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(54) Titre : IDENTIFICATION, OPTIMISATION ET UTILISATION D'EPITOPES HLA-B*0702 PARTAGES POUR
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(54) Title: IDENTIFICATION, OPTIMIZATION AND USE OF SHARED HLA-B*0702 EPITOPES FOR IMMUNOTHERAPY

(57) **Abrégé/Abstract:**

The present invention provides novel methods and materials for efficiently treating patients having an HLA-B*0702 phenotype, based on peptides representing shared epitopes of tumour antigens. In particular, the invention relates to a method for identifying a HLA-B*0702-restricted peptide which can trigger a cytotoxic response against several antigens from one single multigenic family, and to several such epitopes.



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(57) Abstract: The present invention provides novel methods and materials for efficiently treating patients having an HLA-B*0702 phenotype, based on peptides representing shared epitopes of tumour antigens. In particular, the invention relates to a method for identifying a HLA-B*0702-restricted peptide which can trigger a cytotoxic response against several antigens from one single multigenic family, and to several such epitopes.

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IDENTIFICATION, OPTIMIZATION AND USE OF SHARED HLA-B*0702 EPITOPES FOR IMMUNOTHERAPY

The present invention relates to the field of peptide immunotherapy. In particular, the invention provides novel methods and materials for efficiently treating patients having an HLA-B*0702 phenotype, based on peptides representing shared epitopes of tumour antigens.

Peptide immunization or immunotherapy is a therapeutic approach which is currently the subject of great interest in the context of the prevention or treatment of cancers. The principle thereof is based on immunization with peptides which reproduce T epitopes of tumour antigens that are recognized by Cytotoxic T Lymphocytes (CTLs), which play a major role in the elimination of cancer cells expressing these antigens at their surface.

It will be recalled that CTLs do not recognize whole protein antigens, but peptide fragments thereof, presented by the major histocompatibility complex (MHC) molecules expressed at the surface of various cells. These peptide fragments constitute the T epitopes. The peptides presented by the major histocompatibility complex class I (MHC I) generally have 8 to 11 amino acids, and are recognized by CD8⁺ T cells, which represent the major component of the cytotoxic response. During the antigen processing, a peptide selection takes place, which results in a hierarchy of peptides presentation. Peptides that are preferentially presented by the MHC I molecules are called immunodominant while peptides that are weakly presented are called cryptic. Immunodominant peptides exhibit a high affinity for the MHC I and are immunogenic while cryptic peptides exhibit a low affinity for MHC I and are non-immunogenic.

The identification of tumour specific epitopes, and in particular (given the essential role of the CD8⁺ response in cytotoxicity) of those presented by the more frequent MHC I alleles, constitutes an essential step for the development of anti-tumour immunotherapy compositions. Many tumour antigens are known at the current time; some of the T epitopes of these antigens have been identified and the effectiveness of vaccines based on peptides which reproduce these T epitopes has been shown in many cases (Menez-Jamet and Kosmatopoulos, 2009).

However, the expression of the majority of tumour antigens is restricted to certain histological types of tumours, which limits their clinical use. The search for broadly expressed "universal" tumour antigens has been intensified with the identification of antigens with functions essential for the maintenance of the oncogenic phenotype, and effort are being made to identify epitopes expressed by a majority of patients.

Another considerable limitation of peptide immunotherapy comes from the appearance, in certain patients, of tumour variants (escape variants) which no longer express the antigen recognized by the cytotoxic T lymphocytes.

Some tumour antigens belong to multigene families: within the same family, there is a sequence homology, which may result in the existence of shared epitopes common to two or more members of the same family.

5 Generally, various members of the same family of antigens are expressed in various tumour types; the use of an epitope shared by these antigens could make it possible to obtain anti-tumour vaccines with a broad spectrum of activity.

Furthermore, in many cases, several antigens of the same family are co-expressed in the same tumour line; since the probability of loss of the expression of all these antigens is extremely low, the use of an epitope shared by these antigens may avoid
10 the appearance of escape variants.

Among the tumour antigens known to belong to a multigene family, mention will in particular be made of the antigens of the MAGE-A, HER, BAGE or GAGE families.

MAGE-A is a multigene family consisting of 12 homologous genes
15 (MAGE-A1 to A12) located in the q28 region of the X chromosome (De Plaen et al., 1994). Among the members of this family, MAGE-A1, -A2, -A3, -A4, -A6, -A10 and -A12 are strongly expressed by tumours but not by normal tissues, with the exception of the testis and of the placenta.

The MAGE-A1, -A2, -A3, -A4, -A6, -A10 and -A12 antigens are present
20 in a wide spectrum of tumours of very varied histological origin, such as melanomas, lung cancers, breast cancers, head and neck tumours, and sarcomas, myelomas, etc.

MAGE-based cancer vaccines, such as MAGE-A3 Antigen Specific Cancer Immunotherapeutic (ASCI) (GlaxoSmithKline) are currently in late phase of development with encouraging results. For example, this vaccine, which is based on
25 tumour antigens presented to the patient's immune system as recombinant proteins in combination with a GSK proprietary adjuvant system, has completed successfully two clinical trials in melanoma and non small cell lung cancer.

The expression of each MAGE-A antigen can vary from one tumour to another, but overall, the vast majority of tumours express at least one MAGE-A antigen.

30 Despite the potential advantage of using shared T epitopes, this approach has only been very rarely used because of the rarity of the regions of appropriate size (at least 8 amino acids for a peptide presented by MHC I) that are completely identical from one antigen to another.

The inventors have previously described a method for identifying peptide
35 epitopes presented by an HLA class I molecule and shared by several antigens of the same multigen family. This method is characterized by the following steps (EP1 485 719):

a) aligning the sequences of said antigens in order to identify on each of them a sequence of 8 to 10 amino acids comprising at least one common pentapeptide

sequence preceded by 3 amino acids at the N-terminal end and, optionally, followed by one or two amino acids at the C-terminal end; indeed, the authors have found that an identity limited to the sequence of 5 amino acids extending from positions P4 to P8 of the peptide was sufficient.

5 b) preparing the peptides corresponding to the sequences identified and determining the binding affinity of each of the peptides for the HLA class I molecule concerned, and their immunogenicity using human CMH-I transgenic mice

 c) In case a selected peptide is cryptic and consequently non-immunogenic, the method further comprises a step of increasing its immunogenicity.

10 Using this method, the inventors have described an immunogenic peptide defined by the sequence YLEYRQVPV (SEQ ID No: 1), presented by HLA-A*0201 common to the MAGE-A1, -2, -3, -4, -6, -10 and -12 antigens of the MAGE-A family, capable of inducing CTLs which recognize all the MAGE-A antigens, and of lysing tumour cells expressing at least one antigen of the MAGE-A family.

15 Immunodominant peptides have widely been targeted by tumour vaccines in preclinical and clinical studies with disappointing results (Gross et al., 2004). Indeed, tumour antigens are frequently self proteins over-expressed by tumours and expressed at lower levels by normal cells and tissues. The immune system is unable to react against these self antigens because of the self tolerance process. Self-tolerance
20 concerns mainly the immunodominant peptides, thus explaining the incapacity of these peptides to induce tumour immunity.

 Cryptic peptides are much less involved in self tolerance process (Gross et al., 2004) and can therefore induce an efficient tumour immunity providing their immunogenicity is enhanced.

25 The usual strategy for enhancing the immunogenicity of cryptic peptides, which because of their low MHC I affinity are non-immunogenic, consists in increasing their affinity for the MHC I molecules *via* amino acids substitutions. Peptide affinity for MHC I molecules mainly depends on the presence at well defined positions (primary anchor positions) of residues called "primary anchor residues". These residues are MHC I
30 allele specific. The presence of primary anchor residues, although often necessary, is not sufficient to ensure a high MHC I affinity. It has been shown that residues located outside the primary anchor positions (secondary anchor residues) may exert a favourable or unfavourable effect on the affinity of the peptide for the MHC I. The presence of these secondary anchor residues makes it possible to explain the existence, within the peptides
35 having the primary anchor motifs, of a great variability in the binding affinity (Ruppert et al., 1993).

 Moreover, amino acids substitutions aiming at enhancing affinity for MHC I molecule must preserve the antigenicity of such optimized peptides. CTL generated

against optimized peptides must indeed cross-react with the corresponding native peptides, which are those naturally presented at the tumour cell surface.

The inventors have previously described methods for selection of cryptic peptides in tumour antigens and their optimization to induce specific immune response for patients HLA-A*0201 ((Tourdot et al., 2000), EP 1 309 860) and HLA-B*0702 (WO 2008/010098). A method for selecting HLA-A*2402-restricted cryptic epitopes has also been recently described by the inventors, in a patent application which has not been published yet. Briefly, this method consists in selecting, in an antigen, a peptide of 8 to 12 amino acids having a tyrosine in position 2, with the proviso that the peptide does not have, simultaneously, a positively charged amino acid (lysine or arginine) in position 1 and a leucine or an isoleucine or a phenylalanine in C-terminal position. Such a cryptic peptide can then be optimized by substituting its N-terminal residue with an arginine or a lysine, and/or by substituting its C-terminal residue with a leucine (or an isoleucine or a phenylalanine).

HLA-B*0702 is a frequently expressed molecule (25% of the population). Identification and optimization of HLA-B*0702 restricted tumour peptides is therefore necessary in order to develop efficient cancer vaccines for HLA-B*0702 expressing patients.

In order to identify a broad spectrum tumour vaccine for HLA-B*0702 expressing patients, the inventors have aligned the sequences of the MAGE-A antigens and searched for peptides having anchor positions 2 and 3 (respectively a proline and an arginine or a histidine or a methionine or a lysine) and an identical sequence in the region extending from positions P4 to P8 of the peptide. No corresponding sequence was found in conserved MAGE-A regions.

Sequences were then selected as having only one modification in the antigenicity region (position P4 to P8 in 9-mers, and P4 to P9 in 10-mers), and non-immunogenic epitopes were optimized as described in WO 2008/010098. Surprisingly, the inventors have demonstrated that a peptide corresponding to a cryptic HLA-B*0702 epitope modified to increase its antigenicity can raise a cytotoxic response not only against the native peptide, but also against the homologous epitope which is present on other MAGE-A antigens.

Hence, a first aspect of the present invention is a method for identifying a HLA-B*0702-restricted peptide which can trigger a cytotoxic response against at least two antigens from one single multigenic family, comprising at least the following steps:

(i) identifying, in the genes of said multigenic family, peptides of 9 or 10 amino acids having a P in position 2 and an amino acid selected in the group consisting of R, K, H and M in position 3;

(ii) aligning the sequences obtained in (i);

(iii) identifying, amongst the peptides obtained in step (i), a group of at least two peptides, in which at least one peptide is such that its antigenic region differs from those of the other peptides of the group by at most one residue, wherein said antigenic region extends from position 4 to position 8 in a peptide having 9 amino acids, and from position 4 to position 9 in a peptide having 10 amino acids.

A peptide which is such that its antigenic region differs from those of the other peptides of the group identified in step (iii) by at most one residue will be referred to hereafter as an “essentially shared peptide”. Such a peptide triggers a cytotoxic response against at least two antigens from said multigenic family.

According to preferred embodiments of said method, the method enables identification of a HLA-B*0702-restricted peptide which can trigger a cytotoxic response against at least three, four, five, six, seven or more antigens from said multigenic family. This is the case when the group of peptides selected in step (iii) comprises peptides from at least three, four, five, six, seven or more genes of said multigenic family, respectively.

In a particular embodiment of the above method, the group of peptides selected in step (iii) comprises at least two peptides which have different antigenic regions. In this case, illustrated in the examples below, at least two of these peptides exhibit one and only one difference in their antigenic regions.

In a preferred embodiment, the method further comprises a step (iv) of measuring the immunogenicity of the selected essentially shared peptide. This step will be preferentially performed *in vivo* in an appropriate model, *i.e.*, a model which predicts the immunogenicity of the peptide in an individual who expresses HLA-B*0702. An example of such an appropriate model is described in the experimental part and consists of a HLA-B*0702 transgenic mice. In this model, the immunogenicity of a selected peptide is measured by vaccinating the mice and testing if specific CTLs have been generated, by using human cells expressing HLA-B*0702 and loaded with the peptide as target cells. In what follows, a peptide will be considered as a non-immunogenic epitope if none of the vaccinated mice develop a specific immune response against the tested peptide. If some of the mice, but not all of them, develop a specific immune response against the tested peptide, the peptide is considered as immunogenic, but it can be advantageous to further improve its immunogenicity.

In case a selected essentially shared peptide is non-immunogenic or if its immunogenicity has to be enhanced, the method further comprises a step of increasing its immunogenicity, by a method as described in WO 2008/010098. In particular, if the selected essentially shared peptide is non-immunogenic and has any amino acid but P at its N-terminus (especially if the three first residues of said cryptic epitope are APR or APK or APH or APM), then step (v) consists of substituting the C-terminal residue of said epitope with a leucine. In case the selected essentially shared peptide is non-immunogenic and has

an amino acid selected amongst L, A, I, V, M, C or T (especially L, A, I, V or M) at its C-terminus, then step (v) can be performed by substituting the N-terminal residue of said epitope with an alanine. Of course, in this method, the word "substituting" is to be understood as obtaining a peptide the sequence of which is derived from the sequence of said HLA-B*0702-restricted cryptic epitope by the mentioned substitution, whatever the technical method used to obtain said peptide. For example, the peptide can be produced by artificial peptide synthesis or by recombinant expression.

The method according to the invention can be performed for identifying epitopes which can trigger an immunogenic response against several members of any known multigenic family, such as MAGE-A, HER, BAGE or GAGE families. In a preferred embodiment, illustrated in the experimental part below, said multigenic family is the MAGE-A family.

Another aspect of the present invention is an isolated peptide identified by a method as above-described, wherein said selected peptide is selected in the group consisting of MPKTGFLII (SEQ ID No: 2), MPKTGLLII (SEQ ID No: 3), FPKTGLLII (SEQ ID No: 4), VPKTGLLII (SEQ ID No: 5), MPKAGLLII (SEQ ID No: 6), MPKTGILIL (SEQ ID No: 7), MPKTGFLIIV (SEQ ID No: 8), MPKTGFLIII (SEQ ID No: 9), MPKTGLLIIV (SEQ ID No: 10), FPKTGLLIIV (SEQ ID No: 11), VPKTGLLIIV (SEQ ID No: 12), MPKAGLLIIV (SEQ ID No: 13), MPKTGILILI (SEQ ID No: 14), GPRALAETS (SEQ ID No: 15), GPRALIETS (SEQ ID No: 16), GPRALVETS (SEQ ID No: 17), GPRALAETSY (SEQ ID No: 18), GPRALIETSY (SEQ ID No: 19), GPRALVETSY (SEQ ID No: 20), EPRKLLTQD (SEQ ID No: 21), HPRKLLTQD (SEQ ID No: 22), DPKKLLTQH (SEQ ID No: 23), DPKKLLTQY (SEQ ID No: 24), HPKLLMQD (SEQ ID No: 25), EPRKLLTQDL (SEQ ID No: 26), EPRKLLTQDW (SEQ ID No: 27), HPRKLLTQDL (SEQ ID No: 28), HPKLLMQDL (SEQ ID No: 29), DPKKLLTQHF (SEQ ID No: 30), DPKKLLTQYF (SEQ ID No: 31).

Of course, in the present text, the term "isolated peptide" is not to be understood narrowly. To the contrary, this term designates not only molecules in which amino acid residues (in L or D configurations) are joined by peptide (-CO-NH-) linkages, but also synthetic pseudopeptides or peptidomimetics in which the peptide bond is modified, especially to become more resistant to proteolysis, and provided their immunogenicity is not impaired by this modification.

Immunogenic optimized peptides derived from the epitopes of the above list are also part of the present invention. In what follows, the expression "optimized peptide" or "optimized immunogenic HLA-B*0702-restricted epitope" will designate an immunogenic peptide derived from a HLA-B*0702-restricted epitope (called its "cognate native peptide") by a method as described above and in WO 2008/010098. Optimized

peptides according to the invention are peptides of SEQ ID Nos : 32 to 67, disclosed in Table 1 below.

Native peptides			MAGE-A corresponding antigenic sequence	Optimized peptide	
Name	Sequence	Seq ID n°		Sequence	Seq ID n°
MAGE-A 188 (9 mers)	MPKTGFLII	2	MAGE A1, A6	APKTGFLII	32
				MPKTGFLIL	33
	MPKTGLLII	3	MAGE A2, MAGE A4, MAGE A12	APKTGLLII	34
	FPKTGLLII	4		MPKTGLLIL	35
	VPKTGLLII	5		FPKTGLLIL	36
				VPKTGLLIL	37
	MPKAGLLII	6	MAGE A3	APKAGLLII	38
				MPKAGLLIL	39
	MPKTGILIL	7	MAGE A10	APKTGILIL	40
MAGE-A 188 (10 mers)	MPKTGFLIIV	8	MAGE A1, MAGE A6	APKTGFLIIV	41
	MPKTGFLIII	9		APKTGFLIII	42
				MPKTGFLIIL	43
	MPKTGLLIIV	10	MAGE A2, MAGE A4, MAGE A12	APKTGLLIIV	44
	FPKTGLLIIV	11		MPKTGLLIIL	45
	VPKTGLLIIV	12		FPKTGLLIIL	46
				VPKTGLLIIL	47
	MPKAGLLIIV	13	MAGE A3	APKAGLLIIV	48
				MPKAGLLIIL	49
	MPKTGILILI	14	MAGE A10	APKTGILILI	50
				MPKTGILILL	51
MAGE-A 267 (9 mers)	GPRALAETS	15	MAGE A1, A4	GPRALAETL	52
	GPRALIETS	16	MAGE A2, A6	GPRALIETL	53
	GPRALVETS	17	MAGE A3, A12	GPRALVETL	54
MAGE-A 267 (10 mers)	GPRALAETSY	18	MAGE A1, A4	GPRALAETSL	55
	GPRALIETSY	19	MAGE A2, A6	GPRALIETSL	56
	GPRALVETSY	20	MAGE A3, A12	GPRALVETSL	57
MAGE-A 233 (9 mers)	EPRKLLTQD	21	MAGE A1, A4, A10	EPRKLLTQL	58
	HPRKLLTQD	22	MAGE A12	HPRKLLTQL	59
	DPKLLTQH	23	MAGE A3	DPKLLTQL	60
	DPKLLTQY	24	MAGE A6	DPKLLTQL	61
	HPKLLMQD	25	MAGE A2	HPKLLMQL	62
MAGE-A 233 (10 mers)	EPRKLLTQDL	26	MAGE A1	EPRKLLTQDL	63
	EPRKLLTQDW	27	MAGE A4, A10	APRKLLTQDL	64
	HPRKLLTQDL	28	MAGE A12		
	HPKLLMQDL	29	MAGE A2	APKLLMQDL	65
	DPKLLTQHF	30	MAGE A3	DPKLLTQHL	66
	DPKLLTQYF	31	MAGE A6	DPKLLTQYL	67

Table 1: HLA-B7 restricted native and corresponding optimized peptides highly homologous amongst MAGE-A antigens (antigenic sequences are highlighted)

5 Polyspecific tumour vaccination offers a broader control of tumour cells than monospecific vaccination, thereby reducing the risk of emergence of immune escape variants. In most cases, immunotherapy is then more efficient when targeting several epitopes than when targeting only one epitope, provided the tumour is known to express all targeted antigens. The inventors have previously described a polypeptide composed of

10 HLA-A*0201 restricted optimized cryptic peptides derived from three different universal tumour antigens (TERT_{988Y}, HER-2/neu_{402Y} and MAGE-A_{248V9}), named Vx-006

(WO 2007/073768). Vx-006 is able to induce a polyspecific CD8 cell response both *in vivo* in HLA-A*0201 transgenic HHD mice and *in vitro* in humans, whereas the mixture of TERT_{988Y}, HER-2/neu_{402Y} and MAGE-A_{248V9} peptides failed to induce a trispecific response. Hence, a chimeric polypeptide comprising several epitopes can be more efficient
 5 than a mere mixture of the same epitopes to trigger a response against more than one epitope. Depending on the context, a chimeric polypeptide comprising a repetition of one single epitope can also trigger a stronger response against said epitope than a peptide consisting of said epitope. Indeed, a polypeptide organization (either with several different epitopes or with a repetition of one single epitope) can produce new junctional epitopes,
 10 especially CD4 restricted epitopes, able to optimize the targeted peptide(s)-specific immune response. Moreover, when free peptides are subcutaneously injected, peptides bind directly to MHC molecules of every cells present at the site of injection. As polypeptides need to be processed, vaccination with polypeptides is more efficient to target antigenic peptides to professional Antigenic Presenting Cells (APC) as Dendritic Cells.

15 A further aspect of the invention is hence a chimeric polypeptide, comprising one, two, three or more HLA-B*0702-restricted epitopes as above-described. In particular, a chimeric polypeptide according to the invention can comprise one, two, three or more native HLA-B*0702-restricted epitopes described above, or one, two, three or more immunogenic optimized HLA-B*0702-restricted epitopes selected amongst
 20 SEQ ID Nos: 32-67. Of course, optimized HLA-B*0702-restricted epitopes can also be combined, in a chimeric polypeptide, to native HLA-B*0702-restricted epitopes which have been identified as immunogenic epitopes. In a chimeric polypeptide according to the invention, the epitopes can be different from each other, and/or the same epitope can be repeated several times.

25 It is to be noted that when several epitopes specific for the same HLA molecule are used together, either in a mix or in a chimeric polypeptide, the epitopes are in competition for the binding to the corresponding HLA molecule. Contrarily, by using a mix of different HLA-restricted epitopes (HLA-A*0201, HLA-A*2402, HLA-B*0702 or others), or a chimeric polypeptide comprising the same different HLA-restricted epitopes,
 30 there will be no competition for HLA binding, and a polyspecific response will be obtained with certainty, provided all the HLA molecules are expressed in the vaccinated individual.

In a chimeric polypeptide according to the invention, HLA-B*0702-restricted cryptic or immunogenic (native or optimized) epitopes, described above, can hence be advantageously associated to previously described HLA-A*0201 (WO 02/02716)
 35 and/or HLA-B*0702 peptides (WO 2008/010010 and WO 2008/010098), and/or to HLA-A*2402 peptides as disclosed in Table 2 below, and/or to immunogenic epitopes derived from previously described tumour associated antigens, comprising CEA, PRAME, Tyrosinase, TRAG-3, NY-Eso-1, P53, Muc-1, PSA/PSMA, survivin, Melan-A/MART-1,

TRP-1, TRP-2, WT1, EphA1, EphA2, EphA3, EphA4, G250/MN/CAIX, STEAP, alphafoetoprotein, RAGE-1, PAGE-1. Of course, a polyallelic peptides mix, comprising at least a peptide according to the present invention and one different HLA-restricted epitope (HLA-A*0201, HLA-A*2402, HLA-B*0702 or others), is also part of the present invention.

Examples of epitopes which can advantageously be combined to HLA-B*0702-restricted MAGE-A epitopes (either in a mix or in a chimeric polypeptide), as well as examples of optimized immunogenic epitopes which can advantageously be combined to (native or optimized) immunogenic HLA-B*0702-restricted MAGE-A epitopes, are described in Table 2 below. Of course, these lists are not limitative.

HLA-A*0201					
Native peptide			Optimized peptide		
Antigen	Sequence	No	Name	Sequence	No
Mart-1 ₂₇	AAGIGILTV	68	Mart-1 ₂₇ Y1	YAGIGILTV	112
Mart-1 ₂₆	EAAGIGILTV	69	Mart-1 ₂₆ L27	ELAGIGILTV	113
Gp100 ₁₇₇	AMLGTHTMEV	70	Gp100 ₁₇₇ Y1	YMLGTHTMEV	114
Gp100 ₁₇₈	MLGTHTMEV	71	Gp100 ₁₇₈ Y1	YLGTHTMEV	115
Gp100 ₁₅₄	KTWGQYWQV	72	Gp100 ₁₅₄ Y1	YTWGQYWQV	116
			Gp100 ₁₅₄ M155	KMWGQYWQV	117
Gp100 ₅₇₀	SLADTNSLAV	73	Gp100 ₅₇₀ Y1	YLADTNSLAV	118
Gp100 ₂₀₉	TDQVPFSV	74	Gp100 ₂₀₉ Y1	YDQVPFSV	119
			Gp100 ₂₀₉ M210	YMQVPFSV	120
Gp100 ₄₇₆	VLYRYGSFSV	75	Gp100 ₄₇₆ Y1	YLYRYGSFSV	121
Gp100 ₄₅₇	LLDGTATLRL	76	Gp100 ₄₅₇ Y1	YLDGTATLRL	122
HER-2/neu ₇₉₉	QLMPYGCLL	77	HER-2/neu ₇₉₉ Y1	YLMPYGCLL	123
HER-2/neu ₃₆₉	KIFGSLAFL	78	HER-2/neu ₃₆₉ Y1	YIFGSLAFL	124
HER-2/neu ₇₈₉	CLTSTVQLV	79	HER-2/neu ₇₈₉ Y1	YLTSTVQLV	125
HER-2/neu ₄₈	HLYQGCQW	80	HER-2/neu ₄₈ Y1	YLYQGCQW	126
HER-2/neu ₇₇₃	VMAGVGSPYV	81	HER-2/neu ₇₇₃ Y1	YMAGVGSPYV	127
HER-2/neu ₅	ALCRWGLL	82	HER-2/neu ₅ Y1	YLCRWGLL	128
HER-2/neu ₈₅₁	VLVKSPNHV	83	HER-2/neu ₈₅₁ Y1	YLVKSPNHV	129
HER-2/neu ₆₆₁	ILLVVVLGV	84	HER-2/neu ₆₆₁ Y1	YLLVVVLGV	130
HER-2/neu ₆₅₀	PLTSIISAV	85	HER-2/neu ₆₅₀ Y1	YLTSIISAV	131
HER-2/neu ₄₆₆	ALIHNTHL	86	HER-2/neu ₄₆₆ Y1	YLIHNTHL	132
HER-2/neu ₄₀₂	TLEEITGYL	87	HER-2/neu ₄₀₂ Y1	YLEEITGYL	133
HER-2/neu ₃₉₁	PLQPEQLQV	88	HER-2/neu ₃₉₁ Y1	YLQPEQLQV	134
HER-2/neu ₉₇₁	ELVSEFSRM	89	HER-2/neu ₉₇₁ Y1	YLVSEFSRM	135
EphA2 ₆₁	DMPIYMYSV	90	EphA2 ₆₁ Y1	YMPIYMYSV	136

HER2 ₉₁₁	TVWELMTFGA	91	HER _{911Y1V10}	YVWELMTFGV	137
HER4 ₉₁₁	TIWELMTFGG	92			
HER1 ₉₁₁	TVWELMTFGS	93			
HER2 ₇₂₂	KVKVLGSGA	94	HER _{722Y1V9}	YVKVLGSGV	138
HER3 ₇₂₂	KLKVLGSGV	95			
HER4 ₇₂₂	RVKVLGSGA	96			
HER1 ₇₂₂	KIKVLGSGA	97	HER _{845Y1}	YLAARNVLV	139
HER2 ₈₄₅	DLAARNVLV	98			
HER3 ₈₄₅	NLAARNVLL	99			
HER2 ₉₀₄	DVWSYGVTV	100	HER _{904Y1}	YVWSYGVTV	140
HER4 ₉₀₄	DVWSYGVTI	101			
HER2 ₉₃₃	DLLEKGERL	102	HER _{933Y1}	YLLEKGERL	141
HER1 ₉₃₃	SILELKGERL	103			
HER2 ₉₄₅	PICTIDVYMI	104	HER _{945Y1}	YICTIDVYMV	142
HER3 ₉₄₅	QICTIDVYMV	105			
HER4 ₉₄₅	PICTIDVYMV	106			
HER1 ₉₄₅	PICTIDVYKI	107			
MAGE-A _{248G9}	YLEYRQVPG	108	MAGE-A _{248V9}	YLEYRQVPV	143
MAGE-A _{248D9}	YLEYRQVPD	109			
TERT ₉₈₈	DLQVNSLQTV	110	TERT _{988Y1}	YLQVNSLQTV	144
TERT ₅₇₂	RLFFYRKSV	111	TERT _{572Y1}	YLFFYRKSV	145
HLA-B*0702					
Native peptide			Optimized peptide		
Name	Sequence	No	Name	Sequence	No
TERT ₄₄₄	DPRRLVQLL	146	TERT _{444A1}	APRRLVQLL	151
CEA _{188/554}	SPRLQLSNG	147	CEA _{188/554L9}	SPRLQLSNL	152
HER-2/neu ₁₀₆₉	APRSPLAPS	148	HER-2/neu _{1069L9}	APRSPLAPL	153
HER-2/neu ₇₆₀	SPKANKEIL	149	HER-2/neu _{760A1}	APKANKEIL	154
HER-2/neu ₂₄₆	GPKHSDCLA	150	HER-2/neu _{246A1}	APKHSDCLA	155
HLA-A*2402					
Native peptide			Optimized peptide		
Name	Sequence	No	Name	Sequence	No
TERT 403	PYGVLLKTH	156	TERT 403 _{K1L9}	KYGVLLKTL	165
TERT 770	PYMRQFVAH	157	TERT 770 _{R1L9}	RYMRQFVAL	166
HER 780	PYVSRLLLGI	158	HER 780 _{R1}	RYVSRLLLGI	167
EphA2 47	PYGKGWDLM	159	EphA2 47 _{R1L9}	RYGKGWDLL	168

EphA2 502	TYLVQVQAL	160	EphA2 502 _{R1}	RYLVQVQAL	169
EphA2 817	PYWELSNHE	161	EphA2 817 _{RIL9}	RYWELSNHL	170
Her2/neu 922	PYDGIPARE	162			
MAGE 261	RYEFLWGPR	163			
Her2/neu 300	PYNYLSTDV	164			

Table 2: HLA-A2, -B7 and -A24 epitopes which can be combined to HLA-B*0702-restricted MAGE-A epitopes in chimeric polypeptides according to the invention

The skilled artisan can chose any known technique to produce such polypeptides. For example, the polypeptide can be obtained by chemical synthesis, or by using the technology of genetic engineering (Velders et al., 2001).

Another object of the present invention is an isolated nucleic acid molecule designed to cause the expression of a cryptic HLA-B*0702-restricted MAGE-A epitope, or of an immunogenic HLA-B*0702-restricted MAGE-A epitope (either native or optimized), or of a chimeric polypeptide as above-described. By “designed to cause the expression of” a peptide is herein meant that said peptide is expressed as such, isolated from the whole antigen from which its sequence has been selected (and, in appropriate cases, optimized as above-described), when the nucleic acid is introduced in an appropriate cell. The region encoding the epitope or chimeric polypeptide will typically be situated in the polynucleotide under control of a suitable promoter. Bacterial promoters will be preferred for expression in bacteria, which can produce the polypeptide either *in vitro*, or, in particular circumstances, *in vivo*. An example of bacterium that can be used to produce a peptide or polypeptide according to the invention, directly *in vivo*, is *Listeria monocytogenes*, which is a facultative intracellular bacterium that enters professional antigen-presenting cells by active phagocytosis (Paterson and Maciag, 2005). Alternatively, a nucleic acid according to the invention can be administered directly, using an appropriate vector. In this case, a tissue-specific, a strong constitutive, or an endogenous promoter can be used to control the peptide expression. Suitable vector systems include naked DNA plasmids, liposomal compositions to enhance delivery, and viral vectors that cause transient expression. Examples of viral vectors are adenovirus or vaccinia virus vectors and vectors of the herpes family, especially in a non-replicative form.

The present invention also pertains to a pharmaceutical composition comprising at least, as an active principle, a HLA-B*0702-restricted MAGE-A cryptic epitope as above-described, or an immunogenic (optimized or native) HLA-B*0702-restricted MAGE-A epitope as mentioned above, or a chimeric polypeptide according to the invention, or a nucleic acid encoding any of these, and/or a vector carrying said nucleic acid. Formulation of pharmaceutical compositions will accord with contemporary standards and techniques. Medicines intended for human administration will be prepared in adequately sterile conditions, in which the active ingredient(s) are combined with an

isotonic solution or other pharmaceutical carrier appropriate for the recommended therapeutic use. Suitable formulations and techniques are generally described in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing Co, Easton PA).

5 In particular, a HLA-B*0702-restricted MAGE-A epitope or a chimeric polypeptide or a nucleic acid according to the invention can be used for the preparation of a composition for preventive or curative anti-cancer immunotherapy. The peptide GPRALVETL (SEQ ID No: 54), and chimeric polypeptides comprising it, are especially suited for this purpose.

10 In a particular embodiment, a pharmaceutical composition according to the invention is a vaccine. In this latter case, the components described above can be combined with an adjuvant to potentiate the immune response. Classic adjuvants include oil emulsions, like Incomplete Freund's Adjuvant or Montanide, and adherent surfaces such as alum. Adjuvants that recruit and activate dendritic cells particularly via TLR (such as bacterial DNA or bacterial membrane derived proteins) or help elicit cytotoxic T cells
15 are especially useful. Other factors that otherwise boost the immune response or promote apoptosis or elimination of cancer cells can also be included in the composition, such as IL-2 or IL-12 cytokines or GM-CSF.

Multiple doses and/or different combinations of the immunogenic compositions of this invention can be packaged for distribution separately or together.
20 Each composition or set of compositions, such as the kits of parts described below, can be accompanied with written instructions regarding the use of the composition or combination for eliciting an immune response and/or for the treatment of cancer.

In a previous patent application (WO 2006/120038), the Applicant has described a vaccination protocol which enables the initiation and maintenance of a T cell
25 response targeting sub-dominant/cryptic epitopes. The results reported in WO 2006/120038 demonstrate that injection of a native peptide corresponding to a sub-dominant/cryptic epitope, following vaccination with its cognate optimized peptide, can maintain the immune response initiated by said optimized peptide.

According to the invention, a HLA-B*0702-restricted MAGE-A cryptic
30 epitope can hence be used for the preparation of a medicinal composition for maintaining the CTL immune response initiated by its cognate optimized peptide. An immunogenic peptide having an optimized immunogenic HLA-B*0702-restricted MAGE-A epitope sequence derived from a HLA-B*0702-restricted MAGE-A cryptic epitope can also be used, for the preparation of a medicinal composition for initiating a CTL immune response
35 against said HLA-B*0702-restricted MAGE-A cryptic epitope, but also against all the epitopes of the group selected in step (iii) of the above-described method. Of course, mix of peptides from the group selected in step (iii) can also be used for maintaining the CTL immune response initiated by the essentially shared peptide. For example, a mix of

peptides SEQ ID No: 15-17 can be used for maintaining the CTL immune response initiated by the peptide of SEQ ID No: 54.

The present invention also encompasses a method for vaccinating a patient against a tumoral or viral antigen, wherein said method comprises a first step of vaccination with an optimized immunogenic peptide cognate to a native HLA-B*0702-restricted MAGE-A cryptic epitope of said antigen or epitopes of the group selected in step (iii), followed by a second step of vaccination with said native peptide or mix of peptides of the considered group.

In such a method, the first step and/or the second step can be performed by using a chimeric polypeptide comprising one, two, three or more optimized or cryptic peptides as above-described, instead of single-epitope peptides. In particular, a chimeric polypeptide comprising several cryptic epitopes having at most one variant position in their antigenic region, can be used to maintain the CTL immune response initiated by optimized peptide cognate to one of said cryptic epitopes. For example, a chimeric polypeptide comprising the sequences SEQ ID No: 15-17 can be used for maintaining the CTL immune response initiated by the peptide of SEQ ID No: 54. It is to be noted that due to the expression tropism of MAGE-A antigens, if a HLA-B*0702-restricted epitope as described above proves to be immunogenic, the same native immunogenic epitope can be used in both vaccination steps. In particular, a native immunogenic MAGE-A epitope can advantageously be combined with native cryptic epitopes in a first chimeric polypeptide or mix of mono-epitope peptides, and with optimized epitopes, in a second chimeric polypeptide or mix of mono-epitope peptides.

The invention also pertains to a kit of parts comprising, in separate formulations or containers (vials, tubes, etc.):

(i) a first peptide comprising a sequence of a HLA-B*0702-restricted MAGE-A native (preferably cryptic) epitope, and

(ii) a second peptide comprising a sequence corresponding to an optimized immunogenic epitope cognate to the native epitope recited in (i).

Examples of peptides which can be part of a kit according to the invention are the peptides of SEQ ID Nos: 2 to 31 which can constitute the first peptide, the second peptide being then derived from said first peptide by a method for increasing its immunogenicity, as described above and in WO 2008/010098. A preferred kit according to the invention comprises the peptide of SEQ ID No: 54 and, in another container, the peptide of SEQ ID No: 17 or 15 or 16, preferentially the peptide of SEQ ID No: 17. In a variant of this kit, the kit also comprises peptides of SEQ ID Nos: 16 and/or 15, either in the same container as SEQ ID No: 17, or in one or several separate container(s).

Other kits of parts according to the invention comprise at least one chimeric polypeptide. In this embodiment, the kit also comprises at least a peptide cognate

to one of the epitopes comprised in the chimeric polypeptide, wherein said cognate peptide is either isolated or included in another chimeric polypeptide.

Several preferred variants of such kits are contemplated: in a first embodiment, the kit comprises, in separate formulations, a first chimeric polypeptide comprising one, two, three or more HLA-B*0702-restricted MAGE-A cryptic epitopes, and a second chimeric polypeptide corresponding to its cognate HLA-B*0702-restricted MAGE-A immunogenic chimeric polypeptide (which means that it comprises optimized HLA-B*0702-restricted MAGE-A immunogenic epitopes cognate to the cryptic epitopes comprised in the first chimeric polypeptide). In a second embodiment, the kit comprises one, two, three or more peptides corresponding to distinct HLA-B*0702-restricted MAGE-A cryptic epitopes, wherein said peptides are either mixed in one single formulation, or separated in several formulations and, in a separate formulation, a chimeric polypeptide comprising the optimized HLA-B*0702-restricted MAGE-A immunogenic epitopes cognate to said cryptic peptides.

As mentioned above, a polyallelic stimulation (*i.e.*, using epitopes specific for different HLA molecules) can advantageously be performed to obtain a polyspecific response. Accordingly, preferred embodiments of the kits according to the invention comprise, in separate containers:

(i) a polyallelic peptides mix or a polyallelic chimeric polypeptide, comprising at least a HLA-B*0702-restricted MAGE-A native (preferably cryptic) epitope as described above and at least one different HLA-restricted native (preferably cryptic) epitope (from and antigen of the MAGE-A family or from another antigen), and

(ii) a polyallelic peptides mix or a polyallelic chimeric polypeptide, comprising at least a HLA-B*0702-restricted MAGE-A immunogenic epitope cognate to the HLA-B*0702-restricted MAGE-A native epitope recited in (i), and at least another immunogenic epitope cognate to the other native epitope recited in (i).

Alternatively, the kits according to the invention can comprise, instead of at least part the peptides or chimeric polypeptides, nucleic acid(s) encoding said peptides or chimeric polypeptides. In this case, the nucleic acid(s) is(are) as above-described.

In the following description of some specific kits according to the invention, mention will be made only of the peptides (native or optimized) included therein; it is understood that chimeric polypeptide(s) (comprising native cryptic epitopes or optimized epitopes) can be enclosed in the kits instead of single-epitope peptides, and that nucleic acid(s) can also be included in addition or instead of at least part of said peptides or chimeric polypeptides.

In a particular embodiment of the invention, the kit is a vaccination kit, wherein said first (native) and second (cognate optimized) peptides are in separate vaccination doses. In a preferred embodiment, the vaccination kit comprises 2 or 3 doses of

optimized peptide, and 3, 4, 5 or 6 doses of native peptide. A particular vaccination kit according to the invention is adapted for the first vaccination sequence of 6 injections, and comprises 2 or 3 doses of optimized peptide, and 4 or 3 doses of native peptide. In case of long-lasting diseases, it is preferable to maintain the level of immunity obtained after this
5 primo-vaccination, by regular recalls. This can be done, for example, by injections performed every 1 to 6 months. Therefore, complementary kits, comprising at least 2 doses, and up to 40 or 50 doses of native peptide, are also part of the present invention. Alternatively, the vaccination kit can comprise 2 to 3 doses of optimized peptide, and 3 to 40 or up to 50 doses of native peptide. Of course, said native and optimized peptides
10 present in the kit are as described above.

Each dose comprises between 0.1 and 10 mg of peptide, preferably from 1 to 5 mg, or between 1 and 20 mg of polypeptide. In a preferred embodiment, each dose is formulated for subcutaneous injection. For example, each dose can be formulated in 0.3 to 1.5 ml of an emulsion of aqueous solution emulsified with Montanide ISA51, used as an
15 adjuvant. The skilled artisan can choose any other adjuvant(s) in place of (or in addition to) Montanide ISA51. In a particular embodiment, the doses are in the form of an aqueous solution. Alternatively, the doses can be in the form of a lyophilized peptide, for extemporaneous preparation of the liquid solution to be injected. Other possible components of said kits are one or several adjuvants, to be added to the peptide
20 compositions before administration, and a notice describing how to use said kits.

The invention is further illustrated by the following figures and examples.

LEGENDS OF FIGURES

Figure 1 : MAGE-A multigene family sequences. In order to identify
25 one or more epitopes shared by the various MAGE-A antigens and presented by the HLA-B*0702 molecule, the sequences of the MAGE-A antigens were aligned, and regions of at least 5 amino acids were selected on the basis of their homology between these antigens (boxed in black continuous line). Amino acids that are completely identical from MAGE-A1, -A2, -A3, -A4, -A6, -A12 and/or -A10 are highlighted in grey.

**Figure 2: Immunogenicity of HLA-B*0702 restricted optimized
30 cryptic peptides.** HLA-B*0702 transgenic mice were vaccinated with the optimized peptides following the described protocol and generated CTL were tested against T2-B7 targets loaded with the optimized and both corresponding native peptides as indicated. A; Vaccination with the MAGE-A A1L9 peptide of SEQ ID No:171, B; Vaccination with the
35 monomodified MAGE A L9 peptide of SEQ ID No: 54.

EXAMPLES

The examples have been performed using the following materials and methods:

Transgenic Mice. The HLA-B7 H-2 class-I knockout mice were previously described (Rohrlich et al., 2003).

Cells. HLA-B*0702 transfected human T2-B7 cells were previously described (Rohrlich et al., 2003).

Peptides and Plasmids. Peptides were synthesized by Epytop (Nîmes, France). HLA-B*0702 plasmid was provided by Dr. Lemonnier (Institut Pasteur, Paris, France) (Rohrlich et al., 2003).

Measurement of Peptide Relative Affinity to HLA-B*0702. The protocol used has been described previously (Rohrlich et al., 2003). Briefly, T2-B7 cells were incubated at 37°C for 16 hours with peptides concentrations ranging from 100 µM to 0.1 µM, and then stained with ME-1 monoclonal antibody (mAb) to quantify the surface expression of HLA-B*0702. For each peptide concentration, the HLA-B*0702 specific staining was calculated as the percentage of staining obtained with 100 µM of the reference peptide CMV₂₆₅₋₂₇₄ (R10V; RIPHERNGFTV, SEQ ID NO: 172). The relative affinity (RA) was determined as: $RA = (\text{Concentration of each peptide that induces 20 \% of HLA-B*0702-expression} / \text{Concentration of the reference peptide that induces 20 \% of HLA-B*0702 expression})$.

CTL Induction in vivo in HLA-B*0702 Transgenic Mice. Mice were injected subcutaneously with 100 µg of peptide emulsified in Incomplete Freund's Adjuvant (IFA) in the presence of 150 µg of the I-A^b restricted HBVcore₁₂₈ T helper epitope (TPPAYRPPNAPIL, SEQ ID NO: 173). After 11 days, 5×10^7 spleen cells were stimulated *in vitro* with peptide (10 µM). On day 6 of culture, the bulk responder populations were tested for specific cytotoxicity.

Cytotoxic assay. Targets were labelled with 100 µCi of Cr⁵¹ for 60 min, plated in 96-well V-bottomed plates (3×10^3 cell/well in 100 µL of RPMI 1640 medium) and, when necessary, pulsed with peptides (1 µM) at 37°C for 2 hours. Effectors were then added in the wells and incubated at 37°C for 4 hours. Percentage of specific lysis was determined as: $\% \text{ Lysis} = (\text{Experimental Release} - \text{Spontaneous Release}) / (\text{Maximal Release} - \text{Spontaneous Release}) \times 100$.

Example 1: identification of cryptic epitopes presented by the HLA-B*0702 molecule that are shared by the MAGE-A1, -A2, -A3, -A4, -A6, -A12 and/or -A10 antigens, and determination of their affinities with said HLA molecule

In order to identify one or more epitopes shared by the various MAGE-A antigens and presented by the HLA-B*0702 molecule, the sequences of the MAGE-A antigens were aligned (figure 1), and regions of 9 to 10 amino acids were searched on the

basis of their homology between MAGE -A1, -A2, -A3, -A4, -A6, -A12 and/or -A10 antigens (sequences highlighted in grey, figure 1). As MAGE -A10 sequence is less homologous to MAGE-A1, -A2, -A3, -A4, -A6, -A12, shared sequences were not eliminated if no equivalent was found in MAGE-A10 (figure 1).

5 In the following description, these regions of 9 to 10 amino acids are denoted with reference to the position of their first amino acid in the MAGE-A1 sequence. Only two regions of at least 9 amino acids were identified (position 181 and 270). As previously described, as few homologous sequences exist, authors described a method to identify a sequence of 8 to 10 amino acids comprising at least one common pentapeptide
10 sequence preceded by 3 amino acids at the N-terminal end and, optionally, followed by one or two amino acids at the C-terminal end; indeed, the authors have found that an identity limited to the sequence of 5 amino acids extending from positions P4 to P8 of the peptide was sufficient. Sequences of at least 5 common amino acids are boxed in figure 1. Using this method of selection, four additional regions were indentified (position 21, 65, 132,
15 256).

Peptides of 9 or 10 amino acids having a P in position 2 and an amino acid selected in the group consisting of R, K, H and M in position 3 corresponding to HLA-B*0702 restricted peptides were then identified. As shown in figure 1, no sequence completely identical was found.

20 In order to broaden the choice of the candidate peptides, a second search was carried out, according to the described method, to select regions exhibiting complete sequence identity between positions P4 and P8. One more time, no sequence was indentified. Finally, a third search was performed, to select sequences having only one mismatch between positions P4 and P8. Identified sequences are in table 1 above, and are
25 boxed in dotted line in figure 1.

The MAGE-A 269 (9mers) group was selected as only three different sequences allow recognizing all the MAGE-A genes (accept MAGE-A10). This group comprises three peptides: MAGE-A A, SEQ ID No15 (MAGE-A1, -A4), MAGE-A I, SEQ ID No16 (MAGE-A2, -A6) and MAGE-A V, SEQ ID No17 (MAGE-A3, -A12), which
30 differ in terms of their position P6. No corresponding sequence was found in MAGE-A10.

Each peptide was tested for its capacity to bind HLA-B*0702 (table 3).

Peptide	Sequence	RA	SEQ ID No
MAGE-A A	GPRALAETS	-	15
MAGE-A I	GPRALIETS	-	16
MAGE-A V	GPRALVETS	-	17

Table 3. Affinity of the selected cryptic peptides to HLA-B*0702.

35 **RA = Relative Affinity = (Concentration of each peptide that induces 20 % of HLA-B*0702-expression / Concentration of the reference peptide that induces 20 % of HLA-B*0702 expression), (-) means RA>10, (+/-) 1<RA<10, (+) 5<RA<10, (++) RA <1**

None of the three native peptides was shown to bind to HLA-B*0702 molecules, despite the fact that these peptides harbour primary P2R3 anchor positions, showing that they are cryptic peptides. The aim of this study was to find an immunogenic peptide that is capable to induce a specific immune response able to recognize a cell
5 whatever the MAGE-A gene expressed. More precisely, CTL induced by the vaccination with the validated peptide, have to be able to recognize a cell which expresses or presents both MAGE-A A, MAGE-A I and MAGE-A V cryptic native peptide (native peptide cross recognition). Selected peptides were then modified to enhance their immunogenicity.

Example 2: enhancement of the immunogenicity of the selected peptide

10 To enhance HLA-B*0702 affinity and consequently immunogenicity of these low affinity peptides, it is necessary to identify unfavourable secondary anchor motifs and substitute them with favourable motifs. Native peptides were selected to have the P2R3 primary anchor positions; the interest was then focused on secondary anchor position 1 and 9.

15 The first optimized peptide tested was based on the MAGE-A V sequence, modified at both positions respectively by replacing the P1 by an alanine (A) and the P9 by a leucine (L), known to be amino acids favourable for HLA-B*0702 binding.

20 The peptide MAGE-A A1L9 has the sequence APRALVETL (SEQ ID n°171), and was able to bind to MHC (Table 4), confirming that modifications have enhanced its affinity for HLA-B*0702 molecules. HLA-B*0702 transgenic mice were then vaccinated with the modified peptide, and eleven days later, their spleen cells were *in vitro* stimulated with the peptide. As shown in figure 2A and table 4, the modified peptide was immunogenic but MAGE-A A1L9 specific CTLs induced were not able to cross-recognize
25 the native peptides.

The substitutions should however preserve the conformation of the peptide segment that interacts with the TCR, preserving the peptide specificity. As two modifications could modify dramatically the peptide conformation, a new optimized peptide was tested, only modified at position 9. Indeed, a G at position 1 is described as
30 neutral and non unfavourable for the peptide affinity to MHC.

MAGE-A L9 (SEQ ID No: 54) was shown to be strongly immunogenic, as all vaccinated mice developed a specific immune response against the MAGE-A L9. Most importantly, CTLs induced by the MAGE-A L9 peptide were able to recognize a target cell loaded with each of the native cryptic peptides (figure 2B and table 4).

Peptide	Sequence	RA	Immuno genicity	Native peptide crossrecognition	SEQ ID No
MAGE-A A	GPRALAETS	-			15
MAGE-A I	GPRALIETS	-			16
MAGE-A V	GPRALVETS	-			17
MAGE-A A1L9	APRALVETL	+	6/11	MAGE-A A (1/8) MAGE-A I (0/3) MAGE-A V (0/3)	171
MAGE-A L9	GPRALVETL	ND	18/18	MAGE-A A (3/8) MAGE-A I (3/5) MAGE-A V (4/5)	54

Table 4: affinity and immunogenicity of the optimized peptides.

RA = Relative Affinity = (Concentration of each peptide that induces 20 % of HLA-B*0702-expression / Concentration of the reference peptide that induces 20 % of HLA-B*0702 expression), (-) means RA>10, (+/-) 1<RA<10, (+) 5<RA<10, (++) RA <1

5 (X/Y) means that X mice developed a specific response for a total of Y mice vaccinated.

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CLAIMS

1. A method for identifying a HLA-B*0702-restricted peptide which can trigger a cytotoxic response against at least two antigens from one single multigenic family, comprising at least the following steps:

5 (i) identifying, in the genes of said multigenic family, peptides of 9 or 10 amino acids having a P in position 2 and an amino acid selected in the group consisting of R, K, H and M in position 3;

(ii) aligning the sequences obtained in (i);

10 (iii) identifying, amongst the peptides obtained in step (i), a group of at least two peptides, in which at least one peptide is an “essentially shared peptide”, *i.e.*, is such that its antigenic region differs from those of the other peptides of the group by at most one residue, wherein said antigenic region extends from position 4 to position 8 in a peptide having 9 amino acids, and from position 4 to position 9 in a peptide having 10 amino acids;

15 wherein said at least one essentially shared peptide triggers a cytotoxic response against at least two antigens of said multigenic family.

2. The method of claim 1, for identifying a HLA-B*0702-restricted peptide which can trigger a cytotoxic response against at least three antigens from said multigenic family, wherein the group of peptides selected in step (iii) comprises peptides
20 from at least three genes of said multigenic family.

3. The method of claim 1 or claim 2, wherein the group of peptides selected in step (iii) comprises at least two peptides which have different antigenic regions.

4. The method according to any of claims 1 to 3, further comprising a step (iv) of measuring the immunogenicity of the essentially shared peptide.

25 5. The method of claim 4, further comprising a step (v) of increasing the immunogenicity of the essentially shared peptide.

30 6. The method of claim 5, wherein the essentially shared peptide selected in step (iii) is a non-immunogenic epitope with any amino acid but P at its N-terminus, and wherein step (v) consists of substituting the C-terminal residue of said epitope with a leucine.

7. The method of claim 5, wherein the essentially shared peptide selected in step (iii) is a non-immunogenic epitope with a C-terminal amino acid selected in the group consisting of L, A, I, V, M, C and T, and wherein step (v) consists of substituting the N-terminal residue of said epitope with an alanine.

35 8. The method of any of claims 1 to 7, wherein said multigenic family is the MAGE-A family.

9. An isolated peptide identified by a method according to any of claims 1 to 3, wherein said essentially shared peptide is selected in the group consisting of

MPKTGFLII (SEQ ID No: 2), MPKTGLLII (SEQ ID No: 3), FPKTGLLII (SEQ ID No: 4),
 VPKTGLLII (SEQ ID No: 5), MPKAGLLII (SEQ ID No: 6), MPKTGILIL (SEQ ID
 No: 7), MPKTGFLIIV (SEQ ID No: 8), MPKTGFLIII (SEQ ID No: 9), MPKTGLLIIV
 (SEQ ID No: 10), FPKTGLLIIV (SEQ ID No: 11), VPKTGLLIIV (SEQ ID No: 12),
 5 MPKAGLLIIV (SEQ ID No: 13), MPKTGILILI (SEQ ID No: 14), GPRALAETS (SEQ
 ID No: 15), GPRALIETS (SEQ ID No: 16), GPRALVETS (SEQ ID No: 17),
 GPRALAETSY (SEQ ID No: 18), GPRALIETSY (SEQ ID No: 19), GPRALVETSY
 (SEQ ID No: 20), EPRKLLTQD (SEQ ID No: 21), HPRKLLTQD (SEQ ID No: 22),
 DPKKLLTQH (SEQ ID No: 23), DPKKLLTQY (SEQ ID No: 24), HPKLLMQD (SEQ
 10 ID No: 25), EPRKLLTQDL (SEQ ID No: 26), EPRKLLTQDW (SEQ ID No: 27),
 HPRKLLTQDL (SEQ ID No: 28), HPKLLMQDL (SEQ ID No: 29), DPKKLLTQHF
 (SEQ ID No: 30), DPKKLLTQYF (SEQ ID No: 31).

10. An isolated peptide identified by a method according to any of claims
 5 to 7, wherein said isolated peptide is selected in the group consisting of SEQ ID NOS: 32
 15 to 67.

11. A chimeric polypeptide, comprising one, two, three or more HLA-
 B*0702-restricted epitopes according to claim 9.

12. A chimeric polypeptide, comprising one, two, three or more HLA-
 B*0702-restricted epitopes according to claim 10.

20 13. An isolated nucleic acid molecule designed to cause the expression of
 a HLA-B*0702-restricted epitope according to claim 9 or claim 10, or a chimeric
 polypeptide according to claim 11 or claim 12.

25 14. A pharmaceutical composition comprising at least, as an active
 principle, a HLA-B*0702-restricted epitope according to claim 9 or claim 10, or a chimeric
 polypeptide according to claim 11 or claim 12, or a nucleic acid according to claim 13.

15. The pharmaceutical composition of claim 14, which is a vaccine.

16. A kit of parts comprising, in separate containers:

(i) a first peptide comprising a sequence of a HLA-B*0702-restricted
 epitope selected in the group of SEQ ID Nos: 2-31, and

30 (ii) a second peptide comprising a sequence consisting of a HLA-
 A*2402-restricted epitope selected in the group of SEQ ID Nos: 32-67.

17. The kit according to claim 16, wherein said first peptide is an isolated
 epitope selected in the group of SEQ ID Nos: 2-31, and said second peptide is its cognate
 optimized epitope.

35 18. The kit according to claim 16 or claim 17, wherein said first peptide
 comprises a sequence selected amongst GPRALAETS (SEQ ID No: 15), GPRALIETS
 (SEQ ID No: 16) and GPRALVETS (SEQ ID No: 17), and said second peptide comprises
 the sequence GPRALVETL (SEQ ID No: 54).

19. The kit according to any of claims 16 to 18, which is a vaccination kit, wherein said first and second peptides or chimeric polypeptides are in separate vaccination doses.

[illegible]

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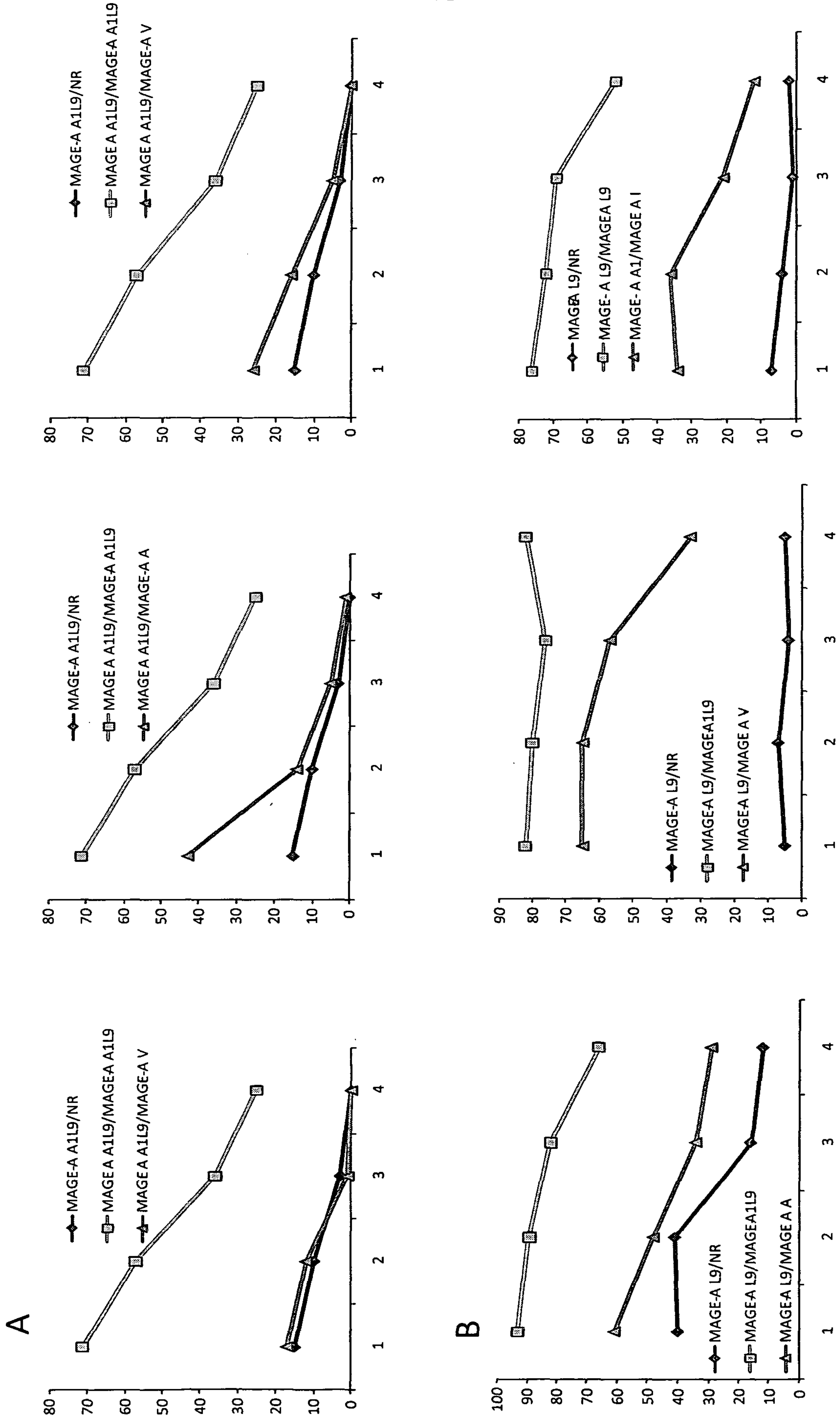


Figure 2