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(54) Title: PROBIOTIC FORMULATIONS AND METHODS FOR USE

(57) **Abrégé/Abstract:**

Provided herein are compositions comprising a biocompatible microsphere, a biofilm-generating probiotic bacterium, a prebiotic, and/or a prebiofilmic. Methods for preparing and formulating the compositions and methods for treating or preventing a disease using the compositions are also provided.



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PROBIOTIC FORMULATIONS AND METHODS FOR USE

CROSS-REFERENCE TO RELATED APPLICATION

[0001] This application claims priority under 35 U.S.C. § 119(e) to U.S. Provisional Application No. 61/949,058, filed March 6, 2014, the content of which is hereby incorporated by reference in its entirety.

FIELD OF THE INVENTION

[0002] This disclosure relates to novel probiotic formulations and methods for using same for treating or preventing disease.

BACKGROUND

[0003] Diarrheal illness is a major worldwide cause of morbidity and mortality, and accounts for approximately 15% of deaths in children. Enterohemorrhagic *Escherichia coli* (EHEC) and enteropathogenic *E. coli* (EPEC) are two primary bacterial causes of pediatric diarrhea. The mechanisms by which these pathogens cause diarrheal disease is not yet completely understood, but is initiated when the pathogens colonize the intestinal epithelium (Nataro and Kaper (1998) Diarrheagenic *Escherichia Coli*, Clin Microbiol Rev. 11:142-201).

[0004] A closely related pathogen, namely *Citrobacter rodentium* is a murine pathogen that is widely used to model human EPEC and EHEC infection, because mice are relatively resistant to both EPEC and EHEC. In mice, *C. rodentium* results in colonic pathology that is nearly indistinguishable from that produced by EPEC and EHEC in humans (Borenshtein, M. et al. (2008) Utility of the *Citrobacter rodentium* Infection Model in Laboratory Mice, Curr Opin Gastroenterol, 24:32-7; Luperchio and Schauer, (2001) Molecular Pathogenesis of *Citrobacter rodentium* and Transmissible Murine Colonic Hyperplasia, Microbes Infect, 3:333-40; Mundy, T. T. et al. (2005) *Citrobacter rodentium* of Mice and Man, Cell Microbiol 7:1697-706). This may not be surprising, since *C. rodentium* possesses a homologue of the locus of enterocyte effacement (LEE) pathogenicity island carried by EPEC and EHEC that encodes for the effector proteins necessary for the development of attaching and effacing (A/E) lesions. These lesions are accompanied by the development of colonic hyperplasia, and pathological colitis marked by epithelial defects and leukocyte infiltration (Luperchio and Schauer (2001) Molecular Pathogenesis of *Citrobacter rodentium* and Transmissible Murine Colonic Hyperplasia, Microbes Infect. 3:333-40).

[0005] The intestinal epithelium provides a formidable barrier to enteric pathogens. In order to cause disease, enteric pathogens must either adhere to or penetrate/invade host epithelial cells. Thus, interaction with epithelial cells is the first step in pathogenicity for all enteric pathogens, and this step can be studied through the use of A/E pathogens by assessing colonic colonization and resultant pathology.

[0006] Colonization of A/E pathogens in the colon is dependent upon the composition of the intestinal microbiota. Inducing dysbiosis (the disruption of the native populations of beneficial bacteria) within the colonic microbiota by administering antibiotics (Wlodarska, B. et al., (2011) Antibiotic Treatment Alters the Colonic Mucus Layer and Predisposes the Host to Exacerbated *Citrobacter rodentium*-Induced Colitis, Infect Immun, 79:1536-45) or by inducing an inflammatory response (Lupp, M. L. et al., (2007) Host-Mediated Inflammation Disrupts the Intestinal Microbiota and Promotes the Overgrowth of *Enterobacteriaceae*, Cell Host Microbe, 2:119-29) has been shown to greatly enhance pathogen colonization.

[0007] Colonic dysbiosis can further exacerbate the inflammatory response to the colonic pathogen (Wlodarska, B. et al., (2011) Antibiotic Treatment Alters the Colonic Mucus Layer and Predisposes the Host to Exacerbated *Citrobacter rodentium*-Induced Colitis, Infect Immun. 79:1536-45), but even in the absence of pathogen challenge, dysbiosis can propagate inflammatory responses in genetically susceptible individuals, as evidenced by the findings of dysbiosis in patients with inflammatory bowel disease (Machiels, et al., (2013) A Decrease of the Butyrate-Producing Species *Roseburia hominis* and *Faecalibacterium prausnitzii* Defines Dysbiosis in Patients with Ulcerative Colitis, Gut, published online first September 10, 2013; Morgan, et al., (2012) Dysfunction of the Intestinal Microbiome in Inflammatory Bowel Disease and Treatment, Genome Biol., 13:R79) or irritable bowel syndrome (Carroll, et al., (2012) Alterations in Composition and Diversity of the Intestinal Microbiota in Patients with Diarrhea-Predominant Irritable Bowel Syndrome, Neurogastroenterol Motil. 24:521-30, e248; Chassard, M. et al., (2012) Functional Dysbiosis within the Gut Microbiota of Patients with Constipated-Irritable Bowel Syndrome, Aliment Pharmacol Ther., 35:828-38).

[0008] Probiotics, are live microbes that when ingested in high enough quantities confer a health benefit for the host (Food and Agriculture Organization of the United Nations and World Health Organization, "Health and Nutritional Properties of Probiotics in Food Including Powdered Milk with Live Bacteria" (2001)), are gaining traction as a viable option for treating enteric diseases (Hemarajata and Versalovic, (2013) Effects of Probiotics on Gut

Microbiota: Mechanisms of Intestinal Immunomodulation and Neuromodulation, Therap Adv Gastroenterol, 6:39-51).

[0009] Many probiotic microbes have the capacity to enhance immune system activity, but fewer probiotic microbes have anti-inflammatory effects. *Lactobacillus reuteri* is a commonly used probiotic that has been shown to regulate the mammalian and avian intestinal immune system (Lin, et al., (2008) Probiotic *Lactobacillus reuteri* Suppress Proinflammatory Cytokines Via C-Jun, Inflamm Bowel Dis., 14:1068-83). Studies *in vitro* demonstrate that some strains of *L. reuteri* (such as PTA6475) can suppress the ability of myeloid cells to produce inflammatory cytokines (such as TNF- α) through a down-regulation of cell signal transduction pathways (e.g., c-Jun-dependent activator protein 1 (AP-1)) (Jones and Versalovic, (2009) Probiotic *Lactobacillus reuteri* Biofilms Produce Antimicrobial and Anti-Inflammatory Factors, BMC Microbiol, 9:35; Lin, et al., (2008) Probiotic *Lactobacillus Reuteri* Suppress Proinflammatory Cytokines Via C-Jun, Inflamm Bowel Dis. 14:1068-83).

[0010] Other strains of *L. reuteri*, such as ATCC23272, can down-regulate both cytokine and chemokine production by colonic epithelial cells stimulated with *C. rodentium*. *L. reuteri* can also reduce colonic inflammation in both juvenile and adult animals (Eaton, A. et al., (2011) Probiotic *Lactobacillus reuteri* Ameliorates Disease Due to Enterohemorrhagic *Escherichia coli* in Germfree Mice, Infect Immun., 79:185-91; Schreiber, et al., (2009) *Lactobacillus reuteri* Prevents Colitis by Reducing P-Selectin-Associated Leukocyte- and Platelet-Endothelial Cell Interactions, Am J Physiol Gastrointest Liver Physiol, 296:G534-42).

[0011] Studies demonstrate that *L. reuteri* attenuates the exacerbating effects of stress on *C. rodentium*-induced colitis as marked by reductions in colonic cytokines and chemokines, inflammatory cell infiltration, colonic epithelial cell defects, and pathogen translocation from the colon to the spleen (Mackos, et al., (2013) Probiotic *Lactobacillus reuteri* Attenuates the Stressor-Enhanced Severity of *Citrobacter rodentium* Infection, Infect Immun, 81:3253-63).

[0012] The effects of *L. reuteri* are most evident when stress leads to mild to moderate *C. rodentium*-induced colitis. However, under stress conditions that lead to severe *C. rodentium*-induced colitis, *L. reuteri* was able to prevent pathogen translocation and the development of systemic inflammatory responses, but it was not able to reduce all aspects of colonic pathology (Mackos, et al., (2013) Probiotic *Lactobacillus reuteri* Attenuates the Stressor-Enhanced Severity of *Citrobacter rodentium* Infection, Infect Immun, 81:3253-63).

[0013] Moreover, the effects of *L. reuteri* on the host were short-lived and no longer evident after daily administration was terminated. These studies demonstrate the immunomodulatory potential of *L. reuteri*.

SUMMARY

[0014] Under the right conditions, many probiotics can effectively prevent pathogen colonization due to either direct (e.g., production of antimicrobial defenses) or indirect (e.g., stimulation of host defenses) mechanisms. Few probiotic species are able to both prevent pathogen colonization and limit excessive inflammatory responses. This is important, however, because excessive colonic inflammation in response to colonic infection can lead to the development of protracted illness, such as post-infectious irritable bowel syndrome. Thus, the development of probiotics that are able to prevent excessive immune responses to colonic pathogens, while still maintaining anti-bacterial immunity would have the ability to prevent both short-term and longer-term health effects of enteric infection.

[0015] Anxiety and depression are common co-morbidities in both adults and children with gastrointestinal disease (Maunder et al., (2008) The role of stress in the development and clinical course of inflammatory bowel disease: epidemiological evidence (Curr Mol Med, 8:247-252; Waler et al., (2008) The Manitoba IBD cohort study: a population-based study of the prevalence of lifetime and 12-month anxiety and mood disorders, Am J Gastroenterol, 103:1989-1997), and studies suggest that reducing gastrointestinal disease can in turn improve anxiety and depression (Guloksuz et al., (2013) Depressive symptoms in Crohn's disease: relationship with immune activation and tryptophan availability; PLoS One, 8(3):e6043).

[0016] Aspects and embodiments of this technology combine the probiotic bacteria with prebiotic substances to help stimulate the exclusive growth of the probiotic species and provide the bacteria in the form of a biofilm on a biocompatible microsphere, which has greater efficacy and duration. It has been shown that probiotic biofilms can be grown on surfaces as a means to introduce bacteria into the site of wounds, where a formulation comprising a plaster or dressing based on a hydrocolloid that is a natural gelatin to treat wounds (i.e., EP2450062). However, there is an unmet need for fewer probiotic doses and greater efficacy of probiotic bacteria and its appropriate formulation in the methods as disclosed herein, to the best of Applicants' knowledge, has not yet heretofore been disclosed.

[0017] This technology provides methods of formulation, which enhance the efficiency and durability of introducing probiotic strains at a site. It specifically bypasses the rate limiting step of biofilm formation. This technology is useful for gastrointestinal gut health and any aspects where probiotic bacteria need to establish, e.g., the gastrointestinal tract, wound healing, skin, vaginal, oral, water purification.

[0018] Probiotics are a natural way to protect and restore gut microbiota to a healthy state. Unfortunately, even under optimal conditions, probiotic bacteria (as typically delivered) fail to establish, or sufficiently persist, minimizing the magnitude and duration of their healthful effects. One of the rate limiting steps is the capacity of introduced bacteria to form a lasting biofilm. When bacteria are already in the form of a biofilm (a surface adhered community) as opposed to planktonic (free-living), they more readily establish and persist. The positive effects of probiotic bacteria can be enhanced by providing them in a biofilm state; this can readily be accomplished by growing the bacteria on the surface of a biocompatible and non-toxic microsphere. Biocompatible microspheres can be biodegradable polymers, non-biodegradable polymers, a metal, or a combination thereof. When this surface is in the form of a microsphere, prebiotic and/or prebiofilmic substances can be added as cargo to facilitate establishment and maintenance of the probiotic bacterial biofilm.

[0019] Microspheres have added value in ideally providing diffusible prebiotic (nutritional supplementation specific/exclusive to probiotic bacteria) cargo that can help promote probiotic bacterial establishment and survival while limiting pathogenic bacterial challenge. At least for the probiotic bacterium *Lactobacillus reuteri*, the biofilm state is advantageous in establishing in the murine gut over the same bacteria in planktonic form.

[0020] Furthermore, *L. reuteri* introduced into mice as biofilms have a more robust and durable prophylactic effect on the pathogenesis of the enteropathogenic bacterium, *Citrobacter rodentium*, than *L. reuteri* in its planktonic form. Based on these results, highly integrated examples are developed that yield novel formulations of probiotics that provide greater and more lasting effects against dysbiosis preventing or even treating gut pathogenesis with a far reduced need for patient compliance.

[0021] The biofilm-generating probiotic bacterium adheres to the surface of the biocompatible microsphere and generates a biofilm. The biocompatible microsphere has either a solid or hollow core. When the biocompatible microsphere has a hollow core, it can carry a prebiotic and any nutritional supplementation for the probiotic bacterium as a cargo.

The prebiotic can be encapsulated within the hollow core. The microsphere can also carry a drug, or a compound, or an agent, which is selective against a pathogen, that in one aspect, may compete with the health-inducing bacterium in the composition. In addition to a biocompatible microsphere, biofilm-generating probiotic and prebiotic, a novel probiotic formulation can also contain a prebiofilmic, which is a substance that supports biofilm formation and/or durability, and in one aspect, the prebiofilmic is a DNA binding polypeptide or protein and/or a DNABII polypeptide or protein or a fragment thereof that supports biofilm formation and/or durability.

[0022] In view of the above advantages, provided herein is a composition comprising, or alternatively consisting essentially of, or yet further consisting of, a biocompatible microsphere, a biofilm-generating probiotic bacterium and a prebiotic, wherein the prebiotic comprises, or alternatively consisting essentially of, or yet consisting of, a nutritional supplementation for the probiotic bacterium. In one aspect, the composition further comprises, or alternatively consists essentially of, or yet further consisting of, a carrier, such as a pharmaceutically acceptable carrier or a biocompatible scaffold.

[0023] In some embodiments, the composition is formulated in a dosage form. Suitable dosage forms include, but are not limited to a suppository, a powder, a liquid, a capsule, a chewable tablet, a swallowable tablet, a buccal tablet, a troche, a lozenge, a soft chew, a solution, a suspension, a spray, a tincture, a decoction, an infusion, and combinations thereof.

[0024] This disclosure also provides a method for preparing the above-noted composition, the method comprising, or alternatively consisting essentially of, or yet further consisting of, admixing a biocompatible microsphere with a biofilm-generating probiotic bacterium, a prebiotic, and in one aspect, further admixing a prebiofilmic. In a further aspect, the method further comprises, or alternatively consists essentially of, or yet further consists of, admixing an effective amount of a nutritional supplement for the probiotic bacterium.

[0025] This disclosure also provides a composition comprising, or alternatively consisting essentially of, or yet further consisting of, a PGLA-biocompatible microsphere, one or more biofilm-generating probiotic bacterium comprising at least *Lactobacillus reuteri* ("*L. reuteri*"), and a nutritional supplementation comprising one or more of sucrose, glycerol, fructose and/or maltose, in an amount to support the growth of the probiotic bacterium. The composition may further comprise, or alternatively consist essentially of, or yet further consist of, an effective amount of an HU polypeptide or protein. The composition can further

comprise a pharmaceutically acceptable carrier or a biocompatible scaffold and is optionally formulated as a suppository.

[0026] This disclosure also provides a method for treating or preventing a disease suitable treated by a biofilm in a subject, such as inflammatory bowel disease, necrotizing enterocolitis (NEC) or psychological disorders or mood disorders, in a subject comprising, or alternatively consisting essentially of, or yet further consisting of, administering to a subject in need thereof, an effective amount of a composition comprising, or alternatively consisting essentially of, or yet further consisting of, a composition as described herein.

[0027] In some embodiments, a kit is provided comprising, or alternatively consisting essentially of, or yet consisting of, a composition as described herein and instructions for use diagnostically or therapeutically.

BRIEF DESCRIPTION OF THE FIGURES

[0028] **FIGS. 1 A and 1B** illustrate that *L. reuteri* biofilm structural integrity relies on the presence of DNABII family proteins. Confocal microscopy images of *in vitro* *L. reuteri* biofilms stained with LIVE/DEAD BacLight Bacterial Viability Kit (Molecular Probes). *L. reuteri* biofilms were grown for 24 hours at 37°C and 5% CO₂, at which time they were treated with a 1:50 dilution of either **A)** rabbit naïve serum, **B)** rabbit anti-integration host factor polypeptide (“IHF”) , or media with nothing added (data not shown) for 16 hours. Anti-IHF treatments resulted in a 20% decrease in maximum height, 35% decrease in average thickness, and 41% decrease in biomass (data not shown).

[0029] **FIG. 2** illustrates that Prebiotic compounds increase probiotic biofilms in average thickness and biomass. Addition of 10 µg/ml *S. mutans* HU to *L. reuteri* biofilm at time of seeding increased average thickness and biomass 33%, and 55%, respectively. Addition of 10 µg/ml calf thymus DNA increased average thickness 44% and biomass 68%. Adding 10 µg/ml of HU and DNA together led to an increased effect compared to either alone, with average thickness increasing 53% and biomass increasing 78%.

[0030] **FIG. 3** illustrates that *L. reuteri* *in vivo* colonization and retention with a single oral administration. Mice (n = 3/condition) were administered *L. reuteri* as planktonic, planktonic + PLGA, biofilm, and biofilm + PLGA cultures via oral gavage. After seven days, mice were sacrificed and *L. reuteri* 16S rRNA genes were PCR amplified from the mouse colon. The

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probiotic was found in a higher percentage of mice that were treated with biofilm cultures or cultures with PLGA present than in planktonic treatments.

[0031] FIG. 4 illustrates that *L. reuteri* biofilm grown with PLGA microspheres and HU reduces *C. rodentium* spleen colonization more effectively than biofilm and planktonic *L. reuteri*. Mice (n = 6/condition) were treated with a single oral gavage of *L. reuteri* in one of the following forms: planktonic, planktonic + PLGA + HU, biofilm, and biofilm + PLGA + HU (0.115µg/ml PLGA, 10µg/ml HU). After 12 hours the mice were gavaged with *C. rodentium*, and sacrificed 12 days post-infection for necropsy. Only *L. reuteri* biofilm + PLGA + HU showed a statistically significant decrease in *C. rodentium* CFU/g(P=0.0343).

[0032] FIG. 5 shows the results of a study establishing that compositions of this disclosure reduce inflammation and antagonize bacterial pathogens in an animal model of NEC.

SEQUENCE LISTING

[0033] SEQ ID NO: 1 Full Length Wild type (wt) 86-028NP *Haemophilus influenzae* IhfA; Genbank accession No.: AAX88425.1, last accessed March 21, 2011:

MATITKLDIIEYLSDKYHLSK

QDTKNVVENFLEEIRLSLESGQDVKLSGFGNFELRDKSSRPGRNPKTGDVVPVSARR
VVTFKPGQKLRARVEKTK

[0034] SEQ ID NO: 2 Full Length wild-type 86-028NP *Haemophilus influenzae* HU, Genbank accession No.: YP_248142.1, last accessed March 21, 2011:

MRFVTIFINHAFNSSQVRLSFAQFLR

QIRKDTFKESNFLFNRRYKFMNKTDLIDAIANAELNKKQAKAALEATLDAITASLK
EGEPVQLIGFGTFKVNERRAARTGRNPQTGAIEIQIAASKVPAFVSGKALKDAIK

[0035] SEQ ID NO: 3 Full Length wt R2846 *Haemophilus influenzae* IhfA, Genbank accession No.: ADO96375, last accessed March 21, 2011:

MATITKLDIIEYLSDKYHLSKQDTKNVVENFL

EEIRLSLESGQDVKLSGFGNFELRDKSSRPGRNPKTGDVVPVSARRVVTFKPGQKLR
ARVEKTK

[0036] SEQ ID NO: 4 Full Length wild-type Rd *Haemophilus influenzae* IhfA; Genbank accession No.: AAC22959.1, last accessed March 21, 2011:

MATITKLDIIEYLSDKYHLSKQDTK

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NVVENFLEEIRLSLESGQDVKLSGFGNFELRDKSSRPGRNPKTGDVVPVSARRVVTF
KPGQKLRARVEKTK;

[0037] SEQ ID NO: 5 Full Length wild-type *E. coli* K12 IhfA; Genbank accession No.: AAC74782.1, last accessed March 21, 2011:

MALTKAEMSEYLFDKLGLSKRDAKELVELFFE
EIRRALENGEQVKLSGFGNFDLRDKNQRPGRNPKTGEDIPITARRVVT
FRPGQKLKSRVENASPKDE; DNA Genbank No. NC_000913

[0038] SEQ ID NO: 6 Full Length wild-type *P. aeruginosa* PA 01 IhfA; Genbank accession No.: AAG06126.1, last accessed March 21, 2011: MGALTKAEIAERLYEELGLNKREA
KELVELFFEEIRQALEHNEQVKLSGFGNFDLRDKNQRPGRNPKTGEEIPITARRVVTF
RPGQKLKARVEAYAGTKS

[0039] SEQ ID NO: 7 β -3 and α -3 portions of (IHF α): TFRPGQ and
KLKSRVENASPKDE

[0040] SEQ ID NO: 8 β -3 and α -3 portions of (IHF β): HFKPGK and ELRDRANIYG

[0041] SEQ ID NO: 9 β -3 and α -3 portions of: TFKPGQ and KLRARVEKTK

[0042] SEQ ID NO: 10 β -3 and α -3 portions of 2019 *Haemophilus influenzae* IhfA:
TFKPGQ and KLRARVENTK

[0043] SEQ ID NO: 11 β -3 and α -3 portions of: TFKPGQ and: KLRARVEKTK

[0044] SEQ ID NO: 12 β -3 and α -3 portions of: TFRPGQ and KLKSRVENASPKDE

[0045] SEQ ID NO: 13 β -3 and α -3: TFRPGQ and KLKARVEAYAGTKS

[0046] SEQ ID NO: 14 *E. coli* hupA, Genbank accession No.: AP_003818, Last accessed March 21, 2011:

MNKTQLIDVIAEKAELSKTQAKAALESTLAAITESLKEGDAVQLVGFGTFK
VNHRAERTGRNPQTGKEIKIAAANVPAFVSGKALKDAVK

[0047] SEQ ID NO: 15 *E. coli* hupB, Genbank accession No.: AP_001090.1, Last accessed March 21, 2011:

MNKSQLIDKIAAGADISKAAAGRALDAIIASVTESLKEGDDVALVGFG
TFAVKERAARTGRNPQTGKEITIAAAKVPSFRAGKALKDAVN

[0048] SEQ ID NO: 16 β -3 and α -3 portions of: AFVSGK and ALKDAVK

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[0049] SEQ ID NO: 17 β -3 and α -3 portions of SFRAGK and ALKDAVN

[0050] SEQ ID NO: 18 C-terminal 20 amino acids of IHF α :
TFRPGQKLKSRVENASPKDE

[0051] SEQ ID NO: 19 C-terminal 20 amino acids of IHF β :
KYVPHFKPGKELRDRANIYG

[0052] SEQ ID NO: 20 DNABII binding consensus sequence: WATCAANNNTTR
wherein W is A or T, N is any base and R is a purine.

[0053] SEQ ID NO: 21 *E. coli* IHF α : GRNPKTGEDIPI

[0054] SEQ ID NO: 22 *E. coli* IHF β : GRNPKTGDKVEL

[0055] SEQ ID NO: 23 *E. coli* HU α : GRNPQTGKEIKI

[0056] SEQ ID NO: 24 *E. coli* HU β : GRNPQTGKEITI

DETAILED DESCRIPTION

[0057] It is to be understood that this invention is not limited to particular embodiments described, as such may, of course, vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to be limiting, since the scope of the present invention will be limited only by the appended claims.

[0058] Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, the preferred methods, devices and materials are now described. All technical and patent publications cited herein are incorporated herein by reference in their entirety. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

[0059] The practice of the present technology will employ, unless otherwise indicated, conventional techniques of tissue culture, immunology, molecular biology, microbiology, cell biology and recombinant DNA, which are within the skill of the art. See, e.g., Sambrook and Russell eds. (2001) *Molecular Cloning: A Laboratory Manual*, 3rd edition; the series Ausubel et al. eds. (2007) *Current Protocols in Molecular Biology*; the series *Methods in Enzymology* (Academic Press, Inc., N.Y.); MacPherson et al. (1991) *PCR 1: A Practical Approach* (IRL

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Press at Oxford University Press); MacPherson et al. (1995) PCR 2: A Practical Approach; Harlow and Lane eds. (1999) Antibodies, A Laboratory Manual; Freshney (2005) Culture of Animal Cells: A Manual of Basic Technique, 5th edition; Gait ed. (1984) Oligonucleotide Synthesis; U.S. Patent No. 4,683,195; Hames and Higgins eds. (1984) Nucleic Acid Hybridization; Anderson (1999) Nucleic Acid Hybridization; Hames and Higgins eds. (1984) Transcription and Translation; Immobilized Cells and Enzymes (IRL Press (1986)); Perbal (1984) A Practical Guide to Molecular Cloning; Miller and Calos eds. (1987) Gene Transfer Vectors for Mammalian Cells (Cold Spring Harbor Laboratory); Makrides ed. (2003) Gene Transfer and Expression in Mammalian Cells; Mayer and Walker eds. (1987) Immunochemical Methods in Cell and Molecular Biology (Academic Press, London); and Herzenberg et al. eds (1996) Weir's Handbook of Experimental Immunology.

[0060] All numerical designations, e.g., pH, temperature, time, concentration and molecular weight, including ranges, are approximations which are varied (+) or (-) by increments of 1.0 or 0.1, as appropriate, or alternatively by a variation of +/- 15 %, or alternatively 10%, or alternatively 5% or alternatively 2%. It is to be understood, although not always explicitly stated, that all numerical designations are preceded by the term "about". It also is to be understood, although not always explicitly stated, that the reagents described herein are merely exemplary and that equivalents of such are known in the art.

[0061] As used in the specification and claims, the singular form "a", "an" and "the" include plural references unless the context clearly dictates otherwise. For example, the term "a bacterium" includes a plurality of bacteria, including mixtures thereof.

[0062] As used herein, the term "comprising" is intended to mean that the compositions and methods include the recited elements, but do not exclude others. "Consisting essentially of" when used to define compositions and methods, shall mean excluding other elements of any essential significance to the combination for the intended use. Thus, a composition consisting essentially of the elements as defined herein would not exclude trace contaminants from the isolation and purification method and pharmaceutically acceptable carriers, such as phosphate buffered saline, preservatives and the like. "Consisting of" shall mean excluding more than trace elements of other ingredients and substantial method steps for administering the compositions of this invention. Embodiments defined by each of these transition terms are within the scope of this invention.

[0063] A “biofilm” intends a thin layer or an organized community of microorganisms that at times can adhere to the surface of a structure, that may be organic or inorganic, together with the polymers, such as DNA, that they secrete and/or release. The biofilms are very resistant to microbiotics and antimicrobial agents. They live on gingival tissues, teeth, and restorations, causing caries and periodontal disease, also known as periodontal plaque disease. They also cause chronic middle ear infections. Biofilms can also form on the surface of dental implants, stents, catheter lines and contact lenses. They grow on pacemakers, heart valve replacements, artificial joints and other surgical implants. The Centers for Disease Control estimate that over 65% of nosocomial (hospital-acquired) infections are caused by biofilms. Fungal biofilms also frequently contaminate medical devices. They cause chronic vaginal infections and lead to life-threatening systemic infections in people with hobbled immune systems. Biofilms also are involved in numerous diseases. For instance, cystic fibrosis patients have *Pseudomonas* infections that often result in antibiotic resistant biofilms.

[0064] A “prebiotic” intends a nutritional supplement for the probiotic bacterium. Prebiotics are food ingredients, for example, oligosaccharides, that are non-digestible by a subject (e.g., by a mammal such as a human), and that stimulates the growth or activity of one or more beneficial bacteria and/or inhibit the growth or activity of one or more pathogenic bacteria. A prebiotic may selectively stimulate the growth and/or activity of one or a limited number of bacteria in the subject.

[0065] A “prebiofilmic” intends a substance that supports biofilm formation and durability, for example the prebiofilmic can be a substance that supports the extracellular matrix of the biofilm like an eDNA binding polypeptide or protein or alternatively a substrate that can be converted into a substance that facilitate adhesion, e.g. sucrose.

[0066] A “DNABII polypeptide or protein” intends a DNA binding protein or polypeptide that is composed of DNA-binding domains and thus have a specific or general affinity for DNA. In one aspect, they bind DNA in the minor groove. Non-limiting examples of DNABII proteins are an integration host factor (IHF) protein and a histone-like protein from *E. coli* strain U93 (HU), examples of which are provided in SEQ ID NOs: 1 to 24 and additional strains and polypeptides are provided in **Table 4**. Also intended are polypeptide fragments and equivalent polypeptides that have amino acid modifications that do not substantially change the biological activity of the protein or polypeptides, or active fragment thereof.

Active fragments thereof include, for example, the c-terminal half or c-terminal third of the protein or polypeptide. Other DNA binding proteins that can be associated with the biofilm include DPS (Genbank Accession No.: CAA49169), H-NS (Genbank Accession No.: CAA47740), Hfq (Genbank Accession No.: ACE63256), CbpA (Genbank Accession No.: BAA03950) and CbpB (Genbank Accession No.: NP_418813), as well as equivalent polypeptides and active fragments thereof.

[0067] A “microsphere” intends a biofilm-carrying and/or compound-carrying (e.g., drug-carrying) particulate or granular material within the particular size range recited. As used herein, a microsphere consisting of particles 50 millimeters or less in diameter, and 1 micron or more (e.g., 1 to 100 or alternatively, or alternatively, 1 to 75 microns, or alternatively 1 to 50, or alternatively 1 to 25, or alternatively 1 to 10 microns) in diameter. Non-limiting examples of such include hollow microspheres that can, in some aspects, contain a pharmaceutical or drug, microcapsules (in which the excipient forms a skin or shell that surrounds and contains a cargo, such as a drug), and microparticles, which are used as a generic term for any particles in the recited size range, whether spherical or not, as those terms are typically used in the art.

[0068] A “biodegradable polymer” intends polymers that are biocompatible and can degrade *in vivo* by bodily processes to products that are readily disposable by the body and should not accumulate in the body.

[0069] By “biocompatible”, it is meant that the components of the delivery system will not cause tissue injury or injury to the human biological system. To impart biocompatibility, polymers and excipients that have had history of safe use in humans or with GRAS (Generally Accepted As Safe) status, are preferentially used. By biocompatibility, it is meant that the ingredients and excipients used in the composition will ultimately be “bioabsorbed” or cleared by the body with no adverse effects to the body. For a composition to be biocompatible, and be regarded as non-toxic, it must not cause toxicity to cells. Similarly, the term “bioabsorbable” refers to microspheres made from materials which undergo bioabsorption *in vivo* over a period of time such that long term accumulation of the material in the patient is avoided. The biocompatible nanoparticle is bioabsorbed over a period of less than 2 years, preferably less than 1 year and even more preferably less than 6 months. The rate of bioabsorption is related to the size of the particle, the material used, and other factors

well recognized by the skilled artisan. A mixture of bioabsorbable, biocompatible materials can be used to form the microspheres used in this invention.

[0070] An “integration host factor” or “IHF” protein is a bacterial protein that is used by bacteriophages to incorporate their DNA into the host bacteria. These are DNA binding proteins that function in genetic recombination as well as in transcription and translational regulation. They also bind extracellular microbial DNA. The genes that encode the IHF protein subunits in *E. coli* are *himA* (Genbank accession No.: POA6X7.1) and *himD* (POA6Y1.1) genes. Non-limiting examples of such are provided in the attached sequence listing and noted in Table 4.

[0071] “HU” or “histone-like protein from *E. coli* strain U93” refers to a class of heterodimeric proteins typically associated with *E. coli*. HU proteins are known to bind DNA junctions. Related proteins have been isolated from other microorganisms. The complete amino acid sequence of *E. coli* HU was reported by Laine et al. (1980) Eur. J. Biochem. 103(3):447-481. Antibodies to the HU protein are commercially available from Abcam. Non-limiting examples of such are provided in the attached sequence listing.

[0072] The term “protein”, “peptide” and “polypeptide” are used interchangeably and in their broadest sense to refer to a compound of two or more subunit amino acids, amino acid analogs or peptidomimetics. The subunits may be linked by peptide bonds. In another embodiment, the subunit may be linked by other bonds, e.g., ester, ether, etc. A protein or peptide must contain at least two amino acids and no limitation is placed on the maximum number of amino acids which may comprise a protein’s or peptide’s sequence. As used herein the term “amino acid” refers to either natural and/or unnatural or synthetic amino acids, including glycine and both the D and L optical isomers, amino acid analogs and peptidomimetics.

[0073] A “c-terminal polypeptide” intends the c-terminal half or c-terminal third of a polypeptide. As an example, for polypeptides containing 90 amino acids, the c-terminal polypeptide would comprise amino acids 46 through 90 or amino acids 60 through 90. In another aspect, the term intends the c-terminal 20 amino acids from the carboxy terminus.

[0074] A “n-terminal polypeptide” intends the n-terminal half of a polypeptide. As an example, for polypeptides containing 90 amino acids, the c-terminal polypeptide would

comprise amino acids 1 through 45. In another aspect, the term intends the c-terminal 20 amino acids from the amino terminus.

[0075] The terms “polynucleotide” and “oligonucleotide” are used interchangeably and refer to a polymeric form of nucleotides of any length, either deoxyribonucleotides or ribonucleotides or analogs thereof. Polynucleotides can have any three-dimensional structure and may perform any function, known or unknown. The following are non-limiting examples of polynucleotides: a gene or gene fragment (for example, a probe, primer, EST or SAGE tag), exons, introns, messenger RNA (mRNA), transfer RNA, ribosomal RNA, RNAi, ribozymes, cDNA, recombinant polynucleotides, branched polynucleotides, plasmids, vectors, isolated DNA of any sequence, isolated RNA of any sequence, nucleic acid probes and primers. A polynucleotide can comprise modified nucleotides, such as methylated nucleotides and nucleotide analogs. If present, modifications to the nucleotide structure can be imparted before or after assembly of the polynucleotide. The sequence of nucleotides can be interrupted by non-nucleotide components. A polynucleotide can be further modified after polymerization, such as by conjugation with a labeling component. The term also refers to both double- and single-stranded molecules. Unless otherwise specified or required, any embodiment of this invention that is a polynucleotide encompasses both the double-stranded form and each of two complementary single-stranded forms known or predicted to make up the double-stranded form.

[0076] A polynucleotide is composed of a specific sequence of four nucleotide bases: adenine (A); cytosine (C); guanine (G); thymine (T); and uracil (U) for thymine when the polynucleotide is RNA. Thus, the term “polynucleotide sequence” is the alphabetical representation of a polynucleotide molecule. This alphabetical representation can be input into databases in a computer having a central processing unit and used for bioinformatics applications such as functional genomics and homology searching.

[0077] The term “isolated” or “recombinant” as used herein with respect to nucleic acids, such as DNA or RNA, refers to molecules separated from other DNAs or RNAs, respectively that are present in the natural source of the macromolecule as well as polypeptides. The term “isolated or recombinant nucleic acid” is meant to include nucleic acid fragments which are not naturally occurring as fragments and would not be found in the natural state. The term “isolated” is also used herein to refer to polynucleotides, polypeptides, antibodies and proteins that are isolated from other cellular proteins and is meant to encompass both purified

and recombinant polypeptides. In other embodiments, the term “isolated or recombinant” means separated from constituents, cellular and otherwise, in which the cell, tissue, polynucleotide, peptide, polypeptide, protein, antibody or fragment(s) thereof, which are normally associated in nature. For example, an isolated cell is a cell that is separated from tissue or cells of dissimilar phenotype or genotype. An isolated polynucleotide is separated from the 3' and 5' contiguous nucleotides with which it is normally associated in its native or natural environment, e.g., on the chromosome. As is apparent to those of skill in the art, a non-naturally occurring polynucleotide, peptide, polypeptide, protein, antibody or fragment(s) thereof, does not require “isolation” to distinguish it from its naturally occurring counterpart.

[0078] It is to be inferred without explicit recitation and unless otherwise intended, that when the present invention relates to a polypeptide, protein, polynucleotide or antibody, an equivalent or a biologically equivalent of such is intended within the scope of this invention. As used herein, the term “biological equivalent thereof” is intended to be synonymous with “equivalent thereof” when referring to a reference protein, antibody, polypeptide, polynucleotide or nucleic acid, intends those having minimal homology while still maintaining desired structure or functionality. Unless specifically recited herein, it is contemplated that any nucleic acid, polynucleotide, polypeptide or protein mentioned herein also includes equivalents thereof. For example, an equivalent intends at least about 70%, or alternatively 80 % homology or identity and alternatively, at least about 85 %, or alternatively at least about 90 %, or alternatively at least about 95 %, or alternatively 98 % percent homology or identity across the protein or a particular fragment thereof, and exhibits substantially equivalent biological activity to the reference protein, polypeptide or nucleic acid.

[0079] A polynucleotide or polynucleotide region (or a polypeptide or polypeptide region) having a certain percentage (for example, 80%, 85%, 90%, or 95%) of “sequence identity” to another sequence means that, when aligned, that percentage of bases (or amino acids) are the same in comparing the two sequences. The alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in Current Protocols in Molecular Biology (Ausubel et al., eds. 1987) Supplement 30, section 7.7.18, Table 7.7.1. Preferably, default parameters are used for alignment. A preferred alignment program is BLAST, using default parameters. In particular, preferred

programs are BLASTN and BLASTP, using the following default parameters: Genetic code = standard; filter = none; strand = both; cutoff = 60; expect = 10; Matrix = BLOSUM62; Descriptions = 50 sequences; sort by = HIGH SCORE; Databases = non-redundant, GenBank + EMBL + DDBJ + PDB + GenBank CDS translations + SwissProtein + SPupdate + PIR. Details of these programs can be found at the following Internet address: ncbi.nlm.nih.gov/cgi-bin/BLAST.

[0080] “Homology” or “identity” or “similarity” refers to sequence similarity between two peptides or between two nucleic acid molecules. Homology can be determined by comparing a position in each sequence which may be aligned for purposes of comparison. When a position in the compared sequence is occupied by the same base or amino acid, then the molecules are homologous at that position. A degree of homology between sequences is a function of the number of matching or homologous positions shared by the sequences. An “unrelated” or “non-homologous” sequence shares less than 40% identity, or alternatively less than 25% identity, with one of the sequences of the present invention.

[0081] As used herein, “expression” refers to the process by which polynucleotides are transcribed into mRNA and/or the process by which the transcribed mRNA is subsequently being translated into peptides, polypeptides, or proteins. If the polynucleotide is derived from genomic DNA, expression may include splicing of the mRNA in an eukaryotic cell.

[0082] The term “encode” as it is applied to polynucleotides refers to a polynucleotide which is said to “encode” a polypeptide if, in its native state or when manipulated by methods well known to those skilled in the art, it can be transcribed and/or translated to produce the mRNA for the polypeptide and/or a fragment thereof. The antisense strand is the complement of such a nucleic acid, and the encoding sequence can be deduced therefrom.

[0083] A “subject” or “patient” of diagnosis or treatment is a cell or an animal such as a mammal or a human. Non-human animals subject to diagnosis or treatment and are those subject to infections or animal models, for example, simians, murines, such as, rats, mice, chinchilla, canine, such as dogs, leporids, such as rabbits, livestock, sport animals and pets.

[0084] As used herein, the terms “treating,” “treatment” and the like are used herein to mean obtaining a desired pharmacologic and/or physiologic effect. The effect may be prophylactic in terms of completely or partially preventing a disorder or sign or symptom

thercof and/or may be therapeutic in terms of a partial or complete cure for a disorder and/or adverse effect attributable to the disorder.

[0085] To “prevent” intends to prevent a disorder or effect *in vitro* or *in vivo* in a system or subject that is predisposed to the disorder or effect. An example of such is preventing the formation of a biofilm in a system that is infected with a microorganism known to produce one.

[0086] The term “culturing” refers to the *in vitro* propagation of cells or organisms on or in media of various kinds. It is understood that the descendants of a cell grown in culture may not be completely identical (i.e., morphologically, genetically, or phenotypically) to the parent cell. By “expanded” is meant any proliferation or division of cells.

[0087] “Pharmaceutically acceptable carriers” refers to any diluents, excipients or carriers that may be used in the compositions of the invention. Pharmaceutically acceptable carriers include ion exchangers, alumina, aluminum stearate, lecithin, serum proteins, such as human serum albumin, buffer substances, such as phosphates, glycine, sorbic acid, potassium sorbate, partial glyceride mixtures of saturated vegetable fatty acids, water, salts or electrolytes, such as protamine sulfate, disodium hydrogen phosphate, potassium hydrogen phosphate, sodium chloride, zinc salts, colloidal silica, magnesium trisilicate, polyvinyl pyrrolidone, cellulose-based substances, polyethylene glycol, sodium carboxymethylcellulose, polyacrylates, waxes, polyethylene-polyoxypropylene-block polymers, polyethylene glycol and wool fat. Suitable pharmaceutical carriers are described in Remington's Pharmaceutical Sciences, Mack Publishing Company, a standard reference text in this field. They are preferably selected with respect to the intended form of administration, that is, oral tablets, capsules, elixirs, syrups and the like and consistent with conventional pharmaceutical practices.

[0088] A “biocompatible scaffold” refers to a scaffold or matrix for with the ability to support biofilm proliferation upon administration to a subject. In other embodiments, a biocompatible scaffold is a precursor to an implantable device which has the ability to perform its intended function, with the desired degree of incorporation in the host, without eliciting an undesirable local or systemic effects in the host. Biocompatible scaffolds are described in U.S. Patent Nos. 6,638,369 and 8,815,276;

[0089] “Administration” can be effected in one dose, continuously or intermittently throughout the course of treatment. Methods of determining the most effective means and dosage of administration are known to those of skill in the art and will vary with the composition used for therapy, the purpose of the therapy, the target cell being treated and the subject being treated. Single or multiple administrations can be carried out with the dose level and pattern being selected by the treating physician. Suitable dosage formulations and methods of administering the agents are known in the art. Route of administration can also be determined and method of determining the most effective route of administration are known to those of skill in the art and will vary with the composition used for treatment, the purpose of the treatment, the health condition or disease stage of the subject being treated and target cell or tissue. Non-limiting examples of route of administration include oral administration, vaginal, nasal administration, injection, topical application and by suppository.

[0090] The term “effective amount” refers to a quantity sufficient to achieve a beneficial or desired result or effect. In the context of therapeutic or prophylactic applications, the effective amount will depend on the type and severity of the condition at issue and the characteristics of the individual subject, such as general health, age, sex, body weight, and tolerance to pharmaceutical compositions. In the context of a therapeutic composition, in some embodiments the effective amount is the amount sufficient to result in a protective response against a pathogen. In other embodiments, the effective amount is the amount sufficient to result in antibody generation against the antigen. In some embodiments, the effective amount is the amount required to confer passive immunity on a subject in need thereof. In some embodiments, the amount is sufficient to accomplish one or more of 1) clear pathogen; 2) restore healthy microbiota; 3) modulate the immune system; and 4) maintain metabolism and metabolic pathways.

[0091] In the case of an *in vitro* application, in some embodiments the effective amount will depend on the size and nature of the application in question. It will also depend on the nature and sensitivity of the *in vitro* target and the methods in use. The skilled artisan will be able to determine the effective amount based on these and other considerations. The effective amount may comprise one or more administrations of a composition depending on the embodiment.

[0092] The agents and compositions can be used in the manufacture of medicaments and for the treatment of humans and other animals by administration in accordance with conventional procedures, such as an active ingredient in pharmaceutical compositions.

[0093] An agent of the present invention can be administered for therapy by any suitable route of administration. It will also be appreciated that the preferred route will vary with the condition and age of the recipient and the disease being treated.

[0094] Necrotizing enterocolitis ("NEC") is a medical condition primarily seen in premature infants where portions of the bowel undergo necrosis (tissue death). It occurs postnatally (i.e., is not seen in stillborn infants) and is the second most common cause of mortality. 7% of all neonatal intensive care unit admissions are NEC related. The mortality rate is 12%

MODES FOR CARRYING OUT THE DISCLOSURE

[0095] Diarrheal illness occurs in approximately four billion individuals per year and causes more than two million deaths worldwide. Among the most important bacterial causes of diarrheal illness in infants and young children are the attaching and effacing (A/E) pathogens, which upon colonization induce diarrheal disease that is associated with an increase in inflammatory cytokines and structural changes to colonic tissue. This acute infection can have a lasting effect on gut health, and infection with A/E pathogens and excessive inflammatory responses are known risk factors for the development of post-infectious irritable bowel syndrome.

[0096] Probiotics are a natural way to protect and restore gut microbiota to a healthy state and have been shown to promote health distal to the site of colonization. See Mackos et al. (2013) *Infection and Immunity* 81, No. 9 (3253-3262). Unfortunately, even under optimal conditions, probiotic bacteria fail to establish, or sufficiently persist, minimizing the magnitude and duration of their healthful effects. One of the rate limiting steps is the capacity of introduced bacteria to form a lasting biofilm. When bacteria are already in the form of a biofilm (a surface adhered community) as opposed to planktonic (free-living), they more readily establish and persist. The positive effects of probiotic bacteria can be enhanced by providing them in a biofilm state; this can readily be accomplished by growing the bacteria on the surface of a biocompatible and non-toxic microsphere. Biocompatible microspheres can be biodegradable polymers, non-biodegradable polymers, a metal, or a

combination thereof. When this surface is in the form of a microsphere, prebiotic and/or prebiofilmic substances can be added as cargo to facilitate establishment and maintenance of the probiotic bacterial biofilm.

[0097] Microspheres have added value in ideally providing diffusible prebiotic (nutritional supplementation specific/exclusive to probiotic bacteria) cargo that can help promote probiotic bacterial establishment and survival while limiting pathogenic bacterial challenge. At least for the probiotic bacterium *Lactobacillus reuteri*, the biofilm state is advantageous in establishing in the murine gut over the same bacteria in planktonic form.

[0098] Furthermore, *L. reuteri* introduced into mice as biofilms have a more robust and durable prophylactic effect on the pathogenesis of the enteropathogenic bacterium, *Citrobacter rodentium*, than *L. reuteri* in its planktonic form. Based on these results, three highly integrated examples are developed that yield novel formulations of probiotics that provide greater and more lasting effects against dysbiosis preventing or even treating gut pathogenesis with a far reduced need for patient compliance.

[0099] The biofilm-generating probiotic bacterium adheres to the surface of the biocompatible microsphere and generates a biofilm. The biocompatible microsphere has either a solid or hollow core. When the biocompatible microsphere has a hollow core, it can carry a prebiotic and any nutritional supplementation for the probiotic bacterium as a cargo. The prebiotic can be encapsulated within the hollow core. The microsphere can also carry a drug, or a compound, or an agent, which is selective against the growth or proliferation of a pathogen. In addition to a biocompatible microsphere, biofilm-generating probiotic and prebiotic, a novel probiotic formulation may also contain a prebiofilmic, which a substance that supports biofilm formation and durability, specifically, the prebiofilmic is a DNA binding polypeptide or protein and/or a DNABII polypeptide or protein, a fragment and/or an equivalent of each thereof. Non-limiting examples of such are provided in the attached sequence listing. One or more drug, compound or agent as well as one or more prebiofilmic can be within a single microsphere.

[0100] The prebiotic can support the growth of any probiotic bacteria, including biofilm-generating bacteria. The prebiotic is usually one or more of a water-soluble carbohydrate, such as inulin, oligofructose, fructo-oligosaccharide, galacto-oligosaccharide, glucose, maltose, maltodextrins, polydextrose, sucrose, fructose, lactose, isomaltulose, polyols, and

glycerol. The combination of various prebiotics can be used to support the growth of probiotics.

[0101] Probiotics are any type of micro-organisms that have health benefits. Probiotics are also commonly consumed as part of fermented foods with specially added active live cultures, such as in yogurt, soy yogurt, or as dietary supplements. Probiotics can also be taken as a suppository. Some limiting examples of probiotics are *L. acidophilus*, *L. crispatus*, *L. gasseri*, group *L. delbrueckii*, *L. salivarius*, *L. casei*, *L. paracasei*, *L. plantarum*, *L. rhamnosus*, *L. reuteri*, *L. brevis*, *L. buchneri*, *L. fermentum*, *L. rhamnosus*, *B. adolescentis*, *B. angulation*, *B. bifidum*, *B. breve*, *B. catenulatum*, *B. infantis*, *B. lactis*, *B. longum*, *B. pseudocatenulatum*, and *S. thermophiles*.

[0102] Probiotics support anti-bacterial immunity by preventing pathogen colonization and/or limiting excessive inflammatory responses. Without being bound by theory, the probiotics down-regulate cytokine and chemokine production.

[0103] The biocompatible microsphere can be one or more of a biodegradable polymer, a non-biodegradable polymer, a metal, or a mixture thereof. The biodegradable polymer can be selected from, but not limited to, poly(lactic-co-glycolic acid) or PLGA; polycaprolactone or PLC; Chitosan; Gelatin; DNA hydrogen; acetalated dextran; poly(lactide); poly(glycolide); poly(lactide-co-glycolide); poly(lactic acid); poly(glycolic acid); poly(lactic acid-co-glycolic acid); poly(lactide)/poly(ethylene glycol) copolymers; poly(glycolide)/poly(ethylene glycol) copolymer; poly(lactide-co-glycolide)/poly(ethylene glycol) copolymers; poly(lactic acid)/poly(ethylene glycol) copolymer; poly(glycolic acid)/poly(ethylene glycol) copolymer; poly(lactic acid-co-glycolic acid)/poly(ethylene glycol) copolymer; poly(caprolactone); poly(caprolactone)/poly(ethylene glycol) copolymer; poly(orthoester); poly(phosphazene); poly(hydroxybutyrate); poly(hydroxybutyrate); poly(lactide-co-caprolactone); polycarbonate; polyesteramide; polyanhydride; poly(dioxanone); poly(alkylene alkylate); polyethylene glycol/polyorthoester copolymer; polyurethane; poly(amino acid); polyetherester; polyacetal; polycyanoacrylate; poly(oxyethylene)/poly(oxypropylene) copolymer; Sephadex® copolymers (made from dextran cross-linked with epichlorohydrine, commercially available from Sigma-Aldrich and noted in Koo and Wankat (1988) Korean Biochem. J. 21(1)) and/or a combination thereof. The non-biodegradable polymer can be selected from, but not limited to, poly(ethylene vinyl acetate), poly(vinyl acetate), silicone polymers, polyurethanes, polysaccharides such as a cellulosic polymers and cellulose derivatives, acyl substituted

cellulose acetates and derivatives thereof, copolymers of poly(ethylene glycol) and poly(butylene terephthalate), polystyrenes, polyvinyl chloride, polyvinyl fluoride, poly(vinyl imidazole), chlorosulphonated polyolefins, polyethylene oxide, and copolymers and blends thereof. The metal can be selected from, but not limited to, cobalt, chromium, gold, nickel, platinum, stainless steel, titanium, tantalum, nickel-titanium, and alloys and combinations thereof.

[0104] The microspheres are selected to facilitate the endurance and robustness of the probiotic biofilms are identified and characterized. It has been shown that probiotic biofilms formed on the biodegradable (and FDA approved) surface, poly(lactic-co-glycolic acid) (PLGA) yields biofilms that are superior at preventing pathogen translocation through the epithelial barrier. Other FDA approved or generally regarded as safe (GRAS) materials that can be used to create surfaces to grow biofilms are also examined. The results using biological effectiveness and durability in animal models and shelf life as the base criteria are prioritized. Finally, to further improve the effectiveness of the introduction and maintenance of the probiotic biofilm, prebiotic substances to the probiotic biofilm surface by way of diffusible cargo within the microspheres are provided.

Compositions

[0105] This disclosure provides a composition comprising, or alternatively consisting essentially of, or yet further consisting of, a biocompatible microsphere, a biofilm-generating probiotic bacterium and a prebiotic, wherein the prebiotic comprises, or alternatively consists essentially of, or yet further consists of a nutritional food source or supplement for the culturing and/or growth of the probiotic bacterium. The composition can further comprise a prebiofilmic. The prebiofilmic comprises a substance that supports biofilm formation and durability, specifically; the prebiofilmic can be a DNA binding polypeptide or protein and/or a DNABII polypeptide or protein. In one aspect, the composition is frozen, for example flash frozen. In another aspect, the composition is lyophilized or dried in powder form. In a further aspect, it is formulated for administration as a suppository or in ingestible form (e.g., tablet). The composition can further comprise a mixture of the above-noted microspheres, e.g., a mixture containing two or more probiotic bacterium and/or two or prebiofilmics and/or two or more nutritional and/or supplement to support the culturing and/or growth of the probiotic bacterium.

[0106] In some embodiments, the prebiotic comprises a water-soluble carbohydrate selected from, but not limited to, one or more of inulin, oligofructose, fructo-oligosaccharide, galacto-oligosaccharide, glucose, maltose, maltodextrins, polydextrose, sucrose, fructose, lactose, isomaltulose, polyols, glycerol, and combinations thereof. In one aspect, the composition further comprises a solid or a liquid carrier, such as a pharmaceutically acceptable carrier.

[0107] As is apparent to those of skill in the art, the prebiotic and prebiofilmic are selected in each composition to specifically support the growth of the probiotic bacterium. By way of example only, when the probiotic bacterium comprises *L. reuteri*, the composition comprises an effective amount of sucrose, glycerol and optionally HU polypeptide or protein, to support the growth and maintenance of the probiotic when administered to the subject or patient. Non-limiting examples of prebiofilmic compositions include, without limitation, one or more of the polypeptides provided in SEQ ID NOs: 1 to 24, a c-terminal fragment thereof, or a n-terminal fragment thereof, or the additional strains and polypeptides and fragments thereof, such as the full length or the c-terminal fragment or the n-terminal fragment of those provided in **Table 4**, and equivalents of each thereof. Additional nutritional supplements for the support of other probiotic bacterium are disclosed in Bergey's Manual of Determinative Bacteriology, 9th Ed, Ed. Holt et al., WilliamsWilkins (1994),

[0108] Non-limiting examples of a probiotic bacterium for use in the composition includes, without limitation, one or more of *L. acidophilus*, *L. crispatus*, *L. gasseri*, group *L. delbrueckii*, *L. salivarius*, *L. casei*, *L. paracasei*, *L. plantarum*, *L. rhamnosus*, *L. reuteri*, *L. brevis*, *L. buchneri*, *L. fermentum*, *L. rhamnosus*, *B. adolescentis*, *B. angulation*, *B. bifidum*, *B. breve*, *B. catenulatum*, *B. infantis*, *B. lactis*, *B. longum*, *B. pseudocatenulatum*, *S. thermophiles*, or a combination thereof. As is apparent to those of skill in the art, one or more bacterium can be combined in a single composition. In some embodiments, the probiotic bacterium is *Lactobacillus reuteri*. The bacteria are available from commercial sources, such as the American Type Culture Collection (ATCC). In one aspect, the one or more probiotic bacterium in the composition supports anti-bacterial immunity. In other aspects, the one or more probiotic bacterium in the composition prevents pathogen colonization and/or limits excessive inflammatory responses by down-regulating cytokine and chemokine production. In some embodiments, the composition further comprises an agent, and the agent is selective against a pathogen, such as a competing pathogen.

[0109] The biocompatible microsphere comprises one or more of a biodegradable polymer, a non-biodegradable polymer, a metal, or a combination thereof. In some embodiments, the microsphere comprises a solid core. In some embodiments, the microsphere comprises a hollow core. In some embodiments, the prebiotic is encapsulated within the hollow core of the microsphere.

[0110] In one aspect, the disclosure provides a composition comprising, or alternatively consisting essentially of, or yet further consisting of, a PGLA-biocompatible microsphere, one or more biofilm-generating probiotic bacterium, and a nutritional supplementation comprising one or more of sucrose or glycerol in an amount to support the growth of the probiotic bacterium. The biofilm-generating probiotic bacterium may comprise *Lactobacillus reuteri* (“*L. reuteri*”). The composition may further comprise, or alternatively consist essentially of, or yet further consist of, an effective amount of HU polypeptide or protein. The composition can further comprise a pharmaceutically acceptable carrier or a biocompatible scaffold and is optionally formulated as a suppository.

[0111] The size of the microsphere can range from about 0.5 microns to about 100 microns. In certain embodiments, the microsphere is less than about 100 microns in diameter. In other embodiments, the microsphere is less than about 50 microns, or less than about 40 microns, or less than about 30 microns, less than about 20 microns, less than about 10 microns, or less than about 5 microns, or less than 3 microns to 0.5 microns in diameter. In further embodiments, the microsphere is from about 0.5 microns to about 90 microns, or to about 80 microns, or to about 70 microns, or to about 60 microns, or to about 50 microns, or to about 40 microns, or to about 30 microns, or to about 20 microns, or about 10 microns, or about 5 microns, or about 3 microns, or about 2 microns, or about 1 micron, in diameter. Alternatively, the diameter is from about 1 to about 100, or alternatively from about 1 to about 75, or alternatively from about 1 to about 50, or alternatively from about 1 to about 25, or alternatively from about 1 to about 15, or alternatively from about 1 to about 10, microns in diameter.

[0112] In some embodiments, the microsphere is a biodegradable polymer, non-limiting examples of such include poly(lactic-co-glycolic acid) (“PLGA”); polycaprolactone (“PLC”); chitosan; gelatin; DNA hydrogen; acetalated dextran; poly(lactide); poly(glycolide); poly(lactide-co-glycolide); poly(lactic acid); poly(glycolic acid); poly(lactic acid-co-glycolic acid); poly(lactide)/poly(ethylene glycol) copolymers; poly(glycolide)/poly(ethylene glycol)

copolymer; poly(lactide-co-glycolide)/poly(ethylene glycol) copolymers; poly(lactic acid)/poly(ethylene glycol) copolymer; poly(glycolic acid)/poly(ethylene glycol) copolymer; poly(lactic acid-co-glycolic acid)/poly(ethylene glycol) copolymer; poly(caprolactone); poly(caprolactone)/poly(ethylene glycol) copolymer; poly(orthoester); poly(phosphazene); poly(hydroxybutyrate); poly(hydroxybutyrate); poly(lactide-co-caprolactone); polycarbonate; polyesteramide; polyanhydride; poly(dioxanone); poly(alkylene alkylate); polyethylene glycol/polyorthoester copolymer; polyurethane; poly(amino acid); polyetherester; polyacetal; polycyanoacrylate; poly(oxyethylene)/poly(oxypropylene) copolymer; and combinations thereof. In some embodiments, the biodegradable polymer is poly(lactic-co-glycolic acid) or PLGA.

[0113] In some embodiments, the microsphere comprises a non-biodegradable polymer. Non-limiting examples of non-biodegradable polymers, include without limitation, of one or more of poly(ethylene vinyl acetate), poly(vinyl acetate), silicone polymers, polyurethanes, polysaccharides such as a cellulosic polymers and cellulose derivatives, acyl substituted cellulose acetates and derivatives thereof, copolymers of poly(ethylene glycol) and poly(butylene terephthalate), polystyrenes, polyvinyl chloride, polyvinyl fluoride, poly(vinyl imidazole), chorosulphonated polyolefins, polyethylene oxide, and copolymers and blends thereof.

[0114] In some embodiments, the microsphere comprises a metal. The metal can be selected from, but not limited to, one or more of cobalt, chromium, gold, nickel, platinum, stainless steel, titanium, tantalum, nickel-titanium, and alloys and combinations thereof.

Pharmaceutical Compositions

[0115] The composition can be formulated as a frozen composition, e.g., flash frozen, dried or lyophilized for storage and/or transport. In addition, the composition can administered alone or in combination with a carrier, such as a pharmaceutically acceptable carrier or a biocompatible scaffold. Compositions of the invention may be conventionally administered rectally as a suppository, parenterally, by injection, for example, intravenously, subcutaneously, or intramuscularly. Additional formulations which are suitable for other modes of administration include oral formulations. Oral formulations include such normally employed excipients such as, for example, pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, cellulose, magnesium carbonate and the like. These compositions take the form of solutions, suppositories, suspensions, tablets, pills, capsules,

sustained release formulations or powders and contain about 10% to about 95% of active ingredient, preferably about 25% to about 70%.

[0116] Typically, compositions are administered in a manner compatible with the dosage formulation, and in such amount as will be therapeutically effective for the disease or condition by treated. The quantity to be administered depends on the subject to be treated. Precise amounts of the composition to be administered depend on the judgment of the practitioner. Suitable regimes for initial administration and boosters are also variable, but are typified by an initial administration followed by subsequent administrations.

[0117] In many instances, it will be desirable to have multiple administrations of the compositions about, at most about or at least about 3, 4, 5, 6, 7, 8, 9, 10 days or more. The administrations will normally range from 2 day to twelve week intervals, more usually from one to two week intervals. Periodic boosters at intervals of 0.5-5 years, usually two years, may be desirable to maintain the condition of the immune system

[0118] In some embodiments, additional pharmaceutical compositions are administered to a subject to support or augment the compositions as described herein. Different aspects of the present invention involve administering an effective amount of the composition to a subject. Additionally, such compositions can be administered in combination with modifiers of the immune system. Such compositions will generally be dissolved or dispersed in a pharmaceutically acceptable carrier or aqueous medium.

[0119] The phrases "pharmaceutically acceptable" or "pharmacologically acceptable" refer to molecular entities and compositions that do not produce an adverse, allergic, or other untoward reaction when administered to an animal, or human. As used herein, "pharmaceutically acceptable carrier" includes any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like. The use of such media and agents for pharmaceutical active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active ingredients, its use in immunogenic and therapeutic compositions is contemplated.

[0120] The carrier may be a solvent or dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid poly(ethylene glycol), and the like), suitable mixtures thereof, and vegetable oils. The proper fluidity can be maintained, for example, by the use of a coating, such as lecithin, by the maintenance of the

required particle size in the case of dispersion, and by the use of surfactants. The prevention of the action of undesirable microorganisms can be brought about by various antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, sorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars or sodium chloride. Prolonged absorption of the injectable compositions can be brought about by the use in the compositions of agents delaying absorption, for example, aluminum monostearate and gelatin.

[0121] An effective amount of therapeutic composition is determined based on the intended goal. The term "unit dose" or "dosage" refers to physically discrete units suitable for use in a subject, each unit containing a predetermined quantity of the composition calculated to produce the desired responses discussed above in association with its administration, i.e., the appropriate route and regimen. The quantity to be administered, both according to number of treatments and unit dose, depends on the result and/or protection desired. Precise amounts of the composition also depend on the judgment of the practitioner and are peculiar to each individual. Factors affecting dose include physical and clinical state of the subject, route of administration, intended goal of treatment (alleviation of symptoms versus cure), and potency, stability, and toxicity of the particular composition. Upon formulation, solutions will be administered in a manner compatible with the dosage formulation and in such amount as is therapeutically or prophylactically effective. The formulations are easily administered in a variety of dosage forms, such as the type of injectable solutions described above.

Processes for Preparing Compositions

[0122] This disclosure also provides a method for preparing a composition as described herein, comprising, or alternatively consisting essentially of, or yet further consists of, the steps of admixing, contacting or culturing a biocompatible microsphere with a biofilm-generating probiotic bacterium and a prebiotic. In one aspect, the method further comprises adding or admixing a prebiofilmic that supports the formation and growth of a biofilm by the bacterium. Non-limiting examples of such include, one or more of a DNA binding polypeptide or protein and/or a DNABII polypeptide or protein. The prebiotic utilized in the method comprises a water-soluble carbohydrate, which can be selected from, but not limited to, one or more of inulin, oligofructose, fructo-oligosaccharide, galacto-oligosaccharide, glucose, maltose, maltodextrins, polydextrose, sucrose, fructose, lactose, isomaltulose, polyols, glycerol, and combinations thereof.

Therapeutic Methods

[0123] In some embodiments, a method for treating or preventing a disease in a subject is provided, comprising administering to a subject an effective amount of a composition as described above, to a subject in need of such treatment. As used herein, a “subject” intends an animal (e.g., murine, bovine, canine, feline, equine, simian) or a human. Non-limiting diseases to be treated include, but not limited to, psychological disorders, such as depression or anxiety, enteric infectious disease, infection-induced colitis, traveler’s diarrhea, inflammatory bowel disease (IBD), colitis, diarrheal illness, vaginosis, wound, burns, psoriasis, dermatitis, tooth decay, periodontitis, sinusitis, or any of chronic and/or recurrent disease that is caused by pathogenic bacteria displacing healthy bacteria or necrotizing enterocolitis (NEC). In addition, the compositions can be administered to support anti-bacterial immunity, enhancing or supporting the gastrointestinal barrier, or antagonizing disease-related bacterial infection. In some embodiments, the disease is vaginosis. In some embodiments, the disease is colitis or traveler’s diarrhea. As is apparent to the skilled artisan, the composition is specifically selected for the disease to be treated. In some embodiments, the composition further comprises a prebiofilmic. In some embodiments, the prebiofilmic comprises a DNA binding polypeptide or protein and/or a DNABII polypeptide or protein, e.g., HU, a fragment thereof and/or an equivalent of each thereof. In some embodiments, the composition is administered as a suppository.

[0124] In some embodiments, the composition of the method is administered to provide from about 1×10^7 to about 1×10^9 CFU/ml of the biofilm-generating probiotic bacterium. In some embodiments, the composition is administered at about 6, 12, 18, 24, 36, 48, and 72 hours. In some embodiments, the composition is administered in a single dose.

[0125] In some embodiments, a method of administering a probiotic is provided, comprising administering a dose of a composition as described above, comprising, or alternatively consisting essentially of, or yet consisting of, a biocompatible microsphere, a biofilm-generating probiotic bacterium, a prebiotic, and a prebiofilmic to a subject in need of such treatment. In some embodiments, the composition of the method is administered to provide from about 1×10^7 to about 1×10^9 CFU/ml of the biofilm-generating probiotic bacterium. In some embodiments, the composition is administered at about 6, 12, 18, 24, 36, 48, and 72 hours. In some embodiments, the composition is administered in a single dose.

Kits

[0126] In some embodiments, a kit containing one or more compositions as described herein is provided. The kit comprises, or alternatively consists essentially of, or yet further consists of, a composition as described above, and instructions for use. Alternatively, the kit comprises a microsphere and instructions to make the composition as described above. In one aspect, the bacteria and prebiotic are also provided in the kit.

Experimental Examples**Example 1**

[0127] To determine if *L. reuteri* in a biofilm state are superior to planktonic bacteria for establishment in the murine gut, *L. reuteri* was introduced via oral gavage, but instead of repeating the gavage daily, which is typically needed for retention of planktonic bacteria and for beneficial effects 15, 41, a single administration of *L. reuteri* was provided. The *L. reuteri* were grown in biofilm cultures or biofilm grown on poly(lactic-co-glycolic acid) microspheres, such as PLGA, or other FDA approved and biodegradable microspheres (hydrolyzed into lactic acid and glycolic acid) with diameters ranging from 20-300 μ m (Beer, et al., (1998) Poly (Lactic-Glycolic) Acid Copolymer Encapsulation of Recombinant Adenovirus Reduces Immunogenicity in Vivo, Gene Ther, 5: 740-6; Kumari, et al., (2010) Biodegradable Polymeric Nanoparticles Based Drug Delivery Systems, Colloids Surf B Biointerfaces, 75:1-18).

[0128] Similar preparations of planktonic bacteria were prepared but PLGA microspheres and prebiofilms were added just prior to gavage. As shown in Figure 3, the number of mice in which *L. reuteri* was detected in the murine colon after 7 days increased when introduced as a biofilm versus planktonic-grown cells. The presence of PLGA also enhanced the number of mice that were positive for *L. reuteri* compared to conditions where PLGA was not present regardless of whether the bacteria were planktonic or in a biofilm state; this could indicate that *L. reuteri* can initiate attachment to the PLGA, a prelude to biofilm formation, even during this brief interaction (<30 minutes). In the stomach, the only conditions where all mice retained *L. reuteri* was biofilm-grown cells with the addition of PLGA. Thus, it is evident that growing *L. reuteri* in a biofilm in the presence of PLGA enhances colonization and persistence within the stomach and colon compared to planktonic-grown cells.

Example 2 - *L. reuteri* vs *C. rodentium* in vitro

[0129] To determine if *L. reuteri* has the capacity to better compete with *C. rodentium* as either a biofilm or in planktonic state *in vitro*, a competition assay was developed. Here *C. rodentium* biofilms in glass chamber slides (LB medium, 24 hours, 37°C, 5% CO₂) were performed. *L. reuteri* (10⁸ colony forming units (CFUs)) was then added as a treatment either as planktonic or in one of three biofilm forms (biofilm, PLGA biofilm, PLGA+HU biofilm; preparation as in Figure 3) in a medium compatible with both organisms. After 16 hours, the biofilm contents of the chamber slides was removed and aliquots were plated on media selective for *L. reuteri* (MRS) and *C. rodentium* (LB). *C. rodentium* treated with *L. reuteri* biofilm showed a >2 fold decrease in CFU/ml compared to untreated (Table 1), regardless of the state of the introduced *L. reuteri*. More interesting, while all the *L. reuteri* proliferated during the 16 hour challenge, the *L. reuteri* introduced in the form of a biofilm yielded >10-fold more CFUs than when added in planktonic form.

Table 1. *L. reuteri* vs. *C. rodentium* in vitro competition assays

Condition ^a	<i>C. rodentium</i> Biofilm (CFU/ml)	<i>L. reuteri</i> Biofilm (CFU/ml)
C.rodentium biofilm		
Untreated	1.71 x 10 ⁹	n/a
+ <i>L.r</i> planktonic	6.00 x 10 ⁸	9.00 x 10 ⁷
+ <i>L.r</i> biofilm	4.65 x 10 ⁸	1.12 x 10 ⁹
+ <i>L.r</i> PLGA biofilm	5.30 x 10 ⁸	1.17 x 10 ⁹
+ <i>L.r</i> PLGA HU biofilm	4.30 x 10 ⁸	1.08 x 10 ⁹
L.reuteri biofilm		
Untreated	n/a	2.00 x 10 ⁹
+ <i>C.r</i> planktonic	9.20 x 10 ⁶	1.40 x 10 ⁹
+ <i>C.r</i> biofilm	7.90 x 10 ⁷	2.60 x 10 ⁹
PLGA + <i>C.r</i> biofilm	5.00 x 10 ⁷	2.50 x 10 ⁹
PLGA HU + <i>C.r</i> biofilm	7.25 x 10 ⁷	3.45 x 10 ⁹

^aBiofilms were treated with 10⁸ CFU of challenge condition

[0130] In the converse experiment, the *L. reuteri* biofilm was introduced first and then treated with *C. rodentium* (10⁸ CFUs) in planktonic and biofilm forms. In contrast to the previous experiment, *L. reuteri* was still able to proliferate increasing in CFUs by >10-fold

regardless of the presence of *C. rodentium* (<2-fold difference between conditions) but *C. rodentium* did not proliferate during the 16 hour challenge and was actually reduced in CFUs when introduced planktonically. These *in vitro* results show that *C. rodentium* biofilms can be effectively challenged with *L. reuteri* and when introduced in the biofilm state, *L. reuteri* persist better than planktonic cells. Moreover, preformed *L. reuteri* biofilms create a poor environment for challenge by planktonic *C. rodentium* to establish.

Example 3 - L. reuteri vs C. rodentium in vivo

[0131] To determine if *L. reuteri* has the capacity to better compete with *C. rodentium* as either a biofilm or in planktonic state *in vivo*, a version of the published competition assay was utilized Mackos, et al., (2013) Probiotic *Lactobacillus reuteri* Attenuates the Stressor-Enhanced Severity of *Citrobacter Rodentium* Infection, Infect Immun, 81:3253-63). Briefly, *L. reuteri* were introduced by oral gavage to mice as described above (*L. reuteri* planktonic vs biofilm *in vivo*). After 12 hours, an equal number of planktonic *C. rodentium* were also added by gavage. After 12 days, all mice were sacrificed for necropsy. Unlike the published work that shows that *C. rodentium* penetration of brush border epithelia and propagation to the spleen can be thwarted by daily doses of planktonic *L. reuteri* (Mackos, et al., (2013) Probiotic *Lactobacillus reuteri* Attenuates the Stressor-Enhanced Severity of *Citrobacter rodentium* Infection, Infect Immun, 81:3253-63), there was a statistically significant 10-fold drop in *C. rodentium* penetration to the spleen in the prebiofilmic treated *L. reuteri* biofilm with a single dose (Figure 4). This result is consistent with the magnitude and robustness of prebiofilmic treated probiotic biofilms of having a more durable phenotype *in vivo*.

Example 4 - Characterization Of Probiotic Therapeutic Biofilms For Endurance And Robustness.

[0132] This example has provided strong evidence that the probiotics in the biofilm state provides a superior formulation to bacteria grown planktonically. It also provides one example of how to prepare these biofilms including the frequency of dosing. In addition, the example examines the nature of the biofilm itself to begin to determine why this state outperforms planktonic bacteria. Finally, it examines the shelf life of the preparations as a prelude to reduction to practice in human hosts. Combined, this example identifies and characterizes the conditions and constituents for probiotic biofilm preparations.

Example 4.1 Effects Of Growth Phase

[0133] *L. reuteri* forms a robust biofilm *in vitro* and that *L. reuteri* in a 24-hour biofilm establishes well in the mouse gut were shown. This Example varies the age of the biofilm to determine the optimal age for biofilm establishment.

[0134] *In vivo L. reuteri* biofilms. *L. reuteri* begins to attach almost immediately when exposed to a surface. After 6 hours sufficient biomass has been produced to be both visible and to start forming the classic biofilm structures (e.g., mushroom, Abee and Kuipers, (2011) Understanding Microbial Behavior within and Outside the Host to Improve Food Functionality and Safety, Curr Opin Biotechnol, 22:133-5). *L. reuteri* biofilms are isolated at about 6, 12, 18, 24, 36, 48 and 72 hours, that have been grown on PGLA microspheres with HU and calf thymus DNA (as described above) normalizing to CFUs (108) and introducing them by gavage into mice (9 per time point from triplicate experiments). Each mouse is assessed by counting total lactobacilli levels in fecal samples daily for 12 days (cultured on MRS agar).

[0135] In addition, this Example uses a real-time PCR method to assess 16S rRNA gene sequence copy numbers for the *Lactobacillus* genus (including some species of *Weisella*, *Pediococcus*, and *Leuconostoc* due to difficulties with primer specificity) and specifically for *L. reuteri*. The 16S rRNA gene copy numbers is determined in the feces daily for 12 days, as well as in the colon, cecum, small intestine (including ileum, jejunum, and duodenum), and stomach (including the forestomach) using real-time PCR on Days 1, 3, 6, and 12 post-oral inoculation. Sham mice with and without planktonic cells serve as controls. A significant increase in *L. reuteri* levels in mice treated with biofilm-grown *L. reuteri* in comparison to sham or planktonic-treated mice is an indicator of durability and robustness.

Example 4.2 Effects Of Growth Conditions

[0136] One set of growth conditions has been used to date, standing cultures in MRS media (Jones, and Versalovic, (2009) Probiotic *Lactobacillus reuteri* Biofilms Produce Antimicrobial and Anti-Inflammatory Factors, BMC Microbiol, 9:35), at 37°C. While not an exhaustive list, here this Example varies the media, the prebiofilms as well as pH and aerobicity.

[0137] Varying growth conditions *in vitro*. In this Example, other media to grow the biofilms instead of MRS including LB, THYE (THB with yeast extract), mTSB (modified

tryptic soy broth) are used as *L. reuteri* grows in each to varying degrees. In addition, the Example also varies the starting pH to about 5.5, 6, 6.5 or 7 as *L. reuteri* growth is favored under more acidic conditions. While *L. reuteri* can be grown microaerophilically under 5% CO₂, stressful conditions of times favor biofilm growth (Flemming, and Wingender, (2010) The Biofilm Matrix, Nat Rev Microbiol, 8:623-33); here *L. reuteri* biofilms are also grown in air or in the absence of oxygen (anaerobic chamber). Finally, the Example varies the prebiofilmics of HU (about 0.1, 1, 10, 100 µg/ml) and calf thymus DNA (about 0.1, 1, 10, 100 µg/ml). All the aforementioned biofilms are assessed by CSLM with LIVE/DEAD® staining in triplicate for height, average thickness and biomass as indicators of robust growth.

[0138] Varying growth conditions *in vivo*. Conditions optimal for biofilm growth are compared against both the initial standard conditions as well as the conditions that create the poorest biofilm (control). Biofilms are introduced by oral gavage into 9 mice (from triplicate experiments) for each trial under the conditions optimized in Example 4.1. Sham mice with and without planktonic cells serve as controls. *L. reuteri* levels are assessed as in Example 4.1, on Days 1, 3, 6, and 12 post-challenge.

Example 4.3 Effects Of Bacterial Dosing

[0139] Dosing of *L. reuteri*; frequency and size. The frequency and or size of dosing improves the durability and robustness of the introduction of *L. reuteri* are determined. *L. reuteri* biofilms are grown on PLGA microspheres with added HU and calf thymus DNA for 24 hours (or an age condition as determined in Example 4.1 and 4.2). *L. reuteri* biofilms are introduced to mice by oral gavage creating a matrix of varying the dose (10^7 , 10^8 and 10^9 CFUs) as well as the frequency (single dose, or daily dose up to 3 days) yielding 9 different conditions. *L. reuteri* levels are assessed *in vivo* on Days 1, 3, 6, and 12 post gavage as outlined in Example 4.1. Nine mice (from triplicate experiments) for each condition at each time point are used. Sham mice with and without planktonic cells serve as controls.

Example 4.4 Testing Dispersed Biofilm Bacteria

[0140] Testing dispersed *L. reuteri* from biofilms. Dispersed bacteria for their endurance and robustness in the mouse gut are examined. *L. reuteri* biofilms can be dispersed by antisera to a DNABII family member (e.g., *E. coli* IHF). Here this Example tests the bacteria released (dispersed) due to anti-IHF treatment. 24 hour *L. reuteri* biofilms (no added PLGA, HU or DNA so as to facilitate dispersal) grown in chamber slides are treated with anti-IHF20.

As the peak of dispersal is about 8 to 12 hours after treatment (Goodman, et al. (2011) Biofilms Can Be Dispersed by Focusing the Immune System on a Common Family of Bacterial Nucleoid-Associated Proteins, *Mucosal Immunol*, 4:625-37), conditioned media containing dispersed *L. reuteri* after 12 hours of antibody treatment are used for introduction into mice by oral gavage. *L. reuteri* levels are assessed *in vivo* on Days 1, 3, 6, and 12 post challenge as outlined in Example 4.1. Nine mice for each time point (from triplicate experiments) with a similar number for controls using planktonic bacteria and optimized biofilm bacteria (Example 4.1 to 4.3) are used.

[0141] Biofilms are found to be superior for establishment, persistence and duration of probiotic bacteria in the gut. It is not the biofilm *per se* that possesses superior features to planktonic bacteria but the bacteria that are dispersed from biofilms. In effect, the biofilm would be acting as a dispersed-bacteria generator. Indeed, physiologic differences in dispersed bacteria as compared to laboratory grown planktonic bacteria (e.g. in antibiotic sensitivity) have been observed.

Example 4.5 Shelf Life

[0142] For reduction to practice and ease of use, *L. reuteri* preparations need to be in a sufficiently stable form.

[0143] Freezing. *L. reuteri* biofilms have been flash frozen and found no diminution in CFUs and minimum inhibitory concentration or MIC (>2 mg/ml ampicillin; MIC for planktonic *L. reuteri* <4 µg/ml) suggesting *L. reuteri* retains at least one property of its biofilm state, enhanced MIC. Optimized *L. reuteri* biofilms (Example 4.1. to 4.3) for ambient air freezing to -20°C and -80°C with and without glycerol (a cryo-protectant; See also Example 2) as well as flash freezing to -80°C (placing storage tubes with fresh bacterial suspensions in dry ice-ethanol) are examined.

[0144] Media are first removed and the resulting biofilm are scraped off and treated to freeze. Bacteria are stored at these temperatures for 1 day, 1 week or 1 month and then thawed at ambient room temperature to be used for introduction into mice by oral gavage. Nine mice from triplicate experiments are used with a similar number for controls using planktonic bacteria and optimized biofilm bacteria (Example 4.1 to 4.3). Each mouse is assessed as in Example 4.1.

[0145] Desiccation. Optimized *L. reuteri* biofilms (Example 4.1. to 4.3) via lyophilization after freezing using the optimized technique in Example 4.5 are examined. Desiccated bacteria are stored at room temperature for about 1 day, 1 week or 1 month and then rehydrated with the original biofilm volume of sterile distilled water at ambient room temperature to be used for introduction into mice by gavage. Nine mice from triplicate experiments are used with a similar number for controls using planktonic bacteria and optimized biofilm bacteria (Example 4.1 to 4.3). Each mouse is assessed as in Example 4.1.

[0146] Finally, a strain of *L. reuteri* (ATCC23272) is utilized. Additional strains of *L. reuteri* (e.g. strain 100-23, ATCCPTA6475, ATCC55730) are also examined to assess strain differences. As an additional control, *L. reuteri* strains that are commercially available (Fleet® Pedia-Lax™ Probiotic Yums™ ~ 100million CFU/tablet, *L. reuteri* Protectis®DSM 17938 and Gerber® Soothe Colic Drops ~ 100 million CFU/serving (5 drops, ~200ul), *L. reuteri* Protectis®DSM 17938) are examined. This Example finds that by dissolving each product in water and using them directly in *in vitro* competition experiments with *C. rodentium* each product is shown to be no better than the strain of *L. reuteri* in planktonic form.

Example 5 - Identification And Characterization Of Biodegradable Surfaces And Pre-Biotic Substances To Facilitate The Endurance And Robustness Of The Probiotic Biofilms

[0147] Other types of microspheres as well as inherent cargo that may facilitate either probiotic growth or inhibit pathogens are explored.

Example 5.1 Testing Empty Microspheres

[0148] Empty microspheres *in vitro*, DNA, gelatin, Polylactic acid, Poly- α -caprolactone, chitosan and acetalated dextran are examined in this Example.

[0149] While PLGA microspheres are utilized as a surface to grow the biofilms, there are other FDA approved or GRAS biodegradable microspheres that may prove advantageous for the goals. As shown in **Table 2**, 5 additional types of microspheres are examined (F. Chellat, et al., (2000) In Vitro and in Vivo Biocompatibility of Chitosan-Xanthan Polyionic Complex, J Biomed Mater Res., 51:107-16; D. Costa, et al., (2012) Swelling Behavior of a New Biocompatible Plasmid DNA Hydrogel, Colloids Surf B Biointerfaces, 92:106-12; Kauffman, et al., (2012) Synthesis and Characterization of Acetalated Dextran Polymer and Microparticles with Ethanol as a Degradation Product, ACS Appl Mater Interfaces, 4:4149-

55; Kumari, et al., (2010) Biodegradable Polymeric Nanoparticles Based Drug Delivery Systems, Colloids Surf B Biointerfaces, 75:1-18; Sinha, et al., (2004) Poly-Epsilon-Caprolactone Microspheres and Nanospheres: An Overview, Int J Pharm, 278:1-23; Topuz, and O. Okay, (2009) Formation of Hydrogels by Simultaneous Denaturation and Cross-Linking of DNA, Biomacromolecules, 10:2652-61). Thus, DNA can be used as the microsphere material as it is the basis of the EPS for biofilms.

[0150] This is an example of an optimization strategy *in vitro* and *in vivo* from Example 4. Microspheres from materials in **Table 2** and repeat Examples 4.1-4.5 are constructed. Microspheres that fail to support *in vitro* robust biofilm growth using height, thickness and biomass, as initial metrics; are no longer be considered. Likewise those microsphere types that subsequently fail to surpass *in vivo* metrics relative to planktonic bacteria are also no longer be considered. Shelf life with and without bacteria, stability at low pH (gastric conditions) are also contemplated.

Table 2. Types Of Biodegradable Polymeric Microspheres To Be Tested

Type of Microsphere	Size Range(μm)	Degradation Products	FDA Approval
PLGA			
(poly-D,L,-lactide-co-glycolide) ^{a,b}	20-300	Lactic acid, Glycolic acid	X
PCL			
(poly-ε-caprolactone) ^{a,c}	10-500	6-hydroxyhexanoic acid 3-(2-hydroxyethoxy)propanoic acid	X
Chitosan ^{a,d}	20-550	Glucosamine,	X
N-acetyl-D-glucosamine			
Gelatin ^a	35-100	Amino acids	X
DNA (hydrogel) ^{e,f}	Variable	DNA, ethylene glycol diglycidyl ether	
Acetalated dextran ^g	0.1-10	Dextran, Acetone, Ethanol	

a. Kumari A, 2010, Colloid Surface B, supra.

b. Beer SJ, 1998, Gene Ther ., supra.

c. Sinha VR, 2004, Int J Pharm., supra.

d. Chellat F, 2000, J Biomed Mater Res., supra.

e. Costa D, 2012, Colloid Surface B., supra.

f. Topuz F, 2009, Biomacromolecules., supra.

g. Kauffman KJ, 2012, App Mater Interfac., supra.

Example 5.2 Testing Prebiotic Nutrients And Additives That Favor Probiotics As Cargo

[0151] The cargo of PLGA is known to diffuse slowly or not even at all relative to the rate of microsphere hydrolysis (Fredenberg, et al. (2011) The Mechanisms of Drug Release in Poly(Lactic-Co-Glycolic Acid)-Based Drug Delivery Systems--a Review, Int J Pharm, 415:34-52). Here microspheres with prebiotic cargo were synthesized and evaluated for their ability to support *L. reuteri* growth *in vitro* and *in vivo* in the mouse models.

[0152] This examines nutrients *in vitro*. As an initial test cargo is loaded into PLGA microsphere during their synthesis (so as to be encapsulated in the interior of the microsphere). These cargos include, but not limited to, inulin, fructo-oligosaccharides, and galacto-oligosaccharides as they support lactobacilli growth. In addition, microspheres with MRS media and/or glycerol are made, as the former is restrictive to Gram-negative bacteria some of which are pathogens and the latter stimulates reuterin production (an antimicrobial molecule believed to give *L. reuteri* an advantage against competing bacteria). *L. reuteri* biofilm growth on these microspheres is performed on the conditions observed in Example 4 (or Example 5.1 with a variant microsphere) and is adjudicated by CSLM for height, thickness and biomass.

[0153] This example tests prebiofilmics *in vitro*. As in Example 4.2, the ability of prebiofilmics (HU and DNA) was examined as cargo in PLGA microspheres (and the microsphere types from Example 5.1) to support *in vitro* biofilm growth. In each case, biofilms are grown under the conditions observed in Example 4 with microspheres synthesized in the presence of HU and or DNA (so as to be encapsulated in the interior of the microsphere) and are adjudicated by CSLM for height, thickness and biomass.

[0154] This example tests a combination of prebiotics and prebiofilmics *in vitro*. Here a matrix of combinations of the two probiotic and two prebiofilmic cargos is created (all 16 combinations of two, all 4 combinations of 3, and the single combination of all 4 equaling 21 total combinations) to find the suitable prebiotics or prebiofilmics. In each case, biofilms are grown under the conditions observed in Example 1 with PLGA microspheres (and the microsphere types from Example 5.1) synthesized in the presence of cargo and are adjudicated by CSLM for height, thickness and biomass.

[0155] This example tests optimized components *in vivo*. Conditions from Example 5.2 that yielded the biofilms are used for *in vivo* experiments. The four most promising

conditions for PLGA microsphere cargo (or the two most promising PLGA and two most promising other type of microsphere from Example 5.1) are tested on nine mice each derived from triplicate experiments. Each mouse is assessed as in Example 4.1 on Days 1, 3, 6, and 12 post-*L. reuteri* introduction. Sham mice (no bacteria) and planktonic bacteria serve as controls.

Example 5.3 Prebiotic Nutrients That Impede Pathogens

[0156] Microspheres containing various probiotic cargos to determine if they support pathogen biofilm growth are examined. The microspheres containing prebiofilmics come into contact with a pathogen (i.e., *C. rodentium* strain DBS120 (pCRP1::Tn5)) as well as probiotic.

[0157] This example tests pathogen impeding nutrients *in vitro*. The same prebiotic and prebiofilmic substances from Example 5.2, are used as cargo to grow *in vitro* biofilms. *C. rodentium* is grown in LB media and used to seed biofilms with PLGA and the aforementioned cargos. Biofilms is adjudicated by CSLM for height, thickness and biomass compared to empty PLGA microspheres.

[0158] This example tests pathogen impeding nutrients *in vivo*. Taking into consideration the results from *in vitro* biofilm data in Example 5.3, four cargos for *C. rodentium* biofilm growth and use them *in vivo* in mouse models are examined. Nine mice for each condition per time point (from triplicate experiments) are used with planktonic *C. rodentium* and sham (no bacteria) as controls. *C. rodentium* levels in the stool is determined via culture on all days post oral *C. rodentium* administration. On Days 1, 6, 12 and 24 post-oral *C. rodentium* administration, the colon is removed and transected longitudinally so that inflammatory cytokines (e.g., TNF- α), inflammatory mediators (e.g., inducible nitric oxide synthase (iNOS)), and chemokines (e.g., CCL2) can be assessed in half of the colon via real-time RT-PCR. In the second half of the tissue, immunohistochemistry is used to assess leukocyte infiltration into the colon (e.g., F4/80+ macrophages; myeloperoxidase (MPO)+ polymorphonuclear cells). While the aforementioned immune components are necessary for protective immunity against *C. rodentium*, when produced in excess, they can lead to tissue-damaging colitis. Thus, colonic pathology is assessed via H&E staining on the second half of the tissue.

[0159] Thus, microsphere biofilm preparations can include alternative types of microspheres and varying cargo. It is Applicants' belief that biofilms (regardless of surface) are superior to planktonic bacteria at seeding probiotic colonization *in vivo*.

[0160] Non-limiting examples cargos, include without limitation specific effectors of innate immunity that reduce inflammation, part of the process leading to dysbiosis. For example, microspheres can comprise conditioned media from *L. reuteri* as *L. reuteri* produce such substances. Likewise other bacteria are within the scope of this disclosure, e.g., *C. rodentium* and *L. reuteri*, in general for pathologies due to dysbiosis.

Experiment No. 6 - Characterization Of L. reuteri's Capacity To Limit Or Displace The Murine Gut Enteropathogenic Bacterium C. rodentium.

[0161] Previous examples have identified and characterized the means to create an *L. reuteri* biofilm with the good endurance and robustness in the murine gut while also examining how these conditions might affect the murine enteropathogenic *C. rodentium*. In this Example, the formulations of *L. reuteri* biofilms to determine if they can reduce the effects of *C. rodentium*, or even partially clear introduced or extant pathogen are examined.

Example 6.1 Testing Optimized L. reuteri Biofilm Growth Conditions In C. rodentium Challenge; Making Of L. reuteri

[0162] *In vitro* challenge of *L. reuteri* with *C. rodentium*. This Example systematically determines which of the conditions improves *L. reuteri* prophylaxis against *C. rodentium* challenge. As shown in **Table 3**, the Example systematically performs *in vitro* experiments where *L. reuteri* is grown in biofilms (about 12, 24, and 48 hours biofilms to reflect varying age) and then treated with varying quantities of planktonic *C. rodentium* (10^7 , 10^8 and 10^9 CFUs). *L. reuteri* biofilm growth conditions from Examples 4.2 (e.g., for prebiofilms as the media for challenge needs to at least facilitate growth of both bacterial species) as well as 2.1, 2.2 and 2.3 are examined. Mixed biofilms are evaluated after 12 or 24 hours of treatment by CSLM and by plate counts on selective media to determine which species' architecture and numbers dominate under each condition. Controls include each bacterial species without the other under each condition (e.g., the addition of *C. rodentium* added to PLGA microspheres without *L. reuteri* in each chamber slide). All experiments are done in triplicate.

Table 3. Systematic Approach To Find Optimal Conditions For *L. Reuteri* Vs. *C. Rodentium* Challenge

Condition	<i>In vitro</i>	<i>In vivo</i>
Example 1		
<i>L. reuteri</i> 6, 12, 18, 24, 36, 48, 72 hours biofilm	X	X
<i>L. reuteri</i> grown in different media (MRS, LB, THYE, mTSB)	X	
<i>L. reuteri</i> + HU at 0.1, 1, 10, 100 µg/ml	X	
<i>L. reuteri</i> + DNA at 0.1, 2, 10, 50 µg/ml	X	
<i>L. reuteri</i> grown at varying pH (5.5, 6, 6.5, 7)	X	
Optimal growth conditions	X	X
<i>L. reuteri</i> dose CFU/ml (10^7 , 10^8 , 10^9)	X	X
<i>L. reuteri</i> dosage frequency (1, 2, 3 days)	X	X
Dispersed <i>L. reuteri</i> bacteria	X	X
Shelf life of <i>L. reuteri</i> biofilm preparations (freezing, desiccation)	X	
Example 2		
<i>L. reuteri</i> + PLGA, PCL, chitosan, gelatin, DNA, acetalated dextran microspheres	X	
<i>L. reuteri</i> + nutrient/prebiofilmic/nutrient-prebiofilmic microspheres	X	
<i>L. reuteri</i> + 4 most promising conditions of loaded microspheres	X	X
Prebiotic nutrients that impede pathogens	X	X
Example 3		
<i>L. reuteri</i> 12, 24, 48 hours biofilm challenge of <i>C. rodentium</i> planktonic 10^7 , 10^8 , 10^9 CFU	X	X
<i>C. rodentium</i> challenge of <i>L. reuteri</i> at 12, 24, 36 hours post-treatment with <i>L. reuteri</i>		X
Established <i>C. rodentium</i> infection challenged by <i>L. reuteri</i> with top 3 conditions at 10^7 , 10^8 , 10^9 CFU	X	X

[0163] *In vivo* challenge of *L. reuteri* with *C. rodentium*. *L. reuteri* biofilm preparations for introduction into animals are prioritized based on the greatest retention or supremacy of *L. reuteri* observed. In addition, *L. reuteri* is prepared based on any successes derived from Examples 4.1, 4.4 and 4.5. In general, *L. reuteri* biofilms are introduced 12 hours prior to oral challenge with *C. rodentium*. Triplicate experiments are conducted for a final sample size of 9 mice for each condition and time point that are assessed at 1, 6, 12, and 24 days post-challenge (peak *C. rodentium* infection occurs at about Day 12). *C. rodentium* levels in

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the stool are assessed and pathogen-induced colitis is assessed as in Example 5.3. *L. reuteri* levels are also assessed as in Example 4.1. In every case, controls include *C. rodentium* without *L. reuteri* and *C. rodentium* challenge plus planktonic *L. reuteri*.

Example 6.2 Testing Dosing Of Challenge Conditions

[0164] Dosing frequency and timing of *L. reuteri* with challenge by *C. rodentium* *in vivo*. This Example tests here how dosing of *L. reuteri* affect its ability to act as a prophylactic against *C. rodentium* challenge. The Example prioritizes the top three *L. reuteri* dosing conditions to reflect the most robust and durable results derived from Example 4.3. The Example then uses these conditions to challenge these *L. reuteri* treated mice with *C. rodentium* (about 12, 24 or 36 hours after the final *L. reuteri* treatment). Nine mice (from triplicate experiments) are used for each condition and time point. Vehicle mice infected with *C. rodentium* and single planktonic *L. reuteri* serve as controls. *C. rodentium* levels and pathogen-induced colitis are assessed on Days 1, 6, 12 and 24 post-challenge as in Example 5.3, with *L. reuteri* levels assessed as in Example 4.1.

Example 6.3 Testing Therapeutic Probiotic Challenge After Pathogenic Treatment Based on The Results In Examples 6.1, and 6.2

[0165] In Examples 6.1 and 6.2, conditions for using *L. reuteri* as a prophylactic against the pathologies caused by *C. rodentium* have been optimized. Here *C. rodentium* was introduced before *L. reuteri* to determine what effects challenge with *L. reuteri* has on extant *C. rodentium* pathogenesis.

[0166] Challenge of *C. rodentium* by *L. reuteri* biofilms *in vitro*. This Example shows that *L. reuteri* biofilms effectively challenged *C. rodentium* biofilms more effectively than planktonic *L. reuteri*. Here *L. reuteri* in biofilm form under conditions patterned after the three conditions from Example 6.1 is used. Briefly, *C. rodentium* biofilms (12, 24 or 36 hours) are challenged with *L. reuteri* biofilms (10^7 , 10^8 and 10^9 CFUs). Mixed biofilms are evaluated after 12 or 24 hours after *L. reuteri* challenge of *C. rodentium* biofilms by CSLM and by plate counts on selective media to determine which species' architecture and numbers dominated under each condition, respectively. Controls include each bacterial species without the other under each condition (e.g., the addition of *L. reuteri* to chamber slides without extant *C. rodentium*). All experiments are done in triplicate.

[0167] Challenge of *C. rodentium* by *L. reuteri* biofilms *in vivo*. Here this Example determines if *L. reuteri* biofilms can challenge prior *C. rodentium* infection in the murine model. Three different *C. rodentium* conditions (single gavage 12, 24 or 36 hours) prior to challenge with *L. reuteri* were examined. Four *L. reuteri* biofilm conditions including dosing (Example 6.2) are used to challenge *C. rodentium*. At least two of these conditions are derived from Example 6.1. Nine mice from triplicate experiments are used to test each of these 12 conditions. Pathogen-induced colitis is assessed as in Example 5.3, with *L. reuteri* levels assessed as in Example 4.1.

[0168] Here, this Example determines how effective *L. reuteri* introduced in the form of a biofilm is as a prophylactic to *C. rodentium* challenge and as a treatment for extant *C. rodentium* infection. To date, *L. reuteri* under the conditions fails to clear pathogens like *C. rodentium*, so it's particularly important if conditions where a probiotic can prevent or even cure an enteropathogenic infection can be found. Results here provide a rationale for future probiotic approaches.

[0169] Finally, the *in vitro* assays are performed on other pathogens as a prelude to future *in vivo* experiments. Pathogens included in an *in vitro* survey are enteric pathogens with different modes of infection, including invasive pathogens (e.g., *Salmonella enterica* subspecies Typhimurium and *Shigella flexneri*), additional A/E pathogens (e.g., Enterohemorrhagic *E. coli* O157:H7; and Enteropathogenic *E. coli*), and toxin-producing pathogens (e.g., *Vibrio cholera* and Enterotoxigenic *E. coli*); the rate limiting step in these experiments is finding co-culturing conditions that sufficiently mimic the *in vivo* state.

Example 7 - Statistical Analyses And Determination Of Sample Size

[0170] Most of the experiments involve multiple parameters and groups. Thus, two, three, or four factor analysis of variance (ANOVA) are primarily used. As an example of the statistical approach, in Example 4.1, a between subject ANOVA is used with probiotic (i.e., probiotic vs. vehicle control), condition (i.e., biofilm vs. planktonic), and time of culture (i.e., 6, 12, 18, 24, or 36 hours) as between subjects variables. Because different groups of mice are harvested on Days 1, 3, 6, and 12 post-oral inoculation, day of harvest is also used as a between subjects variable.

[0171] A significant 4-way interaction is interpreted first using post hoc independent samples t-tests with Modified Bonferroni correction factor applied for multiple comparisons.

Afterward, 3-way and 2-way interactions are interpreted via post hoc testing, followed by interpretation of main effects. This general approach is followed for both *in vitro* and *in vivo* experiments.

[0172] Because of the inherent variability of *in vivo* experiments, considerable time was spent determining the sample size that would be needed to identify statistically significant differences between groups. A power analysis conducted using preliminary data investigating *C. rodentium* levels after *L. reuteri* administration with six different groups (preliminary sample size of 6), a population mean of 3.95, and population variance of .75, indicated that to obtain statistical significance with $\alpha=.05$, while maintaining power at 0.8, a sample size of $n=9$ per condition per time point would be needed. Thus, all animal experiments involve a sample size of nine per treatment and time point. This is accomplished by combining data from triplicate experiments, each containing $n=3$ mice per treatment and time point.

[0173] Probiotics have been widely used for digestive health benefits, although few actually prevent pathogen colonization and reduce the inflammatory response. The effects of probiotic bacteria can be significantly improved by the manner in which they are introduced into the host; specifically by growing them in the form of a biofilm. The data suggest that colonization *in vivo* by the probiotic *L. reuteri* is greatly enhanced when grown as a biofilm compared to planktonic-grown cells. In addition, when *L. reuteri* was grown in the presence of a biodegradable surface (PLGA), colonization was also increased indicating that the conditions were optimized that allowed a vast improvement in regards to *L. reuteri* establishment within the host.

[0174] Unexpectedly and surprisingly, Applicants demonstrated both *in vitro* and *in vivo* that treatment of *L. reuteri* as a biofilm in the presence of PLGA prior to challenging with the bacterial pathogen *C. rodentium*, caused a significant reduction in the number of *C. rodentium* compared to planktonic *L. reuteri* treatment. These data reveal that a probiotic can colonize better when presented as a biofilm, indicating that the way in which bacteria are introduced can greatly reflect the outcome of disease.

Experiment No. 8

[0175] Probiotic microbes have also been shown to reduce anxiety and depression in otherwise healthy humans and laboratory animals. A combination of *Lactobacillus helveticus*

and *B. longum* administered daily for 30 days was shown to reduce anxiety and depression in healthy human volunteers and in healthy rats (Messoudi et al. (2011) Beneficial psychological effects of a probiotic formulation (Lactobacillus helveticus R0052 and Bifidobacterium longum R0175) in healthy human volunteers. Br J Nutri, 105:755-764).

[0176] This experiment tests whether *L. reuteri* preparations are superior at reducing infectious colitis-induced sickness, anxiety-like, and depressive-like behaviors using the same experimental design as the above-noted studies assessing the effects on infectious colitis itself, with minor modifications. The primary difference is that animal behavior will be assessed as well as, circulating cytokines, circulating hormones, and neuronal activation in the brain.

[0177] To determine whether prophylactic *L. reuteri* can prevent *C. rodentium*-induced sickness, such as anxiety-like and depressive-like behavior, prophylactic treatment with *L. reuteri* biofilms is assessed to determine if *C. rodentium* will prevent bacterium-induced sickness, anxiety-, and depressive-like behaviors. Preparations of *L. reuteri* biofilms that are found to be superior in *in vitro* assays are administered to mice via oral gavage 12 hours prior to oral challenge with *C. rodentium*. Triplicate experiments are conducted for a final sample size of 9 mice for each condition and time point that are assessed at 1, 6, 12, and 24 days post-challenge (peak *C. rodentium* infection occurs about Day 12). At each time point, animal behavior is assessed for locomotor activity (such as on the open field test), anxiety-like behavior (such as in the light:dark preference test and elevated plus maze), depressive-like behavior (such as on the tail suspension test and Porsolt forced swim task), and sickness behavior (such as with the sucrose preference test). Blood serum cytokines associated with emotional and illness behavior (e.g., IL-1 α/β and IL-6) are assessed on each day. Circulating corticosterone levels will also be assessed. Neuronal activation in the brain, especially the paraventricular nucleus of the hypothalamus, are assessed using c-Fos immunoreactivity.

[0178] Whether *L. reuteri* can be used as a therapeutic to treat *C. rodentium*-induced sickness, anxiety-like, and depressive-like behavior also is assessed. For example, the compositions are tested to determine whether treating an established *C. rodentium* infection will reduce sickness, anxiety-, and depressive-like behaviors. Preparations of *L. reuteri* that are found to be superior in *in vitro* assays are administered to mice via an oral gavage 12, 24, and/or 36 hours after oral challenge with *C. rodentium*. On days 1, 6, 12, and 24 post-*C. rodentium* challenge, animal behavior is assessed for locomotor activity (such as on the open

field test), anxiety-like behavior (such as in the light:dark preference test and elevated plus maze), depressive-like behavior (such as on the tails suspension test and Porsolt forced swim task), and sickness behavior (such as with the sucrose preference test). Circulating cytokines associated with emotional and illness behavior (e.g., IL-1 α / β and IL-6) are assessed on each day. Circulating corticosterone levels are also assessed. Neuronal activation in the brain, especially the paraventricular nucleus of the hypothalamus, are assessed using c-Fos immunoreactivity.

[0179] These examples permit to modify conditions to create a more robust and long-lasting probiotic, and once established, and allow us to test these conditions in an *in vivo* model that could ultimately reflect treatments for bacterial infections and human disease.

Example 9 – NEC

[0180] Probiotic administration may be beneficial in the prevention of NEC. However, probiotics must be administered daily to achieve beneficial effects. Applicants describe herein a novel probiotic delivery system in which the probiotics are grown as a biofilm on the surface of prebiotic-loaded biocompatible microspheres, allowing enhanced and more durable efficacy with only a single treatment.

[0181] Following cesarean delivery, neonatal rats were subjected to experimental NEC [hypoxia/hypothermia/hypertonic feeds (stress)]. On day 1, pups were randomized to receive a single enteral dose of the following: (1) vehicle only (100 μ L sterile water) (N=32); (2) 1×10^9 CFU/mL *Lactobacillus reuteri* (N=9); (3) prebiotic-loaded biocompatible microspheres (N=12); or (4) 1×10^9 CFU/mL *L. reuteri* coupled with prebiotic-loaded biocompatible microspheres (N=33). Control pups were unstressed (N=10). Pups were sacrificed when clinical signs of NEC developed or by 96 hours after birth. A verified histologic NEC injury grading system was used to measure the incidence and severity of NEC, with Grade 2 or greater injury considered to be consistent with NEC.

[0182] As graphically depicted in FIG. 5, 69% of untreated stressed pups developed NEC. Compared to untreated stressed pups, 67% of pups treated with *L. reuteri* (p=0.329), 50% of pups treated with prebiotic-loaded microspheres (p=0.364), and 33% of pups treated with *L. reuteri* coupled with prebiotic-loaded microspheres (p=0.003) developed NEC. No unstressed pups developed NEC.

[0183] A single dose of a Lactobacillus biofilm coupled with prebiotic-loaded biocompatible microspheres reduces the incidence of NEC and therefore is an effective treatment. Without being bound by theory, the compositions as disclosed herein are prophylactic in their use in subjects in need of such treatment.

Example 10: Dessication Tolerance Assay

[0184] Another advantage of Applicants' invention is improved long-term survival of probiotic bacteria. A dessication tolerance assay was used to test stability and viability of the bacteria combined with the microspheres. The assay can generally be conducted by performing the following steps. To grow the bacteria culture, transfer 1ml to a 1.5ml of the culture to a microcentrifuge tube (1 tube per condition per time period to be tested). Add about 10 μ l of hydrated microspheres, trehalose, or nothing to the tube. Incubate the tube for 30 minutes and then pellet the cells via centrifugation. Remove the supernatant and wash the pellet twice with sterile saline. Afterwards, remove all liquid from the pellet. Place the open tube on top of Drierite within an enclosed container and place the container into an incubator at 40°C. After 7 days, remove the tubes, rehydrate, and suspend the pellet in 1ml of growth medium for 5 minutes. Then, serially dilute and plate for viable colony forming units. Finally, repeat rehydration and plating at 30 days and 90 days.

[0185] *P. fluorescens* and a proprietary *Azospirillum* sp. were placed after 90 days incubation at 40°C while on top of Drierite, a strong desiccant, and then rehydrated and tested for viability. *P. fluorescens* with no microspheres shows a complete loss of colony forming units (CFU) after just one week in these conditions, whereas when incubated with cellulose microspheres, there are 10^5 viable cells after 90 days in these conditions. *Azospirillum* sp. shows significant loss of CFUs after 30 days and complete loss after 90 days when grown without the microsphere formulation; however, when stored in harsh conditions with the microspheres, 10^6 CFU/ml of *Azospirillum* sp. are viable even after 90 days.

Example 11: Acid Tolerance Protocol (48-Well Plate)

[0186] Microspheres filled with *L. reuteri* growth medium as cargo were utilized to provide a surface that leaches buffered nutrients to the bacteria for the formation of a biofilm that enhances survivability at low pH. Bacterial cells with microspheres show over a 2 log increase in viable colony forming units compared to cells without microspheres after sitting in pH 2 gastric acid for 4 hours. Further, *L. reuteri* with microspheres show increased

adherence to mouse colonic cells, addressing the problem of poor colonization and sustainability of orally administered bacteria. Taken together, the novel microsphere formulations not only increase survivability at low pH, but also contribute to colonization of beneficial bacteria in the gut, making *L. reuteri* a more efficient probiotic.

[0187] An acid tolerance protocol assay, such as that used to generate the above information, can generally be conducted by performing the following steps. First, grow 5ml culture overnight at 37°C (5% CO₂ or anaerobically) and then dilute the culture 1:2500 in a fresh medium. Transfer 500ml per condition per time period to be tested into a 48-well plate. Transfer ~10ul of hydrated microspheres or nothing into the well. Afterwards, incubate at 37°C 5% CO₂ (or anaerobically) for 20 hours overnight. At 20 hours, remove the spent media from the biofilm and replace with pH 2 gastric acid. At two and four hours, remove the acid from the biofilm and suspend cells by pipette mixing in the growth medium. Finally, serial dilute and plate the cells.

Example 12: Cellular Adherence Assay

[0188] Microspheres filled with *L. reuteri* growth medium as cargo were utilized to provide a surface that leaches buffered nutrients to the bacteria for the formation of a biofilm that enhances survivability at low pH. Bacterial cells with microspheres show over a 2 log increase in viable colony forming units compared to cells without microspheres after sitting in pH 2 gastric acid for 4 hours. Further, *L. reuteri* with microspheres show increased adherence to mouse colonic cells, addressing the problem of poor colonization and sustainability of orally administered bacteria. These results show that novel microsphere formulations not only increase survivability at low pH, but also contribute to colonization of beneficial bacteria in the gut, making *L. reuteri* a more efficient probiotic.

[0189] A cellular adherence assay, such as that used to generate the above information, can generally be conducted by performing the following steps. First, grow up a mammalian cell culture line and dilute to ~10⁶ cells/ml. Transfer 500ul of the diluted mammalian cell lines to a 48-well plate. Then, grow to confluence (time varies, at least 16 hours) and grow the bacterial culture overnight. Afterwards, transfer 500ul of the bacterial culture to a 1.5ml microcentrifuge tube (1 tube per condition per time period). Pellet the bacterial cells via centrifugation and wash the pellet 2-3 times to remove all growth medium. Resuspend the pelleted bacteria in a cell line culture medium. Add either microspheres hydrated in a cell line culture medium, microspheres hydrated in MRS, or nothing to the suspended bacteria.

[0190] Remove the growth medium from the confluent mammalian cell culture wells. Aspirate the bacterial conditions with cell line growth medium into mammalian cell culture wells. Incubate at 37°C 5% CO₂. After 1 hour, remove the supernatant spent medium from each well and wash cells with sterile PBS twice to remove non-adhered bacteria. Add 500ul trypsin to each well to dislodge adhered mammalian cells from the plastic and incubate at 37°C for 5-10 minutes. Thoroughly mix the liquid in each well to resuspend the mammalian cells. Then, serially dilute and plate to calculate the number of bacteria that remained adhered to the mammalian cells. At 4 and 8 hours, remove the supernatant spent medium from each well and wash cells with sterile PBS twice to remove non-adhered bacteria. Add 500 µl trypsin to each well to dislodge adhered mammalian cells from the plastic and incubate at 37°C for 5-10 minutes. Thoroughly mix the liquid in each well to resuspend the mammalian cells. Then, serially dilute and plate to calculate the number of bacteria that remained adhered to the mammalian cells.

[0191] Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention belongs.

[0192] The inventions illustratively described herein may suitably be practiced in the absence of any element or elements, limitation or limitations, not specifically disclosed herein. Thus, for example, the terms “comprising,” “including,” “containing,” etc. shall be read expansively and without limitation. Additionally, the terms and expressions employed herein have been used as terms of description and not of limitation, and there is no intention in the use of such terms and expressions of excluding any equivalents of the features shown and described or portions thereof, but it is recognized that various modifications are possible within the scope of the invention claimed.

[0193] Thus, it should be understood that the materials, methods, and examples provided here are representative of preferred embodiments, are exemplary, and are not intended as limitations on the scope of the invention.

[0194] The invention has been described broadly and generically herein. Each of the narrower species and sub-generic groupings falling within the generic disclosure also form part of the invention. This includes the generic description of the invention with a proviso or

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negative limitation removing any subject matter from the genus, regardless of whether or not the excised material is specifically recited herein.

[0195] In addition, where features or aspects of the invention are described in terms of Markush groups, those skilled in the art will recognize that the invention is also thereby described in terms of any individual member or subgroup of members of the Markush group.

[0196] All publications, patent applications, patents, and other references mentioned herein are expressly incorporated by reference in their entirety, to the same extent as if each were incorporated by reference individually. In case of conflict, the present specification, including definitions, will control.

[0197] Other embodiments are set forth within the following claims.

Table 4

Gram (+) - only HU, Gram (-) - all have HU some also IHF

Bacteria strain	Abbreviation	Protein name(s)
<i>S. sobrinus</i> 6715	<i>Ss</i>	1310 (HU)
<i>S. pyogenes</i> MGAS10270	<i>Spyog</i>	SPY1239 (HU)
<i>S. gordonii</i> Challis NCTC7868	<i>Sg</i>	SGO_0701 (HlpA)
<i>S. agalactiae</i> (Group B Strep)2603V/R	GBS	SAG_0505 (Hup)
<i>S. mutans</i> UA159	<i>Sm</i>	Smu_589 (HU)
<i>S. pneumoniae</i> R6	<i>Spneu</i>	spr1020 (HU)
<i>S. gallolyticus</i> UCN34 (<i>S. bovis</i>)	<i>Sgall</i>	YP_003430069 (HlpA)
<i>S. aureus</i> MW2	<i>Sa</i>	MW1362 (HU)
<i>S. epidermidis</i> RP62A	<i>Se</i>	SERP1041 (Hup)
<i>E. coli</i> K12-MG1655	<i>Ec</i>	b1712 (HimA) b0912 (HimD) (HupA) (HupB)
<i>H. influenza</i> KW20 Rd	<i>Hi</i>	HI1221 (HimA) HI1313 (HimD) HI0430 (HupA)

TABLE 4 (CONTINUED)

<i>Salmonella enteric serovar typhi CT18</i>	<i>Salm</i>	Sty1771	(HimA)
		Sty0982	(HimD)
<i>Aggregatibacter actinomycetemcomitans D11S-1</i>	<i>Aa</i>	YP_003255965	(IHfalpha)
		YP_003256209	(IHfB)
		YP_003255304	(HU)
<i>P. gingivalis W83</i>	<i>Pg</i>	PG_0121	(Hup-1)
		PG_1258	(Hup-2)
<i>N. gonorrhoeae FA1090 (Oklahoma)</i>	<i>Ng</i>	NGO603	(IHfβ)
		NGO030	(IHfα)
<i>N. meningitides MC58</i>	<i>Nm</i>	NMB_0729	(HimA)
		NMB_1302	(HimA)
<i>P. aeruginosa</i>	<i>Pa</i>	PA3161	(HimD)
		PA1804	(HupB)
		PA2758	(HimA)
<i>H. pylori 26695</i>	<i>Hp</i>	Hp0835	(Hup)
<i>B. burgdorferi B31</i>	<i>Bb</i>	BB_0232	(Hbb)
<i>Moraxella catarrhalis RH4</i>	<i>Mc</i>	YP_003626307	(HimA)
		YP_003627027	(HimD)
		YP_003626775	(HupB)

TABLE 4 (CONTINUED)

<i>V. cholera</i> El Tor N16961	Vc	VC_0273	(HupA)
		VC_1914	(HipB)
		VC_1919	(HupB)
		VC_1222	(HimA)
<i>Burkholderia cenocepacia</i> HI2424	Bc	Bcen2424_1048	(IHFB)
		Bcen2424_1481	(IHFA)
<i>Burkholderia pseudomallei</i> 668	Bp	BURPS668_2881	(IHFB)
		BURPS668_1718	(IHFA)
<i>Mycobacterium tuberculosis</i> CDC1551	Mtb	MT_3064	(HU)
		MSMEG_2389	(Hup)
<i>Mycobacterium smegmatis</i> MC2	Ms	TDE_1709	(HU)
<i>Treponema denticola</i> ATCC 35405	Td	TP_0251	(DNA binding protein II)
<i>Treponema palladium</i> Nichols	Tp	PREME0022_2103	(HupB)
<i>Prevotella melaninogenica</i> ATCC 25845	Pm	PREME0022_0268	(HupA)
		PREME0022_0341	(Hup)
		PREME0022_0340	(HimA)

TABLE 4 (CONTINUED)

<i>Prevotella intermedia</i> 17	<i>Pi</i>	PIN_A0704	(Hup)
		PIN_A1504	(Hup-2)
		PIN_0345	(HimA)
		PIN_0343	(Hypothetical protein)
<i>Bordetella pertusis</i> Tohama 1	<i>Bpert</i>	BP2572	(Ihfa)
		BP3530	(HupB)
		BP0951	(Ihfb)
		Ef1550	(hup)
<i>Enterococcus faecalis</i> V583	<i>Ef</i>		

References

- T. Abee, and O. P. Kuipers, Understanding Microbial Behavior within and Outside the Host to Improve Food Functionality and Safety, *Curr Opin Biotechnol*, 22 (2011), 133-5
- L. T. Axelsson, et al., Production of a Broad Spectrum Antimicrobial Substance by *Lactobacillus Reuteri*, *Microbial Ecology in Health and Disease*, 2 (1989)
- M. T. Bailey, et al., Stressor Exposure Disrupts Commensal Microbial Populations in the Intestines and Leads to Increased Colonization by *Citrobacter Rodentium*, *Infect Immun*, 78 (2010), 1509-19
- S. J. Beer, et al., Poly (Lactic-Glycolic) Acid Copolymer Encapsulation of Recombinant Adenovirus Reduces Immunogenicity in Vivo, *Gene Ther*, 5 (1998), 740-6
- X. M. Ben, et al., Low Level of Galacto-Oligosaccharide in Infant Formula Stimulates Growth of Intestinal Bifidobacteria and Lactobacilli, *World J Gastroenterol*, 14 (2008), 6564-8
- R. E. Black, et al., W. H. O. Child Health Epidemiology Reference Group of, and Unicef, "Global, Regional, and National Causes of Child Mortality in 2008: A Systematic Analysis", *Lancet*, 375 (2010), 1969-87
- D. Borenshtein, et al., Utility of the *Citrobacter Rodentium* Infection Model in Laboratory Mice, *Curr Opin Gastroenterol*, 24 (2008), 32-7
- C. P. Braegger, and T. T. MacDonald, Immune Mechanisms in Chronic Inflammatory Bowel Disease, *Ann Allergy*, 72 (1994), 135-41
- K. A. Brandstetter, et al., Antibodies Directed against Integration Host Factor Mediate Biofilm Clearance from Nasopore, *Laryngoscope* (2013)
- I. M. Carroll, et al., Alterations in Composition and Diversity of the Intestinal Microbiota in Patients with Diarrhea-Predominant Irritable Bowel Syndrome, *Neurogastroenterol Motil*, 24 (2012), 521-30, e248
- C. Chassard, M. et al., Functional Dysbiosis within the Gut Microbiota of Patients with Constipated-Irritable Bowel Syndrome, *Aliment Pharmacol Ther*, 35 (2012), 828-38
- F. Chellat, M. et al., In Vitro and in Vivo Biocompatibility of Chitosan-Xanthan Polyionic Complex, *J Biomed Mater Res*, 51 (2000), 107-16

- D. Costa, J. et al., Swelling Behavior of a New Biocompatible Plasmid DNA Hydrogel, *Colloids Surf B Biointerfaces*, 92 (2012), 106-12
- J. C. De Man, M. et al., A Medium for the Cultivation of Lactobacilli, *J Applied Bacteriology*, 23 (1960), 130-35
- K. A. Eaton, A. et al., Probiotic Lactobacillus Reuteri Ameliorates Disease Due to Enterohemorrhagic Escherichia Coli in Germfree Mice, *Infect Immun*, 79 (2011), 185-91
- L. Eckmann, Animal Models of Inflammatory Bowel Disease: Lessons from Enteric Infections, *Ann N Y Acad Sci*, 1072 (2006), 28-38
- H. C. Flemming, and J. Wingender, The Biofilm Matrix, *Nat Rev Microbiol*, 8 (2010), 623-33
- S. Fredenberg, M. et al., The Mechanisms of Drug Release in Poly(Lactic-Co-Glycolic Acid)-Based Drug Delivery Systems--a Review, *Int J Pharm*, 415 (2011), 34-52
- M. O. Freire, P. et al., Development of an Animal Model for Aggregatibacter Actinomycetemcomitans Biofilm-Mediated Oral Osteolytic Infection: A Preliminary Study, *J Periodontol*, 82 (2011), 778-89
- S. D. Goodman, K. P. et al., Biofilms Can Be Dispersed by Focusing the Immune System on a Common Family of Bacterial Nucleoid-Associated Proteins, *Mucosal Immunol*, 4 (2011), 625-37
- J. E. Gustave, et al., Targeting Bacterial Integration Host Factor to Disrupt Biofilms Associated with Cystic Fibrosis, *J Cyst Fibros*, 12 (2013), 384-9
- National Institutes of Health, 2013, <http://grants2.nih.gov/grants/guide/pa-files/PA-06-537.html>
- P. Hecmarajata, and J. Versalovic, Effects of Probiotics on Gut Microbiota: Mechanisms of Intestinal Immunomodulation and Neuromodulation, *Therap Adv Gastroenterol*, 6 (2013), 39-51
- L. M. Higgins, et al., Citrobacter Rodentium Infection in Mice Elicits a Mucosal Th1 Cytokine Response and Lesions Similar to Those in Murine Inflammatory Bowel Disease, *Infect Immun*, 67 (1999), 3031-9
- F. Ito, H. et al., Factors Affecting the Loading Efficiency of Water-Soluble Drugs in Plga Microspheres, *Colloids Surf B Biointerfaces*, 61 (2008), 25-9

- S. E. Jones, and J. Versalovic, Probiotic *Lactobacillus Reuteri* Biofilms Produce Antimicrobial and Anti-Inflammatory Factors, *BMC Microbiol*, 9 (2009), 35
- S. S. Justice, B. et al., Aberrant Community Architecture and Attenuated Persistence of Uropathogenic *Escherichia Coli* in the Absence of Individual Ihf Subunits, *PLoS One*, 7 (2012), e48349
- K. J. Kauffman, C. et al., Synthesis and Characterization of Acetalated Dextran Polymer and Microparticles with Ethanol as a Degradation Product, *ACS Appl Mater Interfaces*, 4 (2012), 4149-55
- A. Kumari, et al., Biodegradable Polymeric Nanoparticles Based Drug Delivery Systems, *Colloids Surf B Biointerfaces*, 75 (2010), 1-18
- S. L. Lebeis, et al., Protective and Destructive Innate Immune Responses to Enteropathogenic *Escherichia Coli* and Related a/E Pathogens, *Future Microbiol*, 3 (2008), 315-28
- Y. P. Lin, et al., Probiotic *Lactobacillus Reuteri* Suppress Proinflammatory Cytokines Via C-Jun, *Inflamm Bowel Dis*, 14 (2008), 1068-83
- S. A. Luperchio, and D. B. Schauer, Molecular Pathogenesis of *Citrobacter Rodentium* and Transmissible Murine Colonic Hyperplasia, *Microbes Infect*, 3 (2001), 333-40
- C. Lupp, M. L. Robertson, et al., Host-Mediated Inflammation Disrupts the Intestinal Microbiota and Promotes the Overgrowth of Enterobacteriaceae, *Cell Host Microbe*, 2 (2007), 119-29
- K. Machiels, et al., A Decrease of the Butyrate-Producing Species *Roseburia Hominis* and *Faecalibacterium Prausnitzii* Defines Dysbiosis in Patients with Ulcerative Colitis, *Gut* (2013)
- A. R. Mackos, et al., Probiotic *Lactobacillus Reuteri* Attenuates the Stressor-Enhanced Severity of *Citrobacter Rodentium* Infection, *Infect Immun*, 81 (2013), 3253-63
- X. C. Morgan, et al., Dysfunction of the Intestinal Microbiome in Inflammatory Bowel Disease and Treatment, *Genome Biol*, 13 (2012), R79
- R. Mundy, et al., *Citrobacter Rodentium* of Mice and Man, *Cell Microbiol*, 7 (2005), 1697-706
- J. P. Nataro, and J. B. Kaper, Diarrheagenic *Escherichia Coli*, *Clin Microbiol Rev*, 11 (1998), 142-201

L. A. Novotny, et al., Structural Stability of Burkholderia Cenocepacia Biofilms Is Reliant on Edna Structure and Presence of a Bacterial Nucleic Acid Binding Protein, PLoS One, 8 (2013), e67629

Food and Agriculture Organization of the United Nations and World Health Organization, "Health and Nutritional Properties of Probiotics in Food Including Powdered Milk with Live Bacteria", (2001)

G. A. Preidis, et al., Probiotics Stimulate Enterocyte Migration and Microbial Diversity in the Neonatal Mouse Intestine, FASEB J, 26 (2012), 1960-9

L. A. Sarmiento-Rubiano, et al., Dietary Supplementation with Sorbitol Results in Selective Enrichment of Lactobacilli in Rat Intestine, Res Microbiol, 158 (2007), 694-701

O. Schreiber, et al., Lactobacillus Reuteri Prevents Colitis by Reducing P-Selectin-Associated Leukocyte- and Platelet-Endothelial Cell Interactions, Am J Physiol Gastrointest Liver Physiol, 296 (2009), G534-42

V. R. Sinha, et al., Poly-Epsilon-Caprolactone Microspheres and Nanospheres: An Overview, Int J Pharm, 278 (2004), 1-23

N. Takemura, et al., Inulin Prolongs Survival of Intragastrically Administered Lactobacillus Plantarum No. 14 in the Gut of Mice Fed a High-Fat Diet, J Nutr, 140 (2010), 1963-9

F. Topuz, and O. Okay, Formation of Hydrogels by Simultaneous Denaturation and Cross-Linking of DNA, Biomacromolecules, 10 (2009), 2652-61

M. Wlodarska, et al., Antibiotic Treatment Alters the Colonic Mucus Layer and Predisposes the Host to Exacerbated Citrobacter Rodentium-Induced Colitis, Infect Immun, 79 (2011), 1536-45

Q. Xia, et al., Quantitative Analysis of Intestinal Bacterial Populations from Term Infants Fed Formula Supplemented with Fructo-Oligosaccharides, J Pediatr Gastroenterol Nutr, 55 (2012), 314-20

What is claimed is:

1. A composition comprising a biocompatible microsphere, a biofilm-generating probiotic bacterium and a prebiotic, wherein the prebiotic comprises a nutritional supplementation for the probiotic bacterium.
2. The composition of claim 1, further comprising a prebiofilmic.
3. The composition of claim 2, wherein the prebiofilmic comprises an agent that supports biofilm formation and durability.
4. The composition of claim 3, wherein the prebiofilmic is a DNA binding polypeptide or protein and/or a DNABII polypeptide or protein or an equivalent of each thereof, optionally, a polypeptide comprising one or more of SEQ ID NO: 1 to 24, or a biologically active fragment or equivalent of each thereof, alone or in combination.
5. The composition of any one of claims 1-4, wherein the prebiotic comprises a water-soluble carbohydrate, wherein the water-soluble carbohydrate comprises one or more of inulin, oligofructose, fructo-oligosaccharide, galacto-oligosaccharide, glucose, maltose, maltodextrins, polydextrose, sucrose, fructose, lactose, isomaltulose, polyols, glycerol, and combination thereof.
6. The composition of any one of claims 1-5, further comprising a pharmaceutically acceptable carrier or a biocompatible scaffold.
7. The composition of any one of claims 1-6, wherein the probiotic bacterium is one or more of *L. acidophilus*, *L. crispatus*, *L. gasseri*, group *L. delbrueckii*, *L. salivarius*, *L. casei*, *L. paracasei*, *L. plantarum*, *L. rhamnosus*, *L. reuteri*, *L. brevis*, *L. buchneri*, *L. fermentum*, *L. rhamnosus*, *B. adolescentis*, *B. angulation*, *B. bifidum*, *B. breve*, *B. catenulatum*, *B. infantis*, *B. lactis*, *B. longum*, *B. pseudocatenulatum*, *S. thermophiles*, and a combination thereof.
8. The composition of claim 7, wherein the probiotic bacterium is *Lactobacillus reuteri* ("*L. reuteri*").
9. The composition of claim 1 or 8, wherein the probiotic bacterium provides one or more of supporting anti-bacterial immunity, enhancing or supporting the gastrointestinal barrier, or antagonizing disease-related bacterial infections.

10. The composition of claim 1 or 8, wherein the probiotic bacterium prevents pathogen colonization and/or limits excessive inflammatory responses by down-regulating cytokine and chemokine production.
11. The composition of any one of claims 1-10, wherein the biocompatible microsphere comprises one or more of a biodegradable polymer, a non-biodegradable polymer, a metal, or a combination thereof.
12. The composition of claim 11, wherein the microsphere comprises a solid core.
13. The composition of claim 11, wherein the microsphere comprises a hollow core.
14. The composition of claim 13, wherein the prebiotic is encapsulated within the hollow core.
15. The composition of any one of claims 1-14, further comprising an agent, wherein the agent is selective against a pathogen.
16. The composition of any one of claims 1-15, wherein the microsphere is a biodegradable polymer.
17. The composition of claim 16, wherein the biodegradable polymer is one or more of Sephadex, Sephadex G-25, poly(lactic-co-glycolic acid) ("PLGA"); polycaprolactone ("PLC"); chitosan; gelatin; DNA hydrogen; acetalated dextran; poly(lactide); poly(glycolide); poly(lactide-co-glycolide); poly(lactic acid); poly(glycolic acid); poly(lactic acid-co-glycolic acid); poly(lactide)/poly(ethylene glycol) copolymers; poly(glycolide)/poly(ethylene glycol) copolymer; poly(lactide-co-glycolide)/poly(ethylene glycol) copolymers; poly(lactic acid)/poly(ethylene glycol) copolymer; poly(glycolic acid)/poly(ethylene glycol) copolymer; poly(lactic acid-co-glycolic acid)/poly(ethylene glycol) copolymer; poly(caprolactone); poly(caprolactone)/poly(ethylene glycol) copolymer; poly(orthoester); poly(phosphazene); poly(hydroxybutyrate); poly(hydroxybutyrate); poly(lactide-co-caprolactone); polycarbonate; polycyanoacrylate; polyanhydride; poly(dioxanone); poly(alkylene alkylate); polyethylene glycol/polyorthoester copolymer; polyurethane; poly(amino acid); polyetherester; polyacetal; polycyanoacrylate; poly(oxyethylene)/poly(oxypropylene) copolymer; and a combination thereof.

18. The composition of claim 17, wherein the biodegradable is poly(lactic-co-glycolic acid) ("PGLA") and/or Sephadex.
19. The composition of any one of claims 1-15, wherein the microsphere comprises a non-biodegradable polymer.
20. The composition of claim 19, wherein the non-biodegradable polymer is one or more of poly(ethylene vinyl acetate), poly(vinyl acetate), silicone polymers, polyurethanes, polysaccharides such as a cellulosic polymers and cellulose derivatives, acyl substituted cellulose acetates and derivatives thereof, copolymers of poly(ethylene glycol), poly(butylene terephthalate), polystyrenes, polyvinyl chloride, polyvinyl fluoride, poly(vinyl imidazole), chorosulphonated polyolefins, polyethylene oxide, and copolymers and blends thereof.
21. The composition of any one of claims 1-15, wherein the microsphere comprises a metal.
22. The composition of claim 21, wherein the metal comprises one or more of cobalt, chromium, gold, nickel, platinum, stainless steel, titanium, tantalum, nickel-titanium, and alloys and combinations thereof.
23. The composition of any one claims 1-22, wherein the microsphere comprises an average diameter of from about 0.5 microns to about 75 microns.
24. A method for preparing a composition comprising admixing a biocompatible microsphere with a biofilm-generating probiotic bacterium and a prebiotic.
25. The method of claim 24, further comprising admixing a prebiofilmic.
26. The method of claim 25, wherein the prebiofilmic comprises a DNA binding polypeptide or protein and/or a DNABII polypeptide or protein or an equivalent of each thereof.
27. The method of any one of claims 24-26, wherein the prebiotic comprises a water-soluble carbohydrate, which comprises one or more of inulin, oligofructose, fructo-oligosaccharide, galacto-oligosaccharide, glucose, maltose, maltodextrins, polydextrose, sucrose, fructose, lactose, isomaltulose, polyols, glycerol, and combinations thereof.

28. The method of any one of claims 24-27, wherein the probiotic bacterium comprises one or more of *L. acidophilus*, *L. crispatus*, *L. gasseri*, group *L. delbrueckii*, *L. salivarius*, *L. casei*, *L. paracasei*, *L. plantarum*, *L. rhamnosus*, *L. reuteri*, *L. brevis*, *L. buchneri*, *L. fermentum*, *L. rhamnosus*, *B. adolescentis*, *B. angulation*, *B. bifidum*, *B. breve*, *B. catenulatum*, *B. infantis*, *B. lactis*, *B. longum*, *B. pseudocatenulatum*, *S. thermophiles*, and combinations thereof.
29. The method of claim 28, wherein the probiotic bacterium comprises *Lactobacillus reuteri*.
30. The method of any one of claims 24-29, wherein the biocompatible microsphere comprises one or more of a biodegradable polymer, a non-biodegradable polymer, a metal, combinations thereof.
31. The method of claim 30, wherein the microsphere comprises a solid core.
32. The method of claim 30, wherein the microsphere comprises a hollow core.
33. The method of claim 32, wherein the prebiotic is encapsulated within the hollow core.
34. The method of any one of claims 24-33, wherein the microsphere is a biodegradable polymer.
35. The method of claim 34, wherein the biodegradable polymer comprises one or more of Sephadex, Sephadex G-25, poly(lactic-co-glycolic acid); polycaprolactone; chitosan; gelatin; DNA hydrogen; acetalated dextran; poly(lactide); poly(glycolide); poly(lactide-co-glycolide); poly(lactic acid); poly(glycolic acid); poly(lactic acid-co-glycolic acid); poly(lactide)/poly(ethylene glycol) copolymers; poly(glycolide)/poly(ethylene glycol) copolymer; poly(lactide-co-glycolide)/poly(ethylene glycol) copolymers; poly(lactic acid)/poly(ethylene glycol) copolymer; poly(glycolic acid)/poly(ethylene glycol) copolymer; poly(lactic acid-co-glycolic acid)/poly(ethylene glycol) copolymer; poly(caprolactone); poly(caprolactone)/poly(ethylene glycol) copolymer; poly(orthoester); poly(phosphazene); poly(hydroxybutyrate); poly(hydroxybutyrate); poly(lactide-co-caprolactone); polycarbonate; polyesteramide; polyanhydride; poly(dioxanone); poly(alkylene alkylate); polyethylene glycol/polyorthoester copolymer; polyurethane; poly(amino acid); polyetherester; polyacetal;

polycyanoacrylate; poly(oxyethylene)/poly(oxypropylene) copolymer and combinations thereof.

36. The method of claim 35, wherein the biodegradable polymer comprises poly(lactic-co-glycolic acid) and/or Sephadex.

37. The method of any one of claims 24-33, wherein the microsphere comprises a non-biodegradable polymer.

38. The method of claim 37, wherein the non-biodegradable polymer comprises one or more of poly(ethylene vinyl acetate), poly(vinyl acetate), silicone polymers, polyurethanes, polysaccharides such as a cellulosic polymers and cellulose derivatives, acyl substituted cellulose acetates and derivatives thereof, copolymers of poly(ethylene glycol) and poly(butylene terephthalate), polystyrenes, polyvinyl chloride, polyvinyl fluoride, poly(vinyl imidazole), chorosulphonated polyolefins, polyethylene oxide, and copolymers and blends thereof.

39. The method of any one of claims 24-33, wherein the microsphere comprises a metal.

40. The method of claim 39, wherein the metal comprises one or more of cobalt, chromium, gold, nickel, platinum, stainless steel, titanium, tantalum, nickel-titanium, alloys and combinations thereof.

41. A method for treating or preventing a disease suitably treated by a biofilm in a subject in need thereof, comprising administering to the subject an effective amount of the composition of any one of claims 1-23.

42. The method of claim 41, wherein the diseases comprises one or more of inflammatory bowel disease (IBD), colitis, enteric infectious disease, diarrheal illness, vaginosis, necrotizing enterocolitis (NEC), wound, burns, psoriasis, dermatitis, tooth decay, periodontitis, sinusitis, infection-induced colitis, traveler's diarrhea, psychological stress, psychological disorders, or any of chronic or recurrent disease that is caused by pathogenic bacteria displacing healthy bacteria.

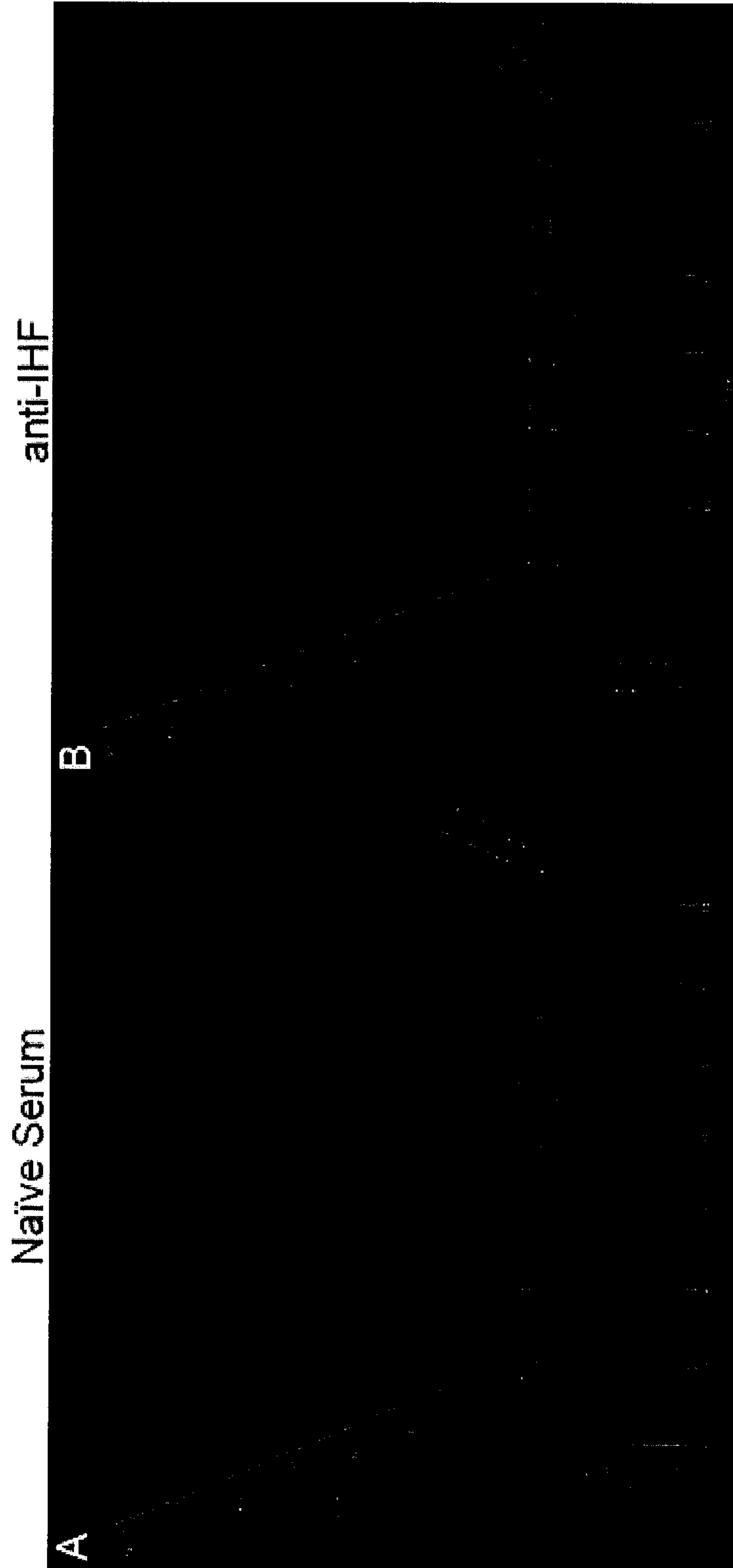
43. The method of claim 42, wherein the disease is vaginosis.

44. The method of claim 42, wherein the disease is colitis or NEC.

45. The method of any one of claims 41-44, wherein the composition is administered as a suppository.
46. A method of administering a probiotic to a subject comprising administering a dose of a composition of any one of claims 1-23 to a subject, thereby administering the probiotic.
47. The method of any one of claims 41-46, wherein the composition is administered to provide from about 1×10^7 to about 1×10^9 CFU/ml of the biofilm-generating probiotic bacterium.
48. The method of any one of claims 41-46, wherein the composition is administered at about 6, 12, 18, 24, 36, 48, and 72 hours.
49. The method of any one of claims 41-46, wherein the composition is administered in a single dose.
50. A formulation comprising the composition of any one of claims 1-23, wherein the composition is formulated in a dosage form selected from the group consisting of: suppository, within a biocompatible scaffold, powder, liquid, capsule, chewable tablet, swallowable tablet, buccal tablet, troche, lozenge, soft chew, solution, suspension, spray, tincture, decoction, infusion, and combinations thereof.
51. A kit comprising the composition of any one of claims 1-23, and instructions for use.
52. A composition comprising a PGLA-biocompatible microsphere, one or more biofilm-generating probiotic bacterium comprising at least *Lactobacillus reuteri* ("*L. reuteri*"), and a nutritional supplementation comprising one or more of sucrose or glycerol in an amount to support the growth of the probiotic bacterium.
53. The composition of claim 1, further comprising an effective amount of HU polypeptide or protein.
54. The composition of claim 52 or 53 further comprising a pharmaceutically acceptable carrier or a biocompatible scaffold.
55. The composition of any one of claims 52-54, wherein the microsphere has a diameter in the range of from about 0.5 microns to 75 microns.

56. A method for preparing the composition of claim 52, comprising admixing a PGLA-biocompatible microsphere with an effective amount of one or more biofilm-generating probiotic bacterium comprising at least *Lactobacillus reuteri* ("*L. reuteri*"), and a nutritional supplementation comprising one or more of sucrose or glycerol in an amount to support the growth of the probiotic bacterium.
57. The method of claim 56, further comprising admixing an effective amount of HU polypeptide or protein.
58. A method for treating a gastrointestinal disorder, comprising administering to a subject in need thereof an effective amount of the composition of 52-55, thereby treating the gastrointestinal disorder.
59. The method of claim 58, wherein the gastrointestinal disorder is one or more of infection-induced colitis, inflammatory bowel disease, necrotizing enterocolitis (NEC) or traveler's diarrhea.
60. The method of claim 58 or 59, wherein the composition is administered as a suppository formulation.
61. A kit comprising the composition of any one of claims 52-55, and instructions for use.

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2/4

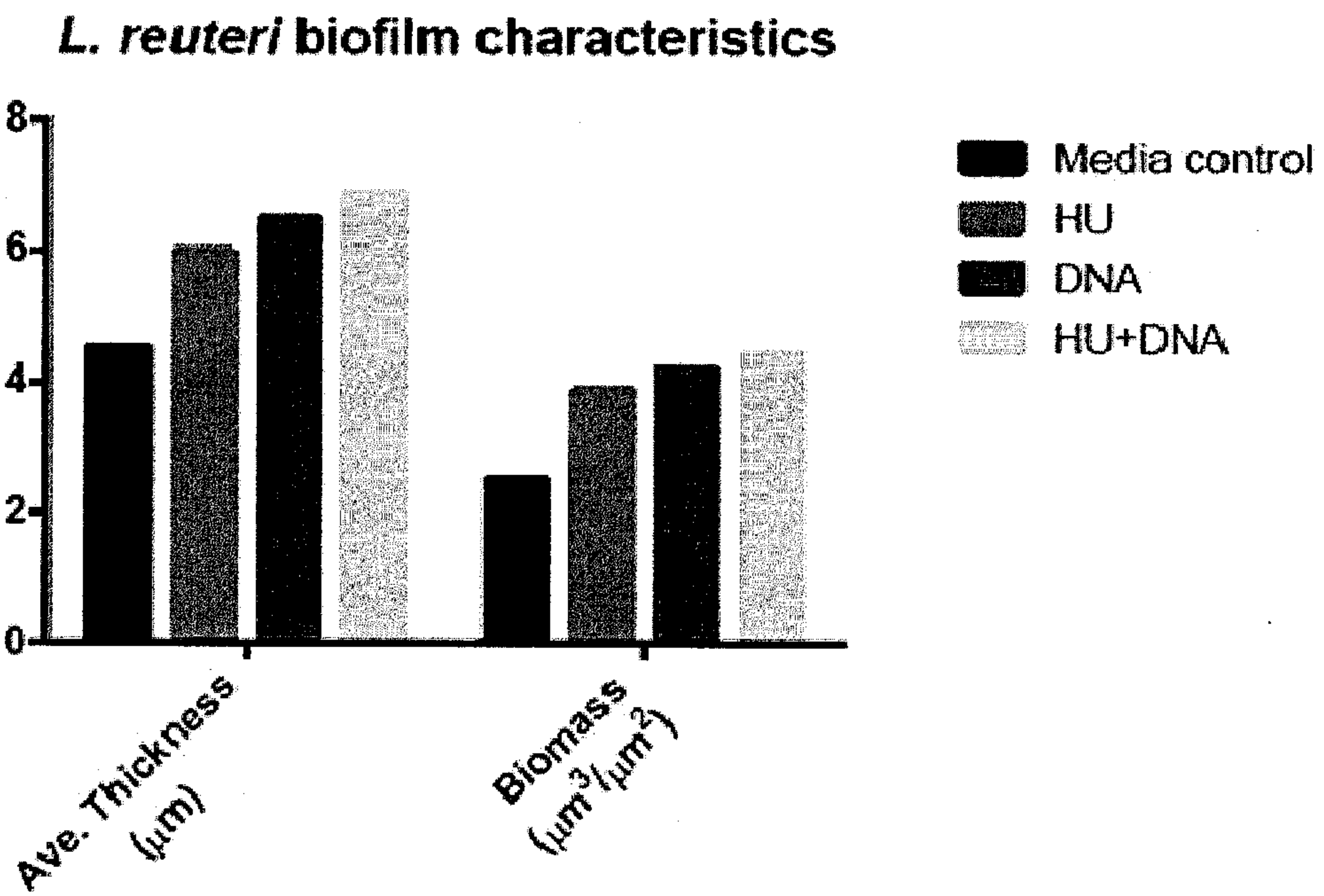


FIG. 2

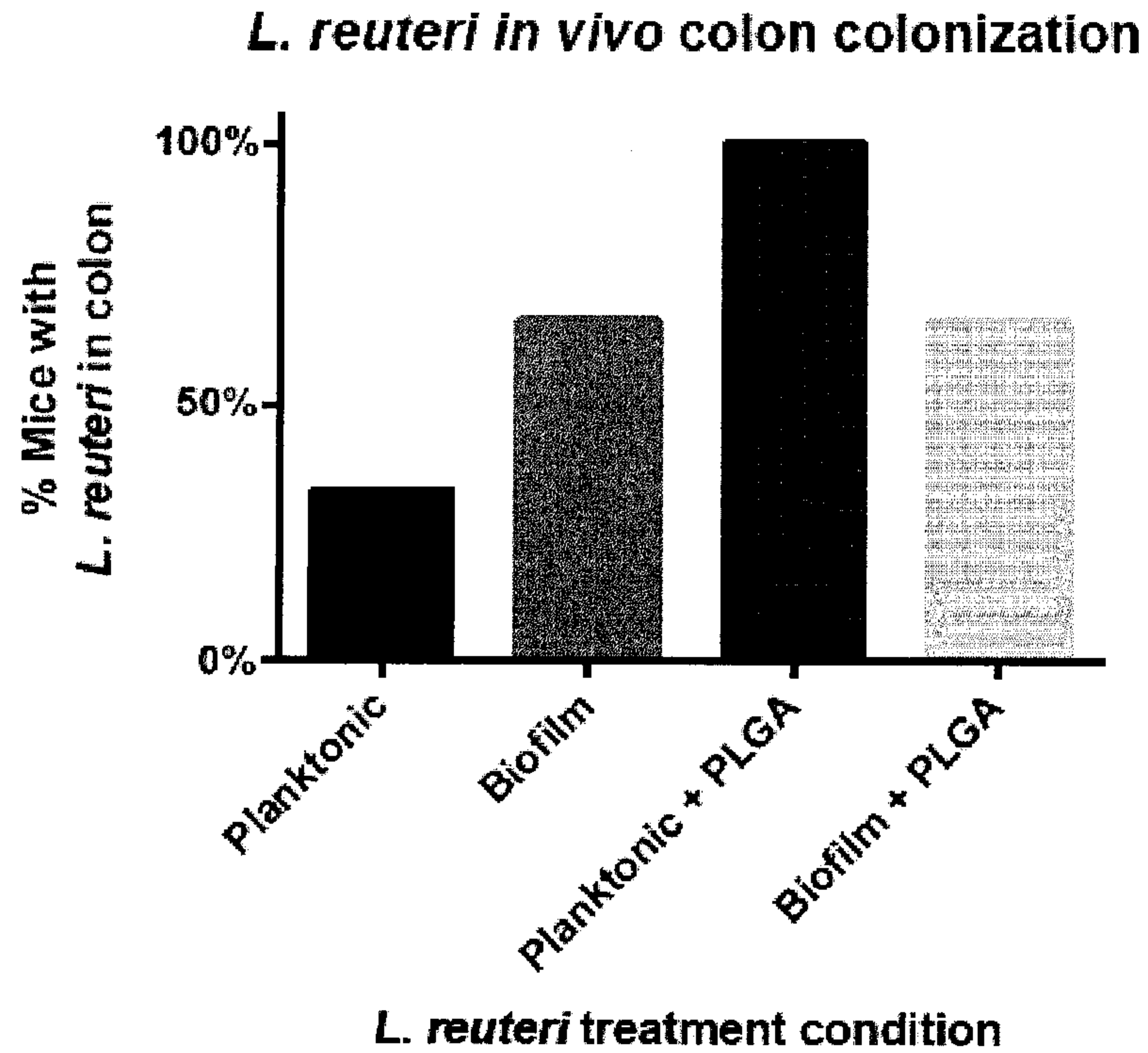


FIG. 3

3/4

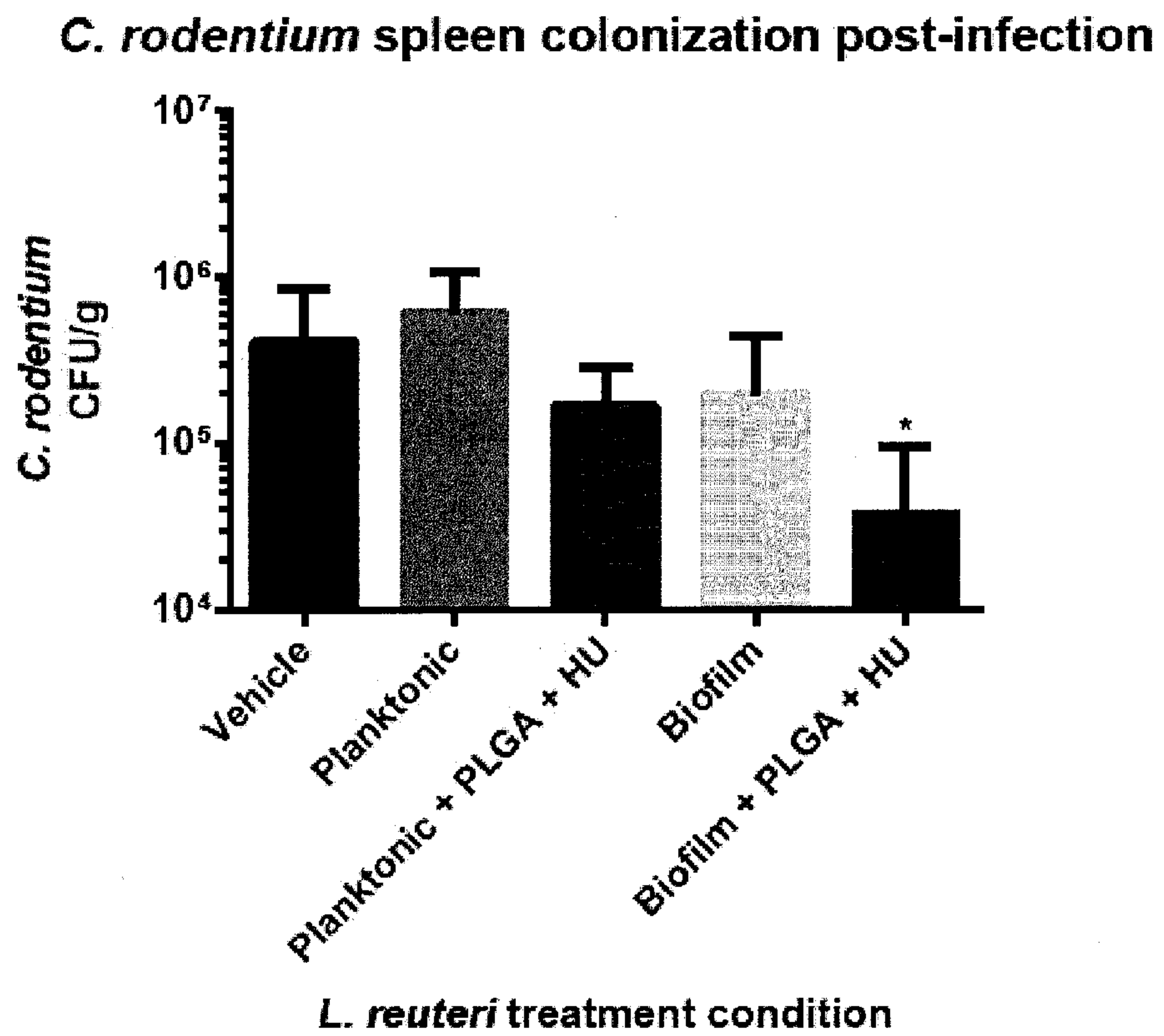


FIG. 4

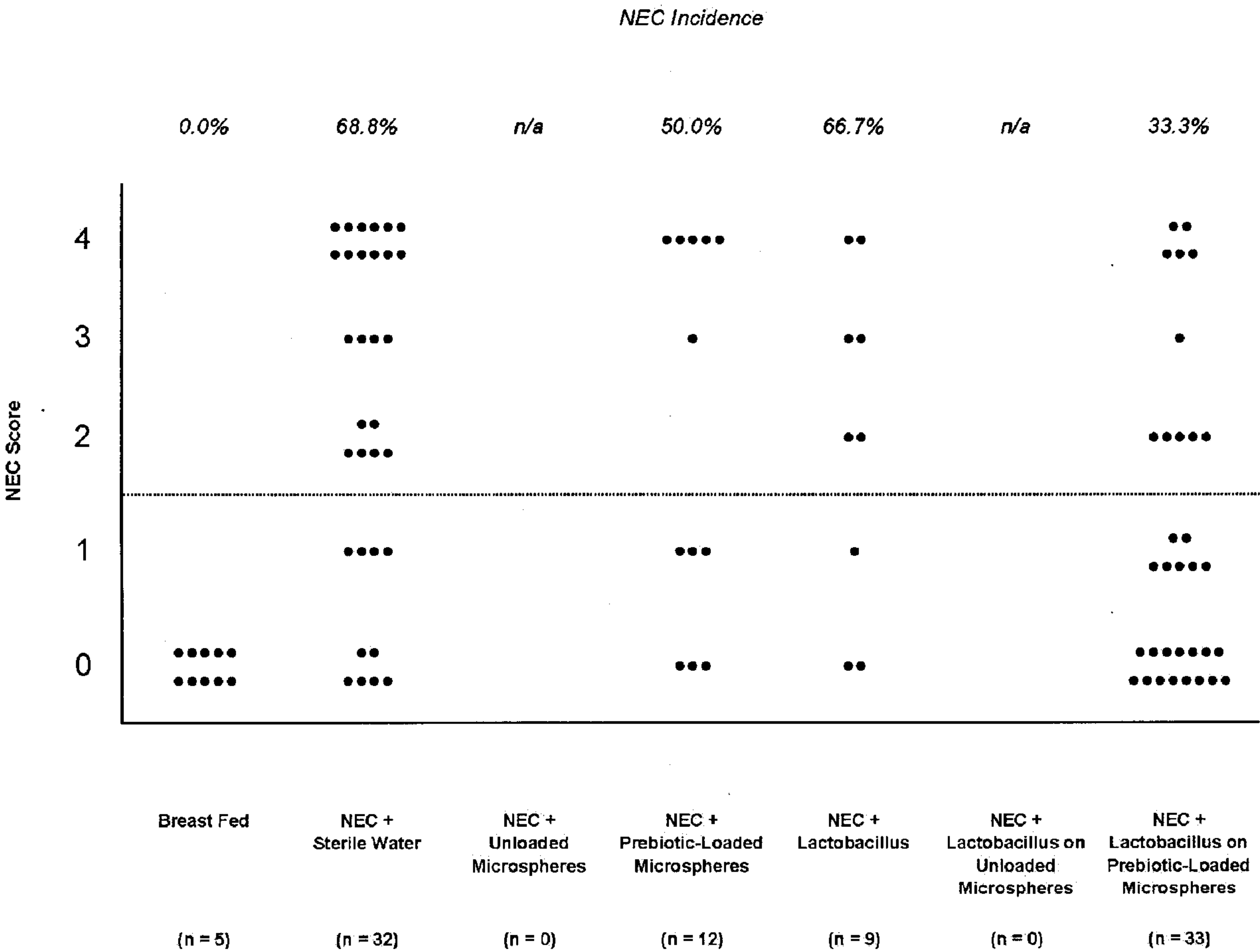


FIG. 5