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Danubia Szabadalmi és Jogi Iroda Kft.,
Budapest(54) **Polipeptid vakcinák hipervirulens meningokokkusz vonalak elleni széleskörű védelemre**

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(54) **Polypeptide vaccines for broad protection against hypervirulent meningococcal lineages**

Poylpeptidimpfstoffe zum breiten Schutz gegen hypervirulente Meningo-kokken-Linien

Vaccins incluant du NadA oligomère de la méningococcie pour une protection élargie contre des lignées hypervirulentes

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Description**TECHNICAL FIELD**

5 **[0001]** This invention is in the fields of immunology and vaccinology. In particular, it relates to antigens from *Neisseria meningitidis* (meningococcus) and their use in immunisation.

BACKGROUND ART

10 **[0002]** *N.meningitidis* is a non-motile, Gram-negative human pathogen that colonises the pharynx and causes meningitis (and, occasionally, septicaemia in the absence of meningitis). It causes both endemic and epidemic disease. Following the introduction of the conjugate vaccine against *Haemophilus influenzae*, *N.meningitidis* is the major cause of bacterial meningitis in the USA.

15 **[0003]** Based on the organism's capsular polysaccharide, various serogroups of *N.meningitidis* have been identified. Serogroup A is the pathogen most often implicated in epidemic disease in sub-Saharan Africa. Serogroups B and C are responsible for the vast majority of cases in the United States and in most developed countries. Serogroups W135 and Y are responsible for the rest of the cases in the USA and developed countries. After serogroup, classification includes serotype, serosubtype and then immunotype, and the standard nomenclature lists serogroup, serotype, serosubtype, and immunotype, each separated by a colon e.g. B:4:P1.15:L3,7,9. Within serogroup B, some lineages cause disease often (hyperinvasive), some lineages cause more severe forms of disease than others (hypervirulent), and others rarely cause disease at all. Seven hypervirulent lineages are recognised, namely subgroups I, III and IV-1, ET-5 complex, ET-37 complex, A4 cluster and lineage 3. These have been defined by multilocus enzyme electrophoresis (MLEE), but multilocus sequence typing (MLST) has also been used to classify meningococci [ref. 1].

25 **[0004]** A polysaccharide vaccine against serogroups A, C, W135 & Y has been known for many years [2, 3] but a vaccine against serogroup B has proved elusive. Vaccines based on outer-membrane vesicles have been tested [e.g. see ref. 4], but the protection afforded by these vaccines is typically restricted to the strain used to make the vaccine. There remains a need, therefore, for a broadly-effective serogroup B vaccine.

30 **[0005]** Genome sequences for meningococcal serogroups A [5] and B [6,7] have been reported, and the serogroup B sequence has been studied to identify vaccine antigens [e.g. refs. 8 to 13]. Candidate antigens have been manipulated to improve heterologous expression [refs. 14 to 16].

[0006] It is an object of the invention to provide further and improved compositions for providing immunity against meningococcal disease and/or infection, and in particular for providing broad immunity against serogroup B meningococcus.

DISCLOSURE OF THE INVENTION

35 **[0007]** Vaccines against pathogens such as hepatitis B virus, diphtheria and tetanus typically contain a single protein antigen (e.g. the HBV surface antigen, or a tetanus toxoid). In contrast, acellular whooping cough vaccines typically contain at least three *B.pertussis* proteins and the Prevenar™ pneumococcal vaccine contains seven separate conjugated saccharide antigens. Other vaccines such as cellular pertussis vaccines, the measles vaccine, the inactivated polio vaccine (IPV) and meningococcal OMV vaccines are by their very nature complex mixtures of a large number of antigens.

40 **[0008]** Whether protection against can be elicited by a single antigen, a small number of defined antigens, or a complex mixture of undefined antigens, therefore depends on the pathogen in question. The invention is based on the discovery that a small number of defined antigens is able to provide broad protection against meningococcal infection, and the invention provides a composition which, after administration to a subject, is able to induce an antibody response in that subject, wherein the antibody response is bactericidal against two or more (e.g. 2 or 3) of hypervirulent lineages A4, ET-5 and lineage 3 of *N.meningitidis* serogroup B.

45 **[0009]** Rather than consisting of a single antigen, it is preferred that the composition of the invention comprises a mixture of 10 or fewer (e.g. 9, 8, 7, 6, 5, 4, 3, 2) purified antigens, and it is particularly preferred that the composition should not include complex or undefined mixtures of antigens e.g. it is preferred not to include outer membrane vesicles in the composition.

50 **[0010]** For serogroup B meningococcus, a mixture of five defined protein antigens has been found to elicit a good protective immune response. The invention thus provides a composition as defined in the claims comprising the following five meningococcal protein antigens: (1) a 'NadA' protein; (2) a '741' protein; (3) a '936' protein; (4) a '953' protein; and
55 (5) a '287' protein. These antigens are referred to herein as the 'five basic antigens'.

NadA protein

[0011] 'NadA' (Neisserial adhesin A) from serogroup B of *N.meningitidis* is disclosed as protein '961' in reference 10 (SEQ IDs 2943 & 2944) and as 'NMB1994' in reference 6 (see also GenBank accession numbers: 11352904 & 7227256). A detailed description of the protein can be found in reference 17. There is no corresponding protein in serogroup A [5, 17].

[0012] When used according to the present invention, NadA is a polypeptide consisting of the amino acid sequence of SEQ ID NO:2.

[0013] NadA may take various forms including truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, NadA without its C-terminal membrane anchor is preferred (e.g. deletion of residues 351-405 for strain 2996 [SEQ ID 1]), which is sometimes distinguished herein by the use of a 'C' superscript e.g. NadA^(C). Expression of NadA without its membrane anchor domain (e.g. SEQ ID 1) in *E.coli* results in secretion of the protein into the culture supernatant with concomitant removal of its 23mer leader peptide (e.g. to leave a 327mer for strain 2996 [SEQ ID 2]). Polypeptides without their leader peptides are sometimes distinguished herein by the use of a 'NL' superscript e.g. NadA^(NL) or NadA^{(C)(NL)}.

[0014] Allelic forms of NadA are shown in Figure 9 of reference 18.

[0015] Where N-terminus residues are deleted, it is preferred that the deletion should not remove the ability of NadA to adhere to human epithelial cells. A preferred fragment of SEQ ID 1 is SEQ ID 2.

[0016] Secreted NadA can conveniently be prepared in highly pure form from culture supernatant by a process comprising the steps of: concentration and diafiltration against a buffer by ultrafiltration; anionic column chromatography; hydrophobic column chromatography; hydroxylapatite ceramic column chromatography; diafiltration against a buffer; and filter sterilisation. Further details of the process are given in the examples.

[0017] NadA is preferably used in an oligomeric form (e.g. in trimeric form).

741 protein

[0018] '741' protein from serogroup B is disclosed in reference 10 (SEQ IDs 2535 & 2536) and as 'NMB1870' in reference 6 (see also GenBank accession number GI:7227128). The corresponding protein in serogroup A [5] has GenBank accession number 7379322. 741 is naturally a lipoprotein.

[0019] When used according to the present invention, 741 protein has $\geq 85\%$ identity to SEQ ID NO: 3.

[0020] 741 protein may take various forms including truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, the N-terminus of 741 may be deleted up to and including its poly-glycine sequence (*i.e.* deletion of residues 1 to 72 for strain MC58 [SEQ ID 3]), which is sometimes distinguished herein by the use of a ' ΔG ' prefix. This deletion can enhance expression. The deletion also removes 741's lipidation site.

[0021] Preferred 741 sequences have 90% or more identity (e.g. 95%, 99% or more) to SEQ ID 3. This includes 741 variants (e.g. allelic variants, homologs, orthologs, paralogs, mutants, *etc.*). Allelic forms of 741 can be found in SEQ IDs 1 to 22 of reference 16, and in SEQ IDs 1 to 23 of reference 19. SEQ IDs 1-299 of reference 20 give further 741 sequences.

[0022] Protein 741 is an extremely effective antigen for eliciting anti-meningococcal antibody responses, and it is expressed across all meningococcal serogroups. Phylogenetic analysis shows that the protein splits into two groups, and that one of these splits again to give three variants in total [21], and while serum raised against a given variant is bactericidal within the same variant group, it is not active against strains which express one of the other two variants *i.e.* there is intra-variant cross-protection, but not inter-variant cross-protection. For maximum cross-strain efficacy, therefore, it is preferred that a composition should include more than one variant of protein 741. An exemplary sequence from each variant is given in SEQ ID 10, 11 and 12 herein, starting with a N-terminal cysteine residue to which a lipid will be covalently attached in the lipoprotein form of 741.

[0023] It is therefore preferred that the composition should include: (1) a first protein, comprising an amino acid sequence having at least $a\%$ sequence identity to SEQ ID 10; and at least one of: (2) a second protein, comprising an amino acid sequence having at least $b\%$ sequence identity to SEQ ID 11 and/or comprising an amino acid sequence consisting of a fragment of at least y contiguous amino acids from SEQ ID 11; and (3) a third protein, comprising an amino acid sequence having at least $c\%$ sequence identity to SEQ ID 12 and/or comprising an amino acid sequence consisting of a fragment of at least z contiguous amino acids from SEQ ID 12.

[0024] The value of a is at least 85 e.g. 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 99.5, or more. The value of b is at least 85 e.g. 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 99.5, or more. The value of c is at least 85 e.g. 86, 87, 88, 89, 90, 91, 92, 93, 94, 95, 96, 97, 98, 99, 99.5, or more. The values of a , b and c are not intrinsically related to each other.

[0025] The value of y is at least 7 e.g. 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 120, 140, 160, 180, 200, 225, 250). The value of z is at least 7 e.g. 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 60, 70, 80, 90, 100, 120, 140,

160, 180, 200, 225, 250). The values of x , y and z are not intrinsically related to each other.

[0026] It is preferred that any given 741 amino acid sequence will not fall into more than one of categories (1), (2) and (3). Any given 741 sequence will thus fall into only one of categories (1), (2) and (3). It is thus preferred that: protein (1) has less than $i\%$ sequence identity to protein (2); protein (1) has less than $j\%$ sequence identity to protein (3); and protein (2) has less than $k\%$ sequence identity to protein (3). The value of i is 60 or more (e.g. 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, etc.) and is at most a . The value of j is 60 or more (e.g. 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, etc.) and is at most b . The value of k is 60 or more (e.g. 61, 62, 63, 64, 65, 66, 67, 68, 69, 70, 71, 72, 73, 74, 75, 76, 77, 78, 79, 80, 81, 82, 83, 84, 85, 86, 87, 88, 89, 90, etc.) and is at most c . The values of i , j and k are not intrinsically related to each other.

936 protein

[0027] '936' protein from serogroup B is disclosed in reference 10 (SEQ IDs 2883 & 2884) and as 'NMB2091' in reference 6 (see also GenBank accession number GI:7227353). The corresponding gene in serogroup A [5] has GenBank accession number 7379093.

[0028] When used according to the present invention, 936 protein has $\geq 85\%$ identity to SEQ ID NO: 4.

[0029] 936 protein may take various forms including truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, the N-terminus leader peptide of 936 may be deleted (*i.e.* deletion of residues 1 to 23 for strain MC58 [SEQ ID 4]) to give 936^(NL).

[0030] Preferred 936 sequences have 90% or more identity (e.g. 95%, 99% or more) to SEQ ID 4. This includes variants (e.g. allelic variants, homologs, orthologs, paralog, mutants etc.).

953 protein

[0031] '953' protein from serogroup B is disclosed in reference 10 (SEQ IDs 2917 & 2918) and as 'NMB1030' in reference 6 (see also GenBank accession number GI:7226269). The corresponding protein in serogroup A [5] has GenBank accession number 7380108.

[0032] When used according to the present invention, 953 protein has $\geq 85\%$ identity to SEQ ID NO: 5.

[0033] 953 protein may take various forms. Preferred forms of 953 are truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, the N-terminus leader peptide of 953 may be deleted (*i.e.* deletion of residues 1 to 19 for strain MC58 [SEQ ID 5]) to give 953^(NL).

[0034] Preferred 953 sequences have 90% or more identity (e.g. 95%, 99% or more) to SEQ ID 5. This includes 953 variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.). Allelic forms of 953 can be seen in Figure 19 of reference 12.

287 protein

[0035] '287' protein from serogroup B is disclosed in reference 10 (SEQ IDs 3103 & 3104), as 'NMB2132' in reference 6, and as 'GNA2132' in reference 13 (see also GenBank accession number GI:7227388). The corresponding protein in serogroup A [5] has GenBank accession number 7379057.

[0036] When used according to the present invention, 287 protein has $\geq 85\%$ identity to SEQ ID NO: 6.

[0037] 287 protein may take various forms including truncation or deletion variants, such as those disclosed in references 14 to 16. In particular, the N-terminus of 287 may be deleted up to and including its poly-glycine sequence (*i.e.* deletion of residues 1 to 24 for strain MC58 [SEQ ID 6]), which is sometimes distinguished herein by the use of a ' ΔG ' prefix. This deletion can enhance expression.

[0038] Preferred 287 sequences have 90% or more identity (e.g. 95%, 99% or more) to SEQ ID 6. This includes 287 variants (e.g. allelic variants, homologs, orthologs, paralog, mutants, etc.). Allelic forms of 287 can be seen in Figures 5 and 15 of reference 12, and in example 13 and figure 21 of reference 10 (SEQ IDs 3179 to 3184).

Fusion proteins

[0039] The five antigens may be present in the composition as five separate proteins, but it is preferred that at least two of the antigens are expressed as a single polypeptide chain (a 'hybrid' protein [refs. 14 to 16]) e.g. such that the five antigens form fewer than five polypeptides. Hybrid proteins offer two principal advantages: first, a protein that may be unstable or poorly expressed on its own can be assisted by adding a suitable hybrid partner that overcomes the problem; second, commercial manufacture is simplified as only one expression and purification need be employed in order to produce two separately-useful proteins.

[0040] A hybrid protein included in a composition of the invention may comprise two or more (*i. e.* 2, 3, 4 or 5) of the five basic antigens. Hybrids consisting of two of the five basic antigens are preferred.

[0041] Within the combination of five basic antigens, an antigen may be present in more than one hybrid protein and/or as a non-hybrid protein. It is preferred, however, that an antigen is present either as a hybrid or as a non-hybrid, but not as both, although it may be useful to include protein 741 both as a hybrid and a non-hybrid (preferably lipoprotein) antigen, particularly where more than one variant of 741 is used.

[0042] Two-antigen hybrids for use in the invention comprise: NadA & 741; NadA & 936; NadA & 953; NadA & 287; 741 & 936; 741 & 953; 741 & 287; 936 & 953; 936 & 287; 953 & 287. Preferred two-antigen hybrids comprise: 741 & 936; 953 & 287.

[0043] Hybrid proteins can be represented by the formula $\text{NH}_2\text{-A-}[\text{X-L}]_n\text{-B-COOH}$, wherein: X is an amino acid sequence of one of the five basic antigens; L is an optional linker amino acid sequence; A is an optional N-terminal amino acid sequence; B is an optional C-terminal amino acid sequence; and *n* is 2, 3, 4 or 5.

[0044] If a -X- moiety has a leader peptide sequence in its wild-type form, this may be included or omitted in the hybrid protein. In some embodiments, the leader peptides will be deleted except for that of the -X- moiety located at the N-terminus of the hybrid protein *i.e.* the leader peptide of X_1 will be retained, but the leader peptides of $X_2 \dots X_n$ will be omitted. This is equivalent to deleting all leader peptides and using the leader peptide of X_1 as moiety -A-.

[0045] For each *n* instances of [-X-L-], linker amino acid sequence -L- may be present or absent. For instance, when *n*=2 the hybrid may be $\text{NH}_2\text{-X}_1\text{-L}_1\text{-X}_2\text{-L}_2\text{-COOH}$, $\text{NH}_2\text{-X}_1\text{-X}_2\text{-COOH}$, $\text{NH}_2\text{-X}_1\text{-L}_1\text{-X}_2\text{-COOH}$, $\text{NH}_2\text{-X}_1\text{-X}_2\text{-L}_2\text{-COOH}$, *etc.* Linker amino acid sequence(s) -L- will typically be short (*e.g.* 20 or fewer amino acids *i.e.* 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples comprise short peptide sequences which facilitate cloning, poly-glycine linkers (*i.e.* comprising Gly_n where *n* = 2, 3, 4, 5, 6, 7, 8, 9, 10 or more), and histidine tags (*i.e.* His_n where *n* = 3, 4, 5, 6, 7, 8, 9, 10 or more). Other suitable linker amino acid sequences will be apparent to those skilled in the art. A useful linker is GSGGGG (SEQ ID 9), with the Gly-Ser dipeptide being formed from a *Bam*HI restriction site, thus aiding cloning and manipulation, and the $(\text{Gly})_4$ tetrapeptide being a typical poly-glycine linker. If X_{n+1} is a Δ G protein and L_n is a glycine linker, this may be equivalent to X_{n+1} not being a Δ G protein and L_n being absent.

[0046] -A- is an optional N-terminal amino acid sequence. This will typically be short (*e.g.* 40 or fewer amino acids *i.e.* 39, 38, 37, 36, 35, 34, 33, 32, 31, 30, 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples include leader sequences to direct protein trafficking, or short peptide sequences which facilitate cloning or purification (*e.g.* histidine tags *i.e.* His_n where *n* = 3, 4, 5, 6, 7, 8, 9, 10 or more). Other suitable N-terminal amino acid sequences will be apparent to those skilled in the art. If X_1 lacks its own N-terminus methionine, -A- is preferably an oligopeptide (*e.g.* with 1, 2, 3, 4, 5, 6, 7 or 8 amino acids) which provides a N-terminus methionine.

[0047] -B- is an optional C-terminal amino acid sequence. This will typically be short (*e.g.* 40 or fewer amino acids *i.e.* 39, 38, 37, 36, 35, 34, 33, 32, 31, 30, 29, 28, 27, 26, 25, 24, 23, 22, 21, 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5, 4, 3, 2, 1). Examples include sequences to direct protein trafficking, short peptide sequences which facilitate cloning or purification (*e.g.* comprising histidine tags *i.e.* His_n where *n* = 3, 4, 5, 6, 7, 8, 9, 10 or more), or sequences which enhance protein stability. Other suitable C-terminal amino acid sequences will be apparent to those skilled in the art.

[0048] Most preferably, *n* is 2. Two preferred proteins of this type are: X_1 is a 936 and X_2 is a 741; X_1 is a 287 and X_2 is a 953.

[0049] Two particularly preferred hybrid proteins of the invention are as follows:

n	A	X_1	L_1	X_2	L_2	B	[SEQ ID]
2	MA	Δ G287	GSGGGG	953 ^(NL)	-	-	7
2	M	936 ^(NL)	GSGGGG	Δ G741	-	-	8

These two proteins may be used in combination with NadA (SEQ ID 2).

[0050] 936- Δ G741 hybrid can conveniently be prepared in highly pure form from expression in *E.coli* by a process comprising the steps of: homogenisation; centrifugation; cationic column chromatography; anionic column chromatography; hydrophobic column chromatography; diafiltration against a buffer; and filter sterilisation. Further details of the process are given in the examples.

[0051] Polypeptides can be prepared by various means (*e.g.* recombinant expression, purification from cell culture, chemical synthesis (at least in part), *etc.*) and in various forms (*e.g.* native, fusions, non-glycosylated, lipidated, *etc.*). They are preferably prepared in substantially pure form (*i.e.* substantially free from other *N.meningitidis* or host cell proteins).

Strains

[0052] Preferred proteins comprise an amino acid sequence found in *N.meningitidis* serogroup B. Within serogroup B, preferred strains are 2996, MC58, 95N477, and 394/98. Strain 394/98 is sometimes referred to herein as 'NZ', as it is a New Zealand strain.

[0053] Protein 287 is preferably from strain 2996 or, more preferably, from strain 394/98.

[0054] Protein 741 is preferably from serogroup B strains MC58, 2996, 394/98, or 95N477, or from serogroup C strain 90/18311. Strain MC58 is more preferred.

[0055] Proteins 936 and 953 are preferably from strain 2996. NadA is from strain 2996.

[0056] Strains may be indicated as a subscript e.g. 741_{MC58} is protein 741 from strain MC58. Unless otherwise stated, proteins mentioned herein (e.g. with no subscript) are from *N.meningitidis* strain 2996, which can be taken as a 'reference' strain. It will be appreciated, however, that the invention is not in general limited by strain. As mentioned above, general references to a protein (e.g. '287', '919' etc.) may be taken to include that protein from any strain. This will typically have sequence identity to 2996 of 90% or more (eg. 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99% or more).

[0057] Where a composition includes a particular protein antigen (e.g. 741 or 287), the composition can include that antigen in more than one variant form e.g. the same protein, but from more than one strain. These proteins may be included as tandem or separate proteins.

[0058] Where hybrid proteins are used, the individual antigens within the hybrid (i.e. individual -X-moieties) may be from one or more strains. Where $n=2$, for instance, X_2 may be from the same strain as X_1 or from a different strain.

Where $n=3$, the strains might be (i) $X_1=X_2=X_3$ (ii) $X_1=X_2 \neq X_3$ (iii) $X_1 \neq X_2 = X_3$ (iv) $X_1 \neq X_2 \neq X_3$ or (v) $X_1 = X_3 \neq X_2$, etc.

Hypervirulent lineages and bactericidal antibody responses

[0059] In general, compositions of the invention are able to induce serum bactericidal antibody responses after being administered to a subject. These responses are conveniently measured in mice and are a standard indicator of vaccine efficacy [e.g. see end-note 14 of reference 13]. Serum bactericidal activity (SBA) measures bacterial killing mediated by complement, and can be assayed using human or baby rabbit complement. WHO standards require a vaccine to induce at least a 4-fold rise in SBA in more than 90% of recipients.

[0060] Rather than offering narrow protection, compositions of the invention can induce bactericidal antibody responses against more than one hypervirulent lineage of serogroup B. In particular, they can induce bactericidal responses against two or three of the following three hypervirulent lineages: (i) cluster A4; (ii) ET5 complex; and (iii) lineage 3. They may additionally induce bactericidal antibody responses against one or more of hypervirulent lineages subgroup I, subgroup III, subgroup IV-1 or ET-37 complex, and against other lineages e.g. hyperinvasive lineages.

[0061] This does not necessarily mean that the composition can induce bactericidal antibodies against each and every strain of serogroup B meningococcus within these hypervirulent lineages e.g. rather, for any given group of four or more strains of serogroup B meningococcus within a particular hypervirulent lineage, the antibodies induced by the composition are bactericidal against at least 50% (e.g. 60%, 70%, 80%, 90% or more) of the group. Preferred groups of strains will include strains isolated in at least four of the following countries: GB, AU, CA, NO, IT, US, NZ, NL, BR, and CU. The serum preferably has a bactericidal titre of at least 1024 (e.g. 2^{10} , 2^{11} , 2^{12} , 2^{13} , 2^{14} , 2^{15} , 2^{16} , 2^{17} , 2^{18} or higher, preferably at least 2^{14}) i.e. the serum is able to kill at least 50% of test bacteria of a particular strain when diluted 1/1024, as described in reference 13.

[0062] Preferred compositions can induce bactericidal responses against the following strains of serogroup B meningococcus: (i) from cluster A4, strain 961-5945 (B:2b:P1.21,16) and/or strain G2136 (B:-); (ii) from ET-5 complex, strain MC58 (B:15:P1.7,16b) and/or strain 44/76 (B:15:P1.7,16); (iii) from lineage 3, strain 394/98 (B:4:P1.4) and/or strain BZ198 (B:NT:-). More preferred compositions can induce bactericidal responses against strains 961-5945, 44/76 and 394/98.

[0063] Strains 961-5945 and G2136 are both *Neisseria* MLST reference strains [ids 638 & 1002 in ref. 22]. Strain MC58 is widely available (e.g. ATCC BAA-335) and was the strain sequenced in reference 6. Strain 44/76 has been widely used and characterised (e.g. ref. 23) and is one of the *Neisseria* MLST reference strains [id 237 in ref. 22; row 32 of Table 2 in ref. 1]. Strain 394/98 was originally isolated in New Zealand in 1998, and there have been several published studies using this strain (e.g. refs. 24 & 25). Strain BZ198 is another MLST reference strain [id 409 in ref. 22; row 41 of Table 2 in ref. 1].

[0064] The composition may additionally induce a bactericidal response against serogroup W135 strain LNP17592 (W135:2a:P1.5,2), from ET-37 complex. This is a Hajj strain isolated in France in 2000.

Heterologous host

[0065] Whilst expression of the proteins may take place in *Neisseria*, preferably a heterologous host is utilised. The

heterologous host may be prokaryotic (e.g. a bacterium) or eukaryotic. It is preferably *E.coli*, but other suitable hosts include *Bacillus subtilis*, *Vibrio cholerae*, *Salmonella typhi*, *Salmonella typhimurium*, *Neisseria lactamica*, *Neisseria cinerea*, *Mycobacteria* (e.g. *M.tuberculosis*), yeast, etc.

[0066] Thus the invention provides a composition as defined in the claims which, after administration to a subject, is able to induce an antibody response in that subject, wherein the antibody response is bactericidal against two or more (e.g. 2 or 3) of hypervirulent lineages A4, ET-5 and lineage 3 of *N.meningitidis* serogroup B, and wherein the immunogens in the composition which give rise to the antibody response are obtained by recombinant expression in a non-Neisserial host. Thus the immunogens in the compositions of the invention are preferably recombinant immunogens. Compositions which do not include OMV preparations may thus be preferred.

Immunogenic compositions and medicaments

[0067] Compositions of the invention are immunogenic, and are more preferably vaccine compositions. Vaccines according to the invention may either be prophylactic (i.e. to prevent infection) or therapeutic (i.e. to treat infection), but will typically be prophylactic.

[0068] The pH of the composition is preferably between 6 and 8, preferably about 7. Stable pH may be maintained by the use of a buffer. Where a composition comprises an aluminium hydroxide salt, it is preferred to use a histidine buffer [26]. The composition may be sterile and/or pyrogen-free. Compositions of the invention may be isotonic with respect to humans.

[0069] Compositions may be presented in vials, or they may be presented in ready-filled syringes. The syringes may be supplied with or without needles. A syringe will include a single dose of the composition, whereas a vial may include a single dose or multiple doses. Injectable compositions will usually be liquid solutions or suspensions. Alternatively, they may be presented in solid form (e.g. freeze-dried) for solution or suspension in liquid vehicles prior to injection.

[0070] Compositions of the invention may be packaged in unit dose form or in multiple dose form. For multiple dose forms, vials are preferred to pre-filled syringes. Effective dosage volumes can be routinely established, but a typical human dose of the composition for injection has a volume of 0.5ml.

[0071] Where a composition of the invention is to be prepared extemporaneously prior to use (e.g. where a component is presented in lyophilised form) and is presented as a kit, the kit may comprise two vials, or it may comprise one ready-filled syringe and one vial, with the contents of the syringe being used to reactivate the contents of the vial prior to injection.

[0072] The invention also provides a composition of the invention for use as a medicament. The medicament is preferably able to raise an immune response in a mammal (i.e. it is an immunogenic composition) and is more preferably a vaccine.

[0073] The invention also provides the use of a composition of the invention in the manufacture of a medicament for raising an immune response in a mammal. The use of a 'NadA' protein, a '741' protein, a '936' protein, a '953' protein, and a '287' protein (and other optional antigens) in the manufacture of a medicament for raising an immune response in a mammal is also disclosed. The medicament is preferably a vaccine.

[0074] Also disclosed is a method for raising an immune response in a mammal comprising the step of administering an effective amount of a composition of the invention. The immune response is preferably protective and preferably involves antibodies. The method may raise a booster response.

[0075] The mammal is preferably a human. Where the vaccine is for prophylactic use, the human is preferably a child (e.g. a toddler or infant); where the vaccine is for therapeutic use, the human is preferably an adult. A vaccine intended for children may also be administered to adults e.g. to assess safety, dosage, immunogenicity, etc.

[0076] These uses and methods are preferably for the prevention and/or treatment of a disease caused by a *Neisseria* (e.g. meningitis, septicaemia, bacteremia, gonorrhoea etc.). The prevention and/or treatment of bacterial or meningococcal meningitis is preferred.

[0077] One way of checking efficacy of therapeutic treatment involves monitoring Neisserial infection after administration of the composition of the invention. One way of checking efficacy of prophylactic treatment involves monitoring immune responses against the five basic antigens after administration of the composition. Immunogenicity of compositions of the invention can be determined by administering them to test subjects (e.g. children 12-16 months age, or animal models [27]) and then determining standard parameters including serum bactericidal antibodies (SBA) and ELISA titres (GMT) of total and high-avidity anti-capsule IgG. These immune responses will generally be determined around 4 weeks after administration of the composition, and compared to values determined before administration of the composition. A SBA increase of at least 4-fold or 8-fold is preferred. Where more than one dose of the composition is administered, more than one post-administration determination may be made.

[0078] Preferred compositions of the invention can confer an antibody titre in a patient that is superior to the criterion for seroprotection for each antigenic component for an acceptable percentage of human subjects. Antigens with an associated antibody titre above which a host is considered to be seroconverted against the antigen are well known, and such titres are published by organisations such as WHO. Preferably more than 80% of a statistically significant sample

of subjects is seroconverted, more preferably more than 90%, still more preferably more than 93% and most preferably 96-100%.

[0079] Compositions of the invention will generally be administered directly to a patient. Direct delivery may be accomplished by parenteral injection (e.g. subcutaneously, intraperitoneally, intravenously, intramuscularly, or to the interstitial space of a tissue), or by rectal, oral, vaginal, topical, transdermal, intranasal, ocular, aural, pulmonary or other mucosal administration. Intramuscular administration to the thigh or the upper arm is preferred. Injection may be via a needle (e.g. a hypodermic needle), but needle-free injection may alternatively be used. A typical intramuscular dose is 0.5 ml.

[0080] The invention may be used to elicit systemic and/or mucosal immunity.

[0081] Dosage treatment can be a single dose schedule or a multiple dose schedule. Multiple doses may be used in a primary immunisation schedule and/or in a booster immunisation schedule. A primary dose schedule may be followed by a booster dose schedule. Suitable timing between priming doses (e.g. between 4-16 weeks), and between priming and boosting, can be routinely determined.

[0082] Neisserial infections affect various areas of the body and so the compositions of the invention may be prepared in various forms. For example, the compositions may be prepared as injectables, either as liquid solutions or suspensions. Solid forms suitable for solution in, or suspension in, liquid vehicles prior to injection can also be prepared (e.g. a lyophilised composition). The composition may be prepared for topical administration e.g. as an ointment, cream or powder. The composition may be prepared for oral administration e.g. as a tablet or capsule, or as a syrup (optionally flavoured). The composition may be prepared for pulmonary administration e.g. as an inhaler, using a fine powder or a spray. The composition may be prepared as a suppository or pessary. The composition may be prepared for nasal, aural or ocular administration e.g. as spray, drops, gel or powder [e.g. refs 28 & 29]. Success with nasal administration of pneumococcal saccharides [30,31], pneumococcal polypeptides [32], Hib saccharides [33], MenC saccharides [34], and mixtures of Hib and MenC saccharide conjugates [35] has been reported.

[0083] Immunogenic compositions used as vaccines comprise an immunologically effective amount of antigen(s), as well as any other components, as needed. By 'immunologically effective amount', it is meant that the administration of that amount to an individual, either in a single dose or as part of a series, is effective for treatment or prevention. This amount varies depending upon the health and physical condition of the individual to be treated, age, the taxonomic group of individual to be treated (e.g. non-human primate, primate, etc.), the capacity of the individual's immune system to synthesise antibodies, the degree of protection desired, the formulation of the vaccine, the treating doctor's assessment of the medical situation, and other relevant factors. It is expected that the amount will fall in a relatively broad range that can be determined through routine trials, and a typical quantity of each meningococcal saccharide antigen per dose is between 1 µg and 20 µg e.g. about 1 µg, about 2.5 µg, about 4 µg, about 5 µg, or about 10 µg (expressed as saccharide).

Further non-antigen components of the composition

[0084] The composition of the invention will typically, in addition to the components mentioned above, comprise one or more 'pharmaceutically acceptable carriers', which include any carrier that does not itself induce the production of antibodies harmful to the individual receiving the composition. Suitable carriers are typically large, slowly metabolised macromolecules such as proteins, polysaccharides, polylactic acids, polyglycolic acids, polymeric amino acids, amino acid copolymers, sucrose [36], trehalose [37], lactose, and lipid aggregates (such as oil droplets or liposomes). Such carriers are well known to those of ordinary skill in the art. The vaccines may also contain diluents, such as water, saline, glycerol, etc. Additionally, auxiliary substances, such as wetting or emulsifying agents, pH buffering substances, and the like, may be present. Sterile pyrogen-free, phosphate-buffered physiologic saline is a typical carrier. A thorough discussion of pharmaceutically acceptable excipients is available in reference 38.

[0085] Compositions of the invention may include an antimicrobial, particularly when packaged in multiple dose format.

[0086] Compositions of the invention may comprise detergent e.g. a Tween (polysorbate), such as Tween 80. Detergents are generally present at low levels e.g. <0.01%.

[0087] Compositions of the invention may include sodium salts (e.g. sodium chloride) to give tonicity. A concentration of 10±2mg/ml NaCl is typical.

[0088] Compositions of the invention will generally include a buffer. A phosphate buffer is typical.

[0089] Compositions of the invention may comprise a sugar alcohol (e.g. mannitol) or a disaccharide (e.g. sucrose or trehalose) e.g. at around 15-30mg/ml (e.g. 25 mg/ml), particularly if they are to be lyophilised or if they include material which has been reconstituted from lyophilised material. The pH of a composition for lyophilisation may be adjusted to around 6.1 prior to lyophilisation.

[0090] Vaccines of the invention may be administered in conjunction with other immunoregulatory agents. In particular, compositions will usually include an adjuvant. Adjuvants which may be used in compositions of the invention include, but are not limited to:

A. Mineral-containing compositions

[0091] Mineral containing compositions suitable for use as adjuvants in the invention include mineral salts, such as aluminium salts and calcium salts. The invention includes mineral salts such as hydroxides (e.g. oxyhydroxides), phosphates (e.g. hydroxyphosphates, orthophosphates), sulphates, etc. [e.g. see chapters 8 & 9 of ref. 39], or mixtures of different mineral compounds, with the compounds taking any suitable form (e.g. gel, crystalline, amorphous, etc.), and with adsorption being preferred. The mineral containing compositions may also be formulated as a particle of metal salt [40].

[0092] Aluminium phosphates are particularly preferred, particularly in compositions which include a *H.influenzae* saccharide antigen, and a typical adjuvant is amorphous aluminium hydroxyphosphate with PO_4/Al molar ratio between 0.84 and 0.92, included at 0.6mg Al^{3+}/ml . Adsorption with a low dose of aluminium phosphate may be used e.g. between 50 and 100 μg Al^{3+} per conjugate per dose. Where there is more than one conjugate in a composition, not all conjugates need to be adsorbed.

B. Oil Emulsions

[0093] Oil emulsion compositions suitable for use as adjuvants in the invention include squalene-water emulsions, such as MF59 [Chapter 10 of ref. 39; see also ref. 41] (5% Squalene, 0.5% Tween 80, and 0.5% Span 85, formulated into submicron particles using a microfluidizer). Complete Freund's adjuvant (CFA) and incomplete Freund's adjuvant (IFA) may also be used.

C. Saponin formulations [chapter 22 of ref. 39]

[0094] Saponin formulations may also be used as adjuvants in the invention. Saponins are a heterologous group of sterol glycosides and triterpenoid glycosides that are found in the bark, leaves, stems, roots and even flowers of a wide range of plant species. Saponin from the bark of the *Quillaia saponaria* Molina tree have been widely studied as adjuvants. Saponin can also be commercially obtained from *Smilax ornata* (sarsapilla), *Gypsophilla paniculata* (brides veil), and *Saponaria officianalis* (soap root). Saponin adjuvant formulations include purified formulations, such as QS21, as well as lipid formulations, such as ISCOMs. QS21 is marketed as Stimulon™.

[0095] Saponin compositions have been purified using HPLC and RP-HPLC. Specific purified fractions using these techniques have been identified, including QS7, QS17, QS18, QS21, QH-A, QH-B and QH-C. Preferably, the saponin is QS21. A method of production of QS21 is disclosed in ref. 42. Saponin formulations may also comprise a sterol, such as cholesterol [43].

[0096] Combinations of saponins and cholesterol can be used to form unique particles called immunostimulating complexes (ISCOMs) [chapter 23 of ref. 39]. ISCOMs typically also include a phospholipid such as phosphatidylethanolamine or phosphatidylcholine. Any known saponin can be used in ISCOMs. Preferably, the ISCOM includes one or more of QuilA, QHA & QHC. ISCOMs are further described in refs. 43-45. Optionally, the ISCOMS may be devoid of additional detergent [46].

[0097] A review of the development of saponin based adjuvants can be found in refs. 47 & 48.

D. Virosomes and virus-like particles

[0098] Virosomes and virus-like particles (VLPs) can also be used as adjuvants in the invention. These structures generally contain one or more proteins from a virus optionally combined or formulated with a phospholipid. They are generally non-pathogenic, non-replicating and generally do not contain any of the native viral genome. The viral proteins may be recombinantly produced or isolated from whole viruses. These viral proteins suitable for use in virosomes or VLPs include proteins derived from influenza virus (such as HA or NA), Hepatitis B virus (such as core or capsid proteins), Hepatitis E virus, measles virus, Sindbis virus, Rotavirus, Foot-and-Mouth Disease virus, Retrovirus, Norwalk virus, human Papilloma virus, HIV, RNA-phages, Q β -phage (such as coat proteins), GA-phage, fr-phage, AP205 phage, and Ty (such as retrotransposon Ty protein p1). VLPs are discussed further in refs. 49-54. Virosomes are discussed further in, for example, ref. 55

E. Bacterial or microbial derivatives

[0099] Adjuvants suitable for use in the invention include bacterial or microbial derivatives such as non-toxic derivatives of enterobacterial lipopolysaccharide (LPS), Lipid A derivatives, immunostimulatory oligonucleotides and ADP-ribosylating toxins and detoxified derivatives thereof.

[0100] Non-toxic derivatives of LPS include monophosphoryl lipid A (MPL) and 3-O-deacylated MPL (3dMPL). 3dMPL

is a mixture of 3 de-O-acylated monophosphoryl lipid A with 4, 5 or 6 acylated chains. A preferred "small particle" form of 3 De-O-acylated monophosphoryl lipid A is disclosed in ref. 56. Such "small particles" of 3dMPL are small enough to be sterile filtered through a 0.22µm membrane [56]. Other non-toxic LPS derivatives include monophosphoryl lipid A mimics, such as aminoalkyl glucosaminide phosphate derivatives *e.g.* RC-529 [57,58].

[0101] Lipid A derivatives include derivatives of lipid A from *Escherichia coli* such as OM-174. OM-174 is described for example in refs. 59 & 60.

[0102] Immunostimulatory oligonucleotides suitable for use as adjuvants in the invention include nucleotide sequences containing a CpG motif (a dinucleotide sequence containing an unmethylated cytosine linked by a phosphate bond to a guanosine). Double-stranded RNAs and oligonucleotides containing palindromic or poly(dG) sequences have also been shown to be immunostimulatory.

[0103] The CpG's can include nucleotide modifications/analogs such as phosphorothioate modifications and can be double-stranded or single-stranded. References 61, 62 and 63 disclose possible analog substitutions *e.g.* replacement of guanosine with 2'-deoxy-7-deazaguanosine. The adjuvant effect of CpG oligonucleotides is further discussed in refs. 64-69.

[0104] The CpG sequence may be directed to TLR9, such as the motif GTCGTT or TTCGTT [70]. The CpG sequence may be specific for inducing a Th1 immune response, such as a CpG-A ODN, or it may be more specific for inducing a B cell response, such as a CpG-B ODN. CpG-A and CpG-B ODNs are discussed in refs. 71-73. Preferably, the CpG is a CpG-A ODN.

[0105] Preferably, the CpG oligonucleotide is constructed so that the 5' end is accessible for receptor recognition. Optionally, two CpG oligonucleotide sequences may be attached at their 3' ends to form "immunomers". See, for example, refs. 70 & 74-76.

[0106] Bacterial ADP-ribosylating toxins and detoxified derivatives thereof may be used as adjuvants in the invention. Preferably, the protein is derived from *E.coli* (*E.coli* heat labile enterotoxin "LT"), cholera ("CT"), or pertussis ("PT"). The use of detoxified ADP-ribosylating toxins as mucosal adjuvants is described in ref. 77 and as parenteral adjuvants in ref. 78. The toxin or toxoid is preferably in the form of a holotoxin, comprising both A and B subunits. Preferably, the A subunit contains a detoxifying mutation; preferably the B subunit is not mutated. Preferably, the adjuvant is a detoxified LT mutant such as LT-K63, LT-R72, and LT-G192. The use of ADP-ribosylating toxins and detoxified derivatives thereof, particularly LT-K63 and LT-R72, as adjuvants can be found in refs. 79-86. Numerical reference for amino acid substitutions is preferably based on the alignments of the A and B subunits of ADP-ribosylating toxins set forth in ref. 87.

F. Human immunomodulators

[0107] Human immunomodulators suitable for use as adjuvants in the invention include cytokines, such as interleukins (*e.g.* IL-1, IL-2, IL-4, IL-5, IL-6, IL-7, IL-12 [88], *etc.*) [89], interferons (*e.g.* interferon-γ), macrophage colony stimulating factor, and tumor necrosis factor.

G. Bioadhesives and Mucoadhesives

[0108] Bioadhesives and mucoadhesives may also be used as adjuvants in the invention. Suitable bioadhesives include esterified hyaluronic acid microspheres [90] or mucoadhesives such as cross-linked derivatives of poly(acrylic acid), polyvinyl alcohol, polyvinyl pyrrolidone, polysaccharides and carboxymethylcellulose. Chitosan and derivatives thereof may also be used as adjuvants in the invention [91].

H. Microparticles

[0109] Microparticles may also be used as adjuvants in the invention. Microparticles (*i.e.* a particle of ~100nm to ~150µm in diameter, more preferably ~200nm to ~30µm in diameter, and most preferably ~500nm to ~10µm in diameter) formed from materials that are biodegradable and non-toxic (*e.g.* a poly(α-hydroxy acid), a polyhydroxybutyric acid, a polyorthoester, a polyanhydride, a polycaprolactone, *etc.*), with poly(lactide-co-glycolide) are preferred, optionally treated to have a negatively-charged surface (*e.g.* with SDS) or a positively-charged surface (*e.g.* with a cationic detergent, such as CTAB).

I. Liposomes (chapters 13 & 14 of ref. 39)

[0110] Examples of liposome formulations suitable for use as adjuvants are described in refs. 92-94.

J. Polyoxyethylene ether and polyoxyethylene ester formulations

[0111] Adjuvants suitable for use in the invention include polyoxyethylene ethers and polyoxyethylene esters [95]. Such formulations further include polyoxyethylene sorbitan ester surfactants in combination with an octoxynol [96] as well as polyoxyethylene alkyl ethers or ester surfactants in combination with at least one additional non-ionic surfactant such as an octoxynol [97]. Preferred polyoxyethylene ethers are selected from the following group: polyoxyethylene-9-lauryl ether (laureth 9), polyoxyethylene-9-stearyl ether, polyoxyethylene-8-stearyl ether, polyoxyethylene-4-lauryl ether, polyoxyethylene-35-lauryl ether, and polyoxyethylene-23-lauryl ether.

K. Polyphosphazene (PCPP)

[0112] PCPP formulations are described, for example, in refs. 98 and 99.

L. Muramyl peptides

[0113] Examples of muramyl peptides suitable for use as adjuvants in the invention include N-acetyl-muramyl-L-threonyl-D-isoglutamine (thr-MDP), N-acetyl-normuramyl-L-alanyl-D-isoglutamine (nor-MDP), and N-acetylmuramyl-L-alanyl-D-isoglutaminyl-L-alanine-2-(1'-2'-dipalmitoyl-sn-glycero-3-hydroxyphosphoryloxy)-ethylamine MTP-PE).

M. Imidazoquinolone Compounds.

[0114] Examples of imidazoquinolone compounds suitable for use as adjuvants in the invention include Imiquimod and its homologues (e.g. "Resiquimod 3M"), described further in refs. 100 and 101.

[0115] The invention may also comprise combinations of aspects of one or more of the adjuvants identified above. For example, the following adjuvant compositions may be used in the invention: (1) a saponin and an oil-in-water emulsion [102]; (2) a saponin (e.g. QS21) + a non-toxic LPS derivative (e.g. 3dMPL) [103]; (3) a saponin (e.g. QS21) + a non-toxic LPS derivative (e.g. 3dMPL) + a cholesterol; (4) a saponin (e.g. QS21) + 3dMPL + IL-12 (optionally + a sterol) [104]; (5) combinations of 3dMPL with, for example, QS21 and/or oil-in-water emulsions [105]; (6) SAF, containing 10% squalane, 0.4% Tween 80™, 5% pluronic-block polymer L121, and thr-MDP, either microfluidized into a submicron emulsion or vortexed to generate a larger particle size emulsion. (7) Ribi™ adjuvant system (RAS), (Ribi Immunochem) containing 2% squalene, 0.2% Tween 80, and one or more bacterial cell wall components from the group consisting of monophosphorylipid A (MPL), trehalose dimycolate (TDM), and cell wall skeleton (CWS), preferably MPL + CWS (Detox™); and (8) one or more mineral salts (such as an aluminum salt) + a non-toxic derivative of LPS (such as 3dMPL).

[0116] Other substances that act as immunostimulating agents are disclosed in chapter 7 of ref. 39.

[0117] The use of an aluminium hydroxide or aluminium phosphate adjuvant is particularly preferred, and antigens are generally adsorbed to these salts. Aluminium hydroxide is preferably avoided as an adjuvant if the composition includes a Hib antigen. Where an aluminium phosphate is used and desired not to adsorb an antigen to the adjuvant, this is favoured by including free phosphate ions in solution (e.g. by the use of a phosphate buffer). Prevention of adsorption can also be achieved by selecting the correct pH during antigen/adjuvant mixing, an adjuvant with an appropriate point of zero charge, and an appropriate order of mixing for different antigens in a composition [106].

[0118] Calcium phosphate is another preferred adjuvant.

Further antigens

[0119] Compositions of the invention contain five basic meningococcal protein antigens. They may also include further antigens, although it may contain no meningococcal protein antigens other than the five basic antigens. Further antigens for inclusion may be, for example:

- a saccharide antigen from *Haemophilus influenzae* B.
- a saccharide antigen from *N.meningitidis* serogroup A, C, W135 and/or Y, such as the oligosaccharide disclosed in ref. 107 from serogroup C or the oligosaccharides of ref. 108.
- a saccharide antigen from *Streptococcus pneumoniae* [e.g. 155, 156 157].
- an antigen from hepatitis A virus, such as inactivated virus [e.g. 109, 110].
- an antigen from hepatitis B virus, such as the surface and/or core antigens [e.g. 110, 111].
- a diphtheria antigen, such as a diphtheria toxoid [e.g. chapter 3 of ref. 112] e.g. the CRM₁₉₇ mutant [e.g. 113].
- a tetanus antigen, such as a tetanus toxoid [e.g. chapter 4 of ref. 112].
- an antigen from *Bordetella pertussis*, such as pertussis holotoxin (PT) and filamentous haemagglutinin (FHA) from *B.pertussis*, optionally also in combination with pertactin and/or agglutinogens 2 and 3 [e.g. refs. 114 & 115]. Cellular

pertussis antigen may be used.

- an outer-membrane vesicle (OMV) preparation from *N.meningitidis* serogroup B, such as those disclosed in refs. 4, 116, 117, 118 *etc.*
- polio antigen(s) [e.g. 119, 120] such as OPV or, preferably, IPV.

[0120] The composition may comprise one or more of these further antigens. Antigens will typically be present at a concentration of at least 1 µg/ml each. In general, the concentration of any given antigen will be sufficient to elicit an immune response against that antigen. It is preferred that the protective efficacy of individual saccharide antigens is not removed by combining them, although actual immunogenicity (e.g. ELISA titres) may be reduced.

[0121] Where a diphtheria antigen is included in the composition it is preferred also to include tetanus antigen and pertussis antigens. Similarly, where a tetanus antigen is included it is preferred also to include diphtheria and pertussis antigens. Similarly, where a pertussis antigen is included it is preferred also to include diphtheria and tetanus antigens. Such DTP combinations can be used to reconstitute lyophilised conjugates.

[0122] Where a saccharide or carbohydrate antigen is used, it is preferably conjugated to a carrier protein in order to enhance immunogenicity (see below).

[0123] Toxic protein antigens may be detoxified where necessary (e.g. detoxification of pertussis toxin by chemical and/or genetic means [115]).

[0124] As an alternative to using protein antigens in the composition of the invention, nucleic acid encoding the antigen may be used [e.g. refs. 121 to 129]. Protein components of the compositions of the invention may thus be replaced by nucleic acid (preferably DNA e.g. in the form of a plasmid) that encodes the protein. Similarly, compositions of the invention may comprise proteins which mimic saccharide antigens e.g. mimotopes [130] or anti-idiotypic antibodies. These may replace individual saccharide components, or may supplement them. As an example, the vaccine may comprise a peptide mimic of the MenC [131] or the MenA [132] capsular polysaccharide in place of the saccharide itself.

[0125] Particularly preferred compositions of the invention include one, two or three of: (a) saccharide antigens from meningococcus serogroups Y, W135, C and (optionally) A; (b) a saccharide antigen from *Haemophilus influenzae* type B; and/or (c) an antigen from *Streptococcus pneumoniae*. A composition comprising the serogroup B antigens and a Hib conjugate is particularly preferred.

Meningococcus serogroups Y, W135, C and (optionally) A

[0126] As mentioned above, polysaccharide vaccines against serogroups A, C, W135 & Y has been known for many years. These vaccines (MENGEVAX ACWY™ and MENOMUNE™) are based on the organisms' capsular polysaccharides and, although they are effective in adolescents and adults, they give a poor immune response and short duration of protection, and they cannot be used in infants.

[0127] In contrast to the unconjugated polysaccharide antigens in these vaccines, the recently-approved serogroup C vaccines (Menjugate™ [133,107], Meningitec™ and NeisVac-C™) include conjugated saccharides. Menjugate™ and Meningitec™ have oligosaccharide antigens conjugated to a CRM₁₉₇ carrier, whereas NeisVac-C™ uses the complete polysaccharide (de-O-acetylated) conjugated to a tetanus toxoid carrier.

[0128] Compositions of the present invention preferably include capsular saccharide antigens from one or more of meningococcus serogroups Y, W135, C and (optionally) A, wherein the antigens are conjugated to carrier protein(s) and/or are oligosaccharides.

[0129] A typical quantity of each meningococcal saccharide antigen per dose is between 1 µg and 20 µg e.g. about 1 µg, about 2.5 µg, about 4 µg, about 5 µg, or about 10 µg (expressed as saccharide).

[0130] Where a mixture comprises capsular saccharides from both serogroups A and C, the ratio (w/w) of MenA saccharide:MenC saccharide may be greater than 1 (e.g. 2:1, 3:1, 4:1, 5:1, 10:1 or higher). Where a mixture comprises capsular saccharides from serogroup Y and one or both of serogroups C and W135, the ratio (w/w) of MenY saccharide:MenW135 saccharide may be greater than 1 (e.g. 2:1, 3:1, 4:1, 5:1, 10:1 or higher) and/or that the ratio (w/w) of MenY saccharide:MenC saccharide may be less than 1 (e.g. 1:2, 1:3, 1:4, 1:5, or lower). Preferred ratios (w/w) for saccharides from serogroups A:C:W135:Y are: 1:1:1:1; 1:1:1:2; 2:1:1:1; 4:2:1:1; 8:4:2:1; 4:2:1:2; 8:4:1:2; 4:2:2:1; 2:2:1:1; 4:4:2:1; 2:2:1:2; 4:4:1:2; and 2:2:2:1. Preferred ratios (w/w) for saccharides from serogroups C:W135:Y are: 1:1:1; 1:1:2; 1:1:1; 2:1:1; 4:2:1; 2:1:2; 4:1:2; 2:2:1; and 2:1:1. Using a substantially equal mass of each saccharide is preferred.

[0131] Capsular saccharides will generally be used in the form of oligosaccharides. These are conveniently formed by fragmentation of purified capsular polysaccharide (e.g. by hydrolysis), which will usually be followed by purification of the fragments of the desired size.

[0132] Fragmentation of polysaccharides is preferably performed to give a final average degree of polymerisation (DP) in the oligosaccharide of less than 30 (e.g. between 10 and 20, preferably around 10 for serogroup A; between 15 and 25 for serogroups W135 and Y, preferably around 15-20; between 12 and 22 for serogroup C; *etc.*). DP can conveniently be measured by ion exchange chromatography or by colorimetric assays [134].

[0133] If hydrolysis is performed, the hydrolysate will generally be sized in order to remove short-length oligosaccharides [135]. This can be achieved in various ways, such as ultrafiltration followed by ion-exchange chromatography. Oligosaccharides with a degree of polymerisation of less than or equal to about 6 are preferably removed for serogroup A, and those less than around 4 are preferably removed for serogroups W135 and Y.

[0134] Preferred MenC saccharide antigens are disclosed in reference 133, as used in Menjugate™.

[0135] The saccharide antigen may be chemically modified. This is particularly useful for reducing hydrolysis for serogroup A [136; see below]. De-O-acetylation of meningococcal saccharides can be performed. For oligosaccharides, modification may take place before or after depolymerisation.

[0136] Where a composition of the invention includes a MenA saccharide antigen, the antigen is preferably a modified saccharide in which one or more of the hydroxyl groups on the native saccharide has/have been replaced by a blocking group [136]. This modification improves resistance to hydrolysis, and means that the serogroup A antigen can be stored and used in a liquid formulation rather than requiring lyophilisation.

[0137] The number of monosaccharide units having blocking groups can vary. For example, all or substantially all the monosaccharide units may have blocking groups. Alternatively, at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80% or 90% of the monosaccharide units may have blocking groups. At least 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29 or 30 monosaccharide units may have blocking groups.

[0138] Likewise, the number of blocking groups on a monosaccharide unit may vary. For example, the number of blocking groups on a monosaccharide unit may be 1 or 2. The blocking group will generally be at the 4 position and/or 3-position of the monosaccharide units.

[0139] The terminal monosaccharide unit may or may not have a blocking group instead of its native hydroxyl. It is preferred to retain a free anomeric hydroxyl group on a terminal monosaccharide unit in order to provide a handle for further reactions (e.g. conjugation). Anomeric hydroxyl groups can be converted to amino groups (-NH₂ or -NH-E, where E is a nitrogen protecting group) by reductive amination (using, for example, NaBH₃CN/NH₄Cl), and can then be regenerated after other hydroxyl groups have been converted to blocking groups.

[0140] Blocking groups to replace hydroxyl groups may be directly accessible via a derivatizing reaction of the hydroxyl group *i.e.* by replacing the hydrogen atom of the hydroxyl group with another group. Suitable derivatives of hydroxyl groups which act as blocking groups are, for example, carbamates, sulfonates, carbonates, esters, ethers (e.g. silyl ethers or alkyl ethers) and acetals. Some specific examples of such blocking groups are allyl, Aloc, benzyl, BOM, t-butyl, trityl, TBS, TBDPS, TES, TMS, TIPS, PMB, MEM, MOM, MTM, THP, *etc.* Other blocking groups that are not directly accessible and which completely replace the hydroxyl group include C₁₋₁₂ alkyl, C₃₋₁₂ alkyl, C₅₋₁₂ aryl, C₅₋₁₂ aryl-C₁₋₆ alkyl, NR¹R² (R¹ and R² are defined in the following paragraph), H, F, Cl, Br, CO₂H, CO₂(C₁₋₆ alkyl), CN, CF₃, CCl₃, *etc.* Preferred blocking groups are electron-withdrawing groups.

[0141] Preferred blocking groups are of the formula: -O-X-Y or -OR³ wherein: X is C(O), S(O) or SO₂; Y is C₁₋₁₂ alkyl, C₁₋₁₂ alkoxy, C₃₋₁₂ cycloalkyl, C₅₋₁₂ aryl or C₅₋₁₂ aryl-C₁₋₆ alkyl, each of which may optionally be substituted with 1, 2 or 3 groups independently selected from F, Cl, Br, CO₂H, CO₂(C₁₋₆ alkyl), CN, CF₃ or CCl₃; or Y is NR¹R²; R¹ and R² are independently selected from H, C₁₋₁₂ alkyl, C₃₋₁₂ cycloalkyl, C₅₋₁₂ aryl, C₅₋₁₂ aryl-C₁₋₆ alkyl; or R¹ and R² may be joined to form a C₃₋₁₂ saturated heterocyclic group; R³ is C₁₋₁₂ alkyl or C₃₋₁₂ cycloalkyl, each of which may optionally be substituted with 1, 2 or 3 groups independently selected from F, Cl, Br, CO₂(C₁₋₆ alkyl), CN, CF₃ or CCl₃; or R³ is C₅₋₁₂ aryl or C₅₋₁₂ aryl-C₁₋₆ alkyl, each of which may optionally be substituted with 1, 2, 3, 4 or 5 groups selected from F, Cl, Br, CO₂H, CO₂(C₁₋₆ alkyl), CN, CF₃ or CCl₃. When R³ is C₁₋₁₂ alkyl or C₃₋₁₂ cycloalkyl, it is typically substituted with 1, 2 or 3 groups as defined above. When R¹ and R² are joined to form a C₃₋₁₂ saturated heterocyclic group, it is meant that R¹ and R² together with the nitrogen atom form a saturated heterocyclic group containing any number of carbon atoms between 3 and 12 (e.g. C₃, C₄, C₅, C₆, C₇, C₈, C₉, C₁₀, C₁₁, C₁₂). The heterocyclic group may contain 1 or 2 heteroatoms (such as N, O or S) other than the nitrogen atom. Examples of C₃₋₁₂ saturated heterocyclic groups are pyrrolidinyl, piperidinyl, morpholinyl, piperazinyl, imidazolidinyl, azetidiny and aziridinyl.

[0142] Blocking groups -O-X-Y and -OR³ can be prepared from -OH groups by standard derivatizing procedures, such as reaction of the hydroxyl group with an acyl halide, alkyl halide, sulfonyl halide, *etc.* Hence, the oxygen atom in -O-X-Y is preferably the oxygen atom of the hydroxyl group, while the -X-Y group in -O-X-Y preferably replaces the hydrogen atom of the hydroxyl group.

[0143] Alternatively, the blocking groups may be accessible via a substitution reaction, such as a Mitsunobu-type substitution. These and other methods of preparing blocking groups from hydroxyl groups are well known.

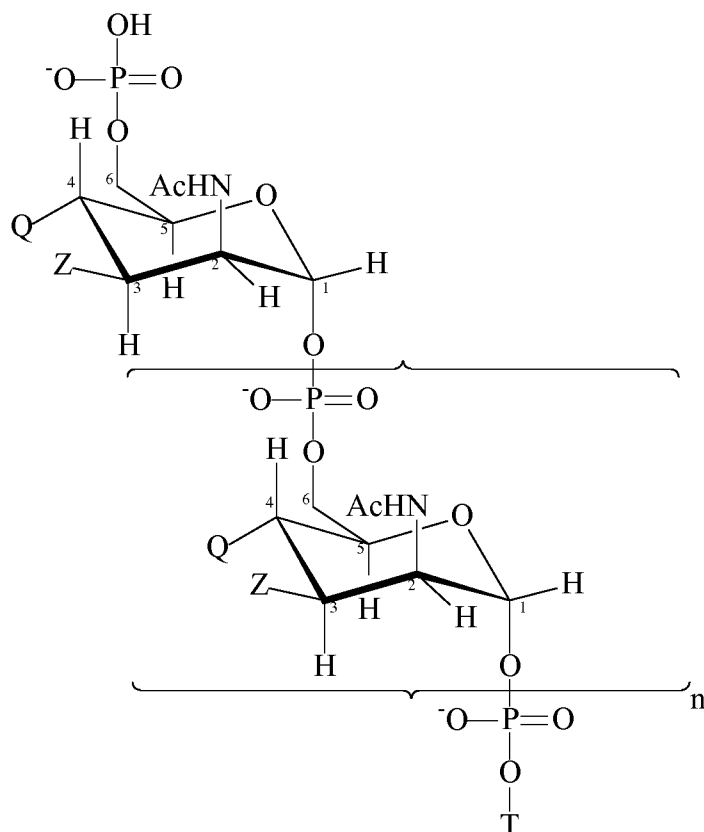
[0144] More preferably, the blocking group is -OC(O)CF₃ [137], or a carbamate group -OC(O)NR¹R², where R¹ and R² are independently selected from C₁₋₆ alkyl. More preferably, R¹ and R² are both methyl *i.e.* the blocking group is -OC(O)NMe₂. Carbamate blocking groups have a stabilizing effect on the glycosidic bond and may be prepared under mild conditions.

[0145] Preferred modified MenA saccharides contain *n* monosaccharide units, where at least *h*% of the monosaccharide units do not have -OH groups at both of positions 3 and 4. The value of *h* is 24 or more (e.g. 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 55, 60, 65, 70, 75, 80, 85, 90, 95, 98, 99 or 100) and is preferably 50 or more. The absent -OH groups are

preferably blocking groups as defined above.

[0146] Other preferred modified MenA saccharides comprise monosaccharide units, wherein at least *s* of the monosaccharide units do not have -OH at the 3 position and do not have -OH at the 4 position. The value of *s* is at least 1 (e.g. 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, 16, 17, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 35, 40, 45, 50, 60, 70, 80, 90). The absent -OH groups are preferably blocking groups as defined above.

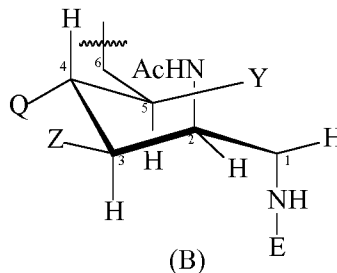
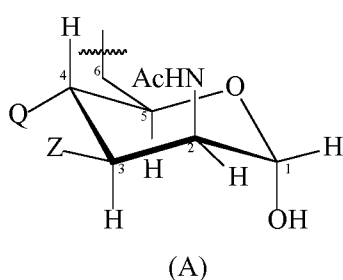
[0147] Suitable modified MenA saccharides for use with the invention have the formula:



wherein

n is an integer from 1 to 100 (preferably an integer from 15 to 25);

T is of the formula (A) or (B):



each Z group is independently selected from OH or a blocking group as defined above; and each Q group is independently selected from OH or a blocking group as defined above;

Y is selected from OH or a blocking group as defined above;

E is H or a nitrogen protecting group;

and wherein more than about 7% (e.g. 8%, 9%, 10% or more) of the Q groups are blocking groups.

[0148] Each of the $n+2$ Z groups may be the same or different from each other. Likewise, each of the $n+2$ Q groups may be the same or different from each other. All the Z groups may be OH. Alternatively, at least 10%, 20, 30%, 40%, 50% or 60% of the Z groups may be OAc. Preferably, about 70% of the Z groups are OAc, with the remainder of the Z groups being OH or blocking groups as defined above. At least about 7% of Q groups are blocking groups. Preferably, at least 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% or even 100% of the Q groups are blocking groups.

[0149] Preferred compositions of the invention can be stored for 28 days at 37°C and, after that period, less than $f\%$ of the initial total amount of conjugated MenA saccharide will be unconjugated, where f is 20, 19, 18, 17, 16, 15, 14, 13, 12, 11, 10, 9, 8, 7, 6, 5 or lower.

[0150] Meningococcal capsular polysaccharides are typically prepared by a process comprising the steps of polysaccharide precipitation (e.g. using a cationic detergent), ethanol fractionation, cold phenol extraction (to remove protein) and ultracentrifugation (to remove LPS) [e.g. ref. 138]. A more preferred process [108], however, involves polysaccharide precipitation followed by solubilisation of the precipitated polysaccharide using a lower alcohol. Precipitation can be achieved using a cationic detergent such as tetrabutylammonium and cetyltrimethylammonium salts (e.g. the bromide salts), or hexadimethrine bromide and myristyltrimethylammonium salts. Cetyltrimethylammonium bromide ('CTAB') is particularly preferred [139]. Solubilisation of the precipitated material can be achieved using a lower alcohol such as methanol, propan-1-ol, propan-2-ol, butan-1-ol, butan-2-ol, 2-methyl-propan-1-ol, 2-methyl-propan-2-ol, diols, etc., but ethanol is particularly suitable for solubilising CTAB-polysaccharide complexes. Ethanol is preferably added to the precipitated polysaccharide to give a final concentration (based on total content of ethanol and water) of between 50% and 95%.

[0151] After re-solubilisation, the polysaccharide may be further treated to remove contaminants. This is particularly important in situations where even minor contamination is not acceptable (e.g. for human vaccine production). This will typically involve one or more steps of filtration e.g. depth filtration, filtration through activated carbon may be used, size filtration and/or ultrafiltration. Once filtered to remove contaminants, the polysaccharide may be precipitated for further treatment and/or processing. This can be conveniently achieved by exchanging cations (e.g. by the addition of calcium or sodium salts).

[0152] As an alternative to purification, capsular saccharides of the present invention may be obtained by total or partial synthesis e.g. Hib synthesis is disclosed in ref. 140, and MenA synthesis in ref. 141.

[0153] Compositions of the invention comprise capsular saccharides from at least two serogroups of *N.meningitidis*. The saccharides are preferably prepared separately (including any fragmentation, conjugation, modification, etc.) and then admixed to give a composition of the invention.

[0154] Where the composition comprises capsular saccharide from serogroup A, however, it is preferred that the serogroup A saccharide is not combined with the other saccharide(s) until shortly before use, in order to minimise the potential for hydrolysis. This can conveniently be achieved by having the serogroup A component (typically together with appropriate excipients) in lyophilised form and the other serogroup component(s) in liquid form (also with appropriate excipients), with the liquid components being used to reconstitute the lyophilised MenA component when ready for use. Where an aluminium salt adjuvant is used, it is preferred to include the adjuvant in the vial containing the with the liquid vaccine, and to lyophilise the MenA component without adjuvant.

[0155] A composition of the invention may thus be prepared from a kit comprising: (a) capsular saccharide from *N.meningitidis* serogroup A, in lyophilised form; and (b) the further antigens from the composition, in liquid form.

[0156] Within each dose, the amount of an individual saccharide antigen will generally be between 1-50 μg (measured as mass of saccharide), with about 2.5 μg , 5 μg or 10 μg of each being preferred. With A:C:W135:Y weight ratios of 1:1:1:1; 1:1:1:2; 2:1:1:1; 4:2:1:1; 8:4:2:1; 4:2:1:2; 8:4:1:2; 4:2:2:1; 2:2:1:1; 4:4:2:1; 2:2:1:2; 4:4:1:2; and 2:2:2:1, therefore, the amount represented by the figure 1 is preferably about 2.5 μg , 5 μg or 10 μg . For a 1:1:1:1 ratio A:C:W:Y composition and a 10 μg per saccharide, therefore, 40 μg saccharide is administered per dose. Preferred compositions have about the following μg saccharide per dose:

A	10	0	0	0	10	5	2.5
C	10	10	5	2.5	5	5	2.5
W135	10	10	5	2.5	5	5	2.5
Y	10	10	5	2.5	5	5	2.5

Preferred compositions of the invention comprise less than 50 μg meningococcal saccharide per dose. Other preferred compositions comprise ≤ 40 μg meningococcal saccharide per dose. Other preferred compositions comprise ≤ 30 μg meningococcal saccharide per dose. Other preferred compositions comprise ≤ 25 μg meningococcal saccharide per dose. Other preferred compositions comprise ≤ 20 μg meningococcal saccharide per dose. Other preferred compositions

comprise ≤ 10 μg meningococcal saccharide per dose but, ideally, compositions of the invention comprise at least 10 μg meningococcal saccharide per dose.

[0157] The Menjugate™ and NeisVac™ MenC conjugates use a hydroxide adjuvant, whereas Meningitec™ uses a phosphate. It is possible in compositions of the invention to adsorb some antigens to an aluminium hydroxide but to have other antigens in association with an aluminium phosphate. For tetravalent serogroup combinations, for example, the following permutations are available:

Serogroup	Aluminium salt (H = a hydroxide; P = a phosphate)														
A	P	H	P	H	H	H	P	P	P	H	H	H	P	P	H
C	P	H	H	P	H	H	P	H	H	P	P	H	P	H	P
W135	P	H	H	H	P	H	H	P	H	H	P	P	P	P	H
Y	P	H	H	H	H	P	H	H	P	P	H	P	H	P	P

[0158] For trivalent *N.meningitidis* serogroup combinations, the following permutations are available:

Serogroup	Aluminium salt (H = a hydroxide; P = a phosphate)							
C	P	H	H	H	P	P	P	H
W135	P	H	H	P	H	P	H	P
Y	P	H	P	H	H	H	P	P

Haemophilus influenzae type B

[0159] Where the composition includes a *H.influenzae* type B antigen, it will typically be a Hib capsular saccharide antigen. Saccharide antigens from *H.influenzae* b are well known.

[0160] Advantageously, the Hib saccharide is covalently conjugated to a carrier protein, in order to enhance its immunogenicity, especially in children. The preparation of polysaccharide conjugates in general, and of the Hib capsular polysaccharide in particular, is well documented [e.g. references 142 to 150 etc.]. The invention may use any suitable Hib conjugate. Suitable carrier proteins are described below, and preferred carriers for Hib saccharides are CRM₁₉₇ ('HbOC'), tetanus toxoid ('PRP-T') and the outer membrane complex of *N.meningitidis* ('PRP-OMP').

[0161] The saccharide moiety of the conjugate may be a polysaccharide (e.g. full-length polyribosylribitol phosphate (PRP)), but it is preferred to hydrolyse polysaccharides to form oligosaccharides (e.g. MW from ~1 to ~5 kDa).

[0162] A preferred conjugate comprises a Hib oligosaccharide covalently linked to CRM₁₉₇ via an adipic acid linker [151, 152]. Tetanus toxoid is also a preferred carrier.

[0163] Administration of the Hib antigen preferably results in an anti-PRP antibody concentration of $\geq 0.15 \mu\text{g/ml}$, and more preferably $\geq 1 \mu\text{g/ml}$.

[0164] Compositions of the invention may comprise more than one Hib antigen.

[0165] Where a composition includes a Hib saccharide antigen, it is preferred that it does not also include an aluminium hydroxide adjuvant. If the composition includes an aluminium phosphate adjuvant then the Hib antigen may be adsorbed to the adjuvant [153] or it may be non-adsorbed [154].

[0166] Hib antigens may be lyophilised e.g. together with meningococcal antigens.

Streptococcus pneumoniae

[0167] Where the composition includes a *S.pneumoniae* antigen, it will typically be a capsular saccharide antigen which is preferably conjugated to a carrier protein [e.g. refs. 155 to 157]. It is preferred to include saccharides from more than one serotype of *S.pneumoniae*. For example, mixtures of polysaccharides from 23 different serotype are widely used, as are conjugate vaccines with polysaccharides from between 5 and 11 different serotypes [158]. For example, PreVNar™ [159] contains antigens from seven serotypes (4, 6B, 9V, 14, 18C, 19F, and 23F) with each saccharide individually conjugated to CRM₁₉₇ by reductive amination, with 2 μg of each saccharide per 0.5ml dose (4 μg of serotype 6B), and with conjugates adsorbed on an aluminium phosphate adjuvant. Compositions of the invention preferably include at least serotypes 6B, 14, 19F and 23F. Conjugates may be adsorbed onto an aluminium phosphate.

[0168] As an alternative to using saccharide antigens from pneumococcus, the composition may include one or more polypeptide antigens. Genome sequences for several strains of pneumococcus are available [160,161] and can be

subjected to reverse vaccinology [162-165] to identify suitable polypeptide antigens [166,167]. For example, the composition may include one or more of the following antigens: PhtA, PhtD, PhtB, PhtE, SpsA, LytB, LytC, LytA, Sp125, Sp101, Sp128, Sp130 and Sp130, as defined in reference 168. The composition may include more than one (e.g. 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13 or 14) of these antigens.

[0169] In some embodiments, the composition may include both saccharide and polypeptide antigens from pneumococcus. These may be used in simple admixture, or the pneumococcal saccharide antigen may be conjugated to a pneumococcal protein. Suitable carrier proteins for such embodiments include the antigens listed in the previous paragraph [168].

[0170] Pneumococcal antigens may be lyophilised e.g. together with meningococcal and/or Hib antigens.

Covalent conjugation

[0171] Capsular saccharides in compositions of the invention will usually be conjugated to carrier protein(s). In general, conjugation enhances the immunogenicity of saccharides as it converts them from T-independent antigens to T-dependent antigens, thus allowing priming for immunological memory. Conjugation is particularly useful for paediatric vaccines and is a well known technique [e.g. reviewed in refs. 169 and 142-150].

[0172] Preferred carrier proteins are bacterial toxins or toxoids, such as diphtheria toxoid or tetanus toxoid. The CRM₁₉₇ diphtheria toxoid [170-172] is particularly preferred. Other suitable carrier proteins include the *N.meningitidis* outer membrane protein [173], synthetic peptides [174,175], heat shock proteins [176,177], pertussis proteins [178,179], cytokines [180], lymphokines [180], hormones [180], growth factors [180], artificial proteins comprising multiple human CD4⁺ T cell epitopes from various pathogen-derived antigens [181], protein D from *H.influenzae* [182,183], pneumococcal surface protein PspA [184], iron-uptake proteins [185], toxin A or B from *C.difficile* [186], etc. Preferred carriers are diphtheria toxoid, tetanus toxoid, *H.influenzae* protein D, and CRM₁₉₇.

[0173] Within a composition of the invention, it is possible to use more than one carrier protein e.g. to reduce the risk of carrier suppression. Thus different carrier proteins can be used for different serogroups e.g. serogroup A saccharides might be conjugated to CRM₁₉₇ while serogroup C saccharides might be conjugated to tetanus toxoid. It is also possible to use more than one carrier protein for a particular saccharide antigen e.g. serogroup A saccharides might be in two groups, with some conjugated to CRM₁₉₇ and others conjugated to tetanus toxoid. In general, however, it is preferred to use the same carrier protein for all saccharides.

[0174] A single carrier protein might carry more than one saccharide antigen [187]. For example, a single carrier protein might have conjugated to it saccharides from serogroups A and C. To achieve this goal, saccharides can be mixed prior to the conjugation reaction. In general, however, it is preferred to have separate conjugates for each serogroup.

[0175] Conjugates with a saccharide:protein ratio (w/w) of between 1:5 (*i. e.* excess protein) and 5:1 (*i. e.* excess saccharide) are preferred. Ratios between 1:2 and 5:1 are preferred, as are ratios between 1:1.25 and 1:2.5 are more preferred. Excess carrier protein may be preferred for MenA and MenC.

[0176] Conjugates may be used in conjunction with free carrier protein [188]. When a given carrier protein is present in both free and conjugated form in a composition of the invention, the unconjugated form is preferably no more than 5% of the total amount of the carrier protein in the composition as a whole, and more preferably present at less than 2% by weight.

[0177] Any suitable conjugation reaction can be used, with any suitable linker where necessary.

[0178] The saccharide will typically be activated or functionalised prior to conjugation. Activation may involve, for example, cyanylating reagents such as CDAP (e.g. 1-cyano-4-dimethylamino pyridinium tetrafluoroborate [189,190,etc.]). Other suitable techniques use carbodiimides, hydrazides, active esters, norborane, p-nitrobenzoic acid, N-hydroxysuccinimide, S-NHS, EDC, TSTU; see also the introduction to reference 148).

[0179] Linkages via a linker group may be made using any known procedure, for example, the procedures described in references 191 and 192. One type of linkage involves reductive amination of the polysaccharide, coupling the resulting amino group with one end of an adipic acid linker group, and then coupling a protein to the other end of the adipic acid linker group [146,193,194]. Other linkers include B-propionamido [195], nitrophenyl-ethylamine [196], haloacyl halides [197], glycosidic linkages [198], 6-aminocaproic acid [199], ADH [200], C₄ to C₁₂ moieties [201] etc. As an alternative to using a linker, direct linkage can be used. Direct linkages to the protein may comprise oxidation of the polysaccharide followed by reductive amination with the protein, as described in, for example, references 202 and 203.

[0180] A process involving the introduction of amino groups into the saccharide (e.g. by replacing terminal =O groups with -NH₂) followed by derivatisation with an adipic diester (e.g. adipic acid N-hydroxysuccinimido diester) and reaction with carrier protein is preferred. Another preferred reaction uses CDAP activation with a protein D carrier e.g. for MenA or MenC.

[0181] After conjugation, free and conjugated saccharides can be separated. There are many suitable methods, including hydrophobic chromatography, tangential ultrafiltration, diafiltration etc. [see also refs. 204 & 205, etc.].

[0182] Where the composition of the invention includes a conjugated oligosaccharide, it is preferred that oligosaccha-

ride preparation precedes conjugation.

Further and alternative serogroup B polypeptides antigens

[0183] The invention provides a composition as defined in the claims which, after administration to a subject, is able to induce an antibody response in that subject, wherein the antibody response is bactericidal against two or three of hypervirulent lineages A4, ET-5 and lineage 3 of *N.meningitidis* serogroup B.

[0184] Although NadA, 741, 936, 953 and 287 are preferred antigens for achieving this broad protection, other MenB polypeptide antigens which may be included in compositions of the invention (in combination with the five basic antigens as defined in the claims) include those comprising one of the following amino acid sequences: SEQ ID NO:650 from ref. 8; SEQ ID NO:878 from ref. 8; SEQ ID NO:884 from ref. 8; SEQ ID NO:4 from ref. 9; SEQ ID NO:598 from ref. 10; SEQ ID NO:818 from ref. 10; SEQ ID NO:864 from ref. 10; SEQ ID NO:866 from ref. 10; SEQ ID NO:1196 from ref. 10; SEQ ID NO:1272 from ref. 10; SEQ ID NO:1274 from ref. 10; SEQ ID NO:1640 from ref. 10; SEQ ID NO:1788 from ref. 10; SEQ ID NO:2288 from ref. 10; SEQ ID NO:2466 from ref. 10; SEQ ID NO:2554 from ref. 10; SEQ ID NO:2576 from ref. 10; SEQ ID NO:2606 from ref. 10; SEQ ID NO:2608 from ref. 10; SEQ ID NO:2616 from ref. 10; SEQ ID NO:2668 from ref. 10; SEQ ID NO:2780 from ref. 10; SEQ ID NO:2932 from ref. 10; SEQ ID NO:2958 from ref. 10; SEQ ID NO:2970 from ref. 10; SEQ ID NO:2988 from ref. 10, or a polypeptide comprising an amino acid sequence which: (a) has 50% or more identity (e.g. 60%, 70%, 80%, 90%, 95%, 99% or more) to said sequences; and/or (b) comprises a fragment of at least *n* consecutive amino acids from said sequences, wherein *n* is 7 or more (eg. 8, 10, 12, 14, 16, 18, 20, 25, 30, 35, 40, 50, 60, 70, 80, 90, 100, 150, 200, 250 or more). Preferred fragments for (b) comprise an epitope from the relevant sequence. More than one (e.g. 2, 3, 4, 5, 6) of these polypeptides may be included.

General

[0185] The term "comprising" means "including" as well as "consisting" e.g. a composition "comprising" X may consist exclusively of X or may include something additional e.g. X + Y.

[0186] The term "about" in relation to a numerical value *x* means, for example, $x \pm 10\%$.

[0187] The word "substantially" does not exclude "completely" e.g. a composition which is "substantially free" from Y may be completely free from Y. Where necessary, the word "substantially" may be omitted from the definition of the invention.

[0188] References to a percentage sequence identity between two amino acid sequences means that, when aligned, that percentage of amino acids are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in section 7.7.18 of reference 206. A preferred alignment is determined by the Smith-Waterman homology search algorithm using an affine gap search with a gap open penalty of 12 and a gap extension penalty of 2, BLOSUM matrix of 62. The Smith-Waterman homology search algorithm is taught in reference 207.

[0189] The term "alkyl" refers to alkyl groups in both straight and branched forms. The alkyl group may be interrupted with 1, 2 or 3 heteroatoms selected from -O-, -NH- or -S-. The alkyl group may also be interrupted with 1, 2 or 3 double and/or triple bonds. However, the term "alkyl" usually refers to alkyl groups having no heteroatom interruptions or double or triple bond interruptions. Where reference is made to C₁₋₁₂ alkyl, it is meant the alkyl group may contain any number of carbon atoms between 1 and 12 (e.g. C₁, C₂, C₃, C₄, C₅, C₆, C₇, C₈, C₉, C₁₀, C₁₁, C₁₂). Similarly, where reference is made to C₁₋₆ alkyl, it is meant the alkyl group may contain any number of carbon atoms between 1 and 6 (e.g. C₁, C₂, C₃, C₄, C₅, C₆).

[0190] The term "cycloalkyl" includes cycloalkyl, polycycloalkyl, and cycloalkenyl groups, as well as combinations of these with alkyl groups, such as cycloalkylalkyl groups. The cycloalkyl group may be interrupted with 1, 2 or 3 heteroatoms selected from -O-, -NH- or -S-. However, the term "cycloalkyl" usually refers to cycloalkyl groups having no heteroatom interruptions. Examples of cycloalkyl groups include cyclopentyl, cyclohexyl, cyclohexenyl, cyclohexylmethyl and adamantyl groups. Where reference is made to C₃₋₁₂ cycloalkyl, it is meant that the cycloalkyl group may contain any number of carbon atoms between 3 and 12 (e.g. C₃, C₄, C₅, C₆, C₇, C₈, C₉, C₁₀, C₁₁, C₁₂).

[0191] The term "aryl" refers to an aromatic group, such as phenyl or naphthyl. Where reference is made to C₅₋₁₂ aryl, it is meant that the aryl group may contain any number of carbon atoms between 5 and 12 (e.g. C₅, C₆, C₇, C₈, C₉, C₁₀, C₁₁, C₁₂).

[0192] The term "C₅₋₁₂ aryl-C₁₋₆ alkyl" refers to groups such as benzyl, phenylethyl and naphthylmethyl.

[0193] Nitrogen protecting groups include silyl groups (such as TMS, TES, TBS, TIPS), acyl derivatives (such as phthalimides, trifluoroacetamides, methoxycarbonyl, ethoxycarbonyl, t-butoxycarbonyl (Boc), benzyloxycarbonyl (Z or Cbz), 9-fluorenylmethoxycarbonyl (Fmoc), 2-(trimethylsilyl)ethoxy carbonyl, 2,2,2-trichloroethoxycarbonyl (Troc)), sulfonfyl derivatives (such as β -trimethylsilyl ethanesulfonyl (SES)), sulfonyl derivatives, C₁₋₁₂ alkyl, benzyl, benzhydryl, trityl, 9-phenylfluorenyl etc. A preferred nitrogen protecting group is Fmoc.

[0194] Sequences included to facilitate cloning or purification, *etc.*, do not necessarily contribute to the invention and may be omitted or removed.

[0195] It will be appreciated that sugar rings can exist in open and closed form and that, whilst closed forms are shown in structural formulae herein, open forms are also encompassed by the invention.

MODES FOR CARRYING OUT THE INVENTION

ΔG287-953 hybrid protein

[0196] DNA encoding protein 287 from meningococcal serogroup B strain 394/98 and protein 953 from meningococcal serogroup B strain 2996 were digested and ligated, together with a short linker sequence, to give a plasmid encoding amino acid sequence SEQ ID 7. The plasmid was transfected into *E.coli* and bacteria were grown to express the protein.

[0197] After adequate growth, bacteria were harvested and the protein was purified. From culture, bacteria were centrifuged and the pellet was homogenized in the presence of 50 mM acetate buffer (pH 5) with a pellet:buffer volume ratio of 1: 8. Lysis was performed using a high pressure homogenizer (AVESTIN, 4 cycles at 14000 psi). After lysis, urea was added at final concentration of 5M, followed by agitation for 1 hour at room temperature. The pH was reduced from 6 to 5 using 200 mM acetate buffer (pH 4) + 5 M urea. The mixture was centrifuged at 16800g for 60 minutes at 2-8°C. The supernatant was collected and filtered by SARTOBRA P (0.45-0.22μm SARTORIUS).

[0198] Protein in the filtered supernatant was stable for ≥30 days at -20°C and for ≥15 days at 2-8°C.

[0199] Protein was further purified on a cationic exchange column (SPFF, Amersham Biosciences) with elution using 350mM NaCl + 50 mM acetate + 5 M urea pH 5.00. The majority of impurities were present in the flow-thru. A pre-elution washing using a lower NaCl concentration (180 mM) advantageously eliminated two contaminating *E.coli* proteins.

[0200] The eluted material was adjusted to pH 8 (using 200 mM TRIS/HCl + 5 M urea pH 9) and further purified on a Q Sepharose HP column (Amersham) with elution using 150 mM NaCl + 20 mM TRIS/HCl pH 8.00 in 5 M urea. Again, a pre-elution washing with reduced salt (90 mM) was useful for eliminating impurities.

[0201] The filtered eluted material from Q HP column was diluted 1:2 using PBS pH 7.00 (150 mM NaCl + 10 mM potassium phosphate, pH 7.00) and then diafiltered against 10 volumes of PBS pH 7.00 by tangential ultrafiltration. At the end of diafiltration the material was concentrated 1.6 times to about 1.2 mg/ml total proteins. Using a 30,000 Da cut-off membrane (Regenerated Cellulose membrane 50cm², Millipore PLCTK 30) it was possible to dialyze the material with a yield of about 90% .

936-ΔG741 hybrid protein

[0202] DNA encoding protein 936 from meningococcal serogroup B strain 2996 and protein 741 from meningococcal serogroup B strain MC58 were digested and ligated, together with a short linker sequence, to give a plasmid encoding amino acid sequence SEQ ID 8. The plasmid was transfected into *E.coli* and bacteria were grown to express the protein. The recombinant protein was not secreted, but remained soluble within the bacteria.

[0203] After adequate growth, bacteria were centrifuged to give a humid paste and treated as follows:

- Homogenisation by high pressure system in presence of 20mM sodium phosphate pH 7.00.
- Centrifugation and clarification by orthogonal filtration.
- Cationic column chromatography (SP Sepharose Fast Flow), with elution by 150mM NaCl in 20mM sodium phosphate pH 7.00.
- Anionic column chromatography (Q Sepharose XL) with flow-through harvesting.
- Hydrophobic column chromatography (Phenyl Sepharose 6 Fast Flow High Sub) with elution by 20mM sodium phosphate, pH 7.00.
- Diafiltration against PBS pH 7.4 with a 10Kd cut-off.
- Final sterile filtration and storing at -20°C

[0204] Protein in the final material was stable for at least 3 months both at -20°C and at 2-8°C.

NadA^{(NL)(C)} protein

[0205] DNA encoding NadA protein from meningococcal serogroup B strain 2996 was digested to remove the sequence encoding its C-terminus, to give a plasmid encoding amino acid sequence SEQ ID 1. The plasmid was transfected into *E.coli* and bacteria were grown to express the protein. The recombinant protein was secreted into the culture medium, and the leader peptide was absent in the secreted protein (SEQ ID 2). The supernatant was treated as follows:

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- Concentration 7X and diafiltration against buffer 20mM TRIS/HCl pH7.6 by cross flow UF (Cut off 30Kd).
- Anionic column chromatography (Q Sepharose XL), with elution by 400mM NaCl in 20mM TRIS/HCl pH 7.6.
- Hydrophobic column chromatography step (Phenyl Sepharose 6 Fast Flow High Sub), with elution by 50mM NaCl in TRIS/HCl pH 7.6.
- Hydroxylapatite ceramic column chromatography (HA Macro. Prep) with elution by 200mM sodium phosphate pH 7.4.
- Diafiltration (cut off 30Kd) against PBS pH 7.4
- Final sterile filtration and storing at -20°C

[0206] Protein in the final material was stable for at least 6 months both at -20°C and at 2-8°C.

[0207] NadA protein is susceptible to degradation, and truncated forms of NadA may be detected by western blot or by mass spectrometry (e.g. by MALDI-TOF) indicating up to 10kDa MW loss. Degradation products can be separated from native NadA by gel filtration (e.g. using column TSK 300SWXL, precolumn TSKSWXL, TOSOHAAS). Such filtration gives three peaks: (i) a first peak with retention time 12.637 min and apparent MW 885.036 Da; (ii) retention time 13.871 min and apparent MW 530.388 Da; (iii) retention time 13.871 min and apparent MW 530.388 Da. Light scattering analysis of the three peaks reveals real MW values of (i) 208500 Da, (ii) 98460 Da, (iii) 78760 Da. Thus the first peak contains NadA aggregates, and the third peak contains degradation products.

[0208] As the predicted molecular weight of NadA^{(NL)(C)} is 34.113 Da, peak (ii) contains a trimeric protein, which is the desired antigen.

Antigenic combinations

[0209] Mice were immunised with a composition comprising the three proteins and an aluminium hydroxide adjuvant. For comparison purposes, the three proteins were also tested singly. Ten mice were used per group. The mixture was able to induce high bactericidal titres against various strains:

	Meningococcal strain (Serogroup)							
	2996 (B)	MC58 (B)	NGH38	394/98 (B)	H44/76 (B)	F6124 (A)	BZ133 (C)	C11 (C)
(1)	32000	16000	130000	16000	32000	8000	16000	8000
(2)	256	131000	128	16000	32000	8000	16000	<4
(3)	32000	8000	-	-	-	8000	-	32000
Mix	32000	32000	65000	16000	260000	65000	>65000	8000

'-' indicates that this strain contains no NadA gene Looking at individual mice, the triple mixture induced high and consistent bactericidal titres against the three serogroup B strains from which the individual antigens are derived:

#	1	2	3	4	5	6	7	8	9	10
2996	32768	16384	65536	32768	32768	65536	65536	32768	65536	8192
MC58	65536	32768	65536	65536	65536	8192	65536	32768	32768	65536
394/98	65536	4096	16384	4096	8192	4096	32768	16384	8192	16384

Combination and comparison with OMVs

[0210] In further experiments, the adjuvanted antigens (20µg of each antigen per dose) were administered in combination with 10µg OMVs prepared either from strain H44/76 (Norway) or strain 394/98 (New Zealand). Positive controls were the anti-capsular SEAM-3 mAb for serogroup B or CRM197-conjugated capsular saccharides for other strains. Results (bactericidal titres) are shown in Table 1. The mixture almost always gives better titres than simple OMVs and, furthermore, the addition of the mixture to OMVs almost always significantly enhances the efficacy of the OMVs. Moreover, in many cases the antigen mixture matches or exceeds the response seen with the positive control.

Hypervirulent lineage tests

[0211] The following antigens were tested against a variety of serogroup B strains from a variety of hypervirulent

lineages:

- (a) NadA^{(NL)(C)}
- (b) ΔG287-953
- (c) 936-ΔG741
- (d) a mixture of (a), (b) and (c)
- (e) OMVs prepared from strain H44/76 (Norway)
- (f) OMVs prepared from strain 394/98 (New Zealand)
- (g) A mixture of ΔG287 and (e)
- (h) A mixture of (d) and (e)
- (i) A mixture of (d) and (f)

SEAM-3 was used as a positive control.

[0212] Results were as follows, expressed as the percentage of strains in the indicated hypervirulent lineage where the serum bactericidal titre exceeded 1024:

	# strains	(a)	(b)	(c)	(d)	(e)	(f)	(g)	(h)	(i)	S-3
A4	4	50	50	0	100	25	25	25	100	100	+
ET-5	8	25	75	88	100	71	14	71	100	100	+
Lineage 3	13	0	75	15	93	8	85	8	92	93	+
ET-37	4	11	22	0	33	0	0	0	22	25	+

[0213] Against particular reference strains, bactericidal titres were as follows:

	Strain	(a)	(b)	(c)	(d)	(e)	(f)	(g)	(h)	(i)	S-3
A4	961-5945	128	2048	<8	2048	262144	8192	262144	262144	4096	8192
ET-5	44/76	<4	2048	32768	131072	524288	8192	524288	524288	524288	16384
Lineage 3	394/98	<4	1024	32	4096	<4	16384	256	16384	16384	16384
ET-37	LPN17592	2048	1024	256	4096	<8	<8	512	16384	65536	1024

[0214] Compositions (d), (h) and (i) therefore induce bactericidal antibody responses against a wide variety of strains of serogroup B meningococcus from within hypervirulent lineages A4, ET-5 and lineage 3. Titres using compositions (h) and (i) were generally higher than with (d), but the coverage of strains within hypervirulent lineages A4, ET-5 and lineage 3 were no better.

[0215] Coverage of untyped strains was also high with compositions (d), (h) and (i).

Analysis of NadA N-terminus domain

[0216] Purified *N.meningitidis* NadA protein is known to bind to human epithelial cells [17] (e.g. Chang cells, HeLa cells, Hep-2 cells), and recombinant *E.coli* which express NadA display an adherent phenotype [18]. These *E.coli* are also able to invade epithelial cells, and intracellular NadA⁺ *E.coli* can be detected in Chang cells by immunofluorescence (after membrane permeabilisation) and by electron microscopy. NadA is thus believed function as an adhesin and an invasin for epithelial cells.

[0217] On the basis of secondary structure analysis, mature NadA has been subdivided into three putative domains: a N-terminal globular domain (aa 24-87), an α -helix internal region (aa 88-350) with high coiled-coil propensity, and a C-terminal membrane anchor (aa 351-405). The role of the N-terminal globular domain in host-cell interaction was investigated.

[0218] A truncated *nadA* gene coding for a protein devoid of amino acids 30-87 was cloned into pET-21 vector (pET-NadAΔ30-87) and expressed in *E.coli* BL21(DE3) strain. Amino acids 24-29 were retained to allow processing of the leader peptide and correct maturation of the protein. Western blot and FACS analysis confirmed that NadAΔ30-87 was expressed and formed oligomers on the *E.coli* cell surface i.e. deletion of the N-terminal domain does not interfere with the expression, export and membrane localization of NadA. However, the recombinant *E.coli* strain completely lost the

capacity to adhere to Chang epithelial cells. The N-terminus domain is thus implicated in adhesin activity.

[0219] To further investigate which part of the N-terminal domain is involved in the interaction, the region was additionally divided into three putative sub-domains: amino acids 24-42, containing a predicted α -helix region with hydrophobic residues; amino acids 43-70, the internal part without a predicted defined secondary structure; and amino acids 71-87 containing an other predicted α -helix structure. Three constructs, each encoding a protein deleted of a single sub-domain, were generated and then introduced into *E. coli* BL21(DE3), obtaining the following strains: BL21(DE3)/pET-NadA Δ 24-42, BL21(DE3)/pET-NadA Δ 43-70 and BL21(DE3)/pET-NadA Δ 71-87. Surface localisation of the oligomers was confirmed by western blot and FACS analysis, but adhesion to Chang epithelial cells was no better than the control BL21(DE3)/pET *E. coli* strain. These results, confirmed also using immunofluorescence microscopy analysis, indicate that the entire globular N-terminal domain of NadA is important in the interaction with human cells.

Combination with meningococcal and/or Hib conjugates

[0220] The triple MenB composition is combined with a mixture of oligosaccharide conjugates for serogroups C, W135 and Y, to give a vaccine containing the following antigens:

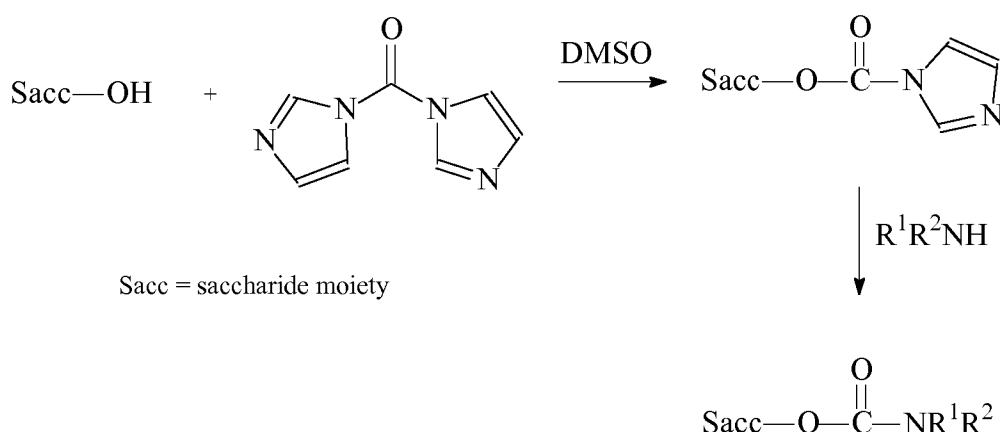
Component	Quantity per 0.5ml dose
Serogroup C conjugate	10 μ g saccharide + 12.5-25 μ g CRM ₁₉₇
Serogroup W135 conjugate	10 μ g saccharide + 6.6-20 μ g CRM ₁₉₇
Serogroup Y conjugate	10 μ g saccharide + 6.6-20 μ g CRM ₁₉₇
Δ G287-953	20 μ g polypeptide
936- Δ G741	20 μ g polypeptide
NadA	20 μ g polypeptide

[0221] A similar vaccine is prepared, including MenA conjugate (10 μ g saccharide + 12.5-33 μ g CRM₁₉₇) and/or a HbOC Hib conjugate (10 μ g saccharide + 2-5 μ g CRM₁₉₇).

Use of modified MenA saccharide

[0222] Capsular polysaccharide was purified from MenA and was hydrolysed to give MenA oligosaccharide. The polysaccharide (2 g) was hydrolyzed at 50°C in 50 mM sodium acetate buffer, pH 4.75, at a polysaccharide concentration of 10 mg/mL for about 4 hours [135]. After hydrolysis, the solution was dried by rotary evaporation.

[0223] The oligosaccharide was activated using the following reaction scheme:



[0224] The oligosaccharide was dissolved in DMSO to give a saccharide concentration of 10 mg/mL. According to a molar ratio of oligosaccharide:CDI being 1:20, 21.262 g of CDI was then added and the reaction mixture stirred for 16 hours at room temperature. The resulting MenA-CDI compound was purified by selective precipitation in a 80:20 (v/v) acetone:DMSO mixture followed by centrifugation. The efficiency of the activation reaction was calculated to be about 67.9% by determining the ratio of free imidazole to bonded imidazole.

[0225] In the second reaction step, the MenA-CDI oligosaccharide was solubilised in DMSO at a saccharide concentration of about 10 mg/mL. According to a molar ratio of MenA-CDI unit:DMA being 1:100, 36.288 g of 99% dimethylamine hydrochloride (*i.e.* R¹ & R² = Me) was added and the reaction mixture stirred for 16 hours at room temperature. The reaction product was freeze-dried and resolubilised in 10 mg/mL water solution.

[0226] To remove the low molecular weight reaction reagent (in particular the dimethylamine (DMA)) from the oligosaccharide preparation, a dialysis step was performed through a 3.5 kDa MWCO membrane (Spectra/Por™). Four dialysis steps were carried out: (i) 16 hours against 2 L of 1 M sodium chloride (dialysis factor 1:20), (ii) 16 hours against 2 L of 0.5 M sodium chloride (dialysis factor 1:20), (iii) and (iv) 16 hours against 2 L of WFI (dialysis factor 1:20). To improve the purification a diafiltration step was also performed through a 1 kDa MWCO membrane (Centricon™).

[0227] The purified MenA-CDI-DMA product was buffered at pH 6.5 in 25 mM L-histidine (Fluka™).

[0228] For preparing conjugates of the modified MenA saccharide (MenA-CDI-DMA), the overall process was as follows:

- hydrolysis of the polysaccharide to give oligosaccharide fragments
- sizing of the oligosaccharide fragments
- reductive amination of terminal aldehyde groups on the sized oligosaccharides
- protection of terminal -NH₂ groups by Fmoc group before the CDI reaction
- intrinsic de-protection of -NH₂ groups during the DMA reaction
- activation of terminal -NH₂ groups by SIDEA (N-hydroxysuccinimide adipic acid)
- covalent attachment to CRM₁₉₇ protein

[0229] The modified MenA oligosaccharide conjugate is much more resistant to hydrolysis than its natural counterpart at elevated temperatures. After 28 days at 37°C, for instance, the percentage of released saccharide is 6.4 % for the modified oligosaccharide vs. 23.5 % for the natural antigen. Moreover, the titres induced by the modified oligosaccharides are not significantly lower than those obtained using the native sugar structures.

[0230] The modified MenA conjugate is combined with MenC, MenW135 and MenY conjugates as a substitute for the conjugate of unmodified oligosaccharide. This tetravalent mixture is mixed with the three MenB polypeptides to give a vaccine effective against serogroups A, B, C, W135 and Y of *N.meningitidis* in a single dose.

Pneumococcal combinations

[0231] The three combined MenB proteins are mixed with pneumococcal saccharide conjugates to give a final concentration of 2 µg/dose of each of the pneumococcal serotypes (double for serotype 6B). The reconstituted vaccine thus contains the following antigens:

Component	Quantity per 0.5ml dose
Serogroup A conjugate	5 µg saccharide + 6.25-16.5 µg CRM ₁₉₇
Serogroup C conjugate	5 µg saccharide + 6.25-12.5 µg CRM ₁₉₇
Serogroup W135 conjugate	5 µg saccharide + 3.3-10 µg CRM ₁₉₇
Serogroup Y conjugate	5 µg saccharide + 3.3-10 µg CRM ₁₉₇
Pneumococcus serotype 4 conjugate	2 µg saccharide + 2.5 µg CRM ₁₉₇
Pneumococcus serotype 9V conjugate	2 µg saccharide + 2.5 µg CRM ₁₉₇
Pneumococcus serotype 14 conjugate	2 µg saccharide + 2.5 µg CRM ₁₉₇
Pneumococcus serotype 18C conjugate	2 µg saccharide + 2.5 µg CRM ₁₉₇
Pneumococcus serotype 19F conjugate	2 µg saccharide + 2.5 µg CRM ₁₉₇
Pneumococcus serotype 23F conjugate	2 µg saccharide + 2.5 µg CRM ₁₉₇
Pneumococcus serotype 6B conjugate	4 µg saccharide + 5 µg CRM ₁₉₇

[0232] It will be understood that the invention has been described by way of example only and modifications may be made whilst remaining within the scope of the claims.

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TABLE 1

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ET:	other	other	n.d.	ET5	ET5	ET5	ET5	lin.3	lin.3	lin.3	lin.3	lin.3	A4	A4	A4		sIII	sl		
Positive control	32768	32768	32768	16384	16384	>16384	8192	16384	8192	32768	8192	16384	8192	32768	8192		1024		1024	4096
Antigen mixture	4096	4096	65536	32768	65536	>65536	>4096	8192	2048	>4096	4096	4096	2048	2048	>4096		8192	16384	4096	>8192
Antigens + H44/76 OMVs	16384	8192	>65536	32768	524288	>65536	>4096	16384	8192	>4096	>4096	>4096	>8192	2048	>4096		32768	32768	16384	>8192
Antigens + 394/98 OMV	8192	8192	>65536	32768	>65536	>65536	>4096	65536	>8192	>4096	>4096	>4096	2048	8192	>4096		65536	65536	65536	>8192
OMVs (Norway)	<4	1024	8192	2048	262144	256	<8	4096	<4	<8	<8	<4	>8192	<8	<8		1024	<4	<8	>4096
OMVs (NZ)	512	<4	128	2048	<4	<8	<8	32768	>8192	4096	1024	4096	<16	n.d.	<8		4096	1024	<8	>4096

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	Thr	Ile	Thr	Lys	Lys	Asp	Ala	Thr	Ala	Ala	Asp	Val	Glu	Ala	Asp	Asp
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	Phe	Lys	Gly	Leu	Gly	Leu	Lys	Lys	Val	Val	Thr	Asn	Leu	Thr	Lys	Thr
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	Asn	Thr	Lys	Ala	Asp	Glu	Ala	Val	Lys	Thr	Ala	Asn	Glu	Ala	Lys	Gln
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		210					215					220					
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		225				230				235						240	
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				245					250						255		
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			35					40					45			
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				100					105					110		
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	Asp	Asn	Pro	Lys	Asn	Glu	Asp	Glu	Val	Ala	Gln	Asn	Asp	Met	Pro	Gln
	65					70					75					80
15	Asn	Ala	Ala	Gly	Thr	Asp	Ser	Ser	Thr	Pro	Asn	His	Thr	Pro	Asp	Pro
					85					90					95	
	Asn	Met	Leu	Ala	Gly	Asn	Met	Glu	Asn	Gln	Ala	Thr	Asp	Ala	Gly	Glu
20				100					105					110		
	Ser	Ser	Gln	Pro	Ala	Asn	Gln	Pro	Asp	Met	Ala	Asn	Ala	Ala	Asp	Gly
			115					120					125			
25	Met	Gln	Gly	Asp	Asp	Pro	Ser	Ala	Gly	Gly	Gln	Asn	Ala	Gly	Asn	Thr
		130					135					140				
	Ala	Ala	Gln	Gly	Ala	Asn	Gln	Ala	Gly	Asn	Asn	Gln	Ala	Ala	Gly	Ser
	145					150					155					160
30	Ser	Asp	Pro	Ile	Pro	Ala	Ser	Asn	Pro	Ala	Pro	Ala	Asn	Gly	Gly	Ser
					165					170					175	
	Asn	Phe	Gly	Arg	Val	Asp	Leu	Ala	Asn	Gly	Val	Leu	Ile	Asp	Gly	Pro
				180					185					190		
35	Ser	Gln	Asn	Ile	Thr	Leu	Thr	His	Cys	Lys	Gly	Asp	Ser	Cys	Ser	Gly
			195					200					205			
	Asn	Asn	Phe	Leu	Asp	Glu	Glu	Val	Gln	Leu	Lys	Ser	Glu	Phe	Glu	Lys
40		210					215					220				
	Leu	Ser	Asp	Ala	Asp	Lys	Ile	Ser	Asn	Tyr	Lys	Lys	Asp	Gly	Lys	Asn
	225					230					235					240
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					245					250					255	
	Asn	Gln	Tyr	Ile	Ile	Phe	Tyr	Lys	Pro	Lys	Pro	Thr	Ser	Phe	Ala	Arg
				260					265					270		
50	Phe	Arg	Arg	Ser	Ala	Arg	Ser	Arg	Arg	Ser	Leu	Pro	Ala	Glu	Met	Pro
			275					280					285			
	Leu	Ile	Pro	Val	Asn	Gln	Ala	Asp	Thr	Leu	Ile	Val	Asp	Gly	Glu	Ala
		290					295					300				
55	Val	Ser	Leu	Thr	Gly	His	Ser	Gly	Asn	Ile	Phe	Ala	Pro	Glu	Gly	Asn
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	Tyr	Arg	Tyr	Leu	Thr	Tyr	Gly	Ala	Glu	Lys	Leu	Pro	Gly	Gly	Ser	Tyr	
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				340					345					350			
	Ala	Ala	Val	Tyr	Asn	Gly	Glu	Val	Leu	His	Phe	His	Thr	Glu	Asn	Gly	
			355					360					365				
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		370					375						380				
	Ser	Lys	Ser	Val	Asp	Gly	Ile	Ile	Asp	Ser	Gly	Asp	Asp	Leu	His	Met	
	385					390					395					400	
15	Gly	Thr	Gln	Lys	Phe	Lys	Ala	Ala	Ile	Asp	Gly	Asn	Gly	Phe	Lys	Gly	
					405					410					415		
	Thr	Trp	Thr	Glu	Asn	Gly	Ser	Gly	Asp	Val	Ser	Gly	Lys	Phe	Tyr	Gly	
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	Pro	Ala	Gly	Glu	Glu	Val	Ala	Gly	Lys	Tyr	Ser	Tyr	Arg	Pro	Thr	Asp	
			435					440					445				
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5	Ala	Pro	Val	Val	Ser	Glu	Lys	Glu	Thr	Glu	Ala	Lys	Glu	Asp	Ala	Pro
				20					25					30		
	Gln	Ala	Gly	Ser	Gln	Gly	Gln	Gly	Ala	Pro	Ser	Ala	Gln	Gly	Gly	Gln
			35					40					45			
10	Asp	Met	Ala	Ala	Val	Ser	Glu	Glu	Asn	Thr	Gly	Asn	Gly	Gly	Ala	Ala
		50					55					60				
	Ala	Thr	Asp	Lys	Pro	Lys	Asn	Glu	Asp	Glu	Gly	Ala	Gln	Asn	Asp	Met
15	65					70					75					80
	Pro	Gln	Asn	Ala	Ala	Asp	Thr	Asp	Ser	Leu	Thr	Pro	Asn	His	Thr	Pro
					85					90					95	
	Ala	Ser	Asn	Met	Pro	Ala	Gly	Asn	Met	Glu	Asn	Gln	Ala	Pro	Asp	Ala
20				100					105					110		
	Gly	Glu	Ser	Glu	Gln	Pro	Ala	Asn	Gln	Pro	Asp	Met	Ala	Asn	Thr	Ala
			115					120					125			
25	Asp	Gly	Met	Gln	Gly	Asp	Asp	Pro	Ser	Ala	Gly	Gly	Glu	Asn	Ala	Gly
		130					135					140				
	Asn	Thr	Ala	Ala	Gln	Gly	Thr	Asn	Gln	Ala	Glu	Asn	Asn	Gln	Thr	Ala
30																
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40																
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					145											150																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																																									
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	Val	Asp	Glu	Tyr	His	Ala	Asn	Ala	Arg	Phe	Ala	Ile	Asp	His	Phe	Asn	
					485					490					495		
5	Thr	Ser	Thr	Asn	Val	Gly	Gly	Phe	Tyr	Gly	Leu	Thr	Gly	Ser	Val	Glu	
				500					505					510			
	Phe	Asp	Gln	Ala	Lys	Arg	Asp	Gly	Lys	Ile	Asp	Ile	Thr	Ile	Pro	Val	
			515					520					525				
10	Ala	Asn	Leu	Gln	Ser	Gly	Ser	Gln	His	Phe	Thr	Asp	His	Leu	Lys	Ser	
		530					535					540					
	Ala	Asp	Ile	Phe	Asp	Ala	Ala	Gln	Tyr	Pro	Asp	Ile	Arg	Phe	Val	Ser	
	545					550					555					560	
15	Thr	Lys	Phe	Asn	Phe	Asn	Gly	Lys	Lys	Leu	Val	Ser	Val	Asp	Gly	Asn	
				565						570					575		
	Leu	Thr	Met	His	Gly	Lys	Thr	Ala	Pro	Val	Lys	Leu	Lys	Ala	Glu	Lys	
20				580					585					590			
	Phe	Asn	Cys	Tyr	Gln	Ser	Pro	Met	Ala	Lys	Thr	Glu	Val	Cys	Gly	Gly	
			595					600					605				
25	Asp	Phe	Ser	Thr	Thr	Ile	Asp	Arg	Thr	Lys	Trp	Gly	Val	Asp	Tyr	Leu	
	610						615					620					
	Val	Asn	Val	Gly	Met	Thr	Lys	Ser	Val	Arg	Ile	Asp	Ile	Gln	Ile	Glu	
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				20					25					30			
	Leu	Arg	Ile	Glu	Thr	Thr	Ala	Arg	Ser	Tyr	Leu	Arg	Gln	Asn	Asn	Gln	
			35					40					45				
10	Thr	Lys	Gly	Tyr	Thr	Pro	Gln	Ile	Ser	Val	Val	Gly	Tyr	Asn	Arg	His	
		50					55					60					
	Leu	Leu	Leu	Leu	Gly	Gln	Val	Ala	Thr	Glu	Gly	Glu	Lys	Gln	Phe	Val	
15	65					70					75					80	
	Gly	Gln	Ile	Ala	Arg	Ser	Glu	Gln	Ala	Ala	Glu	Gly	Val	Tyr	Asn	Tyr	
					85					90					95		
	Ile	Thr	Val	Ala	Ser	Leu	Pro	Arg	Thr	Ala	Gly	Asp	Ile	Ala	Gly	Asp	
20				100					105					110			
	Thr	Trp	Asn	Thr	Ser	Lys	Val	Arg	Ala	Thr	Leu	Leu	Gly	Ile	Ser	Pro	
			115					120					125				
25																	
30																	
35																	
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55																	

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	Ala	Thr	Gln	Ala	Arg	Val	Lys	Ile	Val	Thr	Tyr	Gly	Asn	Val	Thr	Tyr	
	130						135					140					
5	Val	Met	Gly	Ile	Leu	Thr	Pro	Glu	Glu	Gln	Ala	Gln	Ile	Thr	Gln	Lys	
	145					150					155					160	
	Val	Ser	Thr	Thr	Val	Gly	Val	Gln	Lys	Val	Ile	Thr	Leu	Tyr	Gln	Asn	
					165					170					175		
10	Tyr	Val	Gln	Arg	Gly	Ser	Gly	Gly	Gly	Gly	Val	Ala	Ala	Asp	Ile	Gly	
				180					185					190			
	Ala	Gly	Leu	Ala	Asp	Ala	Leu	Thr	Ala	Pro	Leu	Asp	His	Lys	Asp	Lys	
			195					200					205				
15	Gly	Leu	Gln	Ser	Leu	Thr	Leu	Asp	Gln	Ser	Val	Arg	Lys	Asn	Glu	Lys	
	210						215					220					
	Leu	Lys	Leu	Ala	Ala	Gln	Gly	Ala	Glu	Lys	Thr	Tyr	Gly	Asn	Gly	Asp	
20	225					230					235					240	
	Ser	Leu	Asn	Thr	Gly	Lys	Leu	Lys	Asn	Asp	Lys	Val	Ser	Arg	Phe	Asp	
					245					250					255		
25	Phe	Ile	Arg	Gln	Ile	Glu	Val	Asp	Gly	Gln	Leu	Ile	Thr	Leu	Glu	Ser	
				260					265					270			
	Gly	Glu	Phe	Gln	Val	Tyr	Lys	Gln	Ser	His	Ser	Ala	Leu	Thr	Ala	Phe	
			275					280					285				
30	Gln	Thr	Glu	Gln	Ile	Gln	Asp	Ser	Glu	His	Ser	Gly	Lys	Met	Val	Ala	
	290						295					300					
	Lys	Arg	Gln	Phe	Arg	Ile	Gly	Asp	Ile	Ala	Gly	Glu	His	Thr	Ser	Phe	
35	305					310					315					320	
	Asp	Lys	Leu	Pro	Glu	Gly	Gly	Arg	Ala	Thr	Tyr	Arg	Gly	Thr	Ala	Phe	
					325					330					335		
	Gly	Ser	Asp	Asp	Ala	Gly	Gly	Lys	Leu	Thr	Tyr	Thr	Ile	Asp	Phe	Ala	
40				340					345					350			
	Ala	Lys	Gln	Gly	Asn	Gly	Lys	Ile	Glu	His	Leu	Lys	Ser	Pro	Glu	Leu	
			355					360					365				
45	Asn	Val	Asp	Leu	Ala	Ala	Ala	Asp	Ile	Lys	Pro	Asp	Gly	Lys	Arg	His	
	370						375					380					
	Ala	Val	Ile	Ser	Gly	Ser	Val	Leu	Tyr	Asn	Gln	Ala	Glu	Lys	Gly	Ser	
	385					390					395					400	
50	Tyr	Ser	Leu	Gly	Ile	Phe	Gly	Gly	Lys	Ala	Gln	Glu	Val	Ala	Gly	Ser	
				405						410					415		
	Ala	Glu	Val	Lys	Thr	Val	Asn	Gly	Ile	Arg	His	Ile	Gly	Leu	Ala	Ala	
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	1				5					10					15		
5	Ala	Asp	Ala	Leu	Thr	Ala	Pro	Leu	Asp	His	Lys	Asp	Lys	Gly	Leu	Gln	
				20					25					30			
	Ser	Leu	Thr	Leu	Asp	Gln	Ser	Val	Arg	Lys	Asn	Glu	Lys	Leu	Lys	Leu	
			35					40					45				
10	Ala	Ala	Gln	Gly	Ala	Glu	Lys	Thr	Tyr	Gly	Asn	Gly	Asp	Ser	Leu	Asn	
		50					55					60					
	Thr	Gly	Lys	Leu	Lys	Asn	Asp	Lys	Val	Ser	Arg	Phe	Asp	Phe	Ile	Arg	
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	Gln	Ile	Glu	Val	Asp	Gly	Gln	Leu	Ile	Thr	Leu	Glu	Ser	Gly	Glu	Phe	
					85					90					95		
	Gln	Val	Tyr	Lys	Gln	Ser	His	Ser	Ala	Leu	Thr	Ala	Phe	Gln	Thr	Glu	
20				100					105					110			
	Gln	Ile	Gln	Asp	Ser	Glu	His	Ser	Gly	Lys	Met	Val	Ala	Lys	Arg	Gln	
			115					120					125				
25	Phe	Arg	Ile	Gly	Asp	Ile	Ala	Gly	Glu	His	Thr	Ser	Phe	Asp	Lys	Leu	
		130					135					140					
	Pro	Glu	Gly	Gly	Arg	Ala	Thr	Tyr	Arg	Gly	Thr	Ala	Phe	Gly	Ser	Asp	
	145					150					155					160	
30	Asp	Ala	Gly	Gly	Lys	Leu	Thr	Tyr	Thr	Ile	Asp	Phe	Ala	Ala	Lys	Gln	
					165					170					175		
	Gly	Asn	Gly	Lys	Ile	Glu	His	Leu	Lys	Ser	Pro	Glu	Leu	Asn	Val	Asp	
35				180					185					190			
	Leu	Ala	Ala	Ala	Asp	Ile	Lys	Pro	Asp	Gly	Lys	Arg	His	Ala	Val	Ile	
			195					200					205				
	Ser	Gly	Ser	Val	Leu	Tyr	Asn	Gln	Ala	Glu	Lys	Gly	Ser	Tyr	Ser	Leu	
40		210					215					220					
	Gly	Ile	Phe	Gly	Gly	Lys	Ala	Gln	Glu	Val	Ala	Gly	Ser	Ala	Glu	Val	
	225					230					235					240	
45																	
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	Cys	Ser	Ser	Gly	Gly	Gly	Gly	Val	Ala	Ala	Asp	Ile	Gly	Ala	Gly	Leu
	1				5					10					15	
5	Ala	Asp	Ala	Leu	Thr	Ala	Pro	Leu	Asp	His	Lys	Asp	Lys	Ser	Leu	Gln
				20					25					30		
	Ser	Leu	Thr	Leu	Asp	Gln	Ser	Val	Arg	Lys	Asn	Glu	Lys	Leu	Lys	Leu
			35					40					45			
10	Ala	Ala	Gln	Gly	Ala	Glu	Lys	Thr	Tyr	Gly	Asn	Gly	Asp	Ser	Leu	Asn
		50					55					60				
	Thr	Gly	Lys	Leu	Lys	Asn	Asp	Lys	Val	Ser	Arg	Phe	Asp	Phe	Ile	Arg
15	65					70					75					80
	Gln	Ile	Glu	Val	Asp	Gly	Gln	Leu	Ile	Thr	Leu	Glu	Ser	Gly	Glu	Phe
					85					90					95	
	Gln	Ile	Tyr	Lys	Gln	Asp	His	Ser	Ala	Val	Val	Ala	Leu	Gln	Ile	Glu
20				100					105					110		
	Lys	Ile	Asn	Asn	Pro	Asp	Lys	Ile	Asp	Ser	Leu	Ile	Asn	Gln	Arg	Ser
			115					120					125			
25	Phe	Leu	Val	Ser	Gly	Leu	Gly	Gly	Glu	His	Thr	Ala	Phe	Asn	Gln	Leu
		130					135					140				
	Pro	Asp	Gly	Lys	Ala	Glu	Tyr	His	Gly	Lys	Ala	Phe	Ser	Ser	Asp	Asp
	145					150					155					160
30	Ala	Gly	Gly	Lys	Leu	Thr	Tyr	Thr	Ile	Asp	Phe	Ala	Ala	Lys	Gln	Gly
					165					170					175	
	His	Gly	Lys	Ile	Glu	His	Leu	Lys	Thr	Pro	Glu	Gln	Asn	Val	Glu	Leu
35				180					185					190		
	Ala	Ala	Ala	Glu	Leu	Lys	Ala	Asp	Glu	Lys	Ser	His	Ala	Val	Ile	Leu
			195					200					205			
	Gly	Asp	Thr	Arg	Tyr	Gly	Ser	Glu	Glu	Lys	Gly	Thr	Tyr	His	Leu	Ala
40		210					215					220				
	Leu	Phe	Gly	Asp	Arg	Ala	Gln	Glu	Ile	Ala	Gly	Ser	Ala	Thr	Val	Lys
	225					230					235					240
45	Ile	Gly	Glu	Lys	Val	His	Glu	Ile	Gly	Ile	Ala	Gly	Lys	Gln		
					245					250						

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50 <212> PRT

<213> Neisseria meningitidis

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	Cys	Ser	Ser	Gly	Gly	Gly	Gly	Ser	Gly	Gly	Gly	Gly	Val	Ala	Ala	Asp	
	1				5					10					15		
5	Ile	Gly	Thr	Gly	Leu	Ala	Asp	Ala	Leu	Thr	Ala	Pro	Leu	Asp	His	Lys	
				20					25					30			
	Asp	Lys	Gly	Leu	Lys	Ser	Leu	Thr	Leu	Glu	Asp	Ser	Ile	Pro	Gln	Asn	
			35					40					45				
10	Gly	Thr	Leu	Thr	Leu	Ser	Ala	Gln	Gly	Ala	Glu	Lys	Thr	Phe	Lys	Ala	
		50					55					60					
	Gly	Asp	Lys	Asp	Asn	Ser	Leu	Asn	Thr	Gly	Lys	Leu	Lys	Asn	Asp	Lys	
15	65					70					75					80	
	Ile	Ser	Arg	Phe	Asp	Phe	Val	Gln	Lys	Ile	Glu	Val	Asp	Gly	Gln	Thr	
				85						90					95		
	Ile	Thr	Leu	Ala	Ser	Gly	Glu	Phe	Gln	Ile	Tyr	Lys	Gln	Asn	His	Ser	
20				100					105					110			
	Ala	Val	Val	Ala	Leu	Gln	Ile	Glu	Lys	Ile	Asn	Asn	Pro	Asp	Lys	Thr	
			115					120					125				
	Asp	Ser	Leu	Ile	Asn	Gln	Arg	Ser	Phe	Leu	Val	Ser	Gly	Leu	Gly	Gly	
25			130				135					140					
	Glu	His	Thr	Ala	Phe	Asn	Gln	Leu	Pro	Gly	Gly	Lys	Ala	Glu	Tyr	His	
	145					150					155					160	
30	Gly	Lys	Ala	Phe	Ser	Ser	Asp	Asp	Pro	Asn	Gly	Arg	Leu	His	Tyr	Ser	
				165						170					175		
	Ile	Asp	Phe	Thr	Lys	Lys	Gln	Gly	Tyr	Gly	Arg	Ile	Glu	His	Leu	Lys	
35				180					185					190			
	Thr	Leu	Glu	Gln	Asn	Val	Glu	Leu	Ala	Ala	Ala	Glu	Leu	Lys	Ala	Asp	
			195					200					205				
	Glu	Lys	Ser	His	Ala	Val	Ile	Leu	Gly	Asp	Thr	Arg	Tyr	Gly	Ser	Glu	
40		210					215					220					
	Glu	Lys	Gly	Thr	Tyr	His	Leu	Ala	Leu	Phe	Gly	Asp	Arg	Ala	Gln	Glu	
	225					230				235						240	
	Ile	Ala	Gly	Ser	Ala	Thr	Val	Lys	Ile	Gly	Glu	Lys	Val	His	Glu	Ile	
45					245					250					255		
	Gly	Ile	Ala	Gly	Lys	Gln											
50				260													

Claims

1. A composition comprising five meningococcal antigens:

- (1) a NadA protein which is a polypeptide consisting of the amino acid sequence of SEQ ID NO: 2;
- (2) a 741 protein which has $\geq 85\%$ identity to SEQ ID NO: 3;
- (3) a 936 protein which has $\geq 85\%$ identity to SEQ ID NO: 4;

- (4) a 953 protein which has $\geq 85\%$ identity to SEQ ID NO: 5; and
- (5) a 287 protein which has $\geq 85\%$ identity to SEQ ID NO: 6.

2. The composition of claim 1, wherein the composition comprises a polypeptide of formula $\text{NH}_2\text{-A-}[\text{-X-L-}]_n\text{-B-COOH}$, wherein: X is an amino acid sequence of one of the five antigens (1) to (5); L is an optional linker amino acid sequence; A is an optional N-terminal amino acid sequence; B is an optional C-terminal amino acid sequence; and n is 2, 3, 4 or 5.
3. The composition of claim 2, wherein: (a) n is 2, X1 is the 936 protein as defined in claim 1 and X2 is the 741 protein as defined in claim 1; and/or (b) n is 2, X1 is the 287 protein as defined in claim 1 and X2 is the 953 protein as defined in claim 1.
4. The composition of any one of claims 1 to 3, comprising (a) a protein comprising SEQ ID NO: 7 and/or (b) a protein comprising SEQ ID NO: 8.
5. The composition of any one of claims 1 to 4 comprising an outer-membrane vesicle (OMV) preparation from *N. meningitidis* serogroup B.

Patentansprüche

1. Zusammensetzung, umfassend fünf Meningokokken-Antigene:

- (1) ein NadA-Protein, das ein Polypeptid, bestehend aus der Aminosäuresequenz von SEQ ID NO: 2, ist;
- (2) ein 741-Protein, das $\geq 85\%$ Identität zu SEQ ID NO: 3 aufweist;
- (3) ein 936-Protein, das $\geq 85\%$ Identität zu SEQ ID NO: 4 aufweist;
- (4) ein 953-Protein, das $\geq 85\%$ Identität zu SEQ ID NO: 5 aufweist; und
- (5) ein 287-Protein, das $\geq 85\%$ Identität zu SEQ ID NO: 6 aufweist.

2. Zusammensetzung gemäss Anspruch 1, wobei die Zusammensetzung ein Polypeptid der Formel $\text{NH}_2\text{-A-}[\text{-X-L-}]_n\text{-B-COOH}$ aufweist, worin X eine Aminosäuresequenz von einem der fünf Antigene (1) bis (5) ist; L eine optionale Linker-Aminosäuresequenz ist; A eine optionale N-terminale Aminosäuresequenz ist; B eine optionale C-terminale Aminosäuresequenz ist und n 2, 3, 4 oder 5 ist.
3. Zusammensetzung gemäss Anspruch 2, worin: (a) n 2 ist, X1 das 936-Protein ist, wie in Anspruch 1 definiert, und X2 das 741-Protein ist, wie in Anspruch 1 definiert; und/oder (b) n 2 ist, X1 das 287-Protein ist, wie in Anspruch 1 definiert, und X2 das 953-Protein ist, wie in Anspruch 1 definiert.
4. Zusammensetzung gemäss irgendeinem der Ansprüche 1 bis 3, umfassend (a) ein Protein, das SEQ ID NO: 7 umfasst, und/oder (b) ein Protein, das SEQ ID NO: 8 umfasst.
5. Zusammensetzung gemäss irgendeinem der Ansprüche 1 bis 4, umfassend eine Aussenmembranvesikel (OMV)-Zubereitung von *N. Meningitidis* Serogruppe B.

Revendications

1. Composition comprenant cinq antigènes méningococciques :

- (1) une protéine NadA qui est un polypeptide constitué de la séquence d'acides aminés de SEQ ID NO : 2 ;
- (2) une protéine 741 qui présente $\geq 85\%$ d'identité avec SEQ ID NO : 3 ;
- (3) une protéine 936 qui présente $\geq 85\%$ d'identité avec SEQ ID NO : 4 ;
- (4) une protéine 953 qui présente $\geq 85\%$ d'identité avec SEQ ID NO : 5 ; et
- (5) une protéine 287 qui présente $\geq 85\%$ d'identité avec SEQ ID NO : 6.

2. Composition selon la revendication 1, la composition comprenant un polypeptide de formule $\text{NH}_2\text{-A-}[\text{-X-L-}]_n\text{-B-COOH}$, dans laquelle : X représente une séquence d'acides aminés de l'un des cinq antigènes (1) à (5) ; L représente une séquence d'acides aminés de lieu facultatif ; A représente une séquence d'acides aminés N-terminale

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facultative ; B représente une séquence d'acides aminés C-terminale facultative ; et n est égal à 2, 3, 4 ou 5.

3. Composition selon la revendication 2, dans laquelle : (a) n vaut 2, X1 représente la protéine 936 telle que définie dans la revendication 1 et X2 représente la protéine 741 telle que définie dans la revendication 1 ; et/ou (b) n vaut 2, X1 représente la protéine 287 telle que définie dans la revendication 1 et X2 représente la protéine 953 telle que définie dans la revendication 1.
4. Composition selon l'une quelconque des revendications 1 à 3, comprenant (a) une protéine comprenant SEQ ID NO : 7 et/ou (b) une protéine comprenant SEQ ID NO : 8.
5. Composition selon l'une quelconque des revendications 1 à 4 comprenant une préparation de vésicules de la membrane externe (VME) de *N. meningitidis* du sérotype B.

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Szabadalmi igénypontok

1. Készítmény, amely öt meningokokkusz antigént tartalmaz:

- (1) NadA protein, amely a SEQ ID NO: 2 szerinti aminosav-szekvenciából álló polipeptid;
- (2) 741 protein, amely $\geq 85\%$ azonosságú a SEQ ID NO: 3 szerinti szekvenciával;
- (2) 936 protein, amely $\geq 85\%$ azonosságú a SEQ ID NO: 4 szerinti szekvenciával;
- (2) 953 protein, amely $\geq 85\%$ azonosságú a SEQ ID NO: 5 szerinti szekvenciával; és
- (2) 287 protein, amely $\geq 85\%$ azonosságú a SEQ ID NO: 6 szerinti szekvenciával;

2. Az 1. igénypont szerinti készítmény, ahol a készítmény $\text{NH}_2\text{-A-}[-\text{X-L-}]_n\text{-B-COOH}$ képletű polipeptidet tartalmaz, ahol: X jelentése a fenti (1) - (5) antigén egyikének az aminosav-szekvenciája; L jelentése opcionális kapcsoló aminosav-szekvencia; A jelentése opcionális N-terminális aminosav-szekvencia; B jelentése opcionális C-terminális aminosav-szekvencia, és n értéke 2, 3, 4 vagy 5.

3. A 2. igénypont szerinti készítmény, ahol: (a) n értéke 2, X1 jelentése a 936 protein, amint az 1. igénypontban definiálva van és X2 jelentése a 741 protein, amint az 1. igénypontban definiálva van; és/vagy (b) n értéke 2, X1 jelentése a 287 protein, amint az 1. igénypontban definiálva van és X2 jelentése a 953 protein, amint az 1. igénypontban definiálva van;

4. Az 1-3. igénypontok bármelyike szerinti készítmény, amely tartalmaz (a) SEQ ID NO: 7 szerinti szekvenciát tartalmazó proteint és/vagy (b) SEQ ID NO: 8 szerinti szekvenciát tartalmazó proteint.

5. Az 1-4. igénypontok bármelyike szerinti készítmény, amely *N. meningitidis* B szerocsoportból származó külső membrán vezikulum (OMV) készítményt tartalmaz.

