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(71) Applicant(s)
CSL Limited

(72) Inventor(s)
Horn, Carsten;Zollner, Sabine;Metzner, Hubert;Schulte, Stefan

(74) Agent / Attorney
FB Rice, Level 23 44 Market Street, Sydney, NSW, 2000

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(71) Applicant (for all designated States except US): CSL BEHRING GMBH [DE/DE]; Emil-von-Behring-Straße 76, 35041 Marburg (DE).

(72) Inventors; and

(71) Applicants (for US only): HORN, Carsten [DE/DE]; Höhenweg 39, 35041 Marburg (DE). ZOLLNER, Sabine [DE/DE]; Talblickstrasse 6, 59969 Bromskirchen (DE). METZNER, Hubert [DE/DE]; Im Boden 6, 35041 Marburg (DE). SCHULTE, Stefan [DE/DE]; Bauerbacher Straße 46, 35043 Marburg (DE).

(74) Agents: HAUSER, Hans-Peter et al.; Emil-von-Behring-Straße 76, 35041 Marburg (DE).

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(54) Title: METHOD FOR IMPROVING THE STABILITY OF PURIFIED FACTOR VIII AFTER RECONSTITUTION

(57) Abstract: The present invention relates to a method for increasing the stability of a Factor VIII molecule after purification, lyophilization and reconstitution, comprising preventing proteolytic cleavage of the Factor VIII molecule into a first fragment comprising essentially the A1 domain and the A2 domain and a second fragment comprising essentially the A3 domain, the C1 domain and the C2 domain throughout manufacturing of the Factor VIII molecule. The invention further pertains to a method for improving the bioavailability of Factor VIII after intravenous and non- intravenous injection.

5 Method for improving the stability of purified Factor VIII after reconstitution

The present invention relates to a method for increasing the stability of a Factor VIII molecule after purification, lyophilization and reconstitution, comprising preventing proteolytic cleavage of the Factor VIII molecule into a first fragment comprising 10 essentially the A1 domain and the A2 domain and a second fragment comprising essentially the A3 domain, the C1 domain and the C2 domain throughout manufacturing of the Factor VIII molecule. The invention further pertains to a method for improving the bioavailability of Factor VIII after intravenous and non-intravenous injection.

15

BACKGROUND OF THE INVENTION

Classic hemophilia or hemophilia A is an inherited bleeding disorder. It results from 20 a chromosome X-linked deficiency of blood coagulation Factor VIII, and affects almost exclusively males with an incidence of between one and two individuals per 10,000. The X-chromosome defect is transmitted by female carriers who are not 25 themselves hemophiliacs. The clinical manifestation of hemophilia A is an increased bleeding tendency. Before treatment with Factor VIII concentrates was introduced the mean life span for a person with severe hemophilia was less than 20 years. The use of concentrates of Factor VIII from plasma has considerably improved the situation for the hemophilia patients increasing the mean life span extensively, giving most of them the possibility to live a more or less normal life. 30 However, there have been certain problems with the plasma derived concentrates and their use, the most serious of which have been the transmission of viruses. So far, viruses causing AIDS, hepatitis B, and non-A non-B hepatitis have hit the population seriously. Since then different virus inactivation methods and new highly

purified Factor VIII concentrates have recently been developed which established a very high safety standard also for plasma derived Factor VIII.

Several recombinant and plasma-derived, therapeutic polypeptides, e.g. blood 5 coagulation factors, are commercially available for therapeutic and prophylactic use in humans. FVIII is a blood plasma glycoprotein of up to about 280 kDa molecular mass, produced in the liver of mammals. It is a critical component of the cascade of coagulation reactions that lead to blood clotting. Within this cascade is a step in which factor IXa (FIXa), in conjunction with activated factor VIII (FVIIIa), converts 10 factor X (FX) to an activated form, FXa. FVIIIa acts as a cofactor at this step, being required together with calcium ions and phospholipids for maximizing the activity of FIXa.

An important advance in the treatment of hemophilia A has been the isolation of 15 cDNA clones encoding the complete 2,351 amino acid sequence of human FVIII (United States Patent No. 4,757,006) and the provision of the human FVIII gene DNA sequence and recombinant methods for its production).

Factor VIII is synthesized as a single polypeptide chain with a molecular weight of 20 about 280 kDa. The amino-terminal signal peptide is removed upon translocation of factor VIII into the endoplasmatic reticulum, and the mature (i.e. after the cleavage of the signal peptide) native Factor VIII molecule is then proteolytically cleaved after amino acid residues 1313 and 1648 in the course of its secretion. This results in the release of a heterodimer which consists of a C-terminal light chain of about 80 kDa 25 in a metal ion-dependent association with an about 160-200 kDa N-terminal heavy chain fragment. (See review by Kaufman, Transfusion Med. Revs. 6:235 (1992)).

Physiological activation of the heterodimer occurs through proteolytic cleavage of 30 the protein chains by thrombin. Thrombin cleaves the heavy chain to a 90 kDa protein, and then to 54 kDa and 44 kDa fragments. Thrombin also cleaves the 80 kDa light chain to a 72 kDa protein. It is the latter protein, and the two heavy chain

fragments (54 kDa and 44 kDa above), held together by calcium ions, that constitute active FVIII. Inactivation occurs when the 44 kDa A2 heavy chain fragment dissociates from the molecule or when the 72 kDa and 54 kDa proteins are further cleaved by thrombin, activated protein C or FXa. In plasma, FVIII is 5 stabilized by association with a 50-fold molar excess of vWF protein ("vWF"), which appears to inhibit proteolytic destruction of FVIII as described above.

The amino acid sequence of FVIII is organized into three structural domains: a triplicated A domain of 330 amino acids, a single B domain of 980 amino acids, and 10 a duplicated C domain of 150 amino acids. The B domain has no homology to other proteins and provides 18 of the 25 potential asparagine(N)-linked glycosylation sites of this protein. The B domain has apparently no function in coagulation and can be deleted with the B-domain deleted FVIII molecule still having procoagulatory activity.

15

The Factor VIII products on the market are currently presented as a lyophilized formulation of Factor VIII either produced by recombinant technology or purified from pooled plasma. The lyophilized product is reconstituted prior to administration. Once reconstituted, shelf-life of the Factor VIII is relatively short. Factor VIII is a 20 relatively unstable protein, particularly in aqueous solutions. Stabilization during manufacturing and storage by complexing with other plasma proteins, particularly von Willebrand factor (vWF) and albumin, has been described. See, for example, US Patent No. 6,228,613. US Patent No. 5,565,427 discloses a stabilized formulation of Factor VIII comprising an amino acid or one of its salts or 25 homologues and a detergent or an organic polymer such as polyethyleneglycol. US Patent No. 5,605,884 discloses stabilized formulations of Factor VIII in high ionic strength media based on histidine buffer in the presence of calcium chloride and a high concentration of sodium chloride or potassium chloride. Such compositions were shown to improve significantly the stability of Factor VIII in aqueous form 30 following reconstitution. The importance of calcium ions in the formulations of Factor VIII is generally recognized. According to US Patent No. 6,599,724, the

presence of other divalent cations, namely Cu^{2+} and Zn^{2+} , optionally in the presence of Ca^{2+} ions or Mn^{2+} ions improves the stability of Factor VIII. Also WO 2011/027152 A1 describes stable aqueous Factor VIII compositions comprising various additives.

5

In view of the short shelf life of Factor VIII after reconstitution of a lyophilisate, there is a need for methods to increase the stability of reconstituted Factor VIII in aqueous solution. To provide a purified FVIII preparation with increased stability in the liquid phase is desirable for different reasons. First of all, it is of advantage to 10 have a sufficient time span at ambient temperature to support manufacturing of the purified FVIII product at ambient temperature. In particular, the filling step necessitates some storage of a liquid bulk to increase flexibility in manufacturing. Secondly, an increased stability of the liquid purified FVIII would be of advantage for physician and patient if the product could not be applied directly after 15 reconstitution. And finally, the use of FVIII under continuous infusion conditions e.g. upon surgery in hospitalized patients is depending on a preferably high product stability after reconstitution (Takedani H., Haemophilia 2010, 16: 740-746). A FVIII molecule with increased stability would also be an advantage for development of a FVIII preparation suitable for long term storage under liquid conditions.

20

The inventors of this application surprisingly found that the stability of purified Factor VIII after reconstitution of a lyophilisate is substantially enhanced in single-chain Factor VIII constructs. Such constructs can be obtained by preventing the proteolytic cleavage which typically occurs in the Golgi compartment prior to 25 secretion of Factor VIII. The single-chain constructs exhibit a better stability in solution after purification and/or a better bioavailability upon subcutaneous administration.

Any discussion of documents, acts, materials, devices, articles or the like which has been included in the present specification is not to be taken as an admission that any or all of these matters form part of the prior art base or were common general knowledge in the field relevant to the present disclosure as it existed before the 5 priority date of each claim of this application.

Throughout this specification the word "comprise", or variations such as "comprises" or "comprising", will be understood to imply the inclusion of a stated element, integer or step, or group of elements, integers or steps, but not the 10 exclusion of any other element, integer or step, or group of elements, integers or steps.

SUMMARY OF THE INVENTION

15 In a first aspect the present invention provides a method for improving the plasma half-life of a Factor VIII molecule after intravenous or non-intravenous administration relative to human wild-type Factor VIII, comprising fusing a first amino acid selected from the amino acids at positions 740 to 1647 of the Factor VIII sequence with a second amino acid selected from the amino acids at positions 20 1649 to 1690 of the Factor VIII sequence, whereby the proteolytic cleavage site between Arg1648 and Glu1649, and, if present in the FVIII molecule, the cleavage site between Arg1313 and Ala1314 is inactivated.

25 In another aspect the present invention relates to a method for increasing the stability of a Factor VIII molecule after purification, lyophilization and reconstitution, comprising preventing proteolytic cleavage of the Factor VIII molecule into a first

fragment comprising essentially the A1 domain and the A2 domain and a second fragment comprising essentially the A3 domain, the C1 domain and the C2 domain.

A further aspect of the present invention encompasses a method for increasing the 5 stability of a Factor VIII molecule after purification, lyophilization and reconstitution, comprising preventing proteolytic cleavage of the Factor VIII molecule into a first fragment comprising essentially the A1 domain and the A2 domain and a second fragment comprising essentially the A3 domain, the C1 domain and the C2 domain throughout manufacturing of the Factor VIII molecule.

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A futher aspect of the present invention encompasses a method for increasing the stability of a Factor VIII molecule after purification, lyophilization and reconstitution, comprising preventing proteolytic cleavage of the Factor VIII molecule into a first fragment comprising essentially the A1 domain and the A2 domain and a second 15 fragment comprising essentially the A3 domain, the C1 domain and the C2 domain prior to the purification of the Factor VIII molecule.

20

With regard to these methods according to the invention the terms "throughout manufacturing of the Factor VIII molecule" and "prior to the purification of the Factor 20 VIII molecule" are intended to mean that the methods of the invention prevent the cleavage of Factor VIII into a first fragment comprising essentially the A1 domain and the A2 domain and a second fragment comprising essentially the A3 domain, the C1 domain and the C2 domain but the methods according to the invention do not prevent the activation cleavage of Factor VIII which may occur after 25 administration of the reconstituted Factor VIII molecule. The Factor VIII molecules

generated by the methods of the invention can still be activated by thrombin which cleaves the Factor VIII molecule after Arg 372, Arg 740 and Arg 1689.

In a second aspect, the present invention relates to a method for increasing the

5 stability of a Factor VIII molecule after purification, lyophilization and reconstitution, comprising inactivating the proteolytic cleavage sites which are cleaved during secretion of said Factor VIII molecule by the host cell expressing the Factor VIII molecule except the cleavage site between the signal sequence and the mature Factor VIII. Typically, at least 50% of the Factor VIII molecules expressed and

10 secreted by the host cells are single-chain Factor VIII molecules. Preferably, at least 60%, or at least 70%, or at least 80%, or at least 90%, or at least 95% of the Factor VIII molecules expressed and secreted by the host cells are single-chain Factor VIII molecules.

15 Preferably, the method comprises inactivating the proteolytic cleavage site between Arg1648 and Glu1649 and, if present in the FVIII molecule, the proteolytic cleavage site between Arg1313 and Ala1314. The inactivation of the proteolytic cleavage site may be effected by deleting one or more residues of the protease recognition sequence. For example, inactivation step may comprise deleting at least Arg1648

20 from the Factor VIII sequence. In one embodiment, the inactivation step comprises deleting at least the amino acid sequence from Arg1313 to Arg1648 from the Factor VIII sequence.

In another embodiment of the first aspect of the invention the inactivation of the

25 proteolytic cleavage site is effected by substituting one or more amino acid residues forming the protease recognition sequence.

In yet another embodiment (concerning those FVIII variants which retain the part of the B-domain comprising Arg1313) the method further comprises inactivating the

30 proteolytic cleavage site between Arg1313 and Ala1314 by deletion or substitution of one or more residues forming the protease recognition sequence. In a

particularly preferred embodiment, the method comprises deleting at least a portion from the Factor VIII amino acid sequence which comprises both protease cleavage sites after residues Arg1313 and Arg1648.

5 It is further preferred that a first amino acid selected from the amino acids at positions 741 to 1647 of the Factor VIII sequence is fused with a second amino acid selected from the amino acids at positions 1649 to 1690 of the Factor VIII sequence, whereby the proteolytic cleavage site between Arg1648 and Glu1649 and, if present in the FVIII molecule, the cleavage site between Arg1313 and
10 Ala1314 is inactivated.

In another preferred embodiment the Factor VIII molecule stabilized in accordance with the first or second aspect of the invention exhibits an increased stability in aqueous solution. The loss of activity of the modified Factor VIII molecule, in
15 aqueous solution, after storage for 7 days at 25°C is preferably less than 15%.

In another preferred embodiment the Factor VIII molecule stabilized in accordance with the first or second aspect of the invention exhibits an increased stability in aqueous solution after reconstitution.

20 In yet another preferred embodiment the Factor VIII molecule stabilized in accordance with the first or second aspect of the invention exhibits an increased bioavailability after non-intravenous injection, as compared to the bioavailability of human wild type Factor VIII or as compared to a B-domain deleted Factor VIII
25 molecule where Asn745 is fused to Pro1640, administered at the same dose and in the same manner. In yet another preferred embodiment the Factor VIII molecule stabilized in accordance with the first or second aspect of the invention exhibits an increased bioavailability after non-intravenous injection, as compared to the bioavailability of a B-domain deleted Factor VIII molecule where Asn745 is fused to
30 Pro1640, administered at the same dose and in the same manner. The bioavailability of the modified FVIII is preferably increased by at least 25%, as

compared to the bioavailability of human wild type Factor VIII or of a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640, each administered at the same dose and in the same manner. In another preferred embodiment the non-intravenous injection is subcutaneous, transdermal or intramuscular injection.

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Another preferred embodiment is a method wherein (i) the Factor VIII exhibits improved plasma half-life after intravenous administration relative to human wild type Factor VIII; preferably wherein the plasma half-life is improved by at least 40% relative to human wild type Factor VIII, or (ii) wherein the Factor VIII exhibits a 10 longer time period for the thrombin peak level as determined in a thrombin generation assay over time in hemophilia A mice to fall below 50 nM after intravenous administration relative to human wild type Factor VIII; preferably wherein this time period is prolonged by at least 10 hours relative to human wild type Factor VIII, or (iii) wherein the Factor VIII retains a higher activity as 15 determined by a one-stage FVIII:C assay after having been incubated for 4 days in human plasma at 37°C relative to human wild type Factor VIII after having been incubated for 4 days in human plasma at 37°C; preferably wherein the retained activity of the Factor VIII is at least 10% higher relative to that of a human wild type Factor VIII after having been incubated for 4 days in human plasma at 37°C.

20

The methods may further comprise the steps of

- (i) providing a nucleic acid encoding a modified Factor VIII molecule in which the proteolytic cleavage sites between Arg1648 and Glu1649, and between Arg1313 and Ala1314, are inactivated,
- 25 (ii) transforming a host cell with said nucleic acid,
- (iii) culturing the transformed host cell under conditions such that the modified Factor VIII molecule is expressed, and
- (iv) recovering the modified Factor VIII molecule from the host cells or from the culture medium.

30

In another aspect, the present invention relates to a method for improving the bioavailability of a Factor VIII molecule after non-intravenous administration, comprising inactivating the proteolytic cleavage site between Arg1648 and Glu1649 and, if present in the FVIII molecule, the proteolytic cleavage site between Arg1313

5 and Ala1314. Preferably, the non-intravenous injection is subcutaneous injection. The bioavailability after subcutaneous injection is preferably increased by at least 25% as compared to that of human wild type Factor VIII or of a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640, each administered at the same dose and in the same manner.

10

In another aspect, the present invention relates to a method for improving the plasma half-life of a Factor VIII molecule after intravenous administration relative to human wild-type Factor VIII, comprising inactivating the proteolytic cleavage site between Arg1648 and Glu1649, and, if present in the FVIII molecule, the proteolytic cleavage site between Arg1313 and Ala1314.

15 In another aspect, the present invention relates to a method for prolonging the time period for the thrombin peak level as determined in a thrombin generation assay over time in hemophilia A mice to fall below 50 nM after intravenous administration of a Factor VIII molecule relative to human wild type Factor VIII, comprising inactivating the proteolytic cleavage site between Arg1648 and Glu1649, and, if present in the FVIII molecule, the proteolytic cleavage site between Arg1313 and Ala1314.

20 25 In another aspect, the present invention relates to a method for retaining a higher activity for a Factor VIII molecule as determined by a one-stage FVIII:C assay after having been incubated for 4 days in human plasma at 37°C relative to human wild type Factor VIII after having been incubated for 4 days in human plasma at 37°C, comprising inactivating the proteolytic cleavage site between Arg1648 and Glu1649, and, if present in the FVIII molecule, the proteolytic cleavage site between Arg1313 and Ala1314.

A preferred embodiment of the methods described above are methods wherein a first amino acid selected from the amino acids at positions 741 to 1647 of the Factor VIII sequence is fused with a second amino acid selected from the amino acids at 5 positions 1649 to 1690 of the Factor VIII sequence, whereby the proteolytic cleavage site between Arg1648 and Glu1649, and, if present in the FVIII molecule, the proteolytic cleavage site between Arg1313 and Ala1314 is inactivated.

10 The preferred embodiments of the different aspects are applicable *mutatis mutandis*.

In yet another aspect, the present invention relates to a pharmaceutical preparation comprising a single chain Factor VIII molecule for use in the treatment or prophylaxis of a bleeding disorder, preferably hemophilia A, by (i) on the one hand 15 non-intravenous administration, wherein the bioavailability of said single chain Factor VIII molecule is increased by at least 25% as compared to human wild type Factor VIII or as compared to a B-domain deleted human Factor VIII molecule where Asn745 is fused to Pro1640, administered at the same dose and in the same manner, or (ii) on the other hand by intravenous administration, wherein (a) the 20 plasma half-life of said single chain Factor VIII molecule after intravenous administration is increased by at least 40%, relative to human wild type Factor VIII, administered at the same dose and in the same manner, or (b) the single chain Factor VIII molecule exhibits a time period prolonged by at least 10 hours for the thrombin peak level as determined in a thrombin generation assay over time in 25 hemophilia A mice to fall below 50 nM after intravenous administration relative to human wild type Factor VIII, administered at the same dose and in the same manner.

30 In yet another aspect, the present invention relates to a pharmaceutical preparation comprising a single chain Factor VIII molecule for use in the treatment or prophylaxis of a bleeding disorder, preferably hemophilia A, wherein the single

chain Factor VIII molecule retains at least a 10% higher activity as determined by a one-stage FVIII:C assay after having been incubated for 4 days in human plasma at 37°C relative to human wild type Factor VIII after having been incubated for 4 days in human plasma at 37°C.

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In yet another aspect, the present invention relates to a pharmaceutical preparation comprising a single chain Factor VIII molecule for use in the treatment or prophylaxis of a bleeding disorder, preferably hemophilia A, by non-intravenous administration, wherein the dose of said FVIII molecule can be decreased by at 10 least 25% as compared to that of a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640, administered at the same dose and in the same manner to achieve the same hemostatic activity in blood.

In another aspect, the present invention relates to the use of a single chain Factor 15 VIII molecule for achieving an increased stability after reconstitution or a longer shelf life of a pharmaceutical preparation for treating a bleeding disorder, wherein (i) the Factor VIII activity of the pharmaceutical preparation comprising the single chain Factor VIII molecule, after reconstitution and storage at room temperature for 7 days after reconstitution is at least 10% higher than that of a pharmaceutical preparation comprising the same amount of a B-domain deleted Factor VIII 20 molecule where Asn745 is fused to Pro1640, or (ii) wherein the single chain Factor VIII molecule retains at least a 10% higher activity as determined by a one-stage FVIII:C assay when incubated for 4 days in human plasma at 37°C relative to human wild type Factor VIII after having been incubated for 4 days in human 25 plasma at 37°C at the same concentration.

DESCRIPTION OF THE DRAWINGS

Figure 1 depicts the results of Example 1. Various Factor VIII molecules have been provided as aqueous solutions, and their stability has been monitored over a time

5 period of seven days. The loss in activity after seven days of storage is much less for the single chain Factor VIII molecule as compared to heterodimeric (two-chain) full length Factor VIII molecules (Beriate® and Helixate®) and to heterodimeric (two-chain) B-domain deleted constructs (ReFacto®).

10 **Figure 2** depicts the results of Example 2. Various lyophilized Factor VIII preparations were reconstituted to aqueous solutions, and their stability has been monitored over a time period of seven days. The loss in activity after seven days of storage is much less for the single chain Factor VIII molecule as compared to a heterodimeric (two-chain) full length Factor VIII molecule (Advate®) and to a 15 heterodimeric (two-chain) and B-domain deleted construct (ReFacto®).

20 **Figure 3** depicts the results of Example 3. Three different Factor VIII molecules have been injected subcutaneously in mice and their bioavailability has been determined as described in Example 2. The bioavailability of the single chain Factor VIII molecule is substantially higher than that of a two chain and full length Factor VIII (Advate®) or a heterodimeric (two chain) B-domain deleted construct (ReFacto®).

25 **Figure 4** depicts the results of Example 4. The Factor VIII molecule of the invention and 2 commercially obtainable FVIII preparations were incubated at 37°C after purification, lyophilization and reconstitution. The FVIII- samples were incubated at 37°C for varying time periods (0, 0.25, 1, 2, 4 and 8 days) and the FVIII:C activity was determined by an one-stage-coagulation assay. The values shown represent the average and standard deviation of two samples (except 0.25 days only one 30 sample).

Figure 5 depicts part of the results of Example 5. The pharmacokinetic (PK) profiles of scFVIII and full-length rFVIII (Advate[®], Baxter Healthcare) were determined following a single I.V. injection to cynomolgus monkeys at a dose of 250 IU/kg.

5 **Figure 6** depicts part of the results of Example 5. The pharmacokinetic (PK) profiles of full-length rFVIII (Advate[®], Baxter Healthcare) were determined following a single I.V. injection to hemophilia A mice at a dose of 100 IU/kg.

10 **Figure 7** depicts part of the results of Example 6. The average peak thrombin levels from days 1-8 were determined after scFVIII or full-length rFVIII (Advate[®], Baxter Healthcare) were administered to hemophilia A mice at a dose of 250 IU/kg.

15 **Figure 8** depicts the results of Example 7. The pharmacokinetic (PK) profiles of full-length rFVIII (Advate[®], Baxter Healthcare) and of a B-domain deleted Factor VIII (ReFacto[®], Pfizer) was determined following a single I.V. injection to VWF deficient mice at a dose of 100 IU/kg.

DETAILED DESCRIPTION

20

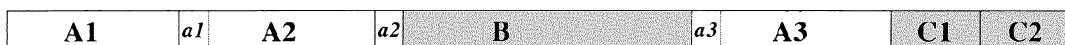
The present invention relates to a method for increasing the stability of a Factor VIII molecule after purification, lyophilization and reconstitution, comprising preventing proteolytic cleavage of the Factor VIII molecule into a first fragment comprising essentially the A1 domain and the A2 domain and a second fragment comprising 25 essentially the A3 domain, the C1 domain and the C2 domain.

30 This invention further pertains to a method for increasing the stability of a Factor VIII molecule after purification, lyophilization and reconstitution, comprising inactivating the proteolytic cleavage site between Arg1648 and Glu1649 and, optionally inactivating the proteolytic cleavage site between Arg1313 and Ala1314, if present in the Factor VIII molecule.

Factor VIII

The terms "blood coagulation Factor VIII", "Factor VIII" and FVIII" are used

5 interchangeably herein. Mature human Factor VIII consists of 2332 amino acids which are arranged in the following domain structure:



10 A1: residues 1-336,
A2: residues 373-710,
B: residues 741-1648,
A3: residues 1690-2019,
C1: residues 2020-2172,
15 and
C2: residues 2173-2332.

In addition, there are three acidic regions a_1 (337-372), a_2 (711-740), and a_3 (1649-1689). It is known that the acidic region a_3 is involved in the binding of the
20 Factor VIII molecule to von Willebrand Factor (vWF) which plays an important role in blood coagulation. During secretion, the FVIII is cleaved between the B-domain and the a_3 acidic region, resulting in a heterodimeric polypeptide. The factor VIII heterodimer consists of a light chain (comprising A3, C1 and C2) and a variably sized heavy chain (comprising A1, A2 and B). The latter is heterogeneous due to
25 limited proteolysis within the B-domain. In case of heterodimeric B-domain deleted constructs the "heavy chain" comprises A1 and A2 but lacks part or all of the B-domain.

The amino acid sequence of the mature wild type form of human blood coagulation
30 Factor VIII is shown in SEQ ID NO:2. The reference to an amino acid position of a specific sequence means the position of said amino acid in the FVIII wild-type

- 15 -

protein and does not exclude the presence of mutations, e.g. deletions, insertions and/or substitutions at other positions in the sequence referred to. For example, a mutation in "Glu2004" referring to SEQ ID NO:2 does not exclude that in the modified homologue one or more amino acids at positions 1 through 2332 of SEQ 5 ID NO:2 are missing. A DNA sequence encoding SEQ ID NO:2 is shown in SEQ ID NO:1.

"Blood coagulation Factor VIII" includes wild type blood coagulation Factor VIII as well as derivatives of wild type blood coagulation Factor VIII having the 10 procoagulant activity of wild type blood coagulation Factor VIII. Derivatives may have deletions, insertions and/or additions compared with the amino acid sequence of wild type Factor VIII. Preferred derivatives are FVIII molecules in which all or part of the B-domain has been deleted. Amino acid positions indicated throughout this application always refer to the position of the respective amino acid in the full length 15 mature (i.e. after signal peptide cleavage) wild-type FVIII.

The term "factor VIII" includes any factor VIII variants or mutants having at least 10%, preferably at least 25%, more preferably at least 50%, most preferably at least 75% of the biological activity of wild type factor VIII. A suitable test to determine the 20 biological activity of Factor VIII is the one stage or the two stage coagulation assay (Rizza et al. 1982. Coagulation assay of FVIII:C and FIXa in Bloom ed. The Hemophilias. NY Churchill Livingston 1992) or the chromogenic substrate FVIII:C assay (S. Rosen, 1984. Scand J Haematol 33: 139-145, suppl.). The content of these references is incorporated herein by reference.

25

As non-limiting examples, Factor VIII molecules include Factor VIII mutants preventing or reducing APC cleavage (Amano 1998. Thromb. Haemost. 79:557-563), albumin-fused FVIII molecules (WO 2011/020866 A2), FVIII-Fc fusion molecules (WO 04/101740 A), Factor VIII mutants further stabilizing the A2 domain 30 (WO 97/40145), FVIII mutants resulting in increased expression (Swaroop et al. 1997. JBC 272:24121-24124), Factor VIII mutants with reduced immunogenicity

(Lollar 1999. Thromb. Haemost. 82:505-508), FVIII reconstituted from differently expressed heavy and light chains (Oh et al. 1999. Exp. Mol. Med. 31:95-100), FVIII mutants reducing binding to receptors leading to catabolism of FVIII like HSPG (heparan sulfate proteoglycans) and/or LRP (low density lipoprotein receptor related protein) (Ananyeva et al. 2001. TCM, 11:251-257), disulfide bond-stabilized FVIII variants (Gale et al., 2006. J. Thromb. Hemost. 4:1315-1322), FVIII mutants with improved secretion properties (Miao et al., 2004. Blood 103:3412-3419), FVIII mutants with increased cofactor specific activity (Wakabayashi et al., 2005. Biochemistry 44:10298-304), FVIII mutants with improved biosynthesis and secretion, reduced ER chaperone interaction, improved ER-Golgi transport, increased activation or resistance to inactivation and improved half-life (summarized by Pipe 2004. Sem. Thromb. Hemost. 30:227-237), and FVIII mutants having a deletion of all or part of the B-domain (see, e.g., WO 2004/067566 A1, WO 02/102850 A2, WO 00/24759 A1 and US patent No. 4,868,112). All of these factor VIII mutants and variants are incorporated herein by reference in their entirety.

The term "single-chain Factor VIII" refers to a Factor VIII molecule which has not been proteolytically cleaved into two chains (e.g. a heavy chain and a light chain) during secretion from the cells expressing said FVIII molecule and, accordingly, is present as a single polypeptide chain.

Preventing cleavage

The method of the invention comprises preventing proteolytic cleavage of the Factor VIII molecule into a first fragment comprising essentially the A1 domain and the A2 domain and a second fragment comprising essentially the A3 domain, the C1 domain and the C2 domain. The term "preventing proteolytic cleavage" includes partially preventing proteolytic cleavage and completely preventing proteolytic cleavage. It further includes the embodiment "reducing proteolytic cleavage". In other words, "preventing proteolytic cleavage of the Factor VIII molecule" does not

require completely abolishing any proteolytic cleavage such that substantially 100% of the Factor VIII molecules expressed and secreted by the host cells are single chain molecules (though this embodiment is encompassed by the method of the invention). Usually, the proteolytic cleavage of the Factor VIII molecule is prevented

5 in a manner such that at least 50%, preferably at least 60%, more preferably at least 70%, more preferably at least 80%, more preferably at least 90%, most preferably at least 95% of the Factor VIII molecules expressed and secreted by the host cells are single chain molecules. The incomplete prevention of cleavage may, at least in part, be due to the fact that there can be some minor cleavage sites

10 within the B domain which can lead to proteolytic cleavage of a small portion of the Factor VIII molecules even if the major cleavage sites (at R1313 and R1648) are absent. This minor cleavage may or may not be prevented in accordance with this invention.

15 The first fragment comprises essentially the A1 domain and the A2 domain of Factor VIII. The first fragment may comprise the A1 domain and the A2 domain, each domain having exactly the amino acid sequence indicated above. For example, the first fragment may comprise at least amino acids 1 to 740 of the amino acid sequence of SEQ ID NO:2. Alternatively, the first fragment may

20 comprise a variant of this sequence, having amino acid deletions, substitutions and/or insertions which do not substantially affect the Factor VIII activity. The first fragment may additionally comprise an N-terminal part of the B domain of Factor VIII.

25 The second fragment comprises essentially the A3 domain, the C1 domain and the C2 domain. The second fragment may comprise the A3 domain, the C1 domain and the C2 domain, each domain having exactly the amino acid sequence indicated above. For example, the second fragment may comprise at least amino acids 1690 to 2332 of the amino acid sequence shown in SEQ ID NO:2. Alternatively, the

30 second fragment may comprise a variant of this sequence, having amino acid deletions, substitutions and/or insertions which do not substantially affect the Factor

VIII activity. The second fragment may additionally comprise a C-terminal part of the acidic a3 region.

The method of the invention comprises preventing the proteolytic cleavage during 5 secretion of the recombinantly expressed FVIII molecule, which would result in a heterodimeric (two-chain) polypeptide. That is, the method includes obtaining a single-chain Factor VIII molecule. This can be achieved in various ways, e.g. by inactivating the proteolytic cleavage sites involved in the intracellular processing of the mature, one-chain FVIII into the heterodimeric FVIII eventually secreted by the 10 host cells.

In one embodiment the step of inactivating the proteolytic cleavage site between Arg1648 and Glu1649 comprises deleting one or more amino acids forming the protease recognition sequence. The cleavage site after residue 1648 is a furin-type 15 cleavage site. The recognition sequence for the protease in the Factor VIII sequence is LKRHQR. Preferably, the inactivation step comprises deleting one, two, three, four, five or more of these amino acid residues forming the recognition sequence. Preferably, the inactivation step comprises deleting at least one basic amino acid within the recognition sequence, more preferably, the inactivation step 20 comprises deleting at least the arginine at position 1648. Still more preferably, the inactivation step comprises deleting at least amino acids 1643 to 1648 of the Factor VIII sequence. If the respective FVIII derivative comprises Arg1313, the inactivation step comprises also deleting at least the arginine at position Arg 1313. Still 25 preferably is deleting at least amino acids 1313 to 1648 of the Factor VIII sequence to inactivate both cleavage sites after 1313 and 1648, respectively.

Most preferably, the inactivation step comprises deleting at least the amino acid sequence from residues 800 to 1648 from the Factor VIII sequence, e.g. the amino acid sequence from residues 741 to 1648 from the Factor VIII sequence. In another 30 preferred embodiment, a first amino acid selected from the amino acids at positions 741 to 1647 of the Factor VIII sequence is fused with a second amino acid selected

from the amino acids at positions 1649 to 1690 of the Factor VIII sequence, whereby the proteolytic cleavage during secretion is prevented. Preferred deletions are as follows:

- 5 – amino acid 740 is fused to amino acid 1650, whereby amino acids 741 to 1649 are deleted;
- amino acid 740 is fused to amino acid 1690, whereby amino acids 741 to 1689 are deleted;
- amino acid 740 is fused to amino acid 1669, whereby amino acids 741 to 1668 are deleted;
- 10 – amino acid 743 is fused to amino acid 1650, whereby amino acids 744 to 1649 are deleted;
- amino acid 764 is fused to amino acid 1650, whereby amino acids 765 to 1649 are deleted;
- 15 – amino acid 764 is fused to amino acid 1653, whereby amino acids 765 to 1652 are deleted;
- amino acid 764 is fused to amino acid 1656, whereby amino acids 765 to 1655 are deleted;
- amino acid 745 is fused to amino acid 1650, whereby amino acids 746 to 1649 are deleted;
- 20 – amino acid 745 is fused to amino acid 1653, whereby amino acids 746 to 1652 are deleted;
- amino acid 745 is fused to amino acid 1656, whereby amino acids 746 to 1655 are deleted;
- 25 – amino acid 757 is fused to amino acid 1650, whereby amino acids 758 to 1649 are deleted;
- amino acid 757 is fused to amino acid 1653, whereby amino acids 758 to 1652 are deleted;
- amino acid 757 is fused to amino acid 1656, whereby amino acids 758 to 1655 are deleted;
- 30 – amino acid 757 is fused to amino acid 1656, whereby amino acids 758 to 1655 are deleted;

- 20 -

- amino acid 793 is fused to amino acid 1649, whereby amino acids 794 to 1648 are deleted;
- amino acid 793 is fused to amino acid 1690, whereby amino acids 794 to 1689 are deleted;

5 – amino acid 747 is fused to amino acid 1649, whereby amino acids 748 to 1648 are deleted;

– amino acid 751 is fused to amino acid 1649, whereby amino acids 752 to 1648 are deleted;

– amino acid 776 is fused to amino acid 1649, whereby amino acids 777 to 1648

10 are deleted;

– amino acid 770 is fused to amino acid 1667, whereby amino acids 771 to 1666 are deleted.

15 The molecules resulting from the deletion are usually obtained in the form of single chain Factor VIII molecules.

Preferred single chain FVIII molecules have a deletion of all or part of the B-domain and a deletion of all or a part of the acidic a3 region, so that the cleavage site at Arg1648 (which is usually cleaved during secretion) is deleted. Single chain FVIII

20 molecules are disclosed in, e.g., WO 2004/067566 A1; US 2002/132306 A1; Krishnan et al. (1991) European Journal of Biochemistry vol. 195, no. 3, pages 637-644; Herlitschka et al. (1998) Journal of Biotechnology, vol. 61, no. 3, pages 165-173; Donath et al. (1995) Biochem. J., vol. 312, pages 49-55. These single-chain Factor VIII molecules described in these references are incorporated herein by

25 reference.

30 The fusions referred to above may be direct fusions or indirect fusions. In the latter case, the deleted amino acids are replaced by a heterologous spacer. This embodiment is described in more detail hereinafter. It is possible that the deleted amino acids are replaced with a peptidic linker consisting of about 1 to about 500

amino acids, or about 2 to 250 amino acids, or about 3 to about 100 amino acids, or about 4 to about 50 amino acids, or about 5 to about 10 amino acids. The peptidic linker should be flexible and not immunogenic (Robinson et al.; PNAS (1998), Vol 95, p5929). The peptidic linkers may consist of Gly preceded N-terminally to said

5 Gly by multimers of the amino acid sequence GlyGlySer or GlyGlySerSer or any combination thereof, in a specific embodiment the peptidic linker consists of 80 to 120 amino acids.

In an alternative embodiment, one or more amino acids which form the protease 10 recognition site at residue 1313 and 1648 may be substituted with another amino acid such that the cleavage does not occur. For example, the basic amino acids may be replaced with hydrophobic amino acids.

Preparation of single-chain Factor VIII

15 The step of "preventing proteolytic cleavage" or "inactivating a proteolytic cleavage site" is carried out prior to the purification, lyophilisation and reconstitution of the Factor VIII. The step of "preventing proteolytic cleavage" or "inactivating a proteolytic cleavage site" is typically carried out during the preparation of the Factor VIII molecule. The method of the invention may include preventing the proteolytic 20 cleavage during expression of the Factor VIII molecule (in host cells), or inactivating the proteolytic cleavage site at Arg1313 and/or Arg1648 during the preparation of the nucleic acid encoding the Factor VIII molecule.

These steps of "preventing proteolytic cleavage" or "inactivating a proteolytic 25 cleavage site" may include removing, from a nucleic acid encoding Factor VIII, a portion encoding the proteolytic cleavage site at Arg1313 and/or Arg1648, in accordance with the embodiments described above. This typically results in a nucleic acid encoding single chain Factor VIII. Generally, the method of the

invention may further include providing a nucleic acid encoding the single-chain Factor VIII, e.g. in an expression plasmid or vector.

The nucleic acid, the expression vector or the expression plasmid may then be
5 introduced into host cells, preferably mammalian host cells, for expression. The method of the invention may further comprise culturing the host cells under suitable conditions such that the modified Factor VIII molecule, e.g. the single chain Factor VIII molecule, is expressed; and optionally recovering (e.g. purifying) the modified Factor VIII molecule from the host cells or from the culture medium. Generally,
10 techniques of manipulating the nucleic acid encoding Factor VIII, of culturing mammalian cells to allow expression of the Factor VIII, and of purifying Factor VIII from the cell culture medium are known in the art.

It is preferred to purify the single chain Factor VIII molecule to $\geq 80\%$ purity, more
15 preferably $\geq 95\%$ purity and particularly preferred is a pharmaceutically pure state that is greater than 99.9% pure with respect to contaminating macromolecules, particularly other proteins or/and nucleic acids, and free of infectious and pyrogenic agents. Preferably, an isolated or purified modified Factor VIII molecule is substantially free of other polypeptides.

20

The methods of the invention may further comprise the steps of purifying, lyophilizing, and reconstituting the single chain Factor VIII. The reconstitution is preferably carried out by using water, e.g. "water for injection".

25 *Stability*

The Factor VIII molecules prepared in accordance with the present invention exhibit enhanced stability relative to full length Factor VIII and/or relative to a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640 (i.e. a B-domain
30 deleted Factor VIII molecule consisting essentially of amino acids 1-745 and 1640-2332 of SEQ ID NO:2).

As used herein, the term "stability" refers to stability in aqueous solution, preferably to stability in aqueous solution after reconstitution of a lyophilized Factor VIII preparation, e.g. by adding water to the lyophilized Factor VIII preparation.

5 Typically, the lyophilized Factor VIII preparation is reconstituted with "water for injection".

The stability in aqueous solution can be determined by providing the Factor VIII molecule in aqueous solution and incubating it for a certain period of time. In a 10 preferred embodiment, the conditions for determining the storage stability of the Factor VIII molecule are as follows:

The Factor VIII molecule is provided in aqueous solution having the following composition:

15	L-histidine	25 mM
	NaCl	225 mM
	calcium chloride	4 mM
	Tween® 80	0.03% (w/w)
	sucrose	2% (w/w)
20	D-mannitol	8% (w/w)
	pH 7.0.	

This solution is referred to hereinafter as "Buffer A". The initial Factor VIII activity in the aqueous solution is preferably between 100 IU/ml and 1,500 IU/ml, preferably it 25 is 100 IU/ml.

The so prepared Factor VIII solution can then be incubated at 25°C for at least 24 hours, preferably for at least two days, more preferably for at least five days, most 30 preferably for seven or eight days. After the incubation period the stability is determined by measuring the Factor VIII activity in the solution, preferably by using a chromogenic substrate assay (e.g. Coamatic® Factor VIII, Chromogenix). The

lower the loss in activity relative to the initial activity, the higher is the stability of the Factor VIII molecule. Most preferably, the stability is determined as in Example 1 or 2 below.

5 According to the present invention the loss in Factor VIII activity of the single-chain Factor VIII after seven days of storage under the above-identified conditions is less than 15%, preferably less than 12%, most preferably less than 10%.

Typically, the initial Factor VIII activity at the start of the incubation period (t=0) is 10 normalized to 100%. The remaining Factor VIII activity after 24 hours of storage in Buffer A at 25°C is preferably at least 95% of the initial Factor VIII activity. The remaining Factor VIII activity after 48 hours of storage in Buffer A at 25°C is preferably at least 95% of the initial Factor VIII activity. The remaining Factor VIII activity after 4 days of storage in Buffer A at 25°C is preferably at least 90%, more 15 preferably at least 95% of the initial Factor VIII activity. The remaining Factor VIII activity after 7 days of storage in Buffer A at 25°C is preferably at least 85%, more preferably at least 90%, most preferably at least 95% of the initial Factor VIII activity. The remaining Factor VIII activity after 8 days of storage in Buffer A at 25°C is preferably at least 85%, more preferably at least 90%, most preferably at least 20 95% of the initial Factor VIII activity.

The remaining Factor VIII activity of the single chain Factor VIII is usually higher than that of two-chain Factor VIII molecules (assuming that both molecules have been incubated under identical conditions for the same period of time).

25 The term "human full length two-chain Factor VIII" is used herein interchangeably with the term "human wild-type Factor VIII".

In one embodiment, the remaining Factor VIII activity of the single chain Factor VIII 30 is higher than that of human full length two-chain Factor VIII. In another embodiment, the remaining Factor VIII activity of the single chain Factor VIII is

higher than that of a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640 (i.e. a B-domain deleted Factor VIII molecule consisting essentially of amino acids 1-745 and 1640-2332 of SEQ ID NO:2).

5 Preferably, the remaining Factor VIII activity of the single chain Factor VIII after 48 hours of storage in Buffer A at 25°C exceeds the remaining Factor VIII activity of human full length two-chain Factor VIII by at least 4 percentage points. It is also preferred that the remaining Factor VIII activity of the single chain Factor VIII after 48 hours of storage in Buffer A at 25°C exceeds the remaining Factor VIII activity of 10 a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640 (i.e. a B-domain deleted Factor VIII molecule consisting essentially of amino acids 1-745 and 1640-2332 of SEQ ID NO:2) by at least 4 percentage points.

In another embodiment, the remaining Factor VIII activity of the single chain Factor 15 VIII after 4 days of storage in Buffer A at 25°C exceeds the remaining Factor VIII activity of human full length two-chain Factor VIII by at least 5 percentage points. It is also preferred that the remaining Factor VIII activity of the single chain Factor VIII after 4 days of storage in Buffer A at 25°C exceeds the remaining Factor VIII activity of a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640 (i.e. 20 a B-domain deleted Factor VIII molecule consisting essentially of amino acids 1-745 and 1640-2332 of SEQ ID NO:2) by at least 5 percentage points.

In another embodiment, the remaining Factor VIII activity of the single chain Factor VIII after 7 days of storage in Buffer A at 25°C exceeds the remaining Factor VIII 25 activity of human full length two-chain Factor VIII by at least 5, preferably by at least 10 percentage points. It is also preferred that the remaining Factor VIII activity of the single chain Factor VIII after 7 days of storage in Buffer A at 25°C exceeds the remaining Factor VIII activity of a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640 (i.e. a B-domain deleted Factor VIII molecule consisting 30 essentially of amino acids 1-745 and 1640-2332 of SEQ ID NO:2) by at least 5, preferably by at least 10 percentage points.

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In another embodiment, the remaining Factor VIII activity of the single chain Factor VIII after 8 days of storage in Buffer A at 25°C exceeds the remaining Factor VIII activity of human full length two-chain Factor VIII by at least 5, preferably by at least 5 10 percentage points. It is also preferred that the remaining Factor VIII activity of the single chain Factor VIII after 8 days of storage in Buffer A at 25°C exceeds the remaining Factor VIII activity of a B-domain deleted Factor VIII molecule where Asn745 is fused to Pro1640 (i.e. a B-domain deleted Factor VIII molecule consisting essentially of amino acids 1-745 and 1640-2332 of SEQ ID NO:2) by at least 5, 10 preferably by at least 10 percentage points.

Alternatively to Buffer A other buffers may also be used like, for example the buffer used in Example 2 of the present invention.

15 The preferred pH range for the buffers of the present invention is a pH range from 5.5 to 9.0, preferably a pH range from 6.0 to 8.5 and especially preferred a pH range from 6.5 to 8.0.

The activity of Factor VIII can be determined by a chromogenic or clotting assay, or any other bioassay. Preferably, the Factor VIII activity is determined as shown in 20 Example 1 below.

Bioavailability

In another embodiment, the Factor VIII molecule stabilized in accordance with the 25 present invention exhibits improved bioavailability after non-intravenous injection, as compared to two chain human wild type Factor VIII or compared to two chain human B-domain deleted Factor VIII. The non-intravenous injection is preferably subcutaneous, transdermal or intramuscular injection. Most preferably, the non-intravenous injection is subcutaneous injection.

The term „bioavailability”, as used herein, refers to the proportion of an administered dose of a Factor VIII or a FVIII-related preparation that can be detected in plasma at predetermined times until a final time point after subcutaneous, intravenous or intradermal administration. Typically, bioavailability is

5 measured in test animals by administering a dose of between 10 IU/kg and 1000 IU/kg of the preparation (e.g. 400 IU/kg body weight); obtaining plasma samples at pre-determined time points after administration; and determining the content of the Factor VIII or Factor VIII-related polypeptides in the samples using one or more of a chromogenic or clotting assay (or any bioassay), an immunoassay, or an equivalent

10 thereof. The bioavailability is expressed as the area under the curve (AUC) of the concentration or activity of the coagulation factor in plasma on the y-axis and the time after administration on the x-axis until a predefined final time point after administration. Preferably, this predefined time point is 72 or 48 hours after administration. Most preferably, the bioavailability is determined as shown in

15 Example 3 herein below. Relative bioavailability of a test preparation refers to the ratio between the AUC of the test preparation (here: single chain Factor VIII) and that of the reference preparation (e.g. full length recombinant two-chain Factor VIII or two-chain B-domain deleted Factor VIII) which is administered in the same dose and way (e.g. intravenous, subcutaneous or intradermal) as the test preparation.

20 According to the present invention, the bioavailability of the single chain Factor VIII after subcutaneous injection is higher than that of the two-chain human wild type Factor VIII or of two-chain human B-domain deleted Factor VIII. Preferably, the bioavailability (AUC over 72 hours after subcutaneous injection) is increased by at

25 least 10%, more preferably by at least 25%, more preferably by at least 50%, most preferably by at least 75%, relative to wild type FVIII. In another embodiment, the bioavailability (AUC over 72 hours after subcutaneous injection) is increased by at least 10%, more preferably by at least 20%, more preferably by at least 30%, most preferably by at least 40%, relative to a B-domain deleted Factor VIII molecule

30 where Asn745 is fused to Pro1640 (i.e. a B-domain deleted Factor VIII molecule consisting essentially of amino acids 1-745 and 1640-2332 of SEQ ID NO:2).

Improvement of plasma half-life (in -vivo)

In another embodiment, the Factor VIII molecule stabilized in accordance with the

5 present invention exhibit increased pharmacokinetic (PK) parameters.

Factor VIII molecules of the invention can be tested by i.v. injection into different species like hemophilia A mice or cynomolgus monkeys e.g. at a dose of 100 IU/kg or 250 IU/kg respectively e.g. as determined in a chromogenic assay. Blood

10 samples are drawn at various time points after administration e.g. until 72 hours (hrs) in hemophilia A mice and e.g. until 24 hrs in cynomolgus monkeys. Citrate plasma is prepared immediately and used for quantification of FVIII:C e.g. by a chromogenic assay system (FVIII:C) (Chromogenix - Instrumentation Laboratory SpA, Milan, Italy).

15

The AUC of the FVIII levels in plasma is calculated using the linear trapezoidal rule to calculate AUC_{last} : from $t=0$ to last observation. Terminal half-life ($t_{1/2\beta}$) is determined by a log-linear regression using the points of the terminal phase selected by the adjusted R2 criterion. AUC: from $t=0$ to infinity (extrapolated by 20 using the regression model of the terminal phase).

The single chain FVIII molecules according to the invention show at least a 40%, preferably at least a 50%, even more preferably at least a 60% increased terminal half life as compared to the terminal half-life a human wild-type Factor VIII 25 administered at the same dose and in the same manner.

Preferably the plasma half-life is determined as shown in Example 5.

Prolongation of efficacy as determined in a thrombin generation assay (in-vivo)

In another embodiment, the Factor VIII molecule stabilized in accordance with the present invention exhibit a longer time period for the thrombin peak level as

5 determined in a thrombin generation assay over time in hemophilia A mice to fall below 50 nM after intravenous administration relative to human wild type Factor VIII. This test show that also the functionality of FVIII is stabilized in the molecules according to the invention.

10 FVIII molecules according to the invention can be tested by first administering the FVIII molecule of the invention at an equimolar dose (e.g. at 250 IU/kg) intravenously into hemophilia A mice. At different time points (e.g. daily from day 1 to 8) citrated blood is collected and a thrombin generation assay (TGA) is performed e.g. by calibrated thrombinography (CAT) (Thrombinoscope, 15 Netherlands) after intrinsic activation in presence of Phospholipid (e.g. Rossix, Mölndal, Sweden) / Pathromtin® SL (Siemens Healthcare Diagnostics Products GmbH, Marburg, Germany) (1:30). Thrombin peak levels are recorded. The average AUC of peak thrombin levels from days 1-8 is calculated by the linear trapezoidal rule. The AUC of the two Factor VIII products are compared using an 20 approximate F-test for the difference in AUC in a linear model with variable variances per time-point and treatment group resulting in a estimated time until peak levels of thrombin drop below a defined limit ranging of 50 nM.

25 Preferably the efficacy in a thrombin generation assay is determined as shown in Example 6.

In hemophilia A mice scFVIII shows a favorable hemostatic activity compared to human wild-type Factor VIII. This translates into an averaged at least 10 hrs longer, preferably at least 15 hours longer and even more preferred at least 20 hours

30 longer thrombin generation activity value for scFVIII versus full-length rFVIII before the thrombin peak level falls below a level of 50 nM.

Retaining higher FVIII:C activity in plasma (ex vivo)

In another embodiment, the Factor VIII molecule stabilized in accordance with the present invention retain a higher activity as determined by a one-stage FVIII:C assay after having been incubated for 4 days in human plasma at 37°C relative to human wild type Factor VIII after having been incubated for 4 days in human plasma at 37°C; preferably wherein the retained activity of the Factor VIII is at least 10% higher relative to that of a human wild type Factor VIII after having been incubated for 4 days in human plasma at 37°C.

10 Samples with Factor VIII molecules according to the invention can be tested by diluting them into with FVIII deficient plasma (e.g. from Siemens Healthcare Diagnostics) to 1 IU/mL FVIII:C (based on values determined by the chromogenic substrate assay). The FVIII- samples are then incubated at 37°C for varying time periods (e.g. for 0, 0.25, 1, 2, 4 and 8 days) in presence of 0.05% Na-azide. After each incubation period, FVIII:C is then determined by a one-stage-coagulation assay e.g. by using Pathromtin-SL (Siemens Healthcare Diagnostics) as activator, normalized to the value at t= 0 (% FVIII:C) and plotted versus the incubation time.

15 20 After a 4 day incubation at 37°C the Factor VIII molecule of the invention has retained at least a 10% higher FVIII:C activity, preferably at least 15% higher FVIII:C activity, preferably at least a 20% higher FVIII:C activity, preferably at least a 25% higher FVIII:C activity, preferably at least a 30% higher FVIII:C activity.

25 Preferably the activity in plasma is determined as shown in Example 4.

Treatment and prophylaxis

The single-chain Factor VIII constructs in accordance with the present invention having increased stability after reconstitution can be administered in the treatment or prophylaxis of bleeding disorders.

As used herein, the term "bleeding disorders" includes familial and acquired hemophilia A and B, familial or acquired von Willebrand disease, familial or acquired deficiency of any coagulation factor, all types of trauma, blunt or penetrating, leading to severe hemorrhage either from a single organ, a bone

5 fraction or from polytrauma, bleeding during surgical procedures including peri- or postoperative haemorrhage, bleeding due to cardiac surgery including patients undergoing extracorporeal circulation and hemodilution in pediatric cardiac surgery, intracerebral hemorrhage, subarachnoid hemorrhage, sub- or epidural bleeding, bleedings due to blood loss and hemodilution, by non-plasmatic volume substitution

10 leading to reduced levels of coagulation factors in affected patients, bleedings due to disseminated intravascular coagulation (DIC) and a consumption coagulopathy, thrombocyte dysfunctions, depletion and coagulopathies, bleeding due to liver cirrhosis, liver dysfunction and fulminant liver failure, liver biopsy in patients with liver disease, bleeding after liver and other organ transplantations, bleeding from

15 gastric varices and peptic ulcer bleeding, gynaecological bleedings as dysfunctional uterine bleeding (DUB), premature detachment of the placenta, periventricular haemorrhage in low birth weight children, post partum haemorrhage, fatal distress of newborns, bleeding associated with burns, bleeding associated with amyloidosis, hematopoietic stem cell transplantation associated with platelet disorder, bleedings

20 associated with malignancies, infections with haemorrhaging viruses, bleeding associated with pancreatitis.

The components of the pharmaceutical preparation may be dissolved in conventional physiologically compatible aqueous buffer solutions to which there

25 may be added, optionally, pharmaceutical excipients to provide the pharmaceutical preparation. The components of the pharmaceutical preparation may already contain all necessary pharmaceutical, physiologically compatible excipients and may be dissolved in water for injection to provide the pharmaceutical preparation.

30 Such pharmaceutical carriers and excipients as well as the preparation of suitable pharmaceutical formulations are well known in the art (see for example

“Pharmaceutical Formulation Development of Peptides and Proteins”, Frokjaer et al., Taylor & Francis (2000) or “Handbook of Pharmaceutical Excipients”, 3rd edition, Kibbe et al., Pharmaceutical Press (2000)). In certain embodiments, a pharmaceutical composition can comprise at least one additive such as a bulking agent, buffer, or stabilizer. Standard pharmaceutical formulation techniques are well known to persons skilled in the art (see, e.g., 2005 Physicians’ Desk Reference®, Thomson Healthcare: Montvale, NJ, 2004; Remington: The Science and Practice of Pharmacy, 20th ed., Gennaro et al., Eds. Lippincott Williams & Wilkins: Philadelphia, PA, 2000). Suitable pharmaceutical additives include, e.g., sugars like mannitol, sorbitol, lactose, sucrose, trehalose, or others, amino acids like histidine, arginine, lysine, glycine, alanine, leucine, serine, threonine, glutamic acid, aspartic acid, glutamine, asparagine, phenylalanine, or others, additives to achieve isotonic conditions like sodium chloride or other salts, stabilizers like Polysorbate 80, Polysorbate 20, Polyethylene glycol, propylene glycol, calcium chloride, or others, physiological pH buffering agents like Tris(hydroxymethyl)aminomethan, and the like. In certain embodiments, the pharmaceutical compositions may contain pH buffering reagents and wetting or emulsifying agents. In further embodiments, the compositions may contain preservatives or stabilizers. In particular, the pharmaceutical preparation comprising the blood coagulation factor may be formulated in lyophilized or stable soluble form. The blood coagulation factor may be lyophilized by a variety of procedures known in the art. Lyophilized formulations are reconstituted prior to use by the addition of one or more pharmaceutically acceptable diluents such as sterile water for injection or sterile physiological saline solution or a suitable buffer solution.

25 The composition(s) contained in the pharmaceutical preparation of the invention may be delivered to the individual by any pharmaceutically suitable means. Various delivery systems are known and can be used to administer the composition by any convenient route. Preferably, the composition(s) contained in the pharmaceutical preparation of the invention are delivered to the individual by non-intravenous injection. More preferably, the composition(s) of the invention are formulated for

subcutaneous, intramuscular, intraperitoneal, intracerebral, intrapulmonar, intranasal, intradermal or transdermal administration, most preferably for subcutaneous, intramuscular or transdermal administration according to conventional methods. The formulations can be administered continuously by 5 infusion or by bolus injection. Some formulations may encompass slow release systems.

The composition(s) of the pharmaceutical preparation of the present invention is/are administered to patients in a therapeutically effective dose, meaning a dose that is 10 sufficient to produce the desired effects, preventing or lessening the severity or spread of the condition or indication being treated without reaching a dose which produces intolerable adverse side effects. The exact dose depends on many factors as e.g. the indication, formulation, mode of administration and has to be determined in preclinical and clinical trials for each respective indication.

15

In one embodiment of the invention, the plasma level of the coagulation factor in the treated subject is, during a period from 5 hours after injection to 8 hours after non-intravenous injection, continuously higher than 2%, preferably higher than 5%, more preferably higher than 8%, most preferably higher than 10%, of the normal plasma 20 level of the coagulation factor in healthy subjects. The plasma level is to be determined as shown hereinafter in Example 3.

In one embodiment of the invention, the plasma level of the coagulation factor in the treated subject is, during a period from 4 hours after injection to 16 hours after non-intravenous injection, continuously higher than 2%, preferably higher than 5%, more 25 preferably higher than 8%, most preferably higher than 10%, of the normal plasma level of the coagulation factor in healthy subjects.

In another embodiment of the invention, the plasma level of the coagulation factor 30 in the treated subject is, during a period from 3 hours after injection to 24 hours after non-intravenous injection, continuously higher than 2%, preferably higher than

4%, more preferably higher than 6%, most preferably higher than 8%, of the normal plasma level of the coagulation factor in healthy subjects.

In another embodiment of the invention, the plasma level of the coagulation factor

5 in the treated subject is, during a period from 2 hours after injection to 32 hours after non-intravenous injection, continuously higher than 2%, preferably higher than 3%, more preferably higher than 4%, most preferably higher than 5%, of the normal plasma level of the coagulation factor in healthy subjects.

10 Preferably, the dose of single-chain Factor VIII for one non-intravenous injection is less than 1,000 IU/kg body weight, or less than 800 IU/kg body weight, or less than 600 IU/kg body weight, or less than 400 IU/kg body weight, e.g. at a dose of from about 10 IU/kg body weight to about 1,000 IU/kg body weight, or from about 20 IU/kg body weight to about 800 IU/kg body weight, or from about 30 IU/kg body weight to about 700 IU/kg body weight, or from about 40 IU/kg body weight to about 600 IU/kg body weight, or from about 50 IU/kg body weight to about 500 IU/kg body weight, or from about 75 IU/kg body weight to about 400 IU/kg body weight, or from about 100 IU/kg body weight to about 300 IU/kg body weight, or from about 50 IU/kg body weight to about 1,000 IU/kg body weight, or from about 50 IU/kg body weight to about 800 IU/kg body weight, or from about 50 IU/kg body weight to about 700 IU/kg body weight, or from about 50 IU/kg body weight to about 600 IU/kg body weight, or from about 50 IU/kg body weight to about 500 IU/kg body weight, or from about 50 IU/kg body weight to about 400 IU/kg body weight, or from about 50 IU/kg body weight to about 300 IU/kg body weight, or about 50 IU/kg body weight to about 200 IU/kg body weight. The FVIII can be administered on its own, or as a complex with VWF.

30 The pharmaceutical composition(s) of the invention may be administered alone or in conjunction with other therapeutic agents. These agents may be incorporated as part of the same pharmaceutical.

Examples

Example 1: Stability of purified Factor VIII molecules after reconstitution

5

The following Factor VIII preparations were used in this Example:

10 Beriate[®], a lyophilized human coagulation Factor VIII concentrate, was obtained from CSL Behring GmbH. Beriate[®] comprises plasma-derived Factor VIII in heterodimeric form.

Helixate[®], a lyophilized, recombinant coagulation Factor VIII was obtained from CSL Behring GmbH. Helixate[®] contains recombinantly produced heterodimeric Factor VIII.

15

ReFacto[®] is a lyophilized Factor VIII preparation containing heterodimeric, B-domain-deleted Factor VIII produced by recombinant technology. It can be obtained from, e.g., Pfizer Pharma GmbH, Germany.

20 Beriate[®], Helixate[®], and ReFacto[®] are predominantly heterodimeric two-chain polypeptides.

25 The construct termed "scFVIII" is a single-chain Factor VIII produced by recombinant expression in mammalian cell culture cells. The single-chain Factor VIII used in this Example was obtained by directly fusing Asn764 with Thr1653, and provided in lyophilized form after purification. That is, "scFVIII" is a single chain polypeptide consisting substantially of amino acids 1-764 and 1653-2332 of SEQ ID NO:2.

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Beriate®, Helixate®, and ReFacto® were reconstituted according the manufacturer's instructions as given in the package insert. "scFVIII" was reconstituted by dissolving the purified and lyophilized FVIII preparation in water for injection resulting in a composition containing 25 mM L-histidine, 225 mM NaCl, 4 mM CaCl₂, 0.03 %

5 Tween 80, 2% sucrose, 8% D-mannitol, pH 7.0.

The reconstituted FVIII products were incubated at 25°C. The FVIII activity of the products was determined in duplicates by a chromogenic substrate assay (Coamatic® Factor VIII, Chromogenix) at the following time points: 0 h, 6h, 1day, 2

10 days, 4 days, 7 days. Activity values were normalized to time point 0.

The results are shown in the following Table and in Figure 1.

Table 1: Factor VIII activity over time

Time (days)	0 d	0.25 d	1 d	2 d	4 d	7 d
Beriate®	100.0	103.8	92.7	97.9	90.0	82.7
Helixate®	100.0	101.4	103.3	95.5	85.0	78.6
ReFacto®	100.0	106.7	83.7	94.0	93.6	77.3
scFVIII	100.0	107.8	99.4	102.0	99.0	93.3

15

As can be seen, "scFVIII" shows the lowest loss in activity and, consequently, is the most stable Factor VIII molecule.

Example 2: Stability of purified Factor VIII molecules after reconstitution

20

The following Factor VIII preparations were used in this Example:

ReFacto® and "scFVIII" were the same as used in Example 1, with the difference that "scFVIII", provided by CSL Behring GmbH, was applied in a formulation 25 containing different excipients. Advate® is a full-length, heterodimeric, recombinant Factor VIII preparation which was purchased in lyophilized form from Baxter.

Advate® and ReFacto® were reconstituted according the manufacturer's instructions as given in the package insert. "scFVIII" was reconstituted in water for injection resulting in a composition containing 20 mM L-histidine, 280 mM NaCl, 3.4 mM CaCl₂, 0.02 % Tween 80, 0.6% sucrose, pH 7.0. The sample "scFVIII 001" had 5 an initial FVIII activity of 100 IU/ml, the sample "scFVIII 0006" had an initial FVIII activity of 400 IU/ml. The reconstituted FVIII products were incubated at 25°C. The FVIII activity of the products was determined in duplicates by a chromogenic substrate assay (Coamatic® Factor VIII, Chromogenix) at the following time points: 0 h, 6h, 1day, 2 days, 4 days, 8 days. Activity values were normalized to time point 10 0.

The results are shown in the following Table and in Figure 2.

Table 2: Factor VIII activity over time

Time (days)	0 d	0.25 d	1 d	2 d	4 d	8 d
Advate	100.0	94.9	100.6	89.8	74.1	73.7
ReFacto	100.0	92.0	88.8	92.1	74.3	80.6
scFVIII 001	100.0	99.2	95.6	99.2	89.0	95.1
scFVIII 006	100.0	96.8	97.6	100.0	88.9	92.3

15

As can be seen, "scFVIII" shows the lowest loss in activity and, consequently, is the most stable Factor VIII molecule.

20

Example 3: Bioavailability of Factor VIII molecules

Advate®, ReFacto® and "scFVIII" were the same as used in Example 2 and reconstituted as described in Example 2.

25 Factor VIII knockout mice were used as animal model for hemophilia A. These mice lack exons 16 and 17 and thus do not express FVIII (Bi L. et al, Nature genetics,

1995, Vol 10(1), 119-121; Bi L. et al, Blood, 1996, Vol 88(9), 3446-3450). This allows the analysis of FVIII levels following treatment by quantification of FVIII activity in the plasma of the ko mice.

To assess whether extravascular injections might be an option for an improved

5 therapy with human FVIII, subcutaneous injection was chosen. The design of the non-clinical pharmacokinetic study performed is detailed in table 3 below. Plasma levels of Factor VIII activity were determined following a single subcutaneous injection of the respective FVIII preparation (detailed treatment groups in table 3) to a hemophilia A model.

10 Corresponding groups were treated with the same dose of FVIII:chromogen activity. For a single application the Factor VIII was provided in a volume of 200 µL (identical volumes for all groups) prior to subcutaneous application to FVIII knockout (ko) mice weighing about 25 g. The treatment groups are summarized in table 3.

15 Under short term anesthesia, blood samples were drawn, anticoagulated using sodium citrate to 10 % citrate blood, processed to plasma and stored at -70°C for the determination of FVIII activity. The sampling time points are detailed in table 4. Quantification of FVIII activity in plasma was performed by a standard, aPTT based approach (Behring Coagulation Timer). The animals were kept at standard housing

20 conditions.

Table 3: Treatment groups

No.	Treatment	FVIII:chromogen / Additive Dose	volume [mL/kg]	schedule	route	N
1	Advate®	400 IU/kg	8	single injection (t=0)	s.c.	25
2	ReFacto®	400 IU/kg	8	single injection (t=0)	s.c.	25
3	"scFVIII"	400 IU/kg	8	single injection (t=0)	s.c.	20

Results

5 The results are summarized in Table 4 and Figure 3.

Table 4:

timepoint [hours]	scFVIII			ReFacto®			Advate®		
	mean	SD	n	mean	SD	n	mean	SD	n
1	5.83	1.84	5	6.35	7.62	5	3.82	3.30	5
4	11.53	5.35	5	4.77	3.47	5	3.16	2.72	5
8	20.03	9.91	5	9.81	6.06	5	5.52	3.92	5
16	7.75	4.78	5	4.91	1.98	5	2.77	2.10	5
24	3.00	3.25	5	0.66	0.69	5	0.31	0.68	5
32	3.30	2.36	5	2.11	1.53	5	3.13	1.78	5
48	3.71	1.52	5	6.01	5.28	5	4.15	1.95	5
72	1.23	2.63	5	0.00	0.00	5	0.00	0.00	5
AUC _{0-72h} (h × % of the norm)	383.9			275.1			195.1		

10 Subcutaneous injection of 400 IU/kg single chain FVIII ("scFVIII") to FVIII ko mice resulted in a significant increase of FVIII activity in plasma level as compared to administration of heterodimeric full length FVIII (Advate®) or heterodimeric B-

domain-deleted FVIII (ReFacto®). That is, the single chain Factor VIII molecule shows the highest in vivo bioavailability after subcutaneous injection into mice. The two chain full length construct Advate®, as well as the two chain B-domain deleted preparation ReFacto® showed substantially lower bioavailability.

5

Example 4: Stability of Factor VIII molecules in plasma (in-vitro)

Different FVIII products (Advate®, ReFacto AF® and two lots of scFVIII as used in Example 2) were diluted with FVIII deficient plasma (Siemens Healthcare 10 Diagnostics) to 1 IU/mL FVIII:C (based on values determined by the chromogenic substrate assay). The FVIII- samples were incubated at 37°C for varying time periods (0, 0.25, 1, 2, 4 and 8 days) in presence of 0.05% Na-azide. After each 15 incubation period, FVIII:C was determined by one-stage-coagulation assay using Pathromtin-SL (Siemens Healthcare Diagnostics) as activator, normalized to the value at t= 0 (% FVIII:C) and plotted versus the incubation time. The values shown represent the average and standard deviation of two samples (except 0.25 days only one sample).

Table 5: Average % Activity Compared to Time 0

20

-	<u>0 d</u>	<u>0,25 d</u>	<u>1 d</u>	<u>2 d</u>	<u>4 d</u>	<u>8 d</u>
Advate	100,00	90,63	81,25	79,69	70,31	59,38
ReFacto	100,00	92,31	86,15	83,08	70,77	58,46
scFVIII 001	100,00	93,33	93,33	96,67	93,33	93,33
scFVIII 006	100,00	96,88	96,88	93,75	93,75	90,63

Example 5: Stability of Factor VIII molecules in plasma (in-vivo)

The pharmacokinetic (PK) profiles of scFVIII and full-length rFVIII (Advate®, Baxter Healthcare) was determined following a single I.V. injection to cynomolgus monkeys (Figure 5 and Table 6) and hemophilia A mice (Figure 6 and Table 7) at doses of 250 IU/kg and 100 IU/kg, respectively. Test items were dosed according to labeled activity for Advate® and chromogenic activity (FVIII:C) for scFVIII. Blood samples were drawn predose (monkeys only) and at various time points after administration until 72 hours (hrs) in hemophilia A mice and until 24 hrs in cynomolgus monkeys. Citrate plasma was prepared immediately and used for quantification of FVIII:C by a chromogenic assay system (FVIII:C) (Chromogenix - Instrumentation Laboratory SpA, Milan, Italy).

The AUC of the FVIII levels in plasma was calculated using the linear trapezoidal rule to calculate AUC_{last} : from $t=0$ to last observation. Terminal half-life ($t_{1/2\beta}$) was determined by a log-linear regression using the points of the terminal phase selected by the adjusted R² criterion. AUC: from $t=0$ to infinity (extrapolated by using the regression model of the terminal phase).

In cynomolgus monkeys scFVIII showed a ~1.6 fold enhanced $AUC_{0-t_{last}}$ or $t_{1/2\beta}$ with a correspondingly ~2 fold lower clearance (CL), while FVIII activity peak levels (C_{max}), representative of in vivo recovery (IVR), and volume of distribution at steady state (V_{ss}) appeared more similar versus full-length rFVIII. These PK parameter results were obtained from n=10 animals after toxicokinetic data from 8 additional monkeys, when dosed during the GLP- toxicity studies with 250 IU/kg of scFVIII, were included (Table 6 and Figure 5).

In hemophilia A mice enhancement of $AUC_{0-t_{last}}$, of mean residence time (MRT), time until 5% FVIII activity trough levels, terminal half-life and a correspondingly lower CL ranged between 1.6-2 fold for scFVIII, while C_{max} , representative of IVR, and V_{ss} appeared similar versus full-length rFVIII. $AUC_{0-t_{last}}$ and $t_{1/2\beta}$ results

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obtained after rVIII-SingleChain treatment were significantly better than for full-length rFVIII with an $AUC_{0-t_{last}}$ ratio of 1.97 (90% confidence interval (CI): 1.7-2.3; p-value(ratio=1): < 0.0001), and a $t_{1/2\beta}$ ratio of 1.65 (90% CI: 1.11-2.70; p-value(ratio=1): 0.036 (Table 7 and Figure 6).

5

Table 6: PK parameters of scFVIII and full-length rFVIII in cynomolgus monkeys

Parameters	scFVIII (n=10)	Full-length rFVIII (n=2)
$AUC_{0-t_{last}}$ (hrs·IU/mL)	78.4	49.1
C_{max} (IU/mL)	7.8	8.7
CL ((mL/hrs)/kg)	2.1	4.7
$t_{1/2\beta}$ (hrs)	11.0	6.8

10

Table 7: PK parameters of scFVIII and full-length rFVIII in hemophilia A mice

Parameters	scFVIII	Full-length rFVIII
AUC _{0-last} (hrs·IU/mL)	35	18
C _{max} (IU/mL)	2.3	2.2
CL ((mL/hrs)/kg)	2.7	5.5
MRT (hrs)	18	10
V _{ss} (mL/kg)	50	57
t _{1/2β} (hrs)	15.9	9.7
Time until 0.05 IU/mL (hrs)	73	39

5

Both sets of PK parameters reflect the increased stability of scFVIII after purification, lyophilization, reconstitution *in vivo* in plasma after administration to the two animal species tested.

10

Example 6: Thrombin generation assay in hemophilia A mice (ex vivo)

Citrate- (10% v/v) hemophilia A mouse blood was terminally collected under deep anesthesia at different time-points (days 1-8) when scFVIII or full-length rFVIII (Advate[®]), were dosed @ a level of 250 IU/kg. TGA was performed by calibrated thrombinography (CAT, Thrombinoscope, Netherlands) after intrinsic activation in presence of Phospholipid (Rossix, Mölndal, Sweden) / Pathromtin[®] SL (Siemens Healthcare Diagnostics Products GmbH, Marburg, Germany) (1:30). Thrombin

peak levels were recorded. The average AUC of peak thrombin levels from days 1-8 was calculated by the linear trapezoidal rule. The AUC of the two Factor VIII products were compared using an approximate F-test for the difference in AUC in a linear model with variable variances per time-point and treatment group resulting in 5 estimated time until peak levels of thrombin drop below a defined limit ranging between 50-250 nM.

In hemophilia A mice scFVIII showed a favorable hemostatic activity compared to full-length rFVIII as indicated by the estimated time until peak levels of thrombin 10 drop below a defined limit ranging from 50-250 nm peak level (Figure 8 and Table 8). This translated into an averaged 20 hrs longer thrombin generation activity value for scFVIII versus full-length rFVIII for the thrombin peak level interval between 50 and 250 nM. When assessing the area under the peak curve between days 1-8 the thrombin generation activity of scFVIII was significantly better with $p(AUC_{TGA\ Peak-}ratio=1) = 0.0002$ (estimated ratio 1.26, 90% CI: 1.14-1.39) compared to full-length 15 FVIII, or in other words it took significantly longer for scFVIII to fall below a thrombin peak level of 50nm after administration than for the human wild-type Factor VIII Advate®.

20 These results again confirmed the increased functional stability of scFVIII after purification, lyophilization and reconstitution.

Table 8: Peak nm Thrombin

time [hrs]	CSL627 (N=8-14)		Advate (N=7-14)	
	mean	SD	mean	SD
0	0	0	0	0
24	325,5	40,2	343,4	40,41
32	294	102,8	305,2	72,13
48	277,1	22,44	279,6	30,92
72	283,1	42,88	163,6	79,87
96	115,4	29,63	75,44	35,28
120	91,85	57,63	55,17	30,89
144	45,6	36,15	24,61	13,52
168	8,453	12,66	14,13	19,44
192	3,901	9,384	4,71	8,1

5 **Example 6:** Stability of Factor VIII molecules in vWF deficient plasma (in-vivo)

scFVIII was reconstituted in 2.5 mL water for injection. ReFacto AF® and Advate® were reconstituted according to the description in the package insert. All test 10 articles were aliquoted and stored immediately frozen at approximately -70°C. Prior to administration test articles were diluted with formulation buffer for CSL 627 to get a minimum practical volume ensuring a reliable administration.

12 VWF ko mice (6 female/6 male) per group received a single i.v. injection of 100 IU/kg of either scFVIII based on chromogenic FVIII activity and ReFacto AF® or 15 Advate® based on the labeled FVIII activity into the lateral tail vein. Following administration of the different test items blood samples were drawn for determination of FVIII plasma levels at 0.083, 0.5, 1, 2, 4, 7, 16 and 24 hours from n= 2-3 mice per time point. Blood samples were processed to 10% citrate (3.13% w/v) plasma and subsequently subjected to FVIII plasma level analysis using the

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chromogenic assays system The chromogenic FVIII activity was determined using the COAMATIC® FVIII test kit from Chromogenix, Italy.

The AUC of the FVIII levels in plasma was calculated using the linear trapezoidal rule to calculate AUClast : from t=0 to last observation.

5 Likewise to results obtained after i.v. administration to FVIII ko mice and normal monkeys as well as s.c. administration to FVIII ko mice analysis the exposure to CSL627 was higher compared to ReFacto AF® and Advate®. Since analysis of the AUC, the most relevant and representative PK parameter for systemic exposure yielded a 30 % higher AUC value after administration of CSL627 compared to both

10 10 ReFacto AF® and Advate®. Again. these observations reflect the increased intrinsic stability of scFVIII after purification, lyophilization, reconstitution in vivo in plasma after administration to mice lacking systemic, circulating VWF, hence in absence of its shielding and protective effect for systemic, circulating FVIII.

15 **Table 9:** Plasma levels of scFVIII compared to Advate® and ReFacto AF® after administration to VWF deficient mice at a dose level of 100 IU/kg

	CSL627	Advate®	ReFacto AF®
Baseline	0,0	0,0	0,0
Total Area	2427	1654	1714
Total Peak Area	2427	1654	1714
Number of Peaks	1,000	1,000	1,000
Peak 1			
First X=	0,0830	0,0830	0,0830
Last X=	24,00	24,00	24,00
Peak X=	0,0830	0,0830	0,0830
Peak Y=	1770	1131	1248
Area under curve=	2427	1654	1714
%Area=	100,0	100,0	100,0

The claims defining the invention are as follows:

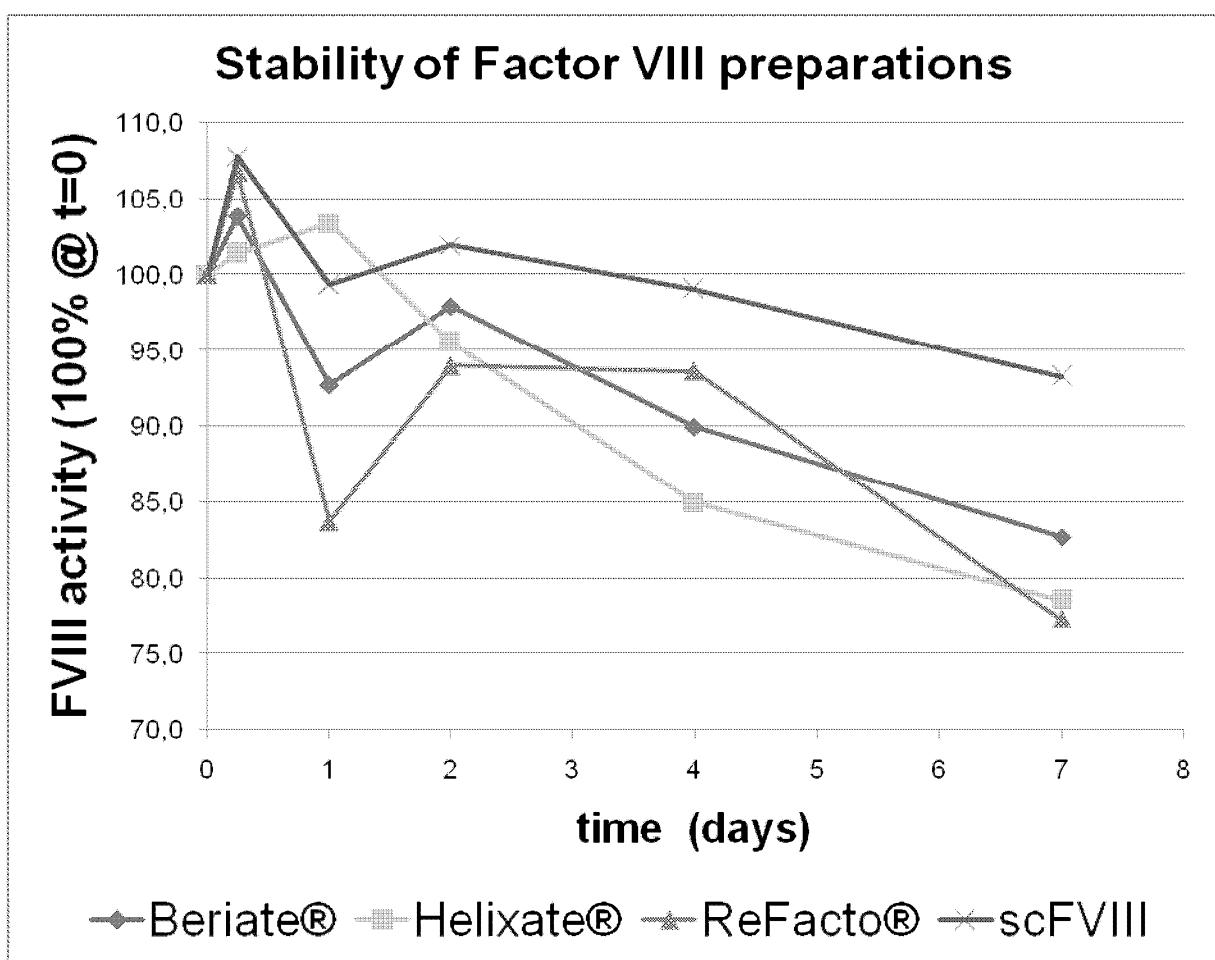
1. A method for improving the plasma half-life of a Factor VIII molecule after intravenous or non-intravenous administration relative to human wild-type Factor VIII, comprising fusing a first amino acid selected from the amino acids at positions 740 to 1647 of the Factor VIII sequence with a second amino acid selected from the amino acids at positions 1649 to 1690 of the Factor VIII sequence, whereby the proteolytic cleavage site between Arg1648 and Glu1649, and, if present in the FVIII molecule, the cleavage site between Arg1313 and Ala1314 is inactivated.
2. The method of claim 1, wherein the inactivation comprises deleting at least the amino acid sequence from Arg1313 to Arg1648 from the Factor VIII sequence.
3. The method of claim 1 or claim 2, wherein
 - amino acid 740 is fused to amino acid 1650, whereby amino acids 741 to 1649 are deleted;
 - amino acid 740 is fused to amino acid 1690, whereby amino acids 741 to 1689 are deleted;
 - amino acid 740 is fused to amino acid 1669, whereby amino acids 741 to 1668 are deleted;
 - amino acid 743 is fused to amino acid 1650, whereby amino acids 744 to 1649 are deleted;
 - amino acid 764 is fused to amino acid 1650, whereby amino acids 765 to 1649 are deleted;
 - amino acid 764 is fused to amino acid 1653, whereby amino acids 765 to 1652 are deleted;
 - amino acid 764 is fused to amino acid 1656, whereby amino acids 765 to 1655 are deleted;
 - amino acid 745 is fused to amino acid 1650, whereby amino acids 746 to 1649 are deleted;
 - amino acid 745 is fused to amino acid 1653, whereby amino acids 746 to 1652 are deleted;
 - amino acid 745 is fused to amino acid 1656, whereby amino acids 746 to 1655 are deleted;

- amino acid 757 is fused to amino acid 1650, whereby amino acids 758 to 1649 are deleted;
- amino acid 757 is fused to amino acid 1653, whereby amino acids 758 to 1652 are deleted;
- amino acid 757 is fused to amino acid 1656, whereby amino acids 758 to 1655 are deleted;
- amino acid 793 is fused to amino acid 1649, whereby amino acids 794 to 1648 are deleted;
- amino acid 793 is fused to amino acid 1690, whereby amino acids 794 to 1689 are deleted;
- amino acid 747 is fused to amino acid 1649, whereby amino acids 748 to 1648 are deleted;
- amino acid 751 is fused to amino acid 1649, whereby amino acids 752 to 1648 are deleted;
- amino acid 776 is fused to amino acid 1649, whereby amino acids 777 to 1648 are deleted; or
- amino acid 770 is fused to amino acid 1667, whereby amino acids 771 to 1666 are deleted.

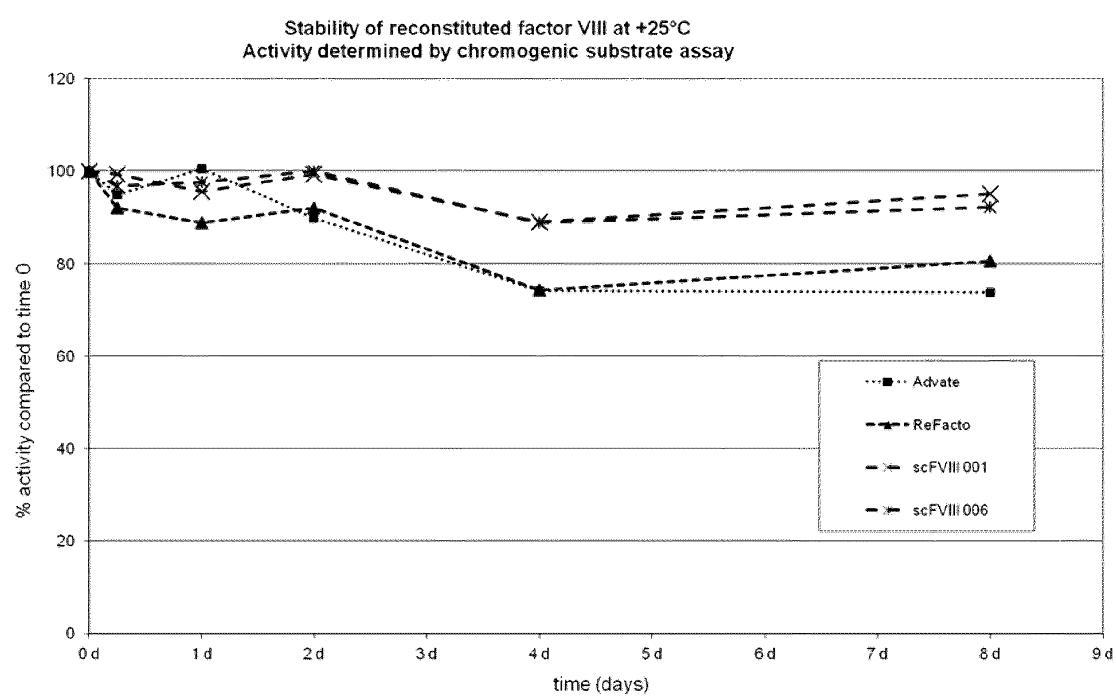
4. The method of any preceding claim, substantially as described herein.

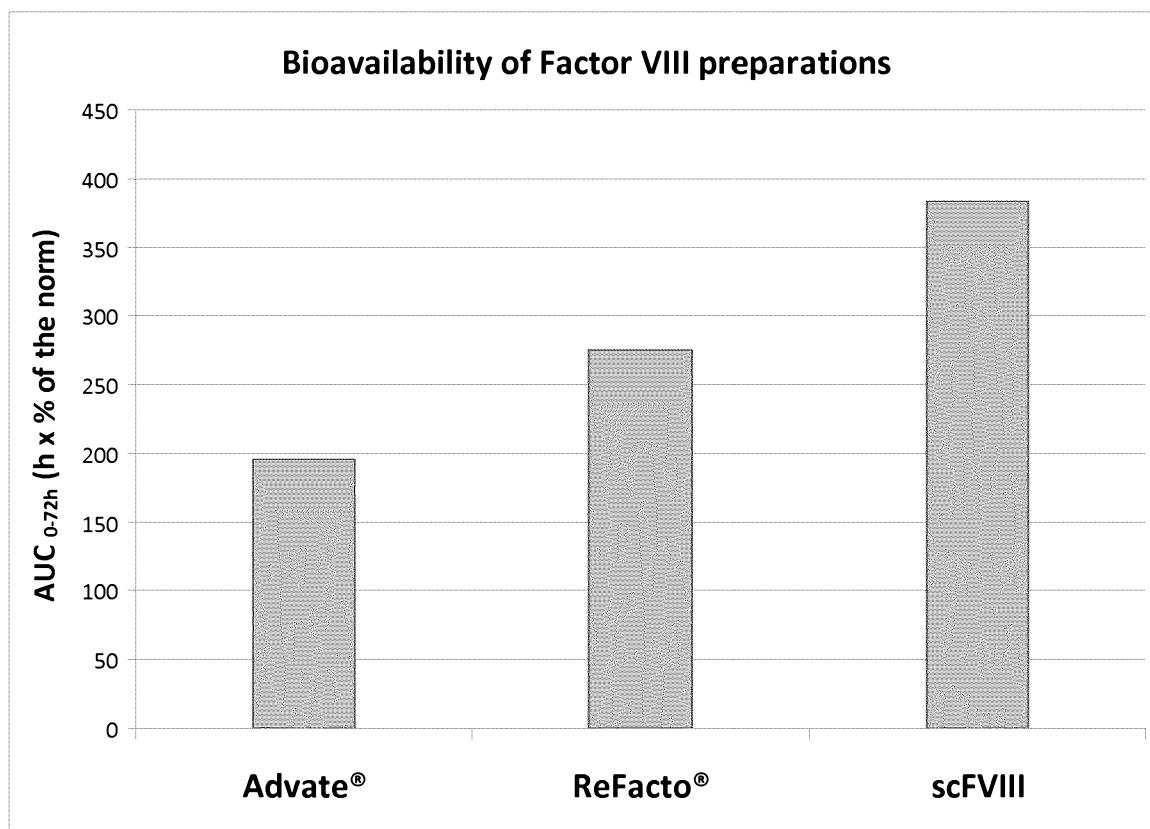
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Figure 1

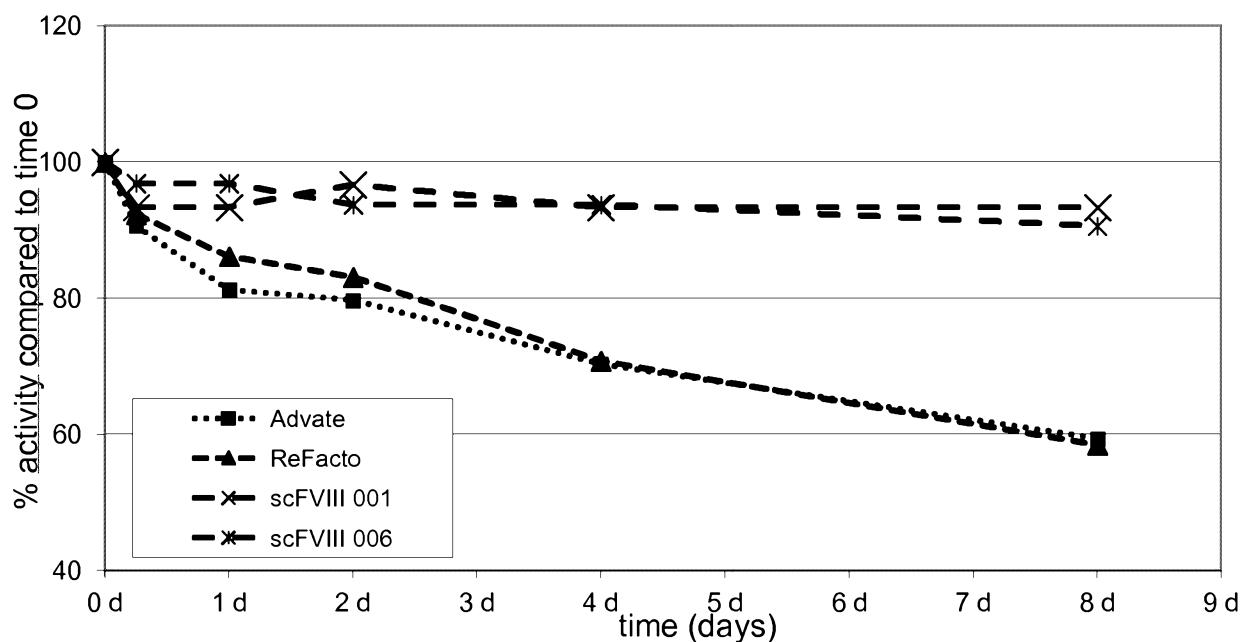


2/8

Figure 2

3/8**Figure 3**

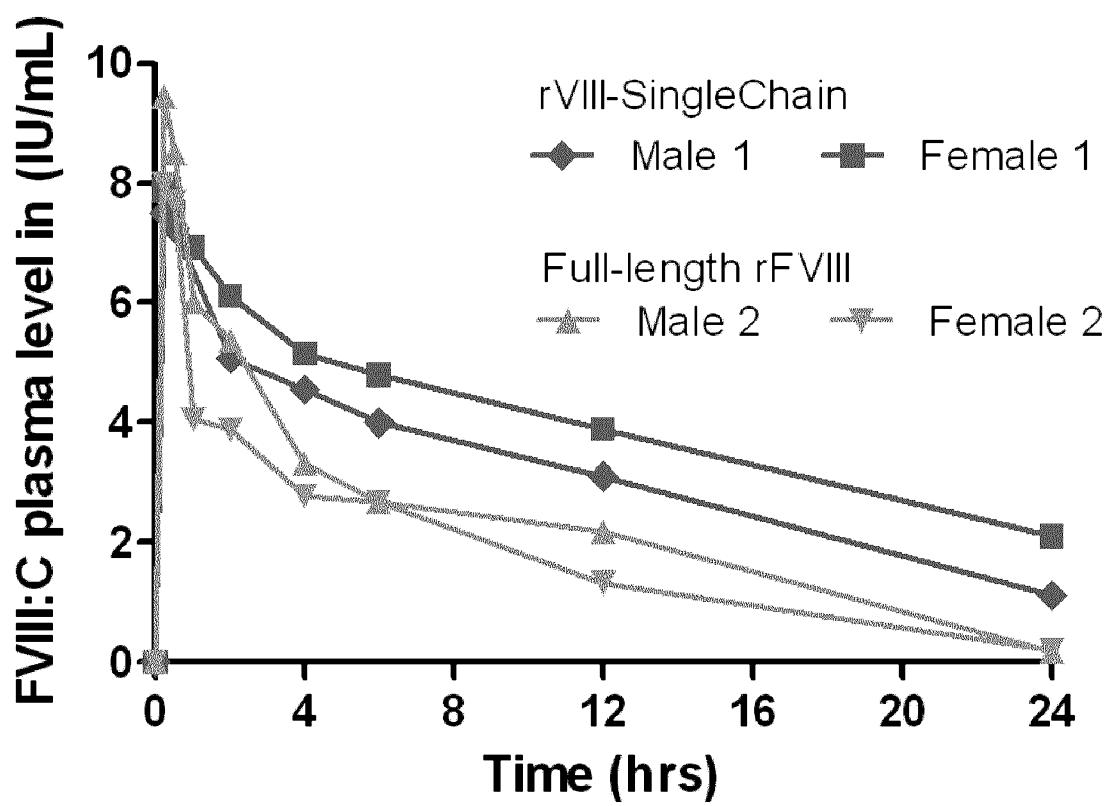
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Figure 4**Stability of FVIII molecules in plasma (ex-vivo)**

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Figure 5

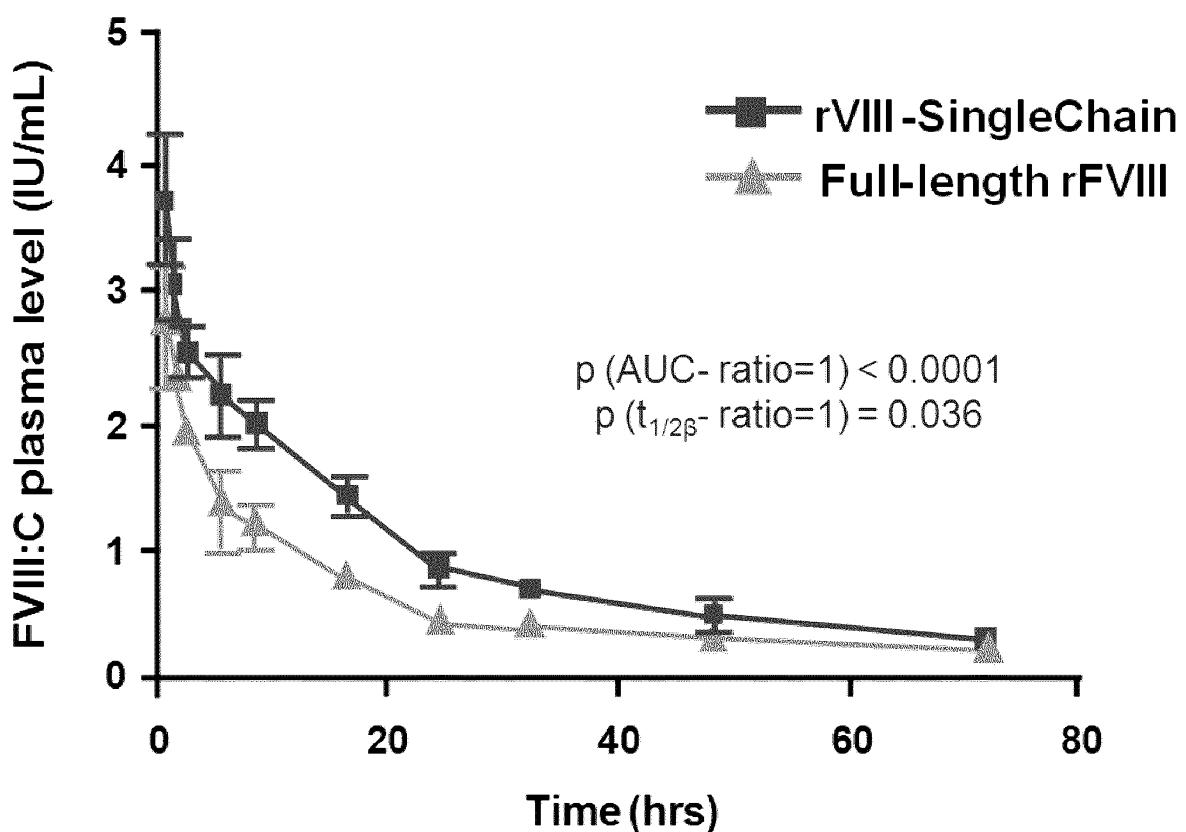
Stability of FVIII molecules in plasma (in-vivo) of cynomolgus monkeys (dosed at 250 IU/kg)



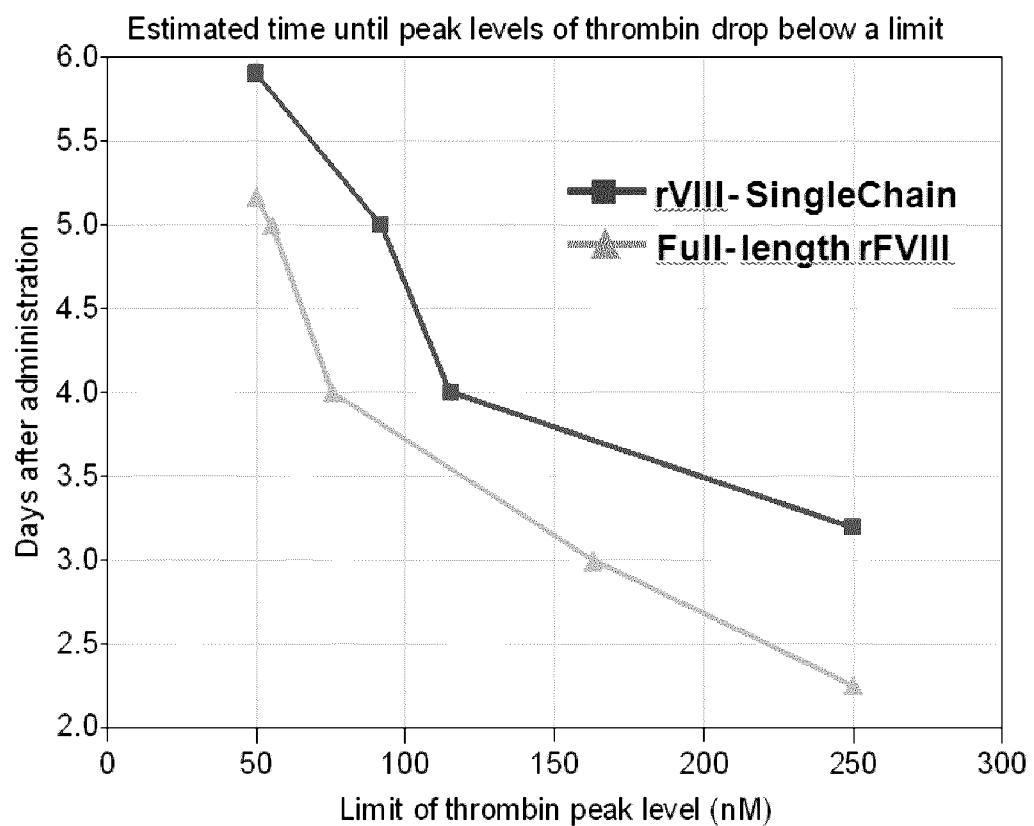
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Figure 6

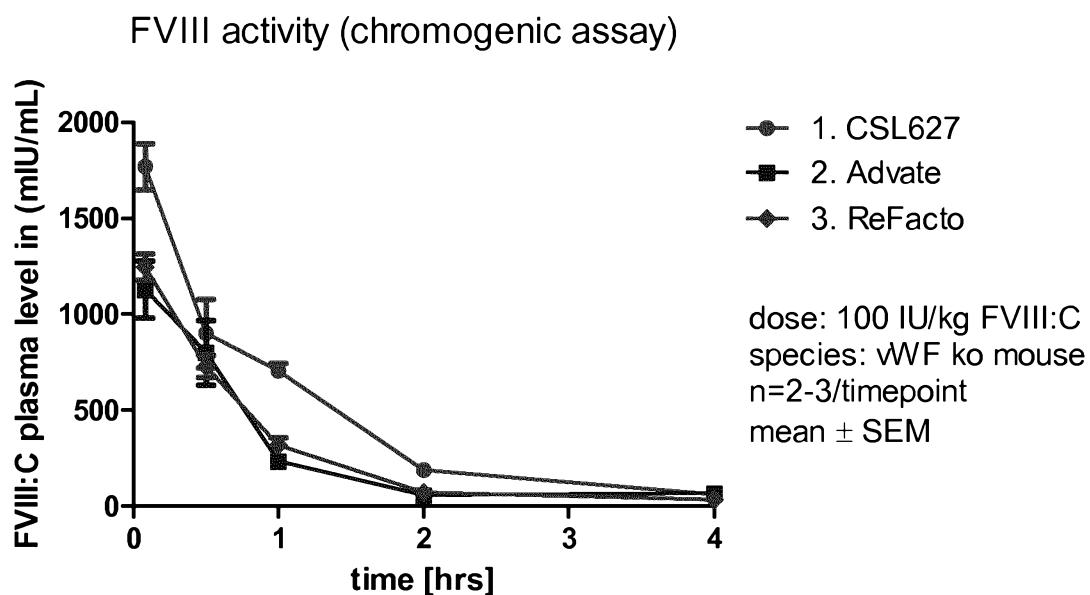
Stability of FVIII molecules in plasma (in-vivo) of in hemophilia A mice (dosed at 100 IU/kg)



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Figure 7**Thrombin generation in hemophilia A mice (ex vivo)**

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Figure 8**Stability of FVIII molecules in plasma (in-vivo) of vWF deficient mice**

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96

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144

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Leu	Val	Cys	Arg	Glu	Gly	Ser	Leu	Ala	Lys	Glu	Lys	Thr	Gln	Thr	Leu		
180							185					190					
cac	aaa	ttt	ata	cta	ctt	ttt	gct	gta	ttt	gat	gaa	ggg	aaa	agt	tgg	624	
His	Lys	Phe	Ile	Leu	Leu	Phe	Ala	Val	Phe	Asp	Glu	Gly	Lys	Ser	Trp		
195							200					205					
cac	tca	gaa	aca	aag	aac	tcc	ttg	atg	cag	gat	agg	gat	gct	gca	tct	672	
His	Ser	Glu	Thr	Lys	Asn	Ser	Leu	Met	Gln	Asp	Arg	Asp	Asp	Ala	Ala	Ser	
210							215					220					
gct	cg	gcc	tgg	cct	aaa	atg	cac	aca	gtc	aat	ggt	tat	gta	aac	agg	720	
Ala	Arg	Ala	Trp	Pro	Lys	Met	His	Thr	Val	Asn	Gly	Tyr	Val	Asn	Arg		
225							230				235				240		
tct	ctg	cca	ggt	ctg	att	gga	tgc	cac	agg	aaa	tca	gtc	tat	tgg	cat	768	
Ser	Leu	Pro	Gly	Leu	Ile	Gly	Cys	His	Arg	Lys	Ser	Val	Tyr	Trp	His		
245							250					255					
gtg	att	gga	atg	ggc	acc	act	cct	gaa	gtg	cac	tca	ata	ttc	ctc	gaa	816	
Val	Ile	Gly	Met	Gly	Thr	Thr	Pro	Glu	Val	His	Ser	Ile	Phe	Leu	Glu		
260							265					270					
ggt	cac	aca	ttt	ctt	gtg	agg	aac	cat	cgc	cag	gcg	tcc	ttg	gaa	atc	864	
Gly	His	Thr	Phe	Leu	Val	Arg	Asn	His	Arg	Gln	Ala	Ser	Leu	Glu	Ile		
275							280					285					
tcg	cca	ata	act	ttc	ctt	act	gct	caa	aca	ctc	ttg	atg	gac	ctt	gga	912	
Ser	Pro	Ile	Thr	Phe	Leu	Thr	Ala	Gln	Thr	Leu	Leu	Met	Asp	Leu	Gly		
290							295					300					
cag	ttt	cta	ctg	ttt	tgt	cat	atc	tct	tcc	cac	caa	cat	gat	ggc	atg	960	
Gln	Phe	Leu	Leu	Phe	Cys	His	Ile	Ser	Ser	His	Gln	His	Asp	Gly	Met		
305							310					315				320	
gaa	gct	tat	gtc	aaa	gta	gac	agc	tgt	cca	gag	gaa	ccc	caa	cta	cga	1008	
Glu	Ala	Tyr	Val	Lys	Val	Asp	Ser	Cys	Pro	Glu	Glu	Pro	Gln	Leu	Arg		
325							330						335				
atg	aaa	aat	aat	gaa	gaa	gcg	gaa	gac	tat	gat	gat	gat	ctt	act	gat	1056	
Met	Lys	Asn	Asn	Glu	Glu	Ala	Glu	Asp	Tyr	Asp	Asp	Asp	Leu	Thr	Asp		
340							345						350				
tct	gaa	atg	gat	gtg	gtc	agg	ttt	gat	gat	gac	aac	tct	cct	tcc	ttt	1104	
Ser	Glu	Met	Asp	Val	Val	Arg	Phe	Asp	Asp	Asp	Asn	Ser	Pro	Ser	Phe		
355							360						365				
atc	caa	att	cgc	tca	gtt	gcc	aag	aag	cat	cct	aaa	act	tgg	gta	cat	1152	
Ile	Gln	Ile	Arg	Ser	Val	Ala	Lys	Lys	His	Pro	Lys	Thr	Trp	Val	His		
370							375						380				
tac	att	gct	gct	gaa	gag	gag	gac	tgg	gac	tat	gct	ccc	tta	gtc	ctc	1200	
Tyr	Ile	Ala	Ala	Glu	Glu	Asp	Trp	Asp	Tyr	Ala	Pro	Leu	Val	Leu			
385							390						395			400	
gcc	ccc	gat	gac	aga	agt	tat	aaa	agt	caa	tat	ttg	aac	aat	ggc	cct	1248	
Ala	Pro	Asp	Asp	Arg	Ser	Tyr	Lys	Ser	Gln	Tyr	Leu	Asn	Asn	Gly	Pro		
405							410							415			
cag	cg	att	ggt	agg	aag	aag	tac	aaa	aaa	gtc	cga	ttt	atg	gca	tac	aca	1296
Gln	Arg	Ile	Gly	Arg	Lys	Tyr	Lys	Lys	Val	Arg	Phe	Met	Ala	Tyr	Thr		
420							425						430				

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gat gaa acc ttt aag act cgt gaa gct att cag cat gaa tca gga atc	1344
Asp Glu Thr Phe Lys Thr Arg Glu Ala Ile Gln His Glu Ser Gly Ile	
435 440 445	
ttg gga cct tta ctt tat ggg gaa gtt gga gac aca ctg ttg att ata	1392
Leu Gly Pro Leu Leu Tyr Gly Glu Val Gly Asp Thr Leu Leu Ile Ile	
450 455 460	
ttt aag aat caa gca agc aga cca tat aac atc tac cct cac gga atc	1440
Phe Lys Asn Gln Ala Ser Arg Pro Tyr Asn Ile Tyr Pro His Gly Ile	
465 470 475 480	
act gat gtc cgt cct ttg tat tca agg aga tta cca aaa ggt gta aaa	1488
Thr Asp Val Arg Pro Leu Tyr Ser Arg Arg Leu Pro Lys Gly Val Lys	
485 490 495	
cat ttg aag gat ttt cca att ctg cca gga gaa ata ttc aaa tat aaa	1536
His Leu Lys Asp Phe Pro Ile Leu Pro Gly Glu Ile Phe Lys Tyr Lys	
500 505 510	
tgg aca gtg act gta gaa gat ggg cca act aaa tca gat cct cgg tgc	1584
Trp Thr Val Thr Val Glu Asp Gly Pro Thr Lys Ser Asp Pro Arg Cys	
515 520 525	
ctg acc cgc tat tac tct agt ttc gtt aat atg gag aga gat cta gct	1632
Leu Thr Arg Tyr Tyr Ser Ser Phe Val Asn Met Glu Arg Asp Leu Ala	
530 535 540	
tca gga ctc att ggc cct ctc atc tgc tac aaa gaa tct gta gat	1680
Ser Gly Leu Ile Gly Pro Leu Leu Ile Cys Tyr Lys Glu Ser Val Asp	
545 550 555 560	
caa aga gga aac cag ata atg tca gac aag agg aat gtc atc ctg ttt	1728
Gln Arg Gly Asn Gln Ile Met Ser Asp Lys Arg Asn Val Ile Leu Phe	
565 570 575	
tct gta ttt gat gag aac cga agc tgg tac ctc aca gag aat ata caa	1776
Ser Val Phe Asp Glu Asn Arg Ser Trp Tyr Leu Thr Glu Asn Ile Gln	
580 585 590	
cgc ttt ctc ccc aat cca gct gga gtg cag ctt gag gat cca gag ttc	1824
Arg Phe Leu Pro Asn Pro Ala Gly Val Gln Leu Glu Asp Pro Glu Phe	
595 600 605	
caa gcc tcc aac atc atg cac agc atc aat ggc tat gtt ttt gat agt	1872
Gln Ala Ser Asn Ile Met His Ser Ile Asn Gly Tyr Val Phe Asp Ser	
610 615 620	
ttg cag ttg tca gtt tgt ttg cat gag gtg gca tac tgg tac att cta	1920
Leu Gln Leu Ser Val Cys Leu His Glu Val Ala Tyr Trp Tyr Ile Leu	
625 630 635 640	
agc att gga gca cag act gac ttc ctt tct gtc ttc ttc tct gga tat	1968
Ser Ile Gly Ala Gln Thr Asp Phe Leu Ser Val Phe Phe Ser Gly Tyr	
645 650 655	
acc ttc aaa cac aaa atg gtc tat gaa gac aca ctc acc cta ttc cca	2016
Thr Phe Lys His Lys Met Val Tyr Glu Asp Thr Leu Thr Leu Phe Pro	
660 665 670	
ttc tca gga gaa act gtc ttc atg tcg atg gaa aac cca ggt cta tgg	2064
Phe Ser Gly Glu Thr Val Phe Met Ser Met Glu Asn Pro Gly Leu Trp	
675 680 685	
att ctg ggg tgc cac aac tca gac ttt cgg aac aga ggc atg acc gcc	2112
Ile Leu Gly Cys His Asn Ser Asp Phe Arg Asn Arg Gly Met Thr Ala	
690 695 700	

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tta ctg aag gtt tct agt tgt gac aag aac act ggt gat tat tac gag	2160
Leu Leu Lys Val Ser Ser Cys Asp Lys Asn Thr Gly Asp Tyr Tyr Glu	
705 710 715 720	
gac agt tat gaa gat att tca gca tac ttg ctg agt aaa aac aat gcc	2208
Asp Ser Tyr Glu Asp Ile Ser Ala Tyr Leu Leu Ser Lys Asn Asn Ala	
725 730 735	
att gaa cca aga agc ttc tcc cag aat tca aga cac cgt agc act agg	2256
Ile Glu Pro Arg Ser Phe Ser Gln Asn Ser Arg His Arg Ser Thr Arg	
740 745 750	
caa aag caa ttt aat gcc acc aca att cca gaa aat gac ata gag aag	2304
Gln Lys Gln Phe Asn Ala Thr Thr Ile Pro Glu Asn Asp Ile Glu Lys	
755 760 765	
act gac cct tgg ttt gca cac aga aca cct atg cct aaa ata caa aat	2352
Thr Asp Pro Trp Phe Ala His Arg Thr Pro Met Pro Lys Ile Gln Asn	
770 775 780	
gtc tcc tct agt gat ttg ttg atg ctc ttg cga cag agt cct act cca	2400
Val Ser Ser Asp Leu Leu Met Leu Leu Arg Gln Ser Pro Thr Pro	
785 790 795 800	
cat ggg cta tcc tta tct gat ctc caa gaa gcc aaa tat gag act ttt	2448
His Gly Leu Ser Leu Ser Asp Leu Gln Glu Ala Lys Tyr Glu Thr Phe	
805 810 815	
tct gat gat cca tca cct gga gca ata gac agt aat aac agc ctg tct	2496
Ser Asp Asp Pro Ser Pro Gly Ala Ile Asp Ser Asn Asn Ser Leu Ser	
820 825 830	
gaa atg aca cac ttc agg cca cag ctc cat cac agt ggg gac atg gta	2544
Glu Met Thr His Phe Arg Pro Gln Leu His His Ser Gln Asp Met Val	
835 840 845	
ttt acc cct gag tca ggc ctc caa tta aga tta aat gag aaa ctg ggg	2592
Phe Thr Pro Glu Ser Gly Leu Gln Leu Arg Leu Asn Glu Lys Leu Gly	
850 855 860	
aca act gca gca aca gag ttg aag aaa ctt gat ttc aaa gtt tct agt	2640
Thr Thr Ala Ala Thr Glu Leu Lys Lys Leu Asp Phe Lys Val Ser Ser	
865 870 875 880	
aca tca aat aat ctg att tca aca att cca tca gac aat ttg gca gca	2688
Thr Ser Asn Asn Leu Ile Ser Thr Ile Pro Ser Asp Asn Leu Ala Ala	
885 890 895	
ggt act gat aat aca agt tcc tta gga ccc cca agt atg cca gtt cat	2736
Gly Thr Asp Asn Thr Ser Ser Leu Gly Pro Pro Ser Met Pro Val His	
900 905 910	
tat gat agt caa tta gat acc act cta ttt ggc aaa aag tca tct ccc	2784
Tyr Asp Ser Gln Leu Asp Thr Thr Leu Phe Gly Lys Lys Ser Ser Pro	
915 920 925	
ctt act gag tct ggt gga cct ctg agc ttg agt gaa gaa aat aat gat	2832
Leu Thr Glu Ser Gly Gly Pro Leu Ser Leu Ser Glu Glu Asn Asn Asp	
930 935 940	
tca aag ttg tta gaa tca ggt tta atg aat agc caa gaa agt tca tgg	2880
Ser Lys Leu Leu Glu Ser Gly Leu Met Asn Ser Gln Glu Ser Ser Trp	
945 950 955 960	
gga aaa aat gta tcg tca aca gag agt ggt agg tta ttt aaa ggg aaa	2928
Gly Lys Asn Val Ser Ser Thr Glu Ser Gly Arg Leu Phe Lys Gly Lys	
965 970 975	

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aga	gct	cat	gga	cct	gct	ttg	ttg	act	aaa	gat	aat	gcc	tta	ttc	aaa	2976
Arg	Ala	His	Gly	Pro	Ala	Leu	Leu	Thr	Lys	Asp	Asn	Ala	Leu	Phe	Lys	
980					985							990				
gtt	agc	atc	tct	ttg	tta	aag	aca	aac	aaa	act	tcc	aat	aat	tca	gca	3024
Val	Ser	Ile	Ser	Leu	Leu	Lys	Thr	Asn	Lys	Thr	Ser	Asn	Asn	Ser	Ala	
995				1000							1005					
act	aat	aga	aag	act	cac	att	gat	ggc	cca	tca	tta	tta	att	gag		3069
Thr	Asn	Arg	Lys	Thr	His	Ile	Asp	Gly	Pro	Ser	Leu	Leu	Ile	Glu		
1010					1015						1020					
aat	agt	cca	tca	gtc	tgg	caa	aat	ata	tta	gaa	agt	gac	act	gag		3114
Asn	Ser	Pro	Ser	Val	Trp	Gln	Asn	Ile	Leu	Gl	Ser	Asp	Thr	Glu		
1025					1030						1035					
ttt	aaa	aaa	gtg	aca	cct	ttg	att	cat	gac	aga	atg	ctt	atg	gac		3159
Phe	Lys	Lys	Val	Thr	Pro	Leu	Ile	His	Asp	Arg	Met	Leu	Met	Asp		
1040					1045						1050					
aaa	aat	gct	aca	gct	ttg	agg	cta	aat	cat	atg	tca	aat	aaa	act		3204
Lys	Asn	Ala	Thr	Ala	Leu	Arg	Leu	Asn	His	Met	Ser	Asn	Lys	Thr		
1055					1060						1065					
act	tca	tca	aaa	aac	atg	gaa	atg	gtc	caa	cag	aaa	aaa	gag	ggc		3249
Thr	Ser	Ser	Lys	Asn	Met	Glu	Met	Val	Gln	Gln	Lys	Lys	Glu	Gly		
1070					1075						1080					
ccc	att	cca	cca	gat	gca	caa	aat	cca	gat	atg	tcg	tcc	ttt	aag		3294
Pro	Ile	Pro	Pro	Asp	Ala	Gln	Asn	Pro	Asp	Met	Ser	Phe	Phe	Lys		
1085					1090						1095					
atg	cta	tcc	ttg	cca	gaa	tca	gca	agg	tgg	ata	caa	agg	act	cat		3339
Met	Leu	Phe	Leu	Pro	Glu	Ser	Ala	Arg	Trp	Ile	Gln	Arg	Thr	His		
1100					1105						1110					
gga	aag	aac	tct	ctg	aac	tct	ggg	caa	ggc	ccc	agt	cca	aag	caa		3384
Gly	Lys	Asn	Ser	Leu	Asn	Ser	Gly	Gln	Gly	Pro	Ser	Pro	Lys	Gln		
1115					1120						1125					
tta	gta	tcc	tta	gga	cca	gaa	aaa	tct	gtg	gaa	ggt	cag	aat	ttc		3429
Leu	Val	Ser	Leu	Gly	Pro	Glu	Lys	Ser	Val	Glu	Gly	Gln	Asn	Phe		
1130					1135						1140					
ttg	tct	gag	aaa	aac	aaa	gtg	gta	gta	gga	aag	ggt	gaa	ttt	aca		3474
Leu	Ser	Glu	Lys	Asn	Lys	Val	Val	Val	Gly	Lys	Gly	Glu	Phe	Thr		
1145					1150						1155					
aag	gac	gta	gga	ctc	aaa	gag	atg	gtt	ttt	cca	agc	agc	aga	aac		3519
Lys	Asp	Val	Gly	Leu	Lys	Glu	Met	Val	Phe	Pro	Ser	Ser	Arg	Asn		
1160					1165						1170					
cta	ttt	ctt	act	aac	ttg	gat	aat	tta	cat	gaa	aat	aat	aca	cac		3564
Leu	Phe	Leu	Thr	Asn	Leu	Asp	Asn	Leu	His	Gl	Asn	Asn	Thr	His		
1175					1180						1185					
aat	caa	gaa	aaa	aaa	att	cag	gaa	gaa	ata	gaa	aag	aag	gaa	aca		3609
Asn	Gln	Glu	Lys	Lys	Ile	Gln	Glu	Glu	Ile	Glu	Lys	Lys	Glu	Thr		
1190					1195						1200					
tta	atc	caa	gag	aat	gta	gtt	ttg	cct	cag	ata	cat	aca	gtg	act		3654
Leu	Ile	Gln	Glu	Asn	Val	Val	Leu	Pro	Gln	Ile	His	Thr	Val	Thr		
1205					1210						1215					
ggc	act	aag	aat	ttc	atg	aag	aac	ctt	ttc	tta	ctg	agc	act	agg		3699
Gly	Thr	Lys	Asn	Phe	Met	Lys	Asn	Leu	Phe	Leu	Leu	Ser	Thr	Arg		
1220					1225						1230					

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caa	aat	gta	gaa	ggt	tca	tat	gac	ggg	gca	tat	gct	cca	gta	ctt	3744
Gln	Asn	Val	Glu	Gly	Ser	Tyr	Asp	Gly	Ala	Tyr	Ala	Pro	Val	Leu	
1235					1240						1245				
caa	gat	ttt	agg	tca	tta	aat	gat	tca	aca	aat	aga	aca	aag	aaa	3789
Gln	Asp	Phe	Arg	Ser	Leu	Asn	Asp	Ser	Thr	Asn	Arg	Thr	Lys	Lys	
1250					1255						1260				
cac	aca	gct	cat	ttc	tca	aaa	aaa	ggg	gag	gaa	gaa	aac	ttg	gaa	3834
His	Thr	Ala	His	Phe	Ser	Lys	Lys	Gly	Gl	Gl	Gl	Asn	Leu	Gl	
1265					1270						1275				
ggc	ttg	gga	aat	caa	acc	aag	caa	att	gta	gag	aaa	tat	gca	tgc	3879
Gly	Leu	Gly	Asn	Gln	Thr	Lys	Gln	Ile	Val	Gl	Lys	Tyr	Ala	Cys	
1280					1285						1290				
acc	aca	agg	ata	tct	cct	aat	aca	agc	cag	cag	aat	ttt	gtc	acg	3924
Thr	Thr	Arg	Ile	Ser	Pro	Asn	Thr	Ser	Gln	Gln	Asn	Phe	Val	Thr	
1295					1300						1305				
caa	cgt	agt	aag	aga	gct	ttg	aaa	caa	ttc	aga	ctc	cca	cta	gaa	3969
Gln	Arg	Ser	Lys	Arg	Ala	Leu	Lys	Gln	Phe	Arg	Leu	Pro	Leu	Gl	
1310					1315						1320				
gaa	aca	gaa	ctt	gaa	aaa	agg	ata	att	gtq	gat	gac	acc	tca	acc	4014
Glu	Thr	Glu	Leu	Glu	Lys	Arg	Ile	Ile	Val	Asp	Asp	Thr	Ser	Thr	
1325					1330						1335				
cag	tgg	tcc	aaa	aac	atg	aaa	cat	ttg	acc	ccg	agc	acc	ctc	aca	4059
Gln	Trp	Ser	Lys	Asn	Met	Lys	His	Leu	Thr	Pro	Ser	Thr	Leu	Thr	
1340					1345						1350				
cag	ata	gac	tac	aat	gag	aag	gag	aaa	ggg	gcc	att	act	cag	tct	4104
Gln	Ile	Asp	Tyr	Asn	Glu	Lys	Glu	Lys	Gly	Ala	Ile	Thr	Gln	Ser	
1355					1360						1365				
ccc	tta	tca	gat	tgc	ctt	acg	agg	agt	cat	agc	atc	cct	caa	gca	4149
Pro	Leu	Ser	Asp	Cys	Leu	Thr	Arg	Ser	His	Ser	Ile	Pro	Gln	Ala	
1370					1375						1380				
aat	aga	tct	cca	tta	ccc	att	gca	aag	gta	tca	tca	ttt	cca	tct	4194
Asn	Arg	Ser	Pro	Leu	Pro	Ile	Ala	Lys	Val	Ser	Ser	Phe	Pro	Ser	
1385					1390						1395				
att	aga	cct	ata	tat	ctg	acc	agg	gtc	cta	ttc	caa	gac	aac	tct	4239
Ile	Arg	Pro	Ile	Tyr	Leu	Thr	Arg	Val	Leu	Phe	Gln	Asp	Asn	Ser	
1400					1405						1410				
tct	cat	ctt	cca	gca	gca	tct	tat	aga	aag	aaa	gat	tct	ggg	gtc	4284
Ser	His	Leu	Pro	Ala	Ala	Ser	Tyr	Arg	Lys	Lys	Asp	Ser	Gly	Val	
1415					1420						1425				
caa	gaa	agc	agt	cat	ttc	tta	caa	gga	gcc	aaa	aaa	aat	aac	ctt	4329
Gln	Glu	Ser	Ser	His	Phe	Leu	Gln	Gly	Ala	Lys	Lys	Asn	Asn	Leu	
1430					1435						1440				
tct	tta	gcc	att	cta	acc	ttg	gag	atg	act	ggt	gat	caa	aga	gag	4374
Ser	Leu	Ala	Ile	Leu	Thr	Leu	Glu	Met	Thr	Gly	Asp	Gln	Arg	Glu	
1445					1450						1455				
gtt	ggc	tcc	ctg	ggg	aca	agt	gcc	aca	aat	tca	gtc	aca	tac	aag	4419
Val	Gly	Ser	Leu	Gly	Thr	Ser	Ala	Thr	Asn	Ser	Val	Thr	Tyr	Lys	
1460					1465						1470				
aaa	gtt	gag	aac	act	gtt	ctc	ccg	aaa	cca	gac	ttg	ccc	aaa	aca	4464
Lys	Val	Glu	Asn	Thr	Val	Leu	Pro	Lys	Pro	Asp	Leu	Pro	Lys	Thr	
1475					1480						1485				

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tct	ggc	aaa	gtt	gaa	ttg	ctt	cca	aaa	gtt	cac	att	tat	cag	aag	4509
Ser	Gly	Lys	Val	Glu	Leu	Leu	Pro	Lys	Val	His	Ile	Tyr	Gln	Lys	
1490					1495						1500				
gac	cta	ttc	cct	acg	gaa	act	agc	aat	ggg	tct	cct	ggc	cat	ctg	4554
Asp	Leu	Phe	Pro	Thr	Glu	Thr	Ser	Asn	Gly	Ser	Pro	Gly	His	Leu	
1505					1510						1515				
gat	ctc	gtg	gaa	ggg	agc	ctt	ctt	cag	gga	aca	gag	gga	gcg	att	4599
Asp	Leu	Val	Glu	Gly	Ser	Leu	Leu	Gln	Gly	Thr	Glu	Gly	Ala	Ile	
1520					1525						1530				
aag	tgg	aat	gaa	gca	aac	aga	cct	gga	aaa	gtt	ccc	ttt	ctg	aga	4644
Lys	Trp	Asn	Glu	Ala	Asn	Arg	Pro	Gly	Lys	Val	Pro	Phe	Leu	Arg	
1535					1540						1545				
gta	gca	aca	gaa	agc	tct	gca	aag	act	ccc	tcc	aag	cta	ttg	gat	4689
Val	Ala	Thr	Glu	Ser	Ser	Ala	Lys	Thr	Pro	Ser	Lys	Leu	Leu	Asp	
1550					1555						1560				
cct	ctt	gct	tgg	gat	aac	cac	tat	ggt	act	cag	ata	cca	aaa	gaa	4734
Pro	Leu	Ala	Trp	Asp	Asn	His	Tyr	Gly	Thr	Gln	Ile	Pro	Lys	Glu	
1565					1570						1575				
gag	tgg	aaa	tcc	caa	gag	aag	tca	cca	gaa	aaa	aca	gct	ttt	aag	4779
Glu	Trp	Lys	Ser	Gln	Glu	Lys	Ser	Pro	Glu	Lys	Thr	Ala	Phe	Lys	
1580					1585						1590				
aaa	aag	gat	acc	att	ttg	tcc	ctg	aac	gct	tgt	gaa	agc	aat	cat	4824
Lys	Lys	Asp	Thr	Ile	Leu	Ser	Leu	Asn	Ala	Cys	Glu	Ser	Asn	His	
1595					1600						1605				
gca	ata	gca	gca	ata	aat	gag	gga	caa	aat	aag	ccc	gaa	ata	gaa	4869
Ala	Ile	Ala	Ala	Ile	Asn	Glu	Gly	Gln	Asn	Lys	Pro	Glu	Ile	Glu	
1610					1615						1620				
gtc	acc	tgg	gca	aag	caa	ggt	agg	act	gaa	agg	ctg	tgc	tct	caa	4914
Val	Thr	Trp	Ala	Lys	Gln	Gly	Arg	Thr	Glu	Arg	Leu	Cys	Ser	Gln	
1625					1630						1635				
aac	cca	cca	gtc	ttg	aaa	cgc	cat	caa	cgg	gaa	ata	act	cgt	act	4959
Asn	Pro	Pro	Val	Leu	Lys	Arg	His	Gln	Arg	Glu	Ile	Thr	Arg	Thr	
1640					1645						1650				
act	ctt	cag	tca	gat	caa	gag	gaa	att	gac	tat	gat	gat	acc	ata	5004
Thr	Leu	Gln	Ser	Asp	Gln	Glu	Glu	Ile	Asp	Tyr	Asp	Asp	Thr	Ile	
1655					1660						1665				
tca	gtt	gaa	atg	aag	aag	gaa	gat	ttt	gac	att	tat	gat	gag	gat	5049
Ser	Val	Glu	Met	Lys	Lys	Glu	Asp	Phe	Asp	Ile	Tyr	Asp	Glu	Asp	
1670					1675						1680				
gaa	aat	cag	agc	ccc	cgc	agc	ttt	caa	aag	aaa	aca	cga	cac	tat	5094
Glu	Asn	Gln	Ser	Pro	Arg	Ser	Phe	Gln	Lys	Lys	Thr	Arg	His	Tyr	
1685					1690						1695				
ttt	att	gct	gca	gtg	gag	agg	ctc	tgg	gat	tat	ggg	atg	agt	agc	5139
Phe	Ile	Ala	Ala	Val	Glu	Arg	Leu	Trp	Asp	Tyr	Gly	Met	Ser	Ser	
1700					1705						1710				
tcc	cca	cat	gtt	cta	aga	aac	agg	gct	cag	agt	ggc	agt	gtc	cct	5184
Ser	Pro	His	Val	Leu	Arg	Asn	Arg	Ala	Gln	Ser	Gly	Ser	Val	Pro	
1715					1720						1725				
cag	tcc	aag	aaa	gtt	gtt	ttc	cag	gaa	ttt	act	gat	ggc	tcc	ttt	5229
Gln	Phe	Lys	Lys	Val	Val	Phe	Gln	Glu	Phe	Thr	Asp	Gly	Ser	Phe	
1730					1735						1740				

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act	cag	ccc	tta	tac	cgt	gga	gaa	cta	aat	gaa	cat	ttg	gga	ctc	5274	
Thr	Gln	Pro	Leu	Tyr	Arg	Gly	Glu	Leu	Asn	Glu	His	Leu	Gly	Leu		
1745						1750					1755					
ctg	ggg	cca	tat	ata	aga	gca	gaa	gtt	gaa	gat	aat	atc	atg	gta	5319	
Leu	Gly	Pro	Tyr	Ile	Arg	Ala	Glu	Val	Glu	Asp	Asn	Ile	Met	Val		
1760						1765					1770					
act	ttc	aga	aat	cag	gcc	tct	cgt	ccc	tat	tcc	ttc	tat	tct	agc	5364	
Thr	Phe	Arg	Asn	Gln	Ala	Ser	Arg	Pro	Tyr	Ser	Phe	Tyr	Ser	Ser		
1775						1780					1785					
ctt	att	tct	tat	gag	gaa	gat	cag	agg	caa	gga	gca	gaa	cct	aga	5409	
Leu	Ile	Ser	Tyr	Glu	Glu	Asp	Gln	Arg	Gln	Gly	Ala	Glu	Pro	Arg		
1790						1795					1800					
aaa	aac	ttt	gtc	aag	cct	aat	gaa	acc	aaa	act	tac	ttt	tgg	aaa	5454	
Lys	Asn	Phe	Val	Lys	Pro	Asn	Glu	Thr	Lys	Thr	Tyr	1815	Phe	Trp	Lys	
1805						1810										
gtg	caa	cat	cat	atg	gca	ccc	act	aaa	gat	gag	ttt	gac	tgc	aaa	5499	
Val	Gln	His	His	Met	Ala	Pro	Thr	Lys	Asp	Glu	Phe	Asp	Cys	Lys		
1820						1825					1830					
gcc	tgg	gct	tat	ttc	tct	gat	gtt	gac	ctg	gaa	aaa	gat	gtg	cac	5544	
Ala	Trp	Ala	Tyr	Phe	Ser	Asp	Val	Asp	Leu	Glu	Lys	Asp	Val	His		
1835						1840					1845					
tca	ggc	ctg	att	gga	ccc	ctt	ctg	gtc	tgc	cac	act	aac	aca	ctg	5589	
Ser	Gly	Leu	Ile	Gly	Pro	Leu	Leu	Val	Cys	His	Thr	Asn	Thr	Leu		
1850						1855					1860					
aac	cct	gct	cat	ggg	aga	caa	gtg	aca	gta	cag	gaa	ttt	gct	ctg	5634	
Asn	Pro	Ala	His	Gly	Arg	Gln	Val	Thr	Val	Gln	Glu	Phe	Ala	Leu		
1865						1870					1875					
ttt	ttc	acc	atc	ttt	gat	gag	acc	aaa	agc	tgg	tac	ttc	act	gaa	5679	
Phe	Phe	Thr	Ile	Phe	Asp	Glu	Thr	Lys	Ser	Trp	Tyr	Phe	Thr	Glu		
1880						1885					1890					
aat	atg	gaa	aga	aac	tgc	agg	gct	ccc	tgc	aat	atc	cag	atg	gaa	5724	
Asn	Met	Glu	Arg	Asn	Cys	Arg	Ala	Pro	Cys	Asn	Ile	Gln	Met	Glu		
1895						1900					1905					
gat	ccc	act	ttt	aaa	gag	aat	tat	cgc	ttc	cat	gca	atc	aat	ggc	5769	
Asp	Pro	Thr	Phe	Lys	Glu	Asn	Tyr	Arg	Phe	His	Ala	Ile	Asn	Gly		
1910						1915					1920					
tac	ata	atg	gat	aca	cta	cct	ggc	tta	gta	atg	gct	cag	gat	caa	5814	
Tyr	Ile	Met	Asp	Thr	Leu	Pro	Gly	Leu	Val	Met	Ala	Gln	Asp	Gln		
1925						1930					1935					
agg	att	cga	tgg	tat	ctg	ctc	agc	atg	ggc	agc	aat	gaa	aac	atc	5859	
Arg	Ile	Arg	Trp	Tyr	Leu	Leu	Ser	Met	Gly	Ser	Asn	Glu	Asn	Ile		
1940						1945					1950					
cat	tct	att	cat	ttc	agt	gga	cat	gtg	ttc	act	gta	cga	aaa	aaa	5904	
His	Ser	Ile	His	Phe	Ser	Gly	His	Val	Phe	Thr	Val	Arg	Lys	Lys		
1955						1960					1965					
gag	gag	tat	aaa	atg	gca	ctg	tac	aat	ctc	tat	cca	ggt	gtt	ttt	5949	
Glu	Glu	Tyr	Lys	Met	Ala	Leu	Tyr	Asn	Leu	Tyr	Pro	Gly	Val	Phe		
1970						1975					1980					
gag	aca	gtg	gaa	atg	tta	cca	tcc	aaa	gct	gga	att	tgg	cgg	gtg	5994	
Glu	Thr	Val	Glu	Met	Leu	Pro	Ser	Lys	Ala	Gly	Ile	Trp	Arg	Val		
1985						1990					1995					

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gaa	tgc	ctt	att	ggc	gag	cat	cta	cat	gct	ggg	atg	agc	aca	ctt	6039
Glu	Cys	Leu	Ile	Gly	Glu	His	Leu	His	Ala	Gly	Met	Ser	Thr	Leu	
2000						2005					2010				
ttt	ctg	gtg	tac	agc	aat	aag	tgt	cag	act	ccc	ctg	gga	atg	gct	6084
Phe	Leu	Val	Tyr	Ser	Asn	Lys	Cys	Gln	Thr	Pro	Leu	Gly	Met	Ala	
2015						2020					2025				
tct	gga	cac	att	aga	gat	ttt	cag	att	aca	gct	tca	gga	caa	tat	6129
Ser	Gly	His	Ile	Arg	Asp	Phe	Gln	Ile	Thr	Ala	Ser	Gly	Gln	Tyr	
2030						2035					2040				
gga	cag	tgg	gcc	cca	aag	ctg	gcc	aga	ctt	cat	tat	tcc	gga	tca	6174
Gly	Gln	Trp	Ala	Pro	Lys	Leu	Ala	Arg	Leu	His	Tyr	Ser	Gly	Ser	
2045						2050					2055				
atc	aat	gcc	tgg	agc	acc	aag	gag	ccc	ttt	tct	tgg	atc	aag	gtg	6219
Ile	Asn	Ala	Trp	Ser	Thr	Lys	Glu	Pro	Phe	Ser	Trp	Ile	Lys	Val	
2060						2065					2070				
gat	ctg	ttg	gca	cca	atg	att	att	cac	ggc	atc	aag	acc	cag	ggt	6264
Asp	Leu	Leu	Ala	Pro	Met	Ile	Ile	His	Gly	Ile	Lys	Thr	Gln	Gly	
2075						2080					2085				
gcc	cgt	cag	aag	tcc	tcc	agc	ctc	tac	atc	tct	cag	ttt	atc	atc	6309
Ala	Arg	Gln	Lys	Phe	Ser	Ser	Leu	Tyr	Ile	Ser	Gln	Phe	Ile	Ile	
2090						2095					2100				
atg	tat	agt	ctt	gat	ggg	aag	aag	tgg	cag	act	tat	cga	gga	aat	6354
Met	Tyr	Ser	Leu	Asp	Gly	Lys	Lys	Trp	Gln	Thr	Tyr	Arg	Gly	Asn	
2105						2110					2115				
tcc	act	gga	acc	tta	atg	gtc	tcc	ttt	ggc	aat	gtg	gat	tca	tct	6399
Ser	Thr	Gly	Thr	Leu	Met	Val	Phe	Phe	Gly	Asn	Val	Asp	Ser	Ser	
2120						2125					2130				
ggg	ata	aaa	cac	aat	att	ttt	aac	cct	cca	att	att	gct	cga	tac	6444
Gly	Ile	Lys	His	Asn	Ile	Phe	Asn	Pro	Pro	Ile	Ile	Ala	Arg	Tyr	
2135						2140					2145				
atc	cgt	ttg	cac	cca	act	cat	tat	agc	att	cgc	agc	act	ctt	cgc	6489
Ile	Arg	Leu	His	Pro	Thr	His	Tyr	Ser	Ile	Arg	Ser	Thr	Leu	Arg	
2150						2155					2160				
atg	gag	ttg	atg	ggc	tgt	gat	tta	aat	agt	tgc	agc	atg	cca	ttg	6534
Met	Glu	Leu	Met	Gly	Cys	Asp	Leu	Asn	Ser	Cys	Ser	Met	Pro	Leu	
2165						2170					2175				
gga	atg	gag	agt	aaa	gca	ata	tca	gat	gca	cag	att	act	gct	tca	6579
Gly	Met	Glu	Ser	Lys	Ala	Ile	Ser	Asp	Ala	Gln	Ile	Thr	Ala	Ser	
2180						2185					2190				
tcc	tac	ttt	acc	aat	atg	ttt	gcc	acc	tgg	tct	cct	tca	aaa	gct	6624
Ser	Tyr	Phe	Thr	Asn	Met	Phe	Ala	Thr	Trp	Ser	Pro	Ser	Lys	Ala	
2195						2200					2205				
cga	ctt	cac	ctc	caa	ggg	agg	agt	aat	gcc	tgg	aga	cct	cag	gtg	6669
Arg	Leu	His	Leu	Gln	Gly	Arg	Ser	Asn	Ala	Trp	Arg	Pro	Gln	Val	
2210						2215					2220				
aat	aat	cca	aaa	gag	tgg	ctg	caa	gtg	gac	ttc	cag	aag	aca	atg	6714
Asn	Asn	Pro	Lys	Glu	Trp	Leu	Gln	Val	Asp	Phe	Gln	Lys	Thr	Met	
2225						2230					2235				
aaa	gtc	aca	gga	gta	act	act	cag	gga	gta	aaa	tct	ctg	ctt	acc	6759
Lys	Val	Thr	Gly	Val	Thr	Thr	Gln	Gly	Val	Lys	Ser	Leu	Leu	Thr	
2240						2245					2250				

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agc atg tat gtg aag gag ttc ctc atc tcc agc agt caa gat ggc	6804
Ser Met Tyr Val Lys Glu Phe Leu Ile Ser Ser Ser Gln Asp Gly	
2255 2260 2265	
cat cag tgg act ctc ttt ttt cag aat ggc aaa gta aag gtt ttt	6849
His Gln Trp Thr Leu Phe Phe Gln Asn Gly Lys Val Lys Val Phe	
2270 2275 2280	
cag gga aat caa gac tcc ttc aca cct gtg gtg aac tct cta gac	6894
Gln Gly Asn Gln Asp Ser Phe Thr Pro Val Val Asn Ser Leu Asp	
2285 2290 2295	
cca ccg tta ctg act cgc tac ctt cga att cac ccc cag agt tgg	6939
Pro Pro Leu Leu Thr Arg Tyr Leu Arg Ile His Pro Gln Ser Trp	
2300 2305 2310	
gtg cac cag att gcc ctg agg atg gag gtt ctg ggc tgc gag gca	6984
Val His Gln Ile Ala Leu Arg Met Glu Val Leu Gly Cys Glu Ala	
2315 2320 2325	
cag gac ctc tac	6996
Gln Asp Leu Tyr	
2330	

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<211> 2332
<212> PRT
<213> homo sapiens

<400> 2

Ala Thr Arg Arg Tyr Tyr Leu Gly Ala Val Glu Leu Ser Trp Asp Tyr
1 5 10 15

Met Gln Ser Asp Leu Gly Glu Leu Pro Val Asp Ala Arg Phe Pro Pro
20 25 30

Arg Val Pro Lys Ser Phe Pro Phe Asn Thr Ser Val Val Tyr Lys Lys
35 40 45

Thr Leu Phe Val Glu Phe Thr Asp His Leu Phe Asn Ile Ala Lys Pro
50 55 60

Arg Pro Pro Trp Met Gly Leu Leu Gly Pro Thr Ile Gln Ala Glu Val
65 70 75 80

Tyr Asp Thr Val Val Ile Thr Leu Lys Asn Met Ala Ser His Pro Val
85 90 95

Ser Leu His Ala Val Gly Val Ser Tyr Trp Lys Ala Ser Glu Gly Ala
100 105 110

Glu Tyr Asp Asp Gln Thr Ser Gln Arg Glu Lys Glu Asp Asp Lys Val
115 120 125

Phe Pro Gly Gly Ser His Thr Tyr Val Trp Gln Val Leu Lys Glu Asn
130 135 140

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Gly Pro Met Ala Ser Asp Pro Leu Cys Leu Thr Tyr Ser Tyr Leu Ser
145 150 155 160

His Val Asp Leu Val Lys Asp Leu Asn Ser Gly Leu Ile Gly Ala Leu
165 170 175

Leu Val Cys Arg Glu Gly Ser Leu Ala Lys Glu Lys Thr Gln Thr Leu
180 185 190

His Lys Phe Ile Leu Leu Phe Ala Val Phe Asp Glu Gly Lys Ser Trp
195 200 205

His Ser Glu Thr Lys Asn Ser Leu Met Gln Asp Arg Asp Ala Ala Ser
210 215 220

Ala Arg Ala Trp Pro Lys Met His Thr Val Asn Gly Tyr Val Asn Arg
225 230 235 240

Ser Leu Pro Gly Leu Ile Gly Cys His Arg Lys Ser Val Tyr Trp His
245 250 255

Val Ile Gly Met Gly Thr Thr Pro Glu Val His Ser Ile Phe Leu Glu
260 265 270

Gly His Thr Phe Leu Val Arg Asn His Arg Gln Ala Ser Leu Glu Ile
275 280 285

Ser Pro Ile Thr Phe Leu Thr Ala Gln Thr Leu Leu Met Asp Leu Gly
290 295 300

Gln Phe Leu Leu Phe Cys His Ile Ser Ser His Gln His Asp Gly Met
305 310 315 320

Glu Ala Tyr Val Lys Val Asp Ser Cys Pro Glu Glu Pro Gln Leu Arg
325 330 335

Met Lys Asn Asn Glu Glu Ala Glu Asp Tyr Asp Asp Asp Leu Thr Asp
340 345 350

Ser Glu Met Asp Val Val Arg Phe Asp Asp Asp Asn Ser Pro Ser Phe
355 360 365

Ile Gln Ile Arg Ser Val Ala Lys Lys His Pro Lys Thr Trp Val His
370 375 380

Tyr Ile Ala Ala Glu Glu Glu Asp Trp Asp Tyr Ala Pro Leu Val Leu
385 390 395 400

Ala Pro Asp Asp Arg Ser Tyr Lys Ser Gln Tyr Leu Asn Asn Gly Pro
405 410 415

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Gln Arg Ile Gly Arg Lys Tyr Lys Lys Val Arg Phe Met Ala Tyr Thr
420 425 430

Asp Glu Thr Phe Lys Thr Arg Glu Ala Ile Gln His Glu Ser Gly Ile
435 440 445

Leu Gly Pro Leu Leu Tyr Gly Glu Val Gly Asp Thr Leu Leu Ile Ile
450 455 460

Phe Lys Asn Gln Ala Ser Arg Pro Tyr Asn Ile Tyr Pro His Gly Ile
465 470 475 480

Thr Asp Val Arg Pro Leu Tyr Ser Arg Arg Leu Pro Lys Gly Val Lys
485 490 495

His Leu Lys Asp Phe Pro Ile Leu Pro Gly Glu Ile Phe Lys Tyr Lys
500 505 510

Trp Thr Val Thr Val Glu Asp Gly Pro Thr Lys Ser Asp Pro Arg Cys
515 520 525

Leu Thr Arg Tyr Tyr Ser Ser Phe Val Asn Met Glu Arg Asp Leu Ala
530 535 540

Ser Gly Leu Ile Gly Pro Leu Leu Ile Cys Tyr Lys Glu Ser Val Asp
545 550 555 560

Gln Arg Gly Asn Gln Ile Met Ser Asp Lys Arg Asn Val Ile Leu Phe
565 570 575

Ser Val Phe Asp Glu Asn Arg Ser Trp Tyr Leu Thr Glu Asn Ile Gln
580 585 590

Arg Phe Leu Pro Asn Pro Ala Gly Val Gln Leu Glu Asp Pro Glu Phe
595 600 605

Gln Ala Ser Asn Ile Met His Ser Ile Asn Gly Tyr Val Phe Asp Ser
610 615 620

Leu Gln Leu Ser Val Cys Leu His Glu Val Ala Tyr Trp Tyr Ile Leu
625 630 635 640

Ser Ile Gly Ala Gln Thr Asp Phe Leu Ser Val Phe Phe Ser Gly Tyr
645 650 655

Thr Phe Lys His Lys Met Val Tyr Glu Asp Thr Leu Thr Leu Phe Pro
660 665 670

Phe Ser Gly Glu Thr Val Phe Met Ser Met Glu Asn Pro Gly Leu Trp
675 680 685

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Ile Leu Gly Cys His Asn Ser Asp Phe Arg Asn Arg Gly Met Thr Ala
690 695 700

Leu Leu Lys Val Ser Ser Cys Asp Lys Asn Thr Gly Asp Tyr Tyr Glu
705 710 715 720

Asp Ser Tyr Glu Asp Ile Ser Ala Tyr Leu Leu Ser Lys Asn Asn Ala
725 730 735

Ile Glu Pro Arg Ser Phe Ser Gln Asn Ser Arg His Arg Ser Thr Arg
740 745 750

Gln Lys Gln Phe Asn Ala Thr Thr Ile Pro Glu Asn Asp Ile Glu Lys
755 760 765

Thr Asp Pro Trp Phe Ala His Arg Thr Pro Met Pro Lys Ile Gln Asn
770 775 780

Val Ser Ser Ser Asp Leu Leu Met Leu Leu Arg Gln Ser Pro Thr Pro
785 790 795 800

His Gly Leu Ser Leu Ser Asp Leu Gln Glu Ala Lys Tyr Glu Thr Phe
805 810 815

Ser Asp Asp Pro Ser Pro Gly Ala Ile Asp Ser Asn Asn Ser Leu Ser
820 825 830

Glu Met Thr His Phe Arg Pro Gln Leu His His Ser Gly Asp Met Val
835 840 845

Phe Thr Pro Glu Ser Gly Leu Gln Leu Arg Leu Asn Glu Lys Leu Gly
850 855 860

Thr Thr Ala Ala Thr Glu Leu Lys Lys Leu Asp Phe Lys Val Ser Ser
865 870 875 880

Thr Ser Asn Asn Leu Ile Ser Thr Ile Pro Ser Asp Asn Leu Ala Ala
885 890 895

Gly Thr Asp Asn Thr Ser Ser Leu Gly Pro Pro Ser Met Pro Val His
900 905 910

Tyr Asp Ser Gln Leu Asp Thr Thr Leu Phe Gly Lys Lys Ser Ser Pro
915 920 925

Leu Thr Glu Ser Gly Gly Pro Leu Ser Leu Ser Glu Glu Asn Asn Asp
930 935 940

Ser Lys Leu Leu Glu Ser Gly Leu Met Asn Ser Gln Glu Ser Ser Trp
945 950 955 960

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Gly Lys Asn Val Ser Ser Thr Glu Ser Gly Arg Leu Phe Lys Gly Lys
965 970 975

Arg Ala His Gly Pro Ala Leu Leu Thr Lys Asp Asn Ala Leu Phe Lys
980 985 990

Val Ser Ile Ser Leu Leu Lys Thr Asn Lys Thr Ser Asn Asn Ser Ala
995 1000 1005

Thr Asn Arg Lys Thr His Ile Asp Gly Pro Ser Leu Leu Ile Glu
1010 1015 1020

Asn Ser Pro Ser Val Trp Gln Asn Ile Leu Glu Ser Asp Thr Glu
1025 1030 1035

Phe Lys Lys Val Thr Pro Leu Ile His Asp Arg Met Leu Met Asp
1040 1045 1050

Lys Asn Ala Thr Ala Leu Arg Leu Asn His Met Ser Asn Lys Thr
1055 1060 1065

Thr Ser Ser Lys Asn Met Glu Met Val Gln Gln Lys Lys Glu Gly
1070 1075 1080

Pro Ile Pro Pro Asp Ala Gln Asn Pro Asp Met Ser Phe Phe Lys
1085 1090 1095

Met Leu Phe Leu Pro Glu Ser Ala Arg Trp Ile Gln Arg Thr His
1100 1105 1110

Gly Lys Asn Ser Leu Asn Ser Gly Gln Gly Pro Ser Pro Lys Gln
1115 1120 1125

Leu Val Ser Leu Gly Pro Glu Lys Ser Val Glu Gly Gln Asn Phe
1130 1135 1140

Leu Ser Glu Lys Asn Lys Val Val Val Gly Lys Gly Glu Phe Thr
1145 1150 1155

Lys Asp Val Gly Leu Lys Glu Met Val Phe Pro Ser Ser Arg Asn
1160 1165 1170

Leu Phe Leu Thr Asn Leu Asp Asn Leu His Glu Asn Asn Thr His
1175 1180 1185

Asn Gln Glu Lys Lys Ile Gln Glu Glu Ile Glu Lys Lys Glu Thr
1190 1195 1200

Leu Ile Gln Glu Asn Val Val Leu Pro Gln Ile His Thr Val Thr
1205 1210 1215

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Gly Thr Lys Asn Phe Met Lys Asn Leu Phe Leu Leu Ser Thr Arg
1220 1225 1230

Gln Asn Val Glu Gly Ser Tyr Asp Gly Ala Tyr Ala Pro Val Leu
1235 1240 1245

Gln Asp Phe Arg Ser Leu Asn Asp Ser Thr Asn Arg Thr Lys Lys
1250 1255 1260

His Thr Ala His Phe Ser Lys Lys Gly Glu Glu Glu Asn Leu Glu
1265 1270 1275

Gly Leu Gly Asn Gln Thr Lys Gln Ile Val Glu Lys Tyr Ala Cys
1280 1285 1290

Thr Thr Arg Ile Ser Pro Asn Thr Ser Gln Gln Asn Phe Val Thr
1295 1300 1305

Gln Arg Ser Lys Arg Ala Leu Lys Gln Phe Arg Leu Pro Leu Glu
1310 1315 1320

Glu Thr Glu Leu Glu Lys Arg Ile Ile Val Asp Asp Thr Ser Thr
1325 1330 1335

Gln Trp Ser Lys Asn Met Lys His Leu Thr Pro Ser Thr Leu Thr
1340 1345 1350

Gln Ile Asp Tyr Asn Glu Lys Glu Lys Gly Ala Ile Thr Gln Ser
1355 1360 1365

Pro Leu Ser Asp Cys Leu Thr Arg Ser His Ser Ile Pro Gln Ala
1370 1375 1380

Asn Arg Ser Pro Leu Pro Ile Ala Lys Val Ser Ser Phe Pro Ser
1385 1390 1395

Ile Arg Pro Ile Tyr Leu Thr Arg Val Leu Phe Gln Asp Asn Ser
1400 1405 1410

Ser His Leu Pro Ala Ala Ser Tyr Arg Lys Lys Asp Ser Gly Val
1415 1420 1425

Gln Glu Ser Ser His Phe Leu Gln Gly Ala Lys Lys Asn Asn Leu
1430 1435 1440

Ser Leu Ala Ile Leu Thr Leu Glu Met Thr Gly Asp Gln Arg Glu
1445 1450 1455

Val Gly Ser Leu Gly Thr Ser Ala Thr Asn Ser Val Thr Tyr Lys
1460 1465 1470

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Lys Val Glu Asn Thr Val Leu Pro Lys Pro Asp Leu Pro Lys Thr
1475 1480 1485

Ser Gly Lys Val Glu Leu Leu Pro Lys Val His Ile Tyr Gln Lys
1490 1495 1500

Asp Leu Phe Pro Thr Glu Thr Ser Asn Gly Ser Pro Gly His Leu
1505 1510 1515

Asp Leu Val Glu Gly Ser Leu Leu Gln Gly Thr Glu Gly Ala Ile
1520 1525 1530

Lys Trp Asn Glu Ala Asn Arg Pro Gly Lys Val Pro Phe Leu Arg
1535 1540 1545

Val Ala Thr Glu Ser Ser Ala Lys Thr Pro Ser Lys Leu Leu Asp
1550 1555 1560

Pro Leu Ala Trp Asp Asn His Tyr Gly Thr Gln Ile Pro Lys Glu
1565 1570 1575

Glu Trp Lys Ser Gln Glu Lys Ser Pro Glu Lys Thr Ala Phe Lys
1580 1585 1590

Lys Lys Asp Thr Ile Leu Ser Leu Asn Ala Cys Glu Ser Asn His
1595 1600 1605

Ala Ile Ala Ala Ile Asn Glu Gly Gln Asn Lys Pro Glu Ile Glu
1610 1615 1620

Val Thr Trp Ala Lys Gln Gly Arg Thr Glu Arg Leu Cys Ser Gln
1625 1630 1635

Asn Pro Pro Val Leu Lys Arg His Gln Arg Glu Ile Thr Arg Thr
1640 1645 1650

Thr Leu Gln Ser Asp Gln Glu Glu Ile Asp Tyr Asp Asp Thr Ile
1655 1660 1665

Ser Val Glu Met Lys Lys Glu Asp Phe Asp Ile Tyr Asp Glu Asp
1670 1675 1680

Glu Asn Gln Ser Pro Arg Ser Phe Gln Lys Lys Thr Arg His Tyr
1685 1690 1695

Phe Ile Ala Ala Val Glu Arg Leu Trp Asp Tyr Gly Met Ser Ser
1700 1705 1710

Ser Pro His Val Leu Arg Asn Arg Ala Gln Ser Gly Ser Val Pro
1715 1720 1725

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Gln Phe Lys Lys Val Val Phe Gln Glu Phe Thr Asp Gly Ser Phe
1730 1735 1740

Thr Gln Pro Leu Tyr Arg Gly Glu Leu Asn Glu His Leu Gly Leu
1745 1750 1755

Leu Gly Pro Tyr Ile Arg Ala Glu Val Glu Asp Asn Ile Met Val
1760 1765 1770

Thr Phe Arg Asn Gln Ala Ser Arg Pro Tyr Ser Phe Tyr Ser Ser
1775 1780 1785

Leu Ile Ser Tyr Glu Glu Asp Gln Arg Gln Gly Ala Glu Pro Arg
1790 1795 1800

Lys Asn Phe Val Lys Pro Asn Glu Thr Lys Thr Tyr Phe Trp Lys
1805 1810 1815

Val Gln His His Met Ala Pro Thr Lys Asp Glu Phe Asp Cys Lys
1820 1825 1830

Ala Trp Ala Tyr Phe Ser Asp Val Asp Leu Glu Lys Asp Val His
1835 1840 1845

Ser Gly Leu Ile Gly Pro Leu Leu Val Cys His Thr Asn Thr Leu
1850 1855 1860

Asn Pro Ala His Gly Arg Gln Val Thr Val Gln Glu Phe Ala Leu
1865 1870 1875

Phe Phe Thr Ile Phe Asp Glu Thr Lys Ser Trp Tyr Phe Thr Glu
1880 1885 1890

Asn Met Glu Arg Asn Cys Arg Ala Pro Cys Asn Ile Gln Met Glu
1895 1900 1905

Asp Pro Thr Phe Lys Glu Asn Tyr Arg Phe His Ala Ile Asn Gly
1910 1915 1920

Tyr Ile Met Asp Thr Leu Pro Gly Leu Val Met Ala Gln Asp Gln
1925 1930 1935

Arg Ile Arg Trp Tyr Leu Leu Ser Met Gly Ser Asn Glu Asn Ile
1940 1945 1950

His Ser Ile His Phe Ser Gly His Val Phe Thr Val Arg Lys Lys
1955 1960 1965

Glu Glu Tyr Lys Met Ala Leu Tyr Asn Leu Tyr Pro Gly Val Phe
1970 1975 1980

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Glu Thr Val Glu Met Leu Pro Ser Lys Ala Gly Ile Trp Arg Val
1985 1990 1995

Glu Cys Leu Ile Gly Glu His Leu His Ala Gly Met Ser Thr Leu
2000 2005 2010

Phe Leu Val Tyr Ser Asn Lys Cys Gln Thr Pro Leu Gly Met Ala
2015 2020 2025

Ser Gly His Ile Arg Asp Phe Gln Ile Thr Ala Ser Gly Gln Tyr
2030 2035 2040

Gly Gln Trp Ala Pro Lys Leu Ala Arg Leu His Tyr Ser Gly Ser
2045 2050 2055

Ile Asn Ala Trp Ser Thr Lys Glu Pro Phe Ser Trp Ile Lys Val
2060 2065 2070

Asp Leu Leu Ala Pro Met Ile Ile His Gly Ile Lys Thr Gln Gly
2075 2080 2085

Ala Arg Gln Lys Phe Ser Ser Leu Tyr Ile Ser Gln Phe Ile Ile
2090 2095 2100

Met Tyr Ser Leu Asp Gly Lys Lys Trp Gln Thr Tyr Arg Gly Asn
2105 2110 2115

Ser Thr Gly Thr Leu Met Val Phe Phe Gly Asn Val Asp Ser Ser
2120 2125 2130

Gly Ile Lys His Asn Ile Phe Asn Pro Pro Ile Ile Ala Arg Tyr
2135 2140 2145

Ile Arg Leu His Pro Thr His Tyr Ser Ile Arg Ser Thr Leu Arg
2150 2155 2160

Met Glu Leu Met Gly Cys Asp Leu Asn Ser Cys Ser Met Pro Leu
2165 2170 2175

Gly Met Glu Ser Lys Ala Ile Ser Asp Ala Gln Ile Thr Ala Ser
2180 2185 2190

Ser Tyr Phe Thr Asn Met Phe Ala Thr Trp Ser Pro Ser Lys Ala
2195 2200 2205

Arg Leu His Leu Gln Gly Arg Ser Asn Ala Trp Arg Pro Gln Val
2210 2215 2220

Asn Asn Pro Lys Glu Trp Leu Gln Val Asp Phe Gln Lys Thr Met
2225 2230 2235

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Lys Val Thr Gly Val Thr Thr Gln Gly Val Lys Ser Leu Leu Thr
2240 2245 2250

Ser Met Tyr Val Lys Glu Phe Leu Ile Ser Ser Ser Gln Asp Gly
2255 2260 2265

His Gln Trp Thr Leu Phe Phe Gln Asn Gly Lys Val Lys Val Phe
2270 2275 2280

Gln Gly Asn Gln Asp Ser Phe Thr Pro Val Val Asn Ser Leu Asp
2285 2290 2295

Pro Pro Leu Leu Thr Arg Tyr Leu Arg Ile His Pro Gln Ser Trp
2300 2305 2310

Val His Gln Ile Ala Leu Arg Met Glu Val Leu Gly Cys Glu Ala
2315 2320 2325

Gln Asp Leu Tyr
2330