

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(10) International Publication Number

WO 2019/051202 A1

(43) International Publication Date
14 March 2019 (14.03.2019)

(51) International Patent Classification:
A61K 48/00 (2006.01) *C12N 9/88* (2006.01)
C07K 14/435 (2006.01)

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(21) International Application Number:
PCT/US2018/049914

(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DJ, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JO, JP, KE, KG, KH, KN, KP, KR, KW, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(22) International Filing Date:
07 September 2018 (07.09.2018)

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
62/556,088 08 September 2017 (08.09.2017) US

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(54) Title: METHOD AND COMPOSITION FOR TREATING NEUROPATHIC PAIN

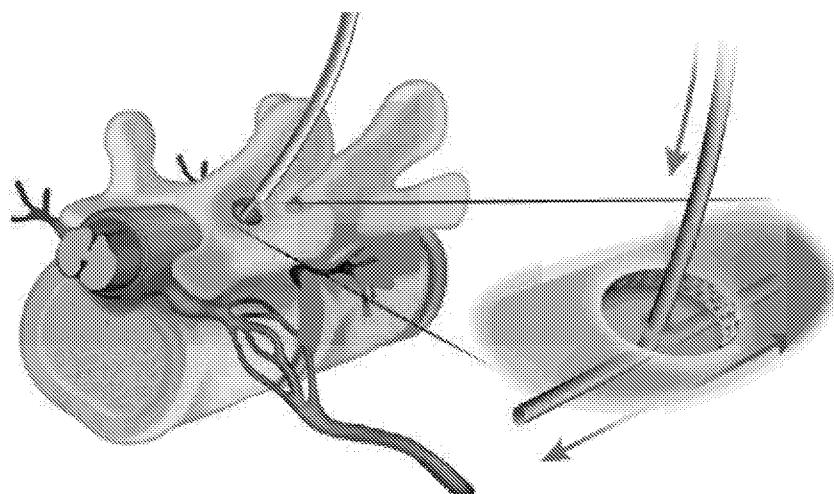


FIG. 1A

(57) Abstract: The present invention provides a therapy for treating neuropathic pain by subpial administration of small quantities of a composition for spinal segment-specific upregulation of GAD65 (glutamatedecarboxylase) gene and VGAT (vesicular GABA transporter) gene, which is effective for induction of nociceptive effects by potentiating release of vesicular GABA from infected dorsal horn neurons into the synaptic cleft.

Published:

- *with international search report (Art. 21(3))*
- *with sequence listing part of description (Rule 5.2(a))*

METHOD AND COMPOSITION FOR TREATING NEUROPATHIC PAIN

CROSS REFERENCE TO RELATED APPLICATION(S)

[0001] This application claims the benefit of priority under 35 U.S.C. § 119(e) of U.S. Serial No. 62/556,088, filed September 8, 2017, the entire content of which is incorporated herein by reference.

SEQUENCE LISTING

[0002] The instant application contains a Sequence Listing which has been submitted electronically in ASCII format and is hereby incorporated by reference in its entirety. The ASCII copy, created on September 6, 2018, is named 20378-201844_SL.txt and is 22KB in size.

BACKGROUND OF THE INVENTION

FIELD OF THE INVENTION

[0003] The invention relates generally to treating neuropathic pain and more specifically to a combined therapeutic regimen involving spinal subpial delivery of one or more genes for treating chronic neuropathic pain.

BACKGROUND INFORMATION

[0004] Neuropathic pain is pain caused by various types of nerve damage. Some examples of neuropathic pain conditions include, but are not limited to, diabetic peripheral neuropathy, herpes zoster, post herpetic neuralgia, trigeminal neuralgia, complex regional pain syndrome, reflex sympathetic dystrophy, migraine headache, phantom limb syndrome, neuropathic pain due to chronic disease (multiple sclerosis, HIV, etc), neuropathic pain due to trauma (causalgia), neuropathic pain due to impingement (i.e. sciatica, carpal tunnel, etc.), neuropathic pain due to drug exposure or toxic chemical exposure, neuropathic pain due to infection or post infection, neuropathic pain due to impaired organ function, neuropathic pain due to vascular disease, neuropathic pain due to metabolic disease, neuropathic pain due to cancer or cancer treatment, neuropathic pain due to autoimmune disease, neuropathic low back pain, neuropathic pain due to fibromyalgia, and neuropathic pain with no known cause (idiopathic). In fact, neuropathic pain is most often diagnosed based on the symptoms, so that any pain is that is characterized by burning sensations and/or shooting pain and/or

numbness and/or tingling and/or allodynia is typically considered neuropathic. Other characteristics of neuropathic pain include hyperpathia (greatly exaggerated pain sensation to stimuli), hyperesthesia (an increased sensitivity to normal stimulation), dysesthesia (unpleasant abnormal sensations as if damage is being done when this is not the case), and paresthesia (an abnormal sensation, such as "pins and needles", whether spontaneous or evoked).

[0005] It is well known that nociceptive pain and neuropathic pain are caused by different mechanisms, and therefore respond to different treatment modalities. Nociceptive pain is mediated by receptors which are located in skin, bone, connective tissue, muscle and viscera. These receptors typically respond to noxious chemical, thermal and mechanical stimuli producing pain that is typically described as sharp, aching, throbbing, or gnawing. In contrast, neuropathic pain is produced by damage to, or pathological changes in, the peripheral or central nervous systems, typically producing pain that is described as "burning", "electric", "tingling", and "shooting" in nature. Finally, nociceptive pain usually responds to opioids and non-steroidal anti-inflammatories (NSAIDS), whereas success treating neuropathic pain with these approaches has been limited.

[0006] At present, intrathecally-infused anti-nociceptive compounds (like gabapentin and opioids) are used to achieve a spinally-restricted anti-nociceptive effect. However, no gene-therapy-based technique is available to treat chronic neuropathic pain. Similarly, no gene therapy-based technique that can effectively be used to suppress nociceptive transmission in specific spinal segments, and which is ipsilateral to the site of peripheral nerve injury, is currently available. In addition, currently available spinal drug delivery systems (such as epidural or intrathecal delivery) do not permit a spinal segment-restricted therapeutic effect. Thus, a need exists for such improved treatments for neuropathic pain.

SUMMARY OF THE INVENTION

[0007] The present invention is based on the observation that a combined treatment composed of spinal segment-specific upregulation of GAD65 (glutamatedecarboxylase) and VGAT (vesicular GABA transporter) is effective for induction of nociceptive effects by potentiating release of vesicular GABA from infected dorsal horn neurons into the synaptic cleft.

[0008] Accordingly, the invention provides a method of treating neuropathic pain in a subject. The method includes subpial administration of a composition comprising: (i) a viral vector sequence, a GAD65 gene sequence (e.g., SEQ ID NO: 2 or 5), and a VGAT gene sequence (e.g., SEQ ID NO: 4 or 6) under the control of a tissue-specific promoter; and (ii) a pharmaceutically acceptable viral carrier, thereby treating neuropathic pain in the subject. In various embodiments, the subject may be a mammal. In various embodiments, the nucleic acid construct is encapsidated with an AAV serotype 9 capsid. In various embodiments, the concentration of the nucleic acid construct in the composition is between about $0.1\text{-}2.0 \times 10^{13}$ gc/ml. In various embodiments, the pharmaceutically acceptable viral carrier is selected from the group consisting of a lentiviral vector, an adenoviral vector (AV), or an adeno-associated vector (AAV). In various embodiments, the AAV is selected from the group consisting of AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV rh8, AAVrh10, AAVrh33, AAV rh34, and AAV Anc80. In various embodiments, the tissue-specific promoter is selected from the group consisting of human ubiquitin promoter and human synapsin promoter.

[0009] In another aspect, the invention provides a method of treating neuropathic pain in a subject. The method includes subpially administering a composition comprising a therapeutically effective amount of a gene therapy construct comprising (i) a viral vector sequence; (ii) a GAD65 gene sequence; and (iii) a VGAT gene sequence under control of a tissue-specific promoter, and a pharmaceutically acceptable carrier, thereby treating neuropathic pain in the subject. In various embodiments, the nucleic acid construct is encapsidated with an AAV serotype 9 capsid. In various embodiments, the concentration of the nucleic acid construct in the composition is between about $0.1\text{-}2.0 \times 10^{13}$ gc/ml. In various embodiments, the pharmaceutically acceptable viral carrier is selected from the group consisting of a lentiviral vector, an adenoviral vector (AV), or an adeno-associated vector (AAV). In various embodiments, the AAV is selected from the group consisting of AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV rh8, AAVrh10, AAVrh33, AAV rh34, and AAV Anc80. In various embodiments, the tissue-specific promoter is selected from the group consisting of human ubiquitin promoter and human synapsin promoter.

[0010] In another aspect, the invention provides a method of treating neuropathic pain in a subject. The method includes subpial administration of a first vector encoding GAD65 (glutamate decarboxylase) gene and a second vector encoding VGAT (vesicular GABA

transporter) gene, thereby treating neuropathic pain in the subject. Administration of the first and second vectors results in spinal-specific upregulation of the GAD65 gene and VGAT gene in the subject. In various embodiments, the first viral vector comprises a polynucleotide encoding GAD65 and the second viral vector comprises a polynucleotide encoding VGAT, wherein GAD65 and VGAT are expressed in the subject, thereby treating neuropathic pain in the subject. The first and second vectors may be a lentiviral vector, adenoviral vector, or an adeno-associated vector (AAV). The AAV may be AAV type 9 (AAV9) or AAV Anc80. In various embodiments, the first vector AAV9-UBI-GAD65 and the second vector is AAV9-UBI-VGAT.

[0011] In another aspect, the invention provides a method of treating neuropathic pain in a subject. The method includes subpially administering to a subject in need thereof a therapeutically effective amount of a viral vector comprising a polynucleotide encoding GAD65 and systemically administering to the subject a GABA agonist, thereby treating spasticity in the subject. The vector may be a lentiviral vector, adenoviral vector, or an adeno-associated vector (AAV), and may be administered directly into the spine of the subject. The AAV may be AAV type 9 (AAV9) or AAV Anc80. In various embodiments, the vector is AAV9-UBI-GAD65.

[0012] In another aspect, the invention provides a nucleic acid construct comprising: a viral vector sequence, a GAD65 gene sequence, and a VGAT gene sequence under the control of a tissue-specific promoter. In various embodiments, the tissue-specific promoter is selected from the group consisting of human ubiquitin promoter and human synapsin promoter. In various embodiments, the viral vector is an adeno-associated viral (AAV) vector, such as, AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV rh8, AAVrh10, AAVrh33, AAV rh34, or AAV Anc80.

BRIEF DESCRIPTION OF THE DRAWINGS

[0013] Figures 1A-1C are pictorial diagrams showing unilateral spinal subpial vector delivery. Figure 1A shows a pictorial diagram of a spinal cord with a subpially placed catheter in an animal model. Figure 1B shows unilateral subpial advancement (3 mm) of an injection needle into the subpial space for delivery of the vector composition. Figure 1C shows unilateral dorsal horn mRNA-GFP signal (fluorescence in situ hybridization; FISH) at

2 weeks after unilateral subpial delivery of AAV9-UBI-GFP (0.5 μ l; 1.2x10¹³ gc/ml) in an adult mouse.

[0014] Figures 2A-2F are pictorial and graphical diagrams showing anti-nociceptive results after subpial delivery of AAV9-UBI-GAD65/VGAT or Anc80-UBI-GAD65/VGAT in mice with developed neuropathic pain. Figures 2A and 2B show significant suppression of tactile and brush-evoked nociceptive response in hind paw in animals receiving unilateral subpial injection of the AAV9-UBI-GAD65/VGAT vector. Figures 2C and 2D show significant improvement of open field motor performance (running distance) and ipsilateral hind paw placement pattern (Cat Walk assay) in neuropathic animals receiving unilateral subpial injection of the AAV9-UBI-GAD65/VGAT vector. Figures 2E and 2F show that a comparable anti-nociceptive effect was seen in neuropathic animals treated with the Anc80-UBI-GAD65/VGAT vector.

[0015] Figures 3A-3E are pictorial and graphical diagrams showing induction of mixed inhibitory-excitatory neurotransmitter phenotype in ipsilateral dorsal horn excitatory interneurons after unilateral (ipsilateral) (L2-L4) subpial AAV9-UBI-GAD65/VGAT delivery in adult mice. Figures 3A-3D show images of staining of ipsilateral dorsal horn neurons with VGLUT2 (Figure 3A), GAD65 (Figure 3B), VGAT (Figure 3C) and colocalization of VGLUT2, GAD65 and VGAT-stained puncta (Figure 3D). A clear co-expression of VGLUT2 with VGAT and VGLUT2 with GAD65 (white arrows) was detected. Figure 3E shows quantitative densitometry results of VGLUT, GAD65 and VGAT expression in the dorsal horn (lamina I-III) in treated (Injured+GAD65+VGAT) and control (Injured+PBS) animals. Mice were injected ipsilaterally (L2-L4) with AAV9-UBI-GAD65/VGAT vector or PBS only. No increased expression in contralateral dorsal horn was seen/measured.

[0016] Figures 4A-4D are pictorial diagrams showing the results of pre-embedding immune-gold staining with VGLUT2 and GAD65 and VGAT antibodies coupled with electron microscopy that show a clear increase in GAD65 and VGAT immunogold-tagged particles in VGLUT2+ terminals in AAV9-UBI-GAD65/VGAT-injected animals (PNI+GAD65+VGAT) if compared to PBS-injected animals (PNI-PBS).

[0017] Figures 4E-4J are pictorial diagrams showing the results from fluorescence in situ hybridization that show a clear appearance of double and triple-tagged neurons (white arrows) with VGLUT2, GAD65 and VGAT mRNA in ipsilateral dorsal horn neurons in AAV9-UBI-GAD65/VGAT-injected animals.

[0018] Figures 5A-5D are pictorial diagrams showing unilateral Anc80-UBI-Rpl22 (3xHA) delivery in an adult pig model. As shown, ipsilateral dorsal horn neuron-specific Rpl22 protein expression was observed after unilateral subpial (L2-L3) Anc80-UBI-Rpl22-3xHA vector delivery (100 μ l; 1.2x10¹³ gc/ml) in adult pig model (DH-dorsal horn; VH-ventral horn).

[0019] Figures 6A-6D are pictorial diagrams showing that unilateral Anc80-UBI-GAD65/VGAT delivery in an adult pig induces mixed excitatory-inhibitory neuronal phenotype in dorsal horn neurons. Staining of ipsilateral dorsal horn neurons with VGLUT2 (Figure 6A), VGAT (Figure 6B), and GAD65 (Figure 6C) antibodies in the animal model (adult pig; 35 kg) injected ipsilaterally with Anc80-UBI-GAD65/VGAT vector (100 μ l; 1.2x10¹³ gc/ml). As shown, a clear co-expression of VGLUT2 with VGAT and GAD65 (white puncta) can be detected (Figure 6D).

[0020] Figure 7 is a pictorial diagram showing exemplary steps of an experimental design to study the therapeutic anti-spasticity effect of spinal subpial delivery of GAD65 and VGAT genes in a rat model with chronic spinal injury-induced muscle spasticity. Steps A and B show that muscle spasticity in hind limbs is induced by Th9 spinal segment transection and spasticity is identified/measured by quantitative change in gastrocnemius muscle EMG response to paw tactile stimulus. Steps C and D show that 2-3 months after induction of muscle spasticity, the animals received lumbar subpial injection of AAV9 (or Anc80)-UBI-GAD65/VGAT vector and the presence of spasticity response was measured for an additional 2 months. Step E shows that after sacrifice the presence of GAD65 and VGAT upregulation was measured by immunofluorescence staining.

[0021] Figures 8A-8D are pictorial and graphical diagrams showing anti-spasticity effect after subpial delivery of AAV9-UBI-GAD65/VGAT in chronic rat model of spinal transection-induced muscle spasticity. Figures 8A and 8B show the results of measurement of muscle spasticity in animals receiving a control vector (AAV9-GFP), which showed a

progressive increase in muscle spasticity for 8 weeks after virus delivery (compared to baseline measured at 2-3 months after spinal transection). In contrast a near complete block of spasticity response was measured in animals receiving AAV9-UBI-GAD65/VGAT vector. Figures 8C and 8D show the results of measurement of rate-dependent depression (RDD) of H-reflex, which showed a significant recovery of RDD in animals treated with AAV9-UBI-GAD65/VGAT vector.

[0022] Figures 9A-9D are pictorial diagrams showing expression of VGLUT2, GAD65 and VGAT in dorsal horn neurons in animals (rats) injected with AAV9-UBI-GFP (control vector) or AAV9-UBI-GAD65/VGAT vector. Figures 9A and 9B show no co-expression of GAD65 and VGAT in VGLUT2 terminals in animals receiving control AAV9-UBI-GFP vector. Figures 9C and 9D show induction of mixed inhibitory-excitatory neurotransmitter phenotype in lumbar excitatory interneurons after bilateral (L2-L4) subpial AAV9-UBI-GAD65/VGAT delivery in an adult rat model of chronic muscle spasticity. The appearance of mixed neurotransmitter phenotype in spinal interneurons as evidenced by co-expression of GAD65 and VGAT in VGLUT2 terminals can be seen.

DETAILED DESCRIPTION OF THE INVENTION

[0023] The present invention is based on the observation that subpial administration of small quantities of a combined treatment composed of spinal segment-specific upregulation of GAD65 (glutamatedecarboxylase) gene and VGAT (vesicular GABA transporter) gene is effective for induction of nociceptive effects by potentiating release of vesicular GABA from infected dorsal horn neurons into the synaptic cleft.

[0024] Before the present compositions and methods are described, it is to be understood that this invention is not limited to particular compositions, methods, and experimental conditions described, as such compositions, methods, and conditions may vary. It is also to be understood that the terminology used herein is for purposes of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only in the appended claims.

[0025] As used in this specification and the appended claims, the singular forms “a”, “an”, and “the” include plural references unless the context clearly dictates otherwise. Thus, for example, references to “the method” includes one or more methods, and/or steps of the type

described herein which will become apparent to those persons skilled in the art upon reading this disclosure and so forth.

[0026] The term “comprising,” which is used interchangeably with “including,” “containing,” or “characterized by,” is inclusive or open-ended language and does not exclude additional, unrecited elements or method steps. The phrase “consisting of” excludes any element, step, or ingredient not specified in the claim. The phrase “consisting essentially of” limits the scope of a claim to the specified materials or steps and those that do not materially affect the basic and novel characteristics of the claimed invention. The present disclosure contemplates embodiments of the invention compositions and methods corresponding to the scope of each of these phrases. Thus, a composition or method comprising recited elements or steps contemplates particular embodiments in which the composition or method consists essentially of or consists of those elements or steps.

[0027] Ranges provided herein are understood to be shorthand for all of the values within the range. For example, a range of 1 to 50 is understood to include any number, combination of numbers, or sub-range from the group consisting of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, 45, 46, 47, 48, 49, and 50.

[0028] The terms “about” and “approximately” shall generally mean an acceptable degree of error for the quantity measured given the nature or precision of the measurements. Typically, exemplary degrees of error are within 20 percent (%), preferably within 10%, and more preferably within 5% of a given value or range of values. Alternatively, and particularly in biological systems, the terms “about” and “approximately” may mean values that are within an order of magnitude, preferably within 10- or 5-fold, and more preferably within 2-fold of a given value. Numerical quantities given herein are approximate unless stated otherwise, meaning that the term “about” or “approximately” can be inferred when not expressly stated.

[0029] Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those

described herein can be used in the practice or testing of the invention, the preferred methods and materials are now described.

[0030] The term “subject” as used herein refers to any individual or patient to which the subject methods are performed. Generally the subject is human, although as will be appreciated by those in the art, the subject may be an animal. Thus other animals, including mammals such as rodents (including mice, rats, hamsters and guinea pigs), cats, dogs, rabbits, farm animals including cows, horses, goats, sheep, pigs, etc., and primates (including monkeys, chimpanzees, orangutans and gorillas) are included within the definition of subject.

[0031] A “therapeutic effect,” as used herein, encompasses a therapeutic benefit and/or a prophylactic benefit as described herein.

[0032] As used herein, the terms “reduce” and “inhibit” are used together because it is recognized that, in some cases, a decrease can be reduced below the level of detection of a particular assay. As such, it may not always be clear whether the expression level or activity is “reduced” below a level of detection of an assay, or is completely “inhibited.” Nevertheless, it will be clearly determinable, following a treatment according to the present methods.

[0033] As used herein, “treatment” or “treating” means to administer a composition to a subject or a system with an undesired condition. The condition can include a disease or disorder. “Prevention” or “preventing” means to administer a composition to a subject or a system at risk for the condition. The condition can include a predisposition to a disease or disorder. The effect of the administration of the composition to the subject (either treating and/or preventing) can be, but is not limited to, the cessation of one or more symptoms of the condition, a reduction or prevention of one or more symptoms of the condition, a reduction in the severity of the condition, the complete ablation of the condition, a stabilization or delay of the development or progression of a particular event or characteristic, or minimization of the chances that a particular event or characteristic will occur.

[0034] The terms “polypeptide,” “peptide,” and “protein” are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical mimetic of a corresponding

naturally occurring amino acid, as well as to naturally occurring amino acid polymers and non-naturally occurring amino acid polymer.

[0035] The term “amino acid” refers to naturally occurring and synthetic amino acids, as well as amino acid analogs and amino acid mimetics that function in a manner similar to the naturally occurring amino acids. Naturally occurring amino acids are those encoded by the genetic code, as well as those amino acids that are later modified, *e.g.*, hydroxyproline, α -carboxyglutamate, and O-phosphoserine. Amino acid analogs refers to compounds that have the same basic chemical structure as a naturally occurring amino acid, *i.e.*, an α carbon that is bound to a hydrogen, a carboxyl group, an amino group, and an R group, *e.g.*, homoserine, norleucine, methionine sulfoxide, methionine methyl sulfonium. Such analogs have modified R groups (*e.g.*, norleucine) or modified peptide backbones, but retain the same basic chemical structure as a naturally occurring amino acid. Amino acid mimetics refers to chemical compounds that have a structure that is different from the general chemical structure of an amino acid, but that functions in a manner similar to a naturally occurring amino acid.

[0036] Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUB Biochemical Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes.

[0037] As used herein, a “regulatory gene” or “regulatory sequence” is a nucleic acid sequence that encodes products (*e.g.*, transcription factors) that control the expression of other genes.

[0038] As used herein, a “protein coding sequence” or a sequence that encodes a particular protein or polypeptide, is a nucleic acid sequence that is transcribed into mRNA (in the case of DNA) and is translated (in the case of mRNA) into a polypeptide *in vitro* or *in vivo* when placed under the control of appropriate regulatory sequences. The boundaries of the coding sequence are determined by a start codon at the 5' terminus (N-terminus) and a translation stop nonsense codon at the 3' terminus (C-terminus). A coding sequence can include, but is not limited to, cDNA from eukaryotic mRNA, genomic DNA sequences from eukaryotic DNA, and synthetic nucleic acids. A transcription termination sequence will usually be located 3' to the coding sequence.

[0039] As used herein, a “promoter” is defined as a regulatory DNA sequence generally located upstream of a gene that mediates the initiation of transcription by directing RNA polymerase to bind to DNA and initiating RNA synthesis. A promoter can be a constitutively active promoter (*i.e.*, a promoter that is constitutively in an active/“ON” state), it may be an inducible promoter (*i.e.*, a promoter whose state, active/“ON” or inactive/“OFF”, is controlled by an external stimulus, *e.g.*, the presence of a particular compound or protein), it may be a spatially restricted promoter (*i.e.*, transcriptional control element, enhancer, etc.) (*e.g.*, tissue specific promoter, cell type specific promoter, etc.), and it may be a temporally restricted promoter (*i.e.*, the promoter is in the “ON” state or “OFF” state during specific stages of embryonic development or during specific stages of a biological process.

[0040] As used herein, the term “gene” means the deoxyribonucleotide sequences comprising the coding region of a structural gene. A “gene” may also include non-translated sequences located adjacent to the coding region on both the 5' and 3' ends such that the gene corresponds to the length of the full-length mRNA. The sequences which are located 5' of the coding region and which are present on the mRNA are referred to as 5' non-translated sequences. The sequences which are located 3' or downstream of the coding region and which are present on the mRNA are referred to as 3' non-translated sequences. The term “gene” encompasses both cDNA and genomic forms of a gene. A genomic form or clone of a gene contains the coding region interrupted with non-coding sequences termed “introns” or “intervening regions” or “intervening sequences.” Introns are segments of a gene which are transcribed into heterogenous nuclear RNA (hnRNA); introns may contain regulatory elements such as enhancers. Introns are removed or “spliced out” from the nuclear or primary transcript; introns therefore are absent in the messenger RNA (mRNA) transcript. The mRNA functions during translation to specify the sequence or order of amino acids in a nascent polypeptide.

[0041] As used herein, the terms “functionally linked” and “operably linked” are used interchangeably and refer to a functional relationship between two or more DNA segments, in particular gene sequences to be expressed and those sequences controlling their expression. For example, a promoter/enhancer sequence, including any combination of cis-acting transcriptional control elements is operably linked to a coding sequence if it stimulates or modulates the transcription of the coding sequence in an appropriate host cell or other

expression system. Promoter regulatory sequences that are operably linked to the transcribed gene sequence are physically contiguous to the transcribed sequence.

[0042] “Conservatively modified variants” applies to both amino acid and nucleic acid sequences. With respect to particular nucleic acid sequences, conservatively modified variants refers to those nucleic acids which encode identical or essentially identical amino acid sequences, or where the nucleic acid does not encode an amino acid sequence, to essentially identical sequences. Because of the degeneracy of the genetic code, a large number of functionally identical nucleic acids encode any given protein. For instance, the codons GCA, GCC, GCG and GCU all encode the amino acid alanine. Thus, at every position where an alanine is specified by a codon, the codon can be altered to any of the corresponding codons described without altering the encoded polypeptide. Such nucleic acid variations are “silent variations,” which are one species of conservatively modified variations. Every nucleic acid sequence herein which encodes a polypeptide also describes every possible silent variation of the nucleic acid. One of skill will recognize that each codon in a nucleic acid (except AUG, which is ordinarily the only codon for methionine, and TGG, which is ordinarily the only codon for tryptophan) can be modified to yield a functionally identical molecule. Accordingly, each silent variation of a nucleic acid which encodes a polypeptide is implicit in each described sequence.

[0043] As to amino acid sequences, one of skill in the art will recognize that individual substitutions, deletions or additions to a nucleic acid, peptide, polypeptide, or protein sequence which alters, adds or deletes a single amino acid or a small percentage of amino acids in the encoded sequence is a “conservatively modified variant” where the alteration results in the substitution of an amino acid with a chemically similar amino acid. Conservative substitution tables providing functionally similar amino acids are well known in the art. Such conservatively modified variants are in addition to and do not exclude polymorphic variants, interspecies homologs, and alleles of the invention.

[0044] The following eight groups each contain amino acids that are conservative substitutions for one another:

[0045] 1) Alanine (A), Glycine (G);

[0046] 2) Aspartic acid (D), Glutamic acid (E);

[0047] 3) Asparagine (N), Glutamine (Q);

[0048] 4) Arginine (R), Lysine (K);

[0049] 5) Isoleucine (I), Leucine (L), Methionine (M), Valine (V);

[0050] 6) Phenylalanine (F), Tyrosine (Y), Tryptophan (W);

[0051] 7) Serine (S), Threonine (T); and

[0052] 8) Cysteine (C), Methionine (M) (see, *e.g.*, Creighton, *Proteins* (1984)).

[0053] A conservative substitution may include substitution such as basic for basic, acidic for acidic, polar for polar, etc. The sets of amino acids thus derived are likely to be conserved for structural reasons. These sets can be described in the form of a Venn diagram (Livingstone C. D. and Barton G. J. (1993) "Protein sequence alignments: a strategy for the hierarchical analysis of residue conservation" *Comput. Appl Biosci.* 9: 745-756; Taylor W. R. (1986) "The classification of amino acid conservation" *J. Theor. Biol.* 119; 205-218). Conservative substitutions may be made, for example, according to the table below which describes a generally accepted Venn diagram grouping of amino acids.

TABLE 1: Grouping of amino acids

Characteristic	Set	Characteristic	Sub-set
Hydrophobic	F W Y H K M I L V A G C	Aromatic Aliphatic	F W Y H I L V
Polar	W Y H K R E D C S T N Q	Charged Positive	H K R E D H K R
		Charged Negative	E D
Small	V C A G S P T N D	Tiny	A G S

[0054] "Percentage of sequence identity" is determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (*i.e.*, gaps) as compared to the reference sequence (*e.g.*, a polypeptide of the invention), which does not comprise additions or deletions, for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions,

dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.

[0055] The terms “identical” or percent “identity,” in the context of two or more nucleic acids or polypeptide sequences, refer to two or more sequences or subsequences that are the same sequences. Two sequences are “substantially identical” if two sequences have a specified percentage of amino acid residues or nucleotides that are the same (*i.e.*, 60% identity, optionally 65%, 70%, 75%, 80%, 85%, 90%, or 95% identity over a specified region, or, when not specified, over the entire sequence), when compared and aligned for maximum correspondence over a comparison window, or designated region as measured using one of the following sequence comparison algorithms or by manual alignment and visual inspection. The invention provides polypeptides that are substantially identical to the polypeptides, respectively, exemplified herein, as well as uses thereof including, but not limited to, use for treating or preventing neurological diseases or disorders, *e.g.*, neurodegenerative diseases or disorders, and/or treating SCI. Optionally, the identity exists over a region that is at least about 50 nucleotides in length, or more preferably over a region that is 100 to 500 or 1000 or more nucleotides in length, or the entire length of the reference sequence.

[0056] For sequence comparison, typically one sequence acts as a reference sequence, to which test sequences are compared. When using a sequence comparison algorithm, test and reference sequences are entered into a computer, subsequence coordinates are designated, if necessary, and sequence algorithm program parameters are designated. Default program parameters can be used, or alternative parameters can be designated. The sequence comparison algorithm then calculates the percent sequence identities for the test sequences relative to the reference sequence, based on the program parameters.

[0057] A “comparison window,” as used herein, includes reference to a segment of any one of the number of contiguous positions selected from the group consisting of from 20 to 600, usually about 50 to about 200, more usually about 100 to about 150 in which a sequence may be compared to a reference sequence of the same number of contiguous positions after the two sequences are optimally aligned. Methods of alignment of sequences for comparison are well known in the art. Optimal alignment of sequences for comparison can be conducted, *e.g.*, by the local homology algorithm of Smith and Waterman (1970) *Adv. Appl. Math.*

2:482c, by the homology alignment algorithm of Needleman and Wunsch (1970) *J. Mol. Biol.* 48:443, by the search for similarity method of Pearson and Lipman (1988) *Proc. Nat'l. Acad. Sci. USA* 85:2444, by computerized implementations of these algorithms (GAP, BESTFIT, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group, 575 Science Dr., Madison, Wis.), or by manual alignment and visual inspection (see, *e.g.*, Ausubel et al., *Current Protocols in Molecular Biology* (1995 supplement)).

[0058] Two examples of algorithms that are suitable for determining percent sequence identity and sequence similarity are the BLAST and BLAST 2.0 algorithms, which are described in Altschul et al. (1977) *Nuc. Acids Res.* 25:3389-3402, and Altschul et al. (1990) *J. Mol. Biol.* 215:403-410, respectively. Software for performing BLAST analyses is publicly available through the National Center for Biotechnology Information. This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold (Altschul *et al.*, *supra*). These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always >0) and N (penalty score for mismatching residues; always <0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W , T , and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) or 10, $M=5$, $N=-4$ and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength of 3, and expectation (E) of 10, and the BLOSUM62 scoring matrix (see Henikoff and Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915) alignments (B) of 50, expectation (E) of 10, $M=5$, $N=-4$, and a comparison of both strands.

[0059] The BLAST algorithm also performs a statistical analysis of the similarity between two sequences (see, *e.g.*, Karlin and Altschul (1993) Proc. Natl. Acad. Sci. USA 90:5873-5877). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance. For example, a nucleic acid is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference nucleic acid is less than about 0.2, more preferably less than about 0.01, and most preferably less than about 0.001.

[0060] “Nucleic acid” refers to deoxyribonucleotides or ribonucleotides and polymers thereof in either single- or double-stranded form, and complements thereof. The term encompasses nucleic acids containing known nucleotide analogs or modified backbone residues or linkages, which are synthetic, naturally occurring, and non-naturally occurring, which have similar binding properties as the reference nucleic acid, and which are metabolized in a manner similar to the reference nucleotides. Examples of such analogs include, without limitation, phosphorothioates, phosphoramidates, methyl phosphonates, chiral-methyl phosphonates, 2-O-methyl ribonucleotides, peptide-nucleic acids (PNAs). In various embodiments, nucleic acids are isolated when purified away from other cellular components or other contaminants (*e.g.*, other nucleic acids or proteins present in the cell) by standard techniques including, including alkaline/SDS treatment, CsCl banding, column chromatography, agarose gel electrophoresis and others well-known in the art. See *e.g.*, F. Ausubel, et al., ed. (1987) Current Protocols in Molecular Biology, Greene Publishing and Wiley Interscience, New York. In various embodiments, a nucleic acid is, for example, DNA or RNA and may or may not contain intronic sequences. In a preferred embodiment, the nucleic acid is a cDNA molecule.

[0061] As used herein, a “vector” is a nucleic acid molecule that when introduced into a host cell produces a transformed host cell. A vector may include nucleic acid sequences that permit it to replicate in the host cell, such as an origin of replication. A vector may also include one or more selectable marker genes and other genetic elements known in the art. A vector may include a “gene transfer vector,” “gene therapy vector,” or “gene therapy construct,” or similar terms, which refer to specific vector constructs that are suitable to conduct gene transfer to administer a desired gene.

[0062] The terms “vector,” “cloning vector,” and “expression vector” therefore refer to the vehicle by which a DNA or RNA sequence (*e.g.*, a foreign gene) can be introduced into a host cell so as to transform the host and promote expression (*e.g.*, transcription and translation) of the introduced sequence. Vectors include any genetic element, such as a plasmid, phage, transposon, cosmid, chromosome, virus, virion, etc., which is capable of replication when associated with the proper control elements and which can transfer gene sequences between cells. Thus, the term includes cloning and expression vehicles, as well as viral vectors.

[0063] As used herein “pharmaceutically acceptable carrier” encompasses any of the standard pharmaceutical carriers, such as a phosphate buffered saline solution, water and emulsions such as an oil/water or water/oil emulsion, and various types of wetting agents. Some examples of materials which can serve as pharmaceutically acceptable carriers include: sugars, such as lactose, glucose and sucrose; starches, such as corn starch and potato starch; cellulose, and its derivatives, such as sodium carboxymethyl cellulose, ethyl cellulose and cellulose acetate; powdered tragacanth; malt; gelatin; talc; excipients, such as cocoa butter and suppository waxes; oils, such as peanut oil, cottonseed oil, safflower oil, sesame oil, olive oil, corn oil and soybean oil; glycols, such as propylene glycol; polyols, such as glycerin, sorbitol, mannitol and polyethylene glycol; esters, such as ethyl oleate and ethyl laurate; agar; buffering agents, such as magnesium hydroxide and aluminum hydroxide; alginic acid; pyrogen-free water; isotonic saline; Ringer's solution; ethyl alcohol; phosphate buffer solutions; and other non-toxic compatible substances employed in pharmaceutical formulations. Suitable pharmaceutical carriers are described in Remington's Pharmaceutical Sciences, Mack Publishing Company, a standard reference text in this field.

[0064] As used herein, the term “gene therapy” refers to a method of changing the expression of an endogenous gene by exogenous administration of a gene, *i.e.*, a GAD65 and/or VGAT gene. As used herein, gene therapy also refers to the replacement of a defective GAD65 and/or VGAT gene, or replacement of a missing GAD65 and/or VGAT gene, by introducing a functional gene or portion of a gene corresponding to the defective or missing GAD65 and/or VGAT gene into the spinal subpial space of an individual in need. For purposes of the present disclosure, gene therapy can be accomplished by “*in vivo*” methods using a broad range of viral vectors (*e.g.*, AAV), liposomes, nanoparticles,

protein:DNA complexes, modified nucleic acids or naked DNA in order to achieve a therapeutic outcome.

[0065] The term “transgene” refers to a polynucleotide that is introduced into a cell of a subject and is capable of being expressed under appropriate conditions and confers a desired property to a cell into which it was introduced, or otherwise leads to a desired therapeutic outcome.

[0066] The terms “genome particles (gp),” “genome equivalents,” or “genome copies (gc)” as used in reference to a viral titer, refer to the number of virions containing the recombinant AAV DNA genome, regardless of infectivity or functionality. The number of genome particles in a particular vector preparation can be measured by procedures such as described elsewhere herein, or for example, in Clark et al. (1999) *Hum. Gene Ther.*, 10:1031-1039; Veldwijk et al. (2002) *Mol. Ther.*, 6:272-278.

[0067] As used herein, the term “neuron” include a neuron and a portion or portions thereof (*e.g.*, the neuron cell body, an axon, or a dendrite). Thus, the term “neuron” as used herein denotes nervous system cells that include a central cell body or soma, and two types of extensions or projections: dendrites, by which, in general, the majority of neuronal signals are conveyed to the cell body, and axons, by which, in general, the majority of neuronal signals are conveyed from the cell body to effector cells, such as target neurons or muscle. Neurons can convey information from tissues and organs into the central nervous system (afferent or sensory neurons) and transmit signals from the central nervous systems to effector cells (efferent or motor neurons). Other neurons, designated interneurons, connect neurons within the central nervous system (the brain and spinal column). Certain specific examples of neuron types that may be subject to treatment or methods according to the invention include cerebellar granule neurons, dorsal root ganglion neurons, and cortical neurons.

[0068] The term “neuronal degeneration” is used broadly and refers to any pathological changes in neuronal cells, including, without limitation, death or loss of neuronal cells, any changes that precede cell death, and any reduction or loss of an activity or a function of the neuronal cells. The pathological changes may be spontaneous or may be induced by any event and include, for example, pathological changes associated with apoptosis. The neurons may be any neurons, including without limitation sensory, sympathetic, parasympathetic, or

enteric, *e.g.*, dorsal root ganglia neurons, motor neurons, and central neurons, *e.g.*, neurons from the spinal cord. Neuronal degeneration or cell loss is a characteristic of a variety of neurological diseases or disorders, *e.g.*, neurodegenerative diseases or disorders. In some embodiments, the neuron is a sensory neuron. In some embodiments, the neuron is a motor neuron. In some embodiments, the neuron is a damaged spinal cord neuron.

[0069] As used herein, “neurodegenerative disorder” or a “neurological disorder” refers to a disorder which causes morphological and/or functional abnormality of a neural cell or a population of neural cells. The neurodegenerative disorder can result in an impairment or absence of a normal neurological function or presence of an abnormal neurological function in a subject. For example, neurodegenerative disorders can be the result of disease, injury, and/or aging. Non-limiting examples of morphological and functional abnormalities include physical deterioration and/or death of neural cells, abnormal growth patterns of neural cells, abnormalities in the physical connection between neural cells, under- or over production of a substance or substances, *e.g.*, a neurotransmitter, by neural cells, failure of neural cells to produce a substance or substances which it normally produces, production of substances, *e.g.*, neurotransmitters, and/or transmission of electrical impulses in abnormal patterns or at abnormal times. Neurodegeneration can occur in any area of the brain of a subject and is seen with many disorders including, for example, head trauma, stroke, ALS, multiple sclerosis, Huntington's disease, Parkinson's disease, and Alzheimer's disease.

[0070] As used herein, the term “nociception” refers to the sensory nervous system’s response to certain harmful or potentially harmful stimuli. In nociception, intense chemical (*e.g.*, chili powder in the eyes), mechanical (*e.g.*, cutting, crushing), or thermal (heat and cold) stimulation of sensory nerve cells called nociceptors produces a signal that travels along a chain of nerve fibers via the spinal cord to the brain. Nociception triggers a variety of physiological and behavioral responses and usually results in a subjective experience of pain in sentient beings.

[0071] Gamma-aminobutyric acid (GABA) and glutamate are the primary inhibitory and excitatory neurotransmitters in mammals. The balance between GABA and glutamate controls diverse processes such as neurogenesis, movement, circadian clocks, tissue development and blood glucose regulation. GABA is synthesized from glutamate by the 65

kDa and 67 kDa isoforms of the pyridoxal phosphate (PLP) dependent enzyme Glutamic Acid Decarboxylase (GAD65 and GAD67, respectively).

[0072] Nucleic acid sequences for rat GAD2/GAD65 are known in the art. See, for example, GenBank Accession No.: NM_012563, *Rattus norvegicus glutamate decarboxylase 2 (Gad2)*, mRNA, which provides the nucleic acid sequence (SEQ ID NO: 5):

[0073] Human GAD65 and GAD67 have been isolated and cloned by Bu et al. (1992) Proc Natl Acad Sci 89:2115-2119. Human GAD65 cDNA encodes a Mr 65,000 polypeptide, with 585 amino acid residues (Genbank Accession No. NM000818; M81882), Human GAD67 encodes a Mr 67,000 polypeptide, with 594 amino acid residues (Genbank Accession No. NM013445; M81883); each of which is incorporated herein by reference). See also, US Pub. No. 2016/0081956, incorporated herein by reference).

[0074] Additional nucleic acid and amino acid sequences for human GAD65 are known in the art. See, for example, GenBank Accession No.: Q05329, human Glutamate decarboxylase 2 (GAD2/GAD65), which provides the amino acid sequence (SEQ ID NO: 1):

MASPGSGFWSGSEDGSGDSENP GTARAWCQVAQKFTGGIGNKL CALLYGDAEKP AESGGSQPPRAAARKAACACDQKPCSKVDVNYAFLHATDLPACDGERPTLAFL QDVMNILLQYVVKS FDRSTKVIDHYPNELLQEYNWELADQPQNLEEILMHCQTTL KYAIKTGHPRYFNQLSTGLDMVGLAADWLTSTANTNMFTYEIAPVFLLEYVTLKK MREIIGWPGGSGDGIFSPGGAISNMYAMMIARFKMFPEVKEKGMAALPRLIAFTSEH SHFSLKKGAAALGIGTDSVILIKCDERGKMI PSDLERRILEAKQKGFPFLVSATAGTT VYGA FDPPLAVADICKKYKJWMHVDAAWGGGLLMSRKHKWKL SGVERANSVTW NPHKMMGVPLQCSALLVREEGLMQNCNQMHASYLFQQDKHYDLSYDTGDKALQC GRHVDVFKLWLMWRAKGTTGFEAHVDKCLEAEYLYNIKNREGYEMVFDGKPQH TNVCFWYIPPSLRTLEDNEERMSRLSKVAPVIKARMMEY GTTMVSYQPLGDKVNFF RMVISNPAATHQDIDFLIEEIERLGQDL.

See also, for example, GenBank Accession No.: X69936, Homo sapiens mRNA for glutamate decarboxylase (GAD2/GAD65), which provides the nucleic acid sequence (SEQ ID NO: 2):

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1 atgtcccccta tacatcacca tcaccatcac ctgggtccgc gtggatccga agcttcgaat
61 tctggctttt ggttttcgg gtcggaaagat ggctctgggg attccgagaa tccccggcaca
121 gcgcgagcct ggtcccaagt ggctcagaag ttacacggcg gcatcgaaaa caaactgtgc
181 gcccgtctc acggagacgc cgagaagccg gcggagagcg gcgggagcca aaaaaa cccgg
241 gcccggcccc ggaaggccgc ctgcgcctgc gaccagaagc cctgcagctg ctccaaagtg
301 gatgtcaact acgcgtttct ccatgcaaca gacctgctgc cggcggtgtga tggagaaagg
361 cccactttgg cgtttctgca agatgttatg aacatttac ttca gatgtatgt ggtgaaaagt
421 ttca gatagat caaccaaaatg gattgatttc cattatccta atgagcttct ccaagaataat
481 aattggaaat tggcagacca accacaaaat ttggaggaaa ttttgcatgca ttgcca aaca
541 actctaaaat atgcaattaa aacagggcat cctagataact tcaatcaact ttctactgg
601 ttggatatgg ttggattagc agcagactgg ctgacatcaa cagcaaatac taacatgttc
661 acctatgaaa ttgc tccagt atttgc ttttgc ttggatatgc tcacactaaa gaaaatgaga
721 gaaatcattt gctggccagg gggctctggc gatggatat tttctcccg tggcgccata
781 tcta acatgt atgcatgat gatgc acacgc ttta agatgt tccca gaaatgtca aacatgtca
841 ggaatggctg ctcttccca gctcattgcc ttca cgtctg aacatgtca ttttctctc
901 aagaaggag ctgc agccctt agggatttga acagacacgc tgattctgat taaatgtat
961 gagagaggaa aatgatttcc atctgatctt gaaagaaggaa ttcttgc aacatgtca aacatgtca
1021 gggtttgc ttttcctcgt gagtgc caca gctggaaacca ccgtgtacgg agcatttgc
1081 cccctcttag ctgtgc tgc catttgc aaggataaaga tctggatgca tggatgc
1141 gcttgggtt ggggattact gatgtccgc aaacacaatg ggaaactgag tggcg tggag
1201 agggccaaact ctgtgacgtg gaatccacac aagatgtat gatgc tccctt gca gatgtctc
1261 gctctccgg ttagagaaga gggatttgc gagaatttgc accaaatgca tgcctc ttc
1321 ctcttcagc aagataaaca ttatgac tgc tcctatgaca ctggagacaa ggc ttc
1381 tgcggacgccc acgttgc tttttaacta tggctgtatg ggaggc aaaa gggactacc
1441 gggtttgc aagatgtat gatggat ttttgc gatggat gatggat gatggat gatggat
1501 aaaaaccgag aaggatataa gatggat ttttgc gatggat gatggat gatggat gatggat
1561 ttcttgc ttcctccaa gcttgc ttttgc ttttgc ttttgc ttttgc ttttgc ttttgc
1621 ctctcgaagg tggctccatg gatggat gatggat gatggat gatggat gatggat gatggat
1681 agctaccaac ctttggaga caaggat ttttgc ttttgc ttttgc ttttgc ttttgc ttttgc
1741 gcaactcacc aagacatttgc ttttgc ttttgc ttttgc ttttgc ttttgc ttttgc ttttgc
1801 taa.

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[0075] GABA acts at inhibitory synapses in the brain by binding to specific transmembrane receptors in the plasma membrane of both pre- and postsynaptic neuronal processes. This binding causes the opening of ion channels to allow the flow of either negatively charged chloride ions into the cell or positively charged potassium ions out of the cell. This action results in a negative change in the transmembrane potential, usually causing hyperpolarization. Two general classes of GABA receptor are known: GABA_A in which the receptor is part of a ligand-gated ion channel complex, and GABA_B metabotropic receptors, which are G protein-coupled receptors that open or close ion channels via intermediaries (G proteins).

[0076] GABA agonists well known in the art include muscimol, progabide, riluzole, baclofen, gabapentin (NEURONTIN®), vigabatrin, valproic acid, tiagabine (GABITRIL®), lamotrigine (LAMICTAL®), pregabalin, phenytoin (DILANTIN®), carbamazepine (TEGRETOL®), topiramate (TOPAMAX®) and analogs, derivatives, prodrugs and pharmaceutically acceptable salts of those GABA agonists. It will be recognized by those skilled in the art that other GABA agonists are also useful in the combinations, pharmaceutical compositions, methods and kits of this invention. GABA agonists have been disclosed to be useful in anti-seizure therapy for central nervous system disorders such as epilepsy, Huntington's chorea, cerebral ischemia, Parkinson's disease, tardive dyskinesia and spasticity. GABA agonists have also been disclosed to be useful as antidepressants, anxiolytics, antipsychotics, and to have utility in the treatment of pain.

[0077] VGAT (vesicular GABA transporter) (also known as vesicular inhibitory amino acid transporter (VIAAT)) is a protein that in humans is encoded by the SLC32A1 gene (also known as the VGAT gene). VGAT is highly concentrated in the nerve endings of GABAergic neurons in the brain and spinal cord but also in glycinergic nerve endings. Caudhry, et al., J. Neurosci., 18(23):9733-9750 (1998), incorporated herein by reference. Nucleic acid and amino acid sequences for human VGAT are known in the art. See, for example, GenBank Accession No.: Q9H598, human Vesicular inhibitory amino acid transporter (VIAAT/VGAT), which provides the amino acid sequence (SEQ ID NO: 3):

MATLLRSKLSNVATSVSNKSQAKMSGMFARMGFQAATDEEAVGFAHCDLDFEHR
QGLQMDILKAEGEPCGDEGAEAPVEGDIHYQRGSGAPLPPSGSKDQVGGGEFGGH
DKPKITAWEAGWNVTNAIQGMFVLGLPYAILHGGYLGLFLIIFAAVVCCYTGKILIAC
LYEENEDGEVVRVRDSYVAIANACCAPRFPTLGGRRVVNAQIIELVMTCILYVVVSG

NLMYNSFPGLPVSQKSWIIATAVLLPCAFLKNLKAVSKFSLLCTLAHFVINILVIAYC
 LSRARDWAWEVKFYIDVKKFPISIGIIVFSYTSQIFLPSLEGNMQQPSEFHCMMNWT
 HIAACVLKGLFALVAYLTWADETKEVITDNLPGSIRAVVNIFLVAKALLSYPLPFFAA
 VEVLEKSLFQEGRSRAFFPACYSGDGRLKSWGLTLRCALVVFLLMAIYVPHFALLMG
 LTGSLTGAGLCFLPLSLFHLRLLWRKLLWHQVFFDVAIFVIGGICSVSGFVHSLEGLIE
 AYRTNAED.

See also, for example, GenBank Accession No.: NM_080552, Homo sapiens solute carrier family 32 member 1 (SLC32A1), mRNA, which provides the nucleic acid sequence (SEQ ID NO: 4):

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 1 gctcgcgccc cgcggcagct ccgcagtgca ctagccacca cgcgcgcgc cggcgctccg
 61 ccagacctgc tgccagctt cccggtccag ccctgagaga gcctcgaacg ccagctgcga
121 gggtcatgag ccagagagcc cccgggcgcc ggcggagag caagcggaga tagcactt
181 ggcgcgcgc cccctcgct tcttcgtatcg ctgtccgc atcctcggtt cttctgtcc
241 ttccgcgtt ccccaaccgcc gccatggcca ctttgctccg cagcaagctg tccaaacgtgg
301 ccacgtccgt gtccaaacaag tcccaaggcca agatgagcgg catgtcgcc aggtatgggtt
361 ttcaggcgcc cacggatgag gaggcgggtt gcttcgcgc ttgcgacgac ctgcacttt
421 agcaccgcgc gggcgtcgag atggacatcc tgaaagccg gggagagccc tgcggggac
481 agggcgctga agcgcgcgtc gagggagaca tccattatca gcgaggcagc gagctcctc
541 tggccgcctc cggctccaag gaccagggtt gaggtgggtt cgaattcggg ggcacgcaca
601 agcccaaaat cacggcggtt gaggcaggct ggaacgtgac caacgcgcac cagggcatgt
661 tcgtgctggg cttaccctac gccatccgc acggcggtt cctgggtt tttctcatca
721 tcttcgcgc cgttgtgtc tgctacaccg gcaagatctt catcgctgc ctgtacgagg
781 agaatgaaga cggcgagggtt gtgcgcgtc gggactcgta cgtggccata gccaacgcct
841 gctgcgcgc cggctccca acgctggcg gccgagtgtt gaacgtagcg cagatcatcg
901 agctggtgat gacgtgcata ctgtacgtt gggtgatgtt caacccatcg tacaacagct
961 tcccgggctt gcccgtgtc cagaagtcctt gttccattat cgccacggcc gtgcgtgc
1021 cttgcgcctt ctttaagaac ctcaaggccg tgcctcaaggat cagtcgtgc tgcactctgg
1081 cccacttcgt catcaatatac ctggctcatag cttactgtt atcgcggcg cgcactggg
1141 cctggggagaa ggtcaagtt tacatcgacg tcaagaaggat ccccatctcc attggcatca
1201 tcgtgttcag ctacacgtt cagatcttc tgccttcgtt ggaggcaat atgcagcagc
1261 ccagcgagtt ccactgtcatg atgaacttgg cgcacatcg acgcctgcgtt ctcaaggggcc
1321 tcttcgcgtt cgtcgccatc ctcacccgtt ccgacgagac caaggagggtt atcacaaggata
1381 acctggccgg ctccatccgc gccgtggta acatctttt ggtggccaag ggcgtgtt
1441 cctatccctt gccattctt ggcgtgtc aggtgttggaa gaagtcgtc ttccagggaa
1501 gcagccgcgc ctttttcccg gctgttaca gcccggacgg ggcctgaag tccctggggc
1561 tgacgtgcgtt ctgcgcgtc gtcgtttca cgtgttcat ggcattttt gtgcgcact
1621 tcgcgtgtt catgggcctt accggcagcc tcaacggccg cggccctgtt ttcttgcgtc
1681 ccagccctt tcacccgtc ctgcgttgc gcaagctgtt gtggcaccat gtccttcgtt
1741 acgtcgccat cttcgatc ggcggcatct gcaagcggtt cggcgttgc cactccctcg
1801 agggcctcat cgaaggcttac cgaaccaacg cggaggacta gggcgcacagg ggcggccccc
1861 gccgcgcctt tgcgtcttcc ccccttcctt ctcacccgc ccccaaccaggc ccagtgcgc
1921 ctgcgcgcgc gcttggggagg ccaagtttta aacatctctg gttccatgtt tctgattatt
1981 cggggatggg ggggatggg ggggacaggattt ctcacgttcat ctcgttgcgtt ggcgttctgt
2041 tgcgtttttt ttccacaaac accctggttt tggggggagg cgggggttcat ttgcgggcag
2101 ggttctctgt cttccaaagt gggggcccgaa cactttggtt ccagtcatcg aggggggttgg
2161 gaaggggaggg agagggggccg cagctcgac ggcgtggcaac ttgacccatgg gggaaatattt
2221 cacatccatc cagactcgatc aatctacatcg gtccttcgtt ccacccatcg aacgcgttcc
2281 attccggaga cgtttcaacc ctgcacgggg aaaggctgtac tggaaatcc attttgggtt
2341 gggcaatttcc ttcaacacgaa ccggaaaggcg agaagccgcg gcccggccag cttgcgttgc
2401 ggttttcagg aatctaaact ctcatcttgc gcaatttac aggtgtggaa ctgttctact
2461 gtgcgtgtgg tgcgttgcgtt gtcataaaga tggaaatgtat atcagaaaaaa aatctatctc
2521 taatttagag tgcgttcatc aattatatcc gcaaaataaaag aagagacaaa ggctaaaaaa
2581 a.
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[0078] In addition, nucleic acid sequences for rat VGAT are known in the art. See, for example, GenBank Accession No.: AF030253.1, *Rattus norvegicus vesicular GABA transporter (VGAT) mRNA, complete cds*, which provides the nucleic acid sequence (SEQ ID NO: 6):

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1  agcggagata gcggcccttg ctgccttgac ggcgcggccgc cgcgtccccca gacccttctg
61 tcctttctc cggccccggcc gcccgcattgg ccaccctgtt cgcgcagcaag ctgaccaacg
121 tggccacccctc tttgttccaaac aagtcccagg ccaagggttag cggcatgttc gccaggatgg
181 ggtttcaggc ggcacccggat gaggaggccg tgggcttcgc gcactgcgcac gatctcgact
241 ttgagccaccg ccaggccctg cagatggaca ttctgaaatc ggaaggcgag ccctgcgggg
301 acgaggccgc agaacacctcc gtcgaggagg acattcattt tcagcgcggc ggcgcgtcccc
361 tgccaccctc gggcttccaaag gaccaggccg tgggagactgg tggggagttc gggggtcac
421 acaaacccaa gatcacggcg tgggaagccg gctggaaacgt gacaaacgcattcaggggca
481 tgttcgtgtt gggcttaccc tacccatcc tccacccggg ctacctgggg ttgttccctca
541 tcatacttcgc cgcgggtgggt tgctgttaca cccggcaagat cctcatcgcc tgcctgttac
601 aggagaacga agatggtagt gttgtgcgc tgaggactc gtatgtggcc atagctaacc
661 cgtgtgcgc tcctcgattt cccacgcgtt gcccgcgcgt ggtcaatgtt gcccagatca
721 tcgagctgggt gatgacgtgtt atcttgcgtt tagtggtagt cggcaacccat atgtacaaca
781 gtttcccccggg gctgcccgtt tcgcagaagt cctggccat catagccacg gcggtgttgc
841 tgccctgcgc ctcccttgcgg aatctcaagg ccgtgtccaa gttcagtctg ctgtgcacgc
901 tggcccaactt cgtcatcaac atccctggtca tccgcctactg tctctcgcc ggcgcgttact
961 gggccctggga gaaggtagt ttctacatcg acgtcaagaa gtttccatcc tccatcgcc
1021 tcatactgtt cagctacacg tcgcagatct ttctgcctt cgtcaaggc aacatgcac
1081 agcccaacgcgaa attccactgc atgatgaact ggacacacat cgcgcctgc gtgctcaagg
1141 gtctcttcgc gctcgccgcg taccttccat gggccgcacca gaccaaggaa gtcatcacgg
1201 ataacacttgc cgggttccatc cgcgcgttgc tcaacatctt cctggggcc aaggcgcttgc
1261 tgccttaccc gttgccttgc ttgcggccg tcgaagtgtt ggagaagtct ctcttccagg
1321 aaggcagtcg tgccttcttc cccgcctgtt acgggtggcgaa cggtcgcctt aagtccctggg
1381 ggctgacgct ggcgtgcgcg ctgggtgtt tcacgctgtt catggccatc tacgtgcac
1441 acttcgcgcgct gctcatgggc ctcacggca gcctcacggg agccggcctc tgcttccgtt
1501 tgcccaacgcctt cttccacttgc cgtttcttgc ggcgcacactt gctgtggcac caggcttct
1561 tcgatgtggc cattttcgtt atcggccggca tctgcagctg gtccggcttc gtgcatttcac
1621 tcgagggcctt catcgaggcc taccaacca acgcagaggg cttagggggcg gggaccctgc
1681 ccccaacttgc ctcccccgc acccccaacttgc ccccttatcc cgcgcgcgc ccccaaccc
1741 cagcccccttgc cgcacccacg ctggggaggc cggactttaa acacccccc ttccctagtt
1801 ctgattattt ggggaccggg cggggggagggg agggggatag acatccaagg tccactgcgt
1861 ctgcgtttctt gtcgttctt ctattccaca tcgttctgtt ttggggggag ggagcagac
1921 gtataagtga agggatattt ctgttcttcc tagaacaccc accaccacca ccaccaaact
1981 ttggctccag tcaatgttag ggggtggaaag ggagggggaa agggaaacacg cagttcgac
2041 gctcgaaac ttgaccttgg ggggtgggtt gggggacattt cacagccatt cagtgcttgg
2101 aatctactgc gtccagccat ttccagcaag agcgccttcc atgccttaga catttcaacc
2161 ttgaggccctg aaaggcttgc cggggaaatcc atttcggggca ggcgcacttcc ctctggagaa
2221 gcccggccag gggcccccgtt tgcctgcgg gttttcaggaa accccaaacttcc atcttgcac
2281 atgtatccgg ttgttggaaact gtatactgtt cgtgtgggtt gtcgttgggtt aataagatga
2341 aatgtatatac agaaaaaaatc tatcttaat ttagagtgcc gtcgcctcgcc cc.

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[0079] In human patients as well as in animal models of chronic neuropathic pain, the mechanism as well as associated neuropathological changes leading to neuropathic pain states are relatively well defined. These can be represented by the presence of peripheral nerve injury-induced neuroma or partial spinal segmental traumatic or ischemic injury. As such, the segmental level as well as the site of the origin of pain-inducing stimuli can be

identified. Accordingly, the present invention provides treatment strategies which would selectively target a key population of spinal neurons responsible for the transmission of nociceptive stimuli at relevant spinal segments, yet not affect other neuronal populations (such as α -motoneurons or interneurons in non-targeted neighboring segments). The data provided herein demonstrates that subpial delivery of GAD65 and VGAT genes into L3-L5 spinal segments of a mouse model of neuropathic pain provides a potent and long-lasting anti-nociceptive effect.

[0080] Preferential expression of GAD65 gene in infected astrocytes (as opposed to neurons) appears to provide a specific advantage with respect to expected GABA mediated anti-spasticity effect (see, *e.g.*, WO2014/116652 and PCT/US2017/024285, each of which is incorporated herein by reference). As has been shown *in vitro*, infection of primary astrocytes led to a Ca^{2+} independent increase in extracellular GABA concentration. Accordingly, it is expected that astrocyte-mediated GABA release in the spinal parenchyma will be independent of the functionality and connectivity of local neuronal inhibitory circuitry and will specifically exert its hyperpolarizing effect on GABA_B receptor expressed on Ia afferents and/or α -motoneurons. The biological activity of astrocyte-produced GABA was confirmed by its depolarization-inducing effect on preferentially GABA_A receptor-expressing cultured hNT neurons.

[0081] The use of a dual GAD65 and VGAT gene therapy represents a novel approach not previously tested in the context of spinal or brain delivery with the goal to increase regional neuronal inhibition. First, this approach uses subpial administration of a vector delivered in small quantities, which leads to only a localized infection of neurons in the dorsal horn of the vector-injected segment that is unilateral to the site of peripheral nerve injury. This is in contrast to an intrathecal delivery technique where the vector is effective in infecting ventral horn neurons and dorsal root ganglion cells throughout the injected regions (left and right and over multiple segments). No dorsal horn neurons (which are the primary neuronal population responsible for spinal transmission of nociceptive stimuli into brain) are infected after intrathecal delivery of the vector. Second, combination of the GAD65 and VGAT genes is required for effective induction of nociceptive effect by potentiating release of vesicular GABA from infected dorsal horn neurons into synaptic cleft.

[0082] Accordingly, in one aspect, the invention provides a method of treating neuropathic pain in a subject by spinal-specific upregulation of the GAD65 gene and VGAT gene. In various embodiments, the method includes subpial administration of a viral vector encoding GAD65 and VGAT, and expressing the GAD65 and VGAT distally, contralaterally, and/or ipsilaterally to the administration site at a therapeutic level of the subject, thereby decreasing neuropathic pain in the subject. In various embodiments, the vector includes a nucleotide sequence encoding GAD65 and VGAT. Also within the scope of the invention is a polypeptide encoded by a nucleotide sequence that has at least 60% homology to GAD65 or a functional fragment thereof (*i.e.*, a polypeptide encoded by nucleotide sequence that has about 70% homology, about 75% homology, about 80% homology, about 85% homology, about 90% homology, about 95% homology, about 99% homology to GAD65 or a fragment thereof). Also within the scope of the invention is a polypeptide encoded by nucleotide sequence that has at least 60% homology to VGAT or a fragment thereof (*i.e.*, a polypeptide encoded by nucleotide sequence that about 70% homology, about 75% homology, about 80% homology, about 85% homology, about 90% homology, about 95% homology, about 99% homology to VGAT or a fragment thereof).

[0083] Viral vectors can be particularly useful for introducing a polynucleotide useful in performing a method of the invention into a target cell. Viral vectors have been developed for use in particular host systems, particularly mammalian systems and include, for example, retroviral vectors, other lentivirus vectors such as those based on the human immunodeficiency virus (HIV), adenovirus vectors (AV), adeno-associated virus vectors (AAV), herpes virus vectors, vaccinia virus vectors, and the like (see Miller and Rosman, *BioTechniques* 7:980-990, 1992; Anderson *et al.*, *Nature* 392:25-30 *Suppl.*, 1998; Verma and Somia, *Nature* 389:239-242, 1997; Wilson, *New Engl. J. Med.* 334:1185-1187 (1996), each of which is incorporated herein by reference). In one aspect of the invention, a lentivirus, an AV, or an AAV is utilized.

[0084] Adenoviruses represent the largest nonenveloped viruses, because they are the maximum size able to be transported through the endosome (*i.e.*, envelope fusion is not necessary). The virion also has a unique “spike” or fibre associated with each penton base of the capsid that aids in attachment to the host cell. AAV is a dependent parvovirus that by definition requires co-infection with another virus (typically an adenovirus or herpesvirus) to initiate and sustain a productive infectious cycle. In the absence of such a helper virus, AAV

is still competent to infect or transduce a target cell by receptor-mediated binding and internalization, penetrating the nucleus in both non-dividing and dividing cells.

[0085] Once in the nucleus, the virus uncoats and the transgene is expressed from a number of different forms--the most persistent of which are circular monomers. AAV will integrate into the genome of 1-5% of cells that are stably transduced (Nakai *et al.*, *J. Virol.* 76: 11343-349, 2002). Expression of the transgene can be exceptionally stable. Because progeny virus is not produced from AAV infection in the absence of helper virus, the extent of transduction is restricted only to the initial cells that are infected with the virus. It is this feature which makes AAV a suitable gene therapy vector for the present invention.

[0086] Additional references describing adenovirus vectors and other viral vectors which could be used in the methods of the present invention include the following: Horwitz, M. S., *Adenoviridae and Their Replication*, in Fields, B., et al. (eds.) *Virology*, Vol. 2, Raven Press New York, pp. 1679-1721, 1990); Graham, F., et al., pp. 109-128 in *Methods in Molecular Biology*, Vol. 7: *Gene Transfer and Expression Protocols*, Murray, E. (ed.), Humana Press, Clifton, N.J. (1991); Miller, N., et al., *FASEB Journal* 9: 190-199, 1995; Schreier, H., *Pharmaceutica Acta Helveticae* 68: 145-159, 1994; Schneider and French, *Circulation* 88:1937-1942, 1993; Curiel D. T., et al., *Human Gene Therapy* 3: 147-154, 1992; Graham, F. L., et al., WO 95/00655 (5 Jan. 1995); Falck-Pedersen, E. S., WO 95/16772 (22 Jun. 1995); Denefle, P. et al., WO 95/23867 (8 Sep. 1995); Haddada, H. et al., WO 94/26914 (24 Nov. 1994); Perricaudet, M. et al., WO 95/02697 (26 Jan. 1995); Zhang, W., et al., WO 95/25071 (12 Oct. 1995). A variety of adenovirus plasmids are also available from commercial sources, including, e.g., Microbix Biosystems of Toronto, Ontario (see, e.g., Microbix Product Information Sheet: Plasmids for Adenovirus Vector Construction, 1996).

[0087] Additional references describing AAV vectors which could be used in the methods of the present invention include the following: Carter, B., *Handbook of Parvoviruses*, vol. I, pp. 169-228, 1990; Berns, *Virology*, pp. 1743-1764 (Raven Press 1990); Carter, B., *Curr. Opin. Biotechnol.*, 3: 533-539, 1992; Muzyczka, N., *Current Topics in Microbiology and Immunology*, 158: 92-129, 1992; Flotte, T. R., et al., *Am. J. Respir. Cell Mol. Biol.* 7:349-356, 1992; Chatterjee et al., *Ann. NY Acad. Sci.*, 770: 79-90, 1995; Flotte, T. R., et al., WO 95/13365 (18 May 1995); Trempe, J. P., et al., WO 95/13392 (18 May 1995); Kotin, R.,

Human Gene Therapy, 5: 793-801, 1994; Flotte, T. R., et al., *Gene Therapy* 2:357-362, 1995; Allen, J. M., WO 96/17947 (13 Jun. 1996); and Du et al., *Gene Therapy* 3: 254-261, 1996.

[0088] As used herein, the term “adeno-associated virus” (AAV), includes but is not limited to, AAV type 1, AAV type 2, AAV type 3 (including types 3A and 3B), AAV type 4, AAV type 5, AAV type 6, AAV type 7, AAV type 8, AAV type 9, AAV type 10, AAV type 11, avian AAV, bovine AAV, canine AAV, equine AAV, ovine AAV, and any other AAV now known. AAV vectors are derived from single-stranded (ss) DNA parvoviruses that are nonpathogenic for mammals (reviewed in Muzyscka (1992) *Curr. Top. Microb. Immunol.*, 158:97-129, incorporated herein by reference). Briefly, AAV-based vectors have the rep and cap viral genes that account for 96% of the viral genome removed, leaving the two flanking 145-basepair (bp) inverted terminal repeats (ITRs), which are used to initiate viral DNA replication, packaging and integration. In the absence of helper virus, wild-type AAV integrates into the human host-cell genome with preferential site-specificity at chromosome 19q 13.3 or it may remain expressed episomally. A single AAV particle can accommodate up to 5 kb of ssDNA, therefore leaving about 4.5 kb for a transgene and regulatory elements, which is typically sufficient. However, trans-splicing systems as described, for example, in U.S. Pat. No. 6,544,785, may nearly double this limit. Adeno-associated virus of many serotypes have been extensively studied and characterized as gene therapy vectors. Those skilled in the art will be familiar with the preparation of functional AAV-based gene therapy vectors. Numerous references to various methods of AAV production, purification and preparation for administration to human subjects can be found in the extensive body of published literature (see, e.g., *Viral Vectors for Gene Therapy: Methods and Protocols*, ed. Machida, Humana Press, 2003, incorporated herein by reference). Additionally, AAV-based gene therapy targeted to cells of the CNS has been described in U.S. Pat. Nos. 6,180,613 and 6,503,888 (each of which are incorporated herein by reference).

[0089] Optionally, the AAV viral capsid is AAV2/9, AAV9, AAVrh8, AAVrh10, AAV Anc80, or AAV PHP.B; however, the serotype of the viral capsid used in certain embodiments of the invention can be selected from among known viral capsids, including AAV viral capsids of other known serotypes.

[0090] Optionally, the gene therapy vector, e.g., AAV or AAV-based vector, can be modified to improve virus uptake into the target tissue of interest, viral stability, and tropism.

For example, the capsid of an AAV vector may be modified with a ligand (*e.g.*, synthetic or naturally occurring small molecule, peptide, or polypeptide, or other biomolecule) that binds to a receptor at or in the tissue of interest. Other modifications are possible to improve and/or enhance the functional properties of the vector being used to both target the tissue of interest and allow the construct to enter and effectively transduce the target cells. Such modifications will be within the skill set of a person having ordinary skill in the art.

[0091] Depending on the host cell/vector system utilized, any of a number of suitable transcription and translation elements, including constitutive and inducible promoters, transcription enhancer elements, transcription terminators, and the like can be used in the expression vector (Bitter *et al.*, *Meth. Enzymol.* 153:516-544, 1987). As defined above, reference to a “promoter” or “promoter sequence” is to be taken in its broadest context and includes a DNA regulatory region capable of binding RNA polymerase in a cell and initiating transcription of a polynucleotide or polypeptide coding sequence such as messenger RNA, ribosomal RNAs, small nuclear or nucleolar RNAs or any kind of RNA transcribed by any class of any RNA polymerase. “Promoters” contemplated herein may also include the transcriptional regulatory sequences of a classical genomic gene, including the Goldberg-Hogness box which is required for accurate transcription initiation in eukaryotic cells, with or without a CAT box sequence and additional regulatory elements (*i.e.*, upstream activating sequences, enhancers and silencers).

[0092] Placing a sequence under the regulatory control of a promoter sequence means positioning said molecule such that expression is controlled by the promoter sequence. Promoters are generally positioned 5' (upstream) to the genes that they control. In the construction of heterologous promoter/structural gene combinations, generally promoter position may be a distance from the gene transcription start site that is approximately the same as the distance between that promoter and the gene it controls in its natural setting, *i.e.*, the gene from which the promoter is derived. As is known in the art, some variation in this distance can be accommodated without loss of promoter function. Similarly, the positioning of a regulatory sequence element with respect to a heterologous gene to be placed under its control is defined by the positioning of the element in its natural setting, *i.e.*, the genes from which it is derived. Again, as is known in the art, some variation in this distance can also occur.

[0093] Promoter sequences having differing characteristics and expression profiles are well known in the art, including those that are tissue-specific, tissue-non-specific, constitutive, and inducible. Reference can be further made to, for example, Papadakis et al., "Promoters and Control Elements: Designing Expression Cassettes for Gene Therapy." Current Gene Therapy, 2004, 4, 89-113, the contents of which are incorporated herein by reference. Promoters contemplated by the present invention include, but are not limited to: Apo A-I, ApoE, serpina (TBG), alpha-1-antitrypsin (hAAT) (liver specific); MCK (muscle specific); GFAP, NSE, Synapsin I, Preproenkephalin, Dopamine b-hydroxylase (dbH), Prolactin, Myelin basic protein (neuronal-specific), GUSB, CBA, CAG, Ankyrin (erythroid specific), human ubiquitin promoter (UBI) and human synapsin promoter. However, other known tissue-specific or cell-specific promoters may be used.

[0094] Suitable host cells for producing recombinant AAV particles include, but are not limited to, microorganisms, yeast cells, insect cells, and mammalian cells, that can be, or have been, used as recipients of a exogenous nucleic acid molecule. Thus, a "host cell" as used herein generally refers to a cell which has been transfected with an exogenous nucleic acid molecule. The host cell includes any eukaryotic cell or cell line so long as the cell or cell line is not incompatible with the protein to be expressed, the selection system chosen or the fermentation system employed.

[0095] The AAV vectors can be formulated into preparations for injection or administration by dissolving, suspending or emulsifying them in appropriate, pharmaceutically acceptable carriers or diluents. Examples of such pharmaceutically acceptable carriers or diluents include an aqueous or nonaqueous solvent, such as oils, synthetic aliphatic acid glycerides, esters of higher aliphatic acids or propylene glycol; and if desired, with conventional additives such as solubilizers, isotonic agents, suspending agents, emulsifying agents, stabilizers and preservatives.

[0096] If a viral vector specific for the cell type is not available, the vector can be modified to express a receptor (or ligand) specific for a ligand (or receptor) expressed on the target cell, or can be encapsulated within a liposome, which also can be modified to include such a ligand (or receptor). A peptide agent can be introduced into a cell by various methods, including, for example, by engineering the peptide to contain a protein transduction domain such as the human immunodeficiency virus TAT protein transduction domain, which can facilitate

translocation of the peptide into the cell. In addition, there are a variety of biomaterial-based technologies such as nano-cages and pharmacological delivery wafers (such as used in brain cancer chemotherapeutics) which may also be modified to accommodate this technology.

[0097] Accordingly, in another aspect, the present invention provides gene therapy vectors or constructs comprising GAD65 and/or VGAT genes, or derivatives and/or mutants thereof, which are operably linked to at least a promoter element that is capable of being expressed in a tissue of the central nervous system. As demonstrated herein using accepted mouse, rat, and pig models, the gene therapy vectors of the present invention were effective in treating neuropathic pain and/or muscle spasticity.

[0098] Thus, in various embodiments, the serotype of the viral vector used in the invention may be selected from the group consisting of AAV1, AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAVrh8, AAVrh10, AAVrh33, AAV rh34, AAV Anc80 (Anc80), AAV PHP.B, and others (see, *e.g.*, Cao et al. (2002) PNAS, 99:11854-11859; and *Viral Vectors for Gene Therapy: Methods and Protocols*, ed. Machida, Humana Press, 2003, incorporated herein by reference). Other serotypes besides those listed herein are also contemplated. In certain exemplary embodiments, AAV9 or Anc80 are used. It is also contemplated that the disclosed compositions and methods may use AAV chimeric vectors, whereby portions of AAV are fused with other similar vectors, such as Adenovirus.

[0099] In various embodiments, the gene therapy constructs described herein may also comprise a vector (or gene therapy expression vector) into which the gene(s) of interest (*e.g.*, GAD65 and/or VGAT) is cloned or otherwise which includes the gene(s) of interest in a manner such that the nucleotide sequences of the vector allow for the expression (constitutive or otherwise regulated in some manner) of the gene(s) of interest. The vector constructs herein described include any suitable gene expression vector that is capable of being delivered to a tissue of interest (*e.g.*, CNS) and which will provide for the expression of the gene of interest in the selected tissue of interest (*e.g.*, CNS).

[0100] Accordingly, the present invention also provides a gene therapy composition comprising the GAD65 and/or VGAT -providing vector(s) as described herein. The gene therapy composition of the invention may therefore include the vector(s) as described herein and one or more pharmaceutically acceptable carriers for gaining entry into a cell or tissue,

e.g., a CNS cell or tissue, for treating neuropathic pain or muscle spasticity. Advantageously, the gene therapy composition of the invention provides for a controlled delivery of an active gene, especially a therapeutic gene, to a site of action at an optimum rate and therapeutic dose. Association of the gene therapy vector and/or viral vector containing such gene therapy vector with a delivery system enables, in particular, its specific delivery to the site of action or its controlled expression of genes after targeting the action site.

[0101] In addition to cells integrating gene transfer after the use of lentiviral vectors, there are reports of successful GAD65 gene overexpression after AAV-GAD65 injections into subthalamic nuclei. In those studies, persistent GAD65 expression was seen up to 4-5 months after AAV-GAD65 injections. More importantly, recent systematic data demonstrate a high efficiency of AAV-based gene delivery into rat or minipig striatum even after a limited number of AAV injections (1-2 injections). Thus, in another embodiment, the present invention employs an AAV-based, genome-non-integrating GAD65-encoding and VGAT-encoding vector to achieve segment-specific GAD65 and VGAT expression.

[0102] As demonstrated herein, by combining spinal delivery (*i.e.*, subpial administration) of GAD65 and VGAT (either by using a single vector encoding both genes, or by using separate vectors encoding each individual gene), a significant and functionally relevant decrease in neuropathic pain was achieved. The potency of spinal inhibition was tested in a well-characterized mouse model of chronic neuropathic pain (see, *e.g.*, Pain 76(1-2): 215-222, 1998, incorporated herein by reference).

[0103] Administering the instant combinational therapy can be effected or performed using any of the various methods and delivery systems known to those skilled in the art. As used herein, the term “administration” or “administering” is defined to include an act of providing a compound or pharmaceutical composition of the invention to a subject in performing the methods of the invention. Exemplary routes of administration include, but are not limited to, intravenously, intraarticularly, intracisternally, intraocularly, intraventricularly, intrathecally, subpially, intramuscularly, intraperitoneally, intradermally, intracavitarily, and the like, as well as combinations of any two or more thereof. In certain embodiments, the vector composition may be delivered directly into the spinal parenchyma, intrathecal space of the spine, into the spinal subpial space of the subject, and/or into the peripheral spastic

muscle to achieve spinal upregulation of the GAD65 gene and VGAT gene. See, e.g., WO2016/122791, incorporated herein by reference.

[0104] Thus, the method provided herein permits spinal subpial gene therapy, such as by use of an AAV, in large animals or in humans. An exemplary delivery system for delivering the vector into the subpial space includes a guiding tube bended at 90° and catheter (e.g., PE-5 or PE-10), which permits precise guidance and placement of the subpial catheter into the dorsal subpial space of targeted spinal cord segments. After placement of the catheter, the vector is infused for a certain amount of time before being removed.

[0105] As used herein, the term “PE-10” refers to polyethylene tubing having an inner diameter of approximately 0.010 inches. In certain embodiments, the inner diameter of the PE-10 tubing will be about 0.011 inches. Likewise, the term “PE-5” refers to polyethylene tubing having an inner diameter of approximately 0.005 inches. In certain embodiments, the inner diameter of the PE-5 tubing will be about 0.008 inches.

[0106] Accordingly, the claimed method provides subpial delivery (*i.e.*, bypassing the pial membrane), which provides near complete spinal parenchymal vector-mediated gene expression in both white and grey matter of the subject being treated. Currently available non-invasive techniques do not permit a comparable level of spinal parenchymal transgene expression or well controlled segment-specific gene silencing.

[0107] An exemplary method for placing the subpial catheter in a mammalian subject, is shown in Figure 7, wherein several sequential procedural steps may be followed to minimize potential spinal injury associated with instruments/catheter manipulation in the vicinity of the exposed “dura-free” spinal cord. As shown in steps A and B of Figure 7, muscle spasticity in hind limbs is induced by Th9 spinal segment transection and spasticity is identified/measured by quantitative change in gastrocnemius muscle EMG response to paw tactile stimulus. 2-3 months after induction of muscle spasticity, the animals received lumbar subpial injection of AAV9 (or Anc80)-UBI-GAD65/VGAT vector and the presence of spasticity response was measured for additional 2 months (Figure 7; steps C and D). After sacrifice the presence of GAD65 and VGAT upregulation is measured by immunofluorescence staining (Figure 7; step E).

[0108] To deliver a gene therapy vector described herein specifically to a particular region of the central nervous system, especially to the spinal subpial space, it may be administered by stereotaxic microinjection. Thus, in various embodiments, the use of caudal and cranial spinal clamps (placed just above and below the laminectomy) may be used to minimize spinal cord pulsation during catheter placement. Also in various embodiments, an “L” shaped catheter stainless steel guiding tube (*e.g.*, a 16-26 G stainless steel tube bended at 90°) mounted on an XYZ manipulator (as described in, for example, US Pub. No. 2015/0224331, incorporated herein by reference) may be used to facilitate subpial catheter placement.

[0109] In certain embodiments, the pia is first punctured using a bent 30G needle. Once the tip of the penetrating needle (*e.g.*, a 30G needle) is in the subpial space for about 1-1.5 mm, the pia may be slightly lifted by 1-2 mm. The subpial catheter is then placed into the subpial space by advancing the catheter from the guiding tube. After the catheter is advanced into the targeted length, the penetrating needle tip of the guiding tube is removed from the subpial space. The vector in a pharmaceutically acceptable carrier may then be injected. Once vector injection is completed (typically over 2-5 min, and in some embodiments, over about 3 min), the catheter is pulled out of the subpial space and the dura is closed. By using this technical approach, placement of the subpial catheter may be accomplished within about 3-5 min from the moment of dura opening. Thereafter, the transgene is expressed distal, contralateral, and/or ipsilateral to the administration site at a therapeutic level.

[0110] The term “therapeutically effective amount” or “effective amount” means the amount of the compound or composition that will elicit the biological or medical response of a tissue, system, animal or human that is being sought by the researcher, veterinarian, medical doctor or other clinician, *e.g.*, spinal upregulation of the GAD65 gene and VGAT gene. Thus, the term “therapeutically effective amount” is used herein to denote any amount of a formulation that causes a substantial improvement in a disease condition when applied to the affected areas repeatedly over a period of time. The amount will vary with the condition being treated, the stage of advancement of the condition, and the type and concentration of formulation applied. Appropriate amounts in any given instance will be readily apparent to those skilled in the art or capable of determination by routine experimentation. For example, a “therapeutically effective amount” of, *e.g.*, a vector encoding the GAD65 gene alone or in combination with the VGAT gene or a composition comprising a vector encoding the GAD65 gene and VGAT gene, with respect to the subject method of treatment, refers to an

amount of the vector in a preparation which, when applied as part of a desired treatment regimen brings about upregulation of the GAD65 gene and VGAT gene. In some cases, multiple doses of the vector are administered. For example, in some embodiments, in addition to the first administration site, a composition comprising a gene therapy vector described herein carrying a transgene is administered to another site that can be contralateral or ipsilateral to the first administration site.

[0111] An effective amount may also depend on the particular vector used. For example, dosages for targeting a CNS tissue may depend on the serotype (*e.g.*, the capsid protein) of the AAV. For example, the AAV may have a capsid protein of an AAV serotype selected from the group consisting of: AAV1, AAV2, AAV5, AAV6, AAV6.2, AAV7, AAV8, AAV9, rh.10, rh.39, rh.43 and CSp3. In certain embodiments, the effective amount of AAV is 10^{10} , 10^{11} , 10^{12} , 10^{13} , or 10^{14} genome copies per kg. In certain embodiments, the effective amount of AAV is 10^{10} , 10^{11} , 10^{12} , 10^{13} , 10^{14} , or 10^{15} genome copies per subject. In experimental mice, the total volume of injected vector, *e.g.*, AAV vector, solution is, for example, between 0.25 μ l and 1.0 μ l; whereas the total volume of injected vector solution in experimental pigs is between 50 μ l and 150 μ l.

[0112] Determining a therapeutically or prophylactically effective amount of the delivery vector can be done based on animal data using routine computational methods. Appropriate doses will depend, among other factors, on the specifics of the transfer vector chosen, on the route of administration, on the mammal being treated (*e.g.*, human or non-human primate or other mammal), age, weight, and general condition of the subject to be treated, the severity of the disorder being treated, the location of the area within the heart being treated and the mode of administration. Thus, the appropriate dosage may vary from patient to patient. An appropriate effective amount can be readily determined by one of skill in the art.

[0113] Human dosage amounts can initially be determined by extrapolating from the amount of compound used in mice, rats and/or pigs, as a skilled artisan recognizes it is routine in the art to modify the dosage for humans compared to animal models. In certain embodiments it is envisioned that the dosage may vary from between about 1 mg compound/Kg body weight to about 5000 mg compound/Kg body weight; or from about 5 mg/Kg body weight to about 4000 mg/Kg body weight or from about 10 mg/Kg body weight to about 3000 mg/Kg body weight; or from about 50 mg/Kg body weight to about 2000

mg/Kg body weight; or from about 100 mg/Kg body weight to about 1000 mg/Kg body weight; or from about 150 mg/Kg body weight to about 500 mg/Kg body weight. In other embodiments this dose may be about 1, 5, 10, 25, 50, 75, 100, 150, 200, 250, 300, 350, 400, 450, 500, 550, 600, 650, 700, 750, 800, 850, 900, 950, 1000, 1050, 1100, 1150, 1200, 1250, 1300, 1350, 1400, 1450, 1500, 1600, 1700, 1800, 1900, 2000, 2500, 3000, 3500, 4000, 4500, 5000 mg/Kg body weight. In other embodiments, it is envisaged that higher doses may be used, such doses may be in the range of about 5 mg compound/Kg body to about 20 mg compound/Kg body. In other embodiments the doses may be about 8, 10, 12, 14, 16 or 18 mg/Kg body weight. Of course, this dosage amount may be adjusted upward or downward, as is routinely done in such treatment protocols, depending on the results of the initial clinical trials and the needs of a particular patient. In certain embodiments, the dosage may be in terms of vector concentration. For example, the concentration of gene therapy vector described herein may be at least: 0.5, 0.75, 1.0, 1.1, 1.2, 1.3, 1.4, 1.5, 1.6, 1.7, 1.8, 1.9 or 2.0 $\times 10^{13}$ gc/ml.

[0114] Dosage treatment may be a single dose schedule or a multiple dose schedule. Moreover, the subject may be administered as many doses as appropriate. One of skill in the art can readily determine an appropriate number of doses. However, the dosage may need to be adjusted to take into consideration an alternative route of administration, or balance the therapeutic benefit against any side effects. Such dosages may vary depending upon the therapeutic application for which the recombinant vector is employed.

[0115] Optionally, AAV-mediated delivery according to the invention may be combined with delivery by other viral and non-viral vectors. Such other viral vectors including, without limitation, adenoviral vectors, retroviral vectors, lentiviral vectors, herpes simplex virus (HSV) vectors, baculovirus vectors, and synthetic vectors may be readily selected and generated according to methods known in the art. Similarly, non-viral vectors, including, without limitation, liposomes, lipid-based vectors, polyplex vectors, molecular conjugates, polyamines and polycation vectors, may be readily selected and generated according to methods known in the art. When administered by these alternative routes, the dosage is desirable in the range described above.

[0116] The gene therapy compositions of the invention can be included in a kit and/or pharmaceutical package, container, pack, or dispenser together with instructions for

administration. Thus, the present disclosure provides kits for the treatment neuropathic pain and/or muscle spasticity. In one embodiment, the kit includes a therapeutic composition containing an effective amount of gene therapy vector in unit dosage form. In some embodiments, the kit comprises a sterile container which contains a therapeutic composition; such containers can be boxes, ampoules, bottles, vials, tubes, bags, pouches, blister-packs, or other suitable container forms known in the art. Such containers can be made of plastic, glass, laminated paper, metal foil, or other materials suitable for holding medicaments. The instructions will generally include information about the use of the composition for the treatment of neuropathic pain and/or muscle spasticity. In other embodiments, the instructions include at least one of the following: description of the composition; dosage schedule and administration for treatment neuropathic pain and/or muscle spasticity or symptoms associated therewith; precautions; warnings; indications; counter-indications; overdosage information; adverse reactions; animal pharmacology; clinical studies; and/or references. The instructions may be printed directly on the container (when present), or as a label applied to the container, or as a separate sheet, pamphlet, card, or folder supplied in or with the container.

[0117] In another aspect, the method of treating neuropathic pain may include a combined therapy wherein local segmental upregulation of the one or more genes described herein is performed in combination with systemic treatment with a GABA uptake inhibitor, such as tiagabine, thereby potentiating the anti-nociceptive effect to only be present in GAD65/VGAT-overexpressing spinal segments and associated dermatomes. Thus, the treatment regimen may include administering a viral vector encoding the GAD65 gene and the VGAT gene in combination with systemic administration of with a GABA uptake inhibitor.

[0118] In addition, the methods of the invention can be used in the treatment of nerve damage, such as peripheral neuropathy, which is caused by exposure to toxic compounds, including heavy metals (*e.g.*, lead, arsenic, and mercury) and industrial solvents, as well as drugs including chemotherapeutic agents (*e.g.*, vincristine and cisplatin), dapsone, HIV medications (*e.g.*, Zidovudine, Didanosine, Stavudine, Zalcitabine, Ritonavir, and Amprenavir), cholesterol lowering drugs (*e.g.*, Lovastatin, Indapamid, and Gemfibrozil), heart or blood pressure medications (*e.g.*, Amiodarone, Hydralazine, Perhexiline), and Metronidazole.

[0119] The methods of the invention can also be used to treat injury to the nervous system caused by physical, mechanical, or chemical trauma. Thus, the methods can be used in the treatment of peripheral nerve damage caused by physical injury (associated with, *e.g.*, burns, wounds, surgery, and accidents), ischemia, prolonged exposure to cold temperature (*e.g.*, frost-bite), as well as damage to the central nervous system due to, *e.g.*, stroke or intracranial hemorrhage (such as cerebral hemorrhage). Likewise, the methods of the invention can be used in the treatment of chronic pain/nociception caused by such trauma.

[0120] The following examples are intended to illustrate but not limit the invention.

EXAMPLE 1

Vector Delivery in Mouse Model

[0121] Figures 1A and 1B show exemplary subpial delivery of an AAV9 (or Anc80) vector encoding GAD65 (glutamate-decarboxylase 65) and VGAT (vesicular GABA transporter) into targeted segments. Naïve C57BL6 mice had a tight ligature placed around 1/3 to 1/2 of the diameter of the sciatic nerve (unilateral ligation) to induce mechanical allodynia. After nerve injury animals were tested for changes in tactile nociceptive threshold using von Frey filaments and brush-evoked allodynia for 10 days.

[0122] At 10 days after induction of sciatic nerve injury animals received a unilateral subpial injection of AAV9 encoding GAD65 and VGAT gene under ubiquitin promoter (UBI). Two control groups were studied. In the first control group only PBS was injected subpially at L2-L3 in sciatic-nerve-injured animals. In another control group (naïve non-injured animals), no treatment was performed. In a separate group of naïve animals (n=6) using AAV9 encoding GFP, it has been demonstrated that subpial unilateral injection of 0.5 μ l of AAV9 (1.2x10¹³ gc/ml; injected into L2-L3 subpial space) selectively infect unilateral dorsal horn neurons in adult mice (Figure 1C). Thus, the data provided herein demonstrate that use of a subpial vector delivery technique achieves targeted transgene expression that is restricted to dorsal horn neurons and is ipsilateral to vector delivery.

[0123] A consistently high degree of tactile hypersensitivity ipsilateral to the site of sciatic nerve ligation was measured in sciatic nerve-injured animals from day 2 after sciatic nerve ligation when the tactile withdrawal threshold decreased from around 1 g (pre-injury baseline) to about 0.2 g. No changes in tactile hypersensitivity were seen in control PBS-

injected sciatic nerve-injured animals and all animals showed continuing signs of neuropathic pain (*i.e.*, tactile and brush-evoked hypersensitivity) for up to 10 weeks after subpial PBS delivery (Figures 2A and 2B). In contrast, in animals receiving subpial delivery of AAV9-UBI-GAD65/VGAT or Anc80-UBI-GAD65/VGAT, a progressive and complete ($P<0.05$) reversal of tactile and brush-evoked hypersensitivity was seen and persisted for 10 weeks (*i.e.*, the duration of study) (Figures 2A, 2B, 2E and 2F). These data demonstrate that unilateral spinal delivery of AAV9-UBI-GAD65/VGAT is highly potent in providing a long-lasting anti-nociceptive effect. Importantly no detectable side effect, such as motor weakness (which is major problem after using systemically or intrathecally delivered anti-nociceptive drugs) was seen.

[0124] Analysis of open field motor performance (running distance) and ipsilateral hind paw placement pattern (Cat Walk assay) showed a significant increase in running distance and normalization of ipsilateral paw placement in animals treated subpially with AAV9-UBI-GAD65/VGAT (Figures 2C and 2D).

[0125] As shown in Figures 3A-3E, unilateral (ipsilateral) (L2-L4) subpial AAV9-UBI-GAD65/VGAT delivery in adult mice results in induction of mixed inhibitory-excitatory neurotransmitter phenotype in ipsilateral dorsal horn excitatory interneurons. However, staining of contralateral dorsal horn neurons with VGLUT2, GAD65 and VGAT antibodies resulted in detection of no co-localization of VGLUT2 with GAD65 or VGAT. Whereas staining of ipsilateral dorsal horn neurons with the same antibodies demonstrated a clear co-expression of VGLUT2 with VGAT and VGLUT with both GAD65 (Figures 3A-3D; white arrows). Likewise, quantitative densitometry analysis showed a significant increase in GAD65 and VGAT expression in ipsilateral dorsal horn (Figure 3E) injected with AAV9-UBI-GAD65/VGAT vector. No increased expression in contralateral dorsal horn was measured.

[0126] Pre-embedding immune-gold staining with VGLUT2, GAD65 and VGAT antibodies coupled with electron microscopy showed a clear increase in GAD65 and VGAT immunogold-tagged particles in VGLUT2+ terminals in AAV9-UBI-GAD65/VGAT-injected animals (Figures 4C and 4D; PNI+GAD65+VGAT:Ipsi images). Minimal or no co-expression of VGLUT2 with GAD65 and VGAT particles was seen in control PBS-injected animals (Figures 4A and 4B; PNI+PBS: Ipsi images). Fluorescence *in situ* hybridization

showed a clear appearance of double and triple-tagged neurons with VGLUT2, GAD65 and VGAT mRNA in AAV9-UBI-GAD65/VGAT-injected animals (PNI: GAD65+VGAT; Ipsi images) (Figures 4E-4J).

[0127] Thus, co-expression of inhibitory neurotransmitter machinery (*i.e.*, GAD65 and VGAT) in excitatory (VGLUT2) neurons/terminals leads to a measured anti-nociceptive effect with improved motor performance in animal models. Further, the induction of a dual excitatory-inhibitory neurotransmitter phenotype leads to a preferential inhibitory effect after activation of peripheral nociceptive afferents.

EXAMPLE 2

Vector Delivery in Pig Model

[0128] In this example, an adult pig model was used to demonstrate that subpial delivery of a vector achieves a well targeted transgene expression in an animal species that has similar spinal cord dimension to that of an adult human. Adult pigs (Yucatan pigs, 15-25 kg; n=3) received unilateral subpial (L2-L3) Anc80-UBI-Rpl22-3xHA vector delivery (100 μ l; 1.2 \times 10¹³ gc/ml). After vector injections, animals survived for 48 hrs and were then perfusion-fixed with 4% paraformaldehyde. Transverse spinal cord sections were then prepared and stained with anti-HA antibody. Stained sections were analyzed with confocal microscopy. As shown in Figures 5A-5D, dorsal horn neuron-specific Rpl22 protein expression ipsilateral to the side of vector injection was observed.

[0129] A different group of adult pigs (Gottingen - Minnesota; 35-45 kg; n=3) received unilateral subpial (L2-L3) Anc80-UBI-GAD65/VGAT (100 μ l; 1.2 \times 10¹³ gc/ml) to demonstrate induction of mixed excitatory-inhibitory neuronal phenotype in infected neurons, similar to what was observed in the mouse model. After vector injections, the animals survived for 8 weeks and were periodically assessed for open field motor performance. No detectable motor weakness was observed in any animal. At 8 weeks animals were perfusion-fixed with 4% paraformaldehyde and transverse spinal cord sections prepared from L1-L4 segments. Sections were stained with anti-VGLUT2, GAD65 and VGAT antibody and analyzed with confocal microscopy.

[0130] As shown in Figures 6A-6D, staining of ipsilateral dorsal horn neurons with VGLUT2, GAD65 and VGAT antibodies showed clear co-expression of VGLUT2 with

VGAT and GAD65. Thus, by manipulating the volume of vector delivered into subpial space a comparable multisegmental expression of targeted transgene can be achieved in a spinal cord that has similar dimensions to that of an adult human spinal cord.

EXAMPLE 3

Vector Delivery in Rat Model of Chronic Muscle Spasticity

[0131] In a rat model for chronic muscle spasticity (*i.e.*, rats with spinal transection-induced muscle spasticity), vector therapy was provided to demonstrate anti-spasticity effect after subpial delivery of AAV9-UBI-GAD65/VGAT. As shown in Figures 8A and 8B, measurement of muscle spasticity in animals receiving a control vector (AAV9-GFP) showed progressive increase in muscle spasticity for 8 weeks after virus delivery (compared to baseline measured at 2-3 months after spinal transection). In contrast a near complete block of spasticity response was measured in animals receiving AAV9-UBI-GAD65/VGAT vector. Measurement of rate-dependent depression (RDD) of H-reflex showed a significant recovery of RDD in animals treated with AAV9-UBI-GAD65/VGAT vector (Figures 8C and 8D).

[0132] Bilateral (L2-L4) subpial AAV9-UBI-GAD65/VGAT delivery in the adult rat model of chronic muscle spasticity induced mixed inhibitory-excitatory neurotransmitter phenotype in lumbar excitatory interneurons. As shown in Figures 9C and 9D, the appearance of mixed neurotransmitter phenotype in spinal interneurons, as evidenced by co-expression of GAD65 and VGAT in VGLUT2 terminals, was observed. However, no co-expression of GAD65 and VGAT in VGLUT2 terminals was seen in animals receiving control AAV9-UBI-GFP vector (Figures 9A and 9B).

[0133] In comparison to control vector-injected animals a highly significant ($p<0.01$) increase in GAD65, VGAT expression and in number of VGLUT1 and VGLUT2 terminals co-expressing GAD65 and VGAT in gray matter was measured in spastic rats receiving lumbar subpial injection of AAV9-UBI-GAD65/VGAT vector (Table 2). Thus, spinal subpial delivery of AAV9-UBI-GAD65/VGAT vector is highly effective in suppressing chronic muscle spasticity in the well-established rat model of spinal transection-induced hind limb spasticity. Thus, the mechanism of measured anti-spasticity effect is based on induction of mixed excitatory-inhibitory neurotransmitter phenotype in spinal excitatory interneurons and resulting suppression of otherwise exacerbated spastic EMG response.

TABLE 2: Quantitative analysis of GAD65 and VGAT expression

Lumbar Subpial AAV9 delivery	GAD65	VGAT	VGLUT1 + GAD65	VGLUT1 + VGAT	VGLUT2+ GAD65	VGLUT2+ VGAT
Experimental Groups	% Normalized Signal (Integrated Density)					
AAV9-GAD65/VGAT (n=4; 6 sections/animal)	208±19*	166±25*	14±8*	9±3.4*	245±97*	331±67*
AAV9-GFP (n=4; 6 sections/animal)	100±7	100±15	2±1	2±0.9	1±0.7	5±1

[0134] Although the invention has been described with reference to the above examples, it will be understood that modifications and variations are encompassed within the spirit and scope of the invention. Accordingly, the invention is limited only by the following claims.

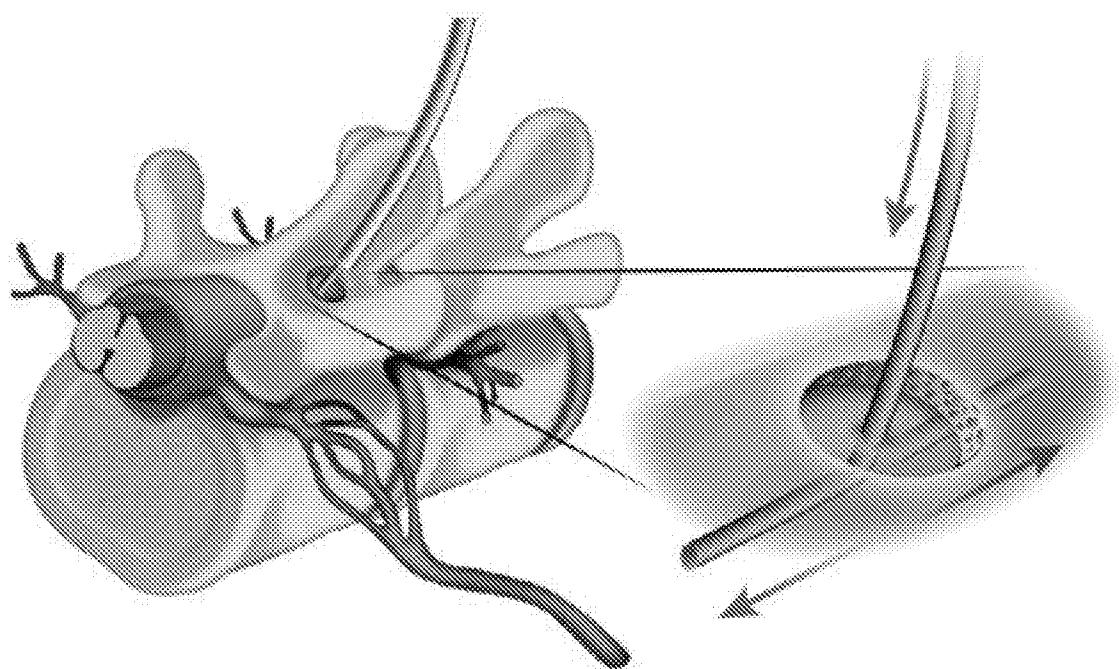
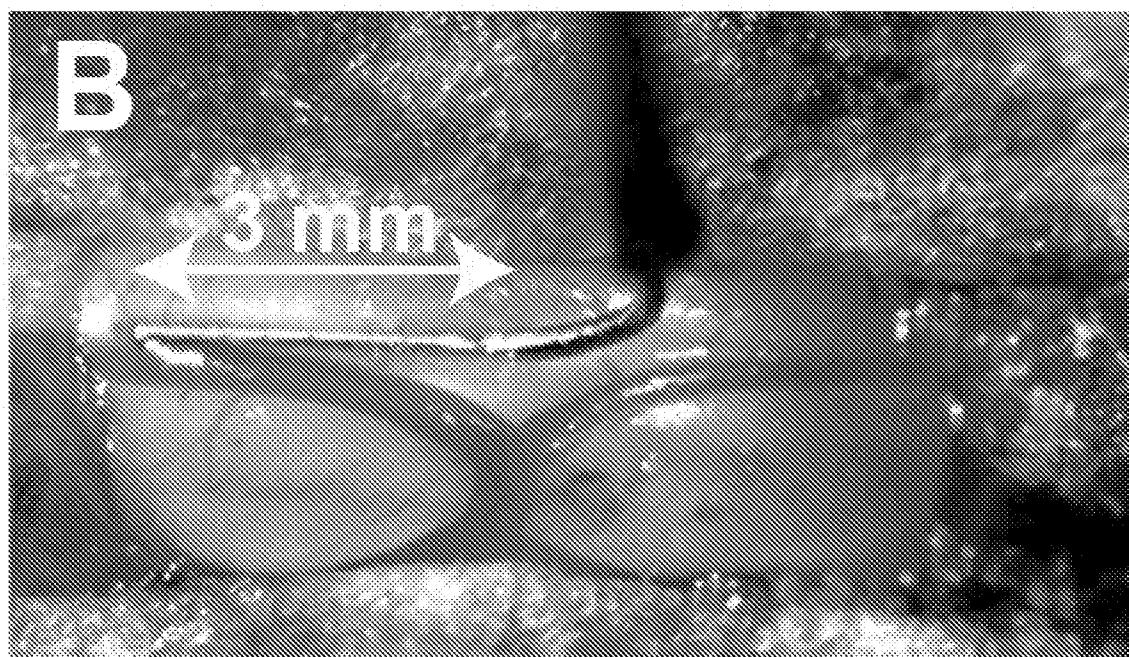
What is claimed is:

1. A method of treating neuropathic pain in a subject comprising subpial administration of a composition comprising: (i) a viral vector sequence, a GAD65 gene sequence, and a VGAT gene sequence under the control of a tissue-specific promoter; and (ii) a pharmaceutically acceptable viral carrier, thereby treating neuropathic pain in the subject.
2. The method of claim 1, wherein the subject is a mammal.
3. The method of claim 1, wherein the nucleic acid construct is encapsidated with an AAV serotype 9 or Anc80 capsid.
4. The method of claim 1, wherein the concentration of the nucleic acid construct in the composition is between about $0.1\text{-}2.0 \times 10^{13}$ gc/ml.
5. The method of claim 1, wherein the pharmaceutically acceptable viral carrier is selected from the group consisting of a lentiviral vector, an adenoviral vector (AV), or an adeno-associated vector (AAV).
6. The method of claim 5, wherein the pharmaceutically acceptable viral carrier is an AAV.
7. The method of claim 6, wherein the AAV is selected from the group consisting of AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV rh8, AAVrh10, AAVrh33, AAV rh34, and AAV Anc80.
8. The method of claim 1, wherein the tissue-specific promoter is selected from the group consisting of human ubiquitin promoter and human synapsin promoter.
9. The method of claim 1, wherein the GAD65 gene sequence is SEQ ID NO: 2 or 5.
10. The method of claim 1, wherein the VGAT gene sequence is SEQ ID NO: 4 or 6.
11. A method for treating neuropathic pain in a subject by gene therapy comprising subpially administering a composition comprising a therapeutically effective amount of a gene therapy construct comprising (i) a viral vector sequence; (ii) a GAD65 gene

sequence; and (iii) a VGAT gene sequence under control of a tissue-specific promoter, and a pharmaceutically acceptable carrier, thereby treating neuropathic pain in the subject.

12. The method of claim 11, wherein the viral vector is selected from the group consisting of a lentiviral vector, an adenoviral vector (AV), or an adeno-associated vector (AAV).
13. The method of claim 12, wherein the viral vector is an AAV.
14. The method of claim 13, wherein the AAV is selected from the group consisting of AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV rh8, AAVrh10, AAVrh33, AAV rh34, and AAV Anc80.
15. The method of claim 11, wherein the gene therapy construct comprises SEQ ID NO: 2 and SEQ ID NO: 4.
16. The method of claim 11, wherein the tissue-specific promoter is selected from the group consisting of human ubiquitin promoter and human synapsin promoter.
17. The method of claim 11, wherein the composition comprises the gene therapy construct at a concentration of about $0.1\text{-}2.0 \times 10^{13}$ gc/ml.
18. The method of claim 11, wherein the subject is a mammal.
19. A method for treating neuropathic pain in a subject comprising subpial administration of a first vector encoding GAD65 and a second vector encoding VGAT, thereby treating neuropathic pain in the subject.
20. The method of claim 19, wherein the subject is a mammal.
21. The method of claim 19, wherein each of the first and second vectors are independently selected from the group consisting of a lentiviral vector, an adenoviral vector (AV), or an adeno-associated vector (AAV).
22. The method of claim 21, wherein each of the first and second vectors is an AAV.

23. The method of claim 22, wherein the AAV is selected from the group consisting of AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV rh8, AAVrh10, AAVrh33, AAV rh34, and AAV Anc80.
24. The method of claim 19, wherein first vector comprises SEQ ID NO: 2 and wherein the second vector comprises SEQ ID NO: 4.
25. A nucleic acid construct comprising: a viral vector sequence, a GAD65 gene sequence, and a VGAT gene sequence under the control of a tissue-specific promoter.
26. The nucleic acid construct of claim 25, wherein the tissue-specific promoter is selected from the group consisting of human ubiquitin promoter and human synapsin promoter.
27. The nucleic acid construct of claim 25, wherein the viral vector is an adeno-associated viral (AAV) vector.
28. The nucleic acid construct of claim 27, wherein the AAV is selected from the group consisting of AAV2, AAV3, AAV4, AAV5, AAV6, AAV7, AAV8, AAV9, AAV rh8, AAVrh10, AAVrh33, AAV rh34, and AAV Anc80.

**FIG. 1A****FIG. 1B**

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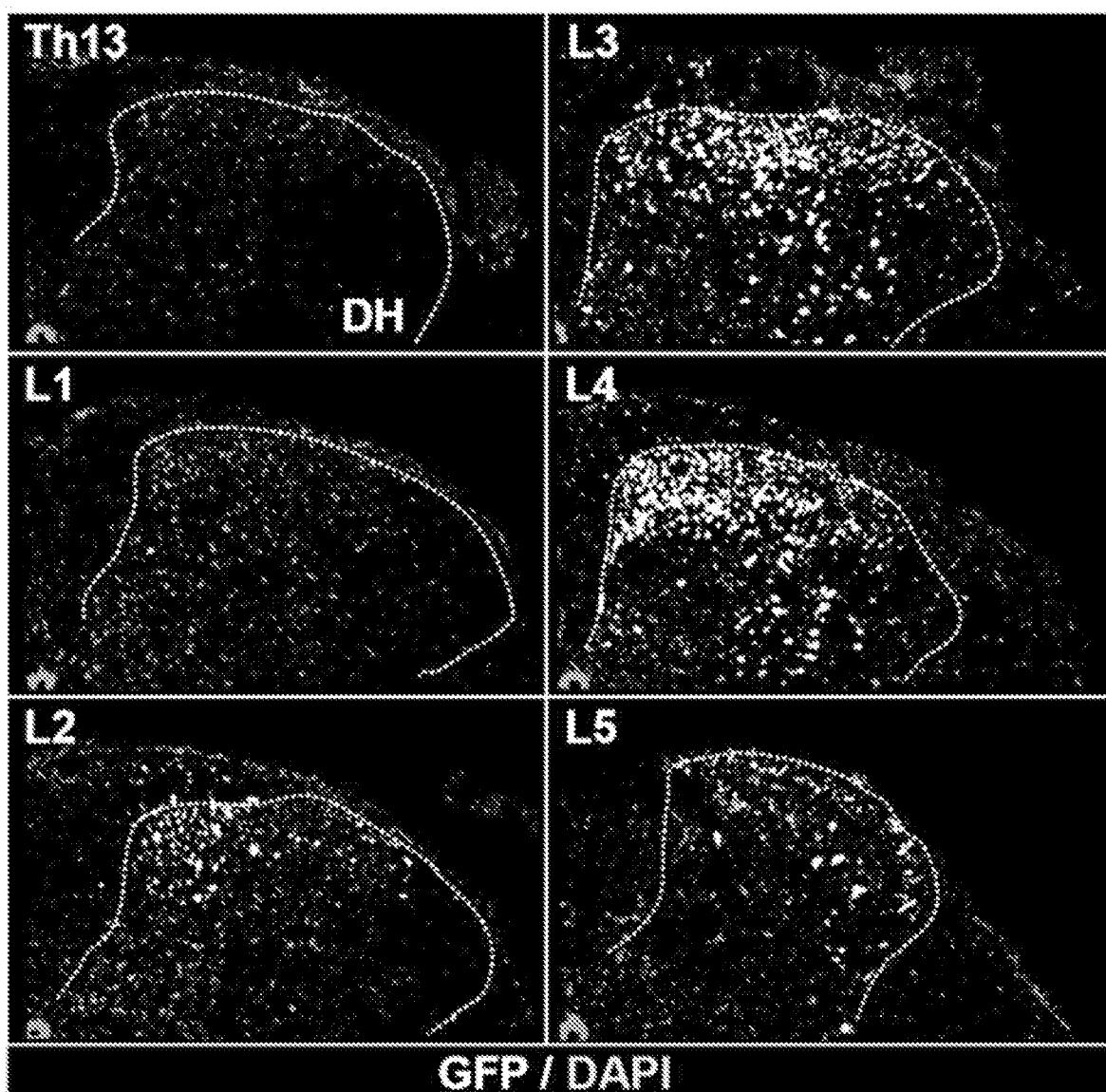


FIG. 1C

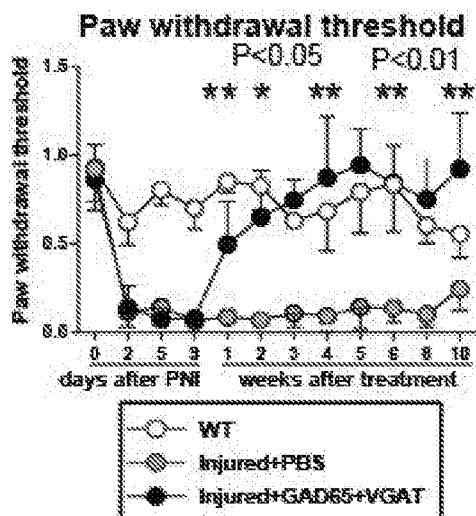


FIG. 2A

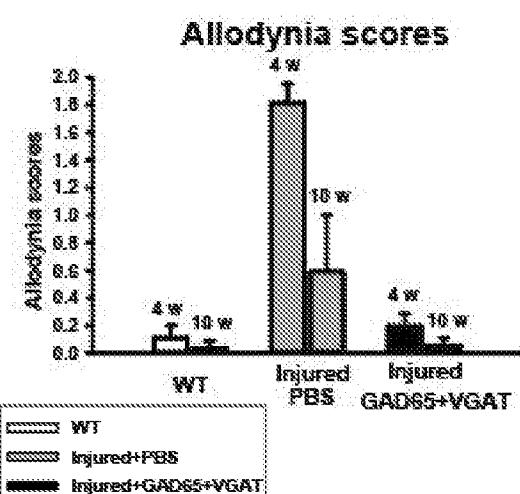


FIG. 2B

3/14

Open Field Motor Performance

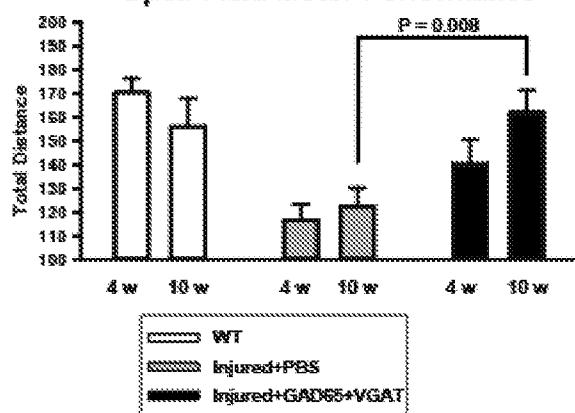


FIG. 2C

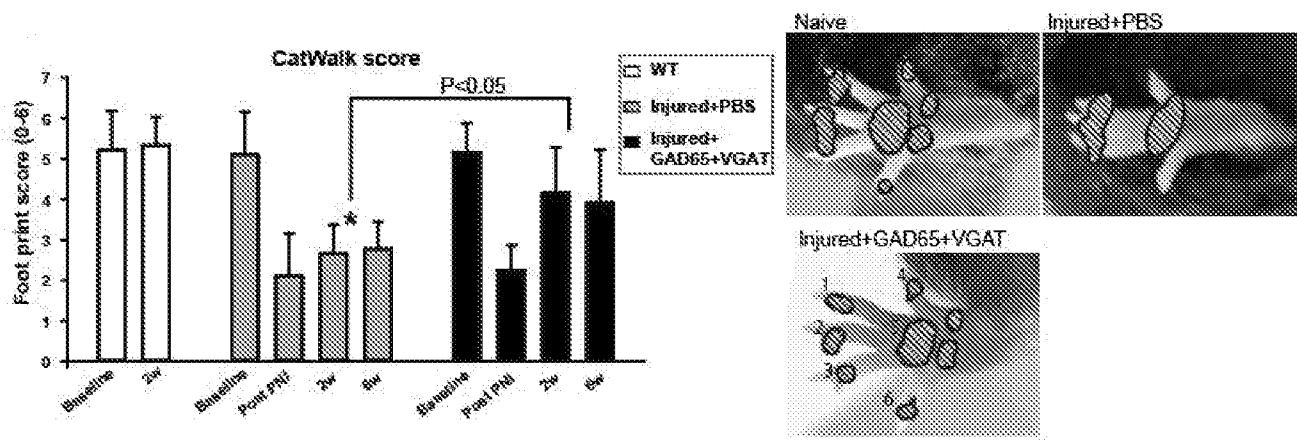


FIG. 2D

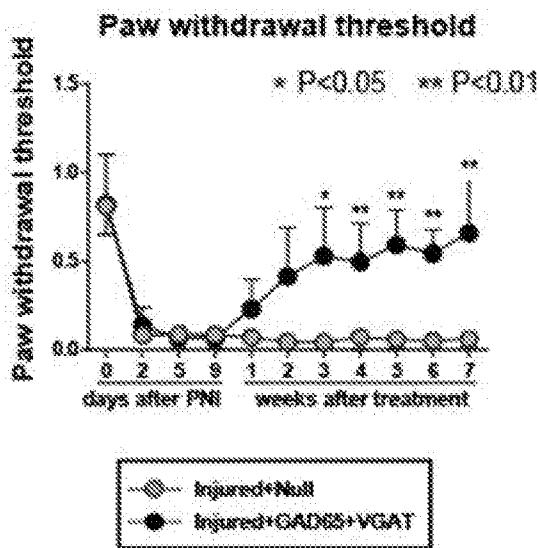


FIG. 2E

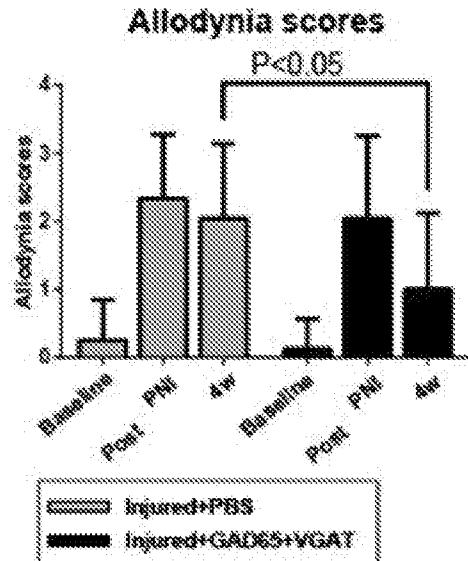


FIG. 2F

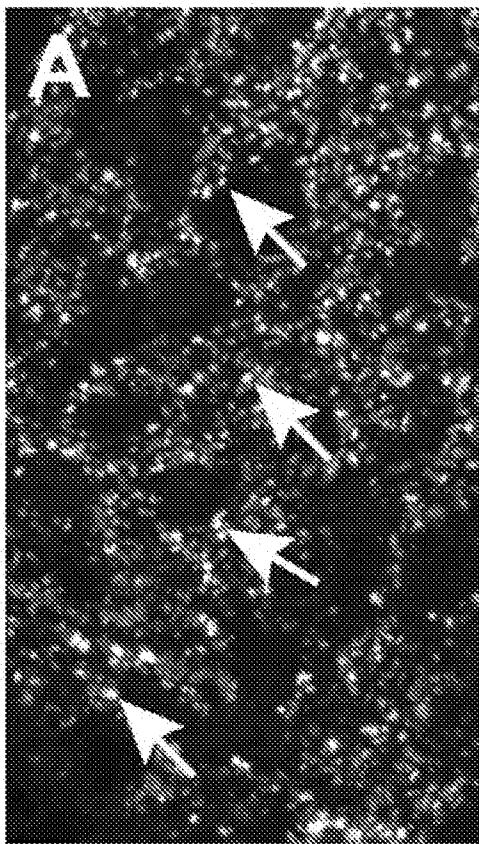


FIG. 3A

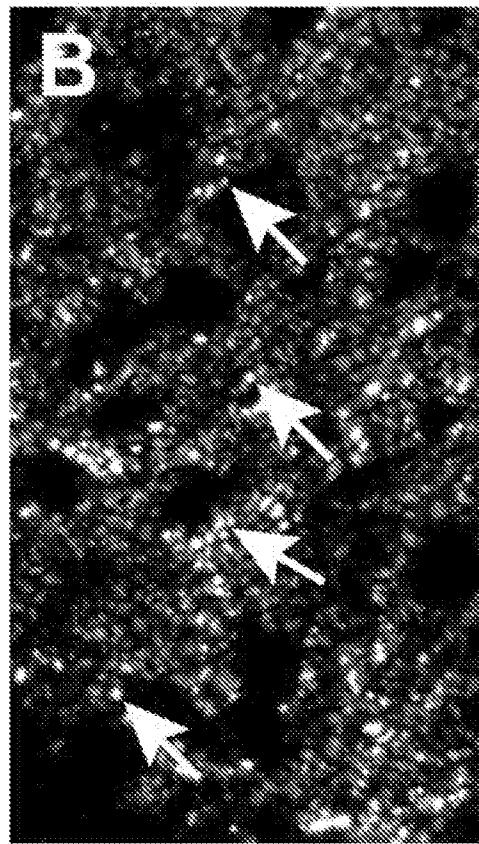


FIG. 3B

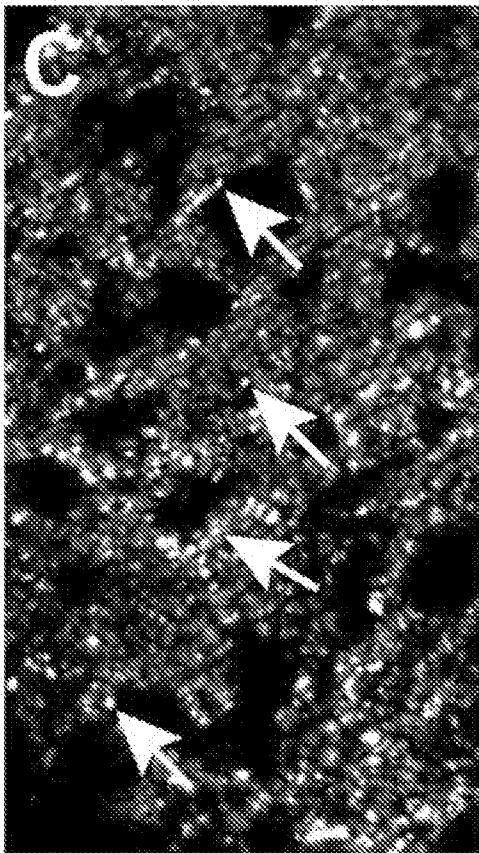


FIG. 3C

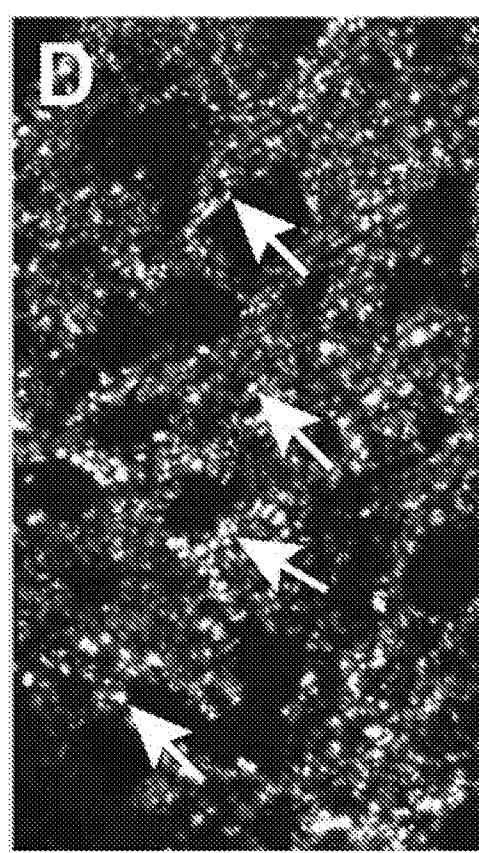


FIG. 3D

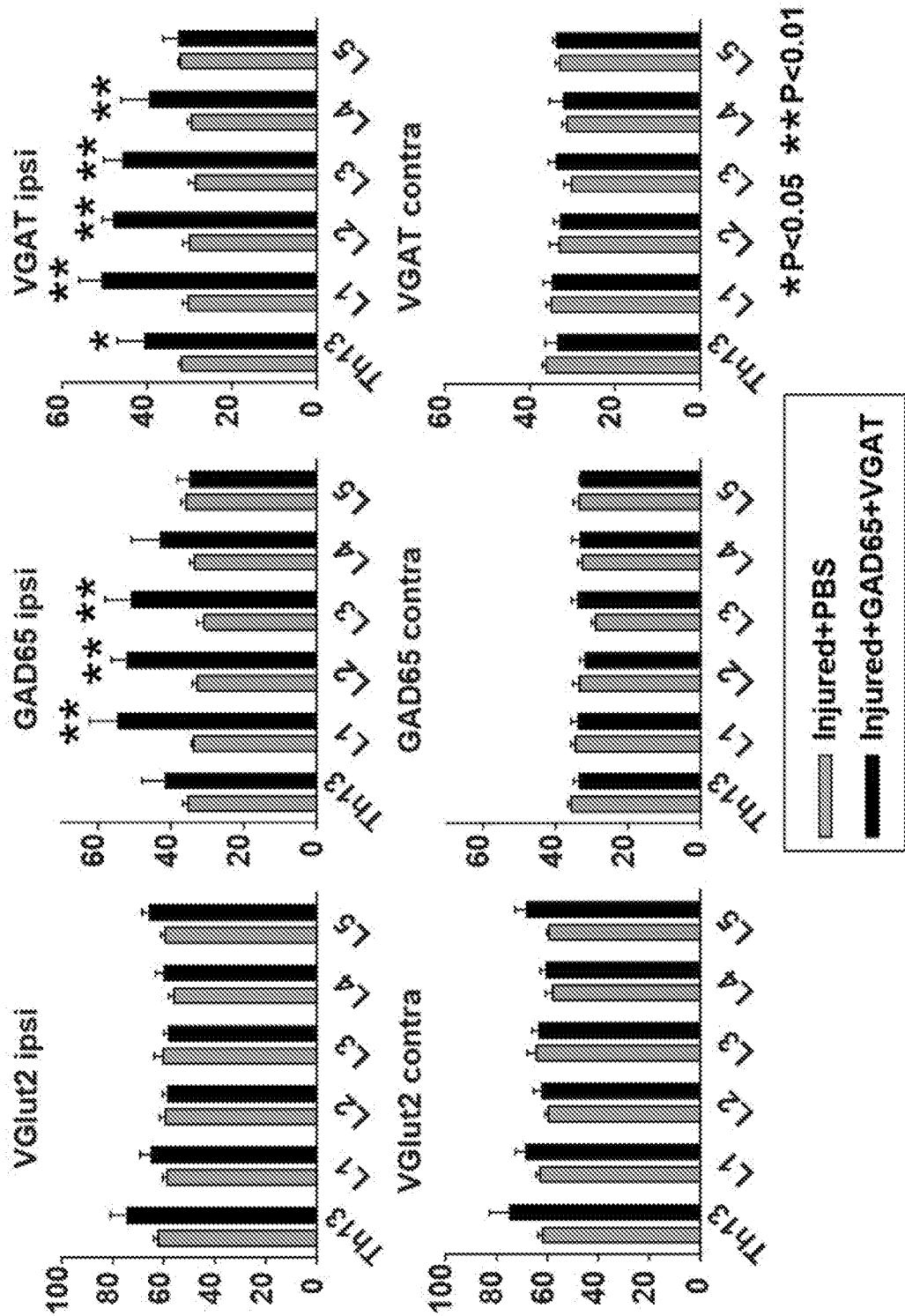
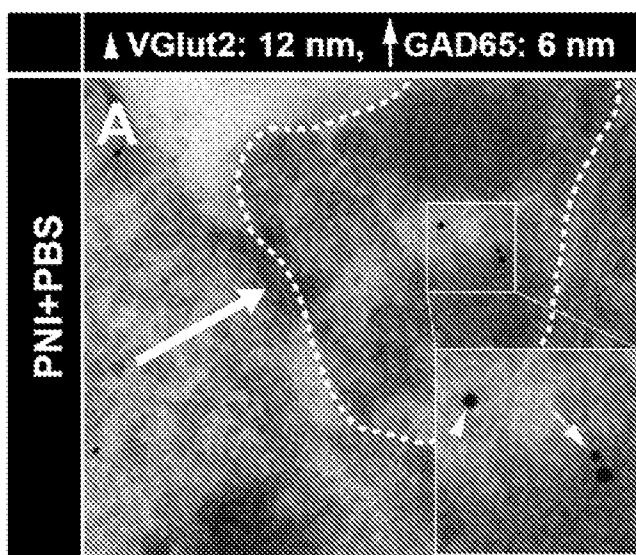
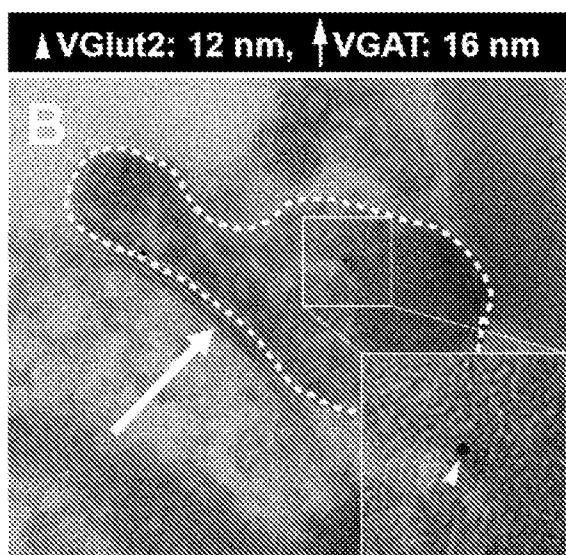
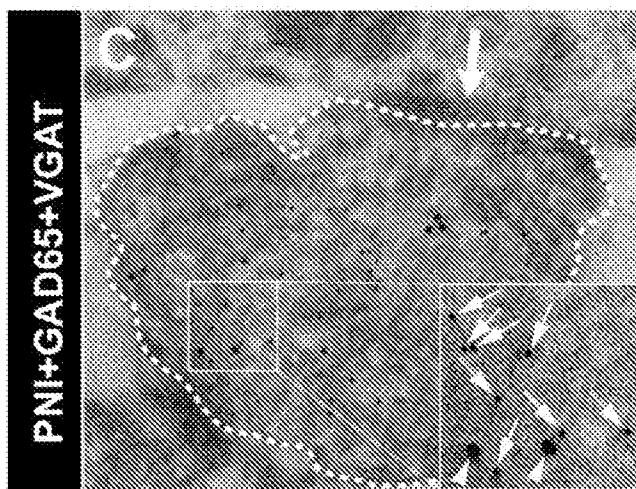
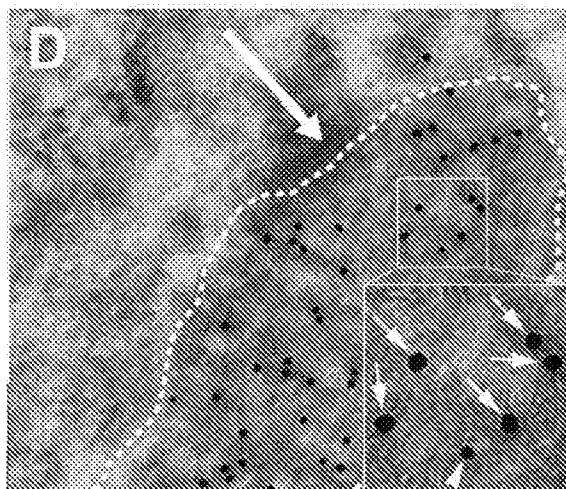
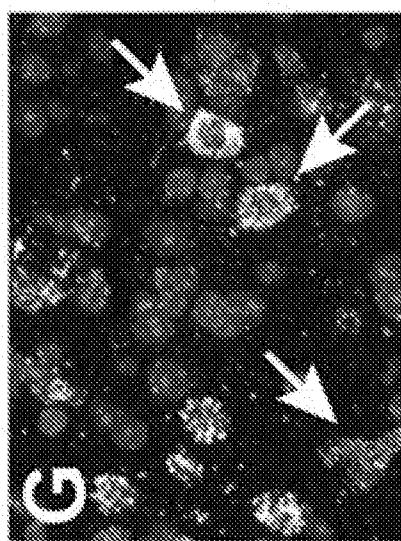


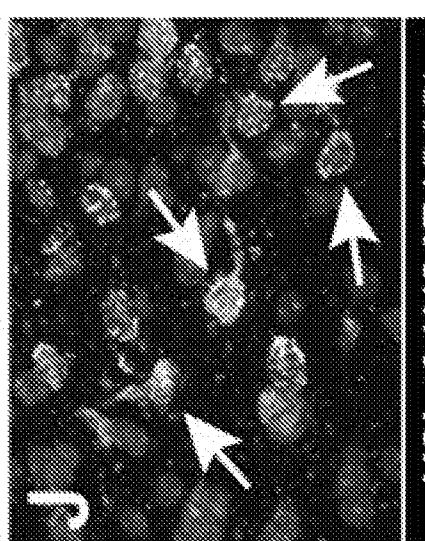
FIG. 3E

**FIG. 4A****FIG. 4B****FIG. 4C****FIG. 4D**



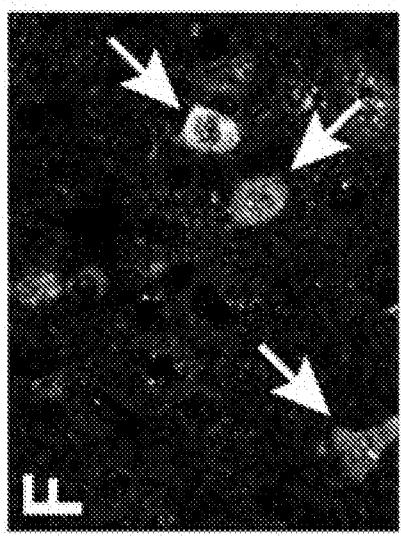
vGlut2 / GAD65 / DAPI

FIG. 4G



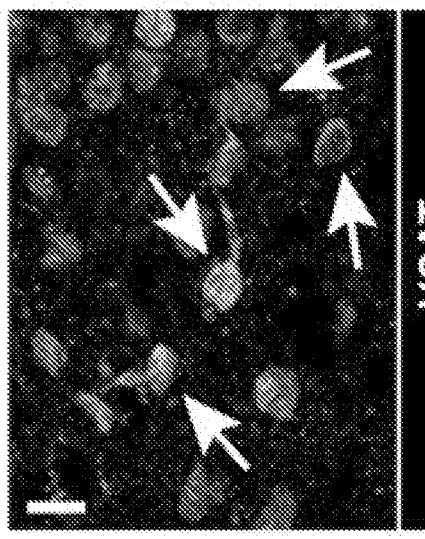
vGlut2 / VGAT / DAPI

FIG. 4J



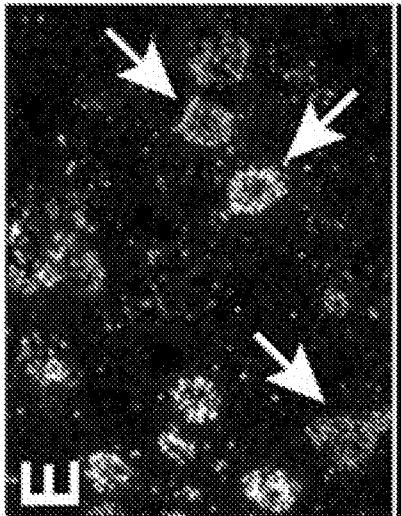
GAD65

FIG. 4F



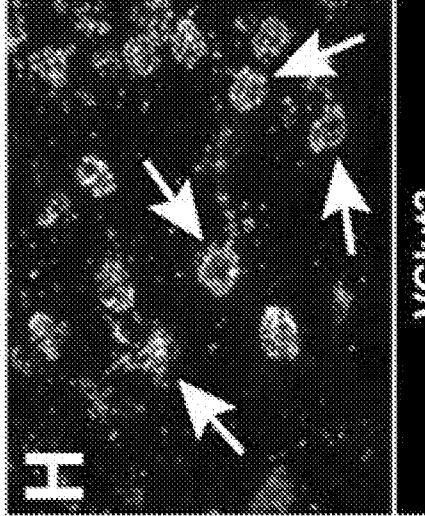
VGAT

FIG. 4I



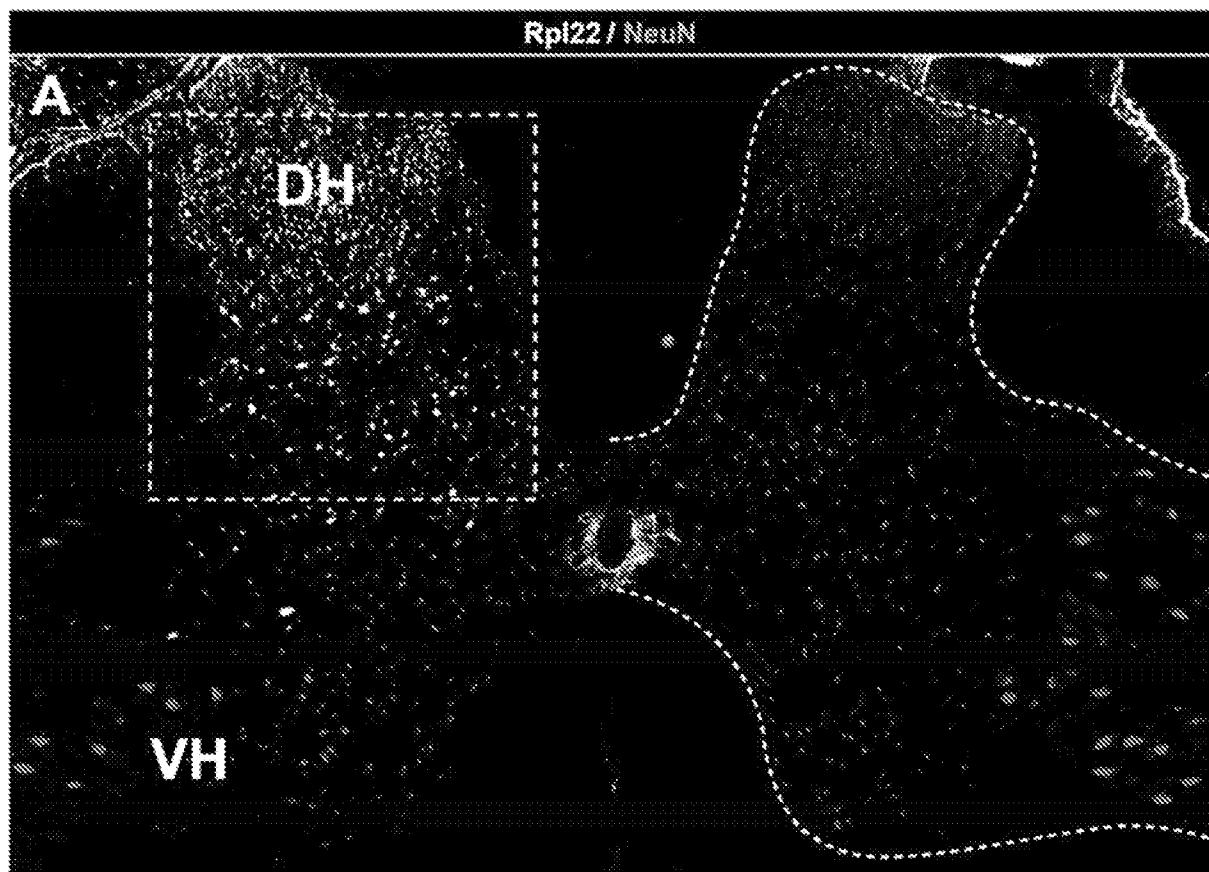
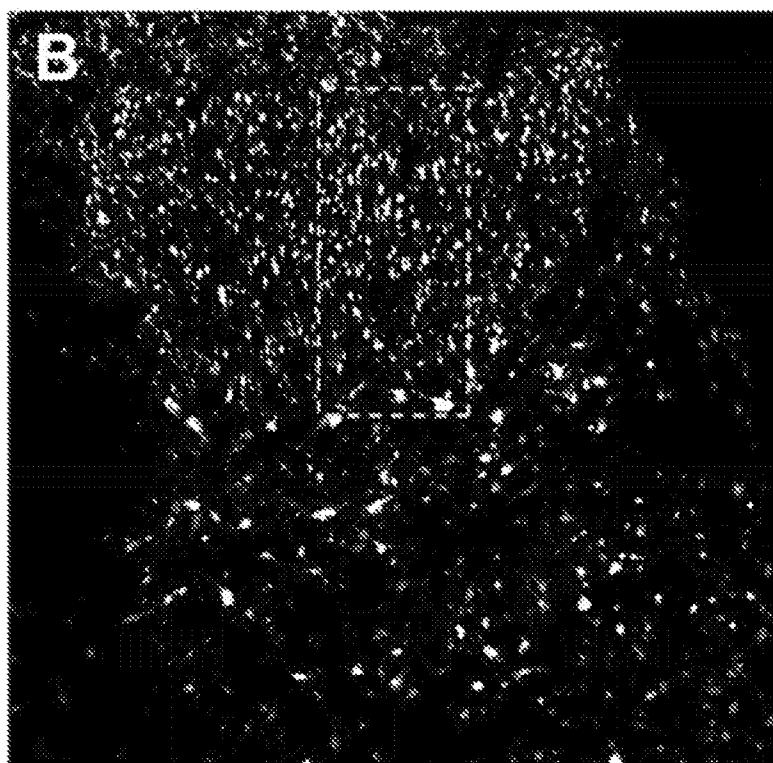
vGlut2

FIG. 4E



vGlut2

FIG. 4H

**FIG. 5A****FIG. 5B**

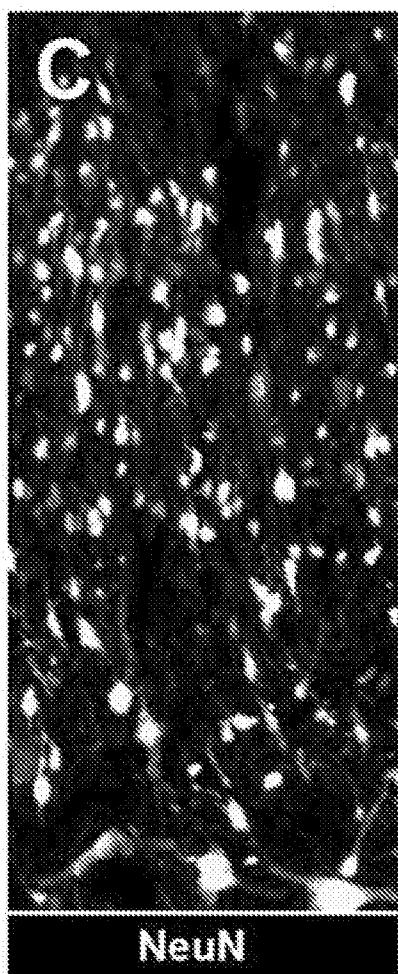


FIG. 5C

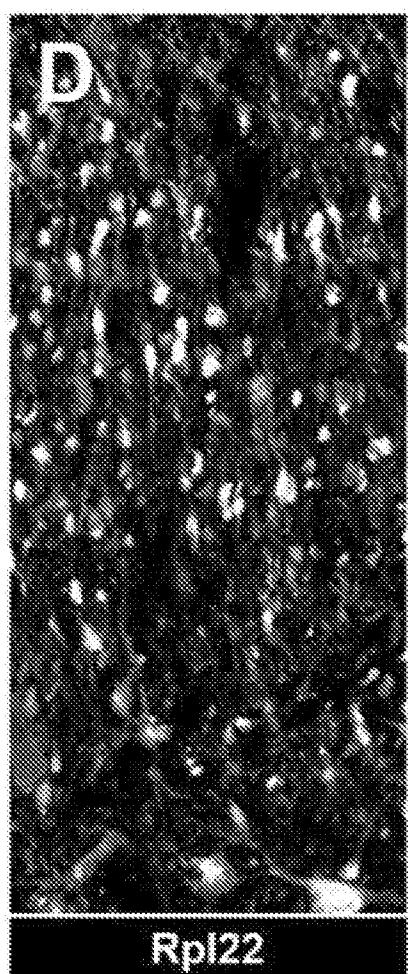


FIG. 5D

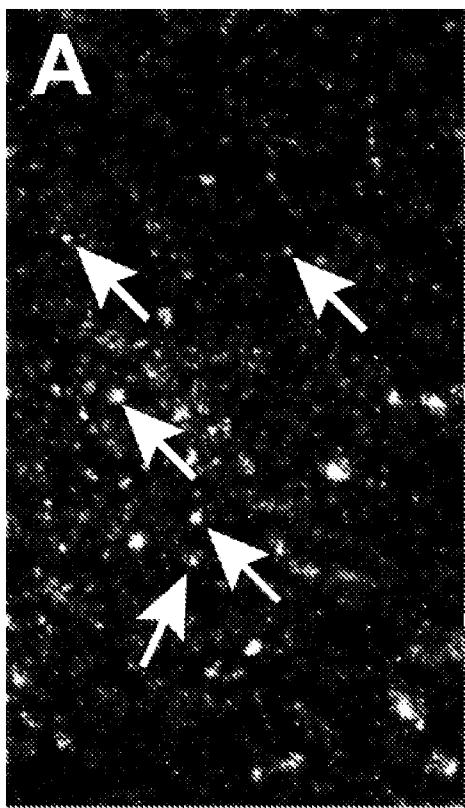


FIG. 6A

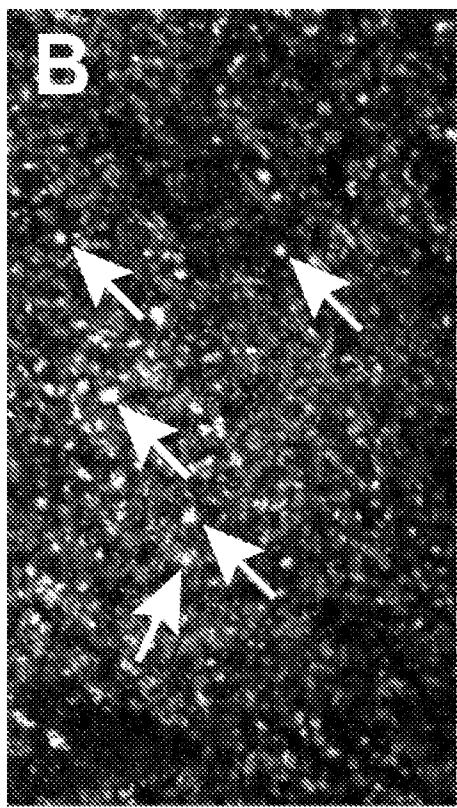


FIG. 6B

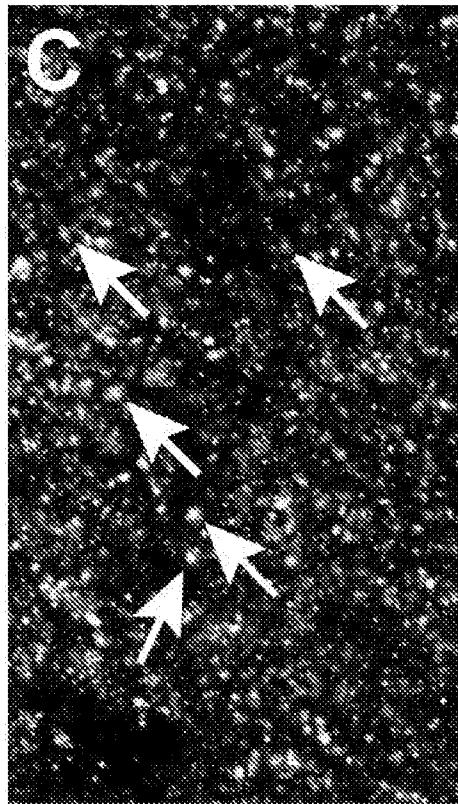


FIG. 6C

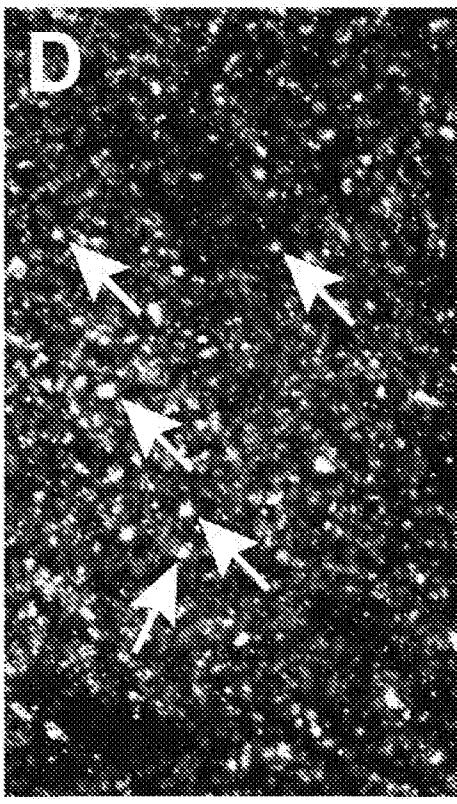
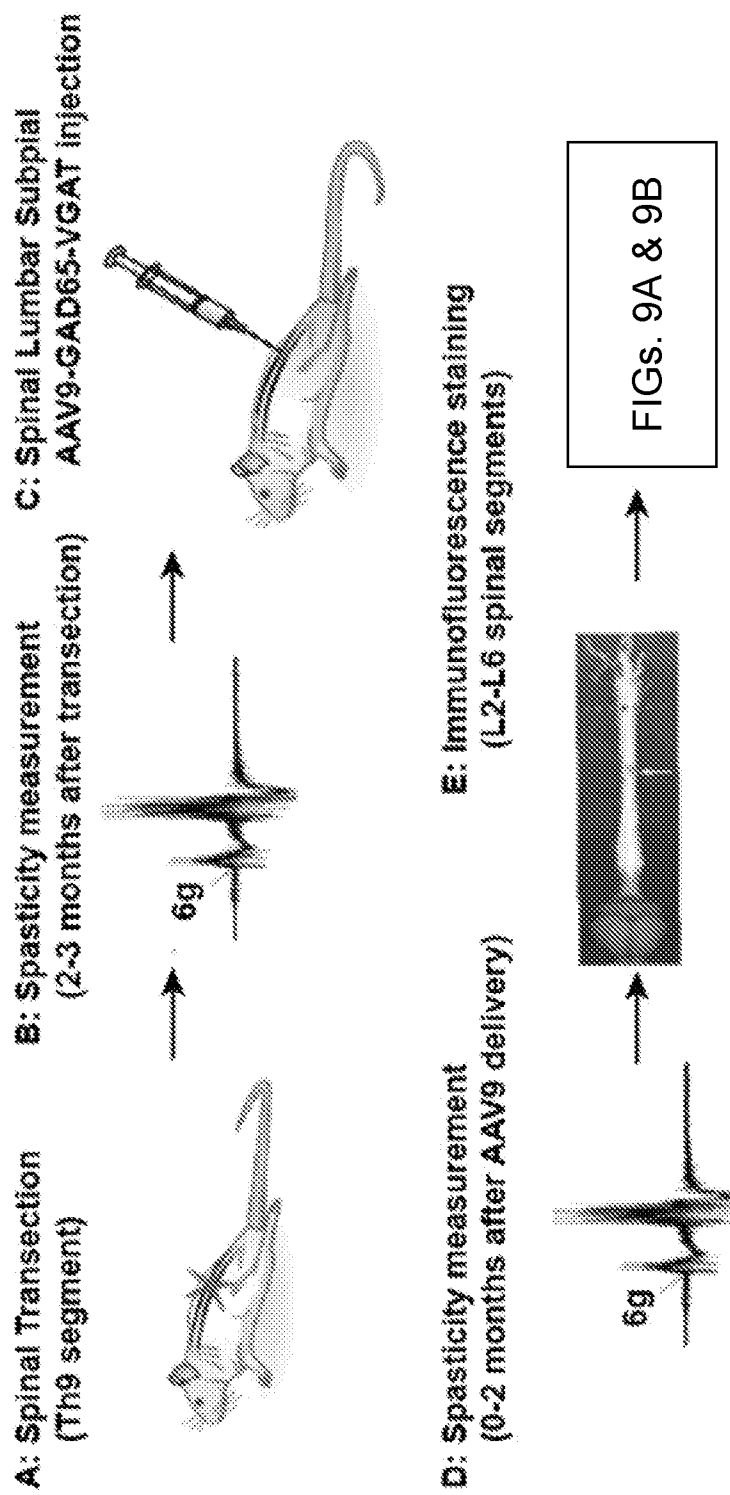


FIG. 6D

**FIG. 7**

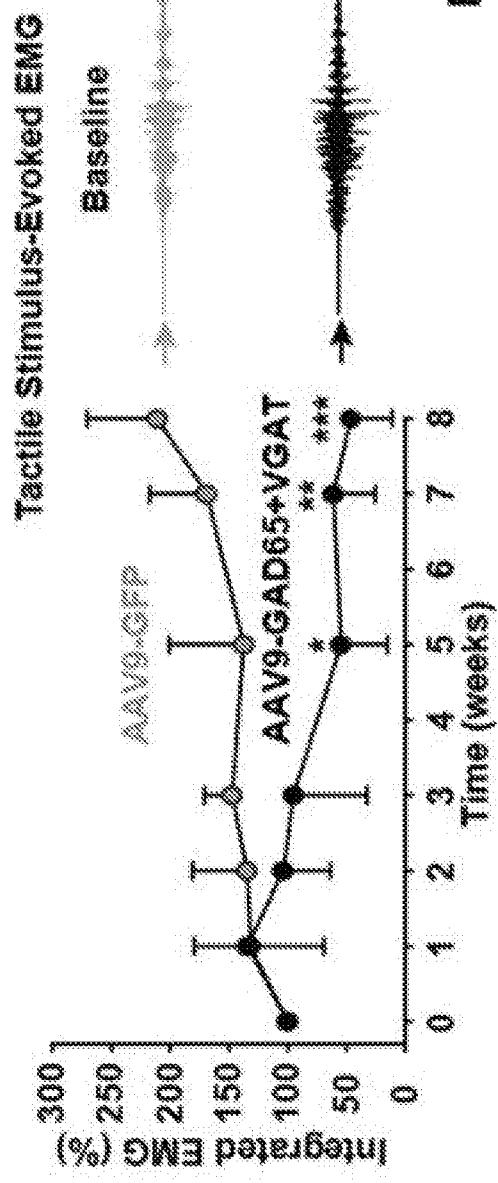
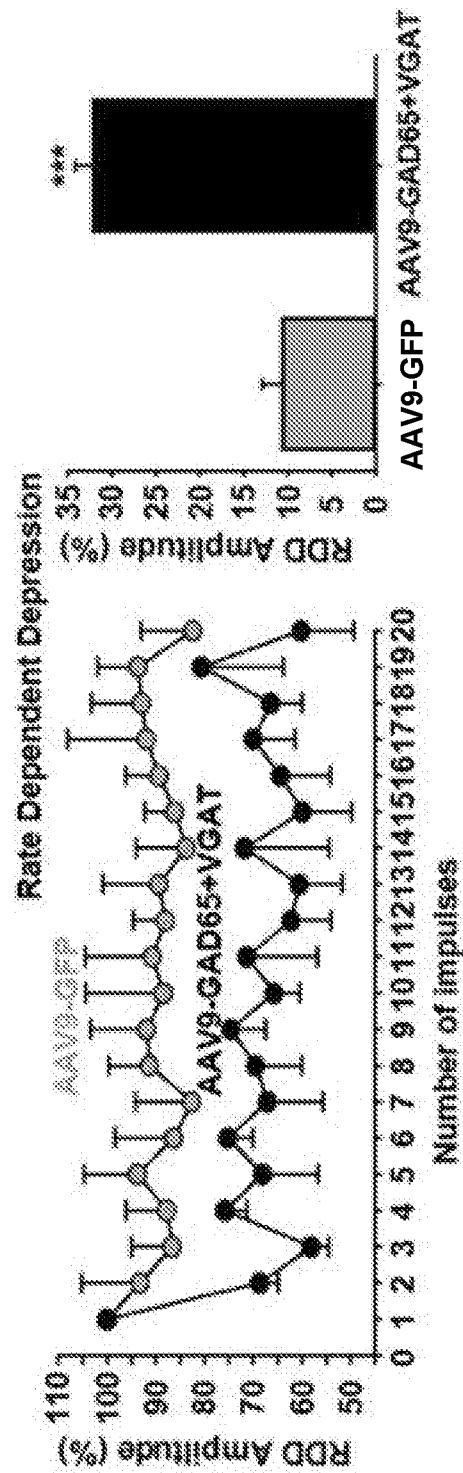
**FIG. 8B**

FIG. 9A

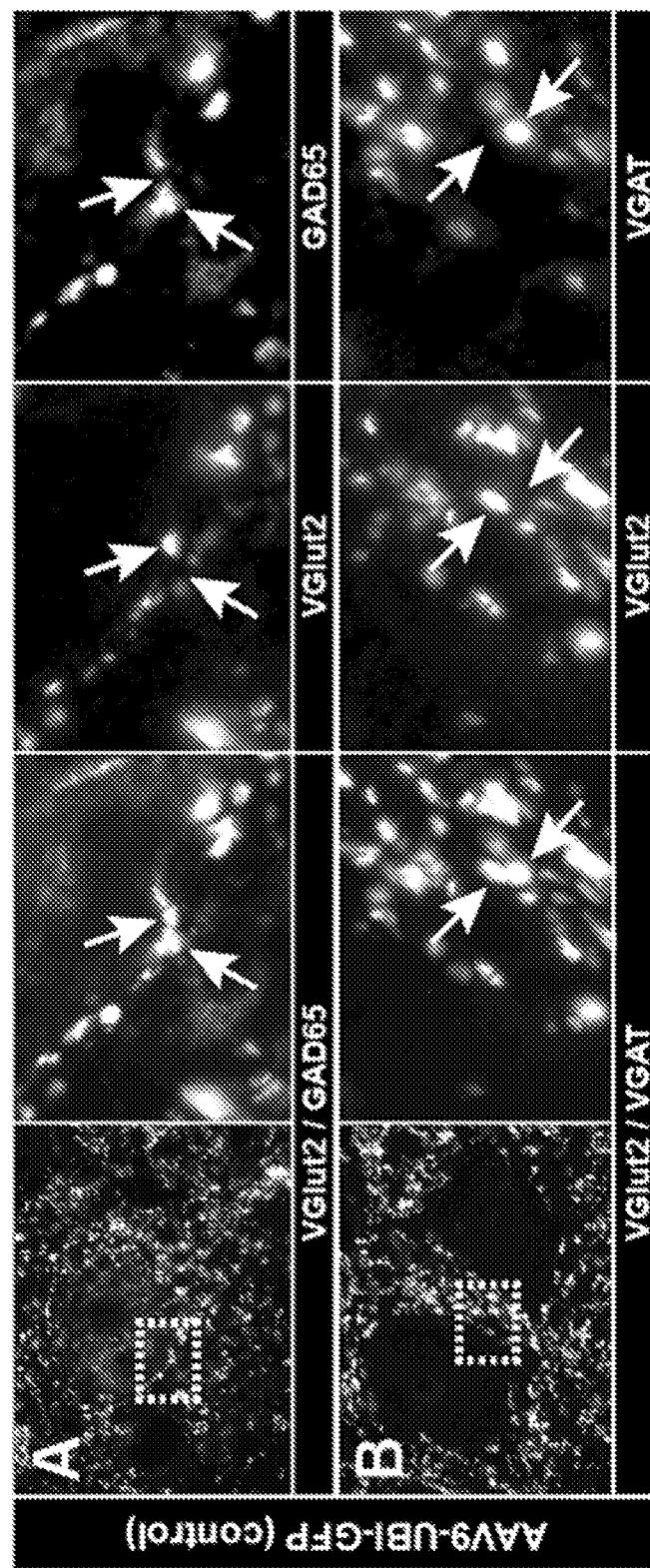
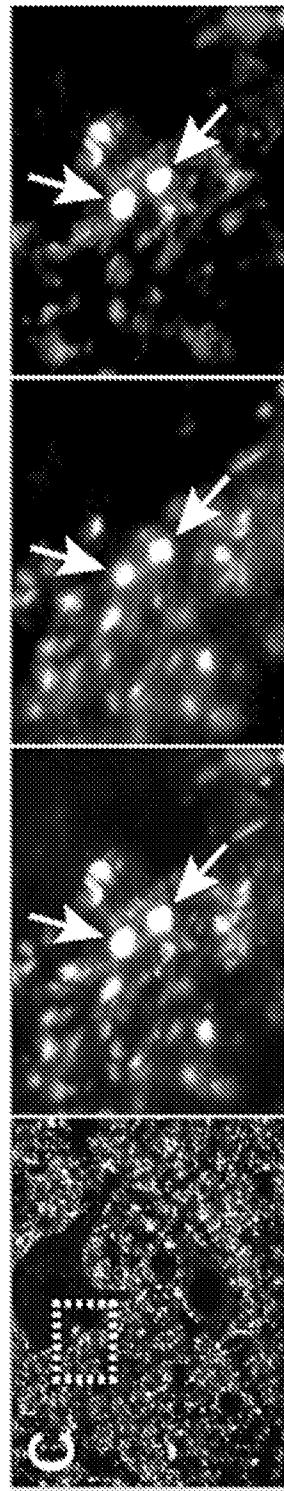


FIG. 9B

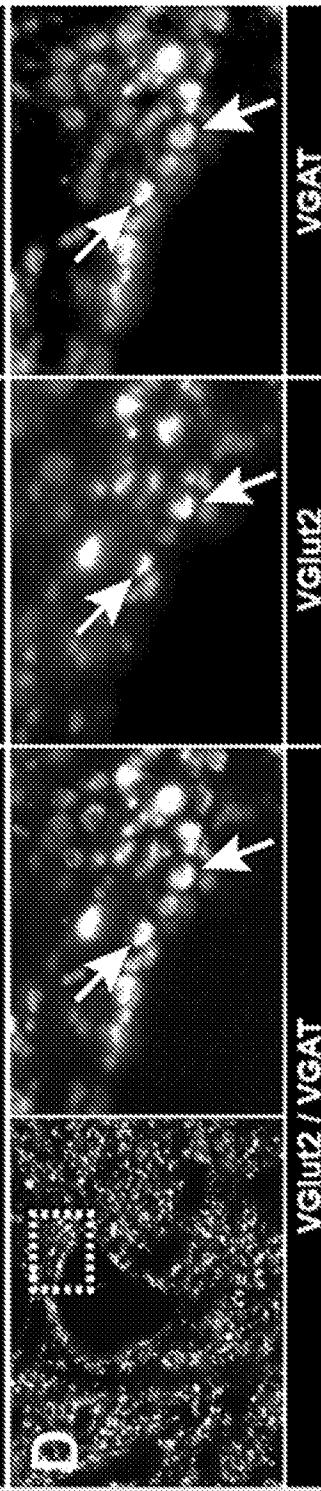
FIG. 9C



VGlut2 / GAD65

AAV9-UBE1-GAD654+VGAT (treated)

FIG. 9D



VGlut2 / VGAT

D

C

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US 18/49914

A. CLASSIFICATION OF SUBJECT MATTER
 IPC(8) - A61K 48/00, C07K 14/435, C12N 9/88 (2018.01)
 CPC - A61K 48/005, C12Y 401/01015, C12N 2750/14111, C12N 2750/14143

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

See Search History Document

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

See Search History Document

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

See Search History Document

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	US 2014/0256801 A1 (UNIVERSITY OF PITTSBURGH - OF THE COMMONWEALTH SYSTEM OF HIGHER EDUCATION et al.) 11 September 2011 (11.09.2011) para [0006], [0029], [0043], [0045], [0050], [0054-0056]. Claims 13-40, 43	1-28
Y	US 2003/0069398 A1 (RIPPMANN et al.) 10 April 2003 (10.04.2003) para [0004], [0013], [0050]	1-28
Y	WO 2016/122791 A1 (THE REGENTS OF THE UNIVERSITY OF CALIFORNIA) 4 August 2016 (04.08.2016) para [0029], [0042], [0052], [0060-0061], [0076], [0087-0088]	1-28
Y	WO 2008/112177 A2 (GENIZON BIOSCIENCES Inc.) 18 September 2008 (18.09.2008) (Claim 24, SEQ ID NO: 1677)	10, 15, 24
Y	X69936, GenBank Accession No: X69936, H.sapiens mRNA for glutamate decarboxylase, 16 March 1993 [online]. [Retrieved on 9 November 2018]. Retrieved from the Internet <URL: https://www.ncbi.nlm.nih.gov/nuccore/X69936> Entire document	9, 15, 24

 Further documents are listed in the continuation of Box C. See patent family annex.

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- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed
- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
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- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
- "&" document member of the same patent family

Date of the actual completion of the international search

9 November 2018

Date of mailing of the international search report

21 NOV 2018

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