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(12) **United States Patent**
O'Keefe

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(45) **Date of Patent:** **Apr. 2, 2019**

(54) **METHOD FOR RAPIDLY DETERMINING EFFECTIVE STERILIZATION, DEIMMUNIZATION, AND/OR DISINFECTION**

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(65) **Prior Publication Data**

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Related U.S. Application Data

(60) Provisional application No. 62/314,617, filed on Mar. 29, 2016.

(51) **Int. Cl.**
C12Q 1/22 (2006.01)

(52) **U.S. Cl.**
CPC **C12Q 1/22** (2013.01)

(58) **Field of Classification Search**
None
See application file for complete search history.

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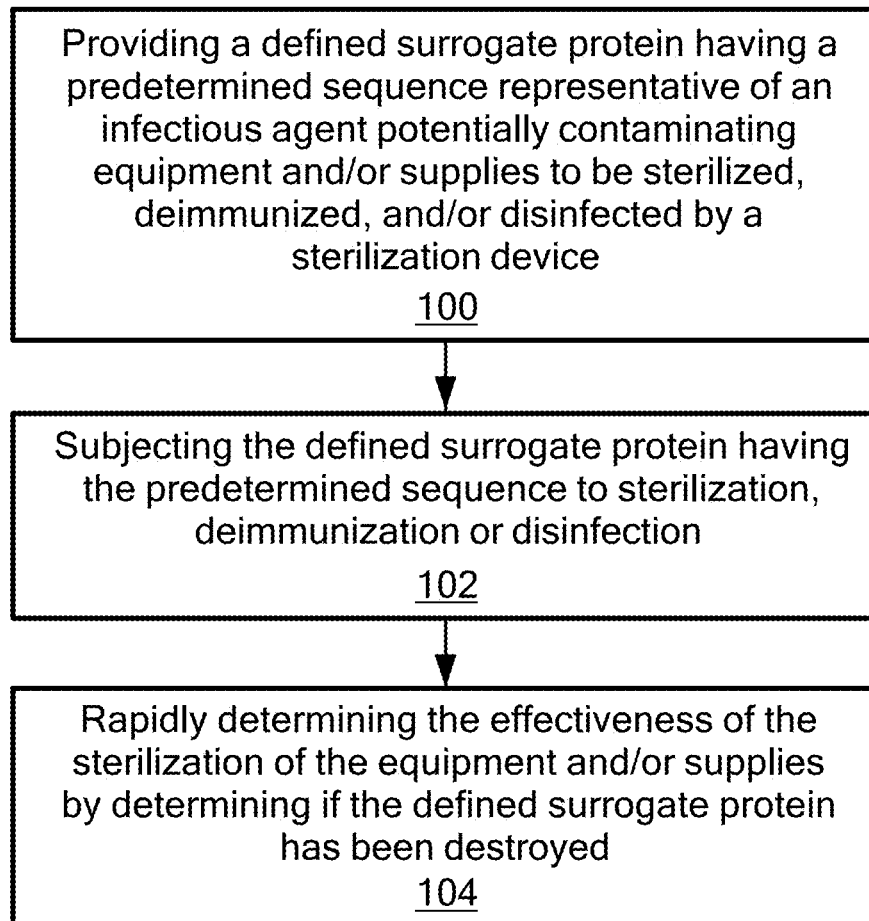
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(57) **ABSTRACT**

A method for rapidly determining effective sterilization, deimmunization, and/or disinfection of equipment and/or supplies by a device. The method includes providing a defined surrogate protein having a predetermined sequence representative of an infectious agent potentially contaminating the equipment and/or the supplies to be sterilized, deimmunized, and/or disinfected by the device. The defined surrogate protein having the predetermined sequence is subjected to sterilization, deimmunization, or disinfection. The effectiveness of the sterilization deimmunization, or disinfection is rapidly determined by determining if the defined surrogate protein has been destroyed.

15 Claims, 29 Drawing Sheets

Specification includes a Sequence Listing.

***FIG. 1***

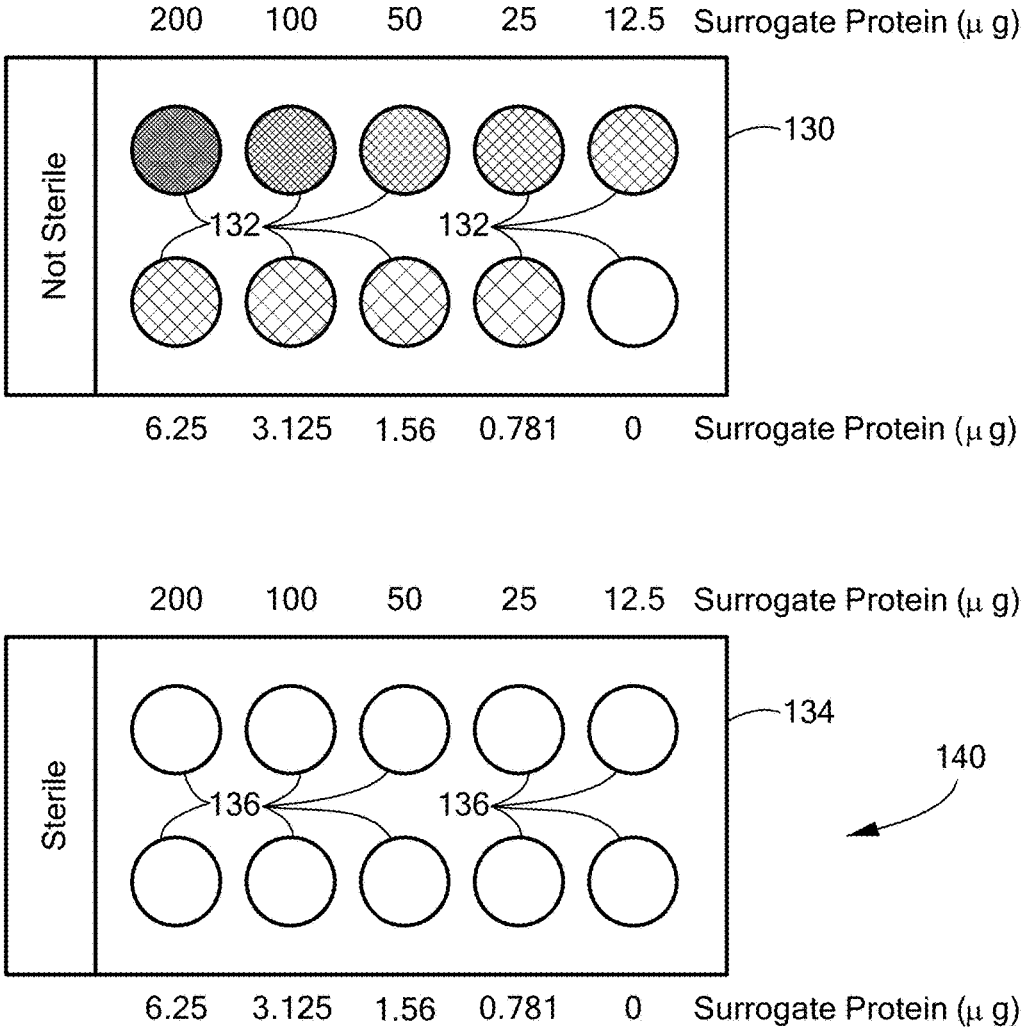
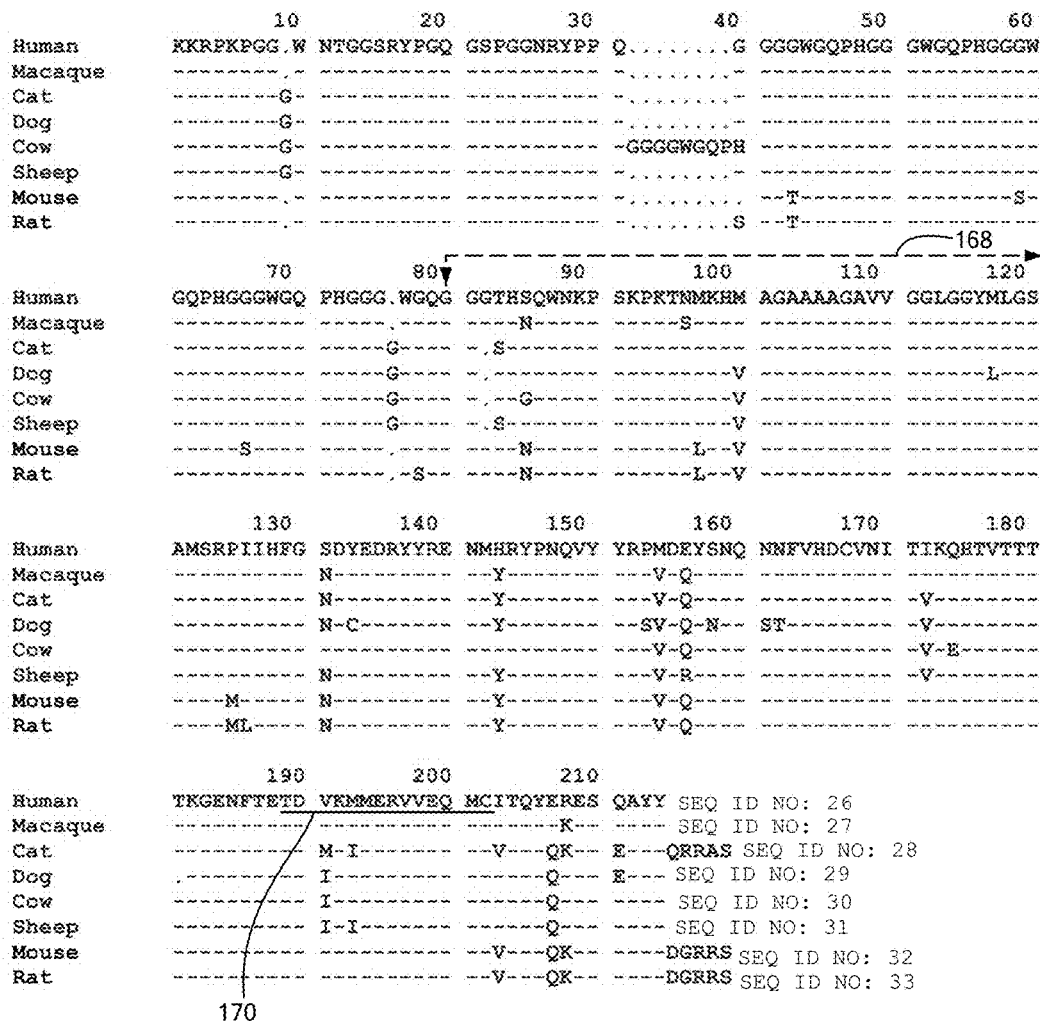
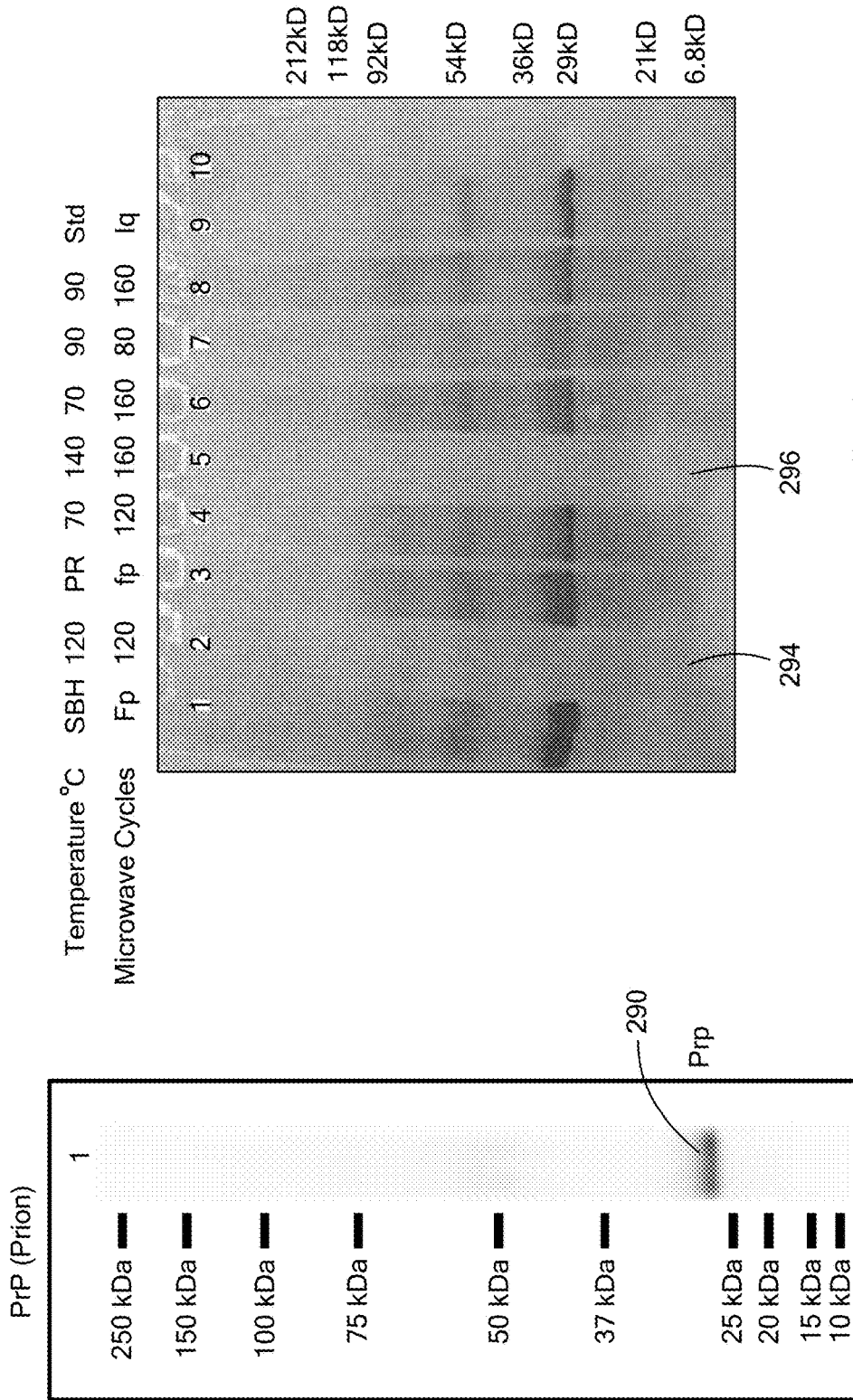


FIG. 2



Amino Acid Sequence Comparison of Human PrP Proteins with a Selection of other Species PrP Proteins. Residues cover only standard mature protein sequences.

FIG. 4



(Clostridium)
 Homology Diagram comparing protein sequence in a research (non-pathogenic) Clostridium species and 2 pathogenic Clostridium species. Standard single letter amino acid abbreviation is used. [-] indicates amino acid homology, [.] indicates gap in sequence.

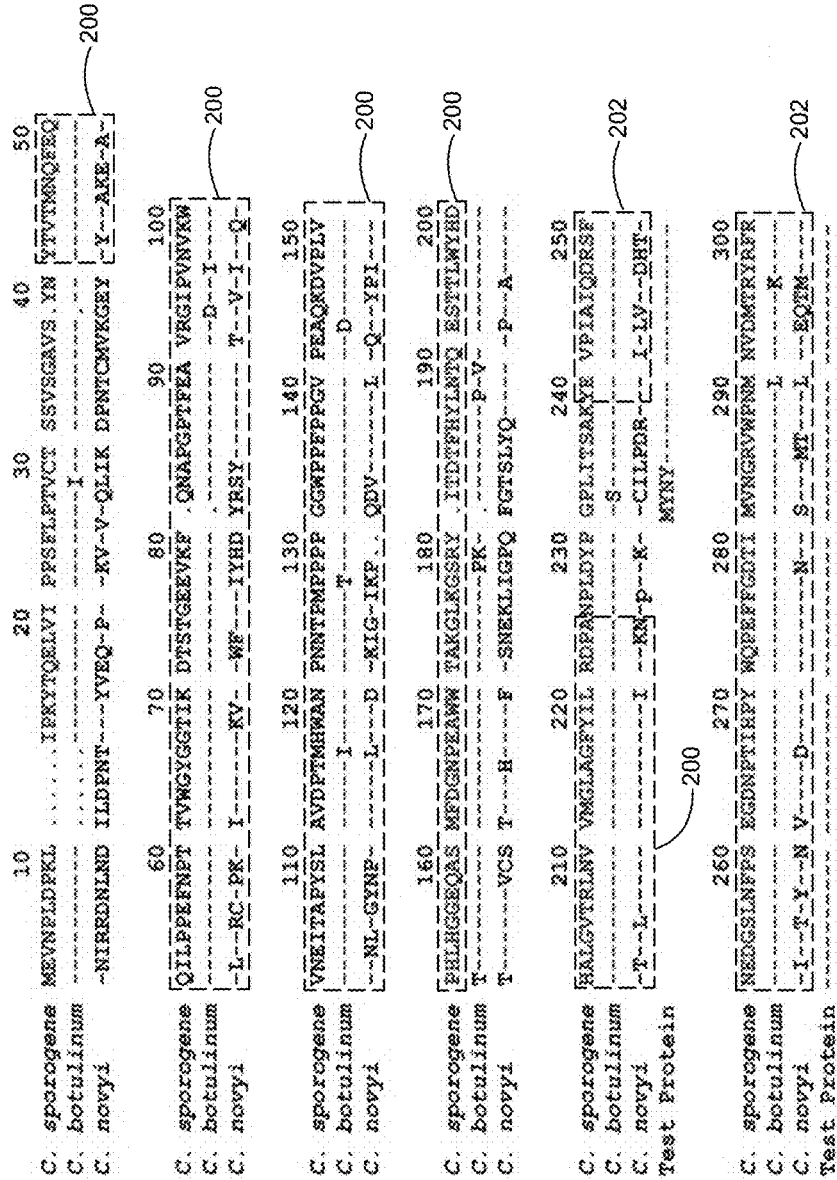


FIG. 7

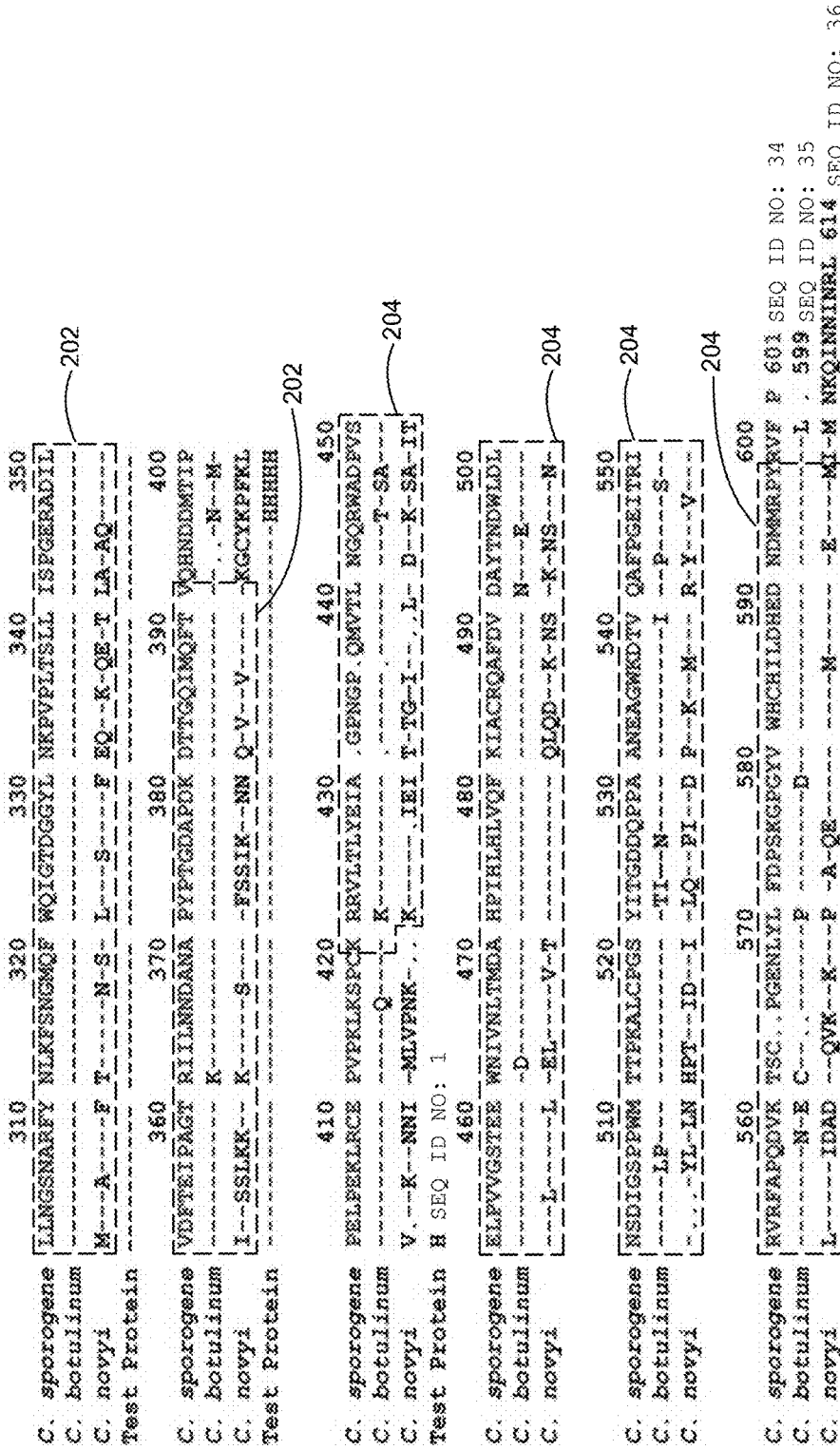


FIG. 7 Continued

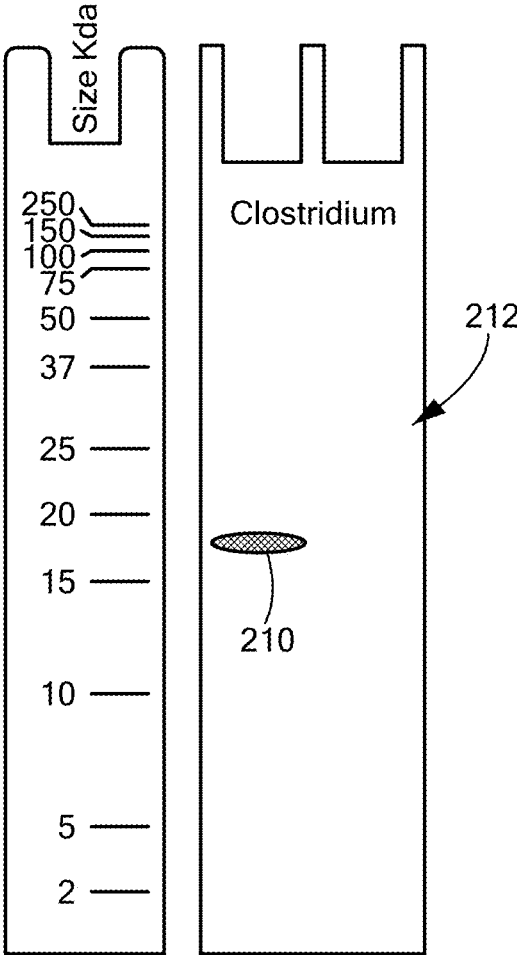


FIG. 8

(Bacillus)

Homology Diagram comparing protein sequence of three Bacillus species that are used in approved sterilization equipment qualification. Standard single letter amino acid abbreviation is used. [-] indicates amino acid homology, [.] indicates gap in sequence.

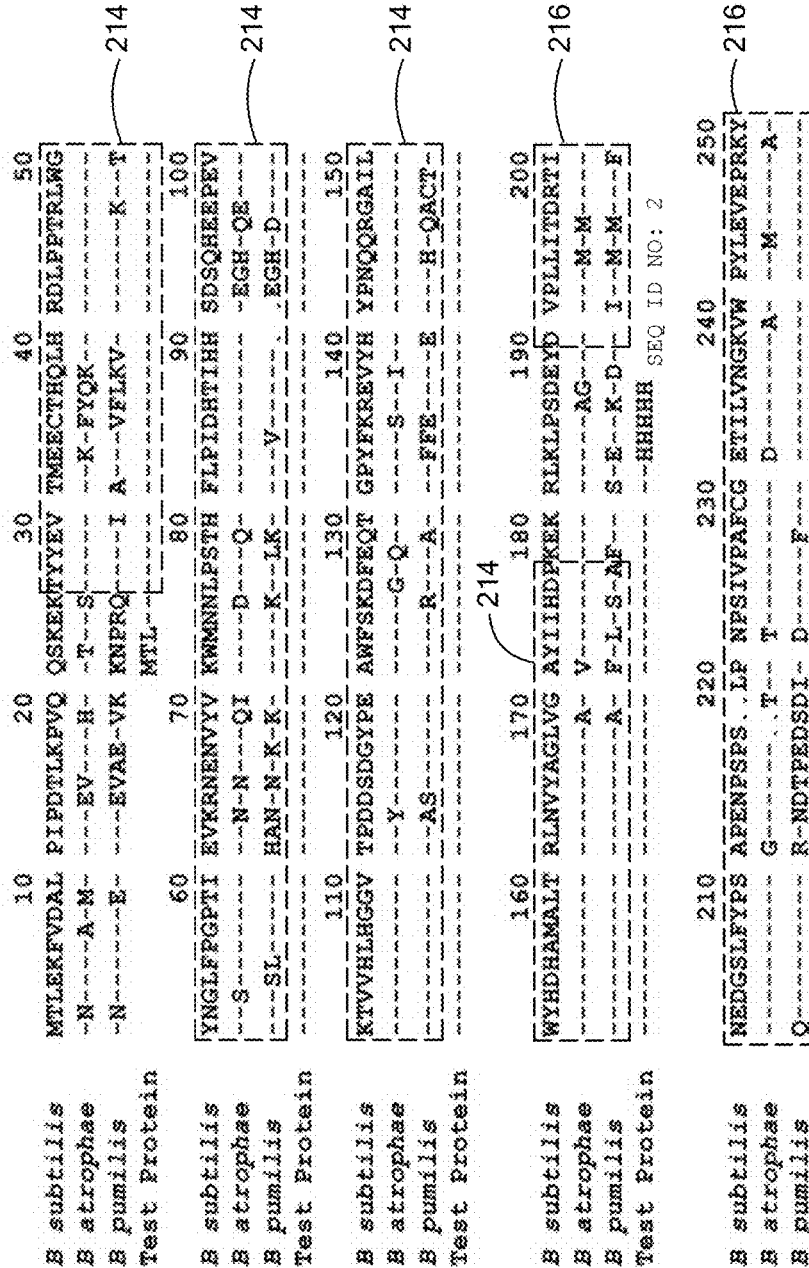


FIG. 9

	260	270	280	290	300	
B subtilis	RFRVINASNT	RTYNLSLNDG	GDFIQIGSDG	GILPERSVKLN	SFSLAPAERY	
B atrophae	---IV---	---E-L-V---	---	---S-I---	---F---	216
B pumilis	---IL---	---E-H---D	ATIM---	---F---P-RHQ	---I---	216
	310	320	330	340	350	
B subtilis	DIIDFTAYE	GESILLANSA	GCGGVNPFET	DANIMQFRVT	KPLAQKTKAE	
B atrophae	---A-F---	---Q---V---	---A---S---	---V---	---KE-D---	
B pumilis	---V---	---S---NKT-T-K-T	---Q---	---K---	---R---KGRVPKT	216
	360	370	380	390	400	
B subtilis	SRSTSPHT	LR YSMKDINIRT	LKLAGTQDEY	GRPVLLLNK	RWHDFVTEFP	
B atrophae	---KPRFL-NLPPV	TDEKIQ-L---	---	---	---S---	218
B pumilis	.LRPIEKPLPP	PSRADRE---	---T-T---	---K---	---I---D-H F	218
	410	420	430	440	450	
B subtilis	KVGTTEINSY	INRHAHILI	HHLVSRVL	DRRPFDIARY	QESGELSYTV	
B atrophae	---L---S---	---PTRGTHP---	---	---	---T-K- A-TN V-F-G	218
B pumilis	HL-SL-V---	V-PTRGTHP---	---Q---	---I---	---TEV- -ST-IV--G	218
	460	470	480	490	500	
B subtilis	RCPAAA	SEKG WKDTIQAHAG	EVLRIATFG	PYSGRYVWHC	HILEHEDYDM	
B atrophae	PAVEPPP---	---V-S---	---I-M-K---	---	---	218
B pumilis	PNE-PPLH-Q	Y---	---I-V-R-V---	---	---	218
	510					
B subtilis	MRPMDTDPH	K	511 SEQ ID NO: 37			
B atrophae	---VW---N	Q	513 SEQ ID NO: 38			
B pumilis	---IQ---		510 SEQ ID NO: 39			

FIG. 9 Continued

218

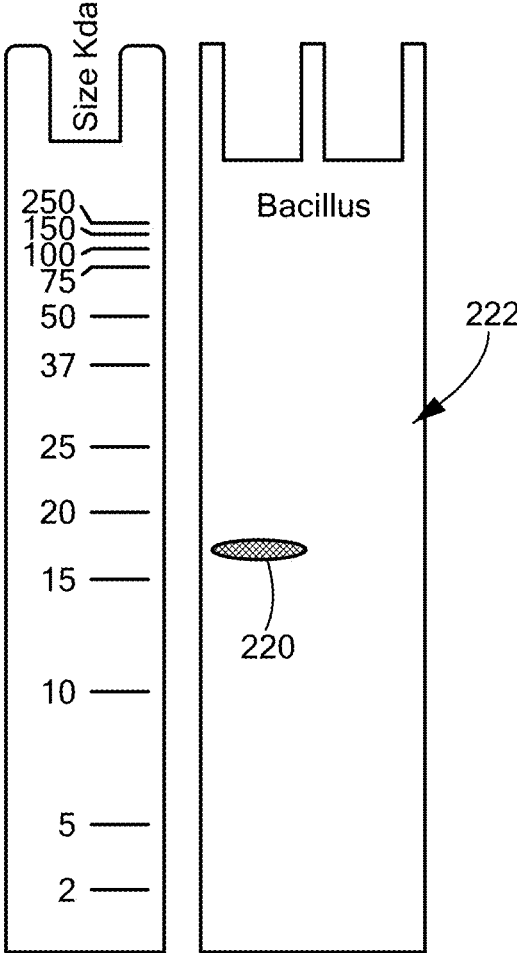


FIG. 10

(Mycobacterium)

Homology Diagram comparing protein sequence in three Mycobacterium species. Standard single letter amino acid abbreviation is used. [-] indicates amino acid homology, [.] indicates gap in sequence.

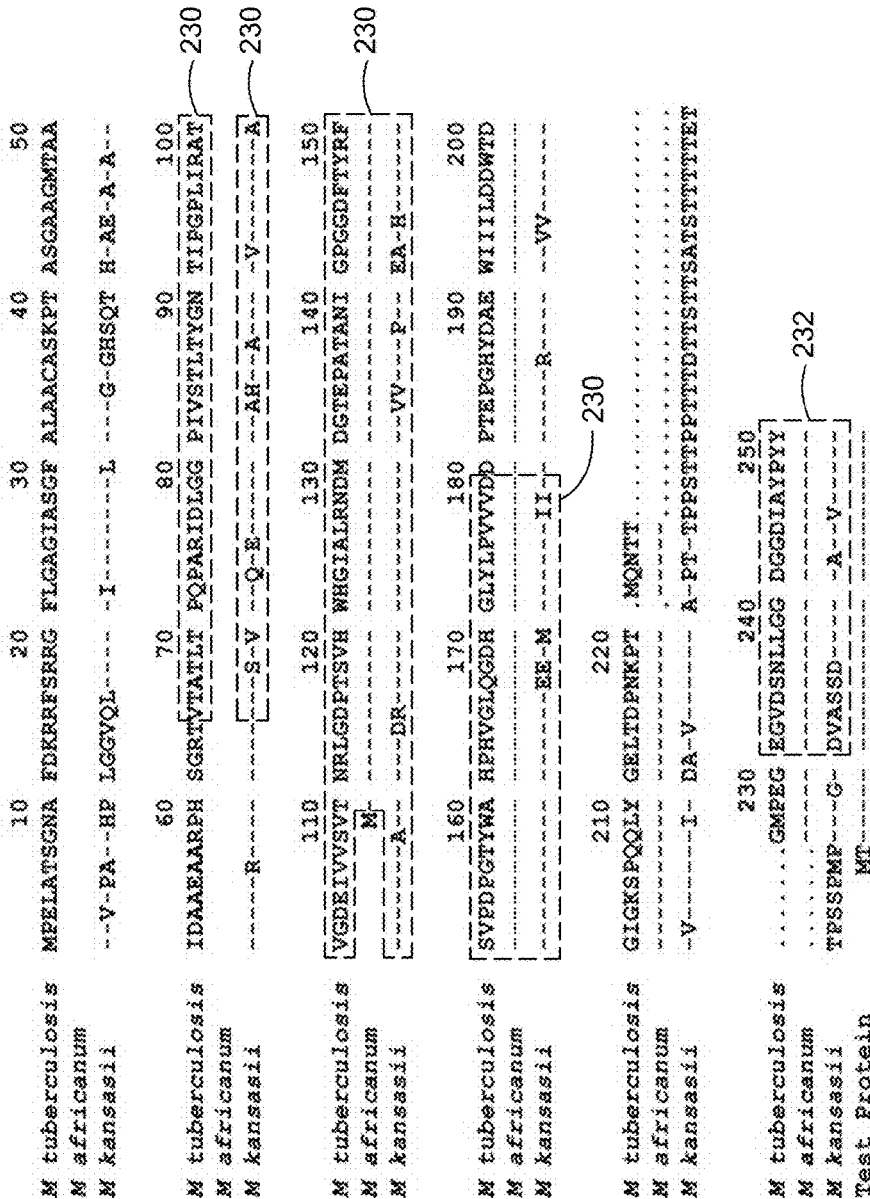


FIG. 11

M tuberculosis	260	270	280	290	300	
M africanum	LINGRIFVAA	TSEKARFQR	IRIRIINSAA	DTAFRIALAG	HSWIVYTHDG	
M kansasii	A-P	T-N	A	V		232
Test Protein						
M tuberculosis	310	320	330	340	350	
M africanum	YEVIPTEVDA	LLICMAERYD	VMVTAAGGVE	FLVALAEGKN	ALARALLSTG	
M kansasii	L-P	G	I-S		V-S	232
Test Protein						
M tuberculosis	360	370	380	390	400	
M africanum	AGSPDPQFR	PDELNWRVGI	VENMFTAANTA	NLGRPEPTHD	LPVTLGGTMA	
M kansasii	A	TKK	I-T-S	A-CLE	V	234
Test Protein	HHH	HH	SEQ ID NO: 3			
M tuberculosis	410	420	430	440	450	
M africanum	KYDWTINGER	YSTINPLHVR	LGQRPTLMFD	NTTMMYHPIH	LHGHTFQMIK	
M kansasii		R-K	QVH Q	QLIV	A-V-W-M	Y-I
Test Protein						
M tuberculosis	460	470	480	490	500	
M africanum	ADGSPGARKD	TVIVLPKQKM	RAVLVADNPG	VVMHCNNY	HQVAGMATRL	
M kansasii	L	V Q	T	M	LA	234
M tuberculosis	DYTL	504 SEQ ID NO: 40				
M africanum		396 SEQ ID NO: 41				
M kansasii	VF	540 SEQ ID NO: 42				

234 FIG. 11 Continued

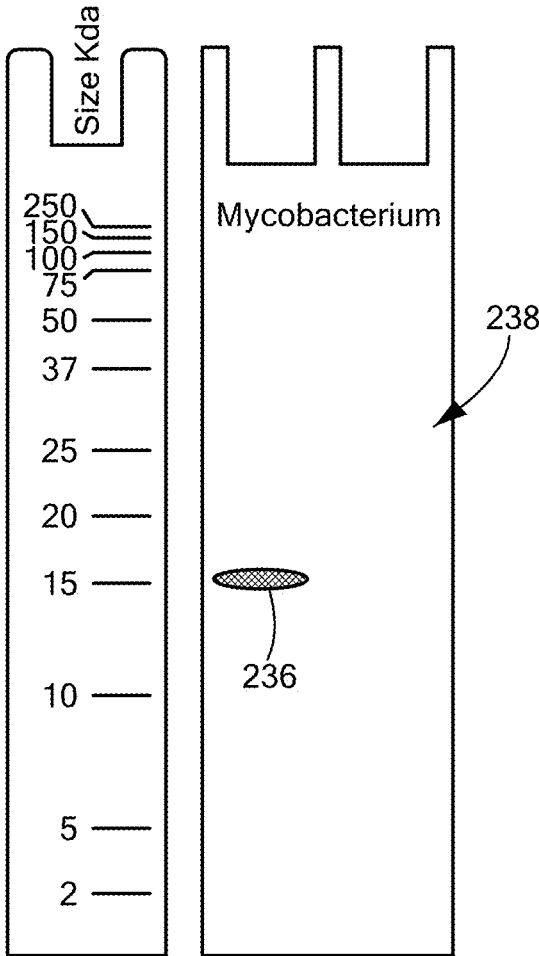


FIG. 12

(Staphylococcus)
 Homology Diagram comparing protein sequence in three Staphylococcus species.
 Standard single letter amino acid abbreviation is used. [-] indicates amino
 acid homology, [.] indicates gap in sequence.

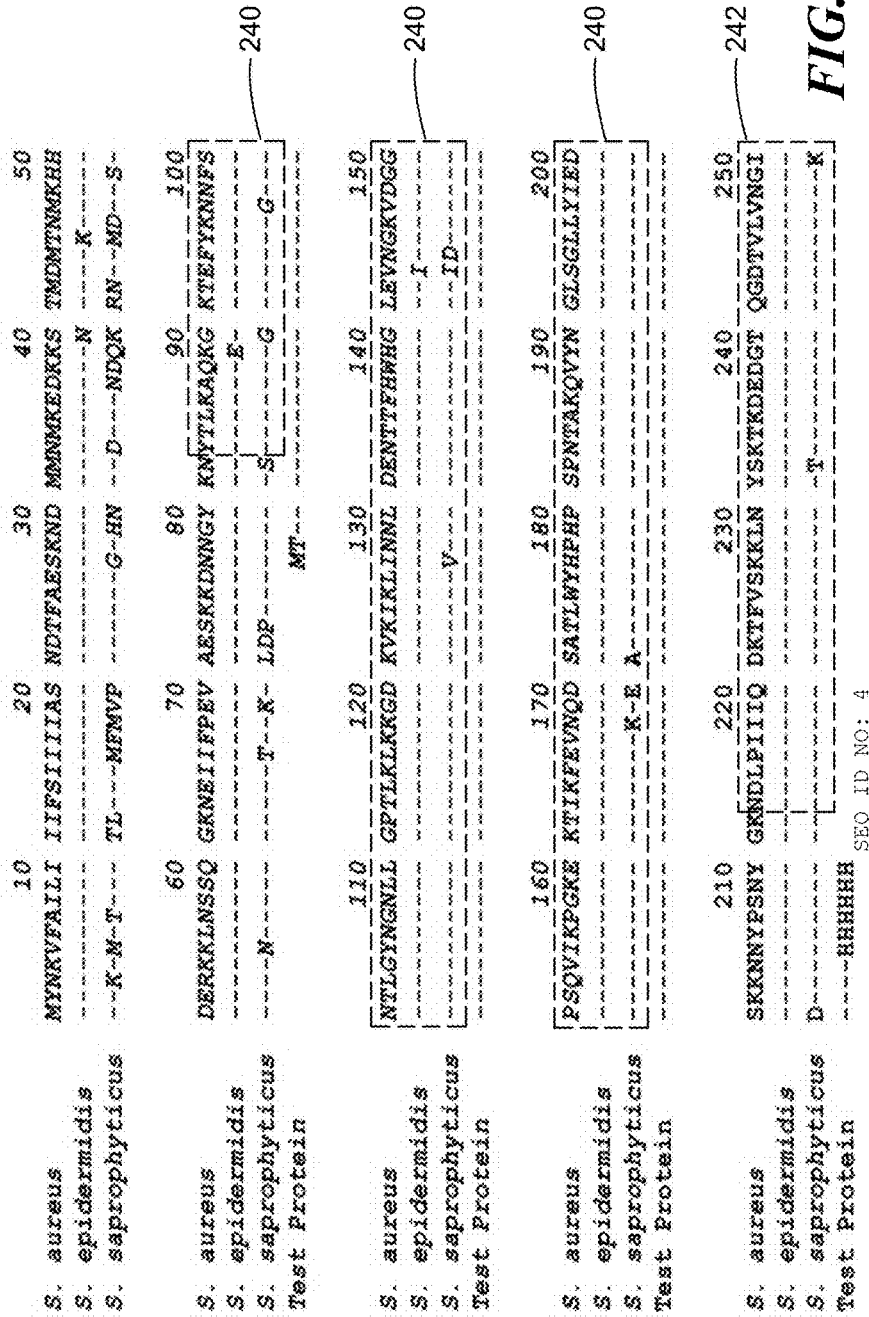


FIG. 13

SEQ ID NO: 4

<i>S. aureus</i>	260	270	280	290	300	
<i>S. epidermidis</i>	VNPKLTAKEE	KIRLRLINGS	NARDINLKLS	NNQSFYIAS	DGGQLKNAKK	
<i>S. saprophyticus</i>	-D--T--G	-----	-----	-----	E--H-EKT--	242
<i>S. aureus</i>	310	320	330	340	350	
<i>S. epidermidis</i>	LKEINLAPSE	RKEIVIDLK	MKGEKISLVD	NDKTVILPIS	NKEKSSNKS	
<i>S. saprophyticus</i>	---A---	---E--VN---	---E---	---I---	---T--DT---	242
<i>S. aureus</i>	360	370	380	390	400	
<i>S. epidermidis</i>	TPKVGKKIKL	EGMNDNVTIN	GNKFDPNRID	FTQKLNQKEV	WEIENVKDKM	
<i>S. saprophyticus</i>	---S---	---H---	---K---	---V-R--T---	-----	246
<i>S. aureus</i>	410	420	430	440	450	
<i>S. epidermidis</i>	GGMKHPFH	GTQFKVLSVD	GEKPPKDMRG	KKDVISLEPG	QKAKIEVVF	
<i>S. saprophyticus</i>	-----	-----	---K--ES---	-----	-----	246
<i>S. aureus</i>	460	470				
<i>S. epidermidis</i>	NTGYMFHCH	ILEHEDNGMM	GQVKVTN	477	SEQ ID NO: 43	
<i>S. saprophyticus</i>	-----	---E---	---I---	477	SEQ ID NO: 44	
	-----	---I---	---K---	477	SEQ ID NO: 45	

FIG. 13 Continued

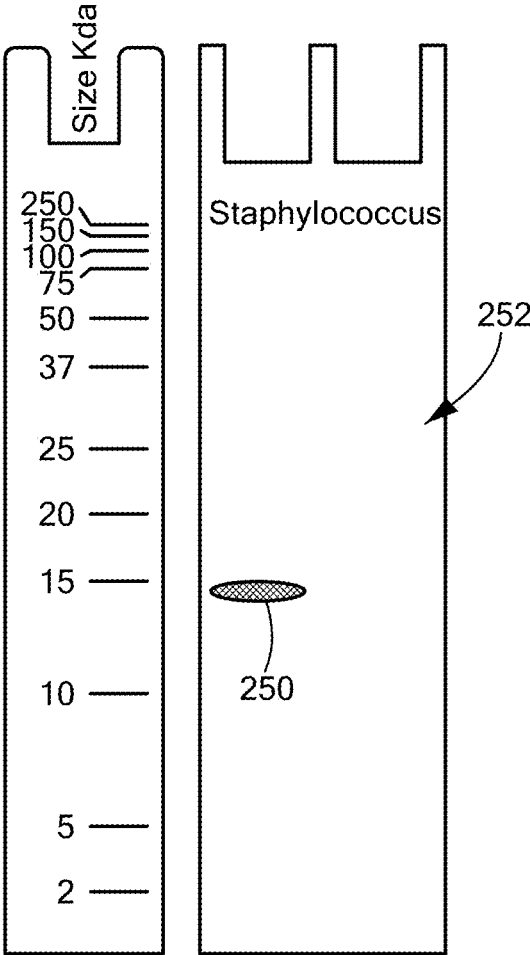


FIG. 14

(Pseudomonas)

Homology Diagram comparing protein sequence in three Pseudomonas species. Standard single letter amino acid abbreviation is used. [-] indicates amino acid homology, [.] indicates gap in sequence.

	10	20	30	40	50
<i>P. aeruginosa</i>	MTFRRQVLG	GLAGLAVVGL	GAGGARL.WLA	RPQ.VAQEYDY	ELIAAPLDLE
<i>P. fluorescens</i>	-S-----I--	-----V---V	-----SRY--G	KMADADAG--	-----V--
<i>P. putida</i>	-S-----M-K	--T--V-----	-----ARY--G	KVEDENAGH--	-----EY--

	60	70	80	90	100
<i>P. aeruginosa</i>	IVPGFSSPAL	AYGGQCPGVE	LRAKQGEWLR	VRFTNRLLDEP	TTIHHHGIRL
<i>P. fluorescens</i>	L---HKTEAW	-F-PSA--T-	--VR-----	---I-H-PVA	-----
<i>P. putida</i>	L---KTEAW	-F-PSA--T-	--VR--T---	---I-H-PVE	-----

	110	120	130	140	150
<i>P. aeruginosa</i>	PIEMDGVPI	SQPFVQPGES	FIYQFKTQDA	GSYWHHELM	SSEQLRGLV
<i>P. fluorescens</i>	-L-----V	--L--L--Y	-D-K-RVP--	-----VS	---E-----
<i>P. putida</i>	-L-----V	--L--K--Y	-D-K-RVP--	-----VS	---E-----

	160	170	180	190	200
<i>P. aeruginosa</i>	GPLIIEEREP	TGERHEKVLG	LKTHVDEQG	AFTFFSVPRQ	AAREGTRGRY
<i>P. fluorescens</i>	-----V-	---KY--T-S	--N--I-DE-	H-VE-----	E---G--A--L
<i>P. putida</i>	-----V-	---Q--RTLS	--N-----	-WL--I--E	---N--A--L

	210	220	230	240	250
<i>P. aeruginosa</i>	STINGKHVPT	IDLPAGQIVR	VRLINVDNTV	TYRLNL.PNCE	ARIYAVDGH
<i>P. fluorescens</i>	-----VPS-V	-E-----T-	-----L--L	-----I--GV-	-Q--L--N--
<i>P. putida</i>	I-----QADSI	TE-----V--	--V--L--W	-----KG-C-	-----L--N--

FIG. 15

	260	270	280	290	300
<i>P. aeruginosa</i>	VEPRGEGQY	WIGPGMRLEL	ALKVPEAGTE	LSLRDGPVRL	ATIRSVASAE
<i>P. fluorescens</i>	---PLGKE--	-L-----IC-	-I-A-P--E-	-----N---	G-L-----NND
<i>P. putida</i>	-T--AL,DE-	-L-----IC-	-IRI-----E-	I-----F---	G-L-----ND
Test Protein	-----	-----	-----	-----	HHH
					262
<i>P. aeruginosa</i>	310	320	330	340	350
<i>P. fluorescens</i>	APAGDWPKPL	PANPVSEPL	ANAEKIGRF	EWVGAMSDTS	GKNPYPSFWQ
<i>P. putida</i>	---T, E---A-	-----A---	---LN-N-	---SV-VNV	DNGKPP-L---
Test Protein	---S, D---PA-	-P--IA-----	ER---LN-N-	---AAGV-VTA	DPAKPS-M---
					264
					HHH SEQ ID NO: 5
	360	370	380	390	400
<i>P. aeruginosa</i>	INGKAWEGGE	EHKHNAPPLA,	KLKEGQSYIF	ELRNMAQYQH	PIHLHGMAFK
<i>P. fluorescens</i>	---VITD	KTCADR-IASL	---	---K--T---	-----S---
<i>P. putida</i>	---Q--DITD	KTCADR-IATL	Q. K-K---	---K--T---	-----S---
					264
	410	420	430	440	450
<i>P. aeruginosa</i>	VILDSRREII	PYFTDTYLLG	KNETARVALV	ADNPGLWMFH	CHVIDHMETG
<i>P. fluorescens</i>	-IA-N-HK--	---	---R-Q---	-----V---	-----
<i>P. putida</i>	-IA-N-HD-K	EPW-----	---R-Q---	-----T---	-----
					264
	460				
<i>P. aeruginosa</i>	LMGTIAVGEA	WCG	463 SEQ ID NO: 46		
<i>P. fluorescens</i>	---AA-E-K		458 SEQ ID NO: 47		
<i>P. putida</i>	---AA---V		459 SEQ ID NO: 48		
					264

FIG. 15 Continued

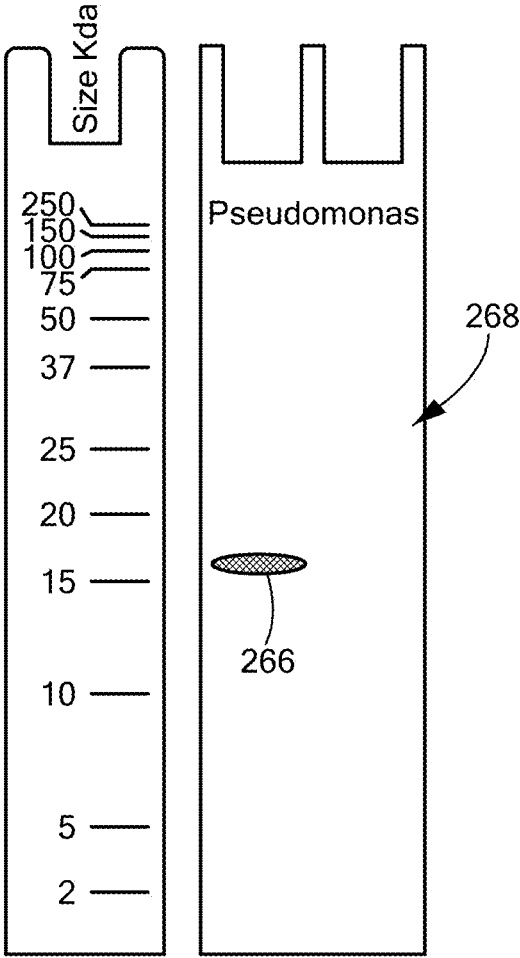


FIG. 16

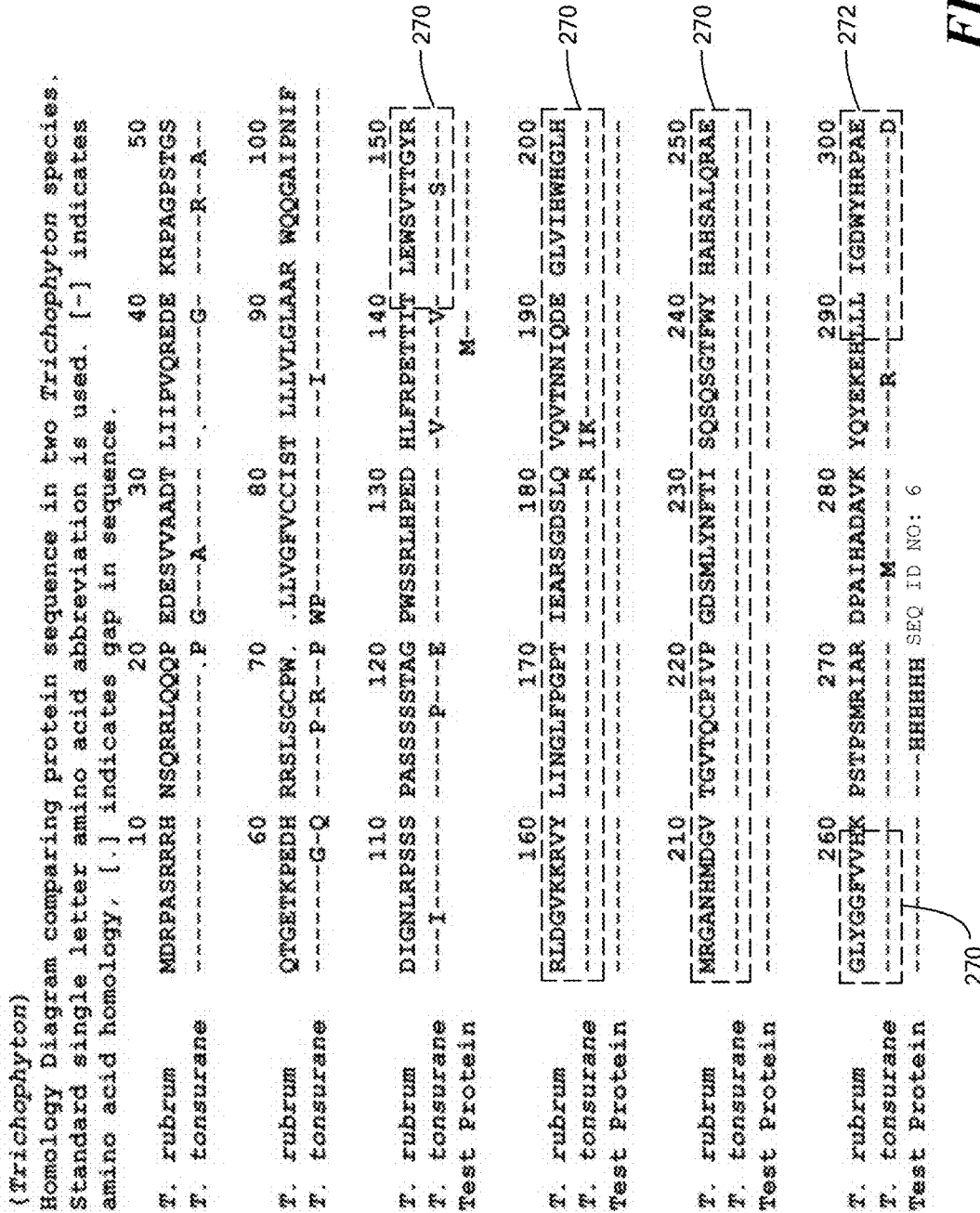


FIG. 17

T. rubrum	310	320	330	340	350	
T. tonsurane	DVLKWFKSLE	ANGQEPVDS	FLINGAGRFN	CSMALPTPI	DCVDEGYTF	272
	N		L			
T. rubrum	360	370	380	390	400	
T. tonsurane	ELLDSST	SY RMRVINVSL	AGVSLGFRG	TVTPIQVDSG	TEVELPSVSP	272
	SSS	V	A			
T. rubrum	410	420	430	440	450	
T. tonsurane	NARSMGIVYP	QORTDFVLRN	FLCETGQSSI	TVELDECFPS	LENPALTRVQ	272
	I	AF-GAE				
T. rubrum	460	470	480	490	500	
T. tonsurane	TFPISGSARK	PS.HPLSDNPI	GEAGTHVDLI	ELTSTASTIS	HIPAKADETF	274
	R	S-T	S		E	
T. rubrum	510	520	530	540	550	
T. tonsurane	LVYTLKLS	SNNVFFAFF	NHISWRPOAD	FELPLISLQR	KWDKXQFTI	274
T. rubrum	560	570	580	590	600	
T. tonsurane	KTSSRASWVD	LIVNNLDEGP	HFFHINGHDF	YVMSLHEADT	GMGSYNFWD	274
	K	V				
T. rubrum	610	620	630	640	650	
T. tonsurane	SNKAPAYDHS	QAILRDTVHI	FARGHAVLRF	KADNFGIWL	HCILMHLAS	274
	Q	N	K			
T. rubrum	660	670				
T. tonsurane	GMAMLVDVMD	SASRPLHGIL	NQTCRYLT			274
		V	P			

FIG. 17 Continued

SEQ ID NO: 49

SEQ ID NO: 50

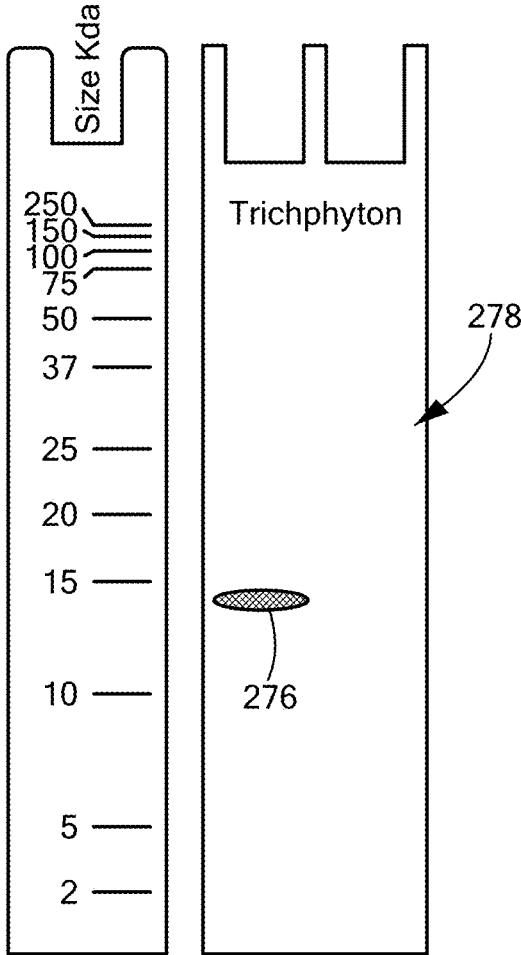


FIG. 18

{Candida}

Homology Diagram comparing protein sequence in four Candida species. Standard single letter amino acid abbreviation is used. [-] indicates amino acid homology, [.] indicates gap in sequence.

	10	20	30	40	50		
<i>Candida albicans</i>	MRPIVSSFIF	FISFLSSLIT	AKHTRWYFKT	SNVDANPDGV	FFPKMIGFND	} 280	
<i>Candida dubliniensis</i>	--RMTL--LVI	S-T-EF--A	[.]	G-N	-E-F-		
<i>Candida tropicalis</i>	--SASLFL..	L-CS-I-V-S	[.]	W-Q	G-N		-ERP-
<i>Candida auris</i>	--NQLSLF-VL	--W.FA-AS	[.]	G-K	-N-E-DV--G		
Test Protein		M-	[.]	[.]	[.]		

	60	70	80	90	100	
<i>Candida albicans</i>	SWELPTLRVK	KGDTVNLVLI	NGFDDRNTSL	RFHGLFQHG	NQMDGPEMVT	} 280
<i>Candida dubliniensis</i>	[.]	-RIQ-	-NL-T	[.]	N-	
<i>Candida tropicalis</i>	T	[.]	-R-Q-N	[.]	M-N-	
<i>Candida auris</i>	[.]	-R-	-T	[.]	M-N-S A	
Test Protein	[.]	[.]	[.]	[.]	[.]	

	110	120	130	140	150		
<i>Candida albicans</i>	QCFIPPGETF	LYNFTVDDQV	GSYWYHSHTS	GQYGDGMRCV	FIIEDDDFPY	} 280	
<i>Candida dubliniensis</i>	[.]	-Y	[.]	-T-A	[.]		V
<i>Candida tropicalis</i>	[.]	[.]	-G	[.]	[.]		[.]
<i>Candida auris</i>	[.]	-Y	-A	-T-A	-AF		EKNKE
Test Protein	[.]	[.]	[.]	[.]	[.]		HHHHHH

SEQ ID NO: 7

	160	170	180	190	200	
<i>Candida albicans</i>	DYD...	EEVVLTL	SEHYHDYSKD	LMPGFLSRFN	FTGAEPIPSN	ILFNETRNNT
<i>Candida dubliniensis</i>	[.]	-D-	GD	-NE II-T-	[.]	Q F-L
<i>Candida tropicalis</i>	[.]	[.]	A	-F-DE -T-K-	[.]	M
<i>Candida auris</i>	--PFDFD	-L-P- G-W-	-PADV	-L-KF-N-Y-	[.]	Q L

	210	220	230	240	250		
<i>Candida albicans</i>	NRVEPGKTYL	LRIANTGRFV	TCYLWMDHE	FTVVEVDGVY	VEKNTTDMLY	} 282	
<i>Candida dubliniensis</i>	[.]	V-V-I-G-	S	-Q	-I		I
<i>Candida tropicalis</i>	[.]	-N-V-V-I-	S-I	-D	-I		-Q-L
<i>Candida auris</i>	[.]	-NT-F V-V-M-G-	S	-Y	-EI		-L
Test Protein	[.]	[.]	[.]	[.]	[.]		[.]

	260	270	280	290	300		
<i>Candida albicans</i>	ITIAQRYGVL	ITPENSTNEN	YAFMNRVDDT	MLDTIFRDLQ	LRGTYIIVYN	} 282	
<i>Candida dubliniensis</i>	[.]	-T-S-	[.]	-D-Q-TD	-V-S-		[.]
<i>Candida tropicalis</i>	[.]	-V-S-	[.]	-E-D-I	-V-G-E		[.]
<i>Candida auris</i>	[.]	V	-K-ERADR-	[.]	-AF-V-I		-S-Q-T
Test Protein	[.]	[.]	[.]	[.]	[.]		[.]

	310	320	330	340	350			
<i>Candida albicans</i>	ESAPLEDAYD	VDSIDDYLD	FYLKPLNKEK	LLDDADYTIT	VDVQMDNLGR	} 284		
<i>Candida dubliniensis</i>	[.]	-D-S-	[.]	-L-S-	[.]		-ND	
<i>Candida tropicalis</i>	[.]	-D-D-EP-L	L	-FF-	-W-S-		[.]	LE
<i>Candida auris</i>	[.]	DDTSM	-E-F I-P-RF-	[.]	-V-RDG-		[.]	-P-SDNQVV I-K-D
Test Protein	[.]	[.]	[.]	[.]	[.]		[.]	[.]

282

FIG. 19

	360	370	380	390	400	
<i>Candida albicans</i>	GVNYAFFNNI	TYMTPEVPTL	LSVLSAGDAS	TNELVYGSNT	NSFVLQGGDV	284
<i>Candida dubliniensis</i>	-----KA-----	-T-----A	-----I--T--	-----E--	-----E--	
<i>Candida tropicalis</i>	-----AH-----	M-----S--DA	S-----T--	-----E--	-----E--	
<i>Candida auris</i>	-----S-VA--I-L-	ATAM--ELA	--SYI--	-----A--	-----KK-ET	
	410	420	430	440	450	
<i>Candida albicans</i>	VDIVLNNLDT	GRHPPHLHGH	VFQLIERHKE	IPOTEDPVSY	NVSDHAEWPE	284
<i>Candida dubliniensis</i>	-----K-----	A-----E-	-----T-	-AT--D-	-----D-	
<i>Candida tropicalis</i>	I--M-----K-	-----EG	VD--D--A-	S	-----S	
<i>Candida auris</i>	-----Q-D--T-	-----G	PEF.G--F	DYNN-S-F--	-----F--	
	460	470	480	490	500	
<i>Candida albicans</i>	YFMSRDTVYV	KPQSYIVMRF	KADNPGVWFF	HCHIEWHLQ	GLAIVLIEEP	284
<i>Candida dubliniensis</i>	-----M-----	R-----E-	-----E-	-----FQ--	-----D-	
<i>Candida tropicalis</i>	-----L--I-I	N--A-L-	-----E-	-----E-	-----V-A-	
<i>Candida auris</i>	-----K-----	N-N-----	T-----E-	-----E-	-----V-A-	
	510	520	530	540	550	
<i>Candida albicans</i>	EAIQENSSQH	LTDNHHQICE	RVGVSWEGNA	AANSNNYLDL	KGENIQVKRL	
<i>Candida dubliniensis</i>	-G--QE--Q	I--E--	-----P--	-GNTG--	-----V-H--	
<i>Candida tropicalis</i>	Q--E--K	I-----	-----F-Q--	-----NRD--N-	D--L--	
<i>Candida auris</i>	-EM--DP--Q	--E-F-DV-S	-G-MNYS--	-G--VDFM--	T-M-T-P--	
	560	570	580	590	600	
<i>Candida albicans</i>	FTGFTARGIV	ALVFSCIAAF	LGIAAIAYYG	MNDIEDVEER	VARDLDVBLD	
<i>Candida dubliniensis</i>	-----K--	-----G--	-M--S--	-----QNM-K-	I-----YF-	
<i>Candida tropicalis</i>	-----K--	-----GV	-LV--S--	-T--KN--Q-	-----Q-	
<i>Candida auris</i>	-A-----	-----GV	-MV--TI--	LA-VK-ID--	-----	
	610	620				
<i>Candida albicans</i>	RENEDEEEAE	IVNEGSSSSG	SNSKQH	SEQ ID NO: 51		
<i>Candida dubliniensis</i>	DDE-EDQS.	-TEQDATG-S	-SPSNK	SEQ ID NO: 52		
<i>Candida tropicalis</i>	DDDVEQLSEE	GSSGSN-KQH	SEQ ID NO: 53			
<i>Candida auris</i>	-IAA--SSQ.	L-PGD--RN	K	SEQ ID NO: 54		

FIG. 19 Continued

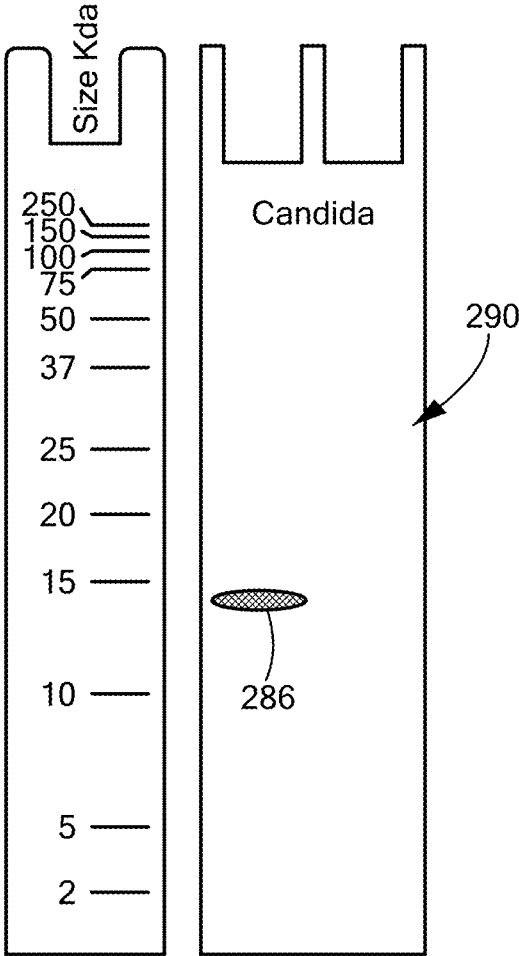


FIG. 20

Homology Diagram for α Gliadin Proteins or Prolamin in Common Grains

Gliadin protein is the immunogenic component of gluten and must be avoided by Celiac patients. Legend - standard single letter abbreviations are used for amino acids. [-] indicates homology. [.] indicates gap in sequence alignment. Amino Acid numbering aligned with example of Bread wheat α Gliadin.

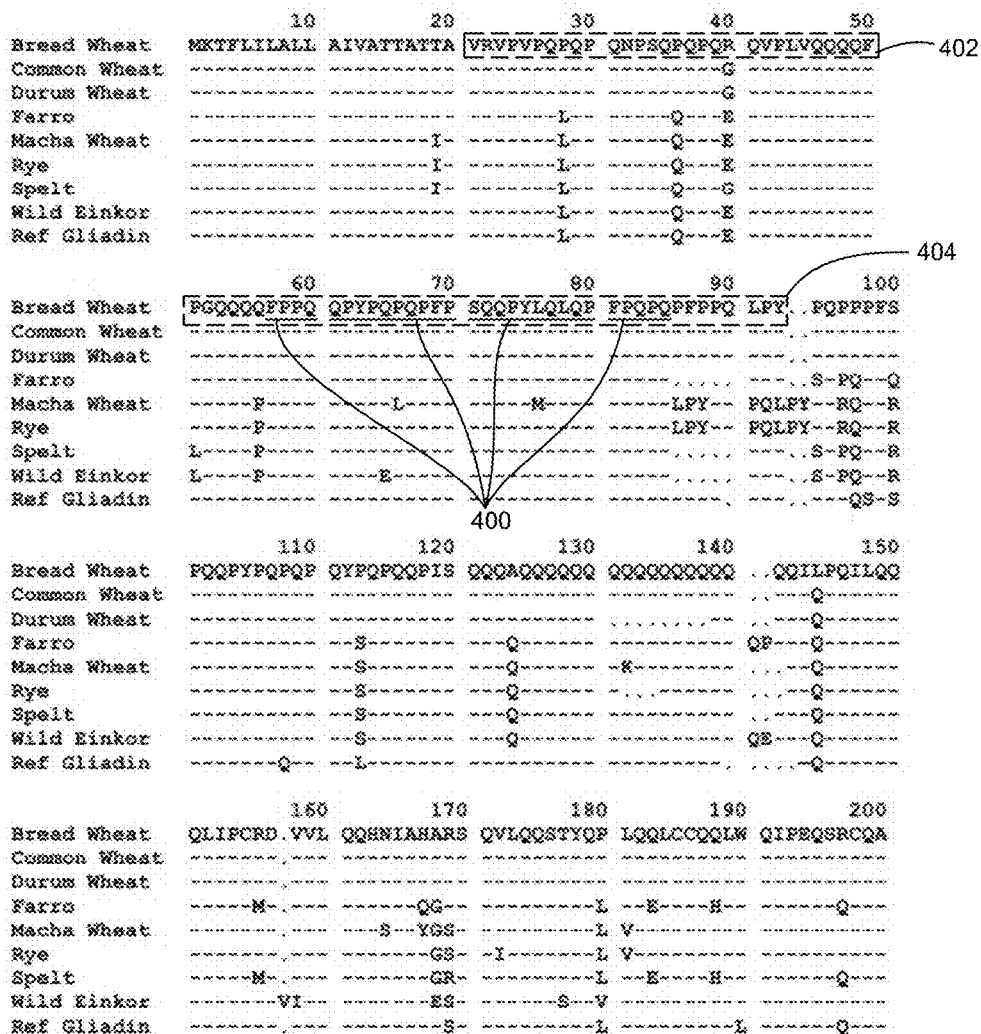


FIG. 21

	210	220	230
Bread Wheat	IHNVVHAILL HQQQQQQQ	PS	SGVELQQFQQ
Common Wheat	-----R-----		
Durum Wheat	-----R--R-----		-----F-----
Farro	-----K--Q---K---Q---		-----F-----
Macha Wheat	-----K--Q---K---Q---		-----L--F-----
Rye	-----K--Q---K---Q---		-----F-----
Spelt	-----K--Q---K---Q---		-----L--F-----
Wild Einkor	-----K--Q---K---Q---		-----Y-----
Ref Gliadin	---A---M-----	EQNQQLQQQQQQQQQLQQQQQQQ	-----F-----

	240	250	260	270	280
Bread Wheat	QVPSGGGFFQ	FSQQNPPQAQG	SVQFPQLPQF	EEIRNLALGT	LPFMCNVYIF
Common Wheat	-----	-----	-----	-----	-----
Durum Wheat	-----	-----	-----	-----	-----
Farro	-----S-R--L-----	-----	-----A-----	-----A-----	-----
Macha Wheat	-----S-----	-----H-----	-----K-----	-----AV-----	-----
Rye	-----S-----	-----	-----	-----A-----	-----
Spelt	-----L--S-R--S-----	-----	-----Q-----	-----A-----	-----
Wild Einkor	-----S-----	-----F--H-----	-----	-----A-----	-----
Ref Gliadin	-----S-VS--L-----	-----	-----A-----	-----A-----	-----

	290	SEQ ID NO: 55	SEQ ID NO: 56
Bread Wheat	PYCSTTTAPF GIPGTN	-----	-----
Common Wheat	-----	-----	-----
Durum Wheat	-----V--S-----	-----	-----
Farro	-----H-----	-----	-----
Macha Wheat	-----	-----	-----
Rye	-----V-----	-----	-----
Spelt	-----	-----	-----
Wild Einkor	-----	-----	-----
Ref Gliadin	FR---I---S---	-----	-----

FIG. 21 Continued

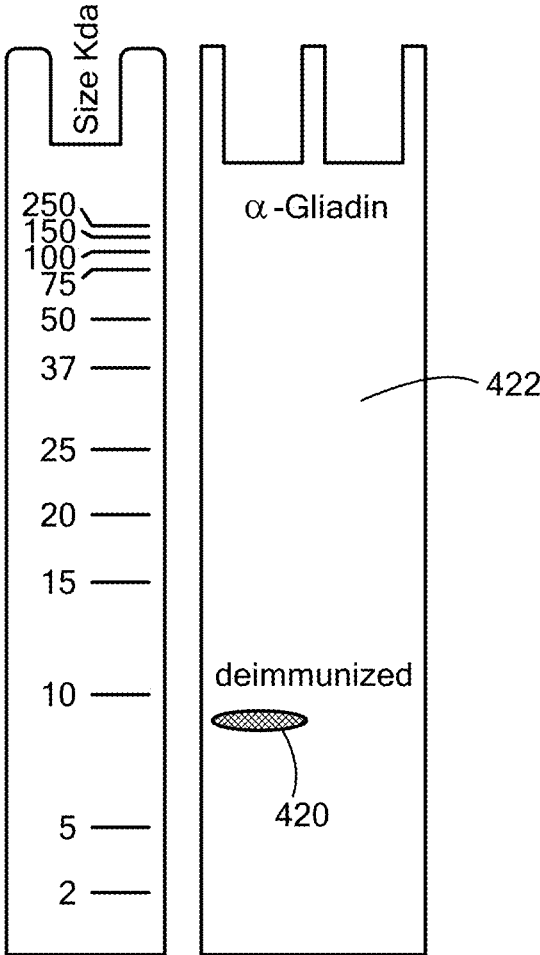


FIG. 22

1

METHOD FOR RAPIDLY DETERMINING EFFECTIVE STERILIZATION, DEIMMUNIZATION, AND/OR DISINFECTION

RELATED APPLICATIONS

This application claims benefit of and priority to U.S. Provisional Application Ser. No. 62/314,617 filed Mar. 29, 2016, under 35 U.S.C. §§ 119, 120, 363, 365, and 37 C.F.R. § 1.55 and § 1.78, which is incorporated herein by this reference.

FIELD OF THE INVENTION

This invention relates to a method for rapidly determining effective sterilization, deimmunization, and/or disinfection.

BACKGROUND OF THE INVENTION

A wide range of infectious agents, including infectious proteins, spore forming bacteria, vegetative bacteria, fungus and viruses have major impacts in medical settings. The process to remove infectious organisms or render them non-infectious from medical equipment makes use of a wide range of sterilization devices or equipment and disinfection devices and processes. The CDC lists examples of infectious agents and microorganisms by resistance to standard disinfection and sterilization processes. See Table 1 below from CDC's Guideline for Disinfection and Sterilization in Healthcare Facilities, 2008.

Some infectious agents, such as the HIV virus, may be easy to remove from medical equipment. Many infectious agents, including vegetative bacteria, are moderately difficult to eliminate. Other infectious agents, such as prions, can only be destroyed by extremely harsh conditions that damage and/or destroy modern medical equipment. Failure to eliminate infectious agents from medical equipment before use can put patients at extreme risk of injury and death.

TABLE 1

Decreasing order of resistance of infectious agents and microorganisms to disinfection and sterilization.	
Agent Category	Example Organisms or Diseases
Prions	Creutzfeldt-Jakob Disease
Bacterial spores	<i>Bacillus atropheus</i>
Coccidica	<i>Cryptosporidium</i>
Mycobacteria	<i>M. tuberculosis</i> , <i>M. terrae</i>
Nonlipid or small viruses	polio, coxsackie
Fungi	<i>Aspergillus</i> , <i>Candida</i>
Vegetative bacteria	<i>S. aureus</i> , <i>P. aeruginosa</i>
Lipid of medium-sized viruses	HIV, herpes, hepatitis B

Some conventional methods to determine if sterilization equipment functions effectively may rely on FDA approved Biologic Indicator process (BI strips) in a multi-step process. This widely accepted conventional process starts with filter papers infused with a defined number of bacterial spores (BI strips). The BI strips are subjected to a standard cycle by the sterilization equipment or device, e.g., an ethylene oxide (EtO) sterilization, radiation, or steam sterilization equipment which is being qualified. After the sterilization process is completed, the treated strips are then placed in a defined bacterial media for growth; frequently for days to weeks. If no growth is seen after the defined period, the sterilization process by the medical equipment

2

being certified is declared a success. Together this combination of supplies and techniques is the approved process to qualify sterilization equipment in positive or negative process. If there is growth, the sterilization equipment fails and if there is no growth the sterilization equipment passes.

The conventional biologic indicator tests may use one of three different species of bacteria. The standard species used to test the effectiveness of ethylene oxide (EtO) sterilization is *B. atropheus*. To test the effectiveness of gamma radiation sterilization, the bacteria species used is *B. pumilis*. To test the effectiveness of steam sterilization the bacteria species used is *G. stearothermophilus*. However, the three species that are used to qualify sterilization capacity of equipment are not bacteria that commonly cause disease in humans. Instead, the species are surrogate species, strains of soil bacteria that form high persistent spores. They are used instead of medically relevant infectious agents, because, inter alia, the spores of the bacteria are extremely difficult to damage such that they can no longer replicate, and if for some reason a health care worker or patient accidentally comes in contact with the spores through use or on improperly cleaned equipment, there is very little chance that the human will become ill. As spores from the surrogate species are scientifically known to be more difficult to destroy than medically relevant species, such as Polio or *S. aureus*, e.g., Methicillin-resistant *Staphylococcus aureus* (MRSA), when the sterilization equipment is qualified to destroy all spores on a BI test strip, the FDA accepts that the equipment is also able to destroy all organisms that rank lower for resistance to sterilization.

The conventional methods discussed above used to qualify effective sterilization, deimmunization, or disinfection may only measure the ability of the surrogate organisms to grow after sterilization treatment. However, such conventional methods do not indicate how the surrogate organisms are damaged and/or destroyed resulting in the absence of growth. Conventional surrogate testing methods also require the accurate production, storage, transport and handling of 10 thousand to 100 million pure bacteria spores, proper control of growth medias, extended period of growth of the specific spores and careful protection of all growth materials for environmental contamination to qualify if all the test surrogate organisms were completely eliminated. If any component of the process is not vigorously controlled, the sterilization qualification could give false positive or false negative results. False positive results will trigger extensive effort to unnecessarily repair sterilization equipment as well as the recall of days or weeks of sterilized medical equipment and the patients treated with such equipment. False negative results are worse because they will result in defective sterilization equipment being used and the resulting contaminated medical equipment endangering patients.

For other infectious organisms, such as members of the bacterial genera *Clostridium*, *Staphylococcus* or fungal genera *Trichophyton* or *Candida*, and the like, specific tests for each genera may be based on similar fundamentals. See Table 2 below for a list of common bacteria and fungus genera and species having an impact on human medicine.

TABLE 2

Common bacteria and fungus genera and species having an impact on human medicine.		
Genus	Example Species	Health Care Application - Problem pathogen or BL1/sterilization testing organism.
<i>Bacillus</i>	<i>Bacillus subtilis</i>	Research organism, used as an “indicator organism” during disinfection testing. BL1.
	<i>Bacillus atrophaeus</i>	Used as an “indicator organism” during gas (EtO) sterilization procedure. BL1.
	<i>Bacillus pumilis</i>	Used as an “indicator organism” during radiation sterilization procedure. BL1.
	<i>Geobacillus stearotherophilus</i> (formerly <i>B. stearotherophilus</i>)	Used as an “indicator organism” during steam sterilization procedure. BL1.
	<i>Bacillus anthracis</i>	Causes anthrax.
	<i>Bacillus cereus</i>	Causes food poisoning similar to that caused by <i>Staphylococcus</i> .
<i>Clostridium</i>	<i>Clostridium sporogenes</i>	Used as a surrogate for <i>C. botulinum</i> when testing the efficacy of commercial sterilization.
	<i>Clostridium tetani</i>	Causes tetanus.
	<i>Clostridium botulinum</i>	Causes botulism poisoning.
	<i>Clostridium perfringens</i>	Causes gas gangrene
	<i>Clostridium difficile</i>	Causes <i>C. dif</i> GI infection.
	<i>Clostridium novyi</i>	Causes a wide range of human and animal infections depending on type.
<i>Mycobacterium</i>	<i>Mycobacterium tuberculosis</i>	Major cause of human tuberculosis.
	<i>Mycobacterium africanum</i>	Slow growing form of tuberculosis.
	<i>Mycobacterium caprae</i>	More rare form of human tuberculosis.
	<i>Mycobacterium kansasii</i>	Chronic human pulmonary disease resembling tuberculosis (involvement of the upper lobe).
	<i>Mycobacterium ulcerans</i>	Infects the skin and subcutaneous tissues, giving rise to indolent nonulcerated and ulcerated lesions.
	<i>Mycobacterium interjectum</i>	Chronic lymphadenitis
	<i>Mycobacterium leprae</i>	Causes leprosy
	<i>Mycobacterium lepromatosis</i>	Causes leprosy
	<i>Mycobacterium terrae</i>	Causes serious skin infections that are relatively resistant to antibiotic therapy.
<i>Staphylococcus</i>	<i>Staphylococcus aureus</i>	Casual resident of human stomachs, but not considered an etiologic agent of disease. (BL1). Causes a variety of infections in the body, including boils, cellulitis, abscesses, wound infections, toxic shock syndrome, pneumonia, and food poisoning.
		Substrain - Methicillin-resistant <i>Staphylococcus aureus</i> (MRSA).
		Substrain - Vancomycin-resistant <i>Staphylococcus aureus</i> (VISA) - acquired gene from VRE.
	<i>Staphylococcus capitis</i>	Associated with prosthetic valve endocarditis, forms biofilms.
	<i>Staphylococcus epidermidis</i>	Hospital-acquired concern as it forms biofilms catheters or other surgical implants.
	<i>Staphylococcus haemolyticus</i>	Second-most frequently isolated hospital-acquired Infection, often associated with the insertion of medical devices; highly antibiotic-resistant phenotype and able to form biofilms.

TABLE 2-continued

Common bacteria and fungus genera and species having an impact on human medicine.		
Genus	Example Species	Health Care Application - Problem pathogen or BL1/sterilization testing organism.
	<i>Staphylococcus lugdunensis</i>	Wide variety of infections including cardiovascular infections, osteomyelitis and prosthetic/native joints infections, skin and soft-tissue infection, central nervous infections, peritonitis, endocephalitis, and urinary tract infections.
	<i>Staphylococcus saccharolyticus</i>	May cause of infective endocarditis.
	<i>Staphylococcus saprophyticus</i>	Common cause of community-acquired urinary tract infections.
	<i>Staphylococcus auricularis</i>	Occasionally can be involved with human skin infections.
<i>Salmonella</i>	<i>Salmonella enterica</i>	Causes food poisoning.
<i>Enterococcus</i>	<i>Enterococcus faecalis</i>	Can cause endocarditis and septicemia, urinary tract infections, meningitis, and other infections. Substrain - Vancomycin-resistant <i>Enterococcus</i> (VRE).
	<i>Enterococcus faecium</i>	Neonatal meningitis or endocarditis. Substrain - Vancomycin-resistant <i>Enterococcus</i> (VRE).
	<i>Enterococcus gallinarum</i>	Known to cause outbreaks and spread in hospitals.
	<i>Enterococcus hirae</i>	Endocarditis and septicemia in humans.
	<i>Enterococcus malodoratus</i>	Frequently the cause of hospital-acquired noscomial infections, bloodstream infections, and urinary tract infections.
<i>Escherichia</i>	<i>Escherichia coli</i>	Some serotypes can cause serious food poisoning in their hosts. Substrain K-12 strain commonly used in recombinant DNA work (BL1). Substrain O157:H7 causes serious illness or death in the elderly, the very young, or the immunocompromised. Substrain O104:H4, can trigger major cause of foodborne illness and lead to hemolytic-uremic syndrome (HUS).
	<i>Escherichia fergusonii</i>	Known to infect open wounds and may also cause bacteraemia or urinary tract infections; highly resistant to the antibiotic ampicillin and some also resistant to gentamicin and chloramphenicol.
<i>Helicobacter</i>	<i>Helicobacter pylori</i>	Cause gastritis and ulcers.
	<i>Helicobacter hepaticus</i>	May be associated with Crohn's disease and ulcerative colitis.
	<i>Helicobacter bilis</i>	May be associated with Crohn's disease and ulcerative colitis.
	<i>Helicobacter ganmani</i>	May be associated with Crohn's disease and ulcerative colitis.
<i>Klebsiella</i>	<i>Klebsiella pneumoniae</i>	Causes pneumonia, urinary tract infections, septicemia, meningitis, diarrhea, and soft tissue infections; naturally resistant to many antibiotics. Substrain - CREs - carbapenem-resistant <i>Klebsiella pneumoniae</i> (CRKP).
	<i>Klebsiella oxytoca</i>	Cause colitis and sepsis.
<i>Neisseria</i>	<i>Neisseria gonorrhoeae</i>	Causes Gonorrhoea.
	<i>Neisseria meningitidis</i>	Causes meningitis.

TABLE 2-continued

Common bacteria and fungus genera and species having an impact on human medicine.		
Genus	Example Species	Health Care Application - Problem pathogen or BL1/sterilization testing organism.
<i>Pseudomonas</i>	<i>Pseudomonas aeruginosa</i>	A multidrug resistant pathogen associated with hospital-acquired infections such as ventilator-associated pneumonia and various sepsis syndromes. Common in CF patients.
	<i>Pseudomonas mendocina</i>	Occasionally causes hospital-acquired infections, such as infective endocarditis and spondylodiscitis.
	<i>Pseudomonas fluorescens</i>	Produces enzymes that cause milk to spoil and occasionally infects immunocompromised patients.
	<i>Pseudomonas putida</i>	Used in bioremediation, or the use of microorganisms to biodegrade oil.
<i>Trichophyton</i> (Fungus)	<i>Trichophyton rubrum</i>	Most common cause of athlete's foot, fungal infection of nail, jock itch, and ringworm.
	<i>Trichophyton tonsurans</i>	Causes ringworm infection of the scalp.
	<i>Trichophyton interdigitale</i>	One of three common fungi which cause ringworm.
	<i>Trichophyton mentagrophytes</i>	Causes tinea infections including athlete's foot, ringworm, jock itch, and similar infections of the nail, beard, skin and scalp.
<i>Candida</i> (Fungus)	<i>Candida albicans</i>	Associated with the skin infection tinea imbricate.
	<i>Candida albicans</i>	Dimorphic fungus that grows both as yeast and filamentous cells; Responsible for 50-90% of all cases of candidiasis in human. Important causes of morbidity and mortality in immunocompromised patients. Biofilms may form on the surface of implantable medical devices. Cause of 85-95% of vaginal infections cases are responsible for physician office visits every year.
	<i>Candida dubliniensis</i>	A fungal opportunistic pathogen originally isolated from AIDS patients. It is also occasionally isolated from immunocompetent individuals.
	<i>Candida tropicalis</i>	Common pathogen in neutropaenic hosts; research suggests that <i>C. tropicalis</i> , working synergistically with <i>Escherichia coli</i> and <i>Serratia marcescens</i> . May cause or contribute to Crohn's disease
	<i>Candida auris</i>	Causes candidiasis in humans; often acquired in the hospital when human immune systems are weakened; causes fungemia, yielding candidemia (systemic candidiasis); attracted clinical attention because of multidrug resistance.

Examples of standardized methods for sterilization or disinfection (A) and standardized testing methods protocols (B) used to determine the effectiveness of sterilization or disinfection, as shown in Table 3 below:

TABLE 3

Approved Methods of Sterilization or Disinfection and Qualifying Test Protocols	
(A) Standard Methods for Preparing Healthcare Equipment	
Disinfection	Sterilization
Alcohol	Steam Sterilization
Chlorine and Chlorine Compounds	Flash Sterilization
Formaldehyde	Ethylene Oxide "Gas" Sterilization
Glutaraldehyde	Hydrogen Peroxide Gas Plasma
Hydrogen Peroxide	Peracetic Acid Sterilization
Iodophors	Ionizing Radiation
Ortho-phthalaldehyde	Dry-Heat Sterilizers
Peracetic Acid	Liquid Chemicals
Peracetic Acid and Hydrogen Peroxide	Performic Acid
Phenolics	Filtration
Quaternary Ammonium Compounds	Microwave
Radiation	Glass Bead "Sterilizer"
Pasteurization	Vaporized Hydrogen Peroxide
Flushing- and Washer-Disinfectors	Ozone
	Formaldehyde
	Gaseous Chlorine Dioxide
	Vaporized Peracetic Acid
	Infrared radiation
(B) Standard Test to Qualify Healthcare Equipment	
Test Name	Example Test Species
BI (ethylene oxide (EtO) sterilization)	<i>B. atrophaeus</i>
BI (gamma radiation sterilization)	<i>B. pumilis</i>
BI (steam sterilization)	<i>G. stearothermophilus</i>
AOAC Sporicidal Efficacy Test Method	<i>Clostridium sporogenes</i> <i>Bacillus subtilis</i>
AOAC Tuberculosis Rate of Kill	<i>Mycobacterium terrae</i>
AOAC Use Dilution Test	<i>Pseudomonas aeruginosa</i> <i>Staphylococcus aureus</i> <i>Salmonella enterica</i>
AOAC Fungicidal Efficacy Test Method	<i>Trichophyton mentagrophytes</i>

For each standard test protocol, a define number of organisms are placed on a carrier, such as tube, filter paper, or coated on and in a test solid surface. The specific organisms may be a particular infectious species or could be a surrogate species of the same genus that is closely related to the infectious species. In all cases, the species, carrier and growth conditions are defined by the FDA and/or the Association of Analytical Communities (AOAC) protocol. Following treatment with sterilization or disinfection equipment, the carrier with the specific species sample is placed into ideal growing conditions for the particular test species. After a required period in culture, usually 2 to 30 days, the culture is monitored. If no growth is observed, the sterilization or disinfection equipment is declared to be operating within required parameters.

In addition to enabling growth and infectivity, protein components of infectious organisms could trigger severe immunogenic or allergic reactions in susceptible individuals even at very low level. Examples include mold proteins that are able to trigger severe allergic reactions even if the mold has been rendered no longer able to grow. Immunogenic proteins can also occur in food such as gliadin, a highly immunogenic protein component of the seed storage protein gluten in wheat and related grains. Gliadin can trigger reactions in most individuals suffering from Crohn's disease. It is critical that immunogenic proteins are completely

removed from any equipment that will be used in conjunction with susceptible individuals.

A wide range of pathogenic organisms use a multicopper oxidase with 3 cupredoxin superfamily domains for growth and survival. As disclosed herein, the loci *suf I* that contains a critical protein that confers different functions depending on the genus (bacteria or fungus) and this critical protein can be targeted. Depending on the genus, the *suf I* loci encoded protein can have different names. The functions of the protein encoded by *suf I* include cell division (FtsP), formation of spore coat proteins (CotA), chromosome partitioning, inorganic ion transport, and metabolism and cell wall, membrane, and envelope formation. As the protein product of the *suf I* loci are absolutely critical for the survival of the spores (in spore forming bacteria) and/or growth (all bacteria and fungus), if the protein product of the *suf I* loci is irreversibly fragmented into short polypeptides and amino acids, the bacteria or fungus cannot survive. Additionally, it is likely that a sterilization method that clearly demonstrates fragmentation of the protein product of the *suf I* loci would also fragment other proteins in the bacterium or fungus. Bacteria and fungus can be divided into distinct genus each containing multiple species. Many species also have subspecies that carry unique characteristics include multi-drug resistance. In human health situations, certain bacteria and fungus species and subspecies are of major concern because they are capable of causing disease. Related species may be used in medical research, e.g., *E. coli* K12, or as indicator species for qualification of sterilization, e.g., *B. atrophaeus* used to qualify gas sterilization. See Table 2 above for a list of common bacteria and fungus genera and species with impact on human medicine.

Prions are a unique category of a transmissible infectious agent that comprised only of protein, without DNA or RNA. Prions can cause a wide range of neurodegenerative diseases known as transmissible spongiform encephalopathies (TSE) or prion diseases including the new variant Creutzfeldt-Jakob disease (nvCJD). See Table 4 below. Infectious prions are in fact an abnormally folded brain protein. This brain protein (Protease resistant Proteins, PrP) can be folded into two different structural (tertiary) forms, the normal brain protein, PrP_c, and the abnormal, disease triggering form, PrP_{sc}. The disease triggering form, PrP_{sc}, is found in high quantity in the brain of infected humans and animals and can be transferred to a new host with the transfer of infected material. Once in the new host, the abnormally folded protein (PrP_{sc}) causes disease symptoms by promoting the unfolding of the normal host protein (PrP_c) and refolding into the disease causing form (PrP_{sc}). PrP proteins can also be partially cleave and still retain their infectious characteristics. Full length mature PrP protein (both PrP_c and PrP_{sc}) is 209 amino acids long. Limited proteolysis of PrP_{sc} will cleave amino acids from the amino terminus resulting in another infectious protein form PrP 27-30 that is approximately 142 amino acids long. Additional cleavage that significantly reduces the 142 amino acid long PrP 27-30 is needed to render the PrP protein irreversibly non-infectious. Although most infectious agents can be permanently rendered non-infectious by heat or steam, these methods are not sufficient to eliminate infectious prions from medical equipment.

TABLE 4

Example of Prion Diseases in Different Species and Potential Origin of the Infectious Protein.					
Disease	Species	Potential Origin	Disease	Species	Potential Origin
Creutzfeldt-Jakob disease (CJD)	Human	Inherited	Scrapie	Sheep and Goat	Inherited/ environmental
New Variant Creutzfeldt-Jakob disease (CJD)	Human	Consumption, Medical Contamination	Bovine Spongiform Encephalopathy (BSE)	Cattle	Consumption
Fatal Familial Insomnia (FFI)	Human	Inherited	Transmissible Mink Encephalopathy (TME)	Mink	Environmental
Gerstmann-Straussler disease (GSD)	Human	Inherited	Chronic Wasting Disease (CWD)	Mule Deer and Elk	Environmental
Huntington disease-like type 1 (HDL1)	Human	Inherited	Feline Spongiform Encephalopathy (FSE)	Cats	Consumption
Kuru	Human	Consumption of Human Brains	Exotic Ungulate Encephalopathy (EUE)	Nyala and Greater Kudu	Environmental

As discussed above, prions are abnormally folded protease resistant proteins (PrP_{sc}) that cause disease symptoms by promoting the unfolding of normal proteins (PrP_c) and refolding into the disease causing protein form (PrP_{sc}). As the level of the PrP_{sc} rises in the patient's brain, symptoms of progressive dementia, myoclonic seizures, abnormalities of high cortical function, cerebellar and corticospinal disturbances develop. The period between infection and development of disease can extend for years to decades. The duration of disease symptoms is variable but is typically 8 to 18 months.

Once prion proteins fold into the infectious form (PrP_{sc}), they are extremely difficult to render non-infectious. Conventional methods to sterilize medical equipment contaminated with prions, such as high heat to promote loss of function of other protein types by triggering loss of tertiary structure, are ineffective because unlike most proteins, the denatured prion proteins, both infectious and non-infectious, will spontaneously refold by themselves back to their pretreatment forms. In some cases, conventional methods may actually result in refolded into infectious form promoting the conversion of the non-infection prion protein into the infectious prion protein.

To render infectious proteins such as prions irreversibly non-infectious, all infectious proteins must be fragmented into small polypeptides, amino acids or components. The only currently approved conventional method for this process is harsh treatment of medical equipment and supplies with caustic soda, an extremely harsh process that frequently damages and/or destroys medical equipment.

Determining whether or not an infectious prion (PrP_{sc}) sample has been permanently destroyed can be extremely difficult and time consuming. Conventional methods for determining whether an infectious prion has been permanently destroyed require that after attempted deactivation, the PrP_{sc} sample is injected into a matched susceptible animal that is then followed for an extended time to see if the animal develops disease. In larger animals, the process can take years, but even in a small animal such as a mouse, the test can take months. As there is a potential for inter-animal variation and poor test accuracy, a large animal test pool is required to obtain relatively accurate results.

Immunogens may include a wide range of molecules including proteins that can trigger dramatic immunologic responses in susceptible individuals. The responses can trigger serious allergic reactions on the skin (e.g., poison ivy

20

rash), in the gut (e.g., triggering a flare-up in Crohn's disease), in the lung (e.g., asthma) or a systemic response (e.g., anaphylaxis). Protein immunogens are a special class of immunogens produced by a wide range of bacteria, fungus (e.g. mold) or plants and can be difficult to destroy. An example of a common plant immunogen is gluten. Common grains such as various strains of wheat, farro, rye and spelt are derived from wild and domesticated grains of the *Triticum*, *Aegilops* and *Secale* genera. Common to all these species is the seed storage protein complex called gluten. When seeds are ground into flour, the gluten protein complex gives bread dough its elastic quality and bread its spongy texture. Unfortunately gluten is comprised of several proteins including Gliadin (also called Prolamin) which triggers severe T cell attack on the gut of patients with the autoimmune disease Celiac disease (CD). Gliadins can be typed as α , γ , and ω with a small protease resistant fragment (p57-73) of α -gliadins triggering the most severe destructive T cell response. As a results CD patients must not only avoid products containing gluten, but also need to be extremely careful to avoid small amounts of residual α -gliadin that may contaminate food preparation utensils.

Protease resistant proteins like α -gliadin are resistant to destruction so it is critical that devices and methods used to destroy them and other immunogens (also called allergens) can be easily checked to ensure they are operating at peak efficiency. If not, residual allergens can trigger life threatening responses in sensitive patients. The process of removing immunogens by deimmunization methods or devices is called deimmunization. The ability to test for the destruction of different immunogens on surfaces is not standardized. Usually affected patients are subjected to skin test regiments to determine their individual reactions to different candidate immunogens/allergens. The patient is then advised to avoid all immunogen contact and discard any materials potential contaminated with the specific immunogen or allergen. In cooking and manufacturing situations, extreme care must be taken to avoid potential cross contamination to the point that food packaging labels frequently carry warning labels about the potential issues.

Thus, a method is needed to determine irreversible destruction of proteins critical for the growth of infectious organisms, immunogenic proteins, and/or infectious proteins (e.g., prions) and thus rapidly and accurately determines the effectiveness of sterilization, deimmunization, and/or disinfection of equipment or supplies by a device.

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With such a method for rapidly determining effective sterilization, deimmunization, and/or disinfection, medical personnel and patients can have confidence that the medical equipment used for patient treatment is not contaminated with potentially lethal or immunogenic proteins. Without such a method, medical personnel may believe they are using properly sterilized equipment and then later discover that they have accidentally exposed their patients to lethal infections and harmful immune reactions

SUMMARY OF THE INVENTION

In one aspect, a method for rapidly determining effective sterilization, deimmunization, and/or disinfection of equipment and/or supplies by a device is featured. The method includes providing a defined surrogate protein having a predetermined sequence representative of an infectious agent potentially contaminating the equipment and/or the

supplies to be sterilized, deimmunized, and/or disinfected by the device. The defined surrogate protein having the predetermined sequence is subjected to sterilization, deimmunization, or disinfection. The effectiveness of the sterilization, deimmunization, or disinfection is rapidly determined by determining if the defined surrogate protein having the predetermined sequence has been destroyed.

In one embodiment, the defined surrogate protein may include proteins critical for stability, growth and/or infectious capacity of infectious agents. The defined surrogate protein may include a protein critical for stability, growth and/or infectious capacity of surrogate organisms of infectious agents. The infectious agent may include one or more of: an infectious protein, an infectious spore forming bacteria, an infectious vegetative bacteria, an infectious fungus, and an infectious virus. The defined surrogate protein may include pathogenic proteins, proteins critical for the growth of infectious agents, and immunogenic proteins. The predetermined sequence may be defined by the sequence:

(SEQ ID NO: 1)

10	20	30	40	50
MNYNTSAKYE	VP IAIQDRSF	NEDGSLNFPS	EGDNPTIHPY	WQPEFFGDTI
MVNGRVVPM	NVDMTRYRFR	LLNGSNARFY	NLKFSGMQF	WQIGTDGGYL
NKPVPLTSL	ISPGERADIL	VDFTEIPAGT	RIILNNDANA	PYPTGDAPDK
DTTGQIMQFT	VQHNDHHHHH	H		

The defined surrogate protein for SEQ ID NO: 1 may be at least 95% homologous the predetermined sequence or substantial fragment of the predetermined sequence. The predetermined sequence may be defined by the sequence:

(SEQ ID NO: 2)

10	20	30	40	50
MTLEKTYEYEV	TMEECTHQLH	RDLPPTRLWG	YNGLFPGPTI	EVKRNENVYV
KWMNNLPSTH	FLPIDHTIHH	SDSQHEEPEV	KTVVHLHGGV	TPDSDSGYPE
AWFSKDFEQT	GPYFKREVYH	YPNQQRGAIL	WYHDHAMALT	RLNVYAGLVG
AYIIHDPKEK	RLKHHHHHH			

The defined surrogate protein for SEQ ID NO: 2 may be at least 95% homologous to the predetermined sequence or substantial fragment of the predetermined sequence. The predetermined sequence may be defined by the sequence:

(SEQ ID NO: 3)

10	20	30	40	50
MTGMPEGEGV	DSNLLGGDGG	DIAYPYYLIN	GRIPVAATSF	KAKPGQRIRI
RIINSAADTA	FRIALAGHSM	TVTHTDGYPV	IPTEVDALLI	GMAERYDVMV
TAAGGVFPLV	ALAEKNALA	RALLSTGAGS	PPDHHHHHH	

The defined surrogate protein for SEQ ID NO: 3 may be at least 95% homologous to the predetermined sequence or substantial fragment of the predetermined sequence. The predetermined sequence may be defined by the sequence:

(SEQ ID NO: 4)

10	20	30	40	50
MTGYKNYTLK	AQKGKTEFYK	NNFSNTLGYN	GNULLGPTLKL	KKGDKVKIKL
INNLDENTTF	HWHGLEVNKG	VDGGPSQVIK	PGKEKTIKFE	VNQDSATLWY
HPHPSPNTAK	QVYNGLSGLL	YIEDSKKNHH	HHHH	

15

The defined surrogate protein SEQ ID NO: 4 may be at least 95% homologous to the predetermined sequence or substantial fragment of the predetermined sequence.

The predetermined sequence may be defined by the sequence:

```

                10          20          30          40          50
(MSEQ ID NO: 5)
MTGFRHEKVL  CLKTWHVDEQ  GAFTPFVSVPR  QAAREGTRGR  YSTINGKHVP
TIDLPAQQIV  RVRLLNVDNT  VTYRLNPNGE  ARIYANDGHP  VEPRGFEGQY
WIGPGMRLEL  ALKVPEAGTE  LSLRDGPVRL  ATIRSVAHHH  HHH
    
```

The defined surrogate protein SEQ ID NO: 5 may be at least 95% homologous to the predetermined sequence or substantial fragment of the predetermined sequence. The predetermined sequence may be defined by the sequence:

```

                10          20          30          40          50
(MSEQ ID NO: 6)
MTITLEWSVT  TGYRRLDGVK  KRUYLINGLF  PGPTIEARSG  DSLQVQVTNN
IQDEGLVIHW  HGLHMRGANH  MDGVTGVTQC  PIVPGDSMLY  NFTISQSQSG
TFWYHAHSAL  QRAEGLYGGF  VVHKPSTHHH  HHH
    
```

The defined surrogate protein SEQ ID NO: 6 may be at least 95% homologous to the predetermined sequence or substantial fragment of the predetermined sequence. The predetermined sequence may be defined by the sequence:

```

                10          20          30          40          50
(MSEQ ID NO: 7)
MTAETHTWYF  KTSWVDANPD  GVFPKRMIGF  NDSWPLPLTR  VKKGDTVNLV
LINGFDDRNT  SLHFHGLFQH  GTNQMDGPEM  VTQCPIPPGE  TFLYNFTVDD
QVGSYWYHSH  TSGQYGDGMR  GVFIIEDHHH  HHH
    
```

The defined surrogate protein SEQ ID NO: 7 may be at least 95% homologous to the predetermined sequence or substantial fragment of the predetermined sequence. The predetermined sequence may be defined by the sequence:

```

                10          20          30          40          50
(MSEQ ID NO: 8)
MKTVRVPVPQ  PQQPNPSQPQ  PQRQVPLVQQ  QQFPGQQQQP  PPQQPYQPQP
PPPSQQPYLQ  LQFPQPQPFP  PPQLPYHHHH  HH
    
```

The defined surrogate protein SEQ ID NO: 8 may be at least 95% homologous to the predetermined sequence or substantial fragment of the predetermined sequence. The rapidly determining may include a sensitive protein analysis procedure. The sensitive protein analysis procedure may include one or more of: a Western Blot analysis, a protein assay analysis, a magnetic separation analysis, a peptide analysis, a mass spectrometry analysis, and a gas chromatography analysis. The sensitive protein analysis procedure may include fluorescence analysis of proteins covalently crosslinked on a solid surface. The sensitive protein analysis procedure may include fluorescence analysis of proteins covalently crosslinked on magnetic beads. The defined surrogate protein having the predetermined sequence may be disposed on a surface, disposed on a test strip, disposed in or on a vessel, on a tube, or in or on a holder. The holder may be disposed to receive a flow of a sterilization agent, a deimmunization agent or a disinfection agent.

16

BRIEF DESCRIPTION OF THE SEVERAL VIEWS OF THE DRAWINGS

Other objects, features and advantages will occur to those skilled in the art from the following description of a pre-

ferred embodiment and the accompanying drawings, in which:

FIG. 1 is a schematic block diagram showing the primary steps of one embodiment of the method for rapidly deter-

mining effective sterilization, deimmunization, and/or disinfection of this invention;

FIG. 2 is a schematic diagram showing examples of multi-well glass slides having samples of one or more of the

defined surrogate protein having a predetermined sequence in wells which are subjected to sterilization, deimmunization, or disinfection to provide a visual depiction of the

effectiveness of sterilization, deimmunization, and/or disinfection;

FIG. 3 is a schematic block diagram showing one example of the multi-well glass slides shown in FIG. 2 placed in a holder disposed at the end of a flow of sterilization, deimmunization, or disinfection agent in accordance with one embodiment of this invention;

FIG. 4 shows an example of an amino acid comparison of human PrP proteins with a selection of other species of PrP proteins;

FIG. 5 shows an example of a Western gel where the recombinant protein runs approximately 28 kDa inside;

FIG. 6 shows an example of a Western Blot where the absence of bands indicates successful sterilization, deimmunization, or disinfection;

17

FIG. 7 is a homology diagram comparing protein sequences of a research *Clostridium* species and two pathogenic *Clostridium* species;

FIG. 8 shows an example of a Western Blot for a defined *Clostridium* surrogate protein having a predetermined sequence that has been subjected to sterilization, deimmunization, or disinfection;

FIG. 9 is a homology diagram comparing protein sequences of a three research *Bacillus* species;

FIG. 10 shows an example of a Western Blot for a defined *Bacillus* surrogate protein having a predetermined sequence that has been subjected to sterilization, deimmunization, or disinfection;

FIG. 11 is a homology diagram comparing protein sequences of three pathogenic *Mycobacterium* species;

FIG. 12 shows an example of a Western Blot for a defined *Mycobacterium* surrogate protein having a predetermined sequence that has been subjected to sterilization, deimmunization, or disinfection;

FIG. 13 is a homology diagram comparing protein sequences of three pathogenic *Staphylococcus* species;

FIG. 14 shows an example of a Western Blot for a defined *Staphylococcus* surrogate protein having a predetermined sequence that has been subjected to sterilization, deimmunization, or disinfection;

FIG. 15 is a homology diagram comparing protein sequences of a research *Pseudomonas* species and two pathogenic *Pseudomonas* species;

FIG. 16 shows an example of a Western Blot for a defined *Pseudomonas* surrogate protein having a predetermined sequence that has been subjected to sterilization, deimmunization, or disinfection;

FIG. 17 is a homology comparing protein sequences of two pathogenic *Trichophyton* species;

FIG. 18 shows an example of a Western Blot for a defined *Trichophyton* surrogate protein having a predetermined sequence that has been subjected to sterilization, deimmunization, or disinfection;

FIG. 19 is a homology comparing protein sequences of four pathogenic *Candida* species;

FIG. 20 shows an example of a Western Blot for a defined *Candida* surrogate protein having a predetermined sequence that has been subjected to sterilization, deimmunization, or disinfection;

FIG. 21 is a homology comparing protein sequences of α -Gliadin from many species of commonly consumed grains;

FIG. 22 shows an example of a Western Blot for a defined α -Gliadin surrogate protein having a predetermined sequence that has been subjected to sterilization, deimmunization, or disinfection.

DETAILED DESCRIPTION OF THE INVENTION

Aside from the preferred embodiment or embodiments disclosed below, this invention is capable of other embodiments and of being practiced or being carried out in various ways. Thus, it is to be understood that the invention is not limited in its application to the details of construction and the arrangements of components set forth in the following description or illustrated in the drawings. If only one embodiment is described herein, the claims hereof are not to be limited to that embodiment. Moreover, the claims hereof are not to be read restrictively unless there is clear and convincing evidence manifesting a certain exclusion, restriction, or disclaimer.

18

The method for rapidly determining effective sterilization, deimmunization, and/or disinfection of equipment or supplies of one or more embodiments of this invention may be utilized to qualify sterilization, deimmunization, and/or disinfection by a device, e.g., a sterilization device, a deimmunization device, or a disinfection device and provide improvements to the conventional methods discussed above. In one example, the method for rapidly detecting effective sterilization, deimmunization, and/or disinfection of equipment or supplies of one or more embodiments of this invention may be based on a specific measuring the complete destruction of a specific protein critical for an organism's growth and requires only a few hours to return absolute results. The method for rapidly detecting effective sterilization, deimmunization, and/or disinfection of equipment or supplies of one or more embodiments of this invention also contains multiple layers of internal controls that enable a clear determination if either false positive and false negative results have occurred. This allows the avoidance of unnecessary repairs to sterilization, deimmunization, or disinfection equipment and eliminates false negative tests and more dangerous exposure of patients to improperly sterilized, deimmunized, or disinfected medical equipment or supplies.

The method for rapidly detecting effective sterilization, deimmunization, and/or disinfection of equipment or supplies of one or more embodiments of this invention may be used to rapidly determine if a sterilization, deimmunization, and/or disinfection device is effectively destroying specific infectious or immunogenic agents, defined herein as infectious or pathogenic proteins, infectious spore forming bacteria, infectious vegetative bacteria, infectious fungus, infectious viruses, and immunogenic proteins.

In one embodiment, the method for rapidly determining effective sterilization, deimmunization and/or disinfection of equipment and/or supplies by a device, such as a sterilization device, e.g., by a device that applies cycles of a solvent and electromagnetic radiation, e.g., microwaves, such as disclosed in U.S. application Ser. No. 15/330,469 by the assignee hereof, hereinafter the '469 patent application, any of the sterilization devices discussed in the Background section above, the devices for the methods of sterilization shown in Table 3 above, an autoclave, of similar type sterilization device known to those skilled in the art, a deimmunization device, e.g., a device that applies cycles of a solvent and electromagnetic radiation, e.g., microwaves, such as disclosed in the '469 patent application and/or disinfection device, e.g., a device that applies cycles of a solvent and electromagnetic radiation, e.g., microwaves, such as disclosed in the '469 patent application, or the devices for the methods of disinfection shown in Table 3 above, includes providing a defined surrogate protein having a predetermined sequence representative of an infectious agent potentially contaminating the equipment and/or supplies to be sterilized, deimmunized and/or disinfected by the device, step 100, FIG. 1. The defined surrogate protein having the predetermined sequence is then subjected to sterilization, deimmunization, or disinfection, step 102. The effectiveness of the sterilization, deimmunization, and/or disinfection by the device is then rapidly determined by determining if the defined surrogate protein having the predetermined sequence has been destroyed, step 106, as discussed further below.

The method preferably directly measures the irreversible destruction of the defined surrogate proteins having the predetermined sequence that are critical for survival and/or growth of such infectious agents.

In one embodiment, the method utilizes one or more prion detection indicator samples configured as a defined surrogate protein. In this example, the defined surrogate protein has the following predetermined sequence:

	(SEQ ID No: 9)
KKRPKPGGWN TGGSRYPGQG SPGGNRYPPQ GGTWGQPHGG GWGQPHGGSW	50
GQPHGGSWGQ PHGGGWGQGG GTHNQWNKPS KPKTNLKHVA GAAAAGAVVG	100
GLGGYMLGSA MSRPMIHFGN DWEDRYREN MYRYPNQVYY RPDQYSNQN	150
NFVHDCVNIT IKQHTVTTTT KGENFTETDV KMMERVVEQM CVTQYQKESQ	200
AYYDGRRS	208

The defined prion surrogate protein having the predetermined sequence above for prion detection is then subjected to sterilization, deimmunization, or disinfection, by a device, e.g., a sterilization device, a deimmunization device or a disinfection device, e.g., by conventional methods discussed in the Background section above, or by applying cycles of a solvent and electromagnetic radiation, e.g., microwaves, such as disclosed in the '469 patent application.

In this example, to rapidly determine the effectiveness of sterilization, deimmunization, or disinfection of equipment or supplies by the device, a determination is made if the defined surrogate protein has been destroyed using a sensitive protein analysis procedure, such as Western Blot analysis, or similar protein analysis techniques, such as fluorescence analysis of proteins covalently crosslinked to solid surfaces used in protein array analysis which are extremely sensitive processes that measure both the amount of full length intact defined surrogate protein having a predetermined sequence and the amount of the destroyed or degraded defined surrogate protein, protein array analysis, magnetic separation analysis, peptide analysis, mass spectrometry analysis, gas chromatography analysis, or similar type analysis. In one example, the defined surrogate protein includes a protein critical for stability, growth, and/or infectious capacity of an infectious agent. The defined surrogate protein may include a protein critical for stability, growth, and/or infectious capacity of a surrogate organism of the infectious agent.

One embodiment of the method for rapidly detecting effective sterilization, deimmunization, and/or disinfection of this invention includes providing the defined surrogate protein having a predetermined sequence that is based on the development of a synthetic recombinant test protein sequence with high homology to a section of a protein encoded by the suf I loci in the targeted pathogenic genus, e.g., one or more or all of the genus shown in Table 2 above, and the development of a monoclonal or polyclonal antibody that is able to detect the defined surrogate recombinant protein using Western Blot or other similar protein analysis techniques discussed above. In genera that form spores, such as *Bacillus* and *Clostridium*, the target protein encoded in the suf I loci or defined surrogate protein having the predetermined sequence of the tests forms the spore coat protein CotA. In non-spore forming genus such as the bacteria *Mycobacterium*, *Staphylococcus* and *Pseudomonas*, or the fungus *Candida*, the target protein encoded in the suf I loci or defined surrogate protein of the tests forms the cell division protein (FtsP). In other genera of bacteria and fungus, the target protein or encoded defined surrogate protein having the predetermined sequence in the suf I loci may have

additional names but all carry the same protein structure of multicopper oxidase with 3 cupredoxin superfamily domains.

For each method of rapidly determining effective sterilization, deimmunization, or disinfection, specific for its unique genus, a predetermined quantity of the defined surrogate having the predetermined protein sequence is placed on a carrier, such as filter paper, or a tube, or on the surface of an object, such as a glass or slide, a microtiter plate, a flexible membrane, magnetic beads, e.g., magnetic beads used for magnetic bead separation or similar type object or surface. For Western analysis, the defined surrogate protein having the predetermined sequence is not covalently linked to the carrier or surface. In other protein analysis processes such as protein array or magnetic bead separation, the defined surrogate protein having the predetermined sequence is covalently linked to the carrier or solid surface. After treatment with the sterilization, deimmunization or disinfection equipment of process and Western analysis, this involves recovering the recombinant defined surrogate protein having the predetermined sequence, both in its intact and fragmented forms, from the carrier, surface tube, or object, treating the sample with denaturing loading buffer and running the sample on an acrylamide gel. A control sample that was not subjected to sterilization, deimmunization, or disinfection is also included. In one example, the samples are transferred nylon membrane and the intact and fragmented samples are visualized using the specific antibody. If the sterilization, deimmunizing or disinfection equipment is operating correctly, the control sample will have protein indicator bands that are easily visualized but the treated sample will be absent any type protein indicator bands indicating the sterilization, deimmunizing or disinfection equipment was able to irreversibly fragment the recombinant defined surrogate protein. If other protein analysis processes such as protein array or magnetic bead separation are used, the solid surface to which defined surrogate protein having the predetermined sequence was covalently linked will be tested such that intact defined surrogate protein having the predetermined sequence can be visualized using the specific antibody and fragment and/or destroyed defined surrogate protein having the predetermined sequence is no longer detected by the process. As the amino acid sequence and structure of the defined surrogate protein having a predetermined sequence is highly homologous to the target protein in the pathogenic members of the genus, irreversible destruction of the defined surrogate protein indicates that the sterilization, deimmunizing or disinfection equipment will have also destroyed the target proteins, resulting in destruction of all members of the genus that may be on the sterilization, deimmunizing or disinfection equipment or supplies.

In accordance with one or more embodiments of the method for rapidly determining effective sterilization, deimmunization, and/or disinfection, the predetermined sequence of the defined surrogate protein and the corresponding peptide used for development of a polyclonal antibody for Western Blot analysis include one or more of the following predetermined sequences:

For *Clostridium*:

```

                                (SEQ ID NO: 1)
      10          20          30          40          50
MNYNYSKAYE  VPVIAIQDRSF  NEDGSLNFPF  EGDNPTIHPY  WQPEFFGDTI
MVNGRVWPNM  NVDMTRYRFR  LLNGSNARFY  NLKFSNGMQF  WQIGTDGGYL
NKPVPLTSL  ISPGERADIL  VDFTEIPAGT  RIILNNDANA  PYPTGDAPDK
DTTGQIMQFT  VQHNDHHHHH  H

```

The peptides used for the development of polyclonal and monoclonal antibodies for use by Western Blot analysis for the above sequence are:

15

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                                (SEQ ID NO: 10)
      KYEVPIAIQDRSFNEDGSLNFPSE
      and
                                (SEQ ID NO: 11)
      YLNKPVPLTSLISPGERADILVD

```

For *Bacillus*:

```

                                (SEQ ID NO: 2)
      10          20          30          40          50
MTLEKTYEYEV  TMEECTHQLH  RDLPPTRLWG  YNGLFPGPTI  EVKRNENVYV
KWMNLPSTH  FLPIDHTIHH  SDSQHEEPEV  KTVVHLHGGV  TPDDSDGYPE
ANFSKDFEQT  GPYFKREVYH  YPNQQRGAIL  WYHDHAMALT  RLNVYAGLVG
AYIIHDPKEK  RLKHHHHHH

```

The peptides used for the development of polyclonal and monoclonal antibodies for use by Western Blot analysis for the above sequence are:

35

```

                                (SEQ ID NO: 12)
      QRGAILWYHDHAMALTRLNVYAGL
      and
                                (SEQ ID NO: 13)
      QLHRDLPPTRLWGYNGLFPGPTIE

```

For *Mycobacterium*:

```

                                (SEQ ID NO: 3)
      10          20          30          40          50
MTGMPEGEGV  DSNLLGGDGG  DIAYPYYLIN  GRIPVAATSF  KAKPGQRIRI
RIINSAADTA  FRIALAGHSM  TVTHTDGYPV  IPTEVDALLI  GMAERYDVMV
TAAGGVFPLV  ALAEGKNALA  RALLSTGAGS  PPDHHHHHH

```

The peptides used for the development of polyclonal and monoclonal antibodies for use by Western Blot analysis for the above sequence are:

55

```

                                (SEQ ID NO: 14)
      DTAFRIALAGHSMTVTHTDGYVPVITEVD
      and
                                (SEQ ID NO: 15)
      VFPLVALAEGKNALARALLSTGAGS

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65

For *Staphylococcus*:

(SEQ ID NO: 4)

10	20	30	40	50
MTGYKNYTLK	AQKGKTEFYK	NNFSNTLGYN	GNULLGPTLKL	KKGDKVKIKL
INNLDENTTF	HWHGLEVNGK	VDGGPSQVIK	PGKEKTIKFE	VNQDSATLWY
HPHPSPNTAK	QVYNGLSGLL	YIEDSKKNHH	HHHH	

The peptides used for the development of polyclonal and monoclonal antibodies for use by Western Blot analysis for the above sequence are:

(SEQ ID NO: 16)

NFSNTLGYNGLLGP TLK LK K G D K V K I K L
and

(SEQ ID NO: 17)

KFEVNQDSATLWYHPHPSPNTAK

For *Pseudomonas*:

(SEQ ID NO: 5)

10	20	30	40	50
MTGFRHEKVL	CLKTWHVDEQ	GAFTPFVSVPR	QAAREGTRGR	YSTINGKHVP
TIDLPAQQIV	RVRLLNVDNT	VTYRLNPNGE	ARIYAVDGHP	VEPRGFEQQY
WIGPGMRLEL	ALKVPEAGTE	LSLRDGPVRL	ATIRSVAHHH	HHH

The peptides used for the development of polyclonal and monoclonal antibodies for use by Western Blot analysis for the above sequence are:

(SEQ ID NO: 18)

DLPAGQIVRVRLLNVDNTV TYRLN
and

(SEQ ID NO: 19)

QYWIGPGMRLELALKVPEAG

For *Trichophyton*:

(SEQ ID NO: 6)

10	20	30	40	50
MTITLEWSVT	TGYRRLDGVK	KRVYLINGLF	PGPTIEARSG	DSLQVQVTNN
IQDEGLVIHW	HGLHMRGANH	MDGVTGVTQC	PIVPGDSMLY	NFTISQSQSG
TFWYHAHSAL	QRAEGLYGGF	VVHKPSTHHH	HHH	

The peptides used for the development of polyclonal and monoclonal antibodies for use by Western Blot analysis for the above sequence are:

(SEQ ID NO: 20)

YRRLDGVKKRVYLINGLFP GPPTIE
and

(SEQ ID NO: 21)

TQCPIVPGDSMLYNFTISQSQSG

For *Candida*:

```

                (SEQ ID NO: 7)
          10      20      30      40      50
MTAETHTWYF  KTSWVDANPD  GVFPKRMIGF  NDSWPLPTLR  VKKGDTVNLV
LINGFDDRNT  SLHFHGLFQH  GTNQMDGPEM  VTQCPIPPGE  TFLYNFTVDD
QVGSYWYHSH  TSGQYGDGMR  GVFIIEDHHH  HHH
    
```

The peptides used for the development of polyclonal and monoclonal antibodies for use by Western Blot analysis for the above sequence are:

```

                (SEQ ID NO: 22)
          15      20      25      30
GFNDSWPLPTLRVKKGDTVNLV
and
                (SEQ ID NO: 23)
          15      20      25      30
WYFKTTSWVDANPDGVFPKRMIG
    
```

For α -Gliadin:

```

                (SEQ ID NO: 8)
          10      20      30      40      50
MKTVRVPVPQ  PQPNPSQPQ  PQRQVPLVQQ  QQFPGQQQF  PPQQPYPPQ
PFPSQQPYLQ  LQPPFPQPF  PPQLPYHHHH  HH
    
```

The peptide used for the development of monoclonal or polyclonal antibody used by Western Blot analysis for the above sequence is:

```

                (SEQ ID NO: 24)
          15      20      25      30      35
FPPQQPYPPQPFPSQQPYLQLQPPFPQ
    
```

Western Blot analysis typically utilizes equipment, e.g., acrylamide gel, a power supply to create an electric field to trigger protein migration where smaller fragments move faster than larger fragments to separate intact proteins from fragmented or degraded proteins, a membrane, transfer equipment, and visualization equipment. Western Blot analysis also preferably utilizes specific reagents, e.g., a positive control protein to show the location of a full length defined surrogate protein and specific antibody for the defined surrogate protein, e.g., any of the defined surrogate proteins having the associated predetermined sequences and the corresponding antibody above. The specific antibody binds to the associated defined surrogate protein, both full length, and fragments, to provide a visualization if the defined surrogate protein was destroyed by sterilization, deimmunization, or disinfection.

Western Blot analysis for infectious organisms may also typically include a defined number of colony forming units (CFU) or spores of the test pathogen which are added to a stable substrate such as filter paper (dried) or other sample holder. The number of CFU or spores will contain a defined quantity of each indication protein to be followed.

Western Blot analysis may also be conducted on defined surrogate proteins having the associated predetermined sequences samples placed on indicator strips or other small sample holders that maybe subjected to sterilization, deimmunization or disinfection. After sterilization, deimmunization or disinfection, the defined surrogate proteins from the indicator strips or other small sample holders are extracted from the filter paper or other sample holders into a loading dye and denatured (eliminating any tertiary protein struc-

10 ture). The samples are then run on the gel, including control wells with (1) size marker, (2) positive control protein, (3) other controls if needed. After transfer to membrane, defined surrogate proteins and protein fragments are visualized with
 15 unique antibodies preferably having a high affinity and specific binding to an indicator region of the protein being tested. Successful sterilization, deimmunization or disinfection will result in the loss of all indicator proteins of defined length. The development process may require side by side
 20 Western Blots and standard growth studies to demonstrate sufficient equivalence.

30 Western Blot analysis may also require defining the defined surrogate proteins having the associated predetermined sequence, positive control proteins, and/or a negative control proteins and may require the development of a polyclonal or monoclonal antibodies that specifically bind to specific defined surrogate proteins discussed above in West-
 35 ern Blot analysis or other protein analysis format, both full-length and fragments, and the positive control protein. The antibody will not bind to the negative control protein. It is also possible that a secondary antibody that is labeled with
 40 an enzyme or other visualization marker will be needed to visualize the detection antibody.

Protein array analysis can be used as a substitution for Western blot analysis in circumstances in which less sensi-
 45 tivity can be tolerated, but faster results are needed. In a Western Blot analysis, a substrate such as a piece of filter paper or tube is used to temporarily hold the recombinant protein sample during testing of sterilization equipment. After sterilization, deimmunization and/or disinfection, the recombinant protein sample is removed from the substrate,
 50 denatured, separated on an acrylamide gel and transferred to nylon membrane before visualizing with antibodies. In protein array analysis, the recombinant protein sample, e.g., one or more of the defined surrogate proteins having the associated predetermined sequence discussed above, is covalently crosslinked to solid surfaces such as glass, plas-
 55 tics or metal beads. After treatment by sterilization, deimmunization or disinfection, the solid surfaces, with the covalently crosslinked recombinant proteins are directly visualized with antibodies. Unlike Western blot analysis that
 60 can visualize the amount of protein fragmentation has occurred, protein array analysis can indicate how much of the protein sample has lost regions that are recognized by visualizing antibodies protein assay analysis but is unable to
 65 determine what other areas of the recombinant proteins were not destroyed. Thus, protein array analysis is highly suitable for testing the presence or absence of intact test proteins, such as the defined surrogate proteins having the associated

predetermined sequence homologous to proteins critical for spore coats or bacteria or fungus survival or growth, e.g., one or more of the defined surrogate proteins having the predetermined sequences for *Clostridium*, *Bacillus*, *Mycobacterium*, *Staphylococcus*, *Pseudomonas*, *Trichophyton*, *Candida*, and α -Gliadin shown above. Because protein assay analysis cannot determine the absolute level of protein fragmentation that has occurred, protein array analysis may not be sensitive enough for quantifying the complete destruction of infectious proteins like prions although it is not excluded as a test method if less sensitivity is tolerated.

To establish a protein array analysis, defined quantities of the defined surrogate proteins for *Clostridium*, *Bacillus*, *Mycobacterium*, *Staphylococcus*, *Pseudomonas*, *Trichophyton*, *Candida*, and α -Gliadin above are covalently linked to a solid surface. The solid surface may include a wide support surface, such as a microscope slide made of glass or silicon, a flexible membrane, a magnetizable bead, a microtiter plate, or other similar solid surface to which a selection of the defined surrogate proteins having the associated predetermined sequence can be permanently attached. The surface used is determined by the type detector that will be used to determine the quantity of protein left on the support after sterilization, deimmunization or disinfection. After sterilization, deimmunization or disinfection, the solid surface is visualized with antibodies or other specific ligands that will specifically bind to the intact defined surrogate protein but not to destroyed defined surrogate proteins. The antibodies or other ligands can contain fluorescent dyes or other detection enabling attachments. Control slides that were not subjected to sterilization, deimmunization and/or disinfection are included as a positive control to enable estimations for the relative amount of test protein destroyed by sterilization, deimmunization and/or disinfection. FIG. 2 shows an example of multi-well glass slide 130 which has the control defined surrogate protein having the associated predetermined sequence in the wells exemplarily, indicated at 132 and multi-well glass slide 134 which has one of the defined surrogate proteins having the predetermined sequence for *Clostridium*, *Bacillus*, *Mycobacterium*, *Staphylococcus*, *Pseudomonas*, *Trichophyton*, *Candida*, and α -Gliadin in the wells, exemplarily indicated at 136. In this example, control multi well glass slide 130 was not subject to sterilization, deimmunization, or disinfection and visualization of the antibodies is indicated by the shading as shown which indicates the defined surrogate protein having the associated predetermined sequence was not sterilized, deimmunized, or disinfected. In this example, multi-well glass slide 134 was subject to sterilization, deimmunization and/or disinfection and visualization shows no presence of antibodies and, therefore, the defined surrogate proteins having the predetermined sequence were effectively sterilized. Thus, in this embodiment, the method for rapidly determining effective sterilization was able to quickly determine effective sterilization, deimmunization and/or disinfection in as little as two hours depending on the type of detection equipment used to rapidly determine effective sterilization, deimmunization or disinfection discussed above.

One key benefit of using protein array analysis in accordance with one embodiment of the method for rapidly determining effective sterilization, deimmunization, and/or disinfection of this invention is the covalently linked surrogate proteins on the surface of an object can be used for testing sterilization, deimmunization or disinfection by a sterilization, deimmunization or disinfection device that may use radiation or heat, as well as devices that use flowing disinfectants or gases to sterilize, deimmunize or disinfect

medical equipment or supplies. In one example, holder 150, FIG. 3, may be used to hold multiwell glass slide 152 having one or more wells, exemplarily indicated at 154, which each holds one or more samples of the defined surrogate protein having the predetermined sequence discussed above, e.g., one of *Clostridium*, *Bacillus*, *Mycobacterium*, *Staphylococcus*, *Pseudomonas*, *Trichophyton*, *Candida* or α -Gliadin. Holder 150 with multi-well glass slide 152 including one or more samples of the defined surrogate protein having the predetermined sequence discussed above may then be placed below flow 156 of sterilization, deimmunization, or disinfection agent at the end of a medical device undergoing sterilization, deimmunization, or disinfection, indicated at 158. After the test is complete, holder 150 is then removed, indicated at 160, and multi-well glass slide 152 can be easily transferred for complete protein analysis and for rapid detection of sterilization, deimmunization, or disinfection as discussed above.

If magnetizable beads are used as the solid surface, one or more of the defined surrogate proteins having the associated predetermined sequence could also be mixed into liquid sterilization, deimmunization and/or disinfection steam and later harvested with a strong magnet. Flowing disinfectants or gases would result in the complete loss of the defined surrogate protein having the associated predetermined sequence samples that are not covalently linked to a support surface and the protein array analysis may be used to rapidly determine effective sterilization, deimmunization or disinfection.

Both Western Blot Analysis and protein array analysis are an indirect measurement of sterilization, deimmunization and/or disinfection. Sterilization is the complete destruction of spore forming infectious organisms so the standard direct measurements are long growth studies to determine the number of organisms that survived the sterilization process. Deimmunization is the complete destruction of immunogenic proteins such that no protein fragments capable of triggering an immune reaction in human or animal remain. Disinfection is the complete destruction of vegetative bacteria, fungus and/or viruses. As disclosed herein, the defined surrogate proteins are considered to be in the form of isolated fragments and destroyed if proteins critical for the survival of the infectious agents are used in quantities 10,000 to 1,000,000 times as much as would occur in a standard sample of intact infectious agents. To correctly establish the scale to determine success ratio between the direct and indirect tests, multiple conditions are preferably utilized to determine the Western Blot analysis and protein array analysis conditions that perfectly align with standard sterilization, deimmunization and/or disinfection studies using intact infectious organisms and post sterilization growth conditions.

The samples prepared for Western Blot analysis and protein array analysis may use a wide range of circumstances. If the sterilization, deimmunization and/or disinfection equipment uses flowing gases or liquid to wash over the surfaces to be sterilized holder 150, FIG. 3, and multi-well glass slide 152 or similar type device, may be used to ensure the defined surrogate protein samples remain in the flowing stream of disinfecting gas or liquid. For example, if the sterilization, deimmunization or disinfecting gas or liquid is washed through medical equipment to sterilize, deimmunize, or disinfect the inside of a lumen, holder 150 with the multi-well glass slide 150 having one or more defined surrogate proteins having the associated predetermined sequence therein, e.g., *Clostridium*, *Bacillus*, *Mycobacterium*, *Staphylococcus*, *Pseudomonas*, *Trichophyton*, *Can-*

did, and α -Gliadin analysis tests could be connected to the end of the stream to better measure the quality of sterilization, deimmunization, or disinfection that occurred through the entire length of the medical equipment lumen.

The following examples are exemplary and explanatory only and do not limit or restrict this invention.

Examples

Example 1: Comparing Amino Acid Sequence of Human PrP Protein to Other Species

It is important to qualify the ability of a sterilization device to destroy prions that may be contaminating medical equipment. For the specific test described herein in accordance with one or more embodiments of the method for rapidly determining effective sterilization, deimmunization, or disinfection of this invention, a defined quantity of the defined surrogate prion protein discussed above is provided and then evaluated using Western Blot analysis and an antibody specific for the defined prion surrogate protein.

To protect the human operators of the test, infectious human prions cannot be used. Instead, the defined surrogate protein is used that incorporates all the characteristics of human prion (PrP) proteins, with the critical exception that it cannot infect humans. In this example, to select the defined surrogate PrP protein to be used, a protein analysis was conducted comparing the amino acid protein sequence of human PrP protein against a selection of the protein sequence databases of other species including a primate, two

companion animals, two food animals and two research animals. From this analysis, the mouse PrP protein was determined to be the best candidate surrogate protein. Structurally, mouse PrP protein is as robust as the human PrP protein and thus will be equivalently resistant to a wide range of destructive methods but is also sufficiently different as to be unable to infect humans. Arrow 168, FIG. 4, indicates the initiation location for Pr27-30, the smallest PrP fragment that retains infectivity. If a PrP protein is cleaved anywhere from this initiation point to the end of the protein sequence, the resulting fragments can no longer cause disease.

To evaluate additional surrogate PrP proteins that could be used, the mouse PrP protein sequence was compared against the protein sequence database. Table 5 below shows the wide diversity of a predetermined list of species PrP proteins that could be used as well as each protein's sequence ID and its homology to Mouse PrP. The human sequence is less than 90% homologous to mouse PrP. The search also demonstrates how conserved the PrP protein is across a wide range of mammalian species contained within the sequence database and any one of these could be used as the surrogate protein in the Protein Indicator Test. The database also contains the sequence for chicken PrP but as it is less than 50% homologous to other mammalian PrP proteins, it is possible that it or other related proteins could be used in the Protein Indicator Test but its divergence could impact its resistance to destruction. This would make it a less suitable surrogate for the test than other proteins, especially any mammalian PrP protein.

TABLE 5

Potential PrP Protein that could be used as Surrogate Protein in Sterilization Indication Tests. Information about each PrP protein includes species, Sequence ID number and homology to Mouse recombinant PrP protein.

Species	Sequence ID Number	Homology to Mouse (%)
Mouse (<i>Mus musculus</i>)	sp P04925.2 PRIO_MOUSE	100
Rat (<i>Rattus norvegicus</i>)	sp P13852.2 PRIO_RAT	98
Cotton Rat (<i>Stomodon hispidus</i>)	sp Q9Z0T3.1 PRIO_SIGHI	97
Chinese Hamster (<i>Cricetulus griseus</i>)	sp Q60506.1 PRIO_CRIGR	97
Grey Dwarf Hamster (<i>Cricetulus migratorius</i>)	sp Q60468.1 PRIO_CRIMI	95
Golden Hamster (<i>Mesocricetus auratus</i>)	sp P04273.1 PRIO_MESAU	95
Greater Kudu (<i>Tragelaphus strepsiceros</i>)	sp P40243.1 PRIO2_TRAST	90
Red-bellied Titi (<i>Callicebus moloch</i>)	sp P40248.1 PRIO_CALMO	96
Three-striped Night Monkey (<i>Aotus trivirgatus</i>)	sp P40245.1 PRIO_AOTTR	94
Black-capped capuchin (<i>Sapajus apella</i>)	sp P40249.1 PRIO_CEBAP	95
Common Marmoset (<i>Callithrix jacchus</i>)	sp P40247.1 PRIO_CALJA	95
Red-faced Spider Monkey (<i>Ateles paniscus</i>)	sp P51446.1 PRIO_ATEPA	94
Geoffrey's Spider Monkey (<i>Ateles geoffroyi</i>)	sp P40246.1 PRIO_ATEGE	91
Nilgai (<i>Boselaphus tragocamelus</i>)	sp Q5UJG7.1 PRIO_BOSTR	86
Alpine Musk Deer (<i>Moschus chrysogaster</i>)	sp Q68G95.1 PRIO_MOSCH	88
Common Squirrel Monkey (<i>Saimiri sciureus</i>)	sp P40258.1 PRIO_SAISC	91
Gelada Baboon (<i>Theropithecus gelada</i>)	sp Q95270.1 PRIO_THEGE	90
Black Crested Mangabey (<i>Lophocebus aterrimus</i>)	sp P67990.1 PRIO_LOPAT	90
Mona Monkey (<i>Cercopithecus mona</i>)	sp P61761.1 PRIO_CERMO	91
Patas Monkey (<i>Erythrocebus patas</i>)	sp Q95174.1 PRIO_ERYPA	91
Grivet (<i>Chlorocebus aethiops</i>)	sp P67988.1 PRIO_CHLAE	90
Mantled Guereza (<i>Colobus guereza</i>)	sp P40251.1 PRIO_COLGU	91
Bornean orangutan (<i>Pongo pygmaeus</i>)	sp P40256.1 PRIO_PONPY	91
François' Langur (<i>Trachypithecus francoisi</i>)	sp P40257.2 PRIO_TRAFR	91
Sooty Mangabey (<i>Cercocebus atys</i>)	sp Q95176.1 PRIO_CERAT	91
Crab-eating Macaque (<i>Macaca fascicularis</i>)	sp P67992.1 PRIO_MACFA	91
Mandrill (<i>Mandrillus sphinx</i>)	sp P40255.1 PRIO_MANSP	91
Gorilla (<i>Gorilla gorilla gorilla</i>)	sp P40252.1 PRIO_GORGO	90
Cat (<i>Felis catus</i>)	sp O18754.3 PRIO_FELCA	87
Human (<i>Homo sapiens</i>)	sp P04156.1 PRIO_HUMAN	89
Lar Gibbon (<i>Hylobates lar</i>)	sp P61766.1 PRIO_HYLLA	89
Bighorn Sheep (<i>Ovis canadensis</i>)	sp Q7JH3.1 PRIO_OVICA	88
Goat (<i>Capra hircus</i>)	sp P52113.1 PRIO_CAPHI	88
Blackbuck (<i>Antelope cervicapra</i>)	sp Q5UJG1.1 PRIO_ANTCE	85
Takin (<i>Budorcas taxicolor</i>)	sp Q95M08.1 PRIO_BUDTA	87

TABLE 5-continued

Potential PrP Protein that could be used as Surrogate Protein in Sterilization Indication Tests. Information about each PrP protein includes species, Sequence ID number and homology to Mouse recombinant PrP protein.		
Species	Sequence ID Number	Homology to Mouse (%)
Sheep (<i>Ovis aries</i>)	sp P23907.1 PRIO_SHEEP	87
Rabbit (<i>Oryctolagus cuniculus</i>)	sp Q95211.1 PRIO_RABIT	90
Water Buffalo (<i>Bubalus bubalis</i>)	sp Q5UJH8.1 PRIO_BUBBU	84
Rocky Mountain Elk (<i>Cervus canadensis nelsoni</i>)	sp P67986.1 PRIO_CEREN	87
Mule Deer (<i>Odocoileus hemionus</i>)	sp P47852.1 PRIO_ODOHE	87
Lesser Kudu (<i>Tragelaphus imberbis</i>)	sp Q5UJG3.1 PRIO_TRAIM	85
Greater Kudu (<i>Tragelaphus strepsiceros</i>)	sp P40242.1 PRIO1_TRAST	84
Cow (<i>Bos taurus</i>)	sp P10279.2 PRIO_BOVIN	84
Common Brushtail Possum (<i>Trichosurus Vulpecula</i>)	sp P51780.1 PRIO_TRIVU	77
Giant Panda (<i>Ailuropoda melanoleuca</i>)	sp Q6EH52.1 PRIO_AILME	84
American Mink (<i>Neovison vison</i>)	sp P40244.1 PRIO_NEOVI	79
Dromedary Camel (<i>Camelus dromedaries</i>)	sp P79141.1 PRIO_CAMDR	88
Pig (<i>Sus scrofa</i>)	sp P49927.1 PRIO_PIG	79
Ferret (<i>Mustela putorius furo</i>)	sp P52114.1 PRIO_MUSPF	78
Artic Fox (<i>Vulpes lagopus</i>)	sp B0FYL5.1 PRIO_VULLA	78
Dog (<i>Canis lupus familiaris</i>)	sp O46501.1 PRIO_CANFA	85
Chicken (<i>Gallus gallus</i>)	sp P27177.2 PRIO_CHICK	43

Example 2: Developing Method for Testing the Ability to Destroy PrP Proteins

One purpose of developing the defined prion surrogate protein test is to provide a rapid method for determining effective sterilization equipment. The method preferably has multiple steps including at least: 1) preparing defined prion surrogate proteins having a predetermined sequence, 2) subjecting the prion surrogate protein samples to sterilization, and 3) using Western Blot analysis to visualize the effects of sterilization of defined prion surrogate protein samples. Successful sterilization has occurred when all of the defined prion surrogate protein sample was fragmented and, as a result of the protein fragmentation, none remains to bind to the visualization antibodies. If sterilization was not successful, protein bands will be seen on the Western Blot analysis.

A defined surrogate protein having a sequence above for the prions, recombinant mouse PrP full-length protein (208 amino acids long) was obtained from Abcam, Cambridge Mass. On a Western gel, the recombinant protein runs at about approximated 28 kDa in size, indicated at **290**, FIG. 5.

The Western Blot analysis discussed above is well-established and is an extremely sensitive method to determine the presence and/or absence of a specific protein. The first step in Western Blot analysis involves preparing the defined prion surrogate protein samples for separation by size. In accordance with one embodiment of the method for rapidly determining effective sterilization, deimmunization, or disinfection of the defined prion surrogate proteins will be run as single denatured proteins and thus must be denatured through the use of a denaturing loading buffer, boiled for 10 minutes and then run on a denaturing gel. Once loaded on to an appropriate acrylamide gel, the samples are subjected to electric current that results in the protein fragments traveling at different rates depending on size with smaller fragments moving faster than larger ones. In one example, the gel contained 8% acrylamide. To be able to monitor the separation of the defined prion surrogate protein samples, a standard sample with a mix of proteins of defined sizes was also included. For some experiments discussed herein, addi-

25

tional protein samples may be included to provide for controls of the sample handling conditions:

After separation, the defined prion surrogate protein samples are transferred to a special nylon membrane before being permanently cross-linked to the membrane. The final steps include incubating the nylon membrane with a primary antibody available from Abcam, Cambridge Mass. that specifically binds to the defined surrogate protein of interest. In the experiments discussed herein, the primary anti-PrP antibody was a rabbit monoclonal antibody that was raised against a synthetic peptide corresponding to residues near the C-terminus of human Prion protein PrP, as indicated by underlined region indicated at **170**, FIG. 4. In addition to binding to both PrP_c and PrP_{sc}, the antibody will bind to the infectious protein form PrP 27-30. The primary antibody binds to Human, Rat and Mouse PrP proteins. The secondary antibody is a HRP-labelled goat anti-rabbit available from Abcam, Cambridge Mass. to enable visualization of the infectious PrP fragments, both intact and fragmented. If the defined prion surrogate proteins have been fragmented into non-infectious fragments that eliminate the C-terminus region of the PrP, the antibody will not bind them. Without this C-terminus region, the degraded PrP protein is no longer infectious. Very small fragments and amino acids which are no longer infectious will be too small to be retained on the gel.

The first tests involved carefully drying defined quantities of recombinant mouse PrP protein onto filter papers to assist in storage, transport to and from location of sterilization equipment and handling during sterilization tests. To create, test strips of filter paper were cut 4 mm wide by 20 mm long and the bottoms were squared. Serial dilutions (1:3) of mouse PrP protein were created to contain protein solutions 0.1 µg/µl to 0.0037 µg/µl. The protein preps were added dropwise to the filter paper strips until 10 µl were added. The filter paper strips were then air dried. Filter paper strips were prepared in duplicate with 1 µg, 0.333 µg, 0.111 µg, or 0.033 µg applied respectively to a pair of strips. When dry, the strips were placed in Eppendorf tubes and 50 µl 1× Loading Buffer where BME was added. The strips and loading buffer were incubated at 95° C. for 5 minutes then given a quick spin in a micro centrifuge. For one of each paired sample,

65

the filter paper strip was taken out of the Eppendorf tube and inserted into the well of the SDS-PAGE gel. For these samples, electroporation was used to extract any residual defined prion surrogate protein that was retained on the filter paper after extraction by boiling and centrifugation. Half of the loading buffer from these tubes was also loaded into the same well. For the other samples of the pair only boiling and centrifugation was used to extract the defined prion surrogate protein from the filter strips. From these, half of the loading buffer was added to the wells without also adding the filter paper. A control sample of 0.5 µg protein (equivalent to half of the 1 µg concentration samples) was also loaded on the gel for comparison. The gel was run and the Western Blot Analysis performed using the procedure described above.

The results demonstrated that there was no significant difference between the protein concentrations that were dried onto the filter paper and the control sample. There was also no significant difference between the extraction process using only boiling and centrifugation and the more difficult process of using boiling, centrifugation and electroporation for extraction. In addition, the Western Blot analysis demonstrated that the Western Blot analysis was sensitive to below 0.05 µg. Hence, a 1 µg of recombinant mouse PrP protein should be used for all sterilization tests. The samples should be extracted from filter paper using boiling in loading the buffer with BME and centrifugation and half of the sterilized samples used for Western Bolt analysis.

Example 3: Evaluation of Destruction of Prion Proteins Using Sterilization Equipment

Experiments were conducted to demonstrate a combination of one or more of heat, a vaporizing solvent, and electromagnetic radiation, e.g., microwaves, e.g., as disclosed in the '469 patent application, could irreversibly destroy the defined prion surrogate protein shown with the sequence shown above. The stable PrP protein was selected for the experiments as it cannot be irreversibly destroyed using a standard sterilization autoclave device. For the experiments, samples were created that each contained 1 µg of a structurally robust mouse PrP protein and wrapped in 100% cotton paper to avoid extraneous contamination. This containment was placed in a second layer of 100% cotton paper to increase stability during treatment. The samples were treated with different temperatures and for differing numbers of moisture saturation and microwave cycles, e.g., as disclosed in the '469 patent application. After treatment, the samples were prepared in loading buffer and boiled. Half of each sample was then run on a denaturing protein gel. After Western Blot analysis, it was possible to see that certain combinations of temperature and treatment cycles completely destroyed the stable protein samples, shown by the absence of protein bands of gel indicated at **294**, **296**, **FIG. 6**. Other treatment conditions did not destroy the stable proteins.

Example 4: Evaluation of Destruction of Prion Proteins Contained within Polypropylene Tubes

Additional experiments were conducted to demonstrate that with the combination of one or more of heat, saturating moisture and microwaves as disclosed in the '469 patent application could destroy the defined surrogate stable PrP proteins if the samples were contained within a vessel such as a polypropylene tube. In this example, samples of the defined prion surrogate proteins were created that each

contained about 1 µg of a structurally robust mouse protein. However, instead of drying the protein samples onto filter paper, the 1 µg samples were dried within polypropylene tubes of different lengths including 0.75 cm and 3.5 cm in length. The tubes were closed at one end (on to which the samples were dried) and open at the other. The tubes were contained within 2 layers of 100% cotton paper to prevent cross contamination during treatment. The samples were subjected to sterilization by a combination of one or more of heat, a solvent, e.g., reverse osmosis (RO) filtered water, and electromagnetic radiation with differing numbers of moisture saturation and radiation cycles, e.g., as disclosed in the '469 patent application. After treatment, the samples were prepared in loading buffer and boiled. Half of each sample was then run on a denaturing protein gel. After Western blot analysis, it was possible to see that certain combinations of temperature and treatment cycles completely destroyed the stable protein samples (as demonstrated by the absence of protein bands of gel). Other treatment conditions did not destroy the stable proteins. The results of the sterilization treatment of PrP samples dried onto filter paper and dried within a polypropylene tube were identical indicating that the method of containing the PrP sample did not alter the results.

Example 5: Comparison of Prion Protein Indicator Test Results and Mouse Model Prion Tests

All mice were kept in an AAALAC-accredited facility and handled in compliance with guidelines provided by the US Guide for the Care and Use of Laboratory Animals.

Creating Prion Infected Brain Homogenate: Brains from terminally ill C57BL/6 mice infected with 22 L prions were prepared as follows: each brain was homogenized (10%, w/v) in phosphate-buffered saline (PBS) by repeatedly passing the material first through an 18-gauge needle and then repeatedly through a 26 gauge needle. The brain homogenates were combined to make a stock preparation, diluted with PBS (1/10), aliquoted into 100 ul preps in 2 ml polypropylene freezing vials and frozen at -80° C. until use.

Division into treated and non-treated preps: frozen prion preps having the defined prion surrogate protein were allowed to thaw and the caps removed. In duplicate, samples were sterilized by a combination of one or more of heat, microwaves and saturating moisture, e.g., as disclosed in the '469 patent application. The used conditions were similar to the process disclosed in Example 4 above. In one example, the sterilization preferably includes: (A) 140° C., 100 cycles of microwave; (B) 100° C., 100 cycles of microwave; and (C) room temperature, 100 cycles with no microwaves. The Prion Protein Indicator Test having the defined surrogate protein samples were analyzed by Western Blot analysis as described in Example 4 above. Depending on treatment conditions, the test sample was completely destroyed (A), partially destroyed (B), or completely intact (C).

Testing infectiveness of Prion Prep: C57BL/6 mice aged 4-5 weeks were divided into 4 cohorts, 10 mice per cohort, to receive the samples that correlated with the following Prion Protein Indicator Test samples having the defined prion surrogate protein: (A) complete destruction; (B) partial destruction; (C) no treatment and (D) PBS control. After anesthesia each mouse was intracerebrally inoculated with a 20 ul-aliquot of the designated inoculum. The mice were observed up to one year after inoculation, unless they displayed terminal symptoms of PrP infection including persistent signs of ataxia, kyphosis, somnolence, and hind leg weakness. Terminally-ill mice were euthanized and their

brains divided sagittally along the midline and place formalin fixation for histological analysis or flash-frozen in liquid nitrogen for protein analysis. At one year, all remaining mice (showed no obvious signs of neurologic disease) were euthanized and their brains also divided for histological analysis or protein analysis.

Results: Over the observation period, none of the mice that received either the PBS control (D) or the brain homogenate treated with the conditions that demonstrated complete destruction on the Prion Protein Indicator Test having the defined prion surrogate protein (A) demonstrated any symptoms of disease. After euthanization, none of the brains demonstrated any signs of prion disease. Western Blot analysis of the brains showed no increase in concentrations of PrP proteins over normal levels. The mice that received the brain homogenate and also received no sterilization treatment (C) or partial destruction (B) as indicated by the Prion Protein Indicator Test all demonstrated terminal symptoms of PrP infection before the completion of the 1 year observation period. Their brains demonstrated obvious signs of prion disease and by Western analysis, the concentration of PrP proteins were greatly increased over normal levels. Together the results of the mouse study indicated a clear correlation between the results from the Prion Protein Indicator Test results and the mouse model results.

Example 6: Comparing Amino Acid Sequence of Multiple Members of the *Clostridium* Genus

It is important to qualify the ability of a sterilization device, a deimmunization device, or a disinfection device to destroy bacteria of any *Clostridium* species that may be contaminating medical equipment or supplies. In this example, a defined quantity of the defined surrogate protein having the predetermined SEQ ID NO: 1 discussed above was subjected to sterilization, deimmunization, or disinfection to rapidly determine the effectiveness of the sterilization, deimmunization, or disinfection using Western Blot analysis, protein array analysis, or similar type analysis, and the antibody for the defined *Clostridium* surrogate protein shown above. In this example, to protect the human operators of the test, the defined surrogate protein needs to incorporate critical characteristics of *Clostridium* proteins that are critical for the survival and growth of members of the *Clostridium* genus while avoiding organisms that can infect humans.

To design the synthetic defined surrogate, *Clostridium* protein having the predetermined sequence, SEQ ID NO: 1, a protein analysis was conducted comparing the amino acid sequences of the suf I loci gene from multiple species of the *Clostridium* genus shown in Table 2 above. In this example, the Sequence IDs (found in Pubmed, www.ncbi.nlm.nih.gov/Pubmed/) for the suf I loci proteins, multiple *Clostridium* species, used for the comparative are shown in Table 5 below:

TABLE 5

(Clostridium). Sequences used to determine regions of high homology in suf I locus of multiple Clostridium Species.	
Species	Sequence ID
<i>Clostridium sporogenes</i>	WP_061905762.1
<i>Clostridium botulinum</i>	WP_011948579.1
<i>Clostridium novyi</i>	WP_039217212.1

The protein produced by the suf I loci is fundamental to survival and growth of a wide range of spores, bacteria and fungus. In species of the *Clostridium* genus, the protein product of the suf I loci is called cotA and is critical for many live stages including strongly contributing to the stability of the spore coat.

To design the defined *Clostridium* surrogate protein having SEQ ID NO: 1, the specific amino acids from the proteins listed in Table 5 were aligned to determine amino acid sequence regions that are highly homologous in all evaluated *Clostridium* species as shown in FIG. 7. The protein encoded by the suf I loci includes three cupredoxin domains that are indicated at 200 for domain 1, 202 for domain 2, and 204 for domain 3. In the *Clostridium* genus, domain 2, indicated at 202, shows high homology between *Clostridium* species so was used to for the design of the synthetic surrogate protein SEQ ID NO: 1 above to be created for the *Clostridium* test and the corresponding peptides discussed above were used to develop the polyclonal and monoclonal antibodies for use by Western Blot analysis.

Example 7: Developing Western Test to Qualify Ability to Destroy *Clostridium* Test Protein

One purpose of developing the synthetic defined *Clostridium* surrogate protein is to provide a method for rapidly determining the effectiveness of sterilization, deimmunization, and/or disinfection by a device, such as sterilization device, deimmunization device, or disinfection device. In this example, the method for rapidly determining effective sterilization, deimmunization, and/or disinfection includes multiple steps including at least: 1) preparing the synthetic defined *Clostridium* surrogate protein test samples, 2) subjecting the defined *Clostridium* surrogate protein test samples to sterilization, deimmunization, or disinfection, and 3) using Western Blot or similar type analysis to visualize the effects of sterilization, deimmunization, or disinfection of defined *Clostridium* surrogate protein test samples. Successful sterilization, deimmunization, or disinfection has occurred when all the defined *Clostridium* surrogate protein test samples are fragmented and as a result of the protein fragmentation, none remains to bind to the visualization antibodies indicating the defined *Clostridium* surrogate protein was destroyed. If sterilization, deimmunization, or disinfection was not successful, protein bands will be seen on the Western Blot analysis.

Following the process more fully described for the Prion test, a similar process was followed to create the *Clostridium* sterilization, deimmunization, and/or disinfection test. First, to create the sample for the test using the defined *Clostridium* surrogate protein, DNA encoding the amino acid SEQ ID NO: 1 for *Clostridium* above was synthesized and cloned into standard vectors both for *E. coli* and yeast expression. Using standard techniques, large quantities of protein were produced in *E. coli* or yeast and isolated by standard recombinant methods. Using a nickel column, a full-length defined *Clostridium* surrogate protein (171 amino acids long) was isolated. To create the samples having the defined *Clostridium* surrogate protein to qualify sterilization, deimmunization, or disinfection of the samples were dried onto small filter papers, dried inside small tubes or a surface of an object subjected to sterilization, deimmunization, or disinfection e.g., using cycles of applying a solvent and microwave energy as disclosed in the '469 patent application.

After sterilization, deimmunization, or disinfection, the treated *Clostridium* samples were transferred to tubes, denatured and separated by size and transferred to nylon membrane before being permanently cross-linked to the membrane. The final steps include incubating the nylon membrane with a primary antibody that specifically binds to the defined *Clostridium* surrogate protein of interest. In the experiments discussed herein, the primary anti-suf I loci encoded protein antibody was a rabbit polyclonal antibody that was raised against a synthetic peptide discussed above for SEQ ID NO: 1 residues. For added sensitivity, addition of antibodies, both monoclonal and polyclonal, were raised against the other synthetic peptide(s). For the Western Blot Analysis, a secondary antibody may be a HRP-labelled goat anti-rabbit to enable visualization of the protein fragments, both intact and fragmented. If the defined *Clostridium* surrogate proteins have completely fragmented no bands will be visualized on the Western blot. Very small fragments and amino acids will be too small to be retained on the gel. When successful sterilization, deimmunization, or disinfection occurred, the visualized Western blot has a dark band in the untreated control sample, e.g., indicated at **210**, FIG. 8, and a complete absence of any bands for the defined *Clostridium* surrogate protein sample subjected to sterilization, deimmunization, or disinfection indicates successful sterilization, deimmunization, or disinfection, e.g., indicated at **212**.

Example 8: Comparing Amino Acid Sequence of Multiple Members of the *Bacillus* Genus

It is important to qualify the ability of a sterilization device, a deimmunization device or disinfection device to destroy bacteria of any *Bacillus* species that may be contaminating medical equipment or supplies. In this example, a defined quantity of the defined *Bacillus* surrogate protein having the predetermined SEQ ID NO: 2 discussed above is subjected to sterilization, deimmunization, or disinfection to rapidly determine the effectiveness of the sterilization, deimmunization, or disinfection using Western Blot analysis, protein array analysis, or similar type analysis, and the antibody for the defined *Bacillus* surrogate protein shown above. In this example, to protect the human operators of the test, the defined surrogate *Bacillus* surrogate protein needs to incorporate critical characteristics of *Bacillus* proteins that are critical for the survival and growth of members of the *Bacillus* genus while avoiding organisms that can infect humans.

To design the synthetic defined surrogate *Bacillus* protein having SEQ ID NO: 2, a protein analysis was conducted comparing the amino acid sequences of the suf I loci gene from multiple species of the *Bacillus* genus shown in Table 2 above. In this example, the Sequence IDs (found in Pubmed, www.ncbi.nlm.nih.gov/Pubmed/) for the suf I loci encoded proteins, multiple *Bacillus* species, used for the comparative are shown in Table 6 below.

TABLE 6

(Bacillus). Sequences used to determine regions of high homology in suf I locus of multiple Bacillus Species.	
Species	Sequence ID
<i>Bacillus subtilis</i>	AAB62305.1
<i>Bacillus atrophaeus</i>	WP_011948579.1
<i>Bacillus pumilus</i>	WP_039217212.1

The protein produced by the suf I loci is fundamental to survival and growth of a wide range of spores, bacteria and fungus. In species of the *Bacillus* genus, the protein product of the suf I loci is called cotA and is critical for many live stages including strongly contributing to the stability of the spore coat.

To design the synthetic defined *Bacillus* surrogate protein for SEQ ID NO: 2, the specific amino acids from the proteins listed in Table 6 were aligned to determine amino acid sequence regions that are highly homologous in all evaluated *Bacillus* species as shown in FIG. 9. The protein encoded by the suf 0.1 loci includes three cupredoxin domains, indicated at **214** for domain 1, **216** for domain 2, and **218** for domain 3. In the *Bacillus* genus, domain 2, indicated at **216**, shows high homology between *Bacillus* species, so SEQ ID NO: 2 above was used as the synthetic defined *Bacillus* surrogate protein to be used for the *Bacillus* test and the corresponding peptide discussed above was used to raise a polyclonal antibody for use by Western Blot analysis. Additional polyclonal and monoclonal antibodies were created as needed.

Example 9: Developing Western Test to Qualify Ability to Destroy *Bacillus* Test Protein

One purpose of developing the synthetic defined *Bacillus* surrogate protein is to provide a method for rapidly determining the effectiveness of sterilization, deimmunization, and/or disinfection by a device, such as a sterilization device, deimmunization device, or disinfection device. The method for rapidly determining effective sterilization, deimmunization, and/or disinfection preferably includes multiple steps including at least: 1) preparing synthetic defined *Bacillus* surrogate protein test samples, 2) subjecting the *Bacillus* surrogate protein test samples to sterilization, deimmunization, or disinfection, and 3) using Western Blot or similar analysis to visualize the effects of sterilization, deimmunization, or disinfection of defined *Bacillus* surrogate protein test samples. Successful sterilization, deimmunization, or disinfection has occurred when all the defined *Bacillus* surrogate protein test samples are fragmented and as a result of the protein fragmentation, none remains to bind to the visualization antibodies indicating the defined *Bacillus* surrogate protein was destroyed. If sterilization, deimmunization, or disinfection was not successful, protein bands will be seen on the Western Blot analysis.

Following the process more fully described for the Prion test, a similar process was followed to create the *Bacillus* sterilization, deimmunization, and/or disinfection test. First, to create the sample for the test using the synthetic defined *Bacillus* surrogate protein, DNA encoding the amino acid for SEQ ID NO: 2 above was synthesized and cloned into standard vectors both for *E. coli* and yeast expression. Using standard techniques, large quantities of the defined *Bacillus* surrogate protein were produced in *E. coli* or yeast and isolated by standard recombinant methods. Using a nickel column, a full-length defined *Bacillus* surrogate protein (171 amino acids long) was isolated. To create the samples having the synthetic defined *Bacillus* surrogate protein to qualify sterilization, deimmunization, and/or disinfection the samples were dried onto small filter papers, dried inside small tubes or a surface of an object subjected to sterilization, deimmunization, or disinfection, e.g., using cycles of applying a solvent and microwave energy, e.g., as disclosed in the '469 patent application.

After sterilization deimmunization and/or disinfection, the treated *Bacillus* samples were transferred to tubes,

denatured and separated by size and transferred to nylon membrane before being permanently cross-linked to the membrane. The final steps include incubating the nylon membrane with a primary antibody that specifically binds to the defined *Bacillus* surrogate proteins. In the experiments discussed herein, the primary anti-suf I loci protein antibody was a rabbit polyclonal antibody that was raised against a synthetic peptide discussed above for SEQ ID NO: 2 residues. For added sensitivity, addition antibodies, both monoclonal and polyclonal, were raised against the other synthetic peptide(s). For the Western Blot Analysis, a secondary antibody may be a HRP-labelled goat anti-rabbit to enable visualization of the protein fragments, both intact and fragmented. If the defined *Bacillus* surrogate proteins have completely fragmented no bands will be visualize on the Western blot. Very small fragments and amino acids will be too small to be retained on the gel. When successful sterilization, deimmunization, or disinfection occurred, the visualized Western blot has a dark ban in the untreated control sample, e.g., indicated at 220, FIG. 10, and a complete absence of any bands for defined *Bacillus* surrogate protein sample subjected to sterilization, deimmunization, or disinfection indicates successful sterilization, deimmunization, or disinfection, indicated at 222.

Example 10: Comparing Amino Acid Sequence of Multiple Members of the *Mycobacterium* Genus

It is important to qualify the ability of a sterilization device, a deimmunization device, or a disinfection device to destroy bacteria of any *Mycobacterium* species that may be contaminating medical equipment or supplies. In this example, a defined quantity of the defined surrogate protein having a SEQ ID NO: 3 discussed above is subjected to sterilization, deimmunization, or disinfection to rapidly determine the effectiveness of the sterilization, deimmunization, or disinfection using Western Blot analysis, protein array analysis, or similar type analysis and using an antibody specific for the protein. In this example, to protect the human operators of the test, the defined *Mycobacterium* surrogate protein needs to incorporate critical characteristics of *Mycobacterium* proteins that are critical for the survival and growth of members of the *Mycobacterium* genus while avoiding organisms that can infect humans.

To design the synthetic defined *Mycobacterium* surrogate protein SEQ ID NO: 3 above, a protein analysis was conducted comparing the amino acid sequences of the suf I loci gene from multiple species of the *Mycobacterium* genus shown in Table 2 above. In this example, the Sequence IDs (found in Pubmed, www.ncbi.nlm.nih.gov/Pubmed/) for the suf 1 loci proteins, multiple *Mycobacterium* species, used for the comparative are shown in Table 7 below:

TABLE 7

(Mycobacteria). Sequences used to determine regions of high homology in suf 1 locus of multiple <i>Mycobacterium</i> Species.	
Species	Sequence ID
<i>Mycobacterium tuberculosis</i>	WP_003404392.1
<i>Mycobacterium africanum</i>	KBG17039.1
<i>Mycobacterium kansasii</i>	WP_023367763.1

The protein produced by the suf I loci is fundamental to survival and growth of a wide range of spores, bacteria and fungus. In species of the *Mycobacterium* genus, the protein

product of the suf I loci is called cumA and is critical for many live stages including cell survival and growth.

To design the synthetic defined *Mycobacterium* surrogate protein having SEQ ID NO: 3, the specific amino acids from the proteins listed in Table 7 were aligned to determine amino acid sequence regions that are highly homologous in all evaluated *Mycobacterium* species as shown in FIG. 11. The protein encoded by the suf 1 loci includes three cupredoxin domains that are indicated at 230 for domain 1, 232 for domain 2, and 234 for domain 3. In the *Mycobacterium* genus, domain 2, indicated at 232, shows high homology between *Mycobacterium* species so SEQ ID NO: 3 above was used to for the design for synthetic defined *Mycobacterium* surrogate protein to be created for the *Mycobacterium* test and the corresponding peptide for the defined *Mycobacterium* surrogate protein discussed above was used for a polyclonal antibody for use by Western Blot analysis.

Example 11: Developing Western Test to Qualify Ability to Destroy *Mycobacterium* Test Protein

One purpose of developing the defined *Mycobacterium* surrogate protein is to provide a method for rapidly determining the effectiveness of sterilization, deimmunization, and/or disinfection by a device, such as a sterilization device, a deimmunization device, or a disinfection device. The method for rapidly determining effective sterilization, deimmunization, and/or disinfection preferably includes multiple steps including at least: 1) preparing the defined *Mycobacterium* surrogate protein test samples, 2) subjecting the defined *Mycobacterium* surrogate protein test samples to sterilization, deimmunization, or disinfection, and 3) using Western Blot or similar analysis to visualize the effects of sterilization, deimmunization, and/or disinfection of synthetic defined *Mycobacterium* surrogate protein test samples. Successful sterilization, deimmunization, and/or disinfection has occurred when all the synthetic defined *Mycobacterium* surrogate protein test samples are fragmented and as a result of the protein fragmentation, none remains to bind to the visualization antibodies indicating the defined *Mycobacterium* protein was destroyed. If sterilization, deimmunization, and/or disinfection was not successful, protein bands will be see on the Western Blot analysis.

Following the process more fully described for the Pion test, a similar process was followed to create the *Mycobacterium* sterilization, deimmunization, and/or disinfection test. First, to create the sample for the test using the defined *Mycobacterium* surrogate protein, DNA encoding the amino acid SEQ ID NO: 3 above was synthesized and cloned into standard vectors both for *E. coli* and yeast expression. Using standard techniques, large quantities of protein were produced in *E. coli* or yeast and isolated by standard recombinant methods. Using a nickel column, a full-length synthetic *Mycobacterium* surrogate protein (171 amino acids long) was isolated. To create the samples having the *Mycobacterium* surrogate protein to qualify sterilization, deimmunization, and/or disinfection the samples were dried onto small filter papers, dried inside small tubes or a surface of an object subjected to sterilization, deimmunization, and/or disinfection, e.g., using cycles of applying a solvent and microwave energy as disclosed in the '469 patent application.

After sterilization, deimmunization, and/or disinfection the treated *Mycobacterium* surrogate protein samples were transferred to tubes, denatured and separated by size and transferred to nylon membrane before being permanently cross-linked to the membrane. The final steps include incu-

bating the nylon membrane with a primary antibody that specifically binds to the protein of interest. In the experiments discussed herein, the primary anti-suf I loci protein antibody was a rabbit polyclonal antibody that was raised against a synthetic peptide discussed above for SEQ ID NO: 3 for *Mycobacterium* above residues. For added sensitivity, addition antibodies, both monoclonal and polyclonal, were raised against the other synthetic peptide(s). For the Western Blot Analysis, a secondary antibody may be a HRP-labelled goat anti-rabbit to enable visualization of the protein fragments, both intact and fragmented. If the *Mycobacterium* surrogate proteins have completely fragmented no bands will be visualize on the Western blot. Very small fragments and amino acids will be too small to be retained on the gel. When successful sterilization, deimmunization, and/or disinfection has occurred, the visualized Western blot has a dark band in the untreated control sample, e.g., indicated at 236, FIG. 12, and a complete absence of any bands for the *Mycobacterium* surrogate protein sample subjected to sterilization, deimmunization, or disinfection indicates successful sterilization, deimmunization, or disinfection, e.g., indicated at 238.

Example 12: Comparing Amino Acid Sequence of Multiple Members of the *Staphylococcus* Genus

It is important to qualify the ability of a sterilization device, a deimmunization device, or a disinfection device to destroy bacteria of any *Staphylococcus* species that may be contaminating medical equipment or supplies. In this example, a defined quantity of the defined *Staphylococcus* surrogate protein having the predetermined SEQ ID NO: 4 discussed above is subjected to sterilization, deimmunization, or disinfection to rapidly determine the effectiveness of the sterilization, deimmunization, or disinfection using Western Blot analysis, protein array analysis, or similar type analysis and the antibody specific for the defined *Staphylococcus* surrogate protein. In this example, to protect the human operators of the test, the synthetic defined *Staphylococcus* surrogate protein needs to incorporate critical characteristics of *Staphylococcus* proteins that are critical for the survival and growth of members of the *Staphylococcus* genus while avoiding organisms that can infect humans.

To design the synthetic defined *Staphylococcus* surrogate protein having SEQ ID NO: 4, a protein analysis was conducted comparing the amino acid sequences of the suf I loci gene from multiple species of the *Staphylococcus* genus shown in Table 2 above. In this example, the Sequence IDs (found in Pubmed, www.ncbi.nlm.nih.gov/Pubmed/) for the suf I loci proteins, multiple *Staphylococcus* species, used for the comparative are shown in Table 8 below:

TABLE 8

(Staphylococcus). Sequences used to determine regions of high homology in Suf I locus of multiple <i>Staphylococcus</i> Species.	
Species	Sequence ID
<i>Staphylococcus aureus</i>	WP_000282432.1
<i>Staphylococcus epidermidis</i>	WP_023567454.1
<i>Staphylococcus saprophyticus</i>	OEK13316.1

The protein produced by the suf I loci is fundamental to survival and growth of a wide range of spores, bacteria and fungus. In species of the *Staphylococcus* genus, the protein

product of the suf I loci is called cueO and is critical for many live stages including strongly contributing to cell survival and growth

To design the synthetic defined *Staphylococcus* surrogate protein, the specific amino acids from the proteins listed in Table 8 were aligned to determine amino acid sequence regions that are highly homologous in all evaluated *Staphylococcus* species as shown in FIG. 13. The protein encoded by the suf I loci includes three cupredoxin domains that are indicated at 240 for domain 1, 242 for domain 2, and 244 for domain 3. In the *Staphylococcus* genus, domain 2 indicated at 242 shows high homology between *Staphylococcus* species, so SEQ ID NO: 4 above was used to for the design of the synthetic defined *Staphylococcus* surrogate protein to be created for the *Staphylococcus* test and the corresponding peptide discussed above was used for a polyclonal antibody for use by Western Blot analysis.

Example 13: Developing Western Test to Qualify Ability to Destroy *Staphylococcus* Test Protein

One purpose of developing the defined *Staphylococcus* surrogate protein is to provide a method for rapidly determining the effectiveness of sterilization, deimmunization, and/or disinfection device, such as sterilization device, deimmunization device, or disinfection device and/or supplies. The method for rapidly determining effective sterilization, deimmunization, and/or disinfection includes multiple steps including at least: 1) preparing synthetic defined *Staphylococcus* surrogate protein test samples, 2) subjecting the defined *Staphylococcus* surrogate protein test samples to sterilization, deimmunization, and/or disinfection, and 3) using Western Blot or similar analysis to visualize the effects of sterilization, deimmunization, and/or disinfection of defined *Staphylococcus* surrogate protein test samples of the defined surrogate protein. Successful sterilization has occurred when all the defined *Staphylococcus* surrogate protein test samples are fragmented and as a result of the protein fragmentation, none remains to bind to the visualization antibodies indicating the defined *Staphylococcus* surrogate protein was destroyed. If sterilization, deimmunization, and/or disinfection was not successful, protein bands will be see on the Western Blot analysis.

Following the process more fully described for the Prion test, a similar process was followed to create the *Staphylococcus* sterilization, deimmunization, or disinfection test. First to create the sample for the test using the synthetic defined *Staphylococcus* surrogate protein, DNA encoding the amino acid SEQ ID NO: 4 above was synthesized and cloned into standard vectors both for *E. coli* and yeast expression. Using standard techniques, large quantities of the defined *Staphylococcus* surrogate protein were produced in *E. coli* or yeast and isolated by standard recombinant methods. Using a nickel column, a full-length defined *Staphylococcus* surrogate protein (171 amino acids long) was isolated. To create the samples having the synthetic defined *Staphylococcus* surrogate protein to qualify sterilization, deimmunization, and/or disinfection the samples were dried onto small filter papers, dried inside small tubes or a surface of an object subjected to sterilization, deimmunization, and/or disinfection e.g., using cycles of applying a solvent and microwave energy as disclosed in the '469 patent application.

After sterilization, deimmunization, and/or disinfection, the treated defined *Staphylococcus* surrogate protein samples were transferred to tubes, denatured and separated by size and transferred to nylon membrane before being

permanently cross-linked to the membrane. The final steps include incubating the nylon membrane with a primary antibody that specifically binds to the *Staphylococcus* surrogate protein of interest. In the experiments discussed herein, the primary anti-suf I loci protein antibody was a rabbit polyclonal antibody that was raised against a synthetic peptide discussed above for SEQ ID NO: 4 residues. For added sensitivity, addition antibodies, both monoclonal and polyclonal, were raised against the other synthetic peptide(s). For the Western Blot analysis, a secondary antibody may be a HRP-labelled goat anti-rabbit to enable visualization of the protein fragments, both intact and fragmented. If the defined *Staphylococcus* surrogate proteins have completely fragmented no bands will be visualize on the Western blot. Very small fragments and amino acids will be too small to be retained on the gel. When successful sterilization, deimmunization, and/or disinfection occurred, the visualized Western blot has a dark band in the untreated control sample, indicated at **250**, FIG. **14**, and a complete absence of any bands for defined *Staphylococcus* surrogate protein sample subjected to sterilization, deimmunization, or disinfection indicates successful sterilization, deimmunization, or disinfection, indicated at **252**.

Example 14: Comparing Amino Acid Sequence of Multiple Members of the *Pseudomonas* Genus

It is important to qualify the ability of a sterilization device, a deimmunization device or disinfection device to destroy bacteria of any *Pseudomonas* species that may be contaminating medical equipment or supplies. In this example, a defined *Pseudomonas* surrogate protein quantity of the defined surrogate protein having a predetermined SEQ ID NO: 5 discussed above is subjected to sterilization, deimmunization, or disinfection to rapidly determine the effectiveness of the sterilization, deimmunization, or disinfection using Western Blot analysis, protein array analysis, or similar type analysis, and using an antibody specific for the defined *Pseudomonas* surrogate protein shown above. In this example, to protect the human operators of the test, the defined *Pseudomonas* surrogate protein needs to incorporate critical characteristics of *Pseudomonas* proteins that are critical for the survival and growth of members of the *Pseudomonas* genus while avoiding organisms that can infect humans.

To design the synthetic defined *Pseudomonas* surrogate protein having SEQ ID NO: 5, a protein analysis was conducted comparing the amino acid sequences of the suf loci gene from multiple species of the *Pseudomonas* genus shown in Table 2 above. In this example, the Sequence IDs (found in Pubmed, www.ncbi.nlm.nih.gov/Pubmed/) for the suf I loci Table 9 below:

TABLE 9

<i>Pseudomonas</i> . Sequences used to determine regions of high homology in suf I locus of multiple <i>Pseudomonas</i> Species.	
Species	Sequence ID
<i>Pseudomonas aeruginosa</i>	WP_023096478.1
<i>Pseudomonas fluorescens</i>	WP_003227851.1
<i>Pseudomonas putida</i>	WP_019750583.1

The protein produced by the suf I loci is fundamental to survival and growth of a wide range of spores, bacteria and fungus. In species of the *Pseudomonas* genus, the protein

product of the suf I loci is called cumA and is critical for many live stages including strongly contributing to the cell survival and growth

To design the defined *Pseudomonas* surrogate protein, the specific amino acids from the proteins listed in Table 9 were aligned to determine amino acid sequence regions that are highly homologous in all evaluated *Pseudomonas* species as shown in FIG. **15**. The protein encoded by the suf I loci includes three cupredoxin domains indicated at **260** for domain 1, **262** for domain 2, and **264** for domain 3. In the *Pseudomonas* genus, domain 2, indicated at **262**, shows high homology between *Pseudomonas* species so SEQ ID NO: 5 above was used to for the design of the synthetic surrogate protein to be created for the defined *Pseudomonas* surrogate protein test and the corresponding peptide shown above was used for a polyclonal antibody for use by Western Blot analysis.

Example 15: Developing Western Test to Qualify Ability to Destroy *Pseudomonas* Test Protein

One purpose of developing the defined *Pseudomonas* surrogate protein test is to create a method for rapidly determining the effectiveness of sterilization, deimmunization, and/or disinfection by a device, such as a sterilization device, a deimmunization device, or a disinfection device. The method for rapidly determining effective sterilization, deimmunization, and/or disinfection preferably includes multiple steps including at least: 1) preparing synthetic defined *Pseudomonas* surrogate protein test samples, 2) subjecting the synthetic *Pseudomonas* surrogate protein test samples to sterilization, deimmunization, and/or disinfection, and 3) using Western Blot or similar analysis to visualize the effects of sterilization, deimmunization, and/or disinfection of synthetic defined *Pseudomonas* surrogate test samples. Successful sterilization has occurred when all the defined *Pseudomonas* surrogate protein test samples are fragmented and as a result of the protein fragmentation, none remains to bind to the visualization antibodies indicating the defined *Pseudomonas* surrogate protein was destroyed. If sterilization, deimmunization, and/or disinfection was not successful, protein bands will be seen on the Western Blot analysis.

Following the process more fully described for the Prion test, a similar process was followed to create the defined *Pseudomonas* sterilization, deimmunization and/or disinfection test. First, to create the sample for the test using the defined *Pseudomonas* surrogate protein, DNA encoding the amino acid SEQ ID NO: 5 defined above was synthesized and cloned into standard vectors both for *E. coli* and yeast expression. Using standard techniques, large quantities of the defined *Pseudomonas* surrogate protein were produced in *E. coli* or yeast and isolated by standard recombinant methods. Using a nickel column, a full-length synthetic defined *Pseudomonas* surrogate protein (171 amino acids long) was isolated. To create the samples having the synthetic defined surrogate protein to qualify sterilization, deimmunization, and/or disinfection of the samples were dried onto small filter papers, dried inside small tubes or a surface of an object subjected to sterilization, deimmunization, and/or disinfection, e.g., using cycles of applying a solvent and microwave energy as disclosed in the '469 patent application.

After sterilization, deimmunization, and/or disinfection, the treated defined *Pseudomonas* surrogate protein samples were transferred to tubes, denatured and separated by size and transferred to nylon membrane before being perma-

nently cross-linked to the membrane. The final steps include incubating the nylon membrane with a primary antibody that specifically binds to the defined *Pseudomonas* surrogate protein. In the experiments discussed herein, the primary anti-suf 1 loci protein antibody was a rabbit polyclonal antibody that was raised against a synthetic peptide discussed above for SEQ ID NO: 5 residues. For added sensitivity, addition antibodies, both monoclonal and polyclonal, were raised against the other synthetic peptide(s). For the Western Blot Analysis, a secondary antibody may be a HRP-labelled goat anti-rabbit to enable visualization of the protein fragments, both intact and fragmented. If the synthetic defined *Pseudomonas* surrogate proteins have completely fragmented no bands will be visualize on the Western blot. Very small fragments and amino acids will be too small to be retained on the gel. When successful sterilization, deimmunization, or disinfection occurred, the visualized Western blot has a dark ban in the untreated control sample, e.g., indicated at **266**, FIG. **16**, and a complete absence of any bands for the defined *Pseudomonas* surrogate protein sample subjected to sterilization, deimmunization, or disinfection indicates successful sterilization, deimmunization, or disinfection, indicated at **268**.

Example 16: Comparing Amino Acid Sequence of Multiple Members of the *Trichophyton* Genus

It is important to qualify the ability of a sterilization device, a deimmunization device, and/or a disinfection device to destroy bacteria of any *Trichophyton* species that may be contaminating medical equipment. In this example, a defined quantity of the defined *Trichophyton* surrogate protein having a predetermined SEQ ID NO: 6 discussed above is subjected to sterilization, deimmunization, or disinfection to rapidly determine the effectiveness of the sterilization, deimmunization, or disinfection using Western Blot analysis, protein array analysis, or similar type analysis and using the antibody specific for the defined *Trichophyton* surrogate protein. In this example, to protect the human operators of the test, the synthetic defined *Trichophyton* surrogate protein needs to incorporate critical characteristics of *Trichophyton* proteins that are critical for the survival and growth of members of the *Trichophyton* genus while avoiding organisms that can infect humans.

To design the synthetic defined *Trichophyton* surrogate protein having the SEQ ID NO: 6, a protein analysis was conducted comparing the amino acid sequences of the suf I loci gene from multiple species of the *Trichophyton* genus shown in Table 2 above. In this example, the Sequence IDs (found in Pubmed, www.ncbi.nlm.nih.gov/Pubmed/) for the suf I loci proteins, multiple *Trichophyton* species, used for the comparative are shown in Table 10 below:

TABLE 10

<i>Trichophyton</i> . Sequences used to determine regions of high homology in suf I locus of <i>Trichophyton</i> Species.	
Species	Sequence ID
<i>Trichophyton rubrum</i>	XP_003236812.1
<i>Trichophyton tonsurane</i>	EGD95875.1

The protein produced by the suf 1 loci is fundamental to survival and growth of a wide range of spores, bacteria and fungus. In species of the *Trichophyton* genus, the protein

product of the suf 1 loci is called laccase and is critical for many live stages including strongly contributing to cell survival and growth

To design the synthetic defined *Trichophyton* surrogate protein, the specific amino acids from the proteins listed in Table 10 above were aligned to determine amino acid sequence regions that are highly homologous in all evaluated *Trichophyton* species as shown in FIG. **17**. The protein encoded by the suf 1 loci includes three cupredoxin domains that are indicated at **270** for domain 1, **272** for domain 2, and **274** for domain 3. In the *Trichophyton* genus, domain 2, indicated at **272**, shows high homology between *Trichophyton* species so SEQ ID NO: 6 above was used to for the design of the synthetic defined *Trichophyton* surrogate protein to be created for the *Trichophyton* test and the corresponding peptide discussed above was used for a polyclonal antibody for use by Western Blot analysis.

Example 17: Developing Western Test to Qualify Ability to Destroy *Trichophyton* Test Protein

One purpose of developing the defined *Trichophyton* surrogate protein is to provide a method for rapidly determining the effectiveness of sterilization, deimmunization, and/or disinfection by a device, such as sterilization device, a deimmunization device, or a disinfection device. The method for rapidly determining effective sterilization, deimmunization, or disinfection preferably includes multiple steps including at least: 1) preparing synthetic defined *Trichophyton* surrogate protein test samples, 2) subjecting the defined *Trichophyton* surrogate protein test samples to sterilization, deimmunization, or disinfection, and 3) using Western Blot or similar analysis to visualize the effects of sterilization, deimmunization, or disinfection of defined *Trichophyton* surrogate protein test samples. Successful sterilization, deimmunization, or disinfection has occurred when all the defined *Trichophyton* surrogate protein test samples are fragmented and as a result of the protein fragmentation, none remains to bind to the visualization antibodies indicating the defined *Trichophyton* surrogate protein was destroyed. If sterilization, deimmunization and/or disinfection was not successful, protein bands will be seen on the Western Blot analysis.

Following the process more fully described for the Prion test, a similar process was followed to create the *Trichophyton* sterilization, deimmunization and/or disinfection test. First, to create the sample for the test using the synthetic defined *Trichophyton* surrogate protein, DNA encoding the amino acid SEQ ID NO: 7 above was synthesized and cloned into standard vectors both for *E. coli* and yeast expression. Using standard techniques, large quantities of defined *Trichophyton* surrogate protein were produced in *E. coli* or yeast and isolated by standard recombinant methods. Using a nickel column, a full-length synthetic defined *Trichophyton* surrogate protein (171 amino acids long) was isolated. To create the samples having the synthetic defined *Trichophyton* surrogate protein to qualify sterilization, deimmunization, or disinfection, the defined *Trichophyton* surrogate protein samples were dried onto small filter papers, dried inside small tubes or a surface of an object subjected to sterilization, deimmunization and/or disinfection, e.g., using cycles of applying a solvent and microwave energy as disclosed in the '469 patent application.

After sterilization, deimmunization, or disinfection, the treated *Trichophyton* samples were transferred to tubes, denatured and separated by size and transferred to nylon membrane before being permanently cross-linked to the

membrane. The final steps include incubating the nylon membrane with a primary antibody that specifically binds to the protein of interest. In the experiments discussed herein, the primary anti-suf 1 loci protein antibody was a rabbit polyclonal antibody that was raised against a synthetic peptide discussed above for SEQ ID NO: 6 residues. For added sensitivity, addition antibodies, both monoclonal and polyclonal, were raised against the other synthetic peptide(s). For the Western Blot analysis, a secondary antibody may be a HRP-labelled goat anti-rabbit to enable visualization of the protein fragments, both intact and fragmented. If the proteins have completely fragmented no bands will be visualize on the Western blot. Very small fragments and amino acids will be too small to be retained, on the gel. When successful sterilization, deimmunization, or disinfection has occurred, the visualized Western blot has a dark ban in the untreated control sample, e.g., indicated at 276, FIG. 18, and a complete absence of any bands for the defined *Trichophyton* surrogate protein sample subjected to sterilization, deimmunization, or disinfection indicates successful sterilization, deimmunization, or disinfection, indicated at 278.

Example 18: Comparing Amino Acid Sequence of Multiple Members of the *Candida Genus*

It is important to qualify the ability of a sterilization device, a deimmunization device, or disinfection device, to destroy bacteria of any *Candida* species that may be contaminating medical equipment. In this example, a defined quantity of the defined *Candida* surrogate protein having a predetermined SEQ ID NO: 7 discussed above is subjected to sterilization, deimmunization, or disinfection to rapidly determine the effectiveness of the sterilization, deimmunization, or disinfection using Western Blot analysis, protein array analysis, or similar type analysis and the antibody specific for the defined *Candida* surrogate protein. In this example, to protect the human operators of the test, the defined *Candida* surrogate protein needs to incorporate critical characteristics of *Candida* proteins that are critical for the survival and growth of members of the *Candida* genus while avoiding organisms that can infect humans.

To design the synthetic defined *Candida* surrogate protein having the SEQ ID NO: 7, a protein analysis was conducted comparing the amino acid sequences of the suf 1 loci gene from multiple species of the *Candida* genus shown in Table 2 above. In this example, the Sequence IDs (found in Pubmed, www.ncbi.nlm.nih.gov/Pubmed/) for the suf 1 loci proteins, multiple *Candida* species, used for the comparative are shown in Table 11 below:

TABLE 11

(Candida). Sequences used to determine regions of high homology in Suf I locus of Candida Species.	
Species	Sequence ID
<i>Candida albicans</i>	KHC71512.1
<i>Candida dubliniensis</i>	XP_002420841.1
<i>Candida tropicalis</i>	XP_002548698.1
<i>Candida auris</i>	XP_018169615.1

The protein produced by the suf I loci is fundamental to survival and growth of a wide range of spores, bacteria and fungus. In species of the *Candida* genus, the protein product of the suf I loci is called laccase and is critical for many live stages including strongly contributing to cell survival and growth

To design the synthetic defined *Candida* surrogate protein, the specific amino acids from the proteins listed in Table 11 were aligned to determine amino acid sequence regions that are highly homologous in all evaluated *Candida* species as shown in FIG. 19. The protein encoded by the suf 1 loci includes three cupredoxin domains, indicated at 280 for domain 1, 282 for domain 2, and 284 for domain 3. In the *Candida* genus, domain 2, indicated at 284, shows high homology between *Candida* species so SEQ ID NO: 7 above was used to for the design of the synthetic defined *Candida* surrogate protein to be created for the *Candida* test and the corresponding peptide discussed above was used for a polyclonal antibody for use by Western Blot analysis.

Example 19: Developing Western Test to Qualify Ability to Destroy *Candida* Test Protein

One purpose of developing the defined *Candida* surrogate protein test is to provide a method for rapidly determining the effectiveness of sterilization, deimmunization, and/or disinfection by a device, such as a sterilization device, a deimmunization device, or a disinfection device. The method for rapidly determining effective sterilization, deimmunization, and/or disinfection includes multiple steps preferably including at least: 1) preparing synthetic defined *Candida* surrogate protein test samples, 2) subjecting the defined *Candida* surrogate protein test samples to sterilization, deimmunization, or disinfection, and 3) using Western Blot or similar analysis to visualize the effects of sterilization, deimmunization, or disinfection of defined *Candida* surrogate protein test samples. Successful sterilization, deimmunization, or disinfection has occurred when all the defined *Candida* surrogate protein test samples are fragmented and as a result of the protein fragmentation, none remains to bind to the visualization antibodies indicating the defined *Candida* surrogate protein was destroyed. If sterilization, deimmunization, or disinfection was not successful, protein bands will be see on the Western Blot analysis.

Following the process more fully described for the Prion test, a similar process was followed to create the defined *Candida* surrogate protein sterilization, deimmunization and/or disinfection test. First, to create the sample for the synthetic defined *Candida* surrogate protein test using the defined *Candida* surrogate protein, DNA encoding the amino acid SEQ ID NO: 7 was synthesized and cloned into standard vectors both for *E. coli* and yeast expression. Using standard techniques, large quantities of protein were produced in *E. coli* or yeast and isolated by standard recombinant methods. Using a nickel column, a full-length synthetic defined *Candida* surrogate protein (171 amino acids long) was isolated. To create the samples having the synthetic defined *Candida* surrogate protein to qualify sterilization, deimmunization, or disinfection, the samples having the defined *Candida* surrogate protein were dried onto small filter papers, dried inside small tubes or a surface of an object subjected to sterilization, deimmunization, and/or disinfection, e.g., using cycles of applying a solvent and microwave energy as discussed in the '469 patent application.

After sterilization, deimmunization, or disinfection, the treated defined *Candida* surrogate protein samples were transferred to tubes, denatured and separated by size and transferred to nylon membrane before being permanently cross-linked to the membrane. The final steps include incubating the nylon membrane with a primary antibody that specifically binds to the defined *Candida* surrogate protein. In the experiments discussed herein, the primary anti-suf 1

loci protein antibody was a rabbit polyclonal antibody that was raised against a synthetic peptide discussed above for SEQ ID NO: 7 residues. For added sensitivity, addition of antibodies, both monoclonal and polyclonal, were raised against the other synthetic peptide(s). For the Western Blot analysis, a secondary antibody may be a HRP-labelled goat anti-rabbit to enable visualization of the protein fragments, both intact and fragmented. If the proteins have completely fragmented no bands will be visualized on the Western blot. Very small fragments and amino acids will be too small to be retained on the gel. When successful sterilization, deimmunization, or disinfection has occurred, the visualized Western blot has a dark band in the untreated control sample, indicated at **286**, FIG. **20**, and a complete absence of any bands for the defined *Candida* surrogate protein sample subjected to sterilization, deimmunization, or disinfection indicates successful sterilization, deimmunization, or disinfection, indicated at **290**.

Example 20: Immobilizing Proteins onto a Solid Surface

Many processes and solid substrates are well known in the art for immobilizing proteins onto a solid surface. In this example, glass slides were obtained having round wells created by printing the glass with highly water-repellent mark, e.g., as shown by multi-well glass slides **130**, **134**, FIG. **2**, or multi-well glass slide **152**, FIG. **3**. In this example, the glass slides were washed with acetone and milli-Q water before soaking overnight in 1 M NaOH (room temperature). To create amino silane-treated slides, washed slides using milli-Q water and 99.5% ethanol, were treated with 3-aminopropyltriethoxysilane for 2 hours at room temperature, washed with milli-Q water, and baked for 2 hr. at 100° C. The multi-well slides were soaked in 1% (v/v) glutaraldehyde overnight at 37° C. to produce glutaraldehyde-treated slides. The slides were then incubated overnight at 37° C. in chitosan (0.05% (w/v)) dissolved in 0.1 M acetic acid buffer (pH 5.0) supplemented with 0.25 mM sodium azide. The treated slides were rinsed twice with 0.1 M acetic acid buffer (pH 5) and then three times with water before being soaked in 1% (v/v) glutaraldehyde overnight at 37° C. The slides were incubated overnight at 37° C. in a 0.05% (w/v) solution of N-(5-Amino-1-carboxy pentyl) iminodiacetic acid (AB-NTA) in 0.1 M HEPES buffer (pH 8.0), then rinsed 3 times with milli-Q water. The slides were then soaked in blocking solution (1% (v/v) glycine) for 1 h at 37° C., then rinsed 3 times with milli-Q water, 3 times with 0.5 M NiCl₂, and 3 times with milli-Q water to produce Ni-NTA immobilized slides ready for protein immobilization.

In this example, the defined *Clostridium* surrogate protein having SEQ ID NO: 1 was diluted in TG buffer (50 mM Tris-HCl (pH 8.0), 10% (v/v) glycerol) to 200 µg/ml. Different quantities of the defined *Clostridium* surrogate protein, e.g., 200 ng, 100 ng, 50 ng, 25 ng, 12.5 ng, 6.25 ng, 3.125 ng, 1.562 ng, 0.781 ng and 0 ng, were spotted in individual wells, e.g., the wells exemplarily indicated at **132** and **136**, FIG. **2**, and incubated for 60 min in a moist, dark chamber and finished by washing with TG buffer. After subjecting the slides **130** and **134** to sterilization, deimmunization, or disinfection using cycles of a solvent and electromagnetic radiation, e.g., microwaves as disclosed in the '469 patent application, slides **130** and **134** were incubated with anti-*Clostridium* antibodies discussed above raised against the peptide selected from SEQ ID NO: 1. To visualize the amount of the defined *Clostridium* surrogate protein remaining after sterilization, deimmunization, or

disinfection, the slides were then treated with anti-Ig antibody labeled with the fluorescence dye fluorescein isothiocyanate (FITC). A matched slide that was not sterilized in this example, slide **130**, was used as a control for the protein array analysis. Using a fluorescence microscope with camera, the loss of protein can be visualized that can be detected with the specific anti-*Clostridium* antibodies. With additional fluorescence detection equipment, such as automated readers, increasing sensitivity and titration of detection can be added. An example of successful sterilization, deimmunization and/or disinfection is indicated at **140**, for slide **134**, FIG. **2**, when compared against control slide **130** which visibly shows color, indicated by the shading in wells **132**. Confirmatory tests were conducted by including samples of intact *C. sporogenes* using cycles of a solvent and electromagnetic energy or microwaves, e.g., as disclosed in the '469 patent application. After treatment, the intact bacteria were processed following industry standard protocols. Similar methods may be used for any of Examples 6 to 18 above.

Example 21: Comparing Amino Acid Sequence of α-Gliadin from Appropriate Human Consumed Grains

To develop a test to qualify the ability of a sterilization device, deimmunization, or disinfection to destroy medically important immunogenic proteins, such as α-gliadin, that may be contaminating medical equipment, the following experiments were conducted. In this example, to rapidly detect effective deimmunization, a defined quality of an isolated protein needs to be deimmunized with a deimmunization device, e.g., using cycles of a combination of a solvent and electromagnetic microwaves radiation, e.g., microwaves, as disclosed in the '469 patent application and then evaluated by Western Blot analysis using an antibody specific for the protein. To design a candidate protein to serve as the representative defined surrogate α-gliadin protein having a predetermined sequence, the translated sequences of representative α-gliadin genes from many commonly consumed human grains were aligned. See Table 13 and Table 14 below. Gliadin protein is the immunogenic component of gluten and must be avoided by Celiac patients. FIG. **21** shows the amino acid numbering aligned with an example of Bread wheat α-Gliadin. The underline region, indicated at **400**, is the polypeptide that acts as common immunogen in a majority of Celiac Patients. Regions at **402**, **404** indicate protein region used to design test immunogen reagent.

TABLE 13

Sequence ID of α-Gliadin Proteins or Prolamins in Common Grains Used to Design Recombinant Protein Sequence.

Common Name	Species Name	Sequence ID Number w/Link
Bread Wheat	<i>Triticum aestivum</i>	pir A27319
Common Wheat	<i>Triticum sphaerococcum</i>	ABQ45316.1
Durum wheat/Pasta Wheat	<i>Triticum turgidum</i> subsp. <i>durum</i>	ADA83698.1
Farro/Emmer Wheat	<i>Triticum dicoccon</i>	AKC91191.1
Macha Wheat	<i>Triticum macha</i>	AKC91223.1
Rye	<i>Secale cereale</i>	AFK32718.1
Spelt/Dinkel Wheat	<i>Triticum spelta</i>	APU92351.1
Red Wild Einkor	<i>Triticum urartu</i>	AKC91171.1
Precursor αGliadin Bateman et al 2004	<i>Triticum aestivum</i>	Q41545

TABLE 14

Additional α -Gliadin Protein Sequences that could be used to design Recombinant Protein.

Species	Common	Sequence ID	Species	Common Name	Sequence ID
<i>Triticum aestivum</i>	Bread Wheat	pir A27319 P04726.1	<i>Triticum turgidum</i> subsp. <i>durum</i>	Durum wheat, pasta wheat or macaroni wheat	ADA83698.1 ADA83690.1
		AED99851.1	<i>Triticum dicoccon</i>	Emmer wheat or Farro	AKC91191.1
		SCW25751.1	<i>Triticum urartu</i>	red wild einkor	AKC91171.1
		AHY37812.1	<i>Triticum macha</i>	Macha wheat	AKC91223.1
		AHY37818.1	<i>Secale cereale</i>	Rye	AFK32718.1
		AFX69612.1			
		ABQ52118.1	<i>Aegilops tauschii</i>	Tausch's goatgrass	AFX69602.1 XP_020186089.1
		AFX69616.1			AKC91337.1
		AFX69586.1			ABQ52112.1
<i>Triticum sphaerococcum</i>	Common wheat	ABQ45316.1			
<i>Triticum spelta</i>	Spelt or dinkel wheat	APU92351.1	<i>Aegilops sharonensis</i>	Sharon goatgrass	AMS25611.1 AMS25610.1 AMS25614.1
		APU92675.1			AKC91312.1
		APU92554.1	<i>Aegilops searsii</i>	Goatgrass	AKC91311.1
		APU92300.1			AHN85624.1
		APU92425.1	<i>Aegilops speltoides</i>	Goatgrass	AHN85626.1
		APU92357.1	<i>Aegilops speltoides</i>	Goatgrass	AEV55370.1
		APU92415.1	<i>Aegilops univariata</i>	Goatgrass	ADP94197.1
		APU92583.1	<i>Thinopyrum bessarabicum</i>	Wild <i>Thinopyrum</i> grasses	
		APU92336.1			
		APU92334.1			

In this example, the defined α -Gliadin surrogate protein has the following predetermined sequence:
For α -Gliadin:

35 application. After treatment, the intact wheat flour samples were processed following industry standard protocols to test for the presence of gluten in food stuffs.

(SEQ ID NO: 8)

10 20 30 40 50

MKTVRVVPVQ PQQNPSQPQ PQRQVPLVQQ QQFPGQQQF PPQQYPQPQ

PFPSQQPYLQ LQPFPPQPF PPQLPYHHH HH

The peptide used for the development of monoclonal or polyclonal antibody used by Western Blot analysis for the above sequence is:

45 Although specific features of the invention are shown in some drawings and not in others, this is for convenience only as each feature may be combined with any or all of the other features in accordance with the invention. The words "including", "comprising", "having", and "with" as used herein are to be interpreted broadly and comprehensively and are not limited to any physical interconnection. Moreover, any embodiments disclosed in the subject application are not to be taken as the only possible embodiments. Other embodiments will occur to those skilled in the art and are within the following claims.

(SEQ ID NO: 24)

50 FPPQQYPQPFPFPSQQPYLQLQPFPPQPQ

A fragment of this peptide (KLQPFPPQPELPYPQPQ (SEQ ID NO: 25)) in form is the medically important immunogen is CD.

If the defined α -Gliadin surrogate proteins have completely fragmented, no bands will be visualized on the Western Blot. Very small fragments in amino acid would be too small to be retained on the gel. When successful deimmunization, in this example, has occurred, the visualized Western Blot has a dark band in the untreated control sample, e.g., indicated at 420, FIG. 22, and a complete absence of any bands for the defined α -Gliadin surrogate protein sample subject to deimmunization indicates successful deimmunization, e.g., indicated at 422.

55 In addition, any amendment presented during the prosecution of the patent application for this patent is not a disclaimer of any claim element presented in the application as filed: those skilled in the art cannot reasonably be expected to draft a claim that would literally encompass all possible equivalents, many equivalents will be unforeseeable at the time of the amendment and are beyond a fair interpretation of what is to be surrendered (if anything), the rationale underlying the amendment may bear no more than a tangential relation to many equivalents, and/or there are many other reasons the applicant cannot be expected to describe certain insubstantial substitutes for any claim element amended.

65 Confirmatory tests were conducted by including samples of intact wheat flour using a combination of cycles of a solvent and microwaves, e.g., as disclosed in the '469 patent

SEQUENCE LISTING

<160> NUMBER OF SEQ ID NOS: 63

<210> SEQ ID NO 1

<211> LENGTH: 171

<212> TYPE: PRT

<213> ORGANISM: Clostridium sporogenes

<400> SEQUENCE: 1

```

Met Tyr Asn Tyr Thr Ser Ala Lys Tyr Glu Val Pro Ile Ala Ile Gln
1           5           10           15
Asp Arg Ser Phe Asn Glu Asp Gly Ser Leu Asn Phe Pro Ser Glu Gly
          20           25           30
Asp Asn Pro Thr Ile His Pro Tyr Trp Gln Pro Glu Phe Phe Gly Asp
          35           40           45
Thr Ile Met Val Asn Gly Arg Val Trp Pro Asn Met Asn Val Asp Met
          50           55           60
Thr Arg Tyr Arg Phe Arg Leu Leu Asn Gly Ser Asn Ala Arg Phe Tyr
          65           70           75           80
Asn Leu Lys Phe Ser Asn Gly Met Gln Phe Trp Gln Ile Gly Thr Asp
          85           90           95
Gly Gly Tyr Leu Asn Lys Pro Val Pro Leu Thr Ser Leu Leu Ile Ser
          100          105          110
Pro Gly Glu Arg Ala Asp Ile Leu Val Asp Phe Thr Glu Ile Pro Ala
          115          120          125
Gly Thr Arg Ile Ile Leu Asn Asn Asp Ala Asn Ala Pro Tyr Pro Thr
          130          135          140
Gly Asp Ala Pro Asp Lys Asp Thr Thr Gly Gln Ile Met Gln Phe Thr
          145          150          155          160
Val Gln His Asn Asp His His His His His His
          165          170

```

<210> SEQ ID NO 2

<211> LENGTH: 169

<212> TYPE: PRT

<213> ORGANISM: Bacillus subtilis

<400> SEQUENCE: 2

```

Met Thr Leu Glu Lys Thr Tyr Tyr Glu Val Thr Met Glu Glu Cys Thr
1           5           10           15
His Gln Leu His Arg Asp Leu Pro Pro Thr Arg Leu Trp Gly Tyr Asn
          20           25           30
Gly Leu Phe Pro Gly Pro Thr Ile Glu Val Lys Arg Asn Glu Asn Val
          35           40           45
Tyr Val Lys Trp Met Asn Asn Leu Pro Ser Thr His Phe Leu Pro Ile
          50           55           60
Asp His Thr Ile His His Ser Asp Ser Gln His Glu Glu Pro Glu Val
          65           70           75           80
Lys Thr Val Val His Leu His Gly Gly Val Thr Pro Asp Asp Ser Asp
          85           90           95
Gly Tyr Pro Glu Ala Trp Phe Ser Lys Asp Phe Glu Gln Thr Gly Pro
          100          105          110
Tyr Phe Lys Arg Glu Val Tyr His Tyr Pro Asn Gln Gln Arg Gly Ala
          115          120          125
Ile Leu Trp Tyr His Asp His Ala Met Ala Leu Thr Arg Leu Asn Val
          130          135          140
Tyr Ala Gly Leu Val Gly Ala Tyr Ile Ile His Asp Pro Lys Glu Lys

```


-continued

<213> ORGANISM: *Pseudomonas aeruginosa*

<400> SEQUENCE: 5

Met Thr Gly Phe Arg His Glu Lys Val Leu Cys Leu Lys Thr Trp His
 1 5 10 15
 Val Asp Glu Gln Gly Ala Phe Thr Pro Phe Ser Val Pro Arg Gln Ala
 20 25 30
 Ala Arg Glu Gly Thr Arg Gly Arg Tyr Ser Thr Ile Asn Gly Lys His
 35 40 45
 Val Pro Thr Ile Asp Leu Pro Ala Gly Gln Ile Val Arg Val Arg Leu
 50 55 60
 Leu Asn Val Asp Asn Thr Val Thr Tyr Arg Leu Asn Pro Asn Gly Glu
 65 70 75 80
 Ala Arg Ile Tyr Ala Val Asp Gly His Pro Val Glu Pro Arg Gly Phe
 85 90 95
 Glu Gly Gln Tyr Trp Ile Gly Pro Gly Met Arg Leu Glu Leu Ala Leu
 100 105 110
 Lys Val Pro Glu Ala Gly Thr Glu Leu Ser Leu Arg Asp Gly Pro Val
 115 120 125
 Arg Leu Ala Thr Ile Arg Ser Val Ala His His His His His His
 130 135 140

<210> SEQ ID NO 6

<211> LENGTH: 133

<212> TYPE: PRT

<213> ORGANISM: *Trichophyton rubrum*

<400> SEQUENCE: 6

Met Thr Ile Thr Leu Glu Trp Ser Val Thr Thr Gly Tyr Arg Arg Leu
 1 5 10 15
 Asp Gly Val Lys Lys Arg Val Tyr Leu Ile Asn Gly Leu Phe Pro Gly
 20 25 30
 Pro Thr Ile Glu Ala Arg Ser Gly Asp Ser Leu Gln Val Gln Val Thr
 35 40 45
 Asn Asn Ile Gln Asp Glu Gly Leu Val Ile His Trp His Gly Leu His
 50 55 60
 Met Arg Gly Ala Asn His Met Asp Gly Val Thr Gly Val Thr Gln Cys
 65 70 75 80
 Pro Ile Val Pro Gly Asp Ser Met Leu Tyr Asn Phe Thr Ile Ser Gln
 85 90 95
 Ser Gln Ser Gly Thr Phe Trp Tyr His Ala His Ser Ala Leu Gln Arg
 100 105 110
 Ala Glu Gly Leu Tyr Gly Gly Phe Val Val His Lys Pro Ser Thr His
 115 120 125
 His His His His His
 130

<210> SEQ ID NO 7

<211> LENGTH: 133

<212> TYPE: PRT

<213> ORGANISM: *Candida albicans*

<400> SEQUENCE: 7

Met Thr Ala Glu Thr His Thr Trp Tyr Phe Lys Thr Ser Trp Val Asp
 1 5 10 15
 Ala Asn Pro Asp Gly Val Phe Pro Arg Lys Met Ile Gly Phe Asn Asp
 20 25 30

-continued

Ser Trp Pro Leu Pro Thr Leu Arg Val Lys Lys Gly Asp Thr Val Asn
 35 40 45
 Leu Tyr Leu Ile Asn Gly Phe Asp Asp Arg Asn Thr Ser Leu His Phe
 50 55 60
 His Gly Leu Phe Gln His Gly Thr Asn Gln Met Asp Gly Pro Glu Met
 65 70 75 80
 Val Thr Gln Cys Pro Ile Pro Pro Gly Glu Thr Phe Leu Tyr Asn Phe
 85 90 95
 Thr Val Asp Asp Gln Val Gly Ser Tyr Trp Tyr His Ser His Thr Ser
 100 105 110
 Gly Gln Tyr Gly Asp Gly Met Arg Gly Val Phe Ile Ile Glu Asp His
 115 120 125
 His His His His His
 130

<210> SEQ ID NO 8
 <211> LENGTH: 82
 <212> TYPE: PRT
 <213> ORGANISM: Unknown
 <220> FEATURE:
 <223> OTHER INFORMATION: surrogate protein sequence

<400> SEQUENCE: 8

Met Lys Thr Val Arg Val Pro Val Pro Gln Pro Gln Asn Pro
 1 5 10 15
 Ser Gln Pro Gln Pro Gln Arg Gln Val Pro Leu Val Gln Gln Gln Gln
 20 25 30
 Phe Pro Gly Gln Gln Gln Gln Phe Pro Pro Gln Gln Pro Tyr Pro Gln
 35 40 45
 Pro Gln Pro Phe Pro Ser Gln Gln Pro Tyr Leu Gln Leu Gln Pro Phe
 50 55 60
 Pro Gln Pro Gln Pro Phe Pro Pro Gln Leu Pro Tyr His His His His
 65 70 75 80
 His His

<210> SEQ ID NO 9
 <211> LENGTH: 208
 <212> TYPE: PRT
 <213> ORGANISM: Artificial Sequence
 <220> FEATURE:
 <223> OTHER INFORMATION: surrogate protein 9

<400> SEQUENCE: 9

Lys Lys Arg Pro Lys Pro Gly Gly Trp Asn Thr Gly Gly Ser Arg Tyr
 1 5 10 15
 Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Gly Gly
 20 25 30
 Thr Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly Gly
 35 40 45
 Ser Trp Gly Gln Pro His Gly Gly Ser Trp Gly Gln Pro His Gly Gly
 50 55 60
 Gly Trp Gly Gln Gly Gly Gly Thr His Asn Gln Trp Asn Lys Pro Ser
 65 70 75 80
 Lys Pro Lys Thr Asn Leu Lys His Val Ala Gly Ala Ala Ala Ala Gly
 85 90 95
 Ala Val Val Gly Gly Leu Gly Gly Tyr Met Leu Gly Ser Ala Met Ser
 100 105 110

-continued

Arg Pro Met Ile His Phe Gly Asn Asp Trp Glu Asp Arg Tyr Tyr Arg
 115 120 125

Glu Asn Met Tyr Arg Tyr Pro Asn Gln Val Tyr Tyr Arg Pro Val Asp
 130 135 140

Gln Tyr Ser Asn Gln Asn Asn Phe Val His Asp Cys Val Asn Ile Thr
 145 150 155 160

Ile Lys Gln His Thr Val Thr Thr Thr Thr Lys Gly Glu Asn Phe Thr
 165 170 175

Glu Thr Asp Val Lys Met Met Glu Arg Val Val Glu Gln Met Cys Val
 180 185 190

Thr Gln Tyr Gln Lys Glu Ser Gln Ala Tyr Tyr Asp Gly Arg Arg Ser
 195 200 205

<210> SEQ ID NO 10
 <211> LENGTH: 24
 <212> TYPE: PRT
 <213> ORGANISM: Clostridium sporogenes

<400> SEQUENCE: 10

Lys Tyr Glu Val Pro Ile Ala Ile Gln Asp Arg Ser Phe Asn Glu Asp
 1 5 10 15

Gly Ser Leu Asn Phe Pro Ser Glu
 20

<210> SEQ ID NO 11
 <211> LENGTH: 24
 <212> TYPE: PRT
 <213> ORGANISM: Clostridium sporogenes

<400> SEQUENCE: 11

Tyr Leu Asn Lys Pro Val Pro Leu Thr Ser Leu Leu Ile Ser Pro Gly
 1 5 10 15

Glu Arg Ala Asp Ile Leu Val Asp
 20

<210> SEQ ID NO 12
 <211> LENGTH: 24
 <212> TYPE: PRT
 <213> ORGANISM: Bacillus subtilis

<400> SEQUENCE: 12

Gln Arg Gly Ala Ile Leu Trp Tyr His Asp His Ala Met Ala Leu Thr
 1 5 10 15

Arg Leu Asn Val Tyr Ala Gly Leu
 20

<210> SEQ ID NO 13
 <211> LENGTH: 24
 <212> TYPE: PRT
 <213> ORGANISM: Bacillus subtilis

<400> SEQUENCE: 13

Gln Leu His Arg Asp Leu Pro Pro Thr Arg Leu Trp Gly Tyr Asn Gly
 1 5 10 15

Leu Phe Pro Gly Pro Thr Ile Glu
 20

<210> SEQ ID NO 14
 <211> LENGTH: 29
 <212> TYPE: PRT

-continued

<213> ORGANISM: Mycobacterium tuberculosis

<400> SEQUENCE: 14

Asp Thr Ala Phe Arg Ile Ala Leu Ala Gly His Ser Met Thr Val Thr
 1 5 10 15

His Thr Asp Gly Tyr Pro Val Ile Pro Thr Glu Val Asp
 20 25

<210> SEQ ID NO 15

<211> LENGTH: 25

<212> TYPE: PRT

<213> ORGANISM: Mycobacterium tuberculosis

<400> SEQUENCE: 15

Val Phe Pro Leu Val Ala Leu Ala Glu Gly Lys Asn Ala Leu Ala Arg
 1 5 10 15

Ala Leu Leu Ser Thr Gly Ala Gly Ser
 20 25

<210> SEQ ID NO 16

<211> LENGTH: 29

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 16

Asn Phe Ser Asn Thr Leu Gly Tyr Asn Gly Asn Leu Leu Gly Pro Thr
 1 5 10 15

Leu Lys Leu Lys Lys Gly Asp Lys Val Lys Ile Lys Leu
 20 25

<210> SEQ ID NO 17

<211> LENGTH: 23

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus aureus

<400> SEQUENCE: 17

Lys Phe Glu Val Asn Gln Asp Ser Ala Thr Leu Trp Tyr His Pro His
 1 5 10 15

Pro Ser Pro Asn Thr Ala Lys
 20

<210> SEQ ID NO 18

<211> LENGTH: 24

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas aeruginosa

<400> SEQUENCE: 18

Asp Leu Pro Ala Gly Gln Ile Val Arg Val Arg Leu Leu Asn Val Asp
 1 5 10 15

Asn Thr Val Thr Tyr Arg Leu Asn
 20

<210> SEQ ID NO 19

<211> LENGTH: 20

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas aeruginosa

<400> SEQUENCE: 19

Gln Tyr Trp Ile Gly Pro Gly Met Arg Leu Glu Leu Ala Leu Lys Val
 1 5 10 15

Pro Glu Ala Gly
 20

-continued

<210> SEQ ID NO 20
 <211> LENGTH: 24
 <212> TYPE: PRT
 <213> ORGANISM: Trichophyton rubrum

<400> SEQUENCE: 20

Tyr Arg Arg Leu Asp Gly Val Lys Lys Arg Val Tyr Leu Ile Asn Gly
 1 5 10 15

Leu Phe Pro Gly Pro Thr Ile Glu
 20

<210> SEQ ID NO 21
 <211> LENGTH: 23
 <212> TYPE: PRT
 <213> ORGANISM: Trichophyton rubrum

<400> SEQUENCE: 21

Thr Gln Cys Pro Ile Val Pro Gly Asp Ser Met Leu Tyr Asn Phe Thr
 1 5 10 15

Ile Ser Gln Ser Gln Ser Gly
 20

<210> SEQ ID NO 22
 <211> LENGTH: 23
 <212> TYPE: PRT
 <213> ORGANISM: Candida albicans

<400> SEQUENCE: 22

Gly Phe Asn Asp Ser Trp Pro Leu Pro Thr Leu Arg Val Lys Lys Gly
 1 5 10 15

Asp Thr Val Asn Leu Tyr Leu
 20

<210> SEQ ID NO 23
 <211> LENGTH: 22
 <212> TYPE: PRT
 <213> ORGANISM: Candida albicans

<400> SEQUENCE: 23

Trp Tyr Phe Lys Thr Ser Trp Val Asp Ala Asn Pro Asp Gly Val Phe
 1 5 10 15

Pro Arg Lys Met Ile Gly
 20

<210> SEQ ID NO 24
 <211> LENGTH: 29
 <212> TYPE: PRT
 <213> ORGANISM: Unknown
 <220> FEATURE:
 <223> OTHER INFORMATION: alpha-Gliadin

<400> SEQUENCE: 24

Phe Pro Pro Gln Gln Pro Tyr Pro Gln Pro Gln Pro Phe Pro Ser Gln
 1 5 10 15

Gln Pro Tyr Leu Gln Leu Gln Pro Phe Pro Gln Pro Gln
 20 25

<210> SEQ ID NO 25
 <211> LENGTH: 16
 <212> TYPE: PRT
 <213> ORGANISM: Unknown
 <220> FEATURE:

-continued

<223> OTHER INFORMATION: alpha-Gliadin

<400> SEQUENCE: 25

Lys Leu Gln Pro Phe Pro Gln Pro Glu Leu Pro Tyr Pro Gln Pro Gln
 1 5 10 15

<210> SEQ ID NO 26

<211> LENGTH: 204

<212> TYPE: PRT

<213> ORGANISM: Homo sapiens

<400> SEQUENCE: 26

Lys Lys Arg Pro Lys Pro Gly Gly Trp Asn Thr Gly Gly Ser Arg Tyr
 1 5 10 15

Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Gly Gly
 20 25 30

Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly
 35 40 45

Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly
 50 55 60

Gly Gly Trp Gly Gln Gly Gly Gly Thr His Ser Gln Trp Asn Lys Pro
 65 70 75 80

Ser Lys Pro Lys Thr Asn Met Lys His Met Ala Gly Ala Ala Ala Ala
 85 90 95

Gly Ala Val Val Gly Gly Leu Gly Gly Tyr Met Leu Gly Ser Ala Met
 100 105 110

Ser Arg Pro Ile Ile His Phe Gly Ser Asp Tyr Glu Asp Arg Tyr Tyr
 115 120 125

Arg Glu Asn Met His Arg Tyr Pro Asn Gln Val Tyr Tyr Arg Pro Met
 130 135 140

Asp Glu Tyr Ser Asn Gln Asn Asn Phe Val His Asp Cys Val Asn Ile
 145 150 155 160

Thr Ile Lys Gln His Thr Val Thr Thr Thr Thr Lys Gly Glu Asn Phe
 165 170 175

Thr Glu Thr Asp Val Lys Met Met Glu Arg Val Val Glu Gln Met Cys
 180 185 190

Ile Thr Gln Tyr Glu Arg Glu Ser Gln Ala Tyr Tyr
 195 200

<210> SEQ ID NO 27

<211> LENGTH: 204

<212> TYPE: PRT

<213> ORGANISM: Macaca mulatta

<400> SEQUENCE: 27

Lys Lys Arg Pro Lys Pro Gly Gly Trp Asn Thr Gly Gly Ser Arg Tyr
 1 5 10 15

Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Gly Gly
 20 25 30

Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly
 35 40 45

Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly
 50 55 60

Gly Gly Trp Gly Gln Gly Gly Gly Thr His Asn Gln Trp Asn Lys Pro
 65 70 75 80

Ser Lys Pro Lys Thr Ser Met Lys His Met Ala Gly Ala Ala Ala Ala
 85 90 95

-continued

Gly Ala Val Val Gly Gly Leu Gly Gly Tyr Met Leu Gly Ser Ala Met
 100 105 110
 Ser Arg Pro Ile Ile His Phe Gly Asn Asp Tyr Glu Asp Arg Tyr Tyr
 115 120 125
 Arg Glu Asn Met Tyr Arg Tyr Pro Asn Gln Val Tyr Tyr Arg Pro Val
 130 135 140
 Asp Gln Tyr Ser Asn Gln Asn Asn Phe Val His Asp Cys Val Asn Ile
 145 150 155 160
 Thr Ile Lys Gln His Thr Val Thr Thr Thr Thr Lys Gly Glu Asn Phe
 165 170 175
 Thr Glu Thr Asp Val Lys Met Met Glu Arg Val Val Glu Gln Met Cys
 180 185 190
 Ile Thr Gln Tyr Glu Lys Glu Ser Gln Ala Tyr Tyr
 195 200

<210> SEQ ID NO 28
 <211> LENGTH: 210
 <212> TYPE: PRT
 <213> ORGANISM: Felis catus

<400> SEQUENCE: 28

Lys Lys Arg Pro Lys Pro Gly Gly Gly Trp Asn Thr Gly Gly Ser Arg
 1 5 10 15
 Tyr Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Gly
 20 25 30
 Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His
 35 40 45
 Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His
 50 55 60
 Gly Gly Gly Gly Trp Gly Gln Gly Gly Ser His Ser Gln Trp Asn Lys
 65 70 75 80
 Pro Ser Lys Pro Lys Thr Asn Met Lys His Met Ala Gly Ala Ala Ala
 85 90 95
 Ala Gly Ala Val Val Gly Gly Leu Gly Gly Tyr Met Leu Gly Ser Ala
 100 105 110
 Met Ser Arg Pro Ile Ile His Phe Gly Asn Asp Tyr Glu Asp Arg Tyr
 115 120 125
 Tyr Arg Glu Asn Met Tyr Arg Tyr Pro Asn Gln Val Tyr Tyr Arg Pro
 130 135 140
 Val Asp Gln Tyr Ser Asn Gln Asn Asn Phe Val His Asp Cys Val Asn
 145 150 155 160
 Ile Thr Val Lys Gln His Thr Val Thr Thr Thr Thr Lys Gly Glu Asn
 165 170 175
 Phe Thr Glu Thr Asp Met Lys Ile Met Glu Arg Val Val Glu Gln Met
 180 185 190
 Cys Val Thr Gln Tyr Gln Lys Glu Ser Glu Ala Tyr Tyr Gln Arg Arg
 195 200 205
 Ala Ser
 210

<210> SEQ ID NO 29
 <211> LENGTH: 205
 <212> TYPE: PRT
 <213> ORGANISM: Canis familiaris

<400> SEQUENCE: 29

-continued

Lys Lys Arg Pro Lys Pro Gly Gly Gly Trp Asn Thr Gly Gly Ser Arg
 1 5 10 15
 Tyr Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Gly
 20 25 30
 Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His
 35 40 45
 Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His
 50 55 60
 Gly Gly Gly Gly Trp Gly Gln Gly Gly Thr His Ser Gln Trp Asn Lys
 65 70 75 80
 Pro Ser Lys Pro Lys Thr Asn Met Lys His Val Ala Gly Ala Ala Ala
 85 90 95
 Ala Gly Ala Val Val Gly Gly Leu Gly Gly Tyr Leu Leu Gly Ser Ala
 100 105 110
 Met Ser Arg Pro Ile Ile His Phe Gly Asn Asp Cys Glu Asp Arg Tyr
 115 120 125
 Tyr Arg Glu Asn Met Tyr Arg Tyr Pro Asn Gln Val Tyr Tyr Arg Ser
 130 135 140
 Val Asp Gln Tyr Asn Asn Gln Ser Thr Phe Val His Asp Cys Val Asn
 145 150 155 160
 Ile Thr Val Lys Gln His Thr Val Thr Thr Thr Thr Lys Gly Glu Asn
 165 170 175
 Phe Thr Glu Thr Asp Ile Lys Met Met Glu Arg Val Val Glu Gln Met
 180 185 190
 Cys Ile Thr Gln Tyr Gln Arg Glu Ser Glu Ala Tyr Tyr
 195 200 205

<210> SEQ ID NO 30
 <211> LENGTH: 213
 <212> TYPE: PRT
 <213> ORGANISM: Bos taurus

<400> SEQUENCE: 30

Lys Lys Arg Pro Lys Pro Gly Gly Gly Trp Asn Thr Gly Gly Ser Arg
 1 5 10 15
 Tyr Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Gly
 20 25 30
 Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His
 35 40 45
 Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His
 50 55 60
 Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Gly Trp Gly Gln Gly
 65 70 75 80
 Gly Thr His Gly Gln Trp Asn Lys Pro Ser Lys Pro Lys Thr Asn Met
 85 90 95
 Lys His Val Ala Gly Ala Ala Ala Gly Ala Val Val Gly Gly Leu
 100 105 110
 Gly Gly Tyr Met Leu Gly Ser Ala Met Ser Arg Pro Ile Ile His Phe
 115 120 125
 Gly Ser Asp Tyr Glu Asp Arg Tyr Tyr Arg Glu Asn Met His Arg Tyr
 130 135 140
 Pro Asn Gln Val Tyr Tyr Arg Pro Val Asp Gln Tyr Ser Asn Gln Asn
 145 150 155 160
 Asn Phe Val His Asp Cys Val Asn Ile Thr Val Lys Glu His Thr Val

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165 170 175

Thr Thr Thr Thr Lys Gly Glu Asn Phe Thr Glu Thr Asp Ile Lys Met
 180 185 190

Met Glu Arg Val Val Glu Gln Met Cys Ile Thr Gln Tyr Gln Arg Glu
 195 200 205

Ser Gln Ala Tyr Tyr
 210

<210> SEQ ID NO 31
 <211> LENGTH: 205
 <212> TYPE: PRT
 <213> ORGANISM: *Ovis aries*

<400> SEQUENCE: 31

Lys Lys Arg Pro Lys Pro Gly Gly Gly Trp Asn Thr Gly Gly Ser Arg
 1 5 10 15

Tyr Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Gly
 20 25 30

Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His
 35 40 45

Gly Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His
 50 55 60

Gly Gly Gly Gly Trp Gly Gln Gly Gly Ser His Ser Gln Trp Asn Lys
 65 70 75 80

Pro Ser Lys Pro Lys Thr Asn Met Lys His Val Ala Gly Ala Ala Ala
 85 90 95

Ala Gly Ala Val Val Gly Gly Leu Gly Gly Tyr Met Leu Gly Ser Ala
 100 105 110

Met Ser Arg Pro Ile Ile His Phe Gly Asn Asp Tyr Glu Asp Arg Tyr
 115 120 125

Tyr Arg Glu Asn Met Tyr Arg Tyr Pro Asn Gln Val Tyr Tyr Arg Pro
 130 135 140

Val Asp Arg Tyr Ser Asn Gln Asn Asn Phe Val His Asp Cys Val Asn
 145 150 155 160

Ile Thr Val Lys Gln His Thr Val Thr Thr Thr Thr Lys Gly Glu Asn
 165 170 175

Phe Thr Glu Thr Asp Ile Lys Ile Met Glu Arg Val Val Glu Gln Met
 180 185 190

Cys Ile Thr Gln Tyr Gln Arg Glu Ser Gln Ala Tyr Tyr
 195 200 205

<210> SEQ ID NO 32
 <211> LENGTH: 209
 <212> TYPE: PRT
 <213> ORGANISM: *Mus musculus*

<400> SEQUENCE: 32

Lys Lys Arg Pro Lys Pro Gly Gly Trp Asn Thr Gly Gly Ser Arg Tyr
 1 5 10 15

Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Gly Gly
 20 25 30

Gly Thr Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly
 35 40 45

Gly Ser Trp Gly Gln Pro His Gly Ser Gly Trp Gly Gln Pro His Gly
 50 55 60

Gly Gly Trp Gly Gln Gly Gly Gly Thr His Asn Gln Trp Asn Lys Pro

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65	70	75	80
Ser Lys Pro Lys Thr Asn Leu Lys His Val Ala Gly Ala Ala Ala Ala	85	90	95
Gly Ala Val Val Gly Gly Leu Gly Gly Tyr Met Leu Gly Ser Ala Met	100	105	110
Ser Arg Met Ile Ile His Phe Gly Asn Asp Tyr Glu Asp Arg Tyr Tyr	115	120	125
Arg Glu Asn Met Tyr Arg Tyr Pro Asn Gln Val Tyr Tyr Arg Pro Val	130	135	140
Asp Gln Tyr Ser Asn Gln Asn Asn Phe Val His Asp Cys Val Asn Ile	145	150	155
Thr Ile Lys Gln His Thr Val Thr Thr Thr Thr Lys Gly Glu Asn Phe	165	170	175
Thr Glu Thr Asp Val Lys Met Met Glu Arg Val Val Glu Gln Met Cys	180	185	190
Val Thr Gln Tyr Gln Lys Glu Ser Gln Ala Tyr Tyr Asp Gly Arg Arg	195	200	205

Ser

<210> SEQ ID NO 33
 <211> LENGTH: 209
 <212> TYPE: PRT
 <213> ORGANISM: Rattus norvegicus

<400> SEQUENCE: 33

Lys Lys Arg Pro Lys Pro Gly Gly Trp Asn Thr Gly Gly Ser Arg Tyr	1	5	10	15
Pro Gly Gln Gly Ser Pro Gly Gly Asn Arg Tyr Pro Pro Gln Ser Gly	20	25	30	
Gly Thr Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly	35	40	45	
Gly Gly Trp Gly Gln Pro His Gly Gly Gly Trp Gly Gln Pro His Gly	50	55	60	
Gly Gly Trp Ser Gln Gly Gly Gly Thr His Asn Gln Trp Asn Lys Pro	65	70	75	80
Ser Lys Pro Lys Thr Asn Leu Lys His Val Ala Gly Ala Ala Ala Ala	85	90	95	
Gly Ala Val Val Gly Gly Leu Gly Gly Tyr Met Leu Gly Ser Ala Met	100	105	110	
Ser Arg Met Leu Ile His Phe Gly Asn Asp Tyr Glu Asp Arg Tyr Tyr	115	120	125	
Arg Glu Asn Met Tyr Arg Tyr Pro Asn Gln Val Tyr Tyr Arg Pro Val	130	135	140	
Asp Gln Tyr Ser Asn Gln Asn Asn Phe Val His Asp Cys Val Asn Ile	145	150	155	160
Thr Ile Lys Gln His Thr Val Thr Thr Thr Thr Lys Gly Glu Asn Phe	165	170	175	
Thr Glu Thr Asp Val Lys Met Met Glu Arg Val Val Glu Gln Met Cys	180	185	190	
Val Thr Gln Tyr Gln Lys Glu Ser Gln Ala Tyr Tyr Asp Gly Arg Arg	195	200	205	

Ser

<210> SEQ ID NO 34

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<211> LENGTH: 601
 <212> TYPE: PRT
 <213> ORGANISM: Clostridium sporogenes
 <400> SEQUENCE: 34

Met Glu Val Asn Pro Leu Asp Pro Lys Leu Ile Pro Lys Tyr Thr Gln
 1 5 10 15
 Glu Leu Val Ile Pro Pro Ser Phe Leu Pro Thr Val Cys Thr Ser Ser
 20 25 30
 Val Ser Gly Ala Val Ser Tyr Asn Tyr Thr Val Thr Met Asn Gln Phe
 35 40 45
 Glu Gln Gln Ile Leu Pro Pro Glu Phe Asn Pro Thr Thr Val Trp Gly
 50 55 60
 Tyr Gly Gly Thr Ile Lys Asp Thr Ser Thr Gly Glu Glu Val Lys Phe
 65 70 75 80
 Gln Asn Ala Pro Gly Pro Thr Phe Glu Ala Val Arg Gly Ile Pro Val
 85 90 95
 Asn Val Lys Trp Val Asn Glu Ile Thr Ala Pro Tyr Ser Leu Ala Val
 100 105 110
 Asp Pro Thr Met His Trp Ala Asn Pro Asn Asn Thr Pro Met Pro Pro
 115 120 125
 Pro Pro Gly Gly Trp Pro Pro Phe Pro Pro Gly Val Pro Glu Ala Gln
 130 135 140
 Lys Asp Val Pro Leu Val Pro His Leu His Gly Gly Glu Gln Ala Ser
 145 150 155 160
 Met Phe Asp Gly Asn Pro Glu Ala Trp Trp Thr Ala Lys Gly Leu Lys
 165 170 175
 Gly Ser Arg Tyr Ile Thr Asp Thr Phe His Tyr Leu Asn Thr Gln Glu
 180 185 190
 Ser Thr Thr Leu Trp Tyr His Asp His Ala Leu Gly Val Thr Arg Leu
 195 200 205
 Asn Val Val Met Gly Leu Ala Gly Phe Tyr Ile Leu Arg Asp Pro Ala
 210 215 220
 Asn Pro Leu Asp Tyr Pro Gly Pro Leu Ile Thr Ser Ala Lys Tyr Glu
 225 230 235 240
 Val Pro Ile Ala Ile Gln Asp Arg Ser Phe Asn Glu Asp Gly Ser Leu
 245 250 255
 Asn Phe Pro Ser Glu Gly Asp Asn Pro Thr Ile His Pro Tyr Trp Gln
 260 265 270
 Pro Glu Phe Phe Gly Asp Thr Ile Met Val Asn Gly Arg Val Trp Pro
 275 280 285
 Asn Met Asn Val Asp Met Thr Arg Tyr Arg Phe Arg Leu Leu Asn Gly
 290 295 300
 Ser Asn Ala Arg Phe Tyr Asn Leu Lys Phe Ser Asn Gly Met Gln Phe
 305 310 315 320
 Trp Gln Ile Gly Thr Asp Gly Gly Tyr Leu Asn Lys Pro Val Pro Leu
 325 330 335
 Thr Ser Leu Leu Ile Ser Pro Gly Glu Arg Ala Asp Ile Leu Val Asp
 340 345 350
 Phe Thr Glu Ile Pro Ala Gly Thr Arg Ile Ile Leu Asn Asn Asp Ala
 355 360 365
 Asn Ala Pro Tyr Pro Thr Gly Asp Ala Pro Asp Lys Asp Thr Thr Gly
 370 375 380
 Gln Ile Met Gln Phe Thr Val Gln His Asn Asp Asp Met Thr Ile Pro

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385                390                395                400
Pro Glu Leu Pro Glu Lys Leu Arg Cys Glu Pro Val Pro Lys Leu Lys
                405                410                415
Ser Pro Cys Lys Arg Arg Val Leu Thr Leu Tyr Glu Ile Ala Gly Pro
                420                425                430
Asn Gly Pro Gln Met Val Thr Leu Asn Gly Gln Arg Trp Ala Asp Pro
                435                440                445
Val Ser Glu Leu Pro Val Val Gly Ser Thr Glu Glu Trp Asn Ile Val
                450                455                460
Asn Leu Thr Met Asp Ala His Pro Ile His Leu His Leu Val Gln Phe
                465                470                475                480
Lys Ile Ala Cys Arg Gln Ala Phe Asp Val Asp Ala Tyr Thr Asn Asp
                485                490                495
Trp Leu Asp Leu Asn Ser Asp Ile Gly Ser Pro Pro Trp Met Thr Thr
                500                505                510
Pro Lys Ala Leu Cys Pro Gly Ser Tyr Ile Thr Gly Asp Asp Gln Pro
                515                520                525
Pro Ala Ala Asn Glu Ala Gly Trp Lys Asp Thr Val Gln Ala Phe Pro
                530                535                540
Gly Glu Ile Thr Arg Ile Arg Val Arg Phe Ala Pro Gln Asp Val Lys
                545                550                555                560
Thr Ser Cys Pro Gly Glu Asn Leu Tyr Leu Phe Asp Pro Ser Lys Gly
                565                570                575
Pro Gly Tyr Val Trp His Cys His Ile Leu Asp His Glu Asp Asn Asp
                580                585                590
Met Met Arg Pro Tyr Arg Val Phe Pro
                595                600

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<210> SEQ ID NO 35
<211> LENGTH: 598
<212> TYPE: PRT
<213> ORGANISM: Clostridium botulinum

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<400> SEQUENCE: 35
Met Glu Val Asn Pro Leu Asp Pro Lys Leu Ile Pro Lys Tyr Thr Gln
1                5                10                15
Glu Leu Val Ile Pro Pro Ser Phe Leu Pro Thr Ile Cys Thr Ser Ser
                20                25                30
Val Ser Gly Ala Val Ser Tyr Asn Tyr Thr Val Thr Met Asn Gln Phe
                35                40                45
Glu Gln Gln Ile Leu Pro Pro Glu Phe Asn Pro Thr Thr Val Trp Gly
                50                55                60
Tyr Gly Gly Thr Ile Lys Asp Thr Ser Thr Gly Glu Glu Val Lys Phe
65                70                75                80
Gln Asn Ala Pro Gly Pro Thr Phe Glu Ala Val Arg Asp Ile Pro Ile
                85                90                95
Asn Val Lys Trp Val Asn Glu Ile Thr Ala Pro Tyr Ser Leu Ala Val
                100               105               110
Asp Pro Thr Ile His Trp Ala Asn Pro Asn Asn Thr Pro Met Thr Pro
                115               120               125
Pro Pro Gly Gly Trp Pro Pro Phe Pro Pro Gly Val Pro Glu Asp Gln
                130               135               140
Lys Asp Val Pro Leu Val Thr His Leu His Gly Gly Glu Gln Ala Ser
145                150                155                160

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Met Phe Asp Gly Asn Pro Glu Ala Trp Trp Thr Ala Lys Gly Leu Lys
165 170 175

Gly Pro Lys Tyr Ile Thr Asp Thr Phe His Tyr Pro Asn Val Gln Glu
180 185 190

Ser Thr Thr Leu Trp Tyr His Asp His Ala Leu Gly Val Thr Arg Leu
195 200 205

Asn Val Val Met Gly Leu Ala Gly Phe Tyr Ile Leu Arg Asp Pro Ala
210 215 220

Asn Pro Leu Asp Tyr Pro Gly Ser Leu Ile Thr Ser Ala Lys Tyr Glu
225 230 235 240

Val Pro Ile Ala Ile Gln Asp Arg Ser Phe Asn Glu Asp Gly Ser Leu
245 250 255

Asn Phe Pro Ser Glu Gly Asp Asn Pro Thr Ile His Pro Tyr Trp Gln
260 265 270

Pro Glu Phe Phe Gly Asp Thr Ile Met Val Asn Gly Arg Val Trp Pro
275 280 285

Asn Leu Asn Val Asp Met Thr Lys Tyr Arg Phe Arg Leu Leu Asn Gly
290 295 300

Ser Asn Ala Arg Phe Tyr Asn Leu Lys Phe Ser Asn Gly Met Gln Phe
305 310 315 320

Trp Gln Ile Gly Thr Asp Gly Gly Tyr Leu Asn Lys Pro Val Pro Leu
325 330 335

Thr Ser Leu Leu Ile Ser Pro Gly Glu Arg Ala Asp Ile Leu Val Asp
340 345 350

Phe Thr Glu Ile Pro Ala Gly Thr Lys Ile Ile Leu Asn Asn Asp Ala
355 360 365

Asn Ala Pro Tyr Pro Thr Gly Asp Ala Pro Asp Lys Asp Thr Thr Gly
370 375 380

Gln Ile Met Gln Phe Thr Val Gln Asp Asn Met Thr Met Pro Pro Glu
385 390 395 400

Leu Pro Glu Lys Leu Arg Cys Glu Pro Val Pro Lys Leu Gln Ser Pro
405 410 415

Cys Lys Lys Arg Val Leu Thr Leu Tyr Glu Ile Ala Gly Pro Asn Gly
420 425 430

Pro Gln Met Val Thr Leu Asn Gly Gln Thr Trp Ser Ala Pro Val Ser
435 440 445

Glu Leu Pro Val Val Gly Ser Thr Glu Glu Trp Asp Ile Val Asn Leu
450 455 460

Thr Met Asp Ala His Pro Ile His Leu His Leu Val Gln Phe Lys Ile
465 470 475 480

Ala Cys Arg Gln Ala Phe Asp Val Asn Ala Tyr Thr Glu Asp Trp Leu
485 490 495

Asp Leu Asn Ser Asp Ile Gly Ser Pro Pro Trp Met Thr Thr Pro Lys
500 505 510

Ala Leu Cys Pro Gly Ser Tyr Thr Ile Gly Asp Asn Gln Pro Pro Ala
515 520 525

Ala Asn Glu Ala Gly Trp Lys Asp Thr Ile Gln Ala Pro Pro Gly Glu
530 535 540

Ile Ser Arg Ile Arg Val Arg Phe Ala Pro Gln Asn Val Glu Cys Ser
545 550 555 560

Cys Pro Gly Glu Asn Leu Tyr Pro Phe Asp Pro Ser Lys Gly Pro Asp
565 570 575

Tyr Val Trp His Cys His Ile Leu Asp His Glu Asp Asn Asp Met Met

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Asp Asp Ser Asp Gly Tyr Pro Glu Ala Trp Phe Ser Lys Asp Phe Glu
 115 120 125
 Gln Thr Gly Pro Tyr Phe Lys Arg Glu Val Tyr His Tyr Pro Asn Gln
 130 135 140
 Gln Arg Gly Ala Ile Leu Trp Tyr His Asp His Ala Met Ala Leu Thr
 145 150 155 160
 Arg Leu Asn Val Tyr Ala Gly Leu Val Gly Ala Tyr Ile Ile His Asp
 165 170 175
 Pro Lys Glu Lys Arg Leu Lys Leu Pro Ser Asp Glu Tyr Asp Val Pro
 180 185 190
 Leu Leu Ile Thr Asp Arg Thr Ile Asn Glu Asp Gly Ser Leu Phe Tyr
 195 200 205
 Pro Ser Ala Pro Glu Asn Pro Ser Pro Ser Leu Pro Asn Pro Ser Ile
 210 215 220
 Val Pro Ala Phe Cys Gly Glu Thr Ile Leu Val Asn Gly Lys Val Trp
 225 230 235 240
 Pro Tyr Leu Glu Val Glu Pro Arg Lys Tyr Arg Phe Arg Val Ile Asn
 245 250 255
 Ala Ser Asn Thr Arg Thr Tyr Asn Leu Ser Leu Asp Asn Gly Gly Asp
 260 265 270
 Phe Ile Gln Ile Gly Ser Asp Gly Gly Leu Leu Pro Arg Ser Val Lys
 275 280 285
 Leu Asn Ser Phe Ser Leu Ala Pro Ala Glu Arg Tyr Asp Ile Ile Ile
 290 295 300
 Asp Phe Thr Ala Tyr Glu Gly Glu Ser Ile Ile Leu Ala Asn Ser Ala
 305 310 315 320
 Gly Cys Gly Gly Asp Val Asn Pro Glu Thr Asp Ala Asn Ile Met Gln
 325 330 335
 Phe Arg Val Thr Lys Pro Leu Ala Gln Lys Thr Lys Ala Glu Ser Arg
 340 345 350
 Ser Thr Ser Pro His Thr Leu Arg Tyr Ser Met Lys Asp Thr Asn Ile
 355 360 365
 Arg Thr Leu Lys Leu Ala Gly Thr Gln Asp Glu Tyr Gly Arg Pro Val
 370 375 380
 Leu Leu Leu Asn Asn Lys Arg Trp His Asp Pro Val Thr Glu Thr Pro
 385 390 395 400
 Lys Val Gly Thr Thr Glu Ile Trp Ser Ile Ile Asn Arg His Ala Glu
 405 410 415
 His Ile Leu Ile His Leu His Leu Val Ser Phe Arg Val Leu Asp Arg
 420 425 430
 Arg Pro Phe Asp Ile Ala Arg Tyr Gln Glu Ser Gly Glu Leu Ser Tyr
 435 440 445
 Thr Val Arg Cys Pro Ala Ala Ala Ser Glu Lys Gly Trp Lys Asp Thr
 450 455 460
 Ile Gln Ala His Ala Gly Glu Val Leu Arg Ile Ala Ala Thr Phe Gly
 465 470 475 480
 Pro Tyr Ser Gly Arg Tyr Val Trp His Cys His Ile Leu Glu His Glu
 485 490 495
 Asp Tyr Asp Met Met Arg Pro Met Asp Ile Thr Asp Pro His Lys
 500 505 510

<210> SEQ ID NO 38

<211> LENGTH: 512

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<212> TYPE: PRT

<213> ORGANISM: Bacillus atrophaeus

<400> SEQUENCE: 38

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Met Asn Leu Glu Lys Phe Ala Asp Met Leu Pro Ile Pro Glu Val Leu
1          5          10          15
Lys Pro His Gln Gln Thr Lys Glu Ser Thr Tyr Tyr Glu Val Thr Met
20          25          30
Lys Glu Phe Tyr Gln Lys Leu His Arg Asp Leu Pro Pro Thr Arg Leu
35          40          45
Trp Gly Tyr Asn Ser Leu Phe Pro Gly Pro Thr Ile Glu Val Asn Arg
50          55          60
Asn Glu Asn Val Gln Ile Lys Trp Met Asn Asp Leu Pro Ser Gln His
65          70          75          80
Phe Leu Pro Ile Asp His Thr Ile His His Ser Glu Gly His His Gln
85          90          95
Glu Pro Glu Val Lys Thr Val Val His Leu His Gly Gly Val Thr Pro
100         105         110
Tyr Asp Ser Asp Gly Tyr Pro Glu Ala Trp Phe Ser Lys Gly Phe Gln
115        120        125
Gln Thr Gly Pro Tyr Phe Ser Arg Glu Ile Tyr His Tyr Pro Asn Gln
130        135        140
Gln Arg Gly Ala Ile Leu Trp Tyr His Asp His Ala Met Ala Leu Thr
145        150        155        160
Arg Leu Asn Val Tyr Ala Gly Leu Ala Gly Val Tyr Ile Ile His Asp
165        170        175
Pro Lys Glu Lys Arg Leu Lys Leu Pro Ala Gly Glu Tyr Asp Val Pro
180        185        190
Leu Met Ile Met Asp Arg Thr Ile Asn Glu Asp Gly Ser Leu Phe Tyr
195        200        205
Pro Ser Gly Pro Glu Asn Pro Ser Pro Thr Leu Pro Thr Pro Ser Ile
210        215        220
Val Pro Ala Phe Cys Gly Asp Thr Ile Leu Val Asn Gly Lys Ala Trp
225        230        235        240
Pro Tyr Met Glu Val Glu Pro Arg Ala Tyr Arg Phe Arg Ile Val Asn
245        250        255
Ala Ser Asn Thr Arg Thr Tyr Asn Leu Ser Leu Asp Asn Gly Gly Glu
260        265        270
Phe Leu Gln Val Gly Ser Asp Gly Gly Leu Leu Pro Arg Ser Val Lys
275        280        285
Leu Ser Ser Ile Ser Leu Ala Pro Ala Glu Arg Phe Asp Ile Ile Ile
290        295        300
Asp Phe Ala Ala Phe Glu Gly Gln Ser Ile Val Leu Ala Asn Ser Glu
305        310        315        320
Gly Cys Gly Gly Asp Ala Asn Pro Glu Ser Asp Ala Asn Val Met Gln
325        330        335
Phe Arg Val Ile Lys Pro Leu Lys Glu Lys Asp Glu Ser Arg Lys Pro
340        345        350
Arg Phe Leu Thr Asn Leu Pro Pro Val Thr Asp Glu Lys Ile Gln Asn
355        360        365
Leu Arg Thr Leu Lys Leu Thr Gly Thr Gln Asp Glu Tyr Gly Arg Pro
370        375        380
Val Leu Leu Leu Asn Asn Lys Arg Trp Ser Asp Pro Val Thr Glu Ala
385        390        395        400

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Pro Lys Leu Gly Thr Ser Glu Ile Trp Ser Ile Ile Asn Pro Thr Arg
 405 410 415
 Gly Thr His Pro Ile His Leu His Leu Ile Ser Phe Arg Val Leu Asp
 420 425 430
 Arg Arg Pro Phe Asp Thr Ala Lys Tyr Ala Glu Thr Asn Val Ser Phe
 435 440 445
 Thr Gly Pro Ala Val Pro Pro Pro Pro Ser Glu Lys Gly Trp Lys Asp
 450 455 460
 Thr Val Gln Ser His Ala Gly Glu Val Ile Arg Ile Met Ala Lys Phe
 465 470 475 480
 Gly Pro Tyr Ser Gly Arg Tyr Val Trp His Cys His Ile Leu Glu His
 485 490 495
 Glu Asp Tyr Asp Met Met Arg Pro Met Asp Val Val Asp Pro Asn Gln
 500 505 510

<210> SEQ ID NO 39

<211> LENGTH: 509

<212> TYPE: PRT

<213> ORGANISM: Bacillus pumilus

<400> SEQUENCE: 39

Met Asn Leu Glu Lys Phe Val Asp Glu Leu Pro Ile Pro Glu Val Ala
 1 5 10 15
 Glu Pro Val Lys Lys Asn Pro Arg Gln Thr Tyr Tyr Glu Ile Ala Met
 20 25 30
 Glu Glu Val Phe Leu Lys Val His Arg Asp Leu Pro Pro Thr Lys Leu
 35 40 45
 Trp Thr Tyr Asn Gly Ser Leu Pro Gly Pro Thr Ile His Ala Asn Arg
 50 55 60
 Asn Glu Lys Val Lys Val Lys Trp Met Asn Lys Leu Pro Leu Lys His
 65 70 75 80
 Phe Leu Pro Val Asp His Thr Ile His Glu Gly His His Asp Glu Pro
 85 90 95
 Glu Val Lys Thr Val Val His Leu His Gly Gly Val Thr Pro Ala Ser
 100 105 110
 Ser Asp Gly Tyr Pro Glu Ala Trp Phe Ser Arg Asp Phe Glu Ala Thr
 115 120 125
 Gly Pro Phe Phe Glu Arg Glu Val Tyr Glu Tyr Pro Asn His Gln Gln
 130 135 140
 Ala Cys Thr Leu Trp Tyr His Asp His Ala Met Ala Leu Thr Arg Leu
 145 150 155 160
 Asn Val Tyr Ala Gly Leu Ala Gly Phe Tyr Leu Ile Ser Asp Ala Phe
 165 170 175
 Glu Lys Ser Leu Glu Leu Pro Ser Asp Asp Tyr Asp Ile Pro Leu Met
 180 185 190
 Ile Met Asp Arg Thr Phe Gln Glu Asp Gly Ser Leu Phe Tyr Pro Ser
 195 200 205
 Arg Pro Asn Asp Thr Pro Glu Asp Ser Asp Ile Pro Asp Pro Ser Ile
 210 215 220
 Val Pro Phe Phe Cys Gly Glu Thr Ile Leu Val Asn Gly Lys Val Trp
 225 230 235 240
 Pro Tyr Leu Glu Val Glu Pro Arg Lys Tyr Arg Phe Arg Ile Leu Asn
 245 250 255
 Ala Ser Asn Thr Arg Thr Tyr Glu Leu His Leu Asp Asn Asp Ala Thr

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Asp Met Asp Gly Thr Glu Pro Ala Thr Ala Asn Ile Gly Pro Gly Gly
 130          135          140

Asp Phe Thr Tyr Arg Phe Ser Val Pro Asp Pro Gly Thr Tyr Trp Ala
 145          150          155

His Pro His Val Gly Leu Gln Gly Asp His Gly Leu Tyr Leu Pro Val
          165          170          175

Val Val Asp Asp Pro Thr Glu Pro Gly His Tyr Asp Ala Glu Trp Ile
          180          185          190

Ile Ile Leu Asp Asp Trp Thr Asp Gly Ile Gly Lys Ser Pro Gln Gln
          195          200          205

Leu Tyr Gly Glu Leu Thr Asp Pro Asn Lys Pro Thr Met Gln Asn Thr
 210          215          220

Thr Gly Met Pro Glu Gly Glu Gly Val Asp Ser Asn Leu Leu Gly Gly
 225          230          235

Asp Gly Gly Asp Ile Ala Tyr Pro Tyr Tyr Leu Ile Asn Gly Arg Ile
          245          250          255

Pro Val Ala Ala Thr Ser Phe Lys Ala Lys Pro Gly Gln Arg Ile Arg
          260          265          270

Ile Arg Ile Ile Asn Ser Ala Ala Asp Thr Ala Phe Arg Ile Ala Leu
          275          280          285

Ala Gly His Ser Met Thr Val Thr His Thr Asp Gly Tyr Pro Val Ile
 290          295          300

Pro Thr Glu Val Asp Ala Leu Leu Ile Gly Met Ala Glu Arg Tyr Asp
 305          310          315

Val Met Val Thr Ala Ala Gly Gly Val Phe Pro Leu Val Ala Leu Ala
          325          330          335

Glu Gly Lys Asn Ala Leu Ala Arg Ala Leu Leu Ser Thr Gly Ala Gly
          340          345          350

Ser Pro Pro Asp Pro Gln Phe Arg Pro Asp Glu Leu Asn Trp Arg Val
          355          360          365

Gly Thr Val Glu Met Phe Thr Ala Ala Thr Thr Ala Asn Leu Gly Arg
 370          375          380

Pro Glu Pro Thr His Asp Leu Pro Val Thr Leu Gly Gly Thr Met Ala
 385          390          395

Lys Tyr Asp Trp Thr Ile Asn Gly Glu Pro Tyr Ser Thr Thr Asn Pro
          405          410          415

Leu His Val Arg Leu Gly Gln Arg Pro Thr Leu Met Phe Asp Asn Thr
          420          425          430

Thr Met Met Tyr His Pro Ile His Leu His Gly His Thr Phe Gln Met
          435          440          445

Ile Lys Ala Asp Gly Ser Pro Gly Ala Arg Lys Asp Thr Val Ile Val
 450          455          460

Leu Pro Lys Gln Lys Met Arg Ala Val Leu Val Ala Asp Asn Pro Gly
 465          470          475

Val Trp Val Met His Cys His Asn Asn Tyr His Gln Val Ala Gly Met
          485          490          495

Ala Thr Arg Leu Asp Tyr Ile Leu
          500

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<210> SEQ ID NO 41

<211> LENGTH: 396

<212> TYPE: PRT

<213> ORGANISM: Mycobacterium africanum

<400> SEQUENCE: 41

-continued

Met Thr Asn Arg Leu Gly Asp Pro Thr Ser Val His Trp His Gly Ile
 1 5 10 15
 Ala Leu Arg Asn Asp Met Asp Gly Thr Glu Pro Ala Thr Ala Asn Ile
 20 25 30
 Gly Pro Gly Gly Asp Phe Thr Tyr Arg Phe Ser Val Pro Asp Pro Gly
 35 40 45
 Thr Tyr Trp Ala His Pro His Val Gly Leu Gln Gly Asp His Gly Leu
 50 55 60
 Tyr Leu Pro Val Val Val Asp Asp Pro Thr Glu Pro Gly His Tyr Asp
 65 70 75 80
 Ala Glu Trp Ile Ile Ile Leu Asp Asp Trp Thr Asp Gly Ile Gly Lys
 85 90 95
 Ser Pro Gln Gln Leu Tyr Gly Glu Leu Thr Asp Pro Asn Lys Pro Thr
 100 105 110
 Met Gln Asn Thr Thr Gly Met Pro Glu Gly Glu Gly Val Asp Ser Asn
 115 120 125
 Leu Leu Gly Gly Asp Gly Gly Asp Ile Ala Tyr Pro Tyr Tyr Leu Ile
 130 135 140
 Asn Gly Arg Ile Pro Val Ala Ala Thr Ser Phe Lys Ala Lys Pro Gly
 145 150 155 160
 Gln Arg Ile Arg Ile Arg Ile Ile Asn Ser Ala Ala Asp Thr Ala Phe
 165 170 175
 Arg Ile Ala Leu Ala Gly His Ser Met Thr Val Thr His Thr Asp Gly
 180 185 190
 Tyr Pro Val Ile Pro Thr Glu Val Asp Ala Leu Leu Ile Gly Met Ala
 195 200 205
 Glu Arg Tyr Asp Val Met Val Thr Ala Ala Gly Gly Val Phe Pro Leu
 210 215 220
 Val Ala Leu Ala Glu Gly Lys Asn Ala Leu Ala Arg Ala Leu Leu Ser
 225 230 235 240
 Thr Gly Ala Gly Ser Pro Pro Asp Pro Gln Phe Arg Pro Asp Glu Leu
 245 250 255
 Asn Trp Arg Val Gly Thr Val Glu Met Phe Thr Ala Ala Thr Thr Ala
 260 265 270
 Asn Leu Gly Arg Pro Glu Pro Thr His Asp Leu Pro Val Thr Leu Gly
 275 280 285
 Gly Thr Met Ala Lys Tyr Asp Trp Thr Ile Asn Gly Glu Pro Tyr Ser
 290 295 300
 Thr Thr Asn Pro Leu His Val Arg Leu Gly Gln Arg Pro Thr Leu Met
 305 310 315 320
 Phe Asp Asn Thr Thr Met Met Tyr His Pro Ile His Leu His Gly His
 325 330 335
 Thr Phe Gln Met Ile Lys Ala Asp Gly Ser Pro Gly Ala Arg Lys Asp
 340 345 350
 Thr Val Ile Val Leu Pro Lys Gln Lys Met Arg Ala Val Leu Val Ala
 355 360 365
 Asp Asn Pro Gly Val Trp Val Met His Cys His Asn Asn Tyr His Gln
 370 375 380
 Val Ala Gly Met Ala Thr Arg Leu Asp Tyr Ile Leu
 385 390 395

<210> SEQ ID NO 42

<211> LENGTH: 539

-continued

<212> TYPE: PRT

<213> ORGANISM: Mycobacterium kansasii

<400> SEQUENCE: 42

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Met Pro Val Leu Pro Ala Ser Gly His Pro Leu Gly Gly Val Gln Leu
1          5          10          15
Ser Arg Arg Gly Phe Ile Gly Ala Gly Ile Ala Ser Gly Leu Ala Leu
20          25          30
Ala Gly Cys Gly His Ser Gln Thr His Ser Ala Glu Ala Ala Met Ala
35          40          45
Ala Ala Ile Asp Ala Ala Glu Arg Ala Arg Pro His Ser Gly Arg Thr
50          55          60
Val Thr Ala Ser Leu Val Pro Gln Gln Ala Glu Ile Asp Leu Gly Gly
65          70          75          80
Pro Ile Ala His Thr Leu Ala Tyr Gly Asn Thr Val Pro Gly Pro Leu
85          90          95
Ile Arg Ala Ala Val Gly Asp Glu Ile Val Val Ala Val Thr Asn Arg
100         105         110
Leu Asp Arg Pro Thr Ser Val His Trp His Gly Ile Ala Leu Arg Asn
115         120         125
Asp Met Asp Gly Val Val Pro Ala Thr Pro Asn Ile Glu Ala Gly His
130         135         140
Asp Phe Thr Tyr Arg Phe Ser Val Pro Asp Pro Gly Thr Tyr Trp Ala
145         150         155         160
His Pro His Val Gly Leu Glu Glu Asp Met Gly Leu Tyr Leu Pro Val
165         170         175
Ile Ile Asp Asp Pro Thr Glu Pro Gly Arg Tyr Asp Ala Glu Trp Ile
180         185         190
Val Val Leu Asp Asp Trp Thr Asp Gly Val Gly Lys Ser Pro Gln Gln
195         200         205
Ile Tyr Asp Ala Leu Val Asp Pro Asn Lys Pro Thr Ala Met Pro Thr
210         215         220
Thr Thr Pro Pro Ser Thr Thr Pro Pro Thr Thr Thr Asp Thr Thr Ser
225         230         235         240
Thr Thr Ser Ala Thr Ser Thr Thr Thr Thr Thr Glu Thr Thr Pro Ser
245         250         255
Ser Pro Met Pro Gly Met Pro Gly Gly Asp Val Ala Ser Ser Asp Leu
260         265         270
Leu Gly Gly Asp Gly Gly Asp Val Ala Tyr Pro Tyr Tyr Leu Ile Asn
275         280         285
Gly Arg Ile Pro Ala Ala Pro Thr Thr Phe Asn Ala Lys Pro Gly Gln
290         295         300
Arg Ile Arg Ile Arg Ile Ile Asn Ala Ala Ala Asp Thr Ala Phe Arg
305         310         315         320
Val Ala Leu Ala Gly His Ser Met Thr Val Thr His Thr Asp Gly Tyr
325         330         335
Pro Val Leu Pro Thr Pro Val Asp Ala Leu Leu Ile Gly Met Gly Glu
340         345         350
Arg Tyr Asp Val Ile Val Thr Ala Ala Ser Gly Val Phe Pro Leu Val
355         360         365
Ala Leu Ala Glu Gly Lys Asn Ala Val Ala Arg Ser Leu Leu Ser Thr
370         375         380
Gly Ala Gly Ser Ala Pro Asp Pro Gln Phe Arg Pro Ala Glu Leu Thr
385         390         395         400

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Asn Leu Asp Glu Asn Thr Thr Phe His Trp His Gly Leu Glu Ile Asn
 130 135 140
 Gly Lys Val Asp Gly Gly Pro Ser Gln Val Ile Lys Pro Gly Lys Glu
 145 150 155 160
 Lys Thr Ile Lys Phe Glu Val Asn Gln Asp Ser Ala Thr Leu Trp Tyr
 165 170 175
 His Pro His Pro Ser Pro Asn Thr Ala Lys Gln Val Tyr Asn Gly Leu
 180 185 190
 Ser Gly Leu Leu Tyr Ile Glu Asp Ser Lys Lys Asn Asn Tyr Pro Ser
 195 200 205
 Asn Tyr Gly Lys Asn Asp Leu Pro Ile Ile Ile Gln Asp Lys Thr Phe
 210 215 220
 Val Ser Lys Lys Leu Asn Tyr Ser Lys Thr Lys Asp Glu Asp Gly Thr
 225 230 235 240
 Gln Gly Asp Thr Val Leu Val Asn Gly Ile Val Asn Pro Lys Leu Thr
 245 250 255
 Ala Lys Glu Glu Lys Ile Arg Leu Arg Leu Leu Asn Gly Ser Asn Ala
 260 265 270
 Arg Asp Leu Asn Leu Lys Leu Ser Asn Asn Gln Ser Phe Glu Tyr Ile
 275 280 285
 Ala Ser Asp Gly Gly Gln Leu Lys Asn Ala Lys Lys Leu Lys Glu Ile
 290 295 300
 Asn Leu Ala Pro Ser Glu Arg Lys Glu Ile Val Ile Asp Leu Ser Lys
 305 310 315 320
 Met Lys Gly Glu Lys Ile Ser Leu Val Asp Asn Asp Lys Thr Val Ile
 325 330 335
 Leu Pro Ile Ser Asn Lys Glu Lys Ser Ser Asn Lys Gly Asn Thr Pro
 340 345 350
 Lys Val Ser Lys Lys Ile Lys Leu Glu Gly Met Asn Asp His Val Thr
 355 360 365
 Ile Asn Gly Asn Lys Phe Asp Pro Asn Arg Ile Asp Phe Thr Gln Lys
 370 375 380
 Leu Asn Gln Lys Glu Val Trp Glu Ile Glu Asn Val Lys Asp Lys Met
 385 390 395 400
 Gly Gly Met Lys His Pro Phe His Ile His Gly Thr Gln Phe Lys Val
 405 410 415
 Leu Ser Val Asp Gly Glu Lys Pro Pro Lys Asp Met Arg Gly Lys Lys
 420 425 430
 Asp Val Ile Ser Leu Glu Pro Gly Gln Lys Ala Lys Ile Glu Ile Val
 435 440 445
 Phe Lys Asn Thr Gly Thr Tyr Met Phe His Cys His Ile Leu Glu His
 450 455 460
 Glu Glu Asn Gly Met Met Gly Gln Val Lys Ile Thr Asn
 465 470 475

<210> SEQ ID NO 45

<211> LENGTH: 477

<212> TYPE: PRT

<213> ORGANISM: Staphylococcus saprophyticus

<400> SEQUENCE: 45

Met Tyr Lys Lys Met Phe Thr Ile Leu Ile Thr Leu Phe Ser Ile Met
 1 5 10 15

Phe Met Val Pro Asn Asp Thr Phe Ala Glu Gly Lys His Asn Met Met
 20 25 30

-continued

Phe Lys Asn Thr Gly Thr Tyr Met Phe His Cys His Ile Leu Glu His
450 455 460

Glu Asp Asn Gly Met Met Gly Gln Ile Lys Val Thr Lys
465 470 475

<210> SEQ ID NO 46
<211> LENGTH: 463
<212> TYPE: PRT
<213> ORGANISM: Pseudomonas aeruginosa

<400> SEQUENCE: 46

Met Thr Phe Thr Arg Arg Gln Val Leu Gly Gly Leu Ala Gly Leu Ala
1 5 10 15
Val Val Gly Leu Gly Ala Gly Gly Ala Arg Leu Trp Leu Ala Arg Pro
20 25 30
Gln Val Ala Gln Glu Tyr Asp Tyr Glu Leu Ile Ala Ala Pro Leu Asp
35 40 45
Leu Glu Ile Val Pro Gly Phe Ser Ser Pro Ala Leu Ala Tyr Gly Gly
50 55 60
Gln Cys Pro Gly Val Glu Leu Arg Ala Lys Gln Gly Glu Trp Leu Arg
65 70 75 80
Val Arg Phe Thr Asn Arg Leu Asp Glu Pro Thr Thr Ile His Trp His
85 90 95
Gly Ile Arg Leu Pro Ile Glu Met Asp Gly Val Pro Tyr Ile Ser Gln
100 105 110
Pro Pro Val Gln Pro Gly Glu Ser Phe Ile Tyr Gln Phe Lys Thr Gln
115 120 125
Asp Ala Gly Ser Tyr Trp Tyr His Pro His Leu Met Ser Ser Glu Gln
130 135 140
Leu Gly Arg Gly Leu Val Gly Pro Leu Ile Ile Glu Glu Arg Glu Pro
145 150 155 160
Thr Gly Phe Arg His Glu Lys Val Leu Cys Leu Lys Thr Trp His Val
165 170 175
Asp Glu Gln Gly Ala Phe Thr Pro Phe Ser Val Pro Arg Gln Ala Ala
180 185 190
Arg Glu Gly Thr Arg Gly Arg Tyr Ser Thr Ile Asn Gly Lys His Val
195 200 205
Pro Thr Ile Asp Leu Pro Ala Gly Gln Ile Val Arg Val Arg Leu Leu
210 215 220
Asn Val Asp Asn Thr Val Thr Tyr Arg Leu Asn Leu Pro Asn Gly Glu
225 230 235 240
Ala Arg Ile Tyr Ala Val Asp Gly His Pro Val Glu Pro Arg Gly Phe
245 250 255
Glu Gly Gln Tyr Trp Ile Gly Pro Gly Met Arg Leu Glu Leu Ala Leu
260 265 270
Lys Val Pro Glu Ala Gly Thr Glu Leu Ser Leu Arg Asp Gly Pro Val
275 280 285
Arg Leu Ala Thr Ile Arg Ser Val Ala Ser Ala Glu Ala Pro Ala Gly
290 295 300
Asp Trp Pro Lys Pro Leu Pro Ala Asn Pro Val Ser Glu Pro Asp Leu
305 310 315 320
Ala Asn Ala Glu Lys Ile Gly Phe Arg Phe Glu Trp Val Gly Ala Met
325 330 335
Ser Asp Thr Ser Gly Lys Asn Pro Tyr Pro Ser Phe Trp Gln Ile Asn
340 345 350

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Gly Lys Ala Trp Glu Gly Gly Glu Glu His Lys His Asn Ala Pro Pro
 355 360 365
 Leu Ala Lys Leu Lys Glu Gly Gln Ser Tyr Ile Phe Glu Leu Arg Asn
 370 375 380
 Met Ala Gln Tyr Gln His Pro Ile His Leu His Gly Met Ala Phe Lys
 385 390 395 400
 Val Leu Asp Ser Asp Arg Arg Glu Ile Ile Pro Tyr Phe Thr Asp Thr
 405 410 415
 Tyr Leu Leu Gly Lys Asn Glu Thr Ala Arg Val Ala Leu Val Ala Asp
 420 425 430
 Asn Pro Gly Leu Trp Met Phe His Cys His Val Ile Asp His Met Glu
 435 440 445
 Thr Gly Leu Met Gly Thr Ile Ala Val Gly Glu Ala Trp Cys Gly
 450 455 460

<210> SEQ ID NO 47
 <211> LENGTH: 458
 <212> TYPE: PRT
 <213> ORGANISM: Pseudomonas fluorescens

<400> SEQUENCE: 47

Met Ser Phe Thr Arg Arg Gln Ile Leu Gly Gly Leu Ala Gly Leu Val
 1 5 10 15
 Val Val Gly Val Gly Ala Gly Gly Ala Ser Arg Tyr Trp Leu Gly Lys
 20 25 30
 Met Ala Asp Ala Asp Ala Gly Tyr Asp Tyr Glu Leu Ile Ala Ala Pro
 35 40 45
 Leu Asp Val Glu Leu Val Pro Gly His Lys Thr Glu Ala Trp Ala Phe
 50 55 60
 Gly Pro Ser Ala Pro Gly Thr Glu Leu Arg Val Arg Gln Gly Glu Trp
 65 70 75 80
 Leu Arg Val Arg Phe Ile Asn His Leu Pro Val Ala Thr Thr Ile His
 85 90 95
 Trp His Gly Ile Arg Leu Pro Leu Glu Met Asp Gly Val Pro Tyr Val
 100 105 110
 Ser Gln Leu Pro Val Leu Pro Gly Glu Tyr Phe Asp Tyr Lys Phe Arg
 115 120 125
 Val Pro Asp Ala Gly Ser Tyr Trp Tyr His Pro His Val Ser Ser Ser
 130 135 140
 Glu Glu Leu Gly Arg Gly Leu Val Gly Pro Leu Ile Val Glu Glu Arg
 145 150 155 160
 Glu Pro Thr Gly Phe Lys Tyr Glu Lys Thr Leu Ser Leu Lys Asn Trp
 165 170 175
 His Ile Asp Asp Glu Gly His Phe Val Glu Phe Ser Val Pro Arg Glu
 180 185 190
 Ala Ala Arg Gly Gly Thr Ala Gly Arg Leu Ser Thr Ile Asn Gly Val
 195 200 205
 Pro Ser Pro Val Ile Glu Leu Pro Ala Gly Gln Ile Thr Arg Val Arg
 210 215 220
 Leu Leu Asn Leu Asp Asn Thr Leu Thr Tyr Arg Leu Asn Ile Pro Gly
 225 230 235 240
 Val Glu Ala Gln Ile Tyr Ala Leu Asp Gly Asn Pro Val Glu Pro Arg
 245 250 255
 Pro Leu Gly Lys Glu Tyr Trp Leu Gly Pro Gly Met Arg Ile Cys Leu

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260				265				270							
Ala	Ile	Lys	Ala	Pro	Pro	Ala	Gly	Glu	Glu	Leu	Ser	Leu	Arg	Asn	Gly
		275					280							285	
Pro	Val	Arg	Leu	Gly	Thr	Leu	Arg	Ser	Val	Ala	Asn	Asn	Asp	Ala	Pro
		290				295					300				
Thr	Glu	Trp	Pro	Lys	Ala	Leu	Pro	Ala	Asn	Pro	Val	Ala	Glu	Pro	Asp
					310					315					320
Leu	Ala	Asn	Ala	Glu	Lys	Leu	Asn	Phe	Asn	Phe	Glu	Trp	Val	Gly	Ser
					325					330					335
Val	Ser	Val	Asn	Val	Asp	Asn	Gly	Lys	Pro	Pro	Ser	Leu	Trp	Gln	Ile
			340						345						350
Asn	Gly	Lys	Ala	Trp	Val	Ile	Thr	Asp	Lys	Thr	Cys	Ala	Asp	Arg	Pro
		355					360						365		
Ile	Ala	Ser	Leu	Lys	Leu	Gly	Gln	Ser	Tyr	Ile	Phe	Glu	Leu	Lys	Asn
						375					380				
Met	Thr	Gln	Tyr	Gln	His	Pro	Ile	His	Leu	His	Gly	Met	Ser	Phe	Lys
					390					395					400
Val	Ile	Ala	Ser	Asn	Arg	His	Lys	Ile	Ile	Pro	Tyr	Phe	Thr	Asp	Thr
					405					410					415
Tyr	Leu	Leu	Gly	Lys	Asn	Glu	Arg	Ala	Gln	Val	Ala	Leu	Val	Ala	Asp
			420						425					430	
Asn	Pro	Gly	Val	Trp	Met	Phe	His	Cys	His	Val	Ile	Asp	His	Met	Glu
		435					440						445		
Thr	Gly	Leu	Met	Ala	Ala	Ile	Glu	Val	Lys						
		450				455									

<210> SEQ ID NO 48

<211> LENGTH: 459

<212> TYPE: PRT

<213> ORGANISM: Pseudomonas putida

<400> SEQUENCE: 48

Met	Ser	Phe	Thr	Arg	Arg	Gln	Met	Leu	Lys	Gly	Leu	Thr	Gly	Leu	Val
1				5					10					15	
Val	Val	Gly	Leu	Gly	Ala	Gly	Gly	Ala	Ala	Arg	Tyr	Trp	Leu	Gly	Lys
			20					25					30		
Val	Glu	Asp	Glu	Asn	Ala	Gly	His	Asp	Tyr	Glu	Leu	Ile	Ala	Ala	Pro
		35					40						45		
Leu	Glu	Val	Glu	Leu	Val	Pro	Gly	Phe	Lys	Thr	Glu	Ala	Trp	Ala	Phe
		50				55						60			
Gly	Pro	Ser	Ala	Pro	Gly	Thr	Glu	Leu	Arg	Val	Arg	Gln	Gly	Thr	Trp
					70					75					80
Leu	Arg	Val	Arg	Phe	Ile	Asn	His	Leu	Pro	Val	Glu	Thr	Thr	Ile	His
				85					90					95	
Trp	His	Gly	Ile	Arg	Leu	Pro	Leu	Glu	Met	Asp	Gly	Val	Pro	Tyr	Val
			100						105					110	
Ser	Gln	Leu	Pro	Val	Lys	Pro	Gly	Glu	Tyr	Phe	Asp	Tyr	Lys	Phe	Arg
			115						120				125		
Val	Pro	Asp	Ala	Gly	Ser	Tyr	Trp	Tyr	His	Pro	His	Val	Ser	Ser	Ser
		130					135					140			
Glu	Glu	Leu	Gly	Arg	Gly	Leu	Val	Gly	Pro	Leu	Ile	Val	Glu	Glu	Arg
					150					155					160
Glu	Pro	Thr	Gly	Phe	Gln	His	Glu	Arg	Thr	Leu	Ser	Leu	Lys	Asn	Trp
					165					170					175

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His Val Asp Glu Gln Gly Ala Trp Leu Pro Phe Ser Ile Pro Arg Glu
 180 185 190

Ala Ala Arg Asn Gly Thr Ala Gly Arg Leu Ile Thr Ile Asn Gly Gln
 195 200 205

Ala Asp Ser Ile Thr Glu Leu Pro Ala Gly Gln Val Val Arg Val Arg
 210 215 220

Val Leu Asn Leu Asp Asn Thr Trp Thr Tyr Arg Leu Asn Leu Lys Gly
 225 230 235 240

Asn Cys Glu Ala Arg Ile Tyr Ala Leu Asp Gly Asn Pro Val Thr Pro
 245 250 255

Arg Ala Leu Asp Glu Tyr Trp Leu Gly Pro Gly Met Arg Ile Cys Leu
 260 265 270

Ala Ile Arg Ile Pro Glu Ala Gly Glu Glu Ile Ser Leu Arg Asp Gly
 275 280 285

Phe Val Arg Leu Gly Thr Leu Arg Ser Val Ala Ser Asn Asp Ala Pro
 290 295 300

Ser Asp Trp Pro Pro Ala Leu Pro Pro Asn Pro Ile Ala Glu Pro Asp
 305 310 315 320

Leu Glu His Ala Glu Lys Leu Asn Phe Asn Phe Glu Trp Ala Ala Gly
 325 330 335

Val Ser Val Thr Ala Asp Pro Ala Lys Pro Ser Ser Met Trp Gln Ile
 340 345 350

Asn Gly Gln Ala Trp Asp Ile Thr Asp Lys Thr Cys Ala Asp Arg Pro
 355 360 365

Ile Ala Thr Leu Gln Lys Gly Lys Ser Tyr Ile Phe Glu Leu Lys Asn
 370 375 380

Met Thr Gln Tyr Gln His Pro Ile His Leu His Gly Met Ser Phe Lys
 385 390 395 400

Val Ile Ala Ser Asn Arg His Asp Ile Lys Glu Pro Trp Phe Thr Asp
 405 410 415

Thr Tyr Leu Leu Gly Lys Asn Glu Arg Ala Gln Val Ala Leu Val Ala
 420 425 430

Asp Asn Pro Gly Thr Trp Met Phe His Cys His Val Ile Asp His Met
 435 440 445

Glu Thr Gly Leu Met Ala Ala Ile Ala Val Val
 450 455

<210> SEQ ID NO 49
 <211> LENGTH: 678
 <212> TYPE: PRT
 <213> ORGANISM: Trichophyton rubrum

<400> SEQUENCE: 49

Met Asp Arg Pro Ala Ser Arg Arg Arg His Asn Ser Gln Arg Arg Leu
 1 5 10 15

Gln Gln Gln Pro Glu Asp Glu Ser Val Val Ala Ala Asp Thr Leu Ile
 20 25 30

Ile Pro Val Gln Arg Glu Asp Glu Lys Arg Pro Ala Gly Pro Ser Thr
 35 40 45

Gly Ser Gln Thr Gly Glu Thr Lys Pro Glu Asp His Arg Arg Ser Leu
 50 55 60

Ser Gly Cys Pro Trp Leu Leu Val Gly Phe Val Cys Cys Ile Ser Thr
 65 70 75 80

Leu Leu Leu Val Leu Gly Leu Ala Ala Arg Trp Gln Gln Gly Ala Ile
 85 90 95

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Pro Asn Ile Phe Asp Ile Gly Asn Leu Arg Pro Ser Ser Ser Pro Ala
 100 105 110

Ser Ser Ser Ser Ser Thr Ala Gly Pro Trp Ser Ser Arg Leu His Pro
 115 120 125

Glu Asp His Leu Phe Arg Pro Glu Thr Thr Ile Thr Leu Glu Trp Ser
 130 135 140

Val Thr Thr Gly Tyr Arg Arg Leu Asp Gly Val Lys Lys Arg Val Tyr
 145 150 155 160

Leu Ile Asn Gly Leu Phe Pro Gly Pro Thr Ile Glu Ala Arg Ser Gly
 165 170 175

Asp Ser Leu Gln Val Gln Val Thr Asn Asn Ile Gln Asp Glu Gly Leu
 180 185 190

Val Ile His Trp His Gly Leu His Met Arg Gly Ala Asn His Met Asp
 195 200 205

Gly Val Thr Gly Val Thr Gln Cys Pro Ile Val Pro Gly Asp Ser Met
 210 215 220

Leu Tyr Asn Phe Thr Ile Ser Gln Ser Gln Ser Gly Thr Phe Trp Tyr
 225 230 235 240

His Ala His Ser Ala Leu Gln Arg Ala Glu Gly Leu Tyr Gly Gly Phe
 245 250 255

Val Val His Lys Pro Ser Thr Pro Ser Met Arg Ile Ala Arg Asp Pro
 260 265 270

Ala Ile His Ala Asp Ala Val Lys Tyr Gln Tyr Glu Lys Glu His Leu
 275 280 285

Leu Leu Ile Gly Asp Trp Tyr His Arg Pro Ala Glu Asp Val Leu Lys
 290 295 300

Trp Phe Lys Ser Leu Glu Ala Asn Gly Gln Glu Pro Val Pro Asp Ser
 305 310 315 320

Phe Leu Ile Asn Gly Ala Gly Arg Phe Asn Cys Ser Met Ala Leu Pro
 325 330 335

Thr Arg Pro Ile Asp Cys Val Asp Glu Gly Tyr Pro Thr Pro Glu Leu
 340 345 350

Leu Leu Asp Ser Ser Thr Ser Tyr Arg Met Arg Val Ile Asn Val Gly
 355 360 365

Ser Leu Ala Gly Val Ser Leu Gly Phe Glu His Gly Thr Val Thr Pro
 370 375 380

Ile Gln Val Asp Gly Gly Thr Glu Val Glu Leu Pro Ser Val Ser Pro
 385 390 395 400

Asn Ala Arg Ser Met Gly Ile Val Tyr Pro Gly Gln Arg Thr Asp Phe
 405 410 415

Val Leu Arg Asn Pro Leu Gly Glu Thr Gly Gln Ser Ser Ile Thr Val
 420 425 430

Glu Leu Asp Pro Glu Cys Phe Ser Leu Pro Asn Pro Ala Leu Thr Arg
 435 440 445

Val Gln Thr Phe Pro Ile Ser Gly Ser Ala Lys Lys Pro Ser His Pro
 450 455 460

Leu Ser Asp Asn Pro Ile Gly Glu Ala Gly Thr His Val Asp Leu Thr
 465 470 475 480

Glu Leu Thr Ser Thr Ala Ser Thr Ile Ser His Ile Pro Ala Lys Ala
 485 490 495

Asp Glu Thr Phe Leu Val Tyr Thr Leu Leu Ser Lys Leu Ser Ser Asn
 500 505 510

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Asn Tyr Val Pro Phe Ala Phe Phe Asn His Thr Ser Trp Arg Pro Gln
   515                               520                               525
Ala Asp Pro Pro Leu Pro Leu Ile Ser Leu Gln Arg Lys Asp Trp Asp
   530                               535                               540
Lys Asn Gln Phe Thr Ile Lys Thr Ser Ser Arg Ala Ser Trp Val Asp
   545                               550                               555                               560
Leu Ile Val Asn Asn Leu Asp Glu Gly Pro His Pro Phe His Ile His
                               565                               570                               575
Gly His Asp Phe Tyr Val Met Ser Leu His Glu Ala Asp Thr Gly Met
   580                               585                               590
Gly Ser Tyr Asn Pro Trp Asp Pro Ser Asn Lys Ala Pro Ala Tyr Asp
   595                               600                               605
His Ser Gln Ala Ile Leu Arg Asp Thr Val His Ile Pro Ala Arg Gly
   610                               615                               620
His Ala Val Leu Arg Phe Arg Ala Asp Asn Pro Gly Ile Trp Leu Phe
   625                               630                               635                               640
His Cys His Ile Leu Trp His Leu Ala Ser Gly Met Ala Met Leu Val
   645                               650                               655
Asp Val Met Asp Ser Ala Ser Arg Pro Leu His Gly Ile Leu Asn Gln
   660                               665                               670
Thr Cys Arg Tyr Leu Thr
   675

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<210> SEQ ID NO 50

<211> LENGTH: 681

<212> TYPE: PRT

<213> ORGANISM: Trichophyton tonsurans

<400> SEQUENCE: 50

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Met Asp Arg Pro Ala Ser Arg Arg Arg His Asn Ser Gln Arg Arg Leu
 1      5      10      15
Gln Gln Pro Gly Asp Glu Ser Ala Val Ala Ala Asp Thr Leu Ile Pro
 20     25     30
Val Gln Arg Glu Gly Glu Lys Arg Pro Ala Arg Pro Ser Ala Gly Ser
 35     40     45
Gln Thr Gly Glu Thr Lys Pro Gly Asp Gln Arg Arg Ser Leu Pro Gly
 50     55     60
Arg Pro Trp Pro Trp Pro Leu Val Gly Phe Val Cys Cys Ile Ser Thr
 65     70     75     80
Leu Leu Ile Val Leu Gly Leu Ala Ala Arg Trp Gln Gln Gly Ala Ile
 85     90     95
Pro Asn Ile Phe Asp Ile Gly Ile Leu Arg Pro Ser Ser Ser Pro Ala
100    105    110
Ser Ser Ser Pro Ser Thr Ala Glu Pro Trp Ser Ser Arg Leu His Pro
115    120    125
Glu Asp His Val Phe Arg Pro Glu Thr Thr Val Thr Leu Glu Trp Ser
130    135    140
Val Ser Thr Gly Tyr Arg Arg Leu Asp Gly Val Lys Lys Arg Val Tyr
145    150    155    160
Leu Ile Asn Gly Leu Phe Pro Gly Pro Thr Ile Glu Ala Arg Ser Gly
165    170    175
Asp Ser Leu Arg Ile Lys Val Thr Asn Asn Ile Gln Asp Glu Gly Leu
180    185    190
Val Ile His Trp His Gly Leu His Met Arg Gly Ala Asn His Met Asp
195    200    205

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Gly Val Thr Gly Val Thr Gln Cys Pro Ile Val Pro Gly Asp Ser Met
 210 215 220
 Leu Tyr Asn Phe Thr Ile Ser Gln Ser Gln Ser Gly Thr Phe Trp Tyr
 225 230 235 240
 His Ala His Ser Ala Leu Gln Arg Ala Glu Gly Leu Tyr Gly Gly Phe
 245 250 255
 Val Val His Lys Pro Ser Thr Pro Ser Met Arg Ile Ala Arg Asp Pro
 260 265 270
 Ala Met His Ala Asp Ala Val Lys Tyr Gln Tyr Glu Arg Glu His Leu
 275 280 285
 Leu Leu Ile Gly Asp Trp Tyr His Arg Pro Ala Asp Asp Val Leu Asn
 290 295 300
 Trp Phe Lys Ser Leu Glu Ala Asn Gly Gln Glu Pro Val Pro Asp Ser
 305 310 315 320
 Phe Leu Ile Asn Gly Ala Gly Arg Phe Asn Cys Ser Met Ala Leu Pro
 325 330 335
 Thr Arg Pro Leu Asp Cys Val Asp Glu Gly Tyr Pro Thr Pro Glu Leu
 340 345 350
 Leu Leu Asp Ser Ser Ser Ser Ser Ser Tyr Arg Met Arg Val Val Asn
 355 360 365
 Val Gly Ser Leu Ala Gly Val Ser Leu Ala Phe Glu His Gly Thr Val
 370 375 380
 Thr Pro Ile Gln Val Asp Gly Gly Thr Glu Val Glu Leu Pro Ser Val
 385 390 395 400
 Ser Pro Asn Ala Arg Ser Ile Gly Ile Val Tyr Pro Gly Gln Arg Thr
 405 410 415
 Asp Phe Val Leu Arg Asn Ala Phe Gly Gly Ala Glu Gln Ser Ser Ile
 420 425 430
 Thr Val Glu Leu Asp Pro Glu Cys Phe Ser Leu Pro Asn Pro Ala Leu
 435 440 445
 Thr Arg Val Gln Thr Phe Pro Ile Arg Gly Ser Ala Lys Lys Pro Ser
 450 455 460
 Ser His Thr Leu Ser Asp Asn Pro Ile Gly Glu Ser Gly Thr His Val
 465 470 475 480
 Asp Leu Thr Glu Leu Thr Ser Thr Ala Ser Thr Ile Ser His Ile Pro
 485 490 495
 Ala Glu Ala Asp Glu Thr Phe Leu Val Tyr Thr Leu Leu Ser Lys Leu
 500 505 510
 Ser Ser Asn Asn Tyr Val Pro Phe Ala Phe Phe Asn His Thr Ser Trp
 515 520 525
 Arg Pro Gln Ala Asp Pro Pro Leu Pro Leu Ile Ser Leu Gln Arg Lys
 530 535 540
 Asp Trp Asp Lys Asn Gln Phe Thr Ile Lys Thr Ser Ser Lys Ala Ser
 545 550 555 560
 Trp Val Asp Leu Val Val Asn Asn Leu Asp Glu Gly Pro His Pro Phe
 565 570 575
 His Ile His Gly His Asp Phe Tyr Val Met Ser Leu His Glu Ala Asp
 580 585 590
 Thr Gly Met Gly Ser Tyr Asn Pro Trp Asp Pro Ser Asn Gln Ala Pro
 595 600 605
 Ala Tyr Asn His Ser Lys Ala Ile Leu Arg Asp Thr Val His Ile Pro
 610 615 620

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Ala Arg Gly His Ala Val Leu Arg Phe Arg Ala Asp Asn Pro Gly Ile
625 630 635 640

Trp Leu Phe His Cys His Ile Leu Trp His Leu Ala Ser Gly Met Ala
645 650 655

Met Leu Val Asp Val Met Asp Ser Val Ser Arg Pro Leu His Gly Ile
660 665 670

Pro Asn Gln Thr Cys Arg Tyr Leu Thr
675 680

<210> SEQ ID NO 51
 <211> LENGTH: 626
 <212> TYPE: PRT
 <213> ORGANISM: Candida albicans

<400> SEQUENCE: 51

Met Arg Phe Ile Val Ser Ser Phe Ile Phe Phe Ile Ser Phe Leu Ser
1 5 10 15

Ser Leu Ile Thr Ala Glu Thr His Thr Trp Tyr Phe Lys Thr Ser Trp
20 25 30

Val Asp Ala Asn Pro Asp Gly Val Phe Pro Arg Lys Met Ile Gly Phe
35 40 45

Asn Asp Ser Trp Pro Leu Pro Thr Leu Arg Val Lys Lys Gly Asp Thr
50 55 60

Val Asn Leu Tyr Leu Ile Asn Gly Phe Asp Asp Arg Asn Thr Ser Leu
65 70 75 80

His Phe His Gly Leu Phe Gln His Gly Thr Asn Gln Met Asp Gly Pro
85 90 95

Glu Met Val Thr Gln Cys Pro Ile Pro Pro Gly Glu Thr Phe Leu Tyr
100 105 110

Asn Phe Thr Val Asp Asp Gln Val Gly Ser Tyr Trp Tyr His Ser His
115 120 125

Thr Ser Gly Gln Tyr Gly Asp Gly Met Arg Gly Val Phe Ile Ile Glu
130 135 140

Asp Asp Asp Phe Pro Tyr Asp Tyr Asp Glu Glu Val Val Leu Thr Leu
145 150 155 160

Ser Glu His Tyr His Asp Tyr Ser Lys Asp Leu Met Pro Gly Phe Leu
165 170 175

Ser Arg Phe Asn Pro Thr Gly Ala Glu Pro Ile Pro Ser Asn Ile Leu
180 185 190

Phe Asn Glu Thr Arg Asn Asn Thr Trp Lys Val Glu Pro Gly Lys Thr
195 200 205

Tyr Leu Leu Arg Ile Ala Asn Thr Gly Arg Phe Val Thr Gln Tyr Leu
210 215 220

Trp Met Glu Asp His Glu Phe Thr Val Val Glu Val Asp Gly Val Tyr
225 230 235 240

Val Glu Lys Asn Thr Thr Asp Met Leu Tyr Ile Thr Ile Ala Gln Arg
245 250 255

Tyr Gly Val Leu Ile Thr Thr Lys Asn Ser Thr Asn Lys Asn Tyr Ala
260 265 270

Phe Met Asn Arg Val Asp Asp Thr Met Leu Asp Thr Ile Pro Lys Asp
275 280 285

Leu Gln Leu Asn Gly Thr Asn Tyr Ile Val Tyr Asn Glu Ser Ala Pro
290 295 300

Leu Pro Asp Ala Tyr Asp Val Asp Ser Ile Asp Asp Tyr Leu Asp Asp
305 310 315 320

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225		230		235		240									
Gln	Asn	Thr	Thr	Asp	Leu	Leu	Tyr	Ile	Thr	Val	Ala	Gln	Arg	Tyr	Ser
				245					250					255	
Val	Leu	Ile	Thr	Thr	Lys	Asn	Glu	Thr	Asp	Lys	Asn	Tyr	Ala	Phe	Met
			260					265					270		
Asn	Arg	Val	Asp	Ile	Thr	Met	Leu	Asp	Val	Ile	Pro	Gly	Asp	Leu	Glu
		275					280					285			
Leu	Asn	Gly	Thr	Asn	Tyr	Ile	Val	Tyr	Asn	Glu	Asp	Ala	Asp	Leu	Pro
	290					295					300				
Glu	Pro	Tyr	Leu	Leu	Asp	Ser	Ile	Asp	Asp	Phe	Phe	Asp	Asp	Phe	Trp
305					310					315					320
Leu	Lys	Pro	Leu	Ser	Lys	Glu	Lys	Leu	Leu	Asp	Asp	Ala	Asp	Tyr	Thr
			325					330						335	
Ile	Thr	Leu	Glu	Val	Gln	Met	Asp	Asn	Leu	Gly	Asn	Gly	Val	Asn	Tyr
		340						345					350		
Ala	Phe	Phe	Asn	Asn	Ile	Thr	Tyr	Ala	His	Pro	Lys	Val	Pro	Thr	Leu
		355					360					365			
Met	Ser	Val	Leu	Ser	Ser	Gly	Asp	Asp	Ala	Ser	Asn	Glu	Leu	Val	Tyr
370						375					380				
Gly	Thr	Asn	Thr	Asn	Ser	Phe	Val	Leu	Gln	Gly	Gly	Glu	Val	Val	Asp
385					390					395					400
Ile	Val	Leu	Asn	Asn	Leu	Asp	Thr	Gly	Lys	His	Pro	Phe	His	Leu	His
			405						410					415	
Gly	His	Ala	Phe	Gln	Leu	Ile	Glu	Arg	His	Glu	Glu	Ile	Pro	Asp	Thr
			420					425					430		
Glu	Asp	Pro	Val	Thr	Tyr	Asn	Ala	Thr	Asp	His	Ala	Asp	Trp	Pro	Glu
		435					440					445			
Tyr	Pro	Met	Leu	Arg	Asp	Thr	Ile	Tyr	Ile	Asn	Pro	Gln	Ser	Tyr	Ala
450						455					460				
Val	Leu	Arg	Phe	Lys	Ala	Asp	Asn	Pro	Gly	Val	Trp	Phe	Phe	His	Cys
465				470						475					480
His	Ile	Glu	Trp	His	Leu	Asp	Gln	Gly	Leu	Ala	Ile	Val	Leu	Ile	Glu
			485						490					495	
Asp	Pro	Gln	Ala	Ile	Gln	Lys	Asn	Glu	Lys	Ile	Thr	Asp	Asn	His	Lys
			500					505					510		
Gln	Ile	Cys	Glu	Lys	Val	Gly	Val	Pro	Trp	Gln	Gly	Asn	Ala	Ala	Ala
		515					520					525			
Asn	Asn	Lys	Asp	Tyr	Leu	Asn	Leu	Asp	Gly	Glu	Asn	Leu	Gln	Val	Lys
530						535					540				
Arg	Leu	Pro	Thr	Gly	Phe	Thr	Ala	Lys	Gly	Ile	Val	Ala	Leu	Val	Phe
545					550					555					560
Ser	Cys	Ile	Ala	Gly	Val	Leu	Gly	Leu	Val	Ala	Ile	Ser	Tyr	Tyr	Gly
			565						570					575	
Met	Thr	Asp	Ile	Lys	Asn	Val	Glu	Gln	Arg	Val	Ala	Arg	Asp	Leu	Asp
			580					585					590		
Val	Asp	Leu	Asp	Asp	Asp	Val	Glu	Gln	Leu	Ser	Glu	Glu	Gly	Ser	
		595				600						605			
Ser	Gly	Ser	Asn	Ser	Lys	Gln	His								
610						615									

<210> SEQ ID NO 54

<211> LENGTH: 617

<212> TYPE: PRT

<213> ORGANISM: Unknown

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<220> FEATURE:

<223> OTHER INFORMATION: Candida auris

<400> SEQUENCE: 54

Met Asn Gln Leu Ser Leu Phe Phe Val Leu Phe Ile Ser Trp Phe Ala
1 5 10 15
Leu Ala Ser Ala Glu Thr His Thr Trp Tyr Phe Lys Thr Gly Trp Val
20 25 30
Lys Ala Asn Pro Asp Gly Asn Phe Glu Arg Asp Val Ile Gly Phe Asn
35 40 45
Gly Ser Trp Pro Leu Pro Thr Leu Arg Val Lys Lys Gly Asp Arg Val
50 55 60
Asn Leu Tyr Leu Thr Asn Gly Phe Asp Asp Arg Asn Thr Thr Leu His
65 70 75 80
Phe His Gly Met Phe Gln Asn Gly Ser Ala Gln Met Asp Gly Pro Glu
85 90 95
Met Val Thr Gln Cys Pro Ile Pro Pro Gly Glu Thr Tyr Leu Tyr Asn
100 105 110
Phe Thr Val Ala Asp Gln Val Gly Thr Tyr Trp Tyr His Ser His Thr
115 120 125
Ala Gly Gln Tyr Gly Asp Gly Met Arg Ala Pro Phe Ile Ile Glu Glu
130 135 140
Lys Asn Lys Glu Asp Tyr Pro Phe Asp Phe Asp Glu Glu Leu Val Leu
145 150 155 160
Pro Leu Gly Glu Trp Tyr His Asp Pro Ala Asp Val Leu Leu Pro Lys
165 170 175
Phe Leu Asn Arg Tyr Asn Pro Thr Gly Ala Glu Pro Ile Pro Gln Asn
180 185 190
Leu Leu Phe Asn Glu Thr Arg Asn Asn Thr Trp Lys Val Glu Pro Asn
195 200 205
Thr Thr Tyr Gly Val Arg Ile Val Asn Met Gly Gly Phe Val Ser Gln
210 215 220
Tyr Leu Tyr Met Glu Asp His Glu Phe Glu Ile Val Glu Val Asp Gly
225 230 235 240
Val Tyr Val Glu Lys Asn Thr Thr Asp Leu Leu Tyr Val Thr Ile Ala
245 250 255
Gln Arg Tyr Gly Val Leu Ile Lys Thr Lys Glu Lys Ala Asp Arg Asn
260 265 270
Tyr Ala Phe Met Asn Ala Phe Asp Asp Thr Met Leu Asp Val Ile Pro
275 280 285
Lys Asp Leu Ile Leu Asn Gly Thr Asn Ser Ile Gln Tyr Thr Asp Asp
290 295 300
Thr Ser Met Pro Asp Glu Tyr Phe Ile Asp Ser Phe Asp Asp Arg Phe
305 310 315 320
Asp Asp Phe Tyr Leu Val Pro Lys Asp Gly Glu Lys Leu Leu Pro Asp
325 330 335
Ser Asp Asn Gln Val Val Ile Asp Val Lys Met Asp Asn Leu Gly Asp
340 345 350
Val Asn Tyr Ala Phe Phe Asn Asn Ile Ser Tyr Val Ala Pro Lys Ile
355 360 365
Pro Leu Leu Ala Thr Ala Met Ser Ala Gly Glu Leu Ala Thr Asn Ser
370 375 380
Tyr Ile Tyr Gly Asn Thr Asn Ala Phe Val Leu Lys Lys Gly Glu Thr
385 390 395 400

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Val Asp Ile Val Leu Asn Asn Gln Asp Asp Gly Thr His Pro Phe His
 405 410 415
 Leu His Gly His Val Phe Gln Leu Ile Glu Arg Gly Pro Glu Phe Gly
 420 425 430
 Asp Pro Val Ser Phe Asp Tyr Asn Asn His Ser Glu Phe Pro Glu Tyr
 435 440 445
 Pro Met Lys Arg Asp Thr Val Tyr Val Asn Pro Asn Ser Tyr Ile Val
 450 455 460
 Met Arg Phe Thr Ala Asp Asn Pro Gly Val Trp Phe Phe His Cys His
 465 470 475 480
 Ile Glu Trp His Leu Glu Gln Gly Leu Ala Ile Val Leu Val Glu Ala
 485 490 495
 Pro Glu Glu Met Gln Lys Asp Pro Ser Gln Gln Leu Thr Glu Asn Phe
 500 505 510
 Lys Asp Val Cys Ser Lys Gly Gly Met Asn Tyr Ser Gly Asn Ala Ala
 515 520 525
 Gly Asn Ser Val Asp Phe Met Asp Leu Lys Gly Met Asn Thr Gln Pro
 530 535 540
 Lys Arg Leu Pro Ala Gly Phe Thr Ala Arg Gly Ile Val Ala Leu Val
 545 550 555 560
 Phe Ser Cys Ile Ala Gly Val Leu Gly Met Val Ala Ile Thr Ile Tyr
 565 570 575
 Gly Leu Ala Asp Val Lys Asp Ile Asp Glu Arg Val Ala Arg Asp Leu
 580 585 590
 Asp Val Asp Leu Asp Glu Ile Ala Ala Asp Glu Ser Ser Gln Leu Val
 595 600 605
 Pro Gly Asp Ser Ser Ser Arg Asn Lys
 610 615

<210> SEQ ID NO 55
 <211> LENGTH: 296
 <212> TYPE: PRT
 <213> ORGANISM: Triticum aestivum

<400> SEQUENCE: 55

Met Lys Thr Phe Leu Ile Leu Ala Leu Leu Ala Ile Val Ala Thr Thr
 1 5 10 15
 Ala Thr Thr Ala Val Arg Val Pro Val Pro Gln Pro Gln Pro Gln Asn
 20 25 30
 Pro Ser Gln Pro Gln Pro Gln Arg Gln Val Pro Leu Val Gln Gln Gln
 35 40 45
 Gln Phe Pro Gly Gln Gln Gln Gln Phe Pro Pro Gln Gln Pro Tyr Pro
 50 55 60
 Gln Pro Gln Pro Phe Pro Ser Gln Gln Pro Tyr Leu Gln Leu Gln Pro
 65 70 75 80
 Phe Pro Gln Pro Gln Pro Phe Pro Pro Gln Leu Pro Tyr Pro Gln Pro
 85 90 95
 Pro Pro Phe Ser Pro Gln Gln Pro Tyr Pro Gln Pro Gln Pro Gln Tyr
 100 105 110
 Pro Gln Pro Gln Gln Pro Ile Ser Gln Gln Gln Ala Gln Gln Gln Gln
 115 120 125
 Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Ile Leu
 130 135 140
 Pro Gln Ile Leu Gln Gln Gln Leu Ile Pro Cys Arg Asp Val Val Leu

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<210> SEQ ID NO 58
 <211> LENGTH: 300
 <212> TYPE: PRT
 <213> ORGANISM: Triticum dicoccum

<400> SEQUENCE: 58

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Met Lys Thr Phe Leu Ile Leu Ala Leu Leu Ala Ile Val Ala Thr Thr
1           5           10           15
Ala Thr Thr Ala Val Arg Val Pro Val Pro Gln Leu Gln Pro Gln Asn
20           25           30
Pro Ser Gln Gln Gln Pro Gln Glu Gln Val Pro Leu Val Gln Gln Gln
35           40           45
Gln Phe Pro Gly Gln Gln Gln Gln Phe Pro Pro Gln Gln Pro Tyr Pro
50           55           60
Gln Pro Gln Pro Phe Pro Ser Gln Gln Pro Tyr Leu Gln Leu Gln Pro
65           70           75           80
Phe Pro Gln Pro Gln Leu Pro Tyr Ser Gln Pro Gln Pro Phe Gln Pro
85           90           95
Gln Gln Pro Tyr Pro Gln Pro Gln Pro Gln Tyr Ser Gln Pro Gln Gln
100          105          110
Pro Ile Ser Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln
115          120          125
Gln Gln Gln Gln Gln Gln Gln Gln Pro Gln Gln Ile Gln Pro Gln Ile
130          135          140
Leu Gln Gln Gln Leu Ile Pro Cys Met Asp Val Val Leu Gln Gln His
145          150          155          160
Asn Ile Ala Gln Gly Arg Ser Gln Val Leu Gln Gln Ser Thr Tyr Gln
165          170          175
Leu Leu Gln Glu Leu Cys Cys Gln His Leu Trp Gln Ile Pro Glu Gln
180          185          190
Ser Gln Cys Gln Ala Ile His Asn Val Val His Ala Ile Ile Leu His
195          200          205
Gln Gln Gln Lys Gln Gln Gln Gln Gln Gln Gln Lys Gln Gln Pro Ser
210          215          220
Ser Gln Val Ser Phe Gln Gln Pro Gln Gln Gln Tyr Pro Ser Gly Gln
225          230          235          240
Gly Ser Phe Arg Pro Ser Leu Gln Asn Pro Gln Ala Gln Gly Ser Val
245          250          255
Gln Pro Gln Gln Leu Pro Gln Phe Ala Glu Ile Arg Asn Leu Ala Leu
260          265          270
Gln Thr Leu Pro Ala Met Cys Asn Val Tyr Ile Pro Pro His Cys Ser
275          280          285
Thr Thr Thr Ala Pro Phe Gly Ile Phe Gly Thr Asn
290          295          300

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<210> SEQ ID NO 59
 <211> LENGTH: 303
 <212> TYPE: PRT
 <213> ORGANISM: Unknown
 <220> FEATURE:
 <223> OTHER INFORMATION: Triticum macha

<400> SEQUENCE: 59

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Met Lys Thr Phe Leu Ile Leu Ala Leu Leu Ala Ile Val Ala Thr Thr
1           5           10           15
Ala Thr Ile Ala Val Arg Val Pro Val Pro Gln Leu Gln Pro Gln Asn

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	20					25										30			
Pro	Ser	Gln	Gln	Gln	Pro	Gln	Glu	Gln	Val	Pro	Leu	Val	Gln	Gln	Gln				
	35						40					45							
Gln	Phe	Pro	Gly	Gln	Gln	Gln	Pro	Phe	Pro	Pro	Gln	Gln	Pro	Tyr	Pro				
	50					55					60								
Gln	Leu	Gln	Pro	Phe	Pro	Ser	Gln	Gln	Pro	Tyr	Met	Gln	Leu	Gln	Pro				
	65				70					75					80				
Phe	Pro	Gln	Pro	Gln	Leu	Pro	Tyr	Pro	Gln	Pro	Gln	Leu	Pro	Tyr	Pro				
				85					90					95					
Gln	Arg	Gln	Pro	Phe	Arg	Pro	Gln	Gln	Pro	Tyr	Pro	Gln	Pro	Gln	Pro				
	100								105					110					
Gln	Tyr	Ser	Gln	Pro	Gln	Gln	Pro	Ile	Ser	Gln	Gln	Gln	Gln	Gln	Gln				
	115						120						125						
Gln	Gln	Gln	Gln	Gln	Lys	Gln	Gln	Gln	Gln	Gln	Gln	Gln	Gln	Gln	Ile				
	130					135							140						
Gln	Pro	Gln	Ile	Leu	Gln	Gln	Gln	Leu	Ile	Pro	Cys	Arg	Asp	Val	Val				
	145				150					155					160				
Leu	Gln	Gln	His	Ser	Ile	Ala	Tyr	Gly	Ser	Ser	Gln	Val	Leu	Gln	Gln				
			165						170					175					
Ser	Thr	Tyr	Gln	Leu	Val	Gln	Gln	Leu	Cys	Cys	Gln	Gln	Leu	Trp	Gln				
			180					185						190					
Ile	Pro	Glu	Gln	Ser	Arg	Cys	Gln	Ala	Ile	His	Asn	Val	Val	His	Ala				
	195						200						205						
Ile	Ile	Leu	His	Gln	Gln	Gln	Lys	Gln	Gln	Gln	Gln	Gln	Gln	Gln	Lys				
	210					215							220						
Gln	Pro	Leu	Ser	Gln	Val	Ser	Phe	Gln	Gln	Pro	Gln	Gln	Gln	Tyr	Pro				
	225				230					235					240				
Ser	Gly	Gln	Gly	Ser	Phe	Gln	Pro	Ser	Gln	Gln	Asn	Pro	Gln	Ala	Gln				
				245					250					255					
Gly	Ser	Val	Gln	His	Gln	Gln	Leu	Pro	Gln	Phe	Glu	Glu	Ile	Arg	Lys				
			260					265						270					
Leu	Ala	Leu	Gln	Thr	Leu	Pro	Ala	Val	Cys	Asn	Val	Tyr	Ile	Pro	Pro				
	275						280						285						
Tyr	Cys	Ser	Thr	Thr	Thr	Ala	Pro	Phe	Gly	Ile	Phe	Gly	Thr	Asn					
	290					295					300								

<210> SEQ ID NO 60
 <211> LENGTH: 290
 <212> TYPE: PRT
 <213> ORGANISM: Secale cereale

<400> SEQUENCE: 60

Met	Lys	Thr	Phe	Leu	Ile	Leu	Ala	Leu	Leu	Ala	Ile	Val	Ala	Thr	Thr				
				5					10					15					
Ala	Thr	Ile	Ala	Val	Arg	Val	Pro	Val	Pro	Gln	Leu	Gln	Pro	Gln	Asn				
			20					25					30						
Pro	Ser	Gln	Gln	Gln	Pro	Gln	Glu	Gln	Val	Pro	Leu	Val	Gln	Gln	Gln				
		35					40					45							
Gln	Phe	Pro	Gly	Gln	Gln	Gln	Pro	Phe	Pro	Pro	Gln	Gln	Pro	Tyr	Pro				
	50					55					60								
Gln	Pro	Gln	Pro	Phe	Pro	Ser	Gln	Gln	Pro	Tyr	Leu	Gln	Leu	Gln	Pro				
	65				70					75					80				
Phe	Pro	Gln	Pro	Gln	Leu	Pro	Tyr	Pro	Gln	Pro	Gln	Leu	Pro	Tyr	Pro				
				85					90					95					

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Gln Arg Gln Pro Phe Arg Pro Gln Gln Pro Tyr Pro Gln Pro Gln Pro
 100 105 110

Gln Tyr Ser Gln Pro Gln Gln Pro Ile Ser Gln Gln Gln Gln Gln
 115 120 125

Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Ile Gln Pro Gln
 130 135 140

Ile Leu Gln Gln Gln Leu Ile Pro Cys Arg Asp Val Val Leu Gln Gln
 145 150 155 160

His Asn Ile Ala His Gly Ser Ser Gln Ile Leu Gln Gln Ser Thr Tyr
 165 170 175

Gln Leu Val Gln Gln Leu Cys Cys Gln Gln Leu Trp Gln Ile Pro Glu
 180 185 190

Gln Ser Arg Cys Gln Ala Ile His Asn Val Val His Ala Ile Ile Leu
 195 200 205

His Gln Gln Gln Gln Gln Pro Ser Ser Gln Val Ser Phe Gln Gln Pro
 210 215 220

Gln Gln Gln Tyr Pro Ser Gly Gln Gly Ser Phe Gln Pro Ser Gln Gln
 225 230 235 240

Asn Pro Gln Ala Gln Gly Ser Val Gln Pro Gln Gln Leu Pro Gln Phe
 245 250 255

Glu Glu Ile Arg Asn Leu Ala Leu Gln Thr Leu Pro Ala Met Cys Asn
 260 265 270

Val Tyr Ile Pro Pro Tyr Cys Thr Thr Ala Pro Val Gly Ile Phe Gly
 275 280 285

Thr Asn
 290

<210> SEQ ID NO 61
 <211> LENGTH: 291
 <212> TYPE: PRT
 <213> ORGANISM: Triticum spelta

<400> SEQUENCE: 61

Met Lys Thr Phe Leu Ile Leu Ala Leu Leu Ala Ile Val Ala Thr Thr
 1 5 10 15

Ala Thr Ile Ala Val Arg Val Pro Val Pro Gln Leu Gln Pro Gln Asn
 20 25 30

Pro Ser Gln Gln Gln Pro Gln Gly Gln Val Pro Leu Val Gln Gln Gln
 35 40 45

Gln Phe Leu Gly Gln Gln Gln Pro Phe Pro Pro Gln Gln Pro Tyr Pro
 50 55 60

Gln Pro Gln Pro Phe Pro Ser Gln Gln Pro Tyr Leu Gln Leu Gln Pro
 65 70 75 80

Phe Pro Gln Pro Gln Leu Pro Tyr Ser Gln Pro Gln Pro Phe Arg Pro
 85 90 95

Gln Gln Pro Tyr Pro Gln Pro Gln Pro Gln Tyr Ser Gln Pro Gln Gln
 100 105 110

Pro Ile Ser Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln
 115 120 125

Gln Gln Gln Gln Gln Gln Gln Gln Gln Ile Gln Pro Gln Ile Leu Gln
 130 135 140

Gln Gln Leu Ile Pro Cys Met Asp Val Leu Gln Gln His Asn Ile Ala
 145 150 155 160

His Gly Arg Ser Gln Val Leu Gln Gln Ser Thr Tyr Gln Leu Leu Gln
 165 170 175

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Glu Leu Cys Cys Gln His Leu Trp Gln Ile Pro Glu Gln Ser Gln Cys
 180 185 190
 Gln Ala Ile His Asn Val Val His Ala Ile Ile Leu His Gln Gln Gln
 195 200 205
 Lys Gln Gln Gln Gln Leu Ser Ser Gln Val Ser Phe Gln Gln Pro Gln
 210 215 220
 Gln Gln Tyr Pro Leu Gly Gln Gly Ser Phe Arg Pro Ser Gln Gln Asn
 225 230 235 240
 Ser Gln Ala Gln Gly Ser Val Gln Pro Gln Gln Leu Pro Gln Phe Gln
 245 250 255
 Glu Ile Arg Asn Leu Ala Leu Gln Thr Leu Pro Ala Met Cys Asn Val
 260 265 270
 Tyr Ile Pro Pro Tyr Cys Ser Thr Thr Thr Ala Pro Phe Gly Ile Phe
 275 280 285
 Gly Thr Asn
 290

<210> SEQ ID NO 62
 <211> LENGTH: 299
 <212> TYPE: PRT
 <213> ORGANISM: Triticum urartu

<400> SEQUENCE: 62

Met Lys Thr Phe Leu Ile Leu Ala Leu Leu Ala Ile Val Ala Thr Thr
 1 5 10 15
 Ala Thr Thr Ala Val Arg Val Pro Val Pro Gln Leu Gln Pro Gln Asn
 20 25 30
 Pro Ser Gln Gln Gln Pro Gln Glu Gln Val Pro Leu Val Gln Gln Gln
 35 40 45
 Gln Phe Leu Gly Gln Gln Gln Pro Phe Pro Pro Gln Gln Pro Tyr Pro
 50 55 60
 Glu Pro Gln Pro Phe Pro Ser Gln Gln Pro Tyr Leu Gln Leu Gln Pro
 65 70 75 80
 Phe Pro Gln Pro Gln Leu Pro Tyr Ser Gln Pro Gln Pro Phe Arg Pro
 85 90 95
 Gln Gln Pro Tyr Pro Gln Pro Gln Pro Gln Tyr Ser Gln Pro Gln Gln
 100 105 110
 Pro Ile Ser Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln
 115 120 125
 Gln Gln Gln Gln Gln Gln Gln Gln Glu Gln Gln Ile Gln Pro Gln Ile
 130 135 140
 Leu Gln Gln Gln Leu Ile Pro Cys Arg Asp Val Ile Val Leu Gln Gln
 145 150 155 160
 His Asn Ile Ala His Glu Ser Ser Gln Val Leu Gln Gln Ser Ser Tyr
 165 170 175
 Gln Val Leu Gln Gln Leu Cys Cys Gln Gln Leu Trp Gln Ile Pro Glu
 180 185 190
 Gln Ser Arg Cys Gln Ala Ile His Asn Val Val His Ala Ile Ile Leu
 195 200 205
 His Gln Gln Gln Gln Gln Gln Gln Gln Val Gln Gln Gln Pro Ser Ser
 210 215 220
 Gln Val Ser Tyr Gln Gln Pro Gln Gln Gln Tyr Pro Ser Gly Gln Gly
 225 230 235 240
 Ser Phe Gln Pro Ser Gln Gln Asn Pro Gln Ala Gln Gly Phe Val Gln

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245	250	255
Pro Gln His Leu Pro Gln Phe Glu Glu Ile Arg Asn Leu Ala Leu Gln 260 265 270		
Thr Leu Pro Ala Met Cys Asn Val Tyr Ile Pro Pro Tyr Cys Ser Thr 275 280 285		
Thr Thr Ala Pro Phe Gly Ile Phe Gly Thr Asn 290 295		
<210> SEQ ID NO 63 <211> LENGTH: 316 <212> TYPE: PRT <213> ORGANISM: Unknown <220> FEATURE: <223> OTHER INFORMATION: Gliadin reference sequence, unknown species <400> SEQUENCE: 63		
Met Lys Thr Phe Leu Ile Leu Ala Leu Leu Ala Ile Val Ala Thr Thr 1 5 10 15		
Ala Thr Thr Ala Val Arg Val Pro Val Pro Gln Leu Gln Pro Gln Asn 20 25 30		
Pro Ser Gln Gln Gln Pro Gln Glu Gln Val Pro Leu Val Gln Gln Gln 35 40 45		
Gln Phe Pro Gly Gln Gln Gln Gln Phe Pro Pro Gln Gln Pro Tyr Pro 50 55 60		
Gln Pro Gln Pro Phe Pro Ser Gln Gln Pro Tyr Leu Gln Leu Gln Pro 65 70 75 80		
Phe Pro Gln Pro Gln Pro Phe Pro Pro Leu Pro Tyr Pro Gln Pro Gln 85 90 95		
Ser Phe Ser Pro Gln Gln Pro Tyr Pro Gln Gln Gln Pro Gln Tyr Leu 100 105 110		
Gln Pro Gln Gln Pro Ile Ser Gln Gln Gln Ala Gln Gln Gln Gln Gln 115 120 125		
Gln Gln Gln Gln Gln Gln Gln Gln Gln Gln Ile Gln Pro Gln Ile Leu 130 135 140		
Gln Gln Gln Leu Ile Pro Cys Arg Asp Val Val Leu Gln Gln His Asn 145 150 155 160		
Ile Ala His Ala Ser Ser Gln Val Leu Gln Gln Ser Thr Tyr Gln Leu 165 170 175		
Leu Gln Gln Leu Cys Cys Gln Gln Leu Leu Gln Ile Pro Glu Gln Ser 180 185 190		
Gln Cys Gln Ala Ile His Asn Val Ala His Ala Ile Ile Met His Gln 195 200 205		
Gln Gln Gln Gln Gln Gln Glu Gln Lys Gln Gln Leu Gln Gln Gln Gln 210 215 220		
Gln Gln Gln Gln Gln Leu Gln Gln Gln Gln Gln Gln Gln Pro Ser 225 230 235 240		
Ser Gln Val Ser Phe Gln Gln Pro Gln Gln Tyr Pro Ser Ser Gln 245 250 255		
Val Ser Phe Gln Pro Ser Gln Leu Asn Pro Gln Ala Gln Gly Ser Val 260 265 270		
Gln Pro Gln Gln Leu Pro Gln Phe Ala Glu Ile Arg Asn Leu Ala Leu 275 280 285		

-continued

Gln Thr Leu Pro Ala Met Cys Asn Val Tyr Ile Pro Pro His Cys Ser
290 295 300

Thr Thr Ile Ala Pro Phe Gly Ile Ser Gly Thr Asn
305 310 315

What is claimed is:

1. A method for determining effective sterilization, deimmunization, and/or disinfection of equipment and/or supplies by a device, the method comprising:
providing a defined surrogate protein having a predetermined sequence defined by a sequence which is at least about 95% homologous to the sequence

6. The method of claim 1 in which the protein analysis procedure includes one or more of: a Western Blot analysis, a protein assay analysis, a magnetic separation analysis, a peptide analysis, a mass spectrometry analysis, and a gas chromatography analysis.

(SEQ ID NO: 1)
10 20 30 40 50
MNYNYSKAYE VP IAIQDRSF NEDGSLNFPS EGDNPTIHPY WQPEFFGDTI
MVNGRVWPNM NVDMTRYRFR LLNGSNARFY NLKFSNGMQF WQIGTDGGYL
NKPVPLTSLI ISPGERADIL VDFTEIPAGT RIILNNDANA PYPTGDAPDK
DTTGQIMQFT VQHNDHHHHH H

representative of an infectious agent potentially contaminating the equipment and/or the supplies to be sterilized, deimmunized, and/or disinfected by the device;

subjecting the defined surrogate protein having the predetermined sequence to sterilization, deimmunization, or disinfection; and

determining the effectiveness of the sterilization, deimmunization, and/or disinfection by determining if the defined surrogate protein having the predetermined sequence has been destroyed; and wherein the sterilization, deimmunization, and/or disinfection occurs when the defined surrogate protein has been fragmented to yield a negative result in a protein analysis procedure.

2. The method of claim 1 in which the defined surrogate protein is a protein critical for stability, growth and/or infectious capacity of infectious agents.

3. The method of claim 1 in which the defined surrogate protein is a protein critical for stability, growth and/or infectious capacity of surrogate organisms of infectious agents.

4. The method of claim 2 in which the infectious agent includes one or more of: an infectious protein, an infectious spore forming bacteria, an infectious vegetative bacteria, an infectious fungus, and an infectious virus.

7. The method of claim 1 in which the protein analysis procedure includes fluorescence analysis of proteins covalently crosslinked on a solid surface.

8. The method of claim 1 in which the protein analysis procedure includes fluorescence analysis of proteins covalently crosslinked on magnetic beads.

9. The method of claim 1 in which the defined surrogate protein having the predetermined sequence is disposed on a surface.

10. The method of claim 1 in which the defined surrogate protein having the predetermined sequence is disposed on a test strip.

11. The method of claim 1 in which the defined surrogate protein having the predetermined sequence is disposed in or on a vessel.

12. The method of claim 1 in which the surrogate protein having the predetermined sequence is disposed on a tube.

13. The method of claim 1 in which the surrogate protein having the predetermined sequence is disposed on a holder.

14. The method of claim 13 in which the holder is disposed to receive a flow of a sterilization agent, a deimmunization agent or a disinfection agent.

15. A method for determining effective sterilization, deimmunization, and/or disinfection of equipment and/or supplies by a device, the method comprising:

providing a defined surrogate protein having a predetermined sequence defined by the sequence:

(SEQ ID NO: 1)
10 20 30 40 50
MNYNYSKAYE VP IAIQDRSF NEDGSLNFPS EGDNPTIHPY WQPEFFGDTI
MVNGRVWPNM NVDMTRYRFR LLNGSNARFY NLKFSNGMQF WQIGTDGGYL
NKPVPLTSLI ISPGERADIL VDFTEIPAGT RIILNNDANA PYPTGDAPDK
DTTGQIMQFT VQHNDHHHHH H

5. The method of claim 1 in which the defined surrogate protein is a pathogenic protein, a protein critical for the growth of infectious agents, and an immunogenic protein.

representative of an infectious agent potentially contaminating the equipment and/or the supplies to be sterilized, deimmunized, and/or disinfected by the device;

subjecting the defined surrogate protein having the predetermined sequence to sterilization, deimmunization, or disinfection; and
determining the effectiveness of the sterilization, deimmunization, and/or disinfection by determining if the defined surrogate protein having the predetermined sequence has been destroyed; and wherein the sterilization, deimmunization, and/or disinfection occurs when the defined surrogate protein has been fragmented to yield a negative result in a protein analysis procedure.

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