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(54) Title: MEDICAL DEVICES COATED WITH SHAPE MEMORY POLYMER FOAMS

(57) Abstract: An embodiment includes a system comprising: a substrate of a medical device; an un-foamed polyurethane coating directly contacting the substrate and fixedly attached to the substrate; a thermoset polyurethane shape memory polymer (SMP) foam, having first and second states, which directly contacts the polyurethane coating and fixedly attaches to the polyurethane coating; wherein the polyurethane coating fixedly attaches the SMP foam to the substrate. Other embodiments are described herein.

MEDICAL DEVICES COATED WITH SHAPE MEMORY POLYMER FOAMSStatements As To Rights To Inventions Made  
Under Federally Sponsored Research And Development

[0001] This invention was made with government support under R01EB000462 awarded by National Institutes of Health, National Institute of Biomedical Imaging and Bioengineering. The government has certain rights in the invention.

Priority Claim

[0002] This application claims priority to United States Provisional Patent Application No. 62/197,181 filed on July 27, 2015 and entitled "Medical Devices Coated With Shape Memory Polymer Foams", the content of which is hereby incorporated by reference.

Background

[0003] A cerebral aneurysm is a weakening of the neurovascular wall that allows localized dilation and ballooning of the artery. Aneurysms are susceptible to rupturing due to wall thinning, which causes an intracranial hemorrhage. It is estimated that 30,000 aneurysms rupture in the United States every year and approximately 3-5 million people in America have or will develop an aneurysm in their lifetime. Current aneurysm treatments include surgical clipping and the endovascular delivery of platinum coils to embolize the aneurysm. Surgical clipping includes a craniotomy and although proven effective, is extremely invasive. An endoscopic endonasal approach for aneurysm clipping has been successful, but it is also fairly invasive and requires pedicled nasoseptal flap reconstruction after the aneurysm clipping.

[0004] Endovascular embolization is now the preferred treatment for many patients. The current standard for embolization utilizes platinum coils to occlude the aneurysm and promote thrombogenesis. Issues with platinum coils include material cost and a significant incidence of recanalization and retreatments due to coil packing over time. Hydrogel coated platinum wires were developed to reduce aneurysm recurrence rates and are the intermediary device between the platinum coil embolism treatments and shape memory polymer (SMP) coated treatments.

### Brief Description Of The Drawings

[0005] Features and advantages of embodiments of the present invention will become apparent from the appended claims, the following detailed description of one or more example embodiments, and the corresponding figures. Where considered appropriate, reference labels have been repeated among the figures to indicate corresponding or analogous elements.

[0006] Figure 1 includes a bulk foam in an embodiment.

[0007] Figure 2 includes vertically oriented substrates in an embodiment (i.e., substrate long axis located generally parallel to path of expanding foam).

[0008] Figure 3 includes a polymer coated substrate in an embodiment.

[0009] Figure 4 shows a top view (with foam rising towards viewer of figure) of void caused by a horizontally oriented substrate (i.e., substrate long axis located generally orthogonal to path of expanding foam).

[0010] Figure 5 includes a foam over wire (FOW) in an embodiment.

[0011] Figure 6 addresses pore size for FOW and bulk foam in an embodiment.

[0012] Figure 7 addresses dry volume expansion for FOW and bulk foam in an embodiment.

[0013] Figure 8 includes a process in an embodiment.

### Detailed Description

[0014] Reference will now be made to the drawings wherein like structures may be provided with like suffix reference designations. In order to show the structures of various embodiments more clearly, the drawings included herein are diagrammatic representations of structures. Thus, the actual appearance of the structures, for example in a photomicrograph, may appear different while still incorporating the claimed structures of the illustrated embodiments. Moreover, the drawings may only show the structures useful to understand the illustrated embodiments. Additional structures known in the art may not have been included to maintain the clarity of the drawings. "An embodiment", "various embodiments" and the like indicate

embodiment(s) so described may include particular features, structures, or characteristics, but not every embodiment necessarily includes the particular features, structures, or characteristics. Some embodiments may have some, all, or none of the features described for other embodiments. “First”, “second”, “third” and the like describe a common object and indicate different instances of like objects are being referred to. Such adjectives do not imply objects so described must be in a given sequence, either temporally, spatially, in ranking, or in any other manner. “Connected” may indicate elements are in direct physical or electrical contact with each other and “coupled” may indicate elements co-operate or interact with each other, but they may or may not be in direct physical or electrical contact.

[0015] Applicant previously practiced a protocol for attaching SMP foams to a backbone wire. The protocol was to mechanically crimp and adhere the foam directly to the wire at both ends using epoxy, thus setting the secondary shape of the SMP around the wire. Using this process, Applicant determined a number of problems. Applicant determined the foams had a tendency to slip along the wire (if no epoxy or ineffective epoxy was used) and cause multiple problems during delivery of the device to the aneurysm site. Further, Applicant determined the use of epoxy could be detrimental to the device as a whole because of the possibility for a lack of biocompatibility. Additionally, Applicant determined the use of adhesive increases the minimum compressed diameter of the foam over the wire (which may complicate delivery of the foam into small areas, such as a patient’s vasculature). In general, the process of adhering previously synthesized foam to a wire was problematic.

[0016] However, an embodiment includes a neurovascular embolism device that addresses these shortcomings. The embodiment uses ultra-low density polyurethane SMP foam adhered to a, for example, nitinol or polymer suture backbone to occlude the aneurysm. SMPs are a class of smart materials that are capable of deforming to a stable secondary shape (programmed state) and recovering to a primary form or state when exposed to a particular stimulus, such as heat. This shape memory property allows for the delivery of a compressed foam to the aneurysm site via micro-catheter and for consistent, high volume recoveries once positioned. The embodiment includes a process for attaching SMP foam to a

backbone wire by synthesizing the foam directly over the nitinol wire or polymer suture backbone, while holding the current properties of the foam constant in the immediate vicinity of the wire. The specimen is then cut to the desired diameter using an Excimer Laser, resistive wire cutter, or biopsy punch. This eliminates the need to optimize the process of crimping the foam to the wire and alleviates multiple issues within the device design and fabrication, including foam slippage and detachment from the coil, and the use of an epoxy like adhesive.

[0017] In an embodiment a coating is added to a nitinol wire in order to facilitate the placement of the SMP foam to the wire. The embodiment aids in the production of a functional embolic device treatment using SMP foams synthesized directly onto a nitinol wire and shaped using an Excimer Laser. The axial length of the foam/wire device is approximately 12 cm in an embodiment but other embodiments include wires of 2, 5, 10, 15, 20, 25, 30 cm or more.

[0018] Embodiments are directed to a porous coating for a wire, wherein the porous coating comprises a porous, shape memory material. In certain embodiments, the porous coating is formed by coating the wire with a shape memory material and then curing the material to the wire. A further embodiment is directed to a medical device comprising a body and a porous coating secured to a surface of the body, wherein the porous coating comprises a porous, shape memory material. In certain embodiments, the porous coating is adapted to volumetrically expand so as to fill gaps between the coil and the wall of the blood vessel and to create volumetric occlusion between the medical device and the wall of the blood vessel. In such an embodiment, volumetric filling is achieved by the shape memory effect of the shape memory material, which causes blood flow stasis and embolization of vascular anatomy.

[0019] Another embodiment is directed to a method for providing therapy to a patient, the method comprising: providing a medical device comprising a body and a porous coating secured to a surface of the body, wherein the porous coating comprises a porous, shape memory material; inserting the medical device into the patient so that the porous coating volumetrically expands so as to fill gaps between the device body and the wall of the adjacent tissue to create volumetric filling

between the medical device and the wall of the adjacent tissue. The interface of the porous coating and adjacent tissue is meant to promote cellular ingrowth to integrate and anchor the device to the tissue.

[0020] An embodiment is directed to the application of a SMP foam to a nitinol wire using a direct foaming process. In accordance with a process in an embodiment, the nitinol wire is coated with an optimized coating for proper foam adhesion, wetting, and stabilization of pores during the foaming process. The coatings used are selected from, but not limited to, the group consisting of a silicone coating, a hydrophilic coating (e.g., siloxane), a polytetrafluoroethylene (PTFE) coating, a thermoplastic polyurethane, and a thermoset neat polyurethane with a bulk chemical composition as the foam. The wires may be acid-etched or unetched prior to coating.

[0021] In an embodiment the coated wires are subjected to direct foaming by exposing the coated wires to a reaction mixture comprising at least one shape memory foam polymer and at least one foaming agent. In certain embodiments, the foaming agent is a blowing agent.

[0022] In an embodiment the reaction mixture is transferred to a container or vessel containing the coated wires. In certain embodiments, the coated wires are oriented axially to the direction of application of the foam. In other embodiments, the coated wires are oriented transversally to the direction of application of the foam.

[0023] Shape memory foam-coated wires of embodiments possess pore sizes and densities comparable to previously created foams. The ratio of bulk foam to FOW foam, excluding the mass of the substrate wire, is nearly one in many cases, indicating little to no change in the ultra low-density nature of the polymers. FOW pore sizes are comparable to their bulk foam counterparts, indicating little change in the nature of the foams.

## WORKING EXAMPLES METHODS

### FOAM SYNTHESIS

[0024] An established protocol for the synthesis of ultra low-density polyurethane SMP foams was used to create foam for device use. See Singhal et al (Controlling the Actuation Rate of Low-Density Shape-Memory Polymer Foams in Water; *Macromol. Chem. Phys.* 2013, 214, 1204–1214). An embodiment for foam synthesis involves combining the alcohols triethanolamine (TEA, Sigma Aldrich, St. Louis MO), and N, N, N', N'- Tetrakis (2-hydroxypropyl) ethylenediamine (HPED, Sigma Aldrich, St. Louis MO), in a ratio of 0.32-0.35 to isocyanates to form an NCO premix, or prepolymer. Isophorone diisocyanate (IPDI, TCI America, Tokyo, Japan) with a racemic mixture of 2,2,4-trimethyl-1,6-hexamethylene diisocyanate and 2,4,4-trimethyl-1,6-hexamethylene diisocyanate (TMHDI TCI America, Tokyo, Japan) was used in varying compositions to create the polyurethane (PU) foams.

[0025] The range of chemical compositions for foam synthesis used in this experiment can be seen in Table 1.

Table 1: Isocyanate content of the five chemical compositions used in this experiment

Chemical Composition	1	2	3	4	5
IPDI content	0%	5%	10%	15%	20%
TMHDI Content	100%	95%	90%	85%	80%

[0026] An isocyanate premix of 32-35% alcohols was allowed to cure for two days in a programmed oven, in which the temperature was ramped to 50°C and subsequently allowed to cool to room temperature. An alcohol premix, containing the balance of remaining OH monomers was created immediately before foaming to react with the excess isocyanates with a 0.96 OH/NCO ratio. The alcohol premix also contained catalysts BL-22 and T-131 as well as the surfactant, DC 5943, for pore stability. Water was added to the OH premix to serve as a chemical blowing agent for the foaming process. Table 2 shows the exact masses of agents added.

Table 2: Exact amounts of accessory chemicals for foaming process of all foam compositions created in this work

	DC 5943	BL-22	T-131	Enovate	Water
Target for one foam	2.8g	0.253g	0.101g	7mL	0.917g

[0027] The OH premix and the NCO premix were combined and mixed for 10 seconds in a speed mixer (SpeedMixer DAC 150 FVZ-K, Flacktek, Landrum, SC), after which the physical blowing agent, Enovate, was added and the polymer was mixed again for 10 additional seconds. The reaction mixture was carefully transferred into a large bucket containing the coated wires, which were previously positioned in the bucket, and placed in an oven. As the foam reacted, it experienced about 30x volume expansion and took the primary shape of the container it was reacting in. The foam was allowed to react for 20 minutes at 90°C, after which time the foam was removed from the oven and allowed to cool to room temperature under a fume hood.

[0028] Once cooled, the buckets were slightly deformed to release the foams from the container. The wires attached through small holes in the bottom were released, the tape discarded. At this point the foams were simply lifted out of the bucket. The foams were cut in half using the Proxxon Thermocut Hotwire Cutter.

#### COATING OPTIMIZATION AND ADHESION

[0029] Some embodiments include an optimized coating for proper foam adhesion. Test coatings were ordered from Coatings2Go.com and tested on multiple samples. Without any etching process, a silicone coating, a hydrophilic coating (e.g., siloxane), and a hydrophobic polytetrafluoroethylene (PTFE) coating were applied onto the 0.0035" diameter nitinol wire using a KimWipe. This process was repeated for acid etched wires, which were etched using a mixture of 1:3 ratio of 6M nitric acid and 6M hydrochloric acid. Lastly, a thermoset neat polymer with the same bulk chemical composition as the foam was used as a wire coating. Bare, untreated, and acid etched wires were used as controls for foam adhesion.

[0030] Neat coatings were created inside a humidity-controlled chamber in order to reduce isocyanate-reactions with ambient moisture. The four monomers were mixed

in stoichiometric amounts with varying IPDI content congruent to the foam to be created. The monomer target values for all compositions can be seen in Table 3 below.

Table 3: Stoichiometric amounts of monomers for varying IPDI Content in Neat Polymers

0% IPDI	Sample ID	130625AMNCMB01	
Components	Target for 1 foam (g)	Scale	Total target value (g)
HPED	9.191	0.25	2.298
TEA	3.080	0.25	0.770
TMHDI	19.729	0.25	4.932
IPDI	0.000	0.25	0.000

5% IPDI	Sample ID	130625AMNCMB01	
Components	Target for 1 foam (g)	Scale	Total target value (g)
HPED	9.175	0.25	2.294
TEA	3.074	0.25	0.769
TMHDI	18.710	0.25	4.677
IPDI	1.041	0.25	0.260

10% IPDI	Sample ID	130625AMNCMB01	
Components	Target for 1 foam (g)	Scale	Total target value (g)
HPED	9.159	0.25	2.290
TEA	3.069	0.25	0.767
TMHDI	17.694	0.25	4.424
IPDI	2.078	0.25	0.520

15% IPDI:	Sample ID	130625AMNCMB01	
Components	Target for 1 foam (g)	Scale	Total target value (g)
HPED	9.143	0.25	2.286
TEA	3.063	0.25	0.766
TMHDI	16.682	0.25	4.170
IPDI	3.112	0.25	0.778

20% IPDI:	Sample ID	130625AMNCMB01	
Components	Target for 1 foam (g)	Scale	Total target value (g)
HPED	9.127	0.25	2.282
TEA	3.058	0.25	0.765
TMHDI	15.673	0.25	3.918
IPDI	4.142	0.25	1.036

[0031] Once the chemicals were massed and measured, the polymer solution was then mixed for one minute, checked for homogeneity, and mixed again for one minute. The wires to be coated were strung into a rectangular frame structure, and the polymer coatings were wiped on by hand using a Kim-Wipe.

#### WIRE ORIENTATION

[0032] To attain ideal foaming, Applicant determined the wire orientation also required optimization in some embodiments. Wires were strung in one bucket transversely to the foaming direction, while the second bucket had wires axial to the foam. This tested the interactions of the foam and wires to determine the least disruptive orientation to foaming. Once determined, an attachment to the foaming vessel was created in order to optimize foam/wire adherence that maintained the stability of the wire.

#### MICROSCOPE IMAGING

[0033] Microscope imaging was used to determine the homogeneity of the foam surrounding the wire and quantitative data such as density and pore size. Dry volume expansion was then used to further quantify the homogeneity of the foams. After foam synthesis, the wires were cut out of the foam using a Proxxon Thermocut Hotwire Cutter to remove extraneous foam. The sample was then imaged using a Leica long-range microscope with Jenoptic camera system and the foam morphology surrounding the wire was used to determine the useful length, with a goal of 12 cm. 12 cm was a parameter derived from the length of the final geometry of the device, a coil of nitinol wire in the aneurysm.

#### DENSITY CALCULATIONS

[0034] The ratio of the foam density surrounding the wire to the density of the bulk foam was measured to determine foam homogeneity in the vicinity of the wire. Bulk foam density was calculated by cutting three small squares from the center of the foam, far away from the wires to ensure minimal interference with the general structure and pores of the samples. The length, width, and height of each of the three squares were measured three times and the averages of each dimension were

taken. These averages were used to calculate the volume of the foam sample. The sample was then massed and the density was found using Equation 1.

$$\rho = m/V \quad \text{Equation (1)}$$

[0035] Where  $\rho$  is the density,  $m$  is the mass, and  $V$  is the volume. This process was then repeated for samples taken within 2 mm of the wire for three wires in the foam. The process used to measure the cube volume described above was repeated and each cube was massed. The calculated density was then averaged and then the bulk foam density was divided by the average wire-foam density, resulting in the ratio of the two foam types, as described by Equation 2.

$$\text{Density Ratio} = \rho_{Bulk} / \rho_{Wire} \quad \text{Equation (2)}$$

[0036] Where *Density Ratio* is the reported value,  $\rho_{Bulk}$  is the bulk foam density (where bulk foam refers to the foam after initial foaming and separate and apart from obstacles such as wires or other substrates, such as Figure 2), and  $\rho_{Wire}$  is the wire foam density.

#### PORE SIZE CALCULATIONS

[0037] Cell images were taken using a Leica long-range microscope with Jenoptic camera system from a sample of foam approximately one cell thick cut transversely to the axis of foaming. The diameter of a specific cell was measured from the collected images, and the diameter of 15 cells was used for the analysis. For non-circular cells, diameters were taken at the longest point between two walls. Calculations assume perfectly circular cell sizes. Pore sizes were calculated using Equation 3:

$$\text{Pore Size} = \frac{\pi}{4} d^2 \quad \text{Equation (3)}$$

Where  $d$  is the measured diameter of the cell.

#### VOLUME RECOVERY IN AIR

[0038] Cylindrical samples with a diameter of 6mm and a height of 4 cm were cut from the bulk foam, making sure that the wire was embedded along the axis of the

sample. The foams were radially crimped using an ST150-42 Machine Solutions stent crimper to the smallest possible diameter. The secondary morphology was set by maintaining the samples at 100°C while in the primary shape for 20 minutes, then crimping to the smallest possible diameter and subsequently cooling the samples back to room temperature. An image was taken using the Leica long-range microscope with Jenoptic camera system and analyzed for the diameter using Image J software. The samples were actuated at 100°C for 20 minutes in an oven, and allowed to cool to room temperature. Once cooled, the samples were again imaged and analyzed for the final diameter. The volume recovery was calculated using Equation 4.

$$V_E = (D_r/D_c)^2 \quad \text{Equation (4)}$$

Where  $V_E$  is the volume expansion,  $D_r$  is the recovered diameter, and  $D_c$  is the crimped diameter.

This was repeated for 5 samples.

## SCANNING ELECTRON MICROSCOPE IMAGES

[0039] Scanning Electron Microscope (SEM) images were taken of two samples of wire. The first sample was the uncoated, 0.0035" nitinol backbone wire. The second sample was the same nitinol wire with the addition of a neat polymer coating. The resulting images were analyzed with ImageJ Software to determine the overall change in diameter caused by the addition of the neat polymer coating.

## RESULTS

### FOAM SYNTHESIS

[0040] Foam Synthesis was completed on a range of chemical compositions (See Table 1). It was noted that as IPDI content increased (15% IPDI and 20% IPDI), the successful length of foam along the wire was lower when holding all other variables constant. The viscosity of the prepolymer increased with increasing IPDI content and resulted in a more viscous reaction mixture than lower IPDI concentrations as the foam progressed up the wire. To account for the increase in viscosity, the ratio

of OH/NCO prepolymer was decreased from 0.35 to 0.32. This change produced for a longer successful foam length along the wire as determined by qualitative imaging.

[0041] It was also discovered that the lower IPDI content foams (0% and 5% IPDI) should have a shorter mixing time than higher IPDI content foams after the addition of the Enovate. The lower IPDI content lends to a lower viscosity prepolymer compared to higher IPDI concentrations. The less viscose foam mixture required less mixing to achieve homogeneity, and rapidly expanded in size. The original 10 second mixing time was reduced to 5 seconds, and this change resulted in greater bulk homogeneity in foam composition, as shown in Figure 1.

[0042] Two different methods of securing wires were tested, shown in Figure 2, in order to determine the best method to attach wires to a foaming vessel. Applicant determined that the top of the foaming vessel needed to secure the coated wires, eliminating as much lateral motion as possible and to allow gaseous escape during the foaming reaction. Attaching wires to the bucket handle allowed for forward/backward movement from the handle, and was not adequate for our purposes. The concept was tedious and wasteful because it required taping spatulas to the top of the foaming bucket. From the given constraints, and the desire to be able to foam over many wires at once, the final structure, as seen in Figure 2, was created with the goal of securing the coated wires to the foaming vessel.

[0043] 1mm diameter holes were cut in the bottom of the 2L bucket, 1.5" from the center (halfway between center and outer diameter) at 90° angles from one another. This allowed four wires to be fed through the bottom of the bucket and taped down using Kapton heat-resistant tape. The wires were then pulled taught through the bucket and attached to the steel bars, seen in Figure 2, using the Kapton tape.

#### COATING OPTIMIZATION AND ADHESION

[0044] Multiple coatings were tested in order to optimize foam-wire adhesion, including uncoated (as a control), PTFE, Hydrophillic, silicone, and neat SMP. When uncoated wires were placed in the foaming vessel and foam was synthesized directly onto them, large caverns in the foams surrounding the wires were created. This was due to contrasting chemical compositions of the nitinol wire and PU foams. The

silicone, PTFE, and hydrophilic coatings were used to coat the 0.0035" wire following the directions provided by the company. The homogeneity of the foam is severely compromised in the immediate vicinity of the wire for silicone, PTFE, and hydrophilic coatings. For example, density near the wire is far greater than bulk foam density. From this, Applicant determined in some embodiments the coating of the wire should have an almost identical chemical composition as the foam. Therefore thermoset neat polymer was used as a coating.

[0045] An SEM image of the neat polymer coated wire can be seen in Figure 3. The polymer formed an uneven distribution of the polymer along the wire. Applicant determined that uneven coatings actually increased homogenous foam adhesion due to the expansion of the foam as it progressed up the wire.

[0046] Additionally, the uneven adhesion caused a varying change in diameter of the wire from the neat polymer. The original diameter of the wire was 0.0035" whereas the highest recorded diameter after two neat polymer coatings was 0.0069". This was almost double the diameter of the wire and could cause significant issues when crimping the foam around the wire because the crimped device diameter should ideally be under 0.016' to accommodate the dimensions of human microcatheters.

#### WIRE ORIENTATION

[0047] At least 4 attempts at wire orientation inside the buckets were foamed and it was determined by inspection that the transverse wires disrupted the foaming process significantly more than the vertical wires. There was a higher probability that the foam would split immediately above transverse placed wires, resulting in poor foam coverage above the wire, as shown in Figure 4. Figure 4 is a perspective taken just above the foam showing a cavity in the "wake" or side of substrate opposite the side that was initially contacted by foam. However, there was no such void when the substrate was oriented vertically with the substrate directly contacting the foam on all sides of the substrate (e.g., Figure 2 set up). In other words, the vertically oriented wire had adequate adhesion and little foam disruption compared to the previous coating used. Figure 5 shows the overall FOW structure. The

specimen seen in Figure 5 was cut to a diameter of 2.5 mm from the bulk foam using an Excimer Laser.

## DENSITY CALCULATIONS

[0048] The results of the density calculations are summarized in Table 4 below.

Table 4: Results of foam density calculations

Composition	Bulk Density (g/cm <sup>3</sup> )	FOW Density (g/cm <sup>3</sup> )	Ratio
0% IPDI	0.0107	0.0113	1.0622
5% IPDI	0.0108	0.0109	1.0141
10% IPDI	0.0127	0.0111	1.0207
15% IPDI	0.0120	0.0111	0.9662
20% IPDI	0.0092	0.0093	1.0075

[0049] The bulk density was the foam density calculated near the center of the foam, where no wires are present. The FOW density was calculated using foam taken within 2 mm of the wire. The ratio calculated is the FOW density to the bulk density. The majority (4 out of 5) of the density ratios were greater than one, implying that the foam surrounding the wires was slightly denser than the density of standard foams of the same composition. This was to be expected since the wire causes an imperfection in the foaming area, causing the foam to congregate more around the wire. Maintaining low density in the vicinity of the wire was of paramount importance because it relates to smaller device diameters when crimping. Since the ratio of densities was only slightly larger than one, this parameter was met.

## Pore Size Calculations

[0050] The data acquired from the pore size experiments are summarized in Figure 6. The pore size data indicates that there were slightly smaller pores in the FOW samples for the majority of compositions, with the largest difference being the 15% IPDI composition. However, Applicant noted that this composition had irregular data among all the experiments performed. Overall, as expected, pore size decreased slightly in the vicinity of the wire. However since this decrease was minimal, it shouldn't have a significant impact on the device as a whole.

## VOLUME RECOVERY IN AIR

[0051] The results of the volume recovery experiments in air are summarized in Figure 7. The data suggested large variability with possibly of decreasing volume recovery with increasing IPDI content. The data was skewed possibly due to the variability of foam quality around wire, and the position of the wire relative to the center of the foam. When cutting the foam with the biopsy punch, it was difficult to place the wire directly in the middle of the cylinder. The data would have been more accurate if all the samples were cut using an Excimer laser, ensuring the nitinol backbone wire was centered within the foam cylinder. The dry volume expansion data also suggested that adhered foam-wire samples may have better volume expansion than their bulk foam counter-part samples, which is excellent for use of this technology in the overall device. It is also important to note that dry volume expansion behavior was not entirely representative of the clinical uses of this material. Future expansion tests will be conducted in a heated water bath to better represent the aqueous vascular environment.

[0052] The following examples pertain to further embodiments.

[0053] Example 1 includes a system comprising: a substrate of a medical device; an un-foamed polyurethane coating directly contacting the substrate and fixedly attached to the substrate; a thermoset polyurethane shape memory polymer (SMP) foam, having first and second states, which directly contacts the polyurethane coating and fixedly attaches to the polyurethane coating; wherein the polyurethane coating fixedly attaches the SMP foam to the substrate.

[0054] While polyurethane examples (for the coating and the foam) are mentioned herein, other types of foams and/or coatings may be used including, for example and without limitation, polycarbonate, polyamide, polyurea, polyester, polycaprolactone, polyolefin, or combinations thereof.

[0055] The substrate may include a polymer suture, a metal backbone coil or wire (platinum, nitinol, nichrome), the exterior ring of a prosthetic heart valve, the exterior surface of a stent graft, a face of a water or air filter, a surface of a safety helmet,

and the like. In other words, examples are not limited to medical devices, much less embolic foams for aneurysms.

[0056] By fixedly attaching the foam to the substrate, the coating alleviates many problems Applicant identified (e.g., a foam that slides along a backbone can be difficult to place in an aneurysm or atrial appendage or anatomical void and the like). For example, a polymer coating having the same or similar chemistry as the foam helps prevent the rising foam from contacting a foreign object (e.g., an uncoated nitinol wire) and bunching or coalescing around the object (e.g., where cells in foam collapse at the wire interface forming a dense region of foam (as compared to the bulk foam) that may lose flexibility and become unsuitable for certain applications like manipulation through tortuous vasculature of a patient).

[0057] Example 2 includes the system of claim 1, wherein the polyurethane coating is a thermoplastic.

[0058] Example 3 includes the system of claim 1, wherein the polyurethane coating is a cured thermoset.

[0059] Thus, embodiments allow, for example, thermoplastic or thermoset coatings.

[0060] Example 4 includes the system of claim 1, wherein the polyurethane coating has a chemical composition equal to a chemical composition of the SMP foam.

[0061] Example 5 includes the system of claim 1, wherein the polyurethane coating and the SMP foam both include N,N,N',N'-tetrakis (2-hydroxypropyl) ethylenediamine (HPED), triethanolamine (TEA), and hexamethylene diisocyanate (HDI) with TEA contributing a higher molar ratio of hydroxyl groups than HPED.

[0062] For example, the coating may include a neat polymer that is un-foamed yet still have the same chemical structure or constituents as the foamed polymer.

[0063] Example 6 includes the system of claim 1, wherein the polyurethane coating and the SMP foam both include N,N,N',N'-tetrakis (2-hydroxypropyl) ethylenediamine (HPED), triethanolamine (TEA), and trimethylhexamethylenediamine (TMHDI) with HPED contributing a higher molar ratio of hydroxyl groups than TEA.

[0064] Example 7 includes the system of claim 1, wherein the polyurethane coating and the SMP foam both include N,N,N',N'-tetrakis (2-hydroxypropyl) ethylenediamine (HPED), Glycerol, pentanediol, and hexamethylene diisocyanate (HDI).

[0065] Example 8 includes the system of claim 1, wherein the polyurethane coating is more hydrophobic than siloxane and less hydrophobic than polytetrafluoroethylene (PTFE).

[0066] Applicant has determined such a level of hydrophobicity helps the rising foam adequately couple to the coating without forming a dense foam area at the substrate/foam interface.

[0067] Example 9 includes the system of claim 1, the SMP foam includes cells and a majority of the cells include unequal major and minor axes and the major axes generally align with each other.

[0068] For example, in an embodiment the cells are elliptical with a major axis generally parallel (+/- 15 degrees) to the long axis of the substrate, such as the long axis of the vertical substrate of the wires of Figure 2. This would not be true for the instance where the wire is orthogonal to the foam path, such as Figure 4. In an embodiment, the elliptical cells are within 2 mm of the substrate.

[0069] Example 10 includes the system of claim 9, wherein substrate includes a major axis unequal to a minor axis and the major axis of the substrate generally aligns with the major axes of the majority of the cells.

[0070] Example 11 includes the system of claim 1, the SMP foam includes cells and a majority of the cells are isotropic in shape.

[0071] In other embodiments, the cells are not as described in Example 9. In some embodiments the cells are elliptical within 2 mm of the substrate but isotropic more than 2 mm from the substrate. 2 mm is a demarcation used for explanation and can be 1, 3, 4, 5, 6 mm or more in other embodiments.

[0072] Example 12 includes the system of claim 1, wherein the substrate is included in a member selected from the group comprising: embolic coil, stent, stent graft, polymer suture, wire, heart valve, and tissue scaffold.

[0073] Example 13 includes the system of claim 1, wherein: the polyurethane coating directly contacts the SMP foam along a first maximum length; and the polyurethane coating directly contacts the substrate along a second maximum length that is at least as long as the first maximum length.

[0074] For example, in Figure 2 the foam would envelope and adhere along the entire length of the wire substrate. Due to masking, the polymer coating may cover the majority of the wire but other portions of the coating may be masked such that they do not adhere to foam. Such masking may allow for placing foam at intervals on the substrate and the like. Regardless, in such a case the foam need not contact every portion of the coating but the coating may still ensure the foam is secured along the entire length of the foam. For example, for an embolic foam a backbone may have enough polymer coating such that the entire length of the embolic foam fixedly adheres to the backbone, thereby facilitating foam placement. Embodiments may include situations where the foam is longer than the polymer coating or vice versa. Portions of the substrate may be masked and connect to neither polymer coating nor foam.

[0075] Example 14 includes the system of claim 13, wherein the first maximum length is at least 2 cm and the substrate includes a wire that is less than .0020" in diameter.

[0076] In other embodiments the length is 4, 6, 8, 10, 12, 14, 16, 18, 20 cm or more and/or .001", .003", 0.004" .005", .006" diameter or more.

[0077] Example 15 includes the system of claim 1, wherein: the SMP foam has inner and outer portions; the inner portion is no more than 2 mm thick, measured orthogonal to the substrate, and directly between the outer portion and the polyurethane coating; the inner portion has a first foam density and the outer half portion has a second foam density; a ratio of the first foam density to the second foam density is less than 1.1.

[0078] Other embodiments include a ratio of .8, .9, 1.2, 1.4, 1.6 and the like. For example, the center of a column or portion of foam may have a density immediately adjacent the substrate that is similar to the density further removed from the substrate.

[0079] Example 16 includes the system of claim 1, wherein: the SMP foam has inner and outer portions; the inner portion is no more than 2 mm thick, measured orthogonal to the substrate, and directly between the outer portion and the polyurethane coating; the inner portion has a first foam density and the outer portion has a second foam density and both of the first and second foam densities are less than 0.06 g/cm<sup>3</sup>.

[0080] Other embodiments include a density less than .04, .08, .1, and .5 g/cm<sup>3</sup>.

[0081] Example 17 includes the system of claim 1, wherein: the SMP foam has inner and outer portions; the inner portion is no more than 2 mm thick, measured orthogonal to the substrate, and directly between the outer portion and the polyurethane coating; the inner portion has a first average cell diameter and the outer half portion has a second average cell diameter that is greater than the first average cell diameter.

[0082] Example 18 includes a method comprising: coating a substrate of a medical device with an un-foamed polyurethane coating that directly contacts the substrate and fixedly attaches to the substrate; forming a mixture by combining N,N,N',N'-tetrakis (2-hydroxypropyl) ethylenediamine (HPED), and at least two members selected from the group consisting of: triethanolamine (TEA), and hexamethylene diisocyanate (HDI), trimethylhexamethylenediamine (TMHDI), Glycerol, and pentanediol; locating the polyurethane coated substrate adjacent the mixture; foaming the mixture into a monolithic foam that expands towards the polyurethane coated substrate, directly contacts the polyurethane coated substrate, substantially surrounds the polyurethane coated substrate, and fixedly attaches to the polyurethane coated substrate; separating the monolithic foam into a first portion of the foam contacting the polyurethane coated substrate and a remaining bulk portion

of the foam; and forming the first portion of the foam into a primary state and a programmed state to form a shape memory polymer (SMP) foam.

[0083] Figure 8 includes method 800. Block 801 includes coating a substrate of a medical device with an un-foamed polyurethane coating that directly contacts the substrate and fixedly attaches to the substrate. Block 802 includes forming a mixture by combining N,N,N',N'-tetrakis (2-hydroxypropyl) ethylenediamine (HPED), and at least two members selected from the group consisting of: triethanolamine (TEA), and hexamethylene diisocyanate (HDI), trimethylhexamethylenediamine (TMHDI), Glycerol, and pentanediol. Block 803 includes locating the polyurethane coated substrate adjacent the mixture. For example, see Figure 2 where the mixture would be in the bottom of the bucket and would move upwards along the substrate. Block 804 includes foaming the mixture into a monolithic foam that expands towards the polyurethane coated substrate, directly contacts the polyurethane coated substrate, substantially surrounds the polyurethane coated substrate, and fixedly attaches to the polyurethane coated substrate. Block 805 includes separating the monolithic foam into a first portion of the foam contacting the polyurethane coated substrate and a remaining bulk portion of the foam. Block 806 includes forming the first portion of the foam into a primary state and a programmed state to form a shape memory polymer (SMP) foam.

[0084] Example 19 includes the method of claim 18, wherein foaming the mixture into a monolithic foam comprises expanding the monolithic foam to first contact a proximal portion of the polyurethane coated substrate, then a middle portion of the polyurethane coated substrate, and then a distal portion of the polyurethane coated substrate.

[0085] Example 20 includes the method of claim 19, wherein foaming the mixture into a monolithic foam comprises simultaneously surrounding the monolithic foam in a plane orthogonal to a long axis of the polyurethane coated substrate.

[0086] Example 21 includes the method of claim 20, wherein the un-foamed polyurethane coating comprises HPED, and at least two members selected from the group consisting of: TEA, HDI, TMHDI, Glycerol, and pentanediol.

[0087] Example 1a includes a porous coating for a wire or coil, wherein the porous coating comprises a porous, shape memory material, wherein the porous coating is adapted to volumetrically expand as to fill gaps between the device body and the wall of the adjacent tissue and to create volumetric filling between the medical device and the adjacent tissue.

[0088] Example 2a includes a porous coating according to example 1 wherein the porous coating is formed by coating the wire with a shape memory material and then heating on to the wire.

[0089] Example 3a includes a medical device comprising a body and a porous coating secured to a surface of the body, wherein the porous coating comprises a porous, shape memory material.

[0090] Example 4a includes the medical device according to example 1 wherein the volumetric expansion is caused by the shape memory effect of the shape memory material.

[0091] Example 5a includes the medical device according to example 1 wherein the volumetric filling by the porous coating causes an interface with the adjacent tissue and promotes cellular ingrowth.

[0092] Example 6a includes a method for providing therapy to a patient, the method comprising: providing a medical device comprising a body and a porous coating secured to a surface of the body, wherein the porous coating comprises a porous, shape memory material; inserting the medical device into the patient so that the porous coating expands against the adjacent tissue so as to fill gaps between the device body and the wall of the adjacent tissue and to promote tissue integration between the medical device and the wall of the blood vessel.

[0093] Example 7a includes the method according to example 6 wherein the volumetric expansion is created by the shape memory effect of the shape memory material.

[0094] Example 8a includes the method according to example 6 wherein the volumetric expansion of the porous coating causes embolization of an aneurysm.

[0095] Example 9a includes a method of applying a shape memory polymer to a nitinol wire using a direct foaming process.

[0096] Example 10a includes the method of example 9, wherein the nitinol wire is coated with a coating application to the application of the shape memory polymer foam.

[0097] Example 11a includes the method of example 10, wherein the coating is selected from, but not limited to, the group consisting of a silicone coating, a hydrophilic coating, a polytetrafluoroethylene (PTFE) coating, a polyurethane, and a thermoset neat polymer with the same bulk chemical composition as the foam.

[0098] Example 12a includes the method of example 11, wherein the wire is acid-etched prior to coating.

[0099] Example 13a includes the method of example 11, wherein the wire is unetched prior to coating.

[0100] Example 14a includes the method of example 11 wherein, the wire is subjected to direct foaming by exposing the wire to a reaction mixture comprising at least one shape memory foam polymer and at least one foaming agent.

[0101] The foregoing description of the embodiments has been presented for the purposes of illustration and description. It is not intended to be exhaustive or to limit the embodiments to the precise forms disclosed. This description and the claims following include terms, such as left, right, top, bottom, over, under, upper, lower, first, second, etc. that are used for descriptive purposes only and are not to be construed as limiting. The embodiments of a device or article described herein can be manufactured, used, or shipped in a number of positions and orientations. Persons skilled in the relevant art can appreciate that many modifications and variations are possible in light of the above teaching. Persons skilled in the art will recognize various equivalent combinations and substitutions for various components shown in the Figures. It is therefore intended that the scope of the embodiments be limited not by this detailed description, but rather by the claims appended hereto.

What is claimed is:

- 1 1. A system comprising:  
2 a substrate of a medical device;  
3 an un-foamed polyurethane coating directly contacting the substrate and  
4 fixedly attached to the substrate;  
5 a thermoset polyurethane shape memory polymer (SMP) foam, having first  
6 and second states, which directly contacts the polyurethane coating and fixedly  
7 attaches to the polyurethane coating;  
8 wherein the polyurethane coating fixedly attaches the SMP foam to the  
9 substrate.
  
- 1 2. The system of claim 1, wherein the polyurethane coating is a thermoplastic.  
1
  
- 1 3. The system of claim 1, wherein the polyurethane coating is a cured  
2 thermoset.
  
- 1 4. The system of claim 1, wherein the polyurethane coating has a chemical  
2 composition equal to a chemical composition of the SMP foam.
  
- 1 5. The system of claim 1, wherein the polyurethane coating and the SMP foam  
2 both include N,N,N',N'-tetrakis (2-hydroxypropyl) ethylenediamine (HPED),  
3 triethanolamine (TEA), and hexamethylene diisocyanate (HDI) with TEA contributing  
4 a higher molar ratio of hydroxyl groups than HPED.
  
- 1 6. The system of claim 1, wherein the polyurethane coating and the SMP foam  
2 both include N,N,N',N'-tetrakis (2-hydroxypropyl) ethylenediamine (HPED),  
3 triethanolamine (TEA), and trimethylhexamethylenediamine (TMHDI) with HPED  
4 contributing a higher molar ratio of hydroxyl groups than TEA.

- 1 7. The system of claim 1, wherein the polyurethane coating and the SMP foam  
2 both include N,N,N',N'-tetrakis (2-hydroxypropyl) ethylenediamine (HPED), Glycerol,  
3 pentanediol, and hexamethylene diisocyanate (HDI).
- 1 8. The system of claim 1, wherein the polyurethane coating is more hydrophobic  
2 than siloxane and less hydrophobic than polytetrafluoroethylene (PTFE).
- 1 9. The system of claim 1, the SMP foam includes cells and a majority of the cells  
2 include unequal major and minor axes and the major axes generally align with each  
3 other.
- 1 10. The system of claim 9, wherein substrate includes a major axis unequal to a  
2 minor axis and the major axis of the substrate generally aligns with the major axes of  
3 the majority of the cells.
- 1 11. The system of claim 1, the SMP foam includes cells and a majority of the cells  
2 are isotropic in shape.
- 1 12. The system of claim 1, wherein the substrate is included in a member  
2 selected from the group comprising: embolic coil, stent, stent graft, polymer suture,  
3 wire, heart valve, and tissue scaffold.
- 1 13. The system of claim 1, wherein:  
2 the polyurethane coating directly contacts the SMP foam along a first  
3 maximum length; and  
4 the polyurethane coating directly contacts the substrate along a second  
5 maximum length that is at least as long as the first maximum length.
- 1 14. The system of claim 13, wherein the first maximum length is at least 2 cm and  
2 the substrate includes a wire that is less than .0020" in diameter.

1 15. The system of claim 1, wherein:  
2 the SMP foam has inner and outer portions;  
3 the inner portion is no more than 2 mm thick, measured orthogonal to the  
4 substrate, and directly between the outer portion and the polyurethane coating;  
5 the inner portion has a first foam density and the outer half portion has a  
6 second foam density;  
7 a ratio of the first foam density to the second foam density is less than 1.1.

1 16. The system of claim 1, wherein:  
2 the SMP foam has inner and outer portions;  
3 the inner portion is no more than 2 mm thick, measured orthogonal to the  
4 substrate, and directly between the outer portion and the polyurethane coating;  
5 the inner portion has a first foam density and the outer portion has a second  
6 foam density and both of the first and second foam densities are less than 0.06  
7 g/cm<sup>3</sup>.

1 17. The system of claim 1, wherein:  
2 the SMP foam has inner and outer portions;  
3 the inner portion is no more than 2 mm thick, measured orthogonal to the  
4 substrate, and directly between the outer portion and the polyurethane coating;  
5 the inner portion has a first average cell diameter and the outer half portion  
6 has a second average cell diameter that is greater than the first average cell  
7 diameter.

1 18. A method comprising:  
2 coating a substrate of a medical device with an un-foamed polyurethane  
3 coating that directly contacts the substrate and fixedly attaches to the substrate;  
4 forming a mixture by combining N,N,N',N'-tetrakis (2-hydroxypropyl)  
5 ethylenediamine (HPED), and at least two members selected from the group  
6 consisting of: triethanolamine (TEA), and hexamethylene diisocyanate (HDI),  
7 trimethylhexamethylenediamine (TMHDI), Glycerol, and pentanediol;  
8 locating the polyurethane coated substrate adjacent the mixture;

9 foaming the mixture into a monolithic foam that expands towards the  
10 polyurethane coated substrate, directly contacts the polyurethane coated substrate,  
11 substantially surrounds the polyurethane coated substrate, and fixedly attaches to  
12 the polyurethane coated substrate;

13 separating the monolithic foam into a first portion of the foam contacting the  
14 polyurethane coated substrate and a remaining bulk portion of the foam; and

15 forming the first portion of the foam into a primary state and a programmed  
16 state to form a shape memory polymer (SMP) foam.

1 19. The method of claim 18, wherein foaming the mixture into a monolithic foam  
2 comprises expanding the monolithic foam to first contact a proximal portion of the  
3 polyurethane coated substrate, then a middle portion of the polyurethane coated  
4 substrate, and then a distal portion of the polyurethane coated substrate.

1 20. The method of claim 19, wherein foaming the mixture into a monolithic foam  
2 comprises simultaneously surrounding the monolithic foam in a plane orthogonal to a  
3 long axis of the polyurethane coated substrate.

1 21. The method of claim 20, wherein the un-foamed polyurethane coating  
2 comprises HPED, and at least two members selected from the group consisting of:  
3 TEA, HDI, TMHDI, Glycerol, and pentanediol.



Figure 1

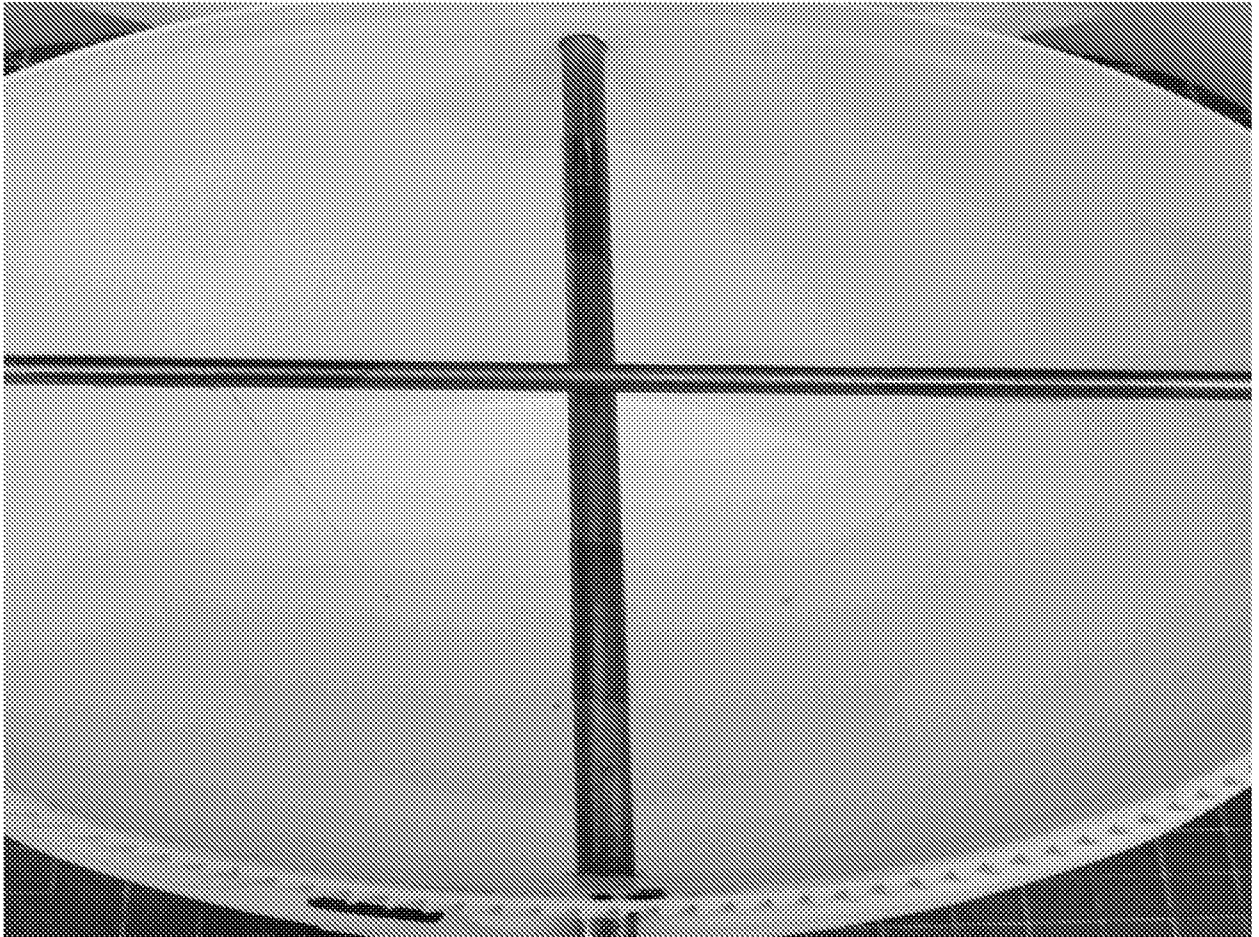


Figure 2

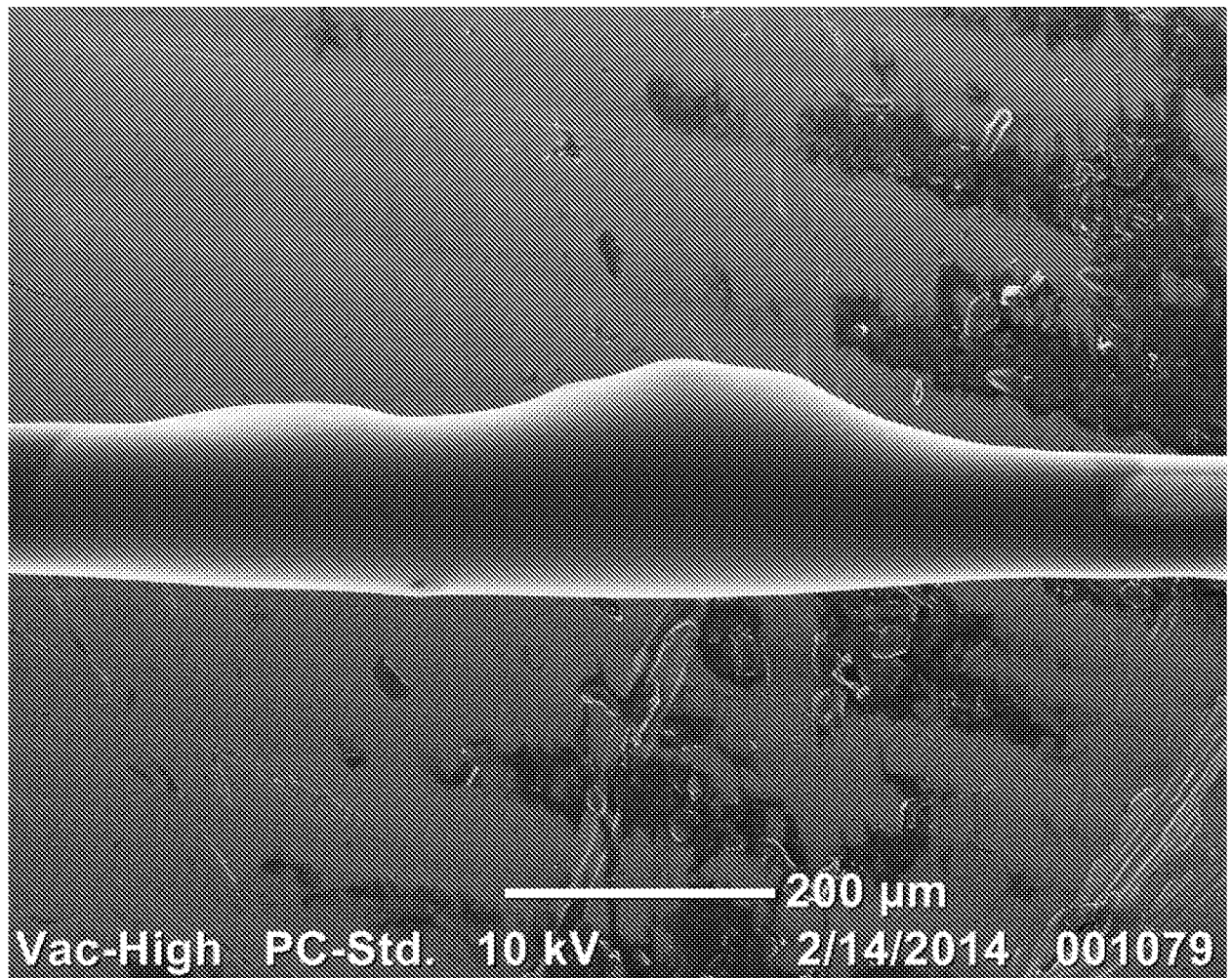


Figure 3

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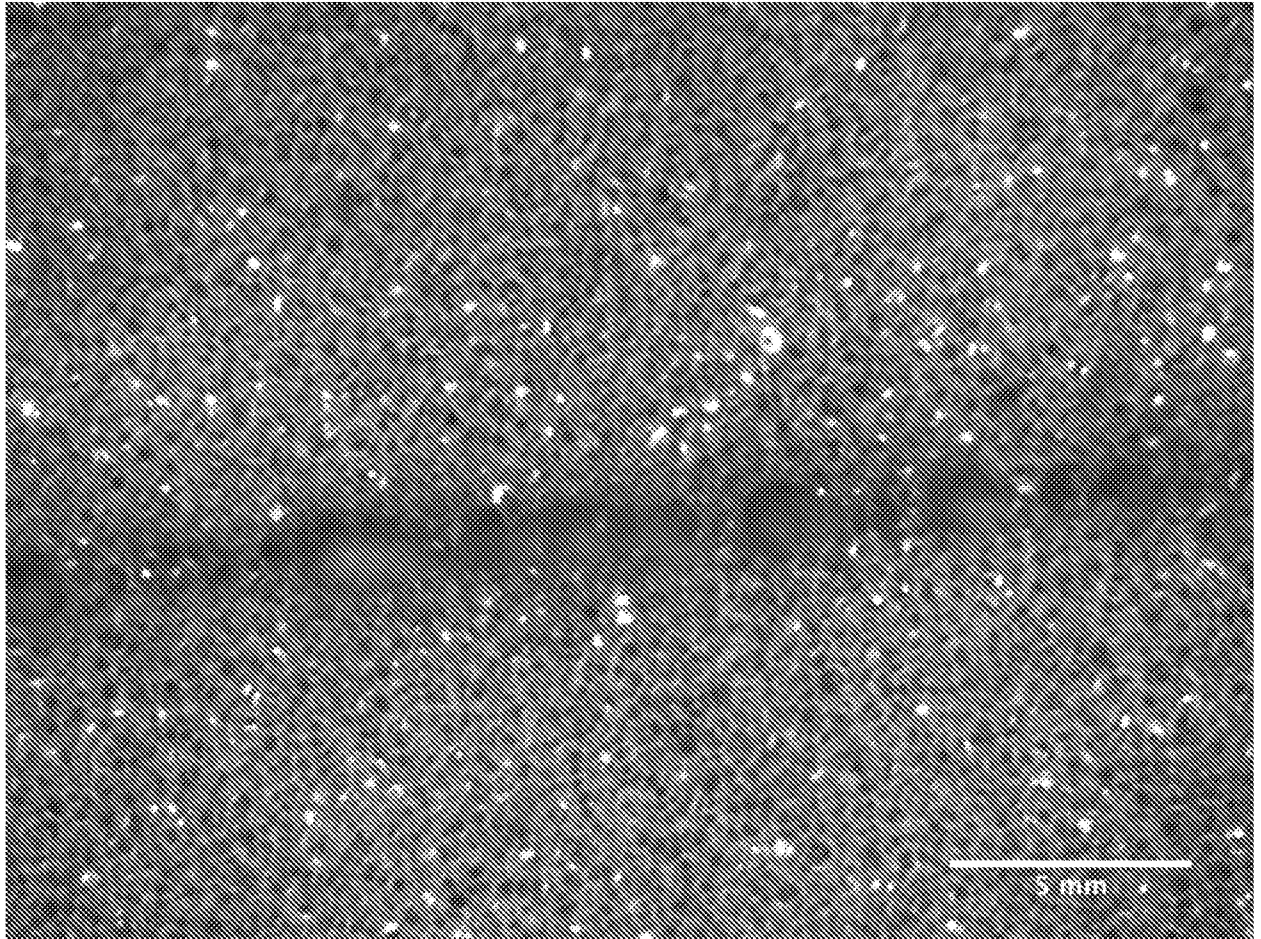


Figure 4

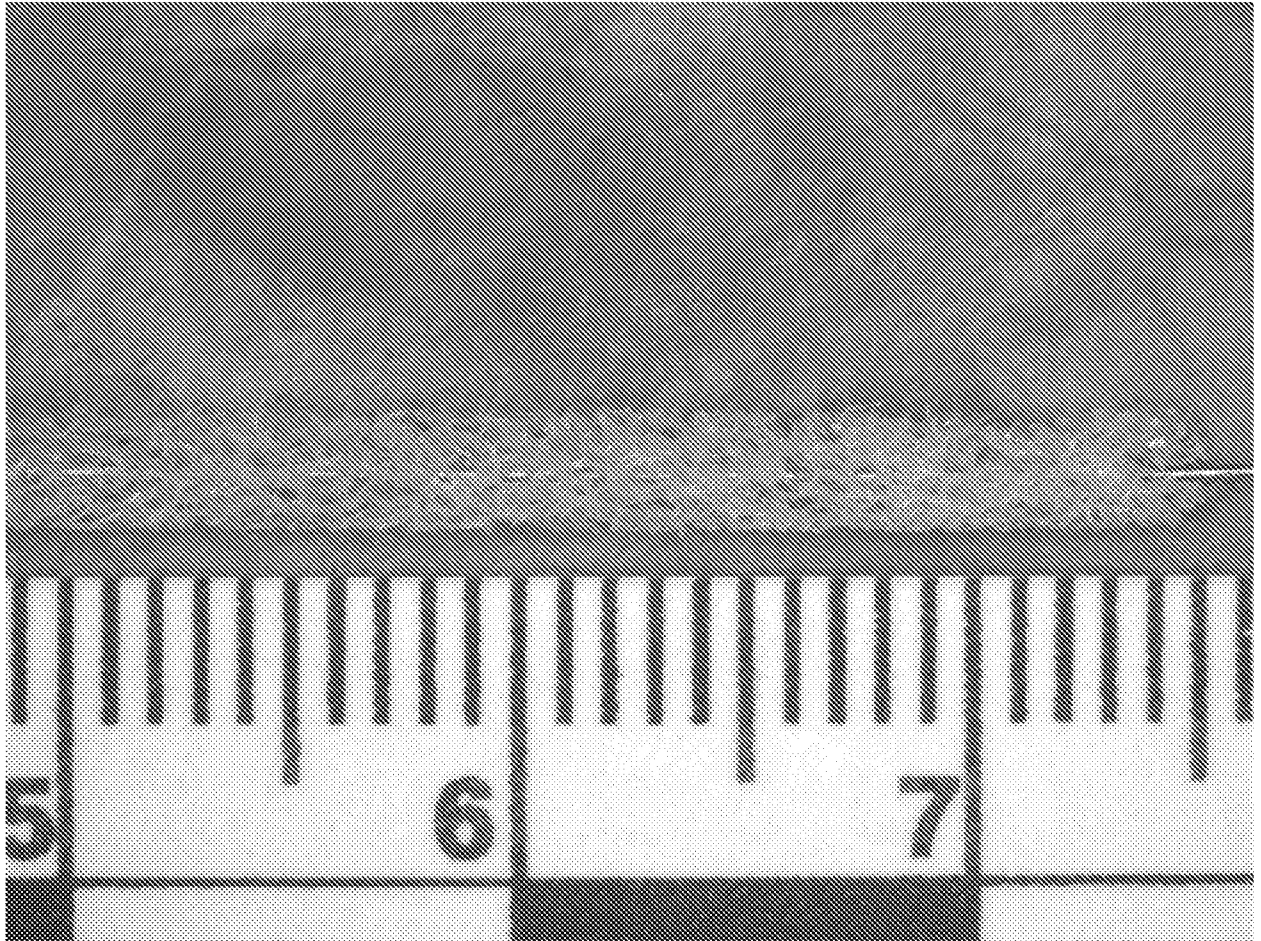


Figure 5

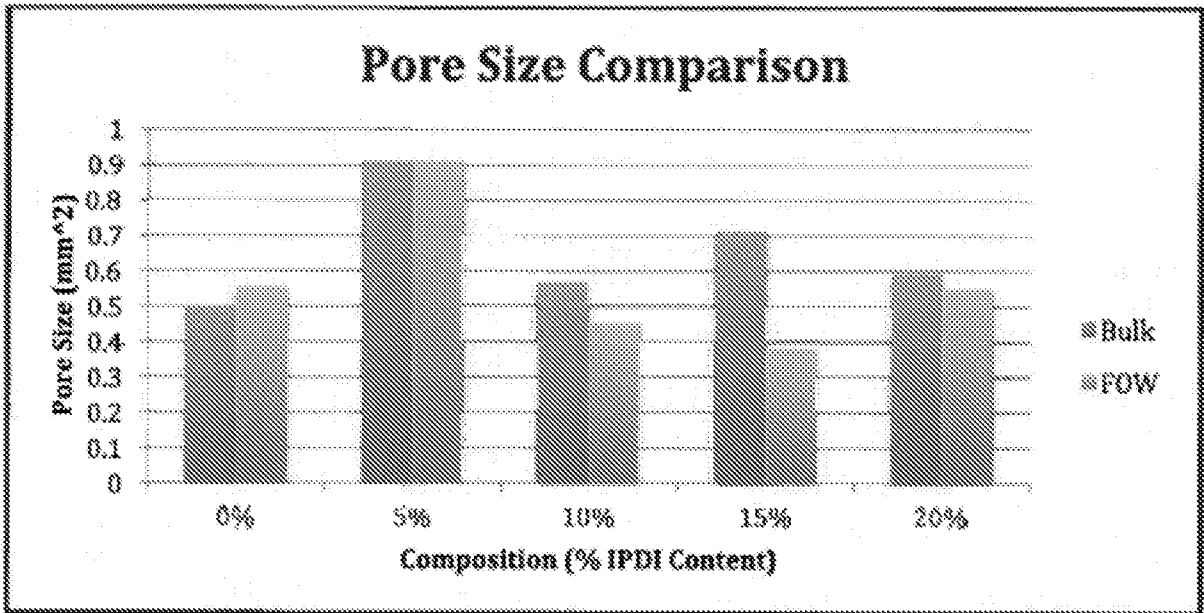


Figure 6

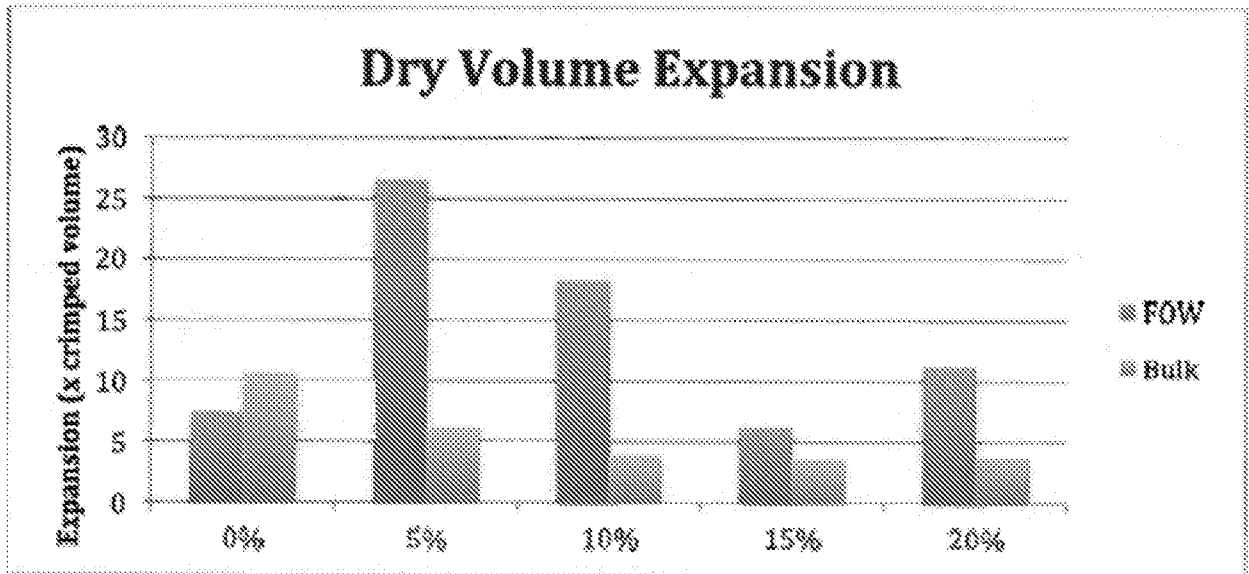


Figure 7

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800

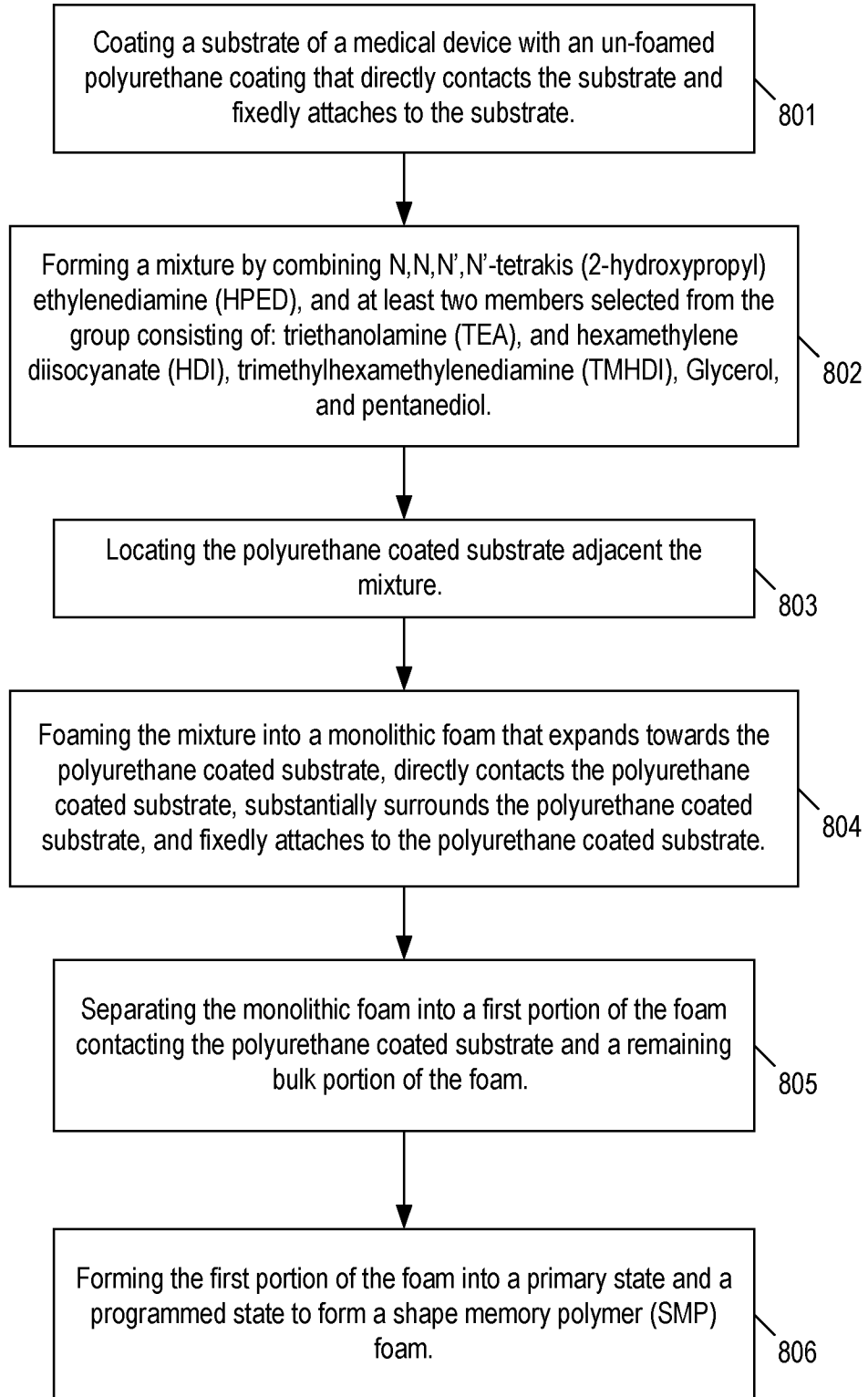


Figure 8

INTERNATIONAL SEARCH REPORT

International application No.  
PCT/US 16/43711

<p>A. CLASSIFICATION OF SUBJECT MATTER IPC(8) - A61L 33/00 (2016.01) CPC - A61L 31/10; A61L 31/16; A61L 27/34 UC = 427/2.1 and 427/407 According to International Patent Classification (IPC) or to both national classification and IPC</p>																	
<p>B. FIELDS SEARCHED</p> <p>Minimum documentation searched (classification system followed by classification symbols) IPC(8) - A61L 33/00 (2016.01) CPC - A61L 31/10; A61L 31/16; A61L 27/34 UC = 427/2.1 and 427/407</p> <p>Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched CPC - C08J9/32, B01J13/18, C08G2280/00, C09J175/04, C08G18/10, Y10T442/2738 and Y10T428/249953 (Search terms - See below)</p> <p>Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) PatBase and Google. Search terms: substrate, surface, coat, cover, finish, medical, device, polyurethane, urethane, unfoam, neat, foamable, thermoset, shape, memory, shape, memory, smp, attach, adhesive and bond.</p>																	
<p>C. DOCUMENTS CONSIDERED TO BE RELEVANT</p> <table border="1"> <thead> <tr> <th>Category*</th> <th>Citation of document, with indication, where appropriate, of the relevant passages</th> <th>Relevant to claim No.</th> </tr> </thead> <tbody> <tr> <td>X --- Y</td> <td>US 2013/0317541 et al. (Singhal et al.) 28 November 2013 (28.11.2013) Entire document, especially (para [0013]-[0019], [0034], [0042], [0046] [0054], [0055] [0059], [0060] and [0067]</td> <td>1, 3-6 and 8-17. ----- 2, 7 and 18-21</td> </tr> <tr> <td>Y</td> <td>Singhal et al. "Ultra Low Density and Highly Crosslinked Biocompatible Shape Memory Polyurethane Foams"; (Published March 4, 2012), Journal of Polymer Science Part B: Polymer Physics; 201,50, pp 724-737 url:&lt; http://onlinelibrary.wiley.com/doi/10.1002/polb.23056/full&gt; Entire document, especially pg 724, col 2 para 2.</td> <td>2</td> </tr> <tr> <td>Y</td> <td>US 9,018,273 B2 (Ito et al.) 28 April 2015 (28.04.2015) Entire document, especially col 2, ln 37-39 and col 5, ln 3-40</td> <td>7</td> </tr> <tr> <td>Y</td> <td>US 9,051,411 B2 (Wilson et al.) 09 June 2015 (09.06.2015) Entire document, especially Col 2, ln 21-23 and col 14, ln 58-63</td> <td>18-21</td> </tr> </tbody> </table>			Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.	X --- Y	US 2013/0317541 et al. (Singhal et al.) 28 November 2013 (28.11.2013) Entire document, especially (para [0013]-[0019], [0034], [0042], [0046] [0054], [0055] [0059], [0060] and [0067]	1, 3-6 and 8-17. ----- 2, 7 and 18-21	Y	Singhal et al. "Ultra Low Density and Highly Crosslinked Biocompatible Shape Memory Polyurethane Foams"; (Published March 4, 2012), Journal of Polymer Science Part B: Polymer Physics; 201,50, pp 724-737 url:< http://onlinelibrary.wiley.com/doi/10.1002/polb.23056/full> Entire document, especially pg 724, col 2 para 2.	2	Y	US 9,018,273 B2 (Ito et al.) 28 April 2015 (28.04.2015) Entire document, especially col 2, ln 37-39 and col 5, ln 3-40	7	Y	US 9,051,411 B2 (Wilson et al.) 09 June 2015 (09.06.2015) Entire document, especially Col 2, ln 21-23 and col 14, ln 58-63	18-21
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<p><input type="checkbox"/> Further documents are listed in the continuation of Box C. <input type="checkbox"/></p>																	
<p>* Special categories of cited documents:</p> <table border="0"> <tr> <td>"A" document defining the general state of the art which is not considered to be of particular relevance</td> <td>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</td> </tr> <tr> <td>"E" earlier application or patent but published on or after the international filing date</td> <td>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</td> </tr> <tr> <td>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</td> <td>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art</td> </tr> <tr> <td>"O" document referring to an oral disclosure, use, exhibition or other means</td> <td>"&amp;" document member of the same patent family</td> </tr> <tr> <td>"P" document published prior to the international filing date but later than the priority date claimed</td> <td></td> </tr> </table>			"A" document defining the general state of the art which is not considered to be of particular relevance	"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention	"E" earlier application or patent but published on or after the international filing date	"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone	"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art	"O" document referring to an oral disclosure, use, exhibition or other means	"&" document member of the same patent family	"P" document published prior to the international filing date but later than the priority date claimed						
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"O" document referring to an oral disclosure, use, exhibition or other means	"&" document member of the same patent family																
"P" document published prior to the international filing date but later than the priority date claimed																	
<p>Date of the actual completion of the international search 19 September 2016</p>		<p>Date of mailing of the international search report <b>13 OCT 2016</b></p>															
<p>Name and mailing address of the ISA/US Mail Stop PCT, Attn: ISA/US, Commissioner for Patents P.O. Box 1450, Alexandria, Virginia 22313-1450 Facsimile No. 571-273-8300</p>		<p>Authorized officer: Lee W. Young</p> <p>PCT Helpdesk: 571-272-4300 PCT OSP: 571-272-7774</p>															