



(12) **DEMANDE DE BREVET CANADIEN
CANADIAN PATENT APPLICATION**

(13) **A1**

(86) Date de dépôt PCT/PCT Filing Date: 2017/02/17
(87) Date publication PCT/PCT Publication Date: 2017/09/14
(85) Entrée phase nationale/National Entry: 2018/08/02
(86) N° demande PCT/PCT Application No.: US 2017/018249
(87) N° publication PCT/PCT Publication No.: 2017/155677
(30) Priorité/Priority: 2016/03/08 (US62/305,382)

(51) Cl.Int./Int.Cl. *A61K 9/18* (2006.01),
A61M 1/00 (2006.01), *B01J 20/22* (2006.01),
B01J 20/28 (2006.01), *B01J 20/285* (2006.01)

(71) Demandeur/Applicant:
CYTOSORBENTS CORPORATION, US

(72) Inventeurs/Inventors:
GRUDA, MARYANN, US;
O'SULLIVAN, PAMELA, US;
GULIASHVILI, TAMAZ, US;
SCHEIRER, ANDREW, US;
GOLOBISH, THOMAS, US;
CAPONI, VINCENT, US;
CHAN, PHILLIP, US

(74) Agent: SMART & BIGGAR

(54) Titre : UTILISATION D'UN SORBANT DE TYPE PERLE DE POLYMERE POREUX HEMOCOMPATIBLE PERMETTANT L'ELIMINATION DES MOLECULES A MOTIFS MOLECULAIRES ASSOCIES A DES AGENTS PATHOGENES (PAMP) ET MOLECULES A MOTIFS MOLECULAIRES ASSOCIES A DES LESIONS (DAMP)
(54) Title: THE USE OF A HEMOCOMPATIBLE POROUS POLYMER BEAD SORBENT FOR REMOVAL OF PAMPS AND DAMPS

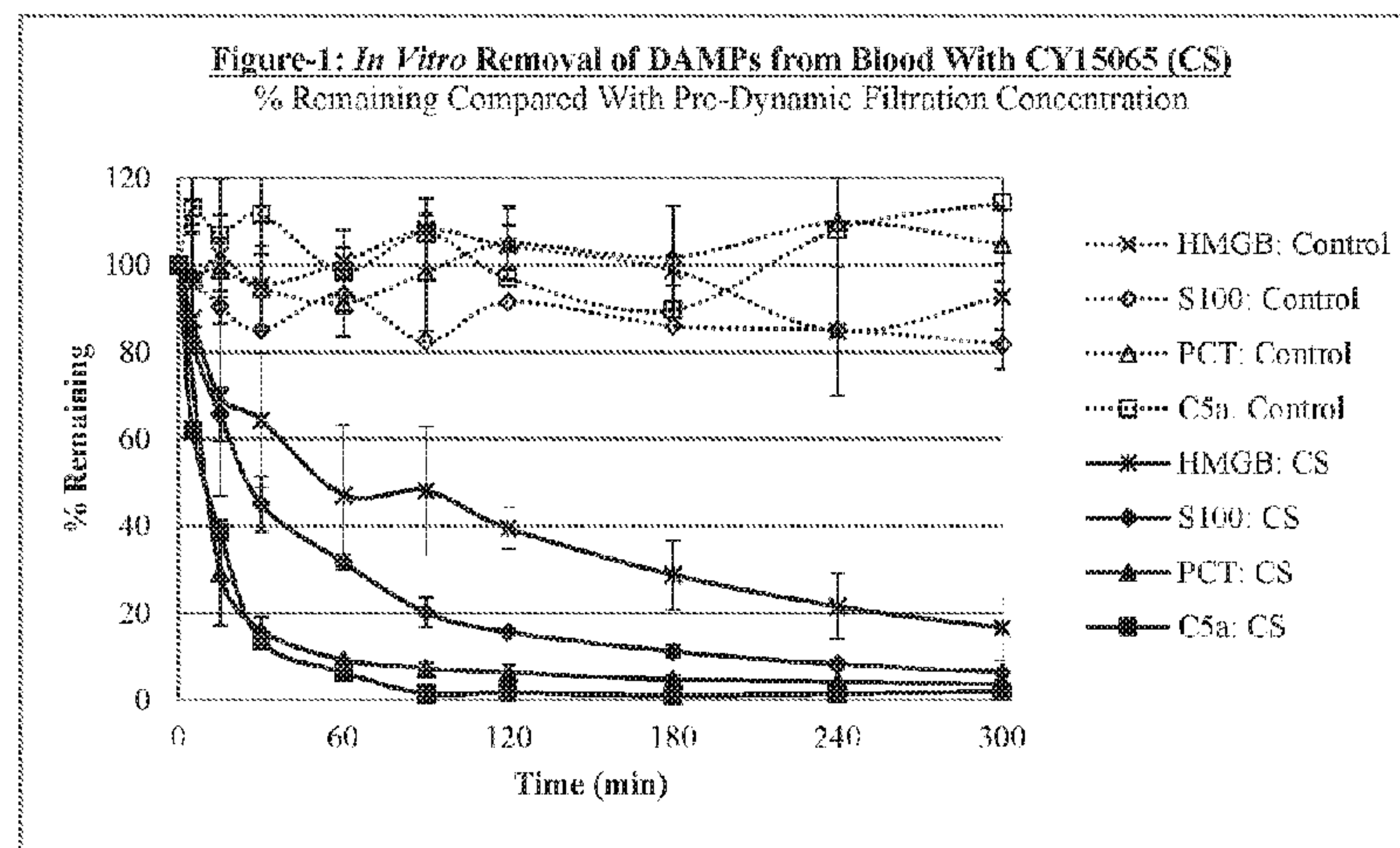


FIGURE 1

(57) **Abrégé/Abstract:**

The invention concerns biocompatible polymer systems comprising at least one polymer sorbent with a plurality of pores, said polymer designed to adsorb pathogen-associated molecular pattern molecules and damage-associated molecular pattern molecules. Also disclosed herein are methods for reducing contamination in a biological substance, or treating contamination in a subject, by one or more pathogen-associated molecular pattern molecules and damage-associated molecular pattern molecules, by contacting the biological substance with an effective amount of sorbent capable of sorbing the toxin.

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property
Organization
International Bureau(43) International Publication Date
14 September 2017 (14.09.2017)

WIPO | PCT

(10) International Publication Number
WO 2017/155677 A1

(51) International Patent Classification:

A61K 9/18 (2006.01) *B01J 20/22* (2006.01)
A61M 1/00 (2006.01) *B01J 20/28* (2006.01)
B01J 20/285 (2006.01)

(21) International Application Number:

PCT/US2017/018249

(22) International Filing Date:

17 February 2017 (17.02.2017)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

62/305,382 8 March 2016 (08.03.2016) US

(71) Applicant: **CYTOSORBENTS CORPORATION**
[US/US]; 7 Deer Park Drive, Suite K, Monmouth Junction,
NJ 08852 (US).(72) Inventors: **GRUDA, Maryann**; c/o 7 Deer Park Drive,
Suite K, Monmouth Junction, NJ 08852 (US). **O'SULLI-
VAN, Pamela**; c/o 7 Deer Park Drive, Suite K, Monmouth
Junction, NJ 08852 (US). **GULIASHVILI, Tamaz**; c/o 7
Deer Park Drive, Suite K, Monmouth Junction, NJ 08852
(US). **SCHEIRER, Andrew**; 3425 Kendall Court, Bethle-
hem, PA 18020 (US). **GOLOBISH, Thomas**; 16 Ever-
green Circle, Princeton, NJ 08540 (US). **CAPPONI, Vin-**cent; 204 Heskens Court, Monmouth Junction, NJ 08852
(US). **CHAN, Phillip**; 1004 Owl Lane, Cherry Hill, NJ
08003 (US).(74) Agents: **ROSEDALE, Jeffrey, H.** et al.; Baker &
Hostetler LLP, 2929 Arch Street, Cira Centre, 12th Floor,
Philadelphia, PA 19104-2891 (US).(81) Designated States (*unless otherwise indicated, for every
kind of national protection available*): AE, AG, AL, AM,
AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY,
BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DJ, DK, DM,
DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT,
HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KH, KN,
KP, KR, KW, KZ, LA, LC, LK, LR, LS, LU, LY, MA,
MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG,
NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS,
RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY,
TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN,
ZA, ZM, ZW.(84) Designated States (*unless otherwise indicated, for every
kind of regional protection available*): ARIPO (BW, GH,
GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ,
TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU,
TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE,
DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU,
LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK,

[Continued on next page]

(54) Title: THE USE OF A HEMOCOMPATIBLE POROUS POLYMER BEAD SORBENT FOR REMOVAL OF PAMPS AND DAMPS

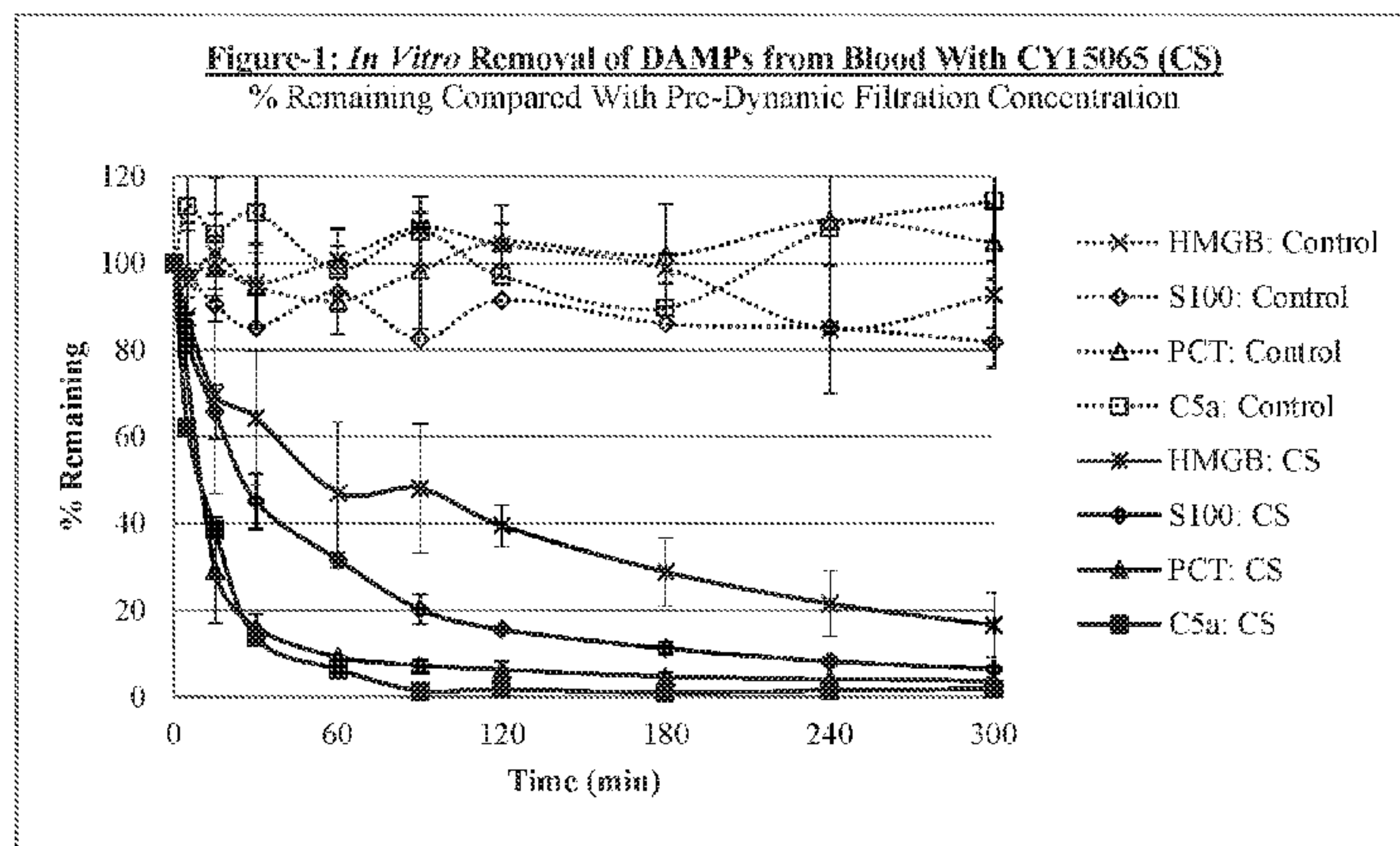


FIGURE 1

(57) **Abstract:** The invention concerns biocompatible polymer systems comprising at least one polymer sorbent with a plurality of pores, said polymer designed to adsorb pathogen-associated molecular pattern molecules and damage-associated molecular pattern molecules. Also disclosed herein are methods for reducing contamination in a biological substance, or treating contamination in a subject, by one or more pathogen-associated molecular pattern molecules and damage-associated molecular pattern molecules, by contacting the biological substance with an effective amount of sorbent capable of sorbing the toxin.

WO 2017/155677 A1



SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Published:
— *with international search report (Art. 21(3))*

THE USE OF A HEMOCOMPATIBLE POROUS POLYMER BEAD SORBENT FOR REMOVAL OF PAMPS AND DAMPS

CROSS REFERENCE TO RELATED APPLICATIONS

[0001] The present application claims benefit of U.S. Patent Application No. 62/305,382 filed on March 8, 2016, the disclosure of which is incorporated herein in its entirety.

GOVERNMENT RIGHTS

[0002] The subject matter disclosed herein was made with government support under contract number N66001-12-C-4199, awarded by The Defense Advanced Research Projects Agency (DARPA). The government may have certain rights in the herein disclosed subject matter.

TECHNICAL FIELD

[0003] The disclosed inventions are in the field of porous polymeric sorbents. The disclosed inventions are also in the field of broadly reducing pathogen-associated molecular pattern molecules and damage-associated molecular pattern molecules in blood, blood products, and other physiologic fluids. Additionally, the disclosed inventions are in the field of broadly removing pathogen-associated molecular pattern molecules and damage-associated molecular pattern molecules by static adsorption, perfusion, or hemoperfusion.

BACKGROUND

[0004] Prolonged and upregulated inflammatory responses may lead to sepsis or systemic inflammatory response syndrome (SIRS), both of which can progress to potentially fatal septic shock and multiple organ dysfunction syndrome (MODS). Sepsis and septic shock result from a life-threatening systemic inflammatory response syndrome (SIRS) to invading pathogens or direct tissue insults. Sepsis is a highly heterogeneous disease with severity and progression dependent upon a myriad of interacting factors, including: the microbial insult, which may be of bacterial (gram-positive and gram-negative), viral, fungal or parasitic origin; the pathogen load, toxin production, virulence; host factors such as age, genetic composition, and comorbidities; the

site of infection as well as the elapsed time since the initial infection. This complexity creates a highly dynamic and unstable situation that has confounded therapeutic efforts targeted to specific factors.

[0005] Examples of pathogens commonly associated with the development of sepsis are *Staphylococcus species including Staphylococcus aureus (S. aureus)*, *Streptococcus species such as Streptococcus pneumonia, Streptococcus pyogenes (S. pyogenes)*, *Klebsiella species*, *Escherichia coli (E. coli)*, *Pseudomonas species such as Pseudomonas aeruginosa (P. aeruginosa)*, *Listeria species*, several fungal species (e.g. *Aspergillus*, *Fusarium* and *Candida* subspecies, as well as viruses (such as Dengue and influenza viruses) and parasites. These pathogens release or cause the release of a daunting array of virulence factors that modulate the immune response and influence the severity of the disease. The host response to pathogenic insults involves multiple sequential and concurrent processes that produce both exaggerated inflammation and immune suppression. Pathogen-associated molecular pattern molecules (PAMPs), such as lipopolysaccharides, lipopeptides, lipoteichoic acid, peptidoglycans, nucleic acids such as double-stranded RNA, toxins and flagellins, trigger an immune response in the host (e.g. the innate immune system) to fight the infection, leading to the production of high levels of inflammatory and anti-inflammatory mediators, such as cytokines. PAMPs and high cytokine levels, as well as direct tissue injury (trauma, burns, etc.), can damage tissue, causing the extracellular release of damage-associated molecular pattern (DAMPs) molecules into the bloodstream. DAMPs are a broad class of endogenous molecules, which like PAMPs, trigger the immune response through pattern recognition receptors (PRRs) such as Toll-like receptors (TLRs).

[0006] DAMPs have also been associated with countless syndromes and diseases. These include complications from trauma, burns, traumatic brain injury and invasive surgery, and also organ-specific illnesses like liver disease, kidney dialysis complications, and autoimmune diseases. DAMPs are host molecules that can initiate and perpetuate noninfectious SIRS and exacerbate infectious SIRS. DAMPs are a diverse family of molecules that are intracellular in physiological conditions and many are nuclear or cytosolic proteins. DAMPs can be divided into two groups: (1) molecules that perform noninflammatory functions in living cells (such as HMGB1) and acquire immunomodulatory properties when released, secreted, modified, or exposed on the cell surface during cellular stress, damage, or injury, or (2) alarmins, i.e.,

molecules that possess cytokine-like functions (such as β -Defensins and Cathelicidin), which can be stored in cells and released upon cell lysis, whereupon they contribute to the inflammatory response. When released outside the cell or exposed on the surface of the cell following tissue injury, they move from a reducing to an oxidizing milieu, which affects their activity. Also, following necrosis, mitochondrial and nuclear DNA fragments are released outside the cell becoming DAMPs.

[0007] DAMPs, such as HMGB-1, heat-shock and S100 proteins are normally found inside cells and are released by tissue damage. DAMPs act as endogenous danger signals to promote and exacerbate the inflammatory response. HMGB-1 is a non-histone nuclear protein that is released under stress conditions. Extracellular HMGB-1 is an indicator of tissue necrosis and has been associated with an increased risk of sepsis and multiple organ dysfunction syndrome (MODS). S100 A8 (granulin A, MRP8) and A9 (granulin B, MRP14) homo and heterodimers bind to and signal directly via the TLR4/lipopolysaccharide receptor complex where they become danger signals that activate immune cells and vascular endothelium. Procalcitonin is a marker of severe sepsis caused by bacteria and its release into circulation is indicative of the degree of sepsis. Serum amyloid A (SAA), an acute-phase protein, is produced predominantly by hepatocytes in response to injury, infection, and inflammation. During acute inflammation, serum SAA levels may rise by 1000-fold. SAA is chemotactic for neutrophils and induces the production of proinflammatory cytokines. Heat shock proteins (HSP) are a family of proteins that are produced by cells in response to exposure to stressful conditions and are named according to their molecular weight (10, 20-30, 40, 60, 70, 90). The small 8-kilodalton protein ubiquitin, which marks proteins for degradation, also has features of a heat shock protein. Hepatoma-derived growth factor (HDGF), despite its name, is a protein expressed by neurons. HDGF can be released actively by neurons via a nonclassical pathway and passively by necrotic cells. Other factors, such as complement factors 3 and 5, are activated as part of the host defense against pathogens but can also contribute to the adverse outcomes in sepsis. Excessive, persistent circulating levels of cytokines and DAMPs contribute to organ injury and identify those patients who have the highest risk of multiple organ dysfunction (MODs) and death in community acquired pneumonia and sepsis.

[0008] *Staphylococcus aureus*, the leading cause of gram positive bacteremia, is associated with higher morbidity and mortality largely due to the increase in methicillin-resistant

S. aureus (MRSA). *S. aureus* is effective in invading the bloodstream and evading the host immunological response due to a variety of PAMPs, such as Pantan-Valentine leukocidin (PVL), a cytotoxin produced by many *S. aureus* clinical isolates that functions as a key virulence factor by forming pores in cell membranes. *Streptococcus pneumoniae* and *Listeria monocytogenes* are also gram-positive bacteria that produce the pore forming toxins pneumolysin, streptolysin and listeriolysin that facilitate infection by damaging host cells and interfering with the host immune response.

[0009] Superantigens are a class of antigens that cause non-specific activation of T-cells resulting in polyclonal T cell activation and massive cytokine release. Superantigens are produced by some pathogenic viruses and bacteria most likely as a defense mechanism against the immune system. *Staphylococcal* and *Streptococcal* superantigens form a large protein family having all evolved from a single primordial superantigen. In particular, *Streptococcus pyrogenic* exotoxins (SPEs) A, C, G-M, *S. aureus* TSST-1 toxin, and *Y. pseudotuberculosis* YPM-a and YPM-b are superantigens. The nucleocapsid (N) protein of rabies virus is reported to be a superantigen in humans, stimulating Vb8T lymphocytes.

[0010] *Staphylococcal* A and B (ETA and ETB), that produces Staphylococcal Scalded Skin Syndrome (SSSS) are serine proteases that belong to the class of exfoliative toxins. Hypotension and possible organ failure can be found in severe cases of SSSS where there are extensive areas of denuded skin with significant fluid loss or with a secondary infecting organism.

[0011] *Streptococcus pyogenes* is a group A streptococcus (GAS) that utilizes several virulence factors, Spe A to G, to establish infection. Of these, the streptococcus pyrogenic exotoxin B (SpeB), cleaves or degrades host immunoglobulin and complement components to evade the immune response by inhibiting phagocytic activity (Kuo 2008).

[0012] Bacterial flagellins are bacterial structural protein that elicits immune response via toll-like receptor 5, a PRR. Flagellins are extraordinarily potent proinflammatory stimuli in the lungs during sepsis. Flagellins induce a local release of proinflammatory cytokines, the accumulation of inflammatory cells, and the development of pulmonary hyperpermeability. Numerous forms of flagellin are made by bacteria with *E. coli* produced flagellin ranging in size from 37 to 69 kDa.

[0013] Over 20 *Aspergillus* species are known to cause human disease. Invasive aspergillosis is a devastating infectious disease that mainly affects critically ill and immunocompromised patients. *Aspergillus fumigatus* is the most prevalent and is largely responsible for the increased incidence of invasive aspergillosis (IA) in the immunocompromised patient population. IA is a devastating illness, with mortality rates in some patient groups reaching as high as 90%. *Aspergillus* species produce a variety of mycotoxins, such as gliotoxin, that contribute to pathogenicity by host immunosuppression, and aflatoxin that can cause acute hepatic injury and liver failure. *Fusarium* species cause a broad spectrum of infections in humans, including superficial, locally invasive, and disseminated infections. *Fusarium* species possess several virulence factors, including mycotoxins, such as T-2 toxin, a trichothecene mycotoxin, which suppresses humoral and cellular immunity and may also cause tissue breakdown.

SUMMARY

[0014] In some aspects, the invention concerns a biocompatible polymer system comprising at least one polymer; the polymer system capable of adsorbing (i) pathogen-associated molecular pattern molecules and (ii) damage-associated molecular pattern molecules having a molecular weight of from less than about 0.5 kDa to about 1,000 kDa (or about 1 kDa to about 1,000 kDa or about 0.1 kDa to about 1,000 kDa in some embodiments). Some preferred polymers are hemocompatible. Certain preferred polymer systems have geometry of a spherical bead.

[0015] Some polymer systems have a polymer pore structure that has a total volume of pore sizes in the range of from 50 Å to 40,000 Å greater than 0.5 cc/g and less than 5.0 cc/g dry polymer.

[0016] In some embodiments, the toxins adsorbed comprise one or more of PAMPs and DAMPS comprised of one or more of flagellins, lipopeptides, formyl peptides, mycotoxins, exotoxins, endotoxins, lipoteichoic acid, cytolysins, superantigens, proteases, lipases, amylases, enzymes, peptides including bradykinin, activated complement, soluble receptors, soluble CD40 ligand, bioactive lipids, oxidized lipids, cellular DNA, mitochondrial DNA, pathogen or host derived RNA, cell-free hemoglobin, cell-free myoglobin, growth factors, peptidoglycans, glycoproteins, released intracellular components, cell wall or viral envelope components,

Polyinosinic:polycytidylic acid (poly I:C), prions, toxins, bacterial and viral toxins, drugs, vasoactive substances, and foreign antigens.

[0017] The polymers can be made by any means known in the art to produce a suitable porous polymer. In some embodiments, the polymer is made using suspension polymerization. Some polymers comprise a hypercrosslinked polymer. Certain spherical beads have a biocompatible hydrogel coating.

[0018] Certain polymers are formed and subsequently modified to be biocompatible. Some modifications comprise forming a biocompatible surface coating or layer.

[0019] Other aspects include methods of perfusion comprising passing a physiologic fluid once, or by way of a suitable extracorporeal circuit, through a device comprising the biocompatible polymer system one or more times described herein.

[0020] Yet another aspect concerns devices for removing (i) pathogen-associated molecular pattern molecules and (ii) damage-associated molecular pattern molecules from less than 0.5 kDa to 1,000 kDa from physiologic fluid comprising the biocompatible polymer system described herein.

BRIEF DESCRIPTION OF THE DRAWINGS

[0021] The accompanying drawings, which are included to provide a further understanding of the disclosure, are incorporated in and constitute a part of this specification, illustrate aspects of the disclosure and together with the detailed description serve to explain the principles of the disclosure. No attempt is made to show structural details of the disclosure in more detail than may be necessary for a fundamental understanding of the disclosure and the various ways in which it may be practiced. In the drawings:

[0022] **Figures 1 and 2** present DAMPs and PAMPs removal data from an *in vitro* dynamic model, using whole blood, expressed as percentage remaining compared to the pre-circulation concentrations for modified polymer CY15065.

[0023] **Figures 3 and 4** presents DAMPs and PAMPs removal data from an *in vitro* dynamic model, using whole blood, expressed as percentage remaining compared to the pre-circulation concentrations for polymer CY15077.

DETAILED DESCRIPTION OF ILLUSTRATIVE EMBODIMENTS

[0024] As required, detailed embodiments of the present invention are disclosed herein; it is to be understood that the disclosed embodiments are merely exemplary of the invention that may be embodied in various forms. Therefore, specific structural and functional details disclosed herein are not to be interpreted as limits, but merely as a basis for teaching one skilled in the art to employ the present invention. The specific examples below will enable the invention to be better understood. However, they are given merely by way of guidance and do not imply any limitation.

[0025] The present invention may be understood more readily by reference to the following detailed description taken in connection with the accompanying figures and examples, which form a part of this disclosure. It is to be understood that this invention is not limited to the specific materials, devices, methods, applications, conditions or parameters described and/or shown herein, and that the terminology used herein is for the purpose of describing particular embodiments by way of example only and is not intended to be limiting of the claimed invention. The term “plurality”, as used herein, means more than one. When a range of values is expressed, another embodiment includes from the one particular value and/or to the other particular value. Similarly, when values are expressed as approximations, by use of the antecedent “about,” it will be understood that the particular value forms another embodiment. All ranges are inclusive and combinable.

[0026] It is to be appreciated that certain features of the invention, which are, for clarity, described herein in the context of separate embodiments, may also be provided in combination in a single embodiment. Conversely, various features of the invention that are, for brevity, described in the context of a single embodiment, may also be provided separately or in any subcombination. Further reference to values stated in ranges includes each and every value within that range.

[0027] The following definitions are intended to assist in understanding the present invention:

[0028] Pathogen-associated molecular pattern molecules (PAMPS) are molecules derived from microorganisms that are recognized by cells of the innate immune system. These molecules have small molecular motifs conserved within a class of microbes that are recognized

by toll-like receptor and pattern recognition receptors that initiate and perpetuate a pathogen-induced inflammatory response.

[0029] Damage-associated molecular pattern molecules (DAMPS) are host biomolecules released by stressed cells that initiate and perpetuate inflammation in response to trauma, ischemia, and tissue damage either in the absence or presence of pathogenic infection.

[0030] The term “biocompatible” is defined to mean the sorbent is capable of coming in contact with physiologic fluids, living tissues, or organisms, without producing unacceptable clinical changes during the time that the sorbent is in contact with the physiologic fluids, living tissues, or organisms.

[0031] The term “hemocompatible” is defined as a condition whereby a biocompatible material when placed in contact with whole blood or blood plasma results in clinically acceptable physiologic changes.

[0032] As used herein, the term “sorbent” includes adsorbents and absorbents.

[0033] For purposes of this invention, the term “sorb” is defined as “taking up and binding by absorption and adsorption”.

[0034] For the purposes of this invention, the term “perfusion” is defined as passing a physiologic fluid, once through or by way of a suitable extracorporeal circuit, through a device containing the porous polymeric adsorbent to remove toxic molecules from the fluid.

[0035] The term “hemoperfusion” is a special case of perfusion where the physiologic fluid is blood.

[0036] The term “dispersant” or “dispersing agent” is defined as a substance that imparts a stabilizing effect upon a finely divided array of immiscible liquid droplets suspended in a fluidizing medium.

[0037] The term “macroreticular synthesis” is defined as a polymerization of monomers into polymer in the presence of an inert precipitant which forces the growing polymer molecules out of the monomer liquid at a certain molecular size dictated by the phase equilibria to give solid nanosized microgel particles of spherical or almost spherical symmetry packed together to give a bead with physical pores of an open cell structure [U.S. Patent 4,297,220, Meitzner and Oline, October 27, 1981; R.L.Albright, Reactive Polymers, 4, 155-174(1986)].

[0038] The term “hypercrosslinked” describes a polymer in which the single repeating unit has a connectivity of more than two. Hypercrosslinked polymers are prepared by

crosslinking swollen, or dissolved, polymer chains with a large number of rigid bridging spacers, rather than copolymerization of monomers. Crosslinking agents may include bis(chloromethyl) derivatives of aromatic hydrocarbons, methylal, monochlorodimethyl ether, and other bifunctional compounds that react with the polymer in the presence of Friedel-Crafts catalysts [Tsyurupa, M. P., Z. K. Blinnikova, N. A. Proskurina, A. V. Pastukhov, L. A. Pavlova, and V. A. Davankov. "Hypercrosslinked Polystyrene: The First Nanoporous Polymeric Material." *Nanotechnologies in Russia* 4 (2009): 665-75.]

[0039] Some preferred polymers comprise residues from one or more monomers, or containing monomers, or mixtures thereof, selected from acrylonitrile, allyl glycidyl ether, butyl acrylate, butyl methacrylate, cetyl acrylate, cetyl methacrylate, 3,4-dihydroxy-1-butene, dipentaerythritol diacrylate, dipentaerythritol dimethacrylate, dipentaerythritol tetraacrylate, dipentaerythritol tetramethacrylate, dipentaerythritol triacrylate, dipentaerythritol trimethacrylate, divinylbenzene, divinylformamide, divinylanthracene, divinylsulfone, 3,4-epoxy-1-butene, 1,2-epoxy-9-decene, 1,2-epoxy-5-hexene, ethyl acrylate, ethyl methacrylate, ethylstyrene, ethylvinylbenzene, glycidyl methacrylate, methyl acrylate, methyl methacrylate, octyl acrylate, octyl methacrylate, pentaerythritol diacrylate, pentaerythritol dimethacrylate, pentaerythritol tetraacrylate, pentaerythritol tetramethacrylate, pentaerythritol triacrylate, pentaerythritol trimethacrylate, styrene, trimethylolpropane diacrylate, trimethylolpropane dimethacrylate, trimethylolpropane triacrylate, trimethylolpropane trimethacrylate, trivinylbenzene, trivinylcyclohexane, vinyl acetate, vinylbenzyl alcohol, 4-vinyl-1-cyclohexene 1,2-epoxide, vinylformamide, vinylanthracene, 2-vinylloxirane, and vinyltoluene.

[0040] Some embodiments of the invention use an organic solvent and/or polymeric porogen as the porogen or pore-former, and the resulting phase separation induced during polymerization yield porous polymers. Some preferred porogens are selected from, or mixtures comprised of any combination of, benzyl alcohol, cyclohexane, cyclohexanol, cyclohexanone, decane, dibutyl phthalate, di-2-ethylhexyl phthalate, di-2-ethylhexylphosphoric acid, ethylacetate, 2-ethyl-1-hexanoic acid, 2-ethyl-1-hexanol, n-heptane, n-hexane, isoamyl acetate, isoamyl alcohol, n-octane, pentanol, poly(propylene glycol), polystyrene, poly(styrene-co-methyl methacrylate), tetraline, toluene, tri-n-butylphosphate, 1,2,3-trichloropropane, 2,2,4-trimethylpentane, and xylene.

[0041] In yet another embodiment, the dispersing agent is selected from a group consisting of hydroxyethyl cellulose, hydroxypropyl cellulose, poly(diethylaminoethyl acrylate), poly(diethylaminoethyl methacrylate), poly(dimethylaminoethyl acrylate), poly(dimethylaminoethyl methacrylate), poly(hydroxyethyl acrylate), poly(hydroxyethyl methacrylate), poly(hydroxypropyl acrylate), poly(hydroxypropyl methacrylate), poly(vinyl alcohol), salts of poly(acrylic acid), salts of poly(methacrylic acid) and mixtures thereof.

[0042] Preferred sorbents are biocompatible. In another further embodiment, the polymer is biocompatible. In yet another embodiment, the polymer is hemocompatible. In still a further embodiment, the biocompatible polymer is hemocompatible. In still a further embodiment, the geometry of the polymer is a spherical bead.

[0043] In another embodiment, the biocompatible polymer comprises poly(N-vinylpyrrolidone).

[0044] The coating/dispersant on the poly(styrene-*co*-divinylbenzene) resin will imbue the material with improved biocompatibility.

[0045] In still yet another embodiment, a group of cross-linkers consisting of dipentaerythritol diacrylates, dipentaerythritol dimethacrylates, dipentaerythritol tetraacrylates, dipentaerythritol tetramethacrylates, dipentaerythritol triacrylates, dipentaerythritol trimethacrylates, divinylbenzene, divinylformamide, divinylanthracene, divinylsulfone, pentaerythritol diacrylates, pentaerythritol dimethacrylates, pentaerythritol tetraacrylates, pentaerythritol tetramethacrylates, pentaerythritol triacrylates, pentaerythritol trimethacrylates, trimethylolpropane diacrylate, trimethylolpropane dimethacrylate, trimethylolpropane triacrylate, trimethylolpropane trimethacrylate, trivinylbenzene, trivinylcyclohexane and mixtures thereof can be used in formation of a hemocompatible hydrogel coating.

[0046] In some embodiments, the polymer is a polymer comprising at least one crosslinking agent and at least one dispersing agent. The dispersing agent may be biocompatible. The dispersing agents can be selected from chemicals, compounds or materials such as hydroxyethyl cellulose, hydroxypropyl cellulose, poly(diethylaminoethyl acrylate), poly(diethylaminoethyl methacrylate), poly(dimethylaminoethyl acrylate), poly(dimethylaminoethyl methacrylate), poly(hydroxyethyl acrylate), poly(hydroxyethyl methacrylate), poly(hydroxypropyl acrylate), poly(hydroxypropyl methacrylate), poly(vinyl alcohol), salts of poly(acrylic acid), salts of poly(methacrylic acid) and mixtures thereof; the

crosslinking agent selected from a group consisting of dipentaerythritol diacrylates, dipentaerythritol dimethacrylates, dipentaerythritol tetraacrylates, dipentaerythritol tetramethacrylates, dipentaerythritol triacrylates, dipentaerythritol trimethacrylates, divinylbenzene, divinylformamide, divinylanthracene, divinylsulfone, pentaerythritol diacrylates, pentaerythritol dimethacrylates, pentaerythritol tetraacrylates, pentaerythritol tetramethacrylates, pentaerythritol triacrylates, pentaerythritol trimethacrylates, trimethylolpropane diacrylate, trimethylolpropane dimethacrylate, trimethylolpropane triacrylate, trimethylolpropane trimethacrylate, trivinylbenzene, trivinylcyclohexane and mixtures thereof. Preferably, the polymer is developed simultaneously with the formation of the coating, wherein the dispersing agent is chemically bound or entangled on the surface of the polymer.

[0047] In still another embodiment, the biocompatible polymer coating is selected from a group consisting of poly(diethylaminoethyl methacrylate), poly(dimethylaminoethyl methacrylate), poly(hydroxyethyl acrylate), poly(hydroxyethyl methacrylate), poly(hydroxypropyl acrylate), poly(hydroxypropyl methacrylate), poly(N-vinylpyrrolidone), poly(vinyl alcohol), salts of poly(acrylic acid), salts of poly(methacrylic acid) and mixtures thereof.

[0048] In still another embodiment, the biocompatible oligomer coating is selected from a group consisting of poly(diethylaminoethyl methacrylate), poly(dimethylaminoethyl methacrylate), poly(hydroxyethyl acrylate), poly(hydroxyethyl methacrylate), poly(hydroxypropyl acrylate), poly(hydroxypropyl methacrylate), poly(N-vinylpyrrolidone), poly(vinyl alcohol), salts of poly(acrylic acid), salts of poly(methacrylic acid) and mixtures thereof.

[0049] Some present biocompatible sorbent compositions are comprised of a plurality of pores. The biocompatible sorbents are designed to adsorb a broad range of toxins from less than 0.5 kDa to 1,000 kDa. While not intending to be bound by theory, it is believed the sorbent acts by sequestering molecules of a predetermined molecular weight within the pores. The size of a molecule that can be sorbed by the polymer will increase as the pore size of the polymer increases. Conversely, as the pore size is increased beyond the optimum pore size for adsorption of a given molecule, adsorption of said protein may or will decrease.

[0050] In certain methods, the solid form is porous. Some solid forms are characterized as having a pore structure having a total volume of pore sizes in the range of from 50Å to 40,000Å greater than 0.5 cc/g and less than 5.0 cc/g dry polymer.

[0051] In one embodiment, the sorbent has a pore structure wherein at least 1/3 of the pore volume in pores having diameters between 50Å and 40,000Å is in pores having diameters between 100Å and 1,000Å.

[0052] In another embodiment, the sorbent has a pore structure wherein at least 1/2 of the pore volume in pores having diameters between 50Å and 40,000Å is in pores having diameters between 1000Å and 10,000Å.

[0053] In still another embodiment, the sorbent has a pore structure wherein at least 1/3 of the pore volume in pores having diameters between 50Å and 40,000Å is in pores having diameters between 10,000Å and 40,000Å.

[0054] In certain embodiments, the polymers can be made in bead form having a diameter in the range of 0.1 micrometers to 2 centimeters. Certain polymers are in the form of powder, beads or other regular or irregularly shaped particulates.

[0055] In some embodiments, the plurality of solid forms comprises particles having a diameter in the range for 0.1 micrometers to 2 centimeters.

[0056] In some methods, the undesirable molecules include PAMPs and DAMPS comprised of one or more of flagellins, lipopeptides, formyl peptides, mycotoxins, exotoxins, cytolysins, superantigens, proteases, lipases, amylases, enzymes, peptides including bradykinin, activated complement, soluble receptors, soluble CD40 ligand, bioactive lipids, oxidized lipids, cell-free hemoglobin, cell-free myoglobin, growth factors, glycoproteins, prions, toxins, bacterial and viral toxins, drugs, vasoactive substances, foreign antigens, and antibodies.

[0057] In one embodiment, the polymers of this invention are made by suspension polymerization in a formulated aqueous phase with free radical initiation in the presence of aqueous phase dispersants that are selected to provide a biocompatible and a hemocompatible exterior surface to the formed polymer beads. In some embodiments, the beads are made porous by the macroreticular synthesis with an appropriately selected porogen (pore forming agent) and an appropriate time-temperature profile for the polymerization in order to develop the proper pore structure.

[0058] In another embodiment, polymers made by suspension polymerization can be made biocompatible and hemocompatible by further grafting of biocompatible and hemocompatible monomers or low molecular weight oligomers. It has been shown that the radical polymerization procedure does not consume all the vinyl groups of DVB introduced into copolymerization. On average, about 30% of DVB species fail to serve as crosslinking bridges and remain involved in the network by only one of two vinyl groups. The presence of a relatively high amount of pendant vinyl groups is therefore a characteristic feature of the adsorbents. It can be expected that these pendant vinyl groups are preferably exposed to the surface of the polymer beads and their macropores, if present, should be readily available to chemical modification. The chemical modification of the surface of DVB-copolymers relies on chemical reactions of the surface-exposed pendant vinyl groups and aims at converting these groups into more hydrophilic functional groups. This conversion via free radical grafting of monomers and/or cross-linkers or low molecular weight oligomers provides the initial hydrophobic adsorbing material with the property of hemocompatibility.

[0059] In yet another embodiment, the radical polymerization initiator is initially added to the dispersed organic phase, not the aqueous dispersion medium as is typical in suspension polymerization. During polymerization, many growing polymer chains with their chain-end radicals show up at the phase interface and can initiate the polymerization in the dispersion medium. Moreover, the radical initiator, like benzoyl peroxide, generates radicals relatively slowly. This initiator is only partially consumed during the formation of beads even after several hours of polymerization. This initiator easily moves toward the surface of the bead and activates the surface exposed pendant vinyl groups of the divinylbenzene moiety of the bead, thus initiating the graft polymerization of other monomers added after the reaction has proceeded for a period of time. Therefore, free-radical grafting can occur during the transformation of the monomer droplets into polymer beads thereby incorporating monomers and/or cross-linkers or low molecular weight oligomers that impart biocompatibility or hemocompatibility as a surface coating.

[0060] The hemoperfusion and perfusion devices consist of a packed bead bed of the polymer beads in a flow-through container fitted with either a retainer screen at both the exit end and the entrance end to maintain the bead bed inside the container, or with a subsequent retainer screen to collect the beads after mixing. The hemoperfusion and perfusion operations are

performed by passing the whole blood, blood plasma or physiologic fluid through the packed bead bed. During the perfusion through the bead bed, the toxic molecules are retained by sorption, torturous path, and/or pore capture, while the remainder of the fluid and intact cell components pass through essentially unchanged in concentration.

[0061] In some other embodiments, an in-line filter is comprised of a packed bead bed of the polymer beads in a flow-through container, fitted with a retainer screen at both the exit end and the entrance end to maintain the bead bed inside the container. Biological fluids are passed from a storage bag once-through the packed bead bed via gravity, during which the toxic molecules are retained by sorption, torturous path, and/or pore capture, while the remainder of the fluid and intact cell components pass through essentially unchanged in concentration.

[0062] Certain polymers useful in the invention (as is or after further modification) are macroporous polymers prepared from the polymerizable monomers of styrene, divinylbenzene, ethylvinylbenzene, and the acrylate and methacrylate monomers such as those listed below by manufacturer. Rohm and Haas Company, (now part of Dow Chemical Company): macroporous polymeric sorbents such as Amberlite™ XAD-1, Amberlite™ XAD-2, Amberlite™ XAD-4, Amberlite™ XAD-7, Amberlite™ XAD-7HP, Amberlite™ XAD-8, Amberlite™ XAD-16, Amberlite™ XAD-16 HP, Amberlite™ XAD-18, Amberlite™ XAD-200, Amberlite™ XAD-1180, Amberlite™ XAD-2000, Amberlite™ XAD-2005, Amberlite™ XAD-2010, Amberlite™ XAD-761, and Amberlite™ XE-305, and chromatographic grade sorbents such as Amberchrom™ CG 71,s,m,c, Amberchrom™ CG 161,s,m,c, Amberchrom™ CG 300,s,m,c, and Amberchrom™ CG 1000,s,m,c. Dow Chemical Company: Dowex™ Optipore™ L-493, Dowex™ Optipore™ V-493, Dowex™ Optipore™ V-502, Dowex™ Optipore™ L-285, Dowex™ Optipore™ L-323, and Dowex™ Optipore™ V-503. Lanxess (formerly Bayer and Sybron): Lewatit™ VPOC 1064 MD PH, Lewatit™ VPOC 1163, Lewatit™ OC EP 63, Lewatit™ S 6328A, Lewatit™ OC 1066, and Lewatit™ 60/150 MIBK. Mitsubishi Chemical Corporation: Diaion™ HP 10, Diaion™ HP 20, Diaion™ HP 21, Diaion™ HP 30, Diaion™ HP 40, Diaion™ HP 50, Diaion™ SP70, Diaion™ SP 205, Diaion™ SP 206, Diaion™ SP 207, Diaion™ SP 700, Diaion™ SP 800, Diaion™ SP 825, Diaion™ SP 850, Diaion™ SP 875, Diaion™ HP 1MG, Diaion™ HP 2MG, Diaion™ CHP 55A, Diaion™ CHP 55Y, Diaion™ CHP 20A, Diaion™ CHP 20Y, Diaion™ CHP 2MGY, Diaion™ CHP 20P, Diaion™ HP 20SS, Diaion™ SP 20SS, Diaion™ SP 207SS. Purolite Company: Purosorb™ AP 250 and Purosorb™

AP 400, and Kaneka Corp. Lixelle and CTR beads.and BioSKY™ MG Blood Perfusion Column and polymers within, BioSKY™ DX Bilirubin Perfusion Column and polymers within, Jafron Columns/Cartridges and polymers within such as BS330, DNA230, HA130, HA230, HA280, HA330, and HA330-II.

[0063] Various DAMPs and PAMPs may be adsorbed by the composition of the instant disclosure. Some of these proteins and their molecular weights are shown in the table below.

Protein	Molecular Weight (Da)	Protein	Molecular Weight (Da)
Gliotoxin	326	PCNA, proliferating cell nuclear antigen	29,000
T-2 toxin	466	Panton–Valentine leukocidin	34,000
PAF (Platelet Activating Factor)	524	Arginase I	35,000
Bilirubin	549	Carboxypeptidase A	35,000
Heme b	616	Thrombin	36,700
Lipopeptides	607-1,536	Flagellin H4	37,000
Formyl peptides	437-3,600	Flagellin H17	39,000
cathelicidin	4,500	Heat shock protein 40	40,000
β-Defensin	7,000	Strep pyrogenic exotoxin B	42,000
Ubiquitin	8000	alpha-1 antitrypsin	44,324
Complement C5a	8,200	Soluble CD14	49,000
Complement C3a	9,089	Rabies nucleocapsid N protein	50,600
Heat shock protein 10	10,000	Soluble TNF-α receptor	55,000
S100B (monomer)	10,000		
Histone H4	11,000	Activated Protein C	56,200
Procalcitonin	13,000	Amylase	57,000
Serum amyloid A	13,000	Hemopexin	57,000
Phospholipase A2, secretory PLA2	14,000	Lysteriolysin	58,000
PLA2G2A	16,083	Heat shock protein 60	60,000
S100 proteins A8 (dimer)	20,000	Streptolysin O	60,500
Trypsin	23,300	Diphtheria toxoid	62,000
Staph Enterotoxin B	24,500	Hemoglobin, oxy	64,000
HMGB1	24,894	Pseudomonas Exotoxin A	66,000
Chymotrypsin	25,000	Flagellin H9	69,000
Elastase (neutrophil)	25,000	Heat shock protein 70	70,000
Staph ETA	26,900	Calpain-1 (human erythrocytes)	112,00
HDGF	27,000	C reactive Protein (5x25 kDa)	115,000
PF4	27,100	Myeloperoxidase (neutrophils)	150,000
Staph ETB	27,300	NOS synthase	150,000

[0064] The following examples are intended to be exemplary and non-limiting.

Example 1: Base Sorbent Synthesis CY14175 & CY15077

[0061] Reactor Setup: a 4-neck glass lid was affixed to a 3L jacketed cylindrical glass reaction vessel using a stainless steel flange clamp and PTFE gasket. The lid was fitted with a PTFE stirrer bearing, RTD adapter, and water-cooled reflux condenser. A stainless steel stirring shaft having five 60° agitators was fit through the stirrer bearing and inserted into a digital overhead stirrer. An RTD was fit through the corresponding adapter, and connected to a PolyStat circulating heating and chilling unit. Compatible tubing was used to connect the inlet and outlet of the reaction vessel jacket to the appropriate ports on the PolyStat. The unused port in the lid was used for charging the reactor and was plugged at all other times.

[0062] Polymerization: Aqueous phase and organic phase compositions are shown below, in Table I and Table II, respectively. Ultrapure water was split into approximately equal parts in two separate Erlenmeyer flasks, each containing a PTFE coated magnetic stir bar. Poly(vinyl alcohol) (PVA), having a degree of hydrolysis of 85.0 to 89.0 mol percent and a viscosity of 23.0 to 27.0cP in a 4% aqueous solution at 20°C, was dispersed into the water in the first flask and heated to 80°C on a hot plate with agitation. Salts (see Table 1, MSP, DSP, TSP and Sodium nitrite) were dispersed into the water in the second flask and heated to 80°C on a hot plate with agitation. Circulation of heat transfer fluid from the PolyStat through the reaction vessel jacket was started, and fluid temperature heated to 60°C. Once PVA and salts dissolved, both solutions were charged to the reactor, one at a time, using a glass funnel. The digital overhead stirrer was powered on and the rpm set to a value to form appropriate droplet sizes upon organic phase addition. Temperature of the aqueous phase in the kettle was set to 70°C. The organic phase was prepared by adding benzoyl peroxide (BPO) to the divinylbenzene (DVB) in a 2L Erlenmeyer flask and swirling until completely dissolved. 2,2,4-trimethylpentane and toluene were added to the flask, which was swirled to mix well. Once the temperature of the aqueous phase in the reactor reached 70°C, the organic phase was charged into the reactor using a narrow-necked glass funnel. Temperature of the reaction volume dropped upon the organic addition. A temperature program for the PolyStat was started, heating the reaction volume from 60 to 77°C over 30 minutes, 77 to 80°C over 30 minutes, holding the temperature at 80°C for 960 minutes, and cooling to 20°C over 60 minutes.

Table I: Aqueous Phase Composition

<u>Reagent</u>	<u>Mass (g)</u>
Ultrapure water	1500.000
Poly(vinyl alcohol) (PVA)	4.448
Monosodium phosphate (MSP)	4.602
Disodium phosphate (DSP)	15.339
Trisodium phosphate (TSP)	9.510
Sodium nitrite	0.046
Total	1533.899

Table II: Organic Phase Compositions

	CY14175	CY15077
<u>Reagent</u>	<u>Mass (g)</u>	<u>Mass (g)</u>
Divinylbenzene, 63% (DVB)	508.751	498.383
2,2,4-trimethylpentane (Isooctane)	384.815	482.745
Toluene	335.004	222.404
Benzoyl peroxide, 98% (BPO)	3.816	3.738
Total (excluding BPO)	1228.571	1203.532

[0063] Work-up: reaction volume level in the reactor was marked. Overhead stirrer agitation was stopped, residual liquid siphoned out of the reactor, and the reactor filled to the mark with ultrapure water at room temperature. Overhead stirrer agitation was restarted and the slurry heated to 70°C as quickly as possible. After 30 minutes, agitation was stopped and residual liquid siphoned out. Polymer beads were washed five times in this manner. During the final wash, the slurry temperature was cooled to room temperature. After the final water wash, polymer beads were washed with 99% isopropyl alcohol (IPA) in the same manner. 99% IPA was siphoned out and replaced with 70% IPA before transferring the slurry into a clean 4L glass container. Unless noted otherwise, on an as-needed basis the polymer was steam stripped in a stainless steel tube for 8 hours, rewet in 70% IPA, transferred into DI water, sieved to obtain only the portion of beads having diameters between 300 and 600 μ m, and dried at 100°C until no further weight loss on drying was observed.

[0064] Cumulative pore volume data for polymers CY14175 and CY15077, measured by nitrogen desorption isotherm and mercury intrusion porosimetry, respectively, are shown below in Tables III and IV, respectively.

Table III: Nitrogen Desorption Isotherm Data for CY14175

<u>Pore Diameter</u> <u>Range (Å)</u>	<u>Pore size</u> <u>Diameter (Å)</u>	<u>Cumulative Pore</u> <u>Volume (cm³/g)</u>	<u>Pore Diameter</u> <u>Range (Å)</u>	<u>Pore size</u> <u>Diameter (Å)</u>	<u>Cumulative Pore</u> <u>Volume (cm³/g)</u>
1411.9 - 1126.5	1236.809577	0.018062878	208.6 - 130.5	151.2300725	1.464027373
1126.5 - 981.7	1043.979923	0.038442381	130.5 - 105.7	115.2567614	1.527062065
981.7 - 752.9	836.7828769	0.141559621	105.7 - 82.8	91.14860242	1.592486039
752.9 - 659.9	700.1024343	0.24336622	82.8 - 67.6	73.42901881	1.641003444
659.9 - 572.0	609.4657394	0.416511969	67.6 - 57.5	61.59836256	1.6763711
572.0 - 483.1	519.8089977	0.646318614	57.5 - 51.6	54.15491457	1.699539142
483.1 - 449.8	465.2234212	0.730406771	51.6 - 45.0	47.72291376	1.728282889
449.8 - 401.4	422.7246485	0.849167577	45.0 - 39.8	42.01726183	1.752728216
401.4 - 354.1	374.6289335	0.956165766	39.8 - 35.8	37.55877213	1.779016164
354.1 - 337.9	345.6019761	0.997336398	35.8 - 31.8	33.51596841	1.8086605
337.9 - 313.5	324.758962	1.0547802	31.8 - 28.7	30.02327371	1.82963357
313.5 - 290.8	301.2432086	1.09667858	28.7 - 26.0	27.18773181	1.850084632
290.8 - 262.8	275.299967	1.164042391	26.0 - 23.3	24.46989555	1.87529426
262.8 - 247.2	254.510376	1.199751164	23.3 - 20.9	21.92055755	1.902736527
247.2 - 233.6	240.0176376	1.228796957	20.9 - 18.5	19.52461159	1.935789448
233.6 - 220.1	226.435352	1.256631669	18.5 - 16.2	17.16324429	1.97779901
220.1 - 208.6	213.9982044	1.283063762			

Table IV: Mercury Intrusion Data for CY15077

<u>Pore size</u> <u>Diameter (Å)</u>	<u>Cumulative</u> <u>Intrusion (mL/g)</u>	<u>Pore size</u> <u>Diameter (Å)</u>	<u>Cumulative</u> <u>Intrusion (mL/g)</u>	<u>Pore size</u> <u>Diameter (Å)</u>	<u>Cumulative</u> <u>Intrusion (mL/g)</u>
226299.0625	3.40136E-30	672.187561	1.581117511	111.9475937	2.162935257
213166.0781	0.001678752	636.7885742	1.60271585	108.8830032	2.167646885
201295.1563	0.002518128	604.7248535	1.621845484	106.6480179	2.174062729
172635.8125	0.004364755	558.1287231	1.651492	104.5217743	2.179908991
139538.0625	0.007554384	518.2624512	1.678913713	102.4295197	2.179908991
113120.7813	0.011919139	483.5536499	1.708594561	100.1580353	2.182951927
90542.36719	0.01645177	453.5110779	1.735918999	98.29322052	2.184018135
78733.25781	0.0203129	426.9998474	1.755934	96.44822693	2.191127539
72446.375	0.022327403	403.1251526	1.783603072	94.42159271	2.198545218
60340.40234	0.027867284	382.7776794	1.793849826	91.52587891	2.209161043
48343.83984	0.035327822	362.7162476	1.817784309	89.25807953	2.209312439
39009.13672	0.040918175	342.3734436	1.838774562	87.0777359	2.215425491
32136.4082	0.04899035	330.1105042	1.851493955	85.42358398	2.221472025
25330.65625	0.063195683	315.5238037	1.869742155	83.62612915	2.232139587
20981.51563	0.079529688	302.2973938	1.885128617	82.11174011	2.237514496
16219.86426	0.108860672	290.2946777	1.895119786	79.91614532	2.239231586
13252.41211	0.141730919	279.1246643	1.912378907	78.01462555	2.239560127
10501.53613	0.193969816	268.7442627	1.924305081	76.19993591	2.239560127
8359.911133	0.262399256	259.1106873	1.936048627	75.09249115	2.239560127

6786.30127	0.345866203	241.8737793	1.955100656	73.41201019	2.239560127
5538.122559	0.438174427	226.7678223	1.972970247	72.23709869	2.240245819
4337.931152	0.563276172	213.3626251	1.988123298	71.09960175	2.242422104
3501.674805	0.681870878	201.4908142	2.007521152	69.86301422	2.243849993
2838.742188	0.804727197	194.9888611	2.022114754	68.40761566	2.257676363
2593.016846	0.865813017	188.9506989	2.033871174	67.13697815	2.259181261
2266.688965	0.938610673	180.582901	2.035052776	66.03359222	2.266284466
1831.041748	1.056586146	172.8530121	2.050720692	65.08189392	2.270181179
1509.850708	1.163395643	164.9621735	2.062945843	64.04368591	2.272682428
1394.006104	1.21002543	157.8110657	2.071056128	62.38490295	2.280714512
1294.780151	1.257248282	151.1540375	2.082133055	61.32764053	2.280714512
1207.692627	1.293158531	143.9185333	2.096480608	60.30379868	2.287917852
1131.860962	1.326992273	138.4670563	2.106938839	59.41370392	2.287917852
1065.099976	1.35812819	132.8492737	2.119287968	58.54679489	2.293802738
953.1816406	1.405935764	129.5760345	2.126605988	57.79866409	2.297607183
884.0358887	1.445426106	126.5438614	2.126605988	56.88977814	2.299046278
823.5491333	1.478719592	124.2635574	2.132267475	55.9213295	2.302111387
770.9108276	1.510579824	120.8976135	2.141504765	54.98665237	2.303381443
722.4724731	1.537048101	117.3792267	2.150759459		
684.6119995	1.564400196	114.791893	2.154810667		

Example 2: Polymer Modification CY15065

[0065] 250mL base polymer CY14175, wetted in DI water, was added to a 500mL jacketed glass reactor which was equipped with a Teflon coated agitator and RTD probe. 90mL excess DI water was added to the reactor, and the slurry mixed at 90 RPM. Reaction temperature was set to 20°C. Three separate additions were prepared; 1.4g ammonium persulfate in 14mL DI water, 0.7g N-vinylpyrrolidinone in 21mL DI water, and 1.5g N,N,N,N-tetramethylethylenediamine in 7mL DI water. Reaction temperature setpoint was increased to 40°C and monitored closely. Ammonium persulfate solution was added once reaction temperature reached 30°C. N,N,N,N-tetramethylethylenediamine solution was added once reaction temperature reached 35°C. N-vinylpyrrolidinone solution was added once reaction temperature reached 39°C. Reaction was then maintained at 40°C for 2 hours, before decreasing temperature to 25°C.

[0066] Work-up: reaction volume level in the reactor was marked. Overhead stirrer agitation was stopped, residual liquid siphoned out of the reactor, and the reactor filled to the mark with ultrapure water at room temperature. Overhead stirrer agitation was restarted. After 30 minutes, agitation was stopped and residual liquid siphoned out. Polymer beads were washed

three times in this manner. The polymer was steam stripped in a stainless steel tube for 8 hours, rewet in 70% IPA, then transferred into DI water.

[0067] Cumulative pore volume data for polymers CY15065, measured by nitrogen desorption isotherm, is shown below in Table V.

Table V: Nitrogen Desorption Isotherm Data for CY15065					
<u>Pore Diameter</u> <u>Range (Å)</u>	<u>Pore size</u> <u>Diameter (Å)</u>	<u>Cumulative Pore</u> <u>Volume (cm³/g)</u>	<u>Pore Diameter</u> <u>Range (Å)</u>	<u>Pore size</u> <u>Diameter (Å)</u>	<u>Cumulative Pore</u> <u>Volume (cm³/g)</u>
3735.2 - 1226.3	1462.700326	0.005546248	224.9 - 209.7	216.7455697	1.046516069
1226.3 - 724.9	851.9599269	0.012549654	209.7 - 141.7	162.0655488	1.225860688
724.9 - 656.2	686.9797148	0.01499268	141.7 - 103.9	116.5968304	1.329424083
656.2 - 595.3	622.6438437	0.020898533	103.9 - 82.9	90.79492123	1.388864147
595.3 - 566.7	580.2249215	0.033282221	82.9 - 69.3	74.73718587	1.431186463
566.7 - 511.2	535.9524733	0.111667343	69.3 - 59.3	63.39311832	1.465462484
511.2 - 459.4	482.4242892	0.244617735	59.3 - 50.9	54.36935206	1.495795233
459.4 - 410.9	432.3534717	0.366033907	50.9 - 44.8	47.39226606	1.521105084
410.9 - 376.5	392.140848	0.468167046	44.8 - 39.5	41.71847385	1.544891234
376.5 - 334.5	352.8905481	0.63111569	39.5 - 35.3	37.12325172	1.568519104
334.5 - 298.7	314.4631626	0.753980937	35.3 - 31.4	33.1071979	1.594220982
298.7 - 291.1	294.7961544	0.786627632	31.4 - 28.2	29.60557504	1.614149255
291.1 - 273.8	281.8886994	0.844237668	28.2 - 25.6	26.73665706	1.633426358
273.8 - 256.6	264.6128479	0.91291745	25.6 - 22.8	24.01512381	1.656221583
256.6 - 228.9	241.1063341	0.990966112	22.8 - 20.5	21.49286805	1.680044553
228.9 - 224.9	226.9020278	1.007532081	20.5 - 18.2	19.15056378	1.707534945

Example 3: Removal from Whole Bovine Blood in a Recirculation Model

[0068] Purified proteins were added to 300 mL 3.8% citrated whole bovine blood (Lampire Biologicals) at expected clinical concentrations and recirculated through a 20 mL polymer-filled device or control (no bead) device at a flow rate of 140 mL/min for five hours. Proteins and initial concentrations were: S100A8 at 50 ng/mL, complement C5a at 25 ng/mL, procalcitonin at 16 ng/mL, HMGB-1 at 100 ng/mL, and SPE B at 100 ng/mL. Plasma was analyzed by enzyme-linked immunosorbent assay (ELISA) following manufacturer instructions (S100, and C5a, duosets (R&D Systems); procalcitonin (Sigma), HMGB-1 (Chondrex ELISA); and toxins (Toxin Technologies). Removal data from experiments using polymer CY15065 are shown below in Figure 1 and Figure 2 for DAMPs and PAMPs, respectively. For brevity, CY15065 is denoted 'CS' in the figure legends. Removal data from experiments using polymer CY15077 are shown below in Figure 3 and Figure 4 for DAMPs and PAMPs, respectively.

What is claimed:

1. A biocompatible polymer system comprising at least one polymer, said polymer system capable of adsorbing (i) pathogen-associated molecular pattern molecules (PAMPS) and (ii) damage-associated molecular pattern molecules (DAMPS) having a molecular weight of from less than about 0.5 kDa to about 1,000 kDa.
2. The biocompatible polymer system of claim 1 wherein the polymer comprises a plurality of pores and the polymer's pore structure has a total volume of pore sizes in the range of from 50Å to 40,000Å greater than 0.5 cc/g and less than 5.0 cc/g dry polymer.
3. The biocompatible polymer system of claim 1 wherein the polymer is hemocompatible.
4. The biocompatible polymer system of claim 1 wherein the geometry of said polymer system is a spherical bead.
5. The biocompatible polymer system of claim 1 wherein the toxins comprise PAMPs and DAMPS comprised of one or more of flagellins, lipopeptides, formyl peptides, mycotoxins, exotoxins, endotoxins, lipoteichoic acid, cytolysins, superantigens, proteases, lipases, amylases, enzymes, peptides including bradykinin, activated complement, soluble receptors, soluble CD40 ligand, bioactive lipids, oxidized lipids, cellular DNA, mitochondrial DNA, pathogen or host derived RNA, cell-free hemoglobin, cell-free myoglobin, growth factors, peptidoglycans, glycoproteins, released intracellular components, cell wall or viral envelope components, Polyinosinic:polycytidylic acid (poly I:C), prions, toxins, bacterial and viral toxins, drugs, vasoactive substances, and foreign antigens.
6. The biocompatible polymer system of claim 1 wherein said polymer is made using suspension polymerization.
7. The biocompatible polymer system of claim 1 wherein said polymer is a hypercrosslinked polymer.
8. The biocompatible polymer system of claim 4 wherein the spherical bead has a biocompatible hydrogel coating.
9. The biocompatible polymer system of claim 1 wherein the polymer is formed and subsequently modified to be biocompatible.

10. A method of perfusion comprising passing a physiologic fluid once through or by way of a suitable extracorporeal circuit through a device once or many times comprising the biocompatible polymer system of any one of claims 1-9.
11. A device for removing (i) pathogen-associated molecular pattern molecules and/or (ii) damage-associated molecular pattern molecules from less than 0.5 kDa to 1,000 kDa from physiologic fluid comprising the biocompatible polymer system of any one of claims 1-9.
12. The polymer in claim 1 housed in a container suitable to retain the polymer and for transfusion of whole blood, packed red blood cells, platelets, albumin, plasma or any combination thereof.
13. The polymer in claim 1 is in a device suitable to retain the polymer and be incorporated into an extracorporeal circuit.
14. The polymer in claim 1 where free polymer (i.e. not contained) is used to treat physiologic fluids.

1/4

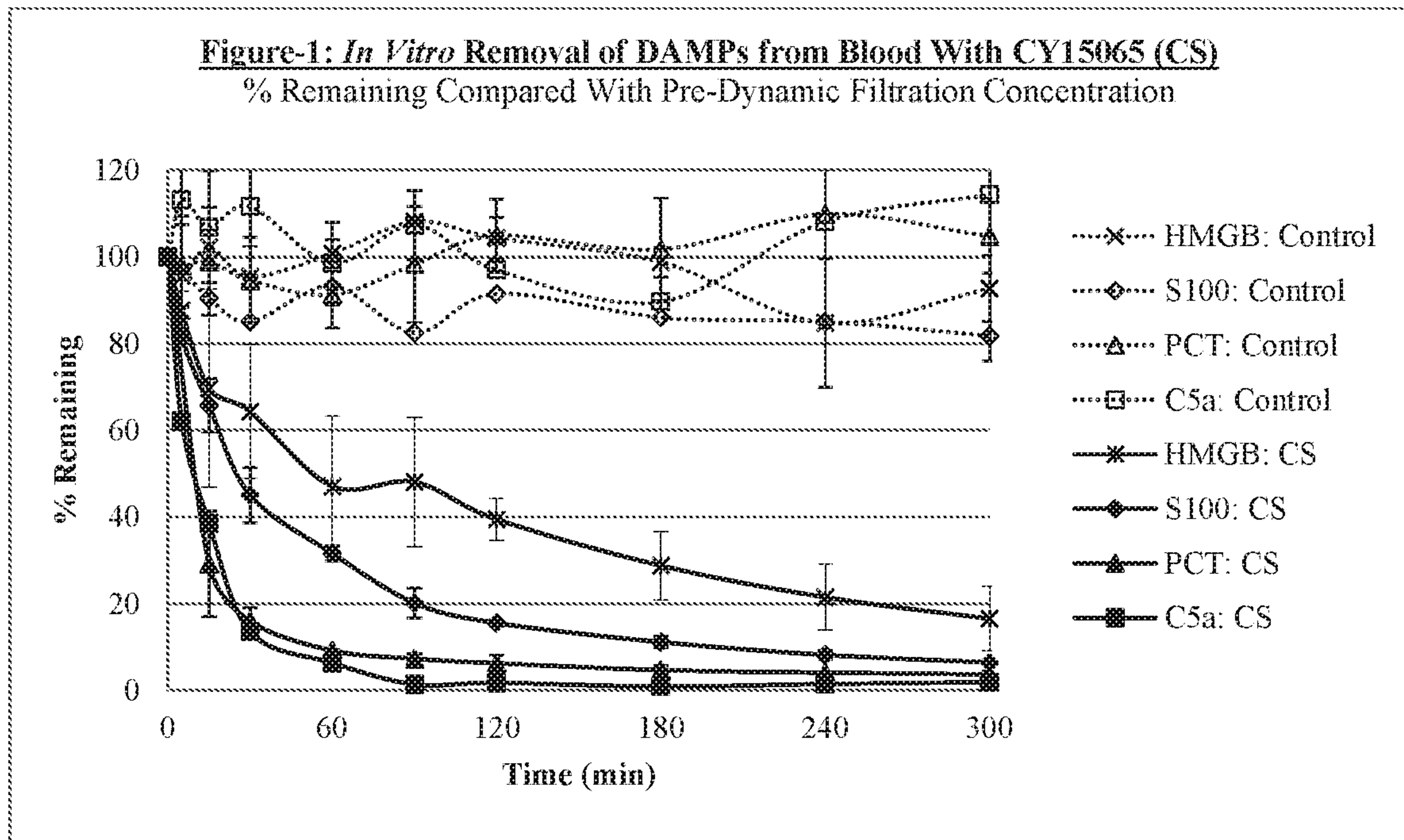


FIGURE 1

2/4

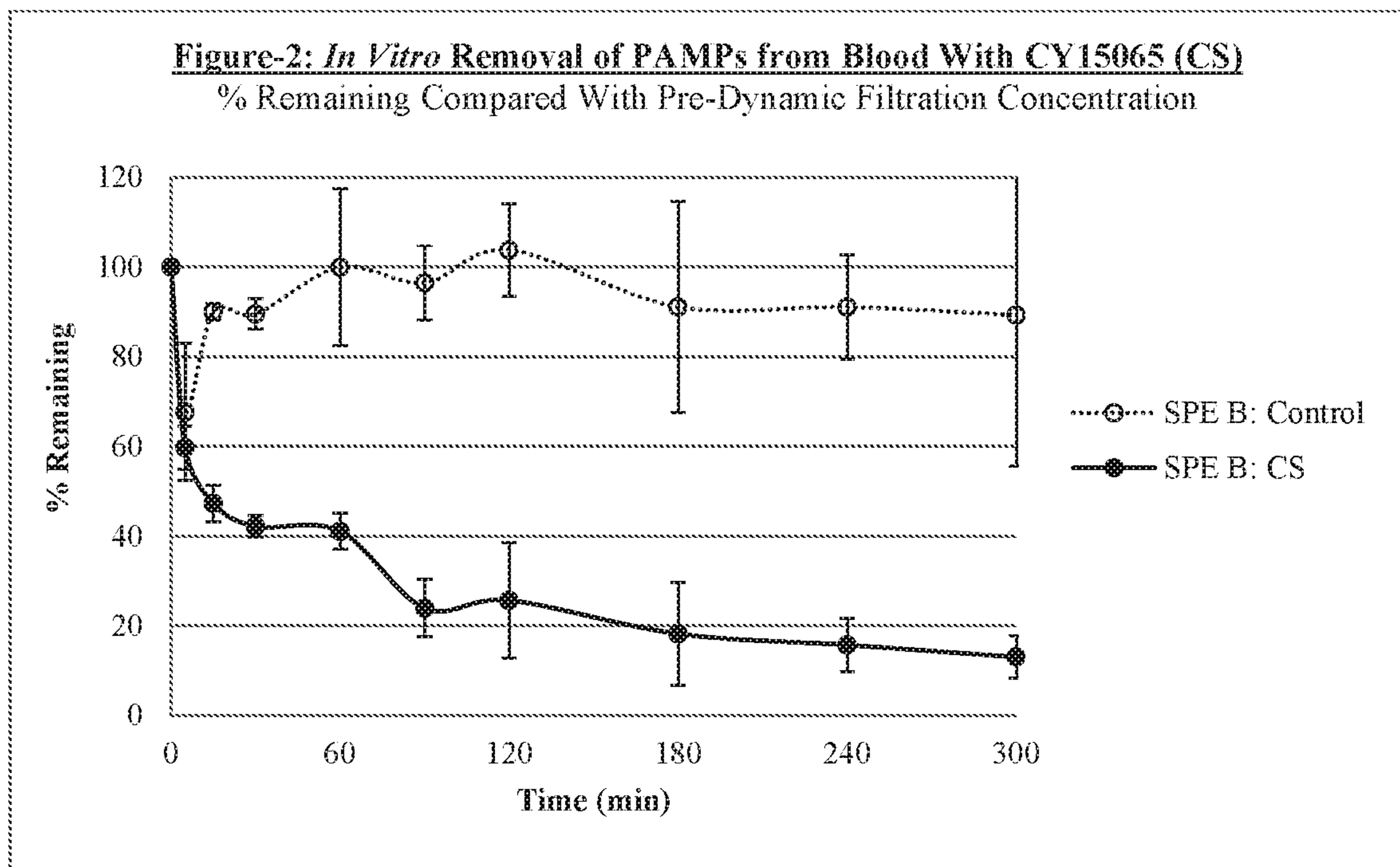


FIGURE 2

3/4

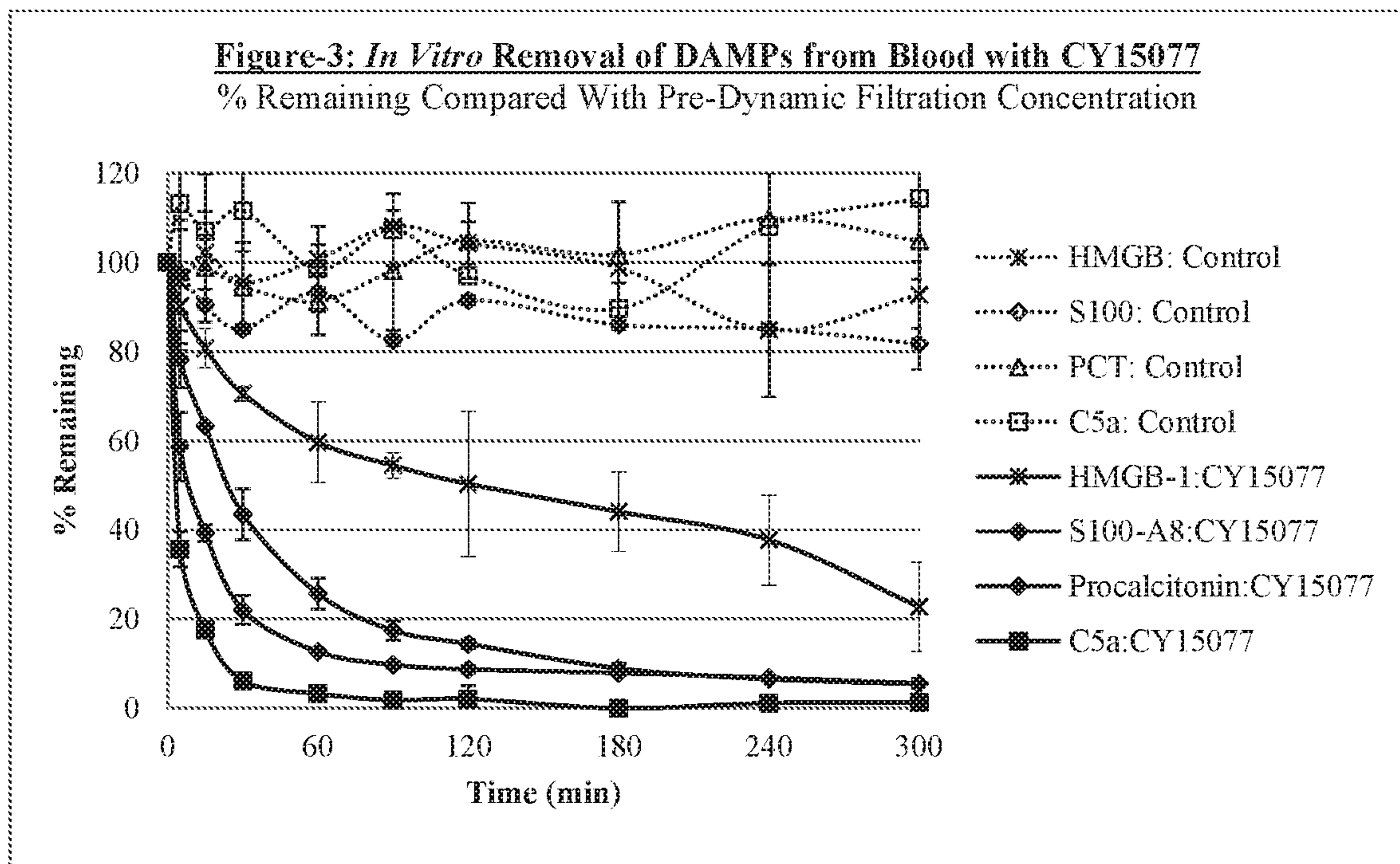


FIGURE 3

4/4

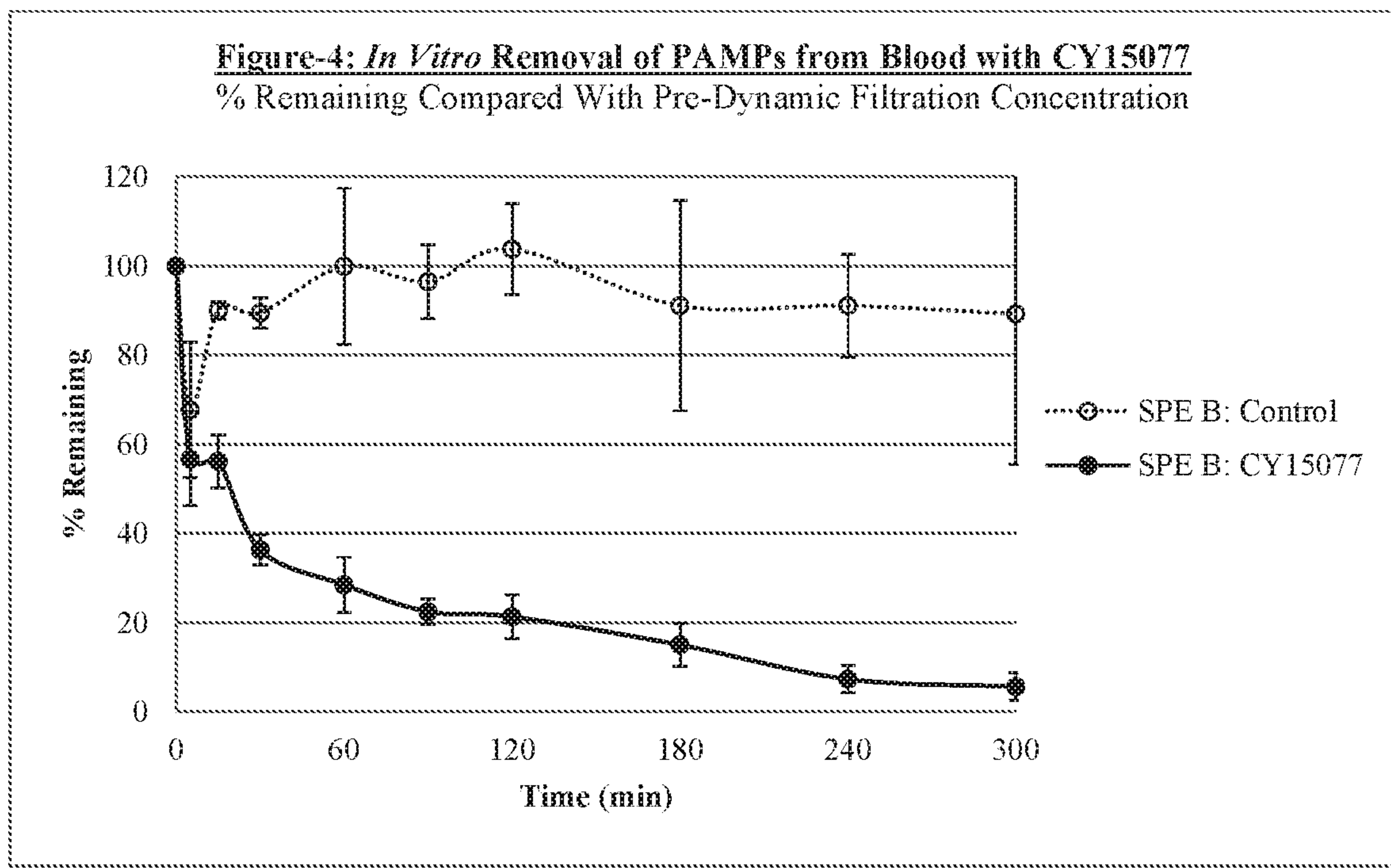


FIGURE 4

Figure-1: *In Vitro* Removal of DAMPs from Blood With CY15065 (CS)

% Remaining Compared With Pre-Dynamic Filtration Concentration

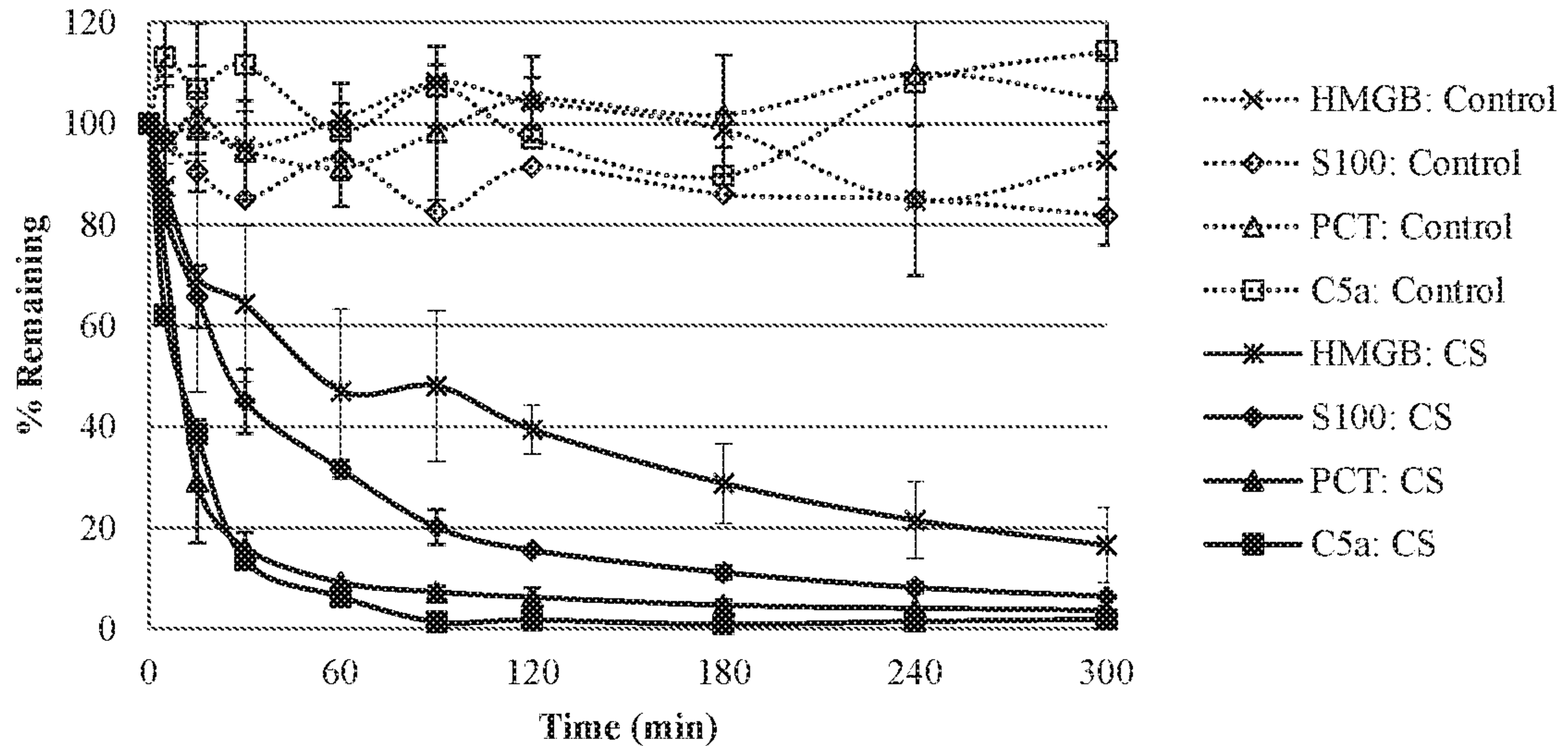


FIGURE 1