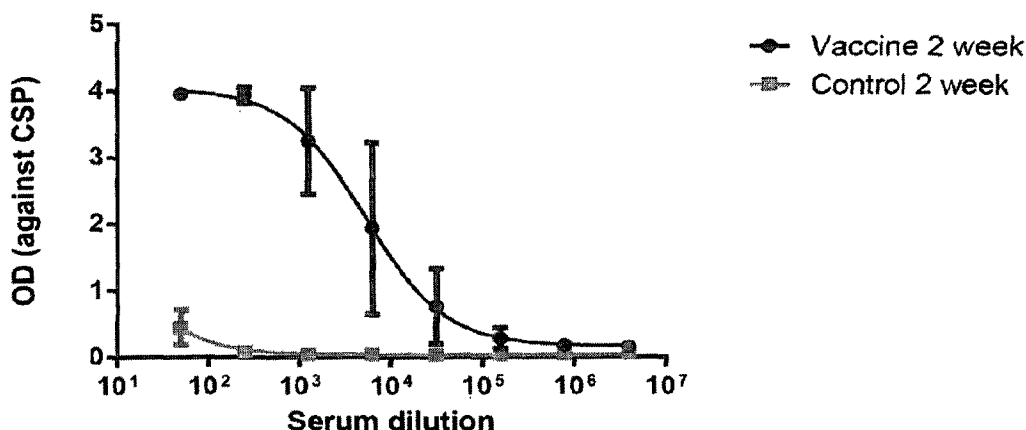




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(54) Titre : PARTICULES PSEUDO-VIRALES DU VIRUS COMPRENANT UN ANTIGENE CONTRE LA MALARIA ET
LEUR UTILISATION COMME UN VACCIN CONTRE LA MALARIA
(54) Title: A VIRUS-LIKE PARTICLE COMPRISING A MALARIA ANTIGEN AND USE THEREOF AS A MALARIA
VACCINE



(57) Abrégé/Abstract:

Disclosed is a particle comprising a polypeptide and at least one malaria antigen, and a composition or vaccine comprising thereof, its use in medicine, particularly in the prevention or treatment of malaria infections.

ABSTRACT

Disclosed is a particle comprising a polypeptide and at least one malaria antigen, and a composition or vaccine
5 comprising thereof, its use in medicine, particularly in the prevention or treatment of malaria infections.

A VIRUS-LIKE PARTICLE COMPRISING A MALARIA ANTIGEN AND USE
THEREOF AS A MALARIA VACCINE

TECHNICAL FIELD

5 [0001] The present invention relates to a particle comprising a polypeptide and at least one malaria antigen, and a composition or vaccine comprising thereof, its use in medicine, particularly in the prevention or treatment of malaria.

10 BACKGROUND

[0002] Malaria is one of the world's most prevalent serious infectious diseases, with approximately 250 million cases and 1 million deaths per year (WHO, 2009). Mortality is primarily in children under the age of five and in pregnant
15 women. Every 45 seconds, an African child dies of malaria. The disease is transmitted from person to person by infected mosquitoes, so past eradication efforts involved massive insecticide campaigns. These were successful in the Southeast U.S. for example, but failed in most poorly
20 developed tropical countries. Current efforts involve distribution of bednets, particularly bednets impregnated with insecticide, to prevent mosquito bites at night. However, resistance to insecticides and to anti-malarial drugs for both prevention and treatment is rapidly rising. Thus, the

need for a malaria vaccine is imperative for protection of millions of people from disease.

[0003] Malaria caused by *Plasmodium falciparum* remains a major public health threat, especially among children and pregnant women in Africa. An effective malaria vaccine would be a valuable tool to reduce the disease burden and could contribute to elimination of malaria in some regions of the world. Current malaria vaccine candidates are directed against human and mosquito stages of the parasite life cycle, but thus far, relatively few proteins have been studied for potential vaccine development.

[0004] The most advanced vaccine candidate, RTS,S, conferred partial protection against malaria in phase II clinical trials and is currently being evaluated in a phase III trial in Africa. (The Journal of Clinical Investigation 120(12) 4168-4178, 2010).

[0005] The CSP is the predominant surface antigen on sporozoites. CSP is composed of an N-terminal region that binds heparin sulfate proteoglycans (RI), a central region containing a four-amino-acid (NPNA) repeat, and a GPI-anchored C-terminal region containing a thrombospondin-like domain (RII). The region of the CSP included in the RTS,S vaccine includes the last 16 NPNA repeats and the entire

flanking C-terminus. HBsAg particles serve as the matrix carrier for RTS,S, 25% of which is fused to the CSP segment (The Journal of Clinical Investigation 120(12) 4168-4178, 2010).

5 [0006] In a series of phase II clinical trials for RTS,S, 30%-50% of malaria-naive adults immunized with RTS,S were protected against challenge by mosquitoes infected with the homologous *P. falciparum* clone. In phase II field trials in the Gambia and Kenya, RTS,S conferred short-lived protection
10 against malaria infection in approximately 35% of adults, although results from the Kenya trial did not reach statistical significance. Approximately 30%-50% of children and infants immunized with RTS,S in phase II trials conducted in Mozambique, Tanzania, and Kenya were protected from
15 clinical malaria, however, protection was generally short-lived (The Journal of Clinical Investigation 120(12) 4168-4178, 2010). Results from a pivotal, large-scale Phase III trial, published November 9, 2012, online in the *New England Journal of Medicine (NEJM)*, show that the RTS,S malaria
20 vaccine candidate can help protect African infants against malaria. When compared to immunization with a control vaccine, infants (aged 6-12 weeks at first vaccination) vaccinated with RTS,S had one-third fewer episodes of both clinical and severe malaria and had similar reactions to the injection.

[0007] There are currently no licensed vaccines against malaria. Highly effective malaria vaccine is strongly desired.

[0008] Virus-like particles (VLPs) are multiprotein
5 structures that mimic the organization and conformation of
authentic native viruses but lack the viral genome,
potentially yielding safer and cheaper vaccine candidates.
A handful of prophylactic VLP-based vaccines are currently
commercialized worldwide: GlaxoSmithKline's Engerix®
10 (hepatitis B virus) and Cervarix® (human papillomavirus),
and Merck and Co., Inc.'s Recombivax HB® (hepatitis B virus)
and Gardasil® (human papillomavirus) are some examples.
Other VLP-based vaccine candidates are in clinical trials or
undergoing preclinical evaluation, such as, influenza virus,
15 parvovirus, Norwalk and various chimeric VLPs. Many others
are still restricted to small-scale fundamental research,
despite their success in preclinical tests. The implications
of large-scale VLP production are discussed in the context
of process control, monitorization and optimization. The
20 main up- and down-stream technical challenges are identified
and discussed accordingly. Successful VLP-based vaccine
blockbusters are briefly presented concomitantly with the
latest results from clinical trials and the recent
developments in chimeric VLP-based technology for either

therapeutic or prophylactic vaccination (Expert Rev. Vaccines 9(10), 1149-1176, 2010).

[0009] Chikungunya virus (CHIKV) has infected millions of people in Africa, Europe and Asia since this alphavirus reemerged from Kenya in 2004. The severity of the disease and the spread of this epidemic virus present a serious public health threat in the absence of vaccines or antiviral therapies. It is reported that a VLP vaccine for epidemic Chikungunya virus protects non-human primates against infection (Nat Med. 2010 March; 16(3): 334-338). US patent publication No. 2012/0003266 discloses a virus-like particle (VLP) comprising one or more Chikungunya virus structural polypeptides which is useful for formulating a vaccine or antigenic composition for Chikungunya that induces immunity to an infection or at least one symptom thereof. WO2012/106356 discloses modified alphavirus or flavivirus virus-like particles (VLPs) and methods for enhancing production of modified VLPs for use in the prevention or treatment of alphavirus and flavivirus-mediated diseases.

SUMMARY

[0009a] Certain embodiments provide a particle comprising a virus structural polypeptide and at least one malaria antigen, wherein said virus structural polypeptide comprises at least one first attachment site and said at least one malaria antigen comprises at least one second attachment site, and wherein said virus structural polypeptide and said malaria antigen are linked through said at least one first and said at least one second attachment site, and wherein said particle is a virus-like particle. In some embodiments there is provided a virus-like particle comprising a virus structural polypeptide and at least one malaria antigen, wherein said virus structural polypeptide is a polypeptide of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) and comprises the capsid and the envelope proteins E1 and E2, wherein said at least one malaria antigen is an antigen comprising (NPNA)_n, where n is an integer from 4 to 30, and/or an antigen comprising (EYLNKIQNLSLSTEWSPCSVT)_y, where y is an integer from 1 to 6, and wherein said at least one malaria antigen is inserted into the virus structural peptide at a position that corresponds to: between residues 509-510, 510- 511, 511-512, 519-520, 529-530, 530-531, or 531-532 of SEQ ID NO. 1 or 2;

or between residues 515-516, 516-517, 517-518, 518-519, 519-520, 536-537, 537-538 or 538-539 of SEQ ID NO. 3.

[0009b] Certain embodiments further provide an isolated nucleic acid molecule consisting of a nucleotide sequence represented by SEQ ID Nos. 26-27, 29-30, 32-33, 35-36, 38, 40 or 42.

[0009c] Certain embodiments further provide an isolated nucleic acid molecule consisting of a nucleotide sequence which has a sequence identity of 90% or more with a nucleotide sequence represented by SEQ ID Nos. 26-27, 29-30, 32-33, 35-36, 38, 40 or 42.

[0010] In the first aspect, the present invention provides a particle which is capable of being self-assembled, comprising a polypeptide and at least one malaria antigen, wherein said polypeptide comprises at least one first attachment site and said at least one malaria antigen comprises at least one second attachment site, and wherein said polypeptide and said malaria antigen are linked through said at least one first and said at least one second attachment site.

[0011] In the second aspect, the present invention provides a nucleic acid molecule which is designed for expression of

a particle provided in the first aspect of the present invention.

[0012] In the third aspect, the present invention provides a composition or vaccine comprising the particle provided in
5 the first aspect of the present invention and/or the nucleic acid molecule provided in the second aspect of the present invention.

[0013] In the fourth aspect, the present invention provides a method of producing an antibody, comprising contacting the particle provided in the first aspect of the present invention and/or the nucleic acid molecule provided in the second aspect of the present invention to a mammal.

[0014] In the fifth aspect, the present invention provides a method of immunomodulation, a method of treating malaria , a method of inducing and/or enhancing immune response against a malaria antigen in a mammal, comprising administering the composition provided in the third aspect of the present invention to a mammal.

[0015] In sixth aspect, the present invention provides a method of passive immunization against a malaria-causing pathogen, comprising administering the antibody provided in the fourth aspect of the present invention to a mammal.

[0016] In seventh aspect, the present invention provides a method of presenting an antigen on macrophage, comprising contacting the particle provided in the first aspect of the present invention and/or the nucleic acid molecule provided in the second aspect of the present invention to a mammal.

[0017] In eighth aspect, the present invention provides a method for producing the particle provided in the first aspect of the present invention, comprising preparing a

vector which is designed for expression of said particle;
culturing a cell which is transfected with said vector to
express said particle; and recovering said particle.

BRIEF DESCRIPTION OF THE INVENTION

5 [0018]

Fig 1 shows pVLP74_15 (VLP_CHI 532 NPNAx6) vector.

Fig 2 shows pVLP78_15 (VLP_CHI 532 NPNAx25) vector.

Fig 3 shows pVLP74_25 (VLP_VEEV 519 NPNAx6) vector.

Fig 4 shows that the serum from individual monkeys immunized
10 with Malaria VLPs after 2 weeks induced high titer of
antibodies against CSP.

Fig 5 shows mean value and SD of the data shown in Fig 4.

Fig 6 shows effects of combined immunization of CHIKV VLP
and VEEV VLP on induction of antibodies against CSP. In the
15 figure, Adj indicates adjuvant.

Fig 7 shows effects of administered VLP fused with no malaria
antigen on induction of antibodies against CSP. In the
figure, 4w, 6w, 10w and 14w indicate 4 weeks after
immunization, 6 weeks after immunization, 10 weeks after
20 immunization and 14 weeks after immunization, respectively.

Fig 8 shows effects of administered VLP fused with malaria
antigen on induction of antibodies against CSP. In the
figure, 4w, 6w, 10w and 14w indicate 4 weeks after

immunization, 6 weeks after immunization, 10 weeks after immunization and 14 weeks after immunization, respectively. Fig 9 shows effects of administered VLP fused with malaria antigen together with adjuvant on induction of antibodies against CSP. In the figure, 4w, 6w, 10w and 14w indicate 4 weeks after immunization, 6 weeks after immunization, 10 weeks after immunization and 14 weeks after immunization, respectively.

Fig 10 shows schedule of the experiment.

Fig 11 shows detection of 18S malaria DNA by means of PCR.

DETAILED DESCRIPTION OF SELECTED EMBODIMENTS

[0019]

(1) A particle comprising a polypeptide and at least one malaria antigen

In the first aspect, the present invention provides a particle which is capable of being self-assembled, comprising a polypeptide and at least one malaria antigen, wherein said polypeptide comprises at least one first attachment site and said at least one antigen comprises at least one second attachment site, and wherein said polypeptide and said malaria antigen are linked through said at least one first and said at least one second attachment site.

[0020] As used herein, "a particle which is capable of being self-assembled" refers to a particle formed by at least one constituent which is spontaneously assembled. The constituent may be a polypeptide or non-peptide chemical compound. In one embodiment, "a particle which is capable of being self-assembled" may be a particle comprising or consisting of at least one polypeptide. The at least one polypeptide consists of one or more kinds of peptide. In one embodiment, said particle has a diameter of at least 10nm, for example, at least 20nm, preferably at least 50nm. In one embodiment, molecular weight of said particle is from 100 kDa to 100,000 kDa, preferably from 400kDa to 30,000kDa.

[0021] A polypeptide used for the present invention may be spontaneously assembled. The polypeptide may be a virus structural polypeptide. Thus, the particle provided by the present invention may be a virus like particle.

[0022] A virus structural polypeptide may be a naturally occurring viral polypeptide or modified polypeptide thereof. In one embodiment, the modified polypeptide has at least 70%, 75%, 80%, 85%, 90%, 95% or 98% amino acid sequence identity to a naturally occurring viral structural polypeptide including capsid and envelope protein. In one embodiment, the modified polypeptide is a mutant where at most 10% of the amino acids are deleted, substituted,

and/or added to a naturally occurring viral structural polypeptide including capsid and envelope protein.

[0023] In one embodiment, virus structural polypeptide used for the present invention consists of or comprises
5 capsid and/or envelope protein or fragment thereof. For example, virus structural polypeptide used for the present invention consists of or comprises capsid and E2 and E1.

An antigen may be inserted into E2. In one embodiment, a particle provided by the first aspect of the present
10 invention can be formed by assembling 240 capsids, 240 E1 proteins and 240 E2 proteins where a malaria antigen is inserted into each of E2 proteins.

[0024] Virus structural polypeptide used for the present invention may be derived from Alphavirus or Flavivirus. Thus,
15 the particle provided by the present invention may be a virus like particle derived from Alphavirus or Flavivirus. Examples of Alphavirus and Flavivirus include, but not limited to, Aura virus, Babanki virus, Barmah Forest virus (BFV), Bebaru virus, Cabassou virus, Chikungunya virus
20 (CHIKV), Eastern equine encephalitis virus (EEEV), Eilat virus, Everglades virus, Fort Morgan virus, Getah virus, Highlands J virus, Kyzylagach virus, Mayaro virus, Me Tri virus, Middelburg virus, Mosso das Pedras virus, Mucambo virus, Ndumu virus, O'nyong-nyong virus, Pixuna virus, Rio
25 Negro virus, Ross River virus (RRV), Salmon pancreas

disease virus, Semliki Forest virus, Sindbis virus, Southern elephant seal virus, Tonate virus, Trocara virus, Una virus, Venezuelan equine encephalitis virus (VEEV), Western equine encephalitis virus (WEEV), Whataroa virus, West Nile virus, dengue virus, tick-borne encephalitis virus and yellow fever virus.

[0025] Malaria is a disease which human or other animal (e.g. mouse) suffers from. Example of malaria include, but are not limited to, a disease caused by Plasmodium (P.) species including P. falciparum, P. malariae, P. ovale, P. vivax, P. knowlesi, P. berghei, P. chabaudi and P. yoelii

[0026] As used herein, the term "malaria antigen" refers to any antigen or fragment thereof. The term antigen or fragment thereof, means any peptide-based sequence that can be recognized by the immune system and/or that stimulates a cell-mediated immune response and/or stimulates the generation of antibodies.

[0027] According to Scand. J. Immunol. 56, 327-343, 2002, considering the whole parasite life cycle, there are essentially six targets for a malaria vaccine: (1) sporozoites; (2) liver stages; (3) merozoites; (4) infected RBC; (5) parasite toxins; (6) sexual stages.

[0028] Table summarizes the main candidate antigens of each stage identified.

Table 1. Main vaccine candidates from the different phases of *Plasmodium* life cycle

Targets	Candidate antigens
Sporozoite	Circumsporozoite protein (CSP)
	Thrombospondin-related adhesive protein (TRAP)
	Sporozoite and liver-stage antigen (SALSA)
	Sporozoite threonine- and asparagine-rich protein (STARP)
Liver stage	CSP
	Liver-stage antigen (LSA)-1 and -3
	SALSA
	STARP
Merozoite	Merozoite surface protein (MSP)-1, -2, -3, -4 and -5
	Erythrocyte-binding antigen (EBA)-175
	Apical membrane antigen (AMA)-1
	Rhoptry-associated protein (RAP)-1 and -2
Blood stage	Acidic-basic repeat antigen (ABRA)
	Duffy-binding protein (DBP) (<i>Plasmodium vivax</i>)
	Ring erythrocyte surface antigen (RESA)
	Serine-rich protein (SERP)

Table 1. Main vaccine candidates from the different phases of *Plasmodium* life cycle

Targets	Candidate antigens
	Erythrocyte membrane protein (EMP)-1, -2 and -3
	Glutamate-rich protein (GLURP)
Toxins	Glycosilphosphatidylinositol (GPI)
Sexual stages	Ps25, Ps28, Ps48/45 and Ps230

(Scand. J. Immunol. 56, 327-343, 2002)

[0029] According to the present invention, one or more antigens listed above can be used as long as it is formed to a particle. For example, a circumsporozoite protein and a fragment thereof can be used as an antigen. Examples of circumsporozoite protein include, but are not limited to, *Plasmodium falciparum* circumsporozoite protein consisting of amino acid sequence described below (SEQ ID No.:56):

Mmrklailsvssflfvealfqeyqcygsssnttrvlnelnydnagtnlynelemnyygkq
 enwyslkknsrslgenddgnnngdngregkdcdkrdggnedneklrkpkhkkklkqpgd
 gnpdpnanpnvdnpnanpnvdnpnanpnvdnpnanpnanpnanpnanpnanpnanpn
 npnanpnanpnanpnanpnanpnanpnanpnvdnpnanpnanpnanpnanpnanpn
 anpnanpnanpnanpnanpnanpnanpnanpnanpnanpnanpnknnggng
 qghnmpndpnrvdenanannavknnnnneepsdkhieqylkkiknsistewspcsvtcg
 ngiqvrikpgsankpkdeldyendiekkickmekcssvfnvnssiglimvlsflflnt
 r.

[0030] In one embodiment, malaria antigen is a Plasmodium falciparum circumsporozoite protein B cell epitope. Example of Plasmodium falciparum circumsporozoite protein B cell epitope may be a repeat sequence of NPNA, including (NPNA)₄₋₃₀ (i.e. 4xNPNA, 5xNPNA, 6xNPNA, 7xNPNA, 8xNPNA, 9xNPNA, 10xNPNA, 11xNPNA, 12xNPNA, 13xNPNA, 14xNPNA, 15xNPNA, 16xNPNA, 17xNPNA, 18xNPNA, 19xNPNA, 20xNPNA, 21xNPNA, 22xNPNA, 23xNPNA, 24xNPNA, 25xNPNA, 26xNPNA, 27xNPNA, 28xNPNA, 29xNPNA or 30xNPNA).

10 [0031] In one embodiment, malaria antigen is a Plasmodium yoelii circumsporozoite protein B cell epitope including (QGPGAP)₃₋₁₂.

[0032] In one embodiment, malaria antigen is a Plasmodium vivax circumsporozoite protein B cell epitope including
15 (ANGAGNQPG)₁₋₁₂.

[0033] In one embodiment, malaria antigen is a Plasmodium malariae circumsporozoite protein B cell epitope including (NAAG)₄₋₃₀.

[0034] In one embodiment, malaria antigen is a Plasmodium falciparum circumsporozoite protein T cell epitope. Example
20 of Plasmodium falciparum circumsporozoite protein T cell epitope may be EYLNKIQNSLSTEWSPCSVT (SEQ ID No.:44). (EYLNKIQNSLSTEWSPCSVT)₁₋₆ may be also used as a malaria antigen.

[0035] In one embodiment, malaria antigen is a Plasmodium
yoelii circumsporozoite protein T cell epitope which is
YNRNIVNRLLGDALNGPEEK (SEQ ID No.45).
(YNRNIVNRLLGDALNGPEEK)₁₋₆ may be also used as a malaria
5 antigen.

[0036] The present invention addresses one or more of the
above needs by providing antigens, vectors encoding the
antigens, and antibodies (and antibody-like molecules
including aptamers and peptides) that specifically bind to
10 the antigen, together with the uses thereof (either alone or
in combination) in the prevention or treatment of malaria
infections. As used herein, the term "antibody" refers to
molecules which are capable of binding an epitope or
antigenic determinant. The term is meant to include whole
15 antibodies and antigen-binding fragments thereof, including
single-chain antibodies. Such antibodies include human
antigen binding antibody fragments and include, but are not
limited to, Fab, Fab' and F(ab')₂, Fd, single-chain Fvs
(scFv), single-chain antibodies, disulfide-linked Fvs (sdFv)
20 and fragments comprising either a VL or VH domain. The
antibodies can be from any animal origin including birds and
mammals. Preferably, the antibodies are mammalian e.g. human,
murine, rabbit, goat, guinea pig, camel, horse and the like,
or other suitable animals e.g. chicken. As used herein,
25 "human" antibodies include antibodies having the amino acid

sequence of a human immunoglobulin and include antibodies isolated from human immunoglobulin libraries or from animals transgenic for one or more human immunoglobulins and that do not express endogenous immunoglobulins, as described, for example, in U.S. Patent No. 5,939,598.

[0037] The antigen used for the present invention can be modified polypeptide derived from a naturally occurring protein. The modified polypeptide may be a fragment of the naturally occurring protein. In one embodiment, the modified polypeptide has at least 70%, 75%, 80%, 85%, 90%, 95% or 98% amino acid sequence identity to a polypeptide derived from a naturally occurring protein. In one embodiment, the modified polypeptide derived is a mutant where at most 10% of the amino acids are deleted, substituted, and/or added based on a polypeptide derived from naturally occurring protein.

[0038] In the particle as provided by the present invention, a polypeptide and an antigen may be linked through at least one first attachment site which is present in the polypeptide and at least one second attachment site which is present in the antigen.

[0039] As used herein, each of "a first attachment site" and "a second attachment site" refers to a site where more than one substance is linked each other.

[0040] In one embodiment, the polypeptide and the antigen are directly fused. Alternatively, one or two linkers may intervene between N-terminal residue of the antigen and the polypeptide and/or between C-terminal residue of the antigen and the polypeptide.

[0041] The antigen or the polypeptide can be truncated and replaced by short linkers. In some embodiments, the antigen or the polypeptide include one or more peptide linkers. Typically, a linker consists of from 2 to 25 amino acids. Usually, it is from 2 to 15 amino acids in length, although in certain circumstances, it can be only one, such as a single glycine residue.

[0042] In one embodiment, a nucleic acid molecule, in which polynucleotide encoding the polypeptide is genetically fused with polynucleotide encoding the antigen, is expressed in a host cell so that the first attachment site and the second attachment site are linked through a peptide bond. In this case, the polypeptide and the antigen are linked through a peptide bond. Relating to this embodiment, the first attachment site and/or the second attachment site may be genetically modified from the original polypeptide or antigen. For example, the first attachment site is modified from the polypeptide so that through a linker peptide including SG, GS, SGG, GGS and SGSG, the polypeptide is conjugated with the antigen.

[0043] When the polypeptide are chemically conjugated with the antigen, the first attachment site and the second attachment site may be linked through a chemical cross-linker which is a chemical compound.

5 [0044] Examples of the cross-linker include, but are not limited to, SMPH, Sulfo-MBS, Sulfo-EMCS, Sulfo-GMBS, Sulfo-SIAB, Sulfo-SMPB, Sulfo-SMCC, SVSB, SIA and other cross-linkers available from the Pierce Chemical Company.

[0045] In one embodiment, the particle provided by the
10 present invention comprises a polypeptide linked to an antigen, wherein spatial distance between the N-terminal residue and C-terminal residue of the antigen is 30Å or less when the distance is determined in a crystal of the antigen or a naturally occurring protein containing the antigen or
15 modified protein therefrom.

[0046] The antigen used for the present invention can be designed by a person skilled in the art. For example, the antigen used for the present invention may be a naturally occurring protein or a fragment thereof. Alternatively, the
20 antigen used for the present invention may be a protein modified from a naturally occurring protein or a fragment thereof. A person skilled in the art can design the antigen so that spatial distance between the N-terminal residue and C-terminal residue of the antigen is 30Å or less when the
25 distance is determined in a crystal of the antigen or a

naturally occurring protein containing the antigen or modified protein therefrom. For example, the antigen used for the particle provided by the present invention can be designed using a free software including PyMOL (e.g. PyMOL v0.99: <http://www.pymol.org>). In one embodiment, the spatial distance between the N-terminal residue and C-terminal residue of the antigen is 30Å (angstrom) or less, 20Å or less, or 10Å or less (e.g. from 5 Å to 15 Å, from 5 Å to 12 Å, from 5 Å to 11 Å, from 5 Å to 10 Å, from 5 Å to 8 Å, from 8 Å to 15 Å, from 8 Å to 13 Å, from 8 Å to 12 Å, from 8 Å to 11 Å, from 9 Å to 12 Å, from 9 Å to 11 Å, from 9 Å to 10 Å or from 10 Å to 11 Å).

[0047]

15 Chikungunya virus like particle or a Venezuelan equine encephalitis virus like particle

In one embodiment, the present invention provides a Chikungunya virus like particle or a Venezuelan equine encephalitis virus like particle comprising a Chikungunya or Venezuelan equine encephalitis virus structural polypeptide and at least one malaria antigen, wherein said Chikungunya virus structural polypeptide or said Venezuelan equine encephalitis virus structural polypeptide comprises at least one first attachment site and said at least one malaria antigen comprises at least one second attachment site, and

wherein said Chikungunya or Venezuelan equine encephalitis virus structural polypeptide and said at least one antigen are linked through said at least one first and said at least one second attachment site.

5

[0048] In one embodiment, a spatial distance between the N-terminal residue and C-terminal residue of the malaria antigen may be 30 Å or less; 25 Å or less; 20 Å or less; 15 Å or less; 14 Å or less; 13 Å or less; 12 Å or less; 10 11 Å or less; 10 Å or less; 9 Å or less; or 8 Å or less (e.g. from 5 Å to 15 Å, from 5 Å to 12 Å, from 5 Å to 11 Å, from 5 Å to 10 Å, from 5 Å to 8 Å, from 8 Å to 15 Å, from 8 Å to 13 Å, from 8 Å to 12 Å, from 8 Å to 11 Å, from 9 Å to 12 Å, from 9 Å to 11 Å, from 9 Å to 10 Å or from 10 Å to 15 11 Å) when the distance is determined in a crystal of the malaria antigen or a naturally occurring protein containing the malaria antigen or modified protein therefrom.

[0049] In one embodiment, the malaria antigen is linked to the Chikungunya or Venezuelan equine encephalitis virus structural polypeptide by way of chemical cross-linking or as a fusion protein produced by way of genetic engineering.

[0050] A Chikungunya or Venezuelan equine encephalitis virus structural polypeptide used in the present invention may comprise a Chikungunya or Venezuelan equine encephalitis virus envelope protein and/or a capsid.

25

[0051] Examples of Chikungunya virus include, but are not limited to, strains of 37997 and LR2006 OPY-1.

Examples of Venezuelan equine encephalitis virus include, but are not limited to, TC-83.

5

[0052] Chikungunya or Venezuelan equine encephalitis virus structural polypeptide used in the present invention may naturally occurring virus structural polypeptide or modified polypeptide thereof. The modified polypeptide may be a fragment of the naturally occurring virus structural polypeptide. In one embodiment, the modified polypeptide has at least 70%, 75%, 80%, 85%, 90%, 95% or 98% amino acid sequence identity to a naturally occurring viral capsid and/or envelope protein. In one embodiment, the modified polypeptide is a mutant where at most 10% of the amino acids are deleted, substituted, and/or added based on a naturally occurring viral capsid and/or envelope protein. For example, K64A or K64N mutation may be introduced into a capsid of Venezuelan equine encephalitis virus structural polypeptide used in the present invention.

[0053] Chikungunya or Venezuelan equine encephalitis virus structural polypeptide may consist of or comprise a capsid, E2 and E1.

[0054] Examples of Chikungunya virus structural polypeptide include, but are not limited to, Capsid- E2-E1 of Chikungunya

25

virus Strain 37997, and Capsid- E2-E1 of Chikungunya virus LR2006 OPY-1.

[0055] Examples of Venezuelan equine encephalitis virus structural polypeptide include, but are not limited to, Capsid- E2-E1 of Venezuelan equine encephalitis virus Strain TC-83.

[0056] An exemplary Chikungunya virus structural polypeptide sequence is provided at Genbank Accession No. ABX40006.1, which is described below (SEQ ID No.:1):

```
mefiptqtfnrryqprpwtprptiqvirprprpqrqagqlaqlisavnkltmravpqq
kprnrknkkqkqkqgapqnnntnqkkqppkkkpaqkkkkpgrrrermcmkiendcifevk
hegkvtgyaclvgdkvmkpahvkgtdnadlaklfrsskydlecaqipvhmksdask
fthekpeggyynwhhgavqysggrftiptgagkpgdsgrpifdnkgrvvaivlgganega
rtalsvvtwnkdivtkitpegaeewslaipvmcllanttfpcsqppctpcceyekepet
lrmllednvmrpgyyqllqasltcsphrqrrstkdnfnvykatrpylahcpdcgeghsch
spvalerirneatdgtlkiqvsllqigiktddshdwtklrymdnhmpadaeraglfvrt
apctitgtmghfilarcpkgetltvgftdsrkishscthpfhhdppvigrekfhsrpqh
gkelpcstyvgstaatteieievhmppdpdrtlmsqsgsnvkitvngqtvrykcncggs
neglttttdkvinncvdqchaavtnhkkwgynsplvrnaelgdrkgkihipfplanvt
crvpkarnptvtygknqvimllypdhptllsyrnmgeepnyqeewvmhkkevltvpte
glevtwgnnepkywpqlstngtahghpheillyyelyptmtvvvsvatfillsmvg
maagmcmcarrrcitpyeltpgatvpflslliccirtakaatyqeaaiylwneqqplfw
lqaliplaalivlcnclrlpcccktlaflavmsvgahtvsayehvtvipntvgvpykt
lvnrpgyspmvlemellsvtleptlsldytceyktvipspyvkcctgaekdknlpdy
sckvftgvypfmwggaycfodaentqlseahveksescktefasayrahtasasaklr
lyqgnnitvtayangdhavtvkdakfivgpmssawtpfdnkivvykgdvynmdyppfga
grpgqfgdiqsrtpeskdvantqlvlqrpavgtvhvpysqapsgfkywlkergaslqh
tapfgcqiatspvravncavgnmpisidipeaaftrvvdapsltdmscevpatchssdf
ggvaiikyaaskkgkcavhsmtnavtireaeievegnsqqlqisfstalasaefrvqvc
tqvhaaeachppkdhivnypashttlgvqdisatamswvqkitggvgllvavaaliliv
vlcvsfsrh
```

10

[0057] Another exemplary Chikungunya virus structural polypeptide sequence is provided at Genbank Accession No. ABX40011.1, which is described below (SEQ ID No.:2):

```
mefiptqtfnrryqprpwprptiqvirprprpqrqagqlaqlisavnkltmravpqq
kprnrknkkqkqkqapqndpkqkkqppqkkpaqkkkkpgrrrermcmkiendc
ifevkhegkvmgyaclvgdkvmkpahvkgtdnadlaklfrsskydlecaqipvh
mksdaskfthekpeggyynwhhgavqysggrftiptgagkpgdsgrpifdnkgrvva
ivlgganegartalsvvtwnkdivtkitpegaeewslalpvcllanttfpcsqppctpccey
```

kepestlrmlednvmrpgyyqllkasltcsphrqrrstkdnfnvykatrpylahepcdeg
 eghschspialerirneatdgtlkiqvsqigiktdsdshdwtklrymdshtpadaeragll
 vrtsapctitgtmghfilarecpkgetltvgftdsrkishtcthpfhheppvigrerfhsrpq
 hgkelpcstyvqstaataeeievhmppdtpdrtlmtqqsgnvkitvngqtvrykencg
 gsnegltttdkvinckidqchaavtnhknwqynsplvprnaelgdrkgkihpfplan
 vtrcrvpkarnptvtygknqvtmllypdhptlsyrnmgqepnyheewvthkkevltlv
 pteglevtwgnnepykywpqmstngtahghpheiillyyelyptmtvvivsvasfvlls
 mvgtavgmecvcarrrcitpyeltpgatvpfllsllccvrttkaatyeeaaaylwneqqplf
 wlqaliplaalivlcnclllpcccktlafavmsigahtvsayehvtvipntvgvpyktlvn
 rpgyspmvlemelqsvtleptlsldyitceyktvipspyvkcgcgaecdkslpdyckvf
 tgvypfmwggaycfdaentqlseahveksescktefasayrahtasasaklvlyqgn
 nitvaayangdhavtvkdakfvvgpmssawtpfdnkivvykgdvynmdyppfgagr
 pgqfgdiqsrtpeskdvyanqlvlqrpaagtvhvpysqapsgfkywlkergaslqhta
 pfgcqiatnpvrvncavgnipisidipdaaftrvvdapsvtdmscevpacthssdfggv
 aii kytaskkgkcavhsmtnavtireadvevegnsqllqisfstalasaefrvqvcestqvhc
 aaachppkdhivnypashttlgvqdisttamswwqkitggvglivavaalilivlvcsfs
 rh

[0058] An exemplary Venezuelan equine encephalitis virus structural polypeptide is described below (SEQ ID No.:3):

mfpfqpmypmqmpyrnpfaaprrpwpfprtdpflamqvqeltrsmantlftkqrrdappe
 gpsaakpkkeasqkqkggggqgkkkknqgkktaktgppnpkaqngnkkktnkkgpkrqrm
 vmklesdktfpimlegkingyacvvggklfrpmhvegkidndvlaalktkkaskydley
 advpqnmradt fkythekpqgyyswhhgavqyengrftvpkgvgakgdsgrpildnqgr
 vvaivlggvnegsrtalsvmmwnekgtvkvypenceqwslvttmcllanvtfpcagpp
 icydrkpaetlamlsvndnpgydelleaavkcpgrkrsteelfneykltrpymarci
 rcavgschspiaieavksdghdgyvrlqtssqyglssgnlkgtrtmrydmhgtikeipl
 hqvslytsrphivdghgyfllarcpgadsitmfkkdsvrhscsvpyevkfnpvgrrel
 ythppehgveqacqvayahdaqnrgayvemhlpqsevdsslvslsgssvtvtpdgtal
 vecceggtkisetinktkqfsqctkkeqcrayrlqndkwvynsdklpkaagatlkgklh
 vpflldagkctvplapepmifgfrsvslklhpkntylitrqladephythelisepa
 vrnftvtekgwefvwnhppkrfwaqetapgnphglphevithyyhrypmstilglsic
 aaiatvsvaastwlfcrsrvacltpyrlltpnaripfclavlc cartaraettwesldhl
 wnnnqgmfwiglpllaalivvtrllrcvccvvpflvmagaagagayehattmpsqagi
 syntivnragyaplpisitptkikliptvnleyvtchytgmdspaikccgsqectpty
 rpdeqckvftgvypfmwggaycfcdtentqvskayvmksddcladhaeaykahtasvqa
 flnitvgehsivttvyvnggetpvnfngvkitagplstawtpfdrkivqyageiynydfp
 eygagqpqgafgdiqsrvtsssdlyantnlvlqrpkagaihvpvtqapsgfeqwkdkap
 slkftapfgceiytnpiraencavgsiplafldipdalftvsetptlsaaectlnecvy
 ssdfggiatvkysasksgkcavhvpsgtatlkeaavelteggssatihfstanihpefrl
 qictsytvckgdchppkdhivthpvyhaqftaavsktawtwltsllggsaviiiiglv
 lativamyvltlnqkhn.

- 5 [0059] In one embodiment, a first attachment site comprises an amino group, preferably an amino group of a lysine residue. In one embodiment, the second attachment site comprises

sulfhydryl group, preferably, a sulfhydryl group of a cysteine.

[0060] In one embodiment, a conjugation of more than two substances (e.g. antigen and Chikungunya or Venezuelan equine encephalitis virus structural polypeptide) through a first attachment site or a second attachment site is achieved using chemical cross linker. Examples of the cross-linker include, but are not limited to, SMPH, Sulfo-MBS, Sulfo-EMCS, Sulfo-GMBS, Sulfo-SIAB, Sulfo-SMPB, Sulfo-SMCC, SVSB, SIA and other cross-linkers available from the Pierce Chemical Company.

[0061] According to the present invention, a Chikungunya or Venezuelan equine encephalitis virus like particle comprising a Chikungunya or Venezuelan equine encephalitis virus structural polypeptide and an antigen, wherein said Chikungunya or Venezuelan equine encephalitis virus structural polypeptide and said antigen are expressed as a fusion protein can be provided.

[0062] In one embodiment, the antigen can be fused with any site of the Chikungunya or Venezuelan equine encephalitis virus structural polypeptide. For example, the antigen may be directly or indirectly linked to N- or C- terminal of the Chikungunya or Venezuelan equine encephalitis virus structural polypeptide, or the antigen may be inserted into

Chikungunya or Venezuelan equine encephalitis virus structural protein.

[0063] In one embodiment, at least one antigen is inserted into E2 of Chikungunya or Venezuelan equine encephalitis virus structural protein. For example, regarding Chikungunya virus structural protein, at least one antigen is inserted between residues 519 and 520 of SEQ ID Nos.1 or 2 (i.e. between G at 519-position and Q at 520-position of SEQ ID Nos.1 or 2); between residues 530 and 531 of SEQ ID Nos.1 or 2 (i.e. between G at 530-position and S at 531-position of SEQ ID Nos.1 or 2); between residues 531 and 532 of SEQ ID Nos.1 or 2 (i.e. between S at 531-position and N at 532-position of SEQ ID Nos.1 or 2); between residues 529 and 530 of SEQ ID Nos.1 or 2 (i.e. between G at 529-position and G at 530-position of SEQ ID Nos.1 or 2); or between residues 510 and 511 of SEQ ID Nos.1 or 2 (i.e. between S at 510-position and G at 511-position of SEQ ID Nos.1 or 2); or between residues 511 and 512 of SEQ ID Nos.1 or 2 (i.e. between G at 511-position and N at 512-position of SEQ ID Nos.1 or 2); or between residues 509 and 510 of SEQ ID Nos.1 or 2 (i.e. between Q at 509-position and S at 510-position of SEQ ID Nos.1 or 2).

[0064] For example, regarding Venezuelan equine encephalitis virus structural protein, at least one antigen is inserted between residues 517 and 518 of SEQ ID No.3 (i.e.

between G at 517-position and S at 518-position of SEQ ID No.3); between residues 518 and 519 of SEQ ID No.3 (i.e. between S at 518-position and S at 519-position of SEQ ID No.3); between residues 519 and 520 of SEQ ID No.3 (i.e. between S at 519-position and V at 520-position of SEQ ID No.3); between residues 515 and 516 of SEQ ID No.3(i.e. between L at 515-position and S at 516-position of SEQ ID No.3); between residues 516 and 517 of SEQ ID No.3 (i.e. between S at 516-position and G at 517-position of SEQ ID No.3); between residues 536 and 537 of SEQ ID No.3(i.e. between C at 536-position and G at 537-position of SEQ ID No.3) ; between residues 537 and 538 of SEQ ID No.3(i.e. between G at 537-position and G at 538-position of SEQ ID No.3) ; between residues 538 and 539 of SEQ ID No.3(i.e. between G at 538-position and T at 539-position of SEQ ID No.3) .

[0065] The fusion protein may be expressed using a conventional technique in the art. A variety of expression systems can be used for the expression of the fusion protein. For example, the fusion protein can be expressed in 293 cells, Sf9 cells or E.coli.

[0066] A polypeptide derived from Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) may be a naturally occurring viral polypeptide or modified polypeptide thereof. In addition, a polypeptide derived

from malaria antigen may be a naturally occurring polypeptide or modified polypeptide of the naturally occurring polypeptide or a fragment of the naturally occurring polypeptide or the modified peptide. The modified
5 polypeptide may be a fragment of the naturally occurring virus structural polypeptide.

[0067] In one embodiment, the modified polypeptide derived from malaria antigen has at least 70%, 75%, 80%, 85%, 90%, 95% or 98% amino acid sequence identity to a naturally
10 occurring polypeptide. In one embodiment, the modified peptide derived from malaria antigen is a mutant where at most 10% of the amino acids are deleted, substituted, and/or added based on a naturally occurring polypeptide derived from malaria antigen.

15 [0068] When a polypeptide derived from a virus is conjugated with a polypeptide derived from an antigen, a linker peptide including SG, GS, SGG, GGS SGSG and TRGGS may be used. Examples of conjugation of the polypeptide derived from a virus (referred to as "PFV" below) with the
20 polypeptide derived from the antigen (referred to as "PFA" below) include, but not limited to: PFV-SG-PFA-GS-PFV; PFV-SG-PFA-GGS-PFV; PFV-SSG-PFA-GS-PFV; PFV-SGG-PFA-GGS-PFV; PFV-SGSG-PFA-GS-PFV; and PFA-SGG-PFA-TRGGS-PFV.

[0069] In one embodiment, the present invention provides a
25 virus like particle comprising a fusion protein of a

polypeptide derived from Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) and a polypeptide derived from malaria antigen, wherein the virus like particle is prepared by transfecting an expression
5 vector comprising a nucleic acid molecule corresponding to the amino acid sequence represented by SEQ ID NO. 28, 31, 34, 37, 39, 41 or 43 into a mammalian cell (e.g. 293F cell). Regarding this embodiment, modified fusion protein can be also used for a virus like particle provided by the present
10 invention, which can be prepared by transfecting an expression vector comprising a nucleic acid molecule corresponding to the amino acid sequence having at least 70%, 75%, 80%, 85%, 90%, 95% or 98% amino acid sequence identity to SEQ ID NO. 28, 31, 34, 37, 39, 41 or 43 into a mammalian
15 cell (e.g. 293F cell).

[0070] In one embodiment, the present invention provides a virus like particle comprising or consisting of:
one or more capsid of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV);
20 one or more E1 of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV); and
one or more E2 of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV), wherein malaria antigen is inserted into E2 of Chikungunya virus (CHIKV) or Venezuelan
25 equine encephalitis virus (VEEV). For example, present

invention provides a virus like particle comprising or consisting of:

240 capsids of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV);

5 240 E1s of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV); and

240 E2s of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV), wherein malaria antigen is inserted into each of E2s of Chikungunya virus (CHIKV) or
10 Venezuelan equine encephalitis virus (VEEV).

[0071] In this embodiment, the E2 into which the antigen is inserted may consist of an amino acid sequence represented by SEQ ID No.50; the E1 may consist of an amino acid sequence represented by SEQ ID No.51; and the capsid may consist of
15 an amino acid sequence represented by SEQ ID NO.: 52; or the E2 into which the antigen is inserted may consist of an amino acid sequence represented by SEQ ID NO.53; the E1 may consist of an amino acid sequence represented by SEQ ID NO.54; and the capsid may consist of an amino acid sequence
20 represented by SEQ ID NO.: 55.

[0072] Further, regarding this embodiment, modified capsid of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV), modified E1 of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) and
25 modified E2 of Chikungunya virus (CHIKV) or Venezuelan

equine encephalitis virus (VEEV) may be used for the virus like particle. For example, the modified capsid of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) may have at least 70%, 75%, 80%, 85%, 90%, 95% or 98% amino acid sequence identity to the amino acid sequence represented by SEQ ID NO.: 52 or SEQ ID No.:55; the modified E1 of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) may have at least 70%, 75%, 80%, 85%, 90%, 95% or 98% amino acid sequence identity to the amino acid sequence represented by SEQ ID NO.: 51 or SEQ ID No.:54; and/or the modified E2 of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) may have at least 70%, 75%, 80%, 85%, 90%, 95% or 98% amino acid sequence identity to the amino acid sequence represented by SEQ ID NO.: 50 or SEQ ID No.:53.

Also, the modified capsid, E1 or E2 may be a mutant where at most 10% of the amino acids are deleted, substituted, and/or added based on the capsid consisting of an amino acid sequence represented by SEQ ID NO.: 52 or SEQ ID No.:55; E1 consisting of an amino acid sequence represented by SEQ ID NO.: 51 or SEQ ID No.:54; and/or E2 consisting of an amino acid sequence represented by SEQ ID NO.: 50 or SEQ ID No.:53.

[0073]

(2) Nucleotide, Vector, Host cell

In the second aspect, the present invention provides a nucleic acid molecule which is designed for expression of a particle as provided in the first aspect of the present invention.

[0074] In one embodiment, the present invention provides a nucleic acid molecule comprising a nucleotide sequence that encodes the Chikungunya or Venezuelan equine encephalitis virus like particle as described above.

[0075] Examples of the nucleotide sequence that encodes the Chikungunya or Venezuelan equine encephalitis virus like particle include, but are not limited to, a nucleotide sequence encoding envelope of Chikungunya virus Strain 37997, a nucleotide sequence encoding Capsid-envelope of Chikungunya virus Strain 37997, a nucleotide sequence encoding envelope of Chikungunya virus Strain LR2006 OPY-1, a nucleotide sequence encoding Capsid-envelope of Chikungunya virus LR2006 OPY-1, a nucleotide sequence encoding envelope of Venezuelan equine encephalitis virus Strain TC-83 and a nucleotide sequence encoding Capsid-envelope of Venezuelan equine encephalitis virus TC-83.

[0076] Regarding Chikungunya virus, an exemplary nucleotide sequence that encodes envelope is described below (SEQ ID No.:4):

Recue/Date Received 2022-07-06

Atgagtcttggccatccagttatgtgctgttggcaaacacacggttccctgctcccagcccccttgca
 cgccctgctgctacgaaaaggaaacggaggaaacccctacgcatgcttgaggacaaagtcattgagacctgg
 gtactatcagctgctacaagcatccttaacatgttctccccacggccagcgacgcagcaaccaaggacaac
 ttcaatgtctataaagccaagaacatacttagctcactgtccgactgtggagaagggcactcgtgcc
 atagtcccgtagcactagaacgcattcagaatgaagcgacagacgggacgctgaaaaaccaggtctcctt
 gcaaatcgggaataaagacggatgacagccacgattggaccaagctgcgttatatggacaaccacatgcca
 gcagacgcagagaggcggggctatttgaagaacatcagcacggtgtacgattactggaacaatgggac
 acttcatcctggcccgatgtccaaaaggggaaactctgacggtgggattcactgacagtaggaagatag
 tcaactcatgtacgcacccatttcaccaagaccctcctgtgataggctgggaaaaattccattccgacgg
 cagcacggtaaaagactaccttgacgcacgtacgtgcagagcaccgcgcaactaccgaggagatagagg
 tacacatgccccagacaccctgatcgcacattaatgtcacacagtcggcaacgtaagatcacagt
 caatggccagacggtgcggtacaagtgttaattgcggtggctcaaatgaaggactaacaaactacagacaaa
 gtgattataaactgcaaggttgatcaatgtcatgcgcgggtcaccaatcacaaaaagtggcagtataact
 cccctctgggtcccgctaatgctgaacttggggaccgaaaaggaaaaattcacatcccgcttccgctggc
 aatgtaacatgcaggggtgctaaagcaaggaaacccacccgtgacgtacgggaaaaaacaagtcacatg
 ctactgtatcctgacaccccaacactcctgtcctaccggaatatgggagaagaaccaaactatcaagaag
 agtgggtgatgcataagaagggaagtgcgtacacgctgccgactgaagggtcgaggccagtggggcaa
 caacgagccgtataagtatggccgagttatctacaaaagggtacagcccatggccaccgcatgagata
 attctgtattatgatgctgtacccactatgactgtagtagttgtgtcagtgggccacggttcatactcc
 tgtcgatggtgggtatggcagcggggaagtgcattgtgtgcacgacgcagatgcatcacccgtatgaact
 gacaccaggagctacggtcccttccctgcttagcctaataatgctgcatacagaacagctaaagcggccaca
 taccagagggtgcgatatacctgtggaacgagcagcaaccccttggtttgggtacaagcccttatccgc
 tggcagccctgattgtctatgcaactgtctgagactcttaccatgctgctgtaaaaagcttggcttttt
 agccgtaatgagcgtcggtgccacactgtgagcgcgtacgaacacgtaacagtgatccgaacacggtg
 ggagtacggtataagactctagtcaatagacctggctacagccccatggtattggagatggaactactgt
 cagtcactttggagccaacactatcgcttgattacatcacgtgcgagtacaaaaacggtcatcccgctcc
 gtacgtgaagtgctgcgtacagcagagtgcaggacaaaaaacctacctgactacagctgtaaggctctc
 accggcgtctacccatttatgtggggcggcgcctactgctctgcgacgctgaaaacacgcagttgagcg
 aagcacacgtggagaagtccgaatcatgcaaacagaatttgcatcagcatacagggctcataccgcattc
 tgcattcagctaaagctccgcgtccctttaaacaaggaaataacatcactgtaactgcctatgcaaacggcgac
 catgcggtcacagttaaaggacgcaaaattcatgtggggccaatgtcttcagcctggacaccttccgaca
 acaaaattgtgggtgtaaaaagggtgacgtctataacatggactaccgccttggcgaggaagaccagg
 acaatttggcgatatccaaagtgcacacctgagagtaagacggtctatgctaatacacaactggtactg
 cagagacccggtgtgggtacgggtacacgtgccatactctcaggcaccatctggctttaagtattggctaa
 aagaacgcggggcgctgcgtgcagcacaagcaccatttggctgccaaatagcaaaaacccggtgaagagc
 ggtgaactgcgcgttagggaaacatgccatctccatcgacataaccggaagcggccttcactagggctgtc
 gagcgcctctttaacggacatgtcgtgcgaggtaccagcctgcacccattcctcagacttgggggctg
 tcgccattattaaatatgcagccagcaagaaaggcaagtgtgcggtgcatcogatgactaacgcctcac
 tattcgggaagctgagatagaagtgaagggaattctcagctgcaaatctcttctcgcagcggccttagcc
 agcgcgaattcccgctacaagttctgtctacacaagtaactgtgcagccgagtgccacccccgaagg
 accacatagtcaactaccggcggtcacataccaccctcggggtccaggacatctccgctacggcgatgtc
 atgggtgcagaagatcacgggaggtgtgggactggtgtgtgtgtgctgttgccgactgattctaatcgtggtg
 ctatgcgtgtcgttcagcaggcac

[0078] Regarding Chikungunya virus, an exemplary nucleotide
 sequence that encodes a Capsid-envelope is described below
 (SEQ ID No.:6):

atggagttcatcccgacgcaaactttctataacagaaggtaccaaccccgaccctgggc
cccacgccctacaattcaagtaattagacctagaccacgtccacagaggcaggctgggc
aactcgcccagctgatctccgcagtcaacaaattgaccatgcgcgcggtacctcaacag
aagcctcgagaaatcggaaaaacaagaagcaaaggcagaagaagcaggcgccgcaaaa
cgacccaaagcaaaagaagcaaccaccacaaaagaagccggctcaaaagaagaagaaac
caggccgtagggagagaatgtgcatgaaaattgaaaatgattgcatcttcgaagtcaag
catgaaggcaaaagtgatgggctacgcatgcctgggtgggggataaaagtaatgaaaccagc
acatgtgaagggaactatcgacaatgccgatctggctaaactggcctttaagcggctcgt
ctaaatacgatcttgaatgtgcacagataccgggtgcacatgaagtctgatgcctcgaag
tttaccacagagaaacccgaggggtactataactggcatcacggagcagtgacagtattc
aggaggccggttcactatcccgacgggtgcaggcaagccgggagacagcggcagaccga
tcttcgacaacaaaggacgggtgggtggccatcgtcctaggagggggccaacgaagggtgcc
cgacggccctctccgtggtgacgtggaacaaagacatcgtcacaaaaattaccctga
gggagccgaagagtggagcctcgccctcccggtcttgtgcctgttggcaaacactacat
tcccctgctctcagccgccttgcacaccctgctgctacgaaaaggaaccggaaagcacc

ttgcgcattgcttgaggacaacgtgatgagacccggatactaccagctactaaaagcatc
 gctgacttgcctccccaccgccaagacgcagtaactaaggacaattttaatgtctata
 aagccacaagaccatatctagctcattgtcctgactgcggagaagggcattcgtgccac
 agccctatcgcattggagcgcacagaaatgaagcaacggacggaacgctgaaaatcca
 ggtctctttgcagatcgggataaagacagatgacagccacgattggaccaagctgcgt
 atatggatagccatacgcagcggacgcggagcgcgcggattgcttgtaaggacttca
 gcaccgtgcacgatcaccgggaccatgggacactttattctcgcccgatgcccgaagg
 agagacgctgacagtgggatttacggacagcagaaagatcagccacacatgcacacacc
 cgttccatcatgaaccacctgtgataggtaggagaggttccactctcgaccacaacat
 ggtaaagagttaccttgcagcacgtacgtgcagagcaccgctgccactgctgaggagat
 agaggtgcatatgccccagatactcctgaccgcacgctgatgacgcagcagctctggca
 acgtgaagatcacagttaatgggcagacgggtgcggtacaagtgcactgcggtggctca
 aacgagggactgacaaccacagacaaaagtgcataaactgcaaaattgatcagtgcca
 tgctgcagtcactaatcacaagaattggcaatacaactcccccttagtccgcgcgaacg
 ctgaactcggggaccgtaaaaggaaagatccacatcccatccattggcaaacgtgact
 tgcagagtgcacaaaagcaagaaaccctacagtaacttacggaaaaaaccaagtcaccat
 gctgctgtatcctgaccatccgacactcctgtcttaccgtaacatgggacaggaaccaa
 attaccacgaggagtggtgacacacaagaaggaggttaccttgaccgtgcctactgag
 ggtctggaggtcacttggggcaacaacgaaccatacaagtaactggccgcagatgtctac
 gaacgggtactgctcatggtcaccacacatgagataatcctgtactattatgagctgtacc
 cactatgactgtagtcatgtgtcgggtggcctcgttcgtgcttctgtcgatgggtgggc
 acagcagtggggaatgtgtgtgtgcgcacgggcgcagatgcattacaccatatgaattaac
 accaggagccactgttcccttccctgctcagcctgctatgctgcgtcagaacgaccaagg
 cggccacatattacgaggctgcggcatatctatggaacgaacagcagccctgttctgg
 ttgcaggctcttatcccgctggccgccttgatcgtcctgtgcaactgtctgaaactctt
 gccatgctgctgtaagaccctggcttttttagccgtaatgagcatcgggtgccacactg
 tgagcgcgtacgaacacgtaacagtgtatccgaacacgggtgggagtaccgtataagact
 cttgtcaacagaccgggttacagcccatggtgttgagatggagctacaatcagtcac
 cttggaaccaacactgtcacttgactacatcacgtgcgagtaaaaaactgtcatccct
 ccccgtagtggaagtgtgtggtacagcagagtgcgaaggacaagagcctaccagactac
 agctgcaaggtctttactggagtctacccatttatgtggggcgggcgcctactgcttttg
 cgagcgcgaaaatacgcatttgagcgaaggcacatgtagagaaatctgaatcttgcaaaa
 cagagtttgcatcggcctacagagcccccacaccgcacatcggcgtcggcggaagctccgcgtc
 ctttaccaggaacaaacattaccgtagctgcctacgctaaccggtgacctgcccgtcac
 agtaaaggacgccaagtttgcgtgggcccattgtcctccgcctggacaccttttgaca
 acaaaatcgtggtgtacaaaggcgacgtctacaacatggactaccacaccttttggcgca
 ggaagaccaggacaatttggtgacattcaaagtcgtacaccggaaagtaagacgttta
 tgccaacactcagttggtactacagaggccagcagcagggcacggtacatgtaccatact
 ctcaggcaccatctggcttcaagtattggctgaaggaacgaggagcatcgtacagcac
 acggcaccgttccggttgccagattgcgacaaacccggtaagagctgtaaattgcgctgt
 ggggaacataccaatttccatcgacataccggatgcggcctttactagggtgtcgtatg
 caccctctgtaacggacatgtcatgcgaagtaccagcctgcactcactcctccgacttt
 gggggcgtcgccatcatcaatacacagctagcaagaaaggtaaatgtgcagtacattc
 gatgaccaacgcggttaccattcgagaagccgacgtagaagtagaggggaaactcccagc
 tgcaaatatccttctcaacagccctggcaagcgcgcaggtttcgcgtgcaagtgtgctcc
 acacaagtacactgcgcagccgcgtgcccacctccaaaggaccacatagtcaattaccc
 agcatcacacaccaccttgggggtccaggatatatccacaacggcaatgtcttgggtgc
 agaagattacgggaggagtaggattaattgttgcgtgttgcgtgccttaattttaattgtg
 gtgctatgcgtgtcgttttagcaggcactaa.

[0079] Regarding Chikungunya virus, another exemplary
 nucleotide sequence that encodes a Capsid-envelope is
 described below (SEQ ID No.:7):

atggagttcatcccaacccaaactttttacaataggaggtaccagcctcgaccctggac
 tccgcgccctactatccaagtcacagggccagaccgcgccctcagaggcaagctgggc
 aacttgcccagctgatctcagcagtttaataaactgacaatgcgcgcggtaccacaacag
 aagccacgcaggaatcggaagaataagaagcaaaagcaaaaacaacaggcgccacaaaa
 caacacaaatcaaaagaagcagccacctaataaagaacccggctcaaaagaaaaagaagc
 cgggcccgcagagagaggatgtgcatgaaaatcgaaaatgattgtattttcgaagtcag
 cacgaaggttaaggtaacaggttacgcgtgcttgggtggggacaaagtaataaaaccagc
 acacgtaaaggggaccatcgataacgcggaccctggccaaactggcctttaagcgggtcat
 ctaagtatgacctgaatgcgcgcagatacccggtgcacatgaagtcgcagcgttcgaag
 ttcacccatgagaaaccggaggggtactacaactggcaccacggagcagtagtactc
 aggaggccgggttcacccatccctacaggtgctggcaaaccaggggacagcggcagaccga
 tcttcgacaacaagggacgcgtgggtggccatagtccttaggaggagctaataagaggacc
 cgtacagccctctcggtgggtgacctggaataaagacattgtcactaaaatcacccccga
 gggggccgaagagtgaggctcttgccatcccagttatgtgcctgttggcaaacaccacgt
 tccctgtctccagcccccttgacgcctctgctgctacgaaaaggaaccggaggaaacc
 ctacgcgtgcttgaggacaacgtcatgagacctgggtactatcagctgctacaagcatc
 cttacactgttctccccaccgcccagcagcagcaccaggaacttcaatgtctata
 aagccacaagaccatacttagctcactgtcccactgtggagaagggcactcgtgccat
 agtcccgtagcactagaacgcacagaaatgaagcgacagacgggacgctgaaaatcca
 ggtctccttgcaaatcggaataaagacggatgacagccacgattggaccaagctgcgtt
 atatggacaaccacatgccagcagacgcagagagggcggggtatattgtaagaacatca
 gcaccgtgtacgattactggaacaatgggacacttcatcctggcccgatgtccaaaagg
 ggaaactctgacgggtgggattcactgacagtaggaagattagtcactcatgtacgcacc
 catttccaccacgaccctcctgtgataggtcgggaaaaattccattccccgaccgcagcac
 ggtaaagagctaccttgacgcagctacgtgcagagcaccgcccgaactaccgaggagat
 agaggtaacacatgccccagacacccctgatcgacacattaatgtcacaacagtcgggca
 acgtaaagatcacagtcattggccagacgggtgcgggtacaagtgaattgcgggtgggtca
 aatgaaggactaacaactacagacaaagtgaattataactgcaagggtgactcaatgtca
 tgccgcgggtcaccaatcacaaaaagtggcagtagtataactccctctgggtcccgcgtaag
 ctgaacttggggaccgaaaaggaaaaattcacatcccggttccgctggcaaatgtaaca
 tgcagggtgcctaaagcaaggaaacccccaccgtgacgtacgggaaaaaccaagtcacat
 gctactgtatcctgaccacccaacactcctgtcctaccggaatatgggagaagaaccaa
 actatcaagaagagtggtgatgcataagaaggagtcgtgctaaccgtgccgactgaa
 gggctcgaggtcacgtggggcaacaacgagccgtataagatttggccgcagttatctac
 aaacgggtacagcccatggccaccgcgatgagataattctgtattattatgagctgtacc
 ccaactatgactgtagtagttgtgtcagtggccacgttcatactcctgtcgatgggtgggt
 atggcagcggggatgtgcatgtgtgcacgacgcagatgcacacccgtatgaactgac
 accaggagctaccgtcccttctcctgcttagcctaataatgctgcatcagaacagctaaag
 cggccacataccaagaggtgcgatatacctgtggaacgagcagcaaccttgttttgg
 ctacaagcccttattccgctggcagccctgattgttctatgcaactgtctgagactctt
 accatgctgctgtaaaaacgttggcttttttagccgtaatgagcgtcgggtgccacactg
 tgagcgcgtacgaacacgtaacagtgatcccgaacacgggtgggagtaccgtataagact
 ctagtcaatagacctggctacagcccatgggtattggagatggaactactgtcagtcac
 tttggagccaacactatcgcttgattacatcacgtgcgagtacaaaaccgtcatcccg
 ctccgtacgtgaagtgctgcgggtacagcagagtgcaaggacaaaaacctacctgactac
 agctgtaaggtcttccaccggcgtctacccatttatgtggggcgggcgccctactgcttctg
 cgacgctgaaaacacgcagttgagcggaagcacacgtggagaagtcgaatcatgcaaaa
 cagaatttgcatcagcatacaggggtcataccgcctctgcatcagctaagctccgcgtc
 ctttaaccaaggaaataacatcactgtaactgcctatgcaaacggcgaccatgccgtcac
 agttaaggacgccaattcattgtggggccaatgtcttcagcctggacaccttccgaca
 acaaaattgtgggtgtacaaaggtgacgtctataacatggactaccgccttggcgca
 ggaagaccaggacaatttggcgatatccaaagtcgcacacctgagagtaagacgtcta

tgctaatacacaaactggtactgcagagaccggctgtgggtacgggtacacgtgccatact
 ctcaggcaccatctggctttaagtattggctaaaagaacgcggggcgctcgctgcagcac
 acagcaccatttggctgccaaatagcaacaaacccggtaagagcggtgaactgcgccgt
 agggaacatgcccatctccatcgacataccggaagcggccttcactagggtcgtcgacg
 cgccctctttaacggacatgtcgtgaggtaccagcctgcaccattcctcagacttt
 gggggcgctcgccattattaaatagcagccagcaagaaaggcaagtgtgcggtgcattc
 gatgactaacgccgtcactattcggaagctgagatagaagttgaagggaattctcagc
 tgcaaactctctttctcgacggccttagccagcgcgaattccgcgtacaagtctgttct
 acacaagtacactgtgcagccgagtgccacccccgaaggaccacatagtcaactaccc
 ggcgtcacataccaccctcggggtccaggacatctccgctacggcgatgtcatgggtgc
 agaagatcacgggaggtgtgggactggtgtgtgctgttgccgcactgattctaatacgtg
 gtgctatgcgtgtcgttcagcaggcactaa.

[0080] In one embodiment, the present invention provides
 a vector comprising the nucleic acid molecule as described
 5 above, wherein the vector optionally comprises an expression
 control sequence operably linked to the nucleic acid molecule.

[0081] Examples of an expression control sequence include,
 but are not limited to, promoter such as CMV promoter, phage
 lambda PL promoter, the E. coli lac, phoA and tac promoters,
 10 the SV40 early and late promoters, and promoters of
 retroviral LTRs.

[0082] In this embodiment, the vector comprising an
 expression control sequence operably linked to the nucleic
 acid molecule as described above can be used as an expression
 15 vector for preparing the particle provided by the first
 aspect of the present invention.

[0083] The expression vectors can be prepared by a person
 skilled in the art based on WO/2012/006180.

[0084] Examples of vectors which can be used for expressing a virus like particle comprising a fusion protein of a polypeptide derived from Chikungunya virus (CHIKV) and a polypeptide of antigen include a vector shown in VLP_CHI 512
5 vector (SEQ ID No.:8) containing CHIKV VLP polynucleotide (SEQ ID No. 13; corresponding amino acid sequence represented by SEQ ID No.:14); and VLP_CHI 532 vector (SEQ ID No.: 9) containing CHIKV VLP polynucleotide (SEQ ID No. 15; corresponding amino acid sequence represented by SEQ ID
10 No.:16).

[0085] The expression vectors can be prepared by a person skilled in the art based on US2012/0003266.

[0086] Examples of vectors which can be used for expressing a virus like particle comprising a fusion protein of a
15 polypeptide derived from Venezuelan equine encephalitis virus (VEEV) and a polypeptide of antigen include a vector shown in VLP_VEEV VLP 518 vector (SEQ ID No.:10) containing VEEV VLP polynucleotide (SEQ ID No. 17; corresponding amino acid sequence represented by SEQ ID No.:18); VLP_VEEV VLP
20 519 vector (SEQ ID No.11) containing VEEV VLP polynucleotide (SEQ ID No. 19; corresponding amino acid sequence represented by SEQ ID No.:20); and VLP_VEEV VLP 538 vector (SEQ ID No.: 12) containing VEEV VLP polynucleotide (SEQ ID No. 21; corresponding amino acid
25 sequence represented by SEQ ID No.:22).

[0087] In one embodiment, the present invention provides a nucleic acid molecule which is designed for expression of a virus like particle comprising a fusion protein of a polypeptide derived from Chikungunya virus (CHIKV) or
5 Venezuela equine encephalitis virus (VEEV) and a polypeptide derived from malaria antigen, which consists of a nucleotide sequence represented by SEQ ID Nos.26-27, 29-30, 32-33, 35-36, 38, 40 or 42.

[0088] In one embodiment, the present invention provides
10 a nucleic acid molecule which is modified from the nucleic acid molecule having a nucleotide sequence represented by any one of SEQ ID Nos.26-27, 29-30, 32-33 or 35-36, 38, 40 or 42. The modified nucleic acid molecule may have at least 70%, 75%, 80%, 85%, 90%, 95% or 98% nucleotide sequence
15 identity to the nucleic acid molecule having a nucleotide sequence represented by any one of SEQ ID Nos.26-27, 29-30, 32-33, 35-36, 38, 40 or 42. Also, the modified nucleic acid molecule may be a mutant where at most 10% of the amino acids are deleted, substituted, and/or added based on the nucleic
20 acid molecule having a nucleotide sequence represented by any one of SEQ ID Nos.26-27, 29-30, 32-33, 35-36, 38, 40 or 42.

[0089]

(3) Composition or vaccine

In the third aspect, the present invention provides a composition or vaccine comprising the particle provided in the first aspect of the present invention and/or the nucleic acid molecule provided in the second aspect of the present invention.

[0090] In one embodiment, the present invention provides a composition or vaccine comprising the Alphavirus or Flavivirus virus like particle (e.g. Chikungunya virus like particle or Venezuelan equine encephalitis virus like particle) as described above or the nucleic acid molecule as described above. The content of the Alphavirus or Flavivirus virus like particle and the content of the nucleic acid molecule may be 0.00001-1 w/w%.

[0091] Dosage amount of the particle provided in the first aspect of the present invention (e.g. CHIKV VLP or VEEV VLP) may be 1-500µg/day.

[0092] One or more malaria antigens may be used for one composition or one vaccine provided by the third aspect of the present invention.

[0093] The composition or vaccine may further comprise a pharmaceutical acceptable carrier and/or adjuvant. Examples of adjuvant include, but are not limited to, aluminium salts, sodium hydroxide, Freund's complete adjuvant, Freund's

incomplete adjuvant and Ribi solution (Sigma Adjuvant system, Sigma-Aldrich). The composition or vaccine provided in the third aspect of the present invention may contain buffering agent such as dibasic sodium phosphate hydrate, sodium dihydrogen phosphate and sodium chloride; and preserving agent such as thimerosal. In one embodiment, the composition or vaccine is an aqueous solution containing 0.001-1 w/w% of the particle provided in the first aspect of the present invention (e.g. CHIKV VLP or VEEV VLP), 1-10w/w% of buffering agent, 0.01-1w/w% of adjuvant and 0.00001-0.001w/w% of preserving agent.

[0094] A skilled person can prepare the pharmaceutical composition and vaccine using conventional technique. For example, the particle provided in the first aspect of the present invention is mixed with buffer solution having physiological pH (e.g. pH 5-9, pH7) to prepare the pharmaceutical composition and vaccine provided in the third aspect of the present invention.

[0095] The pharmaceutical composition of the present invention may contain a single active ingredient or a combination of two or more active ingredients, as far as they are not contrary to the objects of the present invention. For example, cytokines including chemokines, anti-body of cytokines such as anti TNF antibody (e.g. infliximab, adalimumab), anti-VEGF antibody (e.g. bevacizumab and

ranibizumab), cytokine receptor antagonist such as anti HER2 antibody (e.g. Trastuzumab), anti EGF receptor antibody (e.g. Cetuximab), anti VEGF aptamer (e.g. Pegaptanib) and immunomodulator such as cyclosporine, tacrolimus, ubenimex
5 may be used for the combination therapy.

[0096] In a combination of plural active ingredients, their respective contents may be suitably increased or decreased in consideration of their therapeutic effects and safety.

[0097] The term "combination" used herein means two or more
10 active ingredient are administered to a patient simultaneously in the form of a single entity or dosage, or are both administered to a patient as separate entities either simultaneously or sequentially with no specific time limits, wherein such administration provides therapeutically
15 effective levels of the two components in the body, preferably at the same time.

[0098] In one embodiment, the composition is a vaccine composition including a DNA vaccine. In one embodiment, the DNA vaccine provided by the present invention comprises CpG
20 containing oligonucleotide.

[0099] The composition or vaccine provided in the third aspect of the present invention can be administered one or more times. When the composition or vaccine provided in the third aspect of the present invention is administered more
25 than one time, different particle provided in the first

aspect of the present invention (e.g. CHIKV VLP or VEEV VLP) may be used for each of the administration. In one embodiment, combination of immunization using CHIKV VLP provided in the first aspect of the invention and
5 immunization using VEEV VLP provided in the first aspect of the invention is employed. For example, CHIKV VLP provided in the first aspect of the present invention may be used for the 1st immunization and VEEV VLP provided in the first aspect of the present invention may be used for the 2nd
10 immunization, or VEEV VLP provided in the first aspect of the present invention may be used for the 1st immunization and CHIKV VLP provided in the first aspect of the present invention may be used for the 2nd immunization.

[0100] A skilled person can determine timing of
15 immunization using the composition or vaccine provided in the third aspect of the present invention. For example, 2nd immunization is performed 1, 2, 3, 4, 5, 6, 7, 8, 9 or 10 weeks after 1st immunization.

[0101] In one embodiment, the present invention provides
20 a kit comprising

- (a) a vaccine composition comprising the particle provided in the first aspect of the present invention; and
- (b) another vaccine composition comprising the particle provided in the first aspect of the present invention,

wherein the particle contained in (a) is a virus like particle which is different from the particle contained in (b). In this embodiment, the particle contained in (a) may be Chikungunya virus like particle and the particle contained in (b) may be Venezuelan equine encephalitis virus like particle.

[0102] In one embodiment, the present invention provides a kit comprising

(a) a vaccine composition comprising the particle provided in the first aspect of the present invention; and

(b) another vaccine composition comprising the particle provided in the first aspect of the present invention,

(c) one or more vaccine composition, each of which comprises the particle provided in the first aspect of the present invention,

wherein (a) is used for priming immunization and (b) and (c) are used for boosting immunization; and the particle contained in (a) is a virus like particle which is different from the particle contained in (b); and the particle contained in (c) is different from the particle contained in (a) and (b), or the same as the particle contained in (a) or (b).

[0103] The respective vaccine compositions contained in the above-described kit may be administered simultaneously, separately or sequentially.

[0104] The Alphavirus or Flavivirus virus like particle (e.g. Chikungunya virus or Venezuelan equine encephalitis virus) provided in the first aspect of the present invention or the nucleic acid molecule provided by the second aspect of the invention can be used for the composition and vaccine provided in the third aspect of the present invention.

[0105] For example, Chikungunya or Venezuelan equine encephalitis virus like particle comprising or consisting of:

10 one or more (e.g. 240) capsid of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV);

one or more (e.g. 240) E1 of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV); and

one or more (e.g. 240) E2 of Chikungunya virus (CHIKV) or

15 Venezuelan equine encephalitis virus (VEEV), wherein malaria antigen is inserted into E2 of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) may be used for preparing the composition or vaccine provided in the third aspect of the present invention. The E2 into which the

20 antigen is inserted may consist of an amino acid sequence represented by SEQ ID No.50; the E1 may consist of an amino acid sequence represented by SEQ ID No.51; and the capsid may consist of an amino acid sequence represented by SEQ ID NO.: 52; or

the E2 into which the antigen is inserted may consist of an amino acid sequence represented by SEQ ID NO.53; the E1 may consist of an amino acid sequence represented by SEQ ID NO.54; and the capsid may consist of an amino acid sequence represented by SEQ ID NO.: 55.

[0106] The composition or vaccine provided in the third aspect of the present invention can be administered to a mammal (e.g. human) intramuscularly (i.m.), intracutaneously (i.c.), subcutaneously (s.c.), intradermally (i.d.) or intraperitoneally (i.p.).

[0107] The composition or vaccine provided in the third aspect of the present invention may be used for treating or preventing malaria.

[0108] Thus, use of the Alphavirus or Flavivirus (e.g. Chikungunya virus or Venezuelan equine encephalitis virus) virus like particle provided in the first aspect of the present invention or the nucleic acid molecule provided by the second aspect of the invention for manufacturing a pharmaceutical composition or vaccine for treating or preventing malaria is also provided by the present invention.

[0109]

(4) Method of producing an antibody, Method of immunomodulation, Method of treating malaria, Method of inducing and/or enhancing immune response against a malaria antigen in a mammal, Method of passive immunization, Method of presenting an antigen on macrophage, and Method for producing a particle

In the fourth aspect, the present invention provides a method of producing an antibody, comprising contacting the particle provided in the first aspect of the present invention and/or the nucleic acid molecule provided in the second aspect of the present invention to a mammal.

[0110] The antibody produced in the fourth aspect of the present invention may be humanized using a conventional technique. Thus, in one embodiment, the method provided in the fourth aspect of the invention further comprises a step of humanizing non-human mammal produced antibody.

[0111] The particle provided in the first aspect of the present invention and/or the nucleic acid molecule provided in the second aspect of the present invention may be administered directly into the patient, into the affected organ or systemically, or applied ex vivo to cells derived from the patient or a human cell line which are subsequently administered to the patient, or used in vitro to select a

subpopulation from immune cells such as B-cell and T-cell derived from the patient, which are then re-administered to the patient.

[0112] According to the present invention, the virus like
5 particle can be applied for the immune therapy.

[0113] In the fifth aspect, the present invention provides a method of immunomodulation, a method of treating malaria, a method of inducing and/or enhancing immune response against a malaria antigen in a mammal comprising administering the
10 composition provided in the third aspect of the present invention to a mammal.

[0114] In sixth aspect, the present invention provides a method of passive immunization against a malaria-causing pathogen, comprising administering the antibody provided in
15 the fourth aspect of the present invention to a mammal.

[0115] In seventh aspect, the present invention provides a method of presenting a malaria antigen on macrophage, comprising contacting the particle provided in the first aspect of the present invention and/or the nucleic acid
20 molecule provided in the second aspect of the present invention to a mammal.

[0116] In eighth aspect, the present invention provides a method for producing the particle provided in the first aspect of the present invention, comprising preparing a vector designed for expression of said particle; culturing
5 a cell which is transfected with said vector to express said particle; and recovering said particle.

[0117] Examples of mammal include, but are not limited to, a human.

[0118] In one embodiment, the present invention provides a
10 method of producing an antibody against malaria antigen, comprising contacting the Chikungunya or Venezuelan equine encephalitis virus like particle as described above and/or the nucleic acid molecule as described above to a mammal. The produced antibody may be an antibody which can
15 specifically bind to a malaria antigen comprised in the Chikungunya or Venezuelan equine encephalitis virus like particle or a malaria antigen encoded by the nucleic acid molecule. The method of producing an antibody provided by the present invention may be a useful for producing a
20 monoclonal or polyclonal antibody against a malaria antigen.

[0119] In one embodiment, an antibody against malaria antigen obtained by the method of producing an antibody according to the present invention is used for passive

immunization. The method of passive immunization may comprise administering the obtained antibody to a mammal.

[0120] In one preferred embodiment, the immunomodulation provided by the present invention is inducing and/or
5 enhancing immune response against malaria antigen in a mammal. Thus, in one embodiment, the present invention provides a method of inducing and/or enhancing immune response against malaria antigen in a mammal, comprising administering an effective amount of the composition as described above to
10 the mammal.

[0121] Given the symptom of patients infected with Chikungunya or Venezuelan equine encephalitis together with unusual big molecule of Chikungunya or Venezuelan equine encephalitis, this VLP can act effectively and efficiently
15 to target macrophage and its composition such as cytokines and immunomodulative compounds.

[0122] In one aspect, the present invention provides a method of presenting an antigen on macrophage, comprising
20 administering the Chikungunya or Venezuelan equine encephalitis virus like particle as described above and/or the nucleic acid molecule as described above to a mammal. The Chikungunya or Venezuelan equine encephalitis virus like particle provided by the present invention is good to target

macrophage. In one embodiment, the Chikungunya or Venezuelan equine encephalitis virus like particle provided by the present invention is a kind of delivery system of the at least one antigen, which is comprised in the Chikungunya or
5 Venezuelan equine encephalitis virus like particle, to macrophage.

[0123] In one embodiment, the present invention provides a method for producing Chikungunya or Venezuelan equine encephalitis virus like particle provided in the first aspect
10 of the present invention, comprising preparing a vector designed for expression of said particle; culturing a cell which is transfected with said vector to express said particle; and recovering said particle. In this embodiment, transfection can be conducted using a conventional method.
15 Cells using for the transfection may be 293 cells. Recovering VLP may include collecting a conditioned medium after cells are transfected with a vector, and may further include purify VLP from the conditioned medium using ultracentrifugation.

20 [0124] The present invention will be described in detail with reference to the following example, which, however, is not intended to limit the scope of the present invention.

[0125]

EXAMPLES

EXAMPLE 1: Preparation of Chikungunya virus (CHIKV) like
particle comprising a virus structural polypeptide and a
fragment of malaria antigen

The following polynucleotides of malaria CSP1 proteins are used. N terminal linker is SGG and C terminal linker is GGS.

VLP74 (6 repeat of NPNA amino acid sequence)

10 Sgggnpnanpnanpnanpnanpnanpnaggs (SEQ ID No.:46)

(Tccggaggaaacccgaatgccaatcccaacgcgaaccccaatgctaaccctaatgcc
acccaaacgccaaccccaacgctgggtggatcc) (SEQ ID No.:47)

VLP78 (25 repeat of NPNA amino acid sequence)

15 Sggnpnanpnanpnanpnanpnanpnvdnpnanpnanpnanpnanpnanpnanpnanpna
npnanpnanpnanpnanpnanpnanpnanpnanpnanpnanpnanpnaggs (SEQ ID
No.: 48)

(tccggaggaaacccgaatgccaatcccaacgcgaacccccaacgcctaaccaccaacgcga
atccgaatgcaaaccgaacgttgacccaaacgcgaacccgaatgccaatcccaacgcg
aaccaccaatgctaaccacaaatgccaaacccaaacgcgaaccccaacgcctaataccaaacgc
20 caaccctaacgcgaatcccaacgcgaatcctaacgcctaatacccaacgcgaatcccaatg
ctaataccgaacgcgaaccctaatgcaaaccccaacgcgaacccgaacgcctaaccgaac
gctaatacccaacgccggtgatcc) (SEQ ID No.:49)

[0126] The respective polynucleotides was inserted between the codons encoding Ser at 531-position and Asn at 532-position of SEQ ID Nos.15 or 16 (SEQ ID Nos.1 or 2) to construct a plasmid (hereinafter referred to as CHIKV-VLP74 or 78) for expressing Chikungunya virus like particle where
5 the modified VLP74 or 78-derived peptide is inserted into E2 of Chikungunya virus structural polypeptide.

[0127] 293F cells (Lifetechnology) were transfected with the plasmid using PEI (GE Healthcare) or GeneX (ATCC). 4
10 days after the transfection, the conditioned medium was collected and centrifuged at 3000rpm for 15 minutes to separate it from cells. The supernatant was filtrated using 0.45µm filter to obtain virus like particles. The virus like particles were concentrated using TFF column and
15 purified using QXL column (GE Healthcare) to obtain purified virus like particles. When animals were immunized with virus like particles, the purified virus like particles were further concentrated using spin column (Molecular Weight-cutoff: 100kDa) to prepare the virus like particles for the
20 immunization.

[0128] The expression of VLP comprising VLP74 or 78 conjugated with Chikungunya virus structural polypeptide was confirmed by Western Blot using an antibody specific for

CHIKV (ATCC: VR-1241AF) and an antibody specific for VLP74 or 78.

[0129]

EXAMPLE 2: Preparation of Venezuelan equine encephalitis virus (VEEV) like particle comprising a virus structural polypeptide and a fragment of malaria antigen

The same polynucleotides of malaria CSP1 proteins (VLP74 and VLP78) used in EXAMPLE 1 are used. N terminal linker is SGG and C terminal linker is GGS.

[0130] The respective polynucleotides was inserted between the codons encoding Ser at 518-position and Ser at 519-position of SEQ ID Nos.19 or 20 (SEQ ID No.3) to construct a plasmid (hereinafter referred to as VEEV-VLP74 or 78) for expressing Venezuelan equine encephalitis virus like particle where the modified VLP74 or 78 -derived peptide is inserted into E2 of Venezuelan equine encephalitis virus structural polypeptide.

[0131] 293F cells (Lifetechnology) were transfected with the plasmid using PEI (GE Healthcare) or GeneX (ATCC). 4 days after the transfection, the conditioned medium was collected and centrifuged at 3000rpm for 15 minutes to separate it from cells. The supernatant was filtrated using

0.45µm filter to obtain virus like particles. The virus like particles were concentrated using TFF column and purified using QXL column (GE Healthcare) to obtain purified virus like particles. When animals were immunized with virus like particles, the purified virus like particles were further concentrated using spin column (Molecular Weight-cutoff: 100kDa) to prepare the virus like particles for the immunization.

[0132] The expression of VLP comprising VLP 74 or 78 conjugated with Venezuelan equine encephalitis virus structural polypeptide was confirmed by Western Blot using an antibody specific for VEEV and an antibody specific for VLP74 or 78.

[0133]

15 EXAMPLE 3:Immunogenicity in Non-human Primate (Monkey)

The monkeys were immunized with x25-CHI (80ug) at 0 week and x6-VEE (80ug) at 3 week by intramuscular injection with or without adjuvant (Sigma Adjuvant System, Sigma, S6322). X25-CHI means 25 times malaria CSP repeat amino acid NPNA on CHIKV VLP particle (VLP78_15). X6-VEE means 6 times malaria CSP repeat amino acid NPNA on VEEV VLP particle (VLP74_25). The blood is taken at 2 and 5 weeks after the first immunization.

[0134] 96 well ELISA plate were coated with 50ng of Recombinant Circumsporozoite Protein (rCSP) (Reagent Proteins, ATG-422) in 100ul PBS buffer pre well. The Plates after 2 hours incubation were washed three times TBS buffer containing 0.05% Tween-20 and blocked with TBS buffer containing 0.05% Tween-20 and 5% dry milk. The heat inactivated diluted serum from monkeys were added in the blocking buffer and incubated for 1 h at room temperature. After washing three times, peroxidase labeled goat anti-human IgG or anti-mouse IgG was added at 1:4000 dilution and incubated for 1h at room temperature. After washing three times, Peroxidase substrate was added for development and incubated for 10 mins and 2N H2SO4 was added to stop the development. The data were analyzed using Gen5 (BioTek) and GraphPad Prism6 (GraphPad software Inc).

[0135] The Immunogenicities are shown in Figures 4 to 6.

[0136] Induction of antibodies against CSP was found in the serum of all monkeys immunized with Malaria VLPs (see Figure 4). The mean OD values indicating titer of antibodies against CSP is shown in Figure 5. Figure 5 shows that the serum from immunized monkeys induced high titer of antibodies against CSP.

[0137] As seen in Figure 6, higher titer of antibodies against CSP was achieved when CHIKV VLP particle comprising NPNA and VEEV VLP particle comprising NPNA were used for the priming immunization and boosting immunization, respectively, compared to when only CHIKV VLP particle comprising NPNA was used for both of the priming immunization and the boosting immunization. In addition, Figure 6 shows that use of adjuvant further enhanced the titer of antibodies against CSP. Further, Figure 6 shows that administration of 25-repeats of NPNA induces higher titer of antibodies against CSP compared to administration of 6-repeats of NAPA.

[0138] The anti-Pf CSP antibody titer in the serum from the monkeys immunized with x25-CHI (80ug) at 0 week and x6-VEE (80ug) at 3 week without using adjuvant was measured by ELISA at Malaria Serology Lab Malaria Vaccine Branch, WRAIR. In the ELISA performed at Malaria Serology Lab Malaria Vaccine Branch, WRAIR, the plates were coated with CSPrp ((NPNA)6 Peptide) [0.2µg/µL] (Supplier: Eurogentec EP070034) and Goat α-Human IgG (KPL/074-1002 LOT# 120714) was used as 2nd antibody. The final titer was determined by the dilution factor that yields an OD of 1.0 (414nm).

[0139] As a result, the anti-Pf CSP antibody titer in the serum from the monkeys was enhanced after 2nd immunization compared to 1st immunization (see Table 2). Compared to the

anti-Pf CSP antibody titer in the serum from the monkeys immunized with RTS,S (GlaxoSmithKline), the anti-Pf CSP antibody titer in the serum from the monkeys immunized with x25-CHI (80ug) and x6-VEE (80ug) in the absence of adjuvant was considered to be higher even though RTS,S (GlaxoSmithKline) contains adjuvant.

[Table 2]

Animal No.	After 1st immunization	After 2nd immunization
	Week 2	Week 5
1	8990	29420
2	48210	44100
3	80400	51230
4	16260	19640
Geometric mean	27359	33801

[0140]

10 Example 4: Immunogenicity in mouse

The mice immunized with 10ug of VLP78_15 at week 0, 10ug of VLP74_25 at week3 and 10ug of VLP78_15 at week 6 with or without adjuvant (Sigma Adjuvant System, Sigma, S6322) by intramuscular injection.

[0141] The anti-Pf CSP antibody titer in the serum from the immunized mice were measured by ELISA at Malaria Serology Lab Malaria Vaccine Branch, WRAIR, where plates were coated with CSPrp ((NPNA)6 Peptide) [0.2µg/µL] (Supplier: Eurogentec EP070034) and Goat α-Mouse IgG (KPL/074-1806 LOT# 100737) is used as 2nd antibody to detect the antibodies in the serum.

[0142] The final titer was determined by the dilution factor that yields an OD of 1.0 (414nm).

10 [0143] The Immunogenicity are shown in Tables 3 and 4.

[0144] Tables 3 and 4 show that higher titer of antibodies against CSP was achieved after immunizing virus like particle three times. In addition, Tables 3 and 4 show that use of adjuvant enhanced the titer of antibodies against CSP.

15 [Table 3]

Mouse Week 3 (after 1st immunization)

Mouse No.	VLP	VLP+Adjuvant
1	6070	QNS
2	5850	13680
3	9610	7610
4	5440	23370
5	16320	27390
Geometric Mean	7875	16066

QNS=Quantity not sufficient to test

[Table 4]

Mouse Week 9 (after 1st immunization)

Mouse No.	VLP	VLP+Adjuvant
1	35510	729000
2	15040	197800
3	41650	106700
4	37250	436000
5	48200	497600
Geometric Mean	33134	319666

[0145]

Example 5: Immunogenicity of P. yoelii CSP inserted VLP in mice

QGPGAP seen in the rodent malaria CSP (P.yoelii CSP) was used as an antigen. 6x QGPGAP was inserted into CHIKV VLP. The mice were immunized with the CHIKV VLP 2 times at 0 and 8 week (20ug VLP per mouse) by intramuscle injection with or without Adjuvant Ribi.

[0146] Immunogenicity was confirmed at 4, 6, 10 and 14 weeks after the first immunization. The anti- P. yoelii CSP antibody were measured by ELISA. The ELISA was performed in the same way as ELISA described in Example 3 except that the plates were coated with P. Yoelii CSP repeat sequence peptide at 0.1 ng/μl. The secondary antibody was anti-mouse IgG HRP (Cell signal, #7076S). The results are shown in Figures 7-9.

[0147] Figures 7-9 show that higher titer of antibodies against CSP was achieved by intramuscular administration of CHIKV VLP comprising 6X QGPGAP. In addition, Figures 7-9 show that use of adjuvant enhanced the titer of antibodies
5 against CSP.

[0148]

Example 6: Protection of mice against malaria by intramuscle injection of CHIKV VLP comprising P. yoelii CSP epitope: 6x QGPGAP
10

6x QGPGAP was inserted into CHIKV VLP. The mice (n=5) were immunized with the CHIKV VLP 2 times at 0 and 8 week (20ug VLP per mouse) by intramuscle injection with or without Adjuvant Ribi (see Figure 10). Rodent malaria: P. yoelii-
15 challenge (i.v.) was conducted at 17 weeks(see Figure 10).

[0149] Malaria infection was confirmed by PCR. Genomic DNA was purified from the mice blood day 6 after challenge. 18S malaria DNA was amplified by PCR. Figure 11 shows results of the PCR, indicating that all of 5 control mice
20 (PBS injection) were infected with malaria; all of 5 mice immunized with Control VLP were infected with malaria; among 5 mice immunized with CHIKV VLP comprising 6x QGPGAP, 2 mice were infected with malaria and 3 mice were not infected with malaria; and among 5 mice immunized with CHIKV VLP comprising

6x QGPGAP with adjuvant: Ribi, 1 mouse was infected with malaria and 4 mice were not infected with malaria.

[0150]

Example 7: Preparation of vaccine composition comprising
5 Chikungunya virus (CHIKV) like particle comprising NPNA
repeat or Venezuelan equine encephalitis virus (VEEV) like
particle comprising NPNA repeat

Chikungunya virus (CHIKV) like particle comprising 6x
or 25x NPNA was prepared according to Example 1, and
10 Venezuelan equine encephalitis virus (VEEV) like particle
comprising 6x NPNA was prepared according to Example 2.

To prepare a vaccine composition, 80 μ g of each of the
prepared particles was mixed with 1ml of Sucrose Phosphate
Solution, pH 7.2, Endotoxin Free (Teknova, SP buffer).

CLAIMS

1. A virus-like particle comprising a virus structural polypeptide and at least one malaria antigen,

5 wherein said virus structural polypeptide is a polypeptide of Chikungunya virus (CHIKV) or Venezuelan equine encephalitis virus (VEEV) and comprises the capsid and the envelope proteins E1 and E2,

 wherein said at least one malaria antigen is an
10 antigen comprising $(NPNA)_n$, where n is an integer from 4 to 30, and/or an antigen comprising $(EYLNKIQNSLSTEWSPCSVT)_y$, where y is an integer from 1 to 6, and

 wherein said at least one malaria antigen is inserted into the envelope protein E2 of the virus structural
15 polypeptide at a position that corresponds to: between residues 509 and 510, 510 and 511, 511 and 512, 519 and 520, 529 and 530, 530 and 531, or 531 and 532 of SEQ ID NO. 1 or 2; or between residues 515 and 516, 516 and 517, 517 and 518, 518 and 519, 519 and 520, 536 and 537, 537 and 538 or 538
20 and 539 of SEQ ID NO. 3.

2. The virus-like particle according to claim 1, wherein said virus structural polypeptide is a polypeptide of Chikungunya virus (CHIKV).

3. The virus-like particle according to claim 1 or 2, wherein
25 said at least one malaria antigen is inserted into the

envelope protein E2 of the virus structural protein at a position that corresponds to between residues 531 and 532 of SEQ ID Nos.1 or 2, or between residues 518 and 519 of SEQ ID No.3.

5 4. The virus-like particle according to any one of claims 1 to 3, wherein said virus-like particle is expressed by transfecting a nucleic acid molecule encoding the amino acid sequence represented by SEQ ID Nos. 28, 31, 34, 37, 39, 41 or 43 into a mammalian cell.

10 5. The virus-like particle according to any one of claims 1 to 4, wherein said at least one malaria antigen is the antigen comprising (NPNA)_n wherein n is from 4 to 30.

15 6. An isolated nucleic acid molecule comprising a nucleotide sequence for expressing the virus-like particle according to any one of claims 1 to 5.

7. An isolated nucleic acid molecule consisting of a nucleotide sequence represented by SEQ ID No. 26, 27, 29, 30, 32, 33, 35, 36, 38, 40 or 42.

20 8. A vector comprising the nucleic acid molecule according to claim 6 or 7, wherein the vector optionally comprises an expression control sequence operably linked to the nucleic acid molecule.

25 9. A composition comprising the virus-like particle according to any one of claims 1 to 5 and/or the nucleic acid molecule according to claim 6 or 7, and/or the vector

of claim 8, and a pharmaceutically acceptable carrier and/or adjuvant.

10. A pharmaceutical composition comprising:

- 5 (a) the virus-like particle according to any one of claims 1 to 5 and/or the nucleic acid molecule according to claim 6 or 7, and/or the vector of claim 8; and
(b) a pharmaceutically acceptable carrier.

11. A vaccine composition comprising the virus-like particle according to any one of claims 1 to 5, and a pharmaceutically acceptable carrier and/or adjuvant.
10

12. A DNA vaccine composition comprising the nucleic acid molecule according to claim 6 or 7, and/or the vector of claim 8, and a pharmaceutically acceptable carrier and/or adjuvant.

15 13. Use, for production of an antibody in a mammal, of the virus-like particle according to any one of claims 1 to 5 and/or the nucleic acid molecule according to claim 6 or 7, and/or the vector of claim 8.

14. The use according to claim 13, wherein said antibody is a monoclonal antibody.
20

15. Use, for immunomodulation of a mammal, of an immunologically effective amount of the composition of any one of claims 9 to 12.

16. Use, to induce and/or enhance an immune response against a malaria antigen in a mammal, of an effective amount of the composition of any one of claims 9 to 12.
17. Use, to treat malaria in a mammal, of an effective amount
5 of the composition of any one of claims 9 to 12.
18. Use, for passive immunization against a malaria-causing pathogen in a mammal, of the antibody obtained by the use according to claim 13 or 14.
19. Use, to present an antigen on macrophage in a mammal, of
10 the virus-like particle according to any one of claims 1 to 5 and/or the nucleic acid molecule according to claim 6 or 7 and/or the vector of claim 8.
20. An *in vitro* or *ex vivo* method for producing the virus-like particle according to any one of claims 1 to 5,
15 comprising preparing a vector comprising a nucleic acid encoding said virus-like particle; culturing a cell which is transfected with said vector to express said virus-like particle; and recovering said virus-like particle.
21. A vaccine comprising, as separate components:
- 20 (i) a priming composition comprising the virus-like particle according to any one of claims 1 to 5, and/or the nucleic acid molecule according to claim 6 or 7, and/or the vector of claim 8, and a pharmaceutically acceptable carrier and/or adjuvant; and

(ii) a boosting composition comprising the virus-like particle according to any one of claims 1 to 5, and/or the nucleic acid molecule according to claim 6 or 7, and/or the vector of claim 8, and a pharmaceutically acceptable carrier and/or adjuvant, wherein

the at least one malaria antigen comprised in or encoded by the priming composition is the same as or different from the at least one malaria antigen comprised in or encoded by the boosting composition.

22. The vaccine of claim 21, for use in immunomodulation of a mammal.

23. The vaccine of claim 21, for introduction and/or enhancement of an immune response against a malaria antigen in a mammal.

24. The vaccine of claim 21, for use in treatment of malaria in a mammal.

25. The vaccine composition according to claim 11 for use in the prevention or treatment of malaria, wherein the virus-like particle comprises a plurality of malaria antigens.

26. A kit comprising

- (a) a vaccine composition comprising the virus-like particle according to any one of claims 1 to 5; and
- (b) another vaccine composition comprising the virus-like particle according to any one of claims 1 to 5,

wherein the virus-like particle contained in (a) is different from the virus-like particle contained in (b).

27. The kit according to claim 26, wherein the virus-like particle contained in (a) is Chikungunya virus-like particle and the virus-like particle contained in (b) is Venezuelan equine encephalitis virus-like particle, or the virus-like particle contained in (a) is Venezuelan equine encephalitis virus-like particle and the virus-like particle contained in (b) is Chikungunya virus-like particle.

28. The kit according to claim 26 or 27, further comprising (c) one or more vaccine compositions, each of which comprises the virus-like particle according to any one of claims 1 to 5, wherein (a) primes immunization and (b) and (c) boost immunization, and the virus-like particle contained in (c) is different from the virus-like particles contained in (a) and (b), or the same as the virus-like particle contained in (a) or (b).

29. The kit according to any one of claims 26 to 28, wherein the respective vaccine compositions are in a form for administration simultaneously, separately or sequentially.

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Fig. 1

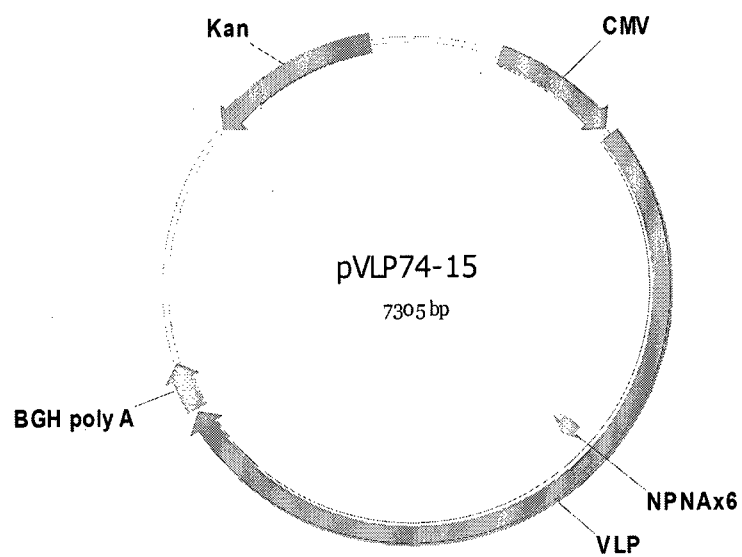
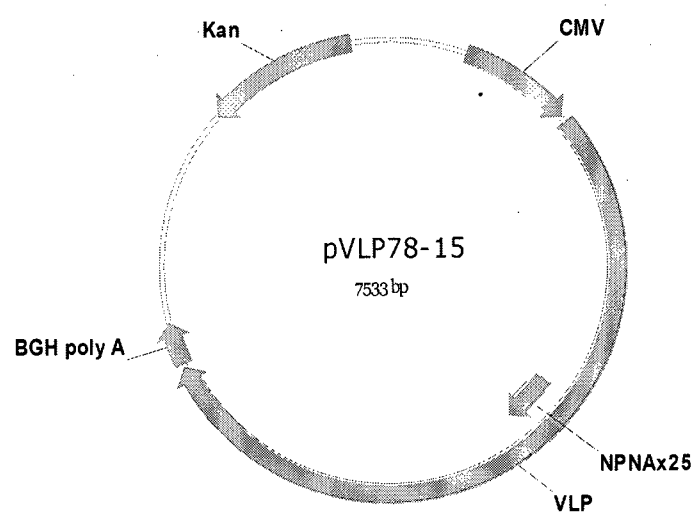


Fig. 2



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Fig. 3

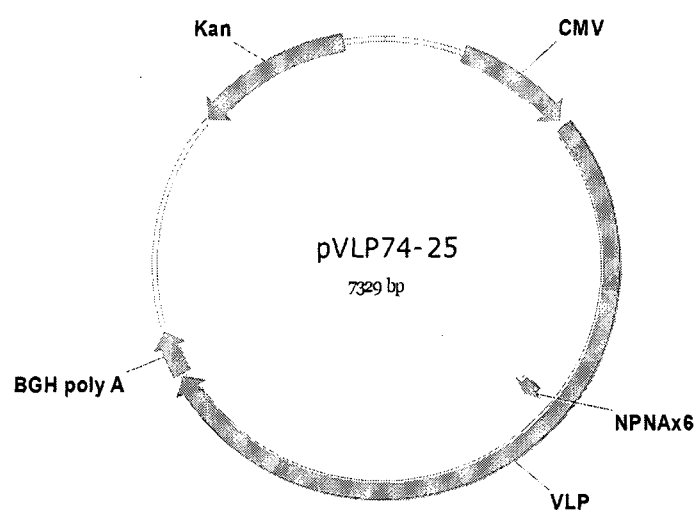
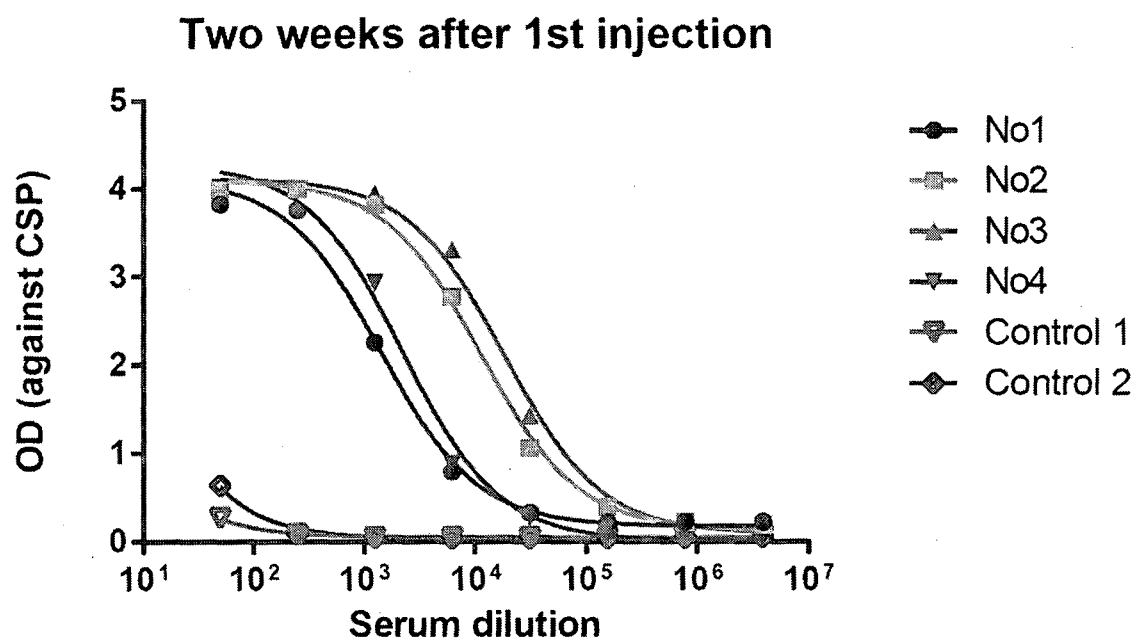


Fig. 4



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Fig. 5

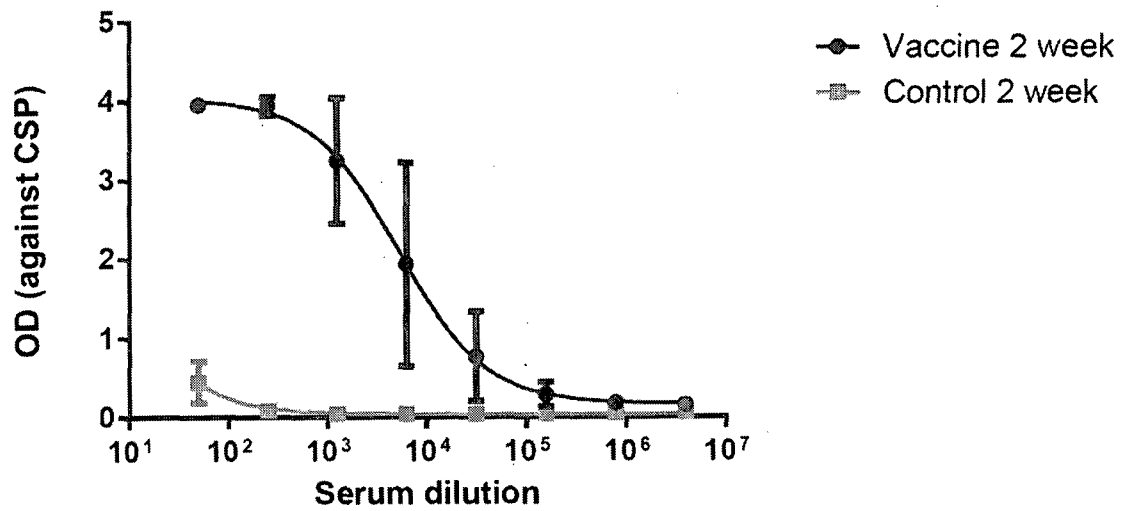
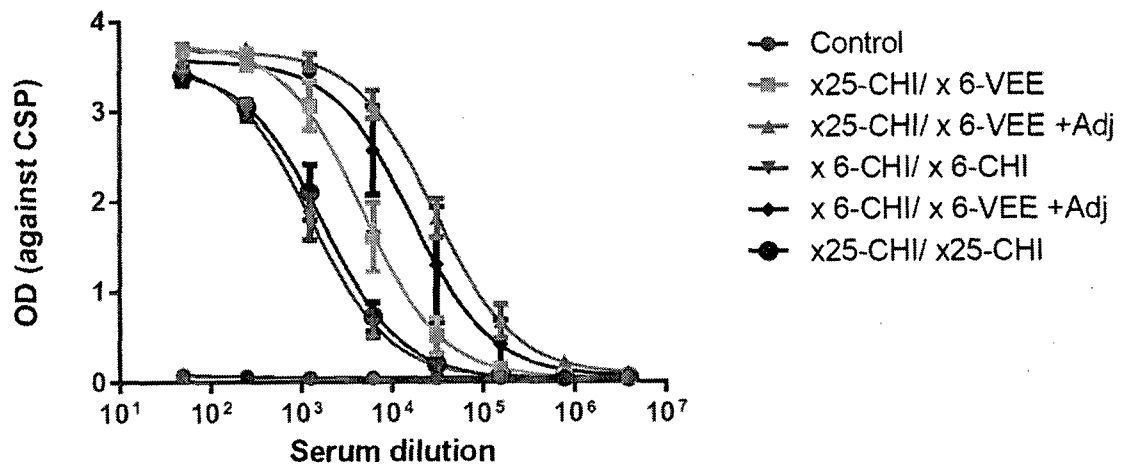


Fig. 6



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Fig. 7

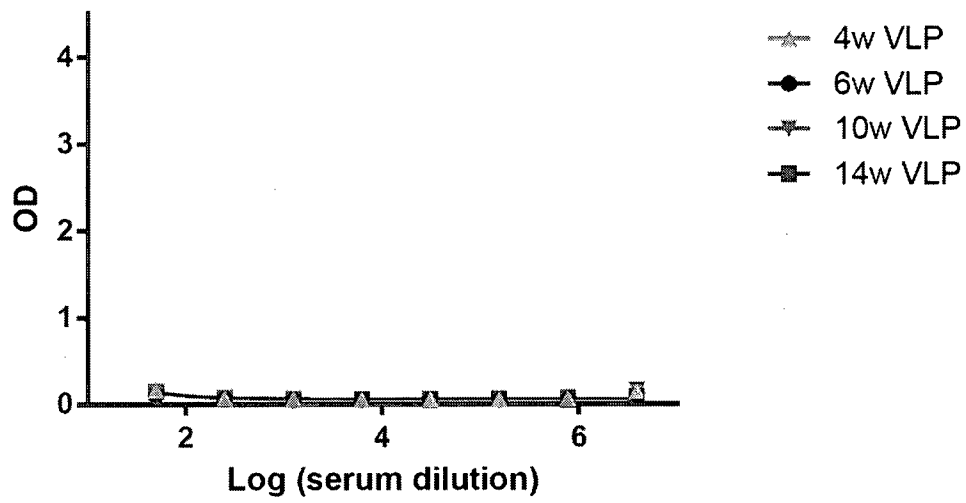


Fig. 8

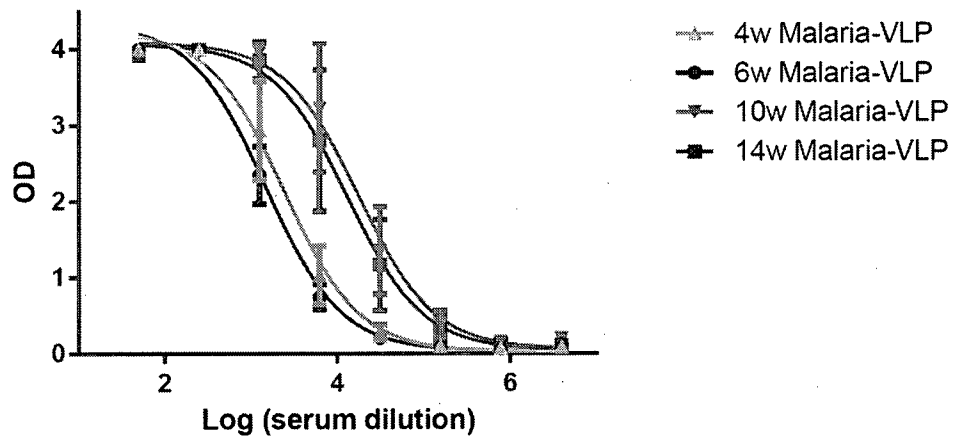


Fig. 9

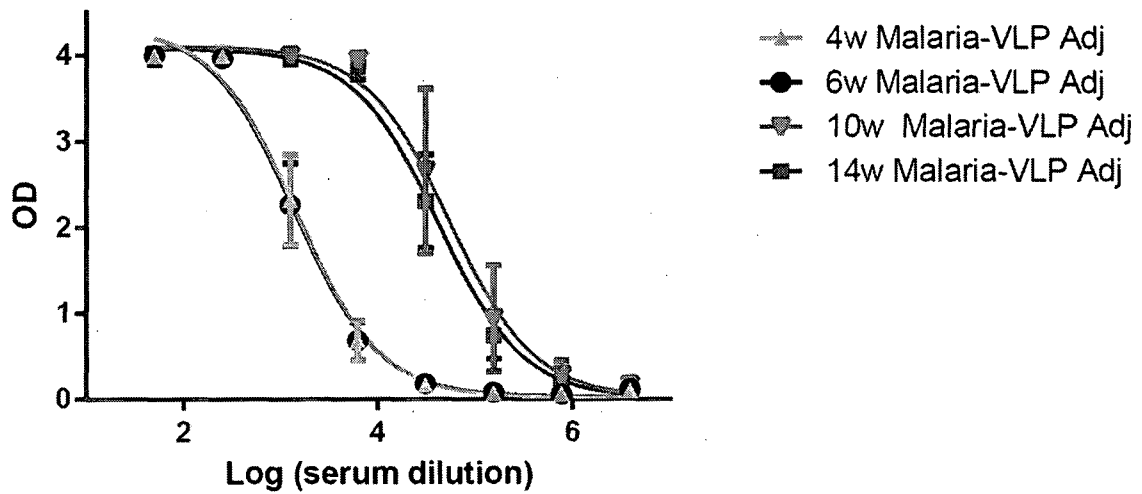


Figure 10

P. yoelii challenge after immunization

BALB/c mice
n=5 x 4 groups

- Control (PBS)
- Empty VLP
- Malaria VLP
- Malaria VLP + Ribi

The mice were challenged with 5,000 PySPZ/mouse

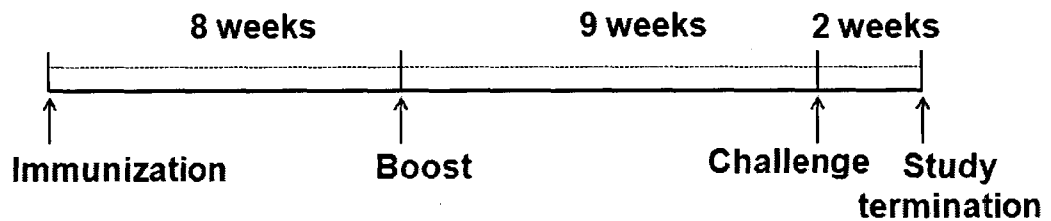


Figure 11

Results from 18S PCR (Day6)

