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(54) IDENTIFICATION AND CHARACTERIZATION OF MULTIPLE SPLICE VARIANTS OF THE MU-OPIOID **RECEPTOR GENE**

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- ABSTRACT (57)

The present invention encompasses novel splice variant forms of the mu-opioid receptor-1 (MOR-1) and the polynucleotide sequences encoding the MOR-1 splice variants. The invention further encompasses methods of screening for compositions regulating the MOR-1 splice variant activities and the development of therapeutic modalities directed to regulating activity. Regulation of the MOR-1 splice variant activities may impact the physiologic processes of analgesia and weight management.

MOR-IC (SEQ ID NO: 17) M D S S A G P G N I S D C S D P L A P A S C S P A P G S W L N L S H V DGNQSDPCGPNRTGLGGSHSLCPQTGSPSMVTAIT I M A L Y S I V C V V G L F G N F L V M Y V I V R Y T K M K T A T N I Y I F N L A L A D A L A T S T L P F Q S V N Y L M G T W P F G N I L C K I V I S I D Y Y N M F T S I F T L C T M S V D R Y I A V C H P V K A LDFRTPRNAKIVNVCNWILSSAIGLPVMFMATTKY RQGSIDCTLTFSHPTWYWENLLKICVFIFAFIMPV LIITVCYGLMILRLKSVRMLSGSKEKDRNLRRITR MVLVVVAVFIVCWTPIHIYVIIKALITIPETTFQT V S W H F C I A L G Y T N S C L N P V L Y A F L D E N F K R C F R E F C I P T S S T I E Q Q N S A R I R Q N T R E H P S T A N T V D R T N H Exon 7 Exon 8 PTLAVSVAQIFTGYPSPTHVEKPCKSCMDR GM 0 Exon 9

<u>RNLLPDDGPRQESGEGQLG</u>

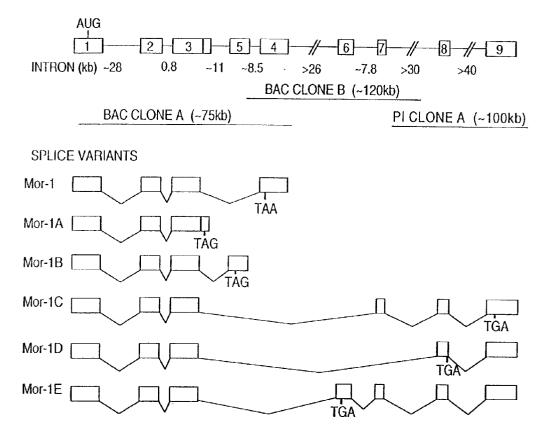


FIG. 1

MOR-1c (SEQ ID NO:1)

AGAACCATGGACAGCAGCGGCCGGCCCAGGGAACATCAGCGACTGCTCTGACCCCTTAGCT CCTGCAAGTTGCTCCCCAGCACCTGGCTCCTGGCTCAACTTGTCCCACGTTGATGGCAAC CAGTCCGACCCATGCGGTCCTAACCGCACGGGGGCTTGGCGGGAGCCACAGCCTGTGCCCT CAGACCGGCAGCCCTTCCATGGTCACAGCCATCACCATCATGGCCCTCTATTCTATCGTG ATGAAGACTGCCACCAACATCTACATTTTCAACCTTGCTCTGGCAGATGCCTTAGCCACT AGCACGCTGCCCTTTCAGAGTGTTAACTACCTGATGGGAACGTGGCCCTTTGGAAACATC CTCTGCAAGATCGTGATCTCAATAGACTACTACAACATGTTCACCAGTATCTTCACCCTC TGCACCATGAGTGTAGACCGCTACATTGCCGTCTGCCACCCGGTCAAGGCCCTGGATTTC CGTACCCCCCGAAATGCCAAAATTGTCAATGTCTGCAACTGGATCCTCTCTGCCATT CTCACGTTCTCTCATCCCACATGGTACTGGGAGAACCTGCTCAAAATCTGTGTCTTCATC TTCGCCTTCATCATGCCGGTCCTCATCATCACTGTGTGTTATGGACTGATGATCTTACGA CTCAAGAGTGTCCGCATGCTGTCGGGCTCCAAAGAAAAGGACAGGAACCTGCGCAGGATC ACCCGGATGGTGCTGGTGGTCGTGGCTGTATTTATTGTCTGCTGGACCCCCATCCACATC TATGTCATCATCAAAGCACTGATCACGATTCCAGAAACCACTTTCCAGACTGTTTCCTGG CACTTCTGCATTGCCTTGGGTTACACAAACAGCTGCCTGAACCCAGTTCTTTATGCGTTC CTGGATGAAAACTTCAAACGATGTTTTAGAGAGTTCTGCATCCCAACTTCCTCCACAATC GAACAGCAAAACTCTGCTCGAATCCGTCAAAACACTAGGGAACACCCCTCCACGGCTAAT Exon 7 ACAGTGGATCGAACTAACCACCAG/CCAACCCTGGCAGTCAGCGTGGCCCAGATCTTTACA Exon 8 <u>GGATATCCTTCTCCGACTCATGTTGAAAAACCCTGCAAGAGTTGCATGGACAG/AGGAATG</u> Exon 9 AGGAACCTTCTTCCTGATGATGGCCCCAAGACAGGAATCCGGGGAAGGCCAGCTTGGCAG/G

TGAATGTCATCCGAACACAGGGATGAGCTGGTGAGCAGTGTGG

FIG. 2A

MOR-1g (SEQ ID NO: 2) Exon la

/TTTTACTGTCCTTGAGAATGGAGAGGATCAGCAAAGCTGGAAGCCCTCCAGGCTCATTTC

AGAGAGAATATTCCACAGAGCTTGAAGGCGCGGGGATCTGGGCCGATGATGGAAGCTTTCT

TCAGATATACCAAAATGAAGACTGCCACCAACATCTACATTTTCAACCTTGCTCTGGCAG ATGCCTTAGCCACTAGCACGCTGCCCTTTCAGAGTGTTAACTACCTGATGGGAACGTGGC CCTTTGGAAACATCCTCTGCAAGATCGTGATCTCAATAGACTACTACAACATGTTCACCA GTATCTTCACCCTCTGCACCATGAGTGTAGACCGCTACATTGCCGTCTGCCACCCGGTCA AGGCCCTGGATTTCCGTACCCCCGAAATGCCAAAATTGTCAATGTCTGCAACTGGATCC CCATAGATTGCACCCTCACGTTCTCTCATCCCACATGGTACTGGGAGAACCTGCTCAAAA TCTGTGTCTTCATCTTCGCCTTCATCATGCCGGTCCTCATCATCACTGTGTGTTATGGAC TGATGATCTTACGACTCAAGAGTGTCCGCATGCTGTCGGGCTCCAAAGAAAAGGACAGGA CCCCCATCCACATCTATGTCATCATCAAAGCACTGATCACGATTCCAGAAACCACTTTCC AGACTGTTTCCTGGCACTTCTGCATTGCCTTGGGTTACACAAACAGCTGCCTGAACCCAG TTCTTTATGCGTTCCTGGATGAAAACTTCAAACGATGTTTTAGAGAGTTCTGCATCCCAA CTTCCTCCACAATCGAACAGCAAAACTCTGCTCGAATCCGTCAAAACACTAGGGAACACC CCTCCACGGCTAATACAGTGGATCGAACTAACCACCAGCTAGAAAATCTGGAAGCAGAAA CTGCTCCATTGCCCTAACTGGGTCCCACGCCATCCAGACCCTCGCTAAACTTAGAGGCTG CCATCTACTTGGAATCAGGTTGCTGTCAGGGTTTGTGGGAGGCTCTGGTTTCCTGGAAAA **GCATCTGATCCTGCATTCAAAGTCATTCTAACTGGGTC**

FIG. 2B

MOR-1d (SEQ ID NO: 3)

AGAACCATGGACAGCAGCGCCGGCCCAGGGAACATCAGCGACTGCTCTGACCCCTTAGCT CCTGCAAGTTGCTCCCCAGCACCTGGCTCCTGGCTCAACTTGTCCCACGTTGATGGCAAC CAGTCCGACCCATGCGGTCCTAACCGCACGGGGGCTTGGCGGGAGCCACAGCCTGTGCCCT CAGACCGGCAGCCCTTCCATGGTCACAGCCATCACCATCATGGCCCTCTATTCTATCGTG ATGAAGACTGCCACCAACATCTACATTTTCAACCTTGCTCTGGCAGATGCCTTAGCCACT AGCACGCTGCCCTTTCAGAGTGTTAACTACCTGATGGGAACGTGGCCCTTTGGAAACATC CTCTGCAAGATCGTGATCTCAATAGACTACTACAACATGTTCACCAGTATCTTCACCCTC TGCACCATGAGTGTAGACCGCTACATTGCCGTCTGCCACCCGGTCAAGGCCCTGGATTTC CTCACGTTCTCTCATCCCACATGGTACTGGGAGAACCTGCTCAAAATCTGTGTCTTCATC TTCGCCTTCATCATGCCGGTCCTCATCATCACTGTGTGTTATGGACTGATGATCTTACGA CTCAAGAGTGTCCGCATGCTGTCGGGCTCCAAAGAAAAGGACAGGAACCTGCGCAGGATC ACCCGGATGGTGCTGGTGGTCGTGGCTGTATTTATTGTCTGCTGGACCCCCATCCACATC TATGTCATCATCAAAGCACTGATCACGATTCCAGAAACCACTTTCCAGACTGTTTCCTGG CACTTCTGCATTGCCTTGGGTTACACAAACAGCTGCCTGAACCCAGTTCTTATGCGTTC CTGGATGAAAACTTCAAACGATGTTTTAGAGAGTTCTGCATCCCAACTTCCTCCACAATC GAACAGCAAAACTCTGCTCGAATCCGTCAAAACACTAGGGAACACCCCTCCACGGCTAAT Exon 8

ACAGTGGATCGAACTAACCACCAG/AGGAATGAGGAACCTTCTTCCTGATGATGGCCCAAG

Exon 9

ACAGGAATCCGGGGAAGGCCAGCTTGGCAG/GTGAATGTCATCCGAACACAGGGATGAGCT

GGTGAGCAGTGTGG

FIG. 2C

MOR-1e (SEQ ID NO: 4)

AGAACCATGGACAGCAGCGCCGGCCCAGGGAACATCAGCGACTGCTCTGACCCCTTAGCT CCTGCAAGTTGCTCCCCAGCACCTGGCTCCTGGCTCAACTTGTCCCACGTTGATGGCAAC CAGTCCGACCCATGCGGTCCTAACCGCACGGGGCTTGGCGGGAGCCACAGCCTGTGCCCT CAGACCGGCAGCCCTTCCATGGTCACAGCCATCACCATCATGGCCCTCTATTCTATCGTG ATGAAGACTGCCACCAACATCTACATTTTCAACCTTGCTCTGGCAGATGCCTTAGCCACT AGCACGCTGCCCTTTCAGAGTGTTAACTACCTGATGGGAACGTGGCCCTTTGGAAACATC CTCTGCAAGATCGTGATCTCAATAGACTACTACAACATGTTCACCAGTATCTTCACCCTC TGCACCATGAGTGTAGACCGCTACATTGCCGTCTGCCACCCGGTCAAGGCCCTGGATTTC CGTACCCCCGAAATGCCAAAATTGTCAATGTCTGCAACTGGATCCTCTCTGCCATT CTCACGTTCTCTCATCCCACATGGTACTGGGAGAACCTGCTCAAAATCTGTTGCTTCATC TTCGCCTTCATCATGCCGGTCCTCATCATCACTGTGTGTTATGGACTGATGATCTTACGA CTCAAGAGTGTCCGCATGCTGTCGGGCTCCAAAGAAAAGGACAGGAACCTGCGCAGGATC ACCCGGATGGTGCTGGTGGTCGTGGCTGTATTTATTGTCTGCTGGACCCCCATCCACATC TATGTCATCATCAAAGCACTGATCACGATTCCAGAAACCACTTTCCAGACTGTTTCCTGG CACTTCTGCATTGCCTTGGGTTACACAACAGCTGCCTGAACCCAGTTCTTATGCGTTC CTGGATGAAAACTTCAAACGATGTTTTAGAGAGTTCTGCATCCCAACTTCCTCCACAATC GAACAGCAAAACTCTGCTCGAATCCGTCAAAACACTAGGGAACACCCCTCCACGGCTAAT Exon 6

ACAGTGGATCGAACTAACCACCAG/AAGAAAAGCTGGACTCCCAGAGAGGGGTGTGTACAG

CATCCAGTGTGACCTGTCCCTTGTCTTTGAGCCTGGGGGGCCATCTTCTTCACAGCATAC Exon 7

ATTTCCTTGTATCCTCTGAAG/CCAACCCTGGCAGTCAGCGTGGCCCAGATCTTACA Exon 8 GGATATCCTTCTCCGACTCATGTTGAAAAACCCTGCAAGAGTTGCATGGACAG/AGGAATG Exon 9 AGGAACCTTCTTCCTGATGATGGCCCAAGAGAGGAATCCGGGGAAGGCCAGCTTGGCAG/G

TGAATGTCATCCGAACACAGGGATGAGCTGGTGAGCAGTGTGG

FIG. 2D

MOR-1h (SEQ 1D NO:5)

Exon la /TTTTACTGTCCTTGAGAATGGAGAGGA<u>TCAGCAAAGCTGGAAGCCC</u>TCCAGGCTCATTTC

AGAGAGAATATTCCACAGAGCTTGAAGGCGCGGGATCTGGGCCGATGATGGAAGCTTTCT

CTAAGTCTGCATTCCAAAAGCTCAGACAGAGAGAGATGGAAATCAAGAGGGGAAGAGCTACC

TCAGATATACCAAAATGAAGACTGCCACCAACATCTACATTTTCAACCTTGCTCTGGCAG ATGCCTTAGCCACTAGCACGCTGCCCTTTCAGAGTGTTAACTACCTGATGGGAACGTGGC CCTTTGGAAACATCCTCTGCAAGATCGTGATCTCAATAGACTACTACAACATGTTCACCA GTATCTTCACCCTCTGCACCATGAGTGTAGACCGCTACATTGCCGTCTGCCACCCGGTCA AGGCCCTGGATTTCCGTACCCCCCGAAATGCCAAAATTGTCAATGTCTGCAACTGGATCC CCATAGATTGCACCCTCACGTTCTCTCATCCCACATGGTACTGGGAGAACCTGCTCAAAA TCTGTGTCTTCATCTTCGCCTTCATCATGCCGGTCCTCATCATCACTGTGTGTTATGGAC TGATGATCTTACGACTCAAGAGTGTCCGCATGCTGTCGGGCTCCAAAGAAAAGGACAGGA CCCCCATCCACATCTATGTCATCATCAAAGCACTGATCACGATTCCAGAAACCACTTTCC AGACTGTTTCCTGGCACTTCTGCATTGCCTTGGGTTACACAAACAGCTGCCTGAACCCAG TTCTTTATGCGTTCCTGGATGAAAACTTCAAACGATGTTTTAGAGAGATTCTGCATCCCAA CTTCCTCCACAATCGAACAGCAAAACTCTGCTCGAATCCGTCAAAACACTAGGGAACACC Exon 7

CCTCCACGGCTAATACAGTGGATCGAACTAACCACCAG/CCAACCCTGGCAGTCAGCGTG

GCCCAGATCTTTACAGGATATCCTTCTCCGACTCATGTTGAAAAACCCCTGCAAGAGTTGC Exon 8

ATGGACAG/AGGAATGAGGAACCTTCTTCCTGATGATGGCCCAAGAGAGGAATCCGGGGA Exon 9

AGGCCAGCTTGGCAG/GTGAATGTCATCCGAACACAGGGATGAGCTGGTGAGCAGTGTGG

FIG. 2E

5' 1 5' 61 5' 121	CCTTTAGTTC ATGAAGACAA TACTTCTGTT	11 AGGGGAAGAG TCCCCTTCTC 71 CACCCTCCCC GTGGGAGGGG 31 TTCCACTTT	21 G TTACCTCAGG AATGGAGTCC 81 G TTTAGAAGAC G AAATCTTCTG	31 G TCTTGTGCAG AGAACACGTC 91 C AGTGCTTCAG	41 GTGCCTGCTC CACGGACGAC 1 AACACTCCCA TTGTGAGGGT	51 G CTGTGAATTC GACACTTAAG 11 A ACTAGCCTCT TGATCGGAGA 71
			FIG. 2	F		
16	1416 (5	DEQ	id Na	(5:0		
5'	CCTCCAGGCT	11 CATTTCAGAG	21 AGAATATTCC	31 ACAGAGCTTG	41 AAGGCGCGGG TTCCGCGCCC	51 ATCTGGGCCG TAGACCCGGC
5' 61	ATGATGGAAG TACTACCTTC	GAAAGAGATT	81 GTCTGCATTC CAGACGTAAG Exon 1b	91 CAAAAGCTCA <u>GTTTTCGAGT</u>	1 GACAGAGAGA CTGTCTCTCT	11 TGGAAATCAA ACCTTTAGTT
5' 121	GAGGGGAAGA CTCCCCTTCT	31 GTTACCTCAG	41 GTCTTGTGCA	51 GGTGCACTGC CCACGTGACG	61 TGCTGTGAAT ACGACACTTA	TCATGAAGAC
5' 181	AACACCCTCC TTGTGGGAGG		TGTCACGAAG	TGTTGTGAGG		
5' 241	TGTTCACTTT ACAAGTGAAA	51 GTCCCCTCTT CAGGGGGAGAA	61 CTGAAGCAGG <u>GACTTCG</u> TCC	71	81 GTAAGAAACT CATTCTTTGA	91 GAGGAGCCTA CTCCTCGGAT
5' 301	GGGCAGCTGT CCCGTCGACA					
5' 361		1 8	31 9	21 1	1	1

FIG. 2G

27	30510 (Exon 1a Exon 1b				
5' 1	ATTCCAAAAG	11 CTCAGACAGA	21 GAGATGGAAG	31 TCAAGAGGGG	41 AAGAGTTACC	51 TCAGGTCTTG AGTCCAGAAC
5' 61					1 CTCCCCTTTA GAGGGGAAAT	11 GTAGACAGCG CATCTGTCGC
5' 121	<u>GAAGTGTTGT</u>			CTGATGTTCA	61 CTTTGTCCCC GAAACAGGGG	
5' 181					21 TTCAACCTTG AAGTTGGAAC	
5' 241		51 ACTAGCACGC TGATCGTGCG		71	81	91
M	DR-1 f(50	EQID	FIG. 2H	\mathbf{i}		
5' 1	GGAACCCGAA	11 CACTCTTGAG	21 TGCTCTCAGT	31 TACAGCCTAC	41 CGAGTCCGCA GCTCAGGCGT	
5' 61					1 Actgctctga Tgacgagact	
5' 121					61 Tgtcccacgt Acagggtgca	
5' 181					21 GGAGCCACAG CCTCGGTGTC	
5' 241					81 TGGCCCTCTA ACCGGGAGAT	
5. 301					41 TGATTGTAAG ACTAACATTC	

FIG. 21

361 ATGAAGACTG CCACCAACAT CTACATTTTC AACCTTGCTC TGGCAGATGC CTTAGCCACT TACTTCTGAC GGTGGTTGTA GATGTAAAAG TTGGAACGAG ACCGTCTACG GAATCGGTGA 421 AGCACGCTGC CCTTTCAGAG TGTTAACTAC CTGATGGGAA CGTGGCCCTT TGGAAACATC TCGTGCGACG GGAAAGTCTC ACAATTGATG GACTACCCTT GCACCGGGAA ACCTTTGTAG 5' 91 1 .11 21 31 481 CTCTGCAAGA TCGTGATCTC AATAGACTAC TACAACATGT TCACCAGTAT CTTCACCCTC GAGACGTTCT AGCACTAGAG TTATCTGATG ATGTTGTACA AGTGGTCATA GAAGTGGGAG 5' 541 TGCACCATGA GTGTAGACCG CTACATTGCC GICTGCCACC CGGTCAAGGC CCTGGATTTC ACGTGGTACT CACATCTGGC GATGTAACGG CAGACGGTGG GCCAGTTCCG GGACCTAAAG 5' 601 CGTACCCCCC GAAATGCCAA AATTGTCAAT GTCTGCAACT GGATCCTCTC TTCTGCCATT GCATGGGGGG CTTTACGGTT TTAACAGTIA CAGACGTTGA CCTAGGAGAG AAGACGGTAA 5' 661 GGTCTGCCCG TAATGTTCAT GGCAACCACA AAATACAGGC AGGGGTCCAT AGATTGCACC CCAGACGGGC ATTACAAGTA CCGTTGGTGT TTTATGTCCG TCCCCAGGTA TCTAACGTGG 5* 721 CTCACGTTCT CTCATCCCAC ATGGTACTGG GAGAACCTGC TCAAAATCTG TGTCTTCATC GAGTGCAAGA GAGTAGGGTG TACCATGACC CTCTTGGACG AGTTTTAGAC ACAGAAGTAG 5' 781 TTCGCCTTCA TCATGCCGGT CCTCATCATC ACTGTGTGTT ATGGACTGAT GATCTTACGA AAGCGGAAGT AGTACGGCCA GGAGTAGTAG TGACACACAA TACCTGACTA CTAGAATGCT 5' 841 CTCAAGAGTG TCCGCATGCT GTCGGGCTCC AAAGAAAAGG ACAGGAACCT GCGCAGGATC GAGTTCTCAC AGGCGTACGA CAGCCCGAGG TTTCTTTTCC TGTCCTTGGA CGCGTCCTAG 5' 901 ACCCGGATGG TGCTGGTGGT CGTGGCTGTA TTTATTGTCT GCTGGACCCC CATCCACATC TGGGCCTACC ACGACCACCA GCACCGACAT AAATAACAGA CGACCTGGGG GTAGGTGTAG 5' 961 TATGTCATCA TCAAAGCACT GATCACGATT CCAGAAACCA CTTTCCAGAC TGTTTCCTGG ATACAGTAGT AGTTTCGTGA CTAGTGCTAA GGTCTTTGGT GAAAGGTCTG ACAAAGGACC 1021 CACTTCTGCA TTGCCTTGGG TTACACAAAC AGCTGCCTGA ACCCAGTTCT TTATGCGTTC GTGAAGACGT AACGGAACCC AATGTGTTTG TCGACGGACT TGGGTCAAGA AATACGCAAG

FIG. 2J

5' 1081	CTGGATGAAA GACCTACTTT	91 ACTTCAAACG TGAAGTTTGC	1 ATGTTTTAGA TACAAAATCT	11 GAGTTCTGCA CTCAAGACGT	21 TCCCAACTTC AGGGTTGAAG	31 CTCCACAATC GAGGTGTTAG
5' 1141				TTGTGATCCC	81 AACACCCCTC TTGTGGGGGAG	
5' 1201			21 CCAGGCACCA	31 TGTGCATGCG	41 TGCCTGGAGC ACGGACCTCG	
5' 1261					1 CAGTTGGGAG GTCAACCCTC	
5' 1321					61 GCGCTGAACC CGCGACTTGG	
5' 1381				11 AAGAAAAAGC	21 TGGACTCCCA <u>ACCTGAGGGT</u>	
5' 1441					81 GGGGGGCCATC CCCCCGGTAG	
5. 1501					41 CAGTCAGCGT GTCAGTCGCA	
5' 1561					1 GCAAGAGTTG CGTTCTCAAC	11 CATGGACAGA
5' 1621					61 AATCCGGGGGA TTAGGCCCCT	
5' 1681				11 GAGCTGGTGA CTCGACCACT		

FIG. 2K

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MOR-IbII (SEQIDNO: 10)									
5	1 GGAACCCGAA	11 CACTCTTGAG	21 F TGCTCTCAG	31 T TACAGCCTA	41 CGAGTCCGCA GCTCAGGCGT	CCAACCATTO			
51 61	AGAACCATGG	ACAGCAGCGC	CGGCCCAGG	G AACATCAGCO	1 G ACTGCTCTGA C TGACGAGACT	T22123			
5' 121	CCTGCAAGTT GGACGTTCAA	GCTCCCCAGC	ACCTGGCTCC	: IGGCTCAACT	61 TGTCCCACGT ACAGGGTGCA	TEATCOCAAC			
5' 181	CAGTCCGACC GTCAGGCTGG	CATGCGGTCC	TAACCGCACG	alaattigaa i	21 GGAGCCACAG CCTCGGTGTC	31 CCTGTGCCCT GGACACGGGA			
	CAGACCGGCA GTCTGGCCGT	GCCCTTCCAT	GGTCACAGCC	ATCACCATCA	81 TGGCCCTCTA ACCGGGAGAT	TTCTATCGTG			
5. 301	TGTGTAGTGG ACACATCACC	GULTUTIG	AAACHICCIG	GICAIGTATG	41 TGATTGTAAG ACTAACATTC	ΑΤΑΤΑΓΓΑΑΑ			
	ATGAAGACTG	CCACCAACAT	CTACATTTTC	91 AACCTTGCTC TTGGAACGAG	1 TGGCAGATGC ACCGTCTACG	11 CTTAGCCACT GAATCGGTGA			
	AGCACGCTGC	CCTTTCAGAG	TGTTAACTAC	CTGATGGGAA	61 CGTGGCCCTT GCACCGGGAA	TGGAAACATC			
	CILIGLAAGA	ICGIGAICIC	AATAGACTAC	TACAACATGT	21 TCACCAGTAT AGTGGTCATA	OTOODADTTO			
5' 541	TGCACCATGA ACGTGGTACT	GTGTAGACCG	61 CTACATTGCC GATGTAACGG	GTCTGCCACC	CGGTCAAGGC	91 CCTGGATTTC GGACCTAAAG			
5' 601	CGTACCCCCC GCATGGGGGGG	GAAATGCCAA A	AATTGTCAAT	GTCTGCAACT	GGATCCTCTC	51 TTCTGCCATT AAGACGGTAA			
5' 661	GGTCTGCCCG CCAGACGGGC	TAATGTTCAT	GGCAACCACA	AAATACAGGC	AGGGGTCCAT	I 1 Agattgcacc Fctaacgtgg			

FIG. 2L

5' 721 CTCACGTTCT CTCATCCCAC ATGGTACTGG GAGAACCTGC TCAAAATCTG TGTCTTCATC GAGTGCAAGA GAGTAGGGTG TACCATGACC CTCTTGGACG AGTTTTAGAC ACAGAAGTAG 5' 781 TTCGCCTTCA TCATGCCGGT CCTCATCATC ACTGTGTGTT ATGGACTGAT GATCTTACGA AAGCGGAAGT AGTACGGCCA GGAGTAGTAG TGACACACAA TACCTGACTA CTAGAATGCT 5' 841 CTCAAGAGTG TCCGCATGCT GTCGGGCTCC AAAGAAAAGG ACAGGAACCT GCGCAGGATC GAGTTCTCAC AGGCGTACGA CAGCCCGAGG TTTCTTTTCC TGTCCTTGGA CGCGTCCTAG 5' 901 ACCCGGATGG TGCTGGTGGT CGTGGCTGTA ITTATTGTCT GCTGGACCCC CATCCACATC TGGGCCTACC ACGACCACCA GCACCGACAT AAATAACAGA CGACCTGGGG GTAGGTGTAG 5 * 961 TATGTCATCA TCAAAGCACT GATCACGATT CCAGAAACCA CTTTCCAGAC TGTTTCCTGG ATACAGTAGT AGTTTCGTGA CTAGTGCTAA GGTCTTTGGT GAAAGGTCTG ACAAAGGACC 1021 CACTTCTGCA TTGCCTTGGG TTACACAAAC AGCTGCCTGA ACCCAGTICT TTATGCGTTC GTGAAGACGT AACGGAACCC AATGTGTTTG TCGACGGACT TGGGTCAAGA AATACGCAAG 5 ' 1081 CTGGATGAAA ACTTCAAACG ATGTTTTAGA GAGTTCTGCA TCCCAACTTC CTCCACAATC GACCTACTTT TGAAGTTTGC TACAAAATCT CTCAAGACGT AGGGTTGAAG GAGGTGTTAG 5 ' 1141 GAACAGCAAA ACTCTGCTCG AATCCGTCAA AACACTAGGG AACACCCCTC CACGGCTAAT CTTGTCGTTT TGAGACGAGC TTAGGCAGTT TTCTGATCCC TTGTGGGGAG GTGCCGATTA Exon 5b 5' 1201 ACAGTGGATC GAACTAACCA CCAGAAGCTT TTAATGTGGA GAGCTATGCC TACATTCAAG TGTCACCTAG CTTGATTGGT GGTCTTCGAA AATTACACCT CTCGATACGG ATGTAAGTTC 5' 1261 AGACACTTGG CTATCATGTT AAGCCTTGAT AATTAGGGCA CCAAAGGGGA CAAGTGTCAA TCTGTGAACC GATAGTACAA TTCGGAACTA TTAATCCCGT GGTTTCCCCT GTTCACAGTT 1321 ATCAAGATGC TGTTTTTGTT TTTGTTTTTT GTTTTTTGTT TTTTCTGGTT CCATCAAGTT <u>ΤΑGTTCTACG ΑCΑΑΑΑΑCAA ΑΑΑCΑΑΑΑΑΑ CΑΑΑΑΑΑΑCAA ΑΑΑΑGACCAA GGTAGTTCAA</u> 5' 1381 CTTGTAGAAC ACTATTATGG TTAGCAATGC TCAATAGACA ATGTCAGGGG GTGTGACATA GAACATCTTG TGATAATACC AATCGTTACG AGTTATCTGT TACAGTCCCC CACACTGTAT

FIG. 2M

5' 1441		AGAAGCACTA		ACTCCATAGT	81 TGGAAGAGCA ACCTTCTCGT	
5' 1501					41 CTGTGAATTG GACACTTAAC	
5' 1561				TCAGAATTCA	1 Agaaattcag <u>Tctttaagtc</u>	
5' 1621		ATCTCTCACT	CCCATGCATT	CAAATGTGTC	61 ČTGAATACAT GACTTATGTA	CCACAGACAC
1681		AAAACTCTCT		AGCTTGTGCT	21 TCGTTTGGGT AGCAAACCCA	
5' 1741					81 GAAGCCTACC CTTCGGATGG	
					41 TTTTCTCTAA AAAAGAGATT	
	<u>ACAACGATTC</u>	ACAATCCGAA	CATTTACTGT	GCAAGAAAAC	1 TTTTGAATAC AAAACTTATG	
5' 1921	Exon 5a CAGAAAATAG GTCITTTATC	ATTTATTTTG	41 AAAAGGCATA TTTTCCGTAT	TACACAGAAC	61 TGGGAGAAGC <u>ACCCTCTTCG</u>	71 ACACCAAAGA TGTGGTTTCT
	TATTTTGTTA ATAAAACAAT	CCATATGGCA	1 AATGTAACCA TTACATTGGT	TAGAGAGCAG	21 Agtacctaat Tcatggatta	31 GCTGGTGCCA <u>CGACCACGGT</u>
	ACCCC TGGGG	51	61	71	81	91

FIG. 2N

MOR-1a (SEQ ID NO. 11)								
5'	GGAACCCGAA	11 CACTCTTGAG	21 TGCTCTCAGT	31 TACAGCCTAC ATGTCGGATG	CGAGTCCGCA	51 GCAAGCATTC CGTTCGTAAG		
	AGAACCATGG TCTTGGTACC	71 ACAGCAGCGC TGTCGTCGCG	CGGCCCAGGG	91 AACATCAGCG TTGTAGTCGC	ACTGCTCTGA	CCCCTTAGCT		
	CCTGCAAGTT GGACGTTCAA	31 GCTCCCCAGC CGAGGGGTCG	ACCTGGCTCC	51 TGGCTCAACT ACCGAGTTGA	TGTCCCACGT	71 TGATGGCAAC ACTACCGTFG		
	CAGTCCGACC GTCAGGCTGG	91 CATGCGGTCC GTACGCCAGG	1 TAACCGCACG ATTGGCGTGC	11 GGGCTTGGCG CCCGAACCGC	GGAGCCACAG	31 CCTGTGCCCT GGACACGGGA		
	CAGACCGGCA GTCTGGCCGT	51 GCCCTTCCAT CGGGAAGGTA	GGTCACAGCC	71 ATCACCATCA TAGTGGTAGT	81 TGGCCCTCTA ACCGGGAGAT	91 TTCTATCGTG AAGATAGCAC		
			AAACTTCCTG	31 GTCATGTATG CAGTACATAC	TGATTGTAAG			
	ATGAAGACTG TACTTCTGAC	71 CCACCAACAT GGTGGTTGTA	CTACATTTTC	91 AACCTTGCTC TTGGAACGAG	1 TGGCAGATGC ACCGTCTACG	11 CTTAGCCACT GAATCGGTGA		
	AGCACGCTGC TCGTGCGACG	CCTTTCAGAG	TGTTAACTAC	51 CTGATGGGAA GACTACCCTT	CGTGGCCCTT	71 TGGAAACATC ACCTTTGTAG		
	CTCTGCAAGA GAGACGTTCT	TCGTGATCTC	1 AATAGACTAC TTATCTGATG	11 TACAACATGT ATGTTGTACA	TCACCAGTAT	31 CTTCACCCTC GAAGTGGGAG		
5' 541	TGCACCATGA Acgtggtact	GTGTAGACCG	61 CTACATTGCC GATGTAACGG	71 GTCTGCCACC CAGACGGTGG	81 CGGTCAAGGC GCCAGTTCCG	91 CCTGGATTTC GGACCTAAAG		
5' 601		GAAATGCCAA		31 GTCTGCAACT CAGACGTTGA	GGATCCTCTC			
5' 661	GGTCTGCCCG CCAGACGGGC	TAATGTTCAT	81 GGCAACCACA CCGTTGGTGT	91 AAATACAGGC TTTATGTCCG	AGGGGTCCAT	11 AGATTGCACC TCTAACGTGG		

FIG. 20

5 ' 721 CTCACGTTCT CTCATCCCAC ATGGTACTGG GAGAACCTGC TCAAAATCTG TGTCTTCATC GAGTGCAAGA GAGTAGGGTG TACCATGACC CTCTTGGACG AGTTTTAGAC ACAGAAGTAG 5' 781 TICGCCTTCA TCATGCCGGT CCTCATCATC ACTGTGTGTT ATGGACTGAT GATCTTACGA AAGCGGAAGT AGTACGGCCA GGAGTAGTAG TGACACACAA TACCTGACTA CTAGAATGCT 5 ' 841 CTCAAGAGTG TCCGCATGCT GTCGGGCTCC AAAGAAAAGG ACAGGAACCT GCGCAGGATC GAGTTCTCAC AGGCGTACGA CAGCCCGAGG TTTCTTTTCC TGTCCTTGGA CGCGTCCTAG 5' 1 901 ACCCGGATGG TGCTGGTGGT CGTGGCTGTA TTTATTGTCT GCTGGACCCC CATCCACATC TGGGCCTACC ACGACCACCA GCACCGACAT AAATAACAGA CGACCTGGGG GTAGGTGTAG 5' 961 TATGTCATCA TCAAAGCACT GATCACGATT CCAGAAACCA CTTTCCAGAC TGTTTCCTGG ATACAGTAGT AGTITCGTGA CTAGTGCTAA GGTCTTTGGT GAAAGGTCTG ACAAAGGACC 5' 1021 CACTTCTGCA TTGCCTTGGG TTACACAAAC AGCTGCCTGA ACCCAGTTCT TTATGCGTTC GTGAAGACGT AACGGAACCC AATGTGTTTG TCGACGGACT TGGGTCAAGA AATACGCAAG 5. 1081 CTGGATGAAA ACTTCAAACG ATGTTTTAGA GAGTTCTGCA TCCCAACTTC CTCCACAATC GACCTACTIT TGAAGTITGC TACAAAATCT CTCAAGACGT AGGGTTGAAG GAGGTGTTAG 1141 GAACAGCAAA ACTCTGCTCG AATCCGTCAA AACACTAGGG AACACCCCTC CACGGCTAAT CTTGTCGTTT TGAGACGAGC TTAGGCAGTT TTGTGATCCC TTGTGGGGAG GTGCCGATTA Exon 3a 5* 1201 ACAGTGGATC GAACTAACCA CCAGGTATGT GCTTTCTAGA ATTATGTATA ACATATAAAA TGTCACCTAG CTTGATTGGT GGTCCATACA CGAAAGATCT TAATACATAT TGTATATTT 1261 ACACAGCACC TGATACCAGT CTAAGATTTA GATCCTTAAG GAGGTCGGTT ACTGGAGAAT TGTGTCGTGG ACTATGGTCA GATTCTAAAT CTAGGAATTC CTCCAGCCAA 1GACCTCTTA 5 ' 1321 CCAGCCAAGC CTAAAAATAG AGAGGGAGTA GGGGACCAAA TTCTG GGTCGGTTCG GATITTATC TCTCCCTCAT CCCCTGGTTT AAGAC

FIG. 2P

MOR-IDI (SEQ 10_{21} NO; 12) 41 51 1 GGAACCCGAA CACTCTTGAG TGCTCTCAGT TACAGCCTAC CGAGTCCGCA GCAAGCATTC CCTTGGGCTT GTGAGAACTC ACGAGAGTCA ATGTCGGATG GCTCAGGCGT CGTTCGTAAG 5' 71 81 91 1 11 61 AGAACCATGG ACAGCAGCGC CGGCCCAGGG AACATCAGCG ACTGCTCTGA CCCCTTAGCT TCTTGGTACC TGTCGTCGCG GCCGGGTCCC TTGTAGTCGC TGACGAGACT GGGGAATCGA 5* 31 41 51 61 71 121 CCTGCAAGTT GCTCCCCAGC ACCTGGCTCC TGGCTCAACT TGTCCCACGT TGATGGCAAC GGACGTTCAA CGAGGGGTCG TGGACCGAGG ACCGAGTTGA ACAGGGTGCA ACTACCGTTG 5' 91 1 11 21 31 181 CAGTCCGACC CATGCGGTCC TAACCGCACG GGGCTTGGCG GGAGCCACAG CCTGTGCCCT GTCAGGCTGG GTACGCCAGG ATTGGCGTGC CCCGAACCGC CCTCGGTGTC GGACACGGGA 5 * 51 61 71 81 91 241 CAGACCGGCA GCCCTTCCAT GGTCACAGCC ATCACCATCA TGGCCCTCTA TTCTATCGTG GTCTGGCCGT CGGGAAGGTA CCAGTGTCGG TAGTGGTAGT ACCGGGAGAT AAGATAGCAC 51 11 21 31 41 51 301 TGTGTAGTGG GCCTCTTTGG AAACTTCCTG GTCATGTATG TGATTGTAAG ATATACCAAA ACACATCACC CGGAGAAACC TITGAAGGAC CAGTACATAC ACTAACATTC TATATGGTTT 5 ' 71 81 91 1 11 361 ATGAAGACTG CCACCAACAT CTACATTTTC AACCTTGCTC TGGCAGATGC CTTAGCCACT TACTTCTGAC GGTGGTTGTA GATGTAAAAG TTGGAACGAG ACCGTCTACG GAATCGGTGA 5' 31 l 41 51 ° 61 71 421 AGCACGCTGC CCTTTCAGAG TGTTAACTAC CTGATGGGAA CGTGGCCCTT TGGAAACATC TCGTGCGACG GGAAAGTCTC ACAATTGATG GACTACCCTT GCACCGGGAA ACCTTTGTAG 5' 91 1 11 21 31 481 CTCTGCAAGA TCGTGATCTC AATAGACTAC TACAACATGT TCACCAGTAT CTTCACCCTC GAGACGTTCT AGCACTAGAG TTATCTGATG ATGTTGTACA AGTGGTCATA GAAGTGGGAG 5' 51 61 71 81 91 541 TGCACCATGA GTGTAGACCG CTACATTGCC GTCTGCCACC CGGTCAAGGC CCTGGATTTC ACGTGGTACT CACATCTGGC GATGTAACGG CAGACGGTGG GCCAGTTCCG GGACCTAAAG 5' 11 21 31 41 51 . 601 CGTACCCCCC GAAATGCCAA AATTGTCAAT GTCTGCAACT GGATCCTCTC TTCTGCCATT GCATGGGGGG CTTTACGGTT TTAACAGTTA CAGACGTTGA CCTAGGAGAG AAGACGGTAA 5' 71 81 91 1 11 661 GGTCTGCCCG TAATGTTCAT GGCAACCACA AAATACAGGC AGGGGTCCAT AGATTGCACC CCAGACGGGC ATTACAAGTA CCGTTGGTGT TTTATGTCCG TCCCCAGGTA TCTAACGTGG

FIG. 2Q

721 CTCACGTTCT CTCATCCCAC ATGGTACTGG GAGAACCTGC TCAAAATCTG TGTCTTCATC GAGTGCAAGA GAGTAGGGTG TACCATGACC CTCTTGGACG AGTTTTAGAC ACAGAAGTAG 781 TTCGCCTTCA TCATGCCGGT CCTCATCATC ACTGTGTGTT ATGGACTGAT GATCTTACGA AAGCGGAAGT AGTACGGCCA GGAGTAGTAG TGACACACAA TACCTGACTA CTAGAATGCT 841 CTCAAGAGTG TCCGCATGCT GTCGGGCTCC · AAAGAAAAGG ACAGGAACCT GCGCAGGATC GAGTTCTCAC AGGCGTACGA CAGCCCGAGG TTTCTTTTCC TGTCCTTGGA CGCGTCCTAG 5 * 901 ACCCGGATGG TGCIGGTGGT CGTGGCTGTA TTTATTGTCT GCTGGACCCC CATCCACATC TGGGCCTACC ACGACCACCA GCACCGACAT AAATAACAGA CGACCTGGGG GTAGGTGTAG 5 * 961 TATGTCATCA TCAAAGCACT GATCACGATT CCAGAAACCA CTTTCCAGAC TGTTTCCTGG ATACAGTAGT AGTTTCGTGA CTAGTGCTAA GGTCTTTGGT GAAAGGTCTG ACAAAGGACC ć 61 1021 CACTTCTGCA TTGCCTTGGG TTACACAAAC AGCTGCCTGA ACCCAGTTCT TTATGCGTTC GTGAAGACGT AACGGAACCC AATGTGTTTG TCGACGGACT TGGGTCAAGA AATACGCAAG 5' 1081 CTGGATGAAA ACTTCAAACG ATGTTTTAGA GAGTTCTGCA TCCCAACTTC CTCCACAATC GACCTACTTT TGAAGTTTGC TACAAAATCT CTCAAGACGT AGGGTTGAAG GAGGTGTTAG 5' 1141 GAACAGCAAA ACTCTGCTCG AATCCGTCAA AACACTAGGG AACACCCCTC CACGGCTAAT CTTGTCGTTT TGAGACGAGC TTAGGCAGTT ITGTGATCCC TTGTGGGGAG GTGCCGATTA Exon 5a 5' 1201 ACAGTGGATC GAACTAACCA CCAGAAAATA GATTTATTTT GAAAAGGCAT ATACACAGAA TGTCACCTAG CTTGATTGGT GGTCITTTAT_CTAAATAAAA CTTTTCCGTA TATGTGTCTT 5' 1261 CTGGGAGAAG CACACCAAAG ATATTTTGTT ACCATATGGC AAATGTAACC ATAGAGAGCA GACCCTCTTC GTGTGGTTTC TATAAAACAA TGGTATACCG TTTACATTGG TATCTCTCGT 5' 1321 GAGTACCTAA TGCTGGTGCC AACCCC CTCATGGATT ACGACCACGG TTGGGG

FIG. 2R

MOR-11 (SEQ 1D NO: 13)

Exon 11 5 * 11 21 31 41 51 1 GGGAACACCC CTCCACGGCT AATACAGTGG ATCGAACTAA CCACCAGTGT GTATGAGTGC CCCTTGTGGG GAGGTGCCGA TTATGTCACC TAGCTTGATT GGTGGTDACA CATACTCACG 5' 71 81 91 1 11 61 TATGCCCACA GGGACCAGAA GATGGTATCA GACCTTCTAG AACTGAAGTA GTGAGCAGTC ATACGGGTGT CCCTGGTCTT CTACCATAGT CTGGAAGATC TTGACTTCAT CACTCGTCAG Exon 5b 5' 31 41 -51 61 71 121 CCCACCCCCA CCCCCGCAA TAAAATAGAT TTATTTTGAA AAGGCATATA CACAGAACTG GGGTGGGGGT GG<u>GGGGGCGTT AITTTATCTA AATAAAACTT TTCCGTATAT GTGTCTTGAC</u> 5' 91 181 GGAGAAGCAC ACC

CCTCTTCGTG TGG

FIG. 2S

MOR-1; (SEO ID NO: 14) Exon 12 1 gggaacacce etceaegget aatacagtgg ategaactaa ceaecaggag eetcagteag 61 <u>cqqaqacatq atqtqaatga acqqactqat taqacaaqqt ttcctqaaca ctqaqataca</u> 121 <u>aaacaaatag agagettaet agagaaaatt egtageeega aaatteaatt atagaaacaa</u> 181 atgaqtqtta gagtagatat ggtaaggcet caqaqaggtt ttattteacq actaacaaca 241 tracccaage cacctaatec ategteatta gattacaaag acaattetag tecetegeee 301 <u>aqaqaaatqt ttqtctccca caqacaagcc tcacacttca gtaatqaaat qaqtaaatta</u> 361 <u>aatcqqtqaq caaqatqqtq qqaqqaqtca aaatattttc atqccttcct qtqqaactcc</u> 421 <u>aaaqqaaqac caacacaqtc aactaacctq qctcttqqtq gctctcaqaq ctqaacaacc</u> <u>aaccaaagag cactcatgag ctagacctag gcctctttta cacgtgtagc agatgtgcgt</u> ctccatcttc atgtgggtcc ccccaacaag taaagtagca gctgtctcta aagctgttgc <u>ctqtctqqct tcqqtqgaaq aaqatqtqat tcqcttaacc ctgaaqtqac ttgatatgca</u> Exon 5a

gggaaaatag atttattttg aaaaggcata tacacagaac tgggagaagc acacc

FIG. 2T

MOR-1-610302 (SEQ ID NO:15) Exon 4c

G GAT AGA ACT AAT CAT CAG / TGC CTA CCT ATA CCT TCC CTG TCT TGC TGG GCT CTA GAG CAT GGC CGC TTG GTT GTG TAC CCT GGA CCA CTG CAA GGA CCT CTT GTC AGA TAT GAC CTC CCA GCT

FIG. 20 MOR-1 (SEO ID NO:16)

gctccctccc ttccactcag agagtggcgc tttggggatg ctaaggatgc gcctccgtgt 1 61 acttetaagg tgggaggggg etacaageag aggagaatat eggaegetea gaegtteeat 121 tctgcctgcc gctcttctct ggttccacta gggcttgtcc ttgtaagaaa ctgacggagc 181 ctagggcagc tgtgagagga agaggctggg gcgcctggaa cccgaacact cttgagtgct 241 ctcagttaca geetaccgag teegcageaa geatteagaa ceatggaeag eagegeegge 301 ccagggaaca tcagcgactg ctctgacccc ttagctcctg caagttgctc cccagcacct 361 ggctcctggc tcaacttgtc ccacgttgat ggcaaccagt ccgacccatg cggtcctaac 421 cgcacggggc ttggcgggag ccacagcetg tgccetcaga ccggcageee ttccatggte 481 acagccatca ccatcatggc cctctattct atcgtgtgtg tagtggggcct ctttggaaac 541 ttcctggtca tgtatgtgat tgtaagatat accaaaatga agactgccac caacatctac 601 attttcaacc ttgctctggc agatgcctta gccactagca cgctgccctt tcagagtgtt 661 aactaeetga tgggaaegtg geeetttgga aacateetet geaagategt gateteaata 721 gactactaca acatgttcac cagtatette accetetgea ceatgagtgt agaeegetae 781 attgccgtct gccacccggt caaggccctg gatttccgta ccccccgaaa tgccaaaatt 841 gtcaatgtct gcaactggat cctctttct gccattggtc tgcccgtaat gttcatggca 901 accacaaaat acaggcaggg gtccatagat tgcaccctca cgttctctca tcccacatgg 961 tactgggaga acctgctcaa aatctgtgtc ttcatcttcg ccttcatcat gccggtcctc 1021 atcatcactg tgtgttatgg actgatgatc ttacgactca agagtgtccg catgctgtcg 1081 ggctccaaag aaaaggacag gaacctgcgc aggatcaccc ggatggtgct ggtggtcgtg 1141 gctgtattta ttgtctgctg gacccccatc cacatctatg tcatcatcaa agcactgatc 1201 acgattecag aaaceaettt ecagaetgtt teetgeaet tetgeattge ettgggttae 1261 acaaacagct gcctgaaccc agttctttat gcgttcctgg atgaaaactt caaacgatgt 1321 tttagagagt tctgcatccc aacttcctcc acaatcgaac agcaaaactc tgctcgaatc 1381 cgtcaaaaca ctagggaaca cccctccacg gctaatacag tggatcgaac taaccaccag 1441 ctagaaaatc tggaagcaga aactgctcca ttgccctaac tgggtcccac gccatccaga 1501 ccctcgctaa acttagaggc tgccatctac ttggaatcag gttgctgtca gggtttgtgg 1561 gaggetetgg ttteetggaa aageatetga teetgeatte aaagteatte

FIG. 2V

MOR-IC (SEQ ID NO:17)

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Q PTLAVSVAQIFTGYPSPTHVEKPCKSCMDR GM

Exon 9

<u>RNLLPDDGPRQESGEGQLG</u> <u>R</u>

FIG. 3A

MOR-1G (SEQ 10 NO: 18)

Exon la

 M E R I S K A G S P P G S F Q R E Y S T E L E G A G S G P M M E A F

 S K S A F Q K L R Q R D G N Q E G K S Y L R Y T K M K T A T N I Y I F

 N L A L A D A L A T S T L P F Q S V N Y L M G T W P F G N I L C K I V

 I S I D Y Y N M F T S I F T L C T M S V D R Y I A V C H P V K A L D F

 R T P R N A K I V N V C N W I L S S A I G L P V M F M A T T K Y R Q G

 S I D C T L T F S H P T W Y W E N L L K I C V F I F A F I M P V L I I

 T V C Y G L M I L R L K S V R M L S G S K E K D R N L R R I T R M V L

 V V A V F I V C W T P I H I Y V I I K A L I T I P E T T F Q T V S W

 H F C I A L G Y T N S C L N P V L Y A F L D E N F K R C F R E F C I P

 T S S T I E Q Q N S A R I R Q N T R E H P S T A N T V D R T N H Q L E

FIG. 3B

MOR-1D (SEQ ID NO: 19)

M D S S A G P G N I S D C S D P L A P A S C S P A P G S W L N L S H V DGNQSDPCGPNRTGLGGSHSLCPQTGSPSMVTAIT I M A L Y S I V C V V G L F G N F L V M Y V I V R Y T K M K T A T N I Y I F N L A L A D A L A T S T L P F Q S V N Y L M G T W P F G N I L C K I V I S I D Y Y N M F T S I F T L C T M S V D R Y I A V C H P V K A LDFRTPRNAKIVNVCNWILSSAIGLPVMFMATTKY RQGSIDCTLTFSHPTWYWENLLKICVFIFAFIMPV LIITVCYGLMILRLKSVRMLSGSKEKDRNLRRITR M V L V V A V F I V C W T P I H I Y V I I K A L I T I P E T T F Q T V S W H F C I A L G Y T N S C L N P V L Y A F L D E N F K R C F R E F C I P T S S T I E Q Q N S A R I R Q N T R E H P S T A N T V D R T N H Exon 8

QIRNEEPS

FIG. 3C

MOR-1E (SEQ 10 NO: 20) M D S S A G P G N I S D C S D P L A P A S C S P A P G S W L N L S H V D G N Q S D P C G P N R T G L G G S H S L C P Q T G S P S M V T A.I T I M A L Y S I V C V V G L F G N F L V M Y V I V R Y T K M K T A T N I Y I F N L A L A D A L A T S T L P F Q S V N Y L M G T W P F G N I L C K I V I S I D Y Y N M F T S I F T L C T M S V D R Y I A V C H P V K A LDFRTPRNAKIVNVCNWILSSAIGLPVMFMATTKY RQGSIDCTLTFSHPTWYWENLLKICVFIFAFIMPV LIITVCYGLMILRLKSVRMLSGSKEKDRNLRRITR M V L V V A V F I V C W T P I H I Y V I I K A L I T I P E T T F Q T V S W H F C I A L G Y T' N S C L N P V L Y A F L D E N F K R C F R E F CIPTSSTIEQQNSARIRQNTREHPSTANTVDRTNH Exon 6

Q | <u>KKKLDSQRGCVOHPV</u>

FIG. 3D

MOR-1H (SEO ID NO: 21) Exon 1a MERISKAGSPPGSFQREYSTELEGAGSGPMMEAF <u>SKSAFQKLRQRDGNQEGKSYLR</u>YTKMKTATNIYIF N L A L A D A L A T S T L P F Q S V N Y L M G T W P F G N I L C K I V I S I D Y Y N M F T S I F T L C T M S V D R Y I A V C H P V K A L D F R T P R N A K I V N V C N W I L S S A I G L P V M F M A T T K Y R Q G SIDCTLTFSHPTWYWENLLKICVFIFAFIMPVLII T V C Y G L M I L R L K S V R M L S G S K E K D R N L R R I T R M V L V V V A V F I V C W T P I H I Y V I I K A L I T I P E T T F Q T V S W H F C I A L G Y T N S C L N P V L Y A F L D E N F K R C F R E F C I P T S S T I E Q Q N S A R I R Q N T R E H P S T A N Ť V D R T N H Q Exon 7 Exon 8 PTLAVSVAQIFTGYPSPTHVEKPCKSCMDR GMR Exon 9 NLLPDDGPRQESGEGQLG | R FIG. 3E MOR-11 (SEQ ID NO:22) Exon 3 Exon 11 EHPSTANTVDRTNHQ CV FIG. 3F MOR-1J (SEQ ID NO:23) Exon 3 Exon 12 EHPSTANTVDRTNHQ EPQSAET FIG. 3G hMOR-1-610302 (SEQ ID NO! 24) Exon 3 Exon 5 DRTNHQ CLPIPSLSCWALEHGRLVVYPGPLQ GPLVRYDLPA

FIG. 3H

MOR-1A (SEQ ID NO! 25) Ν 1 MDSSAGPGNI SDCSDPLAPA SCSPAPGSWL NLSHVDGNQS DPCGPNRTGL GGSHSLCPOT N 61 GSPSMVTAIT IMALYSIVCV VGLFGNFLVM YVIVRYTKMK TATNIYIFNL ALADALATST Ν 121 LPFQSVNYLM GTWPFGNILC KIVISIDYYN MFTSIFTLCT MSVDRYIAVC HPVKALDFRT N 181 PRNAKIVNVC NWILSSAIGL PVMFMATTKY RQGSIDCTLT FSHPTWYWEN LLKICVFIFA Ν 81 -241 FIMPVLIITV CYGLMILRLK SVRMLSGSKE KDRNLRRITR MVLVVVAVFI VCWTPIHIYV Ν 301 IIKALITIPE TTFQTVSWHF CIALGYTNSC LNPVLYAFLD ENFKRCFREF CIPTSSTIEQ Exon 3a N 361 QNSARIRONT REHPSTANTV DRTNHOVCAF FIG. 31 $\operatorname{MOR-1BI}_{N} \left(\operatorname{SGQ}_{21} | \operatorname{OO}_{21}^{\prime} | 26 \right)$ 1 MDSSAGPGNI SDCSDPLAPA SCSPAPGSWL NLSHVDGNQS DPCGPNRTGL GGSHSLCPQT N 61 GSPSMVTAIT IMALYSIVCV VGLFGNFLVM YVIVRYTKMK TATNIYIFNL ALADALATST N 121 LPFQSVNYLM GTWPFGNILC KIVISIDYYN MFTSIFTLCT MSVDRYIAVC HPVKALDFRT Ν 181 PRNAKIVNVC NWILSSAIGL PVMFMATTKY ROGSIDCTLT FSHPTWYWEN LLKICVFIFA N 241 FIMPVLIITV CYGLMILRLK SVRMLSGSKE KDRNLRRITR MVLVVVAVFI VCWTPIHIYV N 301 IIKALITIPE TTFQTVSWHF CIALGYTNSC LNPVLYAFLDE NFKRCFREF CIPTSSTIEQ Exon 5b 361 QNSARIRONT REHPSTANTY DRTNHOKIDL F

FIG. 3J

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MOR-IBII (SEQ ID NO',27) Ν 1 MDSSAGPGNI SDCSDPLAPA SCSPAPGSWL NLSHVDGNQS DPCGPNRTGL GGSHSLCPQT Ν 61 GSPSMVTAIT IMALYSIVCV VGLFGNFLVM YVIVRYTKMK TATNIYIFNL ALADALATST Ν 121 LPFQSVNYLM GTWPFGNILC KIVISIDYYN MFTSIFTLCT MSVDRYIAV CHPVKALDFRT N 181 PRNAKIVNVC NWILSSAIGL PVMFMATTKY RQGSIDCTLT FSHPTWYWEN LLKICVFIFA Ν 241 FIMPVLIITV CYGLMILRLK SVRMLSGSKE KORNLRRITR MVLVVVAVFI VCWTPIHIYV Ν 301 IIKALITIPE TTFQTVSWHF CIALGYTNSC LNPVLYAFLD ENFKRCFREF CIPTSSTIEQ Exon 5b Ν 361 ONSARIRONT REHPSTANTV DRTNHOKLLM WRAMPTEKRH LAIMLSLDN

FIG. 3K

MOR-IF (SEQ ID NO! 28) Ν 1 MDSSAGPGNI SDCSDPLAPA SCSPAPGSWL NLSHVDGNQS DPCGPNRTGL GGSHSLCPQT Ν GSPSMVTAIT IMALYSIVCV VGLFGNFLVM YVIVRYTKMK TATNIYIFNL ALADALATST Ν LPFQSVNYLM GTWPFGNILC KIVISIDYYN MFTSIFTLCT MSVDRYIAVC HPVKALDFRT Ν 21: PRNAKIVNVC NWILSSAIGL PVMFMATTKY RQGSIDCTLT FSHPTWYWEN LLKICVFIFA N FIMPVLIITV CYGLMILRLK SVRMLSGSKE KDRNLRRITR MVLVVVAVFI VCWTPIHIYV Ν IIKALITIPE TTFOTVSWHF CIALGYTNSC LNPVLYAFLD ENFKRCFREF CIPTSSTIEQ Exon 10 N QNSARIRONT REHPSTANTV DRTNHQAPCA_CVPGANRGQT_KASDLLDLEL_ETVGSHQADA Ν ETNPGPYEGS_KCAEPLAISL_VPLY

FIG. 3L

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MOR-1		(5	EG	2 1[) n	00	29)
ggshslcpqt	aladalatst	hpvkaldfrt	llkicvfifa	vcwtpihiyv	ciptsstieq		
dpcgpnrtgl	tatniyifnl	msvdryiavc	fshptwywen	mvlvvvavfi vcwtpihiyv	enfkrcfref		
nlshvdgnqs	yvivrytkmk	c kivisidyyn mftsiftlct	rqgsidctlt	kdrnlrritr	lnpvlyafld	eaetaplp	
scspapgswl	imalysivcv vglfgnflvm	kivisidyyn	pvmfmattky	svrmlsgske	cialgytnsc	drtnhqlenl	FIG. 3M
sdcsdplapa		gtwpfgnil	nwilssaigl	cyglmilrlk	ttfqtvswhf	rehpstantv	
mdssagpgni	gspsmvtait	lpfqsvnylm	prnakivnvc	241 fimpvliitv	iikalitipe	qnsarirqnt	
ب ا	61	121	181	241	301	361	

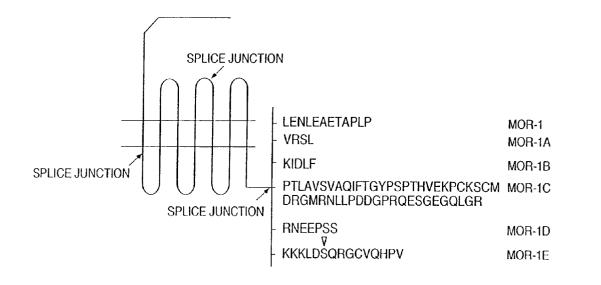
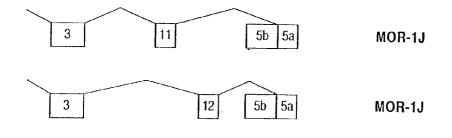


FIG. 4 (SEQ ID NOS! 30-35)





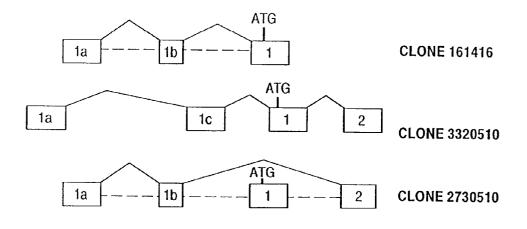


FIG. 6

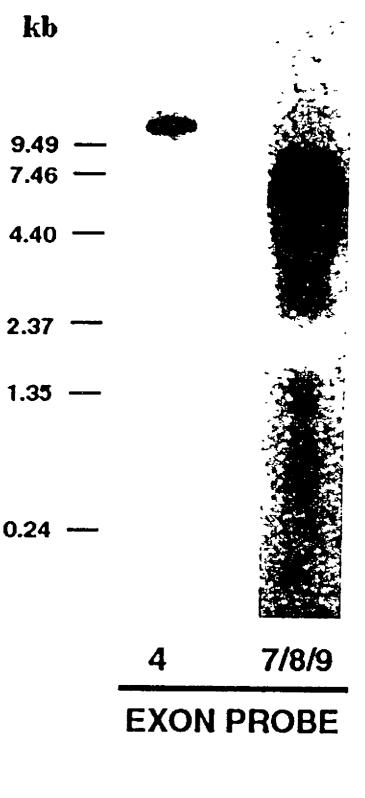


FIG. 7

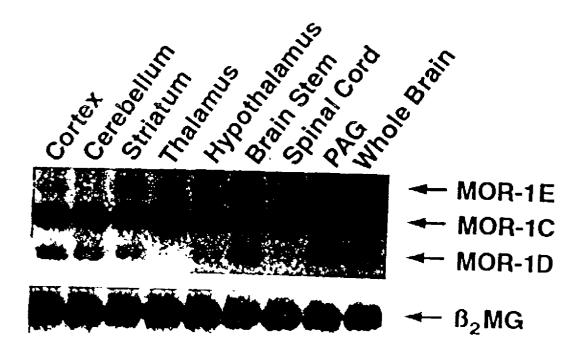


FIG. 8A

$MOR-1 \longrightarrow \texttt{MOR-1} \texttt{MOR-1}$

FIG. 8B

FIG. 9A

FIG. 9B

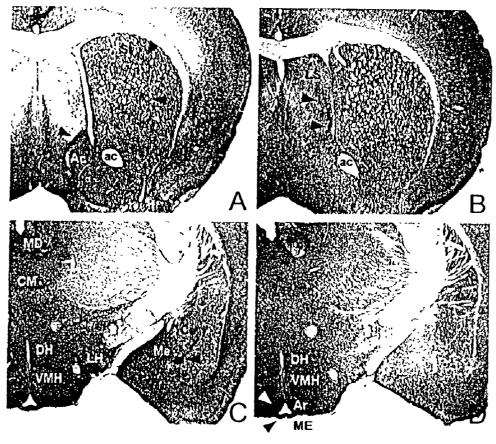
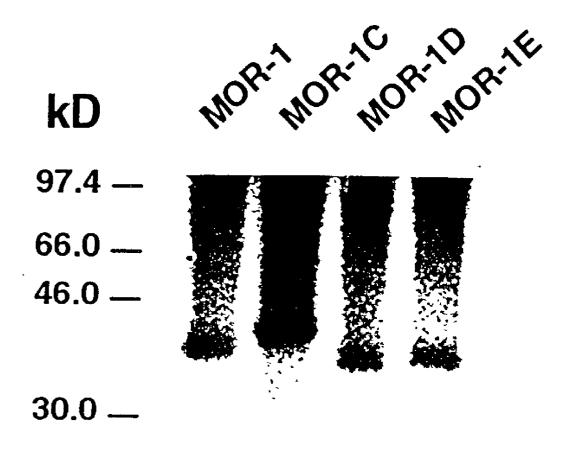


FIG. 9C

FIG. 9D



21.5 -

14.3 —

FIG. 10

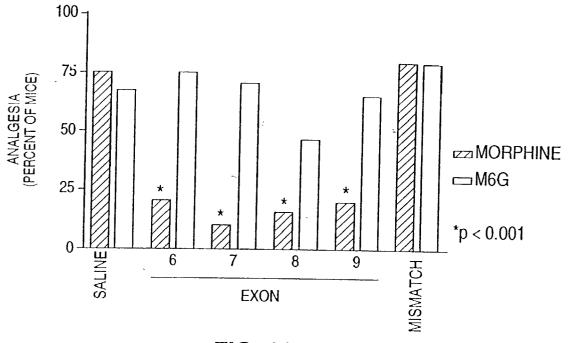


FIG. 11

IDENTIFICATION AND CHARACTERIZATION OF MULTIPLE SPLICE VARIANTS OF THE MU-OPIOID RECEPTOR GENE

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This is a continuation-in-part of the Jan. 16, 2001 U.S. national phase application of International Application PCT/US99/15974, having an international filing date of Jul. 15 1999, and designating the U.S. and claiming priority from U.S. Provisional Application No. 60/092,980, filed Jul. 16 1998.

STATEMENT OF RIGHTS TO INVENTIONS MADE UNDER FEDERALLY SPONSORED RESEARCH

[0002] This work was supported by the government, in part, by grants from the National Institute on Drug Abuse (DA02615, DA06241 and DA07242) and a Senior Scientist Award (DA00220) to Gavril W. Pasternak and a core grant to Memorial Sloan-Kettering Cancer Center, New York, N.Y. (CA08748). The government may have certain rights to this invention.

TECHNICAL FIELD

[0003] The present invention relates to mu-opioid receptor-1 (MOR-1) splice variant polypeptides, to DNA sequences encoding the splice variants, to DNA sequences encompassing non-coding region splice variants, to methods of screening compositions for agonists and antagonists of the splice variant receptor activities and to methods of measuring splice variant binding activities.

BACKGROUND ART

[0004] Opiates are drugs derived from opium and include morphine, codeine and a wide variety of semisynthetic opioid congeners derived from them and from thebaine, another component of opium. Opioids include the opiates and all agonists and antagonists with morphine-like activity and naturally occurring endogenous and synthetic opioid peptides. Morphine and other morphine-like opioid agonists are commonly used pharmaceutically to produce analgesia.

[0005] There are now many compounds with pharmacological properties similar to those produced by morphine, but none has proven to be clinically superior in relieving pain. References to morphine herein will be understood to include morphine-like agonists as well. The effects of morphine on human beings are relatively diverse and include analgesia, drowsiness, changes in mood, respiratory depression, decreased gastrointestinal motility, nausea, vomiting, and alterations of the endocrine and autonomic nervous systems. Pasternak (1993) Clin. Neuropharmacol. 16:1. Doses of morphine need to be tailored based on individual sensitivity to the drug and the pain-sparing needs of the individual. For instance, the typical initial dose of morphine (10 mg/70 kg) relieves post-operative pain satisfactorily in only two-thirds of patients. Likewise, responses of an individual patient may vary dramatically with different morphine-like drugs and patients may have side effects with one such drug and not another. For example, it is known that some patients who are unable to tolerate morphine may have no problems with an equianalgesic dose of methadone. The mechanisms underlying variations in individual responses to morphine and morphine-like agonists have not been defined.

[0006] The analgesic effects of morphine are transduced through opioid receptors in the central nervous system (CNS), located at both spinal and multiple supraspinal sites. Morphine and other agonists induce profound analgesia when administered intrathecally or instilled locally into the dorsal horn of the spinal cord. Several mechanisms of action are believed to mediate the inhibition of nociceptive reflexes from reaching higher centers of the brain, including the inhibition of neurotransmitter release by opioid receptors on the termini of primary afferent nerves and post synaptic inhibitory actions on interneurons and on the out-put neurons of the spinothalamic tract.

[0007] Profound analgesia can also be produced by the instillation of morphine into the third ventricle or within various sites in the midbrain and medulla, most notably the periaqueductal gray matter, the nucleus raphe magnus, and the locus ceruleus. Although the neuronal circuitry responsible has not been defined, these actions produce enhanced activity in the descending aminergic bulbospinal pathways that exert inhibitory effects on the processing of nociceptive information in the spinal cord. Simultaneous administration of morphine at both spinal and supraspinal sites results in a synergized analgesic response, with a ten-fold reduction in the total dose of morphine necessary to produce equivalent analgesia at either site alone.

[0008] Morphine also exerts effects on the neuroendocrine system. Morphine acts in the hypothalamus to inhibit the release of gonadotropin releasing hormone (GnRH) and corticotropin-releasing factor (CRF), thus decreasing circulating concentrations of luteinizing hormone (LH), follicle stimulating hormone (FSH), and adrenocorticotropin (ACTH), and β -endorphin. As a result of the decreased concentrations of pituitary trophic hormones, the concentrations of testosterone and cortisol in the plasma decline. The administration of opiates increases the concentration of prolactin (PRL) in plasma, most likely by reducing the dopaminergic inhibition of PRL secretion. With chronic administration, tolerance eventually develops to the effects of morphine on hypothalamic releasing factors.

[0009] Opiates can interfere with normal gastrointestinal functioning. Morphine decreases both gastric motility and the secretion of hydrochloric acid in the stomach. Morphine may delay passage of gastric contents through the duodenum for as long as 12 hours. Morphine also decreases biliary, pancreatic, and intestinal secretions and delays the digestion of food in the small intestine. Propulsive peristaltic waves in the colon are diminished or abolished after administration of morphine and commonly, constipation occurs. For a detailed review of the physiological effects of morphine, see Reisine and Pasternak (1996) Goodman & Gilman's The pharmacological basis of therapeutics, Ninth Edition (Hardman et al. eds.) McGraw-Hill pp. 521-555.

[0010] Morphine also exerts effects on the immune system. The most firmly established effect of morphine is its ability to inhibit the formation of rosettes by human lymphocytes. The administration of morphine to animals causes suppression of the cytotoxic activity of natural killer cells and enhances the growth of implanted tumors. These effects appear to be mediated by actions within the CNS. By contrast, β -endorphin enhances the cytotoxic activity of

human monocytes in vitro and increases the recruitment of precursor cells into the killer cell population; this peptide also can exert a potent chemotactic effect on these cells. A novel type of receptor (designated ϵ) may be involved. These effects, combined with the synthesis of Proopiomel-anocortin (POMC) and preproenkephalin by various cells of the immune system, have stimulated studies of the potential role of opioids in the regulation of immune function. Sibinga and Goldstein (1988) Annu. Rev. Immunol. 6:219.

[0011] Side effects resulting from the use of morphine range from mild to life-threatening. Morphine causes constriction of the pupil by an excitatory action on the parasympathetic nerve innervating the pupil. Morphine depresses the cough reflex through inhibitory effects on the cough centers in the medulla. Nausea and vomiting occur in some individuals through direct stimulation of the chemoreceptor trigger zone for emesis, in the postrema of the medulla. Therapeutic doses of morphine also result in peripheral vasodilatation, reduced peripheral resistance and an inhibition of baroreceptor reflexes in the cardiovascular system. Additionally, morphine provokes the release of histamines, which can cause hypotension. Morphine depresses respiration, at least in part by direct effects on the brainstem regulatory systems. In humans, death from morphine poisoning is nearly always due to respiratory arrest. Opioid antagonists can produce a dramatic reversal of severe respiratory depression and naloxone is currently the treatment of choice. High doses of morphine and related opioids can produce convulsions that are not always relieved by naloxone.

[0012] The development of tolerance and physical dependence with repeated use is a characteristic feature of all opiates. Dependence seems to be closely related to tolerance, since treatments that block tolerance to morphine also block dependence. In vivo studies in animal models demonstrate the importance of neurotransmitters and their interactions with opioid pathways in the development of tolerance to morphine. Blockade of glutamate actions by noncompetitive and competitive NMDA (N-methyl-D-aspartate) antagonists blocks morphine tolerance. Trujillo and Akil (1991) Science 251:85; and Elliott et al. (1994) Pain 56:69. Blockade of the glycine regulatory site on NMDA receptors has similar effects to block tolerance. Kolesnikov et al. (1994) Life Sci. 55:1393. Administering inhibitors of nitric oxide synthase in morphine-tolerant animals reverses tolerance, despite continued opioid administration. Kolesnikov et al. (1993) Proc. Natl. Acad. Sci. U.S.A. 90:5162. These studies indicate several important aspects of tolerance and dependence. First, the selective actions of drugs on tolerance and dependence demonstrate that analgesia can be dissociated from these two unwanted actions. Second, the reversal of preexisting tolerance by NMDA antagonists and nitric oxide synthase inhibitors indicates that tolerance is a balance between activation of processes and reversal of those processes. These observations suggest that, by use of selective agonists and/or antagonists, tolerance and dependence in the clinical management of pain can be minimized or disassociated from the therapeutic effects.

[0013] In addition to morphine, there are a variety of opioids suitable for clinical use. These include, but are not limited to, Levorphanol, Meperidine, Fentanyl, Methadone, Codeine, Propoxyphene and various opioid peptides. Certain opioids are mixed agonists/antagonists and partial ago-

nists. These include pentazocine, nalbuphine, butorphanol, and buprenorphine. The pharmacological effects of levorphanol closely parallel those of morphine although clinical reports suggest that levorphanol produces less nausea.

[0014] Meperidine exerts its chief pharmacological effects on the central nervous system and the neural elements in the bowel. Meperidine produces a pattern of effects similar but not identical to those described for morphine. In equianalgesic doses, meperidine produces as much sedation, respiratory depression, and euphoria as morphine. The pattern of unwanted side effects that follow the use of meperidine are similar to those observed after equianalgesic doses of morphine, except that constipation and urinary retention are less common.

[0015] Fentanyl is a synthetic opioid estimated to be 80 times as potent as morphine as an analgesic. High doses of fentanyl can result in severe toxicity and produce side effects including muscular rigidity and respiratory depression.

[0016] Methadone is an opioid with pharmacological properties similar to morphine. The properties of methadone include effective analgesic activity, efficacy by the oral route and persistent effects with repeated administration. Side effects include detection of miotic and respiratory-depressant effects for more than 24 hours after a single dose, and marked sedation is seen in some patients. Effects on cough, bowel motility, biliary tone and the secretion of pituitary hormones are qualitatively similar to those of morphine. In contrast to morphine, codeine is approximately 60% as effective orally as parenterally, both as an analgesic and as a respiratory depressant.

[0017] Codeine has an exceptionally low affinity for opioid receptors, and the analgesic effect of codeine is due to its conversion to morphine. However, codeine's antitussive actions probably involve distinct receptors that bind codeine specifically.

[0018] Propoxyphene produces analgesia and other CNS effects that are similar to those seen with morphine. It is likely that at equianalgesic doses the incidence of side effects such as nausea, anorexia, constipation, abdominal pain, and drowsiness would be similar to those of codeine.

[0019] Opioid antagonists have therapeutic utility in the treatment of overdosage with opioids. As understanding of the role of endogenous opioid systems in pathophysiological states increases, additional therapeutic indications for these antagonists will emerge. If endogenous opioid systems have not been activated, the pharmacological actions of opioid antagonists depend on whether or not an opioid agonist has been administered previously, the pharmacological profile of that opioid and the degree to which physical dependence on an opioid has developed. The antagonist naloxone produces no discernible subjective effects aside from slight drowsiness. Naltrexone functions similarly, but with higher oral efficacy and a longer duration of action. Currently, naloxone and naltrexone are used clinically to treat opioid overdoses. Their potential utility in the treatment of shock, stroke, spinal cord and brain trauma, and other disorders that may involve mobilization of endogenous opioids remains to be established.

[0020] The complex interactions of morphine and drugs with mixed agonist/antagonist properties are mediated by multiple classes of opioid receptors. Opioid receptors com-

prise a family of cell surface proteins, which control a range of biological responses, including pain perception, modulation of affective behavior and motor control, autonomic nervous system regulation and neuroendocrinological function. There are three major classes of opioid receptors in the CNS, designated mu, kappa and delta, which differ in their affinity for various opioid ligands and in their cellular distribution. The different classes of opioid receptors are believed to serve different physiologic functions. Olson et al. (1989) Peptides 10:1253; Lutz and Pfister (1992) J. Receptor Res. 12:267; and Simon (1991) Medicinal Res. Rev. 11:357. Morphine produces analgesia primarily through the mu-opioid receptor. However, among the opioid receptors, there is substantial overlap of function as well as of cellular distribution.

[0021] The mu-opioid receptor mediates the actions of morphine and morphine-like opioids, including most clinical analgesics. In addition to, morphine, several highly selective agonists have been developed for mu-opioid receptors, including [D-Ala², MePhe⁴, Gly(ol)⁵]enkephalin (DAMGO), levorphanol and methadone. Differential sensitivity to antagonists, such as naloxonazine, indicates the pharmacological distinctions between the mu-opioid receptor subtypes, mu₁ and mu₂. Several of the opioid peptides will also interact with mu-opioid receptors.

[0022] There are three distinct families of endogenous opioid peptides, the enkephalins, endorphins and dynorphins, where each peptide is derived from a distinct precursor polypeptide. Mu-opioid receptors have a high affinity for the enkephalins as well as β -endorphin and dynorphin A. For review, see Reisine and Pasternak (1996).

[0023] Members of each known class of opioid receptor have been cloned from human cDNA and their predicted amino acid sequences have been determined. Yasuda et al. (1993) Proc. Natl. Acad. Sci. U.S.A. 90:6736; and Chen et al. (1993) Mol. Pharmacol. 44:8. The opioid receptors belong to a class of transmembrane spanning receptors known as G-protein coupled receptors. G-proteins consist of three tightly associated subunits, alpha, beta and gamma (1:1:1) in order of decreasing mass. Following agonist binding to the receptor, a conformational change is transmitted to the G-protein, which causes the G-alpha subunit to exchange a bound GDP for GTP and to dissociate from the beta and gamma subunits. The GTP-bound form of the alpha subunit is typically the effector-modulating moiety. Signal amplification results from the ability of a single receptor to activate many G-protein molecules, and from the stimulation by G-alpha-GTP of many catalytic cycles of the effector.

[0024] Most opioid receptor-mediated functions appear to be mediated through G-protein interactions. Standifer and Pasternak (1997) Cell Signal. 9:237. Antisense oligodeoxy-nucleotides directed against various G-protein alpha subunits were shown to differentially block the analgesic actions of the mu-, delta-, and kappa-opioid agonists in mice. Standifer et al. (1996) Mol. Pharmacol. 50:293.

[0025] The amino acid sequences of the opioid receptors are approximately 65% identical, and they have little sequence similarity to other G-protein-coupled receptors except for somatostatin. Reisine and Bell (1993) Trends Neurosci. 16:506. The regions of highest similarity in sequence are the sequences predicted to lie in the seven transmembrane-spanning regions and the intracellular loops.

Regions of amino acid sequence divergence are the amino and carboxy termini and the second and third extracellular loops.

[0026] Each receptor subtype has a characteristic pattern of expression. Mu-opioid receptor mRNA is present in the periaqueductal gray, spinal trigeminal nucleus, cuneate and gracile nuclei, and thalamus regions of the brain involved in pain perception and associated with morphine analgesia (Defts et al. (1994) J. Comp. Neurol. 345:46); in nuclei involved in control of respiration, consistent with the ability of morphine to depress respiration; and in neurons of the area postrema, where morphine has been shown to cause nausea and induce vomiting. Other consequences of muopioid receptor activation include miosis, reduced gastrointestinal motility, and feelings of well-being or euphoria. Pasternak (1993). The pattern of mu-opioid receptor mRNA expression correlates with the brain centers involved in mediating the biological actions of morphine and mu-selective agonists. Delta-opioid receptor mRNA is found in the dorsal horn of the spinal cord. Kappa₁-opioid receptor mRNA is expressed in the hypothalamic regions, which may account for many of the neuroendocrine effects of the kappa selective agonists.

[0027] Soon after the mu-opioid receptor MOR-1 was cloned (Chen et al. (1993); and Wang et al. (1993) Proc. Natl. Acad. Sci. U.S.A. 90:10230), antisense experiments confirmed its involvement with morphine analgesia. Rossi et al. (1994) Life Sci. 54:375; and Rossi et al. (1995) FEBS Lett. 369:192. Antisense oligonucleotides directed against MOR-1 mRNA blocked the analgesic actions of morphine in rats, demonstrating that proper translation of the MOR-1 mRNA was essential for modulating morphine analgesia. Antisense approaches have also demonstrated a relationship between MOR-1 activity and ingestive responses. Administration of antisense oligonucleotides directed against MOR-1 mRNA significantly reduced food and water intake and subsequently, body weight in rats.

[0028] In recent years, a number of mu-opioid receptor subtypes have been proposed. The first suggestion of mu₁ and mu₂ receptor subtypes came from a combination of binding and pharmacological studies based on the antagonists naloxonazine and naloxazone. Wolozin and Pasternak (1981) Proc. Natl. Acad. Sci. U.S.A. 78:6181; Reisine and Pasternak (1996); and Pasternak (1993). To date, only a single mu receptor gene, MOR-1, has been identified. Min et al. (1994) Proc. Natl. Acad. Sci. U.S.A. 91:9081; Giros et al. (1995) Life Sci. 56:PL369; and Liang et al. (1995) Brain Res. 679:82. The MOR-1 cDNA consists of exons 1-4, which total 1610 bp in length and encode 398 amino acids. More recently, pharmacological and molecular differences between morphine and morphine- 6β -glucuronide (M6G) have suggested yet another mu-opioid receptor subtype. Pasternak and Standifer (1995) Trends Pharmacol. Sci. 16:344; Rossi et al. (1995); and Rossi et al. (1996) Neurosci. Lett. 216:1.

[0029] Antisense oligonucleotides directed against selected exons within the MOR-1 mRNA revealed interesting therapeutic patterns of morphine and M6G analgesia, with some MOR-1 exons implicated in the analgesic actions of one drug, but not the other. Rossi et al. (1997) J. Pharmacol. Exp. Ther. 281:109; and Rossi et al. (1995). Although the two analgesics were known to act through

different receptors, the sensitivity of the effect of both analgesics to at least six different MOR-1 antisense probes implied that both receptors were closely associated with MOR-1, raising the possibility of pharmacologically relevant MOR-1 splice variants. Pasternak and Standifer (1995); and Rossi et al. (1995). Alternative splicing has been observed with a number of G-protein-coupled receptors, including somatostatin 2 (Vanetti et al. (1998) FEBS Lett. 311:290), dopamine D2 (Guiramand et al. (1995) J. Biol. Chem. 270:7354), prostaglandin EP3 (Namba et al. (1993) Trends Pharmacol. Sci. 16:246), serotonin receptor subtypes 5-HT₄ and 5-HT₇ (Lucas and Hen. (1995) Trends Pharmacol. Sci. 16:246) and MOR-1. Bare et al. (1994) FEBS Lett. 354:213; and Zimprich et al. (1995) FEBS Lett. 359:142.

[0030] Several opioid receptor splice variants have been identified and characterized. At least two MOR-1 splice variants are known, the human MOR-1A and the rat MOR-1B_s. Bare et al. (1994); and Zimprich et al. (1995). The hMOR-1A splice variant consists of exons 1, 2, 3 and a new exon 3a, and was determined to possess ligand binding characteristics similar to the full-length MOR-1. Bare et al. (1994). The rMOR-1B_s splice variant consists of exons 1, 2, 3 and a new exon 5, and like hMOR-1A, differs from MOR-1 only in length and amino acid composition at the carboxy-terminal tail. Zimprich et al. (1995). MOR-1B, has affinity to opioid compounds similar to that of MOR-1, but is much more resistant to agonist-induced desensitization than MOR-1. The C-terminal differences between MOR-1 and MOR-1A or MOR-1Bs could have effects on receptor coupling or receptor transport and localization. The MOR-1 splice variants are potential targets for the modulation of physiological effects resulting from mu-opioid receptor activity.

[0031] Availability of polynucleotide sequences of opioid receptor splice variants, and, in the case of splice variants in coding regions, the corresponding polypeptide sequences, will significantly increase the capability to design pharmaceutical compositions, such as analgesics, with enhanced specificity of function. In general, the availability of these polynucleotide and polypeptide sequences will enable efficient screening of candidate compositions. The principle in operation through the screening process is straightforward: natural agonists and antagonists bind to cell-surface receptors and channels to produce physiological effects; certain other molecules can produce physiological effects and act as therapeutic pharmaceutical agents. Thus, the ability of candidate drugs to bind to opioid receptor splice variants can function as an extremely effective screening criterion for the selection of pharmaceutical compositions with desired functional efficacy and specificity.

DISCLOSURE OF THE INVENTION

[0032] The invention encompasses MOR-1 splice variant polypeptides or polypeptide fragments or homologs thereof retaining MOR-1 activity.

[0033] The invention further encompasses a MOR-1 splice variant polynucleotide, encoding MOR-1 splice variant polypeptides or polypeptide fragments or homologs thereof retaining MOR-1 activity, and noncoding mRNA splice variants and complementary strands thereto.

[0034] The invention further encompasses a polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, comprising at least 15 consecutive nucleotides of an MOR-1 splice variant polynucleotide where the polynucleotide contains promoter elements.

[0035] The invention further encompasses methods of screening compositions for an opioid activity by obtaining a control cell that does not express a recombinant or endogenous opioid receptor, obtaining a test cell that expresses a recombinant MOR-1 splice variant polypeptide, contacting the control cell and test cell with an amount of an opioid sufficient to exert a physiologic effect, separately measuring the physiologic effect of the composition on the control cell and test cell and comparing the physiologic effect of the sphere determination of a physiologic effect of the composition is expressed relative to that of the opioid.

[0036] The invention further encompasses methods of screening compositions for an opioid activity by obtaining a control polypeptide that is not a recombinant opioid receptor, obtaining a test polypeptide that is a recombinant MOR-1 splice variant polypeptide, contacting a composition with the control polypeptide and the test polypeptide, contacting the test polypeptide with an amount of an opioid sufficient to measurably bind the test polypeptide, measuring the binding of the composition and the opioid, and comparing the test polypeptide binding of the composition to that of the opioid, where determination of binding of the composition is expressed relative to that of the opioid.

[0037] The invention further encompasses methods of screening compositions for differential or selective opioid activity comprising obtaining a first and second test polypeptide that are MOR-1 splice variant polypeptide fragments and contacting each with a composition, measuring the binding affinity of the composition to the first and second test polypeptides and comparing the binding of the composition and the first test polypeptide to that of the second test polypeptide where differential activity is expressed as a ratio of the two binding affinities.

[0038] The invention further encompasses a non-human animal in which one or both endogenous MOR-1 alleles has been altered by homologous recombination with an exogenously introduced MOR-1 splice variant polynucleotide.

[0039] The invention further encompasses a non-human transgenic animal carrying a transgene comprising an MOR-1 splice variant polynucleotide.

[0040] The invention further encompasses a method for regulating morphine analgesia in a subject by altering the amount of MOR-1 splice variant polypeptide activity. Activity can be regulated by administering antigen binding fragments, agonists, antagonists or small molecule ligands to a subject in an amount and a duration sufficient to regulate morphine analgesia. The antigen binding fragment, agonist, antagonist or small molecule ligand is directed to an MOR-1 splice variant polypeptide fragment or MOR-1 splice variant mRNA.

[0041] The invention further encompasses regulating opioid activity by administering a DNA plasmid vector containing an MOR-1 splice variant polynucleotide. The DNA plasmid vector thereby expresses an mRNA splice variant that may encode an MOR-1 polypeptide in a subject in an amount of and a duration sufficient to regulate morphine analgesia. Activity can also be regulated by adminis-

tering an antisense nucleic acid complementary to an MOR-1 splice variant polynucleotide, thereby blocking gene expression in a subject in an amount and a duration sufficient to regulate morphine analgesia.

[0042] The invention further encompasses a method for regulating body weight in a subject by altering the amount of MOR-1 splice variant polypeptide activity in the subject. Activity can be regulated by administering antigen binding fragments, agonists, antagonists or small molecule ligands to a subject in an amount and a duration sufficient to regulate body weight. The antigen binding fragment, agonist, antagonist or small molecule ligand is directed to an MOR-1 splice variant polypeptide.

[0043] Activity can also be regulated by administering to the subject a DNA plasmid vector containing an MOR-1 splice variant polynucleotide. The DNA plasmid vector thereby expresses an MOR-1 polypeptide fragment or MOR-1 splice variant mRNA in the subject in an amount of and a duration sufficient to regulate body weight of the subject. Activity can also be regulated by administering an antisense nucleic acid complementary to an MOR-1 splice variant polynucleotide, thereby blocking gene expression in a subject in an amount and a duration sufficient to regulate body weight of the subject.

[0044] The invention further encompasses a method for diagnosing an MOR-1 splice variant-associated pharmacological abnormality, comprising measuring the amount of variant activity or tissue distribution thereof in a subject and comparing that activity or tissue distribution to a control sample, wherein a difference in the amount of activity or tissue distribution correlates with the presence of a pharmacologic defect.

[0045] The invention further encompasses a method for diagnosing an MOR-1 splice variant-associated disorder, comprising measuring the amount of variant activity or tissue distribution thereof in a subject and comparing that activity or tissue distribution to a control sample, wherein a difference in the amount of activity or tissue distribution correlates with the presence of a disorder of the neuroendocrine system.

[0046] The invention further encompasses antigen-binding fragments specific for the MOR-1 splice variant polypeptides described herein.

BRIEF DESCRIPTION OF THE DRAWINGS

[0047] FIG. 1 depicts a schematic diagram of MOR-1 gene structure and alternative splicing. Exons and introns are indicated by boxes and horizontal lines, respectively. The translational start codon and termination codon are AUG and TAA or TAG or TGA. Overlapping genomic clones covering the entire MOR-1 gene are shown by heavy horizontal lines on the top panel.

[0048] FIG. 2 depicts the MOR-1 splice variant polynucleotides. These include: MOR-1a; MOR-1bI; MOR-1g; MOR-1h; MOR-1c; MOR-1d; MOR-1e; MOR-1j; MOR-1i; MOR-1bII; MOR-1f; hMOR-1-610302; and clones 3320510, 2730510, and 161416.

[0049] FIG. 3 depicts the MOR-1 splice variant polypeptides. These include: MOR-1C; MOR-1G; MOR-1D; MOR-1E; MOR-1H; MOR-1I; MOR-1J; hMOR-1-610302; MOR- 1A; MOR-1BI; MOR-1BII; MOR-1F; and MOR-1. **FIG. 3** also designates exons where applicable.

[0050] FIG. 4 compares the amino acid sequences of several MOR-1 splice variant polypeptides predicted from the cDNA clones. All are murine variants except MOR-1a and MOR-1b which are human and rat, respectively. In **FIG. 4**, the small solid triangles represent casein kinase phosphorylation sites and the large open triangle represents a protein kinase C phosphorylation site.

[0051] FIG. 5 is a schematic diagram comparing the exons of MOR-1I and MOR-1J.

[0052] FIG. 6 is a schematic diagram comparing the exons of Clones 161416; 3320510 and 2730150.

[0053] FIG. 7 depicts the results of Northern blots performed on mouse brain using an exon 4 probe and a probe including exons 7/8/9.

[0054] FIG. 8 depicts regional distribution of the MOR-1c, MOR-1d and MOR-1e mRNA. In 8A, RT-PCR was performed on the indicated brain regions using the indicated probes. In 8B, RT-PCR was performed on the indicated brain regions using the indicated probe.

[0055] FIG. 9 depicts immunohistochemical localization of MOR-1 and MOR-1C in mouse brain. Sections A and B and Sections C and D were stained with MOR-1 and MOR-1C antisera respectively. Regions were (A and B) St, striatum; ac, anteriorcommissure; Ac, accumbens; and LS, lateral septum; (c) MD, mediodorsal thalamic nucleus; CM, centromedian thalamic nucleus; DH, dorsal hypothalamic nucleus; LH, lateral hypothalamic nucleus; Ce, central amygdaloid nucleus; Ic, intercalated amygdaloid nucleus; and Me, medial amygdaloid nucleus; and (D) Ar, arcuate nucleus; and ME, median eminence.

[0056] FIG. 10 depicts in vitro translation of MOR-1, MOR-1C, MOR-1D and MOR-1E.

[0057] FIG. 11 depicts antisense mapping of exons 6, 7, 8 and 9 of MOR-1. The solid bars represent M6G and the stippled bars represent morphine treatment.

BEST MODE FOR CARRYING OUT THE INVENTION

[0058] In view of the strong pharmacological evidence for distinct mu-opioid receptors, alternative splicing of the MOR-1 gene has been explored further. It has now been determined that the MOR-1 gene is subject to alternative splicing that produces novel splice variant forms of the mRNA and/or receptor. Eleven new exons for the MOR-1 gene have been identified, which combine to yield fifteen novel MOR-1 splice variant polynucleotides. These splice variant polynucleotides and the polypeptides encoded thereby are potential targets for modulating morphine analgesia and opioid-mediated ingestive responses.

[0059] The invention further encompasses isolated MOR-1 splice variant polynucleotide sequences indicated in FIG. 2. In addition to FIG. 2, the polynucleotide sequences can be any sequence of the appropriate genetic code to encode any of the MOR-1 splice variant polypeptides indicated in FIG. 3. Preferably, the polynucleotide is at least 15 consecutive nucleotides.

[0060] A "polynucleotide" is a polymeric form of nucleotides of any length, which contain deoxyribonucleotides, ribonucleotides, and analogs in any combination. Polynucleotides may have three-dimensional structure, and may perform any function, known or unknown. The term "polynucleotide" includes double-, single-stranded, and triplehelical molecules. Unless otherwise specified or required, any embodiment of the invention described herein that is a polynucleotide encompasses both the double stranded form and each of two complementary forms known or predicted to make up the double stranded form of either the DNA, RNA or hybrid molecule.

[0061] The following are non-limiting examples of polynucleotides: a gene or gene fragment, exons, introns, mRNA, tRNA, rRNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes and primers. A polynucleotide may comprise modified nucleotides, such as methylated nucleotides and nucleotide analogs, uracyl, other sugars and linking groups such as fluororibose and thiolate, and nucleotide branches. The sequence of nucleotides may be further modified after polymerization, such as by conjugation, with a labeling component. Other types of modifications included in this definition are caps, substitution of one or more of the naturally occurring nucleotides with an analog, and introduction of means for attaching the polynucleotide to proteins, metal ions, labeling components, other polynucleotides or solid support.

[0062] An "isolated" polynucleotide or polypeptide is one that is substantially free of the materials with which it is associated in its native environment. By substantially free, is meant at least 50%, preferably at least 70%, more preferably at least 80%, and even more preferably at least 90% free of these materials.

[0063] The invention further comprises a complementary strand to the MOR-1 splice variant polynucleotide.

[0064] The complementary strand can be polymeric and of any length, and can contain deoxyribonucleotides, ribonucleotides, and analogs in any combination.

[0065] Hybridization reactions can be performed under conditions of different "stringency." Conditions that increase stringency of a hybridization reaction are well known. See for examples, "Molecular Cloning: A Laboratory Manual", second edition (Sambrook et al. 1989). Examples of relevant conditions include (in order of increasing stringency): incubation temperatures of 25° C., 37° C., 50° C., and 68° C.; buffer concentrations of 10×SSC, 6×SSC, 1×SSC, 0.1×SSC (where SSC is 0.15 M NaCl and 15 mM citrate buffer) and their equivalent using other buffer systems; formamide concentrations of 0%, 25%, 50%, and 75%; incubation times from 5 minutes to 24 hours; 1, 2 or more washing steps; wash incubation times of 1, 2, or 15 minutes; and wash solutions of $6\times$ SSC, 1×SSC, 0.1×SSC, or deionized water.

[0066] The invention further encompasses polynucleotides encoding functionally equivalent variants and derivatives of the MOR-1 splice variant polypeptides and functionally equivalent fragments thereof which may enhance, decrease or not significantly affect properties of the polypeptides encoded thereby. These functionally equivalent variants, derivatives, and fragments display the ability to retain MOR-1 activity. For instance, changes in a DNA sequence that do not change the encoded amino acid sequence, as well as those that result in conservative substitutions of amino acid residues, one or a few amino acid deletions or additions, and substitution of amino acid residues by amino acid analogs are those which will not significantly affect properties of the encoded polypeptide. Conservative amino acid substitutions are glycine/alanine; valine/isoleucine/leucine; asparagine/glutamine; aspartic acid/glutamic acid; serine/ threonine/methionine; lysine/arginine; and phenylalanine/ tyrosine/tryptophan.

[0067] The invention further encompasses the MOR-1 splice variant polynucleotides contained in a vector molecule or an expression vector and operably linked to a promoter element if necessary.

[0068] A "vector" refers to a recombinant DNA or RNA plasmid or virus that comprises a heterologous polynucleotide to be delivered to a target cell, either in vitro or in vivo. The heterologous polynucleotide may comprise a sequence of interest for purposes of therapy, and may optionally be in the form of an expression cassette. As used herein, a vector need not be capable of replication in the ultimate target cell or subject. The term includes cloning vectors for translation of a polynucleotide encoding sequence. Also included are viral vectors.

[0069] The term "recombinant" means a polynucleotide of genomic cDNA, semisynthetic, or synthetic origin which either does not occur in nature or is linked to another polynucleotide in an arrangement not found in nature.

[0070] "Heterologous" means derived from a genetically distinct entity from the rest of the entity to which it is being compared. For example, a polynucleotide, may be placed by genetic engineering techniques into a plasmid or vector derived from a different source, and is a heterologous polynucleotide. A promoter removed from its native coding sequence and operatively linked to a coding sequence other than the native sequence is a heterologous promoter.

[0071] The polynucleotides of the invention may comprise additional sequences, such as additional encoding sequences within the same transcription unit, controlling elements such as promoters, ribosome binding sites, polyadenylation sites, additional transcription units under control of the same or a different promoter, sequences that permit cloning, expression, homologous recombination, and transformation of a host cell, and any such construct as may be desirable to provide embodiments of this invention.

[0072] A "host cell" denotes a prokaryotic or eukaryotic cell that has been genetically altered, or is capable of being genetically altered by administration of an exogenous polynucleotide, such as a recombinant plasmid or vector. When referring to genetically altered cells, the term refers both to the originally altered cell, and to the progeny thereof.

[0073] Polynucleotides comprising a desired sequence can be inserted into a suitable cloning or expression vector, and the vector in turn can be introduced into a suitable host cell for replication and amplification. Polynucleotides can be introduced into host cells by any means known in the art. The vectors containing the polynucleotides of interest can be introduced into the host cell by any of a number of appropriate means, including direct uptake, endocytosis, transfection, f-mating, electroporation, transfection employing calcium chloride, rubidium chloride, calcium phosphate, DEAE-dextran, or other substances; microprojectile bombardment; lipofection; and infection (where the vector is infectious, for instance, a retroviral vector). The choice of introducing vectors or polynucleotides will often depend on features of the host cell.

[0074] Once introduced, the exogenous polynucleotide can be maintained within the cell as a non-integrated vector (such as a plasmid) or integrated into the host cell genome. Amplified DNA can be isolated from the host cell by standard methods. See, e.g., Sambrook et al. (1989). RNA can also be obtained from transformed host cell, or it can be obtained directly from the DNA by using a DNA-dependent RNA polymerase.

[0075] Expression vectors generally are replicable polynucleotide constructs that contain a polynucleotide encoding the polypeptide of interest. Herein, this means any of the MOR-1 splice variant polypeptides. For expression, one or more translational controlling elements are also usually required, such as ribosome binding sites, translation initiation sites and stop codons. These controlling elements (transcriptional and translational) can be derived from the MOR-1 gene, or heterologous (i.e., derived from other genes or other organisms). A number of expression vectors suitable for expression in eukaryotic cells including yeast, avian, and mammalian cells are well known in the art. One example of an expression vector is pcDNA3 (Invitrogen, San Diego, Calif.), in which transcription is driven by the cytomegalovirus (CMV) early promoter/enhancer. This vector also contains recognition sites for multiple restriction enzymes for insertion of an MOR-1 splice variant polypeptide of interest. Another example of an expression vector system is the baculovirus/insect system.

[0076] Cloning and expression vectors typically contain a selectable marker (for example, a gene encoding a protein necessary for the survival or growth of a host cell transformed with the vector), although such a marker gene can be carried on another polynucleotide sequence co-introduced into the host cell. Only those host cells into which a selectable gene has been introduced will grow under selective conditions. Typical selection genes either: (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate; (b) complement auxotrophic deficiencies; or (c) supply critical nutrients not available for complex media. The choice of the proper marker gene will depend on the host cell, and appropriate genes for different hosts are known in the art. Vectors also typically contain a replication system recognized by the host.

[0077] Suitable cloning vectors can be constructed according to standard techniques, or selected from a large number of cloning vectors available in the art. While the cloning vector selected may vary according to the host cell intended to be used, useful cloning vectors will generally have the ability to self-replicate, may possess a single target for a particular restriction endonuclease, or may carry marker genes. Suitable examples include plasmids and bacterial viruses, e.g., pUC18, mp18, mp19, pBR322, pMB9, ColE1, pCR1, RP4, phage DNAs, and shuttle vectors such as pSA3 and pAT28. These and other cloning vectors are available from commercial vendors such as BioRad, Stratagene, and Invitrogen.

[0078] The invention further encompasses an isolated polynucleotide, or a complementary strand thereto that

hybridizes under stringent conditions, comprising at least 15 consecutive nucleotides of the MOR-1 splice variant polynucleotides depicted in (FIG. 2) where the polynucleotide contains promoter elements.

[0079] The MOR-1 splice variant promoter elements, are contained in exons 1a, 1b, and 1c or in any combination thereof. Promoter elements can control the level, tissue specificity, inducibility and, in gene clusters, the sequence of transcriptional activation and repression. Promoter elements include but are not limited to, enhancer sequences and repressor sequences.

[0080] The invention further encompasses non-human animals in which one or both MOR-1 alleles has been altered by homologous recombination with an exogenously introduced nucleic acid.

[0081] Non-human animals devoid of one or more gene products are generated to determine the "loss-of-function" phenotype associated with the loss of that particular gene product. Herein, the gene product is the MOR-1 gene or splice variants thereof. Phenotypic abnormalities can be present, for instance, in anatomical structures, biochemical and genetic pathways and pharmacological responses. Loss-of-function phenotypic analysis has the potential to reveal the function of the gene product.

[0082] Methods of homologous recombination with an exogenously introduced nucleic acid are used to inactivate one or more alleles in non-human animals. These methods, as applied to mice and rats, are well known in the art. Capecchi (1989) Science 244:1288. Usually, an exogenous polynucleotide encoding a selectable marker gene, and having sufficient sequence homology to the targeted site of integration at either end of the polynucleotide, is introduced into the genome of embryonic stem cells (ES cells) derived from the inner cell mass of non-human animal blastocysts. Evans and Kaufman (1981) Nature 292:154. Through homologous recombination, the polynucleotide is incorporated into the genetic locus at the targeted site of integration, replacing the corresponding sequences of the endogenous allele. ES cells are used to generate chimeric animals either by microinjection into, or aggregation with wildtype embryos. Chimeric animals having germ line transmission of the inactivated allele are bred to produce heterozygous, and subsequently, homozygous lines carrying the inactivated allele. Robertson (1991) Biol. Reprod. 44:238.

[0083] The invention further encompasses non-human transgenic animals carrying a transgenic MOR-1 splice variant polynucleotide.

[0084] Non-human animals carrying additional copies of the gene of interest are generated to determine the "gainof-function" phenotype associated with an excess of that particular gene product. Herein, the gene product is any of the MOR-1 splice variant polynucleotides. Phenotypic abnormalities can be present, for instance, in anatomical structures, biochemical and genetic pathways and pharmacological responses. Gain-of-function phenotypic analysis has the potential to reveal the function of the gene product.

[0085] Methods of generating transgenic animals are well known in the art. Jaenisch (1988) Science 240:1468. "Transgenes" are exogenous polynucleotides encoding the gene of interest. Transgenes are introduced into the embryonic genome through microinjection. Alternatively, a transgene encoding the gene of interest and a selectable marker gene is introduced into the ES cell genome through transfection or electroporation. ES cells carrying the transgene are subsequently used to produce animals with multiple copies of the gene of interest.

[0086] The invention encompasses splice variant polypeptides. The exemplary MOR-1 splice variant polypeptides are composed of the amino acids indicated in (**FIG. 3**). Polypeptide fragments comprising 5 amino acids, more preferably 7 amino acids, more preferably 15 amino acids, more preferably 25 amino acids, more preferably 50 amino acids and more preferably 75 amino acids, which are not the same as the known MOR-1 or MOR-1 variants are claimed herein and encompassed in the term "MOR-1 splice variant polypeptides." The exemplary MOR-1 splice variant polypeptide fragments retain MOR-1 activity. The complete cDNA sequences of MOR-1C, MOR-1D, and MOR-1E have been deposited in GenBank, numbers AF062752, AF062753, and AF074974 respectively, in satisfaction of the requirements of the Budapest Treaty.

[0087] The terms "protein", "peptide", "polypeptide" and "polypeptide fragment" are used interchangeably herein to refer to polymers of amino acid residues of any length. The polymer can be linear or branched, it may comprise modified amino acids or amino acid analogs, and it may be interrupted by chemical moieties other than amino acids. The terms also encompass an amino acid polymer that has been modified naturally or by intervention; for example disulfide bond formation, glycosylation, lipidation, acetylation, phosphorylation, or any other manipulation or modification, such as conjugation with a labeling or bioactive component.

[0088] The MOR-1 splice variant polypeptides retain MOR-1 activity. To "retain MOR-1 activity" is to have a similar level of functional activity as the MOR-1 polypeptide **(FIG. 3)**. This activity includes but is not limited to, immunologic and pharmacological activity.

[0089] The "immunologic activity" is binding to antiopioid receptor antigen binding fragments. The antigen binding fragments can be any functional antibody, fragment or derivative thereof, including, but not limited to, whole native antibodies, bispecific antibodies, chimeric antibodies, Fab, F(ab')2, single chain V region fragments (scFv), and fusion polypeptides comprising an antigen binding fragment fused to a chemically functional moiety.

[0090] The "pharmacologic activity" is activation or deactivation of the MOR-1 splice variant polypeptides upon binding of agonists or antagonists.

[0091] The invention further encompasses MOR-1 splice variant polypeptide homologs. A "homolog" is a polypeptide similar in amino acid sequence to other polypeptides among a single species or, a "homolog" in evolution is a polypeptide similar in amino acid sequence to other polypeptides in different species because they have been inherited from a common ancestor. Preferably, homologs of the present invention are human homologs.

[0092] Isolation of MOR-1 splice variant human homolog cDNAs can be carried out by any method known in the art. For instance, methods analogous to the isolation of the mouse MOR-1 splice variants described herein (see Example 1). Using primers corresponding to the human MOR-1 gene and a Marathon-Ready human cDNA Library

to carry out reactions according to the Marathon cDNA Amplification Kit (Clontech), human MOR-1 splice variants can be obtained. Alternatively, screening of human cDNA libraries with probes corresponding to mouse MOR-1 splice variant sequences can be carried out at reduced stringency to identify human MOR-1 splice variant cDNAs.

[0093] The invention further encompasses the MOR-1 splice variant polypeptides in a heterodimeric or homodimeric form. A "heterodimer" is a protein made up of more than one kind of polypeptide. A "homodimer" is a protein made up of more than one kind of polypeptide.

[0094] Pharmaceutical compositions and treatment modalities can be detected by the methods of this invention. The MOR-1 splice variant polypeptide fragments and MOR-1 splice variant nucleic acid sequences can be used in screening for compositions that alter variant activity. Compositions that selectively regulate the MOR-1 splice variant polypeptide fragments or selectively modulate physiological processes can be identified.

[0095] The invention further encompasses methods of screening compositions for opioid activity by obtaining a control cell that does not express a recombinant opioid receptor and obtaining a test cell that is the same as the control cell except that it expresses a recombinant MOR-1 splice variant polypeptide, contacting the control cell and test cell with an amount of an opioid sufficient to exert a physiologic effect, separately measuring the physiologic effect of the composition on the control cell and test cell and comparing the physiologic effect of the opioid, where determination of a physiologic effect of the composition is expressed relative to that of the opioid.

[0096] The invention further comprises a method of screening compositions for opioid activity by obtaining a control polypeptide that is not a recombinant opioid receptor and obtaining a test polypeptide, contacting a composition with the control polypeptide and the test polypeptide, contacting the test polypeptide with an amount of an opioid sufficient to measurably bind the test polypeptide, measuring the binding of the composition and the opioid and comparing the test polypeptide binding of the composition to that of the opioid, where determination of binding of the composition is expressed relative to that of the opioid.

[0097] The invention further encompasses a method of screening compositions for differential opioid activity by obtaining a first test polypeptide that is an MOR-1 splice variant polypeptide and contacting it with a composition and obtaining a second test polypeptide that is an MOR-1 splice variant polypeptide, measuring the binding of the composition to the first and second test polypeptides, and comparing the binding of the composition and the first test polypeptide to that of the second test polypeptide where differential activity is expressed as a ratio of the two binding affinities.

[0098] The compositions screened include but are not limited to chemical, synthetic combinatorial libraries of small molecule ligands, eukaryotic whole cell lysates or extracts, media conditioned by cultured eukaryotic cells, natural products and extracts thereof.

[0099] The opioid can be but is not limited to, morphine, methadone, etorphine, levorphanol, fentanyl, sufentanil,

[D-Ala², MePhe⁴, Gly(ol)⁵]enkephalin (DAMGO), pentazocine, ethylketocyclazocine, bremazocine, spiradoline, [D-Ser², Leu⁵]enkephalin-Thr⁶ (DSLET), Met-enkephalin, Leu-enkephalin, β -endorphin, dynorphin A, dynorphin B, α -neoendorphin analogs and combinatorial chemistry products thereof.

[0100] The physiological effect can be measured by any method known in the art such as changes in the levels of neuroendocrine hormones, including, but not limited to prolactin, growth hormone, gonadotropin-releasing hormone, adrenocorticotropin, corticotropin-releasing factor, luteinizing hormone, follicle stimulating hormone, testosterone or cortisol. The physiological effect can also be measured by changes in the levels of neurotransmitters, including but not limited to, acetylcholine or dopamine.

[0101] Activation of an MOR-1 receptor, and likely, the MOR-1 splice variant polypeptides, stimulates a variety of physiological responses, including analgesia, depression of gastrointestinal motility and respiration, and alterations of the immune, endocrine and autonomic nervous system. Compositions that regulate the activity of the MOR-1 receptor and/or the MOR-1 splice variant polypeptides can elicit responses that have therapeutic effects. The invention is useful in diagnosis, treatment, design and screening of novel reagents. Screening of compounds can result in obtaining those with differential or selective activity. That is, for instance, certain compositions can retain analgesic effects but do not affect peristaltic activity and thus do not cause constipation. Conversely, compositions that lack analgesic effects but affect peristaltic activity would be useful in treating chemotherapy and HIV patients. Other applications relating to the side effects of opiates can be readily envisaged by one of skill in the art.

[0102] The invention further encompasses a method for regulating morphine analgesia in a subject by altering the amount of MOR-1 splice variant polypeptide activity in the subject. Activity can be regulated by administering antigen binding fragments, agonists, antagonists or small molecule ligands to a subject in an amount and a duration sufficient to regulate morphine analgesia. The antigen binding fragment, agonist, antagonist or small molecule ligand is directed to an MOR-1 splice variant.

[0103] Activity can also be regulated by administering a DNA plasmid vector containing an MOR-1 splice variant polynucleotide. The DNA plasmid vector thereby expresses an MOR-1 splice variant polynucleotide in a subject in an amount and a duration sufficient to regulate morphine analgesia. Activity can also be regulated by administering an antisense nucleic acid complementary to an MOR-1 splice variant polynucleotide, thereby blocking gene expression in a subject in an amount and a duration sufficient to regulate morphine analgesia.

[0104] The invention further encompasses a method for regulating body weight in a subject by altering the amount of MOR-1 splice variant polypeptide activity. Activity can be regulated by administering antigen binding fragments, agonists, antagonists or small molecule ligands to a subject in an amount and a duration sufficient to regulate body weight. The antigen binding fragment, agonist, antagonist or small molecule ligand is directed to or specific for an MOR-1 splice variant polypeptide.

[0105] Activity can also be regulated by administering to a subject a DNA plasmid vector containing an MOR-1 splice

variant polynucleotide. The plasmid vector thereby expresses the MOR-1 splice variant polynucleotide in a subject in an amount and a duration sufficient to regulate body weight of the subject. Activity can also be regulated by administering an antisense nucleic acid complementary to an MOR-1 splice variant polynucleotide, thereby blocking gene expression in a subject in an amount and a duration sufficient to regulate body weight.

[0106] Agonists and antagonists of MOR-1 splice variant polypeptide activity can include but are not limited to, morphine, methadone, etorphine, levorphanol, fentanyl, $MePhe^4$, Gly(ol)5]enkephalinsufentanil, $[D-Ala^2,$ (DAMGO), butorphanol, naloxone, naltrexone, D-Phe-Cys-Tyr-D-Trp-Orn-Thr-Pen-Thr-NH₂ (CTOP), diprenorphine, β-funaltrexamine, naloxonazine, nalorphine, pentazocine, nalbuphine, benzoylhydrazone, bremazocine, ethylketocyclazocine, U50488, U69593, spiradoline, naltrindole, [D-Pen², D-Pen⁻⁵]enkephalin (DPDPE), [D-Ala², Glu⁴]deltorphin, [D-Ser², Leu⁵]enkephalin-Thr⁶ (DSLET), Met-enkephalin, Leu-enkephalin, β -endorphin, dynorphin A, dynorphin B, α -neoendorphin and derivatives such as those produced by combinatorial chemistry.

[0107] A "subject" is a vertebrate, preferably a mammal, and more preferably a human. Mammals include but are not limited to humans, farm animals, sport animals, and pets.

[0108] The invention further encompasses a method for diagnosing an MOR-1 splice variant-associated pharmacological abnormality in a subject, comprising measuring the amount of polypeptide activity or tissue distribution of polypeptide and/or polynucleotide in the subject and comparing that activity or tissue distribution to a control sample, wherein a difference in the amount of activity or tissue distribution correlates with the presence of a pharmacological defect. This disorder can be heritable.

[0109] The invention further encompasses a method for diagnosing an MOR-1 splice variant-associated disorder of the neuroendocrine system of a subject, comprising measuring the amount of polypeptide activity or tissue distribution of polypeptide and/or polynucleotide thereof in the subject and comparing that activity or tissue distribution to a control sample, wherein a difference in the amount of activity or tissue distribution correlates with the presence of a disorder of the neuroendocrine system. This disorder can be heritable.

[0110] The invention further encompasses antigen binding fragments specific for an MOR-1 splice variant polypeptide. According to the invention, an MOR-1 splice variant polypeptide can be used as an immunogen to generate antigen binding fragments which immunospecifically bind the immunogen.

[0111] Production of antigen binding fragments such as polyclonal antibodies can be carried out by any method known in the art. Various host animals can be immunized by injection with the immunogen, including but not limited to rabbits, mice and rats. Various adjuvants can be used to increase the immunological response, depending on the host species, and including but not limited to Freund's (complete or incomplete) adjuvant, mineral gels such as aluminum hydroxide, surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, keyhole limpet hemocyanins, dinitrophenol, and potentially

useful human adjuvants such as BCG (Bacille Calmette-Guerin) and *Corynebacterium parvum*.

[0112] For preparation of antigen binding fragments such as monoclonal antibodies, any technique which provides for the production of antibody molecules by continuous cell lines in culture can be used. Examples of such techniques include the original hybridoma technique (Kohler and Milstein (1975) Nature 256:495) as well as the trioma technique, the human B-cell hybridoma technique (Kozbor et al. (1983) Immunol. Today 4:72), and the EBV hybridoma technique to produce human monoclonal antibodies (Cole et al. (1985) in Monoclonal Antibodies and Cancer Therapy, Alan R. Liss, Inc., pp. 77-96). Monoclonal antibodies can also be produced in germ-free animals utilizing known technology (PCT/US90/02545). Human antibodies can be obtained using human hybridomas (Cote et al. (1983) Proc. Natl. Acad. Sci. U.S.A. 80:2026), or by transforming human B cells with EBV virus in vitro (Cole et al. (1985)). Techniques developed for the production of "chimeric antibodies" (Morrison et al. (1984) Proc. Natl. Acad. Sci. U.S.A. 81:6851; Neuberger et al. (1984) Nature 312:604; and Takeda et al. (1985) Nature 314:452) by splicing the genes from a mouse antibody molecule specific for MOR-1 splice variants together with genes from a human antibody of appropriate biological activity can be used.

[0113] Techniques described for the production of single chain antibodies (U.S. Pat. No. 4,946,778) can be adapted to produce MOR-1 splice variant polypeptide-specific single chain antibodies. Techniques described for the production of Fab expression libraries (Huse et al. (1989) Science 246:1275) can be utilized, allowing rapid and easy identification of monoclonal Fab fragments specific for an MOR-1 splice variant polypeptide.

[0114] Antibody fragments which contain the idiotype of the molecule can be generated by known techniques. For example, such fragments include but are not limited to: the F(abl), fragment which can be produced by pepsin digestion of the antibody molecule; the Fab' fragments which can be generated by reducing the disulfide bridges of the F(abl) fragment, the Fab fragments which can be generated by treating the antibody molecule with papain and a reducing agent, and Fv fragments.

[0115] Single chain V region fragments ("scFv") can also be produced. Single chain V region fragments are made by linking L (light) and/or H (heavy) chain V (variable) regions by using a short linking peptide. Bird et al. (1988) Science 242:423. Any peptide having sufficient flexibility and length can be used as a linker in a scFv. Usually the linker is selected to have little to no immunogenicity. An example of a linking peptide is (GGGGS)₃, which bridges approximately 3.5 nm between the carboxy terminus of one V region and the amino terminus of another V region. Other linker sequences can also be used, and can provide additional functions, such as for attaching a drug or a solid support.

[0116] All or any portion of the H or L chain can be used in any combination. Typically, the entire V regions are included in the scFv. For instance, the L chain V region can be linked to the H chain V region. Alternatively, a portion of the L chain V region can be linked to the H chain V region, or a portion thereof. Also contemplated are scFvs in which the H chain V region is from H11, and the L chain V region is from another immunoglobulin. It is also possible to construct a biphasic, scFv in which one component is an MOR-1 splice variant polypeptide and another component is a different polypeptide, such as a T cell epitope.

[0117] The scFvs can be assembled in any order, for example, V_{H} -(linker)- V_{L} or V_{L} -(linker)- V_{H} . There may be a difference in the level of expression of these two configurations in particular expression systems, in which case one of these forms may be preferred. Tandem scFvs can also be made, such as (X)-(linker)-(X)-linker-(X), in which X are MOR-1 splice variant polypeptides, or combinations of MOR-1 splice variant polypeptides with other polypeptides. In another embodiment, single chain antibody polypeptides have no linker polypeptide, or just a short, inflexible linker. Exemplary configurations include V_L - V_H and V_H - V_L . The linkage is too short to permit interaction between V_L and V_H within the chain, and the chains form homodimers with a V_L/V_H antigen binding site at each end. Such molecules are referred to in the art as "diabodies".

[0118] ScFvs can be produced either recombinantly or synthetically. For synthetic production of scFv, an automated synthesizer can be used. For recombinant production of scFv, a suitable plasmid containing a polynucleotide that encodes the scFv can be introduced into a suitable host cell, either eukaryotic, such as yeast, plant, insect or mammalian cells, or prokaryotic, such as *Escherichia coli*, and the protein expressed by the polynucleotide can be isolated using standard protein purification techniques.

[0119] A particularly useful system for the production of scFvs is plasmid pET-22b(+) (Novagen, Madison, Wis.) in *E. coli.* pET-22b(+) contains a nickel ion binding domain consisting of 6 sequential histidine residues, which allows the expressed protein to be purified on a suitable affinity resin. Another example of a suitable vector is pcDNA3 (Invitrogen, San Diego, Calif.), described above.

[0120] The following examples are provided to illustrate but not limit the claimed invention.

EXAMPLE 1

[0121] Identification of MOR-1C, MOR-1D, and MOR-1E cDNA Sequences

[0122] The cDNA clones of the MOR-1 splice variants MOR-1c, MOR-1d, and MOR-1e were isolated using 3'-Rapid Amplification of cDNA Ends (RACE) and Reverse Transcription Polymerase Chain Reaction (RT-PCR). First, standard PCR reactions were performed using a Marathon cDNA Amplification Kit (Clontech) and a Marathon-Ready mouse cDNA Library. A sense primer located at the 3'-end of exon 3, nucleotide position 1338 to 1359 of the mouse mu-opioid receptor, and an antisense adapter primer were used to PCR amplify a mouse brain cDNA template. The PCR products were separated on an agarose gel. Multiple bands were amplified and each band was excised. Individual bands were amplified using a second set of nested primers, including a sense primer located at position 1394-1412 of the MOR-1 receptor, and an antisense adapter primer. The resulting PCR fragments were then subcloned into Bluescript plasmids and sequenced.

[0123] The sequence of one clone, 110222, was approximately 500 bp in length and failed to align with the sequence of MOR-1 (GenBank Accession #U26915). Clone 110222

contained partial 3' MOR-1 exon 3 sequences followed by a novel sequence. The new sequence was 454 bp long and open reading frame analysis predicted 7 amino acids beyond exon 3 followed by a termination codon and a 3' untranslated region (UTR).

[0124] To obtain full length cDNA clones of the 110222 variant, a sense primer corresponding to the 5' UTR of MOR-1, nucleotide position 217 to 240, and an antisense primer corresponding to the 3' UTR of the new sequence, antisense primer A (5' CCA CAC TGC TCA CCA GCT CAT CCC 3'), were used in RT-PCR amplification of mouse brain RNA. Three fragments of approximately 1.3, 1.4 and 1.5 kb in length, respectively, were obtained, subcloned into pCRII-ToPo plasmid (Invitrogen, Carlsbad, Calif.) and sequenced in both directions.

[0125] The three clones obtained are named MOR-1c, MOR-1d and MOR-1e. Through sequence analysis it was determined that all three clones contain the same coding exons 1, 2 and 3 from MOR-1, with novel sequences beginning downstream of exon 3. In addition, MOR-1c and MOR-1e contain an alternate exon. MOR-1d aligned with the original clone 110222. MOR-1c contains an 89 bp insertion between exon 3 and the 454 bp sequence identified in MOR-1d. MOR-1e has a 209 bp insertion between exon 3 and the 454 bp sequence identified in MOR-1d, making it the longest novel sequence. The last 89 bp in this insertion are identical to the 89 bp sequence found in MOR-1c. (FIG. 2).

[0126] The three new variants are derived from combinations of five newly discovered exons located downstream from the original MOR-1exon 4. Exon 6 is 120 bp, exon 7 is 89 bp, exon 8 is 66 bp, and exon 9 is the longest, 388 bp. MOR-1d encodes exons 1, 2, 3, 8 and 9 (FIG. 2), MOR-1c encodes exons 1, 2, 3, 7, 8 and 9 (FIG. 2), and MOR-1e encodes exons 1, 2, 3, 6, 7, 8, and 9. (FIG. 2). All of the new exons have flanking sequences that are consistent with consensus splice junctions. Thus, the MOR-1 gene consists of nine exons spanning at least 200 kb. This is depicted in FIG. 1.

[0127] The predicted amino acid sequences for these new variants differ from MOR-1 and from each other. MOR-1 has 12 predicted amino acids. MOR-1d has only 7 predicted amino acids. Although MOR-1c contains the same new sequence found in MOR-1d, the 89 bp insertion produces a reading-frame shift. As a result, open reading frame analysis of MOR-1c predicts 52 amino acids, which do not include the amino acid sequence from MOR-1d. The termination codon in MOR-1e is found in exon 6, therefore exons 7, 8 and 9 are not translated and MOR-1e is translated into only 15 amino acids.

[0128] Partial human mu opioid splice variant sequences were obtained using an RT-PCR approach. We amplified a cDNA fragment from human brain which contained an alternatively spliced exon 4 of human MOR-1 gene. In the PCR reaction, template was the first-strand cDNA reverse-transcribed from human brain mRNA, and a sense primer derived from exon 3 of the human MOR-1 gene, and an antisense primer derived from exon 7 of the mouse MOR-1 gene (5'TGT CCA TGC AAC TCT TGC AGG GTT TTT CAA CAT GAG TCG GAG AAG GAT3'). Sequence analysis of the fragment indicated that it contains the human exon 3 sequences from the sense primer to the end of exon 3 and

a 104 bp new sequence between exon 3 and the mouse antisense primer. (**FIG. 2**). Translation of the sequence from exon 3 into the new sequence indicates that it encodes 34 AA with no homology to any mouse variants. (**FIG. 3**). However it does not contain a stop codon, which suggests there is more downstream exon sequence. The new sequence has been mapped, in a human genomic BAC clone to 10 kb downstream of human exon 4.

Cloning Strategy for MOR-1I and MOR-1J

[0129] A sense primer designed from exon 3 (5'GGG AAC ACC CCT CCA CGG3') and an antisense primer from exon 5a (5'GGT GTG CTT CTC CCA GTT CTG TGT3') were used in RT-PCR of mouse brain RNA. Two fragments of approximately 0.2 and 0.7 kb in length, respectively, were obtained, subcloned into pcRIIToPo plasmid and sequenced. Sequence analysis indicates that the 0.2 kb fragment, MOR-1I, contains exons 3 and 5a except that there is a 94 kb insertion, exon 11, between exons 3 and 5a. (**FIG. 2**). Exon 11 only encodes 2AA (CV). The 0.7 kb fragment, MOR-1I, also contains exons 3 and 5a sequences, but there is a 617 bp insertion, exon 12, between exons 3 and 5a. Exon 12 encodes 7 AA. (**FIGS. 3 and 5**). Cloning strategy for 161416

[0130] A sense primer designed from exon 1a (5'CCT CCA GGC TCA TTT CAG AGA GA3') and an antisense primer from exon 1 (5'CAG GAA GTT TCC AAA GAG GCC C3') were used in RT-PCR of mouse brain RNA. The PCR fragment obtained was subcloned into pcRIIToPo plasmid and sequenced. Sequence analysis of the fragment indicates that there is a 127 bp insertion sequence, exon 1b, between exons 1a and 1. (FIG. 2). Cloning strategy for 2730510 and 3320510

[0131] The sense primer above (exon 1a) and an antisense primer from exon 2 (5'GGG CAG GTG GTA GTG GCT AAG GC3') were used in RT-PCR of mouse brain RNA. Two fragments of approximately 0.26 and 0.6 kb in length, respectively, were obtained, subcloned into pcRIIToPo plasmid and sequenced. Sequence analysis indicated that the 0.26 kb fragment, clone 2730510, contains both exons 1a, 2 and 1b, with exon 1b between exons 1a and 2. The clone 3320510, however, contains exons 1a, 1c, 1 and 2. (FIGS. 2 and 6).

Cloning Strategy for mMOR-1BI and mMOR-1BII

[0132] Mouse exon 5a sequence was obtained by sequencing mouse Genome BAC clone A using primers derived from rat MOR-1B sequences (Zimprich et al. (1995)). Then an antisense primer designed from the mouse exon 5a and a sense primer from the 5' UTR of MOR-1 nucleotide position 217 to 240 were used in RT-PCR amplification of mouse brain RNA. Two fragments of approximately 1.3 and 2.0 kb in length, respectively, were obtained, subcloned into PCRII-ToPo plasmid and sequenced. Sequence analysis of the fragments indicated that similar to rat MOR-1B, the 1.3 kb fragment contains exons 1, 2, 3 and 5a which encodes 5 AA (KIDLE). However, the 2.0 kb fragment had the same exons 1, 2, 3 and 5a, except that there is a 699 bp insertion sequence, exon 5b, between exons 3 and 5a. Exon 5b encodes 23 AA KLLMWRAMPTFKRHLAIMLSLDN. (FIGS. 2 and 3).

Cloning Strategy for mMOR-1A

[0133] First, we obtained mouse exon 3 a sequence by sequencing mouse Genomic BAC clone A with exon 3

primers. The full length cDNA of mMOR-1A was then obtained by RT-PCR using the first-strand cDNA reverse-transcribed from mouse brain total RNA as template. A sense primer corresponding to the 5' UTR of MOR-1, nucleotide position 217 to 240, and an antisense primer corresponding to the 3' UTR of exon 3a (5'GAT CAG AAT TTG GTG CCC TAC TCC CTC TCT3') were used in PCR. The PCR fragment was subcloned into pcRIIToPo plasmid and sequenced. Sequence analysis of the fragment showed that exon splice pattern was exons 1, 2,,3 and 3 a which encodes 4AA (VCAF). (FIGS. 2 and 3).

EXAMPLE 2

Mapping of the MOR-1 Gene to Mouse Chromosome 10

[0134] In order to obtain genomic clones containing the full-length MOR-1 gene, two mouse genomic BAC libraries (Genome Systems, St. Louis, Mo. and Research Genetics, Huntsville, Ala.) and a mouse genomic P1 library (Genome Systems) were screened using either PCR or standard hybridization methods. Initially, BAC clone A(~75 kb), was obtained from the Genome Systems BAC library using MOR-1 exon 4 primers for PCR amplification. BAC clone A contained only MOR-1 exons 1, 2, 3 and 4. Since no positive clones were obtained by screening the BAC library with a probe corresponding to exons 8 and 9, we screened the P1 library with this probe and obtained P1 clone A(~100 kb in length). P1 clone A contained exon 8 and 9 sequences, however, it shared no overlapping sequences with either BAC clone A or exons 6 and 7. To identify a clone containing these insertions, a second mouse BAC library (Research Genetics, Inc.) was screened by hybridization with a probe corresponding to the insertional sequences (exon 6). One new clone, BAC clone B (~120 kb) contained exons 4, 6 and 7. Alignment of the three genomic clones predicted an MOR-1 gene of approximately 230 kb.

[0135] Chromosomal localization of P1 clone A was carried out using FISH methods developed by Genome Systems, Inc. P1 clone A was labeled with digoxigenin dUTP and hybridized to metaphase chromosomes derived from a mouse embryo fibroblast cell line. Specific hybridization signals were detected by incubating the hybridized slides in fluoresceinated anti-digoxigenin antibodies followed by counterstaining with DAPI. The initial experiment resulted in specific labeling of the proximal portion of a medium sized chromosome, identified as chromosome 10 on the basis of DAPI staining. Cohybridization of a specific probe for the telomeric region of chromosome 10 with the P1 clone A demonstrated conclusively that the P1 clone A was located immediately adjacent to the heterochromatic euchromatic boundary of chromosome 10, an area corresponding to band 10A2. A total of 80 metaphase cells were analyzed with 68 exhibiting specific labeling. Genome Systems Inc. used interphase FISH analysis (van den Engh et al. (1992) Science 257:1410) to estimate the physical distance between the BAC clone A and the P1 clone A. The distance between the BAC clone A and the P1 clone A estimated was approximately 250 kb, with a possible error of approximately 30%. This was in agreement with the size derived from the overlapping genomic clones, which predicted an MOR-1 gene of approximately 230 kb. (FIG. 2)

EXAMPLE 3

Expression Patterns of the MOR-1 Variants

[0136] To determine the lengths of the mRNA transcripts encoding the MOR-1 variants, Northern blot analysis was performed as described previously by Pan et al. (1994). Total RNA was isolated from mouse brain using the guanidinium thiocyanate phenol-chloroform extraction method. Samples of total brain RNA (50 μ g) were separated on a 0.8% formaldehyde agarose gel, and transferred to a Gene Plus membrane. The membrane was hybridized with ³²P-labeled fragments corresponding to sequences from exons 7, 8, and 9 of the MOR-1 variants.

[0137] Northern analysis of the variants indicates mRNA transcripts ranging in size from approximately 6 to 9 kb (**FIG. 7**). A probe specific for exon 7 would detect only MOR-1c and MOR-1e. A probe specific for exon 6 fails to detect MOR-1e mRNA.

[0138] The regional pattern of MOR-1 variant mRNA expression was determined using RT-PCR analysis. Total RNA was extracted from multiple mouse brain regions as described above and reverse-transcribed with Super Script II Reverse Transcriptase (GIBCO) in the presence of random hexamers. RNA loading was estimated by comparison to a parallel PCR reaction using β_2 -microglobulin primers (ClonTech). The agarose gel was stained with ethidium bromide and photographed by a Kodak DC 120 Digital Camera and Imagine System. Three major bands were amplified and the predicted sizes of the PCR products for MOR-1c, MOR-1d and MOR-1e are 246 bp, 157 bp and 366 bp, respectively. Each band was extracted from the agarose gel, subcloned into a pCRII-ToPo plasmid and sequenced, confirming that the amplification products corresponded to their respective variants.

[0139] MOR-1c is the predominant isoform in all of the brain regions examined, but the relative expression of the other variants varied widely (**FIG. 8A**). MOR-1 was expressed in all regions (**FIG. 8B**). MOR-1e and MOR-1d display differential patterns of expression. In the thalamus, there is little evidence for either MOR-1d or MOR-1e expression. MOR-1c mRNA is predominant in the spinal cord, with lower levels of MOR-1e expression and no observable MOR-1d expression present. In contrast, the periaqueductal gray (PAG) and striatum, all three of the variants are detected, with the highest levels of expression displayed by MOR-1c, followed by MOR-1e and then MOR-1d. The cortex has comparably higher levels of MOR-1d expression than MOR-1e expression, as do the cerebellum and brainstem.

[0140] Regional distribution of MOR-1c was analyzed using a polyclonal antibody generated against a unique amino acid sequence in this variant. Mouse brains were sectioned and immunostaining for MOR-1 and MOR-1c determined as described. Abbadie et al. (1996) Neuroscience 70:201; Abbadie et al. (1999) Proc. Natl. Acad. Sci. U.S.A. 96:260); and Abbadie et al. (1999) submitted. Sections A and B and sections C and D were stained with MOR-1 and MOR-1C antisera, respectively. Regions were as follows: A and B) St, striatum; ac, anterior commissure; Ac, accumbens; LS, lateral septum; C) MD, mediodorsal thalamic nucleus; CM, centromedial thalamic nucleus; DH, dorsal hypothalamic nucleus; VMH, ventromedial hypothalamic

nucleus; LH, lateral hypothalamic nucleus; Ce, central amygdaloid nucleus; Ic, intercalated amygdaloid nucleus; Me, medial amygdaloid nucleus; D) Ar, arcuate nucleus; ME, median eminence. Western blotting showed that the polyclonal antibody recognized MOR-1C, but not MOR-1 obtained from transfected cells.

[0141] Sections through the striatum (FIGS. 9A and B) demonstrate marked differences between MOR-1 and MOR-1c. MOR-1 immunolabeling is observed in patches in the striatum, as well as in the subcallosal streak. Dense areas of labeling are also seen in the nucleus accumbens. MOR-1c antiserum fails to label these areas. There is MOR-1c immunoreactivity in regions of the lateral septum which have minimal staining with MOR-1 antiserum. The hypothalamus has significant differences between the two antisera (FIGS 9C and D). While there is some MOR-1 staining, MOR-1c immunoreactivity is far more intense in the arcuate nucleus and median eminence. Additional studies show intense MOR-1c immunoreactivity in the trigeminal tract and the dorsal horn of the spinal cord, as well as the PAG.

EXAMPLE 4

Binding Activity of the Variants

[0142] The cDNA fragments containing the full length MOR-1 or the MOR-1 variants in pCRII-ToPo were subcloned into pcDNA3.1 (Invitrogen, Carlsbad, Calif.), a mammalian expression vector. Synthesis of MOR-1C, MOR-1D, and MOR-1E full-length proteins was carried out in vitro using a TNT coupled reticulocyte lysate kit (Promega, Madison, Wis.). MOR-1/pcDNA3, MOR-1c/ pcDNA3, and MOR-1d/pcDNA3 plasmids were incubated with T7 RNA polymerase and reticulocyte lysate in the presence of 0.04 mCi of [³⁵S]methionine (>1000 Ci/mmol; DuPont-NEN, Boston, Mass.) at 300C. for 1 hour. The translation products were separated by a 12.5% SDS-PAGE gel, which was treated with Amplify (Amersham Life Science), dried and exposed on Kodak BioMax MR film. The MOR-1D and MOR-1E variants had molecular weights similar to that of MOR-1, while the size of MOR-1C was larger than the others, as expected based upon the predicted amino acid sequences (FIGS. 3 and 10).

[0143] CHO cells were stably transfected with plasmids MOR-1/pcDNA3, MOR-1c/pcDNA3, MOR-1d/pcDNA3 or MOR-1e/pcDNA3 using LipofectAMINE reagents (GIBCO, Gaithersburg, Md.). Stable transformants were subcloned two weeks after selection with G418 and positive clones were identified using a ³H-DAMGO binding assay.

[0144] To examine opioid binding, membranes were prepared from pcDNA3 stable transformants as described previously by Pan et al. (1994); and Pan et al. (1996). ³H-DAMGO binding was performed at 25° C. for 60 minutes in 50 mM potassium phosphate buffer, pH 7.4, containing 5 mM magnesium sulfate. Specific binding was defined as the difference between total binding and nonspecific binding, as indicated by levallorphan $(1 \,\mu\text{M})$. K_D and K_I values were calculated by nonlinear regression analysis (Prism, Graph Pad Software). Protein concentrations were determined against bovine serum albumin as the standard curve. Lowry et al. (1951) J. Biol. Chem. 193:265.

[0145] Saturation studies were performed and the binding parameters established by nonlinear regression analysis. ³H-DAMGO binding was examined in stable lines expressing either MOR-1 or MOR-1C.

[0146] In saturation studies ³H-DAMGO displays high affinity for all the variants (Table 1). Indeed, the new variants bind ³H-DAMGO with higher affinities than MOR-1. Results are reported as the means \pm s.e.m of at least 3 independent determinations.

TABLE 1

Clone	$K_{D}\left(nM ight)$
MOR-1 MOR-1C MOR-1D MOR-1E	$\begin{array}{c} 1.75 \pm 0.44 \\ 0.93 \pm 0.19 \\ 0.72 \pm 0.11 \\ 1.2 \pm 0.5 \end{array}$

[0147] Competition studies were performed using at least three concentrations of the indicated competitor. ³H-DAMGO binding was performed in stable transfectants containing the indicated cDNA's. Analysis of variance was performed to determine whether there were differences among the various clones for each competitor, followed by Tukey's post hoc analysis.

[0148] In competition studies, mu ligands such as morphine, DAMGO, M6G and the endorphins bind competitively while the kappa1 opioid U50,488H and the delta opioid ligand [D-Pen², D-Pen⁵]enkephalin (DPDPE) are ineffective. However, the binding selectivity profiles among the variants are significantly different. For example, morphine competes for binding to the MOR-1D variant over 3-fold more potently than against MOR-1 itself (p<0.05). Similarly, the opioid peptide DSLET is twice as potent against binding to the MOR-1D variant than MOR-1 (p<0.05). The most dramatic differences in potency are seen with the endogenous opioids dynorphin A (p<0.0001) and β -endorphin (p<0.0003). The MOR-1D variant has the highest affinity for both dynorphin A and β -endorphin. MOR-1E also has a significantly higher affinity for β -endorphin than MOR-1. Dynorphin A has significantly higher affinity for MOR-1C and MOR-1D than either MOR-1 or MOR-1E. Through competition studies all of the variants have been classified within the mu opioid receptor family (Table 2). Results are reported as the means \pm s.e.m. of at least 3 independent determinations.

TABLE 2

	Selectivity	of MOR-1 ar	nd MOR-1C ir	the receptor	binding as	say
		K _i valu	e (nM)			Tukey
Ligand	MOR-1	MOR-1C	MOR-1D	MOR-1E	ANOVA	MOR:P value
Morphine M6G	5.3 ± 2.0 5.2 ± 1.8	2.4 ± 0.6 4.1 ± 1.2	1.5 ± 0.2 4.8 ± 0.8	2.3 ± 0.4 5.6 ± 0.7	N.S.	1vs1D:P < 0.05

TABLE 2-continued

	Selectivit	y of MOR-1 ar	nd MOR-1C in	the receptor	binding ass	ay
		K _i valu	ie (nM)			Tukey
Ligand	MOR-1	MOR-1C	MOR-1D	MOR-1E	ANOVA	MOR:P value
DAMGO DADLE	1.8 ± 0.5 2.1 ± 0.3	0.93 ± 0.19 3.2 ± 1.9	0.71 ± 0.11 1.3 ± 0.4	1.2 ± 0.5 2.5 ± 0.7	N.S. N.S.	

EXAMPLE 5

Functional Significance of the Variants

[0149] Antisense mapping was used to explore the functional significance of these new variants. Pasternak and Standifer (1995); and Standifer et al. (1994). This method has been used extensively to correlate opioid pharmacology with the function of the MOR-1 receptor. Rossi et al. (1994); Rossi et al. (1995); Rossi et al. (1995); and Kolesnikov et al. (1996). Groups of mice ($n \ge 20$) received antisense oligodeoxynucleotides corresponding to specific MOR-1, MOR-1c, MOR-1d, or MOR-1e exons daily for five days. Following administration of the antisense probes, analgesia was assessed by the radiant heat tailflick assay. Rossi et al. (1996); and Rossi et al. (1995). This assay was performed by exposing tails to a light source and determining the baseline latency (typically between 2 and 3 sec). Analgesia was indicated when doubling of the baseline latency occurred. Significance between groups was assessed using the Fisher Exact Test.

[0150] The remaining activity of the variants was measured following administration of the antisense probes in the presence of both morphine and M6G analgesia (**FIG. 11**),

two mu drugs whose actions have been distinguished using antisense approaches. Rossi et al. (1994); Rossi et al. (1995); and Rossi et al., (1995). All four antisense probes significantly lowered morphine analgesia (FIG. 11). A mismatch control probe targeted against exon 7 was inactive, confirming the specificity of the response.

[0151] In contrast to their significant blockade of morphine analgesia, none of the antisense probes significantly lowered M6G analgesia. Thus, these exons are not a component of the postulated M6G receptor. The reduction in morphine analgesia produced by the antisense probes implies that each of the variant mRNAs, and ultimately the receptor(s) which they encode, are involved in mediating morphine analgesia.

[0152] All references cited herein, are hereby incorporated herein. Although the foregoing invention has been described in some detail, by way of illustration and example for the purposes of clarity and understanding, it will be apparent to those skilled in the art that certain changes and modifications can be practiced. Therefore, the description and examples should not be construed as limiting the scope of the invention, which is delineated by the appended claims.

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US 2003/0103972 A1

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124

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 Ile Thr Val Cys Tyr Gly Leu Met Ile Leu Arg Leu Lys Ser Val Arg

 210
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Ser	His	Val 35	Asp	Gly	Asn	Gln	Ser 40	Asp	Pro	Cys	Gly	Pro 45	Asn	Arg	Thr
Gly	Leu 50	Gly	Gly	Ser	His	Ser 55	Leu	Cys	Pro	Gln	Thr 60	Gly	Ser	Pro	Ser
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Val	Gly	Leu	Phe	Gl y 85	Asn	Phe	Leu	Val	Met 90	Tyr	Val	Ile	Val	Arg 95	Tyr
Thr	Lys	Met	L y s 100	Thr	Ala	Thr	Asn	Ile 105	Tyr	Ile	Phe	Asn	Leu 110	Ala	Leu
Ala	Asp	Ala 115	Leu	Ala	Thr	Ser	Thr 120	Leu	Pro	Phe	Gln	Ser 125	Val	Asn	Tyr
Leu	Met 130	Gly	Thr	Trp	Pro	Phe 135	Gly	Asn	Ile	Leu	C y s 140	Lys	Ile	Val	Ile
Ser 145	Ile	Asp	Tyr	Tyr	Asn 150	Met	Phe	Thr	Ser	Ile 155	Phe	Thr	Leu	Сув	Thr 160
Met	Ser	Val	Asp	Arg 165	Tyr	Ile	Ala	Val	C y s 170	His	Pro	Val	Lys	Ala 175	Leu
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Lys	Tyr 210	Arg	Gln	Gly	Ser	Ile 215	Asp	Суз	Thr	Leu	Thr 220	Phe	Ser	His	Pro
Thr 225	Trp	Tyr	Trp	Glu	Asn 230	Leu	Leu	Lys	Ile	C y s 235	Val	Phe	Ile	Phe	Ala 240
Phe	Ile	Met	Pro	Val 245	Leu	Ile	Ile	Thr	Val 250	Cys	Tyr	Gly	Leu	Met 255	Ile
Leu	Arg	Leu	L y s 260	Ser	Val	Arg	Met	Leu 265	Ser	Gly	Ser	Lys	Glu 270	Lys	Asp
Arg	Asn	Leu 275	Arg	Arg	Ile	Thr	A rg 280	Met	Val	Leu	Val	Val 285	Val	Ala	Val
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Суз	Ile	Ala	Leu	Gly 325	Tyr	Thr	Asn	Ser	C y s 330	Leu	Asn	Pro	Val	Leu 335	Tyr

Ala Phe Leu Asp Glu Asn Phe Lys Arg Cys Phe Arg Glu Phe Cys Ile 340 345 350 Pro Thr Ser Ser Thr Ile Glu Gln Gln Asn Ser Ala Arg Ile Arg Gln 355 360 365 Asn Thr Arg Glu His Pro Ser Thr Ala Asn Thr Val Asp Arg Thr Asn 370 375 380 His Gln Arg Asn Glu Glu Pro Ser 385 390 <210> SEQ ID NO 20 <211> LENGTH: 401 <212> TYPE: PRT <213> ORGANISM: Mus musculus <400> SEOUENCE: 20 Met Asp Ser Ser Ala Gly Pro Gly Asn Ile Ser Asp Cys Ser Asp Pro151015 Leu Ala Pro Ala Ser Cys Ser Pro Ala Pro Gly Ser Trp Leu Asn Leu 20 25 30 Ser His Val Asp Gly Asn Gln Ser Asp Pro Cys Gly Pro Asn Arg Thr 35 40 Gly Leu Gly Gly Ser His Ser Leu Cys Pro Gln Thr Gly Ser Pro Ser 55 50 60 Met Val Thr Ala Ile Thr Ile Met Ala Leu Tyr Ser Ile Val Cys Val65707580 Val Gly Leu Phe Gly Asn Phe Leu Val Met Tyr Val Ile Val Arg Tyr 85 90 Thr Lys Met Lys Thr Ala Thr Asn Ile Tyr Ile Phe Asn Leu Ala Leu 105 Ala Asp Ala Leu Ala Thr Ser Thr Leu Pro Phe Gln Ser Val Asn Tyr 120 115 125 Leu Met Gly Thr Trp Pro Phe Gly Asn Ile Leu Cys Lys Ile Val Ile 130 135 140 Ser Ile Asp Tyr Tyr Asn Met Phe Thr Ser Ile Phe Thr Leu Cys Thr 145 150 155 160 Met Ser Val Asp Arg Tyr Ile Ala Val Cys His Pro Val Lys Ala Leu 165 170 175 Asp Phe Arg Thr Pro Arg Asn Ala Lys Ile Val Asn Val Cys Asn Trp 180 185 190 Ile Leu Ser Ser Ala Ile Gly Leu Pro Val Met Phe Met Ala Thr Thr 200 205 195 Lys Tyr Arg Gln Gly Ser Ile Asp Cys Thr Leu Thr Phe Ser His $\ensuremath{\mathsf{Pro}}$ 215 210 220 Thr Trp Tyr Trp Glu Asn Leu Leu Lys Ile Cys Val Phe Ile Phe Ala 225 230 235 240
 Phe Ile Met Pro Val Leu Ile Ile Thr Val Cys Tyr Gly Leu Met Ile

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 Leu Arg Leu Lys Ser Val Arg Met Leu Ser Gly Ser Lys Glu Lys Asp 265 Arg Asn Leu Arg Arg Ile Thr Arg Met Val Leu Val Val Val Ala Val 285 275 280
 Phe Ile Val Cys Trp Thr Pro Ile His Ile Tyr Val Ile Ile Lys Ala

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Leu Ile Thr Ile Pro Glu Thr Thr Phe Gln Thr Val Ser Trp His Phe 305 310 315 320 Cys Ile Ala Leu Gly Tyr Thr Asn Ser Cys Leu Asn Pro Val Leu Tyr 325 330 335 Ala Phe Leu Asp Glu Asn Phe Lys Arg Cys Phe Arg Glu Phe Cys Ile 340 345 350 Pro Thr Ser Ser Thr Ile Glu Gln Gln Asn Ser Ala Arg Ile Arg Gln 360 Asn Thr Arg Glu His Pro Ser Thr Ala Asn Thr Val Asp Arg Thr Asn 375 380 370 His Gln Lys Lys Leu Asp Ser Gln Arg Gly Cys Val Gln His Pro 385 390 395 400 Val <210> SEQ ID NO 21 <211> LENGTH: 399 <212> TYPE: PRT <213> ORGANISM: Mus musculus <400> SEQUENCE: 21 Met Glu Arg Ile Ser Lys Ala Gly Ser Pro Pro Gly Ser Phe Gln Arg151015 Glu Tyr Ser Thr Glu Leu Glu Gly Ala Gly Ser Gly Pro Met Met Glu 20 25 30 Ala Phe Ser Lys Ser Ala Phe Gln Lys Leu Arg Gln Arg Asp Gly Asn 35 40 45 Gln Glu Gly Lys Ser Tyr Leu Arg Tyr Thr Lys Met Lys Thr Ala Thr 50 55 60 Asn Ile Tyr Ile Phe Asn Leu Ala Leu Ala Asp Ala Leu Ala Thr Ser 70 65 75 80
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Ile His Ile Tyr Val Ile Ile Lys Ala Leu Ile Thr Ile Pro Glu Thr 260 265 270 Thr Phe Gln Thr Val Ser Trp His Phe Cys Ile Ala Leu Gly Tyr Thr 275 280 285 Asn Ser Cys Leu Asn Pro Val Leu Tyr Ala Phe Leu Asp Glu Asn Phe 290 295 300 Lys Arg Cys Phe Arg Glu Phe Cys Ile Pro Thr Ser Ser Thr Ile Glu 305 310 315 320 Gln Gln Asn Ser Ala Arg Ile Arg Gln Asn Thr Arg Glu His Pro Ser 325 330 335 Thr Ala Asn Thr Val Asp Arg Thr Asn His Gln Pro Thr Leu Ala Val 340 345 350 Ser Val Ala Gln Ile Phe Thr Gly Tyr Pro Ser Pro Thr His Val Glu 355 360 365 Lys Pro Cys Lys Ser Cys Met Asp Arg Gly Met Arg Asn Leu Leu Pro 370 375 380 Asp Asp Gly Pro Arg Gln Glu Ser Gly Glu Gly Gln Leu Gly Arg 390 395 385 <210> SEQ ID NO 22 <211> LENGTH: 17 <212> TYPE: PRT <213> ORGANISM: Mus musculus <400> SEQUENCE: 22 Glu His Pro Ser Thr Ala Asn Thr Val Asp Arg Thr Asn His Gln Cys 5 10 Val <210> SEQ ID NO 23 <211> LENGTH: 22 <212> TYPE: PRT <213> ORGANISM: Mus musculus <400> SEQUENCE: 23 Glu His Pro Ser Thr Ala Asn Thr Val Asp Arg Thr Asn His Gln Glu 10 5 15 1 Pro Gln Ser Ala Glu Thr 20 <210> SEQ ID NO 24 <211> LENGTH: 41 <212> TYPE: PRT <213> ORGANISM: Homo sapiens <400> SEQUENCE: 24 Asp Arg Thr Asn His Gln Cys Leu Pro Ile Pro Ser Leu Ser Cys Trp 10 5 1 Ala Leu Glu His Gly Arg Leu Val Val Tyr Pro Gly Pro Leu Gln Gly 25 30 20 Pro Leu Val Arg Tyr Asp Leu Pro Ala 35 40 <210> SEQ ID NO 25 <211> LENGTH: 390 <212> TYPE: PRT

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Val	Gly	Leu	Phe	Gly 85	Asn	Phe	Leu	Val	Met 90	Tyr	Val	Ile	Val	Arg 95	Tyr
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Leu	Met 130	Gly	Thr	Trp	Pro	Phe 135	Gly	Asn	Ile	Leu	C y s 140	Lys	Ile	Val	Ile
Ser 145	Ile	Asp	Tyr	Tyr	Asn 150	Met	Phe	Thr	Ser	Ile 155	Phe	Thr	Leu	Сув	Thr 160
Met	Ser	Val	Asp	Arg 165	Tyr	Ile	Ala	Val	С у в 170	His	Pro	Val	Lys	Ala 175	Leu
Asp	Phe	Arg	Thr 180	Pro	Arg	Asn	Ala	L y s 185	Ile	Val	Asn	Val	С у в 190	Asn	Trp
Ile	Leu	Ser 195	Ser	Ala	Ile	Gly	Leu 200	Pro	Val	Met	Phe	Met 205	Ala	Thr	Thr
Lys	Tyr 210	Arg	Gln	Gly	Ser	Ile 215	Asp	Cys	Thr	Leu	Thr 220	Phe	Ser	His	Pro
Thr 225	Trp	Tyr	Trp	Glu	Asn 230	Leu	Leu	Lys	Ile	С у в 235	Val	Phe	Ile	Phe	Ala 240
Phe	Ile	Met	Pro	Val 245	Leu	Ile	Ile	Thr	Val 250	Сув	Tyr	Gly	Leu	Met 255	Ile
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Arg	Asn	Leu 275	Arg	Arg	Ile	Thr	A rg 280	Met	Val	Leu	Val	Val 285	Val	Ala	Val
Phe	Ile 290	Val	Cys	Trp	Thr	Pro 295	Ile	His	Ile	Tyr	Val 300	Ile	Ile	Lys	Ala
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Сув	Ile	Ala	Leu	Gly 325	Tyr	Thr	Asn	Ser	Cys 330	Leu	Asn	Pro	Val	Leu 335	Tyr
Ala	Phe	Leu	Asp 340	Glu	Asn	Phe	Lys	Arg 345	Сув	Phe	Arg	Glu	Phe 350	Сув	Ile
Pro	Thr	Ser 355	Ser	Thr	Ile	Glu	Gln 360	Gln	Asn	Ser	Ala	Arg 365	Ile	Arg	Gln
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31

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Pro Thr	Ser 355	Ser	Thr	Ile	Glu	Gln 360	Gln	Asn	Ser	Ala	Arg 365	Ile	Arg	Gln
Asn Thr 370	Arg	Glu	His	Pro	Ser 375	Thr	Ala	Asn	Thr	Val 380	Asp	Arg	Thr	Asn
His Gln 385	Lys	Leu	Leu	Met 390	Trp	Arg	Ala	Met	Pro 395		Phe	Lys	Arg	His 400
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Ser His	Val 35	Asp	Gly	Asn	Gln	Ser 40	Asp	Pro	Суз	Gly	Pro 45	Asn	Arg	Thr
Gly Leu 50	Gly	Gly	Ser	His	Ser 55	Leu	Cys	Pro	Gln	Thr 60	Gly	Ser	Pro	Ser
Met Val 65	Thr	Ala	Ile	Thr 70	Ile	Met	Ala	Leu	Ty r 75	Ser	Ile	Val	Cys	Val 80
Val Gly	Leu	Phe	Gly 85	Asn	Phe	Leu	Val	Met 90	Tyr	Val	Ile	Val	Arg 95	Tyr
Thr Lys	Met	L y s 100		Ala	Thr	Asn	Ile 105		Ile	Phe	Asn	Leu 110		Leu
Ala Asp	Ala 115		Ala	Thr	Ser	Thr 120		Pro	Phe	Gln	Ser 125		Asn	Tyr
Leu Met		Thr	Trp	Pro			Asn	Ile	Leu			Ile	Val	Ile
130 Ser Ile	Asp	Tyr	Tyr		135 Met	Phe	Thr	Ser			Thr	Leu	Cys	
145 Met Ser	Val	Asp	Arσ	150 Tyr	Ile	Ala	Val	Cvs	155 His		Val	Lvs	Ala	160 Leu
		-	165	-				170				-	175	
Asp Phe		180					185					190		
Ile Leu	Ser 195	Ser	Ala	Ile	Gly	Leu 200	Pro	Val	Met	Phe	Met 205	Ala	Thr	Thr
Lys Tyr 210	Arg	Gln	Gly	Ser	Ile 215		Cys	Thr	Leu	Thr 220	Phe	Ser	His	Pro
Thr Trp 225	Tyr	Trp	Glu	Asn 230	Leu	Leu	Lys	Ile	C y s 235		Phe	Ile	Phe	Ala 240
Phe Ile	Met	Pro	Val 245	Leu	Ile	Ile	Thr	Val 250	Cys	Tyr	Gly	Leu	Met 255	Ile

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Asp Phe Arg Thr Pro Arg Asn Ala Lys Ile Val Asn Val Cys Asn Trp 180 185 190 Ile Leu Ser Ser Ala Ile Gly Leu Pro Val Met Phe Met Ala Thr Thr 195 200 205 Lys Tyr Arg Gln Gly Ser Ile Asp Cys Thr Leu Thr Phe Ser His Pro 215 210 220 Thr Trp Tyr Trp Glu Asn Leu Leu Lys Ile Cys Val Phe Ile Phe Ala 235 225 230 240 Phe Ile Met Pro Val Leu Ile Ile Thr Val Cys Tyr Gly Leu Met Ile 245 250 255 Leu Arg Leu Lys Ser Val Arg Met Leu Ser Gly Ser Lys Glu Lys Asp 260 265 270 Arg Asn Leu Arg Arg Ile Thr Arg Met Val Leu Val Val Val Ala Val 280 275 285 Phe Ile Val Cys Trp Thr Pro Ile His Ile Tyr Val Ile Ile Lys Ala 295 300 Leu Ile Thr Ile Pro Glu Thr Thr Phe Gln Thr Val Ser Trp His Phe 310 315 320 305 Cys Ile Ala Leu Gly Tyr Thr Asn Ser Cys Leu Asn Pro Val Leu Tyr 325 330 335 Ala Phe Leu Asp Glu Asn Phe Lys Arg Cys Phe Arg Glu Phe Cys Ile 340 345 350 Pro Thr Ser Ser Thr Ile Glu Gln Gln Asn Ser Ala Arg Ile Arg Gln 355 360 365 Asn Thr Arg Glu His Pro Ser Thr Ala Asn Thr Val Asp Arg Thr Asn 370 375 380 His Gln Leu Glu Asn Leu Glu Ala Glu Thr Ala Pro Leu Pro 385 390 395 <210> SEQ ID NO 30 <211> LENGTH: 13 <212> TYPE: PRT <213> ORGANISM: Mus musculus <400> SEQUENCE: 30 Leu Glu Asn Leu Glu Ala Glu Thr Thr Ala Pro Leu Pro 1 5 10 <210> SEQ ID NO 31 <211> LENGTH: 4 <212> TYPE: PRT <213> ORGANISM: Mus musculus <400> SEQUENCE: 31 Val Arg Ser Leu <210> SEQ ID NO 32 <211> LENGTH: 5 <212> TYPE: PRT <213> ORGANISM: Mus musculus <400> SEQUENCE: 32 Lys Ile Asp Leu Phe

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1. An isolated MOR-1C splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

2. An isolated MOR-1D splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

3. An isolated MOR-1E splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

4. An isolated MOR-1F splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

5. An isolated MOR-1G splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

6. An isolated MOR-1A splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

7. An isolated MOR-1H splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

8. An isolated MOR-1B II splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

9. An isolated MOR-1B I splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

10. An isolated MOR-1I splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

11. An isolated MOR-1J splice variant polypeptide that consists essentially of the amino acid residues depicted in **FIG. 3**.

12. An isolated human MOR-1 splice variant polypeptide that consists essentially of the amino acid residues depicted in FIG. 3.

13. The polypeptide as in one of the preceding claims in which the polypeptide comprises a heterodimeric or homodimeric composition.

14. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1c as depicted in **FIG. 2**.

15. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1d as depicted in **FIG. 2**.

16. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1e as depicted in **FIG. 2**.

17. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1f as depicted in **FIG. 2**.

18. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1b II as depicted in FIG. 2.

19. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1a as depicted in **FIG. 2**.

20. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1b I as depicted in **FIG. 2**.

21. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1i as depicted in **FIG. 2**.

22. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions,

wherein the nucleotide fragment consists essentially of MOR-1j as depicted in **FIG. 2**.

23. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1g as depicted in FIG. 2.

24. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of MOR-1h as depicted in FIG. 2.

25. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of human MOR-1 as depicted in FIG. 2.

26. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of 2730510 as depicted in **FIG. 2**.

27. An isolated polynucleotide, or a complementary strand thereto that hybridizes under stringent conditions, wherein the nucleotide fragment consists essentially of 161416 as depicted in **FIG. 2**.

28. A method of screening compositions for opioid activity comprising the steps of a) obtaining a control cell that does not express an MOR-1 splice variant polypeptide; b) obtaining a test cell that is the same as the control cell except that it expresses an MOR-1 splice variant polypeptide as in any one of claims **122-133**; c)contacting the control cell and test cell with an amount of an opioid sufficient to exert a physiologic effect; d) separately measuring the physiologic effect of the composition on the control cell and test cell; and e) comparing the physiologic effect of the composition to the physiologic effect of the opioid, where determination of a physiologic effect of the composition is expressed relative to that of the opioid.

29. The method according to claim **149**, where the composition is selected from the group consisting of synthetic combinatorial libraries of small molecule ligands, eukaryotic whole cell lysates or extracts, or media conditioned by cultured eukaryotic cells.

30. The method according to claim **149**, where the opioid is selected from the group consisting of morphine, methadone, etorphine, levorphanol, fentanyl, sufentanil, [D-Ala2, MePhe4,Gly(ol)5]enkephalin (DAMGO), pentazocine, eth-ylketocyclazocine, bremazocine, spiradoline, [D-Ser2,Leu5]

enkephalin-Thr6 (DSLET), Met-enkephalin, Leuenkephalin, (3-endorphin, dynorphin A, dynorphin B, or a-neoendorphin.

31. The method according to claim **149**, where the physiological effect is measured by changes in the levels of neuroendocrine hormones.

32. The method according to claim **152**, where the hormone is selected from the group consisting of prolactin, growth hormone, gonadotropin-releasing hormone, adreno-corticotropin, corticotropin-releasing factor, luteinizing hormone, follicle stimulating hormone, testosterone or cortisol.

33. A method of screening compositions for opioid binding activity comprising the steps of a) obtaining a control polypeptide that is not an MOR-1 splice variant polypeptide; b) obtaining a test polypeptide that is an MOR-1 splice variant polypeptide as in any one of claims **122-133**; c) contacting a composition with the control polypeptide and the test polypeptide; d) contacting the test polypeptide with an amount of an opioid sufficient to measurably bind the test polypeptide; e) measuring the binding of the composition and the opioid; and f) comparing test polypeptide binding of the composition to that of the opioid, where determination of binding of the composition is expressed relative to that of the opioid.

34. The method according to claim **154** where the composition is selected from the group consisting of synthetic combinatorial libraries of small molecule ligands, eukaryotic whole cell lysates or extracts, or media conditioned by cultured eukaryotic cells.

35. A method for regulating morphine analgesia in a subject comprising altering the amount of MOR-1 polypeptide fragment activity by a) administering antigen binding fragments to a subject in an amount and a duration sufficient to regulate morphine analgesia; or b) administering agonists to a subject in an amount and a duration sufficient to regulate morphine analgesia; or c) administering antagonists to a subject in an amount and a duration sufficient to regulate morphine analgesia; or d) administering small molecule ligands to a subject in an amount and a duration sufficient to regulate morphine analgesia; or d) administering small molecule ligands to a subject in an amount and a duration sufficient to regulate morphine analgesia; and wherein the antigen binding fragment, agonist, antagonist or small molecule ligand is directed to an MOR-1 splice variant as in any one of claims **122-133**.

* * * * *