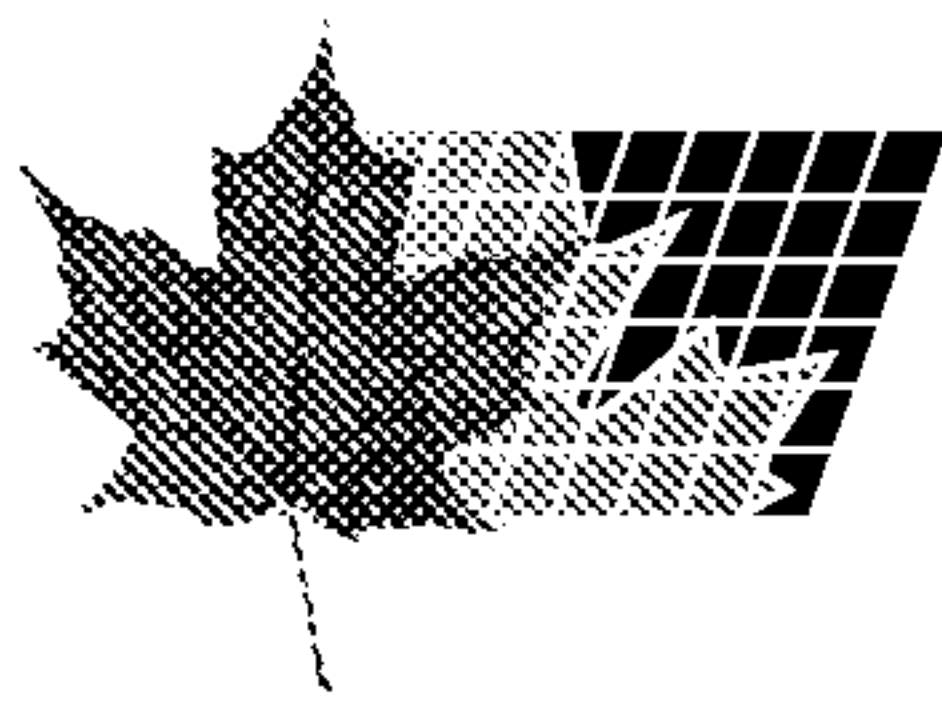




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(54) **ANIMAUX TRANSGENIQUES NON HUMAINS CAPABLES DE
PRODUIRE DES ANTICORPS HETEROLOGUES**

(54) **TRANSGENIC NON-HUMAN ANIMALS CAPABLE OF
PRODUCING HETEROLOGOUS ANTIBODIES**

(57) L'invention porte sur des animaux transgéniques non humains capables de produire des anticorps hétérologues, et sur des procédés de production d'anticorps à séquence humaine qui se fixent aux antigènes humains avec une affinité substantielle.

(57) The invention relates to transgenic non-human animals capable of producing heterologous antibodies and methods for producing human sequence antibodies which bind to human antigens with substantial affinity.

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(54) Title: TRANSGENIC NON-HUMAN ANIMALS CAPABLE OF PRODUCING HETEROLOGOUS ANTIBODIES (57) Abstract The invention relates to transgenic non-human animals capable of producing heterologous antibodies and methods for producing human sequence antibodies which bind to human antigens with substantial affinity.		

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**TRANSGENIC NON-HUMAN ANIMALS CAPABLE OF PRODUCING
HETEROLOGOUS ANTIBODIES**

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TECHNICAL FIELD

The invention relates to transgenic non-human animals capable of producing heterologous antibodies, transgenes used to produce such transgenic animals, transgenes capable of functionally rearranging a heterologous D gene in V-D-J recombination, immortalized B-cells capable of producing heterologous antibodies, methods and transgenes for producing heterologous antibodies of multiple isotypes, methods and transgenes for producing heterologous antibodies wherein a variable region sequence comprises somatic mutation as compared to germline rearranged variable region sequences, transgenic nonhuman animals which produce antibodies having a human primary sequence and which bind to human antigens, hybridomas made from B cells of such transgenic animals, and monclonal antibodies expressed by such hybridomas.

BACKGROUND OF THE INVENTION

One of the major impediments facing the development of in vivo therapeutic and diagnostic applications for monoclonal antibodies in humans is the intrinsic immunogenicity of non-human immunoglobulins. For example, when immunocompetent human patients are administered therapeutic doses of rodent monoclonal antibodies, the patients produce antibodies against the rodent immunoglobulin sequences; these human anti-mouse antibodies (HAMA) neutralize the therapeutic antibodies and can cause acute toxicity. Hence, it is desirable to produce human immunoglobulins that are reactive

with specific human antigens that are promising therapeutic and/or diagnostic targets. However, producing human immunoglobulins that bind specifically with human antigens is problematic.

5 The present technology for generating monoclonal antibodies involves pre-exposing, or priming, an animal (usually a rat or mouse) with antigen, harvesting B-cells from that animal, and generating a library of hybridoma clones. By screening a hybridoma population for antigen binding
10 specificity (idiotype) and also screening for immunoglobulin class (isotype), it is possible to select hybridoma clones that secrete the desired antibody.

 However, when present methods for generating monoclonal antibodies are applied for the purpose of
15 generating human antibodies that have binding specificities for human antigens, obtaining B-lymphocytes which produce human immunoglobulins a serious obstacle, since humans will typically not make immune responses against self-antigens.

 Hence, present methods of generating human
20 monoclonal antibodies that are specifically reactive with human antigens are clearly insufficient. It is evident that the same limitations on generating monoclonal antibodies to authentic self antigens apply where non-human species are used as the source of B-cells for making the hybridoma.

25 The construction of transgenic animals harboring a functional heterologous immunoglobulin transgene are a method by which antibodies reactive with self antigens may be produced. However, in order to obtain expression of therapeutically useful antibodies, or hybridoma clones
30 producing such antibodies, the transgenic animal must produce transgenic B cells that are capable of maturing through the B lymphocyte development pathway. Such maturation requires the

presence of surface IgM on the transgenic B cells, however isotypes other than IgM are desired for therapeutic uses. Thus, there is a need for transgenes and animals harboring such transgenes that are able to undergo functional V-D-J
5 rearrangement to generate recombinational diversity and junctional diversity. Further, such transgenes and transgenic animals preferably include cis-acting sequences that facilitate isotype switching from a first isotype that is required for B cell maturation to a subsequent isotype that
10 has superior therapeutic utility.

A number of experiments have reported the use of transfected cell lines to determine the specific DNA sequences required for Ig gene rearrangement (reviewed by Lewis and Gellert (1989), Cell, 59, 585-588). Such reports have
15 identified putative sequences and concluded that the accessibility of these sequences to the recombinase enzymes used for rearrangement is modulated by transcription (Yancopoulos and Alt (1985), Cell, 40, 271-281). The sequences for V(D)J joining are reportedly a highly conserved,
20 near-palindromic heptamer and a less well conserved AT-rich nanomer separated by a spacer of either 12 or 23 bp (Tonegawa (1983), Nature, 302, 575-581; Hesse, et al. (1989), Genes in Dev., 3, 1053-1061). Efficient recombination reportedly occurs only between sites containing recombination signal
25 sequences with different length spacer regions.

Ig gene rearrangement, though studied in tissue culture cells, has not been extensively examined in transgenic mice. Only a handful of reports have been published describing rearrangement test constructs introduced into mice
30 [Buchini, et al. (1987), Nature, 326, 409-411 (unrearranged chicken λ transgene); Goodhart, et al. (1987), Proc. Natl. Acad. Sci. USA, 84, 4229-4233) (unrearranged rabbit κ gene);

and Bruggemann, et al. (1989), Proc. Natl. Acad. Sci. USA, 86, 6709-6713 (hybrid mouse-human heavy chain)]. The results of such experiments, however, have been variable, in some cases, producing incomplete or minimal rearrangement of the transgene.

Further, a variety of biological functions of antibody molecules are exerted by the Fc portion of molecules, such as the interaction with mast cells or basophils through Fcε, and binding of complement by Fcμ or Fcγ, it further is desirable to generate a functional diversity of antibodies of a given specificity by variation of isotype.

Although transgenic animals have been generated that incorporate transgenes encoding one or more chains of a heterologous antibody, there have been no reports of heterologous transgenes that undergo successful isotype switching. Transgenic animals that cannot switch isotypes are limited to producing heterologous antibodies of a single isotype, and more specifically are limited to producing an isotype that is essential for B cell maturation, such as IgM and possibly IgD, which may be of limited therapeutic utility. Thus, there is a need for heterologous immunoglobulin transgenes and transgenic animals that are capable of switching from an isotype needed for B cell development to an isotype that has a desired characteristic for therapeutic use.

Based on the foregoing, it is clear that a need exists for methods of efficiently producing heterologous antibodies, e.g. antibodies encoded by genetic sequences of a first species that are produced in a second species. More particularly, there is a need in the art for heterologous immunoglobulin transgenes and transgenic animals that are capable of undergoing functional V-D-J gene rearrangement that incorporates all or a portion of a D gene segment which

contributes to recombinational diversity. Further, there is a need in the art for transgenes and transgenic animals that can support V-D-J recombination and isotype switching so that (1) functional B cell development may occur, and (2)

5 therapeutically useful heterologous antibodies may be produced. There is also a need for a source of B cells which can be used to make hybridomas that produce monoclonal antibodies for therapeutic or diagnostic use in the particular species for which they are designed. A heterologous
10 immunoglobulin transgene capable of functional V-D-J recombination and/or capable of isotype switching could fulfill these needs.

In accordance with the foregoing object transgenic nonhuman animals are provided which are capable of producing a
15 heterologous antibody, such as a human antibody.

Further, it is an object to provide B-cells from such transgenic animals which are capable of expressing heterologous antibodies wherein such B-cells are immortalized to provide a source of a monoclonal antibody specific for a
20 particular antigen.

In accordance with this foregoing object, it is a further object of the invention to provide hybridoma cells that are capable of producing such heterologous monoclonal antibodies.

25 Still further, it is an object herein to provide heterologous unrearranged and rearranged immunoglobulin heavy and light chain transgenes useful for producing the aforementioned non-human transgenic animals.

Still further, it is an object herein to provide
30 methods to disrupt endogenous immunoglobulin loci in the transgenic animals.

WO 98/24884

PCT/US97/21803

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Still further, it is an object herein to provide methods to induce heterologous antibody production in the aforementioned transgenic non-human animal.

A further object of the invention is to provide methods to generate an immunoglobulin variable region gene segment repertoire that is used to construct one or more transgenes of the invention.

The references discussed herein are provided solely for their disclosure prior to the filing date of the present application. Nothing herein is to be construed as an admission that the inventors are not entitled to antedate such disclosure by virtue of prior invention.

SUMMARY OF THE INVENTION

Transgenic nonhuman animals are provided which are capable of producing a heterologous antibody, such as a human antibody. Such heterologous antibodies may be of various isotypes, including: IgG1, IgG2, IgG3, IgG4, IgM, IgA1, IgA2, IgA_{sec}, IgD, or IgE. In order for such transgenic nonhuman animals to make an immune response, it is necessary for the transgenic B cells and pre-B cells to produce surface-bound immunoglobulin, particularly of the IgM (or possibly IgD) isotype, in order to effectuate B cell development and antigen-stimulated maturation. Such expression of an IgM (or IgD) surface-bound immunoglobulin is only required during the antigen-stimulated maturation phase of B cell development, and mature B cells may produce other isotypes, although only a single switched isotype may be produced at a time.

Typically, a cell of the B-cell lineage will produce only a single isotype at a time, although cis or trans alternative RNA splicing, such as occurs naturally with the μ_s (secreted μ) and μ_m (membrane-bound μ) forms, and the μ and δ

immunoglobulin chains, may lead to the contemporaneous expression of multiple isotypes by a single cell. Therefore, in order to produce heterologous antibodies of multiple isotypes, specifically the therapeutically useful IgG, IgA, 5 and IgE isotypes, it is necessary that isotype switching occur. Such isotype switching may be classical class-switching or may result from one or more non-classical isotype switching mechanisms.

The invention provides heterologous immunoglobulin 10 transgenes and transgenic nonhuman animals harboring such transgenes, wherein the transgenic animal is capable of producing heterologous antibodies of multiple isotypes by undergoing isotype switching. Classical isotype switching occurs by recombination events which involve at least one 15 switch sequence region in the transgene. Non-classical isotype switching may occur by, for example, homologous recombination between human σ_μ and human Σ_μ sequences (δ -associated deletion). Alternative non-classical switching mechanisms, such as intertransgene and/or interchromosomal 20 recombination, among others, may occur and effectuate isotype switching. Such transgenes and transgenic nonhuman animals produce a first immunoglobulin isotype that is necessary for antigen-stimulated B cell maturation and can switch to encode and produce one or more subsequent heterologous isotypes that 25 have therapeutic and/or diagnostic utility. Transgenic nonhuman animals of the invention are thus able to produce, in one embodiment, IgG, IgA, and/or IgE antibodies that are encoded by human immunoglobulin genetic sequences and which also bind specific human antigens with high affinity.

30 The invention also encompasses B-cells from such transgenic animals that are capable of expressing heterologous antibodies of various isotypes, wherein such B-cells are

immortalized to provide a source of a monoclonal antibody specific for a particular antigen. Hybridoma cells that are derived from such B-cells can serve as one source of such heterologous monoclonal antibodies.

5 The invention provides heterologous unrearranged and rearranged immunoglobulin heavy and light chain transgenes capable of undergoing isotype switching in vivo in the aforementioned non-human transgenic animals or in explanted lymphocytes of the B-cell lineage from such transgenic
10 animals. Such isotype switching may occur spontaneously or be induced by treatment of the transgenic animal or explanted B-lineage lymphocytes with agents that promote isotype switching, such as T-cell-derived lymphokines (e.g., IL-4 and IFN_γ).

15 Still further, the invention includes methods to induce heterologous antibody production in the aforementioned transgenic non-human animal, wherein such antibodies may be of various isotypes. These methods include producing an antigen-stimulated immune response in a transgenic nonhuman animal for
20 the generation of heterologous antibodies, particularly heterologous antibodies of a switched isotype (i.e., IgG, IgA, and IgE).

 This invention provides methods whereby the transgene contains sequences that effectuate isotype
25 switching, so that the heterologous immunoglobulins produced in the transgenic animal and monoclonal antibody clones derived from the B-cells of said animal may be of various isotypes.

 This invention further provides methods that
30 facilitate isotype switching of the transgene, so that switching between particular isotypes may occur at much higher or lower frequencies or in different temporal orders than

typically occurs in germline immunoglobulin loci. Switch regions may be grafted from various C_H genes and ligated to other C_H genes in a transgene construct; such grafted switch sequences will typically function independently of the
5 associated C_H gene so that switching in the transgene construct will typically be a function of the origin of the associated switch regions. Alternatively, or in combination with switch sequences, δ -associated deletion sequences may be linked to various C_H genes to effect non-classical switching by deletion
10 of sequences between two δ -associated deletion sequences. Thus, a transgene may be constructed so that a particular C_H gene is linked to a different switch sequence and thereby is switched to more frequently than occurs when the naturally associated switch region is used.

15 This invention also provides methods to determine whether isotype switching of transgene sequences has occurred in a transgenic animal containing an immunoglobulin transgene.

The invention provides immunoglobulin transgene constructs and methods for producing immunoglobulin transgene
20 constructs, some of which contain a subset of germline immunoglobulin loci sequences (which may include deletions). The invention includes a specific method for facilitated cloning and construction of immunoglobulin transgenes, involving a vector that employs unique XhoI and SalI
25 restriction sites flanked by two unique NotI sites. This method exploits the complementary termini of XhoI and SalI restrictions sites and is useful for creating large constructs by ordered concatemerization of restriction fragments in a vector.

30 The transgenes of the invention include a heavy chain transgene comprising DNA encoding at least one variable gene segment, one diversity gene segment, one joining gene

segment and one constant region gene segment. The immunoglobulin light chain transgene comprises DNA encoding at least one variable gene segment, one joining gene segment and one constant region gene segment. The gene segments encoding the light and heavy chain gene segments are heterologous to the transgenic non-human animal in that they are derived from, or correspond to, DNA encoding immunoglobulin heavy and light chain gene segments from a species not consisting of the transgenic non-human animal. In one aspect of the invention, the transgene is constructed such that the individual gene segments are unrearranged, i.e., not rearranged so as to encode a functional immunoglobulin light or heavy chain. Such unrearranged transgenes permit recombination of the gene segments (functional rearrangement) and expression of the resultant rearranged immunoglobulin heavy and/or light chains within the transgenic non-human animal when said animal is exposed to antigen.

In one aspect of the invention, heterologous heavy and light immunoglobulin transgenes comprise relatively large fragments of unrearranged heterologous DNA. Such fragments typically comprise a substantial portion of the C, J (and in the case of heavy chain, D) segments from a heterologous immunoglobulin locus. In addition, such fragments also comprise a substantial portion of the variable gene segments.

In one embodiment, such transgene constructs comprise regulatory sequences, e.g. promoters, enhancers, class switch regions, recombination signals and the like, corresponding to sequences derived from the heterologous DNA. Alternatively, such regulatory sequences may be incorporated into the transgene from the same or a related species of the non-human animal used in the invention. For example, human immunoglobulin gene segments may be combined in a transgene

with a rodent immunoglobulin enhancer sequence for use in a transgenic mouse.

In a method of the invention, a transgenic non-human animal containing germline unrearranged light and heavy
5 immunoglobulin transgenes - that undergo VDJ joining during D-cell differentiation - is contacted with an antigen to induce production of a heterologous antibody in a secondary repertoire B-cell.

Also included in the invention are vectors and
10 methods to disrupt the endogenous immunoglobulin loci in the non-human animal to be used in the invention. Such vectors and methods utilize a transgene, preferably positive-negative selection vector, which is constructed such that it targets the functional disruption of a class of gene segments encoding
15 a heavy and/or light immunoglobulin chain endogenous to the non-human animal used in the invention. Such endogenous gene segments include diversity, joining and constant region gene segments. In this aspect of the invention, the positive-negative selection vector is contacted with at least
20 one embryonic stem cell of a non-human animal after which cells are selected wherein the positive-negative selection vector has integrated into the genome of the non-human animal by way of homologous recombination. After transplantation, the resultant transgenic non-human animal is substantially
25 incapable of mounting an immunoglobulin-mediated immune response as a result of homologous integration of the vector into chromosomal DNA. Such immune deficient non-human animals may thereafter be used for study of immune deficiencies or used as the recipient of heterologous immunoglobulin heavy and
30 light chain transgenes.

The invention also provides vectors, methods, and compositions useful for suppressing the expression of one or

more species of immunoglobulin chain(s), without disrupting an endogenous immunoglobulin locus. Such methods are useful for suppressing expression of one or more endogenous immunoglobulin chains while permitting the expression of one or more transgene-encoded immunoglobulin chains. Unlike genetic disruption of an endogenous immunoglobulin chain locus, suppression of immunoglobulin chain expression does not require the time-consuming breeding that is needed to establish transgenic animals homozygous for a disrupted endogenous Ig locus. An additional advantage of suppression as compared to endogenous Ig gene disruption is that, in certain embodiments, chain suppression is reversible within an individual animal. For example, Ig chain suppression may be accomplished with: (1) transgenes encoding and expressing antisense RNA that specifically hybridizes to an endogenous Ig chain gene sequence, (2) antisense oligonucleotides that specifically hybridize to an endogenous Ig chain gene sequence, and (3) immunoglobulins that bind specifically to an endogenous Ig chain polypeptide.

The invention provides transgenic non-human animals comprising: a homozygous pair of functionally disrupted endogenous heavy chain alleles, a homozygous pair of functionally disrupted endogenous light chain alleles, at least one copy of a heterologous immunoglobulin heavy chain transgene, and at least one copy of a heterologous immunoglobulin heavy chain transgene, wherein said animal makes an antibody response following immunization with an antigen, such as a human antigen (e.g., CD4). The invention also provides such a transgenic non-human animal wherein said functionally disrupted endogenous heavy chain allele is a J_H region homologous recombination knockout, said functionally disrupted endogenous light chain allele is a J_κ region

homologous recombination knockout, said heterologous immunoglobulin heavy chain transgene is the HC1 or HC2 human minigene transgene, said heterologous light chain transgene is the KC2 or KC1e human κ transgene, and wherein said antigen
5 is a human antigen.

The invention also provides various embodiments for suppressing, ablating, and/or functionally disrupting the endogenous nonhuman immunoglobulin loci.

The invention also provides transgenic mice
10 expressing both human sequence heavy chains and chimeric heavy chains comprising a human sequence heavy chain variable region and a murine sequence heavy chain constant region. Such chimeric heavy chains are generally produced by trans-switching between a functionally rearranged human transgene
15 and an endogenous murine heavy chain constant region (e.g., $\gamma 1$, $\gamma 2a$, $\gamma 2b$, $\gamma 3$). Antibodies comprising such chimeric heavy chains, typically in combination with a transgene-encoded human sequence light chain or endogenous murine light chain, are formed in response to immunization with a predetermined
20 antigen. The transgenic mice of these embodiments can comprise B cells which produce (express) a human sequence heavy chain at a first timepoint and trans-switch to produce (express) a chimeric heavy chain composed of a human variable region and a murine constant region (e.g., $\gamma 1$, $\gamma 2a$, $\gamma 2b$, $\gamma 3$)
25 at a second (subsequent) timepoint; such human sequence and chimeric heavy chains are incorporated into functional antibodies with light chains; such antibodies are present in the serum of such transgenic mice. Thus, to restate: the transgenic mice of these embodiments can comprise B cells
30 which express a human sequence heavy chain and subsequently switch (via trans-switching or cis-switching) to express a chimeric or isotype-switched heavy chain composed of a human

variable region and a alternative constant region (e.g., murine $\gamma 1$, $\gamma 2a$, $\gamma 2b$, $\gamma 3$; human γ , α , ϵ); such human sequence and chimeric or isotype-switched heavy chains are incorporated into functional antibodies with light chains (human or mouse);
5 such antibodies are present in the serum of such transgenic mice.

The invention also provides a method for generating a large transgene, said method comprising:

introducing into a mammalian cell at least three
10 polynucleotide species; a first polynucleotide species having a recombinogenic region of sequence identity shared with a second polynucleotide species, a second polynucleotide species having a recombinogenic region of sequence identity shared with a first polynucleotide species and a recombinogenic
15 region of sequence identity shared with a third polynucleotide species, and a third polynucleotide species having a recombinogenic region of sequence identity shared with said second polynucleotide species.

Recombinogenic regions are regions of substantial
20 sequence identity sufficient to generate homologous recombination in vivo in a mammalian cell (e.g., ES cell), and preferably also in non-mammalian eukaryotic cells (e.g., *Saccharomyces* and other yeast or fungal cells). Typically, recombinogenic regions are at least 50 to 100000 nucleotides
25 long or longer, preferably 500 nucleotides to 10000 nucleotides long, and are often 80-100 percent identical, frequently 95-100 percent identical, often isogenic.

The references discussed herein are provided solely for their disclosure prior to the filing date of the present
30 application. Nothing herein is to be construed as an admission that the inventors are not entitled to antedate such disclosure by virtue of prior invention.

BRIEF DESCRIPTION OF THE FIGURES

Fig. 1 depicts the complementarity determining regions CDR1, CDR2 and CDR3 and framework regions FR1, FR2, FR3 and FR4 in unrearranged genomic DNA and mRNA expressed
5 from a rearranged immunoglobulin heavy chain gene,

Fig. 2 depicts the human λ chain locus,

Fig. 3 depicts the human κ chain locus,

Fig. 4 depicts the human heavy chain locus,

Fig. 5 depicts a transgene construct containing a
10 rearranged IgM gene ligated to a 25 kb fragment that contains human $\gamma 3$ and $\gamma 1$ constant regions followed by a 700 bp fragment containing the rat chain 3' enhancer sequence.

Fig. 6 is a restriction map of the human κ chain locus depicting the fragments to be used to form a light chain
15 transgene by way of in vivo homologous recombination.

Fig. 7 depicts the construction of pGP1.

Fig. 8 depicts the construction of the polylinker contained in pGP1.

Fig. 9 depicts the fragments used to construct a
20 human heavy chain transgene of the invention.

Fig. 10 depicts the construction of pHIG1 and pCON1.

Fig. 11 depicts the human C γ 1 fragments which are inserted into pRE3 (rat enhancer 3') to form pREG2.

Fig. 12 depicts the construction of pHIG3' and PCON.

25 Fig. 13 depicts the fragment containing human D region segments used in construction of the transgenes of the invention.

Fig. 14 depicts the construction of pHIG2 (D segment containing plasmid).

30 Fig. 15 depicts the fragments covering the human J κ and human C κ gene segments used in constructing a transgene of the invention.

Fig. 16 depicts the structure of pE μ .

Fig. 17 depicts the construction of pKapH.

Figs. 18A through 18D depict the construction of a positive-negative selection vector for functionally disrupting
5 the endogenous heavy chain immunoglobulin locus of mouse.

Figs. 19A through 19C depict the construction of a positive-negative selection vector for functionally disrupting the endogenous immunoglobulin light chain loci in mouse.

Figs. 20A through 20E depict the structure of a
10 kappa light chain targeting vector.

Figs. 21A through 21F depict the structure of a mouse heavy chain targeting vector.

Fig. 22 depicts the map of vector pGPe.

Fig. 23 depicts the structure of vector pJM2.

15 Fig. 24 depicts the structure of vector pCOR1.

Fig. 25 depicts the transgene constructs for pIGM1, pHC1 and pHC2.

Fig. 26 depicts the structure of pye2.

Fig. 27 depicts the structure of pVGE1.

20 Fig. 28 depicts the assay results of human Ig expression in a pHC1 transgenic mouse.

Fig. 29 depicts the structure of pJCK1.

Fig. 30 depicts the construction of a synthetic heavy chain variable region.

25 Fig. 31 is a schematic representation of the heavy chain minilocus constructs pIGM₁, pHC1, and pHC2.

Fig. 32 is a schematic representation of the heavy chain minilocus construct pIGG1 and the k light chain minilocus construct pKC1, pKVe1, and pKC2.

30 Fig. 33 depicts a scheme to reconstruct functionally rearranged light chain genes.

Fig. 34 depicts serum ELISA results

Fig. 35 depicts the results of an ELISA assay of serum from 8 transgenic mice.

Fig. 36 is a schematic representation of plasmid pBCE1.

5 Figs. 37A-37C depict the immune response of transgenic mice of the present invention against KLH-DNP, by measuring IgG and IgM levels specific for KLH-DNP (37A), KLH (37B) and BSA-DNP (37C).

10 Fig. 38 shows ELISA data demonstrating the presence of antibodies that bind human carcinoembryonic antigen (CEA) and comprise human μ chains; each panel shows reciprocal serial dilutions from pooled serum samples obtained from mice on the indicated day following immunization.

15 Fig. 39 shows ELISA data demonstrating the presence of antibodies that bind human carcinoembryonic antigen (CEA) and comprise human γ chains; each panel shows reciprocal serial dilutions from pooled serum samples obtained from mice on the indicated day following immunization.

20 Fig. 40 shows aligned variable region sequences of 23 randomly-chosen cDNAs generated from mRNA obtained from lymphoid tissue of HCl transgenic mice immunized with human carcinoembryonic antigen (CEA) as compared to the germline transgene sequence (top line); on each line nucleotide changes relative to germline sequence are shown. The regions
25 corresponding to heavy chain CDR1, CDR2, and CDR3 are indicated. Non-germline encoded nucleotides are shown in capital letters.

Fig. 41 show the nucleotide sequence of a human DNA fragment, designated vk65.3, containing a V_k gene segment; the
30 deduced amino acid sequences of the V_k coding regions are also shown; splicing and recombination signal sequences (heptamer/nonamer) are shown boxed.

Fig. 42 show the nucleotide sequence of a human DNA fragment, designated vk65.5, containing a V_k gene segment; the deduced amino acid sequences of the V_k coding regions are also shown; splicing and recombination signal sequences

5 (heptamer/nonamer) are shown boxed.

Fig. 43 show the nucleotide sequence of a human DNA fragment, designated vk65.8, containing a V_k gene segment; the deduced amino acid sequences of the V_k coding regions are also shown; splicing and recombination signal sequences

10 (heptamer/nonamer) are shown boxed.

Fig. 44 show the nucleotide sequence of a human DNA fragment, designated vk65.15, containing a V_k gene segment; the deduced amino acid sequences of the V_k coding regions are also shown; splicing and recombination signal sequences

15 (heptamer/nonamer) are shown boxed.

Fig. 45 shows formation of a light chain minilocus by homologous recombination between two overlapping fragments which were co-injected.

Fig. 46 shows ELISA results for monoclonal
20 antibodies reactive with CEA and non-CEA antigens showing the specificity of antigen binding.

Fig. 47 shows the DNA sequences of 10 cDNAs amplified by PCR to amplify transcripts having a human VDJ and a murine constant region sequence.

25 Fig. 48 shows ELISA results for various dilutions of serum obtained from mice bearing both a human heavy chain minilocus transgene and a human κ minilocus transgene; the mouse was immunized with human CD4 and the data shown represents antibodies reactive with human CD4 and possessing
30 human κ , human μ , or human γ epitopes, respectively.

Fig. 49 shows relative distribution of lymphocytes staining for human μ or mouse μ as determined by FACS for three mouse genotypes.

Fig. 50 shows relative distribution of lymphocytes staining for human κ or mouse κ as determined by FACS for three mouse genotypes.

Fig. 51 shows relative distribution of lymphocytes staining for mouse λ as determined by FACS for three mouse genotypes.

Fig. 52 shows relative distribution of lymphocytes staining for mouse λ or human κ as determined by FACS for four mouse genotypes.

Fig. 53 shows the amounts of human μ , human γ , human κ , mouse μ , mouse γ , mouse κ , and mouse λ chains in the serum of unimmunized 0011 mice.

Fig. 54 shows a scatter plot showing the amounts of human μ , human γ , human κ , mouse μ , mouse γ , mouse κ , and mouse λ chains in the serum of unimmunized 0011 mice of various genotypes.

Fig. 55 shows the titres of antibodies comprising human μ , human γ , or human κ chains in anti-CD4 antibodies in the serum taken at three weeks or seven weeks post-immunization following immunization of a 0011 mouse with human CD4.

Fig. 56 shows a schematic representation of the human heavy chain minilocus transgenes pHC1 and pHC2, and the light chain minilocus transgenes pKC1, pKC1e, and the light chain minilocus transgene created by homologous recombination between pKC2 and Co4 at the site indicated.

Fig. 57 shows a linkage map of the murine lambda light chain locus as taken from Storb et al. (1989) op.cit.; the stippled boxes represent a pseudogene.

Fig. 58 shows a schematic representation of inactivation of the murine λ locus by homologous gene targeting.

Fig. 59 schematically shows the structure of a homologous recombination targeting transgene for deleting genes, such as heavy chain constant region genes.

Fig. 60 shows a map of the BALB/c murine heavy chain locus as taken from Immunoglobulin Genes, Honjo, T, Alt, FW, and Rabbits TH (eds.) Academic Press, NY (1989) p. 129.

Structural genes are shown by closed boxes in the top line; second and third lines show restriction sites with symbols indicated.

Fig. 61 shows a nucleotide sequence of mouse heavy chain locus α constant region gene.

Fig. 62 shows the construction of a frameshift vector (plasmid B) for introducing a two bp frameshift into the murine heavy chain locus J_4 gene.

Fig. 63 shows isotype specific response of transgenic animals during hyperimmunization. The relative levels of reactive human μ and $\gamma 1$ are indicated by a colorimetric ELISA assay (y-axis). We immunized three 7-10 week old male HC1 line 57 transgenic animals (#1991, #2356, #2357), in a homozygous JHD background, by intraperitoneal injections of CEA in Freund's adjuvant. The figure depicts binding of 250 fold dilutions of pooled serum (collected prior to each injection) to CEA coated microtiter wells.

Fig. 64A and 64B show expression of transgene encoded $\gamma 1$ isotype mediated by class switch recombination. The genomic structure of integrated transgenes in two different human $\gamma 1$ expressing hybridomas is consistent with recombination between the μ and $\gamma 1$ switch regions. Fig. 64A shows a Southern blot of PacI/SfiI digested DNA isolated from

three transgene expressing hybridomas. From left to right: clone 92-09A-5H1-5, human $\gamma 1^+/\mu^-$; clone 92-90A-4G2-2, human $\gamma 1^+/\mu^-$; clone 92-09A-4F7-A5-2, human $\gamma 1^-, \mu^+$. All three hybridomas are derived from a 7 month old male mouse

5 hemizygous for the HC1-57 integration, and homozygous for the JHD disruption (mouse #1991). The blot is hybridized with a probe derived from a 2.3 kb BglII/SfiI DNA fragment spanning the 3' half of the human $\gamma 1$ switch region. No switch product is found in the μ expressing hybridoma, while the two $\gamma 1$

10 expressing hybridomas, 92-09A-5H1-5 and 92-09A-4G2-2, contain switch products resulting in PacI/SfiI fragments of 5.1 and 5.3 kb respectively, Fig. 64B is a diagram of two possible deletional mechanisms by which a class switch from μ to $\gamma 1$ can occur. The human μ gene is flanked by 400 bp direct repeats

15 ($\sigma\mu$ and $\Sigma\mu$) which can recombine to delete μ . Class switching by this mechanism will always generate a 6.4 kb PacI/SfiI fragment, while class switching by recombination between the μ and the $\gamma 1$ switch regions will generate a PacI/SfiI fragment between 4 and 7 kb, with size variation between individual

20 switch events. The two $\gamma 1$ expressing hybridomas examined in Fig. 64A appear to have undergone recombination between the μ and $\gamma 1$ switch regions.

Fig. 65 shows chimeric human/mouse immunoglobulin heavy chains generated by trans-switching. cDNA clones of

25 trans-switch products were generated by reverse transcription and PCR amplification of a mixture of spleen and lymph node RNA isolated from a hyperimmunized HC1 transgenic-JHD mouse (#2357; see legend to Fig. 63 for description of animal and immunization schedule). The partial nucleotide sequence of 10

30 randomly picked clones is shown. Lower case letters indicate germline encoded, capital letters indicate nucleotides that cannot be assigned to known germline sequences; these may be

somatic mutations, N nucleotides, or truncated D segments. Both face type indicates mouse γ sequences.

Figs. 66A and 66B show that the rearranged VH251 transgene undergoes somatic mutation in a hyperimmunized. The partial nucleotide sequence of IgG heavy chain variable region cDNA clones from HC1 line 26 mice exhibiting Fig. 66A primary and Fig. 66B secondary responses to antigen. Germline sequence is shown at the top; nucleotide changes from germline are given for each clone. A period indicates identity with germline sequence, capital letters indicate no identified germline origin. The sequences are grouped according to J segment usage. The germline sequence of each of the J segments is shown. Lower case letters within CDR3 sequences indicate identity to known D segment included in the HC1 transgene. The assigned D segments are indicated at the end of each sequence. Unassigned sequences could be derived from N region addition or somatic mutation; or in some cases they are simply too short to distinguish random N nucleotides from known D segments. Fig. 66A primary response: 13 randomly picked VH251- γ 1 cDNA clones. A 4 week old female HC1 line 26-JHD mouse (#2599) was given a single injection of KLH and complete Freund's adjuvant; spleen cell RNA was isolated 5 days later. The overall frequency of somatic mutations within the V segment is 0.06% (2/3,198 bp). Fig. 66B secondary response: 13 randomly picked VH251- γ 1 cDNA clones. A 2 month old female HC1 line 26-JHD mouse (#3204) was given 3 injections of HEL and Freund's adjuvant over one month (a primary injection with complete adjuvant and boosts with incomplete at one week and 3 weeks); spleen and lymph node RNA was isolated 4 months later. The overall frequency of somatic mutations within the V segment is 1.6% (52/3,198 bp).

Figs. 67A and 67B show that extensive somatic mutation is confined to $\gamma 1$ sequences: somatic mutation and class switching occur within the same population of B cells. Partial nucleotide sequence of VH251 cDNA clones isolated from spleen and lymph node cells of HC1 line 57 transgenic-JHD mouse (#2357) hyperimmunized against CEA (see Fig. 63 for immunization schedule). Fig. 67A: IgM: 23 randomly picked VH251- μ cDNA clones. Nucleotide sequence of 156 bp segment including CDRs 1 and 2 surrounding residues. The overall level of somatic mutation is 0.1% (5/3,744 bp). Fig 67B: IgG: 23 randomly picked VH251- $\gamma 1$ cDNA clones. Nucleotide sequence of segment including CDRs 1 through 3 and surrounding residues. The overall frequency of somatic mutation within the V segment is 1.1% (65/5,658 bp). For comparison with the μ sequences in Fig. 67A: the mutation frequency for first 156 nucleotides is 1.1% (41/3,588 bp). See legend to Figs. 66A and 66B for explanation of symbols.

Fig. 68 indicates that VH51P1 and VH56P1 show extensive somatic mutation of in an unimmunized mouse. The partial nucleotide sequence of IgG heavy chain variable region cDNA clones from a 9 week old, unimmunized female HC2 line 2550 transgenic-JHD mouse (#5250). The overall frequency of somatic mutation with the 19 VH56p1 segments is 2.2% (101/4,674 bp). The overall frequency of somatic mutation within the single VH51p1 segment is 2.0% (5/246 bp). See legend to Figs. 66A and 66B for explanation of symbols.

Fig. 69. Double transgenic mice with disrupted endogenous Ig loci contain human IgMk positive B cells. FACS of cells isolated from spleens of 4 mice with different genotypes. Left column: control mouse (#9944, 6 wk old female JH+/-, JCK+/-; heterozygous wild-type mouse heavy and κ -light chain loci, non-transgenic). Second column: human

WO 98/24884

PCT/US97/21803

24

heavy chain transgenic (#9877, 6 wk old female JH-/-, JCK-/-, HC2 line 2550 +; homozygous for disrupted mouse heavy and κ -light chain loci, hemizygous for HC2 transgene). Third column: human κ -light chain transgenic (#9878, 6 wk old female JH-/-, JCK-/-, KCo4 line 4437 +; homozygous for disrupted mouse heavy and κ -light chain loci, hemizygous for KCo4 transgene). Right column: double transgenic (#9879, 6 wk old female JH-/-m JCK-/-, HC2 line 2550 +, KCo4 line 4437 +; homozygous for disrupted mouse heavy and κ -light chain loci, hemizygous for HC2 and KCo4 transgenes). Top row: spleen cells stained for expression of mouse λ light chain (x-axis) and human κ light chain (y-axis). Second row: spleen cells stained for expression of human μ heavy chain (x-axis) and human κ light chain (y-axis). Third row: spleen cells stained for expression of mouse μ heavy chain (x-axis) and mouse κ light chain (y-axis). Bottom row: histogram of spleen cells stained for expression of mouse B220 antigen (log fluorescence: x-axis; cell number: y-axis). For each of the two color panels, the relative number of cells in each of the displayed quadrants is given as percent of a e-parameter gate based on propidium iodide staining and light scatter. The fraction of B220+ cells in each of the samples displayed in the bottom row is given as a percent of the lymphocyte light scatter gate.

Fig. 70. Secreted immunoglobulin levels in the serum of double transgenic mice. Human μ , γ , and κ , and mouse γ and λ from 18 individual HC2/KCo4 double transgenic mice homozygous for endogenous heavy and κ -light chain locus disruption. Mice: (+) HC2 line 2550 (~5 copies of HC2 per integration), KCo4 line 4436 (1-2 copies of KCo4 per integration); (O) HC2 line 2550, KCo4 line 4437 (~10 copies of KCo4 per integration); (x) HC2 line 2550, KCo4 line 4583 (~5

copies of KCo4 per integration); ($\square$) HC2 line 2572 (30-50 copies of HC2 per integration, KCo4 line 4437; (Δ) HC2 line 5467 (20-30 copies of HC2 per integration, KCo4 line 4437.

Figs. 71A and 71B show human antibody responses to human antigens. Fig. 71A: Primary response to recombinant human soluble CD4. Levels of human IgM and human κ light chain are reported for prebleed (O) and post-immunization ($\bullet$) serum from four double transgenic mice. Fig. 71B: Switching to human IgG occurs *in vivo*. Human IgG (circles) was detected with peroxidase conjugated polyclonal anti-human IgG used in the presence of 1.5 μ /ml excess IgE, κ and 1% normal mouse serum to inhibit non-specific cross-reactivity. Human κ light chain (squares) was detected using a peroxidase conjugated polyclonal anti-human κ reagent in the presence of 1% normal mouse serum. A representative result from one mouse (#9344; HC2 line 2550, KCo4 line 4436) is shown. Each point represents an average of duplicate wells minus background absorbance.

Fig. 72 shows FACS analysis of human PBL with a hybridoma supernatant that discriminates human CD4⁺ lymphocytes from human CD8⁺ lymphocytes.

Fig. 73 shows human α -CD4 IgM and IgG in transgenic mouse serum.

Fig. 74 shows competition binding experiments comparing a transgenic mouse α -human CD4 hybridoma monoclonal, 2C11-8, to the RPA-TA and Leu-3A monoclonals.

Fig. 75 shows production data for Ig expression of cultured 2C11-8 hybridoma.

Fig. 76 shows an overlapping set of plasmid inserts constituting the HCo7 transgene.

Fig. 77A depicts the nucleotide sequence and restriction map of pGP2b plasmid vector.

Fig. 77B depicts the restriction map of pGP2b plasmid vector.

Fig. 78 (parts A and B) depicts cloning strategy for assembling large transgenes.

5 Fig. 79 shows that large inserts are unstable in high-copy pUC derived plasmids.

Fig. 80 shows phage P1 clone P1-570. Insert spans portion of human heavy chain constant region covering $\gamma 3$ and $\gamma 1$, together with switch elements. N, NotI; S, SalI, X, XhoI.

10 Fig. 81 shows serum expression of human μ and $\gamma 1$ in HCo7 transgenic founder animals.

Fig. 82 shows serum expression of human immunoglobulins in HCo7/KCo4 double transgenic/double deletion mice.

15 Fig. 83 shows RT PCR detection of human $\gamma 1$ and $\gamma 3$ transcripts in HCo7 transgenic mouse spleen RNA.

Fig. 84 shows induction of human IgG1 and IgG3 by LPS and IL-4 in vitro.

20 Fig. 85. Agarose gel electrophoresis apparatus for concentration of YAC DNA.

Fig. 86. Two color FACS analysis of bone marrow cells from HC2/KCo5/JHD/JKD and HC2/KCo4/JHD/JKD mice. The fraction of cells in each of the B220⁺/CD43⁻, B220⁺/CD43⁺, and B220⁺/IgM⁺ gates is given as a percent.

25 Fig. 87. Two color FACS analysis of spleen cells from HC2/KCo5/JHD/JKD and HC2/KCo4/JHD/JKD mice. The fraction of cells in each of the B220^{bright}/IgM⁺ and B220^{dull}/IgM⁺ gates is given as a percent.

30 Fig. 88. Binding of IgGk anti-nCD4 monoclonal antibodies to CD4⁺ SupT1 cells.

Fig. 89 Epitope determination for IgG anti-nCD4 monoclonal antibodies by flow cytometry. SupT1 cells were pre-incubated with buffer

(left column), 2.5 mg/ml RPA-T4 (middle column), or 2.5 mg/ml Leu3a (right column) and then with one of the 10 human IgG monoclonal antibodies (in supernatant diluted 1:2), or chimeric Leu3a. Results for 3 representative human IgG monoclonal antibodies are shown in this figure.

5 Fig. 90 Inhibition of an MLR by a human IgGk anti-CD4 monoclonal antibody.

Figure 91 shows the effect of huMAb administration on CD4⁺ cells.

Fig. 92 shows the effect of huMAb administration on CD4⁺ Cells.

10 Fig. 93 shows the effect of huMAb administration on cynomolgus monkey cells.

Figure 94 shows the effect of huMAb administration on lymph node lymphocytes.

Fig. 95 shows serum half-life of huMAbs in cynomolgus monkeys.

15 The data derived from 1E11 and 6G5 were fit to a two compartment model, whereas the data derived from 1G2 were fit to a one compartment model.

Fig. 96 shows inhibition by human anti-CD4 mAbs of human cell responses to tetanus toxoid (TT). Panels (a) and (b) present results for two different assays. Lot numbers of the mAbs are shown in parantheses.

20 Fig. 97 shows human IgM and IgG anti-IL8 serum titers in transgenic mice. Responses of the individual mice to the immunogen were assessed by ELISA, converted to titers (1 = OD >0.1 at 1:50 dilution of serum; 2 = OD >0.1 at 1:250 dilution of serum; 3 = OD >0.1 at 1:1250 dilution of serum; 4 = OD >0.1 at 1:6250 dilution of serum; and 5 = OD >0.1 at 1:31250 dilution of serum) and then averaged.

Fig. 98 shows the effect of human anti-IL8 mAb on IL8-induced neutrophil chemotaxis and elastase release.

Table 1 depicts the sequence of vector pGPe.

Table 2 depicts the sequence of gene V_H49.8.

30 Table 3 depicts the detection of human IgM and IgG in the serum of transgenic mice of this invention.

Table 4 depicts sequences of VDJ joints.

Table 5 depicts the distribution of J segments incorporated into pHCl transgene encoded transcripts to J segments found in adult human peripheral blood lymphocytes (PBL).

35 Table 6 depicts the distribution of D segments incorporated into pHCl transgene encoded transcripts to D segments found in adult human peripheral blood lymphocytes (PBL).

Table 7 depicts the length of the CDR3 peptides from transcripts with in-frame VDJ joints in the pHCl transgenic mouse and in human PBL.

Table 8 depicts the predicted amino acid sequences of the VDJ regions from 30 clones analyzed from a pHCl transgenic.

Table 9 shows transgenic mice of line 112 that were used in the indicated experiments; (+) indicates the presence of the respective transgene, (++) indicates that the animal is homozygous for the J_HD knockout transgene.

Table 10 shows the genotypes of several 0011 mice.

Table 11 shows human variable region usage in hybridomas from transgenic mice.

Table 12 shows transgene V and J segment usage.

Table 13 shows the occurrence of somatic mutation in the HC2 heavy chain transgene in transgenic mice.

Table 14 shows identification of human V_K segments on the YAC 4x17E1.

Table 15. Identification of human vk genes expressed in mouse line KCo5-9272.

Table 16. Secretion levels for human IgGk Anti-nCD4 monoclonal antibodies

Table 17. Rate and affinity constants for monoclonal antibodies that bind to human CD4.

Table 18. Affinity and rate constants of human anti-human CD4 monoclonal antibodies.

Table 19. Avidity and rate constants of human anti-human CD4 monoclonal antibodies.

Table 20. Avidity and rate constants reported for anti CD4 monoclonal antibodies.

Table 21. Avidity constants of human anti-human CD4 monoclonal antibodies as determined by flow cytometry.

Table 22. Partial Nucleotide Sequence for Functional Transcripts.

Table 23 Germline V(D)J Segment Usage in Hybridoma Transcripts.

Table 24. Primers, Vectors and Products Used in Minigene Construction.

Table 25. Effect of Human mAbs on Peripheral Chimpanzee Lymphocytes.

5 Table 26. Summary of Flow Cytometry Studies on Lymph Node Lymphocytes.

Table 27. Monoclonal Antibody Secretion, Avidity and Rate Constants.

10 Table 28. Specificity and Characterization of Human Anti-IL8 mAbs.

DETAILED DESCRIPTION

As has been discussed supra, it is desirable to produce human immunoglobulins that are reactive with specific human antigens that are promising therapeutic and/or diagnostic targets. However, producing human immunoglobulins that bind specifically with human antigens is problematic.

15 First, the immunized animal that serves as the source of B cells must make an immune response against the presented antigen. In order for an animal to make an immune response, the antigen presented must be foreign and the animal must not be tolerant to the antigen. Thus, for example, if it is desired to produce a human monoclonal antibody with an idotype that binds to a human protein, self-tolerance will prevent an immunized human from making a substantial immune response to the human protein, since the only epitopes of the antigen that may be immunogenic will be those that result from polymorphism of the protein within the human population (allogeneic epitopes).

30 Second, if the animal that serves as the source of B-cells for forming a hybridoma (a human in the illustrative given example) does make an immune response against an

WO 98/24884

PCT/US97/21803

30

authentic self antigen, a severe autoimmune disease may result in the animal. Where humans would be used as a source of B-cells for a hybridoma, such autoimmunization would be considered unethical by contemporary standards. Thus, developing hybridomas secreting human immunoglobulin chainsspecifically reactive with predetermined human antigens is problematic, since a reliable source of human antibody-secreting B cells that can evoke an antibody response against predetermined human antigens is needed.

One methodology that can be used to obtain human antibodies that are specifically reactive with human antigens is the production of a transgenic mouse harboring the human immunoglobulin transgene constructs of this invention. Briefly, transgenes containing all or portions of the human immunoglobulin heavy and light chain loci, or transgenes containing synthetic "miniloci" (described *infra*, and in copending applications U.S.S.N. 08/352,322, filed 7 December 1994, U.S.S.N. 07/990,860, filed 16 December 1992, U.S.S.N. 07/810,279 filed 17 December 1991, U.S.S.N. 07/904,068 filed 23 June 1992; U.S.S.N. 07/853,408, filed 18 March 1992, U.S.S.N. 07/574,748 filed August 29, 1990, U.S.S.N. 07/575,962 filed August 31, 1990, and PCT/US91/06185 filed August 28, 1991, each incorporated herein by reference) which comprise essential functional elements of the human heavy and light chain loci, are employed to produce a transgenic nonhuman animal. Such a transgenic nonhuman animal will have the capacity to produce immunoglobulin chains that are encoded by human immunoglobulin genes, and additionally will be capable of making an immune response against human antigens. Thus, such transgenic animals can serve as a source of immune sera reactive with specified human antigens, and B-cells from such transgenic animals can be fused with myeloma cells to produce

hybridomas that secrete monoclonal antibodies that are encoded by human immunoglobulin genes and which are specifically reactive with human antigens.

The production of transgenic mice containing various forms of immunoglobulin genes has been reported previously. Rearranged mouse immunoglobulin heavy or light chain genes have been used to produce transgenic mice. In addition, functionally rearranged human Ig genes including the μ or $\gamma 1$ constant region have been expressed in transgenic mice. However, experiments in which the transgene comprises unrearranged (V-D-J or V-J not rearranged) immunoglobulin genes have been variable, in some cases, producing incomplete or minimal rearrangement of the transgene. However, there are no published examples of either rearranged or unrearranged immunoglobulin transgenes which undergo successful isotype switching between C_H genes within a transgene.

The invention also provides a method for identifying candidate hybridomas which secrete a monoclonal antibody comprising a human immunoglobulin chain consisting essentially of a human VDJ sequence in polypeptide linkage to a human constant region sequence. Such candidate hybridomas are identified from a pool of hybridoma clones comprising: (1) hybridoma clones that express immunoglobulin chains consisting essentially of a human VDJ region and a human constant region, and (2) trans-switched hybridomas that express heterohybrid immunoglobulin chains consisting essentially of a human VDJ region and a murine constant region. The supernatant(s) of individual or pooled hybridoma clones is contacted with a predetermined antigen, typically an antigen which is immobilized by adsorption onto a solid substrate (e.g., a microtitre well), under binding conditions to select antibodies having the predetermined antigen binding

specificity. An antibody that specifically binds to human constant regions is also contacted with the hybridoma supernatant and predetermined antigen under binding conditions so that the antibody selectively binds to at least one human constant region epitope but substantially does not bind to murine constant region epitopes; thus forming complexes consisting essentially of hybridoma supernatant (transgenic monoclonal antibody) bound to a predetermined antigen and to an antibody that specifically binds human constant regions (and which may be labeled with a detectable label or reporter). Detection of the formation of such complexes indicates hybridoma clones or pools which express a human immunoglobulin chain.

In a preferred embodiment of the invention, the anti-human constant region immunoglobulin used in screening specifically recognizes a non- μ , non- δ isotype, preferably a α or ϵ , more preferably a γ isotype constant region. Monoclonal antibodies of the γ isotype are preferred (i) because the characteristics of IgG immunoglobulins are preferable to IgM immunoglobulins for some therapeutic applications (e.g., due to the smaller size of the IgG dimers compared to IgM pentamers) and, (ii) because the process of somatic mutation is correlated with the class switch from the μ constant region to the non- μ (e.g., γ) constant regions. Immunoglobulins selected from the population of immunoglobulins that have undergone class switch (e.g., IgG) tend to bind antigen with higher affinity than immunoglobulins selected from the population that has not undergone class switch (e.g., IgM). See, e.g., Lonberg and Huszar. Intern. Rev. Immunol. 13:65-93 (1995) which is incorporated herein by reference.

In one embodiment the candidate hybridomas are first screened for the γ isotype constant region and the pool of IgG-expressing hybridomas is then screened for specific binding to the predetermined antigen.

5 Thus, according to the method, a transgenic mouse of the invention is immunized with the predetermined antigen to induce an immune response. B cells are collected from the mouse and fused to immortal cells to produce hybridomas. The hybridomas are first screened to identify individual
10 hybridomas secreting Ig of a non-mu, non-delta isotype (e.g., IgG). This set of hybridomas is then screened for specific binding to the predetermined antigen of interest. Screening is carried out using standard techniques as described in, e.g., Harlow and Lane, *Antibodies: A Laboratory Manual*, Cold
15 Spring Harbor, New York (1988). Using this method it is possible to identify high-affinity immunoglobulins (e.g., K_a greater than about 10^7 M^{-1}) practically and efficiently.

Definitions

20 As used herein, the term "antibody" refers to a glycoprotein comprising at least two light polypeptide chains and two heavy polypeptide chains. Each of the heavy and light polypeptide chains contains a variable region (generally the amino terminal portion of the polypeptide chain) which
25 contains a binding domain which interacts with antigen. Each of the heavy and light polypeptide chains also comprises a constant region of the polypeptide chains (generally the carboxyl terminal portion) which may mediate the binding of the immunoglobulin to host tissues or factors including
30 various cells of the immune system, some phagocytic cells and the first component (C1q) of the classical complement system.

As used herein, a "heterologous antibody" is defined in relation to the transgenic non-human organism producing such an antibody. It is defined as an antibody having an amino acid sequence or an encoding DNA sequence corresponding
5 to that found in an organism not consisting of the transgenic non-human animal, and generally from a species other than that of the transgenic non-human animal.

As used herein, a "heterohybrid antibody" refers to an antibody having a light and heavy chains of different
10 organismal origins. For example, an antibody having a human heavy chain associated with a murine light chain is a heterohybrid antibody.

As used herein, "isotype" refers to the antibody class (e.g., IgM or IgG₁) that is encoded by heavy chain
15 constant region genes.

As used herein, "isotype switching" refers to the phenomenon by which the class, or isotype, of an antibody changes from one Ig class to one of the other Ig classes.

As used herein, "nonswitched isotype" refers to the
20 isotypic class of heavy chain that is produced when no isotype switching has taken place; the C_H gene encoding the nonswitched isotype is typically the first C_H gene immediately downstream from the functionally rearranged VDJ gene.

As used herein, the term "switch sequence" refers to
25 those DNA sequences responsible for switch recombination. A "switch donor" sequence, typically a μ switch region, will be 5' (i.e., upstream) of the construct region to be deleted during the switch recombination. The "switch acceptor" region will be between the construct region to be deleted and the
30 replacement constant region (e.g., γ , ϵ , etc.). As there is no specific site where recombination always occurs, the final

gene sequence will typically not be predictable from the construct.

As used herein, "glycosylation pattern" is defined as the pattern of carbohydrate units that are covalently attached to a protein, more specifically to an immunoglobulin protein. A glycosylation pattern of a heterologous antibody can be characterized as being substantially similar to glycosylation patterns which occur naturally on antibodies produced by the species of the nonhuman transgenic animal, when one of ordinary skill in the art would recognize the glycosylation pattern of the heterologous antibody as being more similar to said pattern of glycosylation in the species of the nonhuman transgenic animal than to the species from which the C_H genes of the transgene were derived.

As used herein, "specific binding" refers to the property of the antibody: (1) to bind to a predetermined antigen with an affinity of at least $1 \times 10^7 \text{ M}^{-1}$, and (2) to preferentially bind to the predetermined antigen with an affinity that is at least two-fold greater than its affinity for binding to a non-specific antigen (e.g., BSA, casein) other than the predetermined antigen or a closely-related antigen.

The term "naturally-occurring" as used herein as applied to an object refers to the fact that an object can be found in nature. For example, a polypeptide or polynucleotide sequence that is present in an organism (including viruses) that can be isolated from a source in nature and which has not been intentionally modified by man in the laboratory is naturally-occurring.

The term "rearranged" as used herein refers to a configuration of a heavy chain or light chain immunoglobulin locus wherein a V segment is positioned immediately adjacent

to a D-J or J segment in a conformation encoding essentially a complete V_H or V_L domain, respectively. A rearranged immunoglobulin gene locus can be identified by comparison to germline DNA; a rearranged locus will have at least one
5 recombined heptamer/nonamer homology element.

The term "unrearranged" or "germline configuration" as used herein in reference to a V segment refers to the configuration wherein the V segment is not recombined so as to be immediately adjacent to a D or J segment.

10 For nucleic acids, the term "substantial homology" indicates that two nucleic acids, or designated sequences thereof, when optimally aligned and compared, are identical, with appropriate nucleotide insertions or deletions, in at least about 80% of the nucleotides, usually at least about 90%
15 to 95%, and more preferably at least about 98 to 99.5% of the nucleotides. Alternatively, substantial homology exists when the segments will hybridize under selective hybridization conditions, to the complement of the strand. The nucleic acids may be present in whole cells, in a cell lysate, or in a
20 partially purified or substantially pure form. A nucleic acid is "isolated" or "rendered substantially pure" when purified away from other cellular components or other contaminants, e.g., other cellular nucleic acids or proteins, by standard techniques, including alkaline/SDS treatment, CsCl banding,
25 column chromatography, agarose gel electrophoresis and others well known in the art. See, F. Ausubel, et al., ed. Current Protocols in Molecular Biology, Greene Publishing and Wiley-Interscience, New York (1987).

The nucleic acid compositions of the present
30 invention, while often in a native sequence (except for modified restriction sites and the like), from either cDNA, genomic or mixtures may be mutated, thereof in accordance with

standard techniques to provide gene sequences. For coding sequences, these mutations, may affect amino acid sequence as desired. In particular, DNA sequences substantially homologous to or derived from native V, D, J, constant, switches and other such sequences described herein are contemplated (where "derived" indicates that a sequence is identical or modified from another sequence).

A nucleic acid is "operably linked" when it is placed into a functional relationship with another nucleic acid sequence. For instance, a promoter or enhancer is operably linked to a coding sequence if it affects the transcription of the sequence. With respect to transcription regulatory sequences, operably linked means that the DNA sequences being linked are contiguous and, where necessary to join two protein coding regions, contiguous and in reading frame. For switch sequences, operably linked indicates that the sequences are capable of effecting switch recombination.

Transgenic Nonhuman Animals Capable of Producing Heterologous Antibodies

The design of a transgenic non-human animal that responds to foreign antigen stimulation with a heterologous antibody repertoire, requires that the heterologous immunoglobulin transgenes contained within the transgenic animal function correctly throughout the pathway of B-cell development. In a preferred embodiment, correct function of a heterologous heavy chain transgene includes isotype switching. Accordingly, the transgenes of the invention are constructed so as to produce isotype switching and one or more of the following: (1) high level and cell-type specific expression, (2) functional gene rearrangement, (3) activation of and response to allelic exclusion, (4) expression of a sufficient

primary repertoire, (5) signal transduction, (6) somatic hypermutation, and (7) domination of the transgene antibody locus during the immune response.

As will be apparent from the following disclosure,
5 not all of the foregoing criteria need be met. For example, in those embodiments wherein the endogenous immunoglobulin loci of the transgenic animal are functionally disrupted, the transgene need not activate allelic exclusion. Further, in those embodiments wherein the transgene comprises a
10 functionally rearranged heavy and/or light chain immunoglobulin gene, the second criteria of functional gene rearrangement is unnecessary, at least for that transgene which is already rearranged. For background on molecular immunology, see, Fundamental Immunology, 2nd edition (1989),
15 Paul William E., ed. Raven Press, N.Y., which is incorporated herein by reference.

In one aspect of the invention, transgenic non-human animals are provided that contain rearranged, unrearranged or a combination of rearranged and unrearranged heterologous
20 immunoglobulin heavy and light chain transgenes in the germline of the transgenic animal. Each of the heavy chain transgenes comprises at least one C_H gene. In addition, the heavy chain transgene may contain functional isotype switch sequences, which are capable of supporting isotype switching
25 of a heterologous transgene encoding multiple C_H genes in B-cells of the transgenic animal. Such switch sequences may be those which occur naturally in the germline immunoglobulin locus from the species that serves as the source of the transgene C_H genes, or such switch sequences may be derived
30 from those which occur in the species that is to receive the transgene construct (the transgenic animal). For example, a human transgene construct that is used to produce a transgenic

mouse may produce a higher frequency of isotype switching events if it incorporates switch sequences similar to those that occur naturally in the mouse heavy chain locus, as presumably the mouse switch sequences are optimized to function with the mouse switch recombinase enzyme system, whereas the human switch sequences are not. Switch sequences made be isolated and cloned by conventional cloning methods, or may be synthesized de novo from overlapping synthetic oligonucleotides designed on the basis of published sequence information relating to immunoglobulin switch region sequences (Mills et al., Nucl. Acids Res. 18:7305-7316 (1991); Sideras et al., Intl. Immunol. 1:631-642 (1989), which are incorporated herein by reference).

For each of the foregoing transgenic animals, functionally rearranged heterologous heavy and light chain immunoglobulin transgenes are found in a significant fraction of the B-cells of the transgenic animal (at least 10 percent).

The transgenes of the invention include a heavy chain transgene comprising DNA encoding at least one variable gene segment, one diversity gene segment, one joining gene segment and at least one constant region gene segment. The immunoglobulin light chain transgene comprises DNA encoding at least one variable gene segment, one joining gene segment and at least one constant region gene segment. The gene segments encoding the light and heavy chain gene segments are heterologous to the transgenic non-human animal in that they are derived from, or correspond to, DNA encoding immunoglobulin heavy and light chain gene segments from a species not consisting of the transgenic non-human animal. In one aspect of the invention, the transgene is constructed such that the individual gene segments are unrearranged, i.e., not rearranged so as to encode a functional immunoglobulin light

or heavy chain. Such unrearranged transgenes support recombination of the V, D, and J gene segments (functional rearrangement) and preferably support incorporation of all or a portion of a D region gene segment in the resultant
5 rearranged immunoglobulin heavy chain within the transgenic non-human animal when exposed to antigen.

In an alternate embodiment, the transgenes comprise an unrearranged "mini-locus". Such transgenes typically comprise a substantial portion of the C, D, and J segments as
10 well as a subset of the V gene segments. In such transgene constructs, the various regulatory sequences, e.g. promoters, enhancers, class switch regions, splice-donor and splice-acceptor sequences for RNA processing, recombination signals and the like, comprise corresponding sequences derived from
15 the heterologous DNA. Such regulatory sequences may be incorporated into the transgene from the same or a related species of the non-human animal used in the invention. For example, human immunoglobulin gene segments may be combined in a transgene with a rodent immunoglobulin enhancer sequence for
20 use in a transgenic mouse. Alternatively, synthetic regulatory sequences may be incorporated into the transgene, wherein such synthetic regulatory sequences are not homologous to a functional DNA sequence that is known to occur naturally in the genomes of mammals. Synthetic regulatory sequences are
25 designed according to consensus rules, such as, for example, those specifying the permissible sequences of a splice-acceptor site or a promoter/enhancer motif. For example, a minilocus comprises a portion of the genomic immunoglobulin locus having at least one internal (i.e., not at a terminus of
30 the portion) deletion of a non-essential DNA portion (e.g., intervening sequence; intron or portion thereof) as compared to the naturally-occurring germline Ig locus.

The invention also includes transgenic animals containing germ line cells having a heavy and light transgene wherein one of the said transgenes contains rearranged gene segments with the other containing unrearranged gene segments.

- 5 In the preferred embodiments, the rearranged transgene is a light chain immunoglobulin transgene and the unrearranged transgene is a heavy chain immunoglobulin transgene.

The Structure and Generation of Antibodies

- 10 The basic structure of all immunoglobulins is based upon a unit consisting of two light polypeptide chains and two heavy polypeptide chains. Each light chain comprises two regions known as the variable light chain region and the constant light chain region. Similarly, the immunoglobulin
15 heavy chain comprises two regions designated the variable heavy chain region and the constant heavy chain region.

- The constant region for the heavy or light chain is encoded by genomic sequences referred to as heavy or light constant region gene (C_H) segments. The use of a particular
20 heavy chain gene segment defines the class of immunoglobulin. For example, in humans, the μ constant region gene segments define the IgM class of antibody whereas the use of a γ , γ_2 , γ_3 or γ_4 constant region gene segment defines the IgG class of antibodies as well as the IgG subclasses IgG1 through IgG4.
25 Similarly, the use of a α_1 or α_2 constant region gene segment defines the IgA class of antibodies as well as the subclasses IgA1 and IgA2. The δ and ϵ constant region gene segments define the IgD and IgE antibody classes, respectively.

- The variable regions of the heavy and light
30 immunoglobulin chains together contain the antigen binding domain of the antibody. Because of the need for diversity in this region of the antibody to permit binding to a wide range

of antigens, the DNA encoding the initial or primary repertoire variable region comprises a number of different DNA segments derived from families of specific variable region gene segments. In the case of the light chain variable region, such families comprise variable (V) gene segments and joining (J) gene segments. Thus, the initial variable region of the light chain is encoded by one V gene segment and one J gene segment each selected from the family of V and J gene segments contained in the genomic DNA of the organism. In the case of the heavy chain variable region, the DNA encoding the initial or primary repertoire variable region of the heavy chain comprises one heavy chain V gene segment, one heavy chain diversity (D) gene segment and one J gene segment, each selected from the appropriate V, D and J families of immunoglobulin gene segments in genomic DNA.

In order to increase the diversity of sequences that contribute to forming antibody binding sites, it is preferable that a heavy chain transgene include cis-acting sequences that support functional V-D-J rearrangement that can incorporate all or part of a D region gene sequence in a rearranged V-D-J gene sequence. Typically, at least about 1 percent of expressed transgene-encoded heavy chains (or mRNAs) include recognizable D region sequences in the V region. Preferably, at least about 10 percent of transgene-encoded V regions include recognizable D region sequences, more preferably at least about 30 percent, and most preferably more than 50 percent include recognizable D region sequences.

A recognizable D region sequence is generally at least about eight consecutive nucleotides corresponding to a sequence present in a D region gene segment of a heavy chain transgene and/or the amino acid sequence encoded by such D region nucleotide sequence. For example, if a transgene

includes the D region gene DHQ52, a transgene-encoded mRNA containing the sequence 5'-TAACTGGG-3' located in the V region between a V gene segment sequence and a J gene segment sequence is recognizable as containing a D region sequence, specifically a DHQ52 sequence. Similarly, for example, if a transgene includes the D region gene DHQ52, a transgene-encoded heavy chain polypeptide containing the amino acid sequence -DAF- located in the V region between a V gene segment amino acid sequence and a J gene segment amino acid sequence may be recognizable as containing a D region sequence, specifically a DHQ52 sequence. However, since D region segments may be incorporated in VDJ joining to various extents and in various reading frames, a comparison of the D region area of a heavy chain variable region to the D region segments present in the transgene is necessary to determine the incorporation of particular D segments. Moreover, potential exonuclease digestion during recombination may lead to imprecise V-D and D-J joints during V-D-J recombination.

However, because of somatic mutation and N-region addition, some D region sequences may be recognizable but may not correspond identically to a consecutive D region sequence in the transgene. For example, a nucleotide sequence 5'-CTAAXTGGGG-3', where X is A, T, or G, and which is located in a heavy chain V region and flanked by a V region gene sequence and a J region gene sequence, can be recognized as corresponding to the DHQ52 sequence 5'-CTAACTGGG-3'. Similarly, for example, the polypeptide sequences -DAFDI-, -DYFDY-, or -GAFDI- located in a V region and flanked on the amino-terminal side by an amino acid sequence encoded by a transgene V gene sequence and flanked on the carboxyterminal side by an amino acid sequence encoded by a transgene J gene sequence is recognizable as a D region sequence.

Therefore, because somatic mutation and N-region addition can produce mutations in sequences derived from a transgene D region, the following definition is provided as a guide for determining the presence of a recognizable D region sequence. An amino acid sequence or nucleotide sequence is recognizable as a D region sequence if: (1) the sequence is located in a V region and is flanked on one side by a V gene sequence (nucleotide sequence or deduced amino acid sequence) and on the other side by a J gene sequence (nucleotide sequence or deduced amino acid sequence) and (2) the sequence is substantially identical or substantially similar to a known D gene sequence (nucleotide sequence or encoded amino acid sequence).

The term "substantial identity" as used herein denotes a characteristic of a polypeptide sequence or nucleic acid sequence, wherein the polypeptide sequence has at least 50 percent sequence identity compared to a reference sequence, often at least about 80% sequence identity and sometimes more than about 90% sequence identity, and the nucleic acid sequence has at least 70 percent sequence identity compared to a reference sequence. The percentage of sequence identity is calculated excluding small deletions or additions which total less than 35 percent of the reference sequence. The reference sequence may be a subset of a larger sequence, such as an entire D gene; however, the reference sequence is at least 8 nucleotides long in the case of polynucleotides, and at least 3 amino residues long in the case of a polypeptide. Typically, the reference sequence is at least 8 to 12 nucleotides or at least 3 to 4 amino acids, and preferably the reference sequence is 12 to 15 nucleotides or more, or at least 5 amino acids.

The term "substantial similarity" denotes a characteristic of an polypeptide sequence, wherein the polypeptide sequence has at least 80 percent similarity to a reference sequence. The percentage of sequence similarity is
5 calculated by scoring identical amino acids or positional conservative amino acid substitutions as similar. A positional conservative amino acid substitution is one that can result from a single nucleotide substitution; a first amino acid is replaced by a second amino acid where a codon
10 for the first amino acid and a codon for the second amino acid can differ by a single nucleotide substitution. Thus, for example, the sequence -Lys-Glu-Arg-Val- is substantially similar to the sequence -Asn-Asp-Ser-Val-, since the codon sequence -AAA-GAA-AGA-GUU- can be mutated to -AAC-GAC-AGC-GUU-
15 by introducing only 3 substitution mutations, single nucleotide substitutions in three of the four original codons. The reference sequence may be a subset of a larger sequence, such as an entire D gene; however, the reference sequence is at least 4 amino residues long. Typically, the reference
20 sequence is at least 5 amino acids, and preferably the reference sequence is 6 amino acids or more.

The Primary Repertoire

The process for generating DNA encoding the heavy
25 and light chain immunoglobulin genes occurs primarily in developing B-cells. Prior to the joining of various immunoglobulin gene segments, the V, D, J and constant (C) gene segments are found, for the most part, in clusters of V, D, J and C gene segments in the precursors of primary
30 repertoire B-cells. Generally, all of the gene segments for a heavy or light chain are located in relatively close proximity on a single chromosome. Such genomic DNA prior to

recombination of the various immunoglobulin gene segments is referred to herein as "unrearranged" genomic DNA. During B-cell differentiation, one of each of the appropriate family members of the V, D, J (or only V and J in the case of light chain genes) gene segments are recombined to form functionally rearranged heavy and light immunoglobulin genes. Such functional rearrangement is of the variable region segments to form DNA encoding a functional variable region. This gene segment rearrangement process appears to be sequential.

10 First, heavy chain D-to-J joints are made, followed by heavy chain V-to-DJ joints and light chain V-to-J joints. The DNA encoding this initial form of a functional variable region in a light and/or heavy chain is referred to as "functionally rearranged DNA" or "rearranged DNA". In the case of the heavy

15 chain, such DNA is referred to as "rearranged heavy chain DNA" and in the case of the light chain, such DNA is referred to as "rearranged light chain DNA". Similar language is used to describe the functional rearrangement of the transgenes of the invention.

20 The recombination of variable region gene segments to form functional heavy and light chain variable regions is mediated by recombination signal sequences (RSS's) that flank recombinationally competent V, D and J segments. RSS's necessary and sufficient to direct recombination, comprise a

25 dyad-symmetric heptamer, an AT-rich nonamer and an intervening spacer region of either 12 or 23 base pairs. These signals are conserved among the different loci and species that carry out D-J (or V-J) recombination and are functionally interchangeable. See Oettinger, et al. (1990), Science, 248,

30 1517-1523 and references cited therein. The heptamer comprises the sequence CACAGTG or its analogue followed by a spacer of unconserved sequence and then a nonamer having the

sequence ACAAAAACC or its analogue. These sequences are found on the J, or downstream side, of each V and D gene segment. Immediately preceding the germline D and J segments are again two recombination signal sequences, first the nonamer and then
5 the heptamer again separated by an unconserved sequence. The heptameric and nonameric sequences following a V_L , V_H or D segment are complementary to those preceding the J_L , D or J_H segments with which they recombine. The spacers between the heptameric and nonameric sequences are either 12 base pairs
10 long or between 22 and 24 base pairs long.

In addition to the rearrangement of V, D and J segments, further diversity is generated in the primary repertoire of immunoglobulin heavy and light chain by way of variable recombination between the V and J segments in the
15 light chain and between the D and J segments of the heavy chain. Such variable recombination is generated by variation in the exact place at which such segments are joined. Such variation in the light chain typically occurs within the last codon of the V gene segment and the first codon of the J
20 segment. Similar imprecision in joining occurs on the heavy chain chromosome between the D and J_H segments and may extend over as many as 10 nucleotides. Furthermore, several nucleotides may be inserted between the D and J_H and between the V_H and D gene segments which are not encoded by genomic
25 DNA. The addition of these nucleotides is known as N-region diversity.

After VJ and/or VDJ rearrangement, transcription of the rearranged variable region and one or more constant region gene segments located downstream from the rearranged variable
30 region produces a primary RNA transcript which upon appropriate RNA splicing results in an mRNA which encodes a full length heavy or light immunoglobulin chain. Such heavy

and light chains include a leader signal sequence to effect secretion through and/or insertion of the immunoglobulin into the transmembrane region of the B-cell. The DNA encoding this signal sequence is contained within the first exon of the V segment used to form the variable region of the heavy or light immunoglobulin chain. Appropriate regulatory sequences are also present in the mRNA to control translation of the mRNA to produce the encoded heavy and light immunoglobulin polypeptides which upon proper association with each other form an antibody molecule.

The net effect of such rearrangements in the variable region gene segments and the variable recombination which may occur during such joining, is the production of a primary antibody repertoire. Generally, each B-cell which has differentiated to this stage, produces a single primary repertoire antibody. During this differentiation process, cellular events occur which suppress the functional rearrangement of gene segments other than those contained within the functionally rearranged Ig gene. The process by which diploid B-cells maintain such mono-specificity is termed allelic exclusion.

The Secondary Repertoire

B-cell clones expressing immunoglobulins from within the set of sequences comprising the primary repertoire are immediately available to respond to foreign antigens. Because of the limited diversity generated by simple VJ and VDJ joining, the antibodies produced by the so-called primary response are of relatively low affinity. Two different types of B-cells make up this initial response: precursors of primary antibody-forming cells and precursors of secondary repertoire B-cells (Linton et al., Cell 59:1049-1059 (1989)).

The first type of B-cell matures into IgM-secreting plasma cells in response to certain antigens. The other B-cells respond to initial exposure to antigen by entering a T-cell dependent maturation pathway.

5 During the T-cell dependent maturation of antigen stimulated B-cell clones, the structure of the antibody molecule on the cell surface changes in two ways: the constant region switches to a non-IgM subtype and the sequence of the variable region can be modified by multiple single amino acid
10 substitutions to produce a higher affinity antibody molecule.

 As previously indicated, each variable region of a heavy or light Ig chain contains an antigen binding domain. It has been determined by amino acid and nucleic acid sequencing that somatic mutation during the secondary response
15 occurs throughout the V region including the three complementary determining regions (CDR1, CDR2 and CDR3) also referred to as hypervariable regions 1, 2 and 3 (Kabat et al. Sequences of Proteins of Immunological Interest (1991) U.S. Department of Health and Human Services, Washington, DC,
20 incorporated herein by reference. The CDR1 and CDR2 are located within the variable gene segment whereas the CDR3 is largely the result of recombination between V and J gene segments or V, D and J gene segments. Those portions of the variable region which do not consist of CDR1, 2 or 3 are
25 commonly referred to as framework regions designated FR1, FR2, FR3 and FR4. See Fig. 1. During hypermutation, the rearranged DNA is mutated to give rise to new clones with altered Ig molecules. Those clones with higher affinities for the foreign antigen are selectively expanded by helper
30 T-cells, giving rise to affinity maturation of the expressed antibody. Clonal selection typically results in expression of clones containing new mutation within the CDR1, 2 and/or 3

regions. However, mutations outside these regions also occur which influence the specificity and affinity of the antigen binding domain.

5 Transgenic Non-Human Animals Capable
of Producing Heterologous Antibody

Transgenic non-human animals in one aspect of the invention are produced by introducing at least one of the
10 immunoglobulin transgenes of the invention (discussed hereinafter) into a zygote or early embryo of a non-human animal. The non-human animals which are used in the invention generally comprise any mammal which is capable of rearranging immunoglobulin gene segments to produce a primary antibody
15 response. Such nonhuman transgenic animals may include, for example, transgenic pigs, transgenic rats, transgenic rabbits, transgenic cattle, and other transgenic animal species, particularly mammalian species, known in the art. A particularly preferred non-human animal is the mouse or other
20 members of the rodent family.

However, the invention is not limited to the use of mice. Rather, any non-human mammal which is capable of mounting a primary and secondary antibody response may be used. Such animals include non-human primates, such as
25 chimpanzee, bovine, ovine, and porcine species, other members of the rodent family, e.g. rat, as well as rabbit and guinea pig. Particular preferred animals are mouse, rat, rabbit and guinea pig, most preferably mouse.

In one embodiment of the invention, various gene
30 segments from the human genome are used in heavy and light chain transgenes in an unrearranged form. In this embodiment, such transgenes are introduced into mice. The unrearranged gene segments of the light and/or heavy chain transgene have

DNA sequences unique to the human species which are distinguishable from the endogenous immunoglobulin gene segments in the mouse genome. They may be readily detected in unarranged form in the germ line and somatic cells not
5 consisting of B-cells and in rearranged form in B-cells.

In an alternate embodiment of the invention, the transgenes comprise rearranged heavy and/or light immunoglobulin transgenes. Specific segments of such transgenes corresponding to functionally rearranged VDJ or VJ
10 segments, contain immunoglobulin DNA sequences which are also clearly distinguishable from the endogenous immunoglobulin gene segments in the mouse.

Such differences in DNA sequence are also reflected in the amino acid sequence encoded by such human
15 immunoglobulin transgenes as compared to those encoded by mouse B-cells. Thus, human immunoglobulin amino acid sequences may be detected in the transgenic non-human animals of the invention with antibodies specific for immunoglobulin epitopes encoded by human immunoglobulin gene segments.

20 Transgenic B-cells containing unarranged transgenes from human or other species functionally recombine the appropriate gene segments to form functionally rearranged light and heavy chain variable regions. It will be readily apparent that the antibody encoded by such rearranged
25 transgenes has a DNA and/or amino acid sequence which is heterologous to that normally encountered in the nonhuman animal used to practice the invention.

Unarranged Transgenes

30 As used herein, an "unarranged immunoglobulin heavy chain transgene" comprises DNA encoding at least one variable gene segment, one diversity gene segment, one joining

gene segment and one constant region gene segment. Each of the gene segments of said heavy chain transgene are derived from, or has a sequence corresponding to, DNA encoding immunoglobulin heavy chain gene segments from a species not
5 consisting of the non-human animal into which said transgene is introduced. Similarly, as used herein, an "unrearranged immunoglobulin light chain transgene" comprises DNA encoding at least one variable gene segment, one joining gene segment and at least one constant region gene segment wherein each
10 gene segment of said light chain transgene is derived from, or has a sequence corresponding to, DNA encoding immunoglobulin light chain gene segments from a species not consisting of the non-human animal into which said light chain transgene is introduced.

15 Such heavy and light chain transgenes in this aspect of the invention contain the above-identified gene segments in an unrearranged form. Thus, interposed between the V, D and J segments in the heavy chain transgene and between the V and J segments on the light chain transgene are appropriate
20 recombination signal sequences (RSS's). In addition, such transgenes also include appropriate RNA splicing signals to join a constant region gene segment with the VJ or VDJ rearranged variable region.

In order to facilitate isotype switching within a
25 heavy chain transgene containing more than one C region gene segment, e.g. C μ and C γ 1 from the human genome, as explained below "switch regions" are incorporated upstream from each of the constant region gene segments and downstream from the variable region gene segments to permit recombination between
30 such constant regions to allow for immunoglobulin class switching, e.g. from IgM to IgG. Such heavy and light immunoglobulin transgenes also contain transcription control

sequences including promoter regions situated upstream from the variable region gene segments which typically contain TATA motifs. A promoter region can be defined approximately as a DNA sequence that, when operably linked to a downstream

5 sequence, can produce transcription of the downstream sequence. Promoters may require the presence of additional linked cis-acting sequences in order to produce efficient transcription. In addition, other sequences that participate in the transcription of sterile transcripts are preferably

10 included. Examples of sequences that participate in expression of sterile transcripts can be found in the published literature, including Rothman et al., Intl. Immunol. 2:621-627 (1990); Reid et al., Proc. Natl. Acad. Sci. USA 86:840-844 (1989); Stavnezer et al., Proc. Natl. Acad. Sci. USA 85:7704-7708 (1988); and Mills et al., Nucl. Acids Res. 18:7305-7316 (1991), each of which is incorporated herein by

15 reference. These sequences typically include about at least 50 bp immediately upstream of a switch region, preferably about at least 200 bp upstream of a switch region; and more

20 preferably about at least 200-1000 bp or more upstream of a switch region. Suitable sequences occur immediately upstream of the human S_{V1} , S_{V2} , S_{V3} , S_{V4} , $S_{\alpha1}$, $S_{\alpha2}$, and S_e switch regions; the sequences immediately upstream of the human S_{V1} , and S_{V3} switch regions can be used to advantage, with S_{V1} generally

25 preferred. Alternatively, or in combination, murine Ig switch sequences may be used; it may frequently be advantageous to employ Ig switch sequences of the same species as the transgenic non-human animal. Furthermore, interferon (IFN) inducible transcriptional regulatory elements, such as IFN-

30 inducible enhancers, are preferably included immediately upstream of transgene switch sequences.

In addition to promoters, other regulatory sequences which function primarily in B-lineage cells are used. Thus, for example, a light chain enhancer sequence situated preferably between the J and constant region gene segments on the light chain transgene is used to enhance transgene expression, thereby facilitating allelic exclusion. In the case of the heavy chain transgene, regulatory enhancers and also employed. Such regulatory sequences are used to maximize the transcription and translation of the transgene so as to induce allelic exclusion and to provide relatively high levels of transgene expression.

Although the foregoing promoter and enhancer regulatory control sequences have been generically described, such regulatory sequences may be heterologous to the nonhuman animal being derived from the genomic DNA from which the heterologous transgene immunoglobulin gene segments are obtained. Alternately, such regulatory gene segments are derived from the corresponding regulatory sequences in the genome of the non-human animal, or closely related species, which contains the heavy and light transgene.

In the preferred embodiments, gene segments are derived from human beings. The transgenic non-human animals harboring such heavy and light transgenes are capable of mounting an Ig-mediated immune response to a specific antigen administered to such an animal. B-cells are produced within such an animal which are capable of producing heterologous human antibody. After immortalization, and the selection for an appropriate monoclonal antibody (Mab), e.g. a hybridoma, a source of therapeutic human monoclonal antibody is provided. Such human Mabs have significantly reduced immunogenicity when therapeutically administered to humans.

Although the preferred embodiments disclose the construction of heavy and light transgenes containing human gene segments, the invention is not so limited. In this regard, it is to be understood that the teachings described
5 herein may be readily adapted to utilize immunoglobulin gene segments from a species other than human beings. For example, in addition to the therapeutic treatment of humans with the antibodies of the invention, therapeutic antibodies encoded by appropriate gene segments may be utilized to generate
10 monoclonal antibodies for use in the veterinary sciences.

Rearranged Transgenes

In an alternative embodiment, transgenic nonhuman animals contain functionally at least one rearranged
15 heterologous heavy chain immunoglobulin transgene in the germline of the transgenic animal. Such animals contain primary repertoire B-cells that express such rearranged heavy transgenes. Such B-cells preferably are capable of undergoing somatic mutation when contacted with an antigen to form a
20 heterologous antibody having high affinity and specificity for the antigen. Said rearranged transgenes will contain at least two C_H genes and the associated sequences required for isotype switching.

The invention also includes transgenic animals
25 containing germ line cells having heavy and light transgenes wherein one of the said transgenes contains rearranged gene segments with the other containing unrearranged gene segments. In such animals, the heavy chain transgenes shall have at least two C_H genes and the associated sequences required for
30 isotype switching.

The invention further includes methods for generating a synthetic variable region gene segment repertoire

to be used in the transgenes of the invention. The method comprises generating a population of immunoglobulin V segment DNAs wherein each of the V segment DNAs encodes an immunoglobulin V segment and contains at each end a cleavage recognition site of a restriction endonuclease. The population of immunoglobulin V segment DNAs is thereafter concatenated to form the synthetic immunoglobulin V segment repertoire. Such synthetic variable region heavy chain transgenes shall have at least two C_H genes and the associated sequences required for isotype switching.

Isotype Switching

In the development of a B lymphocyte, the cell initially produces IgM with a binding specificity determined by the productively rearranged V_H and V_L regions. Subsequently, each B cell and its progeny cells synthesize antibodies with the same L and H chain V regions, but they may switch the isotype of the H chain.

The use of μ or δ constant regions is largely determined by alternate splicing, permitting IgM and IgD to be coexpressed in a single cell. The other heavy chain isotypes (γ , α , and ϵ) are only expressed natively after a gene rearrangement event deletes the C μ and C δ exons. This gene rearrangement process, termed isotype switching, typically occurs by recombination between so called switch segments located immediately upstream of each heavy chain gene (except δ). The individual switch segments are between 2 and 10 kb in length, and consist primarily of short repeated sequences. The exact point of recombination differs for individual class switching events. Investigations which have used solution hybridization kinetics or Southern blotting with cDNA-derived

C_H probes have confirmed that switching can be associated with loss of C_H sequences from the cell.

The switch (S) region of the μ gene, S _{μ} , is located about 1 to 2 kb 5' to the coding sequence and is composed of numerous tandem repeats of sequences of the form (GAGCT)_n(GGGGT), where n is usually 2 to 5 but can range as high as 17. (See T. Nikaido et al. Nature 292:845-848 (1981))

Similar internally repetitive switch sequences spanning several kilobases have been found 5' of the other C_H genes. The S _{α} region has been sequenced and found to consist of tandemly repeated 80-bp homology units, whereas murine S_{V2a}, S_{V2b}, and S_{V3} all contain repeated 49-bp homology units very similar to each other. (See, P. Szurek et al., J. Immunol 135:620-626 (1985) and T. Nikaido et al., J. Biol. Chem. 257:7322-7329 (1982), which are incorporated herein by reference.) All the sequenced S regions include numerous occurrences of the pentamers GAGCT and GGGGT that are the basic repeated elements of the S _{μ} gene (T. Nikaido et al., J. Biol. Chem. 257:7322-7329 (1982) which is incorporated herein by reference); in the other S regions these pentamers are not precisely tandemly repeated as in S _{μ} , but instead are embedded in larger repeat units. The S_{V1} region has an additional higher-order structure: two direct repeat sequences flank each of two clusters of 49-bp tandem repeats. (See M. R. Mowatt et al., J. Immunol. 136:2674-2683 (1986), which is incorporated herein by reference).

Switch regions of human H chain genes have been found to be very similar to their mouse homologs. Indeed, similarity between pairs of human and mouse clones 5' to the C_H genes has been found to be confined to the S regions, a fact that confirms the biological significance of these regions.

A switch recombination between μ and α genes produces a composite S_μ - S_α sequence. Typically, there is no specific site, either in S_μ or in any other S region, where the recombination always occurs.

5 Generally, unlike the enzymatic machinery of V-J recombination, the switch machinery can apparently accommodate different alignments of the repeated homologous regions of germline S precursors and then join the sequences at different positions within the alignment. (See, T. H. Rabbits et al.,
10 Nucleic Acids Res. 9:4509-4524 (1981) and J. Ravetch et al.,
Proc. Natl. Acad. Sci. USA 77:6734-6738 (1980), which are incorporated herein by reference.)

The exact details of the mechanism(s) of selective activation of switching to a particular isotype are unknown.
15 Although exogenous influences such as lymphokines and cytokines might upregulate isotype-specific recombinases, it is also possible that the same enzymatic machinery catalyzes switches to all isotypes and that specificity lies in targeting this machinery to specific switch regions.

20 The T-cell-derived lymphokines IL-4 and IFN_γ have been shown to specifically promote the expression of certain isotypes: in the mouse, IL-4 decreases IgM, IgG2a, IgG2b, and IgG3 expression and increases IgE and IgG1 expression; while IFN_γ selectively stimulates IgG2a expression and antagonizes
25 the IL-4-induced increase in IgE and IgG1 expression (Coffman et al., J. Immunol. 136: 949 (1986) and Snapper et al.,
Science 236: 944 (1987), which are incorporated herein by reference). A combination of IL-4 and IL-5 promotes IgA expression (Coffman et al., J. Immunol. 139: 3685 (1987),
30 which is incorporated herein by reference).

Most of the experiments implicating T-cell effects on switching have not ruled out the possibility that the

observed increase in cells with particular switch recombinations might reflect selection of preswitched or precommitted cells; but the most likely explanation is that the lymphokines actually promote switch recombination.

5 Induction of class switching appears to be associated with sterile transcripts that initiate upstream of the switch segments (Lutzker et al., Mol. Cell. Biol. 8:1849 (1988); Stavnezer et al., Proc. Natl. Acad. Sci. USA 85:7704 (1988); Esser and Radbruch, EMBO J. 8:483 (1989); Berton et
10 al., Proc. Natl. Acad. Sci. USA 86:2829 (1989); Rothman et al., Int. Immunol. 2:621 (1990), each of which is incorporated by reference). For example, the observed induction of the $\gamma 1$ sterile transcript by IL-4 and inhibition by IFN- γ correlates with the observation that IL-4 promotes class switching to $\gamma 1$
15 in B-cells in culture, while IFN- γ inhibits $\gamma 1$ expression. Therefore, the inclusion of regulatory sequences that affect the transcription of sterile transcripts may also affect the rate of isotype switching. For example, increasing the transcription of a particular sterile transcript typically can
20 be expected to enhance the frequency of isotype switch recombination involving adjacent switch sequences.

For these reasons, it is preferable that transgenes incorporate transcriptional regulatory sequences within about 1-2 kb upstream of each switch region that is to be utilized
25 for isotype switching. These transcriptional regulatory sequences preferably include a promoter and an enhancer element, and more preferably include the 5' flanking (i.e., upstream) region that is naturally associated (i.e., occurs in germline configuration) with a switch region. This 5'
30 flanking region is typically about at least 50 nucleotides in length, preferably about at least 200 nucleotides in length, and more preferably at least 500-1000 nucleotides.

Although a 5' flanking sequence from one switch region can be operably linked to a different switch region for transgene construction (e.g., a 5' flanking sequence from the human $S_{\gamma 1}$ switch can be grafted immediately upstream of the $S_{\alpha 1}$ switch; a murine $S_{\gamma 1}$ flanking region can be grafted adjacent to a human $\gamma 1$ switch sequence; or the murine $S_{\gamma 1}$ switch can be grafted onto the human $\gamma 1$ coding region), in some embodiments it is preferred that each switch region incorporated in the transgene construct have the 5' flanking region that occurs immediately upstream in the naturally occurring germline configuration.

Monoclonal Antibodies

Monoclonal antibodies can be obtained by various techniques familiar to those skilled in the art. Briefly, spleen cells from an animal immunized with a desired antigen are immortalized, commonly by fusion with a myeloma cell (see, Kohler and Milstein, Eur. J. Immunol., 6:511-519 (1976)). Alternative methods of immortalization include transformation with Epstein Barr Virus, oncogenes, or retroviruses, or other methods well known in the art. Colonies arising from single immortalized cells are screened for production of antibodies of the desired specificity and affinity for the antigen, and yield of the monoclonal antibodies produced by such cells may be enhanced by various techniques, including injection into the peritoneal cavity of a vertebrate host. Various techniques useful in these arts are discussed, for example, in Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor, New York (1988) including: immunization of animals to produce immunoglobulins; production of monoclonal antibodies; labeling immunoglobulins for use as probes; immunoaffinity purification; and immunoassays.

The Transgenic Primary Repertoire

A. The Human Immunoglobulin Loci

An important requirement for transgene function is the generation of a primary antibody repertoire that is
5 diverse enough to trigger a secondary immune response for a wide range of antigens. The rearranged heavy chain gene consists of a signal peptide exon, a variable region exon and a tandem array of multi-domain constant region regions, each of which is encoded by several exons. Each of the constant
10 region genes encode the constant portion of a different class of immunoglobulins. During B-cell development, V region proximal constant regions are deleted leading to the expression of new heavy chain classes. For each heavy chain class, alternative patterns of RNA splicing give rise to both
15 transmembrane and secreted immunoglobulins.

The human heavy chain locus is estimated to consist of approximately 200 V gene segments (current data supports the existence of about 50-100 V gene segments) spanning 2 Mb, approximately 30 D gene segments spanning about 40 kb, six J
20 segments clustered within a 3 kb span, and nine constant region gene segments spread out over approximately 300 kb. The entire locus spans approximately 2.5 Mb of the distal portion of the long arm of chromosome 14.

25 B. Gene Fragment Transgenes

1. Heavy Chain Transgene

In a preferred embodiment, immunoglobulin heavy and light chain transgenes comprise unrearranged genomic DNA from humans. In the case of the heavy chain, a preferred transgene
30 comprises a NotI fragment having a length between 670 to 830 kb. The length of this fragment is ambiguous because the 3' restriction site has not been accurately mapped. It is known,

however, to reside between the $\alpha 1$ and $\psi\alpha$ gene segments. This fragment contains members of all six of the known V_H families, the D and J gene segments, as well as the μ , δ , $\gamma 3$, $\gamma 1$ and $\alpha 1$ constant regions (Berman et al., EMBO J. 7:727-738 (1988),
5 which is incorporated herein by reference). A transgenic mouse line containing this transgene correctly expresses a heavy chain class required for B-cell development (IgM) and at least one switched heavy chain class (IgG₁), in conjunction with a sufficiently large repertoire of variable regions to
10 trigger a secondary response for most antigens.

2. Light Chain Transgene

A genomic fragment containing all of the necessary gene segments and regulatory sequences from a human light
15 chain locus may be similarly constructed. Such transgenes are constructed as described in the Examples and in copending application, entitled "Transgenic Non-Human Animals Capable of Producing Heterologous Antibodies," filed August 29, 1990, under U.S.S.N. 07/574,748.

20

C. Transgenes Generated Intracellularly by In Vivo Recombination

It is not necessary to isolate the all or part of the heavy chain locus on a single DNA fragment. Thus, for
25 example, the 670-830 kb NotI fragment from the human immunoglobulin heavy chain locus may be formed in vivo in the non-human animal during transgenesis. Such in vivo transgene construction is produced by introducing two or more overlapping DNA fragments into an embryonic nucleus of the
30 non-human animal. The overlapping portions of the DNA fragments have DNA sequences which are substantially homologous. Upon exposure to the recombinases contained

within the embryonic nucleus, the overlapping DNA fragments homologously recombined in proper orientation to form the 670-830 kb NotI heavy chain fragment.

In vivo transgene construction can be used to form
5 any number of immunoglobulin transgenes which because of their size are otherwise difficult, or impossible, to make or manipulate by present technology. Thus, in vivo transgene construction is useful to generate immunoglobulin transgenes which are larger than DNA fragments which may be manipulated
10 by YAC vectors (Murray and Szostak, Nature 305:189-193 (1983)). Such in vivo transgene construction may be used to introduce into a non-human animal substantially the entire immunoglobulin loci from a species not consisting of the transgenic non-human animal.

15 In addition to forming genomic immunoglobulin transgenes, in vivo homologous recombination may also be utilized to form "mini-locus" transgenes as described in the Examples.

In the preferred embodiments utilizing in vivo
20 transgene construction, each overlapping DNA fragment preferably has an overlapping substantially homologous DNA sequence between the end portion of one DNA fragment and the end portion of a second DNA fragment. Such overlapping portions of the DNA fragments preferably comprise about 500 bp
25 to about 2000 bp, most preferably 1.0 kb to 2.0 kb. Homologous recombination of overlapping DNA fragments to form transgenes in vivo is further described in commonly assigned U.S. Patent Application entitled "Intracellular Generation of DNA by Homologous Recombination of DNA Fragments" filed August 29,
30 1990, under U.S.S.N. 07/574,747.

D. Minilocus Transgenes

As used herein, the term "immunoglobulin minilocus" refers to a DNA sequence (which may be within a longer sequence), usually of less than about 150 kb, typically
5 between about 25 and 100 kb, containing at least one each of the following: a functional variable (V) gene segment, a functional joining (J) region segment, at least one functional constant (C) region gene segment, and--if it is a heavy chain minilocus--a functional diversity (D) region segment, such
10 that said DNA sequence contains at least one substantial discontinuity (e.g., a deletion, usually of at least about 2 to 5 kb, preferably 10-25 kb or more, relative to the homologous genomic DNA sequence). A light chain minilocus transgene will be at least 25 kb in length, typically 50 to 60
15 kb. A heavy chain transgene will typically be about 70 to 80 kb in length, preferably at least about 60 kb with two constant regions operably linked to switch regions. Furthermore, the individual elements of the minilocus are preferably in the germline configuration and capable of
20 undergoing gene rearrangement in the pre-B cell of a transgenic animal so as to express functional antibody molecules with diverse antigen specificities encoded entirely by the elements of the minilocus. Further, a heavy chain minilocus comprising at least two C_H genes and the requisite
25 switching sequences is typically capable of undergoing isotype switching, so that functional antibody molecules of different immunoglobulin classes will be generated. Such isotype switching may occur in vivo in B-cells residing within the transgenic nonhuman animal, or may occur in cultured cells of
30 the B-cell lineage which have been explanted from the transgenic nonhuman animal.

In an alternate preferred embodiment, immunoglobulin heavy chain transgenes comprise one or more of each of the V_H , D , and J_H gene segments and two or more of the C_H genes. At least one of each appropriate type gene segment is

5 incorporated into the minilocus transgene. With regard to the C_H segments for the heavy chain transgene, it is preferred that the transgene contain at least one μ gene segment and at least one other constant region gene segment, more preferably a γ gene segment, and most preferably $\gamma 3$ or $\gamma 1$. This preference is

10 to allow for class switching between IgM and IgG forms of the encoded immunoglobulin and the production of a secretable form of high affinity non-IgM immunoglobulin. Other constant region gene segments may also be used such as those which encode for the production of IgD, IgA and IgE.

15 Those skilled in the art will also construct transgenes wherein the order of occurrence of heavy chain C_H genes will be different from the naturally-occurring spatial order found in the germline of the species serving as the donor of the C_H genes.

20 Additionally, those skilled in the art can select C_H genes from more than one individual of a species (e.g., allogeneic C_H genes) and incorporate said genes in the transgene as supernumerary C_H genes capable of undergoing isotype switching; the resultant transgenic nonhuman animal

25 may then, in some embodiments, make antibodies of various classes including all of the allotypes represented in the species from which the transgene C_H genes were obtained.

Still further, those skilled in the art can select C_H genes from different species to incorporate into the

30 transgene. Functional switch sequences are included with each C_H gene, although the switch sequences used are not necessarily those which occur naturally adjacent to the C_H gene.

Interspecies C_H gene combinations will produce a transgenic nonhuman animal which may produce antibodies of various classes corresponding to C_H genes from various species.

Transgenic nonhuman animals containing interspecies C_H transgenes may serve as the source of B-cells for constructing hybridomas to produce monoclonals for veterinary uses.

The heavy chain J region segments in the human comprise six functional J segments and three pseudo genes clustered in a 3 kb stretch of DNA. Given its relatively compact size and the ability to isolate these segments together with the μ gene and the 5' portion of the δ gene on a single 23 kb SfiI/SpeI fragment (Sado et al., Biochem. Biophys. Res. Comm. 154:264-271 (1988), which is incorporated herein by reference), it is preferred that all of the J region gene segments be used in the mini-locus construct. Since this fragment spans the region between the μ and δ genes, it is likely to contain all of the 3' cis-linked regulatory elements required for μ expression. Furthermore, because this fragment includes the entire J region, it contains the heavy chain enhancer and the μ switch region (Mills et al., Nature 306:809 (1983); Yancopoulos and Alt, Ann. Rev. Immunol. 4:339-368 (1986), which are incorporated herein by reference). It also contains the transcription start sites which trigger VDJ joining to form primary repertoire B-cells (Yancopoulos and Alt, Cell 40:271-281 (1985), which is incorporated herein by reference). Alternatively, a 36 kb BssHII/SpeI fragment, which includes part on the D region, may be used in place of the 23 kb SfiI/SpeI fragment. The use of such a fragment increases the amount of 5' flanking sequence to facilitate efficient D-to-J joining.

The human D region consists of 4 homologous 9 kb subregions, linked in tandem (Siebenlist, et al. (1981),

Nature, 294, 631-635). Each subregion contains up to 10 individual D segments. Some of these segments have been mapped and are shown in Fig. 4. Two different strategies are used to generate a mini-locus D region. The first strategy involves using only those D segments located in a short contiguous stretch of DNA that includes one or two of the repeated D subregions. A candidate is a single 15 kb fragment that contains 12 individual D segments. This piece of DNA consists of 2 contiguous EcoRI fragments and has been completely sequenced (Ichihara, et al. (1988), EMBO J., 7, 4141-4150). Twelve D segments should be sufficient for a primary repertoire. However, given the dispersed nature of the D region, an alternative strategy is to ligate together several non-contiguous D-segment containing fragments, to produce a smaller piece of DNA with a greater number of segments. Additional D-segment genes can be identified, for example, by the presence of characteristic flanking nonamer and heptamer sequences, supra, and by reference to the literature.

At least one, and preferably more than one V gene segment is used to construct the heavy chain minilocus transgene. Rearranged or unrearranged V segments with or without flanking sequences can be isolated as described in copending applications, U.S.S.N. 07/574,748 filed August 29, 1990, PCT/US91/06185 filed August 28, 1991, and U.S.S.N. 07/810,279 filed December 17, 1991, each of which is incorporated herein by reference.

Rearranged or unrearranged V segments, D segments, J segments, and C genes, with or without flanking sequences, can be isolated as described in copending applications U.S.S.N. 07/574,748 filed August 29, 1990 and PCT/US91/06185 filed August 28, 1991.

A minilocus light chain transgene may be similarly constructed from the human λ or κ immunoglobulin locus.

Thus, for example, an immunoglobulin heavy chain minilocus transgene construct, e.g., of about 75 kb, encoding
5 V, D, J and constant region sequences can be formed from a plurality of DNA fragments, with each sequence being substantially homologous to human gene sequences. Preferably, the sequences are operably linked to transcription regulatory sequences and are capable of undergoing rearrangement. With
10 two or more appropriately placed constant region sequences (e.g., μ and γ) and switch regions, switch recombination also occurs. An exemplary light chain transgene construct can be formed similarly from a plurality of DNA fragments, substantially homologous to human DNA and capable of
15 undergoing rearrangement, as described in copending application, U.S.S.N. 07/574,748 filed August 29, 1990.

E. Transgene Constructs Capable of Isotype Switching

Ideally, transgene constructs that are intended to
20 undergo class switching should include all of the cis-acting sequences necessary to regulate sterile transcripts. Naturally occurring switch regions and upstream promoters and regulatory sequences (e.g., IFN-inducible elements) are preferred cis-acting sequences that are included in transgene
25 constructs capable of isotype switching. About at least 50 basepairs, preferably about at least 200 basepairs, and more preferably at least 500 to 1000 basepairs or more of sequence immediately upstream of a switch region, preferably a human $\gamma 1$ switch region, should be operably linked to a switch sequence,
30 preferably a human $\gamma 1$ switch sequence. Further, switch regions can be linked upstream of (and adjacent to) C_H genes that do not naturally occur next to the particular switch

region. For example, but not for limitation, a human γ_1 switch region may be linked upstream from a human α_2 C_H gene, or a murine γ_1 switch may be linked to a human C_H gene.

An alternative method for obtaining non-classical isotype switching (e.g., δ -associated deletion) in transgenic mice involves the inclusion of the 400 bp direct repeat sequences ($\sigma\mu$ and $\epsilon\mu$) that flank the human μ gene (Yasui et al., Eur. J. Immunol. 19:1399 (1989)). Homologous recombination between these two sequences deletes the μ gene in IgD-only B-cells. Heavy chain transgenes can be represented by the following formulaic description:

$$(V_H)_x - (D)_y - (J_H)_z - (S_D)_m - (C_1)_n - [(T) - (S_A)_p - (C_2)]_q$$

where:

- V_H is a heavy chain variable region gene segment,
- D is a heavy chain D (diversity) region gene segment,
- J_H is a heavy chain J (joining) region gene segment,
- S_D is a donor region segment capable of participating in a recombination event with the S_A acceptor region segments such that isotype switching occurs,
- C_1 is a heavy chain constant region gene segment encoding an isotype utilized in for B cell development (e.g., μ or δ),
- T is a cis-acting transcriptional regulatory region segment containing at least a promoter,
- S_A is an acceptor region segment capable of participating in a recombination event with selected S_D donor region segments, such that isotype switching occurs,
- C_2 is a heavy chain constant region gene segment encoding an isotype other than μ (e.g., γ_1 , γ_2 , γ_3 , γ_4 , α_1 , α_2 , ϵ).

x, y, z, m, n, p, and q are integers. x is 1-100, n is 0-10, y is 1-50, p is 1-10, z is 1-50, q is 0-50, m is 0-10. Typically, when the transgene is capable of isotype switching, q must be at least 1, m is at least 1, n is at least 1, and m is greater than or equal to n.

V_H , D , J_H , S_D , C_1 , T , S_A , and C_2 segments may be selected from various species, preferably mammalian species, and more preferably from human and murine germline DNA.

V_H segments may be selected from various species, but are preferably selected from V_H segments that occur naturally in the human germline, such as V_{H251} . Typically about 2 V_H gene segments are included, preferably about 4 V_H segments are included, and most preferably at least about 10 V_H segments are included.

At least one D segment is typically included, although at least 10 D segments are preferably included, and some embodiments include more than ten D segments. Some preferred embodiments include human D segments.

Typically at least one J_H segment is incorporated in the transgene, although it is preferable to include about six J_H segments, and some preferred embodiments include more than about six J_H segments. Some preferred embodiments include human J_H segments, and further preferred embodiments include six human J_H segments and no nonhuman J_H segments.

S_D segments are donor regions capable of participating in recombination events with the S_A segment of the transgene. For classical isotype switching, S_D and S_A are switch regions such as S_μ , S_{Y1} , S_{Y2} , S_{Y3} , S_{Y4} , S_α , $S_{\alpha2}$, and S_ϵ . Preferably the switch regions are murine or human, more preferably S_D is a human or murine S_μ and S_A is a human or

murine $S_{\gamma 1}$. For nonclassical isotype switching (δ -associated deletion), S_D and S_A are preferably the 400 basepair direct repeat sequences that flank the human μ gene.

C_1 segments are typically μ or δ genes, preferably a
5 μ gene, and more preferably a human or murine μ gene.

T segments typically include S' flanking sequences that are adjacent to naturally occurring (i.e., germline) switch regions. T segments typically at least about at least 50 nucleotides in length, preferably about at least 200
10 nucleotides in length, and more preferably at least 500-1000 nucleotides in length. Preferably T segments are 5' flanking sequences that occur immediately upstream of human or murine switch regions in a germline configuration. It is also evident to those of skill in the art that T segments may
15 comprise cis-acting transcriptional regulatory sequences that do not occur naturally in an animal germline (e.g., viral enhancers and promoters such as those found in SV40, adenovirus, and other viruses that infect eukaryotic cells).

C_2 segments are typically a γ_1 , γ_2 , γ_3 , γ_4 , α_1 , α_2 , or
20 ϵ C_H gene, preferably a human C_H gene of these isotypes, and more preferably a human γ_1 or γ_3 gene. Murine γ_{2a} and γ_{2b} may also be used, as may downstream (i.e., switched) isotype genes from various species. Where the heavy chain transgene contains an immunoglobulin heavy chain minilocus, the total length of the
25 transgene will be typically 150 kilo basepairs or less.

In general, the transgene will be other than a native heavy chain Ig locus. Thus, for example, deletion of unnecessary regions or substitutions with corresponding regions from other species will be present.

30

F. Methods for Determining Functional Isotype Switching in Ig Transgenes

The occurrence of isotype switching in a transgenic nonhuman animal may be identified by any method known to those in the art. Preferred embodiments include the following, employed either singly or in combination:

- 5 1. detection of mRNA transcripts that contain a sequence homologous to at least one transgene downstream C_H gene other than δ and an adjacent sequence homologous to a transgene V_H - D_H - J_H rearranged gene; such detection may be by Northern hybridization, S_1 nuclease protection assays, PCR
10 amplification, cDNA cloning, or other methods;
2. detection in the serum of the transgenic animal, or in supernatants of cultures of hybridoma cells made from B-cells of the transgenic animal, of immunoglobulin proteins encoded by downstream C_H genes, where such proteins can also be shown
15 by immunochemical methods to comprise a functional variable region;
3. detection, in DNA from B-cells of the transgenic animal or in genomic DNA from hybridoma cells, of DNA rearrangements consistent with the occurrence of isotype
20 switching in the transgene, such detection may be accomplished by Southern blot hybridization, PCR amplification, genomic cloning, or other method; or
4. identification of other indicia of isotype switching, such as production of sterile transcripts, production of
25 characteristic enzymes involved in switching (e.g., "switch recombinase"), or other manifestations that may be detected, measured, or observed by contemporary techniques.

Because each transgenic line may represent a different site of integration of the transgene, and a
30 potentially different tandem array of transgene inserts, and because each different configuration of transgene and flanking DNA sequences can affect gene expression, it is preferable to

identify and use lines of mice that express high levels of human immunoglobulins, particularly of the IgG isotype, and contain the least number of copies of the transgene. Single copy transgenics minimize the potential problem of incomplete
5 allelic expression. Transgenes are typically integrated into host chromosomal DNA, most usually into germline DNA and propagated by subsequent breeding of germline transgenic breeding stock animals. However, other vectors and transgenic methods known in the present art or subsequently developed may
10 be substituted as appropriate and as desired by a practitioner.

Trans-switching to endogenous nonhuman heavy chain constant region genes can occur and produce chimeric heavy chains and antibodies comprising such chimeric human/mouse
15 heavy chains. Such chimeric antibodies may be desired for certain uses described herein or may be undesirable.

G. Functional Disruption of
Endogenous Immunoglobulin Loci

20 The expression of successfully rearranged immunoglobulin heavy and light transgenes is expected to have a dominant effect by suppressing the rearrangement of the endogenous immunoglobulin genes in the transgenic nonhuman animal. However, another way to generate a nonhuman that is
25 devoid of endogenous antibodies is by mutating the endogenous immunoglobulin loci. Using embryonic stem cell technology and homologous recombination, the endogenous immunoglobulin repertoire can be readily eliminated. The following describes the functional description of the mouse immunoglobulin loci.
30 The vectors and methods disclosed, however, can be readily adapted for use in other non-human animals.

Briefly, this technology involves the inactivation of a gene, by homologous recombination, in a pluripotent cell

line that is capable of differentiating into germ cell tissue. A DNA construct that contains an altered, copy of a mouse immunoglobulin gene is introduced into the nuclei of embryonic stem cells. In a portion of the cells, the introduced DNA
5 recombines with the endogenous copy of the mouse gene, replacing it with the altered copy. Cells containing the newly engineered genetic lesion are injected into a host mouse embryo, which is reimplanted into a recipient female. Some of these embryos develop into chimeric mice that possess germ
10 cells entirely derived from the mutant cell line. Therefore, by breeding the chimeric mice it is possible to obtain a new line of mice containing the introduced genetic lesion (reviewed by Capecchi (1989), Science, 244, 1288-1292).

Because the mouse λ locus contributes to only 5% of
15 the immunoglobulins, inactivation of the heavy chain and/or κ -light chain loci is sufficient. There are three ways to disrupt each of these loci, deletion of the J region, deletion of the J-C intron enhancer, and disruption of constant region coding sequences by the introduction of a stop codon. The
20 last option is the most straightforward, in terms of DNA construct design. Elimination of the μ gene disrupts B-cell maturation thereby preventing class switching to any of the functional heavy chain segments. The strategy for knocking out these loci is outlined below.

25 To disrupt the mouse μ and κ genes, targeting vectors are used based on the design employed by Jaenisch and co-workers (Zijlstra, et al. (1989), Nature, 342, 435-438) for the successful disruption of the mouse $\beta 2$ -microglobulin gene. The neomycin resistance gene (neo), from the plasmid pMCIneo
30 is inserted into the coding region of the target gene. The pMCIneo insert uses a hybrid viral promoter/enhancer sequence to drive neo expression. This promoter is active in embryonic

stem cells. Therefore, neo can be used as a selectable marker for integration of the knock-out construct. The HSV thymidine kinase (tk) gene is added to the end of the construct as a negative selection marker against random insertion events (Zijlstra, et al., supra.).

A preferred strategy for disrupting the heavy chain locus is the elimination of the J region. This region is fairly compact in the mouse, spanning only 1.3 kb. To construct a gene targeting vector, a 15 kb KpnI fragment containing all of the secreted A constant region exons from mouse genomic library is isolated. The 1.3 kb J region is replaced with the 1.1 kb insert from pMCIneo. The HSV tk gene is then added to the 5' end of the KpnI fragment. Correct integration of this construct, via homologous recombination, will result in the replacement of the mouse J_H region with the neo gene. Recombinants are screened by PCR, using a primer based on the neo gene and a primer homologous to mouse sequences 5' of the KpnI site in the D region.

Alternatively, the heavy-chain locus is knocked out by disrupting the coding region of the μ gene. This approach involves the same 15 kb KpnI fragment used in the previous approach. The 1.1 kb insert from pMCIneo is inserted at a unique BamHI site in exon II, and the HSV tk gene added to the 3' KpnI end. Double crossover events on either side of the neo insert, that eliminate the tk gene, are then selected for. These are detected from pools of selected clones by PCR amplification. One of the PCR primers is derived from neo sequences and the other from mouse sequences outside of the targeting vector. The functional disruption of the mouse immunoglobulin loci is presented in the Examples.

G. Suppressing Expression of

Endogenous Immunoglobulin Loci

In addition to functional disruption of endogenous Ig loci, an alternative method for preventing the expression of an endogenous Ig locus is suppression. Suppression of endogenous Ig genes may be accomplished with antisense RNA produced from one or more integrated transgenes, by antisense oligonucleotides, and/or by administration of antisera specific for one or more endogenous Ig chains.

Antisense Polynucleotides

Antisense RNA transgenes can be employed to partially or totally knock-out expression of specific genes (Pepin et al. (1991) Nature 355: 725; Helene., C. and Toulme, J. (1990) Biochimica Biophys. Acta 1049: 99; Stout, J. and Caskey, T. (1990) Somat. Cell Mol. Genet. 16: 369; Munir et al. (1990) Somat. Cell Mol. Genet. 16: 383, each of which is incorporated herein by reference).

"Antisense polynucleotides" are polynucleotides that: (1) are complementary to all or part of a reference sequence, such as a sequence of an endogenous Ig C_H or C_L region, and (2) which specifically hybridize to a complementary target sequence, such as a chromosomal gene locus or a Ig mRNA. Such complementary antisense polynucleotides may include nucleotide substitutions, additions, deletions, or transpositions, so long as specific hybridization to the relevant target sequence is retained as a functional property of the polynucleotide. Complementary antisense polynucleotides include soluble antisense RNA or DNA oligonucleotides which can hybridize specifically to individual mRNA species and prevent transcription and/or RNA processing of the mRNA species and/or translation of the encoded polypeptide (Ching et al., Proc. Natl. Acad. Sci.

U.S.A. 86:10006-10010 (1989); Broder et al., Ann. Int. Med.
113:604-618 (1990); Loreau et al., FEBS Letters 274:53-56
(1990); Holcenberg et al., WO91/11535; U.S.S.N. 07/530,165
("New human CRIPTO gene"); WO91/09865; WO91/04753; WO90/13641;
5 and EP 386563, each of which is incorporated herein by
reference). An antisense sequence is a polynucleotide
sequence that is complementary to at least one immunoglobulin
gene sequence of at least about 15 contiguous nucleotides in
length, typically at least 20 to 30 nucleotides in length, and
10 preferably more than about 30 nucleotides in length. However,
in some embodiments, antisense sequences may have
substitutions, additions, or deletions as compared to the
complementary immunoglobulin gene sequence, so long as
specific hybridization is retained as a property of the
15 antisense polynucleotide. Generally, an antisense sequence is
complementary to an endogenous immunoglobulin gene sequence
that encodes, or has the potential to encode after DNA
rearrangement, an immunoglobulin chain. In some cases, sense
sequences corresponding to an immunoglobulin gene sequence may
20 function to suppress expression, particularly by interfering
with transcription.

The antisense polynucleotides therefore inhibit
production of the encoded polypeptide(s). In this regard,
antisense polynucleotides that inhibit transcription and/or
25 translation of one or more endogenous Ig loci can alter the
capacity and/or specificity of a non-human animal to produce
immunoglobulin chains encoded by endogenous Ig loci.

Antisense polynucleotides may be produced from a
heterologous expression cassette in a transfectant cell or
30 transgenic cell, such as a transgenic pluripotent
hematopoietic stem cell used to reconstitute all or part of
the hematopoietic stem cell population of an individual, or a

transgenic nonhuman animal. Alternatively, the antisense polynucleotides may comprise soluble oligonucleotides that are administered to the external milieu, either in culture medium in vitro or in the circulatory system or interstitial fluid in vivo. Soluble antisense polynucleotides present in the external milieu have been shown to gain access to the cytoplasm and inhibit translation of specific mRNA species. In some embodiments the antisense polynucleotides comprise methylphosphonate moieties, alternatively phosphorothiolates or O-methylribonucleotides may be used, and chimeric oligonucleotides may also be used (Dagle et al. (1990) Nucleic Acids Res. 18: 4751). For some applications, antisense oligonucleotides may comprise polyamide nucleic acids (Nielsen et al. (1991) Science 254: 1497). For general methods relating to antisense polynucleotides, see Antisense RNA and DNA, (1988), D.A. Melton, Ed., Cold Spring Harbor Laboratory, Cold Spring Harbor, NY).

Antisense polynucleotides complementary to one or more sequences are employed to inhibit transcription, RNA processing, and/or translation of the cognate mRNA species and thereby effect a reduction in the amount of the respective encoded polypeptide. Such antisense polynucleotides can provide a therapeutic function by inhibiting the formation of one or more endogenous Ig chains in vivo.

Whether as soluble antisense oligonucleotides or as antisense RNA transcribed from an antisense transgene, the antisense polynucleotides of this invention are selected so as to hybridize preferentially to endogenous Ig sequences at physiological conditions in vivo. Most typically, the selected antisense polynucleotides will not appreciably hybridize to heterologous Ig sequences encoded by a heavy or light chain transgene of the invention (i.e., the antisense

oligonucleotides will not inhibit transgene Ig expression by more than about 25 to 35 percent).

Antiserum Suppression

5 Partial or complete suppression of endogenous Ig chain expression can be produced by injecting mice with antisera against one or more endogenous Ig chains (Weiss et al. (1984) Proc. Natl. Acad. Sci. (U.S.A.) 81 211, which is incorporated herein by reference). Antisera are selected so
10 as to react specifically with one or more endogenous (e.g., murine) Ig chains but to have minimal or no cross-reactivity with heterologous Ig chains encoded by an Ig transgene of the invention. Thus, administration of selected antisera according to a schedule as typified by that of Weiss et al.
15 op.cit. will suppress endogenous Ig chain expression but permits expression of heterologous Ig chain(s) encoded by a transgene of the present invention. Suitable antibody sources for antibody comprise:

(1) monoclonal antibodies, such as a monoclonal
20 antibody that specifically binds to a murine μ , γ , κ , or λ chains but does not react with the human immunoglobulin chain(s) encoded by a human Ig transgene of the invention;

(2) mixtures of such monoclonal antibodies, so that the mixture binds with multiple epitopes on a single species
25 of endogenous Ig chain, with multiple endogenous Ig chains (e.g., murine μ and murine γ , or with multiple epitopes and multiple chains or endogenous immunoglobulins;

(3) polyclonal antiserum or mixtures thereof, typically such antiserum/antisera is monospecific for binding
30 to a single species of endogenous Ig chain (e.g., murine μ , murine γ , murine κ , murine λ) or to multiple species of endogenous Ig chain, and most preferably such antisera

possesses negligible binding to human immunoglobulin chains encoded by a transgene of the invention; and/or

(4) a mixture of polyclonal antiserum and monoclonal antibodies binding to a single or multiple species of endogenous Ig chain, and most preferably possessing negligible binding to human immunoglobulin chains encoded by a transgene of the invention. Generally, polyclonal antibodies are preferred, and such substantially monospecific polyclonal antibodies can be advantageously produced from an antiserum raised against human immunoglobulin(s) by pre-adsorption with antibodies derived from the nonhuman animal species (e.g., murine) and/or, for example, by affinity chromatography of the antiserum or purified fraction thereof on an affinity resin containing immobilized human Ig (wherein the bound fraction is enriched for the desired anti-human Ig in the antiserum; the bound fraction is typically eluted with conditions of low pH or a chaotropic salt solution).

Cell separation and/or complement fixation can be employed to provide the enhancement of antibody-directed cell depletion of lymphocytes expressing endogenous (e.g., murine) immunoglobulin chains. In one embodiment, for example, antibodies are employed for ex vivo depletion of murine Ig-expressing explanted hematopoietic cells and/or B-lineage lymphocytes obtained from a transgenic mouse harboring a human Ig transgene. Thus, hematopoietic cells and/or B-lineage lymphocytes are explanted from a transgenic nonhuman animal harboring a human Ig transgene (preferably harboring both a human heavy chain transgene and a human light chain transgene) and the explanted cells are incubated with an antibody (or antibodies) which (1) binds to an endogenous immunoglobulin (e.g., murine μ and/or κ) and (2) lacks substantial binding to human immunoglobulin chains encoded by the transgene(s). Such

antibodies are referred to as "suppression antibodies" for clarity. The explanted cell population is selectively depleted of cells which bind to the suppression antibody(ies); such depletion can be accomplished by various methods, such as

5 (1) physical separation to remove suppression antibody-bound cells from unbound cells (e.g., the suppression antibodies may be bound to a solid support or magnetic bead to immobilize and remove cells binding to the suppression antibody), (2)

10 antibody-dependent cell killing of cells bound by the suppression antibody (e.g., by ADCC, by complement fixation, or by a toxin linked to the suppression antibody), and (3) clonal anergy induced by the suppression antibody, and the like.

Frequently, antibodies used for antibody suppression

15 of endogenous Ig chain production will be capable of fixing complement. It is frequently preferable that such antibodies may be selected so as to react well with a convenient complement source for ex vivo/in vitro depletion, such as rabbit or guinea pig complement. For in vivo depletion, it is

20 generally preferred that the suppressor antibodies possess effector functions in the nonhuman transgenic animal species; thus, a suppression antibody comprising murine effector functions (e.g., ADCC and complement fixation) generally would be preferred for use in transgenic mice.

25 In one variation, a suppression antibody that specifically binds to a predetermined endogenous immunoglobulin chain is used for ex vivo/in vitro depletion of lymphocytes expressing an endogenous immunoglobulin. A cellular explant (e.g., lymphocyte sample) from a transgenic

30 nonhuman animal harboring a human immunoglobulin transgene is contacted with a suppression antibody and cells specifically binding to the suppression antibody are depleted (e.g., by

immobilization, complement fixation, and the like), thus generating a cell subpopulation depleted in cells expressing endogenous (nonhuman) immunoglobulins (e.g., lymphocytes expressing murine Ig). The resultant depleted lymphocyte population (T cells, human Ig-positive B-cells, etc.) can be transferred into a immunocompatible (i.e., MHC-compatible) nonhuman animal of the same species and which is substantially incapable of producing endogenous antibody (e.g., SCID mice, RAG-1 or RAG-2 knockout mice). The reconstituted animal (mouse) can then be immunized with an antigen (or reimmunized with an antigen used to immunize the donor animal from which the explant was obtained) to obtain high-affinity (affinity matured) antibodies and B-cells producing such antibodies. Such B-cells may be used to generate hybridomas by conventional cell fusion and screened. Antibody suppression can be used in combination with other endogenous Ig inactivation/suppression methods (e.g., J_H knockout, C_H knockout, D-region ablation, antisense suppression, compensated frameshift inactivation).

20

Complete Endogenous Ig Locus Inactivation

In certain embodiments, it is desirable to effect complete inactivation of the endogenous Ig loci so that hybrid immunoglobulin chains comprising a human variable region and a non-human (e.g., murine) constant region cannot be formed (e.g., by trans-switching between the transgene and endogenous Ig sequences). Knockout mice bearing endogenous heavy chain alleles which are functionally disrupted in the J_H region only frequently exhibit trans-switching, typically wherein a rearranged human variable region (VDJ) encoded by a transgene is expressed as a fusion protein linked to an endogenous murine constant region, although other trans-switched

30

junctions are possible. To overcome this potential problem, it is generally desirable to completely inactivate the endogenous heavy chain locus by any of various methods, including but not limited to the following: (1) functionally
5 disrupting and/or deleting by homologous recombination at least one and preferably all of the endogenous heavy chain constant region genes, (2) mutating at least one and preferably all of the endogenous heavy chain constant region genes to encode a termination codon (or frameshift) to produce
10 a truncated or frameshifted product (if trans-switched), and other methods and strategies apparent to those of skill in the art. Deletion of a substantial portion or all of the heavy chain constant region genes and/or D-region genes may be accomplished by various methods, including sequential deletion
15 by homologous recombination targeting vectors, especially of the "hit-and-run" type and the like. Similarly, functional disruption and/or deletion of at least one endogenous light chain locus (e.g., κ) to ablate endogenous light chain constant region genes is often preferable.

20 Frequently, it is desirable to employ a frameshifted transgene wherein the heterologous transgene comprises a frameshift in the J segment(s) and a compensating frameshift (i.e., to regenerate the original reading frame) in the initial region (i.e., amino-terminal coding portion) of one or
25 more (preferably all) of the transgene constant region genes. Trans-switching to an endogenous IgH locus constant gene (which does not comprise a compensating frameshift) will result in a truncated or missense product that results in the trans-switched B cell being deleted or non-selected, thus
30 suppressing the trans-switched phenotype.

Antisense suppression and antibody suppression may also be used to effect a substantially complete functional

inactivation of endogenous Ig gene product expression (e.g., murine heavy and light chain sequences) and/or trans-switched antibodies (e.g., human variable/murine constant chimeric antibodies).

- 5 Various combinations of the inactivation and suppression strategies may be used to effect essentially total suppression of endogenous (e.g., murine) Ig chain expression.

Trans-Switching

- 10 In some variations, it may be desirable to produce a trans-switched immunoglobulin. For example, such trans-switched heavy chains can be chimeric (i.e., a non-murine (human) variable region and a murine constant region). Antibodies comprising such chimeric trans-switched
- 15 immunoglobulins can be used for a variety of applications where it is desirable to have a non-human (e.g., murine) constant region (e.g., for retention of effector functions in the host, for the presence of murine immunological determinants such as for binding of a secondary antibody which
- 20 does not bind human constant regions). For one example, a human variable region repertoire may possess advantages as compared to the murine variable region repertoire with respect to certain antigens. Presumably the human V_H , D , J_H , V_L , and J_L genes have been selected for during evolution for their
- 25 ability to encode immunoglobulins that bind certain evolutionarily important antigens; antigens which provided evolutionary selective pressure for the murine repertoire can be distinct from those antigens which provided evolutionary pressure to shape the human repertoire. Other repertoire
- 30 advantages may exist, making the human variable region repertoire advantageous when combined with a murine constant region (e.g., a trans-switched murine) isotype. The presence

of a murine constant region can afford advantages over a human constant region. For example, a murine γ constant region linked to a human variable region by trans-switching may provide an antibody which possesses murine effector functions (e.g., ADCC, murine complement fixation) so that such a chimeric antibody (preferably monoclonal) which is reactive with a predetermined antigen (e.g., human IL-2 receptor) may be tested in a mouse disease model, such as a mouse model of graft-versus-host disease wherein the T lymphocytes in the mouse express a functional human IL-2 receptor. Subsequently, the human variable region encoding sequence may be isolated (e.g., by PCR amplification or cDNA cloning from the source (hybridoma clone)) and spliced to a sequence encoding a desired human constant region to encode a human sequence antibody more suitable for human therapeutic uses where immunogenicity is preferably minimized. The polynucleotide(s) having the resultant fully human encoding sequence(s) can be expressed in a host cell (e.g., from an expression vector in a mammalian cell) and purified for pharmaceutical formulation. For some applications, the chimeric antibodies may be used directly without replacing the murine constant region with a human constant region. Other variations and uses of trans-switched chimeric antibodies will be evident to those of skill in the art.

The present invention provides transgenic nonhuman animals containing B lymphocytes which express chimeric antibodies, generally resulting from trans-switching between a human heavy chain transgene and an endogenous murine heavy chain constant region gene. Such chimeric antibodies comprise a human sequence variable region and a murine constant region, generally a murine switched (i.e., non- μ , non- δ) isotype. The transgenic nonhuman animals capable of making chimeric

antibodies to a predetermined antigen are usually also competent to make fully human sequence antibodies if both human heavy chain and human light chain transgenes encoding human variable and human constant region genes are integrated.

5 Most typically, the animal is homozygous for a functionally disrupted heavy chain locus and/or light chain locus but retains one or more endogenous heavy chain constant region gene(s) capable of trans-switching (e.g., γ , α , ϵ) and frequently retains a cis-linked enhancer. Such a mouse is

10 immunized with a predetermined antigen, usually in combination with an adjuvant, and an immune response comprising a detectable amount of chimeric antibodies comprising heavy chains composed of human sequence variable regions linked to murine constant region sequences is produced. Typically, the

15 serum of such an immunized animal can comprise such chimeric antibodies at concentrations of about at least 1 $\mu\text{g/ml}$, often about at least 10 $\mu\text{g/ml}$, frequently at least 30 $\mu\text{g/ml}$, and up to 50 to 100 $\mu\text{g/ml}$ or more. The antiserum containing antibodies comprising chimeric human variable/mouse constant

20 region heavy chains typically also comprises antibodies which comprise human variable/human constant region (complete human sequence) heavy chains. Chimeric trans-switched antibodies usually comprise (1) a chimeric heavy chain composed of a human variable region and a murine constant region (typically

25 a murine gamma) and (2) a human transgene-encoded light chain (typically kappa) or a murine light chain (typically lambda in a kappa knockout background). Such chimeric trans-switched antibodies generally bind to a predetermined antigen (e.g., the immunogen) with an affinity of about at least $1 \times 10^7 \text{ M}^{-1}$,

30 preferably with an affinity of about at least $5 \times 10^7 \text{ M}^{-1}$, more preferably with an affinity of at least $1 \times 10^8 \text{ M}^{-1}$ to $1 \times 10^9 \text{ M}^{-1}$ or more. Frequently, the predetermined antigen is a human

protein, such as for example a human cell surface antigen (e.g., CD4, CD8, IL-8, IL-2 receptor, EGF receptor, PDGF receptor), other human biological macromolecule (e.g., thrombomodulin, protein C, carbohydrate antigen, sialyl Lewis
5 antigen, L-selectin), or nonhuman disease associated macromolecule (e.g., bacterial LPS, virion capsid protein or envelope glycoprotein) and the like.

The invention provides transgenic nonhuman animals comprising a genome comprising: (1) a homozygous functionally
10 disrupted endogenous heavy chain locus comprising at least one murine constant region gene capable of trans-switching (e.g., in cis linkage to a functional switch recombination sequence and typically to a functional enhancer), (2) a human heavy chain transgene capable of rearranging to encode and express a
15 functional human heavy chain variable region and capable of trans-switching (e.g., having a cis-linked RSS); optionally further comprising (3) a human light chain (e.g., kappa) transgene capable of rearranging to encode a functional human light chain variable region and expressing a human sequence
20 light chain; optionally further comprising (4) a homozygous functionally disrupted endogenous light chain locus (κ , preferably κ and λ); and optionally further comprising (5) a serum comprising an antibody comprising a chimeric heavy chain composed of a human sequence variable region encoded by a
25 human transgene and a murine constant region sequence encoded by an endogenous murine heavy chain constant region gene (e.g., $\gamma 1$, $\gamma 2a$, $\gamma 2b$, $\gamma 3$).

Such transgenic mice may further comprise a serum comprising chimeric antibodies which bind a predetermined
30 human antigen (e.g., CD4, CD8, CEA) with an affinity of about at least $1 \times 10^4 \text{ M}^{-1}$, preferably with an affinity of about at least $5 \times 10^4 \text{ M}^{-1}$, more preferably with an affinity of at least

1 x 10⁷ M⁻¹ to 1 x 10⁹ M⁻¹ or more. Frequently, hybridomas can be made wherein the monoclonal antibodies produced thereby have an affinity of at least 8 x10⁷ M⁻¹. Chimeric antibodies comprising a heavy chain composed of a murine constant region
5 and a human variable region, often capable of binding to a nonhuman antigen, may also be present in the serum or as an antibody secreted from a hybridoma.

In some variations, it is desirable to generate transgenic mice which have inactivated endogenous mouse heavy
10 chain loci which retain intact heavy chain constant region genes, and which have a human heavy chain transgene capable of trans-switching, and optionally also have a human light chain transgene, optionally with one or more inactivated endogenous mouse light chain loci. Such mice may advantageously produce
15 B cells capable of alternatively expressing antibodies comprising fully human heavy chains and antibodies comprising chimeric (human variable/mouse constant) heavy chains, by trans-switching. The serum of said mice would contain antibodies comprising fully human heavy chains and antibodies
20 comprising chimeric (human variable/mouse constant) heavy chains, preferably in combination with fully human light chains. Hybridomas can be generated from the B cells of said mice.

Generally, such chimeric antibodies can be generated
25 by trans-switching, wherein a human transgene encoding a human variable region (encoded by productive V-D-J rearrangement in vivo) and a human constant region, typically human μ , undergoes switch recombination with a non-transgene immunoglobulin constant gene switch sequence (RSS) thereby
30 operably linking the transgene-encoded human variable region with a heavy chain constant region which is not encoded by said transgene, typically an endogenous murine immunoglobulin

heavy chain constant region or a heterologous (e.g., human) heavy chain constant region encoded on a second transgene.

Whereas cis-switching refers to isotype-switching by recombination of RSS elements *within* a transgene, trans-switching involves recombination between a transgene RSS and an RSS element outside the transgene, often on a different chromosome than the chromosome which harbors the transgene.

Trans-switching generally occurs between an RSS of an expressed transgene heavy chain constant region gene and either an RSS of an endogenous murine constant region gene (of a non- μ isotype, typically γ) or an RSS of a human constant region gene contained on a second transgene, often integrated on a separate chromosome.

When trans-switching occurs between an RSS of a first, expressed transgene heavy chain constant region gene (e.g., μ) and an RSS of a human heavy chain constant region gene contained on a second transgene, a non-chimeric antibody having a substantially fully human sequence is produced. For example and not limitation, a polynucleotide encoding a human heavy chain constant region (e.g., $\gamma 1$) and an operably linked RSS (e.g., a $\gamma 1$ RSS) can be introduced (e.g., transfected) into a population of hybridoma cells generated from a transgenic mouse B-cell (or B cell population) expressing an antibody comprising a transgene-encoded human μ chain. The resultant hybridoma cells can be selected for the presence of the introduced polynucleotide and/or for the expression of trans-switched antibody comprising a heavy chain having the variable region (idiotype/antigen reactivity) of the human μ chain and having the constant region encoded by the introduced polynucleotide sequence (e.g., human $\gamma 1$). Trans-switch recombination between the RSS of the transgene-encoded human μ chain and the RSS of the introduced polynucleotide encoding a

downstream isotype (e.g., $\gamma 1$) thereby can generate a trans-switched antibody.

The invention also provides a method for producing such chimeric trans-switched antibodies comprising the step of
5 immunizing with a predetermined antigen a transgenic mouse comprising a genome comprising: (1) a homozygous functionally disrupted endogenous heavy chain locus comprising at least one murine constant region gene capable of trans-switching (e.g., $\gamma 2a$, $\gamma 2b$, $\gamma 1$, $\gamma 3$), (2) a human heavy chain transgene capable
10 of rearranging to encode a functional human heavy chain variable region and expressing a human sequence heavy chain and capable of undergoing isotype switching (and/or trans-switching), and optionally further comprising (3) a human light chain (e.g., kappa) transgene capable of rearranging to
15 encode a functional human light (e.g., kappa) chain variable region and expressing a human sequence light chain, and optionally further comprising (4) a homozygous functionally disrupted endogenous light chain locus (typically κ , preferably both κ and λ), and optionally further comprising
20 (5) a serum comprising an antibody comprising a chimeric heavy chain composed of a human sequence variable region encoded by a human transgene and a murine constant region sequence encoded by an endogenous murine heavy chain constant region gene (e.g., $\gamma 1$, $\gamma 2a$, $\gamma 2b$, $\gamma 3$).

25

Affinity Tagging: Selecting for Switched Isotypes

Advantageously, trans-switching (and cis-switching) is associated with the process of somatic mutation. Somatic mutation expands the range of antibody affinities encoded by
30 clonal progeny of a B-cell. For example, antibodies produced by hybridoma cells which have undergone switching (trans- or cis-) represent a broader range of antigen-binding affinities

than is present in hybridoma cells which have not undergone switching. Thus, a hybridoma cell population (typically clonal) which expresses a first antibody comprising a heavy chain comprising a first human heavy chain variable region in polypeptide linkage to a first human heavy chain constant region (e.g., μ) can be screened for hybridoma cell clonal variants which express an antibody comprising a heavy chain containing said first human heavy chain variable region in polypeptide linkage to a second heavy chain constant region (e.g., a human γ , α , or ϵ constant region). Such clonal variants can be produced by natural clonal variation producing cis-switching in vitro, by induction of class switching (trans- or cis-) as through the administration of agents that promote isotype switching, such as T-cell-derived lymphokines (e.g., IL-4 and IFN $_{\gamma}$), by introduction of a polynucleotide comprising a functional RSS and a heterologous (e.g. human) heavy chain constant region gene to serve as a substrate for trans-switching, or by a combination of the above, and the like. Often, polynucleotides containing a human downstream isotype constant region (e.g., γ 1, γ 3, and the like) with an operably linked RSS will also be introduced into hybridoma cells to promote isotype switching via the trans-switch mechanism.

Class switching and affinity maturation take place within the same population of B cells derived from transgenic animals of the present invention. Therefore, identification of class-switched B cells (or hybridomas derived therefrom) can be used as a screening step for obtaining high affinity monoclonal antibodies. A variety of approaches can be employed to facilitate class switching events such as cis-switching (intratransgene switching), trans-switching, or both. For example, a single continuous human genomic fragment

comprising both μ and γ constant region genes with the associated RSS elements and switch regulatory elements (e.g., sterile transcript promoter) can be used as a transgene.

However, some portions of the desired single contiguous human genomic fragment can be difficult to clone efficiently, such as due to instability problems when replicated in a cloning host or the like; in particular, the region between δ and $\gamma 3$ can prove difficult to clone efficiently, especially as a contiguous fragment comprising the μ gene, $\gamma 3$ gene, a V gene, D gene segments, and J gene segments.

Also for example, a discontinuous human transgene (minigene) composed of a human μ gene, human $\gamma 3$ gene, a human V gene(s), human D gene segments, and human J gene segments, with one or more deletions of an intervening (intronic) or otherwise nonessential sequence (e.g., one or more V, D, and/or J segment and/or one or more non- μ constant region gene(s)). Such minigenes have several advantages as compared to isolating a single contiguous segment of genomic DNA spanning all of the essential elements for efficient immunoglobulin expression and switching. For example, such a minigene avoids the necessity of isolating large pieces of DNA which may contain sequences which are difficult to clone (e.g., unstable sequences, poison sequences, and the like). Moreover, miniloci comprising elements necessary for isotype switching (e.g., human γ sterile transcript promoter) for producing cis- or trans-switching, can advantageously undergo somatic mutation and class switching in vivo. As many eukaryotic DNA sequences can prove difficult to clone, omitting non-essential sequences can prove advantageous.

In a variation, hybridoma clones producing antibodies having high binding affinity (e.g., at least 1×10^7 M^{-1} , preferably at least 1×10^8 M^{-1} , more preferably at least 1

x 10^9 M⁻¹ or greater) are obtained by selecting, from a pool of hybridoma cells derived from B cells of transgenic mice harboring a human heavy chain transgene capable of isotype switching (see, supra) and substantially lacking endogenous murine heavy chain loci capable of undergoing productive (in-frame) V-D-J rearrangement, hybridomas which express an antibody comprising a heavy chain comprising a human sequence heavy chain variable region in polypeptide linkage to a human (or mouse) non- μ heavy chain constant region; said antibodies are termed "switched antibodies" as they comprise a "switched heavy chain" which is produced as a consequence of cis-switching and/or trans-switching in vivo or in cell culture. Hybridomas producing switched antibodies generally have undergone the process of somatic mutation, and a pool of said hybridomas will generally have a broader range of antigen binding affinities from which hybridoma clones secreting high affinity antibodies can be selected. Typically, hybridomas secreting a human sequence antibody having substantial binding affinity (greater than 1×10^7 M⁻¹ to 1×10^8 M⁻¹) for a predetermined antigen and wherein said human sequence antibody comprises human immunoglobulin variable region(s) can be selected by a method comprising a two-step process. One step is to identify and isolate hybridoma cells which secrete immunoglobulins which comprise a switched heavy chain (e.g., by binding hybridoma cells to an immobilized immunoglobulin which specifically binds a switched heavy chain and does not substantially bind to an unswitched isotype, e.g., μ). The other step is to identify hybridoma cells which bind to the predetermined antigen with substantial binding affinity (e.g., by ELISA of hybridoma clone supernatants, FACS analysis using labeled antigen, and the like). Typically, selection of hybridomas which secrete switched antibodies is performed

prior to identifying hybridoma cells which bind predetermined antigen. Hybridoma cells which express switched antibodies that have substantial binding affinity for the predetermined antigen are isolated and cultured under suitable growth
5 conditions known in the art, typically as individual selected clones. Optionally, the method comprises the step of culturing said selected clones under conditions suitable for expression of monoclonal antibodies; said monoclonal antibodies are collected and can be administered for therapeutic,
10 prophylactic, and/or diagnostic purposes.

Often, the selected hybridoma clones can serve as a source of DNA or RNA for isolating immunoglobulin sequences which encode immunoglobulins (e.g. a variable region) that bind to (or confer binding to) the predetermined antigen.
15 Subsequently, the human variable region encoding sequence may be isolated (e.g., by PCR amplification or cDNA cloning from the source (hybridoma clone)) and spliced to a sequence encoding a desired human constant region to encode a human sequence antibody more suitable for human therapeutic uses
20 where immunogenicity is preferably minimized. The polynucleotide(s) having the resultant fully human encoding sequence(s) can be expressed in a host cell (e.g., from an expression vector in a mammalian cell) and purified for pharmaceutical formulation.

25

Xenoenhancers

A heterologous transgene capable of encoding a human immunoglobulin (e.g., a heavy chain) advantageously comprises a cis-linked enhancer which is not derived from the mouse
30 genome, and/or which is not naturally associated in cis with the exons of the heterologous transgene. For example, a human κ transgene (e.g., a κ minilocus) can advantageously comprise

a human Vk gene, a human Jk gene, a human Ck gene, and a xenoenhancer, typically said xenoenhancer comprises a human heavy chain intronic enhancer and/or a murine heavy chain intronic enhancer, typically located between a Jk gene and the Ck gene, or located downstream of the Ck gene. For example, the mouse heavy chain J- μ intronic enhancer (Banerji et al. (1983) Cell 33: 729) can be isolated on a 0.9 kb XbaI fragment of the plasmid pKVe2 (see, infra). The human heavy chain J- μ intronic enhancer (Hayday et al. (1984) Nature 307: 334) can be isolated as a 1.4 kb MluI/HindIII fragment (see, infra). Addition of a transcriptionally active xenoenhancer to a transgene, such as a combined xenoenhancer consisting essentially of a human J- μ intronic enhancer linked in cis to a mouse J- μ intronic enhancer, can confer high levels of expression of the transgene, especially where said transgene encodes a light chain, such as human κ . Similarly, a rat 3' enhancer can be advantageously included in a minilocus construct capable of encoding a human heavy chain.

20 Specific Preferred Embodiments

A preferred embodiment of the invention is an animal containing at least one, typically 2-10, and sometimes 25-50 or more copies of the transgene described in Example 12 (e.g., pHCl or pHCl2) bred with an animal containing a single copy of a light chain transgene described in Examples 5, 6, 8, or 14, and the offspring bred with the J_H deleted animal described in Example 10. Animals are bred to homozygosity for each of these three traits. Such animals have the following genotype: a single copy (per haploid set of chromosomes) of a human heavy chain unrearranged mini-locus (described in Example 12), a single copy (per haploid set of chromosomes) of a rearranged human κ light chain construct (described in Example 14), and a

WO 98/24884

PCT/US97/21803

96

deletion at each endogenous mouse heavy chain locus that removes all of the functional J_H segments (described in Example 10). Such animals are bred with mice that are homozygous for the deletion of the J_H segments (Examples 10) to produce
5 offspring that are homozygous for the J_H deletion and hemizygous for the human heavy and light chain constructs. The resultant animals are injected with antigens and used for production of human monoclonal antibodies against these antigens.

10 B cells isolated from such an animal are monospecific with regard to the human heavy and light chains because they contain only a single copy of each gene. Furthermore, they will be monospecific with regards to human or mouse heavy chains because both endogenous mouse heavy
15 chain gene copies are nonfunctional by virtue of the deletion spanning the J_H region introduced as described in Example 9 and 12. Furthermore, a substantial fraction of the B cells will be monospecific with regards to the human or mouse light chains because expression of the single copy of the rearranged
20 human κ light chain gene will allelically and isotypically exclude the rearrangement of the endogenous mouse κ and λ chain genes in a significant fraction of B-cells.

The transgenic mouse of the preferred embodiment will exhibit immunoglobulin production with a significant
25 repertoire, ideally substantially similar to that of a native mouse. Thus, for example, in embodiments where the endogenous Ig genes have been inactivated, the total immunoglobulin levels will range from about 0.1 to 10 mg/ml of serum, preferably 0.5 to 5 mg/ml, ideally at least about 1.0 mg/ml.
30 When a transgene capable of effecting a switch to IgG from IgM has been introduced into the transgenic mouse, the adult mouse ratio of serum IgG to IgM is preferably about 10:1. Of

course, the IgG to IgM ratio will be much lower in the immature mouse. In general, greater than about 10%, preferably 40 to 80% of the spleen and lymph node B cells express exclusively human IgG protein.

5 The repertoire will ideally approximate that shown in a non-transgenic mouse, usually at least about 10% as high, preferably 25 to 50% or more. Generally, at least about a thousand different immunoglobulins (ideally IgG), preferably 10^4 to 10^6 or more, will be produced, depending primarily on
10 the number of different V, J and D regions introduced into the mouse genome. These immunoglobulins will typically recognize about one-half or more of highly antigenic proteins, including, but not limited to: pigeon cytochrome C, chicken lysozyme, pokeweed mitogen, bovine serum albumin, keyhole
15 limpit hemocyanin, influenza hemagglutinin, staphylococcus protein A, sperm whale myoglobin, influenza neuraminidase, and lambda repressor protein. Some of the immunoglobulins will exhibit an affinity for preselected antigens of at least about 10^7M^{-1} , preferably 10^8M^{-1} to 10^9M^{-1} or greater.

20 In some embodiments, it may be preferable to generate mice with predetermined repertoires to limit the selection of V genes represented in the antibody response to a predetermined antigen type. A heavy chain transgene having a predetermined repertoire may comprise, for example, human V_H
25 genes which are preferentially used in antibody responses to the predetermined antigen type in humans. Alternatively, some V_H genes may be excluded from a defined repertoire for various reasons (e.g., have a low likelihood of encoding high affinity V regions for the predetermined antigen; have a low propensity
30 to undergo somatic mutation and affinity sharpening; or are immunogenic to certain humans).

Thus, prior to rearrangement of a transgene containing various heavy or light chain gene segments, such gene segments may be readily identified, e.g. by hybridization or DNA sequencing, as being from a species of organism other
5 than the transgenic animal.

The transgenic mice of the present invention can be immunized with a predetermined antigen, such as a transmembrane proteins, cell surface macromolecule, or other suitable antigen (e.g., TNF, LPS, etc.) for which a human
10 antibody would be desirable. The mice will produce B cells which undergo class-switching via intratransgene switch recombination (cis-switching) and express immunoglobulins reactive with the predetermined antigen. The immunoglobulins can be human sequence antibodies, wherein the heavy and light
15 chain polypeptides are encoded by human transgene sequences, which may include sequences derived by somatic mutation and V region recombinatorial joints, as well as germline-encoded sequences; these human sequence immunoglobulins can be referred to as being substantially identical to a polypeptide
20 sequence encoded by a human V_L or V_H gene segment and a human J_L or J_H segment, even though other non-germline sequences may be present as a result of somatic mutation and differential V-J and V-D-J recombination joints. With respect to such human sequence antibodies, the variable regions of each chain are
25 typically at least 80 percent encoded by human germline V, J, and, in the case of heavy chains, D, gene segments; frequently at least 85 percent of the variable regions are encoded by human germline sequences present on the transgene; often 90 or 95 percent or more of the variable region sequences are
30 encoded by human germline sequences present on the transgene. However, since non-germline sequences are introduced by somatic mutation and VJ and VDJ joining, the human sequence

antibodies will frequently have some variable region sequences (and less frequently constant region sequences) which are not encoded by human V, D, or J gene segments as found in the human transgene(s) in the germline of the mice. Typically, 5 such non-germline sequences (or individual nucleotide positions) will cluster in or near CDRs, or in regions where somatic mutations are known to cluster.

The human sequence antibodies which bind to the predetermined antigen can result from isotype switching, such 10 that human antibodies comprising a human sequence γ chain (such as $\gamma 1$, $\gamma 2a$, $\gamma 2B$, or $\gamma 3$) and a human sequence light chain (such as K) are produced. Such isotype-switched human sequence antibodies often contain one or more somatic mutation(s), typically in the variable region and often in or 15 within about 10 residues of a CDR) as a result of affinity maturation and selection of B cells by antigen, particularly subsequent to secondary (or subsequent) antigen challenge. These high affinity human sequence antibodies may have binding affinities of at least $1 \times 10^9 \text{ M}^{-1}$, typically at least 20 $5 \times 10^9 \text{ M}^{-1}$, frequently more than $1 \times 10^{10} \text{ M}^{-1}$, and sometimes $5 \times 10^{10} \text{ M}^{-1}$ to 1×10^{11} or greater. Such high affinity human sequence antibodies can be made with high binding affinities for human antigens, such as human CD4 and the like human macromolecules (e.g., such as a human transmembrane or cell 25 surface protein or other cell surface antigen).

The B cells from such mice can be used to generate hybridomas expressing monoclonal high affinity (greater than $2 \times 10^9 \text{ M}^{-1}$) human sequence antibodies against a variety of antigens, including human proteins such as CD4 and the like. 30 These hybridomas can be used to generate a composition comprising an immunoglobulin having an affinity constant (K_a)

of at least $2 \times 10^9 \text{ M}^{-1}$ for binding to a predetermined human antigen, wherein said immunoglobulin consists of:

a human sequence light chain composed of (1) a light chain variable region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human V_L gene segment and a human J_L segment, and (2) a light chain constant region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human C_L gene segment; and

a human sequence heavy chain composed of (1) a heavy chain variable region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human V_H gene segment, optionally a D region, and a human J_H segment, and (2) a constant region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human C_H gene segment.

Often, the human sequence heavy chain and human sequence light chain are separately encoded by a human heavy chain transgene and a human light chain transgene, respectively, which are integrated into a mouse cell genome. However, both chains may be encoded on a single transgene, or one or both chains may be encoded on multiple transgenes, such as a human heavy chain transgene (e.g., HC2) which derived a V gene segment from a YAC containing a V_H array which is not integrated at the same locus as the human heavy chain transgene in the mouse germline.

In one embodiment, the composition has an immunoglobulin which comprises a human sequence light chain having a κ constant region and a human sequence heavy chain having a γ constant region.

The mice (and hybridomas derived therefrom) are a source for an immunoglobulin having an affinity constant (K_a)

of at least $1 \times 10^{10} \text{ M}^{-1}$ for binding to a predetermined human antigen, wherein said immunoglobulin consists of:

a human sequence light chain composed of (1) a light chain variable region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human V_L gene segment and a human J_L segment, and (2) a light chain constant region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human C_L gene segment; and

10 a human sequence heavy chain composed of (1) a heavy chain variable region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human V_H gene segment, optionally a D region, and a human J_H segment, and (2) a constant region having a polypeptide
15 sequence which is substantially identical to a polypeptide sequence encoded by a human C_H gene segment.

The invention provides a transgenic mouse comprising: a homozygous pair of functionally disrupted endogenous heavy chain alleles, a homozygous pair of
20 functionally disrupted endogenous light chain alleles, at least one copy of a heterologous immunoglobulin light chain transgene, and at least one copy of a heterologous immunoglobulin heavy chain transgene, and wherein said animal makes an antibody response following immunization with a human
25 antigen wherein the antibody response comprises an immunoglobulin having an affinity constant (K_a) of at least $2 \times 10^9 \text{ M}^{-1}$ for binding to a predetermined human antigen, wherein said immunoglobulin consists of:

a human sequence light chain composed of (1) a light
30 chain variable region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human V_L gene segment and a human J_L segment, and (2) a light

chain constant region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human C_L gene segment; and

a human sequence heavy chain composed of a (1) a heavy chain variable region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human V_H gene segment, optionally a D region, and a human J_H segment, and (2) a constant region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human C_H gene segment.

Such a transgenic mouse can produce a human sequence immunoglobulin which binds to a human surface or transmembrane protein present on at least one somatic cell type of a human, wherein the immunoglobulin binds said human surface or transmembrane protein with an affinity constant (K_a) of between $1.5 \times 10^9 \text{ M}^{-1}$ and $1.8 \times 10^{10} \text{ M}^{-1}$. One example of such a human surface or transmembrane protein is CD4, although others may be used as immunogens as desired.

The development of high affinity human sequence antibodies against predetermined antigens is facilitated by a method for expanding the repertoire of human variable region gene segments in a transgenic mouse having a genome comprising an integrated human immunoglobulin transgene, said method comprising introducing into the genome a V gene transgene comprising V region gene segments which are not present in said integrated human immunoglobulin transgene. Often, the V region transgene is a yeast artificial chromosome comprising a portion of a human V_H or V_L (V_K) gene segment array, as may naturally occur in a human genome or as may be spliced together separately by recombinant methods, which may include out-of-order or omitted V gene segments. Often at least five or more functional V gene segments are contained on the YAC.

In this variation, it is possible to make a transgenic mouse produced by the V repertoire expansion method, wherein the mouse expresses an immunoglobulin chain comprising a variable region sequence encoded by a V region gene segment present on the V region transgene and a C region encoded on the human Ig transgene. By means of the V repertoire expansion method, transgenic mice having at least 5 distinct V genes can be generated; as can mice containing at least about 24 V genes or more. Of course, some V gene segments may be non-functional (e.g., pseudogenes and the like); these segments may be retained or may be selectively deleted by recombinant methods available to the skilled artisan, if desired.

Once the mouse germline has been engineered to contain a functional YAC having an expanded V segment repertoire, substantially not present in the human Ig transgene containing the J and C gene segments, the trait can be propagated and bred into other genetic backgrounds, including backgrounds where the functional YAC having an expanded V segment repertoire is bred into a mouse germline having a different human Ig transgene. Multiple functional YACs having an expanded V segment repertoire may be bred into a germline to work with a human Ig transgene (or multiple human Ig transgenes). Although referred to herein as YAC transgenes, such transgenes when integrated into the genome may substantially lack yeast sequences, such as sequences required for autonomous replication in yeast; such sequences may optionally be removed by genetic engineering (e.g., restriction digestion and pulsed-field gel electrophoresis or other suitable method) after replication in yeast in no longer necessary (i.e., prior to introduction into a mouse ES cell or mouse prozygote).

The invention also provides a method of propagating the trait of human sequence immunoglobulin expression, comprising breeding a transgenic mouse having the human Ig transgene(s), and optionally also having a functional YAC
5 having an expanded V segment repertoire. Both V_H and V_L gene segments may be present on the YAC. The transgenic mouse may be bred into any background desired by the practitioner, including backgrounds harboring other human transgenes, including human Ig transgenes and/or transgenes encoding other
10 human lymphocyte proteins.

The invention also provides a high affinity human sequence immunoglobulin produced by a transgenic mouse having an expanded V region repertoire YAC transgene.

Although the foregoing describes a preferred
15 embodiment of the transgenic animal of the invention, other embodiments are defined by the disclosure herein and more particularly by the transgenes described in the Examples. Four categories of transgenic animal may be defined:

- I. Transgenic animals containing an unrearranged heavy
20 and rearranged light immunoglobulin transgene.
- II. Transgenic animals containing an unrearranged heavy and unrearranged light immunoglobulin transgene
- III. Transgenic animal containing rearranged heavy and an unrearranged light immunoglobulin transgene, and
- 25 IV. Transgenic animals containing rearranged heavy and rearranged light immunoglobulin transgenes.

Of these categories of transgenic animal, the preferred order of preference is as follows II > I > III > IV where the endogenous light chain genes (or at least the κ
30 gene) have been knocked out by homologous recombination (or other method) and I > II > III > IV where the endogenous light

chain genes have not been knocked out and must be dominated by allelic exclusion.

As is discussed supra, the invention provides human sequence monoclonal antibodies that are useful in treatment of human diseases. Therapeutic uses of monoclonal antibodies are discussed in, e.g., Larrick and Bourla, *Journal of Biological Response Modifiers*, 5:379-393, which is incorporated herein by reference. Uses of human monoclonal antibodies include treatment of autoimmune diseases, cancer, infectious diseases, transplant rejection, blood disorders such as coagulation disorders, and other diseases.

The antibodies of this invention may be administered to patients by any method known in the medical arts for delivery of proteins. Antibodies are particularly suited for parenteral administration (i.e., subcutaneous, intramuscular or intravenous administration). The pharmaceutical compositions of the present invention are suitable for administration using alternative drug delivery approaches as well (see, e.g., Langer, *Science*, 249:1527-1533 (1990)).

Pharmaceutical compositions for parenteral administration usually comprise a solution of a monoclonal antibody dissolved in an acceptable carrier, preferably an aqueous carrier. A variety of aqueous carriers can be used, e.g., water, buffered water, 0.4% saline, 0.3% glycine and the like. These solutions are sterile and generally free of particulate matter. These compositions may be sterilized by conventional, well known sterilization techniques. The compositions may contain pharmaceutically acceptable auxiliary substances as required to approximate physiological conditions such as pH-adjusting and buffering agents, tonicity adjusting agents and the like, for example sodium acetate, sodium chloride, potassium chloride, calcium chloride, sodium

lactate, etc. The concentration of antibody in these formulations can vary widely, i.e., from less than about 0.5%, usually at or at least about 0.1% to as much as 1.5% or 2.0% by weight and will be selected primarily based on fluid
5 volumes, viscosities, etc., in accordance with the particular mode of administration selected. Actual methods for preparing parenterally administrable compositions will be known or apparent to those skilled in the art and are described in more detail in, for example, Remington's Pharmaceutical Sciences,
10 17th Ed., Mack Publishing Company, Easton, Pennsylvania (1985), which is incorporated herein by reference.

The compositions containing the present antibodies or a cocktail thereof can be administered for the prophylactic and/or therapeutic treatments. In therapeutic application,
15 compositions are administered to a patient in an amount sufficient to cure or at least partially arrest the infection and its complications. An amount adequate to accomplish this is defined as a "therapeutically effective dose." Amounts effective for this use generally range from about .05 mg/kg
20 body weight to about 5 mg/kg body weight, preferably between about .2 mg/kg body weight to about 1.5 mg/kg body weight.

In some instances it will be desirable to modify the immunoglobulin molecules of the invention to change their biological activity. For example, the immunoglobulins can be
25 directly or indirectly coupled to other chemotherapeutics agent. A variety of chemotherapeutics can be coupled for targeting. For example, anti-inflammatory agents which may be coupled include immunomodulators, platelet activating factor (PAF) antagonists, cyclooxygenase inhibitors, lipoxygenase
30 inhibitors, and leukotriene antagonists. Some preferred moieties include cyclosporin A, indomethacin, naproxen, FK-506, mycophenolic acid, and the like. Similarly, anti-

oxidants, e.g., superoxide dismutase, are useful in treating reperfusion injury. Likewise, anticancer agents, such as daunomycin, doxorubicin, vinblastine, bleomycin, and the like can be targeted.

5 The monoclonal antibodies of the invention may also be used to target amphipaths (e.g., liposomes) to sites in a patient. In these preparations, the drug to be delivered is incorporated as part of a liposome in which a human monoclonal antibody is embedded.

10 The human-sequence monoclonal antibodies of the invention are useful, in part, because they bind specifically to the predetermined antigen against which they are directed. When the predetermined antigen is a human antigen (i.e., a human protein or fragment thereof), it will sometimes be
15 advantageous if the human immunoglobulin of the invention also binds to the *cognate* antigen found in non-human animals, especially animals that are used frequently for drug testing (e.g., preclinical testing of biological activity, pharmacokinetics and safety). These animals include mice,
20 rabbits, rats, dogs, pigs, and, especially, non-human primates such as chimpanzees, apes and monkeys (e.g., Rhesus monkeys and cynomolgus monkeys). The ability to recognize antigens in experimental animals is particularly useful for determining the effect of specific binding on biodistribution of the
25 immunoglobulins. A cognate antigen is an antigen that (i) has a structure (e.g., amino acid sequence) that is substantially similar to the human antigen (i.e., the amino acid sequence of an animal cognate protein will typically be at least about 50% identical to the human protein, usually at least about 70%
30 identical and often at least about 80% identical or more); (ii) has substantially the same function as the human antigen; and, (iii) often is found in the same cellular compartment as

the human antigen. Human and animal cognate antigens typically (but not always) have the same names. Examples of cognate antigens include human tubulin and mouse tubulin, human CD4 and Rhesus CD4, and human IgG and Rat IgG.

5 An other aspect, the invention provides antigen-binding human mABs comprising at least one polypeptide encoded by an artificial gene. An artificial gene comprises a polypeptide-encoding nucleic acid segment that is synthesized *in vitro* by chemical or enzymatic methods that do not require
10 a cell-derived template nucleic acid strand (e.g., a nucleic acid template obtained from a bacterial cell or an immune or hybridoma cell) and the progeny (through replication) of the artificial gene, i.e., a wholly synthetic nucleic acid.

 Although it is routine in genetic engineering to use
15 short synthetic nucleic acids as primers, linkers and the like, it is also possible by chemical and/or enzymatic means to produce wholly synthetic protein-coding nucleic acids that are 30, 50, or more bases in length. The artificial genes of the invention may include both synthetic nucleic acid regions
20 and cell-derived nucleic acid regions. The synthetic nucleic acid region of the artificial gene will generally be at least about 50 bases in length, often at least about 100 bases, typically at least about 200 bases, more often at least about 250 bases and usually over 300 bases or 400 bases in length.
25 Typically the synthetic nucleic acid regions will encode variable gene segments or a portion thereof, e.g., CDR regions, and the constant regions will be encoded by cell-derived nucleic acids. Immunoglobulin polypeptides (i.e., immunoglobulin heavy chains and immunoglobulin light chains)
30 can be conveniently expressed using artificial genes that encode the polypeptides. Usually the artificial genes are operably linked to transcription promoter sequences, e.g.,

promoter sequences derived from immunoglobulin genes or from viruses (e.g., SV40, CMV, HIV, RSV) or hybrid promoters. The artificial gene may be linked to other sequences as well, e.g. polyadenylation sequences and introns. One method for

5 expressing an immunoglobulin polypeptide involves insertion of a synthetic nucleic acid encoding one region of an immunoglobulin polypeptide (e.g., a variable region or portion thereof) into a vector that encodes the remaining segments or parts of the immunoglobulin chain (e.g., a μ , γ , γ_2 , γ_3 , γ_4 ,

10 δ , ϵ , α_1 or α_2 constant region) and, optionally, promoter (e.g., a CMV (cytomegalovirus) promoter), polyadenylation or other sequences. Such vectors are constructed so that upon introduction into a cell, the cellular transcription and translation of the vector sequences results in an immunoglobulin

15 polypeptide.

Functional human sequence immunoglobulin heavy and light chain genes and polypeptides can be constructed using artificial genes, and used to produce immunoglobulins with a desired specificity such as specific binding to a

20 predetermined antigen. This is accomplished by constructing an artificial gene that encodes an immunoglobulin polypeptide substantially similar to a polypeptide expressed by a cell from, or a hybridoma derived from, a transgenic animal immunized with the predetermined antigen. Thus, the invention

25 provides artificial genes encoding immunoglobulin polypeptides and methods for producing a human-sequence immunoglobulin using an artificial gene(s).

According to this method, a transgenic animal (e.g., a transgenic mouse with a homozygous pair of functionally

30 disrupted endogenous heavy chain alleles, a homozygous pair of functionally disrupted endogenous light chain alleles, at least one copy of a human immunoglobulin light chain

transgene, and at least one copy of a human immunoglobulin heavy chain transgene) is immunized with predetermined antigen, e.g., a human protein. Nucleic acid, preferably mRNA, is then collected or isolated from a cell or population
5 of cells in which immunoglobulin gene rearrangement has taken place, and the sequence(s) of nucleic acids encoding the heavy and/or light chains (especially the V segments) of immunoglobulins, or a portion thereof, is determined. This sequence information is used as a basis for the sequence of
10 the artificial gene.

Sequence determination will generally require isolation of at least a portion of the gene or cDNA of interest, e.g., a portion of a rearranged human transgene or corresponding cDNA encoding an immunoglobulin polypeptide.
15 Usually this requires cloning the DNA or, preferably, mRNA (i.e., cDNA) encoding the human immunoglobulin polypeptide. Cloning is carried out using standard techniques (see, e.g., Sambrook et al. (1989) *Molecular Cloning: A Laboratory Guide*, Vols 1-3, Cold Spring Harbor Press, which is incorporated
20 herein by reference). For example, a cDNA library may be constructed by reverse transcription of polyA⁺ mRNA, preferably membrane-associated mRNA, and the library screened using probes specific for human immunoglobulin polypeptide gene sequences. In a preferred embodiment, however, the
25 polymerase chain reaction (PCR) is used to amplify cDNAs (or portions of full-length cDNAs) encoding an immunoglobulin gene segment of interest (e.g., a light chain variable segment). Because the sequences of the human immunoglobulin polypeptide genes are readily available to those of skill, probes or PCR
30 primers that will specifically hybridize to or amplify a human immunoglobulin gene or segment thereof can be easily designed. See, e.g., Taylor et al., *Nuc. Acids. Res.*, 20:6287 (1992)

which is incorporated by reference. Moreover, the sequences of the human transgene of the transgenic mouse will often be known to the practitioner, and primer sequences can be chosen that hybridize to appropriate regions of the transgene. The amplified sequences can be readily cloned into any suitable vector, e.g., expression vectors, minigene vectors, or phage display vectors. It will be appreciated that the particular method of cloning used not critical, so long as it is possible to determine the sequence of some portion of the immunoglobulin polypeptide of interest. As used herein, a nucleic acid that is cloned, amplified, tagged, or otherwise distinguished from background nucleic acids such that the sequence of the nucleic acid of interest can be determined, is considered isolated.

One source for RNA used for cloning and sequencing is a hybridoma produced by obtaining a B cell from the transgenic mouse and fusing the B cell to an immortal cell. An advantage of using hybridomas is that they can be easily screened, and a hybridoma that produces a human monoclonal antibody of interest selected. Alternatively, RNA can be isolated from B cells (or whole spleen) of the immunized animal. When sources other than hybridomas are used, it may be desirable to screen for sequences encoding immunoglobulins or immunoglobulin polypeptides with specific binding characteristics. One method for such screening is the use of phage display technology. Phage display is described in e.g., Dower et al., WO 91/17271, McCafferty et al., WO 92/01047, and Caton and Koprowski, *Proc. Natl. Acad. Sci. USA*, 87:6450-6454 (1990), each of which is incorporated herein by reference. In one embodiment using phage display technology, cDNA from an immunized transgenic mouse (e.g., total spleen cDNA) is isolated, the polymerase chain reaction is used to amplify a

cDNA sequences that encode a portion of an immunoglobulin polypeptide, e.g., CDR regions, and the amplified sequences are inserted into a phage vector. cDNAs encoding peptides of interest, e.g., variable region peptides with desired binding characteristics, are identified by standard techniques such as panning.

The sequence of the amplified or cloned nucleic acid is then determined. Typically the sequence encoding an entire variable region of the immunoglobulin polypeptide is determined, however, it will sometimes be adequate to sequence only a portion of a variable region, for example, the CDR-encoding portion. Typically the portion sequenced will be at least 30 bases in length, more often based coding for at least about one-third or at least about one-half of the length of the variable region will be sequenced.

Sequencing can be carried on clones isolated from a cDNA library, or, when PCR is used, after subcloning the amplified sequence or by direct PCR sequencing of the amplified segment. Sequencing is carried out using standard techniques (see, e.g., Sambrook et al. (1989) *Molecular Cloning: A Laboratory Guide*, Vols 1-3, Cold Spring Harbor Press, and Sanger, F. et al. (1977) *Proc. Natl. Acad. Sci. USA* 74: 5463-5467, which is incorporated herein by reference). By comparing the sequence of the cloned nucleic acid with published sequences of human immunoglobulin genes and cDNAs, one of skill will readily be able to determine, depending on the region sequenced, (i) the germline segment usage of the hybridoma immunoglobulin polypeptide (including the isotype of the heavy chain) and (ii) the sequence of the heavy and light chain variable regions, including sequences resulting from N-region addition and the process of somatic mutation. One source of immunoglobulin gene sequence information is the

National Center for Biotechnology Information, National Library of Medicine, National Institutes of Health, Bethesda, Md.

In an alternative embodiment, the amino acid
5 sequence of an immunoglobulin of interest may be determined by direct protein sequencing.

An artificial gene can be constructed that has a sequence identical to or substantially similar to, at least a portion of the immunoglobulin-expressing gene (i.e.,
10 rearranged transgene). Similarly, the artificial gene can encode an polypeptide that is identical or has substantial similarity to a polypeptide encoded by the sequenced portion of the rearranged transgene. The degeneracy of the genetic code allows the same polypeptide to be encoded by multiple
15 nucleic acid sequences. It is sometimes desirable to change the nucleic acid sequence, for example to introduce restriction sites, change codon usage to reflect a particular expression system, or to remove a glycosylation site. In addition, changes in the hybridoma sequences may be introduced
20 to change the characteristics (e.g., binding characteristics) of the immunoglobulin. For example, changes may be introduced, especially in the CDR regions of the heavy and light chain variable regions, to increase the affinity of the immunoglobulin for the predetermined antigen.

25 Methods for constructing an synthetic nucleic acids are well known. An entirely chemical synthesis is possible but in general, a mixed chemical-enzymatic synthesis is carried out in which chemically synthesized oligonucleotides are used in ligation reactions and/or in the polymerase chain
30 reaction to create longer polynucleotides. In a most preferred embodiment, the polymerase chain reaction is carried out using overlapping primers chosen so that the result of the

amplification is a DNA with the sequence desired for the artificial gene. The oligonucleotides of the present invention may be synthesized in solid phase or in solution. Generally, solid phase synthesis is preferred. Detailed
5 descriptions of the procedures for solid phase synthesis of oligonucleotides by phosphite-triester, phosphotriester, and H-phosphonate chemistries are widely available. See, for example, Itakura, U.S. Pat. No. 4,401,796; Caruthers et al., U.S. Pat. Nos. 4,458,066 and 4,500,707; Beaucage et al.,
10 *Tetrahedron Lett.*, 22:1859-1862; Matteucci et al., *J. Amer. Chem. Soc.*, 103:3185-3191 (1981); Caruthers et al., *Genetic Engineering*, 4:1-17 (1982); Jones, chapter 2, Atkinson et al., chapter 3, and Sproat et al., chapter 4, in Gait, ed. *Oligonucleotide Synthesis: A Practical Approach*, IRL Press,
15 Washington, D.C. (1984); Froehler et al., *Tetrahedron Lett.*, 27:469-472 (1986); Froehler et al., *Nucleic Acids Res.*, 14:5399-5407 (1986); Sinha et al., *Tetrahedron Lett.*, 24:5843-5846 (1983); and Sinha et al., *Nucleic Acids Res.*, 12:4539-4557 (1984) which are incorporated herein by reference.
20 The artificial gene can introduced into a cell and expressed to produce an immunoglobulin polypeptide. The choice of cell type for expression will depend on many factors (e.g., the level of protein glycosylation desired), but cells capable of secreting human immunoglobulins will be preferred.
25 Especially preferred cells include CHO cells and myeloma-derived cells such as the SP20 and NS0 cell lines. Standard cell culture are well known and are also described in Newman, et al., *Biotechnology*, 10:1455-1460 (1992); Bebbington, et al., *Biotechnology*, 10:169-175 (1992); Cockett, et al.,
30 *Biotechnology*, 8:662-667 (1990); Carter, et al., *Biotechnology*, 10:163-167 (1992), each of which is incorporated herein by reference. Methods for introduction of

nucleic acids, e.g., an artificial gene, are well known and include transfection (e.g., by electroporation or liposome-mediated) and transformation. Systems for expression of introduced genes are described generally in Sambrook et al.,
5 *supra*.

It is often desirable to express two immunoglobulin polypeptides (i.e., a heavy chain and a light chain) in the same cell so that an immunoglobulin (e.g., an IgG molecule) is produced *in vivo*. Accordingly it will sometimes be desirable
10 to introduce two artificial genes (i.e., one encoding a heavy chain and one encoding a light chain) into a cell. (The two artificial genes can be introduced on a single vector). Alternatively, one artificial gene encoding one immunoglobulin polypeptide can be introduced into a cell that has been
15 genetically engineered to express the other immunoglobulin polypeptide.

It will be apparent that as the cells into which the artificial gene is transfected propagate, the wholly synthetic nucleic acid portion of the artificial gene, will act as a
20 template for replication and transcription. Nonetheless, the progeny genes will have originated from a synthetic nucleic acid (i.e., a polypeptide-encoding nucleic acid molecule that is synthesized *in vitro* by chemical or enzymatic methods that do not require a cell-derived template nucleic acid strand)
25 and as used herein, are also considered artificial genes. Thus, the relationship of the synthetic portion of the artificial gene to the expressed transgene of the hybridoma is one in which there is an *informational* link (i.e., sequence information) but no direct *physical* link.

30 The invention also provides anti-CD4 monoclonal antibodies useful in therapeutic and diagnostic applications, especially the treatment of human disease. CD4 is a cell

surface protein that is expressed primarily on thymocytes and T cells, and which is involved in T-cell function and MHC Class II recognition of antigen. Antibodies directed against CD4 act to reduce the activity of CD4 cells and thus reduce
5 undesirable autoimmune reactions, inflammatory responses and rejection of transplanted organs.

The ability of a human anti-CD4 mAb to inhibit a T-helper cell dependent immune response in primates can be demonstrated by immunizing the primate with a soluble foreign
10 antigen (e.g., tetanus toxoid (TT)) and measuring the ability of the primate to mount a delayed-type hypersensitivity reaction (DTH) to the antigen (e.g., following injection of the human mAb). The DTH is mediated by CD4⁺ (T-helper) cells (E. Benjamin and S. Lescowitz, Immunology: A Short Course,
15 Second Edition, (1991) Wiley-Liss, Inc., New York, pp. 277-292). Antigen-specific T-helper cells recognize the processed antigen presented by MHC Class II molecules on antigen-presenting cells and become activated. The activated T-helper cells secrete a variety of lymphokines (IL2, INF γ , TNF β , MCF)
20 and thus attract and activate macrophages and T-cytotoxic cells at the injection site. Although most of the effector functions occurring as part of the DTH are performed by macrophages and T-cytotoxic cells, it is the T-helper cells which initiate the response. Therefore, if the T-helper cells
25 can be inhibited, there will be no DTH. Administration of anti-CD4 mABs has been shown to prevent (Wofsy, et al., *J. Exp. Med.*, 161:378-391 (1985)) or reverse (Wofsy, et al., *J. Immunol.*, 138:3247-3253 (1987), Waldor, et al., *Science*, 227:415-417 (1985)) autoimmune disease in animal models.
30 Administration of murine or chimeric anti-CD4 mAbs to patients with rheumatoid arthritis has shown evidence of clinical benefit (Knox, et al., *Blood*, 77:20-30 (1991); Goldbery, et

al., *J. Autoimmunity*, 4:617-630; Herzog, et al., *Lancet*,
ii:1461-1462 ; Horneff, et al., *Arthritis Rheum.*, 34:129-140;
Reiter, et al., *Arthritis Rheum.*, 34:525-536; Wending, et al.,
J. Rheum., 18:325-327; Van der Lubbe, et al., *Arthritis*
5 *Rheum.*, 38:1097-1106; Van der Lubbe, et al., *Arthritis Rheum.*,
36:1375-1379; Moreland, et al., *Arthritis Rheum.*, 36:307-318,
and Choy, et al., *Arthritis and Rheumatism*, 39(1):52-56
(1996); all of which is incorporated herein by reference). In
addition, as noted above, a chimeric anti-CD4 mAB has shown
10 some clinical efficacy in patients with mycosis fungoides
(Knox et al. (1991) *Blood* 77:20; which is incorporated herein
by reference). Anti-CD4 antibodies are also discussed in
Newman, et al., *Biotechnology*, 10:1455-1460 (1992), which is
incorporated herein by reference.

15 The invention also provides anti-interleukin-8
monoclonal antibodies useful in therapeutic and diagnostic
applications, especially the treatment of human diseases.
Interleukin-8 (IL8), a very potent and mostly specific
chemoattractant for neutrophils, is thought to play an
20 important role in the inflammatory response. IL8 also induces
angiogenesis, mediates transendothelial neutrophil migration
and contributes to other inflammatory responses. The
properties of IL8 and related cytokines are discussed in
Baggiolini et al., 1994, *Adv. Immunol.* 55:97-179, which is
25 incorporated herein by reference.

 IL8 has been shown to bind to, and activate,
neutrophils and to induce neutrophil chemotaxis through an
endothelial cell layer *in vitro*. The role of IL8 in inducing
neutrophil transmigration from the vasculature to a site of
30 inflammation has been demonstrated *in vivo* as well. Moreover,
inhibition of the IL8 in those circumstances has prevented
tissue damage resulting from neutrophil recruitment. Anti-

rabbit IL8 antibodies can prevent lung reperfusion injuries resulting from ischemia (Sekido et al., 1993, *Nature* 365:654-7). Anti-rabbit IL8 antibodies can also prevent lung damage resulting from endotoxin-induced pleurisy in rabbits (Broaddus et al., 1994, *J. Immunol.* 152:2960-7). In vivo primate models are also suitable for determining the effects of anti-human IL8 mAbs on migration of neutrophils from the vasculature to the inflammatory site. It has been shown that intradermal injection of rhesus monkeys with endotoxin upregulates IL8 expression at the injection site and ultimately results in neutrophil localization (Silber et al., 1994, *Lab. Invest.* 70:163-75).

Anti-IL8 antibodies have also been shown to reduce tissue damage and prolong survival in animal models of acute inflammation including adult respiratory distress syndrome (ARDS) and acid induced lung injury (Sekido et al., 1993, *Nature* 365:654-7; Mulligan et al., 1993, *J. Immunol.* 150:5585-95; Broaddus et al., 1994, *J. Immunol.* 152:2960-7, all of which are incorporated herein by reference). Consistent with the role of IL-8 in inflammation, human anti-IL-8 monoclonal antibodies can be used for treatment of a variety of conditions caused or aggravated by an inflammatory response, including reperfusion injuries (especially to the lung and heart), vasculitis, septic shock, autoimmune diseases (including glomerulonephritis, inflammatory bowel disease, rheumatoid arthritis and psoriasis), allergic reactions (e.g., asthma) and cystic fibrosis.

Two distinct IL8 receptors, IL8RA and IL8RB, have been identified (Holmes et al., 1991, *Science* 253:1278-80; Murphy et al., 1991, *Science* 253:1280-3). Both receptors bind IL8, but only IL8RB binds other CXC chemokines. Both

receptors are found in approximately equal numbers on neutrophils and some lymphocytes.

EXPERIMENTAL EXAMPLES

5 METHODS AND MATERIALS

Transgenic mice are derived according to Hogan, et al., "Manipulating the Mouse Embryo: A Laboratory Manual", Cold Spring Harbor Laboratory, which is incorporated herein by reference.

10 Embryonic stem cells are manipulated according to published procedures (Teratocarcinomas and embryonic stem cells: a practical approach, E.J. Robertson, ed., IRL Press, Washington, D.C., 1987; Zijlstra et al., Nature 342:435-438 (1989); and Schwartzberg et al., Science 246:799-803 (1989),
15 each of which is incorporated herein by reference).

DNA cloning procedures are carried out according to J. Sambrook, et al. in Molecular Cloning: A Laboratory Manual, 2d ed., 1989, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., which is incorporated herein by
20 reference.

Oligonucleotides are synthesized on an Applied Bio Systems oligonucleotide synthesizer according to specifications provided by the manufacturer.

Hybridoma cells and antibodies are manipulated
25 according to "Antibodies: A Laboratory Manual", Ed Harlow and David Lane, Cold Spring Harbor Laboratory (1988), which is incorporated herein by reference.

EXAMPLE 1

30 Genomic Heavy Chain Human Ig Transgene

This Example describes the cloning and microinjection of a human genomic heavy chain immunoglobulin transgene which is microinjected into a murine zygote.

Nuclei are isolated from fresh human placental tissue as described by Marzluff et al., "Transcription and Translation: A Practical Approach", B.D. Hammes and S.J. Higgins, eds., pp. 89-129, IRL Press, Oxford (1985)). The isolated nuclei (or PBS washed human spermatocytes) are embedded in a low melting point agarose matrix and lysed with EDTA and proteinase K to expose high molecular weight DNA, which is then digested in the agarose with the restriction enzyme NotI as described by M. Finney in Current Protocols in Molecular Biology (F. Ausubel, et al., eds. John Wiley & Sons, Supp. 4, 1988, Section 2.5.1).

The NotI digested DNA is then fractionated by pulsed field gel electrophoresis as described by Anand et al., Nucl. Acids Res. 17:3425-3433 (1989). Fractions enriched for the NotI fragment are assayed by Southern hybridization to detect one or more of the sequences encoded by this fragment. Such sequences include the heavy chain D segments, J segments, μ and $\gamma 1$ constant regions together with representatives of all 6 VH families (although this fragment is identified as 670 kb fragment from HeLa cells by Berman et al. (1988), supra., we have found it to be as 830 kb fragment from human placental and sperm DNA). Those fractions containing this NotI fragment (see Fig. 4) are pooled and cloned into the NotI site of the vector pYACNN in Yeast cells. Plasmid pYACNN is prepared by digestion of pYAC-4 Neo (Cook et al., Nucleic Acids Res. 16: 11817 (1988)) with EcoRI and ligation in the presence of the oligonucleotide 5' - AAT TGC GGC CGC - 3'.

YAC clones containing the heavy chain NotI fragment are isolated as described by Brownstein et al., Science

WO 98/24884

PCT/US97/21803

121

244:1348-1351 (1989), and Green et al., Proc. Natl. Acad. Sci. USA 87:1213-1217 (1990), which are incorporated herein by reference. The cloned NotI insert is isolated from high molecular weight yeast DNA by pulse field gel electrophoresis as described by M. Finney, op cit. The DNA is condensed by the addition of 1 mM spermine and microinjected directly into the nucleus of single cell embryos previously described.

EXAMPLE 2

10 Genomic κ Light Chain Human Ig Transgene
 Formed by In Vivo Homologous Recombination

A map of the human κ light chain has been described in Lorenz et al., Nucl. Acids Res. 15:9667-9677 (1987), which is incorporated herein by reference.

A 450 kb XhoI to NotI fragment that includes all of C_{κ} , the 3' enhancer, all J segments, and at least five different V segments is isolated and microinjected into the nucleus of single cell embryos as described in Example 1.

20

EXAMPLE 3

25 Genomic κ Light Chain Human Ig Transgene
 Formed by In Vivo Homologous Recombination

A 750 kb MluI to NotI fragment that includes all of the above plus at least 20 more V segments is isolated as described in Example 1 and digested with BssHII to produce a fragment of about 400 kb.

30

The 450 kb XhoI to NotI fragment plus the approximately 400 kb MluI to BssHII fragment have sequence overlap defined by the BssHII and XhoI restriction sites. Homologous recombination of these two fragments upon microinjection of a mouse zygote results in a transgene

containing at least an additional 15-20 V segments over that found in the 450 kb XhoI/NotI fragment (Example 2).

EXAMPLE 4

5 Construction of Heavy Chain Mini-Locus

A. Construction of pGP1 and pGP2

pBR322 is digested with EcoRI and StyI and ligated with the following oligonucleotides to generate pGP1 which contains a 147 base pair insert containing the restriction
10 sites shown in Fig. 8. The general overlapping of these oligos is also shown in Fig. 9.

The oligonucleotides are:

oligo-1 5' - CTT GAG CCC GCC TAA TGA GCG GGC TTT
15 TTT TTG CAT ACT GCG GCC - 3'

oligo-2 5' - GCA ATG GCC TGG ATC CAT GGC GCG CTA
 GCA TCG ATA TCT AGA GCT CGA GCA -3'

20 oligo-3 5' - TGC AGA TCT GAA TTC CCG GGT ACC AAG
 CTT ACG CGT ACT AGT GCG GCC GCT -3'

oligo-4 5' - AAT TAG CGG CCG CAC TAG TAC GCG TAA
 GCT TGG TAC CCG GGA ATT - 3'

25 oligo-5 5' - CAG ATC TGC ATG CTC GAG CTC TAG ATA
 TCG ATG CTA GCG CGC CAT GGA TCC - 3'

oligo-6 5' - AGG CCA TTG CGG CCG CAG TAT GCA AAA
30 AAA AGC CCG CTC ATT AGG CGG GCT - 3'

This plasmid contains a large polylinker flanked by rare cutting NotI sites for building large inserts that can be isolated from vector sequences for microinjection. The
35 plasmid is based on pBR322 which is relatively low copy compared to the pUC based plasmids (pGP1 retains the pBR322 copy number control region near the origin of replication). Low copy number reduces the potential toxicity of insert

sequences. In addition, pGP1 contains a strong transcription terminator sequence derived from trpA (Christie et al., Proc. Natl. Acad. Sci. USA 78:4180 (1981)) inserted between the ampicillin resistance gene and the polylinker. This further
5 reduces the toxicity associated with certain inserts by preventing readthrough transcription coming from the ampicillin promoters.

Plasmid pGP2 is derived from pGP1 to introduce an additional restriction site (SfiI) in the polylinker. pGP1 is
10 digested with MluI and SpeI to cut the recognition sequences in the polylinker portion of the plasmid.

The following adapter oligonucleotides are ligated to the thus digested pGP1 to form pGP2.

15 5' CGC GTG GCC GCA ATG GCC A 3'
 5' CTA GTG GCC ATT GCG GCC A 3'

pGP2 is identical to pGP1 except that it contains an additional Sfi I site located between the MluI and SpeI sites.
20 This allows inserts to be completely excised with SfiI as well as with NotI.

B. Construction of pRE3 (rat enhancer 3')

An enhancer sequence located downstream of the rat
25 constant region is included in the heavy chain constructs.

The heavy chain region 3' enhancer described by Petterson et al., Nature 344:165-168 (1990), which is incorporated herein by reference) is isolated and cloned. The rat IGH 3' enhancer sequence is PCR amplified by using the
30 following oligonucleotides:

5' CAG GAT CCA GAT ATC AGT ACC TGA AAC AGG GCT TGC 3'

5' GAG CAT GCA CAG GAC CTG GAG CAC ACA CAG CCT TCC 3'

5 The thus formed double stranded DNA encoding the 3' enhancer is cut with BamHI and SphI and clone into BamHI/SphI cut pGP2 to yield pRE3 (rat enhancer 3').

C. Cloning of Human J- μ Region

10 A substantial portion of this region is cloned by combining two or more fragments isolated from phage lambda inserts. See Fig. 9.

A 6.3 kb BamHI/HindIII fragment that includes all human J segments (Matsuda et al., EMBO J., 7:1047-1051 (1988);
15 Ravetech et al. m Cell, 27:583-591 (1981), which are incorporated herein by reference) is isolated from human genomic DNA library using the oligonucleotide GGA CTG TGT CCC TGT GTG ATG CTT TTG ATG TCT GGG GCC AAG.

An adjacent 10 kb HindIII/BamII fragment that
20 contains enhancer, switch and constant region coding exons (Yasui et al., Eur. J. Immunol., 19:1399-1403 (1989)) is similarly isolated using the oligonucleotide:

CAC CAA GTT GAC CTG CCT GGT CAC AGA CCT GAC CAC CTA TGA

An adjacent 3' 1.5 kb BamHI fragment is similarly
25 isolated using clone pMUM insert as probe (pMUM is 4 kb EcoRI/HindIII fragment isolated from human genomic DNA library with oligonucleotide:

CCT GTG GAC CAC CGC CTC CAC CTT CAT

CGT CCT CTT CCT CCT

30 mu membrane exon 1) and cloned into pUC19.

pGP1 is digested with BamHI and BglII followed by treatment with calf intestinal alkaline phosphatase.

Fragments (a) and (b) from Fig. 9 are cloned in the digested pGP1. A clone is then isolated which is oriented such that 5' BamHI site is destroyed by BamHI/Bgl fusion. It is identified as pMU (see Fig. 10). pMU is digested with BamHI and fragment (c) from Fig. 9 is inserted. The orientation is checked with HindIII digest. The resultant plasmid pHIG1 (Fig. 10) contains an 18 kb insert encoding J and C μ segments.

10 D. Cloning of C μ Region

pGP1 is digested with BamHI and HindIII is followed by treatment with calf intestinal alkaline phosphatase (Fig. 14). The so treated fragment (b) of Fig. 14 and fragment (c) of Fig. 14 are cloned into the BamHI/HindIII cut pGP1. Proper orientation of fragment (c) is checked by HindIII digestion to form pCON1 containing a 12 kb insert encoding the C μ region.

Whereas pHIG1 contains J segments, switch and μ sequences in its 18 kb insert with an SfiI 3' site and a SpeI 5' site in a polylinker flanked by NotI sites, will be used for rearranged VDJ segments. pCON1 is identical except that it lacks the J region and contains only a 12 kb insert. The use of pCON1 in the construction of fragment containing rearranged VDJ segments will be described hereinafter.

25 E. Cloning of γ -1 Constant Region (pREG2)

The cloning of the human γ -1 region is depicted in Fig. 16.

Yamamura et al., Proc. Natl. Acad. Sci. USA 83:2152-2156 (1986) reported the expression of membrane bound human γ -1 from a transgene construct that had been partially deleted on integration. Their results indicate that the 3' BamHI site delineates a sequence that includes the

transmembrane rearranged and switched copy of the gamma gene with a V-C intron of less than 5kb. Therefore, in the unrearranged, unswitched gene, the entire switch region is included in a sequence beginning less than 5 kb from the 5' end of the first γ -1 constant exon. Therefore it is included in the 5' 5.3 kb HindIII fragment (Ellison et al., Nucleic Acids Res. 10:4071-4079 (1982), which is incorporated herein by reference). Takahashi et al., Cell 29: 671-679 (1982), which is incorporated herein by reference, also reports that this fragment contains the switch sequence, and this fragment together with the 7.7 kb HindIII to BamHI fragment must include all of the sequences we need for the transgene construct. An intronic sequence is a nucleotide sequence of at least 15 contiguous nucleotides that occurs in an intron of a specified gene.

Phage clones containing the γ -1 region are identified and isolated using the following oligonucleotide which is specific for the third exon of γ -I (CH3).

5' TGA GCC ACG AAG ACC CTG AGG
TCA AGT TCA ACT GGT ACG TGG 3'

A 7.7 kb HindIII to BglII fragment (fragment (a) in Fig. 11) is cloned into HindIII/BglII cut pRE3 to form pREG1. The upstream 5.3 kb HindIII fragment (fragment (b) in Fig. 11) is cloned into HindIII digested pREG1 to form pREG2. Correct orientation is confirmed by BamHI/SpeI digestion.

F. Combining C_γ and C_μ

The previously described plasmid pHIG1 contains human J segments and the C_μ constant region exons. To provide a transgene containing the C_μ constant region gene segments,

pHIG1 was digested with SfiI (Fig. 10). The plasmid pREG2 was also digested with SfiI to produce a 13.5 kb insert containing human Cy exons and the rat 3' enhancer sequence. These sequences were combined to produce the plasmid pHIG3' (Fig.

5 12) containing the human J segments, the human C μ constant region, the human Cy1 constant region and the rat 3' enhancer contained on a 31.5 kb insert.

A second plasmid encoding human C μ and human Cy1 without J segments is constructed by digesting pCON1 with SfiI and combining that with the SfiI fragment containing the human Cy region and the rat 3' enhancer by digesting pREG2 with SfiI. The resultant plasmid, pCON (Fig. 12) contains a 26 kb NotI/SpeI insert containing human C μ , human γ 1 and the rat 3' enhancer sequence.

15

G. Cloning of D Segment

The strategy for cloning the human D segments is depicted in Fig. 13. Phage clones from the human genomic library containing D segments are identified and isolated using probes specific for diversity region sequences (Ichihara et al., EMBO J. 7:4141-4150 (1988)). The following oligonucleotides are used:

DXP1: 5' - TGG TAT TAC TAT GGT TCG GGG AGT TAT TAT
25 AAC CAC AGT GTC - 3'

DXP4: 5' - GCC TGA AAT GGA GCC TCA GGG CAC AGT GGG
CAC GGA CAC TGT - 3'

30 DN4: 5' - GCA GGG AGG ACA TGT TTA GGA TCT GAG GCC
GCA CCT GAC ACC - 3'

A 5.2 kb XhoI fragment (fragment (b) in Fig. 13) containing DLR1, DXP1, DXP'1, and DA1 is isolated from a phage clone identified with oligo DXP1.

A 3.2 kb XbaI fragment (fragment (c) in Fig. 13) containing DXP4, DA4 and DK4 is isolated from a phage clone identified with oligo DXP4.

Fragments (b), (c) and (d) from Fig. 13 are combined and cloned into the XbaI/XhoI site of pGP1 to form pHIG2 which contains a 10.6 kb insert.

10 This cloning is performed sequentially. First, the 5.2 kb fragment (b) in Fig. 13 and the 2.2 kb fragment (d) of Fig. 13 are treated with calf intestinal alkaline phosphatase and cloned into pGP1 digested with XhoI and XbaI. The resultant clones are screened with the 5.2 and 2.2 kb insert.
15 Half of those clones testing positive with the 5.2 and 2.2 kb inserts have the 5.2 kb insert in the proper orientation as determined by BamHI digestion. The 3.2 kb XbaI fragment from Fig. 13 is then cloned into this intermediate plasmid containing fragments (b) and (d) to form pHIG2. This plasmid
20 contains diversity segments cloned into the polylinker with a unique 5' SfiI site and unique 3' SpeI site. The entire polylinker is flanked by NotI sites.

H. Construction of Heavy Chain Minilocus

25 The following describes the construction of a human heavy chain mini-locus which contain one or more V segments.

An unrearranged V segment corresponding to that identified as the V segment contained in the hybridoma of Newkirk et al., J. Clin. Invest. 81:1511-1518 (1988), which
30 is incorporated herein by reference, is isolated using the following oligonucleotide:

5' - GAT CCT GGT TTA GTT AAA GAG GAT TTT
ATT CAC CCC TGT GTC - 3'

A restriction map of the unrearranged V segment is
5 determined to identify unique restriction sites which provide
upon digestion a DNA fragment having a length approximately 2
kb containing the unrearranged V segment together with 5' and
3' flanking sequences. The 5' prime sequences will include
promoter and other regulatory sequences whereas the 3'
10 flanking sequence provides recombination sequences necessary
for V-DJ joining. This approximately 3.0 kb V segment insert
is cloned into the polylinker of pGB2 to form pVH1.

pVH1 is digested with SfiI and the resultant
fragment is cloned into the SfiI site of pHIG2 to form a
15 pHIG5'. Since pHIG2 contains D segments only, the resultant
pHIG5' plasmid contains a single V segment together with D
segments. The size of the insert contained in pHIG5 is 10.6
kb plus the size of the V segment insert.

The insert from pHIG5 is excised by digestion with
20 NotI and SpeI and isolated. pHIG3' which contains J, C μ and
c γ 1 segments is digested with SpeI and NotI and the 3' kb
fragment containing such sequences and the rat 3' enhancer
sequence is isolated. These two fragments are combined and
ligated into NotI digested pGP1 to produce pHIG which contains
25 insert encoding a V segment, nine D segments, six functional J
segments, C μ , C γ and the rat 3' enhancer. The size of this
insert is approximately 43 kb plus the size of the V segment
insert.

30 I. Construction of Heavy Chain Minilocus
by Homologous Recombination

As indicated in the previous section, the insert of pHIG is approximately 43 to 45 kb when a single V segment is employed. This insert size is at or near the limit of that which may be readily cloned into plasmid vectors. In order to provide for the use of a greater number of V segments, the following describes in vivo homologous recombination of overlapping DNA fragments which upon homologous recombination within a zygote or ES cell form a transgene containing the rat 3' enhancer sequence, the human C μ , the human C γ 1, human J segments, human D segments and a multiplicity of human V segments.

A 6.3 kb BamHI/HindIII fragment containing human J segments (see fragment (a) in Fig. 9) is cloned into MluI/SpeI digested pHIG5' using the following adapters:

5' GAT CCA AGC AGT 3'

5' CTA GAC TGC TTG 3'

5' CGC GTC GAA CTA 3'

5' AGC TTA GTT CGA 3'

The resultant is plasmid designated pHIG5'0 (overlap). The insert contained in this plasmid contains human V, D and J segments. When the single V segment from pVH1 is used, the size of this insert is approximately 17 kb plus 2 kb. This insert is isolated and combined with the insert from pHIG3' which contains the human J, C μ , γ 1 and rat 3' enhancer sequences. Both inserts contain human J segments which provide for approximately 6.3 kb of overlap between the two DNA fragments. When coinjected into the mouse zygote, in vivo homologous recombination occurs generating a transgene equivalent to the insert contained in pHIG.

This approach provides for the addition of a multiplicity of V segments into the transgene formed in vivo. For example, instead of incorporating a single V segment into pHIG5', a multiplicity of V segments contained on (1) isolated genomic DNA, (2) ligated DNA derived from genomic DNA, or (3) DNA encoding a synthetic V segment repertoire is cloned into pHIG2 at the SfiI site to generate pHIG5' V_N. The J segments fragment (a) of Fig. 9 is then cloned into pHIG5' V_N and the insert isolated. This insert now contains a multiplicity of V segments and J segments which overlap with the J segments contained on the insert isolated from pHIG3'. When cointrduced into the nucleus of a mouse zygote, homologous recombination occurs to generate in vivo the transgene encoding multiple V segments and multiple J segments, multiple D segments, the C μ region, the C γ 1 region (all from human) and the rat 3' enhancer sequence.

EXAMPLE 5

Construction of Light Chain Minilocus

20 A. Construction of pE μ 1

The construction of pE μ 1 is depicted in Fig. 16. The mouse heavy chain enhancer is isolated on the XbaI to EcoRI 678 bp fragment (Banerji et al., Cell 33:729-740 (1983)) from phage clones using oligo:

25

5' GAA TGG GAG TGA GGC TCT CTC ATA CCC
TAT TCA GAA CTG ACT 3'

30

This E μ fragment is cloned into EcoRV/XbaI digested pGP1 by blunt end filling in EcoRI site. The resultant plasmid is designated pE μ 1.

B. Construction Of κ Light chain Minilocus

The κ construct contains at least one human V_{κ} segment, all five human J_{κ} segments, the human J- C_{κ} enhancer, human κ constant region exon, and, ideally, the human 3' κ enhancer (Meyer et al., EMBO J. 8:1959-1964 (1989)). The κ enhancer in mouse is 9 kb downstream from C_{κ} . However, it is as yet unidentified in the human. In addition, the construct contains a copy of the mouse heavy chain J- C_{μ} enhancers.

The minilocus is constructed from four component fragments:

(a) A 16 kb SmaI fragment that contains the human C_{κ} exon and the 3' human enhancer by analogy with the mouse locus;

(b) A 5' adjacent 5 kb SmaI fragment, which contains all five J segments;

(c) The mouse heavy chain intronic enhancer isolated from pE μ 1 (this sequence is included to induce expression of the light chain construct as early as possible in B-cell development. Because the heavy chain genes are transcribed earlier than the light chain genes, this heavy chain enhancer is presumably active at an earlier stage than the intronic κ enhancer); and

(d) A fragment containing one or more V segments.

The preparation of this construct is as follows.

Human placental DNA is digested with SmaI and fractionated on agarose gel by electrophoresis. Similarly, human placental DNA is digested with BamHI and fractionated by electrophoresis. The 16 kb fraction is isolated from the SmaI digested gel and the 11 kb region is similarly isolated from the gel containing DNA digested with BamHI.

The 16 kb SmaI fraction is cloned into Lambda FIX II (Stratagene, La Jolla, California) which has been digested

WO 98/24884

PCT/US97/21803

133

with XhoI, treated with klenow fragment DNA polymerase to fill in the XhoI restriction digest product. Ligation of the 16 kb SmaI fraction destroys the SmaI sites and leaves XhoI sites intact.

5 The 11 kb BamHI fraction is cloned into λ EMBL3 (Stratagene, La Jolla, California) which is digested with BamHI prior to cloning.

Clones from each library were probed with the Ck specific oligo:

10

5' GAA CTG TGG CTG CAC CAT CTG TCT
TCA TCT TCC CGC CAT CTG 3'

15 A 16 kb XhoI insert that was subcloned into the XhoI cut pE μ 1 so that Ck is adjacent to the SmaI site. The resultant plasmid was designated pKap1.

20 The above Ck specific oligonucleotide is used to probe the λ EMBL3/BamHI library to identify an 11 kb clone. A 5 kb SmaI fragment (fragment (b) in Fig. 20) is subcloned and subsequently inserted into pKap1 digested with SmaI. Those plasmids containing the correct orientation of J segments, Ck and the E μ enhancer are designated pKap2.

25 One or more Vk segments are thereafter subcloned into the MluI site of pKap2 to yield the plasmid pKapH which encodes the human Vk segments, the human Jk segments, the human Ck segments and the human E μ enhancer. This insert is excised by digesting pKapH with NotI and purified by agarose gel electrophoresis. The thus purified insert is
30 microinjected into the pronucleus of a mouse zygote as previously described.

C. Construction of κ Light Chain Minilocus by
 In Vivo Homologous Recombination

The 11 kb BamHI fragment is cloned into BamHI digested pGP1 such that the 3' end is toward the SfiI site. The resultant plasmid is designated pKAPint. One or more V_k segments is inserted into the polylinker between the BamHI and SpeI sites in pKAPint to form pKapHV. The insert of pKapHV is excised by digestion with NotI and purified. The insert from pKap2 is excised by digestion with NotI and purified. Each of these fragments contain regions of homology in that the fragment from pKapHV contains a 5 kb sequence of DNA that include the J_k segments which is substantially homologous to the 5 kb SmaI fragment contained in the insert obtained from pKap2. As such, these inserts are capable of homologously recombining when microinjected into a mouse zygote to form a transgene encoding V_k, J_k and C_k.

15

EXAMPLE 6

Isolation of Genomic Clones
Corresponding to Rearranged and Expressed
20 Copies of Immunoglobulin κ Light Chain Genes

This example describes the cloning of immunoglobulin κ light chain genes from cultured cells that express an immunoglobulin of interest. Such cells may contain multiple alleles of a given immunoglobulin gene. For example, a hybridoma might contain four copies of the κ light chain gene, two copies from the fusion partner cell line and two copies from the original B-cell expressing the immunoglobulin of interest. Of these four copies, only one encodes the immunoglobulin of interest, despite the fact that several of them may be rearranged. The procedure described in this example allows for the selective cloning of the expressed copy of the κ light chain.

A. Double Stranded cDNA

Cells from human hybridoma, or lymphoma, or other cell line that synthesizes either cell surface or secreted or both forms of IgM with a κ light chain are used for the isolation of polyA⁺ RNA. The RNA is then used for the synthesis of oligo dT primed cDNA using the enzyme reverse transcriptase (for general methods see, Goodspeed et al. (1989) Gene 76: 1; Dunn et al. (1989) J. Biol. Chem. 264: 13057). The single stranded cDNA is then isolated and G residues are added to the 3' end using the enzyme polynucleotide terminal transferase. The G-tailed single-stranded cDNA is then purified and used as template for second strand synthesis (catalyzed by the enzyme DNA polymerase) using the following oligonucleotide as a primer:

5' - GAG GTA CAC TGA CAT ACT GGC ATG CCC
CCC CCC CCC - 3'

The double stranded cDNA is isolated and used for determining the nucleotide sequence of the 5' end of the mRNAs encoding the heavy and light chains of the expressed immunoglobulin molecule. Genomic clones of these expressed genes are then isolated. The procedure for cloning the expressed light chain gene is outlined in part B below.

B. Light Chain

The double stranded cDNA described in part A is denatured and used as a template for a third round of DNA synthesis using the following oligonucleotide primer:

5' - GTA CGC CAT ATC AGC TGG ATG AAG TCA TCA GAT
GGC GGG AAG ATG AAG ACA GAT GGT GCA - 3'

This primer contains sequences specific for the constant portion of the κ light chain message (TCA TCA GAT GGC GGG AAG ATG AAG ACA GAT GGT GCA) as well as unique sequences that can be used as a primer for the PCR amplification of the newly synthesized DNA strand (GTA CGC CAT ATC AGC TGG ATG AAG). The sequence is amplified by PCR using the following two oligonucleotide primers:

10 5' - GAG GTA CAC TGA CAT ACT GGC ATG -3'
 5' - GTA CGC CAT ATC AGC TGG ATG AAG -3'

The PCR amplified sequence is then purified by gel electrophoresis and used as template for dideoxy sequencing reactions using the following oligonucleotide as a primer:

5' - GAG GTA CAC TGA CAT ACT GGC ATG -3'

The first 42 nucleotides of sequence will then be used to synthesize a unique probe for isolating the gene from which immunoglobulin message was transcribed. This synthetic 42 nucleotide segment of DNA will be referred to below as o-kappa.

A Southern blot of DNA, isolated from the Ig expressing cell line and digested individually and in pairwise combinations with several different restriction endonucleases including SmaI, is then probed with the 32-P labelled unique oligonucleotide o-kappa. A unique restriction endonuclease site is identified upstream of the rearranged V segment.

30 DNA from the Ig expressing cell line is then cut with SmaI and second enzyme (or BamHI or KpnI if there is SmaI site inside V segment). Any resulting non-blunted ends are treated with the enzyme T4 DNA polymerase to give blunt ended

DNA molecules. Then add restriction site encoding linkers (BamHI, EcoRI or XhoI depending on what site does not exist in fragment) and cut with the corresponding linker enzyme to give DNA fragments with BamHI, EcoRI or XhoI ends. The DNA is then size fractionated by agarose gel electrophoresis, and the fraction including the DNA fragment covering the expressed V segment is cloned into lambda EMBL3 or Lambda FIX (Stratagene, La Jolla, California). V segment containing clones are isolated using the unique probe o-kappa. DNA is isolated from positive clones and subcloned into the polylinker of pKap1. The resulting clone is called pRKL.

EXAMPLE 7

Isolation of Genomic Clones Corresponding to Rearranged Expressed Copies of Immunoglobulin Heavy Chain μ Genes

This example describes the cloning of immunoglobulin heavy chain μ genes from cultured cells of expressed and immunoglobulin of interest. The procedure described in this example allows for the selective cloning of the expressed copy of a μ heavy chain gene.

Double-stranded cDNA is prepared and isolated as described herein before. The double-stranded cDNA is denatured and used as a template for a third round of DNA synthesis using the following oligonucleotide primer:

5' - GTA CGC CAT ATC AGC TGG ATG AAG ACA GGA GAC
GAG GGG GAA AAG GGT TGG GGC GGA TGC - 3'

This primer contains sequences specific for the constant portion of the μ heavy chain message (ACA GGA GAC GAG GGG GAA AAG GGT TGG GGC GGA TGC) as well as unique sequences

that can be used as a primer for the PCR amplification of the newly synthesized DNA strand (GTA CGC CAT ATC AGC TGG ATG AAG). The sequence is amplified by PCR using the following two oligonucleotide primers:

5

5' - GAG GTA CAC TGA CAT ACT GGC ATG - 3'

5' - GTA CTC CAT ATC AGC TGG ATG AAG - 3'

The PCR amplified sequence is then purified by gel electrophoresis and used as template for dideoxy sequencing reactions using the following oligonucleotide as a primer:

5' - GAG GTA CAC TGA CAT ACT GGC ATG - 3'

The first 42 nucleotides of sequence are then used to synthesize a unique probe for isolating the gene from which immunoglobulin message was transcribed. This synthetic 42 nucleotide segment of DNA will be referred to below as o-mu.

A Southern blot of DNA, isolated from the Ig expressing cell line and digested individually and in pairwise combinations with several different restriction endonucleases including MluI (MluI is a rare cutting enzyme that cleaves between the J segment and mu CH1), is then probed with the 32-P labelled unique oligonucleotide o-mu. A unique restriction endonuclease site is identified upstream of the rearranged V segment.

DNA from the Ig expressing cell line is then cut with MluI and second enzyme. MluI or SpeI adapter linkers are then ligated onto the ends and cut to convert the upstream site to MluI or SpeI. The DNA is then size fractionated by agarose gel electrophoresis, and the fraction including the

DNA fragment covering the expressed V segment is cloned directly into the plasmid pGPI. V segment containing clones are isolated using the unique probe o-mu, and the insert is subcloned into MluI or MluI/SpeI cut plasmid pCON2. The
5 resulting plasmid is called pRMGH.

EXAMPLE 8

Construction of Human κ Miniloci Transgenes

Light Chain Minilocus

10 A human genomic DNA phage library was screened with kappa light chain specific oligonucleotide probes and isolated clones spanning the J_{κ} -C region. A 5.7 kb ClaI/XhoI fragment containing $J_{\kappa}1$ together with a 13 kb XhoI fragment containing $J_{\kappa}2-5$ and C_{κ} into pGP1d was cloned and used to create the
15 plasmid pKcor. This plasmid contains $J_{\kappa}1-5$, the kappa intronic enhancer and C_{κ} together with 4.5 kb of 5' and 9 kb of 3' flanking sequences. It also has a unique 5' XhoI site for cloning V_{κ} segments and a unique 3' SalI site for inserting additional cis-acting regulatory sequences.

20

V_{κ} kappa genes

A human genomic DNA phage library was screened with V_{κ} light chain specific oligonucleotide probes and isolated clones containing human V_{κ} segments. Functional V segments
25 were identified by DNA sequence analysis. These clones contain TATA boxes, open reading frames encoding leader and variable peptides (including 2 cysteine residues), splice sequences, and recombination heptamer-12 bp spacer-nonamer sequences. Three of the clones were mapped and sequenced.
30 Two of the clones, 65.5 and 65.8 appear to be functional, they contain TATA boxes, open reading frames encoding leader and variable peptides (including 2 cysteine residues), splice

WO 98/24884

PCT/US97/21803

140

sequences, and recombination heptamer-12 bp spacer-nonamer sequences. The third clone, 65.4, appears to encode a V_K I pseudogene as it contains a non-canonical recombination heptamer.

5 One of the functional clones, V_K 65-8, which encodes a V_K III family gene, was used to build a light chain minilocus construct.

pKC1

10 The kappa light chain minilocus transgene pKC1 (Fig. 32) was generated by inserting a 7.5 kb XhoI/SalI fragment containing V_K 65.8 into the 5' XhoI site of pKcor. The transgene insert was isolated by digestion with NotI prior to injection.

15 The purified insert was microinjected into the pronuclei of fertilized (C57BL/6 x CBA)F2 mouse embryos and transferred the surviving embryos into pseudopregnant females as described by Hogan et al. (in Methods of Manipulating the Mouse Embryo, 1986, Cold Spring Harbor Laboratory, New York).

20 Mice that developed from injected embryos were analyzed for the presence of transgene sequences by Southern blot analysis of tail DNA. Transgene copy number was estimated by band intensity relative to control standards containing known quantities of cloned DNA. Serum was isolated from these

25 animals and assayed for the presence of transgene encoded human Ig kappa protein by ELISA as described by Harlow and Lane (in Antibodies: A Laboratory Manual, 1988, Cold Spring Harbor Laboratory, New York). Microtiter plate wells were coated with mouse monoclonal antibodies specific for human Ig

30 kappa (clone 6E1, #0173, AMAC, Inc., Westbrook, ME), human IgM (Clone AF6, #0285, AMAC, Inc., Westbrook, ME) and human IgG1 (clone JL512, #0280, AMAC, Inc., Westbrook, ME). Serum

samples were serially diluted into the wells and the presence of specific immunoglobulins detected with affinity isolated alkaline phosphatase conjugated goat anti-human Ig (polyvalent) that had been pre-adsorbed to minimize cross-reactivity with mouse immunoglobulins.

Fig. 35 shows the results of an ELISA assay of serum from 8 mice (I.D. #676, 674, 673, 670, 666, 665, 664, and 496). The first seven of these mice developed from embryos that were injected with the pKC1 transgene insert and the eighth mouse is derived from a mouse generated by microinjection of the pHCl transgene (described previously). Two of the seven mice from KC1 injected embryos (I.D.#'s 666 and 664) did not contain the transgene insert as assayed by DAN Southern blot analysis, and five of the mice (I.D.#'s 676, 674, 673, 670, and 665) contained the transgene. All but one of the KC1 transgene positive animals express detectable levels of human Ig kappa protein, and the single non-expressing animal appears to be a genetic mosaic on the basis of DNA Southern blot analysis. The pHCl positive transgenic mouse expresses human IgM and IgG1 but not Ig kappa, demonstrating the specificity of the reagents used in the assay.

pKC2

The kappa light chain minilocus transgene pKC2 was generated by inserting an 8 kb XhoI/SalI fragment containing V_K 65.5 into the 5' XhoI site of pKC1. The resulting transgene insert, which contains two V_K segments, was isolated prior to microinjection by digestion with NotI.

pKVe2

This construct is identical to pKC1 except that it includes 1.2 kb of additional sequence 5' of J_κ and is missing 4.5 kb of sequence 3' of V_κ 65.8. In addition it contains a 0.9 kb XbaI fragment containing the mouse heavy chain J-μ intronic enhancer (Banerji et al., Cell 33:729-740 (1983)) together with a 1.4 kb MluI/HindIII fragment containing the human heavy chain J-μ intronic enhancer (Hayday et al., Nature 307:334-340 (1984)) inserted downstream. This construct tests the feasibility of initiating early rearrangement of the light chain minilocus to effect allelic and isotypic exclusion. Analogous constructs can be generated with different enhancers, i.e., the mouse or rat 3' kappa or heavy chain enhancer (Meyer and Neuberger, EMBO J. 8:1959-1964 (1989); Petterson et al. Nature 344:165-168 (1990), which are incorporated herein by reference).

Rearranged Light Chain Transgenes

A kappa light chain expression cassette was designed to reconstruct functionally rearranged light chain genes that have been amplified by PCR from human B-cell DNA. The scheme is outlined in Fig. 33. PCR amplified light chain genes are cloned into the vector pK5nx that includes 3.7 kb of 5' flanking sequences isolated from the kappa light chain gene 65.5. The VJ segment fused to the 5' transcriptional sequences are then cloned into the unique XhoI site of the vector pK31s that includes J_κ2-4, the J_κ intronic enhancer, C_κ, and 9 kb of downstream sequences. The resulting plasmid contains a reconstructed functionally rearranged kappa light chain transgene that can be excised with NotI for microinjection into embryos. The plasmids also contain unique SalI sites at the 3' end for the insertion of additional cis-acting regulatory sequences.

Two synthetic oligonucleotides (o-130, o-131) were used to amplify rearranged kappa light chain genes from human spleen genomic DNA. Oligonucleotide o-131 (gga ccc aga (g,c)gg aac cat gga a(g,a)(g,a,t,c)) is complementary to the 5' region of V_KIII family light chain genes and overlaps the first ATC of the leader sequence. Oligonucleotide o-130 (gtg caa tca att ctc gag ttt gac tac aga c) is complementary to a sequence approximately 150 bp 3' of J_K1 and includes an XhoI site. These two oligonucleotides amplify a 0.7 kb DNA fragment from human spleen DNA corresponding to rearranged V_KIII genes joined to J_K1 segments. The PCR amplified DNA was digested with NcoI and XhoI and cloned individual PCR products into the plasmid pNN03. The DNA sequence of 5 clones was determined and identified two with functional VJ joints (open reading frames). Additional functionally rearranged light chain clones are collected. The functionally rearranged clones can be individually cloned into light chain expression cassette described above (Fig. 33). Transgenic mice generated with the rearranged light chain constructs can be bred with heavy chain minilocus transgenics to produce a strain of mice that express a spectrum of fully human antibodies in which all of the diversity of the primary repertoire is contributed by the heavy chain. One source of light chain diversity can be from somatic mutation. Because not all light chains will be equivalent with respect to their ability to combine with a variety of different heavy chains, different strains of mice, each containing different light chain constructs can be generated and tested. The advantage of this scheme, as opposed to the use of unrearranged light chain miniloci, is the increased light chain allelic and isotypic exclusion that comes from having the light chain ready to pair with a heavy chain as soon as heavy chain VDJ joining occurs. This

WO 98/24884

PCT/US97/21803

144

combination can result in an increased frequency of B-cells expressing fully human antibodies, and thus it can facilitate the isolation of human Ig expressing hybridomas.

NotI inserts of plasmids pIGM1, pHCl, pIGG1, pKCl, and pKC2 were isolated away from vector sequences by agarose gel electrophoresis. The purified inserts were microinjected into the pronuclei of fertilized (C57BL/6 x CBA)F2 mouse embryos and transferred the surviving embryos into pseudopregnant females as described by Hogan et al. (Hogan et al., Methods of Manipulating the Mouse Embryo, Cold Spring Harbor Laboratory, New York (1986)).

EXAMPLE 9

Inactivation of the Mouse Kappa Light Chain Gene by Homologous Recombination

This example describes the inactivation of the mouse endogenous kappa locus by homologous recombination in embryonic stem (ES) cells followed by introduction of the mutated gene into the mouse germ line by injection of targeted ES cells bearing an inactivated kappa allele into early mouse embryos (blastocysts).

The strategy is to delete J_K and C_K by homologous recombination with a vector containing DNA sequences homologous to the mouse kappa locus in which a 4.5 kb segment of the locus, spanning the J_K gene and C_K segments, is deleted and replaced by the selectable marker neo.

Construction of the kappa targeting vector

The plasmid pGEM7 (KJ1) contains the neomycin resistance gene (neo), used for drug selection of transfected ES cells, under the transcriptional control of the mouse phosphoglycerate kinase (pgk) promoter (XbaI/TaqI fragment;

Adra et al. (1987) Gene 60: 65) in the cloning vector pGEM-7Zf(+). The plasmid also includes a heterologous polyadenylation site for the neo gene, derived from the 3' region of the mouse pgk gene (PvuII/HindIII fragment; Boer et al., Biochemical Genetics, 28:299-308 (1990)). This plasmid was used as the starting point for construction of the kappa targeting vector. The first step was to insert sequences homologous to the kappa locus 3' of the neo expression cassette.

10 Mouse kappa chain sequences (Fig. 20a) were isolated from a genomic phage library derived from liver DNA using oligonucleotide probes specific for the Ck locus:

5'- GGC TGA TGC TGC ACC AAC TGT ATC CAT CTT CCC ACC ATC CAG
15 -3'

and for the Jk5 gene segment:

5'- CTC ACG TTC GGT GCT GGG ACC AAG CTG GAG CTG AAA CGT AAG -
20 3'.

An 8 kb BglII/SacI fragment extending 3' of the mouse C_k segment was isolated from a positive phage clone in two pieces, as a 1.2 kb BglII/SacI fragment and a 6.8 kb SacI fragment, and subcloned into BglII/SacI digested pGEM7 (KJ1) to generate the plasmid pNEO-K3' (Fig. 20b).

A 1.2 kb EcoRI/SphI fragment extending 5' of the J_k region was also isolated from a positive phage clone. An SphI/XbaI/BglII/EcoRI adaptor was ligated to the SphI site of this fragment, and the resulting EcoRI fragment was ligated into EcoRI digested pNEO-K3', in the same 5' to 3' orientation as the neo gene and the downstream 3' kappa sequences, to generate pNEO-K5'3' (Fig. 20c).

The Herpes Simplex Virus (HSV) thymidine kinase (TK) gene was then included in the construct in order to allow for enrichment of ES clones bearing homologous recombinants, as described by Mansour et al., Nature 336:348-352 (1988), which is incorporated herein by reference. The HSV TK cassette was obtained from the plasmid pGEM7 (TK), which contains the structural sequences for the HSV TK gene bracketed by the mouse pgk promoter and polyadenylation sequences as described above for pGEM7 (KJ1). The EcoRI site of pGEM7 (TK) was modified to a BamHI site and the TK cassette was then excised as a BamHI/HindIII fragment and subcloned into pGP1b to generate pGP1b-TK. This plasmid was linearized at the XhoI site and the XhoI fragment from pNEO-K5'3', containing the neo gene flanked by genomic sequences from 5' of Jk and 3' of Ck, was inserted into pGP1b-TK to generate the targeting vector J/C KI (Fig. 20d). The putative structure of the genomic kappa locus following homologous recombination with J/C KI is shown in Fig. 20e.

20 Generation and analysis of ES cells with targeted inactivation of a kappa allele

The ES cells used were the AB-1 line grown on mitotically inactive SNL76/7 cell feeder layers (McMahon and Bradley, Cell 62:1073-1085 (1990)) essentially as described (Robertson, E.J. (1987) in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach. E.J. Robertson, ed. (Oxford: IRL Press), p. 71-112). Other suitable ES lines include, but are not limited to, the E14 line (Hooper et al. (1987) Nature 326: 292-295), the D3 line (Doetschman et al. (1985) J. Embryol. Exp. Morph. 87: 27-45), and the CCE line (Robertson et al. (1986) Nature 323: 445-448). The success of generating a mouse line from ES cells bearing a specific targeted mutation

depends on the pluripotency of the ES cells (i.e., their ability, once injected into a host blastocyst, to participate in embryogenesis and contribute to the germ cells of the resulting animal).

5 The pluripotency of any given ES cell line can vary with time in culture and the care with which it has been handled. The only definitive assay for pluripotency is to determine whether the specific population of ES cells to be used for targeting can give rise to chimeras capable of
10 germline transmission of the ES genome. For this reason, prior to gene targeting, a portion of the parental population of AB-1 cells is injected into C57Bl/6J blastocysts to ascertain whether the cells are capable of generating chimeric mice with extensive ES cell contribution and whether the
15 majority of these chimeras can transmit the ES genome to progeny.

 The kappa chain inactivation vector J/C K1 was digested with NotI and electroporated into AB-1 cells by the methods described (Hasty et al., Nature, 350:243-246 (1991)).
20 Electroporated cells were plated onto 100 mm dishes at a density of $1-2 \times 10^6$ cells/dish. After 24 hours, G418 (200 µg/ml of active component) and FIAU (0.5 µM) were added to the medium, and drug-resistant clones were allowed to develop over 10-11 days. Clones were picked, trypsinized, divided
25 into two portions, and further expanded. Half of the cells derived from each clone were then frozen and the other half analyzed for homologous recombination between vector and target sequences.

 DNA analysis was carried out by Southern blot
30 hybridization. DNA was isolated from the clones as described (Laird et al., Nucl. Acids Res. 19:4293 (1991)) digested with XbaI and probed with the 800 bp EcoRI/XbaI fragment indicated

in Fig. 20e as probe A. This probe detects a 3.7 kb XbaI fragment in the wild type locus, and a diagnostic 1.8 kb band in a locus which has homologously recombined with the targeting vector (see Fig. 20a and e). Of 901 G418 and FIAU resistant clones screened by Southern blot analysis, 7 displayed the 1.8 kb XbaI band indicative of a homologous recombination into one of the kappa genes. These 7 clones were further digested with the enzymes BglII, SacI, and PstI to verify that the vector integrated homologously into one of the kappa genes. When probed with the diagnostic 800 bp EcoRI/XbaI fragment (probe A), BglII, SacI, and PstI digests of wild type DNA produce fragments of 4.1, 5.4, and 7 kb, respectively, whereas the presence of a targeted kappa allele would be indicated by fragments of 2.4, 7.5, and 5.7 kb, respectively (see Fig. 20a and e). All 7 positive clones detected by the XbaI digest showed the expected BglII, SacI, and PstI restriction fragments diagnostic of a homologous recombination at the kappa light chain. In addition, Southern blot analysis of an NsiI digest of the targeted clones using a neo specific probe (probe B, Fig. 20e) generated only the predicted fragment of 4.2 kb, demonstrating that the clones each contained only a single copy of the targeting vector.

Generation of mice bearing the inactivated kappa chain

Five of the targeted ES clones described in the previous section were thawed and injected into C57Bl/6J blastocysts as described (Bradley, A. (1987) in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach. E.J. Robertson, ed. (Oxford: IRL Press), p. 113-151) and transferred into the uteri of pseudopregnant females to generate chimeric mice resulting from a mixture of cells derived from the input ES cells and the host blastocyst. The

extent of ES cell contribution to the chimeras can be visually estimated by the amount of agouti coat coloration, derived from the ES cell line, on the black C57Bl/6J background. Approximately half of the offspring resulting from blastocyst injection of the targeted clones were chimeric (i.e., showed agouti as well as black pigmentation) and of these, the majority showed extensive (70 percent or greater) ES cell contribution to coat pigmentation. The AB1 ES cells are an XY cell line and a majority of these high percentage chimeras were male due to sex conversion of female embryos colonized by male ES cells. Male chimeras derived from 4 of the 5 targeted clones were bred with C57BL/6J females and the offspring monitored for the presence of the dominant agouti coat color indicative of germline transmission of the ES genome. Chimeras from two of these clones consistently generated agouti offspring. Since only one copy of the kappa locus was targeted in the injected ES clones, each agouti pup had a 50 percent chance of inheriting the mutated locus. Screening for the targeted gene was carried out by Southern blot analysis of Bgl II-digested DNA from tail biopsies, using the probe utilized in identifying targeted ES clones (probe A, Fig. 20e). As expected, approximately 50 percent of the agouti offspring showed a hybridizing Bgl II band of 2.4 kb in addition to the wild-type band of 4.1 kb, demonstrating the germline transmission of the targeted kappa locus.

In order to generate mice homozygous for the mutation, heterozygotes were bred together and the kappa genotype of the offspring determined as described above. As expected, three genotypes were derived from the heterozygote matings: wild-type mice bearing two copies of a normal kappa locus, heterozygotes carrying one targeted copy of the kappa gene and one NT kappa gene, and mice homozygous for the kappa

mutation. The deletion of kappa sequences from these latter mice was verified by hybridization of the Southern blots with a probe specific for J_K (probe C, Fig. 20a). Whereas hybridization of the J_K probe was observed to DNA samples from heterozygous and wild-type siblings, no hybridizing signal was present in the homozygotes, attesting to the generation of a novel mouse strain in which both copies of the kappa locus have been inactivated by deletion as a result of targeted mutation.

10

EXAMPLE 10

Inactivation of the Mouse Heavy Chain Gene by Homologous Recombination

This example describes the inactivation of the endogenous murine immunoglobulin heavy chain locus by homologous recombination in embryonic stem (ES) cells. The strategy is to delete the endogenous heavy chain J segments by homologous recombination with a vector containing heavy chain sequences from which the J_H region has been deleted and replaced by the gene for the selectable marker neo.

15
20Construction of a heavy chain targeting vector

Mouse heavy chain sequences containing the J_H region (Fig. 21a) were isolated from a genomic phage library derived from the D3 ES cell line (Gossler et al., Proc. Natl. Acad. Sci. U.S.A. 83:9065-9069 (1986)) using a J_H4 specific oligonucleotide probe:

25

5' - ACT ATG CTA TGG ACT ACT GGG GTC AAG GAA CCT CAG TCA CCG
-3'

30

A 3.5 kb genomic SacI/StuI fragment, spanning the J_H region, was isolated from a positive phage clone and subcloned

into SacI/SmaI digested pUC18. The resulting plasmid was designated pUC18 J_H. The neomycin resistance gene (neo), used for drug selection of transfected ES cells, was derived from a repaired version of the plasmid pGEM7 (KJ1). A report in the literature (Yenofsky et al. (1990) Proc. Natl. Acad. Sci. (U.S.A.) 87: 3435-3439) documents a point mutation the neo coding sequences of several commonly used expression vectors, including the construct pMC1neo (Thomas and Cappechi (1987) Cell 51: 503-512) which served as the source of the neo gene used in pGEM7 (KJ1). This mutation reduces the activity of the neo gene product and was repaired by replacing a restriction fragment encompassing the mutation with the corresponding sequence from a wild-type neo clone. The HindIII site in the prepared pGEM7 (KJ1) was converted to a SalI site by addition of a synthetic adaptor, and the neo expression cassette excised by digestion with XbaI/SalI. The ends of the neo fragment were then blunted by treatment with the Klenow form of DNA polI, and the neo fragment was subcloned into the NaeI site of pUC18 J_H, generating the plasmid pUC18 J_H-neo (Fig. 21b).

Further construction of the targeting vector was carried out in a derivative of the plasmid pGP1b. pGP1b was digested with the restriction enzyme NotI and ligated with the following oligonucleotide as an adaptor:

5'- GGC CGC TCG ACG ATA GCC TCG AGG CTA TAA ATC TAG AAG AAT
TCC AGC AAA GCT TTG GC -3'

The resulting plasmid, called pGMT, was used to build the mouse immunoglobulin heavy chain targeting construct.

The Herpes Simplex Virus (HSV) thymidine kinase (TK) gene was included in the construct in order to allow for enrichment of ES clones bearing homologous recombinants, as described by Mansour et al. (Nature 336, 348-352 (1988)). The
5 HSV TK gene was obtained from the plasmid pGEM7 (TK) by digestion with EcoRI and HindIII. The TK DNA fragment was subcloned between the EcoRI and HindIII sites of pGMT, creating the plasmid pGMT-TK (Fig. 21c).

To provide an extensive region of homology to the
10 target sequence, a 5.9 kb genomic XbaI/XhoI fragment, situated 5' of the J_H region, was derived from a positive genomic phage clone by limit digestion of the DNA with XhoI, and partial digestion with XbaI. As noted in Fig. 21a, this XbaI site is not present in genomic DNA, but is rather derived from phage
15 sequences immediately flanking the cloned genomic heavy chain insert in the positive phage clone. The fragment was subcloned into XbaI/XhoI digested pGMT-TK, to generate the plasmid pGMT-TK-J_H5' (Fig. 21d).

The final step in the construction involved the
20 excision from pUC18 J_H-neo of the 2.8 kb EcoRI fragment which contained the neo gene and flanking genomic sequences 3' of J_H. This fragment was blunted by Klenow polymerase and subcloned into the similarly blunted XhoI site of pGMT-TK-J_H5'. The resulting construct, J_HKO1 (Fig. 21e), contains 6.9 kb of
25 genomic sequences flanking the J_H locus, with a 2.3 kb deletion spanning the J_H region into which has been inserted the neo gene. Fig. 21f shows the structure of an endogenous heavy chain gene after homologous recombination with the targeting construct.

EXAMPLE 11

Generation and analysis of targeted ES cells

AB-1 ES cells (McMahon and Bradley, Cell 62:1073-1085 (1990)) were grown on mitotically inactive
5 SNL76/7 cell feeder layers essentially as described
(Robertson, E.J. (1987) Teratocarcinomas and Embryonic Stem Cells: A Practical Approach. E.J. Robertson, ed. (Oxford: IRL Press), pp. 71-112). As described in the previous example,
prior to electroporation of ES cells with the targeting
10 construct J_HKO1, the pluripotency of the ES cells was
determined by generation of AB-1 derived chimeras which were
shown capable of germline transmission of the ES genome.

The heavy chain inactivation vector J_HKO1 was
digested with NotI and electroporated into AB-1 cells by the
15 methods described (Hasty et al., Nature 350:243-246 (1991)).
Electroporated cells were plated into 100 mm dishes at a
density of 1-2 x 10⁶ cells/dish. After 24 hours, G418
(200mg/ml of active component) and FIAU (0.5mM) were added to
the medium, and drug-resistant clones were allowed to develop
20 over 8-10 days. Clones were picked, trypsinized, divided into
two portions, and further expanded. Half of the cells derived
from each clone were then frozen and the other half analyzed
for homologous recombination between vector and target
sequences.

25 DNA analysis was carried out by Southern blot
hybridization. DNA was isolated from the clones as described
(Laird et al. (1991) Nucleic Acids Res. 19: 4293), digested
with StuI and probed with the 500 bp EcoRI/StuI fragment
designated as probe A in Fig. 21f. This probe detects a StuI
30 fragment of 4.7 kb in the wild-type locus, whereas a 3 kb band
is diagnostic of homologous recombination of endogenous
sequences with the targeting vector (see Fig. 21a and f). Of

525 G418 and FIAU doubly-resistant clones screened by Southern blot hybridization, 12 were found to contain the 3 kb fragment diagnostic of recombination with the targeting vector. That these clones represent the expected targeted events at the J_H locus (as shown in Fig. 21f) was confirmed by further digestion with HindIII, SpeI and HpaI. Hybridization of probe A (see Fig. 21f) to Southern blots of HindIII, SpeI, and HpaI digested DNA produces bands of 2.3 kb, >10 kb, and >10kb, respectively, for the wild-type locus (see Fig. 21a), whereas bands of 5.3 kb, 3.8 kb, and 1.9 kb, respectively, are expected for the targeted heavy chain locus (see Fig 21f). All 12 positive clones detected by the StuI digest showed the predicted HindIII, SpeI, and HpaI bands diagnostic of a targeted J_H gene. In addition, Southern blot analysis of a StuI digest of all 12 clones using a neo-specific probe (probe B, Fig. 21f) generated only the predicted fragment of 3 kb, demonstrating that the clones each contained only a single copy of the targeting vector.

20 Generation of mice carrying the J_H deletion

Three of the targeted ES clones described in the previous section were thawed and injected into C57BL/6J blastocysts as described (Bradley, A. (1987) in Teratocarcinomas and Embryonic Stem Cells: A Practical Approach, E.J. Robertson, ed. (Oxford: IRL Press), p.113-151) and transferred into the uteri of pseudopregnant females. The extent of ES cell contribution to the chimera was visually estimated from the amount of agouti coat coloration, derived from the ES cell line, on the black C57BL/6J background. Half of the offspring resulting from blastocyst injection of two of the targeted clones were chimeric (i.e., showed agouti as well as black pigmentation); the third targeted clone did not

generate any chimeric animals. The majority of the chimeras showed significant (approximately 50 percent or greater) ES cell contribution to coat pigmentation. Since the AB-1 ES cells are an XY cell line, most of the chimeras were male, due to sex conversion of female embryos colonized by male ES cells. Males chimeras were bred with C57BL/6J females and the offspring monitored for the presence of the dominant agouti coat color indicative of germline transmission of the ES genome. Chimeras from both of the clones consistently generated agouti offspring. Since only one copy of the heavy chain locus was targeted in the injected ES clones, each agouti pup had a 50 percent chance of inheriting the mutated locus. Screening for the targeted gene was carried out by Southern blot analysis of StuI-digested DNA from tail biopsies, using the probe utilized in identifying targeted ES clones (probe A, Fig. 21f). As expected, approximately 50 percent of the agouti offspring showed a hybridizing StuI band of approximately 3 kb in addition to the wild-type band of 4.7 kb, demonstrating germline transmission of the targeted J_H gene segment.

In order to generate mice homozygous for the mutation, heterozygotes were bred together and the heavy chain genotype of the offspring determined as described above. As expected, three genotypes were derived from the heterozygote matings: wild-type mice bearing two copies of the normal J_H locus, heterozygotes carrying one targeted copy of the gene and one normal copy, and mice homozygous for the J_H mutation. The absence of J_H sequences from these latter mice was verified by hybridization of the Southern blots of StuI-digested DNA with a probe specific for J_H (probe C, Fig. 21a). Whereas hybridization of the J_H probe to a 4.7 kb fragment in DNA samples from heterozygous and wild-type siblings was observed,

no signal was present in samples from the J_H-mutant homozygotes, attesting to the generation of a novel mouse strain in which both copies of the heavy chain gene have been mutated by deletion of the J_H sequences.

5

EXAMPLE 12

Heavy Chain Minilocus TransgeneA. Construction of plasmid vectors for cloning large DNA sequences10 1. pGP1a

The plasmid pBR322 was digested with EcoRI and StyI and ligated with the following oligonucleotides:

15 oligo-42 5'- caa gag ccc gcc taa tga gcg ggc ttt ttt ttg cat
act gcg gcc gct -3'

oligo-43 5'- aat tag cgg ccg cag tat gca aaa aaa agc ccg ctc
att agg cgg gct -3'

20

The resulting plasmid, pGP1a, is designed for cloning very large DNA constructs that can be excised by the rare cutting restriction enzyme NotI. It contains a NotI restriction site downstream (relative to the ampicillin resistance gene, AmpR) of a strong transcription termination signal derived from the trpA gene (Christie et al., Proc. Natl. Acad. Sci. USA 78:4180 (1981)). This termination signal reduces the potential toxicity of coding sequences inserted into the NotI site by eliminating readthrough transcription from the AmpR gene. In addition, this plasmid is low copy relative to the pUC plasmids because it retains the pBR322 copy number control region. The low copy number further reduces the potential toxicity of insert sequences and reduces the selection against large inserts due to DNA replication.

WO 98/24884

PCT/US97/21803

157

The vectors pGP1b, pGP1c, pGP1d, and pGP1f are derived from pGP1a and contain different polylinker cloning sites. The polylinker sequences are given below

5

pGP1a

NotI
GCGGCCGC

10

pGP1b

NotI XhoI ClaI BamHI HindIII NotI

15

GCggccgcctcgagatcactatcgattaattaaggatccagcagtaagcttgcGGCCGC

pGP1c

20

NotI SmaI XhoI SalI HindIII BamHI SacII NotI
GCggccgcatcccgggtctcgaggtcgacaagctttcgaggatccgcGGCCGC

25

pGP1d

NotI SalI HindIII ClaI BamHI XhoI NotI
GCggccgctgtcgacaagcttatcgatggatcctcgagtgcGGCCGC

30

pGP1f

NotI SalI HindIII EcoRI ClaI KpnI BamHI XhoI NotI
GCggccgctgtcgacaagcttcgaattcagatcgatgtggtacctggatcctcgagtgcGGCCGC

35

Each of these plasmids can be used for the construction of large transgene inserts that are excisable with NotI so that the transgene DNA can be purified away from vector sequences prior to microinjection.

40

2. pGP1b

WO 98/24884

PCT/US97/21803

158

pGP1a was digested with NotI and ligated with the following oligonucleotides:

5 oligo-47 5'- ggc cgc aag ctt act gct gga tcc tta att aat cga
tag tga tct cga ggc -3'

oligo-48 5'- ggc cgc ctc gag atc act atc gat taa tta agg atc
cag cag taa gct tgc -3'

10

The resulting plasmid, pGP1b, contains a short polylinker region flanked by NotI sites. This facilitates the construction of large inserts that can be excised by NotI digestion.

15

3. pGPe

The following oligonucleotides:

20 oligo-44 5'- ctc cag gat cca gat atc agt acc tga aac agg gct
tgc -3'

oligo-45 5'- ctc gag cat gca cag gac ctg gag cac aca cag cct
tcc -3'

25

were used to amplify the immunoglobulin heavy chain 3' enhancer (S. Petterson, et al., Nature 344:165-168 (1990)) from rat liver DNA by the polymerase chain reaction technique.

30 The amplified product was digested with BamHI and SphI and cloned into BamHI/SphI digested pNNO3 (pNNO3 is a pUC derived plasmid that contains a polylinker with the following restriction sites, listed in order: NotI, BamHI, NcoI, ClaI, EcoRV, XbaI, SacI, XhoI, SphI, PstI, BglII, EcoRI, SmaI, KpnI, HindIII, and NotI). The resulting plasmid, pRE3, was digested
35 with BamHI and HindIII, and the insert containing the rat Ig heavy chain 3' enhancer cloned into BamHI/HindIII digested

WO 98/24884

PCT/US97/21803

159

pGP1b. The resulting plasmid, pGPe (Fig. 22 and Table 1), contains several unique restriction sites into which sequences can be cloned and subsequently excised together with the 3' enhancer by NotI digestion.

WO 98/24884

PCT/US97/21803

160

TABLE 1

WO 98/24884

PCT/US97/21803

161

B. Construction of IgM expressing minilocus transgene, pIGM11. Isolation of J- μ constant region clones and construction of pJM1

5 A human placental genomic DNA library cloned into the phage vector λ EMBL3/SP6/T7 (Clonetech Laboratories, Inc., Palo Alto, CA) was screened with the human heavy chain J region specific oligonucleotide:

10 oligo-1 5'- gga ctg tgt ccc tgt gtg atg ctt ttg atg tct ggg
gcc aag -3'

and the phage clone λ 1.3 isolated. A 6 kb HindIII/KpnI
15 fragment from this clone, containing all six J segments as well as D segment DHQ52 and the heavy chain J- μ intronic enhancer, was isolated. The same library was screened with the human μ specific oligonucleotide:

20 oligo-2 5'- cac caa gtt gac ctg cct ggt cac aga cct gac cac
cta tga -3'

and the phage clone λ 2.1 isolated. A 10.5 kb HindIII/XhoI
25 fragment, containing the μ switch region and all of the μ constant region exons, was isolated from this clone. These two fragments were ligated together with KpnI/XhoI digested pNNO3 to obtain the plasmid pJM1.

30 2. pJM2

A 4 kb XhoI fragment was isolated from phage clone λ 2.1 that contains sequences immediately downstream of the sequences in pJM1, including the so called $\Sigma\mu$ element involved in δ -associated deletion of the μ in certain IgD expressing
35 B-cells (Yasui et al., Eur. J. Immunol. 19:1399 (1989), which

is incorporated herein by reference). This fragment was treated with the Klenow fragment of DNA polymerase I and ligated to XhoI cut, Klenow treated, pJM1. The resulting plasmid, pJM2 (Fig. 23), had lost the internal XhoI site but
 5 retained the 3' XhoI site due to incomplete reaction by the Klenow enzyme. pJM2 contains the entire human J region, the heavy chain J- μ intronic enhancer, the μ switch region and all of the μ constant region exons, as well as the two 0.4 kb direct repeats, $\sigma\mu$ and $\sum\mu$, involved in δ -associated deletion
 10 of the μ gene.

3. Isolation of D region clones and construction of pDH1

The following human D region specific oligonucleotide:

15

oligo-4 5'- tgg tat tac tat ggt tcg ggg agt tat tat aac cac
 agt gtc -3'

20 was used to screen the human placenta genomic library for D region clones. Phage clones λ 4.1 and λ 4.3 were isolated. A 5.5 kb XhoI fragment, that includes the D elements D_{K1} , D_{N1} , and D_{M2} (Ichihara et al., EMBO J. 7:4141 (1988)), was isolated from phage clone λ 4.1. An adjacent upstream 5.2 kb XhoI fragment,
 25 that includes the D elements D_{LR1} , D_{XP1} , $D_{XP'1}$, and D_{A1} , was isolated from phage clone λ 4.3. Each of these D region XhoI fragments were cloned into the SalI site of the plasmid vector pSP72 (Promega, Madison, WI) so as to destroy the XhoI site linking the two sequences. The upstream fragment was then
 30 excised with XhoI and SmaI, and the downstream fragment with EcoRV and XhoI. The resulting isolated fragments were ligated together with SalI digested pSP72 to give the plasmid pDH1.

pDH1 contains a 10.6 kb insert that includes at least 7 D segments and can be excised with XhoI (5') and EcoRV (3').

4. pCOR1

5 The plasmid pJM2 was digested with Asp718 (an isoschizomer of KpnI) and the overhang filled in with the Klenow fragment of DNA polymerase I. The resulting DNA was then digested with ClaI and the insert isolated. This insert was ligated to the XhoI/EcoRV insert of pDH1 and XhoI/ClaI
10 digested pGPe to generate pCOR1 (Fig. 24).

5. pVH251

 A 10.3 kb genomic HindIII fragment containing the two human heavy chain variable region segments V_H251 and V_H105
15 (Humphries et al., Nature 331:446 (1988), which is incorporated herein by reference) was subcloned into pSP72 to give the plasmid pVH251.

6. pIGM1

20 The plasmid pCOR1 was partially digested with XhoI and the isolated XhoI/SalI insert of pVH251 cloned into the upstream XhoI site to generate the plasmid pIGM1 (Fig. 25). pIGM1 contains 2 functional human variable region segments, at least 8 human D segments all 6 human J_H segments, the human J- μ
25 enhancer, the human $\sigma\mu$ element, the human μ switch region, all of the human μ coding exons, and the human $\Sigma\mu$ element, together with the rat heavy chain 3' enhancer, such that all of these sequence elements can be isolated on a single fragment, away from vector sequences, by digestion with NotI
30 and microinjected into mouse embryo pronuclei to generate transgenic animals.

C. Construction of IgM and IgG expressing minilocus transgene, pHCl

1. Isolation of γ constant region clones

The following oligonucleotide, specific for human Ig
5 g constant region genes:

oligo-29 5'- cag cag gtg cac acc caa tgc cca tga gcc cag aca
ctg gac -3'

10 was used to screen the human genomic library. Phage clones
129.4 and λ 29.5 were isolated. A 4 kb HindIII fragment of
phage clone λ 29.4, containing a γ switch region, was used to
probe a human placenta genomic DNA library cloned into the
phage vector lambda FIX™ II (Stratagene, La Jolla, CA). Phage
15 clone λ Sg1.13 was isolated. To determine the subclass of the
different γ clones, dideoxy sequencing reactions were carried
out using subclones of each of the three phage clones as
templates and the following oligonucleotide as a primer:

20 oligo-67 5'- tga gcc cag aca ctg gac -3'

Phage clones λ 29.5 and λ S γ 1.13 were both determined
to be of the γ 1 subclass.

25 2. p γ e1

A 7.8 kb HindIII fragment of phage clone λ 29.5,
containing the γ 1 coding region was cloned into pUC18. The
resulting plasmid, pLT1, was digested with XhoI, Klenow
treated, and religated to destroy the internal XhoI site. The
30 resulting clone, pLT1xk, was digested with HindIII and the
insert isolated and cloned into pSP72 to generate the plasmid
clone pLT1xks. Digestion of pLT1xks at a polylinker XhoI site

WO 98/24884

PCT/US97/21803

165

and a human sequence derived BamHI site generates a 7.6 kb fragment containing the $\gamma 1$ constant region coding exons. This 7.6 kb XhoI/BamHI fragment was cloned together with an adjacent downstream 4.5 kb BamHI fragment from phage clone
5 $\lambda 29.5$ into XhoI/BamHI digested pGPe to generate the plasmid clone p $\gamma e1$. p $\gamma e1$ contains all of the $\gamma 1$ constant region coding exons, together with 5 kb of downstream sequences, linked to the rat heavy chain 3' enhancer.

10 3. p $\gamma e2$

A 5.3 kb HindIII fragment containing the $\gamma 1$ switch region and the first exon of the pre-switch sterile transcript (P. Sideras et al. (1989) International Immunol. 1, 631) was isolated from phage clone $\lambda Sy1.13$ and cloned into pSP72 with
15 the polylinker XhoI site adjacent to the 5' end of the insert, to generate the plasmid clone p $Sy1s$. The XhoI/SalI insert of p $Sy1s$ was cloned into XhoI digested p $\gamma e1$ to generate the plasmid clone p $\gamma e2$ (Fig. 26). p $\gamma e2$ contains all of the $\gamma 1$ constant region coding exons, and the upstream switch region
20 and sterile transcript exons, together with 5 kb of downstream sequences, linked to the rat heavy chain 3' enhancer. This clone contains a unique XhoI site at the 5' end of the insert. The entire insert, together with the XhoI site and the 3' rat enhancer can be excised from vector sequences by digestion
25 with NotI.

4. pHC1

The plasmid pIGM1 was digested with XhoI and the 43
30 kb insert isolated and cloned into XhoI digested p $\gamma e2$ to generate the plasmid pHC1 (Fig. 25). pHC1 contains 2 functional human variable region segments, at least 8 human D

WO 98/24884

PCT/US97/21803

166

segments all 6 human J_H segments, the human $J-\mu$ enhancer, the human $\sigma\mu$ element, the human μ switch region, all of the human μ coding exons, the human $\Sigma\mu$ element, and the human $\gamma 1$ constant region, including the associated switch region and sterile transcript associated exons, together with the rat heavy chain 3' enhancer, such that all of these sequence elements can be isolated on a single fragment, away from vector sequences, by digestion with NotI and microinjected into mouse embryo pronuclei to generate transgenic animals.

10

D. Construction of IgM and IgG expressing minilocus transgene, pH2

1. Isolation of human heavy chain V region gene VH49.8

The human placental genomic DNA library lambda, FIX™ II, Stratagene, La Jolla, CA) was screened with the following human VH1 family specific oligonucleotide:

oligo-49 5'- gtt aaa gag gat ttt att cac ccc tgt gtc ctc tcc
aca ggt gtc -3'

20

Phage clone $\lambda 49.8$ was isolated and a 6.1 kb XbaI fragment containing the variable segment VH49.8 subcloned into pNNO3 (such that the polylinker ClaI site is downstream of VH49.8 and the polylinker XhoI site is upstream) to generate the plasmid pVH49.8. An 800 bp region of this insert was sequenced, and VH49.8 found to have an open reading frame and intact splicing and recombination signals, thus indicating that the gene is functional (Table 2).

WO 98/24884

PCT/US97/21803

167

TABLE 2

WO 98/24884

PCT/US97/21803

168

2. pV2

A 4 kb XbaI genomic fragment containing the human V_HIV family gene V_H4-21 (Sanz et al., EMBO J., 8:3741 (1989)), subcloned into the plasmid pUC12, was excised with SmaI and HindIII, and treated with the Klenow fragment of polymerase I. The blunt ended fragment was then cloned into ClaI digested, Klenow treated, pVH49.8. The resulting plasmid, pV2, contains the human heavy chain gene VH49.8 linked upstream of VH4-21 in the same orientation, with a unique SalI site at the 3' end of the insert and a unique XhoI site at the 5' end.

3. pSy1-5'

A 0.7 kb XbaI/HindIII fragment (representing sequences immediately upstream of, and adjacent to, the 5.3 kb γ 1 switch region containing fragment in the plasmid p γ e2) together with the neighboring upstream 3.1 kb XbaI fragment were isolated from the phage clone λ Sg1.13 and cloned into HindIII/XbaI digested pUC18 vector. The resulting plasmid, pSy1-5', contains a 3.8 kb insert representing sequences upstream of the initiation site of the sterile transcript found in B-cells prior to switching to the γ 1 isotype (P. Sideras et al., International Immunol. 1:631 (1989)). Because the transcript is implicated in the initiation of isotype switching, and upstream cis-acting sequences are often important for transcription regulation, these sequences are included in transgene constructs to promote correct expression of the sterile transcript and the associated switch recombination.

30 4. pVGE1

The pS γ 1-5' insert was excised with SmaI and HindIII, treated with Klenow enzyme, and ligated with the following oligonucleotide linker:

5 5'- ccg gtc gac cgg -3'

The ligation product was digested with SalI and ligated to SalI digested pV2. The resulting plasmid, pVP, contains 3.8 kb of γ 1 switch 5' flanking sequences linked downstream of the two human variable gene segments VH49.8 and VH4-21 (see Table 2). The pVP insert is isolated by partial digestion with SalI and complete digestion with XhoI, followed by purification of the 15 kb fragment on an agarose gel. The insert is then cloned into the XhoI site of p γ e2 to generate the plasmid clone pVGE1 (Fig. 27). pVGE1 contains two human heavy chain variable gene segments upstream of the human γ 1 constant gene and associated switch region. A unique SalI site between the variable and constant regions can be used to clone in D, J, and μ gene segments. The rat heavy chain 3' enhancer is linked to the 3' end of the γ 1 gene and the entire insert is flanked by NotI sites.

5. pHC2

The plasmid clone pVGE1 is digested with SalI and the XhoI insert of pIGM1 is cloned into it. The resulting clone, pHC2 (Fig. 25), contains 4 functional human variable region segments, at least 8 human D segments all 6 human J_H segments, the human J-m enhancer, the human $\sigma\mu$ element, the human μ switch region, all of the human μ coding exons, the human $\Sigma\mu$ element, and the human γ 1 constant region, including the associated switch region and sterile transcript associated exons, together with 4 kb flanking sequences upstream of the

WO 98/24884

PCT/US97/21803

170

sterile transcript initiation site. These human sequences are linked to the rat heavy chain 3' enhancer, such that all of the sequence elements can be isolated on a single fragment, away from vector sequences, by digestion with NotI and
5 microinjected into mouse embryo pronuclei to generate transgenic animals. A unique XhoI site at the 5' end of the insert can be used to clone in additional human variable gene segments to further expand the recombinational diversity of this heavy chain minilocus.

10

E. Transgenic mice

The NotI inserts of plasmids pIGM1 and pHCl were isolated from vector sequences by agarose gel electrophoresis. The purified inserts were microinjected into the pronuclei of
15 fertilized (C57BL/6 x CBA)F2 mouse embryos and transferred the surviving embryos into pseudopregnant females as described by Hogan et al. (B. Hogan, F. Costantini, and E. Lacy, Methods of Manipulating the Mouse Embryo, 1986, Cold Spring Harbor Laboratory, New York). Mice that developed from injected
20 embryos were analyzed for the presence of transgene sequences by Southern blot analysis of tail DNA. Transgene copy number was estimated by band intensity relative to control standards containing known quantities of cloned DNA. At 3 to 8 weeks of age, serum was isolated from these animals and assayed for the
25 presence of transgene encoded human IgM and IgG1 by ELISA as described by Harlow and Lane (E. Harlow and D. Lane. Antibodies: A Laboratory Manual, 1988, Cold Spring Harbor Laboratory, New York). Microtiter plate wells were coated with mouse monoclonal antibodies specific for human IgM (clone
30 AF6, #0285, AMAC, Inc. Westbrook, ME) and human IgG1 (clone JL512, #0280, AMAC, Inc. Westbrook, ME). Serum samples were serially diluted into the wells and the presence of specific

immunoglobulins detected with affinity isolated alkaline phosphatase conjugated goat anti-human Ig (polyvalent) that had been pre-adsorbed to minimize cross-reactivity with mouse immunoglobulins. Table 3 and Fig. 28 show the results of an ELISA assay for the presence of human IgM and IgG1 in the serum of two animals that developed from embryos injected with the transgene insert of plasmid pHCl. All of the control non-transgenic mice tested negative for expression of human IgM and IgG1 by this assay. Mice from two lines containing the pIGM1 NotI insert (lines #6 and 15) express human IgM but not human IgG1. We tested mice from 6 lines that contain the pHCl insert and found that 4 of the lines (lines #26, 38, 57 and 122) express both human IgM and human IgG1, while mice from two of the lines (lines #19 and 21) do not express detectable levels of human immunoglobulins. The pHCl transgenic mice that did not express human immunoglobulins were so-called G₀ mice that developed directly from microinjected embryos and may have been mosaic for the presence of the transgene. Southern blot analysis indicates that many of these mice contain one or fewer copies of the transgene per cell. The detection of human IgM in the serum of pIGM1 transgenics, and human IgM and IgG1 in pHCl transgenics, provides evidence that the transgene sequences function correctly in directing VDJ joining, transcription, and isotype switching. One of the animals (#18) was negative for the transgene by Southern blot analysis, and showed no detectable levels of human IgM or IgG1. The second animal (#38) contained approximately 5 copies of the transgene, as assayed by Southern blotting, and showed detectable levels of both human IgM and IgG1. The results of ELISA assays for 11 animals that developed from transgene injected embryos is summarized in the table below (Table 3).

WO 98/24884

PCT/US97/21803

172

TABLE 3

Detection of human IgM and IgG1 in the serum of transgenic animals by ELISA assay

5

		injected	approximate		
	<u>animal #</u>	<u>transgene</u>	<u>transgene</u>	<u>human IgM</u>	<u>human IgG1</u>
			<u>copies per cell</u>		
10	6	pIGM1	1	++	-
	7	pIGM1	0	-	-
	9	pIGM1	0	-	-
15	10	pIGM1	0	-	-
	12	pIGM1	0	-	-
20	15	pIGM1	10	++	-
	18	pHC1	0	-	-
	19	pHC1	1	-	-
25	21	pHC1	<1	-	-
	26	pHC1	2	++	+
30	38	pHC1	5	++	+

Table 3 shows a correlation between the presence of integrated transgene DNA and the presence of transgene encoded immunoglobulins in the serum. Two of the animals that were found to contain the pHC1 transgene did not express detectable levels of human immunoglobulins. These were both low copy animals and may not have contained complete copies of the transgenes, or the animals may have been genetic mosaics

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(indicated by the <1 copy per cell estimated for animal #21), and the transgene containing cells may not have populated the hematopoietic lineage. Alternatively, the transgenes may have integrated into genomic locations that are not conducive to their expression. The detection of human IgM in the serum of pIGM1 transgenics, and human IgM and IgG1 in pHCl transgenics, indicates that the transgene sequences function correctly in directing VDJ joining, transcription, and isotype switching.

10 F. cDNA clones

To assess the functionality of the pHCl transgene in VDJ joining and class switching, as well the participation of the transgene encoded human B-cell receptor in B-cell development and allelic exclusion, the structure of immunoglobulin cDNA clones derived from transgenic mouse spleen mRNA were examined. The overall diversity of the transgene encoded heavy chains, focusing on D and J segment usage, N region addition, CDR3 length distribution, and the frequency of joints resulting in functional mRNA molecules was examined. Transcripts encoding IgM and IgG incorporating VH105 and VH251 were examined.

Polyadenylated RNA was isolated from an eleven week old male second generation line-57 pHCl transgenic mouse. This RNA was used to synthesize oligo-dT primed single stranded cDNA. The resulting cDNA was then used as template for four individual PCR amplifications using the following four synthetic oligonucleotides as primers: VH251 specific oligo-149, cta gct cga gtc caa gga gtc tgt gcc gag gtg cag ctg (g,a,t,c); VH105 specific o-150, gtt gct cga gtg aaa ggt gtc cag tgt gag gtg cag ctg (g,a,t,c); human gamma1 specific oligo-151, ggc gct cga gtt cca cga cac cgt cac cgg ttc; and human mu specific oligo-152, cct gct cga ggc agc caa cgg cca

WO 98/24884

PCT/US97/21803

174

cgc tgc tcg. Reaction 1 used primers 0-149 and o-151 to amplify VH251-gamma1 transcripts, reaction 2 used o-149 and o-152 to amplify VH251-mu transcripts, reaction 3 used o-150 and o-151 to amplify VH105-gamma1 transcripts, and reaction 4 used o-150 and o-152 to amplify VH105-mu transcripts. The resulting 0.5 kb PCR products were isolated from an agarose gel; the μ transcript products were more abundant than the γ transcript products, consistent with the corresponding ELISA data (Fig. 34). The PCR products were digested with XhoI and cloned into the plasmid pNN03. Double-stranded plasmid DNA was isolated from minipreps of nine clones from each of the four PCR amplifications and dideoxy sequencing reactions were performed. Two of the clones turned out to be deletions containing no D or J segments. These could not have been derived from normal RNA splicing products and are likely to have originated from deletions introduced during PCR amplification. One of the DNA samples turned out to be a mixture of two individual clones, and three additional clones did not produce readable DNA sequence (presumably because the DNA samples were not clean enough). The DNA sequences of the VDJ joints from the remaining 30 clones are compiled in Table 4. Each of the sequences are unique, indicating that no single pathway of gene rearrangement, or single clone of transgene expressing B-cells is dominant. The fact that no two sequences are alike is also an indication of the large diversity of immunoglobulins that can be expressed from a compact minilocus containing only 2 V segments, 10 D segments, and 6 J segments. Both of the V segments, all six of the J segments, and 7 of the 10 D segments that are included in the transgene are used in VDJ joints. In addition, both constant region genes (mu and gamma1) are incorporated into transcripts. The VH105 primer turned out not to be specific

WO 98/24884

PCT/US97/21803

175

for VH105 in the reactions performed. Therefore many of the clones from reactions 3 and 4 contained VH251 transcripts. Additionally, clones isolated from ligated reaction 3 PCR product turned out to encode IgM rather than IgG; however this
5 may reflect contamination with PCR product from reaction 4 as the DNA was isolated on the same gel. An analogous experiment, in which immunoglobulin heavy chain sequences were amplified from adult human peripheral blood lymphocytes (PBL), and the DNA sequence of the VDJ joints determined, was
10 recently reported by Yamada et al. (J. Exp. Med. 173:395-407 (1991), which is incorporated herein by reference). We compared the data from human PBL with our data from the pHCl transgenic mouse.

WO 98/24884

PCT/US97/21803

176

TABLE 4

WO 98/24884

PCT/US97/21803

177

G. J segment choice

Table 5 compared the distribution of J segments incorporated into pHCl transgene encoded transcripts to J segments found in adult human PBL immunoglobulin transcripts.

5 The distribution profiles are very similar, J4 is the dominant segment in both systems, followed by J6. J2 is the least common segment in human PBL and the transgenic animal.

TABLE 5

J. Segment Choice

10

15

20

<u>J. Segment</u>	Percent Usage ($\pm$ 3%)	
	<u>HC1 transgenic</u>	<u>Human PBL</u>
J1	7	1
J2	3	<1
J3	17	9
J4	44	53
J5	3	15
J6	<u>26</u>	<u>22</u>
	100%	100%

H. D segment choice

25

30

35

49% (40 of 82) of the clones analyzed by Yamada et al. incorporated D segments that are included in the pHCl transgene. An additional 11 clones contained sequences that were not assigned by the authors to any of the known D segments. Two of these 11 unassigned clones appear to be derived from an inversion of the DIR2 segments which is included in the pHCl construct. This mechanism, which was predicted by Ichihara et al. (EMBO J. 7:4141 (1988)) and observed by Sanz (J. Immunol. 147:1720-1729 (1991)), was not considered by Yamada et al. (J. Exp. Med. 173:395-407 (1991)).

Table 5 is a comparison of the D segment distribution for the pHCl transgenic mouse and that observed for human PBL transcripts by Yamada et al. The data of Yamada et al. was

WO 98/24884

PCT/US97/21803

178

recompiled to include DIR2 use, and to exclude D segments that are not in the pHCl transgene. Table 6 demonstrates that the distribution of D segment incorporation is very similar in the transgenic mouse and in human PBL. The two dominant human D segments, DXP'1 and DN1, are also found with high frequency in the transgenic mouse. The most dramatic dissimilarity between the two distributions is the high frequency of DHQ52 in the transgenic mouse as compared to the human. The high frequency of DHQ52 is reminiscent of the D segment distribution in the human fetal liver. Sanz has observed that 14% of the heavy chain transcripts contained DHQ52 sequences. If D segments not found in pHCl are excluded from the analysis, 31% of the fetal transcripts analyzed by Sanz contain DHQ52. This is comparable to the 27% that we observe in the pHCl transgenic mouse.

TABLE 6

D Segment Choice

	<u>D. Segment</u>	Percent Usage ($\pm$ 3%)	
		<u>HC1 transgenic</u>	<u>Human PBL</u>
20	DLR1	<1	<1
	DXP1	3	6
	DXP'1	25	19
	DA1	<1	12
25	DK1	7	12
	DN1	12	22
	DIR2	7	4
	DM2	<1	2
	DLR2	3	4
30	DHQ52	26	2
	?	<u>17</u>	<u>17</u>
		100%	100%

35

I. Functionality of VDJ joints

Table 7 shows the predicted amino acid sequences of the VDJ regions from 30 clones that were analyzed from the

WO 98/24884

PCT/US97/21803

179

pHC1 transgenic. The translated sequences indicate that 23 of the 30 VDJ joints (77%) are in-frame with respect to the variable and J segments.

WO 98/24884

PCT/US97/21803

180

TABLE 7

Functionality of V-D-J Joints

WO 98/24884

PCT/US97/21803

181

J. CDR3 length distribution

Table 8 compared the length of the CDR3 peptides from transcripts with in-frame VDJ joints in the pHCl transgenic mouse to those in human PBL. Again the human PBL data comes from Yamada et al. The profiles are similar with the transgenic profile skewed slightly toward smaller CDR3 peptides than observed from human PBL. The average length of CDR3 in the transgenic mouse is 10.3 amino acids. This is substantially the same as the average size reported for authentic human CDR3 peptides by Sanz (J. Immunol. 147:1720-1729 (1991)).

TABLE 8

CDR3 Length Distribution

15

		Percent Occurrence ($\pm$ 3%)	
	<u>#amino acids in CDR3</u>	<u>HC1 transgenic</u>	<u>Human PBL</u>
	3-8	26	14
20	9-12	48	41
	13-18	26	37
	19-23	<1	7
	>23	<u><1</u>	<u>1</u>
25		100%	100%

EXAMPLE 1330 Rearranged Heavy Chain TransgenesA. Isolation of Rearranged Human Heavy Chain VDJ segments.

Two human leukocyte genomic DNA libraries cloned into the phage vector λ EMBL3/SP6/T7 (Clontech Laboratories, Inc., Palo Alto, CA) are screened with a 1 kb PacI/HindIII fragment of λ 1.3 containing the human heavy chain J- μ intronic enhancer. Positive clones are tested for hybridization with a mixture of the following V_H specific oligonucleotides:

WO 98/24884

PCT/US97/21803

182

oligo-7 5'-tca gtg aag gtt tcc tgc aag gca tct gga tac acc
ttc acc-3'

oligo-8 5'-tcc ctg aga ctc tcc tgt gca gcc tct gga ttc acc
5 ttc agt-3'

Clones that hybridized with both V and J- μ probes are isolated and the DNA sequence of the rearranged VDJ segment determined.

10

B. Construction of rearranged human heavy chain transgenes

Fragments containing functional VJ segments (open reading frame and splice signals) are subcloned into the plasmid vector pSP72 such that the plasmid derived XhoI site is adjacent to the 5' end of the insert sequence. A subclone containing a functional VDJ segment is digested with XhoI and PacI (PacI, a rare-cutting enzyme, recognizes a site near the J-m intronic enhancer), and the insert cloned into XhoI/PacI digested pHC2 to generate a transgene construct with a functional VDJ segment, the J- μ intronic enhancer, the μ switch element, the μ constant region coding exons, and the γ 1 constant region, including the sterile transcript associated sequences, the γ 1 switch, and the coding exons. This transgene construct is excised with NotI and microinjected into the pronuclei of mouse embryos to generate transgenic animals as described above.

EXAMPLE 14

Light Chain Transgenes

30 A. Construction of Plasmid vectors

1. Plasmid vector pGP1c

WO 98/24884

PCT/US97/21803

183

Plasmid vector pGP1a is digested with NotI and the following oligonucleotides ligated in:

oligo-81 5'-ggc cgc atc ccg ggt ctc gag gtc gac aag ctt tcg
5 agg atc cgc-3'

oligo-82 5'-ggc cgc gga tcc tcg aaa gct tgt cga cct cga gac
ccg gga tgc-3'

10 The resulting plasmid, pGP1c, contains a polylinker with XmaI, XhoI, SalI, HindIII, and BamHI restriction sites flanked by NotI sites.

2. Plasmid vector pGP1d

15 Plasmid vector pGP1a is digested with NotI and the following oligonucleotides ligated in:

oligo-87 5'-ggc cgc tgt cga caa gct tat cga tgg atc ctc gag
tgc -3'

20

oligo-88 5'-ggc cgc act cga gga tcc atc gat aag ctt gtc gac
agc -3'

The resulting plasmid, pGP1d, contains a polylinker with SalI,
25 HindIII, ClaI, BamHI, and XhoI restriction sites flanked by NotI sites.

B. Isolation of JK and Ck clones

A human placental genomic DNA library cloned into
30 the phage vector λ EMBL3/SP6/T7 (Clonetech Laboratories, Inc., Palo Alto, CA) was screened with the human kappa light chain J region specific oligonucleotide:

WO 98/24884

PCT/US97/21803

184

oligo-36 5'- cac ctt cgg cca agg gac acg act gga gat taa acg
taa gca -3'

and the phage clones 136.2 and 136.5 isolated. A 7.4 kb XhoI
5 fragment that includes the Jk1 segment was isolated from
136.2 and subcloned into the plasmid pNNO3 to generate the
plasmid clone p36.2. A neighboring 13 kb XhoI fragment that
includes Jk segments 2 through 5 together with the Ck gene
segment was isolated from phage clone 136.5 and subcloned into
10 the plasmid pNNO3 to generate the plasmid clone p36.5.
Together these two clones span the region beginning 7.2 kb
upstream of Jk1 and ending 9 kb downstream of Ck.

C. Construction of rearranged light chain transgenes

15 1. pCK1, a Ck vector for expressing rearranged variable
segments

The 13 kb XhoI insert of plasmid clone p36.5
containing the Ck gene, together with 9 kb of downstream
sequences, is cloned into the SalI site of plasmid vector
20 pGP1c with the 5' end of the insert adjacent to the plasmid
XhoI site. The resulting clone, pCK1 can accept cloned
fragments containing rearranged VJk segments into the unique
5' XhoI site. The transgene can then be excised with NotI and
purified from vector sequences by gel electrophoresis. The
25 resulting transgene construct will contain the human J-Ck
intronic enhancer and may contain the human 3' k enhancer.

2. pCK2, a Ck vector with heavy chain enhancers for
expressing rearranged variable segments

30 A 0.9 kb XbaI fragment of mouse genomic DNA
containing the mouse heavy chain J- μ intronic enhancer (J.
Banerji et al., Cell 33:729-740 (1983)) was subcloned into

pUC18 to generate the plasmid pJH22.1. This plasmid was linearized with SphI and the ends filled in with Klenow enzyme. The Klenow treated DNA was then digested with HindIII and a 1.4 kb MluI/HindIII fragment of phage clone λ 1.3 (previous example), containing the human heavy chain J- μ intronic enhancer (Hayday et al., Nature 307:334-340 (1984)), to it. The resulting plasmid, pMHE1, consists of the mouse and human heavy chain J- μ intronic enhancers ligated together into pUC18 such that they are excised on a single BamHI/HindIII fragment. This 2.3 kb fragment is isolated and cloned into pGP1c to generate pMHE2. pMHE2 is digested with SalI and the 13 kb XhoI insert of p36.5 cloned in. The resulting plasmid, pCK2, is identical to pCK1, except that the mouse and human heavy chain J- μ intronic enhancers are fused to the 3' end of the transgene insert. To modulate expression of the final transgene, analogous constructs can be generated with different enhancers, i.e. the mouse or rat 3' kappa or heavy chain enhancer (Meyer and Neuberger, EMBO J., 8:1959-1964 (1989); Petterson et al., Nature, 344:165-168 (1990)).

3. Isolation of rearranged kappa light chain variable segments

Two human leukocyte genomic DNA libraries cloned into the phage vector λ EMBL3/SP6/T7 (Clontech Laboratories, Inc., Palo Alto, CA) were screened with the human kappa light chain J region containing 3.5 kb XhoI/SmaI fragment of p36.5. Positive clones were tested for hybridization with the following Vk specific oligonucleotide:

30

oligo-65 5'-agg ttc agt ggc agt ggg tct ggg aca gac ttc act
ctc acc atc agc-3'

Clones that hybridized with both V and J probes are isolated and the DNA sequence of the rearranged VJk segment determined.

4. Generation of transgenic mice containing rearranged human
5 light chain constructs.

Fragments containing functional VJ segments (open reading frame and splice signals) are subcloned into the unique XhoI sites of vectors pCK1 and pCK2 to generate rearranged kappa light chain transgenes. The transgene
10 constructs are isolated from vector sequences by digestion with NotI. Agarose gel purified insert is microinjected into mouse embryo pronuclei to generate transgenic animals. Animals expressing human kappa chain are bred with heavy chain minilocus containing transgenic animals to generate mice
15 expressing fully human antibodies.

Because not all VJk combinations may be capable of forming stable heavy-light chain complexes with a broad spectrum of different heavy chain VDJ combinations, several different light chain transgene constructs are generated, each
20 using a different rearranged VJk clone, and transgenic mice that result from these constructs are bred with heavy chain minilocus transgene expressing mice. Peripheral blood, spleen, and lymph node lymphocytes are isolated from double transgenic (both heavy and light chain constructs) animals,
25 stained with fluorescent antibodies specific for human and mouse heavy and light chain immunoglobulins (PharMingen, San Diego, CA) and analyzed by flow cytometry using a FACScan analyzer (Becton Dickinson, San Jose, CA). Rearranged light chain transgenes constructs that result in the highest level
30 of human heavy/light chain complexes on the surface of the highest number of B cells, and do not adversely affect the immune cell compartment (as assayed by flow cytometric

analysis with B and T cell subset specific antibodies), are selected for the generation of human monoclonal antibodies.

D. Construction of unrearranged light chain minilocus transgenes

1. pJCK1, a Jk, Ck containing vector for constructing minilocus transgenes

The 13 kb Ck containing XhoI insert of p36.5 is treated with Klenow enzyme and cloned into HindIII digested, Klenow-treated, plasmid pGP1d. A plasmid clone is selected such that the 5' end of the insert is adjacent to the vector derived ClaI site. The resulting plasmid, p36.5-1d, is digested with ClaI and Klenow-treated. The Jk1 containing 7.4 kb XhoI insert of p36.2 is then Klenow-treated and cloned into the ClaI, Klenow-treated p36.5-1d. A clone is selected in which the p36.2 insert is in the same orientation as the p36.5 insert. This clone, pJCK1 (Fig. 34), contains the entire human Jk region and Ck, together with 7.2 kb of upstream sequences and 9 kb of downstream sequences. The insert also contains the human J-Ck intronic enhancer and may contain a human 3' κ enhancer. The insert is flanked by a unique 3' SalI site for the purpose of cloning additional 3' flanking sequences such as heavy chain or light chain enhancers. A unique XhoI site is located at the 5' end of the insert for the purpose of cloning in unrearranged Vk gene segments. The unique SalI and XhoI sites are in turn flanked by NotI sites that are used to isolate the completed transgene construct away from vector sequences.

2. Isolation of unrearranged Vk gene segments and generation of transgenic animals expressing human Ig light chain protein

The Vk specific oligonucleotide, oligo-65 (discussed above), is used to probe a human placental genomic DNA library cloned into the phage vector lEMBL3/SP6/T7 (Clonetech Laboratories, Inc., Palo Alto, CA). Variable gene segments from the resulting clones are sequenced, and clones that appear functional are selected. Criteria for judging functionality include: open reading frames, intact splice acceptor and donor sequences, and intact recombination sequence. DNA fragments containing selected variable gene segments are cloned into the unique XhoI site of plasmid pJCK1 to generate minilocus constructs. The resulting clones are digested with NotI and the inserts isolated and injected into mouse embryo pronuclei to generate transgenic animals. The transgenes of these animals will undergo V to J joining in developing B-cells. Animals expressing human kappa chain are bred with heavy chain minilocus containing transgenic animals to generate mice expressing fully human antibodies.

EXAMPLE 15

20 Genomic Heavy Chain Human Ig Transgene

This Example describes the cloning of a human genomic heavy chain immunoglobulin transgene which is then introduced into the murine germline via microinjection into zygotes or integration in ES cells.

25 Nuclei are isolated from fresh human placental tissue as described by Marzluff, W.F., et al. (1985), Transcription and Translation: A Practical Approach, B.D. Hammes and S.J. Higgins, eds., pp. 89-129, IRL Press, Oxford).

The isolated nuclei (or PBS washed human spermatocytes) are embedded in 0.5% low melting point agarose blocks and lysed with 1 mg/ml proteinase K in 500mM EDTA, 1% SDS for nuclei, or with 1mg/ml proteinase K in 500mM EDTA, 1% SDS, 10mM DTT for

spermatocytes at 50°C for 18 hours. The proteinase K is inactivated by incubating the blocks in 40µg/ml PMSF in TE for 30 minutes at 50°C, and then washing extensively with TE. The DNA is then digested in the agarose with the restriction enzyme NotI as described by M. Finney in Current Protocols in Molecular Biology (F. Ausubel et al., eds. John Wiley & Sons, Supp. 4, 1988, e.g., Section 2.5.1).

The NotI digested DNA is then fractionated by pulsed field gel electrophoresis as described by Anand et al., Nuc. Acids Res. 17:3425-3433 (1989). Fractions enriched for the NotI fragment are assayed by Southern hybridization to detect one or more of the sequences encoded by this fragment. Such sequences include the heavy chain D segments, J segments, and γ 1 constant regions together with representatives of all 6 V_H families (although this fragment is identified as 670 kb fragment from HeLa cells by Berman et al. (1988), supra., we have found it to be an 830 kb fragment from human placental and sperm DNA). Those fractions containing this NotI fragment are ligated into the NotI cloning site of the vector pYACNN as described (McCormick et al., Technique 2:65-71 (1990)). Plasmid pYACNN is prepared by digestion of pYACneo (Clontech) with EcoRI and ligation in the presence of the oligonucleotide 5' - AAT TGC GGC CGC - 3'.

YAC clones containing the heavy chain NotI fragment are isolated as described by Traver et al., Proc. Natl. Acad. Sci. USA, 86:5898-5902 (1989). The cloned NotI insert is isolated from high molecular weight yeast DNA by pulse field gel electrophoresis as described by M. Finney, op. cit. The DNA is condensed by the addition of 1 mM spermine and microinjected directly into the nucleus of single cell embryos previously described. Alternatively, the DNA is isolated by pulsed field gel electrophoresis and introduced into ES cells

by lipofection (Gnirke et al., EMBO J. 10:1629-1634 (1991)), or the YAC is introduced into ES cells by spheroplast fusion.

EXAMPLE 16

5 Discontinuous Genomic Heavy Chain Ig Transgene

An 85 kb SpeI fragment of human genomic DNA, containing V_H6, D segments, J segments, the μ constant region and part of the γ constant region, has been isolated by YAC cloning essentially as described in Example 1. A YAC carrying
10 a fragment from the germline variable region, such as a 570 kb NotI fragment upstream of the 670-830 kb NotI fragment described above containing multiple copies of V₁ through V₅, is isolated as described. (Berman et al. (1988), supra. detected two 570 kb NotI fragments, each containing multiple V
15 segments.) The two fragments are coinjected into the nucleus of a mouse single cell embryo as described in Example 1.

Typically, coinjection of two different DNA fragments result in the integration of both fragments at the same insertion site within the chromosome. Therefore,
20 approximately 50% of the resulting transgenic animals that contain at least one copy of each of the two fragments will have the V segment fragment inserted upstream of the constant region containing fragment. Of these animals, about 50% will carry out V to DJ joining by DNA inversion and about 50% by
25 deletion, depending on the orientation of the 570 kb NotI fragment relative to the position of the 85 kb SpeI fragment. DNA is isolated from resultant transgenic animals and those animals found to be containing both transgenes by Southern blot hybridization (specifically, those animals containing
30 both multiple human V segments and human constant region genes) are tested for their ability to express human

immunoglobulin molecules in accordance with standard techniques.

EXAMPLE 17

5 Identification of functionally rearranged variable region sequences in transgenic B cells

An antigen of interest is used to immunize (see Harlow and Lane, Antibodies: A Laboratory Manual, Cold Spring Harbor, New York (1988)) a mouse with the following genetic
10 traits: homozygosity at the endogenous heavy chain locus for a deletion of J_H (Examples 10); hemizygous for a single copy of unrearranged human heavy chain minilocus transgene (examples 5 and 14); and hemizygous for a single copy of a rearranged human kappa light chain transgene (Examples 6 and 14).

15 Following the schedule of immunization, the spleen is removed, and spleen cells used to generate hybridomas. Cells from an individual hybridoma clone that secretes antibodies reactive with the antigen of interest are used to prepare genomic DNA. A sample of the genomic DNA is digested
20 with several different restriction enzymes that recognize unique six base pair sequences, and fractionated on an agarose gel. Southern blot hybridization is used to identify two DNA fragments in the 2-10 kb range, one of which contains the single copy of the rearranged human heavy chain VDJ sequences
25 and one of which contains the single copy of the rearranged human light chain VJ sequence. These two fragments are size fractionated on agarose gel and cloned directly into pUC18. The cloned inserts are then subcloned respectively into heavy and light chain expression cassettes that contain constant
30 region sequences.

The plasmid clone p γ e1 (Example 12) is used as a heavy chain expression cassette and rearranged VDJ sequences

are cloned into the XhoI site. The plasmid clone pCK1 is used as a light chain expression cassette and rearranged VJ sequences are cloned into the XhoI site. The resulting clones are used together to transfect SP₀ cells to produce antibodies
5 that react with the antigen of interest (Co. et al., Proc. Natl. Acad. Sci. USA 88:2869 (1991), which is incorporated herein by reference).

Alternatively, mRNA is isolated from the cloned hybridoma cells described above, and used to synthesize cDNA.
10 The expressed human heavy and light chain VDJ and VJ sequence are then amplified by PCR and cloned (Larrick et al., Biol. Technology, 7:934-938 (1989)). After the nucleotide sequence of these clones has been determined, oligonucleotides are synthesized that encode the same polypeptides, and synthetic
15 expression vectors generated as described by Queen et al., Proc. Natl. Acad. Sci. USA, 84:5454-5458 (1989).

Immunization of Transgenic Animals with Complex Antigens

The following experiment demonstrates that
20 transgenic animals can be successfully immunized with complex antigens such as those on human red blood cells and respond with kinetics that are similar to the response kinetics observed in normal mice.

Blood cells generally are suitable immunogens and
25 comprise many different types of antigens on the surface of red and white blood cells.

Immunization with human blood

Tubes of human blood from a single donor were
30 collected and used to immunize transgenic mice having functionally disrupted endogenous heavy chain loci (J_HD) and harboring a human heavy chain minigene construct (HC1); these

WO 98/24884

PCT/US97/21803

193

mice are designated as line 112. Blood was washed and resuspended in 50 mls Hanks' and diluted to 1×10^8 cells/ml. 0.2 mls (2×10^7 cells) were then injected interperitoneally using a 28 gauge needle and 1 cc syringe. This immunization protocol was repeated approximately weekly for 6 weeks. Serum titers were monitored by taking blood from retro-orbital bleeds and collecting serum and later testing for specific antibody. A pre-immune bleed was also taken as a control. On the very last immunization, three days before these animals were sacrificed for serum and for hybridomas, a single immunization of 1×10^7 cells was given intravenously through the tail to enhance the production of hybridomas.

Table 915 Animals

	Mouse ID	Line	Sex	HC1-112	JHD
1	2343	112	M	+	++
2	2344	112	M	-	+
3	2345	112	F	-	+
4	2346	112	F	-	++
5	2347	112	F	-	++
6	2348	112	F	+	++
7	2349	112	F	-	+

Mice # 2343 and 2348 have a desired phenotype: human heavy chain mini-gene transgenic on heavy chain knock-out background.

Generation of Hybridomas

Hybridomas were generated by fusing mouse spleen cells of approximately 16 week-old transgenic mice (Table 9) that had been immunized as described (supra) to a fusion partner consisting of the non-secreting HAT-sensitive myeloma cell line, X63 Ag8.653. Hybridoma clones were cultivated and hybridoma supernatants containing immunoglobulins having specific binding affinity for blood cell antigens were identified, for example, by flow cytometry.

Flow cytometry

Serum and hybridoma supernatants were tested using flow cytometry. Red blood cells from the donor were washed 4X in Hanks' balanced salt solution and 50,000 cells were placed in 1.1 ml polypropylene microtubes. Cells were incubated with antisera or supernatant from the hybridomas for 30 minutes on ice in staining media (1x RPMI 1640 media without phenol red or biotin (Irvine Scientific) 3% newborn calf serum, 0.1% Na azide). Controls consisted of littermate mice with other genotypes. Cells were then washed by centrifugation at 4°C in Sorvall RT600B for 5-10 minutes at 1000 rpm. Cells were washed two times and then antibody detected on the cell surface with a fluorescent developing reagent. Two monoclonal reagents were used to test. One was a FITC-labeled mouse anti-human μ heavy chain antibody (Pharmagen, San Diego, CA) and the other was a PE-labeled rat anti-mouse kappa light chain (Becton-Dickenson, San Jose, CA). Both of these reagents gave similar results. Whole blood (red blood cells and white blood cells) and white blood cells alone were used as target cells. Both sets gave positive results.

Serum of transgenic mice and littermate controls was incubated with either red blood cells from the donor, or white

WO 98/24884

PCT/US97/21803

195

blood cells from another individual, washed and then developed with anti-human IgM FITC labeled antibody and analyzed in a flow cytometer. Results showed that serum from mice that are transgenic for the human mini-gene locus (mice 2343 and 2348) show human IgM reactivity whereas all littermate animals (2344, 2345, 2346, 2347) do not. Normal mouse serum (NS) and phosphate buffer saline (PBS) were used as negative controls. Red blood cells were ungated and white blood cells were gated to include only lymphocytes. Lines are drawn on the x and y axis to provide a reference. Flow cytometry was performed on 100 supernatants from fusion 2348. Four supernatants showed positive reactivity for blood cell antigens.

EXAMPLE 18

15 Reduction of Endogenous Mouse Immunoglobulin Expression by Antisense RNA

A. Vector for Expression of Antisense Ig Sequences

1. Construction of the cloning vector pGP1h

The vector pGP1b (referred to in a previous example) is digested with XhoI and BamHI and ligated with the following oligonucleotides:

5'- gat cct cga gac cag gta cca gat ctt gtg aat tcg -3'

5'- tcg acg aat tca caa gat ctg gta cct ggt ctc gag -3'

25

to generate the plasmid pGP1h. This plasmid contains a polylinker that includes the following restriction sites: NotI, EcoRI, BglII, Asp718, XhoI, BamHI, HindIII, NotI.

30 Construction of pBCE1.

A 0.8 kb XbaI/BglII fragment of pVH251 (referred to in a previous example), that includes the promoter leader

sequence exon, first intron, and part of the second exon of the human VH-V family immunoglobulin variable gene segment, was inserted into XbaI/BglII digested vector pNN03 to generate the plasmid pVH251.

5 The 2.2 kb BamHI/EcoRI DNA fragment that includes the coding exons of the human growth hormone gene (hGH; Seeburg, (1982) DNA 1:239-249) is cloned into BglII/EcoRI digested pGH1h. The resulting plasmid is digested with BamHI and the BamHI/BglII of pVH251N is inserted in the same
10 orientation as the hGH gene to generate the plasmid pVhgh.

 A 0.9 kb XbaI fragment of mouse genomic DNA containing the mouse heavy chain J- μ intronic enhancer (Banerji et al., (1983) Cell 33:729-740) was subcloned into pUC18 to generate the plasmid pJH22.1. This plasmid was
15 linearized with SphI and the ends filled in with klenow enzyme. The klenow treated DNA was then digested with HindIII and a 1.4 kb MluI(klenow)/HindIII fragment of phage clone λ 1.3 (previous example), containing the human heavy chain J- μ intronic enhancer (Hayday et al., (1984) Nature 307:334-340),
20 to it. The resulting plasmid, pMHE1, consists of the mouse and human heavy chain J- μ intron enhancers ligated together into pUC18 such that they can be excised on a single BamHI/HindIII fragment.

 The BamHI/HindIII fragment of pMHE1 is cloned into
25 BamHI/HindIII cut pVhgh to generate the B-cell expression vector pBCE1. This vector, depicted in Fig. 36, contains unique XhoI and Asp718 cloning sites into which antisense DNA fragments can be cloned. The expression of these antisense sequences is driven by the upstream heavy chain promoter-
30 enhancer combination the downstream hGH gene sequences provide polyadenylation sequences in addition to intron sequences that promote the expression of transgene constructs. Antisense

transgene constructs generated from pBCE1 can be separated from vector sequences by digestion with NotI.

B. An IgM antisense transgene construct.

5 The following two oligonucleotides:

5'- cgc ggt acc gag agt cag tcc ttc cca aat gtc -3'

5'- cgc ctc gag aca gct gga atg ggc aca tgc aga -3'

10 are used as primers for the amplification of mouse IgM constant region sequences by polymerase chain reaction (PCR) using mouse spleen cDNA as a substrate. The resulting 0.3 kb PCR product is digested with Asp718 and XhoI and cloned into Asp718/XhoI digested pBCE1 to generate the antisense transgene
15 construct pMAS1. The purified NotI insert of pMAS1 is microinjected into the pronuclei of half day mouse embryos-- alone or in combination with one or more other transgene constructs--to generate transgenic mice. This construct expresses an RNA transcript in B-cells that hybridizes with
20 mouse IgM mRNA, thus down-regulating the expression of mouse IgM protein. Double transgenic mice containing pMAS1 and a human heavy chain transgene minilocus such as pHCl (generated either by coinjection of both constructs or by breeding of singly transgenic mice) will express the human transgene
25 encoded Ig receptor on a higher percentage of B-cell than mice transgenic for the human heavy chain minilocus alone. The ratio of human to mouse Ig receptor expressing cells is due in part to competition between the two populations for factors and cells that promoter B-cell differentiation and expansion.
30 Because the Ig receptor plays a key role in B-cell development, mouse Ig receptor expressing B-cells that express reduced levels of IgM on their surface (due to mouse Ig

specific antisense down-regulation) during B-cell development will not compete as well as cells that express the human receptor.

5 C. An IgKappa antisense transgene construct.

The following two oligonucleotides:

5'- cgc ggt acc gct gat gct gca cca act gta tcc -3'

5'- cgc ctc gag cta aca ctc att cct gtt gaa gct -3'

10

are used as primers for the amplification of mouse IgKappa constant region sequences by polymerase chain reaction (PCR) using mouse spleen cDNA as a substrate. The resulting 0.3 kb PCR product is digested with Asp718 and XhoI and cloned into
 15 Asp718/XhoI digested pBCE1 to generate the antisense transgene construct pKAS1. The purified NotI insert of pKAS1 is microinjected into the pronuclei of half day mouse embryos-- alone or in combination with one or more other transgene constructs--to generate transgenic mice. This construct
 20 expresses an RNA transcript in B-cells that hybridizes with mouse IgK mRNA, thus down-regulating the expression of mouse IgK protein as described above for pMAS1.

EXAMPLE 19

25 This example demonstrates the successful immunization and immune response in a transgenic mouse of the present invention.

Immunization of Mice

30 Keyhole limpet hemocyanin conjugated with greater than 400 dinitrophenyl groups per molecule (Calbiochem, La Jolla, California) (KLH-DNP) was alum precipitated according

to a previously published method (Practical Immunology, L. Hudson and F.C. Hay, Blackwell Scientific (Pubs.), p. 9, 1980). Four hundred μ g of alum precipitated KLH-DNP along with 100 μ g dimethyldioctadecyl Ammonium Bromide in 100 μ L of phosphate buffered saline (PBS) was injected intraperitoneally into each mouse. Serum samples were collected six days later by retro-orbital sinus bleeding.

Analysis of Human Antibody Reactivity in Serum

Antibody reactivity and specificity were assessed using an indirect enzyme-linked immunosorbent assay (ELISA). Several target antigens were tested to analyze antibody induction by the immunogen. Keyhole limpet hemocyanin (Calbiochem) was used to identify reactivity against the protein component, bovine serum albumin-DNP for reactivity against the hapten and/or modified amino groups, and KLH-DNP for reactivity against the total immunogen. Human antibody binding to antigen was detected by enzyme conjugates specific for IgM and IgG sub-classes with no cross reactivity to mouse immunoglobulin. Briefly, PVC microtiter plates were coated with antigen drying overnight at 37°C of 5 μ g/mL protein in PBS. Serum samples diluted in PBS, 5% chicken serum, 0.5% Tween-20 were incubated in the wells for 1 hour at room temperature, followed by anti-human IgG Fc and IgG F(ab')-horseradish peroxidase or anti-human IgM Fc-horseradish peroxidase in the same diluent. After 1 hour at room temperature enzyme activity was assessed by addition of ABTS substrate (Sigma, St. Louis, Missouri) and read after 30 minutes at 415-490 nm.

Human Heavy Chain Participation in Immune Response in Transgenic Mice

experiment. The observed human μ chain Ig response followed

WO 98/24884

PCT/US97/21803

200

Figure 10A-10D

WO 98/24884

PCT/US97/21803

202

response which continues to rise is characteristic of the pattern seen with affinity maturation. Analysis of Day 21 samples showed lack of reactivity to an unrelated antigen, keyhole limpet hemocyanin (KLC), indicating that the antibody response was directed against CEA in a specific manner.

These data indicate that animals transgenic for human unrearranged immunoglobulin gene loci: (1) can respond to a human antigen (e.g., the human glycoprotein, CEA), (2) can undergo isotype switching ("class switching) as exemplified by the observed μ to γ class switch, and (3) exhibit characteristics of affinity maturation in their humoral immune responses. In general, these data indicate: (1) the human Ig transgenic mice have the ability to induce heterologous antibody production in response to a defined antigen, (2) the capacity of a single transgene heavy chain variable region to respond to a defined antigen, (3) response kinetics over a time period typical of primary and secondary response development, (4) class switching of a transgene-encoded humoral immune response from IgM to IgG, and (5) the capacity of transgenic animal to produce human-sequence antibodies against a human antigen.

Demonstration of somatic mutation in a human heavy chain transgene minilocus.

Line HC1-57 transgenic mice, containing multiple copies of the HC1 transgene, were bred with immunoglobulin heavy chain deletion mice to obtain mice that contain the HC1 transgene and contain disruptions at both alleles of the endogenous mouse heavy chain (supra). These mice express human μ and γ 1 heavy chains together with mouse kappa and lambda light chains (supra). One of these mice was

DEMANDES OU BREVETS VOLUMINEUX

LA PRÉSENTE PARTIE DE CETTE DEMANDE OU CE BREVET
COMPREND PLUS D'UN TOME.

CECI EST LE TOME 1 DE 2

NOTE: Pour les tomes additionels, veuillez contacter le Bureau canadien des brevets

JUMBO APPLICATIONS/PATENTS

THIS SECTION OF THE APPLICATION/PATENT CONTAINS MORE
THAN ONE VOLUME

THIS IS VOLUME 1 OF 2

NOTE: For additional volumes-please contact the Canadian Patent Office

WHAT IS CLAIMED IS:

1. A hybridoma comprising a B cell obtained from a transgenic mouse having a genome comprising a human heavy chain transgene and a human light chain transgene, said B cell fused to an immortalized cell suitable to generate a hybridoma, wherein said hybridoma produces a detectable amount of an immunoglobulin that specifically binds interleukin-8.
2. The hybridoma of Claim 1 wherein the immunoglobulin specifically binds $\text{GRO}\alpha$.
3. The hybridoma of Claim 1, wherein the immunoglobulin has an affinity constant (K_a) of at least $2 \times 10^9 \text{ M}^{-1}$ for binding human interleukin-8.
4. The hybridoma of Claim 1, wherein the affinity constant is at least $2 \times 10^{10} \text{ M}^{-1}$.
5. The hybridoma of Claim 3 wherein the immunoglobulin is selected from the group consisting of 1F8, 2D11, 2F9, 2G1, 3E5, 5E7, 5F10, 5H8, 2C6, 2D6, 3A1, 4D4, 7C5, and 10A6.
6. A composition comprising a substantially pure immunoglobulin produced by the hybridoma of claim 3.
7. A composition comprising a substantially pure human monoclonal antibody, wherein said antibody has an affinity constant (K_a) of at least $2 \times 10^9 \text{ M}^{-1}$ for binding human interleukin-8, and wherein said immunoglobulin consists of:

a human sequence light chain composed of (1) a light chain variable region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human V_L gene segment and a human J_L segment, and (2) a light chain constant region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a C_L gene segment; and

a human sequence heavy chain composed of (1) a heavy chain variable region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human V_H gene segment, optionally a D region, and a human J_H segment, and (2) a constant region having a polypeptide sequence which is substantially identical to a polypeptide sequence encoded by a human C_H gene segment.

8. The composition of claim 7 wherein the human monoclonal antibody specifically binds $GRO\alpha$.

9. A method of preventing efflux of neutrophils from vasculature in a patient, comprising administering an effective amount of a human monoclonal antibody having an affinity constant (K_a) of at least $2 \times 10^9 M^{-1}$ for binding human interleukin-8.

10. A method of treating reperfusion injury comprising administering to a patient a therapeutically effective dose of a human monoclonal antibody having an affinity constant (K_a) of at least $2 \times 10^9 M^{-1}$ for binding human interleukin-8.

11. A method of suppressing a T-helper cell dependent immune response in a primate, comprising

administering a therapeutically effective dose of a human monoclonal antibody having an affinity constant (K_a) of at least $2 \times 10^9 \text{ M}^{-1}$ for binding human CD-4.

12. The method of claim 11 wherein the primate is a chimpanzee.

13. The method of claim 11 wherein the antibody comprises a VH4-34 segment, a JH5 segment, a heavy chain CDR3 region comprising the sequence VINWFDP, a VkL19 segment, a Jk2 segment, and a light chain CD3 region comprising the sequence QQANSFPYT.

14. The method of claim 13 wherein the antibody is 6G5.

15. The method of claim 11 wherein the antibody comprises a VH5-51 segment, a JH2 segment, a heavy chain CDR3 region comprising the sequence PANWNWYFVL, a VkL18 segment, a Jk4 segment, and a light chain CD3 region comprising the sequence QQFISYPQLT.

16. The method of claim 15 wherein the antibody is 1G2.

WO 98/24884

PCT/US97/21803

1/101

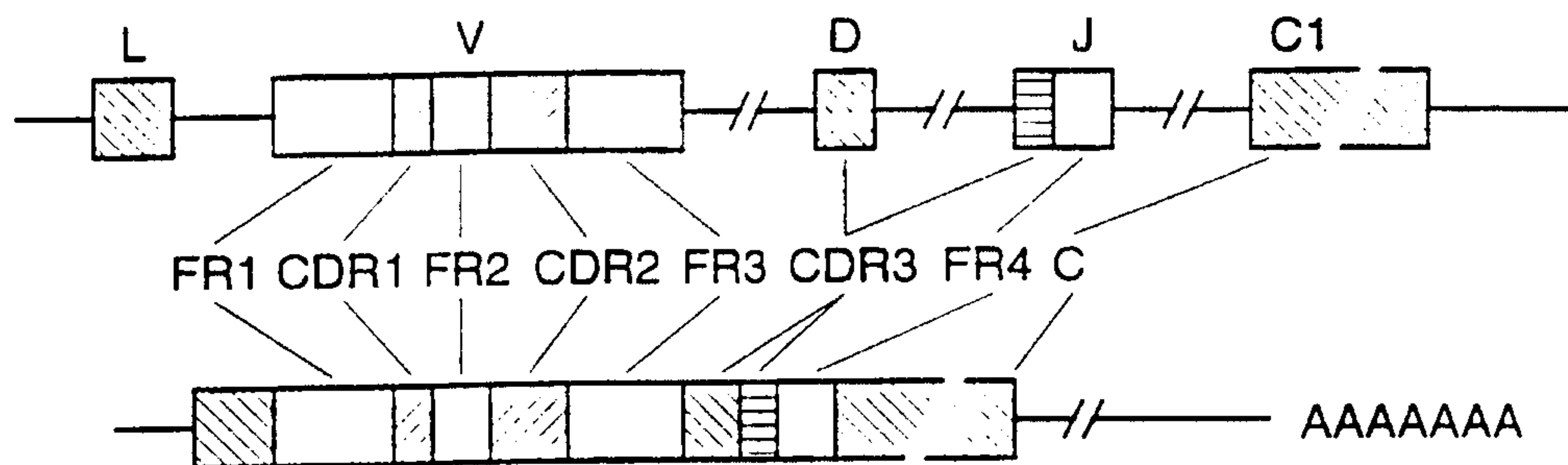


FIG. 1

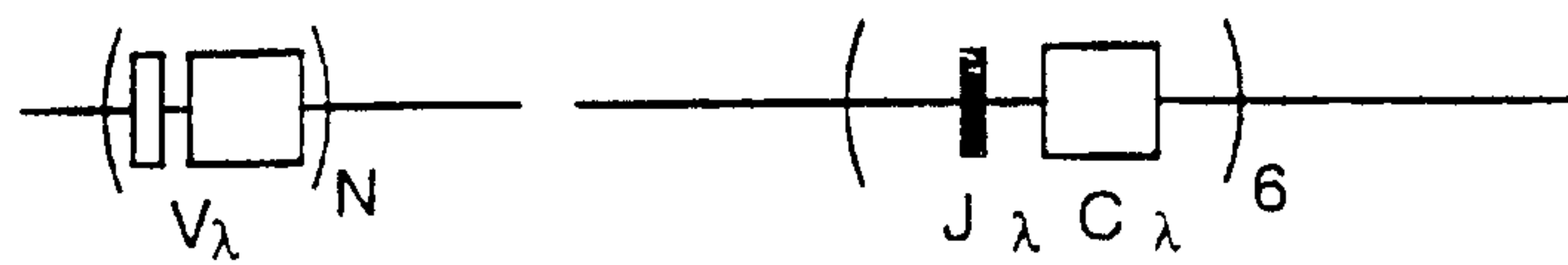


FIG. 2

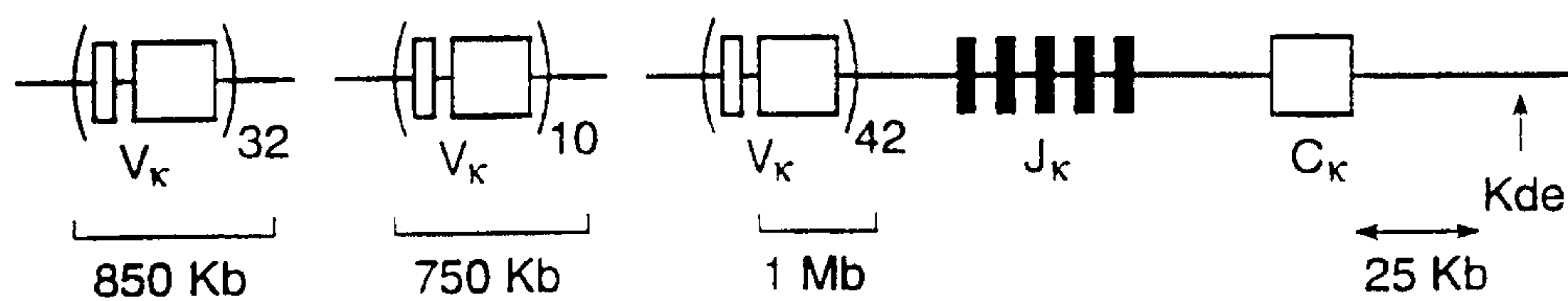


FIG. 3

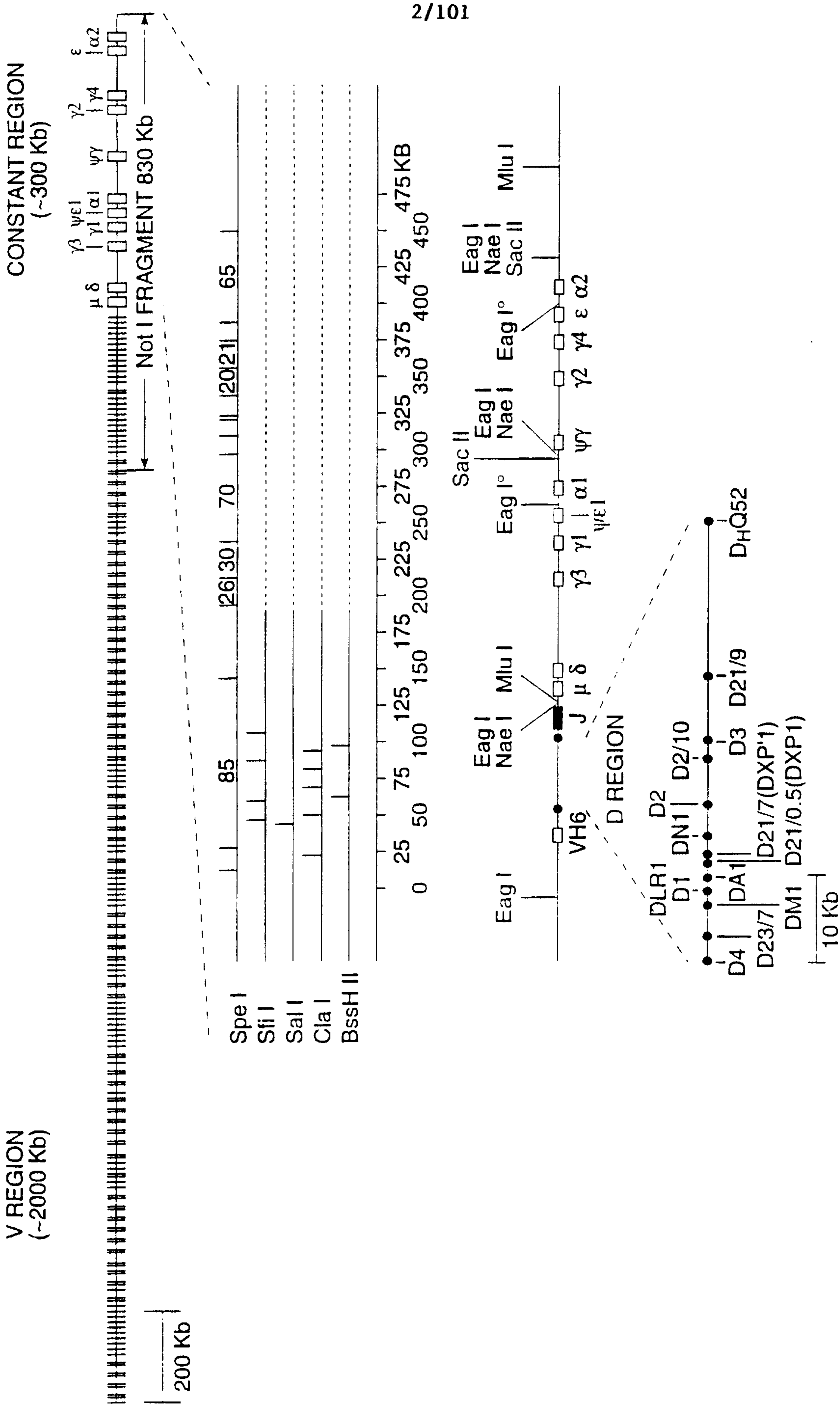


FIG. 4

3/101

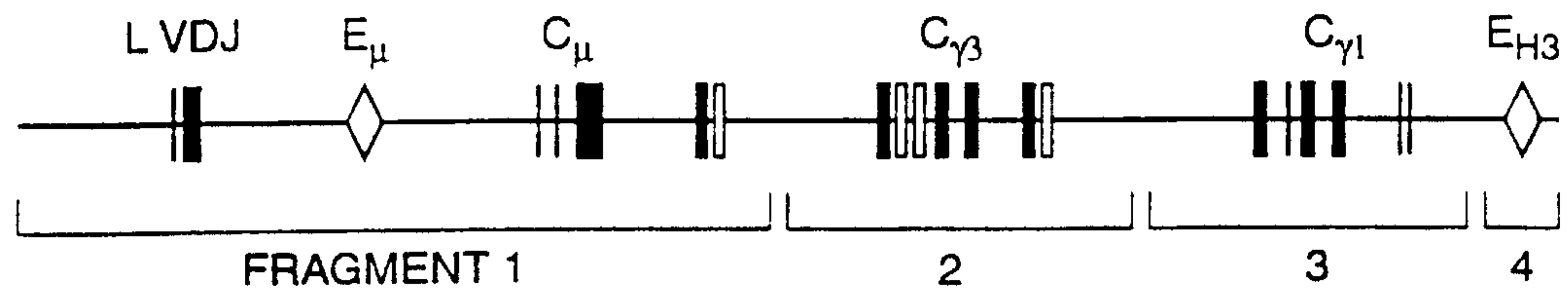


FIG. 5

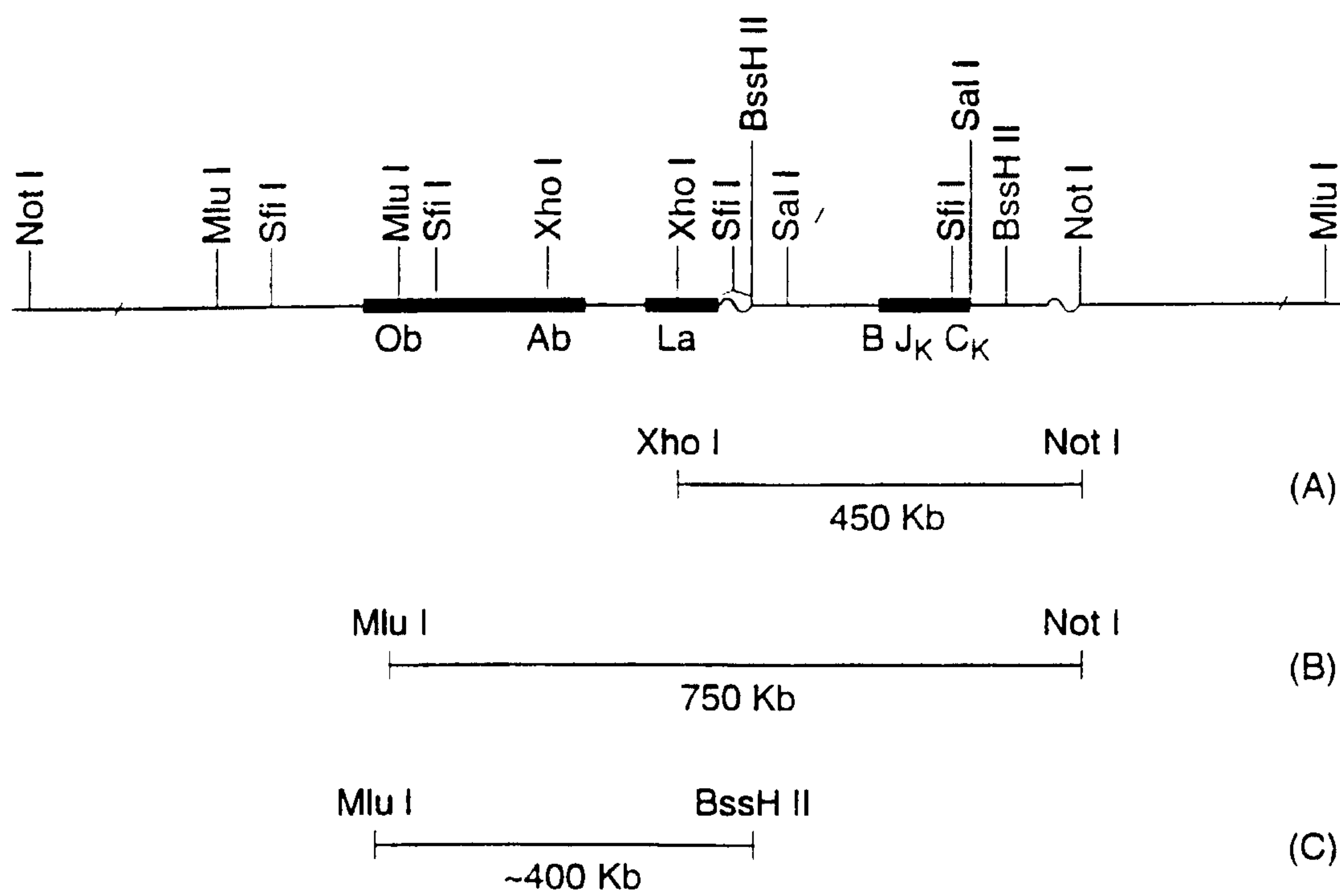


FIG. 6

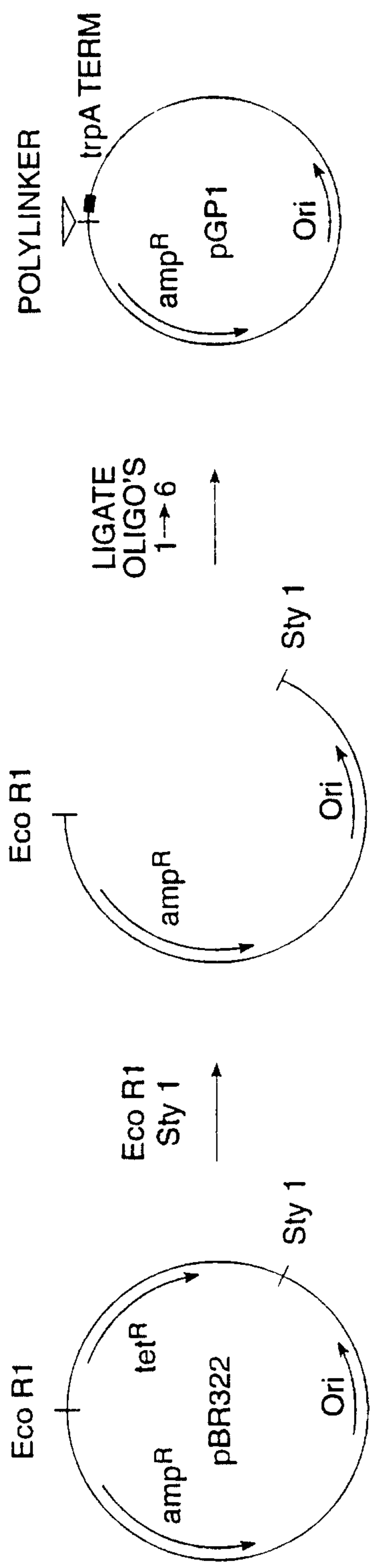


FIG. 7

WO 98/24884

PCT/US97/21803

5/101

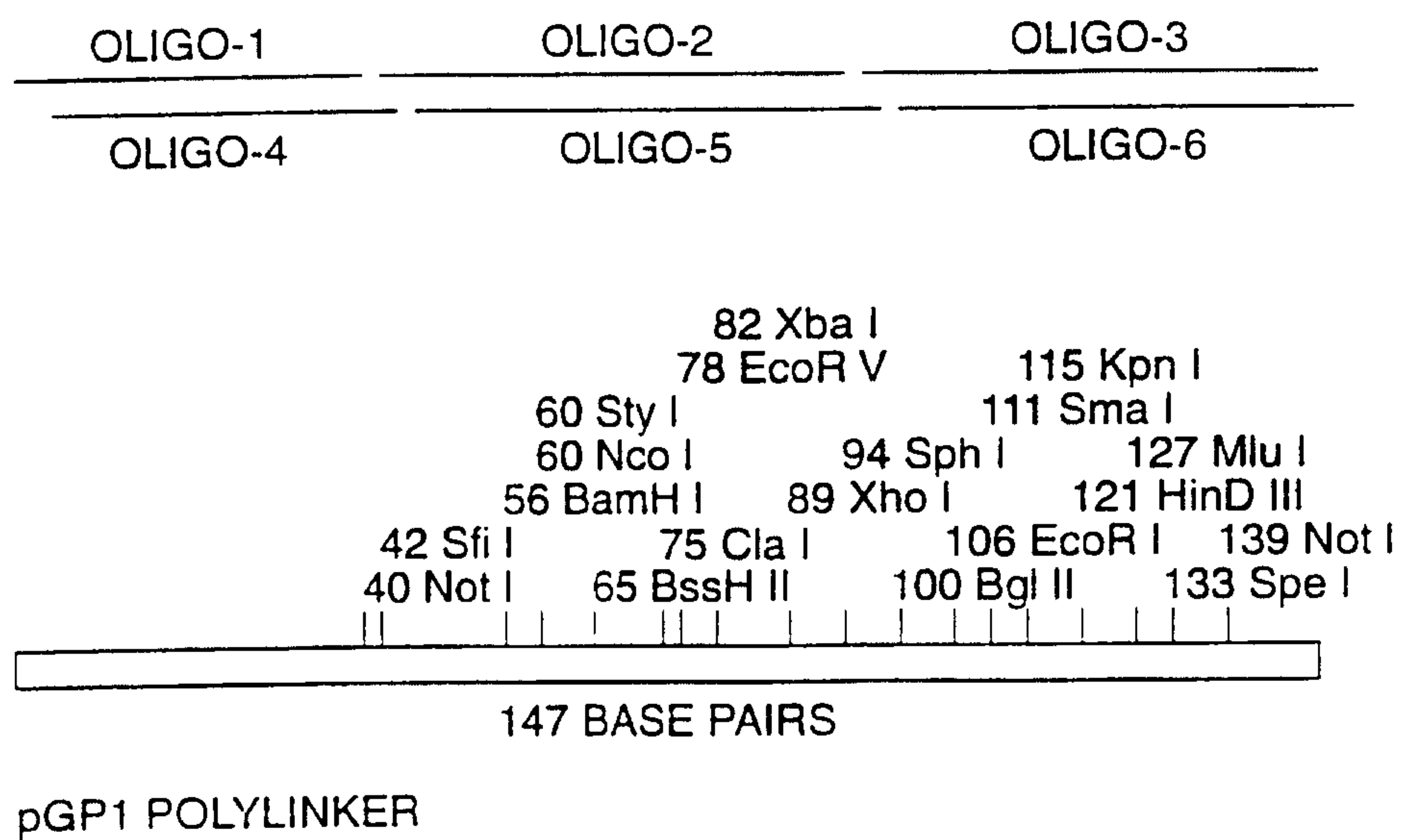


FIG. 8

FIG. 9

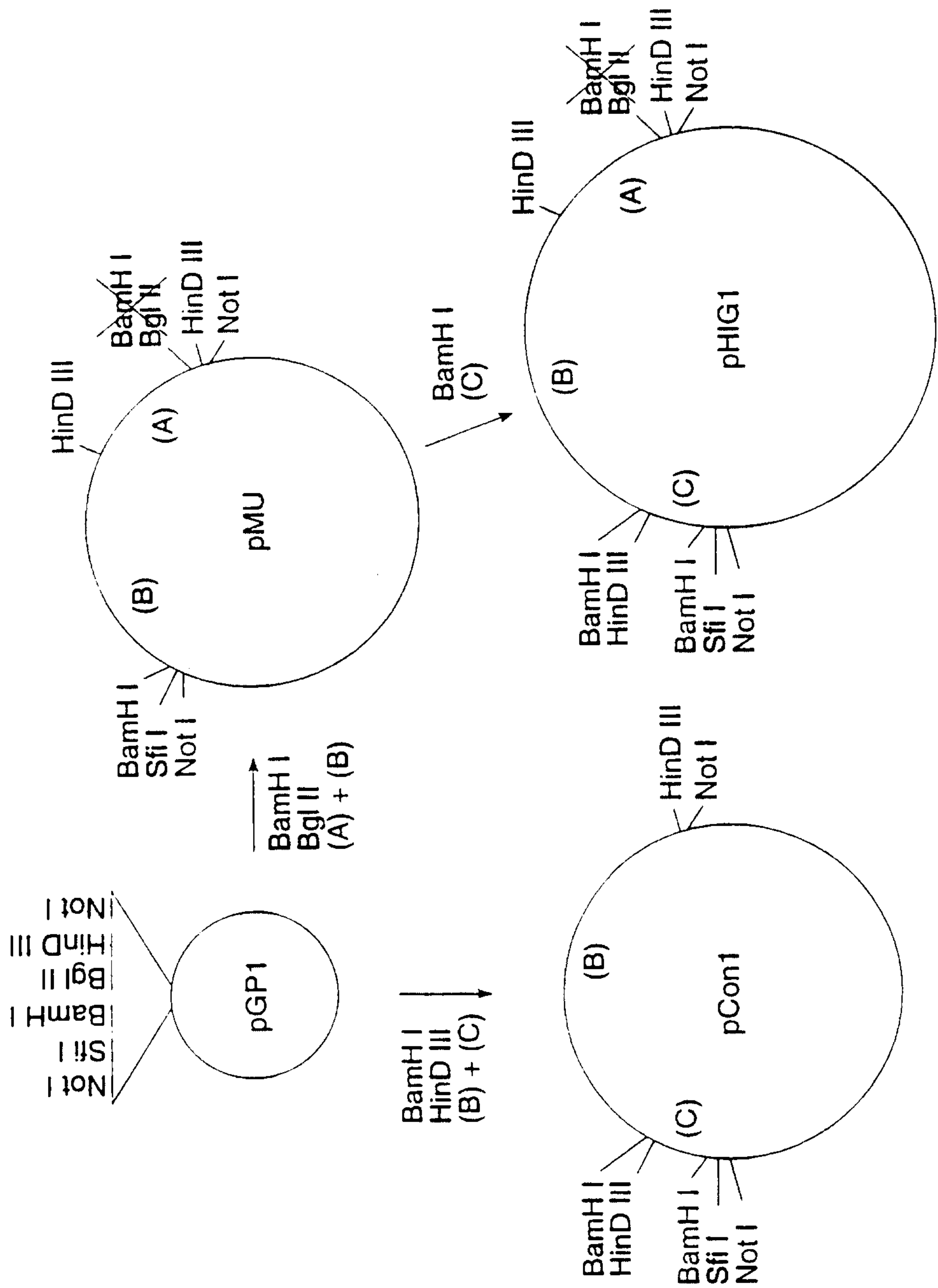


FIG. 10

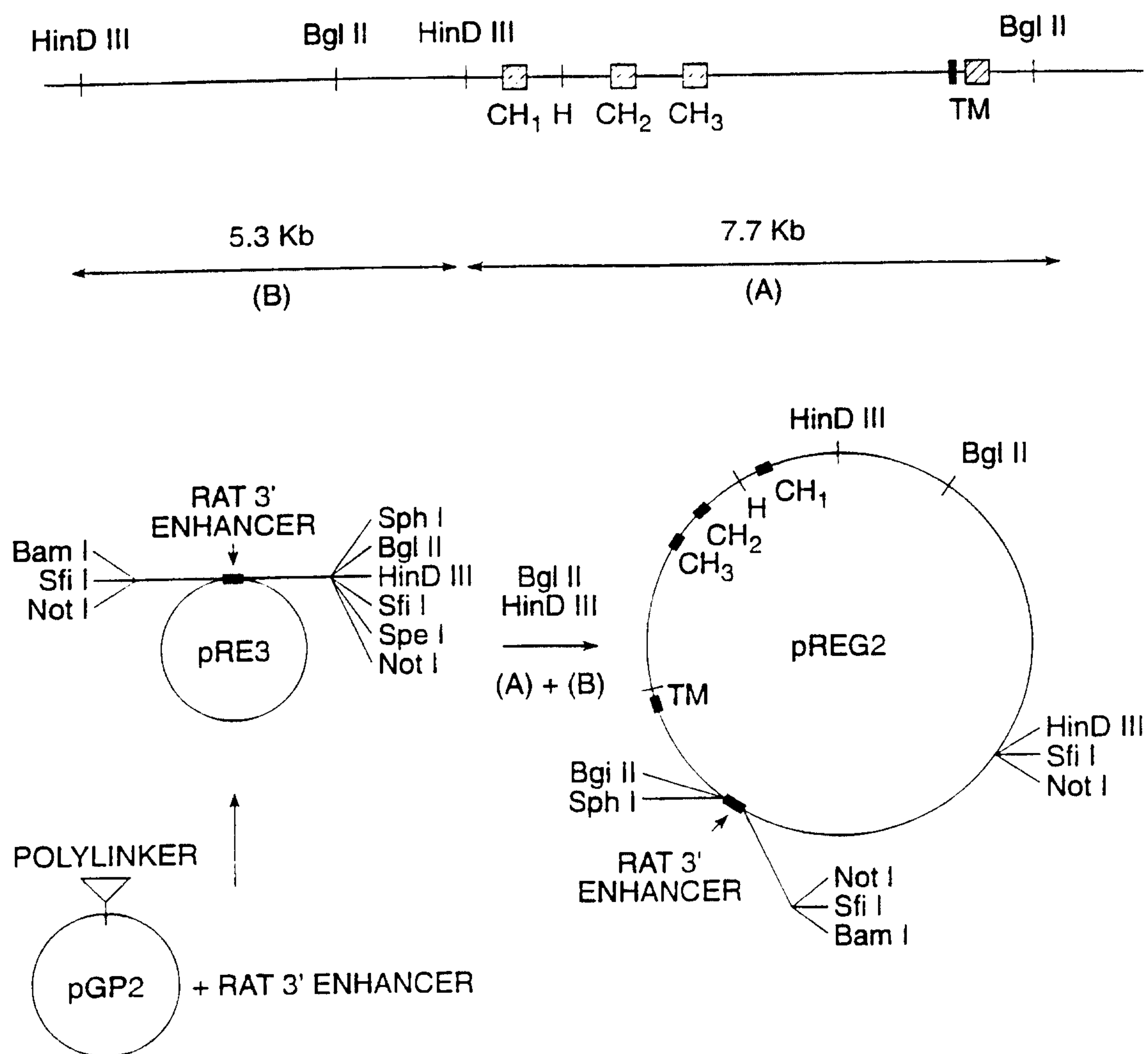
HUMAN C_{γ1} GENE

FIG. 11

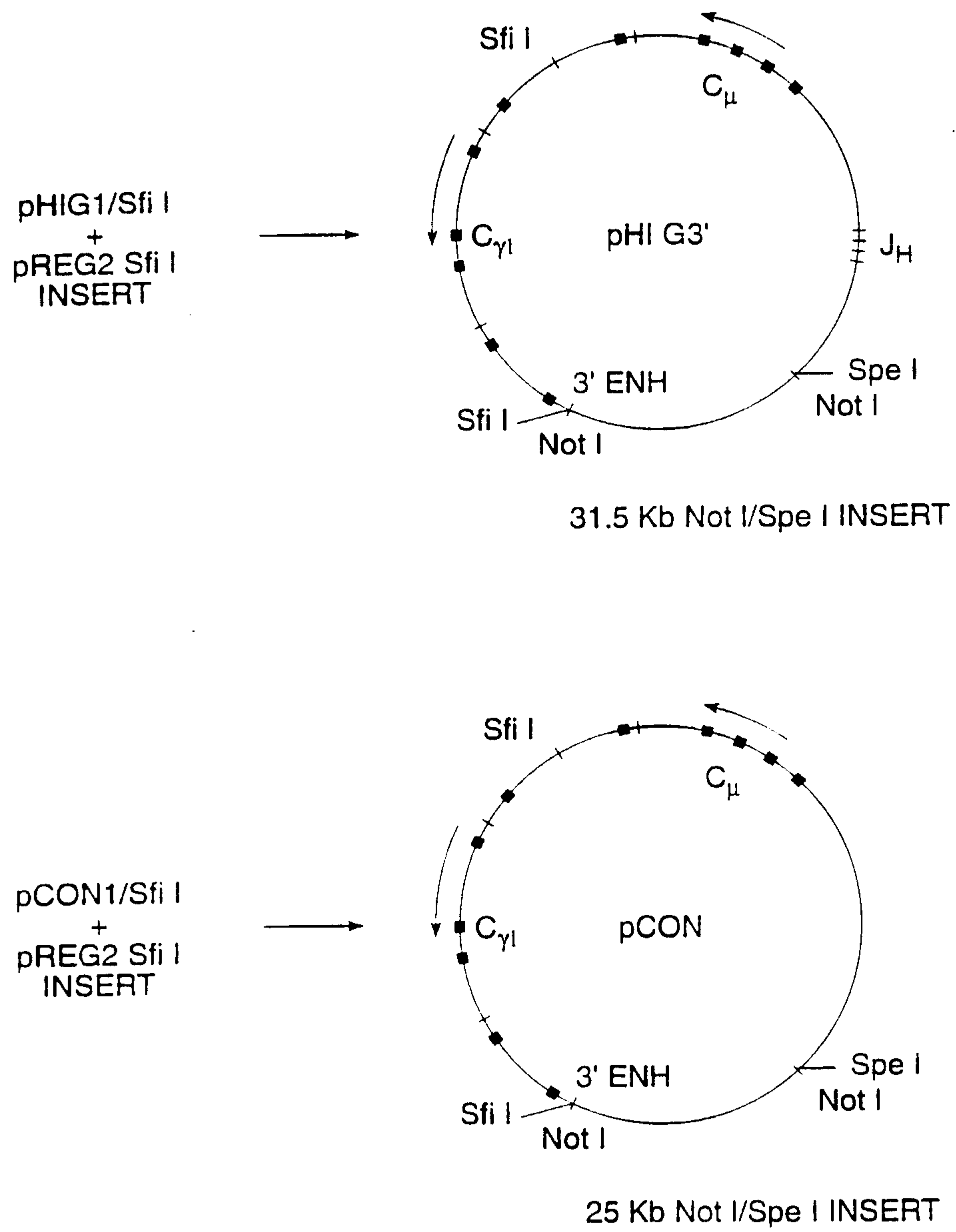


FIG. 12

HUMAN D REGION

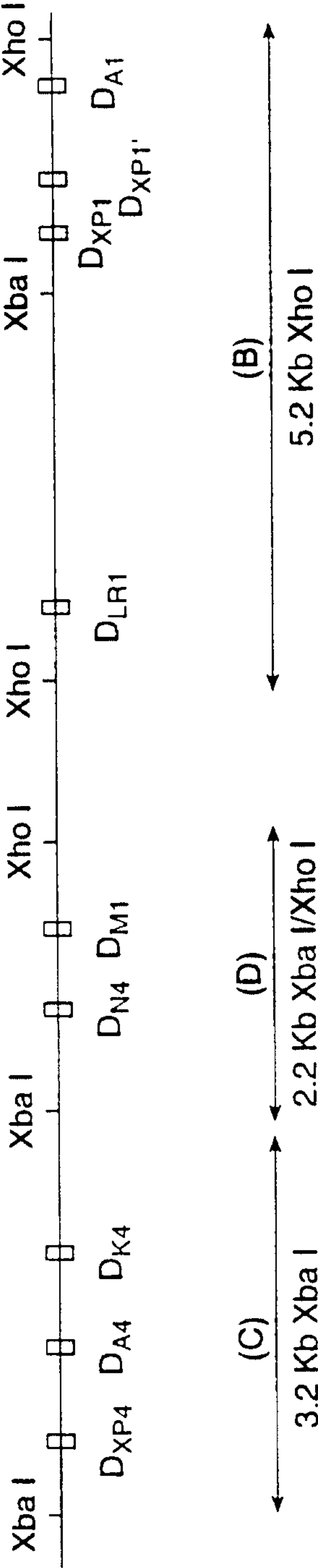


FIG. 13

11/101

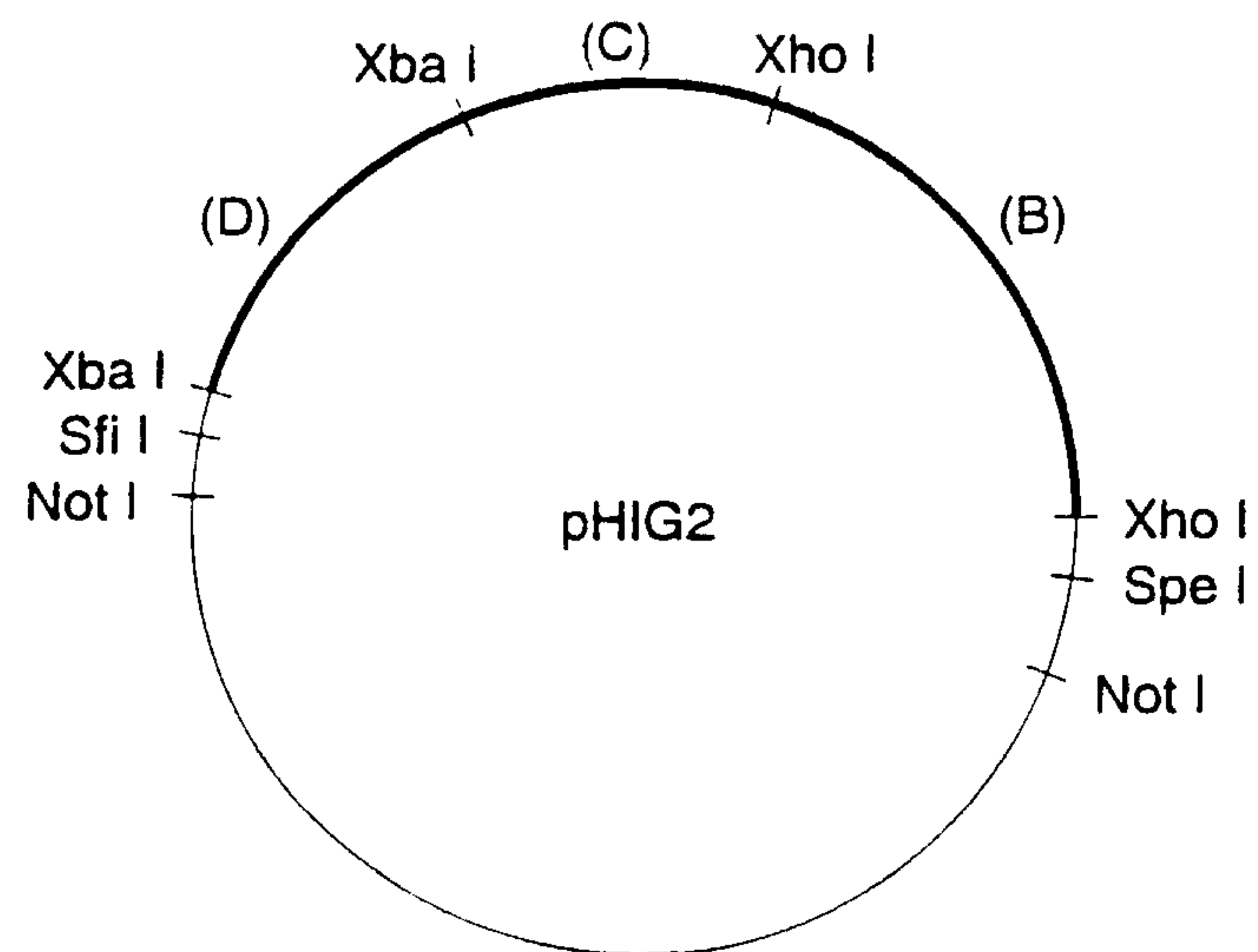


FIG. 14

12/101

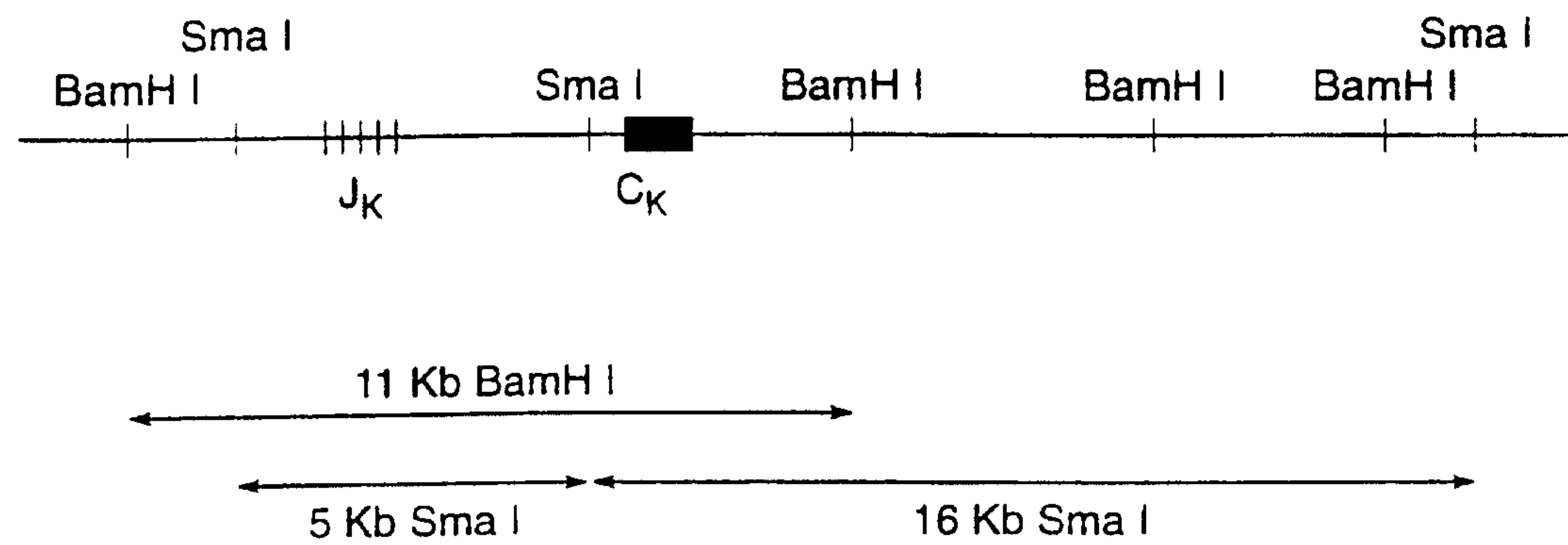


FIG. 15

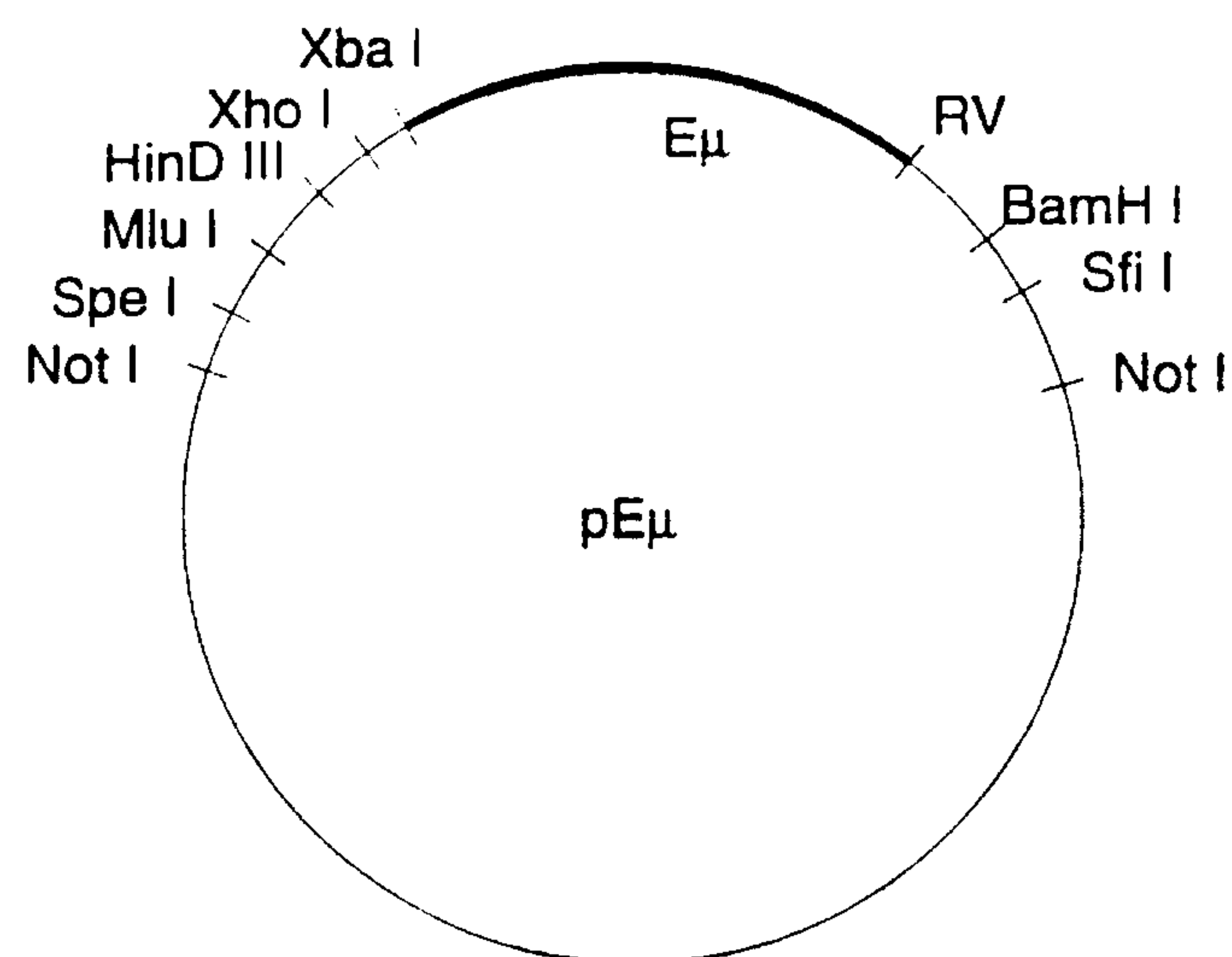
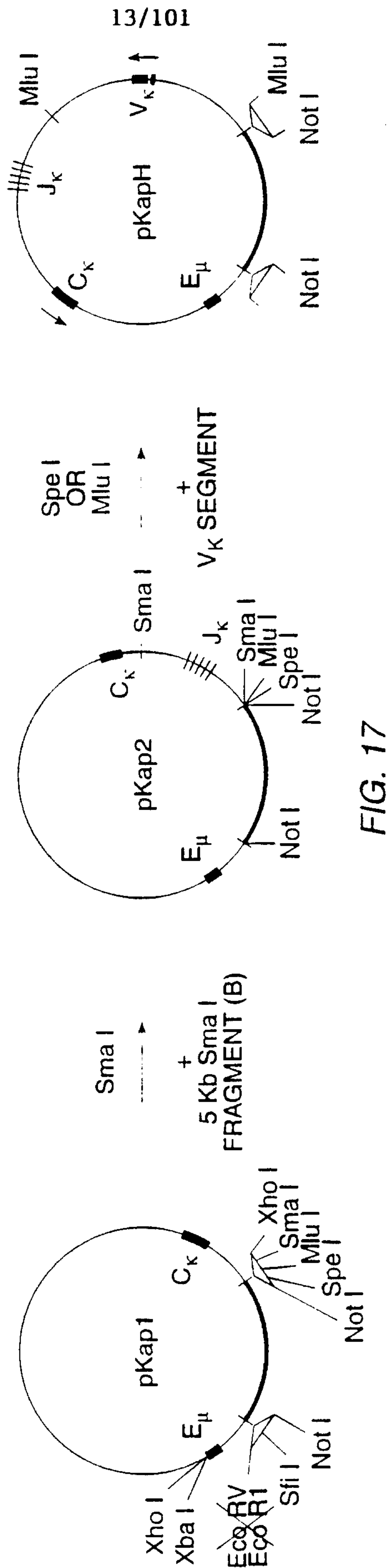


FIG. 16



MOUSE HEAVY CHAIN LOCUS

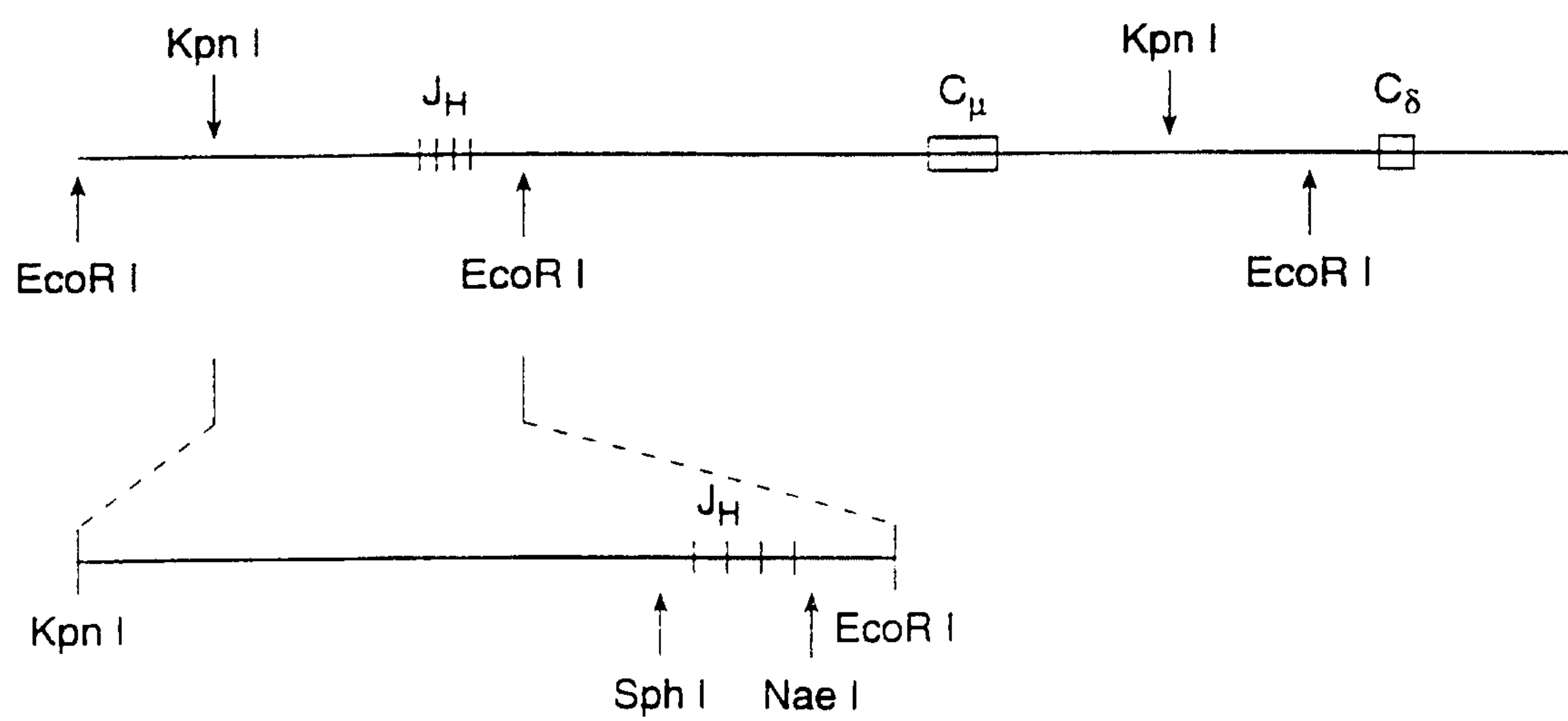


FIG. 18A

15/101

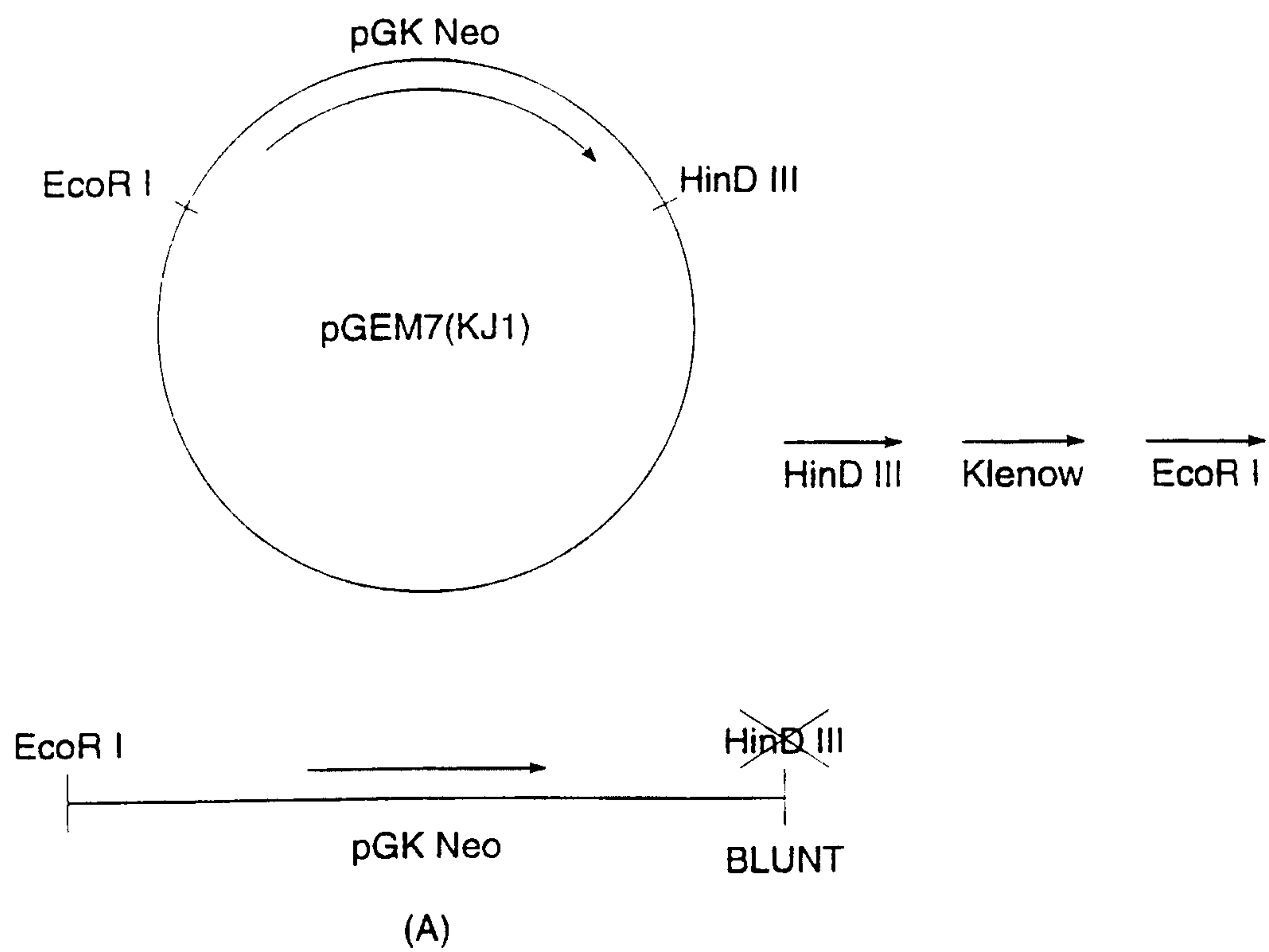


FIG. 18B

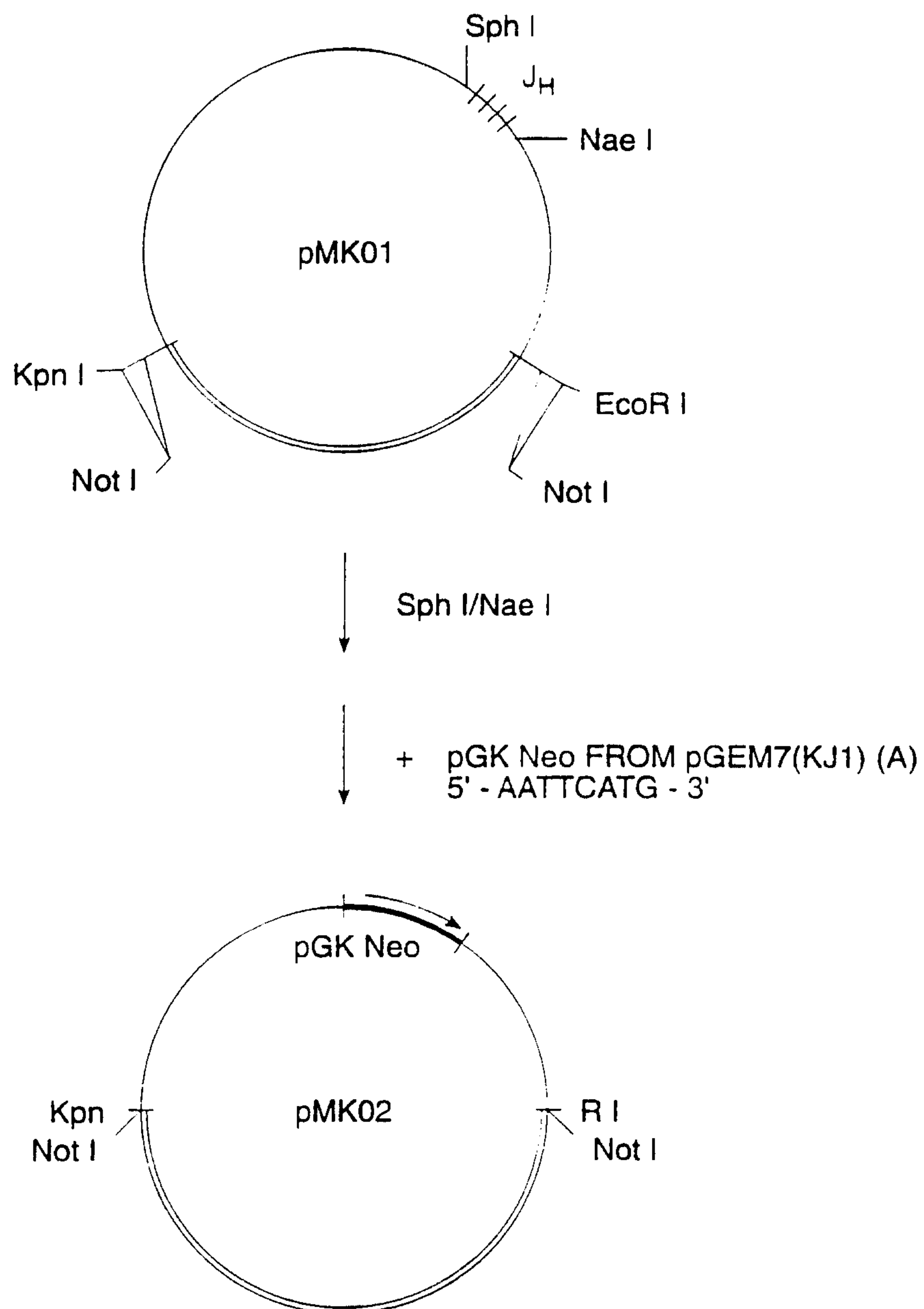


FIG. 18C

17/101

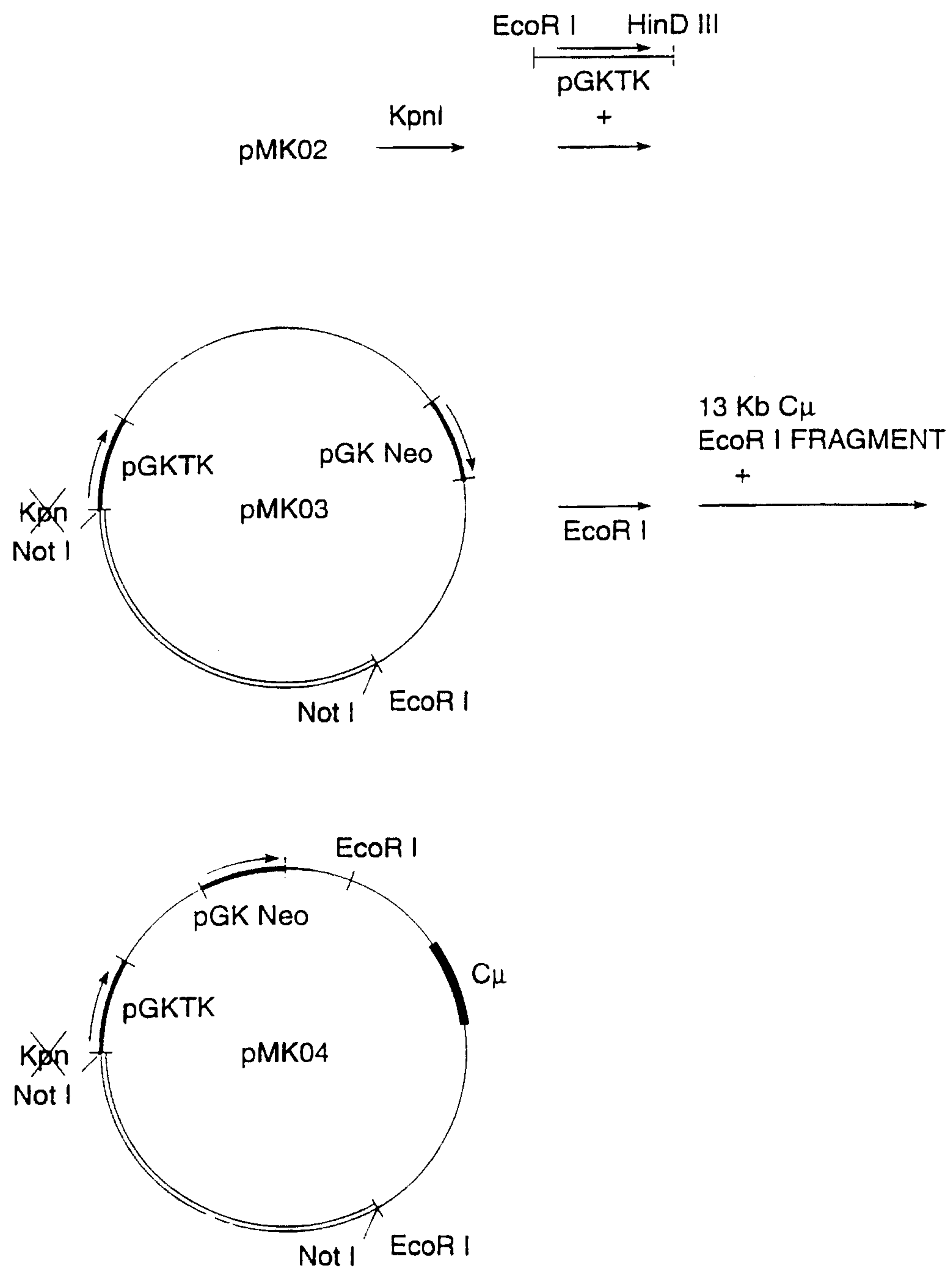


FIG. 18D

MOUSE KAPPA GENE

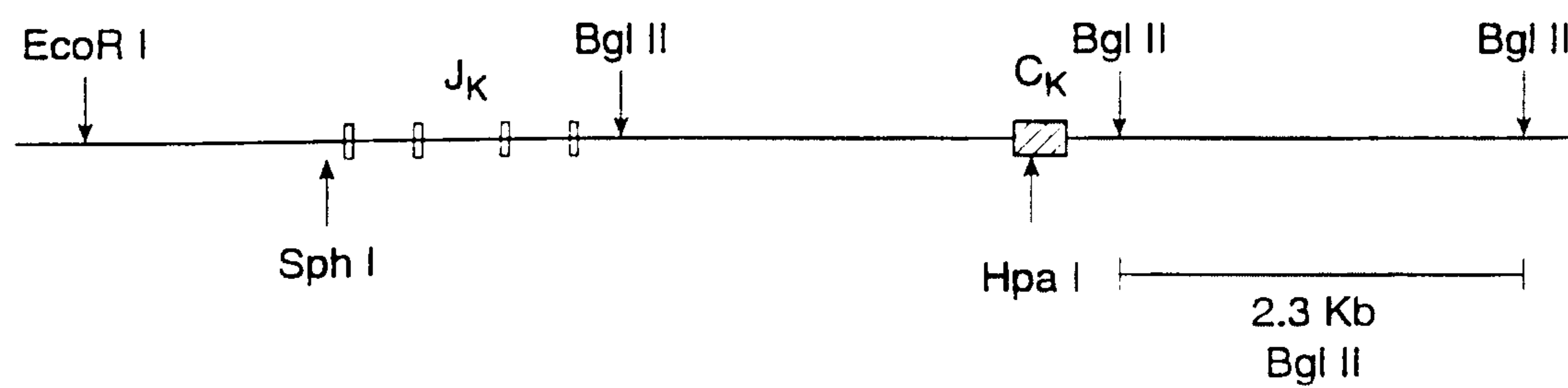


FIG. 19A

19/101

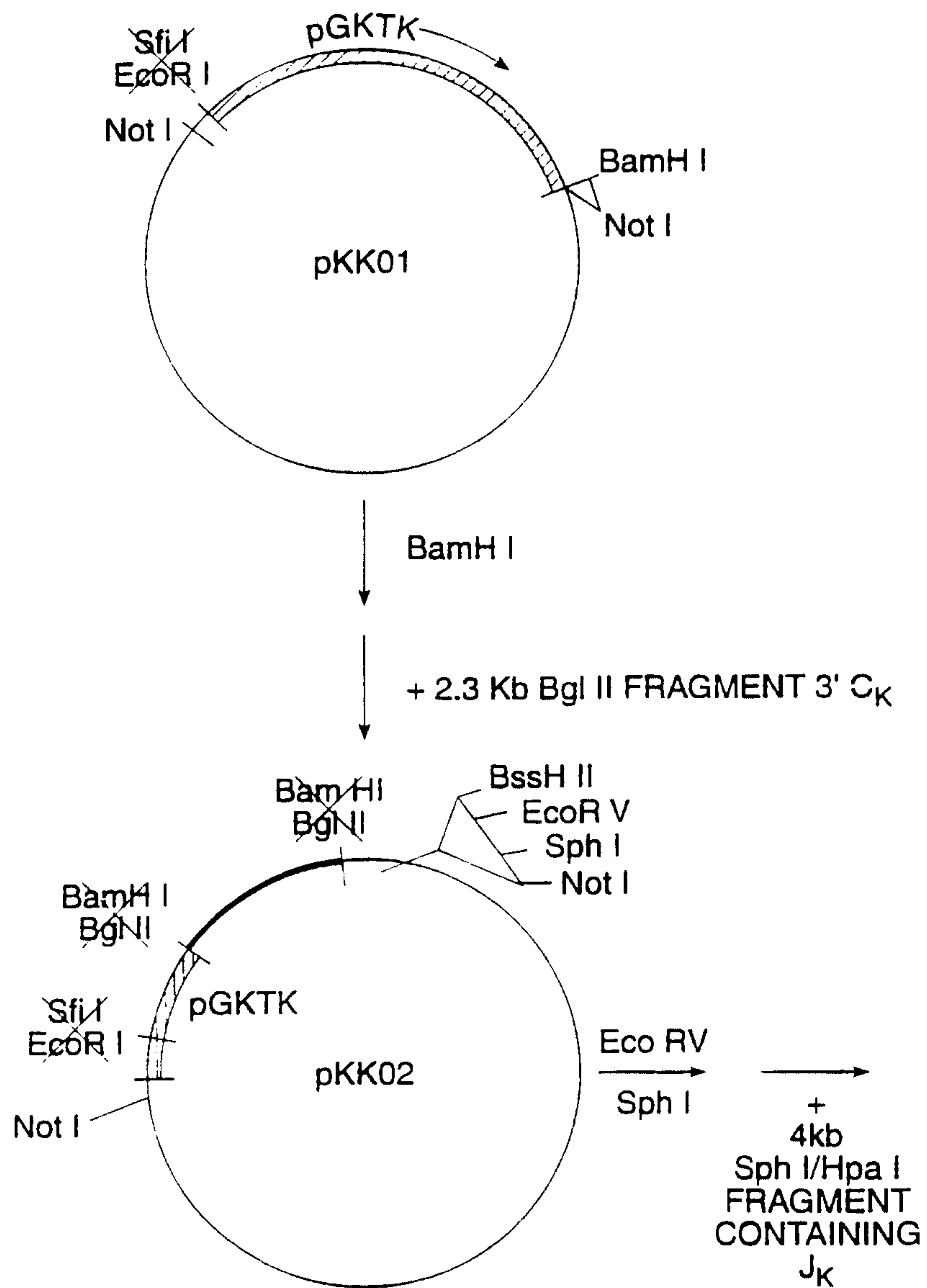


FIG. 19B

20/101

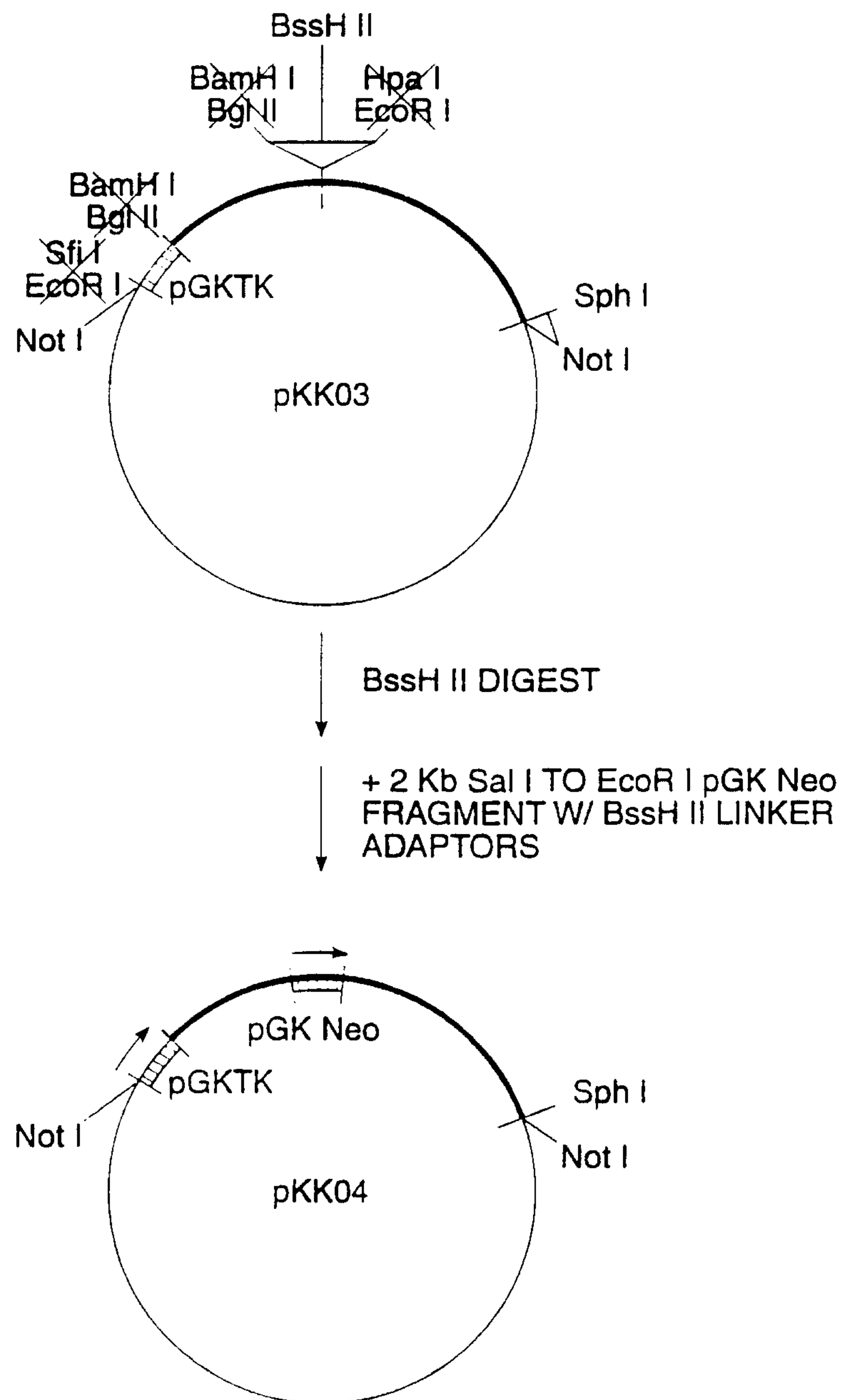


FIG. 19C

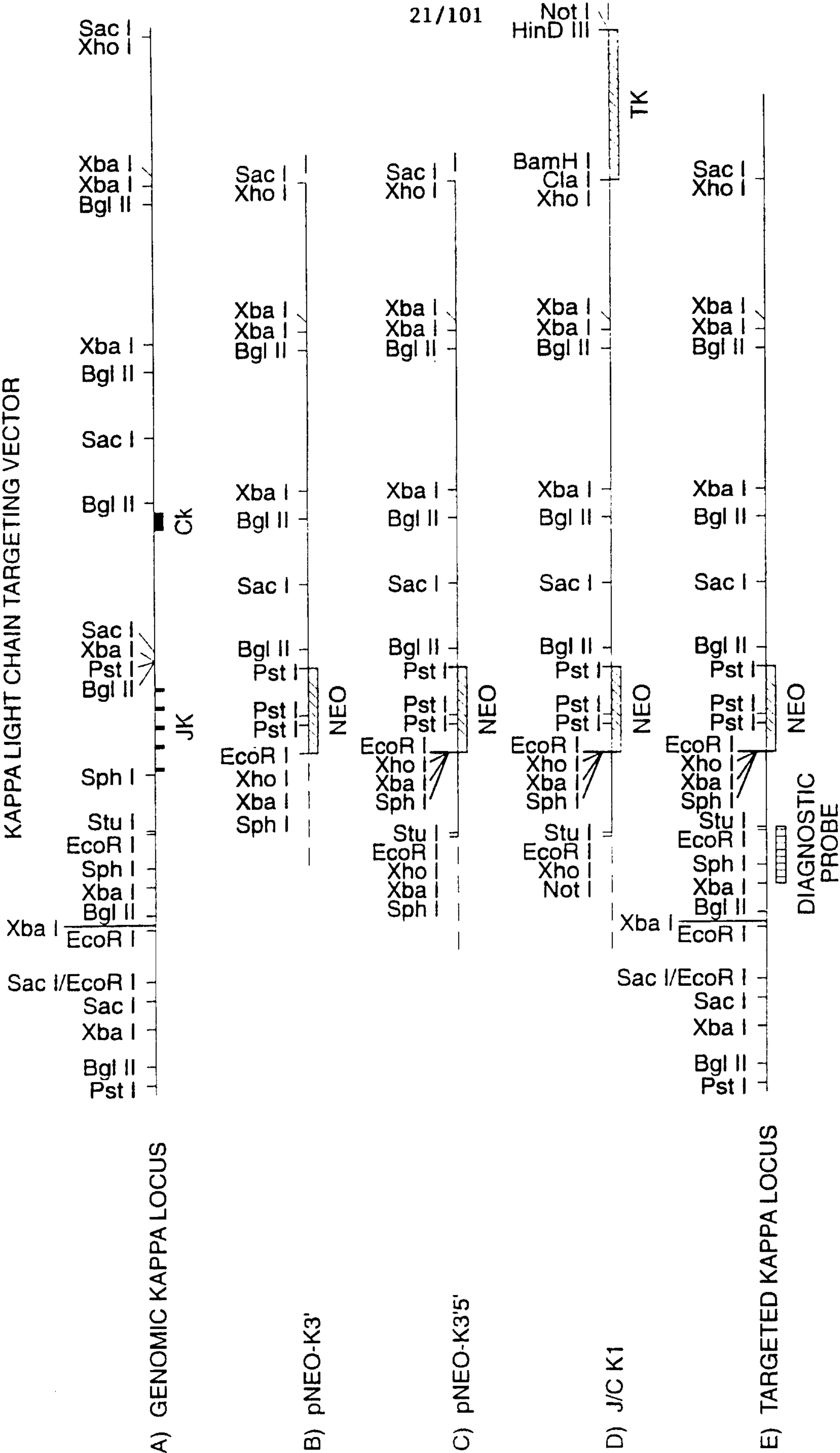
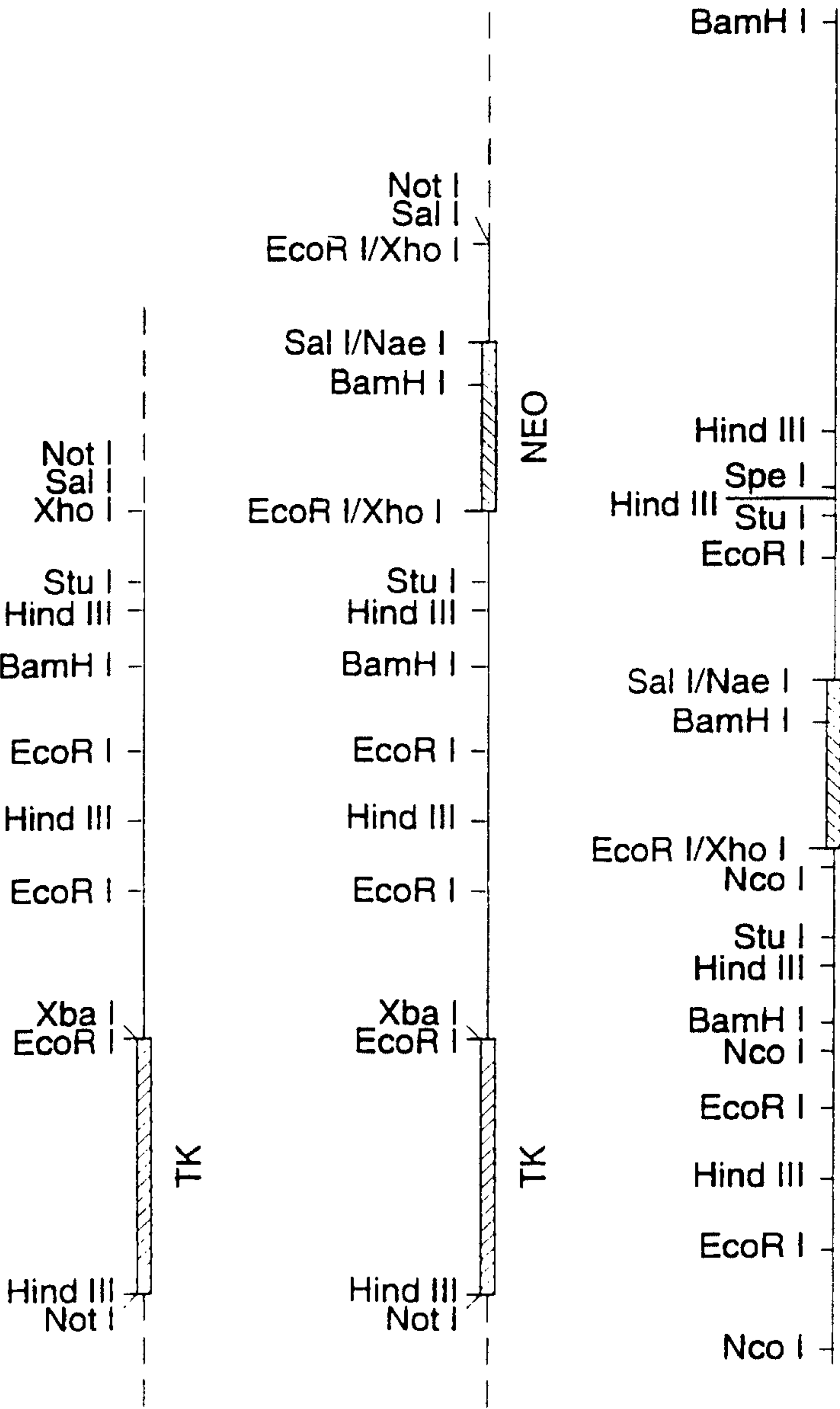


FIG. 20

FIG. 21A-C

23/101

MOUSE HEAVY CHAIN TARGETING VECTOR



DIAGNOSTIC
PROBE

NEO

FIG. 21D-F

D) pGMT-TK-J_H5'

E) J_HK01

F) TARGETED J_H LOCUS

24/101

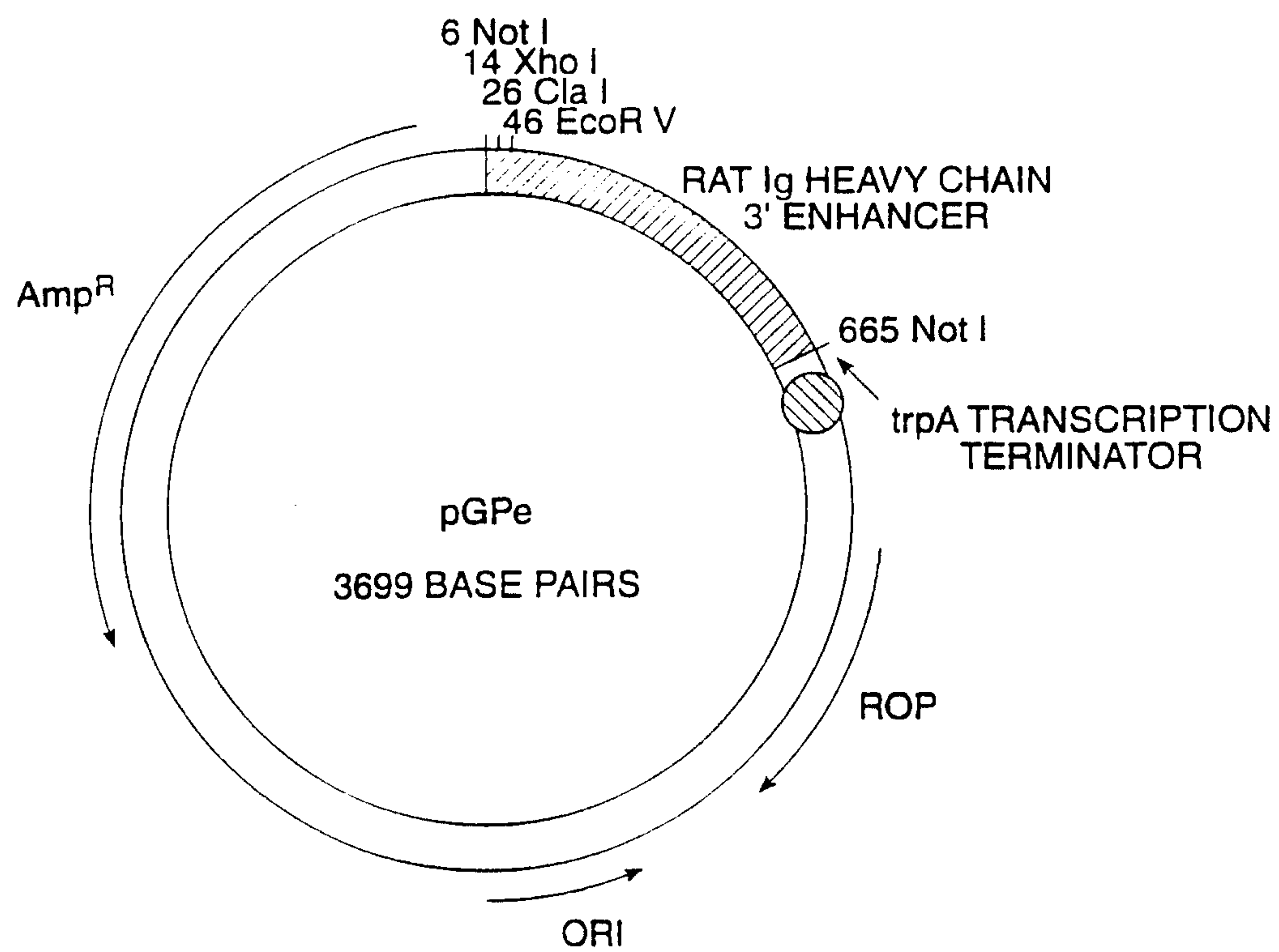
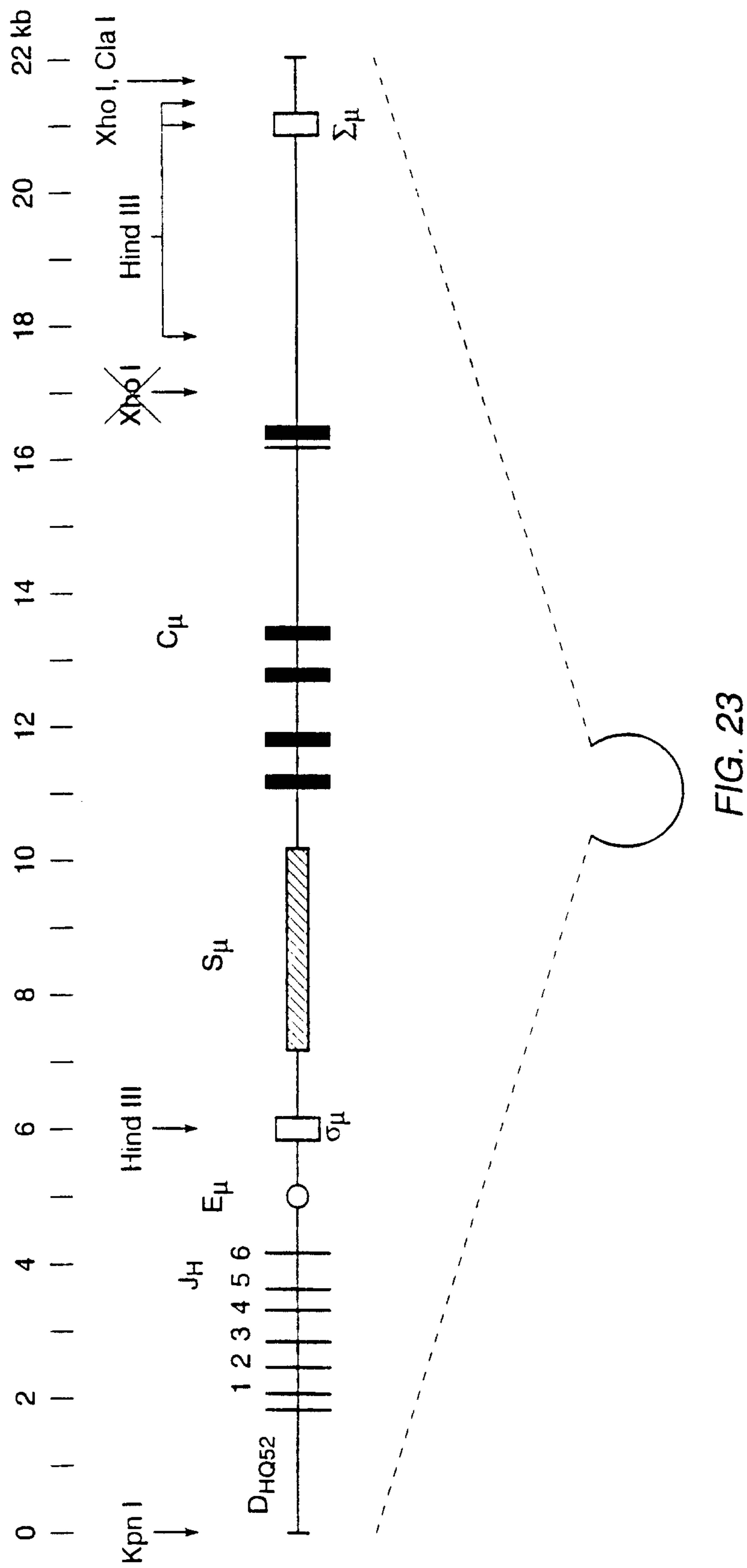


FIG. 22



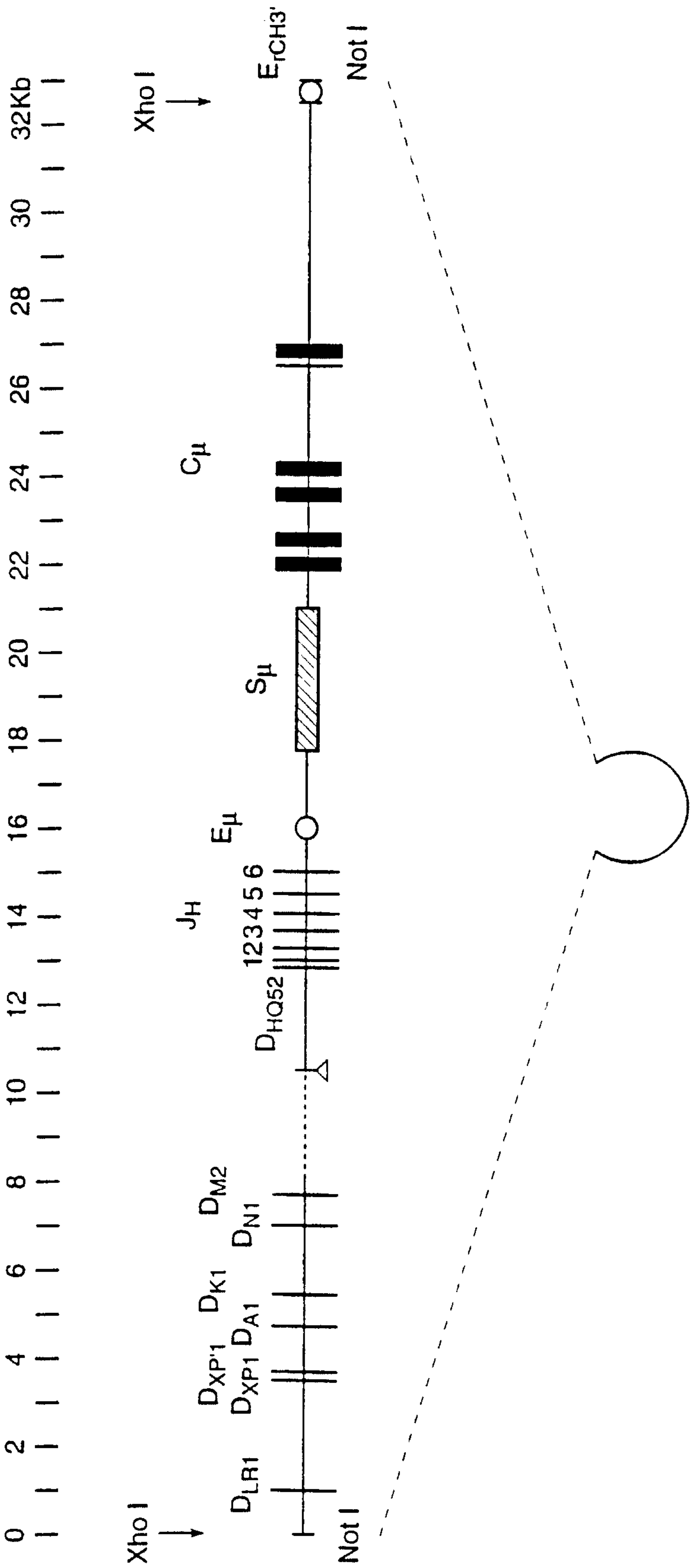


FIG. 24

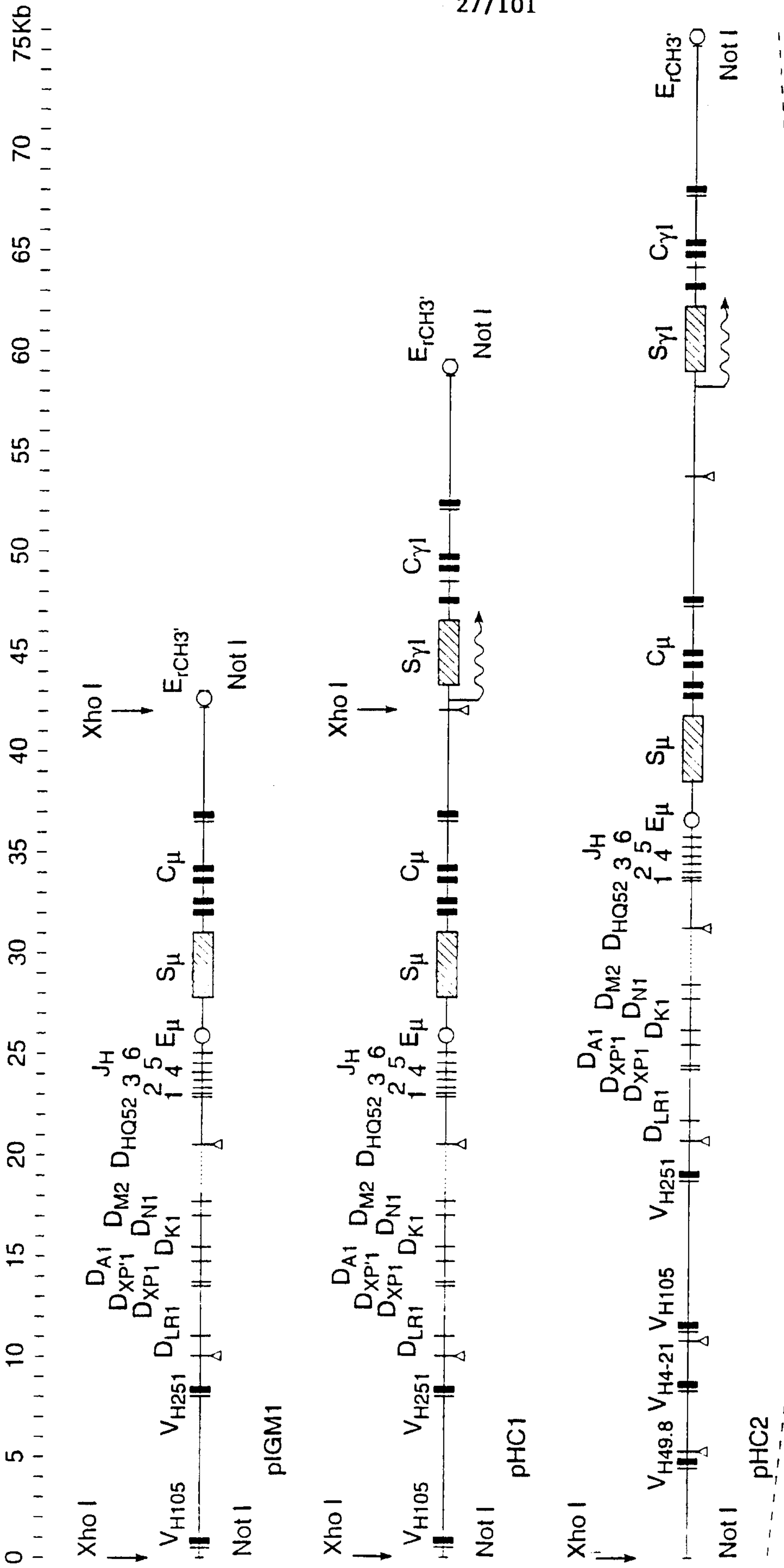


FIG. 25

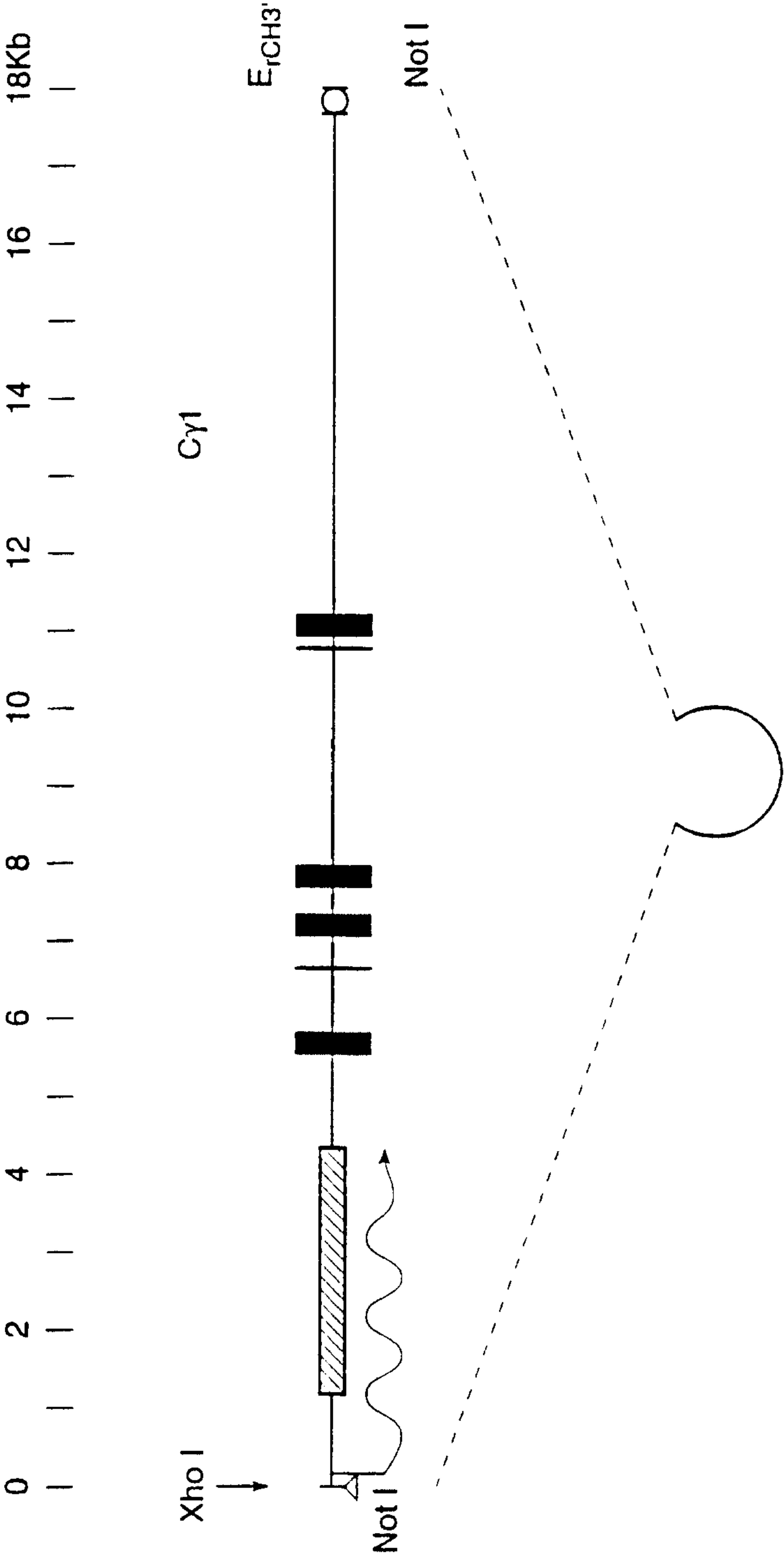
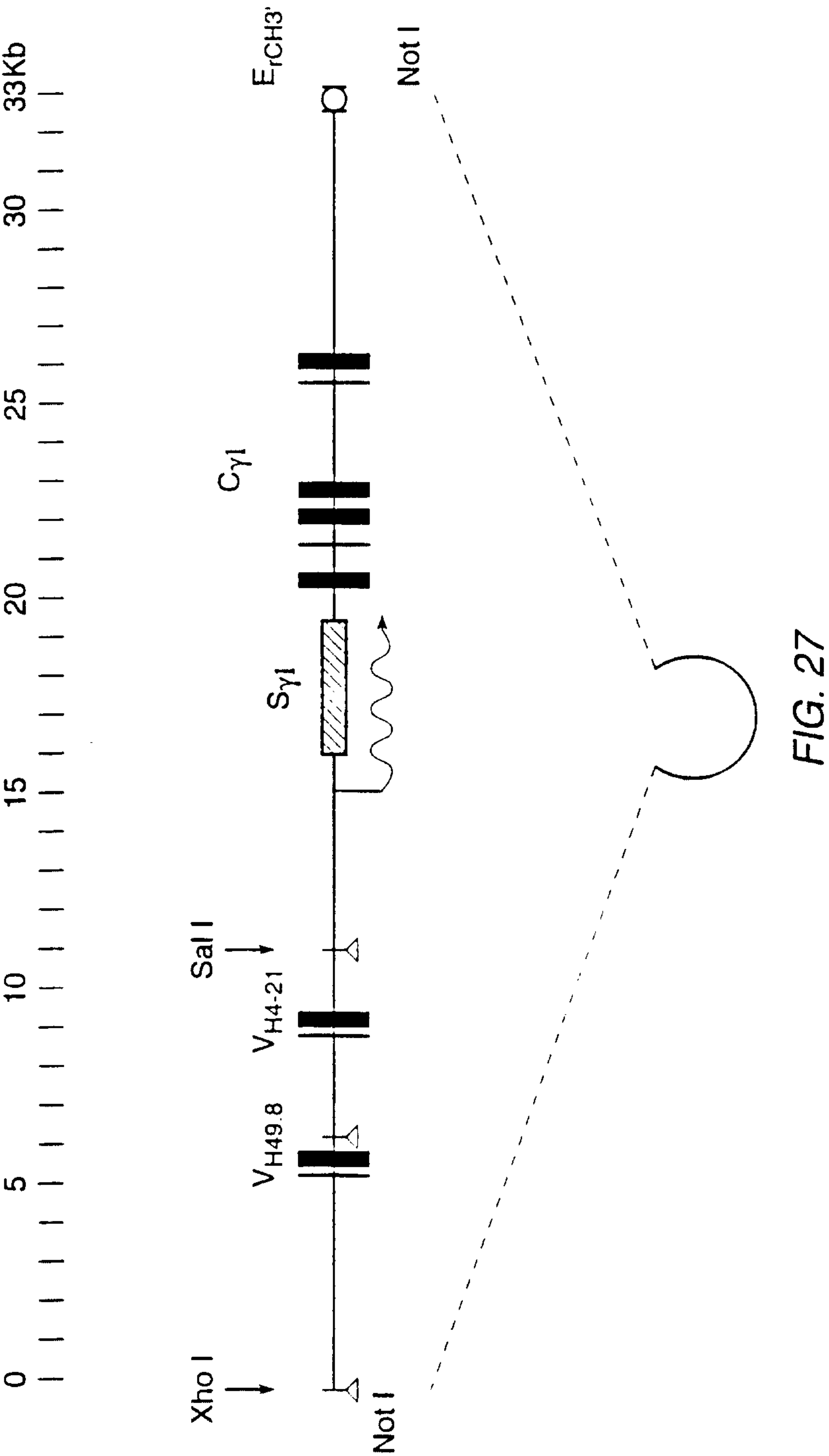


FIG. 26



30/101

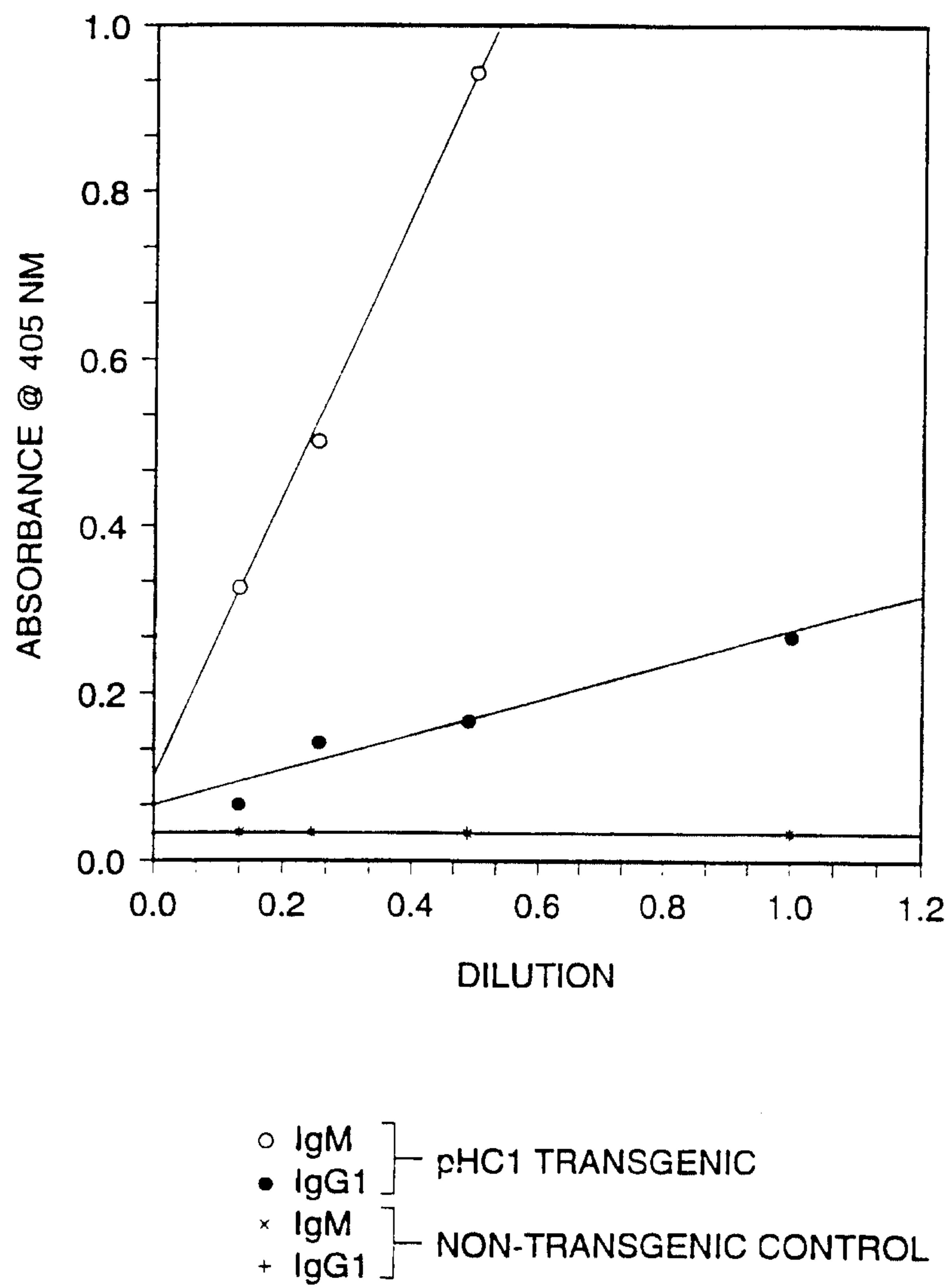


FIG. 28

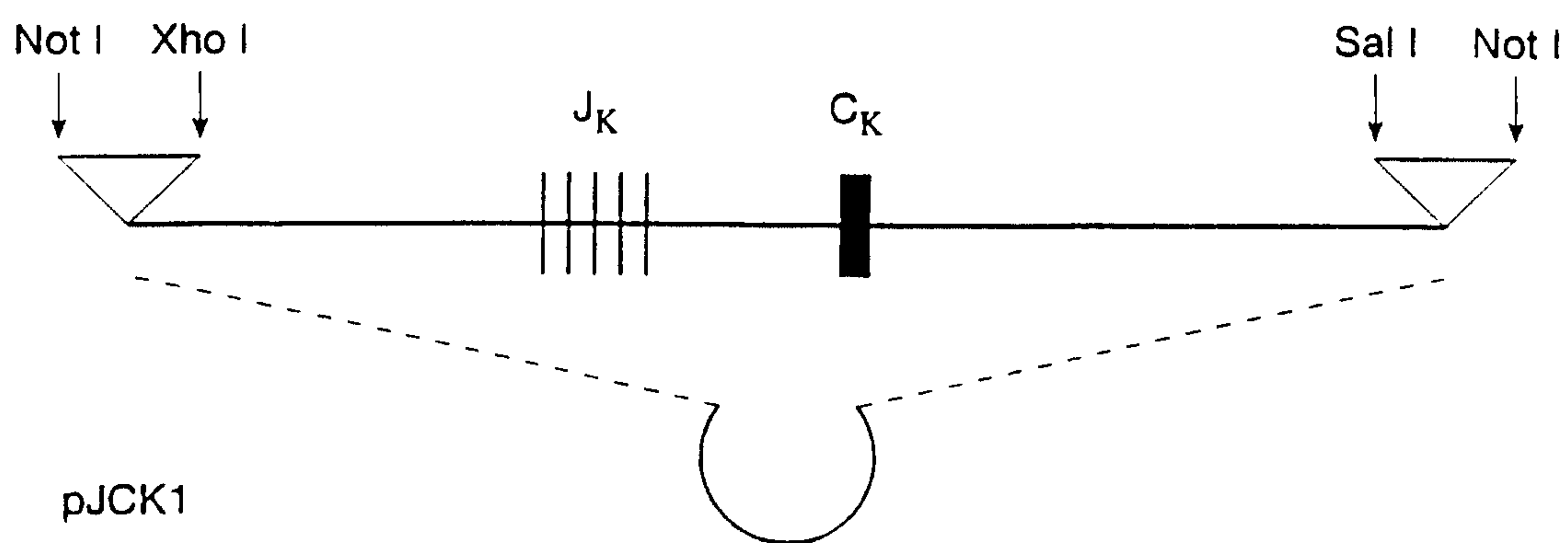
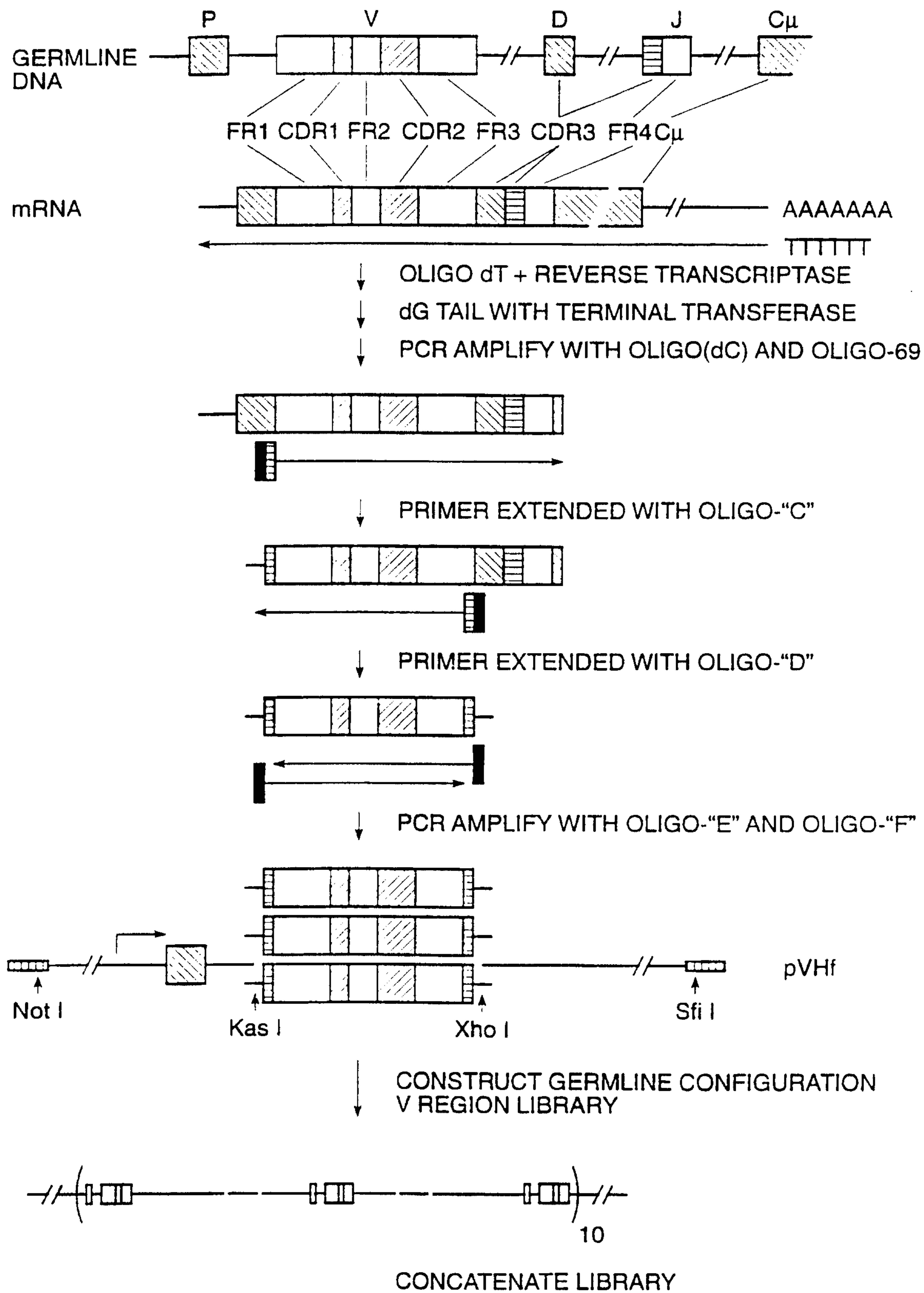


FIG. 29

32/101



SYNTHETIC HEAVY CHAIN VARIABLE REGION

FIG. 30

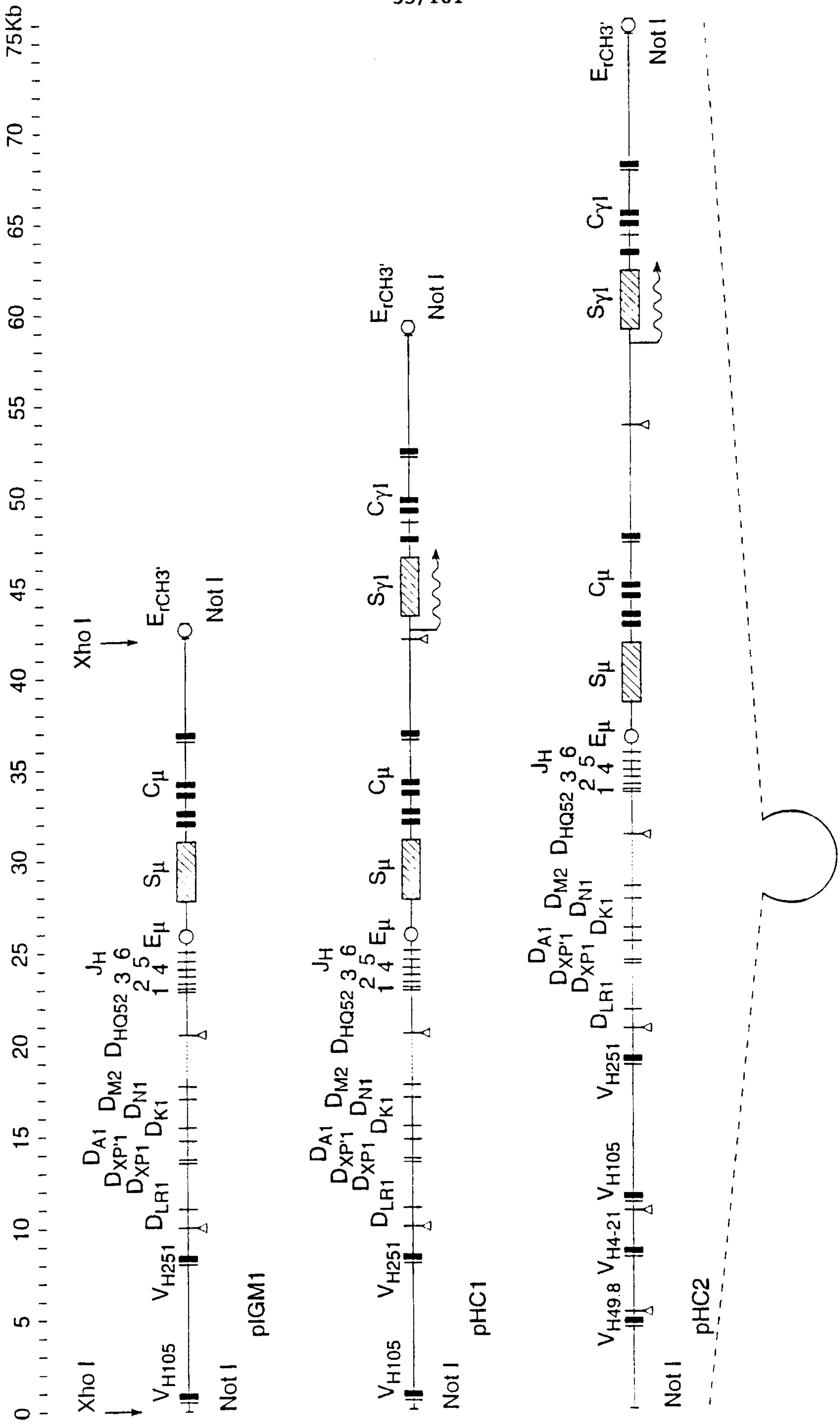


FIG. 31

34/101

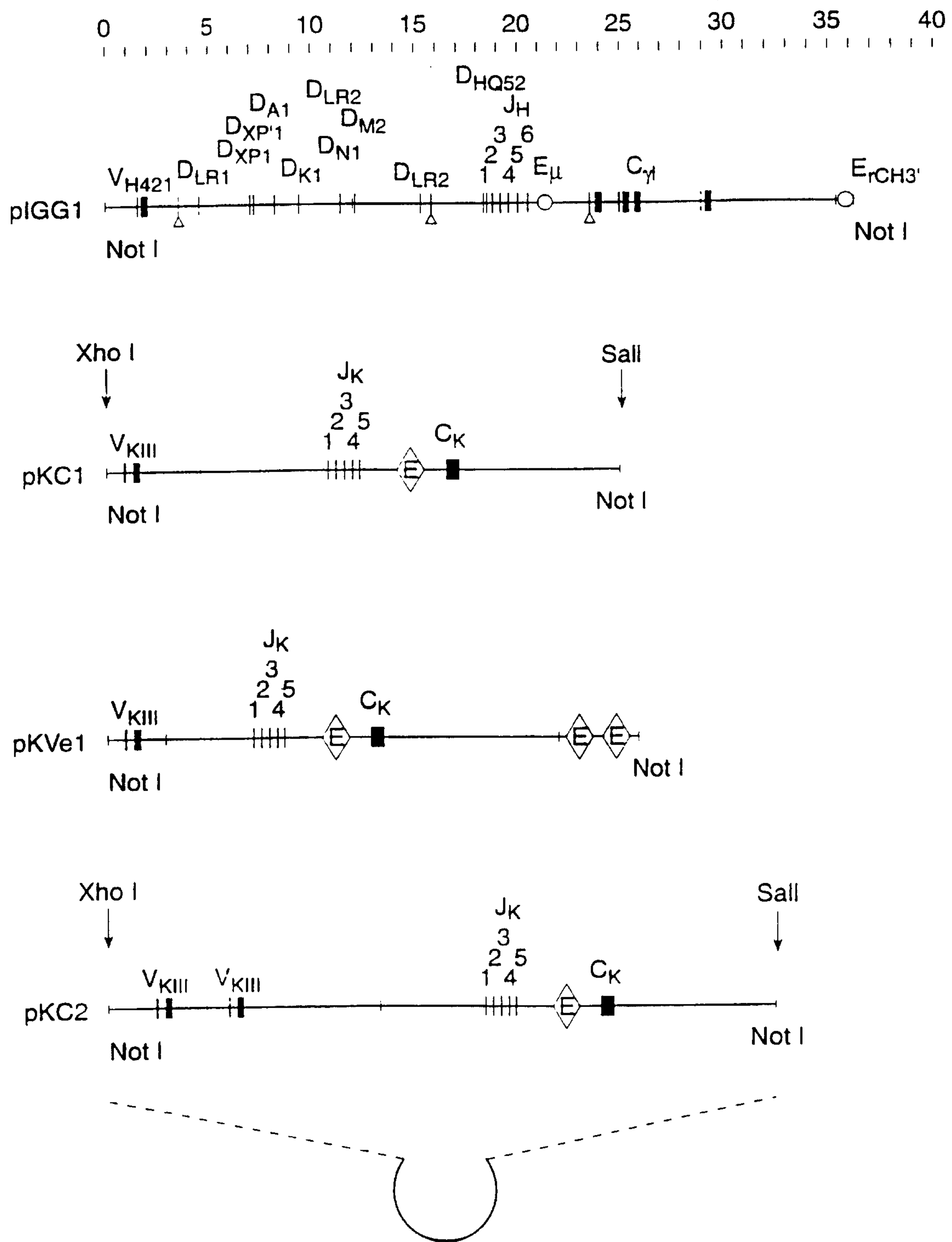


FIG. 32

35/101

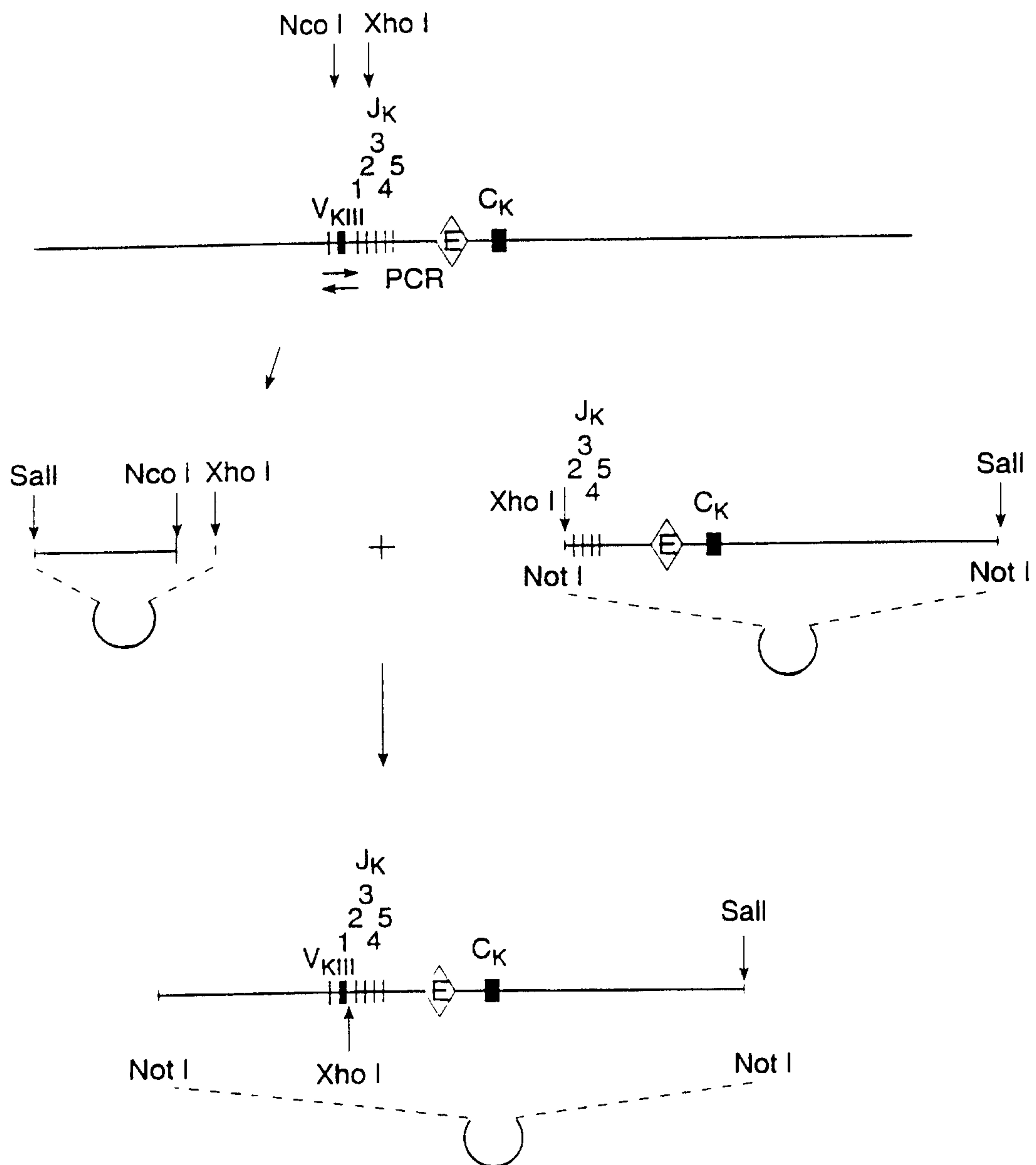


FIG. 33

WO 98/24884

PCT/US97/21803

36/101

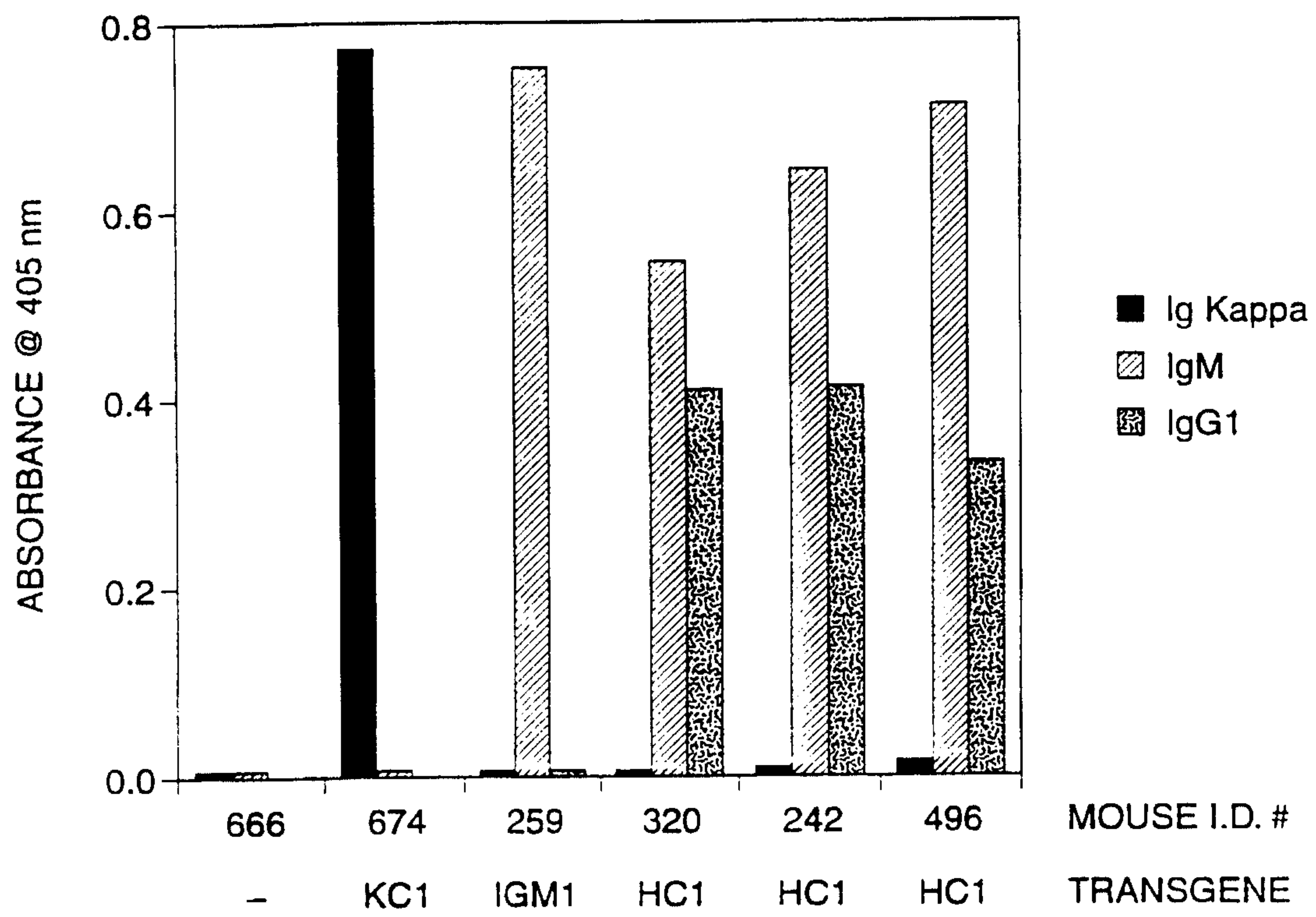


FIG. 34

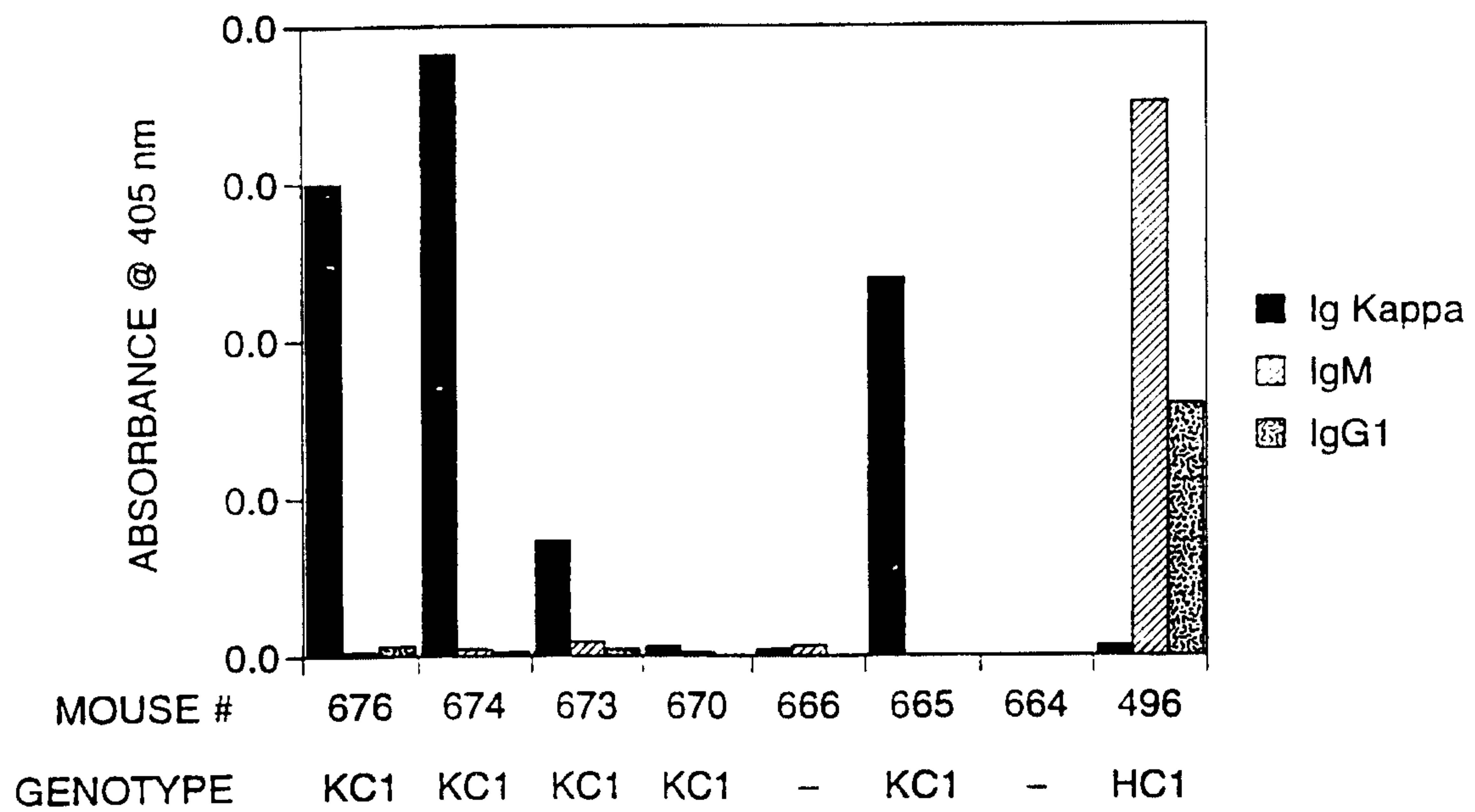
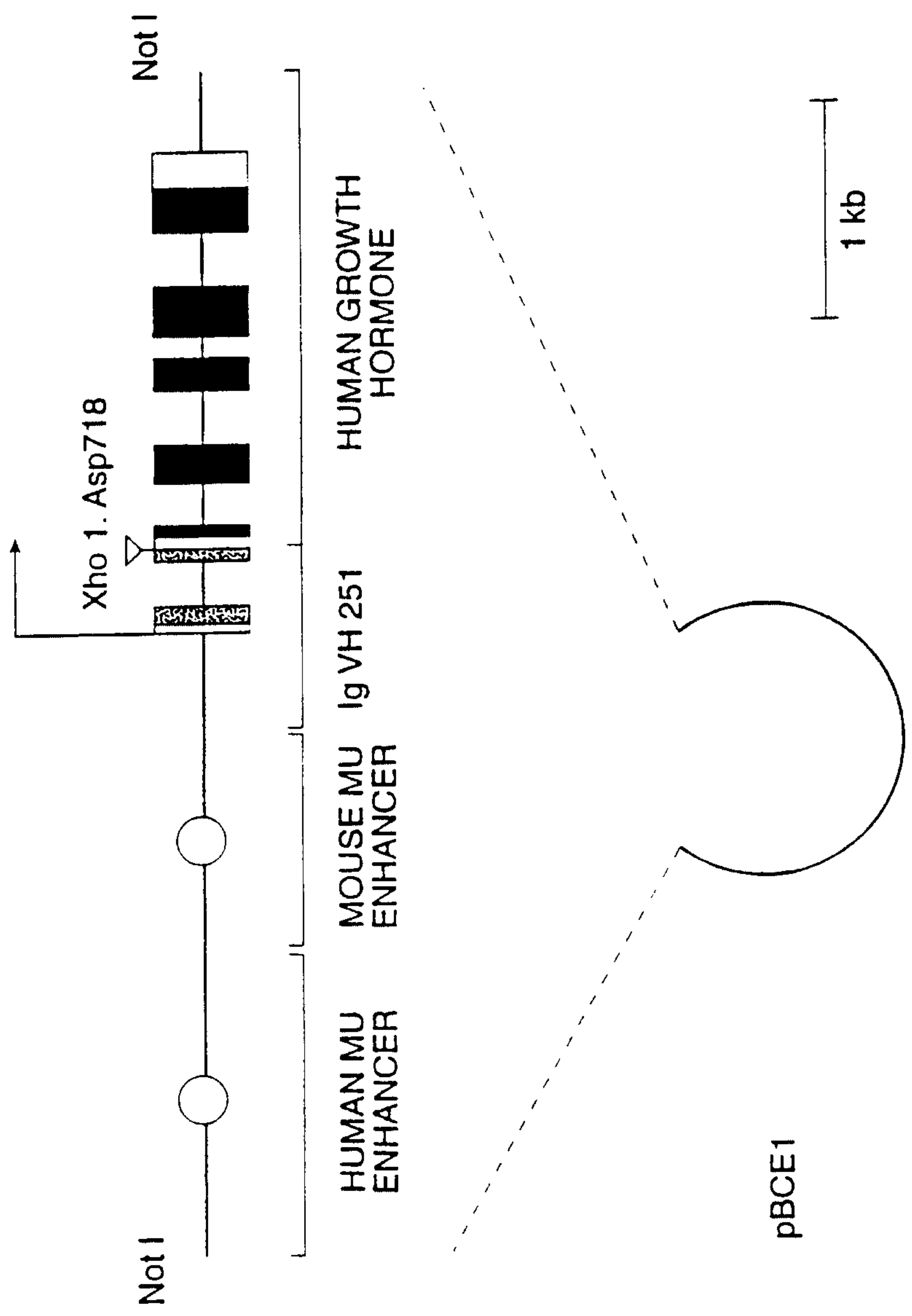


FIG. 35



38/101

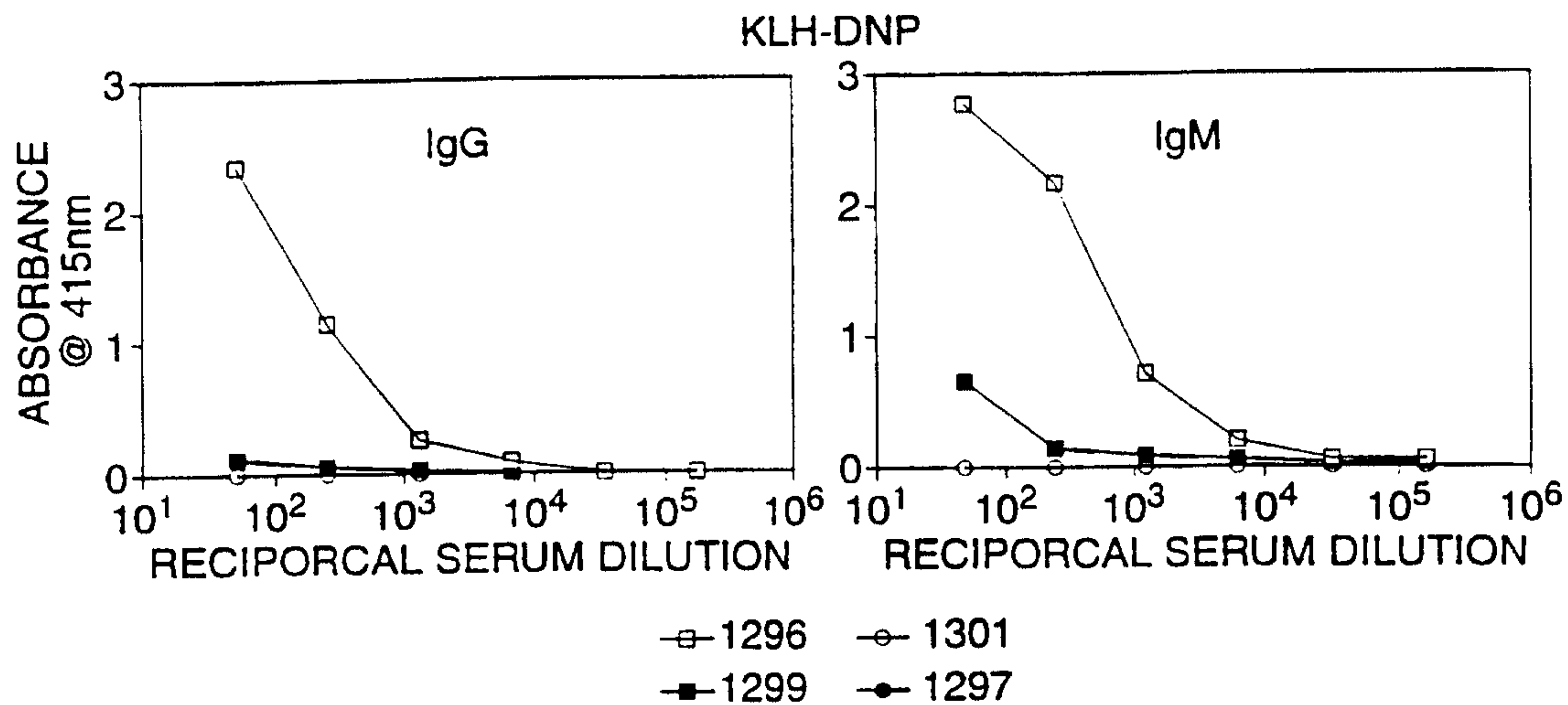


FIG. 37A

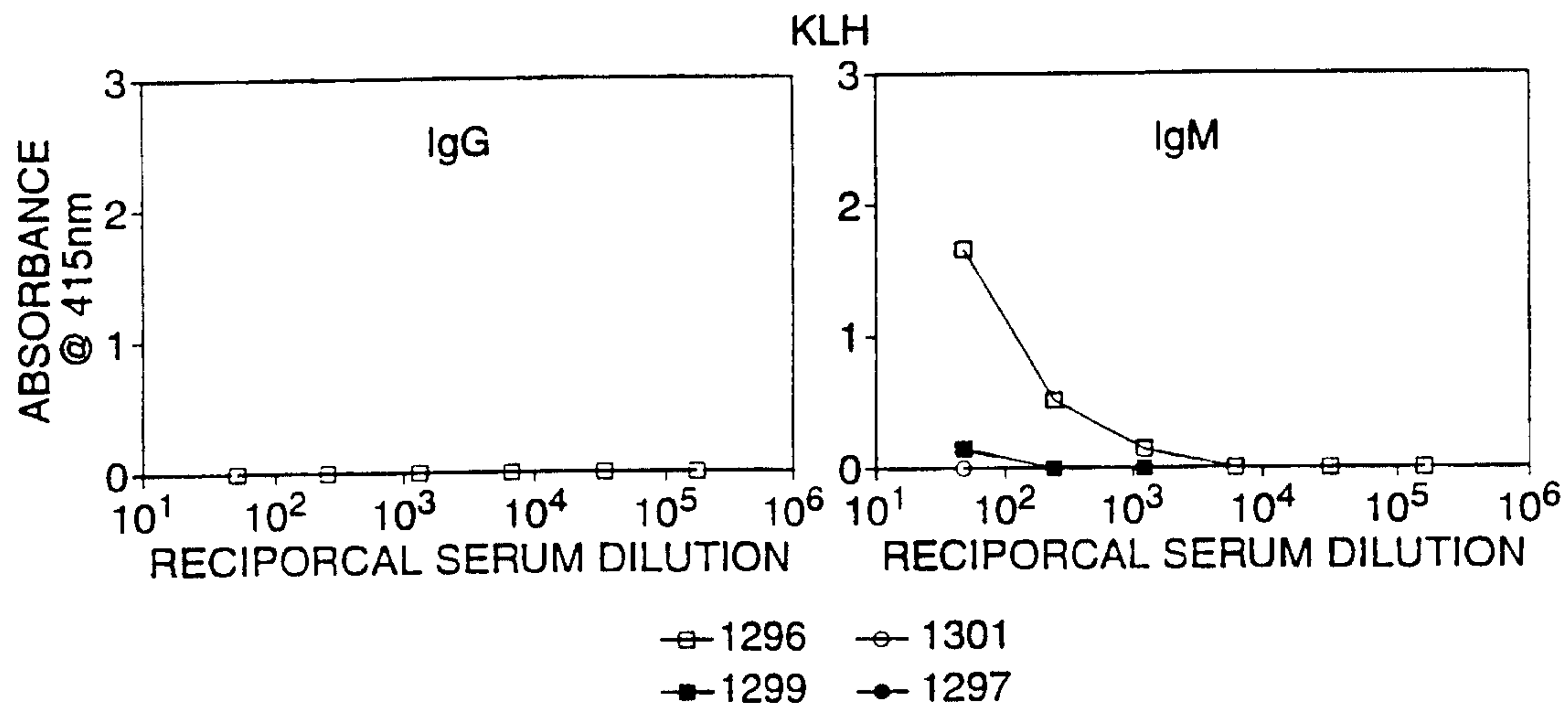


FIG. 37B

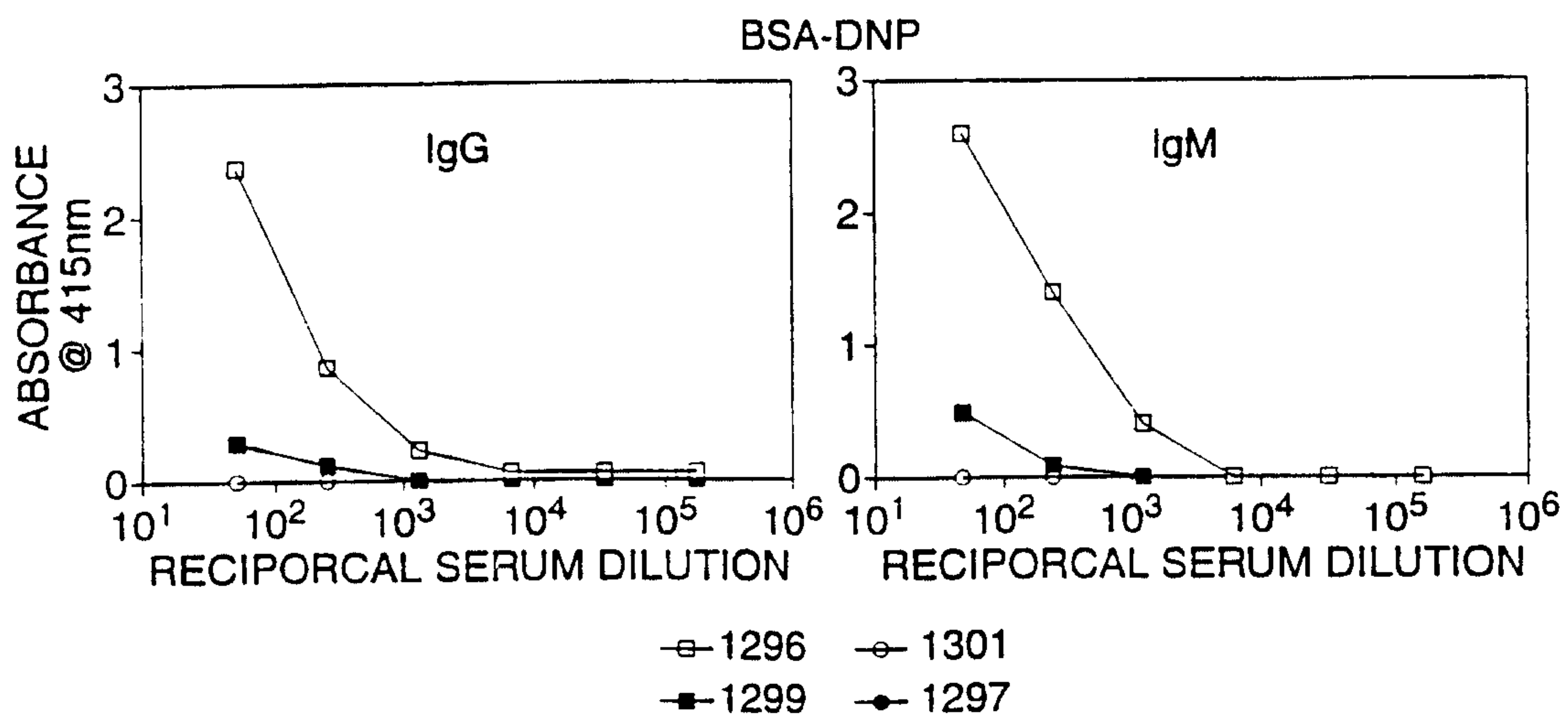


FIG. 37C

39/101

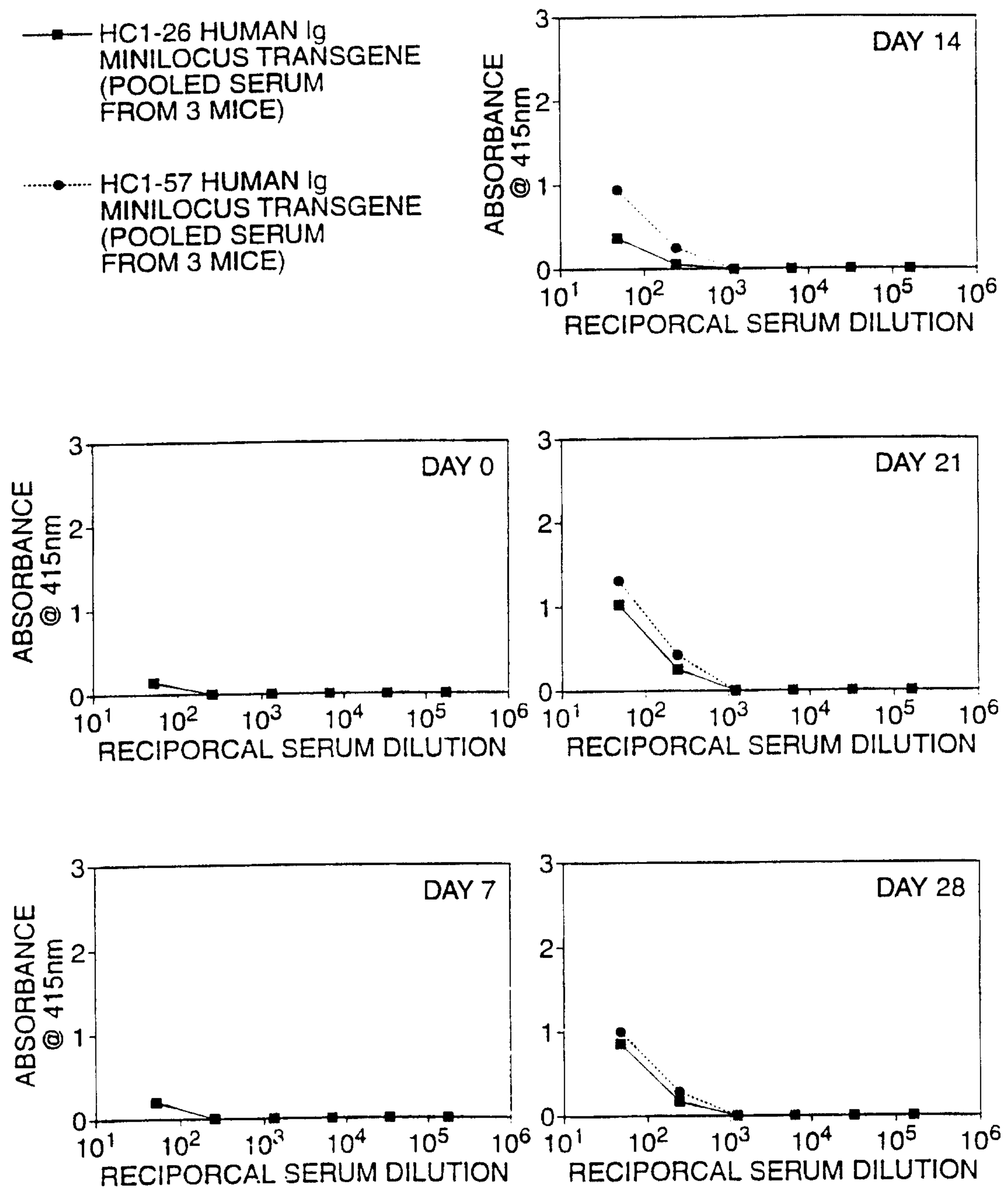


FIG. 38

40/101

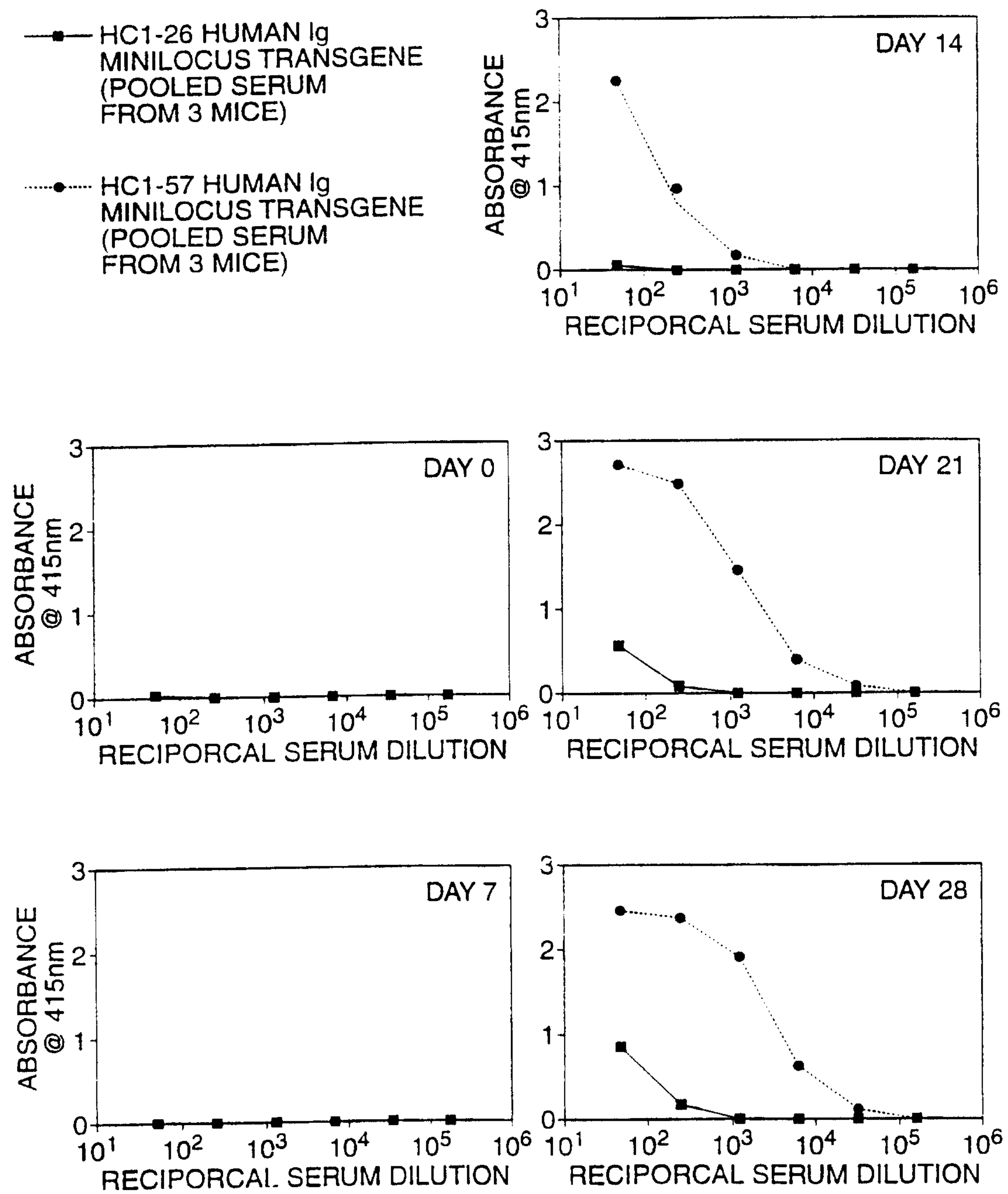


FIG. 39

FIG. 40

42/101

TTTTCTGGCC TGACAACCAG GGTGGCGCAG GATGCTCAGT GCAGAGAGGA 50
 AGAAGCAGGT GGTCTCTGCA GCTGGAAGCT CAGCTCCCAC CCAGCTGCTT 100
 TGCATGTCCC TCCCAGCTGC CCTACCTTCC AGAGCCATA TCAATGCCTG 150
 TGTCAGAGCC CTGGGGAGGA ACTGCTCAGT TAGGACCCAG AGGGAACCAT 200
 GGAAGCCCCA GCTCAGCTTC TCTTCCTCCT GCTACTCTGG CTCCCAGgtg 250
 tGluAlaPro AlaGlnLeuL euPheLeuLe uLeuLeuTrp LeuPro
 agggggaacc atgaggtggt tttgcacatt agtgaaaact cttgccacct 300
 ctgctcagca agaaatataa ttaaaattca aagtatatca acaatttttg 350
 ctctactcaa agacagttgg tttgatcttg attacatgag tgcattttctg 400
 ttttatattcc aatttcagAT ACCACCGGAG AAATTGTGTT GACACAGTCT 450
 Asp ThrThrGlyG luIleValLe uThrGlnSer
 CCAGCCACCC TGTCTTTGTC TCCAGGGGAA AGAGCCACCC TCTCCTGCAG 500
 ProAlaThrL euSerLeuSe rProGlyGlu ArgAlaThrL euSerCysAr
 GGCCAGTCAG AGTGTTAGCA GCTACTTAGC CTGGTACCAA CAGAAACCTG 550
 gAlaSerGln SerValSerS erTyrLeuAl aTrpTyrGln GlnLysProG
 GCCAGGCTCC CAGGCTCCTC ATCTATGATG CATCCAACAG GGCCACTGGC 600
 lyGlnAlaPr oArgLeuLeu IleTyrAspA laSerAsnAr gAlaThrGly
 ATCCCAGCCA GGTTCAAGTGG CAGTGGGTCT GGGACAGACT TCACTCTCAC 650
 IleProAlaA rgPheSerGl ySerGlySer GlyThrAspP heThrLeuTh
 CATCAGCAGC CTAGAGCCTG AAGATTTTGC AGTTTATTAC TGTCAGCAGC 700
 rIleSerSer LeuGluProG luAspPheAl aValTyrTyr CysGlnGlnA
 GTAGCAACTG GCCTCCACACA GTGATTCCAC ATGAAACAAA AACCCCAACA 750
 rgSerAsnTr pPro
 AGACCATCAG TGTTTACTAG ATTATTATAC CAGCTGCTTC CTTTACAGAC 800
 AGCTAGTGGG GT 812

FIG. 41

43/101

AGGGCGGCGC	AGATGCTCAG	TGCAGAGAGA	AGAAACAGGT	GGTCTCTGCA	50
GCTGGAAGCT	CAGCTCCCAC	CCCAGCTGCT	TTGCATGTCC	CTCCCAGCTG	100
CCCTACCTTC	CAGAGCCC	<u>CAT ATCAAT</u> GCCT	GGGTCAGAGC	TCTGGGGAGG	150
AACTGCTCAG	TTAGGACCCA	GACGGAACCA	TGGAAGCCCC	AGCGCAGCTT	200
			M etGluAlaPr oAlaGlnLeu		
CTCTTCCTCC	TGCTACTCTG	GCTCACAGgt	gaggggaata	tgaggtgtct	250
LeuPheLeuL	euLeuLeuTr	pLeuThr			
ttgcacatca	gtgaaaactc	ctgccacctc	tgctcagcaa	gaaatataat	300
taaaattcaa	aatagatcaa	caatttttggc	tctactcaaa	gacagtgggt	350
ttgatttttga	ttacatgagt	gcattttctgt	tttattttcca	atttcagATA	400
			AspT		
CCACCGGAGA	AATTGTGTTG	ACACAGTCTC	CAGCCACCCT	GTCTTTGTCT	450
hrThrGlyGl	uIleValLeu	ThrGlnSerP	roAlaThrLe	uSerLeuSer	
CCAGGGGAAA	GAGCCACCCT	CTCCTGCAGG	GCCAGTCAGG	GTGTTAGCAG	500
ProGlyGluA	rgAlaThrLe	uSerCysArg	AlaSerGlnG	lyValSerSe	
CTACTTAGCC	TGGTACCAGC	AGAAACCTGG	CCAGGCTCCC	AGGCTCCTCA	550
rTyrLeuAla	TrpTyrGlnG	lnLysProGl	yGlnAlaPro	ArgLeuLeuI	
TCTATGATGC	ATCCAACAGG	GCCACTGGCA	TCCCAGCCAG	GTTCAGTGGC	600
leTyrAspAl	aSerAsnArg	AlaThrGlyI	leProAlaAr	gPheSerGly	
AGTGGGCCTG	GGACAGACTT	CACTCTCACC	ATCAGCAGCC	TAGAGCCTGA	650
SerGlyProG	lyThrAspPh	eThrLeuThr	IleSerSerL	euGluProGl	
AGATTTTGCA	GTTTATTACT	GTCAGCAGCG	TAGCAACTGG	CATCC <u>CACAG</u>	700
uAspPheAla	ValTyrTyrC	ysGlnGlnAr	gSerAsnTrp	His	
<u>TGATTCCACA</u>	<u>TGAAACAAAA</u>	<u>ACC</u> CCAACAA	GACCATCAGT	GTTTACTAGA	750
TTATTATACC	AGCTGCTTCC	TTTACAGACA	GCTAGTGGGG	TGGCCACTCA	800
GTGTTAGCAT	CTCAGCTCTA	TTTGGCCATT	TTGGAGTTCA	AGTTGTCAAG	850
TCCAAAATTA	CTTATGTTAG	TCCATTGCAT	CATACCATTT	CAGTGTGGCT	900

FIG. 42

44/101

CCGCCCCAGC TGCTTTGCAT GTCCCTCCCA GCCGCCCTGC AGTCCAGAGC 50
 CCATATCAAT GCCTGGGTCA GAGCTCTGGA GAAGAGCTGC TCAGTTAGGA 100
 ACCCCAGAGG GAACCATGGA AACCCCAGCG CAGCTTCTCT TCCTCCTGCT 150
 MetGl uThrProAla GlnLeuLeuP heLeuLeuLe
 ACTCTGGCTC CCAGgtgagg ggaacatggg atggttttgc atgtcagtga 200
 uLeuTrpLeu Pro
 aaaccctctc aagtcctggt acctggcaac tctgctcagt caatacaata 250
 attaaagctc aatataaagc aataattctg gctcttctgg gaagacaatg 300
 ggttttgattt agattacatg ggtgactttt ctgtttttatt tccaatctca 350
 gATACCACCG GAGAAATTGT GTTGACGCAG TCTCCAGGCA CCCTGTCTTT 400
 AspThrThrG lyGluIleVa lLeuThrGln SerProGlyT hrLeuSerLe
 GTCTCCAGGG GAAAGAGCCA CCCTCTCCTG CAGGGCCAGT CAGAGTGTTA 450
 uSerProGly GluArgAlaT hrLeuSerCy sArgAlaSer GlnSerValS
 GCAGCAGCTA CTTAGCCTGG TACCAGCAGA AACCTGGCCA GGCTCCCAGG 500
 erSerSerTy rLeuAlaTrp TyrGlnGlnL ysProGlyGl nAlaProArg
 CTCCTCATCT ATGGTGCATC CAGCAGGGCC ACTGGCATCC CAGACAGGTT 550
 LeuLeuIleT yrGlyAlaSe rSerArgAla ThrGlyIleP roAspArgPh
 CAGTGGCAGT GGGTCTGGGA CAGACTTCAC TCTCACCATC AGCAGACTGG 600
 eSerGlySer GlySerGlyT hrAspPheTh rLeuThrIle SerArgLeuG
 AGCCTGAAGA TTTTGCAGTG TATTACTGTC AGCAGTATGG TAGCTCACCT 650
 luProGluAs pPheAlaVal TyrTyrCysG lnGlnTyrGl ySerSerPro
 CCACAGTGA TTCAGCTTGA AACAAAAACC TCTGCAAGAC CTTCAATTGTT 700
 TACTAGATTA TACCAGCTGC TTCCTTTACA GATAGCTGCT GCAATGACAA 750
 CTCAATTTAG CATCTCTCTC TGCTTGGGCA TTTTGGGGAT CTTAAAAAAG 800
 TAATCCCTTG ATATATTTTT GACTCTGATT CCTGCATTTT TCCTCAGACC 850
 AAGATGGACA GCCAGGTTTA AGCACAGTTT CACAGTAATG GCCACTGGAT 900

FIG. 43

45/101

AAACACATTC	TCTGCAGACA	AATTTGAGCT	ACCTTGATCT	TACCTGGACA	50
GGTGGGGACA	CTGAGCTGGT	GCTGAGTTAC	TCAGATGCGC	CAGCTCTGCA	100
GCTGTGCCCCA	GCCTGCCCCA	TCCCCTGCTC	ATTTGCATGT	TCCCAGAGCA	150
CAACCTCCTG	CCCTGAAGCC	<u>TTATTAAT</u> AG	GCTGGTCAGA	CTTTGTGCAG	200
GAATCAGACC	CAGTCAGGAC	ACAGCATGGA	CATGAGGGTC	CTCGCTCAGC	250
		MetAs	pMetArgVal	LeuAlaGlnL	
TCCTGGGGCT	CCTGCTGCTC	TGTTTCCCAG	gtaaggatgg	agaacactag	300
euLeuGlyLe	uLeuLeuLeu	CysPhePro			
cagtttactc	agcccagggg	gctcagtact	gcttttactat	tcaggggaaat	350
tctcttataa	catgattaat	tgtgtggaca	tttggttttta	tgttttccaat	400
ctcagGTGCC	AGATGTGACA	TCCAGATGAC	CCAGTCTCCA	TCCTCACTGT	450
GlyAla	ArgCysAspI	leGlnMetTh	rGlnSerPro	SerSerLeuS	
CTGCATCTGT	AGGAGACAGA	GTCACCATCA	CTTGTCGGGC	GAGTCAGGGT	500
erAlaSerVa	lGlyAspArg	ValThrIleT	hrCysArgAl	aSerGlnGly	
ATTAGCAGCT	GGTTAGCCTG	GTATCAGCAG	AAACCAGAGA	AAGCCCCTAA	550
IleSerSerT	rpLeuAlaTr	pTyrGlnGln	LysProGluL	ysAlaProLy	
GTCCCTGATC	TATGCTGCAT	CCAGTTTGCA	AAGTGGGGTC	CCATCAAGGT	600
sSerLeuIle	TyrAlaAlaS	erSerLeuGl	nSerGlyVal	ProSerArgP	
TCAGCGGCAG	TGGATCTGGG	ACAGATTTCA	CTCTCACCAT	CAGCAGCCTG	650
heSerGlySe	rGlySerGly	ThrAspPheT	hrLeuThrIL	eSerSerLeu	
CAGCCTGAAG	ATTTTGCAAC	TTATTACTGC	CAACAGTATA	ATAGTTACCC	700
GlnProGluA	spPheAlaTh	rTyrTyrCys	GlnGlnTyrA	snSerTyrPr	
ACC <u>CACAGTG</u>	TTACACACCC	AA <u>ACATAAAC</u>	CCCCAGGGAA	GCAGATGTGT	750
O					
GAGGCTGGGC	TGCCCCAGCT	GCTTCTCCTG	ATGCCTCCAT	CAGCTGAGAG	800
TGTTCCCTCAG	ATGCAGCCAC	ACTCTGATGG	TGTTGGTAGA	TGGGGAC	847

FIG. 44

46/101

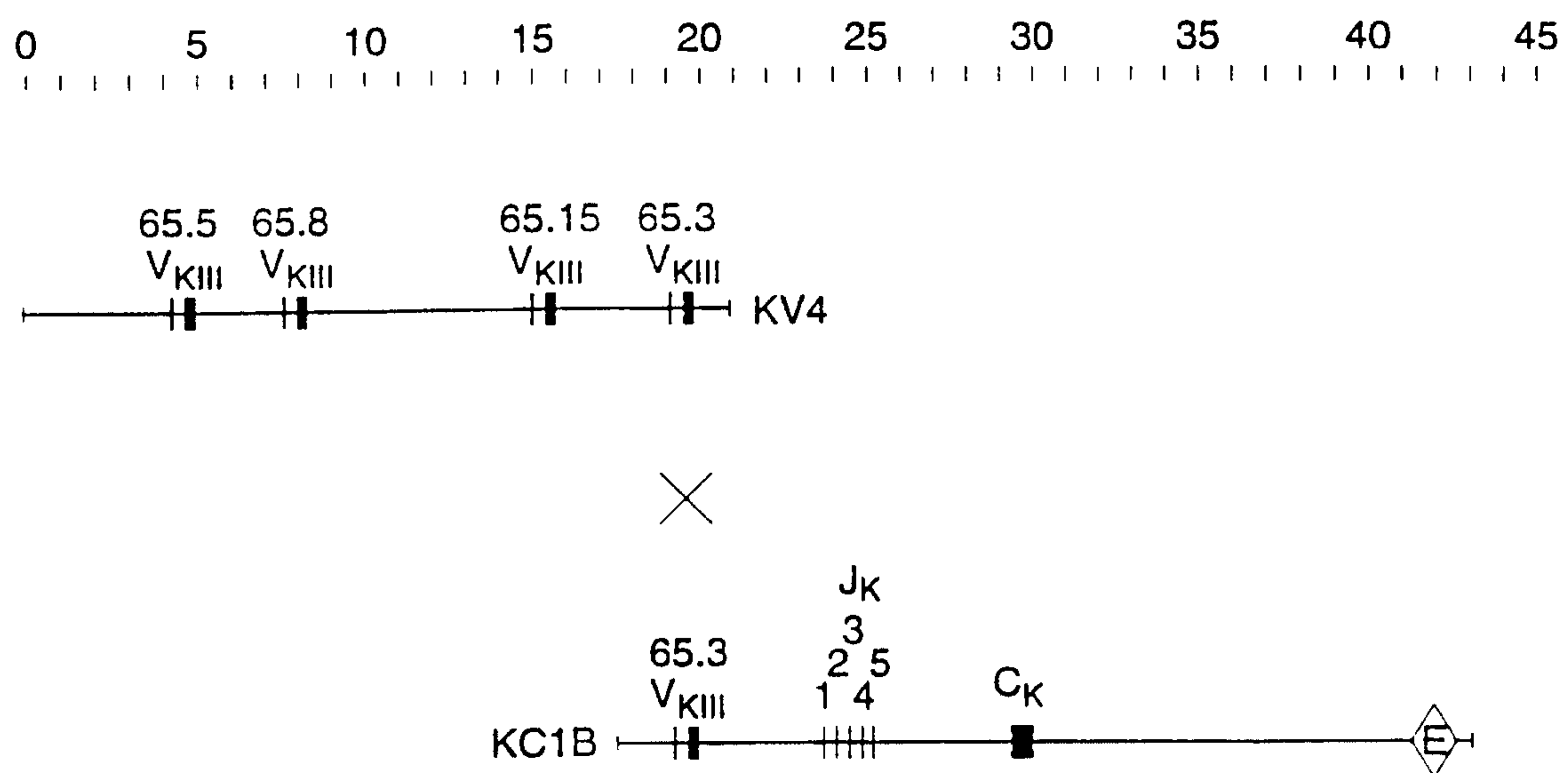


FIG. 45

47/101

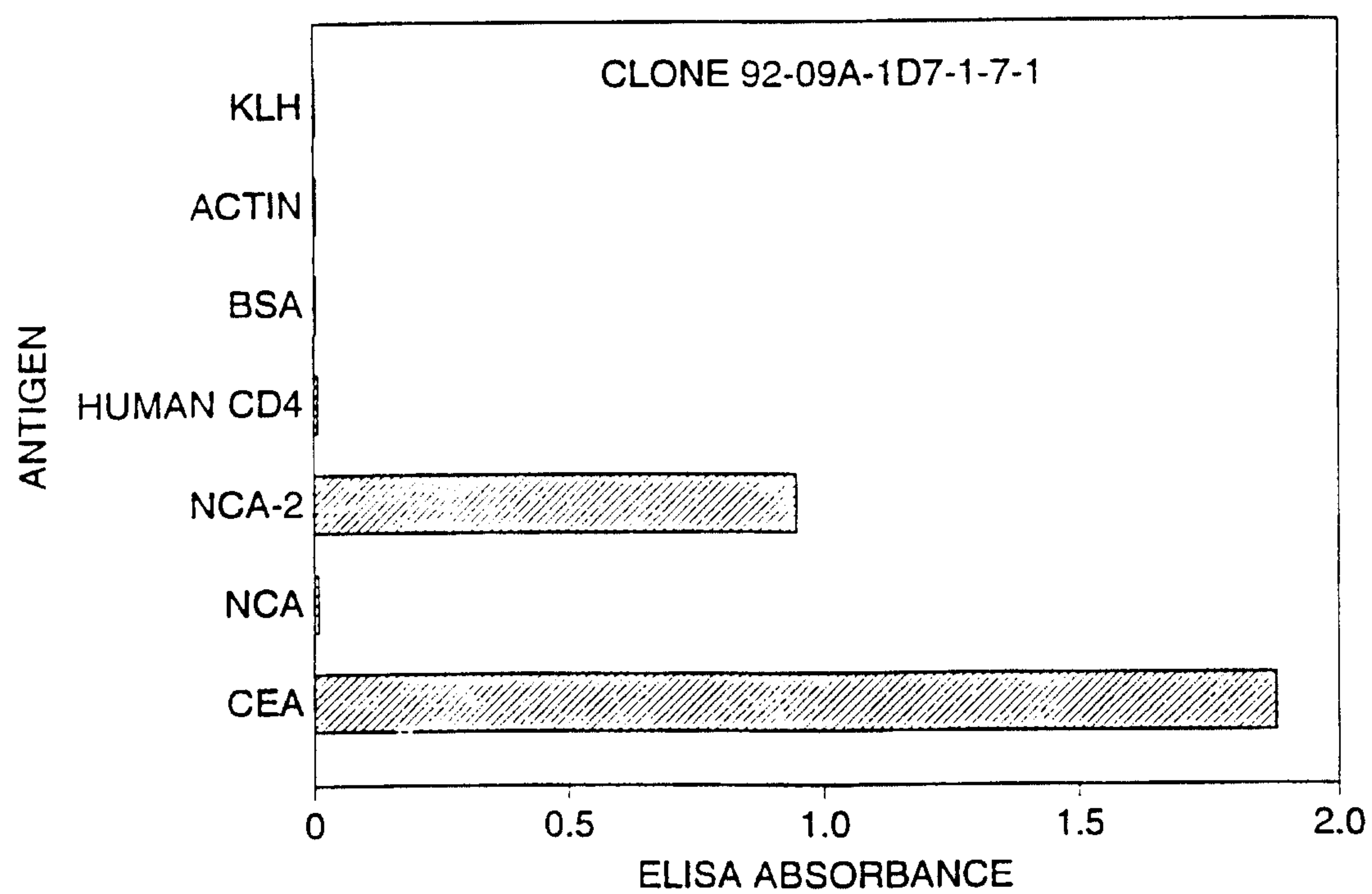
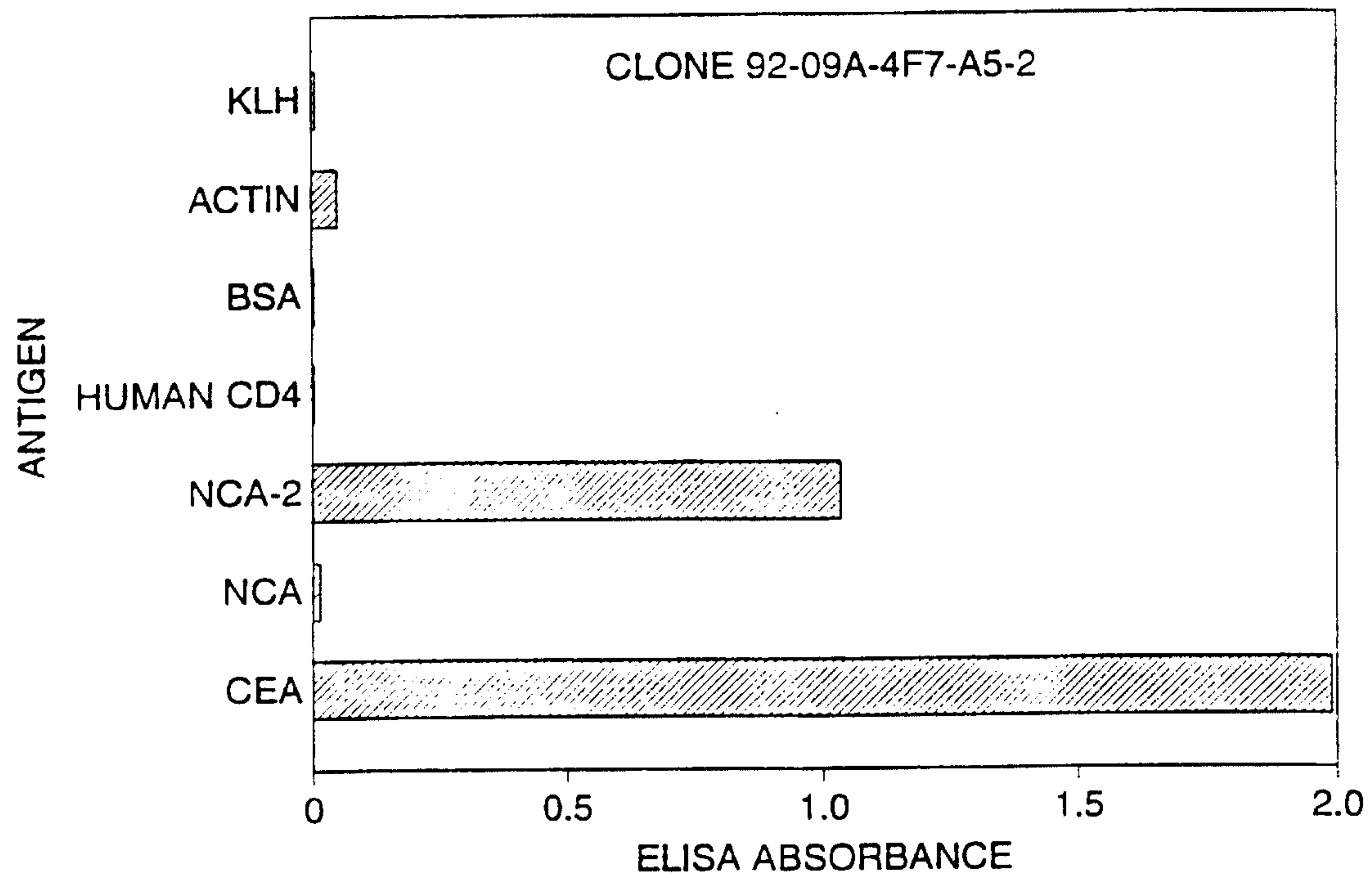


FIG. 46

VH251			HUMAN		MOUSE	
			n D n	J	Cy	
5	DXP'1 J6 G1	GCCTCGGACACCGCATGTATTACTGTGtGAGA	cattTATGGTTCGGGAGTTAcg	GGTgTgaACGTCTGGGGCCAAAGGACCAAGGTACCGTCTCTCTCAG	CCAAACGACACCCCCATCTGTCTATCCACT	
7	DH052 J3 G1	GCCTCGGACACCGCATGTATTACTGTGCGAGA	cACTGGGcatttgat	GCTcTTGATgTCTGGGGCCAAAGGACAAATGcTACCGTCTCTTCAG	CCAAACGACACCCCCATCTGTCTATCCACT	
2	DH052 J3 G2b	GCCTCGGACACCGCATGTATTACTGTGCGAGA	ACTGGGAtgat	GCTTTTGATATCTGGGGCCAAAGGACAAATGGTACCGTCTCTTCAG	CCAAACAACACCCCCATCAGTCTATCCACT	
3	D? J3 G2b	GaCTCGGACACCGCATGTATTACTGTGCGAGA	cagggagagat	GCTTTAGATATCTGGGGCCAAAGGACAAATGGTACCGTCTCTTCAG	CCAAACAACACCCCCATCAGTCTATCCACT	
4	DXP'1 J4 G2b	GCCTCGGACACCGCATGTATTATGTGtGAGA	catagggACtAtatTCGGGGAGTTAttcc	TGACTACTGGGGCCAGGGAACCCIGGTACCGTCTCTCTCAG	CCAAACAACACCCCCATCAGTCTATCCACT	
10	DH052 J3 G2b	GCCTCGGACACCGCATGTATTACTGTGCGAGA	ACTGGGAtgat	GCTTTTGATATCTGGGGCCAAAGGACAAATGGTACCGTCTCTTCAG	CCAAACAACACCCCCATCAGTCTATCCACT	
1	D? J3 G3	GCCTCGGACACCGCATGTATTACTGTGCGAGA	catgggtctatg	GATACTGGGGCCAAAGGACAAATGGTACCGTCTCTTCAG	CTACAACAACAGCCCCATCTGTCTATCCCTT	
6	DH052 J4 G3	GCCTCGGACACCGCATGTATTACTGTGCGAGA	gagagcagtcACTGGGAtcg	TTTGACTATCTGGGGCCAGGGAACCTIGGTACCGTCTCTCTCAG	CTACAACAACAGCCCCATCTGTCTATCCCTT	
8	DIR2 J3 G3	GCCTCGGACACCGCATGTATTACTGTGCGAGA	agggACCCCCctgat	GCTTTTGATATCTGGGGCCAAAGGACAAATGGTACCGTCTCTTCAG	CTACAACAACAGCCCCATCTGTCTATCCCTT	
9	DIR2r J6 G3	GCCTCGGACACCGCATGTATTACTGTGCGAGA	cggGGGcCT	TACTACTACTAGGTATGACGTCTGGGGCCAAAGGACCAAGGTACCGTCTCTTCAG	CTACAACAACAGCCCCATCTGTCTATCCCTT	

FIG. 47

49/101

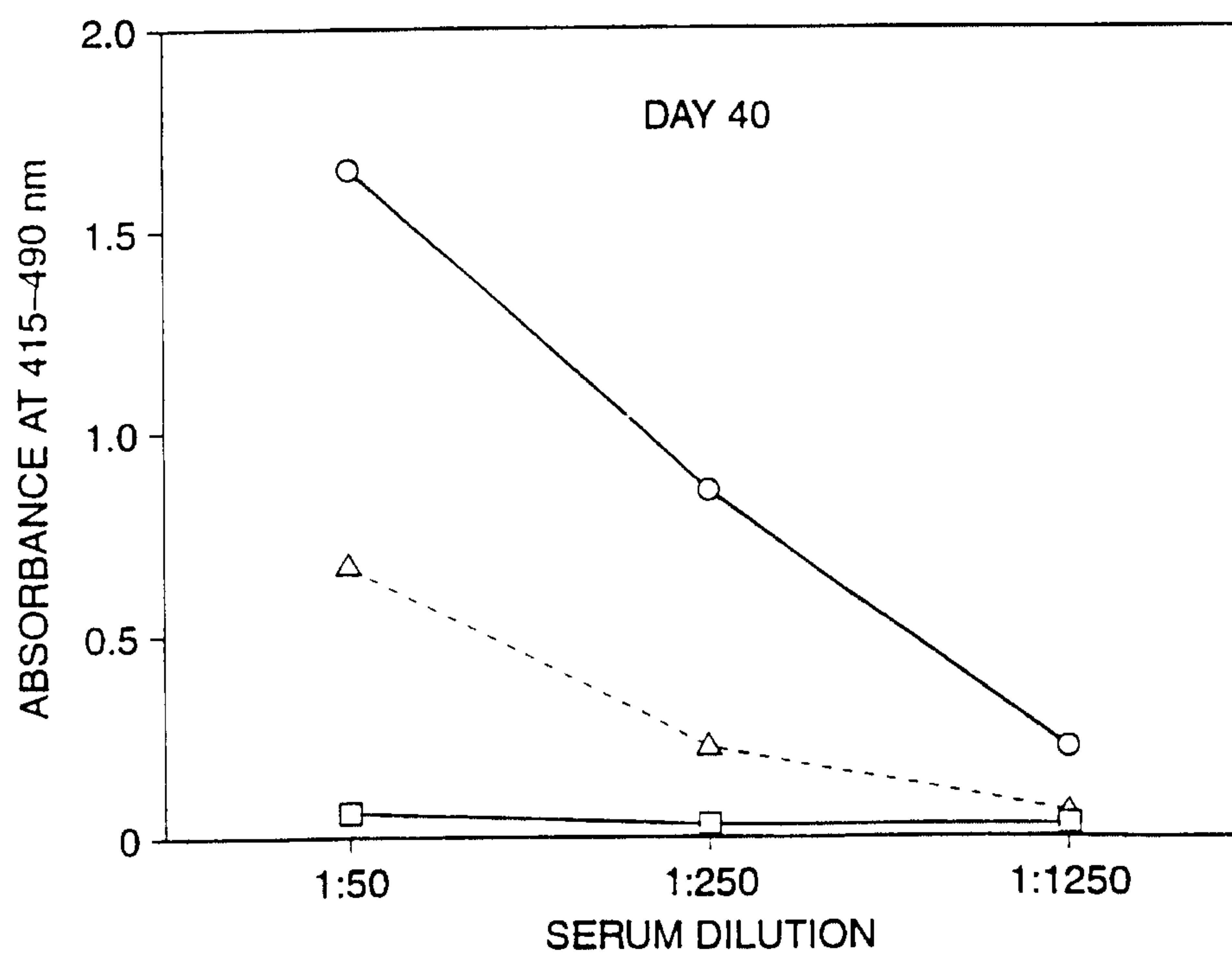
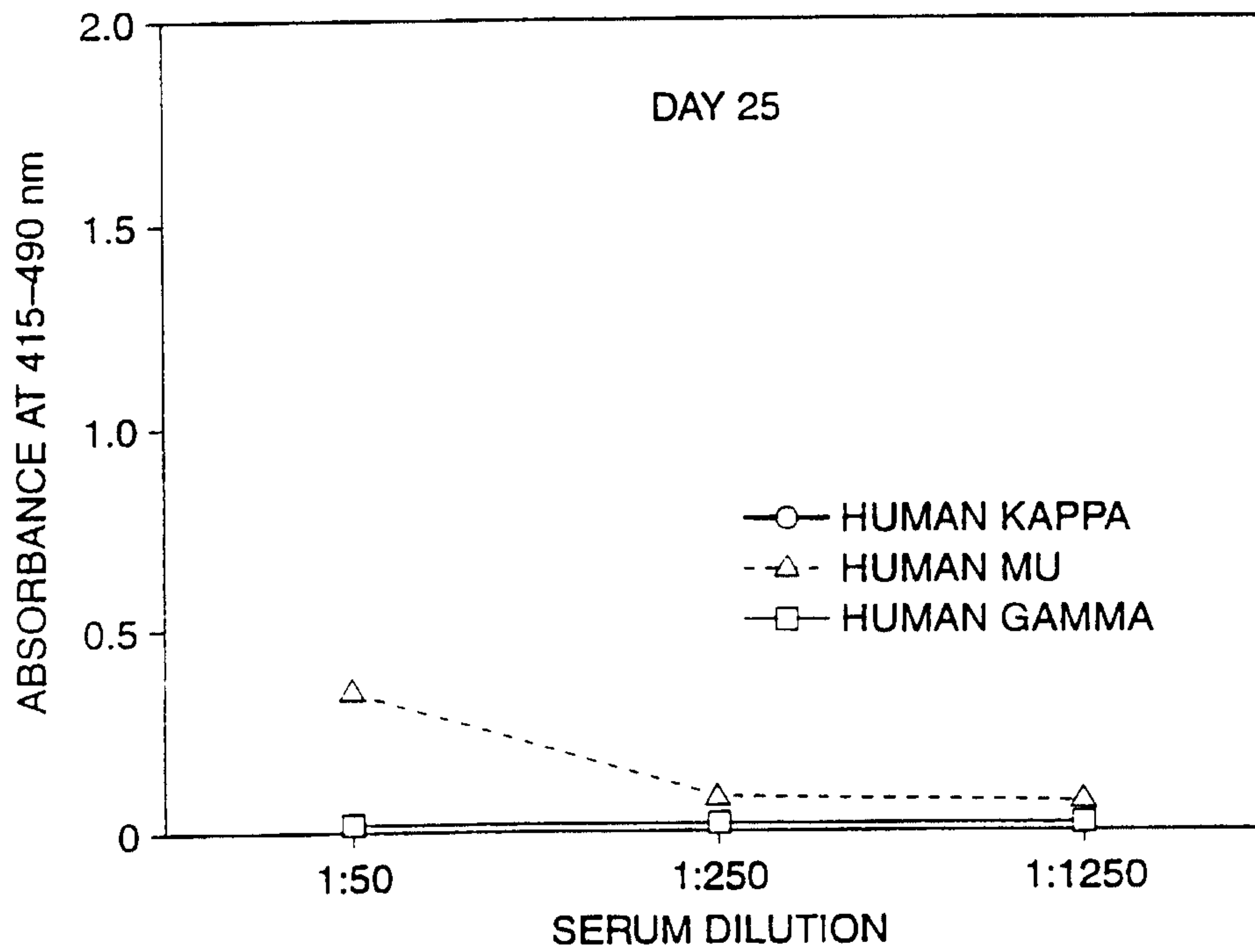


FIG. 48

50/101

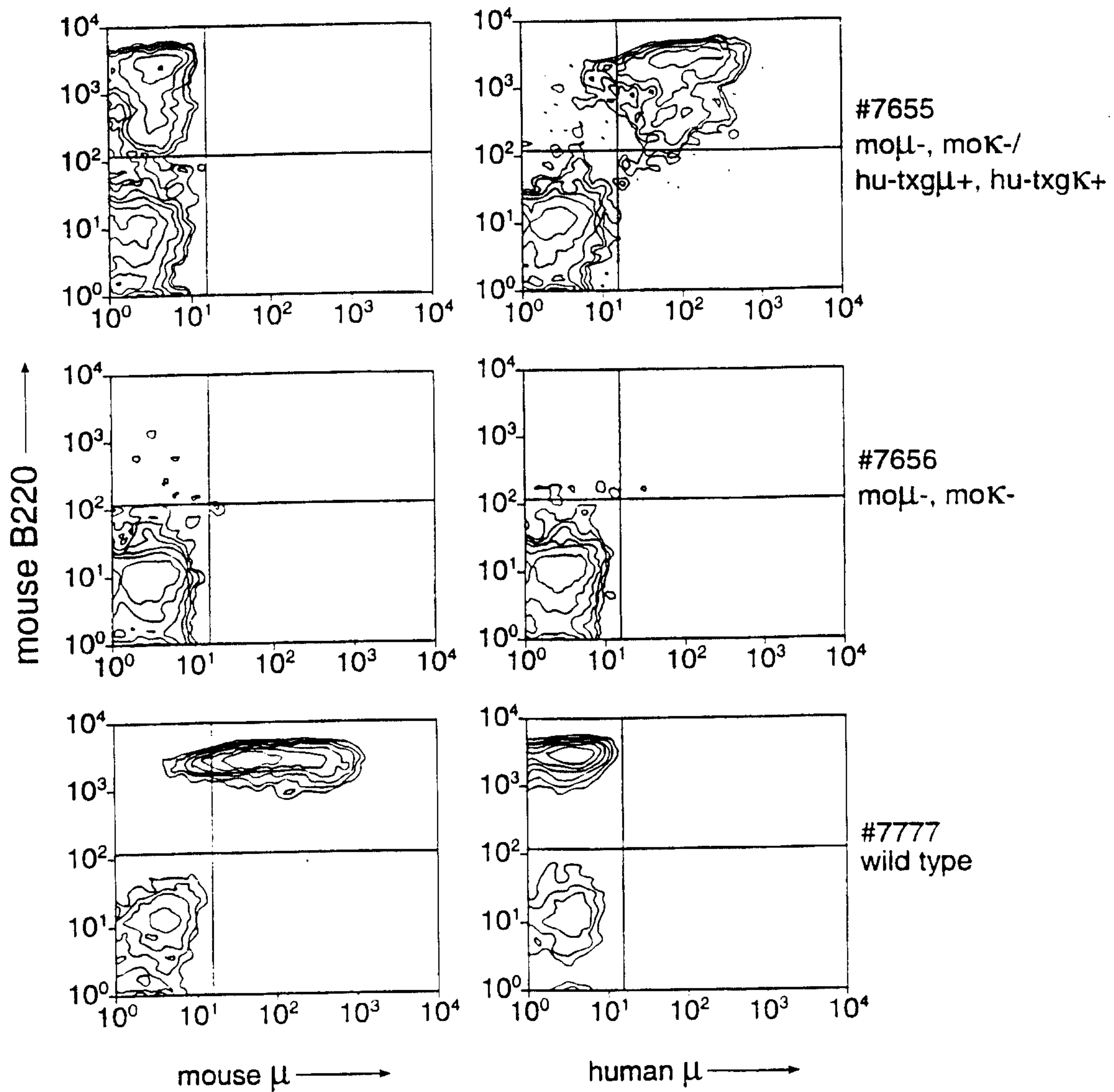


FIG. 49

51/101

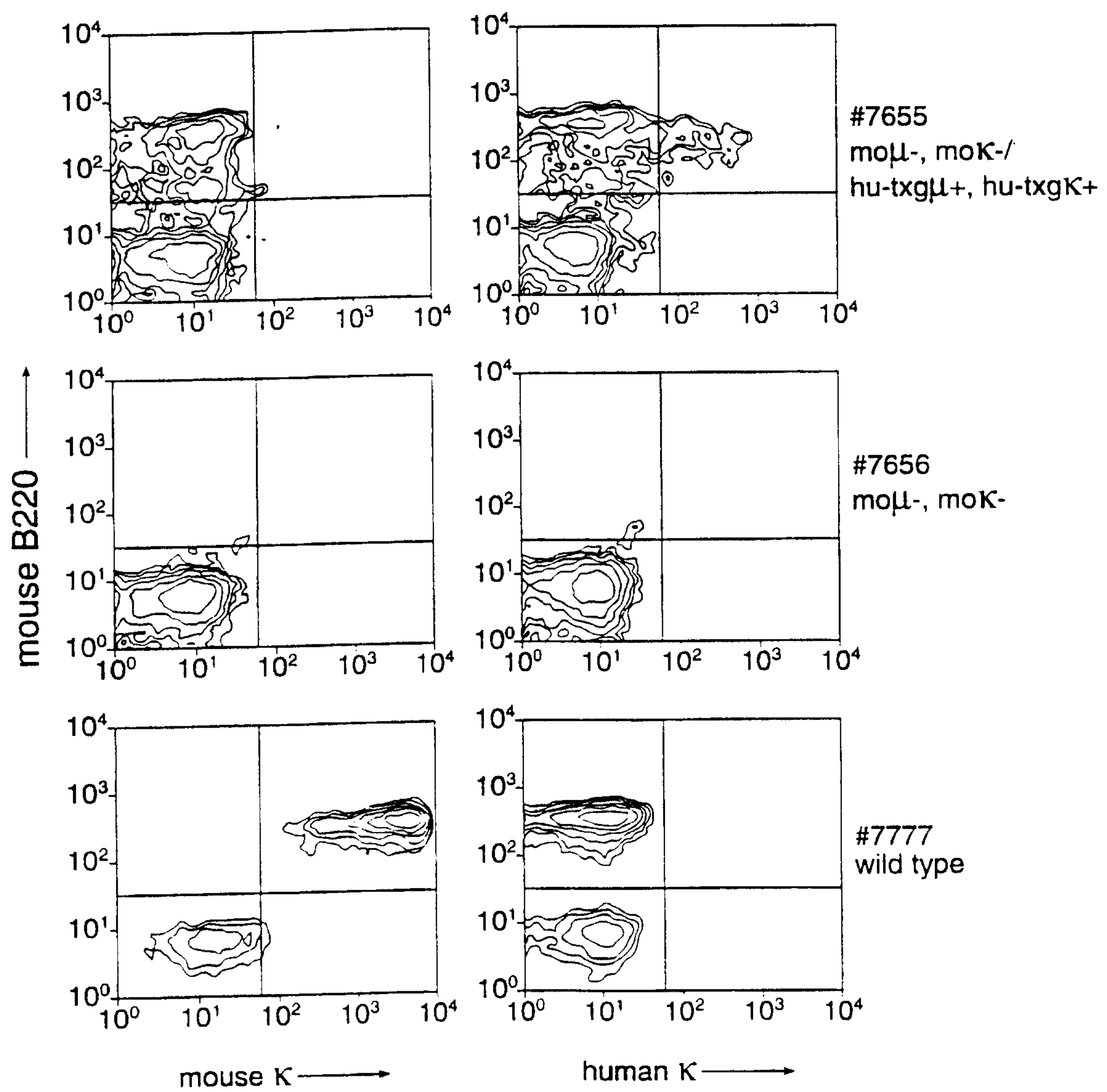


FIG. 50

52/101

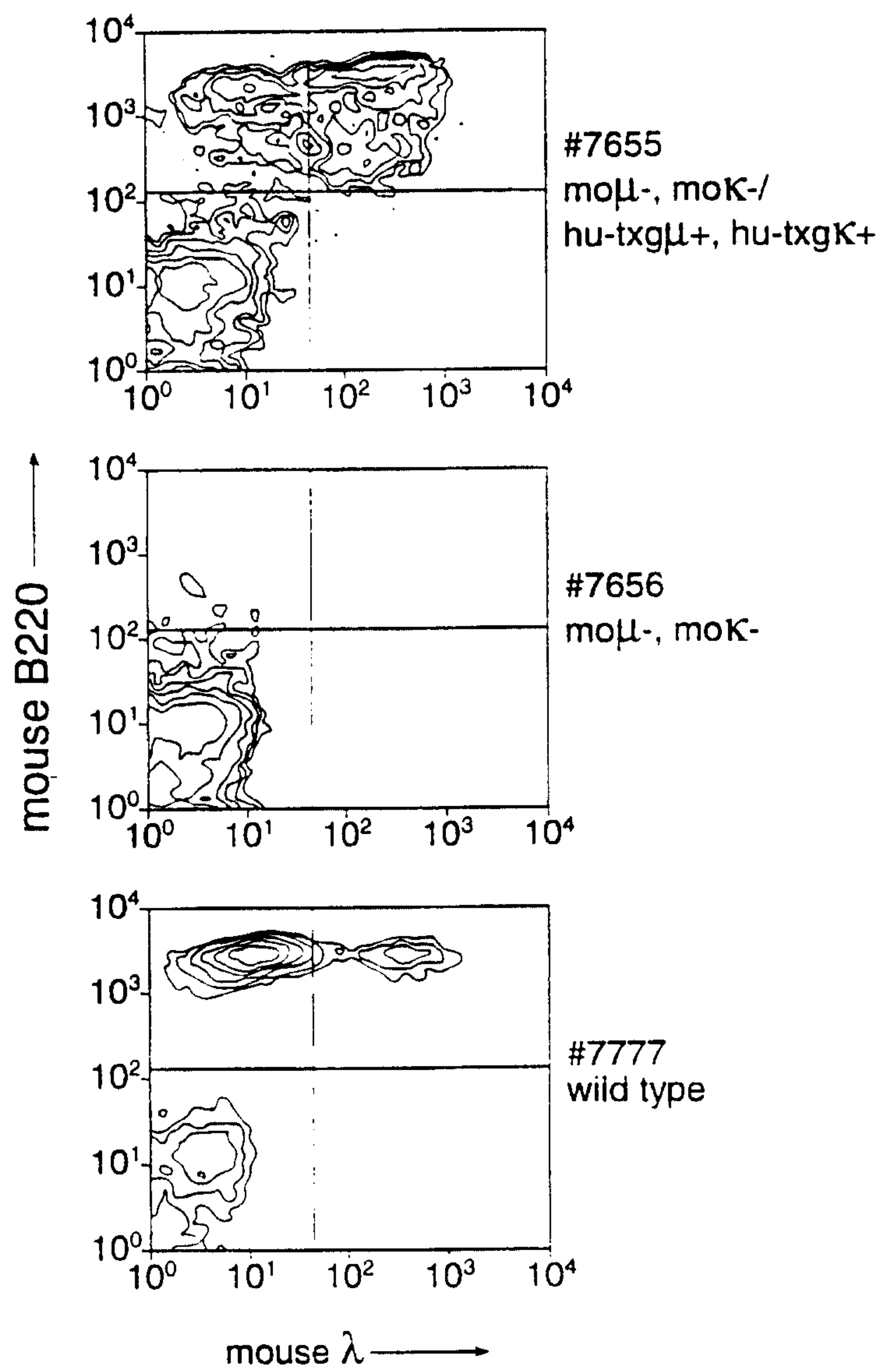


FIG. 51

53/101

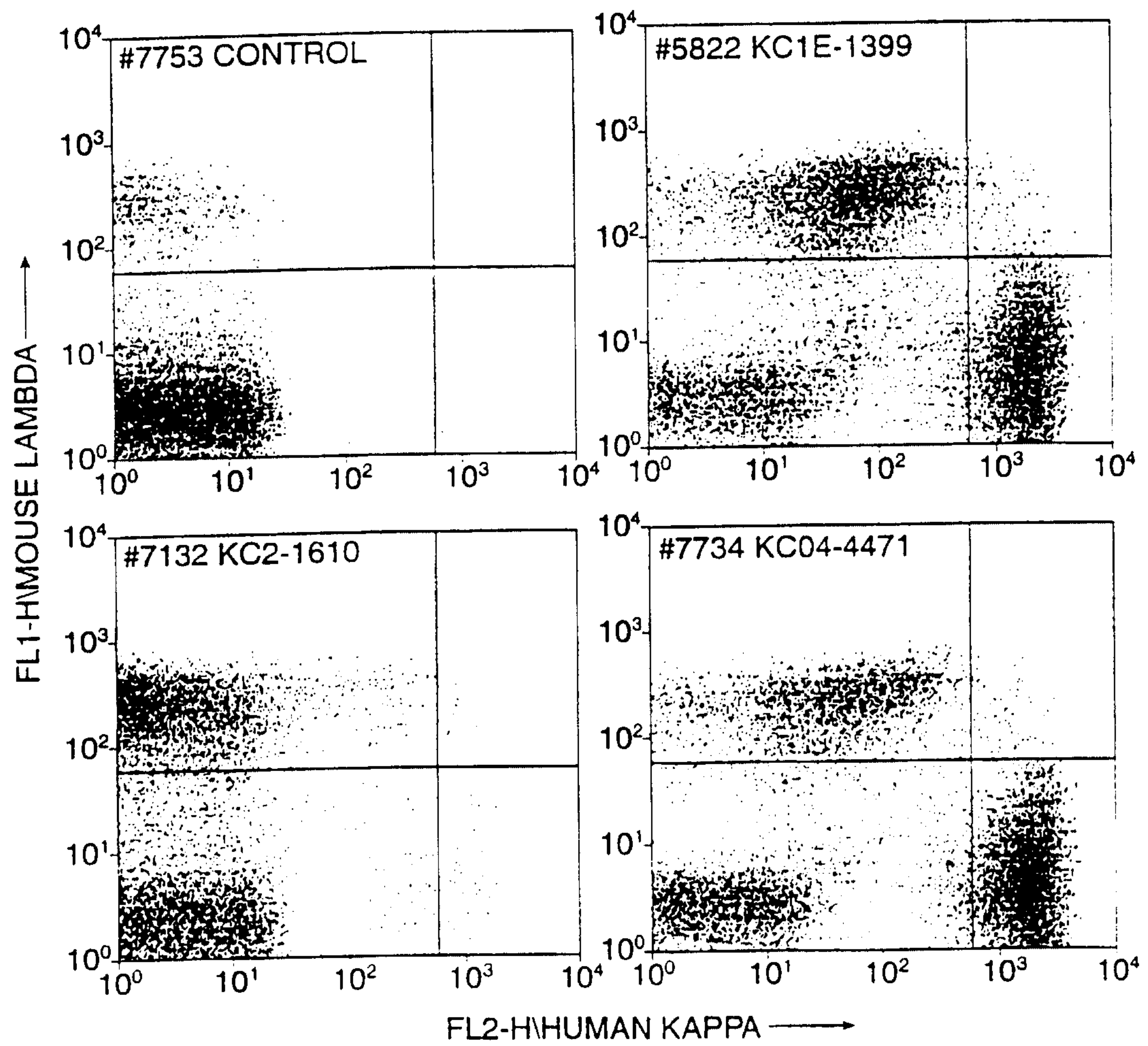


FIG. 52

54/101

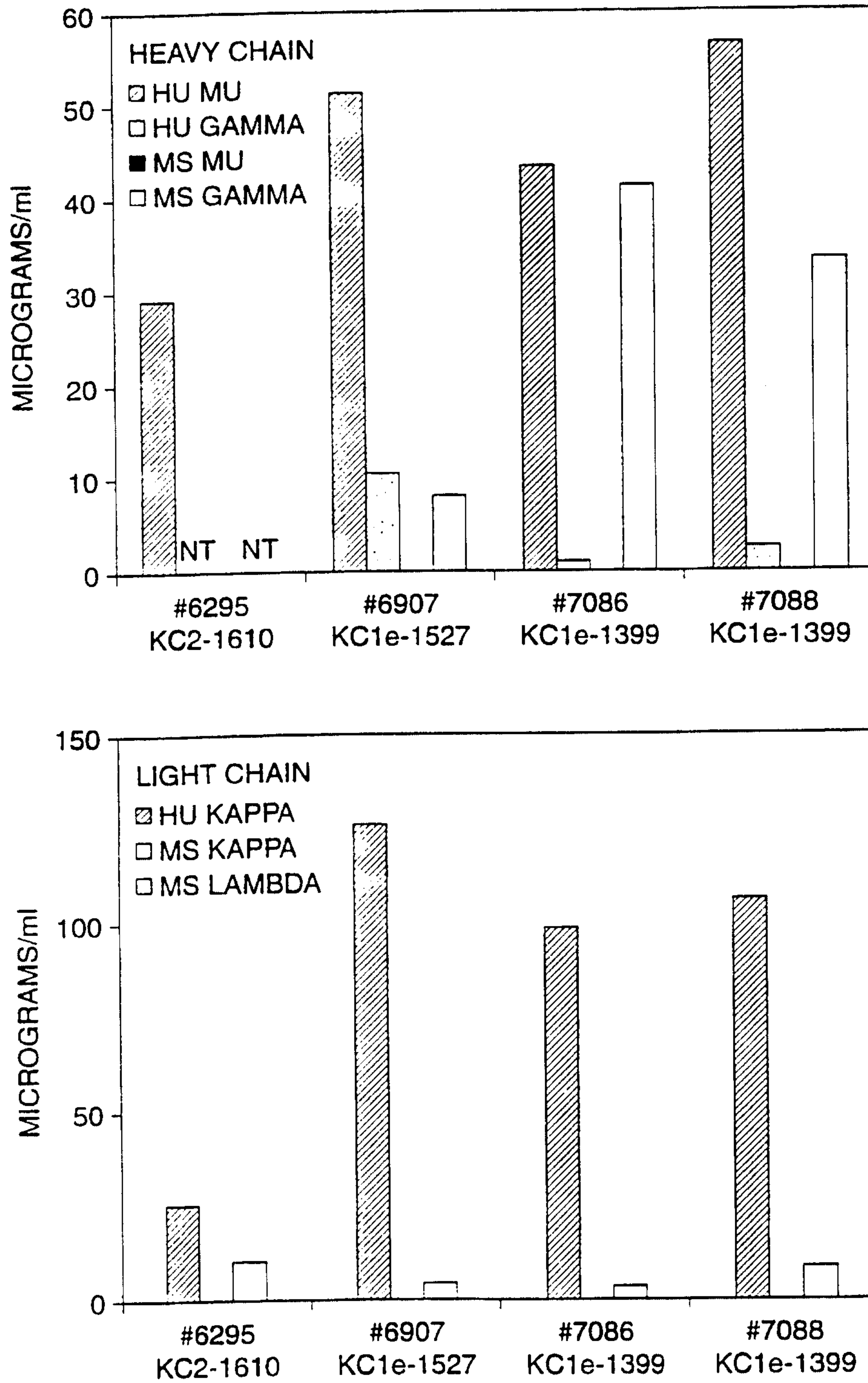


FIG. 53

55/101

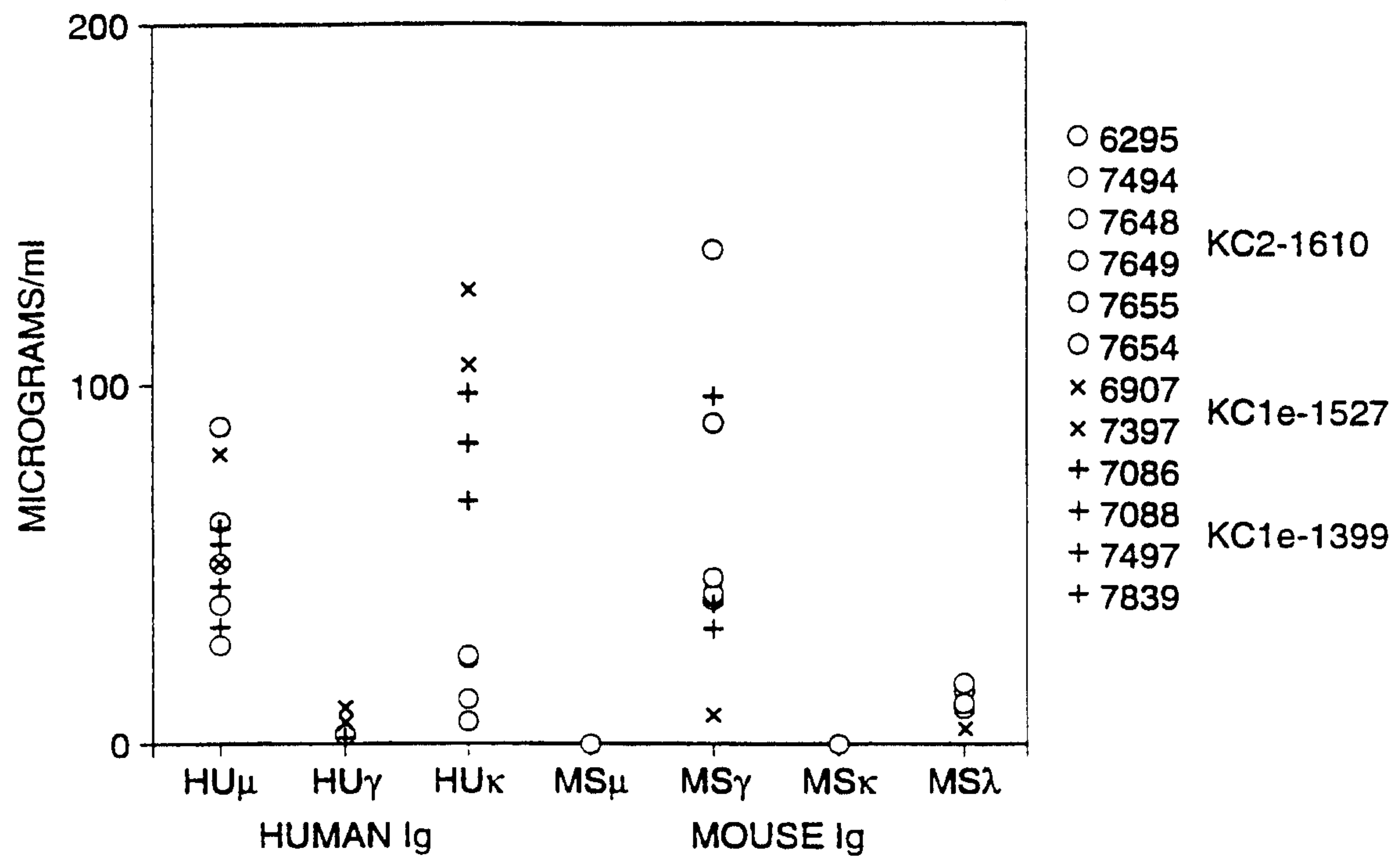


FIG. 54



FIGURE 59

56/101

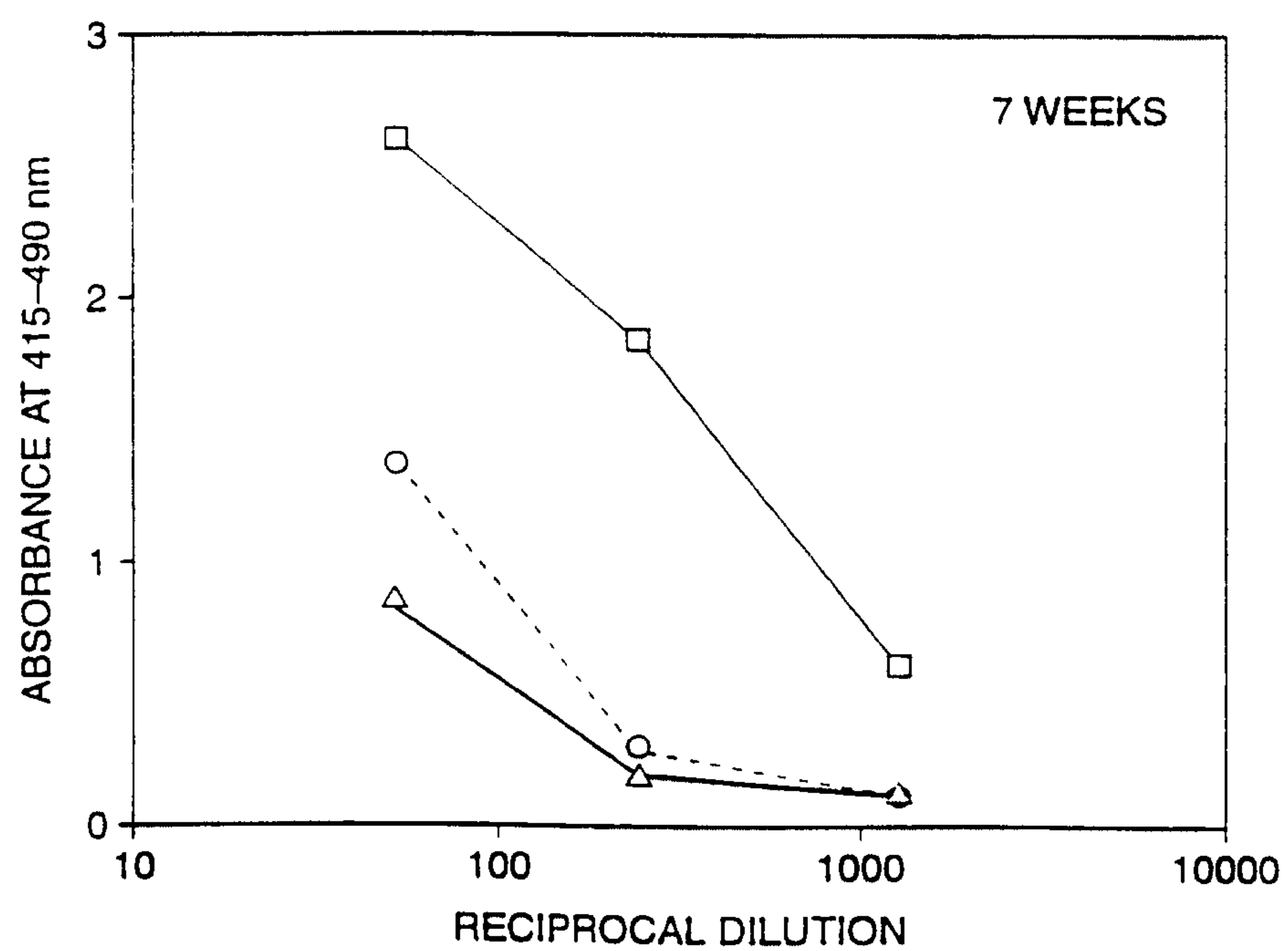
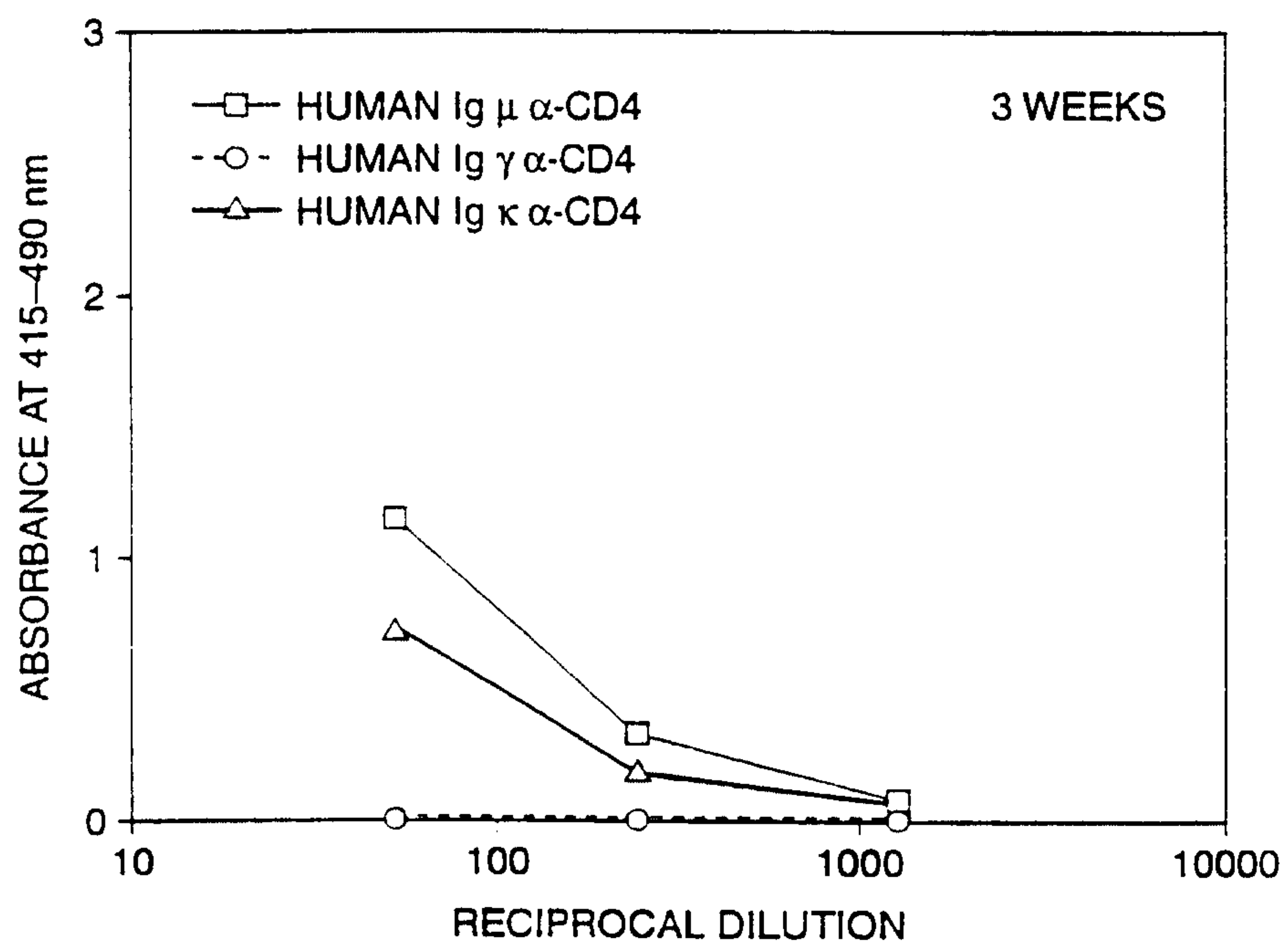
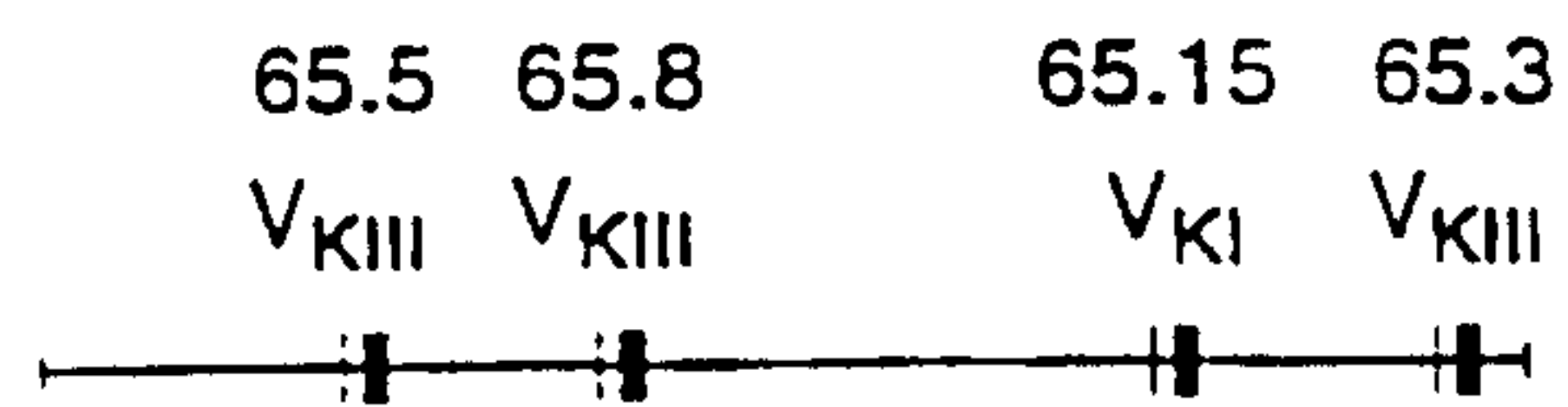
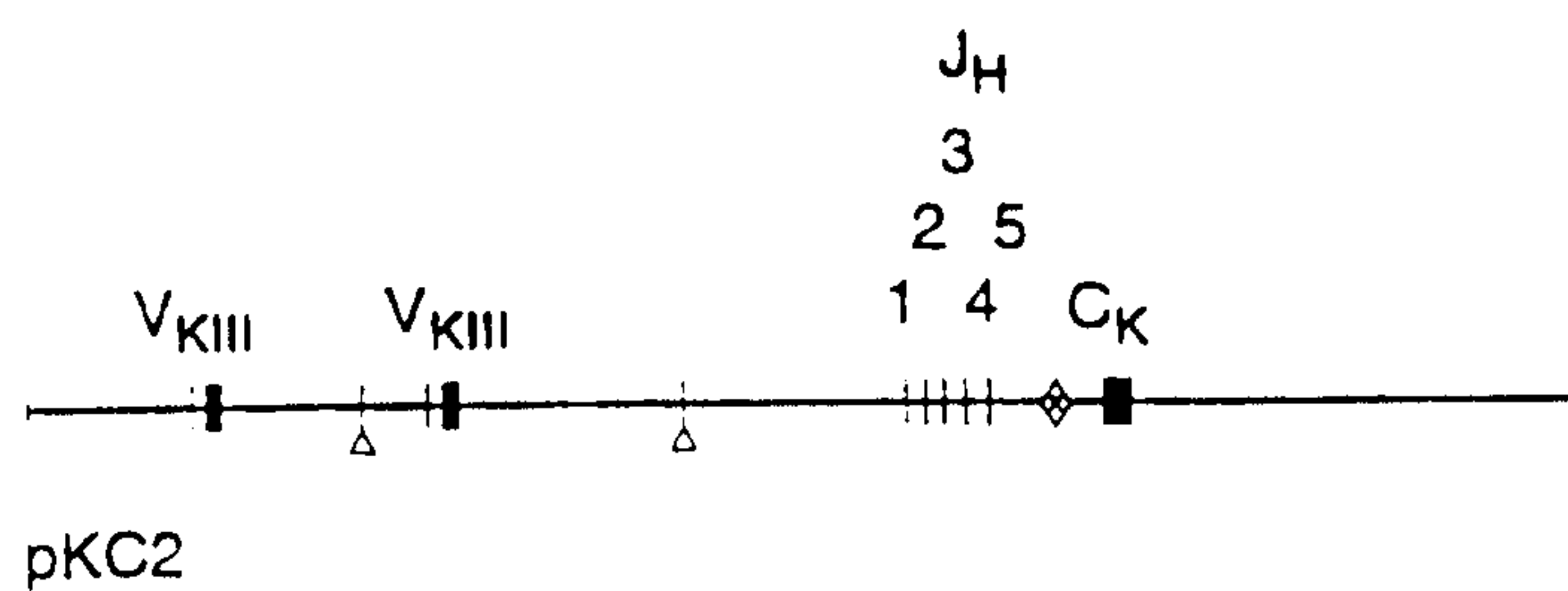
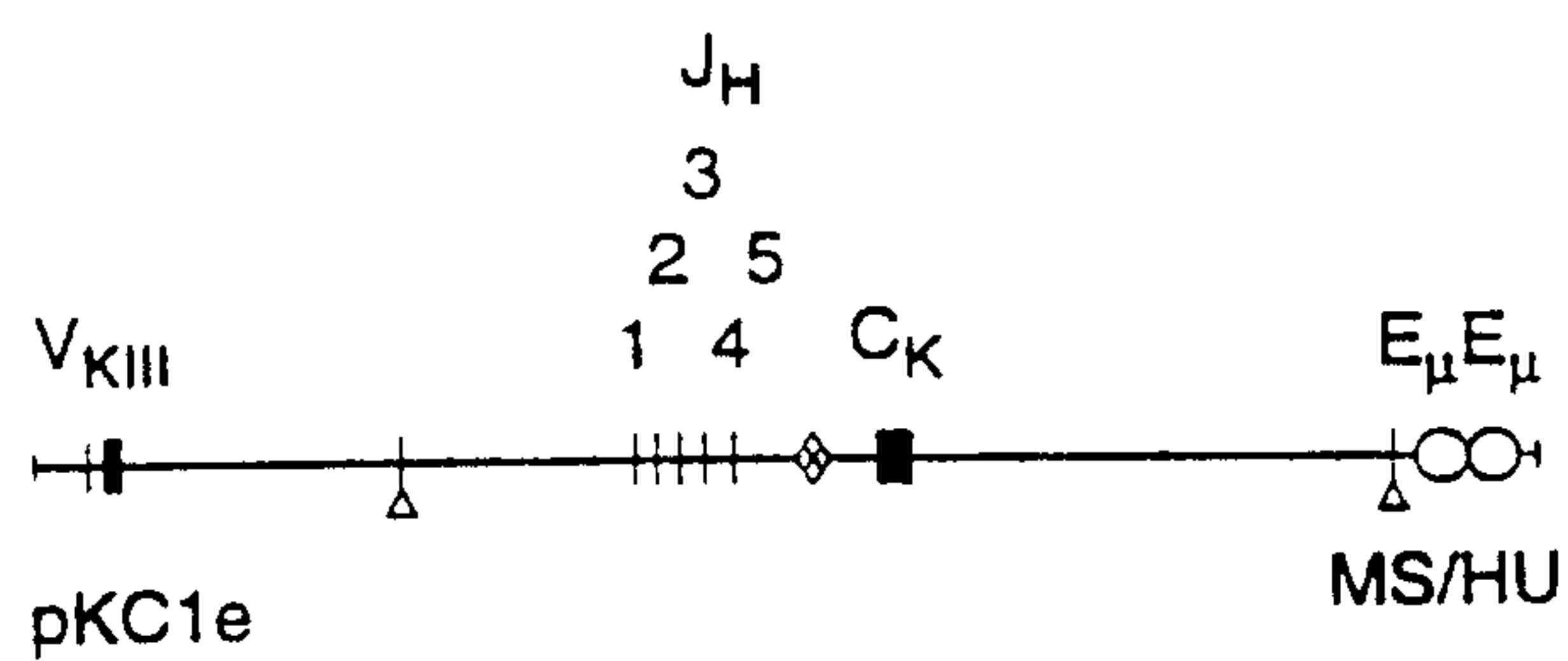
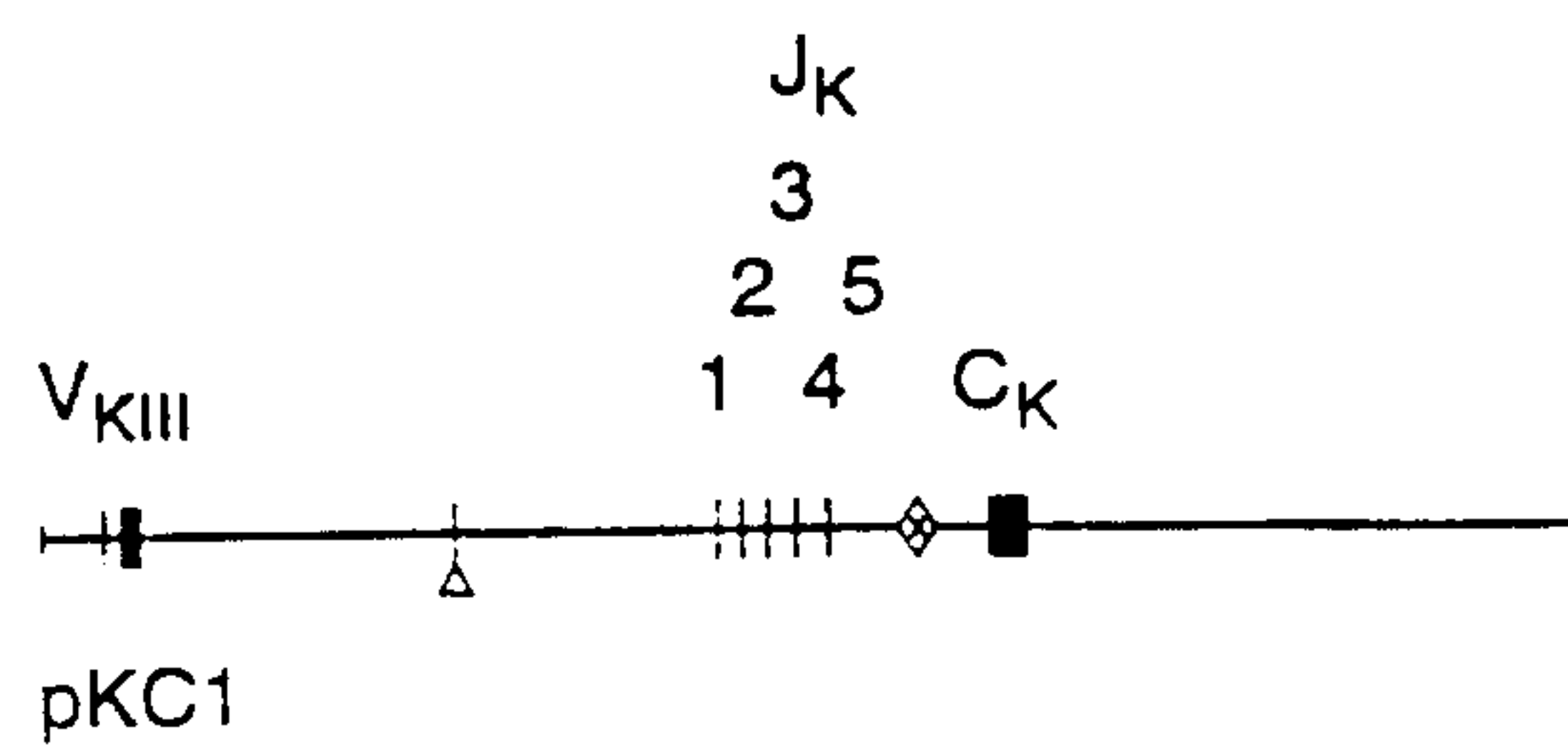


FIG. 55

57/101

0 5 10 15 20 25 30 35 40 45Kb

LIGHT CHAIN MINILOC



X

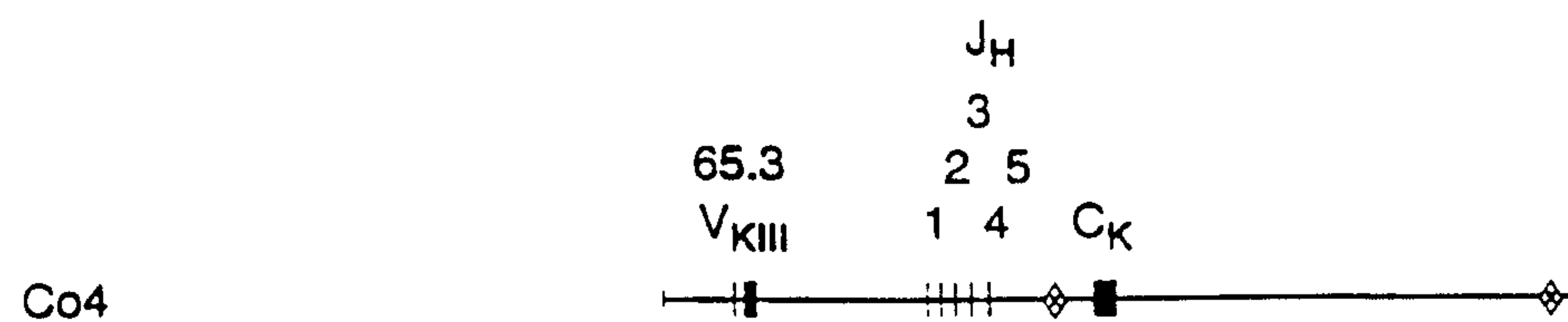


FIG. 56A

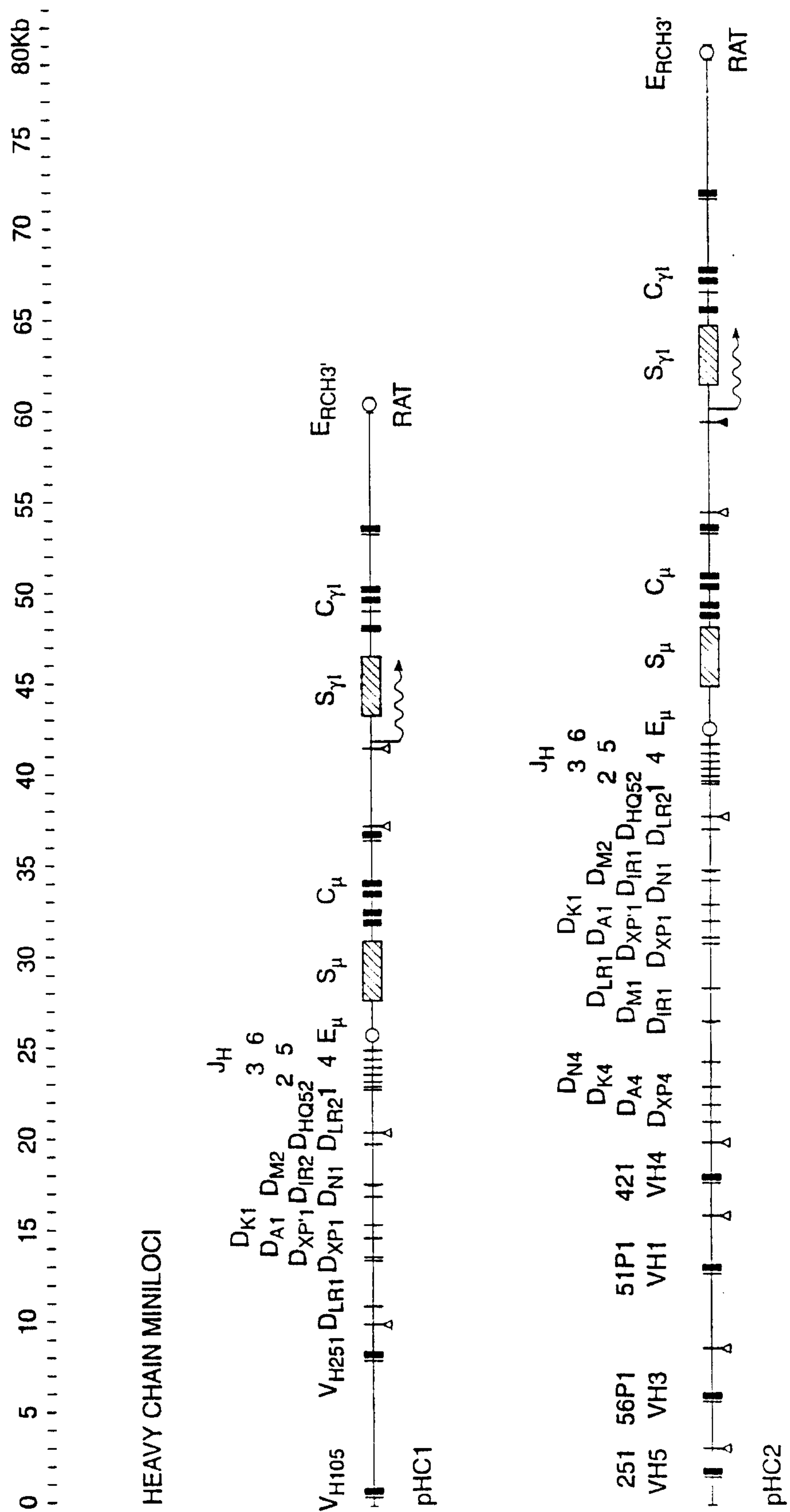


FIG. 56B

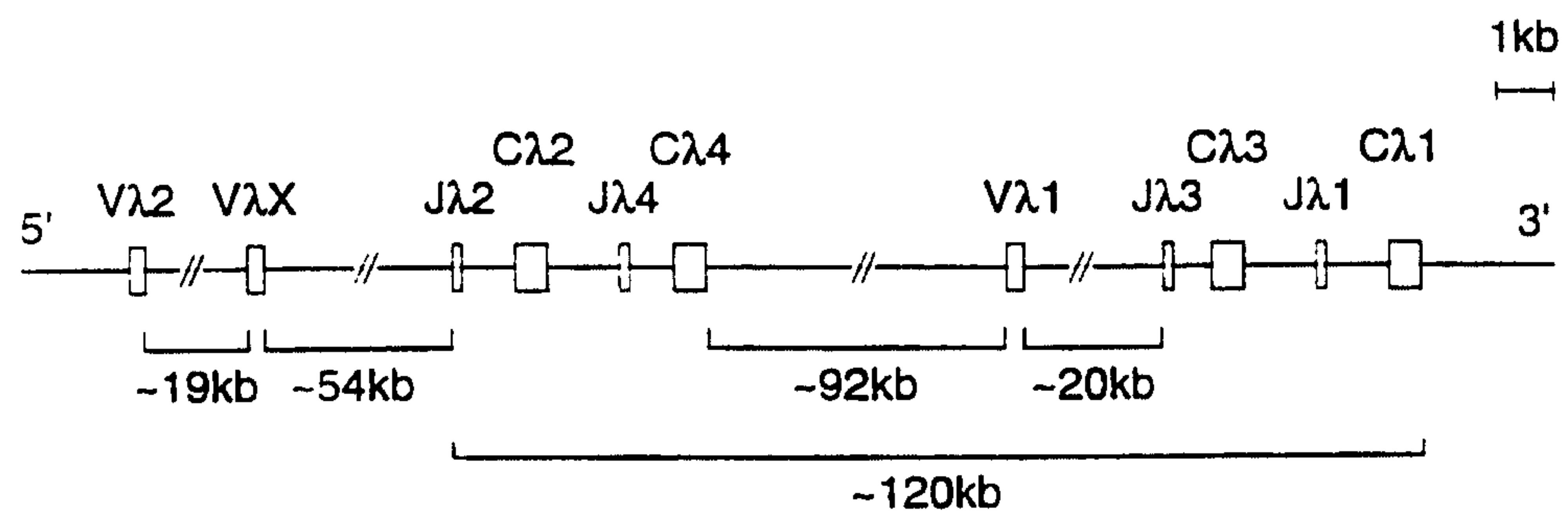
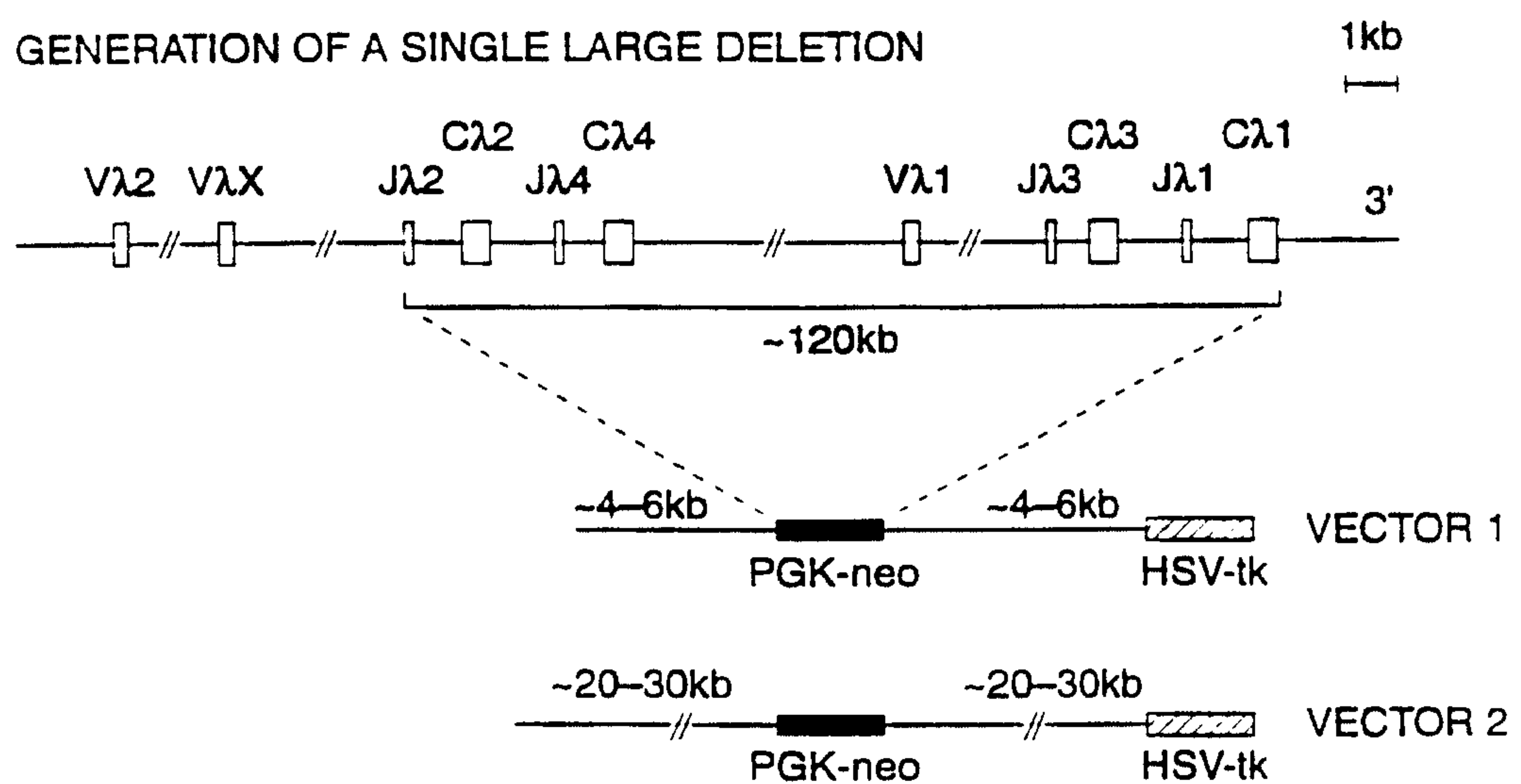


FIG. 57

GENERATION OF A SINGLE LARGE DELETION



GENERATION OF TWO SMALL DELETIONS

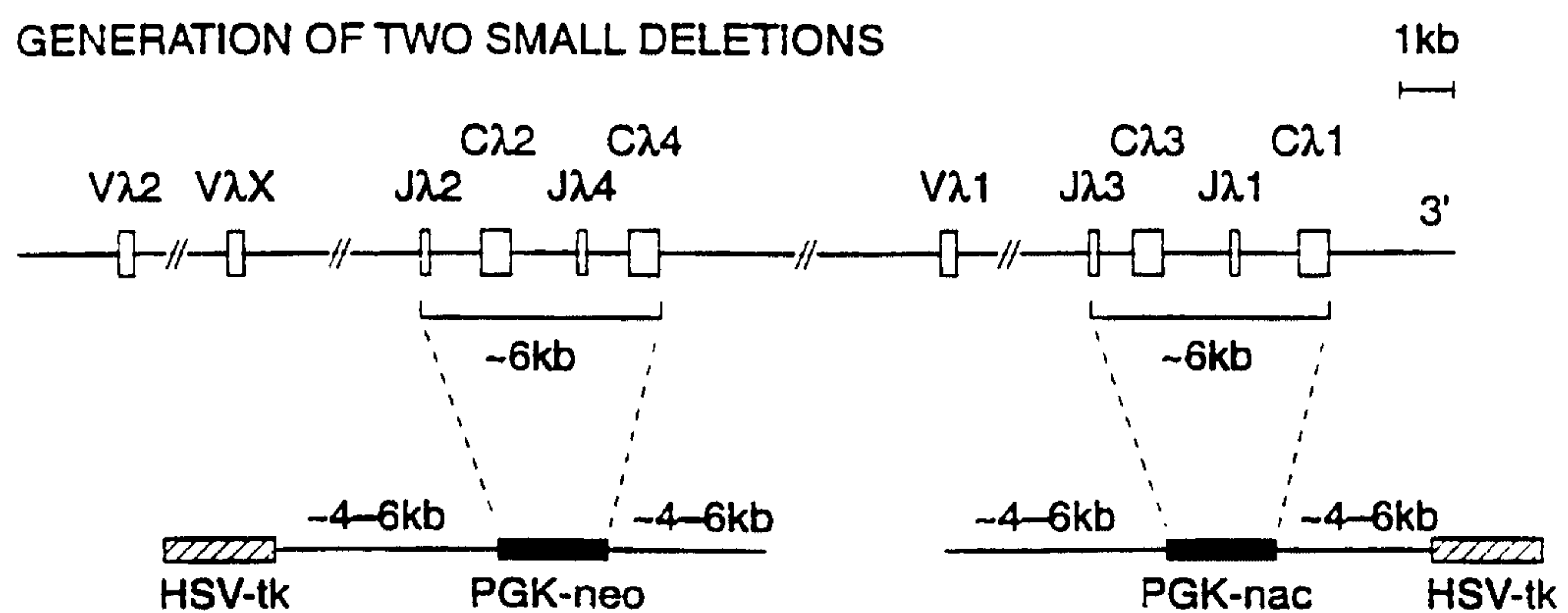


FIG. 58

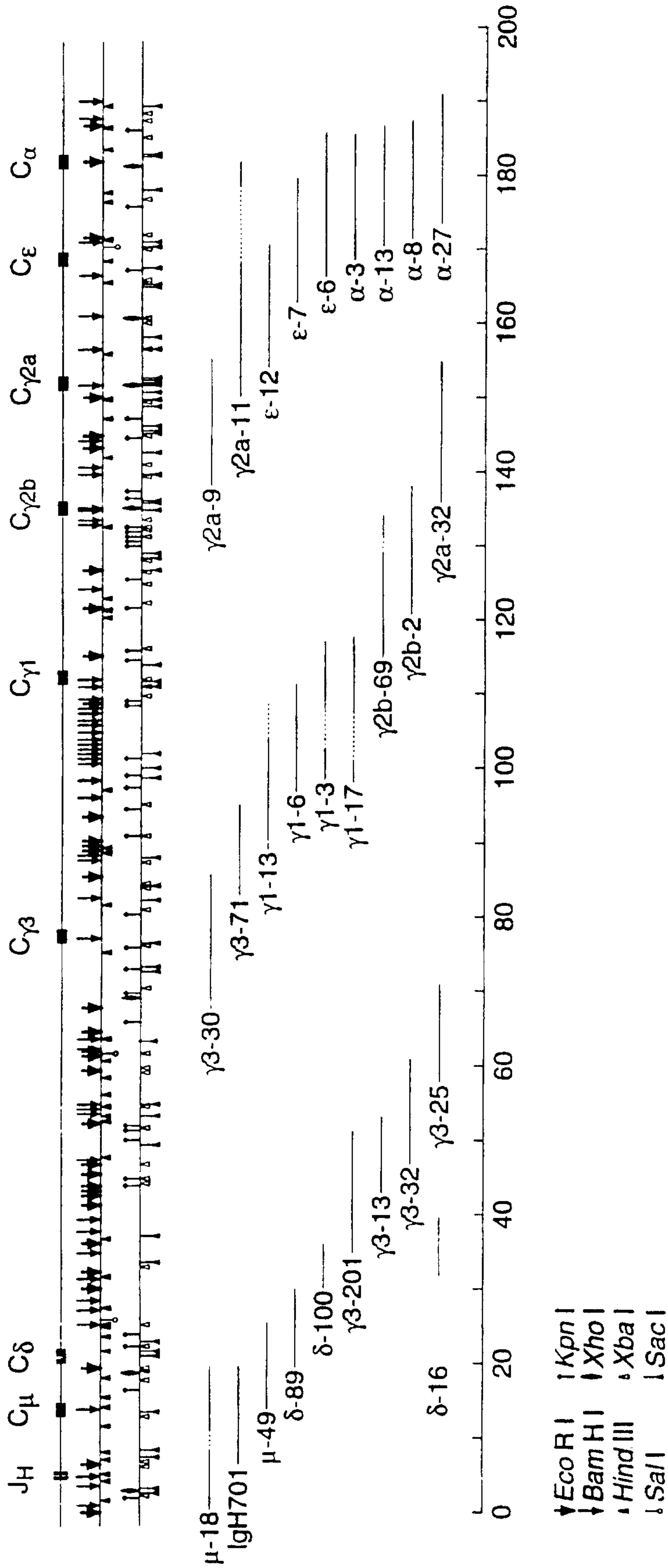


FIG. 60

CGAGAGGGGCGGGGGGAAGACTACTATCCCAGGCAGGTTTTAGGTTCCAGAGTCTGCGAG
AAATCCCACCATCTACCCACTGACACTCCCACCAGTCCTGTGCAGTGATCCCGTGATAAT
CGGCTGCCTGATTCACGATTACTTCCCTTTTCGGCACGATGAATGTGACCTGGGGAAAGAG
TGGGAAGGATATAACCACCGTGAAC TTCCACCTGCCCTCGCCTCTGGGGGACGGTACAC
CATGAGCAGCCAGTTAACCCTGCCAGCTGTCGAGTGCCCAGAAGGAGAGTCCGTGAAATG
TTCCGTGCAACATGACTCTAACCCCGTCCAAGAATTGGATGTGAATTGCTCTGGTAAAGA
ACGTTAGGGGGTCAGCTAGGGGTGGGATAAGTCCTACCTTATCTAGATCCATATATCCCT
CTGATGCACACCCTCACAGGAATCCCTCAGAAACCTCCACTATGGGGATTGGGGGAAGGA
AGCGTAAACAGGTCTAGAAGGAGCTGGAGGCCTCAGAACATCCAGAAACGGGGACAGCAA
AGGAGACAAGGAGAATATACTGATTTGCTAGGACATCTTCTGTTACAGGTCCTACTCCTC
CTCCTCCTATTACTATTCTTCTGCCAGCCCAGCCTGTCACTGCAGCGGCCAGCTCTTG
AGGACCTGCTCCTGGGTTTCAGATGCCAGCATCACATGTACTCTGAATGGCCTGAGAAATC
CTGAGGGAGCTGCTTTTCACCTGGGAGCCCTCCACTGGGAAGGATGCAGTGCAGAAGAAAG
CTGCGCAGAATTCCTGCGGCTGCTACAGTGTGTCCAGCGTCCTGCCTGGCTGTGCTGAGC
GCTGGAACAGTGGCGCATCATTCAAGTGCACAGTTACCCATCCTGAGTCTGGCACCTTAA
CTGGCACAATTGCCAAAGTCACAGGTGAGCTCAGATGCATACCAGGACATTGTATGACGT
TCCCTGCTCACATGCCTGCTTTCTTCTATAATACAGATGCTCAACTAACTGCTCATGTC
CTTATATCACAGAGGGAAATTGGAGCTATCTGAGGAACTGCCCAGAAGGGAAGGGCAGAG
GGGTCTTGCTCTCCTTGCTGAGCCATAACTCTTCTTTCTACCTTCCAGTGAACACCTTC
CCACCCCAGGTCCACCTGCTACCGCCGCCGTGCGAGGAGCTGGCCCTGAATGAGCTCTTG
TCCCTGACATGCCTGGTGCGAGCTTTCAACCCTAAAGAAGTGCTGGTGCGATGGCTGCAT
GGAAATGAGGAGCTGTCCCCAGAAAGCTACCTAGTGTTTGAGCCCCTAAAGGAGCCAGGC
GAGGGAGCCACCACCTACCTGGTGACAAGCGTGTTGCGTGTATCAGCTGAAACCTGGAAA
CAGGGTGACCAGTACTCCTGCATGGTGGGCCACGAGGCCTTGCCCATGAACTTCACCCAG
AAGACCATCGACCGTCTGTGCGGTAAACCCACCAATGTCAGCGTGTCTGTGATCATGTCA
GAGGGAGATGGCATCTGCTACTGAGCCACCCTGCCTGTCCCTACTCCTAGAATAAACTCT
GTGCTCATCCAAAGTATCCCTGCACTTCCACCCAGTGCCTGTCCACCACCCTGGGGTCTA
CGAAACACAGGGAGGGGTCAGGGCCCAGGGAGGGAGAAATACCACCACCTAAGC

FIG. 61

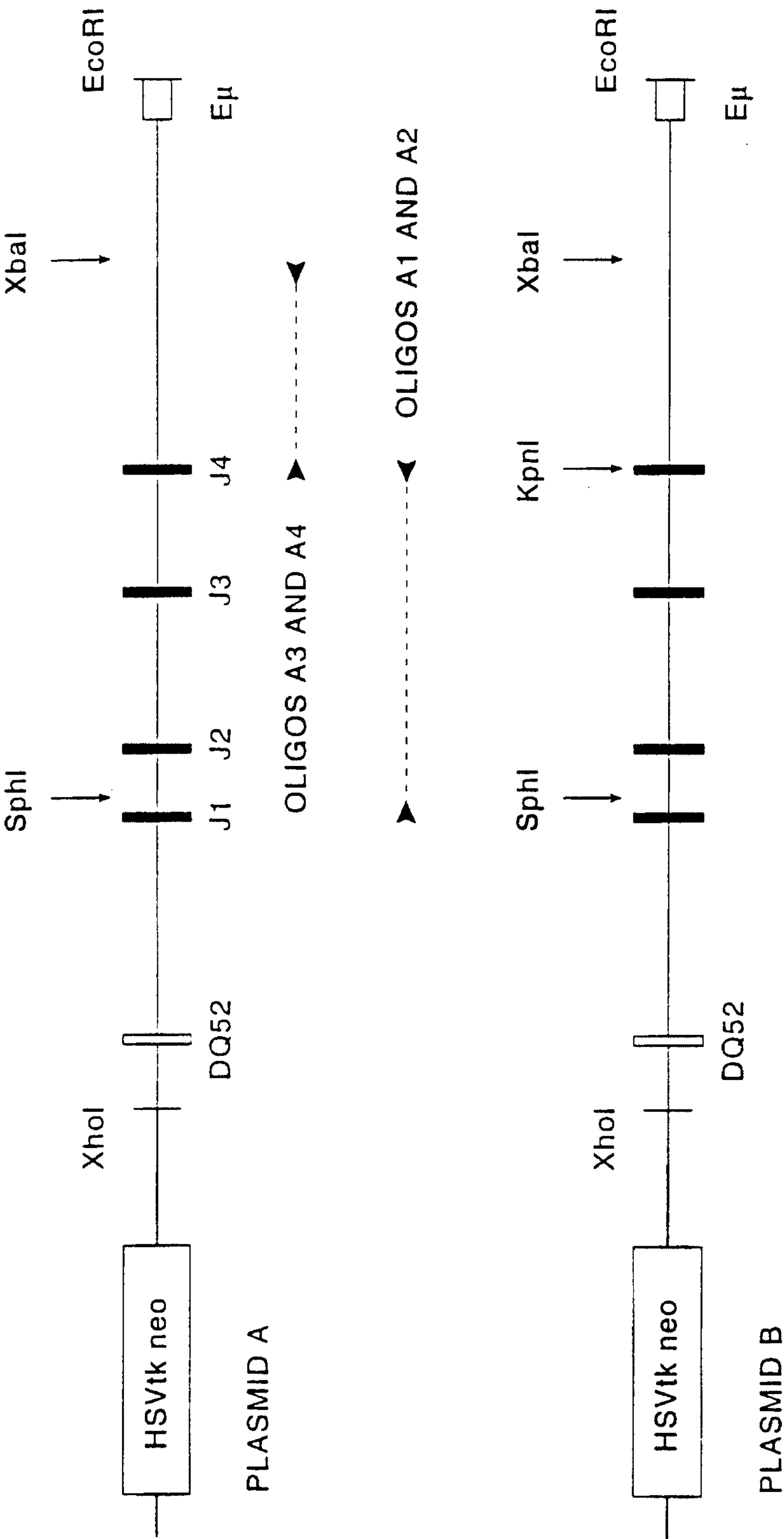


FIG. 62

63/101

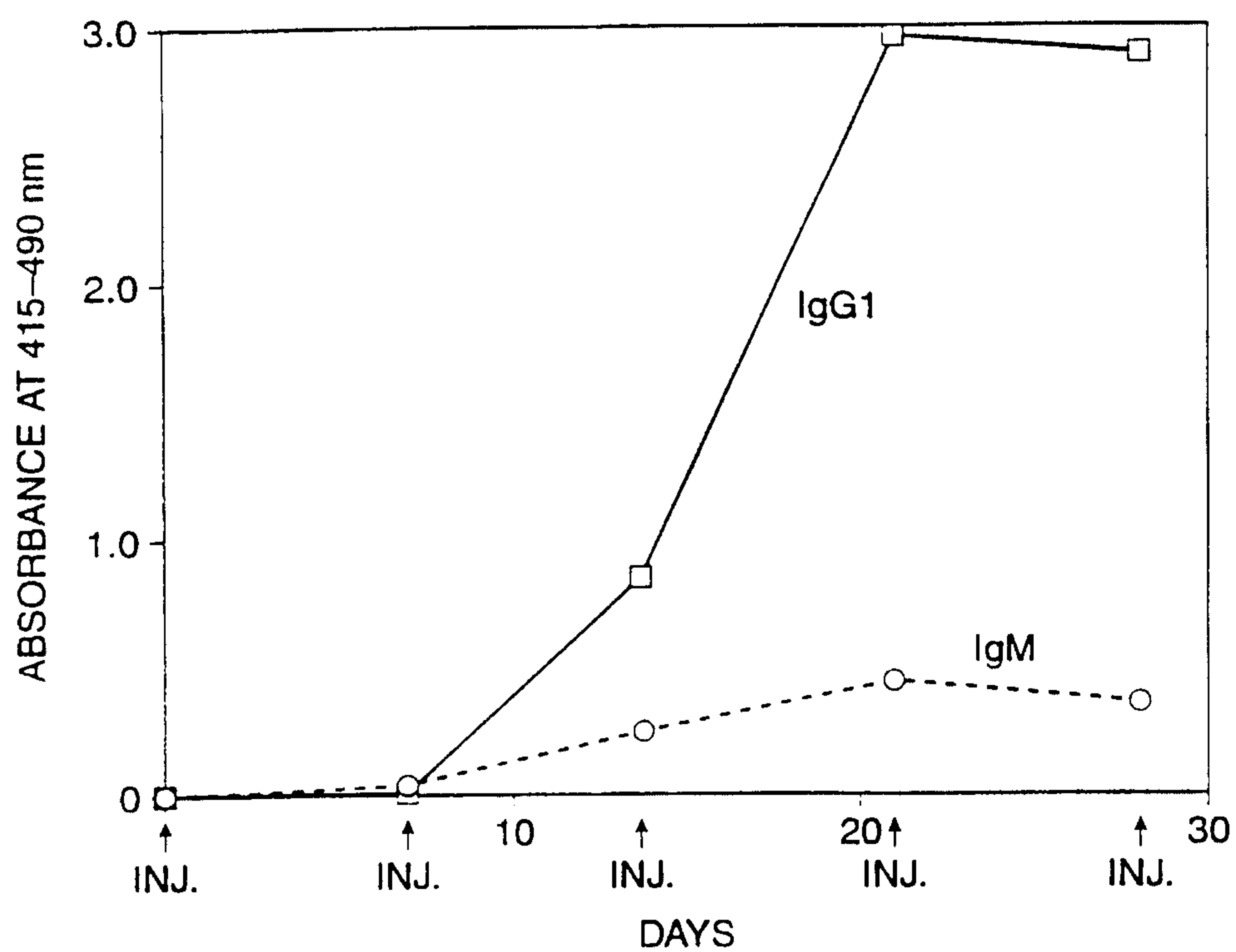


FIG. 63

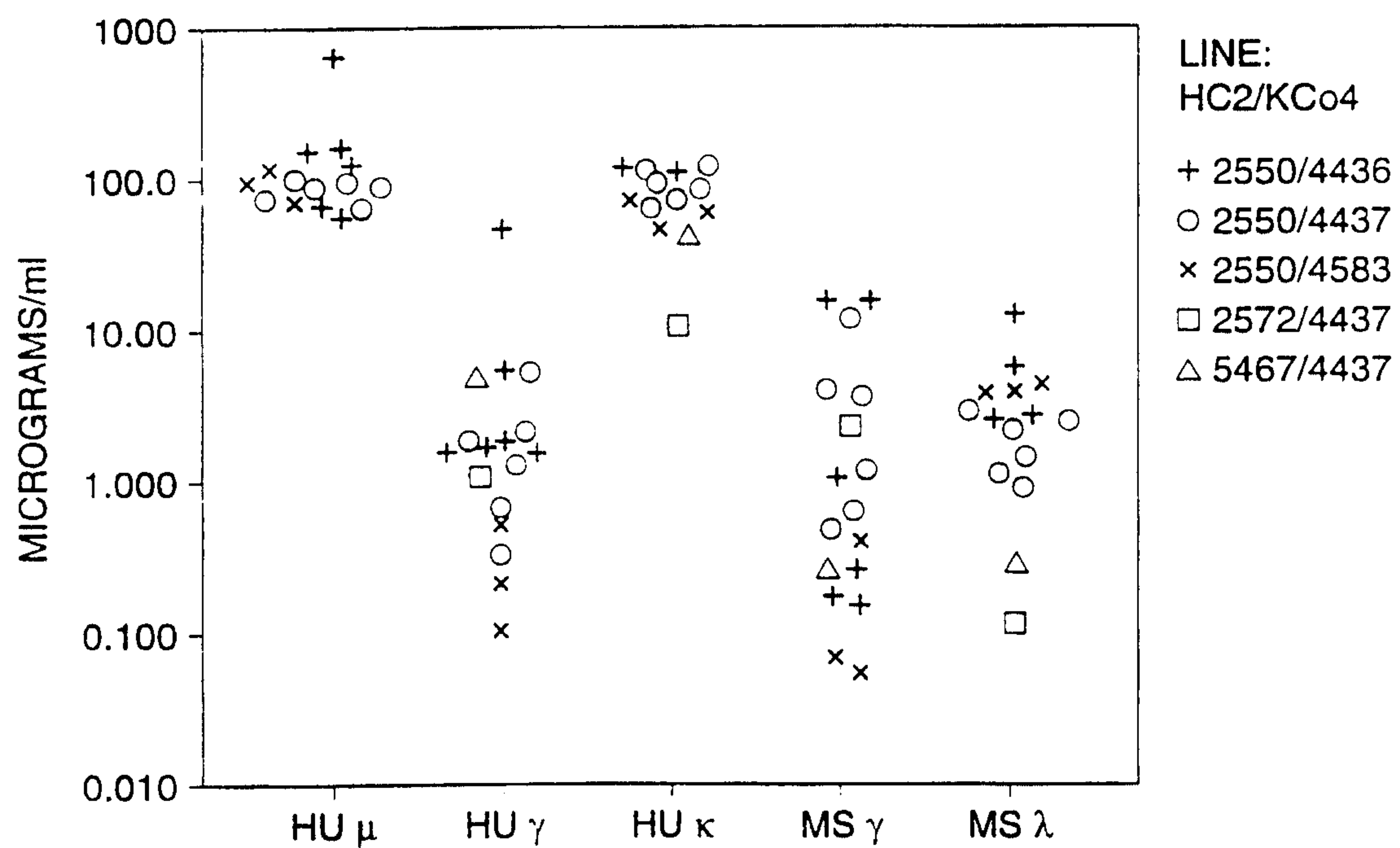


FIG. 70

64/101

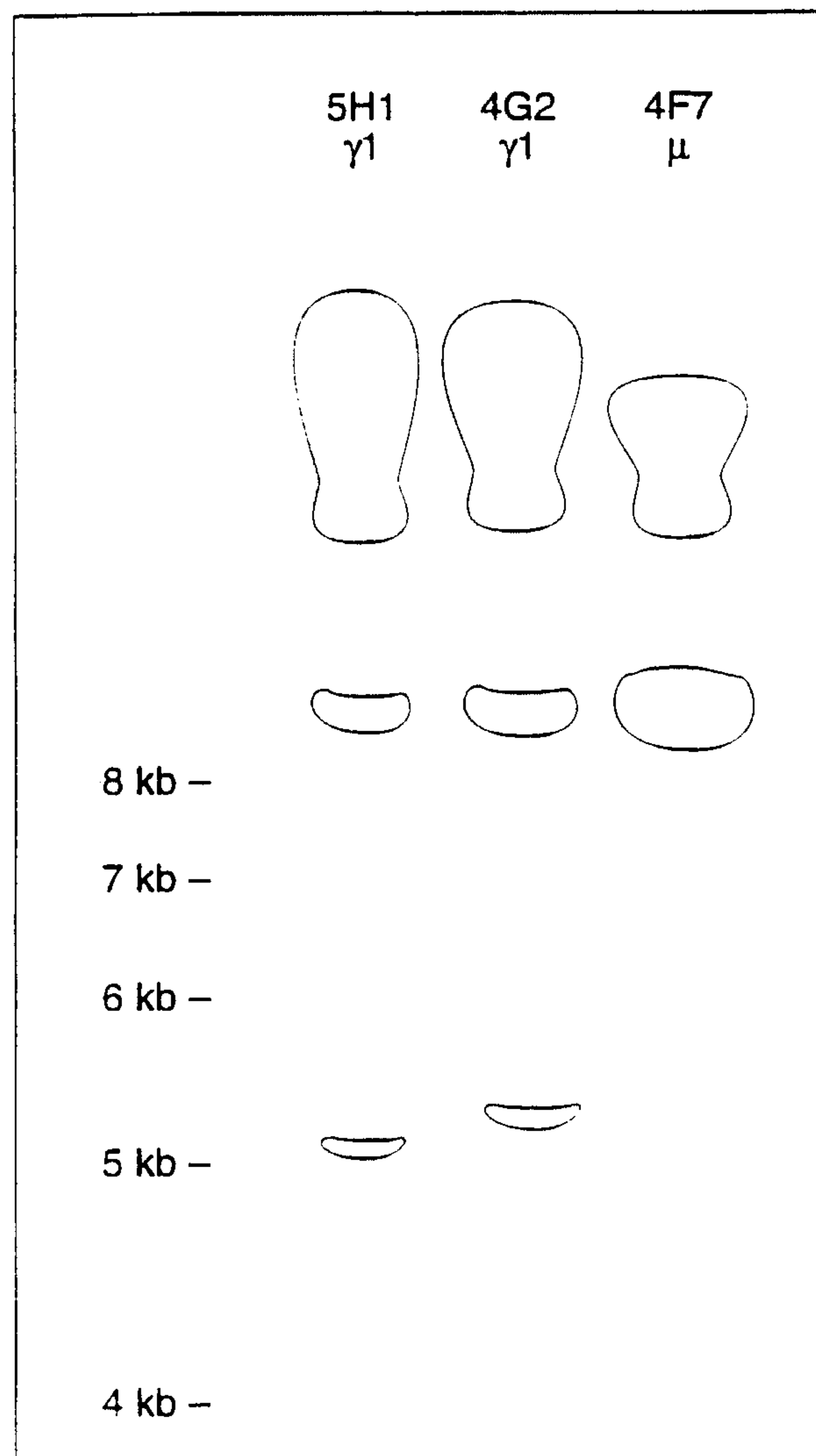


FIG. 64A

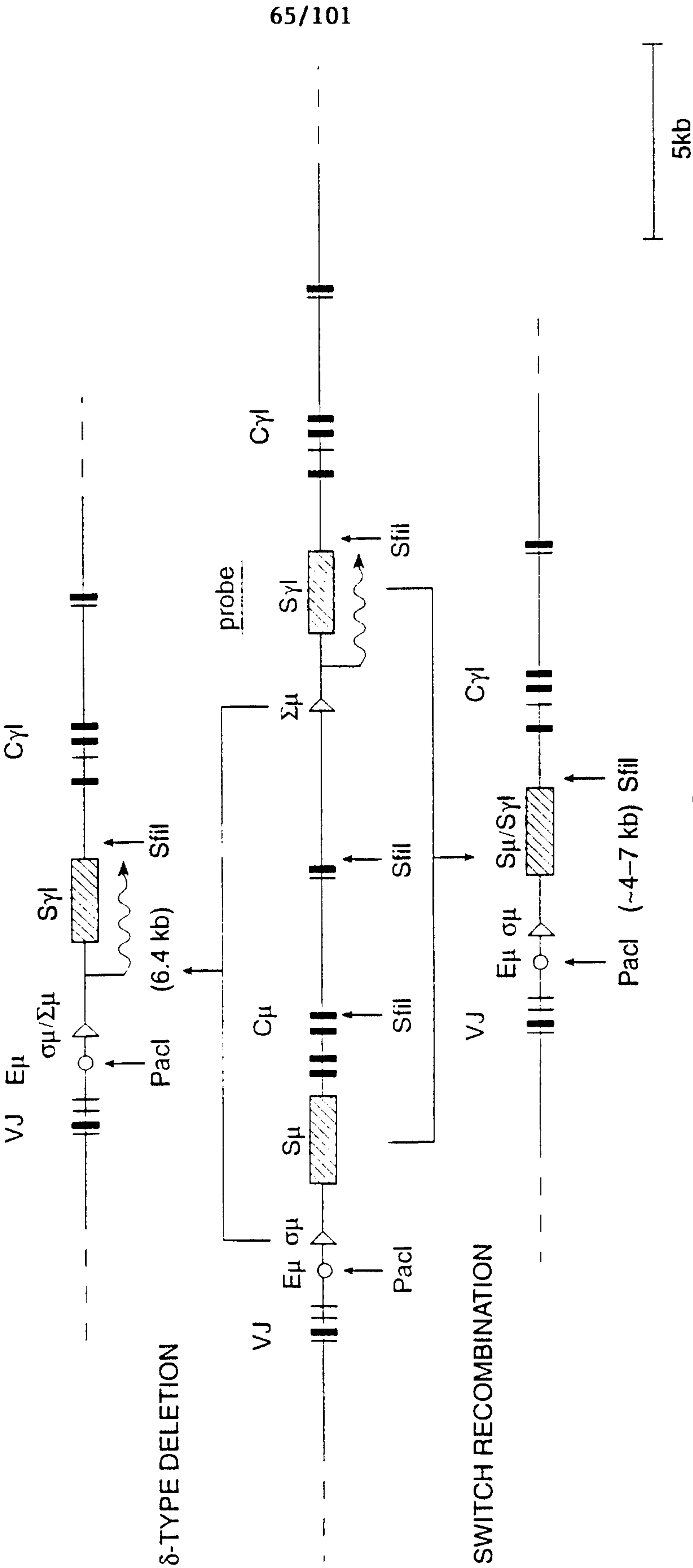


FIG. 64B

V _H 251	N D N	J	C _γ	mouse γ1
2357.t5 DXP'1 J6	gcctcggacacccgccatgtattactgtgTgaga CATTtatgggttcg999agttacG	cggTgTgAacgtctggggccaagggaaccacggTcaccgTctctctcag ccaaaacgacaccccccatctgtctatccac		
2357.t7 DHQ52 J3	gcctcggacacccgccatgtattactgtgcgaga CactgggCATTGGAT	gctCttgatGtctggggccaagggaacaatgCtcacgTctcttcag ccaaaacgacaccccccatctgtctatccac		
2357.t2 DHQ52 J3	gcctcggacacccgccatgtattactgtgcgaga actggggatGAT	gcttttgatatctggggccaagggaacaatggTcacgTctcttcag ccaaaacacacaccccccatcagTctatccac		mouse γ2b
2357.t3 D? J3	gActcggacacccgccatgtattactgtgcgaga CAGGGGAGAGAT	gcttttagatatctggggccaagggaacaatggTcacgTctcttcag ccaaaacacacaccccccatcagTctatccac		
2357.t4 DXP'1 J4	gcctcggacacccgccatgtattaTtgtgTgaga CATAGGgactataTtccggggagtattTTC	tgactactggggccaagggaaccctggTcacgTctctctcag ccaaaacacacaccccccatcagTctatccac		
2357.t10 DHQ52 J3	gcctcggacacccgccatgtattactgtgcgaga actggggatGAT	gcttttgatatctggggccaagggaacaatggTcacgTctcttcag ccaaaacacacaccccccatcagTctatccac		
2357.t1 D? J3	gcctcggacacccgccatgtattactgtgcgaga CATGGGTCTATG	gatatctggggccaagggaacaatggTcacgTctcttcag ctacaacacacagcccccatctgtctatccct		mouse γ3
2357.t6 DHQ52 J4	gcctcggacacccgccatgtattactgtgcgaga GAGAGCGGTcactggggaTCG	tttgactaTtgggggccaagggaaccctggTcacgTctctctcag ctacaacacacagcccccatctgtctatccct		
2357.t8 DIR2 J3	gcctcggacacccgccatgtattactgtgcgaga AGGgacccccCTGAT	gcttttgatatctggggccaagggaacaatggTcacgTctcttcag ctacaacacacagcccccatctgtctatccct		
2357.t9 DIR2R J6	gcctcggacacccgccatgtattactgtgcgaga Cgggggcct	tactactactacggtatggacgtctggggccaagggaaccacggTcacgTctctctcag ctacaacacacagcccccatctgtctatccct		
	human			mouse

FIG. 65

FIG. 66A.1

FIG. 66A.2

FIG. 66B.1

70				80				82				a				b				c				83				90																			
ACCATCTCAGCCGACAAAGTCCATCAGCACCGCCTACCTGCAGTGGAGCAGCCTGAAGGCCCTCGACACCGCCATGTATTACTGTGGGAGA																								TACTGGTACTTCGATCTCTGGGGCCCTGGGCAACCTGGTC																							
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.....																								CAGGgactggggaTCGGGAA																							
.....																								DHQ52																							
.....																								GTAaactgggATCGG																							
.....																								DHQ52																							
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71/101

VH251.GL	20	30	40	50	52 a	53	60
2357.m1	TCTCTGAAGATCTCTCTGTAAGGGTTCTGGATACAGCTTTACCAGCTACTGGATCGGCTGGGTGGTGGCCAGATGCCCCGGGAAGGCCCTGGAGTGGATGGGGATCATCTATCCTGGTGACTCTGATACCAGATACAGCCCGTCTTCC	CAAGGCCAGGTC					
2357.m2							
2357.m5							
2357.m6							
2357.m7							
2357.m12							
2357.m13							
2357.m14							
2357.m16							
2357.m17							
2357.m18							
2357.m19							
2357.m20							
2357.m22							
2357.m26							
2357.m27							
2357.m28							
2357.m29							
2357.m30							
2357.m31							
2357.m32							
2357.m33							
2357.m35							
2357.m36							

CDR II

CDR I

FIG. 67A

72/101

[illegible]

CDR I

COR II

FIG. 67B.1

FIG. 67B.2

74/101

	20	30	40	50	52 a	53	60
VH5P1.GL	TCCCTGAGACTCTCTCTGTGAGCCCTCTGGATTCACTTCAGTAGCTATGCTATGCACCTGGGTCCGCCAGGCTCCAGGCAAGGGGCTGGAGTGGGTGGAGTGGGTATATATATATGATGGAAGCAATAAATACTACGCAGACTCCGTGAAGGGCCGATTCC						
J2							
5250.ln4							
5250.ln5							
5250.ln8		A.A.G.A...A.					
5250.ln10		A.A.G.A...A.					
5250.ln12							
5250.ln27		A.A.G.A.C.A.					
5250.ln115							
5250.ln118		A...A.					
5250.ln202		A.A.G.A...A.					
J4							
5250.ln16							
5250.ln19							
5250.ln22							
5250.sp26							
5250.sp27							
J6							
5250.ln3							
5250.ln24							
5250.sp19							
5250.sp22							
5250.sp28							
VH51P1.GL	TCCGTGAAGGTCTCCTGCAAGGCTTCTGGAGGCACCTTCAGCAGCTATGCTATCAGCTGGTGGACAGGCCCTGGACAAGGGCTTGAGTGGATGGGAAGGATCATCCCTATCTCTGGTATAGCAAACTACGCACAGAAGTTCCAGGGCAGAGTC						
J3							
5250.sp33							
VH4.21.GL	ACCTGTCCCTCACCCTGCGCTGCTATGGTGGTCTTCAGTGGTTACTACTGGAGCTGGATCCGCCAGGCCCCAGGGAAGGGGCTGGAGTGGATTGGGGAATCAATCAT						
J4							
5250.ln2							
5250.sp30							
5250.sp32							

CDR I

CDR II

FIG. 68A

CDR III

FIG. 68B

76/101

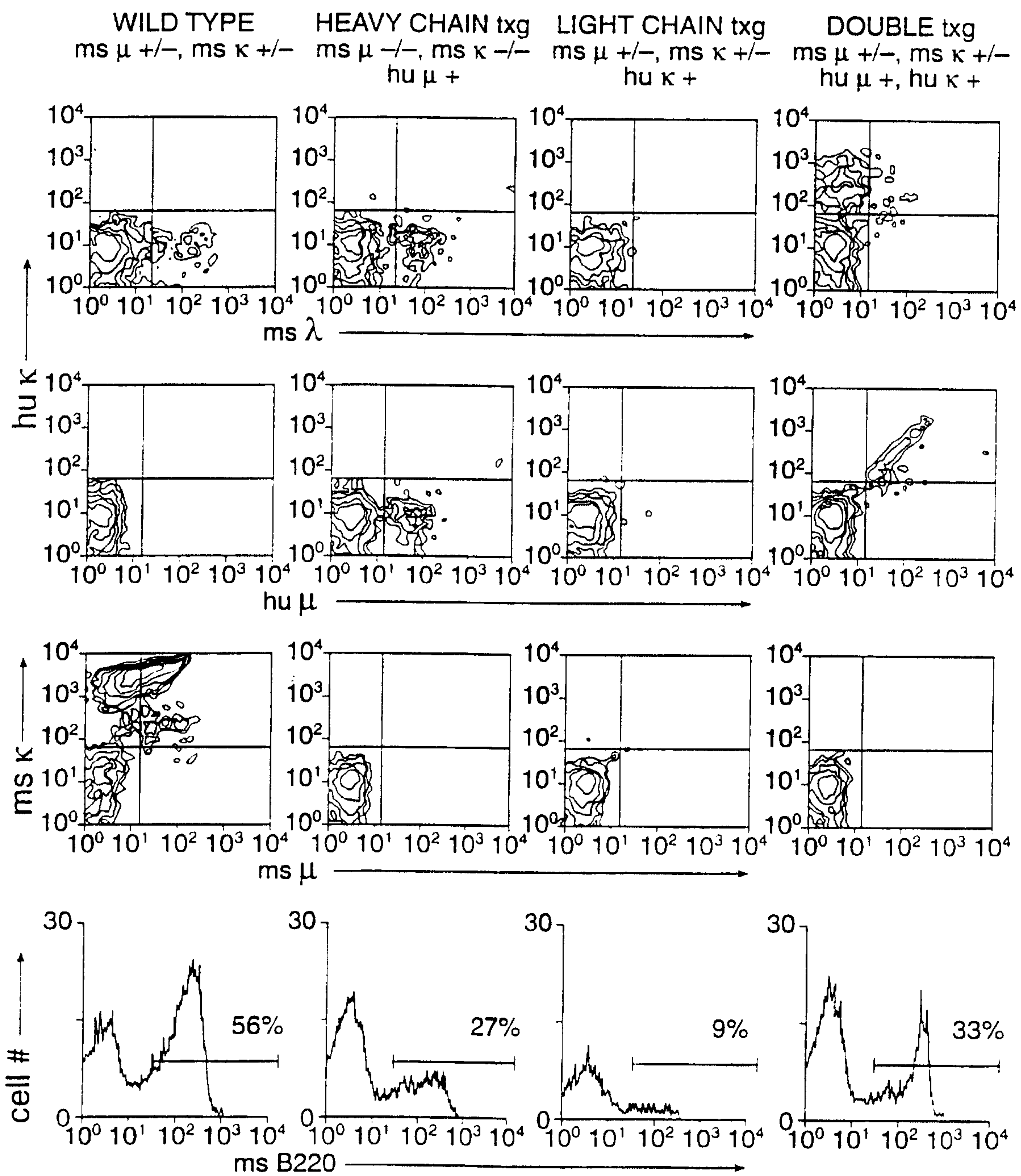


FIG. 69

77/101

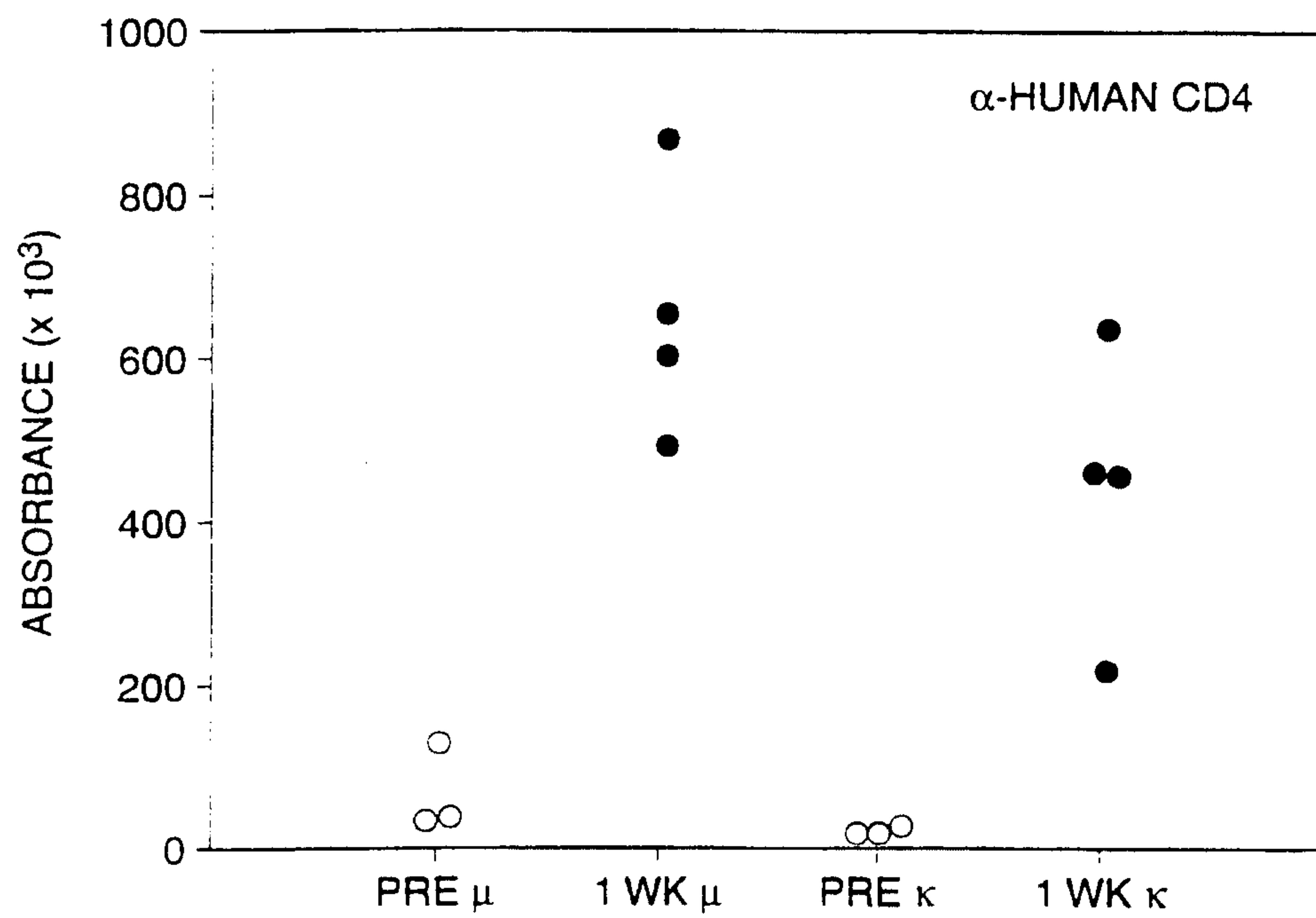


FIG. 71A

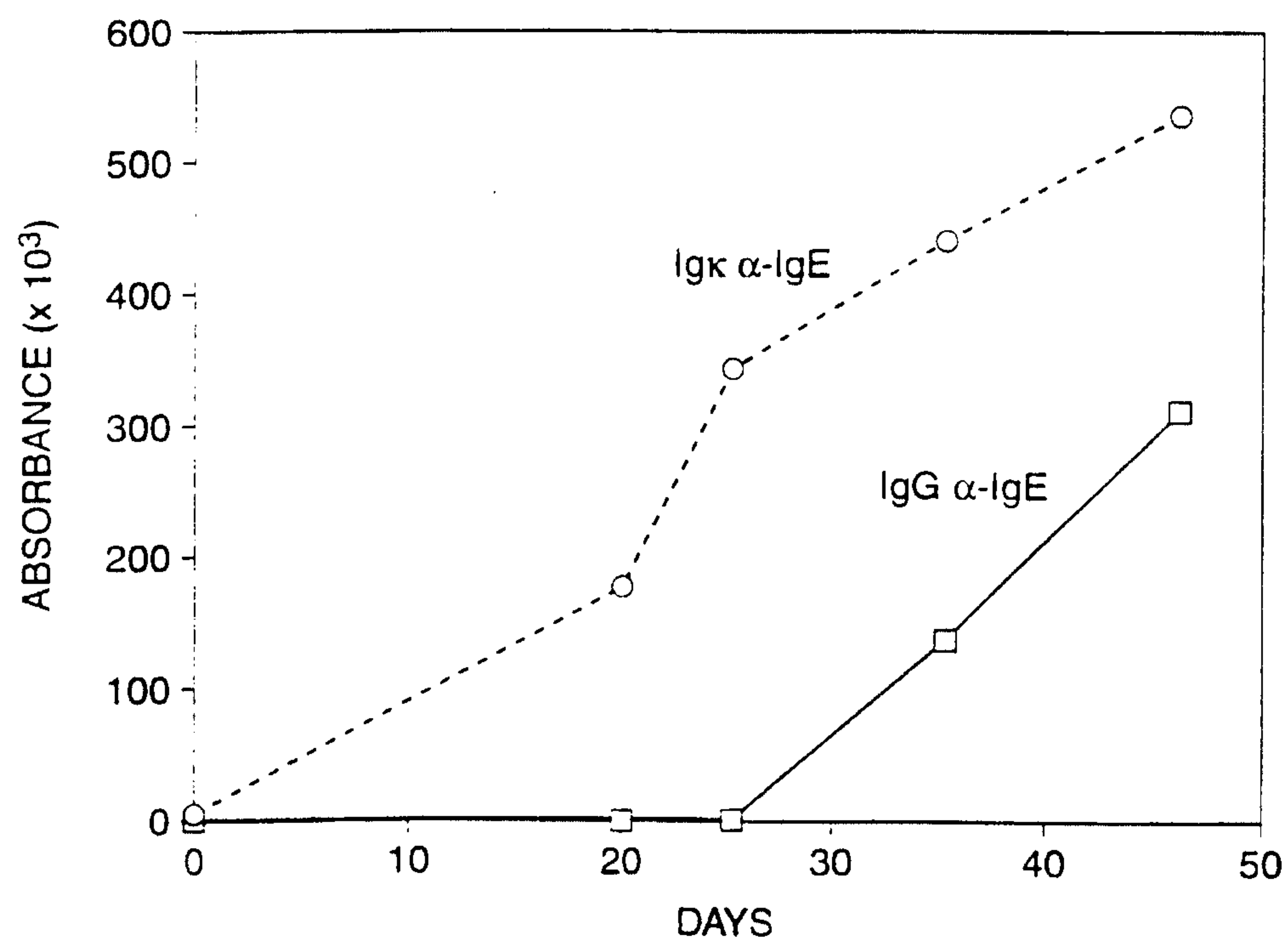


FIG. 71B

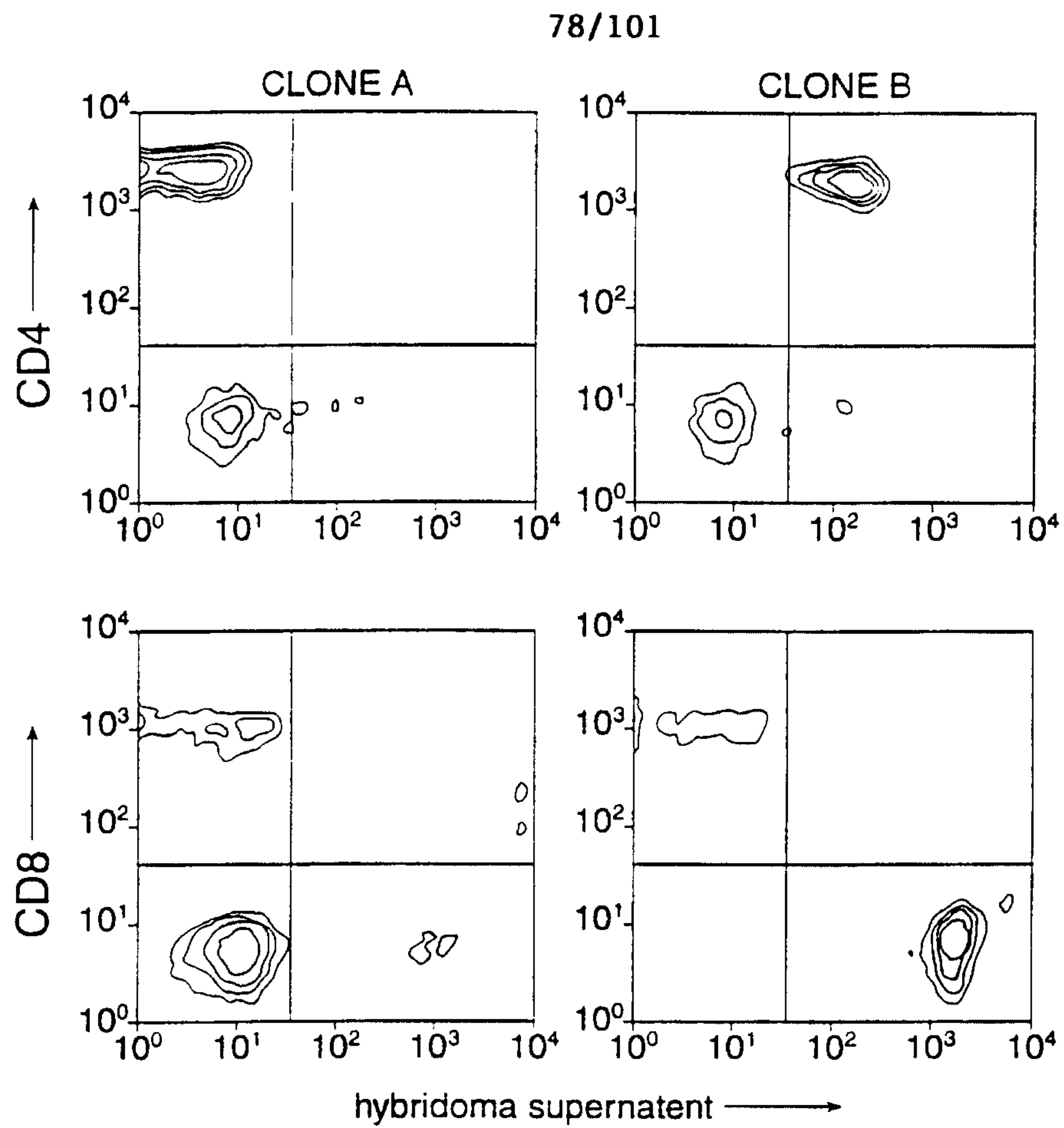


FIG. 72

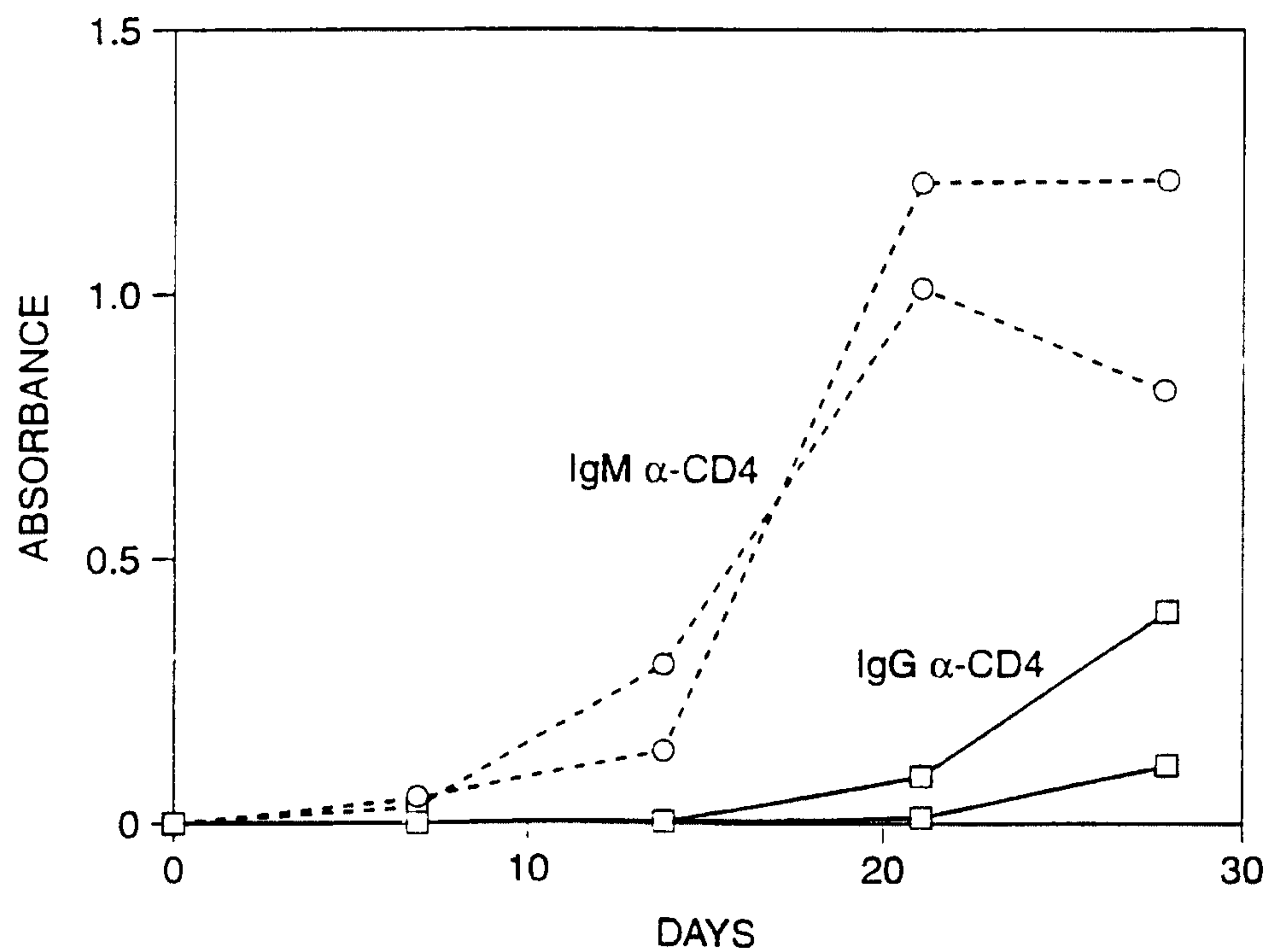


FIG. 73

79/101

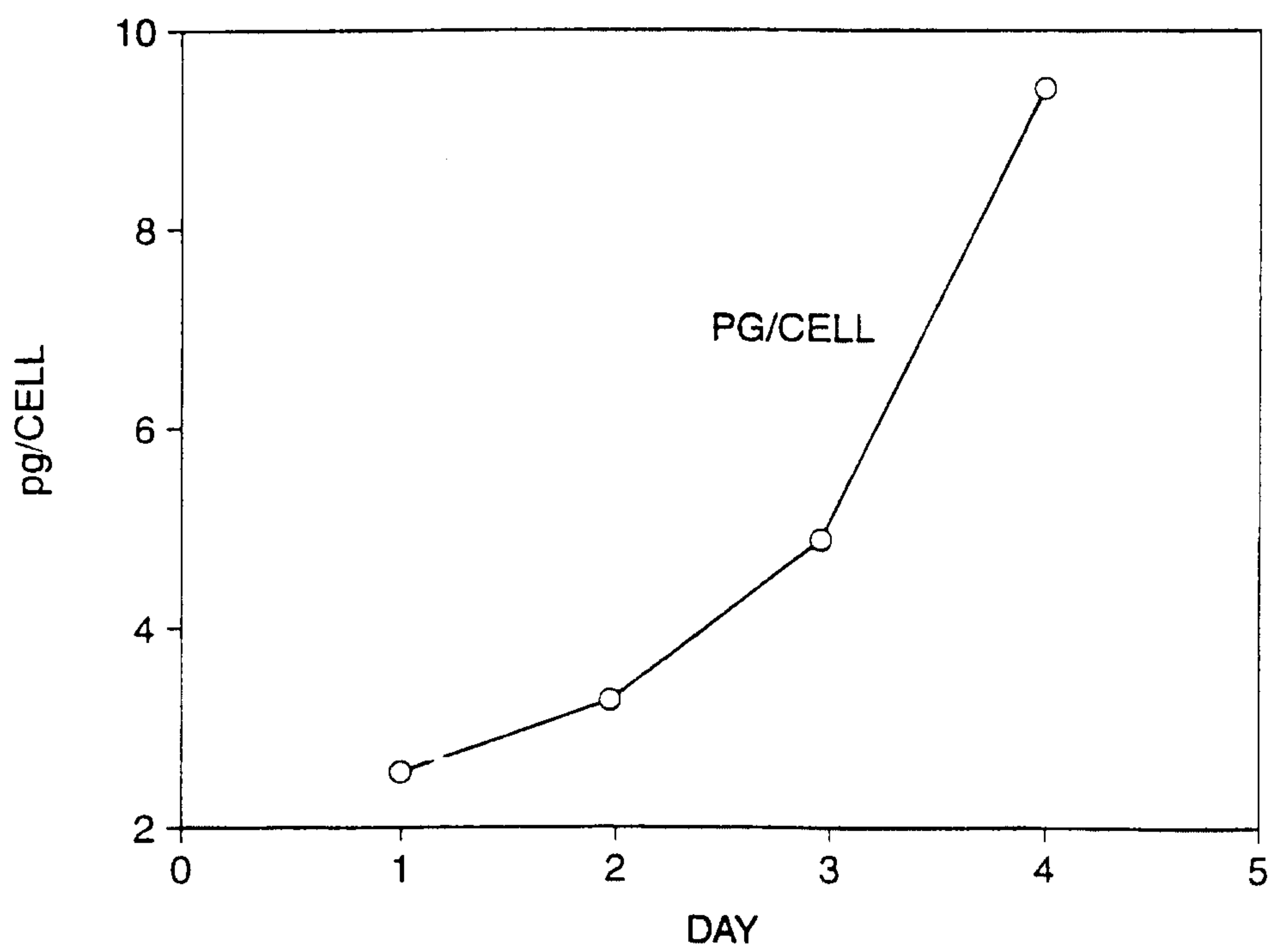
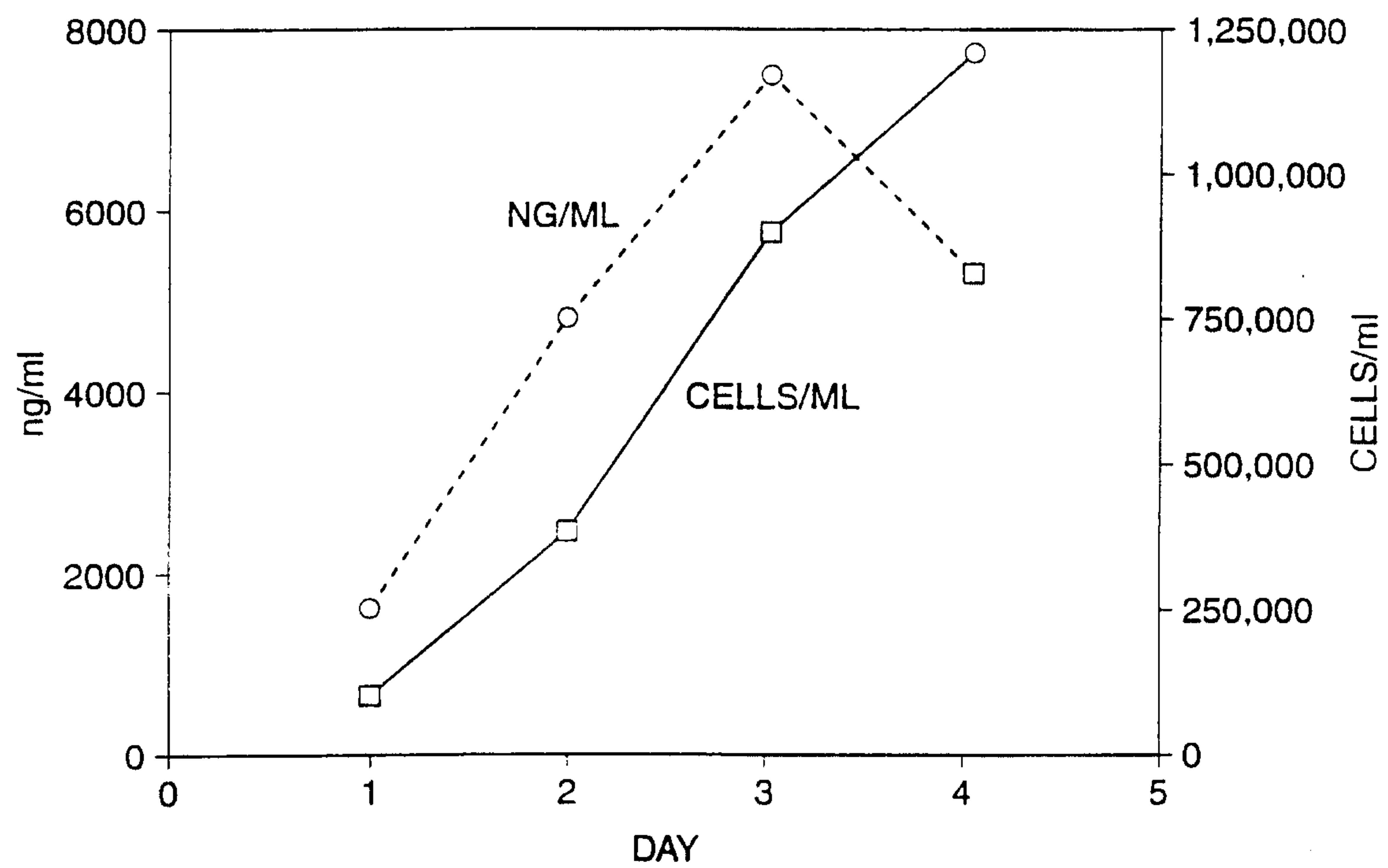
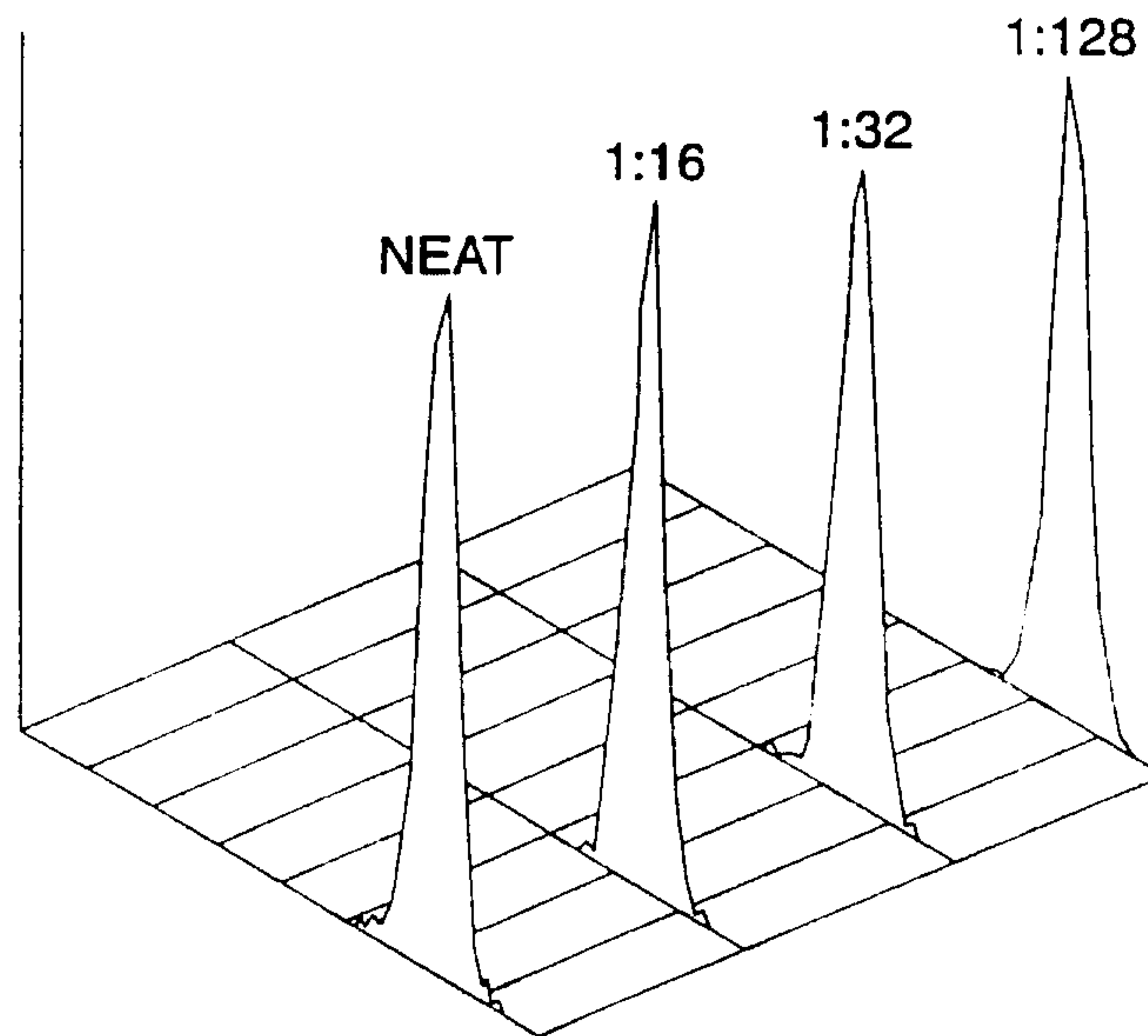


FIG. 74

80/101

RPA-T4/2C11-8

#12:BDPHARMCOMP004\FL2-H\FL2-HEIGHT



Leu-3a/2C11-8

#12:BDPHARMCOMP025\FL2-H\FL2-HEIGHT

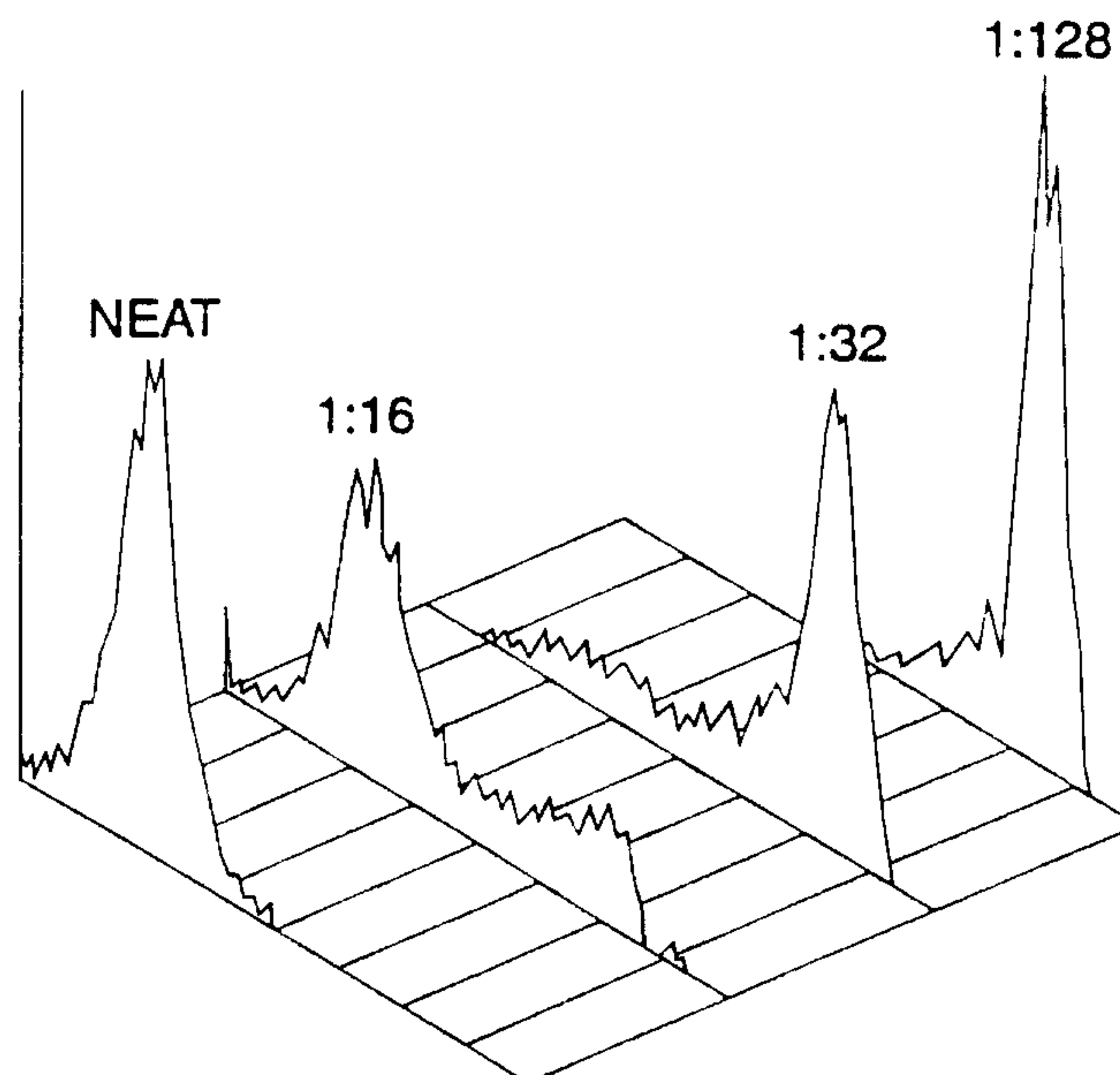


FIG. 75

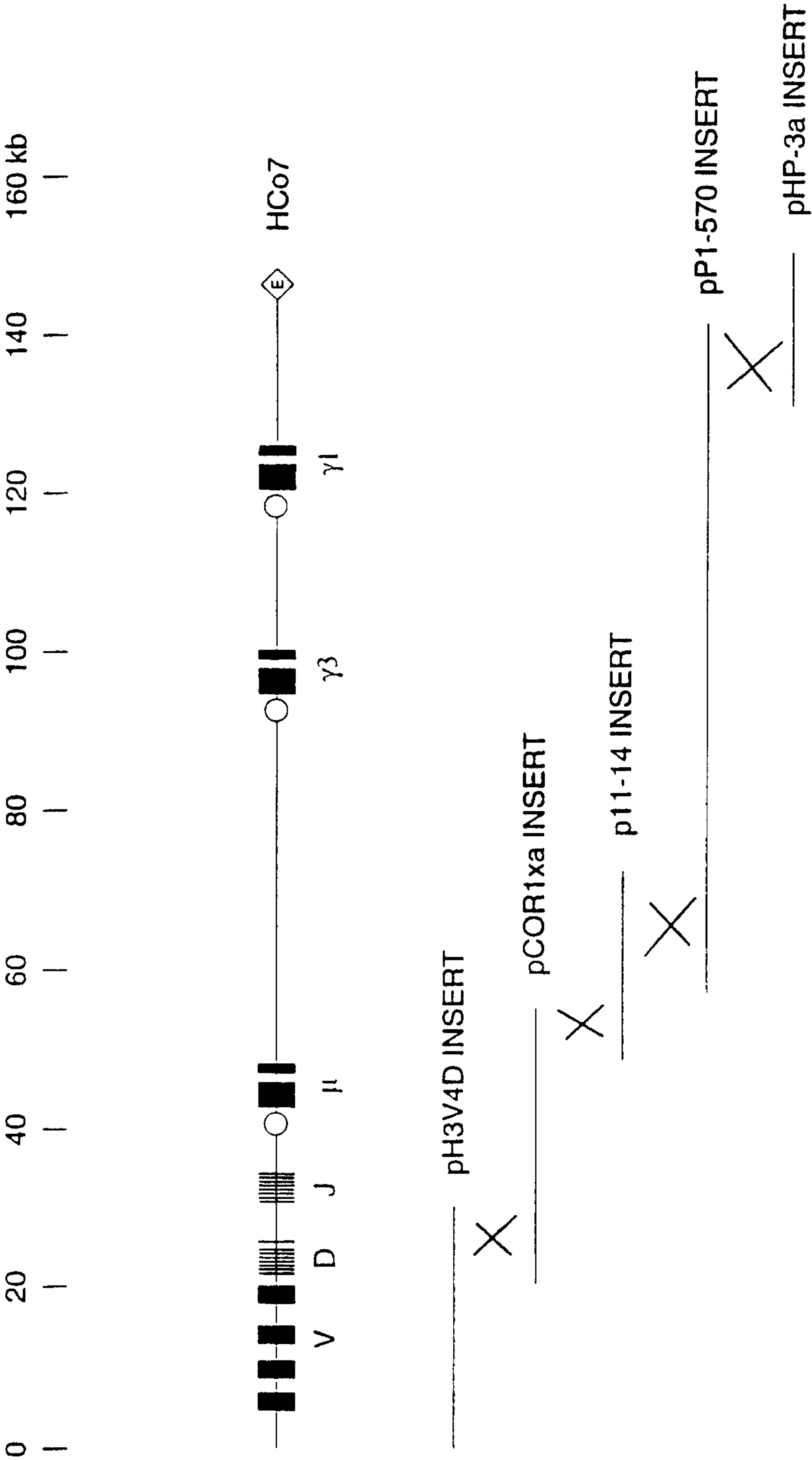


FIG. 76

pGP2b sequence:

AATTAGCggccgctgtcgacaagcttcgaattcagtatcgatgtggtacctggatcctcgagtgcGGCCGCAGTATGCAA
AAAAAGCCCCGCTCATTAGGCGGGCTCTTGGCAGAACATATCCATCGCGTCCGCCATCTCCAGCAGCCGCACGCGGCGCA
TCTCGGGCAGCGTTGGGTCTGGCCACGGGTGCGCATGATCGTGCTCCTGTGCTTGAGGACCCGGCTAGGCTGGCGGGGT
TGCCTTACTGGTTAGCAGAATGAATCACCGATACGCGAGCGAACGTGAAGCGACTGCTGCTGCAAAACGTCTGCGACCTG
AGCAACAACATGAATGGTCTTCGGTTTCCGTGTTTCGTAAAGTCTGGAAACGCGGAAGTCAGCGCCCTGCACCATTATGT
TCCGGATCTGCATCGCAGGATGCTGCTGGCTACCCTGTGGAACACCTACATCTGTATTAACGAAGCGCTGGCATTGACCC
TGAGTGATTTTTCTCTGGTCCCGCCGCATCCATACCGCCAGTTGTTTACCCTCACAACGTTCCAGTAACCGGGCATGTTT
ATCATCAGTAACCCGTATCGTGAGCATCCTCTCTCGTTTCATCGGTATCATTACCCCCATGAACAGAAATTCCCCCTTAC
ACGGAGGCATCAAGTGACCAAACAGGAAAAACCGCCCTTAACATGGCCCCGCTTTATCAGAAGCCAGACATTAACGCTTC
TGGAGAACTCAACGAGCTGGACGCGGATGAACAGGCAGACATCTGTGAATCGCTTCACGACCACGCTGATGAGCTTTAC
CGCAGCTGCCTCGCGCGTTTCGGTGATGACGGTGAAAACCTCTGACACATGCAGCTCCCGGAGACGGTCACAGCTTGTCT
GTAAGCGGATGCCGGGAGCAGACAAGCCCGTCAGGGCGCGTCAGCGGTGTTGGCGGGTGTGCGGGGCGCAGCCATGACCC
AGTCACGTAGCGATAGCGGAGTGTATACTGGCTTAACATGCGGCATCAGAGCAGATTGTAAGTGCAGAGTGCACCATATGC
GGTGTGAAATACCGCACAGATGCGTAAGGAGAAAATACCGCATCAGGCGCTCTTCCGCTTCCTCGCTCACTGACTCGCTG
CGCTCGGTCTGTTCCGGCTGCGGCGAGCGGTATCAGCTCACTCAAAGGCGGTAATACGGTTATCCACAGAATCAGGGGATAA
CGCAGGAAAGAACATGTGAGCAAAAGGCCAGCAAAAGGCCAGGAACCGTAAAAAGGCCGCGTTGCTGGCGTTTTTCCATA
GGCTCCGCCCCCTGACGAGCATCAGAAAAATCGACGCTCAAGTCAGAGGTGGCGAAACCCGACAGGACTATAAAGATAC
CAGGCGTTTTCCCCCTGGAAGCTCCCTCGTGCGCTCTCCTGTTCCGACCCTGCCGCTTACCGGATACCTGTCCGCCTTTCT
CCCTTCGGGAAGCGTGGCGCTTTCTCATAGCTCACGCTGTAGGTATCTCAGTTCGGTGATAGGTGCTTCGCTCCAAGCTGG
GCTGTGTGCACGAACCCCCGTTTACGCCCCAGCGCTGCGCCTTATCCGGTAACATCTGCTTTGAGTCCAACCCGTAAGA
CACGACTTATCGCCACTGGCAGCAGCCAggcgcgcccttggcctaagaggccaCTGGTAACAGGATTAGCAGAGCGAGGTA
TGTAGGCGGTGCTACAGAGTTCTTGAAGTGGTGGCCTAACTACGGCTACACTAGAAGGACAGTATTTGGTATCTGCGCTC
TGCTGAAGCCAGTTACCTTCGGAAAAAGAGTTGGTAGCTCTTGATCCGGCAAACAAACCACCGCTGGTAGCGGTGGTTTT
TTTGTGTGCAAGCAGCAGATTACGCGCAGAAAAAAGGATCTCAAGAAGATCCTTTGATCTTTTCTACGGGGTCTGACGC
TCAGTGGAAACGAAACTCACGTTAAGGGATTTTGGTCATGAGATTATCAAAAAGGATCTTCACCTAGATCCTTTTAAATT
AAAAATGAAGTTTTAAATCAATCTAAAGTATATATGAGTAACTTGGTCTGACAGTTACCAATGCTTAATCAGTGAGGCA
CCTATCTCAGCGATCTGTCTATTTCTGTTTCATCCATAGTTGCCTGACTCCCCGTCGTGTAGATAACTACGATACGGGAGGG
CTTACCATCTGGCCCCAGTGCTGCAATGATACCGCGAGACCCACGCTCACCGGCTCCAGATTTATCAGCAATAAACCAGC
CAGCCGGAAGGGCCGAGCGCAGAAGTGGTCTGCAACTTTATCCGCCTCCATCCAGTCTATTAATTGTTGCCGGGAAGCT
AGAGTAAGTAGTTCCGCCAGTTAATAGTTTCCGCAACGTTGTTGCCATTGCTGCAGGCATCGTGGTGTACGCTCGTCTGTT
TGGTATGGCTTCATTACGCTCCGGTTCCCAACGATCAAGGCGAGTTACATGATCCCCATGTTGTGCAAAAAAGCGGTTA
GCTCCTTCGGTCTCCTCCGATCGTTGTGAGAAGTAAGTTGGCCGCGAGTGTATCACTCATGGTTATGGCAGCACTGCATAAT
TCTCTTACTGTCATGCCATCCGTAAGATGCTTTTCTGTGACTGGTGAGTACTCAACCAAGTCATTCTGAGAATAGTGTAT
GCGGCGACCGAGTTGCTCTTGCCCGGCGTCAACACGGGATAATACCGCGCCACATAGCAGAACTTTAAAAGTGCTCATCA
TTGGAAAACGTTCTTCGGGGCGAAAACCTCTCAAGGATCTTACCGCTGTTGAGATCCAGTTCGATGTAACCCACTCGTGCA
CCCAACTGATCTTCAGCATCTTTTACTTTTACCAGCGTTTCTGGGTGAGCAAAAACAGGAAGGCAAAATGCCGCAAAAAA
GGGAATAAGGGCGACACGGAAATGTTGAATACTCATACTCTTCCTTTTTCAATATTATTGAAGCATTATCAGGGTTATT
GTCTCATGAGCGGATACATATTTGAATGTATTTAGAAAAATAACAAATAGGGGTTCCGCGCACATTTCCCCGAAAAGTG
CCACCTGACGTCTAAGAAACCATTATTATCATGACATTAACCTATAAAAAATAGGCGTATCACGAGGCCCTTTCGTCTTCA
AG

FIG. 77A

83/101

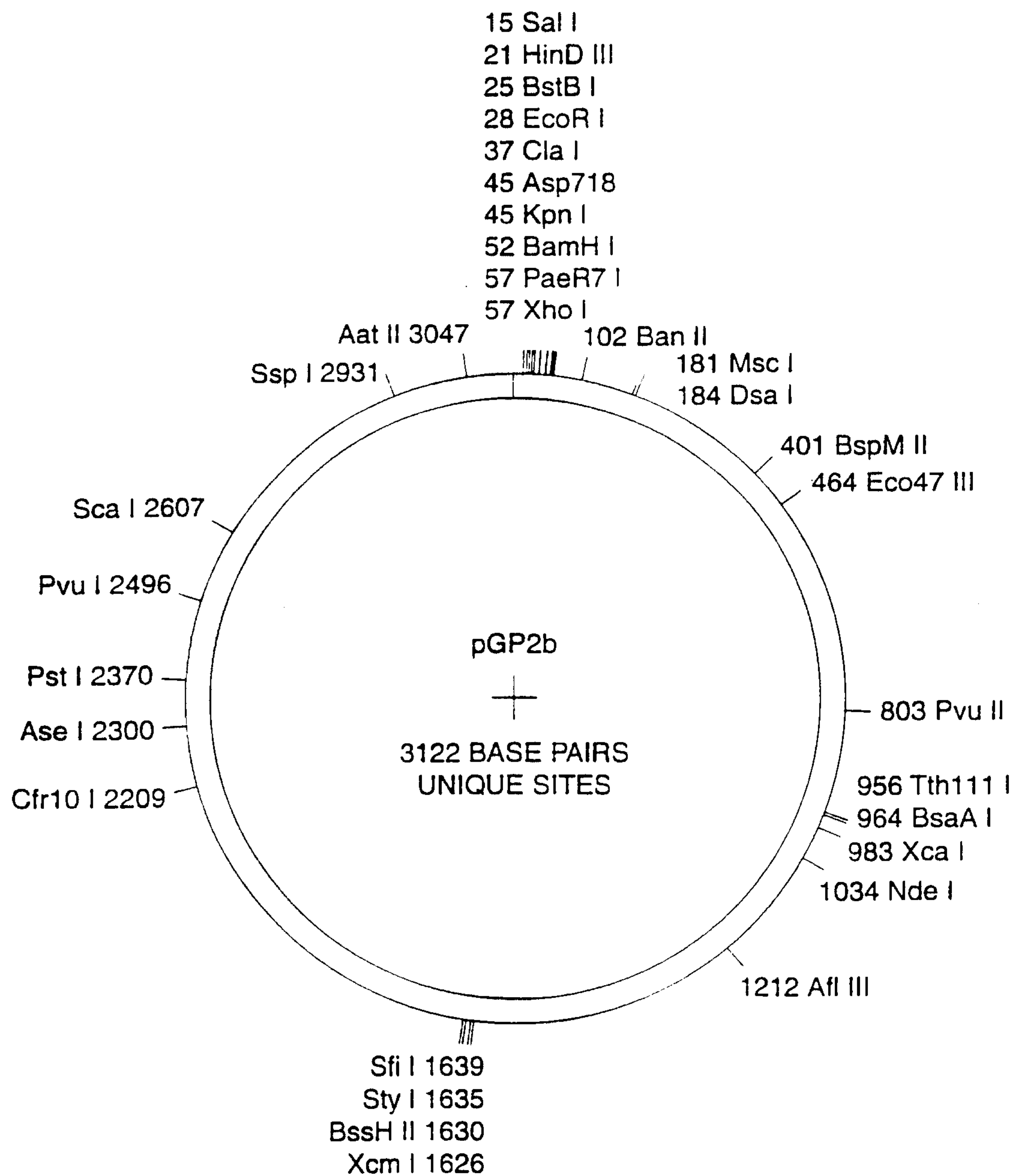


FIG. 77B

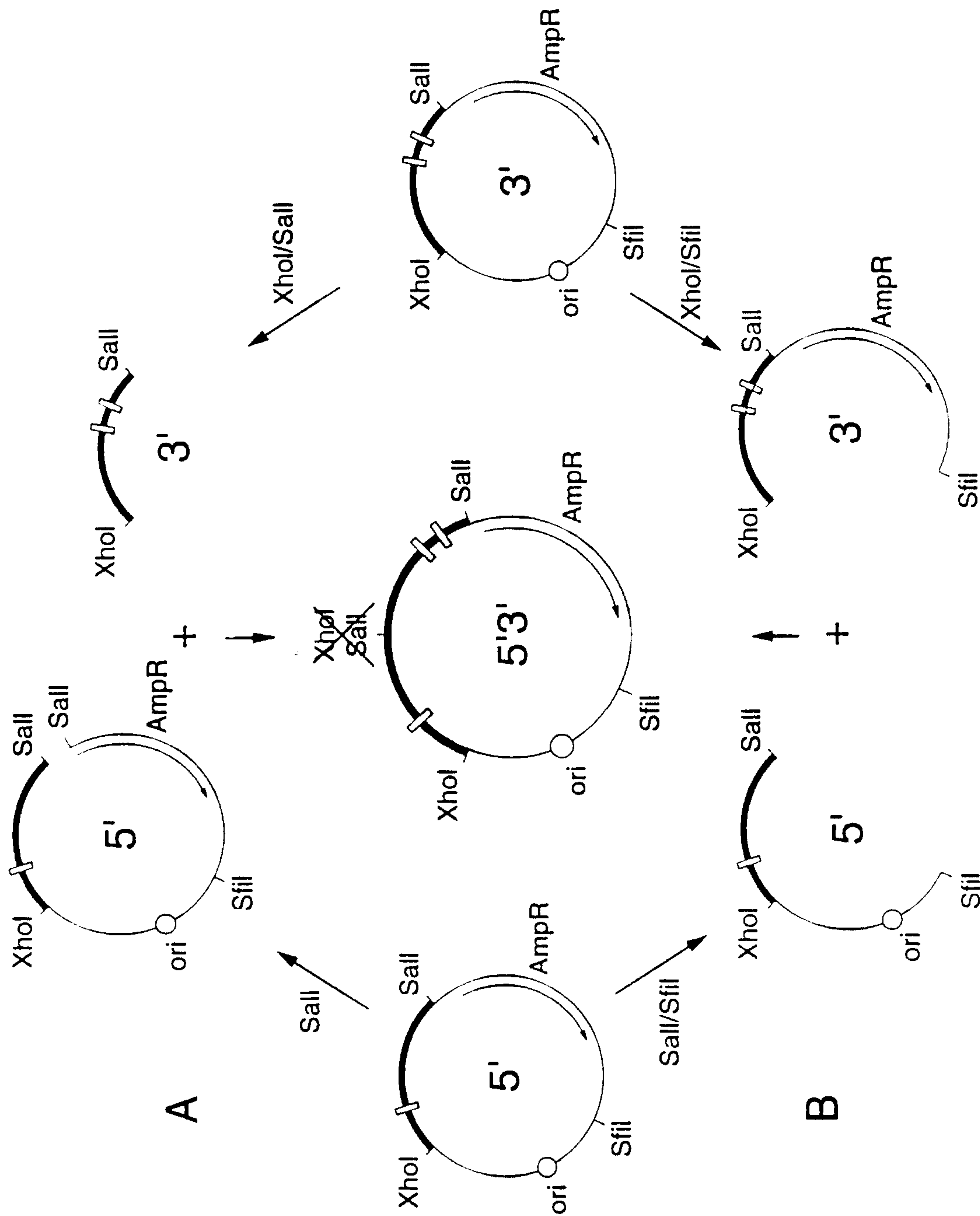


FIG. 78

85/101

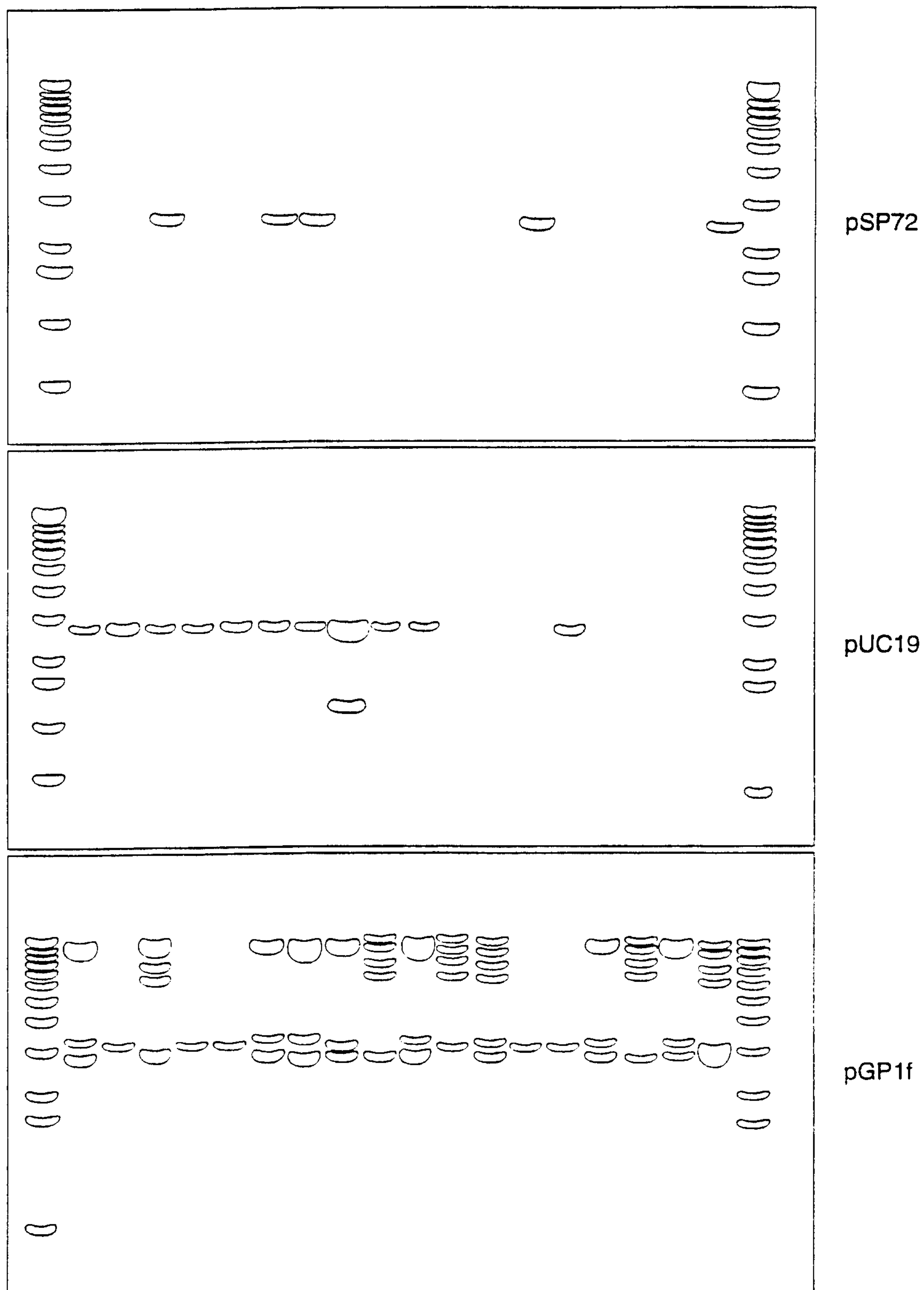


FIG. 79

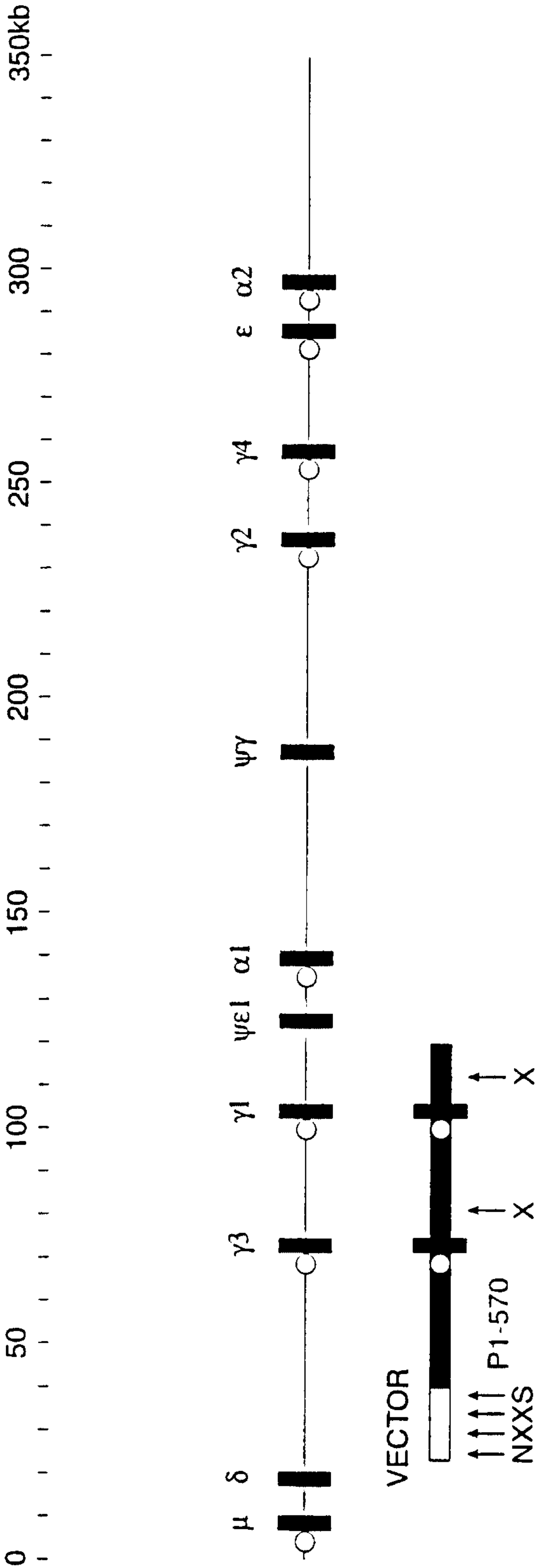


FIG. 80

87/101

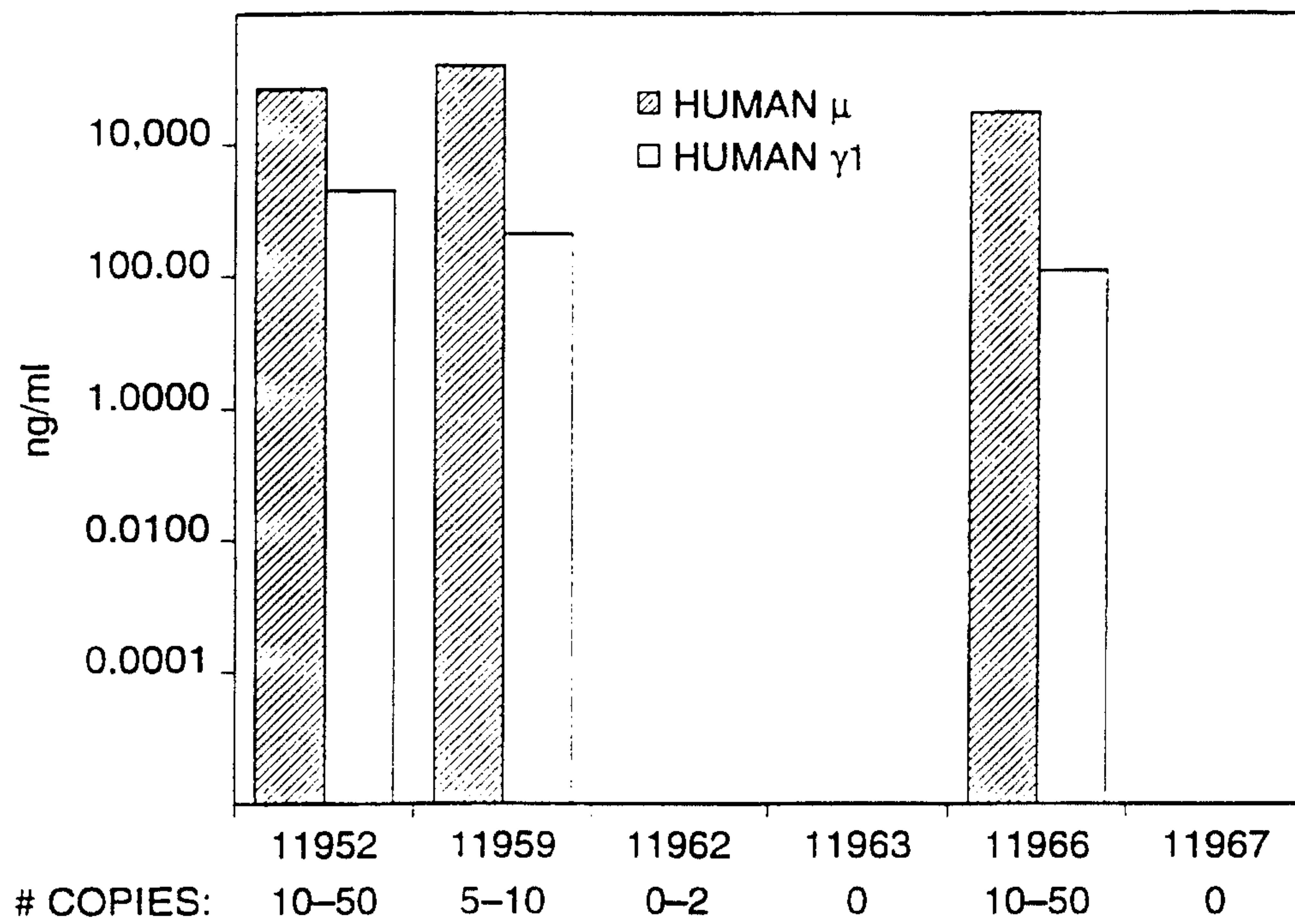


FIG. 81

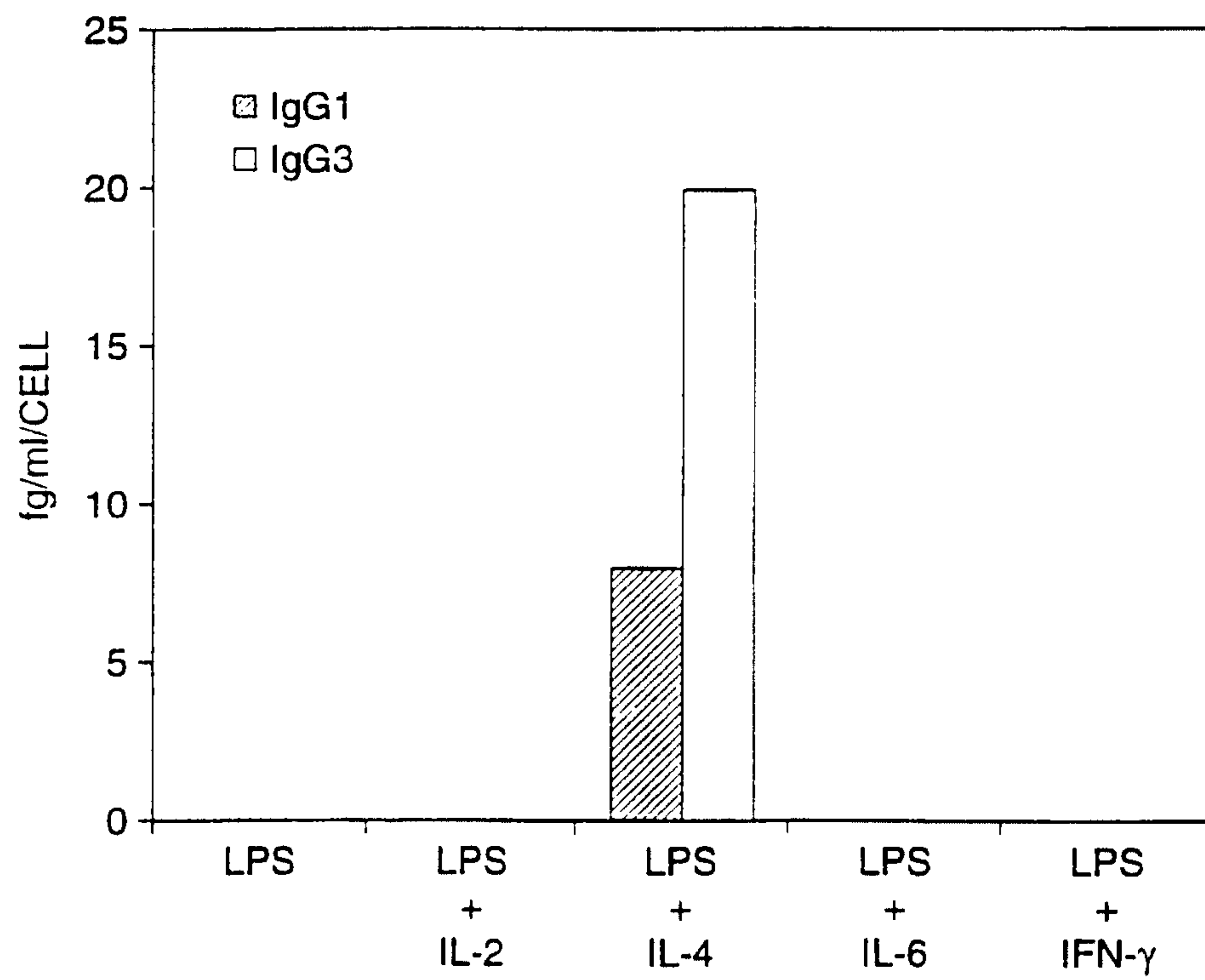


FIG. 84

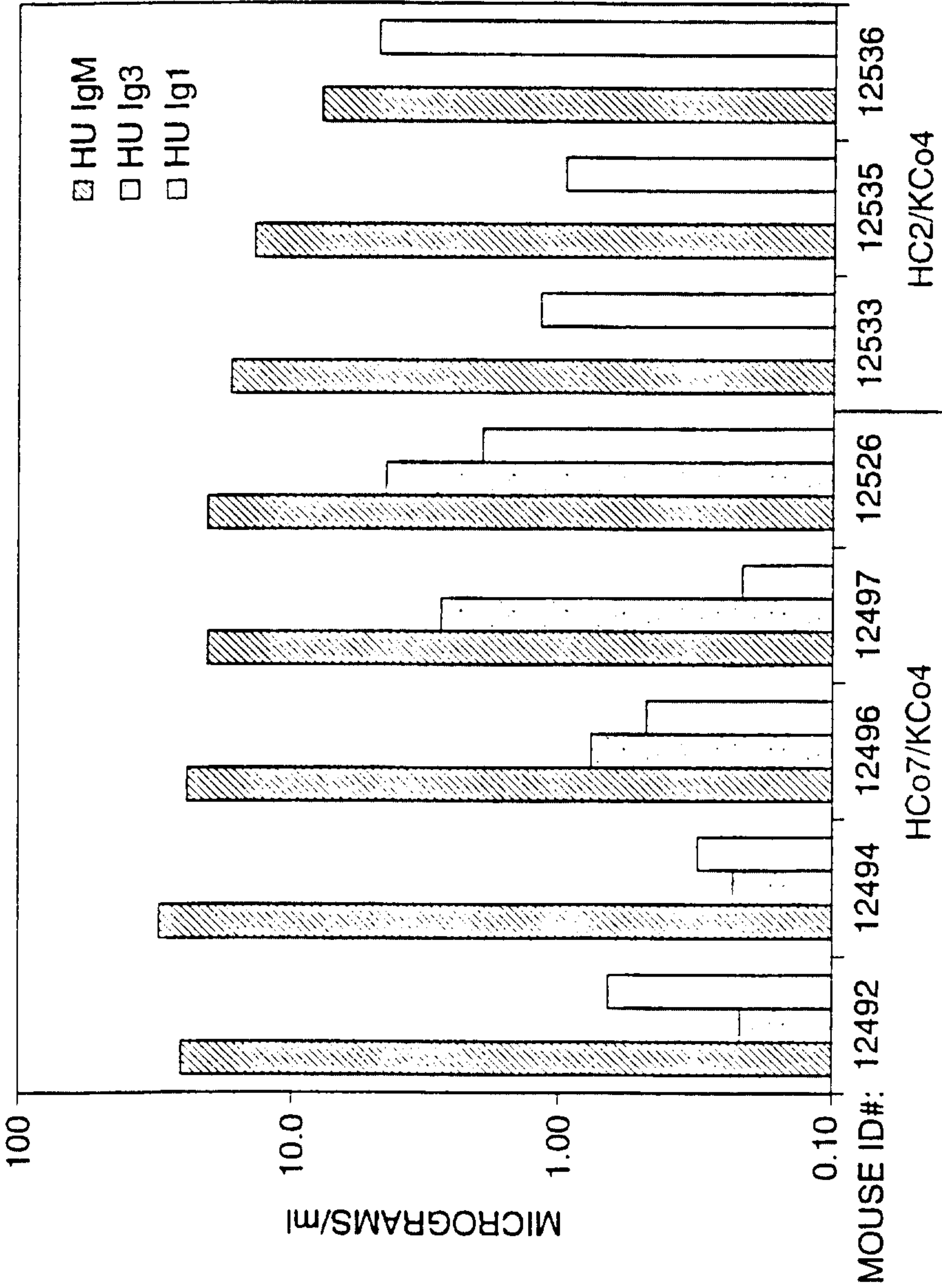


FIG. 82

89/101

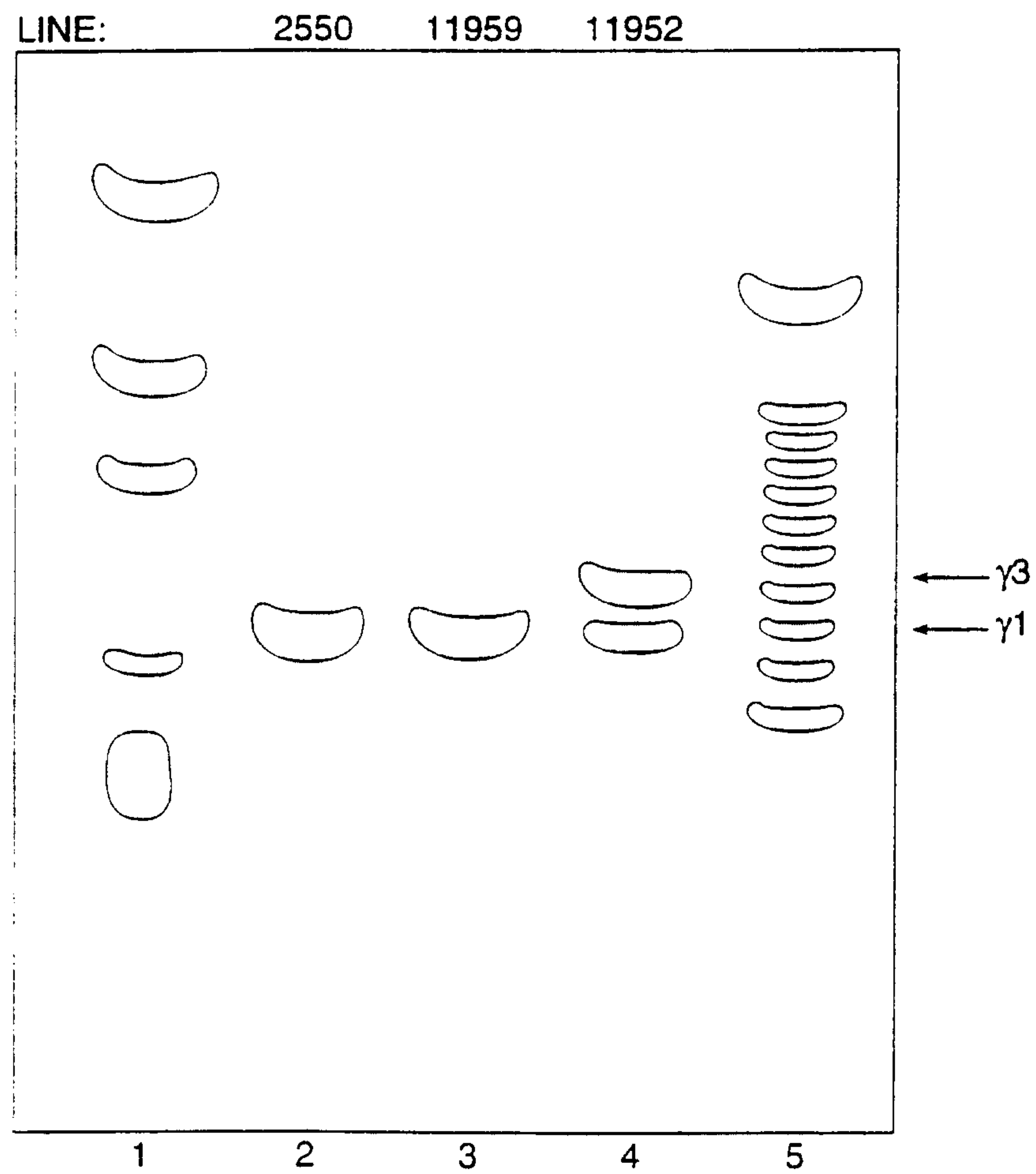


FIG. 83

90/101

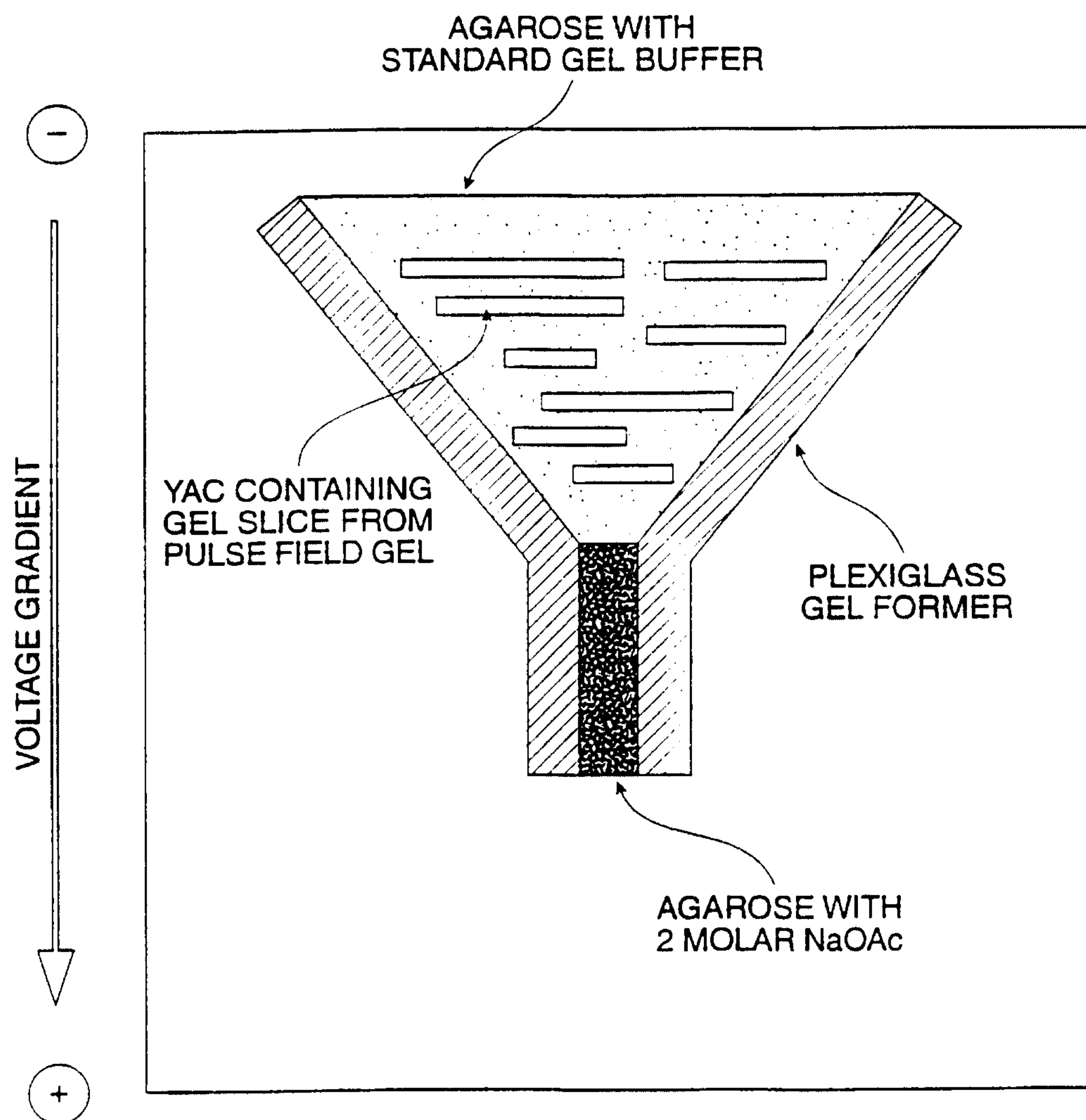


FIG. 85

91/101

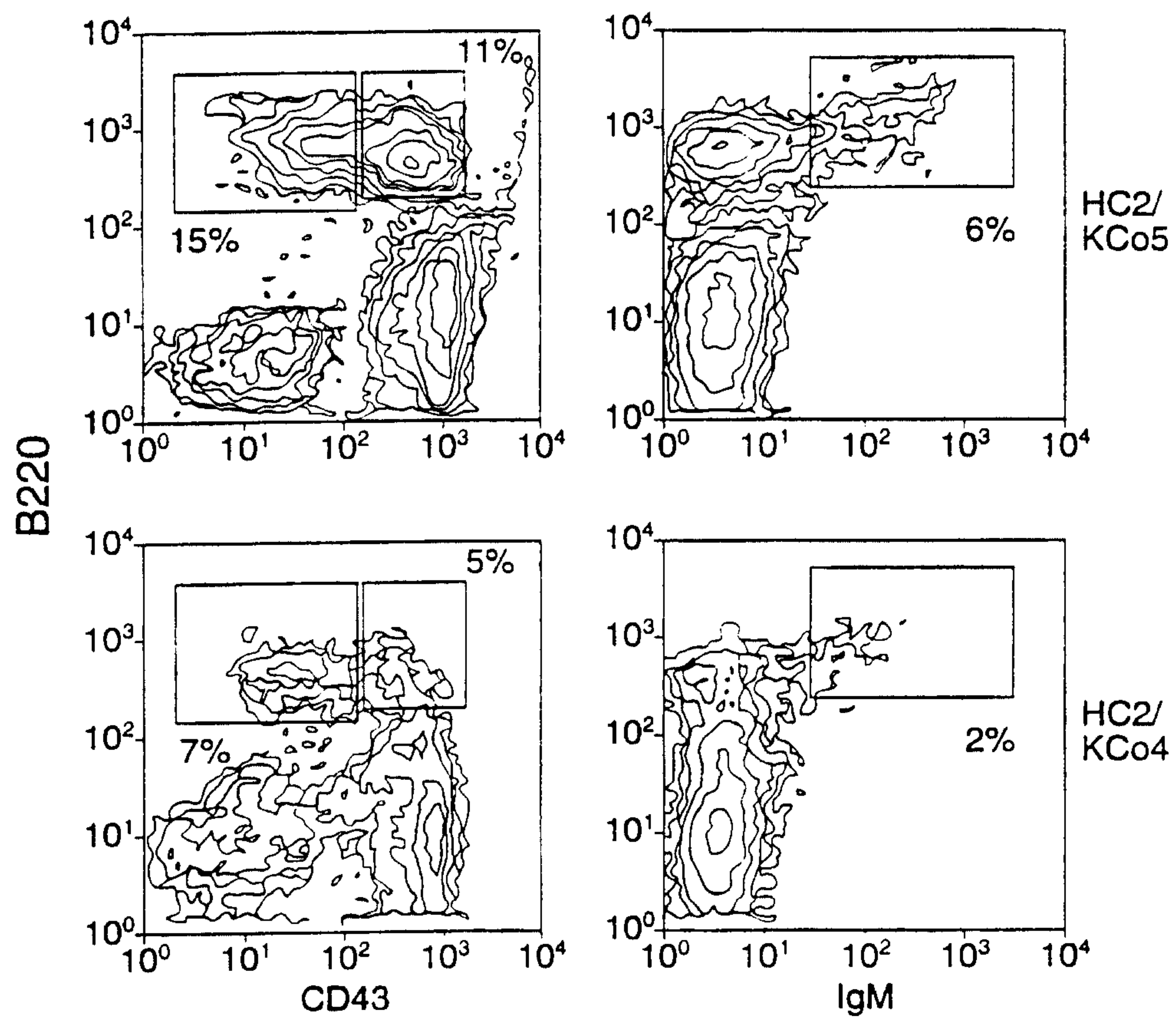


FIG. 86

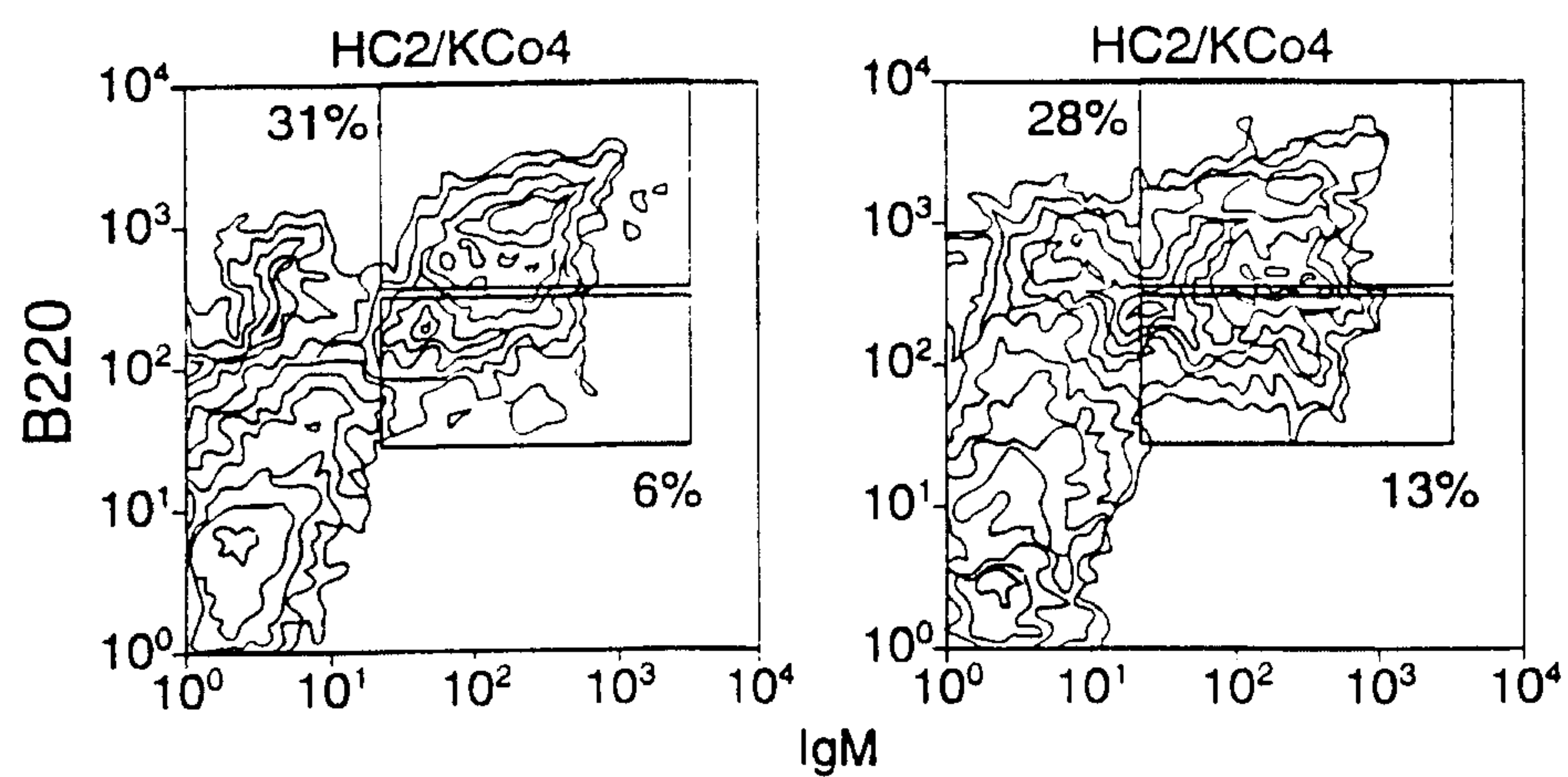


FIG. 87

92/101

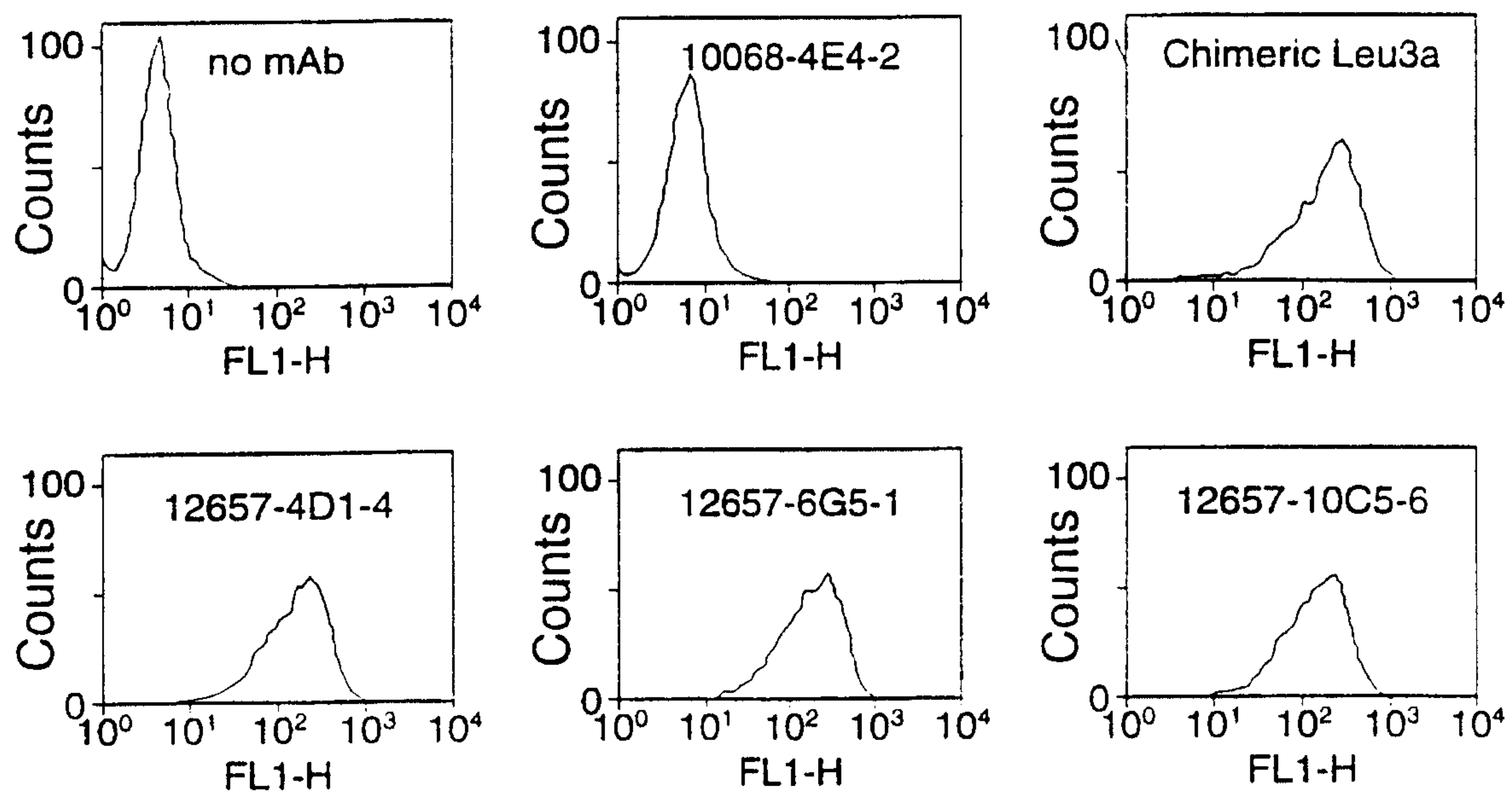


FIG. 88

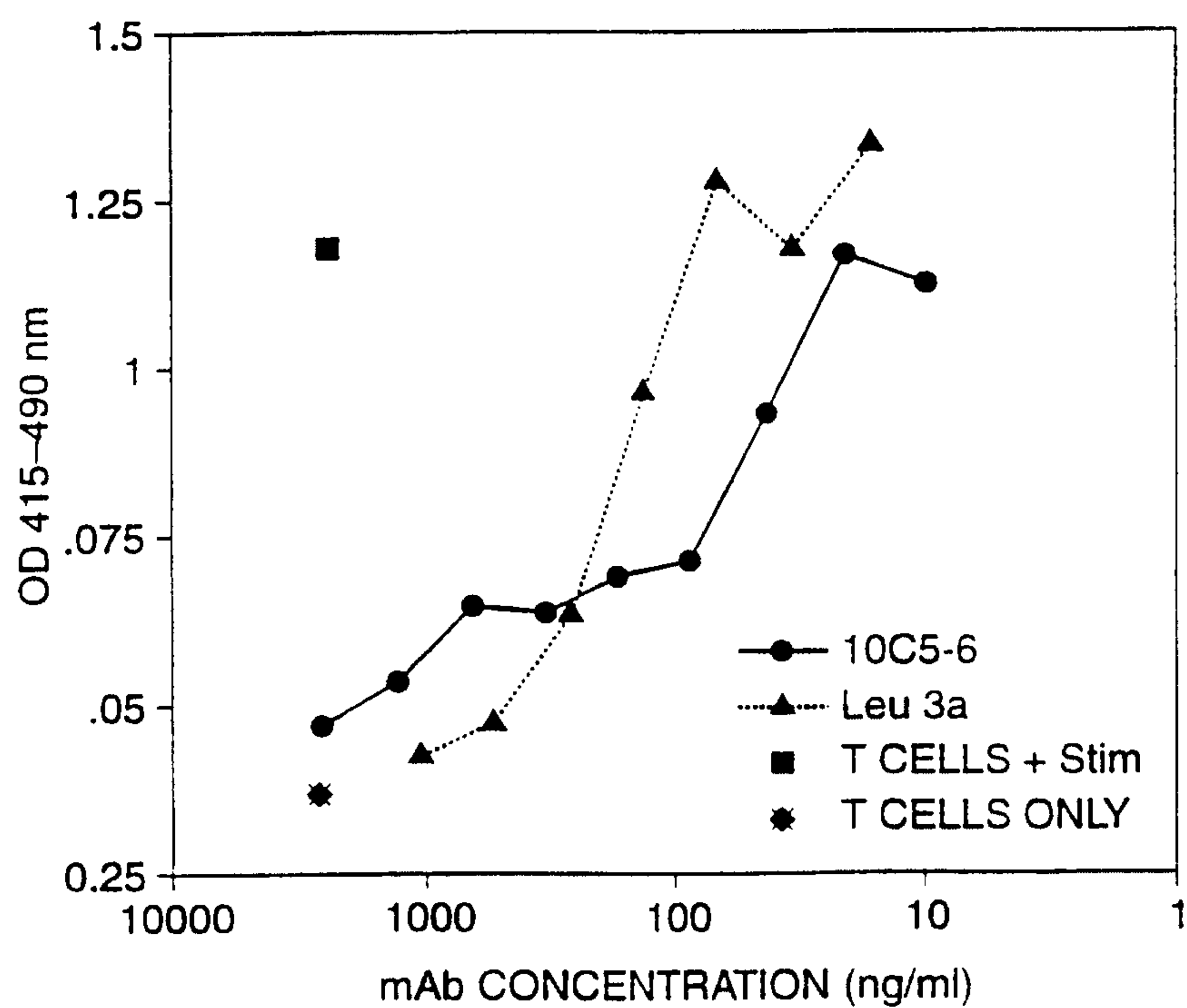


FIG. 90

93/101

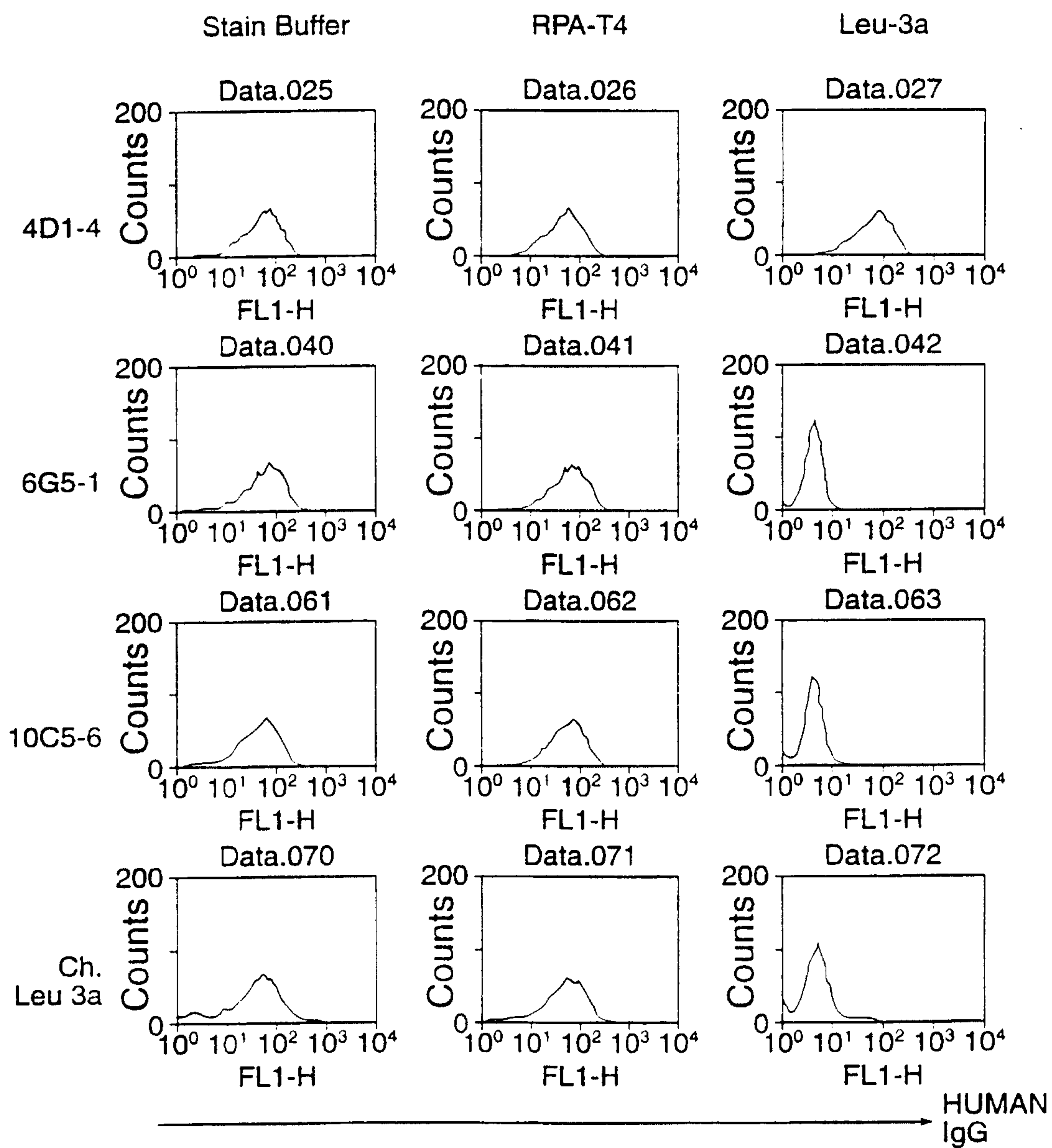
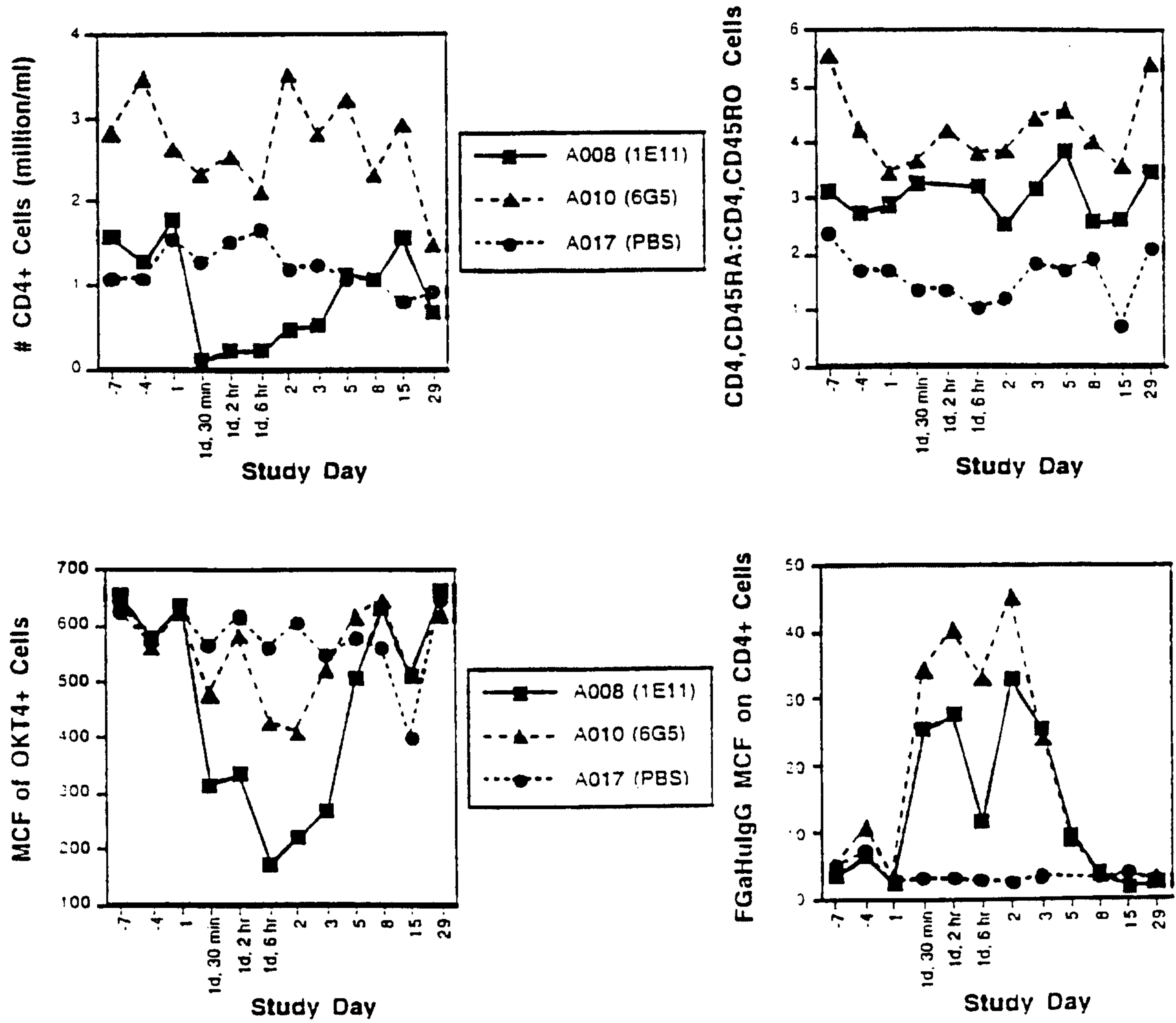


FIG. 89

WO 98/24884

PCT/US97/21803

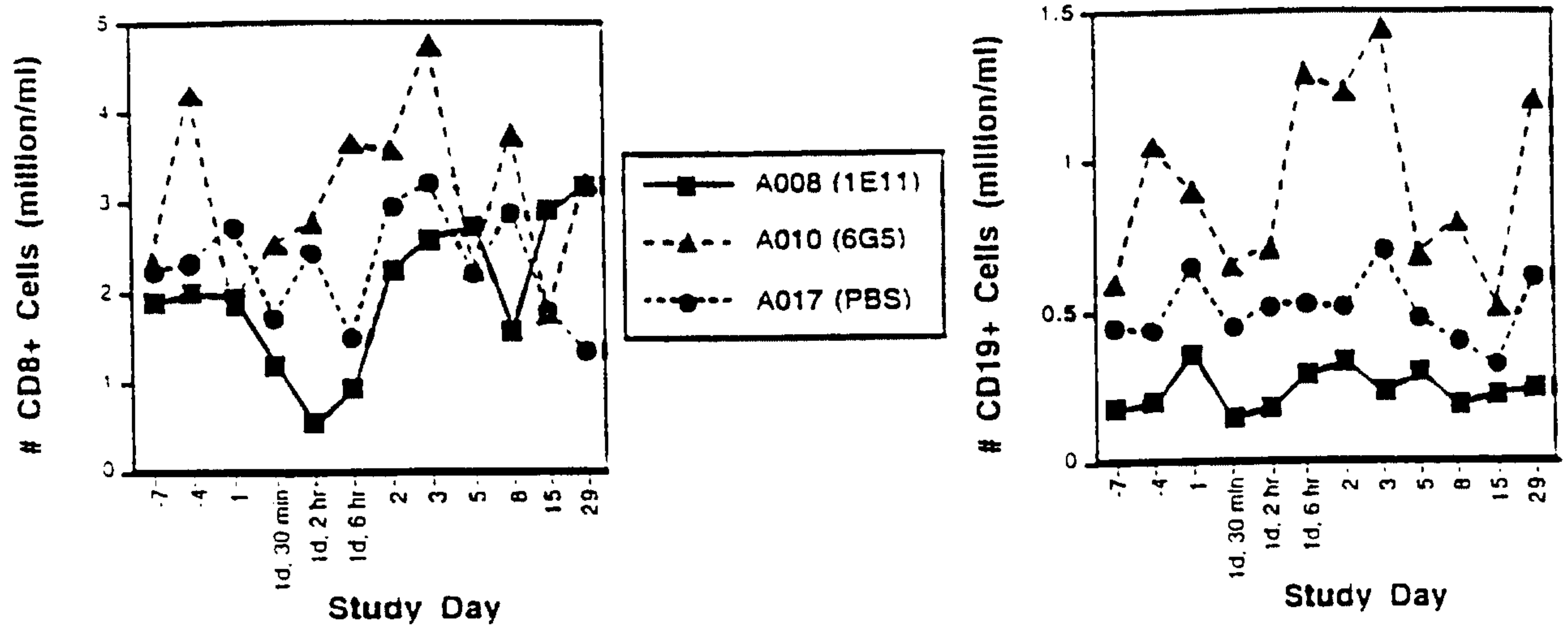
Figure 91 94/101



WO 98/24884

PCT/US97/21803

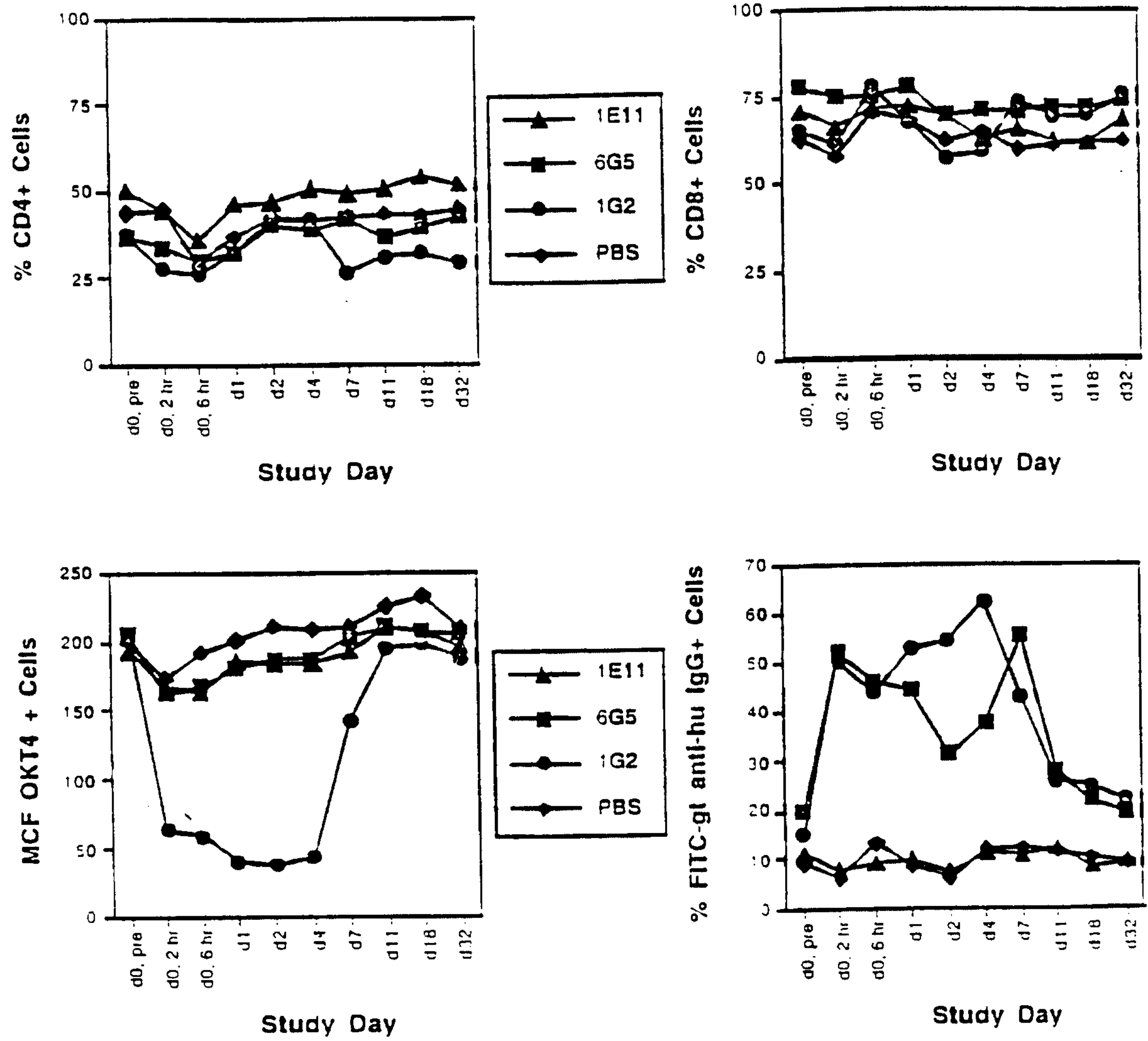
Figure 92 95/101



WO 98/24884

PCT/US97/21803

Figure 93 96/101



WO 98/24884

PCT/US97/21803

Figure 94

97/101

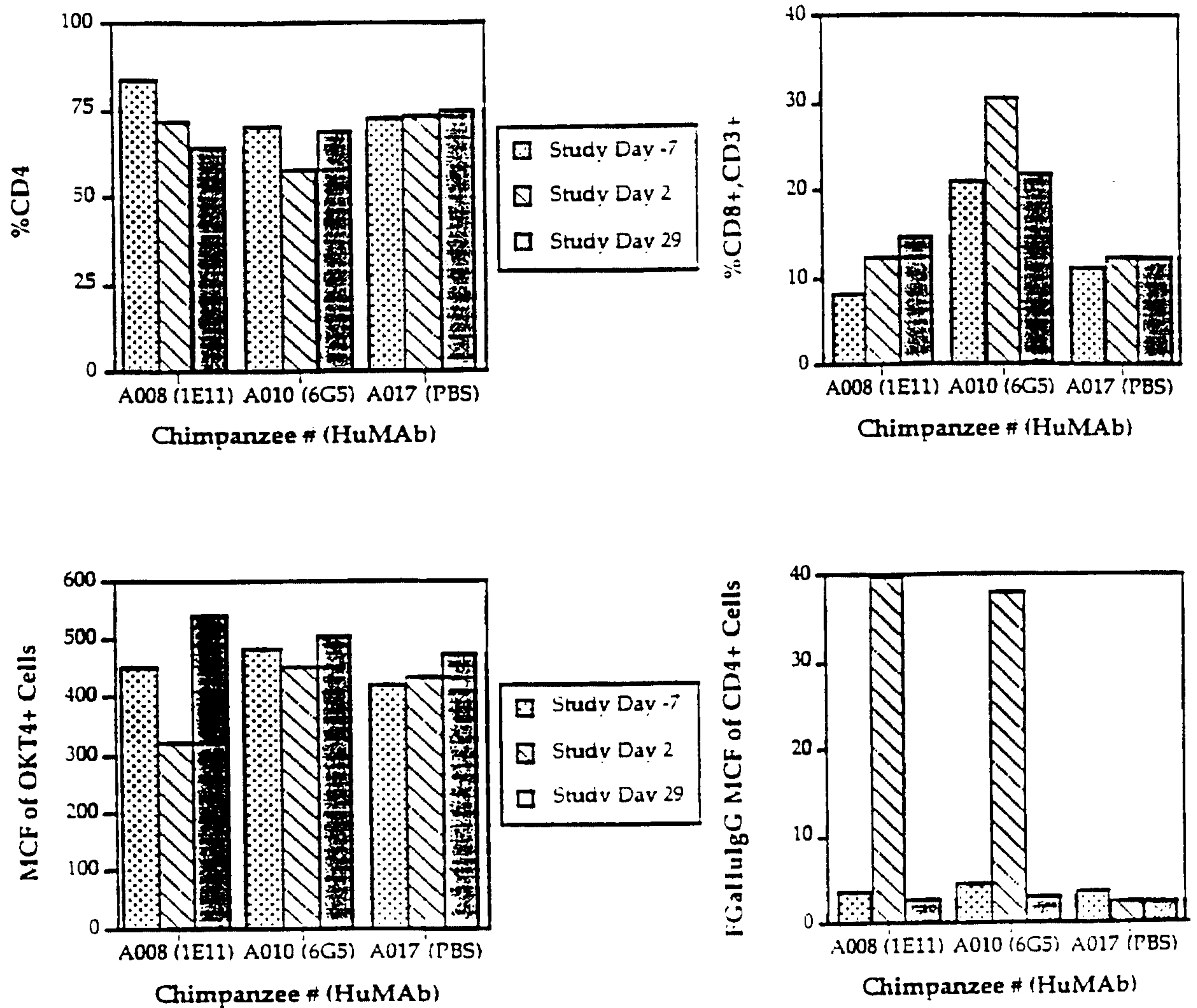


Figure 95 98/101

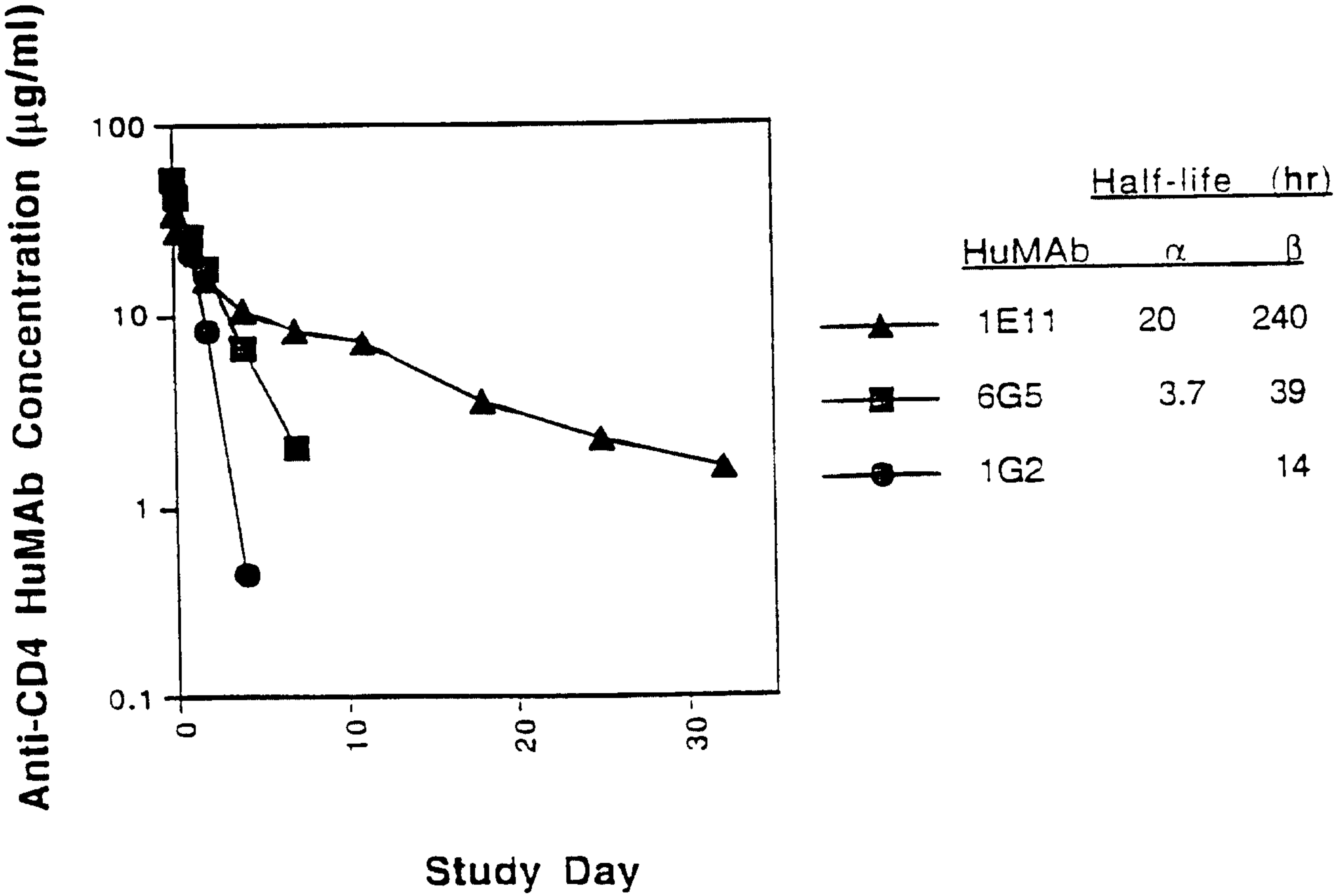
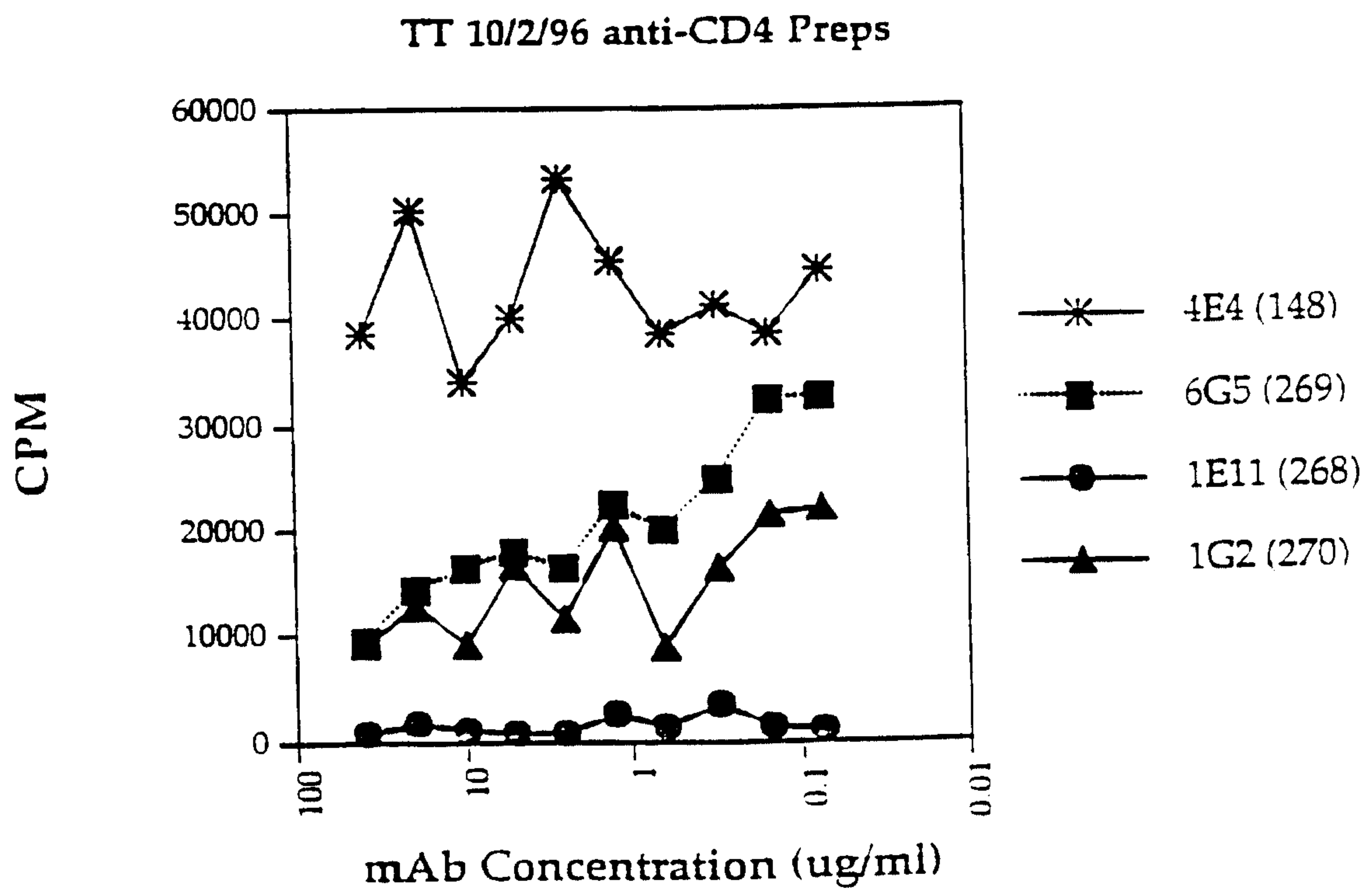
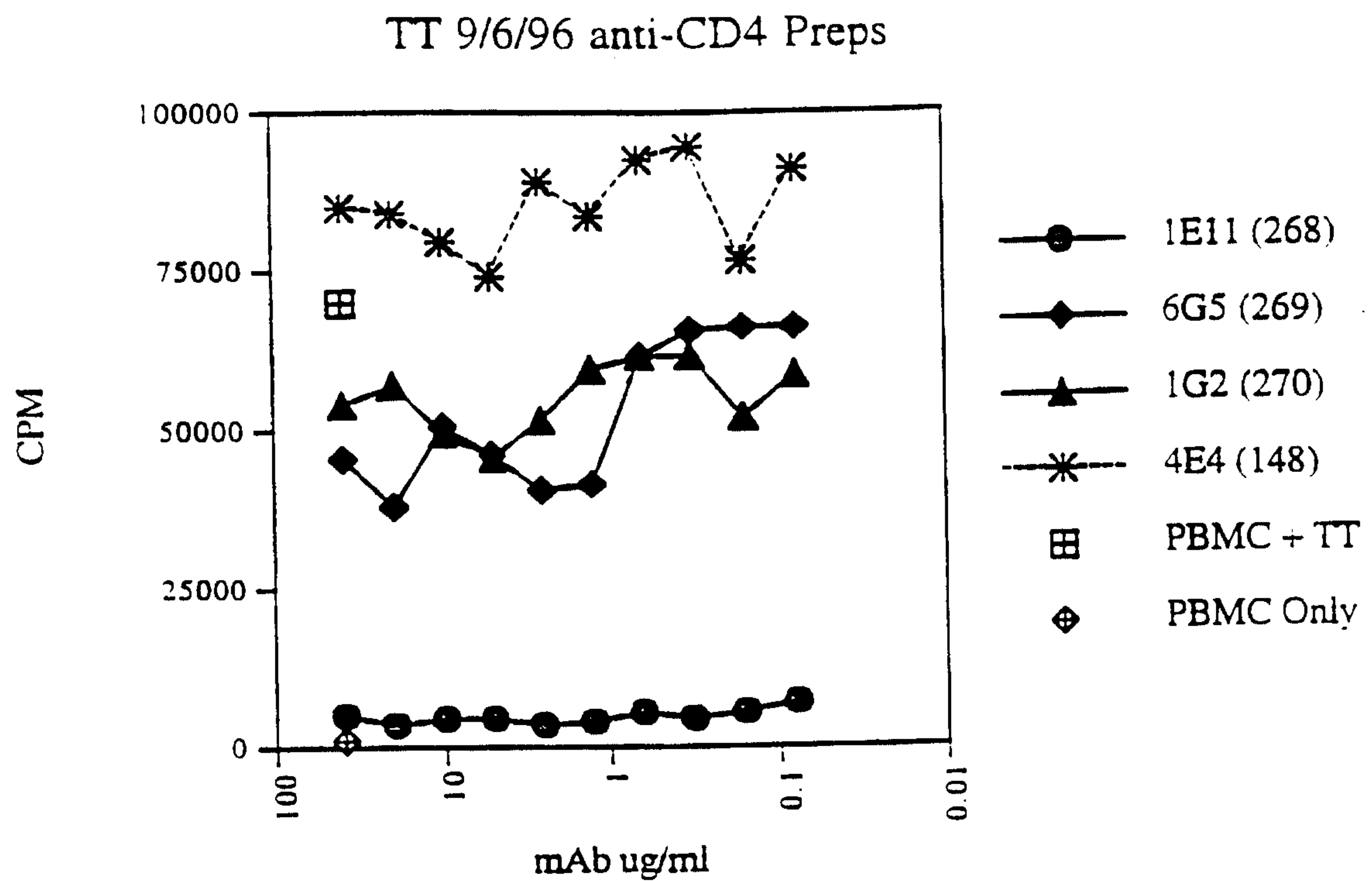


Figure 96

99/101

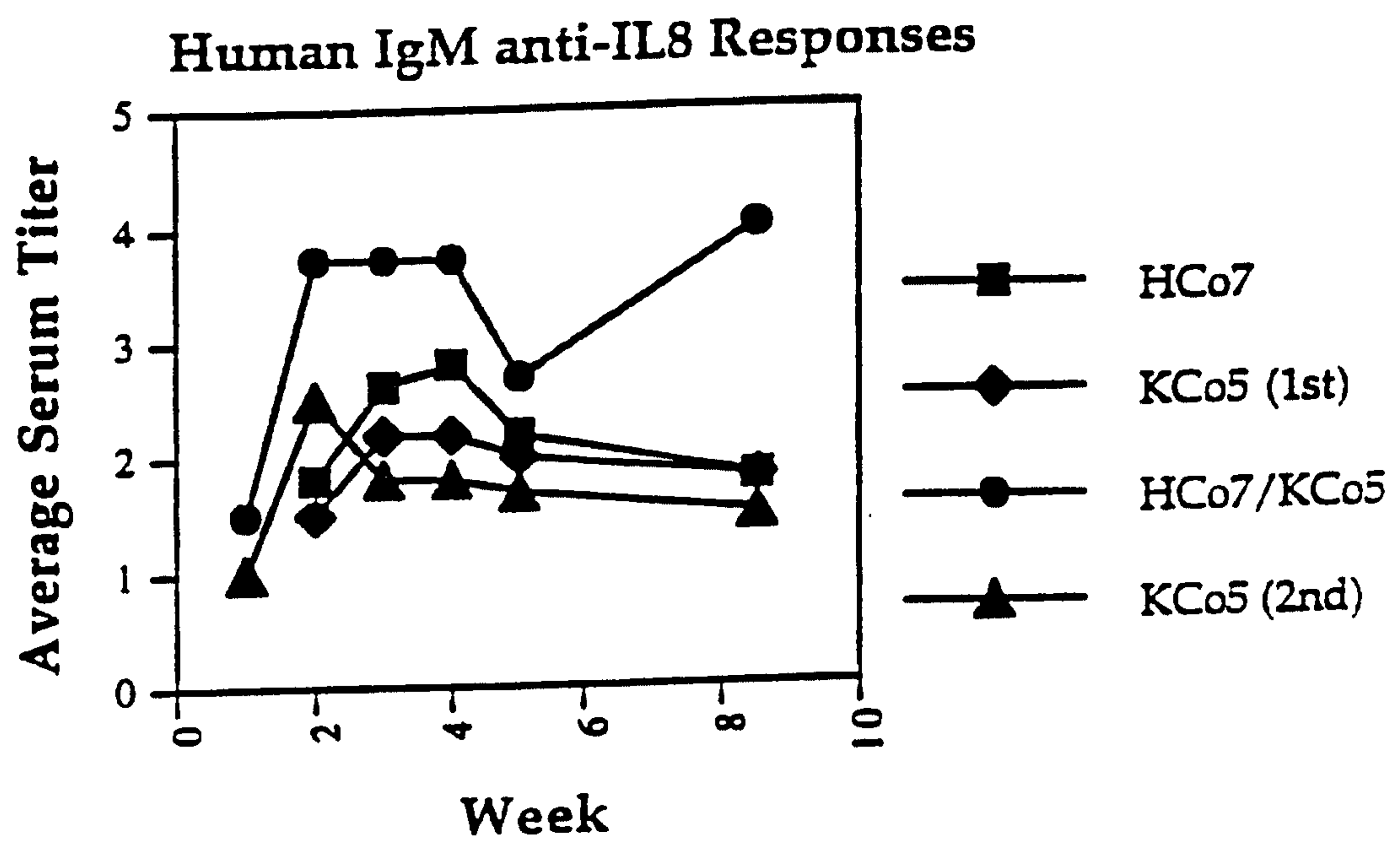


WO 98/24884

PCT/US97/21803

100/101

Fig. 97



WO 98/24884

PCT/US97/21803

101/101

Fig. 98

