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(54) Benævnelse: **GENER INVOLVERET I PRODUKTIONEN AF NOSCAPIN**

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DESCRIPTION

[0001] This disclosure relates to the isolation and sequencing of a nucleic acid molecule that includes a gene cluster comprising 10 genes from a noscapine producing *Papaver somniferum* [opium poppy] cultivar; transgenic cells transformed with said nucleic acid molecule, sequence variants of the genes; the use of said genes/proteins in the production of opiate alkaloids; and the use of the genes as a marker of P. *somniferum* plants that synthesize opiate alkaloids, in particular noscapine.

Background to Disclosure

[0002] Noscapine belongs to the phthalideisoquinoline subclass of the structurally diverse isoquinoline alkaloids whereas codeine, morphine, thebaine and oripavine belong to the morphinan subclass. While the biosynthesis of morphinans has been elucidated at the molecular level our knowledge of noscapine biosynthesis has not advanced significantly since the demonstration using isotope labeling in the 1960s, that it is derived from scoulerine. Understanding the biochemical genetics underpinning noscapine biosynthesis should enable improved production of noscapine and related molecules both in poppy and other expression systems.

[0003] *P. somniferum* is the plant from which opium is extracted. The opium poppy is the only commercially exploited poppy of the family *Papaveraceae* and is the principal source of natural opiates. The opium is extracted from latex harvested from the green seed pods. A further source of opiate alkaloids is the poppy straw which is the dried mature plant. P. *somniferum* is a source of clinically useful opiate alkaloids such as morphine, codeine, thebaine, noscapine [also known as narcotine] and papaverine. The clinical application of these opiate alkaloids and their derivates is broad having use as analgesics, cough suppressants and anti-spasmodics. Although not used as a pharmacological agent in its own right, thebaine is a particularly useful opiate which can be converted into a range of compounds such as hydrocodone, oxycodone, oxymorphone, nalbuphine naltrexone, buprenorphine and etorphine. These intermediates also have broad pharmaceutical applications. For example, oxycodone, oxymorphone and etorphine are widely used as an analgesic for moderate to severe pain and are often combined with other analgesics such as ibuprofen. Buprenorphine is used in the treatment of heroin addiction and chronic pain. Naltrexone is used in the treatment of alcohol and opiate addiction.

[0004] This disclosure relates to transcriptomic analysis of P. *somniferum* noscapine producing cultivars compared to P. *somniferum* cultivars that are non-noscapine producing. The analysis has revealed the exclusive expression of a group of mostly cytochrome P450 and methyltransferase genes in a poppy variety that produces noscapine. These genes are surprisingly absent from the genomes of two non-noscapine producing varieties. Analysis of an F2 mapping population indicated the genes are tightly linked in the noscapine variety and bacterial artificial chromosome sequencing confirmed they exist as a novel gene cluster for the biosynthesis of opiate alkaloids.

Statements of Invention

[0005] According to an aspect of the invention there is provided an isolated nucleic acid molecule comprising or consisting of a nucleotide sequence selected from the group consisting of:

1. i) a nucleotide sequence as represented by the sequence in SEQ ID NO: 8;
2. ii) a nucleotide sequence wherein said sequence is degenerate as a result of the genetic code to the nucleotide sequence defined in (i);
3. iii) a nucleic acid molecule that is at least 80% identical to SEQ ID NO: 8 which encodes a polypeptide that has carboxyltransferase activity;;
4. iv) a nucleotide sequence that encodes a polypeptide comprising an amino acid sequence as represented in SEQ ID NO: 18 or a nucleotide sequence that encodes a polypeptide a polypeptide that has at least 75% amino acid sequence identity across the full length amino acid sequence set forth in SEQ ID NO: 18 wherein said polypeptide has carboxylesterase activity.

[0006] In a preferred aspect or embodiment of the invention said nucleic acid molecule comprises or consists of a nucleotide sequence as represented SEQ ID NO: 8 wherein said nucleic acid molecule encodes a polypeptide with carboxylesterase activity.

[0007] In a preferred embodiment of the invention said nucleic acid molecule further includes one or more nucleotide sequences selected from the group consisting of: SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 9 or 10.

[0008] In a preferred embodiment of the invention said nucleic acid molecule includes 3, 4, 5, 6, 7, 8 or 9 nucleotide sequences selected from the group consisting of: SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9 or 10.

[0009] In a preferred embodiment of the invention said nucleic acid molecule includes each of the nucleotide sequences as represented in SEQ ID NO: 1, 2, 3, 4, 5, 6, 7, 8, 9 and 10.

[0010] According to a further aspect of the invention there is provided an isolated polypeptide selected from the group consisting of:

1. i) a polypeptide comprising or consisting of an amino acid sequence as represented in SEQ ID NO:18; or
2. ii) a modified polypeptide comprising or consisting of a modified amino acid sequence wherein said polypeptide is modified by addition deletion or substitution of at least one amino acid residue of the sequence presented in SEQ ID NO: 18 and which has at least 75% sequence identity to the full length amino acid sequence in SEQ ID NO: 18 and has carboxylesterase activity.

[0011] A modified polypeptide as herein disclosed may differ in amino acid sequence by one or more substitutions, additions, deletions, truncations that may be present in any combination. Among preferred variants are those that vary from a reference polypeptide by conservative amino acid substitutions. Such substitutions are those that substitute a given amino acid by another amino acid of like characteristics. The following non-limiting list of amino acids are considered conservative replacements (similar): a) alanine, serine, and threonine; b) glutamic acid and aspartic acid; c) asparagine and glutamine d) arginine and lysine; e) isoleucine, leucine, methionine and valine and f) phenylalanine, tyrosine and tryptophan. Most highly preferred are variants that retain or enhance the same biological function and activity as the reference polypeptide from which it varies.

[0012] In one embodiment, the modified polypeptides have at least at least75%, 80%, 85%, 90%, 95% identity, and at least 99% identity with most or the full length amino acid sequence illustrated herein.

[0013] According to a further aspect of the invention there is provided a vector comprising a nucleic acid molecule according to the invention.

[0014] Preferably the nucleic acid molecule in the vector is under the control of, and operably linked to, an appropriate promoter or other regulatory elements for transcription in a host cell such as a microbial, (e.g. bacterial, yeast), or plant cell. The vector may be a bifunctional expression vector which functions in multiple hosts. In the case of genomic DNA this may contain its own promoter or other regulatory elements and in the case of cDNA this may be under the control of an appropriate promoter or other regulatory elements for expression in the host cell.

[0015] By "promoter" is meant a nucleotide sequence upstream from the transcriptional initiation site and which contains all the regulatory regions required for transcription. Suitable promoters include constitutive, tissue-specific, inducible, developmental or other promoters for expression in plant cells comprised in plants depending on design. Such promoters include viral, fungal, bacterial, animal and plant-derived promoters capable of functioning in plant cells.

[0016] Constitutive promoters include, for example CaMV 35S promoter (Odell et al. (1985) Nature 313, 9810-812); rice actin (McElroy et al. (1990) Plant Cell 2: 163-171); ubiquitin (Christian et al. (1989) Plant Mol. Biol. 18 (675-689); pEMU (Last et al. (1991) Theor Appl. Genet. 81: 581-588); MAS (Velten et al. (1984) EMBO J. 3. 2723-2730); ALS promoter (U.S. Application Seriel No. 08/409,297), and the like. Other constitutive promoters include those in U.S. Patent Nos. 5,608,149; 5,608,144; 5,604,121; 5,569,597; 5,466,785; 5,399,680, 5,268,463; and 5,608,142.

[0017] Chemical-regulated promoters can be used to modulate the expression of a gene in a plant through the application of an exogenous chemical regulator. Depending upon the objective, the promoter may be a chemical-inducible promoter, where application of the chemical induced gene expression, or a chemical-repressible promoter, where application of the chemical represses gene expression. Chemical-inducible promoters are known in the art and include, but are not limited to, the maize In2-2 promoter, which is activated by benzenesulfonamide herbicide

safeners, the maize GST promoter, which is activated by hydrophobic electrophilic compounds that are used as pre-emergent herbicides, and the tobacco PR-1a promoter, which is activated by salicylic acid. Other chemical-regulated promoters of interest include steroid-responsive promoters (see, for example, the glucocorticoid-inducible promoter in Schena et al. (1991) Proc. Natl. Acad. Sci. USA 88: 10421-10425 and McNellis et al. (1998) Plant J. 14(2): 247-257) and tetracycline-inducible and tetracycline-repressible promoters (see, for example, Gatz et al. (1991) Mol. Gen. Genet. 227: 229-237, and US Patent Nos. 5,814,618 and 5,789,156.

[0018] Where enhanced expression in particular tissues is desired, tissue-specific promoters can be utilised. Tissue-specific promoters include those described by Yamamoto et al. (1997) Plant J. 12(2): 255-265; Kawamata et al. (1997) Plant Cell Physiol. 38(7): 792-803; Hansen et al. (1997) Mol. Gen. Genet. 254(3): 337-343; Russell et al. (1997) Transgenic Res. 6(2): 157-168; Rinehart et al. (1996) Plant Physiol. 112(3): 1331-1341; Van Camp et al. (1996) Plant Physiol. 112(2): 525-535; Canevascni et al. (1996) Plant Physiol. 112(2): 513-524; Yamamoto et al. (1994) Plant Cell Physiol. 35(5): 773-778; Lam (1994) Results Probl. Cell Differ. 20: 181-196; Orozco et al. (1993) Plant Mol. Biol. 23(6): 1129-1138; Mutsuoka et al. (1993) Proc. Natl. Acad. Sci. USA 90 (20): 9586-9590; and Guevara-Garcia et al (1993) Plant J. 4(3): 495-50.

[0019] "Operably linked" means joined as part of the same nucleic acid molecule, suitably positioned and oriented for transcription to be initiated from the promoter. DNA operably linked to a promoter is "under transcriptional initiation regulation" of the promoter. In a preferred aspect, the promoter is a tissue specific promoter, an inducible promoter or a developmentally regulated promoter.

[0020] Particular of interest in the present context are nucleic acid constructs which operate as plant vectors. Specific procedures and vectors previously used with wide success in plants are described by Guerineau and Mullineaux (1993) (Plant transformation and expression vectors. In: Plant Molecular Biology Labfax (Croy RRD ed) Oxford, BIOS Scientific Publishers, pp 121-148. Suitable vectors may include plant viral-derived vectors (see e.g. EP194809). If desired, selectable genetic markers may be included in the construct, such as those that confer selectable phenotypes such as resistance to herbicides (e.g. kanamycin, hygromycin, phosphinotricin, chlorsulfuron, methotrexate, gentamycin, spectinomycin, imidazolinones and glyphosate).

[0021] In a preferred embodiment of the invention said vector is a bacterial artificial chromosome [BACS].

[0022] According to a further aspect of the invention there is provided a transgenic cell transformed or transfected with a nucleic acid molecule or vector according to the invention.

[0023] In a preferred embodiment of the invention said cell is a plant cell.

[0024] In a preferred embodiment of the invention said plant cell is from the genus *Papaver.*

[0025] In a preferred embodiment of the invention said plant cell is a *Papaver somniferum* cell.

[0026] According to a further aspect of the invention there is provided a plant comprising a plant cell according to the invention.

[0027] In a preferred embodiment of the invention said plant is from the genus *Papaver;* preferably *Papaver somniferum.*

[0028] In an alternative preferred embodiment of the invention said cell is a microbial cell; preferably a bacterial or fungal cell [e.g. yeast, *Saccharomyces cerevisae*].

[0029] In a preferred embodiment of the invention said cell is adapted such that the nucleic acid molecule encoding one or more polypeptides according to the invention is over-expressed when compared to a non-transgenic cell of the same species.

[0030] According to a further aspect of the invention there is provided a nucleic acid molecule comprising a transcription cassette wherein said cassette includes the nucleotide sequence of SEQ ID NO: 8, and is adapted for expression by provision of at least one promoter operably linked to said nucleotide sequence such that both sense and antisense molecules are transcribed from said cassette.

[0031] In a preferred embodiment of the invention said cassette is adapted such that both sense and antisense ribonucleic acid molecules are transcribed from said cassette wherein said sense and antisense nucleic acid molecules are adapted to anneal over at least part or all of their length to form a inhibitory RNA or short hairpin RNA.

[0032] In a preferred embodiment of the invention said cassette is provided with at least two promoters adapted to transcribe both sense and antisense strands of said ribonucleic acid molecule.

[0033] In an alternative preferred embodiment of the invention said cassette comprises a nucleic acid molecule wherein said molecule comprises a first part linked to a second part wherein said first and second parts are complementary over at least part of their sequence and further wherein transcription of said nucleic acid molecule produces an ribonucleic acid molecule which forms a double stranded region by complementary base pairing of said first and second parts thereby forming an short hairpin RNA.

[0034] A technique to specifically ablate gene function is through the introduction of double stranded RNA, also referred to as small inhibitory/interfering RNA (siRNA) or short hairpin RNA [shRNA], into a cell which results in the destruction of mRNA complementary to the sequence included in the siRNA/shRNA molecule. The siRNA molecule comprises two complementary strands of RNA (a sense strand and an antisense strand) annealed to each other to form a double stranded RNA molecule. The siRNA molecule is typically derived from exons of the gene which is to be ablated. The mechanism of RNA interference is being elucidated. Many organisms respond to the presence of double stranded RNA by activating a cascade that leads to the formation of siRNA. The presence of double stranded RNA activates a protein complex comprising RNase III which processes the double stranded RNA into smaller fragments (siRNAs,

approximately 21-29 nucleotides in length) which become part of a ribonucleoprotein complex. The siRNA acts as a guide for the RNase complex to cleave mRNA complementary to the antisense strand of the siRNA thereby resulting in destruction of the mRNA.

[0035] In a preferred embodiment of the invention said nucleic acid molecule is part of a vector adapted for expression in a plant cell.

[0036] According to a further aspect of the invention there is provided a plant cell transfected with a nucleic acid molecule or vector according to the invention wherein said cell has reduced expression of a polypeptide according to the invention.

[0037] According to an aspect of the invention there is provided a process for the modification of papaveroxine and desmethylpapaveroxine comprising:

i) providing a transgenic Papaver plant cell according to the invention;

ii) cultivating said Papaver plant cell to produce a transgenic Papaver plant; and optionally

i) harvesting said transgenic Papaver plant, or part thereof.

[0038] In a preferred method of the invention said harvested plant material is dried and opiate alkaloid is extracted.

[0039] According to an alternative aspect of the invention there is provided a process for the modification of papaveroxine or desmethylpapaveroxine comprising :

1. i) providing a transgenic microbial cell according to the invention that expresses one or more nucleic acid molecules according to the invention in culture with at least papaveroxine or desmethylpapaveroxine ;
2. ii) cultivating the microbial cell under conditions that modify papaveroxine or desmethylpapaveroxine; and optionally
3. iii) isolating narcotinehemiacetal or narceotolinehemiacetal from the microbial cell or cell culture.

[0040] In a preferred method of the invention said microbial cell is a bacterial cell or fungal/yeast cell.

[0041] If microbial cells are used as organisms in the process according to the invention they are grown or cultured in the manner with which the skilled worker is familiar, depending on the host organism. As a rule, microorganisms are grown in a liquid medium comprising a carbon source, usually in the form of sugars, a nitrogen source, usually in the form of organic nitrogen sources such as yeast extract or salts such as ammonium sulfate, trace elements such as salts of iron, manganese and magnesium and, if appropriate, vitamins, at temperatures of between 0°C and

100°C, preferably between 10°C and 60°C, while gassing in oxygen.

[0042] The pH of the liquid medium can either be kept constant, that is to say regulated during the culturing period, or not. The cultures can be grown batchwise, semi-batchwise or continuously. Nutrients can be provided at the beginning of the fermentation or fed in semi-continuously or continuously. The methylated opiate alkaloids produced can be isolated from the organisms as described above by processes known to the skilled worker, for example by extraction, distillation, crystallization, if appropriate precipitation with salt, and/or chromatography. To this end, the organisms can advantageously be disrupted beforehand. In this process, the pH value is advantageously kept between pH 4 and 12, preferably between pH 6 and 9, especially preferably between pH 7 and 8.

[0043] The culture medium to be used must suitably meet the requirements of the strains in question. Descriptions of culture media for various microorganisms can be found in the textbook "Manual of Methods for General Bacteriology" of the American Society for Bacteriology (Washington D.C., USA, 1981).

[0044] As described above, these media which can be employed in accordance with the invention usually comprise one or more carbon sources, nitrogen sources, inorganic salts, vitamins and/or trace elements.

[0045] Preferred carbon sources are sugars, such as mono-, di- or polysaccharides. Examples of carbon sources are glucose, fructose, mannose, galactose, ribose, sorbose, ribulose, lactose, maltose, sucrose, raffinose, starch or cellulose. Sugars can also be added to the media via complex compounds such as molasses or other by-products from sugar refining. The addition of mixtures of a variety of carbon sources may also be advantageous. Other possible carbon sources are oils and fats such as, for example, soya oil, sunflower oil, peanut oil and/or coconut fat, fatty acids such as, for example, palmitic acid, stearic acid and/or linoleic acid, alcohols and/or polyalcohols such as, for example, glycerol, methanol and/or ethanol, and/or organic acids such as, for example, acetic acid and/or lactic acid.

[0046] Nitrogen sources are usually organic or inorganic nitrogen compounds or materials comprising these compounds. Examples of nitrogen sources comprise ammonia in liquid or gaseous form or ammonium salts such as ammonium sulfate, ammonium chloride, ammonium phosphate, ammonium carbonate or ammonium nitrate, nitrates, urea, amino acids or complex nitrogen sources such as cornsteep liquor, soya meal, soya protein, yeast extract, meat extract and others. The nitrogen sources can be used individually or as a mixture.

[0047] Inorganic salt compounds which may be present in the media comprise the chloride, phosphorus and sulfate salts of calcium, magnesium, sodium, cobalt, molybdenum, potassium, manganese, zinc, copper and iron.

[0048] Inorganic sulfur-containing compounds such as, for example, sulfates, sulfites, dithionites, tetrathionates, thiosulfates, sulfides, or else organic sulfur compounds such as mercaptans and thiols may be used as sources of sulfur for the production of sulfur-containing fine chemicals, in

particular of methionine.

[0049] Phosphoric acid, potassium dihydrogenphosphate or dipotassium hydrogenphosphate or the corresponding sodium-containing salts may be used as sources of phosphorus.

[0050] Chelating agents may be added to the medium in order to keep the metal ions in solution. Particularly suitable chelating agents comprise dihydroxyphenols such as catechol or protocatechuate and organic acids such as citric acid.

[0051] The fermentation media used according to the invention for culturing microorganisms usually also comprise other growth factors such as vitamins or growth promoters, which include, for example, biotin, riboflavin, thiamine, folic acid, nicotinic acid, panthothenate and pyridoxine. Growth factors and salts are frequently derived from complex media components such as yeast extract, molasses, cornsteep liquor and the like. It is moreover possible to add suitable precursors to the culture medium. The exact composition of the media compounds heavily depends on the particular experiment and is decided upon individually for each specific case. Information on the optimization of media can be found in the textbook "Applied Microbiol. Physiology, A Practical Approach" (Editors P.M. Rhodes, P.F. Stanbury, IRL Press (1997) pp. 53-73, ISBN 0 19 963577 3). Growth media can also be obtained from commercial suppliers, for example Standard 1 (Merck) or BHI (brain heart infusion, DIFCO) and the like.

[0052] All media components are sterilized, either by heat (20 min at 1.5 bar and 121°C) or by filter sterilization. The components may be sterilized either together or, if required, separately. All media components may be present at the start of the cultivation or added continuously or batchwise, as desired.

[0053] The culture temperature is normally between 15°C and 45°C, preferably at from 25°C to 40°C, and may be kept constant or may be altered during the experiment. The pH of the medium should be in the range from 5 to 8.5, preferably around 7.0. The pH for cultivation can be controlled during cultivation by adding basic compounds such as sodium hydroxide, potassium hydroxide, ammonia and aqueous ammonia or acidic compounds such as phosphoric acid or sulfuric acid. Foaming can be controlled by employing antifoams such as, for example, fatty acid polyglycol esters. To maintain the stability of plasmids it is possible to add to the medium suitable substances having a selective effect, for example antibiotics. Aerobic conditions are maintained by introducing oxygen or oxygen-containing gas mixtures such as, for example, ambient air into the culture. The temperature of the culture is normally 20°C to 45°C and preferably 25°C to 40°C. The culture is continued until formation of the desired product is at a maximum. This aim is normally achieved within 10 to 160 hours.

[0054] The fermentation broth can then be processed further. The biomass may, according to requirement, be removed completely or partially from the fermentation broth by separation methods such as, for example, centrifugation, filtration, decanting or a combination of these methods or be left completely in said broth. It is advantageous to process the biomass after its separation.

[0055] However, the fermentation broth can also be thickened or concentrated without separating the cells, using known methods such as, for example, with the aid of a rotary evaporator, thin-film evaporator, falling-film evaporator, by reverse osmosis or by nanofiltration. Finally, this concentrated fermentation broth can be processed to obtain the opiate alkaloids present therein.

[0056] According to a further aspect of the invention there is provided the use of a gene encoded by a nucleic acid molecule as represented by the nucleic acid sequence in SEQ ID NO: 8, or a nucleic acid molecule that is at least 80% identical to SEQ ID NO: 8 and encodes a polypeptide with carboxylesterase activity as a means to identify a locus wherein said locus is associated with altered expression or activity of said carboxylesterase activity.

[0057] Mutagenesis as a means to induce phenotypic changes in organisms is well known in the art and includes but is not limited to the use of mutagenic agents such as chemical mutagens [e.g. base analogues, deaminating agents, DNA intercalating agents, alkylating agents, transposons, bromine, sodium azide] and physical mutagens [e.g. ionizing radiation, psoralen exposure combined with UV irradiation].

[0058] According to a further aspect of the invention there is provided a *P. somniferum* plant that has altered expression of a carboxylesterase polypeptide obtainable by a process comprising the steps of:

1. i) mutagenesis of wild-type seed from a *P. somniferum* plant that does express said polypeptide;
2. ii) cultivation of the seed in i) to produce first and subsequent generations of plants;
3. iii) obtaining seed from the first generation plant and subsequent generations of plants;
4. iv) determining if the seed from said first and subsequent generations of plants has altered nucleotide sequence and/or altered expression of said carboxylesterase polypeptide;
5. v) obtaining the seed or plants with an altered nucleotide sequence encoding an altered carboxylesterase polypeptide wherein the enzyme activity in the modification of papaveroxine or desmethylpapaveroxine is altered when compared to a wild-type *P. somniferum.*

[0059] According to a further aspect of the invention there is provided a method for the analysis of a plant

1. i) obtaining a sample a mutated plant and analysing the nucleic acid sequence of a nucleic acid molecule selected from the group consisting of:
    1. a) a nucleic acid molecule comprising a nucleotide sequence as represented in 8;
    2. b) a nucleic acid molecule that has at least 80% sequence identity to SEQ ID NO: 8 encodes a polypeptide with carboxylesterase activity;
2. ii) comparing the nucleotide sequence of the nucleic acid molecule in said sample to a nucleotide sequence of a nucleic acid molecule of the original wild-type plant; and optionally wherein the nucleic acid molecule is analysed by a method comprising the steps of:
3. iii) extracting nucleic acid from said mutated plants;

4. iv) amplification of a part of said nucleic acid molecule by a polymerase chain reaction;
5. v) forming a preparation comprising the amplified nucleic acid and nucleic acid extracted from wild-type seed to form heteroduplex nucleic acid;
6. vi) incubating said preparation with a single stranded nuclease that cuts at a region of heteroduplex nucleic acid to identify the mismatch in said heteroduplex; and
7. vii) determining the site of the mismatch in said nucleic acid heteroduplex; and
8. viii) obtaining a plant with a mutation in the nucleic acid encoding said carboxylesterase polypeptide.

[0060] According to a further aspect of the invention there is provided a viral vector comprising a nucleic acid molecule according to the invention.

[0061] Virus induced gene silencing [VIGS] is known in the art and exploits a RNA mediated antiviral defence mechanism. Plants that are infected with an unmodified virus induces a mechanism that specifically targets the viral genome. However, viral vectors which are engineered to include nucleic acid molecules derived from host plant genes also induce specific inhibition of viral vector expression and additionally target host mRNA. This allows gene specific gene silencing without genetic modification of the plant genome and is essentially a non-transgenic modification.

[0062] Throughout the description and claims of this specification, the words "comprise" and "contain" and variations of the words, for example "comprising" and "comprises", means "including but not limited to", and is not intended to (and does not) exclude other moieties, additives, components, integers or steps. "Consisting essentially" means having the essential integers but including integers which do not materially affect the function of the essential integers.

[0063] Throughout the description and claims of this specification, the singular encompasses the plural unless the context otherwise requires. In particular, where the indefinite article is used, the specification is to be understood as contemplating plurality as well as singularity, unless the context requires otherwise.

[0064] Features, integers, characteristics, compounds, chemical moieties or groups described in conjunction with a particular aspect, embodiment or example of the invention are to be understood to be applicable to any other aspect, embodiment or example described herein unless incompatible therewith.

[0065] An embodiment of the invention will now be described by example only and with reference to the following figures:
Figure 1: Identification of genes exclusively present in the genome of a noscapine producing poppy variety, HN1 (High Noscapine 1). (A) Relative abundance of the major alkaloids extracted from the capsules of three commercial varieties of poppy, HM1 (High Morphine 1), HT1 (High Thebaine 1) and HN1. M = morphine, C = codeine, T = thebaine, O = oripavine and N = Noscapine. (B) EST libraries from stem and capsule were generated by pyrosequencing and unique contiguous sequences assembled as described in material and methods . Ten genes

(*PSMT1*, *PSMT2*, *PSMT3*, *CYP82X1*, *CYP82X2, CYP82Y1, CYP719A21, PSAT1, PSSDR1* and *PSCXE1*) as defined in the text, were represented only in EST libraries from the HN1 variety. EST abundance of five other functionally characterized *P. somniferum* genes (BBE, TNMT, SalR, SalAT and T6DM) show them to be expressed in all three varieties and at consistently higher levels in stem compared to capsule as is also the case for the HN1 specific genes as shown in colour code (Fig 1). PCR on genomic DNA from all three varieties revealed that the ten HN1 specific genes are absent from the genomes of the HM1 and HT1 varieties (Fig 5);

Figure 2: Segregation analysis of noscapine content in an F2 mapping population demonstrates requirement for the noscapine gene cluster. **(A)** Box plot depiction of noscapine levels as percentage dry weight (DW) in glasshouse grown parental lines HN1 and HM1 and the F1 generation. **(B)** The field grown F2 generation segregated into three classes of zero, low and high noscapine. F2 GC- and F2 GC+ indicate the absence and presence respectively of the noscapine gene cluster. Numbers in brackets indicate number of individuals in each class;

Figure 3: The HN1 gene cluster. The structure and position of the ten HN1 specific genes expressed in stems and capsule tissues is shown above the central black line which represents 401 Kb of genomic sequence. Exons are represented by filled grey boxes and introns by fine black lines. Arrows indicate the 5' to 3' orientation of each gene. Additional open reading frames depicted below the central black line are as defined by the key. None of these ORFs are represented in the stem and capsule EST libraries;

Figure 4: Functional characterisation using virus induced gene silencing of 6 genes from the HN1 gene cluster. Results from both leaf latex and capsules are consistent with each of these genes encoding enzymes involved in noscapine biosynthesis (A-F). All compounds that accumulate, apart from scoulerine, have been putatively identified on the basis of mass spectra as detailed in Fig. 6. The mass-to-charge (m/z) value (M) followed by retention time (T) in seconds is shown for each compound on the horizontal axis. (G) Proposed pathway for noscapine biosynthesis based on VIGS data. Solid arrows depict steps supported by VIGS data, dotted arrows depict additional proposed steps. For the secoberbine intermediates, R1 = H or OH, R2 = H or OH and R3 = CH2OH or CHO or COOH (Fig. 6). The noscapine structure is numbered according to the IUPAC convention;

Figure 5: The ten genes exclusively expressed in the HN1 variety occur in the genome of HN1 but are absent from that of varieties HT1 and HM1. (A) Amplification of fragments from the ten genes exclusively expressed in HN1 using two different primer pairs. **(B)** Amplification of fragments of genes from the protoberberine and morphinan branch pathways that are expressed in all three varieties. Primers used are detailed in Table 3; HyperLadder I (Bioline Reagents, London, UK) was used as molecular size standard;

Figure 6. Evidence for putative identities of intermediates from VIGS experiments. All panels show the mass spectra of the pseudomolecular parent ion at the chromatographic peak apex in black and corresponding MS2 fragmentation spectra in red, scaled to relative abundance. MS2 spectra were generated by targeting the parent ion with a isolation width of 3 m/z and using collisional isolation dissociation energy set to 35%. All mass spectra were obtained at a resolution setting of 7500. Text printed above selected diagnostic ions indicate the exact monoisotopic mass of the ion, the calculated formula within limits C=1:100, O=0:200, N=0:3 and H=1:200, and the number/total number of formulae returned within a 5ppm error window. Fragments were reconciled against theoretical fragments generated by submitting candidate parent structures to Mass Frontier software (version 5.01.2; HighChem, Bratislava, Slovakia). Candidate parent

structures were derived from PubChem searches and the comprehensive review of Papaver spp. alkaloids (Sariyar (2002) Pure Appl. Chem. 74, 557-574). (A) Tetrahydrocolumbamine; this compound was characterized from a peak eluting at 174s from VIGS-silenced CYP719A21. Eight out of ten observed MS2 fragments were calculated as feasible by Mass Frontier; only the two most abundant diagnostic fragments are shown. (B) Secoberbine intermediate 1 (C21H25NO6); this compound was characterized from a peak eluting at 147s from VIGS-silenced CYP82X2. If R1=OH, R2=H, and R3=CH2OH, then this compound is narcotolinol which is consistent with both annotated fragments. Another candidate formula fit would be demethoxylated narcotindiol (R1=H, R2=OH, R3=CH2OH); however this structure would not form the observed fragment at 206.0816. (C) Secoberbine intermediate 2 (C21H23NO6); this compound was characterized from a peak eluting at 103s from VIGS-silenced CYP82X2. If R1=OH , R2=H, and R3=CHO, then this compound would be a desmethylated derivative of macrantaldehyde. (D) Papaveroxine; this compound was characterized from a peak eluting at 214s from VIGS-silenced PSCXE1. The 398.1600 fragment observed is consistent with deacetylation. (E) Narcotinehemiacetal; this compound was characterized from a peak eluting at 121s from VIGS-silenced PSSDR1. (F) Narcotoline (4'-desmethylnoscapine); this compound was characterized from a peak eluting at 208s from VIGS-silenced PSMT2. Other isobaric possibilities were 6- or 7-desmethylnoscapine. However, the 206.0816 fragment observed is consistent with a hydroxylated 4' position. Alternative structures could be discounted by comparing the candidate fragmentation spectra with that from synthetic 7-desmethylnoscapine, which eluted at a different retention time and lacked the characteristic 206.0816 fragment;

Table 1 Illustrates the % identity of CYP82Y1, PSCXE1, PSDFR1 and PSAT1 (SEQ ID 17-20) with their respective closest functionally characterised homologues. Accession numbers given are from GenBank, Swiss-Prot or PDB databases;

Table 2. Genotyping of F3 families derived from two F2 phenotypic classes: low noscapine and high noscapine. The observed versus expected segregation ratios strongly support the hypothesis that individuals in the low noscapine F2 class are heterozygous for the HN1 gene cluster and individuals in the high noscapine class are homozygous;

Table 3. Primer sequences and associated information.

Table 1

Protein	**% Identity**	**Accession number**	**Annotation**
CYP82Y1	54		CYP82X1 from *Papaver somniferum*
	48		CYP82X2 from *Papaver somniferum*
	39	ABM46919.1	CYP82E3, nicotine demethylase from *Nicotiana tomentosiformis*
PSCXE1	45	2O7R_A	AeCXE1, Carboxyl esterase from *Actinidia eriantha*
PSSDR1	46	AAB41550.1	Vestitone reductase from *Medicago sativa*
	45	ABQ97018.1	Dihydroflavonol 4-reductase from *Saussurea medusa*
PSAT1	66	Q94FT4.1	Salutaridinol 7-*O*-acetyltransferase from *Papaver somniferum*

Table 2

Noscapine class and genotyping result of F2 individual	F3 seed family (obtained through self-pollination of F2 individual)	Number of F3 individuals genotyped	Observed segregation of gene cluster in F3 progeny		Expected segregation in F3 if F2 low noscapine class is heterozygous and the high noscapine class is homozygous		Chi-Square	
			GC+	GC-	GC+	GC-	squared	p-value
low noscapine/GC+	S-111809	28	18	10	21	7	1.714	0.190
low noscapine/GC+	S-111835	26	18	8	19.5	6.5	0.462	0.497
high noscapine/GC-	S-111714	28	28		28			
high noscapine/GC-	S-111854	54	54		54			

Primer sequences (5'- to 3'-)			
	Reverse	Notes	Application
	AACACAAAATAGGATTACTTACTTTTGTCC	primer pair 1	Primersfor the amplification of fragments from genomic DNA of HM1, HT1 and HN1 as shown in Figure 5
	GCATGAAATGGATGTAGTTATCTTGG	primer pair 2	
	ATTCCCGTTCAAGTAAACATGCGG	primer pair 1	
	CAGTAAATTCACACATTCCGTATCTTCCC	primer pair 2	
	GAGGGTAAGCCTCAATAACAGACTGG	primer pair 1	
	TCGTTCCATTCGTGAAGAATGC	primer pair 2	
	TGCAATTGAATTTAGCTCATCTCC	primer pair 1	
	ATTCATGATTGTGACCTTTGTAATCC	primer pair 2	
	ACGATTCTGTCATCATCATTTTTCGC	primer pair 1	
	CCCAAGATTTTCATATCCTTTACAA	primer pair 2	
ATGG	GCTCCGTAAGTGCTCCTGTG	primer pair 1	
	CCCTTCACATCTACCATCCCTT	primer pair 2	
C	CGAGTGCCCATGCAGTGG	primer pair 1	
IC	CACTCCATCAGACACACAAGACC	primer pair 2	
	AAATGGCAGTTCCACCGC	primer pair 1	
	CACTGCTGACTTCCATATCAAAGC	primer pair 2	
	AGCTGAATTTGTCGATCAATAAGTGG	primer pair 1	
	ACTGGCATGATATGCAACATTAGC	primer pair 2	
	GATACACTGGGAGGAGGATGGG	primer pair 1	
	CGGCAAAATTCATTCCTTGAGC	primer pair 2	
	CATTCCTCGTCTAATTCATCTGC	sequencing primer	Primers used as sequencing primers to obtain genomic DNA sequence from HN1
	GCTTCACTACTTCTTCTTGAAAAG	sequencing primer	
	CATTATAATTTCCAATGCCGTAGTTC	sequencing primer	
	CATTCGTTGTTGTTGCTGGTAAG	sequencing primer	
	CATCCTCATTGCTTGTGTCC	sequencing primer	
TCC			

TGCCTCATGTTATTTCTGTTGCC		sequencing primer
GCATGAAATGGATGTAGTTATCTTGG		sequencing primer
AAATCGTTCGCTCTTTACCGC		sequencing primer
CACACCAAACTTGATCATTGTC		sequencing primer
ATTGTTGATATTGAATCAGAAACTTTC		sequencing primer
TCAATACCAGTACTGTTAGTTTCCG		sequencing primer
GCAACTGTTTCATTAACAGGCACATCC		sequencing primer
ATTGATGTCGGTGGTGGTCACG		sequencing primer
GCACACTGTCTTTTTCTTCCACC		sequencing primer
ACCGGAATGAGAATGCATAAAGTAAAGG		sequencing primer
CCAATACCCAATCAATTAAACTC		sequencing primer
CAGTAAATTCACACATTCCGTATCTTCCC		sequencing primer
ATTGTATAGCCAAAGTTGCAGGTAGGG		sequencing primer
AGACCGTTTGTACCGAATTCTGC		sequencing primer
GCAGTGAAAGCCATATCCAAAGC		sequencing primer
AACCGTCCCCAAGATGATTCC		sequencing primer
TCGTTCCATTCGTGAAGAATGC		sequencing primer
GAGGGTAAGCCTCAATAACAGACTGG		sequencing primer
GAACCATTAAACACTTGAGTCATGC		sequencing primer
TTGATGAACGACAAGGAACCG		sequencing primer
TCGACAGCGCTTACGAACG		sequencing primer
CAATTATCAAAGAATCAATGC		sequencing primer
TGCAATTGAATTTAGCTCATCT		sequencing primer
ATTCATGATTGTGACCTTTGTAATCC		sequencing primer
GACAGAGGGCCCAAGTTAAGG		sequencing primer
AGCAAACCATTCGTCCATCC		sequencing primer
TACGACAGGTTGCTAGCTTGG		sequencing primer
AATAATGGATCAGTCACGGCTTCC		sequencing primer
AATCCATCAGATTTTCAACCAGAGAGG		sequencing primer
TGTCAGCCAACCATTCGTCCATCCTAAC		sequencing primer
GGCTTCCCGGAGATGACCCAGATTTTAT		sequencing primer
TTGTTATTTTCATGACTATTACCACCAGCTTCCTCTTA		sequencing primer
AGTGGAGGAGGCACAAAAGTTAGGATGGAC		sequencing primer
CCATGTCTGATAAATACGGGTCGGTGTTC		sequencing primer
TTGTTGATAAGGACGACTAAGAATAAGCAGAAGATA		sequencing primer
ACGATTCTGTCATCATCATTTTCGC		sequencing primer
AGTCGTGTATCGTTCGCTTAATGC		sequencing primer
CATGCCTATCTATTTCCTCCCTTGCCCTC		sequencing primer

Table 3

Gene	Forward
PSMT1	GATTCCCGATTTACTCCTGATG
PSMT1	TGCCTCATGTTATTTCTGTTGCC
PSMT2	ATTGATGTCGGTGGTGGTCACG
PSMT2	GCAACTGTTTCATTAACAGGCACATCC
PSMT3	GCTTCAGCATTGGTTAACGAGTGC
PSMT3	AGACCGTTTGTACCGAATTCTGC
CYP82X1	GAACCATTAAACACTTGAGTCATGC
CYP82X1	TTGATGAACGACAAGGAACCG
CYP82X2	ATGTGGAAAACGGTAAGCAAGTGG
CYP82X2	CAACCTCAATCTAGCTAGAGTCG
CYP82Y1	CAATAATTGAGTAATTTCAGTTCATTC
CYP82Y1	GAATTGTGGTAAAAAATTAGATGCAG
CYP719A21	CAAAGAGTCAATCTGACTCAAGCTAG
CYP719A21	TCAAACCCTGCTACTAACACTTACTTG
PSAT1	TTTTATCGACCTTGAGGAACAATTAGG
PSAT1	GACTTCATGATGAAATCAGATGCAC
PSCXE1	ATGCTGTTGATGCTTTAAACTGGG
PSCXE1	AATAAAAATCCAACAATGGCAGATCC
PSSDR1	GGAAGATGTGAGCCACCTTAAAGC
PSSDR1	GAGAGTAACCACATCTTTGTTGTCGG
BBE	GTTTACTCCCACGTGCATC
TNMT	GTTTACTCCCACGTGCATC
SalR	AAACAATGCTGGGGTTGC
SalAT	TAAGAGAGGGAGACCACGAG
T6ODM	CTTATGAAGCTAGGTAATGGTATGGA
PSMT1	CTCTAAAATGCCAAACGCG
PSMT1	GACCCTTTGGGACTTCCTCG
PSMT1	CGTGTTGTTTGGTCCCTCG
PSMT1	TGCCTCATGTTATTTCTGTTGCC
PSMT1	GATTCCCGATTTACTCCTGATGG
PSMT1	AACACAAAATACGATTACTTACTTTTG

he BAC library				
Primer pairs for the amplification and cloning of fragments selected for VIGS				
CYP82X2	TGTCAGCCAACCATTCGTCCATCCTAAC		sequencing primer	
CYP82X2	TGTTCGATCACGTTGTCTCTTTTTGCCATAA		sequencing primer	
CYP82X2	TAACAATAAAAGTACTGATAATGGTGGTCGAAGGAGAA		sequencing primer	
CYP82Y1	TATTGATGTGGACCAGTACC		sequencing primer	
CYP82Y1	TGTAACTCTTGGTCACATGG		sequencing primer	
CYP82Y1	CGCGTACTTGACATTTAACG		sequencing primer	
CYP82Y1	GGATCATCGCCAAAAGAAAC		sequencing primer	
CYP719A21	CAAAGAGTCAATCTGACTCAAGCTAGC		sequencing primer	
CYP719A21	TGAAATGCCTGAGATCACTAAAATCG		sequencing primer	
CYP719A21	TCAAACCCTGCTACTAACACTTACTTGC		sequencing primer	
CYP719A21	TGTAAAGACACTTCATTGATGGGC		sequencing primer	
CYP719A21	TTCGATTTGTGTAAACATTAATGATATTTGG		sequencing primer	
CYP719A21	GAGATGATCAAGTGGTTTAACCATTCC		sequencing primer	
CYP719A21	CGAGTGCCCATGCAGTGG		sequencing primer	
PSCXE1	AATAAAAATCCAACAATGGCAGATCC		sequencing primer	
PSCXE1	ATGCTGTTGATGCTTTAAACTGGG		sequencing primer	
PSCXE1	GGTTAATCGAGAGATGTTTTGTGGTAGG		sequencing primer	
PSCXE1	CGATGACACAGAGCAAGAACGAC		sequencing primer	
PSCXE1	CGCGGGTATATGTGTAGCAATCG		sequencing primer	
PSCXE1	CGGCAACGCCAGTTCCC		sequencing primer	
PSSDR1	CTAACAGGCAAACAATAACAGGTTGC		sequencing primer	
PSSDR1	GGAAGATGTGAGCCACCTTAAAGC		sequencing primer	
PSSDR1	AAAGGTACTGACAGAAAGAGCTTGCC		sequencing primer	
PSSDR1	AGATACACTGGGAGGAGGATGGG		sequencing primer	
PSSDR1	CGGCAAAATTCATTCCTTGAGC		sequencing primer	
PSSDR1	AACATATAGCCAAAGGACTCTTCG		sequencing primer	
PSAT1	AGGATACACAATGACCCAAC		sequencing primer	
PSAT1	TTTTATCGACCTTGAGGAACAATTAGG		sequencing primer	
PSAT1	TGTTCACTAGGTGGAAAGAG		sequencing primer	
PSAT1	AGTACAATACCGAGAAATCCGACAAG		sequencing primer	
PSAT1	GCTCAATTAATGGAACAGTAGTTACCC		sequencing primer	
			specific PCR conditions:	Primer pairs for genotyping of the F2 mapping population
PsMT1	VIC®-CGTGTTGTTTGGTCCCTCG	GCACACTGTCTTTTTCTTCCACC	30 cylces, 20 s extension at 72°	
PsMT2	VIC®-GCAACTGTTTCATTAACAGGCACATCC	GCCAGCGCTAATACAAGGATGTGG	36 cylces, 50 s extension at 72°	
PsMT3	VIC®-GCAGTGAAAGCCATATCCAAAGC	TCGTTCCATTCGTGAAGAATGC	30 cylces, 30 s extension at 72°	
CYP82X1	VIC®-GCTACGAAAGATAATGGTGCAGC	AGCAAACCATTCGTCCATCC	30 cylces, 30 s extension at 72°	
CYP82X2	VIC®-ATGTGGAAAACGGTAAGCAAGTGG	ACGATTCTGTCATCATCATTTTCGC	30 cylces, 50 s extension at 72°	

PSMT1
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CYP719A21	VIC®-TGAAATGCCTGAGATCACTAAAATCG	GGAATGGTTAAACCACTTGATCATCTC	30 cycles, 30 s extension at 72°
PSCXE1	VIC®-ATGCCAGTTTAAGAGCAATAGAAATGG	GGGAACTGGCGTTGCCG	30 cycles, 30 s extension at 72°
PSSDR1	VIC®-GAAGATGTGAGCCACCTTAAAGC	GCTCAAGGAATGAATTTTGCCG	30 cycles, 30 s extension at 72°
CYP82X2	GTTGACCCAGGAAGCTTTTGC	GGAACATAAGATTTAACTCCGCCTC	Primer pair for PCR amplification of t screening probe
PSMT1	aaactcgagaagcttGGTCATAATCATCAATCAG	aaaggtaccCATGTACTACTACATCATCTCC	
PSMT2	aaactcgagaagcttGTGTAACTAAGCCAGCGC	aaaggtaccACTTGAATATATCACCGC	
CYP82X1	aaaggatccTTTGAGTAATGGTGAAAAGA	aaaggtaccAACATCTACTCTCGAGGATTG	
CYP82X2	aaactcgagaagcttTAGGAGGGTATGTCCGGC	aaaggtaccTTAACTCCGCCTCGGCTCC	
CYP82Y1	aaaggatccTTCAGTTCATTCATGCCG	aaaggtaccGTTCATAGTAAATAATAACAGGCG	
CYP719A21	aaactcgagaagcttATGATCATGAGTAACTTATGGA	aaaggtaccCCAACAGGCCATTCCGTTG	
PSCXE1	aaaggatccTGGCAGATCCTTATGAATTCC	aaaggtaccTTATGATAGGAAGCAGCTTATTC	
PSSDR1	aaaggatccGAAATTGACGAGACAATATGG	aaaggtaccCATTCAAAAACGAATATGTGTGC	
PSAT1	aaaggatccCCTAAGAGAGATCCTCCAACTG	aaaggtaccAATACAAGTATGAAAACAAGAGAATAA	
PSPDS	GAGGTGTTCATTGCCATGTCAA	GTTTCGCAAGCTCCTGCATAGT	

Materials and Methods

[0066] Plant Material Three GSK Australia poppy varieties that predominantly accumulate either noscapine (High Noscapine, HN1), morphine (High Morphine, HM1) or thebaine (High Thebaine HT1), were grown in Maxi (Fleet) Rootrainers™ (Haxnicks, Mere, UK) under glass in 16 hour days at the University of York horticulture facilities. The growth substrate consisted of 4 parts John Innes No. 2, 1 part Perlite and 2 parts Vermiculite. The HM1×HN1 F2 mapping population was grown at the GlaxoSmithKline Australia field-trial site, Latrobe, Tasmania from September 2009 to February 2010.

[0067] Crossing and selfing Crosses were carried out between HN1and HM1 individuals to generate F1 hybrid seed. At the hook stage of inflorescence development, immature stamens were removed from selected HN1 flower buds. HN1 stigmas were fertilized with pollen from synchronously developing HM1 flowers shortly after onset of anthesis. To prevent contaminating pollen from reaching the receptive stigmas, emasculated flowers were covered with a muslin bag for four days after pollination. Both the F1 and F2 generations were self-pollinated to produce F2 and F3 seed, respectively. Self-pollination was ensured by covering the flowers shortly before onset of anthesis with a muslin bag.

[0068] RNA isolation and cDNA synthesis Upper stems (defined as the 2 cm section immediately underneath the capsule) and whole capsules were harvested at two developmental stages represented by 1-3 days and 4-6 days, after petal fall. Five plants were used per developmental stage and cultivar. The material was ground to a fine powder in liquid nitrogen using a mortar and pestle. RNA was isolated from the powder using a CTAB-based extraction method (Chang et al (1993) Plant Mol. Biol. Rep. 11, 113-116) with small modifications: (i) three sequential extractions with chloroform:isoamylalcohol (24:1) were performed and (ii) the RNA was precipitated overnight with lithium chloride at 4°C. After spectrophotometric quantification, equal amounts of RNA were pooled from five plants per cultivar, development stage and organ. The pooled samples underwent a final purification step using an RNeasy Plus MicroKit (Qiagen, Crawley, UK). RNA was typically eluted in 30-100 µl water. cDNA was prepared with the SMART cDNA Library Construction Kit (Clontech, Saint-Germainen-Laye, France) according to the manufacturer's instructions but using SuperScript II Reverse Transcriptase (Invitrogen, Paisley, UK) for first strand synthesis. The CDSIII/3'PCR primer was modified to: 5' ATT CTA GAT CCR ACA TGT TTT TTT TTT TTT TTT TTT TVN 3' where R = A or G, V = A, C or G; N = A/T or C/G. Following digestion with MmeI (New England Biolabs, Hitchin, UK) the cDNA was finally purified using a QIAquick PCR Purification kit (Qiagen, Crawley, UK).

[0069] cDNA Pyrosequencing: Pyrosequencing was performed on the Roche 454 GS-FLX sequencing platform (Branford, CT) using cDNA prepared from the following four samples of each of the three varieties:

1. i. upper stem, 1-3 days after petal fall
2. ii. upper stem, 4 -6 days after petal fall
3. iii. capsule, 1-3 days after petal fall
4. iv. capsule, 4 -6 days after petal fall

[0070] Raw sequence analysis, contiguous sequence assembly and annotation The raw sequence datasets were derived from parallel tagged sequencing on the 454 sequencing platform (Meyer et al (2008) Nature Prot. 3, 267-78). Primer and tag sequences were first removed from all individual sequence reads. Contiguous sequence assembly was only performed on sequences longer than 40 nucleotides and containing less than 3% unknown (N) residues. Those high quality Expressed Sequence Tag (EST) sequences were assembled into unique contiguous sequences with the CAP3 Sequence Assembly Program (Huang and Madan (1999) Genome Res. 9, 868-877), and the resulting contigs were annotated locally using the BLAST2 program (Altschul et al.

(1997) Nucleic Acids Res. 25, 3389-3402) against the non-redundant peptide database downloaded from the NCBI.

[0071] Expression profiling: The number of ESTs associated with a specific consensus sequence representing each of the candidate genes detailed in Figure 1 was counted for each EST library. EST numbers were normalised on the basis of total number of ESTs obtained per library. For each variety, EST counts were combined for the two developmental stages from both stems and capsules. Differences in candidate gene expression levels between organs and varieties were visualised as a heat map using Microsoft Excel.

Preparation of genomic DNA from glasshouse grown plants

[0072] In order to amplify and obtain genomic sequences of the candidate genes 30-50 mgs of leaf material was collected from 4-6 week old glasshouse-grown seedlings from each of the three varieties. Genomic DNA was extracted using the BioSprint 96 Plant kit on the BioSprint 96 Workstation (Qiagen, Crawley, UK) according to the manufacturer's protocol. Extracted DNA was quantified using Hoescht 33258 and normalized to10 ng/ul.

Amplification and sequencing of candidate genes from genomic DNA

[0073] Primers for amplification and Sanger-sequencing of the candidate genes from genomic DNA were based on the respective contiguous sequences assembled from the ESTs or on BAC sequences. The primer sequences are shown in Table 3. PCR amplifications were performed on pools of genomic DNA comprising DNA from four individuals. Amplification was typically carried out on 10 ng genomic DNA in 1× Phusion High Fidelity Buffer supplemented with 200 nM forward and reverse primers, 0.2 mM dNTPs, 0.02 units/µl Phusion Hot Start DNA Polymerase (Finnzymes, Vantaa, Finnland). Standard PCR conditions were used throughout with annealing temperatures and times dependent on primers and PCR equipment.

DNA extraction from the field-grown F2 mapping population

[0074] 40-50 mg of leaf tissue was harvested from F2 plants at the 'small rosette' growth stage (~10 leaves present on each plant) into 1.2 ml sample tubes. A 3 mm tungsten carbide bead was added to each tube and samples were kept at -80°C for a minimum of two hours prior to freeze-drying for 18 hours. Following freeze drying, samples were powdered by bead-milling (Model TissueLyser, Qiagen, Hilden, Germany) at 30 Hz for two 60 s cycles separated by plate inversion. DNA extraction was performed with the Nucleospin Plant II kit (Macherey-Nagel, Düren, Germany) using the supplied Buffer Set PL2/3 following the manufacturer's protocol for centrifugal extraction. DNA was quantified by UV- spectroscopy.

Genotyping of the HN1×HM1 F2 mapping population for the presence or absence of the HN1-specific candidate genes

[0075] Plants of the F2 mapping population were genotyped for the presence or absence of eight candidate genes. The gene primer pairs (Table 3) were designed with fluorescent tags (5'-VIC®-labeled) for use on the ABI 3730xl capillary apparatus (Applied Biosystems, Foster City, CA). PCR amplifications were typically carried out on 10 ng genomic DNA in 1x GoTaq buffer supplemented with 1 mM $MgCl_2$, 500 nM forward and reverse primer, 0.125 mM dNTPs, 0.1 U GoTaq (Promega, Southampton, UK). The amplification conditions were: 1 min 94°C, 30-36 cycles of 30 s denaturation at 94°C, 30 s annealing at 62°C and 20-50 s extension at 72°C, followed by a final extension for 5 min at 72°C. Cycle number and extension times depended on the candidate gene (Table 3). Amplification products were diluted 1:20 in H_2O and fractionated on an ABI 3730xl capillary sequencer (Applied Biosystems, Foster City, CA). Data were scored using GeneMarker™ software (Softgenetics, State College, PA).

Poppy straw analysis from field grown F2 plants

[0076] Poppy capsules were harvested by hand from the mapping population once capsules had dried to approximately 10% moisture on the plant. After manually separating the seed from the capsule, the capsule straw samples (Poppy Straw) were then ground in a ball mill (Model MM04, Retsch, Haan, Germany) into a fine powder. Samples of ground poppy straw were then weighed accurately to 2 ± 0.003 g and extracted in 50 ml of a 10% acetic acid solution. The extraction suspension was shaken on an orbital shaker at 200 rpm for a minimum of 10 min, then filtered to provide a clear filtrate. The final filtrate was passed through a 0.22 µm filter prior to analysis. The loss on drying (LOD) of the straw was determined by drying in an oven at 105°C for 3 hours.

[0077] All solutions were analysed using a Waters Acquity UPLC system (Waters Ltd., Elstree, UK). fitted with a Waters Acquity BEH C18 column, 2.1 mm × 100 mm with 1.7 micron packing. The mobile phase used a gradient profile with eluent A consisting of 10 mM ammonium bicarbonate of pH 10.2 and eluent B methanol. The mobile phase gradient conditions used are as listed in the table below with a linear gradient. The flow rate was 0.5 ml per minute and the column maintained at 60°C. The injection volume was 2 µl and eluted peaks were ionised in positive APCI mode and detected within 5 ppm mass accuracy using a Thermo LTQ-Orbitrap. The runs were controlled by Thermo Xcalibur software (Thermo Fisher Scientific Inc., Hemel Hempstead, UK).

- Gradient Flow Program:

[0078]

TIME (minutes)	% Eluent A	% Eluent B	Flow (ml/min)
0.0	98.	2.0	0.50
0.2	98.0	2.0	0.50

TIME (minutes)	% Eluent A	% Eluent B	Flow (ml/min)
0.5	60.0	40	0.50
4.0	20.0	80.0	0.50
4.5	20.0	80.0	0.50

[0079] Mass spectra were collected over the 150-900 m/z range at a resolution setting of 7500. All data analysis was carried out in the R programming language in a 64-bit Linux environment (R 2.11). Peak-picking was performed using the Bioconductor package, XCMS (Smith et al (2006) Anal. Chem. 78, 779-787), employing the centWave algorithm (Tautenhahn et al (2008) BMC Bioinformatics 9, 504). Redundancy in peak lists was reduced using the CAMERA package (Kuhl et al (2012) Anal. Chem. 84, 283-289). Alkaloids were identified by comparing exact mass and retention time values to those of standards and quantified by their pseudomolecular ion areas using custom R scripts.

Bacterial Artificial Chromosome (BAC) library construction

[0080] The HN1 BAC library was constructed from high molecular weight (HMW) genomic DNA processed at Amplicon Express, Inc. (Pullman, WA) from four week old seedlings using the method described (Tao et al (2002) Theor. Appl. Genet. 105, 1058-1066). The HMW DNA was partially digested with the restriction enzyme HindIII and size selected prior to ligation of fragments into the pCC1BAC vector (Epicentre Biotechnologies, Madison, WI) and transformation of DH10B *E. coli* cells, which were then plated on Luria-Bertani (LB) agar with chloramphenicol, X-gal and IPTG at appropriate concentrations. Clones were robotically picked with a Genetix QPIX (Molecular Devices, Sunnyvale, CA) into 240 384-well plates containing LB freezing media. Plates were incubated for 16 hours, replicated and then frozen at -80°C. The replicated copy was used as a source plate for nylon filters that were made and used for screening using the PCR DIG Probe Synthesis Kit (Roche Applied Science, Indianapolis, IN). To estimate insert sizes, DNA aliquots of 10 BAC minipreps were digested with 5U of NotI enzyme for 3 hours at 37°C. The digestion products were separated by pulsed-field gel electrophoresis (CHEF-DRIII system, Bio-Rad, Hercules, CA) in a 1% agarose gel in TBE. Insert sizes were compared to those of the Lambda Ladder MidRange I PFG Marker (New England Biolabs, Ipswich, MA). Electrophoresis was carried out for 18 hours at 14°C with an initial switch time of 5 s, a final switch time of 15 s, in a voltage gradient of 6 V/cm. The average BAC clone size for the library was found to be 150 Kb.

Filter construction and screening

[0081] Filter design and screening was carried out at Amplicon Express, Inc. (Pullman, WA). Bioassay dishes containing LB agar plate media and 12.5 µg/mL chloramphenicol were prepared. Positively charged nylon Amersham Hybond-N^+ membrane (GE Healthcare Bio-Sciences, Piscataway, NJ) was applied to the media surface and the GeneMachines G3 (Genomics

Solutions, Bath, UK) was used to robotically grid 18,432 clones in duplicate on filters. The filters were incubated at 37°C for 12 to 14 hours. The filters were processed using the nylon filter lysis method (Sambrook et al., Molecular Cloning: A Laboratory Manual (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 2001, ed. 3, vol.1, chap. 1) with slight modifications. Following processing, the DNA was linked to the hybridization membrane filters according to the Hybond N+ manual by baking at 80°C for 2 hours. To screen the library a 643 bp digoxigenin (DIG)-labeled probe representing position 2161-2803 in the genomic sequence of *CYP82X2* (SEQ ID NO 6) was generated from 1.5 ng gDNA by PCR reaction using the primers shown in Table 3 and the PCR DIG synthesis kit (Roche Applied Science, Indianapolis, IN) according to the manufacturer's instructions. A non-labeled probe was amplified, diluted and spotted to each filter in the following dilutions of 2 ng, 1 ng, 0.1 ng and 0.0 ng as a positive control. The controls were baked at 80°C for 30 min. Following a 30 min prehybridizing wash in DIG EasyHyb solution at 45°C approximately 0.5 µl of denatured DIG labeled PCR product was added per ml of hybridization solution with the nylon filters and incubated with gentle shaking overnight at 45°C. The nylon filters were washed twice in a 2x standard sodium citrate (SSC), 0.1% sodium dodecyl sulfate (SDS) buffer at room temperature for 5 min each, and twice with a 0.5x SSC, 0.1% SDS buffer at 65°C for 15 minutes each. The hybridized probe was detected using NBT/BCIP stock solution according to the manufacturer's instructions (Roche Applied Science, Indianapolis, IN) and was found to hybridize to six BAC clones.

[0082] BAC sequencing and automated sequence assembly: The six positive BAC clones from the BAC library were sequenced at Amplicon Express, Inc. (Pullman, WA) by Focused Genome Sequencing (FGS) with an average depth of 100x coverage. FGS is a Next Generation Sequencing (NGS) method developed at Amplicon Express that allows very high quality assembly of BAC clone sequence data using the Illumina HiSeq platform (Illumina,Inc, San Diego, CA). The proprietary FGS process makes NGS tagged libraries of BAC clones and generates a consensus sequence of the BAC clones with all reads assembled at 80 bp overlap and 98% identity. The gapped contiguous sequences were ordered and orientated manually based on mate pair sequences from four libraries of insert size 5000, 2000, 500 and 170 bp. Overlapping BAC clones, PS_BAC193L09, PS_BAC179L19, PS_BAC150A23 and PS_BAC164F07, which together encoded all 10 genes from the HN1 cluster, were selected for further sequence assembly. Where possible, gaps and ambiguous regions on both BAC clones were covered by primer walking with traditional Sanger sequencing to validate the assembly. Combination of the four overlapping BAC sequences gave a single continuous consensus sequence assembly of 401 Kb. The sequences of the 10 genes from the HN1 cluster were determined independently by Sanger sequencing and the 100% agreement of the Sanger determined gene sequences with the assembly from FGS provided quality assurance for the whole assembly.

[0083] Annotation of the assembled sequence: The sequences of the four BAC clones were annotated with an automated gene prediction program FGENESH (Salamov and Solovyev (2002) Genome Res. 10, 516-522). The gene structure including exon-intron arrangement for the 10 genes in the HN1 cluster was validated by comparison with cDNA sequence for each gene. cDNA sequence was not available for any of the remaining ORFs detailed in Fig. 3 since they are not represented in any of the EST libraries. The predicted function of all ORFs was evaluated by BLAST analysis (Altschul et al (1997) Nucleic Acids Res. 25, 3389-3402) and those ORFs with

significant hits (e-value less than $1e^{-8}$) were included in Fig.3.

Generation of plasmid constructs for Virus Induced Gene Silencing (VIGS)

[0084] The tobacco rattle virus (TRV) based gene silencing system (Liu et al (2002) Plant J. 30, 415-422) was used to investigate the gene function of *PSMT1, PSMT2, CYP719A21, CYP82X2, PSSDR1* and *PSCXE1.* DNA fragments selected for silencing were amplified by PCR and cloned into the silencing vector pTRV2 (GenBank accession no: AF406991). They were linked to a 129 bp-long fragment (SEQ ID NO: 30) of the *P. somniferum PHYTOENE DESATURASE* gene (*PSPDS*) in order to simultaneously silence the respective candidate genes and *PSPDS.* Plants displaying the photo-bleaching phenotype resulting from *PSPDS* silencing (Hileman et al (2005) Plant J. 44, 334-341) were identified as plants successfully infected with the respective silencing constructs and selected for further analysis.

[0085] Generation of the pTRV2:PDS construct: A 622 bp fragment of *PSPDS* was amplified from cDNA prepared from HN1 using primers shown in Table 3. Sau3AI digestion of the 622 bp PCR product yielded among others a fragment of 129 bp (SEQ ID NO: 30) which was cloned into the BamHI site of the pTRV2 vector. The orientation and fidelity was confirmed by sequencing and the resulting pTRV2:PDS vector was used in the generation of the VIGS construct for each candidate gene. The pTRV2:PDS construct also served as the control in the VIGS experiments.

[0086] DNA fragments selected for silencing the respective candidate genes were amplified from either HN1 genomic or cDNA. Primers used for amplification as well as the positions of the selected sequences within the respective open reading frames are shown in Table 3. The *PSMT1, CYP719A21* and *CYP82X2* fragments were first cloned into pTV00 (Ratcliff et al (2001) Plant J., 237-245) using HindIII and KpnI and then subcloned into pTRV2:PDS using BamHI and KpnI. *PSMT2, PSCXE1* and *PSSDR1* fragments were cloned directly into pTRV2:PDS using BamHI and KpnI. The orientation and fidelity of all constructs was confirmed by sequencing.

[0087] Transformation of *Agrobacterium tumefaciens* with VIGS constructs: VIGS constructs were propagated in *E. coli* strain DH5α and transformed into electrocompetent *Agrobacterium tumefaciens* (strain GV3101) by electroporation.

[0088] Infiltration of plants: Separate overnight liquid cultures of *A. tumefaciens* containing individual VIGS constructs (each consisting of a selected DNA fragment from the target gene linked to the 129 bp-long fragment from the *P. somniferum PHYTOENE DESATURASE* gene) were used to inoculate LB medium containing 10 mM MES, 20 µM acetosyringone and 50µg/ml kanamycin. Cultures were maintained at 28°C for 24 hours, harvested by centrifugation at 3000xg for 20 min, and resuspended in infiltration solution (10 mM MES, 200 µM acetosyringone, 10 mM $MgCl_2$,) to an OD_{600} of 2.5. *A. tumefaciens* harbouring the individual VIGS constructs including the control, pTRV2:PDS, were each mixed 1:1 (v/v) with *A. tumefaciens* containing pTRV1 (GenBank accession no: AF406990), and incubated for two hours at 22°C prior to infiltration. Two week old seedlings of HN1 grown under standard greenhouse conditions (22°C, 16h photoperiod), with emerging first leaves, were infiltrated as described (Hagel and Facchini (2010) Nat. Chem. Biol. 6,

273-275).

[0089] Latex and capsule analysis of silenced plants: Leaf latex of infiltrated plants displaying photo-bleaching as a visual marker for successful infection and silencing was analyzed when the first flower buds emerged (~7 week old plants). Latex was collected from cut petioles, with a single drop dispersed into 500 µl of 10% acetic acid. This was diluted 10x in 1% acetic acid to give an alkaloid solution in 2% acetic acid for further analysis. Capsules were harvested from the same plants used for latex analysis and single capsules were ground to a fine powder in a ball mill (Model MM04, Retsch, Haan, Germany). Samples of ground poppy straw were then weighed accurately to 10 ± 0.1 mg and extracted in 0.5 ml of a 10% acetic acid solution with gentle shaking for 1h at room temperature. Samples were then clarified by centrifugation and a 50 µl subsample diluted 10x in 1% acetic acid to give an alkaloid solution in 2% acetic acid for further analysis. All solutions were analyzed as described for the poppy straw analysis from field grown F2 plants. Likewise, all data analysis was carried out using the R programming language. Putative alkaloid peaks were quantified by their pseudomolecular ion areas using custom scripts. Peak lists were compiled and any peak-wise significant differences between samples were identified using 1-way ANOVA with p-values adjusted using the Bonferroni correction for the number of unique peaks in the data set. For any peak-wise comparisons with adjusted p-values < 0.05, Tukey's HSD test was used to identify peaks that were significantly different between any given sample and the control. Alkaloids were identified by comparing exact mass and retention time values to those of standards. Where standards were not available, the Bioconductor rcdk package (Smith et al (2006) Anal. Chem. 78, 779-787) was used to generate pseudomolecular formulae from exact masses within elemental constraints C = 1 100, H = 1 200, O = 0 200, N = 0 3 and mass accuracy < 5ppm. The hit with the lowest ppm error within these constraints was used to assign a putative formula.

Example 1

Transcriptomic analysis reveals the exclusive expression of 10 genes encoding five distinct enzyme classes in a high noscapine producing poppy variety, HN1. These genes are absent from the genome of two noscapine non-producing varieties.

[0090] Capsule extract from three opium poppy varieties developed in Tasmania for alkaloid production designated as High Morphine 1 (HM1), High Thebaine 1 (HT1) and High Noscapine 1 (HN1) on the basis of the most abundant alkaloid in each case (Fig. 1A) underwent metabolite profiling. Noscapine was found to be unique to HN1 relative to HM1 and HT1. Roche 454 pyrosequencing was performed on cDNA libraries derived from stem and capsule tissue from all three varieties. Analysis of Expressed Sequence Tag (EST) abundance led to the discovery of a number of previously uncharacterized genes that are expressed in the HN1 variety but are completely absent from the HM1 and HT1 EST libraries (Fig. 1B). The corresponding genes were putatively identified as three O-methyltransferases (*PSMT1, PSMT2,* PSMT3), four cytochrome P450s (*CYP82X1, CYP82X2, CYP82X3* and *CYP719A21*), an acetyltransferase (*PSAT1*), a carboxylesterase (*PSCXE1*) and a short-chain dehydrogenase/reductase (*PSSDR1*). In contrast a

number of other functionally characterized genes associated with benzylisoquinoline alkaloid synthesis, including *Berberine Bridge Enzyme* (*BBE*), *Tetrahydroprotoberberine cis-N-MethylTransferase (TNMT), Salutaridine Reductase (SalR), Salutaridinol 7-O-AcetylTransferase* (*SalAT*) and *Thebaine 6-O-demethylase* (*T6ODM*) were expressed in all three varieties (Fig. 1B). PCR analysis on genomic DNA from all three varieties revealed that the genes exclusively expressed in the HN1 variety are present as expected in the genome of HN1 but absent from the genomes of the HM1 and HT1 varieties (Fig.1B and Fig. 5).

Example 2

Analysis of an F2 mapping population shows the genes are tightly linked in HN1 and their presence is associated with the production of noscapine.

[0091] An F2 mapping population of 271 individuals was generated using HN1 and HM1 as parents. Genotyping of the field grown F2 population revealed that the HN1 specific genes are tightly linked and associated with the presence of noscapine suggesting they occur as a gene cluster involved in noscapine biosynthesis (Fig. 2B). Analysis of noscapine levels in field grown F2 capsules revealed that individuals containing this putative gene cluster fall into two classes. The first class containing 150 individuals, have relatively low levels of noscapine and the second class containing 63 individuals exhibit the high noscapine trait of the parental HN1 variety (Fig. 2B). The 58 F2 individuals that lack the putative gene cluster contain undetectable levels of noscapine (Fig. 2B). F3 family analysis confirmed that F2 individuals exhibiting the high noscapine trait were homozygous for the gene cluster while those exhibiting the low noscapine trait were heterozygous (Table 2). Noscapine levels in both the F1 population (Fig. 2A) and the heterozygous F2 class are much lower than the intermediate levels expected for a semi-dominant trait, suggesting involvement of some form of repression. The step change to high noscapine in homozygous F2 class suggests this trait is linked to the gene cluster locus rather than spread quantitatively among other loci.

Example 3

Bacterial artificial chromosome sequencing confirms that the 10 genes exist as a complex gene cluster

[0092] To further characterize the putative noscapine gene cluster, a Bacterial Artificial Chromosome (BAC) library was prepared from genomic DNA isolated from HN1 and six overlapping BACs containing genes from the cluster were identified. Next generation and Sanger sequencing was used to generate a high quality assembly of 401 Kb confirming the arrangement of the 10 genes in a cluster spanning 221 Kb (Fig 3). Only one other homologous gene, a carboxylesterase (*PSCXE2*), was found in the genomic sequence flanking the gene

cluster (Fig 3) but *PSCXE2* was not represented in any of our EST libraries. Interspersed among the ten genes are both retrotransposon and DNA transposable element (TE) sequences (Fig 3), which may have some function in gene rearrangement for cluster formation as thought to be the case for the thalianol and marneral clusters from *A. thaliana* (Field et al (2011) PNAS 108, 16116-16121).

Example 4

Virus induced gene silencing results in accumulation of pathway intermediates allowing gene function to be linked to noscapine synthesis and a novel bifurcated biosynthetic pathway to be proposed

[0093] In order to functionally characterize the genes in the HN1 cluster Virus Induced Gene Silencing (VIGS) was performed on poppy seedlings. VIGS in poppy seedlings persists through to mature plant stages (Hileman et al (2005) Plant J. 44, 334-341), and therefore both leaf latex and capsule extracts were routinely assayed (Fig. 4). Silencing *PSMT1* resulted in accumulation of scoulerine in capsules and also low levels of reticuline in latex, indicating that this gene product is responsible for the first committed step in the pathway to noscapine synthesis (Fig 4A). The predicted product of PSMT1 is tetrahydrocolumbamine (Fig. 6), which accumulated in seedlings and capsules that were silenced for *CYP719A21* (Fig 4B). CYP719A21 shows high homology to cytochrome P450 oxidases that act as methylenedioxy bridge-forming enzymes (Diaz Chávez et al (2011) Arch. Biochem. Biophys. 507, 186193; Ikezawa et al (2009) Plant Cell Rep. 28, 123-133). Therefore CYP719A21 may encode a canadine synthase.Fig. 6). Silencing of a second cytochrome P450 gene, *CYP82X2,* resulted in accumulation of several secoberbine intermediates some of which may represent side products to the main synthetic pathway (Fig. 4C, Fig. 6). Silencing of the carboxylesterase gene *PSCXE1* resulted in accumulation of up to 20% total alkaloid content of putative papaveroxine (Fig. 6) implying acetylation of a secoberbine intermediate as depicted in Fig. 4G. The *PSAT1* gene from the HN1 cluster is an obvious candidate for this reaction. Silencing of *PSSDR1* resulted in accumulation of what was putatively identified as narcotinehemiacetal (Fig. 6), an immediate precursor of noscapine (Fig 4G). These data support a biosynthetic route to noscapine that involves early O-methylation of a secoberbine intermediate at the position equivalent to the C4' hydroxyl group of noscapine (Fig. 4G). However, silencing *PSMT2,* resulted in accumulation of up to 20% narcotoline, indicating that O-methylation at the C4' hydroxyl group can also occur as a final step in noscapine production (Fig. 4F). These results imply bifurcation of the main pathway at the secoberbine intermediate stage with *PSMT2* being responsible for both the O-methylation of a secoberbine intermediate and narcotoline. Silencing *PSMT2* results in accumulation of high levels of narcotoline as flux is directed down the desmethyl branch of the pathway (Fig. 4F).

SEQUENCE LISTING

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<213> Papaver somniferum

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<211> 1915

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<213> Papaver somniferum

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<212> DNA
<213> Papaver somniferum

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<213> Papaver somniferum

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<211> 390

<212> PRT

<213> Papaver somniferum

<400> 11

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Val Cys Tyr Leu Ser Glu Thr Ala Asn Leu Gly Lys Leu Ile Cys Ile
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Pro Met Ala Leu Arg Ala Ala Met Glu Leu Asn Val Phe Gln Leu Ile
    50                  55                  60

Ser Lys Phe Gly Thr Asp Ala Lys Val Ser Ala Ser Glu Ile Ala Ser
65                  70                  75                  80

Lys Met Pro Asn Ala Lys Asn Asn Pro Glu Ala Ala Met Tyr Leu Asp
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Arg Ile Leu Arg Leu Leu Gly Ala Ser Ser Ile Leu Ser Val Ser Thr
            100                 105                 110

Thr Lys Lys Ser Ile Asn Arg Gly Gly Asp Asp Val Val Val His Glu
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Lys Leu Tyr Gly Leu Thr Asn Ser Ser Cys Cys Leu Val Pro Arg Gln
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Glu Asp Gly Val Ser Leu Val Glu Glu Leu Leu Phe Thr Ser Asp Lys
145                 150                 155                 160

Val Val Val Asp Ser Phe Phe Lys Leu Lys Cys Val Val Glu Glu Lys
                165                 170                 175

Asp Ser Val Pro Phe Glu Val Ala His Gly Ala Lys Ile Phe Glu Tyr
            180                 185                 190

Ala Ala Thr Glu Pro Arg Met Asn Gln Val Phe Asn Asp Gly Met Ala
        195                 200                 205

Val Phe Ser Ile Val Val Phe Glu Ala Val Phe Arg Val Tyr Asp Gly
    210                 215                 220

Phe Leu Asp Met Lys Glu Leu Leu Asp Val Gly Gly Gly Ile Gly Thr
225                 230                 235                 240

Ser Val Ser Lys Ile Val Ala Lys Tyr Pro Leu Ile Arg Gly Val Asn
                245                 250                 255

Phe Asp Leu Pro His Val Ile Ser Val Ala Pro Gln Tyr Pro Gly Val
            260                 265                 270
```

```
Glu His Val Ala Gly Asp Met Phe Glu Glu Val Pro Lys Gly Gln Asn
        275                 280                 285

Met Leu Leu Lys Trp Val Leu His Asp Trp Gly Asp Glu Arg Cys Val
    290                 295                 300

Lys Leu Leu Lys Asn Cys Trp Asn Ser Leu Pro Val Gly Gly Lys Val
305                 310                 315                 320

Leu Ile Ile Glu Phe Val Leu Pro Asn Glu Leu Gly Asn Asn Ala Glu
                325                 330                 335

Ser Phe Asn Ala Leu Ile Pro Asp Leu Leu Leu Met Ala Leu Asn Pro
            340                 345                 350

Gly Gly Lys Glu Arg Thr Ile Ser Glu Tyr Asp Asp Leu Gly Lys Ala
        355                 360                 365

Ala Gly Phe Ile Lys Thr Ile Pro Ile Pro Ile Ser Asn Gly Leu His
    370                 375                 380

Val Ile Glu Phe His Lys
385                 390
```

<210> 12

<211> 356

<212> PRT

<213> Papaver somniferum

<400> 12

```
Met Glu Ile His Leu Glu Ser Gln Glu Gln Glu Met Lys Tyr Gln Ser
1               5                   10                  15

Gln Ile Trp Asn Gln Ile Cys Gly Thr Val Asp Thr Ser Val Leu Arg
            20                  25                  30

Cys Ala Ile Gln Leu Gly Ile Phe Asp Ala Ile His Asn Ser Gly Lys
        35                  40                  45

Pro Met Ile Thr Leu Thr Glu Leu Ser Ser Ile Val Ser Ser Pro Ser
    50                  55                  60

Ser Ser Ser Ile Glu Pro Cys Asn Leu Tyr Arg Leu Val Arg Tyr Leu
65                  70                  75                  80

Ser Gln Met Asp Leu Ile Ser Ile Gly Glu Cys Leu Asn Glu Ala Thr
                85                  90                  95

Val Ser Leu Thr Gly Thr Ser Lys Leu Leu Leu Arg Asn Gln Glu Lys
            100                 105                 110

Ser Leu Ile Asp Trp Val Leu Ala Ile Ser Cys Glu Met Met Val Val
        115                 120                 125

Val Trp His Glu Leu Ser Ser Ser Val Ser Thr Pro Ala Asp Glu Pro
    130                 135                 140

Pro Ile Phe Gln Lys Val His Gly Lys Asn Ala Leu Glu Leu Ala Gly
```

145 150 155 160

Glu Phe Pro Glu Trp Asn Asp Leu Ile Asn Asn Ala Met Thr Ser Asp
165 170 175

Thr Ser Val Thr Lys Pro Ala Leu Ile Gln Gly Cys Gly Lys Ile Leu
180 185 190

Asn Gly Val Thr Ser Leu Ile Asp Val Gly Gly Gly His Gly Ala Thr
195 200 205

Met Ala Tyr Ile Val Glu Ala Phe Pro His Ile Lys Gly Ala Val Ile
210 215 220

Asp Leu Pro His Val Val Glu Ala Ala Pro Glu Arg Pro Gly Val Glu
225 230 235 240

Phe Ile Ser Gly Asp Ile Phe Lys Ser Ile Ser Asn Ala Asp Ala Val
245 250 255

Leu Leu Lys Tyr Val Leu His Asn Trp Glu Asp Thr Glu Cys Val Asn
260 265 270

Leu Leu Lys Arg Cys Lys Glu Ala Val Pro Ala Asp Lys Gly Lys Val
275 280 285

Ile Ile Met Asp Leu Val Ile Asp Asp Asp Asp Asn Ser Ile Leu Thr
290 295 300

Gln Ala Lys Leu Ser Leu Asp Leu Thr Val Met Asn His Gly Gly Gly
305 310 315 320

Arg Glu Arg Thr Lys Glu Asp Trp Arg Asn Leu Ile Glu Met Ser Gly
325 330 335

Phe Ser Arg His Glu Ile Ile Pro Ile Ser Ala Met Pro Ser Ile Ile
340 345 350

Val Ala Tyr Pro
355

<210> 13
<211> 339
<212> PRT
<213> Papaver somniferum

<400> 13
Met Glu Val Val Ser Lys Ile Asp Gln Glu Asn Gln Ala Lys Ile Trp
1 5 10 15

Lys Gln Ile Phe Gly Phe Ala Glu Ser Leu Val Leu Lys Cys Ala Val
20 25 30

Gln Leu Glu Ile Ala Glu Thr Leu His Asn Asn Val Lys Pro Met Ser
35 40 45

Leu Ser Glu Leu Ala Ser Lys Leu Pro Ala Gln Pro Val Asn Glu Asp

```
    50                  55                  60

Arg Leu Tyr Arg Ile Leu His Phe Leu Val His Met Lys Leu Phe Asn
65                  70                  75                  80

Lys Asp Ala Thr Thr Gln Lys Tyr Ser Leu Ala Pro Pro Ala Lys Tyr
                85                  90                  95

Leu Leu Lys Gly Trp Glu Lys Ser Met Val Pro Ser Ile Leu Ser Val
            100                 105                 110

Thr Asp Lys Asp Phe Thr Ala Pro Trp Asn His Leu Gly Asp Gly Leu
        115                 120                 125

Thr Gly Asn Cys Asn Ala Phe Glu Lys Ala Leu Gly Lys Gly Ile Arg
    130                 135                 140

Val Tyr Met Arg Glu Asn Pro Glu Lys Asp Gln Leu Phe Asn Glu Gly
145                 150                 155                 160

Met Ala Cys Asp Thr Arg Leu Phe Ala Ser Ala Leu Val Asn Glu Cys
                165                 170                 175

Lys Ser Ile Phe Ser Asp Gly Ile Asn Thr Leu Ala Gly Val Gly Arg
            180                 185                 190

Gly Thr Gly Thr Ala Val Lys Ala Ile Ser Lys Ala Phe Pro Asp Ile
        195                 200                 205

Lys Cys Thr Ile His Asp Leu Pro Glu Val Thr Ser Lys Asn Ser Lys
    210                 215                 220

Ile Pro Arg Asp Val Phe Lys Ser Val Pro Ser Ala Asp Ala Ile Phe
225                 230                 235                 240

Met Lys Ser Ile Leu His Glu Trp Asn Asp Glu Glu Cys Ile Gln Ile
                245                 250                 255

Leu Lys Arg Cys Lys Glu Ala Ile Pro Lys Gly Gly Lys Val Ile Ile
            260                 265                 270

Ala Asp Val Val Ile Asp Met Asp Ser Thr His Pro Tyr Ser Lys Ser
        275                 280                 285

Arg Leu Ala Met Asp Leu Ala Met Met Leu His Thr Gly Gly Lys Glu
    290                 295                 300

Arg Thr Glu Glu Asp Trp Lys Lys Leu Ile Asp Ala Ala Gly Phe Ala
305                 310                 315                 320

Ser Cys Lys Ile Thr Lys Leu Ser Ala Leu Gln Ser Val Ile Glu Ala
                325                 330                 335

Tyr Pro His
```

<210> 14

<211> 540

<212> PRT

<213> Papaver somniferum

<400> 14

```
Met Glu Leu Phe Ile Lys Leu Pro Phe Ile Gln Pro Ile Pro Phe Ser
1               5                  10                  15

Ile Ile Leu Val Thr Thr Val Ser Ile Val Leu Leu Tyr Ser Val Phe
            20                  25                  30

Phe Trp Val Thr Asp Lys Lys Lys Lys Arg Lys Lys Ala Pro Asn Ala
        35                  40                  45

Ala Gly Ala Trp Pro Leu Ile Gly His Leu Arg Leu Leu Met Asn Asp
    50                  55                  60

Lys Glu Pro Leu Tyr Arg Ala Leu Gly Ser Met Ala Asp Lys Tyr Gly
65                  70                  75                  80

Pro Ala Phe Asn Ile Arg Leu Gly Asn Gln Glu Val Leu Val Val Ser
                85                  90                  95

Asn Trp Glu Met Val Lys Gln Cys Phe Gly Asn Gln Asn Asp Lys Leu
            100                 105                 110

Phe Ser Asn Arg Gln Thr Thr Leu Ala Ala Lys Tyr Met Leu Asn Gln
        115                 120                 125

Thr Thr Ser Ser Gly Phe Ala Pro Tyr Gly Pro Tyr Trp Arg Glu Leu
    130                 135                 140

Arg Lys Ile Met Val Gln Gln Leu Leu Ser Lys Gln Ser Leu Glu Ser
145                 150                 155                 160

Trp Lys His Leu Lys Ile Lys Glu Met Asp Ala Ser Phe Ser Lys Leu
                165                 170                 175

Asn Glu Leu Cys Asn Asn Asn Gly Thr Gly Thr Ala Thr Leu Ile Arg
            180                 185                 190

Met Asp Glu Trp Phe Ala Glu Leu Thr Phe Asn Val Ile Ala Arg Asn
        195                 200                 205

Val Phe Gly Tyr Gln Ser Gly Gly Arg Ser Thr Ala Leu Thr Asn Gly
    210                 215                 220

Asp Thr Glu Ser Lys Gly Glu Arg Tyr Lys Lys Thr Leu Glu Glu Ala
225                 230                 235                 240

Leu His Leu Met Ser Ile Phe Ala Val Ser Asp Ile Phe Pro Ser Leu
                245                 250                 255

Glu Trp Val Asp Arg Leu Arg Gly Leu Ile Arg Asn Met Lys Arg Phe
            260                 265                 270

Gly Asp Glu Leu Asn Ser Ile Ala Gly Cys Leu Ile Glu Glu His Arg
        275                 280                 285
```

```
Gln Lys Arg Leu Gln Ser Val Ser Lys Ser Asp Lys Gly Val Gly Asp
    290                 295                 300

Glu Gln Asp Phe Val Asp Val Leu Leu Ser Val Ala Glu Lys Ser Gln
305                 310                 315                 320

Leu Pro Gly Asp Asp Pro Asp Leu Val Ile Lys Ser Met Ile Leu Glu
                325                 330                 335

Ile Val Ser Gly Gly Ser Glu Thr Thr Ser Ser Thr Leu Thr Trp Ala
            340                 345                 350

Leu Cys Leu Leu Leu Asn His Pro His Val Leu Lys Lys Ala Lys Glu
        355                 360                 365

Glu Leu Asp Thr His Val Gly Lys Asp Arg His Val Glu Glu Ser Asp
    370                 375                 380

Thr Pro Lys Leu Val Tyr Ile Asn Ala Ile Ile Lys Glu Ser Met Arg
385                 390                 395                 400

Leu Tyr Pro Asn Gly Ala Met Leu Asp Arg Leu Ala Leu Glu Glu Cys
                405                 410                 415

Glu Val Gly Gly Phe His Val Pro Ala Gly Gly Arg Leu Phe Val Asn

            420                 425                 430

Val Trp Lys Ile Gln Arg Asp Pro Ser Val Trp Glu Asn Pro Leu Glu
        435                 440                 445

Phe Lys Pro Glu Arg Trp Phe Leu Ser Asn Gly Glu Lys Met Asp Val
    450                 455                 460

Asp Tyr Lys Gly His Asn His Glu Phe Ile Pro Phe Gly Ile Gly Arg
465                 470                 475                 480

Arg Met Cys Ala Gly Met Leu Trp Ala Ser Glu Val Ile His Leu Val
                485                 490                 495

Leu Pro Arg Leu Ile His Gly Phe Asp Met Lys Ala Ala Ser Ala Asn
            500                 505                 510

Gly Lys Val Asp Met Ala Glu Met Ala Gly Met Val Ile Cys Phe Lys
        515                 520                 525

Lys Thr Pro Leu Glu Val Met Val Asn Pro Arg Glu
    530                 535                 540
```

<210> 15

<211> 486

<212> PRT

<213> Papaver somniferum

<400> 15

```
Met Ile Met Ser Asn Leu Trp Ile Leu Thr Leu Ile Ser Thr Ile Leu
1               5                   10                  15
```

```
Ala Val Phe Ala Ala Val Leu Ile Ile Phe Arg Arg Arg Ile Ser Ala
            20                  25                  30

Ser Thr Thr Glu Trp Pro Val Gly Pro Lys Thr Leu Pro Ile Ile Gly
        35                  40                  45

Asn Leu His Ile Leu Gly Gly Thr Ala Leu His Val Val Leu His Lys
    50                  55                  60

Leu Ala Glu Val Tyr Gly Ser Val Met Thr Ile Trp Ile Gly Ser Trp
65                  70                  75                  80

Lys Pro Val Ile Ile Val Ser Asp Phe Asp Arg Ala Trp Glu Val Leu
                85                  90                  95

Val Asn Lys Ser Ser Asp Tyr Ser Ala Arg Glu Met Pro Glu Ile Thr

            100                 105                 110

Lys Ile Gly Thr Ala Asn Trp Arg Thr Ile Ser Ser Ser Asp Ser Gly
        115                 120                 125

Pro Phe Trp Ala Thr Leu Arg Lys Gly Leu Gln Ser Val Ala Leu Ser
    130                 135                 140

Pro Gln His Leu Ala Ser Gln Thr Ala His Gln Glu Arg Asp Ile Ile
145                 150                 155                 160

Lys Leu Ile Lys Asn Leu Lys Asp Glu Ala Ala Ser Gly Met Val Lys
                165                 170                 175

Pro Leu Asp His Leu Lys Lys Ala Thr Val Arg Leu Ile Ser Arg Leu
            180                 185                 190

Ile Tyr Gly Gln Asp Phe Asp Asp Asp Lys Tyr Val Glu Asp Met His
        195                 200                 205

Asp Val Ile Glu Phe Leu Ile Arg Ile Ser Gly Tyr Ala Gln Leu Ala
    210                 215                 220

Glu Val Phe Tyr Tyr Ala Lys Tyr Leu Pro Gly His Lys Arg Ala Val
225                 230                 235                 240

Thr Gly Ala Glu Glu Ala Lys Arg Arg Val Ile Ala Leu Val Arg Pro
                245                 250                 255

Phe Leu Gln Ser Asn Pro Ala Thr Asn Thr Tyr Leu His Phe Leu Lys
            260                 265                 270

Ser Gln Leu Tyr Pro Glu Glu Val Ile Ile Phe Ala Ile Phe Glu Ala
        275                 280                 285

Tyr Leu Leu Gly Val Asp Ser Thr Ser Ser Thr Thr Ala Trp Ala Leu
    290                 295                 300

Ala Phe Leu Ile Arg Glu Pro Ser Val Gln Glu Lys Leu Tyr Gln Glu
```

```
305                 310                 315                 320

Leu Lys Asn Phe Thr Ala Asn Asn Asn Arg Thr Met Leu Lys Val Glu
                325                 330                 335

Asp Val Asn Lys Leu Pro Tyr Leu Gln Ala Val Val Lys Glu Thr Met
            340                 345                 350

Arg Met Lys Pro Ile Ala Pro Leu Ala Ile Pro His Lys Ala Cys Lys
        355                 360                 365

Asp Thr Ser Leu Met Gly Lys Lys Val Asp Lys Gly Thr Lys Val Met
    370                 375                 380

Val Asn Ile His Ala Leu His His Thr Glu Lys Val Trp Lys Glu Pro
385                 390                 395                 400

Tyr Lys Phe Ile Pro Glu Arg Phe Leu Gln Lys His Asp Lys Ala Met
                405                 410                 415

Glu Gln Ser Leu Leu Pro Phe Ser Ala Gly Met Arg Ile Cys Ala Gly
            420                 425                 430

Met Glu Leu Gly Lys Leu Gln Phe Ser Phe Ser Leu Ala Asn Leu Val
        435                 440                 445

Asn Ala Phe Lys Trp Ser Cys Val Ser Asp Gly Val Leu Pro Asp Met
    450                 455                 460

Ser Asp Leu Leu Gly Phe Val Leu Phe Met Lys Thr Pro Leu Glu Ala
465                 470                 475                 480

Arg Ile Val Pro Arg Leu
                485
```

<210> 16

<211> 554

<212> PRT

<213> Papaver somniferum

<400> 16

```
Met Lys Ser Leu Met Met Asn Lys Leu Leu Phe Leu Gln Arg Ile Thr
1               5                   10                  15

Asp Ser Pro Ser Thr Thr Ile Ile Ser Thr Phe Ile Val Thr Ile Ile
            20                  25                  30

Ser Ile Val Phe Leu Tyr Thr Val Leu Leu Ile Arg Thr Thr Lys Asn
        35                  40                  45

Lys Gln Lys Ile Ala Ala Pro Lys Ala Ser Gly Ala Trp Pro Phe Ile
    50                  55                  60

Gly His Leu Lys Leu Phe Met Lys Gln Asp Thr Gln Phe Tyr Arg Thr
65                  70                  75                  80

Leu Gly Thr Met Ser Asp Lys Tyr Gly Ser Val Phe Thr Leu Arg Leu
                85                  90                  95
```

85 90 95

Gly Asn Gln Ala Ile Leu Val Val Ser Asn Trp Glu Met Val Lys Glu
100 105 110

Cys Phe Thr Thr Asn Asp Lys Ser Phe Ser Asn Arg Pro Ser Thr Leu
115 120 125

Ser Thr Lys Tyr Met Leu Asn Asp Thr Asn Ser Val Val Phe Ser Pro
130 135 140

Tyr Gly Thr Tyr Trp Arg Glu Met Arg Lys Ile Leu Val Gln Lys Leu
145 150 155 160

Leu Ile Ser Asn Gln Arg Ser Glu Ala Leu Lys Asn Leu Lys Thr Lys
165 170 175

Glu Ile Asp Asn Ser Phe Val Lys Leu Asn Asp Leu Cys Asn Asn Asp
180 185 190

Val Ser Gly Gly Gly Thr Lys Val Arg Met Asp Glu Trp Leu Ala Asp
195 200 205

Met Met Phe Asn Ile Ile Ala Arg Ile Thr Phe Gly Tyr Gln Ser Gly
210 215 220

Gly Gly Asp Ala Pro Gly Ala Ser Thr Thr Ser Lys Asn Val Glu Arg
225 230 235 240

Tyr Lys Lys Thr Leu Asp Glu Met Phe Val Val Leu Ala Thr Arg Phe
245 250 255

Ala Val Ser Asp Ile Phe Pro Ser Leu Glu Phe Ile Asp Arg Leu Arg
260 265 270

Gly Leu Val Lys Asp Met Lys Ile Leu Gly Asp Glu Leu Asn Ser Ile
275 280 285

Ala Gly Cys Phe Ile Glu Glu His Arg Gln Lys Arg Arg Glu Ser Leu
290 295 300

Ser Ser Leu Leu Ser Leu Ser Asn Glu Ser Val Gly Asp Glu Gln Asp
305 310 315 320

Phe Ile Asp Val Leu Leu Ser Ile Met Asp Gln Ser Arg Leu Pro Gly
325 330 335

Asp Asp Pro Asp Phe Ile Ile Lys Ile Met Ile Leu Glu Ala Phe Ala
340 345 350

Gly Gly Thr Asp Ser Leu Ser Ala Thr Leu Thr Trp Val Leu Ser Leu
355 360 365

Leu Leu Asn His Pro Asn Val Leu Lys Arg Ala Arg Glu Glu Ile Asp
370 375 380

Arg His Val Glu Asn Gly Lys Gln Val Glu Val Ser Asp Ile Pro Lys
385 390 395 400

```
Leu Gly Tyr Ile Asp Ala Ile Ile Lys Glu Thr Met Arg Leu Tyr Pro
                405                 410                 415

Val Gly Ala Leu Ser Glu Arg Tyr Thr Thr Glu Glu Cys Glu Val Gly
            420                 425                 430

Arg Phe Asn Val Pro Ala Gly Thr Arg Leu Leu Val Asn Ile Trp Lys
        435                 440                 445

Ile His Arg Asp Pro Ser Val Trp Glu Asn Pro Ser Asp Phe Gln Pro
    450                 455                 460

Glu Arg Phe Leu Cys Ser Asp Lys Val Gly Val Asp Leu Tyr Gly Gln
465                 470                 475                 480

Asn Tyr Glu Leu Ile Pro Phe Gly Ala Gly Arg Arg Val Cys Pro Ala
                485                 490                 495

Ile Val Ser Ser Leu Gln Thr Met His Tyr Ala Leu Ala Arg Leu Ile
            500                 505                 510

Gln Gly Tyr Glu Met Lys Ser Ala Ser Leu Asp Gly Lys Val Asn Met
        515                 520                 525

Glu Glu Met Ile Ala Met Ser Cys His Lys Met Ser Pro Leu Glu Val
    530                 535                 540

Ile Ile Ser Pro Arg Glu Pro Arg Arg Ser
545                 550
```

<210> 17

<211> 556

<212> PRT

<213> Papaver somniferum

<400> 17

```
Met Ala Tyr Leu Met Ile Lys Lys Ser Ile Tyr Leu Phe Phe Asp Gln
1               5                   10                  15

Pro Thr Ala Val Gly Thr Leu Ile Leu Ala Phe Leu Leu Thr Leu Ser
            20                  25                  30

Pro Val Ile Ile Tyr Tyr Glu Gln Lys Lys Arg Gly Leu Arg Arg Asn
        35                  40                  45

Arg Thr Ala Ile Thr Thr Thr Pro Leu Pro Glu Ala Ser Gly Ala Trp
    50                  55                  60

Pro Val Ile Gly His Leu Leu Leu Phe Met Asn Glu Asn Asp Leu Asn
65                  70                  75                  80

His Val Thr Leu Gly His Met Ala Asp Lys Tyr Gly Pro Ile Phe Ser
                85                  90                  95

Leu Arg Phe Gly Arg His Arg Thr Leu Val Val Ser Ser Trp Glu Met
            100                 105                 110
```

```
Val Lys Glu Cys Phe Thr Gly Thr Asn Asp Lys Leu Phe Ser Asn Arg
        115                 120                 125

Pro Ser Ser Leu Ala Val Lys Leu Met Phe Tyr Asp Thr Glu Ser Tyr
    130                 135                 140

Gly Phe Ala Pro Tyr Gly Lys Tyr Trp Arg Glu Leu Arg Lys Ile Ser
145                 150                 155                 160

Thr His Lys Leu Leu Ser Asn Gln Gln Leu Glu Lys Phe Lys His Leu
                165                 170                 175

Arg Ile Ser Glu Val Asp Asn Ser Phe Lys Lys Leu His Glu Leu Cys
            180                 185                 190

Ser Asn Asn Lys Gln Gly Gly Asp Thr Thr Tyr Val Ala Ser Leu Val
        195                 200                 205

Arg Met Asp Asp Trp Phe Ala Tyr Leu Thr Phe Asn Val Ile Gly Arg
    210                 215                 220

Ile Val Ser Gly Phe Gln Ser Asn Ala Val Ala Gly Ala Thr Asn Ser
225                 230                 235                 240

Gln Glu Lys Tyr Lys Leu Ala Ile Asp Glu Val Ser Asn Leu Met Ala
                245                 250                 255

Thr Phe Ala Val Ser Asp Val Val Pro Arg Leu Gly Trp Ile Asp Arg
            260                 265                 270

Leu Thr Gly Leu Thr Gly Lys Met Lys Asn Cys Gly Lys Lys Leu Asp
        275                 280                 285

Ala Val Val Gly Asp Ala Val Glu Asp His Arg Gln Lys Lys Leu Lys
    290                 295                 300

Ile Ser Arg Asn Asn Thr Gly Ala Leu Thr Glu His Glu Glu Glu Asp
305                 310                 315                 320

Phe Ile Asp Val Cys Leu Ser Ile Met Glu Gln Ser Gln Ile Pro Gly
                325                 330                 335

Asn His Pro Glu Ile Ser Val Lys Ser Ile Ala Leu Asp Met Leu Ser
            340                 345                 350

Gly Gly Ser Asp Thr Thr Lys Leu Ile Met Thr Trp Thr Leu Ser Leu
        355                 360                 365

Leu Leu Asn His Pro Asp Ile Leu Asp Lys Ala Lys Glu Glu Val Asp
    370                 375                 380

Thr Tyr Phe Gly Lys Lys Lys Ile Ser Asp Asn Thr Pro Val Val Asp
385                 390                 395                 400

Ala Ala Asp Val Pro Asn Leu Val Tyr Ile Gln Ala Ile Ile Lys Glu
                405                 410                 415

Ser Met Arg Leu Tyr Pro Ala Ser Thr Leu Met Glu Arg Met Thr Ser
            420                 425                 430
```

```
Asp Asp Cys Asp Val Gly Gly Phe His Val Pro Ala Gly Thr Arg Leu
        435                 440                 445

Trp Val Asn Val Trp Lys Met Gln Arg Asp Pro Arg Val Trp Lys Asp
    450                 455                 460

Pro Leu Val Phe Leu Pro Glu Arg Phe Leu Ser Asn Asp Lys Gly Met
465                 470                 475                 480

Val Asp Val Lys Gly Gln Asn Tyr Glu Leu Ile Pro Phe Gly Thr Gly
                485                 490                 495

Arg Arg Ile Cys Pro Gly Ala Ser Phe Ala Leu Glu Val Leu His Leu

            500                 505                 510

Val Leu Thr Arg Leu Ile Leu Glu Phe Glu Met Lys Ala Pro Glu Gly
        515                 520                 525

Lys Ile Asp Met Arg Ala Arg Pro Gly Phe Phe His Asn Lys Val Val
    530                 535                 540

Pro Leu Asp Val Gln Leu Thr Pro Arg Thr Leu Asp
545                 550                 555
```

<210> 18

<211> 320

<212> PRT

<213> Papaver somniferum

<400> 18

```
Met Ala Asp Pro Tyr Glu Phe Leu Met Cys Ile His Asn Pro Glu Glu
1               5                   10                  15

Asp Thr Leu Thr Arg Asn Phe Pro Ile Pro Ala Thr Pro Leu Asp Gln
            20                  25                  30

Asn Thr Lys Asp Ile Ser Leu Asn Pro Asp Arg Lys Thr Ser Leu Arg
        35                  40                  45

Ile Phe Arg Pro Pro Thr Lys Glu Pro Pro Val Thr Lys Asn Lys Leu
    50                  55                  60

Leu Pro Ile Ile Ile Tyr Phe His Gly Gly Gly Phe Ile Leu Phe Asn
65                  70                  75                  80

Ala Asp Ser Thr Met Asn His Asp Phe Cys Gln Ser Ile Ala Thr His
                85                  90                  95

Ile Pro Ala Leu Val Val Ser Val Asp Tyr Arg Leu Ala Pro Glu Asn
            100                 105                 110

Arg Leu Pro Ala Ala Tyr Asp Asp Ala Val Asp Ala Leu Asn Trp Val
        115                 120                 125
```

```
Lys Asp Gln Gly Leu Gly Lys Leu Asn Asn Ser Glu Val Trp Leu Lys
    130                 135                 140

Glu Tyr Gly Asp Phe Ser Lys Cys Phe Ile Met Gly Cys Ser Ser Gly
145                 150                 155                 160

Ala Asn Val Ala Tyr His Ala Ser Leu Arg Ala Ile Glu Met Asp Leu

                165                 170                 175

Glu Pro Ala Lys Ile Asn Gly Leu Ile Leu His Cys Pro Phe Phe Gly
            180                 185                 190

Ser Leu Glu Arg Thr Glu Ser Asp Ser Lys Val Ile Asn Asn Gln Asp
        195                 200                 205

Leu Pro Leu Ala Val Arg Asp Val Met Trp Glu Leu Ala Leu Pro Leu
    210                 215                 220

Gly Ser Thr Arg Asp His Val Tyr Cys Asn Pro Asn Ile Asp His Asp
225                 230                 235                 240

Gly Ser Ser Ser Gly Asn Met Val Gly Leu Ile Glu Arg Cys Phe Val
                245                 250                 255

Val Gly Phe Tyr Gly Asp Pro Leu Ile Asp Arg Gln Ile Gln Leu Val
            260                 265                 270

Lys Met Leu Glu Glu Lys Gly Val Lys Val Glu Thr Trp Ile Glu Gln
        275                 280                 285

Gly Gly Tyr His Gly Val Leu Cys Phe Asp Pro Met Ile Arg Glu Thr
    290                 295                 300

Phe Leu Glu Lys Leu Lys His Phe Ile Leu Asn Asp Glu Phe Ile Tyr
305                 310                 315                 320
```

<210> 19

<211> 348

<212> PRT

<213> Papaver somniferum

<400> 19

```
Met His Gly Gln Lys Asn Ile Ser Glu Arg Tyr Gln Lys Phe Lys Glu
1               5                   10                  15

Met Glu Gly Thr Gly Lys Ile Val Cys Val Thr Gly Gly Ala Gly Tyr
            20                  25                  30

Leu Ala Ser Trp Leu Ile Met Arg Leu Leu Glu Arg Gly Tyr Ser Val
        35                  40                  45

Arg Thr Thr Val Arg Ser Asp Pro Lys Phe Arg Glu Asp Val Ser His
    50                  55                  60

Leu Lys Ala Leu Pro Glu Ala Thr Glu Lys Leu Gln Ile Phe Glu Ala
```

```
65                  70                  75                  80

Asp Leu Glu Asn Pro Glu Ser Phe Asp Asp Ala Ile Asn Gly Cys Val
                85                  90                  95

Gly Val Phe Leu Val Ala Gln Gly Met Asn Phe Ala Glu Glu Tyr Thr
            100                 105                 110

Leu Glu Lys Ile Ile Lys Thr Cys Val Glu Gly Thr Leu Arg Ile Leu
        115                 120                 125

Gln Ser Cys Leu Lys Ser Lys Thr Val Lys Lys Val Val Tyr Thr Ser
    130                 135                 140

Ser Ala Asp Ala Ala Met Met Ile Ser Asn Leu Lys Ala Val Lys Glu
145                 150                 155                 160

Ile Asp Glu Thr Ile Trp Ser Glu Val Asp Asn Phe Ile Ser Lys Pro
                165                 170                 175

Glu Gln Val Ile Pro Gly Leu Pro Ser Tyr Val Val Ser Lys Val Leu
            180                 185                 190

Thr Glu Arg Ala Cys Leu Lys Phe Ser Glu Glu His Gly Leu Asp Val
        195                 200                 205

Val Thr Ile Leu Pro Pro Leu Val Val Gly Pro Phe Ile Thr Pro His
    210                 215                 220

Pro Pro Pro Ser Val Ser Ile Ala Leu Ser Ile Ile Ser Gly Asp Val
225                 230                 235                 240

Ser Met Met Leu Gly Val Arg Leu Glu Asn Ala Val His Ile Asp Asp
                245                 250                 255

Val Ala Leu Ala His Ile Phe Val Phe Glu Cys Glu Lys Ala Lys Gly
            260                 265                 270

Arg His Ile Cys Ser Ser Val Asp Phe Pro Met His Asp Leu Pro Lys
        275                 280                 285

Phe Ile Ser Glu Asn Tyr Pro Glu Phe Asn Val Pro Thr Asp Leu Leu
    290                 295                 300

Lys Asp Ile Glu Glu Gln Glu Pro Val His Leu Ser Ser Asp Lys Leu
305                 310                 315                 320

Leu Ser Met Gly Phe Gln Phe Lys Tyr Asp Phe Ala Glu Ile Phe Gly
                325                 330                 335

Asp Ala Ile Arg Cys Ala Lys Glu Lys Gly Phe Leu
            340                 345
```

<210> 20

<211> 471

<212> PRT

<213> Papaver somniferum

<400> 20

```
Met Ala Thr Met Ser Ser Ala Ala Val Glu Val Ile Ser Lys Glu Thr
1               5                  10                  15

Ile Lys Pro Arg Asn Pro Thr Pro Tyr Gln Leu Arg Asn Tyr Asn Met
            20                  25                  30

Ser Leu Leu Asp Gln Tyr Ser Ser Leu Val Tyr Val Pro Ile Ile Leu
        35                  40                  45

Phe Tyr Pro Ala Ala Ser Asp Ala Asn Ser Thr Gly Ser Lys His His
    50                  55                  60

Asp Asp Leu His Leu Leu Lys Arg Ser Leu Ser Glu Thr Leu Val His
65                  70                  75                  80

Phe Tyr Pro Met Ala Gly Arg Met Lys Asp Asn Met Thr Val Asp Cys
                85                  90                  95

Asn Asp Glu Gly Ile Asp Phe Phe Glu Val Arg Ile Lys Gly Arg Met
            100                 105                 110

Cys Asp Phe Met Met Lys Ser Asp Ala His Leu Ser Leu Leu Leu Pro
        115                 120                 125

Ser Glu Val Ala Ser Thr Asn Phe Val Lys Glu Ala Gln Val Ile Val
    130                 135                 140

Gln Val Asn Met Phe Asp Cys Gly Gly Thr Ala Ile Cys Phe Cys Ile
145                 150                 155                 160

Ser Asn Lys Ile Ala Asp Ala Cys Thr Met Ile Thr Phe Ile Arg Ser
                165                 170                 175

Leu Ala Gly Thr Thr Asn Ile Ala Arg Arg Gly Ser Ser Ile Ala Ala
            180                 185                 190

Pro Thr Thr Asn Gln Asn Leu Val Pro Ser Phe Asp Ser Thr Ser Leu
        195                 200                 205

Phe Pro Pro Ser Glu Gln Leu Ala Ser Gln Val Ser Tyr Pro Thr Gln
    210                 215                 220

Asp Ser Thr Ser Val Asp Lys Leu Val Ser Lys Arg Phe Val Phe Asp
225                 230                 235                 240

Ala Ala Lys Ile Thr Ser Ala Arg Glu Lys Leu Gln Ser Leu Met His
                245                 250                 255

Asp Lys Tyr Lys Cys His Arg Pro Thr Arg Val Glu Val Val Ser Ala
            260                 265                 270

Leu Ile Trp Lys Ser Ala Val Lys Ser Ala Pro Pro Gly Ser Ile Ser
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Thr Val Thr His Ala Met Asn Phe Arg Lys Lys Met Asp Pro Pro Leu
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Pro Ala Thr Thr Ala Thr Thr Thr Asn Pro Ala Thr Lys Lys Val Ser
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Ser Thr Ser Asn Glu Glu Gln Val Ala Leu Asp Glu Leu Ser Asp Phe
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Val Ala Leu Leu Arg Arg Glu Ile Asp Lys Val Lys Gly Asp Lys Gly
        355                 360                 365

Cys Met Glu Lys Ile Ile Gln Lys Phe Ile Tyr Gly His Asp Ala Ser
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Val Ala Lys Asp Ser Asp Val Glu Asp Lys Val Thr Ala Leu Phe Met
385                 390                 395                 400

Thr Ser Trp Cys Lys Phe Gly Phe Tyr Glu Ala Asp Phe Gly Trp Gly
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Thr Pro Val Trp Val Thr Thr Val Pro Leu Ile Glu Pro Lys Tyr Lys
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Asn Met Val Phe Met Asn Asp Met Lys Cys Gly Glu Gly Ile Glu Val
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ggtgctgccc cgtcttattc atgggtttga tatgaaagca gcaagtgcca atgggaaagt 180

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PATENTKRAV

1. Et isoleret nukleinsyremolekyle omfattende eller bestående af en nukleotidsekvens udvalgt fra gruppen:

i) en nukleotidsekvens som angivet i SEQ ID-NR.: 8;

ii) en nukleotidsekvens, hvori den førnævnte sekvens er degenereret som et resultat af den genetiske kode til nukleotidsekvensen, der er defineret i (i);

iii) et nukleinsyremolekyle, der har mindst 80 % sekvensidentitet med SEQ ID-NR.: 8 og koder et carboxylesterasepolypeptid med aktivitet;

iv) en nukleotidsekvens, der koder for et polypeptid, omfattende en aminosyresekvens, som angivet i SEQ ID-NR.: 18, eller en nukleotidsekvens, der koder for et polypeptid, som har mindst 75 % aminosyresekvensidentitet hen over den fulde længde af aminosyresekvensen, der er angivet i SEQ ID-NR.: 18, hvori det førnævnte polypeptid har carboxylesteraseaktivitet.

2. Et isoleret polypeptid udvalgt fra gruppen bestående af:

i) et polypeptid omfattende eller bestående af en aminosyresekvens, som repræsenteret i SEQ ID-NR.:18;

ii) et modificeret polypeptid omfattende eller bestående af en modificeret aminosyresekvens, hvori det førnævnte polypeptid er modificeret ved tilføjelse, sletning eller substitution af mindst én aminosyrerest af sekvensen, der er præsenteret i SEQ ID NR.: 18, og som har mindst 75 % sekvensidentitet med den fulde længde af aminosyresekvensen i SEQ
ID-NR.: 18 og har carboxylesteraseaktivitet;

iii) et modificeret polypeptid omfattende eller bestående af en aminosyresekvens, der er mindst 75 % identisk med den fulde længde af aminosyresekvensen i SEQ ID-NR.: 18, og som koder for et polypeptid med carboxylesteraseaktivitet.

3. Isoleret nukleinsyremolekyle ifølge patentkrav 1, hvori nukleinsyremolekylet yderligere omfatter en eller flere nukleotidsekvenser, der er valgt fra gruppen bestående af:

i) en nukleotidsekvens som repræsenteret ved sekvensen i SEQ ID-NR.: 1, 2, 3, 4, 5, 6, 7, 9 eller 10;

ii) en nukleotidsekvens, hvori den førnævnte sekvens er degenereret som et resultat af den genetiske kode til nukleotidsekvensen, der er defineret i (i);

iii) et nukleinsyremolekyle, der er mindst 80 % identisk med sekvensen i SEQ ID-NR.: 1, 2, 3, 4, 5, 6, 7, 9 eller 10, hvori det førnævnte nukleinsyremolekyle koder polypeptider, der er involveret i biosyntesen af noscapin af P. *somniferum;*

iv) en nukleotidsekvens, der koder et polypeptid, omfattende en aminosyresekvens, som repræsenteret i SEQ ID-NR.: 11, 12, 13, 14, 15, 16, 17, 19 eller 20; og

v) en nukleotidsekvens, der koder et polypeptid, omfattende en aminosyresekvens, hvori den førnævnte aminosyresekvens modificeres ved tilføjelse, sletning eller substitution af mindst én aminosyrerest, som repræsenteret i iv) ovenfor, og som er mindst 75 % identisk med aminosyresekvenserne, der er repræsenteret i SEQ ID-NR.: 11, 12, 13, 14, 15, 16, 17, 19 eller 20, der er involveret i biosyntesen af noscapin.

4. Nukleinsyremolekylet ifølge et af patentkravene 1 eller 3, hvori det førnævnte nukleinsyremolekyle inkluderer SED ID-NR.: 9 yderligere omfatter en eller flere nukleotidsekvenser, der er valgt fra gruppen bestående af: SEQ ID-NR.: 1, 2, 3, 4, 5, 6, 7 og 10.

5. Nukleinsyremolekylet ifølge patentkravene 1 eller 3, hvori det førnævnte nukleinsyremolekyle inkluderer 3, 4, 5, 6, 7, 8, 9 eller 10 nukleotidsekvenser, der er valgt fra gruppen bestående af: SEQ ID-NR.: 1, 2, 3, 4, 5, 6, 7, 8, 9 og 10.

6. En vektor omfattende et nukleinsyremolekyle ifølge et hvilket som helst af patentkravene 1, 3, 4 eller 5.

7. En transgenisk celle, omfattende en vektor ifølge patentkrav 6.

8. Den transgeniske celle ifølge patentkrav 7, hvori den førnævnte celle er en plantecelle.

9. Den transgeniske celle ifølge patentkrav 7, hvori den førnævnte celle er en mikrobiel celle.

10. Et nukleinsyremolekyle, omfattende en transskriptionskassette, hvori kassetten omfatter nukleotidsekvensen af SEQ ID-NR.: 8 og er tilpasset til ekspression ved tilvejebringelse af mindst én promotor, der er operativt forbundet til førnævnte nukleotidsekvens, således at både sanse- og anti-sansemolekyler transskriberes fra den førnævnte kassette.

11. En fremgangsmåde til modifikation af papaveroxin og desmethylpapaveroxin omfattende:

i) at tilvejebringe en transgen papaverplantecelle ifølge 8;

ii) at dyrke den førnævnte plantecelle til at producere en transgenisk papaverplante; og eventuelt

iii) høst af den førnævnte papavertransgenplante eller en del deraf.

12. En fremgangsmåde til modifikation af papaveroxin eller desmethylpapaveroxin omfattende:

i) at tilvejebringe en transgen mikrobiel celle ifølge patentkrav 9, der udtrykker et eller flere nukleinsyremolekyler, der koder for et polypeptid med carboxylesteraseaktivitet og papaveroxin eller desmethylpapaveroxin;

ii) dyrkning af den mikrobielle celle under forhold, der modificerer papaveroxin eller desmethylpapaveroxin og eventuelt

iii) isolering af narkotinhemiacetat eller narcotolinhemiacetat fra den mikrobielle celle eller cellekultur.

13. Anvendelsen af et gen kodet af nukleinsyremolekylet af SEQ ID-NR.: 8 eller et nukleinsyremolekyle, der er mindst 80 % identisk med SEQ ID-NR.: 8 og koder et polypeptid med carboxylesteraseaktivitet som et middel til at identificere et sted, hvori det førnævnte sted er forbundet med ændret ekspression eller aktivitet af carboxylesterase.

14. En *P. somniferum*-plante, der har ændret udtrykket af et carboxylesterasepolypeptid ifølge patentkrav 2, som kan opnås ved hjælp af en fremgangsmåde, omfattende trinnene med:

i) mutagenese af vildtypefrø fra en *P. somniferum*-plante, der udtrykker det førnævnte polypeptid;

ii) dyrkning af frøet i i) til fremstilling af første og efterfølgende generationer af planter;

iii) at få frø fra første generation af planter og efterfølgende generationer af planter;

iv) at bestemme, om frøet fra førnævnte første og efterfølgende generation af planter har ændret nukleotidsekvens og/eller ændret udtryk af det førnævnte carboxylesterasepolypeptid; og

v) at opnå frø eller planter med en ændret nukleotidsekvens, der koder for et ændret carboxylesterasepolypeptid, hvori enzymaktiviteten i modifikationen af papaveroxin eller desmethylpapaveroxin ændres sammenlignet med en vildtype *P. somniferum*-

plante.

15. En fremgangsmåde til analyse af en plante, omfattende:

i) opnåelse af en prøve fra en plante ifølge patentkrav 14 og analyse af nukleinsyresekvensen af et nukleinsyremolekyle udvalgt fra gruppen bestående af:

a) et nukleinsyremolekyle omfattende en sekvens i SEQ ID-NR.: 8; og

b) et nukleinsyremolekyle, der har mindst 80 % sekvensidentitet med SEQ ID-NR.: 8 og koder et carboxylesterasepolypeptid med aktivitet;

ii) sammenligning af nukleotidsekvensen af nukleinsyremolekylet i den førnævnte prøve med en nukleotidsekvens af et nukleinsyremolekyle af den originale vildtypeplante; og eventuelt hvori nukleinsyremolekylet analyseres ved hjælp af en fremgangsmåde omfattende trinnene med:

iii) at ekstrahere nukleinsyre fra de førnævnte, muterede planter;

iv) amplifikation af en del af det førnævnte nukleinsyremolekyle ved en polymerasekædereaktion;

v) at danne et præparat omfattende den amplificerede nukleinsyre og nukleinsyre, der er ekstraheret fra vildtypefrø til at danne heteroduplexnukleinsyre;

vi) at inkubere det førnævnte præparat med en enkeltstrenget nuklease, der skærer i et område med heteroduplexnukleinsyre for at identificere uoverensstemmelsen i førnævnte heteroduplex;

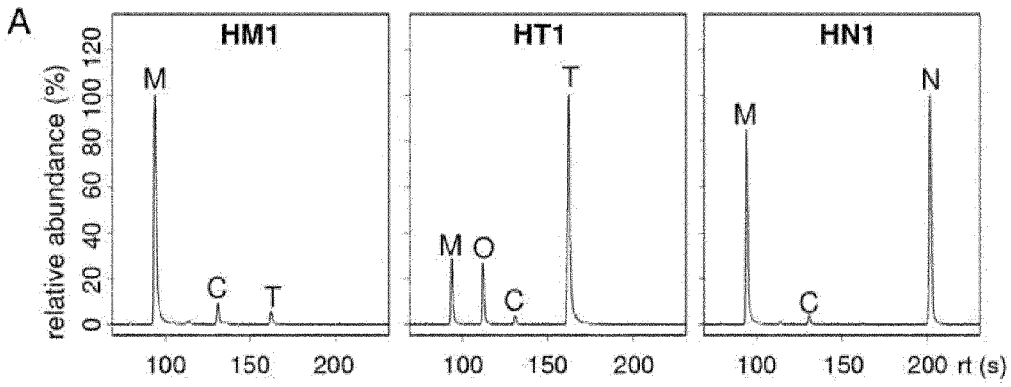
vii) bestemmelse af stedet for uoverensstemmelse i det førnævnte nukleinsyreheteroduplex; og

viii) at opnå en plante med en mutation i nukleinsyren, der koder det førnævnte carboxylesterasepolypeptid

16. En vektor omfattende et nukleinsyremolekyle ifølge patentkrav 1.

DRAWINGS

Figure 1



B

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	Stem	Capsule	Stem	Capsule	Stem	Capsule	HM1	HT1	HN1
PSMT1							-	-	+
PSMT2							-	-	+
PSMT3							-	-	+
CYP82X1							-	-	+
CYP82X2							-	-	+
CYP82Y1							-	-	+
CYP719A21							-	-	+
PSAT1							-	-	+
PSSDR1							-	-	+
PSCXE1							-	-	+
BBE							+	+	+
TNMT							+	+	+
SalR							+	+	+
SalAT							+	+	+
T6ODM							+	+	+

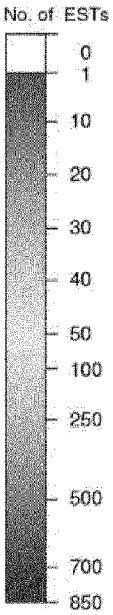


Figure 2

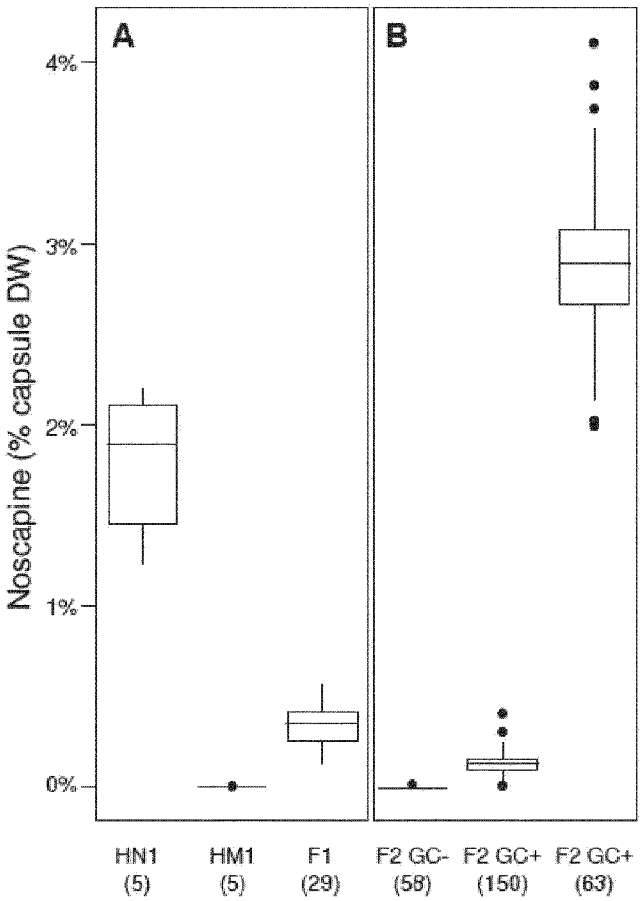

A
B
4%
3%
2%
1%
0%
Noscapine (% capsule DW)
HN1 (5)
HM1 (5)
F1 (29)
F2 GC- (58)
F2 GC+ (150)
F2 GC+ (63)

Figure 3

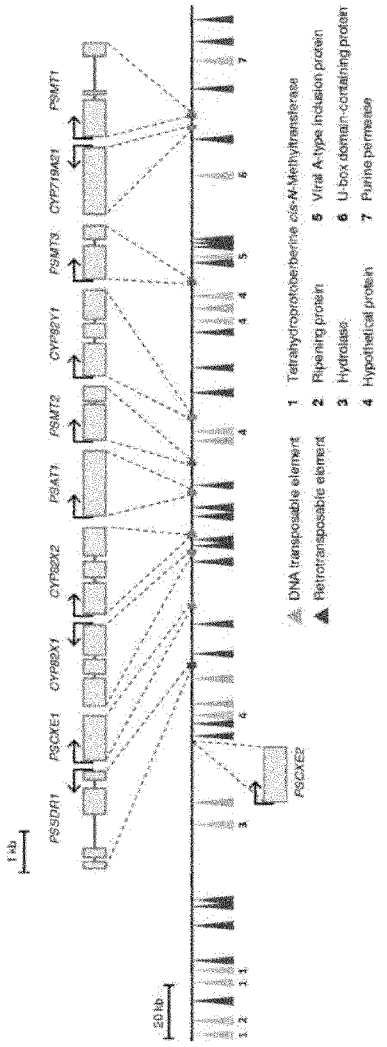
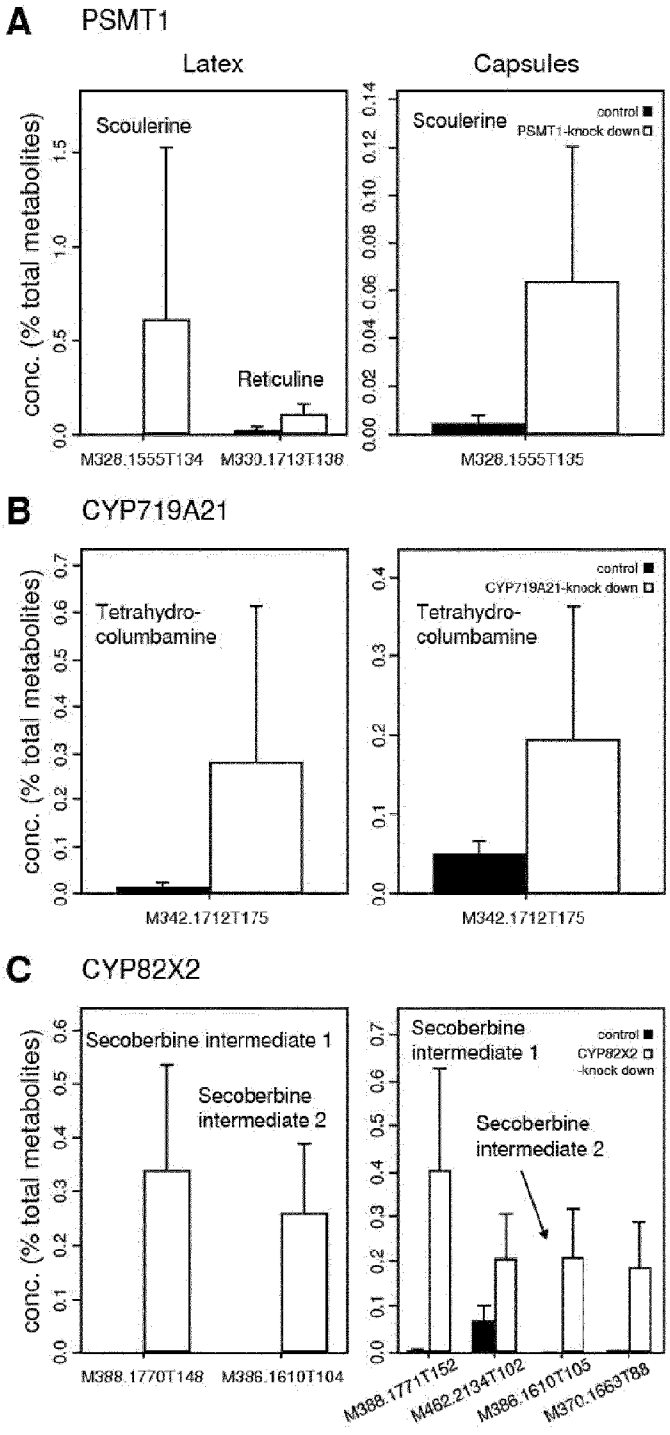
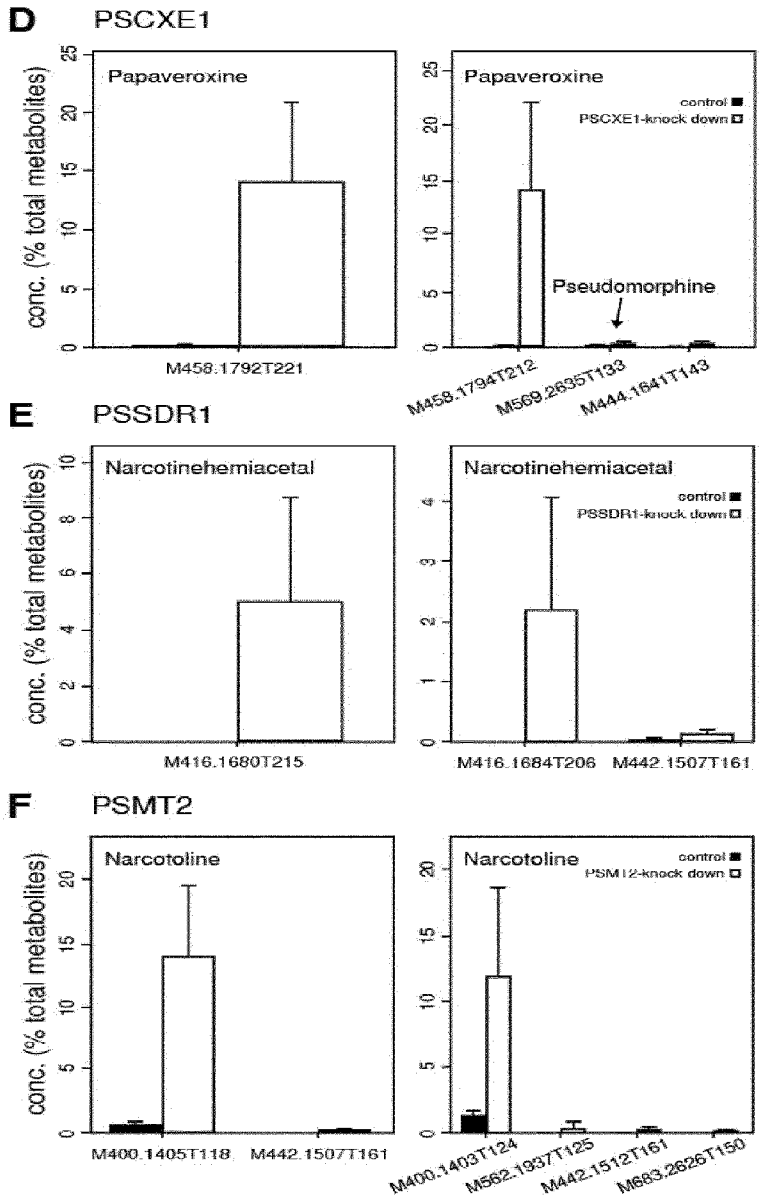

PSIDR1
PSCXE1
CYP82X1
CYP82X2
PSAT1
PSMT2
CYP82Y1
PSMT3
CYP71AK21
PSMT1
PSCXE2
1 kb
20 kb
1 2 3 4 4 4 4 5 6 7
DNA transposable element
Retrotransposable element
1 Tetrahydroprotoberberine cis-N-Methyltransferase
2 Ripening protein
3 Hydrolase
4 Hypothetical protein
5 Viral A-type inclusion protein
6 U-box domain-containing protein
7 Purine permease

Figure 4


A
PSMT1
Latex
Capsules
conc. (% total metabolites)
Scoulerine
Reticuline
control
PSMT1-knock down
M328.1555T134
M330.1713T138
M328.1555T135
B
CYP719A21
Tetrahydro-columbamine
CYP719A21-knock down
M342.1712T175
C
CYP82X2
Secoberbine intermediate 1
Secoberbine intermediate 2
CYP82X2 -knock down
M388.1770T148
M386.1610T104
M388.1771T152
M462.2134T102
M386.1610T105
M370.1663T88


D
PSCXE1
Papaveroxine
conc. (% total metabolites)
M458.1792T221
Papaveroxine
control
PSCXE1-knock down
Pseudomorphine
M458.1794T212
M569.2635T133
M444.1641T143
E
PSSDR1
Narcotinehemiacetal
conc. (% total metabolites)
M416.1680T215
Narcotinehemiacetal
control
PSSDR1-knock down
M416.1684T206
M442.1507T161
F
PSMT2
Narcotoline
conc. (% total metabolites)
M400.1405T118
M442.1507T161
Narcotoline
control
PSMT2-knock down
M400.1403T124
M562.1937T125
M442.1512T161
M683.2625T150

G

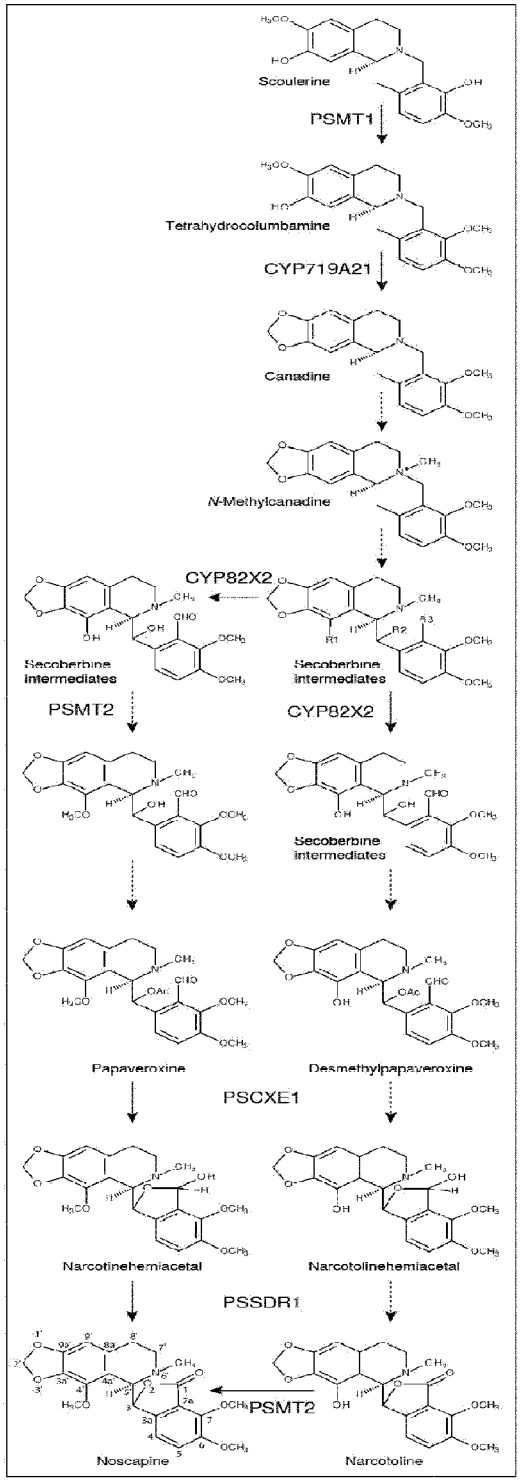

Scoulerine
PSMT1
Tetrahydrocolumbamine
CYP719A21
Canadine
N-Methylcanadine
CYP82X2
Secoberbine intermediates
Secoberbine intermediates
PSMT2
CYP82X2
Secoberbine intermediates
Papaveroxine
Desmethylpapaveroxine
PSCXE1
Narcotinehemiacetal
Narcotolinehemiacetal
PSSDR1
PSMT2
Noscapine
Narcotoline

Figure 5

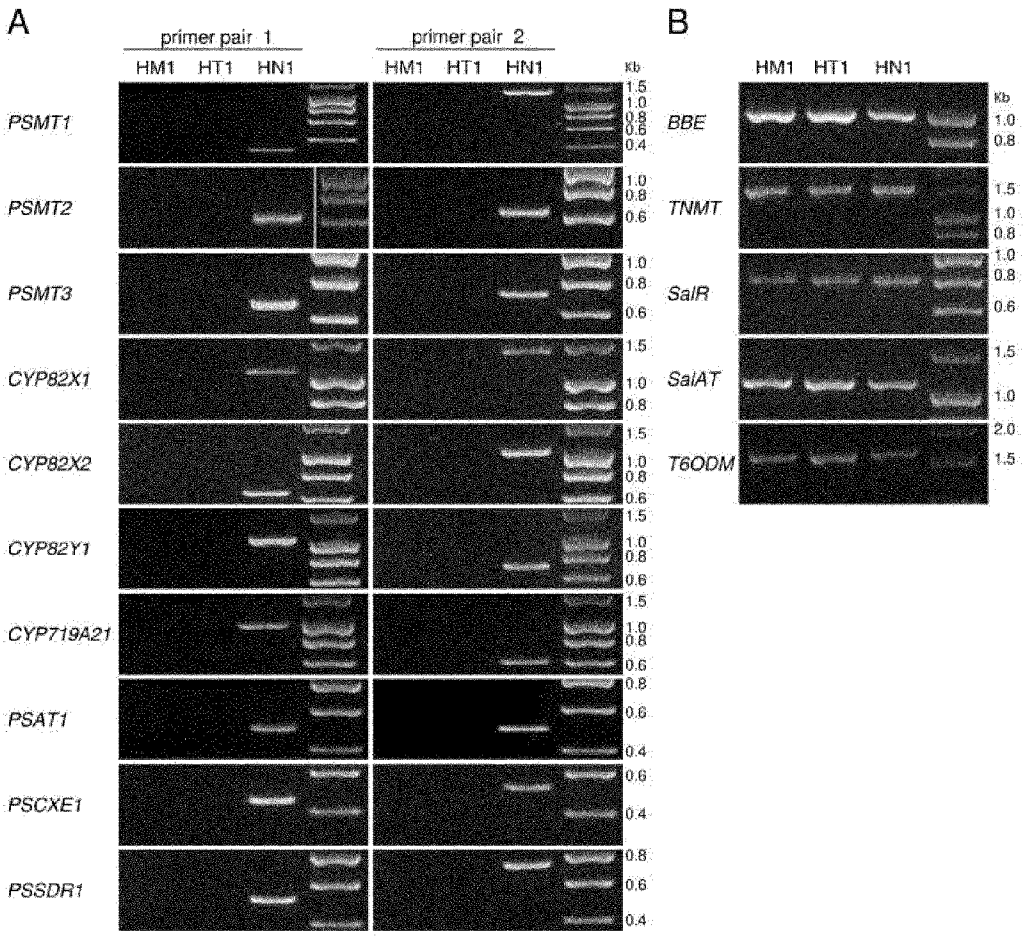
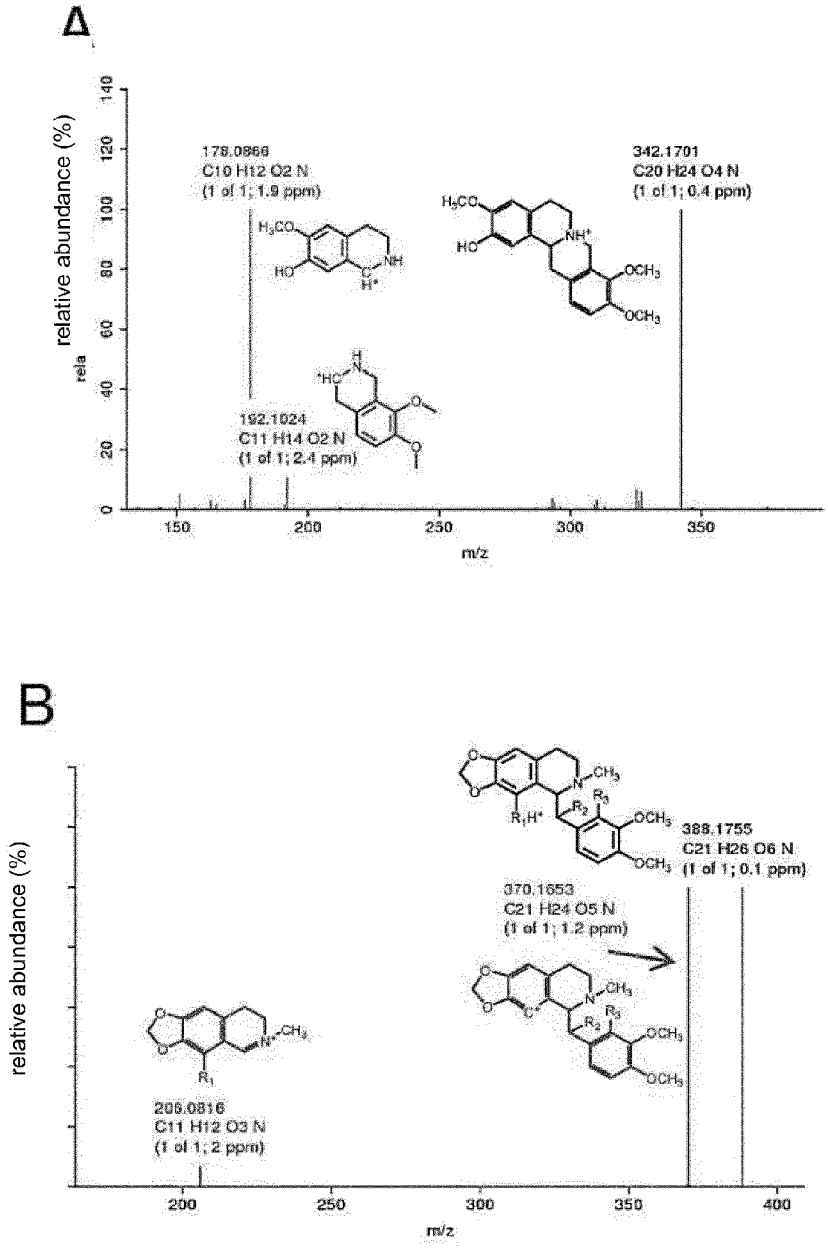
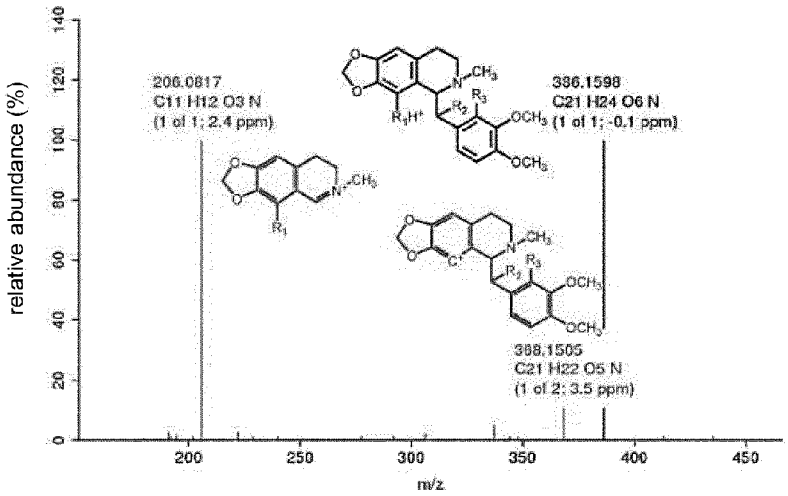

A
primer pair 1
primer pair 2
HM1 HT1 HN1
HM1 HT1 HN1
Kb
PSMT1
PSMT2
PSMT3
CYP82X1
CYP82X2
CYP82Y1
CYP719A21
PSAT1
PSCXE1
PSSDR1
B
HM1 HT1 HN1
Kb
BBE
TNMT
SalR
SalAT
T6ODM

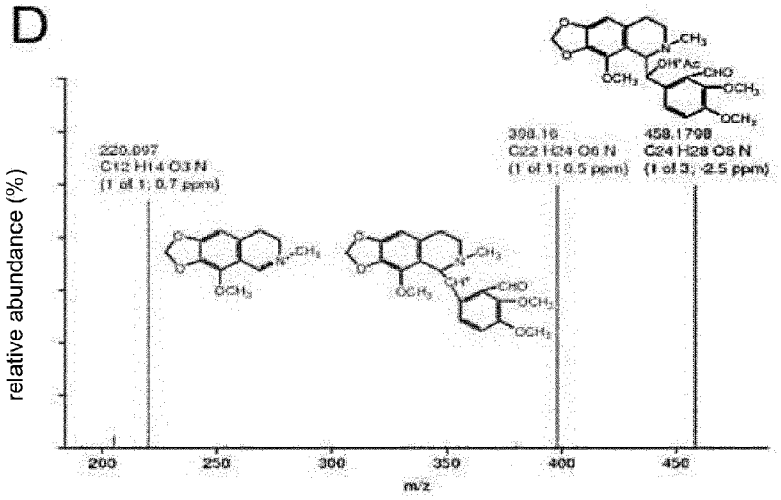
Figure 6


A
relative abundance (%)
rela
178.0866
C10 H12 O2 N
(1 of 1; 1.9 ppm)
342.1701
C20 H24 O4 N
(1 of 1; 0.4 ppm)
192.1024
C11 H14 O2 N
(1 of 1; 2.4 ppm)
m/z
B
relative abundance (%)
388.1755
C21 H26 O6 N
(1 of 1; 0.1 ppm)
370.1653
C21 H24 O5 N
(1 of 1; 1.2 ppm)
206.0816
C11 H12 O3 N
(1 of 1; 2 ppm)
m/z

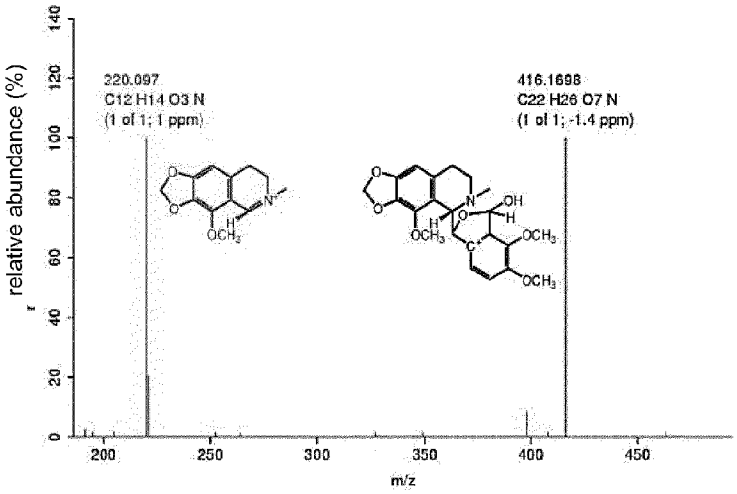
C


relative abundance (%)
206.0817
C11 H12 O3 N
(1 of 1; 2.4 ppm)
386.1598
C21 H24 O6 N
(1 of 1; -0.1 ppm)
388.1505
C21 H22 O5 N
(1 of 2; 3.5 ppm)
m/z

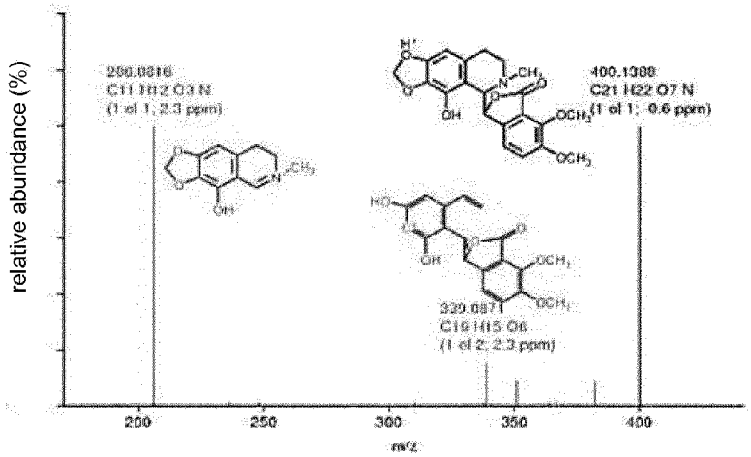
D


relative abundance (%)
220.097
C12 H14 O3 N
(1 of 1; 0.7 ppm)
398.16
C22 H24 O6 N
(1 of 1; 0.5 ppm)
458.1798
C24 H28 O8 N
(1 of 3; -2.5 ppm)
m/z

E


relative abundance (%)
320.097
C12 H14 O3 N
(1 of 1; 1 ppm)
416.1698
C22 H26 O7 N
(1 of 1; -1.4 ppm)
m/z

F


relative abundance (%)
290.0816
C11 H12 O3 N
(1 of 1; 2.3 ppm)
400.1388
C21 H22 O7 N
(1 of 1; 0.6 ppm)
320.0871
C19 H15 O6
(1 of 2; 2.3 ppm)
m/z

SEQ ID NO: 1 ***PSMT1***

Start	End	Feature
1	66	5′ untranslated region
67	877	exon 1
878	993	intron 1
994	1058	exon 2
1059	1841	intron 2
1842	2138	exon 3
2139	2306	3′ untranslated region

> PSMT1

CACACCAAACTTGATCATTGTCATAAAAAACAGTCCTAATTGTCATCAATCAAAAACAGTCCTAACATGGCTACC
AATGGCGAAATTTTCAATACCTATGGTCATAATCATCAATCAGCCACAGTCACTAAAATCACTGCTTCTAATGAA
AGCAGCAATGGTGTCTGTTATCTTTCAGAAACGGCTAACTTGGGGAAGTTAATATGCATTCCAATGGCACTAAGA
GCTGCGATGGAGCTAAATGTGTTCCAACTTATCTCAAAGTTCGGAACTGACGCAAAAGTTTCGGCTTCTGAAATT
GCCTCTAAAATGCCAAACGCGAAGAATAATCCTGAAGCAGCTATGTATTTGGATAGAATTCTTCGACTGCTCGGG
GCAAGTTCTATTCTTTCTGTTTCTACTACAAAAAAATCAATCAACAGAGGAGGAGATGATGTAGTAGTACATGAG
AAGCTTTATGGGTTAACAAATTCGTCGTGTTGTTTGGTCCCTCGACAAGAAGACGGGGTGTCATTAGTCGAAGAA
TTGCTATTCACATCTGACAAGGTTGTTGTGGATAGTTTTTTCAAACTGAAATGTGTGGTGGAAGAAAAAGACAGT
GTGCCATTTGAGGTTGCTCATGGTGCTAAAATCTTTGAGTATGCTGCTACAGAACCAAGAATGAATCAAGTATTT
AACGATGGAATGGCAGTTTTCTCTATTGTTGTTTTTGAGGCTGTTTTTAGAGTTTACGATGGATTTCTTGATATG
AAAGAATTGTTAGATGTTGGTGGTGGTATTGGTACTTCGGTTAGTAAGATTGTTGCTAAATACCCTTTGATTCGC
GGTGTCAACTTCGACTTGCCTCATGTTATTTCTGTTGCCCCTCAATACCCAGGTATACCTTCTTCTTCTTTTTTC
TGAAAAGAACGGGTTCGAATTTTTACAGAATTTTTTTTCTCATTCGATACTCAAGCAACTCTATTAAAGTATACT
GTGTAATAATGCATGCAGGTGTAGAGCATGTTGCAGGAGATATGTTCGAGGAAGTCCCAAAGGGTCAAAACATGT
TGCTAAAAGTAAGCTAACCATACTCAATTTTCTTAATAATTAGGAAAATTGCAAAAACCGTCACAATATTATAAA
GGCATCTGAAGTGCCATCACTCAGATACCGATGCTATGTACTCTATACATTGACAAAATTCCATGGTATCAAGTC
TCAACCTGCCGGTTATAATAATTTTTTTCAGGCTTTCTTTAAAAGAAATTATTTTGAATGGTAAAAATCATCATT
ATATTGGAGAAAAGTGCAGATCTTGCTACATTAAAATTTATAATATAATAAAACATTTGTTTATGGTTGTTTGAA
AAAAAAAATCTCATTGTTAATGCATCTTTCTAAGTTAATGGTGATTAATGGTGAATAATATGATATCTTATTACC
GTCTTGACACTTTTTTTTTTTGTCGTAGACAAAATATTTCCAACTTTTCTATATTAATAAAATCAGAAATATTTCA
TTTATATGAATATTAAAATAAGAAGGTGCATGAGTAATATTCCAAATTTCTTAAAGCGTTTTTTATAGCAGTACG
GCGTTTTCTCAAATCTTATTAACCCATAATTAAAGGGTTTCCGTAAATTAAATTGAGGGATATCAAAACAAAAAC
AAAAAATAGGGTTATTTTGCAGTAAAATCAATAACCCCTTATCATATGAAAAGGATAACTTAGTCTACCCCAATT
TGGAGAGATATGGGCAATTATTGTATTACTAGTTCGTTTGAGCATTGATAATATTTTTCATTAGATTTATACTCA
ATAAAATATATGAACTATATTGATAAAGATTAATAATGCAGTGGGTACTGCACGATTGGGGTGATGAACGATGTG
TGAAGCTGTTAAAGAATTGTTGGAACTCATTACCTGTGGGTGGAAAAGTTTTGATAATCGAGTTTGTTCTCCCGA
ATGAACTTGGTAACAATGCTGAATCATTCAATGCGTTGATTCCCGATTTACTCCTGATGGCTCTGAATCCAGGCG
GTAAAGAGCGAACGATTTCCGAATACGATGATTTAGGCAAAGCAGCTGGATTCATAAAAACTATACCTATCCCTA
TCTCCAATGGTCTTCATGTCATTGAGTTTCACAAATGAATGGTTATTGAGTGCTTTGGTAATTAAACTACCAAGA
TAACTACATCCATTTCATGCATTTGCTTTTTTTTTTTCTTTTTTTTCTTTTTTTTTTCTTTTTGTTTTGTATTCCA
GGTGTGAACTAGTTAGTGTGTTGAGTGGACAAAAGTAAGTAATCGTATTTTGTGTT

SEQ ID NO: 2 ***PSMT2***

Start	End	Feature
1	74	5′ untranslated region
75	851	exon 1
852	938	intron
939	1232	exon 2
1233	1440	3′ untranslated region

> PSMT2

GAATCAGAAACTTTCTTCTAAAATCTTTCAATACCAGTACTGTTAGTTTCCGATAAGAGCCACACTAATCCATTA
TGGAAATTCATTTAGAAAGCCAAGAACAAGAAATGAAATATCAATCTCAAATCTGGAACCAAATATGTGGCACTG
TTGATACCTCTGTTCTGAGATGTGCAATTCAATTAGGTATATTTGATGCCATTCATAACTCTGGCAAACCAATGA
TTACCTTAACCGAATTATCAAGCATTGTTTCATCACCCTCTTCATCTTCAATCGAACCCTGCAACTTGTATAGAT
TAGTGAGATACTTATCCCAAATGGATCTCATTAGTATCGGAGAATGTTTGAATGAAGCAACTGTTTCATTAACAG
GCACATCCAAGTTACTACTTAGAAACCAAGAAAAGAGTTTAATTGATTGGGTATTGGCAATTTCTTGCGAAATGA
TGGTTGTTGTTTGGCACGAACTAAGTAGCTCTGTTTCAACTCCTGCGGATGAGCCTCCAATCTTCCAGAAGGTTC
ATGGTAAAAATGCTTTAGAATTAGCAGGGGAATTTCCAGAATGGAATGATCTGATCAACAATGCTATGACTAGTG
ATACTAGTGTAACTAAGCCAGCGCTAATACAAGGATGTGGCAAAATCCTGAACGGAGTTACATCGTTAATTGATG
TCGGTGGTGGTCACGGTGCCACTATGGCCTACATAGTTGAAGCTTTTCCTCACATAAAAGGTGCGGTAATCGATT
TACCACATGTTGTTGAAGCCGCTCCGGAGCGTCCAGGTGTTGAGTTCATCAGCGGTGATATATTCAAGTCCATTT
CTAACGCTGATGCTGTGTTGTTGAAGGTATGTAAAGAGTAGCTAACCTTAGTGCGTCTAATTTATTCCACAAATT
TTTCTGATGCATTTTATTCTTATTTTTGGTTTTTGCAGTATGTCCTGCACAATTGGGAAGATACGGAATGTGTGA
ATTTACTGAAGAGATGTAAGGAAGCAGTTCCGGCAGACAAAGGAAAAGTGATCATAATGGATTTAGTAATAGACG
ACGATGATAACAGTATTTTAACGCAGGCAAAGTTGAGCCTTGATCTCACTGTGATGAACCATGGAGGAGGTAGAG
AAAGGACTAAAGAAGATTGGAGAAATCTAATTGAGATGTCTGGATTTAGTAGGCATGAAATAATTCCAATATCTG
CCATGCCATCAATTATTGTTGCTTATCCTTAGTTAAGTCACCCGCATGTTTACTTGAACGGGAATAAGTTGGGGG
CGTGTTGAATCTGTTAACATCGCAATTGTGCCTTTACTTTATGCATTCTCATTCCGGTAGAAACTGTTTGGGGCA
TTCGGATTCTGCTGAGCCCTTTTATGTATGTTTGTTTGTTGGTTGGTTGGTTTTCAAGTAACTGAAGTTTCTTCT
CTGTTTTCAAGGCAT

SEQ ID NO: 3 ***PSMT3***

Start	End	Feature
1	120	5′ untranslated region
121	846	exon 1
847	994	intron
995	1288	exon 2
1289	1436	3′ untranslated region

> PSMT3

AAGTTGCAGGTAGGGTTATGAGCAAGCTCAATTATCTCTCCTATAAAAGCTAACATTAGAAAAACTAATAAGCAC
ACAAACCGTAAAAGTTCTGAAGATAGACAAAACAAGAGAAAAAAGATGGAAGTAGTAAGTAAGATTGATCAAGAA
AACCAAGCAAAAATTTGGAAACAAATTTTTGGTTTTGCAGAATCATTAGTTCTAAAATGTGCAGTTCAGTTAGAG
ATTGCTGAAACACTTCATAATAATGTAAAACCCATGTCTTTATCCGAGTTAGCATCTAAACTTCCGGCTCAACCC
GTTAATGAAGACCGTTTGTACCGAATTCTGCATTTCTTAGTTCACATGAAACTCTTCAACAAAGATGCTACCACA
CAGAAATATTCATTAGCTCCACCAGCAAAGTATTTGCTAAAAGGCTGGGAAAAATCAATGGTTCCTTCAATATTA
AGCGTGACTGATAAAGATTTTACAGCTCCATGGAATCATCTTGGGGACGGTTTAACCGGTAACTGTAACGCTTTT
GAGAAAGCGTTAGGAAAGGGCATTCGGGTTTATATGAGAGAAAATCCTGAAAAAGATCAATTGTTTAATGAAGGA
ATGGCTTGTGATACTAGATTATTTGCTTCAGCATTGGTTAACGAGTGCAAAAGTATTTTCAGTGACGGGATCAAT
ACACTTGCCGGTGTTGGCCGTGGTACTGGTACTGCAGTGAAAGCCATATCCAAAGCTTTTCCGGATATTAAGTGC
ACAATCCATGATCTTCCTGAAGTTACCAGTAAAAATAGTAAAATTCCAAGAGATGTTTTTAAGTCCGTTCCTAGT
GCAGACGCCATCTTTATGAAGGTAACTTCTAAGAAATTTTGTTTTAGAATATTCGTTGCAACTCTAATTGACAAC
ATTCATAAAAAATATGTTAATGGTCTTAATTTATTAATTCTAGTAGAGTTACTTAAATGATATACAAAAATTCAA
AATCATATAACATTTGCAGAGCATTCTTCACGAATGGAACGATGAGGAATGTATTCAAATCTTGAAACGATGCAA
AGAAGCAATACCAAAAGGGGGCAAAGTTATCATTGCGGATGTCGTAATAGACATGGACTCGACTCATCCGTATTC
AAAATCTAGACTCGCAATGGATTTGGCTATGATGCTCCACACGGGTGGAAAAGAGAGAACTGAAGAAGATTGGAA
AAAACTTATTGATGCTGCAGGTTTTGCTAGCTGTAAAATTACTAAACTATCTGCTCTCCAGTCTGTTATTGAGGC
TTACCCTCATTGAGGATAATTTTTATCCTTCTGTTTTCCCTTTGGTTAATTGTTGCCTTCTCTTTGGATCATGGT
TGCGTTTATAATAAATGCAGCGTTTCTTTCCTGGCGGTAAGTGCAAGAAAGAAAAAGCTTCCAGAAACTTCCTTG
AGTATGCCTGG

SEQ ID NO: 4 ***CYP82X1***

Start	End	Feature
1	130	5′ untranslated region
131	800	exon 1
801	881	intron 1
882	1216	exon 2
1217	1298	intron 2
1299	1916	exon 3
1917	1921	3′ untranslated region

CTTGAGTCATGCCTTGATATGCTCATATTTTAGTTTGTCATATTCACTATAACTATAAATTTCAATACAATTTCT
AAAACTCATCATCATTCAAGAGAGATACAAATACCTTGATATCCTTTTATCATCAATGGAGTTATTCATAAAGTT
ACCATTTATCCAACCAATTCCTTTCAGTATTATTCTTGTTACTACAGTTTCGATTGTTCTATTATACAGTGTCTT
CTTCTGGGTTACTGATAAGAAAAAGAAGAGGAAGAAAGCACCAAATGCTGCAGGGGCATGGCCGTTAATAGGTCA
TCTCCGTCTATTGATGAACGACAAGGAACCGTTGTATAGAGCACTAGGGAGCATGGCTGATAAGTACGGACCTGC
ATTCAACATCCGATTAGGTAACCAAGAAGTTCTTGTTGTGAGTAACTGGGAGATGGTAAAACAGTGTTTTGGTAA
TCAAAATGATAAGCTATTTTCGAATCGTCAAACTACATTAGCTGCAAAATACATGCTTAATCAAACAACTTCTAG
CGGATTCGCACCATATGGACCATATTGGACAGAGCTACGAAAGATAATGGTGCAGCAATTACTCTCTAAACAATC
TTTAGAATCGTGGAAACATCTGAAAATCAAAGAGATGGATGCTTCATTTAGTAAACTTAACGAGTTATGCAACAA
CAACGGTACTGGAACAGCTACCCTAATTAGGATGGACGAATGGTTTGCTGAGTTGACGTTCAACGTGATCGCAAG
AAATGTCTTTGGCTACCAAAGTGGCGGAAGGTCGACAGCGCTTACGAACGGTAATATGATCATACTCCCTCAATC
TGTATCAATTTAAGGAAATCATTTTGGTCTTGTTATTAACTTGAATTTTCTATTAGGAGATACGGAATCAAAGGG
CGAGAGGTACAAGAAAACATTGGAAGAAGCACTTCATCTTATGTCAATTTTTGCAGTTTCAGACATATTTCCAAG
TCTAGAGTGGGTAGATCGGTTAAGAGGCCTTATAAGGAATATGAAACGCTTTGGAGATGAGCTAAATTCAATTGC
AGGGTGTCTTATTGAAGAGCACCGCCAAAAGAGATTACAATCCGTATCTAAAAGTGATAAAGGAGTTGGTGATGA
ACAAGACTTCGTTGATGTTCTCTTATCGGTTGCTGAAAAATCGCAACTTCCTGGAGATGACCCTGATTTGGTCAT
CAAGTCTATGATTCTGGTTAGGCTATTGATACCAAGTCTATTGCAATTTTGGTTTATGTGCTTGTTCTAACTTTC
GTTTACTGCATATGGATGTGCAGGAAATCCTATCAGGTGGGAGTGAGACCACATCGTCAACCTTAACTTGGGCCC
TCTGTCTGTTACTGAACCATCCGCATGTGTTAAAGAAGGCAAAAGAGGAATTAGATACGCACGTAGGAAAAGATA
GGCATGTAGAAGAGTCAGATACCCCTAAGCTCGTGTACATTAATGCAATTATCAAAGAATCAATGCGATTGTATC
CAAACGGGGCAATGCTTGATCGGTTGGCGTTAGAAGAGTGCGAAGTTGGTGGATTTCATGTACCGGCCGGGGGAC
GCTTATTTGTCAATGTTTGGAAGATTCAGAGAGATCCGAGTGTTTGGGAGAATCCTCTGGAGTTTAAACCAGAGA
GGTGGTTTTTGAGTAATGGTGAAAAGATGCATGTGGATTACAAAGGTCACAATCATGAATTCATACCATTTGGGA
TAGGTCGGAGGATGTGCGCTGGTATGCTTTGGGCATCGGAGGTGATTCATTTGGTGCTGCCCCGTCTTATTCATG
GGTTTGATATGAAAGCAGCAAGTGCCAATGGGAAAGTAGATATGGCAGAAATGGCAGGCATGGTGATTTGTTTTA
AGAAGACACCTCTTGAAGTTATGGTCAATCCTCGAGAGTAGATGTT

SEQ ID NO: 5 ***CYP719A21***

Start	End	Feature
1	69	5′ untranslated region
70	1530	ORF
1531	1688	3′ untranslated region

CATGAAATTCTTTATGCAAAGAGTCAATCTGACTCAAGCTAGCTAGAATATATACCAATCATAAAAGAAATGATC
ATGAGTAACTTATGGATTCTTACGCTCATTTCTACCATATTAGCAGTCTTTGCTGCTGTGTTAATCATTTTCAGG
AGAAGAATATCAGCATCCACAACGGAATGGCCTGTTGGCCCAAAAACATTACCAATCATAGGTAACTTGCACATT
CTTGGAGGCACTGCTCTCCATGTCGTCTTACATAAACTTGCTGAAGTTTACGGCAGTGTAATGACGATATGGATT
GGTAGTTGGAAACCTGTTATTATTGTTTCCGACTTTGATCGAGCCTGGGAAGTTCTTGTTAACAAATCGTCAGAT
TATTCAGCTCGTGAAATGCCTGAGATCACTAAAATCGGCACTGCAAATTGGAGAACAATTTCAAGTTCTGATTCT
GGTCCGTTTTGGGCCACTCTTCGAAAAGGTCTTCAGAGTGTAGCATTATCGCCTCAGCATTTAGCATCGCAAACT
GCACACCAAGAGAGAGATATAATAAAGTTGATCAAAAATTTGAAAGACGAAGCAGCTTCTGGAATGGTTAAACCA
CTTGATCATCTCAAGAAAGCAACTGTAAGATTAATCAGTCGGTTAATCTATGGTCAGGATTTTGATGACGATAAG
TATGTTGAAGATATGCATGACGTGATCGAGTTTTTGATTCGTATTAGTGGTTATGCTCAACTTGCTGAGGTATTC
TATTATGCTAAATATCTACCAGGTCATAAGAGAGCTGTAACTGGCGCCGAAGAAGCAAAAAGAAGAGTAATAGCT
CTGGTGCGTCCTTTTCTTCAGTCAAACCCTGCTACTAACACTTACTTGCATTTTCTCAAATCGCAACTGTATCCT
GAAGAGGTTATCATATTCGCTATATTCGAAGCTTATCTTTTAGGTGTTGATAGCACTTCTTCAACCACTGCATGG
GCACTCGCATTCTTAATACGCGAACCATCTGTTCAAGAGAAACTTTATCAAGAGCTTAAGAATTTCACAGCCAAT

AACAATCGCACAATGCTGAAAGTCGAAGACGTCAACAAATTACCATATTTACAAGCTGTTGTTAAAGAAACAATG
AGGATGAAACCAATTGCACCACTGGCGATTCCTCATAAAGCTTGTAAAGACACTTCATTGATGGGCAAGAAAGTT
GATAAGGGAACTAAAGTTATGGTTAACATTCATGCTTTACATCATACTGAAAAGGTTTGGAAAGAACCTTACAAA
TTCATACCAGAGAGGTTTCTGCAGAAGCACGATAAGGCGATGGAACAATCACTATTACCATTTAGTGCAGGTATG
AGAATTTGTGCAGGAATGGAATTAGGAAAACTTCAGTTTAGTTTTTCTCTTGCTAATCTTGTTAATGCTTTTAAA
TGGTCTTGTGTGTCTGATGGAGTGCTTCCTGATATGAGTGATTTACTGGGGTTTGTTCTGTTCATGAAAACCCCA
CTCGAAGCACGTATAGTTCCTCGTCTGTAGTGATGGAAATTTCATCTCATGTTGTTGTTTCTCTTCATGTTTACT
ATTTCGTACTCGTTTGGTTTTGGTGTAAAAAATAAGATCTAAACTTCCAAATATCATTAATGTTTACACAAATCG
AAATCAATCAACTATGTTATGAAAATTAGTGTTTTCTC

SEQ ID NO: 6 ***CYP82X2***

Start	End	Feature
1	783	promoter sequence
784	893	5′ untranslated region
894	1581	exon 1
1582	1694	intron 1
1695	2050	exon 2
2051	2170	intron 2
2171	2791	exon 3
2792	2918	3′ untranslated region

>CYP82X2

AAGTGTGCCACTAATCTACTGCTAGTGCTACTGCTCACTGACACTTACACATATGATTGATTTATGGCTAAACAG
GATGACCACTAAATTTATTTTGGAAAGCGGAGTGAATTAATTAAGTGGCACATTTTCCATGAGAATTATTGATGG
CATGCATTTAGATGAACAAGATACACCAAATGTAGTGACTGAACAAGATGCTCGATCCTAACCCCACCTGCAACT
TTAGCTAAACTTTAATAATTACATGTCTTATCTTTTTATTGAATCATTTTATCTATCAATGGATGCTGATCAATA
ATATCATATATCTTTGCTTTTTCTTCAATCATTTAGATGAACAAAAAACACAATAAGTGTAGTGGTTGTTCATAA
CCCCACCTTCAACTCATTCTTCCCCTTAATAACAAATATCTTTGCTTTTTCTCCAATCATTTACTTGAACAACCA
ACACTAGTAAGTGTAGTGGTTTCTCATAACCCCACCTGCAATTTTTGCTTACCTTTAATAACATATATCTTTGAT
TTTCTTCGATCATTTTAGCTACCAATGGATGCTGATCCAAAAAGTTATGGCAAAAAGAGACAACGTCATCGAACA
CGAGCCTCTCGTGCACCACAGCATCAAGGTTTGTGGAAATTAACCGCTTGTAAAAAATGGAGTGCGTGATCATAA
TGAGGTATTGCTAAGATATAGTATCAACTTTAGTGAACTGGGCCAACAAAACTCACGAGTTGTTGAAAATTGGAG
ATTATATTTATAAGATAAAAGGGTCACTCCCTACACAACGACTTGCACTGCAAGTGAAAAAGAAAAAAAACAAAC
AACCTCAATCTAGCTAGAGTCGTGAAAAAGTTTTGTGCGACTGTTATTTAGTTAATTATAAAATTTCAATGAAGT
CGTTAATGATGAACAAGTTATTATCTCTCCAACGGATTACTGATTCTCCTTCGACCACCATTATCAGTACTTTTA
TTGTTACAATAATATCCATTGTTTCTCTCTACACTGTCTTGTTGATAAGGACGACTAAGAATAAGCAGAAGATAG
CAGCACCAAAAGCATCGGGGGCGTGGCCGTTCATAGGTCATCTCAAACTATTCATGAAACAAGATACTCAGTTTT
ACAGAACTCTAGGAACCATGTCTGATAAATACGGGTCGGTGTTCACACTTCGATTAGGAAACCAAGCAATCCTAG
TTGTGAGCAACTGGGAGATGGTAAAAGAATGTTTCACAACAAACGACAAGTCATTCTCGAATCGTCCAAGTACGT
TAAGCACTAAATACATGCTGAATGACACTAATTCTGTCGTGTTTTCACCTTACGGAACGTATTGGACAGAAATGC
GGAAGATATTGGTGCAAAAACTACTGATCTCTAACCAAAGATCAGAGGCATTGAAAAATCTGAAAACGAAAGAAA
TCGACAACTCGTTTGTAAAGCTTAATGATTTATGCAACAACGATGTCAGTGGAGGAGGCACAAAAGTTAGGATGG
ACGAATGGTTGGCTGACATGATGTTCAACATTATTGCTAGGATTACATTTGGTTACCAAAGCGGAGGAGGCGATG
CACCTGGTATGTGATCATCAAATTTTCGTTAAAACCAAATTAACTTGTACTATATCTTATGTTTACATGTTATAT
TGATCACTTTGACACGTTCTGATCATTTTCACAAATCGAATTAGGCGCTTCTACAACATCCAAGAATGTCGAGAG
ATACAAGAAAACGTTGGACGAGATGTTTGTTGTTTTAGCGACGAGGTTTGCAGTTTCAGATATATTTCCATCTCT
GGAGTTTATAGACCGATTGAGAGGTCTTGTAAAGGATATGAAAATCTTGGGAGACGAATTAAACTCCATTGCTGG
ATGTTTTATTGAAGAACATCGTCAAAAGAGACGAGAATCATTATCCTCATTGTTATCTTTGTCAAATGAATCCGT
TGGTGATGAACAAGATTTCATTGATGTTCTCTTGTCAATAATGGATCAGTCACGGCTTCCCGGAGATGACCCAGA
TTTTATTATCAAAATTATGATCCTGGTAACATATATTACAACAGTATTTCTTTAAGTTATGGATTAATGGATGTC
GTAACCATGAATATTTTTCTGATCTGGATAAATGTAATCCGGAACTAATATATGAATATTGTTGACGCAGGAAGC
TTTTGCAGGTGGGACGGACAGTTTAAGTGCAACCTTAACTTGGGTCCTCTCTCTACTGCTGAACCACCCAAACGT
GTTAAAGAGGGCAAGGGAGGAAATAGATAGGCATGTGGAAAACGGTAAGCAAGTGGAAGTGTCTGATATTCCGAA
GCTCGGATACATTGATGCAATAATCAAAGAGACGATGAGATTGTATCCAGTCGGAGCATTAAGCGAACGATACAC
GACTGAAGAATGCGAGGTTGGTCGGTTTAACGTACCCGCTGGCACACGCTTACTGGTGAATATATGCAAGATCCA
CAGAGACCCAAGTGTGTGGGAGAATCCATCAGATTTTCAACCAGAGAGGTTTTTGTGCAGCGATAAGGTGGGTGT

GGATTTATATGGCCAGAATTATGAGCTGATACCATTTGGGGCCGGTAGGAGGGTATGTCCGGCTATAGTTTCATC
ACTGCAGACGATGCATTATGCGTTGGCGCGTCTTATTCAAGGATATGAAATGAAATCAGCCAGCCTCGATGGGAA
GGTGAATATGGAAGAAATGATAGCCATGTCGTGCCACAAGATGAGCCCTCTTGAAGTTATTATCAGTCCTCGGGA
GCCGAGGCGGAGTTAAATCTTATGTTCCAATTTTACATTAGCATCTTTGATTATGAAATGTATTGCTCTTAAGTT
TCTTTTTTGTTTTTTATATTTTTAAGCTTGTATGTGATCATCAGCGAAAATGATGATGACAGAATCGT

SEQ ID NO: 7 ***CYP82Y1***

Start	End	Feature
1	12	5′ untranslated region
13	718	exon 1
719	815	intron 1
816	1153	exon 2
1154	1237	intron 2
1238	1864	exon 3
1865	1915	3′ untranslated region

> CYP82Y1
TTCAGTTCATTCATGGCGTATTTGATGATCAAGAAGTCTATCTATTTGTTTTTTGATCAACCAACTGCAGTTGGC
ACTCTTATACTTGCTTTTCTGCTGACACTTTCGCCTGTTATTATTTACTATGAACAGAAGAAGAGGGGTTTGAGG
CGAAATCGCACCGCAATTACAACGACTCCATTACCAGAGGCATCAGGTGCATGGCCAGTGATAGGTCATCTTCTT
CTTTTCATGAACGAAAACGATCTAAATCATGTAACTCTTGGTCACATGGCTGATAAATATGGACCTATTTTCAGC
TTAAGATTCGGTAGACATAGAACTCTAGTTGTTAGTAGTTGGGAGATGGTAAAGGAGTGTTTTACAGGTACCAAT
GACAAGTTGTTCTCAAATCGTCCTTCCTCCTTGGCGGTTAAACTTATGTTTTATGACACTGAATCTTATGGTTTT
GCACCTTATGGGAAATACTGGAGAGAGTTGCGAAAGATATCTACACACAAACTCCTCTCTAATCAGCAATTAGAG
AAGTTCAAGCACTTGCGGATTTCTGAAGTCGATAACTCCTTTAAAAAGCTTCATGAGTTATGCAGCAACAACAAA
CAGGGAGGTGATACTACATATGTGGCTAGTCTTGTGAGAATGGATGATTGGTTCGCGTACTTGACATTTAACGTA
ATAGGACGGATCGTCAGCGGATTCCAATCAAATGCAGTGGCAGGTGAGCTCATATAGCTAGGTTTTTATATGTTT
GGTTTGTACACACACAGCTCATTCATATTCTAAACTGAATTATATGTTATAATTGAACAACATAGGTGCCACAAA
CAGCCAGGAAAAATACAAGCTTGCAATCGATGAAGTGTCAAATCTTATGGCAACGTTTGCCGTTTCAGATGTGGT
TCCACGGCTTGGGTGGATTGATCGATTGACTGGTCTTACAGGAAAGATGAAGAATTGTGGTAAAAAATTAGATGC
AGTAGTTGGGGATGCAGTGGAGGATCATCGCCAAAAGAAACTCAAAATTTCTAGAAATAACACAGGAGCACTTAC
GGAGCACGAAGAAGAAGACTTTATCGATGTTTGCTTGTCGATTATGGAGCAGTCACAGATTCCGGGAAACCACCC
CGAAATCTCTGTCAAATCTATTGCCTTGGTAATACGTCTCATAAGCATGTTAGCAGATTTTACCTCTATATATAC
TTACATATTATTTTTTATCAATCACACATATGTGCAGGACATGTTATCGGGTGGGAGTGACACTACAAAATTGAT
AATGACATGGACCCTTTCTTTGCTGTTGAACCATCCAGACATATTGGACAAGGCTAAAGAAGAAGTAGATACATA
CTTCGGGAAGAAAAAGATATCGGATAACACACCTGTGGTTGATGCTGCCGATGTTCCTAACCTCGTCTACATCCA
AGCAATCATCAAAGAATCAATGCGGTTATACCCTGCTAGCACATTGATGGAGCGAATGACAAGTGATGATTGTGA
TGTTGGTGGCTTCCACGTACCAGCTGGGACACGATTATGGGTTAACGTATGGAAGATGCAACGGGACCCAAGGGT
GTGGAAAGATCCACTGGTATTTCTACCTGAGAGATTCTTGAGCAATGACAAAGGGATGGTAGATGTGAAGGGTCA
GAATTATGAACTGATACCATTTGGAACAGGCAGGCGGATATGTCCTGGTGCATCTTTGCCTTGGAAGTCTTGCA
TTTGGTTCTTACTCGTCTTATTCTTGAGTTCGAGATGAAGGCACCAGAGGGGAAAATTGACATGAGGGCAAGACC
AGGTTTTTTCCACAACAAGGTGGTGCCACTAGATGTTCAACTCACCCCACGCACACTAGATTAAGATTCCTATAT
ATGCTAATTAATTAGATGAATAAAATCTGTGGTCGAGTAA

SEQ ID NO: 8 ***PSCXE1***

Start	End	Feature
1	15	5′ untranslated region
16	978	ORF
979	1333	3′ untranslated region

>PSCXE1
AATAAAAATCCAACAATGGCAGATCCTTATGAATTCCTAATGTGCATTCACAATCCTGAAGAAGATACCCTAACA
ACAAATTTTCCCATTCCTGCTACTCCCTTACATCAAAACACCAAACACATTTCTTTAAATCCTCATACCAAAACC
TCACTTCGAATCTTTCGGCCACCAACCAAAGAACCTCCTGTAACAAAGAATAAGCTGCTTCCTATCATAATTTAT
TTCCATGGTGGAGGTTTCATTCTTTTCAATGCAGATTCAACTATGAACCATGACTTTTGTCAATCGATTGCTACA
CATATACCCGCGCTGGTCGTTTCTGTAGACTACCGTCTTGCTCCTGAAAACCGACTTCCCGCTGCCTATGATGAT
GCTGTTGATGCTTTAAACTGGGTCAAAGACCAAGGTTTAGGCAAACTAAATAATAGTGAAGTATGGTTAAAAGAG
TATGGTGACTTCTCAAAGTGTTTCATTATGGGGTGCAGCTCAGGTGCTAATGTTGCATATCATGCCAGTTTAAGA
GCAATAGAAATGGATCTTGAACCAGCTAAGATTAATGGATTAATATTACACTGCCCTTTTTTTGGTAGTCTTGAG
AGAACTGAATCAGATTCAAAAGTGATCAACAATCAGGACTTGCCGCTTGCCGTAAGGGATGTCATGTGGGAACTG
GCGTTGCCGCTTGGGTCTACTCGTGATCACGTTTATTGTAATCCGAATATTGATCATGATGGATCATCATCTGGA
AATATGGTGGGGTTAATCGAGAGACGTTTTGTGGTAGGATTTTATGGGGATCCACTTATTGATCGACAAATTCAG
CTGGTGAAGATGCTCGAGGAAAAAGGTGTGAAGGTTGAAACTTGGATTGAACAAGGAGGGTATCATGGGGTGCTA
TGCTTTGACCCTATGATACGTGAAACCTTTTTGGAAAAACTAAAACATTTTATTTTAAACGACGAATTTATATAC
TAAAATATATTATTAGTATTAAACAATGAAATTCTTATTTTTTCTAAAATGAGCTTTTGGACGAAACATTGTGTA
CGAACTAGCTGATGTAATTTTTCGCTTTACCGGATTTTTCATTTTTTTTGCTTTCTTTCTGCTCTCTTTTATAAG
TCGTTCTT

SEQ ID NO: 9 ***PSSDR1***

Start	End	Feature
1	254	5′ untranslated region
255	423	exon 1
424	635	intron 1
636	1178	exon 2
1179	1953	intron 2
1954	2146	exon 3
2147	2236	intron 3
2237	2378	exon 4
2379	2488	3′ untranslated region

>PSSDR1
CTAACAGGCAAACAATAACAGGTTGCACCTACAACATTCAATTTTTATTTTGGTAAATGAAGTTCAGTTGGAGAGTAACCACA
TCTTTGTTGTCGGCATCGCCCCCACAATACTGAGTGTTTTGGCTGAGTGTAGTCTGACTGTAGGTAAGCTACAACTGCATGTT
GCAGATAATAATCACTAACTGATTATTCATGCATACCTAACAGTCATATTGTTATAGTTCCCAAAAAAAAATTCTCGAACTATA
AAGGCATGCATGGACAGAAAAATATATCAGAGAGATATCAGAAATTCAAAGAGATGGAAGGAACAGGGAAGATAGTATGTGTA
ACAGGTGGAGCTGGATACTTGGCATCTTGGCTGATCATGAGATTGCTTGAACGTGGTTACTCTGTTCGGACCACCGTTCGGTC
TGACCCAAGTACGTAACAAATTAAATTTCCTGGCATCATTTTCTTCAATATAAATTTCTTATTATCTAGTTCATCATTCTTTA
TTGTTCCAATCATGTCCCCCCAAGTCTAAAAGAAGTAGTAATCTAAAAATAGCTAATTTATGTACGAAATTGTAACAATGATC
TCCTAGCTTATGAGGCCCACCTAATTTCGTTTCTATCATTTTGTGTCTTGAAAGAATTTAGGGAAGATGTGAGCCACCTTAAA
GCTCTTCCTGAAGCTACAGAGAAGCTTCAAATTTTTGAAGCAGATCTTGAAAACCCAGAAAGTTTCGACGATGCGATCAACGG
TTGTGTCGGTGTCTTTCCCGTTGCTCAAGGAATGAATTTTGCCGAAGAATATACTCTTGAAAAAATAATCAAAACATGCGTGG
AAGGAACTCTTAGAATCCTACAGTCATGCTTGAAATCTAAAACAGTGAAAAAGGTTGTGTACACATCTTCTGCTGATGCAGCA
ATGATGATAAGTAATCCCAAAGCTGTTAAAGAAATTGACGAGACAATATGGTCAGAAGTTGACAATTTCATTAGCAAACCGGA
ACAAGTTATTCCTGGACCGCCCTCATATGTGGTTTCAAAGGTACTGACAGAAAGAGCTTGCCTAAAGTTTTCTGAAGAACATG
GTTTGGATGTTGTTACCATACTTCCTCCGTTGGTTGTTGGACCTTTTATCACTCCCCATCCTCCTCCCAGTGTATCTATAGCT
CTTTCGATAATTTCAGGCATCCTCTCCATCCGAAAAATATGCCAATTCCTAAACTTAAAAGGCATATTGATATTTAATAATAC
CTCCATACCTAAAAAAAGAGTTGCTATATAACATTTTTAATTTTCGCCCATTTTTAGGCCTAATTGAAAAAGTTATAATAACA
TTTTAGGAAGGAGGGAGAATGATTTTTGAGCAAACCTTAGAACTGTGTGGTGAGATTTGTCCGTTATCATTGTTGGTATAACT
GTGTATATCATGGTTTTCAAAAGCGCCGCTCACGCTACGCTTCGTACGGTTCGGTCTAGATTTTTTTAATTCGCTCCGAAGCG
TAGTTATGAAGCTACCATGAAGCGCCGCTTCACGCTACGTTTCGTACGCTTCGCTTCAGATTTTTCAAATTCGCTCCGAGGCG
AATCTACCATGAAGTGGAAGATTCCTTTAATTGATTCACTTTTTTACTTAGTCAAGTCTTTTTTAGGGGGTTTCGAAAACTAA
AGTGAACCACTGCGCCCCGCTACTGTTTTTGAAATTAACTAGACTTATATTAAATTGATACAATTATTATATCTTCCTAAATA

TTAAATTATTAATAACAAACTACTACTATTTATAGGAAAAAATTCGCTTCAAATATCAATCATAAAACGACGCTTCACATTTC
AACATGCGCATCGCTTCGTATAAAAAAAAAACGCTTCACGCTTTCAATACCTTGGTGTACATAGATTAATACTTCCTCCTCTG
CGCTGGTGTTAACATTTCTGTGTTTCGTTTATATATATGACCAGGTGATGTGTCGATGATGCTTGGTGTTAGACTTGAAAATG
CGGTACATATAGATGATGTTGCTTTAGCACACATATTCGTTTTTGAATGTGAAAAAGCAAAAGGAAGACATATTTGTTCTTCA
GTTGATTTTCCAATGCATGATCTGCCTAAATTTATATCTGAGAATTATCCGGAATTCAACGTACCGACTGAGTGAGTTTATCT
TTCACCACCTTCTTTATTATTATTCATCAAGTCACTTTGGGTATTTTAACCTTATTGTTTTTACTGAATTATCATCAGTTTAC
TAAAGGATATTGAGGAACAAGAACCAGTTCATCTTTCCTCAGATAAGCTGTTGAGTATGGGATTTCAGTTCAAATATGATTTT
GCAGAGATTTTCGGTGATGCAATACGATGTGCCAAAGAGAAGGGTTTCCTTTAGAGACCAACTATAGTTTGGTTCGGAGGAGA
TGTGGGAGTAGCTAGCCCAAAATGCCCTGCTCGCACTAGCTTATATTATTGTTATTGTTTTTCAAATGAATAAACGGGCAG

SEQ ID NO: 10 *PSAT1*

Start	End	Feature
1	53	5′ untranslated region
54	1469	ORF
1470	1572	3′ untranslated region

> PSAT1
CGCATATAATCCAATTTGCATTGTTTATCGACCTTGAGGAACAATTAGGGGATATGGCAACAATGTCTAGTGCTG
CTGTAGAAGTGATCTCGAAAGAAACGATTAAACCAAGAAATCCAACACCATATCAACTTAGAAACTACAATATGT
CACTTCTCGACCAATATTCTTCTCTAGTTTATGTTCCGATCATTCTTTTCTACCCTGCTGCCTCCGACGCTAATA
GTACCGGAAGTAAGCACCATGATGATCTTCACTTGCTTAAGAGGTCTCTTTCTGAAACGCTAGTTCACTTTTATC
CAATGGCTGGTAGGATGAAAGACAACATGACTGTTGACTGTAACGACGAAGGTATTGACTTTTTCGAAGTAAGAA
TCAAAGGTAGAATGTGTGACTTCATGATGAAATCAGATGCACACTTAAGTCTGCTTCTTCCGTCTGAAGTCGCTT
CCACGAACTTCGTGAAGGAAGCACAGGTGATTGTTCAAGTGAACATGTTTGATTGCGGTGGAACTGCCATTTGTT
TCTGTATATCAAACAAGATTGCAGATGCATGCACCATGATTACCTTCATTCGTAGTTTGGCAGGCACCACCAACA
TAGCTCGTCGTGGGAGCTCTATTGCTGCACCAACCACAAATCAGAATTTGGTTCCTTCTTTCGATTCGACATCAC
TCTTTCCACCTAGTGAACAATTGGCATCTCAAGTTTCCTATCCTACACAGGATAGTACCAGCGTAGATAAACTTG
TCAGCAAAAGATTTGTGTTTGATGCGGCAAAGATTACATCTGCACGTGAAAAATTGCAATCCTTGATGCATGATA
AATACAAATGCCATAGGCCGACAAGGGTTGAGGTAGTTTCCGCTTTGATATGGAAGTCAGCAGTGAAATCTGCTC
CGCCCGGTTCTATATCCACTGTAACCCATGCCATGAACTTTAGAAAGAAAATGGATCCACCATTACAAGATGCGT
CATTCGGGAATCTTTGTGTGGTTGTTACAGCAGTATTACCAGCAACAACGGCGACAACAACAAATCCAGCAACCA
AAAAAGTTAGTAGTACGAGTAATGAAGAGCAAGTGGCACTTGATGAGTTAAGTGATTTTGTAGCCCTATTGAGGC
GCGAAATAGATAAGGTAAAGGGTGATAAAGGTTGCATGGAGAAAATCATTCAAAAGTTCATCTATGGTCATGATG
CTTCCGTAGCGAAAGACAGTGATGTTGAAGATAAGGTGACAGCTTTGTTTATGACTAGCTGGTGCAAGTTTGGAT
TCTACGAAGCTGATTTTGGTTGGGGAACGCCAGTTTGGGTAACTACTGTTCCATTAATTGAGCCAAAGTACAAGA
ACATGGTTTTCATGAACGATATGAAATGTGGTGAAGGAATTGAAGTGTGGGTGAATTTTCTGGAGGATGATATGA
CCAAGTTCGAACACCACCTAAGAGAGATCCTCCAACTGTTTTGATTTTCAACCGTTTCCCTAATAGAGGTCAATT
GTCGTGTTTGTCCATCTTAACTACCATCTTTATTCTCTTGTTTTCATACTTGTATTTGTCTTACTCCGGTAA

SEQ ID NO: 11 PSMT1

MATNGEIFNTYGHNHQSATVTKITASNESSNGVCYLSETANLGKLICIPMALRAAMELNVFQLISKFGTDAKVSA
SEIASKMPNAKNNPEAAMYLDRILRLLGASSILSVSTTKKSINRGGDDVVVHEKLYGLTNSSCCLVPRQEDGVSL
VEELLFTSDKVVVDSFFKLKCVVEEKDSVPFEVAHGAKIFEYAATEPRMNQVFNDGMAVFSIVVFEAVFRVYDGF
LDMKELLDVGGGIGTSVSKIVAKYPLIRGVNFDLPHVISVAPQYPGVEHVAGDMFEEVPKGQNMLLKWVLHDWGD
ERCVKLLKNCWNSLPVGGKVLIIEFVLPNELGNNAESFNALIPDLLLMALNPGGKERTISEYDDLGKAAGFIKTI
PIPISNGLHVIEFHK.

SEQ ID NO: 12 PSMT2

MEIHLESQEQEMKYQSQIWNQICGTVDTSVLRCAIQLGIFDAIHNSGKPMITLTELSSIVSSPSSSSIEPCNLYR
LVRYLSQMDLISIGECLNEATVSLTGTSKLLLRNQEKSLIDWVLAISCEMMVVVWHELSSSVSTPADEPPIFQKV
HGKNALELAGEFPEWNDLINNAMTSDTSVTKPALIQGCGKILNGVTSLIDVGGGHGATMAYIVEAFPHIKGAVID
LPHVVEAAPERPGVEFISGDIFKSISNADAVLLKYVLHNWEDTECVNLLKRCKEAVPADKGKVIIMDLVIDDDDN
SILTQAKLSLDLTVMNHGGGRERTKEDWRNLIEMSGFSRHEIIPISAMPSIIVAYP.

SEQ ID NO: 13 PSMT3

MEVVSKIDQENQAKIWKQIFGFAESLVLKCAVQLEIAETLHNNVKPMSLSELASKLPAQPVNEDRLYRILHFLVH
MKLFNKDATTQKYSLAPPAKYLLKGWEKSMVPSILSVTDKDFTAPWNHLGDGLTGNCNAFEKALGKGIRVYMREN
PEKDQLFNEGMACDTRLFASALVNECKSIFSDGINTLAGVGRGTGTAVKAISKAFPDIKCTIHDLPEVTSKNSKI
PRDVFKSVPSADAIFMKSILHEWNDEECIQILKRCKEAIPKGGKVIIADVVIDMDSTHPYSKSRLAMDLAMMLHT
GGKERTEEDWKKLIDAAGFASCKITKLSALQSVIEAYPH.

SEQ ID NO: 14 CYP82X1

MELFIKLPFIQPIPFSIILVTTVSIVLLYSVFFWVTDKKKKRKKAPNAAGAWPLIGHLRLLMNDKEPLYRALGSM
ADKYGPAFNIRLGNQEVLVVSNWEMVKQCFGNQNDKLFSNRQTTLAAKYMLNQTTSSGFAPYGPYWRELRKIMVQ
QLLSKQSLESWKHLKIKEMDASFSKLNELCNNNGTGTATLIRMDEWFAELTFNVIARNVFGYQSGGRSTALTNGD
TESKGERYKKTLEEALHLMSIFAVSDIFPSLEWVDRLRGLIRNMKRFGDELNSIAGCLIEEHRQKRLQSVSKSDK
GVGDEQDFVDVLLSVAEKSQLPGDDPDLVIKSMILEIVSGGSETTSSTLTWALCLLLNHPHVLKKAKEELDTHVG
KDRHVEESDTPKLVYINAIIKESMRLYPNGAMLDRLALEECEVGGFHVPAGGRLFVNVWKIQRDPSVWENPLEFK
PERWFLSNGEKMDVDYKGHNHEFIPFGIGRRMCAGMLWASEVIHLVLPRLIHGFDMKAASANGKVDMAEMAGMVI
CFKKTPLEVMVNPRE.

SEQ ID NO: 15 CYP719A21

MIMSNLWILTLISTILAVFAAVLIIFRRRISASTTEWPVGPKTLPIIGNLHILGGTALHVVLHKLAEVYGSVMTI
WIGSWKPVIIVSDFDRAWEVLVNKSSDYSAREMPEITKIGTANWRTISSSDSGPFWATLRKGLQSVALSPQHLAS
QTAHQERDIIKLIKNLKDEAASGMVKPLDHLKKATVRLISRLIYGQDFDDDKYVEDMHDVIEFLIRISGYAQLAE
VFYYAKYLPGHKRAVTGAEEAKRRVIALVRPFLQSNPATNTYLHFLKSQLYPEEVIIFAIFEAYLLGVDSTSSTT
AWALAFLIREPSVQEKLYQELKNFTANNNRTMLKVEDVNKLPYLQAVVKETMRMKPIAPLAIPHKACKDTSLMGK
KVDKGTKVMVNIHALHHTEKVWKEPYKFIPERFLQKHDKAMEQSLLPFSAGMRICAGMELGKLQFSFSLANLVNA
FKWSCVSDGVLPDMSDLLGFVLFMKTPLEARIVPRL.

SEQ ID NO: 16 CYP82X2

MKSLMMNKLLFLQRITDSPSTTIISTFIVTIISIVFLYTVLLIRTTKNKQKIAAPKASGAWPFIGHLKLFMKQDT
QFYRTLGTMSDKYGSVFTLRLGNQAILVVSNWEMVKECFTTNDKSFSNRPSTLSTKYMLNDTNSVVFSPYGTYWR
EMRKILVQKLLISNQRSEALKNLKTKEIDNSFVKLNDLCNNDVSGGGTKVRMDEWLADMMFNIIARITFGYQSGG
GDAPGASTTSKNVERYKKTLDEMFVVLATRFAVSDIFPSLEFIDRLRGLVKDMKILGDELNSIAGCFIEEHRQKR
RESLSSLLSLSNESVGDEQDFIDVLLSIMDQSRLPGDDPDFIIKIMILEAFAGGTDSLSATLTWVLSLLLNHPNV
LKRAREEIDRHVENGKQVEVSDIPKLGYIDAIIKETMRLYPVGALSERYTTEECEVGRFNVPAGTRLLVNIWKIH
RDPSVWENPSDFQPERFLCSDKVGVDLYGQNYELIPFGAGRRVCPAIVSSLQTMHYALARLIQGYEMKSASLDGK
VNMEEMIAMSCHKMSPLEVIISPREPRRS.

SEQ ID NO: 17 CYP82Y1

MAYLMIKKSIYLFFDQPTAVGTLILAFLLTLSPVIIYYEQKKRGLRRNRTAITTTPLPEASGAWPVIGHLLLFMN
ENDLNHVTLGHMADKYGPIFSLRFGRHRTLVVSSWEMVKECFTGTNDKLFSNRPSSLAVKLMFYDTESYGFAPYG
KYWRELRKISTHKLLSNQQLEKFKHLRISEVDNSFKKLHELCSNNKQGGDTTYVASLVRMDDWFAYLTFNVIGRI
VSGFQSNAVAGATNSQEKYKLAIDEVSNLMATFAVSDVVPRLGWIDRLTGLTGKMKNCGKKLDAVVGDAVEDHRQ
KKLKISRNNTGALTEHEEEDFIDVCLSIMEQSQIPGNHPEISVKSIALDMLSGGSDTTKLIMTWTLSLLLNHPDI
LDKAKEEVDTYFGKKKISDNTPVVDAADVPNLVYIQAIIKESMRLYPASTLMERMTSDDCDVGGFHVPAGTRLWV
NVWKMQRDPRVWKDPLVFLPERFLSNDKGMVDVKGQNYELIPFGTGRRICPGASFALEVLHLVLTRLILEFEMKA
PEGKIDMRARPGFFHNKVVPLDVQLTPRTLD.

SEQ ID NO: 18 PSCXE1

MADPYEFLMCIHNPEEDTLTRNFPIPATPLDQNTKDISLNPDRKTSLRIFRPPTKEPPVTKNKLLPIIIYFHGGG
FILFNADSTMNHDFCQSIATHIPALVVSVDYRLAPENRLPAAYDDAVDALNWVKDQGLGKLNNSEVWLKEYGDFS
KCFIMGCSSGANVAYHASLRAIEMDLEPAKINGLILHCPFFGSLERTESDSKVINNQDLPLAVRDVMWELALPLG
STRDHVYCNPNIDHDGSSSGNMVGLIERCFVVGFYGDPLIDRQIQLVKMLEEKGVKVETWIEQGGYHGVLCFDPM
IRETFLEKLKHFILNDEFIY.

SEQ ID NO: 19 PSSDR1

MHGQKNISERYQKFKEMEGTGKIVCVTGGAGYLASWLIMRLLERGYSVRTTVRSDPKFREDVSHLKALPEATEKL
QIFEADLENPESFDDAINGCVGVFLVAQGMNFAEEYTLEKIIKTCVEGTLRILQSCLKSKTVKKVVYTSSADAAM
MISNLKAVKEIDETIWSEVDNFISKPEQVIPGLPSYVVSKVLTERACLKFSEEHGLDVVTILPPLVVGPFITPHP
PPSVSIALSIISGDVSMMLGVRLENAVHIDDVALAHIFVFECEKAKGRHICSSVDFPMHDLPKFISENYPEFNVP
TDLLKDIEEQEPVHLSSDKLLSMGFQFKYDFAEIFGDAIRCAKEKGFL.

SEQ ID NO: 20 PSAT1

MATMSSAAVEVISKETIKPRNPTPYQLRNYNMSLLDQYSSLVYVPIILFYPAASDANSTGSKHHDDLHLLKRSLS
ETLVHFYPMAGRMKDNMTVDCNDEGIDFFEVRIKGRMCDFMMKSDAHLSLLLPSEVASTNFVKEAQVIVQVNMFD
CGGTAICFCISNKIADACTMITFIRSLAGTTNIARRGSSIAAPTTNQNLVPSFDSTSLFPPSEQLASQVSYPTQD
STSVDKLVSKRFVFDAAKITSAREKLQSLMHDKYKCHRPTRVEVVSALIWKSAVKSAPPGSISTVTHAMNFRKKM
DPPLQDASFGNLCVVVTAVLPATTATTTNPATKKVSSTSNEEQVALDELSDFVALLRREIDKVKGDKGCMEKIIQ
KFIYGHDASVAKDSDVEDKVTALFMTSWCKFGFYEADFGWGTPVWVTTVPLIEPKYKNMVFMNDMKCGEGIEVWV
NFLEDDMTKFEHHLREILQLF.

SEQ ID NO: 21 VIGS *PSMT1*

TGGTCATAATCATCAATCAGCCACAGTCACTAAAATCACTGCTTCTAATGAAAGCAGCAATGGTGTCTGTTATCT
TTCAGAAACGGCTAACTTGGGGAAGTTAATATGCATTCCAATGGCACTAAGAGCTGCGATGGAGCTAAATGTGTT
CCAACTTATCTCAAAGTTCGGAACTGACGCAAAAGTTTCGGCTTCTGAAATTGCCTCTAAAATGCCAAACGCGAA
GAATAATCCTGAAGCAGCTATGTATTTGGATAGAATTCTTCGACTGCTCGGGGCAAGTTCTATTCTTTCTGTTTC
TACTACAAAAAAATCAATCAACAGAGGAGGAGATGATGTAGTAGTACATG

SEQ ID NO: 22 VIGS *PSMT2*

GTGTAACTAAGCCAGCGCTAATACAAGGATGTGGCAAAATCCTGAACGGAGTTACATCGTTAATTGATGTCGGTG
GTGGTCACGGTGCCACTATGGCCTACATAGTTGAAGCTTTTCCTCACATAAAAGGTGCGGTAATCGATTTACCAC
ATGTTGTTGAAGCCGCTCCGGAGCGTCCAGGTGTTGAGTTCATCAGCGGTGATATATTCAAGT

SEQ ID NO: 23 VIGS *CYP82X1*

TTTGAGTAATGGTGAAAAGATGGATGTGGATTACAAAGGTCACAATCATGAATTCATACCATTTGGGATAGGTCG
GAGGATGTGCGCTGGTATGCTTTGGGCATCGGAGGTGATTCATTTGGTGCTGCCCCGTCTTATTCATGGGTTTGA
TATGAAAGCAGCAAGTGCCAATGGGAAAGTAGATATGGCAGAAATGGCAGGCATGGTGATTTGTTTTAAGAAGAC
ACCTCTTGAAGTTATGGTCAATCCTCGAGAGTAGATGTT

SEQ ID NO: 24 VIGS *CYP719A21*

ATGATCATGAGTAACTTATGGATTCTTACGCTCATTTCTACCATATTAGCAGTCTTTGCTGCTGTGTTAATCATT
TTCAGGAGAAGAATATCAGCATCCACAACGGAATGGCCTGTTGG

SEQ ID NO: 25 VIGS *CYP82X2*

TAGGAGGGTATGTCCGGCTATAGTTTCATCACTGCAGACGATGCATTATGCGTTGGCGCGTCTTATTCAAGGATA
TGAAATGAAATCAGCCAGCCTCGATGGGAAGGTGAATATGGAAGAAATGATAGCCATGTCGTGCCACAAGATGAG
CCCTCTTGAAGTTATTATCAGTCCTCGGGAGCCGAGGCGGAGTTAA

SEQ ID NO: 26 VIGS *CYP82Y1*

TCCTATATATGCTAATTAATTAGATGAATAAAATCTGTGGTCGAGTAAATCTAATTAATGCTAATGAACAAGATG
AATAAAAAATTTTCTTTCTGCTTTTGCTTTGGTTAGGGTTATTTGACCCTCATTTGGTTGTATTCGTTGGCGCAC
AACTTTTGTGCTTCTTAATATAATTCCTTTTGGTGG

SEQ ID NO: 27 VIGS *PSCXE1*

TGGCAGATCCTTATGAATTCCTAATGTGCATTCACAATCCTGAAGAAGATACCCTAACAAGAAATTTTCCGATTC
CTGCTACTCCCTTAGATCAAAACACCAAAGACATTTCTTTAAATCCTGATAGGAAAACCTCACTTCGAATCTTTC
GGCCACCAACCAAAGAACCTCCTGTAACAAAGAATAAGCTGCTTCCTATCATAA

SEQ ID NO: 28 VIGS *PSSDR1*

GAAATTGACGAGACAATATGGTCAGAAGTTGACAATTTCATTAGCAAACCGGAACAAGTTATTCCTGGATTGCCC
TCATATGTGGTTTCAAAGGTACTGACAGAAAGAGCTTGCCTAAAGTTTTCTGAAGAACATGGTTTGGATGTTGTT
ACTATACTTCCTCCGTTGGTTGTTGGACCTTTTATCACTCCCCATCCTCCTCCCAGTGTATCTATAGCTCTTTCG
ATAATTTCAGGTGATGTGTCGATGATGCTTGGTGTTAGACTTGAAAATGCGGTACATATAGATGATGTTGCTTTA
GCACACATATTCGTTTTTGAATG

SEQ ID NO: 29 VIGS *PSAT1*

CCTAAGAGAGATCCTCCAACTGTTTTGATTTTCAACCGTTTCCCTAATAGAGGTCAATTGTCGTGTTTGTCCATC
TTAACTACCATCTTTATTCTCTTGTTTTCATACTTGTATTTG

SEQ ID NO: 30 VIGS *PSPDS*

GATCATCTTCTCTTCAGCAGAAGTCCCCTCTTAAGCGTATACGCTGACATGTCAGTGACATGCAAGGAATATTAT
GACCCAAACAAATCCATGCTTGAGTTGGTATTTGCACCCGCTGAGGAATGGATC