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(54) Title: METHOD OF DETERMINING OR DETECTING DONOR ORGAN DAMAGE FOLLOWING XENOTRANSPLANTATION BASED ON DONOR ORGAN-DERIVED ANALYTES (57) Abstract <p>A method of determining or detecting the presence of a donor organ-derived analyte in a biological fluid indicative of donor organ damage following xenotransplantation and, thereby, enabling identification of xenograft organ damage in a recipient comprises capturing said analyte by an antibody which is: a) specific for the donor organ-derived analyte; or b) capable of cross-reacting with said donor organ-derived analyte and directly or indirectly determining the donor organ-derived analyte. This method enables one to distinguish between host-derived and donor-derived analytes and, thereby, to obtain an indication of xenotransplant status and possibly rejection, so that corrective therapy can be undertaken as appropriate as soon as such a donor organ-derived analyte is identified. The method applies to <i>in vivo</i> and <i>ex vivo</i> xenotransplantation.</p>		

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Method of determining or detecting donor organ damage following xenotransplantation based on donor-organ derived analytes

Technical Field

5 This invention relates to a method of determining or detecting the presence of a donor organ-derived analyte in a biological fluid indicative of donor organ damage following xenotransplantation.

Background Art

10 Currently there exists a worldwide shortage of donor organs for allotransplantation (e.g., liver, kidney and heart) and it has been proposed and demonstrated that this problem can be overcome albeit temporarily by the use of animal organs prior to orthotopic transplant surgery. Chari, R.S., *et al.* (The New England Journal of Medicine (1994); Vol. 33, No. 4 pp 234-237) treated hepatic failure with *ex vivo* pig-liver perfusion followed by orthotopic liver transplantation. One 15 patient was stabilised for 10 days and subsequently underwent successful orthotopic liver transplantation. However, previous attempts at inter-species transplantation of organs (xenotransplantation) have not been successful because of the phenomenon known as *hyperacute rejection* whereby the newly transplanted organ is overwhelmed and 20 rejected by the host immune system within hours of implantation into the recipient. Recent advances in the area of molecular genetics have allowed the development of transgenic animals (mainly pigs) whose organs are genetically engineered to be less immunogenic when transferred into different species (i.e., human/non-human primate) 25 (Transplant News (1995); Vol 5, No. 9). In this report, it is postulated that immunotolerance is achieved by the expression of a human 'decay accelerating factor' gene in the transgenic organ which acts to prevent host activation of complement components (e.g. C3 complex) and therefore minimise hyperacute rejection. However, as with intra-species 30 organ transplantation (allotransplantation), xenograft rejection is still an issue with which the transplant physician must grapple and is further

complicated by the obvious fact that tissues from different species are present in the graft recipient.

Conventional liver function tests are incapable of differentiating between donor and recipient (e.g., porcine and human) organ-derived analytes. For example, serum alanine aminotransferase (ALT)/aspartate aminotransferase (AST) measurement does not distinguish between the porcine and human enzymes thus rendering futile any attempt to identify the enzyme source post-transplant. A similar situation would pertain in the case of renal xenotransplant analysis. In fact there is currently no generally accepted protein marker of kidney integrity.

Glutathione S-transferases (GSTs) comprise a multigene family of proteins consisting mainly of alpha (α GST), mu (μ GST), pi (π GST) and theta-class (θ GST) isoforms as defined by isoelectric point and are responsible for the detoxification of a range of xenobiotics, mainly *via* conjugation to glutathione (Beckett, G.J and Hayes, J.D. *Advances in Clinical Chemistry* (1993); 30, 281-380). In humans, the uniform hepatic distribution of α GST together with relatively high hepatic levels and a short plasma half-life (approximately 1hr) means that this enzyme is more sensitive than the aminotransferases as an indicator of hepatic status following transplantation and drug-induced liver damage.

EP-A 0 640 145 discloses a method which assists in the early diagnosis of rejection in a liver transplant recipient and which comprises measuring an increase in plasma or serum α GST from the recipient in the absence of or preceding any change in plasma or serum transaminase.

Pi Glutathione S-transferase is located in the cytoplasm of bile duct epithelial cells within the liver and is thought to exist only in the homodimeric form. This heterogenous intrahepatic GST distribution suggests that the different isoenzymes have unique *in vivo* functions in different hepatic regions and it can therefore be concluded that measurement of plasma or biliary GST levels will facilitate the monitoring of the hepatic status of an individual. A similar unique

distribution also exists with respect to renal tissues where α GST is located in the proximal tubule region and π GST is confined to the distal tubule region of the nephron (Campbell, J.A.H. *et al.*, Cancer (Philadelphia) (1991); 67, 1608-1613). In a recent report it was
5 proposed that simultaneous detection of α/π GST in human urine could be used to distinguish between cyclosporin A nephrotoxicity and graft rejection respectively, due to the site-specific nature of the respective tissue insult (Sundberg, A.G.M. *et al.*, Nephron, (1994): 67, 308-316).

Accordingly, there is a need for a method of discriminating
10 between host and graft-derived analytes as a means for detecting graft organ damage or, indeed, to distinguish between host and/or graft organ damage.

Disclosure of Invention

The invention provides a method of determining or detecting the
15 presence of a donor organ-derived analyte in a biological fluid indicative of donor organ damage following xenotransplantation and, thereby, enabling identification of xenograft organ damage in said recipient, which method comprises capturing said analyte by an antibody which is:

- 20 a) specific for the donor organ-derived analyte; or
- b) capable of cross-reacting with said donor organ-derived analyte

and directly or indirectly determining the donor organ-derived analyte.

25 Accordingly, the method according to the invention enables one to distinguish between host-derived and donor-derived analytes and, thereby, to obtain an indication of xenotransplant status and possibly rejection, so that corrective therapy can be undertaken as appropriate as soon as such a donor organ-derived analyte is identified in a biological

fluid of the recipient as will be demonstrated in greater detail hereinbelow.

5 By biological fluid herein is meant for example body fluids such as bile, plasma, serum and urine as well as tissue support media and perfusates. The biological fluids herein are also referred to generally as matrices.

By donor organ-derived herein is meant organs *per se* and tissue and cells which are constituent parts thereof or derived therefrom.

10 Likewise by donor material herein is meant organs *per se* and tissue and cells which are constituent parts thereof or derived therefrom.

Thus, by way of example, the donor material can be a liver, a part of a liver such as a lobe or hepatocytes. Typically in the case of hepatocytes, an hepatocyte cartridge would be used.

15 Generally, the recipient will be a human being or a non-human primate.

Accordingly, the donor material will preferably be derived from a non-human primate or an animal such as the pig which is physiologically and biochemically similar to the human being.

20 The donor animal can be a transgenic animal.

The xenotransplantation can be *in vivo* or *ex vivo*

The organ which is xenotransplanted is suitably a heart, kidney or liver.

25 Preferably, the donor organ-derived analyte is a protein, more especially an enzyme.

Preferably, the donor organ-derived analyte will be donor specific. In the event that there is a corresponding recipient analyte, preferably the recipient analyte has a low degree of homology with said donor organ-derived analyte. However, the donor organ-derived analyte can have a high degree of homology (for example, greater than 80% homology) with the corresponding recipient analyte and yet is detectable in accordance with the invention as hereinafter described.

A preferred enzyme is a glutathione S-transferase, more especially α GST.

Further, preferably, determination of the captured donor organ-derived analyte is carried out by an immunoassay format. Suitable immunoassay formats include enzyme immunoassays.

Thus, in the case of porcine α GST in a human biological fluid, the porcine α GST can be captured by IgG [anti-porcine α GST] directly or indirectly bound to a solid phase or, alternatively, by IgG [anti-human α GST] exhibiting cross-reactivity.

We have raised rabbit polyclonal IgG [anti-porcine α GST] which is highly specific for porcine α GST. In the event that the antibody also displays a small degree (~5-10%) cross-reactivity with human α GST, such as in the presence of a high concentration of human α GST, then the human α GST can be separately determined and the amount of porcine α GST correctly calculated as hereinafter described.

We have identified a monoclonal anti-human α GST with almost 100% cross-reactivity with porcine α GST as hereinafter described in Example 3.

The enzyme immunoassay can be a sandwich format or a semi-competitive enzyme immunoassay as hereinafter described in Examples 1 and 2, respectively.

In a further aspect of the invention, a donor material is tested for viability prior to xenotransplantation.

Thus, the invention also enables one to test the viability of donor material by measuring a donor organ-derived analyte in a biological fluid which perfuses the donor material by adopting a method as hereinbefore defined.

In this aspect of the invention one can detect an analyte in accordance with any known method in addition to the methods according to the invention. For example, if the organ-derived analyte is an enzyme, then an enzyme assay based on the use of a substrate for the enzyme can be used to detect said organ-derived analyte.

The invention also provides a test kit or pack containing one or more components for carrying out a method according to the invention as hereinabove described.

15 Brief Description of Drawings

In the accompanying drawings:

Fig. 1 is a schematic diagram of the sandwich enzyme immunoassay of Example 1;

20 Fig. 2 is a plot of absorbance at 450/630 nm *versus* α GST concentration ($\mu\text{g/L}$) according to the sandwich immunometric enzyme immunoassay for porcine α GST of Example 1;

Fig. 3 is a schematic diagram of the semi-competitive enzyme immunoassay of Example 2;

25 Fig. 4 is a plot of absorbance at 450/630 nm *versus* α GST concentration ($\mu\text{g/L}$) according to the competitive immunometric enzyme immunoassay for porcine α GST of Example 2;

Fig. 5 is an SDS-PAGE analysis of human and porcine α GST;

Fig. 6 is an immunoblot analysis of human and porcine α GST using IgG [anti-human α GST] and goat IgG [anti-rabbit IgG]-HRP conjugate;

5 Fig. 7 is an immunoblot analysis of human and porcine α GST using IgG [anti-porcine α GST] and goat IgG [anti-rabbit IgG]-HRP conjugate; and

10 Fig. 8 is an immunoblot analysis of human and porcine α GST using IgG [anti-human α GST] and goat IgG [anti-mouse IgG]-HRP conjugate.

Modes for Carrying Out the Invention

The invention will be further illustrated by the following Examples.

Preparatory Example A

15 Purification of porcine α GST

Alpha GST was purified from porcine liver by affinity chromatography and chromatofocussing (pH 9.5 - 6.0). Precise details of the purification procedure are as follows:

20 a. 30g of porcine liver was homogenised for 2 minutes in homogenisation buffer, at a ratio of one part liver to three parts buffer, using a Waring (Waring is a Trade Mark) blender. The homogenisation buffer had the following composition:

5
10mM Tris-HCl
250mM Sucrose
5mM EDTA pH 7.8
2µg/ml Leupeptin
2µg/ml Pepstatin.

b. The liver homogenate was centrifuged at 10000g for 60 minutes.

c. The supernatant was then loaded on a Glutathione(GSH)-
10 Sepharose Affinity column previously equilibrated in 10mM Tris
pH9.1. Equilibration buffer was reapplied to elute unbound protein.
Finally 50mM Tris pH9.1 containing 5mM GSH was used to elute
bound GST from the affinity column.

d. The eluted material was then dialysed against 25mM
15 ethanolamine pH9.5 and applied to a chromatofocussing column
(PBE94 supplied by Pharmacia). Elution buffer was Polybuffer 96
(Polybuffer 96 is a Trade Mark) prepared according to manufacturer's
instructions.

Preparatory Example B

Antibody Production and Purification:

20 Purified porcine α GST was injected into New Zealand White
rabbits subcutaneously (s.c.) according to the time schedule given below
and serum evaluated for anti- α GST reactivity. Once the IgG [anti-
porcine α GST] titre was sufficient as determined by semi-quantitative
dot blot analysis, the animals were exsanguinated and serum collected.
25 Total IgG was purified from rabbit serum by Protein A affinity
chromatography and was used for both microtitre plate coating (semi-
competitive EIA - Example 2) and biotinylation (sandwich EIA -
Example 1). Monoclonal IgG [anti-human α GST], as ascites, was
obtained from The University Hospital, Nijmegen, The Netherlands and
30 was not purified further prior to use.

Immunisation Schedule (general):

5 Day 1: A test bleed of 5ml preserum from the ear of the rabbit was carried out and then 0.5ml of porcine α GST antigen (100 μ g) was mixed with an equal volume of Freund's Complete Adjuvant. The antigen and adjuvant were homogenised to ensure good emulsion. This mixture was then injected s.c. into multiple sites on the back of the rabbit which had previously been shaved.

10 Day 28: A test bleed of 5ml serum from the ear of the rabbit was carried out and then 0.5ml antigen (100 μ g) was mixed with an equal volume of Freund's Complete Adjuvant. The antigen and adjuvant were homogenised to ensure good emulsion. This mixture was then injected s.c. into multiple sites on the back of the rabbit.

Day 42: A test bleed of 10ml blood was taken from the rabbit's ear.

15 Day 56: A second boost was given to the rabbit as described on Day 28.

Day 70: A test bleed of 10ml of blood was taken from the ear of the rabbit. When the titre was sufficiently high, the rabbit was sacrificed and as much blood as possible was collected

20

Preparatory Example C

Immunoblotting

25 All polyclonal and monoclonal IgGs for use in the following Examples were checked for reactivity and cross - reactivity against both human and porcine α GST, respectively *via* the following immunoblot combinations-

- (a) Rabbit IgG [anti-human α GST] was used to probe nitrocellulose membranes containing immobilised human and porcine α GST.
- 5 (b) Rabbit IgG [anti-porcine α GST] was used to probe nitrocellulose membranes containing immobilised human and porcine α GST.
- (c) Murine IgG [anti-human α GST] was used to probe nitrocellulose membranes containing immobilised human and porcine α GST.
- 10 The method used for Immunoblot detection was as follows:
1. Both human and porcine α GST (0.5 μ g/track) were electrophoresed on 15% SDS-PAGE and molecular weight markers were included.
 - 15 2. After electrophoresis, the polyacrylamide gel was cut and one half stained for protein while the remainder was used for electrophoretic transfer onto nitrocellulose.
 - 20 3. After electrophoretic transfer, the nitrocellulose membranes were blocked for 1 hour with 5%(w/v) Marvel (Marvel is a Trade Mark) in phosphate buffered saline containing 0.05%(w/v) TWEEN-20 (PBST)- blocking buffer.
 - 25 4. The following solutions were then prepared:
 - (i) Rabbit IgG [anti-human α GST] in 1%(w/v) Marvel in PBST
 - (ii) Rabbit IgG [anti-porcine α GST] in 1%(w/v) Marvel in PBST
 - (iii) Murine IgG [anti-human α GST] in 1%(w/v) Marvel in PBST

and added to the membranes, once blocking buffer was decanted. Incubation with antibody solutions was allowed to proceed for one hour.

5 5. The nitrocellulose membranes were then washed in PBST (2x for 5 min. each).

10 6. Anti-rabbit IgG - HRP conjugate was then prepared (1/1000 in 1% (w/v) Marvel in PBST and added to 4(i) and (ii) above. Anti-murine IgG -HRP conjugate was also prepared (1/1000) and added to 4(iii) above. The anti-species (rabbit/mouse) conjugates used for the immunodetection were obtained from Bio-Rad Laboratories Limited.

 7. After one hour incubation with anti - species conjugates, the reagents were discarded and the membranes washed as in 5 above.

15 8. Diaminobenzidine substrate was then prepared and added to the membrane. A positive reaction was indicated by a brown precipitate on the nitrocellulose membrane.

Preparatory Example D

Porcine α GST-horseradish peroxidase (HRP) conjugate synthesis:

20 α GST-HRP conjugates were synthesised using thioether conjugation methodology. Reactive maleimide groups were introduced onto α GST molecules using SMCC (succinimidyl 4-(N-maleimidomethyl) cyclohexane 1-carboxylate) and masked sulphhydryl groups were linked to HRP (Duncan, R.J.S., *et al* (1983); Anal. Biochem. 132, 68-73). After a demasking step to produce reactive sulphhydryl groups, the maleimide-activated α GST and HRP-SH were
25 mixed together and allowed to react for 4.5 hours. The resultant α GST-HRP conjugate, formed by covalent thioether linkage, was brought to 50% (v/v) glycerol and stored at -20°C for use in the semi-competitive EIA of Example 2.

Preparatory Example E

Biotinylation of IgG [anti-porcine α GST]

5 Biotinylated IgG [anti-porcine α GST] was prepared by coupling purified polyclonal IgG[anti-porcine α GST], previously generated by immunisation of rabbits with purified α GST (as described in Preparatory Example B), to N-hydroxysuccinimide-Biotin (NHS-Biotin) under alkaline conditions. Unreacted NHS-Biotin was then removed by extensive dialysis and biotinylated IgG aliquotted and stored frozen at -20°C until required.

10

Example 1

Sandwich enzyme immunoassay

The procedure used is depicted schematically in Fig. 1.

15 a. A Nunc Maxisorp (Nunc Maxisorp is a Trade Mark) microtitre plate was coated with murine monoclonal IgG [anti-human α GST] (referred to in Preparatory Example B) immobilised *via* goat F(ab)₂ fragments [anti-mouse IgG]. This method of antibody coating serves to orientate Mab binding sites and also improves assay sensitivity by minimising adherence - induced denaturation of the capture antibody. Microtitre plates were blocked with a protein/sucrose
20 solution.

b. Porcine α GST, purified from liver as described in Preparatory Example A obtained immediately after expiration and stored at $2-8^{\circ}\text{C}$ or -20°C , was used as the assay calibrator.

25 c. Biotinylated IgG[anti-porcine α GST] conjugates were used to facilitate detection of captured/immobilised α GST.

d. A Streptavidin - horseradish peroxidase (HRP) conjugate was then used to detect the entire antigen sandwich complex in association with the use of tetramethylbenzidine substrate (TMB).

- 5 e. The enzyme reaction was stopped by the addition of 1N H_2SO_4 and the absorbance measured at 450nm using 630nm as a reference wavelength. Colour intensity is proportional to αGST concentration and after generating a plot of $A_{450/630\text{nm}}$ versus concentration ($\mu\text{g/L}$), the concentration of unknown samples can be determined (see Fig. 2). Total assay time was less than 3.5 hours.

Example 2

Semi-Competitive Enzyme Immunoassay

The procedure used is depicted schematically in Fig. 3.

- 10 a. In this case, Nunc Maxisorp plates were coated with polyclonal IgG [anti-porcine αGST] immobilised *via* protein A since it was found that direct coating of polyclonal IgG did not facilitate αGST capture, possibly due to IgG denaturation upon binding to the microtitre plate.
- 15 b. αGST , labelled with HRP was used as the conjugate in this case and was added to the microwell subsequently to the unlabelled calibrator/sample. In this way a competition-type reaction was set up between the HRP-labelled and unlabelled αGST .
- 20 c. After a suitable incubation period, normally 60 minutes, the microtitre plate was washed and TMB substrate was added.
- 25 d. The enzyme reaction was stopped by the addition of 1N H_2SO_4 and the absorbance measured at 450nm using 630nm as a reference wavelength. Colour intensity is inversely proportional to porcine αGST concentration and after plotting $A_{450/630\text{nm}}$ versus concentration ($\mu\text{g/L}$), quantitation of unknown samples is achieved (see Fig. 4).

Example 3Assessment of purity of immunoassay reagents

5 Immunoblotting was carried out in accordance with the procedure described in Preparatory Example C. The results are depicted in Figs. 5-8.

Key to tracks in Figs. 5-8:

Track 1: Molecular weight markers.

Track 2: Human α GST (0.5 μ g).

Track 3: Porcine α GST (0.5 μ g).

10 Fig. 5 illustrates the purity (indicated by the single band in each case) of both human and porcine α GST prior to immunisation into rabbits and confirms the absence of any other human or porcine-derived proteins which might otherwise contribute to reduced assay specificity. Immunoblot analysis of the antibody reactivity reveals that
15 the IgG[anti-human α GST] is highly specific for human α GST and does not exhibit any significant cross-reactivity with porcine α GST (Fig. 6), as indicated by the strong intensity of the human α GST signal relative to the weak intensity of the porcine α GST signal. A finding supported by the lack of porcine α GST (α GST)_p reactivity in a human α GST
20 (α GST)_h-specific enzyme immunoassay marketed by Biotrin International Limited - Mount Merrion, County Dublin, Ireland, under the name HEPKIT. The results are shown in Table 1 which represent an evaluation of α GST_p reactivity in the Biotrin HEPKIT. It is clear that no cross-reactivity occurs when α GST_p is assayed in the HEPKIT.

Table 1

	[α GST]h (μ g/L)	A _{450/630nm}
5		
	0.00	0.069
	1.25	0.156
	2.50	0.332
10	5.00	0.627
	10.0	1.088
	20.0	1.495
	40.0	1.762
15	PC	0.839 (6.9 μ g/L.)
	[α GST]p (μ g/L)	A _{450/630nm}
20	8	0.012
	40	0.010
	200	0.012
	1000	0.014

25 PC = Positive Control.

30 The significance of this fact is of the utmost importance since it implies that the enzyme immunoassay for human α GST quantitation is specific for the detection of human α GST. Thus, any human α GST present in samples for which porcine α GST will also be measured, can be specifically detected without cross-contamination from the porcine antigen. This lack of cross-reactivity in the human α GST enzyme immunoassay and the finding that about 5-10% cross-reactivity is evident between IgG[anti-porcine α GST] and human α GST (Fig. 7), as indicated by the weak intensity of the human α GST signal relative to the strong intensity of the porcine α GST signal, means that

35

simultaneous quantitation of human α GST in the HEPKIT enzyme immunoassay and porcine α GST in the enzyme immunoassay for porcine α GST can be used to correct for any human α GST contribution in the porcine α GST immunoassay.

5 For example, if a sample gives the following readings:

Assay	α GST (μ g/L)
Porcine EIA (porcine α GST)	10000
Hepkit EIA (human α GST)	1000

10 then assuming approximately 7% cross-reactivity of human α GST in porcine EIA implies that human α GST contributes $\frac{1000}{100} \times 7 = 70 \mu\text{g/L}$

to the porcine α GST reading. Therefore, $10000 - 70$

= $9930 \mu\text{g/L}$ porcine α GST present

15 Furthermore as mentioned above, the extensive cross-reactivity between murine IgG [anti-human α GST] and porcine α GST is clearly illustrated in Fig. 8, as indicated by the comparable intensity of the respective bands/signals. This phenomenon is taken advantage of by using this antibody in the sandwich immunoassay of Example 1 for porcine α GST as the 'capture' or plate-coating antibody.

20

Example 4

Assay specificity

25 As the assays of Examples 1 and 2 will primarily be used to detect porcine α GST in non-porcine biological fluids it is essential that assay sensitivity and specificity be evaluated using porcine α GST present in the following matrices:-

- Human urine
- Human serum
- Human plasma
- Human bile
- 5 - Tissue support media

To achieve this analysis, porcine α GST was 'spiked' into all of the above matrices and subsequently assayed in the immunoassays of Examples 1 and 2. In addition to quantitative α GST detection, assay
10 linearity (parallelism) was also evaluated as was assay sensitivity. Furthermore, it is also necessary that assay specificity facilitates detection of porcine α GST only and that any human α GST does not cause significant interference in the immunoassay. To this end potential cross-reactivity of human α GST was investigated by adulterating
15 porcine α GST containing samples with different amounts of human α GST to evaluate potential cross-reactivity. Given that such a significant sequence homology exists between both isoenzymes this is far from a trivial problem and can only be overcome by extensive analysis of antisera [anti-porcine α GST] for potential cross-reactivity
20 with human α GST.

The results of the quantitation of porcine α GST in various sample matrices as determined by the sandwich and semi-competitive enzyme immunoassays are shown in Table 2.

Table 2

**RECOVERY OF SPIKED SAMPLES IN SANDWICH AND
SEMI-COMPETITIVE PORCINE α GST ASSAYS:**

Expected Value (μ g/L)	Sample Diluent (μ g/L)		Plasma-human (μ g/L)		MEM (complete) (μ g/L)		TC-100 (complete) (μ g/L)		Urine-human (μ g/L)	
	sand.	comp.	sand.	comp.	sand.	comp.	sand.	comp.	sand.	comp.
0	61.4	19.4	0	-	0	46.8	0	73	0	84
4000	4530	3853	4299	over	4135	3643	4754	3889	4189.5	3936
2000	2980	2699	2500	over	2262	2420	2059	2071	2165.2	1912
1000	-	-	1041	over	1180	1247	983	953	844.5	857
500	507	446	249	over	512	485	540	562	392.8	454
250	279	260	85	over	290	211	273	266	151.9	218
125	128	140	0	over	136	188	197	163	17.9	126

Key:

1. Sand: Sandwich enzyme immunoassay.
2. Comp: Semi-competitive enzyme immunoassay.
3. Samples diluted 1/5 for Competitive Assay range (0 - 1000 μ g/L) then further diluted to 1/100 for Sandwich Assay (range 0-40 μ g/L).
4. MEM and TC-100: Tissue support media.

It is clear from Table 2 that quantitative recovery of porcine α GST is achievable in all matrices tested. Obviously it is essential that α GST is detectable in both human plasma and urine, respectively so as to facilitate xenograft liver and kidney status, and this is, indeed, the case. However, it is noticeable that the plasma matrix appears to be incompatible with the semi- competitive immunoassay format, possibly due to IgG interaction with immobilised Protein A resulting in anomalous readings. Apart from this all other fluids appear to be compatible with both assay formats. Of particular interest is the ability to detect porcine α GST in tissue support media which should facilitate the possibility of pre-transplant xenograft evaluation or isolated porcine hepatocyte viability analysis (present in cartridges), as these systems are generally maintained in conventional support media. Additionally, this matrix/sample compatibility should greatly facilitate the detection of α GST in the media used for the maintenance of *ex vivo* donor organs whereby the donor organ (e.g., liver) is maintained extra-corporeally in contact with culture medium.

Example 5

Evaluation of human α GST interference in the porcine α GST sandwich enzyme immunoassay

Evaluation of human α GST interference in the porcine α GST sandwich enzyme immunoassay of Example 1 was carried out. No assay interference, as measured by increased absorbance values, was evident at microwell concentrations less than 25 μ g/L (equivalent to 125 μ g/L sample concentration at 1/5 dilution in sample diluent). The results are shown in Table 3.

Table 3

[α GST]p (μ g/L)	[α GST]h	Absorbance _{450/630nm}
20	-	0.869
20	6	0.875
20	12	0.875
20	25	0.881
20	50	0.975

Key:

α GSTp - porcine α GST
 α GSTh - human α GST

5 The results shown in Table 3 demonstrate the specificity of the
immunoassays for the detection of porcine α GST. It is evident from
this data that human α GST does not interfere in the enzyme
immunoassay even at sample concentrations as high as 125 μ g/L
(25 μ g/L x 5) which represents a value of 15 and 6 times the upper limit
10 of normal for human serum and urinary α GST, respectively. At higher
levels of human α GST (>250 μ g/L; 50 μ g/L x 5) it is apparent that some
interference is occurring and it has been calculated that there is about
5-10% cross-reactivity between human α GST and IgG [anti-porcine
 α GST]. However, co-measurement of α GST levels in quantitative
15 immunoassays for both human and porcine α GST should allow the
precise amount of human α GST present to be calculated and thus taken
into account during porcine α GST quantitation in human biological
fluids.

20 It should be further noted that the immunoassays according to
invention can also be used for the detection of porcine GST in porcine
derived biological fluids as would be the case for *in vitro* or *in vivo*
toxicological studies involving pigs or pig organs. This finding is of
significance since it is known that porcine and human
physiology/biochemistry are quite similar and consequently pigs are
25 often used as animal models to predict drug toxicity in humans.

Claims:-

1. A method of determining or detecting the presence of a donor organ-derived analyte in a biological fluid indicative of donor organ damage following xenotransplantation and, thereby, enabling
5 identification of xenograft organ damage in a recipient, which method comprises capturing said analyte by an antibody which is:
- a) specific for the donor organ-derived analyte; or
 - b) capable of cross-reacting with said donor organ-derived
10 analyte
- and directly or indirectly determining the donor organ-derived
10 analyte.
2. A method according to Claim 1, wherein the recipient is a human being.
3. A method according to Claim 1, wherein the recipient is a
15 non-human primate.
4. A method according to any one of Claims 1-3, wherein a donor organ is derived from a pig.
5. A method according to any one of Claims 1-4, wherein the organ is a kidney.
- 20 6. A method according to any one of Claims 1-4, wherein the organ is a liver.
7. A method according to any preceding claim, wherein the donor organ-derived analyte is a protein.
- 25 8. A method according to Claim 7, wherein the donor organ-derived analyte is an enzyme.

9. A method according to Claim 8, wherein the enzyme is a glutathione S-transferase (GST).
10. A method according to Claim 9, wherein the enzyme is an alpha GST.
- 5 11. A method according to any preceding claim, when dependent on Claim 4, wherein the antibody is anti-porcine α GST.
12. A method according to any one of Claims 1-10 when dependent on Claim 2, wherein the antibody is anti-human α GST which exhibits substantially 100% cross-reactivity with porcine α GST.
- 10 13. A method according to any preceding claim, wherein a donor material is tested for viability prior to xenotransplantation.
14. A test kit or pack containing one or more components for carrying out a method according to any one of Claims 1-13.

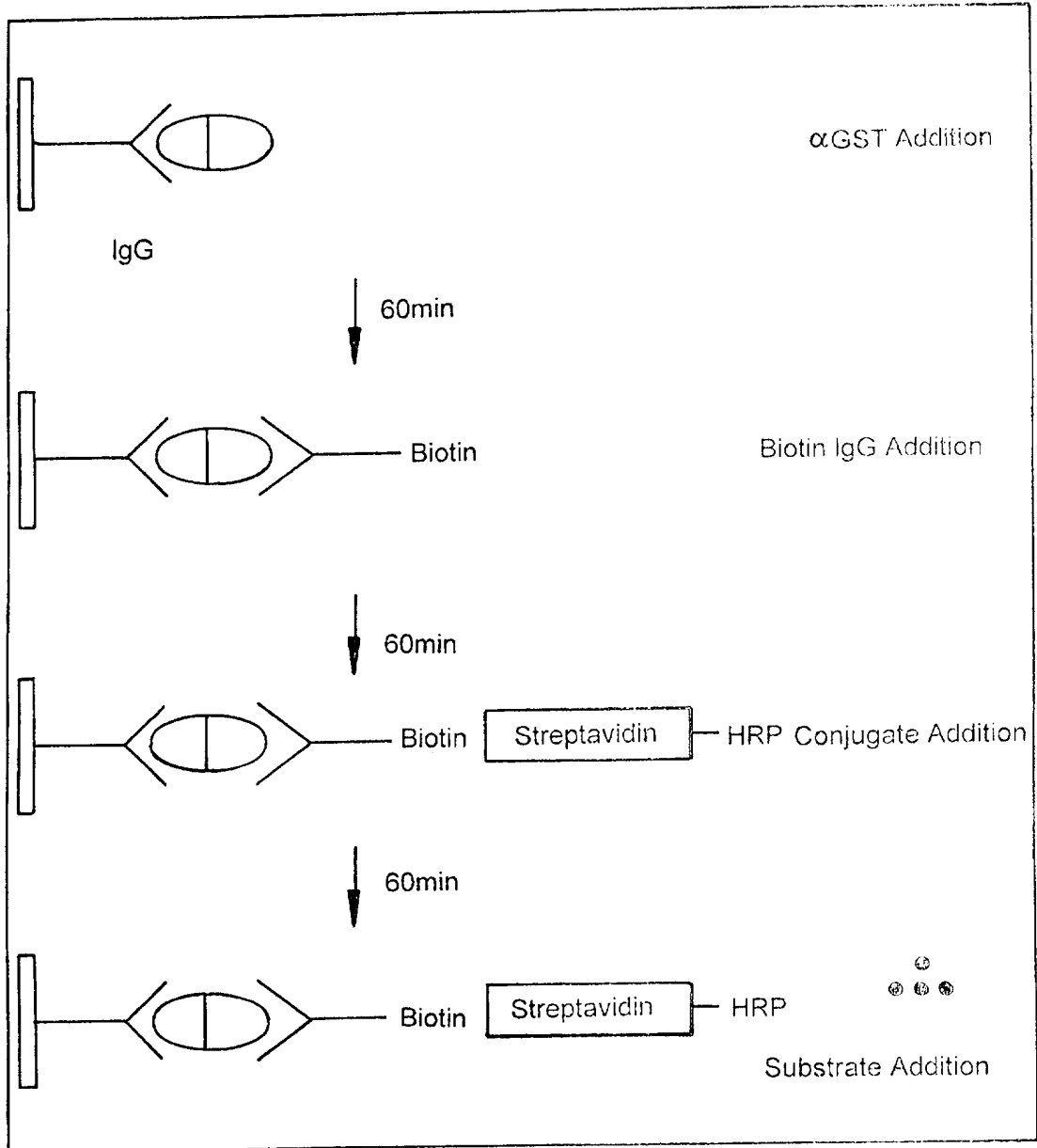


FIG. 1

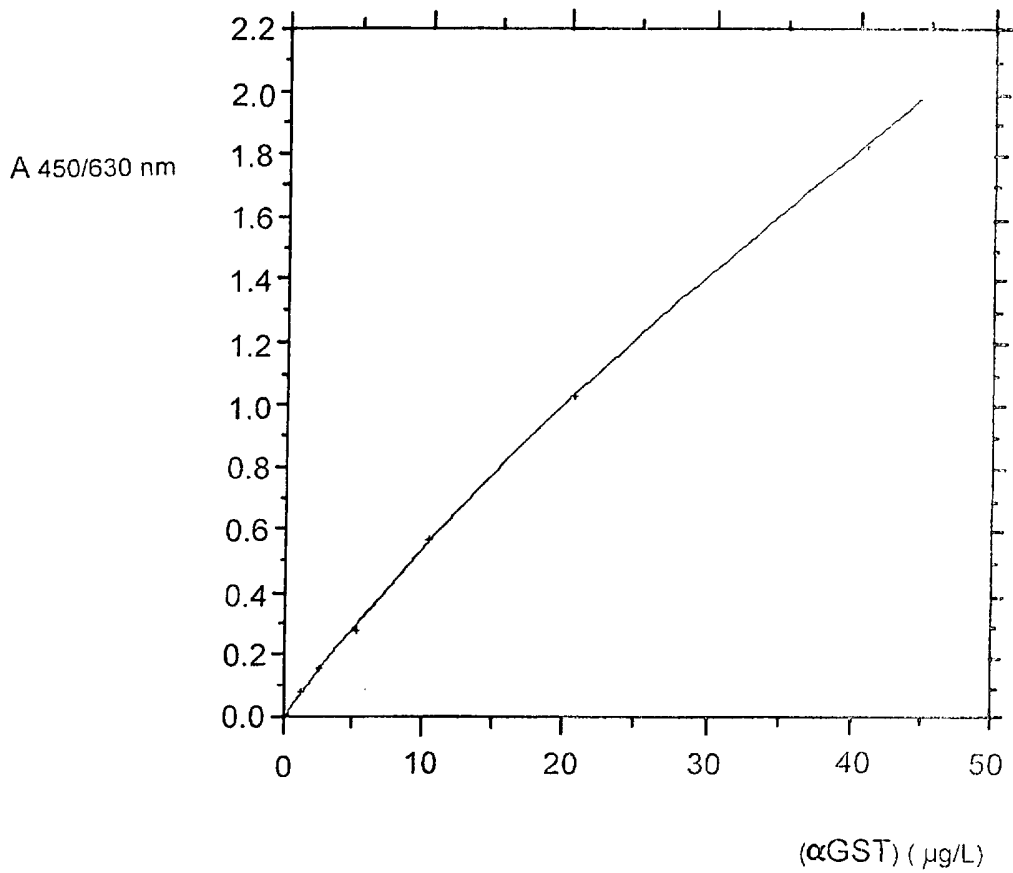


FIG. 2

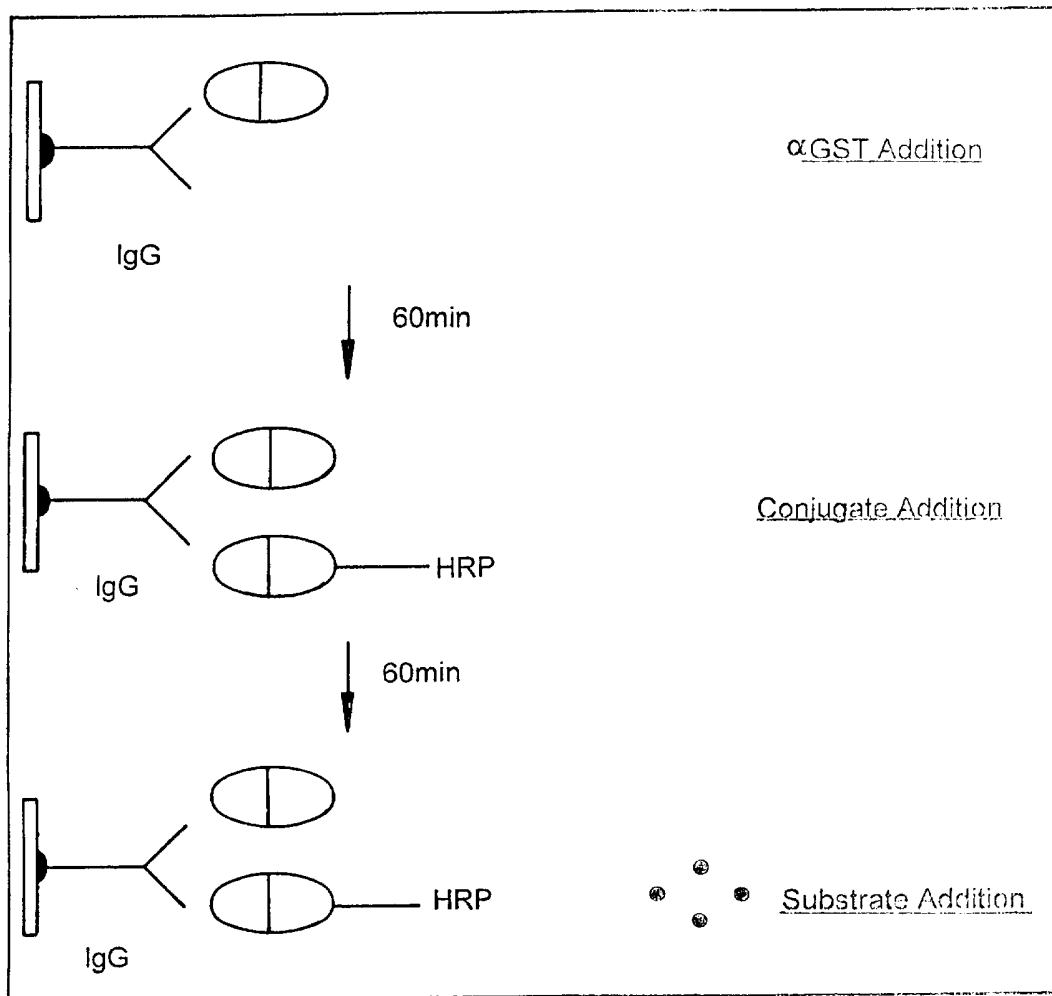


FIG. 3

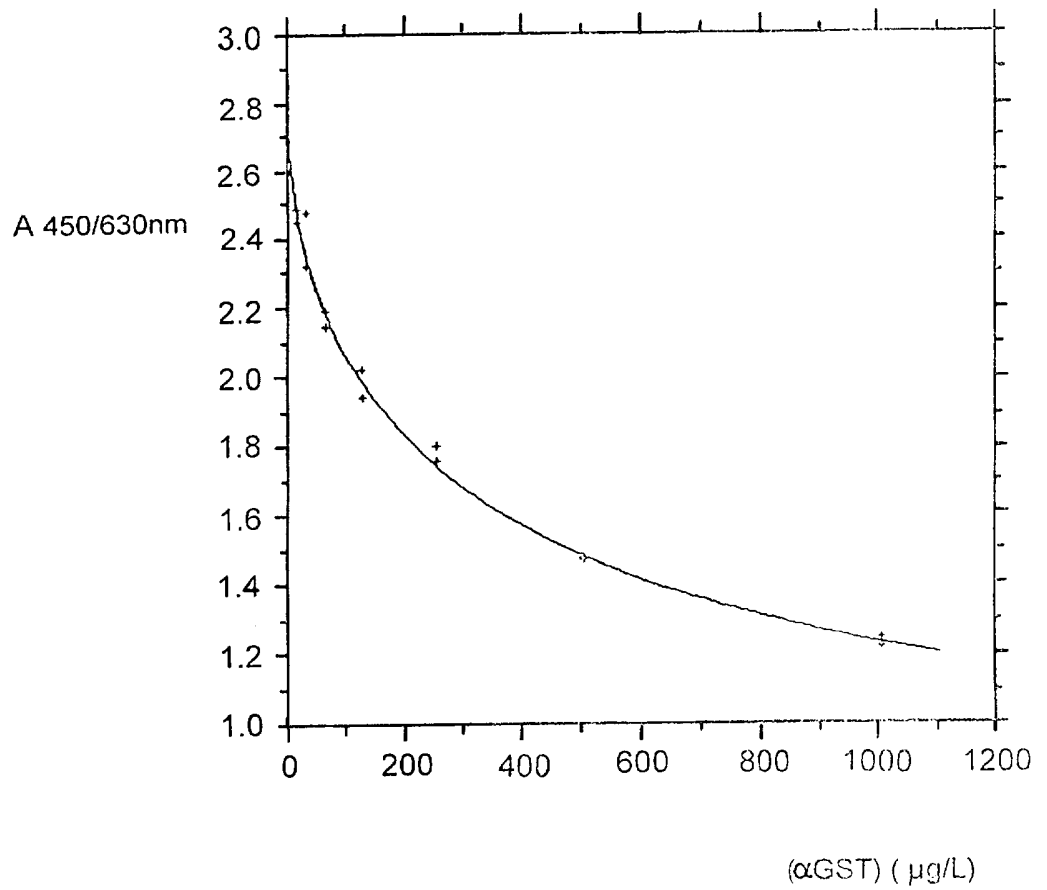


FIG. 4

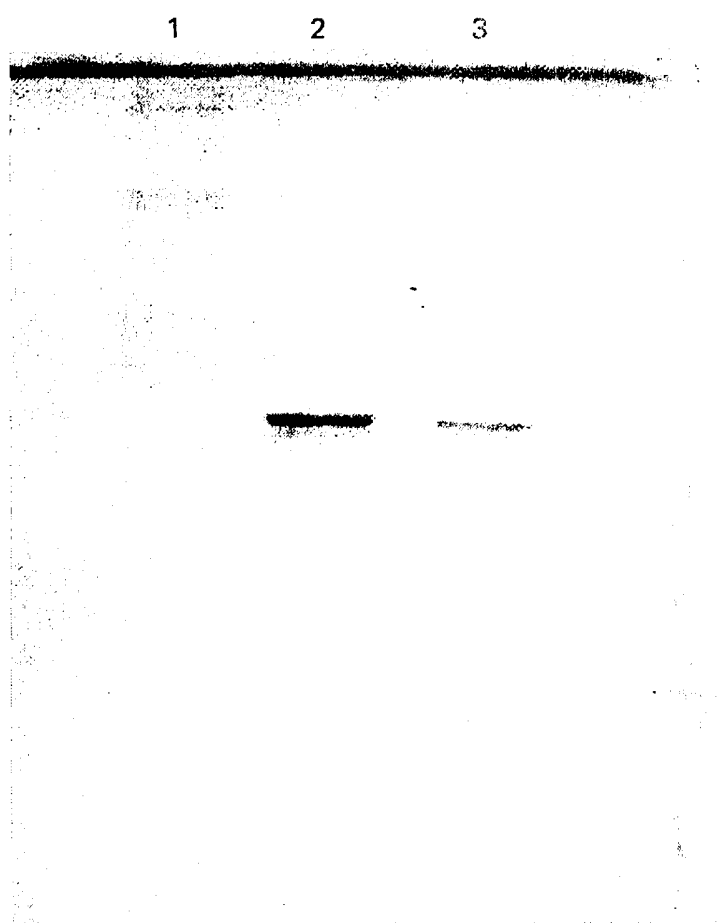


FIG. 5

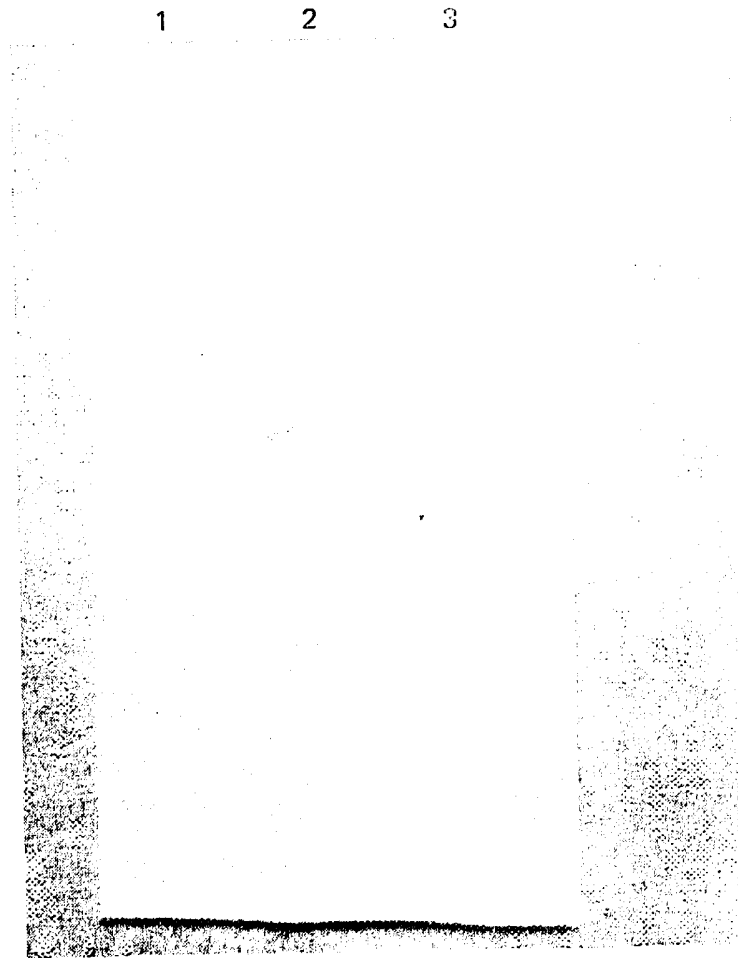


FIG. 6

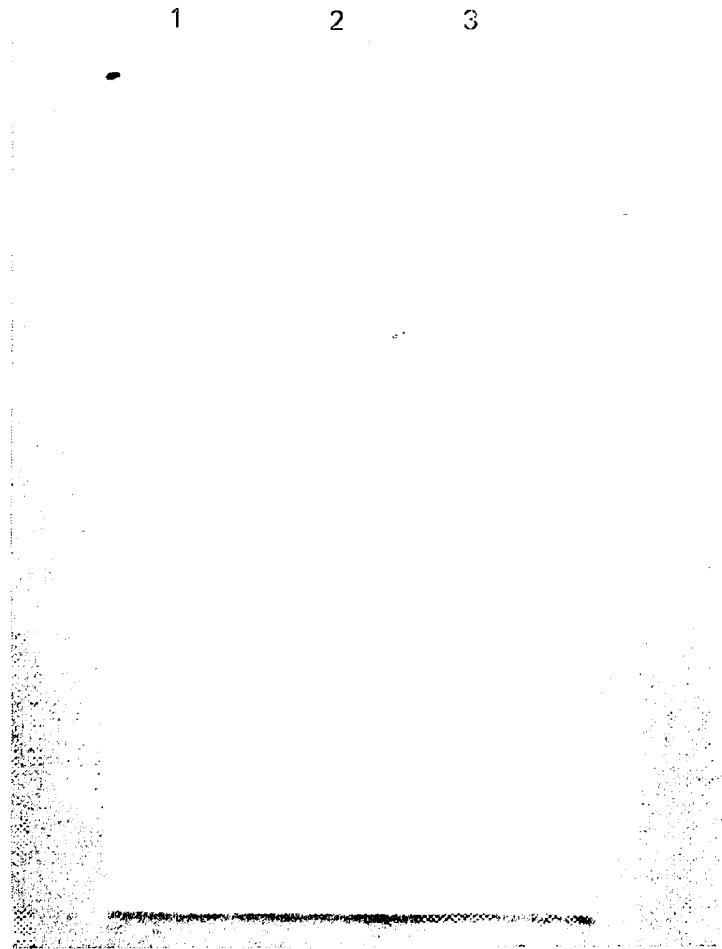


FIG. 7

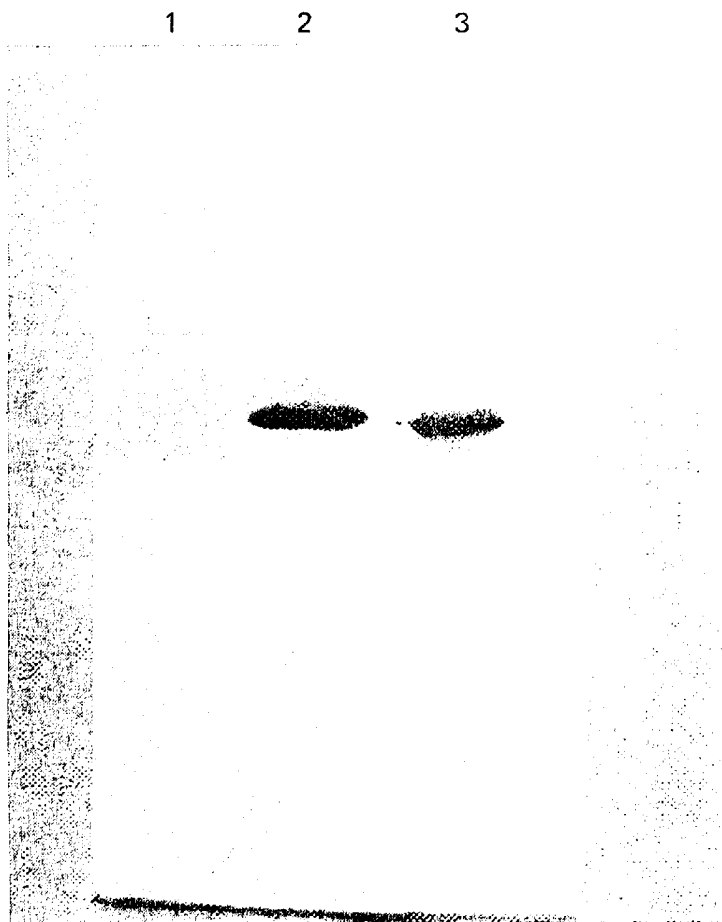


FIG. 8

INTERNATIONAL SEARCH REPORT

International Application No
PC1, IE 95/00061

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 G01N33/53 G01N33/68 G01N33/573 //C12Q1/48

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
IPC 6 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	CELL BIOLOGY AND TOXICOLOGY, vol. 10, no. 5/6, December 1994, pages 407-414, XP000578780 A.E.M. VICKERS.: "Use of human organ slices to evaluate the biotransformation and drug-induced side-effects of pharmaceuticals." ---	
A	WO,A,93 22452 (C. G. KILTY ET AL.) 11 November 1993 cited in the application & EP,A,0 640 145 ---	-/--

Further documents are listed in the continuation of box C.

Patent family members are listed in annex.

* Special categories of cited documents :

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Date of the actual completion of the international search

19 August 1996

Date of mailing of the international search report

09.09.96

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Cartagena y Abella, P

INTERNATIONAL SEARCH REPORT

International Application No
PC 17 IE 95/00061

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>NEPHRON, vol. 67, no. 3, July 1994, pages 308-316, XP000578885 A.G.M. SUNDBERG ET AL.: "Urinary pi-class glutathione transferase as an indicator of tubular damage in the human kidney." cited in the application ---</p>	
E	<p>EP,A,0 692 716 (BAYER AG) 17 January 1996 see the whole document ---</p>	1-14
T	<p>TRANSPLANTATION, vol. 61, no. 6, 27 March 1996, pages 904-908, XP000578779 P. TIAINEN ET AL.: "Hepatocellular integrity in liver donors and recipients indicated by glutathione transferase alpha." -----</p>	

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/IE 95/00061

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9322452	11-11-93	EP-A- 0640145 JP-T- 7506428 US-A- 5217868	01-03-95 13-07-95 08-06-93
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EP-A-692716	17-01-96	NONE	
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