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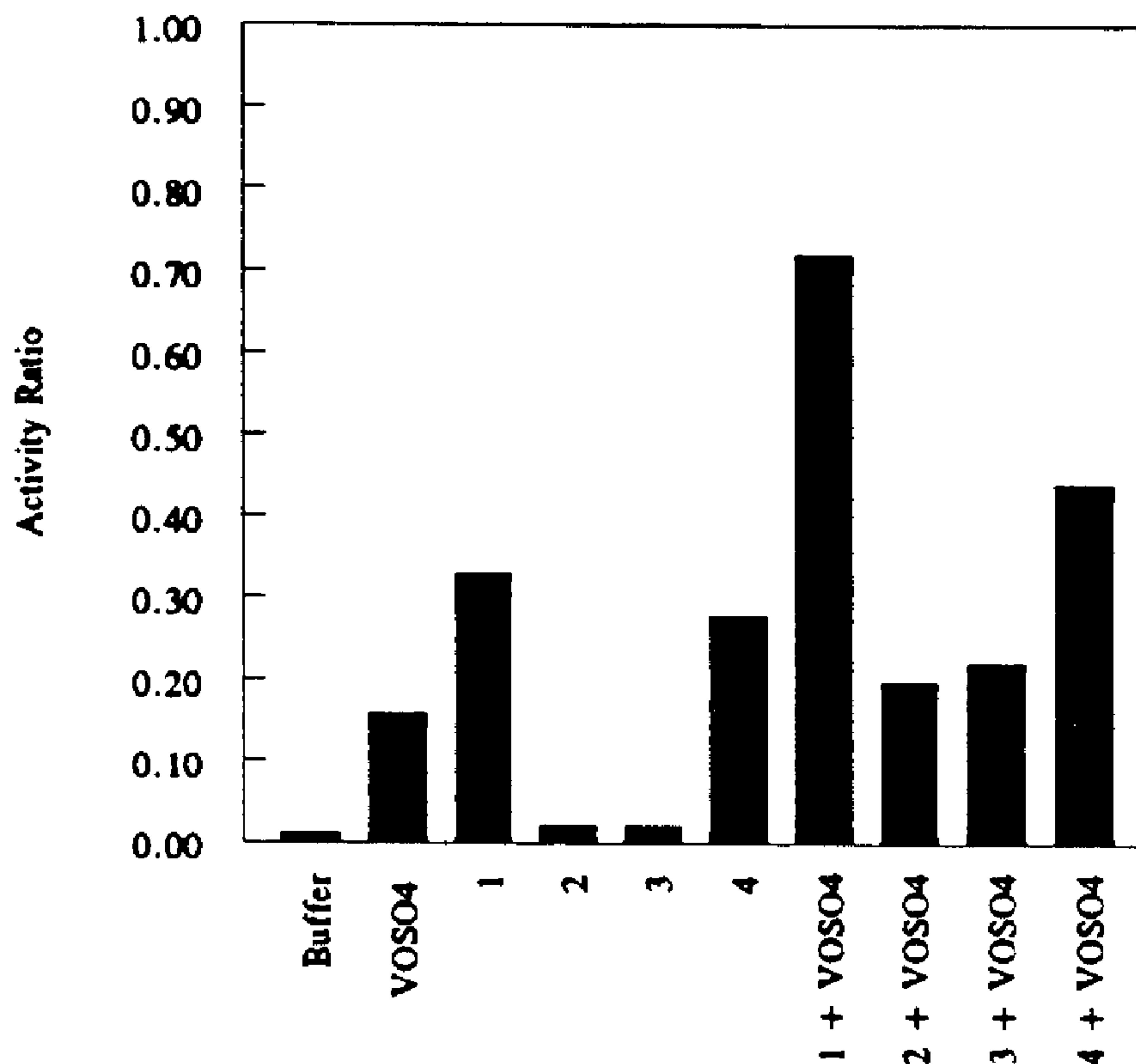
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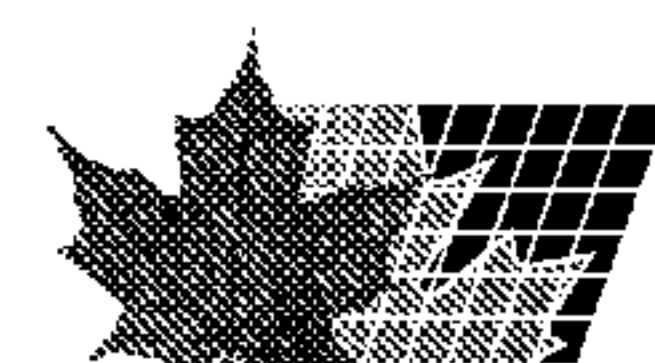
(54) Titre : REDUCTION DE LA DISTORSION DE LA PREMIERE LAME ET AMELIORATION DE LA STABILITE DE L'ENZYME PAR INCORPORATION DE TELLURURES DE DIARYLE DANS LA COUCHE D'IMPRESSON D'ELEMENTS POUR DOSAGE IMMUNOLOGIQUE SUR FILM SEC

(54) Title: REDUCTION IN FIRST SLIDE BIAS AND IMPROVED ENZYME STABILITY BY INCORPORATION OF DIARYL TELLURIDES IN THE GRAVURE LAYER OF DRY-FILM, IMMUNOASSAY ELEMENTS



(57) Abrégé/Abstract:

The present invention relates to a dry immunoassay analytical element for assaying a ligand, comprising a support bearing: (a) a label layer comprising an enzyme labeled ligand or an enzyme labeled receptor; (b) a spreading layer ; (c) a receptor layer



(57) **Abrégé(suite)/Abstract(continued):**

comprising a fixed concentration of an immobilized receptor for the ligand; and (d) a gravure layer comprising a diaryl telluride. A preferred embodiment of the present invention further comprises a vanadyl salt. The present invention further relates to a method for performing an assay using an element as described above.

Abstract of the Disclosure

The present invention relates to a dry immunoassay analytical element for assaying a ligand, comprising a support bearing: (a) a label layer comprising an enzyme labeled ligand or an enzyme labeled receptor; (b) a spreading layer ; (c) a receptor layer comprising a fixed concentration of an immobilized receptor for the ligand; and (d) a gravure layer comprising a diaryl telluride. A preferred embodiment of the present invention further comprises a vanadyl salt. The present invention further relates to a method for performing an assay using an element as described above.

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Reduction in First Slide Bias and Improved Enzyme Stability by Incorporation of Diaryl Tellurides in the Gravure Layer of Dry-film, Immunoassay Elements

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Field of the Invention

This invention relates to dry-film elements and use thereof in an immunoassay. In particular, it relates to improved stability of labeled immunoreactants in dry-film elements.

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Background of the Invention

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Immunoassays, which take advantage of natural immunological reactions, have found widespread use as analytical techniques in clinical chemistry. Because of the specificity of the reactions, they are particularly advantageous in quantifying biological analytes that are present in very low concentration in biological fluids. Such analytes include, for example, antigens, antibodies, therapeutic drugs, narcotics, enzymes, hormones, proteins, etc.

The analyte, which is the target of the assay, is referred to herein as the ligand. Compounds which specifically recognize the ligand and react to form complexes with the ligand are referred to herein as receptors. The receptor and the ligand form a conjugate pair. Any member of the pair can function as a receptor or a ligand.

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In the case of a competitive assay, a labeled analyte, including labeled immuno-competent derivatives and analogs of such analyte, are typical components of the assay; whereas, in the case of a sandwich assay, a labeled receptor for the analyte is typically employed. These are referred to herein as the labeled ligand and labeled receptor, respectively.

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In competitive binding immunoassays, a labeled ligand is placed in competition with unlabeled ligand for reaction with a fixed amount of the appropriate receptor. Unknown concentrations of the ligand can be determined from the measured signal of either the bound or unbound (i.e. free) labeled ligand. The reaction proceeds as follows:

ligand + labeled ligand + receptor - substrate $\rightleftharpoons$

ligand-receptor-substrate + labeled ligand-receptor - substrate.

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In an alternative immunoassay format known as a sandwich immunoassay or immunometric assay, the ligand is contacted with two or more receptor molecules which bind to the ligand at different epitopic sites. One receptor is typically appropriately labeled and the other is either immobilized on a solid substrate or is capable of being immobilized thereon. The amount of ligand is directly proportional to the amount of bound complex among the ligand and the two receptors. This is illustrated as follows:

substrate - receptor₁ + ligand + receptor₂ - label $\rightleftharpoons$

substrate - receptor₁ - ligand - receptor₂ - label

Conventional labels include radioactive tags, enzymes, chromophores, fluorophores, stable free radicals and enzyme cofactors, inhibitors and allosteric effectors.

Immunoassay analytical elements are known from U.S. Patents 4,517,288 and 4,258,001. In general, such elements comprise receptors, such as antibodies for a ligand, immobilized in a particulate layer. In addition the element usually contains a reagent system that, through interaction with a bound or unbound species, results in a signal that can be correlated to the concentration of ligand in a sample. In use, the sample may be combined with an enzyme labeled ligand and applied to the element. After a time, a solution containing a substrate for the labeled ligand is applied to the particulate layer. The reaction with the substrate is catalyzed by the enzyme label to form a reaction product that ultimately causes a signal, for example, color to develop. The reflection density of the color can be correlated to the concentration of the ligand in the sample. Similar signal development systems are known for other known conventional labels such as radioactive tags, chromophores, fluorophores, stable free radicals, and enzyme cofactors, inhibitors and allosteric effectors.

Multilayer immunoassay elements are dry-film elements which use the above described immunoassay principles to measure analytes in fluid samples. In competitive assay elements, the rate of color formation (or other signal) is inversely correlated to the amount of analyte present and in sandwich assay elements, the rate of color formation (or other signal) is directly correlated to the amount of analyte present. Also, the rate of color formation (or other signal) is directly proportional to the activity of the enzyme labeled analyte, e.g. drug, or enzyme labeled receptor bound to the immobilized receptor. For the immunoassays to maintain a stable calibration, none of the enzyme activity (measured rate) can be lost in any of the slides during the specified calibration period.

Frequently, immunoassay elements are supplied to customers in plastic "cartridges" containing 50 separate elements from which one element may be removed at a time as needed. The elements are stacked one on top of another so that the lower 49 elements in the cartridge all have their top surfaces covered by the element above. However, the top element in the stack has no such covering and therefore the surface of that element is exposed to environmental factors to which the

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other 49 elements are not. For example, the top (or first) element is more exposed to air flow and light than the remainder of the elements when the cartridges are being handled during manufacturing or when the cartridges are in the element supplies of the clinical analyzers.

During storage, prior to use, the cartridges themselves are stored in sealed, foil-lined bags. However, the top element is still more exposed to the residual air and humidity inside the sealed bags than the other 49 elements.

It has been found that, when a common test fluid was reacted with the elements in a cartridge, the rate of color formation observed in the top (or first) element was always lower than the rate of color formation observed when the same test fluid was applied to elements below the top element in the same cartridge. This is referred to as first slide bias.

Summary of the Invention

One method for dealing with lower rate of color formation of the first slide involves the use of vanadium ions as described in U.S. Patent 5,516,645 by Daniel et al.

Applicants have discovered the use of diaryl telluride (DAT) or in the plural (DATs) for reducing "first slide bias." The use of diaryl tellurides to prevent first slide bias can be employed in both sandwich and competitive immunorate assays.

According to one aspect of the present invention, there is a dry immunoassay element for assaying a ligand, comprising a support bearing:

A dry immunoassay analytical element for assaying a ligand, comprising a support bearing:

- (a) a label layer comprising an enzyme labeled ligand or an enzyme labeled receptor;
- (b) a spreading layer ;
- (c) a receptor layer comprising a fixed concentration of an immobilized receptor for the ligand; and
- (d) a gravure layer comprising a diaryl telluride.

In one embodiment , the receptor is covalently bonded to polymeric beads having a diameter in the range of 0.1 to 5 μm . In a preferred embodiment enzyme labeled ligand and diaryl telluride are in the same gravure layer or zone, wherein the gravure layer or zone is adjacent to or contiguous with the surface of the spreading layer that provides first or initial contact with a liquid sample.

The element defined above substantially reduces first slide bias, i.e. the altered rate of color development of the top element in a cartridge compared to the other elements in the same cartridge.

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Moreover, all elements in the cartridge exhibit greater long term stability and storage. The examples establish that diaryl telluride compounds provide these benefits.

In addition, the DAT compounds are useful in stabilizing enzyme compositions generally, particularly peroxidases and especially horseradish peroxidase (HRP) and conjugates of enzymes, preferably HRP, with small molecules such as therapeutic drugs, drugs of abuse, steroids, and the like. Another embodiment is conjugates of enzymes, preferably HRP, with large molecules such as proteins, carbohydrates, and the like, in which such enzymes are used as labels. For example, the compounds can be used in solution assays as well as in the dry elements of this invention.

Another embodiment of the present invention provides a method for assaying an immunologically reactive ligand in an aqueous sample, comprising the steps of:

- (1) providing a dry immunoassay analytical element comprising a support bearing:
 - (a) a label layer comprising an enzyme labeled ligand or an enzyme labeled receptor;
 - (b) a spreading layer;
 - (c) a receptor layer comprising a fixed concentration of an immobilized receptor for the ligand;
 - (d) a gravure layer comprising a diaryl telluride;
- (2) contacting a finite area of the spreading layer of the element with the sample thereby forming (i) an immobilized ligand-receptor complex, (ii) an immobilized enzyme-labeled ligand-receptor complex, or (iii) a mixture of (i) and (ii), or an immobilized receptor-ligand-labeled receptor complex;
- (3) contacting the finite area with a substrate solution for the enzyme label thereby catalyzing the development of a signal; and
- (4) determining the concentration of the ligand by detecting the signal or measuring the intensity or amount of said signal.

In the above method, the enzyme labeled receptor or enzyme labeled ligand is separated from immobilized enzyme labeled ligand or immobilized enzyme labeled receptor. Such separation may be effected by any means known in the art, for example by the addition of a substrate solution, e.g. a hydrogen peroxide solution.

One or more layers of an element may each comprise one or more zones. Contiguous layers of an element may have sharp boundaries between them wherein, by optical and/or electron microscopy, they may appear as distinct and separate layers of the element; or they may have a zone or region of contact wherein mixing of the layers occurs and boundaries between layers are blurred or indistinct. In such cases, separate and distinct layers may or may not be visible by optical and/or

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electron microscopy. The formation of zones may be partial or substantially complete (or complete). In the case of partial zone formation, only a portion of a deposited layer penetrates and mixes with a contiguous separately deposited layer (both layers may mutually penetrate each other, but in any event, a zone comprising a portion of both layers forms - and individual and mutual layer penetration are both contemplated, as both result in zone formation as described herein) so as to include a zone (or zones) within the separately deposited contiguous layer (or layers); the layers remaining separate and distinct but having blurred or indistinct boundaries.

On the other hand, most or all of a separately deposited layer may penetrate and mix with a contiguous layer (or layers) to form a zone (or zones) within the contiguous layer (or layers) in such a way as to be contained substantially or entirely within the contiguous layer (or layers), such that it may not appear as a distinct or separate layer apart from the contiguous layer (or layers) of the element by optical and/or electron microscopy. Partial or complete zones may be formed intentionally or may form unavoidably. If formed intentionally, zones within layers may be provided by any process that enables the practitioner to incorporate a specific component or components, to comprise or be included within the zone, within one or more layers. For example a specific component or specific components may be encapsulated within a polymer or other matrix, which matrix may then be incorporated within a layer or layers.

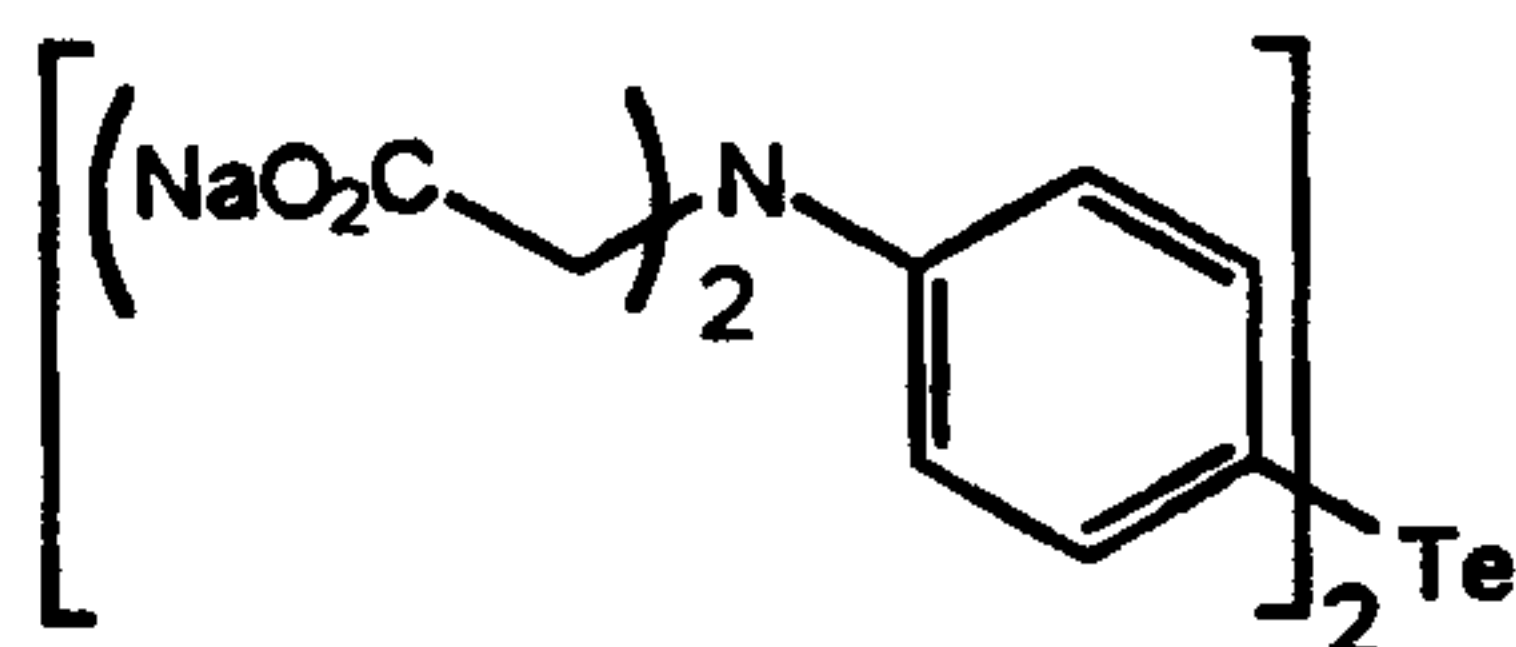
Alternatively, another embodiment of the present invention involves a layer comprising a specific component A (or components) which is contiguous with two adjacent layers may be prepared, such that it is soluble in one of the contiguous layers. During initial contact or subsequently thereafter, a complete or partial zone is formed within the contiguous layer in which it is soluble. The zone in the contiguous layer comprises, therefore, specific component A. The formation of the zone may be partial or complete as defined above. The degree of zone formation can be controlled by adjustment of conditions, such as, solvents which are used to provide the layers, contact time of layers before drying, temperature, etc. as would be known to those of skill in the coating arts. It is to be understood that dry immunoassay elements defined using the terms label layer, receptor layer, gravure layer, and/or other layer, include all embodiments wherein a label, receptor, gravure, and/or other layer of the element may form a separate and distinct layer, or a partial or complete zone (or zones) with a contiguous layer (or layers) during or after deposition of the noted layers.

A gravure layer is a thin layer comprising one or more components deposited upon the surface of another layer, for example, a spreading layer. The term "gravure layer" is used shall mean a thin layer wherein "thin" is a relative term used to denote that the layer is smaller in thickness relative to a contiguous layer (or layers) with which it is in contact. It is a term of convenience, as the gravure coating method represents a preferred means of coating a thin layer comprising diaryl

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telluride (and/or other components). Use of the term is not intended to be restrictive in any manner. Any means that allows coating, deposition or otherwise laying down of a thin layer, as defined hereinabove, is considered as being within the scope of the term "gravure layer". The gravure layer coated directly upon a spreading layer of an element of the present invention may not appear as a distinct or separate layer on the spreading layer; it is anticipated that the gravure layer forms a thin contiguous zone on, on and within, or just below the surface of the spreading layer (or other layer) with which it has initial contact. In any case, liquid sample when contacted with spreading layers comprising a gravure layer and/or zone will directly contact the spreading layer components, beads, etc., because of the porosity of the spreading layer itself, allowing spreading and metering of the sample, as these terms are known in the art. Thus, contacting an element having a gravure layer and/or zone with a liquid sample can be described simply in terms of contacting the spreading layer with the liquid sample, as spreading, metering and all other functions of the spreading layer are unimpeded.

Te5, identified below, represents one of a number of species having the generic formula $Ar^1-Te-Ar^2$ which species are taught in the parent case. Furthermore, we have found that diaryl telluride, especially Te5 of formula



results in significant reduction in first slide bias when incorporated into a gravure layer of the immunoassay element of the carbamazepine (CRBM) competitive immunorate assay at a final rewet level of 1 mM. Applicants have found a significant reduction of first slide bias over a one-year time period. In fact, first slides are indistinguishable from non-first slide controls. When the same level of Te5 is coated directly in the bead spreading layer, rather than in a gravure layer coated over the bead spreading layer, it is insufficient to prevent first slide bias from occurring. It has also been found that the incorporation of 0.25 to 0.50 mM (upon rewet of the element) of Te5 in gravure layers of the dry immunoassay elements designed for determining carbamazepine (CRBM), phenobarbital (PHBR), phenytoin (PHYT), and digoxin (DGXN) significantly reduces first slide bias after six and nine months of freezer storage.

This invention, in general, is drawn to improved dry immunoassay elements comprising diaryl telluride compounds coated in a gravure layer disposed over a spreading layer. However, the gravure layer may be disposed over any layer.

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In another aspect it relates to methods of performing immunoassays using said improved dry-film elements.

Brief Description of the Drawings

5 Figure 1. is a bar-graph showing HRP stability in a model dry-down format when spotting solution variations included: buffer only, 1 mM VOSO₄ and 1 mM each diaryl telluride alone or 1 mM of each with 1 mM VOSO₄ as described in Example 3.

Figure 2. is a bar-graph showing HRP stability over a 24 hour period in a model dry-down format with various concentrations of diaryl tellurides 1 and 4 as described in Example 3.

10 Figure 3. is a bar-graph showing the stability of an anti-CRP-HRP conjugate in a model dry-down format with different concentrations of diaryl tellurides 4 and 5 incorporated in spotting solutions as described in Example 5.

Detailed Description of the Invention

15 The elements of this invention comprise labeled ligand or labeled receptor, spreading layers (or zones), receptor layers (or zones) and gravure layers (or zone). Zones and layers are as earlier defined. The various zones can be in one coated layer or in separate coated layers. For example, the spreading zone and the receptor zone can be in a single layer or they can be in separate layers. The separate layers can be arranged in any order on the support. Or the separate layers can be arranged
20 such that the receptor layer is directly on the support, the spreading layer directly above the receptor layer and the labeled ligand or labeled receptor zone over the spreading layer. When the receptor zone forms an entirely separate layer, the layer will also include a binder of the type described hereinafter. The element can include additional layers such as a gravure layer. All such layers can be coated using coating techniques known in this art.

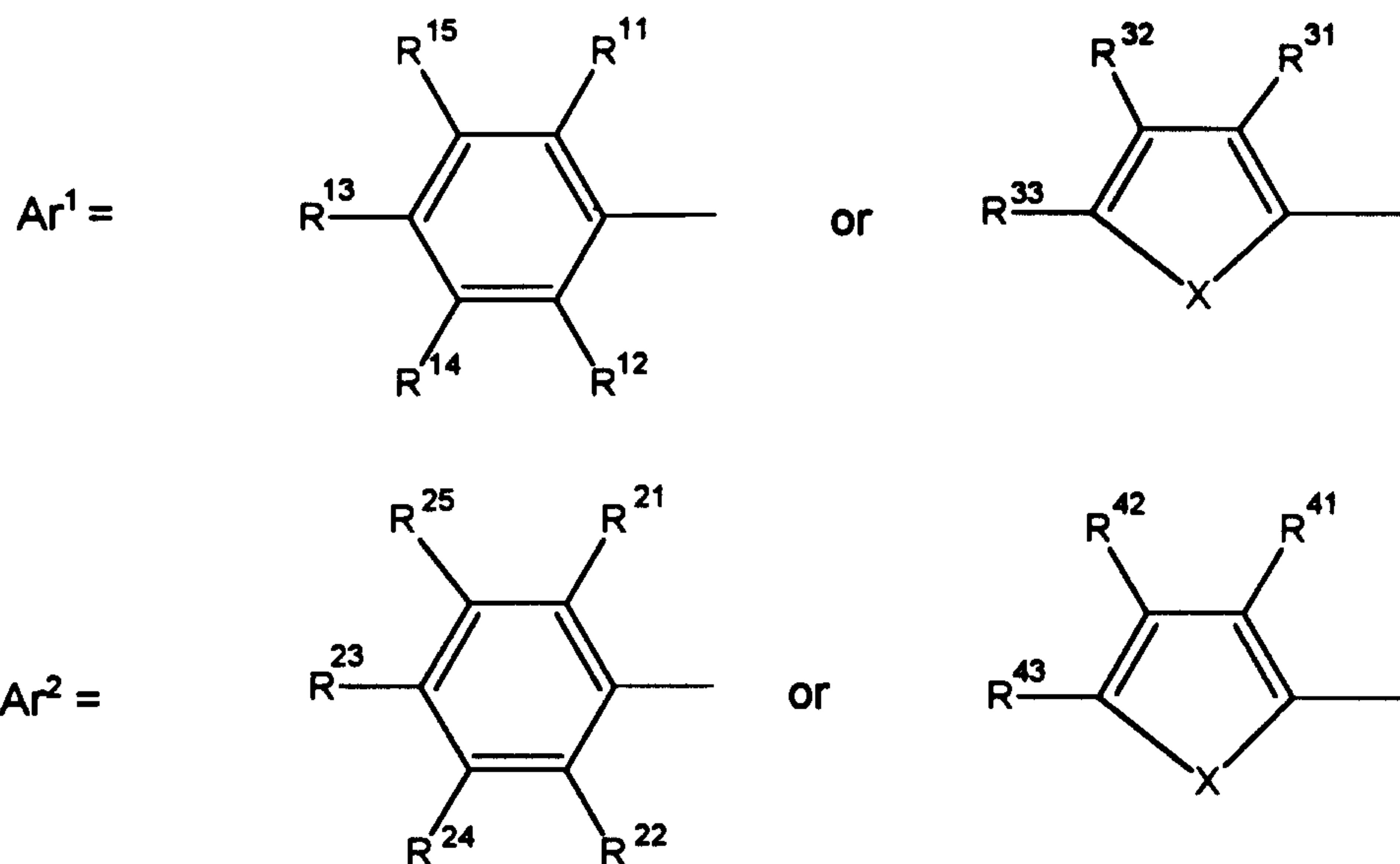
25 Representative compounds which are diaryl tellurides have the formula



The diaryl tellurides result in significant reduction in first slide bias when incorporated into immunoassay elements comprising a horseradish peroxidase labeled immunoreactant such as, the human chorionic gonadotropin (hCG) and C-reactive protein (CRP) immunoassay elements, as
30 described below. A subset of these compounds were tested and shown to also preserve HRP activity in a model system designed to evaluate compounds for their ability to prevent first slide bias.

Useful diorganotellurides include materials where Ar¹ and Ar² are the same or different aryl or heteroaryl groups, which are defined below.

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wherein X is O, S, Se, or Te, and R¹¹, R¹², R¹³, R²¹, R²², R²³, R³¹, R³², R³³, R⁴¹, R⁴² and R⁴³ are the same or different and each selected from the group consisting of hydrogen, alkyl having 1-5 carbon atoms, OH, OR¹, SH, NH₂, NHR¹, NR¹₂, NR¹R², CO₂H or its salts, CO₂R¹, SO₃H and its salts, PO₃H₂ and its salts, and SR¹ wherein R¹ and R² are different and each selected from the group consisting of alkyl having a carbon chain of 1 to 14 carbon atoms optionally carrying one or several hydrophilic groups, phenyl, and substituted phenyl.

R¹⁴, R¹⁵, R²⁴, and R²⁵ are the same or different and each selected from the group consisting of hydrogen, alkyl having 1-5 carbon atoms, alkoxy having 1-5 carbon atoms, CO₂H or its salts, CO₂R¹, SO₃H and its salts, PO₃H₂ and its salts where R¹ is as described above.

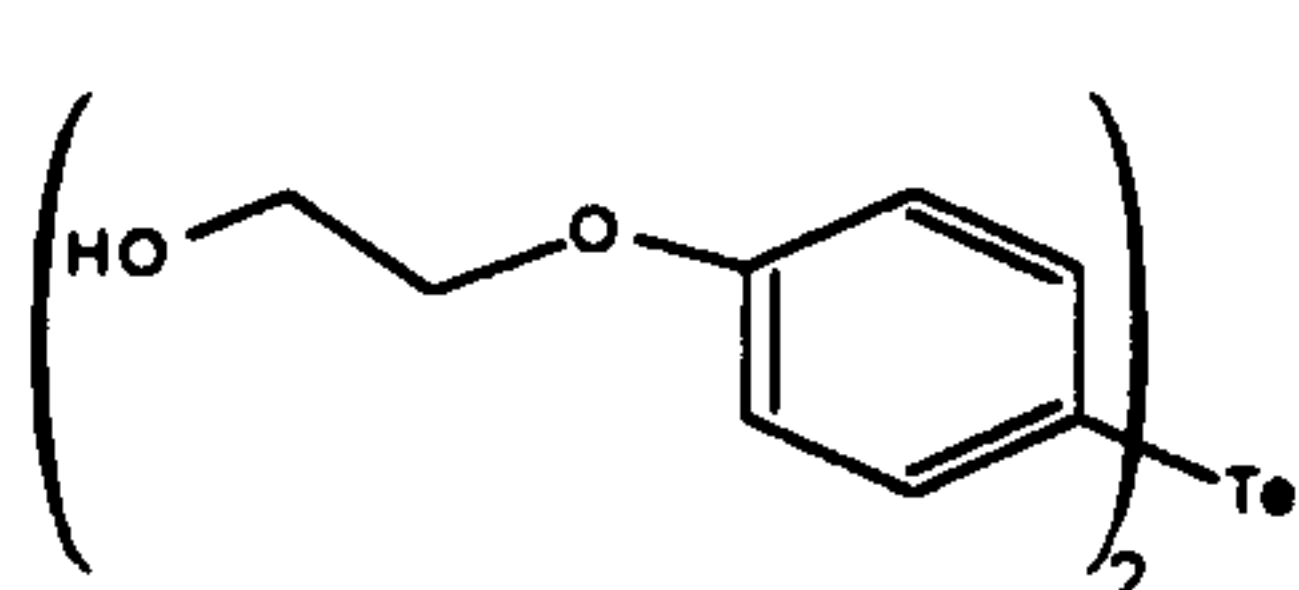
In the above, alkyl shall mean groups such as methyl, ethyl, propyl, isopropyl, n-butyl, sec-butyl, tert-butyl, amyl, isoamyl, or neopentyl. A carbon chain with 1-5 carbon atoms shall mean a straight or branched carbon chain, such as methyl, ethyl, propyl, isopropyl, n-butyl, sec-butyl, tert-butyl, amyl, isoamyl, or neopentyl. A carbon chain with 1-14 carbon atoms shall include, but not be limited to, methyl, ethyl, propyl, isopropyl, n-butyl, sec-butyl, tert-butyl, amyl, isoamyl, neopentyl, hexyl, octyl, decyl, tetradecyl and the like.

Hydrophilic groups mean groups such as sulfonic, phosphonic or carboxylic acid, hydroxyl, and amino groups. Some of these compounds can form salts with either acids or bases. Sodium, potassium, ammonium, calcium and magnesium salts and salts with hydrochloric, hydrobromic,

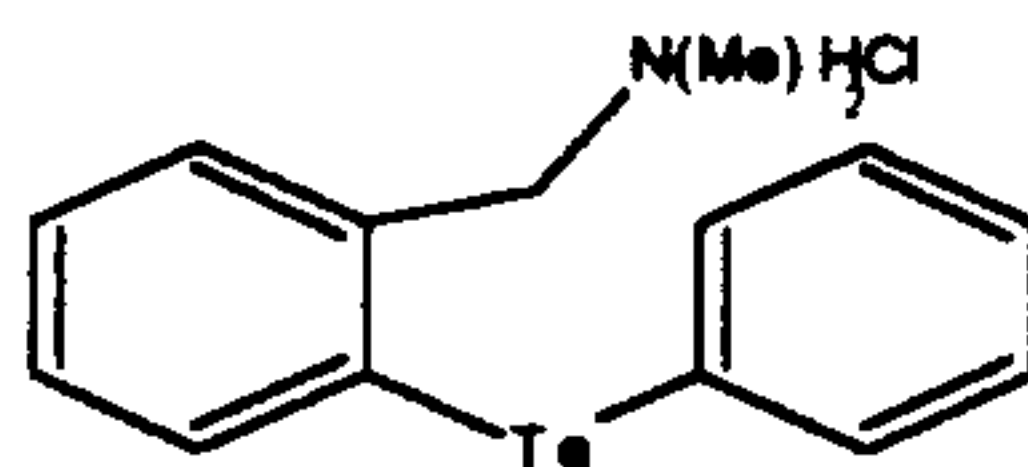
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phosphoric, and sulfuric acids and with organic acids such as oxalic, fumaric, tartaric, malonic, acetic, citric, and succinic acids, are preferred.

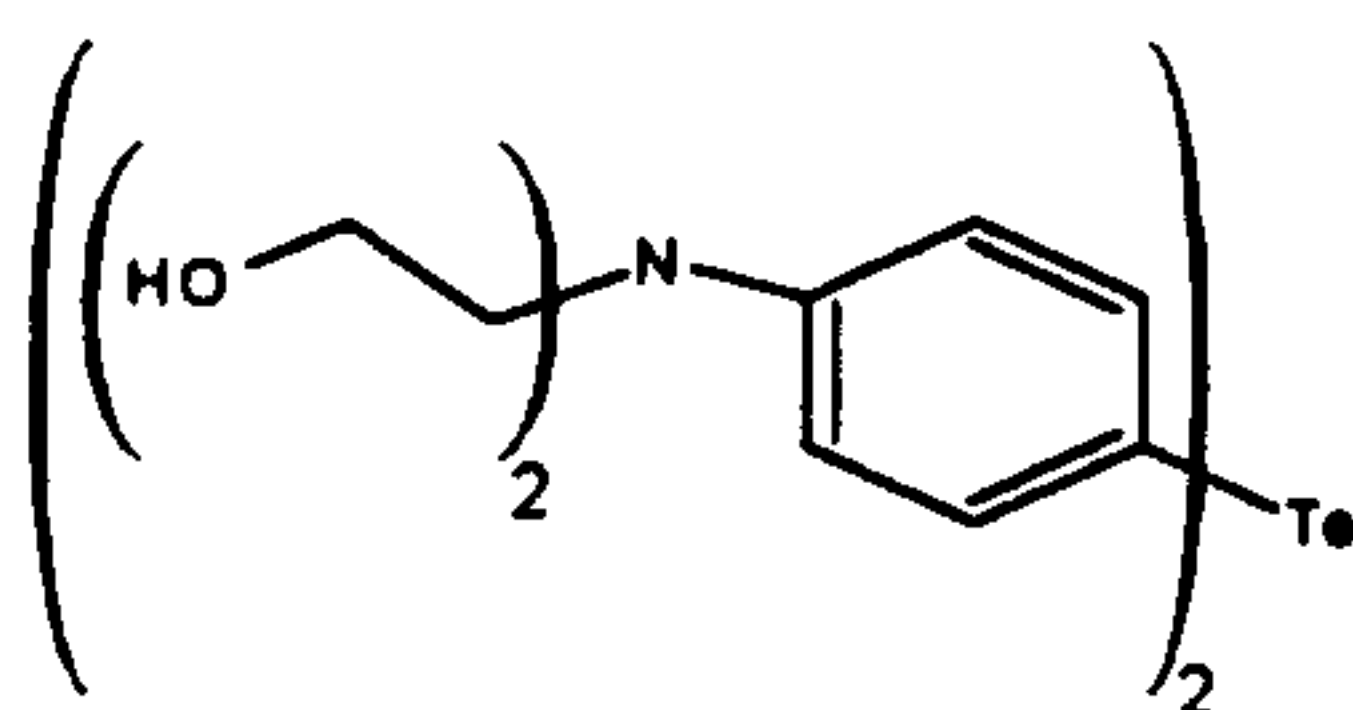
Preferred DATs are those having substituents on the aromatic ring that contain groups that confer water solubility, such as, but not limited to, hydroxyl groups, amines and their salts, and carboxylic acids and their salts. In particular, water soluble DATs that preserve HRP activity either alone or in cooperation with vanadyl compounds are as follows (Te1-Te5):



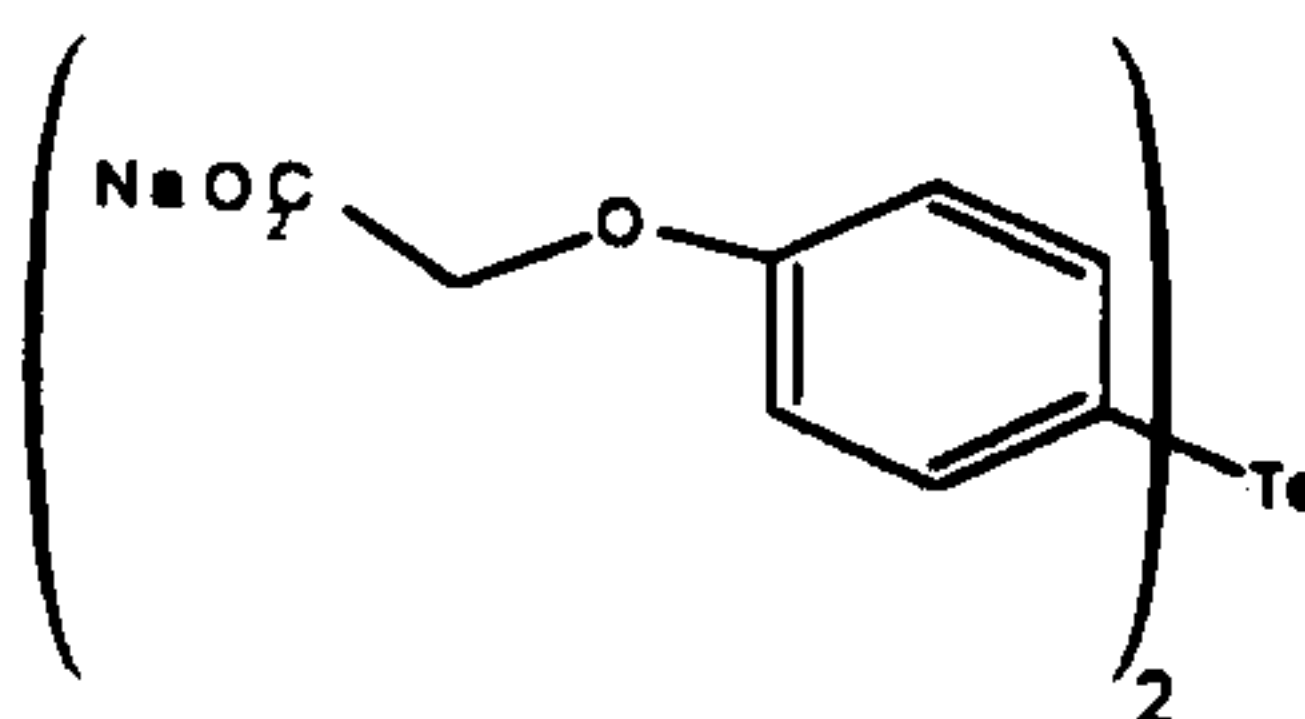
Te1



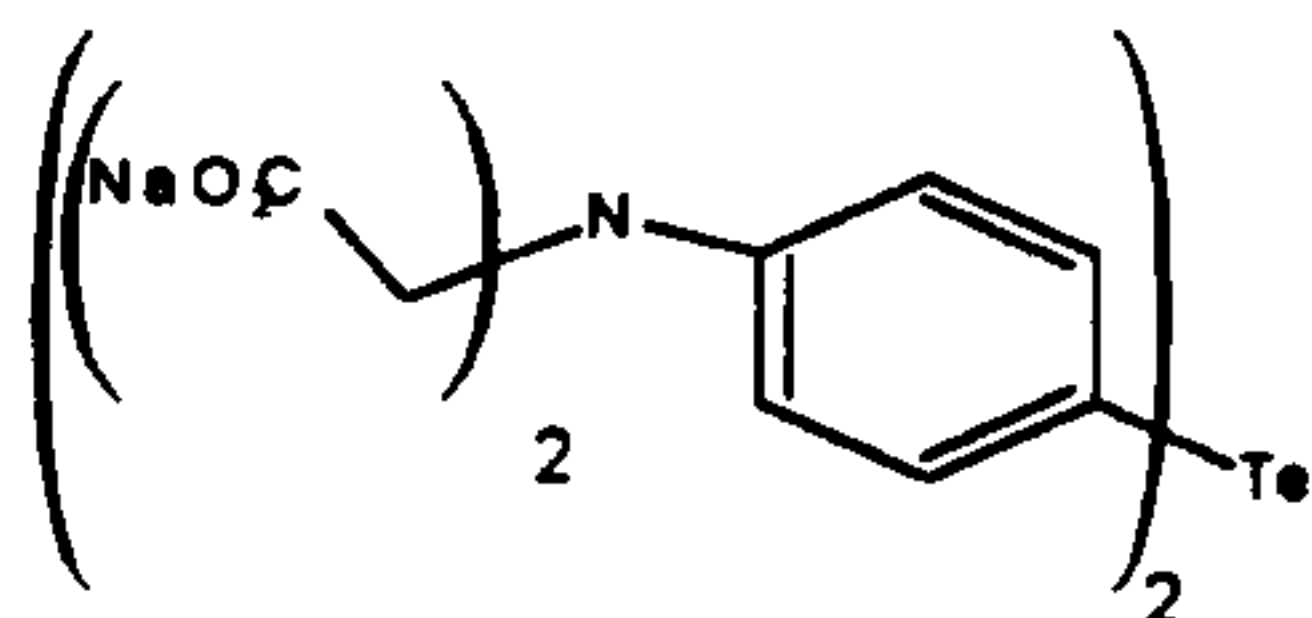
Te2



Te3



Te4



Te5

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Other materials for use in spreading layers are well known in the art of making dry analytical elements as disclosed, for example, in U.S. Patent 4,258,001. Such layers include macroporous layers made from cloth, paper, etc. A preferred particulate layer is a bead spreading layer (BSL). This layer can be easily constructed to have suitable porosity for use in the elements of the present invention to accommodate a test sample (e.g. 1 to 200 μ L), diluted or undiluted. Preferably, the spreading layer is isotropically porous, which property is created by interconnected spaces between the particles comprising the zone. By isotropically porous is meant that the spreading layer uniformly spreads the applied fluid in all directions throughout the layer.

Useful spreading layers, including bead spreading layers are disclosed in U.S. Patents 4,670,381; 4,258,001; and 4,430,436. Particularly useful spreading layers are those having a particulate structure formed by organo-polymeric particles and a polymeric adhesive for those particles described in U.S. Patent 4,258,001. The organo-polymeric particles useful in the spreading layer are generally heatstable, spherical beads having a particle size in the range of from about 10 to 40 μ m in diameter or even smaller.

The particles can be composed of a wide variety of organic polymers, including both natural and synthetic polymers, having the requisite properties. Preferably, however, they are composed of one or more addition polymers described in the aforementioned patents.

When the receptor layer is a separate layer, it is prepared and coated over a support or over a reagent layer or subbing layer on the support. The receptors are covalently bonded to polymer particles through surface reactive groups on the receptor (nucleophilic free amino groups and sulfhydryl groups).

A general procedure for attaching receptors to the small polymer beads includes covalently attaching the selected receptor to the beads using generally known reactions. With many pendant groups, for example the haloalkyl, 2-substituted activated ethylsulfonyl and vinylsulfonyl, the receptor can be directly attached to the beads. Generally, the beads are mixed with the receptor in an aqueous buffered solution, where the pH is generally from about 5 to about 10 and a concentration of from about 0.1 to about 40 weight percent polymer particles, and preferably from about 0.1 to about 10 weight percent. The amount of receptor is at a ratio to polymer of from about 0.1:1000 to about 1:10, and preferably from about 1:100 to about 1:10. Mixing is carried out at a temperature in the range of from about 5 to about 50°C, and preferably at from about 5 to about 40°C, for from about 0.5 to about 48 hours. Any suitable buffer can be used.

In some instances, the pendant reactive groups on the outer surface must be modified or activated in order to cause covalent attachment of the ligand. For example, carboxyl groups must be

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activated using known carbodiimide or carbamoylonium chemistry, described in EP 308235 published 22 July 1992 and U.S. Patent 5,155,166.

The attachment of the receptor to carboxyl group-containing monodispersed polymer beads, however, is carried out in two steps. The first step involves contacting an aqueous suspension of the particles with a carbodiimide or a carbamoylonium compound to produce reactive intermediate polymer particles having intermediate reactive groups in place of the carboxyl groups. This step is carried out at a suitable pH using suitable acids or buffers to provide the desired pH. Generally, the pH is less than 6, but this is not critical as long as the reaction can proceed. More likely, the pH is between about 3.5 and about 7. The molar ratio of carbodiimide or carbamoylonium compound to the carboxyl groups on the surface of the particles is from about 10:1 to 500:1.

In the second step of the method, the reactive intermediate formed in the first step is contacted with a reactive amine- or sulfhydryl-group containing receptor. A covalent linkage is thereby formed between the particles and the receptor. The weight ratio of the receptor to the polymeric particles is generally from about 1:1000 to about 1:1, and preferably from about 1:100 to about 1:10.

In other instances, an epoxy group on the outer surface can be hydrolyzed to form a diol compound capable of reacting with cyanogen bromide which can act as a coupling agent for amine groups in the immunological species. Aldehydes can react directly with amines to form a Schiff's base which can be subsequently reduced to form a covalent link. Alternatively, the aldehyde can be oxidized to an acid and chemistry identified above for carboxyl groups can be used to form an amide linkage.

Any reactive amine- or sulfhydryl-containing receptor can be attached to the monodispersed polymeric beads as long as that receptor contains a reactive amine or sulfhydryl group, respectively, which will react with the reactive groups on the polymer or with the intermediate formed by the reaction of a carbodiimide or a carbamoylonium compound with carboxyl groups on the particles in the case which the polymer has reactive carboxyl groups.

The small polymer beads having reactive groups that readily react directly with the amine or sulfhydryl groups on the receptors are simply mixed with the receptors, in an appropriate buffer if necessary, and allowed to react.

Polymers from which beads for the receptor can be selected include the following: poly(m & p-chloromethylstyrene), poly(styrene-co-m & p chloromethylstyrene-co-2-hydroxyethyl acrylate) (67:30:3 molar ratio), poly(styrene-co-m & p-chloroethylsulfonylmethylstyrene) (95.5:4.5 molar ratio), poly{styrene-co-N-[m & p-(2-chloroethylsulfonylmethyl)phenyl] acrylamide} (99.3:0.7 molar ratio), poly(m & p-chloromethylstyrene-co-methacrylic acid) (95:5, 98:2 and 99.8: 0.2 molar ratio),

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poly(styrene-co-m & p-chloroethylsulfonylmethyl styrene-co-methacrylic acid) (93.5:4.5:2 molar ratio) poly{styrene-co-N-[m & p-(2-chloroethylsulfonylmethyl)phenyl]acrylamide-co-methacrylic acid}(97.3:0.7:2 molar ratio), poly(styrene-co-m & p-chloromethylstyrene)(70:30 molar ratio), poly[styrene-co-3-(p-vinylbenzylthio)propionic acid] (97.6:2.4 molar ratio), poly(styrene-co-vinylbenzyl chloride-co-acrylic acid) (85:10:5 molar ratio), poly(styrene-co-acrylic acid) (99:1 molar ratio), poly(styrene-co-methacrylic acid) (90:10 molar ratio), poly(styrene-co-acrylic acid-co-m & p-divinylbenzene) (89:10:1 molar ratio), poly(styrene-co-2-carboxyethyl acrylate) (90:10 molar ratio), poly(methyl methacrylate-co-acrylic acid) (70:30 molar ratio), poly(styrene-co-m & p-vinylbenzaldehyde)(95:5 molar ratio), and poly(styrene-co-m & p-vinylbenzaldehyde-co-methacrylic acid)(93:5:2 molar ratio).

The layers of the element are carried on a suitable support. The receptor layer is coated over the support although there may be intervening layers, such as a gelatin/buffer layer, between the support and the receptor layer. The support can be any suitable dimensionally stable and, preferably, nonporous and transparent (i.e. radiation transmissive) material which transmits electromagnetic radiation of a wavelength between about 200 and about 900 nm. A support of choice for a particular element should be compatible with the intended mode of detection (reflection, transmission, luminescence or fluorescence spectroscopy). Useful support materials include polystyrene, polyesters [e.g. poly(ethylene terephthalate)], polycarbonates, cellulose esters (e.g. cellulose acetate), etc.

Polymeric binders for the receptor layer are described generally in Canadian patent 1,240,445. Useful polymers are polymers comprising from about 30 to 97 weight percent of polymerized N-alkyl substituted acrylamide such as N- isopropylacrylamide. Other useful N-alkyl-substituted acrylamides include N-n-butylacrylamide, N,N-diethylacrylamide and N-n-propylacrylamide. Poly(N-isopropylacrylamide-co-methacrylic acid-co-N,N'-methylenebisacrylamide is used in the examples to illustrate the utility of these binders.

The polymer binder also comprises from about 3 to 25 weight percent of one or more polymerized crosslinking monomers having at least two addition-polymerizable groups per molecule. These crosslinking monomers are generally well known in the art. The preferred crosslinking monomers contain acrylamido or methacrylamido groups to facilitate polymerization with the N-alkyl-substituted acrylamides.

Examples of useful crosslinking monomers include:

N,N'-methylenebisacrylamide;

N,N'-methylenebismethacrylamide;

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ethylene dimethacrylate;
 2,2-dimethyl-1,3-propylene diacrylate;
 divinylbenzene;
 mono[2,3-bis(methacryloyloxy)propyl] phosphate;
 5 N,N'-bis(methacryloyl)urea;
 triallyl cyanurate;
 5 allyl acrylate;
 allyl methacrylate;
 N-allylmethacrylamide;
 10 4,4'-isopropylidenediphenylene diacrylate;
 1,3-butylene diacrylate;
 1,4-cyclohexylenedimethylene dimethacrylate;
 2,2'-oxydiethylene dimethacrylate;
 divinylloxymethane;
 15 ethylene diacrylate;
 ethylidene diacrylate;
 propylidene dimethacrylate;
 1,6-diacrylamidohexane;
 1,6-hexamethylene diacrylate;
 20 1,6-hexamethylene dimethacrylate;
 phenylethylene dimethacrylate;
 tetramethylene dimethacrylate;
 2,2,2-trichloroethylidene dimethacrylate;
 ethylenebis(oxyethylene) diacrylate;
 25 ethylenebis(oxyethylene) dimethacrylate;
 ethylidyne trimethacrylate;
 propylidyne triacrylate;
 vinyl allyloxyacetate;
 1-vinyloxy-2-allyloxyethane;
 30 2-crotonoyloxyethyl methacrylate,
 diallyl phthalate; and
 2-(5-phenyl-2,4-pentadienoyloxy)ethyl methacrylate.

These polymeric binders can also include 0 to 60 weight percent of polymerized hydrophilic monomers. Amounts of 5 to 35 weight percent are also useful. Hydrophilic monomers

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are disclosed in Canadian patent 1,240,445. In particular, such monomers have one or more groups selected from hydroxy, pyrrolidone, amine, amide, carboxy, sulfo, carboxylate salt, sulfonate salt and sulfate salt groups. Generally the counter ions of the salt groups are alkali metal or ammonium. Useful hydrophilic monomers are acrylic acid and methacrylic acid and their salts, sodium 2-

Further, the recited binders make it possible to form uniform coatings of receptor layers due to the very low binder viscosities achieved from sheer thinning during extrusion hopper coating. A further advantage is achieved with the recited binders in that, immediately after forming uniform coatings, the viscosity of the binders increases substantially, resulting in a "set layer" that remains stable and uniform during wet transport and drying of the binders.

The receptors also can be dispersed in a polymer binder selected from the group consisting of:

- poly(vinyl alcohol);
- bovine serum albumin;
- acacia gum;
- homopolymers of N-vinylpyrrolidone

having a molecular weight in the range 8000 to 400,000; and water-soluble vinyl addition copolymers having two or more monomers selected from the group consisting of acrylamide, methacrylamide, N-alkyl-substituted acrylamides, N-alkyl substituted methacrylamides, 1-vinylimidazole, 2-alkyl substituted- 1-vinylimidazoles, 2-hydroxyalkyl substituted-1-vinylimidazoles, N-vinylpyrrolidone, hydroxyalkyl acrylates, hydroxyalkyl methacrylates, acrylic acid, and methacrylic acid; wherein alkyl and hydroxyalkyl in the copolymers has 1 to 6 carbon atoms such methyl, ethyl, propyl and hexyl.

The element can comprise one or more additional layers, e.g. separate or combined reagent/spreading layer and a gelatin/buffer layer containing other necessary additives such as electron transfer agents.

The gelatin/buffer layer or the reagent layer or the spreading layer of the element can contain the indicator composition comprising one or more reagents dispersed in one or more synthetic or natural binder materials, such as gelatin, or other naturally-occurring colloids, homopolymers and copolymers, such as poly(acrylamide), poly(vinylpyrrolidone), poly(N-isopropylacrylamide), poly(acrylamide-co-N-vinyl-2-pyrrolidone) and similar copolymers. The indicator composition also can be dispersed in the receptor layer.

Other optional layers, e.g. subbing layers, radiation-blocking layers, etc. can be included if desired. All layers of the element are in fluid contact with each other, meaning that fluids and

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reagents and uncomplexed reaction products in the fluids can pass between superposed regions of adjacent layers.

The layers of the element can contain a variety of other desirable but optional components, including surfactants, thickeners, buffers, hardeners, antioxidants, coupler solvents, and other materials known in the art. The amounts of these components are also within the skill of a worker in the art.

The elements can be used to determine low concentrations of immunologically reactive ligands in a liquid, such as a biological fluid (e.g., whole blood, serum, plasma, urine, spinal fluid, suspensions of human or animal tissue, feces, saliva, lymphatic fluid and the like). The ligands can be determined at concentrations as low as about 10^{-15} molar, and most generally at a concentration of from about 10^{-11} to about 10^{-4} molar.

Ligands which can be so determined, either quantitatively or qualitatively, include therapeutic drugs (e.g., phenobarbital, digoxin, digitoxin, theophylline, gentamicin, quinidine, phenytoin, propranolol, carbamazepine, tobramycin, lidocaine, procainamide and the like), natural or synthetic steroids (e.g. cortisol, aldosterone, testosterone, progesterone, estriol, etc.), hormones (e.g. thyroid hormones, peptide hormones, insulin, etc.), proteins (e.g. albumin, IgG, IgM, ferritin, blood clotting factors, C-reactive protein, isoenzymes, hCG, apolipoproteins, etc.), antigens, antibodies including monoclonal antibodies, and other species which will naturally react with a receptor. This invention is particularly useful for the determination of therapeutic drugs, such as digoxin, phenytoin, carbamazepine, theophylline, or phenobarbital, hormones such as thyroxine or triiodothyronine and analytes such as hCG and C-reactive protein. The assay can be carried out using any enzyme label which can be attached to the ligand to form a labeled ligand. Enzymes, such as glucose oxidase, peroxidases such as horseradish peroxidase (HRP), alkaline phosphatase and galactosidase are preferred labels.

It is within the skill of the ordinary worker in clinical chemistry to determine a suitable substrate for a given label. The substrate can be a material which is directly acted upon by the enzyme label or a material that is involved in a series of reactions which involve enzymatic reaction of the label. For example, if the enzyme label is a peroxidase, the substrate is hydrogen peroxide plus an appropriate reducing agent. Using glucose oxidase as an example, the substrate glucose is generally present in the reagent layer or added as a substrate solution to yield about 0.01 mole/m^2 , and preferably from about 0.001 to about 0.1 mole/m^2 . A worker skilled in the art would know how to adjust the amount of a particular substrate for the amount of enzyme label used in the assay.

The reagent layer may contain an indicator composition comprising one or more reagents which provide a detectable species as a result of the reaction catalyzed by the label. The detectable

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species could develop a color, be radioactive, fluorescent, or be chemiluminescent. For present purposes the invention is illustrated using a colorimetric indicator composition which provides a colorimetrically detectable species as a result of enzymatic reaction of an enzyme labeled ligand analog with a substrate.

5 The indicator composition can be a single compound which produces a detectable dye upon enzymatic reaction or a combination of reagents which produce the dye. For example, when glucose is used as the substrate and glucose oxidase as the enzyme label, the colorimetric indicator composition can include a coupler and an oxidizable compound which react to provide a dye. Alternatively, the composition can include a leuco dye and peroxidase or another suitable
10 peroxidative compound which generates a detectable dye as a result of the formation of hydrogen peroxide produced when glucose oxidase converts glucose to gluconic acid. Useful leuco dyes are known in the art and include those, for example, described in U.S. Patent 4,089,747 (issued May 16, 1978 to Bruschi) and U.S. Patent No. 4,670,385 (issued 2 June 1987 to Babb et al). The particular amounts of the colorimetric indicator composition and its various components are within the skill of a
15 worker in the art.

 The labeled ligands can be prepared using known starting materials and procedures, or obtained commercially. Generally, the ligand is attached to the label (e.g. an enzyme moiety) through a covalent bond.

 The immunoassay can be manual or automated. In general, the amount of a ligand in a
20 liquid is determined by taking the element from a supply roll, chip packet or other source and physically contacting a finite area of the spreading layer with a sample of the liquid, e.g. 1 to 200 μL . The finite area which is contacted is generally no more than about 150 mm^2 .

 The amount of ligand is determined by passing the element through a suitable apparatus for detecting the complexed ligand analog directly or the detectable species formed as a result of
25 enzymatic reaction of an enzyme label and a substrate. For example, the species can be detected with suitable spectrophotometric apparatus using generally known procedures. In an enzymatic reaction, the resulting product is determined by measuring, for example, the rate of change of reflection or transmission density in the finite area which was contacted with the test sample. The area which is measured has a diameter of generally from about 3 to about 5 mm. The amount of ligand in the
30 liquid sample is inversely proportional to the amount of label measured in the finite area in the case of a competitive assay, or is directly proportional in the case of a sandwich assay. Generally, label measurement is made after application of a substrate solution.

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Synthesis of the diaryl tellurides.

There exist many synthetic routes to prepare DATs. The route used for a particular compound depends heavily on the substituents on the aromatic rings. Several of the known routes are illustrated in the examples below. Extrapolation of these and other literature methods to the synthesis of other DATs would be readily apparent to one skilled in the art.

Preparation 1. 4,4'-Di(2-hydroxyethoxy)-1,1'-tellurobisbenzene (Te1)

(a) Preparation of [2-(4-Bromophenoxy)ethoxy](1,1-dimethylethyl)dimethylsilane (1a)

To a solution of [2-(4-bromophenoxy)ethanol (10.0 g, 46 mmol) in 150 mL dry dimethylformamide (30 mL) was added t-butyldimethylsilyl chloride (8.34 g, 55 mmol) and imidazole (7.82 g, 115 mmol). Additional dimethylformamide (20 mL) was used to wash these materials into the reaction flask. The resulting solution was stirred overnight at room temperature with a Newman tube. It was poured onto water (200 mL) and extracted with ether (3X 75 mL). The combined extracts were washed with 0.5 N HCl (200 mL), sat. NaHCO₃ (200 mL), water (4X 150 mL), and sat. NaCl (200 mL), dried over MgSO₄, and filtered. The solvent was removed under reduced pressure on a rotary evaporator to give the crude product (16.5 g; >100%). ¹H NMR (CDCl₃) δ 7.34 (2H, d, J=8.9), 6.78 (2H, d, J=8.9), 4.00-3.93 (4H, m), 0.89 (9H, s), 0.08 (6H, s). ¹³C NMR (CDCl₃) δ 158.1, 132.2, 116.4, 112.8, 69.6, 61.9, 25.9, 18.4. FDMS (m/e) 330 (M⁺, ⁷⁹Br). It was used as 100% without purification.

(b) Preparation of 4,4'-Di(2-hydroxyethoxy)-1,1'-tellurobisbenzene (Te1)

A 500 mL 3-neck flask, was placed under argon and equipped with a condenser and an addition funnel. Magnesium turnings (0.89 g, 37 mmol) were added. Bromide 1a (12.3 g, 37 mmol, used as 100%) was taken up in dry tetrahydrofuran (THF) (100 mL), and transferred to the addition funnel. The Grignard reaction was initiated with approximately 10 mL of the bromide solution, 1,2-dibromoethane, and iodine. The remaining bromide solution was added and the reaction was refluxed overnight, after which time no Mg remained. The heating mantle was removed for 30 minutes, then tellurium granules (4.74 g, 37 mmol) were added, and the reaction was heated back to reflux for 7 h, at which time nearly all the Te had been consumed. The reaction was cooled to rt, then poured onto rapidly stirring 10% aq. NH₄Cl (300 mL) and stirred for 15 min. Precipitated Te was removed by filtration through Celite diatomaceous earth and the filter cake was washed with ether. The filtrate was transferred to a separatory funnel and extracted three times with ether (200

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mL, 100 mL, 100 mL). The combined extracts were washed with water and sat. NaCl, dried over MgSO₄, and filtered. The solvent was removed under reduced pressure to give the crude product (12.4 g) as a red oil. It was taken up in toluene (50 mL) with copper powder (2.5 g), and heated to reflux with a Newman tube for 2.5 h, at which time the color had turned from red to gray. The reaction was cooled to rt, filtered through Celite diatomaceous earth, washed with ether, and concentrated, giving an amber oil (12.4 g). By ¹H NMR, it was a mixture of the desired product, quenched Grignard, and residual toluene. It was taken up in methanol (50 mL) and (50 mL) and THF (20 mL) with potassium fluoride (5.0 g, 86 mmol), and refluxed for 24 h. The bulk of the methanol was removed under reduced pressure, and partitioned between ether-ethyl acetate and water. The aqueous phase was extracted twice more with ethyl acetate, and the combined extracts were washed with water and sat. NaCl, dried over Na₂SO₄, and filtered. The solvent was removed under reduced pressure to give the crude product (8g) as a tan solid. Ether trituration gave a white solid that was isolated by filtration, washed with ether, and air dried, to give the crude product (1.74 g). It was adsorbed onto flash silica gel (20 mL) using dichloromethane. Careful flash chromatography on 200 mL flash silica gel, eluting with dichloromethane then 95:5 dichloromethane:methanol (the product elutes just before a small amount of the corresponding biaryl compound), ether trituration and filtration gave pure as a white solid (1.2 g). The ether filtrate from the trituration of the crude product was chromatographed as above. Ether trituration and recrystallization from ethanol gave additional pure Te1 (0.2 g). Total yield: 1.4 g, 19%. ¹H NMR (DMSO-d₆) δ 7.52 (4H, d, J=8.5), 6.80 (4H, d, J=8.5), 4.82 (2H, t, J=5.5), 3.91 (4H, t, J=5.0), 3.65 (4H, app q, J=5.1). ¹³C NMR (DMSO-d₆) δ 159.2, 139.8, 116.5, 104.5, 69.9, 59.9. FDMS (m/e) 404 (M⁺, ¹³⁰Te).

Preparation 2. N,N-Dimethyl-2-(phenyltelluro)benzenemethanamine, Hydrochloride Salt (Te2)

(a) Preparation of N,N-Dimethyl-2-(phenyltelluro)benzenemethanamine (2a)

To a solution of N,N-dimethylbenzenemethanamine (2.70 g, 0.020 mol) in dry ether (50 mL) in a round-bottom flask under argon at rt was added n-BuLi (2.5 M, 10 mL, 0.025 mol) dropwise by syringe. The reaction mixture was stirred 5h at rt, then a solution of phenyltellurenyl bromide in dry tetrahydrofuran (0.5 M) was added dropwise by syringe. After the addition of 38 mL (0.019 mol), the reaction mixture turned the characteristic orange of the phenyltellurenyl bromide, and the addition was stopped. The reaction mixture was poured onto ether (100 mL), and the resulting solution was washed with sat. NaCl (1X 100 mL, 2X 50 mL), dried over MgSO₄, and

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concentrated. The residue was dissolved in acetone (100 mL), and iodine (5.08 g, 0.020 mol) was added. The resulting solution was chilled to precipitate the product. The yellow crystals were collected by filtration, washed with cold acetone, and dried to give the iodine adduct of 2a (5.95 g, 50%, mp 178-179°). The iodine adduct (5.93 g, 0.010 mol) was dissolved in dimethylformamide (100 mL). Sodium bisulfite (5.2 g, 0.05 mol) in water (100 mL) was slowly added, and the reaction mixture was stirred 1 h at rt, during which time the reaction mixture became colorless. The reaction mixture was poured onto water (500 mL), then washed with ether (2X 50). The aqueous layer was made basic with 10% NaOH, and the amine was extracted into ether (3X 100 mL). The combined extracts were washed with sat. NaCl, dried over MgSO₄, and concentrated. The residue was recrystallized from methanol to give the pure product (3.14 g, 93%) as a white solid (mp 51-54°). ¹H NMR (CDCl₃) δ 7.88 (2H, d, J=6.9), 7.35 (1H, t, J=7.3), 7.26 (2H, t, J=7.3), 7.17 (1H, d, J=7.7), 7.10-7.04 (2H, m), 6.94-6.89 (1H, m), 3.53 (2H, s), 2.25 (6H, s).

(b) Preparation of N,N-Dimethyl-2-(phenyltelluro)benzenemethanamine,
Hydrochloride Salt (Te2)

To a slurry of 2-(N,N-dimethylaminomethyl)-1-phenyltellurobenzene (1.02 g, 3.0 mmol) in ether (50 mL) in a water bath was added dropwise by syringe a hydrogen chloride/ether solution (3.15 mL, 1.0 M, 3.15 mmol). The thick slurry was diluted with isopropanol (20 mL) then filtered. The white solid was washed with ether, and air dried, to give the product (1.13 g, 95 %). ¹H NMR (CDCl₃) δ 2.6 (1H, br s), 8.13 (1H, d, J=7.5), 7.80 (1H, d, J=7.7), 7.52-7.48 (3H, m), 7.26-7.17 (4H, m), 4.44 (2H, d, J=5.8), 2.68 (6H, d, J=4.5). Anal. calc'd for C₁₅H₁₈ClN₂Te: C, 48.00; H, 4.83; N, 3.73. Found: C, 47.97; H, 4.87; N, 3.65.

Preparation 3. N,N,N',N'-Tetra(2-hydroxyethyl)-4,4'-tellurobisbenzenamine (Te3)

(a) Preparation of
N,N-Bis[2-[(1,1-dimethylethyl)dimethylsilyl]oxy]ethyl]benzenamine (3a)

A procedure analogous to that used for 1a was used with the following materials: 2,2'-(phenylimino)diethanol (9.0 g, 50 mmol), t-butyltrimethylsilyl chloride (18.1 g, 0.12 mol), imidazole (17.0 g, 0.25 mol), and dimethylformamide (50 mL). The reaction was stirred overnight for convenience. Workup as in 1a gave 3a as a pale oil (21.6 g, >100%). ¹H NMR (CDCl₃) δ 7.21 (2H, d, J=8.4), 6.68 (3H, overlapping d, t), 3.77 (4H, d, J=6.6), 3.52 (4H, d, J=6.6), 0.92 (18H, s),

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0.06 (12H, s). ^{13}C NMR (CDCl_3) δ 147.8, 129.2, 115.7, 111.4, 60.3, 53.5, 25.9, 18.3, -5.3. FDMS (m/e) 409 (M^+). It was used as 100% without purification.

5 (b) Preparation of N,N,N',N'-Tetra[2-[(1,1-dimethylethyl)dimethylsilyl]oxy]ethyl]-4,4'-tellurobisbenzenamine (3b)

To a 500 mL 3-neck flask, oven-dried and put under argon was added tellurium (IV) chloride (5.93 g, 22 mmol). Dry ether (100 mL) was added by syringe, giving a thin yellow slurry, and the flask was placed in a water bath. A solution of 3a (18 g, 44 mmol) in dry ether (50 mL) was prepared under argon, and the solution was transferred by cannula to the reaction mixture, with vigorous stirring. Initially a very thick yellow precipitate formed, which thinned out as the addition of the aniline solution continued. At the completion of the addition, the greenish yellow slurry was stirred overnight. The reaction mixture was filtered, and the filtrate evaporated under reduced pressure. The resulting yellow-green oil was taken up in dichloromethane (200 mL), and a solution of sodium metabisulfite (8.36 g, 44 mmol) in water (200 mL) was added with stirring. The resulting two-phase mixture was stirred 30 min at rt, then filtered through Celite diatomaceous earth. Solid NaHCO_3 was added to pH 9, then the reaction mixture was transferred to a separatory funnel. The dichloromethane layer was split off, and the aqueous layer was extracted with additional dichloromethane. The combined extracts were washed with water / sat. NaCl (200 mL / 50 mL) then sat. NaCl, dried over Na_2SO_4 , and filtered, and the solvent was removed under reduced pressure. The resulting red oil (21.4 g) was taken up in toluene (80 mL), and copper powder (4 g) was added. The reaction mixture was heated to reflux with a Newman tube overnight. The resulting gray reaction mixture was cooled to room temperature, filtered through Celite diatomaceous earth, and the solvent was removed under reduced pressure. The resulting amber oil (18 g) was flash chromatographed twice with 3:1 cyclohexane : dichloromethane to separate the product from recovered starting material, yielding pure 3b as a yellow oil (5.0 g, 48% of theory). ^1H NMR (CDCl_3) δ 7.54 (4H, d, $J=8.6$), 6.53 (4H, d, $J=8.5$), 3.72 (4H, t, $J=6.4$), 3.47 (4H, t, $J=6.5$), 0.88 (18H, s), 0.03 (12H, s). ^{13}C NMR (CDCl_3) δ 147.6, 139.7, 112.7, 98.2, 60.2, 53.4, 25.9, 18.3, -5.3. FDMS (m/e) 946 (M^+ , ^{130}Te).

30 (c) Preparation of N,N,N',N'-Tetra(2-hydroxyethyl)-4,4'-tellurobisbenzenamine (Te3)

A heterogeneous mixture of silyl ether 3b (2.7 g, 2.86 mmol), potassium fluoride (0.66 g, 11.4 mmol), and methanol (20 mL) was heated to reflux for 20 h. The white slurry was

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cooled in an ice bath, then the product was isolated by filtration, washed with cold methanol, and air dried (0.80 g, 57%). A small amount was recrystallized from methanol for an analytical sample (mp 169-170°). ¹H NMR (DMSO-d₆) δ 7.54 (4H, d, J=8.6), 6.53 (4H, d, J=8.5), 3.45 (4H, app q, J=5.7), 3.33 (4H, t, J=5.9). ¹³C NMR (DMSO-d₆) δ 148.1, 139.8, 113.1, 97.6, 58.4, 53.5. IR (KBr) 3350, 1585, 1495, 1350 cm⁻¹. FDMS (m/e) 490 (M⁺, ¹³⁰Te). Anal. calc'd for C₂₀H₂₈N₂O₄Te: C, 49.22; H, 5.78; N, 5.74. Found: C, 48.78; H, 5.72; N, 5.66.

Preparation 4. 2,2'-[Tellurobis(4,1-phenyleneoxy)]bisacetic Acid, Disodium Salt (Te4)

(a) Preparation of 4-(Bromophenoxy)(1,1-dimethylethyl)dimethylsilane (4a)

To a solution of p-bromophenol (104 g, 0.6 mol) in dry dimethylformamide (450 mL) was added t-butyldimethylsilyl chloride (108 g, 0.72 mol) and imidazole (102 g, 1.5 mol). The resulting pale yellow solution was stirred 3h at rt with a Newman tube. It was poured onto water (1.2 L) and extracted three times with ether (900 mL, 300 mL, 300 mL). The combined extracts were washed four times with water (150 mL), once with 1 N HCl (300 mL), sat. NaHCO₃ (300 mL), and sat. NaCl (300 mL), dried over MgSO₄, and filtered. The solvent was removed under reduced pressure on a rotary evaporator then a vacuum pump to give the crude product (183 g; >100%), as a 6:1 ratio with the silanol from hydrolyzed starting material. ¹H NMR (CDCl₃) δ 7.31 (2H, d, J=8.7), 6.71 (2H, d, J=8.7), 0.97 (9H, s), 0.18 (6H, s). It was used as 100% without purification.

(b) Preparation of 4,4'-Di(t-butyldimethylsilyloxy)-1,1'-tellurobisbenzene (4b)

A 2L 3-neck flask, oven dried then placed under argon, was equipped with an overhead stirrer and an addition funnel. Magnesium turnings (15.2 g, 0.66 mol) were added. Bromide 4a (17.2 g, 0.6 mol, used as 100%) was taken up in dry THF (600 mL), and transferred to the addition funnel. The Grignard reaction was initiated with approximately 25 mL of the bromide solution and several crystals of iodine. The remaining bromide solution was added at such a rate that the reaction continued to reflux. The reaction mixture was refluxed for an additional 30 min after the addition was complete, at which time only a small amount of Mg remained. The reaction was cooled slightly in a water bath, tellurium granules (76.8 g, 0.6 mol) were added, and the reaction was heated back to reflux for 2.5 h, at which time nearly all the Te had been consumed. The reaction was cooled to rt, then poured onto rapidly stirring 10% aq. NH₄Cl (2 L) and stirred for 30 min. Precipitated Te was removed by filtration through Celite diatomaceous earth, and the filter cake was washed with

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ether. The filtrate was transferred to a separatory funnel and extracted three times with ether (1200 mL, 300 mL, 300 mL). The combined extracts were washed with water and sat. NaCl, dried over MgSO₄, and filtered through Celite diatomaceous earth to remove both the MgSO₄ and Te which had precipitated during the workup. The solvent was removed under reduced pressure to give the crude product (194 g) as a red oil. It was taken up in toluene (900 mL) with copper powder (38 g), and heated to reflux with a Newman tube for 2.5 h, at which time the color had turned from red to gray. The reaction mixture was cooled to rt, filtered through Celite diatomaceous earth, and concentrated, giving an amber oil (181 g). By ¹H NMR, it was a mixture of the desired product, quenched Grignard, and residual toluene, and was calculated to contain 0.23 mol of 4b. Purification was effected via the iodine adduct of 4b. The crude product was taken up in acetone (0.6 L), and iodine (58 g, 0.23 mol) was added portionwise with vigorous stirring. After 15 min, a very thick precipitate had formed. Ethanol (1.2 L) was added, and the orange solid and dark iridescent crystalline solid were isolated by filtration and air-dried. A second crop was obtained by refiltration of the filtrate. The combined yield was 206 g, and it was pure by NMR, the only contaminant being residual ethanol. ¹H NMR (CDCl₃) δ 7.95 (4H, d, J=8.7), 6.86 (4H, d, J=8.7), 0.98 (18H, s), 0.25 (12H, s). ¹³C NMR (CDCl₃) δ 158.6, 138.4, 121.9, 25.5, 18.2, -4.3. The iodine adduct was taken up in dioxane (500 mL), dichloromethane (500 mL), and 5% aq. NaHSO₃ (1.1 L) and the two-phase system was stirred vigorously. After 1 h, the lower (organic) layer was still red, indicative of incomplete reduction. The bulk of the yellow aqueous layer was decanted and saved, and the organic layer was evaporated under reduced pressure. Dioxane (200 mL), dichloromethane (200 mL), and 5% aq. NaHSO₃ (400 mL) were added to the residue, and the mixture was stirred an additional hour. It was evaporated under reduced pressure to partially remove the organic solvents, and the remaining aqueous dioxane was transferred to a separatory funnel and extracted three times with ether. The combined extracts were washed with water, sat. NaHCO₃, and sat. NaCl, dried over MgSO₄, and filtered. The decanted aqueous layer from above was worked up in the same way, and the two were combined. The solvent was removed under reduced pressure to give pure 4b (97.3 g, 60%) as a mber oil. ¹H NMR (CDCl₃) δ 7.54 (4H, d, J=8.4), 6.80 (4H, d, J=8.4), 0.96 (18H, s), 0.17 (12H, s). ¹³C NMR (CDCl₃) δ 155.8, 139.6, 121.5, 105.2, 25.7, 18.2, -4.4. FDMS 5(m/e) 544 (M⁺, ¹³⁰Te).

(c) Preparation of 4,4'-Tellurobisphenol (4c)

Silyl ether 4b (54.2 g, 0.1 mol) was taken up in methanol (200 mL) in a 500 mL round bottom flask. Potassium fluoride (11.6 g, 0.2 mol) was added, and the reaction mixture was heated to reflux for 1.5 h under argon. After the reaction mixture was cooled to room temperature, it

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was poured into rapidly stirring water (1.2 L), using additional methanol (20 mL) to facilitate the transfer. The pH was adjusted to 14 with 10% NaOH (approx. 40 mL), and the reaction mixture was filtered through Celite diatomaceous earth. The filtrate was transferred to a separatory funnel and washed twice with dichloromethane. The aqueous layer was transferred to a 2 L Erlenmeyer flask in an ice bath, and 25% aq. acetic acid was added to pH 6, resulting in formation of a cream-colored precipitate. The product was isolated by filtration, washed with water, and air dried, to give the crude product (29 g). It was adsorbed onto flash silica gel (150 mL) using ether. Vacuum chromatography on 1 L flash silica gel, eluting with dichloromethane then 1:4 ethyl acetate : dichloromethane gave the pure product. Trituration with dichloromethane gave, in two crops, pure 4c as a yellow solid (24 g, 76%). ¹H NMR (CDCl₃, 5 drops DMSO) δ 8.77 (2H, s), 7.29 (4H, d, J=8.4), 6.47 (4H, d, J=8.4).

(d) Preparation of 4,4'-Di(ethoxycarbonylmethoxy)-1,1'-tellurobisbenzene (4d)

A 250 mL oven-dried 3-neck flask equipped with an overhead stirrer and an addition funnel was put under argon, and sodium hydride (60%, 1.76 g, 44 mmol) was added. It was washed with cyclohexane three times, then dry (sieves) dimethylformamide (60 mL) was added. The flask was placed in a water bath, and a solution of bisphenol 4c (6.28 g, 20 mmol) in dry dimethylformamide (15 mL) was added dropwise from the addition funnel (vigorous H₂ evolution). The resulting slurry was heated in an oil bath to 85° for 20 min (additional H₂ evolution). The slurry was cooled to rt, and a solution of ethyl bromoacetate (6.68 g, 40 mmol) in dry dimethylformamide (5 mL) was added dropwise from the addition funnel, during which time the precipitate cleared. The reaction mixture was stirred 45 min, then was poured onto rapidly stirring water (320 mL). The resulting tan precipitate was isolated by filtration and washed with water. It was partitioned between water (50 mL) and dichloromethane (80 mL). The aqueous phase was extracted twice more with dichloromethane, and the combined extracts were washed with water then sat. NaCl, dried over Na₂SO₄, and filtered. The solvent was removed under reduced pressure, and the resulting solid was recrystallized from ethanol (50 mL), chilled, and filtered. The product was obtained as a white solid (5.3 g, 55%, mp 88.5-89°). ¹H NMR (CDCl₃) δ 7.60 (4H, d, J=8.5), 6.75 (4H, d, J=8.5), 4.58 (4H, s), 4.25 (4H, q, J=7.1), 1.28 (6H, t, J=7.1). IR (KBr) 1765, 1720, 1580, 1480 cm⁻¹. FDMS (m/e) 488 (M⁺, ¹³⁰Te). Anal. calc'd for C₂₀H₂₂O₆Te: C, 49.43; H, 4.56. Found: C, 49.36; H, 4.55.

(e) Preparation of 4,4'-Di(carboxymethoxy)-1,1'-tellurobisbenzene, Disodium Salt (Te4)

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Diester 4d (39.2 g, 81 mmol) was taken up in methanol (400 mL), and 10% aq NaOH (47 mL, 170 mmol) was added. A very thick slurry resulted. The reaction mixture was heated to reflux for 1 h. The resulting slurry was cooled in an ice bath. The solid was isolated by filtration and washed with ethanol, air dried and ground with a mortar and pestle to give Te4 as a pale cream solid (37.6 g, 98%). ¹H NMR (DMSO-d₆, 5 drops D₂O) δ 7.46 (4H, d, J=8.4), 6.66 (4H, d, J=8.5), 4.06 (4H, s). ¹³C NMR (D₂O) δ 176.6, 158.0, 139.8, 115.9, 104.8, 66.5. IR (KBr) 3540 3430, 1595, 1490, 1415, 1230 cm⁻¹. Anal. calc'd for C₁₆H₁₂Na₂O₆Te·1.5 H₂O: C, 38.37; H, 3.02. Found: C, 38.33; H, 3.05.

Preparation 5. N,N,N',N'-Tetra(carboxymethyl)-4,4'-tellurobisbenzenamine, Tetrasodium Salt (Te5)

(a) Preparation of N-(2-Ethoxy-2-oxoethyl)-N-phenylglycine, Ethyl Ester (5a)

Aniline (9.3 g, 0.1 mol), ethyl bromoacetate (36.7 g, 0.22 mol), and 2,6-lutidine (23.5 g, 0.22 mol) in acetonitrile (200 mL) were refluxed 1 day in a flask equipped with a Newman tube. The reaction was complete by TLC (dichloromethane). It was cooled to rt and diluted with ether (200 mL). The precipitated lutidine hydrochloride was removed by filtration, and the filtrate was evaporated under reduced pressure. The residue was partitioned between ether and water, and the water layer was extracted twice more with ether. The combined extracts were washed with 1N HCL, water, sat. NaHCO₃, and sat. NaCl, dried over MgSO₄, and evaporated under reduced pressure to give crude 5a as a dark liquid. It was taken up in dichloromethane and filtered through silica gel to remove color, giving the product (23.2g, 88%) as a pale oil. ¹H NMR (CDCl₃) δ 7.21 (2H, t, J=7.9), 6.77 (1H, t, J=7.3), 6.61 (2H, d, J=8.4), 4.20 (4H, q, J=7.2), 4.13 (4H, s), 1.26 (6H, t, J=7.1).

(b) Preparation of N,N,N',N'-Tetra(ethoxycarbonylmethyl)-4,4'-tellurobisbenzenamine (5b)

To a 500 mL 3-neck flask, oven-dried, equipped with an addition funnel and put under argon, was added tellurium (IV) chloride (11.9 g, 44 mmol). Dry ether (200 mL) was added by syringe, giving a thin yellow slurry, and the flask was placed in an ice bath. A solution of 5a (23.2 g, 89 mmol) in dry ether (50 mL) was prepared under argon, and the solution was transferred by cannula to the addition funnel. It was added dropwise to the reaction mixture, with vigorous stirring. A thick sludge immediately formed and the stirring bar stopped. The remainder of the addition was

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performed while swirling and manually stirring the reaction mixture. The reaction mixture sat without stirring overnight, then was sonicated until all of the sludge was converted to a clumpy solid.

The reaction mixture was filtered, and the filter cake was washed with ether. The filter cake was taken up in dichloromethane (200 mL), and a solution of sodium metabisulfite (17.0 g, 89 mmol) in water (200 mL) was added with stirring. The resulting two-phase mixture was stirred 30 min. at rt. Solid NaHCO_3 was added to pH 7 and the reaction mixture was filtered through Celite diatomaceous earth. The filtrate was transferred to a separatory funnel, the dichloromethane layer was split off, and the aqueous layer was extracted with additional dichloromethane. The combined extracts were washed with water and sat. NaCl, dried over Na_2SO_4 , and filtered, and the solvent was removed under reduced pressure. The resulting red oil (17.9 g) was taken up in toluene (100 mL), and copper powder (9 g) was added. The reaction mixture was heated to reflux with a Newman tube for 2 h. The resulting gray reaction mixture was cooled to room temperature, filtered through Celite diatomaceous earth, and the solvent was removed under reduced pressure. The crude product (18 g amber oil) was flash chromatographed twice with dichloromethane then 98:2 dichloromethane:methanol to separate the product from recovered starting material, yielding pure 5b as a pale orange viscous oil (7.36 g, 50% of theory). ^1H NMR (CDCl_3) δ 7.52 (4H, d, $J=8.6$), 6.43 (4H, d, $J=8.7$), 4.19 (8H, q, $J=7.1$), 4.08 (8H, s), 1.25 (12H, t, $J=7.1$). ^{13}C NMR (CDCl_3) δ 170.7, 147.6, 139.5, 113.6, 101.3, 61.2, 53.3, 14.2. IR (salt plates) 1730, 1580, 1490, 1180 cm^{-1} . FDMS (m/e) 658 (M^+ , ^{130}Te).

(c) Preparation of N,N,N',N'-Tetra(carboxymethyl)
-4,4'-tellurobisbenzenamine, Tetrasodium Salt (Te5)

Diester 5b (4.35 g, 6.63 mmol) was taken up in methanol (60 mL), and 10% aq NaOH (7.4 mL, 26.5 mmol) was added. The reaction mixture was heated to reflux for 1 h. The resulting slurry was cooled in an ice bath, then filtered. The solid was washed with cold methanol and air dried, to give Te5 as a pale cream solid (3.81 g, 91%). ^1H NMR (D_2O) δ 7.56 (4H, d, $J=8.6$), 6.38 (4H, d, $J=8.6$), 3.83 (8H, s). ^{13}C NMR (D_2O) δ 179.4, 148.7, 139.6, 113.0, 98.3, 55.5. IR (KBr) 3250 (br), 1575, 1405, 1210 cm^{-1} . Anal. calc'd for $\text{C}_{20}\text{H}_{16}\text{Na}_4\text{N}_2\text{O}_8\text{Te}\cdot 3\text{H}_2\text{O}$: C, 35.02; H, 3.23; N, 4.08. Found: C, 34.82; H, 3.03; N, 4.04.

The following examples are illustrative but not exhaustive of the invention:

Example 1

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The coatings described were slit and mounted as slides. Cartridges containing the slides were prepared by placing the cartridges on the benchtop for 16-20 hours with the lights in the laboratory remaining on for this time period. On the day following the benchtop incubation, the slides were tested using a prototype automated dry-film immunoassay analyzer. Eleven uL of a human serum matrix solution containing 10,000 m-IU/mL hCG was applied to each slide. Each slide was then incubated for 5 minutes at 37°C after which 12 uL of a wash solution containing Na₂HPO₄ (10 mM, pH6.8), 4'-hydroxyacetanalide (5mM), hexadecylpyridinium chloride (0.1%), H₂O₂ (8 mM), and diethylenetriamine-pentaacetic acid (DTPA) (10mM) was applied to each slide. The wash fluid both washes away (removes) unbound horseradish peroxidase labeled ligand or receptor from the read area and serves to initiate the HRP-catalyzed dye formation reaction. Following addition of the wash solution each slide underwent a second incubation at 37°C during which time reflectance density readings were taken at intervals of 3 seconds at a wavelength of 670nm. The rate of color formation for slides from each coating was calculated from the reflectance density readings.

Dry-film coatings, for the assay of human chorionic gonadotropin (hCG) in serum samples, of the following formulations were prepared on poly(ethylene terephthalate) support. Terms used in all coating structures are described in Table 1.

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Coating 1

<u>Layer</u>		<u>Material</u>	<u>Dry coverage(g/m2)</u>
5	Bead Spread Layer	TES buffer, pH 7	0.219
		3',5'-dicloro-4'-hydroxyacetanilide	0.22
		Dimedone	0.45
		Adhesive Polymer	2.58
		Polymer Beads	130.0
10		BSA	1.0
		Glycerol	2.0
		Mannitol	1.0
		Label	34e-6
15	Receptor layer	TES buffer, pH 7.0	0.10
		TX-100	0.02
		Polymer Binder	0.8
		Leuco Dye	0.2
		Antibody Beads	0.1
		TETRONIC T908	0.02
20		OLIN 10G	0.01
25	Gel	Gelatin	10
		TES Buffer pH 7.0	4.580
		3'5'-dichloro-4'-hydroxyacetanilide	0.44
		BSVME	0.15

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Coating2

<u>Layer</u>		<u>Material</u>	<u>Dry Coverage (g/m2)</u>
5	Bead Spread Layer	TES Buffer. pH 7.0	0.219
		3',5'-dicloro-	
		4'-hydroxyacetanilde	0.22
		Dimedone	0.45
		Adhesive Polymer	2.58
		Polymer Beads	130
		BSA	1.0
		Glycerol	2
		Mannitol	1.0
		Label	34e-6
10		VO SO ₄	.04
15	Receptor layer	TES buffer, pH 7.0	0.1
		TX-100	0.02
		Polymer Binder	0.8
		Leuco Dye	0.2
		Antibody Beads	0.1
		TETRONIC T908	0.02
		OLIN 10G	0.01
20			
25	Gel	Gelatin	10
		TES Buffer pH 7.0	.58
		3'5'-dichloro-4'-	
		hydroxyacetanilide	0.44
		TX-100	0.02
		BSVME	0.15

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Coating 3

	<u>Layer</u>	<u>Material</u>	<u>Dry coverage (g/m2)</u>
5	Bead Spread Layer	TES Buffer , pH 7.0	0.219
		3',5'-dicloro-4'hydroxyacetanilide	0.22
		Dimedone	0.45
		Adhesive Polymer	2.58
		Polymer Beads	130.0
10		BSA	1.0
		Mannitol	1.0
		Label	34e-6
		Tel	0.04
15	Receptor layer	TES Buffer, pH 7.0	0.1
		TX-100	0.02
		Polymer Binder	0.8
		Leuco Dye	0.2
		Antibody Beads	0.1
20		TETRONIC T908	0.02
		OLIN 10G	0.01
25	Gel	Gelatin	10
		TES Buffer pH 7.0	4.58
		3'5'-dichloro-4'-hydroxyacetanilide	0.44
		TX-100	0.02
		BSVME	0.15

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Coating 4

<u>Layer</u>		<u>Material</u>	<u>Dry coverage (g/m2)</u>
5	Bead Spread Layer	TES Buffer, pH 7.0	0.219
		3',5'-dicloro-	
		4'-hydroxyacetanilde	0.22
		Dimedone	0.45
		Adhesive Polymer	2.58
		Polymer Beads	130
		BSA	1.0
		Glycerol	2.0
		Mannitol	1.0
		Label	34e-6
10		Te3	0.012
15	Receptor layer	TES Buffer, pH 7.0	0.1
		TX-100	0.02
		Polymer Binder	0.8
		Leuco Dye	0.2
		Antibody beads	0.1
		TETRONIC T908	0.02
		OLIN 10G	0.01
20			
25	Gel	Gelatin	10
		TES buffer pH 7.0	4.58
		3'5'-dichloro-4'-	
		hydroxyacetanilide	0.44
		TX-100	0.02
		BSVME	0.15

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Coating 5

	<u>Layer</u>	<u>Material</u>	<u>Dry coverage (g/m²)</u>
5	Bead Spread Layer	TES Buffer, pH 7.0	0.219
		3',5'-dicloro-4'hydroxyacetanilide	0.22
		Dimedone	0.45
		Adhesive Polymer	2.58
		Polymer Beads	130
10		BSA	1.0
		Glycerol	2
		Mannitol	1.0
		Label	34e-6
		VO SO _x	0.04
15		Tel	0.04
20	Receptor layer	TES Buffer, pH 7.0	0.10
		TX-100	0.02
		Polymer Binder	0.8
		Leuco Dye	0.2
		Antibody Beads	0.1
		TETRONIC T908	0.02
		OLIN 10G	0.01
25	Gel	Gelatin	10
		TES Buffer pH 7.0	4.58
		3'5'-dichloro-4'-hydroxyacetanilide	0.44
		TX-100	0.02
30		BSVME	0.15

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Coating 6

	<u>Layer</u>	<u>Material</u>	<u>Dry coverage (g/m2)</u>
5	Bead Spread Layer	TES Buffer pH 7.0	0.219
		3',5'-dicloro-4'-hydroxyacetanilide	0.22
		Dimedone	0.45
		Adhesive Polymer	2.58
		Polymer Beads	130
10		BSA	1.0
		Glycerol	2
		Mannitol	1.0
		Label	34e-6
15		VO SO ₄	0.04
		Te3	0.012
20	Receptor layer	TES Buffer, pH 7.0	0.1
		TX-100	0.02
		Polymer Binder	0.8
		Leuco Dye	0.2
		Antibody Beads	0.1
		TETRONIC T908	0.02
25		OLIN 10G	0.01
	Gel	Gelatin	10
		TES Buffer pH 7.0	4.58
		3'5'-dichloro-4'-hydroxyacetanilide	0.44
30		tx-100	0.02
		BSVME	0.15

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Cartridge Slide Comparison for Coatings 1-6

		<u>Rate of Top</u>	<u>Rate of Non-Top</u>	<u>% Rate Loss</u>
5	<u>Coating</u>	<u>Slides (n=3)</u>	<u>Slides (n=15)</u>	<u>of Top</u>
	1	0.186 Dt/Min	0.231 Dt/Min	-19.48%
	2	0.316 Dt/Min	0.317 Dt/Min	-0.32%
	3	0.305 Dt/Min	0.318 Dt/Min	-4.09%
10	4	0.202 Dt/Min	0.211 Dt/Min	-4.27%
	5	0.292 Dt/Min	0.295 Dt/Min	-1.02%
	6	0.211 Dt/Min	0.212 Dt/Min	-0.47%

The cartridge slide comparison clearly shows that a large percent rate loss occurs for the first slides in the cart when either VOSO_4 or the DAT compounds are not incorporated in the coating. This loss of rate of color formation in the first slide would result in an incorrect analyte concentration prediction for the sample being tested. The inclusion of VOSO_4 (coating 2), the DAT compounds (coatings 3 and 4) or the combination of VOSO_4 and the DAT compounds (coatings 5 and 6) results in marked improvement in the percent rate retained for the top slides.

Example 2

In order to further differentiate the effects of the DAT compounds investigated, slides from each coating were placed face-up directly on the benchtop and exposed to the same conditions as the slides which remained in the cartridges (16-20 hour room temperature incubation with fluorescent lighting remaining on). Following the incubation period, the slides were tested on a prototype automated dry-film immunoassay analyzer using the same materials and protocol as described above. The expected result is a greater loss of rate of color formation due to the increased exposure of the slides to environmental factors (light, air, etc.). As a means of observing the effect of increased environmental exposure, the rate of color formation of the benchtop slides was compared to the rate of color formation of the cartridge slides for each condition (excluding the top slides). The results for the slides treated as described are as follows:

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Cartridge Slide Comparison for Coatings 1-6

	<u>Coating</u>	<u>Rate of Benchtop Slides (n=10)</u>	<u>Rate of Cartridge Slides (n=15)</u>	<u>% Rate Loss of Benchtop Slide</u>
5	1	0.161 Dt/Min	0.231 Dt/Min	-30.3%
	2	0.288 Dt/Min	0.317 Dt/Min	-9.15%
	3	0.293 Dt/Min	0.318 Dt/Min	-7.86%
	4	0.179 Dt/Min	0.211 Dt/Min	-15.17%
10	5	0.291 Dt/Min	0.295 Dt/Min	-1.36%
	6	0.198 Dt/Min	0.212 Dt/Min	-6.6%

15 The benchtop slide test shows again that inclusion of VOSO_4 (coating 2), the DAT compounds (coatings 3 and 4), or the combination of VOSO_4 and the DAT compounds (coatings 5 and 6) in the coating provides protection against rate loss due to environmental exposure. This test also shows that a cooperative function occurs when VOSO_4 and Te1 are incorporated in the same coating (coating 5). This results in lower percent rate loss than when either compound is incorporated alone (coatings 2 and 3).

20 Dats, Te1-Te5 were evaluated in a model system designed to accentuate the loss of HRP activity. In this system, DATs Te1, Te4, and Te5 protected HRP activity either alone or in cooperation with a vanadyl salt as shown in the next examples.

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Examples 3-5

Analytical elements of the following formulation were prepared on a poly(ethylene terephthalate) support:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Bead Spread Layer	TES Buffer, pH 7.0	0.219
	Adhesive Polymer	2.58
	Polymer Beads (20-40 mM)	130.0
Receptor Layer	Polymer Binder I	0.60
	TES Buffer, pH 7.0	0.10
	TX-100	0.02
Gel	Gelatin	10.0
	TES Buffer, pH 7.0	4.58
	TX-100	0.020
	BVSME	0.150

- 5 This analytical element was used to measure HRP stability with the following protocol. Ten microliter samples of solutions of 10 mM sodium phosphate buffer, pH 7.0, containing about 3×10^{-8} M HRP and varying amounts of one of the diaryl telluride compounds of the invention were spotted onto each of separate 1 cm² pieces of the above coated element (4 samples for each compound and at each concentration). These elements were dried and placed in a dark drawer. After 30 minutes and again after
- 10 24 hours, i.e., the next day, the elements were removed from the drawer and the HRP was extracted by immersing each element in 1 mL of a solution of 10 mM sodium phosphate buffer, 0.15 M sodium chloride, 0.1% bovine serum albumin, pH 7.0, in a test tube to extract the HRP. After vortexing the test tube for 1 minute (which removed the analytical element components from the poly(ethylene terephthalate) support), the resulting suspension was centrifuged and the solution removed. The amount
- 15 of active HRP in this solution was determined by adding a 100 μ L aliquot to a spectrophotometer cuvette and a reagent solution to provide one mL of a final solution of 50 mM potassium phosphate buffer, pH 7.0, 5 mM 4'-hydroxyacetanilide, 0.625% Triton X-100 surfactant, 0.005%

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4,5-bis(4-dimethylaminophenyl)-2-(4-hydroxy-3-methoxyphenyl)imidazole leuco dye, and 0.85 mM hydrogen peroxide. The assay temperature was 30°C. A blue color was formed whose rate was determined spectrophotometrically at 655 nm. The rate of color formation was directly proportional to the amount of active enzyme in the extract.

5 The data were expressed as activity ratios, calculated by dividing the activity extracted at 24 hours by the activity extracted at 30 minutes for each sample. A ratio of 1 indicated enzyme was fully protected by the additive over a 24 hour time period. Some run to run data variation was observed, which was probably caused by the day to day differences in environment humidity and temperature. This could affect how fast the HRP dried down on the coating and thus affect the enzyme's apparent stability.
10 Therefore, data can be compared within the Examples 3-5, but not between any two of the examples.

Example 3

15 The effectiveness of compounds Te1, Te2, Te3, and Te4 at 1 mM is compared either alone or in the presence of 1 mM VOSO₄ (Figure 1). Compounds Te1 and Te4 protected HRP activity in the model dry-down format relative to no additive. Te2 and Te3 were not effective stabilizers in this format. When both VOSO₄ and either Te1 or Te4 were included, more HRP activity was retained than in the presence of either one alone.

Example 4

20 The effect of concentration on the ability of Te1 and Te4 to stabilize HRP is shown (Figure 2). The following concentrations were used in the HRP solution: 1, 0.1 or 0.01 mM Te1, and 10, 1, or 0.1 mM Te4. In addition, HRP solutions containing buffer only, or 1 mM VOSO₄ were spotted. Activity loss was reagent concentration dependent for both Te1 and Te4. In the presence of 10 mM Te4, HRP activity was fully protected during the 24 hour incubation.

25

Example 5

30 This example shows the effect of Te4 and Te5 on the stability of anti-CRP-HRP enzyme conjugate (2.85×10^{-8} M). Also present in the spotting solution were buffer only; 1, 2, 4, or 8 mM Te4; or 1 or 8 mM Te5. Both Te4 and Te5 protected the activity of conjugated HRP, and the protection was concentration dependent (Figure 3).

Example 6

Coatings 7-10 were prepared as indicated in the following coating structures, 7-10, mounted as slides, conditioned at 70°F/33% RH for two days and frozen.

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Coating 7

A dry-film coating for the assay of C-reactive protein (CRP) in serum samples of the following formulation was prepared on a poly(ethylene terephthalate) support.

	<u>Layer</u>	<u>Material</u>	<u>Dry coverage(g/m²)</u>
5	Bead Spread Layer	TES buffer	0.219
		Dimedone	0.450
		CaCl ₂	1.0
		3',5'-dicloro-	
10		4'-hydroxyacetanilide	0.22
		glycerol	2
		bovine serum albumin	0.5
		magenta dye	0.0538
		adhesive polymer	2.538
15		polymer beads	130
	Receptor layer	TES buffer	0.1
		TX100	0.02
		polymer binder	0.8
20		leuco dye	0.2
		PC beads	0.3
		anti-CRP antibody	0.05
		anti-CRP antibody-	
		HRP conjugate	0.0005
25		Tetronic T908	0.02
		Olin 10G	0.01
	Gel	Gelatin	10
		TES buffer pH 7.0	4.580
30		3'5'-dichloro-4'-	
		hydroxyacetanilide	0.44
		BSVME	0.15

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Coating 8

A dry-film coating for the assay of C-reactive protein (CRP) in serum samples of the following formulation was prepared on a poly(ethylene terephthalate) support.

	<u>Layer</u>	<u>Material</u>	<u>Dry coverage (g/m²)</u>
5	Bead Spread Layer	TES buffer	0.219
		Dimedone	0.450
		CaCl ₂	1.0
		3',5'-dicloro-	
10		4'-hydroxyacetanilde	0.22
		glycerol	2
		bovine serum albumin	0.5
		magenta dye	0.0538
		adhesive polymer	2.538
15		polymer beads	130
		Tel	0.08
	Receptor layer	TES buffer	0.1
		TX100	0.02
20		polymer binder	0.8
		leuco dye	0.2
		PC beads	0.3
		anti-CRP antibody	0.05
		anti-CRP antibody-	
25		HRP conjugate	0.0005
		Tetronic T908	0.02
		Olin 10G	0.01
	Gel	Gelatin	10
30		TES buffer pH 7.0	4.580
		3'5'-dichloro-4'-	
		hydroxyacetanilide	0.44
		BSVME	0.15

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Coating 9

A dry-film coating for the assay of C-reactive protein (CRP) in serum samples of the following formulation was prepared on a poly(ethylene terephthalate) support.

5			
	<u>Layer</u>	<u>Material</u>	<u>Dry coverage (g/m²)</u>
	Bead Spread Layer	TES buffer	0.219
		Dimedone	0.450
		CaCl ₂	1.0
10		3',5'-dicloro-	
		4'hydroxyacetanilide	0.22
		glycerol	2
		bovine serum albumin	0.5
		magenta dye	0.0538
15		adhesive polymer	2.538
		polymer beads	130
		Te2	0.075
	Receptor layer	TES buffer	0.1
20		TX100	0.02
		polymer binder	0.8
		leuco dye	0.2
		PC beads	0.3
		anti-CRP antibody	0.05
25		anti-CRP antibody-	
		HRP conjugate	0.0005
	Gel	Tetronic T908	0.02
		Olin 10G	0.01
30		Gelatin	10
		TES buffer pH 7.0	4.580
		3'5'-dichloro-4'-	
		hydroxyacetanilide	0.44
		BSVME	0.15

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Coating 10

A dry-film coating for the assay of C-reactive protein (CRP) in serum samples of the following formulation was prepared on a poly(ethylene terephthalate) support.

5			
	<u>Layer</u>	<u>Material</u>	<u>Dry coverage (g/m²)</u>
	Bead Spread Layer	TES buffer	0.219
		Dimedone	0.450
10		CaCl ₂	1.0
		3',5'-dicloro-	
		4'-hydroxyacetanilide	0.22
		glycerol	2
		bovine serum albumine	0.5
15		magenta dye	0.0538
		adhesive polymer	2.538
		polymer beads	130
		VOSO ₄	0.04
20	Receptor layer	TES buffer	0.1
		TX100	0.02
		polymer binder	0.8
		leuco dye	0.2
		PC beads	0.3
25		anti-CRP antibody	0.05
		anti-CRP antibody-	
		HRP conjugate	0.0005
		Tetronic T908	0.02
		Olin 10G	0.01
30	Gel	Gelatin	10
		TES buffer pH 7.0	4.580
		3'5'-dichloro-4'-	
		hydroxyacetanilide	0.44
35		BSVME	0.15

Cartridges were thawed and loaded onto an E250 analyzer.

Eleven uL of a human serum sample containing about 20mg/L CRP was applied to each slide.

Each slide was then incubated for 5 minutes at 37°C after which 12uL of a wash solution containing Na₂HPO₄ (10mM, pH 6.8), 4'-hydroxyacetanilide (5mM), hexadecylpyridinium chloride (0.1%), H₂O₂ (8mM), and DTPA (10uM) was applied to each slide to wash (remove) the unbound antibody-HRP conjugate from the read area and initiate the HRP-catalyzed dye formation reaction. After 2.5 min. incubation at 37°C, the reflectance density at 670 nm was measured and converted to a concentration of CRP via a calibration curve.

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The predicted concentration of the top slide was compared with the average predicted concentration of the next six slides. The results of several experiments of this type are shown below:

Bias = prediction top slide - prediction average of slides 2-7;

5	<u>Coating</u>	<u>Agent</u>	<u>Experiment 1</u> <u>1 cart</u>	<u>Experiment 2;</u> <u>Average of</u> <u>2 carts</u>	<u>Experiment 3</u> <u>1 cart</u>
10	7	none	-29.9	-27.5	-34.3
	8	Te1	-6.0	-6.1	-7.2
	9	Te2	-23.2	-19.3	-21.8
	10	VOSO ₄	-13.3	-16.2	-20.6

These results show that in the absence of any protective agent, the top slide has a predicted value substantially less than the subsequent slides. Addition of either Te1, Te2 or VOSO₄ decreases the bias between the top slide and subsequent slides.

The greatest improvement is provided by Te1.

Example 7

Coatings 7-10 were slit, mounted as slides, conditioned at 70°F/33% RH for 2 days, and frozen. Cartridges were thawed. The top slide of each cartridge was ejected and the cartridges were incubated at 70°F/33% RH for 3 days.

The slides were tested with human serum containing about 20mg/L CRP as described in Example 6.

The prediction of the top slide receiving 3 days exposure at 70°F/33% RH was compared with the average prediction of slides 2-7 and the results are shown below:

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Bias = prediction top slide - prediction average of slides 2-7;

	<u>Coating</u>	<u>Agent</u>	<u>average of 2 carts</u>
5	7	none	-26.4
	8	Te1	-4.5
	9	Te2	-8.2
	10	VOSO ₄	-5.6

10 These results show that a newly exposed top slide changes over a 3 day period resulting in a prediction that is substantially lower than subsequent slides in the absence of a protective agent. Diaryl tellurides Te1 and Te2 and VOSO₄ protect the slide and result in a smaller bias compared with subsequent slides in the cartridge, again Te1 showing the greatest improvement.

Example 8

15 Coatings 7-10 were slit, mounted as slides, conditioned at 70°F/33% RH for 2 days and frozen. Cartridges were thawed and incubated at 70°F/33% RH for 7 days.

20 The incubated cartridges and freshly thawed cartridges were analyzed with 3 human serum samples containing CRP in the range of 10-30mg/L as described in Example 6. The predictions of the incubated slides were compared with the predictions of the freshly thawed slides. The average bias of incubated vs freshly thawed slides for the 3 fluids was determined and is shown below.

	<u>Coating</u>	<u>Agent</u>	<u>Average Bias</u>
25	7	none	-7.8
	8	Te1	0.8
	9	Te2	-2.62
	10	VOSO ₄	-0.26

30 These results show that, in the absence of a protective agent, slides exposed to 70°F/33% RH for 7 days develop a bias to freshly thawed slides. In the presence of either Te1 or Te2 or VOSO₄ the bias that results from this exposure is substantially reduced.

The following examples demonstrate results that were superior and unexpected when DAT was coated in a gravure layer compared to when DAT was not used, or coated in a different layer.

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Example 9

Te5 was evaluated in the CRBM assay in the following manner:

5

Coating 1, No Te5

A dry-film coating for the assay of carbamazepine (CRBM) in serum samples of the following formulation was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS	.0045
	BSA	.000215
	A-100	.00108
	3',5'-Dichloro-4'-hydroxyacetanilide	.00995
	Magenta Dye	.05380
	TX-100	.0043
	Carboxymethyl-apo-HRP	.010
	CRBM-HRP Label	0.000012
Bead Spread Layer	TES Buffer, pH 7.0	0.219
	3',5'-Dichloro-4'-hydroxyacetanilide	0.220
	Dimedone	0.450
	Adhesive Polymer	2.583
	Polymer Beads	130.00
	BSA	1.00
	Glycerol	2.00
	Mannitol	1.00
Receptor Layer	Vanadyl sulfate dihydrate	0.04
	TES Buffer, pH 7.0	0.10
	TX-100	0.02
	Polymer Binder	0.50
	Leuco Dye	0.20
	Tetronic T908	0.02
	Olin 10G	0.01
	Antibody Beads	0.20
Gel	Gelatin	10.00
	TES Buffer, pH 7.0	4.580
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.02
	BVSME	0.15
Support	Poly (ethylene terephthalate)	

Example 10**Coating 2**

5

A dry-film coating for the assay of carbamazepine (CRBM) comprising Te5 coated in the spread layer at a final rewet level of 1 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS	.0045
	BSA	.000215
	A-100	.00108
	3',5'-Dichloro-4'-hydroxyacetanilide	.00995
	Magenta Dye	.05380
	TX-100	.0043
	Carboxymethyl-apo-HRP	.010
	CRBM-HRP Label	0.000012
Bead Spread Layer	TES Buffer, pH 7.0	0.219
	3',5'-Dichloro-4'-hydroxyacetanilide	0.220
	Dimedone	0.450
	Adhesive Polymer	2.583
	Polymer Beads	130.00
	BSA	1.00
	Glycerol	2.00
	Mannitol	1.00
	Vanadyl sulfate dihydrate	0.04
Receptor Layer	Te5	0.0632
	TES Buffer, pH 7.0	0.10
	TX-100	0.02
	Polymer Binder	0.50
	Leuco Dye	0.20
	Tetronic T908	0.02
	Olin 10G	0.01
Gel	Antibody Beads	0.20
	Gelatin	10.00
	TES Buffer, pH 7.0	4.580
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.02
Support	BVSME	0.15
	Poly (ethylene terephthalate)	

Example 11**Coating 3, 1mM Te5 (gravure layer)**

5

A dry-film coating for the assay of carbamazepine (CRBM) comprising Te5 coated in the gravure layer at a final rewet level of 1 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS	.0045
	BSA	.000215
	A-100	.00108
	3',5'-Dichloro-4'-hydroxyacetanilide	.00995
	Magenta Dye	.05380
	TX-100	.0043
	Carboxymethyl-apo-HRP	.010
	CRBM-HRP Label	0.000012
	Te5	0.0632
Bead Spread Layer	TES Buffer, pH 7.0	0.219
	3',5'-Dichloro-4'-hydroxyacetanilide	0.220
	Dimedone	0.450
	Adhesive Polymer	2.583
	Polymer Beads	130.00
	BSA	1.00
	Glycerol	2.00
	Mannitol	1.00
	Vanadyl sulfate dihydrate	0.04
Receptor Layer	TES Buffer, pH 7.0	0.10
	TX-100	0.02
	Polymer Binder	0.50
	Leuco Dye	0.20
	Tetronic T908	0.02
	Olin 10G	0.01
	Antibody Beads	0.20
Gel	Gelatin	10.00
	TES Buffer, pH 7.0	4.580
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.02
	BVSME	0.15
Support	Poly (ethylene terephthalate)	

Example 12**Comparative Example 1**

Coatings 1 - 3 were slit and mounted as slides. Cartridges containing the slides were tested for rate response 2 weeks after coating, then stored at in a freezer at -18°C for 1 year. After 1 year of freezer storage, the cartridges were tested for rate response and first slide bias. The slides were tested on a VITROS E250 ANALYZER. Eleven μ l of a human serum matrix solution containing 8.8 μ g/mL CRBM was applied to the first (top) slide from 3 cartridges, and also to slides 2-7 from those same cartridges. Each slide was incubated for 5 minutes at 37°C after which 12 μ l of a wash solution containing Na_2HPO_4 (10 mM, pH 6.8), 4'-hydroxyacetanilide (5 mM), hexadecylpyridinium chloride (0.1%), H_2O_2 (8 mM), and DPTA (10 μ M) was applied to each slide. The wash fluid washes away (or removes) unbound drug-horseradish peroxidase label and serves to initiate the HRP-catalyzed dye formation reaction. Following addition of the wash solution, each slide was reincubated at 37°C during which time reflectance density readings were taken at intervals of 3 seconds for 2.5 minutes at a wavelength of 670 nm. The reflectance measurements were converted to transmittance, Dt, using the Clapper-William transform, and the rate of color formation was calculated. The results are as follows:

<u>Coating</u>	<u>Cartridge</u>	<u>Rate of Top Slide (n=1)</u>	<u>Rate of Non-Top Slides (n=6)</u>	<u>% Rate Loss of Top Slide</u>
1	1	0.05199 Dt/min	0.05682 Dt/min	8.50
1	2	0.05243 Dt/min	0.05738 Dt/min	8.63
1	3	0.05183 Dt/min	0.05687 Dt/min	8.86
2	1	0.05500 Dt/min	0.05975 Dt/min	7.95
2	2	0.05406 Dt/min	0.05973 Dt/min	9.49
2	3	0.05470 Dt/min	0.05924 Dt/min	7.66
3	1	0.05968 Dt/min	0.05927 Dt/min	-0.69
3	2	0.05918 Dt/min	0.05894 Dt/min	-0.41
3	3	0.06020 Dt/min	0.05948 Dt/min	-1.21

The coating comparison clearly shows that a large percent rate loss occurred for first slides in the cartridges when either Te5 was absent from the coating or was placed in the bead spreading layer at a rewet level of 1 mM. This loss of rate of color formation in the first slide would result in an incorrect analyte concentration prediction for the sample being tested. The inclusion of Te5 in the

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gravure layer at a rewet level of 1 mM results in a marked improvement in the percent rate retained for the top slides after 1 year of freezer storage.

Example 13

Te5 was evaluated in the gravure layer of a phenobarbital dry-film element as follows:

5

Coating 4, No Te5

A dry-film coating for the assay of PHBR was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.01
	Magenta dye	0.16
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	PHBR-HRP label	0.00001
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	0.58
	Dimedone	0.50
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Leuco Dye	0.20
	Tetronic T908	0.02
	Olin 10G	0.25
	Polymer beads	130
	Adhesive polymer	2.6
Receptor	TES Buffer	0.10
	Sodium hydroxide, 1N	0.12
	TX-100	0.020
	Polymer Binder	0.80
	Antibody Beads	0.20
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	1.18
Support	Crosslinking agent	0.15
	Poly(ethylene terephthalate)	

Example 14**Coating 5 (0.25 mM Te5 in gravure)**

- 5 A dry-film coating for the assay of PHBR comprising Te5 coated in the gravure layer at a final rewet level of 0.25 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.16
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	PHBR-HRP label	0.000010
	Te5	0.016
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	0.58
	Dimedone	0.50
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Leuco Dye	0.20
	Tetronic T908	0.02
	Olin 10G	0.25
	Polymer beads	130
	Adhesive polymer	2.6
Receptor	TES Buffer	0.10
	Sodium hydroxide, 1N	0.12
	TX-100	0.020
	Polymer Binder	0.80
	Antibody Beads	0.20
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	1.18
	Crosslinking agent	0.15
Support	Poly(ethylene terephthalate)	

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Example 15**Coating 6 (0.5 mM Te5 in gravure)**

5

A dry-film coating for the assay of PHBR comprising Te5 coated in the gravure layer at a final rewet level of 0.5 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.16
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	PHBR-HRP label	0.000010
	Te5	0.032
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	0.58
	Dimedone	0.50
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Leuco Dye	0.20
	Tetronic T908	0.02
	Olin 10G	0.25
	Polymer beads	130
Receptor	Adhesive polymer	2.6
	TES Buffer	0.10
	Sodium hydroxide, 1N	0.12
	TX-100	0.020
	Polymer Binder	0.80
Gel	Antibody Beads	0.20
	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	1.18
Support	Crosslinking agent	0.15
	Poly(ethylene terephthalate)	

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Example 16**Comparative Example 2**

5 Coatings 4-6 were slit and mounted as slides. Cartridges containing the slides were stored in
a freezer at -18°C. After being frozen for 6 months and 9 months the cartridges were tested for rate
response and first slide bias. The slides were tested on a Vitros 250 analyzer. Eleven µl of a human
serum matrix solution containing approximately 27 µg/mL phenobarbital was applied to the first (top)
slide from 3 cartridges, and also to slides 2-7 from those same cartridges. Each slide was incubated
10 for 5 minutes at 37°C after which 12 µl of a wash solution containing Na₂HPO₄ (10 mM, pH 6.8), 4'-
hydroxyacetanilide (5 mM), hexadecylpyridinium chloride (0.1%), H₂O₂ (8 mM), and DPTA (10
µM) was applied to each slide. The wash fluid washes away (or removes) unbound drug-horseradish
peroxidase label and serves to initiate the HRP-catalyzed dye formation reaction. Following addition
of the wash solution, each slide was reincubated at 37°C during which time reflectance density
15 readings weretaken at intervals of 3 seconds for 2.5 minutes at a wavelength of 670 nm. The
reflectance measurements were converted to transmittance, Dt, using the Clapper-Williams
transform, and the rate of color formation was calculated. This was converted to a concentration of
phenobarbital via a calibration curve.

20 The results are as follows: Bias = prediction, top slide - prediction, average of slides 2-7.

<u>Coating</u>	<u>Agent</u>	<u>Bias, 6 mos.</u>	<u>Bias, 9 mos.</u>
4	None	3.43	3.54
5	0.25 Te5	2.96	3.55
6	0.50 Te5	1.02	1.15

Example 17

Te5 was evaluated in the gravure layer of a phenytoin dry-film element as follows:

- 5 **Coating 7, No Te5:** A thin film coating for the assay of PHYT was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.054
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	PHYT-HRP label	0.000013
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	1.90 g / kg batch
	Dimedone	0.45
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Olin 10G	0.24
	Polymer beads	130
Receptor	Adhesive polymer	2.6
	TES Buffer	0.10
	Sulfuric acid, 2N	1.15 g / kg batch
	TX-100	0.020
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.010
	Dimedone	0.050
	Polymer Binder	0.50
	Antibody Beads	0.15
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	13.8 g / kg batch
Support	Crosslinking agent	0.15
	Poly(ethylene terephthalate)	

Example 18**Coating 8 (0.25 mM Te5 in gravure)**

5 A dry-film coating for the assay of PHYT comprising Te5 coated in the gravure layer at a final rewet level of 0.25 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.054
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	PHYT-HRP label	0.000013
	Te5	0.016
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	1.90 g / kg batch
	Dimedone	0.45
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Olin 10G	0.24
	Polymer beads	130
	Adhesive polymer	2.6
Receptor	TES Buffer	0.10
	Sulfuric acid, 2N	1.15 g / kg batch
	TX-100	0.020
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.010
	Dimedone	0.050
	Polymer Binder	0.50
	Antibody Beads	0.15
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	13.8 g / kg batch
	Crosslinking agent	0.15
Support	Poly(ethylene terephthalate)	

Example 19**Coating 9 (0.5 mM Te5 in gravure)**

5 A dry-film coating for the assay of PHYT comprising Te5 coated in the gravure layer at a final rewet level of 0.5 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.054
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	PHYT-HRP label	0.000013
	Te5	0.032
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	1.90 g / kg batch
	Dimedone	0.45
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Olin 10G	0.24
	Polymer beads	130
	Adhesive polymer	2.6
Receptor	TES Buffer	0.10
	Sulfuric acid, 2N	1.15 g / kg batch
	TX-100	0.020
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.010
	Dimedone	0.050
	Polymer Binder	0.50
	Antibody Beads	0.15
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	13.8 g / kg batch
	Crosslinking agent	0.15
Support	Poly(ethylene terephthalate)	

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Example 20**Comparative Example 3**

5 Coatings 7-9 were slit and mounted as slides. Cartridges containing the slides were stored in
a freezer at -18°C. After being frozen for 6 months and 9 months the cartridges were tested for rate
response and first slide bias. The slides were tested on a Vitros 250 analyzer. Eleven µl of a human
serum matrix solution containing approximately 12.5 µg/mL phenytoin was applied to the first (top)
slide from 3 cartridges, and also to slides 2-7 from those same cartridges. Each slide was incubated
10 for 5 minutes at 37°C after which 12 µl of a wash solution containing Na₂HPO₄ (10 mM, pH 6.8), 4'-
hydroxyacetanilide (5 mM), hexadecylpyridinium chloride (0.1%), H₂O₂ (8 mM), and DPTA (10
µM) was applied to each slide. The wash fluid washes away (removes) unbound drug-horseradish
peroxidase label and serves to initiate the HRP-catalyzed dye formation reaction. Following addition
of the wash solution, each slide was reincubated at 37°C during which time reflectance density
15 readings weretaken at intervals of 3 seconds for 2.5 minutes at a wavelength of 670 nm.

The reflectance measurements were converted to transmittance, Dt, using the Clapper-
Williams transform, and the rate of color formation was calculated. This was converted to a
concentration of phenytoin via a calibration curve.

The results areas follows:

Bias = prediction, top slide - prediction, average of slides 2-7.

<u>Coating</u>	<u>Agent</u>	<u>Bias, 6 mos.</u>	<u>Bias, 9 mos.</u>
7	None	0.89	1.10
8	0.25 Te5	0.10	-0.17
9	0.50 Te5	0.19	-0.41

Example 21

Te5 was evaluated in the gravure layer of digoxin as follows:

5

Coating 10 (No Te5)

A dry-film,coating for the assay of DGXN was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.054
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	Digoxin-HRP label	0.0000060
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	1.90 g / kg batch
	Dimedone	0.50
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.25
	Polymer beads	130
	Adhesive polymer	2.6
	Antibody Beads	0.055
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	13.8 g / kg batch
Support	Crosslinking agent	0.15
	Poly(ethylene terephthalate)	

10

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Example 22**Coating 11 (0.25 mM Te5 in gravure)**

- 5 A dry-film coating for the assay of DGXN comprising Te5 coated in the gravure layer at a final rewet level of 0.25 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.054
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	Digoxin-HRP label	0.0000060
	Te5	0.016
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	1.90 g / kg batch
	Dimedone	0.50
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.25
	Polymer beads	130
	Adhesive polymer	2.6
	Antibody Beads	0.055
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	13.8 g / kg batch
	Crosslinking agent	0.15
Support	Poly(ethylene terephthalate)	

10

Example 23**Coating 12 (0.5 mM Te5 in gravure)**

5 A dry-film coating for the assay of DGXN comprising Te5 coated in the gravure layer at a final rewet level of 0.5 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.054
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.00430
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	Digoxin-HRP label	0.0000060
	Te5	0.032
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	1.90 g / kg batch
	Dimedone	0.50
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.25
	Polymer beads	130
	Adhesive polymer	2.6
	Antibody Beads	0.055
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	13.8 g / kg batch
	Crosslinking agent	0.15
Support	Poly(ethylene terephthalate)	

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Example 24**Comparative Example 4**

5 Coatings 10-12 were slit and mounted as slides. Cartridges containing the slides were stored
in a freezer at -18°C. After being frozen for 6 months and 9 months the cartridges were tested for rate
response and first slide bias. The slides were tested on a Vitros 250 analyzer. Eleven µl of a human
serum matrix solution containing approximately 2.0 µg/mL digoxin was applied to the first (top) slide
from 3 cartridges, and also to slides 2-7 from those same cartridges. Each slide was incubated for 5
10 minutes at 37°C after which 12 µl of a wash solution containing Na₂HPO₄ (10 mM, pH 6.8), 4'-
hydroxyacetanilide (5 mM), hexadecylpyridinium chloride (0.1%), H₂O₂ (8 mM), and DPTA (10
µM) was applied to each slide. The wash fluid washes away (removes) unbound drug-horseradish
peroxidase label and serves to initiate the HRP-catalyzed dye formation reaction. Following addition
of the wash solution, each slide was reincubated at 37°C during which time reflectance density
15 readings weretaken at intervals of 3 seconds for 2.5 minutes at a wavelength of 670 nm. The
reflectance measurements were converted to transmittance, Dt, using the Clapper-Williams
transform, and the rate of color formation was calculated. This was converted to a concentration of
digoxin via a calibration curve. The results areas follows:

20 Bias = prediction, top slide - prediction, average of slides 2-7.

<u>Coating</u>	<u>Agent</u>	<u>Bias, 6 mos.</u>	<u>Bias, 9 mos.</u>
10	None	0.16	0.38
11	0.25 Te5	0.063	0.21
12	0.50 Te5	0.003	0.03

Example 25

Te5 was evaluated in the gravure layer of a carbamazepine dry-film element as follows:

- 5 **Coating 13 (No Te5):** A dry-filmcoating for the assay of CRBM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.16
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.0043
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	CRBM-HRP label	0.000012
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	0.57
	Dimedone	0.45
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Olin 10G	0.24
	Polymer beads	130
	Adhesive polymer	2.6
Receptor	TES Buffer	0.10
	Sodium hydroxide, 1 N	0.40 g / kg batch
	TX-100	0.020
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.010
	Dimedone	0.050
	Polymer Binder	0.50
	Antibody Beads	0.20
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	1.18
	Crosslinking agent	0.15
Support	Poly(ethylene terephthalate)	

Example 26**Coating 14 (0.25 mM Te5 in gravure)**

5

A dry-film coating for the assay of CRBM comprising Te5 coated in the gravure layer at a final rewet level of 0.25 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.16
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.0043
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	CRBM-HRP label	0.000012
	Te5	0.016
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	0.57
	Dimedone	0.45
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Olin 10G	0.24
	Polymer beads	130
	Adhesive polymer	2.6
	TES Buffer	0.10
Receptor	Sodium hydroxide, 1 N	0.40 g / kg batch
	TX-100	0.020
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.010
	Dimedone	0.050
	Polymer Binder	0.50
	Antibody Beads	0.20
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	1.18
	Crosslinking agent	0.15
Support	Poly(ethylene terephthalate)	

Example 27**Coating 15 (0.5 mM Te5 in gravure)**

5

A dry-film coating for the assay of CRBM comprising Te5 coated in the gravure layer at a final rewet level of 0.5 mM was prepared having the composition and layer structure indicated below:

<u>Layer</u>	<u>Material</u>	<u>Dry Coverage (g/m²)</u>
Gravure Layer	MOPS buffer	0.0045
	3',5'-Dichloro-4'-hydroxyacetanilide	0.010
	Magenta dye	0.16
	Sodium hydroxide, 1N	(17 g / kg batch)
	BSA	0.00022
	A-100	0.00108
	TX-100	0.0043
	Sulfuric acid, 2N	5.3 g / kg batch
	Carboxymethyl-apo-HRP	0.010
	CRBM-HRP label	0.000012
	Te5	0.032
Bead Spread Layer	TES Buffer	0.22
	Sodium hydroxide, 10N	0.57
	Dimedone	0.45
	3',5'-Dichloro-4'-hydroxyacetanilide	0.22
	Vanadyl sulfate dihydrate	0.040
	Mannitol	1.0
	Glycerol	2.0
	Bovine serum albumin	1.0
	Olin 10G	0.24
	Polymer beads	130
	Adhesive polymer	2.6
Receptor	TES Buffer	0.10
	Sodium hydroxide, 1 N	0.40 g / kg batch
	TX-100	0.020
	Leuco Dye	0.20
	Tetronic T908	0.020
	Olin 10G	0.010
	Dimedone	0.050
	Polymer Binder	0.50
	Antibody Beads	0.20
Gel	Gelatin	10
	TES Buffer	4.6
	3',5'-Dichloro-4'-hydroxyacetanilide	0.44
	TX-100	0.020
	Sodium hydroxide, 10N	1.18
	Crosslinking agent	0.15
Support	Poly(ethylene terephthalate)	

Example 28**Comparative Example 5**

5 Coatings 13-15 were slit and mounted as slides. Cartridges containing the slides were stored in a freezer at -18°C. After being frozen for 6 months and 9 months the cartridges were tested for rate response and first slide bias. The slides were tested on a Vitros 250 analyzer. Eleven µl of a human serum matrix solution containing approximately 9.3 µg/mL carbamazepine was applied to the first (top) slide from 3 cartridges, and also to slides 2-7 from those same cartridges. Each slide was
10 incubated for 5 minutes at 37°C after which 12 µl of a wash solution containing Na₂HPO₄ (10 mM, pH 6.8), 4'-hydroxyacetanilide (5 mM), hexadecylpyridinium chloride (0.1%), H₂O₂ (8 mM), and DPTA (10 µM) was applied to each slide. The wash fluid washes away (removes) unbound drug-horseradish peroxidase label and serves to initiate the HRP-catalyzed dye formation reaction. Following addition of the wash solution, each slide was reincubated at 37°C during which time
15 reflectance density readings were taken at intervals of 3 seconds for 2.5 minutes at a wavelength of 670 nm. The reflectance measurements were converted to transmittance, Dt, using the Clapper-Williams transform, and the rate of color formation was calculated. This was converted to a concentration of carbamazepine via a calibration curve.

20 The results are as follows:

Bias = prediction, top slide - prediction, average of slides 2-7.

<u>Coating</u>	<u>Agent</u>	<u>Bias, 6 mos.</u>	<u>Bias, 9 mos.</u>
13	None	0.35	0.69
14	0.25 Te5	0.53	0.56
15	0.50 Te5	-0.013	0.12

25 Comparative examples 2 - 5 show that the presence of Te5 in the gravure layer results in significant reduction of first-slide bias that is significantly closer to zero than in the absence of the Te5. The level of Te5 required varies, being in some cases 0.25 (PHYT) and in other cases 0.5 (PHBR, DGXN, CRBM). But in all cases the first slide bias can be reduced to near zero with Te5 in the gravure layer.

30 This invention results in significant reduction in first slide bias with lower levels of Te5, when the stabilizer is placed in a gravure layer over the spreading layer as opposed to being coated within the spreading layer in competitive immunorate assays. This is shown to be a general phenomenon by the examples provided herein. Incorporation of even low-levels of diaryl telluride

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5

into the gravure layers of four immunorate assays, compared with diaryl telluride coated directly in the spreading layer, resulted in significant reduction of first slide bias. Since diaryl tellurides can impact the assay negatively in ways which are dose-dependent (by, for example, bleaching the registration dye), the use of lower concentrations of diaryl tellurides in the gravure layer, as opposed to the higher levels required in the spreading layer, confers an important advantage.

The present invention results in improvement in HRP stability either alone or in conjunction with other stabilizers such as vanadyl compounds. A preferred embodiment of the present invention comprises both Te5 and a vanadyl salt.

TABLE 1.

5	Adhesive Polymer	Poly(methyl acrylate-co-sodium 2-acrylamido-2-methylpropanesulfonate-co-2-acetoacetoxyethyl methacrylate).
	Antibody Beads	Polymer particles of poly(styrene-co-3-(p-vinylbenzylthio)propionic acid having an antibody to a ligand bound thereto .
10	PC Beads	Polymer particles of poly(styrene-co-3-(p-vinylbenzylthio) propionic acid having phosphoryl cholin bound thereto.
	BSA	Bovine serum albumin.
15	BVSME	Bis(vinylsulfonylmethyl) ether.
	DTPA	Diethylenetriaminepentaacetic acid.
	Label	A conjugate of a ligand or a receptor for a ligand and horseradish peroxidase.
20	Leuco Dye	4,5-Bis(4-dimethylaminophenyl)-2-(3,5-dimethoxy-4-hydroxyphenyl)imidazole.
	Magenta Dye	4,5-Dihydroxy-3-(6,8-disulfo-2-naphthylazo)-2,7-naphthalenedisulfonic acid, sodium salt
25	MOPS	3-(N-morpholino)propanesulfonic acid buffer.
	OLIN 10G	An isononylphenoxypolyglycidol surfactant averaging about 10 glycidol units per molecule (sold by Olin Chemical Co).
30	Polymer Beads	Poly(vinyltoluene-co-methacrylic acid) particles having an average diameter of 20-40 μm .
35	Polymer Binder	Poly(N-isopropylacrylamide-co-2-acrylamido-2-methylpropane sulfonic acid sodium salt-co-N,N'-methylenebisacrylamide).
	Polymer Binder I	Poly(N-isopropylacrylamide-co-2-hydroxyethyl methacrylate-co-N,N'-methylenebisacrylamide)
40	TES	N-[tris(hydroxymethyl)methyl]-2-aminoethanesulfonic acid buffer.
	TX-100	Triton X-100 surfactant - an octylphenoxy polyethoxy ethanol surfactant (sold by Union Carbide).
45	TETRONIC T908	A nonionic surfactant which is a block copolymer of ethylene oxide and propylene oxide (sold by BASF Corp).

The invention has been described in detail with particular reference to preferred embodiments thereof, but it will be understood that variations and modifications can be effected within the spirit and

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scope of the invention. In particular, one skilled in the art would understand that the use of one or more diaryl tellurides, or the use of a particular diaryl telluride, or combination of specific diaryl tellurides or one or more diaryl tellurides in combination with one or more vanadadyl compounds could be selected in order to optimize Applicants' invention.

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We claim:

1. A dry immunoassay analytical element for assaying a ligand, comprising a support bearing:

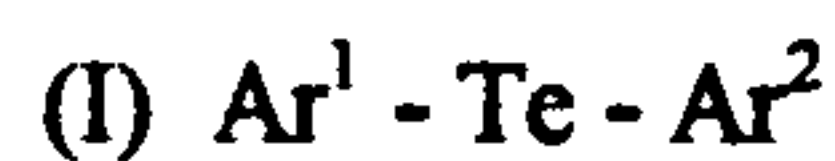
(a) a label layer comprising an enzyme labeled ligand or an enzyme labeled receptor;

(b) a spreading layer ;

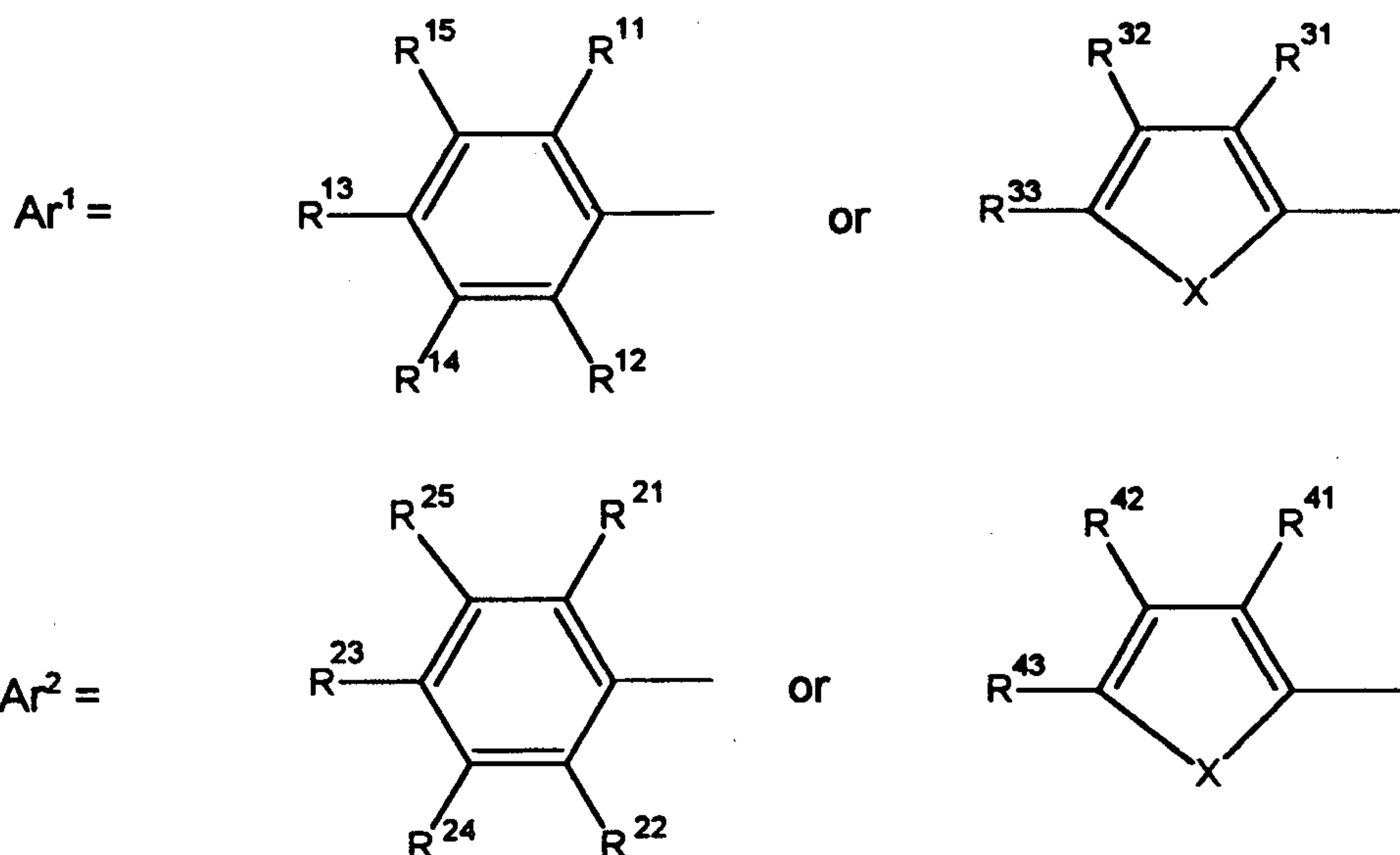
(c) a receptor layer comprising a fixed concentration of an immobilized receptor for the ligand; and

(d) a gravure layer comprising a diaryl telluride.

2. The element according to claim 1 wherein the diaryl telluride compound comprises a structure (I):



wherein Ar^1 and Ar^2 are the same or different aryl or heteroaryl groups where



wherein X is O, S, Se, or Te, and R^{11} , R^{12} , R^{13} , R^{21} , R^{22} , R^{23} , R^{31} , R^{32} , R^{33} , R^{41} , R^{42} and R^{43} are the same or different and each selected from the group consisting of hydrogen, alkyl having 1-5 carbon atoms, OH, OR^1 , SH, NH_2 , NHR^1 , NR_2^1 , NR^1R^2 , CO_2H or its salts, CO_2R^1 , SO_3H or its salts, PO_3H_2 or its

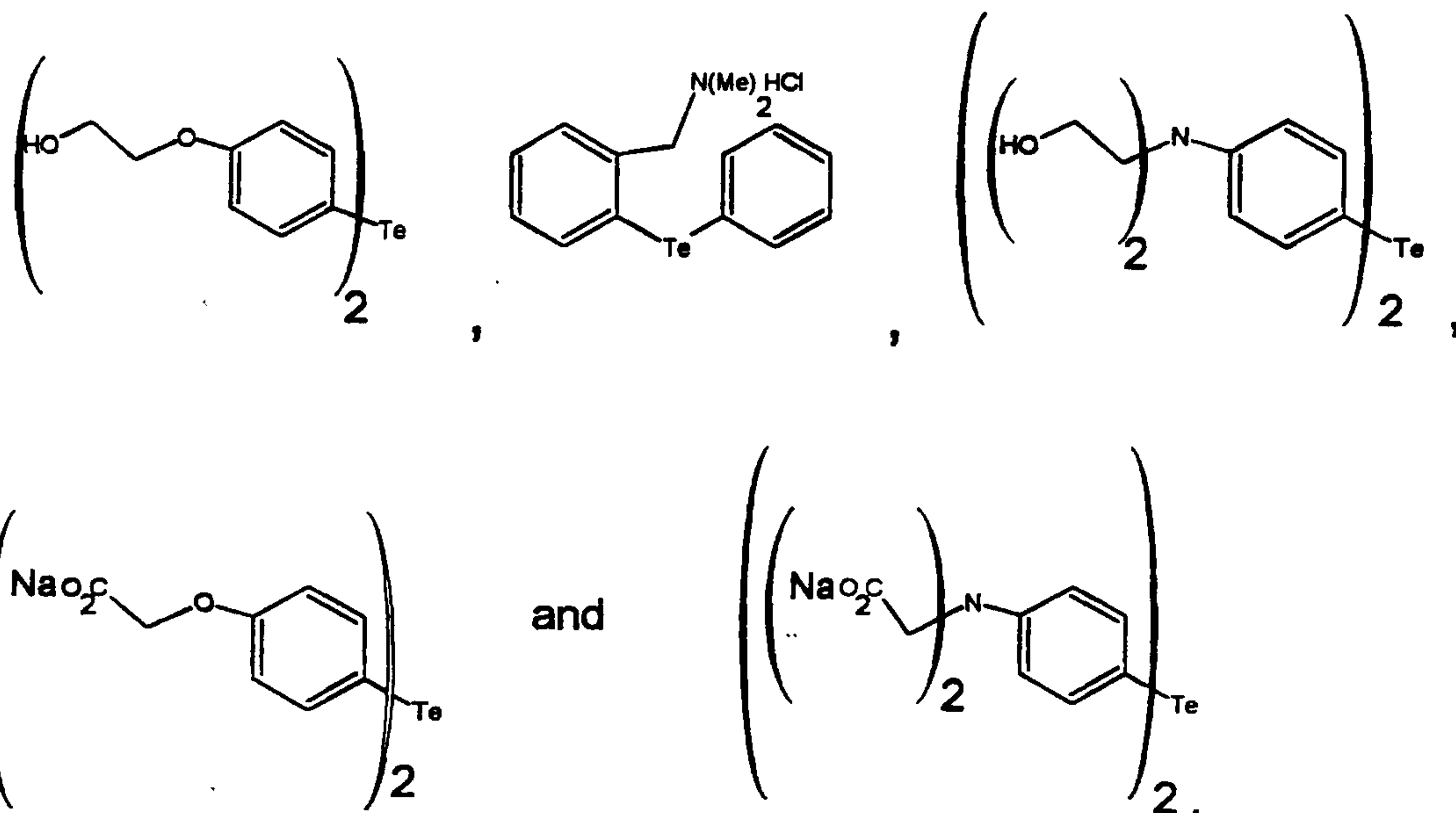
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salts, and SR^1 wherein R^1 and R^2 are different and each selected from the group consisting of alkyl having a carbon chain of 1 to 14 carbon atoms, phenyl, and substituted phenyl;

5 R^{14} , R^{15} , R^{24} , and R^{25} are the same or different and each selected from the group consisting of hydrogen, alkyl having 1-5 carbon atoms, alkoxy having 1-5 carbon atoms, CO_2H or its salts, CO_2R^1 , SO_3H or its salts, and PO_3H_2 or its salts.

3. The element according to claim 2, wherein the alkyl having a carbon chain of 1 to 14 carbon atoms further carries one or more hydrophilic groups.

4. The element according to claim 1 wherein the diaryl telluride is selected from the group consisting of:



15 5. The element according to any one of claims 1 to 4 further comprising a vanadyl salt.

20 6. The element according to claim 1 wherein a peroxidase or a derivative of peroxidase is the label in the labeled ligand or labeled receptor.

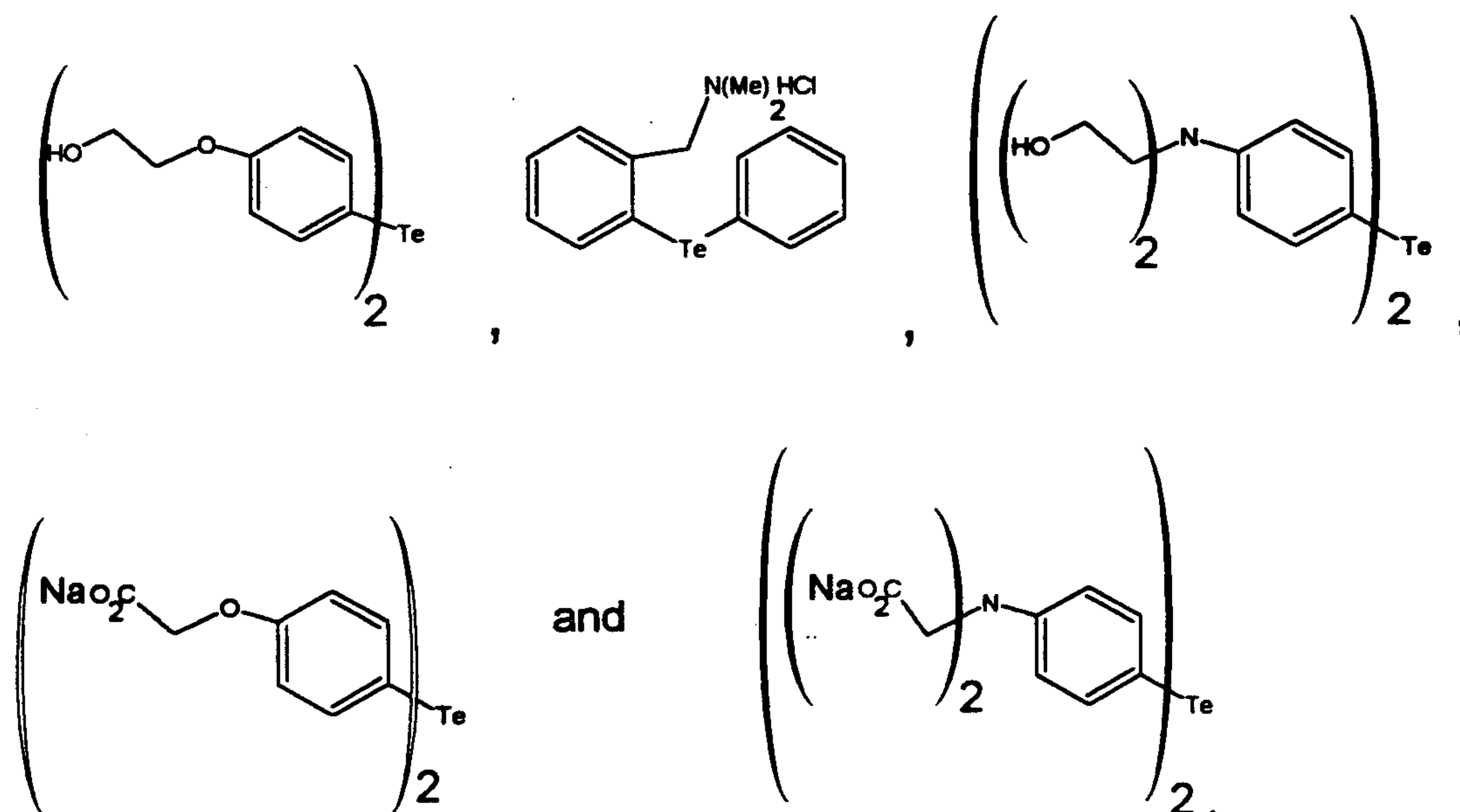
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7. The element according to claim 6 wherein the peroxidase is horseradish peroxidase or a derivative of horseradish peroxidase.

8. The element according to claim 7 wherein the gravure layer also comprises the labeled ligand or labeled receptor.

9. The element according to claim 8 wherein the gravure layer is contiguous with the spreading layer surface that provides contact with a liquid sample.

10. The element according to claim 9 wherein the diaryl telluride is selected from the group consisting of:



11. The element according to claim 10 further comprising a vanadyl salt.

12. A method for assaying an immunologically reactive ligand in an aqueous sample, comprising the steps of:

(1) providing a dry immunoassay analytical element comprising a support bearing:

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(a) a label layer comprising an enzyme labeled ligand or an enzyme labeled receptor;

(b) a spreading layer;

(c) a receptor layer comprising a fixed concentration of an immobilized receptor for the ligand;

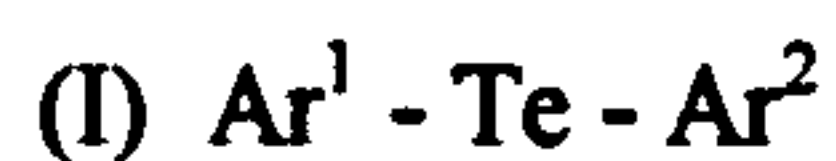
(d) a gravure layer comprising a diaryl telluride;

(2) contacting a finite area of the spreading layer of the element with the sample thereby forming (a) an immobilized ligand-receptor complex, (b) an immobilized enzyme-labeled ligand-receptor complex, or (c) a mixture of (a) and (b), or an immobilized receptor-ligand-labeled receptor complex;

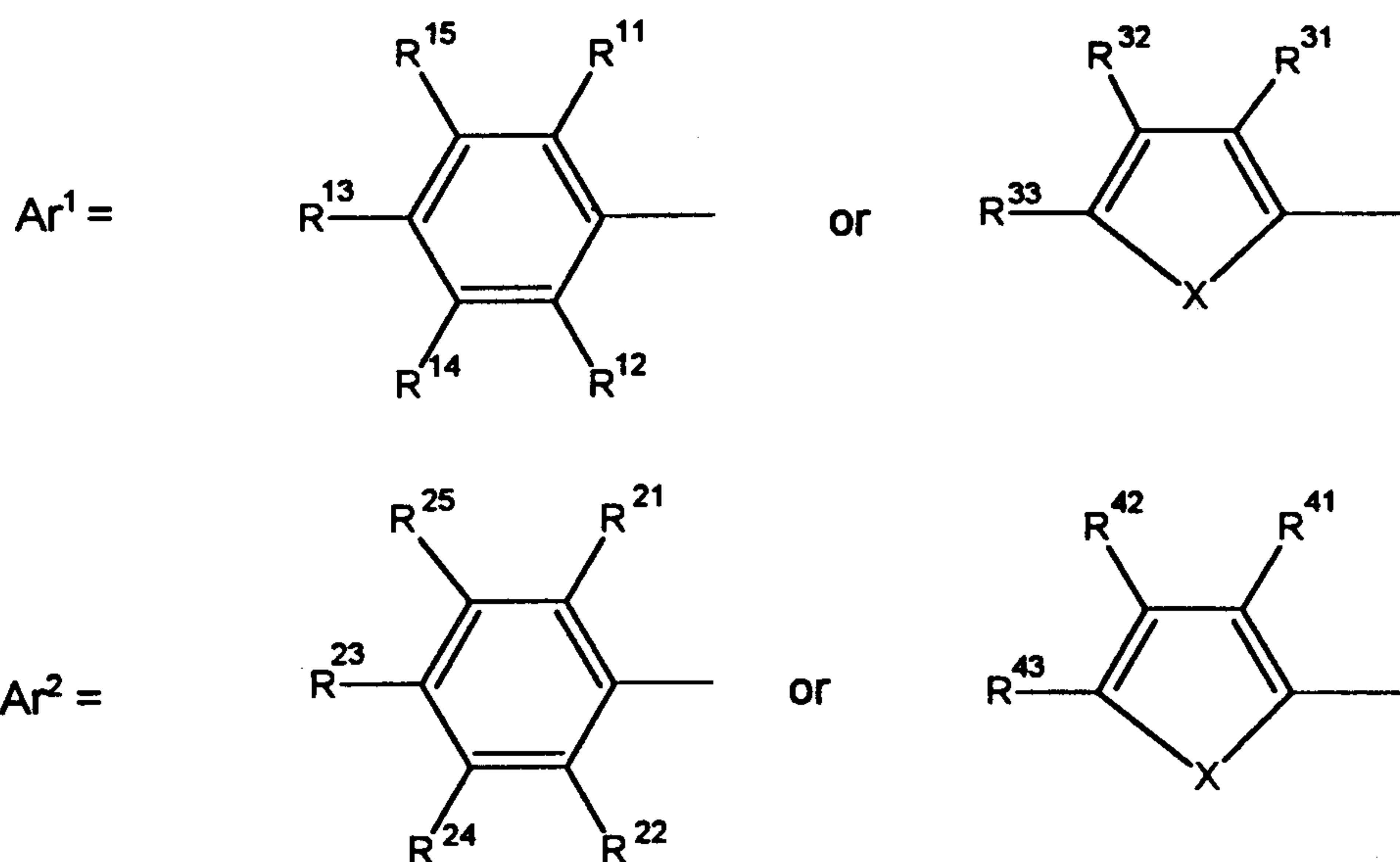
(3) contacting the finite area with a substrate solution for the enzyme label thereby catalyzing the development of a signal ; and

(4) determining the concentration of the ligand by detecting the signal.

13. The method according to claim 12 wherein the diaryl telluride compound comprises a structure (I):



wherein Ar^1 and Ar^2 are the same or different aryl or heteroaryl groups where



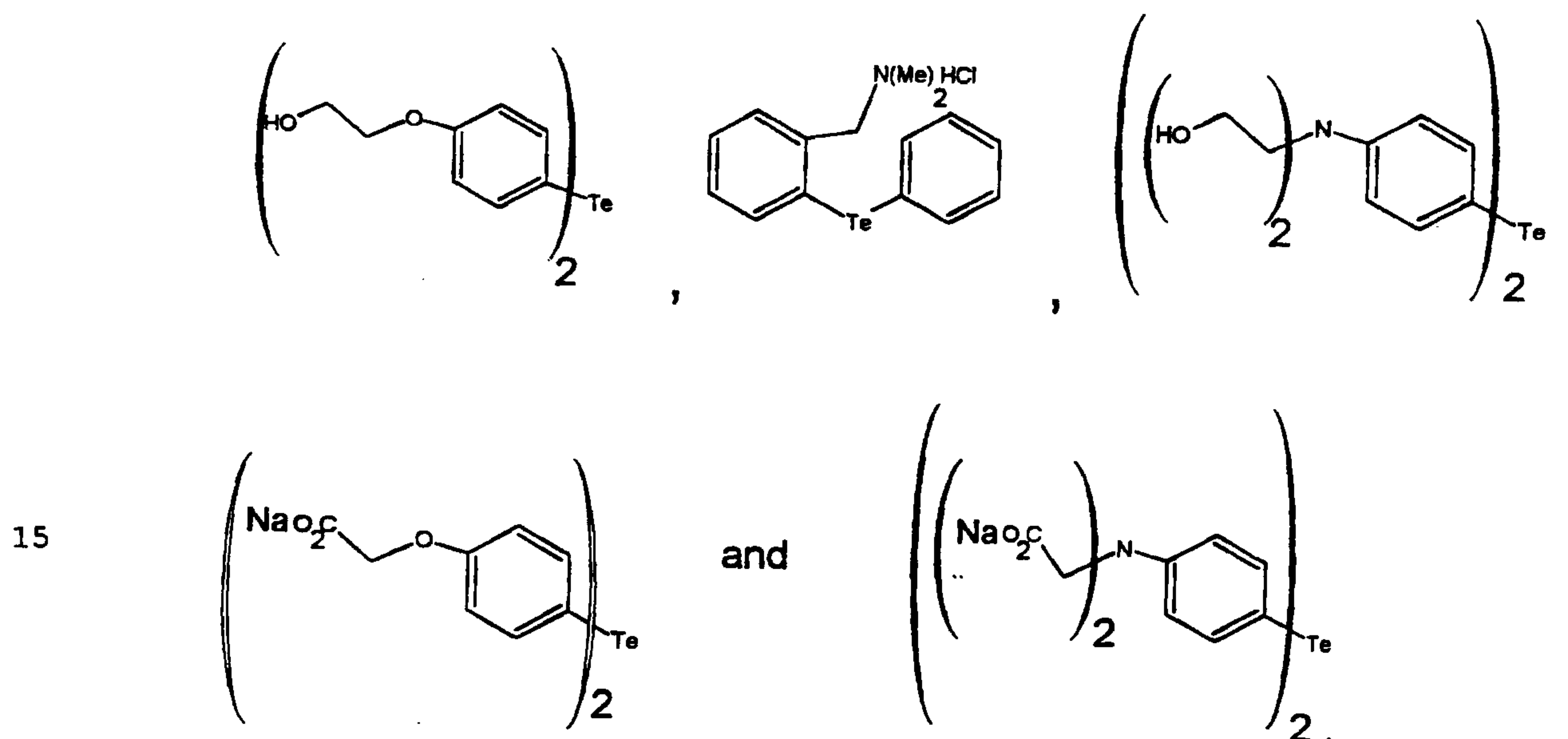
- 71 -

5 wherein X is O, S, Se, or Te, and R^{11} , R^{12} , R^{13} , R^{21} , R^{22} , R^{23} , R^{31} , R^{32} , R^{33} , R^{41} , R^{42} and R^{43} are the same or different and each selected from the group consisting of hydrogen; alkyl having 1-5 carbon atoms; OH, OR^1 , SH, NH_2 , NHR^1 , NR^1_2 , NR^1R^2 , CO_2H or its salts; CO_2R^1 , SO_3H or its salts, PO_3H_2 or its salts, and SR^1 wherein R^1 and R^2 are different and each selected from the group consisting of alkyl having a carbon chain of 1 to 14 carbon atoms, phenyl, and substituted phenyl;

10 R^{14} , R^{15} , R^{24} , and R^{25} are the same or different and each selected from the group consisting of hydrogen, alkyl having 1-5 carbon atoms, alkoxy having 1-5 carbon atoms, CO_2H or its salts, CO_2R^1 , SO_3H or its salts, and PO_3H_2 or its salts.

14. The method according to claim 13, wherein the alkyl having a carbon chain of 1 to 14 carbon atoms further carries one or more hydrophilic groups.

15. The method according to claim 12 wherein the diaryl telluride is selected from the group consisting of:

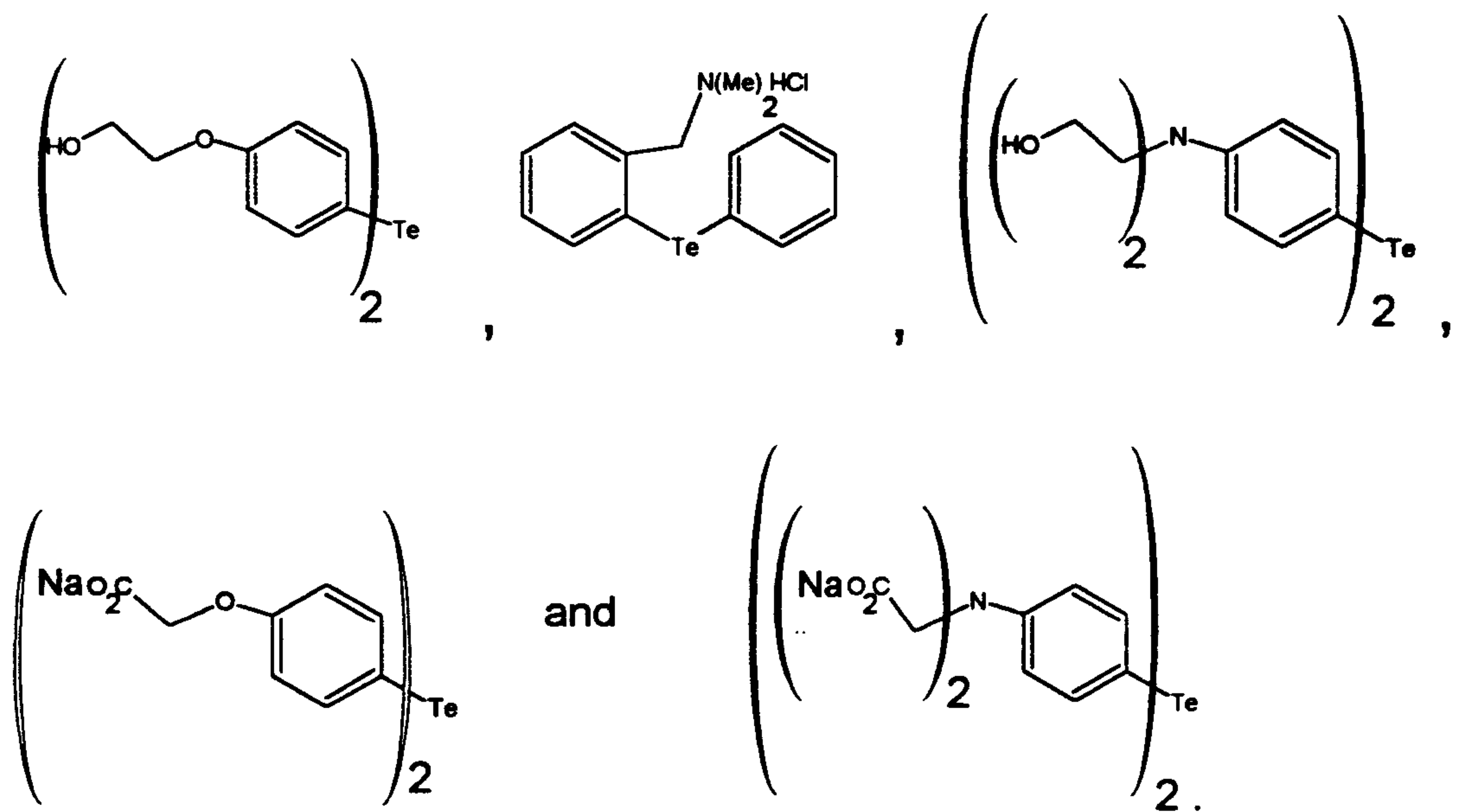


16. The method according to any one of claims 12 to 15 wherein the element further comprises a vanadyl salt.

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- 5 17. The method according to claim 12 wherein a peroxidase or a derivative of peroxidase
is the label in the labeled ligand or labeled receptor.
18. The method according to claim 17 wherein the peroxidase is horseradish peroxidase
or a derivative of horseradish peroxidase.
- 10 19. The method according to claim 18 wherein the signal is a colorimetric signal.
20. The method according to claim 19 wherein the gravure layer further comprises the
labeled ligand or labeled receptor.
- 15 21. The method according to claim 20 wherein the gravure layer is contiguous with the
spreading layer surface that provides contact with a liquid sample.
22. The method according to claim 21 wherein the diaryl telluride is selected from the
group consisting of:
- 20

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23. The method according to claim 22 wherein the element further comprises a vanadyl salt.

FIG. 1

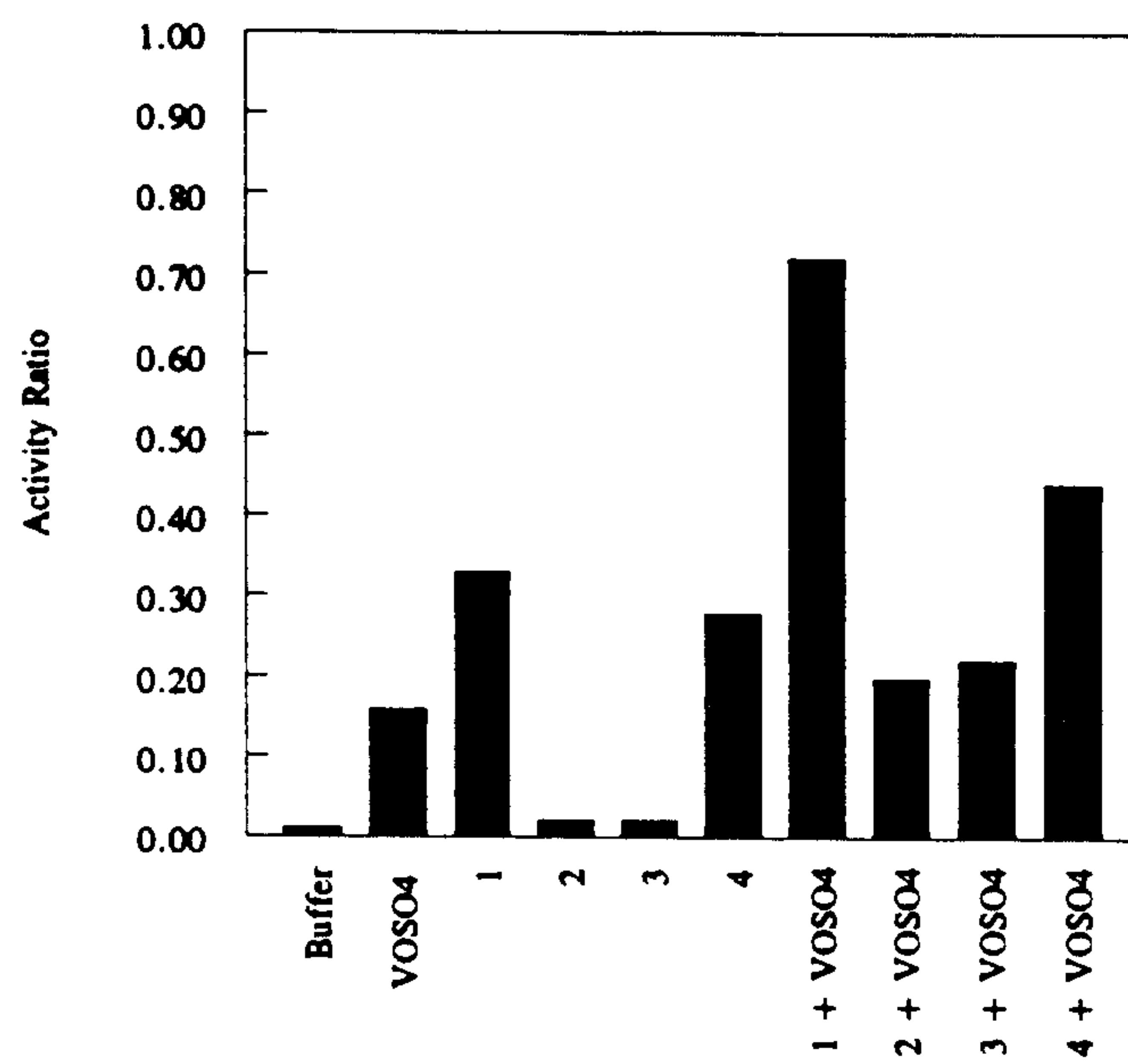


FIG. 2

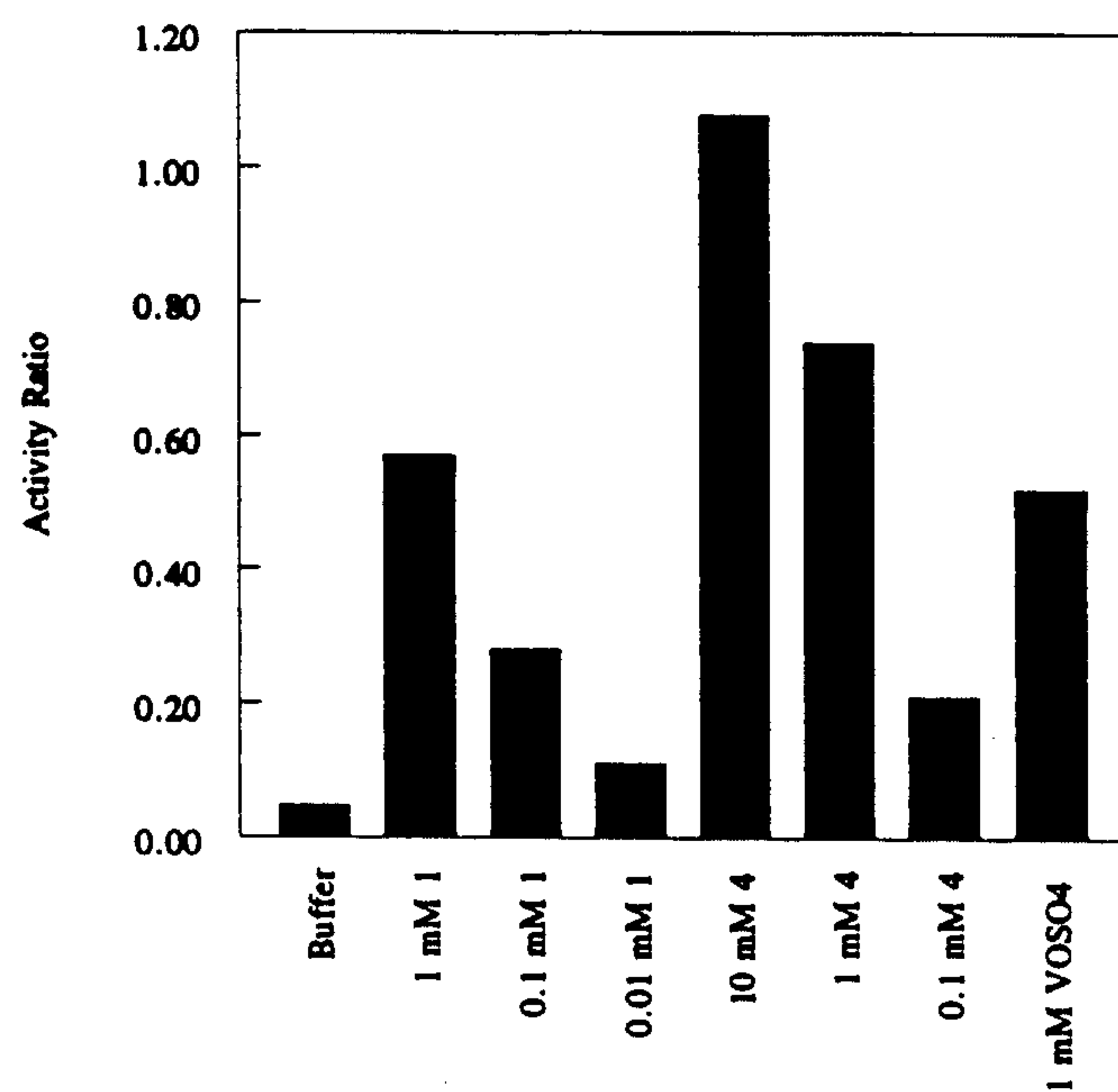
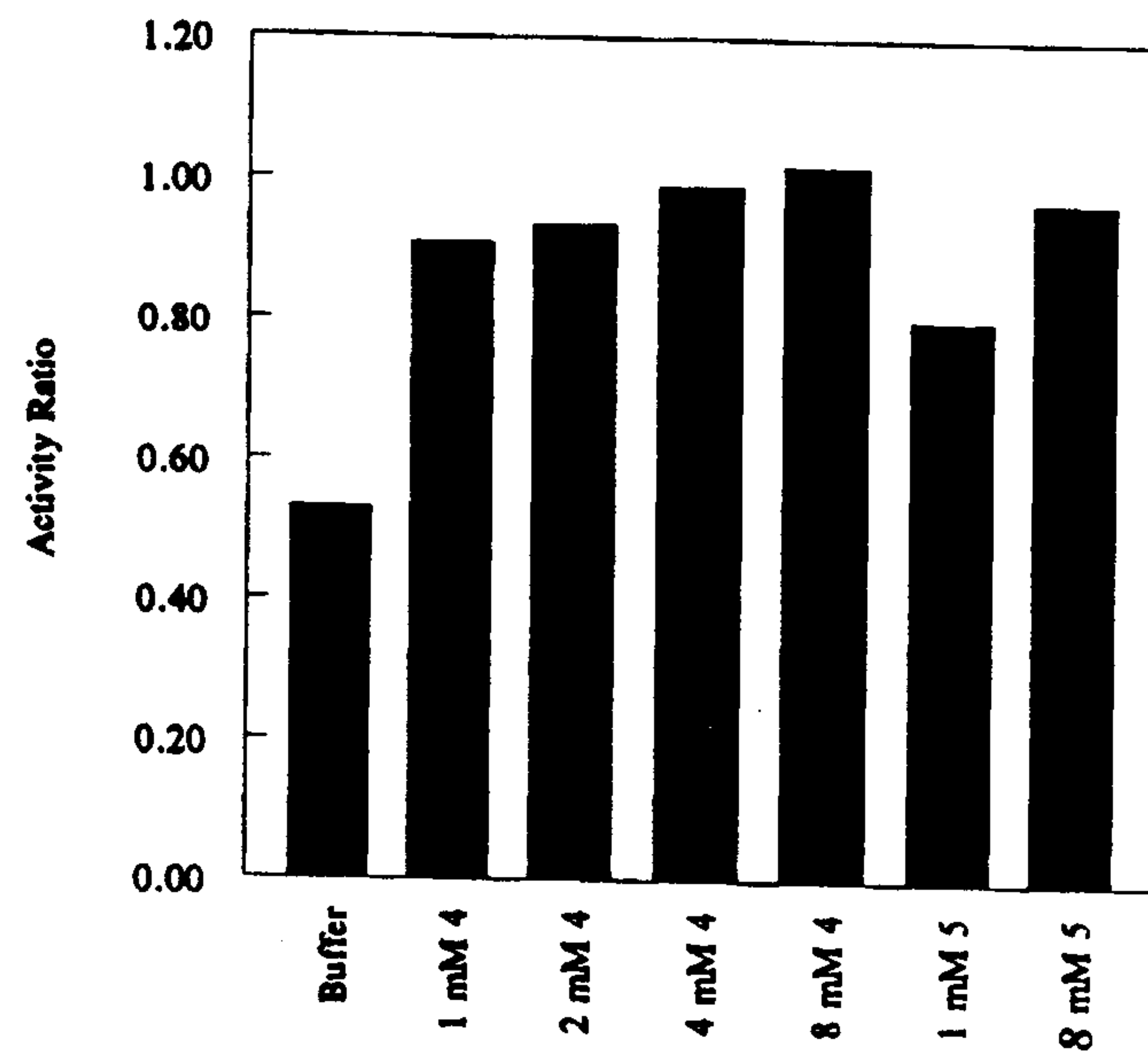


FIG. 3



Activity Ratio

