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(54) Title: IMMUNOASSAY REAGENTS AND METHODS OF USE THEREOF

(57) Abstract: The present invention provides reagents and methods of using the reagents, for example, on automated staining devices, that facilitate detection of two or more antigens in a sample simply and efficiently.

TITLE OF THE INVENTION

IMMUNOASSAY REAGENTS AND METHODS OF USE THEREOF

BACKGROUND OF THE INVENTION

FIELD OF THE INVENTION

The present invention relates to the detection of antigens in a sample using antibody reagents.

DESCRIPTION OF THE BACKGROUND

The evolving field of immunohistochemistry has led to simplified protocols with shorter incubation times, and automation. The ability to stain two or more antibodies on one tissue section has evolved as well. A double or triple stain assay would be much more practical for both pathologist and histo-technologist, reducing error, reducing reagents, reducing amount of tissue needed, and most important—time. Furthermore, this technology leads to a more practical method for a pathologist to make a diagnosis with antibodies that need to be stained in unison as well as situations where there is limit tissue.

In the past, double or triple stains were primarily performed in the research setting. However, the prior double and triple stained procedures required trained personnel, were technically difficult, time consuming, and in most cases, was not used on an automated immunostaining system. Therefore, the use of double or triple staining techniques is not often used in the clinical setting.

Previously, a typical 3-step avidin-biotin horseradish peroxidase (HRP) system was used for the first antibody sequence, followed by an elution (denaturing) step, and then a second antibody sequence with an alkaline phosphatase (AP) system. This product a brown (DAB) and red (Fast Red) color precipitate for each antigen, thus producing a double stain. If the tissue contained endogenous biotin, an avidin-biotin step was required. A double stain could require 30 to 40 steps and a triple stain could take 40 to 50 steps, depending on the

complexity of the assay. Most automated immunohistochemistry stainers were not designed to do these complicated double stain assays. It also required a highly trained technician to properly titer each antibody sequence.

A biotin-free polymer detection kits for both HRP and AP detection system has been recently introduced (DakoCytomation; Zymed Laboratories). These kits eliminate the avidin-biotin blocking steps and require one step for detection versus two. However, depending on the immunostainer and the number of washes required, these kits still require significant technical expertise and IHC staining programming are not design adequately for double stains and especially triple stains. Also, lack of sensitivity with nuclear antigens have been reported with this kind of polymer kit. This new technology has been popular in the research arena, but has not been used much in clinical setting because of these difficulties.

The use of antibody cocktails has been used in the clinical setting for a number of years. LCA, AE1/AE3, CMV, and most recently, CD15, Pan Melanoma (HMB45 + MART-1 + Tyrosinase) and PIN-4 (P504S + HMWCK + p63) are several cocktails that are being used on a routine basis in the clinical setting. Universal detection kits with both anti-mouse and anti-rabbit have also been used since the late 1980's.

Double and triple stain technology using immunohistochemistry in formalin-fixed paraffin-embedded tissues has also been used for many years. Double stains are accomplished by applying primary antibodies and detection in a sequence of steps to achieve multiple labeling on the same tissue. Various methods for detection used include fluorescence, immunoperoxidase, immunogold and in situ methods.

In view of the above, the disadvantages of the previous double stain technology have become apparent. Illustrative of these disadvantages is the typical procedure of the previous technology to perform a double stain immunoassay. For example, the sample is treated with Hydrogen Peroxide for 5 minutes followed by two optional protein block (5 to 10 minutes)

and Avidin-Biotin block (20 to 40 minutes). Then, the primary antibody is applied for 30 to 60 minutes, linked for 10 to 20 minutes, labeled with, e.g., HRP for 10 to 20 minutes, treated with DAB for 5 minutes, and then denatured for 5 minutes. Subsequently, the second primary antibody is applied for 30 to 60 minutes, followed by an optional protein block for 5 to 10 minutes, linking for 10 to 20 minutes, and then labeling the second primary antibody, for example, with AP, for 10 to 20 minutes. The reactivity is detected by applying Fast Red for 10 to 20 minutes followed by counterstaining (plus bluing) for 30 and 60 seconds and coverslipping, which requires a water-base mounting media. The total time for this double stain procedure is about 3 to 4 hours with a total of 11-15 manual steps; plus 12 to 14 washes (32 maximum steps). Further, for a triple stain one would add 5 more steps (40+ maximum steps) and would take a total time of 4 to 5 hours.

Generally, diluting antibodies in a buffer and storing antibodies is known however, those dilutions typically were performed in phosphate buffered saline or other isotonic solution and may also have small quantities of bovine serum albumin and in some instances relatively small amounts of detergents such as Triton X-100 or NP-40 (Antibodies: A Laboratory Manual, Harlow and Lane, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1988; and Current Protocols in Molecular Biology, Ausubel et al (eds.), John Wiley and Sons, Inc. N.Y., 2001). However, the combination of reagents in a primary antibody cocktail as described as the present invention was not described previously. Furthermore, it was not known that these cocktails would stabilize the antibodies and permit the rapid and efficient staining with two or more antibodies.

SUMMARY OF THE INVENTION

Accordingly, the present invention provides reagents containing primary antibodies and a detection system, which accomplish these goals. Thus, the present invention provides a

means to dramatically reduce the amount of steps required and provides the ability to perform automated immunoassays.

In one aspect of the present invention compositions comprising two or more antibodies mixed with various reagents are provided.

In another aspect of the present invention compositions comprising reagents suitable for the detection of the antibody containing reagents are provided.

In another aspect of the present invention, methods of detecting two or more antigens in a single sample using the compositions of the invention are provided. In one embodiment, the method can be performed on an automated device capable of staining and detecting the antigens in the sample.

In an additional aspect of the present invention is a denaturing solution that compensates for tissue pretreatment and methods of use in an antibody detection assay.

DETAILED DESCRIPTION OF THE INVENTION

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described herein. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and are not intended to be limiting.

Reference is made to standard textbooks of molecular biology that contain definitions and methods and means for carrying out basic techniques, encompassed by the present invention. See, for example, Sambrook et al., *Molecular Cloning: A Laboratory Manual*, 3rd

Edition, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York (2001), Current Protocols in Molecular Biology, Ausebel et al (eds.), John Wiley and Sons, Inc., New York (2001), and the various references cited therein.

Primary Antibody compositions

The present invention provides cocktailed primary antibodies, i.e., an antibody composition that are ready to use or concentrates). The advantages of providing such cocktails include one half to one third of the total number primary steps for performing a double staining procedure, i.e., staining with two antibodies; when using a primary cocktail of mouse and rabbit antibodies, the denaturing step may be omitted; a triple stain, i.e., staining with three antibodies, can be accomplished with 8 to 10 steps verses 16 to 24 steps in prior procedures, i.e., 2 to 2 ½ hours for the present invention versus 4 to 5 hours in prior procedures; both double and triple staining procedures can be fully automated; it enables the reduction in the number of slides to be processed; permits a higher level of quality control, for example, when the stained samples are viewed by a pathologist. Overall the present invention also reduces total costs, which is a significant factor in processing samples in the clinical setting.

The present invention is applicable anywhere antibody staining is conventional employed. For example, double, triple and quadruple staining can be performed in Immunocytochemistry; Immunohistochemistry; Frozen sections; Formalin-fixed paraffin embedded tissues; Cell Cultures; Tissue or cell culture microarrays; Paraffin-embedded tissues (any fixation protocol); Cell smears; Cell blocks; Cytospins; PAP smear; Blood smears; and Touch Preps. In certain embodiments, the sample is attached to a solid support, for example, a glass slide, ELISA plate, culture dish, glass dish, plastic dish, glass well, or plastic well.

The antibody compositions or cocktails of the present invention comprise two or more antibodies in double staining procedures, and, for example, in triple staining procedures the compositions would contain three antibodies, etc. The antibodies can be preferably different in that the antibodies react with different antigens and/or different isoforms or epitopes of a particular protein/antigen in a single sample. The antibodies can be monoclonal or polyclonal and can be obtained according to standard antibody technology known in the art. The antibodies can be from any species, such as rat, horse, goat, rabbit, human, mouse, etc. In a preferred embodiment, the antibodies are from rabbit, goat, and/or mouse. In another preferred embodiment, the at least two of the antibodies in the composition are from different sources or species. In another preferred embodiment, at least one of the antibodies in the primary antibody cocktails is a rabbit antibody, and more preferably a rabbit monoclonal antibody. For example, Rabbit monoclonal antibodies can be obtained from Lab Vision Corp; Fremont, CA)

The antibodies can be diluted in one or more of the buffer systems described herein, for example, at least 1:50 and preferably from about 1:50 to 1:6000, including 1:100, 1:150, 1:200, 1:250, 1:300, 1:350, 1:400, 1:450, 1:500, 1:550, 1:600, 1:650, 1:700, 1:750, 1:800, 1:850, 1:900, 1:1000, 1:1100, 1:1200, 1:1300, 1:1400, 1:1500, 1:1600, 1:1700, 1:1800, 1:1900, 1:2000, 1:2100, 1:2200, 1:2300, 1:2400, 1:2500, 1:2600, 1:2700, 1:2800, 1:2900, 1:3000, 1:3100, 1:3200, 1:3300, 1:3400, 1:3500, 1:3600, 1:3700, 1:4000, 1:4100, 1:4200, 1:4300, 1:4400, 1:4500, 1:4600, 1:4700, 1:4800, 1:4900, 1:5000, 1:5100, 1:5200, 1:5300, 1:5400, 1:5500, 1:5600, 1:5700, 1:5800, 1:5900 and all ranges and values therebetween.

According to the invention, the two or more antibodies are prepared in a composition comprising a buffer system that stabilizes the antibodies and facilitates a faster detection after the antibodies have bound to the antigen.

In one embodiment, the composition of the two or more antibodies contains, in one embodiment, sodium phosphate monobasic, monohydrate; potassium phosphate dibasic, trihydrate; Polyoxyethylenesorbitan monolaurate (Tween® 20), bovine serum albumin (BSA), sodium azide, and water and wherein the pH is about from 5.5 to 6.5 and preferably 6.0. In a preferred embodiment, this composition contains approximately 1.5 to 3.0 g/l sodium phosphate monobasic, monohydrate; 0.5 to 0.6 g/l potassium phosphate dibasic, trihydrate; 0.5 to 1.5 ml/l Polyoxyethylenesorbitan monolaurate (Tween® 20) 50 to 100 ml/l of 5% bovine serum albumin (BSA), 0.5 to 1.5 g/l sodium azide, 0.005 g/l, and the remainder water. In a more preferred embodiment, this composition contains approximately 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l Polyoxyethylenesorbitan monolaurate (Tween® 20), 100 ml/l of 5% bovine serum albumin (BSA), 0.9 g/l sodium azide, and the remainder water. In one embodiment, bromthymol blue may be added to these compositions in an amount of from 0.0001 to 0.001 g/l, and preferably 0.005 g/l.

In second embodiment, the composition contains PBS, sodium azide, Polyoxyethylenesorbitan monolaurate (Tween® 20), BSA and the remainder water, and where the pH is from about 7.0 to about 6.5 and preferably about 7.3. In a preferred embodiment, this composition contains 40 to 60 ml/l 10X PBS, 0.75 to 1.25 g/l sodium azide, 0.1 to 0.3 ml/l Polyoxyethylenesorbitan monolaurate (Tween® 20), 45 to 65 g/l BSA, and the remainder water. In a further preferred embodiment, this composition contains 50 ml/l 10X PBS, 0.9 g/l sodium azide, 0.2 ml/l Polyoxyethylenesorbitan monolaurate (Tween® 20), 55 g/l BSA, and the remainder water.

In a third embodiment, the composition contains Tris-HCl, sodium azide, BSA, hydrochloric acid and the remainder water at a pH of from about 5.7 to about 6.5 and preferably about 6.2. In a preferred embodiment, this composition contains 3.0 to 4.0 g/l

Tris-HCl, 0.75 to 1.2 g/l sodium azide, 7.5 to 12.5 g/l BSA, 0.2 to 0.3 ml/l 25 % hydrochloric acid, and the remainder water. In a further preferred embodiment, this composition contains 3.5 g/l Tris-HCl, 0.9 g/l sodium azide, 10 g/l BSA, 0.25 ml/l 25 % hydrochloric acid, and the remainder water.

In a fourth embodiment, the composition contains 1 part of solution A and 1 part of solution B where solution A contains PBS, preservative (Proclin™ 950), Polyoxyethylenesorbitan monolaurate (Tween® 20), purified casein, Prionex (purified Type A gelatin), and water and solution B contains sodium phosphate monobasic, monohydrate; potassium phosphate dibasic, trihydrate; Polyoxyethylenesorbitan monolaurate (Tween® 20), bovine serum albumin (BSA), sodium azide, and water and wherein the pH is about 5.75 to 6.25 and preferably a pH of 6.0. In a preferred embodiment of this composition, solution A contains 75 to 125 ml/l 10 X PBS, 2.5 ml/l preservative (Proclin™ 950), 0.25 to 0.75 ml/l of 50% Polyoxyethylenesorbitan monolaurate (Tween® 20), 2.5 to 7 g/l purified casein, 2.0 to 3.0 ml/l Prionex (purified Type A gelatin), and water; and solution B contains 2.0 to 3.0 g/l sodium phosphate monobasic, monohydrate; 0.5 to 0.6 g/l potassium phosphate dibasic, trihydrate; 0.75 to 1.25 ml/l Polyoxyethylenesorbitan monolaurate (Tween® 20), 75 to 125 ml/l of 2.5 to 7.5 % bovine serum albumin (BSA), 0.7 to 1.1 g/l sodium azide, and the remainder water. In a further preferred embodiment of this composition solution A contains 100 ml/l 10 X PBS, 2.5 ml/l preservative (Proclin™ 950), 0.5 ml/l of 50% Polyoxyethylenesorbitan monolaurate (Tween® 20), 5 g/l purified casein, 2.5 ml/l Prionex (purified Type A gelatin), and water; and solution B contains approximately 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l Polyoxyethylenesorbitan monolaurate (Tween® 20), 100 ml/l of 5% bovine serum albumin (BSA), 0.9 g/l sodium azide, and the remainder water. In a further preferred embodiment,

solution B also contains 0.0001 to 0.001 g/l bromthymol blue, preferably 0.005 g/l bromthymol blue.

In a fifth embodiment, the compositions of primary antibodies comprises sodium phosphate monobasic, monohydrate; potassium phosphate dibasic, trihydrate; Polyoxyethylenesorbitan monolaurate (Tween® 20), bovine serum albumin (BSA), sodium azide, and water and wherein the pH is about from 5.5 to 6.5 and preferably 6.0 and from 40 to 60% glycerol, preferably 50% glycerol. In a preferred embodiment, this composition contains approximately 1.5 to 3.0 g/l sodium phosphate monobasic, monohydrate; 0.5 to 0.6 g/l potassium phosphate dibasic, trihydrate; 0.5 to 1.5 ml/l Polyoxyethylenesorbitan monolaurate (Tween® 20), 50 to 100 ml/l of 5% bovine serum albumin (BSA), 0.5 to 1.5 g/l sodium azide, 0.005 g/l, and the remainder water. In a more preferred embodiment, this composition contains approximately 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l Polyoxyethylenesorbitan monolaurate (Tween® 20), 100 ml/l of 5% bovine serum albumin (BSA), 0.9 g/l sodium azide, and the remainder water. In one embodiment, bromthymol blue may be added to these compositions in an amount of from 0.0001 to 0.001 g/l, and preferably 0.005 g/l.

Non-limiting Examples of primary antibodies and antibody combinations useful in the present invention include those listed in below (AP = Alkaline phosphatase, HRP = Horseradish Peroxidase, Ms + Mouse): T-cell and apoptosis marker (CD3 [PS1]; Caspase-3 (Polyclonal)); B-cell vs T-cell marker (CD20 [L26]; CD3 (Rabbit monoclonal) [SP11]); B-cell vs T-cell marker (CD20 [L26]; CD3 [PS1]); Blood Vessels and cell proliferation (CD31 [JC/70A]; Ki-67 (Rabbit monoclonal) [SP6]); Dermatopathology Marker (CD34 [QBEnd/10]; Factor XIII Subunit a (polyclonal)); Colon cancer verses breast or lung cancer (CDX2 [CDX2-88]; CK7 [K72.7]); G.I vs Lung or breast (CK7 [K72.7]; CK20 [Ks20.8]); Kappa/Lambda (Kappa [KDB-1]; Lambda (polyclonal)); Cell proliferation and Apoptosis

marker (Ki-67 [MIB-1]; Caspase-3 (polyclonal)); Cell proliferation and Apoptosis marker (M30 (CM224A); i-67 (rabbit monoclonal) [SP6]); Screening Marker (LCA [PD7/26/16] + 2B11; S-100 (polyclonal)); Screening Marker (LCA [PD7/26/16 + 2B11]; S100 (Polyclonal); CK8/18 [5D3]) B-cell and cell proliferation (CD20 [L26]; Ki-67 (rabbit monoclonal)[SP6]; Pan Melanoma; Tyrosinase [T311];S-100 (rabbit polyclonal); MART-1 [M2-7C10 + M2-9E3] or A103); Invasive breast cancer (p63 [BC4A4]; CK5 [XM26]; CK8/18 [5D3]); PIN Cocktail-2 (p63 [BC4A4];P504S (rabbit monoclonal) [P504S]); PIN Cocktail-4 (p63 [BC4A4] ; HMW CK [DE-SQ] ; P504S (rabbit monoclonal)); Mesothelioma marker (CK5/6 [CK5/6.007]; Calretinin (rabbit monoclonal) [SP13]); Breast cancer prognostic marker (Estrogen receptor [6F11]; Ki-67 (rabbit monoclonal)); Breast cancer prognostic marker (Estrogen receptor (rabbit monoclonal)[SP1]; Ki-67 [MIB-1]); Prognostic breast cancer marker (CK5 [XM26]; CK17 [Ks17.E3]; CK8/18 [5D3]); Liver cancer marker (CD10 [56C3]; Prostate Specific Antigen (PSA) (polyclonal); Hepatic Specific Antigen (HSA) [OCH1E5]; Pan Neuroendocrine; Chromogranin A [LK2H10 + PHE5]; Synaptophysin (Rabbit Monoclonal) [SP11]); Adenocarcinoma vs Squamous Cell Carcinoma (HMW CK [DE-SQ] or [34 β E12]; LMW CK (CK8/18) [5D3]); B-cell and Apoptosis Marker (CD20 [L26]; Caspase-3 (polyclonal)); T-cell and cell proliferation marker (CD3 [PS1]; Ki-67 (Rabbit Monoclonal)[SP6]); Chronic Lymphocytic Leukemia (CD5 (rabbit monoclonal) [SP9]; PAX-5 [BC/24]); and T-cell Marker (CD4 [1F6]; CD8 (polyclonal))

Further non-limiting examples of antibody cocktails (compositions) are listed in the table below (M=mouse; R=rabbit):

LCA (M) + S100 (R)
 LCA (M)+ S100 (R) + CK8/18 (M) or Pan Cytokeratin (Triple stain)
 Kappa (M) and Lambda (R)
 L26 (M) B-cell + CD3 (R) T cell (Lymphoma)
 L26 (M) + Ki-67 (R)
 CD3 (M) + Ki67 (R)
 Ki-67 (M) + Caspase-3 (R)
 HMWCK (M) + LMWCK (M)
 CK7 (M) + CK20 (M)
 CK7 (M) + CDX-2 (M)
 ER (M) + Ki-67 (R)
 P504S (R) + HMWCK + p63 (M) (PIN)
 CK5 + p63 + CK8/18 (Breast cancer)
 CK5/6 (M) + Calretinin (R)
 MART-1 + Tyrosinase + S100 (R) (Melanoma)
 CD34 (M) + Factor XIIIa (R)
 Apoptotic/CK18 (M) + Ki-67 (R)
 CD31 (M) + Ki-67 (R)
 CD10 + HSA + PSA (Liver cancer and metastasis) (Triple Stain)
 PAX-5 + CD 5 (leukemia)
 CK/5 + CK17 + CK8/18
 CK5/6 + Calretinin
 ER + Ki-67
 CDX2 + CK7

Secondary Antibody compositions

The use of secondary antibodies to detect the binding between a primary antibody and an antigen are known (Antibodies: A Laboratory Manual, Harlow and Lane, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1988; and Current Protocols in Molecular Biology, Ausebel et al (eds.), John Wiley and Sons, Inc. N.Y., 2001). However, in the present invention the formulation of a composition of a secondary antibody composition capable of detecting two different antibodies at the same time was not known.

Accordingly, one embodiment of the present invention is to provide a secondary antibody composition coupled with a biotin-free polymer conjugated to an enzyme or other detectable moiety capable of being detected using various chromogens and/or other

detectable moieties. Conjugating detectable moieties or enzyme systems to secondary antibodies is known (Antibodies: A Laboratory Manual, Harlow and Lane, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1988; and Current Protocols in Molecular Biology, Ausebel et al (eds.), John Wiley and Sons, Inc. N.Y., 2001).

The secondary antibodies are raised to react with the primary antibody based on the species origin of the primary antibody, e.g., if the primary antibody is a mouse antibody then the secondary antibody would be, for example, a rabbit anti-mouse antibody. In the present invention, the two or more secondary antibodies to visualize the binding between the two or more primary antibodies and the two or more antigens may be from the same source species or from different species. However, the two or more secondary antibodies are preferably coupled to a detectable moiety such as horseradish peroxidase (HRP) and alkaline phosphatase (ALP or AP) and in a particularly preferred embodiment; at least a first secondary antibody contains a detectable moiety that is different from the detectable moiety on a second secondary antibody. In one aspect of the present invention, the detectable moiety is poly-HRP or a poly-ALP and is described in Shi et al ((1999) Appl. Immuno & Mol. Morph. 7(3):201-208).

In accordance with the present invention, the secondary antibody composition described above, e.g., containing two or more secondary antibodies specific for the primary antibody composition, is contained in a buffer system suitable to stabilize the HRP and the AP moieties coupled to the secondary antibodies. In one aspect of this embodiment, the buffer system is a Tris buffer system, which may also contain a preservative that prevents microbial growth and does not result in any appreciable degradation of the detectable moieties. In one embodiment, the preservative is Proclin®.

In another embodiment, the buffer system comprises a solution of Tris buffer at a pH of from 7.3 to 7.9, Polyoxyethylenesorbitan monolaurate (Tween® 20) and goat serum. In a

preferred embodiment, the secondary antibody composition comprises 0.8 to 1.2 M Tris buffer, pH from 7.3 to 7.9 with 0.025 to 0.075 % Polyoxyethylenesorbitan monolaurate (Tween® 20) and 2.5 to 3.5 % goat serum; and more preferably the secondary antibody composition comprises 0.1M Tris buffer, pH 7.6 with 0.05% Polyoxyethylenesorbitan monolaurate (Tween® 20) and 3% goat serum. In an alternative embodiment, the above composition may also contain tris-buffered saline (TBS) in amounts from 30 to 37 ml of 10 X TBS/l, preferably 34 ml 10 X TBS/l.

In a preferred embodiment, a conjugated goat anti-mouse poly-alkaline phosphatase (ALP) and a conjugated goat anti-rabbit poly-horseradish peroxidase (HRP) are used as secondary antibodies and react with both heavy and light chains on mouse and rabbit IgG. The ALP- and HRP-polymerization provides a significant increase in staining sensitivity when compared to other conventional ALP- or HRP-conjugated secondary antibodies. Avidin-biotin blocking procedures are not necessary when using the conjugated secondary antibodies. Obviously then, the primary antibody cocktail will contain a rabbit and a mouse antibody.

In one embodiment of the present invention, chromogens are used to detect the antibody-antigen complex. Chromogens have improved dramatically over the last 5 years. In the past, several chromogens required a water-soluble mounting media and/or faded over time. The new generation of chromogens are permanent and do not fade. They come in several different colors. It is now easy to achieve four different chromogens on a single section. For example, chromogens useful in HRP Systems include, DAB (which appears brown), AEC (which appears brick red), and Bajoran Purple (which appears lavender to dark purple). For example, chromogens useful for Alkaline Phosphatase System include Fast Red (which appears pink to fuchsia), and Ferangi Blue (which appear royal blue).

Bajoran Purple is a permanent chromogen and is compatible with streptavidin horseradish peroxidase. Bajoran Purple is not soluble in alcohol or xylene and can be coverslipped just like DAB. When in the presence of a peroxidase enzyme, Bajoran produces lavender-purple stain.

3,3' Diaminobenzidine (DAB) is a widely used chromogen for immunohistochemical staining and immunoblotting. When in the presence of peroxidase enzyme, DAB produces a brown precipitate that is insoluble in alcohol and xylene. In a preferred embodiment, DAB is used with an enhanced to add contrast and increase staining intensity, such as for example *DABENHANCER* (BIOCARE Medical).

Fast Red is a widely used chromogen for immunohistochemical staining. When in the presence of alkaline phosphatase (AP) enzyme, Vulcan Fast Red produces a bright fuchsin-red precipitate that is insoluble in organic solvents and can be coverslipped with a permanent mounting media. When using an alkaline phosphatase system, tris buffer (pH 7.6) rather than PBS should be used as a rinsing buffer.

When in the presence of alkaline phosphatase (AP) enzyme, Ferangi Blue produces a bright royal blue precipitate that is insoluble in organic solvents---and can be coverslipped with a permanent mounting media. When using an alkaline phosphatase system, tris buffer (pH 7.6) rather than PBS should be used as a rinsing buffer.

In preferred embodiments, counterstains are used to visualize the staining patterns of the antibody-antigen complex. Non-limiting examples of those counterstains include Hematoxylin, Nuclear Fast Red, Methyl Green, and Methyl Blue.

In a set of preferred embodiments, cocktails of primary antibodies diluted in various buffers (referred to below as 1-5 corresponding to the first to fifth embodiments of the primary antibody cocktails above, combined with various chromogens are described below:

Methods of performing an immunoassay

To perform an immunoassay using the compositions of the present invention, a sample can be first contacted with a primary antibody composition, i.e., containing two or more primary antibodies and subsequently with a cocktailed detection reagent, i.e., containing two or more secondary antibodies coupled to detectable moieties. In a preferred embodiment of this method, the detectable moieties are at least AP and HRP, more preferably poly (AP) and poly (HRP). The antibody staining method of the present invention is different from prior staining protocols used previously because prior to the present invention performing staining of a sample with two or more antibodies required the application of the first primary antibody followed by detection of this antibody, a subsequent denaturing step to prepare the sample to receive a second primary antibody, then applying a second primary antibody followed by detection of this antibody, etc.

In a further embodiment of the antibody staining method of the present invention, a second composition of primary antibody, either containing a single primary antibody or a cocktail of 2 or more antibodies is added to the same sample following the detection of the first composition of cocktailed primary antibodies. Subsequently, the second composition of primary antibodies is detected using the detection reagents, secondary antibodies and detectable moieties as described herein.

In further embodiments of performing a double stain (or an immunoassay with two different antibodies) using the antibody compositions and the detection reagents of the present invention the following protocol can be employed, washing steps are employed between the various steps as is typically performed in an immunoassay. In some instances it may be preferable to pretreat the sample with a peroxide to facilitate antigen retrieval and/or enzyme digestion, however, as is appreciated by one of skill in the art some retrieval

solutions block peroxidase and therefore the peroxide treatment would not be performed. In addition, in some instances recognized by the skilled artisan, it may be helpful to perform a protein-blocking step to minimize non-specific binding of the antibodies to the sample. If the protein blocking step is used it is typically conducted for about 10 minutes.

The primary antibody cocktail is applied to the sample to be assayed, typically from about 30 to about 60 minutes after which time a secondary-HRP -ALP Polymer cocktail is applied for about 20 to about 30 minutes. To visualize the antibody-antigen complexes a first chromogen, e.g., DAB is applied for about 5 minutes followed by applying a second chromogen, e.g., fast red for about 10 minutes. In preferred embodiments of the present invention, the sample is then counterstained, e.g., with progressive hematoxylin, for approximately 30 seconds to about one minute to facilitate better visualization of the stained antigen. The sample, if on a slide can then be covered with a coverslip to preserve the stained sample. Thus, this procedure can be performed in approximately 2 to 2 ½ hours requiring only about 5 to 7 steps and 6 to 8 wash steps.

To perform a triple stain (or an immunoassay with three different antibodies) using the antibody compositions and the detection reagents of the present invention the following protocol can be employed, washing steps are employed between the various steps as is typically performed in an immunoassay. In some instances it may be preferable to pretreat the sample with a peroxide to facilitate antigen retrieval and/or enzyme digestion, however, as is appreciated by one of skill in the art some retrieval solutions block peroxidase and therefore the peroxide treatment would not be performed. In addition, in some instances recognized by the skilled artisan, it may be helpful to perform a protein-blocking step to minimize non-specific binding of the antibodies to the sample. If the protein blocking step is used it is typically conducted for about 10 minutes.

The primary antibody cocktail is applied to the sample to be assayed, typically from about 30 to about 60 minutes after which time a secondary-HRP -ALP Polymer cocktail is applied for about 20 to about 30 minutes. To visualize the antibody-antigen complexes a first chromogen, e.g., DAB is applied for about 5 minutes followed by applying a second chromogen, e.g., fast red for about 10 minutes. Then the sample is denatured using the appropriate denaturing solution for approximately 2 to 10 minutes (in some embodiments, the use of a graded dilution of the denaturing solution may be employed for about 5 minutes). A third primary antibody is applied to the sample for about 30 minutes and then a goat anti-mouse or rabbit HRP or AP Polymer is applied for about 20 to 30 minutes. A chromogen is applied, which is purple or blue, for 5 minutes. In preferred embodiments of the present invention, the sample is then counterstained, e.g., with progressive hematoxylin, for approximately 30 seconds to about one minute to facilitate better visualization of the stained antigen. The sample, if on a slide can then be covered with a coverslip to preserve the stained sample. Thus, this procedure can be performed in approximately 2 ½ to 3 hours requiring only about 9 to 11 steps and 9 to 11 wash steps.

In a similar manner as described above, immunoassays using four, five, six, etc. antibodies may be employed to visual various antigens in a given sample. When performing the above protocols using an alkaline phosphatase detection system, it is preferable to use a TBS wash buffer rather than a PBS-based wash buffers. In a similar manner, when performing the immunoassay according to the invention, the sample pretreatment must be taken into consideration. In preferred embodiments, the two or more antibodies should require the same sample pretreatment, for example, pretreatments such as none, digestion, and antigen retrieval (e.g., using low and high pH solutions). In some embodiments, no pretreatment procedure can be used for the first antibody composition (e.g., containing two or more antibodies) and then an antigen retrieval procedure can be used for the denaturing step

and then the third antibody or second antibody composition. In some embodiments, antigen retrieval can be used before or after enzyme digestion. If using digestion after antigen retrieval, pepsin or trypsin is preferably used for 30 seconds; or a highly diluted trypsin can be used for 5 minutes at room temperature on automated stainers.

In further preferred embodiments of the invention, a denaturing step is employed when two sequential antibodies and detection systems are used. The denaturing step destroys (elution) the IHC steps of the protein block, primary antibody, secondary antibody, and label from the first antibody sequence. This denaturing step facilitates clean staining, i.e., preventing/minimizing the second antibody sequence from cross-reacting with the first antibody sequence. Depending on the pretreatment that is employed (none, digestion, or antigen retrieval), the time and/or concentration of the denaturing solution should vary. In the past, this procedure was not fully understood. This caused inconsistent staining and inaccurate staining. Typically in the past, laboratories used 0.5 to 1% HCL in water or 70% alcohol for 5 minutes.

A preferred denaturing solution comprises 1 part of solution A to 3 parts of solution B is also provided. Solution A contains 1.1 to 1.3 % hydrochloric acid, 0.020 to 0.030 % preservative (Proclin™ 300), and the balance water. Solution B contains 0.1 to 0.3 % Polyoxyethylenesorbitan monolaurate (Tween® 20), about 0.2 to 0.3 % preservative (Proclin 300) and the balance water. In a preferred embodiment, solution A contains 1.2 % hydrochloric acid, 0.025 % preservative (Proclin™ 300), and the balance water; and solution B contains about 0.2% Polyoxyethylenesorbitan monolaurate (Tween® 20), about 0.25% preservative (Proclin 300) and the balance water. A preferred blocking reagent contains 10% 10X PBS, 0.25 % preservative (Proclin 950), 0.01% 50% solution of Polyoxyethylenesorbitan monolaurate (Tween® 20), 50 grams purified casein, 0.25% Prionex (purified type A gelatin), and the balance water. In one embodiment of antibody

staining with at least two antibodies, the process entails contacting the sample with a first antibody, and subsequent secondary antibody and detection reagents, e.g., chromogens, adding the denaturing solution to the sample, and then contacting the sample with a second antibody with subsequent secondary antibody and detection reagents. The staining patterns can then be visualized. This process may be repeated to visualize, 3, 4, 5, etc. total antibodies on single sample.

Automated applications

The unique formulations and ease of use of the antibody cocktail and/or detection reagents and/or other reagents useful for the immunoassay facilitate performing the immunoassay in one or more automated devices designed to detect antibody staining patterns in a sample. In particular, the compositions of the present invention enable automation of double, triple, quadruple, etc. antibody detections that were difficult and in some instances even impossible with reagents known previously.

In another embodiment of the invention, the method described herein can be applied in the immuno-analysis of tissue microarrays. Tissue microarrays are known in the art and typically contain anywhere from 50 to 500 tissues on a single slide. The advantage of the present invention in the immuno-analysis of the tissue arrays is that the assay can be performed on numerous slides, e.g., 36 to 60 slides, on an automated staining apparatus and perform double, triple, quadruple, etc. stains on each slide. Another advantage of being able to perform such multiple stains on a given slide is cost since the tissue microarray slides typically cost up to \$500.

In one embodiment, a low temperature antigen retrieval protocol can be performed. The low temperature antigen retrieval protocol involves preparing the tissue sample to the substrate, e.g., glass slide prior to staining. The low temperature method produced superior morphology as well as help secures precious tissues to the substrate. Thus, in this

embodiment, the sample is treated, prior to staining, at a temperature from 65°C to 80°C and preferably about 75°C. Typically the sample is treated overnight. In a particularly preferred embodiment, the sample is treated with a device according to the description in U.S. patent no. 6,580,056, the entire contents of which are incorporated herein by reference.

Examples of such automated devices useful according to the present invention include, but are not limited to, BioGenex I6000™ (Biogenex), Dako Autostainer (DakoCytomation) Nemesis™ (BIOCARE), and those from Ventana Medical Systems (Capillary gap stainer, NexES and Benchmark).

For example, an automated double staining procedure using one of the above, e.g., Dako autostainer, is as follows. The sample is can be optionally blocked in hydrogen peroxide for 5 minutes, the sample is treated in a Biocare Decloaker for HIER, background blocking can be attained by adding Background Sniper (Biocare Medical) for 7 minutes and then blown off. A double stain primary antibody cocktail is added to the sample for 30 to 45 minutes and then rinsed; a secondary antibody composition is added for 25 minutes and then rinsed. DAB Chromogen for 5 minutes, rinse and Fast Red chromogen for 10 minutes. The sample is then ready for visualization and analysis.

Kits

The present invention also provides kits for immunoassaying a sample containing the antibody cocktail and/or detection reagents and/or other reagents useful for the immunoassay. In a preferred embodiment, the kit also comes with instructions to use the immunoassay reagents.

Examples

Example 1

A double stain protocol for PIN-4 cocktail (P504S + HMW CK + p63)

1. After the sample is mounted on the slide, it is deparaffinized down to water
2. The antigens are heat retrieved in Reveal or BORG DECLOAKER (Biocare Medical) using a steamer for 60 minutes, or alternatively in a Digital Decloaking Chamber (Biocare Medical) for 30 seconds to 5 minutes.
3. A Protein Block in 4 drops (approx. 150 ul.) using of BACKGROUND SNIPER (Biocare Medical) is applied to the sample for 5-10 minutes. The sample is washed well.
4. The primary antibody PIN-4 Cocktail is applied for 30 to 60 minutes.
5. The sample is washed in two changes of 1X Immunocare TBS (Biocare Medical) for 2 minutes each.
6. Four drops of Goat anti-Mouse (ALP) and Goat anti-Rabbit (HRP) cocktail is added. Incubate for 30 minutes Drain Slides.
7. Wash in two changes of 1X Immunocare TBS wash Buffer (Biocare Medical) for two minutes each. Drain Slides.
8. Prepare DAB Chromogen Substrate by adding 1 drop of DAB Chromogen to 1.0 ml of Substrate Buffer and mix well. Apply four drops of the DAB Chromogen Substrate to the sample and develop for 2-3 minutes at room temperature. (DAB Sparkle (Biocare Medical) may be used for enhancement.)
9. The sample is washed in deionized water. Check specimen under microscope. If it is under-developed DAB Chromogen substrate can be reapplied for another 2 minutes. Rinse in deionized water.
10. Wash in two changes of 1X Immunocare TBS wash Buffer (Biocare Medical) for two minutes each. Drain Slides.
11. Apply 4 drops of Vulcan Fast Red™ Chromogen Substrate (Biocare Medical) for 5-10 minutes. Rinse in deionized water. Place specimen under microscope and check for

light staining. Only if the stain is too light, apply Vulcan Fast Red™ (Biocare Medical) for another 5 minutes and rinse again in deionized water.

12. Add 4 drops of Cat Hematoxylin (Biocare Medical) for 30-60 seconds. Wash in tap water.

13. Blue nuclei in 1X PBS wash buffer for 1 minute. Drain slides.

14. Wash in tap water and rinse in deionized water.

15. Dehydrate in 3 changes of 100% alcohol and clear in 3 changes of xylene.

16. Mount and coverslip.

Example 2

CK5 +p63 + CK8

APPLYING CK5 + P63 WITH A SECONDARY POLYMER HRP/DAB

1. Deparaffinize down to water
2. Heat retrieve in Decloaking Chamber (Biocare Medical) for 2-5 minutes.
3. Come back with a rinse in 1X IMMUNOCARE TBS (Biocare Medical) wash buffer.
4. Protein block for 10 minutes with **BACKGROUNDSNIPER** (Biocare Medical).

Drain protein blocker.

3. Dilute CK5 1:200 and p63 1:500 in Van Gogh Yellow (Biocare Medical). Incubate for 30 minutes.
4. Wash in two changes of 1X TBS wash buffer for 2 minutes each. Drain slides.
5. Apply 4 drops of Goat-anti-Mouse-HRP (Polymer) and incubate for 25 minutes.
Wash in 2 changes of 1X TBS wash buffer for 2 minutes each. Drain slides
6. Prepare Cardassain DAB Chromogen Substrate (Biocare Medical): Add 1 drop of DAB Chromogen

- A and B solutions to 1.0 ml of Substrate Buffer. Mix Well. Apply four drops of the DAB Chromogen Substrate. Develop for 2-5 minutes at room temperature.
9. Wash in deionized water. Check specimen under microscope (manual method only). If it is under-developed you can re-apply DAB Chromogen Substrate for another 2-4 minutes. Rinse in deionized water
 6. Dip section into 0.5% HCL in 70% alcohol 2 to 5 minutes. Rinse well with TBS wash buffer.

APPLYING CK8/18 WITH A POLYMER SECOND ANTIBODY WITH A ALP/FAST RED KIT

1. Dilute CK 8/18 1:100 Van Gogh Yellow (Biocare Medical). Incubate for 30 minutes.
2. Wash in two changes of 1X Immunocare TBS (Biocare Medical) for 2 minutes each.
3. Apply four drops of Goat anti-Mouse-ALP (Polymer). Incubate for 20 minutes.
4. Wash in two changes of 1X Immunocare TBS (Biocare Medical) wash Buffer for two minutes each. Drain Slide.
5. Apply 4 drops of Vulcan Fast Red Chromogen Substrate (Biocare Medical) for 10 minutes. Rinse in deionized water. Place specimen under microscope and check for light staining (manual method only). If the stain is too light, apply Vulcan Fast Red Substrate for another 5 minutes and rinse again in deionized water.
6. Add 4 drops of CAT Hematoxylin for 30-60 seconds. Wash in tap water.
7. Blue nuclei in 1X PBS wash buffer for 1 minute. Drain slides.
8. Wash in tap water and rinse in deionized water.
9. Dehydrate in 3 changes of 100% alcohol and clear in 3 changes of xylene.
10. Mount and coverslip.

Example 3**Tissue Preparation.**

All tissues were formalin-fixed paraffin-embedded and sectioned at 4 microns. Section were dried at 37°C for 30 minutes and then dried at 60°C for 30 minutes. Sections were immediately taken out of the oven and deparaffinized in Slide Brite, (BIOCARE) and then hydrated through a grades series of alcohol. All tissues were placed in a 3% hydrogen peroxide bulk solution for 5 minutes to quench endogenous peroxidase activity. The following tissues were used for double and triple stains: tonsil, colon cancer, liver cancer, breast cancer, dermatofibroma, mesothelioma and melanoma.

Reagents and Equipment

Antigen retrieval was performed using Reveal (RV) (pH 6.0) and BORG Decloaker (BD) (pH, 9.5) in a Decloaking Chamber (pressure cooker) for 30 seconds at 125°C and 20 p.s.i. (BIOCARE Medical). For microarray tissues, antigen retrieval was performed overnight at 75°C and placed in RV or Nuclear Decloaker (pH 9.5) (BIOCARE Medical). Antibody cocktails are listed in Table 1. All antibodies were diluted for optimum titer, followed by biotin-free polymer detection kits and reagents (Table 2). TBS wash buffer was used on the following automated stainers: AutoStainer (Dako), i6000 BioGenex, Nemesis (BIOCARE Medical).

Table 1

Antibody Cocktail	Host	Clone
ApoptoticCK18	Mouse	BC/M30
Ki-67	Rabbit	SP91
CD34	Mouse	Biocare
Factor XIIIa	Rabbit	Polyclonal
L26	Mouse	L26
CD3	Rabbit	Polyclonal
Kappa	Mouse	KDB-1
Lambda	Rabbit	Polyclonal

CK5	Mouse	XM26
P63	Mouse	BC4A4
CK8/18	Mouse	5D3
CK5	Mouse	XM26
CK17	Mouse	Ks 17.E3
CK8/18	Mouse	5D3
HMW CK	Mouse	DE-SQ
p63	Mouse	BC4A4
P504S	Rabbit	P504S
MART-1	Mouse	M2-7C10 + M2-9E3
Tyrosinase	Mouse	T311
S100	Rabbit	Polyclonal
Ki-67	Mouse	DVB-1
Caspase-3	Rabbit	Polyclonal
CK5/6	Mouse	CK5/6.007
Calretinin	Rabbit	SP19
CD10	Mouse	56C6 (HSA)
PSA	Rabbit	Polyclonal
HSA	Mouse	OCH1E5

Antibody cocktails were obtained from Biocare Medical

Table 2

<u>Name</u>	<u>Kit or secondary reagents</u>
Double Stain Detection Kit #1	Goat anti-mouse alkaline phosphatase (AP) Goat anti-rabbit horseradish peroxidase (HRP)
Double Stain Detection Kit #2	Goat anti-mouse HRP Goat anti-rabbit AP
MACH 2™ Mouse-HRP	Goat anti- mouse (HRP)
MACH 2™ Mouse-AP	Goat anti- mouse (AP)
MACH 2™ Rabbit-HRP	Goat anti-rabbit (HRP)
MACH 2™ Rabbit-AP	Goat anti-rabbit (AP)

Double stains kits, reagents and chromogens were obtained from Biocare Medical

Table 3

<u>Antibody Cocktail</u>	<u>Pretreatment</u>	<u>Detection</u>	<u>Chromogens</u>
Apoptotic/CK18 + Ki-67	BORG Decloaker™	DS Kit #1	DAB and Fast Red
CD34 + Factor XIIIa	Trypsin	DS Kit #2	DAB and Fast Red
L26 + CD3	Reveal	DS Kit #1	DAB and Fast Red
Kappa + Lambda	Reveal	DS Kit #2	DAB and Fast Red
CK5 + CK17 + CK8/18	Reveal	DS Kit #2	DAB and Fast Red
CK5 + p63 + CK8/18	Reveal	DS kit #2 + MACH 2 - HRP	DAB and Fast Red
HMW CK + p63 + P504S	BORG Decloaker	DS Kit #2	DAB and Fast Red

MART-1 + Tyrosinase + S100	Reveal	DS Kit #2	DAB and Fast Red
Ki-67 + Caspase-3	BORG Decloaker	DS Kit #2	DAB and Fast Red
CK5/6 + Calretinin	Reveal	DS Kit #2	DAB and Fast Red
CD10 + PSA + HSA	Reveal	DS Kit #2 + M2 Mouse -HRP	B. Purple and Fast Red DAB
LCA + S100 + CK8/18	Reveal	DS Kit #2 + M2 Rabbit -HRP	DAB and Fast Red Bajoran Purple

*Double stains kits, reagents, antibodies, and chromogens were obtained from Biocare Medical

All of the above reagents were tested on tissues mounted on slides and exhibited good double and triple staining patterns.

Obviously, numerous modifications and variations of the present invention are possible in light of the above teachings. It is therefore to be understood that within the scope of the appended claims, the invention may be practiced otherwise than as specifically described herein.

CLAIMS:

1. A composition comprising at least two antibodies and a buffer selected from the group consisting of:

- a. 1.5 to 3.0 g/l sodium phosphate monobasic, monohydrate; 0.5 to 0.6 g/l potassium phosphate dibasic, trihydrate; 0.5 to 1.5 ml/l polyoxyethylenesorbitan monolaurate, 50 to 100 ml/l of 5% bovine serum albumin, 0.5 to 1.5 g/l sodium azide, 0.005 g/l, and water and which is at a pH of from about 5.5 to about 6.5;
- b. 40 to 60 ml/l 10X PBS, 0.75 to 1.25 g/l sodium azide, 0.1 to 0.3 ml/l polyoxyethylenesorbitan monolaurate, 45 to 65 g/l bovine serum albumin, and water and which is at a pH of about 7.0 to about 6.5;
- c. 3.0 to 4.0 g/l Tris-hydrochloride, 0.75 to 1.2 g/l sodium azide, 7.5 to 12.5 g/l bovine serum albumin, 0.2 to 0.3 ml/l 25 % hydrochloric acid, and water and which is at a pH of about 5.7 to about 6.5;
- d. one part of a first solution comprising 75 to 125 ml/l 10 X PBS, 2.5 ml/l preservative, 0.25 to 0.75 ml/l of 50% polyoxyethylenesorbitan monolaurate, 2.5 to 7 g/l purified casein, 2.0 to 3.0 ml/l purified Type A gelatin, and water, and a second solution comprising 2.0 to 3.0 g/l sodium phosphate monobasic, monohydrate; 0.5 to 0.6 g/l potassium phosphate dibasic, trihydrate; 0.75 to 1.25 ml/l polyoxyethylenesorbitan monolaurate, 75 to 125 ml/l of 2.5 to 7.5 % bovine serum albumin, 0.7 to 1.1 g/l sodium azide, and water, and which is at a pH of about 5.75 to about 6.25; and
- e. 1.5 to 3.0 g/l sodium phosphate monobasic, monohydrate; 0.5 to 0.6 g/l potassium phosphate dibasic, trihydrate; 0.5 to 1.5 ml/l polyoxyethylenesorbitan monolaurate, 50 to 100 ml/l of 5% bovine serum albumin, 0.5 to 1.5 g/l sodium azide,

0.005 g/l, 40 to 60% glycerol, and water, and which is at a pH is about 5.5 to about 6.5.

2. The composition of Claim 1, wherein the buffer is a.

3. The composition of Claim 2, wherein the buffer comprises 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l polyoxyethylenesorbitan monolaurate, 100 ml/l of 5% bovine serum albumin, 0.9 g/l sodium azide, and water and which is at a pH of about 6.0.

4. The composition of Claim 1, wherein the buffer is b.

5. The composition of Claim 4, wherein the buffer comprises 50 ml/l 10X PBS, 0.9 g/l sodium azide, 0.2 ml/l polyoxyethylenesorbitan monolaurate, 55 g/l bovine serum albumin, and water, and which is at a pH of about 7.3.

6. The composition of Claim 1, wherein the buffer is c.

7. The composition of Claim 6, wherein the buffer comprises 3.5 g/l Tris-HCl, 0.9 g/l sodium azide, 10 g/l bovine serum albumin, 0.25 ml/l 25 % hydrochloric acid, and water, and which is at a pH of about 6.2.

8. The composition of Claim 1, wherein the buffer is d.

9. The composition of Claim 8, which is at a pH of about 6.0, and wherein the first solution comprises 100 ml/l 10 X PBS, 2.5 ml/l preservative, 0.5 ml/l of 50% polyoxyethylenesorbitan monolaurate, 5 g/l purified casein, 2.5 ml/l purified Type A gelatin, and water; and the second solution comprises 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l polyoxyethylenesorbitan monolaurate, 100 ml/l of 5% bovine serum albumin, 0.9 g/l sodium azide, and water.

10. The composition of Claim 1, wherein the buffer is e.

11. The composition of Claim 10, wherein the buffer comprises 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l polyoxyethylenesorbitan monolaurate, 100 ml/l of 5% bovine serum albumin, 0.9 g/l sodium azide, 50% glycerol, and water, wherein the pH is about 6.0.

12. The composition of Claim 1, wherein at least one antibody is from a different species than at least a second antibody.

13. The composition of Claim 1, wherein at least one antibody is a rabbit antibody.

14. The composition of Claim 1, wherein the at least one rabbit antibody is a rabbit monoclonal antibody.

15. A method of detecting at least two antigens in a sample, comprising contacting the sample with the composition of Claim 1; and detecting the formation of at least two antigen-antibody complexes formed on the sample.

16. The method of Claim 15, wherein the sample is attached to a solid support.

17. The method of Claim 16, wherein the solid support is a glass slide, glass dish, plastic dish, glass well, or plastic well.

18. The method of Claim 16, which further comprises prior to contacting the sample, treating the sample at a temperature of from 65 to 80°C for a time sufficient to adhere the sample to the solid support.

19. The method of Claim 16, wherein the temperature is about 75°C.

20. The method of Claim 15, wherein the method is conducted on an automated staining device.

21. The method of Claim 15, wherein at least one antibody is a rabbit antibody.

22. The method of Claim 21, wherein the at least one rabbit antibody is a rabbit monoclonal antibody.

23. The method of Claim 15, wherein detecting the formation of at least two antigen-antibody complexes comprises contacting the sample with a second composition, which comprises at least two antibodies, which specifically bind to the at least two antibodies in the composition wherein at least one antibody in the second composition is coupled to a poly (horseradish peroxidase) and at least a second antibody in the second composition is coupled to a poly (alkaline phosphatase) and wherein the second composition comprises 0.8 to 1.2 M Tris-hydrochloride, pH from 7.3 to 7.9, 0.025 to 0.075% polyoxyethylenesorbitan monolaurate and 2.5 to 3.5 % goat serum; and visualizing at least two antigen-antibody complexes on the sample.

24. The method of Claim 23, wherein the second composition further comprises from 3.0 to 3.7 ml/l tris-buffered saline.

25. The method of Claim 23, wherein the second composition further comprises from 3.4 ml/l tris-buffered saline.

26. The method of Claim 23, wherein the second composition comprises 0.1M Tris-hydrochloride, pH 7.6, 0.05% polyoxyethylenesorbitan monolaurate, and 3% goat serum.

27. The method of Claim 23, wherein visualizing at least two antigen-antibody complexes on the sample comprising adding at least one chromogen to the sample.

28. A kit comprising the antibody composition of Claim 1 and one or more reagents to detect an antibody-antigen complex.

29. The kit of Claim 28, wherein the buffer is a and comprises 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l polyoxyethylenesorbitan monolaurate, 100 ml/l of 5% bovine serum albumin, 0.9 g/l sodium azide, and water and which is at a pH of about 6.0.

30. The kit of Claim 28, wherein the buffer is b and comprises 50 ml/l 10X PBS, 0.9 g/l sodium azide, 0.2 ml/l polyoxyethylenesorbitan monolaurate, 55 g/l bovine serum albumin, and water, and which is at a pH of about 7.3.

31. The kit of Claim 28, wherein the buffer is c and comprises 3.5 g/l Tris-HCl, 0.9 g/l sodium azide, 10 g/l bovine serum albumin, 0.25 ml/l 25 % hydrochloric acid, and water, and which is at a pH of about 6.2.

32. The kit of Claim 28, wherein the buffer is d and which is at a pH of about 6.0, and wherein the first solution comprises 100 ml/l 10 X PBS, 2.5 ml/l preservative, 0.5 ml/l of 50% polyoxyethylenesorbitan monolaurate, 5 g/l purified casein, 2.5 ml/l purified Type A gelatin, and water; and the second solution comprises 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l polyoxyethylenesorbitan monolaurate, 100 ml/l of 5% bovine serum albumin, 0.9 g/l sodium azide, and water.

33. The kit of Claim 28, wherein the buffer is e and comprises 2.4g/l sodium phosphate monobasic, monohydrate; 0.56g/l potassium phosphate dibasic, trihydrate; 1 ml/l polyoxyethylenesorbitan monolaurate, 100 ml/l of 5% bovine serum albumin, 0.9 g/l sodium azide, 50% glycerol, and water, wherein the pH is about 6.0.

34. The kit of Claim 28, wherein at least one antibody is a rabbit antibody.

35. The kit of Claim 34, wherein the at least one rabbit antibody is a rabbit monoclonal antibody.

36. The kit of Claim 28, which further comprises a second composition comprising at least two antibodies wherein at least one antibody is coupled to a poly (horseradish peroxidase) and at least a second antibody is coupled to a poly(alkaline phosphatase) and a buffer comprising 0.8 to 1.2 M Tris-hydrochloride, pH from 7.3 to 7.9, 0.025 to 0.075% polyoxyethylenesorbitan monolaurate and 2.5 to 3.5 % goat serum.

37. The kit of Claim 36, wherein the second composition further comprises from 3.0 to 3.7 ml/l tris-buffered saline.

38. The kit of Claim 36, wherein the second composition further comprises from 3.4 ml/l tris-buffered saline.

39. The kit of Claim 36, wherein the buffer in the second composition comprises 0.1M Tris-hydrochloride, pH 7.6, 0.05% polyoxyethylenesorbitan monolaurate, and 3% goat serum.

40. A composition comprising at least two antibodies wherein at least one antibody is coupled to a poly (horseradish peroxidase) and at least a second antibody is coupled to a poly(alkaline phosphatase) and a buffer comprising 0.8 to 1.2 M Tris-hydrochloride, pH from 7.3 to 7.9, 0.025 to 0.075% polyoxyethylenesorbitan monolaurate and 2.5 to 3.5 % goat serum.

41. The composition of Claim 40, which further comprises from 3.0 to 3.7 ml/l tris-buffered saline.

42. The composition of Claim 40, which further comprises from 3.4 ml/l tris-buffered saline.

43. The composition of Claim 40, wherein the buffer comprises 0.1M Tris-hydrochloride, pH 7.6, 0.05% polyoxyethylenesorbitan monolaurate, and 3% goat serum.

44. The composition of Claim 43, which further comprises from 3.0 to 3.7 ml/l tris-buffered saline.

45. The composition of Claim 40, which further comprises 3.4 ml/l tris-buffered saline.

46. A method of detecting at least two antigens in a sample, comprising contacting the sample with at least one first antibody that binds to a first antigen and at least one second antibody that binds to a second antigen; contacting the sample with the composition of Claim

40, wherein at least one antibody in the composition binds specifically to the first antibody and another antibody in the composition binds to the second antibody; and detecting the formation of at least two antigen-antibody complexes formed in the sample.

47. The method of Claim 46, wherein the buffer in the composition further comprises from 3.0 to 3.7 ml/l tris-buffered saline.

48. The method of Claim 46, wherein the buffer further comprises 3.4 ml/l tris-buffered saline.

49. The method of Claim 46, wherein the buffer comprises 0.1M Tris-hydrochloride, pH 7.6, 0.05% polyoxyethylenesorbitan monolaurate, and 3% goat serum.

50. The method of Claim 46, wherein the method is conducted on an automated staining device.

51. The method of Claim 46, wherein one of the first or second antibodies is a rabbit antibody.

52. The method of Claim 52, wherein the rabbit antibody is a rabbit monoclonal antibody.

53. The method of Claim 46, wherein the sample is attached to a solid support.

54. The method of Claim 53, wherein the solid support is a glass slide, glass dish, plastic dish, glass well, or plastic well.

55. The method of Claim 53, wherein prior to contacting the sample with the first and second antibody, the method comprises treating the sample at a temperature of from 65 to 80°C for a time sufficient to adhere the sample to the solid support.

56. The method of Claim 55, wherein the temperature is about 75°C.

57. A kit comprising the antibody composition of Claim 40, and one or more reagents to detect an antibody-antigen complex.

58. The kit of Claim 57, wherein the buffer further comprises from 3.0 to 3.7 ml/l tris-buffered saline.

59. The kit of Claim 58, wherein the buffer further comprises from 3.4 ml/l tris-buffered saline.

60. The kit of Claim 58, wherein the buffer comprises 0.1M Tris-hydrochloride, pH 7.6, 0.05% polyoxyethylenesorbitan monolaurate, and 3% goat serum.

61. A method of detecting at least two antigens in a sample, comprising

contacting a sample with a first antibody;

denaturing the sample with a composition comprising 1 part of a first solution which comprises 1.1 to 1.3 % hydrochloric acid, 0.020 to 0.030 % preservative, and water; and 3 parts of a second solution comprising 0.1 to 0.3 %

polyoxyethylenesorbitan monolaurate, about 0.2 to 0.3 % preservative, and water;

contacting the sample with a second antibody; and

visualizing the formation of at least two antigen-antibody complexes on the sample.

62. The method of Claim 61, wherein the first solution comprises 1.2 % hydrochloric acid, 0.025 % preservative, and water.

63. The method of Claim 61, wherein the second solution comprises 0.2% polyoxyethylenesorbitan monolaurate, 0.25% preservative, and water.

64. The method of Claim 61, wherein the sample is attached to a solid support.

65. The method of Claim 64, wherein the solid support is a glass slide, glass dish, plastic dish, glass well, or plastic well.

66. The method of Claim 64, which prior to contacting the sample with the first antibody, the method further comprises treating the sample at a temperature of from 65 to 80°C for a time sufficient to adhere the sample to the solid support.

67. The method of Claim 66, wherein the temperature is about 75°C.

68. The method of Claim 61, wherein the method is conducted on an automated staining device.

69. The method of Claim 61, wherein at least one antibody is a rabbit antibody.

70. The method of Claim 69, wherein the at least one rabbit antibody is a rabbit monoclonal antibody.

71. A method of detecting at least two antigens in a sample, comprising contacting the sample with a composition comprising at least one first primary antibody and at least one second primary antibody; and detecting the formation of at least two antigen-antibody complexes on the sample, wherein the at least one first and second antibodies specifically bind to two antigens selected from the group consisting of:

CD3 and Caspase-3;

CD20 and CD3;

CD31 and Ki-67;

CD34 and Factor XIII subunit a;

CDX2 and CK7;

Ki-67 and Caspase-3;

M30 and Ki-67;

LCA and S-100;

CD20 and Ki-67;

Tyrosinase and S100;

Tyrosinase and MART-1;

Tyrosinase and A103;

P63 and CK5;

P63 and P504S;

P63 and P504S;
CK5/6 and Calretinin;
Estrogen receptor and Ki-67;
CK5 and CK17;
CD10 and Prostate specific antigen;
CD10 and Hepatic specific antigen;
Chromogranin A and Synaptophysin;
HMW CK and LMW CK;
CD20 and Caspase-3;
CD3 and Ki-67;
PAX-5 and CD5;
CD4 and CD8;
Kappa light chain and lambda light chain.

72. The method of Claim 71, which comprises detecting at least three antigens in a sample and wherein the method comprises contacting the sample, after detecting the formation of the antigen-antibody complex, with at least a third primary antibody; and detecting the formation of at least third antibody-antigen complex on the sample.

73. The method of Claim 72, which is performed on an automated staining device.

74. The method of Claim 71, which comprises detecting at least four antigens in a sample and wherein the method comprises contacting the sample, after detecting the formation of the antigen-antibody complexes, with at least a third and a fourth primary antibody; and detecting the formation of at least third and fourth antibody-antigen complexes on the sample.

75. The method of Claim 74, wherein one of the third or fourth primary antibodies is a rabbit antibody.

76. The method of Claim 75, wherein one of the third or fourth primary antibodies is a rabbit monoclonal antibody.

77. The method of Claim 74, which is performed on an automated staining device.

78. The method of Claim 71, wherein one of the first or second primary antibodies is a rabbit antibody.

79. The method of Claim 78, wherein one of the first or second primary antibodies is a rabbit monoclonal antibody.

80. The method of Claim 71, which is performed on an automated staining device.

81. A method of detecting two or more antigens in a sample, comprising contacting a sample, which has been previously contacted with a primary antibody cocktail comprising at least one first primary antibody and at least one second primary antibody, with a composition comprising at least one first secondary antibody and at least one second secondary antibody, wherein the at least one first secondary antibody is coupled to a poly (alkaline phosphatase) moiety and the at least one second secondary antibody is coupled to a poly (horseradish peroxidase) moiety, and wherein the composition comprises a buffer suitable to stabilize the first and second secondary antibodies; and

detecting the formation of at least two antigen-antibody complexes on the sample.

82. The method of Claim 81, which is performed on an automated staining device.

83. The method of Claim 81, wherein at least three antigens are detected in the sample and the method further comprises contacting the sample, after detecting the formation of the at least two antigen-antibody complexes on the sample, with at least a third primary antibody; and detecting the formation of at least a third antibody-antigen complex on the sample.

84. The method of Claim 83, which is performed on an automated staining device.

85. The method of Claim 81, which comprises detecting at least four antigens in a sample and wherein the method comprises contacting the sample, after detecting the formation of the antigen-antibody complexes, with at least a third and a fourth primary antibody; and detecting the formation of at least third and fourth antibody-antigen complexes on the sample.

86. The method of Claim 85, which is performed on an automated staining device.

87. A method of detecting two or more antigens in a sample, comprising contacting a sample with a primary antibody cocktail comprising at least one first primary antibody and at least one second primary antibody, and subsequently contacting the sample with a composition comprising at least one first secondary antibody and at least one second secondary antibody, wherein the at least one first secondary antibody is coupled to a poly (alkaline phosphatase) moiety and the at least one second secondary antibody is coupled to a poly (horseradish peroxidase) moiety, and wherein the composition comprises a buffer suitable to stabilize the first and second secondary antibodies; and

detecting the formation of at least two antigen-antibody complexes on the sample.

88. The method of Claim 87, which is performed on an automated staining device.

89. The method of Claim 87, wherein at least three antigens are detected in the sample and the method further comprises contacting the sample, after detecting the formation of the at least two antigen-antibody complexes on the sample, with at least a third primary antibody; and detecting the formation of at least a third antibody-antigen complex on the sample.

90. The method of Claim 89, which is performed on an automated staining device.

91. The method of Claim 87, which comprises detecting at least four antigens in a sample and wherein the method comprises contacting the sample, after detecting the formation of the antigen-antibody complexes, with at least a third and a fourth primary

antibody; and detecting the formation of at least third and fourth antibody-antigen complexes on the sample.

92. The method of Claim 91, which is performed on an automated staining device.

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US2005/005570

A. CLASSIFICATION OF SUBJECT MATTER IPC 7 H01L31/101				
According to International Patent Classification (IPC) or to both national classification and IPC				
B. FIELDS SEARCHED				
Minimum documentation searched (classification system followed by classification symbols) IPC 7 H01L				
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched				
Electronic data base consulted during the international search (name of data base and, where practical, search terms used) EPO-Internal, INSPEC				
C. DOCUMENTS CONSIDERED TO BE RELEVANT				
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No		
A	OH J ET AL: "INTERDIGITATED GE P-I-N PHOTODETECTORS FABRICATED ON A SI SUBSTRATE USING GRADED SIGE BUFFER LAYERS" IEEE JOURNAL OF QUANTUM ELECTRONICS, IEEE INC. NEW YORK, US, vol. 38, no. 9, September 2002 (2002-09), pages 1238-1241, XP001175548 ISSN: 0018-9197 the whole document ----- -/--	1-62		
<input checked="" type="checkbox"/> Further documents are listed in the continuation of box C <input checked="" type="checkbox"/> Patent family members are listed in annex				
* Special categories of cited documents				
<table style="width: 100%; font-size: x-small;"> <tr> <td style="width: 50%; vertical-align: top;"> *A* document defining the general state of the art which is not considered to be of particular relevance *E* earlier document but published on or after the international filing date *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) *O* document referring to an oral disclosure, use, exhibition or other means *P* document published prior to the international filing date but later than the priority date claimed </td> <td style="width: 50%; vertical-align: top;"> *T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention *X* document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone *Y* document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art *&* document member of the same patent family </td> </tr> </table>			*A* document defining the general state of the art which is not considered to be of particular relevance *E* earlier document but published on or after the international filing date *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) *O* document referring to an oral disclosure, use, exhibition or other means *P* document published prior to the international filing date but later than the priority date claimed	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention *X* document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone *Y* document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art *&* document member of the same patent family
A document defining the general state of the art which is not considered to be of particular relevance *E* earlier document but published on or after the international filing date *L* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) *O* document referring to an oral disclosure, use, exhibition or other means *P* document published prior to the international filing date but later than the priority date claimed	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention *X* document of particular relevance, the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone *Y* document of particular relevance, the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art *&* document member of the same patent family			
Date of the actual completion of the international search <p style="text-align: center; font-size: large;">10 August 2005</p>		Date of mailing of the international search report <p style="text-align: center; font-size: large;">18/08/2005</p>		
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel (+31-70) 340-2040, Tx. 31 651 epo nl, Fax (+31-70) 340-3016		Authorized officer <p style="text-align: center; font-size: large;">Boero, M</p>		

INTERNATIONAL SEARCH REPORT

International Application No
PCT/US2005/005570

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No
A	<p>JONES R E ET AL: "Fabrication and modeling of gigahertz photodetectors in heteroepitaxial Ge-on-Si using a graded buffer layer deposited by low energy plasma enhanced CVD" INTERNATIONAL ELECTRON DEVICES MEETING 2002. IEDM. TECHNICAL DIGEST. SAN FRANCISCO, CA, DEC. 8 - 11, 2002, NEW YORK, NY : IEEE, US, 8 December 2002 (2002-12-08), pages 793-796, XP010626159 ISBN: 0-7803-7462-2 the whole document</p>	1-62
A	<p>US 2003/141565 A1 (HIROSE FUMIHIKO ET AL) 31 July 2003 (2003-07-31) the whole document</p>	1-62

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No
PCT/US2005/005570

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 2003141565	A1	31-07-2003	NONE