

#### (12) United States Patent

#### Berinstein et al.

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(54) TUMOR ANTIGENS BFA4 AND BCY1 FOR

PREVENTION AND / OR TREATMENT OF **CANCER** 

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claimer.

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- (60) Provisional application No. 60/445,342, filed on Feb. 6, 2003, provisional application No. 60/411,833, filed on Sep. 18, 2002, provisional application No. 60/394,503, filed on Jul. 9, 2002, provisional application No. 60/394,346, filed on Jul. 3, 2002.

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	A61K 48/00	(2006.01)
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CPC ...... C07K 14/4748 (2013.01); A61K 39/00 (2013.01); A61K 2039/53 (2013.01); C12N 2799/021 (2013.01); C12N 2799/023 (2013.01) USPC ...... 514/44 R; 435/320.1; 536/23.5

Field of Classification Search

None

See application file for complete search history.

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#### (57)**ABSTRACT**

The present invention relates to a nucleic acid encoding a polypeptide and the use of the nucleic acid or polypeptide in preventing and/or treating cancer. In particular, the invention relates to improved vectors for the insertion and expression of foreign genes encoding tumor antigens for use in immunotherapeutic treatment of cancer.

#### 24 Claims, 6 Drawing Sheets

<sup>\*</sup> cited by examiner

FIGURE

NT GSTCCGGAAAAAGAACCCCCCTCTGAGAAACGTTGCAAGTGAAGGCGAGGGCCAGATCCTGGAGGCCTATAGGTACAAAAGAAGGTATCTGGAAAGAACAAAGAATTCTCTGCAGA | ICAGATGTCAGAAAATACGGATCAGAGTGATGCTGCAGAACTAAATCATAAAGGAGAACCATAGCTTGCATGTTCAAGATCCATCTTCTAGCAGTAAGAAGGACTTGAAAAGCGCAGTTC 

**GGCTACAGAGGAAACAGGGCAAGCCAGAGTGGTCAAGGCCAATTGTCAAGGTTTGAGCCCCAGTTTCAGTGGCCTCAAAAAACCCACAAGTGCCTTCAGATGGGGGGTGTAAAGACTGAATA AATCCAAAACTGACTTACTGGTGAATGACAACCCAGACCCGGCACCTCTGTCTCCAGAGCTTCAGGACTTTAAATGCAATATCTGTGGATATGGTTACTACGGCAACGACCCCACAGAT**  **GCATTGGACGGAAAACACCAGATTGCCAAGGGAACACCAAGTATTTCCGCTGTAAATTCTGCAATTTCACTTATATGGGCAACTCATCCGAATTAGAACAACATTTTCTTCAGC** CACCCAAACAAATAAAAGCTTCTCTCCCCTCCTCTGAGGTTGCAAAACCTTCAGAGAAAAACTCTAACAAGTCCCTGCACTCAATCCAGTGATTTCTGGAGACTTTGGGAAAATG

CTGATTAAGCACTTCCGAAAGTATCACTTAGGACTGCATAACCGGCACCAGGGCAAGATGCTGGACAGGAAAAATCTTGGCCCTTCATAACATGGTGCAGTTCAGCCATTCCAAAGA

CACTGCCAGGAACAGGACATCACTACAGCCAACGGCGAAGGAGGGACGGTCATGCCATATCCACCATAAAGAGGAGCCCAAAATTGACTTCAGGGTCTACAATCTGCTAACTTCAGAACTC GGAGAGGGGCAGACATCCTGCGGGGGAGTCCGTCATACACCCAAGCCTGGTGGGGCTGCTGACGCCTGTGTCTGGCACCCAAGAGAGAAAAAAACACTCTAAGGGATAGTCCCAATGTG **AITTTTGTAGTTTCAGCTGTGAGTCATCTAGCTCACTTAAACTGCTAGAACATTATGGCAAGCAGCACGGAGCAGTGCAGTCAGGCGGCCTTAATCCAGAGTTAAATGATAAGCTTTCC** TTCACAAGTGTACCATTAAACACTGTCCCATTCTGTCCCAGAGGACTTTGCAGCCCAGAAAAGCACCTTGGAGAAATTACTTATCCGTTTGCTTGTAGAAAAGTAATTGTTCCCACTGT **GCACTCTTGCTTCTGCACTTGTCTCCTGGGGGGGGGGGCGCTGGAAGCTCCGAGTCAAACATCAGTGCCATCAGTGTTCATCACCACCCCTGACGTAGAIGTACTCCTCTTTCACTATGAAAG** GAGGCCGCCCATCTGGCGCGACCTATTTATGGCTTTGGCTGTGGAAACCAAGGGATTCCTGCAGGGGGCGCCAGCTGGCGGAGAGAAGTCTGGGGGCCCTCCCCCAGCAGTATCCTGCATC atgagacctgcgaaacacccaaattattcaccaccaggcagccctattgaaaagtaccagctacccactttttggacttacataatgacttccagagtgaagctgattggc CTCATTTTTCAGCTGTTGGATCAAACAATGACATTCCTCTTAGATTTGGCGATCAAGCATTCCAGACCTGGGCCAACTGCAAACGCGTGCCTCCAAGGAGAAACGAAGGCACCACCAAAT GCAGGACAAGATAACAGTCAAAGCAGGAGATGACACTCCTGTTGGGTACTCAGTGCCCCTATAAAGCCCCTCGATTCCTCTAGACAAAATGGTACAGAGGCCACCAGTTACTACTGGTGTA **GGATAATATGGTAACGAGCTATAATTGTCAGTTCTGTGACTTCCGATATTCCAAAAGCCATGGCCCTGATGTAATTGTAGTGGGGCCACTTCTCCCGTCATTATCAACAGCTCCATAACA** CTTAACCCAGAGGCACTTCAGGCTGAGCTCAACAAACAGCAGAGAGGGCAGAATGAGGAGGAAGTCAATGGAAGCCCGTTAGAGAGGAGGTCAGAAGATCATCTAACTGAAAGTCA GGGGTTCTGGAGTAAATATAAGCTCTCCGGTTCCTGGGAATCCGCACTACTTGAGTCACGTGCCTGGCCTACCAAATCCTTGCCAAAACTATGTGCCTTATCCCACCTTCAATCTGC GTAAAAATGAGGTCCCTTGAATGTAGTAAAAACAGAGAAAGTTGATAGAAGTACTCAAGATGATGAACTTTCAACAAATGTGTGGCACTGTGGCATTGTCTTGGATGAAGTGATGA TGCTTTGCATATGAGTTGCCATGGTGACAGTGGACCTTTCCAGTGCAGCATATGCCAGCATCTTTGCACGGACAAATATGACTTCACAACACACATATCCAGAGGGGCCTGCATAGGAACA ATGCACAAGTGGAAAAAATGGAAAACCTAAAGAGTAA\*

## FIGURE

GSGVFCANCLTTKTSLWRKNANGGYVCNACGLYQKLHSTPRPLNIIKQNNGEQIIRRRTRKRLNPEALQAEQLNKQQRGSNEEQVNGSP EKYMRPAKHPNYSPPGSPIEKYQYPLFGLPFVHNDFQSEADWLRFWSKYKLSVPGNPHYLSHVPGLPNPCONYVPYPTFNLPPHFSAVG WVRKKNPPLRNVASEGEGOILEPIGTESKVSGKNKEFSADOMSENTDOSDAAELNHKEEHSLHVQDPSSSSKKDLKSAVLSEKAGFNYE SPSKGGNFPSFPHDEVTDRNMLAFSFPAAGGVCEPLKSPQRAEADDPQDMACTPSGDSLETKEDQKMSPKATEETGQAQSGQANCQGLS HNMVQFSHSKDFQKVNRSVFSGVLQDINSSRPVLLNGTYDVQVTSGGTFIGIGRKTPDCQGNTKYFRCKFCNFTYMGNSSTELEQHFLQ THPNKIKASLPSSEVAKPSEKNSNKSIPALQSSDSGDLGKWQDKITVKAGDDTPVGYSVPIKPLDSSRQNGTEATSYYWCKFCSFSCES SSSLKLLEHYGKQHGAVQSGGLNPELNDKLSRGSVINQNDLAKSSEGETMTKTDKSSSGAKKKDFSSKGAEDNMVTSYNCQFCDFRYSK SHGPDVIVVGPLLRHYQQLHNIHKCTIKHCPFCPRGLCSPEKHLGEITYPFACRKSNCSHCALLLLHLSPGAAGSSRVKHQCHQCSFTT PDVDVLLFHYESVHESQASDVKQEANHLQGSDGQQSVKESKEHSCTKCDFITQVEEEISRHYRRAHSCYKCRQCSFTAADTQSLLEHFN TVHCQEQDITTANGEEDGHAISTIKEEPKIDFRVYNLLTPDSKMGEPVSESVVKREKLEEKDGLKEKVWTESSSDDLRNVTWRGADILR SDNDIPLDLAIKHSRPGPTANGASKEKTKAPPNVKNEGPLNVVKTEKVDRSTQDELSTKCVHCGIVFLDEVMYALHMSCHGDSGPFQCS PVSVASKNPQVPSDGGVRLNKSKTDLLVNDNPDPAPLSPELQDFKCNICGYGYYGNDPTDLIKHFRKYHLGLHNRTRQDAELDSKILAL 3SPSYTQASLGLLTPVSGTQEQTKTLRDSPNVEAAHLARPIYGLAVETKGFLQGAPAGGEKSGALPQQYPASGENKSKDESQSLLRRRR LERRSEDHLTESHQREIPLPSLSKYEAQGSLTKSHSAQQPVLVSQTLDIHKRMQPLHIQIKSPQESTGDPGNSSSVSEGKGSSERGSPI ICQHLCTDKYDFTTHIQRGLHRNNAQVEKNGKPKE

COCTACCOSTORIO O OCITACITACITACITACITACIDA DA GOGO CALO COLICA COCATACO A COLO DA COLO TALA TALO CALO DA GOCOTIDA COCATACACACA

Consensus

#### 946 CTTON GOOCLE AND CONTROL OF THE CONTROL OF CONTROL CONTROL CONTROL OF THE TOO TON CONTROL CONT CTTGAGGGCCAAGACCAACACGTACAAGAACCGGTGAGGGGGGAGGAACCAGTGTTCATGGTGAAGGGAGGAGGAGGAGGAGGAGGACGTGGCCACAGCCGGGCAATGA TOTCAGCAGGGAGCAGTTGTGCATGATGCGTGCCTGCGGGAAGAAGAAGAGGGGGGGCGTTTGGTGTGGGGTCGTGTTTTGCGGGGCGAGTGACCATGGGGGGG TOTCAGCAGAGCACTTCTCCATGATTCCGT9CCTCCGGAACAAGTCAGGCCCCTTTGGTGTGGGCTCCTGCTCTGCCGGCCAGGTGACCATGACGGGCG TOTCAGGAGGAGTACTTCCCATGATCCGTAGCTCCCGAAGAAGAAGAGGCGCCTTTGGGGTCCTGCTCTGTCTCTGCCCGGCCAGGTGACCATGACGTGGGGGG TOTCAGGAGGAGGAGTTCTCCATGATCGGGGCACCAGGAAGAAGAAGAGGCGCGCCTTTGGTGTGGGCTCTGCCGGGCAGGTGACCAGGTGACCAGGTG TOTA A CAROLACTION CATARATION SPOOT COMPANIA ATTA CONTROLATION OF THE STOT CONTROL CONTROL CARACTER CONTROL TO SERVINE TO THE CONTROL OF THE TOTCAGCAGCAGCACTTOTCCATTATCCFFCCTFCCCACAACAAGTCAGGCGCTTTTGGTGFGGCTCTTGCCCGGGCTGCTGGCCCAGGTGACCATCATCAGTGGCGGG CITERESCOCTABERCORRO COTRORIORACROCOSTOROCOSTOROCOCOCORRO PORTICATION COCOROSOCOS COCOROCOCOCOCOCOCOCOS RATION TOTCAGGAGGAGTAGTTCTCCATGATTCCGTGCCTCCCGGAAGAAGTTCAGGGCCGCCTTTGGAGGGTCTGGCTCTTGCCCGGCCAGGTGACCATGACGAGGGGG TOTCASCADGGAGCACTTOTOCATGA FOS TACCATOGGGAACAAGAAGAAGAGGGGGTTTGGAGTTCTGGTTCTGCTCAGCTGAGTGACGATGACGAGGTGAGGGGG OCCUPACIONING GEORGIA STREET GOOD BAAGGOOD TO A COCOMICIA CON TO A CONTRA CONTRA CATA TO CALARIA CONTRA COCONTRA COCONTR OCCTACCENTEGEGONGSTGSTGCCCCCCAAAGGGCCAACCATCAAGGGCATCCAGGAGCAAACCAATATAGATIATCATIATGACGAGGGGGACGCGT OCTACOSOFICITOS INSTINSTINICI OCOMBASSICA TOPAS COSTATORIO BARCO BORA POR TATA TOROA CARIO BOSTINA COSTU A TGGCCSACTIGNOST TARBOGCGGGGTACLACOLACOLACOLAGGAGTGTTTCCCGTGCCCACCTTCCCACCTGGCCGAGATCGTGGGCAAGGCAAGGGTTACAAGTTAAGGG ATGECCERCTRACECTRANSGOGGETA CENNOCACGAGTETTTCCCTACCTACTACTACTCCGETGCCTTGCCGAGATCGTGCCGAGATCGTCGGCAAGCCAAGGCTACAAGGCTAAGGCTAAGGCTAAGGCTAAGGCTAAGGCTAAGGCTAAGGCTAAGGCTAAGGCTAAGGCTAAGGCTAA ATGECCERCTRACTGCGCTTAAAGGGAAGTACAACAACAAGAAGTGTTTCCCSTGTCCCACCTCCGAGCACGTGGCGAGATCGTGGGCAAGGCAAGGCTACAAGATTAAGGC ATGECCEACTION DO CONTRATOR GOAGLACOAGCACOACOAN COACOATON TOTOCOATOCO CACOTOCOA GOACOA GATOCOTOGA GATOCOTOGA GOA ATGBCCBACTGCGCTGAAGGCAGCACAACAACAACAACAACATGTGTTTCCGTGCCACCTCCGAGCACGTGCCGAGATCGTGGGCAAGCTAAGGCTAGGATTAAGGC atgrocsagottranscontaaasconagnagnagnagnancsaccoaterratttootaacontocsaaacaacetssoccaabaateaacaassagotecaagattaag COCTACOSOSTIGITIS GATICATICA GOOCOCA PAGGACIA FOR TORACIO CORTICA RACERA PACORITIA CATILITA CATILITA CAL SACOSTIGA COGGACIO COCATI 88888888 (308) (328) (331) (II 8) (331) (322) (3.25) (331) (331) (331) (223) original BCY1 MCSOAl9 BCY I MCSOA6 BCY I original BCY1 MCSOA6 BCY I MCSOA6 BCY I original BCY1 MCSOB19 BCY I Consensus Consensus Consensus MCSOAls BCY MC54.21 BCY MC55.29 BCX MC55.32 BCX MCSONE BCX MCSOAS BCY MC54.21 BCK MC55.29 3 MC55.32 3 MC5036 MC50A8 MC55,29 MC55,32 MC50A8 MC54.21 MC55.29 MC55.32 MC54.21

FIGURE 3B

1 (641)	(41) TITEBARATEROSCIOCOCARGEOROCOROCOROCOROCOROCOROCOROCOROCOROCORO
.29 BCY I (441) .32 BCY I (441) Consensus (441)	GTICGRGATCACGGGTGCC GTICGRGATCACGGGTGCC GTICGRGATCACGGGTGCC 551
######################################	(78) CORRESTRITATACACOTGOCGAGATIACOCOCOCTUTGEGGGGGCGRGAAGGCGAGGCCAACOTCGITGUTTTTTCCCTYROCITGUTTCCTTCCTTCCTTCCTTCCTTCCTTCCTTCCTTCCT

original BCY1	(878)	881 CTTCCCCAAGGCCCGCGTGGGCCCCGGGGGGGAACGCTCCCCTGCCACTTCCGCGGGAACCGGAGCTGGCCGGAGTGCCGAGGCGCCCCGGGGAAGCCGCTTCCGG
MC50A19 BCY I	(881)	CITCGCCAAGGCCGGGGTGGGCCCCCGGGGGGCACGCICCCCTGCCACTTCCGCGGGACCCGAGTGCCGGACTCCCGAGGCGCCCCCGGGGAGGCGCTCCAG
MC50A6 BCY I	(881)	CTTCCGCCAAGGCCCGCGCTGGGCCCCCGGGCGCACACCGCTCCCCTGCCACTTCCGCGGAACCCGAAGCTGGCCGGAATCCCGAAGGCCCCCCGGGAAGAGCCGCTCCAG
BCY	(881)	CTTCCGCCAAGGCCGGCGCTGGGCCCCCGGGGCGCACACCGCTCCCCTGCCACTTCCGCGGAACCCGAAGTCGGGGACTCCCGAGGCGCCCCCCGGGGAGAGCCGCTCCAG
BCY	(881)	CTTCCGCCAAGGCCCGCGCTGGGCCCCCCGGGCGCACACCGCTCCCCTGCCACTTCCGCGGACCCGGACTCCCGAGGCGCCCCCCCGGGAAGAGCGCTCCAG
MC55.29 BCY I	(881)	CITCCGCCAAGGCCCGCGCTGGGCCCCCCGGGCGCACACCGCTCCCCTGCCACTTCCGCGGAACTGGCGGGCCGCGGAACTCCCGAAGGCGCCCCCCGGGAAGAGCCGCTCCAG
MC55.32 BCY I	(881)	CTTCCGCCAAGGCCGGCGCTGGGCCCCCCGGGGCGCACACCGCTCCCCTGCCGCGGGACCCGAGGCCGGACTCCCGAGGCGCCCCCCGGGGAGAGCCGCTCCAG
Consensus	(881)	CCGCCAAGGCCCGCGCTGGGCCCCCCGGGGCGCACACGCTCCCCTGCCACTTCCGCGGACCCGAGCTGGCCGGACTCCCGAGGCGCGCCCCCGGGAAGCCGCT
		991
original BCY1	(886)	GCTTCTCTAAACTTGGTGGGGGGGGGCCCTGCGGAGCCCCGCAGCCGGGGGGGATTGCATGGTCTGCTTTGAGAGCGAAGTGACTGCCCCTTGTGCCCTGCGGACAA
MC50A19 BCY I	(991)	GGCTTCTCTAAACTIGGTGGGGGGGGGCCTGCGGAGCCCCGGCGGCGGGGGGATTGCATGGTCTGCTTTGAGAGCGAAGTGACTGCCGCCCTTGTGCCTGCGGAAA
MC50A6 BCY I	(991)	GCTTCTCTAAACTIGGIGGGGGGGGCCTGCGGAGCCCCGGCGGCGGGGGATTGCAIGGTTTGATTGAGAGCGAAGIGACTGCCCCTTGTGCCCTGCGGAAA
MC50A8 BCY I	(991)	GGCTTCTCTAAACTTGGTGGGGGGGGGCGGCCTGCGGAGCCCCGGCGGCGGGGGATTGCATGGTCTGCTTTTGAGAGCGAAGTGACTGCCCCTTGTGCCCTTGCGAACAA
MC54.21 BCY I	(991)	GGCTTCTCTAAACTTGGTGGGGGGGGGGCCTGCGGAGCCCCGGCGGCGGGGGATTGCATGGTCTGCTTTGAGAGCGAAGTGACTGCCCCTTGTGCCGGACAA
MC55.29 BCY I	(991)	GCTTCTCTAAACTIGGIGGGGGGGGCCTGCGGAGCCCCGGCGGCGGGGGATIGCAIGGITTGCAIGGTTTGAGAGCGAAGIGACTGCCCCTIGIGCCCTGCGGACAA
MC55.32 BCY I	(991)	GCTTCTCTAAACTTGGTGGGGGGGGCCTGCGGAGCCCCGGCGGCGGGGGGATTGCATGGTCTGCTTTTGAGAGCGAAGTGACTGCCCCTTGTGCCGGCAAA
Consensus	(991)	GGCTTCTCTAAACTIGGIGGGGGGGGGCCTGCGGAGCCCCGG CGGCGGGGGATIGCAIGGICTGCTTTGAGAGCGAAGIGACTGCCCCTTGIGCGAAAA
		1101
original BCY1	(1098)	CAACCTGTTCTGCATGGAGTGTGCAGTACGCATCTGCGAGAGGACCCAGAGTGTCCCGTCTGCCACTCACAGCCACGCAAGCCATCCAAATATTCTCCTAA
MC50A19 BCY I	(1098)	CAACCTGTICIGCAIGGAGIGIGCAGIACGCAICTGCGAGAGGACCCAGAGIGIICCCGTCIGCCACATCACAGCGCGCAAGCCAICCGAAIAIITCTCCIAAGGAI
MC50A6 BCY I	(1098)	CAACCTGTICTGCATGGAGTGTGCAGTACGCATCTGCGAGAGGACCAGAGTGTCCCGTCTGCCACATCACAGCGCGCAAGCCATCCGAATATTCTCCTAAGGAT
MC50A8 BCY I	(1098)	CAACCTGTTCTGCATGGAGTGTGCAGTACGCATCTGCGAGAGGACCGGACCCAGAGTGTCCCGTCTGCCACTCACAGCCGCAGCCATCCGAATATTCTCCTAAGGAT
MC54.21 BCY I	(1098)	CAACCTGTICIGCAIGGAGIGIGCAGIACGCAICIGCGAGAGGACCCAGAGIGIICCCGICIGCCACAICCAGCAGCCGCAAGCCAICCGAAIAIITCICCIAA
MC55.29 BCY I	(1098)	CAACCTGTICIGCATGGAGTGTGCAGTACGCATCTGCGAGAGGACGACCCAGAGTGTCCCGTCTGCCACATCACAGCGCGCAGCAGCCATCCGAATATTCTCCTAA
MC55.32 BCY I	(1098)	CAACCTGTTCTGCATGGAGTGTGCAGTATGCGAGAGGAGGAGGACCCAGAGTGTCCCGTCTGCCACATCACAGCGCGCAAGCCATCCGAATATTTCTCCTAA
Consensus	(1101)	CAACCTGTICIGCAIGGAGIGIGCAGIACGCAICIGCGAGAGGACCCAGAGIGIICCCGICIGCCACATCACAGCGCGCGCAAGCCAICCGAAIAIITCICCIAA
		1211 1214
original BCY1	(1204)	(SEQ ID NO.:222)
MC50A19 BCY I	(1208)	CCACTAGTCCAGTGGGAGATTCTGCAGATATCCA- (SEQ ID NO.:223)
MC50A6 BCY I	(1208)	CCACTAGTCCAGTGTGGAATTCTGCAGATATCCA- (SEQ ID NO.:224)
MC50A8 BCY I	(1208)	CCACTAGTCCAGTGTGGAATTCTGCAGATATCCAG (SEQ ID NO.:225)
MC54.21 BCY I	(1204)	
BCY	(1204)	
MC55.32 BCY I	(1204)	(SEQ ID NO.:228)
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Feb. 3, 2015

# FIGURE 3D

RIGKILEYNN YYGVAEISPP RSPGGGRDCM VCFESEVTAA SMIRASRNKS REIISAAEHF REEIETHIAV PGYGVGKQDV TGAPGNVERA 1 GDFGYGGYLF 1 GRREDVATAR GFSKLGGGGL KINTYIKTPV RGEEPVFMVT TSAGPELAGL PRRPPGEPLQ CIGECGVDSG FEAPRLGEQG SRDRDPVFEI (SEQ ID NO.:4) QQTNTYIITP PKGATIKRIQ LSTFRQNSLG QGCKIKALRA GPPGAHRSPA TAAQAIRIFS PYRVVGLVVG DWRVHQPGCKP SEHVAEIVGR SSSSSAKARA TDPECPVCHI LWAGQENATP TSVLFSSASS LVPCGHNLFC MECAVRICER PGQVTIRVRV NTTECVPVPT AAIDSRYSDA MAELRLKGSS GAAFGVAPAL ENDFLAGSPD

### TUMOR ANTIGENS BFA4 AND BCY1 FOR PREVENTION AND / OR TREATMENT OF CANCER

#### RELATED APPLICATIONS

This application is a divisional application of U.S. Ser. No. 10/611,440 filed Jul. 1, 2003 now U.S. Pat. No. 7,851,213 which claims priority to U.S. Ser. Nos. 60/394,346 filed Jul. 3, 2002; 60/394,503 filed Jul. 9, 2002; 60/411,833 filed Sep. 18, 2002; and, 60/445,342 filed Feb. 6, 2003, all of which being hereby incorporated by reference.

#### FIELD OF THE INVENTION

The present invention relates to a nucleic acid encoding a polypeptide and the use of the nucleic acid or polypeptide in preventing and/or treating cancer. In particular, the invention relates to improved vectors for the insertion and expression of foreign genes encoding tumor antigens for use in immunotherapeutic treatment of cancer.

#### BACKGROUND OF THE INVENTION

There has been tremendous increase in last few years in the development of cancer vaccines with Tumour-associated antigens (TAAs) due to the great advances in identification of molecules based on the expression profiling on primary tumours and normal cells with the help of several techniques 30 such as high density microarray, SEREX, immunohistochemistry (IHC), RT-PCR, in-situ hybridization (ISH) and laser capture microscopy (Rosenberg, Immunity, 1999; Sgroi et al, 1999, Schena et al, 1995, Offringa et al, 2000). The TAAs are antigens expressed or over-expressed by tumour cells and 35 could be specific to one or several tumours for example CEA antigen is expressed in colorectal, breast and lung cancers. Sgroi et al (1999) identified several genes differentially expressed in invasive and metastatic carcinoma cells with combined use of laser capture microdissection and cDNA 40 microarrays. Several delivery systems like DNA or viruses could be used for therapeutic vaccination against human cancers (Bonnet et al, 2000) and can elicit immune responses and also break immune tolerance against TAAs. Tumour cells can be rendered more immunogenic by inserting transgenes 45 encoding T cell co-stimulatory molecules such as B7.1 or cytokines such as IFN-γ, IL2, or GM-CSF, among others. Co-expression of a TAA and a cytokine or a co-stimulatory molecule can develop effective therapeutic vaccine (Hodge et al, 95, Bronte et al, 1995, Chamberlain et al, 1996).

There is a need in the art for reagents and methodologies useful in stimulating an immune response to prevent or treat cancers. The present invention provides such reagents and methodologies which overcome many of the difficulties encountered by others in attempting to treat cancer.

#### SUMMARY OF THE INVENTION

The present invention provides an immunogenic target for administration to a patient to prevent and/or treat cancer. In 60 particular, the immunogenic target is a tumor antigen ("TA") and/or an angiogenesis-associated antigen ("AA"). In one embodiment, the immunogenic target is encoded by SEQ ID NO.: 1 or 3 or has the amino acid sequence of SEQ ID NO.: 2 or 4. In certain embodiments, the TA and/or AA are administered to a patient as a nucleic acid contained within a plasmid or other delivery vector, such as a recombinant virus. The

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TA and/or AA may also be administered in combination with an immune stimulator, such as a co-stimulatory molecule or adjuvant.

#### BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1. BFA4 cDNA sequence.

FIG. 2. BFA4 amino acid sequence.

FIG. 3. BCY1 nucleotide (A-C) and amino acid (D)  $^{\rm 10}$  sequences.

#### DETAILED DESCRIPTION

The present invention provides reagents and methodolo-15 gies useful for treating and/or preventing cancer. All references cited within this application are incorporated by reference.

In one embodiment, the present invention relates to the induction or enhancement of an immune response against one or more tumor antigens ("TA") to prevent and/or treat cancer. In certain embodiments, one or more TAs may be combined. In preferred embodiments, the immune response results from expression of a TA in a host cell following administration of a nucleic acid vector encoding the tumor antigen or the tumor antigen itself in the form of a peptide or polypeptide, for example.

As used herein, an "antigen" is a molecule (such as a polypeptide) or a portion thereof that produces an immune response in a host to whom the antigen has been administered. The immune response may include the production of antibodies that bind to at least one epitope of the antigen and/or the generation of a cellular immune response against cells expressing an epitope of the antigen. The response may be an enhancement of a current immune response by, for example, causing increased antibody production, production of antibodies with increased affinity for the antigen, or an increased cellular response (i.e., increased T cells). An antigen that produces an immune response may alternatively be referred to as being immunogenic or as an immunogen. In describing the present invention, a TA may be referred to as an "immunogenic target".

TA includes both tumor-associated antigens (TAAs) and tumor-specific antigens (TSAs), where a cancerous cell is the source of the antigen. A TAA is an antigen that is expressed on the surface of a tumor cell in higher amounts than is observed on normal cells or an antigen that is expressed on normal cells during fetal development. A TSA is an antigen that is unique to tumor cells and is not expressed on normal cells. TA further includes TAAs or TSAs, antigenic fragments thereof, and modified versions that retain their antigenicity.

TAs are typically classified into five categories according to their expression pattern, function, or genetic origin: cancertestis (CT) antigens (i.e., MAGE, NY-ESO-1); melanocyte differentiation antigens (i.e., Melan A/MART-1, tyrosinase, 55 gp100); mutational antigens (i.e., MUM-1, p53, CDK-4); overexpressed 'self' antigens (i.e., HER-2/neu, p53); and, viral antigens (i.e., HPV, EBV). For the purposes of practicing the present invention, a suitable TA is any TA that induces or enhances an anti-tumor immune response in a host to whom the TA has been administered. Suitable TAs include, for example, gp100 (Cox et al., Science, 264:716-719 (1994)), MART-1/Melan A (Kawakami et al., J. Exp. Med., 180:347-352 (1994)), gp75 (TRP-1) (Wang et al., J. Exp. Med., 186:1131-1140 (1996)), tyrosinase (Wolfel et al., Eur. J. Immunol., 24:759-764 (1994); WO 200175117; WO 200175016; WO 200175007), NY-ESO-1 (WO 98/14464; WO 99/18206), melanoma proteoglycan (Hellstrom et al., J.

Immunol., 130:1467-1472 (1983)), MAGE family antigens (i.e., MAGE-1, 2,3,4,6,12, 51; Van der Bruggen et al., Science, 254:1643-1647 (1991); U.S. Pat. No. 6,235,525; CN 1319611), BAGE family antigens (Boel et al., *Immunity*, 2:167-175 (1995)), GAGE family antigens (i.e., GAGE-1,2; 5 Van den Eynde et al., J. Exp. Med., 182:689-698 (1995); U.S. Pat. No. 6,013,765), RAGE family antigens (i.e., RAGE-1; Gaugler et at., Immunogenetics, 44:323-330 (1996); U.S. Pat. No. 5,939,526), N-acetylglucosaminyltransferase-V (Guilloux et al., J. Exp. Med., 183:1173-1183 (1996)), p15 (Robbins et al., J. Immunol. 154:5944-5950 (1995)), β-catenin (Robbins et al., J. Exp. Med., 183:1185-1192 (1996)), MUM-1 (Coulie et al., Proc. Natl. Acad. Sci. USA, 92:7976-7980 (1995)), cyclin dependent kinase-4 (CDK4) (Wolfel et al., Science, 269:1281-1284 (1995)), p21-ras (Fossum et al., 15 Int. J. Cancer, 56:40-45 (1994)), BCR-abl (Bocchia et al., Blood, 85:2680-2684 (1995)), p53 (Theobald et al., Proc. Natl. Acad. Sci. USA, 92:11993-11997 (1995)), p185 HER2/ neu (erb-B1; Fisk et al., J. Exp. Med., 181:2109-2117 (1995)), epidermal growth factor receptor (EGFR) (Harris et al., 20 Breast Cancer Res. Treat, 29:1-2 (1994)), carcinoembryonic to antigens (CEA) (Kwong et al., J. Natl. Cancer Inst., 85:982-990 (1995) U.S. Pat. Nos. 5,756,103; 5,274,087; 5,571,710; 6,071,716; 5,698,530; 6,045,802; EP 263933; EP 346710; and, EP 784483); carcinoma-associated mutated 25 mucins (i.e., MUC-1 gene products; Jerome et al., J. Immunol., 151:1654-1662 (1993)); EBNA gene products of EBV (i.e., EBNA-1; Rickinson et al., Cancer Surveys, 13:53-80 (1992)); E7, E6 proteins of human papillomavirus (Ressing et al., J. Immunol, 154:5934-5943 (1995)); prostate specific 30 antigen (PSA; Xue et al., *The Prostate*, 30:73-78 (1997)); prostate specific membrane antigen (PSMA; Israeli, et al., Cancer Res., 54:1807-1811 (1994)); idiotypic epitopes or antigens, for example, immunoglobulin idiotypes or T cell receptor idiotypes (Chen et al., J. Immunol., 153:4775-4787 35 (1994)); KSA (U.S. Pat. No. 5,348,887), kinesin 2 (Dietz, et al. Biochem Biophys Res Commun 2000 Sep. 7; 275(3):731-8), HIP-55, TGFβ-1 anti-apoptotic factor (Toomey, et al. Br J Biomed Sci 2001; 58(3):177-83), tumor protein D52 (Bryne J. A., et al., Genomics, 35:523-532 (1996)), HIFT, NY-BR-1 40 (WO 01/47959), NY-BR-62, NY-BR-75, NY-BR-85, NY-BR-87, NY-BR-96 (Scanlan, M. Serologic and Bioinformatic Approaches to the Identification of Human Tumor Antigens, in Cancer Vaccines 2000, Cancer Research Institute, New York, N.Y.), BFA4 (SEQ ID NOS.: 26 and 27), or BCY1 45 (SEQ ID NOS.: 28 and 29), including "wild-type" (i.e., normally encoded by the genome, naturally-occurring), modified, and mutated versions as well as other fragments and derivatives thereof. Any of these TAs may be utilized alone or in combination with one another in a co-immunization pro- 50 tocol.

In certain cases, it may be beneficial to co-immunize patients with both TA and other antigens, such as angiogenesis-associated antigens ("AA"). An AA is an immunogenic molecule (i.e., peptide, polypeptide) associated with cells 55 involved in the induction and/or continued development of blood vessels. For example, an AA may be expressed on an endothelial cell ("EC"), which is a primary structural component of blood vessels. Where the cancer is cancer, it is preferred that that the AA be found within or near blood vessels that supply a tumor. Immunization of a patient against an AA preferably results in an anti-AA immune response whereby angiogenic processes that occur near or within tumors are prevented and/or inhibited.

Exemplary AAs include, for example, vascular endothelial 65 growth factor (i.e., VEGF; Bernardini, et al. *J. Urol.*, 2001, 166(4): 1275-9; Starnes, et al. *J. Thorac. Cardiovasc. Surg.*,

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2001, 122(3): 518-23; Dias, et al. Blood, 2002, 99: 2179-2184), the VEGF receptor (i.e., VEGF-R, flk-1/KDR; Starnes, et al. J. Thorac. Cardiovasc. Surg., 2001, 122(3): 518-23), EPH receptors (i.e., EPHA2; Gerety, et al. 1999, Cell, 4: 403-414), epidermal growth factor receptor (i.e., EGFR; Ciardeillo, et al. Clin. Cancer Res., 2001, 7(10): 2958-70), basic fibroblast growth factor (i.e., bFGF; Davidson, et al. Clin. Exp. Metastasis 2000, 18(6): 501-7; Poon, et al. Am J. Surg., 2001, 182(3):298-304), platelet-derived cell growth factor (i.e., PDGF-B), platelet-derived endothelial cell growth factor (PD-ECGF; Hong, et al. J. Mol. Med., 2001, 8(2):141-8), transforming growth factors (i.e., TGF-α; Hong, et al. J. Mol. Med., 2001, 8(2):141-8), endoglin (Balza, et al. Int. J. Cancer, 2001, 94: 579-585), Id proteins (Benezra, R. Trends Cardiovasc. Med., 2001, 11(6):237-41), proteases such as uPA, uPAR, and matrix metalloproteinases (MMP-2, MMP-9; Djonov, et al. J. Pathol., 2001, 195(2):147-55), nitric oxide synthase (Am. J. Ophthalmol., 2001, 132(4):551-6), aminopeptidase (Rouslhati, E. Nature Cancer, 2: 84-90, 2002), thrombospondins (i.e., TSP-1, TSP-2; Alvarez, et al. Gynecol. Oncol., 2001, 82(2):273-8; Seki, et al. Int. J. Oncol., 2001, 19(2):305-10), k-ras (Zhang, et al. Cancer Res., 2001, 61(16):6050-4), Wnt (Zhang, et al. Cancer Res., 2001, 61(16):6050-4), cyclin-dependent kinases (CDKs; Drug Resist. Updat. 2000, 3(2):83-88), microtubules (Timar, et al. 2001. Path. Oncol. Res., 7(2): 85-94), heat shock proteins (i.e., HSP90 (Timar, supra)), heparin-binding factors (i.e., heparinase; Gohji, et al. Int. J. Cancer, 2001, 95(5):295-301), synthases (i.e., ATP synthase, thymidilate synthase), collagen receptors, integrins (i.e.,  $\alpha \nu \beta 3$ ,  $\alpha \nu \beta 5$ ,  $\alpha 1\beta 1$ ,  $\alpha 2\beta 1$ ,  $\alpha 5\beta 1$ ), or surface proteolglycan NG2, among others, including "wildtype" (i.e., normally encoded by the genome, naturally-occurring), modified, mutated versions as well as other fragments and derivatives thereof. Any of these targets may be suitable in practicing the present invention, either alone or in combination with one another or with other agents.

In certain embodiments, a nucleic acid molecule encoding an immunogenic target is utilized. The nucleic acid molecule may comprise or consist of a nucleotide sequence encoding one or more immunogenic targets, or fragments or derivatives thereof, such as that contained in a DNA insert in an ATCC Deposit. The term "nucleic acid sequence" or "nucleic acid molecule" refers to a DNA or RNA sequence. The term encompasses molecules formed from any of the known base analogs of DNA and RNA such as, but not limited to 4-acetylcytosine, 8-hydroxy-N6-methyladenosine, aziridinyl-cypseudoisocytosine, 5-(carboxyhydroxylmethyl) tosine. uracil, 5-fluorouracil, 5-bromouracil, 5-carboxymethylaminomethyl-2-thiouracil, 5-carboxy-methylaminomethyluracil, dihydrouracil, inosine, N6-iso-pentenyladenine, 1-methyladenine. 1-methylpseudouracil, 1-methylguanine, 1-methylinosine, 2,2-dimethyl-guanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-methyladenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyamino-methyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarbonyl-methyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5oxyacetic acid methylester, uracil-5-oxyacetic acid, oxybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, N-uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid, pseudouracil, queosine, 2-thiocytosine, and 2,6diaminopurine, among others.

An isolated nucleic acid molecule is one that: (1) is separated from at least about 50 percent of proteins, lipids, carbohydrates, or other materials with which it is naturally found when total nucleic acid is isolated from the source cells; (2) is

not be linked to all or a portion of a polynucleotide to which the nucleic acid molecule is linked in nature; (3) is operably linked to a polynucleotide which it is not linked to in nature; and/or, (4) does not occur in nature as part of a larger polynucleotide sequence. Preferably, the isolated nucleic acid molecule of the present invention is substantially free from any other contaminating nucleic acid molecule(s) or other contaminants that are found in its natural environment that would interfere with its use in polypeptide production or its therapeutic, diagnostic, prophylactic or research use. As used herein, the term "naturally occurring" or "native" or "naturally found" when used in connection with biological materials such as nucleic acid molecules, polypeptides, host cells, and the like, refers to materials which are found in nature and are not manipulated by man. Similarly, "non-naturally occurring" or "non-native" as used herein refers to a material that is not found in nature or that has been structurally modified or synthesized by man.

The identity of two or more nucleic acid or polypeptide 20 molecules is determined by comparing the sequences. As known in the art, "identity" means the degree of sequence relatedness between nucleic acid molecules or polypeptides as determined by the match between the units making up the molecules (i.e., nucleotides or amino acid residues). Identity 25 measures the percent of identical matches between the smaller of two or more sequences with gap alignments (if any) addressed by a particular mathematical model or computer program (i.e., an algorithm). Identity between nucleic acid sequences may also be determined by the ability of the 30 related sequence to hybridize to the nucleic acid sequence or isolated nucleic acid molecule. In defining such sequences, the term "highly stringent conditions" and "moderately stringent conditions" refer to procedures that permit hybridization of nucleic acid strands whose sequences are complementary, 35 and to exclude hybridization of significantly mismatched nucleic acids. Examples of "highly stringent conditions" for hybridization and washing are 0.015 M sodium chloride, 0.0015 M sodium citrate at 65-68° C. or 0.015 M sodium chloride, 0.0015 M sodium citrate, and 50% formamide at 42° 40 C. (see, for example, Sambrook, Fritsch & Maniatis, Molecular Cloning: A Laboratory Manual (2nd ed., Cold Spring Harbor Laboratory, 1989); Anderson et al., Nucleic Acid Hybridisation: A Practical Approach Ch. 4 (IRL Press Limited)). The term "moderately stringent conditions" refers to 45 conditions under which a DNA duplex with a greater degree of base pair mismatching than could occur under "highly stringent conditions" is able to form. Exemplary moderately stringent conditions are 0.015 M sodium chloride, 0.0015 M sodium citrate at 50-65° C. or 0.015 M sodium chloride, 50 0.0015 M sodium citrate, and 20% formamide at 37-50° C. By way of example, moderately stringent conditions of 50° C. in 0.015 M sodium ion will allow about a 21% mismatch. During hybridization, other agents may be included in the hybridization and washing buffers for the purpose of reducing 55 non-specific and/or background hybridization. Examples are 0.1% bovine serum albumin, 0.1% polyvinyl-pyrrolidone, 0.1% sodium pyrophosphate, 0.1% sodium dodecylsulfate, NaDodSO<sub>4</sub>, (SDS), ficoll, Denhardt's solution, sonicated salmon sperm DNA (or another non-complementary DNA), 60 and dextran sulfate, although other suitable agents can also be used. The concentration and types of these additives can be changed without substantially affecting the stringency of the hybridization conditions. Hybridization experiments are usually carried out at pH 6.8-7.4; however, at typical ionic strength conditions, the rate of hybridization is nearly independent of pH.

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In preferred embodiments of the present invention, vectors are used to transfer a nucleic acid sequence encoding a polypeptide to a cell. A vector is any molecule used to transfer a nucleic acid sequence to a host cell. In certain cases, an expression vector is utilized. An expression vector is a nucleic acid molecule that is suitable for transformation of a host cell and contains nucleic acid sequences that direct and/or control the expression of the transferred nucleic acid sequences. Expression includes, but is not limited to, processes such as transcription, translation, and splicing, if introns are present. Expression vectors typically comprise one or more flanking sequences operably linked to a heterologous nucleic acid sequence encoding a polypeptide. Flanking sequences may be homologous (i.e., from the same species and/or strain as the host cell), heterologous (i.e., from a species other than the host cell species or strain), hybrid (i.e., a combination of flanking sequences from more than one source), or synthetic, for example.

A flanking sequence is preferably capable of effecting the replication, transcription and/or translation of the coding sequence and is operably linked to a coding sequence. As used herein, the term operably linked refers to a linkage of polynucleotide elements in a functional relationship. For instance, a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the coding sequence. However, a flanking sequence need not necessarily be contiguous with the coding sequence, so long as it functions correctly. Thus, for example, intervening untranslated yet transcribed sequences can be present between a promoter sequence and the coding sequence and the promoter sequence may still be considered operably linked to the coding sequence. Similarly, an enhancer sequence may be located upstream or downstream from the coding sequence and affect transcription of the sequence.

In certain embodiments, it is preferred that the flanking sequence is a trascriptional regulatory region that drives highlevel gene expression in the target cell. The transcriptional regulatory region may comprise, for example, a promoter, enhancer, silencer, repressor element, or combinations thereof. The transcriptional regulatory region may be either constitutive, tissue-specific, cell-type specific (i.e., the region is drives higher levels of transcription in a one type of tissue or cell as compared to another), or regulatable (i.e., responsive to interaction with a compound such as tetracycline). The source of a transcriptional regulatory region may be any prokaryotic or eukaryotic organism, any vertebrate or invertebrate organism, or any plant, provided that the flanking sequence functions in a cell by causing transcription of a nucleic acid within that cell. A wide variety of transcriptional regulatory regions may be utilized in practicing the present invention.

Suitable transcriptional regulatory regions include the CMV promoter (i.e., the CMV-immediate early promoter); promoters from eukaryotic genes (i.e., the estrogen-inducible chicken ovalbumin gene, the interferon genes, the glucocorticoid-inducible tyrosine aminotransferase gene, and the thymidine kinase gene); and the major early and late adenovirus gene promoters; the SV40 early promoter region (Bernoist and Chambon, 1981, Nature 290:304-10); the promoter contained in the 3' long terminal repeat (LTR) of Rous sarcoma virus (RSV) (Yamamoto, et al., 1980, Cell 22:787-97); the herpes simplex virus thymidine kinase (HSV-TK) promoter (Wagner et al., 1981, Proc. Natl. Acad. Sci. U.S.A. 78:1444-45); the regulatory sequences of the metallothionine gene (Brinster et al., 1982, Nature 296:39-42); prokaryotic expression vectors such as the beta-lactamase promoter (Villa-Kamaroff et al., 1978, Proc. Natl. Acad. Sci. U.S.A.,

75:3727-31); or the tac promoter (DeBoer et al., 1983, Proc. Natl. Acad. Sci. U.S.A., 80:21-25). Tissue- and/or cell-type specific transcriptional control regions include, for example, the elastase 1 gene control region which is active in pancreatic acinar cells (Swift et al., 1984, Cell 38:639-46; Ornitz et al., 5 1986, Cold Spring Harbor Symp. Quant. Biol. 50:399-409 (1986); MacDonald, 1987, Hepatology 7:425-515); the insulin gene control region which is active in pancreatic beta cells (Hanahan, 1985, Nature 315:115-22); the immunoglobulin gene control region which is active in lymphoid cells (Gross-10 chedl et al., 1984, Cell 38:647-58; Adames et al., 1985, Nature 318:533-38; Alexander et al., 1987, Mol. Cell. Biol., 7:1436-44); the mouse mammary tumor virus control region in testicular, breast, lymphoid and mast cells (Leder et al., 1986, Cell 45:485-95); the albumin gene control region in 15 liver (Pinkert et al., 1987, Genes and Devel. 1:268-76); the alpha-feto-protein gene control region in liver (Krumlauf et al., 1985, Mol. Cell. Biol., 5:1639-48; Hammer et al., 1987, Science 235:53-58); the alpha 1-antitrypsin gene control region in liver (Kelsev et al., 1987, Genes and Devel. 1:161-20 71); the beta-globin gene control region in myeloid cells (Mogram et al., 1985, Nature 315:338-40; Kollias et al., 1986, Cell 46:89-94); the myelin basic protein gene control region in oligodendrocyte cells in the brain (Readhead et al., 1987, Cell 48:703-12); the myosin light chain-2 gene control region 25 in skeletal muscle (Sani, 1985, Nature 314:283-86); the gonadotropic releasing hormone gene control region in the hypothalamus (Mason et al., 1986, Science 234:1372-78), and the tyrosinase promoter in melanoma cells (Hart, I. Semin Oncol 1996 February; 23(1):154-8; Siders, et al. Cancer Gene Ther 30 1998 September-October; 5(5):281-91), among others. Inducible promoters that are activated in the presence of a certain compound or condition such as light, heat, radiation, tetracycline, or heat shock proteins, for example, may also be utilized (see, for example, WO 00/10612). Other suitable 35 promoters are known in the art.

As described above, enhancers may also be suitable flanking sequences. Enhancers are cis-acting elements of DNA, usually about 10-300 by in length, that act on the promoter to increase transcription. Enhancers are typically orientation- 40 and position-independent, having been identified both 5' and 3' to controlled coding sequences. Several enhancer sequences available from mammalian genes are known (i.e., globin, elastase, albumin, alpha-feto-protein and insulin). Similarly, the SV40 enhancer, the cytomegalovirus early pro- 45 moter enhancer, the polyoma enhancer, and adenovirus enhancers are useful with eukaryotic promoter sequences. While an enhancer may be spliced into the vector at a position 5' or 3' to nucleic acid coding sequence, it is typically located at a site 5' from the promoter. Other suitable enhancers are 50 known in the art, and would be applicable to the present invention.

While preparing reagents of the present invention, cells may need to be transfected or transformed. Transfection refers to the uptake of foreign or exogenous DNA by a cell, 55 and a cell has been transfected when the exogenous DNA has been introduced inside the cell membrane. A number of transfection techniques are well known in the art (i.e., Graham et al., 1973, *Virology* 52:456; Sambrook et al., *Molecular Cloning, A Laboratory Manual* (Cold Spring Harbor Laboratories, 60 1989); Davis et al., *Basic Methods in Molecular Biology* (Elsevier, 1986); and Chu et al., 1981, *Gene* 13:197). Such techniques can be used to introduce one or more exogenous DNA moieties into suitable host cells.

In certain embodiments, it is preferred that transfection of 65 a cell results in transformation of that cell. A cell is transformed when there is a change in a characteristic of the cell,

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being transformed when it has been modified to contain a new nucleic acid. Following transfection, the transfected nucleic acid may recombine with that of the cell by physically integrating into a chromosome of the cell, may be maintained transiently as an episomal element without being replicated, or may replicate independently as a plasmid. A cell is stably transformed when the nucleic acid is replicated with the division of the cell.

The present invention further provides isolated immunogenic targets in polypeptide form. A polypeptide is considered isolated where it: (1) has been separated from at least about 50 percent of polynucleotides, lipids, carbohydrates, or other materials with which it is naturally found when isolated from the source cell; (2) is not linked (by covalent or noncovalent interaction) to all or a portion of a polypeptide to which the "isolated polypeptide" is linked in nature; (3) is operably linked (by covalent or noncovalent interaction) to a polypeptide with which it is not linked in nature; or, (4) does not occur in nature. Preferably, the isolated polypeptide is substantially free from any other contaminating polypeptides or other contaminants that are found in its natural environment that would interfere with its therapeutic, diagnostic, prophylactic or research use.

Immunogenic target polypeptides may be mature polypeptides, as defined herein, and may or may not have an amino terminal methionine residue, depending on the method by which they are prepared. Further contemplated are related polypeptides such as, for example, fragments, variants (i.e., allelic, splice), orthologs, homologues, and derivatives, for example, that possess at least one characteristic or activity (i.e., activity, antigenicity) of the immunogenic target. Also related are peptides, which refers to a series of contiguous amino acid residues having a sequence corresponding to at least a portion of the polypeptide from which its sequence is derived. In preferred embodiments, the peptide comprises about 5-10 amino acids, 10-15 amino acids, 15-20 amino acids, 20-30 amino acids, or 30-50 amino acids. In a more preferred embodiment, a peptide comprises 9-12 amino acids, suitable for presentation upon Class I MHC molecules, for example.

A fragment of a nucleic acid or polypeptide comprises a truncation of the sequence (i.e., nucleic acid or polypeptide) at the amino terminus (with or without a leader sequence) and/or the carboxy terminus. Fragments may also include variants (i.e., allelic, splice), orthologs, homologues, and other variants having one or more amino acid additions or substitutions or internal deletions as compared to the parental sequence. In preferred embodiments, truncations and/or deletions comprise about 10 amino acids, 20 amino acids, 30 amino acids, 40 amino acids, 50 amino acids, or more. The polypeptide fragments so produced will comprise about 10 amino acids, 25 amino acids, 30 amino acids, 40 amino acids, 50 amino acids, 60 amino acids, 70 amino acids, or more. Such polypeptide fragments may optionally comprise an amino terminal methionine residue. It will be appreciated that such fragments can be used, for example, to generate antibodies or cellular immune responses to immunogenic target polypeptides.

A variant is a sequence having one or more sequence substitutions, deletions, and/or additions as compared to the subject sequence. Variants may be naturally occurring or artificially constructed. Such variants may be prepared from the corresponding nucleic acid molecules. In preferred embodiments, the variants have from 1 to 3, or from 1 to 5, or from 1 to 10, or from 1 to 15, or from 1 to 20, or from 1 to 25, or from 1 to 30, or from 1 to 40, or from 1 to 50, or more than 50 amino acid substitutions, insertions, additions and/or deletions.

An allelic variant is one of several possible naturally-occurring alternate forms of a gene occupying a given locus on a chromosome of an organism or a population of organisms. A splice variant is a polypeptide generated from one of several RNA transcript resulting from splicing of to a primary transcript. An ortholog is a similar nucleic acid or polypeptide sequence from another species. For example, the mouse and human versions of an immunogenic target polypeptide may be considered orthologs of each other. A derivative of a sequence is one that is derived from a parental sequence those sequences having substitutions, additions, deletions, or chemically modified variants. Variants may also include fusion proteins, which refers to the fusion of one or more first sequences (such as a peptide) at the amino or carboxy terminus of at least one other sequence (such as a heterologous peptide).

"Similarity" is a concept related to identity, except that similarity refers to a measure of relatedness which includes both identical matches and conservative substitution matches. If two polypeptide sequences have, for example, 10/20 identical amino acids, and the remainder are all non-conservative substitutions, then the percent identity and similarity would both be 50%. If in the same example, there are five more positions where there are conservative substitutions, then the percent identity remains 50%, but the percent similarity would be 75% (15/20). Therefore, in cases where there are conservative substitutions, the percent similarity between two polypeptides will be higher than the percent identity between those two polypeptides.

Substitutions may be conservative, or non-conservative, or any combination thereof. Conservative amino acid modifications to the sequence of a polypeptide (and the corresponding modifications to the encoding nucleotides) may produce polypeptides having functional and chemical characteristics similar to those of a parental polypeptide. For example, a "conservative amino acid substitution" may involve a substitution of a native amino acid residue with a non-native residue such that there is little or no effect on the size, polarity, charge, hydrophobicity, or hydrophilicity of the amino acid residue at that position and, in particular, does not result in decreased immunogenicity. Suitable conservative amino acid substitutions are shown in Table I.

TABLE I

Original Residues	Exemplary Substitutions	Preferred Substitutions
Ala	Val, Leu, Ile	Val
Arg	Lys, Gln, Asn	Lys
Asn	Gln	Gln
Asp	Glu	Glu
Cys	Ser, Ala	Ser
Gln	Asn	Asn
Glu	Asp	Asp
Gly	Pro, Ala	Ala
His	Asn, Gln, Lys, Arg	Arg
Ile	Leu, Val, Met, Ala, Phe, Norleucine	Leu
Leu	Norleucine, Ile, Val, Met, Ala, Phe	Ile
Lys	Arg, 1,4 Diamino-butyric Acid, Gln, Asn	Arg
Met	Leu, Phe, Ile	Leu
Phe	Leu, Val, Ile, Ala, Tyr	Leu
Pro	Ala	Gly
Ser	Thr, Ala, Cys	Thr
Thr	Ser	Ser
Trp	Tyr, Phe	Tyr
Tyr	Trp, Phe, Thr, Ser	Phe
Val	Ile, Met, Leu, Phe, Ala, Norleucine	Leu

A skilled artisan will be able to determine suitable variants of polypeptide using well-known techniques. For identifying

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suitable areas of the molecule that may be changed without destroying biological activity (i.e., MHC binding, immunogenicity), one skilled in the art may target areas not believed to be important for that activity. For example, when similar polypeptides with similar activities from the same species or from other species are known, one skilled in the art may compare the amino acid sequence of a polypeptide to such similar polypeptides. By performing such analyses, one can identify residues and portions of the molecules that are conserved among similar polypeptides. It will be appreciated that changes in areas of the molecule that are not conserved relative to such similar polypeptides would be less likely to adversely affect the biological activity and/or structure of a polypeptide. Similarly, the residues required for binding to MHC are known, and may be modified to improve binding. However, modifications resulting in decreased binding to MHC will not be appropriate in most situations. One skilled in the art would also know that, even in relatively conserved regions, one may substitute chemically similar amino acids for the naturally occurring residues while retaining activity. Therefore, even areas that may be important for biological activity or for structure may be subject to conservative amino acid substitutions without destroying the biological activity or without adversely affecting the polypeptide structure.

Other preferred polypeptide variants-include glycosylation variants wherein the number and/or type of glycosylation sites have been altered compared to the subject amino acid sequence. In one embodiment, polypeptide variants comprise a greater or a lesser number of N-linked glycosylation sites than the subject amino acid sequence. An N-linked glycosylation site is characterized by the sequence Asn-X-Ser or Asn-X-Thr, wherein the amino acid residue designated as X may be any amino acid residue except proline. The substitution of amino acid residues to create this sequence provides a potential new site for the addition of an N-linked carbohydrate chain. Alternatively, substitutions that eliminate this sequence will remove an existing N-linked carbohydrate chain. Also provided is a rearrangement of N-linked carbohydrate chains wherein one or more N-linked glycosylation sites (typically those that are naturally occurring) are eliminated and one or more new N-linked sites are created. To affect O-linked glycosylation of a polypeptide, one would modify serine and/or threonine residues.

Additional preferred variants include cysteine variants, wherein one or more cysteine residues are deleted or substituted with another amino acid (e.g., serine) as compared to the subject amino acid sequence set. Cysteine variants are useful when polypeptides must be refolded into a biologically active conformation such as after the isolation of insoluble inclusion bodies. Cysteine variants generally have fewer cysteine residues than the native protein, and typically have an even number to minimize interactions resulting from unpaired cysteines

In other embodiments, the isolated polypeptides of the current invention include fusion polypeptide segments that assist in purification of the polypeptides. Fusions can be made either at the amino terminus or at the carboxy terminus of the subject polypeptide variant thereof. Fusions may be direct with no linker or adapter molecule or may be through a linker or adapter molecule may be one or more amino acid residues, typically from about 20 to about 50 amino acid residues. A linker or adapter molecule may also be designed with a cleavage site for a DNA restriction endonuclease or for a protease to allow for the separation of the fusion polypeptides can be derivatized according to the methods described herein. Suitable fusion segments include,

among others, metal binding domains (e.g., a poly-histidine segment), immunoglobulin binding domains (i.e., Protein A, Protein G, T cell, B cell, Fc receptor, or complement protein antibody-binding domains), sugar binding domains (e.g., a maltose binding domain), and/or a "tag" domain (i.e., at least 5 a portion of  $\alpha$ -galactosidase, a strep tag peptide, a T7 tag peptide, a FLAG peptide, or other domains that can be purified using compounds that bind to the domain, such as monoclonal antibodies). This tag is typically fused to the polypeptide upon expression of the polypeptide, and can serve as a 10 means for affinity purification of the sequence of interest polypeptide from the host cell. Affinity purification can be accomplished, for example, by column chromatography using antibodies against the tag as an affinity matrix. Optionally, the tag can subsequently be removed from the purified 15 sequence of interest polypeptide by various means such as using certain peptidases for cleavage. As described below, fusions may also be made between a TA and a co-stimulatory components such as the chemokines CXC10 (IP-10), CCL7 (MCP-3), or CCL5 (RANTES), for example.

A fusion motif may enhance transport of an immunogenic target to an MHC processing compartment, such as the endoplasmic reticulum. These sequences, referred to as tranduction or transcytosis sequences, include sequences derived from HIV tat (see Kim et al. 1997 J. Immunol. 159:1666), 25 Drosophila antennapedia (see Schutze-Redelmeier et al. 1996 J. Immunol. 157:650), or human period-1 protein (hPER1; in particular, SRRHHCRSKAKRSRHH (SEQ ID NO: 219).

In addition, the polypeptide or variant thereof may be fused 30 to a homologous polypeptide to form a homodimer or to a heterologous polypeptide to form a heterodimer. Heterologous peptides and polypeptides include, but are not limited to: an epitope to allow for the detection and/or isolation of a fusion polypeptide; a transmembrane receptor protein or a 35 portion thereof, such as an extracellular domain or a transmembrane and intracellular domain; a ligand or a portion thereof which binds to a transmembrane receptor protein; an enzyme or portion thereof which is catalytically active; a polypeptide or peptide which promotes oligomerization, such 40 as a leucine zipper domain; a polypeptide or peptide which increases stability, such as an immunoglobulin constant region; and a polypeptide which has a therapeutic activity different from the polypeptide or variant thereof.

In certain embodiments, it may be advantageous to com- 45 bine a nucleic acid sequence encoding an immunogenic target, polypeptide, or derivative thereof with one or more costimulatory component(s) such as cell surface proteins, cytokines or chemokines in a composition of the present invention. The co-stimulatory component may be included in 50 the composition as a polypeptide or as a nucleic acid encoding the polypeptide, for example. Suitable co-stimulatory molecules include, for instance, polypeptides that bind members of the CD28 family (i.e., CD28, ICOS; Hutloff, et al. Nature 2049-2058) such as the CD28 binding polypeptides B7.1 (CD80; Schwartz, 1992; Chen et al, 1992; Ellis, et al. J. *Immunol.*, 156(8): 2700-9) and B7.2 (CD86; Ellis, et al. J. Immunol., 156(8): 2700-9); polypeptides which bind members of the integrin family (i.e., LFA-1 (CD11a/CD18); Sed- 60 wick, et al. J Immunol 1999, 162: 1367-1375; Wülfing, et al. Science 1998, 282: 2266-2269; Lub, et al. Immunol Today 1995, 16: 479-483) including members of the ICAM family (i.e., ICAM-1, -2 or -3); polypeptides which bind CD2 family members (i.e., CD2, signalling lymphocyte activation molecule (CDw150 or "SLAM"; Aversa, et al. J Immunol 1997, 158: 4036-4044)) such as CD58 (LFA-3; CD2 ligand; Davis,

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et al. Immunol Today 1996, 17: 177-187) or SLAM ligands (Sayos, et al. Nature 1998, 395: 462-469); polypeptides which bind heat stable antigen (HSA or CD24; Zhou, et al. Eur J Immunol 1997, 27: 2524-2528); polypeptides which bind to members of the TNF receptor (TNFR) family (i.e., 4-1BB (CD137; Vinay, et al. Semin Immunol 1998, 10: 481-489), OX40 (CD134; Weinberg, et al. Semin Immunol 1998, 10: 471-480; Higgins, et al. *J Immunol* 1999, 162: 486-493), and CD27 (Lens, et al. Semin Immunol 1998, 10: 491-499)) such as 4-1BBL (4-1BB ligand; Vinay, et al. Semin Immunol 1998, 10: 481-48; DeBenedette, et al. J Immunol 1997, 158: 551-559), TNFR associated factor-1 (TRAF-1; 4-1BB ligand; Saoulli, et al. JExp Med 1998, 187: 1849-1862, Arch, et al. Mol Cell Biol 1998, 18: 558-565), TRAF-2 (4-1BB and OX40 ligand; Saoulli, et al. *JExp Med* 1998, 187: 1849-1862; Oshima, et al. Int Immunol 1998, 10: 517-526, Kawamata, et al. J Biol Chem 1998, 273: 5808-5814), TRAF-3 (4-1BB and OX40 ligand; Arch, et al. Mol Cell Biol 1998, 18: 558-565; Jang, et al. Biochem Biophys Res Commun 1998, 242: 613-20 620; Kawamata S, et al. *J Biol Chem* 1998, 273: 5808-5814). OX40L (OX40 ligand; Gramaglia, et al. J Immunol 1998, 161: 6510-6517), TRAF-5 (OX40 ligand; Arch, et al. Mol Cell Biol 1998, 18: 558-565; Kawamata, et al. J Biol Chem 1998, 273: 5808-5814), and CD70 (CD27 ligand; Couderc, et al. Cancer Gene Ther., 5(3): 163-75). CD154 (CD40 ligand or "CD40L"; Gurunathan, et al. J Immunol., 1998, 161: 4563-4571; Sine, et al. Hum. Gene Ther., 2001, 12: 1091-1102) may also be suitable.

One or more cytokines may also be suitable co-stimulatory components or "adjuvants", either as polypeptides or being encoded by nucleic acids contained within the compositions of the present invention (Parmiani, et al. Immunol Lett 2000 Sep. 15; 74(1): 41-4; Berzofsky, et al. Nature Immunol. 1: 209-219). Suitable cytokines include, for example, interleukin-2 (IL-2) (Rosenberg, et al. Nature Med. 4: 321-327 (1998)), IL-4, IL-7, IL-12 (reviewed by Pardoll, 1992; Harries, et al. J. Gene Med. 2000 July-August; 2(4):243-9; Rao, et al. J. Immunol. 156: 3357-3365 (1996)), IL-15 (Xin, et al. Vaccine, 17:858-866, 1999), IL-16 (Cruikshank, et al. J. Leuk Biol. 67(6): 757-66, 2000), IL-18 (J. Cancer Res. Clin. Oncol. 2001. 127(12): 718-726), GM-CSF (CSF (Disis, et al. Blood, 88: 202-210 (1996)), tumor necrosis factor-alpha (TNF- $\alpha$ ), or interferons such as IFN- $\alpha$  or INF- $\gamma$ . Other cytokines may also be suitable for practicing the present invention, as is known in the art.

Chemokines may also be utilized. For example, fusion proteins comprising CXCL10 (IP-10) and CCL7 (MCP-3) fused to a tumor self-antigen have been shown to induce anti-tumor immunity (Biragyn, et al. Nature Biotech. 1999, 17: 253-258). The chemokines CCL3 (MIP- $1\alpha$ ) and CCL5 (RANTES) (Boyer, et al. Vaccine, 1999, 17 (Supp. 2): S53-S64) may also be of use in practicing the present invention. Other suitable chemokines are known in the art.

It is also known in the art that suppressive or negative 1999, 397: 263-265; Peach, et al. J Exp Med 1994, 180: 55 regulatory immune mechanisms may be blocked, resulting in enhanced immune responses. For instance, treatment with anti-CTLA-4 (Shrikant, et al. Immunity, 1996, 14: 145-155; Sutmuller, et al. J. Exp. Med., 2001, 194: 823-832), anti-CD25 (Sutmuller, supra), anti-CD4 (Matsui, et al. J. Immunol., 1999, 163: 184-193), the fusion protein IL13Ra2-Fc (Terabe, et al. Nature Immunol., 2000, 1: 515-520), and combinations thereof (i.e., anti-CTLA-4 and anti-CD25, Sutmuller, supra) have been shown to upregulate anti-tumor immune responses and would be suitable in practicing the present invention.

Any of these components may be used alone or in combination with other agents. For instance, it has been shown that

a combination of CD80, ICAM-1 and LFA-3 ("TRICOM") may potentiate anti-cancer immune responses (Hodge, et al. Cancer Res. 59: 5800-5807 (1999). Other effective combinations include, for example, IL-12+GM-CSF (Ahlers, et al. J. Immunol., 158: 3947-3958 (1997); Iwasaki, et al. J. Immunol. 5 158: 4591-4601 (1997)), IL-12+GM-CSF+TNF-α (Ahlers, et al. Int. Immunol. 13: 897-908 (2001)), CD80+IL-12 (Fruend, et al. Int. J. Cancer, 85: 508-517 (2000); Rao, et al. supra), and CD86+GM-CSF+IL-12 (Iwasaki, supra). One of skill in the art would be aware of additional combinations 10 useful in carrying out the present invention. In addition, the skilled artisan would be aware of additional reagents or methods that may be used to modulate such mechanisms. These reagents and methods, as well as others known by those of skill in the art, may be utilized in practicing the present 15 invention.

Additional strategies for improving the efficiency of nucleic acid-based immunization may also be used including, for example, the use of self-replicating viral replicons (Caley, et al. 1999. Vaccine, 17: 3124-2135; Dubensky, et al. 2000. 20 Mol. Med. 6: 723-732; Leitner, et al. 2000. Cancer Res. 60: 51-55), codon optimization (Liu, et al. 2000. Mol. Ther., 1: 497-500; Dubensky, supra; Huang, et al. 2001. J. Virol. 75: 4947-4951), in vivo electroporation (Widera, et al. 2000. J. Immunol. 164: 4635-3640), incorporation of CpG stimula- 25 tory motifs (Gurunathan, et al. Ann. Rev. Immunol., 2000, 18: 927-974; Leitner, supra; Cho, et al. J. Immunol. 168(10): 4907-13), sequences for targeting of the endocytic or ubiquitin-processing pathways (Thomson, et al. 1998. J. Virol. 72: 2246-2252; Velders, et al. 2001. J. Immunol. 166: 5366- 30 5373), Marek's disease virus type 1 VP22 sequences (J. Virol. 76(6):2676-82, 2002), prime-boost regimens (Gurunathan, supra; Sullivan, et al. 2000. Nature, 408: 605-609; Hanke, et al. 1998. Vaccine, 16: 439-445; Amara, et al. 2001. Science, 292: 69-74), and the use of mucosal delivery vectors such as 35 Salmonella (Darji, et al. 1997. Cell, 91: 765-775; Woo, et al. 2001. Vaccine, 19: 2945-2954). Other methods are known in the art, some of which are described below.

Chemotherapeutic agents, radiation, anti-angiogenic compreventing cancer using immunogenic targets (Sebti, et al. Oncogene 2000 Dec. 27; 19(56):6566-73). For example, in treating metastatic breast cancer, useful chemotherapeutic agents include cyclophosphamide, doxorubicin, paclitaxel, docetaxel, navelbine, capecitabine, and mitomycin C, among 45 others. Combination chemotherapeutic regimens have also proven effective including cyclophosphamide+methotrexate+5-fluorouracil; cyclophosphamide+doxorubicin+5-fluorouracil; or, cyclophosphamide+doxorubicin, for example. Other compounds such as prednisone, a taxane, navelbine, 50 mitomycin C, or vinblastine have been utilized for various reasons. A majority of breast cancer patients have estrogenreceptor positive (ER+) tumors and in these patients, endocrine therapy (i.e., tamoxifen) is preferred over chemotherapy. For such patients, tamoxifen or, as a second line 55 therapy, progestins (medroxyprogesterone acetate or megestrol acetate) are preferred. Aromatase inhibitors (i.e., aminoglutethimide and analogs thereof such as letrozole) decrease the availability of estrogen needed to maintain tumor growth and may be used as second or third line endo- 60 crine therapy in certain patients.

Other cancers may require different chemotherapeutic regimens. For example, metastatic colorectal cancer is typically treated with Camptosar (irinotecan or CPT-11), 5-fluorouracil or leucovorin, alone or in combination with one 65 another. Proteinase and integrin inhibitors such as as the MMP inhibitors marimastate (British Biotech), COL-3 (Col14

lagenex), Neovastat (Aeterna), AG3340 (Agouron), BMS-275291 (Bristol Myers Squibb), CGS 27023A (Novartis) or the integrin inhibitors Vitaxin (Medimmune), or MED1522 (Merck KgaA) may also be suitable for use. As such, immunological targeting of immunogenic targets associated with colorectal cancer could be performed in combination with a treatment using those chemotherapeutic agents. Similarly, chemotherapeutic agents used to treat other types of cancers are well-known in the art and may be combined with the immunogenic targets described herein.

Many anti-angiogenic agents are known in the art and would be suitable for co-administration with the immunogenic target vaccines (see, for example, Timar, et al. 2001. Pathology Oncol. Res., 7(2): 85-94). Such agents include, for example, physiological agents such as growth factors (i.e., ANG-2, NK1,2,4 (HGF), transforming growth factor beta (TGF- $\beta$ )), cytokines (i.e., interferons such as IFN- $\alpha$ , - $\beta$ , - $\gamma$ , platelet factor 4 (PF-4), PR-39), proteases (i.e., cleaved AT-III, collagen XVIII fragment (Endostatin)), HmwKallikreind5 plasmin fragment (Angiostatin), prothrombin-F1-2, TSPprotease inhibitors (i.e., tissue inhibitor metalloproteases such as TIMP-1, -2, or -3; maspin; plasminogen activator-inhibitors such as PAI-1; pigment epithelium derived factor (PEDF)), Tumstatin (available through ILEX, Inc.), antibody products (i.e., the collagen-binding antibodies HUIV26, HUI77, XL313; anti-VEGF; anti-integrin (i.e., Vitaxin, (Lxsys))), and glycosidases (i.e., heparinase-I, -III). "Chemical" or modified physiological agents known or believed to have anti-angiogenic potential include, for example, vinblastine, taxol, ketoconazole, thalidomide, dolestatin, combrestatin A, rapamycin (Guba, et al. 2002, Nature Med., 8: 128-135), CEP-7055 (available from Cephalon, Inc.), flavone acetic acid, Bay 12-9566 (Bayer Corp.), AG3340 (Agouron, Inc.), CGS 27023A (Novartis), tetracylcine derivatives (i.e., COL-3 (Collagenix, Inc.)), Neovastat (Aeterna), BMS-275291 (Bristol-Myers Squibb), low dose 5-FU, low dose methotrexate (MTX), irsofladine, radicicol, pounds, or other agents may also be utilized in treating and/or 40 cyclosporine, captopril, celecoxib, D45152-sulphated polysaccharide, cationic protein (Protamine), cationic peptide-VEGF, Suramin (polysulphonated napthyl urea), compounds that interfere with the function or production of VEGF (i.e., SU5416 or SU6668 (Sugen), PTK787/ZK22584 (Novartis)), Distamycin A, Angiozyme (ribozyme), isoflavinoids, staurosporine derivatives, genistein, EMD121974 (Merck KcgaA), tyrphostins, isoquinolones, retinoic acid, carboxyamidotriazole, TNP-470, octreotide, 2-methoxyestradiol, aminosterols (i.e., squalamine), glutathione analogues (i.e., N-acteyl-L-cysteine), combretastatin A-4 (Oxigene), Eph receptor blocking agents (Nature, 414:933-938, 2001), Rh-Angiostatin, Rh-Endostatin (WO 01/93897), cyclic-RGD peptide, accutin-disintegrin, benzodiazepenes, humanized anti-avb3 Ab, Rh-PAI-2, amiloride, p-amidobenzamidine, anti-uPA ab, anti-uPAR Ab, L-phanylalanin-Nmethylamides (i.e., Batimistat, Marimastat), AG3340, and minocycline. Many other suitable agents are known in the art and would suffice in practicing the present invention.

The present invention may also be utilized in combination with "non-traditional" methods of treating cancer. For example, it has recently been demonstrated that administration of certain anaerobic bacteria may assist in slowing tumor growth. In one study, Clostridium novyi was modified to eliminate a toxin gene carried on a phage episome and administered to mice with colorectal tumors (Dang, et al. P.N.A.S. USA, 98(26): 15155-15160, 2001). In combination with chemotherapy, the treatment was shown to cause tumor necrosis

in the animals. The reagents and methodologies described in this application may be combined with such treatment methodologies.

Nucleic acids encoding immunogenic targets may be administered to patients by any of several available tech- 5 niques. Various viral vectors that have been successfully utilized for introducing a nucleic acid to a host include retrovirus, adenovirus, adeno-associated virus (AAV), herpes virus, and poxvirus, among others. It is understood in the art that many such viral vectors are available in the art. The vectors of 10 the present invention may be constructed using standard recombinant techniques widely available to one skilled in the art. Such techniques may be found in common molecular biology references such as Molecular Cloning: A Laboratory Manual (Sambrook, et al., 1989, Cold Spring Harbor Labo- 15 ratory Press), Gene Expression Technology (Methods in Enzymology, Vol. 185, edited by D. Goeddel, 1991. Academic Press, San Diego, Calif.), and PCR Protocols: A Guide to Methods and Applications (Innis, et al. 1990. Academic Press, San Diego, Calif.).

Preferred retroviral vectors are derivatives of lentivirus as well as derivatives of murine or avian retroviruses. Examples of suitable retroviral vectors include, for example, Moloney murine leukemia virus (MoMuLV), Harvey murine sarcoma virus (HaMuSV), murine mammary tumor virus (MuMTV), 25 SIV, BIV, HIV and Rous Sarcoma Virus (RSV). A number of retroviral vectors can incorporate multiple exogenous nucleic acid sequences. As recombinant retroviruses are defective, they require assistance in order to produce infectious vector particles. This assistance can be provided by, for example, 30 helper cell lines encoding retrovirus structural genes. Suitable helper cell lines include  $\Psi$ 2, PA317 and PA12, among others. The vector virions produced using such cell lines may then be used to infect a tissue cell line, such as NIH 3T3 cells, to produce large quantities of chimeric retroviral virions. Ret- 35 roviral vectors may be administered by traditional methods (i.e., injection) or by implantation of a "producer cell line" in proximity to the target cell population (Culver, K., et al., 1994, Hum. Gene Ther., 5 (3): 343-79; Culver, K., et al., Cold Spring Harb. Symp. Quant. Biol., 59: 685-90); Oldfield, E., 40 1993, Hum. Gene Ther., 4 (1): 39-69). The producer cell line is engineered to produce a viral vector and releases viral particles in the vicinity of the target cell. A portion of the released viral particles contact the target cells and infect those cells, thus delivering a nucleic acid of the present invention to 45 the target cell. Following infection of the target cell, expression of the nucleic acid of the vector occurs.

Adenoviral vectors have proven especially useful for gene transfer into eukaryotic cells (Rosenfeld, M., et al., 1991, Science, 252 (5004): 431-4; Crystal, R., et al., 1994, Nat. 50 Genet., 8 (1): 42-51), the study eukaryotic gene expression (Levrero, M., et al., 1991, Gene, 101 (2): 195-202), vaccine development (Graham, F. and Prevec, L., 1992, Biotechnology, 20: 363-90), and in animal models (Stratford-Perricaudet, L., et al., 1992, Bone Marrow Transplant., 9 (Suppl. 1): 55 151-2; Rich, D., et al., 1993, Hum. Gene Ther., 4 (4): 461-76). Experimental routes for administrating recombinant Ad to different tissues in vivo have included intratracheal instillation (Rosenfeld, M., et al., 1992, Cell, 68 (1): 143-55) injection into muscle (Quantin, B., et al., 1992, Proc. Natl. Acad. 60 Sci. U.S.A., 89 (7): 2581-4), peripheral intravenous injection (Herz, J., and Gerard, R., 1993, Proc. Natl. Acad. Sci. U.S.A. 90 (7): 2812-6) and stereotactic inoculation to brain (Le Gal La Salle, G., et al., 1993, Science, 259 (5097): 988-90), among others.

Adeno-associated virus (AAV) demonstrates high-level infectivity, broad host range and specificity in integrating into

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the host cell genome (Hermonat, P., et al., 1984, *Proc. Natl. Acad. Sci. U.S.A.*, 81 (20): 6466-70). And Herpes Simplex Virus type-1 (HSV-1) is yet another attractive vector system, especially for use in the nervous system because of its neurotropic property (Geller, A., et al., 1991, *Trends Neurosci.*, 14 (10): 428-32; Glorioso, et al., 1995, *Mol. Biotechnol.*, 4 (1): 87-99; Glorioso, et al., 1995, *Annu. Rev. Microbial.*, 49: 675-710).

Poxvirus is another useful expression vector (Smith, et al. 1983, *Gene*, 25 (1): 21-8; Moss, et al, 1992, *Biotechnology*, 20: 345-62; Moss, et al, 1992, *Curr. Top. Microbiol. Immunol.*, 158: 25-38; Moss, et al. 1991. *Science*, 252: 1662-1667). Poxviruses shown to be useful include vaccinia, NYVAC, avipox, fowlpox, canarypox, ALVAC, and ALVAC(2), among others.

NYVAC (vP866) was derived from the Copenhagen vaccine strain of vaccinia virus by deleting six nonessential regions of the genome encoding known or potential virulence factors (see, for example, U.S. Pat. Nos. 5,364,773 and 5,494, 807). The deletion loci were also engineered as recipient loci for the insertion of foreign genes. The deleted regions are: thymidine kinase gene (TK; J2R); hemorrhagic region (u; B13R+B14R); A type inclusion body region (ATI; A26L); hemagglutinin gene (HA; A56R); host range gene region (C7L-K1L); and, large subunit, ribonucleotide reductase (14L). NYVAC is a genetically engineered vaccinia virus strain that was generated by the specific deletion of eighteen open reading frames encoding gene products associated with virulence and host range. NYVAC has been show to be useful for expressing TAs (see, for example, U.S. Pat. No. 6,265, 189). NYVAC (vP866), vP994, vCP205, vCP1433, placZH6H4Lreverse, pMPC6H6K3E3 and pC3H6FHVB were also deposited with the American Type Culture Collection (ATCC), P.O. Box 1549, 10801 University Boulevard, Manassas, Va. 20110-2209, USA under the terms of the Budapest Treaty, as accession numbers VR-2559, VR-2558, VR-2557, VR-2556, ATCC-97913, ATCC-97912, and ATCC-97914, respectively, all deposits made on Mar. 6,

ALVAC-based recombinant viruses (i.e., ALVAC-1 and ALVAC-2) are also suitable for use in practicing the present invention (see, for example, U.S. Pat. No. 5,756,103). ALVAC(2) is identical to ALVAC(1) except that ALVAC(2) genome comprises the vaccinia E3L and K3L genes under the control of vaccinia promoters (U.S. Pat. No. 6,130,066; Beattie et al., 1995a, 1995b, 1991; Chang et al., 1992; Davies et al., 1993). Both ALVAC(1) and ALVAC(2) have been demonstrated to be useful in expressing foreign DNA sequences, such as TAs (Tartaglia et al., 1993 a,b; U.S. Pat. No. 5,833, 975). ALVAC was deposited under the terms of the Budapest Treaty with the American Type Culture Collection (ATCC), P.O. Box 1549, 10801 University Boulevard, Manassas, Va. 20110-2209, USA, as ATCC accession number VR-2547, on Nov. 14, 1996.

Another useful poxvirus vector is TROVAC. TROVAC refers to an attenuated fowlpox that was a plaque-cloned isolate derived from the FP-1 vaccine strain of fowlpoxvirus which is licensed for vaccination of 1 day old chicks. TROVAC was likewise deposited under the terms of the Budapest Treaty with the American Type Culture Collection (ATCC), P.O. Box 1549, 10801 University Boulevard, Manassas, Va. 20110-2209, USA, as accession number 2553, on Feb. 6, 1997

"Non-viral" plasmid vectors may also be suitable in practicing the present invention. Preferred plasmid vectors are compatible with bacterial, insect, and/or mammalian host cells. Such vectors include, for example, PCR-II, pCR3, and pcDNA3.1 (Invitrogen, San Diego, Calif.), pBSII (Stratagene, La Jolla, Calif.), pET15 (Novagen, Madison, Wis.), pGEX (Pharmacia Biotech, Piscataway, N.J.), pEGFP-N2

(Clontech, Palo Alto, Calif.), pETL (BlueBacII, Invitrogen), pDSR-alpha (PCT pub. No. WO 90/14363) and pFast-BacDual (Gibco-BRL, Grand Island, N.Y.) as well as Bluescript® plasmid derivatives (a high copy number COLE1based phagemid, Stratagene Cloning Systems, La Jolla, Calif.), PCR cloning plasmids designed for cloning Taq-amplified PCR products (e.g., TOPOTM TA cloning® kit, PCR2.1® plasmid derivatives, Invitrogen, Carlsbad, Calif.). Bacterial vectors may also be used with the current invention. These vectors include, for example, Shigella, Salmonella, Vibrio cholerae, Lactobacillus, Bacille calmette guérin (BCG), and Streptococcus (see for example, WO 88/6626; WO 90/0594; WO 91/13157; WO 92/1796; and WO 92/21376). Many other non-viral plasmid expression vectors 15 and systems are known in the art and could be used with the current invention.

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ids or other lipids may also be used. The physical characteristics of liposomes depend on pH, ionic strength, and the presence of divalent cations. Examples of lipids useful in liposome production include phosphatidyl compounds, such as phosphatidylglycerol, phosphatidylcholine, phosphatidylserine, phosphatidylethanolamine, sphingolipids, cerebrosides, and gangliosides. Particularly useful are diacylphosphatidylglycerols, where the lipid moiety contains from 14-18 carbon atoms, particularly from 16-18 carbon atoms, and is saturated. Illustrative phospholipids include egg phosphatidylcholine, dipalmitoylphosphatidylcholine and distearoylphosphatidylcholine.

An immunogenic target may also be administered in combination with one or more adjuvants to boost the immune response. Exemplary adjuvants are shown in Table II below:

#### TABLE II

	Types of Immunologi	c Adjuvants
Type of Adjuvant	General Examples	Specific Examples/References
Gel-type	Aluminum hydroxide/phosphate ("alum adjuvants")	(Aggerbeck and Heron, 1995)
	Calcium phosphate	(Relyveld, 1986)
Microbial	Muramyl dipeptide (MDP)	(Chedid et al., 1986)
	Bacterial exotoxins	Cholera toxin (CT), E. coli labile toxin (LT)(Freytag and Clements, 1999)
	Endotoxin-based adjuvants	Monophosphoryl lipid A (MPL) (Ulrich and Myers, 1995)
	Other bacterial	CpG oligonucleotides (Corral and Petray, 2000), BCG sequences (Krieg, et al. Nature, 374: 576), tetanus toxoid (Rice, et al. J. Immunol., 2001, 167: 1558-1565)
Particulate	Biodegradable Polymer microspheres	(Gupta et al., 1998)
	Immunostimulatory complexes (ISCOMs)	(Morein and Bengtsson, 1999)
	Liposomes	(Wassef et al., 1994)
Oil-emulsion	Freund's incomplete adjuvant	(Jensen et al., 1998)
and	Microfluidized emulsions	MF59 (Ott et al., 1995)
surfactant-		SAF (Allison and Byars, 1992)
based		(Allison, 1999)
adjuvants	Saponins	QS-21 (Kensil, 1996)
Synthetic	Muramyl peptide derivatives	Murabutide (Lederer, 1986)
	, I I	Threony-MDP (Allison, 1997)
	Nonionic block copolymers	L121 (Allison, 1999)
	Polyphosphazene (PCPP)	(Payne et al., 1995)
	Synthetic polynucleotides	Poly A: U, Poly I: C (Johnson, 1994)
	Thalidomide derivatives	CC-4047/ACTIMID (J. Immunol., 168(10): 4914-9)

Suitable nucleic acid delivery techniques include DNAligand complexes, adenovirus-ligand-DNA complexes, direct injection of DNA, CaPO<sub>4</sub> precipitation, gene gun techniques, electroporation, and colloidal dispersion systems, among others. Colloidal dispersion systems include macro- 55 molecule complexes, nanocapsules, microspheres, beads, and lipid-based systems including oil-in-water emulsions, micelles, mixed micelles, and liposomes. The preferred colloidal system of this invention is a liposome, which are artificial membrane vesicles useful as delivery vehicles in vitro 60 and in vivo. RNA, DNA and intact virions can be encapsulated within the aqueous interior and be delivered to cells in a biologically active form (Fraley, R., el al., 1981, Trends Biochem. Sci., 6: 77). The composition of the liposome is usually a combination of phospholipids, particularly high-phase- 65 transition-temperature phospholipids, usually in combination with steroids, especially cholesterol. Other phospholip-

The immunogenic targets of the present invention may also be used to generate antibodies for use in screening assays or for immunotherapy. Other uses would be apparent to one of skill in the art. The term "antibody" includes antibody fragments, as are known in the art, including Fab, Fab<sub>2</sub>, single chain antibodies (Fv for example), humanized antibodies, chimeric antibodies, human antibodies, produced by several methods as are known in the art. Methods of preparing and utilizing various types of antibodies are well-known to those of skill in the art and would be suitable in practicing the present invention (see, for example, Harlow, et al. Antibodies: A Laboratory Manual, Cold Spring Harbor Laboratory, 1988; Harlow, et al. Using Antibodies: A Laboratory Manual, Portable Protocol No. 1, 1998; Kohler and Milstein, Nature, 256:495 (1975)); Jones et al. Nature, 321:522-525 (1986); Riechmann et al. Nature, 332:323-329 (1988); Presta (Curr. Op. Struct. Biol., 2:593-596 (1992); Verhoeyen et al. (Sci-

ence, 239:1534-1536 (1988); Hoogenboom et al., J. Mol. Biol., 227:381 (1991); Marks et al., J. Mol. Biol., 222:581 (1991); Cole et al., Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, p. 77 (1985); Boerner et al., J. Immunol., 147(1):86-95 (1991); Marks et al., Bio/Technology 10, 779-783 (1992); Lonberg et al., Nature 368 856-859 (1994); Morrison, Nature 368 812-13 (1994); Fishwild et al., Nature Biotechnology 14, 845-51 (1996); Neuberger, Nature Biotechnology 14, 826 (1996); Lonberg and Huszar, Intern. Rev. Immunol. 13 65-93 (1995); as well as U.S. Pat. Nos. 4,816, 567; 5,545,807; 5,545,806; 5,569,825; 5,625,126; 5,633,425; and, 5,661,016). The antibodies or derivatives therefrom may also be conjugated to therapeutic moieties such as cytotoxic drugs or toxins, or active fragments thereof such as diptheria A chain, exotoxin A chain, ricin A chain, abrin A chain, curcin, crotin, phenomycin, enomycin, among others. Cytotoxic agents may also include radiochemicals. Antibodies and their derivatives may be incorporated into compositions of the invention for use in vitro or in vivo.

Nucleic acids, proteins, or derivatives thereof representing an immunogenic target may be used in assays to determine the presence of a disease state in a patient, to predict prognosis, or to determine the effectiveness of a chemotherapeutic or other treatment regimen. Expression profiles, performed as is 25 known in the art, may be used to determine the relative level of expression of the immunogenic target. The level of expression may then be correlated with base levels to determine whether a particular disease is present within the patient, the patient's prognosis, or whether a particular treatment regimen 30 is effective. For example, if the patient is being treated with a particular chemotherapeutic regimen, an decreased level of expression of an immunogenic target in the patient's tissues (i.e., in peripheral blood) may indicate the regimen is decreasing the cancer load in that host. Similarly, if the level of 35 expression is increasing, another therapeutic modality may need to be utilized. In one embodiment, nucleic acid probes corresponding to a nucleic acid encoding an immunogenic target may be attached to a biochip, as is known in the art, for the detection and quantification of expression in the host.

It is also possible to use nucleic acids, proteins, derivatives therefrom, or antibodies thereto as reagents in drug screening assays. The reagents may be used to ascertain the effect of a drug candidate on the expression of the immunogenic target profiling technique may be combined with high throughput screening techniques to allow rapid identification of useful compounds and monitor the effectiveness of treatment with a drug candidate (see, for example, Zlokarnik, et al., Science 279, 84-8 (1998)). Drug candidates may be chemical com- 50 pounds, nucleic acids, proteins, antibodies, or derivatives therefrom, whether naturally occurring or synthetically derived. Drug candidates thus identified may be utilized, among other uses, as pharmaceutical compositions for administration to patients or for use in further screening 55 assays.

Administration of a composition of the present invention to a host may be accomplished using any of a variety of techniques known to those of skill in the art. The composition(s) may be processed in accordance with conventional methods 60 of pharmacy to produce medicinal agents for administration to patients, including humans and other mammals (i.e., a "pharmaceutical composition"). The pharmaceutical composition is preferably made in the form of a dosage unit containing a given amount of DNA, viral vector particles, 65 polypeptide or peptide, for example. A suitable daily dose for a human or other mammal may vary widely depending on the

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condition of the patient and other factors, but, once again, can be determined using routine methods.

The pharmaceutical composition may be administered orally, parentally, by inhalation spray, rectally, intranodally, or topically in dosage unit formulations containing conventional pharmaceutically acceptable carriers, adjuvants, and vehicles. The term "pharmaceutically acceptable carrier" or "physiologically acceptable carrier" as used herein refers to one or more formulation materials suitable for accomplishing or enhancing the delivery of a nucleic acid, polypeptide, or peptide as a pharmaceutical composition. A "pharmaceutical composition" is a composition comprising a therapeutically effective amount of a nucleic acid or polypeptide. The terms "effective amount" and "therapeutically effective amount" each refer to the amount of a nucleic acid or polypeptide used to induce or enhance an effective immune response. It is preferred that compositions of the present invention provide for the induction or enhancement of an anti-tumor immune response in a host which protects the host from the develop-20 ment of a tumor and/or allows the host to eliminate an existing tumor from the body.

For oral administration, the pharmaceutical composition may be of any of several forms including, for example, a capsule, a tablet, a suspension, or liquid, among others. Liquids may be administered by injection as a composition with suitable carriers including saline, dextrose, or water. The term parenteral as used herein includes subcutaneous, intravenous, intramuscular, intrasternal, infusion, or intraperitoneal administration. Suppositories for rectal administration of the drug can be prepared by mixing the drug with a suitable non-irritating excipient such as cocoa butter and polyethylene glycols that are solid at ordinary temperatures but liquid at the rectal temperature.

The dosage regimen for immunizing a host or otherwise treating a disorder or a disease with a composition of this invention is based on a variety of factors, including the type of disease, the age, weight, sex, medical condition of the patient, the severity of the condition, the route of administration, and the particular compound employed. For example, a poxviral vector may be administered as a composition comprising  $1\times10^{\circ}$  infectious particles per dose. Thus, the dosage regimen may vary widely, but can be determined routinely using standard methods.

A prime-boost regimen may also be utilized (WO in a cell line, or a cell or tissue of a patient. The expression 45 01/30382 A1) in which the targeted immunogen is initially administered in a priming step in one form followed by a boosting step in which the targeted immunogen is administered in another form. The form of the targeted immunogen in the priming and boosting steps are different. For instance, if the priming step utilized a nucleic acid, the boost may be administered as a peptide. Similarly, where a priming step utilized one type of recombinant virus (i.e., ALVAC), the boost step may utilize another type of virus (i.e., NYVAC). This prime-boost method of administration has been shown to induce strong immunological responses.

While the compositions of the invention can be administered as the sole active pharmaceutical agent, they can also be used in combination with one or more other compositions or agents (i.e., other immunogenic targets, co-stimulatory molecules, adjuvants). When administered as a combination, the individual components can be formulated as separate compositions administered at the same time or different times, or the components can be combined as a single composition.

Injectable preparations, such as sterile injectable aqueous or oleaginous suspensions, may be formulated according to known methods using suitable dispersing or wetting agents and suspending agents. The injectable preparation may also

be a sterile injectable solution or suspension in a non-toxic parenterally acceptable diluent or solvent. Suitable vehicles and solvents that may be employed are water, Ringer's solution, and isotonic sodium chloride solution, among others. For instance, a viral vector such as a poxvirus may be prepared in 0.4% NaCl. In addition, sterile, fixed Oils are conventionally employed as a solvent or suspending medium. For this purpose, any bland fixed oil may be employed, including synthetic mono- or diglycerides. In addition, fatty acids such as oleic acid find use in the preparation of injectables.

For topical administration, a suitable topical dose of a composition may be administered one to four, and preferably two or three times daily. The dose may also be administered with intervening days during which no does is applied. Suitable compositions may comprise from 0.001% to 10% w/w, for example, from 1% to 2% by weight of the formulation, although it may comprise as much as 10% w/w, but preferably not more than 5% w/w, and more preferably from 0.1% to 1% of the formulation. Formulations suitable for topical administration include liquid or semi-liquid preparations suitable for penetration through the skin (e.g., liniments, lotions, ointments, creams, or pastes) and drops suitable for administration to the eye, ear, or nose.

The pharmaceutical compositions may also be prepared in a solid form (including granules, powders or suppositories). 25 The pharmaceutical compositions may be subjected to conventional pharmaceutical operations such as sterilization and/ or may contain conventional adjuvants, such as preservatives, stabilizers, wetting agents, emulsifiers, buffers etc. Solid dosage forms for oral administration may include capsules, tab- 30 lets, pills, powders, and granules. In such solid dosage forms, the active compound may be admixed with at least one inert diluent such as sucrose, lactose, or starch. Such dosage forms may also comprise, as in normal practice, additional substances other than inert diluents, e.g., lubricating agents such 35 as magnesium stearate. In the case of capsules, tablets, and pills, the dosage forms may also comprise buffering agents. Tablets and pills can additionally be prepared with enteric coatings. Liquid dosage forms for oral administration may include pharmaceutically acceptable emulsions, solutions, 40 suspensions, syrups, and elixirs containing inert diluents commonly used in the art, such as water. Such compositions may also comprise adjuvants, such as wetting sweetening, flavoring, and perfuming agents.

Pharmaceutical compositions comprising a nucleic acid or polypeptide of the present invention may take any of several forms and may be administered by any of several routes. In preferred embodiments, the compositions are administered via a parenteral route (intradermal, intramuscular or subcutaneous) to induce an immune response in the host. Alternatively, the composition may be administered directly into a lymph node (intranodal) or tumor mass (i.e., intratumoral administration). For example, the dose could be administered subcutaneously at days 0, 7, and 14. Suitable methods for

immunization using compositions comprising TAs are known in the art, as shown for p53 (Hollstein et al., 1991), p21-ras (Almoguera et al., 1988), HER-2 (Fendly et al., 1990), the melanoma-associated antigens (MAGE-1; MAGE-2) (van der Bruggen et al., 1991), p97 (Hu et al., 1988), melanoma-associated antigen E (WO 99/30737) and carcinoembryonic antigen (CEA) (Kantor et al., 1993; Fishbein et al., 1992; Kaufman et al., 1991), among others.

Preferred embodiments of administratable compositions include, for example, nucleic acids or polypeptides in liquid preparations such as suspensions, syrups, or elixirs. Preferred injectable preparations include, for example, nucleic acids or polypeptides suitable for parental, subcutaneous, intradermal, intramuscular or intravenous administration such as sterile suspensions or emulsions. For example, a recombinant poxvirus may be in admixture with a suitable carrier, diluent, or excipient such as sterile water, physiological saline, glucose or the like. The composition may also be provided in lyophilized form for reconstituting, for instance, in isotonic aqueous, saline buffer. In addition, the compositions can be co-administered or sequentially administered with other antineoplastic, anti-tumor or anti-cancer agents and/or with agents which reduce or alleviate ill effects of antineoplastic, anti-tumor or anti-cancer agents.

A kit comprising a composition of the present invention is also provided. The kit can include a separate container containing a suitable carrier, diluent or excipient. The kit can also include an additional anti-cancer, anti-tumor or antineoplastic agent and/or an agent that reduces or alleviates ill effects of antineoplastic, anti-tumor or anti-cancer agents for co-or sequential-administration. Additionally, the kit can include instructions for mixing or combining ingredients and/or administration.

A better understanding of the present invention and of its many advantages will be had from the following examples, given by way of illustration.

#### **EXAMPLES**

#### Example 1

#### BFA4 Tumor Antigen

The BFA4 sequence was found to be the "trichorhinophalangeal syndrome 1" (TRPS-1) gene (Genebank ID #6684533; Momeniet et al, Nature Genetics, 24(1), 71-74, 2000), a known transcription factor with no function attributed previously in any form of cancer. The BFA4 cDNA sequence is shown in FIG. 1 and the deduced amino acid sequence is shown in FIG. 2.

<sup>0</sup> A. BFA4 Peptides and Polyclonal Antisera

For monitoring purposes, rabbit anti-BFA4 polyclonal antibodies were generated. Six peptides (22-mers) were designed and synthesized to elicit antibody response to BFA4, as shown below:

```
      CLP
      2589 MVRKKNPPLRNVASEGEGQILE
      BFA4
      (1-22)
      (SEQ ID NO.: 5)

      CLP
      2590 SPKATEETGQAQSGQANCQGLS
      BFA4
      (157-178)
      (SEQ ID NO.: 6)

      CLP
      2591 VAKPSEKNSNKSIPALQSSDSG
      BFA4
      (371-392)
      (SEQ ID NO.: 7)

      CLP
      2592 NHLQGSDGQQSVKESKEHSCTK
      BFA4
      (649-670)
      (SEQ ID NO.: 9)

      CLP
      2593 NGEQIIRRRTRKRLNPEALQAE
      BFA4
      (940-961)
      (SEQ ID NO.: 5)

      CLP
      2594 ANGASKEKTKAPPNVKNEGPLNVBFA4
      (1178-1199)
      (SEQ ID NO.: 10)
```

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Rabbits were immunized with the peptides, serum was isolated, and the following antibody titers were observed:

Rabbit #	Peptide	Titer (Bleed 2)	Titer (Final Bleed)
1, 2	CLP2589	800000, 1600000	2560000, 2560000
3, 4	CLP2590	12800, 6400	40000, 40000
5, 6	CLP2591	400000, 400000	320000, 320000
7, 8	CLP2592	25600, 12800	80000, 40000
9, 10	CLP2593	3200000, 51200	2560000, 160000
11, 12	CLP2594	409600, 409600	320000, 320000

These peptides were also modified by coupling with KLH peptides to enhance immune responses as shown below:

BFA4	(1-22)	KLH-MVRKKNPPLRNVASEGEGQILE (CLP-2589; SEQ ID NO.: 5)
BFA4	(157-178)	KLH-SPKATEETGQAQSGQANCQGLS (CLP-2590; SEQ ID NO.: 6)
BFA4	(371-392)	KLH-VAKPSEKNSNKSIPALQSSDSG (CLP-2591; SEQ ID NO.: 7)
BFA4	(649-670)	KLH-NHLQGSDGQQSVKESKEHSCTK (CLP-2592; SEQ ID NO.: 8)
BFA4	(940-961)	KLH-NGEQIIRRRTRKRLNPEALQAE (CLP-2593; SEQ ID NO.: 9)
BFA4	(1178-1200)	KLH-ANGASKEKTKAPPNVKNEGPLNV (CLP-2594; SEQ ID NO.: 10)

The pcDNA3.2BFA4 (3.6 mg) was also used for DNA immunization to generate polyclonal sera in chickens. B. Cloning of BFA4

Complete cDNA sequence for BFA4 is ~10kb and gene is expressed in BT474 ductal carcinoma cells. Primers 7717 (forward primer) and 7723 (reverse primer) were designed to amplify full-length BFA4 gene by amplification of 4kb, 7kb or 10kb products by RT-PCR.

Ten mg of total RNA isolated and frozen in different 50 batches from BT-474 cells using Trizol as indicated by the manufacturer (Gibco BRL) was used in RT-PCR to amplify the BFA4 gene. RT-PCR conditions were optimized using Taq Platinum High Fidelity enzyme, OPC (Oligo Purification Cartridge; Applied Biosystems) purified primers and purified 55 total RNA/polyA mRNA (BT 474 cells). Optimization resulted in a 4.0 kb fragment as a single band.

To re-amplify the BFA4 sequence, mRNA was treated with DNase per manufacturers' instructions (Gibco BRL). The 4 kb DNA was reamplified using PCR using primers 7717 and 60 7723 primers (10 pmole/microliter) and Taq Platinum High Fidelity polymerase (GIBCO BRL) enzyme. Thermocycler conditions for both sets of reactions were as under: 94° C. (2 min), followed by 30 cycles of 94° C. (30 sec), 52° C. (30 sec), 67° C. (4 min) and 67° C. (5 min) and finally 40° C. for 10 65 min. Three BFA4 clones were identified after pCR2.1/TOPO-TA cloning.

Several mutations were identified during analysis of the BFA4 sequence. To correct these sequences, the BamHI/XhoI fragment (5') of the BFA4 gene from clone JB-3552-1-2 (pCR2.1/TOPO/BFA4) was exchanged with the XhoI/BamHI fragment (3') of the BFA4 gene from clone JB-3552-1-4 (pCR2.1/TOPO/BFA4). This recombined fragment was then ligated into pMCS5 BamHI/CAP. Clone JB-3624-1-5 was generated and found to contain the correct sequence.

Nucleotide 344 of the isolated BFA4 clone was different 10 from the reported sequence (C in BFA4, T in TRPS-1). The change resulted in a phe to ser amino acid change. To change this sequence to the reported sequence, the EcoRI/BgIII fragment (5') of the BFA4 gene from clone JB-3552-1-2 (pCR2.1/ TOPO/BFA4) was subcloned into pUC8:2 to generate clone JB-3631-2. This clone was used as a template for Quickchange (Stratagene) mutagenesis to change amino acid 115 of the BFA4 protein from a serine to a phenylalanine as in the TRPS1 protein. The selected clone was JB-3648-2-3. Mutagenesis was also repeated with pMCS5 BFA4 (BT474) 20 as a template for Ouickchange (Stratagene) mutagenesis to change amino acid 115 of the BFA4 protein from a serine to a phenylalanine as in the TRPS1 protein. Several clones were found to be correct by DNA sequencing and one of the clones (JB-3685-1-18) was used for further subcloning.

JB-3685-1-18 was then used to subclone the BFA4 coding sequence into the BamHI sites of four different expression vectors: 1) the poxviral (NYVAC) vector pSD554VC (CO-PAK/H6; JB-3707-1-7); 2) pcDNA3.1/Zeo (+) (JB-3707-3-2); 3) pCAMycHis (JB-3707-5-1); and, 4) Semiliki Forest virus alphaviral replicon vector pMP76 (JB-3735-1-23). The BFA4 coding sequence within JB-3707-1-7, JB-3707-5-1, and JB-3735-1-23 was confirmed by DNA sequencing.

A stop codon was introduced near the end of the cloned sequence in the pcDNA3.1/Zeo/BFA4 construct (JB-3707-3-35 2). A unique EcoR1 site was opened and filled in to introduce a stop codon in-frame with BFA4 coding sequence. Several putative clones were identified by the loss of EcoR1 site, however three clones (JB-3756-1-2; JB-3756-3-1; and JB-3756-4-1) were sequenced. All three were found to be correct in the area of the fill-in. Clone JB-3756-3-1 identified as having the correct sequence and orientation.

Myc and myc/his tags (Evans et al, 1985) were introduced using oligonucleotides, which were annealed and ligated into the pcDNA3.1/Zeo/BFA4 construct (JB-3707-3-2) at the EcoRI/EcoRV sites. Several clones were obtained for these constructs. Three clones having the correct sequences and orientations were obtained: 1) PcDNA3.1/Zeo/BFA4/mychis-tag (JB-3773-1-2); 2) PcDNA3.1/Zeo/BFA4/mychis-tag (JB-3773-2-1); and, 3) PcDNA3.1/Zeo/BFA4/mychis-tag (JB-3773-2-2).

#### C. Expression of BFA4

#### 1. Expression from Poxviral Vectors

The pSD554VC (COPAK/H6; JB-3707-1-7) vector was used to generate NYVAC-BFA4 virus. In vitro recombination was performed with plasmid COPAK/H6/BFA4 and NYVAC in RK13/CEF cells. NYVAC-BFA4 (vP2033-NYVAC-RK13) was generated and amplified to P3 level after completion of three enrichments with final stock concentrations of 1.12×10<sup>9</sup>/ml (10 ml). Vero cells were infected with NYVAC-BFA4 at an M.O.I. of 0.5 pfu/cell. Lysates and media were harvested 24 h post-infection to confirm expression of BFA4 protein. One-twentieth of the concentrated media and 1/40 of the lysate were loaded onto a western blot and incubated with rabbit antisera against the BFA4 peptides CLP 2589, 2591, 2598 and 2594 (see above for peptide sequences and preparation of anti-BFA4 antisera). An approximate 120 kD band was detected in both the lysate and the concentrated media of

NYVAC-BFA4-infected Vero cells which was not evident in either Vero control cells ("mock-infected"), Vero cells infected with the parental NYVAC virus, or concentrated media

#### 2. Expression from pcDNA3.1-based Vectors

Transient transfection studies were performed to verify expression of BFA4 from the pcDNA-based vectors and to analyze quality of polyclonal sera raised against BFA4 peptides. The following constructs were used to study expression of BFA4 gene: pcDNA 3.1 zeo<sup>R</sup>/BFA4, pMP76/BFA4, pcDNA 3.1 zeo<sup>R</sup>/BFA4/Myc tag and pcDNA 3.1 zeo<sup>R</sup>/BFA4/ MycHis tag. BFA4 expression plasmids (5 μg and 10 μg) were co-transfected with pGL3 Luciferase (1 g) (Promega) with the Gene porter reagent (Gene Therapy Systems) as the transfection reagent. At 48 h post-transfection, whole cell extract was prepared by scraping cells in cell lysis reagent (200 µl) and 1 cycle of freeze-thaw (-20° C. freeze, 37° C. thaw). Transfection efficiency was quantitated by analyzing expression of the luciferase reporter gene by measuring Relative 20 Luciferase Units (RLU) in duplicate. Similar RLU values were obtained in the samples co-transfected with luciferase construct in the presence and absence of BFA4 expression vectors. There was no significant difference observed in toxicity or RLU values with differential amount (5 µg and 10 µg) 25 of BFA4 expression vectors. Preliminary western blot analysis using alkaline phosphatase system with the CHOK1 cell extracts (pCDNA3.1/zeo/BFA4/MycHisTag) and an anti-BFA4 polyclonal antisera, revealed a band at approximately 120 kDa band in extracts of BFA4 vector-transfected cells.

A stable transfection study was initiated to obtain stable clones of BFA4 expressing COS A2 cells. These cells are useful for in vitro stimulation assays. pcDNA 3.1 zeo<sup>R</sup>/BFA4 (2.5 μg and 20 μg), and pcDNA 3.1 zeo<sup>R</sup>/BFA4/MycHis tag (2.5 µg) were used to study expression of BFA4). pGL3 35 Luciferase (2.5 µg) was used as a control vector to monitor transfection efficiency. The Gene porter reagent was used to facilitate transfection of DNA vectors. After 48 h post-transfection, whole cell extract were prepared by scraping cells in the cell lysis reagent (200 µl) and 1 cycle of freeze-thaw at 40 -20° C./37° C. for first experiment. Transfected cells obtained from the second experiment were trypsinized, frozen stock established and cells were plated in increasing concentrations of Zeocin (0, 250, 500, 750 and 1000 µg/ml). Non-transfected CosA2cells survived at 60-80% confluency 45 for three weeks at 100 µg/ml (Zeocin) and 10% confluency at 250 ug/ml (Zeocin). However, after three weeks, at higher drug concentration (500-1000 µg/ml), live cells were not observed in the plates containing non-transfected cells and high Zeocin concentration (500-1000 μg/ml).

Several Zeocin-resistant clones growing in differential drug concentrations (Zeocin-250, 500, 750 and 1000 µg/ml) were picked from 10 cm plates after three weeks. These clones were further expanded in a 3.5 cm plate(s) in the presence of Zeocin at 500, 750 and 1000 µg/ml. Frozen lots of 55 these clones were prepared and several clones from each pool (pcDNA 3.1 zeo<sup>R</sup>/BFA4, and pcDNA 3.1 zeo<sup>R</sup>/BFA4/MycHis tag) were expanded to T75 cm<sup>2</sup> flasks in the presence of Zeocin at 1 mg/ml. Five clones from each pool (pcDNA 3.1 zeo<sup>R</sup>/BFA4, and pcDNA 3.1 zeo<sup>R</sup>/BFA4/MycHis tag) were 60 expanded to T75 cm<sup>2</sup> flasks in the presence of Zeocin at 1 mg/ml. Cells are maintained under Zeocin drug (1 mg/ml) selection. Six clones were used in BFA4 peptide-pulsed target experiment, and two clones were found to express BFA4 at a moderate level by immunological assays. The non-adherent 65 cell lines K562A2 and EL4A2 were also transfected with these vectors to generate stable cell lines.

#### 3. Prokaryotic Expression Vector

The BamHI-Xho-1 fragment (1.5 Kbp) fragment encoding N-terminal 54 kDa BFDA4 to from pCDNA3.1/BFA4 was cloned into pGEX4T1-6His (Veritas) plasmid. This vector contains the tac promoter followed by the N-terminal glutathione S-transferase (GST~26kDa) and a hexahistidine tag to C terminus of the GST fusion protein.

The BFA4-N54 expression plasmid was transformed into BL21 cells and grown at 25° C. in antibiotic selection medium (2 L culture) to an OD (600 nm) and thereafter induced with 1 mM IPTG. GST-BFA4-N54 was found to be soluble protein. Clarified extract of the soluble fraction was adsorbed batchwise to glutathione-Sepharose 4B and eluted with 10 mM reduced glutathione. Fractions were analyzed after estimation of protein concentration and TCA precipitation. Specific polypeptide of Mr=85 kDa in the eluate was confirmed by SDS-PAGE. The recombinant protein was purified by gluathione-Sepharose was absorbed on a NiNTA column for further purification. The bound protein was eluted with 0.25M imidazole. The protein was dialyzed versus TBS containing 40% Glycerol, resulting in 4.5 mg GST-BFA4-N54-6 His (N terminus BFA4 protein) protein. Expression of BFA4 was confirmed using the rabbit anti-BFA4 polyclonal antibody by western blot.

#### D. Therapeutic Anti-BFA4 Immune Responses

#### 1. RFA4 Peptides

In addition to genetic immunization vectors for BFA4, immunological reagents for BFA4 have been generated. A library of 100 nonamer peptides spanning the BFA4 gene product was synthesized. The peptides were chosen based on their potential ability to bind to HLA-A\*0201.

TABLE V
lists 100 nonamer peptide epitopes for

HLA-A\*0201 from the BFA4 protein tested

	(see below):	
SEQUENCE	POSITION IN PROTEIN	SEQ ID.
MVRKKNPPL	BFA4 (1-9)	13
KKNPPLRNV	BFA4 (4-12)	14
VASEGEGQI	BFA4 (12-20)	15
QILEPIGTE	BFA4 (19-27)	16
RNMLAFSFP	BFA4 (108-116)	17
NMLAFSFPA	BFA4 (109-117)	18
MLAFSFPAA	BFA4 (110-118)	19
FSFPAAGGV	BFA4 (113-121)	20
AAGGVCEPL	BFA4 (117-125)	21
SGQANCQGL	BFA4 (170-178)	22
ANCQGLSPV	BFA4 (172-180)	23
GLSPVSVAS	BFA4 (176-184)	24
SVASKNPQV	BFA4 (181-189)	25
RLNKSKTDL	BFA4 (196-204)	26
NDNPDPAPL	BFA4 (207-215)	27
DPAPLSPEL	BFA4 (211-219)	28
ELQDFKCNI	BFA4 (218-216)	29

28
TABLE V-continued

	TABLE V-continued				TABLE V-continued	
	100 nonamer peptide epito 201 from the BFA4 protei (see below):	-	— <sub>5</sub>		.00 nonamer peptide epit 201 from the BFA4 protei (see below):	
EQUENCE	POSITION IN PROTEIN	SEQ ID.	3	SEQUENCE	POSITION IN PROTEIN	SEQ ID.
LHNRTRQD	BFA4 (249-257)	30		STIKEEPKI	BFA4 (734-742)	66
LDSKILAL	BFA4 (259-267)	31	10	KIDFRVYNL	BFA4 (741-749)	67
ILALHNMV	BFA4 (263-271)	32		NLLTPDSKM	BFA4 (748-756)	68
				VTWRGADIL	BFA4 (792-800)	69
LHNMVQFS	BFA4 (266-284)	33	15	ILRGSPSYT	BFA4 (799-807)	70
NRSVFSGV	BFA4 (282-290)	34		YTQASLGLL	BFA4 (806-814)	71
SGVLQDIN	BFA4 (287-295)	35		ASLGLLTPV	BFA4 (809-817)	72
INSSRPVL	BFA4 (293-301)	36	20	GLLTPVSGT	BFA4 (812-820)	73
LLNGTYDV	BFA4 (300-308)	37	20	GTQEQTKTL	BFA4 (819-827)	74
CNFTYMGN	BFA4 (337-345)	38		KTLRDSPNV	BFA4 (825-833)	75
MGNSSTEL	BFA4 (342-350)	39		HLARPIYGL	BFA4 (837-845)	76
LQTHPNKI	BFA4 (354-362)	40	25	PIYGLAVET	BFA4 (841-849)	77
ASLPSSEV	BFA4 (363-371)	41		LAVETKGFL	BFA4 (845-853)	78
LGKWQDKI	BFA4 (393-401)	42		FLQGAPAGG	BFA4 (852-860)	79
KAGDDTPV	BFA4 (403-411)	43	30			
SCESSSSL	BFA4 (441-449)	44		AGGEKSGAL	BFA4 (858-866)	80
LEHYGKQ	BFA4 (450-458)	45		GALPQQYPA	BFA4 (864-872)	81
NPELNDK	BFA4 (466-474)	46	35	ALPQQYPAS	BFA4 (865-873)	82
SVINQNDL	BFA4 (478-486)	47		FCANCLTTK	BFA4 (895-903)	83
/INQNDLA	BFA4 (479-487)	48		ANGGYVCNA	BFA4 (911-919)	84
CDFRYSKS	BFA4 (527-535)	49		NACGLYQKL	BFA4 (918-926)	85
HGPDVIVV	BFA4 (535-543)	50	40	GLYQKLHST	BFA4 (921-929)	86
LLRHYQQL	BFA4 (545-553)	51		KLHSTPRPL	BFA4 (925-933)	87
LCSPEKHL	BFA4 (570-578)	52		STPRPLNII	BFA4 (928-936)	88
LGEITYPF	BFA4 (577-585)	53	45	RLNPEALQA	BFA4 (952-960)	89
				VLVSQTLDI	BFA4 (1020-1028)	90
GEITYPFA	BFA4 (578-586)	54		DIHKRMQPL	BFA4 (1027-1035)	91
CALLLHL	BFA4 (594-602)	55	50	RMQPLHIQI	BFA4 (1031-1039)	92
LLLLHLSP	BFA4 (596-604)	56		YPLFGLPFV	BFA4 (1092-1100)	93
LLLHLSPG	BFA4 (597-605)	57		GLPFVHNDF	BFA4 (1096-1104)	94
LLHLSPGA	BFA4 (598-606)	58	55	FVHNDFQSE	BFA4 (1099-1107)	95
HLSFGAA	BFA4 (599-607)	59	33	SVPGNPHYL	BFA4 (1120-1128)	96
TPDVDVL	BFA4 (621-629)	60		GNPHYLSHV	BFA4 (1123-1131)	97
PDVDVLL	BFA4 (622-630)	61		HYLSHVPGL	BFA4 (1126-1134)	98
LLFHYESV	BFA4 (628-636)	62	60	YVPYPTFNL	BFA4 (1141-1149)	99
ITQVEEEI	BFA4 (673-681)	63				
TAADTQSL	BFA4 (699-707)	64		FNLPPHFSA	BFA4 (1147-1155)	100
LEHFNTV	BFA4 (706-714)	65	65	NLPPHFSAV	BFA4 (1148-1156)	101
				SAVGSDNDI	BFA4 (1154-1162)	102

KLLEHYGKQ

GLNPELNDK

GSVINQNDL

SVINQNDLA

FCDFRYSKS

45

46

47

48

49

TABLE V-continued

	100 nonamer peptide epitor 0201 from the BFA4 protein (see below):			PEPTIDE GROUP	SEQUENCE	SEQ ID
	POSITION	SEQ	5		VNRSVFSGV	34
SEQUENCE	IN PROTEIN	ID.			FSGVLQDIN	35
KNEGPLNVV	BFA4 (1192-1200)	103			DINSSRPVL	36
TKCVHCGIV	BFA4 (1215-1223)	104	10		VLLNGTYDV	37
CVHCGIVFL	BFA4 (1217-1225)	105			FCNFTYMGN	38
CGIVFLDEV	BFA4 (1220-1228)	106			YMGNSSTEL	39
FLDEVMYAL	BFA4 (1224-1232)	107	15		FLQTHPNKI	40
VMYALHMSC	BFA4 (1228-1236)	108			KASLPSSEV	41
FQCSICQHL	BFA4 (1243-1251)	109			DLGKWQDKI	42
GLHRNNAQV	BFA4 (1265-1273)	110	20	4	VKAGDDTPV	43
	. ,		_		FSCESSSSL	44

The peptide library was pooled into separate groups containing 7-10 different peptides for immunological testing as shown in Table VI (see below). In addition to a peptide library spanning BFA4, a recombinant protein spanning the N-terminal 300 amino acids (positions 1-300) has been synthesized and purified from  $E.\ coli.$ 

PEPTIDE GROUP	SEQUENCE	SEQ ID	50		SHGPDVIVV	50	
1	MVRKKNPPL	13			PLLRHYQQL	51	
-	KKNPPLRNV	14			GLCSPEKHL	52	
			35	5	HLGEITYPF	53	
	VASEGEGQI	15			LGEITYPFA	54	
	QILEPIGTE	16			HCALLLLHL	55	
	RNMLAFSFP	17	40		ALLLLHLSP	56	
	NMLAFSFPA	18					
	MLAFSFPAA	19			LLLLHLSPG	57	
	FSFPAAGGV	20	45		LLLHLSPGA	58	
	AAGGVCEPL	21	43		LLHLSPGAA	59	
	SGQANCQGL	22			FTTPDVDVL	60	
2	ANCOGLSPV	23			TTPDVDVLL	61	
2	GLSPVSVAS	24	50		VLLFHYESV	62	
				6	FITQVEEEI	63	
	SVASKNPQV	25			FTAADTQSL	64	
	RLNKSKTDL	26	55		SLLEHFNTV	65	
	NDNPDPAPL	27	55			66	
	DPAPLSPEL	28			STIKEEPKI		
	ELQDFKCNI	29			KIDFRVYNL	67	
	GLHNRTRQD	30	60		NLLTPDSKM	68	
	ELDSKILAL	31			KMGEPVSES	220	
	KILALHNMV	32			GLKEKVWTE	221	
3	ALHNMVQFS	33	65		VTWRGADIL	69	
	~						

PEPTIDE SEQUENCE ID  ILRGSPSYT 70  7 YTQASLGLL 71	5	PEPTIDE GROUP	SEQUENCE VMYALHMSC	SEQ ID 108
7 YTQASLGLL 71			VMYALHMSC	108
~	10			
a of or i mari	10		FQCSICQHL	109
ASLGLLTPV 72	10		GLHRNNAQV	110
GLLTPVSGT 73				1.5
GTQEQTKTL 74		2. Immune Reactivity of I Human Effector T Cells:	BFA4 Peptid	es and Generation of
KTLRDSPNV 75		The BFA4 peptides we		
HLARPIYGL 76		7-10 peptides for immuno pools were pulsed onto at		
PIYGLAVET 77		cells and used to activate a	autologous T	-cell-enriched PBMC
LAVETKGFL 78		preparations. Activated 3 stimulated culture were re		
FLQGAPAGG 79	20	using CD40L-activated au	tologous B-c	ells. IFN-γ ELISPOT
AGGEKSGAL 80	20	analysis and assays for Cl cells was performed to de		
8 GALPQQYPA 81		these epitopes from BFA4		
ALPQOYPAS 82		Human T cells demonstr number of pools of peptide		
- FCANCLTTK 83	25	by their ability to secrete	IFN-γ in EL	ISPOT assays. These
ANGGYVCNA 84		experiments were repeated stimulation resulting in the		
NACGLYQKL 85		Peptide groups 1, 2, 4, 5, 6 immunoreactive in these as		
GLYQKLHST 86	30	peptide groups were de-	-convoluted	in additional IFN-γ
KLHSTPRPL 87		ELISPOT assays in which were tested separately. The		
		peptide groups 1, 5 6, 7, 8,	9, and 10 in E	ELISPOT assays. This
STPRPLNII 88	35	analysis revealed a numb peptides from the BFA4 pr		
RLNPEALQA 89		It was also observed that i	nany of these	e single peptides also
VLVSQTLDI 90		induced CTL activity killi phoma cell targets. These		
9 DIHKRMQPL 91	40			
RMQPLHIQI 92			BLE VII	
YPLFGLPFV 93		List of highly i	lmmunoreacti rom BFA4	ive peptides
GLPFVHNDF 94	45	Strong IFN-γ	SEQ ID	Strong CTL
FVHNDFQSE 95	43	Killing	NO.	Killing
SVPGNPHYL 96		RNMLAFSFP	17	RNMLAFSFP
GNPHYLSHV 97		NMLAFSFPA	18	NMLAFSFPA
HYLSHVPGL 98	50	MLAFSFPAA	19	MLAFSFPAA
YVPYPTFNL 99		HLGEITYPF	53	
FNLPPHFSA 100		FTTPDVDVL	60	FTTPDVDVL
10 NLPPHFSAV 101	55	VLLFHYESV	62	VLLFHYYESV
SAVGSDNDI 102		KIDFRVYNL	66	
KNEGPLNVV 103		ASLGLLTPV	72	ASLGLLTPV
TKCVHCGIV 104	60	HLARPIYGL	76	HLARPIYGL
CVHCGIVFL 105		NACGLYQKL	85	NACGLYQKL
CGIVFLDEV 106		KLHSTPRPL	87	
FLDEVMYAL 107	65	RLNPEALQA	89	RLNPEALQA
22000000		YPLFGLPFV	93	

9617SXC:

sequencing the clones:

TABLE VII-continued

List of highly	immunoreacti from BFA4	ive peptides
Strong IFN-γ Killing	SEQ ID NO.	Strong CTL Killing
YVPYPTFNL	99	YVPYPTFNL
NLPPHFSAV	101	
VMYALHMSC	108	
GLHRNNAQV	110	GLHRNNAQV

D. Immune Responses against BFA4 after Immunization in vivo:

The pcDNA3.1/Zeo-BFA4 plasmid was used to immunize transgenic mice expressing a hybrid HLA-A\*0201  $\alpha$ 1 $\alpha$ 2 domain fused to a murine Kb  $\alpha$ 3 domain in C57BL/6 mice (A2-Kb mice). IFN- $\gamma$  ELISPOT analysis using the groups of pooled peptides after DNA immunization and removal of activated spleen cells revealed a number of reactive BFA4 peptide groups. Some of these groups (especially group 7 and 8) also reacted strongly in human T-cell cultures suggesting 25 that overlapping groups of peptides are recognized by human T cells and are naturally processed and presented on HLA-A2 after vaccination.

Vaccination experiments were also performed with the NYVAC-BFA4 and the MP76-18-BFA4 vectors in A2-Kb mice. Mice were immunized subcutaneously with 10-29 μg of MP-76-18-BFA4 and 1-2×10<sup>7</sup> pfu vP2033 (NYVAC-BFA4) and boosted 28 days later with the same amounts of each vector. Re-stimulation of spleen cells from the immunized mice with the pools of BFA4 peptides revealed induction of IFN-γ production in response to BFA4 peptide groups 2, 3, 4, 5, 7, 9, and 10 in ELISPOT assays. Thus, the BFA4 gene encoded in a CMV promoter driven eukaryotic plasmid, NYVAC, or a Semliki replicase-based DNA plasmid, were all capable of inducing T-cell responses against the BFA4 protein in vivo.

Example 2

#### BCY1 Tumor Antigen

The BCY1 gene was detected as a partial open reading frame (ORF) homologous to a to nematode gene called "posterior-expressed maternal gene-3" (PEM-3) playing a role in posterior to anterior patterning in *Caenorhabtidis elegans* embryos. No previous involvement of this gene in cancer has been documented.

#### A. BCY1 and Amino Acid DNA Sequences

A partial DNA sequence was originally determined for BCY1. Primers, 9616SXC and 9617SXC, are derived from the BCY I partial DNA sequence and are designed to clone BCY I by RT-PCR from Calu 6 total RNA. The primers were designed such that the PCR product has BamHI sites at both ends and an ATG start codon and a Kozak sequence at the 5' 60 end, as shown below:

9616SXC: (SEQ ID NO.: 113)

5' CAGTACGGATCCACCATGGCCGAGCTGCGCCTGAAGGGC 3'

34

-continued

(SEQ ID NO.: 114) 5' CCACGAGGATCCTTAGGAGAATATTCGGATGGCTTGCG 3'

The 1.2 Kb expected amplicon was obtained using ThermoScript RT-PCR System (Invitrogen) under optimized conditions. The PCR products from three separate RT-PCR's were digested with BamHI and respectively inserted in pcDNA3.1/Zeo(+). The resulting clones were MC50A6, MC50A8 and MC50A19 from the first RT-PCR; MC54.21 from the second RT-PCR and MC55.29; and, MC55.32 from the third RT-PCR. The following primers were utilized in

9620MC: 5	5'	(SEQ ID NO.: 115)
9621MC: 5	5 '	(SEQ ID NO.: 116)
9618MC: 5	5 '	(SEQ ID NO.: 117)
9619MC: 5	5'	(SEQ ID NO.: 118) GCTCACCCAGGCGTGGGGCCTC 3'

DNA sequencing of all six clones indicated a consensus sequence, as shown in FIGS. 3A-C, having the following differences from the original partial BCY1 sequence: a C to G substitution at position 1031 resulting in an amino acid change of Ala to Gly; a GC deletion at position 1032-1034 resulting in a Thr deletion; and, an A to G substitution at position 1177 resulting in an amino acid change of Thr to Ala. Clones MC50A8 and MC55.29 are identical to the consensus sequence. The amino acid sequence of BCY1 is shown in FIG. 3D.

B. Immunological Reagents for BCY1 Breast Cancer Antigen:

A library of 100 nonamer peptides spanning the BCY1 gene product was synthesized. The peptides were chosen based on their potential ability to bind to HLA-A\*0201. Table VIII lists 100 nonamer peptide epitopes for HLA-A\*0201 from the BCY1 protein tested (see below):

TABLE VIII

_				
	Sequence	Position in Protein	SEQ ID	
•	VPVPTSEHV	2	119	
	PTSEHVAEI	5	120	
	EIVGRQCKI	12	121	
	KIKALRAKT	19	122	
	KALRAKTNT	21	123	
	ALRAKTNTY	22	124	
	LRAKTNTYI	23	125	
	TNTYIKTPV	27	126	
	YIKTPVRGE	30	127	
	TPVRGEEPV	33	128	
	RGEEPVFMV	36	129	
	MVTGRREDV	43	130	

36
TABLE VIII-continued

	17.	EDDE VIII CONCIN	iucu		11	ADDD VIII CONCI	IIaca
Ī	Sequence	Position in Protein	SEQ ID		Sequence	Position in Protein	SEQ ID
-	VTGRREDVA	44	131	5	T 1700117 3 1 170 00	140	150
	GRREDVATA	46	132		IETHIAVRT	149	170
	DVATARREI	50	133		HIAVRTGKI	152	171
	VATARREII	51	134	10	IAVRTGKIL	153	172
	TARREIISA	53	135		KILEYNNEN	159	173
	ARREIISAA	54	136		YNNENDFLA	163	174
	IISAAEHFS	58	137	15	NENDFLAGS	165	175
	ISAAEHFSM	59	138		FLAGSPDAA	169	176
	SAAEHFSMI	60	139		LAGSPDAAI	170	177
	AEHFSMIRA	62	140	20	AIDSRYSDA	177	178
	SMIRASRNK	66	141	20	SRYSDAWRV	180	179
	RASRNKSGA	69	142		VHQPGCKPL	188	180
	NKSGAAFGV	73	143		LSTFRQNSL	196	181
	GAAFGVAPA	76	144	25	LGCIGECGV	204	182
	AAFGVAPAL	77	145		CGVDSGFEA	210	183
	GVAPALPGQ	80	146		GFEAPRLDV	215	184
	VAPALPGQV	81	147	30	RLDVYYGVA	220	185
	PALPGQVTI	83	148		DVYYGVAET	222	186
	ALPGQVTIR	84	149		GVAETSPPL	226	187
	LPGQVTIRV	85	150	35	AETSPPLWA	228	188
	GQVTIRVRV	87	151		PLWAGQENA	233	189
	RVRVPYRVV	92	152		AGQENATPT	236	190
	RVPYRVVGL	94	153	40	QENATPTSV	238	191
	VPYRVVGLV	95	154		VLFSSASSS	246	192
	RVVGLVVGP	98	155		KARAGPPGA	260	193
	GLVVGPKGA	101	156	45	PATSAGPEL	272	194
	LVVGPKGAT	102	157		ATSAGPELA	273	195
	VVGPKGATI	103	158		SAGPELAGL	275	196
	RIQQQTNTY	113	159	50	GLPRRPPGE	282	197
	IQQQTNTYI	114	160		EPLQGFSKL	290	198
	QQQTNTYII	115	161		FSKLGGGGL	295	199
	QQTNTYIIT	116	162	55	KLGGGGLRS	297	200
	YIITPSRDR	121	163	55	GLRSPGGGR	302	201
	TPSRDRDPV	124	164		CMVCFESEV	312	202
	RDRDPVFEI	127	165		MVCFESEVT	313	203
	EITGAPGNV	134	166	60	VCFESEVTA	314	204
	GAPGNVERA	137	167		FESEVTAAL	316	205
	NVERAREEI	141	168		EVTAALVPC	319	206
	EEIETHIAV	147	169	65	VTAALVPCG	320	207

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TABLE IX-continued

TAI	BLE VIII-contin	nued		TA	ABLE IX-continu	ed
Sequence	Position in Protein	SEQ ID			groups of peptides	
ALVPCGHNL	323	208	5	Peptide Group	Peptide Sequence	SEQ ID
LVPCGHNLF	324	209		3	GRREDVATA	132
VPCGHNLFC	325	210			DVATARREI	133
NLFCMECAV	330	211	10		TARREIISA	135
FCMECAVRI	332	212			GVAPALPGQ	146
CAVRICERT	336	213			RVVGLVVGP	155
RICERTDPE	339	214	15		VHQPGCKPL	180
RTDPECPVC	343	215			PATSAGPEL	194
					VTAALVPCG	207
CPVCHITAT	348	216	20	4	VHVPTSEHV	119
VCHITATQA	350	217			ARREIISAA	136
ITATQAIRI	353	218			RIQQQTNTY	159
			25		NVERAREEI	168
	TABLE IX				GFEAPRLDV	184
	groups of peptide				ATSAGPELA	195
	mmunological testir		30		FSKLGGGGL	199
Peptide Group	Peptide Sequence	SEQ ID			GLRSPGGGR	201
1	EPLQGFSKL	198		5	PTSEHVAEI	120
	EVTAALVPC	206	35		EIVGRQCKI	121
	CPVCHITAT	216			LRAKTNTYI	125
	KIKALRAKT	122			VTGRREDVA	131
	IISAAEHFS	137	40		SMIRASRNK	141
	RASRNKSGA	142			CMVCFESEV	202
	GAAFGVAPA	144			LVPCGHNLF	209
	LVVGPKGAT	157	45		NLFCMECAV	211
	EITGAPGNV	166			RICERTDPE	214
	GAPGNVERA	167			RTDPECPVC	215
2	ALRAKTNTY	124	50	6	MVTGRREDV	130
	VATARREII	134	30		GLVVGPKGA	156
	PALPGQVTI	148			IQQQTNTYI	160
	ALPGQVTIR	149	55		FLAGSPDAA	176
	RVRVPYRVV	152	55		GVAETSPPL	187
	RDRDPVFEI	165			FESEVTAAL	205
	RVRVPYRVV	152			FCMECAVRI	212
	HIAVRTGKI	171	60	7	KALRAKTNT	123
	NENDFLAGS	175			RGEEPVFMV	129
	CAVRICERT	213			SAAEHFSMI	139
	VCHITATQA	217	65		AAFGVAPAL	145

40

TA	ABLE IX-continue	ed		TA	ABLE IX-continu	ed
	groups of peptides				groups of peptides	
Peptide Group	Peptide Sequence	SEQ ID	5	Peptide Group	Peptide Sequence	SEQ ID
	VVGPKGATI	158			TPSRDRDPV	164
	YNNENDFLA	174			IAVRTGKIL	172
	LGCIGECGV	182	10		SRYSDAWRV	179
	QENATPTSV	191			LSTFRQNSL	181
	VCFESEVTA	204			RLDVYYGVA	185
8	TNTYIKTPV	126	15		AGQENATPT	190
	NKSGAAFGV	143			MVCFESEVT	203
	QQTNTYIIT	162		Immuna Dagatir	vitre of DCV1 Domtidae	and Conquetion of
	KILEYNNEN	173	<sup>20</sup> Ні	ıman Effector T		
	CGVDSGFEA	183			O peptides from BCY1 peptides for immunol	
	AETSPPLWA	188	so	lved peptide poo	ols were pulsed onto	autologous HLA-
	PLWAGQENA	189	25 en	riched PBMC pr	ells and used to activat reparations.  Activated	T cells from each
	VLFSSASSS	192			lated culture were re-s D40L-activated autolo	
	SAGPELAGL	196	EI	LISPOT analysis	and assays for CTL	killing of peptide-
9	ISAAEHFSM	138	30 no	genicity of these	was performed to dem epitopes from BCY1	•
	QQQTNTYII	161			emonstrated effector copeptides from the BCY	
	EEIETHIAV	169	by	their ability to s	secrete IFN-γ in ELIS repeated after differe	POT assays. These
	IETHIAVRT	170	35 sti	mulation resultii	ng in the same reacti	ve peptide groups.
	LAGSPDAAI	177			2, 3, 4, 5, 6, and 7 these assays. Subsequ	

immunoreactive in these assays. Subsequently, these reactive peptide groups were de-convoluted in additional IFN-y ELISPOT assays in which single peptides from each group were tested separately. This analysis revealed a number of individual strongly reactive peptides from the BCY1 protein recognized by human T cells. Many of these single peptides also induced CTL activity killing peptide-loaded human T2 lymphoma cell targets. Table IX lists these peptides.

While the present invention has been described in terms of the preferred embodiments, it is understood that variations and modifications will occur to those skilled in the art. Therefore, it is intended that the appended claims cover all such equivalent variations that come within the scope of the invention as claimed.

SEQUENCE LISTING

178

186

218

128

140

147

AIDSRYSDA

DVYYGVAET

VPCGHNLFC

ITATQAIRI

TPVRGEEPV

AEHFSMIRA

VAPALPGQV

10

<sup>&</sup>lt;160> NUMBER OF SEQ ID NOS: 229

<sup>&</sup>lt;210> SEQ ID NO 1

<sup>&</sup>lt;211> LENGTH: 3846

<sup>&</sup>lt;212> TYPE: DNA

<sup>&</sup>lt;213> ORGANISM: Homo sapiens

<sup>&</sup>lt;400> SEQUENCE: 1

atggtccgga aaaagaaccc ccctctgaga aacgttgcaa gtgaaggcga gggccagatc 60 ctggagccta taggtacaga aagcaaggta tctggaaaga acaaagaatt ctctgcagat 120 cagatgtcag aaaatacgga tcagagtgat gctgcagaac taaatcataa ggaggaacat

		71				
				-contir	nued	
agcttgcatg	ttcaagatcc	atcttctagc	agtaagaagg	acttgaaaag	cgcagttctg	240
agtgagaagg	ctggcttcaa	ttatgaaagc	cccagtaagg	gaggaaactt	tccctccttt	300
ccgcatgatg	aggtgacaga	cagaaatatg	ttggctttct	catttccagc	tgctggggga	360
gtctgtgagc	ccttgaagtc	tccgcaaaga	gcagaggcag	atgaccctca	agatatggcc	420
tgcaccccct	caggggactc	actggagaca	aaggaagatc	agaagatgtc	accaaaggct	480
acagaggaaa	cagggcaagc	acagagtggt	caagccaatt	gtcaaggttt	gagcccagtt	540
tcagtggcct	caaaaaaccc	acaagtgcct	tcagatgggg	gtgtaagact	gaataaatcc	600
aaaactgact	tactggtgaa	tgacaaccca	gacccggcac	ctctgtctcc	agagcttcag	660
gactttaaat	gcaatatctg	tggatatggt	tactacggca	acgaccccac	agatctgatt	720
aagcacttcc	gaaagtatca	cttaggactg	cataaccgca	ccaggcaaga	tgctgagctg	780
gacagcaaaa	tcttggccct	tcataacatg	gtgcagttca	gccattccaa	agacttccag	840
aaggtcaacc	gttctgtgtt	ttctggtgtg	ctgcaggaca	tcaattcttc	aaggcctgtt	900
ttactaaatg	ggacctatga	tgtgcaggtg	acttcaggtg	gaacattcat	tggcattgga	960
cggaaaacac	cagattgcca	agggaacacc	aagtatttcc	gctgtaaatt	ctgcaatttc	1020
acttatatgg	gcaactcatc	caccgaatta	gaacaacatt	ttcttcagac	tcacccaaac	1080
aaaataaaag	cttctctccc	ctcctctgag	gttgcaaaac	cttcagagaa	aaactctaac	1140
aagtccatcc	ctgcacttca	atccagtgat	tctggagact	tgggaaaatg	gcaggacaag	1200
ataacagtca	aagcaggaga	tgacactcct	gttgggtact	cagtgcccat	aaagcccctc	1260
gattcctcta	gacaaaatgg	tacagaggcc	accagttact	actggtgtaa	attttgtagt	1320
ttcagctgtg	agtcatctag	ctcacttaaa	ctgctagaac	attatggcaa	gcagcacgga	1380
gcagtgcagt	caggeggeet	taatccagag	ttaaatgata	agctttccag	gggctctgtc	1440
attaatcaga	atgatctagc	caaaagttca	gaaggagaga	caatgaccaa	gacagacaag	1500
agctcgagtg	gggctaaaaa	gaaggacttc	tccagcaagg	gagccgagga	taatatggta	1560
acgagctata	attgtcagtt	ctgtgacttc	cgatattcca	aaagccatgg	ccctgatgta	1620
attgtagtgg	ggccacttct	ccgtcattat	caacagctcc	ataacattca	caagtgtacc	1680
attaaacact	gtccattctg	teccagagga	ctttgcagcc	cagaaaagca	ccttggagaa	1740
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Lys Gln Asp Val Tyr Tyr Gly Val Ala Glu Thr Ser Pro Pro Leu Trp
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What is claimed is:

- 1. An expression vector comprising the nucleic acid consisting of SEQ ID NO.:3.
- 2. The expression vector of claim 1 wherein the vector is a plasmid or a viral vector.
- 3. The expression vector of claim 2 wherein the viral vector is selected from the group consisting of poxvirus, alphavirus, adenovirus, retrovirus, herpesvirus, and adeno-associated  $^{30}$  virus.
- **4**. The expression vector of claim **3** wherein the poxvirus selected from the group consisting of vaccinia, NYVAC, avipox, canarypox, ALVAC, ALVAC(2), fowlpox, and TRO-VAC.
- **5**. The expression vector of claim **4** wherein the poxvirus selected from the group consisting of NYVAC, ALVAC, and ALVAC(2).
- 6. An expression vector encoding a polypeptide consisting  $_{\mbox{\scriptsize 40}}$  of SEQ ID NO.:4.
- 7. The expression vector of claim 6 wherein the vector is a plasmid or a viral vector.
- **8.** The expression vector of claim **7** wherein the viral vector is selected from the group consisting of poxvirus, alphavirus, adenovirus, retrovirus, herpesvirus, and adeno-associated virus.
- 9. The expression vector of claim 8 wherein the poxvirus selected from the group consisting of vaccinia, NYVAC, avipox, canarypox, ALVAC, ALVAC(2), fowlpox, and TRO-  $_{50}$
- 10. The expression vector of claim 9 wherein the poxvirus selected from the group consisting of NYVAC, ALVAC, and ALVAC(2).
- 11. The expression vector of claim 1 or 6 further comprising at least one nucleic acid encoding a tumor-associated antigen.

- 12. The expression vector of claim 1 or 6 further comprising at least one nucleic sequence encoding an angiogenesis-associated antigen.
- 13. The expression vector of claim 1 or 6 further comprising at least one nucleic acid sequence encoding a co-stimulatory component.
- 14. The expression vector of claim 13 wherein the costimulatory component is B7.1.
- **15**. The expression vector of claim **13** wherein the costimulatory component is TRICOM comprising B7.1, LFA-3 and ICAM-1.
- 16. The expression vector of claim 13 wherein the vector is a plasmid or a viral vector.
- 17. The expression vector of claim 16 wherein the viral vector is selected from the group consisting of poxvirus, alphavirus, adenovirus, retrovirus, herpesvirus, and adenoassociated virus.
- 18. The expression vector of claim 17 wherein the poxvirus is selected from the group consisting of vaccinia, NYVAC, avipox, canarypox, ALVAC, ALVAC(2), fowlpox, and TRO-VAC.
- 19. The expression vector of claim 18 wherein the poxvirus is selected from the group consisting of NYVAC, ALVAC, and ALVAC(2).
- **20**. A pharmaceutical composition comprising an expression vector of claim **1**.
- 21. A pharmaceutical composition comprising an expression vector of claim 6.
- 22. A pharmaceutical composition comprising an expression vector of claim 14.
- 23. A pharmaceutical composition comprising an expression vector of claim 16.
- 24. A pharmaceutical composition comprising an expression vector of claim 18.

\* \* \* \* \*