

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau



(43) International Publication Date
12 November 2009 (12.11.2009)

(10) International Publication Number
WO 2009/135861 A2

(51) International Patent Classification:
C07K 16/24 (2006.01) C07K 16/46 (2006.01)

(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BR, BW, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IS, JP, KE, KG, KM, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LT, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PG, PH, PL, PT, RO, RS, RU, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(21) International Application Number:
PCT/EP2009/055448

(22) International Filing Date:
6 May 2009 (06.05.2009)

(25) Filing Language:
English

(26) Publication Language:
English

(30) Priority Data:
08103847.3 7 May 2008 (07.05.2008) EP

(71) Applicant (for all designated States except US): **NOVO NORDISK A/S** [DK/DK]; Novo Allé, DK-2880 Bagsværd (DK).

(72) Inventors; and

(75) Inventors/Applicants (for US only): **SVENSSON, Lars Anders** [SE/SE]; Roskildevägen 17B, S-217 46 Malmö (SE). **PADKJÆR, Søren** [DK/DK]; Rosen Allé 2, Hareskov, DK-3500 Værløse (DK). **FRIEDRICHSEN, Birgitte** [DK/DK]; Agertoften 12, DK-2800 Gentofte (DK). **OLSEN KROGH, Berit** [DK/DK]; Vamdrupvej 57, DK-2610 Rødovre (DK). **LUND PEDERSEN, Inger** [DK/DK]; Vangsåvej 10, DK-2720 Vanløse (DK).

(74) Common Representative: **NOVO NORDISK A/S**; Novo Allé, DK-2880 Bagsværd (DK).

(84) Designated States (unless otherwise indicated, for every kind of regional protection available): ARIPO (BW, GH, GM, KE, LS, MW, MZ, NA, SD, SL, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European (AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, SE, SI, SK, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, ML, MR, NE, SN, TD, TG).

Published:

- without international search report and to be republished upon receipt of that report (Rule 48.2(g))
- with sequence listing part of description (Rule 5.2(a))



WO 2009/135861 A2

(54) Title: HUMANIZED ANTIBODIES AGAINST HUMAN INTERFERON-ALPHA

(57) Abstract: The present invention provides humanized anti-human IFN- α monoclonal antibodies useful for therapeutic applications in humans. Preferred antibodies are humanized versions of murine antibodies ACO-1 and ACO-2, as well as variants thereof.

HUMANIZED ANTIBODIES AGAINST HUMAN INTERFERON-ALPHA**FIELD OF THE INVENTION**

The present invention relates to humanized antibodies against human interferon al-
5 pha (IFN- α) and their use in treating or preventing various diseases and disorders in human patients.

BACKGROUND OF THE INVENTION

Based on a variety of different observations, interferon alpha (IFN- α) is a cytokine believed to be involved in a number of autoimmune diseases. Although systemic lupus 10 erythematosus (SLE) patients often do not have measurable serum levels of IFN- α , they appear to have a clear IFN- α gene signature. In addition, induction of dendritic cell (DC) maturation by treatment of DCs with SLE patient serum can be inhibited by an anti-IFN- α antibody. It has also been shown that knockout of the IFN- α / β receptor in New Zealand Black 15 (NZB) mice having an SLE phenotype results in a near normal phenotype (Santiago-Raber et al., J Exp Med. 2003;197(6):777-88).

Antibodies against IFN- α have therefore been suggested as tools to neutralize the activity of this cytokine for the treatment of such autoimmune diseases, alone or in combination. Specific murine antibodies (ACO-1 to ACO-6) that recognize a wide range of different 20 IFN- α subtypes were generated and characterized as described in the international patent application published as WO20060086586. However, murine antibodies are not suitable for use in humans because of their immunogenicity, and it is therefore desirable to generate humanized antibodies where the murine CDRs are grafted onto a human scaffold antibody. However humanized antibodies often suffer from functional deficiencies as compared to the murine parent, such as, e.g., a lower affinity and/or stability and/or undesirable immunogenic- 25 ity. Such deficiencies in humanized antibodies can in some cases be compensated for by making one or a few back point mutations. It is usually desirable to perform no or only a very few back point mutations since the presence of too many back mutations tend to result in undesirable low stability and/or an undesirable degree of immunogenicity. The provision of a safe and stable humanized anti-IFN- α antibody having desirable biological properties such as 30 e.g. retaining affinity and potency of the humanized anti-IFN- α antibody to a large number of IFN- α subtypes is thus desirable.

There is thus a need in the art for humanized anti-IFN- α antibodies having desirable features with respect to features such e.g. stability, specificity, safety, immunogenicity, etc. Furthermore, there is a need in the art for efficient methods for producing such antibodies.

SUMMARY OF THE INVENTION

5

In a first aspect, the present invention relates to a humanized antibody that specifically binds human interferon- α (IFN- α), or an antigen-binding fragment thereof, which humanized antibody is a humanized version of murine antibody ACO-1 or ACO-2, or of a combination thereof, comprising fewer donor amino acid residues than the murine complementary determining regions (CDRs) according to Kabat.

10

In another aspect, the present invention furthermore relates to a humanized antibody that specifically binds IFN- α , or an antigen-binding fragment thereof, wherein said antibody is capable of binding IFN- α subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b and WA, but not subtypes 1 or D, and wherein said antibody comprises fewer donor amino acid residues than the 15 non-human CDRs according to Kabat.

15

The present invention furthermore relates to methods for obtaining such antibodies as well as use of such antibodies for therapeutic purposes and compositions comprising such antibodies.

20

The antibodies according to the present invention can be suitable for treatment of various inflammatory diseases.

DESCRIPTION OF THE DRAWINGS

Figure 1 shows analysis of murine ACO-1 VH (A) and VL (B) sequences for humanization (hz=humanized), where the mask is shown in shaded text, Kabat CDRs are shown in bold, differences between the mouse sequence and the human germline sequence are shown in underlined text, potential somatic hypermutated residues are shown in bold underlined text, and potential back-mutation residues in shaded underlined text. ACO-1 VH = SEQ ID NO:1; human germline VH1_46/JH4 = SEQ ID NO:2; hzACO-1 VH= SEQ ID NO:3; ACO-1 VL = SEQ ID NO:4; human germline VKIII_L6/JK2 = SEQ ID NO:5; hzACO-1 VL = SEQ ID NO:6.

25

Figure 2 shows an alignment between ACO-1 and ACO-2 VH (A) and VL (B) sequences, as well as the corresponding mouse germline sequences. ACO-2 VH = SEQ ID

NO:7;mouse germline J558.33/D/_JH3_1 = SEQ ID NO:8; ACO-2 VL = SEQ ID NO:9; mouse germline ae4/ JK4_1 = SEQ ID NO:10.

5 **Figure 3** shows the location of ACO-2 residues selected for introduction into hzACO-1.

Figure 4 shows hzACO-1 inhibition of the protective effect of all interferon subtypes tested except for IFN- α D and IFN- α 1 in a CPE assay.

10 **Figure 5** shows hzACO-1 inhibition of 12 IFN- α species by a reporter gene (RG) assay. Values for each antibody concentration were normalized and the average of four repetitions were calculated. Data are shown as average +/- standard errors. Best fit sigmoidal response curves were calculated using Prism software. R2 values were for all dataset above 0.98 (except for IFN- α D for which no curve fitting was made).

15 **Figure 6** shows an RG assay comparison of hzACO-1 with a hzACO-1 variants having the single ACO-2-derived mutation A93V, using IFN- α A (A) or IFN- α F (B). Data calculations were made as described for Figure 5.

Figure 7 shows transition temperatures for hzACO-1 and variants at pH 3.5 (A), 4.5 (B), and 5.5 (C), without additives.

20 **Figure 8** shows the structure of hzACO-1 Fab fragment chains (H, L) bound to IFN- α 8, determined by X-ray crystallography.

25 **Figure 9** shows the IFN- α 8 binding epitopes for IFNAR1 and IFNAR2 (Quadt-Akabayov S.R. et al. Protein Sci. 15, 2656-2668, 2006 and Roisman L.C et al. J. Mol. Biol. 353, 271-281, 2005), as indicated by colored boxes below the IFN- α 8 sequence. Residues indicated by boxes colored gray are partly conserved among all IFN- α subtypes while residues indicated by black colored boxes are fully conserved. The hzACO-1 binding epitope, using a 4 \AA distance cut-off, on IFN- α 8 is indicated by "*" above the amino acid sequence of IFN- α 8.

30 **Figure 10** shows comparison of the mouse ACO-1 mAb to hzACO-1 as well as two variants hereof in the RG assay. One variant is a humanized ACO-1 harboring the entire CDRH2 (designated hzACO-1-kabat CDRH2) whereas the hzACO-1 was constructed with a shorter CDRH2 as described in example 2. in addition the figure shows another mutated hzACO-1 which has been optimized for interaction with IFN- α s (hzACO-1 Y32E, T30R) through rational design. These four recombinant mAb variants were compared with respect to inhibition of five different representative IFN- α subtypes, as indicated.

35 **Figure 11** shows a protein stability study of hzACO-1 expressed with human IgG1, IgG2 and IgG4 isotypes. Aggregation was determined by HPLC after incubation in histidine buffer for 5 weeks.

Figure 12 shows a ^{51}Cr release assay illustrating lack of ADCC by hzACO-1 IgG4, IFN- α and different combinations hereof, at different effector:target cell ratios (E:T). Cells + PBMCs alone without IFN- α pr hzACO-1 determines background lysis and Triton-X 100 illustrates maximal lysis. Rituxan was included as a positive control and induces detectable cell lysis at all E:T ratios.

Figure 13 shows a complement binding study by Elisa. The hzACO-1 expressed as an IgG4 was unable to fix complement when bound to IFN- α . As a positive control the hzACO-1 was cross bound with an anti-IgG4 pAb, and a clear dose dependent binding of C4 to the anti-IgG4 was detected.

DESCRIPTION OF THE INVENTION

The present invention is based, in part, on anti-IFN- α antibodies with properties suitable for treating human patients suffering from an IFN- α -related condition or disease, such as, e.g., a lupus disease or disorder such as, e.g., SLE; graft versus host disease; type 15 1 diabetes, AIDS, autoimmune thyroiditis, psoriasis, juvenile dermatomyositis, and Sjögren's syndrome. The antibodies are typically based on humanized versions of the murine ACO-1 and/or ACO-2 antibodies.

ACO-1 and ACO-2 were identified as capable of blocking the bioactivity of thirteen recombinant IFN- α subtypes as well as two complex mixtures of IFN- produced upon viral 20 infection (see WO2006086586). ACO-1 and ACO-2 also consistently blocked the bioactivity of serum from SLE patients that exhibited IFN- α signatures by microarray analysis. ACO-1 and ACO-2 did not significantly neutralize the bioactivity of IFN- α protein subtypes D and 1, but did neutralize the IFN- α bioactivity of SLE serum. Though not limited to theory, it is therefore possible that subtypes D and 1 are not significantly involved in the etiology of SLE.

25 As described in the Examples, structural modelling of the variable regions revealed that it was possible to humanize ACO-1 and ACO-2 using fewer donor (murine) residues than the Kabat CDRs, thus further reducing the risk for an adverse immune response in a human patient. The analysis also identified advantageous sites for back-mutations. It was further discovered that, possibly due to the high sequence similarity between the ACO-1 and 30 ACO-2 variable regions (differing only at 13 sites), certain amino acid residues in the humanized ACO-1 (hzACO-1) sequence could be replaced by ACO-2 residues at the corresponding position. Within CDR regions, mutations can normally be made without rendering the anti-body sequence less human. This humanization procedure resulted in improved functional

properties such as affinity, stability, expression level and IFN- α -inhibitory activity of the humanized antibody.

In a humanized ACO-1 antibody, exemplary mutations in the hzACO-1 VH (SEQ ID NO:3) include V5Q, T28S, M69L, R71V, T73K, S76I, S76N, T77I, V78A, Y79F and A93V, as well as any combination thereof, using Kabat numbering. Exemplary mutations in the hzACO-1 VL (SEQ ID NO:6) include E1Q, D29G, L33F, L47W, S50G, I58V, and F71Y, as well as any combination thereof. In one embodiment, the hzACO-1 VH region comprises a mutation selected from T28S, N31S, and A93V. In another embodiment, the hzACO-1 VH region comprises a mutation selected from T28S, N31S, and A93V, and any combination thereof, such as, e.g., T28S and N31S, T28S and A93V, and N31S and A93V. In another embodiment, the hzACO-1 VH region comprises a mutation selected from T28S, N31S, and A93V, or a combination thereof, such as, e.g., T28S and N31S, T28S and A93V, or N31S and A93V; and at least one additional mutation.

Definitions

To facilitate the understanding of this invention, a number of terms are defined below.

“ACO-1 and ACO-2 antibodies” are characterized and described in WO20060086586. ACO-1 is deposited with ATCC accession no. PTA-6557 (WO2006086586) and ACO-2 is deposited as ATCC accession No. PTA-7778 (WO2008021976). The antibodies according to the present invention are humanized variants of ACO-1 and ACO-2. However, the humanized versions of ACO-1 and ACO-2 according to the present invention do not comprise the full length murine Kabat sequences. In a preferred embodiment, at least one of the CDR sequences comprise a truncation of about 3-10 amino acids, preferably 3-8, more preferably 4-7 amino acids. The antibody preferably comprises a truncation in the CDR H2, said CDR H2 preferably being truncated by 3-10, preferably 3-8, more preferably 4-7, and most preferably by 6 amino acids. And it furthermore follows that occasional point mutations may be introduced in one or more CDR sequences as well as within the human scaffold antibody. The term “ACO-1 and ACO-2 antibodies” may however furthermore embrace any IFN- α antibody capable of binding IFN- α subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b and WA, but not subtypes 1 or D. It should however be understood that ACO-1 and ACO-2 as such may be perceived as only one antibody since the differences in their CDR sequences are only a few amino acids. It is plausible that ACO-1 and ACO-2 thus represent two stages of *in vivo* somatic hypermutation of the same antibody. An ACO-1/ACO-2 antibody according to the present invention is thus a humanized antibody comprising

ing CDR sequences having at least 90% identity with the CDR sequences of ACO-1 and ACO-2, more preferably at least 92%, and most preferably at least 95%.

The terms: “CDR truncation”, “forward mutation”, and “shortening of CDRs” may be used interchangeably throughout the document. In connection with the present invention such terms generally refer to the fact that CDR-truncation may be perceived as a number of forward mutations in a row – meaning that a shortened murine CDR fragment can be grafted onto the human framework. While it may not be surprising that grafting of shorter CDRs tend to result in antibodies with reduced degree of immunogenicity, it is actually surprising that other advantageous features of the humanized antibody may be retained such as e.g. stability, specificity, etc. “Back mutations” always refer to mutations in the framework (i.e. not in the CDRs) – and back mutations are typically introduction of one or more “murine” amino acid residue at selected sites e.g. in order to stabilise the antibody structure.

The term “interferon alpha” (IFN- α), as used herein, refers to a family of proteins that include some of the main effectors of innate immunity. There are at least 15 known subtypes of human IFN- α . The names of the IFN- α protein subtypes and corresponding encoding genes are listed below in Table 1.

Table 1 IFN- α protein subtypes and genes

IFN- α protein subtype	Corresponding IFN- α gene
A	2a
2	2b
B2	8
C	10
D (Val ¹¹⁴)	1
F	21
G	5
H2	14
I	17
J1	7
K	6
4a	4a
4b	4b
WA	16
1 (Ala ¹¹⁴)	1

See Pestka *et al.* (1997) "Interferon Standardization and Designations" *J Interferon Cytokine Res* 17: Supplement 1, S9-S14. IFN- α B2 is sometimes also referred to as IFN- α B, and is not to be confused with IFN- β . Natural IFN- α from leukocytes (leukocyte IFN-), as well as recombinant human IFN- α protein subtypes are available from PBL Biomedical Labs, Piscataway, NJ (interferonsource.com). Natural IFN- α is a complex mixture of IFN- α subtypes. Methods for detecting and quantifying these interferons, such as ELISA and RIA, are known in the art.

The term "antibody" herein is used in the broadest sense and specifically includes full-length monoclonal antibodies, polyclonal antibodies, and, unless otherwise stated or contradicted by context, antigen-binding fragments, antibody variants, and multispecific molecules thereof, so long as they exhibit the desired biological activity. Generally, a full-length antibody is a glycoprotein comprising at least two heavy (H) chains and two light (L) chains inter-connected by disulfide bonds, or an antigen binding portion thereof. Each heavy chain is comprised of a heavy chain variable region (abbreviated herein as VH) and a heavy chain constant region. The heavy chain constant region is comprised of three domains, CH1, CH2 and CH3. Each light chain is comprised of a light chain variable region (abbreviated herein as VL) and a light chain constant region. The light chain constant region is comprised of one domain, CL. The VH and VL regions can be further subdivided into regions of hypervariability, termed complementarily determining regions (CDR), interspersed with regions that are more conserved, termed framework regions (FR). Each VH and VL is composed of three CDRs and four FRs, arranged from amino-terminus to carboxy-terminus in the following order: FR1, CDR1, FR2, CDR2, FR3, CDR3, FR4. The variable regions of the heavy and light chains contain a binding domain that interacts with an antigen.

An "antigen-binding fragment" of an antibody is a molecule that comprises a portion of a full-length antibody which is capable of detectably binding to the antigen. Antigen-binding fragments include multivalent molecules comprising one, two, three, or more antigen-binding portions of an antibody, and single-chain constructs wherein the VL and VH regions, or selected portions thereof, are joined by synthetic linkers or by recombinant methods to form a functional, antigen-binding molecule.

The terms "antibody derivative" and "immunoconjugate" are used interchangeably herein to denote molecules comprising a full-length antibody or an antigen-binding fragment thereof, wherein one or more amino acids are chemically modified, e.g., by alkylation, PEGylation, acylation, ester formation or amide formation or the like, e.g., for linking the antibody to a second molecule. Exemplary modifications include PEGylation, cysteine-PEGylation,

biotinylation, radiolabelling, and conjugation with a second agent, such as a detectable or cytotoxic agent.

A "multispecific molecule" comprises an antibody, or an antigen-binding fragment thereof, which is associated with or linked to at least one other functional molecule (e.g. an 5 other peptide or protein such as another antibody or ligand for a receptor) to generate a molecule that binds to at least two different binding sites or target molecules. Exemplary multispecific molecules include bi-specific antibodies and antibodies linked to soluble receptor fragments or ligands.

A "humanized" antibody is a human/non-human chimeric antibody that contains a 10 minimal sequence (CDR regions) derived from non-human immunoglobulin. Humanized antibodies are thus human immunoglobulins (recipient antibody) in which residues from a hyper-variable region of the recipient are replaced by residues from a hypervariable region of a non-human species (donor antibody) such as mouse, rat, rabbit, or non-human primate having the desired specificity, affinity, and capacity. In some instances, FR residues of the human 15 immunoglobulin are replaced by corresponding non-human residues. Furthermore, humanized antibodies may comprise residues that are not found in the recipient antibody or in the donor antibody. These modifications are made to further refine antibody performance. In general, a humanized antibody will comprise substantially all of at least one, and typically two, variable domains, in which all or substantially all of the hypervariable loops correspond 20 to those of a non-human immunoglobulin and all or substantially all of the FR residues are those of a human immunoglobulin sequence. The humanized antibody can optionally also comprise at least a portion of an immunoglobulin constant region (Fc), typically that of a human immunoglobulin.

The term "hypervariable region" when used herein refers to the amino acid residues 25 of an antibody that are responsible for antigen binding. The hypervariable region generally comprises amino acid residues from a "complementarity-determining region" or "CDR" (residues 24-34 (L1), 50-56 (L2) and 89-97 (L3) in the light-chain variable domain and 31-35 (H1), 50-65 (H2) and 95-102 (H3) in the heavy-chain variable domain; (Kabat *et al.* (1991) Sequences of Proteins of Immunological Interest, Fifth Edition, U.S. Department of Health 30 and Human Services, NIH Publication No. 91-3242) and/or those residues from a "hypervariable loop" (residues 26-32 (L1), 50-52 (L2) and 91-96 (L3) in the light-chain variable domain and 26-32 (H1), 53-55 (H2) and 96-101 (H3) in the heavy-chain variable domain; Chothia and Lesk, *J. Mol. Biol.* 1987;196:901-917). Typically, the numbering of amino acid residues in this region is performed by the method described in Kabat *et al.*, *supra*. Phrases such as 35 "Kabat position", "Kabat residue", and "according to Kabat" herein refer to this numbering

system for heavy chain variable domains or light chain variable domains. Using the Kabat numbering system, the actual linear amino acid sequence of a peptide may contain fewer or additional amino acids corresponding to a shortening of, or insertion into, a FR or CDR of the variable domain. For example, a heavy chain variable domain may include amino acid insertions (residue 52a, 52b and 52c according to Kabat) after residue 52 of CDR H2 and inserted residues (e.g. residues 82a, 82b, and 82c, etc. according to Kabat) after heavy chain FR residue 82. The Kabat numbering of residues may be determined for a given antibody by alignment at regions of homology of the sequence of the antibody with a "standard" Kabat numbered sequence.

10 "Framework region" or "FR" residues are those VH or VL residues other than the CDRs as herein defined.

"Corresponding" amino acid positions in two substantially identical amino acid sequences are those aligned by any of the protein analysis software referred to herein, typically using default parameters.

15 An "isolated" molecule is a molecule that is the predominant species in the composition wherein it is found with respect to the class of molecules to which it belongs (i.e., it makes up at least about 50% of the type of molecule in the composition and typically will make up at least about 70%, at least about 80%, at least about 85%, at least about 90%, at least about 95%, or more of the species of molecule, e.g., peptide, in the composition).

20 Commonly, a composition of an antibody molecule will exhibit 98%, 98%, or 99% homogeneity for antibody molecules in the context of all present peptide species in the composition or at least with respect to substantially active peptide species in the context of proposed use.

 The terms, "selectively neutralizes" and "selectively neutralizing", as used herein, refer to an isolated and purified antibody (such as, but not limited to a monoclonal antibody), or 25 an antigen-binding fragment thereof, that neutralizes selectively at least about 40%, at least about 50%, or at least about 60% of a bioactivity of one or more IFN- α protein subtypes, but does not significantly neutralize at least one bioactivity of another IFN- α protein subtype, wherein the bioactivity can be, e.g., activation of the MxA promoter and/or antiviral activity.

 In the context of the present invention, "treatment" or "treating" refers to preventing, 30 alleviating, managing, curing or reducing one or more symptoms or clinically relevant manifestations of a disease or disorder, unless contradicted by context. For example, "treatment" of a patient in whom no symptoms or clinically relevant manifestations of a disease or disorder have been identified is preventive or prophylactic therapy, whereas "treatment" of a patient in whom symptoms or clinically relevant manifestations of a disease or disorder have 35 been identified generally does not constitute preventive or prophylactic therapy.

The phrase "IFN- α -related condition or disease", as used herein, refers to an abnormal condition, disease, or pre-clinical disease state that has been linked with elevated levels of IFN- α in a patient's serum. Examples of such include, but are not limited to, lupus diseases or disorders such as SLE, graft versus host disease (GVHD), type 1 diabetes, AIDS 5 (caused by human immunodeficiency virus (HIV)), autoimmune thyroiditis, and psoriasis. Methods for determining the level of IFN α are known in the art.

Humanized Anti-IFN- α antibodies

The antibodies of the invention are humanized version of the anti-IFN- α mouse antibodies ACO-1 or ACO-2, variants thereof, and/or antigen-binding fragments thereof, characterized by particular functional and/or structural features or properties. Recombinant antibodies can be produced in suitable host cell lines by standard techniques and be characterized by various assays to evaluate their functional activities, as described below. In fact, it turns out that IFN-alpha antibodies according to the present invention may be produced with a significantly improved yield compared to IFN-alpha antibodies that were humanized in a traditional way.

According to the so-called "best-fit" method, the sequence of the variable domain of a rodent antibody is screened against a library of such known human variable-domain sequences or libraries of human germline sequences. The human sequence that is closest to that of the rodent can then be accepted as the human framework region for the humanized 20 antibody (Sims *et al.*, J. Immunol. 1993;151:2296 et seq.; Chothia *et al.*, Chothia and Lesk, J. Mol. Biol 1987;196:901-917). Another method uses a particular framework region derived from the consensus sequence of all human antibodies of a particular subgroup of light or heavy chains. The same framework may be used for several different humanized antibodies.

Preferred framework sequences for use in the antibodies of the invention are those 25 that are structurally similar to the framework sequences used by ACO-1 or ACO-2. Thus, in one embodiment, the invention provides a humanized ACO-1 or ACO-2 antibody comprising VH framework residues derived from a human VH1_46 gene and a human JH4 gene, and VL framework residues derived from a human VKIII_L6 gene and a human JK2 gene, and specifically binds human IFN- α .

30 Example 1 below describes the design of an exemplary humanized ACO-1 antibody, hzACO-1, comprising such framework sequences.

Functional properties

The humanized antibodies of the invention bind specifically to IFN- α subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, and WA (Table 2). In one embodiment, a humanized ACO-

1 or ACO-2 antibody of the invention binds to an IFN- α protein subtype such as IFN- α A with high affinity, for example with a KD of about 10^{-7} M or less, a KD of about 10^{-8} M or less, a KD of about 5×10^{-9} M or less, or a KD of about 2×10^{-9} M or less. In one embodiment, the humanized antibody is a hzACO-1 variant which binds to IFN- α A, IFN- α F, and/or another 5 IFN- α protein subtypes with an affinity comparable to or higher than that of hzACO-1.

Table 2. Kinetic parameters of hzACO-1 for a range of human IFN- α subtypes

Sample	k_a (1/Ms)	k_d (1/s)	KD (M)
hIFN- α A	2,97E+05	3,94E-04	1,33E-09
hIFN- α 1	No binding	-	-
hIFN- α 2	3,58E+05	3,51E-04	9,81E-10
hIFN- α 4b	3,74E+05	6,22E-04	1,67E-09
hIFN- α G	4,63E+05	4,26E-04	9,20E-10
hIFN- α H2	3,78E+05	1,21E-03	3,21E-09
hIFN- α I	7,23E+05	2,03E-03	2,81E-09
hIFN- α J1	6,81E+05	3,27E-03	4,81E-09
hIFN- α WA	7,09E+05	2,91E-03	4,10E-09
hIFN- α 4a	3,33E+04	1,15E-04	3,45E-09
hIFN- α C	7,19E+05	7,53E-04	1,05E-09
hIFN- α K	5,74E+05	8,27E-04	1,44E-09

10

For example, the ratio between the KD of the hzACO-1 variant and the KD of hzACO-1 to a IFN- α A protein subtype can be about 1.0, about 0.8, about 0.7, or about 0.6. In another embodiment, the humanized antibody is a hzACO-1 variant which binds to IFN- α A, IFN- α F, and/or another IFN- α protein subtype with an affinity comparable to or higher 15 than that of recombinantly produced ACO-1. In another embodiment, the humanized antibody is a hzACO-1 variant which binds to IFN- α A, IFN- α F, and/or another IFN- α protein subtypes with an affinity comparable to or higher than that of hybridoma-produced ACO-1.

Furthermore, the humanized antibodies of the invention are capable of selectively neutralizing a bioactivity of one or more IFN- α protein subtypes. For example, a humanized 20 ACO-1 or ACO-2 variant can be capable of selectively neutralizing at least about 40%, at least about 50%, or at least about 60% of a bioactivity of IFN- α protein subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, or WA, or any combination thereof, as compared to a control. In a particular embodiment, the humanized antibody does not significantly neutralize the bioactivity of IFN- α subtypes D and/or 1. Exemplary bioactivities include, but are not limited to, activation of the MxA promoter, antiviral activity, or both. The ability of a humanized antibody to 25 neutralize such IFN- α bioactivity can be evaluated using, e.g., the reporter gene (RG) an cy-

topathic inhibition (CPE) assays described herein. In one embodiment, the humanized antibody is a hzACO-1 variant which has an IC₅₀ comparable to or lower than the IC₅₀ of hzACO-1 in an RG assay. In a specific embodiment, the hzACO-1 variant has a lower IC₅₀ than hzACO-1 in an RG assay.

5 In one embodiment, the humanized antibodies of the invention compete with and/or bind to the same epitope on an IFN- α protein subtype as ACO-1 and/or ACO-2. Such antibodies can be identified based on their ability to cross-compete with ACO-1 and/or ACO-2 in standard IFN- α binding assays as described herein. The ability of a test humanized antibody antibody to inhibit the binding of ACO-1 or ACO-2 to one or more IFN- α protein subtypes 10 demonstrates that the test antibody can compete with ACO-1 or ACO-2 for binding to IFN- α and thus can bind to the same epitope on the IFN- α protein subtype as ACO-1 and/or ACO-2 (WO02066649 and WO2005059106). In a particular embodiment, the humanized antibody binds to a different human IFN- α epitope than any of the murine monoclonal antibodies 9F3, and MMHA-1, -2, -3, -6, -8, -9, -11, -13, and -17 (PBL Biomedical Laboratories, NJ, USA) 15 and/or human monoclonal antibodies 13H5, 13H7, and 7H9, and/or cross-competes more with ACO-1 or ACO-2 than with one or more of the listed murine and human monoclonal antibodies.

In one embodiment, hzACO-1, hzACO-2, hzACO-1 variants or hzACO-2 variants provided by the invention have an immunogenicity comparable to or lower than that of a humanized ACO-1 or ACO-2 antibody comprising murine CDRs according to Kabat (Kabat 20 ACO-1). Immunogenicity of a humanized antibody can be evaluated by, e.g., one or more of the methods described in Wadwha et al., *Dev Biol (Basel)*. 2005;122:155-70), which is hereby incorporated by reference in its entirety.

In further aspect, the humanized antibodies of the invention are stable in a formulation suitable for administration to a human patient. In one embodiment, the humanized ACO-25 1 or ACO-2 antibody according to the invention is at least as stable as the IFN-alpha antibodies humanized in a traditional way comprising the full length murine Kabat sequences. Stability of an antibody can be evaluated using known methods in the art, including the thera- maflour analyses described in Example 11.

30 Preferred humanized antibodies of the invention exhibit at least one, more preferably two, three, four, five or more, of the following properties: (a) bind specifically to IFN- α subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, and WA; (b) selectively neutralize one or more bioactivities of IFN- α protein subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, or WA; any 35 combination thereof, or all thereof; (c) do not significantly neutralize a bioactivity of IFN- α 1 or D; (d) compete with and/or bind to the same epitope on an IFN- α protein subtype as ACO-1

and/or ACO-2; (e) compete more with ACO-1 or ACO-2 than with any of 9F3, 13H5, 13H7, and 7H9; (f) are less likely to elicit an immune response than a hzACO-1 or hzACO-2 antibody comprising murine CDRs according to Kabat; (g) are stable in pharmaceutical formulations; and (h) binds to at least one of IFN- α protein subtypes A, 2, B2, C, F, G, H2, I, J1, K, 5 4a, 4b, or WA with a KD of 10^{-8} M or less.

Structural properties

Preferred antibodies of the invention are humanized versions of the murine monoclonal antibodies ACO-1 and ACO-2. Such antibodies can be produced, isolated, and structurally and functionally characterized as described in the Examples. Full-length, variable region, and Kabat CDR sequences of ACO-1, hzACO-1 and ACO-2 are set forth in Table 3 and 10 described in Figures 1-3.

Table 3 Sequence numbering for primers, protein and antibodies

Antibody portion	Sequence composition	Sequence	SEQ ID NO:
ACO-1 VH	Protein		1
VH1_46/JH4	Protein		2
hzACO-1 VH	Protein		3
ACO-1 VL	Protein		4
VKIII_L6/JK2	Protein		5
hzACO-1 VL	Protein		6
ACO-2 VH	Protein		7
J558.33/D_/JH3_1	Protein		8
ACO-2 VL	Protein		9
ae4/ JK4_1	Protein		10
PCR Primer ACO-1 cloning	DNA		11
PCR Primer ACO-1 cloning	DNA		12
ACO-1 VH	DNA		13
ACO-1 VL	DNA		14
ACO-1 CDR_H1	Protein	NYWMH	15
ACO-1 CDR_H2	Protein	EINPSHGRTIYNENFKS	16
ACO-1 CDR_H3	Protein	GGLGPAWFAY	17
ACO-1 CDR_L1	Protein	SAGSSVDSSYLY	18

ACO-1 CDR_L2	Protein	STSNLAS	19
ACO-1 CDR_L3	Protein	HQWSSYPFT	20
hzACO-1 CDR_H2	Protein	EINPSHGRTIYAQKFQG	21
ACO-2 CDR_H1	Protein	SYWMH	22
ACO-2 CDR_H2	Protein	EINPSHGRTSYNENFKS	23
ACO-2 CDR_L1	Protein	SAGSSVGSSYFY	24
ACO-2 CDR_L2	Protein	GTSNLAS	25
PCR Primer ACO-2 cloning	DNA		26
PCR Primer ACO-2 cloning	DNA		27
ACO-2 VH	DNA		28
ACO-2 VL	DNA		29
hIFN- α 8	Protein		30
hzACO-1 Fab HC	Protein		31
hzACO-1 LC	Protein		32

The sequences of hzACO-1 CDR H1, H3, L1, L2, and L3 are identical to the corresponding ACO-1 sequences. ACO-2 CDR H3 and L3 are identical to the corresponding ACO-1 CDR sequences. The amino acids shown in italics in the hzACO-1 CDR_H2 sequence correspond to the human framework sequence – in traditionally humanized antibodies the full length Kabat sequence correspond to the ACO-1 CDR_H2 sequence, where all amino acids are derived from the murine antibody.

In one aspect, the invention provides humanized versions of murine ACO-1 and ACO-2 antibodies with fewer donor residues than the Kabat CDRs, *i.e.*, fewer murine residues than a humanized ACO-1 or ACO-2 antibody produced by grafting of the kabat CDRs.

In one embodiment, the humanized antibody specifically binds human IFN- α and is a humanized version of murine antibody ACO-1 or ACO-2, or of a combination thereof, comprising fewer donor amino acid residues than the murine complementary determining regions (CDRs) according to Kabat. The CDR H2 sequence may, for example, comprise fewer donor amino acid residues than those corresponding to Kabat residues 50-65, 50-64, 50-63, 50-62, 50-61, or 50-60. The CDR H2 donor residues may comprise Kabat residues 50-59. Additionally or alternatively, the CDR H2 donor amino acid residues may consist of Kabat residues 50-59. Kabat residues 50-59 correspond to residues 1-11 of SEQ ID NOS:16, 21, and 23. In one embodiment, the remaining VH CDRs may comprise or consist of the Kabat CDRs (see Figures 1-3), *i.e.*, a CDR H1 sequence comprising donor amino acid residues

corresponding to Kabat residues 31-35, and a CDR H3 sequence comprising donor amino acid residues corresponding to Kabat residues 95-102.

In one embodiment, the humanized ACO-1 or ACO-2 antibody may comprise a CDR L1 comprising donor amino acid residues corresponding to Kabat residues 24-34 of the variable region of the ACO-1 light chain (VL), a CDR L2 comprising donor amino acid residues corresponding to Kabat residues 50-56 of the ACO-1 VL region, and a CDR L3 comprising donor amino acid residues corresponding to Kabat residues 89-97 of the ACO-1 VL region (SEQ ID NO:4) or ACO-2 VL region (SEQ ID NO:9). Additionally or alternatively, the antibody may comprise CDR L1 donor amino acid residues consisting of Kabat residues 24-34, CDR L2 donor residues consisting of Kabat residues 50-56, and CDR L3 donor amino acid residues consisting of Kabat residues 89-97. The corresponding amino acid sequences are shown in Table 3.

In one aspect, the invention provides specific humanized ACO-1 antibodies. The humanized ACO-1 antibody specifically binds human IFN- α , and comprises VH CDR sequences substantially identical to the sequences of Kabat residues 31-35, 50-65, and 95-102 of SEQ ID NO:3, with an optional N31S mutation. The antibody may, e.g., comprise a CDR H1 sequence comprising SEQ ID NO:15; a CDR H2 sequence comprising SEQ ID NO:21; and a CDR H3 sequence comprising SEQ ID NO:17. Additionally or alternatively, the antibody may comprise a CDR H1 sequence consisting of SEQ ID NO:15; a CDR H2 sequence consisting of SEQ ID NO:21; and a CDR H3 sequence consisting of SEQ ID NO:17. In one embodiment, the humanized ACO-1 comprises VH framework residues derived from a human VH1_46 gene and/or a human JH4 gene, preferably both. In a specific embodiment, the humanized antibody comprises a VH sequence corresponding to SEQ ID NO:3.

The humanized ACO-1 antibody may further comprise VL CDR sequences substantially identical to the sequences of Kabat residues 24-34, 50-56, and 89-97 of SEQ ID NO:6. The antibody may, e.g., comprise a CDR_L1 sequence comprising SEQ ID NO:18; a CDR_L2 sequence comprising SEQ ID NO:19; and a CDR_L3 sequence comprising SEQ ID NO:20. Additionally or alternatively, the may comprise a CDR_L1 sequence consisting of SEQ ID NO:18; a CDR_L2 sequence consisting of SEQ ID NO:19; and a CDR_L3 sequence consisting of SEQ ID NO:20. In one embodiment, the humanized ACO-1 antibody comprises VL framework residues derived from a human VKIII_L6 gene and/or a human JK2 gene, preferably both. In a specific embodiment, the humanized antibody comprises a VL sequence corresponding to SEQ ID NO:6.

In one aspect, the invention provides an antibody comprising the CDR sequences of ACO-2. The antibody can specifically bind human IFN- α and comprises VH CDR sequences

substantially identical to the sequences of Kabat residues 31-35, 50-59, and 95-102 of SEQ ID NO:7. In one embodiment, the antibody comprises a CDR_H1 sequence comprising SEQ ID NO:22; a CDR_H2 sequence comprising SEQ ID NO:23; and a CDR_H3 sequence comprising SEQ ID NO:17. In an additional or alternative embodiment, the antibody comprises a

5 CDR_H1 sequence consisting of SEQ ID NO:22; a CDR_H2 sequence consisting of SEQ ID NO:23; and a CDR_H3 sequence consisting of SEQ ID NO:17. The antibody may further comprise VL CDR sequences substantially identical to the sequences of residues Kabat residues 24-34, 50-56, and 89-97 of SEQ ID NO:9. In one embodiment, the antibody comprises a CDR_L1 sequence comprising SEQ ID NO:24; a CDR_L2 sequence comprising

10 SEQ ID NO:25; and a CDR_L3 sequence comprising SEQ ID NO:20. Additionally or alternatively, the antibody comprises a CDR_L1 sequence consisting of SEQ ID NO:24; a CDR_L2 sequence consisting of SEQ ID NO:25; and a CDR_L3 sequence consisting of SEQ ID NO:20. The antibody may, in one aspect, be a humanized ACO-2 antibody.

A humanized ACO-1 or ACO-2 antibody may further comprise at least a portion of a human Fc-region (unless the antibody is an antigen-binding fragment not comprising any Fcportion). Typically, the size of the Fc-region is selected to achieve the desired pharmacokinetic properties of the antibody; the larger Fc-portion, the slower clearance. In one embodiment, the humanized antibody is a full-length antibody, preferably comprising an IgG4 isotype Fc-region. In a particular embodiment, the IgG4 Fc-region comprises an S241P mutation, with numbering according to Kabat; corresponding to residue 228 per the EU numbering system (Edelman G.M. et AL., Proc. Natl. Acad. USA 63, 78-85 (1969)).

Given that both ACO-1 and ACO-2 can bind to IFN- α and are similar, the humanized VH and VL sequences can be "mixed and matched" to create other anti-IFN- α binding molecules of the invention. IFN- α binding of such "mixed and matched" antibodies can be tested using the binding assays described herein (e.g. flow cytometry, Biacore, ELISAs) and/or using one or more functional assays as described herein. Preferably, when VH and VL chains are mixed and matched, a VH sequence from a particular VH/VL pairing is replaced with a structurally similar VH sequence. Likewise, a VL sequence from a particular VH/VL pairing is preferably replaced with a structurally similar VL sequence.

30 Accordingly, in one aspect, the invention provides an humanized monoclonal antibody, or antigen binding portion thereof, comprising: (a) a VH region comprising ACO-1 or ACO-2 VH CDRs and (b) a VL region comprising ACO-1 or ACO-2 VL CDRs; wherein the antibody specifically binds IFN- α . Preferred heavy and light chain combinations include: (a) a VH region comprising SEQ ID NOS:15-17, optionally omitting some or all of the 5 C-terminal amino acids of SEQ ID NO:16, and (b) a light chain variable region comprising SEQ

35

ID NOS:18-20; (a) a VH region comprising SEQ ID NOS:15-17, optionally omitting some or all of the 5 C-terminal amino acids of SEQ ID NO:16, and (b) a light chain variable region comprising SEQ ID NOS:24, 25, and 20; (a) a VH region comprising SEQ ID NOS:22, 23, and 17, optionally omitting some or all of the 5 C-terminal amino acids of SEQ ID NO:23, and (b) a light chain variable region comprising SEQ ID NOS:18-20; and (a) a VH region comprising SEQ ID NOS:22, 23, and 17, optionally omitting some or all of the 5 C-terminal amino acids of SEQ ID NO:23, and (b) a light chain variable region comprising SEQ ID NOS:24, 25, and 20. Other preferred heavy and light chain combinations include (a) a VH region comprising the sequence of SEQ ID NO:3 and (b) a VL region comprising the amino acid sequence of SEQ ID NO:4; (a) a VH comprising SEQ ID NOS:15, 21, and 17, and (b) a VL comprising SEQ ID NOS: 18-20; and (a) a VH comprising SEQ ID NOS:15, 21, and 17, and (b) a VL comprising SEQ ID NOS:24, 25, and 20.

In another aspect, the invention provides antibodies that comprise the heavy chain and light chain CDR1s, CDR2s and/or CDR3s of ACO-1 or ACO-2, or combinations thereof. Given that each of these antibodies can bind to IFN- α and that antigen-binding specificity is provided primarily by the CDR1, 2 and 3 regions, the CDR H1, H2 and H3 sequences and CDR L1, L2 and L3 sequences can be "mixed and matched" (i.e., CDRs from different antibodies can be mixed and match, although each antibody can contain a CDR H1, H2 and H3 and a CDR L1, L2 and L3) to create other anti-IFN- α binding molecules of the invention. IFN- α -binding of such "mixed and matched" antibodies can be tested using the binding assays described below and in the Examples (e.g. flow cytometry, Biacore, or ELISAs). Preferably, when VH CDR sequences are mixed and matched, the CDR H1, H2 and/or H3 sequence from a particular VH sequence is replaced with a structurally similar CDR sequence(s). Likewise, when VL CDR sequences are mixed and matched, the CDR L1, L2 and/or L3 sequence from a particular VL sequence preferably is replaced with a structurally similar CDR sequence(s). For example, the CDRs of ACO-1 and ACO-2 share substantial structural similarity and therefore are amenable to mixing and matching.

Accordingly, in another aspect, the invention provides a humanized monoclonal antibody, or antigen binding portion thereof comprising: (a) a CDR H1 comprising an amino acid sequence selected from the group consisting of SEQ ID NOS:15 and 22; (b) a CDR H2 comprising an amino acid sequence selected from the group consisting of at least residues 1-12 of SEQ ID NOS:16 and 23, (c) a CDR H3 comprising SEQ ID NO:17; (d) a CDR L1 comprising an amino acid sequence selected from the group consisting of SEQ ID NOS:18 and 24; (e) a CDR L2 comprising an amino acid sequence selected from the group consist-

ing of SEQ ID NOS:19 and 25; and (f) a CDR L3 comprising SEQ ID NO:20; wherein the antibody specifically binds IFN- α .

In a preferred embodiment, the antibody comprises: (a) a CDR H1 comprising SEQ ID NO:15; (b) a CDR H2 comprising at least residues 1-12 of SEQ ID NO:16; (c) a CDR H3 comprising SEQ ID NO:17; (d) a CDR L1 comprising SEQ ID NO:18; (e) a CDR L2 comprising SEQ ID NO:19; and (f) a CDR L3 comprising SEQ ID NO:20.

In another preferred embodiment, the antibody comprises: (a) a CDR H1 comprising SEQ ID NO: 22; (b) a CDR H2 comprising at least residues 1-12 of SEQ ID NO:23; (c) a CDR H3 comprising SEQ ID NO:17; (d) a CDR L1 comprising SEQ ID NO:24; (e) a CDR L2 comprising SEQ ID NO:25; and (f) a CDR L3 comprising SEQ ID NO:20.

In a preferred embodiment, the antibody comprises: (a) a CDR H1 comprising SEQ ID NO:15; (b) a CDR H2 comprising SEQ ID NO:21; (c) a CDR H3 comprising SEQ ID NO:17; (d) a CDR L1 comprising SEQ ID NO:18; (e) a CDR L2 comprising SEQ ID NO:19; and (f) a CDR L3 comprising SEQ ID NO:20.

In a preferred embodiment, the antibody comprises: (a) a CDR H1 comprising SEQ ID NO:22; (b) a CDR H2 comprising at least residues 1-12 of SEQ ID NO:16; (c) a CDR H3 comprising SEQ ID NO:17; (d) a CDR L1 comprising SEQ ID NO:18; (e) a CDR L2 comprising SEQ ID NO:19; and (f) a CDR L3 comprising SEQ ID NO:20.

In a preferred embodiment, the antibody comprises: (a) a CDR H1 comprising SEQ ID NO:15; (b) a CDR H2 comprising at least residues 1-12 of SEQ ID NO:16; (c) a CDR H3 comprising SEQ ID NO:17; (d) a CDR L1 comprising SEQ ID NO:24; (e) a CDR L2 comprising SEQ ID NO:19; and (f) a CDR L3 comprising SEQ ID NO:20.

In a preferred embodiment, the antibody comprises: (a) a CDR H1 comprising SEQ ID NO:15; (b) a CDR H2 comprising at least residues 1-12 of SEQ ID NO:16; (c) a CDR H3 comprising SEQ ID NO:17; (d) a CDR L1 comprising SEQ ID NO:18; (e) a CDR L2 comprising SEQ ID NO:25; and (f) a CDR L3 comprising SEQ ID NO:20.

Humanized Anti-IFN- α antibody variants

Though an antibody variant or derivative typically has at least one altered property as compared to the parent antibody, antibody variants or derivatives can retain one, some, most, or all of the functional properties of the parent anti-IFN- α antibody, including, but not limited to: (a) bind specifically to IFN- α subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, and WA; (b) selectively neutralize one or more bioactivities of IFN- α protein subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, or WA; any combination thereof, or all thereof; (c) do not significantly neutralize a bioactivity of IFN- α 1 or D; (d) compete with and/or bind to the same epi-

tope on an IFN- α protein subtype as ACO-1 and/or ACO-2; (e) compete more with ACO-1 or ACO-2 than with any of 9F3, 13H5, 13H7, and 7H9; (f) are less likely to elicit an immune response than a hzACO-1 or hzACO-2 antibody comprising murine CDRs according to Kabat; (g) are stable in pharmaceutical formulations; and (h) binds to at least one of IFN- α protein 5 subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, or WA with a KD of 10^{-8} M or less. Any combination of the above-described functional features, and/or the functional features as described in the Examples, may be exhibited by an antibody of the invention.

In certain embodiments, a humanized antibody of the invention comprises a VH region comprising CDR H1-H3 sequences and a VL region comprising CDR L1-L3 sequences, 10 wherein one or more of these CDR sequences comprise specified amino acid sequences based on the preferred antibodies described herein; ACO-1 and ACO-2, or conservative modifications thereof, and wherein the antibodies have retained or improved the desired functional properties of the anti-IFN- α antibodies of the invention. Accordingly, the invention provides an isolated monoclonal antibody, or antigen-binding fragment thereof, comprising a 15 heavy chain variable region comprising CDR H1, CDR H2, and CDR H3 sequences and a light chain variable region comprising CDR H1, CDR H2, and CDR H3 sequences, wherein: (a) a CDR H1 comprising an amino acid sequence selected from the group consisting of SEQ ID NOS:15 and 22, and conservative modifications thereof; (b) a CDR H2 comprising an amino acid sequence selected from the group consisting of at least residues 1-12 of SEQ 20 ID NOS:16 and 23, and conservative modifications thereof, (c) a CDR H3 comprising SEQ ID NO:17, and conservative modifications thereof; (d) a CDR L1 comprising an amino acid sequence selected from the group consisting of SEQ ID NOS:18 and 24, and conservative modifications thereof; (e) a CDR L2 comprising an amino acid sequence selected from the group consisting of SEQ ID NOS:19 and 25, and conservative modifications thereof; and (f) a 25 CDR L3 comprising SEQ ID NO:20, and conservative modifications thereof; wherein the antibody specifically binds IFN- α .

Thus, one or more amino acid residues within the CDR or FR regions of an antibody of the invention can be replaced with other amino acid residues from the same side chain family and the altered antibody can be tested for retained function (*i.e.*, the functions set forth 30 in (c), (d) and (e) above) using the functional assays described herein.

The functional properties of the antibody variants can be assessed using standard assays available in the art and/or described herein. For example, the ability of the antibody to bind IFN- α can be determined using standard binding and biological effects (e.g. reporter gene) assays, such as those set forth in the Examples (e.g., Biacore, or ELISAs).

Variable region modifications

In one aspect, the invention provides a humanized ACO-1 or ACO-2 antibody with mutations in the CDR or framework regions.

Specific exemplary mutations in humanized ACO-1 and their identification are described in Examples 2 and 3 (see, also Figures 1 and 3). These include both back-mutations; introducing ACO-1 residues into humanized ACO-1, as well as mutations where ACO-2-derived residues are introduced into humanized ACO-1. Exemplary back-mutations in the hzACO-1 VH (SEQ ID NO:3) include V5Q, M69L, R71V, T73K, S76I, and V78A, as well as any combination thereof, using Kabat numbering. Exemplary back-mutations in the hzACO-1 VL (SEQ ID NO:6) include E1Q, L47W, I58V, and F71Y, as well as any combination thereof. Exemplary ACO-2-derived mutations in the hzACO-1 VH (SEQ ID NO:3) include T28S, N31S, I58S, S76N, T77I, and A93V, as well as any combination thereof. Exemplary ACO-2 derived mutations in the hzACO-1 VL (SEQ ID NO:6) include D29G, L33F, and S50G, as well as any combination thereof.

Further, various hzACO-1 VH and VL variant sequences can be "mixed and matched" with variant sequences or parent sequences to create a library of hzACO-1 variants of the invention. IFN- α -binding of such "mixed and matched" antibodies can be tested using the binding assays described herein (e.g., Biacore, ELISAs) and/or using one or more functional assays as described herein.

In one embodiment, the invention provides a humanized antibody that specifically binds human IFN- α and contains a variable domain having, incorporated into a human antibody variable domain, amino acids from a donor non-human antibody that binds human IFN- α , comprising a donor antibody amino acid residue at one or more sites selected from 5, 28, 31, 58, 69, 71, 73, 76, 78, 79, and 93 in the heavy chain variable domain.?

In one embodiment, the invention provides a humanized antibody that specifically binds human IFN- α and contains a variable domain having, incorporated into a human antibody variable domain, amino acids from a donor non-human antibody that binds human IFN- α , comprising a donor antibody amino acid residue at one or more sites selected from 1, 29, 33, 47, 50, 58, and 71 in the light chain variable domain.

In one embodiment, the invention provides a humanized ACO-1 antibody that specifically binds human IFN- α and contains a variable domain having, incorporated into a human antibody variable domain, CDR sequences from ACO-1 that bind human IFN- α , and further comprising ACO-2 amino acid residues at one or more sites selected from 28, 31, 58, 76, 77, 78, 79, and 93 in the heavy chain variable domain. In a specific embodiment, the ACO-2 amino acid residues are at one or more sites selected from 28, 31, and 93.

In one embodiment, the invention provides a humanized ACO-1 antibody that specifically binds human IFN- α and contains a variable domain having, incorporated into a human antibody variable domain, CDR sequences from ACO-1 that bind human IFN- α , and further comprising ACO-2 amino acid residues at one or more sites selected from 29, 33, and 5 50 in the light chain variable domain.

In one embodiment, the invention provides a hzACO-1 variant that specifically binds human IFN- α , and comprises VH CDR sequences substantially identical to the sequences of Kabat residues 31-35, 50-65, and 95-102 of SEQ ID NO:3, with an N31S mutation. The antibody may, e.g., comprise a CDR H1 sequence comprising SEQ ID NO:15 with an N31S 10 mutation; a CDR H2 sequence comprising SEQ ID NO:21; and a CDR H3 sequence comprising SEQ ID NO:17. Additionally or alternatively, the antibody may comprise a CDR H1 sequence consisting of SEQ ID NO:15 with an N31S mutation; a CDR H2 sequence consisting of SEQ ID NO:21; and a CDR H3 sequence consisting of SEQ ID NO:17. In one embodiment, the humanized ACO-1 comprises VH framework residues derived from a human 15 VH1_46 gene and/or a human JH4 gene, preferably both. In a specific embodiment, the humanized antibody comprises a VH sequence corresponding to SEQ ID NO:3, with an N to S mutation at Kabat position 31. As shown in the Examples, an N31S mutation in hzACO-1 increased the binding affinity to IFN- α A to levels comparable to that of ACO-1, and increased the stability at pH3.5 and 4.5. Moreover, since residue 31 is in a "donor" CDR residue, the 20 mutation does not introduce a further murine residue into the hzACO-1 sequence, thus not increasing the risk for an immune response against the antibody when administered to a human. Other CDR mutations with the same advantage include D29G and S50G in the hzACO-1 VL.

In one embodiment, the invention provides a hzACO-1 variant that specifically binds 25 human IFN- α , and comprises VH CDR sequences substantially identical to the sequences of Kabat residues 31-35, 50-65, and 95-102 of SEQ ID NO:3, with a T28S mutation. The antibody may, e.g., comprise a CDR H1 sequence comprising SEQ ID NO:15; a CDR H2 sequence comprising SEQ ID NO:21; and a CDR H3 sequence comprising SEQ ID NO:17. Additionally or alternatively, the antibody may comprise a CDR H1 sequence consisting of 30 SEQ ID NO:15; a CDR H2 sequence consisting of SEQ ID NO:21; and a CDR H3 sequence consisting of SEQ ID NO:17. In one embodiment, the humanized ACO-1 comprises VH framework residues derived from a human VH1_46 gene, further comprising a T28S mutation. In a specific embodiment, the humanized antibody comprises a VH sequence corresponding to SEQ ID NO:3, with an T to S mutation at Kabat position 28. As shown in the

Examples, a T28S mutation in hzACO-1 increased the binding affinity to IFN- α A to levels comparable to that of ACO-1, and increased stability at pH3.5 and 4.5.

In one embodiment, the invention provides a hzACO-1 variant that specifically binds human IFN- α , and comprises VH CDR sequences substantially identical to the sequences of Kabat residues 31-35, 50-65, and 95-102 of SEQ ID NO:3, with an A93V mutation. The antibody may, e.g., comprise a CDR H1 sequence comprising SEQ ID NO:15; a CDR H2 sequence comprising SEQ ID NO:21; and a CDR H3 sequence comprising SEQ ID NO:17. Additionally or alternatively, the antibody may comprise a CDR H1 sequence consisting of SEQ ID NO:15; a CDR H2 sequence consisting of SEQ ID NO:21; and a CDR H3 sequence consisting of SEQ ID NO:17. In one embodiment, the humanized ACO-1 comprises VH framework residues derived from a human VH1_46 gene and a human JH4 gene, further comprising an A93V mutation. In a specific embodiment, the humanized antibody comprises a VH sequence corresponding to SEQ ID NO:3, with an A to V mutation at Kabat position 93. As shown in the Examples, an A93V mutation in hzACO-1 increased the binding affinity to IFN- α A to levels comparable to that of ACO-1, increased the potency for inhibition of IFN- effects as measured in the RG assay, and increased stability at pH3.5 and 4.5.

In one embodiment, the invention provides a hzACO-1 variant that specifically binds human IFN- α , and comprises VH CDR sequences substantially identical to the sequences of Kabat residues 31-35, 50-65, and 95-102 of SEQ ID NO:3, further comprising a mutation in one of the VH CDR sequences, wherein the mutation is not in Kabat residue 58. The antibody may, e.g., comprise VH CDRs consisting of Kabat residues 31-35, 50-65, and 95-102 of SEQ ID NO:3, further comprising a mutation in one of the VH CDR sequences, wherein Kabat residue 58 is I. In one embodiment, the humanized ACO-1 comprises VH framework residues derived from a human VH1_46 gene and a human JH4 gene. As shown in the Examples, an I58S mutation in hzACO-1 substantially decreased the binding affinity to IFN- α A and decreased the potency for inhibition of IFN- effects as measured in the RG assay.

Another type of framework modification involves mutating one or more residues within the framework region, or even within one or more CDR regions, to remove T cell epitopes to thereby reduce the potential immunogenicity of the antibody. This approach is also referred to as "deimmunization" and is described in further detail in U.S. Patent Publication No. 20030153043 by Carr *et al.*

Fc modifications

In addition or as an alternative to modifications made within the framework or CDR regions, antibodies of the invention may be engineered to include modifications within the Fc region, typically to alter one or more functional properties of the antibody, such as serum

half-life, complement fixation, Fc receptor binding, protein stability and/or antigen-dependent cellular cytotoxicity, or lack thereof. Furthermore, an antibody of the invention may be chemically modified (e.g., one or more chemical moieties can be attached to the antibody) or be modified to alter its glycosylation, again to alter one or more functional properties of the antibody. Each of these embodiments is described in further detail below. The residues in the Fc region are numbered according to Kabat.

If desired, the class of an antibody may be "switched" by known techniques. Such techniques include, e.g., the use of direct recombinant techniques (see e.g., US Patent 4,816,397) and cell-cell fusion techniques (see e.g., US Patent 5,916,771). For example, an antibody that was originally produced as an IgM molecule may be class switched to an IgG antibody. Class switching techniques also may be used to convert one IgG subclass to another, e.g., from IgG1 to IgG2. Thus, the effector function of the antibodies of the invention may be changed by isotype switching to, e.g., an IgG1, IgG2, IgG3, IgG4, IgD, IgA, IgE, or IgM antibody for various therapeutic uses. Exemplary cDNA sequences for constant regions are available via, e.g., GenBank, each of which incorporated by reference in its entirety, are as follows:

Human IgG1 constant heavy chain region: GenBank accession No.: J00228;

Human IgG2 constant heavy chain region: GenBank accession No.: J00230;

Human IgG3 constant heavy chain region: GenBank accession No.: X04646;

Human IgG4 constant heavy chain region: GenBank accession No.: K01316; and

Human kappa light chain constant region: GenBank accession No.: J00241.

In one embodiment, the hinge region of CH1 is modified such that the number of cysteine residues in the hinge region is altered, e.g., increased or decreased. This approach is described further in U.S. Patent No. 5,677,425 by Bodmer *et al.* The number of cysteine residues in the hinge region of CH1 is altered to, for example, facilitate assembly of the light and heavy chains or to increase or decrease the stability of the antibody.

In another embodiment, the Fc hinge region of an antibody is mutated to decrease the biological half life of the antibody. More specifically, one or more amino acid mutations are introduced into the CH2-CH3 domain interface region of the Fc-hinge fragment such that the antibody has impaired Staphylococcal protein A (SpA) binding relative to native Fc-hinge domain SpA binding. This approach is described in further detail in U.S. Patent No. 6,165,745 by Ward *et al.* In another embodiment, the antibody is modified to increase its biological half life. Various approaches are possible. For example, one or more of the following mutations can be introduced: T252L, T254S, T256F, as described in U.S. Patent No. 6,277,375 to Ward. Alternatively, to increase the biological half life, the antibody can be al-

tered within the CH1 or CL region to contain a salvage receptor binding epitope taken from two loops of a CH2 domain of an Fc region of an IgG, as described in U.S. Patent Nos. 5,869,046 and 6,121,022 by Presta *et al.* In yet other embodiments, the Fc region is altered by replacing at least one amino acid residue with a different amino acid residue to alter the 5 effector function(s) of the antibody. For example, one or more amino acids selected from amino acid residues 234, 235, 236, 237, 297, 318, 320 and 322 can be replaced with a different amino acid residue such that the antibody has an altered affinity for an effector ligand but retains the antigen-binding ability of the parent antibody. The effector ligand to which affinity is altered can be, for example, an Fc receptor or the C1 component of complement.

10 This approach is described in further detail in U.S. Patent Nos. 5,624,821 and 5,648,260, both to Winter *et al.* In another example, one or more amino acids selected from amino acid residues 329, 331 and 322 can be replaced with a different amino acid residue such that the antibody has altered C1q binding and/or reduced or abolished complement dependent cytotoxicity (CDC). This approach is described in further detail in U.S. Patent Nos. 6,194,551 by 15 Idusogie *et al.* In another example, one or more amino acid residues within amino acid positions 231 and 239 are altered to thereby alter the ability of the antibody to fix complement. This approach is described further in PCT Publication WO 94/29351 by Bodmer *et al.* In yet another example, the Fc region is modified to increase the ability of the antibody to mediate antibody dependent cellular cytotoxicity (ADCC) and/or to increase the affinity of the antibody 20 for an Fcγ receptor by modifying one or more amino acids at the following positions: 238, 239, 248, 249, 252, 254, 255, 256, 258, 265, 267, 268, 269, 270, 272, 276, 278, 280, 283, 285, 286, 289, 290, 292, 293, 294, 295, 296, 298, 301, 303, 305, 307, 309, 312, 315, 320, 322, 324, 326, 327, 329, 330, 331, 333, 334, 335, 337, 338, 340, 360, 373, 376, 378, 382, 388, 389, 398, 414, 416, 419, 430, 434, 435, 437, 438 or 439. This approach is described 25 further in PCT Publication WO 00/42072 by Presta. Moreover, the binding sites on human IgG1 for FcγRI, FcγRII, FcγRIII and FcRn have been mapped and variants with improved binding have been described (see Shields, R.L. *et al.* (2001) *J. Biol. Chem.* 276:6591-6604). Specific mutations at positions 256, 290, 298, 333, 334 and 339 were shown to improve 30 binding to FcγRIII. Additionally, the following combination mutants were shown to improve FcγRIII binding: T256A/S298A, S298A/E333A, S298A/K224A and S298A/E333A/K334A.

The constant region may further be modified to stabilize the antibody, e.g., to reduce the risk of a bivalent antibody separating into two monovalent VH-VL fragments. For example, in an IgG4 constant region, residue S241 may be mutated to a proline (P) residue to allow complete disulphide bridge formation at the hinge (see, e.g., Angal *et al.*, *Mol Immunol.* 35 1993;30:105-8).

Glycosylation modifications

In still another embodiment, the glycosylation of an antibody is modified. For example, an aglycoslated antibody can be made (*i.e.*, the antibody lacks glycosylation). Glycosylation can be altered to, for example, increase the affinity of the antibody for antigen. Such carbohydrate modifications can be accomplished by, for example, altering one or more sites of glycosylation within the antibody sequence.

Antigen-binding fragments

The anti-IFN- α antibodies of the invention may be prepared as full-length antibodies or antigen-binding fragments thereof. Examples of antigen-binding fragments include Fab, 10 Fab', F(ab)2, F(ab')2, F(ab)3, Fv (typically the VL and VH domains of a single arm of an antibody), single-chain Fv (scFv; see *e.g.*, Bird *et al.*, *Science* 1988;242:423-426; and Huston *et al.* *PNAS* 1988;85:5879-5883), dsFv, Fd (typically the VH and CH1 domain), and dAb (typically a VH domain) fragments; VH, VL, VhH, and V-NAR domains; monovalent molecules comprising a single VH and a single VL chain; minibodies, diabodies, triabodies, tetrabodies, 15 and kappa bodies (see, *e.g.*, Ill *et al.*, *Protein Eng* 1997;10:949-57); camel IgG; IgNAR; as well as one or more isolated CDRs or a functional paratope, where the isolated CDRs or antigen-binding residues or polypeptides can be associated or linked together so as to form a functional antibody fragment. Various types of antibody fragments have been described or reviewed in, *e.g.*, Holliger and Hudson, *Nat Biotechnol* 2005;23:1126-1136; 20 WO2005040219, and published U.S. Patent Applications 20050238646 and 20020161201.

Antibody fragments can be obtained using conventional recombinant or protein engineering techniques, and the fragments can be screened for antigen-binding or other function in the same manner as are intact antibodies.

Various techniques have been developed for the production of antibody fragments. 25 Traditionally, these fragments were derived via proteolytic digestion of full-length antibodies (see, *e.g.*, Morimoto *et al.*, *Journal of Biochemical and Biophysical Methods*, 24:107-117 (1992); and Brennan *et al.*, *Science*, 229:81 (1985)). However, these fragments can now be produced directly by recombinant host cells. Alternatively, Fab'-SH fragments can be directly recovered from *E. coli* and chemically coupled to form F(ab')2 fragments (Carter *et al.*, 30 *Bio/Technology*, 10:163-167 (1992)). According to another approach, F(ab')2 fragments can be isolated directly from recombinant host cell culture. In other embodiments, the antibody of choice is a single-chain Fv fragment (scFv). See WO 1993/16185; U.S. Pat. No. 5,571,894; and U.S. Pat. No. 5,587,458. The antibody fragment may also be a "linear antibody", *e.g.*, as

described in U.S. Pat. No. 5,641,870, for example. Such linear antibody fragments may be monospecific or bispecific.

Multispecific Molecules

In another aspect, the present invention features multispecific molecules comprising 5 an anti-IFN- α antibody, or an antigen-fragment thereof, of the invention. Such multispecific molecules include bispecific molecules comprising at least one first binding specificity for IFN- α and a second binding specificity for a second target epitope.

One type of bispecific molecules are bispecific antibodies. Bispecific antibodies are antibodies that have binding specificities for at least two different epitopes. Methods for making 10 bispecific antibodies are known in the art, and traditional production of full-length bispecific antibodies is usually based on the coexpression of two immunoglobulin heavy-chain-light-chain pairs, where the two chains have different specificities (Millstein *et al.*, *Nature*, 305: 537-539 (1983)). Bispecific antibodies can be prepared as full-length antibodies or antibody 15 fragments (e.g. F(ab')2 bispecific antibodies) or any other antigen-binding fragments described herein.

Other multispecific molecules include those produced from the fusion of a IFN- α -binding antibody moiety to one or more other non-antibody proteins. Such multispecific proteins and how to construct them have been described in the art. See, e.g., Dreier *et al.* (Bioconjug. Chem. 9(4): 482-489 (1998)); U.S. Patent 6,046,310; U.S. Patent Publication No. 20 20030103984; European Patent Application 1 413 316; US Patent Publication No. 20040038339; von Strandmann *et al.*, *Blood* (2006;107:1955-1962.), and WO 2004056873.

Multispecific molecules with more than two valencies are also contemplated. For example, trispecific antibodies can be prepared. Tutt *et al.*, *J. Immunol.*, 147: 60 (1991).

The multispecific molecules of the present invention can be prepared by conjugating 25 the constituent binding specificities using methods known in the art. For example, each binding specificity of the multispecific molecule can be generated separately and then conjugated to one another. When the binding specificities are proteins or peptides, a variety of coupling or cross-linking agents can be used for covalent conjugation. Examples of cross-linking agents include protein A, carbodiimide, N-succinimidyl-S-acetyl-thioacetate (SATA), 5,5'-30 dithiobis(2-nitrobenzoic acid) (DTNB), o-phenylenedimaleimide (oPDM), N-succinimidyl-3-(2-pyridyldithio)propionate (SPDP), and sulfosuccinimidyl 4-(N-maleimidomethyl) cyclohexane-1-carboxylate (sulfo-SMCC) (see e.g., Karpovsky *et al.* (1984) *J. Exp. Med.* 160:1686; Liu, MA *et al.* (1985) *Proc. Natl. Acad. Sci. USA* 82:8648). Other methods include those described in Paulus (1985) *Behring Ins. Mitt.* No. 78, 118-132; Brennan *et al.* (1985) *Science*

229:81-83), and Glennie *et al.* (1987) *J. Immunol.* 139: 2367-2375). Preferred conjugating agents are SATA and sulfo-SMCC, both available from Pierce Chemical Co. (Rockford, IL).

When the binding specificities are antibodies, they can be conjugated via sulphydryl bonding of the C-terminus hinge regions of the two heavy chains. In a particularly preferred 5 embodiment, the hinge region is modified to contain an odd number of sulphydryl residues, preferably one, prior to conjugation.

Alternatively, both binding specificities can be encoded in the same vector and expressed and assembled in the same host cell. This method is particularly useful where the bispecific molecule is a mAb x mAb, mAb x Fab, Fab x F(ab')2 or ligand x Fab fusion protein.

10 A bispecific molecule of the invention can be a single chain molecule comprising one single chain antibody and a binding determinant, or a single chain bispecific molecule comprising two binding determinants. Bispecific molecules may comprise at least two single chain molecules. Methods for preparing bispecific molecules are described or reviewed in, for example in U.S. Patent Number 5,260,203; U.S. Patent Number 5,455,030; U.S. Patent Number 15 4,881,175; U.S. Patent Number 5,132,405; U.S. Patent Number 5,091, 513; U.S. Patent Number 5,476,786; U.S. Patent Number: 5,013,653; U.S. Patent Number 5,258,498; U.S. Patent Number 5,482,858; U.S. Patent application publication 20030078385, Kontermann *et al.*, (2005) *Acta Pharmacological Sinica* 26(1):1-9; Kostelny *et al.*, (1992) *J. Immunol.* 148(5):1547-1553; Hollinger *et al.*, (1993) *PNAS (USA)* 90:6444-6448; and Gruber *et al.* 20 (1994) *J. Immunol.* 152: 5368.

Antibody derivatives

Antibody derivatives (or immunoconjugates) within the scope of this invention include anti-IFN- α antibodies conjugated or covalently bound to a second agent.

For example, in one aspect, the invention provides immunoconjugates comprising 25 an antibody conjugated or covalently bonded to a cytotoxic agent, which cytotoxic agent can be selected from therapeutic radioisotopes, toxic proteins, toxic small molecules, such as drugs, toxins, immunomodulators, hormones, hormone antagonists, enzymes, oligonucleotides, enzyme inhibitors, therapeutic radionuclides, angiogenesis inhibitors, chemotherapeutic drugs, vinca alkaloids, anthracyclines, epidophyllotoxins, taxanes, antimetabolites, alkylating agents, antibiotics, COX-2 inhibitors, SN-38, antimitotics, antiangiogenic and apoptotic agents, particularly doxorubicin, methotrexate, taxol, CPT-11, camptothecans, nitrogen mustards, gemcitabine, alkyl sulfonates, nitrosoureas, triazenes, folic acid analogs, pyrimidine analogs, purine analogs, platinum coordination complexes, *Pseudomonas* exotoxin, ricin, abrin, 5-fluorouridine, ribonuclease (RNase), DNase I, Staphylococcal enterotoxin-A, poke-

weed antiviral protein, gelonin, diphtherin toxin, *Pseudomonas* exotoxin, and *Pseudomonas* endotoxin and others (see, e.g., Remington's Pharmaceutical Sciences, 19th Ed. (Mack Publishing Co. 1995); Goodman and Gilman's The Pharmacological Basis of Therapeutics (McGraw Hill, 2001); Pastan *et al.* (1986) *Cell* 47:641; Goldenberg (1994) *Cancer Journal for Clinicians* 44:43; U.S. Pat. No. 6,077,499; the entire disclosures of which are herein incorporated by reference). It will be appreciated that a toxin can be of animal, plant, fungal, or microbial origin, or can be created *de novo* by chemical synthesis.

In another embodiment, the antibody is derivatized with a radioactive isotope, such as a therapeutic radionuclide or a radionuclide suitable for detection purposes. Any of a number of suitable radioactive isotopes can be used, including, but not limited to, I-131, Indium-111, Lutetium-171, Bismuth-212, Bismuth-213, Astatine-211, Copper-62, Copper-64, Copper-67, Yttrium-90, Iodine-125, Iodine-131, Phosphorus-32, Phosphorus-33, Scandium-47, Silver-111, Gallium-67, Praseodymium-142, Samarium-153, Terbium-161, Dysprosium-166, Holmium-166, Rhenium-186, Rhenium-188, Rhenium-189, Lead-212, Radium-223, Actinium-225, Iron-59, Selenium-75, Arsenic-77, Strontium-89, Molybdenum-99, Rhodium-105, Palladium-109, Praseodymium-143, Promethium-149, Erbium-169, Iridium-194, Gold-198, Gold-199, and Lead-211. In general, the radionuclide preferably has a decay energy in the range of 20 to 6,000 keV, preferably in the ranges 60 to 200 keV for an Auger emitter, 100-2,500 keV for a beta emitter, and 4,000-6,000 keV for an alpha emitter. Also preferred are radionuclides that substantially decay with generation of alpha-particles.

The antibody conjugates of the invention can be used to modify a given biological response, where the drug moiety is not to be construed as limited to classical chemical therapeutic agents. For example, the drug moiety may be a protein or polypeptide possessing a desired biological activity. Such proteins may include, for example, an enzymatically active toxin, or active fragment thereof, such as abrin, ricin A, *pseudomonas* exotoxin, or diphtheria toxin; a protein such as tumor necrosis factor or interferon- γ ; or, biological response modifiers such as, for example, lymphokines, interleukin-1 ("IL-1"), interleukin-2 ("IL-2"), interleukin-6 ("IL-6"), granulocyte macrophage colony stimulating factor ("GM-CSF"), granulocyte colony stimulating factor ("G-CSF"), or other growth factors.

The second agent can be linked to the antibody directly or indirectly, using any of a large number of available methods. For example, an agent can be attached at the hinge region of the reduced antibody component via disulfide bond formation, using cross-linkers such as N-succinyl 3-(2-pyridyldithio)propionate (SPDP), or via a carbohydrate moiety in the Fc region of the antibody (see, e.g., Yu *et al.* (1994) *Int. J. Cancer* 56: 244; Wong, *Chemistry of Protein Conjugation and Cross-linking* (CRC Press 1991); Upeslasis *et al.*, "Modification of

Antibodies by Chemical Methods," in *Monoclonal antibodies: principles and applications*, Birch *et al.* (eds.), pages 187-230 (Wiley-Liss, Inc. 1995); Price, "Production and Characterization of Synthetic Peptide-Derived Antibodies," in *Monoclonal antibodies: Production, engineering and clinical application*, Ritter *et al.* (eds.), pages 60-84 (Cambridge University Press 1995), Cattel *et al.* (1989) *Chemistry today* 7:51-58, Delprino *et al.* (1993) *J. Pharm. Sci* 82:699-704; Arpicco *et al.* (1997) *Bioconjugate Chernistry* 8:3; Reisfeld *et al.* (1989) *Antibody, Immunicon. Radiopharrn.* 2:217; the entire disclosures of each of which are herein incorporated by reference). See, also, e.g. Arnon *et al.*, "Monoclonal Antibodies For Immuno-targeting Of Drugs In Cancer Therapy", in *Monoclonal Antibodies And Cancer Therapy*, Reisfeld *et al.* (eds.), pp. 243-56 (Alan R. Liss, Inc. 1985); Hellstrom *et al.*, "Antibodies For Drug Delivery", in *Controlled Drug Delivery* (2nd Ed.), Robinson *et al.* (eds.), pp. 623-53 (Marcel Dekker, Inc. 1987); Thorpe, "Antibody Carriers Of Cytotoxic Agents In Cancer Therapy: A Review", in *Monoclonal Antibodies '84: Biological And Clinical Applications*, Pinchera *et al.* (eds.), pp. 475-506 (1985); "Analysis, Results, And Future Prospective Of The Therapeutic Use Of Radiolabeled Antibody In Cancer Therapy", in *Monoclonal Antibodies For Cancer Detection And Therapy*, Baldwin *et al.* (eds.), pp. 303-16 (Academic Press 1985), and Thorpe *et al.*, "The Preparation And Cytotoxic Properties Of Antibody-Toxin Conjugates", *Immunol. Rev.*, 62:119-58 (1982).

For further discussion of types of cytotoxins, linkers and methods for conjugating therapeutic agents to antibodies, see also Saito, G. *et al.* (2003) *Adv. Drug Deliv. Rev.* 55:199-215; Trail, P.A. *et al.* (2003) *Cancer Immunol. Immunother.* 52:328-337; Payne, G. (2003) *Cancer Cell* 3:207-212; Allen, T.M. (2002) *Nat. Rev. Cancer* 2:750-763; Pastan, I. and Kreitman, R. J. (2002) *Curr. Opin. Investig. Drugs* 3:1089-1091; Senter, P.D. and Springer, C.J. (2001) *Adv. Drug Deliv. Rev.* 53:247-264.

In other embodiments, the second agent is a detectable moiety, which can be any molecule that can be quantitatively or qualitatively observed or measured. Examples of detectable markers useful in the conjugated antibodies of this invention are radioisotopes, fluorescent dyes, or a member of a complementary binding pair, such as a member of any one of: and antigen/antibody (other than an antibody to IFN- α), lectin/carbohydrate; avidin/biotin; receptor/ligand; or molecularly imprinted polymer/print molecule systems.

The second agent may also or alternatively be a polymer, intended to, e.g., increase the circulating half-life of the antibody. Exemplary polymers and methods to attach such polymers to peptides are illustrated in, e.g., U.S. Pat. Nos. 4,766,106; 4,179,337; 4,495,285; and 4,609,546. Additional illustrative polymers include polyoxyethylated polyols and polyethylene glycol (PEG) moieties. As used herein, the term "polyethylene glycol" is intended to

encompass any of the forms of PEG that have been used to derivatize other proteins, such as mono (C1-C10) alkoxy- or aryloxy-polyethylene glycol or polyethylene glycol-maleimide. For example, a full-length antibody or antibody fragment can be conjugated to one or more PEG molecules with a molecular weight of between about 1,000 and about 40,000, such as 5 between about 2000 and about 20,000, e.g., about 3,000-12,000. To pegylate an antibody or fragment thereof, the antibody or fragment typically is reacted with polyethylene glycol (PEG), such as a reactive ester or aldehyde derivative of PEG, under conditions in which one or more PEG groups become attached to the antibody or antibody fragment. Preferably, the pegylation is carried out via an acylation reaction or an alkylation reaction with a reactive 10 PEG molecule (or an analogous reactive water-soluble polymer). In certain embodiments, the antibody to be pegylated is an aglycosylated antibody. Methods for pegylating proteins are known in the art and can be applied to the antibodies of the invention. See for example, EP 0 154 316 by Nishimura *et al.* and EP 0 401 384 by Ishikawa *et al.*

Antibody characterization

15 After production or purification, or as part of a screening or selection procedure, the functional characteristics of an anti-IFN- α antibody of the invention can be investigated.

The following are brief descriptions of exemplary assays for antibody characterization. Some are further described in other sections and/or described in the Examples.

Binding Assays

20 The present invention provides for antibodies, and antigen-binding fragments and immunoconjugates thereof, that bind IFN- α . Any of a wide variety of assays can be used to assess binding of an antibody to IFN- α . Protocols based upon ELISAs, radioimmunoassays, Western blotting, BIACORE, and competition assays, *inter alia*, are suitable for use and are well known in the art.

25 For example, simple binding assays can be used, in which a test antibody is incubated in the presence of a target protein or epitope (e.g., an IFN- α protein subtype selected from A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, WA, 1 and D, a portion thereof, or a combination of any thereof), unbound antibodies are washed off, and the presence of bound antibodies is assessed using, e.g., RIA, ELISA, etc. Such methods are well known to those of skill in the 30 art. Any amount of binding above the amount seen with a control, non-specific antibody indicates that the antibody binds specifically to the target.

In such assays, the ability of the test antibody to bind to human IFN- α can be compared with the ability of a (negative) control protein, e.g. an antibody raised against a structurally unrelated antigen, or a non-Ig peptide or protein, to bind to the same target. Antibod-

ies or fragments that bind to IFN- α using any suitable assay with 25%, 50%, 100%, 200%, 1000%, or higher increased affinity capacity ?relative to the control protein, are said to "specifically bind to" or "specifically interact with" the target, and are preferred for use in the therapeutic methods described below. The ability of a test antibody to affect the binding of a 5 (positive) control antibody against IFN- α , e.g. a humanized ACO-1 or ACO-2 antibody, may also be assessed.

In one aspect, the invention provides for humanized anti-IFN- α antibodies sharing biological characteristics and/or substantial VH and/or VL sequence identity with humanized ACO-1 or ACO-2 antibodies. One exemplary biological characteristic is the binding to the 10 ACO-1 or ACO-2 epitope, *i.e.*, the respective regions in the extracellular domain of certain IFN- α protein subtypes to which the ACO-1 and ACO-2 antibodies bind. To screen for antibodies that bind to the ACO-1 or ACO-2 epitope, a routine cross-blocking assay, such as that described in Antibodies, A Laboratory Manual, Cold Spring Harbor Laboratory, Ed Harlow and David Lane (1988), can be performed.

15 In an exemplary cross-blocking or competition assay, ACO-1 or ACO-2 (control) antibody and a test antibody are admixed (or pre-adsorbed) and applied to a sample containing IFN- α . In certain embodiments, one would pre-mix the control antibodies with varying amounts of the test antibody (e.g., 1:10 or 1:100) for a period of time prior to applying to the IFN- α -containing sample. In other embodiments, the control and varying amounts of test antibody 20 can simply be admixed during exposure to the antigen/target sample. As long as one can distinguish bound from free antibodies (e.g., by using separation or washing techniques to eliminate unbound antibodies) and the control antibody from test antibody (e.g., by using species- or isotype-specific secondary antibodies, by specifically labeling the control antibody with a detectable label, or by using physical methods such as mass spectrometry to distinguish 25 between different compounds) one will be able to determine if the test antibody reduces the binding of the control antibody to the antigen, indicating that the test antibody recognizes substantially the same epitope as the control. In this assay, the binding of the (labeled) control antibody in the presence of a completely irrelevant antibody is the control high value. The control low value is be obtained by incubating the labeled (positive) control antibody 30 with unlabeled control antibody, where competition would occur and reduce binding of the labeled antibody.

In a test assay, a significant reduction in labeled antibody reactivity in the presence of a test antibody is indicative of a test antibody that recognizes the same epitope, *i.e.*, one that "cross-reacts" with the labeled control antibody. Any test antibody or compound that reduces the binding of the labeled control to the antigen/target by at least 50% or more pref-

erably 70%, at any ratio of control:test antibody or compound between about 1:10 and about 1:100 is considered to be an antibody or compound that binds to substantially the same epitope or determinant as the control. Preferably, such test antibody or compound will reduce the binding of the control to the antigen/target by at least 90%. Nevertheless, any compound 5 or antibody that reduces the binding of a control antibody or compound to any measurable extent can be used in the present invention.

Biological Activity

Differentiation of Monocytes. The generation of activated T and B lymphocytes requires the recruitment and maturation of antigen presenting cells ("APCs"). These APCs 10 include B cells, monocytes/macrophages and dendritic cells. The serum of SLE patients contains IFN- α which can activate DCs and the activated activity can be blocked with humanized antibody preparations according to the invention. Methods to detect and quantitate this activity are described in the scientific and patent literature (see, e.g., paragraphs 0136 through 15 0150 of patent publication number US20040067232A1, relevant portions of which are hereby incorporated herein by reference).

Activation of the MxA promoter. The ability of IFN- α to activate the MxA promoter, and the ability of the anti-IFN- α monoclonal antibodies of the invention to block this activation can be measured using reporter gene (RG) assays where the MxA promoter is fused to a reporter gene, such as chioramphenicol acetyltransferase (CAT) or luciferase (luc), preferably luciferase. Assays for CAT and luciferase are known to those of skill in the art. Preferably, the activity of the MxA promoter is measured in A549 cells stably transformed with an MxA promoter/reporter gene fusion construct. A549 cells are a lung carcinoma cell line available through the ATCC (product number CC1-185). The MxA (a.k.a. Mxl) promoter can be human, mouse or rat. The sequence and structure of the human MxA promoter is disclosed 20 in Genbank Accession number X55639, Chang *et al.* (1991) Arch Virol. 117:1-15; and Ronni *et al.* (1998) J Interferon Cytokine Res. 18:773-781. Human MxA promoter/luciferase fusion constructs and luciferase assays are disclosed in patent publication US20040209800 and Rosmorduc *et al.* (1999) J of Gen Virol 80:1253-1262. Human MxA promoter/CAT fusion constructs and CAT assays are disclosed in Fernandez *et al.* (2003) J Gen Virol 84:2073-25 2082 and Fray *et al.* (2001) J Immunol Methods 249:235-244. The mouse MxA (Mxl) promoter is disclosed in Genbank accession number M21104; Hug *et al.* (1988) Mol Cell Biol 8:3065-3079; and Lleonart *et al.* (1990) Biotechnology 8:1263-1267. A mouse MxA promoter/luciferase fusion construct and a luciferase assay are disclosed in Canosi *et al.* (1996) J Immunol Methods 199:69-67.

Cytopathic effect inhibition (CPE) assays. CPE assays are based on the antiviral activity of interferon. In general, a suitable cell line is infected with a virus in the presence of interferon, and the inhibitory activity of interferon is quantified on viral propagation or replicative processes. The readout of the assay may be based on reduction of virus yield, reduction 5 of viral cytopathic effect, reduction of viral protein of RNA synthesis, reduction of viral plaque formation. The cytopathic assay may be used to determine the neutralizing effect of antibodies on the activity of interferon. Exemplary CPE assays are described in Meager, A. 1987. Quantification of interferons by anti-viral assays and their standardization. In : Clemens, M.J., Morris, A.G., Gearing, A.J.H. (Eds), Lymphokines and interferons: A Practical Approach. IRL, 10 Press, Oxford, p. 129 and Grossberg and Sedmak, 1984. Assays of interferons In: Billiau, A. (Ed) Interferon, vol.1: General and Applied Aspects. Elsevier, Amsterdam, p. 189, and in Example 6, paragraphs 157-164, and Figure 1 of PCT publication WO2006086586.

Pharmaceutical Formulations

Another object of the present invention is to provide a pharmaceutical formulation 15 comprising a [the protein] compound which is present in a concentration from 10-500 mg/ml, such as e.g. 20-300 mg/ml, preferably 30-100 mg/ml, and most preferably 50-100 mg/ml., and wherein said formulation has a pH from 2.0 to 10.0. The formulation may further comprise a buffer system, preservative(s), tonicity agent(s), chelating agent(s), stabilizers and surfactants. In one embodiment of the invention the pharmaceutical formulation is an aqueous 20 formulation, i.e. formulation comprising water. Such formulation is typically a solution or a suspension. In a further embodiment of the invention the pharmaceutical formulation is an aqueous solution. The term "aqueous formulation" is defined as a formulation comprising at least 50 %w/w water. Likewise, the term "aqueous solution" is defined as a solution comprising at least 50 %w/w water, and the term "aqueous suspension" is defined as a suspension 25 comprising at least 50 %w/w water.

In another embodiment the pharmaceutical formulation is a freeze-dried formulation, whereto the physician or the patient adds solvents and/or diluents prior to use.

In another embodiment the pharmaceutical formulation is a dried formulation (e.g. freeze-dried or spray-dried) ready for use without any prior dissolution.

30

Diagnostic applications

The IFN- α -antibodies of the invention also have non-therapeutic applications. For example, anti-IFN- α antibodies may also be useful in diagnostic assays for IFN- α protein, e.g. detecting its expression in specific cells, tissues, or serum. For example, anti-IFN- α an-

tibodies could be used in assays selecting patients for anti-IFN- α treatment. For such purposes, the anti-IFN- α antibodies could be used for analyzing for the presence of IFN- α in serum or tissue specimens. For diagnostic applications, the antibody typically will be labeled with a detectable moiety.

5 Therapeutic applications

Methods of treating a patient using a humanized anti-IFN- α antibody as described herein are also provided for by the present invention. In one embodiment, the invention provides for the use of a humanized antibody as described herein in the preparation of a pharmaceutical composition for administration to a human patient. Typically, the patient suffers 10 from, or is at risk for, an autoimmune or inflammatory disease or disorder associated with abnormal expression of at least one IFN- α subtype selected from the group consisting of subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b, and WA.

Exemplary conditions or disorders to be treated with the antibodies of the invention, include, but are not limited to lupus (e.g., systemic lupus erythematosis (SLE)), rheumatoid 15 arthritis, juvenile chronic arthritis, osteoarthritis, spondyloarthropathies, systemic sclerosis (scleroderma), idiopathic inflammatory myopathies (dermatomyositis, polymyositis), Sjogren's syndrome, vasculitis, systemic vasculitis, sarcoidosis, autoimmune hemolytic anemia (immune pancytopenia, paroxysmal nocturnal hemoglobinuria), autoimmune thrombocytopenia (idiopathic thrombocytopenic purpura, immune-mediated thrombocytopenia), thyroiditis (Grave's disease, Hashimoto's thyroiditis, juvenile lymphocytic thyroiditis, atrophic thyroiditis), diabetes mellitus, immune-mediated renal disease (glomerulonephritis, tubulointerstitial nephritis), demyelinating diseases of the central and peripheral nervous systems such 20 as multiple sclerosis, idiopathic demyelinating polyneuropathy or Guillain-Barre syndrome, and chronic inflammatory demyelinating polyneuropathy, hepatobiliary diseases such as infectious hepatitis (hepatitis A, B, C, D, E and other non-hepatotropic viruses), autoimmune 25 chronic active hepatitis, primary biliary cirrhosis, granulomatous hepatitis, and sclerosing cholangitis, inflammatory bowel disease (ulcerative colitis, Crohn's disease), celiac disease, gluten-sensitive enteropathy, and Whipple's disease, autoimmune or immune-mediated skin diseases including bullous skin diseases, erythema multiforme and contact dermatitis, psoriasis, allergic diseases such as asthma, allergic rhinitis, atopic dermatitis, food hypersensitivity 30 and urticaria, immunologic diseases of the lung such as eosinophilic pneumonias, idiopathic pulmonary fibrosis and hypersensitivity pneumonitis, and transplantation associated diseases including graft rejection and graft-versus-host-disease. In a specific embodiment, the disease, condition or disorder is selected from lupus, Sjogren's syndrome, psoriasis, dia-

betes mellitus, rheumatoid arthritis, and juvenile dermatomyotosis. In another specific embodiment, the disease, condition, or disorder is SLE. For example, in one aspect, the anti-IFN- α antibody is used in combination with one or more other anti-inflammatory agents, including, but not limited to, analgesic agents, immunosuppressive agents, corticosteroids, and 5 anti-TNF α agents or other anti-cytokine or anti-cytokine receptor agents, and anti-angiogenic agents. Specific examples include methotrexate, TSG-6, Rituxan \circledR , and CTLA4-Fc fusion proteins. Further examples of combination therapies are provided below.

Articles of manufacture

In another embodiment of the invention, an article of manufacture containing materials useful for the treatment of the disorders described above is provided. For example, the 10 article of manufacture can comprise a container containing a humanized anti-IFN- α antibody as described herein together with instructions directing a user to treat a disorder such as an autoimmune or inflammatory disease or disorder in a human with the antibody in an effective amount. The article of manufacture typically comprises a container and a label or package 15 insert on or associated with the container. Suitable containers include, for example, bottles, vials, syringes, etc. The containers may be formed from a variety of materials such as glass or plastic. The container holds a composition that is effective for treating the condition and may have a sterile access port (for example, the container may be an intravenous solution bag or a vial having a stopper pierceable by a hypodermic injection needle). At least one active 20 agent in the composition is the humanized anti-IFN- α antibody herein, or an antigen-binding fragment or antibody derivative (e.g., an immunoconjugate) comprising such an antibody. The label or package insert indicates that the composition is used for treating the condition of choice, such as, e.g., SLE.

Moreover, the article of manufacture may comprise (a) a first container with a composition 25 contained therein, wherein the composition comprises the human or humanized antibody herein, and (b) a second container with a composition contained therein, wherein the composition comprises a therapeutic agent other than the human or humanized antibody. The article of manufacture in this embodiment of the invention may further comprise a package insert indicating that the first and second compositions can be used in combination to 30 treat an autoimmune or inflammatory disease or disorder. Such therapeutic agents may be any of the adjunct therapies described in the preceding section.. Alternatively, or additionally, the article of manufacture may further comprise a second (or third) container comprising a pharmaceutically acceptable buffer, such as bacteriostatic water for injection (BWFI), phosphate-buffered saline, Ringer's solution and dextrose solution. It may further include other

materials desirable from a commercial and user standpoint, including other buffers, diluents, filters, needles, and syringes.

In a first aspect, the present invention thus relates to a humanized antibody, or an antigen-binding fragment thereof, that specifically binds human interferon- α (IFN- α), which humanized antibody is a humanized version of murine antibody ACO-1 or ACO-2, or of a combination thereof, comprising fewer donor amino acid residues than the murine complementary determining regions (CDRs) according to Kabat.

In a second aspect, the present invention thus relates to a humanized antibody that specifically binds IFN- α , or an antigen-binding fragment thereof, wherein said antibody is capable of binding IFN- α subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b and WA, but not subtypes 1 or D, and wherein said antibody comprises fewer donor amino acid residues than the non-human CDRs according to Kabat.

According to one embodiment, the the CDR H2 donor residues comprise Kabat residues 50-59. In another embodiment, said antibody competes with and/or binds to the same epitope on an IFN- α subtype as ACO-1 and/or ACO-2 antibody.

According to a preferred embodiment, the antibody is an IgG4 subtype. In a preferred embodiment, the antibody comprises a CDR H2 sequence according to SEQ ID NO: 21.

In a third aspect, the present invention relates to a method for producing an antibody according to the invention, wherein said method comprises incubating a host cell encoding said antibody under appropriate conditions and subsequently isolating said antibody. The invention furthermore relates to antibodies obtained by or obtainable by such methods.

In a third aspect, the present invention relates to a composition comprising an antibody according to the invention. The invention furthermore relates to a process for the preparation of a composition according to the invention, wherein said method comprises mixing antibody or a fragment thereof with excipients. The invention furthermore relates to compositions obtained by or obtainable by such methods.

In a fifth aspect, the present invention relates to a method of preventing, managing, treating or ameliorating an IFN- α related IFNlammatory disease or disorder, said method comprising administering to a subject in need thereof a prophylactically or therapeutically effective amount of an antibody according to the invention.

Finally, the present invention relates to use of an antibody according to the invention for preparation of a medicament suitable for treatment of an inflammatory disease.

EXAMPLES

5 Further details of the invention are illustrated by the following non-limiting Examples.

Example 1 - Sequencing of murine ACO-1 and ACO-2 antibodies

This example describes sequencing and recombinant expression of the murine antibodies ACO-1 and ACO-2, described in WO20060086586, as well as BLAST searches on the ACO-2 VH and VL sequences.

10

Antibody cloning and sequencing

Total RNA was extracted from hybridomas (ACO-1.5.2 and ACO-2.2.1) using RNeasy (#634914) from Qiagen. cDNA was synthesized from 1 µg total RNA using SMART-RACE cDNA amplification kit from Clonetech. The reaction was run at 42 °C for 1.5h and the samples were diluted in 75 µl tricine-EDTA. PCR amplification of the target was carried out in 50 µl reactions using 5 µl of cDNA as template. The forward primer for both heavy and light chain was universal primer mix (UPM) that was included in the SMART RACE kit. The reverse primer sequence for ACO-1 heavy chain (HC) was designed as follows:

15 5'- CTGGGCCAGGTGCTGGAGG (SEQ ID NO:11) and, for ACO-1 light chain (LC)
20 5'-CTAACACTCATTCCCTGTTGAAGCTC (SEQ ID NO:12).

The reverse primer sequence for ACO-2 heavy chain (HC) was designed as follows:

25 5'- CTAGCTAGCTCATTACCCGGAGACCGGGAGATGG (SEQ ID NO:26) and, for
ACO-2 light chain (LC): 5'-GCTCTAACACTCATTCCCTGTTGAAGCTCTTG (SEQ ID NO:27).

25 The PCR reactions were carried out using Advantage HF PCR kit from Clonetech and the PCR program was run with a single denaturing step at 94 °C/2 min followed by 24 cycles as given: 94 °C/30 sec.; 55 °C/30 sec.; 72 °C/1.5 min. The final extension step was 72 °C/10 min. The PCR products were identified on a 1 % agarose gel containing ethidiumbromide. The PCR products were purified from the gel using GFX Purification kit from GE Healthcare followed by cloning into Zero Blunt TOPO PCR Cloning Kit (#K2875-40) and transformed into TOP10 *E. coli* cells from Invitrogen.

DNA was extracted from *E. coli* colonies using the miniprep kit (#27106) from Qiagen. Plasmids were sequenced at MWG Biotech, Martinsried, Germany using the sequencing primers M13 rev (-29) and M13 uni (-21). HC and LC were verified from the identified sequences by using VectorNTI. All procedures based on kits were performed according 5 to manufacturer directions.

From the ACO-1.5.2 hybridoma cells a single kappa LC and a single IgG2a HC were cloned, having the following nucleic acid and amino acid sequences.

ACO-1 VH sequence (SEQ ID NO:13 (signal peptide included)):

atgggatggagctatatcatgcttttggtagcaacagctacagatgtccactcccaggtccaaactgcagcagcctg
10 gggctgaactggtaagcctggggcttcagtgaagctgtcctgttaaggctctggctacaccctaccaactactggatgcactgg
gtgaaggcagaggcctggacaaggcctgagtggtggagagattaatccatgccacggctgtactatctacaatgaaaactca
agagcaaggccacactgactgttagacaaatccatcacagcctcatgcactcagcagcctgacatctgaggactctgcgg
tctatttctgtcaagagggggactgggacccgcctggttgtactgggccaaggactctggtactgtctgtca

ACO-1 VL sequence (SEQ ID NO:14 (signal peptide included)):

15 atggatttcaagtgcagatttcagcttcctgctaattcagtgtctcagtcataatgtccagaggacaattgttctcaccc
agtctccagcaatcatgtctgtctccctggggagaagggtcacctgacctgcgtgcggctcaagtgttagattccagctattgtac
tggttaccagcagaaggcaggatcctcccccactctggatttagcacatccaaacctggcttctggagtccctgctgcgttcagtg
ggcagtgggtctgggaccttactcttcacaatcagcagcatggaggctgaagatgctgccttatttctccatcagtggagta
gttaccatttcacgttccgtcggtggacaaaattggaaataaaacgg

ACO-1 VH (SEQ ID NO:1 (signal peptide excluded))

QVQLQQPGAEVKPGASVKLSCKASGYTFTNYWMHWVKQRPGQGLEWIGEINPS
HGRTIYNENFKSKATLTVDKSSITAFMQLSSLTSEDSAVYFCARGGLGPAWFAYWGQGTLV
TVSA

ACO-1 VL (SEQ ID NO:4 (signal peptide excluded))

25 QIVLTQSPAAMSASPGEKVLTCAGSSVDSSYLYWYQQKPGSSPKLWIYSTSNLA
SGVPARFSGSGSGTYSLSITISSMEAEDAASYFCHQWSSYPFTFGSGTKLEIKR

From the ACO-2.2.1 hybridoma cells a single kappa type ACO-2 light chain and a single IgG2b ACO-2 heavy chain were cloned, with the following nucleic acid and amino acid sequences.

ACO-2 VH sequence (SEQ ID 28 (signal peptide included))

30 atgggatggagctatatcatccttttggtagcagcagctacagatgtccactcccaggtccaaactgcagcagcctg
gggctgaactggtaagcctggggcttcagtgaagctgtcctgtcaaggcctggctacagcttaccagctactggatgcactg
gtgtgaaggcagaggcctggacaaggcctgagtggtggagagattaatccatgccacggctgtactagctacaatgagaactt
caagagcaaggccacactgactgttagacaaatccatcacatgcactcagcagcctgacatctgaggactctgc
35 ggtctattactgttaagagggggactgggacccgcctggttgtactgggccaaggactctggtactgtctgtca

ACO-2 VL sequence (SEQ ID NO:29 (signal peptide included))

atggatttcaagtgcagatttcagttccctgctaatcagtgtctcagtcataatgtccagaggacaaattgttctcaccc
agtctccagcaatcatgtctgcattccctggggagaaggcacctgaccgcacgtgcaggccggctcaagtgttaggtccagctacttt
ctggtaccagcagaagccaggatctcccccactctggatttatggcacatccaacctggctctggagtcctgctcgcttcag
5 tggcagtggtctggacacttactctcacaatcagcagcatggaggctgaagatgcgccttattctgccatcagtgag
agttatccattcacgttcggctgggacaaaattggaaataaaacgg

ACO-2 VH sequence (SEQ ID NO:7 (signal peptide excluded)):

QVQLQQPGAEVKPGASVKLSCKASGYSFTSYWMHWVKQRPGQGLEWIGEINPS

10 HGRTSYNENFKSKATLTVDKSSNIVYMQQLSSLTSEDSAVYYCVRGGLGPAWFAYWGQQTL
VTVSV

ACO-2 VL sequence (SEQ ID NO:9 (signal peptide excluded)):

QIVLTQSPAAMSASPGEKVTLTCSAGSSVGSSYFYWYQQKPGSSPKLWIYGTSNLA
SGVPARFSGSGSGTSYSLTISSMEAEDAASYFCHQWSSYPFTFGSGTKLEIKR

15 The ACO-2 CDR sequences according to the Kabat definitions were found to be as
follows.

CDR-H1: SYWMH (SEQ ID NO:22)

CDR-H2: EINPSHGRTSYNENFKS (SEQ ID NO:23)

CDR-H3: GGLGPAWFAY (SEQ ID NO:17)

20 CDR-L1: SAGSSVGSSYFY (SEQ ID NO:24)

CDR-L2: GTSNLAS (SEQ ID NO:25)

CDR-L3: HQWSSYPFT (SEQ ID NO:20)

Example 2 – Design of humanized ACO-1 and identification of potential back-mutation residues

25 Identification and characterization of mouse ACO-1 CDRs

The ACO-1 CDR sequences according to the Kabat definitions were found to be as
follows.

CDR-H1: NYWMH (SEQ ID NO:15)

CDR-H2: EINPSHGRTIYNENFKS (SEQ ID NO:16)

30 CDR-H3: GGLGPAWFAY (SEQ ID NO:17)

CDR-L1: SAGSSVDSSYLY (SEQ ID NO:18)

CDR-L2: STSNLAS (SEQ ID NO:19)

CDR-L3: HQWSSYPFT (SEQ ID NO:20)

Identification of human germline

A 3D protein structure model was built using MOE (Molecular Operating Environment; available at www.chemcomp.com) with a structural template from the Protein Data Bank (PDB): 1Z3G. The PDB is described in Berman et al. (Nucl Acids Res 2000;28:235-242), and is available at www.rcsb.org/pdb. Based on an analysis of antibody-antigen complexes in the PDB database, the most probable residues in the paratope were found to be residues 23-35, 49-58, 93-102 of the ACO-1 VH, and 24-34, 49-56, 89-97 of the ACO-1 VL. Using MOE, residues interacting (hydrophobic, hydrogen binding, charge interaction) with the paratope were identified and the combined set of residues (paratope + interacting residues) were taken as the so-called mask of ACO-1 shown in Figure 1.

Searching the germline V databases with the ACO-1 VH and ACO-1 VL returned the following potential framework templates (E-value given in parenthesis):

VH: VH1_46 (3e-038) VH1_f (6e-037), VH1_02 (6e-037), VH1_03 (1e-036), VH1_24 (2e-034),

VL: VKIII_L6 (9e-035), VKIII_A11 (4e-034), VKIII_A27 (8e-034), VKIII_L25 (1e-033), VKI_L8 (1e-033).

Searching the germline databases with the mask returned the following potential framework templates (E-value given in parenthesis):

VH: VH1_46 (3e-011) VH1_02 (6e-011), VH1_f (1e-010), VH5_a (4e-010), VH1_03 (4e-010),

VL: VKIII_A11 (5e-009), VKIII_L6 (7e-009), VKIII_A27 (9e-009), VKIII_L25 (3e-008), VKI_L9 (6e-008).

After manual inspections of the alignments and the hits, VH1_46 and VKIII_L6 were selected as the human scaffolds. Other templates could be chosen to alter or optimize, e.g., the physical-chemical properties of the humanized antibody. JH4 and JK2 were selected as germline J-segments (SEQ ID NO:2 and 5, respectively).

Design of optimal humanized ACO-1

Humanization was performed with the following rules:

- Residues outside the mask were taken as human.
- Residues inside the mask and inside the Kabat CDR were taken as murine.
- Residues inside the mask and outside the Kabat CDR with mouse/germline consensus were taken as the consensus sequence.

- Residues inside the mask and outside the Kabat CDR with mouse/germline were taken as the germline sequence, but the murine difference were subject to potential back mutations.

5 The CDRs of the optimal hzACO-1 antibody obtained were (according to the Kabat definitions):

	CDR_H1	NYWMH	(SEQ ID NO:15)
	CDR_H2	EINPSHGRTIY AQKFQG	(SEQ ID NO:21)
	CDR_H3	GGLGPAWFAY	(SEQ ID NO:17)
10	CDR_L1	SAGSSVDSSYLY	(SEQ ID NO:18)
	CDR_L2	STSNLAS	(SEQ ID NO:19)
	CDR_L3	HQWSSYPFT	(SEQ ID NO:20)

Using the above humanization method, designing a mask of residues predicted to 15 constitute the paratope based on a 3D model of hzACO-1 and IFN- α A, in contrast to simple CDR grafting, a hzACO-1 antibody with fewer murine residues was obtained, since the peptide comprising the 5 C-terminal amino acids of the optimized hzACO-1 CDR H2 sequence (highlighted in bold above) was identical to the corresponding human framework sequence, while the corresponding peptide in the ACO-1 CDR H2 sequence according to the Kabat 20 definitions was of murine origin. Additionally, the CDR H1 sequence for a humanized ACO-1 antibody identified in the present analysis was shorter than the one described in WO2006/086586. The optimized hzACO-1 antibody, or an antibody or antigen-binding fragment comprising at least a portion of the hzACO-1 VH sequence, can thus provide for a reduced risk for a human-anti-mouse-antibody (HAMA) -response in a human patient.

25 In addition, the replacement of the sequence AQK instead of NEN in position 60-62 in heavy chain has the advantage of avoiding two asparagine residues which may be prone to deamination.

Identification of potential backmutations.

30 The analysis of ACO-1 VH and VL sequences is illustrated in Figure 1. In Figure 1, the resulting humanized ACO-1 (hzACO-1) VH (SEQ ID NO:3) and VL (SEQ ID NO:6) sequences are shown with potential back-mutation residues as human, *i.e.*, without any back-mutations. The following back-mutation variants in the framework regions were identified for obtaining one or more optimized hzACO-1 antibodies, which are often required in order to 35 retain the affinity of the original mouse antibody:

- hzACO-1 VH: wild-type (i.e., no back-mutation), V5Q, M69L, R71V, T73K, S76I, V78A and any combination thereof;
- hzACO-1 VL: wild-type, E1Q, L47W, I58V, F71Y and any combination of any thereof;

5 - in various heavy-light chain combinations.

Example 3 – Design of ACO-2-based variants of hzACO-1 for affinity maturation of hzACO-1

As shown in Figure 2, amino acid sequence alignments of the ACO-1 and ACO-2 VH and VL sequences revealed a high sequence identity between the respective light and 10 heavy chains. Without being limited to theory, it is possible that the antibodies derived from the same precursor cell as they had the same V-D-J rearrangement and contained 3 identical mutations compared to the germline sequence. In addition, the antibodies differed at 13 amino acid residues, possibly due to subsequent somatic hyper-mutations.

Out of the 13 non-identical amino acid residues in the ACO-1 and ACO-2 VH and VL 15 domains, 9 residues were selected for mutational analysis in order to improve the affinity of the humanized ACO-1 antibody (Figure 3). Single additions of ACO-2 derived hypermutations could potentially identify deleterious and beneficial amino acid residues and by allowing the introduction of only the beneficial amino acids improve the affinity beyond the original 20 mouse ACO-1 and ACO-2 antibodies. The targeted residues were chosen based on their position within one of the light or heavy chain CDRs (according to the Kabat definition) or based on their location within regions outlined as potential antigen-interacting regions based on antigen-antibody 3D-modelling.

The following variants were identified for obtaining one or more optimized hzACO-1 25 antibodies:

- hzACO-1 VH: wild-type, T28S, N31S, I58S, S76N, T77I and A93V, and any combination of at least two mutations selected from T28S, N31S, I58S, S76N, T77I and A93V;
- hzACO-1 VL: wild-type, D29G, L33F, S50G and any combination of at least two mutations selected from D29G, L33F, and S50G,

30 - in various heavy-light chain combinations.

Residues were mutated separately from ACO-1 sequence to ACO-2 sequence within the hzACO-1 light and heavy chain constructs, in order to evaluate the individual contribution of each residue to antigen binding. A series of combination mutants was also generated.

Example 4 – Cloning of ACO-1, ACO-2, hzACO-1 and site-directed mutagenesis**ACO-1, ACO-2, and hzACO-1**

The VH and VL sequences were transferred to CMV promoter-based expression vectors (pTT vectors) suitable for transient expression in the HEK293-EBNA (HEK293-6E) expression system described by Durocher *et al.* (Nucleic Acids Res. 2002;30(2):E9). In addition to the CMV-promoter, the vectors contain a pMB1 origin, an EBV origin and the AmpR gene. The VH of ACO-1 and ACO-2 were cloned into the CMV-based vector containing the constant region for mouse IgG2a and IgG2b, respectively. The full length LC was transferred to the empty CMV-based vector for both ACO-1 and ACO-2. The DNA sequences for the variable regions of hzACO-1 were ordered from Geneart, Regensburg, Germany. The delivered sequence for the hzACO-1 VH was transferred to the expression vector containing the constant region for human IgG4(S241P) (containing a S241P mutation in the hinge region). The delivered hzACO-1 VL sequence was transferred to the vector containing the constant region for a human kappa light chain.

15 Generation of back-mutation variants of hzACO-1

The 10 potential back-mutations identified in Example 2 were introduced separately to hzACO-1 HC and LC constructs in order to gauge the individual contribution of each residue due to antigen binding. A few combinations were included as well. A variant of hzACO-1 with an extended CDR H2 (hzACO-1-Kabat CDRH2), equivalent to the Kabat definition of the murine CDR H2 (SEQ ID NO:16) was also generated by site-directed mutagenesis.

Site-directed mutagenesis was performed on the hzACO-1 LC and HC expression vectors. To generate single mutants the QuickChange® Site-Directed Mutagenesis kit from Stratagene (cat.# 200518) was used according to the manufacturer's protocol. Introduction of desired mutations was verified by sequencing plasmid DNA preparations (MWG Biotech, Matinsried, Germany) for each mutant.

The mutated LC constructs were combined with the hzACO-1 HC for expression, and mutated HC constructs were combined with the hzACO-1 LC for antibody expression.

Light Chain Variants with Back-Mutations:

hzACO-1-E1Q
hzACO-1-L47W
hzACO-1-I58V
hzACO-1-F71Y
hzACO-1-L47W,I58V

Heavy Chain Variants with Back-Mutations:

hzACO-1-V5Q
hzACO-1-M69L
hzACO-1-R71V
hzACO-1-T73K
5
hzACO-1-S76I
hzACO-1-V78A
hzACO-1-R71V,T73K
hzACO-1-M69L, R71V, T73K, S76I, V78A
hzACO-1-Kabat CDRH2

10

Generation of ACO-2-based variants of hzACO-1

Site-directed mutagenesis introducing ACO-2 specific residues into the hzACO-1 antibody in order to improve affinity, as described in example 3, was performed on the hzACO-1 LC and HC expression vectors. To generate single mutants the QuickChange® 15 Site-Directed Mutagenesis kit from Stratagene (cat.# 200518) was used according to the manufactures protocol. Combination mutants were generated using both the QuickChange® Site-Directed Mutagenesis and QuickChange® Multi Site-Directed Mutagenesis kits according to the manufactures protocols (cat.# 200513).

Introduction of desired mutations was verified by sequencing plasmid DNA preparations (MWG Biotech, Matinsried, Germany) for each mutant.

The mutated light chain constructs were combined with the hzACO-1 heavy chain for expression and mutated heavy chain constructs were combined with the hzACO-1 light chain constructs for antibody expression.

Light Chain Variants with ACO-2-Derived Mutations:

25 hzACO-1-D29G
hzACO-1-L33F
hzACO-1-S50G
hzACO-1-D29G,L33F,S50G

Heavy Chain Variants with ACO-2-Derived Mutations:

30 hzACO-1-T28S
hzACO-1-N31S
hzACO-1-I58S
hzACO-1-S76N
hzACO-1-T77I
35 hzACO-1-A93V

hzACO-1-N31S,I58S
hzACO-1-T28S,N31S,I58S,A93V
hzACO-1-S76N,T77I
hzACO-1-T28S,N31S,I58S,S76N,T77I,A93V
5 hzACO-1-T28S,A93V
hzACO-1-N31S,A93V
hzACO-1-T28S,N31S,A93V
hzACO-1-T28S,N31S

10 **Example 5 - Expression of recombinant ACO derived antibodies**

The ACO-1, ACO-2, hzACO-1, and hzACO-1 variants were expressed in transiently transfected HEK293 cells following a generic antibody expression protocol. The following describes the transfection protocol for suspension adapted HEK293 cells.

Cell maintenance: Suspension adapted HEK293 cells were grown in GIBCO®
15 FreeStyle™ 293 Expression medium (Invitrogen cat. #: 12338-026) supplemented with 25 µg/ml Geneticin® (Invitrogen cat. #: 10131-019), 0.1% v/v Pluronic® F-68 (Invitrogen cat. #: 12347-019) surfactant & 1% v/v Penicillin-Streptomycin (Optional) (Invitrogen cat. # 15140-122). Cells were maintained in Erlenmeyer shaker flasks at cell densities between 0.2-2 x 10⁶ cells/ml in an incubator shaker at 37°C, 8 % CO₂ and 125 rpm.
20 DNA Transfection: The cell density of cultures used for transfection was 0.8-1.5 x 10⁶ cells/ml. 0.5 µg light chain vector DNA + 0.5 µg heavy chain vector DNA were used per ml cell culture. 293fectin™ (Invitrogen cat. #: 12347-019) was used as transfection reagent at a concentration of 1µl reagent per µg transfected DNA. The 293fectin™ was diluted in 30x vol. Opti-MEM® (Invitrogen cat. #: 51985-034), mixed and left at room temperature (23-25 °C) for 5 min. The DNA was diluted in 30µl Opti-MEM® per µg total DNA, mixed and left at room temperature (23-25 °C) for 5 min. The DNA and transfection reagent dilutions were mixed 1:1 and left at room temperature (23-25 °C) for 25 min. The DNA-293fectin™ mix was added directly to the cell culture. The transfection cell culture was transferred to an incubator shaker at 37°C, 8 % CO₂ and 125 rpm. After 4-7 days, cell culture supernatants were harvested by centrifugation followed by filtration through a 0.22 µm PES filter (Corning cat. #: 431098). The antibodies were analyzed as supernatants or purified using standard protein A 25 purification techniques.
30

Expression level comparison of hzACO-1 and hzACO-1-Kabat CDRH2

Transient expression levels in HEK293-6E cells were compared for hzACO-1 and hzACO-1-Kabat CDRH2 to determine if the distal CDR H2 residues had any effect on the ability of the cells to expressed either of the two antibody variants.

HEK293-6E cells were transfected as described above with pTT-based expression vectors for hzACO-1 light chain and hzACO-1 or hzACO-1-Kabat CDRH2 heavy chains. The transfections was performed in triplicates. For each antibody variant, three cultures (25 ml) were transfected using a DNA-293Fectin master mix to minimize the influence from pipetting inaccuracy. The transfected cultures were incubated in a shaker incubator for 4 days as described above. At day 4, samples were extracted from the cultures for cell viability and cell density mesurements and the remaining cell culture supernatants were harvested by centrifugation. Quantitation analysis of antibody production was performed by Biolayer Interferometry directly on clarified cell culture supernatants using the FortéBio Octet system and protein A biosensors. Cell culture densities and viabilities were measured using a Cedex HiRes automated cell culture analyzer. Results are show in Table 4 below.

Table 4. Expression analysis

	Expression Yield (mg/L)	Standard deviation (Expression Yield)	Viable Cell Density (x 10E5 cell/ml)	Standard deviation (Viable Cell Density)	Cell viability (%)	Standard deviation (Cell viability)
hzACO-1	41	0.9	19	0.2	65	2.1
hzACO-1-Kabat CDRH2	22	0.7	18	0.5	65	3.2

The results in Table 4. unexpectedly show a significant difference in the transient expression levels for hzACO-1 (humanized IFN-alpha antibody according to the present invention) compared with hzACO-1-Kabat CDRH2 (humanized IFN-alpha antibody humanized using traditional procedures and hence full length Kabat sequences). No difference in cell viability or density was observed for the cell cultures transfected with either of the two hzACO-1 variants. The expression level for the hzACO-1 was approximately 2-fold higher compared to the expression level for hzACO-1-Kabat CDRH2 antibody variant

By grafting a shorter version of CDR H2 compared to the Kabat-defined CDRH2, expression levels of the antibody were surprisingly significantly increased.

Without being bound by theory, it may hypothesized that the human germline-derived residues in the hzACO-1 as compared to the hzACO-1-Kabat CDRH2 antibody vari-

ant, carrying the extended CDR H2 (SEQ ID:16) affect HC folding and potentially LC-HC interaction, results in an improved protein stability and thus expression yield.

Such an improved level of protein expression resulting from an improved protein stability observed in the transient expression levels will be reflected in stable CHO-based 5 production cell lines. Therefore, by grafting the short version of CDRH2 (SEQ ID:21) generation of a high-producing stable production cell line is thus possible.

Expression level comparison of IgG4, 1 and 2 variants of hzACO-1

Transient expression levels in HEK293-6E cells were compared for the lead 10 IgG4(S241P) variant of hzACO-1 and hzACO-1(IgG1) and hzACO-1(IgG2) to determine if heavy chain subclass switching had any effect on antibody expression levels.

The experiment was performed similarly to the expression assay described above, but with the following changes: The experiment was performed in duplicates in 5 ml cultures. The cultures were incubated in filter capped 50 ml falcon tubes in a shaker incubator at 37°C, 8 % CO₂ and 250 rpm.

15 Unpredictably, the average expression levels for hzACO-1(IgG1) and hzACO-1(IgG2) were approximately 65% of the expression level for hzACO-1 (IgG4). Based on the observed reduction in protein expression (~35%) we conclude that the combination of hzACO-1 variable domains and IgG4 constant domain is superior to combinations with other chain subclasses and the development of this molecule greatly facilitate generation of a high- 20 producing stable production cell line.

Example 6: Crystal structure of IFN- α 8 in complex with hzACO-1-Fab

The crystal structure of IFN- α 8 in complex with the Fab fragment of hzACO-1 was determined and refined to 3.3 Å resolution using X-ray crystallography (Figure 8).

25 *Materials and Methods*

IFN- α 8 (amino acid 1-166 of SEQ ID NO: 30, and hzACO-1 Fab (with the light chain sequence of SEQ ID NO: 32 and heavy chain sequence of residues 1-221 of SEQ ID NO: 31) were mixed, with a slight excess of IFN- α 8, and the complex was purified on a gel-filtration column. The protein complex hzACO-1-Fab/IFN- α 8 was put in a buffer of 25 mM 30 HEPES buffer, pH 7.5, + 25 mM NaCl and concentrated to 5 mg/ml. The complex was crystallized in 100 mM HEPES buffer, pH 7.5, 15 % PEG 10,000 and 15 % Ethylene glycol, the precipitant solution. Prior to diffraction data collection, the crystal was flash frozen in liquid N₂. The crystal was first transferred to a cryo solution which was a mix of 25 % v/v 99 % glycerol and 75 % of the precipitant solution. Diffraction data were collected at beamline

BLI911-3, MAX-Lab, Lund, Sweden. Diffraction data were indexed and integrated using the XDS program package (Kabsch, *J. Appl. Crystallogr.* 1993;26:795-800).

The three-dimensional structure was determined using the Molecular Replacement (MR) method using the PHASER program (Read, 2001, *Acta Crystallogr. Sect. D-Biol. Crystallogr.* 57, 1373-1382) of the CCP4 package (Baily, 1994, *Acta Crystallogr. Sect. D-Biol. Crystallogr.* 50, 760-763). The crystal structure of the hzACO-1 Fab, un-complexed, was earlier been determined to 1.52 Å resolution (R- and R-free 0.18 and 0.21, respectively), data not shown. Those 3D coordinates were subsequently used in the MR calculations for the hzACO-1/IFN- α 8 complex. The search models were divided into three parts: 1) the variable domain of hzACO-1 Fab), 2) the constant domain of the hzACO-1 Fab and 3) the PDB deposited IFNtau model, Protein Data Bank (Berman et al., 2000, *Nucleic Acids Res.* 28, 235-242) accession code 1B5L (RADHAKRISHNAN et al, 1999, *J.MOL.BIOL.v.286pp.151*), mutated by the COOT program to obtain the sequence of IFN- α 8, SEQ ID NO: 30. The final space group determination was made by the PHASER program. The highest scores were obtained for space group P4₁ with rotation function peaks, RZ's, of 10.7, 4.4 and 2.6 σ, respectively, translation peaks, TZ's, of 24.1, 26.4 and 8.0 σ, respectively, log-likelyhood gains, LLG's, of 383, 918 and 1134, respectively, and with no overlaps to symmetry related molecules.

Molecular replacement was followed by some adjustments to the model in the COOT molecular graphics program after which torsional simulated annealing up to 2000 K was applied twice, without refinement of individual temperature factors. Original R- and R-free values were 0.416 and 0.439, respectively, and final values after simulated annealing were 0.314 and 0.427, respectively. The model was then subject to manual intervention in the COOT program followed refinements using the REFMAC5 program (Murshudov et al., 1997, *Acta Crystallogr. Sect. D-Biol. Crystallogr.* 53, 240-255) resulting in R- and R-free values of 0.216 and 0.348, respectively. The model comprised residues 1-21, while residues 22-23 were refined as Ala residues, 28-101 and 114-164 of IFN- α 8, 1-215 of the hzACO-1 light chain and 1-219 of the hzACO-1 heavy chain.

The relatively large difference between R- and R-free seen in the refinement are due to the limited resolution of the data, 3.3 Å, and that there are substantial stretches of the IFN- α 8 X-ray model that are completely missing in the interpretation of the electron density map. The electron density maps clearly define the residues in the stabilised interface of IFN- α 8 to hzACO-1-Fab, while details of the IFN- α 8 X-ray structure model away from the antibody site are less well defined and therefore less accurately determined.

Results

The contacts were identified by the CONTACT computer program of the CCP4 suite using a cut-off distance of 4.0 Å between the Fab and IFN- α 8 molecules. The resulting epitope for human hzACO-1 was found to comprise the following residues of IFN- α 8 (SEQ ID

5 NO: 30): Ser 55, His 58, Glu 59, Gln 62, Gln 63, Asn 66, Glu 97, Leu 118, Arg 121, Lys 122, Phe 124, Gln 125, Arg 126, Thr 128, Leu 129, Thr 132). Residues of hzACO-1 involved in interactions with IFN- α 8, the paratope, included Ser 32, Tyr 33, Tyr 35, Tyr 50, Ser 51, Trp 92, Ser 93, Tyr 95 and Phe 97 of the hzACO-1 light (L) chain, (Numbering according to SEQ 10 ID NO: 32, not Kabat, and Thr 30, Asn 31, Tyr 32, Trp 33, His 35, Glu 50, Asn 52, Ser 54, His 55, Arg 57, Leu 101, Gly 102, Trp 105 of the heavy (H) chain, Table 9 (Numbering according to SEQ ID NO: 31, not Kabat).

SEQ ID NO: 30

>IFN_ α 8

15 CDLPQTHSLGNRRALILLAQMRRISPFSCLKDRHDFEFQEEFDDKQFQKAQAI SVLHEM
IQQTFNLFSTKDSSAALDETLLEDEFYIELDQQLNDLESCVMQEVGVIIESPLMYEDSILAV
RKYFQRITLYLTEKKYSSCAWEVVRAEIMRSFSLSINLQKRLKSKE

SEQ ID NO: 32

20 hzACO-1 LC

1 EIVLTQSPAT LSLSPGERAT LSCSAGSSVD SSYLYWYQQK PGQAPRLLIY
5 STSNLASGIP ARFSGSGSGT DFTLTISLLE PEDFAVYYCH QWSSYPFTFG
10 QGTKLEIKRT VAAPSVFIFP PSDEQLKSGT ASVVCLLNNF YPREAKVQWK
15 VDNALQSGNS QESVTEQDSK DSTYSLSSTL TLSKADYEKH KVYACEVTHQ
25 201 GLSSPVTKSF NRGEC

SEQ ID NO: 31

hzACO-1 Fab HC

30 1 QVQLVQSGAE VKKPGASVKV SCKASGYTFT NYWMHWVRQA PGQGLEWMGE
5 INPSHGRTIY AQKFQGRVTM TRDTSTSTVY MELSSLRSED TAVYYCARGG
10 LGPAWFAYWG QGTLTVSSA STKGPSVFPL APCSRSTSES TAALGCLVKD
15 YFPEPVTVSW NSGALTSGVH TFPAVLQSSG LYSLSSVVTV PSSSLGTKTY
20 201 TCNVDHKPSN TKVDKRVESK

35 As can be seen in figure 9 the hzACO-1 interaction epitope on IFN- α 8 overlaps, partly, the IFNAR1 binding epitope while the IFNAR2 binding epitope is distant from the hzACO-1 binding epitope. That suggests that the neutralization of IFN- α by hzACO-1 occurs by neutralization of IFN- α binding to IFNAR1, but not IFNAR2. Accordingly, it may be envi-

sioned that hzACO-1 binds the IFN- α /IFNAR2 complex but inhibits formation of the ternary receptor complex IFN- α /IFNAR1/IFNAR2 responsible for intracellular signaling.

Table 5. Parameters of the IFN- α 8: hzACO-1-Fab complex crystal used for structure determination

Space Group:	P4 ₁		
Cell Parameters [Å]:	a	b	c
	112.74	112.74	60.55
Molecular complexes/asymmetric unit:	1		

5

Table 6. X-ray data statistics from the program XSCALE of the XDS package

RESOLU- TION [Å]	NUMBER OF REFLECTIONS			COMPLE- TENESS	R-FACTOR		COMPARED	I/SIGMA	R-	Rmrgd-	S_norm/
	OBSERVED	UNIQUE	POSSIBLE		observed	expected			S_ano		
10.00	984	353	453	77.9%	4.4%	4.9%	984	18.84	5.4%	4.1%	-1%
6.00	4237	1461	1562	93.5%	6.8%	7.2%	4237	12.45	8.2%	7.8%	1%
5.00	3922	1343	1411	95.2%	9.7%	10.1%	3922	9.83	11.7%	11.1%	3%
4.00	8747	3063	3182	96.3%	11.6%	11.9%	8747	8.59	14.0%	13.7%	8%
3.50	8588	3106	3212	96.7%	24.8%	24.9%	8588	4.43	30.3%	31.3%	9%
3.45	1102	397	410	96.8%	36.9%	37.8%	1102	3.17	45.0%	44.4%	7%
3.40	1198	448	464	96.6%	41.8%	41.3%	1198	2.82	52.3%	48.7%	-12%
3.35	1271	468	495	94.5%	56.0%	51.5%	1271	2.33	69.9%	66.9%	1%
3.30	1317	486	502	96.8%	59.6%	51.2%	1317	2.14	74.9%	67.4%	-8%
total	31366	11125	11691	95.2%	14.1%	14.2%	31366	7.44	17.2%	20.2%	6%

Table 7. Statistics from the last refinement cycle of IFN- α 8:hzACO-1-Fab of the REFMAC program.

DATA USED IN REFINEMENT.	
RESOLUTION RANGE HIGH (ANGSTROMS)	3.30
RESOLUTION RANGE LOW (ANGSTROMS)	20.23
DATA CUTOFF (SIGMA(F))	NONE
COMPLETENESS FOR RANGE (%)	95.66
NUMBER OF REFLECTIONS	10571
FIT TO DATA USED IN REFINEMENT.	
CROSS-VALIDATION METHOD	THROUGHOUT
FREE R VALUE TEST SET SELECTION	RANDOM
R VALUE (WORKING + TEST SET)	0.22272
R VALUE (WORKING SET)	0.21646
FREE R VALUE	0.34845
FREE R VALUE TEST SET SIZE (%)	5.0

FREE R VALUE TEST SET COUNT	552		
FIT IN THE HIGHEST RESOLUTION BIN.			
TOTAL NUMBER OF BINS USED	20		
BIN RESOLUTION RANGE HIGH	3.300		
BIN RESOLUTION RANGE LOW	3.384		
REFLECTION IN BIN (WORKING SET)	761		
BIN COMPLETENESS (WORKING+TEST) (%)	96.06		
BIN R VALUE (WORKING SET)	0.290		
BIN FREE R VALUE SET COUNT	44		
BIN FREE R VALUE	0.446		
B VALUES.			
FROM WILSON PLOT (A**2)	NULL		
MEAN B VALUE (OVERALL, A**2)	52.314		
OVERALL ANISOTROPIC B VALUE.			
B11 (A**2)	-0.96		
B22 (A**2)	-0.96		
B33 (A**2)	1.92		
B12 (A**2)	0.00		
B13 (A**2)	0.00		
B23 (A**2)	0.00		
ESTIMATED OVERALL COORDINATE ERROR.			
ESU BASED ON R VALUE (A)	NULL		
ESU BASED ON FREE R VALUE (A)	0.776		
ESU BASED ON MAXIMUM LIKELIHOOD (A)	0.574		
ESU FOR B VALUES BASED ON MAXIMUM LIKELIHOOD (A**2)	33.433		
CORRELATION COEFFICIENTS.			
CORRELATION COEFFICIENT FO-FC	0.901		
CORRELATION COEFFICIENT FO-FC FREE	0.727		
RMS DEVIATIONS FROM IDEAL VALUES	COUNT	RMS	WEIGHT
BOND LENGTHS REFINED ATOMS (A)	4628	0.014	0.022
BOND ANGLES REFINED ATOMS (DEGREES)	6285	1.738	1.953
TORSION ANGLES, PERIOD 1 (DEGREES)	577	8.885	5.000
TORSION ANGLES, PERIOD 2 (DEGREES)	195	40.409	24.205
TORSION ANGLES, PERIOD 3 (DEGREES)	770	23.636	15.000
TORSION ANGLES, PERIOD 4 (DEGREES)	22	18.930	15.000
CHIRAL-CENTER RESTRAINTS (A**3)	704	0.110	0.200
GENERAL PLANES REFINED ATOMS (A)	3478	0.007	0.021

Table 8. IFN- α 8 – hzACO-1 Fab L chain interactions.

A cut-off of 4.0 Å was used. The contacts were identified by the CONTACT computer program of the CCP4 suite. In the last column "****" indicates a strong possibility for a hydrogen bond at this contact (distance < 3.3 Å) as calculated by CONTACT, " *" indicates a

weak possibility (distance > 3.3 Å). Blank indicates that the program considered there to be no possibility of a hydrogen bond.

IFN- α 8 Atoms	hzACO-1 Fab L Atoms	Distance (Å)
Leu 118I CB	Tyr 95L CE2	3.78
Leu 118I CG	Tyr 95L CE2	3.68
	Tyr 95L CD2	3.90
Leu 118I CD1	Tyr 95L CG	3.95
	Tyr 95L CE2	3.53
	Tyr 95L CD2	3.21
	Ser 93L O	3.70
	Tyr 95L N	3.96
	Phe 97L CE1	3.62
Leu 118I CD2	Trp 92L O	3.84
Arg 121I NH2	Tyr 95L OH	3.93 *
Lys 122I CA	Tyr 33L OH	3.38
Lys 122I CB	Tyr 33L OH	3.12
Lys 122I CG	Tyr 33L CE1	3.75
	Tyr 33L CZ	3.37
	Tyr 33L OH	2.93
Lys 122I CD	Tyr 33L CZ	3.74
	Tyr 33L OH	3.40
Lys 122I CE	Tyr 33L OH	3.33
Lys 122I C	Tyr 33L OH	3.70
Lys 122I O	Tyr 33L OH	3.41 *
Gln 125I CG	Trp 92L CH2	3.18
	Tyr 33L CE1	3.80
	Trp 92L CZ2	3.63
Gln 125I CD	Trp 92L CH2	3.70
	Tyr 35L OH	3.81
	Trp 92L CZ2	3.58
Gln 125I OE1	Tyr 35L OH	3.50 *
Gln 125I NE2	Ser 32L O	3.24 ***
	Ser 51L OG	3.37 *
	Tyr 33L CE1	3.94
	Tyr 35L OH	3.59 *
	Trp 92L CZ2	3.67
Arg 126I CD	Tyr 33L OH	3.94
Leu 129I CD1	Ser 51L CB	3.80
	Ser 51L OG	3.77
	Ser 32L OG	3.40
Leu 129I CD2	Ser 51L OG	3.90
Thr 132I OG1	Tyr 50L OH	3.90 *

5 **Table 9. IFN- α 8 – hzACO-1 Fab H chain interactions.**

A cut-off of 4.0 Å was used. The contacts were identified by the CONTACT computer program of the CCP4 suite. In the last column "****" indicates a strong possibility for a

hydrogen bond at this contact (distance < 3.3 Å) as calculated by CONTACT, " * " indicates a weak possibility (distance > 3.3 Å). Blank indicates that the program considered there to be no possibility of a hydrogen bond.

IFN- α 8 Atoms	hzACO-1 Fab H Atoms	Distance (Å)
Ser 55I CB	Arg 57H NH2	3.58
Ser 55I OG	Arg 57H NE	3.18 ***
	Arg 57H CZ	3.30
	Arg 57H NH2	2.80 ***
His 58I CG	His 55H CD2	3.73
His 58I CE1	Asn 52H ND2	3.94
	Ser 54H CB	3.90
	Ser 54H OG	3.79
His 58I NE2	His 55H CD2	3.53
	Ser 54H CB	3.60
	Ser 54H OG	3.20 ***
His 58I CD2	His 55H CD2	3.19
Glu 59I CD	Trp 33H NE1	3.90
Glu 59I OE2	Trp 33H CD1	3.74
	Trp 33H NE1	3.01 ***
Gln 62I CD	Asn 52H OD1	3.44
	Thr 30H O	3.71
Gln 62I OE1	Asn 52H CG	3.91
	Asn 52H OD1	3.02 ***
	Ser 54H CB	3.67
	Thr 30H C	3.69
	Thr 30H O	2.55 ***
	Asn 31H CA	3.87
Gln 62I NE2	Asn 52H CG	3.77
	Asn 52H OD1	3.13 ***
	Asn 52H ND2	3.95 *
Gln 63I CD	Leu 101H CD1	3.83
Gln 63I OE1	Leu 101H CG	3.87
	Leu 101H CD1	2.96
Gln 63I NE2	Leu 101H CD2	3.79
Asn 66I CG	Asn 31H O	3.94
Asn 66I OD1	Tyr 32H CE1	3.54
	Asn 31H CB	3.89
	Asn 31H C	3.86
	Asn 31H O	2.80 ***
	Tyr 32H CZ	3.82
Asn 66I ND2	Asn 31H OD1	3.79 *
Glu 97I CG	Ser 54H OG	3.85
Glu 97I CD	Ser 54H O	3.97
	Ser 54H OG	3.24
Glu 97I OE1	Ser 54H O	3.54 *
Glu 97I OE2	Ser 54H CB	3.88
	Ser 54H OG	2.56 ***

Arg	121I CZ	Glu	50H CD	3.73
		Glu	50H OE1	3.31
		Glu	50H OE2	3.30
Arg	121I NH1	His	35H CE1	3.89
		Glu	50H CD	3.78
		Glu	50H OE1	3.06 ***
		Glu	50H OE2	3.66 *
		Trp	105H CZ3	3.82
Arg	121I NH2	Glu	50H CD	3.08
		Glu	50H OE1	2.80 ***
		Glu	50H OE2	2.73 ***
		Trp	33H CG	3.66
		Trp	33H CE2	3.33
		Trp	33H CD2	3.51
		Trp	33H CZ2	3.88
		Trp	33H CD1	3.54
		Trp	33H NE1	3.35 *
Phe	124I CB	Leu	101H CD1	3.79
Gln	125I CD	Gly	102H N	3.69
Gln	125I OE1	Leu	101H CA	3.76
		Leu	101H CB	3.38
		Leu	101H C	3.57
		Gly	102H N	2.55 ***
		Gly	102H CA	3.24
Thr	128I OG1	Leu	101H CB	3.84

Example 7: Design of structure based mutations for affinity maturation of hzACO-1

If the epitope and paratope of an antibody/antigen complex is not known, a large number of possible amino acid residues may be prone for mutations in order to improve the affinity of an antibody and moreover, they can be converted into any of the 20 remaining amino acid residues to identify the optimal residue at the particular position. The CDR region holds approximately 54 residues available for mutations. Thus, in order to improve the affinity of the interaction $54 \times 20 = 1080$ analogues may be generated only within the CDRs. In addition, mutations outside the CDRs may be made in order to improve affinity.

However, when the structure of the antibody/antigen is known, a more limited number of qualified mutations that improve affinity may be predicted and analysed, based on structural predictions. Accordingly, based on the crystal structure of IFN- α 8 in complex with the Fab fragment of hzACO-1, as described in Example 6, three mutants that improve hzACO-1 binding to all IFN- α subtypes were identified. The 3 mutants are hzACO-1 HC T30R, hzACO-1 LC Y32E and the combined hzACO-1 Y32E, T30R (residue numbering according to Kabat). The identification of the mutants is described below.

LC Y32E: It can be seen that the electron density for residues Lys 122 of IFN- α 8, which is part of the binding epitope to hzACO-1, indicates a rather high mobility. Moreover, the atom O γ of residue Tyr 32 (Kabat notation) of the hzACO-1 light chain is directed towards the C β and C γ atoms of the Lys 122 side chain. That interaction is not an optimal residue-residue interaction. Mutating the light chain Tyr 32 to a negatively charged residue like Glu, or Asp, makes the possibility of forming a strong ionic bond between the antibody light chain Tyr 32 residue and the positively charged Lys 122 residue of IFN- α 8. For that reason Y was exchanged for E in order to improve the affinity of the hzACO-1 antibody.

HC T30R: For each hzACO-1 residue close to IFN- α 8 in the X-ray structure, the following properties were calculated: number of core side chain atoms (core), number of periphery sidechain atoms (peri), number of charged interactions (char), number of hydrogen bonds (hybo) and number of hydrophobic interactions (hyph) and given in **Error! Reference source not found.** below:

Table 10. Properties of amino acids of the hzACO-1 mAb

Light Chain						
Kabat	Res	core	peri	char	hybo	hyph
29	D	1	2	1		
31	S	2				1
32	Y	5	3			6

34	Y	3				2
49	Y	3	1		1	2
50	S	2				8
53	N	1	3			1
91	W	5	2			9
92	S		1			
93	S					4
94	Y	6	2			
96	F	2	2			
Heavy Chain						
28	T		3			
30	T		3			
31	N	4				1
32	Y	6	2			1
33	W	10				3
35	H	2	2			
50	E	4	1	1	1	
52	N	3	1		1	
52A	P		1			
53	S	2			2	
54	H	4	2			6
56	R	5	2	1	1	
58	I	2	2			2
64	Q		1			
71	R		1	1		
97	L	4				3
98	G				1	
99	P		2			
100A	W	2	3			1

The number of core and periphery atoms were calculated by overlaying the X-ray structure onto a 101 x 101 x 101 node grid and for each grid point calculating the number of atoms < 2.5 Å (N_{ex}) and atoms < 3.5 Å (N_{in}) from the node. Core nodes have N_{ex} > 0 covering atoms from both hzACO-1 and IFN-8. Periphery nodes have N_{ex} = 0 and N_{in} > 0 covering atoms from both hzACO-1 and IFN- α 8. Core sidechain atoms are now atoms < 2.5 Å from any node. Periphery sidechain atoms are now atoms \geq 2.5 Å & < 3.5 Å from any node.

5

Finally periphery residues are defined as residues with only periphery atoms and no interactions, so they can be modified to create binding. LC S92, HC T28, HC T30, HC P52, HC Q64 and HC P99 have been identified. Focusing on heavy chain non-prolines, it is seen by visual inspection that the mutation HC T28R would have a possible interaction with D90, HC 5 T30R would have a possible interaction with both D90 and E97 and Q64R would have a possible interaction with E114, but other similar mutations can also be applied. Since only E97 is conserved across all interferon alphas, HC T30R is expected to give the best binding with a similar profile as hzACO-1.

10 The specific mutation HC T30R was designed to establish a charge-charge interaction in the periphery of the binding site to improve the affinity of hzACO-1.

Furthermore, a double mutant containing both the LC Y32E and the HC T30R mutations, termed hzACO-1 Y32E, T30R was generated and analyzed to determine if the two mutations would have additive effects.

15 **Example 8 – Determination of the kinetic parameters for the interaction between hzACO-1, hzACO-1 variants and recombinant human IFN- α subtypes.**

Protein interactions can be monitored in real-time using surface plasmon resonance (SPR) analysis. In this study, SPR analysis was performed on Biacore 3000 and Biacore T100 instruments in order to characterize the anti-IFN α monoclonal antibody hzACO-1 and 20 variants thereof, with respect to affinity towards various subtypes of recombinant human Interferon alpha (IFN- α).

Affinity studies were performed using a direct binding procedure, with the respective monoclonal antibody covalently coupled via free amine groups to the carboxymethylated dextrane membrane (CM5) on the sensor chip surface. Recombinant IFN- α subtypes (PBL 25 Biomedical Laboratories, NJ, USA.) were injected in various concentrations, followed by a dissociation period with constant buffer flow over the sensor chip surface. Using this experimental design, the binding of IFN- α to the immobilized monoclonal antibody can be regarded as a 1:1 binding, with one IFN- α molecule binding to one antibody binding site. The kinetic parameters for the interaction can be calculated using a 1:1 interaction Langmuir fitting 30 model.

The purified monoclonal antibodies were immobilized in individual flow cells on a CM5 type sensor chip. Immobilizations were performed using a standard amine coupling procedure, aiming for an immobilization level of 1000 Resonance Units (RU).

35 HBS-EP pH 7.4 (10mM HEPES, 150mM NaCl, 3mM EDTA and 0.005% Polysorbate P20) was used as running buffer, and diluent for the recombinant IFN- α 's. Association (injec-

tion) was 4 min., followed by a 12 to 30 min. dissociation period. Flow rate was 50 μ l/min. Experiments were performed at 25°C. Detection in all flow cells simultaneously. Flow cell #1 contained no immobilized antibody, and was used for subtraction of background and bulk.

The kinetic parameters were calculated by global fitting of the data for a given antibody - antigen combination using a 1:1 langmuir binding model. Data was inspected for mass-transport limitations prior to calculation of the kinetic parameters.

Experiments were performed on Biacore 3000 and T100 instruments. Data was evaluated using Biaeval 4.1 and Biacore T100 evaluation software.

The kinetic parameters obtained are valid only in the buffer used, and with the recombinant form of the antigen.

Results

In order to generate a humanized ACO-1 antibody with retained affinity a number of humanized variants were generated as described in example 2, 3 and 7, using different strategies. The kinetic parameters for the interaction between recombinant human IFN- α subtypes and variants of hzACO-1 was obtained by SPR analysis. As seen in Table 11 eventhough hzACO-1, generated as described in example 2 contains a truncated CDR H2 and no backmutations, the affinity of the hzACO-1 has been retained as compared to the murine ACO-1, as shown by the KD of the hzACO-1 being within two-fold of the mouse antibody. Accordingly, no further backmutations from human to mouse ACO-1 in the framework regions were required for humanization, as identified and described in example 2. In addition, the IFN- α subtype profile of the hzACO-1 antibody had been retained, as shown in Table 2.

Table 11. Kinetic parameters for the interaction of recombinant IFN- α A with ACO-1 and hzACO-1 variants.

KD is the equilibrium dissociation constant, k_a is the association rate constant and k_d is the dissociation rate constant.

Anti-IFN- α mAb	KD (M)	k_a (1/Ms)	k_d (1/s)	KD mAb/KD hzACO-1
ACO-1	3.09E-09	1.24E+05	3.75E-04	0.60
hzACO-1	4.46E-09	1.24E+05	5.56E-04	1
hzACO-1-L33F	4.86E-09	1.89E+05	9.18E-04	1.17
hzACO-1-S50G	4.94E-09	1.98E+05	9.78E-04	1.19
hzACO-1-T28S	2.97E-09	1.58E+05	4.68E-04	0.63
hzACO-1-N31S	2.20E-09	1.16E+05	2.55E-04	0.61
hzACO-1-I58S	1.93E-08	1.93E+05	3.74E-03	4.09

Anti-IFN- α mAb	KD (M)	ka (1/Ms)	kd (1/s)	KD mAb/KD hzACO-1
hzACO-1-S76N	5.07E-09	1.16E+05	5.88E-04	1.09
hzACO-1-T77I	6.79E-09	9.80E+04	6.65E-04	1.46
hzACO-1-A93V	2.16E-09	1.08E+05	2.34E-04	0.60
hzACO-1-T28S,N31S	2.22E-09	1.56E+05	3.45E-04	0.77
hzACO-1-N31S,A93V	1.51E-09	1.59E+05	2.40E-04	0.52
hzACO-1-T28S,A93V	1.66E-09	1.61E+05	2.68E-04	0.69
hzACO-1-T28S,N31S,A93V	1.57E-09	1.85E+05	2.91E-04	0.65

The kinetic parameters for the interaction of hzACO-1 as compared to a humanized ACO-1 molecule generated by traditional CDR grafting and accordingly contains a large murine CDR H2 (designated hzACO-1-kabat CDRH2), with recombinant human IFN- α A are

5 listed in Table 12 . As shown, the affinity of the hzACO-1 molecule, humanized as described in example 2 and containing a shorter CDR H2 than the hzACO-1-kabat CDRH2 molecule, were equal, showing that the humanization process described in example 2 generates a humanized variant as good as that generated by simple CDR grafting, while having a more human sequence.

10

Table 12. Kinetic parameters for the interaction of recombinant IFN- α A with hzACO-1 and hzACO-1-kabat CDRH2.

KD is the equilibrium dissociation constant, ka is the association rate constant and kd is the dissociation rate constant.

15

mAb	KD (M)	ka (1/Ms)	kd (1/s)	KD mAb/KD hzACO-1
hzACO-1	1.9E+09	2.22E+05	4.15E-04	1
hzACO-1-kabat CDRH2	1.7E+09	2.05E+05	3.41E-04	0,9

Furthermore, in order to investigate if the hzACO-1 and the hzACO-1-kabat CDRH2 had comparable kinetic parameters of the binding to various subtypes of human IFN- α , a dissociation comparison experiment was performed on selected human IFN- α subtypes

20 (Table 13). This shows that the affinity of the hzACO-1 appears to be retained as compared to hzACO-1-kabat CDRH2 to all tested subtypes despite having a shorter mouse CDR H2 sequence.

25 **Table 13. Comparison of dissociation rate constants (kd) of the interaction between various subtypes of recombinant human IFN- α A with hzACO-1 and**

hzACO-1-kabat CRH2 respectively.

Subtype	mAb	hzACO-1	hzACO-1-kabat CDRH2	hzACO-1/hzACO-1- kabat CDRH2
		kd (1/s)	kd (1/s)	Ratio
IFN- α H2		4.13E-04	3.28E-04	1,26
IFN- α K		2.89E-04	3.41E-04	0,85
IFN- α 4b		2.17E-04	1.55E-04	1,40
IFN- α WA		2.91E-04	3.66E-04	0,80

5

In order to try to improve the affinity of hzACO-1 beyond that of the mouse ACO-1 antibody, the kinetic of the parameters between IFN- α A and different hzACO-1 variants containing mutations based on the ACO-2 sequence were measured (Table 11). In order to correlate the parameters obtained in the separate experiments performed, the KD value of each 10 individual antibody was normalized against that of hzACO-1 in the same experiment, and the relation of the KD value of the individual mAb to the KD of hzACO-1 in the same experiment is shown in the column “KD mAb vs. KD hzACO-1”.

The affinity determination further demonstrated KD values of all ACO2 derived hzACO-1 antibody variants (except hzACO-1-I58S) in the lower nM range. The minor variations in the KD values are predominantly related to differences in the kd. Introduction of the 15 single amino acid substitutions N31S, A93V, or T28S, and combinations thereof, in the hzACO-1 slightly increased the affinity to a level similar to that of ACO-1. The hzACO-1-I58S mutation has a pronounced negative effect on the kd value, demonstrating the importance of this particular amino acid for the stability of the hzACO-1/IFN α -A complex.

20 Based on the hzACO-1 Fab / IFN- α 8 crystal structure, a number of amino acid substitutions were introduced in hzACO-1 in order to increase the affinity even further (see example7). Of these, two single substitutions, Y32E of the light chain and T30R of the heavy chain, had significant positive effects on the affinity (Table 14) increasing the affinity approximately 2 and 6 fold respectively, against IFN- α A. Remarkably, by combining the two mutations 25 the hzACO-1 construct containing both the Y32E and T30R an approximately 10 fold increase in affinity was observed against IFN- α A (Table 15).

Table 14. Kinetic parameters for the interaction of recombinant IFN- α A with hzACO-1 and hzACO-1 variants respectively.

KD is the equilibrium dissociation constant, ka is the association rate constant and kd is the dissociation rate constant.

mAb	KD (M)	ka (1/Ms)	kd (1/s)	KD mAb/KD hzACO-1
hzACO-1	3.20E-09	1.39E+05	4.43E-04	1
hzACO-1 LC Y32E	1.54E-09	2.47E+05	3.81E-04	0.5
hzACO-1 HC T30R	5.40E-10	1.31E+05	7.08E-05	0.16

5 **Table 15. Kinetic parameters for the interaction of recombinant IFN- α A with hzACO-1 and hzACO-1 Y32E,T30R respectively.**

KD is the equilibrium dissociation constant, ka is the association rate constant and kd is the dissociation rate constant.

mAb	KD (M)	ka (1/Ms)	kd (1/s)	KD mAb/KD hzACO-1
hzACO-1	2.72E-09	1.78E+05	4.85E-04	1
hzACO-1 Y32E,T30R	2.97E-10	1.49E+05	4.43E-05	0.1

10 The kinetic data in **Table 16** illustrate, that a hzACO-1 Y32E,T30R construct generated based on a rational design approach, had retained its IFN- α subtype profile as it does not bind to IFN- α 1 but does bind to the remaining subtypes tested. In order to validate that the effect on the kinetic parameters caused by the Y32E,T30R mutations were reflected in the binding to various subtypes of human IFN- α , a dissociation comparison experiment was performed on selected human IFN- α subtypes (**Table 16**). Although the suggested mutations 15 of the double mutant hzACO-1 construct were based on the structure of the IFN- α 8, unpredictably the improved off-rates were observed for all tested human IFN- α subtypes varying between 6-64 fold.

15 **Table 16. Comparison of dissociation rate constants (kd) of the interaction between various subtypes of recombinant human IFN- α with hzACO-1 and hzACO-1 Y32E,T30R respectively.**

Subtype	hzACO-1	hzACO-1 Y32E,T30R	hzACO-1/hzACO-1 Y32E,T30R
	kd (1/s)	kd (1/s)	Ratio
IFN- α A	6.24E-04	6.85E-05	9,1
IFN- α 1	No binding	No binding	-
IFN- α 4b	1.12E-03	1.74E-05	64,4
IFN- α 1	1.55E-03	3.15E-05	49,2

IFN- α J1	4.54E-03	1.95E-04	23,3
IFN- α WA	2.47E-03	4.05E-04	6,1

Example 9 - Analysis of hzACO-1 constructs in in a CPE-assay

This example shows that hzACO-1 was able to inhibit the protective effect of all IFN- α subtypes tested, except for those of IFN- α 1 and IFN- α D, which were unaffected by hzACO-1.

5 Materials & Methods

This anti-IFN- α neutralization assay used is based upon the lytic effect of EMC virus on A549 cells. All IFN- α subtypes can inhibit the EMC virus replication in A549 cells, resulting in cell survival, which can be measured as cellular DNA staining. The neutralizing effect of an anti-IFN- α antibody on different IFN- α subtypes can be measured by diminished cellular DNA staining, corresponding to increased cell lysis.

The assay was performed in plates with 96 wells (Nunc, Cat. No. 167008), where each well contained a final volume of 200 μ L. All IFN- α preparations were from PBL Biomedical Laboratories, NJ, USA.

To each well, four solutions were added (IFN- α , hzACO-1, cells and virus) at a volume of 50 μ L each. All solutions were prepared in F12Kaihn's medium (Gibco, Cat.no.21127) with 10% FCS. The specific concentration of each IFN- α subtype (listed in Table 17 below) was derived from previous studies. The antibody concentrations used in the assay were selected based on existing data obtained from use of, e.g., murine antibodies ACO-1.5.2 and ACO-2.2.

20 Each IFN- α subtype was pre-incubated with hzACO-1 for 2 hours at 37°C, 5% CO₂. The anti-interferon antibody was diluted as shown in Table 17 below. After pre-incubation of antibody with IFN- α , 50 μ L of cell-solution (300000 cells/mL) were added to obtain 15000 cells/well. After 4.5 hours incubation at 37°C, 5% CO₂, 50 μ L EMC virus at a concentration of 10^{3.5} TCID₅₀ were added, followed by incubation for 48 hours at 37°C, 5% CO₂.

25 The supernatant was subsequently carefully removed, and 50 μ L crystal violet solution (0,5% crystal violet, 25% methanol) were added. After 15 min of incubation at room temperature, the wells were washed in water and dried overnight.

To the dried plates were then added 200 μ L/well of pure methanol for 15 min to extract the crystal violet from the cells. After extraction, 100 μ L of the supernatant were carefully transferred to a new 96-well plate (Nunc, Cat. No. 256510), and 100 μ L Milli-Q water added to each well. The plate was then measured in an ELISA reader at 590nm.

The raw data retrieved from the ELISA reader was corrected for methanol and plate background prior to analysis.

Table 17. CPE assay parameters.

IFN α Subtype	[IFN α] (pg/ μ L)*	[hzACO-1] (ng/mL)
IFN- α A	1,25x 10-1	2.500->0
IFN- α 2	3,125x 10-2	2.500->0
IFN- α F	6,25x 10-2	2.500->0
IFN- α K	6,25x 10-2	2.500->0
IFN- α WA	6,25x 10-2	2.500->0
IFN- α B2	2,5x 10-2	2.500->0
IFN- α H2	1,25x 10-1	2.500->0
IFN- α I	5,0x 10-2	2.500->0
IFN- α J1	1,0x 10-1	2.500->0
IFN- α 4a	2,5x 10-1	250->0
IFN- α C	2,5x 10-2	250->0
IFN- α G	2,5x 10-1	250->0
IFN- α 4b	1,25x 10-1	250->0
IFN- α D	3,75	50.000->0
IFN- α 1	1,0	50.000->0

5 Results and discussion

Six different controls were used on all plates in the study (cells, cells+antibody, cells+IFN- α , cells+antibody+IFN- α , cells+virus, and cells+IFN- α +virus) to ensure that the cytopathic effect observed in the assay was not caused by cytotoxicity of the antibody and/or IFN- α , nor that any lack of IFN- α -protection of the cells against the virus was causing the cytopathic effect. No significant cytotoxic effect was observed in the assays, and at no level of interferon was there a sign of a cytopathic effect in the controls.

As shown in Figure 4, this CPE-assay showed that the protective effect of almost all interferon subtypes could be inhibited by hzACO-1. However, the protective effects of IFN- α D and IFN- α 1 were not inhibited by hzACO-1, even at antibody concentrations of 50000 ng/mL. Accordingly, the specificity of the mouse ACO-1 was retained in the hzACO-1 construct.

Example 10 – Analysis of ACO-derived antibodies in a Reporter Gene (RG) Bioassay

A luciferase-based reporter gene assay was utilized to evaluate the ability of hzACO-1 antibody variants to neutralize the biological activity of recombinant IFN α subtypes.

Materials

5 Dulbecco's Modified Eagle's Medium "complete": DMEM incl. phenol red + 10 % FCS + 2 mM L-glutamine + penicillin + streptomycin + 2-me., Nunc 96-well optical bottom plate, black tissue culture treated, PBS including 1mM Ca²⁺ and 1 mM Mg²⁺, Steady-Glo Luciferase assay system (Promega).

10 The 93D7 cell line was derived by stable transfection of the A549 cell line (CLL-185, ATCC) with an IFN-inducible construct, harboring the Mx promotor driving a luciferase reporter gene. The Mx promotor consist of 1.6 kb BamHI fragment containing the murine MxA promotor and IFN response elements excised from pSP64-Mxp(PstI-Pvull)-r β glo (Leonart *et al.* (1990) Biotechnology 8: 1263-1267).

15 The IFN- α subtypes (all from PBL Biomedical Laboratories, NJ, USA) used are listed in Figure 5.

Methods

RG assays were performed in quadruplicate wells in opaque Nunc 96-well optical bottom plates. For every assay, a positive control IFN α was included as well as negative control wells containing non-stimulated cells and cells treated with antibody in the absence of 20 IFN α stimulation.

Specifically, adherent 93D7 cells were harvested from flasks by removing the culture media, washing once with PBS, and trypsinizing. Trypsinization was stopped using DMEM complete. Cells were counted and adjusted to 600,000/ml in complete DMEM.

25 Purified anti-IFN- α mAbs were pre-incubated with recombinant IFN subtypes for 1 h in a total volume of 100 μ l DMEM complete at 37 °C + 5% CO₂. Following antibody-IFN incubation, 50 μ l 93D7 cells were added and incubated for 5 h at 37 °C + 5% CO₂.

To assay for MxA-driven luciferase induction by recombinant IFN subspecies, the concentration of IFN was adjusted to contain the amount to be placed in each well in 100 μ l. 100 μ l of IFN was placed in quadruplicate wells, incubated for 1 h at 37 °C + 5% CO₂. Subsequently, 50 μ l cells were added and incubation continued for additionally 5 h.

30 To assay for inhibition of MxA-driven luciferase induction by recombinant IFN sub-species using purified mAbs, 50 μ l of a desired dilution of recombinant IFN was added per well. 50 μ l of antibody diluted in DMEM complete was then added to the wells and incubated

for 1 h at 37 °C + 5% CO₂. After this incubation, cells were added and incubation continued for additionally 5 h.

After the 5 h incubations, medium was carefully removed from the cells using a multichannel pipet. Next, an adhesive black blocker was affixed to the bottom of the 96 well plate. 100 µl PBS with Ca²⁺ and Mg²⁺ ions was added to each well. 100 µl reconstituted Steady-Glo reagent was added to each well, making sure that contents in each well were mixed thoroughly. The plates were sealed with a clear adhesive strip. Following 5 min. incubation at room temperature in the dark, luminescence was read in a Topcount luminescence counter (Perkin Elmer).

10 For calculation of the degree of inhibition exerted by the antibody to IFN induced (MxA driven) luciferase activity, counts were when comparing Ab inhibition of various IFN- α subtypes normalized to activity levels in the absence of antibody and this value was set to 100 %. For comparing variant form of antibody inhibition of single IFN- α s data are shown as raw luciferase counts. IFNs were initially titrated in the absence of antibody to determine the 15 EC50 and the IFN concentration at which plateau in the assay was reached. When testing for antibody inhibition, IFN was generally used at 80 % of maximum stimulation levels to ensure both a solid induction of luciferase as well as operating at a level below saturation in the assay. Prism (GraphPad Software, Inc., San Diego) software was used for calculations and data display.

20 Results and discussion

Humanized ACO-1 constructs were evaluated for their ability to inhibit various human IFN- α subtypes in the reporter gene assay. Figure 5 shows normalized data for the inhibition of 12 IFN- α subtypes by the hzACO-1 antibody. IFN- α stimulation in the absence of antibody was set to 100% whereas mock treated cells (receiving medium only) was set to 0 %. Data points are shown with standard error. Curves were calculated as best fit sigmoidal response curves using the Prism software. hzACO-1 was capable of inhibiting all tested sub-species of IFN- α s except for IFN- α D, and accordingly the specificity of the parent mouse ACO-1 antibody was retained during humanization in the hzACO-1 antibody. Inhibition was complete in that, at high antibody concentrations, IFN- α activity was reduced to background 25 levels. IC50 for the inhibition of the various IFN- α subtypes ranged in this study from 28 ng/ml to 314 ng/ml. IFN- α D could not be inhibited even at higher hzACO-1 concentrations.

30 Figure 10 shows a comparison of the mouse ACO-1 Ab to the humanized ACO-1 (hzACO-1) as well as two variants hereof in the RG assay. One variant is a humanized ACO-1 harboring the entire CDRH2 (designated hzACO-1-kabat CDRH2) whereas the hzACO-1 was constructed with a shorter CDRH2 as described in example 2. In addition the figure 35

shows another mutated hzACO-1 which has been optimized for interaction with IFN- α s (designated hzACO-1 Y32E, T30R) through rational design, as described in example 7. These four recombinant mAb variants were compared with respect to inhibition of five different representative IFN- α subtypes in the RG assay.

5 hzACO-1 displayed for all five IFN subtypes almost comparable IC50 values as mouse ACO-1, quantitatively the IC50s being less than twofold (see Table 18 for relative IC50 values) in accordance with the observed KDs reported in example 8.. The humanization was thus accomplished with an affinity loss less than twofold of the parent ACO-1 antibody for functional inhibition. In conclusion, the mAb humanization of ACO-1 has produced an antibody that has retained the affinity for IFN- α s. Accordingly, both the affinity and the potency was considered retained in of the hzACO-1 antibody as compared to the original mouse ACO-1, and no further backmutations were required.

10 15 As described in example 2, the humanization method of ACO-1 resulted in an antibody with relatively more human amino acids in the CDR H2 as compared to commonly humanization by simple CDR grafting.. Whereas this could be expected to lead to reduced potency in the humanized ACO-1 for neutralization of IFN- α subtypes, as shown in figure 10 and Table 18, surprisingly this is not the case, as the hzACO-1 and hzACO-1-kabat CDRH2 are equipotent for all the IFN- α subtypes tested. This is in agreement with the affinities reported for the two ACO-1 variants as described in example 8.

20 **Table 18. IC50 values for IFN- α subtypes inhibition by hzACO-1 variants.**

Normalized data. **IC50 values in were for each IFN- α species normalized to that of hzACO-1.** A value less than one therefore indicates a more potent antibody than hzACO-1 inhibition of IFN- α activity, whereas conversely values higher than one indicates inhibition with lesser potency.

mAb Variant	IC50 values relative to hzACO-1				
	IFN- α A	IFN- α B2	IFN- α F	IFN- α G	IFN- α J1
hzACO-1	1,00	1,00	1,00	1,00	1,00
ACO-1	0,60	0,47	0,82	0,56	0,58
hzACO-1 Y32E, T30R	0,03	0,02	0,05	0,06	0,02
hzACO-1-kabat CDRH2	0,95	0,88	0,95	0,78	1,15

25

To determine the potency of ACO-2 derived hzACO-1 variants, as described in example 3, these were compared to hzACO-1 using the RG assay. Comparisons were made using two IFN- α subspecies (IFN- α F and IFN- α A). Four different single amino acid substitutions were tested: T28S (i.e., at position 28,(according to Kabat) threonine was substituted 30 with serine), I58S, N31S, and A93V.

Whereas the I58S variant inhibited the IFN- action with reduced potency, and possibly also with reduced efficacy (IFN- α A), the T28S and N31S variants both displayed potencies similar to that of hzACO-1. The A93V substituted variant, however, showed increased potency for inhibition of IFN- effects as measured in the RG assay (Figure 6). Although differences in the effects of the substitutions could be detected between different IFN- α subtypes, the trend was the same for the two forms in all four cases.

Through rational design using the crystal structure in Example 6, a mutant was constructed having two amino acids changed, HC T30R, LC Y32E of hzACO-1 (designated ACO-1 Y32E, T30R), as described in example 7, to improve binding to IFN- α . Even though the mutations were based on the structure of IFN- α 8, as seen from figure 10 (A-E) surprisingly this mutant had increased potency for inhibition of all the tested IFN- α s. Furthermore, Table 18 shows that whereas the increment in potency is dependent on the specific subtype it is in the order of 16 fold and up to 50 fold improvement of potency.

It follows that the epitope information obtained from the crystal structure in Example 6 can be used to design other antibody variants with improved binding affinity to this epitope. It also follows that such humanized antibody variants according to the present invention are embraced by the scope of the present invention.

Example 11 – Protein characterization of humanized ACO-derived antibodies

This Example concerns the thermal stability of hzACO-1 and different variants.

20 **Materials**

Antibodies

hzACO-1 expressed as with human IgG1, IgG2 and IgG4 isotypes

hzACO-1-T28S

hzACO-1-N31S

25 hzACO-1-A93V

hzACO-1-T28S-N31S

The following is a list of the buffers (100 mM) and their pH values used in the study: citric acid/sodium citrate, pH 3.0, 3.5; sodium acetate, pH 4.0, 4.5, and 5.0; histidine, pH 6.0, 30 6.5; imidazole, pH 7.0; glycine-glycine pH 8.0, 9.0, 10.0.

The following is a list of additives and their concentrations used in the study: NaCl, 100 mM; sucrose 0.25 M, 0.50 M; phenol, 0.5 %; Tween 80, 0.01%; glycerol 10%.

Thermofluor stability measurements

Solutions of 10 μ l 400X SYPRO Orange protein gel stain 5000X Concentrate in DMSO (Invitrogen Molecular Probes), 25 μ l buffer and 10 μ l protein (10 μ M) were added to wells of a 96-well PCR-plate (Biorad). The plates were sealed with Microseal B Adhesive sealer MSB-1001 (Biorad) and heated in a MyiQ Single-Color Real-Time PCR Detection system (Biorad) from 25 to 95°C in increments of 0.5 °C. Fluorescence changes in the wells of the plate were monitored simultaneously with a charge-coupled device (CCD) camera. The wavelengths for excitation and emission were 490 and 575 nm, respectively. The midpoint temperature for protein unfolding transition was determined as the first derivative maximum of the fluorescence intensity as a function of temperature

The thermofluor graphs for hzACO-1 and variants showed the two expected temperature transitions for IgG proteins, reflecting the two main domains, Fc and Fab. The proteins showed a lower Tm at lower pH. Above pH 5.5 the transition midpoint was rather constant.

The effect of additives was the same for the different antibodies. Generally, sucrose at 0.5M had the most stabilizing effect, while addition of NaCl, phenol and Tween 80 seemed to have a destabilizing effect.

At lower pH (pH 3.5), a difference in stability between hzACO-1 and the different variants was observed (Figure 7A). The double mutant hzACO-1-T28S-N31S and single mutant hzACO-1-A93V showed the highest stability at this pH. This could be important in a purification process for a therapeutic antibody product, as virus inactivation steps often are carried out at pH 3.5-4.0. At higher pH (pH 4.5, 5.5; Figure 7B and 7C, respectively) this difference in stability decreased.

Solution stability study

Solutions of hzACO-1-IgG4, hzACO-1-IgG1 and hzACO-1-IgG2 in 15 mM Histidine pH 6.5, Sucrose 20 mg/ml and tween 80 0.01 % was incubated at 40°C and samples were withdrawn for analysis at initial and after 5 weeks. Distribution of intact protein, soluble aggregate and/or fragments of hzACO-1 was determined using size exclusion chromatography (SEC-HPLC). A high-performance liquid chromatography (HPLC) system model 1100 or 1200 liquid chromatography system (Agilent Technologies, Palo Alto, CA) was used with a BIOSEP SEC 3000 (Phenomenex) column at a flow rate of 0.8 mL/min using pH 7.2 and with phosphate-buffered saline (PBS) as the mobile phase. Protein was detected by monitoring the OD at 215 nm. The percentage of each peak in total protein area was calculated.

The amount of formed high molecular weight forms of the hzACO-1-isotypes is shown in Figure 11 and shows clearly that the IgG4 construct showed no aggregate formation during the incubation while both the IgG1 and IgG2 isotypes formed high molecular weight variants.

5 Furthermore, the hzACO-1-IgG1 variant showed a low molecular weight fragment after incubation. The amount of the fragment was 1.3 %.

Accordingly, from a stability point of view the hzACO-1 IgG4 is a more attractive therapeutic molecule than the corresponding IgG2 and IgG1 antibodies.

10 **Examples 12 - ADCC analysis.**

As described in example 6 the ACO-1 mAb and humanized versions hereof may block the activity of IFN- α by inhibiting binding of IFN- α to the type I interferon alpha receptor subunit 1 (IFNAR1). Accordingly, a humanized therapeutic ACO-1 mAb may bind to the cell surface making a complex consisting of the antibody, IFN- α and IFNAR2. This raises the risk 15 of ACO-1 inducing antibody dependent cellular cytotoxicity (ADCC).

To this end a ADCC experiment was performed to asses the ability of the hzACO-1 antibody expressed as an IgG4 subtype in the presence of IFN- α 2A.

Materials and methods.

20 Raji cells (human B cell line (ATCC # CCL-86)) were used as target cells. Raji cells were cultured in RPMI1640 supplemented with 10% fetal calf serum, 10 mM HEPES, 1 mM sodium pyrovate, 1 mM glutamine, 2,5 g/l glucose and 1% penicillin/streptamycin. Highly purified interferon- α 2A was used for the assay. The protein was tested in a reporter gene assay for biological activity prior to use. Rituxan® (Nomeco A/S, Denmark) was used as a positive 25 control for ADCC, when using Raji cells, which is a B cell line that expresses the B cell surface antigen CD-20.

Target cells were harvested and counted in a hemocytometer. 1.5×10^6 cells were transferred to a 15 mL tube and centrifuged. The supernatant was completely removed and the cell pellet was resuspended in $100 \mu\text{Ci}$ ^{51}Cr (Chromium-51) per 10^6 cells (volume were 30 adjusted according to decay table). During a 1 hour incubation period at 37°C with IFN- α the vial was tapped every 15 minutes. Subsequently the cells were washed twice in medium (RPMI1640, 10% FCS), and resuspended in 2 mL medium of assay medium. 5000 ^{51}Cr labelled cells were plated in a volume of 50 μL in 96 well plates (flat bottom). All samples were analyzed in triplicate Maximum – and minimum releases were determined in wells without 35 effector cells. Maximum release : 5000 target cells/well + 1% Triton X-100. Minimum release:

5000 target cells/well. The effector cells were purified from 'buffy coats' by Ficoll density centrifugation using standard techniques. Graded numbers of freshly isolated human PBMCs were added to each well. The following effector to target (E:T) cell ratios were used :10, 20, 40 and 80. In all experiments hzACO-1 was tested at saturating concentration of 10 µg/mL
5 (66 nM). IFN- α 2A was tested at 0.5, 2.5, 5, and 10 nM. The final assay volume was 200 µL. After 4 hours of incubation at 37°C 30 µL of the supernatant was transferred to a Luma-Plate™ and let to air dry over night. The radioactivity was determined in TopCount NXT (PerkinElmer, USA). Data were entered into the GraphPrism® program and the average counts per minute of triplicates and the corresponding standard deviation were calculated.

10

Results.

15 As seen in figure 12, no induction of ADCC above background (cells or cells with IFN- α) could be observed for the hzACO-1 molecule, expressed as an IgG4 in the presence or absence of IFN- α . In contrast, Rituxumab, which was used as a positive control did induce cell lysis at different ratios of effector and target cell ratios (E:T)

Example 13 – Complement binding ELISA assay.

20 As described in example 6 the ACO-1 mAb and humanized versions hereof may block the activity of IFN- α by inhibiting binding of IFN- α to the type I interferon alpha receptor subunit 1 (IFNAR1). Accordingly, a humanized therapeutic ACO-1 mAb may bind to the cell surface making a complex consisting the antibody, IFN- α and IFNAR2. This raises the risk for activating the complement system and inducing complement dependent cytotoxicity (CDC).

25 The purpose of the present complement binding study was to test whether the classical complement pathway is activated when hzACO-1 expressed as a human IgG4 isotype binds to and forms a complex with a corresponding epitope on hIFN- α . This is accomplished by using an ELISA, which measures the binding of antibodies to C4. Binding of C4 indicates that the C1s is changed and the complement cascade has started. Once the C4 has bound, the other complement components from the plasma will in turn be activated, bind, and enzymatically cleave the next components of the cascade.

Materials and methods.

30 Streptavidin-coated microtiter plates (236001, NUNC) were used as ELISA plates. Biotinylated hIFN- α 2A was used as antigen source and the plates were coated in 100 µl/well by 0.25 µg/ml protein diluted in washing buffer (10 mM Na3PO4 + 145 mM NaCl + 0.05 %

Tween 20). This hIFN- α concentration was shown to be the optimal coating concentration in an ELISA. The plates were incubated for 60 min. at RT and gentle shaking and then washed five times in washing buffer, leaving the buffer from the last wash in the plates for 30 min. to block possible residual binding sites on the plates. The buffer was discarded from the 5 plates and 100 μ l hzACO-1mAb diluted in washing buffer were added to the plates at 1 μ g/ml. The plates were incubated for 60 min. at RT and gentle shaking. The plates were washed five times in washing buffer. A polyclonal anti-IgG4 pAb was used as a positive control by crosslinking of the hzACO-1 IgG4 mAb. The anti-IgG4 pAb mAb was diluted in washing buffer and added to the plates in serial dilutions from 32 μ g/ml to 32ng/ml in 100 μ l/well. Two 10 different purifications of the anti-IgG4 pAb were used, one affinity purified antibody and one proteinA purified antibody. The plates were incubated for 60 min. at RT and gentle shaking. The plates were washed five times in washing buffer. Human plasma diluted 1:200 in plasma buffer (PBS w/ 0.3mM Ca²⁺, 1mM Mg²⁺) was added at 100 μ l/well. The plates were incubated for 60 min. at 37°C with gentle shaking. The plates were washed five times and mouse 15 anti-human C4 (HYB162-02 + HYB162-04, SSI), each diluted 1:2000 in washing buffer, was added to the plates at 100 μ l/well. The plates were incubated for 60 min at RT with gentle shaking. The plates were washed five times in washing buffer before addition of 100 μ l/well of HRP-rabbit anti-mouse IgG (DAKO P0260), diluted 1:1000 in washing buffer. The plates 20 were washed five times and all wells added 100 μ l of TMB substrate. After about 6 min. of incubation, 100 μ l of 4M H₃PO₄ was added to all wells to stop the enzyme reaction. The colour was measured spectrophotometrically by a Victor plate reader (Wallac).

Results.

hzACO-1 expressed as a human IgG4 isotype, was tested for the ability to bind complement components by ELISA using plates coated with IFN- α . This was visualised by 25 detecting binding to C4 which is one of the components in the classical complement cascade. As shown in figure 13, hzACO-1 IgG4 was unable to fix complement.. As a positive control a polyclonal anti-IgG4 pAb was used to induce binding by cross-linking of the hzACO-1 IgG4 antibody. If the hzACO-1 was cross bound with an anti-IgG4 pAb, a clear dose dependent binding of C4 to the anti IgG4 was detected.

30

Example 14 - Selection of Antibody Isotype.

When expressing a humanized monoclonal antibody, a human antibody isotype needs to be selected for expression of the full length human antibody. For the development of a neutralizing anti-IFN- α antibody, no Fc-mediated effector functions are required.

Furthermore, although the ACO-1 derived antibodies are capable of neutralizing IFN- α activity (example 9 and example 10), the binding epitope of the ACO-1 antibody variants, as described in example 6, is compatible with simultaneous binding of the IFNAR2 receptor subunit and IFN- α . Accordingly, although the therapeutic hzACO-1 mAb binds to the 5 soluble IFN- α cytokine, the mAb may in fact be able to bind to cell surfaces through the IFN- α /IFNAR2 complex and cause cytotoxicity by recruitment of Fc mediated effector functions. In addition to this, for antibodies against Type I IFNs (IFN- α and IFN- β) the selection of the antibody isotype is of particular importance, as it has been described that the Fc part of anti-IFN mAbs may in fact cause undesired biological effects resulting in potentiation instead of 10 neutralization of IFN in certain cell types as described below (Moll HP. et al J Immunol 180, 1594-604, 2008).

In the presence of Type I IFNs, antibodies against human IFN- α or IFN- β that normally neutralize IFN activity in other cell types, instead induce IFN activity in human endothelial cells and PMBCs in the presence of IFN- α or IFN- β . The antibodies used in these studies 15 are of the mouse IgG1 isotype, which are known to cross bind to human Fc receptors. The induction of IFN activity is only seen for intact antibodies and not Fab or Fab2 fragments of the same antibodies and appears to be inhibited by blockade of Fc receptors with an antibody isotype control mAb. Accordingly, although the molecular mechanism of this phenomenon is not known, it appears to require that these mAbs bind to cell surface receptors through 20 the Fc part. Furthermore, this phenomenon appears to be specific to Type I IFNs as antibodies against Type II IFN, IFN- γ , and a similar phenomenon is not observed for antibodies against the Type I IFN receptor, IFNAR (Moll HP. et al J Immunol 180, 1594-604, 2008).

In summary, for the development of a therapeutic anti-IFN mAb for neutralization of IFN- α Fc mediated effector function are not required and may in fact cause undesired biological effects by causing cytotoxicity or potentiating IFN activity in certain cell types in patients treated with an anti-IFN- α mAb. Consequently, generation of a humanized mAb that is 25 unable to induce effector functions may be crucial.-.

Four different human antibody isotypes exists i.e. IgG1, IgG2 IgG3 and IgG4. Of these IgG1 and IgG3 are the most efficient in binding to Fc receptors and complement and 30 consequently cause activation of Fc mediated effector functions such as -dependent cellular cytotoxicity (ADCC) and complement-dependent cytotoxicity (CDC) (Salfeld JG. Biotechnol 25, 1369-1372, 2007). Mutations that abrogate binding of various Fc receptors and C1q have been described in the literature (Hezareh M. et al J Virol 75, 12161-12168, 2001. Idusogie EE. Et al. J Immunol 164, 4178-4184, 2000. Lund J. et al J Immunol.;147:2657-6, 35 1991. & Chappel et al 1991, Canfield MS et al J Exp Med. 173:1483-91, 1991). However se-

lection of e.g. an IgG1 isotype and inclusion of several mutations will potentially be able to result in immunogenicity and cause an immune response against the Fc part in humans, as such a modified Fc region not will be naturally occurring in humans.

For that reason the hzACO-1 anti-IFN- α mAb was expressed both with IgG2 and IgG4 isotypes, which are both used in number of therapeutic antibodies (Salfeld JG. Biotechnol 25, 1369–1372, 2007). Unexpectedly, the expression levels of the IgG4 construct was significantly higher than of the IgG2 hzACO-1 variant (example 5) which will improve the production yield considerably. Furthermore, the IgG2 hzACO-1 molecule but not the IgG4 variant was prone to aggregation (example 11). This is considered a serious problem in drug development, as aggregation may lower yield during production, limit shelf-life and result in reduced potency in patients due to increased immunogenicity.

Accordingly, the hzACO-1 IgG4 construct was selected for further characterization in an ADCC assay in the presence of IFN- α , as well as a complement fixation ELISA assay, as described in example 12 and 13, respectively. These data confirm that the hzACO-1 IgG4 molecule is indeed unable to cause undesired cytotoxicity. The expected lack of potentiation of IFN- α activity may also be confirmed in PBMCs and ECs, as described in Moll et al 2008.

In summary, the hzACO-1 IgG4 molecule appears to be a particular suitable therapeutic molecule as it is capable of neutralizing IFN- α without induction of undesired side-effects caused by Fc-mediated effector functions including cytotoxicity and potentiation of IFN activity, and as it is a stable and well expressed molecule making it suitable for manufacturing and administration to patients.

Example 15 – Analysis of t-cell epitopes

Based on standard technologies for immunogenicity predictions (De Groot,A.S. and Moise,L. Curr. Opin. Drug Discov. Devel. 10, 332-340, 2007) the pocket profile method (Sturniolo,T. et al. Nat. Biotechnol. 17, 555-561,1999) as extended by ProPred (Singh,H. & Raghava,G.P. ProPred. BioINNormatics. 17, 1236-1237, 2001) was used to predict linear T-Cell epitopes among 51 HLA-DRB alleles. The method calculates the number of alleles a given 9 residue long peptide within the protein under evaluation can bind. If a given peptide that binds many alleles is of non-human origin, this can be used as an indirect measure of immunogenicity. If the peptide is of human origin, no immune response is expected due to the early negative selection of corresponding T-Cells.

It is the purpose of this example to compare the predicted immunogenicity of a humanization procedure using the full length hzACO-1-kabat CDRH2 to the actual humanization procedure applied for hzACO-1.

As input sequences to the ProPred algorithm, full length ACO-1 CDR_H2 with 10 + 10 residues added around hzACO-1-kabat CDRH2 is used:

APGQGLEWMG/EINPSHGRTIYNENFKS/RVTMTRDTST (CDR_H2_Full), together with the comparable hzACO-1 sequence

5 APGQGLEWMG/EINPSHGRTIYAQKFQG/RVTMTRDTST (CDR_H2_Human) in the same area.

Running CDR_H2_Full through the T-Cell epitope predictor gives the following 3 epitopes: WMGEINPSH (binding to 4% of HLA-DRB alleles), INPSHGRTI (6%) and FKSRTVTMTR (24%). For CDR_H2_Human the first two minor epitopes are identical, 10 whereas the last major epitope is converted to FQGRVTMTR (27%). Even though it is still a T-cell epitope, it is now a fully human sequence and from the assumption, that self reactive T-Cells are deleted by negative selection in the Thymus, this potential major epitope in CDR_H2_Full has been removed by the CDR_H2_Human sequence.

Accordingly hzACO-1 is expected to be less immunogenic, than a traditional CDR 15 grafted humanized ACO-1.

Example 16 – CDR truncation

As described in example 2 the mouse ACO-1 antibody was humanized by an untraditional method. This constitutes the designing of a mask of residues predicted to comprise 20 the paratope, based on a 3D model of hzACO-1 and IFN- α A. The application of this humanization method resulted in a hzACO-1 antibody with fewer murine residues than an antibody humanized by simple CDR grafting, since the peptide comprising the 5 C-terminal amino acids of the optimized hzACO-1 CDR H2 sequence was identical to the corresponding human framework sequence. In contrast, the corresponding peptide sequence in a traditionally CDR 25 grafted humanized antibody (the hzACO-1-kabat CDRH2) was of murine origin. Accordingly, this humanization procedure resulted in a humanized antibody with a more human sequence, less likely to cause immunogenicity in patients. Analysis of the sequences of the CDR H2 revealed that by reducing the mouse amino acid residues in the hzACO-1, an MHC class II T cell epitope containing the mouse amino acids is removed and replaced with a fully human, 30 which patients would be expected to be tolerant to. Accordingly, this confirmed that hzACO-1 is expected to be less immunogenic, than a traditional CDR grafted humanized ACO-1 antibody.

The affinity of the hzACO-1 antibody was retained, within two-fold of the mouse ACO-1 antibody, as shown in example 8. Accordingly, no further mouse backmutations were 35 required. Furthermore, the IFN- α subtype profile of the antibody, binding and neutralizing all

IFN- α subtypes except the IFN- α 1/D had been retained, as described in example 8, 9 and 10. Despite containing fewer mouse amino acids in the CDR H2, the affinity of the hzACO-1 was identical to the affinity and the potency of the hzACO-1-kabat CDRH2 antibody, containing the full length mouse CDR H2 from the mouse ACO-1 antibody (Example 8 and 9, respectively).

Furthermore, the replacement of the sequence AQK instead of NEN in position 60-62 in heavy chain by use of the described humanization method has the advantage of avoiding two asparagine that may be prone to deamidation. Deamidation change the net charge of proteins which may effect stability and/or specificity. By keeping the sequence AQK the homogeneity of hzACO-1 will be better preserved. Comparison of the hzACO-1 to the traditionally CDR grafted hzACO-1-kabat CDRH2 version, revealed that the hzACO-1 is a highly stable protein, whereas the hzACO-1-kabat CDRH2 antibody unexpectedly had a tendency to aggregate (example 11). Aggregation is considered a serious problem in drug development, as aggregation may result in lower yield during production, limited shelf-life and reduced potency in patients due to increased immunogenicity. In addition, the expression levels of the hzACO-1 construct was unexpectedly twice that of the hzACO-1-kabat CDRH2 variant (example 5).

In summary, the hzACO-1 IgG4 antibody was humanized by a novel approach resulting in a therapeutic antibody with less mouse amino acids which is less likely to cause immunogenicity in patients. Despite containing fewer amino acids from the original mouse mAb, this humanized antibody had a comparable affinity, potency and IFN- α subtype profile as the mouse antibody and a humanized version generated by traditional CDR grafting. Furthermore, the hzACO-1 antibody is less prone to deamidation and has a higher expression level. Thus, the hzACO-1 antibody it is a stable and well expressed molecule suitable for manufacturing and administration to patients.

CLAIMS

1. A humanized antibody, or an antigen-binding fragment thereof, that specifically binds human interferon- α (IFN- α), which humanized antibody is a humanized version of murine antibody ACO-1 or ACO-2, or of a combination thereof, comprising fewer donor amino acid residues than the murine complementary determining regions (CDRs) according to Kabat.
5
2. A humanized antibody that specifically binds IFN- α , or an antigen-binding fragment thereof, wherein said antibody is capable of binding IFN- α subtypes A, 2, B2, C, F, G, H2, I, J1, K, 4a, 4b and WA, but not subtypes 1 or D, and wherein said antibody comprises fewer donor amino acid residues than the non-human CDRs according to Kabat.
10
3. An antibody according to any one of claims 1 or 2, wherein the CDR H2 donor residues comprise Kabat residues 50-59.
15
4. An antibody according to any one of claims 1-3, wherein said antibody competes with and/or binds to the same epitope on an IFN- α subtype as ACO-1 and/or ACO-2 antibody.
20
5. An antibody according to any one of claims 1-4, wherein the antibody is an IgG4 subtype.
25
6. An antibody according to any one of claims 1-5, wherein said antibody comprises a CDR H2 sequence according to SEQ ID NO: 21.
7. A method for producing an antibody according to any one of claims 1-6, wherein said method comprises incubating a host cell encoding said antibody under appropriate conditions and subsequently isolating said antibody.
25
8. An antibody obtained by a method according to claim 7.
9. A composition comprising an antibody according to any one of claims 1-6 and 8.
10. A process for the preparation of a composition according to claim 9, wherein said method comprises mixing antibody or a fragment thereof with excipients.
11. A composition obtainable by a method according to claim 9.

12. A method of preventing, managing, treating or ameliorating an IFN- α related IFNlammatory disease or disorder, said method comprising administering to a subject in need thereof a prophylactically or therapeutically effective amount of an antibody according to any one of claims 1-6 and 8.
- 5 13. Use of an antibody according to any one of claims 1-6 and 8 for preparation of a medicament suitable for treatment of an inflammatory disease.

A

Heavy Chain

1	2	3	4	5	6
12345678901234567890123456789012345AB6789012345678901234567890					
QVQLQQPGAEVLVKPGASVKLSCKASGYTFTNYWMH	--WVKQRPGQGLEWIGEINP	--SHGRTIYN	ACO-1_VH		
QVQLVQSGAEVKKPGASVKVSCKASGYTFTSYMMH	--WVRQAPGQGLEWMGI	INP	--SGGSTSYA	VH1_46	
QVQLVQSGAEVKKPGASVKVSCKASGYTFTNYWMH	--WVRQAPGQGLEWMGEINP	--SHGRTIYA	hzACO-1_VH		
7	8	9	10	11	
1234567890123456789012345678901234567890ABCDEF	ABCDEF	GHIIJKL	1234567890	<- Kabat	
ENFKSKATLTVDKSSITAFMQLSSLTSEDSAVYFCARGGLGPA			WFAYWGQGTLTVSA	ACO-1_VH	
QKFQGRVTMTRDTSTSTVYMEMLSSLRSEDTAVYYCAR/			/YFDYWGQGTLTVSS	VH1_46/JH4	
QKFQGRVTMTRDTSTSTVYMEMLSSLRSEDTAVYYCARGGLGPA			WFAYWGQGTLTVSS	hzACO-1_VH	

B

Light Chain

1	2	3	4	5	6
12345678901234567890123456789012345678901234567890					
QIVLTQSPAIMSASPGEKVTLTCSAGSS	-----	VDSSYLYWYQQKPGSSPKLWIYSTSNLASGVPA			
EIVLTQSPATLSSLSPGERATLSCRASQ		SVSSYLAWYQQKPGQAPRLLIYDASN RATGIP			
EIVLTQSPATLSSLSPGERATLSCSAGSS	-----	VDSSYLYWYQQKPGQAPRLIYSTSNLASGPA	hzACO-1_VL		
7	8	9	10		
12345678901234567890123456789012345AB67890123456789	<- The Kabat Scheme				
RFSGSGSGTSYSLTISSMEAEDAASYFCHQWSSYP	FTFGSGTKLEIKR	ACO-1_VL			
RFSGSGSGTDFTLTISSLEPEDFAVYYCQQRSNWP	YTFGQGTLKLEIK	VKIII_L6/JK2			
RFSGSGSGTDFTLTISSLEPEDFAVYYCHQWSSYP	FTFGQGTLKLEIKR	hzACO-1_VL			

Fig. 1

A

Heavy Chain

1	2	3	4	5	6
12345678901234567890123456789012345AB67890123456789012ABC34567890					
QVQLQQPGAEVLVKGASVVLCKASGYTFTNYWMH	WVKQRPGQGLEWIGEINP	SHGRTIYN	ACO-1	VH	
QVQLQQPGAEVLVKGASVVLCKASGYSFTSYWMH	WVKQRPGQGLEWIGEINP	SHGRTSYN	ACO-2	VH	
QVQLQQPGAEVLVKGASVVLCKASGYTFTSYWMH	WVKQRPGQGLEWIGEINP	SINGRTN	YNN	J558.33	(germ-line)
7	8	9	10	11	
1234567890123456789012ABC345678901234567890ABCDEFGHJK1234567890					<- Kabat
ENFKSKATLTVDKSSITAFMQLSSLTSEDSAVYFCARGGLGPAWF				AYWGQGTLTVSA	ACO-1 VH
ENFKSKATLTVDKSSNIVYMQLSSLTSEDSAVYYCVRGGLGPAWF				AYWGQGTLTVSV	ACO-2 VH
EKFKSKATLTVDKSSSTAYMQLSSLTSEDSAVYCAR/ /AWF				AYWGQGTLTVVS	
J558.33/D?/JH3_1					

B

Light Chain

1	2	3	4	5	6
123456789012345678901234567ABCDEF890123456789012345678901234567890					
QIVLTQSPAIMSASPGEKVLTCAGSS	VDSSYLYWYQQKPGSSPKLWIYSTSNLASGVPA				ACO-1 VL
QIVLTQSPAIMSASPGEKVLTCAGSS	VGSSYFYWYQQKPGSSPKLWIYGTSNLASGVPA				ACO-2 VL
QIVLTQSPAIMSASPGEKVLTCASSS	VSSSYLYWYQQKPGSSPKLWIYSTSNLASGVPA				ae4 (germline)
7	8	9	10		
12345678901234567890123456789012345AB67890123456789					<- The Kabat Scheme
RFSGSGSGTYSLSLTISMEAEDAASYFCHQWSSYP	FTFGSGTKLEIKR				ACO-1 VL
RFSGSGSGTYSLSLTISMEAEDAASYFCHQWSSYP	FTFGSGTKLEIKR				ACO-2 VL
RFSGSGSGTYSLSLTISMEAEDAASYFCHQWSSYP //FTFGSGTKLEIKR					ae4/ JK4_1 (germline)

Black : germline sequence
 Grey : somatic hyper mutations

Fig. 2

A

Heavy Chain

1	2	3	4	5	6
1234567890123456789012345678901 12345AB67890123456789012ABC34567890					
QVQLQQPGAEVLVKPGASVVLCKASGYTFTNYWMH	WVKQRPQGLEWIGEINP	SHGRTEYN	ACO-1	VH	
QVQLQQPGAEVLVKPGASVVLCKASGYTFTSYWMH	WVKQRPQGLEWIGEINP	SHGRTEYN	ACO-2	VH	
QVQLVQSGAEVKKPGASVVKVSCKASGYTFTNYWMH	WVRQAPGQGLEWMGEINP	SHGRTIYA	hzACO-1	VH	
7	8	9	10	11	
1234567890123456789012ABC345678901234567890ABCDEFGHIJK1234567890					<- Kabat
ENFKSKATLTVVKSS ENFKSKATLTVVKSS	AFMQLSSLTSEDAVYFC AFMQLSSLTSEDAVYFC	RGGLGPAWF	AYWGQGTLTVSA	ACO-1	VH
ENFKSKATLTVVKSS ENFKSKATLTVVKSS	VYMQLSSLTSEDAVYYC VYMQLSSLTSEDAVYYC	RGGLGPAWF	AYWGQGTLTVSV	ACO-2	VH
QKFQGRVTMTRDTSTSTVYMEMLSSLRSEDTAVYYCARGGLGPAWF			AYWGQGTLTVSS	hzACO-1	VH

B

Light Chain

1	2	3	4	5	6
123456789012345678901 1234567ABCDEF890123456789012345678901234567890					
QIVLTQSPAAMSASPGEKVTLTCAGSS	V DSSYIY WYQQKPGSSPKLWI YTSNLASGVPA				ACO-1 VL
QIVLTQSPAAMSASPGEKVTLTCAGSS	V GSSYIY WYQQKPGSSPKLWI YTSNLASGVPA				ACO-2 VL
EIVLTQSPATLSPGERATLSCSAGSS	VDSSYLYWYQQKPGQAPRLLI YTSNLASGIPA				hzACO-1 VL
7	8	9	10		
1234567890123456789012345678901 12345AB67890123456789					<- The Kabat Scheme
RFSGSGSGTSYSLTISSMEAEDAASYFCHQWSSYP	FTFGSGTKLEIKR	ACO-1	VL		
RFSGSGSGTSYSLTISSMEAEDAASYFCHQWSSYP	FTFGSGTKLEIKR	ACO-2	VL		
RFSGSGSGTDFTLTISSLEPEDFAVYYCHQWSSYP	FTFGQGTKLEIKR	hzACO-1	VL		

Grey : residues mutated to ACO2 residues in hz-ACO1

Fig. 3

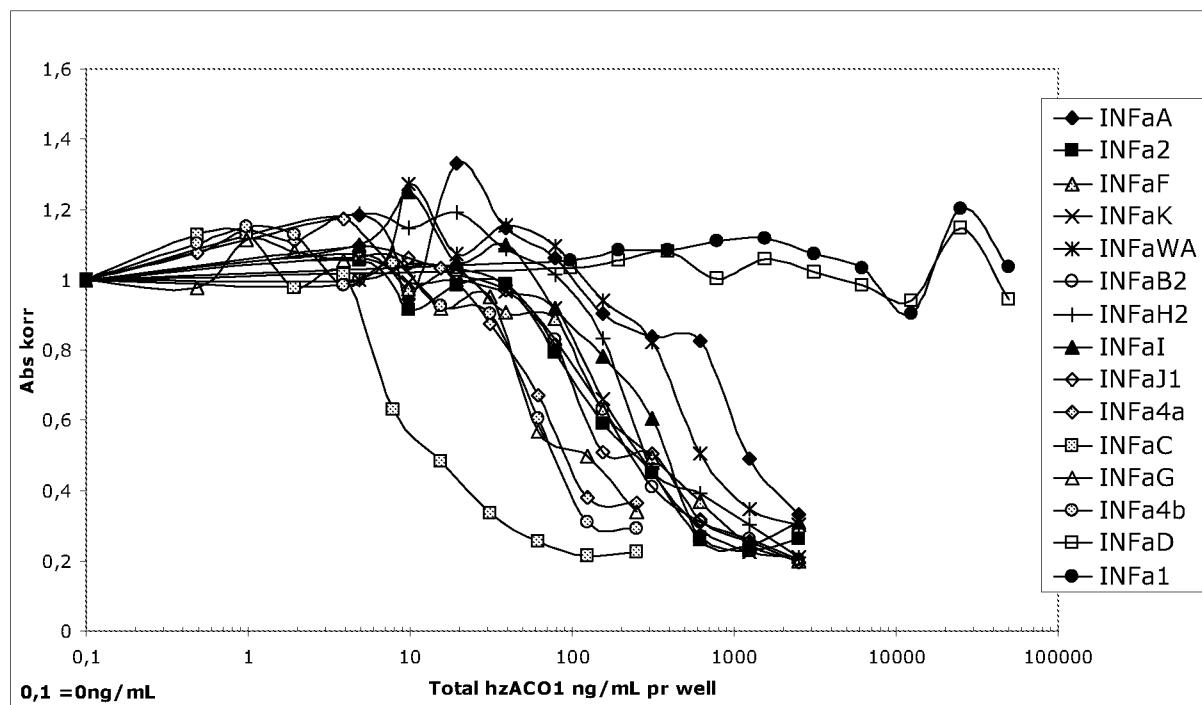


Fig. 4

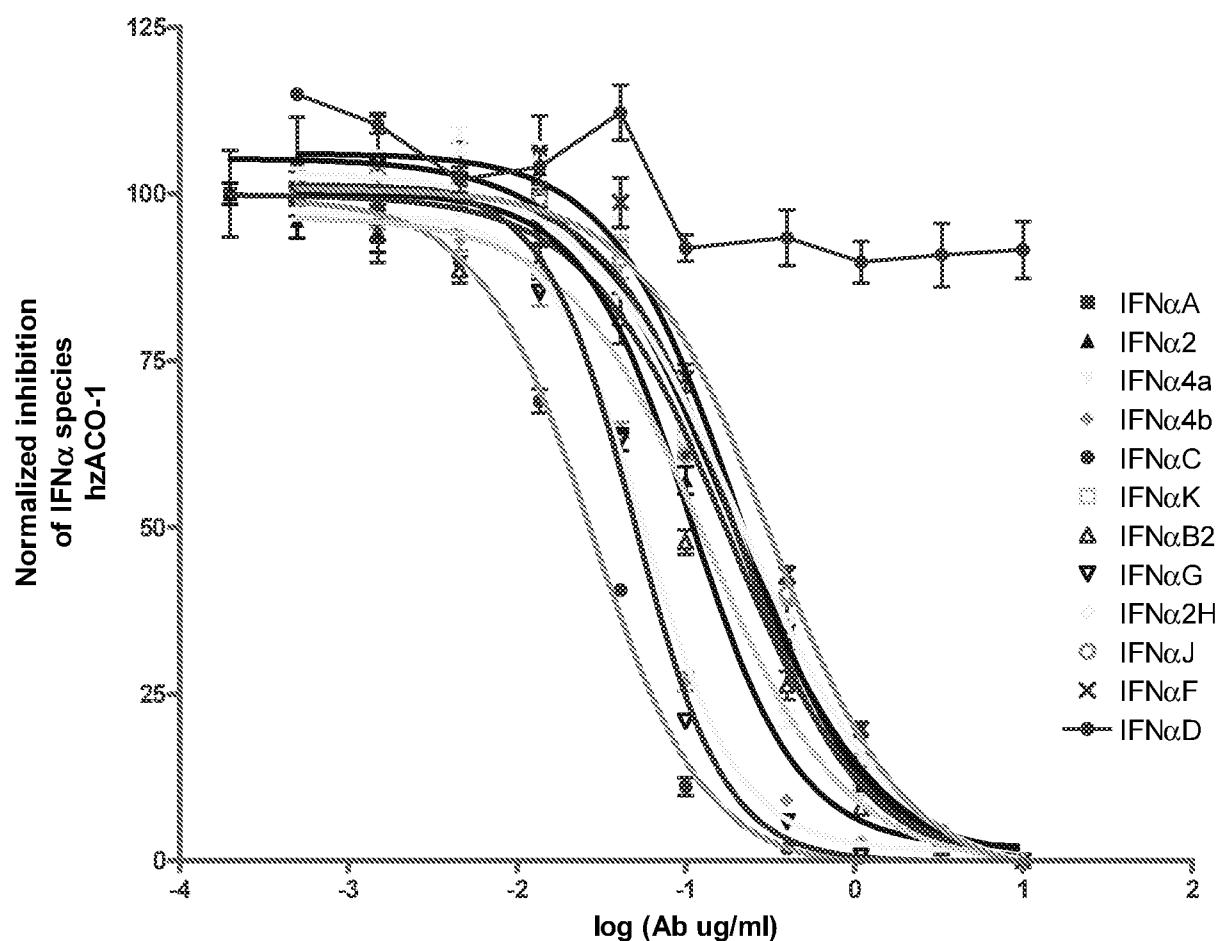
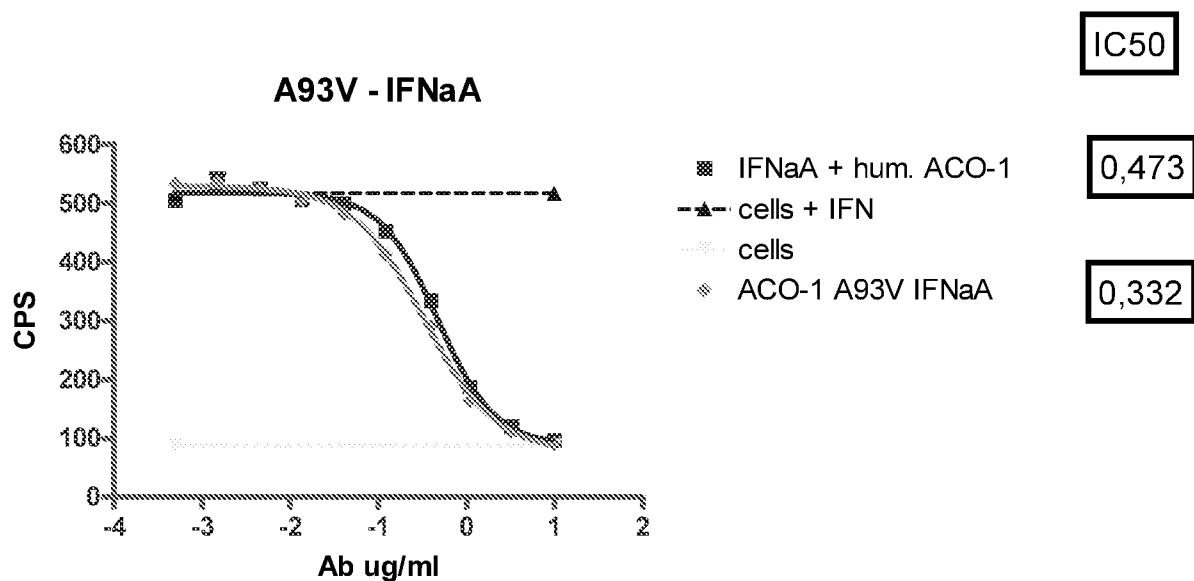
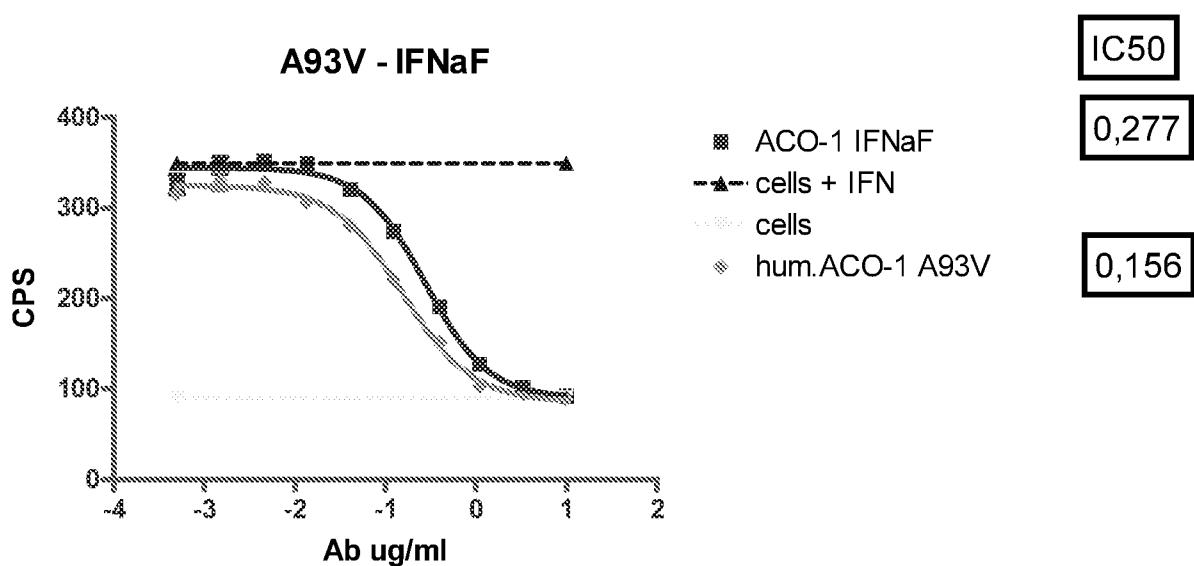
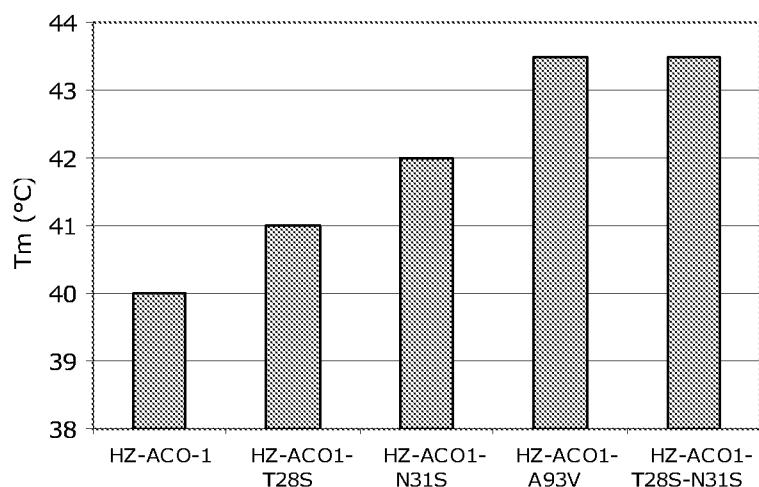
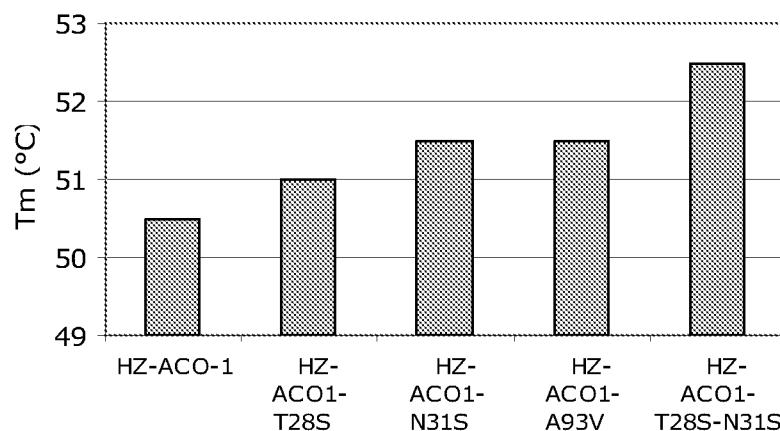
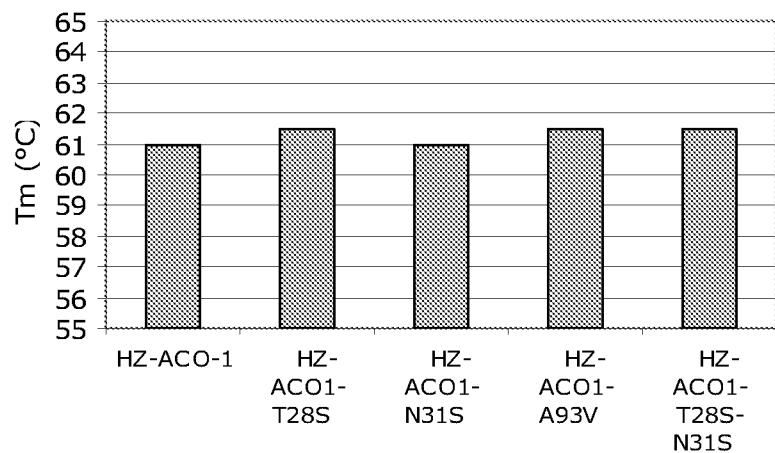


Fig. 5

A**B****Fig. 6**

A**B****C****Fig. 7**

8/13

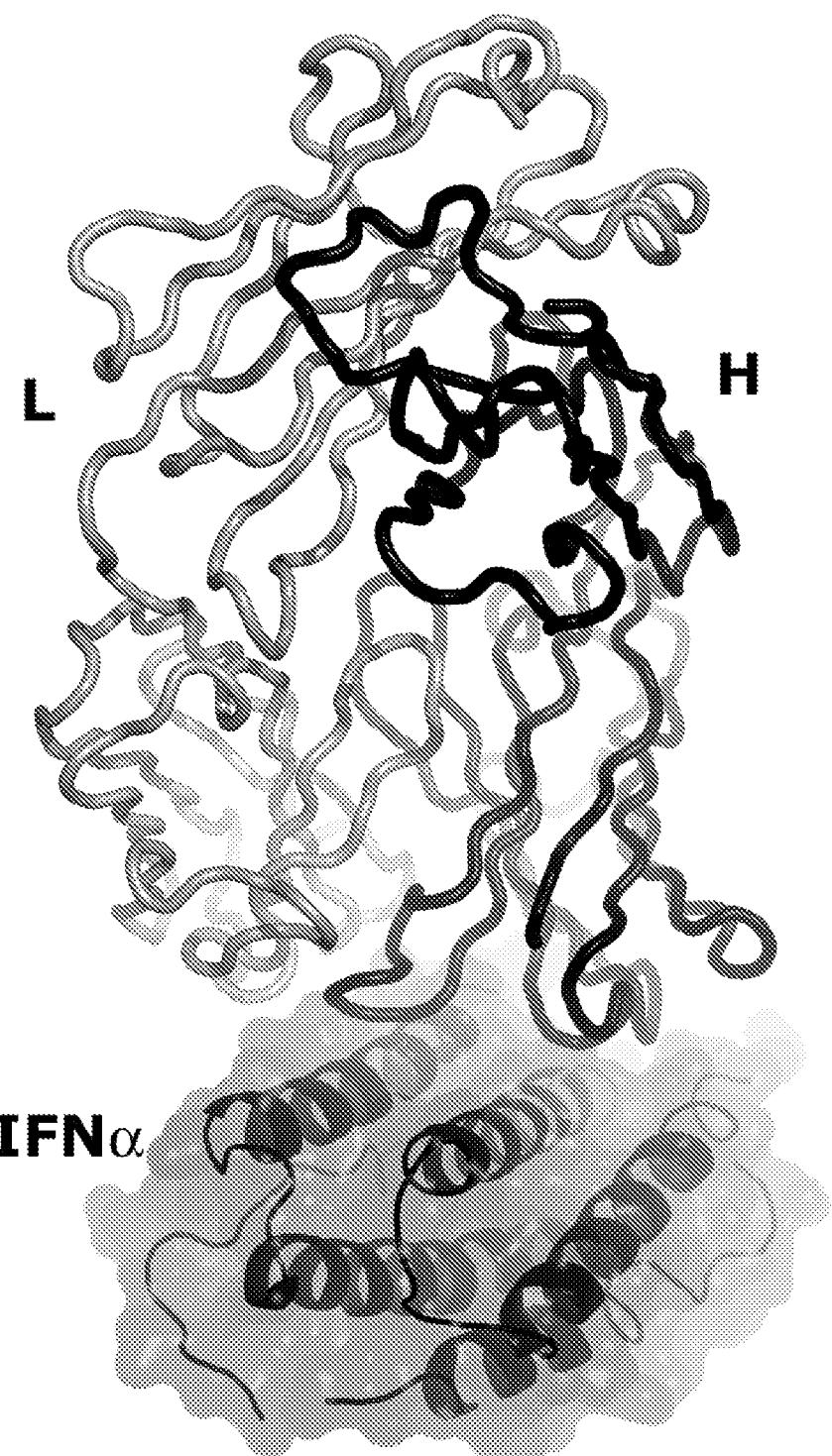
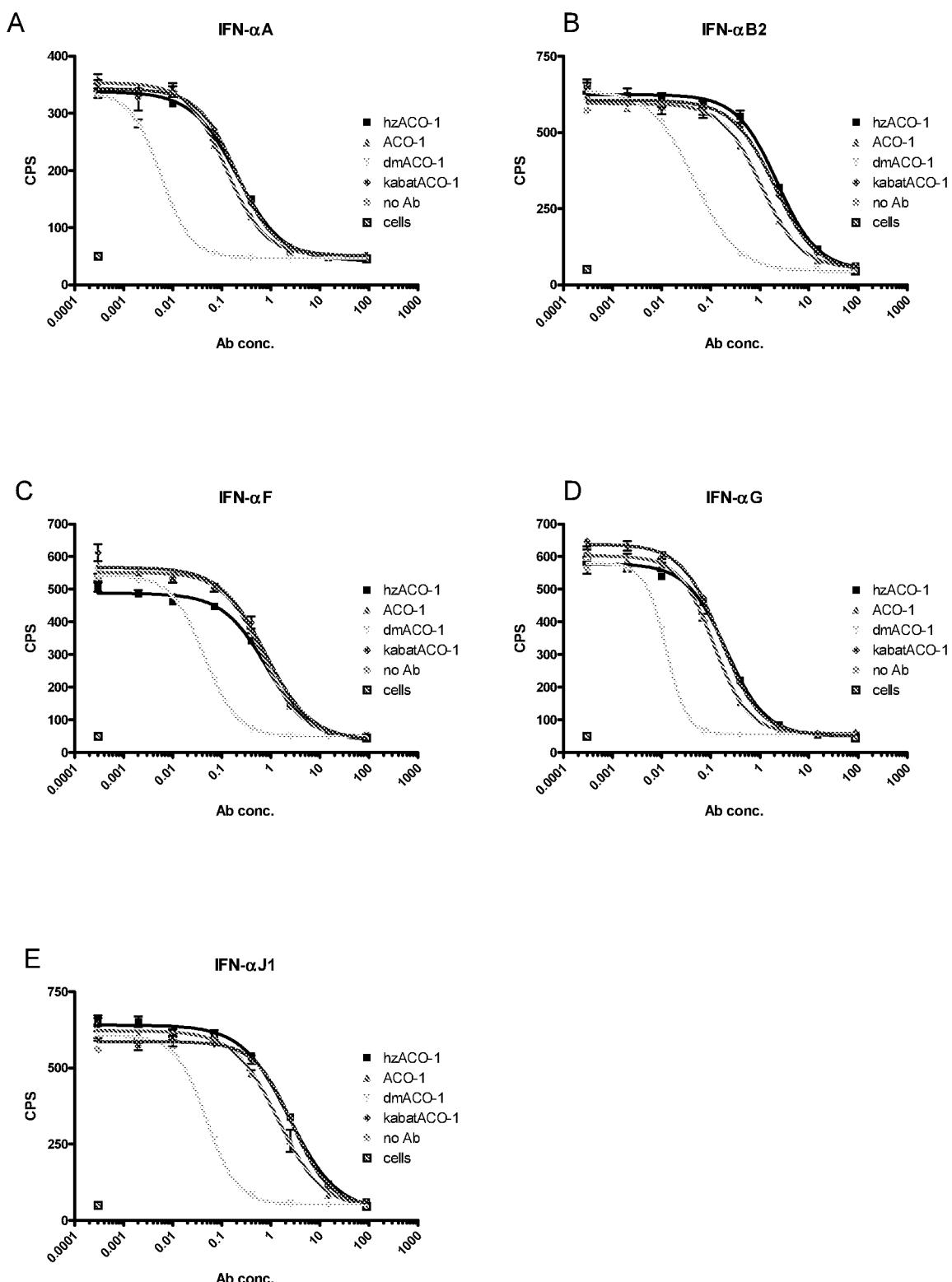
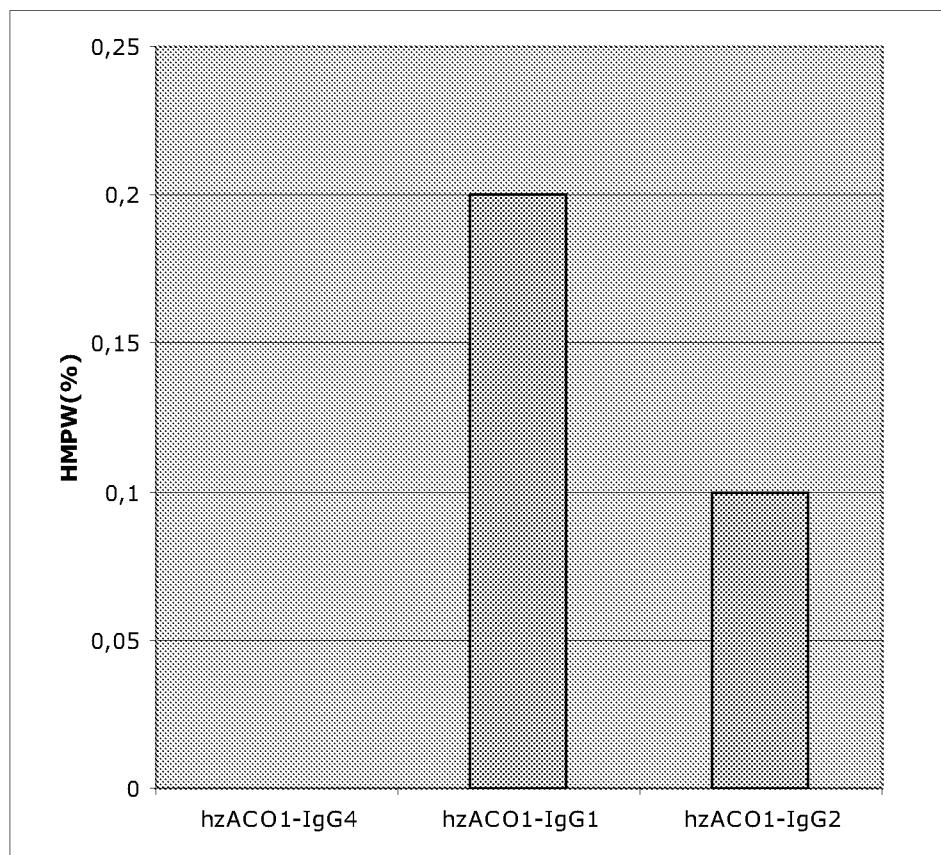


Fig. 8

IFN-a2	1	50
	(1) CDLPQTHSLGSRRTLMLLAQMRRISLFSC	KLDRHDFGFPQEFGN-QFQK
IFNAR1	-----	-----
IFNAR2	-----	-----
hzACO-1	-----	-----
IFN-a8	(1) CDLPQTHSLGNRALILLAQMRRISPFSCL	KDRHDFEFQEEFDDKQFQK
	51	100
IFN-a2	(50) AETIPV	LHEMIQQIFNLF
IFNAR1	-----	-----
IFNAR2	-----	-----
hzACO-1	-----	-----
IFN-a8	(51) AQAISVLHEMIQQT	FNLF
	101	150
IFN-a2	(100) IQGVGV	TETPLMKEDS
IFNAR1	-----	-----
IFNAR2	-----	-----
hzACO-1	-----	-----
IFN-a8	(101) MQE	GVVIESPLMYEDS
	151	166
IFN-a2	(150) SFSL	STNLQES
IFNAR1	-----	-----
IFNAR2	-----	-----
hzACO-1	-----	-----
IFN-a8	(151) SFSL	SINLQKRLKSKE

Figure 9

**Figur 10**

**Figure 11**

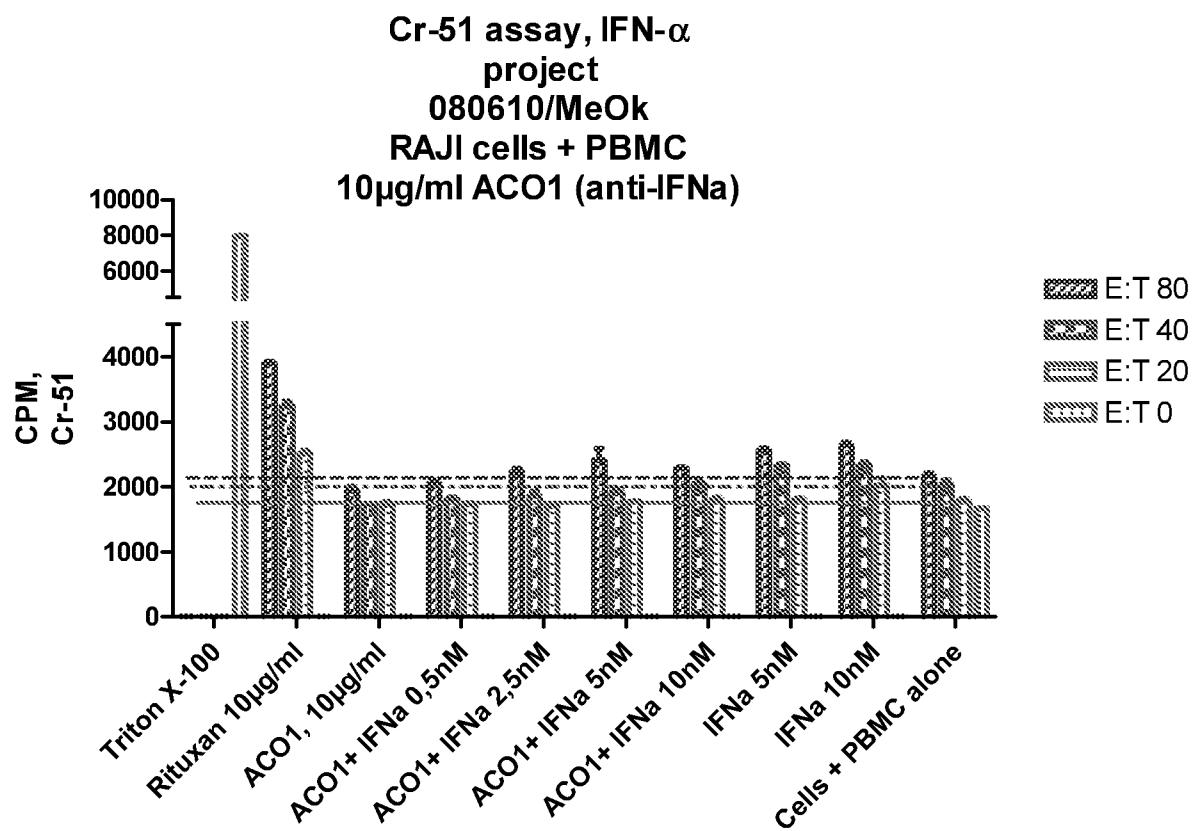
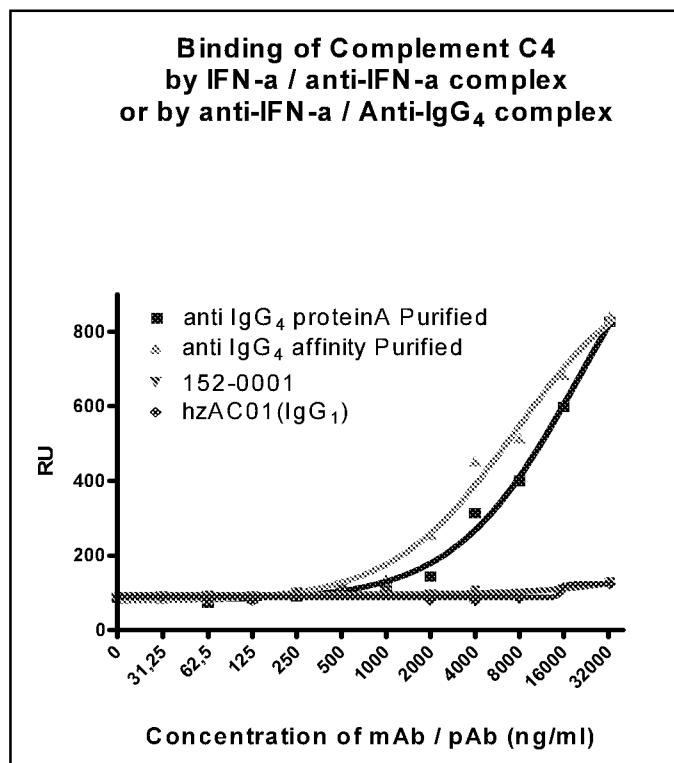


Figure 12

**Figure 13**