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(54) Title: USE OF CELL-PERMEABLE PEPTIDE INHIBITORS OF THE JNK SIGNAL TRANSDUCTION PATHWAY FOR THE TREATMENT OF CHRONIC OR NON-CHRONIC INFLAMMATORY EYE DISEASES

(57) Abstract: The present invention refers to the use of protein kinase inhibitors and more specifically to the use of inhibitors of the protein kinase c-Jun amino terminal kinase, JNK inhibitor (poly-)peptides, chimeric peptides, or of nucleic acids encoding same as well as pharmaceutical compositions containing same, for the treatment of non-chronic or chronic inflammatory eye diseases, such as inflammatory diseases of the blephara, conjunctiva, cornea, sclera, the vitreous body, uvea, ciliary body, choroid, orbital bone, lacrimal gland, or iris, in particular wherein the inflammatory disease is selected from hordeolum, chalazion, conjunctivitis, keratitis, scleritis, episcleritis, endophthalmitis, panophthalmitis, iritis, uveitis, cyclitis, chorioiditis, orbital phlegmon, and myositis of the eye muscle etc..



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Use of cell-permeable peptide inhibitors of the JNK signal transduction pathway for the treatment of chronic or non-chronic inflammatory eye diseases

10 The present invention refers to the use of protein kinase inhibitors and more specifically to the use of inhibitors of the protein kinase c-Jun amino terminal kinase, JNK inhibitor (poly-)peptides, chimeric peptides, or of nucleic acids encoding same as well as pharmaceutical compositions containing same, for the treatment of non-chronic or chronic inflammatory eye diseases, such as hordeolum, chalazion, conjunktivitis, keratitis, scleritis, episcleritis, 15 endophthalmitis, panophthalmitis, iritis, uveitis, cyclitis, chorioiditis, orbital phlegmon, and/or myositis of the eye muscle, etc..

The number of ophthalmological (eye) diseases, particularly of non-chronic and chronic ophthalmological (eye) diseases represents a considerable challenge for the public health 20 care systems. Ophthalmological diseases are diseases that pertain to the eye. The present invention focuses on non-chronic or chronic inflammatory eye diseases. These include for example inflammatory diseases of the blephara, conjunctiva, cornea, sclera, the vitreous body, uvea, ciliary body, choroid, orbital bone, lacrimal gland, iris, etc. Examples of such inflammatory diseases are hordeolum, chalazion, conjunktivitis, keratitis, scleritis, 25 episcleritis, endophthalmitis, panophthalmitis, iritis, uveitis, cyclitis, chorioiditis, orbital phlegmon, and/or myositis of the eye muscle.

The c-Jun NH2-terminal kinases (JNKs) have been identified as stress-activated protein kinases that phosphorylate c-Jun on two sites in its NH2-terminal activation domain. The 30 JNK pathway is activated by certain cytokines, mitogens, osmotic stress and irradiation. The phosphorylation of the c-Jun component of the activator protein AP-1 transcription factor results in pro-inflammatory cytokines production. During inflammation, leukocytes infiltration and rolling result from the early activation of the vascular endothelium that releases important chemotactic factors such as RANTES, IL-8, ICAM and VCAM. Infiltrating

cells in turn release distinct sets of pro- or anti- inflammatory products that contribute to tissue damages and inflammation. Many of the gene products involved in the inflammatory response are regulated by the transcription factor activator protein-1 (AP-1), and the c-Jun NH2-terminal kinase (JNK) pathway: COX-2, cyclooxygenase-2; IFN-g, interferon-gamma; iNOS, inducible nitric oxide synthase; TNF-a, tumor-necrosis factor-alpha; MCP-1, membrane cofactor protein-1; MIP-1, major intrinsic protein-1; IL-2, interleukin-2, ... In lipopolysaccharide (LPS)-stimulated monocytes and tissue macrophages, TNF-a is produced through the JNK pathway activation and modulated by its inhibition.

- 10 JNK inhibitors have been therefore used in various models of inflammation and shown to exert anti-inflammatory and beneficial effects in inflammatory diseases such arthritis and asthma.

The object of the present invention is thus to provide alternative or improved therapies, which allow new and preferably improved cure of non-chronic or chronic (inflammatory) eye diseases, such as hordeolum, chalazion, conjunktivitis, keratitis, scleritis, episcleritis, endophthalmitis, panophtalmitis, iritis, uveitis, cyclitis, chorioiditis, orbital phlegmon, myositis of the eye muscle,, etc..

- 20 This object is solved by the use of a JNK inhibitor (poly-)peptide comprising less than 150 amino acids in length for the preparation of a pharmaceutical composition for treating non-chronic or chronic inflammatory eye diseases in a subject.

The term "non-chronic or chronic inflammatory eye disease" as used herein typically denotes non-chronic or chronic inflammatory diseases that pertain to the eye. This includes diseases of the blephara, conjunctiva, cornea, sclera, the vitreous body, uvea, ciliary body, choroid, orbital bone, lacrimal gland, iris, etc.. Preferably included in this respect are hordeolum, chalazion, conjunktivitis, keratitis, scleritis, episcleritis, endophthalmitis, panophtalmitis, iritis, uveitis, cyclitis, chorioiditis, orbital phlegmon, myositis of the eye muscle. Particularly preferred in the context of the present invention is the treatment of uveitis, for example treatment of anterior uveitis, intermediate uveitis, posterior uveitis and panuveitis.

The present inventors surprisingly found, that JNK inhibitor (poly-)peptides are particularly suitable for treating such chronic or non-chronic inflammatory eye diseases in a subject. This was neither obvious nor suggested by the prior art, even though JNK inhibitor (poly-)peptides in general have been known from the art.

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In the context of the present invention, a JNK inhibitor (poly-)peptide may be typically derived from a human or rat IB1 sequence, preferably from an amino acid sequence as defined or encoded by any of sequences according to SEQ ID NO: 102 (depicts the IB1 cDNA sequence from rat and its predicted amino acid sequence), SEQ ID NO: 103 (depicts the IB1 protein sequence from rat encoded by the exon-intron boundary of the *rlB1* gene – splice donor), SEQ ID NO: 104 (depicts the IB1 protein sequence from *Homo sapiens*), or SEQ ID NO: 105 (depicts the IB1 cDNA sequence from *Homo sapiens*), more preferably from an amino acid sequence as defined or encoded by any of sequences according to SEQ ID NO: 104 (depicts the IB1 protein sequence from *Homo sapiens*), or SEQ ID NO: 105 (depicts the IB1 cDNA sequence from *Homo sapiens*), or from any fragments or variants thereof. In other words, the JNK inhibitor (poly-)peptide comprises a fragment, variant, or variant of such fragment of a human or rat IB1 sequence. Human or rat IB sequences are defined or encoded, respectively, by the sequences according to SEQ ID NO: 102, SEQ ID NO: 103, SEQ ID NO: 104 or SEQ ID NO: 105.

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Preferably, such a JNK inhibitor (poly-)peptide as used herein comprises a total length of less than 150 amino acid residues, preferably a range of 5 to 150 amino acid residues, more preferably 10 to 100 amino acid residues, even more preferably 10 to 75 amino acid residues and most preferably a range of 10 to 50 amino acid residues, e.g. 10 to 30, 10 to 20, or 10 to 15 amino acid residues.

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More preferably, such a JNK inhibitor (poly-)peptide and the above ranges may be selected from any of the above mentioned sequences, even more preferably from an amino acid sequence as defined according to SEQ ID NO: 104 or as encoded by SEQ ID NO: 105, even more preferably in the region between nucleotides 420 and 980 of SEQ ID NO: 105 or amino acids 105 and 291 of SEQ ID NO: 104, and most preferably in the region between nucleotides 561 and 647 of SEQ ID NO: 105 or amino acids 152 and 180 of SEQ ID NO: 104.

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According to a particular embodiment, a JNK inhibitor (poly-)peptide as used herein typically binds JNK and/or inhibits the activation of at least one JNK activated transcription factor, e.g. c-Jun or ATF2 (see e.g. SEQ ID NOs: 15 and 16, respectively) or Elk1.

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Likewise, the JNK inhibitor (poly-)peptide as used herein preferably comprises or consists of at least one amino acid sequence according to any one of SEQ ID NOs: 1 to 4, 13 to 20 and 33 to 100, or a fragment, derivative or variant thereof. More preferably, the JNK inhibitor (poly-)peptide as used herein may contain 1, 2, 3, 4 or even more copies of an amino acid sequence according to SEQ ID NOs: 1 to 4, 13 to 20 and 33 to 100, or a variant, fragment or derivative thereof. If present in more than one copy, these amino acid sequences according to SEQ ID NOs: 1 to 4, 13 to 20 and 33 to 100, or variants, fragments, or derivatives thereof as used herein may be directly linked with each other without any linker sequence or via a linker sequence comprising 1 to 10, preferably 1 to 5 amino acids.

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Amino acids forming the linker sequence are preferably selected from glycine or proline as amino acid residues. More preferably, these amino acid sequences according to SEQ ID NOs: 1 to 4, 13 to 20 and 33 to 100, or fragments, variants or derivatives thereof, as used herein, may be separated by each other by a hinge of two, three or more proline residues.

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The JNK inhibitor (poly-)peptides as used herein may be composed of L-amino acids, D-amino acids, or a combination of both. Preferably, the JNK inhibitor (poly-)peptides as used herein comprise at least 1 or even 2, preferably at least 3, 4 or 5, more preferably at least 6, 7, 8 or 9 and even more preferably at least 10 or more D- and/or L-amino acids, wherein the D- and/or L-amino acids may be arranged in the JNK inhibitor sequences as used herein in a blockwise, a non-blockwise or in an alternate manner.

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According to one preferred embodiment the JNK inhibitor (poly-)peptides as used herein may be exclusively composed of L-amino acids. The JNK inhibitor (poly-)peptides as used herein may then comprise or consist of at least one „native JNK inhibitor sequence“ according to SEQ ID NO: 1 or 3. In this context, the term “native” or “native JNK inhibitor sequence(s)” is referred to non-altered JNK inhibitor (poly-)peptide sequences according to any of SEQ ID NOs: 1 or 3, as used herein, entirely composed of L-amino acids.

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Accordingly, the JNK inhibitor (poly-)peptide as used herein may comprise or consist of at least one (native) amino acid sequence $\text{NH}_2\text{-X}_n^b\text{-X}_n^a\text{-RPTTLXLXXXXXXXQD-X}_n^b\text{-COOH}$ (L-IB generic (s)) [SEQ ID NO: 3] and/or the JNK binding domain (JBDs) of IB1 XRPTTLXLXXXXXXXQDS/TX (L-IB (generic)) [SEQ ID NO: 19]. In this context, each X typically represents an amino acid residue, preferably selected from any (native) amino acid residue. X_n^a typically represents one amino acid residue, preferably selected from any amino acid residue except serine or threonine, wherein n (the number of repetitions of X) is 0 or 1. Furthermore, each X_n^b may be selected from any amino acid residue, wherein n (the number of repetitions of X) is 0-5, 5-10, 10-15, 15-20, 20-30 or more, provided that if n (the number of repetitions of X) is 0 for X_n^a , X_n^b does preferably not comprise a serine or threonine at its C-terminus, in order to avoid a serine or threonine at this position. Preferably, X_n^b represents a contiguous stretch of peptide residues derived from SEQ ID NO: 1 or 3. X_n^a and X_n^b may represent either D or L amino acids. Additionally, the JNK inhibitor (poly-)peptide as used herein may comprise or consist of at least one (native) amino acid sequence selected from the group comprising the JNK binding domain of IB1 DTYRPKRPTTLNLFQVPRSQDT (L-IB1) [SEQ ID NO: 17]. More preferably, the JNK inhibitor (poly-)peptide as used herein further may comprise or consist of at least one (native) amino acid sequence $\text{NH}_2\text{-RPKRPTTLNLFQVPRSQD-COOH}$ (L-IB1(s)) [SEQ ID NO: 1]. Furthermore, the JNK inhibitor (poly-)peptide as used herein may comprise or consist of at least one (native) amino acid sequence selected from the group comprising the JNK binding domain of IB1 L-IB1(s1) ($\text{NH}_2\text{-TLNLFQVPRSQD-COOH}$, SEQ ID NO: 33); L-IB1(s2) ($\text{NH}_2\text{-TTLNLFQVPRSQ-COOH}$, SEQ ID NO: 34); L-IB1(s3) ($\text{NH}_2\text{-PTTLNLFQVPRS-COOH}$, SEQ ID NO: 35); L-IB1(s4) ($\text{NH}_2\text{-RPTTLNLFQVPR-COOH}$, SEQ ID NO: 36); L-IB1(s5) ($\text{NH}_2\text{-KRPTTLNLFQVP-COOH}$, SEQ ID NO: 37); L-IB1(s6) ($\text{NH}_2\text{-PKRPTTLNLFQV-COOH}$, SEQ ID NO: 38); L-IB1(s7) ($\text{NH}_2\text{-RPKRPTTLNLFQ-COOH}$, SEQ ID NO: 39); L-IB1(s8) ($\text{NH}_2\text{-LNLFPQVPRSQD-COOH}$, SEQ ID NO: 40); L-IB1(s9) ($\text{NH}_2\text{-TLNLFQVPRSQ-COOH}$, SEQ ID NO: 41); L-IB1(s10) ($\text{NH}_2\text{-TTLNLFQVPRS-COOH}$, SEQ ID NO: 42); L-IB1(s11) ($\text{NH}_2\text{-PTTLNLFQVPR-COOH}$, SEQ ID NO: 43); L-IB1(s12) ($\text{NH}_2\text{-RPTTLNLFQVP-COOH}$, SEQ ID NO: 44); L-IB1(s13) ($\text{NH}_2\text{-KRPTTLNLFQV-COOH}$, SEQ ID NO: 45); L-IB1(s14) ($\text{NH}_2\text{-PKRPTTLNLFQ-COOH}$, SEQ ID NO: 46); L-IB1(s15) ($\text{NH}_2\text{-RPKRPTTLNLF-COOH}$, SEQ ID NO: 47); L-IB1(s16) ($\text{NH}_2\text{-NLFPQVPRSQD-COOH}$, SEQ ID NO: 48); L-IB1(s17) ($\text{NH}_2\text{-LNLFPQVPRSQ-COOH}$, SEQ ID NO: 49); L-IB1(s18) ($\text{NH}_2\text{-TLNLFQVPRS-COOH}$, SEQ ID NO: 50); L-IB1(s19) ($\text{NH}_2\text{-TTLNLFQVPR-COOH}$, SEQ ID NO: 51); L-IB1(s20) ($\text{NH}_2\text{-$

PTTLNLFQVP-COOH, SEQ ID NO: 52); L-IB1(s21) (NH₂-RPTTLNLFQV-COOH, SEQ ID NO: 53); L-IB1(s22) (NH₂-KRPTTLNLFQ-COOH, SEQ ID NO: 54); L-IB1(s23) (NH₂-PKRPTTLNLF-COOH, SEQ ID NO: 55); L-IB1(s24) (NH₂-RPKRPTTLNLF-COOH, SEQ ID NO: 56); L-IB1(s25) (NH₂-LFPQVPRSQD-COOH, SEQ ID NO: 57); L-IB1(s26) (NH₂-NLFPQVPRSQ-COOH, SEQ ID NO: 58); L-IB1(s27) (NH₂-LNLFPQVPRS-COOH, SEQ ID NO: 59); L-IB1(s28) (NH₂-TLNLFQVPR-COOH, SEQ ID NO: 60); L-IB1(s29) (NH₂-TTLNLFQVP-COOH, SEQ ID NO: 61); L-IB1(s30) (NH₂-PTTLNLFQV-COOH, SEQ ID NO: 62); L-IB1(s31) (NH₂-RPTTLNLFQ-COOH, SEQ ID NO: 63); L-IB1(s32) (NH₂-KRPTTLNLF-COOH, SEQ ID NO: 64); L-IB1(s33) (NH₂-PKRPTTLNLF-COOH, SEQ ID NO: 65); and L-IB1(s34) (NH₂-RPKRPTTLNL-COOH, SEQ ID NO: 66).

Additionally, the JNK inhibitor (poly-)peptide as used herein may comprise or consist of at least one (native) amino acid sequence selected from the group comprising the (long) JNK binding domain (JBDs) of IB1 PGTGCGDTYRPKRPTTLNLFQVPRSQDT (IB1-long) [SEQ ID NO: 13], the (long) JNK binding domain of IB2 IPSPSVEEPHKHRPTTLRLTLGAQDS (IB2-long) [SEQ ID NO: 14], the JNK binding domain of c-Jun GAYGYSNPILKQSMTLNLADPVGNLKPH (c-Jun) [SEQ ID NO: 15], the JNK binding domain of ATF2 TNEDHLAVHKKHHEMTLKFPGPARNDVIV (ATF2) [SEQ ID NO: 16] (see e.g. FIGS. 1A-1C). In this context, an alignment revealed a partially conserved 8 amino acid sequence (see e.g. FIG. 1A) and a further comparison of the JBDs of IB1 and IB2 revealed two blocks of seven and three amino acids that are highly conserved between the two sequences.

According to another preferred embodiment the JNK inhibitor (poly-)peptides as used herein may be composed in part or exclusively of D-amino acids as defined above. More preferably, these JNK inhibitor (poly-)peptides composed of D-amino acids are non-native D retro-inverso sequences of the above (native) JNK inhibitor sequences. The term "retro-inverso (poly-)peptides" refers to an isomer of a linear peptide sequence in which the direction of the sequence is reversed and the chirality of each amino acid residue is inverted (see e.g. Jameson *et al.*, Nature, 368,744-746 (1994); Brady *et al.*, Nature, 368,692-693 (1994)). The advantage of combining D-enantiomers and reverse synthesis is that the positions of carbonyl and amino groups in each amide bond are exchanged, while the position of the side-chain groups at each alpha carbon is preserved. Unless specifically

stated otherwise, it is presumed that any given L-amino acid sequence or peptide as used according to the present invention may be converted into an D retro-inverso sequence or peptide by synthesizing a reverse of the sequence or peptide for the corresponding native L-amino acid sequence or peptide.

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The D retro-inverso (poly-)peptides as used herein and as defined above have a variety of useful properties. For example, D retro-inverso (poly-)peptides as used herein enter cells as efficiently as L-amino acid sequences as used herein, whereas the D retro-inverso sequences as used herein are more stable than the corresponding L-amino acid sequences.

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Accordingly, the JNK inhibitor (poly-)peptides as used herein may comprise or consist of at least one D retro-inverso sequence according to the amino acid sequence $\text{NH}_2\text{-X}_n^b\text{-DQXXXXXXXXLXLTPR-X}_n^a\text{-X}_n^b\text{-COOH}$ (D-IB1 generic (s)) [SEQ ID NO: 4] and/or $\text{XS/TDQXXXXXXXXLXLTPRX}$ (D-IB (generic)) [SEQ ID NO: 20]. As used in this context, X,

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X_n^a and X_n^b are as defined above (preferably, representing D amino acids), wherein X_n^b preferably represents a contiguous stretch of residues derived from SEQ ID NO: 2 or 4.

Additionally, the JNK inhibitor (poly-)peptides as used herein may comprise or consist of at least one D retro-inverso sequence according to the amino acid sequence comprising the JNK binding domain (JBDs) of IB1 $\text{TDQSRPVQPFLNLTPRKPRYTD}$ (D-IB1) [SEQ ID NO:

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18]. More preferably, the JNK inhibitor (poly-)peptides as used herein may comprise or consist of at least one D retro-inverso sequence according to the amino acid sequence $\text{NH}_2\text{-DQSRPVQPFLNLTPRKPR-COOH}$ (D-IB1(s)) [SEQ ID NO: 2]. Furthermore, the JNK inhibitor (poly-)peptides as used herein may comprise or consist of at least one D retro-

inverso sequence according to the amino acid sequence comprising the JNK binding

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domain (JBDs) of IB1 D-IB1(s1) ($\text{NH}_2\text{-QPFLNLTPRKPR-COOH}$, SEQ ID NO: 67); D-IB1(s2) ($\text{NH}_2\text{-VQPFLNLTPRKPR-COOH}$, SEQ ID NO: 68); D-IB1(s3) ($\text{NH}_2\text{-PVQPFLNLTPRK-COOH}$, SEQ ID NO: 69); D-IB1(s4) ($\text{NH}_2\text{-RPVQPFLNLTPR-COOH}$, SEQ ID NO: 70); D-

IB1(s5) ($\text{NH}_2\text{-SRPVQPFLNLTP-COOH}$, SEQ ID NO: 71); D-IB1(s6) ($\text{NH}_2\text{-QSRPVQPFLNLTP-COOH}$, SEQ ID NO: 72); D-IB1(s7) ($\text{NH}_2\text{-DQSRPVQPFLNLTP-COOH}$,

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SEQ ID NO: 73); D-IB1(s8) ($\text{NH}_2\text{-PFLNLTPRKPR-COOH}$, SEQ ID NO: 74); D-IB1(s9) ($\text{NH}_2\text{-QPFLNLTPRKPR-COOH}$, SEQ ID NO: 75); D-IB1(s10) ($\text{NH}_2\text{-VQPFLNLTPRK-COOH}$, SEQ

ID NO: 76); D-IB1(s11) ($\text{NH}_2\text{-PVQPFLNLTPR-COOH}$, SEQ ID NO: 77); D-IB1(s12) ($\text{NH}_2\text{-RPVQPFLNLTP-COOH}$, SEQ ID NO: 78); D-IB1(s13) ($\text{NH}_2\text{-SRPVQPFLNLTP-COOH}$, SEQ

ID NO: 79); D-IB1(s14) (NH₂-QSRPVQPFLNLT-COOH, SEQ ID NO: 80); D-IB1(s15) (NH₂-DQSRPVQPFLNL-COOH, SEQ ID NO: 81); D-IB1(s16) (NH₂-FLNLTTTPRKPR-COOH, SEQ ID NO: 82); D-IB1(s17) (NH₂-PFLNLTTTPRK-COOH, SEQ ID NO: 83); D-IB1(s18) (NH₂-QPFLNLTTTPRK-COOH, SEQ ID NO: 84); D-IB1(s19) (NH₂-VQPFLNLTTTPR-COOH, SEQ ID NO: 85); D-IB1(s20) (NH₂-PVQPFLNLTTTP-COOH, SEQ ID NO: 86); D-IB1(s21) (NH₂-RPVQPFLNLTT-COOH, SEQ ID NO: 87); D-IB1(s22) (NH₂-SRPVQPFLNLT-COOH, SEQ ID NO: 88); D-IB1(s23) (NH₂-QSRPVQPFLNL-COOH, SEQ ID NO: 89); D-IB1(s24) (NH₂-DQSRPVQPFLN-COOH, SEQ ID NO: 90); D-IB1(s25) (NH₂-DQSRPVQPFL-COOH, SEQ ID NO: 91); D-IB1(s26) (NH₂-QSRPVQPFLN-COOH, SEQ ID NO: 92); D-IB1(s27) (NH₂-SRPVQPFLNL-COOH, SEQ ID NO: 93); D-IB1(s28) (NH₂-RPVQPFLNLT-COOH, SEQ ID NO: 94); D-IB1(s29) (NH₂-PVQPFLNLTT-COOH, SEQ ID NO: 95); D-IB1(s30) (NH₂-VQPFLNLTTTP-COOH, SEQ ID NO: 96); D-IB1(s31) (NH₂-QPFLNLTTTPR-COOH, SEQ ID NO: 97); D-IB1(s32) (NH₂-PFLNLTTTPRK-COOH, SEQ ID NO: 98); D-IB1(s33) (NH₂-FLNLTTTPRK-COOH, SEQ ID NO: 99); and D-IB1(s34) (NH₂-LNLTTTPRKPR-COOH, SEQ ID NO: 100).

The JNK inhibitor (poly-)peptides as used herein and as disclosed above are presented in Table 1 (SEQ ID NO:s 1-4, 13-20 and 33-100). The table presents the name of the JNK inhibitor (poly-)peptides/sequences as used herein, as well as their sequence identifier number, their length, and amino acid sequence. Furthermore, Table 1 shows sequences as well as their generic formulas, e.g. for SEQ ID NO's: 1, 2, 5, 6, 9 and 11 and SEQ ID NO's: 3, 4, 7, 8, 10 and 12, respectively. Table 1 furthermore discloses the chimeric sequences SEQ ID NOs: 9-12 and 23-32 (see below), L-IB1 sequences SEQ ID NOs: 33 to 66 and D-IB1 sequences SEQ ID NOs: 67 to 100.

TABLE 1

SEQUENCE/PEPTIDE NAME	SEQ ID NO	AA	SEQUENCE
L-IB1(s)	1	19	RPKRPTTLNLFQVPRSQD (NH ₂ -RPKRPTTLNLFQVPRSQD-COOH)
D-IB1(s)	2	19	DQSRPVQPFLNLTTTPRKPR (NH ₂ -DQSRPVQPFLNLTTTPRKPR-COOH)
L-IB (generic) (s)	3	19	NH ₂ -X _n ^b -X _n ^a -RPTTLXLXXXXXXQD-X _n ^b -COOH
D-IB (generic) (s)	4	19	NH ₂ -X _n ^b -DQXXXXXXXXLXLTTPR-X _n ^a -X _n ^b -COOH
L-TAT	5	10	GRKKRRQRRR (NH ₂ -GRKKRRQRRR-COOH)
D-TAT	6	10	RRRQRRKKRG (NH ₂ -RRRQRRKKRG-COOH)
L-generic-TAT (s)	7	11	NH ₂ -X _n ^b -RKKRRQRRR-X _n ^b -COOH

D-generic-TAT (s)	8	11	NH ₂ -X _n ^b -RRRQRRKKR-X _n ^b -COOH
L-TAT-IB1(s)	9	31	GRKKRRQRRRPPRPKRPTTLNLFQVPRSQD (NH ₂ -GRKKRRQRRRPPRPKRPTTLNLFQVPRSQD-COOH)
L-TAT-IB (generic) (s)	10	29	NH ₂ -X _n ^b -RKKRRQRRR-X _n ^b -X _n ^a -RPTTLXLXXXXXXXXXQD-X _n ^b -COOH
D-TAT-IB1(s)	11	31	DQSRPVQPFLNLTTPRKPRPPRRRQRRKKRG (NH ₂ -DQSRPVQPFLNLTTPRKPRPPRRRQRRKKRG-COOH)
D-TAT-IB (generic) (s)	12	29	NH ₂ -X _n ^b -DQXXXXXXXXLXLTPR-X _n ^a -X _n ^b -RRRQRRKKR-X _n ^b -COOH
IB1-long	13	29	PGTGCGDTYRKRPTTLNLFQVPRSQDT (NH ₂ - PGTGCGDTYRKRPTTLNLFQVPRSQDT -COOH)
IB2-long	14	27	IPSPSVEEPHKHRPTTLRLTLGAQDS (NH ₂ - IPSPSVEEPHKHRPTTLRLTLGAQDS -COOH)
c-Jun	15	29	GAYGYSNPILKQSMTLNLADPVGNLKPH (NH ₂ - GAYGYSNPILKQSMTLNLADPVGNLKPH -COOH)
ATF2	16	29	TNEDHLAVHKKHKHEMTLKFGRNDSVIV (NH ₂ - TNEDHLAVHKKHKHEMTLKFGRNDSVIV -COOH)
L-IB1	17	23	DTYRKRPTTLNLFQVPRSQDT (NH ₂ - DTYRKRPTTLNLFQVPRSQDT -COOH)
D-IB1	18	23	TDQSRPVQPFLNLTTPRKPRYTD (NH ₂ - TDQSRPVQPFLNLTTPRKPRYTD -COOH)
L-IB (generic)	19	19	XRPTTLXLXXXXXXXXQDS/TX (NH ₂ - XRPTTLXLXXXXXXXXQDS/TX -COOH)
D-IB (generic)	20	19	XS/TDQXXXXXXXXLXLTPRX (NH ₂ - XS/TDQXXXXXXXXLXLTPRX -COOH)
L-generic-TAT	21	17	XXXXRKKRRQRRRXXXX (NH ₂ - XXXXRKKRRQRRRXXXX -COOH)
D-generic-TAT	22	17	XXXXRRRQRRKKRXXXX (NH ₂ - XXXXRRRQRRKKRXXXX -COOH)
L-TAT-IB1	23	35	GRKKRRQRRRPPDTYRKRPTTLNLFQVPRSQDT (NH ₂ - GRKKRRQRRRPPDTYRKRPTTLNLFQVPRSQDT -COOH)
L-TAT-IB (generic)	24	42	XXXXXXXXRKKRRQRRRXXXXXXXXXRPTTLXLXXXXXXXXQDS/TX (NH ₂ - XXXXXXXXRKKRRQRRRXXXXXXXXXRPTTLXLXXXXXXXXQDS/TX COOH)
D-TAT-IB1	25	35	TDQSRPVQPFLNLTTPRKPRYTDPPRRRQRRKKRG (NH ₂ - TDQSRPVQPFLNLTTPRKPRYTDPPRRRQRRKKRG -COOH)
D-TAT-IB (generic)	26	42	XT/SDQXXXXXXXXLXLTPRXXXXXXXXRRRQRRKKRXXXXXXXX (NH ₂ - XT/SDQXXXXXXXXLXLTPRXXXXXXXXRRRQRRKKRXXXXXXXX COOH)
L-TAT-IB1(s1)	27	30	RKKRRQRRRPPRPKRPTTLNLFQVPRSQD (NH ₂ -RKKRRQRRRPPRPKRPTTLNLFQVPRSQD-COOH)
L-TAT-IB1(s2)	28	30	GRKKRRQRRRX _n ^c RPKRPTTLNLFQVPRSQD (NH ₂ -GRKKRRQRRRX _n ^c RPKRPTTLNLFQVPRSQD-COOH)
L-TAT-IB1(s3)	29	29	RKKRRQRRRX _n ^c RPKRPTTLNLFQVPRSQD (NH ₂ -RKKRRQRRRX _n ^c RPKRPTTLNLFQVPRSQD-COOH)
D-TAT-IB1(s1)	30	30	DQSRPVQPFLNLTTPRKPRPPRRRQRRKKR (NH ₂ -DQSRPVQPFLNLTTPRKPRPPRRRQRRKKR-COOH)
D-TAT-IB1(s2)	31	30	DQSRPVQPFLNLTTPRKPRX _n ^c RRRQRRKKRG (NH ₂ -DQSRPVQPFLNLTTPRKPRX _n ^c RRRQRRKKRG-COOH)
D-TAT-IB1(s3)	32	29	DQSRPVQPFLNLTTPRKPRX _n ^c RRRQRRKKR (NH ₂ -DQSRPVQPFLNLTTPRKPRX _n ^c RRRQRRKKR-COOH)
L-IB1(s1)	33	13	TLNLFQVPRSQD (NH ₂ -TLNLFQVPRSQD-COOH)

L-IB1(s2)	34	13	TTLNLFQVPRSQ (NH ₂ -TTLNLFQVPRSQ-COOH)
L-IB1(s3)	35	13	PTTLNLFQVPRS (NH ₂ -PTTLNLFQVPRS-COOH)
L-IB1(s4)	36	13	RPTTLNLFQVPR (NH ₂ -RPTTLNLFQVPR-COOH)
L-IB1(s5)	37	13	KRPTTLNLFQVP (NH ₂ -KRPTTLNLFQVP-COOH)
L-IB1(s6)	38	13	PKRPTTLNLFQV (NH ₂ -PKRPTTLNLFQV-COOH)
L-IB1(s7)	39	13	RPKRPTTLNLFQ (NH ₂ -RPKRPTTLNLFQ-COOH)
L-IB1(s8)	40	12	LNLFQVPRSQD (NH ₂ -LNLFQVPRSQD-COOH)
L-IB1(s9)	41	12	TLNLFQVPRSQ (NH ₂ -TLNLFQVPRSQ-COOH)
L-IB1(s10)	42	12	TTLNLFQVPRS (NH ₂ -TTLNLFQVPRS-COOH)
L-IB1(s11)	43	12	PTTLNLFQVPR (NH ₂ -PTTLNLFQVPR-COOH)
L-IB1(s12)	44	12	RPTTLNLFQVP (NH ₂ -RPTTLNLFQVP-COOH)
L-IB1(s13)	45	12	KRPTTLNLFQV (NH ₂ -KRPTTLNLFQV-COOH)
L-IB1(s14)	46	12	PKRPTTLNLFQ (NH ₂ -PKRPTTLNLFQ-COOH)
L-IB1(s15)	47	12	RPKRPTTLNLF (NH ₂ -RPKRPTTLNLF-COOH)
L-IB1(s16)	48	11	NLFQVPRSQD (NH ₂ -NLFQVPRSQD-COOH)
L-IB1(s17)	49	11	LNLFQVPRSQ (NH ₂ -LNLFQVPRSQ-COOH)
L-IB1(s18)	50	11	TLNLFQVPRS (NH ₂ -TLNLFQVPRS-COOH)
L-IB1(s19)	51	11	TTLNLFQVPR (NH ₂ -TTLNLFQVPR-COOH)
L-IB1(s20)	52	11	PTTLNLFQVP (NH ₂ -PTTLNLFQVP-COOH)
L-IB1(s21)	53	11	RPTTLNLFQV (NH ₂ -RPTTLNLFQV-COOH)
L-IB1(s22)	54	11	KRPTTLNLFQ (NH ₂ -KRPTTLNLFQ-COOH)
L-IB1(s23)	55	11	PKRPTTLNLF (NH ₂ -PKRPTTLNLF-COOH)
L-IB1(s24)	56	11	RPKRPTTLNLF (NH ₂ -RPKRPTTLNLF-COOH)
L-IB1(s25)	57	10	LFPQVPRSQD (NH ₂ -LFPQVPRSQD-COOH)
L-IB1(s26)	58	10	NLFQVPRSQ (NH ₂ -NLFQVPRSQ-COOH)
L-IB1(s27)	59	10	LNLFQVPRS (NH ₂ -LNLFQVPRS-COOH)
L-IB1(s28)	60	10	TLNLFQVPR

			(NH ₂ -TLNLFQVPR-COOH)
L-IB1(s29)	61	10	TTLNLFQVP (NH ₂ -TTLNLFQVP-COOH)
L-IB1(s30)	62	10	PTTLNLFQV (NH ₂ -PTTLNLFQV-COOH)
L-IB1(s31)	63	10	RPTTLNLFQ (NH ₂ -RPTTLNLFQ-COOH)
L-IB1(s32)	64	10	KRPTTLNLF (NH ₂ -KRPTTLNLF-COOH)
L-IB1(s33)	65	10	PKRPTTLNF (NH ₂ -PKRPTTLNF-COOH)
L-IB1(s34)	66	10	RPKRPTTLNL (NH ₂ -RPKRPTTLNL-COOH)
D-IB1(s1)	67	13	QPFLNLTPRKPR (NH ₂ -QPFLNLTPRKPR-COOH)
D-IB1(s2)	68	13	VQPFLNLTPRKP (NH ₂ -VQPFLNLTPRKP-COOH)
D-IB1(s3)	69	13	PVQPFLNLTPRK (NH ₂ -PVQPFLNLTPRK-COOH)
D-IB1(s4)	70	13	RPVQPFLNLTPR (NH ₂ -RPVQPFLNLTPR-COOH)
D-IB1(s5)	71	13	SRPVQPFLNLTP (NH ₂ -SRPVQPFLNLTP-COOH)
D-IB1(s6)	72	13	QSRPVQPFLNLTT (NH ₂ -QSRPVQPFLNLTT-COOH)
D-IB1(s7)	73	13	DQSRPVQPFLNLTT (NH ₂ -DQSRPVQPFLNLTT-COOH)
D-IB1(s8)	74	12	PFLNLTPRKPR (NH ₂ -PFLNLTPRKPR-COOH)
D-IB1(s9)	75	12	QPFLNLTPRKPR (NH ₂ -QPFLNLTPRKPR-COOH)
D-IB1(s10)	76	12	VQPFLNLTPRK (NH ₂ -VQPFLNLTPRK-COOH)
D-IB1(s11)	77	12	PVQPFLNLTPR (NH ₂ -PVQPFLNLTPR-COOH)
D-IB1(s12)	78	12	RPVQPFLNLTP (NH ₂ -RPVQPFLNLTP-COOH)
D-IB1(s13)	79	12	SRPVQPFLNLTT (NH ₂ -SRPVQPFLNLTT-COOH)
D-IB1(s14)	80	12	QSRPVQPFLNLTT (NH ₂ -QSRPVQPFLNLTT-COOH)
D-IB1(s15)	81	12	DQSRPVQPFLNL (NH ₂ -DQSRPVQPFLNL-COOH)
D-IB1(s16)	82	11	FLNLTPRKPR (NH ₂ -FLNLTPRKPR-COOH)
D-IB1(s17)	83	11	PFLNLTPRKPR (NH ₂ -PFLNLTPRKPR-COOH)
D-IB1(s18)	84	11	QPFLNLTPRK (NH ₂ -QPFLNLTPRK-COOH)
D-IB1(s19)	85	11	VQPFLNLTPR (NH ₂ -VQPFLNLTPR-COOH)
D-IB1(s20)	86	11	PVQPFLNLTP (NH ₂ -PVQPFLNLTP-COOH)

D-IB1(s21)	87	11	RPVQPFLNLT (NH ₂ -RPVQPFLNLT-COOH)
D-IB1(s22)	88	11	SRPVQPFLNLT (NH ₂ -SRPVQPFLNLT-COOH)
D-IB1(s23)	89	11	QSRPVQPFLNL (NH ₂ -QSRPVQPFLNL-COOH)
D-IB1(s24)	90	11	DQSRPVQPFLN (NH ₂ -DQSRPVQPFLN-COOH)
D-IB1(s25)	91	10	DQSRPVQPFL (NH ₂ -DQSRPVQPFL-COOH)
D-IB1(s26)	92	10	QSRPVQPFLN (NH ₂ -QSRPVQPFLN-COOH)
D-IB1(s27)	93	10	SRPVQPFLNL (NH ₂ -SRPVQPFLNL-COOH)
D-IB1(s28)	94	10	RPVQPFLNLT (NH ₂ -RPVQPFLNLT-COOH)
D-IB1(s29)	95	10	PVQPFLNLT (NH ₂ -PVQPFLNLT-COOH)
D-IB1(s30)	96	10	VQPFLNLTP (NH ₂ -VQPFLNLTP-COOH)
D-IB1(s31)	97	10	QPFLNLTPR (NH ₂ -QPFLNLTPR-COOH)
D-IB1(s32)	98	10	PFLNLTPRK (NH ₂ -PFLNLTPRK-COOH)
D-IB1(s33)	99	10	FLNLTPRK (NH ₂ -FLNLTPRK-COOH)
D-IB1(s34)	100	10	LNLTPRKPR (NH ₂ -LNLTPRKPR-COOH)

It will be understood by a person skilled in the art that a given sequence herein which is composed exclusively of D-amino acids is identified by "D-name". For example, SEQ ID NO:100 has the sequence/peptide name "D-IB1 (s34)". The given amino acid sequence is LNLTPRKPR. However, all amino acids are here D-amino acids.

It will be also understood by a person skilled in the art that the terms "entirely composed of L-amino acids"; "exclusively composed of D-amino acids" "entirely composed of D-amino acids" and/or "exclusively composed of D-amino acids" and the like refer to sequences which need not (but may) exclude the presence of glycine residues. Glycine is the only amino acid which is non-chiral. Therefore, the terms "entirely composed of L-amino acids"; "exclusively composed of D-amino acids" "entirely composed of D-amino acids" and/or "exclusively composed of D-amino acids" are intended to make clear that L-amino acids or D-amino acids, respectively, are used where possible. Nevertheless, if presence of a glycine is necessary or favored at a given position in the amino acid sequence, then it may remain there. A good example is L-TAT (SEQ ID NO:5). As used herein said sequence is considered

to be exclusively composed of L-amino acids "although" said sequence comprises a non chiral glycine residue. Likewise, D-TAT (SEQ ID NO:6), as used herein, may be considered to be exclusively composed of D-amino acids "although" said sequence comprises a non chiral glycine residue.

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According to another preferred embodiment, the JNK inhibitor (poly-)peptide as used herein comprises or consists of at least one variant, fragment and/or derivative of the above defined native or non-native amino acid sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100. Preferably, these variants, fragments and/or derivatives retain biological activity of the

10 above disclosed native or non-native JNK inhibitor (poly-)peptides as used herein, particularly of native or non-native amino acid sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100, i.e. binding JNK and/or inhibiting the activation of at least one JNK activated transcription factor, e.g. c-Jun, ATF2 or Elk1. Functionality may be tested by various tests, e.g. binding tests of the peptide to its target molecule or by biophysical

15 methods, e.g. spectroscopy, computer modeling, structural analysis, etc.. Particularly, an JNK inhibitor (poly-)peptide or variants, fragments and/or derivatives thereof as defined above may be analyzed by hydrophilicity analysis (see e.g. Hopp and Woods, 1981. Proc Natl Acad Sci USA 78: 3824-3828) that can be utilized to identify the hydrophobic and hydrophilic regions of the peptides, thus aiding in the design of substrates for experimental

20 manipulation, such as in binding experiments, or for antibody synthesis. Secondary structural analysis may also be performed to identify regions of an JNK inhibitor (poly-)peptide or of variants, fragments and/or derivatives thereof as used herein that assume specific structural motifs (see e.g. Chou and Fasman, 1974, Biochem 13: 222-223). Manipulation, translation, secondary structure prediction, hydrophilicity and

25 hydrophobicity profiles, open reading frame prediction and plotting, and determination of sequence homologies can be accomplished using computer software programs available in the art. Other methods of structural analysis include, e.g. X-ray crystallography (see e.g. Engstrom, 1974. Biochem Exp Biol 11: 7-13), mass spectroscopy and gas chromatography (see e.g. METHODS IN PROTEIN SCIENCE, 1997, J. Wiley and Sons, New York, NY) and

30 computer modeling (see e.g. Fletterick and Zoller, eds., 1986. Computer Graphics and Molecular Modeling, In: CURRENT COMMUNICATIONS IN MOLECULAR BIOLOGY, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY) may also be employed.

Accordingly, the JNK inhibitor (poly-)peptide as used herein may comprise or consist of at least one variant of (native or non-native) amino acid sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100. In the context of the present invention, a "variant of a (native or non-native) amino acid sequence according to SEQ ID NOs: 1-4, 13-20 and 33-100" is preferably a sequence derived from any of the sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100, wherein the variant comprises amino acid alterations of the amino acid sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100. Such alterations typically comprise 1 to 20, preferably 1 to 10 and more preferably 1 to 5 substitutions, additions and/or deletions of amino acids according to SEQ ID NOs: 1-4, 13-20 and 33-100, wherein the variant exhibits a sequence identity with any of the sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100 of at least about 30%, 50%, 70%, 80%, 90%, 95%, 98% or even at least about 99%.

If variants of (native or non-native) amino acid sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100 as defined above and used herein are obtained by substitution of specific amino acids, such substitutions preferably comprise conservative amino acid substitutions. Conservative amino acid substitutions may include synonymous amino acid residues within a group which have sufficiently similar physicochemical properties, so that a substitution between members of the group will preserve the biological activity of the molecule (see e.g. Grantham, R. (1974), *Science* **185**, 862-864). It is evident to the skilled person that amino acids may also be inserted and/or deleted in the above-defined sequences without altering their function, particularly if the insertions and/or deletions only involve a few amino acids, e.g. less than twenty, and preferably less than ten, and do not remove or displace amino acids which are critical to functional activity. Moreover, substitutions shall be avoided in variants as used herein, which lead to additional threonines at amino acid positions which are accessible for a phosphorylase, preferably a kinase, in order to avoid inactivation of the JNK-inhibitor (poly-)peptide as used herein or of the chimeric peptide as used herein *in vivo* or *in vitro*.

Preferably, synonymous amino acid residues, which are classified into the same groups and are typically exchangeable by conservative amino acid substitutions, are defined in Table 2.

TABLE 2

Preferred Groups of Synonymous Amino Acid Residues

	<u>Amino Acid</u>	<u>Synonymous Residue</u>
	Ser	Ser, Thr, Gly, Asn
	Arg	Arg, Gln, Lys, Glu, His
5	Leu	Ile, Phe, Tyr, Met, Val, Leu
	Pro	Gly, Ala, (Thr), Pro
	Thr	Pro, Ser, Ala, Gly, His, Gln, Thr
	Ala	Gly, Thr, Pro, Ala
	Val	Met, Tyr, Phe, Ile, Leu, Val
10	Gly	Ala, (Thr), Pro, Ser, Gly
	Ile	Met, Tyr, Phe, Val, Leu, Ile
	Phe	Trp, Met, Tyr, Ile, Val, Leu, Phe
	Tyr	Trp, Met, Phe, Ile, Val, Leu, Tyr
	Cys	Ser, Thr, Cys
15	His	Glu, Lys, Gln, Thr, Arg, His
	Gln	Glu, Lys, Asn, His, (Thr), Arg, Gln
	Asn	Gln, Asp, Ser, Asn
	Lys	Glu, Gln, His, Arg, Lys
	Asp	Glu, Asn, Asp
20	Glu	Asp, Lys, Asn, Gln, His, Arg, Glu
	Met	Phe, Ile, Val, Leu, Met
	Trp	Trp

A specific form of a variant of SEQ ID NOs: 1-4, 13-20 and 33-100 as used herein is a
 25 fragment of the (native or non-native) amino acid sequences according to SEQ ID NOs: 1,
 1-4, 13-20 and 33-100" as used herein, which is typically altered by at least one deletion as
 compared to SEQ ID NOs 1-4, 13-20 and 33-100. Preferably, a fragment comprises at least
 4 contiguous amino acids of any of SEQ ID NOs: 1-4, 13-20 and 33-100, a length typically
 sufficient to allow for specific recognition of an epitope from any of these sequences. Even
 30 more preferably, the fragment comprises 4 to 18, 4 to 15, or most preferably 4 to 10
 contiguous amino acids of any of SEQ ID NOs: 1-4, 13-20 and 33-100, wherein the lower
 limit of the range may be 4, or 5, 6, 7, 8, 9, or 10. Deleted amino acids may occur at any
 position of SEQ ID NOs: 1-4, 13-20 and 33-100, preferably N- or C-terminally.

35 Furthermore, a fragment of the (native or non-native) amino acid sequences according to
 SEQ ID NOs: 1-4, 13-20 and 33-100, as described above, may be defined as a sequence
 sharing a sequence identity with any of the sequences according to SEQ ID NOs: 1-4, 13-20
 and 33-100 as used herein of at least about 30%, 50%, 70%, 80%, 90%, 95%, 98%, or
 even 99%.

The JNK inhibitor (poly-)peptides/sequences as used herein may further comprise or consist of at least one derivative of (native or non-native) amino acid sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100 as defined above. In this context, a "derivative of an (native or non-native) amino acid sequence according to SEQ ID NOs: 1-4, 13-20 and 33-100" is preferably an amino acid sequence derived from any of the sequences according to SEQ ID NOs: 1-4, 13-20 and 33-100, wherein the derivative comprises at least one modified L- or D-amino acid (forming non-natural amino acid(s)), preferably 1 to 20, more preferably 1 to 10, and even more preferably 1 to 5 modified L- or D-amino acids. Derivatives of variants or fragments also fall under the scope of the present invention.

"A modified amino acid" in this respect may be any amino acid which is altered e.g. by different glycosylation in various organisms, by phosphorylation or by labeling specific amino acids. Such a label is then typically selected from the group of labels comprising:

- (i) radioactive labels, i.e. radioactive phosphorylation or a radioactive label with sulphur, hydrogen, carbon, nitrogen, etc.;
- (ii) colored dyes (e.g. digoxigenin, etc.);
- (iii) fluorescent groups (e.g. fluorescein, etc.);
- (iv) chemoluminescent groups;
- (v) groups for immobilization on a solid phase (e.g. His-tag, biotin, strep-tag, flag-tag, antibodies, antigen, etc.); and
- (vi) a combination of labels of two or more of the labels mentioned under (i) to (v).

In the above context, an amino acid sequence having a sequence "sharing a sequence identity" of at least, for example, 95% to a query amino acid sequence of the present invention, is intended to mean that the sequence of the subject amino acid sequence is identical to the query sequence except that the subject amino acid sequence may include up to five amino acid alterations per each 100 amino acids of the query amino acid sequence. In other words, to obtain an amino acid sequence having a sequence of at least 95% identity to a query amino acid sequence, up to 5% (5 of 100) of the amino acid residues in the subject sequence may be inserted or substituted with another amino acid or deleted.

For sequences without exact correspondence, a "% identity" of a first sequence may be determined with respect to a second sequence. In general, these two sequences to be compared are aligned to give a maximum correlation between the sequences. This may include inserting "gaps" in either one or both sequences, to enhance the degree of alignment. A % identity may then be determined over the whole length of each of the sequences being compared (so-called global alignment), that is particularly suitable for sequences of the same or similar length, or over shorter, defined lengths (so-called local alignment), that is more suitable for sequences of unequal length.

Methods for comparing the identity and homology of two or more sequences, particularly as used herein, are well known in the art. Thus for instance, programs available in the Wisconsin Sequence Analysis Package, version 9.1 (Devereux *et al.*, 1984, Nucleic Acids Res. *12*, 387-395.), for example the programs BESTFIT and GAP, may be used to determine the % identity between two polynucleotides and the % identity and the % homology between two polypeptide sequences. BESTFIT uses the "local homology" algorithm of (Smith and Waterman (1981), J. Mol. Biol. *147*, 195-197.) and finds the best single region of similarity between two sequences. Other programs for determining identity and/or similarity between sequences are also known in the art, for instance the BLAST family of programs (Altschul *et al.*, 1990, J. Mol. Biol. *215*, 403-410), accessible through the home page of the NCBI at world wide web site ncbi.nlm.nih.gov) and FASTA (Pearson (1990), Methods Enzymol. *183*, 63-98; Pearson and Lipman (1988), Proc. Natl. Acad. Sci. U. S. A *85*, 2444-2448.).

JNK-inhibitor (poly-)peptides /sequences as used according to the present invention and as defined above may be obtained or produced by methods well-known in the art, e.g. by chemical synthesis or by genetic engineering methods as discussed below. For example, a peptide corresponding to a portion of an JNK inhibitor sequence as used herein including a desired region of said JNK inhibitor sequence, or that mediates the desired activity *in vitro* or *in vivo*, may be synthesized by use of a peptide synthesizer.

JNK inhibitor (poly-)peptide as used herein and as defined above, may be furthermore be modified by a trafficking (poly-)peptide, allowing the JNK inhibitor (poly-)peptide as used

herein and as defined above to be transported effectively into the cells. Such modified JNK inhibitor (poly-)peptides are preferably provided and used as chimeric (poly-)peptides.

According to a second aspect the present invention therefore provides the use of a chimeric (poly-)peptide including at least one first domain and at least one second domain, for the preparation of a pharmaceutical composition for treating non-chronic or chronic inflammatory eye diseases in a subject, wherein the first domain of the chimeric peptide comprises a trafficking sequence, while the second domain of the chimeric (poly-)peptide comprises an JNK inhibitor sequence as defined above, preferably of any of sequences according to SEQ ID NO: 1-4, 13-20 and 33-100 or a derivative or a fragment thereof.

Typically, chimeric (poly-)peptides as used according to the present invention have a length of at least 25 amino acid residues, e.g. 25 to 250 amino acid residues, more preferably 25 to 200 amino acid residues, even more preferably 25 to 150 amino acid residues, 25 to 100 and most preferably amino acid 25 to 50 amino acid residues.

As a first domain the chimeric (poly-)peptide as used herein preferably comprises a trafficking sequence, which is typically selected from any sequence of amino acids that directs a peptide (in which it is present) to a desired cellular destination. Thus, the trafficking sequence, as used herein, typically directs the peptide across the plasma membrane, e.g. from outside the cell, through the plasma membrane, and into the cytoplasm. Alternatively, or in addition, the trafficking sequence may direct the peptide to a desired location within the cell, e.g. the nucleus, the ribosome, the endoplasmic reticulum (ER), a lysosome, or peroxisome, by e.g. combining two components (e.g. a component for cell permeability and a component for nuclear location) or by one single component having e.g. properties of cell membrane transport and targeted e.g. intranuclear transport. The trafficking sequence may additionally comprise another component, which is capable of binding a cytoplasmic component or any other component or compartment of the cell (e.g. endoplasmic reticulum, mitochondria, Golgi apparatus, lysosomal vesicles). Accordingly, e.g. the trafficking sequence of the first domain and the JNK inhibitor sequence of the second domain may be localized in the cytoplasm or any other compartment of the cell. This allows to determine localization of the chimeric peptide in the cell upon uptake.

Preferably, the trafficking sequence (being included in the first domain of the chimeric peptide as used herein) has a length of 5 to 150 amino acid sequences, more preferably a length of 5 to 100 and most preferably a length of from 5 to 50, 5 to 30 or even 5 to 15 amino acids.

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More preferably, the trafficking sequence (contained in the first domain of the chimeric peptide as used herein) may occur as a continuous amino acid sequence stretch in the first domain. Alternatively, the trafficking sequence in the first domain may be splitted into two or more fragments, wherein all of these fragments resemble the entire trafficking sequence and may be separated from each other by 1 to 10, preferably 1 to 5 amino acids, provided that the trafficking sequence as such retains its carrier properties as disclosed above. These amino acids separating the fragments of the trafficking sequence may e.g. be selected from amino acid sequences differing from the trafficking sequence. Alternatively, the first domain may contain a trafficking sequence composed of more than one component, each component with its own function for the transport of the cargo JNK inhibitor sequence of the second domain to e.g. a specific cell compartment.

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The trafficking sequence as defined above may be composed of L-amino acids, D-amino acids, or a combination of both. Preferably, the trafficking sequences (being included in the first domain of the chimeric peptide as used herein) may comprise at least 1 or even 2, preferably at least 3, 4 or 5, more preferably at least 6, 7, 8 or 9 and even more preferably at least 10 or more D- and/or L-amino acids, wherein the D- and/or L-amino acids may be arranged in the JNK trafficking sequences in a blockwise, a non-blockwise or in an alternate manner.

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According to one alternative embodiment, the trafficking sequence of the chimeric (poly-)peptide as used herein may be exclusively composed of L-amino acids. More preferably, the trafficking sequence of the chimeric peptide as used herein comprises or consists of at least one „native“ trafficking sequence as defined above. In this context, the term “native” is referred to non-altered trafficking sequences, entirely composed of L-amino acids.

30

According to another alternative embodiment the trafficking sequence of the chimeric (poly-)peptide as used herein may be exclusively composed of D-amino acids. More preferably,

the trafficking sequence of the chimeric peptide as used herein may comprise a D retro-inverso peptide of the sequences as presented above.

The trafficking sequence of the first domain of the chimeric (poly-)peptide as used herein
5 may be obtained from naturally occurring sources or can be produced by using genetic engineering techniques or chemical synthesis (see e.g. Sambrook, J., Fritsch, E. F., Maniatis, T. (1989) Molecular cloning: A laboratory manual. 2nd edition. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y.).

10 Sources for the trafficking sequence of the first domain may be employed including, e.g. native proteins such as e.g. the TAT protein (e.g. as described in U.S. Patent Nos. 5,804,604 and 5,674,980, each of these references being incorporated herein by reference), VP22 (described in e.g. WO 97/05265; Elliott and O'Hare, Cell 88 : 223-233 (1997)), non-viral proteins (Jackson et al, Proc. Natl. Acad. Sci. USA 89 : 10691-10695 (1992)), trafficking
15 sequences derived from Antennapedia (e.g. the antennapedia carrier sequence) or from basic peptides, e.g. peptides having a length of 5 to 15 amino acids, preferably 10 to 12 amino acids and comprising at least 80 %, more preferably 85 % or even 90 % basic amino acids, such as e.g. arginine, lysine and/or histidine. Furthermore, variants, fragments and derivatives of one of the native proteins used as trafficking sequences are disclosed
20 herewith. With regard to variants, fragments and derivatives it is referred to the definition given above for JNK inhibitor sequences as used herein. Variants, fragments as well as derivatives are correspondingly defined as set forth above for JNK inhibitor sequences as used herein. Particularly, in the context of the trafficking sequence, a variant or fragment or derivative may be defined as a sequence sharing a sequence identity with one of the native
25 proteins used as trafficking sequences as defined above of at least about 30%, 50%, 70%, 80%, 90%, 95%, 98%, or even 99%.

In a preferred embodiment of the chimeric (poly-)peptide as used herein, the trafficking sequence of the first domain comprises or consists of a sequence derived from the human
30 immunodeficiency virus (HIV)1 TAT protein, particularly some or all of the 86 amino acids that make up the TAT protein.

For a trafficking sequence (being included in the first domain of the chimeric peptide as used herein), partial sequences of the full-length TAT protein may be used forming a functionally effective fragment of a TAT protein, i.e. a TAT peptide that includes the region that mediates entry and uptake into cells. As to whether such a sequence is a functionally effective fragment of the TAT protein can be determined using known techniques (see e.g. Franked *et al.*, Proc. Natl. Acad. Sci, USA 86 : 7397-7401 (1989)). Thus, the trafficking sequence in the first domain of the chimeric peptide as used herein may be derived from a functionally effective fragment or portion of a TAT protein sequence that comprises less than 86 amino acids, and which exhibits uptake into cells, and optionally the uptake into the cell nucleus. More preferably, partial sequences (fragments) of TAT to be used as carrier to mediate permeation of the chimeric peptide across the cell membrane, are intended to comprise the basic region (amino acids 48 to 57 or 49 to 57) of full-length TAT.

According to a more preferred embodiment, the trafficking sequence (being included in the first domain of the chimeric peptide as used herein) may comprise or consist of an amino acid sequence containing TAT residues 48-57 or 49 to 57, and most preferably a generic TAT sequence $\text{NH}_2\text{-X}_n^b\text{-RKKRRQRRR-X}_n^b\text{-COOH}$ (L-generic-TAT (s)) [SEQ ID NO: 7] and/or XXXXRKKRRQ RRRXXXX (L-generic-TAT) [SEQ ID NO: 21], wherein X or X_n^b is as defined above. Furthermore, the number of " X_n^b " residues in SEQ ID NOs :8 is not limited to the one depicted, and may vary as described above. Alternatively, the trafficking sequence being included in the first domain of the chimeric peptide as used herein may comprise or consist of a peptide containing e.g. the amino acid sequence $\text{NH}_2\text{-GRKKRRQRRR-COOH}$ (L-TAT) [SEQ ID NO: 5].

According to another more preferred embodiment the trafficking sequence (being included in the first domain of the chimeric peptide as used herein) may comprise a D retro-inverso peptide of the sequences as presented above, i.e. the D retro-inverso sequence of the generic TAT sequence having the sequence $\text{NH}_2\text{-X}_n^b\text{-RRRQRRKKR-X}_n^b\text{-COOH}$ (D-generic-TAT (s)) [SEQ ID NO : 8] and/or XXXXRRRQRRKKRXXXX (D-generic-TAT) [SEQ ID NO: 22]. Also here, X_n^b is as defined above (preferably representing D amino acids). Furthermore, the number of " X_n^b " residues in SEQ ID NOs :8 is not limited to the one depicted, and may vary as described above. Most preferably, the trafficking sequence as

used herein may comprise the D retro-inverso sequence NH₂-RRRQRRKKRG-COOH (D-TAT) [SEQ ID NO: 6].

According to another embodiment the trafficking sequence being included in the first domain of the chimeric peptide as used herein may comprise or consist of variants of the trafficking sequences as defined above. A "variant of a trafficking sequence" is preferably a sequence derived from a trafficking sequence as defined above, wherein the variant comprises a modification, for example, addition, (internal) deletion (leading to fragments) and/or substitution of at least one amino acid present in the trafficking sequence as defined above. Such (a) modification(s) typically comprise(s) 1 to 20, preferably 1 to 10 and more preferably 1 to 5 substitutions, additions and/or deletions of amino acids. Furthermore, the variant preferably exhibits a sequence identity with the trafficking sequence as defined above, more preferably with any of SEQ ID NOs: 5 to 8 or 21-22, of at least about 30%, 50%, 70%, 80%, 90%, 95%, 98% or even 99%.

Preferably, such a modification of the trafficking sequence being included in the first domain of the chimeric peptide as used herein leads to a trafficking sequence with increased or decreased stability. Alternatively, variants of the trafficking sequence can be designed to modulate intracellular localization of the chimeric peptide as used herein. When added exogenously, such variants as defined above are typically designed such that the ability of the trafficking sequence to enter cells is retained (i.e. the uptake of the variant of the trafficking sequence into the cell is substantially similar to that of the native protein used a trafficking sequence). For example, alteration of the basic region thought to be important for nuclear localization (see e.g. Dang and Lee, J. Biol. Chem. 264 : 18019-18023 (1989); Hauber *et al.*, J. Virol. 63 : 1181-1187 (1989) ; *et al.*, J. Virol. 63 : 1-8 (1989)) can result in a cytoplasmic location or partially cytoplasmic location of the trafficking sequence, and therefore, of the JNK inhibitor sequence as component of the chimeric peptide as used herein. Additional to the above, further modifications may be introduced into the variant, e.g. by linking e.g. cholesterol or other lipid moieties to the trafficking sequence to produce a trafficking sequence having increased membrane solubility. Any of the above disclosed variants of the trafficking sequences being included in the first domain of the chimeric peptide as used herein can be produced using techniques typically known to a skilled

person (see e.g. Sambrook, J., Fritsch, E. F., Maniatis, T. (1989) Molecular cloning: A laboratory manual. 2nd edition. Cold Spring Harbor Laboratory, Cold Spring Harbor, N.Y.)

5 As a second domain the chimeric peptide as used herein typically comprises an JNK inhibitor sequence, selected from any of the JNK inhibitor sequences as defined above, including variants, fragments and/or derivatives of these JNK inhibitor sequences.

10 Both domains, i.e. the first and the second domain(s), of the chimeric peptide as used herein, may be linked such as to form a functional unit. Any method for linking the first and second domain(s) as generally known in the art may be applied.

15 According to one embodiment, the first and the second domain(s) of the chimeric peptide as used herein are preferably linked by a covalent bond. A covalent bond, as defined herein, may be e.g. a peptide bond, which may be obtained by expressing the chimeric peptide as defined above as a fusion protein. Fusion proteins, as described herein, can be formed and used in ways analogous to or readily adaptable from standard recombinant DNA techniques, as described below. However, both domains may also be linked via side chains or may be linked by a chemical linker moiety.

20 The first and/or second domains of the chimeric peptide as used herein may occur in one or more copies in said chimeric peptide. If both domains are present in a single copy, the first domain may be linked either to the N-terminal or the C-terminal end of the second domain. If present in multiple copies, the first and second domain(s) may be arranged in any possible order. E.g. the first domain can be present in the chimeric peptide as used herein in a multiple copy number, e.g. in two, three or more copies, which are preferably arranged in consecutive order. Then, the second domain may be present in a single copy occurring at the N- or C-terminus of the sequence comprising the first domain. Alternatively, the second domain may be present in a multiple copy number, e.g. in two, three or more copies, and the first domain may be present in a single copy. According to both alternatives, first and second domain(s) can take any place in a consecutive arrangement. Exemplary arrangements are shown in the following: e.g. first domain – first domain – first domain – second domain; first domain – first domain – second domain – first domain; first domain – second domain – first domain – first domain; or e.g. second domain – first domain – first domain – first domain – second domain.

domain – first domain. It is well understood for a skilled person that these examples are for illustration purposes only and shall not limit the scope of the invention thereto. Thus, the number of copies and the arrangement may be varied as defined initially.

- 5 Preferably, the first and second domain(s) may be directly linked with each other without any linker. Alternatively, they may be linked with each other via a linker sequence comprising 1 to 10, preferably 1 to 5 amino acids. Amino acids forming the linker sequence are preferably selected from glycine or proline as amino acid residues. More preferably, the first and second domain(s) may be separated by each other by a hinge of
10 two, three or more proline residues between the first and second domain(s).

The chimeric peptide as defined above and as used herein, comprising at least one first and at least one second domain, may be composed of L-amino acids, D-amino acids, or a combination of both. Therein, each domain (as well as the linkers used) may be composed
15 of L-amino acids, D-amino acids, or a combination of both (e.g. D-TAT and L-IB1(s) or L-TAT and D-IB1(s), etc.). Preferably, the chimeric peptide as used herein may comprise at least 1 or even 2, preferably at least 3, 4 or 5, more preferably at least 6, 7, 8 or 9 and even more preferably at least 10 or more D- and/or L-amino acids, wherein the D- and/or L-amino acids may be arranged in the chimeric peptide as used herein in a blockwise, a non-
20 blockwise or in an alternate manner.

According to a specific embodiment the chimeric peptide as used herein comprises or consists of the L-amino acid chimeric peptides according to the generic L-TAT-IB peptide $\text{NH}_2\text{-X}_n^b\text{-RKKRRQRRR-X}_n^b\text{-X}_n^a\text{-RPTTLXLXXXXXXXQD-X}_n^b\text{-COOH}$ (L-TAT-IB (generic) (s))
25 [SEQ ID NO: 10], wherein X, X_n^a and X_n^b are preferably as defined above. More preferably, the chimeric peptide as used herein comprises or consists of the L-amino acid chimeric peptide $\text{NH}_2\text{-GRKKRRQRRRPPRPKRPTTLNLFQVPRSQD-COOH}$ (L-TAT-IB1 (s)) [SEQ ID NO: 9]. Alternatively or additionally, the chimeric peptide as used herein comprises or consists of the L-amino acid chimeric peptide sequence GRKKRRQRRR PPDTYRPKRP
30 TTLNLFQVP RSQDT (L-TAT-IB1) [SEQ ID NO: 23], or XXXXXXXXRRK RRQRRRXXXX XXXXRPTTLX LXXXXXXXQD S/TX (L-TAT-IB generic) [SEQ ID NO: 24], wherein X is preferably also as defined above, or the chimeric peptide as used herein comprises or consists of the L-amino acid chimeric peptide sequence

RKKRRQRRRPPRKRPTTLNLFQVPRSQD (L-TAT-IB1(s1)) [SEQ ID NO: 27],
 GRKKRRQRRRX_n^cRPKRPTTLNLFQVPRSQD (L-TAT-IB1(s2)) [SEQ ID NO: 28], or
 RKKRRQRRRX_n^cRPKRPTTLNLFQVPRSQD (L-TAT-IB1(s3)) [SEQ ID NO: 29]. In this
 context, each X typically represents an amino acid residue as defined above, more
 preferably X_n^c represents a contiguous stretch of peptide residues, each X independently
 selected from each other from glycine or proline, e.g. a monotonic glycine stretch or a
 monotonic proline stretch, wherein n (the number of repetitions of X_n^c) is typically 0-5, 5-
 10, 10-15, 15-20, 20-30 or even more, preferably 0-5 or 5-10. X_n^c may represent either D
 or L amino acids.

According to an alternative specific embodiment the chimeric peptide as used herein
 comprises or consists of D-amino acid chimeric peptides of the above disclosed L-amino
 acid chimeric peptides. Exemplary D retro-inverso chimeric peptides according to the
 present invention are e.g. the generic D-TAT-IB peptide NH₂-X_n^b-DQXXXXXXXXLXLTPR-X_n^a-
 X_n^b-RRRQRRKKR-X_n^b-COOH (D-TAT-IB (generic) (s)) [SEQ ID NO: 12]. Herein, X, X_n^a and
 X_n^b are preferably as defined above (preferably representing D amino acids). More
 preferably, the chimeric peptide as used herein comprises or consists of D-amino acid
 chimeric peptides according to the TAT-IB1 peptide NH₂-
 DQSRPVQPFLNLTPRKPRPPRRRQRRKKRG-COOH (D-TAT-IB1(s)) [SEQ ID NO: 11].
 Alternatively or additionally, the chimeric peptide as used herein comprises or consists of
 the D-amino acid chimeric peptide sequence
 TDQSRPVQPFLNLTPRKPRYTDPPIRRRQRRKKRG (D-TAT-IB1) [SEQ ID NO: 25], or
 XT/SDQXXXXXXXXLXLTPRXXXXXXXXRRRQRRKKRXXXXXXXX (D-TAT-IB generic) [SEQ ID
 NO: 26], wherein X is preferably also as defined above, or the chimeric peptide as used
 herein comprises or consists of the D-amino acid chimeric peptide sequence
 DQSRPVQPFLNLTPRKPRPPRRRQRRKKR (D-TAT-IB1(s1)) [SEQ ID NO: 30],
 DQSRPVQPFLNLTPRKPRX_n^cRRRQRRKKRG (D-TAT-IB1(s2)) [SEQ ID NO: 31], or
 DQSRPVQPFLNLTPRKPRX_n^cRRRQRRKKR (D-TAT-IB1(s3)) [SEQ ID NO: 32]. X_n^c may be
 as defined above.

The first and second domain(s) of the chimeric peptide as defined above may be linked to
 each other by chemical or biochemical coupling carried out in any suitable manner known
 in the art, e.g. by establishing a peptide bond between the first and the second domain(s)

e.g. by expressing the first and second domain(s) as a fusion protein, or e.g. by crosslinking the first and second domain(s) of the chimeric peptide as defined above.

Many known methods suitable for chemical crosslinking of the first and second domain(s) of the chimeric peptide as defined above are non-specific, i.e. they do not direct the point of coupling to any particular site on the transport polypeptide or cargo macromolecule. As a result, use of non-specific crosslinking agents may attack functional sites or sterically block active sites, rendering the conjugated proteins biologically inactive. Thus, preferably such crosslinking methods are used, which allow a more specific coupling of the first and second domain(s).

In this context, one way to increasing coupling specificity is a direct chemical coupling to a functional group present only once or a few times in one or both of the first and second domain(s) to be crosslinked. For example, cysteine, which is the only protein amino acid containing a thiol group, occurs in many proteins only a few times. Also, for example, if a polypeptide contains no lysine residues, a crosslinking reagent specific for primary amines will be selective for the amino terminus of that polypeptide. Successful utilization of this approach to increase coupling specificity requires that the polypeptide have the suitably rare and reactive residues in areas of the molecule that may be altered without loss of the molecule's biological activity. Cysteine residues may be replaced when they occur in parts of a polypeptide sequence where their participation in a crosslinking reaction would otherwise likely interfere with biological activity. When a cysteine residue is replaced, it is typically desirable to minimize resulting changes in polypeptide folding. Changes in polypeptide folding are minimized when the replacement is chemically and sterically similar to cysteine. For these reasons, serine is preferred as a replacement for cysteine. As demonstrated in the examples below, a cysteine residue may be introduced into a polypeptide's amino acid sequence for crosslinking purposes. When a cysteine residue is introduced, introduction at or near the amino or carboxy terminus is preferred. Conventional methods are available for such amino acid sequence modifications, wherein the polypeptide of interest is produced by chemical synthesis or via expression of recombinant DNA.

Coupling of the first and second domain(s) of the chimeric peptide as defined above and used herein can also be accomplished via a coupling or conjugating agent. There are several intermolecular crosslinking reagents which can be utilized (see for example, Means and Feeney, CHEMICAL MODIFICATION OF PROTEINS, Holden-Day, 1974, pp. 39-43).

5 Among these reagents are, for example, N-succinimidyl 3-(2-pyridyldithio) propionate (SPDP) or N,N'-(1,3-phenylene) bismaleimide (both of which are highly specific for sulfhydryl groups and form irreversible linkages); N, N'-ethylene-bis-(iodoacetamide) or other such reagent having 6 to 11 carbon methylene bridges (which are relatively specific for sulfhydryl groups); and 1,5-difluoro-2,4-dinitrobenzene (which forms irreversible
10 linkages with amino and tyrosine groups). Other crosslinking reagents useful for this purpose include: p,p'-difluoro-m, m'-dinitrodiphenylsulfone which forms irreversible crosslinkages with amino and phenolic groups); dimethyl adipimidate (which is specific for amino groups); phenol-1,4 disulfonylchloride (which reacts principally with amino groups); hexamethylenediisocyanate or diisothiocyanate, or azophenyl-p-diisocyanate (which reacts
15 principally with amino groups); glutaraldehyde (which reacts with several different side chains) and disdiazobenzidine (which reacts primarily with tyrosine and histidine).

Crosslinking reagents used for crosslinking the first and second domain(s) of the chimeric peptide as defined above may be homobifunctional, i.e. having two functional groups that
20 undergo the same reaction. A preferred homobifunctional crosslinking reagent is bismaleimidohexane ("BMH"). BMH contains two maleimide functional groups, which react specifically with sulfhydryl-containing compounds under mild conditions (pH 6.5-7.7). The two maleimide groups are connected by a hydrocarbon chain. Therefore, BMH is useful for irreversible crosslinking of polypeptides that contain cysteine residues.

25

Crosslinking reagents used for crosslinking the first and second domain(s) of the chimeric peptide as defined above may also be heterobifunctional. Heterobifunctional crosslinking agents have two different functional groups, for example an amine-reactive group and a thiol-reactive group, that will crosslink two proteins having free amines and thiols,
30 respectively. Examples of heterobifunctional crosslinking agents are succinimidyl 4-(N-maleimidomethyl)cyclohexane-1-carboxylate ("SMCC"), m-maleimidobenzoyl-N-hydroxysuccinimide ester ("MBS"), and succinimide 4-(p-maleimidophenyl)butyrate ("SMPB"), an extended chain analog of MBS. The succinimidyl group of these crosslinkers

reacts with a primary amine, and the thiol-reactive maleimide forms a covalent bond with the thiol of a cysteine residue.

Crosslinking reagents suitable for crosslinking the first and second domain(s) of the chimeric peptide as defined above often have low solubility in water. A hydrophilic moiety, such as a sulfonate group, may thus be added to the crosslinking reagent to improve its water solubility. In this respect, Sulfo-MBS and Sulfo-SMCC are examples of crosslinking reagents modified for water solubility, which may be used according to the present invention.

Likewise, many crosslinking reagents yield a conjugate that is essentially non-cleavable under cellular conditions. However, some crosslinking reagents particularly suitable for crosslinking the first and second domain(s) of the chimeric peptide as defined above contain a covalent bond, such as a disulfide, that is cleavable under cellular conditions. For example, Traut's reagent, dithiobis(succinimidylpropionate) ("DSP"), and N-succinimidyl 3-(2-pyridyldithio)propionate ("SPDP") are well-known cleavable crosslinkers. The use of a cleavable crosslinking reagent permits the cargo moiety to separate from the transport polypeptide after delivery into the target cell. Direct disulfide linkage may also be useful.

Numerous crosslinking reagents, including the ones discussed above, are commercially available. Detailed instructions for their use are readily available from the commercial suppliers. A general reference on protein crosslinking and conjugate preparation is: Wong, CHEMISTRY OF PROTEIN CONJUGATION AND CROSSLINKING, CRC Press (1991).

Chemical crosslinking of the first and second domain(s) of the chimeric peptide as defined above may include the use of spacer arms. Spacer arms provide intramolecular flexibility or adjust intramolecular distances between conjugated moieties and thereby may help preserve biological activity. A spacer arm may be in the form of a polypeptide moiety that includes spacer amino acids, e.g. proline. Alternatively, a spacer arm may be part of the crosslinking reagent, such as in "long-chain SPDP" (Pierce Chem. Co., Rockford, IL., cat. No. 21651 H).

Furthermore, variants, fragments or derivatives of one of the above disclosed chimeric peptides may be used herein. With regard to fragments and variants it is generally referred to the definition given above for JNK inhibitor sequences.

- 5 Particularly, in the context of the present invention, a “variant of a chimeric peptide” is preferably a sequence derived from any of the sequences according to SEQ ID NOs: 9 to 12 and 23 to 32, wherein the chimeric variant comprises amino acid alterations of the chimeric peptides according to SEQ ID NOs: 9 to 12 and 23 to 32 as used herein. Such alterations typically comprise 1 to 20, preferably 1 to 10 and more preferably 1 to 5 substitutions,
10 additions and/or deletions (leading to fragments) of amino acids according to SEQ ID NOs: 9 to 12 and 23 to 32, wherein the altered chimeric peptide as used herein exhibits a sequence identity with any of the sequences according to SEQ ID NOs: 9-12 and 23 to 32 of at least about 30%, 50%, 70%, 80%, or 95%, 98%, or even 99%. Preferably, these variants retain the biological activity of the first and the second domain as contained in the
15 chimeric peptide as used herein, i.e. the trafficking activity of the first domain as disclosed above and the activity of the second domain for binding JNK and/or inhibiting the activation of at least one JNK activated transcription factor.

- Accordingly, the chimeric peptide as used herein also comprises fragments of the afore
20 disclosed chimeric peptides, particularly of the chimeric peptide sequences according to any of SEQ ID NOs: 9 to 12 and 23 to 32. Thus, in the context of the present invention, a “fragment of the chimeric peptide” is preferably a sequence derived any of the sequences according to SEQ ID NOs: 9 to 12 and 23 to 32, wherein the fragment comprises at least 4 contiguous amino acids of any of SEQ ID NOs: 9 to 12 and 23 to 32. This fragment
25 preferably comprises a length which is sufficient to allow specific recognition of an epitope from any of these sequences and to transport the sequence into the cells, the nucleus or a further preferred location. Even more preferably, the fragment comprises 4 to 18, 4 to 15, or most preferably 4 to 10 contiguous amino acids of any of SEQ ID NOs: 9 to 12 and 23 to 32. Fragments of the chimeric peptide as used herein further may be defined as a sequence
30 sharing a sequence identity with any of the sequences according to any of SEQ ID NOs: 99 to 12 and 23 to 32 of at least about 30%, 50%, 70%, 80%, or 95%, 98%, or even 99%.

Finally, the chimeric peptide as used herein also comprises derivatives of the afore disclosed chimeric peptides, particularly of the chimeric peptide sequences according to any of SEQ ID NOs: 9 to 12 and 23 to 32.

- 5 A particularly preferred use of the present invention is the use of a JNK inhibitor (poly-)peptide consisting of or comprising the amino acid sequence of SEQ ID NO: 11, or consisting of or comprising an amino acid sequence sharing a sequence identity of at least about 30%, 50%, 70%, 80%, 90%, 92% or even 95% with SEQ ID NO: 11, for the treatment of inflammatory eye diseases, in particular for the treatment of uveitis, for
10 example for the treatment of anterior uveitis, intermediate uveitis, posterior uveitis or panuveitis. The JNK inhibitor (poly-)peptide consisting of or comprising the amino acid sequence of SEQ ID NO: 11, or consisting of or comprising an amino acid sequence sharing a sequence identity of at least about 30%, 50%, 70%, 80%, 90%, 92% or even 95% with SEQ ID NO: 11 may be administered for example locally to the eye or systemically.
15 However, the present application also clearly contemplates the use of other JNK inhibitor chimeric (poly-)peptides, i.e. where the JNK inhibitor poly-)peptide used does not consist of or comprise the amino acid sequence of SEQ ID NO: 11 for the treatment of inflammatory eye diseases, in particular for the treatment of uveitis, for example for the treatment of anterior uveitis, intermediate uveitis, posterior uveitis or panuveitis.

20

- Furthermore, the inventors also clearly contemplate the use of the JNK inhibitor (poly-)peptides of the present invention, in particular where the JNK inhibitor poly-)peptide used consists of or comprises the amino acid sequence of SEQ ID NO: 11 or consists of or comprises an amino acid sequence sharing a sequence identity of at least about 30%, 50%,
25 70%, 80%, 90%, 92% or even 95% with SEQ ID NO: 11, for the treatment of inflammatory eye diseases other than inflammation of the uvea and/or retina, e.g. for the treatment of inflammatory eye diseases which are not uveitis and/or retinitis. Moreover, it must be noted that the present invention does in particular not contemplate the treatment of (non-inflammatory) retinopathy.

30

The present invention additionally refers to the use of nucleic acid sequences encoding JNK inhibitor sequences as defined above, chimeric peptides or their fragments, variants or derivatives, all as defined above, for the preparation of a pharmaceutical composition for

treating non-chronic or chronic inflammatory eye diseases in a subject as defined herein. A preferable suitable nucleic acid encoding an JNK inhibitor sequence as used herein is typically chosen from human IB1 nucleic acid (GenBank Accession No. (AF074091), rat IB1 nucleic acid (GenBank Accession No. AF 108959), or human IB2 (GenBank Accession No AF218778) or from any nucleic acid sequence encoding any of the sequences as defined above, i.e. any sequence according to SEQ ID NO: 1-26.

Nucleic acids encoding the JNK inhibitor sequences as used herein or chimeric peptides as used herein may be obtained by any method known in the art (e.g. by PCR amplification using synthetic primers hybridizable to the 3'- and 5'-termini of the sequence and/or by cloning from a cDNA or genomic library using an oligonucleotide sequence specific for the given gene sequence).

Additionally, nucleic acid sequences are disclosed herein as well, which hybridize under stringent conditions with the appropriate strand coding for a (native) JNK inhibitor sequence or chimeric peptide as defined above. Preferably, such nucleic acid sequences comprise at least 6 (contiguous) nucleic acids, which have a length sufficient to allow for specific hybridization. More preferably, such nucleic acid sequences comprise 6 to 38, even more preferably 6 to 30, and most preferably 6 to 20 or 6 to 10 (contiguous) nucleic acids.

"Stringent conditions" are sequence dependent and will be different under different circumstances. Generally, stringent conditions can be selected to be about 5°C lower than the thermal melting point (T_M) for the specific sequence at a defined ionic strength and pH. The T_M is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. Typically, stringent conditions will be those in which the salt concentration is at least about 0.02 molar at pH 7 and the temperature is at least about 60°C. As other factors may affect the stringency of hybridization (including, among others, base composition and size of the complementary strands), the presence of organic solvents and the extent of base mismatching, the combination of parameters is more important than the absolute measure of any one.

"High stringency conditions" may comprise the following, e.g. Step 1: Filters containing DNA are pretreated for 8 hours to overnight at 65°C in buffer composed of 6*SSC, 50 mM

Tris-HCl (pH 7.5), 1 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.02% BSA, and 500 µg/ml denatured salmon sperm DNA. Step 2: Filters are hybridized for 48 hours at 65°C. in the above prehybridization mixture to which is added 100 mg/ml denatured salmon sperm DNA and 5-20*10⁶ cpm of ³²P-labeled probe. Step 3: Filters are washed for 1 hour at 37°C
5 in a solution containing 2*SSC, 0.01% PVP, 0.01% Ficoll, and 0.01% BSA. This is followed by a wash in 0.1*SSC at 50°C for 45 minutes. Step 4: Filters are autoradiographed. Other conditions of high stringency that may be used are well known in the art (see e.g. Ausubel *et al.*, (eds.), 1993, Current Protocols in Molecular Biology, John Wiley and Sons, NY; and Kriegler, 1990, Gene Transfer and Expression, a Laboratory Manual, Stockton Press, NY).

10

“Moderate stringency conditions” can include the following: Step 1: Filters containing DNA are pretreated for 6 hours at 55°C. in a solution containing 6*SSC, 5*Denhardt's solution, 0.5% SDS and 100 mg/ml denatured salmon sperm DNA. Step 2: Filters are hybridized for 18-20 hours at 55°C in the same solution with 5-20*10⁶ cpm ³²P-labeled probe added. Step
15 3: Filters are washed at 37°C for 1 hour in a solution containing 2*SSC, 0.1% SDS, then washed twice for 30 minutes at 60°C in a solution containing 1*SSC and 0.1% SDS. Step 4: Filters are blotted dry and exposed for autoradiography. Other conditions of moderate stringency that may be used are well-known in the art (see e.g. Ausubel *et al.*, (eds.), 1993, Current Protocols in Molecular Biology, John Wiley and Sons, NY; and Kriegler, 1990,
20 Gene Transfer and Expression, a Laboratory Manual, Stockton Press, NY).

25

Finally, “low stringency conditions” can include: Step 1: Filters containing DNA are pretreated for 6 hours at 40°C in a solution containing 35% formamide, 5X SSC, 50 mM Tris-HCl (pH 7.5), 5 mM EDTA, 0.1% PVP, 0.1% Ficoll, 1% BSA, and 500 µg/ml denatured
25 salmon sperm DNA. Step 2: Filters are hybridized for 18-20 hours at 40°C in the same solution with the addition of 0.02% PVP, 0.02% Ficoll, 0.2% BSA, 100 µg/ml salmon sperm DNA, 10% (wt/vol) dextran sulfate, and 5-20 x 10⁶ cpm ³²P-labeled probe. Step 3: Filters are washed for 1.5 hours at 55 C in a solution containing 2X SSC, 25 mM Tris-HCl (pH 7.4), 5 mM EDTA, and 0.1% SDS. The wash solution is replaced with fresh solution and
30 incubated an additional 1.5 hours at 60°C. Step 4: Filters are blotted dry and exposed for autoradiography. If necessary, filters are washed for a third time at 65-68°C and reexposed to film. Other conditions of low stringency that may be used are well known in the art (e.g. as employed for cross-species hybridizations). See e.g. Ausubel *et al.*, (eds.), 1993,

CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley and Sons, NY; and Kriegler, 1990, GENE TRANSFER AND EXPRESSION, A LABORATORY MANUAL, Stockton Press, NY.

- 5 The nucleic acid sequences as defined above according to the present invention can be used to express peptides, i.e. an JNK inhibitor sequence as used herein or an chimeric peptide as used herein for analysis, characterization or therapeutic use; as markers for tissues in which the corresponding peptides (as used herein) are preferentially expressed (either constitutively or at a particular stage of tissue differentiation or development or in
10 disease states). Other uses for these nucleic acids include, e.g. molecular weight markers in gel electrophoresis-based analysis of nucleic acids.

According to a further embodiment of the present invention, expression vectors may be used for the above purposes for recombinant expression of one or more JNK inhibitor
15 sequences and/or chimeric peptides as defined above. The term "expression vector" is used herein to designate either circular or linear DNA or RNA, which is either double-stranded or single-stranded. It further comprises at least one nucleic acid as defined above to be transferred into a host cell or into a unicellular or multicellular host organism. The expression vector as used herein preferably comprises a nucleic acid as defined above
20 encoding the JNK inhibitor sequence as used herein or a fragment or a variant thereof, or the chimeric peptide as used herein, or a fragment or a variant thereof. Additionally, an expression vector according to the present invention preferably comprises appropriate elements for supporting expression including various regulatory elements, such as enhancers/promoters from viral, bacterial, plant, mammalian, and other eukaryotic sources
25 that drive expression of the inserted polynucleotide in host cells, such as insulators, boundary elements, LCRs (e.g. described by Blackwood and Kadonaga (1998), *Science* 281, 61-63) or matrix/scaffold attachment regions (e.g. described by Li, Harju and Peterson, (1999), *Trends Genet.* 15, 403-408). In some embodiments, the regulatory elements are heterologous (i.e. not the native gene promoter). Alternately, the necessary transcriptional
30 and translational signals may also be supplied by the native promoter for the genes and/or their flanking regions.

The term "promoter" as used herein refers to a region of DNA that functions to control the transcription of one or more nucleic acid sequences as defined above, and that is structurally identified by the presence of a binding site for DNA-dependent RNA-polymerase and of other DNA sequences, which interact to regulate promoter function. A functional expression promoting fragment of a promoter is a shortened or truncated promoter sequence retaining the activity as a promoter. Promoter activity may be measured by any assay known in the art (see e.g. Wood, de Wet, Dewji, and DeLuca, (1984), *Biochem Biophys. Res. Commun.* 124, 592-596; Seliger and McElroy, (1960), *Arch. Biochem. Biophys.* 88, 136-141) or commercially available from Promega®).

An "enhancer region" to be used in the expression vector as defined herein, typically refers to a region of DNA that functions to increase the transcription of one or more genes. More specifically, the term "enhancer", as used herein, is a DNA regulatory element that enhances, augments, improves, or ameliorates expression of a gene irrespective of its location and orientation vis-à-vis the gene to be expressed, and may be enhancing, augmenting, improving, or ameliorating expression of more than one promoter.

The promoter/enhancer sequences to be used in the expression vector as defined herein, may utilize plant, animal, insect, or fungus regulatory sequences. For example, promoter/enhancer elements can be used from yeast and other fungi (e.g. the GAL4 promoter, the alcohol dehydrogenase promoter, the phosphoglycerol kinase promoter, the alkaline phosphatase promoter). Alternatively, or in addition, they may include animal transcriptional control regions, e.g. (i) the insulin gene control region active within pancreatic beta-cells (see e.g. Hanahan, *et al.*, 1985. *Nature* 315: 115-122); (ii) the immunoglobulin gene control region active within lymphoid cells (see e.g. Grosschedl, *et al.*, 1984, *Cell* 38 : 647-658); (iii) the albumin gene control region active within liver (see e.g. Pinckert, *et al.*, 1987. *Genes and Dev* 1: 268-276; (iv) the myelin basic protein gene control region active within brain oligodendrocyte cells (see e.g. Readhead, *et al.*, 1987, *Cell* 48: 703-712); and (v) the gonadotropin-releasing hormone gene control region active within the hypothalamus (see e.g. Mason, *et al.*, 1986, *Science* 234: 1372-1378), and the like.

Additionally, the expression vector as defined herein may comprise an amplification marker. This amplification marker may be selected from the group consisting of, e.g. adenosine deaminase (ADA), dihydrofolate reductase (DHFR), multiple drug resistance gene (MDR), ornithine decarboxylase (ODC) and N-(phosphonacetyl)-L-aspartate resistance (CAD).

Exemplary expression vectors or their derivatives suitable for the present invention particularly include, e.g. human or animal viruses (e.g. vaccinia virus or adenovirus); insect viruses (e.g. baculovirus); yeast vectors; bacteriophage vectors (e.g. lambda phage); plasmid vectors and cosmid vectors.

The present invention additionally may utilize a variety of host-vector systems, which are capable of expressing the peptide coding sequence(s) of nucleic acids as defined above. These include, but are not limited to: (i) mammalian cell systems that are infected with vaccinia virus, adenovirus, and the like; (ii) insect cell systems infected with baculovirus and the like; (iii) yeast containing yeast vectors or (iv) bacteria transformed with bacteriophage, DNA, plasmid DNA, or cosmid DNA. Depending upon the host-vector system utilized, any one of a number of suitable transcription and translation elements may be used.

Preferably, a host cell strain, suitable for such a host-vector system, may be selected that modulates the expression of inserted sequences of interest, or modifies or processes expressed peptides encoded by the sequences in the specific manner desired. In addition, expression from certain promoters may be enhanced in the presence of certain inducers in a selected host strain; thus facilitating control of the expression of a genetically-engineered peptide. Moreover, different host cells possess characteristic and specific mechanisms for the translational and post-translational processing and modification (e.g. glycosylation, phosphorylation, and the like) of expressed peptides. Appropriate cell lines or host systems may thus be chosen to ensure the desired modification and processing of the foreign peptide is achieved. For example, peptide expression within a bacterial system can be used to produce a non-glycosylated core peptide; whereas expression within mammalian cells ensures "native" glycosylation of a heterologous peptide.

The present invention further provides the use of antibodies directed against the JNK inhibitor sequences and/or chimeric peptides as described above, for preparing a pharmaceutical composition for the treatment of non-chronic or chronic inflammatory eye diseases as defined herein. Furthermore, efficient means for production of antibodies specific for JNK inhibitor sequences according to the present invention, or for chimeric peptides containing such an inhibitor sequence, are described and may be utilized for this purpose.

According to the invention, JNK inhibitor sequences and/or chimeric peptides as defined herein, as well as, fragments, variants or derivatives thereof, may be utilized as immunogens to generate antibodies that immunospecifically bind these peptide components. Such antibodies include, e.g. polyclonal, monoclonal, chimeric, single chain, Fab fragments and a Fab expression library. In a specific embodiment the present invention provides antibodies to chimeric peptides or to JNK inhibitor sequences as defined above. Various procedures known within the art may be used for the production of these antibodies.

By way of example, various host animals may be immunized for production of polyclonal antibodies by injection with any chimeric peptide or JNK inhibitor sequence as defined above. Various adjuvants may be used thereby to increase the immunological response which include, but are not limited to, Freund's (complete and incomplete) adjuvant, mineral gels (e.g. aluminum hydroxide), surface active substances (e.g. lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, dinitrophenol, etc.), CpG, polymers, Pluronics, and human adjuvants such as Bacille Calmette-Guerin and *Corynebacterium parvum*.

For preparation of monoclonal antibodies directed towards a chimeric peptide or a JNK inhibitor sequence as defined above, any technique may be utilized that provides for the production of antibody molecules by continuous cell line culture. Such techniques include, but are not limited to, the hybridoma technique (see Kohler and Milstein, 1975. *Nature* 256: 495-497); the trioma technique; the human B-cell hybridoma technique (see Kozbor, *et al.*, 1983, *Immunol Today* 4: 72) and the EBV hybridoma technique to produce human monoclonal antibodies (see Cole, *et al.*, 1985. In: *Monoclonal Antibodies and Cancer Therapy*, Alan R. Liss, Inc., pp. 77-96). Human monoclonal antibodies may be utilized in

the practice of the present invention and may be produced by the use of human hybridomas (see Cote, *et al.*, 1983. Proc Natl Acad Sci USA 80: 2026-2030) or by transforming human B-cells with Epstein Barr Virus *in vitro* (see Cole, *et al.*, 1985. In: Monoclonal Antibodies and Cancer Therapy (Alan R. Liss, Inc., pp. 77-96).

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According to the invention, techniques can be adapted for the production of single-chain antibodies specific to the JNK inhibitor sequences and/or chimeric peptides (see e.g. U. S. Patent No. 4,946,778) as defined herein. In addition, methods can be adapted for the construction of Fab expression libraries (see e.g. Huse *et al.*, 1989. Science 246: 1275-1281) to allow rapid and effective identification of monoclonal Fab fragments with the desired specificity for these JNK inhibitor sequences and/or chimeric peptides. Non-human antibodies can be "humanized" by techniques well known in the art (see e.g. U. S. Patent No. 5,225,539). Antibody fragments that contain the idiotypes to a JNK inhibitor sequences and/or chimeric peptide as defined herein may be produced by techniques known in the art including, e.g. (i) a F(ab')₂ fragment produced by pepsin digestion of an antibody molecule; (ii) a Fab fragment generated by reducing the disulfide bridges of an F(ab')₂ fragment ; (iii) a Fab fragment generated by the treatment of the antibody molecule with papain and a reducing agent and (iv) Fv fragments.

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In one embodiment of this invention, methods, that may be utilized for the screening of antibodies and which possess the desired specificity include, but are not limited to, enzyme-linked immunosorbent assay (ELISA) and other immunologically-mediated techniques known within the art. In a specific embodiment, selection of antibodies that are specific to a particular epitope of an JNK inhibitor sequence and/or an chimeric peptide as defined herein (e.g. a fragment thereof typically comprising a length of from 5 to 20, preferably 8 to 18 and most preferably 8 to 11 amino acids) is facilitated by generation of hybridomas that bind to the fragment of an JNK inhibitor sequence and/or an chimeric peptide, as defined herein, possessing such an epitope. These antibodies that are specific for an epitope as defined above are also provided herein.

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The antibodies as defined herein may be used in methods known within the art referring to the localization and/or quantification of an JNK inhibitor sequence (and/or correspondingly to a chimeric peptide as defined above), e.g. for use in measuring levels of the peptide

within appropriate physiological samples, for use in diagnostic methods, or for use in imaging the peptide, and the like.

The JNK inhibitor sequences, chimeric peptides, nucleic acids, vectors, host cells and/or antibodies as defined according to the invention can be formulated in a pharmaceutical composition, which may be applied in the prevention or treatment of any of the diseases as defined herein, particularly in the prevention or treatment of non-chronic or chronic inflammatory eye diseases as defined herein. Typically, such a pharmaceutical composition used according to the present invention includes as an active component, e.g.: (i) any one or more of the JNK inhibitor sequences and/or chimeric peptides as defined above, and/or variants, fragments or derivatives thereof, particularly JNK inhibitor sequences according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 and/or chimeric peptides according to any of sequences of SEQ ID NOs: 9 to 12 and 23 to 32, and/or JNK inhibitor sequences according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 comprising a trafficking sequence according to any of SEQ ID NOs: 5 to 8 and 21 to 22, or variants or fragments thereof within the above definitions; and/or (ii) nucleic acids encoding an JNK inhibitor sequence and/or an chimeric peptide as defined above and/or variants or fragments thereof, and/or (iii) cells comprising any one or more of the JNK inhibitor sequences and/or chimeric peptides, and/or variants, fragments or derivatives thereof, as defined above and/or (iv) cells transfected with a vector and/or nucleic acids encoding an JNK inhibitor sequence and/or an chimeric peptide as defined above and/or variants or fragments thereof.

According to a preferred embodiment, such a pharmaceutical composition as used according to the present invention typically comprises a safe and effective amount of a component as defined above, preferably of at least one JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 and/or at least one chimeric peptide according to any of sequences of SEQ ID NOs: 9 to 12 and 23 to 32, and/or at least one JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 comprising a trafficking sequence according to any of SEQ ID NOs: 5-8 and 21 to 22, or variants or fragments thereof within the above definitions, or at least one nucleic acids encoding same, or at least one vector, host cell or antibody as defined above.

The amount of a JNK-inhibitor sequence and chimeric peptide, respectively, in the pharmaceutical composition to be administered to a subject, may –without being limited thereto - have a very low dose. Thus, the dose may be much lower than for peptide drugs
5 known in the art, such as DTS-108 (Florence Meyer-Losic et al., Clin Cancer Res., 2008, 2145-53). This has several positive aspects, for example a reduction of potential side reactions and a reduction in costs.

Preferably, the dose (per kg bodyweight) is in the range of up to 10 mmol/kg, preferably up
10 to 1 mmol/kg, more preferably up to 100 μ mol/kg, even more preferably up to 10 μ mol/kg, even more preferably up to 1 μ mol/kg, even more preferably up to 100 nmol/kg, most preferably up to 50 nmol/kg.

Thus, the dose range may preferably be from about 1 pmol/kg to about 1 mmol/kg, from
15 about 10 pmol/kg to about 0,1 mmol/kg, from about 10 pmol/kg to about 0,01 mmol/kg, from about 50 pmol/kg to about 1 μ mol/kg, from about 100 pmol/kg to about 500 nmol/kg, from about 200 pmol/kg to about 300 nmol/kg, from about 300 pmol/kg to about 100 nmol/kg, from about 500 pmol/kg to about 50 nmol/kg, from about 750 pmol/kg to about 30 nmol/kg, from about 250 pmol/kg to about 5 nmol/kg, from about 1 nmol/kg to about 10
20 nmol/kg, or a combination of any two of said values.

In this context, prescription of treatment, e.g. decisions on dosage etc. when using the above pharmaceutical composition is typically within the responsibility of general practitioners and other medical doctors, and typically takes account of the disorder to be
25 treated, the condition of the individual patient, the site of delivery, the method of administration and other factors known to practitioners. Examples of the techniques and protocols mentioned above can be found in REMINGTON'S PHARMACEUTICAL SCIENCES, 16th edition, Osol, A. (ed), 1980. Accordingly, a "safe and effective amount" as defined above for components of the pharmaceutical compositions as used according to the
30 present invention means an amount of each or all of these components, that is sufficient to significantly induce a positive modification of a non-chronic or chronic inflammatory eye diseases as defined herein. At the same time, however, a "safe and effective amount" is small enough to avoid serious side-effects, that is to say to permit a sensible relationship

between advantage and risk. The determination of these limits typically lies within the scope of sensible medical judgment. A "safe and effective amount" of such a component will vary in connection with the particular condition to be treated and also with the age and physical condition of the patient to be treated, the severity of the condition, the duration of the treatment, the nature of the accompanying therapy, of the particular pharmaceutically acceptable carrier used, and similar factors, within the knowledge and experience of the accompanying doctor. The pharmaceutical compositions according to the invention can be used according to the invention for human and also for veterinary medical purposes.

10 The pharmaceutical composition as used according to the present invention may furthermore comprise, in addition to one of these substances, a (compatible) pharmaceutically acceptable carrier, excipient, buffer, stabilizer or other materials well known to those skilled in the art.

15 In this context, the expression "(compatible) pharmaceutically acceptable carrier" preferably includes the liquid or non-liquid basis of the composition. The term "compatible" means that the constituents of the pharmaceutical composition as used herein are capable of being mixed with the pharmaceutically active component as defined above and with one another component in such a manner that no interaction occurs which would substantially reduce the pharmaceutical effectiveness of the composition under usual use conditions. Pharmaceutically acceptable carriers must, of course, have sufficiently high purity and sufficiently low toxicity to make them suitable for administration to a person to be treated.

25 If the pharmaceutical composition as used herein is provided in liquid form, the pharmaceutically acceptable carrier will typically comprise one or more (compatible) pharmaceutically acceptable liquid carriers. The composition may comprise as (compatible) pharmaceutically acceptable liquid carriers e.g. pyrogen-free water; isotonic saline or buffered (aqueous) solutions, e.g. phosphate, citrate etc. buffered solutions, vegetable oils, such as, for example, groundnut oil, cottonseed oil, sesame oil, olive oil, 30 corn oil and oil from theobroma; polyols, such as, for example, polypropylene glycol, glycerol, sorbitol, mannitol and polyethylene glycol; alginic acid, etc.. Particularly for injection of the pharmaceutical composition as used herein, a buffer, preferably an aqueous buffer, may be used.

If the pharmaceutical composition as used herein is provided in solid form, the pharmaceutically acceptable carrier will typically comprise one or more (compatible) pharmaceutically acceptable solid carriers. The composition may comprise as (compatible) pharmaceutically acceptable solid carriers e.g. one or more compatible solid or liquid fillers or diluents or encapsulating compounds may be used as well, which are suitable for administration to a person. Some examples of such (compatible) pharmaceutically acceptable solid carriers are e.g. sugars, such as, for example, lactose, glucose and sucrose; starches, such as, for example, corn starch or potato starch; cellulose and its derivatives, such as, for example, sodium carboxymethylcellulose, ethylcellulose, cellulose acetate; powdered tragacanth; malt; gelatin; tallow; solid glidants, such as, for example, stearic acid, magnesium stearate; calcium sulphate, etc..

The precise nature of the (compatible) pharmaceutically acceptable carrier or other material may depend on the route of administration. The choice of a (compatible) pharmaceutically acceptable carrier may thus be determined in principle by the manner in which the pharmaceutical composition as used according to the invention is administered. The pharmaceutical composition as used according to the invention can be administered, for example, systemically. Routes for administration include, for example, parenteral routes (e.g. via injection), such as intravenous, intramuscular, subcutaneous, intradermal, or transdermal routes, etc., enteral routes, such as oral, or rectal routes, etc., topical routes, such as nasal, or intranasal routes, etc., or other routes, such as epidermal routes or patch delivery. Particularly preferred is also the local administration at/in the eye, e.g. intravitreal administration, subconjunctival administration and/or instillation.

The suitable amount of the pharmaceutical composition to be used can be determined by routine experiments with animal models. Such models include, without implying any limitation, rabbit, sheep, mouse, rat, dog and non-human primate models. Preferred unit dose forms for injection include sterile solutions of water, physiological saline or mixtures thereof. The pH of such solutions should be adjusted to about 7.4. Suitable carriers for injection include hydrogels, devices for controlled or delayed release, polylactic acid and collagen matrices. Suitable pharmaceutically acceptable carriers for topical application include those, which are suitable for use in lotions, creams, gels and the like. If the

compound is to be administered perorally, tablets, capsules and the like are the preferred unit dose form. The pharmaceutically acceptable carriers for the preparation of unit dose forms, which can be used for oral administration are well known in the prior art. The choice thereof will depend on secondary considerations such as taste, costs and storability, which
5 are not critical for the purposes of the present invention, and can be made without difficulty by a person skilled in the art.

Pharmaceutical compositions for oral administration may be in tablet, capsule, powder or liquid form. A tablet may include a solid carrier as defined above, such as gelatin, and
10 optionally an adjuvant. Liquid pharmaceutical compositions for oral administration generally may include a liquid carrier as defined above, such as water, petroleum, animal or vegetable oils, mineral oil or synthetic oil. Physiological saline solution, dextrose or other saccharide solution or glycols such as ethylene glycol, propylene glycol or polyethylene glycol may be included.

15 For intravenous, cutaneous or subcutaneous injection, or injection at the site of affliction, the active ingredient will be in the form of a parenterally acceptable aqueous solution which is pyrogen-free and has suitable pH, isotonicity and stability. Those of relevant skill in the art are well able to prepare suitable solutions using, for example, isotonic vehicles
20 such as Sodium Chloride Injection, Ringer's Injection, Lactated Ringer's Injection. Preservatives, stabilizers, buffers, antioxidants and/or other additives may be included, as required. Whether it is a polypeptide, peptide, or nucleic acid molecule, other pharmaceutically useful compound according to the present invention that is to be given to an individual, administration is preferably in a "prophylactically effective amount or a
25 "therapeutically effective amount" (as the case may be), this being sufficient to show benefit to the individual. The actual amount administered, and rate and time-course of administration, will depend on the nature and severity of what is being treated.

Prevention and/or treatment of a disease as defined herein typically includes administration
30 of a pharmaceutical composition as defined above. The term "modulate" includes the suppression of expression of JNK when it is over-expressed in any of the above diseases. It also includes, without being limited thereto, suppression of phosphorylation of c-jun, ATF2 or NFAT4 in any of the above diseases, for example, by using at least one JNK inhibitor

sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 and/or at least one chimeric peptide according to any of sequences of SEQ ID NOs: 9 to 12 and 23 to 32, and/or at least one JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 comprising a trafficking sequence according to any of SEQ ID NOs: 5 to 8 and 21 to 22, or variants or fragments thereof within the above definitions, as a competitive inhibitor of the natural c-jun, ATF2 and NFAT4 binding site in a cell. The term "modulate" also includes suppression of hetero- and homomeric complexes of transcription factors made up of, without being limited thereto, c-jun, ATF2, or NFAT4 and their related partners, such as for example the AP-1 complex that is made up of c-jun, AFT2 and c-fos. When a non-chronic or chronic inflammatory eye disease is associated with JNK overexpression, such suppressive JNK inhibitor sequences can be introduced to a cell. In some instances, "modulate" may then include the increase of JNK expression, for example by use of an IB peptide-specific antibody that blocks the binding of an IB-peptide to JNK, thus preventing JNK inhibition by the IB-related peptide.

Prevention and/or treatment of a subject with the pharmaceutical composition as disclosed above may be typically accomplished by administering (*in vivo*) an ("therapeutically effective") amount of said pharmaceutical composition to a subject, wherein the subject may be e.g. any mammal, e.g. a human, a primate, mouse, rat, dog, cat, cow, horse or pig. The term "therapeutically effective" means that the active component of the pharmaceutical composition is of sufficient quantity to ameliorate the non-chronic or chronic inflammatory eye disease.

Accordingly, peptides as defined above, e.g. at least one JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 and/or at least one chimeric peptide according to any of sequences of SEQ ID NOs: 9 to 12 and 23 to 32, and/or at least one JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 comprising a trafficking sequence according to any of SEQ ID NOs: 5 to 8 and 21 to 22, or variants or fragments thereof within the above definitions, may be utilized in a specific embodiment of the present invention to treat non-chronic or chronic inflammatory eye diseases.

Peptides as defined above and as contained in the inventive pharmaceutical composition may be also encoded by nucleic acids. This is particularly advantageous, if the above peptides are administered for the purpose of gene therapy. In this context, gene therapy refers to therapy that is performed by administration of a specific nucleic acid as defined
5 above to a subject, e.g. by way of a pharmaceutical composition as defined above, wherein the nucleic acid(s) exclusively comprise(s) L-amino acids. In this embodiment of the present invention, the nucleic acid produces its encoded peptide(s), which then serve(s) to exert a therapeutic effect by modulating function of the disease or disorder. Any of the methods relating to gene therapy available within the art may be used in the practice of the present
10 invention (see e.g. Goldspiel, *et al.*, 1993. Clin Pharm 12: 488-505).

In a preferred embodiment, the nucleic acid as defined above and as used for gene therapy is part of an expression vector encoding and expressing any one or more of the IB-related peptides as defined above within a suitable host, i.e. an JNK inhibitor sequence according
15 to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 and/or a chimeric peptide according to any of sequences of SEQ ID NOs: 9 to 12 and 23 to 32, and/or an JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 comprising a trafficking sequence according to any of SEQ ID NOs: 5 to 8 and 21 to 22, or variants or fragments thereof within the above definitions. In a specific embodiment,
20 such an expression vector possesses a promoter that is operably-linked to coding region(s) of a JNK inhibitor sequence. The promoter may be defined as above, e.g. inducible or constitutive, and, optionally, tissue-specific.

In another specific embodiment, a nucleic acid molecule as defined above is used for gene
25 therapy, in which the coding sequences of the nucleic acid molecule (and any other desired sequences thereof) as defined above are flanked by regions that promote homologous recombination at a desired site within the genome, thus providing for intra-chromosomal expression of these nucleic acids (see e.g. Koller and Smithies, 1989. Proc Natl Acad Sci USA 86: 8932-8935).

30 Delivery of the nucleic acid as defined above according to the invention into a patient for the purpose of gene therapy, particular in the context of the above mentioned non-chronic or chronic inflammatory eye diseases as defined above may be either direct (i.e. the patient

is directly exposed to the nucleic acid or nucleic acid-containing vector) or indirect (i.e. cells are first transformed with the nucleic acid *in vitro*, then transplanted into the patient). These two approaches are known, respectively, as *in vivo* or *ex vivo* gene therapy. In a specific embodiment of the present invention, a nucleic acid is directly administered *in vivo*, where it is expressed to produce the encoded product. This may be accomplished by any of numerous methods known in the art including, e.g. constructing the nucleic acid as part of an appropriate nucleic acid expression vector and administering the same in a manner such that it becomes intracellular (e.g. by infection using a defective or attenuated retroviral or other viral vector; see U. S. Patent No. 4,980,286); directly injecting naked DNA; using microparticle bombardment (e.g. a "GeneGun" ; Biolistic, DuPont); coating the nucleic acids with lipids; using associated cell-surface receptors/transfecting agents; encapsulating in liposomes, microparticles, or microcapsules; administering it in linkage to a peptide that is known to enter the nucleus; or by administering it in linkage to a ligand predisposed to receptor-mediated endocytosis (see e.g. Wu and Wu, 1987.J Biol Chem 262: 4429-4432), which can be used to "target" cell types that specifically express the receptors of interest, etc.

An additional approach to gene therapy in the practice of the present invention involves transferring a nucleic acid as defined above into cells in *in vitro* tissue culture by such methods as electroporation, lipofection, calcium phosphate-mediated transfection, viral infection, or the like. Generally, the method of transfer includes the concomitant transfer of a selectable marker to the cells. The cells are then placed under selection pressure (e.g. antibiotic resistance) so as to facilitate the isolation of those cells that have taken up, and are expressing, the transferred gene. Those cells are then delivered to a patient. In a specific embodiment, prior to the *in vivo* administration of the resulting recombinant cell, the nucleic acid is introduced into a cell by any method known within the art including e.g. transfection, electroporation, microinjection, infection with a viral or bacteriophage vector containing the nucleic acid sequences of interest, cell fusion, chromosome-mediated gene transfer, microcell-mediated gene transfer, spheroplast fusion, and similar methods that ensure that the necessary developmental and physiological functions of the recipient cells are not disrupted by the transfer. See e.g. Loeffler and Behr, 1993. Meth Enzymol 217 : 599-618. The chosen technique should provide for the stable transfer of the nucleic acid to the

cell, such that the nucleic acid is expressible by the cell. Preferably, the transferred nucleic acid is heritable and expressible by the cell progeny.

In preferred embodiments of the present invention, the resulting recombinant cells may be delivered to a patient by various methods known within the art including, e.g. injection of epithelial cells (e.g. subcutaneously), application of recombinant skin cells as a skin graft onto the patient, and intravenous injection of recombinant blood cells (e.g. hematopoietic stem or progenitor cells). The total amount of cells that are envisioned for use depend upon the desired effect, patient state, and the like, and may be determined by one skilled within the art. Cells into which a nucleic acid can be introduced for purposes of gene therapy encompass any desired, available cell type, and may be xenogeneic, heterogeneic, syngeneic, or autogeneic. Cell types include, but are not limited to, differentiated cells such as epithelial cells, endothelial cells, keratinocytes, fibroblasts, muscle cells, hepatocytes and blood cells, or various stem or progenitor cells, in particular embryonic heart muscle cells, liver stem cells (International Patent Publication WO 94/08598), neural stem cells (Stemple and Anderson, 1992, Cell 71 : 973-985), hematopoietic stem or progenitor cells, e.g. as obtained from bone marrow, umbilical cord blood, peripheral blood, fetal liver, and the like. In a preferred embodiment, the cells utilized for gene therapy are autologous to the patient.

Alternatively and/or additionally, for treating diseases as mentioned herein targeting therapies may be used to deliver the JNK inhibitor sequences, chimeric peptides, and/or nucleic acids as defined above more specifically to certain types of cell, by the use of targeting systems such as (a targeting) antibody or cell specific ligands. Antibodies used for targeting are typically specific for cell surface proteins of cells associated with any of the diseases as defined below. By way of example, these antibodies may be directed to cell surface antibodies such as e.g. B cell-associated surface proteins such as MHC class II DR protein, CD18 (LFA-1 beta chain), CD45RO, CD40 or Bgp95, or cell surface proteins selected from e.g. CD2, CD2, CD4, CD5, CD7, CD8, CD9, CD10, CD13, CD16, CD19, CD20, CD21, CD22, CD23, CD24, CD25, CD30, CD33, CD34, CD38, CD39, CD4, CD43, CD45, CD52, CD56, CD68, CD71, CD138, etc.. Targeting constructs may be typically prepared by covalently binding the JNK inhibitor sequences, chimeric peptides, and nucleic acids as defined herein according to the invention to an antibody specific for a cell surface

protein or by binding to a cell specific ligand. Proteins may e.g. be bound to such an antibody or may be attached thereto by a peptide bond or by chemical coupling, crosslinking, etc.. The targeting therapy may then be carried out by administering the targeting construct in a pharmaceutically efficient amount to a patient by any of the administration routes as defined below, e.g. intraperitoneal, nasal, intravenous, oral and patch delivery routes. Preferably, the JNK inhibitor sequences, chimeric peptides, or nucleic acids as defined herein according to the invention, being attached to the targeting antibodies or cell specific ligands as defined above, may be released *in vitro* or *in vivo*, e.g. by hydrolysis of the covalent bond, by peptidases or by any other suitable method. Alternatively, if the JNK inhibitor sequences, chimeric peptides, or nucleic acids as defined herein according to the invention are attached to a small cell specific ligand, release of the ligand may not be carried out. If present at the cell surface, the chimeric peptides may enter the cell upon the activity of its trafficking sequence. Targeting may be desirable for a variety of reasons; for example if the JNK inhibitor sequences, chimeric peptides, and nucleic acids as defined herein according to the invention are unacceptably toxic or if it would otherwise require a too high dosage.

Instead of administering the JNK inhibitor sequences and/or chimeric peptides as defined herein according to the invention directly, they could be produced in the target cells by expression from an encoding gene introduced into the cells, e.g. from a viral vector to be administered. The viral vector typically encodes the JNK inhibitor sequences and/or chimeric peptides as defined herein according to the invention. The vector could be targeted to the specific cells to be treated. Moreover, the vector could contain regulatory elements, which are switched on more or less selectively by the target cells upon defined regulation. This technique represents a variant of the VDEPT technique (virus-directed enzyme prodrug therapy), which utilizes mature proteins instead of their precursor forms.

Alternatively, the JNK inhibitor sequences and/or chimeric peptides as defined herein could be administered in a precursor form by use of an antibody or a virus. These JNK inhibitor sequences and/or chimeric peptides may then be converted into the active form by an activating agent produced in, or targeted to, the cells to be treated. This type of approach is sometimes known as ADEPT (antibody-directed enzyme prodrug therapy) or VDEPT (virus-directed enzyme prodrug therapy); the former involving targeting the activating agent to the

cells by conjugation to a cell-specific antibody, while the latter involves producing the activating agent, e.g. a JNK inhibitor sequence or the chimeric peptide, in a vector by expression from encoding DNA in a viral vector (see for example, EP-A-415731 and WO 90/07936).

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According to a further embodiment, the JNK inhibitor sequences, chimeric peptides, nucleic acid sequences or antibodies to JNK inhibitor sequences or to chimeric peptides as defined herein, e.g. an JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 and/or a chimeric peptide according to any of sequences of SEQ ID NOs: 9 to 12 and 23 to 32, and/or an JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 comprising a trafficking sequence according to any of SEQ ID NOs: 5 to 8 and 21 to 22, or variants or fragments thereof within the above definitions, may be utilized in (*in vitro*) assays (e.g. immunoassays) to detect, prognose, diagnose, or monitor various conditions and disease states selected from non-chronic or chronic inflammatory eye diseases as defined above, or monitor the treatment thereof. The immunoassay may be performed by a method comprising contacting a sample derived from a patient with an antibody to an JNK inhibitor sequence, a chimeric peptide, or a nucleic acid sequence, as defined above, under conditions such that immunospecific-binding may occur, and subsequently detecting or measuring the amount of any immunospecific-binding by the antibody. In a specific embodiment, an antibody specific for an JNK inhibitor sequence, a chimeric peptide or a nucleic acid sequence may be used to analyze a tissue or serum sample from a patient for the presence of JNK or a JNK inhibitor sequence; wherein an aberrant level of JNK is indicative of a diseased condition. The immunoassays that may be utilized include, but are not limited to, competitive and non-competitive assay systems using techniques such as Western Blots, radioimmunoassays (RIA), enzyme linked immunosorbent assay (ELISA), "sandwich" immunoassays, immunoprecipitation assays, precipitin reactions, gel diffusion precipitin reactions, immunodiffusion assays, agglutination assays, fluorescent immunoassays, complement-fixation assays, immunoradiometric assays, and protein-A immunoassays, etc.. Alternatively, (*in vitro*) assays may be performed by delivering the JNK inhibitor sequences, chimeric peptides, nucleic acid sequences or antibodies to JNK inhibitor sequences or to chimeric peptides, as defined above, to target cells typically selected from e.g. cultured

animal cells, human cells or micro-organisms, and to monitor the cell response by biophysical methods typically known to a skilled person. The target cells typically used therein may be cultured cells (*in vitro*) or *in vivo* cells, i.e. cells composing the organs or tissues of living animals or humans, or microorganisms found in living animals or humans.

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The present invention additionally provides the use of kits for diagnostic or therapeutic purposes, particular for the treatment, prevention or monitoring of non-chronic or chronic inflammatory eye diseases as defined above, wherein the kit includes one or more containers containing JNK inhibitor sequences, chimeric peptides, nucleic acid sequences and/or antibodies to these JNK inhibitor sequences or to chimeric peptides as defined above, e.g. an anti-JNK inhibitor sequence antibody to an JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100, to a chimeric peptide according to any of sequences of SEQ ID NOs: 9 to 12 and 23 to 32, to an JNK inhibitor sequence according to any of sequences of SEQ ID NOs: 1 to 4 and 13 to 20 and 33-100 comprising a trafficking sequence according to any of SEQ ID NOs: 5 to 8 and 21 to 22, or to or variants or fragments thereof within the above definitions, or such an anti-JNK inhibitor sequence antibody and, optionally, a labeled binding partner to the antibody. The label incorporated thereby into the antibody may include, but is not limited to, a chemiluminescent, enzymatic, fluorescent, colorimetric or radioactive moiety. In another specific embodiment, kits for diagnostic use in the treatment, prevention or monitoring of non-chronic or chronic inflammatory eye diseases as defined above are provided which comprise one or more containers containing nucleic acids that encode, or alternatively, that are the complement to, an JNK inhibitor sequence and/or a chimeric peptide as defined above, optionally, a labeled binding partner to these nucleic acids, are also provided. In an alternative specific embodiment, the kit may be used for the above purposes as a kit, comprising one or more containers, a pair of oligonucleotide primers (e.g. each 6-30 nucleotides in length) that are capable of acting as amplification primers for polymerase chain reaction (PCR; see e.g. Innis, *et al.*, 1990. PCR PROTOCOLS, Academic Press, Inc., San Diego, CA), ligase chain reaction, cyclic probe reaction, and the like, or other methods known within the art used in context with the nucleic acids as defined above. The kit may, optionally, further comprise a predetermined amount of a purified JNK inhibitor sequence as defined above, a chimeric peptide as defined above, or nucleic acids encoding these, for use as a diagnostic, standard, or control in the assays for the above purposes.

The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described herein
5 will become apparent to those skilled in the art from the foregoing description and accompanying figures. Such modifications fall within the scope of the appended claims.

Various publications are cited herein, the disclosures of which are incorporated by reference in their entirety.

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Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and
15 materials are described below. All publications, patent applications, patents, and other references mentioned herein are incorporated by reference in their entirety. In the case of conflict, the present specification, including definitions, will control. In addition, the materials, methods, and examples are illustrative only and not intended to be limiting. Other features and advantages of the invention will be apparent from the following detailed
20 description and claims.

DESCRIPTION OF FIGURES

Figures 1 Clinical efficacy of SEQ ID NO: 11 in LPS-induced uveitis:

Clinical scores (expressed in arbitrary units, A.U.) were evaluated at the peak of the disease, 24 hours after (A) intravenous (IV) or (B) intravitreal (IVT) injection of JNK-inhibitor (poly-)peptide of SEQ ID NO: 11. Comparison was made with untreated uveitic eyes (LPS) and IV/IVT treatment with vehicle (n=10 eyes per group). Clinical manifestations of uveitis were reduced after (A) IV injection of JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (**
 p<0.001 vs LPS; ### p<0.001 vs Vehicle) and (B) IVT injection of (poly-)peptide of SEQ ID NO: 11 (**
 p<0.001 vs LPS; ## p<0.01 vs Vehicle) and dexamethasone (**
 p<0.05 vs LPS). No statistical difference (ns) was observed between IVT injection of JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 and dexamethasone that was used as positive control.

Figure 2 Inhibition of the JNK pathway by SEQ ID NO: 11 in LPS-induced uveitis:

Western-Blot analysis of c-Jun phosphorylation in RPE/choroid/sclera complexes 24 hours after IV or IVT injections of JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 and vehicle (n=2 eyes per group) in Endotoxin-Induced Uveitis (EIU) conditions. Inhibition of c-Jun phosphorylation by JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was visualized on (A) immunoblot (upper lane: Phospho c-Jun (Ser63); bottom lane: β -tubulin reporter protein; and confirmed by (B) densitometric quantitation.

Figure 3 Effects of SEQ ID NO: 11 on the LPS-ERK pathway activation:

Immunohistochemistry against phospho-p44/42 MAPK (Erk1/2) (green) was carried out on ocular histological sections from untreated uveitic control eyes and IV or IVT injected animals (vehicle or SEQ ID NO: 11) (n=3 eyes analyzed per condition; time point: 24h). Nuclei (in blue) were stained with DAPI. p-Erk1/2 was strongly expressed in the iris epithelium after IVT administration of vehicle (A, a) and SEQ ID NO: 11 (B, b), with no detectable difference between the two. Only a faint positive signal could be detected in retinal müller glial cells in EIU eyes treated by IV injection of

either the vehicle (E, e) or SEQ ID NO:11 (F, f) (see arrows). Scale bar: 100µm. c: cornea; ir: iris; st: stroma; ep: epithelium; ONH: optic nerve head; INL: inner nuclear layer; ONL: outer nuclear layer.

- 5 **Figure 4** Ocular biodistribution of JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 in healthy and uveitic eyes:
- 10 Immunohistochemistry against SEQ ID NO: 11 was carried out on ocular histological sections from untreated and IV or IVT injected animals (vehicle or SEQ ID NO: 11), both in (A-L) healthy and (M-W) LPS-induced inflammatory conditions (n=3 eyes analyzed per condition; time point: 24h). (A-C) SEQ ID NO: 11 was undetectable in ocular tissues 24 hours after IV injection in healthy eyes. (D-L) After IVT injection in healthy eyes, SEQ ID NO: 11 was found in iris epithelium (D, G), ciliary body epithelium (E, J), GCL (H), INL (K), IS (I) and RPE (L). In eyes with uveitis, SEQ ID NO: 11 was
- 15 detected in infiltrating inflammatory cells after IV injection of SEQ ID NO: 11 (M-P), but not after IV or IVT injections of vehicle (not shown) or in untreated EIU eyes (V,W). (Q-U) In uveitic eyes treated by IVT of SEQ ID NO: 11, distribution of SEQ ID NO: 11 was similar to that of healthy treated eyes but was also detected in migrating resident inflammatory cells (S-T). Scale bar:
- 20 50µm. c: cornea; ir: iris; l: lens; cb: ciliary body; st: stroma; ep: epithelium; GCL: ganglion cell layer; INL: inner nuclear layer; ONL: outer nuclear layer; RPE: retinal pigment epithelium; IS: photoreceptor inner segment; aq. h: aqueous humor.
- 25 **Figure 5** Reduction of LPS-induced inflammatory cell infiltration by the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11:
- 30 Infiltration of (A) macrophages (ED1 immunopositive cells) and (B) polymorphonuclear leukocytes (PMNs) was quantified on histological sections (n=6 sections per group) stained by immunohistochemistry (illustrated on Figure 6). (A) Intravenous (IV) (** p<0.005 vs LPS) and intravitreal (IVT) (** p<0.005 vs LPS; ## p<0.009 vs Vehicle) injections of the (poly-)peptide of SEQ ID NO: 11 reduced the number of ED1+. (B) the (poly-)peptide of SEQ ID NO: 11 decreased the number of PMNs after

intravenous (IV) (** $p < 0.005$ vs LPS) and intravitreal (IVT) (** $p < 0.005$ vs LPS; ## $p < 0.009$ vs Vehicle) administrations. No statistical difference (ns) was observed between IVT of the (poly-)peptide of SEQ ID NO: 11 and dexamethasone that was used as a positive control.

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Figure 6 Effect of the (poly-)peptide of SEQ ID NO: 11 on ED1+ cells, polymorphonuclear leukocytes and inducible nitric oxide synthase (iNOS) expression in LPS-induced uveitis:

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ED1 and iNOS antigens expression was analyzed by immunohistochemistry on eye cryosections of untreated or treated (IV or IVT, vehicle or SEQ ID NO: 11) uveitic rats (n=3 eyes per condition; time point: 24h). Nuclei were stained with DAPI. Numerous inflammatory cells expressing ED1 (A, D, G, J) and/or iNOS (B, E, H, K) infiltrated the anterior (A-I) and the posterior segment (J-L) of untreated EIU eyes. A few number of ED1+/iNOS+ cells were found in the iris/ciliary body (yellow cells, panels c1-2 and f1-2) but most iNOS+ cells were ED- cells suggesting that mostly PMNs produced iNOS. IV (M-U) and IVT (not shown) injections of SEQ ID NO:11 reduced the inflammatory infiltrate expressing ED1 (M, P, S) and iNOS (N, Q, T). In eyes treated by IV (M-U) and IVT (not shown) of SEQ ID NO:11, a reduced number of ED1+ cells (M, P, S) and iNOS+ cells (N, Q, T) was observed. Scale bar: 50 μ m. c: cornea; ir: iris; cb: ciliary body; ret: retina; ON: optic nerve.

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Figure 7 Down-regulation of LPS-induced iNOS expression by the (poly-)peptide of SEQ ID NO: 11:

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RT-PCR analysis of inducible nitric oxide synthase (iNOS) mRNA levels in neuroretinas 24 hours after IV or IVT injections of SEQ ID NO:11 or vehicle (n=2 eyes per group) in EIU conditions. Down-regulation of iNOS mRNA was visualized on (A) agarose gel under ultraviolet transilluminator (upper lane: 657 bp iNOS cDNA amplification product; bottom lane: 162 bp GAPDH cDNA amplification product) and confirmed by (B) densitometric quantitation

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Figure 8 Modulation of intraocular LPS-induced Chemokine/Cytokine profiles following intravenous (IV) administration of (poly-)peptide of SEQ ID NO: 11:

Multiplex analysis was performed on ocular fluids collected 6h, 24h and 48h after EIU induction. Comparison was made between uninjected control uveitic rats or with (poly-)peptide of SEQ ID NO: 11 IV treated rats (n=10 eyes analyzed per time point and per condition). Results from rats treated by IV injection of vehicle were not represented for more clarity. P values of statistical analysis are indicated on each graph (p). IV injection of (poly-)peptide of SEQ ID NO: 11 decreased chemokines production (A), and decreased pro-inflammatory and Th1 cytokines production at specific time points (B). Levels of IFN- γ and IL-10 were not represented at 48h because of being below detectable levels.

Figure 9 Modulation of intraocular LPS-induced Chemokine/Cytokine profiles following intravitreal (IVT) administration of (poly-)peptide of SEQ ID NO: 11:

Multiplex analysis was performed on ocular fluids collected 6h, 24h and 48h after EIU induction. Comparison was made between rats treated by IVT injection of vehicle and (poly-)peptide of SEQ ID NO: 11 (n=10 eyes analyzed per time point and per condition). P values of statistical analysis are indicated on each graph (p). No significant changes were observed on chemokines expression between vehicle IVT or (poly-)peptide of SEQ ID NO: 11 IVT injections except a decrease of MCP-1 (A). Little changes in cytokines expression were induced by IVT injection of the (poly-)peptide of SEQ ID NO: 11: lower levels of TNF- α , IL-6 and IL-2 at 6 hours, a lower level of IL-2 and a greater level of IL-13 at 24 hours (B)..

Figure 10 shows the the IB1 cDNA sequence from rat and its predicted amino acid sequence (SEQ ID NO:102)

Figure 11 shows the IB1 protein sequence from rat encoded by the exon-intron boundary of the rIB1 gene – splice donor (SEQ ID NO:103)

Figure 12 shows the IB1 protein sequence from *Homo sapiens* (SEQ ID NO:104)

Figure 13 shows the IB1 cDNA sequence from *Homo sapiens* (SEQ ID NO:105)

EXAMPLES

Example 1:

5 Solutions and products

An all-D-retro-inverso JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was produced by Polypeptide Laboratories (France) and purified by High Performance Liquid Chromatography (HPLC). It was analyzed by mass spectrometry for identity and RP-HPLC for purity (Polypeptide Laboratories, France). Once lyophilized, the powder was stored at 2-
10 8°C. One day prior to the experiment, the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 powder was dissolved under sterile conditions at the concentration of 10 µM in saline (NaCl 0.9%, Versol®, Aguettant) in a National Scientific (NSC) deactivated glass vial (NSC-C4015-S1) and stored at 4°C until use.

For each experiment, a fraction of freshly dissolved the JNK-inhibitor (poly-)peptide of SEQ
15 ID NO: 11 was stored at -20°C and its concentration was confirmed by High Performance Liquid Chromatography (HPLC) analysis.

Dexamethasone sodium phosphate 4 mg/mL (Soludecadron; Laboratoire Roussel, Paris, France) was used as positive control for anti inflammatory activity on EIU.10

20 Animals

7 weeks old female Lewis rats weighing 175 g (Elevage Janvier, Le Genest Saint Isle, France) were used and handled in accordance with the ARVO Statement for the Use of Animals in Ophthalmic and Vision Research. Rats were anesthetized with intramuscular injection of Ketamine (88 mg/kg) (Virbac, France) and Largactil (0.6 mg/kg) (Sanofi-Aventis, France)
25 before intravenous or ocular injection.

Injections

For intravenous (IV) injection, 100 µL of saline (NaCl 0.9%) or the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (20 µg/kg in saline) were injected in a tail vein using a 25G-
30 needle connected on a 1 mL syringe (Becton Dickinson, France). For intravitreal (IVT) injection, 5 µL of saline or the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (0.2 µg/injection in saline) were injected in both eyes using a 30G disposable needle (BD-microfine syringe, nm Médical, Asnières, France). The IV dose of 20 µg/kg (i.e. 3.5 µg/rat in

rats weighing 175g) was chosen according to studies showing that the JNK-inhibitor (poly-peptide of SEQ ID NO: 11 is active at very low doses in other models. For intravitreal injections, the inventors used the minimal dose used in direct ear application after acute noise trauma in patients. This corresponds to 5% of the dose injected intravenously.

- 5 Immediately after intravenous or intravitreal treatment, Endotoxin-Induced Uveitis (EIU) was induced by a single footpad injection of 100 μ L sterile pyrogen-free saline containing 200 μ g of LPS (Lipopolysaccharides from *Salmonella typhimurium*, Sigma-Aldrich, Saint-Quentin Fallavier, France). At the end the experiments, i.e. 6, 24 or 48h after LPS challenge, rats were anesthetized by intraperitoneal injection of pentobarbital (30 mg/kg) (Sanofi-
10 Aventis, France) before blood was collected by intracardiac puncture. Rats were then killed with a lethal dose of pentobarbital and both eyes were enucleated.

Samples collection

- Aqueous humor and vitreous were collected and pooled from each enucleated eye. Ocular
15 fluids were immediately centrifuged and the cell-free fractions were collected and frozen at -20°C before analysis by Multiplex assay. Blood samples were first clotted at room temperature for 2 hours and then at 4°C overnight. Serum was collected, centrifuged and the clear supernatant was collected and frozen at -20°C before Multiplex analysis.
- 20 Retinas and RPE/choroid/sclera complexes were carefully dissected out on enucleated eyes, snap frozen and stored at -80°C until being used for RT-PCR and Western-Blot analyses.

- For immunohistochemistry, eyeballs were collected and fixed for 1h at room temperature in phosphate buffered saline (PBS) containing 4% paraformaldehyde before being rinsed
25 overnight in PBS. The next day, samples were embedded and frozen in optimal cutting-temperature (OCT) compound (Tissue-Tek®, Sakura Finetek, Zoeterwoude, Netherland) and stored at -80°C. Frozen antero-posterior sections of eyes (10 μ m thick) were performed at the optic nerve level using a cryostat (Leica CM 3050S, Rueil-Malmaison, France) and mounted on super-frost slides for immunohistochemical analysis.

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Experimental design

In a first set of experiment, 70 rats were randomized into 14 experimental groups with 5 rats per group. Uveitis was induced in each group and rats were killed 6 hours (4 groups), 24

hours (6 groups) and 48 hours (4 groups) after LPS challenge. For each time point tested (i.e. 6h, 24h and 48h), rats treated by intravenous or intravitreal injections of vehicle (NaCl 0.9%) or the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 were compared to untreated control uveitic rats. Two additional groups, treated by intravenous injection of vehicle and intravitreal injection of dexamethasone were used at 24 hours. Clinical ocular inflammation was recorded only at 24 hours (see Scoring of Endotoxin-Induced Uveitis (EIU) section). At each time point, intraocular fluids from each eye (n=10 per group) and serum from each animal (n=5 per group) were used for Chemokine/Cytokine Multiplex Assay.

Retinas and RPE/choroid/sclera complexes were also collected at 24 hours to analyze iNOS mRNA levels by RT-PCR and c-Jun phosphorylation state by Western-Blot. Tissues were collected only from eyes treated by IV (intravenous) injection of vehicle, IV injection of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11, IVT (intravitreal) injection of vehicle and IVT injection of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (n=2 eyes per condition collected from separate rats). Eyes were selected so that their EIU clinical score was representative of the mean of the experimental group they belong to, i.e 3 for eyes treated by IV and IVT injection of vehicle and 2 for eyes treated by IV or IVT injection of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11.

A second set of experiment was designed to evaluate the anti-inflammatory effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 at the cellular and tissue level as well as the biodistribution of this molecule 24 hours after administration. Rats were randomized into 11 experimental groups. 6 groups of rats with uveitis: untreated uveitic rats, rats injected intravenously with NaCl or the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 and rats injected intravitreally with the vehicle, the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 or dexamethasone. The 5 additional groups without uveitis were: untreated healthy rats, rats treated by NaCl IV or the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 IV and rats injected IVT with NaCl or the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11. Three eyes from separate rats were collected per group and used for immunohistochemistry.

Note that, for clinical and histological analyses, dexamethasone was used as a reference treatment.

Scoring of Endotoxin-Induced Uveitis (EIU)

Animals were examined by slit lamp at 24 hours, the clinical peak of the disease in our experiments. The intensity of clinical ocular inflammation was scored on a scale from 0 to 5 for each eye as described previously¹⁰: grade (0) indicates no inflammation; grade (1) indicates the presence of a minimal iris and conjunctival vasodilation but without the observation of flare or cells in the anterior chamber (AC); grade (2) indicates the presence of moderate iris and conjunctival vessel dilation but without evident flare or cells in the AC; grade (3) indicates the presence of intense iris vessels dilation, flare and less than 10 cells per slit lamp field in the AC; grade (4) indicates the presence of more severe clinical signs than grade 3, with more than 10 cells in the AC with or without the formation of a hypopion; grade (5) indicates the presence of intense inflammatory reaction, fibrin formation in the AC and total seclusion of the pupil. Clinical evaluation was performed in a masked manner.

Western-Blot analysis

RPE/choroid/sclera complexes and neuroretinas (2 per experimental group) were snap frozen immediately after dissection and stored at -80°C until use. Tissues were homogenized in 500 µL of lysis buffer (MOPS SDS Running Buffer, Invitrogen, Cergy-Pontoise, France) supplemented with protease inhibitor cocktail (Roche Diagnostics, Meylan, France) (one tablet for 50 mL). After addition of LDS Sample Buffer (Invitrogen) and heating for 5 min at 100°C, equal amounts of proteins were subjected to electrophoresis in a NuPAGE 4-12% Bis-Tris gel (Invitrogen) using MOPS SDS Running Buffer. The bands obtained were then electrotransferred onto nitrocellulose membranes (Schleicher & Schuell BioScience, Dassel, Germany).

Western-blot analyses were carried out to analyze the effect of the JNK-inhibitor (poly-peptide of SEQ ID NO: 11 on the three mitogen-activated protein kinase (MAPK) pathways. To analyze the JNK pathway, blots were sequentially incubated with a rabbit Phospho-c-Jun (Ser63) primary antibody (or Phospho-c-Jun (Ser73) antibody) and an anti-rabbit IgG HRP-linked secondary antibody according to the manufacturer's instruction (PhosphoPlus c-Jun (Ser63) II and c-Jun (Ser73) antibody kit (9260) purchased from Cell Signaling Technology (Ozyme, St Quentin Yvelines, France)). Bands were visualized using the ECL Western Blotting Detection Reagents Kit (Amersham Biosciences, Orsay, France). Blots were then

dehybridized and rehybridized successively with a mouse anti α -tubulin (D-10) (sc-5274) primary antibody (dilution 1:400) and a HRP conjugated goat anti-mouse IgG secondary antibody (sc-3697) (dilution 1:5000) (both purchased from Santa Cruz Biotechnology (Tebu-bio, Le Perray en Yvelines Cedex, France)). The relative band intensity for phospho c-Jun (Ser 63 or Ser73) was calculated in comparison to that for α -tubulin after densitometry analysis (ImageJ software).

To analyze the ERK and p38 MAPK pathways, blots were sequentially incubated with a rabbit phospho-p44/42 MAPK (Erk1/2) (Thr202/Tyr204) (4370) primary antibody (or rabbit phospho-p38 MAPK (Thr180/Tyr182) (9215) antibody) and a horseradish peroxidase-conjugated goat anti-rabbit IgG (H+L) (PI-1000 - Vector Laboratories, Clinisciences, Montrouge, France) secondary antibody at dilution 1:5000. Blots were then dehybridized and rehybridized successively with a rabbit p44/42 MAPK (Erk1/2) (4695) (or rabbit p38 MAP Kinase (9212) antibody) and the same secondary antibody as above. Primary antibodies were purchased from Cell Signaling Technology (Ozyme, St Quentin Yvelines, France) and all steps performed following the manufacturer's instruction.

Immunohistochemistry

To characterize the cellular infiltrate, sections were double-stained with ED1 and iNOS. Briefly, after permeabilization with 0.1% TritonX-100 in phosphate buffered saline (PBS) for 30 min, specimens were rinsed and saturated for 30 min with 5% skimmed milk in PBS. They were incubated overnight at 4°C with the two following primary antibodies: a 1:50 mouse monoclonal anti-macrosialin CD68 (clone ED1), directed against a cytoplasmic antigen in rat monocytes, macrophages and dendritic cells (purchased from Serotec Ltd. (Oxford, UK)) and a polyclonal rabbit anti-iNOS (1/75e; Transduction Laboratories, Lexington, KY). After washing, sections were incubated for 1 hour at room temperature with a secondary Alexa Fluor 594 (red)-conjugated donkey anti-mouse monoclonal antibody (mAb) and a secondary Alexa Fluor 488 (green)-conjugated goat anti-rabbit mAb each at dilution 1:250 (Invitrogen, Cergy Pontoise, France). For each step, antibodies were diluted in PBS-1% skimmed milk - 0.1% TritonX100. Different controls were included in every staining run: negative controls without primary antibodies and isotype controls by incubation with normal mouse or rabbit serum immunoglobulin (Ig) in place of primary antibodies. After staining nuclei with DAPI (Sigma-Aldrich, Saint-Quentin Fallavier, France),

sections were mounted in PBS/Glycerol (1/1) and observed by fluorescence photomicroscopy (FXA, Microphot, Nikon, Melville, USA). Digitized micrographs were obtained using a digital camera (Spot, BFI Optilas, Evry, France). ED1 positive cells and polymorphonuclear cells, identified by the shape of their nuclei stained with DAPI, were quantified on histological sections. The analysis was performed on 3 eyes per experimental group, with 2 different sections per eye at the optic nerve head level. Results were expressed as mean \pm standard error of the mean (SEM).

Immunostaining of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was performed on healthy and uveitic eyes to study its ocular biodistribution after systemic or local administration. Briefly, sections were permeabilized as described above before being sequentially incubated with an anti- SEQ ID NO: 11 purified rabbit IgG and a secondary Alexa 594 (red)-conjugated goat anti-rabbit IgG (Invitrogen, Cergy Pontoise, France) diluted 1:100 and 1:250 in PBS respectively. Immunostaining of untreated healthy and uveitic eyes were used as negative controls. Nuclei were stained with DAPI before mounting and observation.

Immunostaining of p-Erk1/2 was performed to evaluate the effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 on the ERK pathway after IV or IVT administration. Sections were permeabilized as described above and incubated in blocking solution containing 0.1% Triton X-100 and 10% FCS (fetal calf serum) in PBS for 1 hour at room temperature. Sections were incubated overnight at 4°C with a rabbit anti-phospho-p44/42 MAPK (Erk1/2) primary antibody (4370) purchased from Cell Signaling Technology (Ozyme, St Quentin Yvelines, France) diluted 1:400 in blocking solution. After having been rinsed three times in PBS, sections were incubated with a secondary Alexa Fluor 488 (green)-conjugated goat anti-rabbit mAb (diluted 1:300 in blocking solution) for 2 hours at room temperature. Nuclei were stained with DAPI before mounting and observation.

Evaluation of iNOS expression in ocular tissues using semi-quantitative PCR

Two eyes per group were used for this analysis. Immediately after dissection, retinas extracted from each eye were separately snap frozen and stored at -80°C until use. Total RNA was extracted from tissues (RNeasy minikit, Qiagen, Courtaboeuf, France) according to the manufacturer's instructions. Reverse transcription was performed on 1 μ g of total RNA

in a total volume of 20 μ L using Superscript II Reverse Transcriptase (Invitrogen, Cergy-Pontoise, France) following the manufacturer's instructions. To amplify GAPDH and iNOS cDNA, Polymerase-Chain Reaction (PCR) was conducted in a total volume of 25 μ L containing 2 μ L of first-strand reaction product, 0.4 μ M forward and 0.4 μ M reverse primers, 5 0.4 μ M dNTP Mix, 1.5 mM MgCl₂, 1X PCR buffer and 2.5 U Taq DNA polymerase (Invitrogen, Cergy Pontoise, France). Primers specific for GAPDH (Forward: 5'-ATGCCCCCATGTTTGTGATG-3'; Reverse: 5'-ATGGCATGGACTGTGGTCAT-3') and iNOS (Forward: 5'-TTTCTCTTCAAAGTCAAATCCTACCA-3'; Reverse: 5'-TGTGTCTGCAGATGTGCTGAAAC-3') were obtained from Invitrogen. After an initial 10 denaturation (3 min at 94°C), 30 to 32 PCR cycles of denaturation (30 s, 94°C), annealing (1 min, 58°C (GAPDH) and 52°C (iNOS)) and elongation (1 to 2 min, 72°C) were performed on a Crocodile III (Appligene Oncor). The final cycle was completed by 5 min of elongation at 72°C. PCR fragments (162 bp for GAPDH and 657 bp for iNOS) were analyzed by 2.5% agarose gel electrophoresis and visualized by ethidium bromide staining under UV light. 15 The relative band intensity for iNOS was calculated in comparison to that for GAPDH after densitometry analysis (ImageJ software).

Chemokine/Cytokine Multiplex Assay

Intraocular fluids (diluted to obtain a final volume of 25 μ L) and sera (25 μ L of 1:5 dilution) 20 were subjected to multiplex bead analysis. This method uses microspheres as the solid support for immunoassays¹² and allows the titration of a greater number of cytokines with increased sensitivity than occurs with ELISA.¹³ For each sample, seventeen analytes were quantified simultaneously using the rat Cytokine/Chemokine-17plex kit (Milliplex Map Kit, Millipore, Saint-Quentin-en-Yvelines, France) according to the manufacturer's instructions: 25 Chemokines MCP-1/CCL2, MIP-1 α /CCL3, RANTES/CCL5, IP-10/CXCL10 (IFN-inducible protein-10) and GRO/KC ; proinflammatory mediators IL-1 , IL-18 and TNF- ; Th1/Th2/Th17 cytokines IL-2 and IFN- / IL-4, IL-5, IL-6, IL-10 and IL-13 / IL-17. The assay was performed in a 96-well filter plate and standard curves for each cytokine were generated with a Rat Cytokine Standard provided in the kit. All incubation steps were 30 performed under medium orbital agitation and in the dark to protect the beads from light. Data acquisition and analysis were performed with the manager software version 4.1 (Bioplex; Bio-Rad) with four or five logistic parameters for standard curves. Detection thresholds for all the analytes were estimated around 1 to 10 pg/mL.

Statistical analysis

Numerical results were expressed as mean \pm standard error of the mean (SEM). Data were compared using the nonparametric Mann-Whitney U-test. $P < 0.05$ was considered statistically significant.

Results:

The all-D-retro-inverso JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 significantly reduced endotoxins induced uveitis (EIU). The JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 significantly reduced EIU clinical scores after 20 $\mu\text{g/kg}$ intravenous (IV) injection (2.0 ± 0.1) compared to untreated uveitic eyes (3.2 ± 0.1 , $p < 0.001$) and vehicle IV (3.2 ± 0.1 , $p < 0.001$) (Figure 1A). In a similar manner, clinical scores were significantly decreased after 0.2 $\mu\text{g/injection}$ intravitreal (IVT) administration of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (2.2 ± 0.2) in comparison to untreated uveitic eyes (3.2 ± 0.1 , $p < 0.001$) and vehicle IVT (3.0 ± 0.1 , $p < 0.01$) (Figure 1B). The effect of IVT injection of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 on clinical signs of EIU was not statistically different from that observed after IVT of dexamethasone (1.8 ± 0.4) suggesting that the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was as efficient as dexamethasone in reducing EIU when administered at this time point (Figure 1B).

Efficacy of the (poly-)peptide of SEQ ID NO: 11 resulted from JNK pathway inhibition

To determine whether the clinical effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was related to its mode of action, i.e. its ability to interfere with JNK signaling, c-Jun phosphorylation state was analyzed in ocular tissues by Western-Blot. Phosphorylation of c-Jun on Ser63 (Figure 2A) and Ser73 residues was reduced in RPE/ choroid extracts 24 hours after the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was injected intravenously or intravitreally. In the neuroretina, phospho c-Jun could only be faintly detected. An approximately 3-fold decrease in c-Jun phosphorylation was observed in RPE/ choroid either after IV (0.28 ± 0.01 vs 0.77 ± 0.26 in IV of vehicle) or IVT (0.35 ± 0.08 vs 0.79 ± 0.25 in IVT of vehicle) administration of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (Figure 2B). The ability of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 to block c-Jun

NH2-terminal kinases (JNK) activity in the eye tissues demonstrated the specific intraocular activity of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11.

To determine whether the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 could have any effect on the other MAPK pathways, the phosphorylation state of Erk1/2 and p38 was evaluated. Whereas Erk1/2 and p38 were detected in RPE/choroid complexes at similar levels among all groups, the phosphorylation form of these two MAPK could not be detected by western-blot analysis (data not shown). These results demonstrate that JNK is the predominantly activated MAPK pathway in RPE/choroid during EIU. Using histochemical analysis, performed without any signal amplification, we found an intense p-Erk1/2 signal in inflammatory cells infiltrating in the anterior and the posterior segments of the eye in the control LPS and saline treated eyes (Figure 3C, 3D). The effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 either administered intravenously or intravitreously could not be evaluated on those cells, since the infiltration was almost absent in the treated eyes. However, in the neuroretina, where p-Erk1/2 could be detected and located in retinal Müller glial (RMG) cells in the control and saline treated eyes (Figure 3E), no effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 administered by either route was observed (Figure 3F). Interestingly, in the iris, an intense p-Erk1/2 signal was observed in the epithelium of the control and saline injected eyes (Figure 3A) with no effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 treated on the p-Erk1/2 signal in these cells (Figure 3B), strongly suggesting that the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 does not seem to directly act on p-Erk1/2 phosphorylation during EIU in our model, at least in resident cells.

Differential distribution of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 in ocular tissues after IV and IVT administrations

Immunohistochemistry was carried out on histological sections to evaluate the biodistribution of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 in ocular tissues 24 hours after systemic (IV) or local (IVT) administration, both in healthy eyes and in uveitic conditions (Figure 4). No inflammatory cell infiltration was observed in healthy eyes either after IV or IVT of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 or vehicle. No immunoreactivity against the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was detected in untreated control eyes or in eyes treated by vehicle, demonstrating the specificity of the

signal observed in the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11-treated eyes. Whereas no signal was observed in normal eyes after systemic (IV) injection (Figure 4 A-C) at the dose used, the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was distributed in almost all ocular tissues of normal rats after IVT administration (Figure 4 D-L). Interestingly, an accumulation of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was detected mainly in the iris/ ciliary body epithelium (panels G and J) and in the retinal pigment epithelium (panel L). Penetration of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was also detected in the iris stroma (panel G) as well as in the neural retina in the ganglion cell layer (GCL, panel H), the inner nuclear layer (INL, panel K) and the inner segment (IS, panel I) of photoreceptor cells (PR). In all cell types, the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 accumulated within the cytoplasm. Occasional staining was found in the corneal endothelium and in the lens capsule.

In uveitic conditions, no the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 staining was detected in ocular tissues and in infiltrating inflammatory cells of untreated eyes (Figure 4, panels V-W). IV or IVT of vehicle gave similar results to those from untreated eyes . In EIU eyes treated by IV injection of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11, it was not detected in ocular tissues, but occasional infiltrating inflammatory cells were immunopositive in the iris (panel O) and in the aqueous humor (panel P). In uveitic eyes treated by IVT injection, the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was mostly found in ocular tissues like in healthy eyes and in resident cells that are mobilized and participate actively to the inflammatory processes in pathological conditions such as microglial cells (panels S-T).

A significant reduction in cells infiltrating the ocular tissues resulted from the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 administration

To further characterize the effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 in uveitis, the infiltrated inflammatory cells were quantified in ocular tissues (Figure 5) by numeration on histological sections immunostained with ED1 and iNOS antibodies (Figure 6). 24 hours after LPS challenge, the number of ED1 positive cells was significantly reduced in eyes treated with IV injection of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (137 ± 7) (Figure 5A, Figure 6M, P, S) as compared to untreated uveitic eyes (LPS) (187 ± 13 , $p < 0.005$) (Figure 5A, Figure 6A, D, G, J) or vehicle injected eyes . Similarly, IVT of the JNK-

inhibitor (poly-)peptide of SEQ ID NO: 11 significantly reduced ED1 positive infiltrating cells (93 ± 8) as compared to vehicle IVT injected eyes (175 ± 15 , $p < 0.009$) and untreated uveitic eyes ($p < 0.005$) (Figure 5A). The reducing effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 on ED1 positive cell infiltration (93 ± 8) did not differ from that induced by dexamethasone (79 ± 15), suggesting that both treatments have a similar efficacy on this parameter.

The number of polymorphonuclear cells (PMN) (Figure 5B) was also significantly reduced at 24 hours after IV administration of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (60 ± 6) as compared to control eyes (237 ± 15 , $p < 0.005$), and after IVT injection of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (40 ± 5) as compared to IVT injection of the vehicle (152 ± 31 , $p < 0.009$) and control uveitic eyes ($p < 0.005$). Again, the effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 on PMN ocular tissue infiltration did not significantly differ from that of dexamethasone (42 ± 11).

The JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 down regulates iNOS expression. Since iNOS (inducible nitric oxide synthase) has been described as a key mediator in the pathogenesis of uveitis,^{14,15} the effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 on its expression was investigated at both the protein and mRNA levels.

As shown on Figure 6, the number of iNOS positive cells was reduced in eyes treated with injection of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 IV (Figure 6N, Q, T) or IVT compared to control eyes (panels B E, H, K). Among iNOS positive cells observed in control eyes, a few number were ED1+ cells while most of them were ED1- suggesting that mostly PMNs produced iNOS (insets c, f, i). In eyes from the JNK-inhibitor -treated rats, the only cells still expressing iNOS were intra tissular ED1 positive cells located at the ciliary body root (inset r2).

The effect of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 on iNOS expression was confirmed by RT-PCR on ocular tissues (Figure 7). Levels of iNOS mRNA were down-regulated from 1.02 ± 0.21 to 0.40 ± 0.11 after IV of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 and from 1.18 ± 0.05 to 0.27 ± 0.09 in eyes treated by IVT injection. Comparisons were made with IV or IVT of vehicle respectively.

Chemokine/ cytokine profiles in ocular media of eyes treated with the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11

To evaluate the effect of the treatment on the production of pro- and anti- inflammatory mediators, chemokines and cytokines were dosed by multiplex analysis on ocular media (Figures 8 and 9) and sera .

Among the 17 chemokines/ cytokines tested, some were below detectable levels both in control or treated eyes: IP-10, IL-5, IL-17. Other did not differ in treated versus untreated eyes at any of the tested time points: IL-18, IL-4, IL-1 β . In the serum , while some cytokines tended to change after IV administration of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 (reduction of MIP-1 α and IL-2) or after IVT (reduction of IL-2), this was not statistically significant. For the other chemokines/ cytokines, their profile was different in ocular fluids from eyes treated with IV administration of the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 as compared to the IVT administration. Indeed, when the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was injected systemically at the time of LPS challenge, it induced a significant reduction of MCP-1, MIP-1 α and RANTES at 6 and 24 hours (Figure 8A). GRO/KC was also significantly reduced at 6 hours. Th1 cytokines such as TNF- α , IL-6 and INF- γ were significantly reduced at different time points while IL-10 tended to increase at 6 hours in treated eye (but not significantly), suggesting a switch towards a Th2 profile (Figure 8B). No statistical differences were noticed between eyes from vehicle IV injected rats and untreated uveitic control eyes.

When the JNK-inhibitor (poly-)peptide of SEQ ID NO: 11 was injected into the vitreous at the time of LPS challenge, the chemokine/ cytokine profiles was not strikingly different from that of eyes injected with vehicle (Figure 9). It is interesting to note though that IVT of vehicle had a marked effect on the cytokine profile as compared to untreated uveitic control eyes. At 6 hours, a trend to a decrease of MCP-1, TNF- α , IL-6, IL-2 and at 24 hours, a marked decrease of IL-2 and an increase of IL-13 were detected suggesting again a switch towards a Th2 profile.

CLAIMS

1. Use of a JNK inhibitor (poly-)peptide comprising less than 150 amino acids in length for the preparation of a pharmaceutical composition for treating a chronic or non-chronic inflammatory eye disease in a subject.
5
2. The use of a JNK inhibitor (poly-)peptide according to claim 1, wherein the JNK inhibitor (poly-)peptide comprises a range of 5 to 150 amino acid residues, more preferably 10 to 100 amino acid residues, even more preferably 10 to 75 amino acid residues and most preferably a range of 10 to 50 amino acid residues.
10
3. The use of a JNK inhibitor (poly-)peptide of any of claims 1 or 2, wherein the JNK inhibitor (poly-)peptide binds c-jun amino terminal kinase (JNK).
- 15 4. The use of a JNK inhibitor (poly-)peptide of any of claims 1 to 3, wherein the JNK inhibitor (poly-)peptide inhibits the activation of at least one JNK targeted transcription factor when the JNK inhibitor (poly-)peptide is present in a JNK expressing cell.
- 20 5. The use of a JNK inhibitor (poly-)peptide of any of claims 1 to 4, wherein the JNK targeted transcription factor is selected from the group consisting of c-Jun, ATF2, and Elk1.
- 25 6. The use of a JNK inhibitor (poly-)peptide of any of claims 1 to 5, wherein the JNK inhibitor (poly-)peptide alters a JNK effect when the peptide is present in a JNK expressing cell.
- 30 7. The use of a JNK inhibitor sequence of any of claims 1 to 6, wherein the JNK inhibitor (poly-)peptide is composed of L-amino acids, D-amino acids, or a combination of both, preferably comprises at least 1 or even 2, preferably at least 3, 4 or 5, more preferably at least 6, 7, 8 or 9 and even more preferably at least 10 or more D- and/or L-amino acids, wherein the D- and/or L-amino acids may be

arranged in the JNK inhibitor (poly-)peptide in a blockwise, a non-blockwise or in an alternate manner.

- 5 8. The use according to any of claims 1 to 7, wherein the JNK inhibitor (poly-)peptide comprises a fragment, variant, or variant of such fragment of a human or rat IB1 (poly-)peptide as defined or encoded by any of sequences according to SEQ ID NO: 102, SEQ ID NO: 103, SEQ ID NO: 104 or SEQ ID NO: 105.
- 10 9. The use of a JNK inhibitor (poly-)peptide of any of claims 1 to 8, wherein the inhibitor sequence comprises or consists of at least one amino acid sequence according to SEQ ID NOs: 1 to 4, 13 to 20 and 33 to 100, or a fragment, derivative or variant thereof.
- 15 10. Use of a chimeric (poly-)peptide comprising at least one first domain and at least one second domain linked by a covalent bond, the first domain comprising a trafficking (poly-)peptide, and the second domain comprising a JNK inhibitor (poly-)peptide as defined in any of claims 1 to 9 for the preparation of a pharmaceutical composition for treating a chronic or non-chronic inflammatory eye disease in a subject.
- 20 11. The use of the chimeric (poly-)peptide of claim 10, wherein the chimeric (poly-)peptide is composed of L-amino acids, D-amino acids, or a combination of both, preferably comprises at least 1 or even 2, preferably at least 3, 4 or 5, more preferably at least 6, 7, 8 or 9 and even more preferably at least 10 or more D- and/or L-amino acids, wherein the D- and/or L-amino acids may be arranged in the chimeric peptide in a blockwise, a non-blockwise or in an alternate manner.
- 25 12. The use of the chimeric (poly-)peptide of claim 10 or 11, wherein the trafficking (poly-)peptide comprises the amino acid sequence of a human immunodeficiency virus TAT polypeptide.
- 30

13. The use of the chimeric (poly-)peptide of any of claims 10 to 12, wherein the trafficking sequence consists of or comprises the amino acid sequence of SEQ ID NO: 5, 6, 7, 8, 21 or 22.
- 5 14. The use of the chimeric (poly-)peptide of any of claims 10 to 13, wherein the trafficking (poly-)peptide augments cellular uptake of the peptide.
15. The use of the chimeric (poly-)peptide of any of claims 10 to 14, wherein the trafficking (poly-)peptide directs nuclear localization of the peptide.
- 10 16. The use of the chimeric (poly-)peptide of any of claims 10 to 15, wherein the chimeric (poly-)peptide consists of or comprises the amino acid sequence of any of SEQ ID NOs: 9 to 12 and 23 to 32, or a fragment, or variant thereof.
- 15 17. The use of the chimeric peptide of any of claims 10 to 15, wherein the chimeric (poly-)peptide consists of or comprises the amino acid sequence of SEQ ID NO: 9 or 11.
- 20 18. Use of an isolated nucleic acid encoding a JNK inhibitor (poly-)peptide as defined in any of claims 1 to 9 or a chimeric (poly-)peptide as defined in any of claims 10 to 17 for the preparation of a pharmaceutical composition for treating chronic or non-chronic inflammatory eye diseases in a subject.
- 25 19. Use of a vector comprising the nucleic acid as defined in claim 18 for the preparation of a pharmaceutical composition for treating chronic or non-chronic inflammatory eye diseases in a subject.
- 30 20. Use of a cell comprising the vector as defined in claim 19 for the preparation of a pharmaceutical composition for treating chronic or non-chronic inflammatory eye diseases in a subject.
21. Use of an antibody which binds immunospecifically to a JNK inhibitor (poly-)peptide according to any of claims 1 to 9 or to a chimeric (poly-)peptide according

to any of claims 10 to 17 for the preparation of a pharmaceutical composition for treating chronic or non-chronic inflammatory eye diseases in a subject.

22. Use according to any of the preceding claims, wherein the pharmaceutical composition is to be administered by an administration route selected from the group consisting of parenteral routes, including intravenous, intramuscular, subcutaneous, intradermal, transdermal, enteral routes, including orally, rectally, topical routes, including nasal, intranasal, other routes, including epidermal or patch delivery, and local administration to the eye, in particular intravitreal administration, subconjunctival administration and/or instillation.
23. Use according to any of the preceding claims, wherein the non-chronic or chronic inflammatory eye disease is selected from inflammatory diseases of the blephara, conjunctiva, cornea, sclera, the vitreous body, uvea, ciliary body, choroid, orbital bone, lacrimal gland, or iris, in particular wherein the inflammatory disease is selected from hordeolum, chalazion, conjunktivitis, keratitis, scleritis, episcleritis, endophthalmitis, panophtalmitis, iritis, uveitis, cyclitis, chorioiditis, orbital phlegmon, and myositis of the eye muscle.
24. The use according to anyone of the preceding claims, wherein a dose (per kg bodyweight) of the JNK inhibitor (poly-)peptide and/or chimeric (poly-)peptide is in the range of up to 10 mmol/kg, preferably up to 1 mmol/kg, more preferably up to 100 μ mol/kg, even more preferably up to 10 μ mol/kg, even more preferably up to 1 μ mol/kg, even more preferably up to 100 nmol/kg, most preferably up to 50 nmol/kg.
25. The use according to anyone of the preceding claims, wherein a dose of the JNK inhibitor (poly-)peptide and/or chimeric (poly-)peptide is in the range of from about 1 pmol/kg to about 1 mmol/kg, from about 10 pmol/kg to about 0,1 mmol/kg, from about 10 pmol/kg to about 0,01 mmol/kg, from about 50 pmol/kg to about 1 μ mol/kg, from about 100 pmol/kg to about 500 nmol/kg, from about 200 pmol/kg to about 300 nmol/kg, from about 300 pmol/kg to about 100 nmol/kg, from about 500 pmol/kg to about 50 nmol/kg, from about 750 pmol/kg to about 30 nmol/kg, from

about 250 pmol/kg to about 5 nmol/kg, from about 1 nmol/kg to about 10 nmol/kg, or a combination of any two of said values.

- 5 26. The use according to anyone of the preceeding claims, wherein the inflammatory eye disease is uveitis and wherein the (poly-)peptide consists of or comprises the amino acid sequence of SEQ ID NO: 11, or consists of or comprises a amino acid sequence sharing a sequence identity of at least about 30%, 50%, 70%, 80%, 90%, 92% or even 95% with SEQ ID NO: 11.

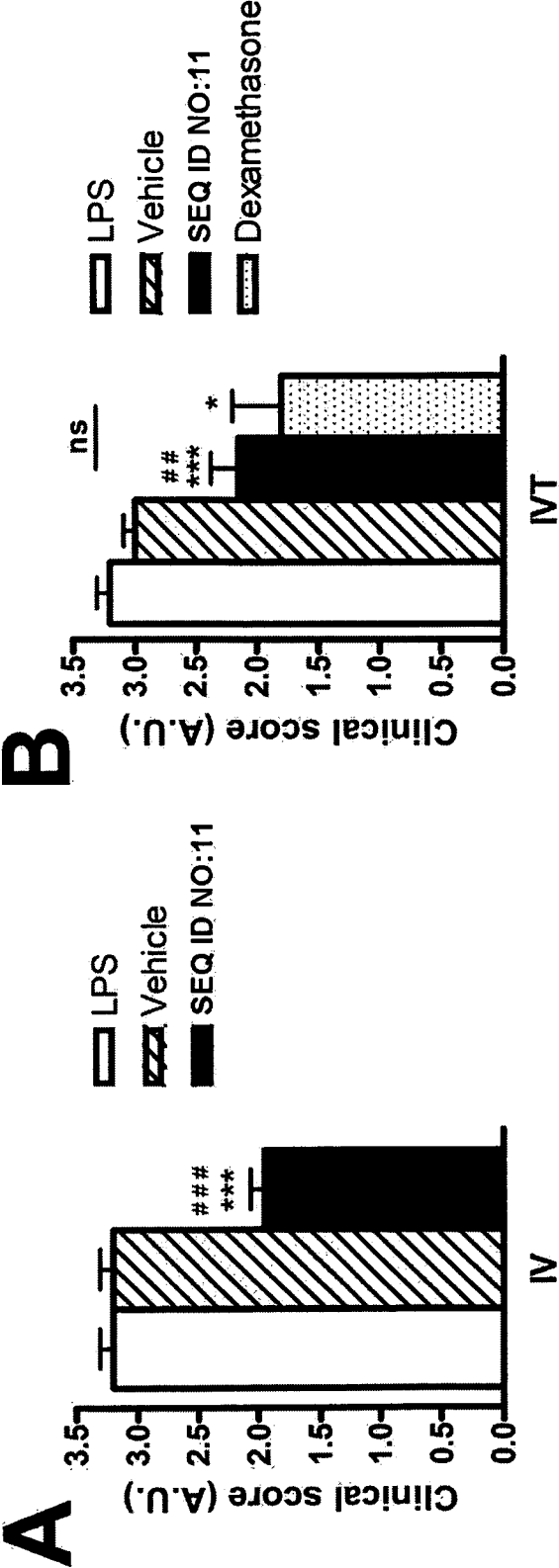


Fig. 1

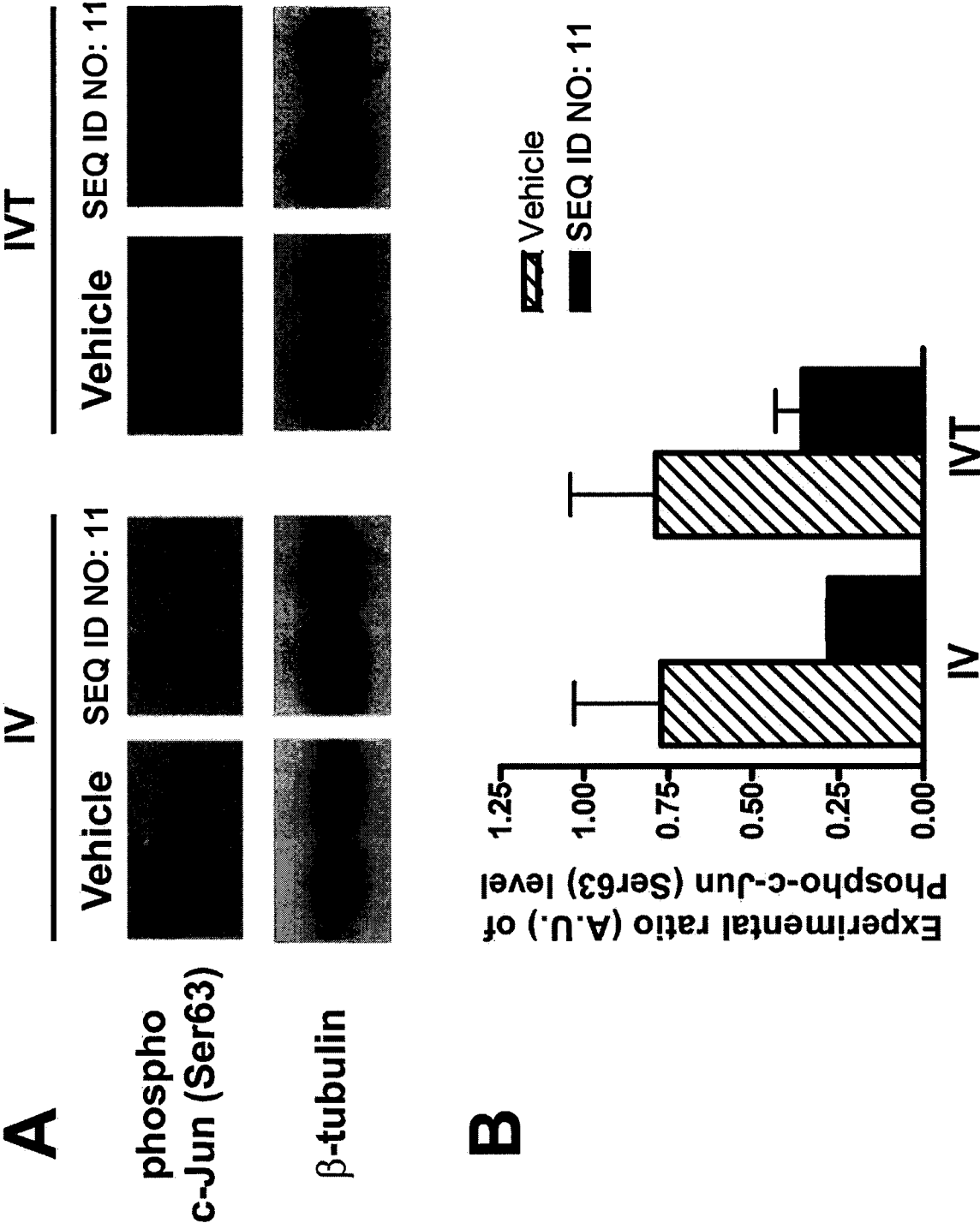


Fig. 2

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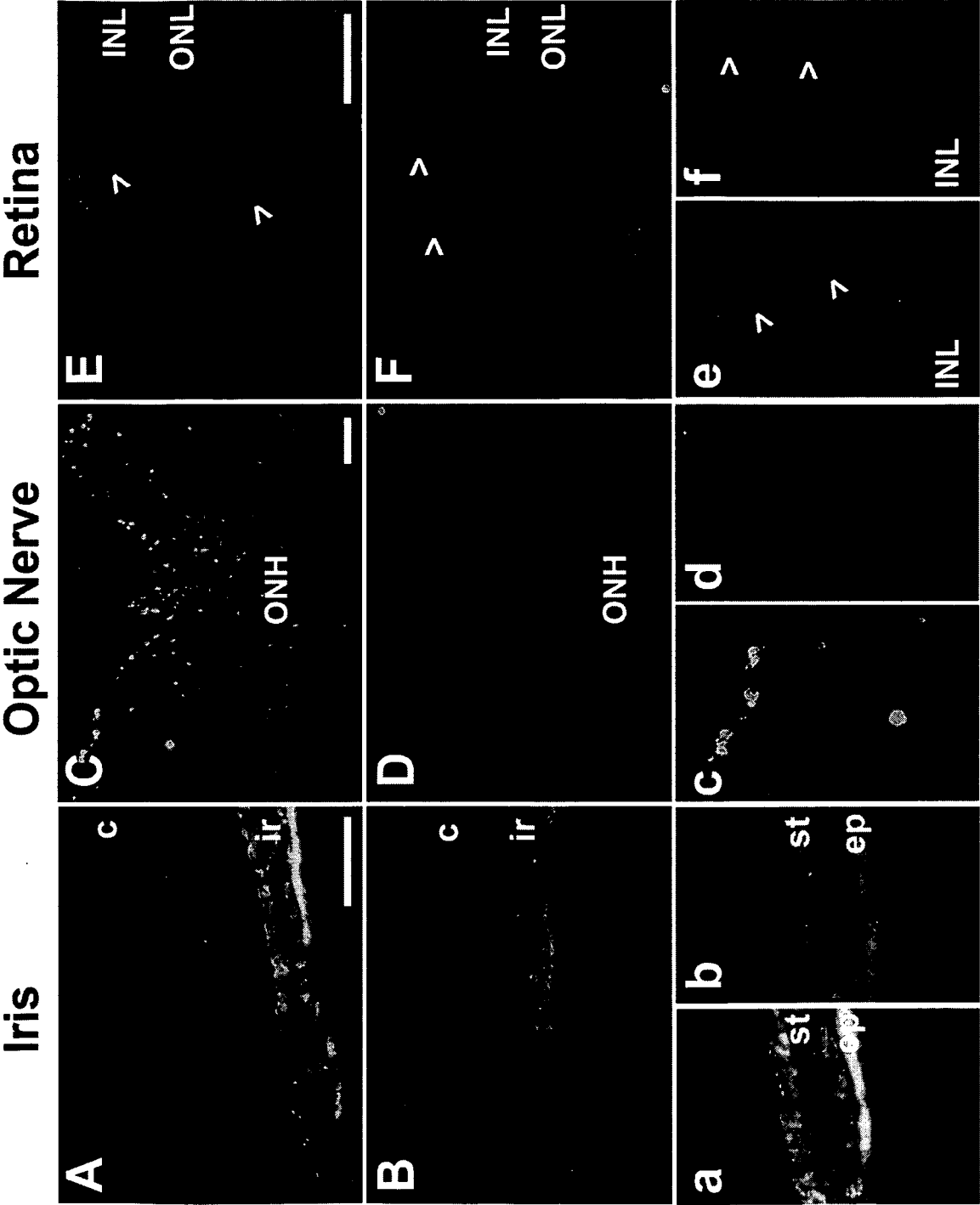


Fig. 3

4/21

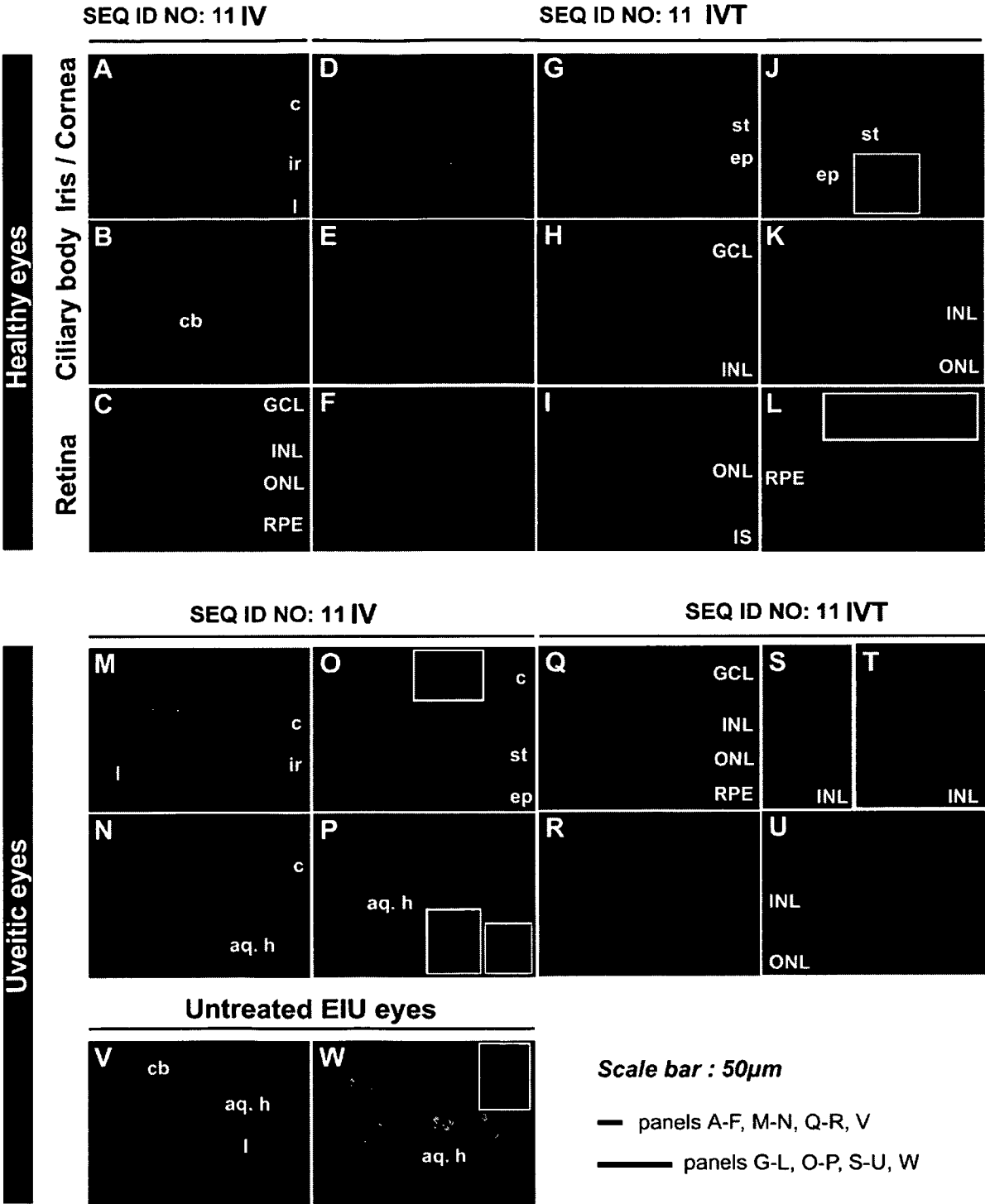


Fig. 4

5/21

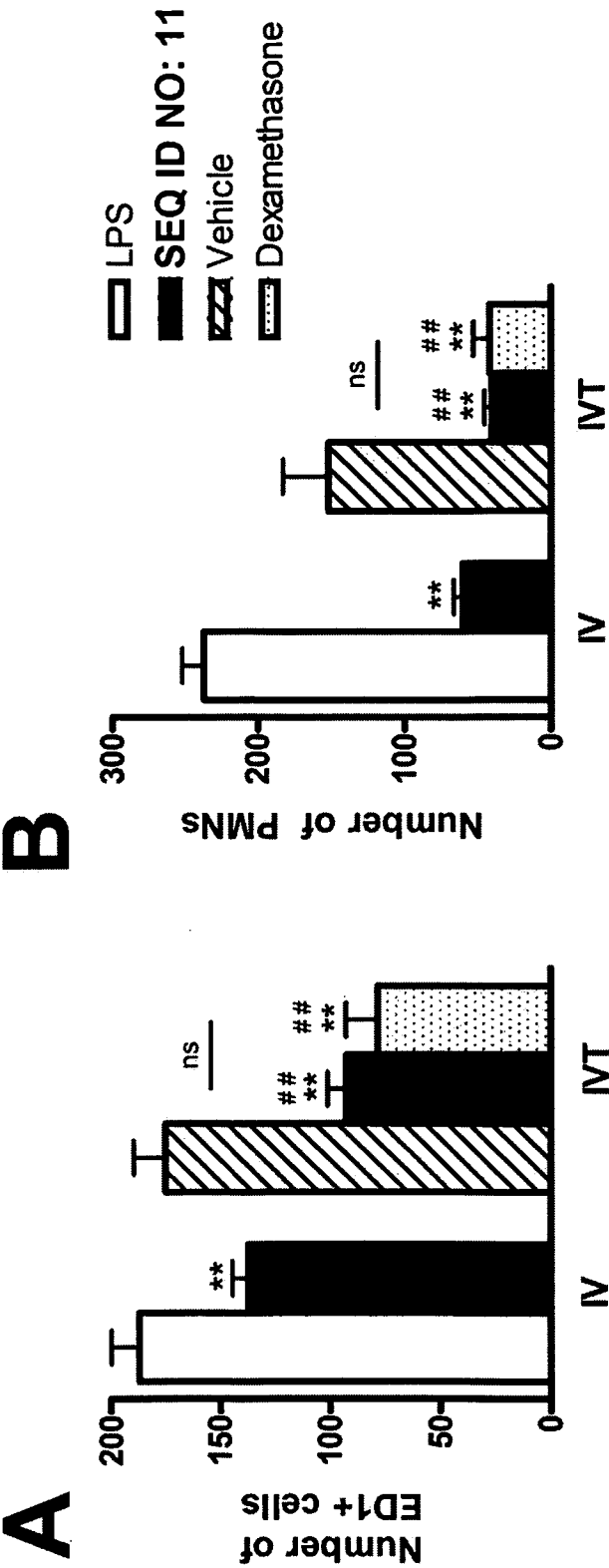


Fig. 5

6/21

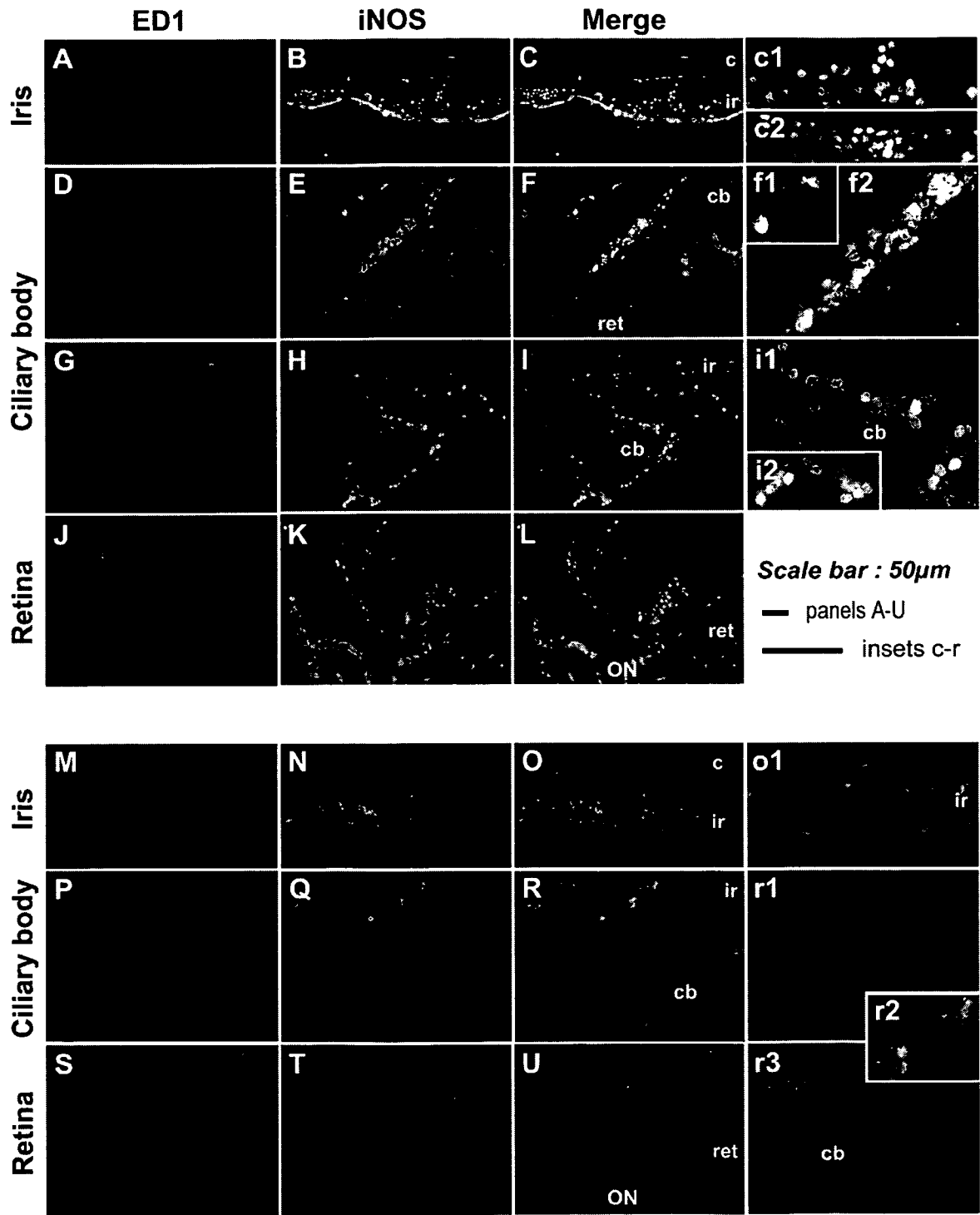


Fig. 6

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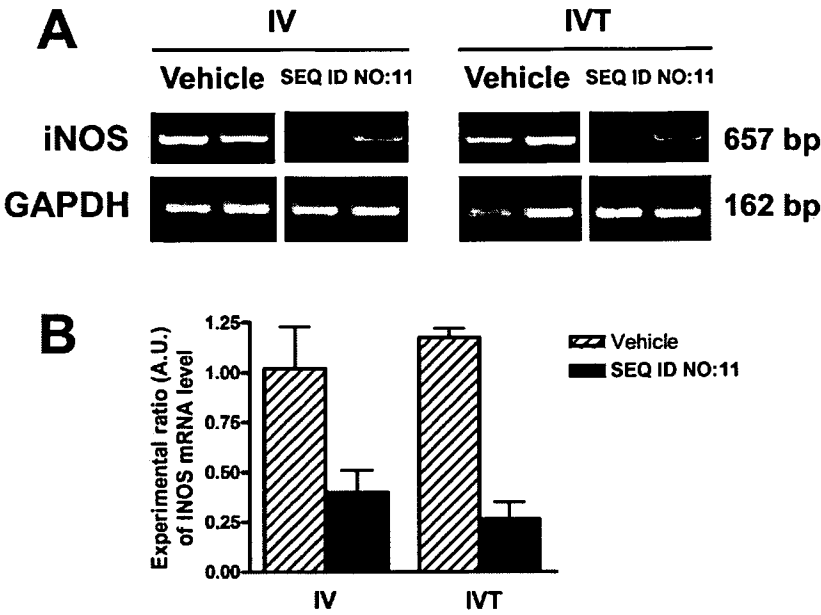


Fig. 7

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A : Chemokines

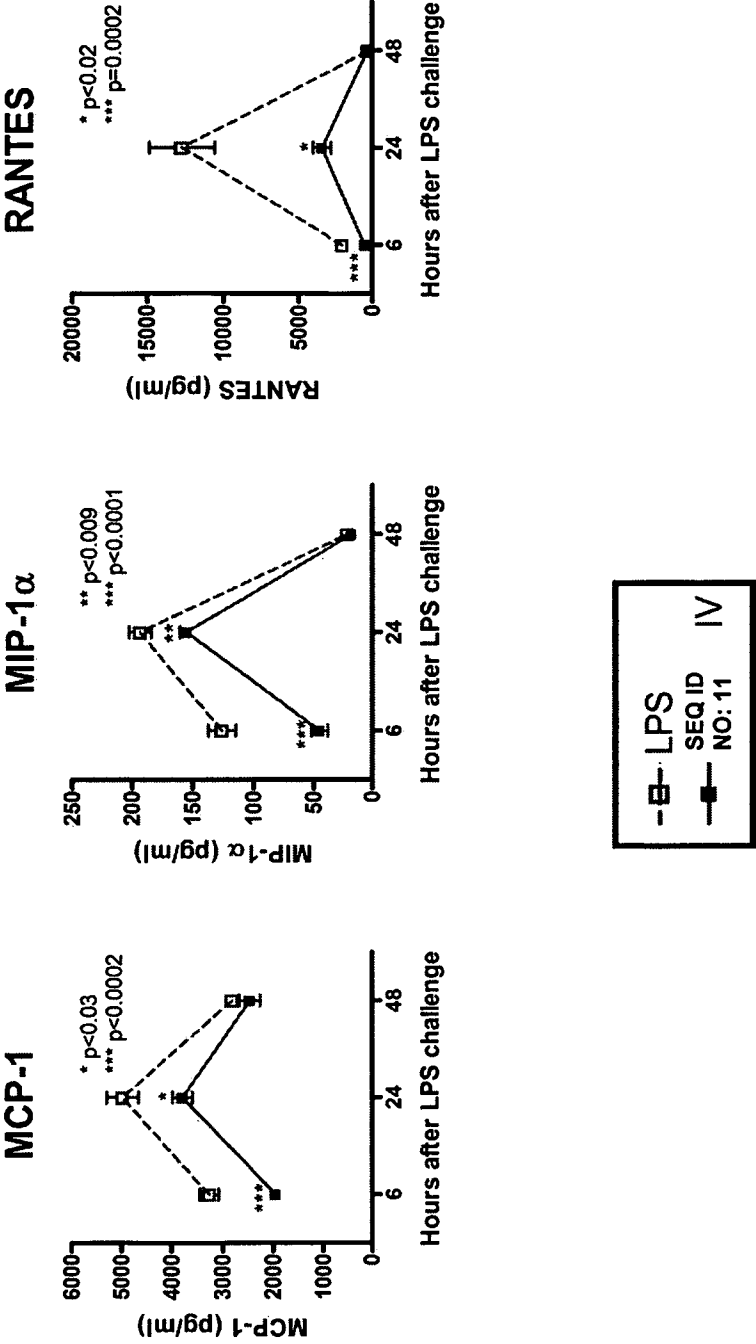


Fig. 8A

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B : Cytokines

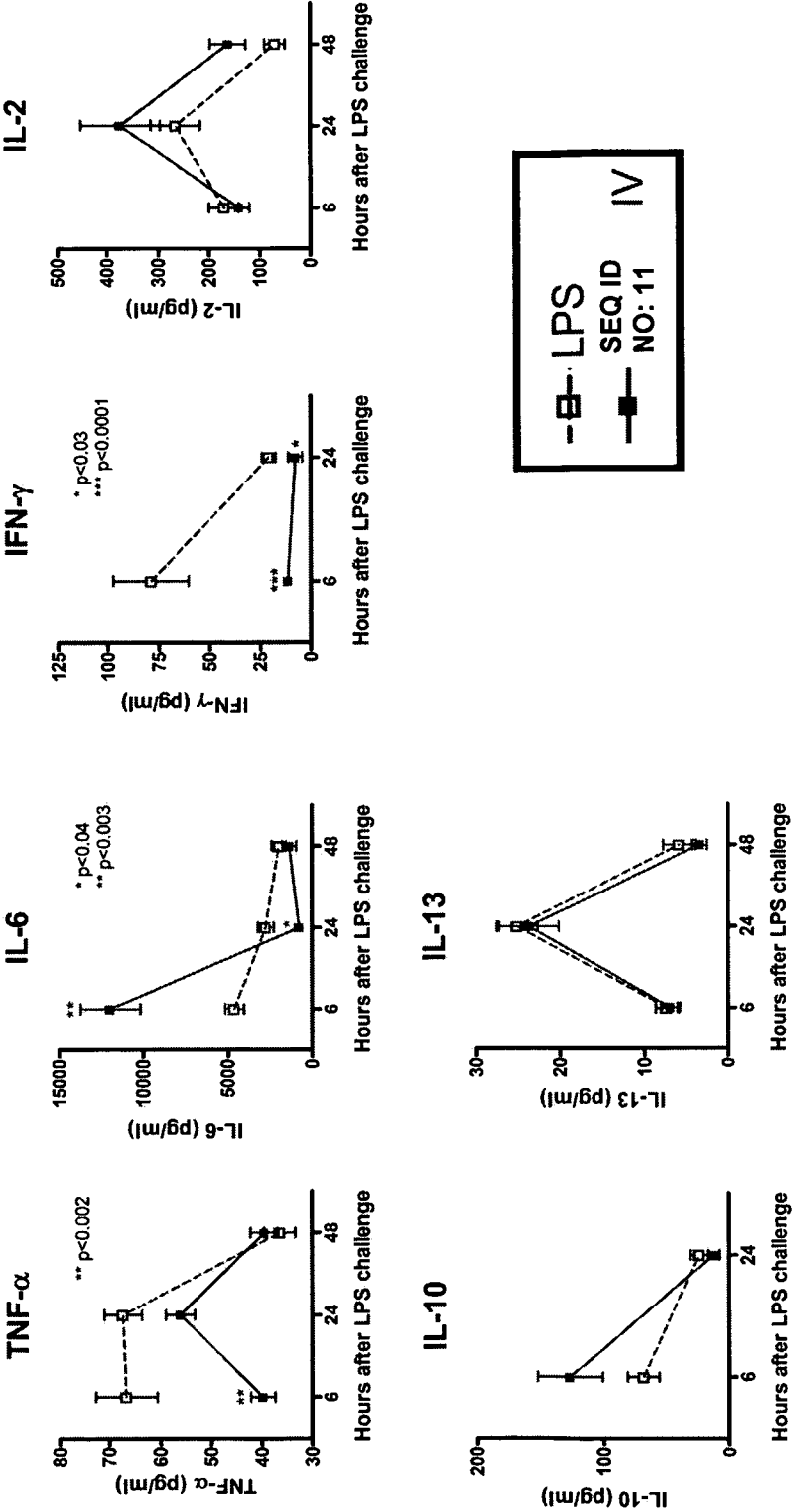


Fig. 8B

10/21

A : Chemokines

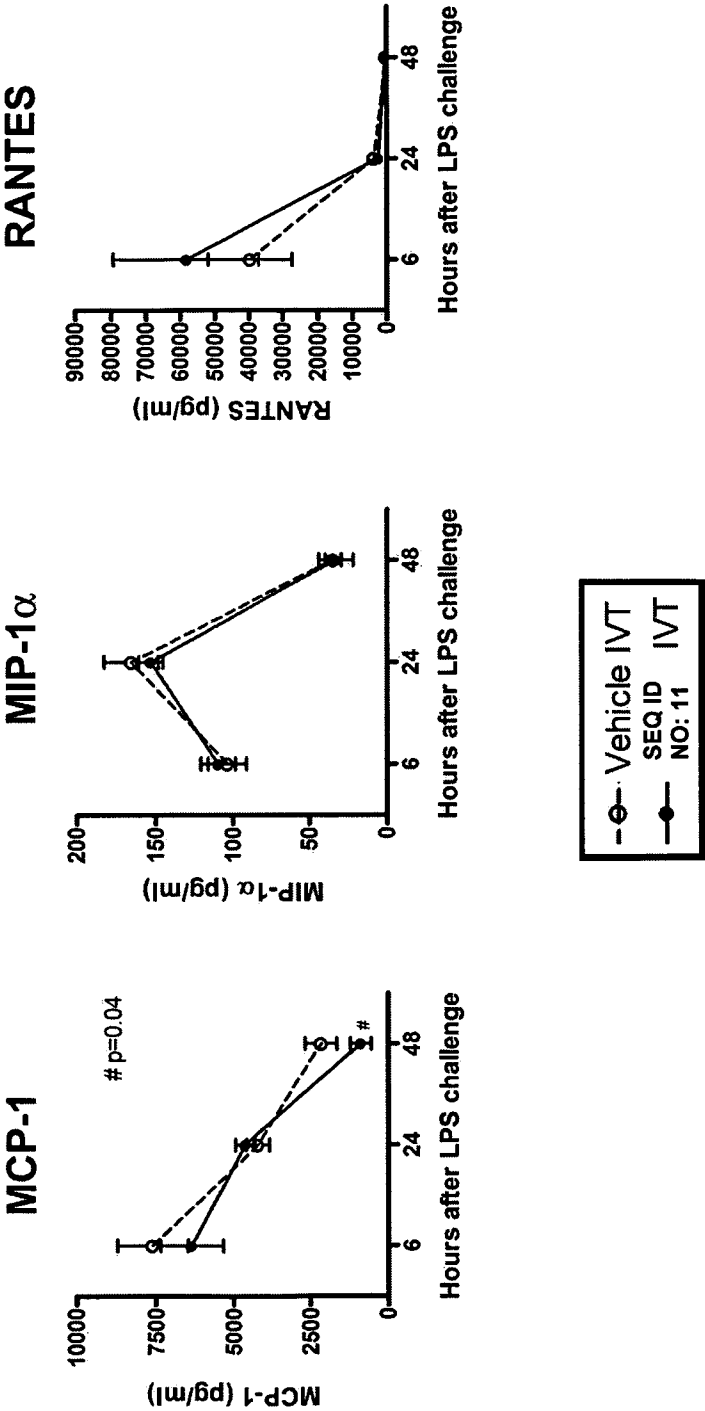


Fig. 9A

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B : Cytokines

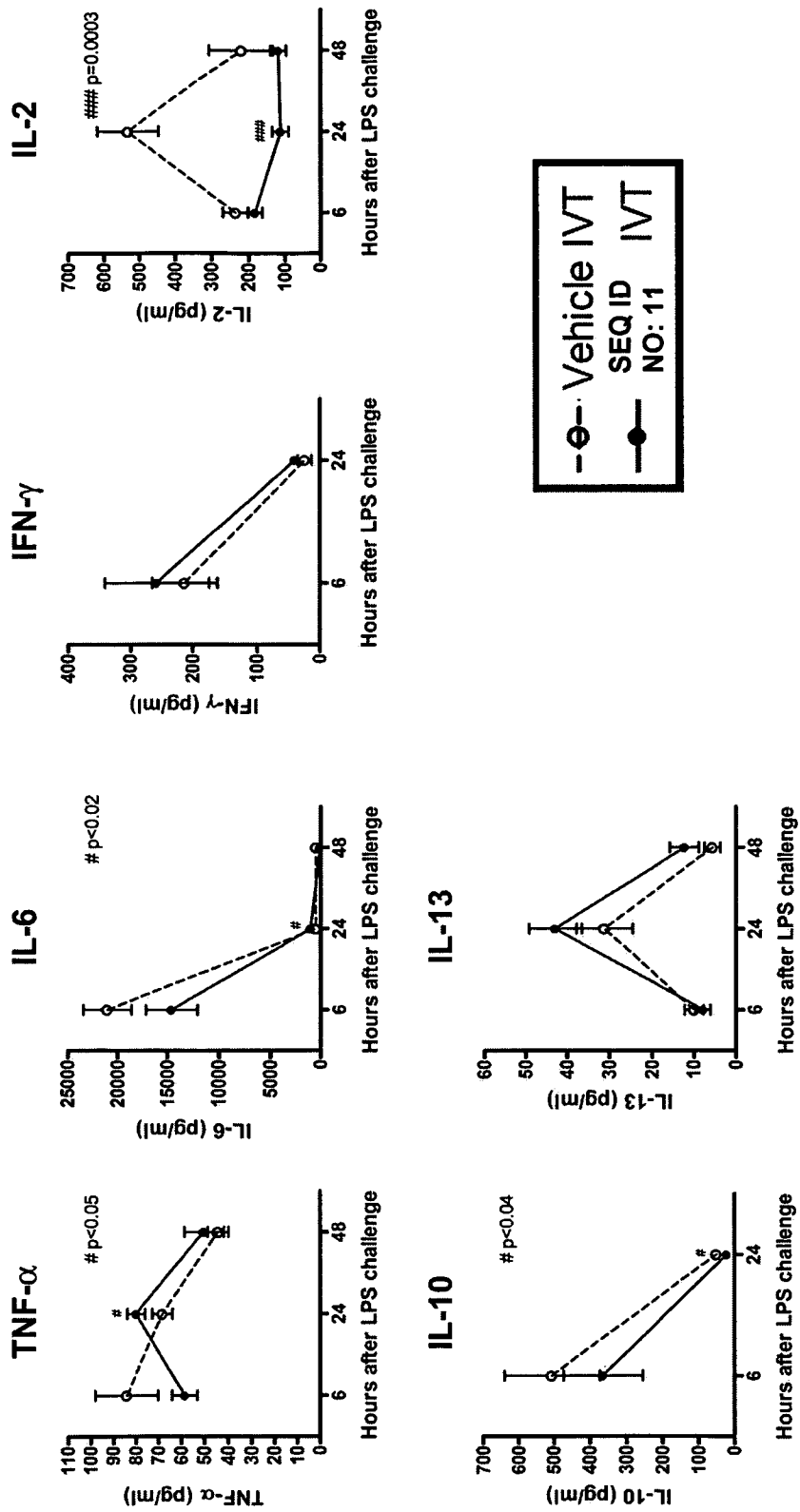


Fig. 9B

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ccactgagag	ccggatgtct	gtcagctcgg	atcctgacct	tgccgcttac	tctgtaactg	1080
cagggcgacc	gcacccttcc	atcagtgaag	aggatgaggg	cttcgactgt	ctgtcatccc	1140
cagagcaagc	tgagccacca	ggtggagggg	ggcggggaag	cctcggggag	ccaccaccgc	1200
ctccacgggc	ctcactgagc	tcggacacca	gcgcaactgt	ctacgactct	gtcaagtaca	1260
cactggtggt	ggatgagcat	gcccagcttg	agttggtgag	cctgcggcca	tgttttggag	1320
attacagtga	cgaaagcgac	tctgccactg	tctatgacaa	ctgtgcctct	gcctcctcgc	1380
cctacgagtc	agccattggt	gaggaatatg	aggaggcccc	tcaaccccgg	cctcccacct	1440
gcctgtcaga	ggactccaca	ccggatgagc	ctgacgtcca	cttctctaag	aagtttctga	1500
atgtcttcat	gagtggccgc	tctcgttcct	ccagtgccga	gtcctttggg	ctgttctcct	1560
gtgtcatcaa	tggggaggag	catgagcaaa	cccatcgggc	tatattcagg	tttgtgcctc	1620
ggcatgaaga	tgaacttgag	ctggaagtgg	acgaccctct	gctggtggag	ctgcaggcag	1680

Fig. 10

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aagactattg gtatgaggcc tataacatgc gcactggagc cegtggtgtc tttcctgcct	1740
actatgccat tgaggtcacc aaggagcctg agcacatggc agcccttgcc aaaaacagcg	1800
actggattga ccagttccgg gtgaagttcc tgggctctgt ccaggttcct tatcacaagg	1860
gcaatgatgt cctctgtgct gctatgcaaa agatcgccac caccgcccgg ctcaccgtgc	1920
actttaaccc gccctccagc tgtgtccttg aaatcagcgt taggggtgtc aagataggtg	1980
tcaaagctga tgaagctcag gaggccaagg gaaataaatg tagccacttt ttccagctaa	2040
aaaacatctc tttctgtggg taccatccaa agaacaacaa gtactttggg tttatcacta	2100
agcacctgc tgaccaccgg tttgcctgcc atgtctttgt gtctgaagat tccaccaaag	2160
ccctggcaga gtctgtgggg cgtgcatttc agcagtctca caagcaattt gtggaatata	2220
cctgtcctac agaagatatc tacttgaggc agcagcaacc cccctctctg cagccccctca	2280
gccccaggcc agtactagga cagctgactg ctgacaggat gttgtactgc cagagagaa	2340
tgggggagtg agggctgttg gggcggggg gcaggggtt ggggagaggc agatgcagtt	2400
tattgtaata tatgggggta gattaatcta tggaggacag tacaggctct ctcggggctg	2460
gggaagggca gggctggggg gggggtcagg catctggcca caaaggggtc ccctagggac	2520
agaggcgctg caccatcctg ggcttgtttc atactagagg ccctggcttt ctggctcttg	2580
ggctctgcct tgacaaagcc cagccacctg gaagtgtcac cttcccttgt ccacctcacc	2640
cagtgccctg agctcatgct gagcccaagc acctccgaag gactttccag taaggaaatg	2700
gcaacatgtg acagtgagac cctgttctca tctgtggggc tccggcagct ccgacccccca	2760
gcctggccag cacgctgacc ctggcaagct tgtgtgttca aagaaggaga gggccacagc	2820
aagccctgcc tgccagggaa ggttccctct cagctggccc cagccaactg gtcactgtct	2880
tgtcacctgg ctactactat taaagtgcc tttcttgtct gaaaaaaaaa aaaaaaaaaa	2940
aaaaaaaaactc gag	2953

Fig. 10 (cont.)

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Met Ala Arg Leu Ser Pro Gly Met Ala Glu Arg Glu Ser Gly Leu Ser
 1 5 10 15
 Gly Gly Ala Ala Ser Pro Pro Ala Ala Ser Pro Phe Leu Gly Leu His
 20 25 30
 Ile Ala Ser Pro Pro Asn Phe Arg Leu Thr His Asp Ile Ser Leu Glu
 35 40 45
 Glu Phe Glu Asp Glu Asp Leu Ser Glu Ile Thr Asp Glu Cys Gly Ile
 50 55 60
 Ser Leu Gln Cys Lys Asp Thr Leu Ser Leu Arg Pro Pro Arg Ala Gly
 65 70 75 80
 Leu Leu Ser Ala Gly Ser Ser Gly Ser Ala Gly Ser Arg Leu Gln Ala
 85 90 95
 Glu Met Leu Gln Met Asp Leu Ile Asp Ala Ala Ser Asp Thr Pro Gly
 100 105 110
 Ala Glu Asp Asp Glu Glu Asp Asp Asp Glu Leu Ala Ala Gln Arg Pro
 115 120 125
 Gly Val Gly Pro Ser Lys Ala Glu Ser Gly Gln Glu Pro Ala Ser Arg
 130 135 140
 Ser Gln Gly Gln Gly Gln Gly Pro Gly Thr Gly Cys Gly Asp Thr Tyr
 145 150 155 160
 Arg Pro Lys Arg Pro Thr Thr Leu Asn Leu Phe Pro Gln Val Pro Arg
 165 170 175
 Ser Gln Asp Thr Leu Asn Asn Asn Ser Leu Gly Lys Lys His Ser Trp
 180 185 190
 Gln Asp Arg Val Ser Arg Ser Ser Ser Pro Leu Lys Thr Gly Glu Gln
 195 200 205
 Thr Pro Pro His Glu His Ile Cys Leu Ser Asp Glu Leu Pro Pro Gln
 210 215 220
 Gly Ser Pro Val Pro Thr Gln Asp Arg Gly Thr Ser Thr Asp Ser Pro
 225 230 235 240
 Cys Arg Arg Thr Ala Ala Thr Gln Met Ala Pro Pro Ser Gly Pro Pro
 245 250 255

Fig. 11

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Ala Thr Ala Pro Gly Gly Arg Gly His Ser His Arg Asp Arg Ser Ile
260 265 270

Ser Ala Asp Val Arg Leu Glu Ala Thr Glu Glu Ile Tyr Leu Thr Pro
275 280 285

Val Gln Arg Pro Pro Asp Pro Ala Glu Pro Thr Ser Thr Phe Leu Pro
290 295 300

Pro Thr Glu Ser Arg Met Ser Val Ser Ser Asp Pro Asp Pro Ala Ala
305 310 315 320

Tyr Ser Val Thr Ala Gly Arg Pro His Pro Ser Ile Ser Glu Glu Asp
325 330 335

Glu Gly Phe Asp Cys Leu Ser Ser Pro Glu Gln Ala Glu Pro Pro Gly
340 345 350

Gly Gly Trp Arg Gly Ser Leu Gly Glu Pro Pro Pro Pro Pro Arg Ala
355 360 365

Ser Leu Ser Ser Asp Thr Ser Ala Leu Ser Tyr Asp Ser Val Lys Tyr
370 375 380

Thr Leu Val Val Asp Glu His Ala Gln Leu Glu Leu Val Ser Leu Arg
385 390 395 400

Pro Cys Phe Gly Asp Tyr Ser Asp Glu Ser Asp Ser Ala Thr Val Tyr
405 410 415

Asp Asn Cys Ala Ser Ala Ser Ser Pro Tyr Glu Ser Ala Ile Gly Glu
420 425 430

Glu Tyr Glu Glu Ala Pro Gln Pro Arg Pro Pro Thr Cys Leu Ser Glu
435 440 445

Asp Ser Thr Pro Asp Glu Pro Asp Val His Phe Ser Lys Lys Phe Leu
450 455 460

Asn Val Phe Met Ser Gly Arg Ser Arg Ser Ser Ser Ala Glu Ser Phe
465 470 475 480

Gly Leu Phe Ser Cys Val Ile Asn Gly Glu Glu His Glu Gln Thr His
485 490 495

Arg Ala Ile Phe Arg Phe Val Pro Arg His Glu Asp Glu Leu Glu Leu
500 505 510

Fig. 11 (cont.)

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Glu Val Asp Asp Pro Leu Leu Val Glu Leu Gln Ala Glu Asp Tyr Trp
 515 520 525

Tyr Glu Ala Tyr Asn Met Arg Thr Gly Ala Arg Gly Val Phe Pro Ala
 530 535 540

Tyr Tyr Ala Ile Glu Val Thr Lys Glu Pro Glu His Met Ala Ala Leu
 545 550 555 560

Ala Lys Asn Ser Asp Trp Ile Asp Gln Phe Arg Val Lys Phe Leu Gly
 565 570 575

Ser Val Gln Val Pro Tyr His Lys Gly Asn Asp Val Leu Cys Ala Ala
 580 585 590

Met Gln Lys Ile Ala Thr Thr Arg Arg Leu Thr Val His Phe Asn Pro
 595 600 605

Pro Ser Ser Cys Val Leu Glu Ile Ser Val Arg Gly Val Lys Ile Gly
 610 615 620

Val Lys Ala Asp Glu Ala Gln Glu Ala Lys Gly Asn Lys Cys Ser His
 625 630 635 640

Phe Phe Gln Leu Lys Asn Ile Ser Phe Cys Gly Tyr His Pro Lys Asn
 645 650 655

Asn Lys Tyr Phe Gly Phe Ile Thr Lys His Pro Ala Asp His Arg Phe
 660 665 670

Ala Cys His Val Phe Val Ser Glu Asp Ser Thr Lys Ala Leu Ala Glu
 675 680 685

Ser Val Gly Arg Ala Phe Gln Gln Phe Tyr Lys Gln Phe Val Glu Tyr
 690 695 700

Thr Cys Pro Thr Glu Asp Ile Tyr Leu Glu
 705 710

Fig. 11 (cont.)

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Met Ala Glu Arg Glu Ser Gly Gly Leu Gly Gly Gly Ala Ala Ser Pro
 1 5 10 15
 Pro Ala Ala Ser Pro Phe Leu Gly Leu His Ile Ala Ser Pro Pro Asn
 20 25 30
 Phe Arg Leu Thr His Asp Ile Ser Leu Glu Glu Phe Glu Asp Glu Asp
 35 40 45
 Leu Ser Glu Ile Thr Asp Glu Cys Gly Ile Ser Leu Gln Cys Lys Asp
 50 55 60
 Thr Leu Ser Leu Arg Pro Pro Arg Ala Gly Leu Leu Ser Ala Gly Gly
 65 70 75 80
 Gly Gly Ala Gly Ser Arg Leu Gln Ala Glu Met Leu Gln Met Asp Leu
 85 90 95
 Ile Asp Ala Thr Gly Asp Thr Pro Gly Ala Glu Asp Asp Glu Glu Asp
 100 105 110
 Asp Asp Glu Glu Arg Ala Ala Arg Arg Pro Gly Ala Gly Pro Pro Lys
 115 120 125
 Ala Glu Ser Gly Gln Glu Pro Ala Ser Arg Gly Gln Gly Gln Ser Gln
 130 135 140
 Gly Gln Ser Gln Gly Pro Gly Ser Gly Asp Thr Tyr Arg Pro Lys Arg
 145 150 155 160
 Pro Thr Thr Leu Asn Leu Phe Pro Gln Val Pro Arg Ser Gln Asp Thr
 165 170 175
 Leu Asn Asn Asn Ser Leu Gly Lys Lys His Ser Trp Gln Asp Arg Val
 180 185 190
 Ser Arg Ser Ser Ser Pro Leu Lys Thr Gly Glu Gln Thr Pro Pro His
 195 200 205
 Glu His Ile Cys Leu Ser Asp Glu Leu Pro Pro Gln Ser Gly Pro Ala
 210 215 220
 Pro Thr Thr Asp Arg Gly Thr Ser Thr Asp Ser Pro Cys Arg Arg Ser
 225 230 235 240
 Thr Ala Thr Gln Met Ala Pro Pro Gly Gly Pro Pro Ala Ala Pro Pro
 245 250 255

Fig. 12

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Gly Gly Arg Gly His Ser His Arg Asp Arg Ile His Tyr Gln Ala Asp
 260 265 270

Val Arg Leu Glu Ala Thr Glu Glu Ile Tyr Leu Thr Pro Val Gln Arg
 275 280 285

Pro Pro Asp Ala Ala Glu Pro Thr Ser Ala Phe Leu Pro Pro Thr Glu
 290 295 300

Ser Arg Met Ser Val Ser Ser Asp Pro Asp Pro Ala Ala Tyr Pro Ser
 305 310 315 320

Thr Ala Gly Arg Pro His Pro Ser Ile Ser Glu Glu Glu Glu Gly Phe
 325 330 335

Asp Cys Leu Ser Ser Pro Glu Arg Ala Glu Pro Pro Gly Gly Gly Trp
 340 345 350

Arg Gly Ser Leu Gly Glu Pro Pro Pro Pro Arg Ala Ser Leu Ser
 355 360 365

Ser Asp Thr Ser Ala Leu Ser Tyr Asp Ser Val Lys Tyr Thr Leu Val
 370 375 380

Val Asp Glu His Ala Gln Leu Glu Leu Val Ser Leu Arg Pro Cys Phe
 385 390 395 400

Gly Asp Tyr Ser Asp Glu Ser Asp Ser Ala Thr Val Tyr Asp Asn Cys
 405 410 415

Ala Ser Val Ser Ser Pro Tyr Glu Ser Ala Ile Gly Glu Glu Tyr Glu
 420 425 430

Glu Ala Pro Arg Pro Gln Pro Pro Ala Cys Leu Ser Glu Asp Ser Thr
 435 440 445

Pro Asp Glu Pro Asp Val His Phe Ser Lys Lys Phe Leu Asn Val Phe
 450 455 460

Met Ser Gly Arg Ser Arg Ser Ser Ser Ala Glu Ser Phe Gly Leu Phe
 465 470 475 480

Ser Cys Ile Ile Asn Gly Glu Glu Gln Glu Gln Thr His Arg Ala Ile
 485 490 495

Phe Arg Phe Val Pro Arg His Glu Asp Glu Leu Glu Leu Glu Val Asp
 500 505 510

Fig. 12 (cont.)

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Asp Pro Leu Leu Val Glu Leu Gln Ala Glu Asp Tyr Trp Tyr Glu Ala
 515 520 525

Tyr Asn Met Arg Thr Gly Ala Arg Gly Val Phe Pro Ala Tyr Tyr Ala
 530 535 540

Ile Glu Val Thr Lys Glu Pro Glu His Met Ala Ala Leu Ala Lys Asn
 545 550 555 560

Ser Asp Trp Val Asp Gln Phe Arg Val Lys Phe Leu Gly Ser Val Gln
 565 570 575

Val Pro Tyr His Lys Gly Asn Asp Val Leu Cys Ala Ala Met Gln Lys
 580 585 590

Ile Ala Thr Thr Arg Arg Leu Thr Val His Phe Asn Pro Pro Ser Ser
 595 600 605

Cys Val Leu Glu Ile Ser Val Arg Gly Val Lys Ile Gly Val Lys Ala
 610 615 620

Asp Asp Ser Gln Glu Ala Lys Gly Asn Lys Cys Ser His Phe Phe Gln
 625 630 635 640

Leu Lys Asn Ile Ser Phe Cys Gly Tyr His Pro Lys Asn Asn Lys Tyr
 645 650 655

Phe Gly Phe Ile Thr Lys His Pro Ala Asp His Arg Phe Ala Cys His
 660 665 670

Val Phe Val Ser Glu Asp Ser Thr Lys Ala Leu Ala Glu Ser Val Gly
 675 680 685

Arg Ala Phe Gln Gln Phe Tyr Lys Gln Phe Val Glu Tyr Thr Cys Pro
 690 695 700

Thr Glu Asp Ile Tyr Leu Glu
 705 710

Fig. 12 (cont.)

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atggcggagc gagaaagcgg cggcctggga gggggggccg cgtccccgcc cgccgcctcc	60
ccgttccttg ggctgcacat cgcttcgcct cccaatttca ggctcaccca tgacatcagc	120
ctggaggagt ttgaggatga agacctctcg gagatcactg atgagtgtgg catcagctta	180
cagtgcaaag acaccctgtc cttacggccc ccgtgcgccg ggctgctctc tgcgggcggc	240
ggcggcgccg ggagccggtt gcaggccgag atgctgcaga tggacctgat cgacgcgacg	300
ggggacactc ccggggccga ggacgacgag gaggacgacg acgaggagcg cgcgccccgg	360
cggccgggag cggggccgcc caaggccgag tccggccagg agccggcgtc ccgcggccag	420
ggccagagcc aaggccagag ccaggggccc ggacgcgggg acacgtaccg gcccaagcgg	480
cccaccacgc tcaacctctt tccgcagggt ccgcggctctc aggacacact gaataataat	540
tctctgggca aaaagcacag ttggcaggat cgggtgtctc gatcatcctc acccctgaag	600
acaggggagc agacaccacc gcatgaacac atctgcctga gcgatgagct gccccccag	660
agcggccccg cccccaccac agatcgaggc acctccaccg acagcccttg ccgccgcagc	720
acagccaccc agatggcacc tccgggtggt cccctgctg ccccgcttg gggtcggggc	780
cactcgcacg gagaccgaat ccactaccag gccgatgtgc gactagaggc cactgaggag	840
atctacctga cccagtgca gaggccccca gacgctgcag agcccacctc cgccttcctg	900
ccgcccactg agagccggat gtcagtcagc tccgatccag accctgccgc ctaccctcc	960
acggcagggc ggccgcaccc ctccatcagt gaagaggaag agggcttcga ctgcctgtcg	1020
tccccagagc gggctgagcc cccaggcgga ggggtggcgg ggagcctggg ggagccgccg	1080
ccacctccac gggcctctct gagctcggac accagcgcgc tgtcctatga ctctgtcaag	1140
tacacgctgg tggtagatga gcatgcacag ctggagctgg tgagcctgcg gccgtgcttc	1200
ggagactaca gtgacgagag tgactctgcc accgtctatg acaactgtgc ctccgtctcc	1260
tcgccctatg agtcggccat cggagaggaa tatgaggagg ccccgcggcc ccagccccct	1320
gcctgcctct ccgaggactc cagcctgat gaacccgacg tccatttctc caagaaattc	1380
ctgaacgtct tcatgagtgg ccgctcccg tcctccagtg ctgagtcctt cgggctgttc	1440
tcctgcatca tcaacgggga ggagcaggag cagaccaccc gggccatatt caggtttgtg	1500
cctcgacacg aagacgaact tgagctggaa gtggatgacc ctctgctagt ggagctccag	1560
gctgaagact actggtacga ggcctacaac atgcgcactg gtgccccggg tgtctttcct	1620
gcctattacg ccatcgaggt caccaaggag cccgagcaca tggcagccct ggccaaaaac	1680
agtgactggg tggaccagtt ccgggtgaag ttctgggct cagtccaggt tccctatcac	1740

Fig. 13

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aagggcaatg acgtcctctg tgctgctatg caaaagattg ccaccacccg ccggctcacc	1800
gtgcacttta acccgccctc cagctgtgtc ctggagatca gcgtgcgggg tgtgaagata	1860
ggcgtcaagg ccgatgactc ccaggaggcc aaggggaata aatgtagcca ctttttccag	1920
ttaaaaaaca tctctttctg cggatatcat ccaaagaaca acaagtactt tgggttcac	1980
accaagcacc ccgccgacca ccggtttgcc tgccacgtct ttgtgtctga agactccacc	2040
aaagccctgg cagagtccgt ggggagagca ttccagcagt tctacaagca gtttgtggag	2100
tacacctgcc ccacagaaga tatctacctg gagtag	2136

Fig. 13 (cont.)

INTERNATIONAL SEARCH REPORT

International application No.

PCT/EP2010/006284

Box No. I Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

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☒

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 together with the international application in electronic form
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International application No
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A. CLASSIFICATION OF SUBJECT MATTER

INV. A61K38/00 A61K38/03 A61K38/04 A61P27/02
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

A61K A61P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, EMBASE

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 2009/143865 A1 (XIGEN S A [CH]; BONNY CHRISTOPHE [CH]) 3 December 2009 (2009-12-03)	1-22, 24-26
Y	abstract page 6, line 8 - line 11 page 22 - page 30 sequences 1-100,102-105 examples 12,13 claims 1-32 ----- -/--	1-26



Further documents are listed in the continuation of Box C.



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INTERNATIONAL SEARCH REPORT

International application No

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C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

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