

PATENT SPECIFICATION

(11) 1 567 341

1 567 341

- (21) Application No. 23695/77 (22) Filed 3 June 1977
- (31) Convention Application No. 693282
- (32) Filed 7 June 1976
- (31) Convention Application No. 785533
- (32) Filed 7 April 1977 in
- (33) United States of America (US)
- (44) Complete Specification published 14 May 1980
- (51) INT CL³ C07C 103/52
- (52) Index at acceptance
C3H 313 314 350 370 A3



(54) SOMATOSTATIN ANALOGS WITH DISSOCIATED BIOLOGICAL ACTIVITIES

(71) We, THE SALK INSTITUTE FOR BIOLOGICAL STUDIES, a Corporation organized and existing under the laws of the State of California, United States of America, of 10010 North Torrey Pines Road, La Jolla, State of California, United States of America, do hereby declare the invention, for which we pray that a patent may be granted to us, and the method by which it is to be performed to be particularly described in and by the following statement:—

The present invention relates generally to peptides having dissociated biological activity in respect to the inhibition of growth hormone, insulin and glucagon secretion. More particularly, the present invention is directed to peptides which are effective to selectively inhibit only the release of growth hormone by the pituitary gland or the release of glucagon or insulin by the pancreas.

A peptide having inhibitory effect on the secretion of growth hormone has been characterized and is described in United States Patent Specification No. 3,904,594. This peptide has been named "somatostatin". Somatostatin (also known as somatotropin release inhibiting factor) is the tetradecapeptide:



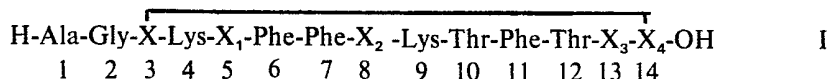
Somatostatin, the linear form of somatostatin (dihydrosomatostatin) and various acylated derivatives of somatostatin and dihydrosomatostatin are described in the aforementioned United States Patent Specification.

Somatostatin and many analogs of somatostatin exhibit activity in respect to the inhibition of growth hormone (GH) secretion from cultured, dispersed, rat anterior pituitary cells *in vitro* and inhibition of insulin and glucagon secretion *in vivo* in the rat. It has been considered highly desirable in the use of somatostatin to selectively inhibit only the secretion of GH, insulin or glucagon. Efforts have been made to develop analogs of somatostatin which possess dissociated biological activity and which inhibit only GH, insulin or glucagon secretion. Although there have been reports citing differences in the amounts of somatostatin required for inhibition of insulin compared to glucagon in the human and the perfused rat pancreas *in vitro*, somatostatin and some somatostatin analogs exhibit similar potencies on the inhibition of these two hormones. By "dissociated biological activity" as used herein is meant a pattern of activity with respect to inhibition of growth hormone, insulin and glucagon secretion which varies significantly from that of somatostatin. By way of example, comparing dosages which have generally the same effect with regard to inhibition of glucagon secretion, some somatostatin analogs have been found to exhibit a significantly greater or lesser effect with regard to inhibiting secretion of growth hormone and/or insulin.

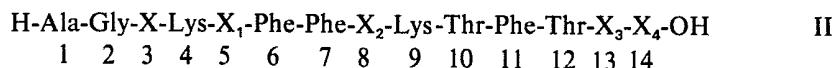
The present invention relates to the discovery that certain amino acids can be substituted for amino acid substituents in somatostatin and dihydrosomatostatin to provide peptides which possess dissociated biological activity in respect to the inhibition of GH, insulin or glucagon secretion. As a convenient shorthand form the novel peptides of the present invention are described in terms of the amino acid moiety which is substituted, the position of substitution and whether the substitution is made in somatostatin (SS) or dihydrosomatostatin (DHSS). The nomenclature used to describe the peptides of the present invention is in accordance with the conventional practice and in accordance with such practice, it

is the L form of the amino acid that is intended, unless otherwise expressly indicated.

The novel peptides of the invention are defined by the formulae herein below, where the individual amino acids of the peptide are numbered from left to right for ease of identification:



and



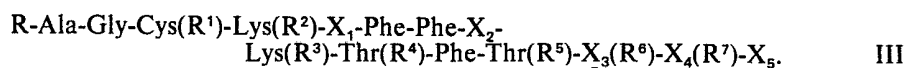
wherein X is Cys or D-Cys, X₁ is Asn or des-Asn (i.e. a direct bond), X₂ is Trp or D-Trp, X₃ is Ser or D-Ser and X₄ is Cys or D-Cys; with the proviso that whenever X₂ is D-Trp, X₃ is D-ser and/or X₄ is D-Cys, and that X₃ is D-Ser whenever X₁ is Asn and X₄ is Cys, and that X₄ is DCys whenever X₁ is Asn and X₃ is Ser. X can be either L-Cys or D-Cys without change in the potency or specificity of the peptides.

The novel peptides of the present invention having specific biological activity in respect to release of growth hormone, insulin and glucagon are:

[D-Ser¹³]-SS; [D-Ser¹³]-DHSS; [D-Trp⁸]-[D-Ser¹³]-SS; [D-Trp⁸]-[D-Ser¹³]-DHSS; [D-Cys¹⁴]-SS; [D-Cys¹⁴]-DHSS; [D-Trp⁸]-[D-Cys¹⁴]-SS and [D-Trp⁸]-[D-Cys¹⁴]-DHSS.

Pharmaceutically acceptable or veterinarily acceptable, acid addition salts of the peptides are also within the scope of the present invention. Such acid addition salts include but are not limited to hydrochlorides, hydrobromides, sulfates, phosphates, maleates, acetates, citrates, benzoates, succinates, malates, ascorbates and tartrates.

Also considered to be within the scope of the present invention are intermediates of the formula:



In the above formula III R is either hydrogen or an α -amino protecting group. The α -amino protecting groups contemplated by R are those known to be useful in the art in the step-wise synthesis of polypeptides. Among the classes of α -amino protecting groups covered by R are (1) acyl type protecting groups such as e.g. formyl, trifluoroacetyl, phthalyl, toluenesulfonyl, (tosyl), benzenesulfonyl, nitrophenylsulfonyl, tritylsulfonyl, o-nitrophenoxycarbonyl, chloroacetyl, acetyl and γ -chlorobutyryl; (a) aromatic urethan type protecting groups such as e.g. benzyloxycarbonyl and substituted benzyloxycarbonyl such as p-chlorobenzyloxycarbonyl, p-nitrobenzyloxycarbonyl, p-bromobenzyloxycarbonyl and p-methoxybenzyloxycarbonyl; (3) aliphatic urethan protecting groups such as e.g. α -t-butyloxycarbonyl, diisopropylmethoxycarbonyl, isopropylloxycarbonyl, ethoxycarbonyl and allyloxycarbonyl; (4) cycloalkyl urethan type protecting groups such as e.g. cyclopenthyloxycarbonyl, adamantyloxycarbonyl and cyclohexyloxycarbonyl; (5) thiourethan type protecting groups such as e.g. phenylthiocarbonyl; (6) alkyl type protecting groups such as e.g. triphenylmethyl (trityl) and benzyl; and (7) trialkylsilyl groups such as e.g. trimethylsilyl. The preferred α -amino protecting group defined by R is t-butyloxycarbonyl.

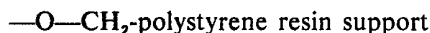
R¹ and R⁷ are each a thio protecting group for Cys or D-Cys selected from p-methoxybenzyl, p-methylbenzyl, acetamidomethyl, trityl and benzyl. The preferred protecting group is p-methoxybenzyl. R¹ and/or R⁷ can be hydrogen which means that there is no protecting group on the sulfur group.

R² and R³ are each a protecting group for the side chain amino substituent of lysine or R² and/or R³ are hydrogen which means there is no protecting group on the side chain amino substituent. Illustrative of suitable side chain amino protecting groups are benzyl, chlorobenzyloxycarbonyl (e.g. 2-chlorobenzyloxycarbonyl), benzyloxycarbonyl, tosyl, t-amyloxycarbonyl and t-

butyloxycarbonyl. The selection of such a side chain amino protecting group is not critical except that it must be one which is not removed during deprotection of the α -amino groups during the synthesis. Hence, the α -amino protecting and side chain amino protecting group cannot be the same.

R^4 , R^5 , and R^6 are protecting groups for the hydroxyl group of Thr and Ser and are selected from acetyl, benzoyl, tert-butyl, trityl, tetrahydropyranyl, benzyl, 2,6 - dichlorobenzyl and benzyloxycarbonyl. The preferred protecting group is benzyl. R^4 and/or R^5 and/or R^6 can be hydrogen which means there is no protecting group on the hydroxyl group.

X_1 , X_2 , X_3 and X_4 are as previously defined. X_5 is selected from OH, ester groups (e.g. OCH_3) amide groups, hydrazide groups and the benzyl ester or hydroxymethyl ester anchoring bond used in solid phase synthesis linked to a solid resin support represented by the formulae:



and



The polymer is preferably a copolymer of styrene with 0.5 to 2% (by weight) divinyl benzene as a cross linking agent which causes the polystyrene polymer to be completely insoluble in certain organic solvents. In formula III at least one of R , R^1 , R^2 , R^3 , R^4 , R^5 , R^6 and R^7 is a protecting group.

Examples of intermediates of formula III are those wherein X_1 is Asn, X_2 is Trp, X_3 is Ser and X_4 is D-Cys; wherein X_1 is Asn, X_2 is D-Trp, X_3 is Ser and X_4 is D-Cys; wherein X_1 is Asn, X_2 is Trp, X_3 is D-Ser and X_4 is Cys; and wherein X_1 is Asn, X_2 is D-Trp, X_3 is D-Ser and X_4 is Cys.

In selecting a particular side chain protecting group to be used in the synthesis of the peptides of formula I or formula II, the following rules should be followed:

(a) the protecting group must be stable to the reagent and under the reaction conditions selected for removing the α -amino protecting group at each step of the synthesis,

(b) the protecting group must retain its protecting properties and not be split off under coupling conditions, and

(c) the side chain protecting group must be removable upon the completion of the synthesis containing the desired amino acid sequence under reaction conditions that will not alter the peptide chain.

The peptides of formula I and formula II can be prepared using solid phase synthesis. The synthesis is commenced from the C-terminal end of the peptide using an α -amino protected resin. Such a starting material can be prepared by attaching an α -amino and S-protected Cys to a chloromethylated resin or a hydroxymethyl resin. The preparation of the hydroxymethyl resin is described by Bodanszky et al., Chem. Ind. (London) 38 1597-98 (1966). A chloromethylated resin is commercially available from Bio Rad Laboratories, Richmond, California and the preparation of such a resin is described by Stewart et al., "Solid Phase Peptide Synthesis" (Freeman & Co., San Francisco 1969), Chapter 1, pp 1-6. The α -amino and S-protected Cys is coupled to the chloromethylated resin according to the procedure of Monahan and Gilon, Biopolymer 12, pp 2513-19, 1973. Following the coupling of the α -amino and S-protected Cys to the resin support, the α -amino protecting group is removed such as by using trifluoroacetic acid in methylene chloride, trifluoroacetic acid alone or HCl in dioxane. The deprotection is carried out at a temperature between 0°C and room temperature.

Other standard cleaving reagents and conditions for removal of specific α -amino protecting groups may be used as described in Schroder & Lubke, "The Peptides", 1 pp 72-75 (Academic Press 1965).

After removal of the α -amino protecting group of Cys the remaining α -amino and side chain protected amino acids are coupled step-wise in the desired order to obtain a compound of formula III or as an alternate to adding each amino acid separately to the synthesis, some of them may be coupled prior to addition to the solid phase reactor. The selection of an appropriate coupling reagent is within the skill of the art. Particularly suitable as a coupling reagent is N,N' - dicyclohexylcarbodiimide.

The activating reagents used in the solid phase synthesis of the peptides are those well known in the peptide art. Examples of suitable activating reagents are: (1) carbodiimides such as N,N - diisopropylcarbodiimide, N - ethyl N' - (dimethylaminopropyl)carbodiimide; (2) cyanamides such as N,N -

dibenzylcyanamide; (3) keteimines; (4) isoxazolium salts such as N - ethyl - 5 - phenyl isoxazolium - 3' - sulfonate; (5) monocyclic nitrogen containing heterocyclic amides of aromatic character containing one through four nitrogens in the ring such as imidazolides, pyrazolides, 1,2,4 - triazolides. Specific heterocyclic amides that are useful include N,N' - carbonyl diimidazole, N,N' - carbonyl - di - 1,2,4 - triazole; (6) alkoxylated acetylene such as ethoxyacetylene; (7) reagents which form a mixed anhydride with the carboxyl moiety of the amino acid such as ethylchloroformate and isobutylchloroformate and (8) nitrogen-containing heterocyclic compounds having a hydroxy group on one ring nitrogen such as N - hydroxyphthalimide, N - hydroxysuccinimide and 1 - hydroxybenzotriazole. Other activating reagents and their use in peptide coupling are described by Schroder & Lubke supra, in Chapter III and by Kapoor, J. Pharm. Sci., 59, pp 1—27. (1970).

Each protected amino acid or amino acid sequence is introduced into the solid phase reactor in about a fourfold excess and the coupling is carried out in a medium of dimethylformamide: methylene chloride (1:1 by volume) or in dimethylformamide or methylene chloride alone. In cases where incomplete coupling occurred the coupling procedure is repeated before removal of the α -amino protecting group, prior to the coupling of the next amino acid. The success of the coupling reaction at each stage of the synthesis is monitored by the ninhydrin reaction, as described by E. Kaiser et al., *Analyt. Biochem.*, 34, 595 (1970).

After the desired amino acid sequence of formula III has been synthesized, the peptide is removed from the resin support by treatment with a reagent such as liquid hydrogen fluoride which not only cleaves the peptide from the resin but also cleaves all remaining side chain protecting groups R¹, R², R³, R⁴, R⁵, R⁶ and R⁷ and the α -amino protecting group R to obtain directly a peptide of formula II. Peptides in accordance with formula I are obtained by oxidizing formula II peptides in accordance with known procedures. As an alternate route, the peptide linked to the resin support may be separated from the resin by alcoholysis after which the recovered C-terminal methyl ester is converted to the acid by hydrolysis. Any side chain protecting group may then be cleaved as previously described or by other procedures such as catalytic reduction (e.g. Pd on BaSO₄) using conditions which will keep the Trp moiety intact. When using hydrogen fluoride for cleaving, anisole is included in the reaction vessel as a scavenger.

The solid phase synthesis procedure discussed above is well known in the art and has been essentially described by Merrifield J. Am. Chem. Soc., 85, p 2149 (1964).

The peptides of the present invention having dissociated effects in respect to inhibition of release of growth hormone, insulin and glucagon are considered to be particularly important in connection with the treatment of diabetes. The traditional view of diabetes has been that it is a disease resulting from impaired insulin production alone. As clinical and research experience has become more extensive, it has become apparent that some factor in addition to impairment of insulin secretion is operative in diabetes. It is known that, while insulin is normally deficient in diabetes, glucagon is normally present in excess. It is now believed that the presence of glucagon is at least as important a factor in diabetes as the absence of insulin.

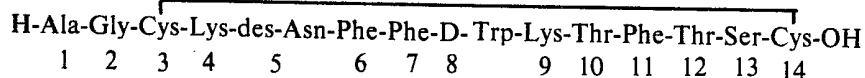
The fact that a deficiency in insulin is normally accompanied by an excess of glucagon has made it difficult to study the role of glucagon in diabetes. While it is easy to add extra quantities of a hormone such as insulin, it has proved very difficult to lower the concentration of glucagon. The discovery of somatostatin has facilitated research in respect to the role of glucagon in diabetes. Somatostatin inhibits the release of both insulin and glucagon. The role of somatostatin in diabetes research is detailed in an article appearing in *Science*, Vol. 188, pp 920—923, 30 May 1975. However, there are several problems in respect to the use of somatostatin as a treatment in diabetes. Somatostatin inhibits the release of insulin in addition to glucagon. Thus, the need for a peptide having a dissociated effect on the inhibition of release of insulin and glucagon has been recognized in connection with diabetes treatment. The novel peptides of the present invention provide such dissociative effect. More particularly, certain of the peptides of the present invention are effective to inhibit secretion of glucagon while having less effect on the inhibition of secretion of insulin.

The following Example illustrates the present invention but is not intended to limit the scope of the invention which is defined in the appended claims.

EXAMPLE

The peptides of the present invention were synthesized by solid phase techniques, generally in accordance with the procedure described in United States Patent Specification No. 3904594. The synthesis was conducted in a stepwise manner on chloromethylated resin. The resin was composed of fine beads (20—70 microns in diameter) of a synthetic resin prepared by copolymerization of styrene with one to two percent divinylbenzene. The benzene rings in the resin were chloromethylated in a Friedel-Crafts reaction with chloromethyl methyl ether and stannic chloride. The chlorine thus introduced is a reactive benzyl chloride type of linkage. The Friedel-Crafts reaction is continued until the resin contains 0.5 to 2 millimoles of chlorine per gram of resin. In the further description of the synthesis of the peptides, the reagents used will be first described by their chemical name with their common abbreviation in parenthesis. Thereafter, the reagent will be referred to by the common abbreviation. Furthermore, the specific peptide synthesized in the detailed description given hereinafter as illustrative of the procedure used is, of course, not included within the scope of the invention. Nonetheless the description of the solid phase methodology used is fully applicable to peptides of the invention.

A peptide having the structure:



was synthesized by the following solid phase methodology. Other peptides (these peptides being in accordance with the invention), described hereinafter, were synthesized by a similar technique.

The t - butyloxycarbonyl - S - p - methoxybenzyl derivative of Cys [Boc-Cys (S-pOMeBzl)-OH] was linked to the resin by any of the three known methods: (1) reflux in ethanol in presence of triethyl amine, (2) cesium salt of the Boc protected amino acid is kept at 50°C in dimethylformamide (DMF) overnight, (3) the potassium salt of the Boc-protected amino acid is kept at 80°C in dimethyl sulfoxide (DMSO) for 2 hours. Only one milliequivalent of the protected Cys per milliequivalent of Cl on the resin is used.

Method (3) is described hereinbelow in more detail: to a slurry of the resin and the dissolved protected Cys in DMSO is added 0.9 mEq of potassium t-butoxide (KOTBut) per mEq of amino acid. The reaction mixture is exposed to air as little as possible so that no amber coloration is observed. Reaction at 80°C for 2 hours yields a suitable substituted resin for synthesis of the peptides (approx. 0.2 meq of amino acid derivative per g of resin). After deprotection and neutralization, the peptide chain is built on the resin. Deprotection, neutralization and addition of each amino acid is performed in accordance with schedule I. $N\alpha$ - t - butyloxycarbonyl (Boc) derivative of each amino acid is used with the exception that any α -amino protecting group can be used for the alanine 1 residue provided it is cleaved by HF (benzyloxycarbonyl; Z; Boc and others). After $N\alpha$ -deprotection of the first residue according to schedule I (steps 3 to 8 inclusive) the $N\alpha$ Boc derivative of Ser is next added along with a coupling agent which is dicyclohexylcarbodiimide (DCC), (step 9 of schedule I). The side chain of Ser is protected with a benzyl group (Bzl). The benzyl(Bzl) protecting group is also used for protection of the threonine side chain. p - Nitro - phenyl ester (ONp) was used to activate the carboxyl end of Asn, if present. o - Nitrophenyl ester can also be used for this purpose. Formyl groups can be used for the protection of the indole N-H. Benzyloxycarbonyl (Z) or 2 - chlorobenzyloxycarbonyl [Z(2CL)] was used as the protecting group for the Lys side chain.

I Schedule for coupling of amino acids other than Asn in solid phase synthesis (5—10 g resin)

	Step	Reagents and operations (percentages are by weight)	Mix times Min.	
	1	CH ₂ Cl ₂ wash 80 ml (2 times)	3	
	2	Methanol (MeOH) wash 30 ml (2 times)	3	
5	3	CH ₂ Cl ₂ wash 80 ml (3 times)	3	5
	4	50 percent trifluoroacetic acid (TFA) containing 5 percent 1,2-ethanedithiol in CH ₂ Cl ₂ 70 ml (2 times)	10	
	5	CH ₂ Cl ₂ wash 80 ml (2 times)	3	
10	6	Triethylamine (Et ₃ N) 12.5 percent in CH ₂ Cl ₂ 70 ml (2 times)	5	10
	7	MeOH wash 40 ml (2 times)	2	
	8	CH ₂ Cl ₂ wash 80 ml (3 times)	3	
15	9	Boc-amino acid (10 mmoles) in 10 ml DMF (1 times) and 30 ml CH ₂ Cl ₂ plus DCC (10 mmoles) in CH ₂ Cl ₂ (2M)	30 to 120	15
	10	MeOH wash 40 ml (2 times)	3	
	11	Et ₃ N 12.5 percent in CH ₂ Cl ₂ 70 ml (2 times)	3	
	12	MeOH wash 30 ml (2 times)	3	
20	13	CH ₂ Cl ₂ wash 80 ml (2 times)	3	20

After step 13, an aliquot is taken for a ninhydrin test:

25	if the test is negative, go back to step 1 for coupling of the next amino acid; if the test is positive, go back to steps 9 to 13. Schedule I was used for coupling of each of the amino acids of the peptide to Cys with the exception of Asn, when present. For peptides of the invention containing Asn, steps 1 through 8 are the same and schedule II is used for the remainder of the coupling reaction:			25
----	--	--	--	----

II Schedule for Boc-Asn-ONp or for any active ester coupling in solid phase synthesis (5—10 g resin)

	Step	Reagents and operations	Mix times Min.	
30	9	DMF wash 60 ml (3 times)	3	30
	10	Boc-Asn-ONp (15 mmoles) in 20 ml DMF (1 time)	800	
	11	MeOH wash 30 ml (4 times)	3	
35	12	Et ₃ N 12.5 percent (by weight) in DMF 30 ml (2 times)	3	35
	13	MeOH wash 30 ml (2 times)	3	
	14	CH ₂ Cl ₂ wash 80 ml (3 times)	3	

After step 14, an aliquot is taken for a ninhydrin test:

40	If the test is negative go back to step 1 for coupling of the next amino acid; if the test is positive, go back to steps 9 to 14.			40
----	---	--	--	----

45	Cleavage of the peptides from the resin (5 grams) and deprotection of the side chain protecting groups of the peptide was performed in hydrofluoric acid (75 ml) in the presence of anisole (8 ml). After elimination of hydrofluoric acid under high vacuum, the resin-peptide was washed with diethyl ether.			45
----	--	--	--	----

50	The dried resin was immediately extracted with 25% (by weight) acetic acid (150 ml) and diluted to 3000 ml with degassed H ₂ O(N ₂). The pH of the solution was adjusted to 6.6—7.0 with NH ₄ OH. The solution was titrated dropwise under stirring with potassium ferricyanide solution (1 g/500 ml H ₂ O) until a permanent yellow color was observed. The solution sat for 10 minutes and pH was adjusted to 5.0 with glacial acetic acid; Bio-Rad (registered Trade Mark) AG 3-X4A resin (100—200 mesh—U.S. Standard Sieve Series, chloride form, 10—15 g) was added to the turbid solution and stirred for 15 minutes. The solution was filtered over Celite (registered Trade Mark) and applied successively onto two columns; a) Bio-Rad AG 3-X4A resin chloride form (10 ml); b) Biorex (Registered Trade Mark) -70 resin (100 ml) cation form. The Celite+resin cake was thoroughly washed with water (500 ml) which was applied onto columns a) and b) as a wash. The peptide material was then eluted from the Biorex-70 resin column with pyridine:acetic acid:water (30:4:66 by volume) or 50% (by weight) acetic acid. Fractions were collected; only the ones containing peptide (ninhydrin positive) were diluted with water and immediately lyophilized. 1.2 g of crude cream colored material was			50
55				55
60				60

obtained. It was applied onto a Sephadex (registered Trade Mark) G-25 F gel column (3×200 cm) equilibrated and eluted with 2N acetic acid.

The elution pattern as observed at 280 nm showed one major symmetrical peak centered at V_0 (400 mg). It was subsequently submitted to counter current distribution (solvent system n-butanol:acetic acid:water, 4:1:5 by volume) 10 ml lower phase per tube. 237 transfers were performed and the major peak was found in tubes 48—64. The compound (250 mg) appeared homogeneous on tlc.

The specific optical rotation was $[\alpha]^{23}_D = -38.2 \pm 1^\circ$ ($c=1$ in 1% by weight acetic acid). Amino acid analysis of this material showed the expected ratio for the different amino acids.

Active esters can be used in solid phase synthesis and the classical method of synthesis can also be used to prepared the peptides of the invention.

In vitro Bioassay: The effects of the various peptides of the invention were tested *in vitro* on the secretion of growth hormone by primary cultures of enzymatically dissociated rat anterior pituitary cells by the method of Vale et al., Endocrinology 91: p 562—571 (1972). The assay is made by treating pituitary glands removed from rats to separate cells therefrom. The cells are placed in culture dishes in a modification of Dulbecco's Modified Eagle Medium. (Vale et al, Methods in Enzymology). Carbon dioxide gas and oxygen are supplied to the cell cultures which are maintained at 37°C for 4—5 days prior to use in the assay. Following media changes, cell cultures are incubated for a period of 4 hours and particular somatostatin peptides are added thereto. Radioimmunoassay analysis is used to determine the rate of growth hormone secretion which is expressed in nanograms per dish per hour.

An investigation of the effect of somatostatin, dihydrosomatostatin, (as controls) and the peptides of the invention to inhibit the release of glucagon and insulin was made as follows:

In vivo Bioassay: Male Sprague-Dawley-CD rats weighing 180—200 g housed in temperature and humidity controlled quarters with 14h light and 10h dark (light 0700—21100) were used in all experiments. Animals were fed a standard ration and tap water *ad libitum*. Experiments were carried out at least 5 days after arrival of rats from the supplier between the hours 1400 and 1600. After either anesthesia, peptides or saline were administered in a volume of 0.2 ml. via the external jugular vein. Animals remained anesthetized until the time of blood collection from the portal vein. The blood samples were placed into chilled tubes containing 10 mg EDTA and 50 μ l of 2M benzamidine per ml of blood.

Plasma was stored at -20°C for insulin and glucagon determinations. Insulin levels were determined by the method of Herbet *et al*, J. Chem. Endocr. Metab. 25:1375, 1965, utilizing porcine insulin antisera and (125I) iodinated insulin tracer. Human insulin standard was obtained from Schwarz-Mann, Orangeburg, New York. Glucagon was determined by the method of Faloona and Unger, in Jaffe et al ed., Method of Hormone Radioimmunoassay, Academic Press, New York, 1974, p. 317, utilizing glucagon antisera 30K. Glucose was determined by the glucose oxidase method, utilizing a Beckman (registered Trade Mark) Glucose Analyzer.

GH determinations were performed on tissue culture media utilizing the following reagents: NIAMDD rat GH standard (GH-RP-1), NIAMDD monkey anti-rat GH (GH-Serum-3), and highly purified rat GH for iodination.

All experiments are carried on in a randomized block design. Following analysis of variance difference between treatments were determined by the multiple range tests of Dunnett and Dunca. Potency values were calculated from four or six point bioassays.

Various peptides in accordance with the invention were prepared in accordance with the solid phase methodology described above. The composition of the peptides is reported hereinbelow in Table I. Table I also sets forth the ratio of effectiveness of the peptide for inhibiting secretion of growth hormone (GH), insulin and glucagon, with inhibition of glucagon taken as the base. Also reported in Table I is the percent potency of the peptide in respect to growth hormone inhibition with somatostatin considered as being 100 percent effective.

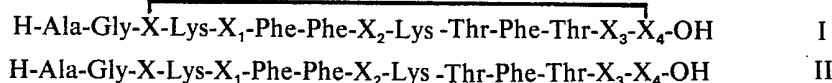
TABLE I

	Somatostatin (control)	Ratio of Effectiveness			%Potency Growth Hormone Base 100	
		GH	Insulin	Glucagon		
5		1	1	1		
	[D-Ser ¹³]SS	10	10	1	10	5
	[D-Trp ⁸]-[D-Ser ¹³]-SS	18	261	<1		
	[D-Cys ¹⁴]-SS	2.7	.1	1	270	
	[D-Trp ⁸]-[D-Cys ¹⁴]-SS	.7	.13	1	650	
10	[D-Cys ³]-[D-Ser ¹³]-SS	:	7	<1		10

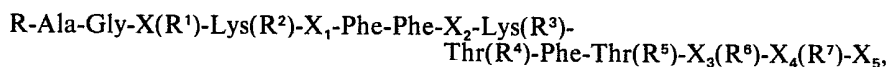
It should be clearly understood that we make no claim herein to a method for the treatment or prevention of disease in a human being.

Subject to the foregoing disclaimer,
WHAT WE CLAIM IS:—

- 15 1. A peptide selected from those of the formulae: 15



and



- 20 wherein X is Cys or D-Cys; X₁ is Asn or a direct bond; X₂ is Trp or D-Trp; X₃ is Ser or D-Ser; and X₄ is Cys or D-Cys; provided that whenever X₂ is D-Trp, X₃ is D-Ser and/or X₄ is D-Cys, and that X₃ is D-Ser whenever X₁ is Asn and X₄ is Cys, and that X₄ is D-Cys whenever X₁ is Asn and X₃ is Ser; R is H or an alpha-amino protecting group; R¹ and R⁷ are each independently H or a thio protecting group for Cys or D-Cys selected from p-methoxybenzyl, p-methylbenzyl, acetamidomethyl, trityl and benzyl; R² and R³ are each independently H or a side chain amino protecting group; R⁴, R⁵ and R⁶ are each independently H or a hydroxyl protecting group selected from acetyl, benzoyl, tert-butyl, trityl, tetrahydropyranyl, 2,6-dichlorobenzyl, benzyl and benzyloxycarbonyl; provided that at least one of R, R¹, R², R³, R⁴, R⁵, R⁶ and R⁷ is other than hydrogen; and X₅ is selected from hydroxy, ester groups, amide groups, hydrazide groups, —O—CH₂-polystyrene resin support and O—CH₂-benzyl-polystyrene resin support.
- 25 2. A peptide in accordance with formula I of Claim 1, wherein X₁ is Asn, X₂ is Trp, X₃ is Ser and X₄ is D-Cys.
- 35 3. A peptide in accordance with formula I of Claim 1, wherein X₁ is Asn, X₂ is D-Trp, X₃ is Ser and X₄ is D-Cys.
4. A peptide in accordance with formula I of Claim 1, wherein X₁ is Asn, X₂ is Trp, X₃ is D-Ser and X₄ is Cys.
- 40 5. A peptide in accordance with formula I of Claim 1, wherein X₁ is Asn, X₂ is D-Trp, X₃ is D-Ser and X₄ is Cys.
6. A peptide in accordance with formula II of Claim 1, wherein X₁ is Asn, X₂ is Trp, X₃ is Ser and X₄ is D-Cys.
7. A peptide in accordance with formula II of Claim 1, wherein X₁ is Asn, X₂ is D-Trp, X₃ is Ser and X₄ is D-Cys.
- 45 8. A peptide in accordance with formula II of Claim 1, wherein X₁ is Asn, X₂ is Trp, X₃ is D-Ser and X₄ is Cys.
9. A peptide in accordance with formula II of Claim 1, wherein X₁ is Asn, X₂ is D-Trp, X₃ is D-Ser and X₄ is Cys.
- 50 10. A peptide in accordance with formula III of Claim 1, wherein X₁ is Asn, X₂ is Trp, X₃ is Ser and X₄ is D-Cys.
11. A peptide in accordance with formula III of Claim 1, wherein X₁ is Asn, X₂ is D-Trp, X₃ is Ser and X₄ is D-Cys.
12. A peptide in accordance with formula III of Claim 1, wherein X₁ is Asn, X₂ is Trp, X₃ is D-Ser and X₄ is Cys.
- 55 13. A peptide in accordance with formula III of Claim 1, wherein X₁ is Asn, X₂ is D-Trp, X₃ is D-Ser and X₄ is Cys.

14. A peptide in accordance with formula III of Claim 1, wherein X_5 is $-\text{O}-\text{CH}_2-$ - polystyrene resin support.

15. A peptide in accordance with formula III of Claim 1, wherein X_5 is $-\text{O}-\text{CH}_2-$ - benzyl - polystyrene resin support.

16. A peptide in accordance with formula III or Claim 1, wherein X_5 is methoxy.

17. A peptide as claimed in Claim 1, wherein R , R^2 , and/or R^3 is/are a group/groups referred to hereinbefore in exemplification of R , R^2 , and/or R^3 .

18. A peptide as claimed in Claim 14, wherein R is t-butyloxycarbonyl, R^4 , R^5 and R^6 are benzyl, R^2 and R^3 are 2 - chlorobenzyloxycarbonyl and R^1 and R^7 are p-methoxybenzyl.

19. A peptide as claimed in any one of claims 1 to 18, wherein X is D-Cys.

20. A method for the synthesis of a peptide as claimed in any one of the preceding claims and substantially as hereinbefore described.

21. A method for the synthesis of a peptide in accordance with formula I, II or III of Claim 1 and substantially as hereinbefore described in any one of the individual runs of the foregoing Example.

22. A peptide which has been synthesised by a method as claimed in claim 20 or claim 21.

23. A pharmaceutically acceptable or veterinarily acceptable acid addition salt of a peptide as claimed in any one of claims 1 to 19 or 22.

24. A pharmaceutical or veterinary formulation which comprises a peptide in accordance with formula I or II and as claimed in any one of claims 1 to 19 or 22 or a salt as claimed in Claim 23 being a salt of a peptide in accordance with formula I or II formulated for pharmaceutical or veterinary use, respectively.

25. A pharmaceutical or veterinary composition, which comprises a peptide in accordance with formula I or II and as claimed in any one of claims 1 to 19 or 22 or a salt as claimed in Claim 23 being a salt of a peptide in accordance with formula I or II together with a pharmaceutically acceptable or veterinarily acceptable, respectively, diluent, carrier or excipient.

26. A formulation as claimed in Claim 24 or a composition as claimed in Claim 25 and in unit dosage form.

27. A method for inhibiting the secretion of growth hormone, insulin or glucagon in an animal, which method comprises administering to the animal a peptide in accordance with formula I or II and as claimed in any one of claims 1 to 19 or 22 or a salt as claimed in Claim 26 being a salt of a peptide in accordance with formula I or II, a formulation as claimed in Claim 24 or Claim 26, or a composition as claimed in Claim 25 or Claim 26.

THE SALK INSTITUTE FOR BIOLOGICAL STUDIES

Per: Boulton Wade & Tennant,
27 Furnival Street
London EC4A 1PQ
Chartered Patent Agents

Printed for Her Majesty's Stationery Office, by the Courier Press, Leamington Spa, 1980
Published by The Patent Office, 25 Southampton Buildings, London, WC2A 1AY, from
which copies may be obtained.