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[Continued on next page]

(54) Title: GSK3 POLYPEPTIDES

A

GSK-3 β 557 Construct Sequence

Met Glu Tyr Met Pro Met Glu Gly Gly Gly

Met Ser Gly Arg Pro Arg Thr Thr Ser Phe Ala Glu Ser Cys Lys Pro
1 5 10 15

Val Gln Gln Pro Ser Ala Phe Gly Ser Met Lys Val Ser Arg Asp Lys
20 25 30

Asp Gly Ser Lys Val Thr Thr Val Val Ala Thr Pro Gly Gln Gly Pro
35 40 45

Asp Arg Pro Gln Glu Val Ser Tyr Thr Asp Thr Lys Val Ile Gly Asn
50 55 60

Gly Ser Phe Gly Val Val Tyr Gln Ala Lys Leu Cys Asp Ser Gly Glu
65 70 75 80

Leu Val Ala Ile Lys Lys Val Leu Gln Asp Lys Arg Phe Lys Asn Arg
85 90 95

Glu Leu Gln Ile Met Arg Lys Leu Asp His Cys Asn Ile Val Arg Leu
100 105 110

Arg Tyr Phe Tyr Ser Ser Gly Glu Lys Lys Asp Glu Val Tyr Leu
115 120 125

Asn Leu Val Leu Asp Tyr Val Pro Glu Thr Val Tyr Arg Val Ala Arg
130 135 140

His Tyr Ser Arg Ala Lys Gln Thr Leu Pro Val Ile Tyr Val Lys Leu
145 150 155 160

Tyr Met Tyr Gln Leu Phe Arg Ser Leu Ala Tyr Ile His Ser Phe Gly
165 170 175

Ile Cys His Arg Asp Ile Lys Pro Gln Asn Leu Leu Asp Pro Asp
180 185 190

Thr Ala Val Leu Lys Leu Cys Asp Phe Gly Ser Ala Lys Gln Leu Val
195 200 205

Arg Gly Glu Pro Asn Val Ser Tyr Ile Cys Ser Arg Tyr Tyr Arg Ala
210 215 220

Pro Glu Leu Ile Phe Gly Ala Thr Asp Tyr Thr Ser Ser Ile Asp Val
225 230 235 240

Trp Ser Ala Gly Cys Val Leu Ala Glu Leu Leu Leu Gly Gln Pro Ile
245 250 255

B

Phe Pro Gly Asp Ser Gly Val Asp Gln Leu Val Glu Ile Ile Lys Val
260 265 270

Leu Gly Thr Pro Thr Arg Glu Gln Ile Arg Glu Met Asn Pro Asn Tyr
275 280 285

Thr Glu Phe Lys Phe Pro Gln Ile Lys Ala His Pro Trp Thr Lys Val
290 295 300

Phe Arg Pro Arg Thr Pro Pro Glu Ala Ile Ala Leu Cys Ser Arg Leu
305 310 315 320

Leu Glu Tyr Thr Pro Thr Ala Arg Leu Thr Pro Leu Glu Ala Cys Ala
325 330 335

His Ser Phe Phe Asp Glu Leu Arg Asp Pro Asn Val Lys His Pro Asn
340 345 350

Gly Arg Asp Thr Pro Ala Leu Phe Asn Phe Thr Gln Glu Leu Ser
355 360 365

Ser Asn Pro Pro Leu Ala Thr Ile Leu Ile Pro Pro His Ala Arg Ile
370 375 380

(57) Abstract: The invention provides truncated GSK3 polypeptides capable of crystallization, including GSK3 α and GSK3 β polypeptides, and use of these polypeptides to identify and optimize GSK3 inhibitors. Also provided are GSK3 polypeptides having at least one substituted amino acid that differs from wild-type GSK3, wherein the substituted amino acid is incapable of being phosphorylated. The invention finds use in providing methods of identifying and optimizing compounds useful for treating diseases mediated by GSK3 activity, including Alzheimer's disease, type 2 diabetes, and inflammation.

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GSK3 POLYPEPTIDES

BACKGROUND OF THE INVENTION

Field of the Invention

The invention provides materials and methods relating to identification
5 and optimization of selective inhibitors of glycogen synthase kinase 3 (GSK3), and also
relates to methods of treating a condition mediated by GSK3 activity. Such conditions
include Alzheimer's disease, type 2 diabetes, and inflammation.

Description of the Related Art

Glycogen synthase kinase 3 (GSK3) is a proline-directed
10 serine/threonine kinase originally identified as an activity that phosphorylates glycogen
synthase as described in Woodgett, *Trends Biochem Sci.* 16:177-181 (1991). The role
in glucose metabolism has been elaborated recently in Summers et al., *J. Biol. Chem.*
274:17934-17940 (1999). GSK3 consists of two isoforms, α and β , and is
constitutively active in resting cells, inhibiting glycogen synthase by direct
15 phosphorylation. Upon insulin activation, GSK3 is inactivated, thereby allowing the
activation of glycogen synthase and possibly other insulin-dependent events. GSK3 is
inactivated by other growth factors or hormones that, like insulin, signal through
receptor tyrosine kinases. Examples of such signaling molecules include IGF-1 and
EGF as described in Saito et al., *Biochem. J.* 303:27-31 (1994), Welsh et al., *Biochem.*
20 *J.* 294:625-629 (1993), and Cross et al., *Biochem. J.* 303:21-26 (1994). GSK3 has been
shown to phosphorylate β -catenin as described in Peifer et al., *Develop. Biol.*
166:543-56 (1994). Other activities of GSK3 in a biological context include GSK3's
ability to phosphorylate tau protein *in vitro* as described in Mandelkow and
Mandelkow, *Trends in Biochem. Sci.* 18:480-83 (1993), Mulot et al., *Febs Lett* 349:
25 359-64 (1994), and Lovestone et al., *Curr. Biol.* 4:1077-86 (1995), and in tissue culture
cells as described in Latimer et al., *Febs Lett* 365:42-6 (1995). Selective inhibition of
GSK3/may be useful to treat or inhibit disorders mediated by GSK3 activity.

There is a need in the art for compositions and molecules that bind to or interact with GSK3, thereby mediating GSK3 activity. The invention meets this need by providing crystallizable GSK3 polypeptides useful for design and optimization of GSK3 inhibitors.

5 BRIEF SUMMARY OF THE INVENTION

The invention provides GSK3 β molecules with N- and C-terminal truncations, wherein the molecules are capable of crystallization.

The invention further provides GSK3 β molecules truncated at amino acid R³⁴⁴, R³⁵⁴, T³⁶⁴, A³⁷⁴, and I³⁸⁴.

10 The invention provides a polypeptide consisting essentially of SEQ ID NO:2 or SEQ ID NO:3, polynucleotides encoding these polypeptides, and vectors comprising these polynucleotides.

The invention still further provides GSK3 β molecules wherein translation of the molecule begins at G³⁴, T³⁹, P⁴⁴, D⁴⁹ or V⁵⁴.

15 The invention also provides GSK3 α molecules with N- and C-terminal truncations, wherein the molecules are capable of crystallization.

The invention further provides a GSK3 α molecule wherein translation of the molecule begins at S⁹⁷ and ends at S⁴⁴⁷, polynucleotides encoding this polypeptide, and vectors comprising these polynucleotides.

20 The invention further provides a method of identifying a GSK3 polypeptide capable of crystallization, comprising: (a) providing a truncated GSK3 polypeptide; (b) testing the polypeptide for formation of crystals.

The invention also provides GSK3 polypeptides capable of interacting with inhibitors of GSK3.

25 The invention further provides a method of identifying an enzymatically active GSK3 polypeptide, comprising: (a) providing a truncated GSK3 polypeptide; (b) contacting the polypeptide with a substrate of GSK3; and (c) measuring the kinase activity of the polypeptide after contacting the polypeptide with the substrate, wherein

the polypeptide is active if it shows $>0.01 \times$ the activity of the full-length enzyme and preferably $>0.1 \times$ the activity of the full-length enzyme.

BRIEF DESCRIPTION OF THE DRAWINGS AND SEQUENCE IDENTIFIERS

Figure 1 provides the polypeptide sequence of human GSK3 β (SEQ ID
5 NO:1).

Figure 2 provides the polypeptide sequence of truncated GSK3 β polypeptide 557 (SEQ ID NO:2). The first ten amino acids represent a Glu-tag, followed by a Gly linker before Met at position 1.

Figure 3 provides the polypeptide sequence of truncated GSK3 β
10 polypeptide 580 (SEQ ID NO:3). The first ten amino acids represent a Glu-tag, followed by a Gly linker before Gly at position 34.

Figure 4 provides the polypeptide sequence of human GSK3 α (SEQ ID NO:4).

Figure 5 provides the polypeptide sequence of human GSK3 α truncated
15 at position 447 (SEQ ID NO:5).

Figure 6 provides the polypeptide sequence of human GSK3 α truncated at position 97 (SEQ ID NO:6).

Figure 7 provides the polypeptide sequence of human GSK3 α from position 97 to position 447 (SEQ ID NO:7).

20 DETAILED DESCRIPTION OF THE INVENTION

The invention provides materials and methods for identifying and optimizing inhibitors of GSK3, including GSK3 α and GSK3 β . The provided materials include C- and N-terminal truncated GSK3 β molecules that are capable of crystallization and may, but need not, retain GSK3 kinase activity, preferably more than
25 $0.01 \times$ the activity of the full-length enzyme and more preferably more than $0.1 \times$ the activity of the full-length enzyme. There is a need in the art for such inhibitors, in view of the role of GSK3 in a variety of diseases and conditions, including Alzheimer's disease, type 2 diabetes and inflammation. Such inhibitors can be identified, and

identified inhibitors can be optimized, using the crystallizable GSK3 polypeptides of the invention.

The invention provides a variety of GSK3 β polypeptides that differ from the native polypeptide at the C- and/or N-terminus. The amino acid sequence of GSK3 β is shown in Figure 1 (SEQ ID NO:1). Included within the scope of the invention are any and all truncations of GSK3 β polypeptide wherein the truncated polypeptide is capable of crystallization and may, but need not, retain kinase activity as measured using the kinase assays described herein. Persons of skill in the art will realize that limited mutation of the protein, or certain post-translational modifications, might be sufficient to inactivate the kinase yet retain the essential 3D structure. Such inactive but structurally related molecules would also be useful for the design and optimization of inhibitors. Kinase assays are disclosed in U.S. Patent Nos. 6,057,117 and 6,057,286, which are incorporated herein by reference. The percent activity that is retained, if any, is not crucial. Methods of assaying activity in the presence and absence of an inhibitor are described herein.

The invention provides numerous truncated GSK3 β polypeptides that meet these criteria. A preferred polypeptide is designated BV557 in which the C-terminal amino acid is R³⁸⁴. This molecule has been successfully crystallized. Additional active polypeptides include those with truncations at amino acid R³⁴⁴, R³⁵⁴, A³⁷⁴, and I³⁸⁴.

The invention also provides truncated GSK3 α polypeptides, including a GSK3 α polypeptide beginning at S⁹⁷ and ending at S⁴⁴⁷.

Additional truncated GSK3 polypeptide include those beginning with an N-terminal amino acid that differs from that of the native protein in that 1 or more amino acids are deleted from the N-terminus. Preferred N-terminal truncations include GSK3 β molecules wherein translation of the molecule begins at G³⁴, T³⁹, P⁴⁴, D⁴⁹ or V⁵⁴. An example is BV580 (amino acids 34 to 384) which has been crystallized.

The invention is not limited to these disclosed truncated molecules. Using the methods and assays described herein, one of skill can construct additional truncated molecules, such as those having 36-76 amino acids deleted from the C-

terminus, and/or 35-54 amino acids deleted from the N-terminus. Such deletions can occur individually, or a polypeptide can have both an N-terminal deletion and a C-terminal deletion. It is preferable but not necessary that the kinase domain remain relatively intact as reflected by the detection of enzymatic activity, such as by using the
5 assays described herein. It is also desirable, although not essential, that the enzymatic activity be capable of inhibition by a known GSK3 inhibitor, such as lithium. A truncated molecule meeting these criteria will be suitable for testing GSK3 inhibitors as potential therapeutic agents, and for optimizing GSK3 inhibitors.

A truncated GSK3 β polypeptide of the invention can consist of between
10 about 250 and 419 contiguous amino acids of SEQ ID NO:1; preferably between about 278 and 419 contiguous amino acids of SEQ ID NO: 1; more preferably between about 285 and 384 contiguous amino acids of SEQ ID NO:1; and most preferably between about 351 and 384 contiguous amino acids of SEQ ID NO:1. Preferred truncated GSK3 β polypeptides include those beginning at amino acid 1, 2, 3, 4, 5, 6, 7, 8, 9, 10,
15 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, 50, 51, 52, 53, 54, 55, 56, 57, 58, 59, 60, 61, or 62 of SEQ ID NO:1, and ending at amino acid 340, 341, 342, 343, 344, 345, 346, 347, 348, 349, 350, 351, 352, 353, 354, 355, 356, 357, 358, 359, 360, 361, 362, 363, 364, 365, 366, 367, 368, 369, 370, 371, 372, 373, 374, 375, 376, 377,
20 378, 379, 380, 381, 382, 383, 384, 385, 386, 387, 388, 389, 390, 391, 392, 393, 394, 395, 396, 397, 398, 399, 400, 401, 402, 403, 404, 405, 406, 407, 408, 409, 410, 411, 412, 413, 414, 415, 416, 417, 418 or 419 of SEQ ID NO:1. The polypeptide can begin with any one of the listed beginning amino acids and end with any one of the ending amino acids. Exemplary and non-limiting embodiments begin at amino acid 34, 39, 44
25 or 54 and end at amino acid 420. Other particularly preferred embodiments begin at about amino acid 1 and end at amino acid 340, 344, 354, 374, 384, 385, 386, 387, 388, 389, 390, 391, 392, 393, 394, 395, 396, 397, 398, 399, 400, 401, 402, 403, 404, 405, 406, 407, 408, 409, 410, 411, 412, 413, 414, 415, 416, 417, 418, 419, or 420.

The truncated GSK3 α polypeptide of the invention can consist of
30 between about 182 and 482 contiguous amino acids of SEQ ID NO: 4, preferably

between about 182 and 386 contiguous amino acids of SEQ ID NO:4, more preferably between about 182 and 351 contiguous amino acids of SEQ ID NO:4, and most preferably from about S⁹⁷ to S⁴⁴⁷ of SEQ ID NO:4.

The truncated GSK3 polypeptides can be prepared by any method known in the art. One method involves expression of a suitably prepared polynucleotide encoding a polypeptide having the desired truncation. For example, a preferred polypeptide of the invention, BV557, was prepared by creating a construct encoding GSK3 β starting at M¹ and ending at I³⁸⁴, as described in the Examples. Briefly, insect cells were transfected with baculovirus vector (designated pBlueBac4.5.Glu.GSK3B.DC.I384#28), which encodes BU557, and the protein was extracted from the lysed cells. The protein was purified by affinity chromatography using an anti glu-tag monoclonal antibody immobilized on a Sepharose column. Activity of the purified protein was assayed using the *in vitro* kinase assay described in U.S. Patent No. 6,057,286.

The Examples herein describe the production of BV557, BV580, and other truncated GSK3 polypeptides by expression of vectors encoding the polypeptides, followed by isolation and purification of the polypeptides. The polypeptide can also be produced by enzymatic cleavage of a native GSK3 protein, using methods known in the art. Other suitable methods include expression of a polynucleotide encoding a truncated polypeptide in a variety of cell types, including mammalian, bacterial, or yeast cells. However, the preferred cell for expression of the polypeptide is an insect cell, preferably a baculovirus-infectable insect cell, such as a Sf9 cell.

The invention also provides unphosphorylated forms of GSK3 wherein the ATP binding site is identical to that of the wild-type protein. Such forms include Y216 non-phosphorylated GSK3 β and Y279 non-phosphorylated GSK3 α . Other forms include constructs with at least one amino acid change that prevents phosphorylation, such as GSK3 β in which Y216 is changed to F216, and GSK3 α in which Y279 is changed to F279. These forms are suitable for inhibitor binding assays to identify inhibitors of GSK3. The invention provides a GSK3 β molecule in which position 216 is not phosphorylated. We have demonstrated that a GSK3 β peptide with Y216 mutated

to F216 crystalized and exhibits a structure in which the ATP-binding site is not substantially different from the un-mutated peptide.

Additional single and multiple amino acid changes include S⁹ to A⁹ in GSK3 β and S²¹ to A²¹ in GSK3 α .

5 These changes in phosphorylation, or ability to be phosphorylated, are optionally incorporated into the truncated forms of GSK3 α and GSK3 β disclosed herein.

The invention therefore provides GSK3 molecules suitable for design and optimization of inhibitors of GSK3 as pharmaceutical agents.

10 The GSK3 constructs of the invention are capable of crystallization. In purified form the constructs bind to inhibitors in a manner that is comparable to inhibitor binding to the native GSK3 polypeptide, due to the retention of the correct folding conformation at the inhibitor binding site. Potential to crystallize is measured using a variety of assays including specific activity, aggregation, microheterogeneity.

15 (See, for example, Table 1). These parameters are indicative of the purity of the preparation and of the solubility of the construct. The specific activity is also a preferred assay for detecting binding of an inhibitor to the correct binding site of the GSK3 construct. Another suitable method is fluorescence polarization. Briefly, a putative inhibitor, with an attached fluorophore, tumbles freely in solution. Thus when

20 the fluorophore is excited by polarized light, the emitted light which is produced after a finite delay now has random polarity and the emitted light is no longer polarized. In the presence of a GSK3 construct with an intact inhibitor binding site, the tumbling rate is slowed sufficiently to ensure that, even though the light emission is delayed with respect to the excitation, the fluorophore has only moved very slightly. Thus, the

25 excited light maintains polarization. A measurement of fluorescence polarization therefore indicates whether or not the GSK3 construct is suitable for identifying and optimizing an inhibitor. The fluorophore can be attached to a compound such as staurosporine (ICN Pharmaceuticals, Inc., Costa Mesa, CA). GSK3 constructs may not retain kinase activity, but their inhibitor binding can still be assessed using fluorescence

30 polarization assays.

The term "truncated glycogen synthase kinase 3" or "truncated GSK3" as used herein refers to GSK3 α or GSK3 β . GSK3 is a protein originally identified by its phosphorylation of glycogen synthase as described in Woodgett et al, *Trends Biochem Sci*, 16:177-181 (1991). Synonyms of GSK3 are tau protein kinase I (TPK I), FA kinase and kinase FA. Mammalian forms of GSK3 have been cloned as described in Woodgett, *EMBO J.* 9(8):2431-2438 (1990). Inhibitors of truncated GSK3 polypeptides can be inhibitors of any of the known forms of GSK3, including either GSK3 α or GSK3 β or both. Truncated polypeptides of the invention possess one or more of the bioactivities of the GSK3 protein, including kinase activities such as polymerizing tau protein, or phosphorylating glycogen synthase, for example. Thus, truncated GSK3 polypeptides useful for designing and optimizing inhibitors of GSK3 can have sequence identity of at least 40%, preferably 50%, preferably 60%, preferably 70%, more preferably 80%, and most preferably 90% to the amino acid sequence of the native protein, wherever derived, from human or nonhuman sources. The polynucleotides encoding a GSK3 polypeptide can have 60%, preferably 70%, more preferably 80%, more preferably 90% and most preferably 95% sequence identity to a native polynucleotide sequence of GSK3. Also included, therefore, are alleles and variants of the native polynucleotide sequence such that the polynucleotide encodes an amino acid sequence with substitutions, deletions, or insertions, as compared to the native sequence.

The term "peptide substrate" refers to a peptide or a polypeptide or a synthetic peptide derivative that can be phosphorylated by GSK3 activity in the presence of an appropriate amount of ATP or a phosphate donor. Detection of the phosphorylated substrate is generally accomplished by the addition of a labeled phosphate that can be detected by some means common in the art of labeling, such as radiolabeled phosphate. The peptide substrate may be a peptide that resides in a molecule as a part of a larger polypeptide, or may be an isolated peptide designed for phosphorylation by GSK3.

As disclosed in U.S. Patent Nos. 6,057,117 and 6,057,286, *in vitro* methods of assaying GSK3 activity include constructing peptide substrates. The

peptide substrate can be any peptide substrate phosphorylatable by GSK3, and may be a peptide substrate including the formula: anchor ligand-(X)_nSXXXS(X)_m (SEQ ID NO:8) (wherein X is any amino acid, n is any integer, m is any integer, and preferably n + m + 5 < 20, i.e. n + m < 15) prephosphorylated at the C terminal serine. The assay is performed by contacting the prephosphorylated substrate with truncated GSK3 polypeptide in the presence of radiolabeled γphosphate-ATP, a substrate anchor, and optionally a candidate inhibitor. The *in vitro* method of identifying an inhibitor of GSK3 kinase activity includes contacting a peptide substrate coupled to an anchor ligand with truncated GSK3 polypeptide in the presence of radiolabeled γphosphate-ATP, a substrate anchor, and candidate inhibitor, measuring an incorporation of radiolabel into the peptide substrate, then, in a separate assay vessel contacting a peptide substrate coupled to an anchor ligand with truncated GSK3 in the presence of radiolabeled γphosphate-ATP, and a substrate anchor, and measuring incorporation of radiolabel into said peptide substrate; ultimately an inhibitor of truncated GSK3 kinase activity is identified by a reduction of label incorporation in the assay with the candidate inhibitor as compared to the assay without the candidate inhibitor.

To conduct the *in vitro* kinase assay of the invention using microwells, scintillant may be present by pre-coating the wells with a scintillant material, or by adding it later following a wash step, as described in Example 4. The scintillant can be obtained from Packard, Meridian, Conn. Wells coated with scintillant are then in addition coated with streptavidin as a substrate anchor, where biotin is the anchor ligand on the peptide. Alternatively, the streptavidin can be present on agarose beads containing scintillant or may be coated on an otherwise untreated plate to which scintillant is added subsequently. In any event, the streptavidin in the wells binds the biotin that contacts it. Following an assay using radiolabeled ATP, the radiolabel incorporated into the phosphorylated substrate that has been conjugated to the biotin will cause the scintillant to emit light. Where the streptavidin is attached to agarose beads containing scintillant, binding a biotin-conjugated radiolabeled peptide substrate will cause the beads to scintillate. In both the case of the wells lined with the scintillant, and the agarose beads containing scintillant, a reduction in scintillation as

compared to a control amount of scintillation measured under non-inhibitory conditions, indicates the presence of a functional inhibitor of GSK3 activity. If the peptide has been phosphorylated by GSK3 with ^{32}P -labeled or ^{33}P -labeled phosphate, radioactive decay will cause the scintillant present in a microwell or mixed in agarose
5 beads that are present in the reaction mixture to emit light and the measure of the amount of light emitted will be a measure of the activity of GSK3 in the assay. Low activity of GSK3 observed in the presence of a candidate inhibitor, as compared to the activity of GSK3 in the absence of the inhibitor, may indicate that the inhibitor is functional and can inhibit GSK3 kinase activity. In any case, an excess of streptavidin
10 over peptide should be loaded into each well or should be affixed to the agarose beads.

GSK3 inhibitory activity can be measured using a cell-free assay as disclosed in publication WO 99/65897, and as described in Example 4 herein. Activity can also be measured using a cell-based assay. Briefly, a cell line, such as a Cos cell line, is transfected with Tau and with a GSK3 polypeptide. The phosphorylation of Tau
15 at a specific site is monitored using a monoclonal antibody, as phosphorylation at that site is dependent on GSK3 activity.

Exemplary polypeptides of the invention include the following truncated polypeptides with reference to SEQ ID NO:1:

20 GSK3 β truncated at R³⁴⁴
GSK3 β truncated at R³⁵⁴
GSK3 β truncated at T³⁶⁴
GSK3 β truncated at A³⁷⁴
GSK3 β truncated at I³⁸⁴
25 GSK3 β beginning at G³⁴
GSK3 β beginning at T³⁹
GSK3 β beginning at P⁴⁴
GSK3 β beginning at D⁴⁹
GSK3 β beginning at V⁵⁴

The above truncations can be combined, providing a GSK3 β polypeptide beginning at any of G³⁴, T³⁹, P⁴⁴, D⁴⁹, or V⁵⁴, and ending at any of R³⁴⁴, R³⁵⁴, T³⁶⁴, A³⁷⁴, or I³⁸⁴.

Other exemplary polypeptides of the invention include the following
5 truncated polypeptides with reference to SEQ ID NO:4:

GSK3 α truncated at S⁴⁴⁷.

GSK3 α beginning at S⁹⁷.

GSK3 α beginning at S⁹⁷ and truncated at S⁴⁴⁷.

10 A truncated GSK3 polypeptide of the invention can be selected on the basis of one or more parameters. A polypeptide will preferably crystallize in a form that is similar to that of native GSK3, with correct folding at and around the inhibitor binding site. Crystallization can be performed using a Crystal Screen Kit (Hampton Research, Laguna Niguel, CA), or methods described by Jancarik, J. et al., *J. Appl.*
15 *Cryst.* 24:409-411, 1991. The potential of a polypeptide to form crystals can be evaluated on the basis of specific activity, purity, homogeneity, mass spectrometry, aggregation, and dynamic light scattering. A preferred truncated polypeptide will meet the following parameters: purity of at least 90%; less than 100% aggregation at 4°C at two weeks; and less than 50% heterogeneity (50% or greater of the desired
20 form). A most preferred truncated polypeptide will have a purity of at least 98%, no aggregation at 4°C at two weeks; and less than 5% heterogeneity (unphosphorylated form). Such parameters indicate that the polypeptide preparation is likely to crystallize, making it suitable for discovering and optimizing GSK3 inhibitors.

A prerequisite for crystallization is to obtain a sufficiently concentrated
25 stock of protein. Not all GSK3 constructs will remain soluble at the required concentration. A preferred concentration is >1 mg/ml, more preferred is >5 mg/ml, and most preferred is >10 mg/ml.

The polypeptides disclosed herein as 557 (SEQ ID NO:2), 580 (SEQ ID NO:3), 458, and 524 meet the criteria described above (see Example 3). Polypeptide
30 458 consists of amino acids 1-420 of SEQ ID NO:1 plus the following addition at the

N-terminus: EFMPTEAMAAPKRVI (SEQ ID NO:8). Polypeptide 524 consists of amino acids 1-420 of SEQ ID NO:1 plus the following addition at the N-terminus: EYMPMEGGG (SEQ ID NO:9). Other modified or truncated GSK3 polypeptides can be prepared and tested as described herein.

EXAMPLES

The following examples are exemplary only, and are not intended to
5 limit the invention.

EXAMPLE 1

PREPARATION AND PURIFICATION OF GSK3 β CONSTRUCT 557

10 Lysis and Extraction. Insect cell slurry from Sf9 cells (about 10 g) from
a 1 liter flask growth was combined with 30 ml of lysate buffer: 20mM Tris, pH 8.0 /
80mM NaCl / 1mM MgCl₂ / 1mM Arsenate / 1mM Tungstenate / 1mM PMSF / 0.5mg
Leupeptin / 0.2 mg Aprotinin. Cells were lysed using a Dounce homogenizer.
Improved extraction of the protein was accomplished by the addition of 5% glycerol
15 and 0.2% octylglucoside. The mixture was allowed to stir, on ice, for 30 minutes. The
total lysate was centrifuged at 39000 x g for 25 minutes at 4°C. The resulting
supernatant contained the extracted GSK3- β #557.

Ion Exchange Chromatography. The following materials and conditions
were used: The resin was Fractogel EMD SO₃- (M); the column diameter was 1.6cm
20 and the column volume was 10ml. The column was run at a flowrate of 90cm/hour
using equilibration buffer of 20mM Na Phosphate / 5% Glycerol, pH 7.5.
Chromatography was carried out at 4°C.

 The lysate supernatant was diluted 1:1 with S-fractogel equilibration
buffer, and loaded onto the equilibrated column. The column was washed with a total
25 of 14 column volumes of equilibration buffer. The GSK3- β was eluted with a linear
salt gradient, over 20 column volumes, to equilibration buffer plus 1M NaCl.
3ml/fraction was collected during gradient elution. The pool was made based on
SDS-PAGE and Western blot results of the fractions collected. Fractions 13-24 were
pooled.

Affinity Chromatography was performed using the following materials and procedures: The resin was anti glu-tag monoclonal antibody immobilized onto Protein G Sepharose, and the equilibration buffer was PBS / 0.3M NaCl / 0.2% octylglucoside / 10% Glycerol. The column diameter was 1.6cm and the column
5 volume was 13ml. The flow rate was 30cm/hour during load and wash, and 15cm/hour during elution.

The S-Fractogel pool was loaded at 30cm/hour onto equilibrated column. The column was washed down to absorbance baseline with approximately 6 column volumes of equilibration buffer, and GSK3 β was eluted with 50 ml of equilibration
10 buffer containing 2mg of elution peptide (EYMPTD). The flow rate during elution was lowered to 15cm/hour. 2ml/fractions were collected during the elution. Based on SDS-PAGE results, elution fractions 6-17 were pooled with a total volume of 24 ml.

Final Yield. The affinity column pool, at a concentration of 0.17mg/ml, contained 4.1 mg of GSK3 β #557. This translates to a final yield of 4.1mg purified 557
15 / liter of growth. Purity, after this 2 column purification, was estimated at >95% by visual inspection of SDS-PAGE results.

EXAMPLE 2

PREPARATION AND PURIFICATION OF GSK3 β CONSTRUCT 580

20

Extraction. SF9 cell paste from a 10 L fermentation was washed with 100 mL PBS (10 mM NaPi, pH7.5, 150 mM NaCl) and then resuspended with 300 mL of Buffer H (20 mM Tris, pH 7.5, 1 mM Tungstate, 1 mM Arsenate, 5 mM DTT, 10 μ g/mL Leupeptin, 1 μ g/mL pepstatin A, 10% glycerol, 0.35% Octyl glucoside, 1 mM
25 Mg²⁺). Cells were homogenized in a 100-mL Dounce Homogenizer (20 strokes with pestle B). The combined homogenate was centrifuged in a Ti45 rotor at 40,000 rpm for 35 minutes to remove cell debris and nuclei. The supernatant from the centrifugation were carefully decanted and filtered through 0.45 μ filter.

S-Fractogel. 100 mL S-Fractogel (EM Science, Cat #18882) was packed
30 into a 3.2 cm x 12.5 cm column and equilibrated with > 1 L of buffer A (20 mM Tris, pH 7.5, 10% glycerol). The filtrate from the previous step was loaded at 15 mL/min

onto the column. The column was washed with 1 L of buffer A and then eluted with a linear gradient from 0 to 1 M NaCl in buffer A over 20 column volumes. The eluant was fractionated into 20 mL each. Fractions containing GSK3 were detected by Western Blot using anti-GSK antibody (Santa Cruz Biotech, Cat # SC-7291). The Western-Blot positive fractions were pooled and mixed with equal volume of buffer M (20 mM Tris, pH 7.5, 10% glycerol, 3.1 M NaCl) and filtered through a 0.45 μ filter. The filtrate was saved for Phenyl-650 M chromatography.

Phenyl-650 M. 37.5 mL Phenyl-650 M (Tosohass, Cat # 014943) was packed into a 2.2 x 10 cm column and equilibrated with 500 mL of buffer C (20 mM Tris, pH 7.5, 10% glycerol, 1.6 M NaCl). Filtrate from S-fractogel step was loaded onto the column at 7.5 mL/min. After the loading was completed, the column was washed with 6.5 cv buffer C and eluted with a linear gradient from 0% to 100% Buffer D (20 mM Tris, pH 7.5, 10% glycerol) over 20 column volumes. Fractions were collected at 15 mL each and GSK containing fractions were detected by Western Blot using anti-GSK antibody. The Western positive fractions were pooled and loaded onto a Glu-tag antibody affinity column.

Glu-tag antibody Affinity Chromatography. Use of a Glu-tag is described in Rubinfeld et al., *Cell* 65:1033-1042, 1991, and a hybridoma expressing anti-Glu-tag antibody is described in Grussenmyer et al., *PNAS* 82:7952-7954 (1985). 50 mg of the Glu-tag antibody was immobilized onto 25 mL of Affi-Gel 10 (BioRAD, Cat #153-6046) and packed into a 2.2 x 6.5 cm column. The column was equilibrated with 200 mL of buffer E (20 mM Tris, pH 7.5, 10% glycerol, 0.3 M NaCl, 0.2% Octylglucoside) and the fraction pool from the Phenyl-650 M step was loaded at 1.0 mL/min. After the loading was completed, the column was washed with 100 mL of buffer E and then eluted with 60 mL Glu-tag peptide (100 μ g/mL) in Buffer E and fractionated into 5 mL each. GSK containing fractions were detected with SDS-PAGE and Coomassie Blue staining. These fractions were pooled and concentrated to approximately 6 mg/mL in an Amicon concentrator using a 10 k MWCO YM10 membrane. The concentrated material was then ready for crystallization.

EXAMPLE 3

ACTIVITY OF TRUNCATED GSK3 β POLYPEPTIDES

- 5 A reaction mixture was prepared containing 5.9 μ M prephosphorylated SGSG-linked CREB peptide (Wang et al., *Anal. Biochem.*, 220:397-402 (1994)) μ in reaction buffer (5mM Tris, pH 7.5, 5mM DTT; 1mM MgCl₂, 0.01% BSA) containing the desired amount of truncated GSK3 polypeptide. ATP was added (specific activity 5.3 Ci/mmol) to 25 μ M final concentration and the mixture was incubated for 20 min. at
- 10 room temperature. The reaction was stopped by transferring 30 μ l onto a P81 filter disc (Whatman). The disc was washed four times in 150 ml of 75 mM H₃PO₄ for 5 minute each. The filter was air dried and counted under 5 ml scintillation fluid. The specific activity was counted by determining the ratio of counts (in cpm) by the mass of GSK3 in the reaction (in μ g).
- 15 The specific activity for construct 557 was 4.3×10^7 cpm/ μ g; for construct 458, 2.8×10^7 cpm/ μ g; and for construct 524, 2.2×10^7 cpm/ μ g.

Table 1

Construct	Purity	Mean Specific Activity	N	Concentration	Aggregation at 4 Degrees	Aggregation at RT	Heterogeneity
		cpm/ μ g		mg/ml			%
458	>98%	2.8×10^7	35	11.5	11% @ > 2 weeks	overnight	10-20% unphosphorylated
557	>98%	4.3×10^7	7	12.7	none @ > 2 weeks	overnight	5% unphosphorylated
524	>98%	2.2×10^7	24	10	ND		< 5% unphosphorylated

N = number of assays used to determine "mean specific activity."

EXAMPLE 4

SCREENING FOR GSK3 INHIBITORY ACTIVITY USING A CELL-FREE ASSAY

Compounds to be tested as GSK3 inhibitors are dissolved in DMSO, then tested for inhibition of human GSK3 β . Expression of GSK3 β is described, for example, in Hughes et al., *Eur. J. Biochem.*, 203:305-11 (1992), which is incorporated herein by reference. An aliquot of 300 μ l of substrate buffer (30 mM tris-HCl, 10 mM MgCl₂, 2 mM DTT, 3 μ g/ ml GSK3 β) and 0.5 μ M biotinylated prephosphorylated SGSG-linked CREB peptide (Chiron Technologies PTY Ltd., Clayton, Australia) is dispensed into wells of a 96 well polypropylene microtiter plate. 3.5 μ l /well of DMSO containing varying concentrations of each compound to be assayed or staurosporine (a known kinase inhibitor used as a positive control, or a negative control) (*i.e.*, DMSO only), is added and mixed thoroughly. The reactions is then initiated by adding 50 μ l/well of 1 μ M unlabeled ATP and 1-2 x 10⁷ cpm γ ³³P-labeled ATP, and the reaction is allowed to proceed for about three hours at room temperature.

While the reaction is proceeding, streptavidin-coated Labsystems "Combiplate 8" capture plates (Labsystems, Helsinki, Finland) are blocked by incubating them with 300 μ l/well of PBS containing 1% bovine serum albumin for at least one hour at room temperature. The blocking solution is then removed by aspiration, and the capture plates are filled with 100 μ l/well of stopping reagent (50 μ M ATP/20 mM EDTA).

When the three hour enzyme reaction is finished, triplicate 100 μ l aliquots of each reaction mix are transferred to three wells containing stopping solution, one well on each of the three capture plates, and the well contents are mixed well. After one hour at room temperature, the wells of the capture plates are emptied by aspiration and washed five times using PBS and a 12 channel Corning 430474 ELISA plate washer. Finally, 200 μ l of Microscint-20 scintillation fluid is added to each well of the plate. The plates are coated with plate sealers, then left on a shaker for 30 minutes. Each capture plate is counted in a Packard TopCount scintillation counter

(Meridian, Connecticut) and the results are plotted as a function of compound concentration.

Compounds identified using this method can be further optimized by testing their ability to bind to truncated GSK3 polypeptides of the invention, using the
5 fluorescence polarization assay, for example, for truncated polypeptides that do not exhibit GSK3 kinase activity. Alternatively, a truncated GSK3 polypeptide of the invention can be used in place of the native GSK3 protein.

EXAMPLE 5

10 SCREENING FOR INHIBITION OF TAU PROTEIN PHOSPHORYLATION

A. Transient Transfection of COS Cells with Expression Plasmid Encoding Truncated GSK3 and Tau Expression Plasmid Construction

COS cells are maintained in T25 tissue culture flasks in high glucose MEM medium / 5% fetal bovine serum. Cells from a confluent T25 flask are harvested
15 and 80,000 cells/well are seeded into Corning 6-well tissue culture plates in a final volume of 2 ml/well of medium. The cells are left to grow at 37°C for 48 hours. The cells are then washed twice in Opti-MEM containing no fetal bovine serum, and finally the cells are left in 1 ml of Opti-MEM.

Polynucleotide encoding tau protein is subcloned into plasmid pSG5
20 under an early SV40 promoter to generate a tau expression plasmid. The cloning of cDNA encoding tau protein is generally described in Goedert et al., *EMBO Journal*, 8(2):393-399 (1989), which is incorporated herein by reference. A GSK3 expression plasmid is prepared by subcloning polynucleotide encoding truncated GSK3 into pCG, which is an ApEVRF derivative described in Giese et al., *Genes & Development*,
25 9:995-1008 (1995) and Matthias et al., *Nucleic Acid Research*, 17:6418 (1989), both of which are incorporated herein by reference. The polynucleotide can encode any of the truncated GSK3 polypeptides of the invention.

The following solutions are prepared in 1.5 ml Eppendorf tubes:

Solution A: for each transfection, 2 µg of DNA (tau expression plasmid) and 0.7 µg of DNA (GSK3 expression plasmid) are diluted into 100 µl of Opti-MEM (Gibco BRL); Solution B: for each transfection, 8 µl of Lipofectamine reagent is diluted into 100 µl of Opti-MEM. The two solutions are combined, mixed gently, and
5 incubated at room temperature for 45 minutes to allow DNA-liposome complexes to form. For each transfection, 0.8 ml of Opti-MEM is added to the tube containing the complexes. The diluted solution is mixed gently and overlaid onto the rinsed cells. The cells are incubated with the complexed DNA / Lipofectamine for 6 hours at 37°C in a CO₂ incubator. Following incubation, 1 ml of growth medium (high glucose MEM)
10 with 20% FBS is added to each well and incubated at 37°C overnight. The medium is replaced with fresh, complete medium at 18 hours following the start of transfection, and the cells are left to grow at 37 °C for another 48 hours.

B. Tau Phosphorylation Inhibition assay

Two hours before harvesting, 2 µl of GSK3 inhibitor dissolved in
15 DMSO is added to each well and incubated at 37°C. After 2 hours the medium is removed and the cells are rapidly frozen on the plates on dry ice and stored at -70°C. Cells are thawed on ice in the presence of 200 µl of lysing buffer (1% Triton[®] X-100, 20 mM Tris pH 7.5, 137 mM NaCl, 15% glycerol, 25 µg/ml leupeptin, 1 µg ml pepstatin-A, 1 µM PMSF, 21 µg/ml aprotinin, 50 mM NaF, 50 mM β-
20 glycerophosphate, 15 mM sodium pyrophosphate, 1 mM sodium orthovanadate). The contents of each well are centrifuged at 14,000 g, 4°C for 5 minutes and the supernatants transferred to clean tubes. At this point the lysates may be stored at -20°C.

C. ELISA to detect phosphorylated tau in cell lysates

Immulon 4 strips (Dynatech) are coated with monoclonal anti-
25 phosphorylated tau (AT8, Polymedco, Inc.) at 5 µg/ml in PBS containing Ca⁺⁺ and Mg⁺⁺, 100 µl/well. After overnight incubation at 4°C, the strips are washed twice with washing buffer (PBS containing 0.05% Tween[®] 20) and blocked with PBS containing 1% BSA, 5% normal mouse serum and 0.05% Tween[®] 20 at room temperature for 1

hour. The strips are washed 5 times with washing buffer. Lysate (100 μ l) diluted 1:10 in PBS containing 1% BSA, 0.1% NaN_3 is added into each well and incubated at room temperature for 1 hour. After washing, 100 μ l of 0.5 μ g/ml biotinylated monoclonal anti-(non-phosphorylated) tau (HT7, Polymedco, Inc.) in PBS-BSA is added into each
5 well. Strips are washed 5 times and HRP-conjugated streptavidin is added, incubated at room temperature for 30 minutes and washed extensively with washing buffer. TMB substrate (Pierce) is used for color development and the reaction is stopped by adding an equal volume of 0.8 M sulfuric acid. Strips are read on an ELISA plate reader using a 450 nm filter. The concentration of compound that inhibits tau phosphorylation to
10 50% of the maximal level (i.e., IC_{50}) is determined by fitting a sigmoidal curve to the plotted data.

Those skilled in the art will recognize, or be able to ascertain, using not more than routine experimentation, many equivalents to the specific embodiments of
15 the invention described herein. Such specific embodiments and equivalents are intended to be encompassed by the following claims.

All patents, published patent applications, and publications cited herein are incorporated by reference as if set forth fully herein.

CLAIMS

1. An isolated nucleic acid comprising a polynucleotide encoding a polypeptide consisting essentially of SEQ ID NO:2, wherein said polypeptide will crystallize and will have at least one biological activity selected from the group consisting of (a) binding a GSK3 inhibitor; and (b) kinase activity.
2. An isolated nucleic acid comprising a polynucleotide encoding a polypeptide consisting essentially of SEQ ID NO:3, wherein said polypeptide will crystallize and will have at least one biological activity selected from the group consisting of (a) binding a GSK3 inhibitor; and (b) kinase activity.
3. A vector comprising the polynucleotide of claim 1 or claim 2.
4. A polypeptide comprising between about 250 and 419 contiguous amino acids of SEQ ID NO:1, wherein said polypeptide is phosphorylated on tyrosine 216, said polypeptide will crystallize, and said polypeptide will have at least one biological activity selected from the group consisting of (a) binding a GSK3 inhibitor; and (b) kinase activity.
5. A polypeptide consisting essentially of between about 278 and 419 contiguous amino acids of SEQ ID NO:1, wherein said polypeptide exhibits at least 1% of the kinase activity of human GSK3 β .
6. A polypeptide consisting essentially of between about 285 and 384 contiguous amino acids of SEQ ID NO:1, wherein said polypeptide exhibits at least 1% of the kinase activity of human GSK3 β .

7. A polypeptide consisting essentially of between about 351 and 384 contiguous amino acids of SEQ ID NO:1, wherein said polypeptide exhibits at least 1% of the kinase activity of human GSK3 β .

8. A polypeptide consisting of the amino acid sequence of SEQ ID NO:2.

9. A polypeptide consisting of the amino acid sequence of SEQ ID NO:3.

10. An isolated nucleic acid comprising a polynucleotide encoding a polypeptide consisting essentially of SEQ ID NO:5, wherein said polypeptide will crystallize and will have at least one biological activity selected from the group consisting of (a) binding to a GSK3 inhibitor; and (b) kinase activity.

11. The nucleic acid of claim 10 wherein said polypeptide is phosphorylated on tyrosine 279.

12. A vector comprising the polynucleotide of claim 10 or claim 11.

13. A polypeptide comprising between about 182 and 482 contiguous amino acids of SEQ ID NO:4, wherein said polypeptide will crystallize, and said polypeptide will have at least one biological activity selected from the group consisting of (a) binding a GSK3 inhibitor; and (b) kinase activity.

14. A polypeptide consisting essentially of between about 182 and 386 contiguous amino acids of SEQ ID NO:4, wherein said polypeptide exhibits at least 1% of the kinase activity of human GSK3 α .

15. A polypeptide consisting essentially of between about 182 and 351 contiguous amino acids of SEQ ID NO:4, wherein said polypeptide exhibits at least 1% of the kinase activity of human GSK3 α .
16. A polypeptide consisting essentially of contiguous amino acids S⁹⁷ to S⁴⁴⁷ of SEQ ID NO:1.
17. A polypeptide consisting essentially of the amino acid sequence of SEQ ID NO:5.
18. A polynucleotide encoding a polypeptide consisting essentially of SEQ ID NO:6.
19. A polypeptide consisting essentially of the amino acid sequence of SEQ ID NO:6.
20. A polynucleotide encoding a polypeptide consisting essentially of SEQ ID NO:7.
21. A polypeptide consisting essentially of the amino acid sequence of SEQ ID NO:7.
22. A polynucleotide encoding a non-phosphorylated human GSK3 polypeptide, wherein said non-phosphorylated polypeptide differs from native GSK3 in at least one and not more than ten amino acids.
23. The polynucleotide of claim 22 wherein tyrosine at position 216 of SEQ ID NO:1 is substituted for by a non-phosphorylatable amino acid.

24. The polynucleotide of claim 23 wherein said non-phosphorylatable amino acid is phenylalanine.

25. The polynucleotide of claim 22 wherein tyrosine at position 279 of SEQ ID NO:4 is substituted for by a non-phosphorylatable amino acid.

26. The polynucleotide of claim 25 wherein said non-phosphorylatable amino acid is phenylalanine.

FIGURE 1A

GSK-3 β Sequence

Met	Ser	Gly	Arg	Pro	Arg	Thr	Thr	Ser	Phe	Ala	Glu	Ser	Cys	Lys	Pro	1	5	10	15
Val	Gln	Gln	Pro	Ser	Ala	Phe	Gly	Ser	Met	Lys	Val	Ser	Arg	Asp	Lys	20	25	30	
Asp	Gly	Ser	Lys	Val	Thr	Thr	Val	Val	Ala	Thr	Pro	Gly	Gln	Gly	Pro	35	40	45	
Asp	Arg	Pro	Gln	Glu	Val	Ser	Tyr	Thr	Asp	Thr	Lys	Val	Ile	Gly	Asn	50	55	60	
Gly	Ser	Phe	Gly	Val	Val	Tyr	Gln	Ala	Lys	Leu	Cys	Asp	Ser	Gly	Glu	65	70	75	80
Leu	Val	Ala	Ile	Lys	Lys	Val	Leu	Gln	Asp	Lys	Arg	Phe	Lys	Asn	Arg	85	90	95	
Glu	Leu	Gln	Ile	Met	Arg	Lys	Leu	Asp	His	Cys	Asn	Ile	Val	Arg	Leu	100	105	110	
Arg	Tyr	Phe	Phe	Tyr	Ser	Ser	Gly	Glu	Lys	Lys	Asp	Glu	Val	Tyr	Leu	115	120	125	
Asn	Leu	Val	Leu	Asp	Tyr	Val	Pro	Glu	Thr	Val	Tyr	Arg	Val	Ala	Arg	130	135	140	
His	Tyr	Ser	Arg	Ala	Lys	Gln	Thr	Leu	Pro	Val	Ile	Tyr	Val	Lys	Leu	145	150	155	160
Tyr	Met	Tyr	Gln	Leu	Phe	Arg	Ser	Leu	Ala	Tyr	Ile	His	Ser	Phe	Gly	165	170	175	
Ile	Cys	His	Arg	Asp	Ile	Lys	Pro	Gln	Asn	Leu	Leu	Leu	Asp	Pro	Asp	180	185	190	
Thr	Ala	Val	Leu	Lys	Leu	Cys	Asp	Phe	Gly	Ser	Ala	Lys	Gln	Leu	Val	195	200	205	
Arg	Gly	Glu	Pro	Asn	Val	Ser	Tyr	Ile	Cys	Ser	Arg	Tyr	Tyr	Arg	Ala	210	215	220	
Pro	Glu	Leu	Ile	Phe	Gly	Ala	Thr	Asp	Tyr	Thr	Ser	Ser	Ile	Asp	Val	225	230	235	240
Trp	Ser	Ala	Gly	Cys	Val	Leu	Ala	Glu	Leu	Leu	Leu	Gly	Gln	Pro	Ile	245	250	255	
Phe	Pro	Gly	Asp	Ser	Gly	Val	Asp	Gln	Leu	Val	Glu	Ile	Ile	Lys	Val	260	265	270	

FIGURE 1B

Leu	Gly	Thr	Pro	Thr	Arg	Glu	Gln	Ile	Arg	Glu	Met	Asn	Pro	Asn	Tyr
275						280						285			
Thr	Glu	Phe	Lys	Phe	Pro	Gln	Ile	Lys	Ala	His	Pro	Trp	Thr	Lys	Val
290						295						300			
Phe	Arg	Pro	Arg	Thr	Pro	Pro	Glu	Ala	Ile	Ala	Leu	Cys	Ser	Arg	Leu
305						310						315			
Leu	Glu	Tyr	Thr	Pro	Thr	Ala	Arg	Leu	Thr	Pro	Leu	Glu	Ala	Cys	Ala
			325						330						
His	Ser	Phe	Phe	Asp	Glu	Leu	Arg	Asp	Pro	Asn	Val	Lys	His	Pro	Asn
			340						345						
Gly	Arg	Asp	Thr	Pro	Ala	Leu	Phe	Asn	Phe	Thr	Thr	Gln	Glu	Leu	Ser
355						360						365			
Ser	Asn	Pro	Pro	Leu	Ala	Thr	Ile	Leu	Ile	Pro	Pro	His	Ala	Arg	Ile
370						375						380			
Gln	Ala	Ala	Ala	Ser	Thr	Pro	Thr	Asn	Ala	Thr	Ala	Ala	Ser	Asp	Ala
385						390						395			
Asn	Thr	Gly	Asp	Arg	Gly	Gln	Thr	Asn	Asn	Ala	Ala	Ser	Ala	Ser	Ala
			405						410						
Ser	Asn	Ser	Thr												
			420												

FIGURE 2A

GSK-3 β 557 Construct Sequence

Met Glu Tyr Met Pro Met Glu Gly Gly Gly

Met	Ser	Gly	Arg	Pro	Arg	Thr	Thr	Ser	Phe	Ala	Glu	Ser	Cys	Lys	Pro	1	5	10	15
Val	Gln	Gln	Pro	Ser	Ala	Phe	Gly	Ser	Met	Lys	Val	Ser	Arg	Asp	Lys	20	25	30	
Asp	Gly	Ser	Lys	Val	Thr	Thr	Val	Val	Ala	Thr	Pro	Gly	Gln	Gly	Pro	35	40	45	
Asp	Arg	Pro	Gln	Glu	Val	Ser	Tyr	Thr	Asp	Thr	Lys	Val	Ile	Gly	Asn	50	55	60	
Gly	Ser	Phe	Gly	Val	Val	Tyr	Gln	Ala	Lys	Leu	Cys	Asp	Ser	Gly	Glu	65	70	75	80
Leu	Val	Ala	Ile	Lys	Lys	Val	Leu	Gln	Asp	Lys	Arg	Phe	Lys	Asn	Arg	85	90	95	
Glu	Leu	Gln	Ile	Met	Arg	Lys	Leu	Asp	His	Cys	Asn	Ile	Val	Arg	Leu	100	105	110	
Arg	Tyr	Phe	Phe	Tyr	Ser	Ser	Gly	Glu	Lys	Lys	Asp	Glu	Val	Tyr	Leu	115	120	125	
Asn	Leu	Val	Leu	Asp	Tyr	Val	Pro	Glu	Thr	Val	Tyr	Arg	Val	Ala	Arg	130	135	140	
His	Tyr	Ser	Arg	Ala	Lys	Gln	Thr	Leu	Pro	Val	Ile	Tyr	Val	Lys	Leu	145	150	155	160
Tyr	Met	Tyr	Gln	Leu	Phe	Arg	Ser	Leu	Ala	Tyr	Ile	His	Ser	Phe	Gly	165	170	175	
Ile	Cys	His	Arg	Asp	Ile	Lys	Pro	Gln	Asn	Leu	Leu	Leu	Asp	Pro	Asp	180	185	190	
Thr	Ala	Val	Leu	Lys	Leu	Cys	Asp	Phe	Gly	Ser	Ala	Lys	Gln	Leu	Val	195	200	205	
Arg	Gly	Glu	Pro	Asn	Val	Ser	Tyr	Ile	Cys	Ser	Arg	Tyr	Tyr	Arg	Ala	210	215	220	
Pro	Glu	Leu	Ile	Phe	Gly	Ala	Thr	Asp	Tyr	Thr	Ser	Ser	Ile	Asp	Val	225	230	235	240
Trp	Ser	Ala	Gly	Cys	Val	Leu	Ala	Glu	Leu	Leu	Leu	Gly	Gln	Pro	Ile	245	250	255	

FIGURE 2B

Phe	Pro	Gly	Asp	Ser	Gly	Val	Asp	Gln	Leu	Val	Glu	Ile	Ile	Lys	Val	
			260					265					270			
Leu	Gly	Thr	Pro	Thr	Arg	Glu	Gln	Ile	Arg	Glu	Met	Asn	Pro	Asn	Tyr	
		275					280					285				
Thr	Glu	Phe	Lys	Phe	Pro	Gln	Ile	Lys	Ala	His	Pro	Trp	Thr	Lys	Val	
	290					295					300					
Phe	Arg	Pro	Arg	Thr	Pro	Pro	Glu	Ala	Ile	Ala	Leu	Cys	Ser	Arg	Leu	
305					310					315					320	
Leu	Glu	Tyr	Thr	Pro	Thr	Ala	Arg	Leu	Thr	Pro	Leu	Glu	Ala	Cys	Ala	
				325					330					335		
His	Ser	Phe	Phe	Asp	Glu	Leu	Arg	Asp	Pro	Asn	Val	Lys	His	Pro	Asn	
			340					345					350			
Gly	Arg	Asp	Thr	Pro	Ala	Leu	Phe	Asn	Phe	Thr	Thr	Gln	Glu	Leu	Ser	
		355					360					365				
Ser	Asn	Pro	Pro	Leu	Ala	Thr	Ile	Leu	Ile	Pro	Pro	His	Ala	Arg	Ile	
	370					375					380					

FIGURE 3A

GSK-3 β 580 Construct Sequence

Met Glu Tyr Met Pro Met Glu Gly Gly Gly

Gly	Ser	Lys	Val	Thr	Thr	Val	Val	Ala	Thr	Pro	Gly	Gln	Gly	Pro	35	40	45		
Asp	Arg	Pro	Gln	Glu	Val	Ser	Tyr	Thr	Asp	Thr	Lys	Val	Ile	Gly	Asn	50	55	60	
Gly	Ser	Phe	Gly	Val	Val	Tyr	Gln	Ala	Lys	Leu	Cys	Asp	Ser	Gly	Glu	65	70	75	80
Leu	Val	Ala	Ile	Lys	Lys	Val	Leu	Gln	Asp	Lys	Arg	Phe	Lys	Asn	Arg	85	90	95	
Glu	Leu	Gln	Ile	Met	Arg	Lys	Leu	Asp	His	Cys	Asn	Ile	Val	Arg	Leu	100	105	110	
Arg	Tyr	Phe	Phe	Tyr	Ser	Ser	Gly	Glu	Lys	Lys	Asp	Glu	Val	Tyr	Leu	115	120	125	
Asn	Leu	Val	Leu	Asp	Tyr	Val	Pro	Glu	Thr	Val	Tyr	Arg	Val	Ala	Arg	130	135	140	
His	Tyr	Ser	Arg	Ala	Lys	Gln	Thr	Leu	Pro	Val	Ile	Tyr	Val	Lys	Leu	145	150	155	160
Tyr	Met	Tyr	Gln	Leu	Phe	Arg	Ser	Leu	Ala	Tyr	Ile	His	Ser	Phe	Gly	165	170	175	
Ile	Cys	His	Arg	Asp	Ile	Lys	Pro	Gln	Asn	Leu	Leu	Leu	Asp	Pro	Asp	180	185	190	
Thr	Ala	Val	Leu	Lys	Leu	Cys	Asp	Phe	Gly	Ser	Ala	Lys	Gln	Leu	Val	195	200	205	
Arg	Gly	Glu	Pro	Asn	Val	Ser	Tyr	Ile	Cys	Ser	Arg	Tyr	Tyr	Arg	Ala	210	215	220	
Pro	Glu	Leu	Ile	Phe	Gly	Ala	Thr	Asp	Tyr	Thr	Ser	Ser	Ile	Asp	Val	225	230	235	240
Trp	Ser	Ala	Gly	Cys	Val	Leu	Ala	Glu	Leu	Leu	Leu	Gly	Gln	Pro	Ile	245	250	255	
Phe	Pro	Gly	Asp	Ser	Gly	Val	Asp	Gln	Leu	Val	Glu	Ile	Ile	Lys	Val	260	265	270	
Leu	Gly	Thr	Pro	Thr	Arg	Glu	Gln	Ile	Arg	Glu	Met	Asn	Pro	Asn	Tyr	275	280	285	

FIGURE 3B

Thr	Glu	Phe	Lys	Phe	Pro	Gln	Ile	Lys	Ala	His	Pro	Trp	Thr	Lys	Val
290						295					300				
Phe	Arg	Pro	Arg	Thr	Pro	Pro	Glu	Ala	Ile	Ala	Leu	Cys	Ser	Arg	Leu
305					310					315					320
Leu	Glu	Tyr	Thr	Pro	Thr	Ala	Arg	Leu	Thr	Pro	Leu	Glu	Ala	Cys	Ala
				325					330					335	
His	Ser	Phe	Phe	Asp	Glu	Leu	Arg	Asp	Pro	Asn	Val	Lys	His	Pro	Asn
			340					345					350		
Gly	Arg	Asp	Thr	Pro	Ala	Leu	Phe	Asn	Phe	Thr	Thr	Gln	Glu	Leu	Ser
		355					360					365			
Ser	Asn	Pro	Pro	Leu	Ala	Thr	Ile	Leu	Ile	Pro	Pro	His	Ala	Arg	Ile
370						375					380				

FIGURE 4Human GSK3 α

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MSGGGPSSGGG PGGSGRARTS SFAEPGGGGG GGGGGPGGSA SGPGGTGGGK
1 50
ASVGAMGGGV GASSSGGGPG GSGGGGSGGP GAGTSFPPPG VKLGRDSSGKV
51 100
TTVVATLGQG PERSQEVAYT DIKVINGSF GVVYQARLAE TRELVAIKKV
101 150
LQDKRFKNRE LQIMRKLDHC NIVRLRYFFY SSGEKKDELY LNLVLEYVPE
151 200
TVYRVARHFT KAKLTIPILY VKVYMYQLFR SLAYIHSQGV CHRDIKPQNL
201 250
LVDPDTAVLK LCDFGSAQOL VRGEPNVSYI CSRYYRAPEL IFGATDYTSS
251 300
IDVWSAGCVL AELLGQPIF PGDSGVDQLV EIIKVLGTPT REQUIREMNP
301 350
YTEFKFPQIK AHPWTKVFKS RTPPEAIALC SSLEYTPSS RLSPLEACAH
351 400
SFFDELRLCG TQLPNNRPLP PLFNFSAGEL SIQPSLNAIL IPPHLRSPAG
401 450
TTTLTPSSQA LTETPTSSDW QSTDATPTLT NSS
451 483
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FIGURE 5Human GSK3 α

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MSGGGPSGGG PGGSGRARTS SFAEPGGGGG GGGGGPGGSA SGPGGTGGGK
1 50
ASVGAMGGGV GASSSGGGPG GSGGGGSGGP GAGTSFPPPG VKLGRDSGKV
51 100
TTVVATLGQG PERSQEVAYT DIKVIENG SF GVYQARLAE TRELVAIKKV
101 150
LQDKRFKNRE LQIMRKLDHC NIVRLRYFFY SSGKKDELY LNLVLEYVPE
151 200
TVYRVARHFT KAKLTIPILY VKVYMYQLFR SLAYIHSQGV CHRDIKPQNL
201 250
LVDPDTAVLK LCDFGSAKQL VRGEPNVSII CSRIYRAPEL IFGATDYTSS
251 300
IDVWSAGCVL AELLGQPIF PGDSGVDQLV EIIKVLGTPT REQUIREMNP
301 350
YTEFKFPQIK AHPWTKVFKS RTPPEAIALC SSLLEYTPSS RLSPLEACAH
351 400
SFFDELRC LG TQLPNNRPLP PLFNFSAGEL SIQPSLNAIL IPPHLRS
401
```

FIGURE 6Human GSK3 α

	<u>SGKV</u>
TTVVATLGQG PERSQEVAYT DIKVINGSF GVVYQARLAE TRELVAIKKV	100
101	150
LQDKRFRKNRE LQIMRKLDHC NIVRLRYFFY SSGEKKDELY LNLVLEYVPE	
151	200
TVYRVARHFT KAKLTIPILY VKVYMYQLFR SLAYIHSQGV CHRDIKPQNL	
201	250
LVDPDTAVLK LCDFGSAKQL VRGEPNVSII CSRYRRAPEL IFGATDYTSS	
251	300
IDVWSAGCVL AELLGQPIF PGDSGVDQLV EIIKVLGTPT REQUIREMNP	
301	350
YTEFKFPQIK AHPWTKVFKS RTPPEAIALC SSLEYTPSS RLSPLEACAH	
351	400
SFFDELRCLG TQLPNNRPLP PLFNFSAGEL SIQPSLNAIL IPPHLRSPAG	
401	450
TTTLTPSSQA LTETPTSSDW QSTDATPTLT NSS	
451	483

FIGURE 7Human GSK3 α

	SGKV
TTVVATLGQG PERSQEVAYT DIKVINGSF GVVYQARLAE TRELVAIKKV	100
101	150
LQDKRFKNRE LQIMRKLDHC NIVRLRYFFY SSGEKKDELY LNLVLEYVPE	
151	200
TVYRVARHFT KAKLTIPILY VKVYMYQLFR SLAYIHSQGV CHRDIKPQNL	
201	250
LVDPDTAVLK LCDFGSAKQL VRGEPNVSII CSRYYRAPEL IFGATDYTSS	
251	300
IDVWSAGCVL AELLGQPIF PGDSGVDQLV EIIKVLGTPT REQUIREMNP	
301	350
YTEFKFPQIK AHPWTKVFKS RTPPEAIALC SSLLEYTPSS RLSPLEACAH	
351	400
SFFDELRLCG TQLPNNRPLP PLFNFSAGEL SIQPSLNAIL IPPHLRS	
401	