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(54) TUBERCULOSIS NUCLEIC ACIDS, POLYPEPTIDES AND IMMUNOGENIC COMPOSITIONS

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(63) Continuation of application No. 11/291,616, filed on Nov. 30, 2005, now Pat. No. 7,608,277. (60) Provisional application No. 60/730,951, filed on Oct. 26, 2005, provisional application No. 60/632,573, filed on Dec. 1, 2004.

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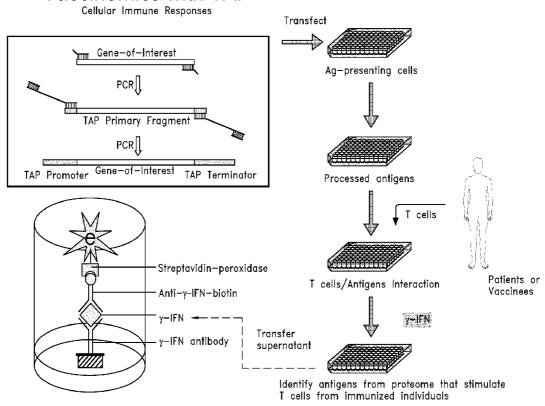
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(57) ABSTRACT

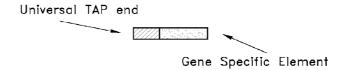
The present invention provides transcriptionally active Mtb polynucleotides, recombinant Mtb peptides and polypeptides, and immunogenic Mtb antigens. Immunogenic compositions are also provided that may be useful as recombinant, subunit and DNA vaccines. In addition the invention provides diagnostic kits for Mtb.

Vaccinomics with TAP

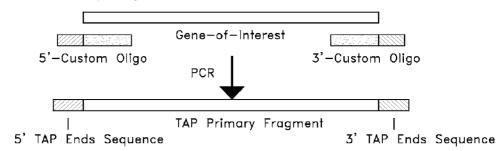


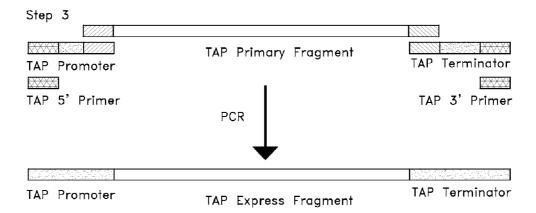
Step 1 Synthesize gene—specific custom primers containing the universal TAP ends

5' and 3' —Custom Oligos



Step 2 Amplify the gene—of—interest with the custom primers to create the TAP Primary Fragment





This fragment is transcriptionally active ready for transfection into cultured cells, or injection into animals

FIG. 1

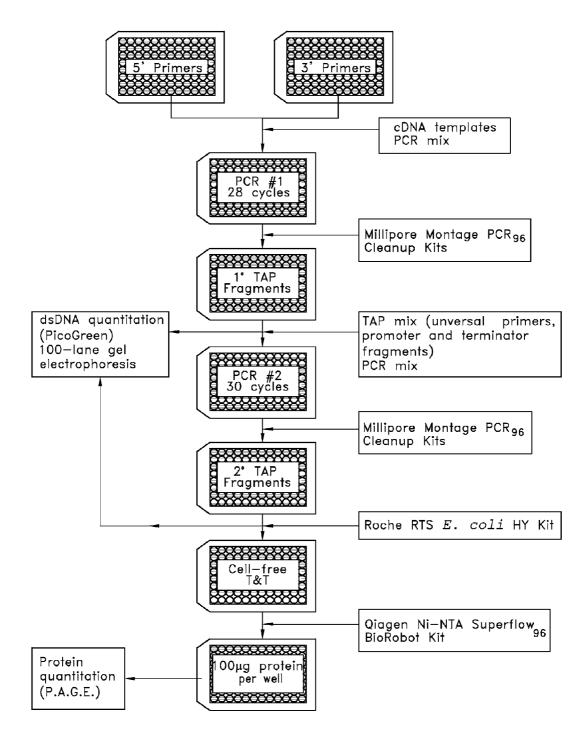
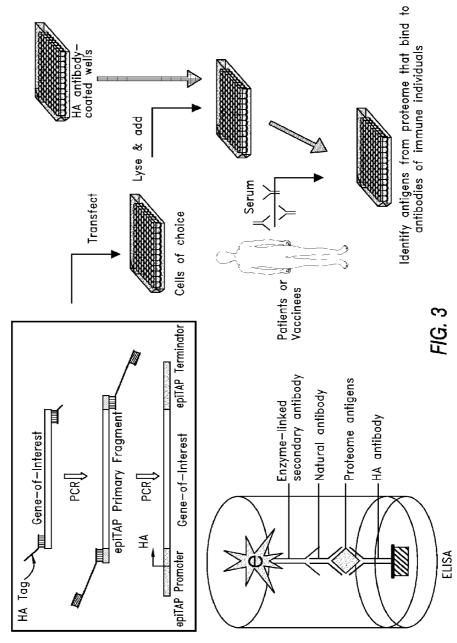
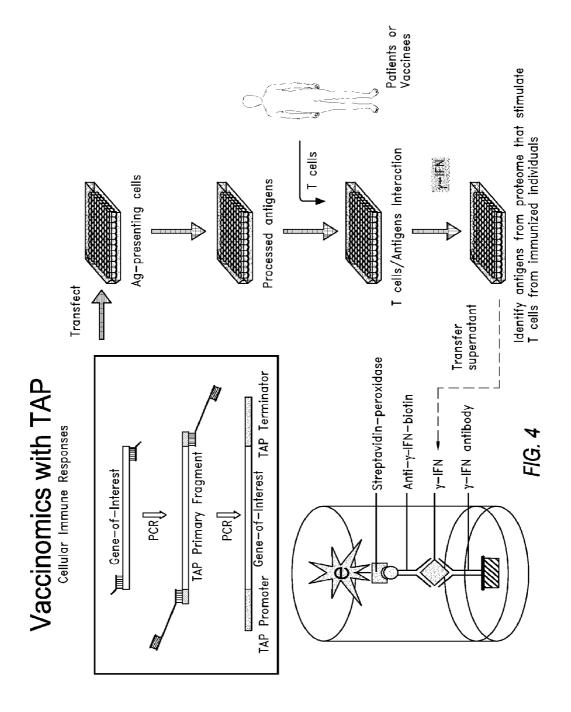


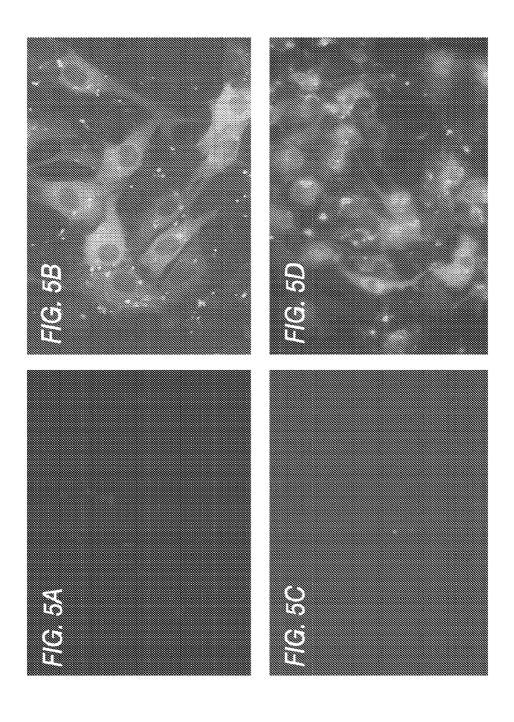
FIG. 2

Vaccinomics with TAP

Humoral Immune Responses







1.833.8 15080384 286252xi3 18 \$98348 9508098 883848 909848 939848 S62398 308C8V83 880849 2800343 640000 2 380143 10080618 Me94 305 83 \$\$00.9 1903/98 2018/02/9 2 889049 0689/8 94999AB 848948 -032204F 5.8556938 2 2500Z48 891046 3 Screening of Immunogenic TB proteins Western blot and ELISA with rabbit anti-TB serum (1:20,000) 900,9696 SEPSSAN š 31111 231.000 363333 3 33 3 1 2 38430vR 190348 P65594 575,628.33 2598348 1 💥 ********* č: **E**Y8344 8 400008 3900AS *** 2030848 >98\$868 O 70099B \$\$\$\$\$\& 3 848888 \$1256AIS 200,00A) 986643 1881/48 4 \$256045E 98.614.94 3 25068435 ,8010x3 9600788 19888198 25 1048 10008994 >\$\$C\$\3 1 2004.924B ×298994 ⁵বেচকে শ্ৰ 3,0848 2011/3 9853399 \$903465 S808% 22 8005500 1208213 861848 1040401 3 601098 \$00068 P\$\$88745 22870YA N86848 613063 824098 1100% 3,583485 989043 £98949 803348 3 083049 3990968 8 580800B 894848 \$ 362048 102693 803948 P063648 Colloidal Blue Western Western Well ID Well ID ELISA ELISA Gene Gene

TUBERCULOSIS NUCLEIC ACIDS, POLYPEPTIDES AND IMMUNOGENIC COMPOSITIONS

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application is a continuation of and claims the benefit of priority to U.S. application Ser. No. 11/291,616 filed on Nov. 30, 2005, entitled TUBERCULOSIS NUCLEIC ACIDS, POLYPEPTIDES AND IMMUNOGENIC COMPOSITIONS, which claims the benefit of priority to U.S. Application Ser. No. 60/632,573 filed on Dec. 1, 2004 and U.S. Application Ser. No. 60/730,951, filed Oct. 26, 2005. The contents of all the aforementioned applications are hereby expressly incorporated by reference, in their entireties.

STATEMENT REGARDING FEDERALLY SPONSORED R&D

[0002] The research leading to the present invention was supported, at least in part, by a grant from the National Institute of Allergy And Infectious Diseases. Accordingly, the Government may have certain rights in the invention.

REFERENCE TO SEQUENCE LISTING, TABLE, OR COMPUTER PROGRAM LISTING

[0003] The present application is being filed along with a sequence listing in electronic format. The sequence listing is provided as a file entitled GTSYS.033C1.txt, created Oct. 27, 2009 which is 101 KB in size. The information in the electronic format of the sequence listing is incorporated herein by reference in its entirety.

BACKGROUND OF THE INVENTION

[0004] Traditional vaccine technology suffers from the problem that it often produces various degrees of immunogenicity in different hosts. Often, the only reliably immunogenic composition is a pathogenic microorganism. But the manufacture and administration of the pathogenic organism carries a risk of infection by the very pathogen the vaccine is designed to treat. Furthermore, recent well-publicized problems with influenza vaccine production highlight the difficulties in producing large quantities of conventional vaccines and the precarious state of worldwide vaccine supplies. In light of general health concerns and the growing threat of bioterrorism, there is a need to develop recombinant and subunit vaccines capable of inducing an appropriate immune response in the context of multiple and genetically diverse hosts. This approach requires the identification of a number of specific antigenic polypeptides. One of the most difficult tasks in developing a protective or therapeutic vaccine, be it a recombinant or genetic, subunit or multi-valent vaccine, is the identification of the appropriate antigens that can stimulate the most rapid, sustained and efficacious immune responses against a particular pathogen for protection and/or therapeutic effect. This is especially challenging when the genome of the pathogen is large and screening for immunogenic antigens is tedious.

[0005] Tuberculosis is a chronic infectious disease that kills approximately 3 million people per year. It has been estimated that two billion people are infected with *M. tubercu*-

losis worldwide, including 7.5 million with active cases of tuberculosis. In recent years there has been an unexpected rise in tuberculosis cases.

[0006] In the U.S., tuberculosis continues to be a major problem especially among the homeless, Native Americans, African-Americans, immigrants, and the elderly. Immunocompromised individuals are particularly susceptible to tuberculosis. Of the 88 million, new cases of tuberculosis projected in this decade, approximately 10% are expected to be attributable to HIV infection. The emergence of AIDS has reactivated millions of dormant cases of tuberculosis (Mtb), causing a sharp rise in the number of tuberculosis-associated deaths.

[0007] The only available vaccine for tuberculosis, BCG, is both unpredictable and highly variable in protective efficacy. Hundreds of millions of children and newborns have been vaccinated with BCG, yet this has not consistently stopped the spread of the disease. Tuberculosis has become one of the fastest spreading infectious diseases in both industrialized and developing countries worldwide. Doubtful efficacy of vaccination has spurred interest in developing effective alternatives to BCG.

[0008] The emergence of multi-drug resistant strains of *M. tuberculosis* e.g. or Mtb, has complicated matters further, with some experts predicting a new tuberculosis epidemic. In the U.S. about 14% of *M. tuberculosis* isolates are resistant to at least one drug, and approximately 3% are resistant to at least two drugs. Some *M. tuberculosis* strains have been isolated that are resistant to as many as seven drugs in the repertoire of drugs commonly used to combat tuberculosis. Resistant strains make treatment of tuberculosis extremely difficult, leading to a mortality rate of about 90%, which is one of the reasons it has gained priority as a defined CDC—Category C Biodefense organism.

[0009] In the current age, where treatment of tuberculosis is becoming more challenging and immunosuppressive diseases are more prevalent, new vaccines are essential. Thus, there is a need for developing and commercializing effective and reliable Mtb vaccines. In addition, there is a considerable need for additional diagnostic tests or tests to detect active TB in the face of other diseases such as HIV.

SUMMARY OF THE INVENTION

[0010] The present invention provides isolated polynucleotides encoding a Mtb polypeptide that is antigenic in any mammal, including SEQ ID NOS: 46-64, 110-121, and fragments thereof, that encode antigenic polypeptides. The mammal can be, for example, a mouse, rabbit, non-human primate, or human. The invention also provides isolated polynucleotides encoding highly immunogenic Mtb antigens including SEQ ID NOS: 46-64, 110-121 and fragments thereof that encode highly immunogenic polypeptides. In some embodiments, highly immunogenic Mtb antigens react with polyclonal antibodies directed to Mtb bacteria (Mtb) from at least two different species. In another embodiment, highly immunogenic Mtb antigens are detected by ELISA, Western blotting, or both using polyclonal antibodies that are directed to Mtb bacteria.

[0011] The present invention also provides TAP polynucleotides, e.g. polynucleotides produced by Transcriptionally-Activated PCR (TAP) technology as described in U.S. Pat. No. 6,280,977, which is expressly incorporated herein by reference. Such polynucleotides include a 5' TAP polynucleotide sequence, a Mtb polynucleotide sequence, and a 3' TAP polynucleotide sequence. The Mtb polynucleotide sequence can, for example, comprise one of SEQ ID NOS: 46-64 and 110-121. In some embodiments, the 5' TAP polynucleotide sequence comprises a promoter. In certain embodiments, the 5' TAP polynucleotide sequence is selected from SEQ ID NOS: 2, 3, 6, and 84. In some embodiments the 3' TAP polynucleotide sequence comprises a terminator. In certain embodiments, the 3' TAP polynucleotide sequence is selected from SEQ ID NOS: 4, 5, 7, and 85.

[0012] The present invention also provides primer pairs for amplifying an Mtb polynucleotide. These primer pairs include SEQ ID NOS: 8 and 9; 10 and 11; 12 and 13; 14 and 15; 16 and 17; 18 and 19; 20 and 21; 22 and 23; 24 and 25; 26 and 27; 28 and 29; 30 and 31; 32 and 33; 34 and 35; 36 and 37; 38 and 39; 40 and 41; 42 and 43; 44 and 45; 86 and 87; 88 and 89; 90 and 91; 92 and 93; 94 and 95; 96 and 97; 98 and 99; 100 and 101; 102 and 103; 104 and 105; 106 and 107; 108 and 109

[0013] Isolated antigenic Mtb peptides and polypeptides are encompassed by the invention, including SEQ ID NOS: 65-83, 122-133, and fragments thereof. Isolated Mtb peptides and polypeptides that are highly immunogenic, including SEQ ID NOS: 65-83, 122-133, and fragments thereof, that are highly immunogenic, are also included in the invention. In one embodiment immunogenic peptides and polypeptides react with polyclonal antibodies that are directed to Mtb bacteria (Mtb). In one aspect of this embodiment, the peptides and polypeptides react with polyclonal antibodies that are directed to Mtb bacteria from at least two different species. In another embodiment, highly immunogenic peptides and polypeptides are detected by ELISA, Western blotting or both using polyclonal antibodies that are directed to Mtb bacteria.

[0014] The present invention also includes recombinant Mtb peptides and polypeptides, wherein the amino terminus of the peptide or polypeptide comprises an HA tag or a (6.times.)His tag, and the carboxy terminus of the polypeptide is selected from the group consisting of: SEQ ID NOS: 65-83 and 122-133. Also included are recombinant Mtb peptides and polypeptides, wherein the carboxy terminus of the polypeptide comprises a HA tag or a His tag, and the amino terminus of the polypeptide is selected from the group consisting of: SEQ ID NOS: 65-83 and 122-133. The peptides and polypeptides of the invention may be expressed in an appropriate in vitro transcription and translation system, such as a T7 polymerase system.

[0015] Immunogenic compositions for inducing an immunological response in a mammalian host against Mtb are also included in the invention. In one embodiment, the immunogenic compositions comprise nucleic acids that encode and express in vivo in a mammalian host cell at least one immunogenic peptide or polypeptide, which may be any one of SEQ ID NOS: 65-83, 122-133, or immunogenic fragments thereof. The nucleic acids can be, for example, plasmids or TAP fragments. The compositions can induce either a humoral- or cell-mediated immune response. Furthermore, the immunogenic compositions can include additional components, such as adjuvants, as well as other applications such as serodiagnostics.

[0016] Immunogenic compositions for inducing an immunological response in a mammalian host against Mtb of the invention can also comprise isolated Mtb peptides and/or polypeptides, such as SEQ ID NOS: 65-83, 122-133 and immunogenic fragments thereof. In one embodiment, the immunogenic peptides and polypeptides are expressed in an

in vitro transcription and translation system. The immunogenic peptide and polypeptide compositions can induce either a humoral- or cell-mediated immune response. Furthermore, the immunogenic peptide and polypeptide compositions can include additional components, such as adjuvants, and include other applications such as diagnostics.

[0017] Similarly, detection of Mtb, its constituent proteins, and/or its immunologically reactive products (e.g. antibodies) is clinically relevant for the diagnosis of Mtb, and to track the efficacy of therapeutic treatments for Mtb, especially as translated to serodiagnostic tests. The present application therefore provides antigens for detection of Mtb for immune assays, including humoral immune assays. These antigens are applicable to detection of active Mtb in the face of HIV- and other Mycobacterial-coinfections, multi-drug resistant infections by Mtb (MDR1), and rapidly mutating forms of Mtb depending on genetic makeup, geographical location, and immunocompetency status.

[0018] As such, the present invention also provides diagnostic compositions, including one or more antibodies directed against the peptide epitopes identified herein. Also, the present invention provides diagnostic kits that include at least one or more of such antibodies.

[0019] In addition, the invention provides a method of generating an immune response in a mammalian host against Mtb. The method includes administering to said mammalian host an immunogenic composition comprising at least one nucleic acid selected from the group of SEQ ID NO: 46-64, 110-121, fragments thereof or combinations thereof, wherein said nucleic acid encodes and expresses in vivo at least one immunogenic peptide or polypeptide, whereby said immune response against Mtb is generated.

[0020] Also, the invention provides a method of generating an immune response in a mammalian host against Mtb, the method including administering to said mammalian host an immunogenic composition comprising at least one nucleic acid encoding and expressing in vivo at least one immunogenic peptide or polypeptide selected from the group of SEQ ID NO: 65-83, 122-133, fragments thereof or combinations thereof, whereby said immune response against Mtb is generated

[0021] In addition, the invention provides a method of generating an immune response in a mammalian host against Mtb comprising administering to said mammalian host an immunogenic composition comprising at least one immunogenic peptide or polypeptide selected from SEQ ID NO: 65-83, 122-133, fragment thereof or combinations thereof, whereby said immune response against Mtb is generated.

[0022] Also, the invention provides kits. In one embodiment the kits include at least one Mtb immunogenic composition selected from a nucleic acid selected from the group consisting of SEQ ID NO: 46-64, 110-121, fragments thereof or combinations thereof, or a peptide selected from the group consisting of SEQ ID NO: 65-83, 122-133, fragments thereof or combinations thereof and an adjuvant. The kit may also include an expression system. In addition the kit may include at least 2, 5, 10, 15, 20 or more of said immunogenic compositions, including nucleic acids and/or peptides, combinations or fragments thereof. In addition, the kit may include controls, e.g. positive and/or negative controls.

BRIEF DESCRIPTION OF THE DRAWINGS

[0023] FIG. 1 illustrates one method used to generate TAP Expression Fragments.

[0024] FIG. 2 displays a method of amplifying multiple genes using TAP technology, expressing said genes products, then purifying and quantifying the resulting polypeptides.

[0025] FIG. 3 demonstrates how a plurality of polypeptides from a target organism can be assayed to determine each polypeptide's ability to elicit a humoral immune response.

[0026] FIG. 4 demonstrates how a plurality of polypeptides from a target organism can be assayed to determine each polypeptide's ability to elicit a cell-mediated response.

[0027] FIG. 5 demonstrates that fluorescent proteins (goat IgG antibody) can be more effectively delivered into either NIH-3 T3 cells (A&B) and human dendritic cells (C&D) with a protein delivery reagent (B&D) as opposed to without a protein delivery reagent (A&C).

[0028] FIG. 6 shows the results of scanning the Mtb proteome for antigenic targets of humoral immunity by ELISA and Western blotting.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENT

[0029] Before proceeding further with a description of the specific embodiments of the present invention, a number of terms will be defined and described in detail.

[0030] Unless specific definitions are provided, the nomenclature utilized in connection with, and the laboratory procedures, techniques and methods described herein are those known in the art to which they pertain. Standard chemical symbols and abbreviations are used interchangeably with the full names represented by such symbols. Thus, for example, the terms "carbon" and "C" are understood to have identical meaning. Standard techniques may be used for chemical syntheses, chemical analyses, pharmaceutical preparation, formulation, delivery, and treatment of patients. Standard techniques may be used for recombinant DNA methodology, oligonucleotide synthesis, tissue culture and the like. Reactions and purification techniques may be performed e.g., using kits according to manufacturer's specifications, as commonly accomplished in the art or as described herein. The foregoing techniques and procedures may be generally performed according to conventional methods well known in the art and as described in various general or more specific references that are cited and discussed throughout the present specification. All references cited herein are incorporated by reference in their entirety and are not admitted to be prior art. See e.g., Sambrook et al. Molecular Cloning: A Laboratory Manual (3rd ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (2000)), Harlow & Lane, Antibodies: A Laboratory Manual (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. (1988)), which are incorporated herein by reference in their entirety for any purpose.

[0031] The terms "polynucleotide" and "nucleic acid (molecule)" are used interchangeably to refer to polymeric forms of nucleotides of any length. The polynucleotides may contain deoxyribonucleotides, ribonucleotides and/or their analogs. Nucleotides may have any three-dimensional structure, and may perform any function, known or unknown. The term "polynucleotide" includes single-stranded, double-stranded and triple helical molecules. The following are non-limiting embodiments of polynucleotides: a gene, a gene fragment, exons, introns, mRNA, tRNA, rRNA, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, cosmids, viruses and other vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes and primers. A nucleic acid molecule may also com-

prise modified nucleic acid molecules, such as methylated nucleic acid molecules and nucleic acid molecule analogs. Analogs of purines and pyrimidines are known in the art, and include, but are not limited to, aziridinylcytosine, 4-acetylcytosine, 5-fluorouracil, 5-bromouracil, 5-carboxymethylaminomethyl-2-thiouracil, 5-carboxymethyl-aminomethyluracil, inosine, N6-isopentenyladenine, 1-methyladenine, 1-methylpseudouracil, 1-methylguanine, 1-methylguanine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, pseudouracil, 5-pentyluracil and 2,6-diaminopurine.

[0032] The use of uracil as a substitute for thymine in a deoxyribonucleic acid is also considered an analogous form of pyrimidine.

[0033] Sugar modifications (e.g., 2'-o-methyl, 2-fluoro and the like) and phosphate backbone modifications (e.g., morpholino, PNA', thioates, dithioates and the like) can be incorporated singly, or in combination, into the nucleic acid molecules of the present invention. In one embodiment, for example, a nucleic acid of the invention may comprise a modified sugar and a modified phosphate backbone. In another embodiment, a nucleic acid of the invention may comprise modifications to sugar, base and phosphate backbone.

[0034] "Oligonucleotide" refers generally to polynucleotides of between 5 and about 100 nucleotides of single- or double-stranded nucleic acid, typically DNA. Oligonucleotides are also known as oligomers or oligos and may be isolated from genes, or synthesized (e.g., chemically or enzymatically) by methods known in the art. A "primer" refers to an oligonucleotide, usually single-stranded, that provides a 3'-hydroxyl end for the initiation of enzyme-mediated nucleic acid synthesis.

[0035] "Peptide" generally refers to a short chain of amino acids linked by peptide bonds. Typically peptides comprise amino acid chains of about 2-100, more typically about 4-50, and most commonly about 6-20 amino acids. "Polypeptide" generally refers to individual straight or branched chain sequences of amino acids that are typically longer than peptides. "Polypeptides" usually comprise at least about 100 to about 1000 amino acids in length, more typically at least about 150 to about 600 amino acids, and frequently at least about 200 to about 500 amino acids. "Proteins" include single polypeptides as well as complexes of multiple polypeptide chains, which may be the same or different. Multiple chains in a protein may be characterized by secondary, tertiary and quaternary structure as well as the primary amino acid sequence structure; may be held together, for example, by disulfide bonds; and may include post-synthetic modifications such as, without limitation, glycosylation, phosphorylation, truncations or other processing. Antibodies such as IgG proteins, for example, are typically comprised of four polypeptide chains (i.e., two heavy and two light chains) that are held together by disulfide bonds. Furthermore, proteins may include additional components such as associated metals (e.g., iron, copper and sulfur), or other moieties. The definitions of peptides, polypeptides and proteins include, without limitation, biologically active and inactive forms; denatured and native forms; as well as variant, modified, truncated, hybrid, and chimeric forms thereof. The peptides, polypeptides and proteins of the present invention may be derived from any source or by any method, including, but not limited to extraction from naturally occurring tissues or other materials; recombinant production in host organisms such as bacteria, fungi, plant, insect or animal cells; and chemical synthesis using methods that will be well known to the skilled artisan.

[0036] "Polyclonal antibodies" or "antisera" are heterogeneous populations of antibody molecules derived from the sera of animals immunized with an antigen, or an antigenic functional derivative thereof. For the production of polyclonal antibodies, host animals such as rabbits, mice and goats, may be immunized by injection with an antigen or hapten-carrier conjugate optionally supplemented with adjuvants. Polyclonal antibodies may also be derived from the sera of humans or non-human animals exposed to a pathogen or vaccinated against a pathogen using a commercially available or experimental vaccine. An antiserum against TB (Mtb), for example, may be obtained from a human patient vaccinated with a TBvaccine, or from an animal, such as a mouse, rabbit, goat or sheep immunized with Mtb bacteria or a Mtb preparation.

[0037] "Monoclonal antibodies," which are abbreviated MAb, are homogeneous populations of antibodies to a particular antigen, may be obtained by any technique that provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to the hybridoma technique of Kohler and Milstein, Nature, 256: 495-7 (1975); and U.S. Pat. No. 4,376,110, the human B-cell hybridoma technique (Kosbor, et al., Immunology Today, 4:72 (1983); Cote, et al., Proc. Natl. Acad. Sci. USA, 80:2026-30 (1983), and the EBV-hybridoma technique (Cole, et al., in Monoclonal Antibodies And Cancer Therapy, Alan R. Liss, Inc., New York, pp. 77-96 (1985)). Such antibodies may be of any immunoglobulin class including IgG, IgM, IgE, IgA, IgD and any subclass thereof. The hybridoma producing the MAb of this invention may be cultivated in vitro or in vivo. Production of high titers of MAbs in vivo makes this a presently preferred method of production

[0038] In addition, techniques developed for the production of "chimeric antibodies" (Morrison, et al., Proc. Natl. Acad. Sci., 81:6851-6855 (1984); Takeda, et al., Nature, 314:452-54 (1985)) by splicing the genes from a mouse antibody molecule of appropriate antigen specificity together with genes from a human antibody molecule of appropriate biological activity can be used. A chimeric antibody is a molecule in which different portions are derived from different sources, such as those having a variable region derived from a murine MAb and a human immunoglobulin constant region. Humanized antibodies can also be generated in which certain parts (e.g., framework regions) of a non-human antibody are altered to make the antibody more like a human antibody, while retaining antigen binding features of the parent molecule.

[0039] Alternatively, techniques described for the production of single chain antibodies (U.S. Pat. No. 4,946,778; Bird, Science 242:423-26 (1988); Huston, et al., Proc. Natl. Acad. Sci. USA, 85:5879-83 (1988); and Ward, et al., Nature, 334: 544-46 (1989)) can be adapted to produce single chain antibodies. Single chain antibodies are typically formed by linking the heavy and light chain fragments of the Fv region via an amino acid bridge, resulting in a single chain polypeptide.

[0040] Antibody fragments that recognize specific epitopes may be generated by known techniques. For example, such fragments include but are not limited to: the Fab fragments that can be produced by papain digestion of the antibody molecule, the $F(ab')_2$ fragments that can be produced by pepsin digestion of the antibody molecule and the Fab' fragments

that can be generated by reducing the disulfide bridges of the $F(ab')_2$ fragments. Alternatively, Fab expression libraries may be constructed (Huse, et al., Science, 246:1275-81 (1989)) to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity.

[0041] The term "hapten" as used herein, refers to a small proteinaceous or non-protein antigenic determinant that is capable of being recognized by an antibody. Typically, haptens do not elicit antibody formation in an animal unless part of a larger species. For example, small peptide haptens are frequently coupled to a carrier protein such as keyhole limpet hemocyanin in order to generate an anti-hapten antibody response. "Antigens" are macromolecules capable of generating an antibody response in an animal and being recognized by the resulting antibody. Both antigens and haptens comprise at least one antigenic determinant or "epitope," which is the region of the antigen or hapten that binds to the antibody. Typically, the epitope on a hapten is the entire molecule.

[0042] By the terms "specifically binding" and "specific binding" as used herein is meant that an antibody or other molecule, binds to a target such as an antigen, with greater affinity than it binds to other molecules under the specified conditions of the present invention. Antibodies or antibody fragments, as known in the art, are polypeptide molecules that contain regions that can bind other molecules, such as antigens. In various embodiments of the invention, "specifically binding" may mean that an antibody or other specificity molecule, binds to a target molecule with at least about a 10⁶-fold greater affinity, preferably at least about a 107-fold greater affinity, more preferably at least about a 108-fold greater affinity, and most preferably at least about a 109-fold greater affinity than it binds molecules unrelated to the target molecule. Typically, specific binding refers to affinities in the range of about 106-fold to about 109-fold greater than nonspecific binding. In some embodiments, specific binding may be characterized by affinities greater than 10⁹-fold over nonspecific binding. Whenever a range appears herein, as in "1-10 or one to ten, the range refers without limitation to each integer or unit of measure in the given range. Thus, by 1-10 it is meant that each of 1, 2, 3, 4, 5, 6, 7, 8, 9, 10 and any subunit in between.

[0043] "Immunogenic compositions" of the present invention are preparations that, when administered to a human or non-human animal, elicit a humoral and/or cellular immune response. "Vaccine," as used herein, refers to immunogenic compositions that are administered to a human or non-human patient for the prevention, amelioration or treatment of diseases, typically infectious diseases. "Traditional vaccines" or "whole vaccines" typically may be live, attenuated or killed microorganisms, such as bacteria or viruses. Vaccines also encompass preparations that elicit or stimulate an immune response that may be useful in the prevention, amelioration or treatment of non-infectious diseases. For example, a cancer cell vaccine may be administered to stimulate or supplement a patient's immune response to neoplastic disease. "Subunit vaccines" may be prepared from purified or partially purified proteins or other antigens from a microorganism, cancer cell or other vaccine target. The term "recombinant vaccine" refers to any vaccine that is prepared using recombinant DNA technology and includes certain subunit vaccines (for example, where subunits are cloned and expressed in vitro prior to administration) and "polynucleotide vaccines" such as DNA vaccines that may encode immunogenic polypeptides. Vaccines typically contain at least one immunogenic

component (e.g. a cell, virus, polypeptide, polynucleotide, and the like) but may also include additional agents such as adjuvants, which may enhance or stimulate the patient's immune response to the immunogenic component. In certain embodiments, vaccines or components of vaccines may be conjugated e.g. to a polysaccharide or other molecule, to improve stability or immunogenicity of one or more vaccine components.

[0044] As used herein, the term "promoter" refers to a DNA sequence having a regulatory function, which is recognized (directly or indirectly) and bound by a DNA-dependent RNA polymerase during the initiation of transcription. Promoters are typically adjacent to the coding sequence of a gene and extend upstream from the transcription initiation site. The promoter regions may contain several short (<10 base pair) sequence elements that bind transcription factors, generally located within the first 100-200 nucleotides upstream of the transcription initiation site. Sequence elements that regulate transcription from greater distances are generally referred to as "enhancers" and may be located several hundred or thousand nucleotides away from the gene they regulate. Promoters and enhancers may be cell- and tissue-specific; they may be developmentally programmed; they may be constitutive or inducible e.g., by hormones, cytokines, antibiotics, or by physiological and metabolic states. For example, the human metallothionein (MT) promoter is upregulated by heavy metal ions and glucocorticoids. Inducible promoters and other elements may be operatively positioned to allow the inducible control or activation of expression of the desired TAP fragment. Examples of such inducible promoters and other regulatory elements include, but are not limited to, tetracycline, metallothionine, ecdysone, and other steroidresponsive promoters, rapamycin responsive promoters, and the like (see e.g., No, et al., Proc. Natl. Acad. Sci. USA, 93:3346-51 (1996); Furth, et al., Proc. Natl. Acad. Sci. USA, 91:9302-6 (1994)). Certain promoters are operative in prokaryotic cells, while different promoter sequences are required for transcription in eukaryotic cells. Additional control elements that can be used include promoters requiring specific transcription factors, such as viral promoters that may require virally encoded factors. Promoters can be selected for incorporation into TAP fragments based on the intended use of the polynucleotide, as one skilled in the art will readily appreciate. For example, if the polynucleotide encodes a polypeptide with potential utility in human cells, then a promoter capable of promoting transcription in mammalian cells can be selected. Typical mammalian promoters include muscle creatine kinase promoter, actin promoter, elongation factor promoter as well as those found in mammalian viruses such as CMV, SV40, RSV, MMV, HIV, and the like. In certain embodiments, it may be advantageous to incorporate a promoter from a plant or a plant pathogen (e.g., cauliflower mosaic virus promoter), a promoter from a fungus such as yeast (e.g., Gal 4 promoter), a promoter from a bacteria or bacterial virus, such as bacteriophage lambda, T3, T7, SP6, and the like.

[0045] The term "terminator" refers to DNA sequences, typically located at the end of a coding region, that cause RNA polymerase to terminate transcription. As used herein, the term "terminator" also encompasses terminal polynucle-otide sequences that direct the processing of RNA transcripts prior to translation, such as, for example, polyadenylation signals. Any type of terminator can be used for the methods and compositions of the invention. For example, TAP termi-

nator sequences can be derived from a prokaryote, eukaryote, or a virus, including, but not limited to animal, plant, fungal, insect, bacterial and viral sources. In one embodiment, artificial mammalian transcriptional terminator elements are used. A nonexclusive list of terminator sequences that may be used in the present invention include the SV40 transcription terminator, bovine growth hormone (BGH) terminator, synthetic terminators, rabbit .beta.-globin terminator, and the like. Terminators can also be a consecutive stretch of adenine nucleotides at the 3' end of a TAP fragment.

[0046] By "pharmaceutically acceptable" or "pharmacologically acceptable" is meant a material which is not biologically or otherwise undesirable, i.e., the material may be administered to an individual in a formulation or composition without causing any undesirable biological effects or interacting in a deleterious manner with any of the components of the composition in which it is contained.

[0047] By "serodiagnostic test" or grammatical equivalents herein is meant diagnostic tests to detect Mtb by serum of infected organisms, animals or patients.

[0048] By "diagnostic test" or grammatical equivalents herein is meant an assay or test to detect Mtb by any scientific technique from infected organisms, animals or patients.

Overview

[0049] The present invention generally relates to Mtb polypeptide libraries, methods of determining the immunogenic effect of Mtb polypeptides, and methods of developing vaccines against Mtb, as well as immunogenic and pharmaceutical compositions. The invention also provides immunogenic Mtb polypeptides and mixtures of polypeptides, polynucleotides encoding immunogenic Mtb polypeptides and immunogenic compositions comprising Mtb polypeptides or polynucleotides.

Polypeptide Libraries

[0050] According to a method of the present invention, a library or array of Mtb polypeptides, oligonucleotides, or polynucleotides is generated. The immunogenicity of individual polypeptides in the library or array is determined by immunological screening where suitable, highly immunogenic Mtb polypeptides are selected for vaccine development. Conveniently, individual polypeptides in the library may be arranged in an array to facilitate screening in a rapid and high throughput manner.

[0051] The term "array" includes any arrangement wherein a plurality of different molecules, compounds or other species are contained, held, presented, positioned, situated, or supported. Arrays can be arranged on microtiter plates, such as 48-well, 96-well, 144-well, 192-well, 240-well, 288-well, 336-well, 384-well, 432-well, 480-well, 576-well, 672-well, 768-well, 864-well, 960-well, 1056-well, 1152-well, 1248well, 1344-well, 1440-well, or 1536-well plates, tubes, slides, chips, flasks, or any other suitable laboratory apparatus. In one embodiment, molecules arranged in an array are peptides, polypeptides or proteins. In another embodiment, the molecules are oligonucleotides or polynucleotides. In one aspect of the invention, polypeptides or polynucleotides in solution are arranged in 96 well plate arrays. In another embodiment, polypeptides or polynucleotides are immobilized on a solid support in an array format. Furthermore, an array can be sub-divided into a plurality of sub-arrays, as for

example, where multiple 96-well plates (each an individual sub-array) are required to hold all of the samples of a single, large array.

[0052] The term "library" is likewise to be construed broadly, and includes any non-naturally occurring collection of molecules, whether arranged or not. A library therefore encompasses an array but the two terms are not necessarily synonymous.

TAP Technology

[0053] Libraries of Mtb polypeptides may be prepared by any method known in the art. Conveniently, GTS' patented Transcriptionally Active PCR ("TAP") products can be used to amplify DNA in preparation for producing Mtb polypeptide libraries. With TAP technology, a particular polynucleotide of interest can be made transcriptionally active and ready for expression in less than one day. "TAP fragments" are transcriptionally active coding sequences prepared using TAP technology, and the two terms can be used interchangeably. TAP fragments encompass polynucleotides that can be readily expressed, for example, by transfection into animal cells or tissues by any nucleic acid transfection technique, without the need for subcloning into expression vectors or purification of plasmid DNA from bacteria. TAP fragments can be synthesized by amplification (e.g., polymerase chain reaction, or PCR) of any polynucleotide of interest using nested oligonucleotide primers. Two polynucleotide sequences are typically incorporated into TAP fragments, one of which comprises an active transcriptional promoter and the other comprises a transcriptional terminator.

[0054] TAP fragments and methods of making the same are described in detail in U.S. Pat. No. 6,280,977, entitled "Method for Generating Transcriptionally Active DNA Fragments" which is hereby incorporated by reference in its entirety. In one embodiment, methods for creating TAP fragments include the steps of: i) designing oligonucleotide primers; ii) amplifying TAP primary fragments; and iii) amplifying TAP expression fragments. FIG. 1 illustrates one method for generating TAP fragments.

[0055] TAP fragments can be prepared using custom oligonucleotide primers designed to amplify a target polynucleotide sequence of interest from the Mtb genome. Primers complementary to the 5' and 3' ends of the polynucleotide of interest can be designed and synthesized using methods well known in the art, and can include any suitable number of nucleotides to permit amplification of the coding region. Typically, the polynucleotide sequence of interest is an open reading frame (ORF) that consists of an uninterrupted stretch of triplet amino acid codons, without stop codons. In certain embodiments, the polynucleotide is a Mtb polypeptide-encoding sequence.

[0056] In one embodiment of the invention, 5'-custom oligonucleotide primers of about 41, 42, 43, 44, 45 or 46 nucleotides are designed and synthesized; about 6 nucleotides of which comprise the 5'-TAP end universal sequence 5'-GAAGGAGATATACCATGCATCATCATCATCATCATCAT-3' (SEQ ID NO: 84) and about 15 to 20 nucleotides are complementary to the Mtb sequence. Accordingly, the target-specific sequence can be, for example, about 15, 16, 17, 18, 19, or 20 nucleotides in length. The 5' oligonucleotide may also incorporate a Kozak consensus sequence (A/GC-CAUGG) near an ATG start codon (initiator methionine) for more efficient translation of mRNA. In one embodiment, an ATG start codon is included in the target-specific primer

sequence. In another embodiment, an ATG start codon is incorporated into the custom 5'-oligonucleotide when the target sequence encoding a polypeptide of interest lacks an initiation methionine codon at its 5' end

[0057] In one embodiment of the invention, 3'-custom oligonucleotide primers comprise about 35, 36, 37, 38, 39, 40, 41, 42, 43, 44, or 45 nucleotides; of these, about 20 nucleotides comprise the 3'-TAP end universal sequence 5'-TGAT-GATGAGAACCCCCCCC-3' (SEQ ID NO: 85) and about 20 nucleotides are complementary to the target Mtb sequence. In one aspect, a stop codon sequence, can be added to the end of the target Mtb sequence to achieve proper translational termination by incorporating a TCA, TTA, or CTA into the 3'-custom oligonucleotide.

Bioinformatics Analysis of Mtb Polynucleotides

[0058] In one embodiment of the invention, a bioinformatics approach is used to identify, prioritize and select Mtb genes, coding sequences, ORFs and other sequences of interest for TAP amplification and to design custom 5' and 3' oligonucleotide primers. According to this approach, a database of Mtb genomic information is compiled from available nucleic acid and amino acid sequence information, including the polynucleotide, gene, locus, polypeptide, and protein names, locations and sizes. In certain embodiments, the location of known coding sequences is included in the database. The sequence information may also be analyzed for unidentified ORFs and putative coding sequences. Any method can be used to identify ORFs and coding sequences including free or commercially available sequence analysis software. For example, the GLIMMER program may be used to predict putative coding regions or genes in prokaryotic nucleotide sequences. See e.g., Salzberg et al., Nucleic Acids Res. 26: 544-548 (1998); Delcher, et al., Nucleic Acids Res. 27:4636-

[0059] In certain aspects of the invention, the genome database includes the entire genomic DNA sequence of Mtb. In one embodiment, the sequence information is obtained from information that is in the public domain. In other embodiments, some or all of the sequence information can be obtained by nucleotide and/or amino acid sequencing.

[0060] As previously described in U.S. Ser. No. 10/159, 428, which is hereby incorporated by reference in its entirety, the methods of the present invention, particularly TAP technology, enable the skilled artisan to prepare a library representing all or substantially all of the polypeptides expressed in an organism or cell type. In certain embodiments of the present invention, however, it may be preferable to prepare a library of polypeptides with selected properties. Thus, one aspect of the present invention utilizes a set of ranking criteria to identify polypeptides predicted to have properties desirable e.g., for vaccine development. Polypeptide ranking criteria, which may be identified using bioinformatics tools, include but are not limited to, the presence of membrane domains, ORF size, secreted proteins signatures, signal sequences, hydrophobicity, B-cell and T-cell epitopes, homology to human proteins, protein and gene expression levels. The ranking criteria may be assigned a numerical score based on relative importance. Coding regions or putative coding regions identified in the database of Mtb sequences are then scored using the numerical ranking criteria and the sum of the scores for each sequence is used to establish a rank order. According to this aspect of the invention, primers are designed to amplify Mtbpolynucleotides in rank order. A library may be constructed, for example, from the top 5%, 10%, 20%, 30%, 40% or 50% by rank of Mtb polynucle-otides.

Amplification of Mtb Polynucleotides

[0061] Using the custom 5' and 3' oligonucleotide primers, TAP primary fragment may be amplified by methods well known in the art. The term "TAP primary fragment" refers to an amplified Mtb polynucleotide, and in one embodiment relates to a polynucleotide sequence that has been amplified but is not transcriptionally active. Generation of TAP primary fragments involves performing PCR, which generates a polynucleotide fragment that contains the Mtb polynucleotide sequence with 5'- and 3'-TAP universal end sequences and may contain other sequences incorporated into the custom 5' and 3' oligonucleotide primers. The 5'- and 3'-TAP universal end sequences are particularly useful for incorporating one or more nucleotide sequences into TAP primary fragment that confer transcriptional activity. In one embodiment, these sequences can include TAP ExpressTM promoter and terminator fragments (e.g., SEQ ID NOS: 2-7). The skilled artisan will be familiar with methods for amplifying polynucleotides, (e.g. by using PCR) and can adjust the above methods in order to optimize the amplification reaction.

[0062] An additional step in the generation of TAP fragments involves incorporating at least one polynucleotide sequence that confers transcriptional activity into the TAP primary fragment. Typically, at least one polynucleotide sequence is incorporated by performing a second PCR reaction. Examples of polynucleotide sequences that confer transcriptional activity are promoter sequences (e.g., prokaryotic Pribnow boxes and eukaryotic TATA box sequences) binding sites for transcription factors, and enhancers. In one embodiment, one promoter and one terminator sequence are added to the TAP fragment. These promoter and terminator sequences can be obtained in numerous ways. For example, one can use restriction enzyme digestion of commercially available plasmids and cDNA molecules, or one can synthesize these sequences with an automated DNA synthesizer by methods well known in the art.

[0063] The end product of the second PCR reaction is referred to as a "TAP expression fragment," which is a transcriptionally active polynucleotide, and which is generally a transcriptionally active coding sequence. In certain embodiments, the TAP expression fragments are used directly for in vivo or in vitro (e.g. cell-free) expression. In other embodiments, TAP expression fragments are transfected into cultured cells or injected into animals.

[0064] Generating TAP fragments is a rapid and efficient way of making a large number of polynucleotide sequences transcriptionally active. Accordingly, a plurality of different genes from Mtb can be made transcriptionally active using TAP technology. Thus, a library representing all, substantially all, or a selected subset of the coding sequences in the Mtb genome can be constructed using TAP technology.

TAP Tags and Linker Molecules

[0065] As described above, TAP technology provides powerful methods for amplifying and expressing Mtb polynucleotides. Coding sequences can be rendered transcriptionally active by the PCR-mediated addition of promoter sequences, enhancers, terminators and other regulatory sequences.

[0066] In addition, Mtb polynucleotides can be amplified with additional coding or non-coding sequences that can facilitate rapid screening, characterization, purification and study of the polypeptides that they encode. These additional sequences include, for example, reporter genes, affinity tags, antibody tags, PNA binding sites, secretory signals, and the like.

[0067] According to the present invention, Mtb polynucleotides can be synthesized with an epitope tag. An "epitope tag" is a short stretch of polynucleotide sequence encoding an epitope. In one embodiment, this epitope is preferably recognized by a well-characterized antibody. By incorporating an epitope tag into TAP fragments, the Mtb polynucleotide of interested can be fused in-frame to an epitope-encoding sequence. Expression of an epitope-tagged TAP fragment produces a fusion protein comprising a tagged Mtb polypeptide. Suitable epitope tags will be well known to those skilled in the art, including the hemagglutinin (HA), the 6.times.His epitope tag, and the Flag epitope tag. The HA epitope tag is well characterized and highly immunoreactive. Upon transfection of an HA-tagged TAP fragment into cells, the resulting HA-tagged polypeptides can be identified with commercially available anti-HA antibodies. Epitope tagging of TAP fragments is useful for rapidly and conveniently detecting expression of TAP fragments. Epitope tagging of TAP fragments can also help determine the intracellular distribution of Mtb polypeptides and help characterize and purify the Mtb polypeptide. Furthermore, epitope-tagged expression products can be quickly captured and/or purified using antibodies specific for the specific epitope. Antibodies directed against the HA epitope can used in the full range of immunological techniques for detection and analysis of tagged polypeptides including but not limited to Western blotting, ELISAs, radioimmune assays, immunoprecipitation, immunocytochemistry and imnuunofluorescence, fluorescence assisted cell sorting (FACS) and immunoaffinity purification of the desired fusion polypeptides.

[0068] The present invention also provides Mtb polypeptides fused to affinity tags. For example, a polynucleotide sequence encoding a histidine tag can be incorporated into the TAP fragment to enable the expressed gene product to be conveniently purified. These His tags consist of six consecutive histidine residues (6.times.His) and are a powerful tool for recombinant polypeptide purification. The 6.times. His tag interacts with metals, such as nickel. Thus, polypeptides fused to a 6.times. His tag can be purified by metal affinity chromatography, for example, using a nickel nitrilotriacetic (Ni-NTA) resin. The 6.times. His tag is much smaller than most other affinity tags and is uncharged at physiological pH. It rarely alters or contributes to polypeptide immunogenicity, rarely interferes with polypeptide structure or function, does not interfere with secretion, does not require removal by protease cleavage, and is compatible with denaturing buffer systems. Accordingly, this tag is a powerful adjunct to expression and purification of recombinant proteins.

[0069] In one aspect of this embodiment, TAP primers can be designed to include the nucleotide sequence encoding the 6.times.His epitope tag: to add the 6.times.His epitope to the 5' end of a Mtb polynucleotide, a sequence encoding histidine residues can be included along with the promoter-containing primer; to add the 6.times.His epitope to the 3' end, the His sequence can be included in the terminator-containing primer.

[0070] Commercially available nickel affinity resins can be used to purify 6.times. His tagged polypeptides. For example, the well-established QIAexpress Protein Expression and Purification Systems are based on the remarkable selectivity and affinity of patented nickel-nitrilotriacetic acid (Ni-NTA) metal-affinity chromatography matrices for polypeptides tagged with 6 consecutive histidine residues (6.times. His tag) available from QIAGEN (Seattle, Wash.). The QLAexpress System is based on the remarkable selectivity of Ni-NTA (nickel-nitrilotriacetic acid) for polypeptides with an affinity tag of six consecutive histidine residues—the 6.times. His tag. This technology allows purification, detection, and assay of almost any 6.times. His-tagged polypeptide from any expression system. Polypeptides with a 6.times. His tag can be purified through nickel nitrilotriacetic (Ni-NTA) resin

[0071] The HA and the 6.times. His epitope embodiments are not to be construed as limiting, and are provided for illustrative purposes only. Those skilled in the art will appreciate that any type of tag can be attached to the expressed products such as for example, a 7.times., 8.times., 9.times., or 10.times. histidine tag, GST tag, fluorescent protein tag, and the like.

[0072] In addition to providing a convenient means for detection and purification of Mtb polypeptides, various tags can provide a "linker" through which the polypeptides of the invention can be immobilized on a solid support. The term "linker molecule," as used herein, encompasses any molecule that is capable of immobilizing the polypeptides to a solid support.

TAP Fragment with Secretory Signal

[0073] For many gene therapy and DNA vaccine applications it may be beneficial for the gene product to be secreted from the transfected cells. Thus, one embodiment of the invention provides a version of the TAP system designed to express Mtb polypeptides containing a fused a secretory signal. A commonly used signal peptide is the first 23 amino acids from human tissue plasminogen activator (tPA) with the coding sequence as follows: ATG GAT GCA ATG AAG AGA GGG CTC TGC TGT GTG CTG CTG TGT GGA GCA GTC TTC GTT TCG CCC AGC. (SEQ ID NO: 1) This sequence can be built into the TAP promoter fragment to create a new TAP fragment in a fashion similar to the construction of the tagged polypeptides described above.

Incorporating TAP Fragments into a Plasmid Vector

[0074] Once the function or immunogenicity of an Mtb polypeptide is identified, it may be of desirable to clone the TAP fragment into a plasmid or other vector to facilitate further gene characterization and manipulation. Standard cloning techniques can involve the use of restriction enzymes to digest the plasmid and the gene fragment to be inserted. Annealing and ligation of the compatible ends can lead to insertion of the gene into the vector. An alternative method of restriction ends-directed cloning is to prepare a linearized plasmid with T overhangs on the 3' ends of the double-stranded DNA to accommodate DNA fragments amplified with the aid of specific polymerases through PCR. This method is sometimes called "T/A cloning". Other methods of cloning TAP fragments will be well known to those of skill in the art.

[0075] In certain embodiments, the TAP Cloning systems, methods, and kits can further simplify the cloning process by taking advantage of the universal 5' and 3' sequences that are present on the TAP Express fragment after the first or second PCR step. These regions overlap with the end sequences of

our linearized TAP Express Cloning Vector. When the TAP fragment and the linearized plasmid are mixed together and directly electroporated into TAP Express Electro-Comp cells, endogenous bacterial recombinase activity recombines the two DNA fragments resulting in a plasmid with the inserted TAP Express fragment. This process can replace conventional cloning with two simple PCR steps. In some embodiments it does not require cutting, pasting and ligating DNA fragments. In addition, this process can be highly suited for fast and convenient cloning of TAP PCR fragments without having to resort to restriction enzymes, DNA ligase, Topoisomerase or other DNA modifying enzymes. "TAP" systems, vectors and cells are readily available from Gene Therapy Systems, Inc., San Diego, Calif.

[0076] GeneGrip PNA compatible TAP system can also be used to couple polypeptides onto DNA through PNA-Dependent Gene Chemistry, thereby avoiding many of the limitations of previously described methodologies. GeneGrip is available through Gene Therapy Systems, Inc., San Diego, Calif. This approach takes advantage of the property of peptide nucleic acids (PNA) to hybridize with duplex DNA in a sequence specific and very high affinity manner. PNA binding sites can be used for attaching a series of peptides onto DNA in order to target the transfected plasmid and improve transgene expression, for example. This can facilitate a rational approach to improve the efficiency and efficacy of gene delivery by adding elements intended to increase nuclear uptake, facilitate endosomal escape, or target gene delivery to the cell surface or to intracellular receptors.

[0077] Incorporating a GeneGrip site into TAP enables peptide nucleic acids (PNAs) to be hybridized to the TAP gene product. Ligands can then be attached to the PNA in order to improve the bioavailability and DNA vaccine potency of the gene.

System for Performing TAP Method

[0078] In another embodiment of the invention, a system can be used to perform every step involved in generating TAP fragments from a Tb, and in particular the Mtb genome. Additionally, each individual step is capable of being controlled by a system. For example, a system can design customized PCR primers, obtain said primers, perform PCR reactions utilizing TAP technology, attach promoters and terminators, and attach sequences that encode linker molecules to the primary or expression fragment. The system can be either automated or non-automated. In one embodiment of the invention, the system comprises a computer program linked to robotic technologies for rapid and high throughput gene amplification of the genome.

Expression of the TAP Fragment

[0079] TAP fragments can be used directly as templates in various expression systems in order to obtain the corresponding polypeptide for each coding sequence in the Mtb genome. The invention provides simple, efficient methods for generating TAP fragments from Mtb that can be readily transfected into animal cells or tissues by any nucleic acid transfection technique. The methods of the invention can avoid the need for subcloning into expression vectors and for purification of plasmid DNA from bacteria. As skilled artisans can appreciate, TAP fragments can be rapidly expressed using in vivo or in vitro (e.g. cell-free) expression systems. For example, the amplified TAP fragments can be directly transfected into a

eukaryotic or prokaryotic cell for expression. Examples of eukaryotic cells that can be used for expression include mammalian, insect (e.g. Baculovirus expression systems), yeast (e.g. *Picchia pastoris*), and the like. An example of a prokaryotic cell expression system includes *E. coli*.

[0080] Alternatively, expression can be accomplished in cell-free systems, for example, a T7 transcription and translation system. Cell-free translation systems can include extracts from rabbit reticulocytes, wheat germ and *Escherichia coli*. These systems can be prepared as crude extracts containing the macromolecular components (30S, 70S or 80S ribosomes, tRNAs, aminoacyl-tRNA synthetases, initiation, elongation and termination factors, etc.) required for translation of exogenous RNA. To promote efficient translation, each extract can be supplemented with amino acids, energy sources (ATP, GTP), energy regenerating systems (creatine phosphate and creatine phosphokinase for eukaryotic systems, and phosphoenol pyruvate and pyruvate kinase for the *E. coli* lysate), and other co-factors (Mg. ²⁺, K⁺, etc.)

[0081] The use of TAP technology allows skilled artisans to rapidly express polypeptides from a plurality of polynucleotides. After a particular Mtb polynucleotide of interest is rendered transcriptionally active, other Mtb polynucleotides can also be made to be transcriptionally active according to the methods of the invention. Accordingly, in one embodiment of the invention, a plurality of polynucleotides from Mtb are amplified and expressed in order to generate a library or array of Mtb polypeptides.

[0082] Other embodiments of the invention relate to expressing the product of a Mtb polynucleotide that encodes an epitope tag, affinity tag or other tags, and which may function as linkers. A polynucleotide sequence encoding a linker molecule can be incorporated into a TAP primary fragment or a TAP expression fragment. Accordingly, the linker molecule can be expressed as a fusion to the Mtb polypeptide. [0083] The generation of Mtb polypeptide libraries according to the methods of the invention allows skilled artisans to easily use them in subsequent research and study. For example, it is possible to organize the expressed Mtb polypeptides into an array for further analysis. The expressed polypeptide arrays can be screened in order to identify, for example, new vaccine and drug targets against microbial, neoplastic disease and the like. The expressed polypeptides can be used to screen antibody libraries, to develop unique research reagents, for functional proteomic studies, and the like. These steps can be rapidly accomplished at rates far exceeding traditional methods.

Adapter Technology

[0084] In addition to amplifying Mtb polynucleotides of interest using TAP technology, the present invention also encompasses amplifying Mtb polynucleotides using "adapter technology". In some embodiments adapter technology is performed using a one-step PCR reaction. The term "adapter technology" as used herein relates to methods of cloning a desired polynucleotide into a vector by flanking a desired nucleic acid sequence, a Mtb TAP fragment for example, with first and second adapter sequences. The resulting fragment can be contacted with the vector having sequences homologous to the first and second adapter sequences under conditions such that the nucleic acid fragment is incorporated into the vector by homologous recombination in vivo in a host cell. Accordingly, adapter technology allows for fast and enzyme-less cloning of nucleic acid fragments into vectors

and can also be used for forced cloning selection for successful transformation. Adapter technology is described in more detail in U.S. patent application Ser. No. 09/836,436, entitled "Fast and Enzymeless Cloning of Nucleic Acid Fragments", U.S. patent application Ser. No. 10/125,789, entitled "Rapid and Enzymeless Cloning of Nucleic Acid Fragments", and PCT Application No. PCTUS 02/12334, all of which are hereby incorporated by reference in their entirety

[0085] The nucleic acid fragment can be incorporated into any vector using adaptor technology. In certain embodiments, the vector that the fragment is incorporated into can be, for example, a plasmid, a cosmid, a bacterial artificial chromosome (BAC), and the like. The plasmid can be CoE1, PR100, R2, pACYC, and the like. The vector can also include a functional selection marker. The functional selection marker can be, for example, a resistance gene for kanamycin, ampicillin, blasticidin, carbonicillin, tetracycline, chloramphenicol, and the like. The vector further can include a dysfunctional selection marker that lacks a critical element, and wherein the critical element is supplied by said nucleic acid fragment upon successful homologous recombination. The dysfunctional selection marker can be, for example, kanamycin resistance gene, ampicillin resistance gene, blasticidin resistance gene, carbonicillin resistance gene, tetracycline resistance gene, chloramphenicol resistance gene, and the like. Further, the dysfunctional selection marker can be, for example, a reporter gene, such as the lacZ gene, and the like. [0086] The vector can include a negative selection element detrimental to host cell growth. The negative selection element can be disabled by said nucleic acid fragment upon successful homologous recombination. The negative selection element can be inducible. The negative selection element can be, for example, a mouse GATA-1 gene. The vector can include a dysfunctional selection marker and a negative selection element.

[0087] The host cell used in adapter technology can be a bacterium. The bacterium can be capable of in vivo recombination. Examples of bacterium include JC8679, TB1, DHx, DH5, HB101, JM101, JM109, LE392, and the like. The plasmid can be maintained in the host cell under the selection condition selecting for the functional selection marker.

[0088] The first and second adapters can be any length sufficient to bind to the homologous sequences of the vector such that the desired nucleic acid sequence is incorporated into the vector. The first and second adapter sequences can be, for example, at least 11 bp, 12 bp, 13, bp, 14 bp, 15 bp, 16 bp, 17 bp, 18 bp, 19 bp, 20 bp, 21 bp, 22 bp, 23 bp, 24 bp, 25 bp, 26 bp, 27 bp, 28 bp, 29 bp, 30 bp, 31 bp, 32 bp, 33 bp, 34 bp, 35 bp, 36 bp, 37 bp, 38 bp, 40 bp, 50 bp, 60 by and the like. Furthermore, the first and second adapter sequences can be greater than 60 bp.

[0089] The first and second adapter sequences further can include a functional element. The functional element can include a promoter, a terminator, a nucleic acid fragment encoding a selection marker gene, a nucleic acid encoding a linker molecule, a nucleic acid fragment encoding a known protein, a fusion tag, a nucleic acid fragment encoding a portion of a selection marker gene, a nucleic acid fragment encoding a growth promoting protein, a nucleic acid fragment encoding a transcription factor, a nucleic acid fragment encoding an autofluorescent protein (e.g. GFP), and the like. [0090] When the common sequences on both the 5' and 3' ends of the nucleic acid fragment are complimentary with terminal sequences in a linearized empty vector, and the

fragment and linearized vector are introduced, by electroporation, for example, together into a host cell, they recombine resulting in a new expression vector with the fragment directionally inserted. In alternative embodiments the host cell can include the linearized empty vector so that only the nucleic acid fragment is introduced into the host cell. It should be noted that in alternative embodiments of the present invention the vector can be circularized, and as used herein a vector can be either linearized or circular. The host cell is converted into an expression vector through homologous recombination. In principle this approach can be applied generally as an alternative to conventional cloning methods.

[0091] A nucleic acid fragment having first and second adapter sequences can be generated by methods well known to those of skill in the art. For example, a gene of interest with known 5' and 3' sequences undergoes PCR along with overlapping 5' and 3' priming oligonucleotides. The priming oligonucleotides can be obtained by methods known in the art, including manufacture by commercial suppliers. A primary fragment with adapter sequences can be generated. The adapter sequences flanking the gene of interest can be homologous to sequences on a vector or to sequences from other 5' or 3' fragments to be used in a subsequent PCR

[0092] In some embodiments of the invention, a particular polynucleotide of interest from Mtb can be amplified with an adapter sequence on both the 3' and 5' ends. In other embodiments adapters can be attached to a plurality of polynucleotides, for example every coding region in the Mtb genome. In certain embodiments adapters can make the desired coding regions transcriptionally active. Once incorporated into the desired vector, the Mtb coding region can be rapidly replicated and expressed, such that a plurality of Mtb's genes, for example every gene, is expressed.

[0093] Pluralities of expression products can be stored in libraries or arrays and can be assayed for their immunogenic properties as will be discussed below. While most embodiments relating to the assay methodologies are discussed in terms of TAP technology, all of the following assays can be used on adapter technology expression products as well. Once the appropriate assays are conducted on the adapter technology expression products, methods of developing vaccines can be utilized. While most of the embodiments relating to developing vaccines, discussed below, pertain to TAP technology, all of the vaccine embodiments can also be used with polypeptide libraries and arrays resulting from adapter technology.

Identifying Immunogenic Polypeptides

[0094] Libraries and arrays of polypeptides, prepared through TAP or adapter technology with subsequent expression can be useful in the development of polypeptide or nucleic acid subunit vaccines. DNA vaccines are effective vaccines that are inexpensive to manufacture, easy and safe to deliver, and can be widely distributed. It has been found that plasmid DNA, when injected into mice without being associated with any adjuvant, can generate antibody and CTL responses to viral antigens encoded by the plasmid DNA, and elicit protective immunity against viral infection (Ulmer at al., Science, 259:1745, 1993). Starting from this, there have been reported many research results regarding the induction of humoral and cellular immune responses resulting from the introduction of DNA vaccines containing various viral genes in animal models (Chow et al., J. Virol., 71:169, 1997; McClements et al., Proc. Natl. Acad. Sci. USA, 93:11414,

1996; Xiang et al., Virology, 199:132, 1994; Wang et al., Virology, 211:102, 1995; Lee et al., Vaccine, 17:473, 1999; Lee et al., J. Virol., 72:8430, 1998). As well, DNA delivery by electroporation techniques has been well-described (Heller et al., Expert Opin. Drug Deliv. 2(2): 1-14, 2005.

[0095] One of the most difficult tasks in developing a DNA vaccine (or any recombinant subunit vaccine) to a pathogen such as Mtb, is the identification of antigens that can stimulate the most effective immune response against the pathogen, particularly when the genome of the organism is large.

[0096] A comprehensive means to accomplish this task, which is embodied by the present invention, is to obtain a plurality of polypeptides from the particular pathogen in the mode of a library or array. These polypeptides can be tested to determine their capability to evoke a humoral and/or a cell-mediated immune response. Polypeptides that evoke immunogenic responses can be tested individually or with other antigens for effectiveness as subunit vaccines. In addition, nucleic acids that encode identified antigenic polypeptides can be used alone or with other nucleic acids that encode antigens to develop a recombinant vaccine, such as a DNA vaccine, for the particular pathogen.

Mtb Scanning

[0097] One embodiment of the invention, incorporates a Rapid High-Throughput Vaccine Antigen Scanning approach, using TAP Express, that is able to systematically screen and identify all, substantially all, or a subset of the antigens in Mtb that give rise to a humoral and cell-mediated immune response. The identification of the Mtb antigens allows for the development of a highly specific subunit vaccine

[0098] FIG. 2 illustrates a method for amplifying multiple Mtb polynucleotides using TAP technology, expressing the gene products of the resultant TAP fragments, purifying, and quantifying the resulting polypeptides. FIG. 2 further illustrates a method of preparing polypeptides, which can be assayed to identify their ability to evoke a cell-mediated or humoral immune response

[0099] In certain methods of developing a Mtb vaccine, a plurality of Mtb polynucleotides can be made transcriptionally active. In one embodiment, all of the open reading frames from Mtb genome can be made transcriptionally active using TAP technology. The present invention thus provides Mtb polynucleotides (SEQ ID NOS: 46-64, 110-121) that have been made transcriptionally active.

[0100] The resulting Mtb TAP fragments of the present invention can be purified and expressed in vitro or in vivo according to any method known in the art. The expression products, which encompass SEQ ID NOS: 65-83, 122-133, can be assayed by various methods to determine their ability to evoke a humoral and/or a cell-mediated immunogenic response. Polypeptides that are identified as capable of evoking an immune response can be used as candidates to develop polynucleotide or polypeptide subunit vaccines. The complete method will be described in more detail below.

[0101] According to one embodiment of the present invention, TAP fragments from Mtb are used to generate a DNA array, and then, if desired, a protein array. In certain embodiments, primers are designed for every gene in the Mtb genome. In another embodiment, designing the primers allows a skilled artisan to make any given Mtb polynucleotide transcriptionally active using TAP technology. In yet another embodiment, coding regions, ORFs and other polynucleotide

sequences of interest are ranked according to and the top Mtb polynucleotides are made transcriptionally active using TAP technology.

[0102] As mentioned above, the custom PCR primers can be designed by using an automated system, such as a computerized robotics system. For example, in order to design custom primers for use in the TAP process, a robotic workstation can be interfaced with a dual Pentium III CPU (1.4 GHz) computer running the Linux operating system. In addition, a customized MySQL database can manage the input sequence data from GenBank and from other sources. This database can track all the operations, samples and analytical data generated by the robot. In another embodiment, PCR primers, PCR products and polypeptides can be tracked by the database. For example, PCR primers, PCR products and polypeptides can be tracked by using bar coded 96-well plates. While the embodiments below discuss using 96-well plates in certain embodiments, those skilled in the art can appreciate that any sized well plate can be used. For example, the well plates can consist of about 48, about 96, about 144, about 192, about 240, about 288, about 336, about 384, about 432, about 480, about 576, about 672, about 768, about 864, about 960, about 1056, about 1152, about 1248, about 1344, about 1440, about 1536 or more wells. In addition to well plates, the PCR products and polypeptides can be tracked using any suitable receptacles, for example test tubes.

[0103] Custom oligonucleotide pairs of the present invention (SEQ ID NOS: 8 and 9; 10 and 11; 12 and 13; 14 and 15; 16 and 17; 18 and 19; 20 and 21; 22 and 23; 24 and 25; 26 and 27; 28 and 29; 30 and 31; 32 and 33; 34 and 35; 36 and 37; 38 and 39; 40 and 41; 42 and 43; 44 and 45; 86 and 87; 88 and 89; 90 and 91; 92 and 93; 94 and 95; 96 and 97; 98 and 99; 100 and 101; 102 and 103; 104 and 105; 106 and 107; 108 and 109), which are needed for the TAP PCR reactions, can be synthesized or obtained in order to perform the TAP technology. In certain embodiments, the Mtb genome sequence data and primer design software (e.g., Primer 3) can be used by the system to generate custom primer pairs for all, substantially all, or a subset of the genes in the Mtb genome. The primers can be organized into arrays of about 48, about 96, about 144, about 192, about 240, about 288, about 336, about 384, about 432, about 480, about 576, about 672, about 768, about 864, about 960, about 1056, about 1152, about 1248, about 1344, about 1440, or about 1536 5' primers and 3' primers according to polynucleotide size and GC content, such PCR reaction conditions can be optimized on a plate by plate basis. The present invention further contemplates that sequences for each of the custom Mtb primer pairs can be sent to an oligonucleotide synthesis provider (e.g., MWG Biotech, Inc., High Point, N.C.) where they can be synthesized. Synthesized primers can be organized and dispensed into bar-coded plates at a desired concentration, such as 100 pmole/.mu.l, frozen and shipped to the practitioner. In one embodiment, 600 Mtbspecific PCR primers, which are capable of amplifying 300 Mtb coding sequences are designed, generated, ordered, and

[0104] After obtaining or generating the custom Mtb PCR primers, the Mtb polynucleotides of interest can be amplified. In one embodiment, the primers can be organized into arrays of 96 5' primers and 96 3' primers according to polynucleotide size, and placed onto a robotic workstation. The robot can be programmed to generate a plate of about 48, about 96, about 144, about 192, about 240, about 288, about 336, about 384, about 432, about 480, about 576, about 672, about 768, about

864, about 960, about 1056, about 1152, about 1248, about 1344, about 1440, about 1536 PCR reactions by mixing the appropriate 5' and 3' primers with Taq polymerase and Mtb genomic DNA. In addition to Taq, any thermally stable polymerase can be used in the PCR reactions. For example, Vent, Pfu, Tfl, Tth, and Tgo polymerases can be used. The robotic workstation can transfer the PCR reaction plate containing the mixed reagents to a PCR machine for amplification. In one embodiment, the robotic workstation can use a robotic arm to transfer the PCR reaction plate to the PCR machine.

[0105] The first TAP PCR procedure can be run for any number of cycles. In one embodiment, the PCR machine is run for about 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40, or more cycles. The first TAP PCR reactions can be transferred robotically to a Millipore Montage 96-well cleanup kit, for example, when desired. Any method, kit or system can, however, be used to purify the PCR products from these reactions. According to one embodiment, a vacuum station of the robotic platform can carry out the purification step. In some embodiments, an aliquot of the resulting product can be transferred robotically to an analysis plate containing the Pico-Green fluorescent probe (Molecular Probes, Eugene, Oreg.) that reacts only with the dsDNA products. Depending on the number of wells, the plate can be transferred to an about 48, about 96, about 144, about 192, about 240, about 288, about 336, about 384, about 432, about 480, about 576, about 672, about 768, about 864, about 960, about 1056, about 1152, about 1248, about 1344, about 1440, about 1536 or more well fluorescent plate reader. The fluorescent signal can be compared to a standard curve to determine the amount of double stranded PCR product generated in this first PCR step. Persons with skill in the art can adjust the above methods in order to optimize their particular PCR reaction, should the need arise.

[0106] In addition to the first TAP PCR procedure, a second TAP PCR reaction can be performed to add at least one sequence that confers transcriptional activity to the primary TAP primary fragment. In one embodiment, a robot can be programmed to transfer an aliquot of each TAP primary fragment from the first TAP PCR reaction into a PCR reaction containing a promoter- and a terminator-containing primers. In a particular embodiment, the promoter can be a T7-His tag terminator sequence and the terminator can be a T7-His tag terminator sequence. Those with skill in the art can appreciate that any promoter or terminator sequence can be added to the primary transcript. In addition, any polynucleotide sequence that encodes a tag or linker allowing the expressed polypeptide to be detected or purified is also contemplated.

[0107] Like the first TAP PCR reaction, the second TAP PCR reaction can be run for any desired number of cycles. In one embodiment, the second TAP PCR reaction is run for about 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 36, 37, 38, 39, 40 cycles or more. Furthermore, any type of thermally stable polymerase can be used for the second TAP PCR reaction. In a particular embodiment the polymerase can be Taq. In some embodiments Vent, Pfu, Tfl, Tth, and Tgo polymerases can be used. The resulting TAP Express PCR fragments from the second PCR reaction can be cleaned by any kit, method or system. A particular kit that can be used to clean the resulting TAP fragments is a Millipore Montage 96-well cleanup kit. Additionally, as discussed above, the level of PCR product recovered can be determined using any detection agent, for example, Pico-Green.

[0108] The resulting TAP fragments can be expressed by using any method of gene expression. In one embodiment, the TAP fragments can be expressed using in vivo or in vitro (e.g. cell-free) systems. For example, the fragments can be directly transfected into any eukaryotic or prokaryotic cell for expression. Examples of eukaryotic cells that can be used for expression include mammalian, insect, yeast, and the like. An example of a prokaryotic cell expression system includes E. Coli. The TAP fragments can also be expressed by a cell-free system. According to one embodiment of the invention, the resulting TAP fragments can be expressed in a high-throughput cell-free expression machine, such as, for example, the Roche RTS (Rapid Translation System)-100. In a further embodiment, the TAP fragments can be incubated in Roche RTS100 system at 30° C. for 5 hours. A person with skill in the art can readily appreciate the utility in following the particular cell-free translation machine's instructions. If a T7-histadine promoter or terminator fragment is added to a primary transcript, translation of the TAP fragment can result in histidine tagged polypeptides, which can be purified as discussed below. As discussed herein, any tag can be used.

[0109] The expressed Mtb polypeptides can be purified using any purification method for purifying expressed polypeptides. In one embodiment histidine tagged polypeptides can be purified with Qiagen nickel columns, such as Ni-NTA Superflow 96 Biorobot Kit. A person with skill in the art can readily appreciate the utility in following the instructions of the particular polypeptide purification system. Other methods that can be used to purify polypeptides include ultrafiltration, extraction, and chromatography.

[0110] The identity, quantity and purity of the purified Mtb polypeptides can be verified by SDS gel electrophoresis. According to one embodiment of the invention, MALDI-TOF MS (Matrix Assisted Laser Desorption/Ionization-Time of Flight Mass Spectrometry) can be employed to confirm the fidelity of the purified polypeptides. According to this embodiment, aliquots of each polypeptide (1-2.mu.g) can be aliquoted into about 48, about 96, about 144, about 192, about 240, about 288, about 336, about 384, about 432, about 480, about 576, about 672, about 768, about 864, about 960, about 1056, about 1152, about 1248, about 1344, about 1440, about 1536 or more well plates and digested with modified trypsin. The resulting material can be mixed with matrix (alpha-cyano-4-hydroxycinnamic acid (CHCA)) and spotted onto any target plate with a suitable number of spots, for example, 48, about 96, about 144, about 192, about 240, about 288, about 336, about 384, about 432, about 480, about 576, about 672, about 768, about 864, about 960, about 1056, about 1152, about 1248, about 1344, about 1440, about 1536 or more spots. In one embodiment, a 384-spot "anchor chip" target plate (Bruker Daltonics, Billerica, Mass.) can be used. The plate can be transferred to the sample stage of a Bruker Autoflex MALDI-TOF mass spectrometer. The spectrometer can be set up to automatically scan the plate and search the Mascot polypeptide database via the Internet. Accordingly, a very rapid verification system can verify purity, identity, and quantity in less than a day, for example, depending on the amount of polypeptides. Purified Mtb polypeptides can be placed in libraries or organized into arrays for subsequent testing and analysis.

Humoral Immune Response

[0111] Use of the Mtb polypeptide libraries and arrays prepared, for example, according the methods above (e.g. using

TAP or adapter technology) can be used to identify antigenic targets of humoral immunity in Mtb non-human animals and human patients. A humoral immune response relates to the generation of antibodies and their ability to bind to a particular antigen. In general, the humoral immune system uses white blood cells (B-cells), which have the ability to recognize antigens, to generate antibodies that are capable of binding to the antigens.

[0112] In one embodiment, the Mtb polypeptides of the invention are generated according to the methods described above. In certain aspects of this embodiment additional polynucleotide sequences that encode linker molecules are added to the TAP primary fragment or the TAP expression fragment such that the expressed Mtb polypeptides are fused to a linker molecule. As discussed previously, the term "linker molecule" encompasses molecules that are capable of immobilizing the polypeptides to a solid support.

[0113] In a particular embodiment, a Mtb polynucleotide of interest is fused to a HA epitope tag such that the expressed product can include the Mtb gene product fused to the HA epitope. In another embodiment, a Mtb polynucleotide of interest is combined with a histidine (His) coding sequence, such that the expressed product can include the Mtb gene product and a 6.times., 7.times., 8.times., 9.times., or 10.times.histidine tag. In other embodiments a Mtb polynucleotide is combined with a sequence that codes for a GST tag, fluorescent protein tag, or Flag tag. Using these methods it is possible to express and tag every Mtb polypeptide encoded by its genome. In another embodiment, the tagged Mtb polypeptide can be attached to a solid support, such as a 96-well plate. The immobilize polypeptides can be contacted with an antiserum or other fluid containing antibodies from an animal that has been immunized with one or more antigens from Mtb. In one embodiment, ELISA and Western blot assays are performed in parallel to detect the presence of immunogenic Mtb polypeptides.

[0114] As an example of an ELISA assay, tagged Mtb polypeptides can be immobilized on a solid support, such as a 96-well plate. The immobilized Mtb polypeptides are then incubated with serum from an animal that has been immunized with one or more antigens from Mtb, or has been infected directly with Mtb by inoculation, aerosol delivery, or the like. The reaction mixture can be washed to remove any unbound serum antibodies. The ability of the serum antibodies to bind to the bound Mtb polypeptides can then be detected using any one of a number of methods. For example, enzyme linked secondary antibodies can be added to detect the presence of an antigen specific antibody. Any enzyme linked secondary antibody can be used in this invention, depending on the source of the serum. For example, if vaccinated mouse serum is used to provide the primary antibody, enzyme linked anti-mouse antibody can be used as a secondary antibody. Likewise if human serum is used to provide the primary antibody, enzyme linked anti-human serum can be used as a secondary enzyme.

[0115] Any suitable assay can be used to determine the amount of bound polypeptide specific antibody. Also, skilled artisans can develop the enzyme assay to determine the amount of polypeptide specific antibody that is bound. In one embodiment, the readout from an assay can show the presence of different levels of antibody in each of the 96 wells. For example, while some Mtb polypeptides are not able to elicit any serum antibodies, other Mtb polypeptides can elicit intermediate levels of antibodies, and some can elicit high anti-

body levels. In one embodiment, polypeptides that generate high antibody titers can be further researched to determine which polypeptides are present on the surface of the virus. In a particular embodiment of the invention Mtb polypeptides that generate high antibody titers and that are located on the surface of the virus are candidates for use in the development of a subunit Mtb vaccine.

[0116] In addition, serodiagnostic tests may be developed using antigens identified and characterized by these methods. That is, the peptide (epitopes) identified herein find use in detecting antibodies in serum from Mtb infected or exposed organisms, animals or patients.

[0117] FIG. 3 demonstrates one embodiment of determining the humoral immune response generated by an array of polypeptides. One of skill in the art may deviate in certain details from those shown in FIG. 3. For example, the HA tag, or any other tag as described above, may be placed at either the C-terminal or N-terminal end of the polypeptide to insure that epitopes are not concealed due to binding to the plate. Instead of HA tagged polypeptides, a histidine tag can be used, and the polypeptides can be bound to nickel coated plates. For example a 6.times., 7.times., 8.times., 9.times., or 10.times.histidine tag can be used. Alternatively, histidine tagged polypeptides can be purified from either transfected cells or from the in vitro transcription translation system. Furthermore, purified Mtb polypeptides can be attached nonspecifically to polypeptide-absorbing plates such as Immulon plates, for example.

[0118] In one aspect of the present invention, highly immunogenic Mtb antigens are detected by comparing the results of Western blotting analysis with ELISA. Western blotting and ELISA are two independent yet complementary methods that may be used to detect immunogenic Mtb in qualitative and quantitative ways. Western blotting is often used to examine the quality of a polypeptide or protein sample, including such parameters as purity, protein integrity, and degradation. Western blotting detects polypeptides in their denatured form. In one aspect of this embodiment, ELISA, which detects native polypeptides, is used to further examine Western-positive Mtb polypeptides in a more quantitative fashion, to illustrate the strength of the Mtb epitope's immunogenicity

Cell-Mediated Immune Response

[0119] Use of the TAP-expressed Mtb polypeptide libraries and arrays prepared according the methods above (e.g. using TAP or adapter technology) can also be exploited to identify the highly immunogenic targets of cell-mediated immunity in Mtb vaccinated non-human animals. In contrast to a humoral immune response, where an antibody binds directly binding to an antigen, a cell-mediated immune response relates to T-cells binding to the surface of other cells that display the antigen. When certain T-cells come into contact with a presented antigen, they produce and release cytokines such as interferon-.gamma. (IFN-.gamma.) or Tumor Necrosis Factor-alpha (TNF-.alpha.). Cytokines are cellular signals that can alter the behavior or properties of another cell. For example, cytokines may inhibit viral replication, induce increased expression of WIC class I and peptide transporter molecules in infected cells, or activate macrophages. Accordingly, cytokines released by T-cells, associated with the binding to an antigen, can be used to identify and detect T-cell/ antigen interactions.

[0120] Some cells have WIC molecules on their membranes to present antigens to T-cells. Efficient T-cell function

relies on proper recognition of the WIC-antigen complex. There are two types of WIC molecules: Class I and Class II. The two different classes of WIC molecules bind peptides from different sources inside the cell for presentation at the cell surface to different classes of T-cells. Any T-cell can be used in the present invention, and include for example both CD4+ and CD8+ T-cells. CD8+ cells (cytotoxic T-cells) bind epitopes that are part of class I WIC molecules. CD4+ T-Cells, which includes inflammatory CD4 T-cells and helper CD4 T-cells, bind epitopes that are part of class II MI-IC molecules. Only specialized antigen-presenting cells express class II molecules.

[0121] There are three main types of antigen-presenting cells: B cells, macrophages and dendritic cells. Each of these cell types is specialized to process and present antigens from different sources to T-cells, and two of them, the macrophages and the B cells, are also the targets of subsequent actions of armed effector T-cells. These three cell types can express the specialized co-stimulatory molecules that enable them to activate naive T-cells, although macrophages and B cells express those molecules only when suitably activated by infection.

[0122] Embodiments of the present invention relate to detecting Mtb polypeptides capable of evoking a cell-mediated immune response in order to identify potential candidates for use in a subunit vaccine or other pharmaceutical composition. According to one method of detecting a cellmediated immune response, an Mtb polypeptide is delivered to an antigen-presenting cell where it can be presented in a manner that is recognized by antigen specific T-cells. In another embodiment of the invention, a transcriptionally active gene can be delivered to an antigen-presenting cell where expressed and presented in a manner that can be recognized by antigen specific T-cells. Mtb antigen specific T-cells can be acquired from numerous sources. For example, animals that have been infected, or immunized with one or more antigens from Mtb virus are a good source of antigen specific T-cells. Alternatively, human Mtb patients and volunteers immunized with Mtb can be a source of antigen specific T-cells.

[0123] FIG. 4 demonstrates one embodiment of determining the cell-mediated immune response generated by an array of polypeptides. One of skill in the art may deviate in certain details from those shown in FIG. 4.

[0124] In order to test the ability of Mtb polypeptides to elicit a cell-mediated response, a plurality of Mtb polynucleotides can be amplified and made transcriptionally active using TAP technology. In one embodiment about 10, about 20, about 30, about 40, about 50, about 60, about 70, about 80, about 90, about 100, about 110, about 120, about 130, about 140, about 150, about 160, about 170, about 180, about 190, about 200, about 210, about 220, about 230, about 240, about 250, about 260, about 266 Mtb polynucleotides are made transcriptionally active using TAP technology.

[0125] In one embodiment, transcriptionally active Mtb polynucleotides can be transfected into an antigen-presenting cell and expressed within the cell. In another embodiment, instead of transfecting the genes into an antigen-presenting cell, the Mtb TAP fragments can be expressed in an in vivo or in vitro (cell-free) expression system and the expressed polypeptide can be delivered into the antigen-presenting cell. The polypeptide can be delivered into the antigen-presenting cell according to any method. In one embodiment, the polypeptide can be delivered using the technology described

in U.S. patent application Ser. No. 09/738,046, entitled "Intracellular Protein Delivery Reagent" and U.S. patent application Ser. No. 10/141,535, entitled "Intracellular Protein Delivery Compositions and Methods of Use," both of which are hereby incorporated by reference in their entirety. The reagents described therein are capable of delivering any type of polypeptide into any type of cell. Furthermore, the results of FIG. 5 demonstrate that dendritic cells can present antigens to T-cells supplied from an immunized host after antigenic polypeptides were delivered to the dendritic cells with reagents from the above mentioned applications.

[0126] In certain embodiments of the invention, reagents used to deliver polypeptides into cultured cells can be a cationic lipid formulation. In one embodiment, these reagents can deliver fluorescently labeled antibodies, high and low molecular weight dextrans, phycoerythrin-BSA, caspase 3, caspase 8, granzyme B, and .beta.-galactosidase into the cytoplasm of a variety of different adherent and suspension cells. Caspases delivered to cells with are functional, since they can be shown to send cells into apoptosis. In one embodiment, Mtb polypeptides are delivered into dendritic cells using these reagents.

[0127] Detecting a T-cell's ability to bind to an antigenpresenting cell, after the antigen-presenting cell has processed a particular polypeptide, is useful in determining whether the particular polypeptide evokes a cell-mediated immune response. Once a particular polypeptide is delivered into or expressed in the antigen-presenting cell, an assay can be performed to identify T-cell interaction with the WICantigen complex. In one embodiment, it can be determined if T-cells obtained from an animal that was immunized with Mtb can bind to a particular antigen presented by an antigenpresenting cell. For example, an ELIspot assay (Enzyme-Linked Immuno spotting; ELIspot) can be performed to identify antigen specific T-cells. Similar immunoassays can be performed to identify Mtb antigens (presented by an antigenpresenting cells) that stimulate T-cells from active Mtb patients or immunized individuals.

[0128] One method of detecting a T-cell/antigen interaction is to measure the amount of a particular cytokine released by the T-cell when it interacts with a WIC-antigen complex. The skilled artisan can appreciate that other cellular signals can be used to indicate a cell-mediated immune response. In one embodiment, the levels of IFN-.gamma. released by T-cells can indicate whether a particular peptide is capable of evoking a cell-mediated immune response. In a particular embodiment, an antibody specific for IFN-.gamma. can be coated onto a solid support. Unbound antibodies can be washed away and IFN-.gamma. obtained from the supernatant containing T-cells plus antigen-presenting cells or antigen transduced antigen-presenting cells, can be added to the wells. A biotinylated secondary antibody specific for IFN-y can be added. Excess secondary antibody can be removed and Streptavidin-Peroxidase can be added to the mixture. Streptavidin-Peroxidase is capable of binding to the biotinylated antibody to complete the four-member immunoassay "sandwich." Excess or unbound Streptavidin-Peroxidase is easily removed from the mixture. In order to detect amount of bound Streptavidin-Peroxidase, a substrate solution can be added which reacts with the Streptavidin-Peroxidase to produce color. The intensity of the colored product is directly proportional to the concentration of IFN-.gamma. present in the T-cell/antigen-presenting cell supernatant. Kits for performing these types of immunoassay are readily available from many commercial suppliers or the necessary reagents composing such kits can be purchased separately or produced in-house. In one embodiment, processed and presented Mtb polypeptide that evokes T-cells to produce a high level of IFN-gamma. can be considered a strong candidate for use in developing a subunit vaccine.

[0129] Those with skill in the art will appreciate that other methods can be used to detect T-cell/Antigen interactions. These methods include bead based assays, flow-based assays, RT-PCR based assays, cytokine ELISAs, lymphoproliferation assays, cytotoxic T cell assays, or any other assay that can detect the interaction of a T-cell with a responder cell (e.g. macrophage).

Developing a Subunit Vaccine, Pharmaceutical Composition, or Immunogenic Composition

[0130] A particular Mtb polypeptide that has been identified to elicit a humoral or cell-mediated immune response, can be further explored to determine its ability to be used in a subunit vaccine, pharmaceutical composition, or immunogenic composition. The terms "subunit vaccine," "DNA vaccine," "recombinant vaccine" and "immunogenic composition" encompass vaccines that are comprised of polypeptides, nucleic acids or a combination of both. Further exploration of a Mtb polypeptide vaccine candidate includes testing the Mtb polypeptide or nucleic acid encoding the Mtb polypeptide in a large number of animal subjects, volunteers or patients. In a particular embodiment, surface antigens can be studied closely because of the likelihood that they can inhibit virus infectivity. In one embodiment, every polypeptide encoded by the Mtb genome is assayed to determine its immunogenic effect. Polypeptides that elicit an immune response, whether cell-mediated or humoral, can be more closely studied to determine potential use alone or in conjunction with other polypeptides and genes as a subunit vaccine, pharmaceutical composition, or immunogenic composition. Suitable methodologies for electing and detecting an immune response are well established in the art.

Uses of Vaccine Compositions

[0131] As noted previously, the present invention provides peptide immunogens and nucleic acids encoding the immunogens. As such, the present invention also provides methods of using the immunogens to generate an immune response in a mammalian host.

[0132] Methods of generating immune responses in a host are known in the art. However, according to the present invention, the method includes administering to the host an immunogenic composition. The immunogenic composition includes at least one nucleic acid selected from SEQ ID NO: 46-64 and/or 110-121. In addition, fragments of these sequences can be used. Also, it should be noted that combinations of these sequences may be used to generate an immune response against Mtb. When using nucleic acids to generate an immune response the nucleic acids preferably encode peptides found in SEQ ID NO: 65-83 and/or 122-133. In addition, fragments of these sequences can be used. Also, combinations of these sequences can be used.

[0133] When combinations of the above immunogenic compositions are to be used at least 2, 3, 4 or 5 or more of the nucleic acids or fragments thereof can be combined to generate an immunogenic composition. Any combination of the nucleic acids finds use in this method.

[0134] Also, methods of generating an immune response include administering to the host at least one peptide selected from the peptides found in SEQ ID NO: 65-83 and/or 122-133. In addition, fragments of these sequences can be used. Also, it should be noted that combinations of these sequences may be used to generate an immune response against Mtb.

When combinations of the above immunogenic compositions are to be used at least 2, 3, 4 or 5 or more of the nucleic acids or fragments thereof can be combined to generate an immunogenic composition. Any combination of screened nucleic acids finds use in this method.

Kits

[0135] Various nucleic acids and peptides have been identified that generate an immune response. As such, the nucleic acids and peptides find use in kits. The kits of the invention are useful for a variety of applications including combining reagents necessary for producing vaccine compositions. Such vaccine compositions include the polypeptides and polynucleotides described herein as well as carriers, diluents and other pharmaceutically acceptable carriers. It should be noted, as described above, that the kits may include fragments of the nucleic acids or peptides described herein as well as combinations of the nucleic acids and/or peptides described herein. Preferably the kits include at least 2, 3, 5, 10, 15, 20, 25, 30 or more nucleic acids or peptides described herein. Any combination of the nucleic acids or peptides can be used. In addition, the kits may include adjuvants. In addition, the kits may include instructions for preparing and administering the

[0136] In addition, the kits of the invention find use as diagnostic kits. In particular, the kits find use as serodiagnostic kits. As such, the kits include at least one peptide as described herein. Preferably, however, the kits include a plurality of peptides, such as at least 2, 3, 5, 10, 15 or 20 or more peptides for diagnosis of Mtb infection or exposure of an organism, animal or patient.

[0137] In some embodiments, the nucleic acids encoding the polypeptides find use in diagnostic kits. The nucleic acids encoding the antigenic peptides find use as probes to detect complementary nucleic acids of Mtb. However, in an alternative embodiment the kits include the polypeptides produced from the in vitro transcription-translation reaction find use in detecting antibodies from an organism, animal or patient exposed to Mtb.

EXAMPLES

Example 1

Procedure for Generating Histidine Tagged TAP Express Fragments

[0138] A detailed procedure that is used to produce tagged T7-TAP Express fragments is as follows: 96 different genes were amplified from a mixture of plasmid templates. A first PCR reaction was run with customized 5' and 3' primers. The 5' primers contained between 43-48 bases. In particular, the T-7-His TAP ends contained 28 bases while the gene-specific component contained between 15-20 bases. The 3' primers contained between 45-50 bases. Specifically, the T7-terminator TAP ends contained 30 bases while the gene specific component contained between 15-20 bases. The reaction temperature and times for the first PCR reaction were: 94° C. for 2 minutes, followed by 28 cycles of: 94° C. for 20 seconds, 58° C. for 35 seconds, and 70° C. for 2 minutes (for genes that contained more than 2 kb, 1 minute was added for each kb). [0139] After the first PCR reaction was performed, an aliquot of each PCR reaction from the previous step was transferred into a PCR reaction containing the T7-histidine promoter fragment and T7 terminator fragment. The T7 promoter primer contained 25 bases, while the T7-promoter-His tag fragment contained a 104 base EcoRV/BglII fragment. The T7-terminator fragment was a 74 base oligonucleotide. The reaction temperature and times for the second PCR reaction were: 94° C. for 2 minutes, followed by 30 cycles of: 94° C. for 20 seconds for 20 seconds, 60° C. for 35 seconds, and 70° C. for 2 minutes (for genes that contained more than 2 kb, 1 minute was added for each kb).

Example 2

Using the Mtb Proteome to Identify the Antigenic Targets of Humoral Immunity in Mtb Mice and Humans

[0140] The following is a method used to systematically screen and identify antigens in Mtb that give rise to a protective humoral immune response. A bioinformatics approach was used to order the *M. tuberculosis* polynucleotide sequences for amplification. The Mtb genome was first analyzed for hydrophobicity by the method of Doolittle. Hydrophilic polynucleotides sequences were then further grouped by size. Hydrophilic open reading frames/coding regions longer than 500 by were selected for TAP amplification. Initially, three hundred Mtb genes were synthesized by TAP and about 100 proteins were translated and purified in arrays; as described below.

TAP PCR

[0141] The PCR reactions were performed such that a nucleotide sequence encoding a 6.times. His tag was fused to these amplified transcriptionally active genes. The resulting His tagged TAP fragments were expressed to produce .about. 100 Mtb polypeptides containing the His tag.

[0142] A detailed procedure that was used to produce tagged T7-TAP Express fragments is as follows: groups of 96 Mtb polynucleotide sequences were amplified from Mtb genomic DNA. A first PCR reaction was performed using customized 5' and 3' primers, as shown in Table 1 (SEQ ID NOS: 8 and 9; 10 and 11; 12 and 13; 14 and 15; 16 and 17; 18 and 19; 20 and 21; 22 and 23; 24 and 25; 26 and 27; 28 and 29; 30 and 31; 32 and 33; 34 and 35; 36 and 37; 38 and 39; 40 and 41; 42 and 43; 44 and 45; 86 and 87; 88 and 89; 90 and 91; 92 and 93; 94 and 95; 96 and 97; 98 and 99; 100 and 101; 102 and 103; 104 and 105; 106 and 107; 108 and 109). The 5' primers contained between 43-48 bases. In particular, the T-7-His TAP ends contained 28 bases while the gene-specific component contained between 15-20 bases. The 3[†] primers contained between 45-50 bases. Specifically, the T7-terminator TAP ends contained 30 bases while the gene specific component contained between 15-20 bases.

TABLE 1

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

Rv2031c HEAT SHOCK PROTEIN HSPX (ALPHA-CRSTALLIN HOMOLOG) 14 kDa ANTIGEN) (HSP16.3)

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATATGGCCACCACCCTT (SEQ ID NO: 8)

3' primer:

TGATGATGAGAACCCCCCCCGTTGGTGGACCGGATCTGAA (SEQ ID NO: 9)

Polynucleotide sequence: ATGGCCACCACCCTTCCCCGTTCAGCGCCACCCGCGGTCCTCTTCCCCGAGTTTTCTGAGCTGTTCGCGGGCCTTCCCGTCATTCGCCGGACTCCCGGCCCA

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

Amino acid sequence:

MATTLPVQRHPRSLFPEFSELFAAFPSFAGLRPTFDTRLMRLEDEMKEGR YEVRAELPGVDPDKDVDIMVRDGQLTIKAERTEQKDFDGRSEFAYGSFVR TVSLPVGADEDDIKATYDKGILTVSVAVSEGKPTEKHIQIRSTN (SEQ ID NO: 65)

RV3763 19 KDA LIPOPROTEIN ANTIGEN PRECURSOR LPQH

5' primer

GAAGGAGATATACCATGCATCATCATCATCATGTGAAGCGTGGACTG (SEO ID No: 10)

3' primer

TGATGATGAGAACCCCCCCGGAACAGGTCACCTCGATTT (SEO ID NO: 11)

Polynucleotide sequence:

Amino acid sequence:

VKRGLTVAVAGAAILVAGLSGCSSNKSTTGSGETTTAAGTTASPGAASGP KVVIDGKDQNVTGSVVCTTAAGNVNIAIGGAATGIAAVLTDGNPPEVKSV GLGNVNGVTLGYTSGTGQGNASATKDGSHYKITGTATGVDMANPMSPVNK SFEIEVTCS

(SEQ ID NO: 66)

Rv2744c CONSERVED 35 KDA ALANINE RICH PROTEIN

5' primer:

 $\begin{array}{lll} {\tt GAAGGAGATATACCATGCATCATCATCATCATCATATGGCCAATCCGTTC} \\ ({\tt SEQ~ID~NO:~12}) \end{array}$

3' primer:

TGATGATGAGAACCCCCCCCTGACCGTAGGGCTGCTCGG (SEQ ID NO: 13)

Polynucleotide sequence:

TABLE 1-continued

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

ACCCCCAGACCGGCCACCGAGACTTCTGGCGGGGCTATTGCCGAGCAGCC CTACGGTCAG 813 bp (SEO ID NO: 48)

Amino acid sequence:

MANPFVKAWKYLMALFSSKIDEHADPKVQIQQAIEEAQRTHQALTQQAAQ VIGNQRQLEMRLINRQLADIEKLQVNVRQALTLADQATAAGDAKATEYNN AAEAFAAQLVTAEQSVEDLKTLHDQALSAAAQAKKAVERNAMVLQQKIAE RTKLLSQLEQAKMQEQVSASLRSMSELAAPGNTPSLDEVRDKIERRYANA IGSAELAESSVQGRMLEVEQAGIQMAGHSRLEQIPASMRGEALPAGGTTA TPRPATETSGGAIAEQPYGQ

(SEQ ID NO: 67)

Rv0097 POSSIBLE OXIDOREDUCTASE

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATCATATGACGCTTAAGGTC (SEQ ID NO: 14)

3' primer:

TGATGATGAGAACCCCCCCCTGCCGCGTATCCCGGCGTCT (SEQ ID No: 15)

Polynucleotide sequence:

 $\tt ATGACGCTTAAGGTCAAAGGCGAGGGACTCGGTGCGCAGGTCACAGGGGT$ $\tt CGATCCCAAGAATCTGGACGATATAACCACCGACGAGATCCGGGATATCG$ $\tt TTTACACGAACAAGCTCGTTGTGCTAAAAGACGTCCATCCGTCTCCGCGG$ GAGTTCATCAAACTCGGCAGGATAATTGGACAAATCGTTCCGTATTACGA ${\tt ACCCATGTACCATCACGAAGACCACCCGGAGATCTTTGTCTCCTCCACTG}$ ${\tt AGGAAGGTCAGGGGGTCCCAAAAACCGGCGCGTTCTGGCATATCGACTAT}$ ATGTTTATGCCGGAACCTTTCGCGTTTTCCATGGTGCTGCCGCTGGCGGT $\tt GCCTGGACACGACCGCGGGACCTATTTCATCGATCTCGCCAGGGTCTGGC$ AGTCGCTGCCCGCCAAGCGAGACCCGGCCCGCGGAACCGTCAGCACC CACGACCCTCGACGCCACATCAAGATCCGACCCAGCGACGTCTACCGGCC CATCGGAGAGGTATGGGACGAGATCAACCGGACCACGCCCCCAATAAAGT GGCCTACGGTCATCCGGCACCCAAAGACCGGCCAAGAGATCCTCTACATC TGCGCGACGGCACCACCAAGATCGAGGACAAGGACGGCAATCCGGTTGA TCCGGAGGTGCTGCAAGAACTCATGGCCGCGACCGGACAGCTCGATCCTG AGTACCAGTCGCCGTTCATACATACTCAGCACTACCAGGTTGGCGACATC ATCTTGTGGGACAACCGGGTTCTCATGCACCGAGCGAAGCACGGCAGCGC CGCGGGCACTCTGACGACCTACCGCCTGACCATGCTTGATGGCCTCAAGA CGCCGGGATACGCGGCA 870 (SEQ ID NO: 49)

Amino acid sequence:

MTLKVKGEGLGAQVTGVDPKNLDDITTDEIRDIVYTNKLVVLKDVHPSPR EFIKLGRIIGQIVPYYEPMYHHEDHPEIFVSSTEEGQGVPKTGAFWHIDY MFMPEPFAFSMVLPLAVPGHDRGTYFIDLARVWQSLPAAKRDPARGTVST HDPRRHIKIRPSDVYRPIGEVWDEINRTTPPIKWPTVIRHPKTGQEILYI CATGTTKIEDKDGNPVDPEVLQELMAATGQLDPEYQSPFIHTQHYQVGDI ILWDNRVLMHRAKHGSAAGTLTTYRLTMLDGLKTPGYAA (SEQ ID NO: 68)

Rv0475 IRON-REGULATED HEPARIN BINDING HEMAGGLUTININ HBHA (ADHESIN)

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATATGGCTGAAAACTCG (SEQ ID NO: 16)

3' primer

TGATGAGGAACCCCCCCCTTCTGGGTGACCTTCTTGG (SEQ ID NO: 17)

Polynucleotide sequence:

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

(SEQ ID NO: 50)

Amino acid sequence:

MAENSNIDDIKAPLLAALGAADLALATVNELITNLRERAEETRTDTRSRV EESRARLTKLQEDLPEQLTELREKFTAEELRKAABGYLEAATSRYNELVE RGEAALERLRSQQSFEEVSARAEGYVDQAVELTQEALGTVASQTPAVGEP AAKLVGIELPKKAAPAKKAAPAKKAAPAKKAAPAKKAAAKKAYTQK (SEQ ID NO: 69)

RV3117 PROBABLE THIOSULFATE SULFURTRANSFERASE CYSA3 (RHODANESE-LIKE PROTEIN) (THIOSULFATE CYANIDE TRANSSULFURASE) (THIOSULFATE THIOTRANSFERASE)

5' primer

GAAGGAGATATACCATCATCATCATCATCATCATATGGCACGCTGCGAT (SEQ ID NO: 18)

3' primer:

TGATGATGAGAACCCCCCCCGCTTCCCAACTCGATCGGGG (SEQ ID NO: 19)

Polynucleotide sequence:

ATGGCACGCTGCGATGTCCTGGTCTCCGCCGACTGGGCTGAGAGCAATCT GCACGCCGAAGGTCGTTTTCGTCGAAGTGGACGAGGACACCAGTGCAT ATGACCGTGACCATATTGCCGGCGCGATCAAGTTGGACTGGCGCACCGAC $\tt CTGCAGGATCCGGTCAAACGTGACTTCGTCGACGCCCAGCAATTCTCCAA$ GCTGCTGTCCGAGCGTGGCATCGCCAACGAGGACACGGTGATCCTGTACG GCGGCAACAACAATTGGTTCGCCGCCTACGCGTACTGGTATTTCAAGCTC ${\tt TACGGCCATGAGAAGGTCAAGTTGCTCGACGGCGGCCGCAAGAAGTGGGA}$ CCTACACCGCCTCCCCGCCGGATAACACGATTCGGGCATTCCGCGACGAG GTCCTGGCGGCCATCAACGTCAAGAACCTCATCGACGTGCGCTCTCCCGA CGAGTTCTCCGGCAAGATCCTGGCCCCCGCGCACCTGCCGCAGGAACAAA GCCAGCGGCCCGGACACATTCCTGGTGCCATCAACGTGCCGTGGAGCAGG GCCGCCAACGAGGACGCACCTTCAAGTCCGATGAGGAGTTGGCCAAGCT TTACGCCGACGCCTAGACAACAGCAAGGAAACGATTGCCTACTGCC GAATCGGGGAACGGTCCTCGCACACCTGGTTCGTGTTGCGGGAATTACTC GGACACCAAAACGTCAAGAACTACGACGGCAGTTGGACAGAATACGGCTC CCTGGTGGGCCCCCGATCGAGTTGGGAAGC 834 bp (SEQ ID NO: 51)

Amino acid sequence:

MARCDVLVSADWAESNLHAPKVVFVEVDEDTSAYDRDHIAGAIKLDWRTD LQDPVKRDFVDAQQFSKLLSERGIANEDTVILYGGNNNWFAAYAYWYFKL YGHEKVKLLDGGRKKWELDGRPLSSDPVSRPVTSYTASPPDNTIRAFRDE VLAAINVKNLIDVRSPDESGKILAPAHLPQEQSQRPGHIPGAINVPWSR AANEDGTFKSDEELAKLYADAGLDNSKETIAYCRIGERSSHTWFVLRELL GHQNVKNYDGSWTEYGSLVGAPIELGS (SEO ID NO: 70)

(SEQ ID NO: 70)

Rv1347c CONSERVED HYPOTHETICAL PROTEIN

5' primer

GAAGGAGATATACCATGCATCATCATCATCATCATATGACCAAACCCACA (SEO ID NO: 20)

3' primer:

TGATGATGAGAACCCCCCCCCGCAGCCGTGGTCGGAGCTT (SEQ ID NO: 21)

Polynucleotide sequence:

TABLE 1-continued

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

ACCGACGCGGAGATGTTGGCGGAGTGATGAACCGTCCTCATCTGGCGGC
GGCCTGGGAGTACGACTGGCCGGCGTCACGTTGGCGTCAACACCTGAACG
CCCAACTTGAGGGAACCTATTCGTTGCCATTGGCGTCAACACCTGAACG
CCCAACTTGAGGGAACCTATTCGTTGCCATTGATTCGCCACAGCGAAGGATTTGATTTC
TCACTACTACGACGCAGACCCCTACGATTTGGGCCTGCACGCGGCCATCG
CGGACTTGTCGAAGGTCAATCGGGGCTTCGCCCGCTGCTGCTACCGCGG
ATCGTGGCCAGCGTCTTTGCCAACGAGCCGGTTGCCGGCGGATCATGTT
CGACCCCGATCACCGCAACACCGCGACCCGTTGTTGTGAGTGGGCCG
GATGCAAGTTCCTCGGTGAGCATGACACCGCGACAAACCGGCGCATGGCCGT
TACGCTTTGGAAGGTCCGACCACGGCTGCG 633 bp
(SEQ ID NO: 52)

Amino acid sequence:

MTKPTSAGQADDALVRLARERFDLPDQVRRLARPPVPSLEPPYGLRVAQL TDAEMLAEWMIRPHLAAAWEYDWPASRWRQHLNIAQLEGTYSLPLIGSWHG TDGGYLELYWAAKDLISHYYDADPYDLGLHAAIADLSKVNRGFGPLLLPR IVASVFANEPRCRRIMFDPDHRNTATRRLCEWAGCKFLGEHDTTNRMAL YALEAPTTAA

(SEQ ID NO: 71)

RV0815c PROBABLE THIOSULFATE SULFURTRANSFERASE CYSA2 (RHODANESE-LIKE PROTEIN) (THIOSULFATE CYA-NIDE TRANSSULFURASE) (THIOSULFATE THIOTRANSFERASE)

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATCATATGGCACGCTGCGAT (SEQ ID NO: 22)

3' primer:

TGATGATGAGAACCCCCCCGCTTCCCAACTCGATCGGGG (SEQ ID NO: 23)

Polynucleotide sequence:

ATGGCACGCTGCGATGTCCTGGTCTCCGCCGACTGGGCTGAGAGCAATCT GCACGCCGAAGGTCGTTTTCGTCGAAGTGGACGAGGACACCAGTGCAT ATGACCGTGACCATATTGCCGGCGCGATCAAGTTGGACTGGCGCACCGAC $\tt CTGCAGGATCCGGTCAAACGTGACTTCGTCGACGCCCAGCAATTCTCCAA$ GCTGCTGTCCGAGCGTGGCATCGCCAACGAGGACACGGTGATCCTGTACG GCGGCAACAACTTGGTTCGCCGCCTACGCGTACTGGTATTTCAAGCTC TACGGCCATGAGAAGGTCAAGTTGCTCGACGGCGGCCGCAAGAAGTGGGA CCTACACCGCCTCCCCGCCGGATAACACGATTCGGGCATTCCGCGACGAG GTCCTGGCGGCCATCAACGTCAAGAACCTCATCGACGTGCGCTCTCCCGA CGAGTTCTCCGGCAAGATCCTGGCCCCCGCGCACCTGCCGCAGGAACAAA GCCAGCGGCCCGGACACATTCCTGGTGCCATCAACGTGCCGTGGAGCAGG GCCGCCAACGAGGACGCACCTTCAAGTCCGATGAGGAGTTGGCCAAGCT TTACGCCGACGCCGGCCTAGACAACAGCAAGGAAACGATTGCCTACTGCC GAATCGGGGAACGGTCCTCGCACACCTGGTTCGTGTTGCGGGAATTACTC GGACACCAAAACGTCAAGAACTACGACGGCAGTTGGACAGAATACGGCTC CCTGGTGGGCGCCCCGATCGAGTTGGGAAGC 834 bp (SEQ ID NO: 53)

Amino acid sequence:

MARCDVLVSADWAESNLHAPKVVFVEVDEDTSAYDRDHIAGAIKLDWRTD LQDPVKRDFVDAQQFSKLLSERGIANEDTVILYGGNNNWFAAYAYWYFKL YGHEKVKLLDGGRKKWELDGRPLSSDPVSRPVTSYTASPPDNTIRAFRDE VLAAINVKNLIDVRSPDEFSGKILAPAHLPQEQSQRPGHIPGAINVPWSR AANEDGTFKSDEELAKLYADAGLDNSKETIAYCRIGERSSHTWFVLRELL GHQNVKNYDGSWTEYGSLVGAPIELGS

(SEQ ID NO: 72)

Rv2613c CONSERVED HYPOTHETICAL PROTEIN

5' primer:

3' primer:

 ${\tt TGATGAGAAACCCCCCCTGGTTGCCGAGCCCACTCGG} \\ ({\tt SEQ\ ID\ NO:\ 25})$

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

Polynucleotide sequence:

Amino acid sequence:

VSDEDRTDRATEDHTIFDRGVGQRDQLQRLWTPYRMNYLAEAPVKRDPNS SASPAQPFTEIPQLSDEEGLVVARGKLVVAVLNLYPYNPGHLMVVPYRRV SELEDLTDLESAELMAFTQKAIRVIKNVSRPHGFNVGLNLGTSAGGSLAE HLHVHVVPRWGGDANFITIIGGSKVIPQLLRDTRRLLATEWARQP (SEQ ID NO: 73)

Rv3226c CONSERVED HYPOTHETICAL PROTEIN

5' primer:

GAAGGAGATATACCATCATCATCATCATCATATGTGCGGACGGTTT (SEQ ID NO: 26)

3' primer:

TGATGATGAGAACCCCCCCCAGCAGCTGGATCTGCTCGG (SEQ ID NO: 27)

Polynucleotide sequence:

ATGTGCGGACGGTTTGCGGTCACCACTGATCCGGCCCAGCTGGCCGAGAA AATCACGGCCATAGACGAGGCCACCGGGTGCGGTGGCGGGAAGACGAGCT ACAACGTGGCACCCACCGACACGATCGCGACAGTGGTGTCCCGCCACAGC GAGCCCGACGACCACCCGCCGGGTGCGGCTCATGCGCTGGGGACT GATTCCGTCGTGGATCAAGGCCGGGCCCCGGCGCGCGCCCCGATGCCAAAG GCCCACCGCTGATCAACGCCCGCGCCGATAAGGTCGCCACGTCGCCGGCG TTCCGGAGTGCGGTCAGAAGTAAGCGTTGCCTGGTGCCGATGGACGGCTG GTACGAATGGCGCGTCGACCCCGACGCCACCCCGGGGAGGCCGAACGCCA AGACGCCGTTCTTCCTGCACCGCCACGACGCCCCTGTTGTTCACGGCC GGGCTGTGGTCGGTTTGGAAGTCTTACAGGTCCGCCCCACCGCTGCTGAG CTGCACGGTGATCACCACCGATGCCGTGGGCGAGCTGGCCGAGATCCATG ACCGGATGCCGCTGCTGCCGGCCGAAGAGGACTGGGACGACTGGCTGAAT CGACATCGCGCTGCGCCAAGTGTCCACGTTGGTCAACAACGTGCGCAACA ACGGGCCTGAGCTGTTGGAGCCGGCCAGGTCGCAGCCCGAGCAGATCCAG CTGCTG 759 bp (SEQ ID NO: 55)

Amino acid sequence:

MCGRFAVTTDPAQLAEKITAIDEATGCGGGKTSYNVAPTDTIATVVSRHS EPDDEPTRRVRLMRWGLIPSWIKAGPGGAPDAKTGPFLINARADKVATSPA FRSAVRSKRCLVPMDGWYEWRVDPDATPGRPNAKTPFFLHRHDGALLFTA GLWSVWKSYRSAPPLLSCTVITTDAVGELAEIHDRMPLLLAEEDWDDWLN PDAPPDPELLARPPDVRDIALRQVSTLVNNVRNNGPELLEPARSQPEQIQ LI.

(SEQ ID NO: 74)

Rv0349 HYPOTHETICAL PROTEIN

5' primer:

 ${\tt GAAGGAGATATACCATGCATCATCATCATCATGTGCCAGAGCTGGAG} \\ ({\tt SEQ\ ID\ NO:\ 28})$

3' primer:

TGATGATGAGAACCCCCCCGTCCGCCAGCTTGACCGACT (SEQ ID NO: 29)

TABLE 1-continued

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

Polynucleotide sequence:

(SEQ ID NO: 56) Amino acid sequence:

VPELETPDDPESIYLARLEDVGEHRPTFTGDIYRLGDGRMVMILQHPCAL RHGVDLHPRLLVAPVRPDSLRSNWARAPFGTMPLPKLIDGQDHSADFINL ELIDSPTLPTCERIAVLSQSGVNLVMQRWVYHSTRLAVPTHTYSDSTVGP FDEADLIEEWVTDRVDDGADPQAAEHECASWLDERISGRTRPALLSDRQH ASSIRREARSHRKSVKLAD

(SEO ID NO: 75)

RV0009 PROBABLE IRON-REGULATED PEPTIDYL-PROLYL CIS-TRANS ISOMERASE A PPIA (PPIase A) (ROTAMASE A)

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATATGGCAGACTGTGAT (SEQ ID NO: 30)

3' primer:

TGATGATGAGAACCCCCCCGGAGATGGTGATCGACTCGA (SEQ ID NO: 31)

Polynucleotide sequence:

(SEQ ID NO: 57)

Amino acid sequence:

 ${\tt MADCDSVTNSPLATATATLHTNRGDIKIALFGNHAPKTVANFVGLAQGTK} \\ {\tt DYSTQNASGGPSGPFYDGAVFHRVIQGFMIQGGDPTGTGRGGPGYKFADE} \\ {\tt FHPELQFDKPYLLAMANAGPGTNGSQFFITVGKTPHLNRRHTIFGEVIDA} \\ {\tt ESQRVVEAISKTATDGNDRPTDPVVIESITIS} \\ {\tt TOTAL CONTROL C$

(SEQ ID NO: 76)

Rv1073 CONSERVED HYPOTHETICAL PROTEIN

5' primer

 ${\tt GAAGGAGATATACCATGCATCATCATCATCATATGGGGGGCGCAGCCG} \\ ({\tt SEQ\ ID\ No:\ 32})$

3' primer:

TGATGATGAGAACCCCCCCCACGTCGTGATGTCAACGTGT (SEQ ID NO: 33)

Polynucleotide sequence:

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

ATCTGGACAAGCGGCTGAAGCCCTCCCTGCGGCAACGCGTTATCGCGGCC TGGCTGTGGTCGGGCCGCAAAGGGGTGATCGCCGGCGCTTCGGCATCAGC $\tt GCTGCACGGCGCAAATGGGTCGATGACCACGCATTGGTGGAGTTGATCT$ GGCGCAACGCCAGGGCGCCGAACGGGGTGCGGACTAAGGATGAGCTACTG $\tt CTCGACGGCGAAGTCCAGCGCTTGTGCGGGCTTACTGTGACTACCGTTGA$ ACGTACGGCCTTCGACTTGGGCAGGCGTCCACCCTTAGGTCAGGCGATAA $\tt CCAGACTGGATGCGCTTGCCAATGCCACCGATTTCAAGATCAACGATGTT$ AGGGAGCTCGCGAGGAAGCACCCCCATACTCGCGGGCTGCGTCAACTAGA ${\tt CAAGGCGCTGGATCTCGTCGACCCAGGTGCGCAGTCGCCGAAGGAGACGT}$ GGCTGCGGCTCTTGCTGATAAACGCCGGCTTTCCACGGCCGTCCACTCAG $\tt ATCCCCTTGCTCGGCGTCTACGGGCATCCAAAGTATTTCCTCGACATGGG$ $\tt ATGGGAGGACATCATGCTCGCGGTCGAGTACGACGGCGAGCAACACCGTC$ ${\tt TCAGCCGAGACCAGTTCGTCAAAGACGTCGAACGCCTGGAATACATCCGG}$ $\tt CGCGCCGGCTGGACTCACATCAGGGTGCTGGCAGACCACAAGGGACCCGA$ $\tt CGTCGTCCGCCGGGTTCGGCAGGCTTGGGACACGTTGACATCACGACGT$ 852 bp

(SEQ ID NO: 58)

Amino acid sequence:

MGAQPFIGSEALAAGLISWHELGKYYTAIMPNVYLDKRLKPSLRQRVIAA WLWSGRKGVIAGASASALHGAKWVDDHALVELIWRNARAPNGVRTKDELL LDGEVQRLCGLTVTTVERTAFDLGRRPPLGQAITRLDALANATDFKINDV RELARKHPHTRGLRQLDKALDLVDPGAQSPKETWLRLLLINAGFPRPSTQ IPLLGYYGHPKYFLDMGWEDIMLAVEYDGEQHRLSRDQFVKDVERLEYIR RAGWTHIRVLADHKGPDVVRRVRQAWDTLTSRR

(SEQ ID NO: 77)

Rv0781 PROBABLE PROTEASE II PTRBA [FIRST PART] (OLIGOPEPTIDASE B)

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATCATATGATGCACCGAACC (SEQ ID NO: 34)

3' primer:

TGATGATGAGAACCCCCCCTCGGCTTCGTGGTAAACCCG (SEQ ID NO: 35)

Polynucleotide sequence:

ATGATGCACCGAACCGCACTACCCTCACCGCCCGTGGCCAAGCGGGTGCA GACCCGC: CGGGAGCACCACGGCGACGTCTTTGTCGACCCATATGAATGG TTGCGCGACAAGGACAGCCCTGAAGTAATCGCCTACCTCGAAGCTGAAAA CGACTACACCGAACGGACCACCGCGCACCTTGAGCCATTGCGGCAAAAGA TCTTCCACGAAATCAAAGCGCGTACCAAGGAAACCGACTTATCGGTGCCG ACGCGACGTGGCAACTGGTGGTACTACGCGCGGACCTTTGAGGGAAAGCA GTATGGCGTACACTGTCGTTGCCCGGTAACCGATCCCGACGACTGGAACC CACCAGAGTTCGACGAGCGCACCGAAATACCCGGTGAACAGCTTCTGCTC CAGCGTCAGCCTGGACGATAACCTCTTAGCGTATTCCGTTGATGTCGTAG $\tt GTGACGAACGATATACCTTGCGGTTCAAGGATTTACGCACCGGAGAACAG$ ${\tt TACCCGGACGAGATCGCCGGGATCGGAGCTGACCTGGGCAGCTGA}$ CAACCACTGTCTACTACACCACCGTGGACGCGGCCTGGCGTCCGGACACA $\tt GTGTGGCGATACCGACTAGGGTCCGGCGAATCGTCGGAGCGGGTTTACCA$ CGAAGCCGA 711 bp (SEQ ID NO: 59)

Amino acid sequence:

MMHRTALPSPPVAKRVQTRREHHGDVFVDPYEWLRDKDSPEVIAYLEAEN DYTERTTAHLEPLRQKIFHEIKARTKETDLSVPTRRGNWYYARTFEGKQ YGVHCRCPVTDPDDWNPPEFDERTEIPGEQLLLDENVEADGHDFFALGAA SVSLDDNLLAYSVDVVGDERYTLRFKDLRTGEQYPDEIAGIGAGVTWAAD NHCLLHHRGRGLASGHSVAIPTRVRRIVGAGLPRSR

(SEQ ID NO: 78)

Rv2108 PPE FAMILY PROTEIN

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATATGCCCAATTTCTGG (SEQ ID NO: 36)

TABLE 1-continued

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

3' primer:

TGATGATGAGAACCCCCCCCAAACTTAGGATGTTCCTTGT (SEO ID NO: 37)

Polynucleotide sequence:

ATGCCCAATTTCTGGGCGTTGCCGCCCGAGATCAACTCCACCCGGATATA ${\tt TCTCGGCCCGGGTTCTGGCCCGATACTGGCCGCCCCAGGGATGGAACG}$ CTCTGGCCAGTGAGCTGGAAAAGACGAAGGTGGGGTTGCAGTCAGCGCTC ${\tt GACACGTTGCTGGAGTCGTATAGGGGTCAGTCGTCGCAGGCTTTGATACA}$ GCAGACCTTGCCGTATGTGCAGTGGCTGACCACGACCGCCGAGCACGCCC $\tt ATAAGACCGCGATCCAGCTCACGGCAGCGGCGAACGCCTACGAGCAGGCT$ ${\tt AGAGCGGCGATGGTGCCGCCGGCGATGGTGCGCGCGAACCGCGTGCAGAC}$ ${\tt CACAGTGTTGAAGGCAATCAACTGGTTCGGGCAATTCTCCACCAGGATCG}$ $\tt CCGACAAGGAGGCCGACTACGAACAGATGTGGTTCCAAGACGCGCTAGTG$ ATGGAGAACTATTGGGAAGCCGTGCAAGAGGCGATACAGTCGACGTCGCA $\tt TTTTGAGGATCCACCGGAGATGGCCGACGACTACGACGAGGCCTGGATGC$ TCAACACCGTGTTCGACTATCACAACGAGAACGCAAAAGAGGGGGTCATC ${\tt CATCTCGTGCCCGACGTGAACAAGGAGAGGGGGCCCATCGAACTCGTAAC}$ CAAGGTAGACAAAGAGGGGACCATCAGACTCGTCTACGATGGGGAGCCCA CGTTTTCATACAAGGAACATCCTAAGTTT 732 bp (SEQ ID NO: 60)

Amino acid sequence:

MPNFWALPPEINSTRIYLGPGSGPILAAAQGWNALASELEKTKVGLQSAL DTLLESYRGQSSQALIQQTLPYVQWLTTTAEHAHKTAIQLTAAANAYEQA RAAMVPPAMVRANRVQTTVLKAINWFGQFSTRIADKEADYEQMWFQDALV MENYWEAVQEAIQSTSHFEDPPEMADDYDEAWMLNTVFDYHNENAKEEVI HLVPDVNKERGPIELVTKVDKEGTIRLVYDGEPTFSYKEHPKF (SEQ ID NO: 79)

 ${\tt Rv3920c}$ HYPOTHETICAL PROTEIN SIMILAR TO JAG PROTEIN

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATCATATGGCCGACGCTGAC (SEO ID NO: 38)

3' primer:

Polynucleotide sequence:

ATGGCCGACGCTGACACCACCGACTTCGACGTCGACGCAGAAGCACCGGG
TGGAGGCGTCCGGGAGGACACGGCGACGATGCTGACGAGGCCGACGATC
AAGAACAGAGATTGGTCGCCGAGGGCGACTTGCAGCGCGACTACCTGGAA
GAGTTATTGGACGTTGGACTTCGATGGCGACATCGACCTCGATGTCGA
AGGCAATCGTGCGGTGGAGCACTCGACGCAGTGACCAGT
TGGTCGGGCGCGGGGGCGAGGTGCTCGACGACTTGAACAAGT
TGGTCGGGTGCATCAGAAGACCGGTGTGCGGAGCCGGTTGATGCTAGACAT
CGCGAGGTGCATCAGAAGACCGGTGTCGGAGCCGGTTGATGCTAGACAT
CGCGAGGTGGCGACGCGCGCGGGAGGAATTGGCGGCGCTGGCCGAC
AGGTGGCCGCGCGAAACCGGTGACCGCGAGAACTCGTTCCA
ATGACGCCGTTCGAACGAAGACTCGTCCACGATGCGGTTGCACCGGTGC
AGGTGTCCACAGCGAAAGCGAGCGTGACCAGAACCCCGAGTCGTTG
TGCTCCGCGAC 564
(SEQ ID NO: 61)

Amino acid sequence:

MADADTTDFDVDAEAPGGGVREDTATDADEADDQEERLVAEGEIAGDYLE ELLDVLDFDGDIDLDVEGNRAVVSIDGSDDLNKLVGRGGEVLDALQELTR LAVHQKTGVRSRLMLDI ARWRRRREELAALADEVARRVAETGDREELVP MTPFERKIVHDAVAAVPGVHSESEGVEPERRVVVLRD (SEQ ID NO: 80)

Rv1044 CONSERVED HYPOTHETICAL PROTEIN

5' primer:

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

3' primer:

TGATGATGAGAACCCCCCCCCCCGCCGATGCTCGCTTCGGCC (SEO ID NO: 41)

Polynucleotide sequence:

(222 12 1.0. 02)

Amino acid sequence:

LCAKPYLIDTIAHMAIWDRLVEVAAEQHGYVTTRDARDIGVDPVQLRLLA GRGRLERVGRGYYRVPVLPRGEHDDLAAAVSWTLGRGVISHESALALHAL ADVNPSRIHLTVPRNNHPRAAGGELYRVHRRDLQAAHVTSVDGIPVTTVA RTIKDCVKTGTDPYQLRAAIERAEAEGTLRRGSAAELRAALDETTAGLRA RPKRASA

(SEQ ID NO: 81)

Rv2882c RIBOSOME RECYCLING FACTOR FRR (RIBOSOME RELEASING FACTOR) (RRF)

5' primer:

3' primer:

TGATGATGAGAACCCCCCCGACCTCCAGCAGCTCGCCTT (SEQ ID NO: 43)

Polynucleotide sequence:

(SEQ ID NO: 63)

Amino acid sequence:

MIDEALFDAEEKMEKAVAVARDDLSTIRTGRANPGMFSRITIDYYGAATP ITQLASINVPEARLVVIKPYEANQLRAIETAIRNSDLGVMPTNDGALIRV AVPQLTEERRRELVKQAKHKGEEAKVSVRNIRRKAMEELHRIRKEGEAGE DEVGRAEKDLDKTTHQYVTQIDELVKHKEGELLEV (SEQ ID NO: 82)

Rv3733c CONSERVED HYPOTHETICAL PROTEIN

5' primer:

3' primer:

TGATGATGAGAACCCCCCCGCGAGGCAGGGATTCTGGTC (SEQ ID NO: 45)

TABLE 1-continued

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

Polynucleotide sequence:

(SEQ ID NO: 64)

Amino acid sequence:

(SEO ID NO: 83)

Rv0138 CONSERVED HYPOTHETICAL PROTEIN

5' primer

GAAGGAGATATACCATCATCATCATCATCATGTGAGCGCTTCGGAG (SEQ ID NO: 86)

3' primer:

TGATGAGAACCCCCCCAGGACCTCCATGCCGGCGCA (SEQ ID NO: 87)

Polynucleotide sequence:

SEQ ID NO: 110)

Amino acid sequence:

VSASEFSRAELAAAFEKFEKTVARAAATRDWDCWVQHYTPDVEYIEHAAG IMRGRQRVRAMIQETMTTFPGSHMVAFFSLWSVIDESTGRIICELDNPML DPGDGSVISATNISIITYAGNGQWCRQEDIYNPLRFLRAAMKWCRKAQEL GTLDEDAARWWRRHGGP

(SEQ ID NO: 122)

Rv0740 CONSERVED HYPOTHETICAL PROTEIN

5' primer:

 ${\tt GAAGGAGATATACCATGCATCATCATCATCATATGCTGCCGAAGAAC} \\ ({\tt SEQ~ID~NO:~88})$

3' primer

TGATGATGAGAACCCCCCCCCCCCCCCGCCTTTTCG (SEQ ID No: 89)

Polynucleotide sequence:

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

Amino acid sequence:

MLPKNTRPTSETAEEFWDNSLWCSWGDRETGYTRTVTVSICQVADGEREA EGVRDMMRLECPAGLDLRTPNPEAYEITGQRPGEFVFVLGYLGHVRAIVG NCYIEIMPMGTRVELSKLADVALDIGRSVGCSAYENDFTLPDIPTQWRNQ PLGWYTQGLAPYLPGLSDPKDAAEG

(SEQ ID NO: 123)

Rv0733 PROBABLE ADENYLATE KINASE ADK (ATP-AMP TRANSPHOSPHORYLASE)

5' primer

 ${\tt GAAGGAGATATACCATGCATCATCATCATCATGTGAGAGTTTTGTTG} \\ ({\tt SEQ\ ID\ No:\ 90})$

3' primer:

TGATGATGAGAACCCCCCCCTTTCCCAGAGCCCGCAACG (SEO ID NO: 91)

Polynucleotide sequence:

Amino acid sequence:

VRVLLLGPPGAGKGTQAVKLAEKLGIPQISTGELFRRNIEEGTKLGVEAK RYLDAGDLVPSDLTNELVDDRLNNPDAANGFILDGYPRSVEQAKALHEML ERRGTDIDAVLEFRVSEEVLLERLKGRGRADDTDDVILNRMKVYRDETAP LLEYYRDQLKTVDAVGTMDEVFARALRALG K (SEQ ID NO: 124)

Rv1065 CONSERVED HYPOTHETICAL PROTEIN

5' primer:

GAAGGAGATATACCATCATCATCATCATCATGTGGTTATGCCTCTT (SEQ ID NO: 92)

3' primer:

TGATGATGAGAACCCCCCCCTCCCGACCCTTCGGGCTGGT (SEQ ID NO: 93)

Polynucleotide sequence:

(SEQ ID NO: 113)

TABLE 1-continued

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

Amino acid sequence:

VVMPLVTPTTAVPSPGPTRLRVADLLRATDQAADDVLGGRCDHLLPDGGV PQTQRWYTRIHGDEELDIWLISWVPGQPTELHDHGGSLGALTVLSGSLNE YRWDGRRLRRRRLDAGDQAGFPLGWVHDVVWAPRPIGGPDAAGMAVAPTL SVHAYSPPLTAMSYYEITERNTLRRQRTELTDQPEGSG (SEO ID NO: 125)

Rv2114 HYPOTHETICAL PROTEIN

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATCATATGTCGGCTCCCGAA (SEQ ID NO: 94)

3' primer:

TGATGATGAGAACCCCCCCCGGCGGTCACCAGCGAGTAGC (SEQ ID NO: 95)

Polynucleotide sequence:

(SEQ ID NO: 114)

Amino acid sequence:

MSAPERVTGLSGQRYGEVLLVTPGEAGPQATVYNSFPLNDCPAELWSALD PQALATEHKAATALLNGPRYWLMNAIEKAPQGPPVTKTFGGIEMLQQATV LLSSMNPAPYTVSQVSRNTVFVPNAGEEVYELQDPKGQRWVMQTWSQVVD PNLSRADLPKLGERLNLPAGWSYHTRVLTSELRVDTTNREARVLQDDLTN SYSLVTA

(SEQ ID NO: 126)

Rv2466c CONSERVED HYPOTHETICAL PROTEIN

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATATGCTCGAGAAGGCC (SEQ ID NO: 96)

3' primer:

TGATGATGAGAACCCCCCCCGTCGAACTGAGGCGGCTCGG (SEQ ID NO: 97)

Polynucleotide sequence:

ATGCTCGAGAAGGCCCCCCAGAAGTCTGTCGCCGATTTCTGGTTCGATCC
GCTGTGCCCGTGGTGCTGGATCACGTCGGCTGGATCCTGAGGTGGCAA
AGGTCCGCGACATCGAGGTGAACTTCCACGTCATGAGCCTGGCAATACTC
AACGAAAACCGTGACGACCTGCCCGAGCAATACCGCGAGGCCCATGGGG
GGCATGGGGACCGGTACGGGTGGCGATCGCCGCAGCAAGCCCATGGGG
CGAAACTCCTGGACCAGGTTACACCGCGATGGGCAACCGGATTCACAAC
CAGGCAACCACGAACTCGACGAGGTCATCACCCAGTCGCTTGCGAACG
CGGTTTGCCCGGGAGTTGGCCAAGGCCGCTACCAGCGACGCTTACACAC
ACGCCCTGCGAAAAGCCACCACGCCGGATGGACGCGTTGGGCAAGGAGC
GTCGGTACGCCGACGATCCATGTCAATGGTGTGGCGTTCTTCGGGCCGGT
GCTCTCGAAGATTCCGCGCGCGAGGAAGCCGCCAAGCTCTGGGATGCCT
CGGTTACCTTCCTACCGCACTTTTTTGAGCTCAAGCGGACCCGC
ACCGACCCCTCAGTTCGAC

(SEQ ID NO: 115)

Amino acid sequence:

MLEKAPQKSVADPWFDPLCPWCWITSRWILEVAKVRDIEVNFHVMSLAIL NENRDDLPEQYREGMARAWGPVRVAIAAEQAHGAKVLDPLYTAMGNRIHN QGNHELDEVITQSLADAGLPAELAKAATSDAYDNALRKSHHAGMDAVGED

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

 ${\tt VGTPTIHVNGVAFFGPVLSKIPRGEEAGKLWDASVTFASYPHFFELKRTR} \\ {\tt TEPPOFD}$

(SEQ ID NO: 127)

RV0158 PROBABLE TRANSCRIPTIONAL REGULATORY PROTEIN (POSSIBLY TETR-FAMILY)

5' primer:

 ${\tt GAAGGAGATATACCATGCATCATCATCATCATATGCCATCCGACACC} \\ ({\tt SEQ\ ID\ NO:\ 98})$

3' primer:

TGATGATGAGAACCCCCCCCCGTTTCCTTCCGAGTTCCAA (SEQ ID NO: 99)

Polynucleotide sequence:

Amino acid sequence:

MPSDTSPNGLSRREELLAVATKLFAARGYHGTRMDDVADVIGLNKATVYH YYASKSLILFDIYRQAAEGTLAAVHDDPSWTAREALYQYTVRLLTAIASN PEPAAVYFQEQPYITEWFTSEQVAEVREKEQQVYEHVHGLIDRGIASGEF YECDSHVVALGYIGMTLGSYRWLRPSGRRTAKEIAAEFSTALLRGLIRDE SIRNQSPLGTRKET

(SEQ ID NO: 128)

Rv3676 PROBABLE TRANSCRIPTIONAL REGULATORY PROTEIN (PROBABLY CRP/FNR-FAMILY)

5' primer:

 $\begin{array}{lll} {\tt GAAGGAGATATACCATGCATCATCATCATCATGTGGACGAGATCCTG} \\ ({\tt SEQ~ID~NO:~100}) \end{array}$

3' primer:

TGATGATGAGAACCCCCCCCCCCTCGCTCGGCGGGCCAGTC (SEQ ID NO: 101)

Polynucleotide sequence:

(SEQ ID NO: 117)

Amino acid sequence:

VDEILARAGIFQGVEPSAIAALTKQLQPVDFPRGHTVFAEGEPGDRLYII ISGKVKIGRRAPDGRENLLTIMGPSDMFGELSIFDPGPRTSSATTITEVR AVSMDRDALRSWIADRPEISEQLLRVLARRLRRTNNNLADLIFTDVPGRV

TABLE 1-continued

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

 ${\tt AKQLLQLAQRFGTQEGGALRVTHDLTQEEIAQLVGASRETVNKALADFAH} \\ {\tt RGWIRLEGKSVLISDSERLARRAR} \\$

(SEO ID NO: 129)

Rv2821c CONSERVED HYPOTHETICAL PROTEIN

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATATGACTACGAGCTAC (SEQ ID NO: 102)

3' primer:

TGATGATGAGAACCCCCCCAACAGCCGCGAGTTCATGGT (SEQ ID NO: 103)

Polynucleotide sequence:

ATGACTACGAGCTACGCCAAGATCGAGATAACCGGGACACTGACCGTCCT
GACGGGCCTGCAGATCGGGGCCGCGATGGCTTCTCCGCCATCGGCGGG
GCGACAAGCCTGTCGTTCCGTGATCCGCTGAGCAGGCTGCCGATGATTCCG
GGTACCAGCCTGAAGGGCAAGGTCCGCACCTTGCTGTCCCGCCAATACGG
CGCCGACACAGAAACGTTTTACAGGAAGCCCGAATGAGGCCCAATA
TCCGTCGGCTTTTCGGCGACACCGAGGAGTACATGACGGCCGACCCATA
TCCGTCGGCTTTTCGGCGACACCAACAAAGACGACCTCGAAGCCCGCGGCGC
TAAGACTCTCACCGAGTGAAATTCGAGAACGCCATCAACCGGGTGACCG
CAAAGGCAAACCTTCGCCAGATGGAACGCGTGATCCCCGGCAGGAGTC
GCGTTCTCACTTGTCTACGAGGTCTCCTTCGGCACCCCGCGGAGAACA
GAAGGCGTCTCTCCTCCGATGAGATCACTCGAGGACTTCAACGCCA
TCGCGCGCGCGCCTGAAGTTGCTCGAACTCGACTACCTCGGCAGCGAGACA
ACCCGTGGCTACGACTCCCAGATGACTCACCTGAAAGCCCCCGCGCGAGACACA
ACCCGTGGCTACGACTCCTCGAACTTCACCAACCTGAAACCCCCCGCCGCAGGAACA
ACCCGTGGCTACGGCAGGTCAACTTCACCAACCTGAAACCCCCCGCCGCCGC
CGGCTGTT

(SEQ ID NO: 118)

Amino acid sequence:

MTTSYAKIEITGTLTVLTGLQIGAGDGFSAIGAVDKPVVRDPLSRLPMIP GTSLKGKVRTLLSRQYGADTETFYRKPNEDHAHIRRLFGDTEEYMTGRLV FRDTKLTNKDDLEARGAKTLTEVKFENAINRVTAKANLRQMERVIPGSEF AFSLVYEVSPGTPGEEQKASLPSSDEIIEDFNAIARGLKLLELDYLGGSG TRGYGQVKFSNLKARAAVGALDGSLLEKLNHELAAV (SEQ ID NO: 130)

Rv1056 CONSERVED HYPOTHETICAL PROTEIN

5' primer:

GAAGGAGATATACCATGCATCATCATCATCATCATATGAGCGTGGATTAC (SEQ ID NO: 104)

3' primer:

Polynucleotide sequence:

ATGAGCGTGGATTACCCCCAAATGGCTGCTACCCGGGGAAGAATAGAACC GGCCCCGCGGCGAGTTCGCGGCTATCTCGGACATGTGCTCGTCTTCGACA CCAGTGCGGCGCTATGTCTGGGAGGTTCCCTACTACCCGCAGTACTAC GCAGCGAGTGCAGCTGGGTCCGTCGCGGCTGCACTCCTTGGTAAGCGCCG GTCAGACCCACCGATCGGCGGCGCGGGTATTCGATGTCGACGGCGACAGC $\tt CCGGTGGCGGCACCGTGCGTTTCAACTGGGATCCGCTGCGGTGGTTCGA$ $\tt GGAGGACGAGCCGATCTACGGCCATCCGCGCAATCCCTATCAGCGGGCCG$ $\tt ATGCGCTGCGCACCGACACGTCCGTGTCGAGCTGGACGGCATTGTG$ CTCGCTGACACCCGATCGCCCGTTCTGCTATTCGAAACTGGGATACCCAC AAGGTATTACATCGATCCGGCCGACATCGCTTTCGAGCATCTGGAGCCCA $\tt CCTCGACGCAGACGTTGTGTCCGTACAAGGGGACGACGTCGGGCTATTGG$ ${\tt TCTGTGCGCGTCGGCGACGCGTGCACCGCGACCTGGCCTGGACGTATCA}$ CTATCCACTGCCCGCCGTTGCCCCGATCGCCGGCCTGGTGGCGTTTTACA ${\tt ACGAGAAGGTCGACCTCACCGTCGACGGCGTCGCCCTGCCGCGCCGCAC}$ ACTCAGTTCAGC

(SEQ ID NO: 119)

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

Amino acid sequence:

MSVDYPQMAATRGRIEPAPRRVRGYLGHVLVFDTSAARYVWEVPYYPQYY
IPLADVRMEFLRDENHPQRVQLGPSRLHSLVSAGQTHRSAARVFDVDGDS
PVAGTTVRFNWDPLRWFEEDEPIYGHPRNPYQRADALRSHRHVRVELDGIV
LADTRSPVLLFETGIPTRYYIDPADIAFEHLEPTSTQTLCPYKGTTSGYW
SVRVGDAVHRDLAWTYHYPLPAVAPIAGLVAFYNEKVDLTVDGVALPRPH
TOFS

(SEQ ID NO: 131)

Rv1353c PROBABLE TRANSCRIPTIONAL REGULATORY PROTEIN

5' primer

 $\begin{array}{lll} {\tt GAAGGAGATATACCATGCATCATCATCATCATATGCAGACAACCCCA} \\ ({\tt SEQ~ID~NO:~106}) \end{array}$

3' primer:

TGATGATGAGAACCCCCCCACGCGCCACCGCTTTGGCCC (SEQ ID NO: 107)

Polynucleotide sequence:

ATGCAGACAACCCCAGGCAAGCGTCAACGACGGCAGCGCGGATCCATCAA $\tt CCCCGAGGACATCATCAGCGGCGCATTCGAACTCGCCCAGCAGGTATCGA$ TAGACAACTTGAGCATGCCATTGCTCGGCAAACACCTTGGCGTCGGGGTC ACCAGCATCTACTGGTACTTCCGCAAGAAGGACGATCTGCTCAACGCGAT ${\tt GACCGACCGCGCTTTGAGCAAGTACGTGTTCGCTACCCCGTACATCGAAG}$ $\tt CCGGCGACTGGCGCAAACGTTGCGCAATCATGCCCGCTCGATGCGGAAG$ ACGTTCGCGGACAACCCCGTACTGTGCGATCTGATACTGATTCGAGCGGC GCTGTCCCCGAAAACGGCGCGGTTGGGCGCCCAAGAGATGGAGAAGGCCA ${\tt TCGCCAATCTGGTGACGGCGGGCCTGTCGCTCGAAGACGCTTTCGACATC}$ ${\tt TACTCGGCGGTTTCGGTCCACGTGCGCGGATCGGTGCTAGATCGGCT}$ $\tt CCGTGGCCATCGATCCCGCGACGACTCCGCTGCTTGCTCACGCAACTGGG$ ${\tt AGGGGGCATCGGATCGGGGCCCCCGATGAAACCAATTTCGAATATGGTCT}$ CGAATGCATCCTCGACCATGCTGGCCGGTTGATCGAACAAGCTCGAAAG CCGCTGGTGAGGTCGCAGTGCGCCGCCCACGGCCACCGCCGATGCGCCT ACGCCGGGCGCGGGCCAAAGCGGTGGCGCGT (SEQ ID NO: 120)

Amino acid sequence:

MQTTPGKRQRRQRSINPEDIISGAFELAQQVSIDNLSMPLLGKHLGVGV TSIYWYFRKKDDLLNAMTDRALSKYVFATPYIEAGDWRETLRNHARSMRK TFADNPVLCDLILIRAALSPKTARLGAQEMEKAIANLVTAGLSLEDAFDI YSAVSVHVRGSVVLDRLSRKSQSAGSGPSAIEHPVAIDPATTPLLAHATG RGHRIGAPDETNFEYGLECILDHAGRLIEQSSKAAGEVAVRRPTATADAP TPGARAKAVAR

(SEQ ID NO: 132)

Rv2528c PROBABLE RESTRICTION SYSTEM PROTEIN MRR

5' primer:

GAAGGAGATATACCATCATCATCATCATCATATGACGATCCCTGAT (SEQ ID NO: 108)

3' primer:

TGATGATGAGAACCCCCCCCAGGCCATCAAAAAAGTCCT (SEQ ID NO: 109)

Polynucleotide sequence:

ATGACGATCCCTGATGCCCAGACGTTGATGCGGCCGATTCTCGCGTATCT
TGCCGATGGACAAGCGAAGTCGGCCAAGGACGTCATCGCGGCGATGTCCG
ACGATTCGGTCTGTCCGACGACGACGAGGGCGCAGATGTTCCCCACGGGT
CGGCAAAGGACCATGTACGACAGGGGGGCACATGTTCTCACACATGTC
GCAGGCCGGATTGCTCGAACGCCTCCCACGGGGGCCACGTCCACGGT
ACACGGGCCGTCAAGTCCTGAAGGCGCATCCCGAGCGGTCGACATGGCT
GTGCTGCGGGGATTCCTCGTACATCGCTTTTCGTGAGCGAACCAAAGC
CAAGCAGCCAGTCGACGGCCCAAGCGACCGTCCGGGGACGATGGC
AGGTCTCACCCGAGGATCTCATCGACGCTTGCGGAGAACCAGAG
GCCGTCGAGGGGAACCTCTCAAGAGAAGCACTCACGTTGTCGCCACCGG
GTTTGAAGATCTGTTATCAACTTTTTGAAGGCATTGGTCCCACCGG
GTTTGAAGATCTGGTTATCAGACTTTTTCGACGCATGGTTACGGCCGAG
CCGGCCGGTGGAACGGACGACTCCCGGTGACGCTTGCGGACACCGAA
ATCATCAGCCAGGACCGCCTCCGGTGACCTTGCACGGA

TABLE 1-continued

Immunogenic Mtb Polypeptides: Primers, Polynucleotide Sequences, and Amino Acid Sequences All polynucleotide sequences are shown in the 5' to 3' orientation

GCGATACGCCGTCGACCAAACGATTGGCCGGCCGAAGATCCACGAGTTCG CCGGCGCCCTCCTGGGCAAGCAGGGCGACCGGGGCGTCTACATCACCACG TCATCGTTTTCCCGCCGTGCCCGCGAGGAAGCTGAGCGGATCAACGCCCG GATCGAACTCATCGACGGCGCTCGGCTGGCCGAGCTGCTCGTGCGGTATC GAGTCGGTGTCCAGGCGGTGCAGACCGTCGAACTCTTACGGCTCGACGAG GACTTTTTTGATGGCCTG

(SEQ ID NO: 121)

Amino acid sequence:

MTIPDAQTLMRPILAYLADGQAKSAKDVIAAMSDEFGLSDDERAQMLPSG RQRTMYDRVHWSLTHMSQAGLLDRPTRGHVQVTDTGRQVLKAHPERVDMA VLREFPSYIAFRERTKAKQPVDATAKRPSGDDVQVSPEDLIDAALAENRA AVEGEILKKALTLSPTGFEDLVIRLLEAMGYGRAGAVERTSASGDAGIDG IISQDPLGLDRIYVQAKRYAVDQTIGRPKIHEFAGALLGKQGDRGVYITT SSFSRGAREEAERINARIELIDGARLAELLVRYRVGVQAVQTVELLRLDE DFFDGL

(SEQ ID NO: 133)

[0143] The PCR reactions contained 100 ng Mtb genomic DNA, 25 nM final concentration of 5' and 3' primers. Polymerase, PCR buffer and nucleotides were from Clontech. The reaction temperature and times for the first PCR reaction were: 94° C. for 2 minutes, followed by 30 cycles of: 94° C. for 30 seconds, 48° C. for 1 min., and 68° C. for 2.5 minutes. [0144] Following the first PCR reaction, an aliquot of each PCR reaction containing 100 ng of PCR product from the previous step was transferred into a PCR reaction containing the TAP promoter and terminator fragments. The sequences of these fragments were:

[0145] Promoter Fragment: (SEQ ID NO:6)

5'CGGTCACGCTTGGGACTGCCATAGGCTGGCCCGGTGATGCCGGCCACG
ATGCGTCCGGCGTAGAGGATCTCGATCCCGCGAAATTAATACGA
CTCACTATAGGGAGACCACAACGGTTTCCCTCTAGAAATAATTTTGTTTA
ACTTTAAGAAGGAGGATATACC 3'

[0146] Terminator Fragment: TABLE-US-00003 (SEQ ID NO: 7)

5'GGGGGGGTTCTCATCATCATCATCATCATTAATAAAAGGGCGAATTC

CAGCACACTGGCGGCCGTTACTAGTGGATCCGGCTGCTAACAAAGCCCGA

AAGGAAGCTGAGTTGGCTGCTGCCACCGCTGAGCAATAACTAGCATAACC

CCTTGGGGCCTCTAAACGGGTCTTGAGGGGTTTTTTTGCTGAAAGGAGGAA

CTATATCCGGAGCGACTCCCACGGCACGTTGGCAAGCTCG 3'

[0147] The reaction temperature and times for the second PCR reaction were: 94° C. for 2 minutes, followed by 30 cycles of: 94° C. for 30 seconds, 48° C. for 60 seconds, and 68° C. for 2.5 minutes.

Protein Expression

[0148] The TAP fragments generated by PCR were used as templates for in vitro protein expression using a Roche RTS100 transcription/translation kit according to manufacturer's instructions. Approximately 0.5.about.1.0.mu.g PCR

product was used as template, producing approximately 0.5. about 5.0.mu.g of protein per template.

Protein Purification

[0149] MagneHis nickel-coated magnetic beads (Promega) were used to purify the expressed proteins. 15.mu.l of Nimagnetic beads (Promega) were pipetted into each well of a microtiter plate. To each well 50 μ l wash buffer (50 mM NaHPO₄, pH 8.0, 300 mM NaCl, 100 mM imidazole) was added with mixing and the plates were placed on a magnetic stand. The supernatant was removed and wash was repeated. 50 μ l of the Protein mixture was added with gentle pipetting. The mixture was incubated at room temperature for 2 minutes. The beads were then separated using a magnetic stand, washed 3 times with 150.mu.l wash buffer and the bound protein was eluted from the beads with 50 μ l of 50 mM NaHPO₄, pH 8.0, 300 mM NaCl, 250 mM imidazole.

Western Blot.

[0150] 15 μ l of the purified proteins were resolved on 4-12% SDS-polyacrylamide gels and transferred to nitrocellulose membranes The membranes were blocked in TBST/1% BSA, followed by incubation with TBST/1% BSA containing 1000-fold diluted rabbit anti-Mtb serum. The blots were washed and then incubated with alkaline phosphatase-conjugated goat-anti rabbit serum secondary antibody. Colorimetric development was used to develop the blots. The results of these analyses are shown in FIG. 6.

FLISA

[0151] The wells of Nunc-Immuno MaxiSorp 96-well plates were coated with 5 µl of expressed protein diluted in 95 ul PBS. The plates were mixed well on a shaker and then incubated overnight at 4° C. The plates were washed with PBS+0.05% Tween-20 for 5 min. with shaking at 200 rpm. 200 µl of PBS+1% BSA blocking solution was added and the plates incubated for 1 hr at room temperature, with shaking at 150 rpm. The blocking buffer was removed and 100 μl primary antibody (04.E293.1.11.WCL(-)LAM rabbit polyclonal antibodies 1:200000 diluted in blocking solution) was added. Following 1 hr incubation at room temperature with shaking at 150 rpm, the plates were washed 3 times with PBS+0.05% Tween-20. 100 µl second antibody (Anti-Rabbit IgG(H+L)-HRP conjugated, (Promega) diluted 1:2500 in blocking solution was added to each well and the plates were incubated for 1 hr at room temperature, with shaking at 150 rpm. After washing 3 times, 100 µl TMB substrate solution (Promega) was added to each well and the blue color was allowed to develop for 15 min at room temperature without shaking. 100 µl of 1N HCl was added to each well to stop the reaction and change the blue color to yellow. The plates were read in a spectrophotometer at 450 nm after 30 min. The results of this analysis are shown in FIG. 6.

[0152] As shown in FIG. 6, rabbit anti-Mtb serum identified 19 and 12 proteins that were reactive to the anti-serum Western blot and ELISA, respectively. The results showed a strong correlation in 'hits' between the two methods. In addition, a few antigen proteins at low abundance exhibited high reactivity relative to the others, suggesting the presence of strong B-cell epitopes, thus making them premier candidates for additional study.

Example 3

Using the Mtb Proteome to Identify the Antigenic Targets of Cell-Mediated Immunity in Mtb Vaccinated Mice and Humans

[0153] The following is a method that is used to systematically screen and identify antigens in Mtb that give rise to a

protective cell-mediated immune response. Through the use of TAP technology coding sequences of the Mtb genome are amplified. The PCR reactions are performed such that each amplified coding sequence becomes transcriptionally active. The resulting TAP fragments are expressed to produce Mtb polypeptides. Each of the polypeptides is delivered into dendritic cells, located in 96-well plates, using a polypeptide delivery reagent. Serum from Mtb immunized humans is added to each of the different wells.

[0154] An IFN-gamma. ELIspot assay is run using the following materials and method:

Materials:

[0155] Millipore 96-well multi-screen filtration plates (Millipore #MAIP S45-10) (Millipore, Bedford, Mass.)

[0156] Anti-IFN-.gamma. purified MAb (Clone 1-DIK) (MABTECH #3420-3) (Mabtech, Naka, Sweden)

[0157] Anti-EFN-g Biotinylated MAb (Clone 7-B6-1) (MABTECH #3420-6) (Mabtech, Naka, Sweden)

[0158] Streptavidin-Alkaline Phosphatase (MABTECH #3310-8) (Mabtech, Naka, Sweden

[0159] Alkaline Phosphate Substrate Kit (BIO-RAD #170-6432) (Bio-Rad, Hercules, Calif.)

[0160] Carbonate Buffer pH 9.6 (0.2.mu.M sterile filtered [0161] RPMI-1640 Medium (GIBCO #22400-089) (Gibco, Grand Island, N.Y.)

[0162] Fetal Bovine Serum (Sigma #F4135-500 mL) (Sigma, St. Louis, Mo.)

[0163] 1×PBS (Prepared from 10×PBS DIGENE #3400-1010) (DIGENE, Gaithersburg, Md.

 ${\bf [0164]}$ TWEEN® 20 (J. T. Baker #X251-07) (J. T. Baker, Phillipsburg, N.J.)

Method:

[0165] 96-well plates are coated with Coating Antibody (anti-IFN-g Clone 1-DIK) at 10-15 µg/mL (100 µL/well) and incubated at 4° C. overnight. Using aseptic technique, plates are flicked to remove Coating Antibody and washed 6 times with RPMI-1640. Plates are blocked with 100 µL/well of RPMI-1640+10% FBS (or Human AB serum) for 1-2 hours at room temperature. Plates are flicked to remove blocking buffer and $100\,\mu\text{L/well}$ of antigen specific or control peptides are added at a final concentration of 10 µg/well. Peripheral blood lymphocytes (PBL) are added at $4\times10^5/\text{well}$ and $1\times10^5/\text{well}$. Plates are incubated at 37° C./5% CO $_2$ for 36 hours. Plates are flicked to remove cells and washed 6 times with PBS+0.05% TWEEN® 20 at 200-250 µL/well. Plates are blot dried on paper towels.

[0166] Biotinylated antibody (anti-IFN-g Clone 7-B6-1) diluted 1:1,000 in 1.times. PBS at 100 $\mu\text{L/well}$ is added. The resulting solution is incubated for 3 hours at room temperature. Plates are flicked to remove biotinylated antibody and washed 6 times with PBS+0.05% TWEEN®. 20 at 200-250 $\mu\text{L/well}$. Plates are blot dried on paper towels. Streptavidin alkaline phosphatase is added at 100 $\mu\text{L/well}$ diluted 1:1,000 in 1×PBS. The plates are incubated for 1 hour at room temperature. Plates are flicked to remove the streptavidin alkaline phosphatase and washed 6 times with 0.05% TWEEN® 20 at 200-250 $\mu\text{L/well}$. The plates are washed again 3 times with 1×PBS at 200-250 $\mu\text{L/well}$. The plates are blot dried on paper towels

[0167] Substrate is added at $100\,\mu\text{L/well}$ for 10-15 minutes at room temperature. The substrate is prepared according to

25

manufacturer's protocol. The 25× substrate buffer is diluted in dH2O to a 1× concentration. Reagent A & B are each diluted 1:100 in the 1× substrate buffer. Rinsing plates with generous amounts of tap water (flooding plate and flicking several times) stops colorimetric substrate. Plates are allowed to dry overnight at room temperature in the dark. Spots corresponding to IFN-.gamma. producing cells are determined visually using a stereomicroscope (Zeiss KS ELIspot). Results can be expressed as the number of IFN-.gamma. secreting cells per 10⁶ spleen cells. Responses are considered positive if the response to test Mtb peptide epitope is significantly different (p<0.05) as compared with the response to no peptide and if the stimulation index (SI=response with test peptide/response with control peptide) is greater than 2.0.

Example 4

Cellular Vaccine Antigen Screen

[0168] A human volunteer was immunized with irradiated sporozoites from *P. falciparum*, the infectious agent responsible for malaria. Dendritic cells from the volunteer were isolated and cultured. Recombinant CSP polypeptide from *P. falciparum* was delivered to dendritic cells with or without polypeptide delivery reagents described in U.S. patent application Ser. No. 09/738,046, entitled "Intracellular Protein Delivery Reagent," which is hereby incorporated by reference in its entirety. T-cells isolated from the immunized volunteer were added to the cultures. The EliSpotassay identified 120 CSP antigen specific T-cells out of 250,000 T-cells that were added to the culture when CSP was added to the culture together with said delivery reagents. When CSP was added without said delivery reagents, the signal was barely above background.

Example 5

DNA Immunization of Mice

[0169] Experiments were set up with five animals per group, consisting of four week old BALB/c female mice, averaging 40 animals per experiment. These mice were immunized IM in each tibialis anterior muscle with 50 μ g plasmid DNA or transcriptionally active PCR fragment encoding selected Mtb antigens, 3 times at 3 week intervals. [0170] Sera was collected 10 days after each immunization for antibody studies. Blood samples (.about.50 ul) were collected from the mice by orbital bleed with a sterilized pasture pipette. The mice were bled about once a week at a volume of approximately 50 μ l.

[0171] Splenocytes were harvested at 14 days after the 3rd immunization and pooled for T-cell studies such as IFN-gamma. ELIspot assays. Tissue collections were performed on animals euthanized via $\rm CO_2$ (SOP 98.19) at the end of the experiment. The experiments can be five animals/group, averaging 40 animals/experiment×4 experiments for a total of 160 mice.

Example 6

Preparation of Human Dendritic Cells

[0172] Dendritic cells were ordered from Allcells: Cat #PB002 (NPB-Mononuclear Cells). The cells were in 50 mL buffer. The cells were counted immediately, the total number was 312.5×10⁶. The cells were pelleted, and resuspended in 25 mL RPMI-1640 containing DNAse. This solution (30

μg/mL) was incubated for 5 minutes at room temperature. The cells were washed twice with complete medium. The cells were resuspended at 10×10^6 cells/3 mL. Twelve 10 mm dishes containing 10 mL complete medium in each dish were used. The cells were incubated at 37° C. for 3 hours. The non-adherent cells were removed by gently shaking plates and aspirating the supernatant. Afterwards, the dishes containing adherent cells were washed 3 times with 10 mL of RPMI-1640 containing 2% Human Serum. 10 mL of culture medium were added to each plate containing 50 ng/mL GM-CSF and 500 u/mL IL-4. This culture medium was added until day 4. After day 4, culture medium without GM-CSF and IL-4 was added. The transfection was done on day 5. The complete medium consisted of RPMI-1640 (455 mL), 5% Human AB Serum (25 mL), Non-essential Amino Acids (5 mL), Sodium Pyruvate (5 mL), L-Glutamine (5 mL), and Penicillin-Streptomycin (5 mL).

Example 7

Generation of Dendritic Cells from Mouse Bone Marrow

[0173] Cells were taken from the bones of one mouse (2 femur and 2 tibiae without removing the macrophages). The red blood cells were obtained from the bone marrow and lysed. The cells were counted (51×10^6 cells, total) and cultured in a growth medium (2.5×10⁶ cells/plate, 10 mL/plate) for 8 days before transfection. On day 4 another 10 mL of growth medium was added. On day 6, 10 mL of the old medium was taken from each plate and the cells were pelleted. The cells were resuspended in 10 mL medium with 10 ng/mL GM-CSF and 2.5 ng/mL IL-4. The cells were placed back into the culture. The cells were cultured until transfection on day 8. On the day of transfection, 2.5×10^6 cells were harvested from each dish. The growth medium for mbmDC contained DMEM/Iscove, 10% FCS, 50 uM β-mercaptoethanol, 1× Penicillin/Streptomycin, 2 mM L-Glutamine, 10 mM Hepes, 1× Non-essential amino acids, 20 ng/mL rmGM-CSF, and 5 ng/mL rmIL-4.

Example 8

Adding an HA Epitope Tag

[0174] Oligos were designed using TAP promoter and terminator fragments from pCMVm and pTP-SV40, respectively, and adding the nucleotide sequence encoding the HA epitope tag. For adding the HA epitope to the 5' end of the coding sequence the following sequences is used: TABLE-US-00004 Promoter 5': CCGCCATGTTGACATTG (SEQ ID NO: 2) Promoter 3': GGCAGATCTGGGAGGCTAGCGTAATCCGGAACATCG (SEQ ID NO: 3) TATGGGTACATTGTTAAGTCGACGGTGC

[0175] For adding the HA epitope to the 3' end of the coding sequence, the following sequences is used:

TABLE-US-00005

Terminator 5':

 ${\tt GATCCCGGGTACCCATACGATGTTCCGGATTACGCT~(SEQ~ID~NO:~4)}\\ {\tt TAGGGGAGATCTCAGACATG}$

Terminator 3':

 ${\tt CAGGATATCATGCCTGCAGGACGACTCTAGAG} \qquad \quad ({\tt SEQ\ ID\ NO:\ 5})$

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[0176] The method includes:

[0177] PCR is used to amplify a new HA-promoter utilizing pCMVm as a template and a new HA-terminator utilizing pTP-SV40 as a template. The resulting PCR products are gel purified using QIAGEN QIAquick Gel Extraction Kit (Qiagen, Seattle, Wash.). The PCR products and both plasmids (pCMVm & pTP-SV40) are digested with EcoRV and BgIII restriction enzymes. All digested products are gel purified using QIAquick Gel Extraction Kit. The HA-promoter and HA-terminator are ligated separately into the digested pCMVm and pTP-SV40 plasmids. These plasmids are transformed into DH5, grown overnight on LB plates containing Kanamycin, colonies are selected and grown in LB media containing Kanamycin. QIAGEN QIAprep Spin Miniprep Kit is used to isolate plasmids. Plasmids are digested using EcoRV and BgIII Digests are run on a gel to identify clones containing plasmid with insert of correct size. The plasmids are sequenced to confirm inserts are correct. A prep culture is grown, plasmids are isolated, plasmids are digested with EcoRV and BgIII, and promoter and terminator fragments are gel purified. Epi-TAP-5 HA and Epi-TAP-3'HA kits are used.

Example 9

ICS

Intracellular Cytokine Staining (ICS)

[0178] Bone marrow derived dendritic cells (BMDCs) were prepared by culturing bone marrow cell suspensions with RPMI tissue culture media plus 10% fetal bovine serum and GM-CSF (20 ng/ml) for 6-7 days at 37° C., 5% CO₂. Cells were then primed with 1.mu.g/ml of antigen for 4 hrs at 37° C., 5% CO₂.

[0179] Cell suspensions obtained from naive or M. tuberculosis infected mice were used as a source of CD4 T cells. CD4 T cells are isolated by magnetic cell sorting and overlaid onto BMDC primed with specific antigens and cultured at 37° C. for 24 hrs. After this time T cells were harvested and stained for CD3/CD4/intracellular IFN.gamma. and analyzed by flow cytometry.

[0180] The sequences disclosed in Table 1 yielded positive results in at least one assay described herein, e.g. Western blot, ELISA or ICS.

Example 10

[0181] In One Embodiment the Method Includes Detection of Antigen-Specific CD4+ T-Cell Responses by Intracellular Cytokine Staining (ICS).

[0182] A panel of immunogenic Mtb proteins discovered in Phase I studies that were recognized by rabbit anti-TB sera was selected for further analysis to determine if these proteins could lead to enhanced induction of CD4+ T-cells. Thirty-six purified Mtb proteins along with positive controls, culture filtrate proteins (CFP), and recombinant ESAT-6, were included in the ICS assay. The results are summarized in Table 2 and demonstrate that 11 of the 36 proteins significantly stimulated CD4+ T-cell responses. Moreover, with equal protein amounts used, 6 Mtb proteins showed greater stimulatory activity than that of ESAT-6. TABLE-US-00006 TABLE 2 Antigen-specific stimulation of CD4⁺ T-cells ID Rv3733c Rv0138 Rv0740 Rv0733 Rv0009 Rv2882c Rv1065 Rv2613c Rv0475 Rv2114 Rv2466c Rv3763 Rv2031c % T-cells 4.3 4.3 8.3 3.7 4.0 3.6 2.8 2.0 2.1 2.9 2.6 2.5 2.2 ID Rv1347c Rv0158Rv3676 Rv2821Rv2108 Rv3226c Rv1056 Rv0815c Rv3117 Rv1073 Rv0097 ESAT-6 Media % T-cells 2.0 2.1 1.5 1.5 1.3 2.5 2.2 1.7 1.9 1.6 1.4 3.6 1.4

[0183] One µg each from 36 purified Mtb proteins, along with the control protein ESAT-6, were incubated with mouse dendritic cells for 24 hr. Spleen cells harvested from Mtbinfected mice were added and incubated for an additional 72 hr. The splenocytes were labeled with cychrome-conjugated anti-CD4 antibody and then stained with fluorescein-conjugated anti-.yIFN.gamma.he cells were washed, fixed and analyzed by flow cytometry. The "% T-cells" indicates the percentage of CD4+ T-cells that released yIFN. Based on previous studies, the percent value at or above 2.5% is significant.

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gtcaccggct ccgtggtgtg cacaaccgcg gccggcaatg tcaacatcgc gatcggcggg 2	40
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gggctcggta acgtcaacgg cgtcacgctg ggatacacgt cgggcaccgg acagggtaac 3	60
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cgactcaacc gacagetgge ggacategaa aagetteagg teaatgtgeg eeaageeetg 2	40
	00
	60
	20
	80
	40
	00
ateggttegg etgaacttge egagagtteg gtgeagggee ggatgetega ggtggageag 6	60

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caaatcgttc cgtattacga acccatgtac catcacgaag accacccgga gatctttgtc	240
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gaccgcggga cctatttcat cgatctcgcc agggtctggc agtcgctgcc cgccgccaag	420
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ccaataaagt ggcctacggt catccggcac ccaaagaccg gccaagagat cctctacatc	600
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catactcage actaccaggt tggcgacate atettgtggg acaaccgggt tetcatgcac	780
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caggaagate tgcccgagca gctcaccgag ctgcgtgaga agttcaccgc cgaggagctg	240
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egeggtgagg eegetetaga geggetgege ageeageaga gettegagga agtgteggeg	360
cgcgccgaag gctacgtgga ccaggcggtg gagttgaccc aggaggcgtt gggtacggtc	420
gcatcgcaga cccgcgcggt cggtgagcgt gccgccaagc tggtcggcat cgagctgcct	480
aagaaggetg eteeggeeaa gaaggeeget eeggeeaaga aggeegetee ggeeaagaag	540
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tacggccatg agaaggtcaa gttgctcgac ggcggccgca agaagtggga gctcgacgga	360
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gataacacga ttcgggcatt ccgcgacgag gtcctggcgg ccatcaacgt caagaacctc	480
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caggaacaaa gccagcggcc cggacacatt cctggtgcca tcaacgtgcc gtggagcagg	600
gccgccaacg aggacggcac cttcaagtcc gatgaggagt tggccaagct ttacgccgac	660
geeggeetag acaacageaa ggaaacgatt geetaetgee gaategggga aeggteeteg	720
cacacctggt tcgtgttgcg ggaattactc ggacaccaaa acgtcaagaa ctacgacggc	780
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ccgccatacg ggttgcgggt cgcacagctg accgacgcgg agatgttggc ggagtggatg	180
aaccgtcctc atctggcggc ggcctgggag tacgactggc cggcgtcacg ttggcgtcaa	240
cacctgaacg cccaacttga gggaacctat tcgttgccat tgatcggcag ctggcacgga	300
acagatggtg gttatctcga attatactgg gcagcaaagg atttgatttc tcactactac	360
gacgcagacc cctacgattt ggggctgcac gcggccatcg cggacttgtc gaaggtcaat	420
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cgttgccggc ggatcatgtt cgaccccgat caccgcaaca ccgcgacccg tcggttgtgt	540
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gataacacga ttcgggcatt ccgcgacgag gtcctggcgg ccatcaacgt caagaacctc	480
ategaegtge geteteeega egagttetee ggeaagatee tggeeeeege geacetgeeg	540
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gccgccaacg aggacggcac cttcaagtcc gatgaggagt tggccaagct ttacgccgac	660
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540

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38

600

660

720 756

60

60

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<212> TYPE: DNA

<213> ORGANISM: Mycobacterium tuberculosis

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<400> SEQUENCE: 56

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<211> LENGTH: 546

<212> TYPE: DNA

<213> ORGANISM: Mycobacterium tuberculosis

<400> SEQUENCE: 57

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<211> LENGTH: 849

<212> TYPE: DNA

<213 > ORGANISM: Mycobacterium tuberculosis

<400> SEQUENCE: 58

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aaccactgtc tactacacca ccgtggacgc ggcctggcgt ccggacacag tgtggcgata	660
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aagacgaagg tggggttgca gtcagcgctc gacacgttgc tggagtcgta taggggtcag	180

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621

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<212> TYPE: DNA
<213> ORGANISM: Mycobacterium tuberculosis
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Pro Thr Phe Asp Thr Arg Leu Met Arg Leu Glu Asp Glu Met Lys Glu
Gly Arg Tyr Glu Val Arg Ala Glu Leu Pro Gly Val Asp Pro Asp Lys
Asp Val Asp Ile Met Val Arg Asp Gly Gln Leu Thr Ile Lys Ala Glu
```

90 Ser Phe Val Arg Thr Val Ser Leu Pro Val Gly Ala Asp Glu Asp Asp 100 105 Ile Lys Ala Thr Tyr Asp Lys Gly Ile Leu Thr Val Ser Val Ala Val Ser Glu Gly Lys Pro Thr Glu Lys His Ile Gln Ile Arg Ser Thr Asn 135 140 <210> SEQ ID NO 66 <211> LENGTH: 159 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 66 Val Lys Arg Gly Leu Thr Val Ala Val Ala Gly Ala Ala Ile Leu Val Ala Gly Leu Ser Gly Cys Ser Ser Asn Lys Ser Thr Thr Gly Ser Gly Glu Thr Thr Thr Ala Ala Gly Thr Thr Ala Ser Pro Gly Ala Ala Ser Gly Pro Lys Val Val Ile Asp Gly Lys Asp Gln Asn Val Thr Gly Ser Val Val Cys Thr Thr Ala Ala Gly Asn Val Asn Ile Ala Ile Gly Gly Ala Ala Thr Gly Ile Ala Ala Val Leu Thr Asp Gly Asn Pro Pro Glu Val Lys Ser Val Gly Leu Gly Asn Val Asn Gly Val Thr Leu Gly Tyr 105 Thr Ser Gly Thr Gly Gln Gly Asn Ala Ser Ala Thr Lys Asp Gly Ser 120 His Tyr Lys Ile Thr Gly Thr Ala Thr Gly Val Asp Met Ala Asn Pro 135 Met Ser Pro Val Asn Lys Ser Phe Glu Ile Glu Val Thr Cys Ser 150 <210> SEQ ID NO 67 <211> LENGTH: 270 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 67 Met Ala Asn Pro Phe Val Lys Ala Trp Lys Tyr Leu Met Ala Leu Phe 10 Ser Ser Lys Ile Asp Glu His Ala Asp Pro Lys Val Gln Ile Gln Gln Ala Ile Glu Glu Ala Gln Arg Thr His Gln Ala Leu Thr Gln Gln Ala Ala Gln Val Ile Gly Asn Gln Arg Gln Leu Glu Met Arg Leu Asn Arg Gln Leu Ala Asp Ile Glu Lys Leu Gln Val Asn Val Arg Gln Ala Leu Thr Leu Ala Asp Gln Ala Thr Ala Ala Gly Asp Ala Ala Lys Ala Thr

Arg Thr Glu Gln Lys Asp Phe Asp Gly Arg Ser Glu Phe Ala Tyr Gly

105 Glu Gln Ser Val Glu Asp Leu Lys Thr Leu His Asp Gln Ala Leu Ser 120 Ala Ala Ala Gln Ala Lys Lys Ala Val Glu Arg Asn Ala Met Val Leu Gln Gln Lys Ile Ala Glu Arg Thr Lys Leu Leu Ser Gln Leu Glu Gln 150 155 Ala Lys Met Gln Glu Gln Val Ser Ala Ser Leu Arg Ser Met Ser Glu 170 Leu Ala Ala Pro Gly Asn Thr Pro Ser Leu Asp Glu Val Arg Asp Lys 185 Ile Glu Arg Arg Tyr Ala Asn Ala Ile Gly Ser Ala Glu Leu Ala Glu Ser Ser Val Gln Gly Arg Met Leu Glu Val Glu Gln Ala Gly Ile Gln 215 Met Ala Gly His Ser Arg Leu Glu Gln Ile Arg Ala Ser Met Arg Gly Glu Ala Leu Pro Ala Gly Gly Thr Thr Ala Thr Pro Arg Pro Ala Thr Glu Thr Ser Gly Gly Ala Ile Ala Glu Gln Pro Tyr Gly Gln <210> SEQ ID NO 68 <211> LENGTH: 289 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 68 Met Thr Leu Lys Val Lys Gly Glu Gly Leu Gly Ala Gln Val Thr Gly 10 Ile Val Tyr Thr Asn Lys Leu Val Val Leu Lys Asp Val His Pro Ser Pro Arg Glu Phe Ile Lys Leu Gly Arg Ile Ile Gly Gln Ile Val Pro Tyr Tyr Glu Pro Met Tyr His His Glu Asp His Pro Glu Ile Phe Val Ser Ser Thr Glu Glu Gly Gln Gly Val Pro Lys Thr Gly Ala Phe Trp His Ile Asp Tyr Met Phe Met Pro Glu Pro Phe Ala Phe Ser Met Val Leu Pro Leu Ala Val Pro Gly His Asp Arg Gly Thr Tyr Phe Ile Asp 120 Leu Ala Arg Val Trp Gln Ser Leu Pro Ala Ala Lys Arg Asp Pro Ala Arg Gly Thr Val Ser Thr His Asp Pro Arg Arg His Ile Lys Ile Arg 145 $\,$ 150 $\,$ 155 $\,$ 160 Pro Ser Asp Val Tyr Arg Pro Ile Gly Glu Val Trp Asp Glu Ile Asn Arg Thr Thr Pro Pro Ile Lys Trp Pro Thr Val Ile Arg His Pro Lys

Glu Tyr Asn Asn Ala Ala Glu Ala Phe Ala Ala Gln Leu Val Thr Ala

180 185 190												
Thr Gly Gln Glu Ile Leu Tyr Ile Cys Ala Thr Gly Thr Thr Lys Ile 195 200 205												
Glu Asp Lys Asp Gly Asn Pro Val Asp Pro Glu Val Leu Gln Glu Leu 210 215 220												
Met Ala Ala Thr Gly Gln Leu Asp Pro Glu Tyr Gln Ser Pro Phe Ile 225 230 240												
His Thr Gln His Tyr Gln Val Gly Asp Ile Ile Leu Trp Asp Asn Arg 245 250 255												
Val Leu Met His Arg Ala Lys His Gly Ser Ala Ala Gly Thr Leu Thr 260 265 270												
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Ala												
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Thr Asn Leu Arg Glu Arg Ala Glu Glu Thr Arg Thr Asp Thr Arg Ser 35 40 45												
Arg Val Glu Glu Ser Arg Ala Arg Leu Thr Lys Leu Gln Glu Asp Leu 50 55 60												
Pro Glu Gln Leu Thr Glu Leu Arg Glu Lys Phe Thr Ala Glu Glu Leu 65 70 75 80												
Arg Lys Ala Ala Glu Gly Tyr Leu Glu Ala Ala Thr Ser Arg Tyr Asn 85 90 95												
Glu Leu Val Glu Arg Gly Glu Ala Ala Leu Glu Arg Leu Arg Ser Gln 100 105 110												
Gln Ser Phe Glu Glu Val Ser Ala Arg Ala Glu Gly Tyr Val Asp Gln 115 120 125												
Ala Val Glu Leu Thr Gln Glu Ala Leu Gly Thr Val Ala Ser Gln Thr 130 135 140												
Arg Ala Val Gly Glu Arg Ala Ala Lys Leu Val Gly Ile Glu Leu Pro 145 150 155 160												
Lys Lys Ala Ala Pro Ala Lys Lys Ala Ala Pro Ala Lys Lys Ala Ala 165 170 175												
Pro Ala Lys Lys Ala Ala Ala Lys Lys Ala Pro Ala Lys Lys Ala Ala 180 185 190												
Ala Lys Lys Val Thr Gln Lys 195												
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<212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis												
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Met Ala Arg Cys Asp Val Leu Val Ser Ala Asp Trp Ala Glu Ser Asn

10

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65					70					/5					80
His	: Leu	Asn	Ala	Gln 85	Leu	Glu	Gly	Thr	Tyr 90	Ser	Leu	Pro	Leu	Ile 95	Gly
Sei	Trp	His	Gly 100	Thr	Asp	Gly	Gly	Tyr 105	Leu	Glu	Leu	Tyr	Trp 110	Ala	Ala
Lys	a Asp	Leu 115	Ile	Ser	His	Tyr	Tyr 120	Asp	Ala	Asp	Pro	Tyr 125	Asp	Leu	Gly
Let	His 130	Ala	Ala	Ile	Ala	Asp 135	Leu	Ser	Lys	Val	Asn 140	Arg	Gly	Phe	Gly
Pro 149	Leu	Leu	Leu	Pro	Arg 150	Ile	Val	Ala	Ser	Val 155	Phe	Ala	Asn	Glu	Pro 160
Arç	g Cys	Arg	Arg	Ile 165	Met	Phe	Asp	Pro	Asp 170	His	Arg	Asn	Thr	Ala 175	Thr
Arg	g Arg	Leu	Cys 180	Glu	Trp	Ala	Gly	Сув 185	Lys	Phe	Leu	Gly	Glu 190	His	Asp
Thi	Thr	Asn 195	Arg	Arg	Met	Ala	Leu 200	Tyr	Ala	Leu	Glu	Ala 205	Pro	Thr	Thr
Ala	Ala 210														
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Le	ı His	Ala	Pro 20	Lys	Val	Val	Phe	Val 25	Glu	Val	Asp	Glu	Asp 30	Thr	Ser
Ala	Tyr	Asp 35	Arg	Asp	His	Ile	Ala 40	Gly	Ala	Ile	ГЛа	Leu 45	Asp	Trp	Arg
Thi	7 Asp	Leu	Gln	Asp	Pro	Val 55	Lys	Arg	Asp	Phe	Val 60	Asp	Ala	Gln	Gln
Phe 65	e Ser	Lys	Leu	Leu	Ser 70	Glu	Arg	Gly	Ile	Ala 75	Asn	Glu	Asp	Thr	Val 80
Ile	e Leu	Tyr	Gly	Gly 85	Asn	Asn	Asn	Trp	Phe 90	Ala	Ala	Tyr	Ala	Tyr 95	Trp
Ту	. Phe	Lys		Tyr	Gly		Glu	-		ГЛа		Leu	-	-	Gly
Arç	l Fàa	Lys 115	Trp	Glu	Leu	Asp	Gly 120	Arg	Pro	Leu	Ser	Ser 125	Asp	Pro	Val
Sei	130	Pro	Val	Thr	Ser	Tyr 135	Thr	Ala	Ser	Pro	Pro 140	Asp	Asn	Thr	Ile
Arç 149	g Ala	Phe	Arg	Asp	Glu 150	Val	Leu	Ala	Ala	Ile 155	Asn	Val	Lys	Asn	Leu 160
Ile	e Asp	Val	Arg	Ser 165	Pro	Asp	Glu	Phe	Ser 170	Gly	Lys	Ile	Leu	Ala 175	Pro
Ala	Nis	Leu	Pro 180	Gln	Glu	Gln	Ser	Gln 185	Arg	Pro	Gly	His	Ile 190	Pro	Gly
Ala	a Ile	Asn 195	Val	Pro	Trp	Ser	Arg 200	Ala	Ala	Asn	Glu	Asp 205	Gly	Thr	Phe

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Lys Ser Asp Glu Glu Leu Ala Lys Leu Tyr Ala Asp Ala Gly Leu Asp
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Asn Ser Lys Glu Thr Ile Ala Tyr Cys Arg Ile Gly Glu Arg Ser Ser
                    230
His Thr Trp Phe Val Leu Arg Glu Leu Leu Gly His Gln Asn Val Lys
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Asn Tyr Asp Gly Ser Trp Thr Glu Tyr Gly Ser Leu Val Gly Ala Pro
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Ile Glu Leu Gly Ser
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Pro Tyr Arg Met Asn Tyr Leu Ala Glu Ala Pro Val Lys Arg Asp Pro
Asn Ser Ser Ala Ser Pro Ala Gln Pro Phe Thr Glu Ile Pro Gln Leu
Ser Asp Glu Glu Gly Leu Val Val Ala Arg Gly Lys Leu Val Tyr Ala 65 \phantom{000}70\phantom{000} 70 \phantom{0000}75\phantom{000} Rou Val Byr Ala 80 \phantom{0000}
Val Leu Asn Leu Tyr Pro Tyr Asn Pro Gly His Leu Met Val Val Pro
Tyr Arg Arg Val Ser Glu Leu Glu Asp Leu Thr Asp Leu Glu Ser Ala
           100
                               105
Glu Leu Met Ala Phe Thr Gln Lys Ala Ile Arg Val Ile Lys Asn Val
Ser Arg Pro His Gly Phe Asn Val Gly Leu Asn Leu Gly Thr Ser Ala
                135
Gly Gly Ser Leu Ala Glu His Leu His Val His Val Val Pro Arg Trp
                  150
                                        155
Gly Gly Asp Ala Asn Phe Ile Thr Ile Ile Gly Gly Ser Lys Val Ile
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Pro Gln Leu Leu Arg Asp Thr Arg Arg Leu Leu Ala Thr Glu Trp Ala
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Arg Gln Pro
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Lys Ile Thr Ala Ile Asp Glu Ala Thr Gly Cys Gly Gly Gly Lys Thr
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Uio															
1115	Ser 50	Glu	Pro	Asp	Asp	Glu 55	Pro	Thr	Arg	Arg	Val 60	Arg	Leu	Met	Arg
Trp 65	Gly	Leu	Ile	Pro	Ser 70	Trp	Ile	ГЛа	Ala	Gly 75	Pro	Gly	Gly	Ala	Pro 80
Asp	Ala	Lys	Gly	Pro 85	Pro	Leu	Ile	Asn	Ala 90	Arg	Ala	Asp	Lys	Val 95	Ala
Thr	Ser	Pro	Ala 100	Phe	Arg	Ser	Ala	Val 105	Arg	Ser	Lys	Arg	Cys 110	Leu	Val
Pro	Met	Asp 115	Gly	Trp	Tyr	Glu	Trp 120	Arg	Val	Asp	Pro	Asp 125	Ala	Thr	Pro
Gly	Arg 130	Pro	Asn	Ala	Lys	Thr 135	Pro	Phe	Phe	Leu	His 140	Arg	His	Asp	Gly
Ala 145	Leu	Leu	Phe	Thr	Ala 150	Gly	Leu	Trp	Ser	Val 155	Trp	Lys	Ser	Tyr	Arg 160
Ser	Ala	Pro	Pro	Leu 165	Leu	Ser	Cys	Thr	Val 170	Ile	Thr	Thr	Asp	Ala 175	Val
Gly	Glu	Leu	Ala 180	Glu	Ile	His	Asp	Arg 185	Met	Pro	Leu	Leu	Leu 190	Ala	Glu
Glu	Asp	Trp 195	Asp	Asp	Trp	Leu	Asn 200	Pro	Asp	Ala	Pro	Pro 205	Asp	Pro	Glu
Leu	Leu 210	Ala	Arg	Pro	Pro	Asp 215	Val	Arg	Asp	Ile	Ala 220	Leu	Arg	Gln	Val
		т	Val	Asn	Asn	Val	Arg	Asn	Asn	_	Pro	Glu	Leu	Leu	
Ser 225	Thr	ьец	741		230					235					240
225		Arg			230	Glu	Gln	Ile	Gln 250		Leu				240
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225 Pro <210 <211 <212 <213 <400 Val 1 Arg Tyr Ala Val 65 Thr	Ala >> SE >> Le >> TY >> OF Pro Leu Arg Leu Arg Met Ile	Arg GQ III ENGTH (PE: GGAN) Glu Glu Leu 35 Arg Pro	Ser NO NO H: 21 PRT FSM: Leu Asp Gly His Asp Leu Leu Leu	Gln 245 75 19 Mycc 75 Glu 5 Val Asp Gly Ser Pro 85 Glu	230 Pro Dbact Thr Gly Val Leu 70 Lys Leu	Pro Glu Arg Asp 55 Arg Leu	Asp His Met 40 Leu Ser Ile Asp	Asp Arg 25 Val His Asn Asp	250 Pro 10 Pro Met Pro Gly 90 Pro	Leu Glu Thr Ile Arg Ala 75 Gln Thr	Ser Phe Leu Leu Arg Asp Leu	Thr Gln 45 Leu Ala His	Gly 30 His Val Pro Ser Thr	Asp Pro Ala Phe Ala 95 Cys	Ala Ile Cys Pro Gly 80 Asp

Ser Tyr Asn Val Ala Pro Thr Asp Thr Ile Ala Thr Val Val Ser Arg 35 40 45

Asp Ser Thr Val Gly Pro Phe Asp Glu Ala Asp Leu Ile Glu Glu Trp 150 155 Val Thr Asp Arg Val Asp Asp Gly Ala Asp Pro Gln Ala Ala Glu His 165 Glu Cys Ala Ser Trp Leu Asp Glu Arg Ile Ser Gly Arg Thr Arg Arg Ala Leu Leu Ser Asp Arg Gln His Ala Ser Ser Ile Arg Arg Glu Ala 200 205 Arg Ser His Arg Lys Ser Val Lys Leu Ala Asp <210> SEQ ID NO 76 <211> LENGTH: 182 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 76 Met Ala Asp Cys Asp Ser Val Thr Asn Ser Pro Leu Ala Thr Ala Thr Asn His Ala Pro Lys Thr Val Ala Asn Phe Val Gly Leu Ala Gln Gly Thr Lys Asp Tyr Ser Thr Gln Asn Ala Ser Gly Gly Pro Ser Gly Pro 50Phe Tyr Asp Gly Ala Val Phe His Arg Val Ile Gln Gly Phe Met Ile 65 70 75 80 Gln Gly Gly Asp Pro Thr Gly Thr Gly Arg Gly Gly Pro Gly Tyr Lys Phe Ala Asp Glu Phe His Pro Glu Leu Gln Phe Asp Lys Pro Tyr Leu 105 Leu Ala Met Ala Asn Ala Gly Pro Gly Thr Asn Gly Ser Gln Phe Phe 120 Ile Thr Val Gly Lys Thr Pro His Leu Asn Arg Arg His Thr Ile Phe Gly Glu Val Ile Asp Ala Glu Ser Gln Arg Val Val Glu Ala Ile Ser 150 155 Lys Thr Ala Thr Asp Gly Asn Asp Arg Pro Thr Asp Pro Val Val Ile 170 Glu Ser Ile Thr Ile Ser 180 <210> SEQ ID NO 77 <211> LENGTH: 283 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 77 Met Gly Ala Gln Pro Phe Ile Gly Ser Glu Ala Leu Ala Ala Gly Leu Ile Ser Trp His Glu Leu Gly Lys Tyr Tyr Thr Ala Ile Met Pro Asn Val Tyr Leu Asp Lys Arg Leu Lys Pro Ser Leu Arg Gln Arg Val Ile \$35\$

Ala Ser Ala Leu His Gly Ala Lys Trp Val Asp Asp His Ala Leu Val Glu Leu Ile Trp Arg Asn Ala Arg Ala Pro Asn Gly Val Arg Thr Lys Asp Glu Leu Leu Asp Gly Glu Val Gln Arg Leu Cys Gly Leu Thr 105 Val Thr Thr Val Glu Arg Thr Ala Phe Asp Leu Gly Arg Arg Pro Pro 120 Leu Gly Gln Ala Ile Thr Arg Leu Asp Ala Leu Ala Asn Ala Thr Asp 135 Phe Lys Ile Asn Asp Val Arg Glu Leu Ala Arg Lys His Pro His Thr Arg Gly Leu Arg Gln Leu Asp Lys Ala Leu Asp Leu Val Asp Pro Gly Ala Gln Ser Pro Lys Glu Thr Trp Leu Arg Leu Leu Leu Ile Asn Ala Gly Phe Pro Arg Pro Ser Thr Gln Ile Pro Leu Leu Gly Val Tyr Gly 195 200 205 His Pro Lys Tyr Phe Leu Asp Met Gly Trp Glu Asp Ile Met Leu Ala Val Glu Tyr Asp Gly Glu Gln His Arg Leu Ser Arg Asp Gln Phe Val 230 235 Lys Asp Val Glu Arg Leu Glu Tyr Ile Arg Arg Ala Gly Trp Thr His 250 245 Ile Arg Val Leu Ala Asp His Lys Gly Pro Asp Val Val Arg Arg Val 265 Arg Gln Ala Trp Asp Thr Leu Thr Ser Arg Arg <210> SEQ ID NO 78 <211> LENGTH: 236 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 78 Met Met His Arg Thr Ala Leu Pro Ser Pro Pro Val Ala Lys Arg Val 10 Gln Thr Arg Arg Glu His His Gly Asp Val Phe Val Asp Pro Tyr Glu 25 Trp Leu Arg Asp Lys Asp Ser Pro Glu Val Ile Ala Tyr Leu Glu Ala Glu Asn Asp Tyr Thr Glu Arg Thr Thr Ala His Leu Glu Pro Leu Arg Gln Lys Ile Phe His Glu Ile Lys Ala Arg Thr Lys Glu Thr Asp Leu Ser Val Pro Thr Arg Arg Gly Asn Trp Trp Tyr Tyr Ala Arg Thr Phe Glu Gly Lys Gln Tyr Gly Val His Cys Arg Cys Pro Val Thr Asp Pro Asp Asp Trp Asn Pro Pro Glu Phe Asp Glu Arg Thr Glu Ile Pro Gly

Ala Ala Trp Leu Trp Ser Gly Arg Lys Gly Val Ile Ala Gly Ala Ser

55

		115					120					125			
Glu	Gln 130	Leu	Leu	Leu	Asp	Glu 135	Asn	Val	Glu	Ala	Asp 140	Gly	His	Asp	Phe
Phe 145	Ala	Leu	Gly	Ala	Ala 150	Ser	Val	Ser	Leu	Asp 155	Asp	Asn	Leu	Leu	Ala 160
Tyr	Ser	Val	Asp	Val 165	Val	Gly	Asp	Glu	Arg 170	Tyr	Thr	Leu	Arg	Phe 175	ГЛа
Asp	Leu	Arg	Thr 180	Gly	Glu	Gln	Tyr	Pro 185	Asp	Glu	Ile	Ala	Gly 190	Ile	Gly
Ala	Gly	Val 195	Thr	Trp	Ala	Ala	Asp 200	Asn	His	СЛа	Leu	Leu 205	His	His	Arg
Gly	Arg 210	Gly	Leu	Ala	Ser	Gly 215	His	Ser	Val	Ala	Ile 220	Pro	Thr	Arg	Val
Arg 225	Arg	Ile	Val	Gly	Ala 230	Gly	Leu	Pro	Arg	Ser 235	Arg				
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Asn	Ala	Leu 35	Ala	Ser	Glu	Leu	Glu 40	Lys	Thr	Lys	Val	Gly 45	Leu	Gln	Ser
Ala	Leu 50	Asp	Thr	Leu	Leu	Glu 55	Ser	Tyr	Arg	Gly	Gln 60	Ser	Ser	Gln	Ala
Leu 65	Ile	Gln	Gln	Thr	Leu 70	Pro	Tyr	Val	Gln	Trp 75	Leu	Thr	Thr	Thr	Ala 80
Glu	His	Ala	His	Lys 85	Thr	Ala	Ile	Gln	Leu 90	Thr	Ala	Ala	Ala	Asn 95	Ala
Tyr	Glu	Gln	Ala 100	Arg	Ala	Ala	Met	Val 105	Pro	Pro	Ala	Met	Val 110	Arg	Ala
Asn	Arg	Val 115	Gln	Thr	Thr	Val	Leu 120	Lys	Ala	Ile	Asn	Trp 125	Phe	Gly	Gln
Phe	Ser 130	Thr	Arg	Ile	Ala	Asp 135	Lys	Glu	Ala	Asp	Tyr 140	Glu	Gln	Met	Trp
Phe 145	Gln	Asp	Ala	Leu	Val 150	Met	Glu	Asn	Tyr	Trp 155	Glu	Ala	Val	Gln	Glu 160
Ala	Ile	Gln	Ser	Thr 165	Ser	His	Phe	Glu	Asp 170	Pro	Pro	Glu	Met	Ala 175	Asp
Asp	Tyr	Asp	Glu 180	Ala	Trp	Met	Leu	Asn 185	Thr	Val	Phe	Asp	Tyr 190	His	Asn
Glu	Asn	Ala 195	ГЛа	Glu	Glu	Val	Ile 200	His	Leu	Val	Pro	Asp 205	Val	Asn	ГЛа
Glu	Arg 210	Gly	Pro	Ile	Glu	Leu 215	Val	Thr	Lys	Val	Asp 220	Lys	Glu	Gly	Thr
Ile 225	Arg	Leu	Val	Tyr	Asp 230	Gly	Glu	Pro	Thr	Phe 235	Ser	Tyr	ГЛа	Glu	His 240

Pro Lys Phe

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			-0011011	iuea
115		120	125	
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Thr Asp Pro Tyr	Gln Leu Arg 165	Ala Ala Ile 170	Glu Arg Ala Gl	ı Ala Glu 175
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Thr Pro Ile Thr 50	Gln Leu Ala 55	Ser Ile Asn	Val Pro Glu Ala	a Arg Leu
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Ala Ile Arg Asn	Ser Asp Leu 85	Gly Val Asn 90	Pro Thr Asn Asp	Gly Ala 95
Leu Ile Arg Val 100	Ala Val Pro	Gln Leu Thr 105	Glu Glu Arg Arg	-
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Arg Asn Ile Arg 130	Arg Lys Ala 135		Leu His Arg Ilo 140	e Arg Lys
Glu Gly Glu Ala 145	Gly Glu Asp 150	Glu Val Gly	Arg Ala Glu Ly: 155	s Asp Leu 160
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<210> SEQ ID NO 88

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accaaactat tegeggegeg eggttateae ggeaccegga tggacgaegt egeegatgtg 120	
atcgggctca acaaagcaac ggtctatcac tactacgcca gcaagtcgct gatcctgttc 180	
gacatttacc gtcaggcggc cgagggcacc ctggccgccg tgcacgacga tccgtcctgg 240	
acggcccgtg aagcgctgta ccagtacacg gtccggctgc tcactgcgat cgcgagcaac 300	
cccgagcggg ccgccgtgta cttccaggag cagccctaca tcaccgagtg gttcaccagc 360	
gageaggteg eggaggteeg eggagaggag eageaagtet aegageaegt acaeggeetg 420	
atcgaccgcg ggattgccag cggcgagttc tatgagtgcg actcgcatgt ggtggcgctg 480	

gggtacatcg ggatgacgct gggcagctac cgctggctgc ggccgagcgg gcgccgaacg

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gccaaggaga tegeggegga gtteageaeg geaetgetge gegggetgat eegegaegaa	600
tegateegea accagtetee gettggaact eggaaggaaa eg	642
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gcactgacga aacagetgca geeegtegae tteeecegtg gacacaeggt ettegeggaa	120
ggggagccgg gcgatcggct gtacatcatc atctcgggga aggtcaagat cggtcgccgg	180
gcaccagacg gccgagaaaa cctgttaacc atcatgggcc cgtcggacat gttcggcgag	240
ttgtcgatct tcgacccggg tccgcgcacg tccagcgcga ccacgatcac cgaggtgcgg	300
geggtgtega tggacegega egegetgegg teatggateg eegategtee egaaatetee	360
gaacagetge tgegggtget ggeeegeegg etgegeegea ceaacaacaa eetggeegae	420
ctcatcttca ccgatgtgcc cggtcgggtg gccaagcagc tgttgcagct cgcccagcgt	480
ttcggcaccc aggaaggtgg cgcattgcgg gtcacccacg acctgacaca ggaagaaatc	540
gcccagctgg tcggggcctc acgcgagacg gtgaacaagg cactggctga tttcgctcac	600
cgcggctgga tccgccttga gggcaagagt gtgctgatct ctgactccga aagactggcc	660
cgccgagcga gg	672
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gateegetga geaggetgee gatgatteeg ggtaceagee tgaagggeaa ggteegeace	180
ttgctgtccc gccaatacgg cgccgacaca gaaacgtttt acaggaagcc gaatgaggac	240
cacgcccata tccgtcggct tttcggcgac accgaggagt acatgacggg ccgactcgtc	300
ttccgcgaca cgaagetcac caacaaagac gacetcgaag cccgcggcgc taagactete	360
accgaggtga aattcgagaa cgccatcaac cgggtgaccg caaaggcaaa ccttcgccag	420
atggaacgeg tgateecegg cagegagtte gegtteteae ttgtetaega ggteteette	480
ggcacccccg gcgaggaaca gaaggcgtct ctgccttcct ccgatgagat catcgaggac	540
ttcaacgcca tcgcgcgcgg cctgaagttg ctcgaactcg actacctcgg cggcagcgga	600
accegtgget aegggeaggt caagtteage aacetgaaag eeegegeege agteggegee	660
ctcgacggtt ctctgctgga gaagctaaac catgaactcg cggctgtt	708
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<210> SEQ ID NO 119 <211> LENGTH: 762 <212> TYPE: DNA <213> ORGANISM: Mycobacterium tuberculosis

<400> SEQUENCE: 119

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tgggaggttc cctactaccc gcagtactac atcccgctgg cggatgtccg catggagttc	180
ctgcgcgacg agaaccaccc gcagcgagtg cagctgggtc cgtcgcggct gcactccttg	240
gtaagegeeg gteagaeeea eegateggeg gegegggtat tegatgtega eggegaeage	300
ccggtggcgg gcaccgtgcg tttcaactgg gatccgctgc ggtggttcga ggaggacgag	360
ccgatctacg gccatccgcg caatccctat cagcgggccg atgcgctgcg ctcgcaccga	420
cacgteegtg tegagetgga eggeattgtg etegetgaca eeegategee egttetgeta	480
ttcgaaactg ggatacccac aaggtattac atcgatccgg ccgacatcgc tttcgagcat	540
ctggagccca cctcgacgca gacgttgtgt ccgtacaagg ggacgacgtc gggctattgg	600
tetgtgegeg teggegaege egtgeaeege gaeetggeet ggaegtatea etatecaetg	660
cccgccgttg ccccgatcgc cggcctggtg gcgttttaca acgagaaggt cgacctcacc	720
gtegaeggeg tegeeetgee geggeegeae acteagttea ge	762
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ttgctcggca aacaccttgg cgtcggggtc accagcatct actggtactt ccgcaagaag	180
gacgatetge teaacgegat gacegacege getttgagea agtacgtgtt egetaceeeg	240
tacategaag eeggegactg gegegaaacg ttgegeaate atgeeegete gatgeggaag	300
acgttegegg acaacecegt actgtgegat etgatactga ttegagegge getgteeceg	360
aaaacggcgc ggttgggcgc ccaagagatg gagaaggcca tcgccaatct ggtgacggcg	420
ggcctgtcgc tcgaagacgc tttcgacatc tactcggcgg tttcggtcca cgtgcgcgga	480
teggtggtgc tagategget etceegeaag ageeagtegg egggeagegg accateegee	540
attgaacacc cegtggecat egatecegeg acgaeteege tgettgetea egeaactggg	600
agggggcatc ggatcggggc ccccgatgaa accaatttcg aatatggtct cgaatgcatc	660
ctcgaccatg ctggccggtt gatcgaacaa agctcgaaag ccgctggtga ggtcgcagtg	720
egeegeeeca eggeeaeege egatgegeet aegeegggeg egegggeeaa ageggtggeg	780
cgt	783
<210> SEQ ID NO 121 <211> LENGTH: 918 <212> TYPE: DNA <213> ORGANISM: Mycobacterium tuberculosis	
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caagegaagt eggeeaagga egteategeg gegatgteeg aegagttegg tetgteegae	120
	100

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<400> SEQUENCE: 123

Met Leu Pro Lys Asn Thr Arg Pro Thr Ser Glu Thr Ala Glu Glu Phe

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caggtcacgg acacgggccg tcaagtcctg aaggcgcatc ccgagcgcgt cgacatggct	300
gtgctgcggg agttcccgtc gtacatcgct tttcgtgagc gaaccaaagc caagcagcca	360
gtcgacgcga ccgccaagcg accgtccggg gacgatgtgc aggtctcacc cgaggatctc	420
atcgacgctg cgcttgcgga gaaccgggca gccgtcgagg gggagatcct gaagaaggca	480
ctcacgttgt cgcccaccgg gtttgaagat ctggttatca gacttttgga ggcgatgggt	540
tacgggcgag ccggcgcggt ggaacggacg agtgcctccg gtgacgctgg catcgacgga	600
atcatcagec aggacceget egggetggae egeatetaeg tgeaggeeaa gegataegee	660
gtcgaccaaa cgattggccg gccgaagatc cacgagttcg ccggcgccct cctgggcaag	720
cagggcgacc ggggcgtcta catcaccacg tcatcgtttt cccgcggtgc ccgcgaggaa	780
gctgagcgga tcaacgcccg gatcgaactc atcgacggcg ctcggctggc cgagctgctc	840
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gacttttttg atggcctg	918
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Lys Phe Glu Lys Thr Val Ala Arg Ala Ala Ala Thr Arg Asp Trp Asp 20 25 30	
Cys Trp Val Gln His Tyr Thr Pro Asp Val Glu Tyr Ile Glu His Ala 35 40 45	
Ala Gly Ile Met Arg Gly Arg Gln Arg Val Arg Ala Trp Ile Gln Glu 50 55 60	
Thr Met Thr Thr Phe Pro Gly Ser His Met Val Ala Phe Pro Ser Leu 65 70 75 80	
Trp Ser Val Ile Asp Glu Ser Thr Gly Arg Ile Ile Cys Glu Leu Asp 85 90 95	
Asn Pro Met Leu Asp Pro Gly Asp Gly Ser Val Ile Ser Ala Thr Asn	
Ile Ser Ile Ile Thr Tyr Ala Gly Asn Gly Gln Trp Cys Arg Gln Glu	
115 120 125	
Asp Ile Tyr Asn Pro Leu Arg Phe Leu Arg Ala Ala Met Lys Trp Cys 130 135 140	
Arg Lys Ala Gln Glu Leu Gly Thr Leu Asp Glu Asp Ala Ala Arg Trp 145 150 155 160	
Met Arg Arg His Gly Gly Pro 165	
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Thr	Arg	Thr 35	Val	Thr	Val	Ser	Ile 40	Сув	Gln	Val	Ala	Asp 45	Gly	Glu	Arg
Glu	Ala 50	Glu	Gly	Val	Arg	Asp 55	Met	Met	Arg	Leu	Glu 60	CAa	Pro	Ala	Gly
Leu 65	Asp	Leu	Arg	Thr	Pro 70	Asn	Pro	Glu	Ala	Tyr 75	Glu	Ile	Thr	Gly	Gln 80
Arg	Pro	Gly	Glu	Phe 85	Val	Phe	Val	Leu	Gly 90	Tyr	Leu	Gly	His	Val 95	Arg
Ala	Ile	Val	Gly 100	Asn	CAa	Tyr	Ile	Glu 105	Ile	Met	Pro	Met	Gly 110	Thr	Arg
Val	Glu	Leu 115	Ser	Lys	Leu	Ala	Asp 120	Val	Ala	Leu	Asp	Ile 125	Gly	Arg	Ser
Val	Gly 130	Сув	Ser	Ala	Tyr	Glu 135	Asn	Asp	Phe	Thr	Leu 140	Pro	Asp	Ile	Pro
Thr 145	Gln	Trp	Arg	Asn	Gln 150	Pro	Leu	Gly	Trp	Tyr 155	Thr	Gln	Gly	Leu	Ala 160
Pro	Tyr	Leu	Pro	Gly 165	Leu	Ser	Asp	Pro	Lys 170	Asp	Ala	Ala	Glu	Gly 175	
<213 <400	2 > TY 3 > OF 0 > SE	RGAN: EQUEI	ISM: ICE:	124							a-		~-	m)	a-
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Ala	Val	ГЛа	Leu 20	Ala	Glu	Lys	Leu	Gly 25	Ile	Pro	Gln	Ile	Ser 30	Thr	Gly
Glu	Leu	Phe 35	Arg	Arg	Asn	Ile	Glu 40	Glu	Gly	Thr	Lys	Leu 45	Gly	Val	Glu
Ala	Lys 50	Arg	Tyr	Leu	Asp	Ala 55	Gly	Asp	Leu	Val	Pro 60	Ser	Asp	Leu	Thr
Asn 65	Glu	Leu	Val	Asp	Asp 70	Arg	Leu	Asn	Asn	Pro 75	Asp	Ala	Ala	Asn	Gly 80
Phe	Ile	Leu	Asp	Gly 85	Tyr	Pro	Arg	Ser	Val 90	Glu	Gln	Ala	ГÀв	Ala 95	Leu
His	Glu	Met	Leu 100	Glu	Arg	Arg	Gly	Thr 105		Ile	Asp	Ala	Val 110	Leu	Glu
Phe	Arg	Val 115	Ser	Glu	Glu	Val	Leu 120	Leu	Glu	Arg	Leu	Lys 125	Gly	Arg	Gly
Arg	Ala 130	Asp	Asp	Thr	Asp	Asp 135	Val	Ile	Leu	Asn	Arg 140	Met	Lys	Val	Tyr
Arg 145	Asp	Glu	Thr	Ala	Pro 150	Leu	Leu	Glu	Tyr	Tyr 155	Arg	Asp	Gln	Leu	Lys 160
Thr	Val	Asp	Ala	Val 165	Gly	Thr	Met	Asp	Glu 170	Val	Phe	Ala	Arg	Ala 175	Leu
Arg	Ala	Leu	Gly 180	Lys											

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Ala Asp Asp Val Leu Gly Gly Arg Cys Asp His Leu Leu Pro Asp Gly
                           40
Gly Val Pro Gln Thr Gln Arg Trp Tyr Thr Arg Ile His Gly Asp Glu
Glu Leu Asp Ile Trp Leu Ile Ser Trp Val Pro Gly Gln Pro Thr Glu
Leu His Asp His Gly Gly Ser Leu Gly Ala Leu Thr Val Leu Ser Gly
Ser Leu Asn Glu Tyr Arg Trp Asp Gly Arg Arg Leu Arg Arg Arg Arg
Leu Asp Ala Gly Asp Gln Ala Gly Phe Pro Leu Gly Trp Val His Asp $115$ $120$ $125$
Val Val Trp Ala Pro Arg Pro Ile Gly Gly Pro Asp Ala Ala Gly Met 130 $135$
Ala Val Ala Pro Thr Leu Ser Val His Ala Tyr Ser Pro Pro Leu Thr
Ala Met Ser Tyr Tyr Glu Ile Thr Glu Arg Asn Thr Leu Arg Arg Gln
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Arg Thr Glu Leu Thr Asp Gln Pro Glu Gly Ser Gly
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<210> SEQ ID NO 126
<211> LENGTH: 207
<212> TYPE: PRT
<213> ORGANISM: Mycobacterium tuberculosis
<400> SEQUENCE: 126
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Tyr Asn Ser Phe Pro Leu Asn Asp Cys Pro Ala Glu Leu Trp Ser Ala
Leu Asp Pro Gln Ala Leu Ala Thr Glu His Lys Ala Ala Thr Ala Leu
Leu Asn Gly Pro Arg Tyr Trp Leu Met Asn Ala Ile Glu Lys Ala Pro
Gln Gly Pro Pro Val Thr Lys Thr Phe Gly Gly Ile Glu Met Leu Gln
Gln Ala Thr Val Leu Leu Ser Ser Met Asn Pro Ala Pro Tyr Thr Val
                     105
Ser Gln Val Ser Arg Asn Thr Val Phe Val Phe Asn Ala Gly Glu Glu
                           120
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Val Tyr Glu Leu Gln Asp Pro Lys Gly Gln Arg Trp Val Met Gln Thr 135 Trp Ser Gln Val Val Asp Pro Asn Leu Ser Arg Ala Asp Leu Pro Lys 150 Leu Gly Glu Arg Leu Asn Leu Pro Ala Gly Trp Ser Tyr His Thr Arg 170 165 Val Leu Thr Ser Glu Leu Arg Val Asp Thr Thr Asn Arg Glu Ala Arg 185 Val Leu Gln Asp Asp Leu Thr Asn Ser Tyr Ser Leu Val Thr Ala 195 200 <210> SEQ ID NO 127 <211> LENGTH: 207 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 127 Met Leu Glu Lys Ala Pro Gln Lys Ser Val Ala Asp Phe Trp Phe Asp Pro Leu Cys Pro Trp Cys Trp Ile Thr Ser Arg Trp Ile Leu Glu Val Ala Lys Val Arg Asp Ile Glu Val Asn Phe His Val Met Ser Leu Ala Ile Leu Asn Glu Asn Arg Asp Asp Leu Pro Glu Gln Tyr Arg Glu Gly 50 $\,$ 60 $\,$ Met Ala Arg Ala Trp Gly Pro Val Arg Val Ala Ile Ala Ala Glu Gln 65 70 75 75 80 Ala His Gly Ala Lys Val Leu Asp Pro Leu Tyr Thr Ala Met Gly Asn 90 Arg Ile His Asn Gln Gly Asn His Glu Leu Asp Glu Val Ile Thr Gln 105 Ser Leu Ala Asp Ala Gly Leu Pro Ala Glu Leu Ala Lys Ala Ala Thr Ser Asp Ala Tyr Asp Asn Ala Leu Arg Lys Ser His His Ala Gly Met 135 Asp Ala Val Gly Glu Asp Val Gly Thr Pro Thr Ile His Val Asn Gly 150 155 Val Ala Phe Phe Gly Pro Val Leu Ser Lys Ile Pro Arg Gly Glu Glu 170 Ala Gly Lys Leu Trp Asp Ala Ser Val Thr Phe Ala Ser Tyr Pro His Phe Phe Glu Leu Lys Arg Thr Arg Thr Glu Pro Pro Gln Phe Asp 200 <210> SEQ ID NO 128 <211> LENGTH: 214 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 128 Met Pro Ser Asp Thr Ser Pro Asn Gly Leu Ser Arg Arg Glu Glu Leu Leu Ala Val Ala Thr Lys Leu Phe Ala Ala Arg Gly Tyr His Gly Thr

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Tyr	His 50	Tyr	Tyr	Ala	Ser	Lув 55	Ser	Leu	Ile	Leu	Phe 60	Aap	Ile	Tyr	Arg
Gln 65	Ala	Ala	Glu	Gly	Thr 70	Leu	Ala	Ala	Val	His 75	Asp	Asp	Pro	Ser	Trp 80
Thr	Ala	Arg	Glu	Ala 85	Leu	Tyr	Gln	Tyr	Thr 90	Val	Arg	Leu	Leu	Thr 95	Ala
Ile	Ala	Ser	Asn 100	Pro	Glu	Arg	Ala	Ala 105	Val	Tyr	Phe	Gln	Glu 110	Gln	Pro
Tyr	Ile	Thr 115	Glu	Trp	Phe	Thr	Ser 120	Glu	Gln	Val	Ala	Glu 125	Val	Arg	Glu
ràs	Glu 130	Gln	Gln	Val	Tyr	Glu 135	His	Val	His	Gly	Leu 140	Ile	Asp	Arg	Gly
Ile 145	Ala	Ser	Gly	Glu	Phe 150	Tyr	Glu	CÀa	Asp	Ser 155	His	Val	Val	Ala	Leu 160
Gly	Tyr	Ile	Gly	Met 165	Thr	Leu	Gly	Ser	Tyr 170	Arg	Trp	Leu	Arg	Pro 175	Ser
Gly	Arg	Arg	Thr 180	Ala	Lys	Glu	Ile	Ala 185	Ala	Glu	Phe	Ser	Thr 190	Ala	Leu
Leu	Arg	Gly 195	Leu	Ile	Arg	_	Glu 200	Ser	Ile	Arg	Asn	Gln 205	Ser	Pro	Leu
Gly	Thr 210	Arg	Lys	Glu	Thr										
)> SE -> LE														
	2 > T\ 3 > OF			Mycc	bact	eriu	ım tu	ıberc	ulos	sis					
< 400)> SE	EQUEN	ICE :	129											
)> SE Asp				Ala	Arg	Ala	Gly	Ile 10	Phe	Gln	Gly	Val	Glu 15	Pro
Val 1		Glu	Ile	Leu 5		_		_	10					15	
Val 1 Ser	qaA	Glu Ile	Ile Ala 20	Leu 5 Ala	Leu	Thr	Гуз	Gln 25	10 Leu	Gln	Pro	Val	Asp 30	15 Phe	Pro
Val 1 Ser Arg	Asp Ala	Glu Ile His 35	Ile Ala 20 Thr	Leu 5 Ala Val	Leu Phe	Thr Ala	Lys Glu 40	Gln 25 Gly	10 Leu Glu	Gln Pro	Pro Gly	Val Asp 45	Asp 30 Arg	15 Phe Leu	Pro Tyr
Val 1 Ser Arg	Asp Ala Gly Ile	Glu Ile His 35 Ile	Ile Ala 20 Thr	Leu 5 Ala Val Gly	Leu Phe Lys	Thr Ala Val 55	Lys Glu 40 Lys	Gln 25 Gly Ile	10 Leu Glu Gly	Gln Pro Arg	Pro Gly Arg 60	Val Asp 45 Ala	Asp 30 Arg Pro	15 Phe Leu Asp	Pro Tyr Gly
Val 1 Ser Arg Ile Arg 65	Asp Ala Gly Ile 50	Glu Ile His 35 Ile Asn	Ile Ala 20 Thr Ser	Leu 5 Ala Val Gly Leu	Leu Phe Lys Thr 70	Thr Ala Val 55	Lys Glu 40 Lys Met	Gln 25 Gly Ile Gly	10 Leu Glu Gly Pro	Gln Pro Arg Ser 75	Pro Gly Arg 60 Asp	Val Asp 45 Ala Met	Asp 30 Arg Pro	15 Phe Leu Asp Gly	Pro Tyr Gly Glu 80
Val 1 Ser Arg Ile Arg 65 Leu	Asp Ala Gly Ile 50 Glu	Glu Ile His 35 Ile Asn Ile	Ile Ala 20 Thr Ser Leu Phe	Leu 5 Ala Val Gly Leu Asp 85	Leu Phe Lys Thr 70	Thr Ala Val 55 Ile	Lys Glu 40 Lys Met	Gln 25 Gly Ile Gly	10 Leu Glu Gly Pro Thr 90	Gln Pro Arg Ser 75 Ser	Pro Gly Arg 60 Asp	Val Asp 45 Ala Met	Asp 30 Arg Pro Phe	15 Phe Leu Asp Gly Thr 95	Pro Tyr Gly Glu 80 Ile
Val 1 Ser Arg Ile Arg 65 Leu	Asp Ala Gly Ile 50 Glu Ser	Glu Ile His 35 Ile Asn Ile Val	Ile Ala 20 Thr Ser Leu Phe Arg 100	Leu 5 Ala Val Gly Leu Asp 85 Ala	Leu Phe Lys Thr 70 Pro	Thr Ala Val 55 Ile Gly Ser	Lys Glu 40 Lys Met Pro	Gln 25 Gly Ile Gly Arg Asp	10 Leu Glu Gly Pro Thr 90 Arg	Gln Pro Arg Ser 75 Ser Asp	Pro Gly Arg 60 Asp Ser	Val Asp 45 Ala Met Ala	Asp 30 Arg Pro Phe Thr Arg 110	15 Phe Leu Asp Gly Thr 95 Ser	Pro Tyr Gly Glu 80 Ile
Val 1 Ser Arg Ile Arg 65 Leu Thr	Asp Ala Gly Ile 50 Glu Ser Glu	Glu Ile His 35 Ile Asn Ile Val Asp 115	Ala 20 Thr Ser Leu Phe Arg 100 Arg	Leu 5 Ala Val Gly Leu Asp 85 Ala	Leu Phe Lys Thr 70 Pro Val	Thr Ala Val 55 Ile Gly Ser Ile	Lys Glu 40 Lys Met Pro Met Ser 120	Gln 25 Gly Ile Gly Arg Asp 105 Glu	10 Leu Glu Gly Pro Thr 90 Arg	Gln Pro Arg Ser 75 Ser Asp	Pro Gly Arg 60 Asp Ser Ala Leu	Val Asp 45 Ala Met Ala Leu Arg 125	Asp 30 Arg Pro Phe Thr Arg 110 Val	15 Phe Leu Asp Gly Thr 95 Ser Leu	Pro Tyr Gly Glu 80 Ile Trp Ala
Val 1 Ser Arg Ile Arg 65 Leu Thr	Asp Ala Gly Ile 50 Glu Ser Glu Ala Arg	Glu Ile His 35 Ile Asn Ile Val Asp 115 Leu	Ala 20 Thr Ser Leu Phe Arg 100 Arg Arg	Leu 5 Ala Val Gly Leu Asp 85 Ala Pro Arg	Leu Phe Lys Thr 70 Pro Val Glu Thr	Thr Ala Val 55 Ile Gly Ser Ile Asn 135	Lys Glu 40 Lys Met Pro Met Ser 120 Asn	Gln 25 Gly Ile Gly Arg Asp 105 Glu Asn	10 Leu Glu Gly Pro Thr 90 Arg Gln Leu	Gln Pro Arg Ser 75 Ser Asp Leu Ala	Pro Gly Arg 60 Asp Ser Ala Leu Asp 140	Val Asp 45 Ala Met Ala Leu Arg 125 Leu	Asp 30 Arg Pro Phe Thr Arg 110 Val	15 Phe Leu Asp Gly Thr 95 Ser Leu Phe	Pro Tyr Gly Glu 80 Ile Trp Ala
Val 1 Ser Arg Ile Arg 65 Leu Thr Ile Arg Asp 145	Asp Ala Gly Ile 50 Glu Ser Glu Ala Arg 130	Glu Ile His 35 Ile Asn Ile Val Asp 115 Leu Pro	Ala 20 Thr Ser Leu Phe Arg 100 Arg Gly	Leu 5 Ala Val Gly Leu Asp 85 Ala Pro Arg	Leu Phe Lys Thr 70 Pro Val Glu Thr Val 150	Thr Ala Val 55 Ile Gly Ser Ile Asn 135 Ala	Lys Glu 40 Lys Met Pro Met Ser 120 Asn	Gln 25 Gly Ile Gly Arg 105 Glu Asn	10 Leu Glu Gly Pro Thr 90 Arg Gln Leu	Gln Pro Arg Ser 75 Ser Asp Leu Ala Leu 155	Pro Gly Arg 60 Asp Ser Ala Leu Asp 140 Gln	Val Asp 45 Ala Met Ala Leu Arg 125 Leu Leu	Asp 30 Arg Pro Phe Thr Arg 110 Val Ile	15 Phe Leu Asp Gly Thr 95 Ser Leu Phe	Pro Tyr Gly Glu 80 Ile Trp Ala Thr

Arg Met Asp Asp Val Ala Asp Val Ile Gly Leu Asn Lys Ala Thr Val

Gln Glu Glu Ile Ala Gln Leu Val Gly Ala Ser Arg Glu Thr Val Asn 185 Lys Ala Leu Ala Asp Phe Ala His Arg Gly Trp Ile Arg Leu Glu Gly 200 Lys Ser Val Leu Ile Ser Asp Ser Glu Arg Leu Ala Arg Arg Ala Arg 215 <210> SEQ ID NO 130 <211> LENGTH: 236 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 130 Met Thr Thr Ser Tyr Ala Lys Ile Glu Ile Thr Gly Thr Leu Thr Val Leu Thr Gly Leu Gln Ile Gly Ala Gly Asp Gly Phe Ser Ala Ile Gly Ala Val Asp Lys Pro Val Val Arg Asp Pro Leu Ser Arg Leu Pro Met Ile Pro Gly Thr Ser Leu Lys Gly Lys Val Arg Thr Leu Leu Ser Arg Gln Tyr Gly Ala Asp Thr Glu Thr Phe Tyr Arg Lys Pro Asn Glu Asp His Ala His Ile Arg Arg Leu Phe Gly Asp Thr Glu Glu Tyr Met Thr 85 90 95 Glu Ala Arg Gly Ala Lys Thr Leu Thr Glu Val Lys Phe Glu Asn Ala 120 Ile Asn Arg Val Thr Ala Lys Ala Asn Leu Arg Gln Met Glu Arg Val 140 135 Ile Pro Gly Ser Glu Phe Ala Phe Ser Leu Val Tyr Glu Val Ser Phe 150 155 Gly Thr Pro Gly Glu Glu Gln Lys Ala Ser Leu Pro Ser Ser Asp Glu Ile Ile Glu Asp Phe Asn Ala Ile Ala Arg Gly Leu Lys Leu Leu Glu 185 Leu Asp Tyr Leu Gly Gly Ser Gly Thr Arg Gly Tyr Gly Gln Val Lys 200 Phe Ser Asn Leu Lys Ala Arg Ala Ala Val Gly Ala Leu Asp Gly Ser 215 Leu Leu Glu Lys Leu Asn His Glu Leu Ala Ala Val <210> SEQ ID NO 131 <211> LENGTH: 254 <212> TYPE: PRT <213 > ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 131 Met Ser Val Asp Tyr Pro Gln Met Ala Ala Thr Arg Gly Arg Ile Glu Pro Ala Pro Arg Arg Val Arg Gly Tyr Leu Gly His Val Leu Val Phe $20 \hspace{1cm} 25 \hspace{1cm} 30 \hspace{1cm}$

Asn His Pro Gln Arg Val Gln Leu Gly Pro Ser Arg Leu His Ser Leu Val Ser Ala Gly Gln Thr His Arg Ser Ala Ala Arg Val Phe Asp Val Asp Gly Asp Ser Pro Val Ala Gly Thr Val Arg Phe Asn Trp Asp Pro 105 Leu Arg Trp Phe Glu Glu Asp Glu Pro Ile Tyr Gly His Pro Arg Asn Pro Tyr Gln Arg Ala Asp Ala Leu Arg Ser His Arg His Val Arg Val Glu Leu Asp Gly Ile Val Leu Ala Asp Thr Arg Ser Pro Val Leu Leu 155 Phe Glu Thr Gly Ile Pro Thr Arg Tyr Tyr Ile Asp Pro Ala Asp Ile Ala Phe Glu His Leu Glu Pro Thr Ser Thr Gln Thr Leu Cys Pro Tyr Lys Gly Thr Thr Ser Gly Tyr Trp Ser Val Arg Val Gly Asp Ala Val His Arg Asp Leu Ala Trp Thr Tyr His Tyr Pro Leu Pro Ala Val Ala 215 Pro Ile Ala Gly Leu Val Ala Phe Tyr Asn Glu Lys Val Asp Leu Thr 230 235 Val Asp Gly Val Ala Leu Pro Arg Pro His Thr Gln Phe Ser 245 <210> SEQ ID NO 132 <211> LENGTH: 261 <212> TYPE: PRT <213> ORGANISM: Mycobacterium tuberculosis <400> SEQUENCE: 132 Met Gln Thr Thr Pro Gly Lys Arg Gln Arg Arg Gln Arg Gly Ser Ile 10 Asn Pro Glu Asp Ile Ile Ser Gly Ala Phe Glu Leu Ala Gln Gln Val 25 Ser Ile Asp Asn Leu Ser Met Pro Leu Leu Gly Lys His Leu Gly Val Gly Val Thr Ser Ile Tyr Trp Tyr Phe Arg Lys Lys Asp Asp Leu Leu Asn Ala Met Thr Asp Arg Ala Leu Ser Lys Tyr Val Phe Ala Thr Pro Tyr Ile Glu Ala Gly Asp Trp Arg Glu Thr Leu Arg Asn His Ala Arg Ser Met Arg Lys Thr Phe Ala Asp Asn Pro Val Leu Cys Asp Leu Ile Leu Ile Arg Ala Ala Leu Ser Pro Lys Thr Ala Arg Leu Gly Ala Gln Glu Met Glu Lys Ala Ile Ala Asn Leu Val Thr Ala Gly Leu Ser Leu

Asp Thr Ser Ala Ala Arg Tyr Val Trp Glu Val Pro Tyr Tyr Pro Gln

Tyr Tyr Ile Pro Leu Ala Asp Val Arg Met Glu Phe Leu Arg Asp Glu

130					135					140				
Glu Asp 145	Ala	Phe	Asp	Ile 150	Tyr	Ser	Ala	Val	Ser 155	Val	His	Val	Arg	Gly 160
Ser Val	Val	Leu	Asp 165	Arg	Leu	Ser	Arg	Lys 170	Ser	Gln	Ser	Ala	Gly 175	Ser
Gly Pro	Ser	Ala 180	Ile	Glu	His	Pro	Val 185	Ala	Ile	Asp	Pro	Ala 190	Thr	Thr
Pro Leu	Leu 195	Ala	His	Ala	Thr	Gly 200	Arg	Gly	His	Arg	Ile 205	Gly	Ala	Pro
Asp Glu 210	Thr	Asn	Phe	Glu	Tyr 215	Gly	Leu	Glu	CAa	Ile 220	Leu	Asp	His	Ala
Gly Arg 225	Leu	Ile	Glu	Gln 230	Ser	Ser	Lys	Ala	Ala 235	Gly	Glu	Val	Ala	Val 240
Arg Arg	Pro	Thr	Ala 245	Thr	Ala	Asp	Ala	Pro 250	Thr	Pro	Gly	Ala	Arg 255	Ala
Lys Ala	Val	Ala 260	Arg											
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Met Thr	Ile	Pro	Asp 5	Ala	Gln	Thr	Leu	Met 10	Arg	Pro	Ile	Leu	Ala 15	Tyr
Leu Ala	Asp	Gly 20	Gln	Ala	Lys	Ser	Ala 25	Lys	Asp	Val	Ile	Ala 30	Ala	Met
Ser Asp	Glu 35	Phe	Gly	Leu	Ser	Asp 40	Asp	Glu	Arg	Ala	Gln 45	Met	Leu	Pro
Ser Gly 50	Arg	Gln	Arg	Thr	Met 55	Tyr	Asp	Arg	Val	His 60	Trp	Ser	Leu	Thr
His Met 65	Ser	Gln	Ala	Gly 70	Leu	Leu	Asp	Arg	Pro 75	Thr	Arg	Gly	His	Val 80
Gln Val	Thr	Asp	Thr 85	Gly	Arg	Gln	Val	Leu 90	Lys	Ala	His	Pro	Glu 95	Arg
Val Asp	Met	Ala 100	Val	Leu	Arg	Glu	Phe 105	Pro	Ser	Tyr	Ile	Ala 110	Phe	Arg
Glu Arg	Thr 115	Lys	Ala	Lys	Gln	Pro 120	Val	Asp	Ala	Thr	Ala 125	Lys	Arg	Pro
Ser Gly 130	Asp	Asp	Val	Gln	Val 135	Ser	Pro	Glu	Asp	Leu 140	Ile	Asp	Ala	Ala
Leu Ala 145	Glu	Asn	Arg	Ala 150	Ala	Val	Glu	Gly	Glu 155	Ile	Leu	Lys	Lys	Ala 160
Leu Thr	Leu	Ser	Pro 165	Thr	Gly	Phe	Glu	Asp 170	Leu	Val	Ile	Arg	Leu 175	Leu
Glu Ala	Met	Gly 180	Tyr	Gly	Arg	Ala	Gly 185	Ala	Val	Glu	Arg	Thr 190	Ser	Ala
Ser Gly	Asp 195	Ala	Gly	Ile	Asp	Gly 200	Ile	Ile	Ser	Gln	Asp 205	Pro	Leu	Gly
Leu Asp 210	Arg	Ile	Tyr	Val	Gln 215	Ala	Lys	Arg	Tyr	Ala 220	Val	Asp	Gln	Thr

	[le 225	Gly	Arg	Pro	Lys	Ile 230	His	Glu	Phe	Ala	Gly 235	Ala	Leu	Leu	Gly	Lys 240
(Gln	Gly	Asp	Arg	Gly 245	Val	Tyr	Ile	Thr	Thr 250	Ser	Ser	Phe	Ser	Arg 255	Gly
ž	Ala	Arg	Glu	Glu 260	Ala	Glu	Arg	Ile	Asn 265	Ala	Arg	Ile	Glu	Leu 270	Ile	Asp
(Gly	Ala	Arg 275	Leu	Ala	Glu	Leu	Leu 280	Val	Arg	Tyr	Arg	Val 285	Gly	Val	Gln
Ž	Ala	Val 290	Gln	Thr	Val	Glu	Leu 295	Leu	Arg	Leu	Asp	Glu 300	Asp	Phe	Phe	Asp
	31y 305	Leu														

What is claimed is:

- 1. An isolated polynucleotide encoding a Mtb polypeptide that is antigenic in a mammal, wherein the polynucleotide is selected from the group consisting of: (a) SEQ ID NOS: 46-64, 110-121; or (b) a fragment thereof, wherein the fragment encodes an antigenic peptide epitope.
- 2. The isolated polynucleotide of claim 1, wherein said mammal is a rabbit, human or mouse.
 - 3. (canceled)
- **4.** The polynucleotide of claim **1**, wherein said Mtb polypeptide reacts with polyclonal antibodies directed to Mtb.
- **5**. The polynucleotide of claim **1**, wherein said Mtb polypeptide is detected by either ELISA or Western blotting using a polyclonal antibody directed to Mtb.
- **6**. The isolated polynucleotide of claim **1**, further comprising: (a) a 5' TAP polynucleotide sequence; and (b) a 3' TAP polynucleotide sequence and said 5' TAP polynucleotide sequence and said 3' TAP polynucleotide sequence are operably coupled to said isolated polynucleotide.
- 7. The isolated polynucleotide of claim 6, wherein the Mtb polynucleotide sequence is selected from the group consisting of: SEQ ID NOS: 46-64, 110-121.
- **8**. The isolated polynucleotide of claim **6**, wherein the 5' TAP polynucleotide sequence comprises a promoter.
- 9. The isolated polynucleotide of claim 6, wherein the 5' TAP polynucleotide sequence is selected from the group consisting of: SEQ ID NOS: 2, 3, 6, and 84.
- 10. The isolated polynucleotide of claim 6, wherein the 3' TAP polynucleotide sequence comprises a terminator.
- 11. The isolated polynucleotide of claim 6, wherein the 3' TAP polynucleotide sequence is selected from the group consisting of: SEQ ID NOS: 4, 5, 7, and 85.

- 12. A primer pair for amplifying a Mtb polynucleotide selected from the group consisting of: SEQ ID NOS: 8 and 9; 10 and 11; 12 and 13; 14 and 15; 16 and 17; 18 and 19; 20 and 21; 22 and 23; 24 and 25; 26 and 27; 28 and 29; 30 and 31; 32 and 33; 34 and 35; 36 and 37; 38 and 39; 40 and 41; 42 and 43; 44 and 45; 86 and 87; 88 and 89; 90 and 91; 92 and 93; 94 and 95; 96 and 97; 98 and 99; 100 and 101; 102 and 103; 104 and 105; 106 and, 107; 108 and 109.
- 13. A method of generating an immune response in a mammalian host against Mtb comprising: administering to said mammalian host an immunogenic composition comprising at least one nucleic acid selected from the group of SEQ ID NO: 46-64, 110-121, fragments thereof or combinations thereof, wherein said nucleic acid encodes and expresses in vivo at least one immunogenic peptide or polypeptide, whereby said immune response against Mtb is generated.
 - 14. (canceled)
- 15. The method according to claim 13, wherein said immunogenic composition further comprises and adjuvant.
 - 16. (canceled)
 - 17. (canceled)
- **18**. A kit comprising: a) at least one Mtb immunogenic composition selected from a nucleic acid selected from the group consisting of SEQ ID NO: 46-64, 110-121, fragments thereof or combinations thereof.
- 19. The kit of claim 18, wherein the kit further comprises an expression system.
- 20. The kit of claim 18 comprising at least two of said immunogenic compositions.
- 21. The kit of claim 20, comprising at least two of said nucleic acids.

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