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WO-A-01/14566
US-A- 5 846 794
US-A- 5 872 247
SHELTON M C ET AL: "2-Keto-3-deoxy-6-phosphogluconate aldolases as catalysts for stereocontrolled carbon-carbon bond formation" JOURNAL OF THE AMERICAN CHEMICAL SOCIETY, AMERICAN CHEMICAL SOCIETY, WASHINGTON, DC, US, vol. 118, no. 9, 6 March 1996 (1996-03-06), pages 2117-2125, XP002263455 ISSN: 0002-7863
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DESCRIPTION

[0001] This invention relates to a process for preparing 2'-deoxynucleoside compounds or 2'-deoxynucleoside precursors using 2-dehydro-3-deoxy-D-gluconic acid (usually abbreviated as KDG) or its salts as a starting material. A variety of 2'-deoxynucleosides and their analogues are used as a starting material for synthesis or drug formulation in production of an antiviral, anticancer or antisense agent.

[0002] Specifically, the invention relates to a method in which KDG or a derivative of KDG is subjected to a decarboxylation step to remove the original carboxy group of KDG. In a preferred embodiment, the KDG used in the method according to the invention is enzymatically produced from D-gluconate or D-glucosamine.

[0003] 2'-deoxynucleosides and 2'-deoxynucleoside precursors including 2-deoxy-D-ribose are used as starting material for synthesis or drug formulation, for instance, in production of antiviral and anticancer agent. 2'-deoxynucleosides or derivatives thereof and 2'-deoxynucleoside precursors are also used as reagents for research, diagnosis and synthesis of therapeutic antisense molecules.

[0004] In one method of the prior art, deoxynucleosides are generated from biological materials such as testis (WO 99/49074) or yeast or fish sperm by enzymatic cleavage of DNA. This method, however, involves several disadvantages, in particular regarding difficulties of obtaining the starting material in sufficient quantity and quality.

[0005] The main production process of 2-deoxy-D-ribose currently consists in chemical hydrolysis of DNA. In this case, the deoxyribosyl moiety originates in ribonucleotide reductase activity. No synthesis of 2-deoxy-D-ribose from KDG has been yet described.

[0006] In most living cells, deoxyribonucleosides result from a "salvage pathway" of the nucleotide metabolism. The deoxyribose moiety of deoxyribonucleosides is obtained through the reduction of a ribosyl moiety into di- or triphosphate ribonucleotides catalyzed by ribonucleotide reductases. However, the deoxyribose moiety is not recycled, but is degraded into D-glyceraldehyde-3-phosphate and acetaldehyde following the reactions of central metabolism:

- deoxynucleoside is cleaved into deoxyribose-1-phosphate and nucleobase through phosphorolysis mediated by products of the genes encoding thymidine phosphorylase (deoA), purine-nucleoside phosphorylase (deoD), uridine phosphorylase (udp) or xanthosine phosphorylase (xapA).
- deoxyribose-1-phosphate is converted into deoxyribose-5-phosphate through a reaction catalyzed by deoxyribose phosphate mutase (deoB),
- which is further degraded to D-glyceraldehyde-3-phosphate and acetaldehyde through a reaction catalyzed by deoxyribose-5-phosphate aldolase (deoC).

[0007] It has been shown that the deo enzymes also catalyze in vitro the reverse anabolic reactions: Deoxyribose-5-phosphate is obtained in vitro in the presence of purified *Escherichia coli* or *Lactobacillus plantarum* deoxyribose aldolase starting from acetaldehyde and D-glyceraldehyde-3-phosphate (Rosen et al., J. Biol. Chem., 240, (1964), 1517-1524; Pricer, J. Biol. Chem., 235, (1960), 1292-1298). Deoxyribose can also be obtained with acetaldehyde and glyceraldehyde as enzyme substrates, but only with a very low yield (Barbas, J. Am. Chem. Soc. 112 (1990), 2013-2014).

[0008] The patent application WO 01/14566 describes the enzymatic synthesis of deoxynucleosides starting from deoxyribose-1-phosphate through the combined activities of three enzymes of the deo operon, i.e. deoxyribose aldolase, deoxyribomutase and phosphorylase (thymidine or purine nucleoside phosphorylase) in a one-pot reaction, using as starting substrates glyceraldehyde-3-phosphate, acetaldehyde and a nucleobase. D-glyceraldehyde-3-phosphate can be obtained from fructose-1,6-bisphosphate by an enzymatic process.

[0009] The patent application EP 1179598 describes the use of phosphorylase to catalyze the enzymatic production of deoxynucleosides starting from deoxyribose-1-phosphate and nucleobase. The yield of deoxynucleoside synthesis is improved by precipitation of phosphate.

[0010] However, methods using enzymes of the deo operon working in the reverse direction compared to their biological function show low yields, which indicates serious drawbacks for their use.

[0011] In view of the above-described ineffectiveness of the currently applied processes for producing deoxynucleosides and deoxynucleoside precursors, it is an object of the present invention to provide means and methods for the biosynthetic production of deoxynucleosides and deoxynucleoside precursors starting from cheap and commercially available compounds without being dependent on unreliable natural sources.

[0012] In particular, there is a need for alternative methods for the production of deoxynucleosides and deoxynucleoside precursors which allow efficient and economical synthesis of deoxyribonucleosides, by means of which the drawbacks of prior art processes are eliminated.

[0013] The present invention relates to a method for producing 2'-deoxynucleosides and precursors thereof starting from 2-dehydro-3-deoxy-D-gluconic acid (KDG) or its salts and comprising a decarboxylation step.

[0014] In particular, this method is useful for producing 2-deoxy-D-ribose (DRI) as well as synthetically versatile enamine derivatives of DRI as 2'-deoxynucleoside precursors.

[0015] The decarboxylation step takes place by reacting either KDG or its salts directly, or a derivative of KDG, usually to cleave the C1-C2 bond of the KDG.

[0016] In one embodiment of the invention, KDG or one of its salts undergoes (oxidative) decarboxylation leading to 2-deoxy-D-ribonic acid (DRN) or its salts, itself being further converted into 2-deoxy-D-ribose (DRI) or 2-deoxy-D-ribitol (DRL).

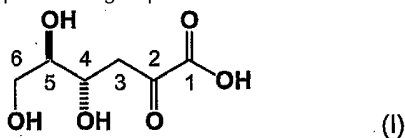
[0017] In another embodiment of the invention, decarboxylation takes place by reacting KDG or its salts with an amine, leading to an enamine derivative. This high energy enamine derivative can be further converted into DRI by hydrolysis.

[0018] In another embodiment of the invention, (oxidative) decarboxylation is carried out on 3-deoxy-D-gluconic acid (DGN) or its salts and/or 3-deoxy-D-mannonic acid (DMN) or its salts as derivatives of KDG, leading to DRI. Production of a mixture of DGN and DMN takes place by reduction of KDG. The decarboxylation is preferably carried out via reaction with hydrogen peroxide.

[0019] In another embodiment of the invention, (oxidative) decarboxylation is carried out on 3-deoxy-D-glucosaminic acid (DGM) or its salts and/or 3-deoxy-D-mannosaminic acid (DMM) or its salts, leading to DRI. Production of a mixture of DGM and DMM takes place from KDG by reductive amination.

[0020] Another aspect of the invention is a convenient and cost-effective method for preparing KDG or its salts to be used in the above methods. This method starts either from D-gluconate or from D-glucosamine through the use of recombinant enzymes. The invention provides a novel nucleotide sequence encoding a polypeptide having D-gluconate dehydratase activity and a nucleotide sequence encoding a polypeptide having D-glucosamine deaminase activity.

[0021] The starting material used for the method of the present invention is KDG, represented by formula (I) below or one of its salts, or a protected derivative thereof wherein one or more of the hydroxyl groups at positions 4, 5 and/or 6 are protected by a protection group known in the art.



[0022] The term "2'-deoxynucleoside" as used herein relates to 2'-deoxyribonucleosides which are N-glycosides, and wherein the basic N-atom of the nucleobase or nucleobase analog is bound to the anomeric carbon atom of 2-deoxy-D-ribose, or one of its derivatives. Examples of a suitable nucleobase are adenine, cytosine, guanine, thymine, uracil, 2,6-diaminopurine, and hypoxanthine. Examples of nucleobase analogs are 5-azacytosine, 2-chloro-adenine, 5-iodo-cytosine, 8-azaguanine, 5-iodo-uracil, 5-bromo-uracil, 5-fluoro-uracil, 5-ethyl-uracil and 5-trifluoromethyl-uracil.

[0023] The term "2'-deoxynucleoside precursors" as used herein, relates to compounds which can be easily converted into 2'-deoxynucleosides by applying methods known in the prior art. Preferred 2'-deoxynucleoside precursors are 2-deoxy-D-ribose (DRI) or carbohydrate compounds which can be converted into the 2-deoxy-D-ribosyl moiety of 2'-deoxynucleosides, for instance, those established in the prior art 1-phospho-2-deoxy-D-ribose, 5-phospho-2-deoxy-D-ribose and those established by the

present invention 2-deoxy-D-ribitol, 2-deoxy-D-ribonic acid, 2-deoxy-D-ribono-1,4-lactone, 1-N-morpholino-3,4,5-trihydroxypentene-1, and their derivatives.

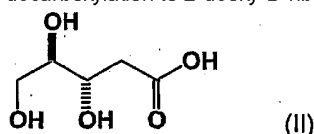
[0024] The method of the invention encompasses methods wherein the decarboxylation step is directly carried out on KDG or its salts or on compounds derived from KDG. Preferred KDG derivatives are 3-deoxy-D-gluconic acid, 3-deoxy-D-mannonic acid, 3-deoxy-D-glucosaminic acid and 3-deoxy-D-mannosaminic acid and their respective salts.

[0025] Furthermore, KDG and its salts or protected forms of these wherein one or more of the hydroxyl groups at the positions 4,5 and /or 6 are replaced by protecting groups known for that purpose in the art are also suitable starting materials for the decarboxylation reaction of the present invention. Unless noted otherwise, any reference to KDG in the following specification embraces protected forms of KDG, just as reference to KDG derivatives is intended to embrace protected forms of these derivatives. Similarly, any reference to the products obtained in the methods of the invention is intended to encompass protected forms of these products. Preferred protection groups for the purpose of the invention are those which replace the respective hydroxyl groups by acetate ester, benzoate ester, allyl ether, benzyl ether, trityl ether, ter-butyldimethylsilyl (TBDMS) ether, isopropylidene or a benzylidene acetal.

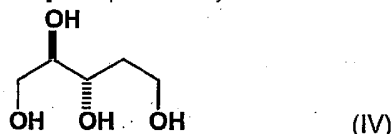
[0026] It should be understood that, depending on suitable reaction conditions for the embodiments of the invention, the carboxylic groups contained in the organic acids used as reactants or obtained as products can be in a protonated form or in their salt form, or may be present in equilibrium. Exemplary salts of these acids are those which have metal or ammonium ions as counterions, particularly alkali metal ions such as sodium and/or potassium.

[0027] Most of the carbohydrate compounds and their derivatives described in the present invention exist under several cyclic form but for simplicity reasons have been represented by open chain formulas. It is understood that the present invention encompasses all these isomeric or tautomeric forms.

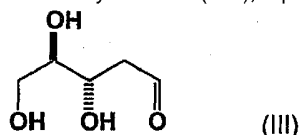
[0028] In a first embodiment of the invention, KDG or its salts is reacted with hydrogen peroxide and undergoes (oxidative) decarboxylation to 2-deoxy-D-ribonic acid (DRN), a compound of formula (II) or its salts.



[0029] The product may be further converted into or 2-deoxy-D-ribitol (DRL), represented by formula (IV)



or 2-deoxy-D-ribose (DRI), represented by formula (III)



DRN, DRL and particularly DRI are among preferred 2'-deoxynucleoside precursors for the purpose of the present invention. Conversion of DRN to DRI may proceed directly or via DRL as an intermediate.

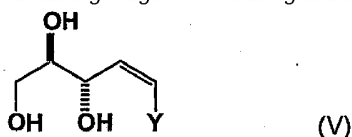
[0030] Preferably, the preparation of DRN is carried out by oxidative decarboxylation of sodium or potassium 2-dehydro-3-deoxy-D-gluconate in aqueous solution with hydrogen peroxide at room temperature as described in example 5. A general method for the preparation of aldonic acids by oxidative decarboxylation of 2-ketoaldonic acids is described in patent EP 1 038 860 A1.

[0031] Preferably, the preparation of DRL is carried out by hydrogenation of 2-deoxy-D-ribonolactone in aqueous solution with Rhodium catalyst on carbon at a temperature of 130°C under a pressure of 80 bars as described in example 6. 2-Deoxy-D-ribonolactone can be easily prepared by converting a 2-deoxy-D-ribonate (DRN salt) into 2-deoxy-D-ribonic acid, which is in equilibrium with its lactonic form in aqueous solutions (Han, Tetrahedron. 1993. 49, 349-362; Han, Tetrahedron Asymmetry. 1994. 5, 2535-62).

[0032] Preferably the preparation of 2-deoxy-D-ribose (DRI) is carried out by oxidization of 2-deoxy-D-ribitol (DRL), e.g. with

chromium oxide in pyridine.

[0033] In another embodiment of the invention, decarboxylation takes place by reacting (KDG) or its salts with an amino group-containing reagent Y-H leading to a compound of formula (V).



or its respective trans isomer or a protected form thereof, as a 2'-deoxynucleoside precursor. Y-H represents an amine with the hydrogen atom H bound to the nitrogen of the amino group.

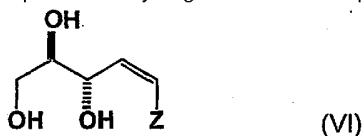
[0034] In a preferred embodiment of the invention, the amino group-containing reagent represented by Y-H is a linear or cyclic secondary amine; a primary amine that possess a β -carbonyl group, preferably 3-amino-2-indolinone which was found to be effective for the decarboxylation of α -keto acids (Hanson, J. Chem. Education, 1987, 591-595). In each of these cases, -Y in formula (V) represents the respective nitrogen containing residue derived from these amino-group containing reagent.

[0035] Preferably, the compound of formula (V) represents an enamine produced via reaction of a linear or cyclic secondary amine as Y-H.

[0036] Preferred cyclic secondary amines are morpholine, pyrrolidine, piperidine, or N-methyl piperazine; preferred non-cyclic amines are those of the formula R_1-NH-R_2 , wherein R_1 and R_2 independently represent a linear or branched alkyl group of 1-8, preferably 1 to 4 carbon atoms. Particularly preferred as a non-cyclic amine is diethylamine.

[0037] Particularly preferred as a cyclic amine is morpholine.

[0038] The compound of formula (V) or its trans isomer or a protected form thereof can be further reacted with Z-H, wherein H represents a hydrogen atom and Z represents a leaving group, to produce a compound of formula (VI)

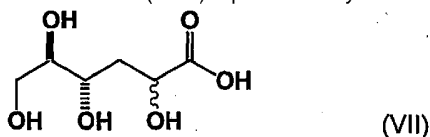


or its respective trans isomer or a protected form thereof, as a 2'-deoxynucleoside precursor. Z-H is preferably water, in which case the compound of formula (VI) is DRI or a protected form thereof (keto-enol-tautomerism).

[0039] Preferably, the preparation of the compound of formula (V) is carried out by reacting KDG in benzene with the amine, e.g. morpholine under reflux using the method described in example 7, leading to 1-N-morpholino-3,4,5-trihydroxy-pentene-1. Acid catalysed hydrolysis with water yields 2-deoxy-D-ribose (DRI)

[0040] A general route to aldehydes via enamines from α -oxocarboxylic acids carrying β -hydrogens is described by Stamos (Tetrahedron Lett. 23 (1982), 459-462). Other methods for the preparation and hydrolysis of enamines have been described elsewhere (Stork, J. Am. Chem. Soc. 85 (1963), 207-222; Stamhuis, J. Org. Chem. 30 (1965), 2156-2160).

[0041] In another embodiment of the invention, KDG or its salt is converted to 3-deoxy-D-gluconic acid (DGN) and/or 3-deoxy-D-mannonic acid (DMN) represented by formula (VII) or the salts of these compounds



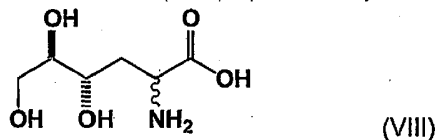
[0042] The products resulting from this reaction undergo (oxidative) decarboxylation, preferably using hydrogen peroxide, to yield DRI. Production of a mixture of DGN and DMN or their salts takes place from KDG or its salts by reduction.

[0043] Preferably the preparation of 2-deoxy-D-ribose (DRI) is carried out by non-stereoselective reduction of 2-dehydro-3-deoxy-D-gluconic acid in water with sodium borohydride at room temperature using the method described for 2-keto-3-deoxyheptonic acid by Weissbach (J. Biol. Chem. 234 (1959), 705-709), followed by oxidative decarboxylation of 3-deoxy-D-gluconate and 3-deoxy-D-mannonate with hydrogen peroxide as described e.g. in US patent 3,312,683; Richards J. Chem. Soc.

(1954), 3638-3640; Sowden J. Am. Chem. Soc. 76 (1954), 3541-3542.

[0044] In another preferred embodiment, the preparation of a mixture of DGN and DMN is carried out by hydrogenation of 2-dehydro-3-deoxy-D-gluconate in aqueous solution with 6% mol Nickel Raney catalyst or Platinum oxide at room temperature under a pressure of 6 bars.

[0045] In another embodiment of the invention, KDG or its salt is converted to 3-deoxy-D-glucosamine (DGM) or 3-deoxy-D-mannosamine (DMM) represented by formula (VIII) or the salts of these compounds



[0046] The products resulting from this reaction undergo (oxidative) decarboxylation, preferably using ninhydrin, to yield DRI. Production of a mixture of DGM and DMM or their salts takes place from KDG or its salts by reductive amination.

[0047] Preferably the preparation of 2-deoxy-D-ribose is carried out by non-stereoselective reductive amination of sodium or potassium 2-dehydro-3-deoxy-D-gluconate in aqueous solution with ammonia and sodium cyanoborohydride at room temperature, followed by oxidative decarboxylation of 3-deoxy-D-2-glucosamine and 3-deoxy-D-2-mannosamine with ninhydrin using the method described for the synthesis of 2-deoxy-D-allose by Shelton (J. Am. Chem. Soc. 118 (1996), 2117-2125 ; and Borch, J. Am. Chem. Soc. 93 (1971), 2897; Durrwachter, J. Am. Chem. Soc. 108 (1986), 7812 referenced therein).

[0048] Furthermore, the present invention provides a method for producing the compound of formula (III) (2-deoxy-D-ribose) by converting the compound of formula (I) or one of its salts (KDG) in one single step. Preferably this conversion is achieved through enzymatic catalysis. This conversion is preferably catalysed by a keto acid decarboxylase. Preferred keto acid decarboxylases are thiamin pyrophosphate (TPP) dependent keto acid decarboxylases. Examples of TPP dependent keto acid decarboxylases are pyruvate decarboxylase (EC 4.1.1.1), a benzoylformate decarboxylase (EC 4.1.1.7), an indolepyruvate decarboxylase (EC 4.1.1.74), a phosphonopyruvate decarboxylase, a sulfopyruvate decarboxylase (EC 4.1.1.79), an oxalyl-coenzymeA decarboxylase (EC 4.1.1.8), an oxoglutarate decarboxylase (EC 4.1.1.71) or a phenylpyruvate decarboxylase (EC 4.1.1.43). It could be shown that keto acid decarboxylases, e.g., pyruvate decarboxylase enzymes from different organisms, can convert KDG into 2-deoxy-D-ribose (see Examples 8 to 12). In principle any keto acid decarboxylase can be used in connection with the present invention.

[0049] In a preferred embodiment of the method according to the invention KDG is converted into 2-deoxy-D-ribose by use of an enzyme having pyruvate decarboxylase activity.

[0050] A pyruvate decarboxylase catalyses the following reaction:



[0051] Several pyruvate decarboxylases (PDC) have been characterized as well as the corresponding pdc genes, for instance PDC from *Zymomonas mobilis* (Genbank accession number AAD19711; Neale et al., J. Bacteriol. 1987, 169:1024-1028), PDC from *Saccharomyces cerevisiae* (Genbank accession number NP013145; Candy et al., J. Gen. Microbiol. 1991, 137:2811-2815), PDC from *Acetobacter pasteurianus* (Genbank accession number AAM21208; Raj et al., Arch. Microbiol. 2001, 176:443-451), PDC from *Zymobacter palmae* (Genbank accession number AAM49566; Raj et al., Appl. Environ. Microbiol. 2002, 68:2869-2876), PDC from *Sarcina ventriculi* (Genbank accession number AAL18557; Lowe et al., J. Gen. Microbiol. 1992, 138:803-807). Many other pyruvate decarboxylases seems to occur in plants, fungi and bacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with well-established pdc genes. Examples of such putative pyruvate decarboxylases are:

PDC from plants:

Arabidopsis thaliana (Genbank accession number T48155)

Echinochloa crus-galli (Genbank accession number AAM18119)

Oryza sativa (Genbank accession number NP922014)

Rhizopus oryzae (Genbank accession number AAM73540)

Lotus corniculatus (Genbank accession number AAO72533)

Zea mays (Genbank accession number BAA03354)

Pisum sativum (Genbank accession number CAA91445)

Garden pea (Genbank accession number S65470)

Nicotiana tabaccum (Genbank accession number CAA57447)

Solanum tuberosum (Genbank accession number BAC23043)

Fragaria ananassa (Genbank accession number AAL37492)

Cucumis melo (Genbank accession number AAL33553)

Vitis vinifera (Genbank accession number AAG22488)

PDC from Fungi:

Saccharum officinarum (Genbank accession number CAB61763)

Aspergillus oryzae (Genbank accession number AAD16178)

Aspergillus parasiticus (Genbank accession number P51844)

Saccharomyces cerevisiae (Genbank accession number NP013145)

Flammulina velutipes (Genbank accession number AAR00231)

Saccharomyces kluyveri (Genbank accession number AAP75899)

Schizosaccharomyces pombe (Genbank accession number CAB75873)

Candida glabrata (Genbank accession number AAN77243)

Neurospora crassa (Genbank accession number JN0782)

Pichia stipitis (Genbank accession number AAC03164)

Kuyveromyces lactis (Genbank accession number CAA61155)

Emericella nidulans (Genbank accession number AAB63012)

PDC from Prokaryotes:

Mycobacterium bovis (Genbank accession number CAD93738)

Mycobacterium leprae (Genbank accession number CAC31122)

Mycobacterium tuberculosis (Genbank accession number NP215368)

Mycoplasma penetrans (Genbank accession number NP758077)

Clostridium acetobutylicum (Genbank accession number NP149189)

Acetobacter pasteurianus (Genbank accession number AAM21208)

Zymobacter palmae (Genbank accession number AAM49566)

Zymomonas mobilis (Genbank accession number AAD19711)

Sarcina ventriculi (Genbank accession number AAL18557)

Nostoc punctiforme (Genbank accession number ZP00110850)

[0052] Such enzymes can be easily produced by recombinant microorganisms overexpressing the corresponding gene. Examples of genes coding for TPP dependent keto acid decarboxylases are *pdc* from *Zymomonas mobilis* (Genbank accession number AF124349), *pdc* from *Saccharomyces cerevisiae* (Genbank accession number NC001144), *pdc* from *Acetobacter pasteurianus* (Genbank accession number AF368435), *pdc* from *Zymobacter palmae* (Genbank accession number AF474145), *pdc* from *Sarcina ventriculi* (Genbank accession number AF354297). Other *pdc* genes can be found at Genbank corresponding to the above list of putative pyruvate decarboxylases.

[0053] In a preferred embodiment the pyruvate decarboxylase is of eukaryotic origin, more preferably it is from yeast and most preferably it is from *Saccharomyces cerevisiae*. In a particularly preferred embodiment the pyruvate decarboxylase is the pyruvate decarboxylase from *S. cerevisiae* which has the amino acid sequence as shown in SEQ ID NO: 21 (see also GenBank accession number NP013145).

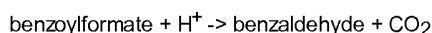
[0054] In another preferred embodiment the pyruvate decarboxylase is of prokaryotic origin, more preferably it is from an organism of the genus *Zymomonas* and most preferably from *Zymomonas mobilis*. In a particularly preferred embodiment the pyruvate decarboxylase is the pyruvate decarboxylase from *Z. mobilis* which has the amino acid sequence as shown in SEQ ID NO: 19 (see also GenBank accession number AAD19711).

[0055] In another preferred embodiment the prokaryotic pyruvate decarboxylase is from an organism of the genus *Acetobacter*, more preferably from the species *Acetobacter pasteurianus*. Particularly preferred the pyruvate decarboxylase is that of *A. pasteurianus* which shows the amino acid sequence as given in SEQ ID NO: 25 (see also GenBank accession number AAM21208).

[0056] In a further preferred embodiment the pyruvate decarboxylase is from an organism of the genus *Zymobacter*, more preferably of the species *Zymobacter palmae*. Particularly preferred is a pyruvate decarboxylase from *Z. palmae* which shows the amino acid sequence given in SEQ ID NO: 29 (see also GenBank accession number AAM49566).

[0057] In another preferred embodiment of the method according to the invention KDG is converted into 2-deoxy-D-ribose by use of an enzyme having benzoylformate decarboxylase activity.

[0058] A benzoylformate decarboxylase catalyses the following reaction:



[0059] A benzoylformate decarboxylase (BDC) from *Pseudomonas putida* (Genbank accessing number AAC15502; Tsou et al., Biochemistry. 1990, 29:9856-9862) has been characterized as well as the corresponding gene *mdlC* (Genbank accessing number AY143338). This enzyme has been shown to decarboxylate both D and L isomers of 2-keto-4,5-dihydroxyvalerate into the respective isomers of 3,4-dihydroxybutanal (Niu et al., J. Am. Chem. Soc. 125 (2003), 12998-12999). Many other benzoylformate decarboxylases seems to occur in bacteria and archaeobacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with genes coding for well-established BDC. Examples of such putative benzoylformate decarboxylases are:

BDC from bacteria:

Pseudomonas aeruginosa (Genbank accession number NP_253588)

Rhodopseudomonas palustris (Genbank accession number NP_946955)

Streptomyces coelicolor (Genbank accession number NP_631486)

Chromobacterium violaceum (Genbank accession number NP_902771)

Bradyrhizobium japonicum (Genbank accession number NP_774243)

BDC from archaeobacteria:

Sulfolobus solfataricus (Genbank accession number NP_343070)

Thermoplasma acidophilum (Genbank accession number NP_393976)

Thermoplasma volcanium (Genbank accession number NP_111716)

[0060] Such enzymes can be easily produced by recombinant microorganisms overexpressing the corresponding *bdc* gene. Such genes can be found at Genbank corresponding to the above list of putative benzoylformate decarboxylases.

[0061] Another example for a thiamine dependent decarboxylase which can be used in the method according to the invention is phosphonopyruvate decarboxylase. Several phosphonopyruvate decarboxylases (PPD) have been characterized as well as the corresponding genes, for instance PPD from *Bacteroides fragilis* (Genbank accession number AAG26466; Zhang et al., J. Biol. Chem. 2003, 278:41302-41308), PPD from *Streptomyces wedmorensis* (Genbank accession number BAA32496; Nakashita et al., J. Antibiot. 1997, 50:212-219). Many other phosphonopyruvate decarboxylases seem to occur in bacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with genes coding for well-established PPD. Examples of such putative phosphonopyruvate decarboxylases are: PPD from *Bacteroides thetaiotaomicron* (Genbank accession number NP_810632), PPD from *Amycolatopsis orientalis* (Genbank accession number CAB45023), PPD from *Clostridium tetani* E88 (Genbank accession number NP_782297), PPD from *Streptomyces viridochromogenes* (Genbank accession number CAA74722), PPD from *Streptomyces hygroscopicus* (Genbank accession number BAA07055), PPD from *Streptomyces coelicolor* A3 (Genbank accession number NP_733715), *Streptomyces rishiriensis* (Genbank accession number AAG29796), *Bordetella pertussis* (Genbank accession number CAE 41214). Such enzymes can be easily produced by recombinant microorganisms overexpressing the corresponding gene.

[0062] A further example of a thiamine dependent decarboxylases which can be used in the method according to the present invention is sulfopyruvate decarboxylase. A sulfopyruvate decarboxylases (SPD) from *Methanococcus jannaschii* (Graupner et al., J. Bacteriol. 2000, 182:4862-4867) consisting of two subunits ComD (Genbank accession number P58415) and ComE (Genbank accession number P58416) has been characterized as well as the corresponding genes. Many other sulfopyruvate decarboxylases seems to occur in archaeobacteria and in bacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with genes coding for well-established SPD.

[0063] Another further example of thiamine dependent decarboxylase which can be used in the method according to the present invention is indolepyruvate decarboxylase. Several indolepyruvate decarboxylases (IPD) have been characterized as well as the corresponding genes, for instance IPD from *Enterobacter cloacae* (Genbank accession number BAA14242; Scutz et al., 2003, Eur. J. Biochem. 270:2322-2331), IPD from *Azospirillum brasilense* (Genbank accession number AAC36886; Costacurta et al., Mol. Gen. Genet. 1994, 243:463-472), IPD from *Erwinia herbicola* (Genbank accession number AAB06571; Brandl et al., Appl. Environ. Microbiol. 1996, 62:4121-4128). Many other indolepyruvate decarboxylases seem to occur in bacteria as evidenced by the occurrence in these organisms of genes sharing sequence homologies with genes coding for well-established IPD.

[0064] Still another further example of a thiamine dependent decarboxylases which can be used in the method according to the present invention is phenylpyruvate decarboxylase. A phenylpyruvate decarboxylase from yeast (Genbank accession number NP010668; Vuralhan et al., Appl. Environ. Microbiol. 2003, 69:4534-41) has been characterized as well as the corresponding gene ARO10 (Genbank accession number NC001136).

[0065] In a preferred embodiment of the method according to the invention in which the decarboxylation step is effected by an enzymatic reaction, the pH value is regulated by addition of an acid to be between pH 5 and pH 9, preferably between pH 6 and pH 8. In principle, any suitable acid can be used for this purpose. Preferred acids are HCl, H₂SO₄, D-gluconic acid or 2-dehydro-3-deoxy-D-gluconic acid.

[0066] Another aspect of the invention is a convenient and cost-effective method for preparing KDG either from D-gluconate (GCN) or from D-glucosaminatate through the use of recombinant enzymes.

[0067] In a preferred embodiment of the method of the invention, the compound of formula (I) is produced in a preliminary step from a D-gluconate salt by the use of a D-gluconate dehydratase activity. Preferred salts are potassium or sodium D-gluconate. Preferably the D-gluconate dehydratase is encoded by a polynucleotide comprising the nucleotide sequence selected from the group consisting of:

1. (a) nucleotide sequences encoding a polypeptide comprising the amino acid sequence of SEQ ID N°2;
2. (b) nucleotide sequences comprising the coding sequence of SEQ ID N°1;

3. (c) nucleotide sequences encoding a fragment encoded by a nucleotide sequence of (a) or (b);
4. (d) nucleotide sequences hybridising with a nucleotide sequence of any one of (a) to (c); and
5. (e) nucleotide sequences which deviate from the nucleoside sequence of (d) as a result of degeneracy of the genetic code.

[0068] The enzymatic synthesis of KDG or its salts using D-gluconate dehydratase proceeds according to the following reaction: D-gluconate is converted into KDG by the elimination of one water molecule. The activity of a D-gluconate dehydratase has been characterized in different bacterial species e.g. in *Alcaligenes* (Kerstens, *Methods in Enzymology* 42 (1975), 301-304); *Clostridium pasteurianum*, (Gottschalk, *Methods in Enzymology* 90 (1982), 283-287); *Thermoplasma acidophilum* (Budgen, *FEBS Letters* 196 (1986), 207-210) and *Sulfolobus solfataricus* (Nicolaus, *Biotechnology Letters* 8(7) (1986), 497-500). The preferred D-gluconate dehydratase was identified by screening several collection strains for D-gluconate dehydratase activity. The gene encoding a D-gluconate dehydratase, which was designated *gcnD* was selected from a genomic library of *Agrobacterium tumefaciens* strain C58, and further inserted in a multi copy vector optimised for expression. It was shown that a crude extract from *E. coli* cells over-expressing the *gcnD* gene catalysed the total conversion of D-gluconate into KDG (see Example 2).

[0069] In a further preferred embodiment of the method of the invention, the compound of formula (I) is produced in a preliminary step from D-glucosamine by the use of a D-glucosamine deaminase activity. Preferably the D-glucosamine deaminase is encoded by a polynucleotide comprising the nucleotide sequence selected from the group consisting of:

- (f) nucleotide sequences encoding a polypeptide comprising the amino acid sequence of SEQ ID N°4;
- (g) nucleotide sequences comprising the coding sequence of SEQ ID N°3;
- (h) nucleotide sequences encoding a fragment encoded by a nucleotide sequence of (a) or (b);
- (i) nucleotide sequences hybridising with a nucleotide sequence of any one of (a) to (c); and
- (j) nucleotide sequences which deviate from the nucleoside sequence of (d) as a result of degeneracy of the genetic code.

[0070] The enzymatic synthesis of KDG or its salts using D-glucosamine deaminase proceeds according to the following reaction: D-glucosamine is converted into KDG by the elimination of one molecule water and one molecule of ammonia. The activity of a D-glucosamine deaminase has been characterized in different bacterial species e.g. in *Pseudomonas fluorescens* (Iwamoto, *Agric. Biol. Chem.* 53 (1989), 2563-2569) *Agrobacterium radiobacter* (Iwamoto, *FEBS Letters* 104 (1979), 131-134; Iwamoto, *J. Biochem.* 91 (1982), 283-289), and its requirement for Mn^{2+} ion was shown (Iwamoto, *Biosci. Biotech. Biochem.* 59 (1995), 408-411).

[0071] The preferred D-glucosamine deaminase was identified by screening several collection strains for D-glucosamine deaminase activity. The gene encoding a D-glucosamine deaminase, which was designated *gmaA* was isolated from *Agrobacterium tumefaciens* strain C58 by cloning a gene annotated as a putative D-serine deaminase. The *gmaA* gene was further inserted in a multi copy vector optimised for expression. It was shown that a crude extract from *E. coli* cells overexpressing the *gmaA* gene catalysed the conversion of D-glucosamine into KDG (see Example 4).

[0072] In a preferred embodiment the present invention relates to a method for producing a compound of formula III, in particular 2-deoxy-D-ribose, starting from D-gluconate or D-glucosamine by enzymatic reactions which, in a first step, convert D-gluconate or D-glucosamine into KDG as described above and, in a second step, convert KDG into 2-deoxy-D-ribose as described above.

[0073] Thus, the enzymatic conversion of D-gluconate into KDG can be achieved by the use of a D-gluconate dehydratase. The enzymatic conversion of D-glucosamine into KDG can be achieved by the use of a D-glucosamine deaminase. With respect to the preferred embodiments the same applies as has already been set forth above. The enzymatic conversion of the resulting KDG into 2-deoxy-D-ribose can be achieved by the use of a keto acid decarboxylase. With respect to the preferred embodiments the same applies as has been set forth above.

[0074] The enzymatic two step method of converting D-gluconate or D-glucosamine into 2-deoxy-D-ribose via KDG can be carried out in vitro by using cell extracts of cells expressing the corresponding enzymes or by using purified or partially purified enzymes. The enzymes can be enzymes which are naturally expressed in an organism or they may be recombinantly produced. Methods of preparing and isolating corresponding (recombinant) enzymes are well-known to the person skilled in the art.

[0075] In a preferred embodiment the enzymatic two step method of converting D-gluconate or D-glucosamine into 2-deoxy-D-ribose via KDG is carried out in vivo, i.e. by using a suitable organism, which expresses the required enzyme activities. This organism may be any type of organism, preferably it is a cell, e.g. a plant, an animal, a fungal cell or a bacterial cell. Most preferably fungal or bacterial cells are used. Preferred fungi are yeasts, such as *Saccharomyces cerevisiae*; preferred bacterial cells are, e.g. *E. coli*, *Zymomonas mobilis*, *Zymobacter palmae*, *Acetobacter pasteurianus*, *Acinetobacter calcoaceticus*, *Agrobacterium tumefaciens* and *Bacillus subtilis*. The organism may be an organism which endogenously already expresses one of the enzymatic activities, i.e. a D-gluconate dehydratase or a D-glucosamine deaminase for producing KDG, or a keto acid decarboxylase for converting KDG into 2-deoxy-D-ribose, and in which the respective other enzymatic activity is expressed due to the introduction of a corresponding exogenous nucleic acid molecule encoding the corresponding enzyme. Alternatively, the organism may also be an organism which naturally does not express the enzyme activities required for converting D-gluconate or D-glucosamine into KDG and further into 2-deoxy-D-ribose and in which corresponding foreign nucleic acid molecules have been introduced encoding D-gluconate dehydratase or D-glucosamine deaminase and a keto acid decarboxylase, respectively.

[0076] In a particularly, preferred embodiment the organism is an organism which does not express a KDG kinase (kdgK) activity. Such an enzyme activity would lead to a phosphorylation of KDG to KDPG, which in turn is cleaved by an aldolase into pyruvate and glyceraldehyde-phosphate, thereby diverting KDG into a different unwanted metabolic pathway. It is possible to use for the method according to the invention organisms which naturally do not express a kdgK gene. If the used organism naturally expresses a kdgK, means and methods are well-known to the skilled person to produce mutants or variants of such an organism in which the corresponding kdgK gene is inactivated.

[0077] If the described method according to the invention is carried out in vivo by using an organism which expresses a D-gluconate dehydratase for converting D-gluconate into KDG and a keto acid decarboxylase for converting KDG into 2-deoxy-D-ribose, this has the advantage that one can provide D-gluconate as a substrate in the culture medium used to culture the organism. D-gluconate is taken up by the organism and is converted into 2-deoxy-D-ribose.

[0078] In another particularly, preferred embodiment the organism is an organism which does not express a KDG aldolase (encoded by the *eda* gene in *E. coli*) activity. Such an enzyme activity would lead to cleavage of KDG into pyruvate and glyceraldehydes, thereby diverting KDG into a different unwanted metabolic pathway. It is possible to use for the method according to the invention organisms which naturally do not express an *eda* gene. If the used organism expresses an *eda* gene, means and methods are well-known to the skilled person to produce mutants or variants of such an organism in which the corresponding *eda* gene is inactivated.

[0079] In still another particularly, preferred embodiment the organism is an organism which does not express a 2-deoxy-D-ribose aldolase (encoded by the *deoC* gene in *E. coli*) activity. Such an enzyme activity would lead to cleavage of 2-deoxy-D-ribose into acetaldehyde and glyceraldehyde, thereby diverting 2-deoxy-D-ribose into a different unwanted metabolic pathway. It is possible to use for the method according to the invention organisms which naturally do not express a *deoC* gene. If the used organism expresses a *deoC* gene, means and methods are well-known to the skilled person to produce mutants or variants of such an organism in which the corresponding *deoC* gene is inactivated. For instance a *deoC* mutant of *E. coli* has been reported (Valentin-Hansen, EMBO J. 1 (1982), 317-322) as well as a method for deleting the *deo* operon in *E. coli* (Kaminski, J. Biol. Chem. 277 (2002), 14400-14407; Valentin-Hansen, Molec. Gen. Genet. 159 (1978), 191-202).

[0080] The present invention also relates to organisms which are capable of enzymatically converting D-gluconate into KDG due to the expression of a D-gluconate dehydratase and/or of enzymatically converting D-glucosamine into KDG due to the expression of a D-glucosamine deaminase and which are furthermore capable of enzymatically converting KDG into 2-deoxy-D-ribose by a decarboxylation reaction catalysed by a pyruvate decarboxylase or a benzoylformate decarboxylase. The organisms are fungal cells or bacterial cells. Preferred fungi are yeasts, e.g. *Saccharomyces cerevisiae*. Preferred bacteria are *Escherichia coli*, *Zymomonas mobilis*, *Zymobacter palmae*, *Acetobacter pasteurianus*, *Acinetobacter calcoaceticus*, *Agrobacterium tumefaciens* and *Bacillus subtilis*. In one aspect, the organism is an organism which already endogenously expresses a D-gluconate dehydratase or a D-glucosamine deaminase and into which a foreign nucleic acid molecule has been introduced which encodes a pyruvate decarboxylase or a benzoylformate decarboxylase which can catalyse the decarboxylation of KDG to 2-deoxy-D-ribose. With respect to the preferred embodiments of the pyruvate decarboxylase or a benzoylformate decarboxylase the same applies as has been set forth previously.

[0081] In another aspect, the organism is an organism which already expresses a pyruvate decarboxylase or a benzoylformate decarboxylase which is capable of converting KDG into 2-deoxy-D-ribose by a decarboxylation reaction but which does not naturally express a D-gluconate dehydratase or a D-glucosamine deaminase, and into which a foreign nucleic acid molecule has been introduced which encodes a D-gluconate dehydratase and/or which encodes a D-glucosamine deaminase. i.e. the

organism can be genetically modified so as to express a D-gluconate dehydratase or a D-glucosamine deaminase or both enzymes.

[0082] In a further aspect, the organism is an organism, which naturally does not express a D-gluconate dehydratase, a D-glucosamine deaminase and a pyruvate decarboxylase or a benzoylformate decarboxylase which is capable of converting KDG by decarboxylation into 2-deoxy-D-ribose, and into which foreign nucleic acid molecules have been introduced encoding a D-gluconate dehydratase or a D-glucosamine deaminase, or both, and a nucleic acid molecule which encodes a pyruvate decarboxylase or a benzoylformate decarboxylase which is capable of converting KDG into 2-deoxy-D-ribose by decarboxylation.

[0083] With respect to the preferred embodiments of the D-gluconate dehydratase, the D-glucosamine deaminase and the pyruvate decarboxylase or a benzoylformate decarboxylase to be expressed in the organisms according to the invention, the same applies which has been set forth above in connection with the method according to the invention.

[0084] In a particularly preferred embodiment the organism according to the invention does not express a KDG kinase (kdgK) activity. It can either be an organism which naturally does not express kdgK or it can be an organism which naturally expresses a kdgK but in which the corresponding gene has been inactivated, e.g. by gene disruption or other suitable methods well-known to the person skilled in the art.

[0085] The present invention also relates to the use of an enzyme having keto acid decarboxylase activity or of a polynucleotide encoding such an enzyme in a method for converting KDG into 2-deoxy-D-ribose. With respect to the preferred embodiments the same applies as has already been set forth in connection with the method according to the present invention.

[0086] These and other embodiments are disclosed and encompassed by the description and examples of the present invention. The disclosure content of any references cited above or below is herewith incorporated into the present application. Further literature concerning any one of the methods, uses and compounds to be employed in accordance with the present invention may be retrieved from public libraries, using for example electronic devices. For example the public database "Medline" may be utilized which is available on the Internet, for example under <http://www.ncbi.nlm.nih.gov/PubMed/medline.html>. Further databases and addresses, such as <http://www.ncbi.nlm.nih.gov/>, <http://www.infobiogen.fr/>, http://www.fmi.ch/biology/research_tools.html, <http://www.tigr.org/>, are known to the person skilled in the art and can also be obtained using, e.g., <http://www.google.de>. An overview of patent information in biotechnology and a survey of relevant sources of patent information useful for retrospective searching and for current awareness is given in Berks, TIBTECH 12 (1994), 352-364.

[0087] Furthermore, the term "and/or" when occurring herein includes the meaning of "and", "or" and "all or any other combination of the elements connected by said term".

EXAMPLES

Example 1: Cloning of a gene encoding a D-gluconate dehydratase from *Agrobacterium tumefaciens* strain C58 (CIP 104333)

[0088] *Agrobacterium tumefaciens* strain C58 (CIP 104333) was obtained from Institut Pasteur Collection (CIP, Paris, France). Chromosomal DNA was extracted and a D-gluconate dehydratase gene was amplified by PCR according to standard protocols using the following primers:

5'-CCCTTAATTAATGACGACATCTGATAATCTTC-3', depicted in SEQ ID N° 5;

5'-TTTGCGGCCGCTTAGTGTTATCGCGCGGC-3', depicted in SEQ ID N° 6;

5'-CCCGGTACCATGACGACATCTGATAATCTTC-3', depicted in SEQ ID N° 7;

[0089] A first DNA fragment amplified using the two primers depicted in SEQ ID N° 5 and SEQ ID N° 6, was ligated into a pUC18-derived vector previously digested by *PacI* and *NotI* to yield the plasmid pVDM80. A second DNA fragment amplified using the two primers depicted in SEQ ID N° 6 and SEQ ID N° 7, was ligated into a pET29a vector (Novagen) previously digested by *KpnI* and *NotI* to yield the plasmid pVDM82. The nucleotide sequence of the cloned gene is depicted in SEQ ID N° 1 and the sequence of

the polypeptide encoded by this gene is depicted in SEQ ID N° 2.

Example 2: Expression of a D-gluconate dehydratase activity in *Escherichia coli* and preparation of 2-dehydro-3-deoxy-D-gluconate from D-gluconate

[0090] Competent cells of *E. coli* BL21 were transformed with the pVDM82 plasmid constructed as described in example 1 yielding strain +1289. Strain + 1289 was cultivated at 30°C in Luria-Bertani (LB) medium (Difco) containing 30 mg/l kanamycin until OD(600 nm) reached a value of 0.6. Then isopropyl-β-D-thiogalactopyranoside (IPTG) was added to a 0.5 mM final concentration. After a further cultivation period of 2 hours and 30 minutes, cells were collected by centrifugation and washed once with 20 mM sodium phosphate buffer pH 7.2. A cell extract was prepared by suspending about 5 g of cells in 10 ml of Tris-HCl 50 mM pH 8.5 buffer containing 10000 units lysozyme (Ready-Lyse, Epicentre, Madison, Wisconsin) and 1 mM EDTA, and incubating the suspension at 30°C for 15 minutes. Then 10000 kUnits deoxyribonuclease I (DNase I, Sigma) as well as 5 mM MgCl₂ were added to the preparation which was incubated at 30°C for an additional period of 15 minutes. The cell extract thus obtained was kept frozen at 20°C before use.

[0091] 1.5 ml of the cell extract was mixed with 2M sodium or potassium D-gluconate in a total volume of 10 ml. This preparation was incubated at 37°C after the pH has been adjusted to 8.5. The progression of 2-dehydro-3-deoxy-D-gluconate (KDG) synthesis was followed by analysing aliquots taken after increasing periods of incubation. Several dilution parts of these aliquots were deposited on silica plates and chromatographed in the following solvent system: isopropanol / water (90/10). A yellow spot of KDG (Rf ~0.40) was detected after revelation with p-anisaldehyde.

[0092] KDG was also quantitated using a spectrophotometric assay based on the reaction with semicarbazide hydrochloride as described by Mac Gee (J. Biol. Chem. 1954. 210, 617-626). Typically, after a 30h period of incubation and using the spectrophotometric assay, KDG concentration ranged from 1.5 to 2 M.

[0093] The sodium or potassium 2-dehydro-3-deoxy-D-gluconate solution thus obtained could be used as such for further synthetic steps. 2-Dehydro-3-deoxy-D-gluconic acid could also be prepared from such a solution applying published protocols (Bender, Anal. Biochem. 1974. 61, 275-279). A crude preparation of a mixture of 2-dehydro-3-deoxy-D-gluconic acid and KCl could also be obtained by adding one equivalent of HCl to a potassium 2-dehydro-3-deoxy-D-gluconate solution which was then evaporated.

Example 3: Cloning of a gene encoding a D-glucosamine deaminase from *Agrobacterium tumefaciens* strain C58 (CIP 104333)

[0094] *Agrobacterium tumefaciens* strain C58 (CIP 104333) was obtained from Institut Pasteur Collection (CIP, Paris, France). Chromosomal DNA was extracted and a D-glucosamine deaminase gene was amplified by PCR according to standard protocols using the following primers:

5'-CCCTTAATTAATGCAGTCTTCTTCAGCTCTTC-3', depicted in SEQ ID N° 8;

5'-TTTGCGGCCCGCCTAGTGAAAGAAGGTTGTGTAGAT-3', depicted in SEQ ID N° 9;

5'-AAATCATGACTATGCAGTCTTCTTCAGCTCTTCG-3', depicted in SEQ ID N° 10;

5'-TATAGATCTCTAGTGAAAGAAGGTTGTGTAGAT-3', depicted in SEQ ID N° 11;

[0095] A first DNA fragment amplified using the two primers depicted in SEQ ID N° 8 and SEQ ID N° 9, was ligated into a pUC18-derived vector previously digested by *PacI* and *NotI* to yield the plasmid pKDGb1. A second DNA fragment amplified using the two primers depicted in SEQ ID N° 10 and SEQ ID N° 11, was ligated into a pQE60 vector (Qiagen) previously digested by *BspH1* and *BglII* to yield the plasmid pEP18. The nucleotide sequence of the cloned gene is depicted in SEQ ID N° 3 and the sequence of the polypeptide encoded by this gene is depicted in SEQ ID N° 4.

Example 4: Expression of a D-glucosamine deaminase activity in *Escherichia coli* and preparation of 2-dehydro-3-deoxy-D-gluconic acid from D-glucosamine

[0096] Competent cells of *E. coli* MG1655 were transformed with the pEP18 plasmid constructed as described in example 1 and pREP4 (Qiagen) yielding strain +1068. Strain + 1068 was cultivated at 37°C in LB medium containing 30 mg/l kanamycin and 100 mg/l ampicillin until OD(600 nm) reached a value of 0.6. Then IPTG was added to a 0.5 mM final concentration. After a further cultivation period of 2 hours and 30 minutes, cells were collected by centrifugation and washed once with 20 mM sodium phosphate buffer pH 7.2. A cell extract was prepared using the protocol described in example 2.

[0097] 2 ml of the cell extract was mixed with 100 mM sodium or potassium D-glucosaminat and 0.1 mM pyridoxal phosphate in a total volume of 5 ml. This preparation was incubated at 37°C after the pH has been adjusted to 7.5.

[0098] The progression of 2-dehydro-3-deoxy-D-gluconate (KDG) synthesis was followed using the protocols described in example 2. Typically, after a 30h period of incubation and using the spectrophotometric assay described in example 2, KDG concentration ranged from 50 to 100 mM.

Example 5: Preparation of 2-deoxy-D-ribonate from 2-dehydro-3-deoxy-D-gluconate

[0099] 0.5 ml of a 31% hydrogen peroxyde solution were added to 5 ml of a 1M potassium 2-dehydro-3-deoxy-D-gluconate (KDG) solution at 25°C. The progression of KDG decarboxylation was followed both by the observation of bubbles resulting from the release of carbon dioxide and by the disappearance of KDG using the thin layer chromatography protocol described in example 2. Typically, after a 3h period of reaction the concentration of residual KDG was less than 10 mM.

Example 6: Preparation of 2-deoxy-D-ribitol from 2-deoxy-D-ribonolactone

[0100] 0.2 g of Rhodium (5 % on carbon) catalyst was added to an aqueous solution of 1 g 2-deoxy-D-ribonolactone prepared following a method described by Deriaz (J. Chem. Soc. (1949), 1879-1883) for the synthesis of 2-deoxy-L-ribonolactone. Hydrogenation of 2-deoxy-D-ribonolactone was performed at 130°C under a pressure of 80 bars. The solution obtained after filtration of the reaction mixture was evaporated. The residue was dissolved in ethyl acetate and further purified by chromatography on a silica column. The solvent was removed in vacuo leading to a yellow oil (yield 85%). The compound thus obtained was identical with 2-deoxy-D-ribitol obtained by reduction of 2-deoxy-D-ribose as described by Rabow (J. Am. Chem. Soc. 122 (1999), 3196-3203).

Example 7: Preparation of 1-N-morpholino-3,4,5-trihydroxypentene-1 from 2-dehydro-3-deoxy-D-gluconate

[0101] 2 g of 2-dehydro-3-deoxy-D-gluconic acid were suspended in 150 ml benzene. 1.1 ml morpholine and 100 mg p-toluenesulfonic acid were added to the suspension and the reaction mixture was refluxed for 3 hours. Water formed by this reaction was removed by distillation. Benzene was decanted. Solid compounds attached to the vessel were collected, washed with acetone and dried. The main compound present in this preparation (yield 40%) was further purified by column chromatography on a silica column using a gradient of methanol in chloroform. Fractions containing 1-N-morpholino-3,4,5-trihydroxypentene-1 were pooled and solvent was removed in vacuo.

¹H-NMR (D₂O): δ = 3.15 ppm (4H, t, morpholine), 3.8 ppm (4H, t, morpholine), 3.4 to 4 ppm, (4H, m, 5a-H, 5b-H, 4-H, 3-H), 6.3 and 6.8 ppm (2H, 2d, 1-H and 2-H, J = 4 Hz).

Example 8: Cloning of a gene encoding a pyruvate decarboxylase from *Zymomonas mobilis*

[0102] *Zymomonas mobilis* strain B-806 (CIP 102538T) was obtained from Institut Pasteur Collection (CIP, Paris, France). Chromosomal DNA was extracted and a pyruvate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-GCGTTAATTAATGAGTTACTGTCCGTACC-3', depicted in SEQ ID N° 12;

5'-TATGCGGCCGCTTAGAGGAGCTTGTTAACAGG-3', depicted in SEQ ID N° 13;

[0103] The DNA fragment amplified using the two primers depicted in SEQ ID N° 12 and SEQ ID N° 13, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described below) previously digested by PacI and NotI to yield respectively plasmid pEVL107 and plasmid pEVL420. The nucleotide sequence of the cloned gene as well as the encoded sequence of the corresponding polypeptide can be found at GenBank (accession number AF124349) and is shown in SEQ ID NO: 18 and SEQ ID NO: 19, respectively.

[0104] Plasmid pSP100 was obtained by introducing a ribosomal binding site, a PacI and a NotI restriction sites into a pUC18 vector previously digested by EcoRI and BamHI using standard protocols. The complete nucleotide sequence of pSP100 is depicted in SEQ ID N° 14.

[0105] Plasmid pEVL5 was obtained by introducing a ribosomal binding site, a PacI and a NotI restriction sites into a pQE70 vector (Qiagen) previously digested by EcoRI and BamHI using standard protocols. The complete nucleotide sequence of pEVL5 is depicted in SEQ ID N° 15.

Example 9: Cloning of a gene encoding a pyruvate decarboxylase from *Saccharomyces cerevisiae*

[0106] Chromosomal DNA was extracted from *Saccharomyces cerevisiae* strain S288C (ATCC 204508) and a pyruvate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-ATATTTAATTAATGTCTGAAATTACTTTGG-3', depicted in SEQ ID N° 16;

5'-ATATGCGGCCGCTTATTGCTTAGCGTTGGT-3', depicted in SEQ ID N° 17;

[0107] The DNA fragment amplified using the two primers depicted in SEQ ID N° 16 and SEQ ID N° 17, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described in example 8) previously digested by PacI and NotI to yield respectively plasmid pVDM61 and plasmid pEVL419. The nucleotide sequence of the cloned gene as well as the encoded sequence of the corresponding polypeptide can be found at GenBank (accession number NC001144) and is shown in SEQ ID NO: 20 and SEQ ID NO: 21, respectively.

Example 10: Expression of a pyruvate decarboxylase activity in *Escherichia coli* and enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate

Expression of pyruvate decarboxylase and preparation of cell-free extracts

[0108] Competent cells of *E. coli* MG1655 strain were transformed with either pEVL107 or pVDM61 (constructed as described in examples 8 and 9) yielding respectively strain +1735 and strain +844. These strains were cultivated at 37°C in Luria-Bertani (LB) medium (Difco) containing 100 mg/l ampicillin until OD(600 nm) reached a value around 1.5.

[0109] Competent cells of *E. coli* MG1655 strain harbouring pREP4 plasmid (Qiagen) were transformed with either pEVL420 or pEVL419 (constructed as described in Examples 8 and 9) yielding respectively strain +3150 and +3148. These strains were cultivated at 37°C in Luria-Bertani (LB) medium (Difco) containing 100 mg/l ampicillin and 30 mg/l kanamycin until OD(600 nm) reached a value of 0.6. Then isopropyl-β-D-thiogalactopyranoside (IPTG) was added to a 0.5 mM final concentration. After a further cultivation period of 2 hours and 30 minutes, cells were collected by centrifugation and washed once with 20 mM sodium phosphate buffer pH 7.2.

[0110] For each strain a cell-free extract was prepared using the same protocol as described in Example 2. Then crude cell-free extracts were passed through a PD-10 column (Amersham) equilibrated with 50 mM Tris-acetate buffer pH 6 and stored at -20°C.

Enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate

[0111] 1.0 ml of cell-free extract was mixed with 20 mM sodium 2-dehydro-3-deoxy-D-gluconate, 0.5 mM thiamine pyrophosphate and 5 mM MgCl₂ in a total volume of 1.5 ml of 50 mM tris-acetate buffer pH 6. The progression of 2-deoxy-D-ribose (DRI) synthesis was followed by analysing aliquots taken after increasing periods of incubation at 37°C. About 1 µl of each aliquot which had been previously concentrated five-fold by evaporation was deposited on a silica plate and chromatographed in the following solvent system: butanol / triethylamine / water (10/2/5). A blue spot of DRI (R_f -0.50) was detected after revelation with orcinol when using cell-free extracts of either strain +3150 or +3148 after a period of incubation of 65 hours. The crude preparation containing the spot corresponding to DRI was concentrated and passed through a 1.5 ml silica column equilibrated with isopropanol. The fractions containing the expected DRI compound were pooled, concentrated and the resulting sample analysed by mass spectrometry. The results of such an analysis confirmed the identity of the isolated compound with DRI, and the production of DRI from KDG catalysed by pyruvate decarboxylase either from *Zymomonas mobilis* or from *Saccharomyces cerevisiae*.

Example 11: Cloning of a gene encoding a pyruvate decarboxylase from *Acetobacter pasteurianus*, expression of encoded pyruvate decarboxylase activity in *Escherichia coli* and enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate

[0112] *Acetobacter pasteurianus* strain NCIB 8618 (DSMZ 2347) was obtained from DSMZ Collection (Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany). Chromosomal DNA was extracted from the cells and a pyruvate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-TCTTTAATTAATGGGTTGTCCGTCATTCATATA-3', depicted in SEQ ID N° 22;

5'-CTAAAGCTTTTAGGCCAGAGTGGTCTTGCGCG-3', depicted in SEQ ID N° 23;

[0113] The DNA fragment amplified using the two primers depicted in SEQ ID N° 22 and SEQ ID N° 23, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described in example 8) previously digested by PaeI and NotI to yield respectively plasmid pEVL541 and plasmid pEVL560. The nucleotide sequence SEQ ID N° 24 of the cloned gene as well as the encoded sequence of the corresponding polypeptide SEQ ID N° 25 can be found at GenBank (accession number AF368435).

[0114] Competent cells of *E. coli* MG1655 strain were transformed with pEVL541 yielding strain +3559. Competent cells of *E. coli* MG1655 strain harbouring pREP4 plasmid (Qiagen) were transformed with pEVL560 yielding strain +3924. These strains were cultivated and cell-free extracts were prepared as described in Example 10. Cell-free extracts were incubated with KDG and the progression of 2-deoxy-D-ribose (DRI) synthesis was followed as described in Example 10. A spot corresponding to DRI was observed indicating that pyruvate decarboxylase from *Acetobacter pasteurianus* was able to decarboxylate KDG into DRI.

Example 12: Cloning of a gene encoding a pyruvate decarboxylase from *Zymobacter palmae*, expression of encoded pyruvate decarboxylase activity in *Escherichia coli* and enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate

[0115] *Zymobacter palmae* strain T109 (DSMZ10491) was obtained from DSMZ Collection (Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany). Chromosomal DNA was extracted from the cells and a pyruvate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-ATCTTAATTAATGTATACCGTTGGTATGTACT-3', depicted in SEQ ID N° 26;

5'-TATGCGGCCGCTTACGCTTGTGGTTTGCAGAGT-3', depicted in SEQ ID N° 27;

[0116] The DNA fragment amplified using the two primers depicted in SEQ ID N° 26 and SEQ ID N° 27, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described in example 8) previously digested by PaeI and NotI to yield respectively plasmid pEVL546 and plasmid pEVL561. The nucleotide sequence of the cloned gene as well as the encoded sequence of the corresponding polypeptide is shown in SEQ ID NOs: 28 and 29, respectively and can be

found at GenBank (accession number AF474145).

[0117] Competent cells of *E. coli* MG1655 strain were transformed with pEVL546 yielding strain +3568. Competent cells of *E. coli* MG1655 strain harbouring pREP4 plasmid (Qiagen) were transformed with pEVL560 yielding strain +3923. These strains were cultivated and cell-free extracts were prepared as described in Example 10. Cell-free extracts were incubated with KDG and the progression of 2-deoxy-D-ribose (DRI) synthesis was followed as described in Example 10. A spot corresponding to DRI was observed indicating that pyruvate decarboxylase from *Zymobacter palmae* was able to decarboxylate KDG into DRI.

Example 13. Cloning of a gene encoding a benzoylformate decarboxylase from *Pseudomonas putida*, expression of encoded benzoylformate decarboxylase activity in *Escherichia coli* and enzymatic synthesis of 2-deoxy-D-ribose from 2-dehydro-3-deoxy-D-gluconate.

[0118] *Pseudomonas putida* strain Migula (DSMZ 291) was obtained from DSMZ Collection (Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH, Braunschweig, Germany). Chromosomal DNA was extracted from and a benzoylformate decarboxylase gene was amplified by PCR according to standard protocols using the following primers:

5'-CTATTAATTAATGGCTTCGGTACACGGCACCA-3', depicted in SEQ ID N° 30;

5'-TATGCGGCCGCTTACTTCACCGGGCTTACGGTGC-3', depicted in SEQ ID N° 31;

[0119] The DNA fragment amplified using the two primers depicted in SEQ ID N° 30 and SEQ ID N° 31, was ligated either into pSP100 or into pEVL5 (respectively a pUC18-derived or a pQE70-derived vector as described in example 8) previously digested by *PacI* and *NotI* to yield respectively plasmid pEVL681 and plasmid pEVL670. The nucleotide sequence SEQ ID N° 32 of the cloned gene as well as the encoded sequence of the corresponding polypeptide SEQ ID N° 33 can be found at GenBank (accessing number AY143338).

[0120] Competent cells of *E. coli* MG1655 strain were transformed with pEVL681 yielding strain +4050. Competent cells of *E. coli* MG1655 strain harbouring pREP4 plasmid (Qiagen) were transformed with pEVL670 yielding strain +3927. Those strains were cultivated and cell-free extracts were prepared as described in example 10. Cell-free extracts were incubated with KDG and the progression of 2-deoxy-D-ribose (DRI) synthesis was followed as described in example 10. A spot corresponding to DRI was observed indicating that benzoylformate decarboxylase from *Pseudomonas putida* was able to decarboxylate KDG into DRI.

Preparative enzymatic synthesis of 2-deoxy-D-ribose

[0121] 100 µl of cell-free extract from strain +3927 (containing 2.5 mg of bacterial proteins) were mixed with 300 mM sodium 2-dehydro-3-deoxy-D-gluconate, 0.5 mM thiamine pyrophosphate and 5 mM MgCl₂ in a total volume of 0.5 ml of 80 mM potassium phosphate buffer pH 6. After a period of incubation of 16 and 40 hours, few µl of a solution of HCl 2N were added to the incubation mixture until the pH reached a value of 6. The progression of 2-deoxy-D-ribose (DRI) synthesis was also followed by analysing aliquots taken after increasing periods of incubation at 37°C. About 1 µl of each aliquot was deposited on a silica plate and chromatographed as described in example 10. The concentration of 2-deoxy-D-ribose was estimated to be about 200 mM by comparison with standard solutions. ¹³C NMR analysis of the crude mixture confirmed that the compound formed from 2-dehydro-3-deoxy-D-gluconate was 2-deoxy-D-ribose, and that the concentration of 2-deoxy-D-ribose was closed to 25 g/l. Another preparative enzymatic synthesis was performed in the same conditions except that no addition of acid was made along the incubation period. In those conditions, the concentration of 2-deoxy-D-ribose was closed to 10 g/l, far lower than the concentration reached in the preceding experiment for which the pH had been controlled and regularly adjusted to a value of 6.

Example 14. Enzymatic synthesis of 2-deoxy-D-ribose from D-gluconate

[0122] One pot enzymatic synthesis of 2-deoxy-D-ribose from D-gluconate was achieved as follows, using D-gluconate dehydratase encoded by *gcnD* gene of *Agrobacterium tumefaciens* and pyruvate decarboxylase from *Zymomonas mobilis*:

50 µl of cell-free extract from strain +1289 (containing 1.5 mg of bacterial proteins) and 400 µl of cell-free extract from strain +3150 (containing 17 mg of bacterial proteins after concentration by ultrafiltration) prepared as described respectively in example

2 and in example 10, were mixed with 50 mM potassium D-gluconate, 0.5 mM thiamine pyrophosphate and 5 mM MgCl₂ in a total volume of 0.5 ml of 50 mM N-(2-hydroxyethyl)piperazine-N'-(2-ethanesulfonic acid) (HEPES) buffer pH 7. The progression of 2-deoxy-D-ribose (DRI) synthesis was also followed by analysing aliquots taken after increasing periods of incubation at 37°C. After a period of incubation of 18 hours, about 1 µl of the incubation mixture was deposited on a silica plate and chromatographed as described in example 10. The concentration of 2-deoxy-D-ribose was estimated to be about 1 g/l by comparison with standard solutions.

SEQUENCE LISTING

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Maliere Technologies Société Civile

Rhodia Chimie

Marliere, Phillipe

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gac gcc cgc aac cta ctc gca tcc ggc aag gtt gtc ttc aag gca tcc      912
Asp Ala Arg Asn Leu Leu Ala Ser Gly Lys Val Val Phe Lys Ala Ser
      290      295      300

gag gca ttt cag ccc tca atg cga atc tgg gcg gag gtc atc tcc gtt      960
Glu Ala Phe Gln Pro Ser Met Arg Ile Trp Ala Glu Val Ile Ser Val
      305      310      315      320

cct gag ccg ggg ctg gcg atc gtc gcc atg gcc atg ccg gat gta tgc      1008
Pro Glu Pro Gly Leu Ala Ile Val Gly Met Gly Met Arg Asp Val Ser
      325      330      335

ttc gat cag gac ctg ccc gtg gcg ctt cgg ctc cat agg gac gga cat      1056
Phe Asp Gln Asp Leu Pro Val Ala Leu Arg Leu His Arg Asp Gly His

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ctg gtc gaa gct gat ctc tct tca tcc gcg aag gtc ggc aag ctc aat      1104
Leu Val Glu Ala Asp Leu Ser Ser Ser Ala Lys Val Gly Lys Leu Asn
      355      360      365

gac cag cat gcc ttc ttg tcc ttc ggg aac ggc agc agt ctg gca atc      1152
Asp Gln His Ala Phe Leu Ser Phe Gly Asn Gly Ser Ser Leu Ala Ile
      370      375      380

ggc gat gtc ata gaa ttc ggc atc tcg cat ccc tgc acg tgc ttc gat      1200
Gly Asp Val Ile Glu Phe Gly Ile Ser His Pro Cys Thr Cys Phe Asp
      385      390      395      400

cgc tgg cgc gtc ttt cac gga atc gat gga tca ggc cgg atc cag cgc      1248
Arg Trp Arg Val Phe His Gly Ile Asp Gly Ser Gly Arg Ile Gln Arg
      405      410      415

atc tac aca acc ttc ttt cac tag      1272
Ile Tyr Thr Thr Phe Phe His
      420

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<210> 4

<211> 423

<212> PRT

<213> Agrobacterium tumefaciens

<400> 4

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Leu Ala Leu Pro Leu Leu Thr Val Asp Leu Ala Val Tyr Arg Gly Asn
      35      40      45

Arg Asp Arg Phe Leu Ala Leu Val Ser Ala His Gly Ala Lys Ala Ala
      50      55      60

Pro His Ala Lys Thr Pro Met Cys Pro Glu Ile Ala Ile Asp Leu Ile
      65      70      75      80

Glu Ala Gly Ala Trp Gly Ala Thr Val Ala Asp Leu Phe Gln Ala Glu
      85      90      95

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Val Leu Leu Lys Ala Gly Val Ser Asn Ile Leu Ile Ala Asn Gln Ile
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 Gly Gly Leu Thr Ser Ala Arg Arg Leu Arg Met Leu Ala Asp Ala Phe
 115 120 125
 Pro Lys Ala Glu Ile Ile Cys Cys Val Asp Ser Val Gln Ala Ser Ala
 130 135 140
 Asn Leu Val Gln Ala Phe Gln Gly Arg Val Asp Ala Pro Phe Lys Val
 145 150 155 160
 Phe Ile Glu Val Gly Val Gly Arg Thr Gly Ala Arg Thr Leu Asn Val
 165 170 175
 Ala Lys Asp Ile Ile Asp Thr Ile Ser Thr Ser Ala Glu Ile Val Leu
 180 185 190
 Ala Gly Val Ser Thr Tyr Glu Gly Ser Val Ser Gly Glu Thr Ser Glu
 195 200 205
 Ala Leu Asp Ala Asn Met Ala Ala Leu Phe Asp Leu Leu Thr Asp Ser
 210 215 220
 Leu Ala Ser Ile Arg Glu Lys Asp Pro Gly Arg Pro Leu Thr Val Ser
 225 230 235 240
 Ala Gly Gly Ser Ile His Phe Asp Arg Val Leu Ala Ala Leu Val Pro
 245 250 255
 Val Cys Glu Ala Asp Gly Asn Ala Thr Leu Leu Leu Arg Ser Gly Ala
 260 265 270
 Ile Phe Phe Ser Asp His Gly Val Tyr Gln Arg Gly Phe Gln Ala Val
 275 280 285
 Asp Ala Arg Asn Leu Leu Ala Ser Gly Lys Val Val Phe Lys Ala Ser
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 Glu Ala Phe Gln Pro Ser Met Arg Ile Trp Ala Glu Val Ile Ser Val
 305 310 315 320
 Pro Glu Pro Gly Leu Ala Ile Val Gly Met Gly Met Arg Asp Val Ser
 325 330 335
 Phe Asp Gln Asp Leu Pro Val Ala Leu Arg Leu His Arg Asp Gly His
 340 345 350
 Leu Val Glu Ala Asp Leu Ser Ser Ser Ala Lys Val Gly Lys Leu Asn
 355 360 365
 Asp Gln His Ala Phe Leu Ser Phe Gly Asn Gly Ser Ser Leu Ala Ile
 370 375 380
 Gly Asp Val Ile Glu Phe Gly Ile Ser His Pro Cys Thr Cys Phe Asp
 385 390 395 400
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<400> 6

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<212> DNA

<213> artificial sequence

<220>

<223> oligonucleotide primer

<400> 7

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<212> DNA

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<400> 10

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<212> DNA

<213> artificial sequence

<220>

<223> oligonucleotide primer

<400> 11

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<212> DNA

<213> vector

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<212> DNA

<213> vector

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<220>

<223> artificial sequence

<400> 16

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<210> 17

<211> 30

<212> DNA

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<220>

<223> artificial sequence

<400> 17

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<210> 18

<211> 1707

<212> DNA

<213> *Zymomonas mobilis*

<220>

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<222> (1)..(1707)

<223>

<400> 18

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Gly Leu Lys His His Phe Ala Val Ala Gly Asp Tyr Asn Leu Val Leu	
20 25 30	
ctt gac aac ctg ctt ttg aac aaa aac atg gag cag gtt tat tgc tgt	144
Leu Asp Asn Leu Leu Leu Asn Lys Asn Met Glu Gln Val Tyr Cys Cys	
35 40 45	
aac gaa ctg aac tgc ggt ttc agt gca gaa ggt tat gct cgt gcc aaa	192
Asn Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ala Lys	
50 55 60	
ggc gca gca gca gcc gtc gtt acc tac agc gtc ggt gcg ctt tcc gca	240
Gly Ala Ala Ala Ala Val Val Thr Tyr Ser Val Gly Ala Leu Ser Ala	
65 70 75 80	
ttt gat gct atc ggt gcc gcc tat gca gaa aac ctt ccg gtt atc ctg	288
Phe Asp Ala Ile Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu	
85 90 95	
atc tcc ggt gct ccg aac aac aat gat cac gct gct ggt cac gtg ttg	336
Ile Ser Gly Ala Pro Asn Asn Asn Asp His Ala Ala Gly His Val Leu	
100 105 110	
cat cac gct ctt gcc aaa acc gac tat cac tat cag ttg gaa atg gcc	384
His His Ala Leu Gly Lys Thr Asp Tyr His Tyr Gln Leu Glu Met Ala	
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aag aac atc acg gcc gcc gct gaa gcg att tac acc ccg gaa gaa gct	432
Lys Asn Ile Thr Ala Ala Ala Glu Ala Ile Tyr Thr Pro Glu Glu Ala	
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Pro Ala Lys Ile Asp His Val Ile Lys Thr Ala Leu Arg Glu Lys Lys	
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Pro Val Tyr Leu Glu Ile Ala Cys Asn Ile Ala Ser Met Pro Cys Ala	
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Ala Pro Gly Pro Ala Ser Ala Leu Phe Asn Asp Glu Ala Ser Asp Glu	
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gct tct ttg aat gca gcg gtt gaa gaa acc ctg aaa ttc atc gcc aac	624
Ala Ser Leu Asn Ala Ala Val Glu Glu Thr Leu Lys Phe Ile Ala Asn	
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Arg Asp Lys Val Ala Val Leu Val Gly Ser Lys Leu Arg Ala Ala Gly	
210 215 220	
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Ala Glu Glu Ala Ala Val Lys Phe Ala Asp Ala Leu Gly Gly Ala Val
 225 230 235 240

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tac atc ggc acc tca tgg ggt gaa gtc agc tat ccg ggc gtt gaa aag 816
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 260 265 270

acg atg aaa gaa gcc gat gcg gtt atc gct ctg gct cct gtc ttc aac 864
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 275 280 285

gac tac tcc acc act ggt tgg acg gat att cct gat cct aag aaa ctg 912
 Asp Tyr Ser Thr Thr Gly Trp Thr Asp Ile Pro Asp Pro Lys Lys Leu
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 Val Leu Ala Glu Pro Arg Ser Val Val Val Asn Gly Ile Arg Phe Pro
 305 310 315 320

agc gtc cat ctg aaa gac tat ctg acc cgt ttg gct cag aaa gtt tcc 1008
 Ser Val His Leu Lys Asp Tyr Leu Thr Arg Leu Ala Gln Lys Val Ser
 325 330 335

aag aaa acc ggt gca ttg gac ttc ttc aaa tcc ctg aat gca ggt gaa 1056
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ctg aag aaa gcc gct ccg gct gat ccg agt gct ccg ttg gtc aac gca 1104
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 355 360 365

gaa atc gcc cgt cag gtc gaa gct ctt ctg acc ccg aac acg acg gtt 1152
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 370 375 380

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 420 425 430

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gaa gtc gct cag atg gtt cgc ctg aaa ctg ccg gtt atc atc ttc ttg 1392
 Glu Val Ala Gln Met Val Arg Leu Lys Leu Pro Val Ile Ile Phe Leu
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 Ile Asn Asn Tyr Gly Tyr Thr Ile Glu Val Met Ile His Asp Gly Pro
 465 470 475 480

tac aac aac atc aag aac tgg gat tat gcc ggt ctg atg gaa gtg ttc 1488
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 485 490 495

aac ggt aac ggt ggt tat gac agc ggt gct ggt aaa ggc ctg aag gct 1536
 Asn Gly Asn Gly Gly Tyr Asp Ser Gly Ala Gly Lys Gly Leu Lys Ala
 500 505 510

aaa acc ggt ggc gaa ctg gca gaa gct atc aag gtt gct ctg gca aac 1584
 Lys Thr Gly Gly Glu Leu Ala Glu Ala Ile Lys Val Ala Leu Ala Asn
 515 520 525

acc gac ggc cca acc ctg atc gaa tgc ttc atc ggt cgt gaa gac tgc 1632
 Thr Asp Gly Pro Thr Leu Ile Glu Cys Phe Ile Gly Arg Glu Asp Cys
 530 535 540

act gaa gaa ttg gtc aaa tgg ggt aag cgc gtt gct gcc gcc aac agc 1680
 Thr Glu Glu Leu Val Lys Trp Gly Lys Arg Val Ala Ala Asn Ser
 545 550 555 560

cgt aag cct gtt aac aag ctg ctg tag 1707
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<210> 19

<211> 568

<212> PRT

<213> Zymomonas mobilis

<400> 19

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Leu Asp Asn Leu Leu Leu Asn Lys Asn Met Glu Gln Val Tyr Cys Cys
35          40          45

Asn Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ala Lys
50          55          60

Gly Ala Ala Ala Ala Val Val Thr Tyr Ser Val Gly Ala Leu Ser Ala
65          70          75          80

Phe Asp Ala Ile Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu
85          90          95

Ile Ser Gly Ala Pro Asn Asn Asn Asp His Ala Ala Gly His Val Leu
100         105         110

His His Ala Leu Gly Lys Thr Asp Tyr His Tyr Gln Leu Glu Met Ala
115         120         125

Lys Asn Ile Thr Ala Ala Ala Glu Ala Ile Tyr Thr Pro Glu Glu Ala
130         135         140

Pro Ala Lys Ile Asp His Val Ile Lys Thr Ala Leu Arg Glu Lys Lys
145         150         155         160

Pro Val Tyr Leu Glu Ile Ala Cys Asn Ile Ala Ser Met Pro Cys Ala
165         170         175

Ala Pro Gly Pro Ala Ser Ala Leu Phe Asn Asp Glu Ala Ser Asp Glu
180         185         190

Ala Ser Leu Asn Ala Ala Val Glu Glu Thr Leu Lys Phe Ile Ala Asn
195         200         205

Arg Asp Lys Val Ala Val Leu Val Gly Ser Lys Leu Arg Ala Ala Gly
210         215         220

Ala Glu Glu Ala Ala Val Lys Phe Ala Asp Ala Leu Gly Gly Ala Val
225         230         235         240

Ala Thr Met Ala Ala Ala Lys Ser Phe Phe Pro Glu Glu Asn Pro His
245         250         255

Tyr Ile Gly Thr Ser Trp Gly Glu Val Ser Tyr Pro Gly Val Glu Lys
260         265         270

Thr Met Lys Glu Ala Asp Ala Val Ile Ala Leu Ala Pro Val Phe Asn
275         280         285

Asp Tyr Ser Thr Thr Gly Trp Thr Asp Ile Pro Asp Pro Lys Lys Leu
290         295         300

Val Leu Ala Glu Pro Arg Ser Val Val Val Asn Gly Ile Arg Phe Pro

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305 310 315 320
 Ser Val His Leu Lys Asp Tyr Leu Thr Arg Leu Ala Gln Lys Val Ser
 325 330 335
 Lys Lys Thr Gly Ala Leu Asp Phe Phe Lys Ser Leu Asn Ala Gly Glu
 340 345 350
 Leu Lys Lys Ala Ala Pro Ala Asp Pro Ser Ala Pro Leu Val Asn Ala
 355 360 365
 Glu Ile Ala Arg Gln Val Glu Ala Leu Leu Thr Pro Asn Thr Thr Val
 370 375 380
 Ile Ala Glu Thr Gly Asp Ser Trp Phe Asn Ala Gln Arg Met Lys Leu
 385 390 395 400
 Pro Asn Gly Ala Arg Val Glu Tyr Glu Met Gln Trp Gly His Ile Gly
 405 410 415
 Trp Ser Val Pro Ala Ala Phe Gly Tyr Ala Val Gly Ala Pro Glu Arg
 420 425 430
 Arg Asn Ile Leu Met Val Gly Asp Gly Ser Phe Gln Leu Thr Ala Gln
 435 440 445
 Glu Val Ala Gln Met Val Arg Leu Lys Leu Pro Val Ile Ile Phe Leu
 450 455 460
 Ile Asn Asn Tyr Gly Tyr Thr Ile Glu Val Met Ile His Asp Gly Pro
 465 470 475 480
 Tyr Asn Asn Ile Lys Asn Trp Asp Tyr Ala Gly Leu Met Glu Val Phe
 485 490 495
 Asn Gly Asn Gly Gly Tyr Asp Ser Gly Ala Gly Lys Gly Leu Lys Ala
 500 505 510
 Lys Thr Gly Gly Glu Leu Ala Glu Ala Ile Lys Val Ala Leu Ala Asn
 515 520 525
 Thr Asp Gly Pro Thr Leu Ile Glu Cys Phe Ile Gly Arg Glu Asp Cys
 530 535 540
 Thr Glu Glu Leu Val Lys Trp Gly Lys Arg Val Ala Ala Ala Asn Ser
 545 550 555 560
 Arg Lys Pro Val Asn Lys Leu Leu
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<210> 20

<211> 1692

<212> DNA

<213> *Saccharomyces cerevisiae*

<220>

<221> CDS

<222> (1)..(1692)

<223>

<400> 20

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atg tct gaa att act ttg ggt aaa tat ttg ttc gaa aga tta aag caa      48
Met Ser Glu Ile Thr Leu Gly Lys Tyr Leu Phe Glu Arg Leu Lys Gln
1      5      10      15

gtc aac gtt aac acc gtt ttc ggt ttg cca ggt gac ttc aac ttg tcc      96
Val Asn Val Asn Thr Val Phe Gly Leu Pro Gly Asp Phe Asn Leu Ser
      20      25      30

ttg ttg gac aag atc tac gaa gtt gaa ggt atg aga tgg gct ggt aac      144
Leu Leu Asp Lys Ile Tyr Glu Val Glu Gly Met Arg Trp Ala Gly Asn
      35      40      45

gcc aac gaa ttg aac gct gct tac gcc gct gat ggt tac gct cgt atc      192
Ala Asn Glu Leu Asn Ala Ala Tyr Ala Ala Asp Gly Tyr Ala Arg Ile
      50      55      60

aag ggt atg tct tgt atc atc acc acc ttc ggt gtc ggt gaa ttg tct      240
Lys Gly Met Ser Cys Ile Ile Thr Thr Phe Gly Val Gly Glu Leu Ser
      65      70      75      80

gct ttg aac ggt att gcc ggt tct tac gct gaa cac gtc ggt gtt ttg      288
Ala Leu Asn Gly Ile Ala Gly Ser Tyr Ala Glu His Val Gly Val Leu
      85      90      95

cac gtt gtt ggt gtc cca tcc atc tct gct caa gct aag caa ttg ttg      336
His Val Val Gly Val Pro Ser Ile Ser Ala Gln Ala Lys Gln Leu Leu
      100      105      110

ttg cac cac acc ttg ggt aac ggt gac ttc act gtt ttc cac aga atg      384
Leu His His Thr Leu Gly Asn Gly Asp Phe Thr Val Phe His Arg Met
      115      120      125

tct gcc aac att tct gaa acc act gct atg atc act gac att gct acc      432
Ser Ala Asn Ile Ser Glu Thr Thr Ala Met Ile Thr Asp Ile Ala Thr
      130      135      140

gcc cca gct gaa att gac aga tgt atc aga acc act tac gtc acc caa      480
Ala Pro Ala Glu Ile Asp Arg Cys Ile Arg Thr Thr Tyr Val Thr Gln
      145      150      155      160

aga cca gtc tac tta ggt ttg cca gct aac ttg gtc gac ttg aac gtc      528
Arg Pro Val Tyr Leu Gly Leu Pro Ala Asn Leu Val Asp Leu Asn Val
      165      170      175

cca gct aag ttg ttg caa act cca att gac atg tct ttg aag cca aac      576
Pro Ala Lys Leu Leu Gln Thr Pro Ile Asp Met Ser Leu Lys Pro Asn
      180      185      190

gat gct gaa tcc gaa aag gaa gtc att gac acc atc ttg gct ttg gtc      624
Asp Ala Glu Ser Glu Lys Glu Val Ile Asp Thr Ile Leu Ala Leu Val
      195      200      205

aag gat gct aag aac cca gtt atc ttg gct gat gct tgt tgt tcc aga      672
Lys Asp Ala Lys Asn Pro Val Ile Leu Ala Asp Ala Cys Cys Ser Arg
      210      215      220

cac gac gtc aag gct gaa act aag aag ttg att gac ttg act caa ttc      720
His Asp Val Lys Ala Glu Thr Lys Lys Leu Ile Asp Leu Thr Gln Phe
      225      230      235      240

cca gct ttc gtc acc cca atg ggt aag ggt tcc att gac gaa caa cac      768
Pro Ala Phe Val Thr Pro Met Gly Lys Gly Ser Ile Asp Glu Gln His
      245      250      255

cca aga tac ggt ggt gtt tac gtc ggt acc ttg tcc aag cca gaa gtt      816
Pro Arg Tyr Gly Val Tyr Val Gly Thr Leu Ser Lys Pro Glu Val
      260      265      270

aag gaa gcc gtt gaa tct gct gac ttg att ttg tct gtc ggt gct ttg      864
Lys Glu Ala Val Glu Ser Ala Asp Leu Ile Leu Ser Val Gly Ala Leu
      275      280      285

ttg tct gat ttc aac acc ggt tct ttc tct tac tct tac aag acc aag      912
Leu Ser Asp Phe Asn Thr Gly Ser Phe Ser Tyr Ser Tyr Lys Thr Lys
      290      295      300

aac att gtc gaa ttc cac tcc gac cac atg aag atc aga aac gcc act      960
Asn Ile Val Glu Phe His Ser Asp His Met Lys Ile Arg Asn Ala Thr
      305      310      315      320

ttc cca ggt gtc caa atg aaa ttc gtt ttg caa aag ttg ttg acc act      1008
Phe Pro Gly Val Gln Met Lys Phe Val Leu Gln Lys Leu Leu Thr Thr
      325      330      335

att gct gac gcc gct aag ggt tac aag cca gtt gct gtc cca gct aga      1056
Ile Ala Asp Ala Ala Lys Gly Tyr Lys Pro Val Ala Val Pro Ala Arg
      340      345      350

act cca gct aac gct gct gtc cca gct tct acc cca ttg aag caa gaa      1104

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Thr Pro Ala Asn Ala Ala Val Pro Ala Ser Thr Pro Leu Lys Gln Glu	
355 360 365	
tgg atg tgg aac caa ttg ggt aac ttc ttg caa gaa ggt gat gtt gtc	1152
Trp Met Trp Asn Gln Leu Gly Asn Phe Leu Gln Gly Asp Val Val	
370 375 380	
att gct gaa acc ggt acc tcc gct ttc ggt atc aac caa acc act ttc	1200
Ile Ala Glu Thr Gly Thr Ser Ala Phe Gly Ile Asn Gln Thr Thr Phe	
385 390 395 400	
cca aac aac acc tac ggt atc tct caa gtc tta tgg ggt tcc att ggt	1248
Pro Asn Asn Thr Tyr Gly Ile Ser Gln Val Leu Trp Gly Ser Ile Gly	
405 410 415	
ttc acc act ggt gct acc ttg ggt gct gct ttc gct gct gaa gaa att	1296
Phe Thr Thr Gly Ala Thr Leu Gly Ala Phe Ala Ala Glu Glu Ile	
420 425 430	
gat cca aag aag aga gtt atc tta ttc att ggt gac ggt tct ttg caa	1344
Asp Pro Lys Lys Arg Val Ile Leu Phe Ile Gly Asp Gly Ser Leu Gln	
435 440 445	
ttg act gtt caa gaa atc tcc acc atg atc aga tgg ggc ttg aag cca	1392
Leu Thr Val Gln Glu Ile Ser Thr Met Ile Arg Trp Gly Leu Lys Pro	
450 455 460	
tac ttg ttc gtc ttg aac aac gat ggt tac acc att gaa aag ttg att	1440
Tyr Leu Phe Val Leu Asn Asn Asp Gly Tyr Thr Ile Glu Lys Leu Ile	
465 470 475 480	
cac ggt cca aag gct caa tac aac gaa att caa ggt tgg gac cac cta	1488
His Gly Pro Lys Ala Gln Tyr Asn Glu Ile Gln Gly Trp Asp His Leu	
485 490 495	
tcc ttg ttg cca act ttc ggt gct aag gac tat gaa acc cac aga gtc	1536
Ser Leu Leu Pro Thr Phe Gly Ala Lys Asp Tyr Glu Thr His Arg Val	
500 505 510	
gct acc acc ggt gaa tgg gac aag ttg acc caa gac aag tct ttc aac	1584
Ala Thr Thr Gly Glu Trp Asp Lys Leu Thr Gln Asp Lys Ser Phe Asn	
515 520 525	
gac aac tct aag atc aga atg att gaa atc atg ttg cca gtc ttc gat	1632
Asp Asn Ser Lys Ile Arg Met Ile Glu Ile Met Leu Pro Val Phe Asp	
530 535 540	
gct cca caa aac ttg gtt gaa caa gct aag ttg act gct gct acc aac	1680
Ala Pro Gln Asn Leu Val Glu Gln Ala Lys Leu Thr Ala Ala Thr Asn	
545 550 555 560	
gct aag caa taa	1692
Ala Lys Gln	

<210> 21

<211> 563

<212> PRT

<213> *Saccharomyces cerevisiae*

<400> 21

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 Leu Leu Asp Lys Ile Tyr Glu Val Glu Gly Met Arg Trp Ala Gly Asn
 35 40 45
 Ala Asn Glu Leu Asn Ala Ala Tyr Ala Ala Asp Gly Tyr Ala Arg Ile
 50 55 60
 Lys Gly Met Ser Cys Ile Ile Thr Thr Phe Gly Val Gly Glu Leu Ser
 65 70 75 80
 Ala Leu Asn Gly Ile Ala Gly Ser Tyr Ala Glu His Val Gly Val Leu
 85 90 95
 His Val Val Gly Val Pro Ser Ile Ser Ala Gln Ala Lys Gln Leu Leu
 100 105 110
 Leu His His Thr Leu Gly Asn Gly Asp Phe Thr Val Phe His Arg Met
 115 120 125
 Ser Ala Asn Ile Ser Glu Thr Thr Ala Met Ile Thr Asp Ile Ala Thr
 130 135 140
 Ala Pro Ala Glu Ile Asp Arg Cys Ile Arg Thr Thr Tyr Val Thr Gln
 145 150 155 160
 Arg Pro Val Tyr Leu Gly Leu Pro Ala Asn Leu Val Asp Leu Asn Val
 165 170 175
 Pro Ala Lys Leu Leu Gln Thr Pro Ile Asp Met Ser Leu Lys Pro Asn
 180 185 190
 Asp Ala Glu Ser Glu Lys Glu Val Ile Asp Thr Ile Leu Ala Leu Val
 195 200 205

Lys Asp Ala Lys Asn Pro Val Ile Leu Ala Asp Ala Cys Cys Ser Arg
 210 215 220
 His Asp Val Lys Ala Glu Thr Lys Lys Leu Ile Asp Leu Thr Gln Phe
 225 230 235 240
 Pro Ala Phe Val Thr Pro Met Gly Lys Gly Ser Ile Asp Glu Gln His
 245 250 255
 Pro Arg Tyr Gly Gly Val Tyr Val Gly Thr Leu Ser Lys Pro Glu Val
 260 265 270
 Lys Glu Ala Val Glu Ser Ala Asp Leu Ile Leu Ser Val Gly Ala Leu
 275 280 285
 Leu Ser Asp Phe Asn Thr Gly Ser Phe Ser Tyr Ser Tyr Lys Thr Lys
 290 295 300
 Asn Ile Val Glu Phe His Ser Asp His Met Lys Ile Arg Asn Ala Thr
 305 310 315 320
 Phe Pro Gly Val Gln Met Lys Phe Val Leu Gln Lys Leu Leu Thr Thr
 325 330 335
 Ile Ala Asp Ala Ala Lys Gly Tyr Lys Pro Val Ala Val Pro Ala Arg
 340 345 350
 Thr Pro Ala Asn Ala Ala Val Pro Ala Ser Thr Pro Leu Lys Gln Glu
 355 360 365
 Trp Met Trp Asn Gln Leu Gly Asn Phe Leu Gln Glu Gly Asp Val Val
 370 375 380
 Ile Ala Glu Thr Gly Thr Ser Ala Phe Gly Ile Asn Gln Thr Thr Phe
 385 390 395 400
 Pro Asn Asn Thr Tyr Gly Ile Ser Gln Val Leu Trp Gly Ser Ile Gly
 405 410 415
 Phe Thr Thr Gly Ala Thr Leu Gly Ala Ala Phe Ala Ala Glu Glu Ile
 420 425 430
 Asp Pro Lys Lys Arg Val Ile Leu Phe Ile Gly Asp Gly Ser Leu Gln

435 440 445
 Leu Thr Val Gln Glu Ile Ser Thr Met Ile Arg Trp Gly Leu Lys Pro
 450 455 460
 Tyr Leu Phe Val Leu Asn Asn Asp Gly Tyr Thr Ile Glu Lys Leu Ile
 465 470 475 480
 His Gly Pro Lys Ala Gln Tyr Asn Glu Ile Gln Gly Trp Asp His Leu
 485 490 495
 Ser Leu Leu Pro Thr Phe Gly Ala Lys Asp Tyr Glu Thr His Arg Val
 500 505 510
 Ala Thr Thr Gly Glu Trp Asp Lys Leu Thr Gln Asp Lys Ser Phe Asn
 515 520 525
 Asp Asn Ser Lys Ile Arg Met Ile Glu Ile Met Leu Pro Val Phe Asp
 530 535 540
 Ala Pro Gln Asn Leu Val Glu Gln Ala Lys Leu Thr Ala Ala Thr Asn
 545 550 555 560

Ala Lys Gln

<210> 22

<211> 33

<212> DNA

<213> artificial sequence

<220>

<223> artificial sequence

<400> 22

tctttaatta atgggttgtc cgtcattcat ata 33

<210> 23

<211> 32

<212> DNA

<213> artificial sequence

<220>

<223> artificial sequence

<400> 23

ctaaagcttt taggccagag tggcttgcg cg 32

<210> 24

<211> 1674

<212> DNA

<213> Acetobacter pasteurianus

<220>

<221> CDS

<222> (1)..(1674)

<223>

<400> 24

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1 5 10 15

ggg ctg aag cat cac ttc gcc gtg ggc ggc gac tac aat ctc gtt ctt 96
Gly Leu Lys His His Phe Ala Val Gly Gly Asp Tyr Asn Leu Val Leu
20 25 30

ctg gat cag ttg ctc ctc aac aag gac atg aaa cag atc tat tgc tgc 144
Leu Asp Gln Leu Leu Leu Asn Lys Asp Met Lys Gln Ile Tyr Cys Cys
35 40 45

aat gag ttg aac tgt ggc ttc agc gcg gaa ggc tac gcc cgt tct aac 192
Asn Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ser Asn
50 55 60

ggg gct gcg gca gcg gtt gtc acc ttc agc gtt ggc gcc att tcc gcc 240
Gly Ala Ala Ala Ala Val Val Thr Phe Ser Val Gly Ala Ile Ser Ala
65 70 75 80

atg aac gcc ctc ggc ggc gcc tat gcc gaa aac ctg ccg gtt atc ctg 288
Met Asn Ala Leu Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu
85 90 95

att tcc ggc gcg ccc aac agc aat gat cag ggc aca ggt cat atc ctg 336
Ile Ser Gly Ala Pro Asn Ser Asn Asp Gln Gly Thr Gly His Ile Leu
100 105 110

cat cac aca atc ggc aag acg gat tac agc tac cag ctt gaa atg gcc 384
His His Thr Ile Gly Lys Thr Asp Tyr Ser Tyr Gln Leu Glu Met Ala
115 120 125

cgt cag gtc acc tgt gcc gcc gaa agc att acc gac gct cac tcc gcc 432
Arg Gln Val Thr Cys Ala Ala Glu Ser Ile Thr Asp Ala His Ser Ala
130 135 140

ccg gcc aag att gac cac gtc att cgc acg gcg ctg cgc gag cgt aag 480
Pro Ala Lys Ile Asp His Val Ile Arg Thr Ala Leu Arg Glu Arg Lys
145 150 155 160

ccg gcc tat ctg gac atc gcg tgc aac att gcc tcc gag ccc tgc gtg 528
Pro Ala Tyr Leu Asp Ile Ala Cys Asn Ile Ala Ser Glu Pro Cys Val
165 170 175

cgg cct ggc cct gtc agc agc ctg ctg tcc gag cct gaa atc gac cac 576
Arg Pro Gly Pro Val Ser Ser Leu Leu Ser Glu Pro Glu Ile Asp His
180 185 190

acg agc ctg aag gcc gca gtg gac gcc acg gtt gcc ttg ctg aaa aat 624
Thr Ser Leu Lys Ala Ala Val Asp Ala Thr Val Ala Leu Leu Lys Asn
195 200 205

cgg cca gcc ccc gtc atg ctg ctg ggc agc aag ctg cgg gcc gcc aac 672
Arg Pro Ala Pro Val Met Leu Leu Gly Ser Lys Leu Arg Ala Ala Asn
210 215 220

gca ctg gcc gca acc gaa acg ctg gca gac aag ctg caa tgc gcg gtg 720
Ala Leu Ala Ala Thr Glu Thr Leu Ala Asp Lys Leu Gln Cys Ala Val
225 230 235 240

acc atc atg gcg gcc gcg aaa ggc ttt ttc ccc gaa gac cac gcg ggt 768
Thr Ile Met Ala Ala Ala Lys Gly Phe Phe Pro Glu Asp His Ala Gly
245 250 255

ttc cgc ggc ctg tac tgg ggc gaa gtc tgc aac ccc ggc gtg cag gaa 816
Phe Arg Gly Leu Tyr Trp Gly Glu Val Ser Asn Pro Gly Val Gln Glu
260 265 270

ctg gtg gag acc tcc gac gca ctg ctg tgc atc gcc ccc gta ttc aac 864
Leu Val Glu Thr Ser Asp Ala Leu Leu Cys Ile Ala Pro Val Phe Asn
275 280 285

gac tat tca aca gtc ggc tgg tgc ggc atg ccc aag ggc ccc aat gtg 912
Asp Tyr Ser Thr Val Gly Trp Ser Gly Met Pro Lys Gly Pro Asn Val
290 295 300

att ctg gct gag ccc gac cgc gta acg gtc gat ggc cgc gcc tat gac 960
Ile Leu Ala Glu Pro Asp Arg Val Thr Val Asp Gly Arg Ala Tyr Asp
305 310 315 320

ggc ttt acc ctg cgc gcc ttc ctg cag gct ctg gcg gaa aaa gcc ccc 1008
Gly Phe Thr Leu Arg Ala Phe Leu Gln Ala Leu Ala Glu Lys Ala Pro
325 330 335

gcg cgc ccg gcc tcc gca cag aaa agc agc gtc ccg acg tgc tgc ctc 1056

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Ala Arg Pro Ala Ser Ala Gln Lys Ser Ser Val Pro Thr Cys Ser Leu
340                      345                      350

acc gcg aca tcc gat gaa gcc ggt ctg acg aat gac gaa atc gtc cgt 1104
Thr Ala Thr Ser Asp Glu Ala Gly Leu Thr Asn Asp Glu Ile Val Arg
355                      360                      365

cat atc aac gcc ctg ctg aca aac acg acg ctg gty gca gaa acc 1152
His Ile Asn Ala Leu Leu Thr Ser Asn Thr Thr Leu Val Ala Glu Thr
370                      375                      380

ggc gat tca tgg ttc aat gcc atg cgc atg acc ctg gcc ggt gcg cgc 1200
Gly Asp Ser Trp Phe Asn Ala Met Arg Met Thr Leu Ala Gly Ala Arg
385                      390                      395                      400

gtg gaa ctg gaa atg cag tgg ggc cat atc ggc tgg tcc gtg ccc tcc 1248
Val Glu Leu Glu Met Gln Trp Gly His Ile Gly Trp Ser Val Pro Ser
405                      410                      415

gcg ttc ggc aat gcc atg ggc tgg cag gac cgc cag cat gtg gtg atg 1296
Ala Phe Gly Asn Ala Met Gly Ser Gln Asp Arg Gln His Val Val Met
420                      425                      430

gta ggc gat ggc tcc ttc cag ctt acc gcg cag gaa gtg gct cag atg 1344
Val Gly Asp Gly Ser Phe Gln Leu Thr Ala Gln Glu Val Ala Gln Met
435                      440                      445

gtg cgc tac gaa ctg ccc gtc att atc ttt ctg atc aac aac cgt ggc 1392
Val Arg Tyr Glu Leu Pro Val Ile Ile Phe Leu Ile Asn Asn Arg Gly
450                      455                      460

tat gtc att gaa atc gcc att cat gac ggc ccg tac aac tat atc aag 1440
Tyr Val Ile Glu Ile Ala Ile His Asp Gly Pro Tyr Asn Tyr Ile Lys
465                      470                      475                      480

aac tgg gat tac gcc ggc ctg atg gaa gtc ttc aac gcc gga gaa ggc 1488
Asn Trp Asp Tyr Ala Gly Leu Met Glu Val Phe Asn Ala Gly Glu Gly
485                      490                      495

cat gga ctt ggc ctg aaa gcc acc acc ccg aag gaa ctg aca gaa gcc 1536
His Gly Leu Gly Leu Lys Ala Thr Thr Pro Lys Glu Leu Thr Glu Ala
500                      505                      510

atc gcc agg gca aaa gcc aat acc cgc ggc ccg acg ctg atc gaa tgc 1584
Ile Ala Arg Ala Lys Ala Asn Thr Arg Gly Pro Thr Leu Ile Glu Cys
515                      520                      525

cag atc gac cgc acg gac tgc acg gat atg ctg gtt caa tgg ggc cgc 1632
Gln Ile Asp Arg Thr Asp Cys Thr Asp Met Leu Val Gln Trp Gly Arg
530                      535                      540

aag gtt gcc tca acc aac gcg cgc aag acc act ctg gcc tga 1674
Lys Val Ala Ser Thr Asn Ala Arg Lys Thr Thr Leu Ala
545                      550                      555

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<210> 25

<211> 557

<212> PRT

<213> Acetobacter pasteurianus

<400> 25

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 Gly Leu Lys His His Phe Ala Val Gly Gly Asp Tyr Asn Leu Val Leu
 20 25 30
 Leu Asp Gln Leu Leu Leu Asn Lys Asp Met Lys Gln Ile Tyr Cys Cys
 35 40 45
 Asn Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ser Asn
 50 55 60
 Gly Ala Ala Ala Ala Val Val Thr Phe Ser Val Gly Ala Ile Ser Ala
 65 70 75 80
 Met Asn Ala Leu Gly Gly Ala Tyr Ala Glu Asn Leu Pro Val Ile Leu
 85 90 95
 Ile Ser Gly Ala Pro Asn Ser Asn Asp Gln Gly Thr Gly His Ile Leu
 100 105 110
 His His Thr Ile Gly Lys Thr Asp Tyr Ser Tyr Gln Leu Glu Met Ala
 115 120 125
 Arg Gln Val Thr Cys Ala Ala Glu Ser Ile Thr Asp Ala His Ser Ala
 130 135 140
 Pro Ala Lys Ile Asp His Val Ile Arg Thr Ala Leu Arg Glu Arg Lys
 145 150 155 160
 Pro Ala Tyr Leu Asp Ile Ala Cys Asn Ile Ala Ser Glu Pro Cys Val
 165 170 175
 Arg Pro Gly Pro Val Ser Ser Leu Leu Ser Glu Pro Glu Ile Asp His
 180 185 190
 Thr Ser Leu Lys Ala Ala Val Asp Ala Thr Val Ala Leu Leu Lys Asn
 195 200 205

Arg Pro Ala Pro Val Met Leu Leu Gly Ser Lys Leu Arg Ala Ala Asn
 210 215 220
 Ala Leu Ala Ala Thr Glu Thr Leu Ala Asp Lys Leu Gln Cys Ala Val
 225 230 235 240
 Thr Ile Met Ala Ala Ala Lys Gly Phe Phe Pro Glu Asp His Ala Gly
 245 250 255
 Phe Arg Gly Leu Tyr Trp Gly Glu Val Ser Asn Pro Gly Val Gln Glu
 260 265 270
 Leu Val Glu Thr Ser Asp Ala Leu Leu Cys Ile Ala Pro Val Phe Asn
 275 280 285
 Asp Tyr Ser Thr Val Gly Trp Ser Gly Met Pro Lys Gly Pro Asn Val
 290 295 300
 Ile Leu Ala Glu Pro Asp Arg Val Thr Val Asp Gly Arg Ala Tyr Asp
 305 310 315 320
 Gly Phe Thr Leu Arg Ala Phe Leu Gln Ala Leu Ala Glu Lys Ala Pro
 325 330 335
 Ala Arg Pro Ala Ser Ala Gln Lys Ser Ser Val Pro Thr Cys Ser Leu
 340 345 350
 Thr Ala Thr Ser Asp Glu Ala Gly Leu Thr Asn Asp Glu Ile Val Arg
 355 360 365
 His Ile Asn Ala Leu Leu Thr Ser Asn Thr Thr Leu Val Ala Glu Thr
 370 375 380
 Gly Asp Ser Trp Phe Asn Ala Met Arg Met Thr Leu Ala Gly Ala Arg
 385 390 395 400
 Val Glu Leu Glu Met Gln Trp Gly His Ile Gly Trp Ser Val Pro Ser
 405 410 415
 Ala Phe Gly Asn Ala Met Gly Ser Gln Asp Arg Gln His Val Val Met
 420 425 430
 Val Gly Asp Gly Ser Phe Gln Leu Thr Ala Gln Glu Val Ala Gln Met
 435 440 445
 Val Arg Tyr Glu Leu Pro Val Ile Ile Phe Leu Ile Asn Asn Arg Gly
 450 455 460
 Tyr Val Ile Glu Ile Ala Ile His Asp Gly Pro Tyr Asn Tyr Ile Lys
 465 470 475 480
 Asn Trp Asp Tyr Ala Gly Leu Met Glu Val Phe Asn Ala Gly Glu Gly
 485 490 495
 His Gly Leu Gly Leu Lys Ala Thr Thr Pro Lys Glu Leu Thr Glu Ala
 500 505 510
 Ile Ala Arg Ala Lys Ala Asn Thr Arg Gly Pro Thr Leu Ile Glu Cys
 515 520 525
 Gln Ile Asp Arg Thr Asp Cys Thr Asp Met Leu Val Gln Trp Gly Arg
 530 535 540
 Lys Val Ala Ser Thr Asn Ala Arg Lys Thr Thr Leu Ala
 545 550 555

<210> 26

<211> 32
 <212> DNA
 <213> artificial sequence

<220>
 <223> artificial sequence

<400> 26
 atcttaatta atgtataccg ttggtatgta ct 32

<210> 27
 <211> 34
 <212> DNA
 <213> artificial sequence

<220>
 <223> artificial sequence

<400> 27
 tatgcggccg cttacgcttg tggttgcga gagt 34

<210> 28
 <211> 1671
 <212> DNA
 <213> Zymobacter palmae

<220>
 <221> CDS
 <222> (1)..(1671)
 <223>

<400> 28
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 Met Tyr Thr Val Gly Met Tyr Leu Ala Glu Arg Leu Ala Gln Ile Gly
 1 5 10 15
 ctg aaa cac cac ttt gcc gtg gcc ggt gac tac aac ctg gtg ttg ctt 96
 Leu Lys His His Phe Ala Val Ala Gly Asp Tyr Asn Leu Val Leu Leu
 20 25 30
 gat cag ctc ctg ctg aac aaa gac atg gag cag gtc tac tgc tgt aac 144
 Asp Gln Leu Leu Leu Asn Lys Asp Met Glu Gln Val Tyr Cys Cys Asn
 35 40 45
 gaa ctt aac tgc ggc ttt agc gcc gaa ggt tac gct cgt gca cgt ggt 192
 Glu Leu Asn Cys Gly Phe Ser Ala Glu Gly Tyr Ala Arg Ala Arg Gly
 50 55 60
 gcc gcc gct gcc atc gtc acg ttc agc gta ggt gct atc tct gca atg 240
 Ala Ala Ala Ala Ile Val Thr Phe Ser Val Gly Ala Ile Ser Ala Met
 65 70 75 80
 aac gcc atc ggt ggc gcc tat gca gaa aac ctg ccg gtc atc ctg atc 288
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REFERENCES CITED IN THE DESCRIPTION

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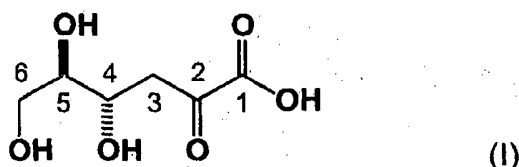
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PATENTKRAV

1. Fremgangsmåde til fremstilling af 2'-desoxynucleosider eller 2'-desoxynucleosid-forstadier ud fra en forbindelse med formlen I eller dens salte

5



eller en beskyttet form deraf ved en fremgangsmåde, der omfatter et decarboxylerings-trin.

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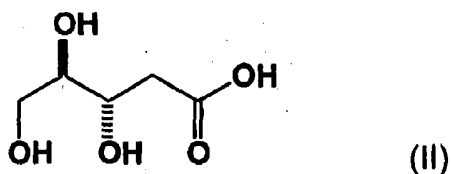
2. Fremgangsmåde ifølge krav 1, ved hvilken decarboxyleringstrinnet spalter C1-C2-bindingen i forbindelsen med formlen I eller dens salte eller en beskyttet form deraf.

3. Fremgangsmåde ifølge krav 1 eller 2, ved hvilken decarboxyleringstrinnet udføres direkte på forbindelsen med formlen I eller dens salte eller en beskyttet form deraf.

15

4. Fremgangsmåde ifølge et hvilket som helst af kravene 1 til 3, ved hvilken decarboxyleringstrinnet foregår ved omsætning af forbindelsen med formlen I eller dens salte eller en beskyttet form deraf med hydrogenperoxid til fremstilling af en forbindelse med formlen II eller dens salte

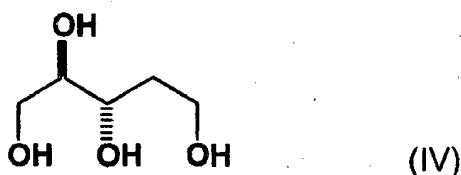
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eller en beskyttet form deraf som 2'-desoxynucleosidforstadium.

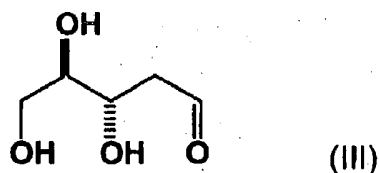
25

5. Fremgangsmåde ifølge krav 4, som endvidere omfatter omdannelse af forbindelsen med formlen II eller dens salte eller en beskyttet form deraf til en forbindelse med formlen IV



eller en beskyttet form deraf som 2'-desoxynucleosidforstadium.

- 5 6. Fremgangsmåde ifølge krav 4, som endvidere omfatter omdannelse af forbindelsen med formlen II eller dens salte eller en beskyttet form deraf til en forbindelse med formlen III

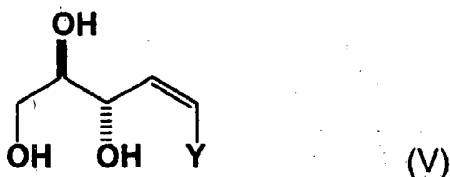


10

eller en beskyttet form deraf som 2'-desoxynucleosidforstadium.

7. Fremgangsmåde ifølge krav 6 omfattende omdannelse af forbindelsen med formlen II eller dens salte eller en beskyttet form deraf til forbindelsen med formlen IV eller en
- 15 beskyttet form deraf som mellemprodukt, som derpå omdannes til forbindelsen med formlen III eller en beskyttet form deraf.

8. Fremgangsmåde ifølge et hvilket som helst af kravene 1 til 3, ved hvilken decarboxyleringstrinnet foregår ved omsætning af forbindelsen med formlen I eller dens
- 20 salte eller en beskyttet form deraf med en amin Y-H, hvori H betyder et til nitrogenatomet i aminogruppen bundet hydrogenatom, til fremstilling af en forbindelse med formlen V

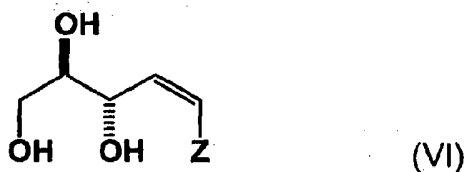


- eller dens respektive trans-isomere eller en beskyttet form deraf som 2'-desoxy-
- 25 nucleosidforstadium.

9. Fremgangsmåde ifølge krav 8, ved hvilken Y-H betyder en ligekædet eller cyclisk sekundær amin.

5 10. Fremgangsmåde ifølge krav 8 eller 9, ved hvilken Y-H er morpholin, pyrrolidin, piperidin, N-methylpiperazin eller diethylamin.

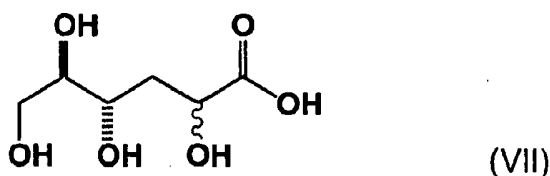
11. Fremgangsmåde ifølge et hvilket som helst af kravene 8 til 10, som endvidere omfatter det trin at omsætte en forbindelse med formlen V eller dens trans-isomere eller
10 en beskyttet form deraf med Z-H, hvori H betyder et hydrogenatom, og Z betyder en fraspaltelig gruppe, til fremstilling af en forbindelse med formlen VI



15 eller dens respektive trans-isomere eller en beskyttet form deraf som 2'-desoxy-nucleosidforstadium.

12. Fremgangsmåde ifølge krav 11, ved hvilken Z-H er vand, til fremstilling af en forbindelse med formlen III eller en beskyttet form deraf som 2'-desoxynucleosid-
20 forstadium.

13. Fremgangsmåde ifølge krav 1 eller 2, ved hvilken forbindelsen med formlen I eller dens salte eller en beskyttet form deraf omdannes til en forbindelse med formlen VII eller dens salte eller en beskyttet form deraf eller en blanding af de respektive epimere
25



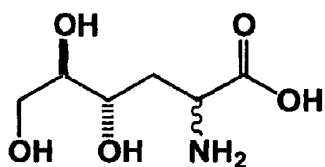
som derpå decarboxyleres til fremstilling af en forbindelse med formlen III eller en beskyttet form deraf som 2'-desoxynucleosidforstadium.

14. Fremgangsmåde ifølge krav 13, ved hvilken omdannelsen af I eller dens salte eller en beskyttet form deraf til VII eller en beskyttet form deraf foregår ved reduktion med natriumborhydrid eller ved hydrogenering med Raney-nikkel eller en platinoxid-

5 katalysator.

15. Fremgangsmåde ifølge krav 13 eller 14, ved hvilken decarboxyleringstrinnet foregår ved omsætning med hydrogenperoxid.

10 16. Fremgangsmåde ifølge krav 1 eller 2, ved hvilken forbindelsen med formelen I eller dens salte eller en beskyttet form deraf omdannes til en forbindelse med formelen VIII eller dens salte eller en beskyttet form deraf eller en blanding af de respektive epimere



(VIII)

15

som derpå decarboxyleres til fremstilling af en forbindelse med formelen III eller en beskyttet form deraf som 2'-desoxynucleosidforstadium.

17. Fremgangsmåde ifølge krav 16, ved hvilken en forbindelse med formelen VIII eller en

20 beskyttet form deraf eller en blanding af de respektive epimere omsættes med ninhydrin, hvilket fører til forbindelsen med formelen III eller en beskyttet form deraf.

18. Fremgangsmåde ifølge krav 16 eller 17, ved hvilken omdannelsen af I eller dens salte eller en beskyttet form deraf til VIII eller en beskyttet form deraf foregår ved reduktiv

25 aminering med ammoniak og natriumcyanoborhydrid.

19. Fremgangsmåde ifølge et hvilket som helst af kravene 1 til 18, ved hvilken beskyttelsesgruppen eller -grupperne indbyrdes uafhængigt er valgt blandt acetatester, benzoatester, allylether, benzylether, tritylether, tert.butyltrimethylsilyl-(TBDMS)-ether,

30 isopropyliden eller en benzylidenacetal.

20. Fremgangsmåde ifølge et hvilket som helst af kravene 1 til 3, ved hvilken decarboxyleringstrinnet udføres ved en enzymatisk reaktion, der omfatter et enkelt trin.

21. Fremgangsmåde ifølge krav 20, ved hvilken den enzymatiske reaktion katalyseres af et enzym med ketosyredecarboxylaseaktivitet.

5 22. Fremgangsmåde ifølge krav 21, ved hvilken det enzym, der har ketosyre-decarboxylaseaktivitet, er en thiaminpyrophosphat-(TPP)-afhængig ketosyre-decarboxylase.

23. Fremgangsmåde ifølge krav 22, ved hvilken den TPP-afhængige ketosyre-decarboxylase er en pyruvatdecarboxylase (EC 4.1.1.1), en benzoylformiat-decarboxylase (EC 4.1.1.7), en indolpyruvatdecarboxylase (EC 4.1.1.74), en phosphonopyruvatdecarboxylase, en sulfopyruvatdecarboxylase (EC 4.1.1.79), en oxalyl-coenzym A-decarboxylase (EC 4.1.1.8), en oxoglutaratdecarboxylase (EC 4.1.1.71) eller en phenylpyruvatdecarboxylase (EC 4.1.1.43).

15

24. Fremgangsmåde ifølge krav 23, ved hvilken pyruvatdecarboxylasen er af eukaryot oprindelse.

25. Fremgangsmåde ifølge krav 24, ved hvilken den eukaryote organisme er en
20 gærorganisme.

26. Fremgangsmåde ifølge krav 25, ved hvilken gæren er *Saccharomyces cerevisiae*.

27. Fremgangsmåde ifølge krav 23, ved hvilken pyruvatdecarboxylasen er af prokaryot
25 oprindelse.

28. Fremgangsmåde ifølge krav 27, ved hvilken den prokaryote organisme er af slægten *Zymomonas*, *Zymobacter* eller *Acetobacter*.

30 29. Fremgangsmåde ifølge krav 28, ved hvilken organismen er af arten *Zymomonas mobilis*, *Zymobacter palmae* eller *Acetobacter pasteurianus*.

30. Fremgangsmåde ifølge krav 23, ved hvilken benzoylformiatdecarboxylasen er af prokaryot oprindelse.

35

31. Fremgangsmåde ifølge krav 30, ved hvilken den prokaryote organisme er af slægten *Pseudomonas*.
32. Fremgangsmåde ifølge krav 31, ved hvilken organismen er af arten *Pseudomonas putida*.
33. Fremgangsmåde ifølge et hvilket som helst af kravene 20 til 32, ved hvilken pH-værdien ved tilsætning af en syre reguleres til en værdi mellem pH 5 og pH 9.
34. Fremgangsmåde ifølge krav 33, ved hvilken pH-værdien reguleres til en værdi mellem pH 6 og pH 8.
35. Fremgangsmåde ifølge krav 33 eller 34, ved hvilken syren er HCl, H₂SO₄, D-gluconsyre eller 2-dehydro-3-desoxy-D-gluconsyre.
36. Fremgangsmåde ifølge et hvilket som helst af kravene 1 til 35 omfattende det forberedende trin at fremstille forbindelsen med formelen I ud fra D-gluconat eller et D-gluconatsalt ved anvendelse af en gluconatdehydrataseaktivitet.
37. Fremgangsmåde ifølge krav 36, ved hvilken D-gluconatsaltet er kalium- eller natrium-D-gluconat.
38. Fremgangsmåde ifølge krav 36 eller 37, ved hvilken der kodes for gluconatdehydratasen af et polynucleotid, der omfatter en nucleotidsekvens valgt fra gruppen bestående af:
- (a) nucleotidsekvenser, der koder for et polypeptid, som omfatter aminosyresekvensen SEQ ID NO: 2;
 - (b) nucleotidsekvenser, der omfatter den kodende sekvens SEQ ID NO: 1;
 - (c) nucleotidsekvenser, der koder for et fragment, der kodes for af en nucleotidsekvens ifølge (a) eller (b);
 - (d) nucleotidsekvenser, der hybridiserer med en nucleotidsekvens ifølge en hvilket som helst af (a) til (c); og
 - (e) nucleotidsekvenser, der afviger fra nucleotidsekvensen ifølge (d) som følge af den genetiske kodes degeneration.

39. Fremgangsmåde ifølge et hvilket som helst af kravene 1 til 35 omfattende det forberedende trin at fremstille forbindelsen med formlen I ud fra D-glucosaminat ved anvendelse af en glucosaminatdeaminaseaktivitet.

- 5 40. Fremgangsmåde ifølge krav 39, ved hvilken der kodes for glucosamindeaminasen af et polynucleotid, der omfatter en nucleotidsekvens valgt fra gruppen bestående af:
- (a) nucleotidsekvenser, der koder for et polypeptid, som omfatter aminosyresekvensen SEQ ID NO: 4;
 - (b) nucleotidsekvenser, der omfatter den kodende sekvens SEQ ID NO: 3;
 - 10 (c) nucleotidsekvenser, der koder for et fragment, der kodes for af en nucleotidsekvens ifølge (a) eller (b);
 - (d) nucleotidsekvenser, der hybridiserer med en nucleotidsekvens ifølge en hvilket som helst af (a) til (c); og
 - (e) nucleotidsekvenser, der afviger fra nucleotidsekvensen ifølge (d) som følge af
 - 15 den genetiske kodes degeneration.

41. Organisme, der er i stand til enzymatisk at omdanne D-gluconat til 2-dehydro-3-desoxy-D-gluconat som følge af ekspression af en D-gluconatdehydratase og/eller er i stand til enzymatisk at omdanne D-glucosaminat til 2-dehydro-3-desoxy-D-gluconat

20 som følge af ekspression af en D-glucosaminatdeaminase, og som er i stand til enzymatisk at omdanne 2-dehydro-3-desoxy-D-gluconat til 2-desoxy-D-ribose ved decarboxylering som følge af ekspression af en pyruvatdecarboxylase eller en benzoylformiatdecarboxylase, idet en sådan organisme er en svampecelle eller en bakteriecelle.

25 42. Organisme ifølge krav 41, som er en organisme, der endogent udtrykker en D-gluconatdehydratase eller en D-glucosaminatdeaminase, og i hvilken der er blevet indført et fremmed nucleinsyremolekyle, som koder for en pyruvatdecarboxylase eller en benzoylformiatdecarboxylase.

30 43. Mikroorganisme ifølge krav 41, som naturligt ikke udtrykker en D-gluconatdehydratase, en D-glucosaminatdeaminase og en pyruvatdecarboxylase eller en benzoylformiatdecarboxylase, og i hvilken der er blevet indført fremmede nucleinsyremolekyler, som koder for en D-gluconatdehydratase eller en D-glucosaminatdeaminase

35 eller begge, og et nucleinsyremolekyle, der koder for en pyruvatdecarboxylase eller en benzoylformiatdecarboxylase.

44. Organisme ifølge krav 41, som er en organisme, der endogent udtrykker en pyruvatdecarboxylase eller en benzoylformiatdecarboxylase, men som ikke naturligt udtrykker en D-gluconatdehydratase eller en D-glucosaminatdeaminase, og i hvilken
5 der er blevet indført et fremmed nucleinsyremolekyle, som koder for en D-gluconatdehydratase og/eller en D-glucosaminatdeaminase.
45. Organisme ifølge krav 42 eller 43, hvori pyruvatdecarboxylasen er af eukaryot oprindelse.
10
46. Organisme ifølge krav 45, hvori pyruvatdecarboxylasen er en decarboxylase fra *Saccharomyces cerevisiae*.
47. Organisme ifølge krav 42 eller 43, hvori pyruvatdecarboxylasen er af prokaryot
15 oprindelse.
48. Organisme ifølge krav 47, hvori pyruvatdecarboxylasen er en decarboxylase fra *Zymomonas mobilis*.
- 20 49. Organisme ifølge et hvilket som helst af kravene 41 til 48, som ikke udtrykker en 2-dehydro-3-desoxy-D-gluconatkinaseaktivitet.
50. Organisme ifølge et hvilket som helst af kravene 41 til 49, som ikke udtrykker en 2-dehydro-3-desoxy-D-gluconataldolaseaktivitet.
25
51. Organisme ifølge et hvilket som helst af kravene 41 til 50, som ikke udtrykker en 2-desoxy-D-ribosealdolaseaktivitet.
52. Fremgangsmåde ifølge et hvilket som helst af kravene 20 til 40, som udføres ved
30 anvendelse af en organisme ifølge et hvilket som helst af kravene 41 til 51.
53. Anvendelse af et polynucleotid ifølge krav 38 eller af en gluconatdehydratase, der kodes for af et sådant polynucleotid, ved en fremgangsmåde ifølge krav 36 eller 37.
- 35 54. Anvendelse af et polynucleotid ifølge krav 40 eller af en glucosaminatdeaminase, der kodes for af et sådant polynucleotid, ved en fremgangsmåde ifølge krav 39.

55. Anvendelse af et enzym med ketosyredecarboxylaseaktivitet eller af et polynucleotid, der koder for et sådant enzym, ved en fremgangsmåde til omdannelse af en forbindelse med formlen I til 2-desoxy-D-ribose.