn
Leu	His	Asn 435	Leu	His	His	Phe	Ser 440	Ser	Leu	Gly	Tyr	Thr 445	His	Asn	Phe
Phe	Gln 450	Gln	Ser	Asp	Val	Pro 455	Asp	Val	Thr	Gly	Phe 460	Val	Asp	Ala	Pro
Ser 465	Arg	Ser	Ser	Asp	Ser 470	Tyr	Ser	Phe	Arg	Tyr 475	Asn	Gly	Thr	Asn	Gly 480
Phe	His	Gly	Leu	Pro 485	Gly	Gly	Ile	Ser	Tyr 490	Ala	Met	Pro	Val	Ala 495	Thr
Ala	Val	Asp	Gln 500	Gly	Gln	Gly	Ile	His 505	Gly	Tyr	Gly	Glu	Asp 510	Gly	Val
Ala	Gly	Ile 515	Asp	Thr	Thr	His	Asp 520	Leu	Tyr	Gly	Ser	Arg 525	Asn	Val	Tyr
Tyr	Leu 530	Ser	Glu	Gly	Ser	Leu 535	Leu	Ala	Asp	Val	Glu 540	Lys	Glu	Gly	Asp
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<210> 92
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 $\langle 222 \rangle$ (1) ... (2112)

[illegible]

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Glu	Pro	Lys	Leu	Glu	Asp	Phe	Leu	Gly	Gly	Asn	Ser	Phe	Val	Ser	Glu		
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Gln	Asp	His	His	Ala	Ala	Gly	Gly	Phe	Leu	Phe	Ser	Gly	Val	Pro	Met		
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gcc	agc	agc	acc	aac	agc	aac	agc	ggg	agc	aac	act	at g	gag	ct c	t cc	480	
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165				170				175									
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Gln	Pro	Gln	Gln	Gln	Gln	Pro	Ala	Ala	Ala	Ala	Ala	Ala	Ala	Ala	Gln		
180				185				190									
cag	gcg	cac	gag	gcg	gcg	gag	at g	agc	acc	gac	gcg	agc	gcg	agc	agc	624	
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Phe	Gly	Cys	Ser	Ser	Asp	Ala	Met	Gly	Arg	Ser	Asn	Asn	Gly	Gly	Ala		
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260				265				270									
cgg	gcc	agc	ggc	gcc	at g	gat	t cg	ccg	ggc	ggt	ggc	gcg	at a	gag	gcc	864	
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275				280				285									
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Val	Pro	Arg	Lys	Ser	Ile	Asp	Thr	Phe	Gly	Gln	Arg	Thr	Ser	Ile	Tyr		
290				295				300									
cga	ggt	gt a	aca	agg	cat	aga	t gg	aca	ggg	cga	t at	gag	gct	cat	ct c	960	
Arg	Gly	Val	Thr	Arg	His	Arg	Trp	Thr	Gly	Arg	Tyr	Glu	Ala	His	Leu		
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Trp	Asp	Asn	Ser	Cys	Arg	Arg	Glu	Gly	Gln	Ser	Arg	Lys	Gly	Arg	Gln		
325				330				335									
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Val	Tyr	Leu	Gly	Gly	Tyr	Asp	Lys	Glu	Asp	Lys	Ala	Ala	Arg	Ala	Tyr		
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5312WOPCT_SEQ_LI STI NG TXT

355	360	365	
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agg cag gag t at att gca t ac ct a aga agg aat agc agt gga t t t t ct Arg Gn Gu Tyr Ile Ala Tyr Leu Arg Arg Asn Ser Ser Gly Phe Ser 385 390 395 400			1200
cgt ggt gca t cg aaa t at cgt ggt gt a acc agg cac cat cag cat ggg Arg Gy Ala Ser Lys Tyr Arg Gy Val Thr Arg His His Gn His Gy 405 410 415			1248
aga t gg caa gca agg at a ggg agg gt t gca gga aac aag gac ct c t ac Arg Trp Gn Ala Arg Ile Gy Arg Val Ala Gy Asn Lys Asp Leu Tyr 420 425 430			1296
t t a ggc acc t t c agc acc gag gag gag gcg gcg gag gcg t ac gac at c Leu Gy Thr Phe Ser Thr Glu Glu Glu Ala Ala Glu Ala Tyr Asp Ile 435 440 445			1344
gcg gcg at c aag t t c cgg ggg ct c aac gcc gt c acc aac t t t gac at g Ala Ala Ile Lys Phe Arg Gy Leu Asn Ala Val Thr Asn Phe Asp Met 450 455 460			1392
agc cgc t ac gac gt c aag agc at c ct g gag agc agc acg ct g ccg gt g Ser Arg Tyr Asp Val Lys Ser Ile Leu Glu Ser Ser Thr Leu Pro Val 465 470 475 480			1440
ggc ggc gcg gcg agg cgg ct g aag gag gcg gcg gac cac gcg gag gcg Gy Gy Ala Ala Arg Arg Leu Lys Glu Ala Ala Asp His Ala Glu Ala 485 490 495			1488
gcc gcc gcc acc at c t gg cgc gcc gcc gac at g gac gcc gcc gcc gt c Ala Gy Ala Thr Ile Trp Arg Ala Ala Asp Met Asp Gy Ala Gy Gy Val 500 505 510			1536
at c t cc ggc ct g gcc gac gt c ggg at g ggc gcc t ac gcc gcc t cg t ac Ile Ser Gy Leu Ala Asp Val Gy Met Gy Ala Tyr Ala Ala Ser Tyr 515 520 525			1584
cac cac cac cac cac cac ggc t gg ccg acc at c gcg t t c cag cag ccg His His His His His His Gy Trp Pro Thr Ile Ala Phe Gn Gn Pro 530 535 540			1632
ccg ccg ct c gcc gt g cac t ac ccg t ac ggc cag gcg ccg gcg gcg ccg Pro Pro Leu Ala Val His Tyr Pro Tyr Gy Gn Ala Pro Ala Ala Pro 545 550 555 560			1680
t cg cgc ggg t gg t gc aag ccc gag cag gac gcc gcc gt c gct gcc gcc Ser Arg Gy Trp Cys Lys Pro Glu Gn Asp Ala Ala Val Ala Ala Ala 565 570 575			1728
gcg cac agc ct c cag gac ct c cag cag ct g cac ct c ggc agc gcc gcc Ala His Ser Leu Gn Asp Leu Gn Gn Leu His Leu Gy Ser Ala Ala 580 585 590			1776
gcc cac aac t t c t t c cag gcg t cg t cg agc t cg acg gt c t ac aac ggc Ala His Asn Phe Phe Gn Ala Ser Ser Ser Ser Thr Val Tyr Asn Gy 595 600 605			1824
ggc ggc ggc ggg t ac cag ggc ct c ggt ggc aac gcc t t c t t g at g ccg Gy Gy Gy Gy Tyr Gn Gy Leu Gy Gy Asn Ala Phe Leu Met Pro 610 615 620			1872
gcg agc acc gt c gt g gcc gac cag ggg cac agc agc acg gcc acc aac Ala Ser Thr Val Val Ala Asp Gn Gy His Ser Ser Thr Ala Thr Asn 625 630 635 640			1920

5312WOPCT_SEQ_LI STI NG TXT

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ggg	tac	gac	gcc	atg	gcg	atg	gcg	agc	ggc	gcc	gcc	ggc	ggc	ggg	tac	2016					
Gly	Tyr	Asp	Ala	Met	Ala	Met	Ala	Ser	Gly	Ala	Ala	Gly	Gly	Gly	Tyr						
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cag	ctg	tcg	cag	ggc	tcg	gcg	tcg	acg	gtg	agc	atc	gcg	agg	gcg	aac	2064					
Gln	Leu	Ser	Gln	Gly	Ser	Ala	Ser	Thr	Val	Ser	Ile	Ala	Arg	Ala	Asn						
			675				680					685									
ggc	tac	tcg	gcc	aac	tgg	agc	tcg	cct	ttc	aat	ggc	gcc	atg	gga	tga	2112					
Gly	Tyr	Ser	Ala	Asn	Trp	Ser	Ser	Pro	Phe	Asn	Gly	Ala	Met	Gly							
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 <212> PRT
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<400> 93

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			20					25					30		
Ser	Gly	Ala	Gly	Asp	Phe	Tyr	Gly	Leu	Pro	Thr	Ser	Gln	Pro	Thr	Ala
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Gly	Ile	Met	Glu	Ala	Phe	Asn	Arg	Gly	Ala	Gln	Glu	Ala	Gln	Asp	Trp
65					70				75					80	
Asn	Met	Arg	Gly	Leu	Asp	Tyr	Asn	Gly	Gly	Ala	Ser	Glu	Leu	Ser	Met
			85					90					95		
Leu	Val	Gly	Ser	Ser	Gly	Gly	Lys	Arg	Ala	Ala	Ala	Val	Glu	Glu	Thr
			100				105						110		
Glu	Pro	Lys	Leu	Glu	Asp	Phe	Leu	Gly	Gly	Asn	Ser	Phe	Val	Ser	Glu
		115					120					125			
Gln	Asp	His	His	Ala	Ala	Gly	Gly	Phe	Leu	Phe	Ser	Gly	Val	Pro	Met
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Ala	Ser	Ser	Thr	Asn	Ser	Asn	Ser	Gly	Ser	Asn	Thr	Met	Glu	Leu	Ser
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			165						170					175	
Gln	Pro	Gln	Gln	Gln	Gln	Pro	Ala	Ala	Ala	Ala	Ala	Ala	Ala	Gln	Gln
			180					185					190		
Gln	Ala	His	Glu	Ala	Ala	Glu	Met	Ser	Thr	Asp	Ala	Ser	Ala	Ser	Ser
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Phe	Gly	Cys	Ser	Ser	Asp	Ala	Met	Gly	Arg	Ser	Asn	Asn	Gly	Gly	Ala
	210					215					220				
Val	Ser	Ala	Ala	Ala	Gly	Gly	Thr	Ser	Ser	Gln	Ser	Leu	Ala	Leu	Ser
225					230					235				240	
Met	Ser	Thr	Gly	Ser	His	Ser	His	Leu	Pro	Ile	Val	Val	Ala	Gly	Gly
			245						250					255	
Gly	Asn	Ala	Ser	Gly	Gly	Ala	Ala	Glu	Ser	Thr	Ser	Ser	Glu	Asn	Lys
			260					265					270		
Arg	Ala	Ser	Gly	Ala	Met	Asp	Ser	Pro	Gly	Gly	Gly	Ala	Ile	Glu	Ala
		275					280					285			
Val	Pro	Arg	Lys	Ser	Ile	Asp	Thr	Phe	Gly	Gln	Arg	Thr	Ser	Ile	Tyr
	290					295					300				
Arg	Gly	Val	Thr	Arg	His	Arg	Trp	Thr	Gly	Arg	Tyr	Glu	Ala	His	Leu
305					310					315				320	
Trp	Asp	Asn	Ser	Cys	Arg	Arg	Glu	Gly	Gln	Ser	Arg	Lys	Gly	Arg	Gln
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5312 WOPCT_SEQ_LI STING.TXT

Val Tyr Leu Gly Gly Tyr Asp Lys Glu Asp Lys Ala Ala Arg Ala Tyr
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 Asp Leu Ala Ala Leu Lys Tyr Trp Gly Thr Thr Thr Thr Thr Asn Phe
 355 360 365
 Pro Ile Ser Asn Tyr Glu Lys Glu Leu Asp Glu Met Lys His Met Thr
 370 375 380
 Arg Gln Glu Tyr Ile Ala Tyr Leu Arg Arg Asn Ser Ser Gly Phe Ser
 385 390 395 400
 Arg Gly Ala Ser Lys Tyr Arg Gly Val Thr Arg His His Gln His Gly
 405 410 415
 Arg Trp Gln Ala Arg Ile Gly Arg Val Ala Gly Asn Lys Asp Leu Tyr
 420 425 430
 Leu Gly Thr Phe Ser Thr Glu Glu Glu Ala Ala Glu Ala Tyr Asp Ile
 435 440 445
 Ala Ala Ile Lys Phe Arg Gly Leu Asn Ala Val Thr Asn Phe Asp Met
 450 455 460
 Ser Arg Tyr Asp Val Lys Ser Ile Leu Glu Ser Ser Thr Leu Pro Val
 465 470 475 480
 Gly Gly Ala Ala Arg Arg Leu Lys Glu Ala Ala Asp His Ala Glu Ala
 485 490 495
 Ala Gly Ala Thr Ile Trp Arg Ala Ala Asp Met Asp Gly Ala Gly Val
 500 505 510
 Ile Ser Gly Leu Ala Asp Val Gly Met Gly Ala Tyr Ala Ala Ser Tyr
 515 520 525
 His His His His His His Gly Trp Pro Thr Ile Ala Phe Gln Gln Pro
 530 535 540
 Pro Pro Leu Ala Val His Tyr Pro Tyr Gly Gln Ala Pro Ala Ala Pro
 545 550 555 560
 Ser Arg Gly Trp Cys Lys Pro Glu Gln Asp Ala Ala Val Ala Ala Ala
 565 570 575
 Ala His Ser Leu Gln Asp Leu Gln Leu His Leu Gly Ser Ala Ala
 580 585 590
 Ala His Asn Phe Phe Gln Ala Ser Ser Ser Ser Thr Val Tyr Asn Gly
 595 600 605
 Gly Gly Gly Gly Tyr Gln Gly Leu Gly Gly Asn Ala Phe Leu Met Pro
 610 615 620
 Ala Ser Thr Val Val Ala Asp Gln Gly His Ser Ser Thr Ala Thr Asn
 625 630 635 640
 His Gly Asn Thr Cys Ser Tyr Gly Asn Glu Glu Gln Gly Lys Leu Ile
 645 650 655
 Gly Tyr Asp Ala Met Ala Met Ala Ser Gly Ala Ala Gly Gly Tyr
 660 665 670
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<400> 94
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 aac cca cag cat cac cag aac ggc tcg ccg tct gcc gcc ggc gac gcc 96
 Asn Pro Gln His His Gln Asn Gly Ser Pro Ser Ala Ala Gly Asp Ala
 20 25 30
 gcc atc gac atc tcc ggc tca ggc gac ttc tat ggt ctg cca acg ccg 144
 Ala Ile Asp Ile Ser Gly Ser Gly Asp Phe Tyr Gly Leu Pro Thr Pro

5312WOPCT_SEQ_LI STI NG.TXT

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agg ggt t t g gac t ac ggc ggc ggc t cc t cc gac ct c t cg at g ct c gt c 288	Arg Gly Leu Asp Tyr 85	Gly Gly Gly Ser Ser 90	Asp Leu Ser Met Leu 95
ggc t cg agc ggc ggc ggg agg agg acg gt g gcc ggc gac ggc gt c ggc 336	Gly Ser Ser Gly 100	Gly Gly Arg Arg Thr 105	Ala Gly Asp Gly Val Gly
gag gcg ccg aag ct g gag aac t t c ct c gac ggc aac t ca t t c t cc gac 384	Gu Ala Pro Lys Leu Gu Asn Phe 120	Leu Asp Gly Asn Ser 125	Phe Ser Asp
gt g cac ggc caa gcc gcc ggc ggg t ac ct c t ac t cc gga agc gct gt c 432	Val His 130	Gly Gn Ala Ala Gly Gly Tyr Leu Tyr Ser 140	Gly Ser Ala Val
ggc ggc gcc ggt ggt t ac agt aac ggc gga t gc ggc ggc gga acc at a 480	Gly Gly Ala Gly Gly Tyr 150	Ser Asn Gly Gly Cys 155	Gly Gly Gly Thr Ile 160
gag ct g t cc at g at c aag acg t gg ct c cgg agc aac cag t cg cag cag 528	Gu Leu Ser Met Ile 165	Lys Thr Trp Leu Arg 170	Ser Asn Gn Ser Gn Gn
cag cca t cg ccg ccg cag cac gct gat cag ggc at g agc acc gac gcc 576	Gn Pro Ser Pro Pro Gn His Ala Asp 185	Gn Gly Met Ser Thr 190	Asp Ala
agc gcg agc agc t ac gcg t gc t cc gac gt g ct g gt g ggg agc t gc ggc 624	Ser Ala Ser 195	Ser Tyr Ala Cys Ser 200	Asp Val Leu Val Gly Ser Cys Gly
ggc ggc ggc gcc ggg ggc acg gcg agc t cg cat ggg cag ggc ct g gcg 672	Gly Gly Gly Ala Gly Gly Thr 215	Ala Ser Ser His 220	Gn Gn Gly Leu Ala
ct g t cg at g agc acg ggg t cg gt g gcc gcc gcc gga ggg ggc ggc gcc 720	Leu Ser Met Ser Thr 230	Ser Val Ala Ala 235	Gly Gly Gly Gly Ala 240
gt c gt c gcg gcc gag agc t cg t cg t cg gag aac aag cgg gt g gat t cg 768	Val Val Ala Ala Gu Ser Ser Ser 245	Gu Asn Lys Arg Val 255	Asp Ser
ccg ggc ggc gcc gt g gac ggc gcc gt c ccg agg aaa t cc at c gac acc 816	Pro Gly Gly Ala Val 260	Asp Gly Ala Val 265	Pro Arg Lys Ser Ile Asp Thr
t t c ggg caa agg acg t ct at a t ac cga ggt gt a aca agg cat aga t gg 864	Phe Gly Gn Arg Thr Ser Ile Tyr 280	Arg Gly Val Thr 285	His Arg Trp
aca gga aga t at gaa gct cat ct g t gg gat aat agc t gt agg aga gaa 912	Thr Gly Arg Tyr Gu Ala His 295	Leu Trp Asp Asn Ser 300	Cys Arg Arg Gu
ggc caa agt cgc aag ggg aga cag gt t t at t t g ggc ggt t at gac aaa 960	Gly Gn Ser Arg Lys Gly Arg Gn Val Tyr Leu Gly Gly Tyr Asp Lys		

5312WOPCT_SEQ_LI STI NG TXT

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ggc G y	acg T hr	acc T hr	aca T hr 340	aca T hr	aca T hr	aat A sn	t t c P he	cca P ro 345	at g M et	agt S er	aat A sn	t at T yr	gaa G u 350	aag L ys	gag G u	1056				
ct a L eu	gag G u	gaa G u 355	at g M et	aaa L ys	cac H i s	at g M et	acc T hr 360	agg A rg	cag G n	gag G u	t ac T yr	at t I l e 365	gca A l a	cat H i s	ct t L eu	1104				
aga A rg	agg A rg 370	aat A sn	agc S er	agt S er	gga G y	t t t P he 375	t ct S er	cgt A rg	ggt G y	gca A l a	t cc S er 380	aaa L ys	t at T yr	cgt A rg	ggt G y	1152				
gt t V al 385	act T hr	agg A rg	cat H i s	cat H i s	cag G n 390	cat H i s	ggg G y	aga A rg	t gg T rp	cag G n 395	gca A l a	agg A rg	at a I l e	ggg G y	cga A rg 400	1200				
gt t V al	gca A l a	ggc G y	aac A sn	aag L ys 405	gat A sp	at c I l e	t ac T yr	ct a L eu	ggc G y 410	acc T hr	t t c P he	agc S er	acc T hr	gag G u 415	gag G u	1248				
gag G u	gcc A l a	gcc A l a	gag G u 420	gcg A l a	t ac T yr	gac A sp	at c I l e	gcc A l a 425	gcc A l a	at c I l e	aag L ys	t t c P he	cgc A rg 430	ggg G y	ct c L eu	1296				
aac A sn	gcc A l a	gt c V al 435	acc T hr	aac A sn	t t c P he	gac A sp	at g M et 440	agc S er	cgg A rg	t ac T yr	gac A sp	gt c V al 445	aag L ys	agc S er	at c I l e	1344				
ct g L eu	gac A sp 450	agc S er	agc S er	acg T hr	ct g L eu	ccg P ro 455	gt c V al	ggc G y	ggc G y	gcg A l a	gcg A l a 460	cgg A rg	cgg A rg	ct c L eu	aag L ys	1392				
gag G u 465	gcg A l a	gag G u	gt c V al	gcc A l a	gcc A l a 470	gcc A l a	gcc A l a	gcg A l a	ggc G y	ggc G y 475	ggc G y	gt g V al	at c I l e	gt c V al	t cc S er 480	1440				
cac H i s	ct g L eu	gcc A l a	gac A sp	ggc G y 485	ggt G y	gt g V al	ggt G y	ggg G y	t ac T yr 490	t ac T yr	t ac T yr	ggg G y	t gc C ys	ggc G y 495	ccg P ro	1488				
acc T hr	at c I l e	gcg A l a	t t c P he 500	ggc G y	ggc G y	ggc G y	ggc G y	cag G n 505	cag G n	ccg P ro	gcg A l a	ccg P ro	ct c L eu 510	gcc A l a	gt g V al	1536				
cac H i s	t ac T yr	ccg P ro 515	t cg S er	t ac T yr	ggc G y	cag G n	gcc A l a 520	agc S er	ggg G y	t gg T rp	t gc C ys	aag L ys 525	ccg P ro	gag G u	cag G n	1584				
gac A sp	gcg A l a 530	gt g V al	at c I l e	gcg A l a	gcc A l a	ggg G y 535	cac H i s	t gc C ys	gcg A l a	acg T hr	gac A sp 540	ct c L eu	cag G n	cac H i s	ct g L eu	1632				
cac H i s 545	ct c L eu	ggg G y	agc S er	ggc G y	ggc G y 550	gcc A l a	gcc A l a	gcc A l a	acc T hr	cac H i s 555	aac A sn	t t c P he	t t c P he	cag G n 560	cag G n 560	1680				
ccg P ro	gcg A l a	t ca S er	agc S er	t cg S er 565	gcc A l a	gt c V al	t ac T yr	ggc G y	aac A sn 570	ggc G y	ggc G y	ggc G y	ggc G y	ggc G y 575	ggc G y	1728				
aac A sn	gcg A l a	t t c P he	at g M et	at g M et	ccg P ro	at g M et	ggc G y	gcc A l a	gt g V al	gt g V al	gcc A l a	gcc A l a	gcc A l a	gat A sp	cac H i s	1776				

5312 WOPCT_SEQ_LI STI NG TXT

580

585

590

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gt c	gt g	ggg	t ac	gac	ggc	gt c	gt c	gac	ccg	t ac	gcg	gcc	at g	aga	agc	1872
Val	Val	G y	Tyr	Asp	G y	Val	Val	Asp	Pro	Tyr	Al a	Al a	Met	Arg	Ser	
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gcg	t ac	gag	ct c	t cg	cag	ggc	t cg	t cg	t cg	t cg	t cg	gt g	agc	gt c	gcg	1920
Al a	Tyr	G u	Leu	Ser	G n	G y	Ser	Ser	Ser	Ser	Ser	Val	Ser	Val	Al a	
625					630					635					640	
aag	gcg	gcg	aac	ggg	t ac	ccg	gac	aac	t gg	agc	t cg	ccg	t t c	aac	ggc	1968
Lys	Al a	Al a	Asn	G y	Tyr	Pro	Asp	Asn	Tr p	Ser	Ser	Pro	Phe	Asn	G y	
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at g	gga	t ga														1977
Met	G y															

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<400> 95

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		20					25					30			
Al a	I le	Asp	I le	Ser	G y	Ser	G y	Asp	Phe	Tyr	G y	Leu	Pro	Thr	Pro
	35						40					45			
Asp	Al a	Hi s	Hi s	I le	G y	Met	Al a	G y	G u	Asp	Al a	Pro	Tyr	G y	Val
	50					55					60				
Met	Asp	Al a	Phe	Asn	Arg	G y	Thr	Hi s	G u	Thr	G n	Asp	Tr p	Al a	Met
65				70						75					80
Arg	G y	Leu	Asp	Tyr	G y	G y	G y	Ser	Ser	Asp	Leu	Ser	Met	Leu	Val
			85						90					95	
G y	Ser	Ser	G y	G y	Arg	Arg	Thr	Val	Al a	G y	Asp	G y	Val	G y	
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G u	Al a	Pro	Lys	Leu	G u	Asn	Phe	Leu	Asp	G y	Asn	Ser	Phe	Ser	Asp
		115					120					125			
Val	Hi s	G y	G n	Al a	Al a	G y	G y	Tyr	Leu	Tyr	Ser	G y	Ser	Al a	Val
	130					135					140				
G y	G y	Al a	G y	G y	Tyr	Ser	Asn	G y	G y	Cys	G y	G y	G y	Thr	I le
145					150					155					160
G u	Leu	Ser	Met	I le	Lys	Thr	Tr p	Leu	Arg	Ser	Asn	G n	Ser	G n	G n
			165						170					175	
G n	Pro	Ser	Pro	Pro	G n	Hi s	Al a	Asp	G n	G y	Met	Ser	Thr	Asp	Al a
			180					185					190		
Ser	Al a	Ser	Ser	Tyr	Al a	Cys	Ser	Asp	Val	Leu	Val	G y	Ser	Cys	G y
		195					200					205			
G y	G y	G y	Al a	G y	G y	Thr	Al a	Ser	Ser	Hi s	G y	G n	G y	Leu	Al a
	210					215					220				
Leu	Ser	Met	Ser	Thr	G y	Ser	Val	Al a	Al a	Al a	G y	G y	G y	G y	Al a
225					230					235					240
Val	Val	Al a	Al a	G u	Ser	Ser	Ser	Ser	G u	Asn	Lys	Arg	Val	Asp	Ser
			245						250					255	
Pro	G y	G y	Al a	Val	Asp	G y	Al a	Val	Pro	Arg	Lys	Ser	I le	Asp	Thr
			260					265					270		
Phe	G y	G n	Arg	Thr	Ser	I le	Tyr	Arg	G y	Val	Thr	Arg	Hi s	Arg	Tr p
	275						280					285			
Thr	G y	Arg	Tyr	G u	Al a	Hi s	Leu	Tr p	Asp	Asn	Ser	Cys	Arg	Arg	G u
	290					295					300				
G y	G n	Ser	Arg	Lys	G y	Arg	G n	Val	Tyr	Leu	G y	G y	Tyr	Asp	Lys

5312WOPCT_SEQ LI STI NG TXT

305 310 315 320
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 G y Thr Thr Thr 325 Thr Asn Phe Pro 330 Met Ser Asn Tyr G u Lys G u
 Leu G u G u Met 340 Lys H i s Met Thr 345 Arg G n G u Tyr I l e Al a H i s Leu
 Arg Arg Asn Ser Ser G y Phe Ser Arg G y Al a Ser 365 Lys Tyr Arg G y
 Val 370 Thr Arg H i s H i s G n 375 H i s G y Arg Tr p G n Al a Arg I l e G y Arg
 385 Val Al a G y Asn Lys Asp I l e Tyr Leu G y Thr Phe Ser Thr G u G u
 G u Al a Al a G u Al a Tyr Asp I l e Al a Al a I l e Lys Phe Arg G y Leu
 Asn Al a Val 420 Thr Asn Phe Asp Met 425 Ser Arg Tyr Asp Val 430 Lys Ser I l e
 Leu Asp Ser Ser Thr Leu Pro Val 440 G y G y Al a Al a Arg Arg Leu Lys
 G u Al a G u Val Al a Al a Al a Al a G y G y G y Val I l e Val Ser
 465 H i s Leu Al a Asp G y G y Val G y G y Tyr Tyr Tyr G y Cys G y Pro
 Thr I l e Al a Phe 485 G y G y G y G y G n G n Pro Al a Pro Leu Al a Val
 H i s Tyr Pro 500 Ser Tyr G y G n Al a 505 Ser G y Tr p Cys Lys Pro G u G n
 Asp Al a Val I l e Al a Al a G y H i s Cys Al a Thr Asp Leu G n H i s Leu
 H i s Leu G y Ser G y G y Al a Al a Al a Thr H i s Asn Phe Phe G n G n
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 Asn Al a Phe Met Met Pro Met G y Al a Val Val Al a Al a Al a Asp H i s
 G y G y G n Ser Ser Al a Tyr G y G y G y Asp G u Ser G y Arg Leu
 Val Val 595 G y Tyr Asp G y Val Val Asp Pro Tyr Al a Al a Met Arg Ser
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 ct g ccg ccc acc cag acg gac tcc acc ct c at c tct gcc gcc acc acc 96
 Leu Pro Pro Thr 20 G n Thr Asp Ser Thr 25 Leu I l e Ser Al a Al a Thr Thr
 gac gat gt c tcc ggc gat gt c tgc ttc aac at c ccc caa gat t gg agc 144
 Asp Asp Val 35 Ser G y Asp Val Cys 40 Phe Asn I l e Pro G n Asp Tr p Ser
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gac Asp	t t c Phe	ct c Leu	ggc Gly	gga Gly	at c Ile	t cc Ser	t t c Phe	t cc Ser	gag Glu	cag Gln	cac His	cac His	aag Lys	gcc Ala	aac Asn	240
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Thr	Val	Ser	Ala	Pro	Pro	Ala	Ala	Ser	Ser	Asn	Asp	Asn	Met	Ala	Asp	
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<213> Sorghum bicolor

<400> 97

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 A r g T y r G u A l a H i s L e u T r p A s p A s n S e r C y s A r g A r g G u G y G n
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 T h r A r g L y s G y A r g G n V a l T y r L e u G y G y T y r A s p L y s G u G u
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 L y s A l a A l a A r g A l a T y r A s p L e u A l a A l a L e u L y s T y r T r p G y P r o
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 A r g H i s H i s G n H i s G y A r g T r p G n A l a A r g I l e G y A r g V a l A l a
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 G u A l a A l a A l a S e r A l a G n H i s H i s A l a G y V a l V a l S e r T y r A s p
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cggcct caac	gccgt cacaa	act t cgacat	gagccgct ac	gacgt caaga	gcat cct gga	3000
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cat cgcgct t c	cagccgagcg	cggccacggg	cct gt accac	ccgt acgcgc	agccgat gcg	3240
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ct ccaact cc	gt cgt gt aca	acgggt gt t gg	t gat agcaac	ggcagcaccg	t cgt cggcag	3480
t ggt ggct ac	at gat gcct a	t gagcgct gc	cacggcgacg	gct accacgg	caat ggt gag	3540
ccacgagcag	gt gcat gcac	gggcacaggg	t gat caccac	gacgaagcca	agcaggct gc	3600
t cagat gggg	t acgagagct	acct ggt gaa	cgcagagaac	t at ggcggcg	ggaggat gt c	3660
t cgggcct gg	gcgact gt ct	cagcgccacc	ggcggcaagc	agcaacgat a	acat ggcgga	3720
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 <212> DNA
 <213> Sorghum bicolor

<220>
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5312WOPCT_SEQ_LI STI NG TXT

<222> (1) . . . (2082)

<400> 99

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Asp	Asn	Pro	Gln	Pro	Asn	His	Gln	Asp	Ser	Ser	Pro	Ala	Ala	Ala	Gly	
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ggc	t cc	gac	ggg	aat	ct c	ggc	gt g	ccg	ggc	ct g	cgg	gac	gat	cac	gct	192
Gly	Ser	Asp	Gly	Asn	Leu	Gly	Val	Pro	Gly	Leu	Arg	Asp	Asp	His	Ala	
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Ser	Tyr	Gly	Ile	Met	Glu	Ala	Phe	Asn	Arg	Val	Pro	Gln	Glu	Thr	Gln	
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gat	t gg	aac	at g	agg	gga	t t g	gac	t ac	aac	ggc	ggc	ggc	t cg	gaa	ct c	288
Asp	Trp	Asn	Met	Arg	Gly	Leu	Asp	Tyr	Asn	Gly	Gly	Gly	Ser	Glu	Leu	
				85					90					95		
t cg	at g	ct t	gt g	ggg	t cc	agc	ggc	ggc	ggc	ggc	ggc	ggc	ggc	aag	agg	336
Ser	Met	Leu	Val	Gly	Ser	Ser	Gly	Gly	Gly	Gly	Gly	Gly	Gly	Lys	Arg	
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gcc	gt g	gaa	gac	agc	gag	ccc	aag	ct c	gaa	gat	t t c	ct c	ggc	ggc	aac	384
Ala	Val	Glu	Asp	Ser	Glu	Pro	Lys	Leu	Glu	Asp	Phe	Leu	Gly	Gly	Asn	
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Ser	Phe	Val	Ser	Glu	His	Asp	Gln	Ser	Gly	Gly	Tyr	Leu	Phe	Ser	Gly	
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Val	Pro	Met	Ala	Ser	Ser	Thr	Asn	Ser	Asn	Ser	Gly	Ser	Asn	Thr	Met	
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Glu	Leu	Ser	Met	Ile	Lys	Thr	Trp	Leu	Arg	Asn	Asn	Gln	Val	Pro	Gln	
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ccg	cag	ccg	cca	gca	gct	ccg	cat	cag	gcg	ccg	cag	act	gag	gag	at g	576
Pro	Gln	Pro	Pro	Ala	Ala	Pro	His	Gln	Ala	Pro	Gln	Thr	Glu	Glu	Met	
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agc	acc	gac	gcc	aac	gcc	agc	gcc	agc	agc	t t t	ggc	t gc	t cg	gat	t cg	624
Ser	Thr	Asp	Ala	Asn	Ala	Ser	Ala	Ser	Ser	Phe	Gly	Cys	Ser	Asp	Ser	
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Met	Gly	Arg	Asn	Gly	Thr	Val	Ala	Ala	Ala	Gly	Ser	Ser	Gln	Ser	Leu	
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gcg	ct c	t cg	at g	agc	acg	ggc	t cg	cac	ct g	ccg	at g	gt t	gt g	gcc	ggc	720
Ala	Leu	Ser	Met	Ser	Thr	Gly	Ser	His	Leu	Pro	Met	Val	Val	Ala	Gly	
225					230					235					240	
ggc	ggc	gcc	agc	gga	gcg	gcc	t cg	gag	agc	acg	t ca	t cg	gag	aac	aag	768
Gly	Gly	Ala	Ser	Gly	Ala	Ala	Ser	Glu	Ser	Thr	Ser	Ser	Glu	Asn	Lys	
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cga	gcg	agc	ggc	gcc	at g	gat	t cg	ccc	ggc	agc	gcg	gt a	gaa	gcc	gt c	816

5312 WOPCT_SEQ LISTING.TXT

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Pro	Arg	Lys	Ser	Ile	Asp	Thr	Phe	Gly	Gln	Arg	Thr	Ser	Ile	Tyr	Arg	
		275					280					285				
ggt	gta	aca	aga	cat	aga	tgg	aca	ggg	cga	tat	gag	gct	cat	cta	tgg	912
Gly	Val	Thr	Arg	His	Arg	Trp	Thr	Gly	Arg	Tyr	Glu	Ala	His	Leu	Trp	
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Tyr	Leu	Gly	Gly	Tyr	Asp	Lys	Glu	Asp	Lys	Ala	Ala	Arg	Ala	Tyr	Asp	
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Leu	Ala	Ala	Leu	Lys	Tyr	Trp	Gly	Thr	Thr	Thr	Thr	Thr	Asn	Phe	Pro	
			340					345					350			
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Ile	Ser	Asn	Tyr	Glu	Lys	Glu	Leu	Glu	Glu	Met	Lys	His	Met	Thr	Arg	
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Gln	Glu	Tyr	Ile	Ala	Tyr	Leu	Arg	Arg	Asn	Ser	Ser	Gly	Phe	Ser	Arg	
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Trp	Gln	Ala	Arg	Ile	Gly	Arg	Val	Ala	Gly	Asn	Lys	Asp	Leu	Tyr	Leu	
				405					410					415		
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Arg	Tyr	Asp	Val	Lys	Ser	Ile	Leu	Glu	Ser	Ser	Thr	Leu	Pro	Val	Gly	
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5312WOPCT_SEQ_LI STING.TXT

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Leu	Gly	Ser	Ala	Ala	His	Asn	Phe	Phe	Gln	Ala	Ser	Ser	Ser	Ser	Ala	575	
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Ala	Gly	Ala	Asp	Gln	Gly	His	Ser	Ser	Ser	Thr	Ala	Asn	Gln	Gly	Ser	610	
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Thr	Cys	Ser	Tyr	Gly	Asp	Asp	His	Gln	Glu	Gly	Lys	Leu	Ile	Gly	Tyr	625	
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Ser	Ile	Ala	Arg	Ala	Asn	Gly	Tyr	Ser	Asn	Asn	Trp	Ser	Ser	Pro	Phe	675	
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 35 40 45
 Gly Ser Asp Gly Asn Leu Gly Val Pro Gly Leu Arg Asp Asp His Ala
 50 55 60
 Ser Tyr Gly Ile Met Glu Ala Phe Asn Arg Val Pro Gln Glu Thr Gln
 65 70 75
 Asp Trp Asn Met Arg Gly Leu Asp Tyr Asn Gly Gly Gly Ser Glu Leu
 85 90 95
 Ser Met Leu Val Gly Ser Ser Gly Gly Gly Gly Gly Lys Arg
 100 105 110
 Ala Val Glu Asp Ser Glu Pro Lys Leu Glu Asp Phe Leu Gly Gly Asn
 115 120 125
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5312WOPCT_SEQ_LI STING TXT

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Glu	Leu	Ser	Met	Ile 165	Lys	Thr	Trp	Leu	Arg 170	Asn	Asn	Gln	Val	Pro 175	Gln
Pro	Gln	Pro	Pro 180	Ala	Ala	Pro	His	Gln 185	Ala	Pro	Gln	Thr	Glu 190	Glu	Met
Ser	Thr	Asp 195	Ala	Asn	Ala	Ser	Ala 200	Ser	Ser	Phe	Gly	Cys 205	Ser	Asp	Ser
Met	Gly 210	Arg	Asn	Gly	Thr	Val 215	Ala	Ala	Ala	Gly	Ser 220	Ser	Gln	Ser	Leu
Ala 225	Leu	Ser	Met	Ser	Thr 230	Gly	Ser	His	Leu	Pro 235	Met	Val	Val	Ala	Gly 240
Gly	Gly	Ala	Ser	Gly 245	Ala	Ala	Ser	Glu	Ser 250	Thr	Ser	Ser	Glu	Asn 255	Lys
Arg	Ala	Ser	Gly 260	Ala	Met	Asp	Ser	Pro 265	Gly	Ser	Ala	Val	Glu 270	Ala	Val
Pro	Arg	Lys 275	Ser	Ile	Asp	Thr	Phe 280	Gly	Gln	Arg	Thr	Ser 285	Ile	Tyr	Arg
Gly	Val 290	Thr	Arg	His	Arg	Trp 295	Thr	Gly	Arg	Tyr	Glu 300	Ala	His	Leu	Trp
Asp 305	Asn	Ser	Cys	Arg	Arg 310	Glu	Gly	Gln	Ser	Arg 315	Lys	Gly	Arg	Gln	Val 320
Tyr	Leu	Gly	Gly	Tyr 325	Asp	Lys	Glu	Asp	Lys 330	Ala	Ala	Arg	Ala	Tyr 335	Asp
Leu	Ala	Ala	Leu 340	Lys	Tyr	Trp	Gly	Thr 345	Thr	Thr	Thr	Thr	Asn 350	Phe	Pro
Ile	Ser	Asn 355	Tyr	Glu	Lys	Glu	Leu 360	Glu	Glu	Met	Lys	His 365	Met	Thr	Arg
Gln	Glu 370	Tyr	Ile	Ala	Tyr	Leu 375	Arg	Arg	Asn	Ser	Ser 380	Gly	Phe	Ser	Arg
Gly 385	Ala	Ser	Lys	Tyr	Arg 390	Gly	Val	Thr	Arg	His 395	His	Gln	His	Gly	Arg 400
Trp	Gln	Ala	Arg	Ile 405	Gly	Arg	Val	Ala	Gly 410	Asn	Lys	Asp	Leu	Tyr 415	Leu
Gly	Thr	Phe	Ser 420	Thr	Glu	Glu	Glu	Ala 425	Ala	Glu	Ala	Tyr	Asp 430	Ile	Ala
Ala	Ile	Lys 435	Phe	Arg	Gly	Leu	Asn 440	Ala	Val	Thr	Asn	Phe 445	Asp	Met	Ser
Arg	Tyr	Asp 450	Val	Lys	Ser	Ile 455	Leu	Glu	Ser	Ser	Thr 460	Leu	Pro	Val	Gly
Gly 465	Ala	Ala	Arg	Arg	Leu 470	Lys	Asp	Ala	Val	Asp 475	His	Val	Glu	Ala	Gly 480
Ala	Thr	Ile	Trp	Arg 485	Ala	Asp	Met	Asp	Gly 490	Gly	Val	Ile	Ser	Gln 495	Leu
Ala	Glu	Ala	Gly 500	Met	Gly	Gly	Tyr	Ala 505	Ser	Tyr	Gly	His 510	His	Ala	Trp
Pro	Thr	Ile 515	Ala	Phe	Gln	Gln	Pro 520	Ser	Pro	Leu	Ser	Val 525	His	Tyr	Pro
Tyr	Gly 530	Gln	Pro	Pro	Ser	Arg 535	Gly	Trp	Cys	Lys	Pro 540	Glu	Gln	Asp	Ala
Ala 545	Val	Ala	Ala	Ala	Ala 550	His	Ser	Leu	Gln	Asp 555	Leu	Gln	Gln	Leu	His 560
Leu	Gly	Ser	Ala	Ala 565	His	Asn	Phe	Phe	Gln 570	Ala	Ser	Ser	Ser	Ser	Ala
Val	Tyr	Asn	Ser 580	Gly	Gly	Gly	Gly	Ala 585	Ser	Gly	Gly	Tyr	His 590	Gln	Gly
Leu	Gly	Gly 595	Gly	Ser	Ser	Ser	Phe 600	Leu	Met	Pro	Ser	Ser 605	Thr	Val	Val
Ala	Gly 610	Ala	Asp	Gln	Gly	His 615	Ser	Ser	Ser	Thr	Ala 620	Asn	Gln	Gly	Ser
Thr 625	Cys	Ser	Tyr	Gly	Asp 630	Asp	His	Gln	Glu	Gly 635	Lys	Leu	Ile	Gly	Tyr 640
Asp	Ala	Met	Val	Ala 645	Ala	Thr	Ala	Ala	Gly 650	Gly	Asp	Pro	Tyr	Ala 655	Ala
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Asn Gly Gly Met Gly
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<212> DNA
<213> Zea mays

<220>
<221> CDS
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Asn Pro Gn Pro Asn Gn Asp Ser Ser Pro Ala Ala Gly lle Asp lle
20 25 30
t cc ggc gcc agc gac ttc tat ggc ct g ccc acg cag cag ggc t cc gac 144
Ser Gly Ala Ser Asp Phe Tyr Gly Leu Pro Thr Gn Gn Gly Ser Asp
35 40 45
ggg cat ct c ggc gt g ccg ggc ct g cgg gac gat cac gct t ct tat ggt 192
Gly His Leu Gly Val Pro Gly Leu Arg Asp Asp His Ala Ser Tyr Gly
50 55 60
at c at g gag gcc tac aac agg gt t cct caa gaa acc caa gat t gg aac 240
lle Met Gu Ala Tyr Asn Arg Val Pro Gn Gu Thr Gn Asp Trp Asn
65 70 75 80
at g agg ggc ttg gac tac aac ggc ggt ggc t cg gag ct c t cg at g ct t 288
Met Arg Gly Leu Asp Tyr Asn Gly Gly Ser Gu Leu Ser Met Leu
85 90 95
gt g ggg t cc agc ggc ggc ggc ggg ggc aac ggc aag agg gcc gt g gaa 336
Val Gly Ser Ser Gly Gly Gly Gly Asn Gly Lys Arg Ala Val Gu
100 105 110
gac agc gag ccc aag ct c gaa gat ttc ct c ggc ggc aac t cg ttc gt c 384
Asp Ser Gu Pro Lys Leu Gu Asp Phe Leu Gly Gly Asn Ser Phe Val
115 120 125
t cc gat caa gat cag t cc ggc ggt tac ct g ttc t ct gga gt c ccg at a 432
Ser Asp Gn Asp Gn Ser Gly Gly Tyr Leu Phe Ser Gly Val Pro lle
130 135 140
gcc agc agc gcc aat agc aac agc ggg agc aac acc at g gag ct c t cc 480
Ala Ser Ser Ala Asn Ser Asn Ser Gly Ser Asn Thr Met Gu Leu Ser
145 150 155 160
at g at c aag acc t gg ct a cgg aac aac cag gt g gcc cag ccc cag ccg 528
Met lle Lys Thr Trp Leu Arg Asn Asn Gn Val Ala Gn Pro Gn Pro
165 170 175
cca gct cca cat cag ccg cag cct gag gaa at g agc acc gac gcc agc 576
Pro Ala Pro His Gn Pro Gn Pro Gu Gu Met Ser Thr Asp Ala Ser
180 185 190
ggc agc agc ttt gga tgc t cg gat t cg at g gga agg aac agc at g gt g 624
Gly Ser Ser Phe Gly Cys Ser Asp Ser Met Gly Arg Asn Ser Met Val
195 200 205
gcg gct ggt ggg agc t cg cag agc ct g gcg ct c t cg at g agc acg ggc 672
Ala Ala Gly Gly Ser Ser Gn Ser Leu Ala Leu Ser Met Ser Thr Gly

5312WOPCT_SEQ_LI STI NG TXT

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agc Ser 240	gga Gly 240	gcg Ala 240	gcc Ala 240
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t cg Ser 250	gag Glu 250	aac Asn 250	aag Lys 250
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gcc Ala 255	at g Met 255	gat Asp 255	768
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gag Ala 265	gt a Val 265	gaa Glu 265	gcc Ala 265
gt a Val 265	cgg Pro 265	agg Arg 265	aag Lys 265
t cc Ser 270	at c Ile 270	gac Asp 270	acg Thr 270
816			
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acc Thr 280	t ct Ser 280	ile Ile 280	t at Tyr 280
cga Arg 285	ggt Gly 285	gt a Val 285	aca Thr 285
agg Arg 285	cat His 285	aga Arg 285	t gg Trp 285
864			
aca Thr 290	ggg Gly 290	cgg Arg 290	t at Tyr 290
gag Glu 295	gct Ala 295	cat His 295	ct a Leu 295
t gg Trp 295	gat Asp 295	aat Asn 295	agt Ser 295
t gt Cys 300	aga Arg 300	agg Arg 300	gaa Glu 300
912			
ggg Gly 305	cag Gln 305	agt Ser 305	cgc Arg 305
aag Lys 310	ggg Gly 310	agg Arg 310	caa Gln 310
gt t Val 315	t ac Tyr 315	ct t Leu 315	ggt Gly 315
ggc Gly 320	t at Tyr 320	gac Asp 320	aag Lys 320
960			
gag Glu 325	gac Asp 325	aag Lys 325	gca Ala 325
agg Arg 330	gct Ala 330	t at Tyr 330	t gg Trp 330
1008			
ggc Gly 335	act Thr 335	acg Thr 335	aca Thr 335
aca Thr 340	aca Thr 340	aca Thr 340	aat Asn 340
t t c Phe 345	cct Pro 345	at a Ile 345	agc Ser 345
aac Asn 350	t ac Tyr 350	gaa Glu 350	aag Lys 350
gag Glu 355	ct a Leu 355	t ac Tyr 355	ct a Leu 355
1104			
aga Arg 360	aga Arg 360	gag Glu 360	t ac Tyr 360
at t Ile 365	gca Ala 365	t ac Tyr 365	ct a Leu 365
1152			
aga Arg 370	aga Arg 370	aat Asn 370	agc Ser 370
agt Ser 375	gga Gly 375	t t t Phe 375	t ct Ser 375
cgt Arg 380	ggg Gly 380	gag Glu 380	t ca Ser 380
aag Lys 385	t at Tyr 385	cgt Arg 385	gga Gly 385
1200			
gt a Val 385	act Thr 385	aga Arg 385	cat His 385
cat His 390	cag Gln 390	cat His 390	ggg Gly 390
aga Arg 395	t gg Trp 395	caa Gln 395	gca Ala 395
agg Arg 400	at a Ile 400	ggg Gly 400	aga Arg 400
1248			
gt t Val 405	gca Ala 405	gga Gly 405	aac Asn 405
aag Lys 410	gat Asp 410	ct c Leu 410	t ac Tyr 410
t t g Leu 415	ggc Gly 415	aca Thr 415	t t c Phe 415
agc Ser 420	acc Thr 420	gag Glu 420	gag Glu 420
1296			
gag Glu 425	gag Glu 425	gag Glu 425	gag Glu 425
gag Glu 430	gag Glu 430	gag Glu 430	gag Glu 430
1344			
aac Asn 435	gag Glu 435	agc Ser 435	aca Thr 435
ct g Leu 440	cct Pro 440	gt c Val 440	gag Glu 440
ggc Gly 445	ggt Gly 445	gag Glu 445	gag Glu 445
acc Thr 450	at c Ile 450	t gg Trp 450	cgc Arg 450
gcc Ala 455	agg Arg 455	cgc Arg 455	ct c Leu 455
aag Lys 460	agc Ser 460	at c Ile 460	ggc Gly 460
1392			
gag Glu 465	gag Glu 465	gag Glu 465	gag Glu 465
gag Glu 470	gag Glu 470	gag Glu 470	gag Glu 470
gag Glu 475	gag Glu 475	gag Glu 475	gag Glu 475
1440			
at g Met 480	gag Glu 480	gag Glu 480	gag Glu 480
gag Glu 485	gag Glu 485	gag Glu 485	gag Glu 485
gag Glu 490	gag Glu 490	gag Glu 490	gag Glu 490
1488			

5312WOPCT_SEQ_LI STI NG TXT

485										490					495					
t ac	gcc	t cg	t ac	ggc	cac	cac	ggc	t gg	ccg	acc	at c	gcg	t t c	cag	cag	1536				
Tyr	Al a	Ser	Tyr	G y	Hi s	Hi s	G y	Tr p	Pro	Thr	I l e	Al a	Phe	G n	G n					
			500					505					510							
ccg	t cg	ccg	ct c	t cc	gt c	cac	t ac	ccg	t ac	ggc	cag	ccg	t cc	cgc	ggg	1584				
Pro	Ser	Pro	Leu	Ser	Val	Hi s	Tyr	Pro	Tyr	G y	G n	Pro	Ser	Arg	G y					
		515					520					525								
t gg	t gc	aaa	ccc	gag	cag	gac	gcg	gcc	gcc	gcc	gcg	gcg	cac	agc	ct g	1632				
Tr p	Cys	Lys	Pro	G u	G n	Asp	Al a	Al a	Al a	Al a	Al a	Al a	Hi s	Ser	Leu					
	530					535					540									
cag	gac	ct c	cag	cag	ct g	cac	ct c	ggc	agc	gcg	gcc	cac	aac	t t c	t t c	1680				
G n	Asp	Leu	G n	G n	Leu	Hi s	Leu	G y	Ser	Al a	Al a	Hi s	Asn	Phe	Phe					
545					550					555					560					
cag	gcg	t cg	t cg	agc	t cc	aca	gt c	t ac	aac	ggc	ggc	gcc	ggc	gcc	agt	1728				
G n	Al a	Ser	Ser	Ser	Ser	Thr	Val	Tyr	Asn	G y	G y	Al a	G y	Al a	Ser					
				565					570					575						
ggc	ggg	t ac	cag	ggc	ct c	ggc	ggc	ggc	agc	t ct	t t c	ct c	at g	ccg	t cg	1776				
G y	G y	Tyr	G n	G y	Leu	G y	G y	G y	Ser	Ser	Phe	Leu	Met	Pro	Ser					
			580					585					590							
agc	act	gt c	gt g	gcg	gcg	gcc	gac	cag	ggg	cac	agc	agc	acg	gcc	aac	1824				
Ser	Thr	Val	Val	Al a	Al a	Al a	Asp	G n	G y	Hi s	Ser	Ser	Thr	Al a	Asn					
		595					600					605								
cag	ggg	agc	acg	t gc	agc	t ac	ggg	gac	gac	cac	cag	gag	ggg	aag	ct c	1872				
G n	G y	Ser	Thr	Cys	Ser	Tyr	G y	Asp	Asp	Hi s	G n	G u	G y	Lys	Leu					
	610					615					620									
at c	ggc	t ac	gac	gcc	gcc	at g	gt g	gcg	acc	gca	gct	ggt	gga	gac	ccg	1920				
I l e	G y	Tyr	Asp	Al a	Al a	Met	Val	Al a	Thr	Al a	Al a	G y	G y	Asp	Pro					
625					630					635					640					
t ac	gct	gcg	gcg	agg	aac	ggg	t ac	cag	t t c	t cg	cag	ggc	t cg	gga	t cc	1968				
Tyr	Al a	Al a	Al a	Arg	Asn	G y	Tyr	G n	Phe	Ser	G n	G y	Ser	G y	Ser					
				645					650					655						
acg	gt g	agc	at c	gcg	agg	gcg	aac	ggg	t ac	gct	aac	aac	t gg	agc	t ct	2016				
Thr	Val	Ser	I l e	Al a	Arg	Al a	Asn	G y	Tyr	Al a	Asn	Asn	Tr p	Ser	Ser					
			660					665					670							
cct	t t c	aac	aac	ggc	at g	ggg	t ga									2040				
Pro	Phe	Asn	Asn	G y	Met	G y														
		675																		

<210> 102
 <211> 679
 <212> PRT
 <213> Zea mays

<400> 102
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 Asn Pro G n Pro Asn G n Asp Ser Ser Pro Al a Al a G y I l e Asp I l e
 20 25 30
 Ser G y Al a Ser Asp Phe Tyr G y Leu Pro Thr G n G n G y Ser Asp
 35 40 45
 G y Hi s Leu G y Val Pro G y Leu Arg Asp Asp Hi s Al a Ser Tyr G y
 50 55 60
 I l e Met G u Al a Tyr Asn Arg Val Pro G n G u Thr G n Asp Trp Asn
 65 70 75 80
 Met Arg G y Leu Asp Tyr Asn G y G y G y Ser G u Leu Ser Met Leu

5312WOPCT_SEQ_LI STI NG TXT

Val	Gly	Ser	Ser	85 Gly	Gly	Gly	Gly	Gly	90 Asn	Gly	Lys	Arg	Ala	95 Val	Glu
Asp	Ser	Glu	100 Pro	Lys	Leu	Glu	105 Asp	Phe	Leu	Gly	Gly	Asn	110 Ser	Phe	Val
Ser	Asp	Gln	115 Asp	Gln	Ser	Gly	120 Gly	Tyr	Leu	Phe	Ser	125 Gly	Val	Pro	Ile
Ala	Ser	Ser	130 Ser	Ala	Asn	Ser	135 Asn	Ser	Gly	Ser	Asn	140 Thr	Met	Glu	Leu
145 Met	Ile	Lys	150 Thr	Trp	Leu	Arg	155 Asn	Asn	Gln	Val	Ala	Gln	Pro	Gln	160 Pro
Pro	Ala	Pro	165 His	Gln	Pro	Gln	170 Pro	Glu	Glu	Met	Ser	175 Thr	Asp	Ala	Ser
Gly	Ser	Ser	180 Phe	Gly	Cys	Ser	185 Asp	Ser	Met	Gly	Arg	190 Asn	Ser	Met	Val
Ala	Ala	210 Gly	Gly	Ser	Ser	Gln	200 Ser	Leu	Ala	Leu	Ser	205 Met	Ser	Thr	Gly
Ser	His	Leu	Pro	Met	Val	Val	Pro	Ser	Gly	Ala	Ala	Ser	Gly	Ala	Ala
225 Ser	Glu	Ser	230 Thr	Ser	Glu	Asn	Lys	Arg	Ala	Ser	Gly	Ala	Met	Asp	240 Asp
Ser	Pro	Gly	245 Ser	Ala	Val	Glu	Ala	Val	Pro	Arg	Lys	Ser	Ile	Asp	Thr
Phe	Gly	Gln	260 Arg	Thr	Ser	Ile	Tyr	Arg	Gly	Val	Thr	Arg	His	Arg	Trp
Thr	Gly	Arg	275 Tyr	Glu	Ala	His	280 Leu	Trp	Asp	Asn	Ser	285 Cys	Arg	Arg	Glu
Gly	Gln	Ser	Arg	Lys	Gly	Arg	Gln	Val	Tyr	Leu	Gly	Gly	Tyr	Asp	Lys
305 Glu	Asp	Lys	Ala	Ala	Arg	Ala	Tyr	Asp	Leu	Ala	Ala	Leu	Lys	Tyr	Trp
Gly	Thr	Thr	325 Thr	Thr	Asn	Phe	Pro	Ile	Ser	Asn	Tyr	Glu	Lys	Glu	
Leu	Glu	Glu	340 Met	Lys	His	Met	Thr	Arg	Gln	Glu	Tyr	Ile	Ala	Tyr	Leu
Arg	Arg	Asn	355 Ser	Ser	Gly	Phe	360 Ser	Arg	Gly	Ala	Ser	365 Lys	Tyr	Arg	Gly
Val	Thr	Arg	370 His	His	Gln	His	Gly	Arg	Trp	Gln	Ala	Arg	Ile	Gly	Arg
385 Val	Ala	Gly	Asn	Lys	Asp	Leu	Tyr	Leu	Gly	Thr	Phe	Ser	Thr	Glu	Glu
Glu	Ala	Ala	405 Glu	Tyr	Asp	Ile	Ala	Ile	Lys	Phe	Arg	415 Gly	Leu		
Asn	Ala	Val	420 Thr	Asn	Phe	Asp	Met	Ser	Arg	Tyr	Asp	Val	Lys	Ser	Ile
Leu	Glu	Ser	435 Ser	Thr	Leu	Pro	Val	Gly	Gly	Ala	Ala	Arg	Arg	Leu	Lys
Asp	Ala	Val	450 Asp	His	Val	Glu	Ala	Gly	Ala	Thr	Ile	Trp	Arg	Ala	Asp
465 Met	Asp	Gly	470 Ala	Val	Ile	Ser	Gln	Leu	Ala	Glu	Ala	Gly	Met	Gly	Gly
Tyr	Ala	Ser	485 Tyr	Gly	His	His	Gly	Trp	Pro	Thr	Ile	Ala	Phe	Gln	Gln
Pro	Ser	Pro	500 Leu	Ser	Val	His	Tyr	Pro	Tyr	Gly	Gln	Pro	Ser	Arg	Gly
Trp	Cys	Lys	515 Pro	Glu	Gln	Asp	Ala	Ala	Ala	Ala	Ala	Ala	His	Ser	Leu
Gln	Asp	Leu	530 Gln	Gln	Leu	His	535 Leu	Gly	Ser	Ala	Ala	His	Asn	Phe	Phe
545 Gln	Ala	Ser	Ser	Ser	Thr	Val	Tyr	Asn	Gly	Gly	Ala	Gly	Ala	Ser	
Gly	Gly	Tyr	565 Gln	Gly	Leu	Gly	Gly	Gly	Ser	Ser	Phe	Leu	Met	Pro	Ser
Ser	Thr	Val	580 Val	Ala	Ala	Ala	Asp	Gln	Gly	His	Ser	Ser	Thr	Ala	Asn
Gln	Gly	Ser	595 Thr	Cys	Ser	Tyr	Gly	Asp	Asp	His	Gln	Glu	Gly	Lys	Leu
Ile	Gly	Tyr	610 Asp	Ala	Ala	Met	Val	Ala	Thr	Ala	Ala	Gly	Gly	Asp	Pro

5312WOPCT_SEQ LISTING.TXT

625 Tyr Ala Ala Ala Arg Asn Gly Tyr Gln Phe Ser Gln Gly Ser Gly Ser
 630 635 640
 Thr Val Ser Ile Ala Arg Ala Asn Gly Tyr Ala Asn Asn Trp Ser Ser
 645 650 655
 Pro Phe Asn Asn Gly Met Gly
 660 665 670
 675

<210> 103
 <211> 975
 <212> DNA
 <213> Zea mays

<220>
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 <222> (1)...(975)

<400> 103
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 1 5 10 15
 cac ggg cag gac gac ggc ggg t cg ccg ccg at g t cg ccg gcc t cc gcc 96
 His G y G n Asp Asp G y G y Ser Pro Pro Met Ser Pro Ala Ser Ala
 20 25 30
 gcg gcg gcg gcg ct g gcg aac gcg ccg t gg aac ccg acc aag gag cag 144
 Ala Ala Ala Ala Leu Ala Asn Ala Arg Trp Asn Pro Thr Lys G u G n
 35 40 45
 gt g gcc gt g ct g gag ggg ct g t ac gag cac ggc ct g cg c acc ccc agc 192
 Val Ala Val Leu G u G y Leu Tyr G u His G y Leu Arg Thr Pro Ser
 50 55 60
 gcg gag cag at a cag cag at c acg ggc agg ct g ccg gag cac ggc gcc 240
 Ala G u G n Ile G n G n Ile Thr G y Arg Leu Arg G u His G y Ala
 65 70 75 80
 at c gag ggc aag aac gt c t t c t ac t gg t t c cag aac cac aag gcc cgc 288
 Ile G u G y Lys Asn Val Phe Tyr Trp Phe G n Asn His Lys Ala Arg
 85 90 95
 cag cgc cag agg cag aag cag gac agc t t c gcc t ac t t c agc agg ct c 336
 G n Arg G n Arg G n Lys G n Asp Ser Phe Ala Tyr Phe Ser Arg Leu
 100 105 110
 ct c cgc ccg ccc ccg ccg ct g ccc gt g ct c t cc at g ccc ccc gcg cca 384
 Leu Arg Arg Pro Pro Pro Leu Pro Val Leu Ser Met Pro Pro Ala Pro
 115 120 125
 ccg t ac cat cac gcc cgc gt c ccg gcg ccg ccc gcg at a ccg at g ccg 432
 Pro Tyr His His Ala Arg Val Pro Ala Pro Pro Ala Ile Pro Met Pro
 130 135 140
 at g gcg ccg ccg ccg ccc gct gca t gc aac gac aac ggc ggc gcg cgt 480
 Met Ala Pro Pro Pro Pro Ala Ala Cys Asn Asp Asn G y G y Ala Arg
 145 150 155 160
 gt g at c t ac agg aac cca t t c t ac gt g gct gcg ccg cag gcg ccc cct 528
 Val Ile Tyr Arg Asn Pro Phe Tyr Val Ala Ala Pro G n Ala Pro Pro
 165 170 175
 gca aat gcc gcc t ac t ac t ac cca cag cca cag cag cag cag cag 576
 Ala Asn Ala Ala Tyr Tyr Tyr Pro G n Pro G n G n G n G n G n G n
 180 185 190
 cag gt g aca gt c at g t ac cag t ac ccg aga at g gag gt a gcc ggc cag 624

5312WOPCT_SEQ LI STI NG TXT

G n	Val	Thr	Val	Met	Tyr	G n	Tyr	Pro	Arg	Met	G u	Val	Al a	G y	G n	
		195					200					205				
gac	aag	at g	at g	acc	agg	gcc	gcg	gcg	cac	cag	cag	cag	cag	cac	aac	672
Asp	Lys	Met	Met	Thr	Arg	Al a	Al a	Al a	His	G n	G n	G n	G n	His	Asn	
	210					215					220					
ggc	gcc	ggg	caa	caa	ccg	gga	gcg	gcc	ggc	cac	ccc	agc	cgc	gag	acg	720
G y	Al a	G y	G n	G n	Pro	G y	Arg	Al a	G y	His	Pro	Ser	Arg	G u	Thr	
225					230					235					240	
ct c	cag	ct g	t t c	ccg	ct c	cag	ccc	acc	t t c	gt g	ct g	cgg	cac	gac	aag	768
Leu	G n	Leu	Phe	Pro	Leu	G n	Pro	Thr	Phe	Val	Leu	Arg	His	Asp	Lys	
				245					250					255		
ggg	cg c	gcc	gcc	aac	ggc	agt	aat	aac	gac	t cc	ct g	acg	t cg	acg	t cg	816
G y	Arg	Al a	Al a	Asn	G y	Ser	Asn	Asn	Asp	Ser	Leu	Thr	Ser	Thr	Ser	
			260					265					270			
acg	gcg	act	gcg	aca	gcg	aca	gcg	aca	gcg	aca	gcg	t cc	gct	t cc	at c	864
Thr	Al a	Thr	Al a	Thr	Al a	Thr	Al a	Thr	Al a	Thr	Al a	Ser	Al a	Ser	I l e	
		275						280				285				
t cc	gag	gac	t cg	gat	ggc	ct g	gag	agc	ggc	agc	t cc	ggc	aag	ggc	gt c	912
Ser	G u	Asp	Ser	Asp	G y	Leu	G u	Ser	G y	Ser	Ser	G y	Lys	G y	Val	
	290					295					300					
gag	gag	gcg	ccc	gcg	ct g	ccg	t t c	t at	gac	t t c	t t c	ggg	ct c	cag	t cc	960
G u	G u	Al a	Pro	Al a	Leu	Pro	Phe	Tyr	Asp	Phe	Phe	G y	Leu	G n	Ser	
305					310					315					320	
t cc	gga	ggc	cg c	t ga												975
Ser	G y	G y	Arg													

<210> 104
 <211> 324
 <212> PRT
 <213> Zea mays

<400> 104

Met	G u	Thr	Pro	G n	G n	G n	Ser	Al a	Al a	Al a	Al a	Al a	Al a	Al a	Al a	
1				5					10					15		
His	G y	G n	Asp	Asp	G y	G y	Ser	Pro	Pro	Met	Ser	Pro	Al a	Ser	Al a	
			20					25					30			
Al a	Al a	Al a	Al a	Leu	Al a	Asn	Al a	Arg	Trp	Asn	Pro	Thr	Lys	G u	G n	
		35					40					45				
Val	Al a	Val	Leu	G u	G y	Leu	Tyr	G u	His	G y	Leu	Arg	Thr	Pro	Ser	
	50					55					60					
Al a	G u	G n	I l e	G n	G n	I l e	Thr	G y	Arg	Leu	Arg	G u	His	G y	Al a	
65				70						75					80	
I l e	G u	G y	Lys	Asn	Val	Phe	Tyr	Trp	Phe	G n	Asn	His	Lys	Al a	Arg	
				85					90					95		
G n	Arg	G n	Arg	G n	Lys	G n	Asp	Ser	Phe	Al a	Tyr	Phe	Ser	Arg	Leu	
			100					105					110			
Leu	Arg	Arg	Pro	Pro	Pro	Leu	Pro	Val	Leu	Ser	Met	Pro	Pro	Al a	Pro	
		115					120					125				
Pro	Tyr	His	His	Al a	Arg	Val	Pro	Al a	Pro	Pro	Al a	I l e	Pro	Met	Pro	
	130					135					140					
Met	Al a	Pro	Pro	Pro	Pro	Al a	Al a	Cys	Asn	Asp	Asn	G y	G y	Al a	Arg	
145					150					155					160	
Val	I l e	Tyr	Arg	Asn	Pro	Phe	Tyr	Val	Al a	Al a	Pro	G n	Al a	Pro	Pro	
				165					170					175		
Al a	Asn	Al a	Al a	Tyr	Tyr	Tyr	Pro	G n	Pro	G n	G n	G n	G n	G n	G n	
			180					185					190			
G n	Val	Thr	Val	Met	Tyr	G n	Tyr	Pro	Arg	Met	G u	Val	Al a	G y	G n	
		195					200					205				

5312WOPCT_SEQ_LI STING.TXT

Asp Lys Met Met Thr Arg Ala Ala Ala His Gln Gln Gln Gln His Asn
 210 215 220
 Gly Ala Gly Gln Gln Pro Gly Arg Ala Gly His Pro Ser Arg Gu Thr
 225 230 235
 Leu Gln Leu Phe Pro Leu Gln Pro Thr Phe Val Leu Arg His Asp Lys
 245 250 255
 Gly Arg Ala Ala Asn Gly Ser Asn Asn Asp Ser Leu Thr Ser Thr Ser
 260 265 270
 Thr Ala Thr Ala Thr Ala Thr Ala Thr Ala Thr Ala Ser Ala Ser Ile
 275 280 285
 Ser Gu Asp Ser Asp Gly Leu Gly Ser Ser Ser Gly Lys Gly Val
 290 295 300
 Gu Gu Ala Pro Ala Leu Pro Phe Tyr Asp Phe Phe Gly Leu Gln Ser
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 Ser Gly Gly Arg

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 <213> Zea mays

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 1 5 10 15
 ggc agc gtg gct gcg ccg gcg gtg tgc cgc ccc agc ggc tgc cgg tgg 96
 Gly Ser Val Ala 20 Ala Pro Ala Val Cys 25 Arg Pro Ser Gly 30 Ser Arg Trp
 20 25 30
 acg ccg acg ccg gag cag atc agg atg ctg aag gag ctg tac tac ggc 144
 Thr Pro Thr Pro Gu Gln Ile Arg Met Leu Lys Gu Leu Tyr Tyr Gly
 35 40 45
 tgc ggc atc cgg tgc ccc agc tgc gag cag atc cag cgc atc acc gcc 192
 Cys Gly Ile Arg Ser Pro Ser Ser Gu Gln Ile Gln Arg Ile Thr Ala
 50 55 60
 atg ctg cgg cag cac ggc aag atc gag ggc aag aac gtc ttc tac tgg 240
 Met Leu Arg Gln His Gly Lys Ile Gu Gly Lys Asn Val Phe Tyr Trp
 65 70 75 80
 ttc cag aac cac aag gcc cgc gag cgc cag aag cgc cgc ctg acc agc 288
 Phe Gln Asn His Lys 85 Ala Arg Gu Arg Gln Lys Arg Arg Leu Thr Ser
 85 90 95
 ctg gac gtc aac gtg ccc gcc gcc gcc ggc gcc gac gcc acc acc agc 336
 Leu Asp Val Asn Val Pro Ala Ala Gly Ala Ala Asp Ala Thr Thr Ser
 100 105 110
 caa ctg ggc gtc ctg tgc ctg tgc tgc ccg ccg cct tca ggc gcg gcg 384
 Gln Leu Gly Val Leu Ser Leu Ser Pro Pro Pro Ser 125 Gly Ala Ala
 115 120 125
 cct ccc tgc ccc acc ctg gcc ttc tac gcc gcc ggc aat ggc ggc gga 432
 Pro Pro Ser Pro Thr Leu Gly Phe Tyr Ala Ala Gly Asn Gly Gly Gly
 130 135 140
 tgc gct gtg ctg ctg gac acg agt tcc gac tgg ggc agc agc ggc gct 480
 Ser Ala Val Leu Leu Asp Thr Ser Ser Asp Trp Gly Ser Ser Gly Ala
 145 150 155 160

5312WOPCT_SEQ_LI STI NG TXT

gcc	at g	gcc	acc	gag	aca	tgc	ttc	ctg	cag	gac	tac	at g	ggc	gt g	acg	528
Al a	Met	Al a	Thr	Gl u	Thr	Cys	Phe	Leu	Gl n	Asp	Tyr	Met	Gl y	Val	Thr	
				165					170					175		
gac	acg	ggc	agc	tcg	tgc	cag	tgg	cca	cgc	ttc	tgc	tgc	tgc	gac	acg	576
Asp	Thr	Gl y	Ser	Ser	Ser	Gl n	Trp	Pro	Arg	Phe	Ser	Ser	Ser	Asp	Thr	
			180					185					190			
at a	at g	gcg	gcg	gcc	gcg	gcg	cgg	gcg	gcg	acg	acg	cgg	gcg	ccc	gag	624
Ile	Met	Al a	Al a	Al a	Al a	Al a	Arg	Al a	Al a	Thr	Thr	Arg	Al a	Pro	Gl u	
		195					200					205				
acg	ctc	cct	ctc	ttc	ccg	acc	tgc	ggc	gac	gac	ggc	ggc	agc	ggc	agc	672
Thr	Leu	Pro	Leu	Phe	Pro	Thr	Cys	Gl y	Asp	Asp	Gl y	Gl y	Ser	Gl y	Ser	
	210					215					220					
agc	agc	tac	tig	ccg	ttc	tgg	ggc	gcc	gcg	tcc	aca	act	gcc	ggc	gcc	720
Ser	Ser	Tyr	Leu	Pro	Phe	Trp	Gl y	Al a	Al a	Ser	Thr	Thr	Al a	Gl y	Al a	
225				230						235					240	
act	tct	tcc	gtt	gcg	atc	cag	cag	caa	cac	cag	ctg	cag	gag	cag	tac	768
Thr	Ser	Ser	Val	Al a	Ile	Gl n	Gl n	Gl n	His	Gl n	Leu	Gl n	Gl u	Gl n	Tyr	
				245					250					255		
agc	ttt	tac	agc	aac	agc	aac	agc	acc	cag	ctg	gcc	ggc	acc	ggc	aac	816
Ser	Phe	Tyr	Ser	Asn	Ser	Asn	Ser	Thr	Gl n	Leu	Al a	Gl y	Thr	Gl y	Asn	
			260					265					270			
caa	gac	gta	tgc	gca	aca	gca	gca	gca	gcc	gcc	gcc	ctg	gag	ctg	agc	864
Gl n	Asp	Val	Ser	Al a	Thr	Al a	Al a	Al a	Al a	Al a	Al a	Leu	Gl u	Leu	Ser	
		275				280						285				
ctc	agc	tca	tgg	tgc	tcc	cct	tac	cct	gct	gca	ggg	agt	at g	tga		909
Leu	Ser	Ser	Trp	Cys	Ser	Pro	Tyr	Pro	Al a	Al a	Gl y	Ser	Met			
	290					295					300					

<210> 106
 <211> 302
 <212> PRT
 <213> Zea mays

<400> 106

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1			5						10				15		
Gl y	Ser	Val	Al a	Al a	Pro	Al a	Val	Cys	Arg	Pro	Ser	Gl y	Ser	Arg	Trp
			20					25					30		
Thr	Pro	Thr	Pro	Gl u	Gl n	Ile	Arg	Met	Leu	Lys	Gl u	Leu	Tyr	Tyr	Gl y
		35					40					45			
Cys	Gl y	Ile	Arg	Ser	Pro	Ser	Ser	Gl u	Gl n	Ile	Gl n	Arg	Ile	Thr	Al a
	50					55					60				
Met	Leu	Arg	Gl n	His	Gl y	Lys	Ile	Gl u	Gl y	Lys	Asn	Val	Phe	Tyr	Trp
65				70					75						80
Phe	Gl n	Asn	His	Lys	Al a	Arg	Gl u	Arg	Gl n	Lys	Arg	Arg	Leu	Thr	Ser
			85					90					95		
Leu	Asp	Val	Asn	Val	Pro	Al a	Al a	Gl y	Al a	Al a	Asp	Al a	Thr	Thr	Ser
			100					105					110		
Gl n	Leu	Gl y	Val	Leu	Ser	Leu	Ser	Ser	Pro	Pro	Pro	Ser	Gl y	Al a	Al a
		115					120					125			
Pro	Pro	Ser	Pro	Thr	Leu	Gl y	Phe	Tyr	Al a	Al a	Gl y	Asn	Gl y	Gl y	Gl y
	130					135					140				
Ser	Al a	Val	Leu	Leu	Asp	Thr	Ser	Ser	Asp	Trp	Gl y	Ser	Ser	Gl y	Al a
145					150				155						160
Al a	Met	Al a	Thr	Gl u	Thr	Cys	Phe	Leu	Gl n	Asp	Tyr	Met	Gl y	Val	Thr
				165					170					175	
Asp	Thr	Gl y	Ser	Ser	Ser	Gl n	Trp	Pro	Arg	Phe	Ser	Ser	Ser	Asp	Thr
			180					185					190		

5312 WOPCT_SEQ LISTING.TXT

I l e	M e t	A l a	A l a	A l a	A l a	A l a	A r g	A l a	A l a	T h r	T h r	A r g	A l a	P r o	G l u
		195					200					205			
T h r	L e u	P r o	L e u	P h e	P r o	T h r	C y s	G l y	A s p	A s p	G l y	G l y	S e r	G l y	S e r
	210					215					220				
S e r	S e r	T y r	L e u	P r o	P h e	T r p	G l y	A l a	A l a	S e r	T h r	T h r	A l a	G l y	A l a
225					230					235					240
T h r	S e r	S e r	V a l	A l a	I l e	G l n	G l n	G l n	H i s	G l n	L e u	G l n	G l u	G l n	T y r
				245					250					255	
S e r	P h e	T y r	S e r	A s n	S e r	A s n	S e r	T h r	G l n	L e u	A l a	G l y	T h r	G l y	A s n
			260					265					270		
G l n	A s p	V a l	S e r	A l a	T h r	A l a	A l a	A l a	A l a	A l a	A l a	L e u	G l u	L e u	S e r
		275					280					285			
L e u	S e r	S e r	T r p	C y s	S e r	P r o	T y r	P r o	A l a	A l a	G l y	S e r	M e t		
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M e t	A l a	A l a	A s n	A l a	G l y	G l y	G l y	G l y	A l a	G l y	G l y	G l y	S e r	G l y	S e r		
1				5				10						15			
g g c	a g c	g t g	g c t	g c g	c c g	g c g	g t g	t g c	c g c	c c c	a g c	g g c	t c g	c g g	t g g		96
G l y	S e r	V a l	A l a	A l a	P r o	A l a	V a l	C y s	A r g	P r o	S e r	G l y	S e r	A r g	T r p		
			20					25					30				
a c g	c c g	a c g	c c g	g a g	c a g	a t c	a g g	a t g	c t g	a a g	g a g	c t c	t a c	t a c	g g c		144
T h r	P r o	T h r	P r o	G l u	G l n	I l e	A r g	M e t	L e u	L y s	G l u	L e u	T y r	T y r	G l y		
			35				40					45					
t g c	g g c	a t c	c g g	t c g	c c c	a g c	t c g	g a g	c a g	a t c	c a g	c g c	a t c	a c c	g c c		192
C y s	G l y	I l e	A r g	S e r	P r o	S e r	S e r	G l u	G l n	I l e	G l n	A r g	I l e	T h r	A l a		
	50					55					60						
a t g	c t g	c g g	c a g	c a c	g g c	a a g	a t c	g a g	g g c	a a g	a a c	g t c	t t c	t a c	t g g		240
M e t	L e u	A r g	G l n	H i s	G l y	L y s	I l e	G l u	G l y	L y s	A s n	V a l	P h e	T y r	T r p		
	65				70					75					80		
t t c	c a g	a a c	c a c	a a g	g c c	c g c	g a g	c g c	c a g	a a g	c g c	c g c	c t c	a c c	a g c		288
P h e	G l n	A s n	H i s	L y s	A l a	A r g	G l u	A r g	G l n	L y s	A r g	A r g	L e u	T h r	S e r		
				85					90					95			
c t c	g a c	g t c	a a c	g t g	c c c	g c c	g c c	g g c	g c g	g c c	g a c	g c c	a c c	a c c	a g c		336
L e u	A s p	V a l	A s n	V a l	P r o	A l a	A l a	G l y	A l a	A l a	A s p	A l a	T h r	T h r	S e r		
			100					105					110				
c a a	c t c	g g c	g t c	c t c	t c g	c t g	t c g	t c g	c c g	c c t	t c a	g g c	g c g	g c g	c c t		384
G l n	L e u	G l y	V a l	L e u	S e r	L e u	S e r	S e r	P r o	P r o	S e r	G l y	A l a	A l a	P r o		
		115				120						125					
c c c	t c g	c c c	a c c	c t c	g g c	t t c	t a c	g c c	g c c	g g c	a a t	g g c	g g c	g g a	t c g		432
P r o	S e r	P r o	T h r	L e u	G l y	P h e	T y r	A l a	A l a	G l y	A s n	G l y	G l y	G l y	S e r		
	130				135						140						
g c t	g g g	c t g	c t g	g a c	a c g	a g t	t c c	g a c	t g g	g g c	a g c	a g c	g g c	g c t	g c t		480
A l a	G l y	L e u	L e u	A s p	T h r	S e r	S e r	A s p	T r p	G l y	S e r	S e r	G l y	A l a	A l a		
	145				150					155					160		
a t g	g c c	a c c	g a g	a c a	t g c	t t c	c t g	c a g	g a c	t a c	a t g	g g c	g t g	a c g	g a c		528
M e t	A l a	T h r	G l u	T h r	C y s	P h e	L e u	G l n	A s p	T y r	M e t	G l y	V a l	T h r	A s p		

5312WOPCT_SEQ_LI STI NG TXT

165

170

175

acg	ggc	agc	t cg	t cg	cag	t gg	cca	t gc	t t c	t cg	t cg	t cg	gac	acg	at a	576
Thr	G y	Ser	Ser	Ser	G n	Tr p	Pro	Cys	Phe	Ser	Ser	Ser	Asp	Thr	I l e	
			180					185					190			
at g	g cg	g cg	g cg	g cg	g cc	g cg	g cg	c gg	gt g	g cg	ac g	ac g	c gg	g cg	ccc	624
Met	Al a	Al a	Al a	Al a	Al a	Al a	Al a	Arg	Val	Al a	Thr	Thr	Arg	Al a	Pro	
		195					200					205				
gag	aca	ct c	c ct	ct c	t t c	c cg	acc	t gc	g gc	gac	gac	gac	gac	gac	gac	672
G u	Thr	Leu	Pro	Leu	Phe	Pro	Thr	Cys	G y	Asp	Asp	Asp	Asp	Asp	Asp	
	210					215				220						
agc	cag	ccc	c cg	c cg	c gg	c cg	c gg	cac	g ca	gt c	cca	gt c	c cg	g ca	g gc	720
Ser	G n	Pro	Pro	Pro	Arg	Pro	Arg	Hi s	Al a	Val	Pro	Val	Pro	Al a	G y	
225					230					235					240	
gag	acc	at c	c gc	g gc	g gc	g gc	g gc	agc	agc	agc	agc	t ac	t t g	c cg	t t c	768
G u	Thr	I l e	Arg	G y	G y	G y	G y	Ser	Ser	Ser	Ser	Tyr	Leu	Pro	Phe	
				245				250					255			
t gg	g gt	g cc	g gt	g cc	g cg	t cc	aca	act	g cc	g gc	g cc	act	t ct	t cc	gt t	816
Tr p	G y	Al a	G y	Al a	Al a	Ser	Thr	Thr	Al a	G y	Al a	Thr	Ser	Ser	Val	
			260					265					270			
g cg	at c	cag	cag	caa	cac	cag	ct g	cag	gag	cag	t ac	agc	t t t	t ac	agc	864
Al a	I l e	G n	G n	G n	Hi s	G n	Leu	G n	G u	G n	Tyr	Ser	Phe	Tyr	Ser	
		275					280					285				
aac	agc	acc	cag	ct g	g cc	g gc	acc	g gc	agc	caa	gac	gt a	t cg	g ct	t ca	912
Asn	Ser	Thr	G n	Leu	Al a	G y	Thr	G y	Ser	G n	Asp	Val	Ser	Al a	Ser	
	290					295					300					
g cg	g cc	g cc	ct g	gag	ct g	agc	ct c	agc	t ca	t gg	t gc	t cc	c ct	t ac	c ct	960
Al a	Al a	Al a	Leu	G u	Leu	Ser	Leu	Ser	Ser	Tr p	Cys	Ser	Pro	Tyr	Pro	
305				310						315					320	
g ct	g ca	g gg	agc	at g	t ga											978
Al a	Al a	G y	Ser	Met												
				325												

<210> 108

<211> 325

<212> PRT

<213> Zea mays

<400> 108

Met	Al a	Al a	Asn	Al a	G y	G y	G y	G y	Al a	G y	G y	G y	Ser	G y	Ser	
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G y	Ser	Val	Al a	Al a	Pro	Al a	Val	Cys	Arg	Pro	Ser	G y	Ser	Arg	Tr p	
			20					25					30			
Thr	Pro	Thr	Pro	G u	G n	I l e	Arg	Met	Leu	Lys	G u	Leu	Tyr	Tyr	G y	
		35					40					45				
Cys	G y	I l e	Arg	Ser	Pro	Ser	Ser	G u	G n	I l e	G n	Arg	I l e	Thr	Al a	
	50					55					60					
Met	Leu	Arg	G n	Hi s	G y	Lys	I l e	G u	G y	Lys	Asn	Val	Phe	Tyr	Tr p	
65					70					75					80	
Phe	G n	Asn	Hi s	Lys	Al a	Arg	G u	Arg	G n	Lys	Arg	Arg	Leu	Thr	Ser	
			85						90					95		
Leu	Asp	Val	Asn	Val	Pro	Al a	Al a	G y	Al a	Al a	Asp	Al a	Thr	Thr	Ser	
			100					105					110			
G n	Leu	G y	Val	Leu	Ser	Leu	Ser	Pro	Pro	Ser	G y	Al a	Al a	Pro		
		115					120				125					
Pro	Ser	Pro	Thr	Leu	G y	Phe	Tyr	Al a	Al a	G y	Asn	G y	G y	G y	Ser	
	130					135					140					
Al a	G y	Leu	Leu	Asp	Thr	Ser	Ser	Asp	Tr p	G y	Ser	Ser	G y	Al a	Al a	

5312WOPCT_SEQ LI STI NG TXT

145 Met Ala Thr Glu Thr 150 Cys Phe Leu Glu Asp 155 Tyr Met Gly Val Thr 160 Asp
 Thr Gly Ser Ser 165 Glu Trp Pro Cys 170 Phe Ser Ser Ser Asp 175 Thr Ile
 Met Ala Ala Ala Ala Ala Ala Ala Arg Val Ala Thr Thr Arg Ala Pro
 Glu Thr Leu Pro Leu Phe Pro Thr Cys Gly Asp Asp Asp Asp Asp
 Ser Glu Pro Pro Pro Arg 215 Arg His Ala Val Pro Val Pro Ala Gly
 225 Glu Thr Ile Arg Gly 230 Gly Gly Gly Ser Ser Ser Ser Tyr Leu Pro Phe
 Trp Gly Ala Gly Ala Ala Ser Thr Thr Ala Gly Ala Thr Ser Ser Val
 Ala Ile Glu 260 Glu Glu His Glu Leu 280 Glu Glu Tyr Ser Phe Tyr Ser
 Asn Ser Thr Glu Leu Ala Gly Thr Gly Ser Glu Asp Val Ser Ala Ser
 Ala Ala Ala Leu Glu Leu Ser Leu Ser Ser Trp Cys Ser Pro Tyr Pro
 305 Ala Ala Gly Ser Met 325

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<220>
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 cct acg gcg gag cag gtg aag gtc ctg acg gag ctg ttc cgc gcg ggg 96
 Pro Thr Ala Glu Glu Val Lys Val Leu Thr Glu Leu Phe Arg Ala Gly 20 25 30
 ctg cgg acg ccc agc acg gag cag atc cag cgc atc tcc acc cac ctg 144
 Leu Arg Thr 35 Pro Ser Thr Glu Glu Ile Glu Arg Ile Ser 45 Thr His Leu 40
 agc gcc ttc ggc aag gtg gag agc aag aac gtc ttc tac tgg ttc cag 192
 Ser Ala Phe Gly Lys Val Glu Ser Lys Asn Val Phe Tyr Trp Phe Glu 50 55 60
 aac cac aag gcc cgc gag cgc cac cac cac aag aag cgc cgc cgc gcc 240
 Asn His Lys Ala Arg Glu 70 Arg His His His Lys 75 Lys Arg Arg Arg Gly 80 65
 gcg tcg tcg tcc tcc ccc gac agc ggc agc ggc agg gga agc aac aac 288
 Ala Ser Ser Ser 85 Pro Asp Ser Gly Ser 90 Gly Arg Gly Ser Asn Asn 95
 gag gaa gac ggc cgt ggt gcc gcc tcg cag tcg cac gac gcc gac gcc 336
 Glu Glu Asp Gly Arg Gly Ala Ala Ser Glu Ser His Asp Ala Asp Ala 100 105 110
 gac gcc gac ctg gtg ctg caa ccg cca gag agc aag cgg gag gcc aga 384
 Asp Ala Asp Leu Val Leu Glu Pro Pro Glu Ser Lys Arg 115 120 125 Glu Ala Arg
 agc tat ggc cac cat cac cgg ctg gtg aca tgc tac gtc agg gac gtg 432

5312WOPCT_SEQ_LI STING.TXT

Ser	Tyr	Gly	His	His	His	Arg	Leu	Val	Thr	Cys	Tyr	Val	Arg	Asp	Val	
	130					135					140					
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Val	Glu	Gln	Gln	Glu	Ala	Ser	Pro	Ser	Trp	Glu	Arg	Pro	Thr	Arg	Glu	160
	145				150					155						
gtg	gag	acg	cta	gag	ctc	ttc	ccc	ctc	aag	tcg	tac	ggc	gac	ctc	gag	528
Val	Glu	Thr	Leu	Glu	Leu	Phe	Pro	Leu	Lys	Ser	Tyr	Gly	Asp	Leu	Glu	175
				165					170					175		
gcg	gcg	gag	aag	gtc	cgg	tcg	tac	gtc	aga	ggc	atc	gcc	gcc	acc	agc	576
Ala	Ala	Glu	Lys	Val	Arg	Ser	Tyr	Val	Arg	Gly	Ile	Ala	Ala	Thr	Ser	
			180					185					190			
gag	cag	tgc	agg	gag	ttg	tcc	ttc	ttc	gac	gtc	tcc	gcc	ggc	cgg	gat	624
Glu	Gln	Cys	Arg	Glu	Leu	Ser	Phe	Phe	Asp	Val	Ser	Ala	Gly	Arg	Asp	
		195					200					205				
ccg	ccg	ctc	gag	ctc	agg	ctc	tgc	agc	ttc	ggc	ccc	tag				663
Pro	Pro	Leu	Glu	Leu	Arg	Leu	Cys	Ser	Phe	Gly	Pro					220
	210					215										

<210> 110
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 <212> PRT
 <213> Zea mays

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			20					25					30			
Leu	Arg	Thr	Pro	Ser	Thr	Glu	Gln	Ile	Gln	Arg	Ile	Ser	Thr	His	Leu	
		35				40						45				
Ser	Ala	Phe	Gly	Lys	Val	Glu	Ser	Lys	Asn	Val	Phe	Tyr	Trp	Phe	Gln	
	50					55					60					
Asn	His	Lys	Ala	Arg	Glu	Arg	His	His	His	Lys	Lys	Arg	Arg	Arg	Gly	
65				70						75					80	
Ala	Ser	Ser	Ser	Pro	Asp	Ser	Gly	Ser	Gly	Arg	Gly	Ser	Asn	Asn		
			85					90					95			
Glu	Glu	Asp	Gly	Arg	Gly	Ala	Ala	Ser	Gln	Ser	His	Asp	Ala	Asp	Ala	
		100						105					110			
Asp	Ala	Asp	Leu	Val	Leu	Gln	Pro	Pro	Glu	Ser	Lys	Arg	Glu	Ala	Arg	
		115					120					125				
Ser	Tyr	Gly	His	His	His	Arg	Leu	Val	Thr	Cys	Tyr	Val	Arg	Asp	Val	
	130					135					140					
Val	Glu	Gln	Gln	Glu	Ala	Ser	Pro	Ser	Trp	Glu	Arg	Pro	Thr	Arg	Glu	
145					150					155					160	
Val	Glu	Thr	Leu	Glu	Leu	Phe	Pro	Leu	Lys	Ser	Tyr	Gly	Asp	Leu	Glu	
				165					170					175		
Ala	Ala	Glu	Lys	Val	Arg	Ser	Tyr	Val	Arg	Gly	Ile	Ala	Ala	Thr	Ser	
		180					185					190				
Glu	Gln	Cys	Arg	Glu	Leu	Ser	Phe	Phe	Asp	Val	Ser	Ala	Gly	Arg	Asp	
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	210					215					220					

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 <211> 896
 <212> DNA
 <213> Zea mays

<400> 111
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5312WOPCT_SEQ_LI STI NG.TXT

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gt gt t t t aga	gaat cat at a	aat gaacagt	t agacat ggt	ct aaaggaca	at t gagt at t	240
t t gacaacag	gact ct acag	t t t t at ct t t	t t agt gt gca	t gt gt t ct cc	t t t t t t t t g	300
caaat agct t	cacct at at a	at act t cat c	cat t t t at t a	gt acat ccat	t t aggg t t t a	360
gggt t aat gg	t t t t t at aga	ct aat t t t t t	t agt acat ct	at t t t at t ct	at t t t agcct	420
ct aaat t aag	aaaact aaaa	ct ct at t t t a	gt t t t t t at	t t aat aat t t	agat at aaaa	480
t agaat aaaa	t aaagt gact	aaaaat t aaa	caaat accct	t t aagaaat t	aaaaaaact a	540
aggaaacat t	t t t ct t gt t t	cgagt agat a	at gccagcct	gt t aaacgcc	gt cgacgagt	600
ct aacggaca	ccaaccagcg	aaccagcagc	gt cgct cgg	gccaagcgaa	gcagacggca	660
cggcat ct ct	gt cgct gcct	ct ggaccct	ct cgagagt t	ccgct ccacc	gt t ggact t g	720
ct ccgct gt c	ggcat ccaga	aat t gcgt gg	cggagcggca	gacgt gagcc	ggcacggcag	780
gcgccct cct	cct cct ct ca	cggcaccggc	agct acgggg	gat t cct t t c	ccaccgct cc	840
t t cgct t t cc	ct t cct cgcc	cgccgt aat a	aat agacacc	ccct ccacac	cct ct t	896

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 <211> 82
 <212> DNA
 <213> Zea mays

<400> 112	t ccccaacct	cgt gt t gt t c	ggagcgcaca	cacacacaac	cagat ct ccc	ccaaat ccac	60
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<210> 113
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	at gcat ggt t	agggcccggt	agt t ct act t	ct gt t cat gt	t t gt gt t aga	t ccgt gt t t g	120
	t gt t agat cc	gt gct gct ag	cgt t cgt aca	cggat gcgac	ct gt acgt ca	gacacgt t ct	180
	gat t gct aac	t t gccagt gt	t t ct ct t t gg	ggaat cct gg	gat ggct ct a	gccgt t ccgc	240
	agacgggat c	gat t t cat ga	t t t t t t t gt	t t cgt t gcat	aggg t t t ggt	t t gccct t t t	300
	cct t t at t t c	aat at at gcc	gt gcaact t gt	t t gt cgggt c	at ct t t t cat	gct t t t t t t t	360
	gt ct t ggt t g	t gat gat gt g	gt ct ggt t gg	gcgggt cgt t c	t agat cggag	t agaat t ct g	420
	t t t caaact a	cct ggt ggat	t t at t aat t t	t ggat ct gt a	t gt gt gt gcc	at acat at t c	480
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