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(54) Titre : EXPRESSION DE GENES CHEZ DES PLANTES TRANSGENIQUES
(54) Title: EXPRESSION OF GENES IN TRANSGENIC PLANTS

(57) **Abrégé/Abstract:**

A-gene promoter sequence has been isolated and characterised from the plant gene encoding the enzyme cinnamyl alcohol dehydrogenase (CAD), an enzyme which participates in the biosynthesis of lignins. The promoter may be used to control expression of exogenous genes placed under its control. Genes controlled by the CAD promoter are inducible by wounding.





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<p>(21) International Application Number: PCT/GB93/01098 (22) International Filing Date: 27 May 1993 (27.05.93) (30) Priority data: 9211416.4 29 May 1992 (29.05.92) GB (71) Applicant (for all designated States except US): ZENECA LIMITED [GB/GB]; Imperial Chemical House, 9 Millbank, London SW1P 3JF (GB). (72) Inventor; and (75) Inventor/Applicant (for US only) : WALTER, Michael, Herbert [DE/DE]; Talstrasse 103, D-7024 Filderstadt 1 (DE). (74) Agent: HUSKISSON, Frank, Mackie; Zeneca Limited, Group Patent Services Department, P.O. Box 6, Bessemer Road, Welwyn Garden City, Herts AL7 1HD (GB).</p>		<p>(81) Designated States: AU, BB, BG, BR, CA, CZ, FI, HU, JP, KP, KR, LK, MG, MN, MW, NO, NZ, PL, RO, RU, SD, SK, UA, US, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG).</p> <p>Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i></p>

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(57) Abstract

A-gene promoter sequence has been isolated and characterised from the plant gene encoding the enzyme cinnamyl alcohol dehydrogenase (CAD), an enzyme which participates in the biosynthesis of lignins. The promoter may be used to control expression of exogenous genes placed under its control. Genes controlled by the CAD promoter are inducible by wounding.

EXPRESSION OF GENES IN TRANSGENIC PLANTS

The present invention relates to the regulation of gene expression in transgenic plants. In particular it is concerned with the isolation and use of DNA sequences which control the expression of genes in lignifying tissues and in response to pathogen attack.

The ability to isolate and manipulate plant genes has opened the way to gain understanding about the mechanisms involved in the regulation of plant gene expression. This knowledge is important for the exploitation of genetic engineering techniques to practical problems such as the expression of genes in genetically manipulated crop plants exhibiting improved quality and production characteristics.

Many examples have been reported in the literature of plant DNA sequences which have been used to drive the expression of foreign genes in plants. In most instances the regions immediately 5' to the coding regions of genes have been used in gene constructs. These regions are referred to as promoter sequences. They may be derived from plant DNA; or from other sources, eg, viruses. It has been demonstrated that sequences up to 500-1000 bases in most instances are sufficient to allow for the regulated expression of foreign genes. The

types of regulation exemplified include tissue-specificity, regulation by external factors such as light, heat treatment, chemicals, hormones, and developmental regulation. However, it has also
5 been shown that sequences much longer than 1 kb may have useful features which permit high levels of gene expression in transgenic plants.

These experiments have largely been carried out using gene fusions between the promoter
10 sequences and foreign structural genes such as bacterial genes, etc. This has led to the identification of useful promoter sequences.

In the work leading to the present invention we have identified a gene which expresses an enzyme
15 involved in biosynthesis of lignin in vascular plants. We have shown that this gene encodes cinnamyl alcohol dehydrogenase (CAD) which is one of the enzymes specific to the lignin branch pathway of phenylpropanoid metabolism. The gene in
20 question is encoded by the cDNA clone pTCAD19 among others which are the subject of our published copending International Patent Application Number WO 93/05159.

The enzyme encoded by the CAD gene catalyses
25 the oxidation of cinnamyl aldehydes to cinnamyl alcohols which are the direct precursors of the lignin polymer. We have isolated the gene encoding this enzyme from tobacco.

We have shown that CAD mRNA is expressed
30 throughout plant development with the highest levels found in lignifying tissues. In these tissues the CAD enzyme can be found in high concentration in the xylem of plants.

An object of the present invention is to

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provide a promoter sequence suitable for the control of foreign gene expression in plants.

5 According to the present invention there is provided a DNA construct for use in transforming plant cells which comprises an exogenous coding sequence under the control of upstream promoter and downstream terminator sequences, characterised in that the upstream promoter is or has functional homology to a promoter of a gene of the lignin biosynthesis pathway.

10 Thus, the present invention comprises the use of the promoter(s) of the cinnamyl alcohol dehydrogenase or other genes involved in the lignin biosynthesis pathway to control the expression of novel and exogenous proteins and genes in these tissues.

Preferably, the promoter is the CAD promoter.

20 The clone gNtCAD9-6.3SH contains the promoter fragment of this gene. This clone has been deposited at the National Collections of Industrial and Marine Bacteria (NCIB), at 23 St. Machar Drive, Aberdeen AB2 1RY, Scotland, on 2nd April 1992 under the reference NCIB Number 40499.

25 We further provide novel plant cells, and plants transformed with constructs according to the present invention. We illustrate the invention hereinafter using tobacco as a model plant species.

30 The constructs can be used in the expression of exogenous genes and proteins in vascular tissues, particularly but not exclusively, such as poplar, eucalyptus, pine, and other woody plants as well as forages such as festuca, alfalfa, maize, sorghum, penesitum, amongst others.

Not only will this promoter be expressed in

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modified plants in a given tissue but it will also be induced in response to environmental and endogenous signals such as wounding, infection, ethylene production and others.

5 Promoters for use in the invention may be derived from genes such as methyl transferase, and cinnamyl CoA reductase. Such promoters may be isolated from genomic libraries by the use of cDNA probes, as has been done in the case of CAD. We
10 particularly prefer to use the promoter of the CAD gene.

The downstream (3') terminator sequences may also be derived from the CAD gene or they may be derived from other genes. Many possibilities are
15 available from the literature: the selection of the terminator being of rather lesser importance.

By the term 'exogenous coding sequence' we mean a sequence of DNA, other than that which follows the promoter region in the natural CAD
20 gene, that is adapted to be transcribed into functional RNA under the action of plant cell enzymes such as RNA polymerase. Functional RNA is RNA which affects the biochemistry of the cell: it may for example be mRNA which is translated into
25 protein by ribosomes; or antisense RNA which inhibits the translation of mRNA complementary (or otherwise related) to it into protein. In principle any exogenous coding sequence may be used in the present invention.

30 Where the exogenous coding sequence codes for mRNA for a protein, this protein may be of bacterial origin (such as enzymes involved in cell wall modification and cell wall metabolism), of eukaryotic origin (such as pharmaceutically active

5 polypeptides) or of plant origin (such as enzymes involved in the synthesis of phenolic compounds, ethylene synthesis, sugar metabolism, cell wall metabolism,), or genes or parts thereof in sense and antisense orientation.

10 A wide variety of exogenous coding sequences is known from the literature, and the present invention is applicable to these as well as many others yet to be reported. As well as functional mRNA, the exogenous gene may code for RNA that interferes with the function of any kind of mRNA produced by the plant cell: for example, antisense RNA complementary to mRNA for genes such as stilbene synthesis, phytoalexin synthesis and
15 flavour and pigment synthesis.

Of particular interest is the ability of the CAD gene promoter to respond to exogenous stimuli.

20 The construction of vectors and constructs of the present invention will be described in more detail in the Examples below. For convenience it will be generally found suitable to use promoter sequences (upstream - i.e. 5' - of the coding sequence of the gene) of between 100 and 2000 bases in length.

25 A particularly preferred embodiment of the invention is a promoter for use in the expression of exogenous genes in plants, comprising the promoter of cinnamyl alcohol dehydrogenase.

30 More particularly the promoter of the invention comprises the nucleotide sequence shown in Figure 2. The invention includes modifications or the use of only parts of the said sequence which, while retaining sufficient homology to the said sequence in order to maintain functionality,

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enhance or alter its tissue-specificity or response to external stimuli.

5 The invention further provides a recombinant gene construct comprising, in sequence, a promoter according to this invention, a coding region and a gene terminator.

The invention further comprises a recombinant plant genome containing the said construct.

10 Plant cells may be transformed with constructs of the invention according to a variety of known methods (Agrobacterium Ti plasmids, electro-
poration, microinjection, microprojectile bombardment, etc). The transformed cells may then be regenerated into whole plants in which the new
15 nuclear material is stably incorporated into the genome. Both transformed monocot and dicot plants may be obtained in this way. The transformation and regeneration methods employed are not particularly germane to this invention and may
20 simply be selected from those available from the literature.

Examples of genetically modified plants according to the present invention include tomatoes, fruits such as mangoes, peaches, apples,
25 pears, strawberries, bananas and melons; and field crops such as maize (corn), sunflowers, sugarbeet, canola, and small grained cereals such as wheat, barley and rice, ornamental plants such carnations, petunias, roses, chrysanthemums etc.

30 Plants produced by the process of the invention may contain more than one recombinant construct. As well as one or more constructs containing the cinnamyl alcohol dehydrogenase promoter, they may contain a wide variety of other

recombinant constructs, for example constructs having different effects on plant development, structure and defense. In particular, constructs which affect lignin structure, composition and quality and quantity, cellulose and hemicellulose structure and amount, and plant defense genes are included in this invention.

Of particular interest is the ability of the CAD gene promoter to respond to exogenous stimuli. Thus, a further aspect of the present invention is a process of activating exogenous coding sequences in plants under the control of the CAD promoter which comprises the application of exogenous stimuli such as viruses, fungi and bacteria, as well as ethylene and other chemicals. This may find particular use when the expression of novel characters in the plant may need to be controlled exogenously.

We now describe the isolation of genomic clones from a tobacco genomic library encoding the cinnamyl alcohol dehydrogenase gene and related sequences. Genomic clones representing one of the closely related CAD genes found in tobacco have been isolated and characterised by DNA sequence analysis. The clone gTCAD9 represents the whole of the gene with exon sequence identical to the clone pTCAD14 with only two mismatches. The genomic clones described in the Examples cover all of the coding region and the complete transcriptional initiation region of the CAD gene. The subclone gNTCAD9 -6.4SH contains approximately 2800 bp of the promoter fragment of this gene.

The invention will be further described with reference to the following drawings, in which:

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Figure 1 shows a restriction map of gNtCAD9 and a diagram of the structure of the CAD gene;

Figure 2 shows the nucleotide sequence of the CAD promoter and the CAD structural gene, most of which is contained in the 6.4 kb SalI -HindIII fragment of gNtCAD9 (SEQ ID: 1).

Figure 3 outlines a scheme for construction of the plant transformation vector pNtCAD9Prom-GUS1.

EXAMPLE 1

10 1.1 Isolation of CAD genes

A tobacco genomic library (N.tabacum cv NK 326) was purchased from Clontech. This library contains MboI partially digested genomic DNA cloned into lambda EMBL3. The library was screened with the pTCAD14 cDNA insert and positive phages were purified by four successive cycles of plaque purification. Four positive clones were isolated.

Restriction fragment mapping and DNA sequence analysis of these clones indicated that all 4 clones are overlapping clones of the CAD gene and are related. Only clones 2 and 9 contain promoter sequences and only clone 9 was characterised in detail. A restriction map of this clone is shown in Figure 1.

25 1.2 Characterisation of the CAD gene promoter sequence

A 6.4 kb SalI - HindIII fragment was isolated from gNtCAD9 and the nucleotide sequence covering 2765 bp of the 5'-flanking (promoter) sequences and most of the coding sequences has been determined (Figure 2). Primer extension experiments place the transcription start point at nucleotide position 2766.

Full sequence data of about 6.8kb have been

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obtained of a tobacco CAD gene contained on clone gNtCAD9. The following are thereby confirmed:

5 (A) Transcription start site mapped to position 2766 from the upstream Sal I border, now designated position + 1, placing the ATG start codon at + 110

(B) Position and exact size of the four introns:
10 (I) at + 198, size 149 bp; (II) at + 312, size 1290 bp; (III) at + 540, size 563 bp; (IV) at 980, size 632bp. The intron position and sizes differ from the ones in ADH genes.

(C) Three nucleotide mismatches to cDNA cNtCAD14, which are all silent with respect to the deduced protein. They can be ascribed to cultivar differences between cDNA and genomic library.

15 1.3 Fusion of CAD promoter to the GUS reporter gene

A XbaI site at position +84 was used to isolate the CAD promoter from gNtCAD9 and create a transcriptional fusion of 2849 bp of promoter and leader sequences to the GUS reporter gene in pBI 20 101.4. The promoter fragment was isolated after partial XbaI digest due to a second XbaI site in the promoter. Correct fusion and fusion borders were confirmed by restriction analysis and DNA 25 sequencing.

1.4 Construction of plant transformation vector - pCAD-gus

The plasmid subclone gNtCAD9-6.4SH containing the 6.4 kb SalI-HindIII fragment was digested with SalI and partially digested with XbaI to recover a 30 2853 bp SalI-XbaI fragment, which contains all of the available CAD promoter sequence and 87 bp of leader sequence. This promoter fragment was inserted into the appropriately cut polylinker of

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the promoterless GAS cassette of the binary plant transformation vector pBI101.4 in the 5'-3' orientation (Figure 3). The correct insertion has been confirmed by DNA sequencing of the fusion borders.

EXAMPLE 2

Generation of transformed plants

The vector pNtCAD9Prom-GUS1 (from Example 1.4) is transferred to Agrobacterium tumefaciens LBA4404 (a micro-organism widely available to plant biotechnologists) and was used to transform tobacco plants. The fusion construct was introduced into Agrobacterium tumefaciens LBA 4404. Tobacco cv. SR1 leaf discs were transformed via cocultivation with Agrobacteria harbouring the construct. Transgenic shoots were rooted and further propagated by standard procedures. Transformation of tobacco leaf disks follows established methods. Transformed plants were identified by their ability to grow on media containing the antibiotic kanamycin. Plants were regenerated and grown to maturity.

The tissue-specific and developmental expression and expression in response to external stimuli of the β -glucuronidase (GUS) gene as determined by the CAD gene promoter is demonstrated by analysis of stems, roots, leaves, seeds, flowers, pollen and in response to wounding and ethylene treatment for GUS enzyme activity.

EXAMPLE 3

Expression of CAD promoter - GUS fusion in transgenic tobacco

From 20 transformants regenerated 11 have been analysed in some detail. Quantitatively

(fluorometric analysis), GUS activity is highest in roots followed by stems and leaves. Histochemical staining by X-gluc indicates strong expression in the vascular system of stems and leaves preferentially in the developing xylem, where lignification is known to occur. No activity was detectable in pith or epidermal tissue. However, leaf and stem hairs ((trichomes)) exhibit considerable GUS expression in certain stages of their development. One example is the region around axillary buds, where strong staining of trichomes was observed. Upon wounding of stems a local induction of promoter activity was seen during the development of a wound plug but only 10-20 days after wounding.

Claims:

1. A DNA construct for use in transforming plant cells which comprises an
5 exogenous coding sequence under the control of upstream promoter and downstream
terminator sequences, characterised in that the upstream promoter is a promoter of a
cinnamyl alcohol dehydrogenase gene, which promoter has a nucleotide sequence
comprising the first 2765 nucleotides of the sequence shown in Figure No. 2.
- 10 2. An isolated cinnamyl dehydrogenase gene promoter, having the nucleotide
sequence comprising the first 2765 nucleotides of the sequence shown in Figure No.
3. A plant cell transformed with a DNA construct as claimed in claim 1.

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Claims:

1. **A DNA construct for use in transforming plant cells which comprises an
5 exogenous coding sequence under the control of upstream promoter and downstream
terminator sequences, characterised in that the upstream promoter is a promoter of a
cinnamyl alcohol dehydrogenase gene, which promoter has a nucleotide sequence
comprising the first 2765 nucleotides of the sequence shown in Figure No. 2.**
- 10 **2. An isolated cinnamyl dehydrogenase gene promoter, having the nucleotide
sequence comprising the first 2765 nucleotides of the sequence shown in Figure No.**
- 15 **3. A plant cell transformed with a DNA construct as claimed in claim 1.**

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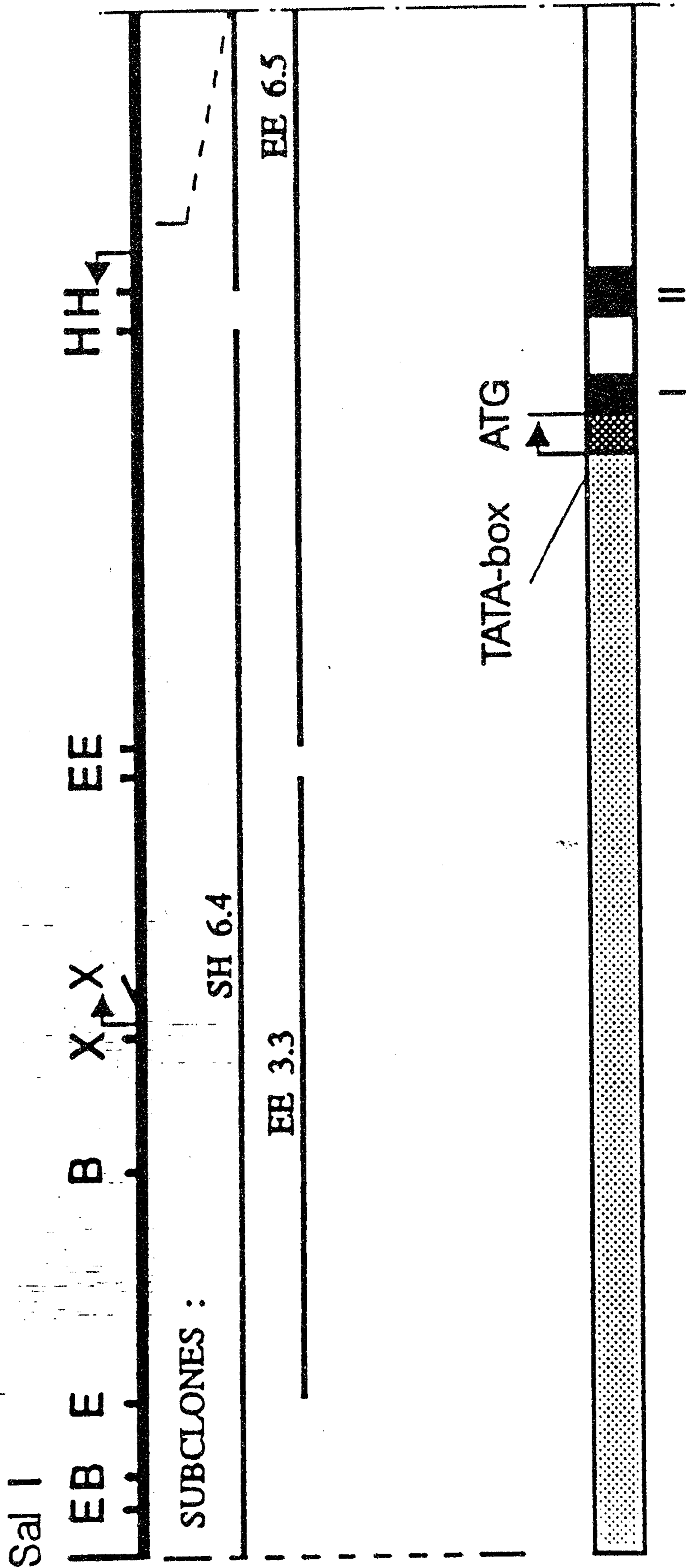
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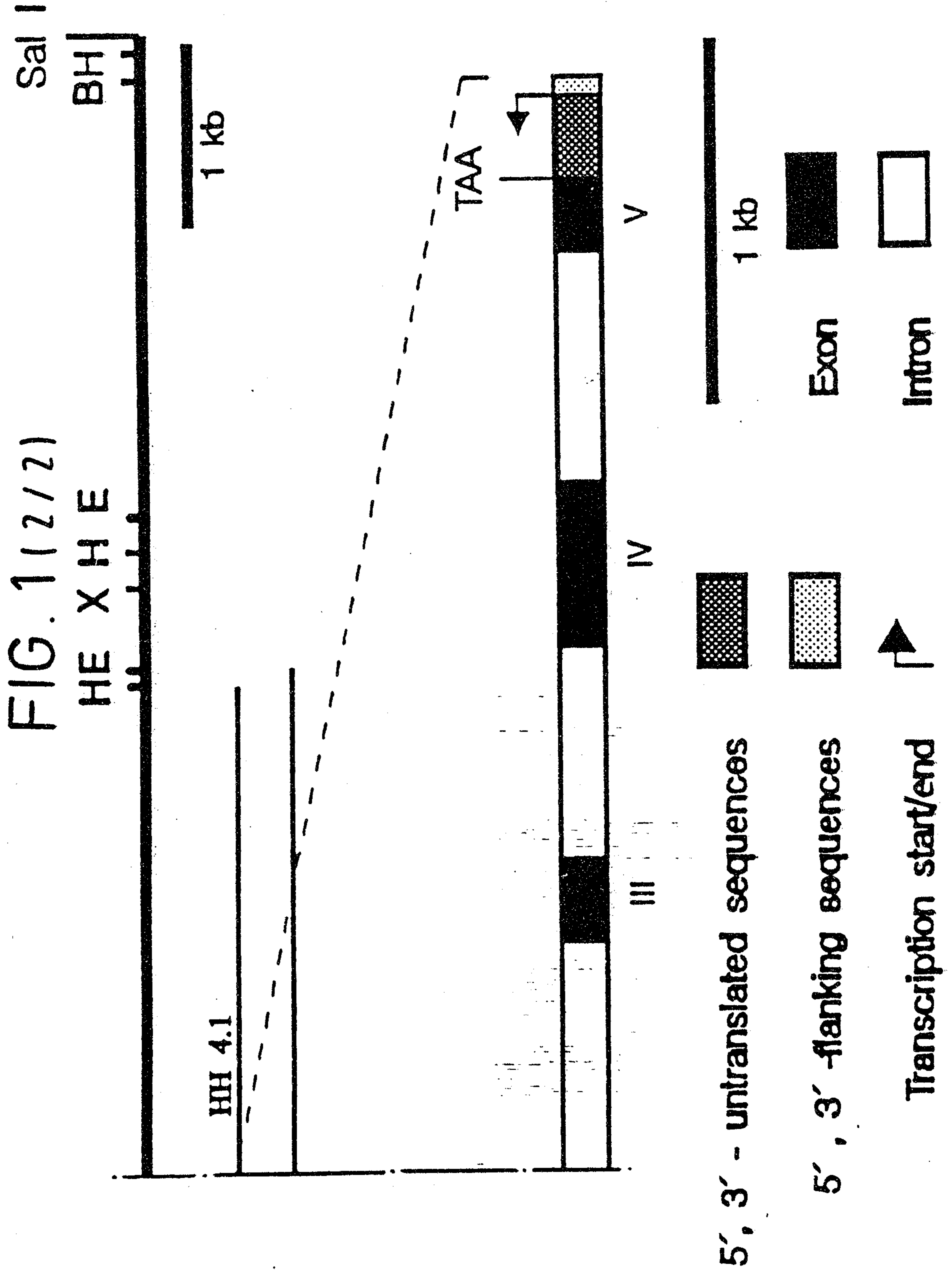
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FIG. 1(1/2)

Tobacco Genomic CAD Clone gNtCAD9 (14 kb Insert)



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FIG. 2 (1/5)

SEQ ID NO: 1
 SEQUENCE TYPE: Nucleotide
 SEQUENCE LENGTH: 6877
 STRANDEDNESS: single
 TOPOLOGY: linear
 MOLECULAR TYPE: genomic

ORIGINAL SOURCE ORGANISM: tobacco cv NK326
 IMMEDIATE EXPERIMENTAL: tobacco genomic DNA library

FEATURES: from 1 - 2765 promoter
 from 2875 - 6579 coding sequence and introns

PROPERTIES: genomic DNA of cinnamyl alcohol dehydrogenase - gNtCAD9

1 GTCGACCTGC AGGTCAACGG ATCATATATT ATGTGTCTTT ACCTATACTA TTATCACTAA
 61 ATCAATAAAT GCTAACTGC AATGATCGTG TTAAATTGTG TATCAGAGAA TTGAGTGACT
 121 GATATATAGT AGGCATTGA AAGTTGAAAG ATGAATTTC AACTAAACTC AGATAAGATG
 181 ATTTTGTGGG GTAGGCATTT GATTAAATGC AAAACCTACT CTCATGAATG TCAGTTTAAAC
 241 AACACAATCA ACCAAATGAA TTCTAAAAC AAGAATTTA CTGGGGAAA AAACGTTTCG
 301 AATAAATAAG AATATCGTAA AATTAATATT AACCAACAAA TTGTCATTGG TTATAACTTA
 361 TAATCACCGT AATGATTTG TACCAATACC TTATTTGGAT CCAAGCCTG GCTAATTGT
 421 TTCTTACCTT CTTTTCACA AGCAACCTCT TTTTAGCTTC TTCTACCTAT ATTACCTACT

FIG. 2
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481	AACITTTTAAG	AGTCCAAAAC	CTAAAATAAA	TCAAAAGTTT	TTTTTAAAAA	AAAAAAA
541	AAAAAATAAA	CCTTACCCTAA	AACATGACGA	TCTAGTGACG	CATACTTCCA	CCTAACCCCG
601	GGGACCCCAA	ACTGCTTGTC	CAGAACAATT	AATGCTTTCT	TTTAGGAAGT	TATCATTCAC
661	ATAATTTCATC	TAGGCCCTGTG	ATAAAGATAA	TTGGTCAATTA	TTGGAAAAGT	CGTCTCACGT
721	AGGAAGAGAT	TGCTAAACTA	GAATTTGAAA	TTTGAGTCCT	CTGAAATTGA	CGTCGAATTC
781	ACTGCATATT	TTAATTACTG	AGTTCATAAT	TATATATTAG	TATAAATTTA	ATTGATTTCTC
841	TCGTATAAAT	ATAGACTTTA	TGCAAAAGCT	ACGAGTFCGA	TCAAACCCCGT	AAATAAATATG
901	TTACCTCCGT	TTCCTAGTAG	GAAGACAGAA	ACAAATATCA	AAAAGAAAAGG	AAAAAAAACAT
961	TTCACACTCT	ACGTACCCTTT	CTATAATTTC	AAAATAAAG	TTGGTGAAAAG	GGAAAAAATGG
1021	TTAGAGGAAA	AATAAGATAG	GAGAGAACAG	AAACTATGTA	TTCCGTAAGGA	ATTTAAAAATT
1081	AAAATATATT	AATTTTGT	TACAAGTTAT	TTAAGAACTT	TAGAAGACGA	ATAACAACAA
1141	TGTTTTTTTT	CTGTTTTTAGT	AATTTTCATA	ACATATGAAC	TAAATGCTCT	TTTTAAAAAGT
1201	TTTTAGGGAA	AAGAAAACG	AAAAATTAAA	GTGGTTGTAT	CAAGAAAAGCA	AAACATATGA
1261	TCAGATTTTC	CGGCATATTT	ATAGGATTTAT	TAACGATGGT	TTACTCAGTT	ACAAGGTTCC
1321	CGAGAAATTA	ATTTTGAAA	TATTAACAGA	AAAAGCCCCA	AATAAACACT	TTTTTCTTTA
1381	AGATTTATTT	ATTTAGGCCT	CGTTAGTTTG	CATGGAGGTT	TGAATCTTAA	TCATTCAGAT
1441	CTCAGATATT	GACTGCCGTTT	CTTTTTAAGT	ATAAATCTTA	ATTATTTCAGA	TCTTAATCAT
1501	TAAATTTT	TAAACCACTG	AATGAGTCTG	AAATCTGAAT	AGGTCCTTAT	GATTACGATT
1561	TATAACAGAG	ACTTAATTTT	ATTAAGATGT	TATCATCCAT	ATTCACTACT	AACCACCGCC
1621	ATTATCACCT	ACCACCATGC	ACCATCCAAC	CAATACCAA	CTCAACCCACC	ACCAAATTC
1681	AACACCATCA	CTAACCACCA	CTATCTCACT	GCTATCATA	TCGACTACCA	TCACCCTCCAA
1741	CCCCACCAT	ATCGATTACC	ATCACCCACC	ACCCCATAA	TATCAACTAA	TACCACCTACC
1801	ATCAATCACA	AACATCTATC	CACCTCACCTC	AACTACTACC	ATCAACCCCC	CCACCACCAT
1861	AATCAACCAT	AGCTACCATT	GTCCTCCACG	CCACCATCAC	CTTCCCTCGAC	CATAACTACT
1921	ACTACCCCAAT	CACCACCTAGT	TATTACTTGG	AACCATCAC	AACCCGCCGC	TCCAGTCCGA
1981	TCCATAAGCC	TCACCCGTCAA	CCATCACTAC	ACCAACTATC	ATATTATTTT	TCAAAAATAT
2041	ATTTATTTAT	AATAGAATAT	TAAATTAAT	AATATTTTAT	TTGAAATTTT	TATTTATTTAT
2101	TTTTTAGATAA	TGATAAATTT	TATACATTC	GATGTTGAAA	ATCAAACAGT	CTTAATCAT
2161	TAGTATTTCA	ATATTAATAT	ATATCTTAAA	TAATCAGATG	TCTATTCTGA	TTCAAAACGTT
2221	TTAATCTTAA	TACACATCTT	AATATTTACA	TGTCCTTTA	TCTTTAAATA	AACAAAATGAG
2281	GCCTTAGCTG	TCTTCACATA	ATTCCTTTAT	GCTGTGCTAT	TAATGAGTAC	GAATTAGTAA
2341	TTCTCTTAAC	GAACAGTGAG	TAGTTTGCTA	ACTTTATATT	CGTTTATTGT	ACGTATATTG

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FIG. 2 (3/5)

2401	TGTAATTATA	GAATAAAAA	ATATTTAGCA	CTTATTTGGT	TAGTATTAAT	TTCCACGGAC
2461	ATTTTAAAG	CATATTTTCC	AAGCTGTATC	TATTTTFTT	TCCTTCCTCT	CATCTCAAGA
2521	TGCAAGTGC	ATTTAATTG	TACAACATGT	CGATTAATAT	AGTGATTACA	ATTATTACCT
2581	TACTGTTTAC	GTATAGGTTG	GAAATAAAT	ATAAGCATCT	ACTCAATCCA	ATTATAAAAC
2641	TTGATGTCTA	TATCTTAGCT	ACTGAGCAA	AAATCAACTT	TTGTAATTC	TAGAAGAAAC
2701	GCCACCCAAC	CTCTGTCTC	CCCTTCTTA	AACTATAAT	ATGTGGTGT	ATTAACCAAA
2761	CTGTAATAA	ATAGCTCACC	ACTATTCCTT	TCTCTTTCCC	TTGAACTGTG	TTTTCGTTT
2821	TTCTGCTCTA	AAACAATAGT	GTGTTCCCTC	TAGATTTTAA	GTTTAAAGAA	CATCATGGGT
2881	GGCTTGGAG	TTGAGAAAC	AACTATTGGT	TGGGCTGCTA	GAGACCCTTC	TGGTGTACTT
2941	TCACCCTTATA	CCTATACTCT	CAGGTACATT	ACATTACTAC	TTATGTCTT	TCTATAATCG
3001	TAATATCCGA	ATCAGCTTAT	GCTTACCCTG	ACTACTTATT	TTTATCTTAT	AATAACTATT
3061	CTACCAATTT	TAAGATGAGT	TATGAACTTT	GTFTTTTGT	TTTTCATGTCC	AGAAACACAG
3121	GACCTGAAGA	TGTGGAAGTC	AAAGTTTGT	ATFTGGGCT	CTGCCACACT	GATCTTCACC
3181	AAGTTAAAA	TGATCTTGGC	ATGTCCAAC	ACCCTCTGGT	TCCTGGGTAT	TTGCTCTTCC
3241	CTCTCTATTT	TCCCCTGTTT	TTCCTCTCT	CTATATATAT	ATATAAACA	TAATTTGAGA
3301	GTTCAAGTTT	TGTGGACAAA	ATGAGGATCA	CTATFGATCC	CCTAAGGATG	GAGGAAAATT
3361	GATTAATTAG	AAGAGAAACA	AGAGTTTGAC	AGTAAFTTTT	TCAAGGATAA	TTTAACTTGT
3421	TTTAGCATAG	GTGTTGCAG	TATTCCTTGAG	ATTACGAATT	TCATTTATGA	TAAATACTTG
3481	TAAGTATATT	TTAAGTGATC	TGATAGCTAG	AAATTCCTTA	CACGCTTTTA	AAATATATGAA
3541	TGTTAAAAAT	CTCCTACAAT	GGTACCCTTT	GAATCTTTGT	TTGATGTCAA	TTTCAGTTT
3601	TTCCCTTAGCC	ACTGAATAAA	TCAAGAGAAA	TGTAGTGGTC	CTTGTGCCCTT	TTAGTACCAA
3661	TCCACCATCC	TCCGGTGACTT	AATAAATTTG	CAACTGTTAG	TTGTTAGGAT	TTGATTTATT
3721	ATTCCAATG	TCTTTTCTTC	CTTCCCTTTAT	TAAACGAATG	ATGAGTACTA	TTGAATTGTT
3781	TAATTAGGAG	TTGTGGCAGA	CCACGTATTT	TCGTGGCCCTA	AATGCCAAAG	CAACATATA
3841	ATAACGTACC	ACTAAGTTCA	CTTCTTTTCAT	TGGATTTGTA	AAAGTTGTTG	GTGACACTTA

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FIG. 2 (4/5)

3901	CGTTTTGTTTC	GAATCCAATG	CTATTATTGT	TAGATAAATT	TTTACTTTTA	GTCCCTAAAA
3961	TTATTGATAA	GTTATAAATT	TAATCCTTGT	ACTATTTGGC	GAAGTACATT	TAGTCTTCAA
4021	TTTAATTAAA	ATTAGACATT	TGATCCTGCT	TCTGCTTGTG	AATCTTCACA	AAGTTAATCA
4081	ATTATGAATT	CATGAATTAT	GTTAAATTTG	TGTTGTTACT	CCCTATCTAA	GGATACTAGA
4141	GTTCTAATTC	ATAAGTAGCG	GTGGTAAAT	AGATAAATA	AATAATTATC	CATTCATTTT
4201	ATCCAATAA	AAATAAATA	CCAATGAATT	TAACTTTTAT	ATTTACAAC	CCTTAATTGA
4261	AGGGTTTCTC	AAGTTTGAGA	TACTAAGAAT	TCTCTCAAAA	GTAATCATA	TGAAGAAGTT
4321	ATGAATCATA	TGTATATCCA	TATTATCTGT	CAATTAACATA	TTTTTTTATCC	ATATTAAATA
4381	TAGATAAGGT	TGAATATTAT	ATTTATTAGA	CGCCCATTTG	TCACCCCTAT	AATCACAATT
4441	GCAATATCCC	TTTTGTTTTT	TACTATGTGT	TGATCCTTAT	CCCTTTGTAA	CTTCATATTT
4501	GTGGTGAATG	ATGTAGACAT	GAAGTGGTGG	GAGAAGTGGT	GGAGGTAGGA	CCAGATGTGT
4561	CAAAATTCAA	AGTGGGGGAC	ACAGTTGGAG	TTGGATTACT	CGTTGGAAGT	TGCAGGAACT
4621	GTGGCCCTTG	CAAGAGAGAT	ATAGAGCAAT	ATTGCAACAA	GAAAGATTGG	AACTGCAATG
4681	ATGTCTACAC	TGATGGCAA	CCCACCCCAAG	GTGGTTTTCG	TAAATCCATG	GTTGTTGATC
4741	AAAAGTAAGT	CTTTTACCCT	TCCATATATA	TAGAGAAGTA	CCTTCTTTA	ATATTAACTT
4801	ATAGCAAGTG	ACATTGTATA	TAAGTTACAC	CCTGTAAATT	AAAGAAACTA	TCAGTATAAC
4861	TTACCTGTCC	TAATAGGTTG	GTTGCCTTAT	TTTGAGGTAA	TTAGTTTAC	TTATTGTGGA
4921	CAGAGCTACA	ACTATTTAGA	TAATAGGATG	GTATAAACA	ATTTCTACAC	TATCATTTGTG
4981	TTTGTTC AAC	TCTTTTTPAA	ATCCATTGGG	TATCGAATCA	TGACAAATGT	TACTAGGACG
5041	ATCAAGAAAT	CATGTAGGC	AGCCCTGTAC	ACTAAGCTCG	CGCTATGCCG	TCCGGGGAAA
5101	GTCGGACCAC	AAAGGTATAT	TGTATGTAGT	CTAACCCCTGC	ATTTCTGCAG	AGGTGCTCGA
5161	ACCCGTATCC	TCTTTGGTCA	CATGGTAATA	ACTTAACCAG	TTACGCCAAG	GTTCCCCCTTC
5221	AAGAAACCAT	ATAATTAAAG	TGGTTAATTC	ACATTTCTTG	AATTAAGA	TCACAATGGA
5281	CTGAATGTTA	TTTTGCCCTTG	TATATAGGTT	TGTGGTGAAA	ATTCAGAGG	GTATGGCACC
5341	AGAACAAGCA	GCACCTCTAT	TATGTGCTGG	TATAACAGTA	TACAGTCCAT	TGAACCATTT
5401	TGGTTTCAA	CAGAGTGGAT	TAAGAGGAGG	AATTTTGGGA	TTAGGAGGAG	TGGGACACAT
5461	GGGAGTGAAA	ATAGCAAAGG	CAATGGGACA	TCATGTTACT	GTCAATTAGTT	CTTCAAATAA

5521 GAAGAGACAA GAGGCATTGG AACATCTTGG TGCAGATGAT TATCTTGTCA GTTCAGACAC
5581 TGATAAAATG CAAGAGGCTT CTGATTCACT TGACTATATT ATTGATACTG TCCCTGTTGG
5641 CCATCCTCTT GAACCTTATC TTTCTTTGCT TAAAATTGAT GGCAAACTTA TCTTGATGGG
5701 AGTTATCAAC ACCCCCTTGC AATTATCTC CCCCATGGTT ATGCTCGGTA AGTCATTAA
5761 CTTATTTACG CTGACAGTGT AACACGTTCC GACGAAATGC ATTATTACAA CAAGTTAAAA
5821 TTGGTTTCTA ACAATATATC GTGTTAGTAC TTGCTCTAAA AAGTCGATTG AAAATTAAAA
5881 CGTTGAATTT GTCTATATAT GAAAAAAGAT AGAACAAAAG GATTGGAAGA GTGGTCATTT
5941 TAGTCAATGA AAGCCATGAT TTGTTGAAAT ATGCAGGTGC CATTGGTCTA TTTTTCGGCC
6001 ATAATAATGT TATGAGCAGA ATTAATATAT AGACAAATTC TAGGCTTGTA GGGTTGTGTT
6061 ATTTAAGGAA CCATGCTGTC TATTGATTA ACATTTGTTT AATGCAACCT CTTATTACTA TTATGTTTAA
6121 AAAGCCAATG TATTGATTA AGCACAGACA GTGTGTAAGT TTTTATTATAC CATCAAGCAA AGTTGATATA
6181 AGATTTTTHA AAAAGTTGAT GTACATAAAT AGATAACATT TCATTAGTGT GGTATGGTAA
6241 TTCCGAAAAG GTACTCGGTA TAATTTTCA TTTTACATTG TCGGCCGATA TAACTTTTAG
6301 ACTTTATTAT TATGATCAGG GAGAAAGAGC ATCACAGGAA GCTTTATTGG TAGCATGAAG
6361 GAAACAGAGG AAATGCTAGA TTTCTGCAAA GAGAGGTTGG AGAAAATGA TGTGAGGATA
6421 GTGAAAATGG ATTATATCAA CACTGCAATG GAGAGGTTGG AGAAAATGA TGTGAGGATA
6481 AGATTTGTGG TTGATGTTAT ATGGTTCACI ATTATACAAG GCTGTGAGAA TACTAAACTT TGATGTCGTC
6601 ACAACATGGA ATGGTTCACI TTTTGTTHA TTTGCCACCT GTATTTTCTT ATTTGGTGAT CGAGAGTGAC
6661 TTTTGTATCC TTTTGTTHA TATTTTCTTT CTTCAAAACA ATTTAATGTA TGAATTTGGA TGTGAGGATA
6721 GTTTATGTAT TATTTTCTTT TATACCAACC AAAACTTTGT TTGGTACGTC AAGCTATCTC CACTTCTCGT
6781 GATTTTGAAA TATACCAACC AAAACTTTGT TTGGTACGTC AAGCTATCTC CACTTCTCGT
6841 GACTAGGGTG TCGATCCATC TTTAAAATCGA TTAACCG 6877

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FIG. 2(5/5)

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Tobacco CAD Promoter Gene Fusion With GUS in pBI 101.4

RB

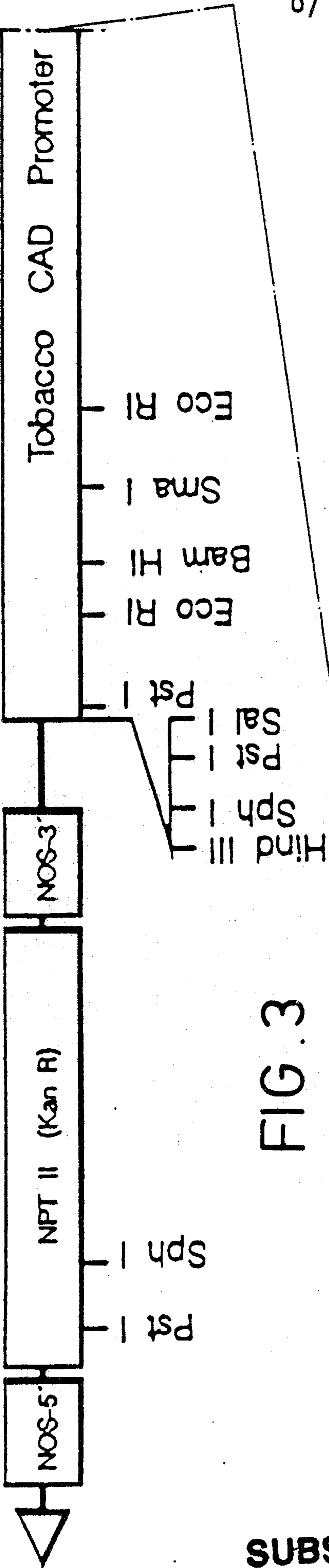


FIG. 3

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