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(71) Applicant: **BOARD OF REGENTS, THE UNIVERSITY OF TEXAS SYSTEM** [US/US]; 210 West 7th Street, Austin, TX 78701 (US).

(72) Inventors: **MCLELLAN, Jason**; 5408 Ketch Ct., Austin, TX 78730 (US). **HSIEH, Ching-lin**; 100 E. 24th St., Austin, TX 78712 (US).

(74) Agent: **SCHNEPP, Amanda, S.J.**; Parker Highlander PLLC, 1120 S. Capital Of Texas Highway, Bldg. One, Suite 200, Austin, TX 78746 (US).

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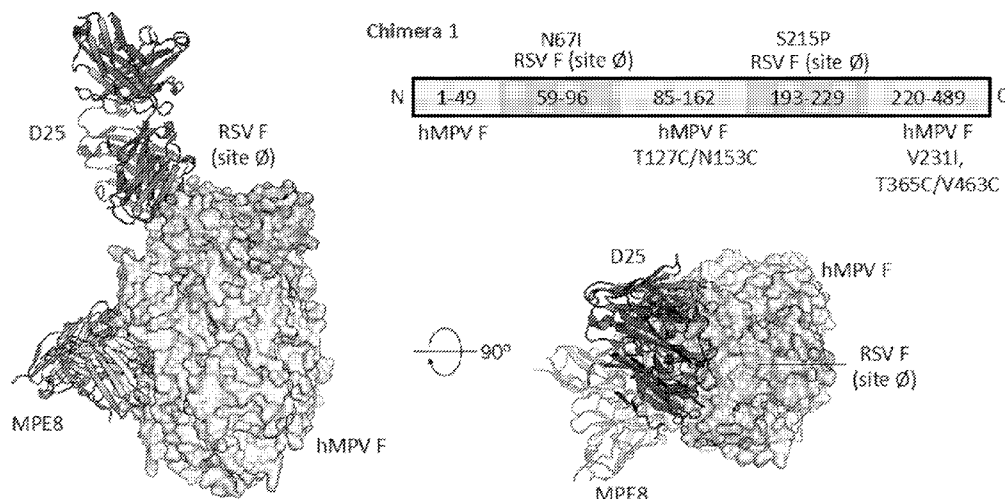


FIG. 1

(57) Abstract: Provided herein are chimeric hMPV/RSV F proteins. In some aspects, the chimeric hMPV/RSV F proteins exhibit enhanced conformational stability, enhanced thermostability, and/or increased expression. Methods are also provided for use of the chimeric F proteins as diagnostics, in screening platforms, and/or in vaccine compositions.



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DESCRIPTION

PREFUSION-STABILIZED CHIMERIC HMPV-RSV F PROTEINS

REFERENCE TO RELATED APPLICATIONS

[0001] The present application claims the priority benefit of United States provisional
5 application number 63/222,140, filed July 15, 2021, the entire contents of which are
incorporated herein by reference.

REFERENCE TO A SEQUENCE LISTING

[0002] The instant application contains a Sequence Listing SML, which has been
submitted electronically and is hereby incorporated by reference in its entirety. Said
10 Sequence Listing XML, created on July 13, 2022, is named UTFBP1284WO_ST26.xml and
is 14,947 bytes in size.

BACKGROUND

1. Field

[0003] The present disclosure relates generally to the fields of medicine, virology,
15 immunology, and protein engineering. More particularly, the disclosure relates to chimeric
human metapneumovirus (hMPV)/respiratory syncytial virus (RSV) F proteins and the use
thereof in vaccine formulation and immune response detection.

2. Description of Related Art

[0004] Pneumoviridae are a family of negative-sense, single-stranded RNA viruses
20 that cause respiratory tract infections. Respiratory syncytial virus (RSV) is an enveloped non-
segmented negative-strand RNA virus in the family Pneumoviridae. It is the most common
cause of bronchiolitis and pneumonia among children in their first year of life. RSV also
causes repeated infections including severe lower respiratory tract disease, which may occur
at any age, especially among the elderly or those with compromised cardiac, pulmonary, or
25 immune systems. Passive immunization currently is used to prevent severe illness caused by
RSV infection, especially in infants with prematurity, bronchopulmonary dysplasia, or
congenital heart disease. Despite the burden of RSV infection in certain populations,
development of an effective RSV vaccine remains elusive.

[0005] Human metapneumovirus (hMPV) is also a negative-sense enveloped virus of the Pneumoviridae family and was discovered in 2001 but has been circulating for at least a half century before its discovery. There is near ubiquitous infection by the age of five and re-infections continue to be a burden throughout life. However, infants (6-12 months), the elderly, and immunocompromised populations are at an increased risk of hospitalization with more severe disease such as pneumonia and bronchiolitis. Despite the disease burden hMPV presents, there are no vaccines or therapeutics that have been approved for prevention or treatment. Similar to RSV, development of an effective hMPV vaccine remains elusive.

SUMMARY

[0006] In one embodiment, provided herein are engineered human metapneumovirus (hMPV)–respiratory syncytial virus (RSV) chimeric F proteins comprising the following segments, in order from N-terminus to C-terminus: (1) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity to amino acids 19-49 of any one of SEQ ID NOs: 5-8; (2) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity to amino acids 59-96 of SEQ ID NO: 10; (3) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity to amino acids 85-162 or 85-164 of any one of SEQ ID NOs: 5-8; (4) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity to amino acids 193-229, 193-231, 195-229, or 195-231 of SEQ ID NO: 10; and (5) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity to amino acids 200-489 or 202-489 of any one of SEQ ID NOs: 5-8.

[0007] In some aspects, the chimeric F proteins comprise a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity to amino acids 19-492 of any one of SEQ ID NOs: 1-4.

[0008] In some aspects, the chimeric F proteins comprise the following segments, in order from N-terminus to C-terminus: (1) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at

least 99%, or 100% identity to amino acids 19-49 of any one of SEQ ID NOs: 5-8; (2) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% identity to amino acids 59-96 of SEQ ID NO: 10; (3) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% identity to amino acids 85-162 of any one of SEQ ID NOs: 5-8; (4) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% identity to amino acids 193-229 of SEQ ID NO: 10; and (5) a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% identity to amino acids 200-489 of any one of SEQ ID NOs: 5-8. In some aspects, the chimeric F proteins comprise a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% identity to amino acids 19-492 of SEQ ID NO: 1.

[0009] In some aspects, the chimeric F proteins comprise a substitution corresponding to Q100R and S101R within segment (3), with the positions being relative to SEQ ID NOs: 5-8. In some aspects, the chimeric F proteins comprise an engineered disulfide bond comprising paired cysteine substitutions at positions corresponding to: T127C and N153C and/or T365C and V463C within segments (3) and (5), respectively, with the positions being relative to SEQ ID NOs: 5-8. In some aspects, the chimeric F proteins comprise a substitution at position V231 within segment (5), with the position being relative to SEQ ID NOs: 5-8. In some aspects, the chimeric F proteins comprise a substitution corresponding to V231I within segment (5), with the position being relative to SEQ ID NOs: 5-8.

[0010] In some aspects, the chimeric F proteins comprise a substitution at position N67 and/or S215 within segments (2) and (4), respectively, with the positions being relative to SEQ ID NO: 10. In some aspects, the chimeric F proteins comprise a substitution corresponding to N67I and/or S215P within segments (2) and (4), respectively, with the positions being relative to SEQ ID NO: 10.

[0011] In some aspects, the chimeric F proteins comprise (i) substitutions corresponding to N67I and S215P within segments (2) and (4), respectively, with the positions being relative to SEQ ID NO: 10; (ii) a substitution corresponding to V231I within segment (5), with the position being relative to SEQ ID NOs: 5-8; and (iii) engineered

disulfide bonds comprising paired cysteine substitutions at positions corresponding to: (a) T127C and N153C and (b) T365C and V463C within segments (3) and (5), respectively, with the positions being relative to SEQ ID NOs: 5-8.

[0012] In some aspects, the chimeric F proteins comprise a substitution at position T91 or L95 within segment (2), with the positions being relative to SEQ ID NO: 10. In some aspects, the chimeric F proteins comprise a substitution corresponding to T91R or L95R within segment (2), with the positions being relative to SEQ ID NO: 10. In some aspects, the chimeric F proteins comprise a substitution corresponding to L95R within segment (2), with the position being relative to SEQ ID NO: 10.

[0013] In some aspects, the chimeric F proteins comprise an engineered disulfide bond comprising paired cysteine substitutions at positions corresponding to: L110C and N322C within segments (3) and (5), respectively, with the positions being relative to SEQ ID NOs: 5-8.

[0014] In some aspects, the chimeric F proteins comprise a substitution at position A107 within segment (3), with the position being relative to SEQ ID NOs: 5-8. In some aspects, the chimeric F proteins comprise a substitution corresponding to A107P within segment (3), with the position being relative to SEQ ID NOs: 5-8.

[0015] In some aspects, the chimeric F proteins comprise a substitution at position L219 within segment (5), with the position being relative to SEQ ID NOs: 5-8. In some aspects, the chimeric F proteins comprise a substitution corresponding to L219K within segment (5), with the position being relative to SEQ ID NOs: 5-8.

[0016] In some aspects, the chimeric F proteins comprise a substitution at position E453 within segment (5), with the position being relative to SEQ ID NOs: 5-8. In some aspects, the chimeric F proteins comprise a substitution corresponding to E453Q within segment (5), with the position being relative to SEQ ID NOs: 5-8.

[0017] In some aspects, the chimeric F proteins comprise engineered disulfide bonds comprising paired cysteine substitutions at positions corresponding to: T130C and N156C, T368C and V466C, and L113C and N325C; and substitutions corresponding to N58I, L86R, A110P, S188P, L222K, V234I, and E456Q, with the positions being relative to SEQ ID NO:

1.

[0018] In some aspects, the chimeric proteins are fused or conjugated to a trimerization domain. In some aspects, the chimeric proteins are fused to a trimerization domain. In some aspects, the trimerization domain comprises a T4 fibrin trimerization domain.

5 [0019] In some aspects, the chimeric proteins are fused or conjugated to a transmembrane domain. In some aspects, the chimeric proteins are fused to a transmembrane domain. In some aspects, the transmembrane domain comprises a metapneumovirus (MPV) F protein transmembrane domain.

[0020] In some aspects, the chimeric proteins comprise an N-terminal signal
10 sequence. In some aspects, the N-terminal signal sequence is MSWKVVIIFSLITPQHG (amino acids 1-18 of SEQ ID NO: 1).

[0021] In some aspects, the chimeric proteins comprise a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% identity to amino acids 19-554 of SEQ ID NO: 1. In
15 some aspects, the chimeric proteins comprise a sequence having at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, at least 99%, or 100% identity to amino acids 1-554 of SEQ ID NO: 1.

[0022] In one embodiment, provided herein are engineered human metapneumovirus (hMPV)–respiratory syncytial virus (RSV) chimeric F protein trimers comprising at least one
20 subunit according to any one of the present engineered, chimeric protein embodiments. In some aspects, the trimer is stabilized in a prefusion conformation relative to a trimer of wildtype metapneumovirus (MPV) F subunits. In some aspects, the trimer comprises at least one engineered disulfide bond between subunits.

[0023] In one embodiment, provided herein are pharmaceutical compositions
25 comprising a pharmaceutically acceptable carrier; and (i) an engineered, chimeric protein or (ii) an engineered trimer of any one of the present embodiments. In some aspects, the pharmaceutical compositions further comprise an adjuvant.

[0024] In one embodiment, provided herein are nucleic acid molecules comprising a nucleotide sequence that encodes an amino acid sequence of an engineered, chimeric protein

of any one of the present embodiments. In some aspects, the nucleic acid comprises a DNA expression vector. In some aspects, the nucleic acid comprises a mRNA.

[0025] In one embodiment, provided herein are methods of preventing metapneumovirus (MPV) or respiratory syncytial virus (RSV) infection or a disease associate
5 with MPV or RSV infection in a subject, comprising administering to the subject an effective amount of the pharmaceutical composition or a nucleic acid molecule of any one of the present embodiments.

[0026] In one embodiment, provided herein are compositions comprising (i) an engineered protein or (ii) an engineered trimer of any one of the present embodiments bound
10 to an antibody.

[0027] Other objects, features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples, while indicating preferred embodiments of the invention, are given by way of illustration only, since various changes and modifications
15 within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

[0028] The following drawings form part of the present specification and are included to further demonstrate certain aspects of the present invention. The invention may be better
20 understood by reference to one or more of these drawings in combination with the detailed description of specific embodiments presented herein.

[0029] **FIG. 1.** Representative design and model for hMPV/RSV F chimera.

[0030] **FIG. 2.** Characterization of hMPV/RSV F chimeras by size exclusion chromatography (left panel) and prefusion F-preferred antibody binding (right panel).

[0031] **FIG. 3.** Additional salt bridge designs further increase the protein expression
25 (left panel) while retaining the prefusion-specific epitopes from hMPV/RSV F (right panel).

[0032] **FIG. 4.** The combinatorial substitutions carrying two different types of designs boost the protein expression (top right panel) and enhance the thermostability (bottom right panel).

[0033] **FIG. 5.** The best chimera construct DS-CavEsP has the highest protein expression and exhibits the prefusion conformation bound by RSV F-specific antibody D25 and hMPV F cross-reactive antibody MPE8. SDS-PAGE analysis of protein expression from prefusion-stabilized chimera variants (left panel). Representative 2D classes of chimera DS-CavEsP bound to D25 and MPE8 (middle panel). A 3D reconstruction of chimera DS-CavEsP bound to D25 and MPE8 (right panel).

[0034] **FIGS. 6A-6D.** Neutralizing antibodies binding to DS-CavEsP. FIG. 6A shows binding of DS-CavEsP to antibody D25 assessed by biolayer interferometry. FIG. 6B shows binding of PRDM to antibody D25 assessed by biolayer interferometry. FIG. 6C shows binding of DS-CavEsP to antibody MPE8 assessed by biolayer interferometry. FIG. 6D shows binding of DS-CavES2 to antibody MPE8 assessed by biolayer interferometry.

[0035] **FIGS. 7A-7C.** Cryo-EM structure of D25 and MPE8 bound to DS-CavEsP. FIG. 7A shows the cryo-EM structure of DS-CavEsP bound to D25 and MPE8 at 3.2Å. FIGS. 7B and 7C show the binding interfaces of MPE8-DS-CavEsP and D25-DS-CavEsP, respectively.

DETAILED DESCRIPTION

[0036] Provided herein are prefusion-stabilized hMPV-RSV chimeric F proteins that can be used as vaccine antigens, for both hMPV and RSV infection, and as reagents for detecting antibodies in sera. The chimeric proteins can be used in diagnostic assays to detect the presence of “prefusion-specific” hMPV F or RSV F protein-directed antibodies. Both the junctions between the hMPV and the RSV portions of the chimeric F proteins, and additional substitutions made in the chimeric F proteins, result in decent expression, stability, and desired prefusion conformation.

I. Definitions

[0037] It is to be understood that both the foregoing general description and the following detailed description are exemplary and explanatory only and are not restrictive of

the invention as claimed. In this application, the use of the singular includes the plural unless specifically stated otherwise. In this application, the use of “or” means “and/or” unless stated otherwise. Furthermore, the use of the term “including”, as well as other forms, such as “includes” and “included”, is not limiting. Also, terms such as “element” or “component” encompass both elements and components comprising one unit and elements and components that comprise more than one subunit unless specifically stated otherwise. Also, the use of the term “portion” can include part of a moiety or the entire moiety.

[0038] As used herein the specification, “a” or “an” may mean one or more. As used herein in the claim(s), when used in conjunction with the word “comprising,” the words “a” or “an” may mean one or more than one.

[0039] The use of the term “or” in the claims is used to mean “and/or” unless explicitly indicated to refer to alternatives only or the alternatives are mutually exclusive, although the disclosure supports a definition that refers to only alternatives and “and/or.” As used herein “another” may mean at least a second or more.

[0040] The term “about” as used herein when referring to a measurable value such as an amount, a temporal duration, and the like, is meant to encompass variations of up to $\pm 10\%$ from the specified value. Unless otherwise indicated, all numbers expressing quantities of ingredients, properties such as molecular weight, reaction conditions, and so forth used in the specification and claims are to be understood as being modified in all instances by the term “about.” Accordingly, unless indicated to the contrary, the numerical parameters set forth in the following specification and attached claims are approximations that may vary depending upon the desired properties sought to be obtained by the disclosed subject matter. At the very least, and not as an attempt to limit the application of the doctrine of equivalents to the scope of the claims, each numerical parameter should at least be construed in light of the number of reported significant digits and by applying ordinary rounding techniques. Notwithstanding that the numerical ranges and parameters setting forth the broad scope of the invention are approximations, the numerical values set forth in the specific examples are reported as precisely as possible. Any numerical value, however, inherently contain certain errors necessarily resulting from the standard deviation found in their respective testing measurements.

[0041] As used herein, “essentially free,” in terms of a specified component, is used herein to mean that none of the specified component has been purposefully formulated into a composition and/or is present only as a contaminant or in trace amounts. The total amount of the specified component resulting from any unintended contamination of a composition is therefore well below 0.05%, preferably below 0.01%. Most preferred is a composition in which no amount of the specified component can be detected with standard analytical methods.

[0042] The term “antibody” refers to an intact immunoglobulin of any isotype, or a fragment thereof that can compete with the intact antibody for specific binding to the target antigen, and includes, for instance, chimeric, humanized, fully human, and bispecific antibodies. An “antibody” is a species of an antigen binding protein. An intact antibody will generally comprise at least two full-length heavy chains and two full-length light chains, but in some instances can include fewer chains such as antibodies naturally occurring in camelids which can comprise only heavy chains. Antibodies can be derived solely from a single source, or can be “chimeric,” that is, different portions of the antibody can be derived from two different antibodies as described further below. The antigen binding proteins, antibodies, or binding fragments can be produced in hybridomas, by recombinant DNA techniques, or by enzymatic or chemical cleavage of intact antibodies. Unless otherwise indicated, the term “antibody” includes, in addition to antibodies comprising two full-length heavy chains and two full-length light chains, derivatives, variants, fragments, and muteins thereof, examples of which are described below. Furthermore, unless explicitly excluded, antibodies include monoclonal antibodies, bispecific antibodies, minibodies, domain antibodies, synthetic antibodies (sometimes referred to herein as “antibody mimetics”), chimeric antibodies, humanized antibodies, human antibodies, antibody fusions (sometimes referred to herein as “antibody conjugates”), and fragments thereof, respectively. In some embodiments, the term also encompasses peptibodies.

[0043] Naturally occurring antibody structural units typically comprise a tetramer. Each such tetramer typically is composed of two identical pairs of polypeptide chains, each pair having one full-length “light” (in certain embodiments, about 25 kDa) and one full-length “heavy” chain (in certain embodiments, about 50-70 kDa). The amino-terminal portion of each chain typically includes a variable region of about 100 to 110 or more amino acids that typically is responsible for antigen recognition. The carboxy-terminal portion of each

chain typically defines a constant region that can be responsible for effector function. Human light chains are typically classified as kappa and lambda light chains. Heavy chains are typically classified as mu, delta, gamma, alpha, or epsilon, and define the antibody's isotype as IgM, IgD, IgG, IgA, and IgE, respectively. IgG has several subclasses, including, but not limited to, IgG1, IgG2, IgG3, and IgG4. IgM has subclasses including, but not limited to, IgM1 and IgM2. IgA is similarly subdivided into subclasses including, but not limited to, IgA1 and IgA2. Within full-length light and heavy chains, typically, the variable and constant regions are joined by a "J" region of about 12 or more amino acids, with the heavy chain also including a "D" region of about 10 more amino acids. See, e.g., *Fundamental Immunology*, Ch. 7 (Paul, W., ed., 2nd ed. Raven Press, N.Y. (1989)) (incorporated by reference in its entirety for all purposes). The variable regions of each light/heavy chain pair typically form the antigen binding site.

[0044] The term "variable region" or "variable domain" refers to a portion of the light and/or heavy chains of an antibody, typically including approximately the amino-terminal 120 to 130 amino acids in the heavy chain and about 100 to 110 amino terminal amino acids in the light chain. In certain embodiments, variable regions of different antibodies differ extensively in amino acid sequence even among antibodies of the same species. The variable region of an antibody typically determines specificity of a particular antibody for its target.

[0045] The variable regions typically exhibit the same general structure of relatively conserved framework regions (FR) joined by three hyper variable regions, also called complementarity determining regions or CDRs. The CDRs from the two chains of each pair typically are aligned by the framework regions, which can enable binding to a specific epitope. From N-terminal to C-terminal, both light and heavy chain variable regions typically comprise the domains FR1, CDR1, FR2, CDR2, FR3, CDR3 and FR4. The assignment of amino acids to each domain is typically in accordance with the definitions of Kabat Sequences of Proteins of Immunological Interest (National Institutes of Health, Bethesda, Md. (1987 and 1991)), Chothia & Lesk, *J. Mol. Biol.*, 196:901-917 (1987) or Chothia et al., *Nature*, 342:878-883 (1989).

[0046] In certain embodiments, an antibody heavy chain binds to an antigen in the absence of an antibody light chain. In certain embodiments, an antibody light chain binds to an antigen in the absence of an antibody heavy chain. In certain embodiments, an antibody binding region binds to an antigen in the absence of an antibody light chain. In certain

embodiments, an antibody binding region binds to an antigen in the absence of an antibody heavy chain. In certain embodiments, an individual variable region specifically binds to an antigen in the absence of other variable regions.

[0047] In certain embodiments, definitive delineation of a CDR and identification of residues comprising the binding site of an antibody is accomplished by solving the structure of the antibody and/or solving the structure of the antibody-ligand complex. In certain embodiments, that can be accomplished by any of a variety of techniques known to those skilled in the art, such as X-ray crystallography. In certain embodiments, various methods of analysis can be employed to identify or approximate the CDR regions. Examples of such methods include, but are not limited to, the Kabat definition, the Chothia definition, the AbM definition and the contact definition.

[0048] The Kabat definition is a standard for numbering the residues in an antibody and is typically used to identify CDR regions. See, e.g., Johnson & Wu, *Nucleic Acids Res.*, 28: 214-8 (2000). The Chothia definition is similar to the Kabat definition, but the Chothia definition takes into account positions of certain structural loop regions. See, e.g., Chothia et al., *J. Mol. Biol.*, 196: 901-17 (1986); Chothia et al., *Nature*, 342: 877-83 (1989). The AbM definition uses an integrated suite of computer programs produced by Oxford Molecular Group that model antibody structure. See, e.g., Martin et al., *Proc Natl Acad Sci (USA)*, 86:9268-9272 (1989); "AbM™, A Computer Program for Modeling Variable Regions of Antibodies," Oxford, UK; Oxford Molecular, Ltd. The AbM definition models the tertiary structure of an antibody from primary sequence using a combination of knowledge databases and ab initio methods, such as those described by Samudrala et al., "Ab Initio Protein Structure Prediction Using a Combined Hierarchical Approach," in *PROTEINS, Structure, Function and Genetics Suppl.*, 3:194-198 (1999). The contact definition is based on an analysis of the available complex crystal structures. See, e.g., MacCallum et al., *J. Mol. Biol.*, 5:732-45 (1996).

[0049] By convention, the CDR regions in the heavy chain are typically referred to as H1, H2, and H3 and are numbered sequentially in the direction from the amino terminus to the carboxy terminus. The CDR regions in the light chain are typically referred to as L1, L2, and L3 and are numbered sequentially in the direction from the amino terminus to the carboxy terminus.

[0050] The term “light chain” includes a full-length light chain and fragments thereof having sufficient variable region sequence to confer binding specificity. A full-length light chain includes a variable region domain, VL, and a constant region domain, CL. The variable region domain of the light chain is at the amino-terminus of the polypeptide. Light chains
5 include kappa chains and lambda chains.

[0051] The term “heavy chain” includes a full-length heavy chain and fragments thereof having sufficient variable region sequence to confer binding specificity. A full-length heavy chain includes a variable region domain, VH, and three constant region domains, CH1, CH2, and CH3. The VH domain is at the amino-terminus of the polypeptide, and the
10 CH domains are at the carboxyl-terminus, with the CH3 being closest to the carboxy-terminus of the polypeptide. Heavy chains can be of any isotype, including IgG (including IgG1, IgG2, IgG3 and IgG4 subtypes), IgA (including IgA1 and IgA2 subtypes), IgM and IgE.

[0052] A bispecific or bifunctional antibody typically is an artificial hybrid antibody
15 having two different heavy/light chain pairs and two different binding sites. Bispecific antibodies can be produced by a variety of methods including, but not limited to, fusion of hybridomas or linking of Fab' fragments. See, e.g., Songsivilai et al., Clin. Exp. Immunol., 79: 315-321 (1990); Kostelny et al., J. Immunol., 148:1547-1553 (1992).

[0053] The term “antigen” refers to a substance capable of inducing adaptive immune
20 responses. Specifically, an antigen is a substance which serves as a target for the receptors of an adaptive immune response. Typically, an antigen is a molecule that binds to antigen-specific receptors but cannot induce an immune response in the body by itself. Antigens are usually proteins and polysaccharides, less frequently also lipids. As used herein, antigens also include immunogens and haptens.

[0054] An “Fc” region comprises two heavy chain fragments comprising the CH1 and
25 CH2 domains of an antibody. The two heavy chain fragments are held together by two or more disulfide bonds and by hydrophobic interactions of the CH3 domains.

[0055] The “Fv region” comprises the variable regions from both the heavy and light chains but lacks the constant regions.

[0056] An antibody that “specifically binds to” or is “specific for” a particular polypeptide or an epitope on a particular polypeptide is one that binds to that particular polypeptide or epitope on a particular polypeptide without substantially binding to any other polypeptide or polypeptide epitope. For example, the hMPV F protein specific antibodies of the present invention are specific to hMPV F protein. In some embodiments, the antibody that binds to hMPV F protein has a dissociation constant (Kd) of ≤ 100 nM, ≤ 10 nM, ≤ 1 nM, ≤ 0.1 nM, ≤ 0.01 nM, or ≤ 0.001 nM (e.g., 10^{-8} M or less, e.g., from 10^{-8} M to 10^{-13} M, e.g., from 10^{-9} M to 10^{-13} M).

[0057] The term “compete” when used in the context of antigen binding proteins (e.g., antibody or antigen-binding fragment thereof) that compete for the same epitope means competition between antigen binding proteins as determined by an assay in which the antigen binding protein (e.g., antibody or antigen-binding fragment thereof) being tested prevents or inhibits (e.g., reduces) specific binding of a reference antigen binding protein (e.g., a ligand, or a reference antibody) to a common antigen (e.g., hMPV F or a fragment thereof). Numerous types of competitive binding assays can be used to determine if one antigen binding protein competes with another, for example: solid phase direct or indirect radioimmunoassay (RIA), solid phase direct or indirect enzyme immunoassay (EIA), sandwich competition assay (see, e.g., Stahli et al., 1983, Methods in Enzymology 9:242-253); solid phase direct biotin-avidin EIA (see, e.g., Kirkland et al., 1986, J. Immunol. 137:3614-3619) solid phase direct labeled assay, solid phase direct labeled sandwich assay (see, e.g., Harlow and Lane, 1988, Antibodies, A Laboratory Manual, Cold Spring Harbor Press); solid phase direct label RIA using 125 I label (see, e.g., Morel et al., 1988, Molec. Immunol. 25:7-15); solid phase direct biotin-avidin EIA (see, e.g., Cheung, et al., 1990, Virology 176:546-552); and direct labeled RIA (Moldenhauer et al., 1990, Scand. J. Immunol. 32:77-82). Typically, such an assay involves the use of purified antigen bound to a solid surface or cells bearing either of these, an unlabeled test antigen binding protein and a labeled reference antigen binding protein. Competitive inhibition is measured by determining the amount of label bound to the solid surface or cells in the presence of the test antigen binding protein. Usually the test antigen binding protein is present in excess. Antigen binding proteins identified by competition assay (competing antigen binding proteins) include antigen binding proteins binding to the same epitope as the reference antigen binding proteins and antigen binding proteins binding to an adjacent epitope sufficiently proximal to the epitope bound by the reference antigen binding protein for steric hindrance to occur. Additional

details regarding methods for determining competitive binding are provided in the examples herein. Usually, when a competing antigen binding protein is present in excess, it will inhibit (e.g., reduce) specific binding of a reference antigen binding protein to a common antigen by at least 40-45%, 45-50%, 50-55%, 55-60%, 60-65%, 65-70%, 70-75% or 75% or more. In some instances, binding is inhibited by at least 80-85%, 85-90%, 90-95%, 95-97%, or 97% or more.

[0058] The term “epitope” as used herein refers to the specific group of atoms or amino acids on an antigen to which an antibody binds. The epitope can be either linear epitope or a conformational epitope. A linear epitope is formed by a continuous sequence of amino acids from the antigen and interacts with an antibody based on their primary structure. A conformational epitope, on the other hand, is composed of discontinuous sections of the antigen’s amino acid sequence and interacts with the antibody based on the 3D structure of the antigen. In general, an epitope is approximately five or six amino acid in length. Two antibodies may bind the same epitope within an antigen if they exhibit competitive binding for the antigen.

[0059] The term “host cell” means a cell that has been transformed, or is capable of being transformed, with a nucleic acid sequence and thereby expresses a gene of interest. The term includes the progeny of the parent cell, whether or not the progeny is identical in morphology or in genetic make-up to the original parent cell, so long as the gene of interest is present.

[0060] The term “identity” refers to a relationship between the sequences of two or more polypeptide molecules or two or more nucleic acid molecules, as determined by aligning and comparing the sequences. “Percent identity” means the percent of identical residues between the amino acids or nucleotides in the compared molecules and is calculated based on the size of the smallest of the molecules being compared. For these calculations, gaps in alignments (if any) are preferably addressed by a particular mathematical model or computer program (i.e., an “algorithm”). Methods that can be used to calculate the identity of the aligned nucleic acids or polypeptides include those described in Computational Molecular Biology, (Lesk, A. M., ed.), 1988, New York: Oxford University Press; Biocomputing Informatics and Genome Projects, (Smith, D. W., ed.), 1993, New York: Academic Press; Computer Analysis of Sequence Data, Part I, (Griffin, A. M., and Griffin, H. G., eds.), 1994, New Jersey: Humana Press; von Heinje, G., 1987, Sequence Analysis in Molecular Biology,

New York: Academic Press; Sequence Analysis Primer, (Gribskov, M. and Devereux, J., eds.), 1991, New York: M. Stockton Press; and Carillo et al., 1988, SIAM J. Applied Math. 48:1073.

[0061] In calculating percent identity, the sequences being compared are typically aligned in a way that gives the largest match between the sequences. One example of a computer program that can be used to determine percent identity is the GCG program package, which includes GAP (Devereux et al., 1984, Nucl. Acid Res. 12:387; Genetics Computer Group, University of Wisconsin, Madison, Wis.). The computer algorithm GAP is used to align the two polypeptides or polynucleotides for which the percent sequence identity is to be determined. The sequences are aligned for optimal matching of their respective amino acid or nucleotide (the “matched span”, as determined by the algorithm). A gap opening penalty (which is calculated as 3× the average diagonal, wherein the “average diagonal” is the average of the diagonal of the comparison matrix being used; the “diagonal” is the score or number assigned to each perfect amino acid match by the particular comparison matrix) and a gap extension penalty (which is usually 1/10 times the gap opening penalty), as well as a comparison matrix such as PAM 250 or BLOSUM 62 are used in conjunction with the algorithm. In certain embodiments, a standard comparison matrix (see, Dayhoff et al., 1978, Atlas of Protein Sequence and Structure 5:345-352 for the PAM 250 comparison matrix; Henikoff et al., 1992, Proc. Natl. Acad. Sci. U.S.A. 89:10915-10919 for the BLOSUM 62 comparison matrix) is also used by the algorithm.

[0062] Examples of parameters that can be employed in determining percent identity for polypeptides or nucleotide sequences using the GAP program can be found in Needleman et al., 1970, J. Mol. Biol. 48:443-453.

[0063] Certain alignment schemes for aligning two amino acid sequences may result in matching of only a short region of the two sequences, and this small aligned region may have very high sequence identity even though there is no significant relationship between the two full-length sequences. Accordingly, the selected alignment method (GAP program) can be adjusted if so desired to result in an alignment that spans at least 50 or other number of contiguous amino acids of the target polypeptide.

[0064] The term “link” as used herein refers to the association via intramolecular interaction, e.g., covalent bonds, metallic bonds, and/or ionic bonding, or inter-molecular interaction, e.g., hydrogen bond or noncovalent bonds.

5 [0065] The term “operably linked” refers to an arrangement of elements wherein the components so described are configured so as to perform their usual function. Thus, a given signal peptide that is operably linked to a polypeptide directs the secretion of the polypeptide from a cell. In the case of a promoter, a promoter that is operably linked to a coding sequence will direct the expression of the coding sequence. The promoter or other control elements need not be contiguous with the coding sequence, so long as they function to direct the
10 expression thereof. For example, intervening untranslated yet transcribed sequences can be present between the promoter sequence and the coding sequence and the promoter sequence can still be considered “operably linked” to the coding sequence.

[0066] The term “polynucleotide” or “nucleic acid” includes both single-stranded and double-stranded nucleotide polymers. The nucleotides comprising the polynucleotide can be
15 ribonucleotides or deoxyribonucleotides or a modified form of either type of nucleotide. Said modifications include base modifications such as bromouridine and inosine derivatives, ribose modifications such as 2',3'-dideoxyribose, and internucleotide linkage modifications such as phosphorothioate, phosphorodithioate, phosphoroselenoate, phosphorodiselenoate, phosphoroanilothioate, phosphoraniladate and phosphoroamidate.

20 [0067] As used herein, a “vector” refers to a nucleic acid molecule as introduced into a host cell, thereby producing a transformed host cell. A vector may include nucleic acid sequences that permit it to replicate in the host cell, such as an origin of replication. A vector may also include one or more therapeutic genes and/or selectable marker genes and other genetic elements known in the art. A vector can transduce, transform or infect a cell, thereby
25 causing the cell to express nucleic acids and/or proteins other than those native to the cell. A vector optionally includes materials to aid in achieving entry of the nucleic acid into the cell, such as a viral particle, liposome, protein coating or the like.

[0068] The terms “polypeptide” or “protein” means a macromolecule having the amino acid sequence of a native protein, that is, a protein produced by a naturally-occurring
30 and non-recombinant cell; or it is produced by a genetically engineered or recombinant cell, and comprise molecules having the amino acid sequence of the native protein, or molecules

having deletions from, additions to, and/or substitutions of one or more amino acids of the native sequence. The term also includes amino acid polymers in which one or more amino acids are chemical analogs of a corresponding naturally occurring amino acid and polymers. The terms “polypeptide” and “protein” specifically encompass hMPV F protein binding proteins, antibodies, or sequences that have deletions from, additions to, and/or substitutions of one or more amino acid of antigen-binding protein. The term “polypeptide fragment” refers to a polypeptide that has an amino-terminal deletion, a carboxyl-terminal deletion, and/or an internal deletion as compared with the full-length native protein. Such fragments can also contain modified amino acids as compared with the native protein. In certain embodiments, fragments are about five to 500 amino acids long. For example, fragments can be at least 5, 6, 8, 10, 14, 20, 50, 70, 100, 110, 150, 200, 250, 300, 350, 400, or 450 amino acids long. Useful polypeptide fragments include immunologically functional fragments of antibodies, including binding domains. In the case of a hMPV F protein-binding antibody, useful fragments include but are not limited to a CDR region, a variable domain of a heavy and/or light chain, a portion of an antibody chain or just its variable region including two CDRs, and the like.

[0069] The pharmaceutically acceptable carriers useful in this invention are conventional. Remington’s Pharmaceutical Sciences, by E. W. Martin, Mack Publishing Co., Easton, PA, 15th Edition (1975), describes compositions and formulations suitable for pharmaceutical delivery of the fusion proteins herein disclosed. In general, the nature of the carrier will depend on the particular mode of administration being employed. For instance, parenteral formulations usually comprise injectable fluids that include pharmaceutically and physiologically acceptable fluids such as water, physiological saline, balanced salt solutions, aqueous dextrose, glycerol or the like as a vehicle. For solid compositions (e.g., powder, pill, tablet, or capsule forms), conventional non-toxic solid carriers can include, for example, pharmaceutical grades of mannitol, lactose, starch or magnesium stearate. In addition to biologically- neutral carriers, pharmaceutical compositions to be administered can contain minor amounts of non-toxic auxiliary substances, such as wetting or emulsifying agents, preservatives, and pH buffering agents and the like, for example sodium acetate or sorbitan monolaurate.

[0070] As used herein, the term “subject” refers to a human or any non-human animal (e.g., mouse, rat, rabbit, dog, cat, cattle, swine, sheep, horse or primate). A human includes

pre- and post-natal forms. In many embodiments, a subject is a human being. A subject can be a patient, which refers to a human presenting to a medical provider for diagnosis or treatment of a disease. The term “subject” is used herein interchangeably with “individual” or “patient.” A subject can be afflicted with or is susceptible to a disease or disorder but may or
5 may not display symptoms of the disease or disorder.

[0071] The term “therapeutically effective amount” or “effective dosage” as used herein refers to the dosage or concentration of a drug effective to treat a disease or condition. For example, with regard to the use of the monoclonal antibodies or antigen-binding fragments thereof disclosed herein to treat viral infection.

10 [0072] “Treating” or “treatment” of a condition as used herein includes preventing or alleviating a condition, slowing the onset or rate of development of a condition, reducing the risk of developing a condition, preventing or delaying the development of symptoms associated with a condition, reducing or ending symptoms associated with a condition, generating a complete or partial regression of a condition, curing a condition, or some
15 combination thereof.

II. Chimeric hMPV/RSV F Proteins

[0073] Human metapneumovirus (hMPV) is a negative-sense enveloped virus of the *Pneumoviridae* family and was discovered in 2001 but has been circulating for at least a half century before its discovery. The hMPV fusion (F) protein is one of three surface
20 glycoproteins encoded by the viral genome. As a class I fusion, hMPV F is first translated as a single polypeptide precursor (F0). Initially nonfunctional, a proteolytic cleavage event is necessary to form the F1 and F2 subunits which are covalently linked by disulfide bonds. The new N-terminal of the F2 polypeptide contains a hydrophobic sequence which gets inserted into the host-cell membrane during the process that eventually fuses the viral and host-cell
25 membranes. At some point, either in transit or at the membrane surface, the F protein associates with itself to form a metastable trimer in what is termed to be the prefusion conformation. The hMPV fusion protein is cleaved extracellularly by a trypsin-like protease. An unknown triggering event occurs that influences the F protein to undergo a dramatic conformational change, extending the fusion peptide into the host-cell membrane, before
30 collapsing back onto itself to form a six-helix bundle in what is termed the postfusion

conformation. The energy differential between the elongated intermediate and the postfusion conformation provides the energy necessary for membrane fusion.

[0074] Historically the structures of the paramyxovirus fusion proteins have been divided into domains generally segmenting the first solved prefusion protein into a head, neck, and stalk. The nomenclature of assigning three general domains (DI, DII, and DIII) to the prefusion structures of additional structures has continued. Since the pneumoviridae family was previously a subfamily of the paramyxoviruses, they too retain this convention. The domains of hMPV are vaguely broken into the same three domains with an additional two regions of heptad repeats defined as well (supplemental figure). Heptad repeat A (HRA) is at the N-terminal of F1 located within DIIIa and heptad repeat B is at the C-terminal outside of the defined domain regions and prior to the transmembrane domain of the protein. However, transferring the antigenic site nomenclature used for the respiratory syncytial virus (RSV), another member of the pneumoviridae family, fusion protein can be used for a more precise description of protein areas due to their structural similarity. DIIIa experiences the largest conformational rearrangement when the fusion peptide is released from the internal cavity of the trimer and HRA forms a three-coil bundle as the fusion peptide is inserted into the target membrane. HRB eventually associates around the outside of the HRA bundle to form the 6HB in the postfusion conformation.

[0075] The RSV F glycoprotein is a type I fusion protein that facilitates fusion of viral and cellular membranes. After initial synthesis, RSV F adopts a metastable prefusion conformation that stores folding energy, which is released during a structural rearrangement to a highly stable postfusion conformation after contact with host cell membranes. Three antigenic sites (I, II, and IV) on RSV F protein have been found to elicit neutralizing activity, and all exist on the postfusion form of RSV F protein as determined by structural and biophysical studies. Absorption of human sera with postfusion RSV F, however, fails to remove the majority of F-specific neutralizing activity, suggesting that the prefusion form of RSV F harbors novel neutralizing antigenic sites.

[0076] Stabilization of class I fusion proteins in their prefusion conformation has recently seen promising results as vaccine antigens in clinical trials, with both RSV and SARS-Coronavirus-2 vaccines using this approach. RSV shares ~38% sequence identity with hMPV with a nearly identical quaternary structure. Based on its solved prefusion structure, a few different strategies have been used to stabilize its fusion protein. For DS-Cav1, internal

cavities were filled more optimally by hydrophobic residue substitutions, and the introduction of a disulfide bond was introduced into the fusion peptide region. For PR-DM, a proline residue was introduced to prevent a loop region from rearranging into the extended alpha helix seen in the postfusion conformation. In RSV F areas of charge repulsion were also identified and reduced. Similarly, a solved prefusion structure allowed for the stabilization of coronaviruses by introducing two proline substitutions (S-2P) into a hinge region. There are also examples within the HIV (SOSIP) and influenza field where engineering stabilizing mutations have resulted in well behaved prefusion reagents. Recently hMPV F prefusion structure has been solved by inserting a proline into the loop region that was exploited by the RSV PR-DM construct. Variants that combine prefusion-stabilized epitopes from both hMPV F and RSV F are needed to elicit pan-pneumovirus antibodies from a single antigen. To this end, provided herein are chimeric fusions of RSV and hMPV F proteins that serve as a stabilized antigen. Substitutions of the chimeric proteins analyzed and provided herein are detailed in Table 1.

Table 1. Chimeric hMPV/RSV F protein substitutions.

Designation	hMPV substitution positions relative to any of SEQ ID NOs: 5-8	RSV substitution positions relative to any of SEQ ID NOs: 10	Positions of hMPV and RSV combined substitutions relative to SEQ ID NO: 1
	Q100R; S101R		Q103R; S104R
	T127C/N153C		T130C/N156C
	V231I		V234I
	T365C/V463C		T368C/N466C
	L110C/N322C		L113C/N325C
	A107P		A110P
	L219K		L222K
	E453Q		E456Q
		N67I	N58I
		T91R	T82R
		L95R	L86R
		S215P	S188P
Chimera 1	Q100R; S101R; T127C/N153C; V231I; T365C/V463C	N67I; S215P	Q103R; S104R; T130C/N156C; V234I; T368C/N466C; N58I; S188P
Chimera 1_L95R	Q100R; S101R; T127C/N153C; V231I; T365C/V463C	N67I; L95R; S215P	Q103R; S104R; T130C/N156C; V234I; T368C/N466C; N58I; L86R; S188P
Chimera 1_L95R_L219K	Q100R; S101R; T127C/N153C; V231I; T365C/V463C; L219K	N67I; L95R; S215P	Q103R; S104R; T130C/N156C; V234I; T368C/N466C; L222K; N58I; L86R; S188P
Chimera 1_L95R_E453Q	Q100R; S101R; T127C/N153C; V231I; T365C/V463C; E453Q	N67I; L95R; S215P	Q103R; S104R; T130C/N156C; V234I; T368C/N466C; E456Q; N58I; L86R; S188P
Chimera 1_L95R_A107P	Q100R; S101R; T127C/N153C; V231I; T365C/V463C; A107P	N67I; L95R; S215P	Q103R; S104R; T130C/N156C; V234I; T368C/N466C; A110P; N58I; L86R; S188P

Chimera 1_L95R_L11 0C/N322C	Q100R; S101R; T127C/N153C; V231I; T365C/V463C; L110C/N322C	N67I; L95R; S215P	Q103R; S104R; T130C/N156C; V234I; T368C/N466C; L113C/N325C; N58I; L86R; S188P
DS-CavEsP	Q100R; S101R; T127C/N153C; V231I; T365C/V463C; L110C/N322C; A107P; L219K; E453Q;	N67I; L95R; S215P	Q103R; S104R; T130C/N156C; V234I; T368C/N466C; L113C/N325C; A110P; L222K; E456Q; N58I; L86R; S188P

III. Pharmaceutical Formulations

[0077] The present disclosure provides pharmaceutical compositions comprising a chimeric hMPV/RSV F protein. Such compositions can be used for stimulating an immune response, such as part of vaccine formulation.

5 [0078] In the case that a nucleic acid molecule encoding a chimeric hMPV/RSV F protein is used in a pharmaceutical composition, the nucleic acid molecule may comprise or consist of deoxyribonucleotides and/or ribonucleotides, or analogs thereof, covalently linked together. A nucleic acid molecule as described herein generally contains phosphodiester bonds, although in some cases, nucleic acid analogs are included that may have at least one
10 different linkage, *e.g.*, phosphoramidate, phosphorothioate, phosphorodithioate, or O-methylphosphoroamidite linkages, and peptide nucleic acid backbones and linkages. Mixtures of naturally occurring polynucleotides and analogs can be made; alternatively, mixtures of different polynucleotide analogs, and mixtures of naturally occurring polynucleotides and analogs may be made. A nucleic acid molecule may comprise modified
15 nucleotides, such as methylated nucleotides and nucleotide analogs. If present, modifications to the nucleotide structure may be imparted before or after assembly of the polymer. The sequence of nucleotides may be interrupted by non-nucleotide components. A polynucleotide may be further modified after polymerization, such as by conjugation with a labeling component. The term also includes both double- and single-stranded molecules. Unless
20 otherwise specified or required, the term polynucleotide encompasses both the double-stranded form and each of two complementary single-stranded forms known or predicted to make up the double-stranded form. A nucleic acid molecule is composed of a specific sequence of four nucleotide bases: adenine (A), cytosine (C), guanine (G), thymine (T), and uracil (U) for thymine when the polynucleotide is RNA. Thus, the term “nucleic acid
25 sequence” is the alphabetical representation of a nucleic acid molecule. Unless otherwise indicated, a particular nucleic acid sequence also implicitly encompasses conservatively modified variants thereof (*e.g.*, degenerate codon substitutions) and complementary sequences as well as the sequence explicitly indicated. Specifically, degenerate codon substitutions may be achieved by generating sequences in which the third position of one or
30 more selected (or all) codons is substituted with mixed-base and/or deoxyinosine residues.

[0079] In some embodiments, the nucleic acids of the present disclosure comprise one or more modified nucleosides comprising a modified sugar moiety. Such compounds

comprising one or more sugar-modified nucleosides may have desirable properties, such as enhanced nuclease stability or increased binding affinity with a target nucleic acid relative to an oligonucleotide comprising only nucleosides comprising naturally occurring sugar moieties. In some embodiments, modified sugar moieties are substituted sugar moieties. In some embodiments, modified sugar moieties are sugar surrogates. Such sugar surrogates may comprise one or more substitutions corresponding to those of substituted sugar moieties.

[0080] In some embodiments, modified sugar moieties are substituted sugar moieties comprising one or more non-bridging sugar substituent, including but not limited to substituents at the 2' and/or 5' positions. Examples of sugar substituents suitable for the 2'-position, include, but are not limited to: 2'-F, 2'-OCH₃ ("OMe" or "O-methyl"), and 2'-O(CH₂)₂OCH₃ ("MOE"). In certain embodiments, sugar substituents at the 2' position is selected from allyl, amino, azido, thio, O-allyl, O--C₁-C₁₀ alkyl, O--C₁-C₁₀ substituted alkyl; OCF₃, O(CH₂)₂SCH₃, O(CH₂)₂--O--N(R_m)(R_n), and O--CH₂--C(=O)--N(R_m)(R_n), where each R_m and R_n is, independently, H or substituted or unsubstituted C₁-C₁₀ alkyl. Examples of sugar substituents at the 5'-position, include, but are not limited to: 5'-methyl (R or S); 5'-vinyl, and 5'-methoxy. In some embodiments, substituted sugars comprise more than one non-bridging sugar substituent, for example, T-F-5'-methyl sugar moieties (see, e.g., PCT International Application WO 2008/101157, for additional 5',2'-bis substituted sugar moieties and nucleosides).

[0081] Nucleosides comprising 2'-substituted sugar moieties are referred to as 2'-substituted nucleosides. In some embodiments, a 2'-substituted nucleoside comprises a 2'-substituent group selected from halo, allyl, amino, azido, SH, CN, OCN, CF₃, OCF₃, O, S, or N(R_m)-alkyl; O, S, or N(R_m)-alkenyl; O, S or N(R_m)-alkynyl; O-alkylenyl-O-alkyl, alkynyl, alkaryl, aralkyl, O-alkaryl, O-aralkyl, O(CH₂)₂SCH₃, O(CH₂)₂--O--N(R_m)(R_n) or O--CH₂--C(=O)--N(R_m)(R_n), where each R_m and R_n is, independently, H, an amino protecting group or substituted or unsubstituted C₁-C₁₀ alkyl. These 2'-substituent groups can be further substituted with one or more substituent groups independently selected from hydroxyl, amino, alkoxy, carboxy, benzyl, phenyl, nitro (NO₂), thiol, thioalkoxy (S-alkyl), halogen, alkyl, aryl, alkenyl and alkynyl.

[0082] In some embodiments, a 2'-substituted nucleoside comprises a 2'-substituent group selected from F, NH₂, N₃, OCF₃, O--CH₃, O(CH₂)₃NH₂, CH₂—CH=CH₂, O--CH₂—CH=CH₂, OCH₂CH₂OCH₃, O(CH₂)₂SCH₃, O--(CH₂)₂--O--N(R_m)(R_n),

O(CH₂)₂O(CH₂)₂N(CH₃)₂, and N-substituted acetamide (O--CH₂--C(=O)--N(R_m)(R_n) where each R_m and R_n is, independently, H, an amino protecting group or substituted or unsubstituted C1-C10 alkyl.

[0083] In some embodiments, a 2'-substituted nucleoside comprises a sugar moiety comprising a 2'-substituent group selected from F, OCF₃, O--CH₃, OCH₂CH₂OCH₃, O(CH₂)₂SCH₃, O(CH₂)₂--O--N(CH₃)₂, --O(CH₂)₂O(CH₂)₂N(CH₃)₂, and O--CH₂--C(=O)--N(H)CH₃.

[0084] In some embodiments, a 2'-substituted nucleoside comprises a sugar moiety comprising a 2'-substituent group selected from F, O--CH₃, and OCH₂CH₂OCH₃.

[0085] In some embodiments, nucleosides of the present disclosure comprise one or more unmodified nucleobases. In certain embodiments, nucleosides of the present disclosure comprise one or more modified nucleobases.

[0086] In some embodiments, modified nucleobases are selected from: universal bases, hydrophobic bases, promiscuous bases, size-expanded bases, and fluorinated bases as defined herein. 5-substituted pyrimidines, 6-azapyrimidines and N-2, N-6 and O-6 substituted purines, including 2-aminopropyladenine, 5-propynyluracil; 5-propynylcytosine; 5-hydroxymethyl cytosine, xanthine, hypoxanthine, 2-aminoadenine, 6-methyl and other alkyl derivatives of adenine and guanine, 2-propyl and other alkyl derivatives of adenine and guanine, 2-thiouracil, 2-thiothymine and 2-thiocytosine, 5-halouracil and cytosine, 5-propynyl CH₃) uracil and cytosine and other alkynyl derivatives of pyrimidine bases, 6-azouracil, cytosine and thymine, 5-uracil (pseudouracil), 4-thiouracil, 8-halo, 8-amino, 8-thiol, 8-thioalkyl, 8-hydroxyl and other 8-substituted adenines and guanines, 5-halo particularly 5-bromo, 5-trifluoromethyl and other 5-substituted uracils and cytosines, 7-methylguanine and 7-methyladenine, 2-F-adenine, 2-amino-adenine, 8-azaguanine and 8-azaadenine, 7-deazaguanine and 7-deazaadenine, 3-deazaguanine and 3-deazaadenine, universal bases, hydrophobic bases, promiscuous bases, size-expanded bases, and fluorinated bases as defined herein. Further modified nucleobases include tricyclic pyrimidines such as phenoxazine cytidine([5,4-b][1,4]benzoxazin-2(3H)-one), phenothiazine cytidine (1H-pyrimido[5,4-b][1,4]benzothiazin-2(3H)-one), G-clamps such as a substituted phenoxazine cytidine (e.g., 9-(2-aminoethoxy)-H-pyrimido[5,4-13][1,4]benzoxazin-2(3H)-one), carbazole cytidine (2H-pyrimido[4,5-b]indol-2-one), pyridoindole cytidine (H-pyrido[3',2':4,5]pyrrolo[2,3-

d]pyrimidin-2-one). Modified nucleobases may also include those in which the purine or pyrimidine base is replaced with other heterocycles, for example 7-deaza-adenine, 7-deazaguanosine, 2-aminopyridine and 2-pyridone. Further nucleobases include those disclosed in U.S. Patent 3,687,808, those disclosed in The Concise Encyclopedia Of Polymer
5 Science And Engineering, Kroschwitz, J. I., Ed., John Wiley & Sons, 1990, 858-859.

[0087] Representative United States Patents that teach the preparation of certain of the above noted modified nucleobases as well as other modified nucleobases include without limitation, U.S. Patents 3,687,808; 4,845,205; 5,130,302; 5,134,066; 5,175,273; 5,367,066; 5,432,272; 5,457,187; 5,459,255; 5,484,908; 5,502,177; 5,525,711; 5,552,540; 5,587,469;
10 5,594,121; 5,596,091; 5,614,617; 5,645,985; 5,681,941; 5,750,692; 5,763,588; 5,830,653 and 6,005,096, each of which is herein incorporated by reference in its entirety.

[0088] Additional modifications may also be made at other positions on the oligonucleotide, particularly the 3' position of the sugar on the 3' terminal nucleotide and the 5' position of 5' terminal nucleotide. For example, one additional modification of the ligand
15 conjugated oligonucleotides of the present disclosure involves chemically linking to the oligonucleotide one or more additional non-ligand moieties or conjugates which enhance the activity, cellular distribution or cellular uptake of the oligonucleotide. Such moieties include but are not limited to lipid moieties such as a cholesterol moiety, cholic acid, a thioether, e.g., hexyl-5-tritylthiol, a thiocholesterol, an aliphatic chain, e.g., dodecandiol or undecyl residues,
20 a phospholipid, e.g., di-hexadecyl-rac-glycerol or triethylammonium 1,2-di-O-hexadecyl-rac-glycero-3-H-phosphonate, a polyamine or a polyethylene glycol chain, or adamantane acetic acid, a palmityl moiety, or an octadecylamine or hexylamino-carbonyl-oxycholesterol moiety. In some aspects, a nucleic acid molecule encoding a chimeric hMPV/RSV F protein is a modified RNA, such as, for example, a modified mRNA. Modified (m)RNA
25 contemplates certain chemical modifications that confer increased stability and low immunogenicity to mRNAs, thereby facilitating expression of therapeutically important proteins. For instance, N1-methyl-pseudouridine (N1m Ψ) outperforms several other nucleoside modifications and their combinations in terms of translation capacity. In some embodiments, the (m)RNA molecules used herein may have the uracils replaced with
30 psuedouracils such as 1-methyl-3'-pseudouridylyl bases. In some embodiments, some of the uracils are replaced, but in other embodiments, all of the uracils have been replaced. The

(m)RNA may comprise a 5' cap, a 5' UTR element, an optionally codon optimized open reading frame, a 3' UTR element, and a poly(A) sequence and/or a polyadenylation signal.

[0089] The nucleic acid molecule, whether native or modified, may be delivered as a naked nucleic acid molecule or in a delivery vehicle, such as a lipid nanoparticle. A lipid nanoparticle may comprise one or more nucleic acids present in a weight ratio to the lipid nanoparticles from about 5:1 to about 1:100. In some embodiments, the weight ratio of nucleic acid to lipid nanoparticles is from about 5:1, 2.5:1, 1:1, 1:5, 1:10, 1:15, 1:20, 1:25, 1:30, 1:35, 1:40, 1:45, 1:50, 1:60, 1:70, 1:80, 1:90, or 1:100, or any value derivable therein.

[0090] In some embodiments, the lipid nanoparticles used herein may contain one, two, three, four, five, six, seven, eight, nine, or ten lipids. These lipids may include triglycerides, phospholipids, steroids or sterols, a PEGylated lipids, or a group with a ionizable group such as an alkyl amine and one or more hydrophobic groups such as C6 or greater alkyl groups.

[0091] In some aspects of the present disclosure, the lipid nanoparticles are mixed with one or more steroid or a steroid derivative. In some embodiments, the steroid or steroid derivative comprises any steroid or steroid derivative. As used herein, in some embodiments, the term "steroid" is a class of compounds with a four ring 17 carbon cyclic structure, which can further comprises one or more substitutions including alkyl groups, alkoxy groups, hydroxy groups, oxo groups, acyl groups, or a double bond between two or more carbon atoms.

[0092] In some aspects of the present disclosure, the lipid nanoparticles are mixed with one or more PEGylated lipids (or PEG lipid). In some embodiments, the present disclosure comprises using any lipid to which a PEG group has been attached. In some embodiments, the PEG lipid is a diglyceride which also comprises a PEG chain attached to the glycerol group. In other embodiments, the PEG lipid is a compound which contains one or more C6-C24 long chain alkyl or alkenyl group or a C6-C24 fatty acid group attached to a linker group with a PEG chain. Some non-limiting examples of a PEG lipid includes a PEG modified phosphatidylethanolamine and phosphatidic acid, a PEG ceramide conjugated, PEG modified dialkylamines and PEG modified 1,2-diacyloxypropan-3-amines, PEG modified diacylglycerols and dialkylglycerols. In some embodiments, PEG modified diastearoylphosphatidylethanolamine or PEG modified dimyristoyl-sn-glycerol. In some

embodiments, the PEG modification is measured by the molecular weight of PEG component of the lipid. In some embodiments, the PEG modification has a molecular weight from about 100 to about 15,000. In some embodiments, the molecular weight is from about 200 to about 500, from about 400 to about 5,000, from about 500 to about 3,000, or from about 1,200 to about 3,000. The molecular weight of the PEG modification is from about 100, 200, 400, 500, 600, 800, 1,000, 1,250, 1,500, 1,750, 2,000, 2,250, 2,500, 2,750, 3,000, 3,500, 4,000, 4,500, 5,000, 6,000, 7,000, 8,000, 9,000, 10,000, 12,500, to about 15,000. Some non-limiting examples of lipids that may be used in the present disclosure are taught by U.S. Patent 5,820,873, WO 2010/141069, or U.S. Patent 8,450,298, which is incorporated herein by reference.

[0093] In some aspects of the present disclosure, the lipid nanoparticles are mixed with one or more phospholipids. In some embodiments, any lipid which also comprises a phosphate group. In some embodiments, the phospholipid is a structure which contains one or two long chain C6-C24 alkyl or alkenyl groups, a glycerol or a sphingosine, one or two phosphate groups, and, optionally, a small organic molecule. In some embodiments, the small organic molecule is an amino acid, a sugar, or an amino substituted alkoxy group, such as choline or ethanolamine. In some embodiments, the phospholipid is a phosphatidylcholine. In some embodiments, the phospholipid is distearoylphosphatidylcholine or dioleoylphosphatidylethanolamine. In some embodiments, other zwitterionic lipids are used, where zwitterionic lipid defines lipid and lipid-like molecules with both a positive charge and a negative charge.

[0094] In some aspects of the present disclosure, lipid nanoparticle containing compounds containing lipophilic and cationic components, wherein the cationic component is ionizable, are provided. In some embodiments, the cationic ionizable lipids contain one or more groups which is protonated at physiological pH but may deprotonated and has no charge at a pH above 8, 9, 10, 11, or 12. The ionizable cationic group may contain one or more protonatable amines which are able to form a cationic group at physiological pH. The cationic ionizable lipid compound may also further comprise one or more lipid components such as two or more fatty acids with C6-C24 alkyl or alkenyl carbon groups. These lipid groups may be attached through an ester linkage or may be further added through a Michael addition to a sulfur atom. In some embodiments, these compounds may be a dendrimer, a dendron, a polymer, or a combination thereof.

[0095] In some aspects of the present disclosure, composition containing compounds containing lipophilic and cationic components, wherein the cationic component is ionizable, are provided. In some embodiments, ionizable cationic lipids refer to lipid and lipid-like molecules with nitrogen atoms that can acquire charge (pKa). These lipids may be known in the literature as cationic lipids. These molecules with amino groups typically have between 2 and 6 hydrophobic chains, often alkyl or alkenyl such as C6-C24 alkyl or alkenyl groups, but may have at least 1 or more than 6 tails.

[0096] In some embodiments, the amount of the lipid nanoparticle with the nucleic acid molecule encapsulated in the pharmaceutical composition is from about 0.1% w/w to about 50% w/w, from about 0.25% w/w to about 25% w/w, from about 0.5% w/w to about 20% w/w, from about 1% w/w to about 15% w/w, from about 2% w/w to about 10% w/w, from about 2% w/w to about 5% w/w, or from about 6% w/w to about 10% w/w. In some embodiments, the amount of the lipid nanoparticle with the nucleic acid molecule encapsulated in the pharmaceutical composition is from about 0.1% w/w, 0.25% w/w, 0.5% w/w, 1% w/w, 2.5% w/w, 5% w/w, 7.5% w/w, 10% w/w, 15% w/w, 20% w/w, 25% w/w, 30% w/w, 35% w/w, 40% w/w, 45% w/w, 50% w/w, 55% w/w, 60% w/w, 65% w/w, 70% w/w, 75% w/w, 80% w/w, 85% w/w, 90% w/w, to about 95% w/w, or any range derivable therein.

[0097] In some aspects, the present disclosure comprises one or more sugars formulated into pharmaceutical compositions. In some embodiments, the sugars used herein are saccharides. These saccharides may be used to act as a lyoprotectant that protects the pharmaceutical composition from destabilization during the drying process. These water-soluble excipients include carbohydrates or saccharides such as disaccharides such as sucrose, trehalose, or lactose, a trisaccharide such as fructose, glucose, galactose comprising raffinose, polysaccharides such as starches or cellulose, or a sugar alcohol such as xylitol, sorbitol, or mannitol. In some embodiments, these excipients are solid at room temperature. Some non-limiting examples of sugar alcohols include erythritol, threitol, arabitol, xylitol, ribitol, mannitol, sorbitol, galactitol, fucitol, iditol, inositol, volemitol, isomalt, maltitol, lactitol, maltotritol, maltotetraitol, or a polyglycitol.

[0098] In some embodiments, the amount of the sugar in the pharmaceutical composition is from about 25% w/w to about 98% w/w, 40% w/w to about 95% w/w, 50% w/w to about 90% w/w, 50% w/w to about 70% w/w, or from about 80% w/w to about 90%

w/w. In some embodiments, the amount of the sugar in the pharmaceutical composition is from about 10% w/w, 15% w/w, 20% w/w, 25% w/w, 30% w/w, 35% w/w, 40% w/w, 45% w/w, 50% w/w, 52.5% w/w, 55% w/w, 57.5% w/w, 60% w/w, 62.5% w/w, 65% w/w, 67.5% w/w, 70% w/w, 75% w/w, 80% w/w, 82.5% w/w, 85% w/w, 87.5% w/w, 90% w/w, to about 5 95% w/w, or any range derivable therein.

[0099] In some embodiments, the pharmaceutically acceptable polymer is a copolymer. The pharmaceutically acceptable polymer may further comprise one, two, three, four, five, or six subunits of discrete different types of polymer subunits. These polymer subunits may include polyoxypropylene, polyoxyethylene, or a similar subunit. In particular, 10 the pharmaceutically acceptable polymer may comprise at least one hydrophobic subunit and at least one hydrophilic subunit. In particular, the copolymer may have hydrophilic subunits on each side of a hydrophobic unit. The copolymer may have a hydrophilic subunit that is polyoxyethylene and a hydrophobic subunit that is polyoxypropylene.

[00100] In some embodiments, expression cassettes are employed to express a 15 chimeric hMPV/RSV F protein, either for subsequent purification and delivery to a cell/subject, or for use directly in a viral-based delivery approach. Provided herein are expression vectors which contain one or more nucleic acids encoding a chimeric hMPV/RSV F protein.

[00101] Expression requires that appropriate signals be provided in the vectors 20 and include various regulatory elements such as enhancers/promoters from both viral and mammalian sources that drive expression of a chimeric hMPV/RSV F protein in cells. Throughout this application, the term “expression cassette” is meant to include any type of genetic construct containing a nucleic acid coding for a gene product in which part or all of the nucleic acid encoding sequence is capable of being transcribed and translated, *i.e.*, is 25 under the control of a promoter. A “promoter” refers to a DNA sequence recognized by the synthetic machinery of the cell, or introduced synthetic machinery, required to initiate the specific transcription of a gene. The phrase “under transcriptional control” means that the promoter is in the correct location and orientation in relation to the nucleic acid to control RNA polymerase initiation and expression of the gene. An “expression vector” is meant to 30 include expression cassettes comprised in a genetic construct that is capable of replication, and thus including one or more of origins of replication, transcription termination signals, poly-A regions, selectable markers, and multipurpose cloning sites.

[00102] The term promoter will be used here to refer to a group of transcriptional control modules that are clustered around the initiation site for RNA polymerase II. Much of the thinking about how promoters are organized derives from analyses of several viral promoters, including those for the HSV thymidine kinase (tk) and
5 SV40 early transcription units. These studies, augmented by more recent work, have shown that promoters are composed of discrete functional modules, each consisting of approximately 7-20 bp of DNA, and containing one or more recognition sites for transcriptional activator or repressor proteins.

[00103] At least one module in each promoter functions to position the start site
10 for RNA synthesis. The best known example of this is the TATA box, but in some promoters lacking a TATA box, such as the promoter for the mammalian terminal deoxynucleotidyl transferase gene and the promoter for the SV40 late genes, a discrete element overlying the start site itself helps to fix the place of initiation.

[00104] Additional promoter elements regulate the frequency of transcriptional
15 initiation. Typically, these are located in the region 30-110 bp upstream of the start site, although a number of promoters have recently been shown to contain functional elements downstream of the start site as well. The spacing between promoter elements frequently is flexible, so that promoter function is preserved when elements are inverted or moved relative to one another. In the tk promoter, the spacing between promoter elements can be increased
20 to 50 bp apart before activity begins to decline. Depending on the promoter, it appears that individual elements can function either co-operatively or independently to activate transcription.

[00105] In certain embodiments, viral promoters such as the human cytomegalovirus (CMV) immediate early gene promoter, the SV40 early promoter, the Rous
25 sarcoma virus long terminal repeat, rat insulin promoter and glyceraldehyde-3-phosphate dehydrogenase can be used to obtain high-level expression of the coding sequence of interest. The use of other viral or mammalian cellular or bacterial phage promoters which are well-known in the art to achieve expression of a coding sequence of interest is contemplated as well, provided that the levels of expression are sufficient for a given purpose. By employing a
30 promoter with well-known properties, the level and pattern of expression of the protein of interest following transfection or transformation can be optimized. Further, selection of a

promoter that is regulated in response to specific physiologic signals can permit inducible expression of the gene product.

5 **[00106]** Enhancers are genetic elements that increase transcription from a promoter located at a distant position on the same molecule of DNA. Enhancers are organized much like promoters. That is, they are composed of many individual elements, each of which binds to one or more transcriptional proteins. The basic distinction between enhancers and promoters is operational. An enhancer region as a whole must be able to stimulate transcription at a distance; this need not be true of a promoter region or its component elements. On the other hand, a promoter must have one or more elements that
10 direct initiation of RNA synthesis at a particular site and in a particular orientation, whereas enhancers lack these specificities. Promoters and enhancers are often overlapping and contiguous, often seeming to have a very similar modular organization.

[00107] Below is a list of promoters/enhancers and inducible promoters/enhancers that could be used in combination with the nucleic acid encoding a gene
15 of interest in an expression construct. Additionally, any promoter/enhancer combination (as per the Eukaryotic Promoter Data Base EPDB) could also be used to drive expression of the gene. Eukaryotic cells can support cytoplasmic transcription from certain bacterial promoters if the appropriate bacterial polymerase is provided, either as part of the delivery complex or as an additional genetic expression construct.

20 **[00108]** The promoter and/or enhancer may be, for example, immunoglobulin light chain, immunoglobulin heavy chain, T-cell receptor, HLA DQ a and/or DQ β , β -interferon, interleukin-2, interleukin-2 receptor, MHC class II 5, MHC class II HLA-Dra, β -Actin, muscle creatine kinase (MCK), prealbumin (transthyretin), elastase I, metallothionein (MTII), collagenase, albumin, α -fetoprotein, t-globin, β -globin, c-fos, c-HA-
25 ras, insulin, neural cell adhesion molecule (NCAM), α 1-antitrypsin, H2B (TH2B) histone, mouse and/or type I collagen, glucose-regulated proteins (GRP94 and GRP78), rat growth hormone, human serum amyloid A (SAA), troponin I (TN I), platelet-derived growth factor (PDGF), SV40, polyoma, retroviruses, papilloma virus, hepatitis B virus, human immunodeficiency virus, cytomegalovirus (CMV), and gibbon ape leukemia virus.

30 **[00109]** Where a cDNA insert is employed, one will typically desire to include a polyadenylation signal to effect proper polyadenylation of the gene transcript. Any

polyadenylation sequence may be employed such as human growth hormone and SV40 polyadenylation signals. Also contemplated as an element of the expression cassette is a terminator. These elements can serve to enhance message levels and to minimize read through from the cassette into other sequences.

5 **[00110]** There are a number of ways in which expression vectors may be introduced into cells. In certain embodiments, the expression construct comprises a virus or engineered construct derived from a viral genome. The ability of certain viruses to enter cells via receptor-mediated endocytosis, to integrate into host cell genome and express viral genes stably and efficiently have made them attractive candidates for the transfer of foreign genes
10 into mammalian cells. These have a relatively low capacity for foreign DNA sequences and have a restricted host spectrum. Furthermore, their oncogenic potential and cytopathic effects in permissive cells raise safety concerns. They can accommodate only up to 8 kB of foreign genetic material but can be readily introduced in a variety of cell lines and laboratory animals.

15 **[00111]** One method for *in vivo* delivery involves the use of an adenovirus expression vector. "Adenovirus expression vector" is meant to include those constructs containing adenovirus sequences sufficient to (a) support packaging of the construct and (b) to express a chimeric hMPV/RSV F protein that has been cloned therein. In this context, expression does not require that the gene product be synthesized.

20 **[00112]** The expression vector comprises a genetically engineered form of adenovirus. Knowledge of the genetic organization of adenovirus, a 36 kB, linear, double-stranded DNA virus, allows substitution of large pieces of adenoviral DNA with foreign sequences up to 7 kB. In contrast to retrovirus, the adenoviral infection of host cells does not result in chromosomal integration because adenoviral DNA can replicate in an episomal
25 manner without potential genotoxicity. Also, adenoviruses are structurally stable, and no genome rearrangement has been detected after extensive amplification. Adenovirus can infect virtually all epithelial cells regardless of their cell cycle stage. So far, adenoviral infection appears to be linked only to mild disease such as acute respiratory disease in humans.

[00113] Adenovirus is particularly suitable for use as a gene transfer vector
30 because of its mid-sized genome, ease of manipulation, high titer, wide target cell range and high infectivity. Both ends of the viral genome contain 100-200 base pair inverted repeats

(ITRs), which are *cis* elements necessary for viral DNA replication and packaging. The early (E) and late (L) regions of the genome contain different transcription units that are divided by the onset of viral DNA replication. The E1 region (E1A and E1B) encodes proteins responsible for the regulation of transcription of the viral genome and a few cellular genes.

5 The expression of the E2 region (E2A and E2B) results in the synthesis of the proteins for viral DNA replication. These proteins are involved in DNA replication, late gene expression and host cell shut-off. The products of the late genes, including the majority of the viral capsid proteins, are expressed only after significant processing of a single primary transcript issued by the major late promoter (MLP). The MLP, (located at 16.8 m.u.) is particularly
10 efficient during the late phase of infection, and all the mRNAs issued from this promoter possess a 5'-tripartite leader (TPL) sequence which makes them preferred mRNAs for translation. In one system, recombinant adenovirus is generated from homologous recombination between shuttle vector and provirus vector. Due to the possible recombination between two proviral vectors, wild-type adenovirus may be generated from this process.
15 Therefore, it is critical to isolate a single clone of virus from an individual plaque and examine its genomic structure.

[00114] Generation and propagation of the current adenovirus vectors, which are replication deficient, depend on a unique helper cell line, designated 293, which was transformed from human embryonic kidney cells by Ad5 DNA fragments and constitutively
20 expresses E1 proteins. Since the E3 region is dispensable from the adenovirus genome, the current adenovirus vectors, with the help of 293 cells, carry foreign DNA in either the E1, the D3 or both regions. In nature, adenovirus can package approximately 105% of the wild-type genome, providing capacity for about 2 extra kb of DNA. Combined with the approximately 5.5 kb of DNA that is replaceable in the E1 and E3 regions, the maximum capacity of the
25 current adenovirus vector is under 7.5 kb, or about 15% of the total length of the vector. More than 80% of the adenovirus viral genome remains in the vector backbone and is the source of vector-borne cytotoxicity. Also, the replication deficiency of the E1-deleted virus is incomplete.

[00115] Helper cell lines may be derived from human cells such as human
30 embryonic kidney cells, muscle cells, hematopoietic cells or other human embryonic mesenchymal or epithelial cells. Alternatively, the helper cells may be derived from the cells of other mammalian species that are permissive for human adenovirus. Such cells include,

e.g., Vero cells or other monkey embryonic mesenchymal or epithelial cells. As stated above, the preferred helper cell line is 293.

[00116] The adenoviruses of the disclosure are replication defective, or at least conditionally replication defective. The adenovirus may be of any of the 42 different known serotypes or subgroups A-F. Adenovirus type 5 of subgroup C is one exemplary starting material that may be used to obtain the conditional replication-defective adenovirus vector for use in the present disclosure.

[00117] Other viral vectors may be employed as expression constructs in the present disclosure. Vectors derived from viruses such as vaccinia virus, adeno-associated virus (AAV) and herpesviruses may be employed. They offer several attractive features for various mammalian cells.

[00118] In embodiments, particular embodiments, the vector is an AAV vector. AAV is a small virus that infects humans and some other primate species. AAV is not currently known to cause disease. The virus causes a very mild immune response, lending further support to its apparent lack of pathogenicity. In many cases, AAV vectors integrate into the host cell genome, which can be important for certain applications, but can also have unwanted consequences. Gene therapy vectors using AAV can infect both dividing and quiescent cells and persist in an extrachromosomal state without integrating into the genome of the host cell, although in the native virus some integration of virally carried genes into the host genome does occur. These features make AAV a very attractive candidate for creating viral vectors for gene therapy, and for the creation of isogenic human disease models. Recent human clinical trials using AAV for gene therapy in the retina have shown promise. AAV belongs to the genus *Dependoparvovirus*, which in turn belongs to the family *Parvoviridae*. The virus is a small (20 nm) replication-defective, nonenveloped virus.

[00119] Wild-type AAV has attracted considerable interest from gene therapy researchers due to a number of features. Chief amongst these is the virus's apparent lack of pathogenicity. It can also infect non-dividing cells and has the ability to stably integrate into the host cell genome at a specific site (designated AAVS1) in the human chromosome 19. This feature makes it somewhat more predictable than retroviruses, which present the threat of a random insertion and of mutagenesis, which is sometimes followed by development of a cancer. The AAV genome integrates most frequently into the site mentioned, while random

incorporations into the genome take place with a negligible frequency. Development of AAVs as gene therapy vectors, however, has eliminated this integrative capacity by removal of the *rep* and *cap* from the DNA of the vector. The desired gene together with a promoter to drive transcription of the gene is inserted between the inverted terminal repeats (ITR) that aid in concatemer formation in the nucleus after the single-stranded vector DNA is converted by host cell DNA polymerase complexes into double-stranded DNA. AAV-based gene therapy vectors form episomal concatemers in the host cell nucleus. In non-dividing cells, these concatemers remain intact for the life of the host cell. In dividing cells, AAV DNA is lost through cell division, since the episomal DNA is not replicated along with the host cell DNA. Random integration of AAV DNA into the host genome is detectable but occurs at very low frequency. AAVs also present very low immunogenicity, seemingly restricted to generation of neutralizing antibodies, while they induce no clearly defined cytotoxic response. This feature, along with the ability to infect quiescent cells present their dominance over adenoviruses as vectors for human gene therapy.

[00120] The AAV genome is built of single-stranded deoxyribonucleic acid (ssDNA), either positive- or negative-sensed, which is about 4.7 kilobase long. The genome comprises inverted terminal repeats (ITRs) at both ends of the DNA strand, and two open reading frames (ORFs): *rep* and *cap*. The former is composed of four overlapping genes encoding Rep proteins required for the AAV life cycle, and the latter contains overlapping nucleotide sequences of capsid proteins: VP1, VP2 and VP3, which interact together to form a capsid of an icosahedral symmetry.

[00121] The Inverted Terminal Repeat (ITR) sequences comprise 145 bases each. They were named so because of their symmetry, which was shown to be required for efficient multiplication of the AAV genome. The feature of these sequences that gives them this property is their ability to form a hairpin, which contributes to so-called self-priming that allows primase-independent synthesis of the second DNA strand. The ITRs were also shown to be required for both integration of the AAV DNA into the host cell genome (19th chromosome in humans) and rescue from it, as well as for efficient encapsidation of the AAV DNA combined with generation of a fully assembled, deoxyribonuclease-resistant AAV particles.

[00122] With regard to gene therapy, ITRs seem to be the only sequences required *in cis* next to the therapeutic gene: structural (*cap*) and packaging (*rep*) proteins can

be delivered *in trans*. With this assumption many methods were established for efficient production of recombinant AAV (rAAV) vectors containing a reporter or therapeutic gene. However, it was also published that the ITRs are not the only elements required *in cis* for the effective replication and encapsidation. A few research groups have identified a sequence
5 designated *cis-acting Rep-dependent element* (CARE) inside the coding sequence of the *rep* gene. CARE was shown to augment the replication and encapsidation when present *in cis*.

[00123] In some aspects, the present disclosure provides pharmaceutical compositions that contain one or more salts. The salts may be an inorganic potassium or sodium salt such as potassium chloride, sodium chloride, potassium phosphate dibasic,
10 potassium phosphate monobasic, sodium phosphate dibasic, or sodium phosphate monobasic. The pharmaceutical composition may comprise one or more phosphate salts such to generate a phosphate buffer solution. The phosphate buffer solution may be comprise each of the phosphates to buffer a solution to a pH from about 6.8, 6.9, 7.0, 7.1, 7.2, 7.3, 7.4, 7.5, 7.6, 7.7, 7.8, 7.9, or 8.0, or any range derivable therein.

15 [00124] In some aspects, the present disclosure comprises one or more excipients formulated into pharmaceutical compositions. An “excipient” refers to pharmaceutically acceptable carriers that are relatively inert substances used to facilitate administration or delivery of an API into a subject or used to facilitate processing of an API into drug formulations that can be used pharmaceutically for delivery to the site of action in a
20 subject. Furthermore, these compounds may be used as diluents in order to obtain a dosage that can be readily measured or administered to a patient. Non-limiting examples of excipients include polymers, stabilizing agents, surfactants, surface modifiers, solubility enhancers, buffers, encapsulating agents, antioxidants, preservatives, nonionic wetting or clarifying agents, viscosity increasing agents, and absorption-enhancing agents.

25 [00125] In a specific embodiment, the term “pharmaceutically acceptable” means approved by a regulatory agency of the Federal or a state government or listed in the U.S. Pharmacopeia or other generally recognized pharmacopeia for use in animals, and more particularly in humans. The term “carrier” refers to a diluent, excipient, or vehicle with which the therapeutic is administered. Such pharmaceutical carriers can be sterile liquids, such as
30 water and can preferably include an adjuvant. Water is a particular carrier when the pharmaceutical composition is administered by injections, such an intramuscular injection. Saline solutions and aqueous dextrose and glycerol solutions can also be employed as liquid

carriers, particularly for injectable solutions. Other suitable pharmaceutical excipients include starch, glucose, lactose, sucrose, gelatin, malt, rice, flour, chalk, silica gel, sodium stearate, glycerol monostearate, talc, sodium chloride, dried skim milk, glycerol, propylene, glycol, water, ethanol and the like.

5 **[00126]** The composition, if desired, can also contain minor amounts of wetting or emulsifying agents, or pH buffering agents. These compositions can take the form of solutions, suspensions, emulsion, tablets, pills, capsules, powders, sustained-release formulations and the like. Oral formulations can include standard carriers such as pharmaceutical grades of mannitol, lactose, starch, magnesium stearate, sodium saccharine, 10 cellulose, magnesium carbonate, etc. Examples of suitable pharmaceutical agents are described in “Remington’s Pharmaceutical Sciences.” Such compositions will contain a prophylactically or therapeutically effective amount of the antibody or fragment thereof, preferably in purified form, together with a suitable amount of carrier so as to provide the form for proper administration to the patient. The formulation should suit the mode of 15 administration, which can be oral, intravenous, intraarterial, intrabuccal, intranasal, nebulized, bronchial inhalation, or delivered by mechanical ventilation.

[00127] Chimeric proteins of the present disclosure, as described herein, can be formulated for parenteral administration, e.g., formulated for injection via the intradermal, intravenous, intramuscular, subcutaneous, intra-tumoral or even intraperitoneal routes. The 20 antibodies could alternatively be administered by a topical route directly to the mucosa, for example by nasal drops, inhalation, or by nebulizer. Pharmaceutically acceptable salts include the acid salts and those which are formed with inorganic acids such as, for example, hydrochloric or phosphoric acids, or such organic acids as acetic, oxalic, tartaric, mandelic, and the like. Salts formed with the free carboxyl groups may also be derived from inorganic 25 bases such as, for example, sodium, potassium, ammonium, calcium, or ferric hydroxides, and such organic bases as isopropylamine, trimethylamine, 2-ethylamino ethanol, histidine, procaine, and the like.

[00128] Generally, the ingredients of compositions of the disclosure are supplied either separately or mixed together in unit dosage form, for example, as a dry 30 lyophilized powder or water-free concentrate in a hermetically sealed container such as an ampoule or sachette indicating the quantity of active agent. Where the composition is to be administered by infusion, it can be dispensed with an infusion bottle containing sterile

pharmaceutical grade water or saline. Where the composition is administered by injection, an ampoule of sterile water for injection or saline can be provided so that the ingredients may be mixed prior to administration.

5 **[00129]** The compositions of the disclosure can be formulated as neutral or salt forms. Pharmaceutically acceptable salts include those formed with anions such as those derived from hydrochloric, phosphoric, acetic, oxalic, tartaric acids, *etc.*, and those formed with cations such as those derived from sodium, potassium, ammonium, calcium, ferric hydroxides, isopropylamine, triethylamine, 2-ethylamino ethanol, histidine, procaine, *etc.*

10 **[00130]** Dosage can be by a single dose schedule or a multiple dose schedule. Multiple doses may be used in a primary immunization schedule and/or in a booster immunization schedule. In a multiple dose schedule the various doses may be given by the same or different routes. Multiple doses will typically be administered at least 1 week apart (e.g., about 2 weeks, about 3 weeks, about 4 weeks, about 6 weeks, about 8 weeks, about 10 weeks, about 12 weeks, about 16 weeks, *etc.*).

15 **[00131]** The compositions disclosed herein may be used to treat both children and adults. Thus, a human subject may be less than 1 year old, 1-5 years old, 5-16 years old, 16-55 years old, 55-65 years old, or at least 65 years old.

20 **[00132]** Preferred routes of administration include, but are not limited to, intramuscular, intraperitoneal, intradermal, subcutaneous, intravenous, intraarterial, and intraocular injection. Particularly preferred routes of administration include intramuscular, intradermal and subcutaneous injection.

IV. Immunodetection Methods

25 **[00133]** In still further embodiments, the present disclosure concerns immunodetection methods for binding, purifying, removing, quantifying and otherwise generally detecting an hMPV F protein-, an RSV F protein-, or a chimeric hMPV/RSV F protein-binding antibodies. While such methods can be applied in a traditional sense, another use will be in quality control and monitoring of vaccine stocks, where antibodies can be used to assess the amount or integrity (i.e., long term stability) of antigens. Alternatively, the methods may be used to screen various antibodies for appropriate/desired reactivity profiles.

[00134] Some immunodetection methods include enzyme linked immunosorbent assay (ELISA), radioimmunoassay (RIA), immunoradiometric assay, fluoroimmunoassay, chemiluminescent assay, bioluminescent assay, and Western blot to mention a few. In particular, a competitive assay for the detection and quantitation of hMPV/RSV and/or hMPV and/or RSV F protein-binding antibodies also is provided. In general, the immunobinding methods include obtaining a sample suspected of containing hMPV/RSV and/or hMPV and/or RSV F protein-binding antibodies, and contacting the sample with an antigen in accordance with the present disclosure, as the case may be, under conditions effective to allow the formation of immunocomplexes.

[00135] These methods include methods for detecting or purifying hMPV/RSV and/or hMPV and/or RSV F protein-binding antibodies or hMPV/RSV and/or hMPV and/or RSV F protein from a sample. The antigen will preferably be linked to a solid support, such as in the form of a column matrix, and the sample suspected of containing the hMPV/RSV and/or hMPV and/or RSV F protein-binding antibodies will be applied to the immobilized antigen. The unwanted components will be washed from the column, leaving the hMPV/RSV and/or hMPV and/or RSV F protein-binding antibodies immunocomplexed to the immobilized antigen, which is then collected by removing the antigen from the column.

[00136] The immunobinding methods also include methods for detecting and quantifying the amount of hMPV/RSV and/or hMPV and/or RSV F protein or related components in a sample and the detection and quantification of any immune complexes formed during the binding process. Here, one would obtain a sample suspected of containing hMPV/RSV and/or hMPV and/or RSV F protein and contact the sample with an antibody that binds hMPV/RSV and/or hMPV and/or RSV F protein or components thereof, followed by detecting and quantifying the amount of immune complexes formed under the specific conditions. In terms of antigen detection, the biological sample analyzed may be any sample that is suspected of containing hMPV/RSV and/or hMPV and/or RSV F protein, such as a tissue section or specimen, a homogenized tissue extract, a biological fluid (e.g., a nasal swab), including blood and serum, or a secretion, such as feces or urine.

[00137] Contacting the chosen biological sample with the antigen under effective conditions and for a period of time sufficient to allow the formation of immune complexes (primary immune complexes) is generally a matter of simply adding the antigen composition to the sample and incubating the mixture for a period of time long enough for

the antigen to form immune complexes with, i.e., to bind to hMPV/RSV and/or hMPV and/or RSV F protein-binding antibodies. After this time, the sample-antibody composition, such as a tissue section, ELISA plate, dot blot or Western blot, will generally be washed to remove any non-specifically bound antibody species, allowing only those antibodies specifically
5 bound within the primary immune complexes to be detected.

[00138] In general, the detection of immunocomplex formation is well known in the art and may be achieved through the application of numerous approaches. These methods are generally based upon the detection of a label or marker, such as any of those radioactive, fluorescent, biological and enzymatic tags. Patents concerning the use of such
10 labels include U.S. Patents 3,817,837, 3,850,752, 3,939,350, 3,996,345, 4,277,437, 4,275,149 and 4,366,241. Of course, one may find additional advantages through the use of a secondary binding ligand such as a second antibody and/or a biotin/avidin ligand binding arrangement, as is known in the art.

[00139] An antibody employed in the detection may itself be linked to a
15 detectable label, wherein one would then simply detect this label, thereby allowing the amount of the primary immune complexes in the composition to be determined. Alternatively, the first antibody that becomes bound within the primary immune complexes may be detected by means of a second binding ligand that has binding affinity for the antibody. In these cases, the second binding ligand may be linked to a detectable label. The
20 second binding ligand is itself often an antibody, which may thus be termed a “secondary” antibody. The primary immune complexes are contacted with the labeled, secondary binding ligand, or antibody, under effective conditions and for a period of time sufficient to allow the formation of secondary immune complexes. The secondary immune complexes are then generally washed to remove any non-specifically bound labeled secondary antibodies or
25 ligands, and the remaining label in the secondary immune complexes is then detected.

[00140] Further methods include the detection of primary immune complexes by a two-step approach. A second binding ligand, such as an antibody that has binding affinity for the antibody, is used to form secondary immune complexes, as described above. After washing, the secondary immune complexes are contacted with a third binding ligand or
30 antibody that has binding affinity for the second antibody, again under effective conditions and for a period of time sufficient to allow the formation of immune complexes (tertiary immune complexes). The third ligand or antibody is linked to a detectable label, allowing

detection of the tertiary immune complexes thus formed. This system may provide for signal amplification if this is desired.

[00141] One method of immunodetection uses two different antibodies. A first biotinylated antibody is used to detect the target antigen, and a second antibody is then used to detect the biotin attached to the complexed biotin. In that method, the sample to be tested is first incubated in a solution containing the first step antibody. If the target antigen is present, some of the antibody binds to the antigen to form a biotinylated antibody/antigen complex. The antibody/antigen complex is then amplified by incubation in successive solutions of streptavidin (or avidin), biotinylated DNA, and/or complementary biotinylated DNA, with each step adding additional biotin sites to the antibody/antigen complex. The amplification steps are repeated until a suitable level of amplification is achieved, at which point the sample is incubated in a solution containing the second step antibody against biotin. This second step antibody is labeled, as for example with an enzyme that can be used to detect the presence of the antibody/antigen complex by histoenzymology using a chromogen substrate. With suitable amplification, a conjugate can be produced which is macroscopically visible.

[00142] Another known method of immunodetection takes advantage of the immuno-PCR (Polymerase Chain Reaction) methodology. The PCR method is similar to the Cantor method up to the incubation with biotinylated DNA, however, instead of using multiple rounds of streptavidin and biotinylated DNA incubation, the DNA/biotin/streptavidin/antibody complex is washed out with a low pH or high salt buffer that releases the antibody. The resulting wash solution is then used to carry out a PCR reaction with suitable primers with appropriate controls. At least in theory, the enormous amplification capability and specificity of PCR can be utilized to detect a single antigen molecule.

1. ELISAs

[00143] Immunoassays, in their most simple and direct sense, are binding assays. Certain preferred immunoassays are the various types of enzyme linked immunosorbent assays (ELISAs) and radioimmunoassays (RIA) known in the art. Immunohistochemical detection using tissue sections is also particularly useful. However, it will be readily appreciated that detection is not limited to such techniques, and western blotting, dot blotting, FACS analyses, and the like may also be used.

[00144] In one exemplary ELISA, the antibodies of the disclosure are immobilized onto a selected surface exhibiting protein affinity, such as a well in a polystyrene microtiter plate. Then, a test composition suspected of containing the hMPV/RSV and/or hMPV and/or RSV F protein is added to the wells. After binding and washing to remove non-specifically bound immune complexes, the bound antigen may be detected. Detection may be achieved by the addition of another anti-hMPV/RSV and/or hMPV and/or RSV F protein antibody that is linked to a detectable label. This type of ELISA is a simple “sandwich ELISA.” Detection may also be achieved by the addition of a second anti-hMPV/RSV and/or hMPV and/or RSV F protein antibody, followed by the addition of a third antibody that has binding affinity for the second antibody, with the third antibody being linked to a detectable label.

[00145] In another exemplary ELISA, the samples suspected of containing the hMPV/RSV and/or hMPV and/or RSV F protein (e.g., potentially infected cells) are immobilized onto the well surface and then contacted with the anti-hMPV/RSV and/or hMPV and/or RSV F protein antibodies of the disclosure. After binding and washing to remove non-specifically bound immune complexes, the bound anti-hMPV/RSV and/or hMPV and/or RSV F protein antibodies are detected. Where the initial anti-hMPV/RSV and/or hMPV and/or RSV F protein antibodies are linked to a detectable label, the immune complexes may be detected directly. Again, the immune complexes may be detected using a second antibody that has binding affinity for the first anti-hMPV/RSV and/or hMPV and/or RSV F protein antibody, with the second antibody being linked to a detectable label.

[00146] Irrespective of the format employed, ELISAs have certain features in common, such as coating, incubating and binding, washing to remove non-specifically bound species, and detecting the bound immune complexes. These are described below.

[00147] In coating a plate with either antigen or antibody, one will generally incubate the wells of the plate with a solution of the antigen or antibody, either overnight or for a specified period of hours. The wells of the plate will then be washed to remove incompletely adsorbed material. Any remaining available surfaces of the wells are then “coated” with a nonspecific protein that is antigenically neutral with regard to the test antisera. These include bovine serum albumin (BSA), casein or solutions of milk powder. The coating allows for blocking of nonspecific adsorption sites on the immobilizing surface and thus reduces the background caused by nonspecific binding of antisera onto the surface.

[00148] In ELISAs, it is probably more customary to use a secondary or tertiary detection means rather than a direct procedure. Thus, after binding of a protein or antibody to the well, coating with a non-reactive material to reduce background, and washing to remove unbound material, the immobilizing surface is contacted with the biological sample to be tested under conditions effective to allow immune complex (antigen/antibody) formation. Detection of the immune complex then requires a labeled secondary binding ligand or antibody, and a secondary binding ligand or antibody in conjunction with a labeled tertiary antibody or a third binding ligand.

[00149] “Under conditions effective to allow immune complex (antigen/antibody) formation” means that the conditions preferably include diluting the antigens and/or antibodies with solutions such as BSA, bovine gamma globulin (BGG) or phosphate buffered saline (PBS)/Tween. These added agents also tend to assist in the reduction of nonspecific background.

[00150] The “suitable” conditions also mean that the incubation is at a temperature or for a period of time sufficient to allow effective binding. Incubation steps are typically from about 1 to 2 to 4 hours or so, at temperatures preferably on the order of 25°C to 27°C, or may be overnight at about 4°C or so.

[00151] Following all incubation steps in an ELISA, the contacted surface is washed so as to remove non-complexed material. A preferred washing procedure includes washing with a solution such as PBS/Tween, or borate buffer. Following the formation of specific immune complexes between the test sample and the originally bound material, and subsequent washing, the occurrence of even minute amounts of immune complexes may be determined.

[00152] To provide a detecting means, the second or third antibody will have an associated label to allow detection. Preferably, this will be an enzyme that will generate color development upon incubating with an appropriate chromogenic substrate. Thus, for example, one will desire to contact or incubate the first and second immune complex with a urease, glucose oxidase, alkaline phosphatase or hydrogen peroxidase-conjugated antibody for a period of time and under conditions that favor the development of further immune complex formation (e.g., incubation for 2 hours at room temperature in a PBS-containing solution such as PBS-Tween).

[00153] After incubation with the labeled antibody, and subsequent to washing to remove unbound material, the amount of label is quantified, e.g., by incubation with a chromogenic substrate such as urea, or bromocresol purple, or 2,2'-azino-di-(3-ethylbenzthiazoline-6-sulfonic acid (ABTS), or H₂O₂, in the case of peroxidase as the enzyme label. Quantification is then achieved by measuring the degree of color generated, e.g., using a visible spectra spectrophotometer.

2. Western Blot

[00154] The Western blot (alternatively, protein immunoblot) is an analytical technique used to detect specific proteins in a given sample of tissue homogenate or extract. It uses gel electrophoresis to separate native or denatured proteins by the length of the polypeptide (denaturing conditions) or by the 3-D structure of the protein (native/ non-denaturing conditions). The proteins are then transferred to a membrane (typically nitrocellulose or PVDF), where they are probed (detected) using antibodies specific to the target protein.

[00155] Samples may be taken from whole tissue or from cell culture. In most cases, solid tissues are first broken down mechanically using a blender (for larger sample volumes), using a homogenizer (smaller volumes), or by sonication. Cells may also be broken open by one of the above mechanical methods. Assorted detergents, salts, and buffers may be employed to encourage lysis of cells and to solubilize proteins. Protease and phosphatase inhibitors are often added to prevent the digestion of the sample by its own enzymes. Tissue preparation is often done at cold temperatures to avoid protein denaturing.

[00156] The proteins of the sample are separated using gel electrophoresis. Separation of proteins may be by isoelectric point (pI), molecular weight, electric charge, or a combination of these factors. The nature of the separation depends on the treatment of the sample and the nature of the gel. This is a very useful way to determine a protein. It is also possible to use a two-dimensional (2-D) gel which spreads the proteins from a single sample out in two dimensions. Proteins are separated according to isoelectric point (pH at which they have neutral net charge) in the first dimension, and according to their molecular weight in the second dimension.

[00157] In order to make the proteins accessible to antibody detection, they are moved from within the gel onto a membrane made of nitrocellulose or polyvinylidene

difluoride (PVDF). The membrane is placed on top of the gel, and a stack of filter papers placed on top of that. The entire stack is placed in a buffer solution which moves up the paper by capillary action, bringing the proteins with it. Another method for transferring the proteins is called electroblotting and uses an electric current to pull proteins from the gel into the PVDF or nitrocellulose membrane. The proteins move from within the gel onto the membrane while maintaining the organization they had within the gel. As a result of this blotting process, the proteins are exposed on a thin surface layer for detection (see below). Both varieties of membrane are chosen for their non-specific protein binding properties (i.e., binds all proteins equally well). Protein binding is based upon hydrophobic interactions, as well as charged interactions between the membrane and protein. Nitrocellulose membranes are cheaper than PVDF, but are far more fragile and do not stand up well to repeated probings. The uniformity and overall effectiveness of transfer of protein from the gel to the membrane can be checked by staining the membrane with Coomassie Brilliant Blue or Ponceau S dyes. Once transferred, proteins are detected using labeled primary antibodies, or unlabeled primary antibodies followed by indirect detection using labeled protein A or secondary labeled antibodies binding to the Fc region of the primary antibodies.

3. Immunohistochemistry

[00158] The antibodies of the present disclosure may also be used in conjunction with both fresh-frozen and/or formalin-fixed, paraffin-embedded tissue blocks prepared for study by immunohistochemistry (IHC). The method of preparing tissue blocks from these particulate specimens has been successfully used in previous IHC studies of various prognostic factors, and is well known to those of skill in the art (Brown et al., 1990; Abbondanzo et al., 1990; Allred et al., 1990).

[00159] Briefly, frozen-sections may be prepared by rehydrating 50 ng of frozen “pulverized” tissue at room temperature in phosphate buffered saline (PBS) in small plastic capsules; pelleting the particles by centrifugation; resuspending them in a viscous embedding medium (OCT); inverting the capsule and/or pelleting again by centrifugation; snap-freezing in -70°C isopentane; cutting the plastic capsule and/or removing the frozen cylinder of tissue; securing the tissue cylinder on a cryostat microtome chuck; and/or cutting 25-50 serial sections from the capsule. Alternatively, whole frozen tissue samples may be used for serial section cuttings.

[00160] Permanent-sections may be prepared by a similar method involving rehydration of the 50 mg sample in a plastic microfuge tube; pelleting; resuspending in 10% formalin for 4 hours fixation; washing/pelleting; resuspending in warm 2.5% agar; pelleting; cooling in ice water to harden the agar; removing the tissue/agar block from the tube; infiltrating and/or embedding the block in paraffin; and/or cutting up to 50 serial permanent sections. Again, whole tissue samples may be substituted.

4. Immunodetection Kits

[00161] In still further embodiments, the present disclosure concerns immunodetection kits for use with the immunodetection methods described above. As antibodies may be used to detect hMPV/RSV and/or hMPV and/or RSV F protein, antibodies may be included in the kit. The immunodetection kits will thus comprise, in suitable container means, a first antibody that binds to an hMPV/RSV and/or hMPV and/or RSV F protein, and optionally an immunodetection reagent. Alternatively, the hMPV/RSV and/or hMPV and/or RSV F protein antigen may be used to detect hMPV/RSV and/or hMPV and/or RSV F protein-binding antibodies. In this case, the immunodetection kits will thus comprise, in suitable container means, an hMPV/RSV and/or hMPV and/or RSV F protein antigen.

[00162] In certain embodiments, the antibody or antigen may be pre-bound to a solid support, such as a column matrix and/or well of a microtitre plate. The immunodetection reagents of the kit may take any one of a variety of forms, including those detectable labels that are associated with or linked to an antibody. Detectable labels that are associated with or attached to a secondary binding ligand are also contemplated. Exemplary secondary ligands are those secondary antibodies that have binding affinity for the first antibody.

[00163] Further suitable immunodetection reagents for use in the present kits include the two-component reagent that comprises a secondary antibody that has binding affinity for the first antibody, along with a third antibody that has binding affinity for the second antibody, the third antibody being linked to a detectable label. As noted above, a number of exemplary labels are known in the art and all such labels may be employed in connection with the present disclosure.

[00164] The kits may further comprise a suitably aliquoted composition of hMPV/RSV and/or hMPV and/or RSV F protein, whether labeled or unlabeled, as may be

used to prepare a standard curve for a detection assay. The kits may contain antibody-label conjugates either in fully conjugated form, in the form of intermediates, or as separate moieties to be conjugated by the user of the kit. The components of the kits may be packaged either in aqueous media or in lyophilized form.

5 **[00165]** The container means of the kits will generally include at least one vial, test tube, flask, bottle, syringe or other container means, into which the antibody may be placed, or preferably, suitably aliquoted. The kits of the present disclosure will also typically include a means for containing the antibody, antigen, and any other reagent containers in close confinement for commercial sale. Such containers may include injection or blow-
10 molded plastic containers into which the desired vials are retained.

5. **Flow Cytometry and FACS**

[00166] The antibodies of the present disclosure may also be used in flow cytometry or FACS. Flow cytometry is a laser- or impedance-based technology employed in many detection assays, including cell counting, cell sorting, biomarker detection and protein
15 engineering. The technology suspends cells in a stream of fluid and passing them through an electronic detection apparatus, which allows simultaneous multiparametric analysis of the physical and chemical characteristics of up to thousands of particles per second. Flow cytometry is routinely used in the diagnosis of disorders, but has many other applications in basic research, clinical practice and clinical trials.

20 **[00167]** Fluorescence-activated cell sorting (FACS) is a specialized type of cytometry. It provides a method for sorting a heterogenous mixture of biological cells into two or more containers, one cell at a time, based on the specific light scattering and fluorescent characteristics of each cell. In general, the technology involves a cell suspension entrained in the center of a narrow, rapidly flowing stream of liquid. The flow is arranged so
25 that there is a large separation between cells relative to their diameter. A vibrating mechanism causes the stream of cells to break into individual droplets. Just before the stream breaks into droplets, the flow passes through a fluorescence measuring station where the fluorescence of each cell is measured. An electrical charging ring is placed just at the point where the stream breaks into droplets. A charge is placed on the ring based immediately prior
30 to fluorescence intensity being measured, and the opposite charge is trapped on the droplet as it breaks form the stream. The charged droplets then fall through an electrostatic deflection system that diverts droplets into containers based upon their charge.

[00168] In certain embodiments, to be used in flow cytometry or FACS, the antibodies of the present disclosure are labeled with fluorophores and then allowed to bind to the cells of interest, which are analyzed in a flow cytometer or sorted by a FACS machine.

5 V. Examples

[00169] The following examples are included to demonstrate preferred embodiments of the invention. It should be appreciated by those of skill in the art that the techniques disclosed in the examples which follow represent techniques discovered by the inventor to function well in the practice of the invention, and thus can be considered to
10 constitute preferred modes for its practice. However, those of skill in the art should, in light of the present disclosure, appreciate that many changes can be made in the specific embodiments which are disclosed and still obtain a like or similar result without departing from the spirit and scope of the invention.

Example 1 – Chimeric hMPV/RSV F Proteins

[00170] Four different designs for the junctions were used to generate four
15 different chimera constructs. The residues within the antigenic site Ø (residues 50-84) in the F2 subunit of hMPV F (see SEQ ID NO: 5) were replaced with residues 59-96 from RSV F (see SEQ ID NO: 10) in all chimeras. For chimera 1, the residues within the antigenic site Ø (residues 163-199) in F1 subunit of hMPV F (see SEQ ID NO: 5) were replaced with the
20 residues 193-229 from RSV F (see SEQ ID NO: 10) (FIG. 1). For chimera 2, residues 165-201 from hMPV F were replaced with residues 195-231 from RSV F. For chimera 3, residues 163-201 from hMPV F were replaced with residues 193-231 from RSV F. For chimera 4, residues 165-199 from hMPV F were replaced with residues 195-229 from RSV F. All chimeras contained two substitutions (N67I, S215P, positions relative to SEQ ID NO: 10)
25 from RSV F and three substitutions (T127C/N153C, V231I, T365C/V463C, positions relative to SEQ ID NO: 5) from hMPV F.

[00171] The protein expression and monodispersity of chimera 1-4 were evaluated by SDS-PAGE and size exclusion column (SEC) (FIG. 2, left panel). The integrity of prefusion-specific epitopes on chimeras was also characterized by binding to RSV F
30 prefusion-specific antibody D25 and hMPV/RSV F cross reactive antibody MPE8 (FIG. 2,

right panel). Based on the protein expression and the binding affinity to D25 and MPE8, chimera 1 was selected for further engineering.

[00172] For the first iteration of chimera optimization, Thr91 or Leu95 from RSV F were individually substituted with Arg on the backbone of chimera 1, aiming to form an inter-subunit salt bridge with Asp224 from hMPV F (see FIG. 4, left panel). The L95R substitution not only increased the protein expression relative to chimera 1, but also maintained the integrity of prefusion epitopes from RSV F and hMPV F (FIG. 3).

[00173] For the second iteration of chimera optimization (FIG. 4), an inter-protomer salt bridge (L219K), a charge repulsion reduction design (E453Q), a proline substitution (A107P) or a disulfide bond design (L110C/N322C) were individually added on the parental construct chimera1_L95R. The L219K, E453Q, and A107P substitutions all increased the protein expression relative to the parental construct (FIG. 4, top right panel). Notably, only the L110C/N322C substitution substantially enhanced the thermostability (FIG. 4, bottom right panel).

[00174] Finally, all ten beneficial substitutions were combined to generate DS-CavEsP. It had the highest protein expression (FIG. 5, left panel), the highest thermostability, and retained binding to RSV F-specific antibody D25 and hMPV F cross-reactive antibody MPE8. From nsEM analysis, DS-CavEsP exhibits a trimeric, prefusion conformation bound by both D25 and MPE8 Fab (FIG. 5, right panel). The affinity of DS-CavEsP to RSV F-specific antibody D25 was compared to that of a prefusion RSV F construct (PRDM) (FIGS. 6A-6B). The affinity of DS-CavEsP to prefusion hMPVF specific antibody MPE8 (Wen et al., 2017) was compared to that of a prefusion stabilized hMPVF construct (DS-Cav-ES2) (FIGS. 6C-6D). The affinity of DS-CavEsP to neutralizing antibodies that bind to site III (II), V, or I (VI) of hMPVF was determined to be comparable to that of a prefusion stabilized hMPVF construct (DS-Cav-ES2). A Cryo-EM structure of D25 and MPE8 complexed with DS-CavEsP was determined at a resolution of 3.2 Å (FIG. 7A). The binding interfaces of each antibody and DS-CavEsP were also resolved (FIGS. 7B-7C).

* * *

[00175] All of the methods disclosed and claimed herein can be made and executed without undue experimentation in light of the present disclosure. While the compositions and methods of this invention have been described in terms of preferred embodiments, it will be

apparent to those of skill in the art that variations may be applied to the methods and in the steps or in the sequence of steps of the method described herein without departing from the concept, spirit and scope of the invention. More specifically, it will be apparent that certain agents which are both chemically and physiologically related may be substituted for the
5 agents described herein while the same or similar results would be achieved. All such similar substitutes and modifications apparent to those skilled in the art are deemed to be within the spirit, scope and concept of the invention as defined by the appended claims.

WHAT IS CLAIMED IS:

1. An engineered human metapneumovirus (hMPV)–respiratory syncytial virus (RSV) chimeric F protein comprising an ectodomain having the following segments, in order from N-terminus to C-terminus:
 - (1) a sequence having at least 90% identity to amino acids 19-49 of any one of SEQ ID NOs: 5-8;
 - (2) a sequence having at least 90% identity to amino acids 59-96 of SEQ ID NO: 10;
 - (3) a sequence having at least 90% identity to amino acids 85-162 or 85-164 of any one of SEQ ID NOs: 5-8;
 - (4) a sequence having at least 90% identity to amino acids 193-229, 193-231, 195-229, or 195-231 of SEQ ID NO: 10; and
 - (5) a sequence having at least 90% identity to amino acids 200-489 or 202-489 of any one of SEQ ID NOs: 5-8.
2. The engineered protein of claim 1, comprising a sequence having at least 90% identity to amino acids 19-492 of any one of SEQ ID NOs: 1-4.
3. The engineered protein of claim 1, comprising a sequence having at least 90% identity to amino acids 19-492 of SEQ ID NO: 1.
4. The engineered protein of claim 1, comprising a substitution corresponding to Q100R and S101R within segment (3), with the positions being relative to SEQ ID NOs: 5-8.
5. The engineered protein of any one of claims 1-4, comprising an engineered disulfide bond comprising paired cysteine substitutions at positions corresponding to: T127C and N153C and/or T365C and V463C within segments (3) and (5), respectively, with the positions being relative to SEQ ID NOs: 5-8.
6. The engineered protein of any one of claims 1-4, comprising a substitution at position V231 within segment (5), with the position being relative to SEQ ID NOs: 5-8.
7. The engineered protein of any one of claims 1-4, comprising a substitution corresponding to V231I within segment (5), with the position being relative to SEQ ID NOs: 5-8.

8. The engineered protein of any one of claims 1-7, comprising a substitution at position N67 and/or S215 within segments (2) and (4), respectively, with the positions being relative to SEQ ID NO: 10.
9. The engineered protein of any one of claims 1-8, comprising a substitution corresponding to N67I and/or S215P within segments (2) and (4), respectively, with the positions being relative to SEQ ID NO: 10.
10. The engineered protein of any one of claims 1-9, comprising: (i) substitutions corresponding to N67I and S215P within segments (2) and (4), respectively, with the positions being relative to SEQ ID NO: 10; (ii) a substitution corresponding to V231I within segment (5), with the position being relative to SEQ ID NOs: 5-8; and (iii) engineered disulfide bonds comprising paired cysteine substitutions at positions corresponding to: (a) T127C and N153C and (b) T365C and V463C within segments (3) and (5), respectively, with the positions being relative to SEQ ID NOs: 5-8.
11. The engineered protein of any one of claims 1-10, comprising a substitution at position T91 or L95 within segment (2), with the positions being relative to SEQ ID NO: 10.
12. The engineered protein of any one of claims 1-10, comprising a substitution corresponding to T91R or L95R within segment (2), with the positions being relative to SEQ ID NO: 10.
13. The engineered protein of any one of claims 1-12, comprising a substitution corresponding to L95R within segment (2), with the position being relative to SEQ ID NO: 10.
14. The engineered protein of any one of claims 1-13, comprising an engineered disulfide bond comprising paired cysteine substitutions at positions corresponding to: L110C and N322C within segments (3) and (5), respectively, with the positions being relative to SEQ ID NOs: 5-8.
15. The engineered protein of any one of claims 1-14, comprising a substitution at position A107 within segment (3), with the position being relative to SEQ ID NOs: 5-8.

16. The engineered protein of any one of claims 1-14, comprising a substitution corresponding to A107P within segment (3), with the position being relative to SEQ ID NOs: 5-8.
17. The engineered protein of any one of claims 1-16, comprising a substitution at position L219 within segment (5), with the position being relative to SEQ ID NOs: 5-8.
18. The engineered protein of any one of claims 1-16, comprising a substitution corresponding to L219K within segment (5), with the position being relative to SEQ ID NOs: 5-8.
19. The engineered protein of any one of claims 1-18, comprising a substitution at position E453 within segment (5), with the position being relative to SEQ ID NOs: 5-8.
20. The engineered protein of any one of claims 1-18, comprising a substitution corresponding to E453Q within segment (5), with the position being relative to SEQ ID NOs: 5-8.
21. The engineered protein of any one of claims 1-20, comprising a sequence having at least 95% identity to amino acids 19-492 of SEQ ID NO: 1.
22. The engineered protein of claim 21, comprising engineered disulfide bonds comprising paired cysteine substitutions at positions corresponding to: T130C and N156C, T368C and V466C, and L113C and N325C; and substitutions corresponding to N58I, L86R, A110P, S188P, L222K, V234I, and E456Q, with the positions being relative to SEQ ID NO: 1.
23. The engineered protein of any one of claims 1-22, wherein the protein is fused or conjugated to a trimerization domain.
24. The engineered protein of claim 23, wherein the protein is fused to a trimerization domain.
25. The engineered protein of claim 24, wherein the trimerization domain is positioned C-terminally relative to the chimeric F protein ectodomain.
26. The engineered protein of claim 24 or 25, wherein the trimerization domain comprises a T4 fibritin trimerization domain.

27. The engineered protein of any one of claims 1-26, wherein the protein is fused or conjugated to a transmembrane domain.
28. The engineered protein of claim 27, wherein the protein is fused to a transmembrane domain.
29. The engineered protein of claim 27, wherein the transmembrane domain comprises a metapneumovirus (MPV) F protein transmembrane domain.
30. The engineered protein of any one of claims 1-29, comprising an N-terminal signal sequence.
31. The engineered protein of claim 30, wherein the N-terminal signal sequence is MSWKVVIIIFSLITPQHG.
32. The engineered protein of any one of claims 1-31, comprising a sequence having at least 95% identity to amino acids 19-554 of SEQ ID NO: 1.
33. The engineered protein of any one of claims 1-31, comprising a sequence having at least 95% identity to amino acids 1-554 of SEQ ID NO: 1.
34. An engineered human metapneumovirus (hMPV)–respiratory syncytial virus (RSV) chimeric F protein trimer comprising at least one subunit according to any one of claims 1-33.
35. The engineered trimer of claim 34, wherein the trimer is stabilized in a prefusion conformation relative to a trimer of wildtype metapneumovirus (MPV) F subunits.
36. The engineered trimer of claim 34, wherein the trimer comprises at least one engineered disulfide bond between subunits.
37. A pharmaceutical composition comprising a pharmaceutically acceptable carrier; and (i) an engineered protein of any one of claims 1-33, or (ii) an engineered trimer of any one of claims 34-36.
38. The composition of claim 37, further comprising an adjuvant.
39. A nucleic acid molecule comprising a nucleotide sequence that encodes an amino acid sequence of an engineered protein of any one of claims 1-33.

40. The nucleic acid of claim 39, wherein the nucleic acid comprises a DNA expression vector.
41. The nucleic acid of claim 39, wherein the nucleic acid comprises a mRNA.
42. A method of preventing metapneumovirus (MPV) or respiratory syncytial virus (RSV) infection or a disease associate with MPV or RSV infection in a subject, comprising administering to the subject an effective amount of the pharmaceutical composition of any one of claims 37-38 or a nucleic acid molecule of any one of claims 39-41.
43. A composition comprising (i) an engineered protein of any one of claims 1-33 or (ii) an engineered trimer of any one of claims 34-36 bound to an antibody.

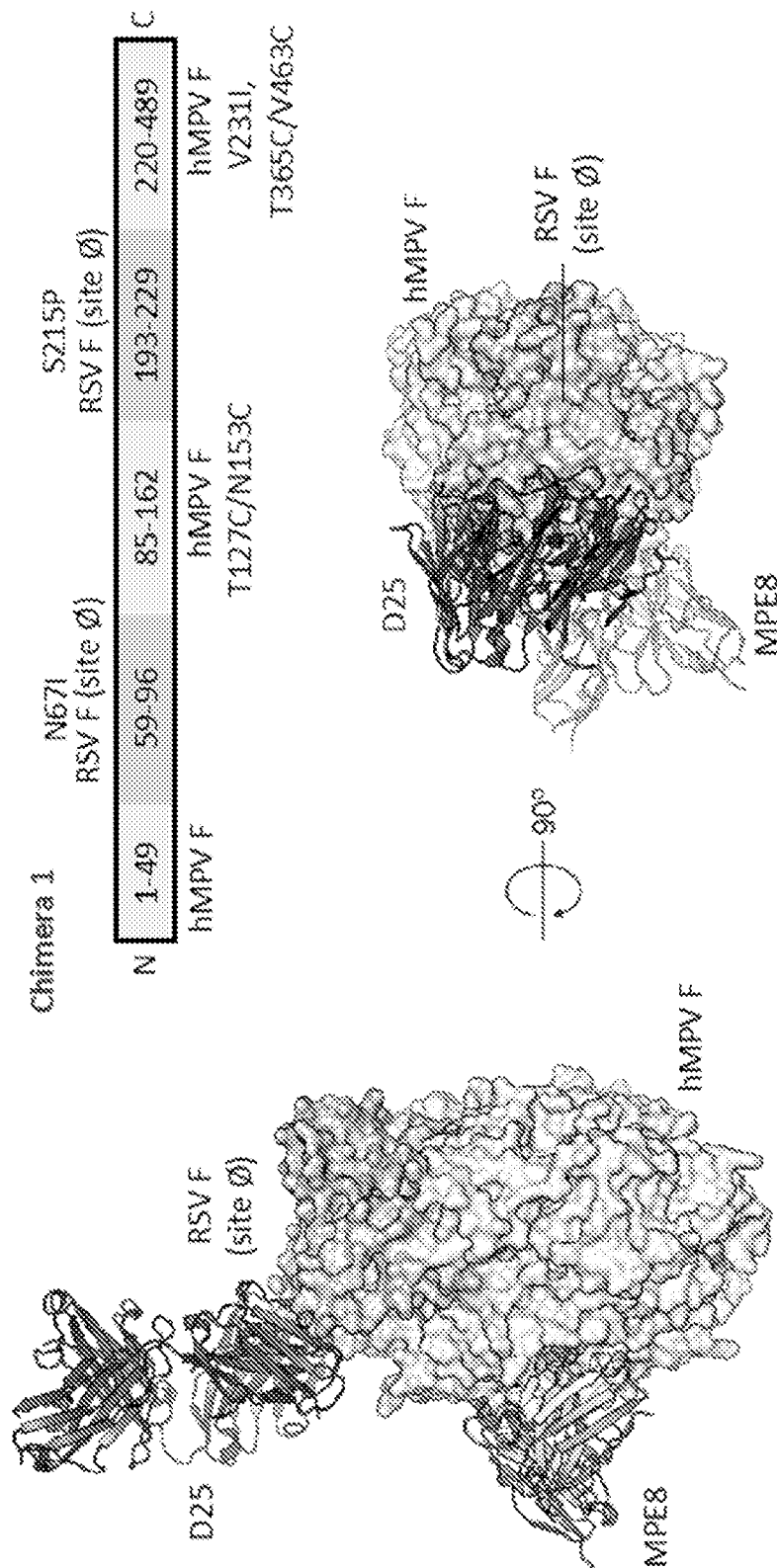


FIG. 1

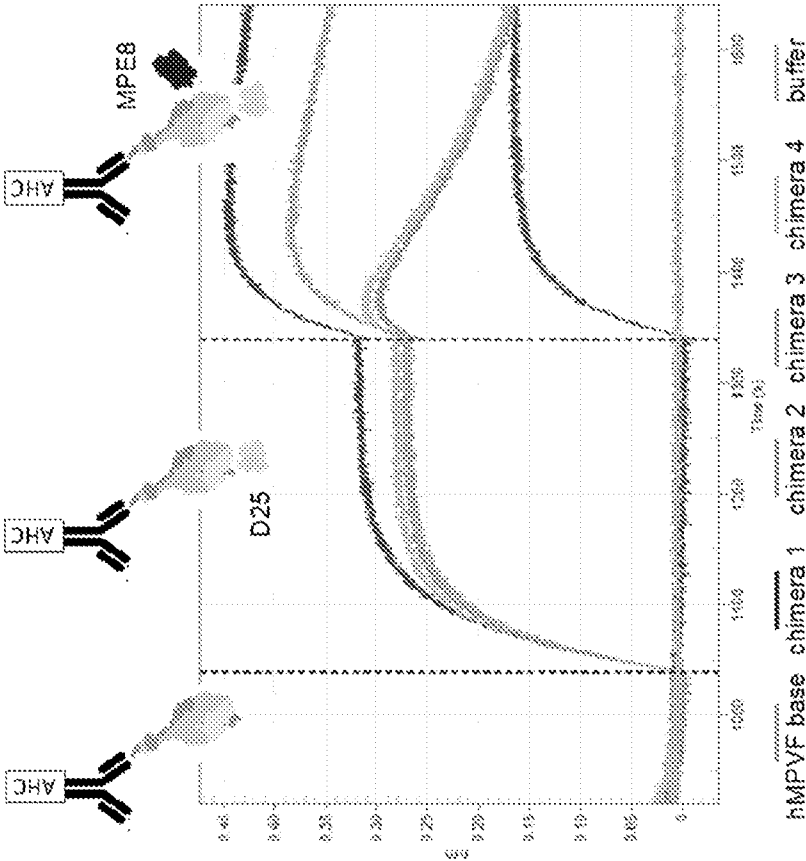
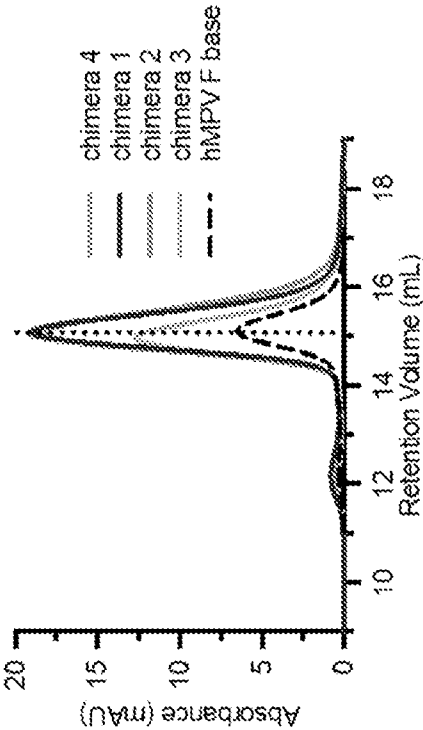


FIG. 2



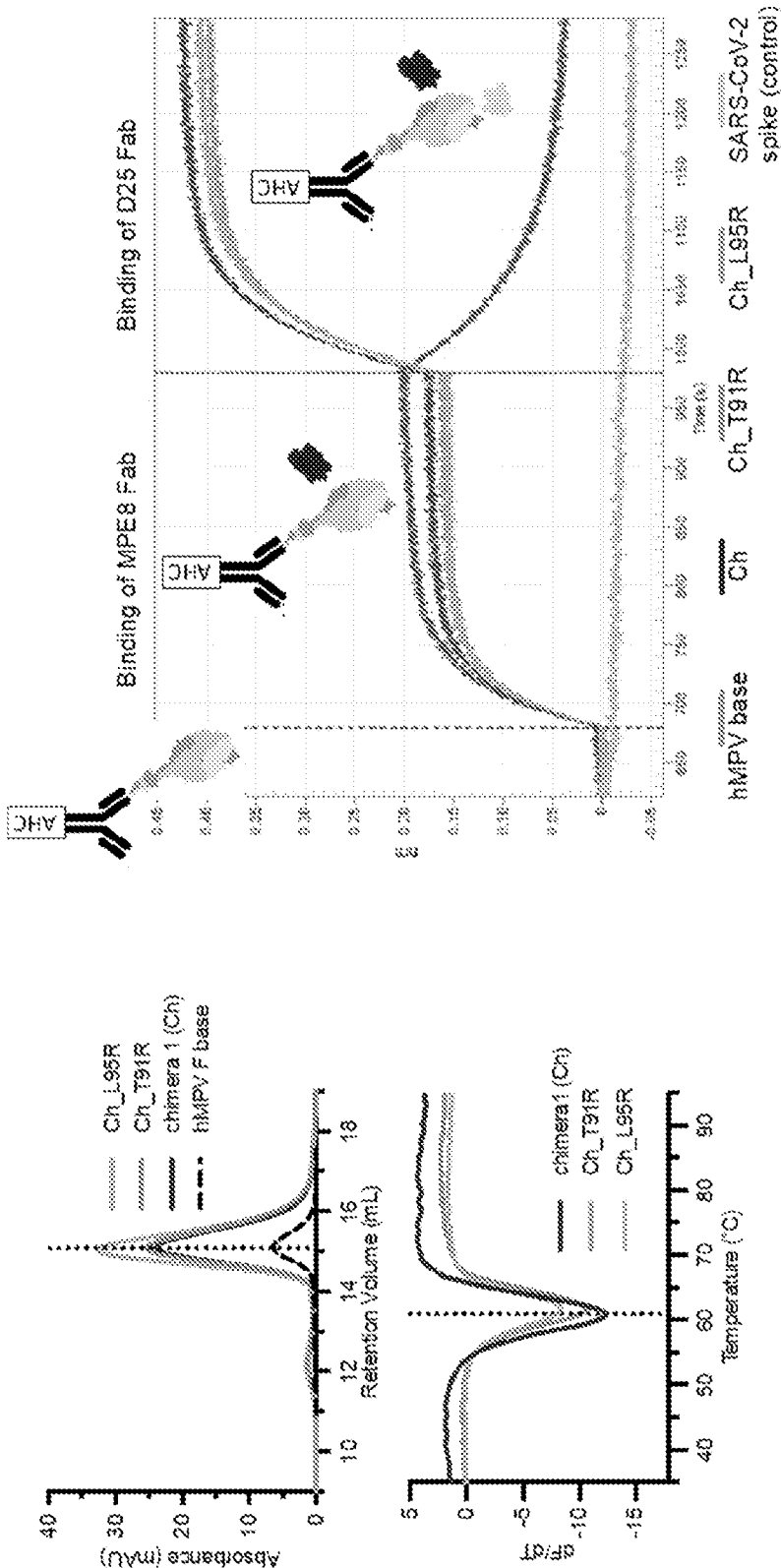


FIG. 3

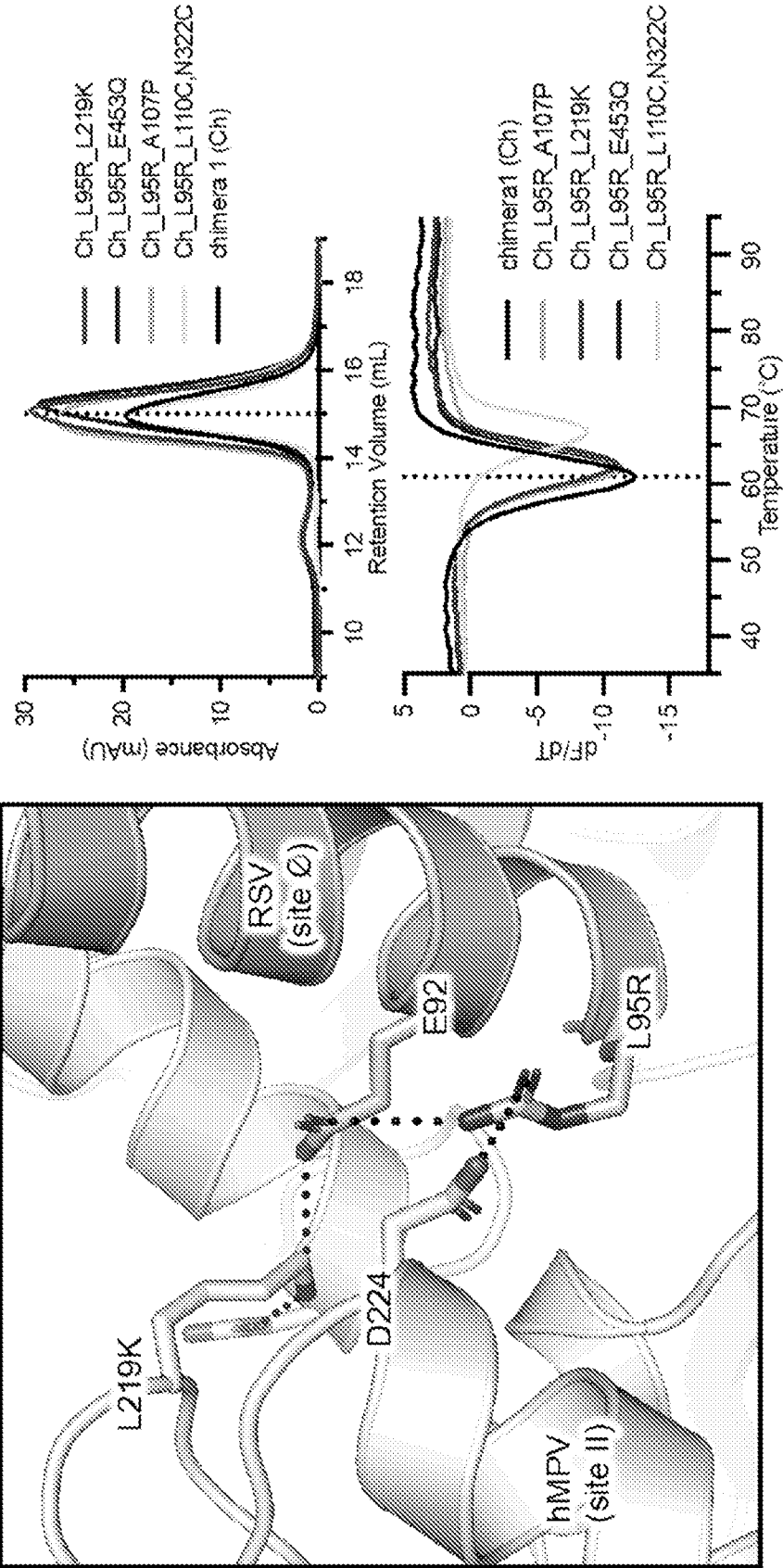


FIG. 4

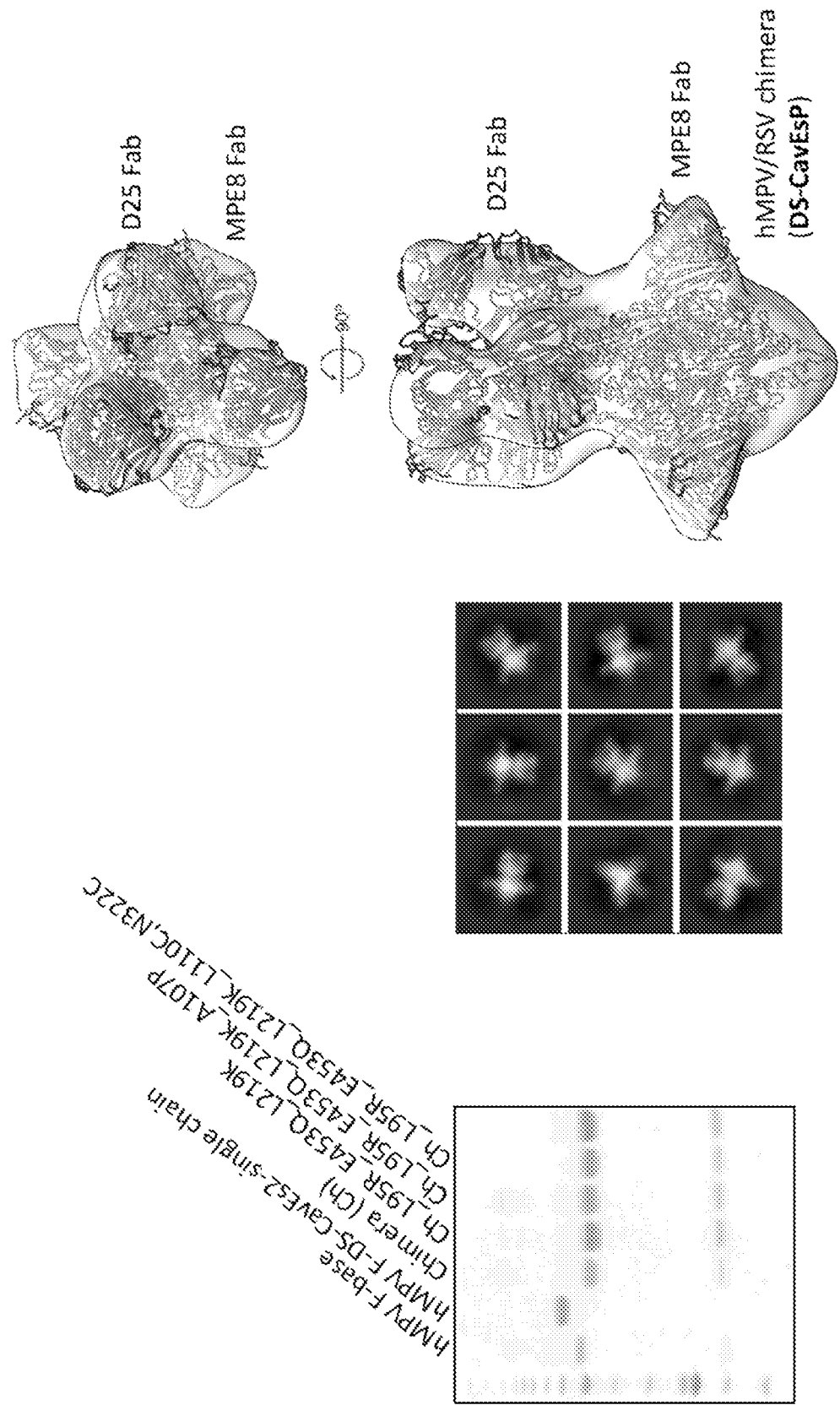
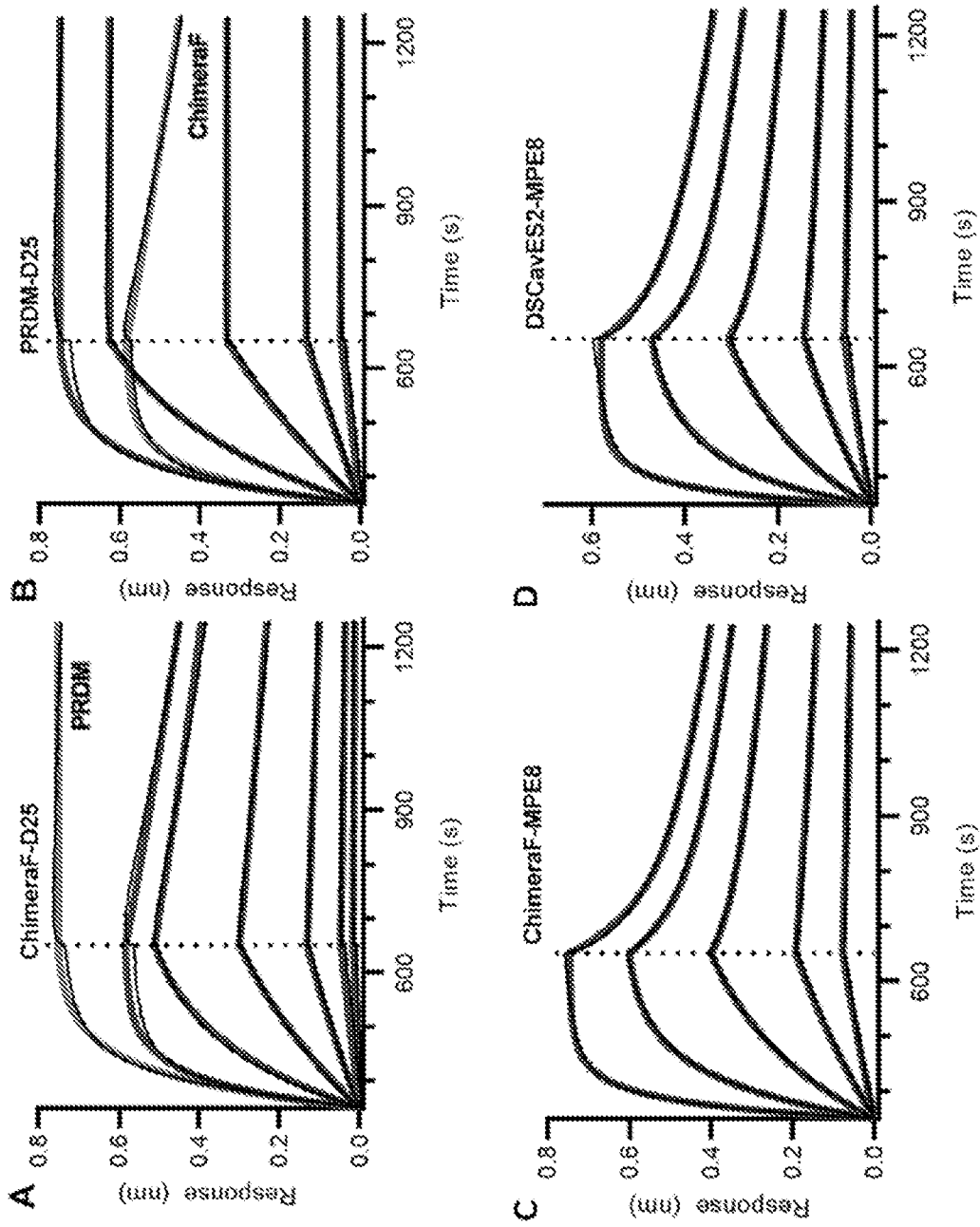


FIG. 5



FIGS. 6A-6D

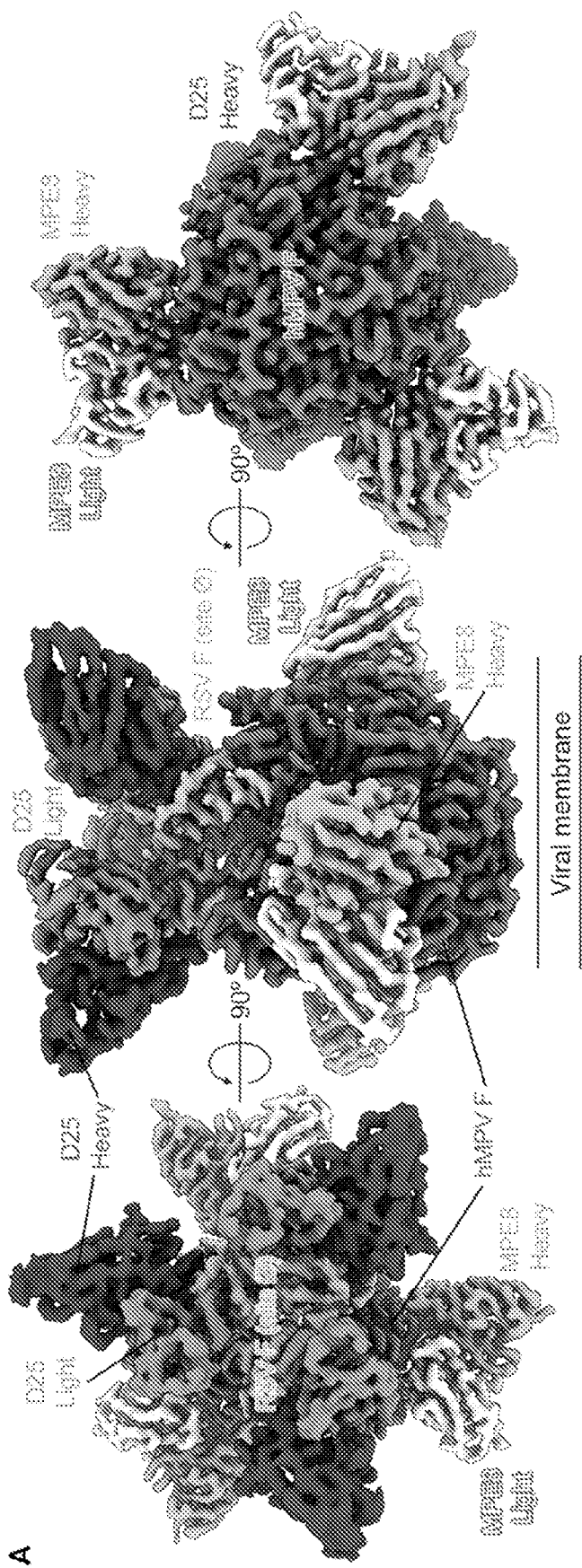
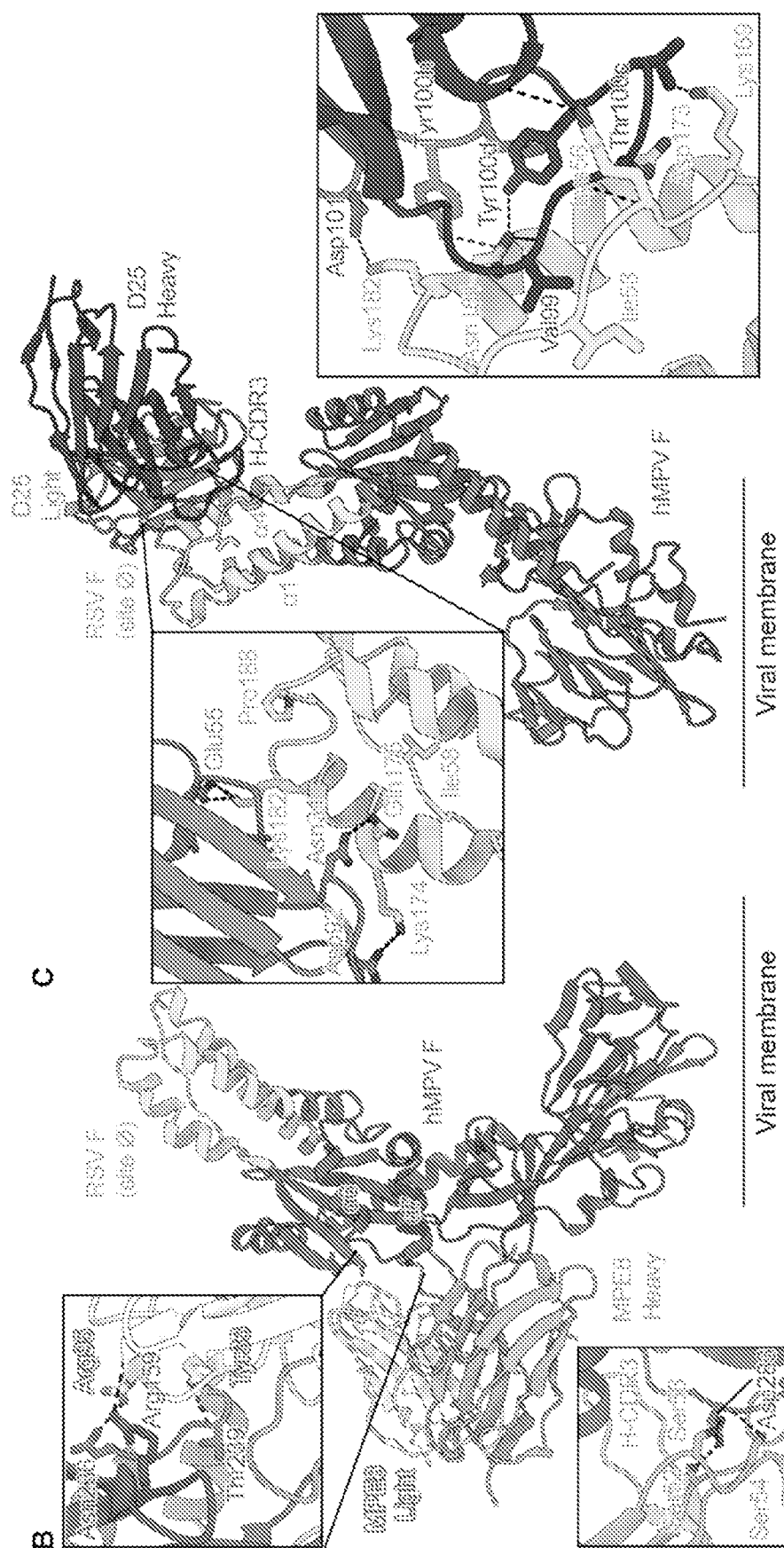


FIG. 7A



FIGS. 7B-7C

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2022/073763

A. CLASSIFICATION OF SUBJECT MATTER

IPC(8) - INV. - C12N 15/62; A61K 39/12; C07K 14/005; C07K 14/135; C07K 19/00 (2022.01)
ADD.

CPC - INV. - C12N 15/62; A61K 39/12 (2022.08)

ADD. - C12N 2760/18311 (2022.08)

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
See Search History document

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
See Search History document

Electronic database consulted during the international search (name of database and, where practicable, search terms used)
See Search History document

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 2020/167813 A1 (EMORY UNIVERSITY et al) 20 August 2020 (20.08.2020) entire document	1-4
A	WO 2018/176103 A1 (THE UNIVERSITY OF QUEENSLAND) 04 October 2018 (04.10.2018) entire document	1-4
A	US 2021/0198323 A1 (ICAHN SCHOOL OF MEDICINE AT MOUNT SINAI et al) 01 July 2021 (01.07.2021) entire document	1-4
A	US 2018/0312544 A1 (THE UNITED STATES OF AMERICA AS REPRESENTED BY THE SECRETARY DEPT. OF HEALTH AND HUMAN SERVICES) 01 November 2018 (01.11.2018) entire document	1-4

☐ Further documents are listed in the continuation of Box C.

☐ See patent family annex.

* Special categories of cited documents:

"A" document defining the general state of the art which is not considered to be of particular relevance

"D" document cited by the applicant in the international application

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search

31 October 2022

Date of mailing of the international search report

NOV 23 2022

Name and mailing address of the ISA/

Mail Stop PCT, Attn: ISA/US, Commissioner for Patents

P.O. Box 1450, Alexandria, VA 22313-1450

Facsimile No. 571-273-8300

Authorized officer

Taina Matos

Telephone No. PCT Helpdesk: 571-272-4300

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2022/073763

Box No. I Nucleotide and/or amino acid sequence(s) (Continuation of item 1.c of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application, the international search was carried out on the basis of a sequence listing:
 - a. ☒ forming part of the international application as filed.
 - b. ☐ furnished subsequent to the international filing date for the purposes of international search (Rule 13ter.1(a)),
☐ accompanied by a statement to the effect that the sequence listing does not go beyond the disclosure in the international application as filed.
2. ☐ With regard to any nucleotide and/or amino acid sequence disclosed in the international application, this report has been established to the extent that a meaningful search could be carried out without a WIPO Standard ST.26 compliant sequence listing.
3. Additional comments:
 SEQ ID NO:1 was searched.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2022/073763

Box No. II Observations where certain claims were found unsearchable (Continuation of item 2 of first sheet)

This international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☒ Claims Nos.: 8-43
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box No. III Observations where unity of invention is lacking (Continuation of item 3 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

See extra sheet(s).

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying additional fees, this Authority did not invite payment of additional fees.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
1-4

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest and, where applicable, the payment of a protest fee.
- ☐ The additional search fees were accompanied by the applicant's protest but the applicable protest fee was not paid within the time limit specified in the invitation.
- ☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US2022/073763

Continued from Box No. III Observations where unity of invention is lacking

This application contains the following inventions or groups of inventions which are not so linked as to form a single general inventive concept under PCT Rule 13.1. In order for all inventions to be examined, the appropriate additional examination fees need to be paid.

Group I+: claims 1-7 are drawn to chimeric hMPV-RSV proteins.

The first invention of Group I+ is restricted to amino acids 19-492 of SEQ ID NO:1. It is believed that claims 1-3 read on this first named invention and thus these claims will be searched without fee to the extent that they read on amino acids 19-492 of SEQ ID NO:1.

Applicant is invited to elect additional hMPV-RSV chimeric F proteins, substitutions, and their respective corresponding, SEQ ID NOs to be searched in a specific combination by paying additional fee for each set of election. An exemplary election would be amino acids 19-492 of SEQ ID NO:2. Additional hMPV-RSV chimeric F proteins, substitutions, and their respective corresponding, SEQ ID NOs will be searched upon the payment of additional fees. Applicants must specify the claims that read on any additional elected inventions. Applicants must further indicate, if applicable, the claims which read on the first named invention if different than what was indicated above for this group. Failure to clearly identify how any paid additional invention fees are to be applied to the "+" group(s) will result in only the first claimed invention to be searched/examined.

The inventions listed in Groups I+ do not relate to a single general inventive concept under PCT Rule 13.1, because under PCT Rule 13.2 they lack the same or corresponding special technical features for the following reasons:

The Groups I+ formulas do not share a significant structural element responsible for proteins exhibiting enhanced conformational stability, enhanced thermostability, and/or increased expression, requiring the selection of alternative engineered human metapneumovirus (hMPV)-respiratory syncytial virus (RSV) chimeric F proteins where "An engineered human metapneumovirus (hMPV)-respiratory syncytial virus (RSV) chimeric F protein comprising an ectodomain having the following segments, in order from N-terminus to C-terminus: (1) a sequence having at least 90% identity to amino acids 19-49 of any one of SEQ ID NOs: 5-8; (2) a sequence having at least 90% identity to amino acids 59-96 of SEQ ID NO: 10; (3) a sequence having at least 90% identity to amino acids 85-162 or 85-164 of any one of SEQ ID NOs: 5-8; (4) a sequence having at least 90% identity to amino acids 193-229, 193-231, 195-229, or 195-231 of SEQ ID NO: 10; and (5) a sequence having at least 90% identity to amino acids 200-489 or 202-489 of any one of SEQ ID NOs: 5-8."

Additionally, even if Groups I+ were considered to share the technical features of an engineered human metapneumovirus (hMPV)-respiratory syncytial virus (RSV) chimeric F protein comprising an ectodomain. However, these shared technical features do not represent a contribution over the prior art.

Specifically, US 2018/0312544 A1 to The United States Of America As Represented By The Secretary Dept. Of Health And Human Services discloses an engineered human metapneumovirus (hMPV)-respiratory syncytial virus (RSV) chimeric F protein comprising an ectodomain (he RSV F insert was engineered to be stabilized, Para. [0075]; including one or more heterologous genes encoding an ectodomain of one or more heterologous envelope proteins ...human metapneumovirus ...subgroup A or subgroup B respiratory syncytial virus F, Para. [0191]).

The inventions listed in Groups I+ therefore lack unity under Rule 13 because they do not share a same or corresponding special technical features.