



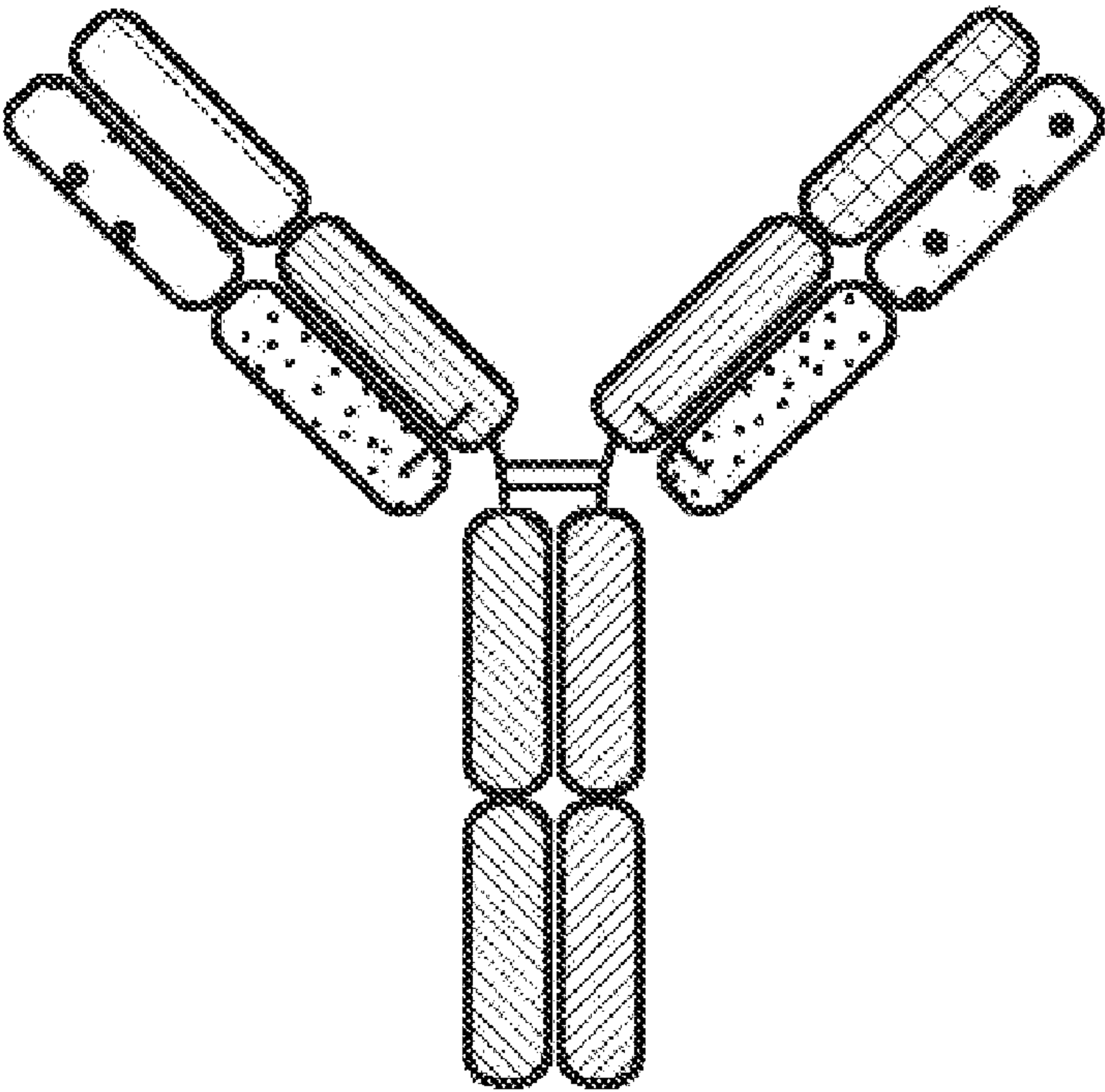
(12) **DEMANDE DE BREVET CANADIEN
CANADIAN PATENT APPLICATION**

(13) **A1**

(86) Date de dépôt PCT/PCT Filing Date: 2018/02/20 (87) Date publication PCT/PCT Publication Date: 2018/08/23 (85) Entrée phase nationale/National Entry: 2019/08/19 (86) N° demande PCT/PCT Application No.: US 2018/018771 (87) N° publication PCT/PCT Publication No.: 2018/152518 (30) Priorité/Priority: 2017/02/20 (US62/461,146)	(51) Cl.Int./Int.Cl. <i>C07K 16/46</i> (2006.01), <i>A61K 39/395</i> (2006.01), <i>A61P 35/00</i> (2006.01), <i>C07K 16/28</i> (2006.01), <i>C12N 5/10</i> (2006.01) (71) Demandeur/Applicant: DRAGONFLY THERAPEUTICS, INC., US (72) Inventeurs/Inventors: CHANG, GREGORY P., US; CHEUNG, ANN F., US; HANEY, WILLIAM, US; LUNDE, BRADLEY M., US; PRINZ, BIANKA, US (74) Agent: BORDEN LADNER GERVAIS LLP
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(54) Titre : PROTEINES SE LIANT A HER2, NKG2D ET CD16
(54) Title: PROTEINS BINDING HER2, NKG2D AND CD16

FIG. 1



(57) **Abrégé/Abstract:**
Multi-specific binding proteins that bind HER2, the NKG2D receptor, and CD 16 are described, as well as pharmaceutical compositions and therapeutic methods useful for the treatment of cancer.

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property
Organization
International Bureau

(43) International Publication Date
23 August 2018 (23.08.2018)



(10) International Publication Number
WO 2018/152518 A1

(51) International Patent Classification:

A61K 39/00 (2006.01) C12P 21/04 (2006.01)
A61K 39/395 (2006.01) C12P 21/08 (2006.01)
C12N 5/16 (2006.01) C07K 16/00 (2006.01)

(21) International Application Number:

PCT/US2018/018771

(22) International Filing Date:

20 February 2018 (20.02.2018)

(25) Filing Language:

English

(26) Publication Language:

English

(30) Priority Data:

62/461,146 20 February 2017 (20.02.2017) US

(71) Applicant: ADIMAB, LLC [US/US]; 7 Lucent Drive, Lebanon, NH 03766 (US).

(72) Inventors: CHANG, Gregory, P.; 143 Saunders Street, Medford, MA 02155 (US). CHEUNG, Ann, F.; 25 Morningside Lane, Lincoln, MA 01773 (US). HANEY, William; 61 Lincoln Road, Wayland, MA 01778 (US). LUNDE, Bradley, M.; 7 Lucent Drive, Lebanon, NH 03766 (US). PRINZ, Bianka; 7 Lucent Drive, Lebanon, NH 03766 (US).

(74) Agent: ASHRAF, Shovon et al.; Goodwin Procter LLP, 100 Northern Avenue, Boston, MA 02210 (US).

(81) Designated States (unless otherwise indicated, for every kind of national protection available): AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DJ, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JO, JP, KE, KG, KH, KN, KP, KR, KW, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME,

MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States (unless otherwise indicated, for every kind of regional protection available):

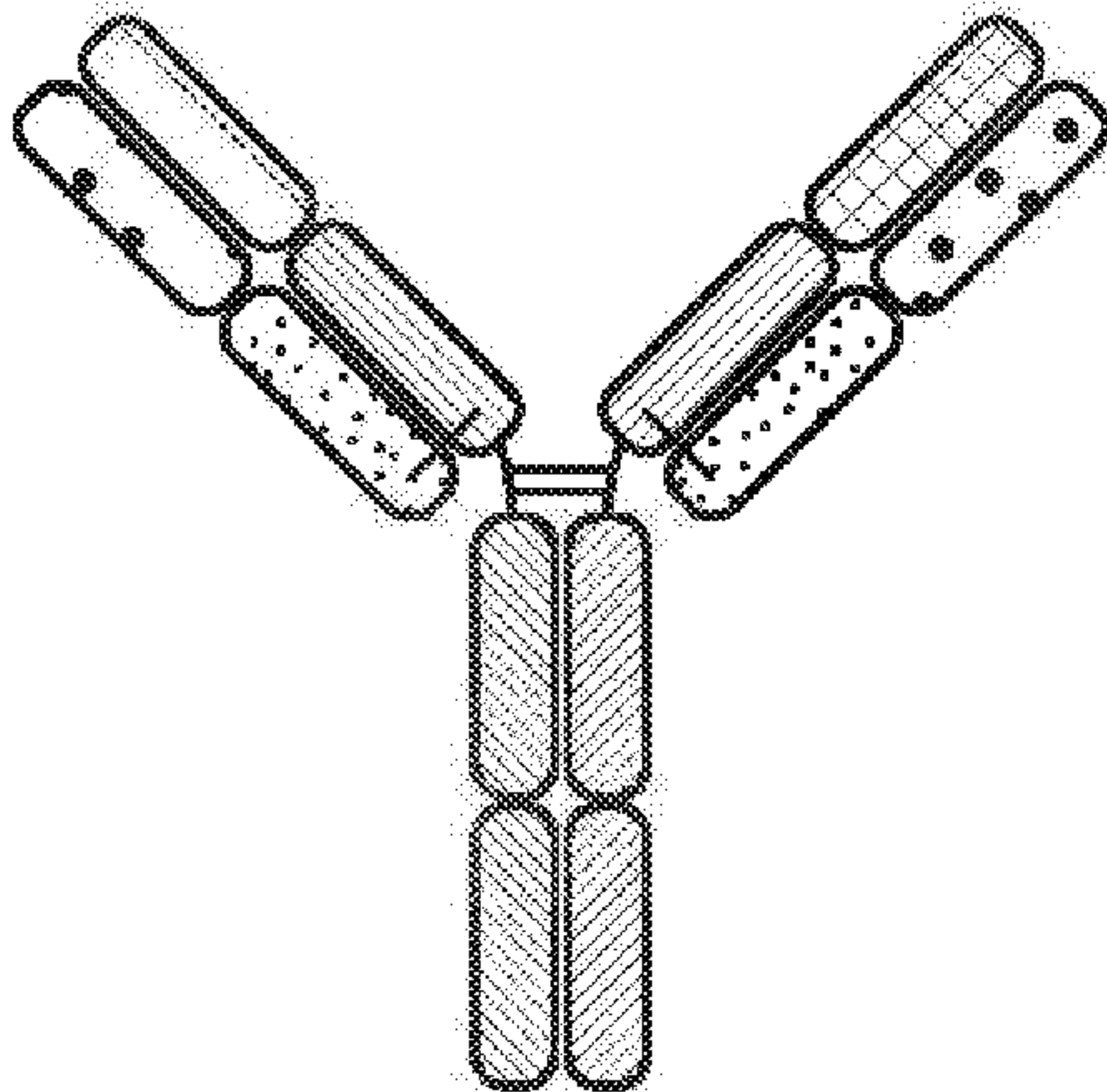
ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Published:

- with international search report (Art. 21(3))
- with sequence listing part of description (Rule 5.2(a))

(54) Title: PROTEINS BINDING HER2, NKG2D AND CD16

FIG. 1



(57) Abstract: Multi-specific binding proteins that bind HER2, the NKG2D receptor, and CD 16 are described, as well as pharmaceutical compositions and therapeutic methods useful for the treatment of cancer.

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PROTEINS BINDING HER2, NKG2D AND CD16

CROSS-REFERENCE TO RELATED APPLICATIONS

[0001] This application claims the benefit of and priority to U.S. Provisional Patent Application No. 62/461,146, filed February 20, 2017, the entire contents of which are
5 incorporated by reference herein for all purposes.

SEQUENCE LISTING

[0002] The instant application contains a Sequence Listing which has been submitted electronically in ASCII format and is hereby incorporated by reference in its entirety. Said ASCII copy, created on February 16, 2018, is named DFY-008PC_SL.txt and is 92,807 bytes
10 in size.

FIELD OF THE INVENTION

[0003] The invention relates to multi-specific binding proteins that bind to human epidermal growth factor receptor 2 (HER2 or ErbB2), the NKG2D receptor, and CD16.

BACKGROUND

15 [0004] Cancer continues to be a significant health problem despite the substantial research efforts and scientific advances reported in the literature for treating this disease. Some of the most frequently diagnosed cancers include prostate cancer, breast cancer, and lung cancer. Prostate cancer is the most common form of cancer in men. Breast cancer remains a leading cause of death in women. Current treatment options for these cancers are
20 not effective for all patients and/or can have substantial adverse side effects. Other types of cancer also remain challenging to treat using existing therapeutic options.

[0005] Cancer immunotherapies are desirable because they are highly specific and can facilitate destruction of cancer cells using the patient's own immune system. Fusion proteins such as bi-specific T-cell engagers are cancer immunotherapies described in the literature that
25 bind to tumor cells and T-cells to facilitate destruction of tumor cells. Antibodies that bind to certain tumor-associated antigens and to certain immune cells have been described in the literature. *See, e.g.*, WO 2016/134371 and WO 2015/095412.

[0006] Natural killer (NK) cells are a component of the innate immune system and make up approximately 15% of circulating lymphocytes. NK cells infiltrate virtually all tissues and
30 were originally characterized by their ability to kill tumor cells effectively without the need for prior sensitization. Activated NK cells kill target cells by means similar to cytotoxic T

cells – *i.e.*, via cytolytic granules that contain perforin and granzymes as well as via death receptor pathways. Activated NK cells also secrete inflammatory cytokines such as IFN- γ and chemokines that promote the recruitment of other leukocytes to the target tissue.

[0007] NK cells respond to signals through a variety of activating and inhibitory receptors on their surface. For example, when NK cells encounter healthy self-cells, their activity is inhibited through activation of the killer-cell immunoglobulin-like receptors (KIRs). Alternatively, when NK cells encounter foreign cells or cancer cells, they are activated via their activating receptors (*e.g.*, NKG2D, NCRs, DNAM1). NK cells are also activated by the constant region of some immunoglobulins through CD16 receptors on their surface. The overall sensitivity of NK cells to activation depends on the sum of stimulatory and inhibitory signals.

[0008] HER2 (ErbB2) is a transmembrane glycoprotein, which belongs to the epidermal growth factor receptor family. It is a receptor tyrosine kinase and regulates cell survival, proliferation, and growth. HER2 plays an important role in human malignancies.

The *erbB2* gene is amplified or overexpressed in approximately 30% of human breast cancers. Patients with HER2-overexpressing breast cancer have substantially lower overall survival rates and shorter disease-free intervals than patients whose cancer does not overexpress HER2. Moreover, overexpression of HER2 leads to increased breast cancer metastasis. Over-expression of HER2 is also known to occur in many other cancer types, including breast, ovarian, esophageal, bladder and gastric cancer, salivary duct carcinoma, adenocarcinoma of the lung and aggressive forms of uterine cancer, such as uterine serous endometrial carcinoma.

SUMMARY

[0009] The invention provides multi-specific binding proteins that bind to HER2 on a cancer cell and to the NKG2D receptor and CD16 receptor on natural killer cells. Such proteins can engage more than one kind of NK activating receptor, and may block the binding of natural ligands to NKG2D. In certain embodiments, the proteins can agonize NK cells in humans, and in other species such as rodents and cynomolgus monkeys. Various aspects and embodiments of the invention are described in further detail below.

[0010] Accordingly, one aspect of the invention provides a protein that incorporates a first antigen-binding site that binds NKG2D; a second antigen-binding site that binds to HER2; and an antibody Fc domain, a portion thereof sufficient to bind CD16, or a third antigen-binding site that binds CD16. The antigen-binding sites may each incorporate an

antibody heavy chain variable domain and an antibody light chain variable domain (*e.g.*, arranged as in an antibody, or fused together to form an scFv, or one or more of the antigen-binding sites may be a single domain antibody, such as a V_HH antibody like a camelid antibody or a V_{NAR} antibody like those found in cartilaginous fish.

5 [0011] The first antigen-binding site, which binds to NKG2D, in one embodiment, can incorporate a heavy chain variable domain related to SEQ ID NO:1, such as by having an amino acid sequence at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:1, and/or incorporating amino acid sequences identical to the CDR1 (SEQ ID NO:62), CDR2 (SEQ ID NO:63), and CDR3 (SEQ ID NO:64) sequences of SEQ ID NO:1. Alternatively, the first antigen-binding site can incorporate a heavy chain variable domain related to SEQ ID NO:41 and a light chain variable domain related to SEQ ID NO:42. For example, the heavy chain variable domain of the first antigen binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:41, and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:65), CDR2 (SEQ ID NO:66), and CDR3 (SEQ ID NO:67) sequences of SEQ ID NO:41. Similarly, the light chain variable domain of the second antigen-binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:42, and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:68), CDR2 (SEQ ID NO:69), and CDR3 (SEQ ID NO:70) sequences of SEQ ID NO:42. In other embodiments, the first antigen-binding site can incorporate a heavy chain variable domain related to SEQ ID NO:43 and a light chain variable domain related to SEQ ID NO:44. For example, the heavy chain variable domain of the first antigen-binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:43, and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:71), CDR2 (SEQ ID NO:72), and CDR3 (SEQ ID NO:73) sequences of SEQ ID NO:43. Similarly, the light chain variable domain of the second antigen-binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:44, and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:74), CDR2 (SEQ ID NO:75), and CDR3 (SEQ ID NO:76) sequences of SEQ ID NO:44.

[0012] Alternatively, the first antigen-binding site can incorporate a heavy chain variable domain related to SEQ ID NO:45 and a light chain variable domain related to SEQ ID NO:46, such as by having amino acid sequences at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:45 and at least 90%

(*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:46 respectively. In another embodiment, the first antigen-binding site can incorporate a heavy chain variable domain related to SEQ ID NO:47 and a light chain variable domain related to SEQ ID NO:48, such as by having amino acid sequences at least 90% (*e.g.*, 90%,
 5 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:47 and at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:48 respectively.

[0013] The second antigen-binding site can optionally incorporate a heavy chain variable domain related to SEQ ID NO:49 and a light chain variable domain related to SEQ ID
 10 NO:53. For example, the heavy chain variable domain of the second antigen-binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:49, and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:50), CDR2 (SEQ ID NO:51), and CDR3 (SEQ ID NO:52) sequences of SEQ ID NO:49. Similarly, the light chain variable domain of the second antigen-binding site can
 15 be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:53 and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:54), CDR2 (SEQ ID NO:55), and CDR3 (SEQ ID NO:56) sequences of SEQ ID NO:53.

[0014] Alternatively, the second antigen-binding site can incorporate a heavy chain
 20 variable domain related to SEQ ID NO:57 and a light chain variable domain related to SEQ ID NO:58. For example, the heavy chain variable domain of the second antigen-binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:57, and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:77), CDR2 (SEQ ID NO:78), and CDR3 (SEQ ID NO:79) sequences of SEQ
 25 ID NO:57. Similarly, the light chain variable domain of the second antigen-binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:58, and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:80), CDR2 (SEQ ID NO:81), and CDR3 (SEQ ID NO:82) sequences of SEQ ID NO:58.

[0015] In another embodiment, the second antigen-binding site can incorporate a heavy
 30 chain variable domain related to SEQ ID NO:59 and a light chain variable domain related to SEQ ID NO:60. For example, the heavy chain variable domain of the second antigen-binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:59, and/or incorporate amino acid sequences identical to the

CDR1 (SEQ ID NO:83), CDR2 (SEQ ID NO:84), and CDR3 (SEQ ID NO:85) sequences of SEQ ID NO:59. Similarly, the light chain variable domain of the second antigen-binding site can be at least 90% (*e.g.*, 90%, 91%, 92%, 93%, 94%, 95%, 96%, 97%, 98%, 99%, or 100%) identical to SEQ ID NO:60, and/or incorporate amino acid sequences identical to the CDR1 (SEQ ID NO:86), CDR2 (SEQ ID NO:87), and CDR3 (SEQ ID NO:88) sequences of SEQ ID NO:60.

[0016] In some embodiments, the second antigen-binding site incorporates a light chain variable domain having an amino acid sequence identical to the amino acid sequence of the light chain variable domain present in the first antigen-binding site.

[0017] In some embodiments, the protein incorporates a portion of an antibody Fc domain sufficient to bind CD16, wherein the antibody Fc domain comprises hinge and CH2 domains, and/or amino acid sequences at least 90% identical to amino acid sequence 234-332 of a human IgG antibody.

[0018] Formulations containing one of these proteins; cells containing one or more nucleic acids expressing these proteins, and methods of enhancing tumor cell death using these proteins are also provided.

[0019] Another aspect of the invention involves a method of treating cancer in a patient. The method comprises administering to a patient in need thereof a therapeutically effective amount of the multi-specific binding protein described herein. Exemplary cancers for treatment using the multi-specific binding proteins include, for example, breast, ovarian, esophageal, bladder and gastric cancer, salivary duct carcinoma, adenocarcinoma of the lung and aggressive forms of uterine cancer, such as uterine serous endometrial carcinoma.

BRIEF DESCRIPTION OF THE DRAWINGS

[0020] **FIG. 1** is a representation of a multi-specific binding protein that contains an NKG2D-binding domain (right arm), a tumor associated antigen-binding domain (left arm) and an Fc domain or a portion thereof that binds to CD16.

[0021] **FIG. 2** is a representation of a multi-specific binding protein that contains an NKG2D-binding domain in a scFv format (right arm), a tumor associated antigen-binding domain (left arm) and an Fc domain or a portion thereof that binds to CD16.

[0022] **FIG. 3** are line graphs demonstrating the binding affinity of NKG2D-binding domains (listed as clones) to human recombinant NKG2D in an ELISA assay.

[0023] **FIG. 4** are line graphs demonstrating the binding affinity of NKG2D-binding domains (listed as clones) to cynomolgus recombinant NKG2D in an ELISA assay.

[0024] FIG. 5 are line graphs demonstrating the binding affinity of NKG2D-binding domains (listed as clones) to mouse recombinant NKG2D in an ELISA assay.

[0025] FIG. 6 are bar graphs demonstrating the binding of NKG2D-binding domains (listed as clones) to EL4 cells expressing human NKG2D by flow cytometry showing mean
5 fluorescence intensity (MFI) fold over background.

[0026] FIG. 7 are bar graphs demonstrating the binding of NKG2D-binding domains (listed as clones) to EL4 cells expressing mouse NKG2D by flow cytometry showing mean fluorescence intensity (MFI) fold over background.

[0027] FIG. 8 are line graphs demonstrating specific binding affinity of NKG2D-binding
10 domains (listed as clones) to recombinant human NKG2D-Fc by competing with natural ligand ULBP-6.

[0028] FIG. 9 are line graphs demonstrating specific binding affinity of NKG2D-binding domains (listed as clones) to recombinant human NKG2D-Fc by competing with natural ligand MICA.

15 [0029] FIG. 10 are line graphs demonstrating specific binding affinity of NKG2D-binding domains (listed as clones) to recombinant mouse NKG2D-Fc by competing with natural ligand Rae-1 delta.

[0030] FIG. 11 are bar graphs showing activation of human NKG2D by NKG2D-binding domains (listed as clones) by quantifying the percentage of TNF α -positive cells, which
20 express human NKG2D-CD3 zeta fusion proteins.

[0031] FIG. 12 are bar graphs showing activation of mouse NKG2D by NKG2D-binding domains (listed as clones) by quantifying the percentage of TNF α -positive cells, which express mouse NKG2D-CD3 zeta fusion proteins.

[0032] FIG. 13 are bar graphs showing activation of human NK cells by NKG2D-
25 binding domains (listed as clones).

[0033] FIG. 14 are bar graphs showing activation of human NK cells by NKG2D-binding domains (listed as clones).

[0034] FIG. 15 are bar graphs showing activation of mouse NK cells by NKG2D-binding domains (listed as clones).

30 [0035] FIG. 16 are bar graphs showing activation of mouse NK cells by NKG2D-binding domains (listed as clones).

[0036] FIG. 17 are bar graphs showing the cytotoxic effect of NKG2D-binding domains (listed as clones) on tumor cells.

[0037] FIG. 18 are bar graphs showing the melting temperature of NKG2D-binding domains (listed as clones) measured by differential scanning fluorimetry.

[0038] FIG. 19 is a graph showing enhanced activation of human NK cells by multi-specific binding proteins.

5 [0039] FIG. 20 is a graph showing multi-specific binding proteins induced higher levels of cytotoxicity towards tumor target cells by human NK cells.

[0040] FIG. 21 is a graph showing multi-specific binding proteins induced higher levels of cytotoxicity towards tumor target cells by human NK cells.

[0041] FIG. 22 is a graph showing multi-specific binding proteins induced higher levels
10 of cytotoxicity towards tumor target cells by human NK cells.

[0042] FIG. 23 is a graph showing multi-specific binding proteins induced higher levels of cytotoxicity towards tumor target cells by human NK cells.

[0043] FIG. 24 is a graph showing multi-specific binding proteins induced higher levels of cytotoxicity towards tumor target cells by mouse NK cells.

15 [0044] FIG. 25 is a graph showing multi-specific binding proteins induced higher levels of cytotoxicity towards tumor target cells by mouse NK cells.

[0045] FIG. 26 is a binding profile of HER2-targeting TriNKETs to NKG2D expressed on EL4 cells. FIG. 26 represents the same two NKG2D-binding domains now paired with a HER2 second targeting arm.

20 [0046] FIG. 27A is a binding profile of HER2-targeting TriNKETs to HER2 expressed on human 786-O renal cell carcinoma cells; FIG. 27B shows that NKG2D binding clone C26 containing TriNKET binds to RMA cells transduced with human HER2; FIG. 27C shows NKG2D binding clone F04 containing TriNKET binds to RMA cells transduced with human HER2.

25 [0047] FIGs. 28A – 28C are bar graphs demonstrating that TriNKETs and trastuzumab were able to activate primary human NK cells in co-culture with HER2-positive human tumor cells, indicated by an increase in CD107a degranulation and IFN γ cytokine production. Compared to the monoclonal antibody trastuzumab, both TriNKETs showed superior activation of human NK cells with a variety of human HER2 cancer cells. FIG. 28A shows
30 that human NK cells are activated by TriNKETs when cultured with SkBr-3 cells. FIG. 28B shows that human NK cells are activated by TriNKETs when cultured with Colo201 cells. FIG. 28C shows that human NK cell are activated by TriNKETs when cultured with HCC1954 cells.

[0048] **FIGs. 29A-29B** are graphs demonstrating TriNKETs provide the greater advantage against HER2 medium and low cancers compared to trastuzumab. FIG. 29A shows activated human NK cell killing of HER2 high-SkBr-3 tumor cells. FIG. 29B shows human NK cell killing of HER2 low-786-O tumor cells. TriNKETs provide a greater advantage compared to trastuzumab against cancer cells with low HER2 expression.

[0049] **FIGs. 30A-30C** are bar graphs of synergistic activation of NK cells using CD16 and NKG2D. FIG. 30A demonstrates levels of CD107a; FIG. 30B demonstrates levels of IFN γ ; FIG. 30C demonstrates levels of CD107a and IFN γ . Graphs indicate the mean (n = 2) $\pm$ SD. Data are representative of five independent experiments using five different healthy donors.

[0050] **FIG. 31** is a bar graph showing activation of NK cells using TriNKETs targeting NKG2D and CD16. Antibodies tested were of human IgG1 isotypes. Graphs indicate the mean (n = 2) $\pm$ SD.

[0051] **FIGs. 32A – 32C** are graphs demonstrating TriNKET enhancement of cytotoxic activity using IL-2-activated and rested human NK cells. FIG. 32A shows percent specific lysis of SkBr-3 tumor cells by rested human NK cells. FIG. 32B shows percent specific lysis of SkBr-3 tumor cells by IL-2-activated human NK cells. FIG. 32C shows percent specific lysis of NCI-H661 lung cancer cells by IL-2-activated human NK cells.

[0052] **FIGs. 33A & 33B** are bar graphs showing B cells from a health donor are sensitive to TriNKET-mediated lysis. **FIGs. 33C & 33D** are bar graphs showing myeloid cells are resistant to TriNKET-mediated lysis.

[0053] **FIG. 34** are line graphs of TriNKETs-mediated hPBMC killing of SkBr-3 tumor cells in long-term co-cultures.

[0054] **FIG. 35** is a line graph showing tri-specific binding in one molecule is important for maximal NK cell activity.

[0055] **FIG. 36** is a flowchart of study design of RMA/S-HER2 subcutaneous SC2.2 efficacy.

[0056] **FIG. 37** are line graphs showing that SC2.2 has no effect on subcutaneous RMA/S-HER2 tumor growth.

[0057] **FIG. 38A** shows that HER2-TriNKET-C26 bridges hNKG2D-Fc to RMA-HER2 cells. **FIG. 38B** shows HER2-TriNKET-F04 bridges hNKG2D-Fc to RMA-HER2 cells. Dotted line represents isotype control. Solid line without fill represents unstained control. Solid line with fill represents the TriNKETs.

[0058] FIG. 39 is a representation of a TriNKET in the Triomab form, which is a trifunctional, bispecific antibody that maintains an IgG-like shape. This chimera consists of two half antibodies, each with one light and one heavy chain, that originate from two parental antibodies. Triomab form may be an heterodimeric construct containing 1/2 of rat antibody
5 and 1/2 of mouse antibody.

[0059] FIG. 40 is a representation of a TriNKET in the KiH Common Light Chain (LC) form, which involves the knobs-into-holes (KIHS) technology. KiH is a heterodimer containing 2 Fabs binding to target 1 and 2, and an Fc stabilized by heterodimerization mutations. TriNKET in the KiH format may be an heterodimeric construct with 2 fabs
10 binding to target 1 and target 2, containing two different heavy chains and a common light chain that pairs with both heavy chains.

[0060] FIG. 41 is a representation of a TriNKET in the dual-variable domain immunoglobulin (DVD-IgTM) form, which combines the target binding domains of two monoclonal antibodies via flexible naturally occurring linkers, and yields a tetravalent IgG -
15 like molecule. DVD-IgTM is an homodimeric construct where variable domain targeting antigen 2 is fused to the N terminus of variable domain of Fab targeting antigen 1 Construct contains normal Fc.

[0061] FIG. 42 is a representation of a TriNKET in the Orthogonal Fab interface (Ortho-Fab) form, which is an heterodimeric construct that contains 2 Fabs binding to target1 and
20 target 2 fused to Fc. LC-HC pairing is ensured by orthogonal interface. Heterodimerization is ensured by mutations in the Fc.

[0062] FIG. 43 is a representation of a TrinKET in the 2-in-1 Ig format.

[0063] FIG. 44 is a representation of a TriNKET in the ES form, which is an heterodimeric construct containing two different Fabs binding to target 1 and target 2 fused to
25 the Fc. Heterodimerization is ensured by electrostatic steering mutations in the Fc.

[0064] FIG. 45 is a representation of a TriNKET in the Fab Arm Exchange form: antibodies that exchange Fab arms by swapping a heavy chain and attached light chain (half-molecule) with a heavy-light chain pair from another molecule, resulting in bispecific antibodies. Fab Arm Exchange form (cFae) is a heterodimer containing 2 Fabs binding to
30 target 1 and 2, and an Fc stabilized by heterodimerization mutations.

[0065] FIG. 46 is a representation of a TriNKET in the SEED Body form, which is an heterodimer containing 2 Fabs binding to target 1 and 2, and an Fc stabilized by heterodimerization mutations.

[0066] FIG. 47 is a representation of a TriNKET in the LuZ-Y form, in which leucine zipper is used to induce heterodimerization of two different HCs. LuZ-Y form is a heterodimer containing two different scFabs binding to target 1 and 2, fused to Fc. Heterodimerization is ensured through leucine zipper motifs fused to C-terminus of Fc.

5 [0067] FIG. 48 is a representation of a TriNKET in the Cov-X-Body form.

[0068] FIGs. 49A-49B are representations of TriNKETs in the $\kappa\lambda$ -Body forms, which are an heterodimeric constructs with two different Fabs fused to Fc stabilized by heterodimerization mutations: Fab1 targeting antigen 1 contains kappa LC, while second Fab targeting antigen 2 contains lambda LC. FIG. 49A is an exemplary representation of one form
10 of a $\kappa\lambda$ -Body; FIG. 49B is an exemplary representation of another $\kappa\lambda$ -Body.

[0069] FIG. 50 is an Oasc-Fab heterodimeric construct that includes Fab binding to target 1 and scFab binding to target 2 fused to Fc. Heterodimerization is ensured by mutations in the Fc.

[0070] FIG. 51 is a DuetMab, which is an heterodimeric construct containing two
15 different Fabs binding to antigens 1 and 2, and Fc stabilized by heterodimerization mutations. Fab 1 and 2 contain differential S-S bridges that ensure correct light chain (LC) and heavy chain (HC) pairing.

[0071] FIG. 52 is a CrossmAb, which is an heterodimeric construct with two different Fabs binding to targets 1 and 2 fused to Fc stabilized by heterodimerization. CL and CH1
20 domains and VH and VL domains are switched, *e.g.*, CH1 is fused in-line with VL, while CL is fused in-line with VH.

[0072] FIG. 53 is a Fit-Ig, which is an homodimeric constructs where Fab binding to antigen 2 is fused to the N terminus of HC of Fab that binds to antigen 1. The construct contains wild-type Fc.

25

DETAILED DESCRIPTION

[0073] The invention provides multi-specific binding proteins that bind a HER2 on a cancer cell and the NKG2D receptor and CD16 receptor on natural killer cells to activate the natural killer cell, pharmaceutical compositions comprising such multi-specific binding
30 proteins, and therapeutic methods using such multi-specific proteins and pharmaceutical compositions, including for the treatment of cancer. Various aspects of the invention are set forth below in sections; however, aspects of the invention described in one particular section are not to be limited to any particular section.

[0074] To facilitate an understanding of the present invention, a number of terms and phrases are defined below.

[0075] The terms “a” and “an” as used herein mean “one or more” and include the plural unless the context is inappropriate. As used herein, the term “antigen-binding site” refers to the part of the immunoglobulin molecule that participates in antigen binding. In human antibodies, the antigen-binding site is formed by amino acid residues of the N-terminal variable (“V”) regions of the heavy (“H”) and light (“L”) chains. Three highly divergent stretches within the V regions of the heavy and light chains are referred to as “hypervariable regions” which are interposed between more conserved flanking stretches known as “framework regions,” or “FRs.” Thus the term “FR” refers to amino acid sequences which are naturally found between and adjacent to hypervariable regions in immunoglobulins. In a human antibody molecule, the three hypervariable regions of a light chain and the three hypervariable regions of a heavy chain are disposed relative to each other in three dimensional space to form an antigen-binding surface. The antigen-binding surface is complementary to the three-dimensional surface of a bound antigen, and the three hypervariable regions of each of the heavy and light chains are referred to as “complementarity-determining regions,” or “CDRs.” In certain animals, such as camels and cartilaginous fish, the antigen-binding site is formed by a single antibody chain providing a “single domain antibody.” Antigen-binding sites can exist in an intact antibody, in an antigen-binding fragment of an antibody that retains the antigen-binding surface, or in a recombinant polypeptide such as an scFv, using a peptide linker to connect the heavy chain variable domain to the light chain variable domain in a single polypeptide.

[0076] The term “tumor associated antigen” as used herein means any antigen including but not limited to a protein, glycoprotein, ganglioside, carbohydrate, lipid that is associated with cancer. Such antigen can be expressed on malignant cells or in the tumor microenvironment such as on tumor-associated blood vessels, extracellular matrix, mesenchymal stroma, or immune infiltrates.

[0077] As used herein, the terms “subject” and “patient” refer to an organism to be treated by the methods and compositions described herein. Such organisms preferably include, but are not limited to, mammals (*e.g.*, murines, simians, equines, bovines, porcines, canines, felines, and the like), and more preferably include humans.

[0078] As used herein, the term “effective amount” refers to the amount of a compound (*e.g.*, a compound of the present invention) sufficient to effect beneficial or desired results. An effective amount can be administered in one or more administrations, applications or

dosages and is not intended to be limited to a particular formulation or administration route. As used herein, the term “treating” includes any effect, *e.g.*, lessening, reducing, modulating, ameliorating or eliminating, that results in the improvement of the condition, disease, disorder, and the like, or ameliorating a symptom thereof.

5 [0079] As used herein, the term “pharmaceutical composition” refers to the combination of an active agent with a carrier, inert or active, making the composition especially suitable for diagnostic or therapeutic use *in vivo* or *ex vivo*.

[0080] As used herein, the term “pharmaceutically acceptable carrier” refers to any of the standard pharmaceutical carriers, such as a phosphate buffered saline solution, water,
10 emulsions (*e.g.*, such as an oil/water or water/oil emulsions), and various types of wetting agents. The compositions also can include stabilizers and preservatives. For examples of carriers, stabilizers and adjuvants, *see e.g.*, Martin, Remington's Pharmaceutical Sciences, 15th Ed., Mack Publ. Co., Easton, PA [1975].

[0081] As used herein, the term “pharmaceutically acceptable salt” refers to any
15 pharmaceutically acceptable salt (*e.g.*, acid or base) of a compound of the present invention which, upon administration to a subject, is capable of providing a compound of this invention or an active metabolite or residue thereof. As is known to those of skill in the art, “salts” of the compounds of the present invention may be derived from inorganic or organic acids and bases. Exemplary acids include, but are not limited to, hydrochloric, hydrobromic, sulfuric,
20 nitric, perchloric, fumaric, maleic, phosphoric, glycolic, lactic, salicylic, succinic, toluene-p-sulfonic, tartaric, acetic, citric, methanesulfonic, ethanesulfonic, formic, benzoic, malonic, naphthalene-2-sulfonic, benzenesulfonic acid, and the like. Other acids, such as oxalic, while not in themselves pharmaceutically acceptable, may be employed in the preparation of salts useful as intermediates in obtaining the compounds of the invention and their
25 pharmaceutically acceptable acid addition salts.

[0082] Exemplary bases include, but are not limited to, alkali metal (*e.g.*, sodium) hydroxides, alkaline earth metal (*e.g.*, magnesium) hydroxides, ammonia, and compounds of formula NW_4^+ , wherein W is C_{1-4} alkyl, and the like.

[0083] Exemplary salts include, but are not limited to: acetate, adipate, alginate,
30 aspartate, benzoate, benzenesulfonate, bisulfate, butyrate, citrate, camphorate, camphorsulfonate, cyclopentanepropionate, digluconate, dodecylsulfate, ethanesulfonate, fumarate, flucoheptanoate, glycerophosphate, hemisulfate, heptanoate, hexanoate, hydrochloride, hydrobromide, hydroiodide, 2-hydroxyethanesulfonate, lactate, maleate, methanesulfonate, 2-naphthalenesulfonate, nicotinate, oxalate, palmoate, pectinate,

persulfate, phenylpropionate, picrate, pivalate, propionate, succinate, tartrate, thiocyanate, tosylate, undecanoate, and the like. Other examples of salts include anions of the compounds of the present invention compounded with a suitable cation such as Na^+ , NH_4^+ , and NW_4^+ (wherein W is a C_{1-4} alkyl group), and the like.

5 [0084] For therapeutic use, salts of the compounds of the present invention are contemplated as being pharmaceutically acceptable. However, salts of acids and bases that are non-pharmaceutically acceptable may also find use, for example, in the preparation or purification of a pharmaceutically acceptable compound.

[0085] Throughout the description, where compositions are described as having,
10 including, or comprising specific components, or where processes and methods are described as having, including, or comprising specific steps, it is contemplated that, additionally, there are compositions of the present invention that consist essentially of, or consist of, the recited components, and that there are processes and methods according to the present invention that consist essentially of, or consist of, the recited processing steps.

15 [0086] As a general matter, compositions specifying a percentage are by weight unless otherwise specified. Further, if a variable is not accompanied by a definition, then the previous definition of the variable controls.

I. PROTEINS

[0087] The invention provides multi-specific binding proteins that bind HER2 on a
20 cancer cell and the NKG2D receptor and CD16 receptor on natural killer cells to activate the natural killer cell. The multi-specific binding proteins are useful in the pharmaceutical compositions and therapeutic methods described herein. Binding of the multi-specific binding protein to the NKG2D receptor and CD16 receptor on natural killer cell enhances the activity of the natural killer cell toward destruction of a cancer cell. Binding of the multi-specific
25 binding protein to HER2 on a cancer cell brings the cancer cell into proximity with the natural killer cell, which facilitates direct and indirect destruction of the cancer cell by the natural killer cell. Further description of exemplary multi-specific binding proteins is provided below.

[0088] The first component of the multi-specific binding proteins binds to NKG2D
30 receptor-expressing cells, which can include but are not limited to NK cells, $\gamma\delta$ T cells and $\text{CD8}^+ \alpha\beta$ T cells. Upon NKG2D binding, the multi-specific binding proteins may block natural ligands, such as ULBP6 and MICA, from binding to NKG2D and activating NKG2D receptors.

[0089] The second component of the multi-specific binding proteins binds to HER2-expressing cells, which can include but are limited to breast, ovarian, esophageal, bladder and gastric cancer, salivary duct carcinoma, adenocarcinoma of the lung and aggressive forms of uterine cancer, such as uterine serous endometrial carcinoma.

5 **[0090]** The third component for the multi-specific binding proteins binds to cells expressing CD16, an Fc receptor on the surface of leukocytes including natural killer cells, macrophages, neutrophils, eosinophils, mast cells, and follicular dendritic cells.

[0091] The multi-specific binding proteins described herein can take various formats. For example, one format is a heterodimeric, multi-specific antibody including a first
 10 immunoglobulin heavy chain, a first immunoglobulin light chain, a second immunoglobulin heavy chain and a second immunoglobulin light chain (FIG. 1). The first immunoglobulin heavy chain includes a first Fc (hinge-CH2-CH3) domain, a first heavy chain variable domain and optionally a first CH1 heavy chain domain. The first immunoglobulin light chain includes a first light chain variable domain and a first light chain constant domain. The first
 15 immunoglobulin light chain, together with the first immunoglobulin heavy chain, forms an antigen-binding site that binds NKG2D. The second immunoglobulin heavy chain comprises a second Fc (hinge-CH2-CH3) domain, a second heavy chain variable domain and optionally a second CH1 heavy chain domain. The second immunoglobulin light chain includes a second light chain variable domain and a second light chain constant domain. The second
 20 immunoglobulin light chain, together with the second immunoglobulin heavy chain, forms an antigen-binding site that binds HER2. The first Fc domain and second Fc domain together are able to bind to CD16 (FIG. 1). In some embodiments, the first immunoglobulin light chain can be identical to the second immunoglobulin light chain.

[0092] Another exemplary format involves a heterodimeric, multi-specific antibody
 25 including a first immunoglobulin heavy chain, a second immunoglobulin heavy chain and an immunoglobulin light chain (FIG. 2). The first immunoglobulin heavy chain includes a first Fc (hinge-CH2-CH3) domain fused via either a linker or an antibody hinge to a single-chain variable fragment (scFv) composed of a heavy variable domain and light chain variable domain which pair and bind NKG2D or HER2. The second immunoglobulin heavy chain
 30 includes a second Fc (hinge-CH2-CH3) domain, a second heavy chain variable domain and optionally a CH1 heavy chain domain. The immunoglobulin light chain includes a light chain variable domain and a constant light chain domain. The second immunoglobulin heavy chain pairs with the immunoglobulin light chain and binds to NKG2D or HER2. The first Fc domain and the second Fc domain together are able to bind to CD16 (FIG. 2).

[0093] One or more additional binding motifs may be fused to the C-terminus of the constant region CH3 domain, optionally via a linker sequence. In certain embodiments, the antigen-binding site could be a single-chain or disulfide-stabilized variable region (scFv) or could form a tetravalent or trivalent molecule.

5 [0094] In some embodiments, the multi-specific binding protein is in the Triomab form, which is a trifunctional, bispecific antibody that maintains an IgG-like shape. This chimera consists of two half antibodies, each with one light and one heavy chain, that originate from two parental antibodies.

[0095] In some embodiments, the multi-specific binding protein is the KiH Common
 10 Light Chain (LC) form, which involves the knobs-into-holes (KIHs) technology. The KIH involves engineering C_H3 domains to create either a “knob” or a “hole” in each heavy chain to promote heterodimerization. The concept behind the “Knobs-into-Holes (KiH)” Fc technology was to introduce a “knob” in one CH3 domain (CH3A) by substitution of a small residue with a bulky one (*e.g.*, T366W_{CH3A} in EU numbering). To accommodate the “knob,”
 15 a complementary “hole” surface was created on the other CH3 domain (CH3B) by replacing the closest neighboring residues to the knob with smaller ones (*e.g.*, T366S/L368A/Y407V_{CH3B}). The “hole” mutation was optimized by structured-guided phage library screening (Atwell S, Ridgway JB, Wells JA, Carter P., Stable heterodimers from remodeling the domain interface of a homodimer using a phage display library, *J. Mol.*
 20 *Biol.* (1997) 270(1):26–35). X-ray crystal structures of KiH Fc variants (Elliott JM, Ultsch M, Lee J, Tong R, Takeda K, Spiess C, *et al.*, Antiparallel conformation of knob and hole aglycosylated half-antibody homodimers is mediated by a CH2-CH3 hydrophobic interaction. *J. Mol. Biol.* (2014) 426(9):1947–57; Mimoto F, Kadono S, Katada H, Igawa T, Kamikawa T, Hattori K. Crystal structure of a novel asymmetrically engineered Fc variant
 25 with improved affinity for Fcγ₁R. *Mol. Immunol.* (2014) 58(1):132–8) demonstrated that heterodimerization is thermodynamically favored by hydrophobic interactions driven by steric complementarity at the inter-CH3 domain core interface, whereas the knob–knob and the hole–hole interfaces do not favor homodimerization owing to steric hindrance and disruption of the favorable interactions, respectively.

30 [0096] In some embodiments, the multi-specific binding protein is in the dual-variable domain immunoglobulin (DVD-IgTM) form, which combines the target binding domains of two monoclonal antibodies via flexible naturally occurring linkers, and yields a tetravalent IgG-like molecule.

[0097] In some embodiments, the multi-specific binding protein is in the Orthogonal Fab interface (Ortho-Fab) form. In the ortho-Fab IgG approach (Lewis SM, Wu X, Pustilnik A, Sereno A, Huang F, Rick HL, *et al.*, Generation of bispecific IgG antibodies by structure-based design of an orthogonal Fab interface. *Nat. Biotechnol.* (2014) 32(2):191–8), structure-based regional design introduces complementary mutations at the LC and HC_{VH-CH1} interface in only one Fab, without any changes being made to the other Fab.

[0098] In some embodiments, the multi-specific binding protein is in the 2-in-1 Ig format. In some embodiments, the multi-specific binding protein is in the ES form, which is a heterodimeric construct containing two different Fabs binding to targets 1 and target 2 fused to the Fc. Heterodimerization is ensured by electrostatic steering mutations in the Fc. In some embodiments, the multi-specific binding protein is in the κλ-Body form, which is an heterodimeric constructs with two different Fabs fused to Fc stabilized by heterodimerization mutations: Fab1 targeting antigen 1 contains kappa LC, while second Fab targeting antigen 2 contains lambda LC. FIG. 49A is an exemplary representation of one form of a κλ-Body; FIG. 49B is an exemplary representation of another κλ-Body.

[0099] In some embodiments, the multi-specific binding protein is in Fab Arm Exchange form (antibodies that exchange Fab arms by swapping a heavy chain and attached light chain (half-molecule) with a heavy-light chain pair from another molecule, which results in bispecific antibodies). In some embodiments, the multi-specific binding protein is in the SEED Body form. The strand-exchange engineered domain (SEED) platform was designed to generate asymmetric and bispecific antibody-like molecules, a capability that expands therapeutic applications of natural antibodies. This protein engineered platform is based on exchanging structurally related sequences of immunoglobulin within the conserved CH3 domains. The SEED design allows efficient generation of AG/GA heterodimers, while disfavoring homodimerization of AG and GA SEED CH3 domains. (Muda M. *et al.*, *Protein Eng. Des. Sel.* (2011, 24(5):447-54)). In some embodiments, the multi-specific binding protein is in the LuZ-Y form, in which a leucine zipper is used to induce heterodimerization of two different HCs. (Wranik, BJ. *et al.*, *J. Biol. Chem.* (2012), 287:43331-9).

[00100] In some embodiments, the multi-specific binding protein is in the Cov-X-Body form. In bispecific CovX-Bodies, two different peptides are joined together using a branched azetidinone linker and fused to the scaffold antibody under mild conditions in a site-specific manner. Whereas the pharmacophores are responsible for functional activities, the antibody scaffold imparts long half-life and Ig-like distribution. The pharmacophores can be

chemically optimized or replaced with other pharmacophores to generate optimized or unique bispecific antibodies. (Doppalapudi VR *et al.*, *PNAS* (2010), 107(52);22611-22616).

[00101] In some embodiments, the multi-specific binding protein is in an Oasc-Fab heterodimeric form that includes Fab binding to target 1, and scFab binding to target 2 fused to Fc. Heterodimerization is ensured by mutations in the Fc.

[00102] In some embodiments, the multi-specific binding protein is in a DuetMab form, which is an heterodimeric construct containing two different Fabs binding to antigens 1 and 2, and Fc stabilized by heterodimerization mutations. Fab 1 and 2 contain differential S-S bridges that ensure correct LC and HC pairing.

[00103] In some embodiments, the multi-specific binding protein is in a CrossmAb form, which is an heterodimeric construct with two different Fabs binding to targets 1 and 2, fused to Fc stabilized by heterodimerization. CL and CH1 domains and VH and VL domains are switched, *e.g.*, CH1 is fused in-line with VL, while CL is fused in-line with VH.

[00104] In some embodiments, the multi-specific binding protein is in a Fit-Ig form, which is an homodimeric constructs where Fab binding to antigen 2 is fused to the N terminus of HC of Fab that binds to antigen 1. The construct contains wild-type Fc.

[00105] Additional formats of the multi-specific binding proteins can be devised by combining various formats of NKG2D- and HER2-binding fragments described herein.

[00106] Table 1 lists peptide sequences of heavy chain variable domains and light chain variable domains that, in combination, can bind to NKG2D.

Clones	Heavy chain variable region amino acid sequence	Light chain variable region amino acid sequence
ADI-27705	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKS RVTVISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:1) CDR1 (SEQ ID NO:62) – GSFSGYYWS CDR2 (SEQ ID NO:63) – EIDHSGSTNYNPSLKS	DIQMTQSPSTLSASVGDRVTITCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESQVPSRFSGSGSGTEFT LTISLQPDDEFATYYCQQYNSYP TFGGGTKVEIK (SEQ ID NO:2)

	CDR3 (SEQ ID NO:64) – ARARGPWSFDP	
ADI-27724	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:3)	EIVLTQSPGTLSSLSPGERATLSCRA SQSVSSSYLAWYQQKPGQAPRL IYGASSRATGIPDRFSGSGSGTDFT LTISRLEPEDFAVYYCQQYGSSPIT FGGGTKVEIK (SEQ ID NO:4)
ADI-27740 (A40)	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:5)	DIQMTQSPSTLSASVGDRVITTCR ASQSIGSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYHSFYT FGGGTKVEIK (SEQ ID NO:6)
ADI-27741	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:7)	DIQMTQSPSTLSASVGDRVITTCR ASQSIGSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQSNSYYT FGGGTKVEIK (SEQ ID NO:8)
ADI-27743	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:9)	DIQMTQSPSTLSASVGDRVITTCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYNSYPT FGGGTKVEIK (SEQ ID NO:10)
ADI-28153	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW GFDPWGQGTLVTVSS (SEQ ID NO:11)	ELQMTQSPSSLSASVGDRVITTCR TSQSISSYLNWYQQKPGQPPKLLI YWASTRESGVDPDRFSGSGSGTDF TLTISSLQPEDSATYYCQQSYDIP YTFGQGTKLEIK (SEQ ID NO:12)

ADI-28226 (C26)	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:13)	DIQMTQSPSTLSASVGDRVITICR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYGSFPIT FGGGTKVEIK (SEQ ID NO:14)
ADI-28154	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:15)	DIQMTQSPSTLSASVGDRVITICR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTDFT LTISSLQPDDFATYYCQQSKEVP WTFGQGTKVEIK (SEQ ID NO:16)
ADI-29399	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:17)	DIQMTQSPSTLSASVGDRVITICR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYNSFPT FGGGTKVEIK (SEQ ID NO:18)
ADI-29401	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:19)	DIQMTQSPSTLSASVGDRVITICR ASQSIGSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYDIYPT FGGGTKVEIK (SEQ ID NO:20)
ADI-29403	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:21)	DIQMTQSPSTLSASVGDRVITICR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYDSYPT FGGGTKVEIK (SEQ ID NO:22)
ADI-29405	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ	DIQMTQSPSTLSASVGDRVITICR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT

	FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:23)	LTISLQPDDEFATYYCQQYGSFPT FGGGTKVEIK (SEQ ID NO:24)
ADI-29407	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:25)	DIQMTQSPSTLSASVGDRVTITCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISLQPDDEFATYYCQQYQSFP FGGGTKVEIK (SEQ ID NO:26)
ADI-29419	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:27)	DIQMTQSPSTLSASVGDRVTITCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISLQPDDEFATYYCQQYSSFS FGGGTKVEIK (SEQ ID NO:28)
ADI-29421	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:29)	DIQMTQSPSTLSASVGDRVTITCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISLQPDDEFATYYCQQYESYST FGGGTKVEIK (SEQ ID NO:30)
ADI-29424	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:31)	DIQMTQSPSTLSASVGDRVTITCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISLQPDDEFATYYCQQYDSFITF GGGKVEIK (SEQ ID NO:32)
ADI-29425	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRVTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:33)	DIQMTQSPSTLSASVGDRVTITCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISLQPDDEFATYYCQQYQSYPT FGGGTKVEIK (SEQ ID NO:34)

ADI-29426	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:35)	DIQMTQSPSTLSASVGDRVITCR ASQSIGSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYHSFPT FGGGTKVEIK (SEQ ID NO:36)
ADI-29429	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:37)	DIQMTQSPSTLSASVGDRVITCR ASQSIGSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYELYSY TFGGGTKVEIK (SEQ ID NO:38)
ADI-29447 (F47)	QVQLQQWGAGLLKPSETLSLTCAVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:39)	DIQMTQSPSTLSASVGDRVITCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESGVPSRFSGSGSGTEFT LTISSLQPDDFATYYCQQYDTFIT FGGGTKVEIK (SEQ ID NO:40)
ADI-27727	QVQLVQSGAEVKKPGSSVKVSCKAS GGTFSSYAISWVRQAPGQGLEWMGG IIPFGTANYAQKFQGRVTITADESTS TAYMELSSLRSEDVAVYYCARGDSSI RHAYYYYGMDVWGQGTITVTVSS (SEQ ID NO:41) CDR1 (SEQ ID NO:65) – GTFSSYAIS CDR2 (SEQ ID NO:66) – GIIPFGTANYAQKFQG CDR3 (SEQ ID NO:67) – ARGDSSIRHAYYYYGMDV	DIVMTQSPDSLAVSLGERATINCK SSQSVLYSSNNKNYLAWYQQKP GQPPKLLIWASTRESGVPRFSG SGSGTDFTLTISLQAEDVAVYYC QQYYSTPITFGGGTKVEIK (SEQ ID NO:42) CDR1 (SEQ ID NO:68) – KSSQSVLYSSNNKNYLA CDR2 (SEQ ID NO:69) – WASTRES CDR3 (SEQ ID NO:70) – QQYYSTPIT
ADI-29443 (F43)	QLQLQESGPGLVKPSETLSLTCTVSG GSISSSSYWGWIRQPPGKGLEWIGSI YYSGSTYYNPSLKSRTISVDTSKNQ	EIVLTQSPATLSLSPGERATLSCRA SQSVSRYLAWYQQKPGQAPRLLI YDASNRAITGIPARFSGSGSGTDFT

	FSLKLSSVTAADTAVYYCARGSDRF HPYFDYWGQGTLVTVSS (SEQ ID NO:43) CDR1 (SEQ ID NO:71) – GSISSSSYWYG CDR2 (SEQ ID NO:72) – SIYYSGSTYYNPSLKS CDR3 (SEQ ID NO:73) – ARGSDRFHPYFDY	LTISSLEPEDFAVYYCQQFDTWPP TFGGGTKVEIK (SEQ ID NO:44) CDR1 (SEQ ID NO:74) – RASQSVSRYLA CDR2 (SEQ ID NO:75) – DASNRAT CDR3 (SEQ ID NO:76) – QQFDTWPPT
ADI-29404 (F04)	QVQLQQWGAGLLKPSETLSLTCVY GGSFSGYYWSWIRQPPGKGLEWIGEI DHSGSTNYNPSLKSRTISVDTSKNQ FSLKLSSVTAADTAVYYCARARGPW SFDPWGQGTLVTVSS (SEQ ID NO:89)	DIQMTQSPSTLSASVGDRVTITCR ASQSISSWLAWYQQKPGKAPKLL IYKASSLESQVPSRFSGSGSGTEFT LTISSLQPDDFATYYCEQYDSYPT FGGGGTKVEIK (SEQ ID NO:90)
ADI-28200	QVQLVQSGAEVKKPGSSVKVSCKAS GGTFSSYAISWVRQAPGQGLEWMGG IIPFGTANYAQKFQGRVTITADESTS TAYMELSSLRSEDVAVYYCARRGRK ASGSFYFYYGMDVWGQGTTVTVSS (SEQ ID NO:91)	DIVMTQSPDSLAVSLGERATINCE SSQSLNLSGNQKNYLTWYQQKP GQPPKPLIYWASTRESQVPDRFSG SGSGTDFTLTISLQAEDVAVYYC QNDYSYPYTFGQGTKLEIK (SEQ ID NO:92)
ADI-29379 (E79)	QVQLVQSGAEVKKPGASVKVSCKAS GYTFTSYMHVVRQAPGQGLEWM GIINPSGGSTSYAQKFQGRVTMTRDT STSTVYMELSSLRSEDVAVYYCARG APNYGDTTHDYYYMDVWGKGTTVT VSS (SEQ ID NO:94) CDR1 (SEQ ID NO:96) - YTFTSYMH CDR2 (SEQ ID NO:97) - IINPSGGSTSYAQKFQG CDR3 (SEQ ID NO:98) - ARGAPNYGDTTHDYYYMDV	EIVMTQSPATLSVSPGERATLSCR ASQSVSSNLAWYQQKPGQAPRLL IYGASTRATGIPARFSGSGSGTEFT LTISSLQSEDFAVYYCQQYDDWP FTFGGGGTKVEIK (SEQ ID NO:95) CDR1 (SEQ ID NO:99) - RASQSVSSNLA CDR2 (SEQ ID NO:100) - GASTRAT CDR3 (SEQ ID NO:101) - QQYDDWPFT

ADI-27749 (A49)	EVQLVESGGGLVKPGGSLRLSCAAS GFTFSSYSMNWVRQAPGKGLEWVSS ISSSSYIYYADSVKGRFTISRDNANKN SLYLQMNSLRAEDTAVYYCARGAP MGAAAGWFDPWGQGTLVTVSS (SEQ ID NO:102) CDR1 (SEQ ID NO:104) - FTFSSYSMN CDR2 (SEQ ID NO:105) - SISSSSSYIYYADSVKGR CDR3 (SEQ ID NO:106) - ARGAPMGAAAGWFDP	DIQMTQSPSSVSASVGDRVTITCR ASQGISSWLAWYQQKPGKAPKLL IYAASSLQSGVPSRFSGSGSGTDF TLTISSLQPEDFATYYCQQGVSP RTFGGGTKVEIK (SEQ ID NO:103) CDR1 (SEQ ID NO:107) - RASQGISSWLA CDR2 (SEQ ID NO:108) - AASSLQS CDR3 (SEQ ID NO:109) - QQGVSPFRT
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[00107] Alternatively, a heavy chain variable domain defined by SEQ ID NO:45 can be paired with a light chain variable domain defined by SEQ ID NO:46 to form an antigen-binding site that can bind to NKG2D, as illustrated in US 9,273,136.

5 QVQLVESGGGLVKPGGSLRLSCAASGFTFSSYGMHWVRQAPGKGLEWVAFIRYDGS
NKYYADSVKGRFTISRDNANKNTLYLQMNSLRAEDTAVYYCAKDRGLGDGTYFDYW
GQGTTVTVSS (SEQ ID NO:45)

QSALTQPASVSGSPGQSITISCSGSSSNIGNNAVNWYQQLPGKAPKLLIYYDDLPSG
VSDRFSGSKSGTSAFLAISGLQSEDEADYYCAAWDDSLNGPVFGGGTKLTVL (SEQ
10 ID NO:46)

[00108] Alternatively, a heavy chain variable domain defined by SEQ ID NO:47 can be paired with a light chain variable domain defined by SEQ ID NO:48 to form an antigen-binding site that can bind to NKG2D, as illustrated in US 7,879,985.

15 QVHLQESGPGLVKPSETLSLTCTVSDDSISSYYWSWIRQPPGKGLEWIGHISYSGSAN
YNPSLKSRTISVDTSKNQFSLKLSSVTAADTAVYYCANWDDAFNIWGQGTMTVTS
S (SEQ ID NO:47)

EIVLTQSPGTLSPGERATLSCRASQSVSSSYLAWYQQKPGQAPRLLIYGASSRATGI
PDRFSGSGSGTDFTLTISRLEPEDFAVYYCQQYGSSPWTFGGGTKVEIK (SEQ ID
20 NO:48)

[00109] Table 2 lists peptide sequences of heavy chain variable domains and light chain variable domains that, in combination, can bind to HER2.

Table 2		
Clones	Heavy chain variable domain amino acid sequence	Light chain variable domain amino acid sequence
Trastuzumab	EVQLVESGGGLVQPGGSLRLSCAA SGFNIKDTYIHWVRQAPGKGLEWV ARIYPTNGYTRYADSVKGRFTISAD TSKNTAYLQMNSLRAEDTAVYYCS RWGGDGFYAMDYWGQGTLVTVS S (SEQ ID NO:49) CDR1(SEQ ID NO:50) - GFNIKDT CDR2 (SEQ ID NO:51) - YPTNGY CDR3 (SEQ ID NO:52) - WGGDGFYAMDY	DIQMTQSPSSLSASVGDRVTITCRA SQDVNTAVAWYQQKPGKAPKLLI YSASFLYSGVPSRFSGSRSGTDFTL TISSLQPEDFATYYCQQHYTTPPTF GQGTKVEIK (SEQ ID NO:53) CDR1(SEQ ID NO:54) - QDVNTAVA CDR2 (SEQ ID NO:55) - SASFLYS CDR3 (SEQ ID NO:56) - QQHYTTPPT
Pertuzumab	EVQLVESGGGLVQPGGSLRLSCAA SGFTFTDYTMDWVRQAPGKGLEW VADVNPNSGGSIYNQRFKGRFTLS VDRSKNTLYLQMNSLRAEDTAVY YCARNLGPSFYFDYWGQGTLVTVS SA (SEQ ID NO:57) CDR1 (SEQ ID NO:77) - GFTFTDY CDR2 (SEQ ID NO:78) - NPNSGG CDR3 (SEQ ID NO:79) - NLGPSFYFDY	DIQMTQSPSSLSASVGDRVTITCKA SQDVSIGVAWYQQKPGKAPKLLIY SASYRYTGVPSRFSGSGSGTDFTLT ISSLQPEDFATYYCQQYYIYPYTFG QGTKVEIKR (SEQ ID NO:58) CDR1 (SEQ ID NO:80) - QDVSIGVA CDR2 (SEQ ID NO:81) - SASYRYT CDR3 (SEQ ID NO:82) - QQYYIYPYT
MGAH22 (US 8,802,093)	QVQLQQSGPELVKPGASLKLSCA SGFNIKDTYIHWVKQRPEQGLEWI GRIYPTNGYTRYDPKFQDKATITAD TSSNTAYLQVSRLTSEDVAVYYCS	DIVMTQSHKFMSTSVGDRVSITCK ASQDVNTAVAWYQQKPGHSPKLL IYSASFRYTGVDPDRFTGSRSGTDFT FTISSVQAEDLAVYYCQQHYTTPP

	RWGGDGFYAMDYWGQGASVTVS SA (SEQ ID NO:59) CDR1 (SEQ ID NO:83) - GFNIKDT CDR2 (SEQ ID NO:84) - YPTNGY CDR3 (SEQ ID NO:85) - WGGDGFYAMDY	TFGGGGTKVEIKR (SEQ ID NO:60) CDR1 (SEQ ID NO:86) - QDVNTAVA CDR2 (SEQ ID NO:87) - SASFRYT CDR3 (SEQ ID NO:88) - QQHYTTPPT
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[00110] Alternatively, novel antigen-binding sites that can bind to HER2 can be identified by screening for binding to the amino acid sequence defined by SEQ ID NO:61.

5 MELAALCRWGLLLALLPPGAASTQVCTGTDMLRLPASPETHDMLRHLYQGCQV
VQGNLELTYLPTNASLSFLQDIQEVQGYVLIAHNQVRQVPLQRLRIVRGTQLFEDNY
ALAVLDNGDPLNNTTPVTGASPGGLRELQLRSLTEILKGGVLIQRNPQLCYQDTILW
KDIFHKNNQLALTIDTNRSRACHPCSPMCKGSRGWGESSEDCQSLTRTVCAAGGCAR
CKGPLPTDCCHEQCAAGCTGPKHSDCLACLFHNHSGICELHCPALVTYNTDTFESMP
NPEGRYTFGASCVTACPYNYLSTDVGSTLVCPLHNQEVTAEEDGTQRCEKCSKPCA
10 RVCYGLGMEHLREVRAVTSANIQEFAGCKKIFGSLAFLPESFDGDPASNTAPLQPEQL
QVFETLEEITGYLYISAWPDSLPLDSVFQNLQVIRGRILHNGAYSLLTLQGLGISWLGLR
SLRELGSGLALIHHTHLCFVHTVPWDQLFRNPHQALLHTANRPEDECVGEGLACH
QLCARGHCWGPGPTQCVNCSQFLRGQECVEECRVLQGLPREYVNARHCLPCHPECQ
PQNGSVTCFGPEADQCVACAHYKDPPFCVARCPSGVKPDLSPYMPIWKFPDEEGACQ
15 PCPINCTHSCVDLDDKGCPAEQRASPLTSIISAVVGILLVVVLGVVFGILIKRRQKIR
KYTMRRLLQETELVEPLTPSGAMPNQAQMRILKETELRKVKVLGSGAFGTVYKGIWI
PDGENVKIPVAIKVLRENTSPKANKEILDEAYVMAGVGSPPYVSRLLGICLTSTVQLVT
QLMPYGCLLDHVRENRRGLGSQDLLNWCMIQAKGMSYLEDVRLVHRDLAARNVL
VKSPNHVKITDFGLARLLDIDETEHADGGKVPIKWMALLESILRRRFTHQSDVWSYG
20 VTVWELMTFGAKPYDGIPAREIPDLLEKGERLPQPPICTIDVYMIMVKCWMIDSECRP
RFRELVSEFSRMARDPQRFVVIQNEDLGPASPLDSTFYRSLLEDDDMGDLVDAEEYL
VPQQGFFCPDPAPGAGGMVHHRHRSSSTRSGGGDLTLGLEPSEEEAPRSPLAPSEGA
GSDVFDGDLGMGAAKGLQSLPHTDPSPLQRYSEDPTVPLPSETDGYVAPLTCSPQPE
YVNQPDVRPQPPSPREGPLPAARPAGATLERPKTLSPGKNGVVKDVFAFGGAVENPE
25 YLTPQGGGAAPQPHPPPAFSPAFDNLYYWDQDPPERGAPPSTFKGTPTAENPEYLGLD
VPV (SEQ ID NO:61).

[00111] Within the Fc domain, CD16 binding is mediated by the hinge region and the CH2 domain. For example, within human IgG1, the interaction with CD16 is primarily focused on
30 amino acid residues Asp 265 – Glu 269, Asn 297 – Thr 299, Ala 327 – Ile 332, Leu 234 – Ser 239, and carbohydrate residue N-acetyl-D-glucosamine in the CH2 domain (*see*,
Sondermann *et al*, Nature, 406 (6793):267-273). Based on the known domains, mutations can be selected to enhance or reduce the binding affinity to CD16, such as by using phage-

displayed libraries or yeast surface-displayed cDNA libraries, or can be designed based on the known three-dimensional structure of the interaction.

[00112] The assembly of heterodimeric antibody heavy chains can be accomplished by expressing two different antibody heavy chain sequences in the same cell, which may lead to the assembly of homodimers of each antibody heavy chain as well as assembly of heterodimers. Promoting the preferential assembly of heterodimers can be accomplished by incorporating different mutations in the CH3 domain of each antibody heavy chain constant region as shown in US13/494870, US16/028850, US11/533709, US12/875015, US13/289934, US14/773418, US12/811207, US13/866756, US14/647480, and US14/830336. For example, mutations can be made in the CH3 domain based on human IgG1 and incorporating distinct pairs of amino acid substitutions within a first polypeptide and a second polypeptide that allow these two chains to selectively heterodimerize with each other. The positions of amino acid substitutions illustrated below are all numbered according to the EU index as in Kabat.

[00113] In one scenario, an amino acid substitution in the first polypeptide replaces the original amino acid with a larger amino acid, selected from arginine (R), phenylalanine (F), tyrosine (Y) or tryptophan (W), and at least one amino acid substitution in the second polypeptide replaces the original amino acid(s) with a smaller amino acid(s), chosen from alanine (A), serine (S), threonine (T), or valine (V), such that the larger amino acid substitution (a protuberance) fits into the surface of the smaller amino acid substitutions (a cavity). For example, one polypeptide can incorporate a T366W substitution, and the other can incorporate three substitutions including T366S, L368A, and Y407V.

[00114] An antibody heavy chain variable domain of the invention can optionally be coupled to an amino acid sequence at least 90% identical to an antibody constant region, such as an IgG constant region including hinge, CH2 and CH3 domains with or without CH1 domain. In some embodiments, the amino acid sequence of the constant region is at least 90% identical to a human antibody constant region, such as a human IgG1 constant region, an IgG2 constant region, IgG3 constant region, or IgG4 constant region. In some other embodiments, the amino acid sequence of the constant region is at least 90% identical to an antibody constant region from another mammal, such as rabbit, dog, cat, mouse, or horse. One or more mutations can be incorporated into the constant region as compared to human IgG1 constant region, for example at Q347, Y349, L351, S354, E356, E357, K360, Q362, S364, T366, L368, K370, N390, K392, T394, D399, S400, D401, F405, Y407, K409, T411 and/or K439. Exemplary substitutions include, for example, Q347E, Q347R, Y349S,

Y349K, Y349T, Y349D, Y349E, Y349C, T350V, L351K, L351D, L351Y, S354C, E356K, E357Q, E357L, E357W, K360E, K360W, Q362E, S364K, S364E, S364H, S364D, T366V, T366I, T366L, T366M, T366K, T366W, T366S, L368E, L368A, L368D, K370S, N390D, N390E, K392L, K392M, K392V, K392F, K392D, K392E, T394F, T394W, D399R, D399K,
 5 D399V, S400K, S400R, D401K, F405A, F405T, Y407A, Y407I, Y407V, K409F, K409W, K409D, T411D, T411E, K439D, and K439E.

[00115] In certain embodiments, mutations that can be incorporated into the CH1 of a human IgG1 constant region may be at amino acid V125, F126, P127, T135, T139, A140, F170, P171, and/or V173. In certain embodiments, mutations that can be incorporated into
 10 the C κ of a human IgG1 constant region may be at amino acid E123, F116, S176, V163, S174, and/or T164.

[00116] Amino acid substitutions could be selected from the following sets of substitutions shown in Table 3.

Table 3		
	First Polypeptide	Second Polypeptide
Set 1	S364E/F405A	Y349K/T394F
Set 2	S364H/D401K	Y349T/T411E
Set 3	S364H/T394F	Y349T/F405A
Set 4	S364E/T394F	Y349K/F405A
Set 5	S364E/T411E	Y349K/D401K
Set 6	S364D/T394F	Y349K/F405A
Set 7	S364H/F405A	Y349T/T394F
Set 8	S364K/E357Q	L368D/K370S
Set 9	L368D/K370S	S364K
Set 10	L368E/K370S	S364K
Set 11	K360E/Q362E	D401K
Set 12	L368D/K370S	S364K/E357L
Set 13	K370S	S364K/E357Q
Set 14	F405L	K409R
Set 15	K409R	F405L

15 **[00117]** Alternatively, amino acid substitutions could be selected from the following sets of substitutions shown in Table 4.

Table 4		
	First Polypeptide	Second Polypeptide
Set 1	K409W	D399V/F405T
Set 2	Y349S	E357W
Set 3	K360E	Q347R
Set 4	K360E/K409W	Q347R/D399V/F405T
Set 5	Q347E/K360E/K409W	Q347R/D399V/F405T
Set 6	Y349S/K409W	E357W/D399V/F405T

[00118] Alternatively, amino acid substitutions could be selected from the following set of substitutions shown in Table 5.

Table 5		
	First Polypeptide	Second Polypeptide
Set 1	T366K/L351K	L351D/L368E
Set 2	T366K/L351K	L351D/Y349E
Set 3	T366K/L351K	L351D/Y349D
Set 4	T366K/L351K	L351D/Y349E/L368E
Set 5	T366K/L351K	L351D/Y349D/L368E
Set 6	E356K/D399K	K392D/K409D

[00119] Alternatively, at least one amino acid substitution in each polypeptide chain could
5 be selected from Table 6.

Table 6	
First Polypeptide	Second Polypeptide
L351Y, D399R, D399K, S400K, S400R, Y407A, Y407I, Y407V	T366V, T366I, T366L, T366M, N390D, N390E, K392L, K392M, K392V, K392F K392D, K392E, K409F, K409W, T411D and T411E

[00120] Alternatively, at least one amino acid substitutions could be selected from the following set of substitutions in Table 7, where the position(s) indicated in the First

Polypeptide column is replaced by any known negatively-charged amino acid, and the position(s) indicated in the Second Polypeptide Column is replaced by any known positively-charged amino acid.

Table 7	
First Polypeptide	Second Polypeptide
K392, K370, K409, or K439	D399, E356, or E357

5 **[00121]** Alternatively, at least one amino acid substitutions could be selected from the following set of in Table 8, where the position(s) indicated in the First Polypeptide column is replaced by any known positively-charged amino acid, and the position(s) indicated in the Second Polypeptide Column is replaced by any known negatively-charged amino acid.

Table 8	
First Polypeptide	Second Polypeptide
D399, E356, or E357	K409, K439, K370, or K392

[00122] Alternatively, amino acid substitutions could be selected from the following set in Table 9.

Table 9	
First Polypeptide	Second Polypeptide
T350V, L351Y, F405A, and Y407V	T350V, T366L, K392L, and T394W

10 **[00123]** Alternatively, or in addition, the structural stability of a heteromultimer protein may be increased by introducing S354C on either of the first or second polypeptide chain, and Y349C on the opposing polypeptide chain, which forms an artificial disulfide bridge within the interface of the two polypeptides.

15 **[00124]** The multi-specific proteins described above can be made using recombinant DNA technology well known to a skilled person in the art. For example, a first nucleic acid sequence encoding the first immunoglobulin heavy chain can be cloned into a first expression vector; a second nucleic acid sequence encoding the second immunoglobulin heavy chain can be cloned into a second expression vector; a third nucleic acid sequence encoding the immunoglobulin light chain can be cloned into a third expression vector; the first, second,

and third expression vectors can be stably transfected together into host cells to produce the multimeric proteins.

[00125] To achieve the highest yield of the multi-specific protein, different ratios of the first, second, and third expression vector can be explored to determine the optimal ratio for transfection into the host cells. After transfection, single clones can be isolated for cell bank generation using methods known in the art, such as limited dilution, ELISA, FACS, microscopy, or Clonepix.

[00126] Clones can be cultured under conditions suitable for bio-reactor scale-up and maintained expression of the multi-specific protein. The multi-specific proteins can be isolated and purified using methods known in the art including centrifugation, depth filtration, cell lysis, homogenization, freeze-thawing, affinity purification, gel filtration, ion exchange chromatography, hydrophobic interaction exchange chromatography, and mixed-mode chromatography.

II. Characteristics of the multi-specific proteins

[00127] In certain embodiments, the multi-specific binding proteins described herein, which include an NKG2D-binding domain and a HER2-binding domain, bind to cells expressing human NKG2D. In certain embodiments, the multi-specific binding proteins which include an NKG2D-binding domain and a HER2-binding domain, bind to HER2 at a comparable level to that of a monoclonal antibody having the same HER2-binding domain. For example, the multi-specific binding proteins that include an NKG2D-binding domain and a HER2-binding domain from Trastuzumab can bind to HER2 expressed on cells at a level comparable to that of Trastuzumab.

[00128] However, the multi-specific binding proteins described herein are more effective in reducing tumor growth and killing cancer cells. For example, a multi-specific binding protein of the present disclosure that targets HER2-expressing tumor/cancer cells is more effective than SC2.2 — a single chain bispecific molecule built from an scFv derived from trastuzumab linked to ULBP-6, a ligand for NKG2D. SC2.2 binds HER2+ cancer cells and NKG2D+ NK cells simultaneously. Therefore, effectiveness of SC2.2 in reducing HER2+ cancer cell number was investigated. *In vitro* activation and cytotoxicity assays demonstrated that SC2.2 was effective in activating and killing NK cells. However, SC2.2 failed to demonstrate efficacy in the RMA/S-HER2 subcutaneous tumor model. The efficacy of SC2.2 was also tested *in vivo* using an RMA/S-HER2 overexpressing syngeneic mouse model (FIG. 36). In this mouse model, SC2.2 failed to demonstrate control of tumor growth compared to

vehicle control (FIG. 37). Thus, although SC2.2 was able to activate and kill NK cells, and binds to HER2+ cancer cells, these properties were insufficient to effectively control HER2+ tumor growth.

[00129] In certain embodiments, the multi-specific binding proteins described herein, which include an NKG2D-binding domain and a binding domain for tumor associated antigen, activate primary human NK cells when culturing with tumor cells expressing the antigen. NK cell activation is marked by the increase in CD107a degranulation and IFN γ cytokine production. Furthermore, compared to a monoclonal antibody that includes the tumor associated antigen-binding domain, the multi-specific binding proteins show superior activation of human NK cells in the presence of tumor cells expressing the antigen. For example, compared to the monoclonal antibody trastuzumab, the multi-specific binding proteins of the present disclosure having a HER2-binding domain, have a superior activation of human NK cells in the presence of HER2-expressing cancer cells.

[00130] In certain embodiments, the multi-specific binding proteins described herein, which include an NKG2D-binding domain and a binding domain for a tumor associated antigen, enhance the activity of rested and IL-2-activated human NK cells in the presence of tumor cells expressing the antigen. Rested NK cells showed less background IFN γ production and CD107a degranulation than IL-2-activated NK cells. In certain embodiments, rested NK cells show a greater change in IFN γ production and CD107a degranulation compared to IL-2-activated NK cells. In certain embodiments, IL-2-activated NK cells show a greater percentage of cells becoming IFN γ +; CD107a+ after stimulation with TriNKETs.

[00131] In certain embodiments, the multi-specific binding proteins described herein, which include an NKG2D-binding domain and a binding domain for a tumor associated antigen (non-limiting examples of tumor associated antigens including CD20, BCMA, and HER2), enhance the cytotoxic activity of rested and IL-2-activated human NK cells in the presence of tumor cells expressing the antigen. Furthermore, the multi-specific binding proteins (*e.g.*, A40-multi-specific binding protein, A49-multi-specific binding protein, C26-multi-specific binding protein, F04-multi-specific binding protein, F43-multi-specific binding protein, F47-multi-specific binding protein, and E79-multi-specific binding protein), which include a binding domain for HER2, more potently direct, activated and rested NK cell responses against the tumor cells, compared to a monoclonal antibody that includes HER2-binding site. In certain embodiments, the multi-specific binding proteins offer advantage against tumor cells expressing medium and low HER2, compared to monoclonal antibodies

that HER2-binding site. Therefore, a therapy including multi-specific binding proteins can be superior to a monoclonal antibody therapy.

[00132] In certain embodiments, compared to monoclonal antibodies, the multi-specific binding proteins described herein (*e.g.*, A40-multi-specific binding protein, A49-multi-specific binding protein, C26-multi-specific binding protein, F04-multi-specific binding protein, F43-multi-specific binding protein, F47-multi-specific binding protein, and E79-multi-specific binding protein), which include a binding domain for HER2 are advantageous in treating cancers with high expression of Fc receptor (FcR), or cancers residing in a tumor microenvironment with high levels of FcR. Monoclonal antibodies exert their effects on tumor growth through multiple mechanisms including ADCC, CDC, phagocytosis, and signal blockade amongst others. Amongst FcγRs, CD16 has the lowest affinity for IgG Fc; FcγRI (CD64) is the high-affinity FcR, which binds about 1000 times more strongly to IgG Fc than CD16. CD64 is normally expressed on many hematopoietic lineages such as the myeloid lineage, and can be expressed on tumors derived from these cell types, such as acute myeloid leukemia (AML). Immune cells infiltrating into the tumor, such as MDSCs and monocytes, also express CD64 and are known to infiltrate the tumor microenvironment. Expression of CD64 by the tumor or in the tumor microenvironment can have a detrimental effect on monoclonal antibody therapy. Expression of CD64 in the tumor microenvironment makes it difficult for these antibodies to engage CD16 on the surface of NK cells, as the antibodies prefer to bind the high-affinity receptor. The multi-specific binding proteins, through targeting two activating receptors on the surface of NK cells, can overcome the detrimental effect of CD64 expression (either on tumor or tumor microenvironment) on monoclonal antibody therapy. Regardless of CD64 expression on the tumor cells, the multi-specific binding proteins are able to mediate human NK cell responses against all tumor cells, because dual targeting of two activating receptors on NK cells provides stronger specific binding to NK cells.

[00133] In some embodiments, the multi-specific binding proteins described herein (*e.g.*, A40-multi-specific binding protein, A49-multi-specific binding protein, C26-multi-specific binding protein, F04-multi-specific binding protein, F43-multi-specific binding protein, F47-multi-specific binding protein, and E79-multi-specific binding protein), which include a binding domain for HER2 provide a better safety profile through reduced on-target off-tumor side effects. Natural killer cells and CD8 T cells are both able to directly lyse tumor cells, although the mechanisms through which NK cells and CD8 T cell recognize normal self from tumor cells differ. The activity of NK cells is regulated by the balance of signals from

activating (NCRs, NKG2D, CD16, *etc.*) and inhibitory (KIRs, NKG2A, *etc.*) receptors. The balance of these activating and inhibitory signals allow NK cells to determine healthy self-cells from stressed, virally infected, or transformed self-cells. This “built-in” mechanism of self-tolerance will help protect normal healthy tissue from NK cell responses. To extend this principle, the self-tolerance of NK cells will allow the multi-specific binding proteins to target antigens expressed both on self and tumor without off tumor side effects, or with an increased therapeutic window. Unlike natural killer cells, T cells require recognition of a specific peptide presented by MHC molecules for activation and effector functions. T cells have been the primary target of immunotherapy, and many strategies have been developed to redirect T cell responses against the tumor. T cell bispecifics, checkpoint inhibitors, and CAR-T cells have all been approved by the FDA, but often suffer from dose-limiting toxicities. T cell bispecifics and CAR-T cells work around the TCR-MHC recognition system by using binding domains to target antigens on the surface of tumor cells, and using engineered signaling domains to transduce the activation signals into the effector cell. Although effective at eliciting an anti-tumor immune response these therapies are often coupled with cytokine release syndrome (CRS), and on-target off-tumor side effects. The multi-specific binding proteins are unique in this context as they will not “override” the natural systems of NK cell activation and inhibition. Instead, the multi-specific binding proteins are designed to sway the balance, and provide additional activation signals to the NK cells, while maintaining NK tolerance to healthy self.

[00134] In some embodiments, the multi-specific binding proteins described herein including an NKG2D-binding domain (*e.g.*, A40-multi-specific binding protein, A49-multi-specific binding protein, C26-multi-specific binding protein, F04-multi-specific binding protein, F43-multi-specific binding protein, F47-multi-specific binding protein, and E79-multi-specific binding protein), which include a binding domain for HER2 delay progression of the tumor more effectively than monoclonal antibodies that include the same tumor antigen-binding domain. In some embodiments, the multi-specific binding proteins including an NKG2D-binding domain and a tumor antigen-binding domain are more effective against cancer metastases than monoclonal antibodies that include the same tumor antigen-binding domain.

III. THERAPEUTIC APPLICATIONS

[00135] The invention provides methods for treating cancer using a multi-specific binding protein described herein and/or a pharmaceutical composition described herein. The methods

may be used to treat a variety of cancers which express HER2 by administering to a patient in need thereof a therapeutically effective amount of a multi-specific binding protein described herein.

[00136] The therapeutic method can be characterized according to the cancer to be treated.

5 For example, in certain embodiments, the cancer is breast, ovarian, esophageal, bladder or gastric cancer, salivary duct carcinoma, salivary duct carcinomas, adenocarcinoma of the lung or aggressive forms of uterine cancer, such as uterine serous endometrial carcinoma.

[00137] In certain other embodiments, the cancer is brain cancer, breast cancer, cervical cancer, colon cancer, colorectal cancer, endometrial cancer, esophageal cancer, leukemia, 10 lung cancer, liver cancer, melanoma, ovarian cancer, pancreatic cancer, rectal cancer, renal cancer, stomach cancer, testicular cancer, or uterine cancer. In yet other embodiments, the cancer is a squamous cell carcinoma, adenocarcinoma, small cell carcinoma, melanoma, neuroblastoma, sarcoma (*e.g.*, an angiosarcoma or chondrosarcoma), larynx cancer, parotid cancer, biliary tract cancer, thyroid cancer, acral lentiginous melanoma, actinic keratoses, 15 acute lymphocytic leukemia, acute myeloid leukemia, adenoid cystic carcinoma, adenomas, adenosarcoma, adenosquamous carcinoma, anal canal cancer, anal cancer, anorectum cancer, astrocytic tumor, Bartholin gland carcinoma, basal cell carcinoma, biliary cancer, bone cancer, bone marrow cancer, bronchial cancer, bronchial gland carcinoma, carcinoid, cholangiocarcinoma, chondrosarcoma, choroid plexus papilloma/carcinoma, chronic 20 lymphocytic leukemia, chronic myeloid leukemia, clear cell carcinoma, connective tissue cancer, cystadenoma, digestive system cancer, duodenum cancer, endocrine system cancer, endodermal sinus tumor, endometrial hyperplasia, endometrial stromal sarcoma, endometrioid adenocarcinoma, endothelial cell cancer, ependymal cancer, epithelial cell cancer, Ewing's sarcoma, eye and orbit cancer, female genital cancer, focal nodular 25 hyperplasia, gallbladder cancer, gastric antrum cancer, gastric fundus cancer, gastrinoma, glioblastoma, glucagonoma, heart cancer, hemangioblastomas, hemangioendothelioma, hemangiomas, hepatic adenoma, hepatic adenomatosis, hepatobiliary cancer, hepatocellular carcinoma, Hodgkin's disease, ileum cancer, insulinoma, intraepithelial neoplasia, interepithelial squamous cell neoplasia, intrahepatic bile duct cancer, invasive squamous cell 30 carcinoma, jejunum cancer, joint cancer, Kaposi's sarcoma, pelvic cancer, large cell carcinoma, large intestine cancer, leiomyosarcoma, lentigo maligna melanomas, lymphoma, male genital cancer, malignant melanoma, malignant mesothelial tumors, medulloblastoma, medulloepithelioma, meningeal cancer, mesothelial cancer, metastatic carcinoma, mouth cancer, mucoepidermoid carcinoma, multiple myeloma, muscle cancer, nasal tract cancer,

nervous system cancer, neuroepithelial adenocarcinoma nodular melanoma, non-epithelial skin cancer, non-Hodgkin's lymphoma, oat cell carcinoma, oligodendroglial cancer, oral cavity cancer, osteosarcoma, papillary serous adenocarcinoma, penile cancer, pharynx cancer, pituitary tumors, plasmacytoma, pseudosarcoma, pulmonary blastoma, rectal cancer, renal
 5 cell carcinoma, respiratory system cancer, retinoblastoma, rhabdomyosarcoma, sarcoma, serous carcinoma, sinus cancer, skin cancer, small cell carcinoma, small intestine cancer, smooth muscle cancer, soft tissue cancer, somatostatin-secreting tumor, spine cancer, squamous cell carcinoma, striated muscle cancer, submesothelial cancer, superficial spreading melanoma, T cell leukemia, tongue cancer, undifferentiated carcinoma, ureter
 10 cancer, urethra cancer, urinary bladder cancer, urinary system cancer, uterine cervix cancer, uterine corpus cancer, uveal melanoma, vaginal cancer, verrucous carcinoma, VIPoma, vulva cancer, well differentiated carcinoma, or Wilms tumor.

[00138] In certain other embodiments, the cancer is non-Hodgkin's lymphoma, such as a B-cell lymphoma or a T-cell lymphoma. In certain embodiments, the non-Hodgkin's
 15 lymphoma is a B-cell lymphoma, such as a diffuse large B-cell lymphoma, primary mediastinal B-cell lymphoma, follicular lymphoma, small lymphocytic lymphoma, mantle cell lymphoma, marginal zone B-cell lymphoma, extranodal marginal zone B-cell lymphoma, nodal marginal zone B-cell lymphoma, splenic marginal zone B-cell lymphoma, Burkitt lymphoma, lymphoplasmacytic lymphoma, hairy cell leukemia, or primary central nervous
 20 system (CNS) lymphoma. In certain other embodiments, the non-Hodgkin's lymphoma is a T-cell lymphoma, such as a precursor T-lymphoblastic lymphoma, peripheral T-cell lymphoma, cutaneous T-cell lymphoma, angioimmunoblastic T-cell lymphoma, extranodal natural killer/T-cell lymphoma, enteropathy type T-cell lymphoma, subcutaneous panniculitis-like T-cell lymphoma, anaplastic large cell lymphoma, or peripheral T-cell
 25 lymphoma.

[00139] The cancer to be treated can be characterized according to the presence of a particular antigen expressed on the surface of the cancer cell. In certain embodiments, the cancer cell can express one or more of the following in addition to HER2: CD2, CD19, CD20, CD30, CD38, CD40, CD52, CD70, EGFR/ERBB1, IGF1R, HER3/ERBB3,
 30 HER4/ERBB4, MUC1, cMET, SLAMF7, PSCA, MICA, MICB, TRAILR1, TRAILR2, MAGE-A3, B7.1, B7.2, CTLA4, and PD1.

IV. COMBINATION THERAPY

[00140] Another aspect of the invention provides for combination therapy. Multi-specific binding proteins described herein be used in combination with additional therapeutic agents to treat the cancer.

5 **[00141]** Exemplary therapeutic agents that may be used as part of a combination therapy in treating cancer, include, for example, radiation, mitomycin, tretinoin, ribomustin, gemcitabine, vincristine, etoposide, cladribine, mitobronitol, methotrexate, doxorubicin, carboquone, pentostatin, nitracrine, zinostatin, cetorelix, letrozole, raltitrexed, daunorubicin, fadrozole, fotemustine, thymalfasin, sobuzoxane, nedaplatin, cytarabine, bicalutamide,
10 vinorelbine, vesnarinone, aminoglutethimide, amsacrine, proglumide, elliptinium acetate, ketanserin, doxifluridine, etretinate, isotretinoin, streptozocin, nimustine, vindesine, flutamide, drogenil, butocin, carmofur, razoxane, sizofilan, carboplatin, mitolactol, tegafur, ifosfamide, prednimustine, picibanil, levamisole, teniposide, improsulfan, enocitabine, lisuride, oxymetholone, tamoxifen, progesterone, mepitiostane, epitiostanol, formestane,
15 interferon-alpha, interferon-2 alpha, interferon-beta, interferon-gamma, colony stimulating factor-1, colony stimulating factor-2, denileukin diftitox, interleukin-2, luteinizing hormone releasing factor and variations of the aforementioned agents that may exhibit differential binding to its cognate receptor, and increased or decreased serum half-life.

[00142] An additional class of agents that may be used as part of a combination therapy in
20 treating cancer is immune checkpoint inhibitors. Exemplary immune checkpoint inhibitors include agents that inhibit one or more of (i) cytotoxic T-lymphocyte-associated antigen 4 (CTLA4), (ii) programmed cell death protein 1 (PD1), (iii) PDL1, (iv) LAG3, (v) B7-H3, (vi) B7-H4, and (vii) TIM3. The CTLA4 inhibitor ipilimumab has been approved by the United States Food and Drug Administration for treating melanoma.

25 **[00143]** Yet other agents that may be used as part of a combination therapy in treating cancer are monoclonal antibody agents that target non-checkpoint targets (*e.g.*, herceptin) and non-cytotoxic agents (*e.g.*, tyrosine-kinase inhibitors).

[00144] Yet other categories of anti-cancer agents include, for example: (i) an inhibitor selected from an ALK Inhibitor, an ATR Inhibitor, an A2A Antagonist, a Base Excision
30 Repair Inhibitor, a Bcr-Abl Tyrosine Kinase Inhibitor, a Bruton's Tyrosine Kinase Inhibitor, a CDC7 Inhibitor, a CHK1 Inhibitor, a Cyclin-Dependent Kinase Inhibitor, a DNA-PK Inhibitor, an Inhibitor of both DNA-PK and mTOR, a DNMT1 Inhibitor, a DNMT1 Inhibitor plus 2-chloro-deoxyadenosine, an HDAC Inhibitor, a Hedgehog Signaling Pathway Inhibitor,

an IDO Inhibitor, a JAK Inhibitor, a mTOR Inhibitor, a MEK Inhibitor, a MELK Inhibitor, a MTH1 Inhibitor, a PARP Inhibitor, a Phosphoinositide 3-Kinase Inhibitor, an Inhibitor of both PARP1 and DHODH, a Proteasome Inhibitor, a Topoisomerase-II Inhibitor, a Tyrosine Kinase Inhibitor, a VEGFR Inhibitor, and a WEE1 Inhibitor; (ii) an agonist of OX40, CD137, CD40, GITR, CD27, HVEM, TNFRSF25, or ICOS; and (iii) a cytokine selected from IL-12, IL-15, GM-CSF, and G-CSF.

[00145] Proteins of the invention can also be used as an adjunct to surgical removal of the primary lesion.

[00146] The amount of multi-specific binding protein and additional therapeutic agent and the relative timing of administration may be selected in order to achieve a desired combined therapeutic effect. For example, when administering a combination therapy to a patient in need of such administration, the therapeutic agents in the combination, or a pharmaceutical composition or compositions comprising the therapeutic agents, may be administered in any order such as, for example, sequentially, concurrently, together, simultaneously and the like. Further, for example, a multi-specific binding protein may be administered during a time when the additional therapeutic agent(s) exerts its prophylactic or therapeutic effect, or *vice versa*.

V. PHARMACEUTICAL COMPOSITIONS

[00147] The present disclosure also features pharmaceutical compositions that contain a therapeutically effective amount of a protein described herein. The composition can be formulated for use in a variety of drug delivery systems. One or more physiologically acceptable excipients or carriers can also be included in the composition for proper formulation. Suitable formulations for use in the present disclosure are found in Remington's Pharmaceutical Sciences, Mack Publishing Company, Philadelphia, Pa., 17th ed., 1985. For a brief review of methods for drug delivery, *see, e.g.*, Langer (Science 249:1527-1533, 1990).

[00148] The intravenous drug delivery formulation of the present disclosure may be contained in a bag, a pen, or a syringe. In certain embodiments, the bag may be connected to a channel comprising a tube and/or a needle. In certain embodiments, the formulation may be a lyophilized formulation or a liquid formulation. In certain embodiments, the formulation may freeze-dried (lyophilized) and contained in about 12-60 vials. In certain embodiments, the formulation may be freeze-dried and 45 mg of the freeze-dried formulation may be contained in one vial. In certain embodiments, the about 40 mg – about 100 mg of freeze-dried formulation may be contained in one vial. In certain embodiments, freeze dried

formulation from 12, 27, or 45 vials are combined to obtain a therapeutic dose of the protein in the intravenous drug formulation. In certain embodiments, the formulation may be a liquid formulation and stored as about 250 mg/vial to about 1000 mg/vial. In certain embodiments, the formulation may be a liquid formulation and stored as about 600 mg/vial.

5 In certain embodiments, the formulation may be a liquid formulation and stored as about 250 mg/vial.

[00149] This present disclosure could exist in a liquid aqueous pharmaceutical formulation including a therapeutically effective amount of the protein in a buffered solution forming a formulation.

10 **[00150]** These compositions may be sterilized by conventional sterilization techniques, or may be sterile filtered. The resulting aqueous solutions may be packaged for use as-is, or lyophilized, the lyophilized preparation being combined with a sterile aqueous carrier prior to administration. The pH of the preparations typically will be between 3 and 11, more preferably between 5 and 9 or between 6 and 8, and most preferably between 7 and 8, such as
15 7 to 7.5. The resulting compositions in solid form may be packaged in multiple single dose units, each containing a fixed amount of the above-mentioned agent or agents. The composition in solid form can also be packaged in a container for a flexible quantity.

[00151] In certain embodiments, the present disclosure provides a formulation with an extended shelf life including the protein of the present disclosure, in combination with
20 mannitol, citric acid monohydrate, sodium citrate, disodium phosphate dihydrate, sodium dihydrogen phosphate dihydrate, sodium chloride, polysorbate 80, water, and sodium hydroxide.

[00152] In certain embodiments, an aqueous formulation is prepared including the protein of the present disclosure in a pH-buffered solution. The buffer of this invention may have a
25 pH ranging from about 4 to about 8, *e.g.*, from about 4.5 to about 6.0, or from about 4.8 to about 5.5, or may have a pH of about 5.0 to about 5.2. Ranges intermediate to the above recited pH's are also intended to be part of this disclosure. For example, ranges of values using a combination of any of the above recited values as upper and/or lower limits are intended to be included. Examples of buffers that will control the pH within this range
30 include acetate (*e.g.* sodium acetate), succinate (such as sodium succinate), gluconate, histidine, citrate and other organic acid buffers.

[00153] In certain embodiments, the formulation includes a buffer system which contains citrate and phosphate to maintain the pH in a range of about 4 to about 8. In certain embodiments the pH range may be from about 4.5 to about 6.0, or from about pH 4.8 to about

5.5, or in a pH range of about 5.0 to about 5.2. In certain embodiments, the buffer system includes citric acid monohydrate, sodium citrate, disodium phosphate dihydrate, and/or sodium dihydrogen phosphate dihydrate. In certain embodiments, the buffer system includes about 1.3 mg/ml of citric acid (*e.g.*, 1.305 mg/ml), about 0.3 mg/ml of sodium citrate (*e.g.*, 0.305 mg/ml), about 1.5 mg/ml of disodium phosphate dihydrate (*e.g.*, 1.53 mg/ml), about 0.9 mg/ml of sodium dihydrogen phosphate dihydrate (*e.g.*, 0.86), and about 6.2 mg/ml of sodium chloride (*e.g.*, 6.165 mg/ml). In certain embodiments, the buffer system includes 1-1.5 mg/ml of citric acid, 0.25 to 0.5 mg/ml of sodium citrate, 1.25 to 1.75 mg/ml of disodium phosphate dihydrate, 0.7 to 1.1 mg/ml of sodium dihydrogen phosphate dihydrate, and 6.0 to 6.4 mg/ml of sodium chloride. In certain embodiments, the pH of the formulation is adjusted with sodium hydroxide.

[00154] A polyol, which acts as a tonicifier and may stabilize the antibody, may also be included in the formulation. The polyol is added to the formulation in an amount which may vary with respect to the desired isotonicity of the formulation. In certain embodiments, the aqueous formulation may be isotonic. The amount of polyol added may also be altered with respect to the molecular weight of the polyol. For example, a lower amount of a monosaccharide (*e.g.*, mannitol) may be added, compared to a disaccharide (such as trehalose). In certain embodiments, the polyol which may be used in the formulation as a tonicity agent is mannitol. In certain embodiments, the mannitol concentration may be about 5 to about 20 mg/ml. In certain embodiments, the concentration of mannitol may be about 7.5 to 15 mg/ml. In certain embodiments, the concentration of mannitol may be about 10-14 mg/ml. In certain embodiments, the concentration of mannitol may be about 12 mg/ml. In certain embodiments, the polyol sorbitol may be included in the formulation.

[00155] A detergent or surfactant may also be added to the formulation. Exemplary detergents include nonionic detergents such as polysorbates (*e.g.*, polysorbates 20, 80 etc.) or poloxamers (*e.g.*, poloxamer 188). The amount of detergent added is such that it reduces aggregation of the formulated antibody and/or minimizes the formation of particulates in the formulation and/or reduces adsorption. In certain embodiments, the formulation may include a surfactant which is a polysorbate. In certain embodiments, the formulation may contain the detergent polysorbate 80 or Tween 80. Tween 80 is a term used to describe polyoxyethylene (20) sorbitanmonooleate (see Fiedler, Lexikon der Hfisstoffe, Editio Cantor Verlag Aulendorf, 4th edi., 1996). In certain embodiments, the formulation may contain between about 0.1 mg/mL and about 10 mg/mL of polysorbate 80, or between about 0.5 mg/mL and

about 5 mg/mL. In certain embodiments, about 0.1% polysorbate 80 may be added in the formulation.

[00156] In embodiments, the protein product of the present disclosure is formulated as a liquid formulation. The liquid formulation may be presented at a 10 mg/mL concentration in either a USP / Ph Eur type I 50R vial closed with a rubber stopper and sealed with an aluminum crimp seal closure. The stopper may be made of elastomer complying with USP and Ph Eur. In certain embodiments vials may be filled with 61.2 mL of the protein product solution in order to allow an extractable volume of 60 mL. In certain embodiments, the liquid formulation may be diluted with 0.9% saline solution.

[00157] In certain embodiments, the liquid formulation of the disclosure may be prepared as a 10 mg/mL concentration solution in combination with a sugar at stabilizing levels. In certain embodiments the liquid formulation may be prepared in an aqueous carrier. In certain embodiments, a stabilizer may be added in an amount no greater than that which may result in a viscosity undesirable or unsuitable for intravenous administration. In certain embodiments, the sugar may be disaccharides, *e.g.*, sucrose. In certain embodiments, the liquid formulation may also include one or more of a buffering agent, a surfactant, and a preservative.

[00158] In certain embodiments, the pH of the liquid formulation may be set by addition of a pharmaceutically acceptable acid and/or base. In certain embodiments, the pharmaceutically acceptable acid may be hydrochloric acid. In certain embodiments, the base may be sodium hydroxide.

[00159] In addition to aggregation, deamidation is a common product variant of peptides and proteins that may occur during fermentation, harvest/cell clarification, purification, drug substance/drug product storage and during sample analysis. Deamidation is the loss of NH₃ from a protein forming a succinimide intermediate that can undergo hydrolysis. The succinimide intermediate results in a 17 dalton mass decrease of the parent peptide. The subsequent hydrolysis results in an 18 dalton mass increase. Isolation of the succinimide intermediate is difficult due to instability under aqueous conditions. As such, deamidation is typically detectable as 1 dalton mass increase. Deamidation of an asparagine results in either aspartic or isoaspartic acid. The parameters affecting the rate of deamidation include pH, temperature, solvent dielectric constant, ionic strength, primary sequence, local polypeptide conformation and tertiary structure. The amino acid residues adjacent to Asn in the peptide chain affect deamidation rates. Gly and Ser following an Asn in protein sequences results in a higher susceptibility to deamidation.

[00160] In certain embodiments, the liquid formulation of the present disclosure may be preserved under conditions of pH and humidity to prevent deamination of the protein product.

[00161] The aqueous carrier of interest herein is one which is pharmaceutically acceptable (safe and non-toxic for administration to a human) and is useful for the preparation of a liquid formulation. Illustrative carriers include sterile water for injection (SWFI), bacteriostatic water for injection (BWFI), a pH buffered solution (*e.g.*, phosphate-buffered saline), sterile saline solution, Ringer's solution or dextrose solution.

[00162] A preservative may be optionally added to the formulations herein to reduce bacterial action. The addition of a preservative may, for example, facilitate the production of a multi-use (multiple-dose) formulation.

[00163] Intravenous (IV) formulations may be the preferred administration route in particular instances, such as when a patient is in the hospital after transplantation receiving all drugs via the IV route. In certain embodiments, the liquid formulation is diluted with 0.9% Sodium Chloride solution before administration. In certain embodiments, the diluted drug product for injection is isotonic and suitable for administration by intravenous infusion.

[00164] In certain embodiments, a salt or buffer components may be added in an amount of 10 mM - 200 mM. The salts and/or buffers are pharmaceutically acceptable and are derived from various known acids (inorganic and organic) with "base forming" metals or amines. In certain embodiments, the buffer may be phosphate buffer. In certain embodiments, the buffer may be glycinate, carbonate, citrate buffers, in which case, sodium, potassium or ammonium ions can serve as counterion.

[00165] A preservative may be optionally added to the formulations herein to reduce bacterial action. The addition of a preservative may, for example, facilitate the production of a multi-use (multiple-dose) formulation.

[00166] The aqueous carrier of interest herein is one which is pharmaceutically acceptable (safe and non-toxic for administration to a human) and is useful for the preparation of a liquid formulation. Illustrative carriers include sterile water for injection (SWFI), bacteriostatic water for injection (BWFI), a pH buffered solution (*e.g.*, phosphate-buffered saline), sterile saline solution, Ringer's solution or dextrose solution.

[00167] This present disclosure could exist in a lyophilized formulation including the proteins and a lyoprotectant. The lyoprotectant may be sugar, *e.g.*, disaccharides. In certain embodiments, the lyoprotectant may be sucrose or maltose. The lyophilized formulation may also include one or more of a buffering agent, a surfactant, a bulking agent, and/or a preservative.

[00168] The amount of sucrose or maltose useful for stabilization of the lyophilized drug product may be in a weight ratio of at least 1:2 protein to sucrose or maltose. In certain embodiments, the protein to sucrose or maltose weight ratio may be of from 1:2 to 1:5.

5 [00169] In certain embodiments, the pH of the formulation, prior to lyophilization, may be set by addition of a pharmaceutically acceptable acid and/or base. In certain embodiments the pharmaceutically acceptable acid may be hydrochloric acid. In certain embodiments, the pharmaceutically acceptable base may be sodium hydroxide.

10 [00170] Before lyophilization, the pH of the solution containing the protein of the present disclosure may be adjusted between 6 to 8. In certain embodiments, the pH range for the lyophilized drug product may be from 7 to 8.

[00171] In certain embodiments, a salt or buffer components may be added in an amount of 10 mM - 200 mM. The salts and/or buffers are pharmaceutically acceptable and are derived from various known acids (inorganic and organic) with “base forming” metals or amines. In certain embodiments, the buffer may be phosphate buffer. In certain
15 embodiments, the buffer may be glycinate, carbonate, citrate buffers, in which case, sodium, potassium or ammonium ions can serve as counterion.

[00172] In certain embodiments, a “bulking agent” may be added. A “bulking agent” is a compound which adds mass to a lyophilized mixture and contributes to the physical structure of the lyophilized cake (*e.g.*, facilitates the production of an essentially uniform lyophilized
20 cake which maintains an open pore structure). Illustrative bulking agents include mannitol, glycine, polyethylene glycol and sorbitol. The lyophilized formulations of the present invention may contain such bulking agents.

[00173] A preservative may be optionally added to the formulations herein to reduce bacterial action. The addition of a preservative may, for example, facilitate the production of
25 a multi-use (multiple-dose) formulation.

[00174] In certain embodiments, the lyophilized drug product may be constituted with an aqueous carrier. The aqueous carrier of interest herein is one which is pharmaceutically acceptable (*e.g.*, safe and non-toxic for administration to a human) and is useful for the preparation of a liquid formulation, after lyophilization. Illustrative diluents include sterile
30 water for injection (SWFI), bacteriostatic water for injection (BWFI), a pH buffered solution (*e.g.*, phosphate-buffered saline), sterile saline solution, Ringer's solution or dextrose solution.

[00175] In certain embodiments, the lyophilized drug product of the current disclosure is reconstituted with either Sterile Water for Injection, USP (SWFI) or 0.9% Sodium Chloride Injection, USP. During reconstitution, the lyophilized powder dissolves into a solution.

5 [00176] In certain embodiments, the lyophilized protein product of the instant disclosure is constituted to about 4.5 mL water for injection and diluted with 0.9% saline solution (sodium chloride solution).

[00177] Actual dosage levels of the active ingredients in the pharmaceutical compositions of this invention may be varied so as to obtain an amount of the active ingredient which is effective to achieve the desired therapeutic response for a particular patient, composition, and
10 mode of administration, without being toxic to the patient.

[00178] The specific dose can be a uniform dose for each patient, for example, 50-5000 mg of protein. Alternatively, a patient's dose can be tailored to the approximate body weight or surface area of the patient. Other factors in determining the appropriate dosage can include the disease or condition to be treated or prevented, the severity of the disease, the route of
15 administration, and the age, sex and medical condition of the patient. Further refinement of the calculations necessary to determine the appropriate dosage for treatment is routinely made by those skilled in the art, especially in light of the dosage information and assays disclosed herein. The dosage can also be determined through the use of known assays for determining dosages used in conjunction with appropriate dose-response data. An individual patient's
20 dosage can be adjusted as the progress of the disease is monitored. Blood levels of the targetable construct or complex in a patient can be measured to see if the dosage needs to be adjusted to reach or maintain an effective concentration. Pharmacogenomics may be used to determine which targetable constructs and/or complexes, and dosages thereof, are most likely to be effective for a given individual (Schmitz *et al.*, *Clinica Chimica Acta* 308: 43-53, 2001; Steimer *et al.*, *Clinica Chimica Acta* 308: 33-41, 2001).
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[00179] In general, dosages based on body weight are from about 0.01 µg to about 100 mg per kg of body weight, such as about 0.01 µg to about 100 mg/kg of body weight, about 0.01 µg to about 50 mg/kg of body weight, about 0.01 µg to about 10 mg/kg of body weight, about 0.01 µg to about 1 mg/kg of body weight, about 0.01 µg to about 100 µg/kg of body weight, about 0.01 µg to about 50 µg/kg of body weight, about 0.01 µg to about 10 µg/kg of body weight, about 0.01 µg to about 1 µg/kg of body weight, about 0.01 µg to about 0.1 µg/kg of body weight, about 0.1 µg to about 100 mg/kg of body weight, about 0.1 µg to about 50 mg/kg of body weight, about 0.1 µg to about 10 mg/kg of body weight, about 0.1 µg to about 1 mg/kg of body weight, about 0.1 µg to about 100 µg/kg of body weight, about 0.1 µg to
30

about 10 µg/kg of body weight, about 0.1 µg to about 1 µg/kg of body weight, about 1 µg to about 100 mg/kg of body weight, about 1 µg to about 50 mg/kg of body weight, about 1 µg to about 10 mg/kg of body weight, about 1 µg to about 1 mg/kg of body weight, about 1 µg to about 100 µg/kg of body weight, about 1 µg to about 50 µg/kg of body weight, about 1 µg to about 10 µg/kg of body weight, about 10 µg to about 100 mg/kg of body weight, about 10 µg to about 50 mg/kg of body weight, about 10 µg to about 10 mg/kg of body weight, about 10 µg to about 1 mg/kg of body weight, about 10 µg to about 100 µg/kg of body weight, about 10 µg to about 50 µg/kg of body weight, about 50 µg to about 100 mg/kg of body weight, about 50 µg to about 50 mg/kg of body weight, about 50 µg to about 10 mg/kg of body weight, about 50 µg to about 1 mg/kg of body weight, about 50 µg to about 100 µg/kg of body weight, about 100 µg to about 100 mg/kg of body weight, about 100 µg to about 50 mg/kg of body weight, about 100 µg to about 10 mg/kg of body weight, about 100 µg to about 1 mg/kg of body weight, about 1 mg to about 100 mg/kg of body weight, about 1 mg to about 50 mg/kg of body weight, about 1 mg to about 10 mg/kg of body weight, about 10 mg to about 100 mg/kg of body weight, about 10 mg to about 50 mg/kg of body weight, about 50 mg to about 100 mg/kg of body weight.

[00180] Doses may be given once or more times daily, weekly, monthly or yearly, or even once every 2 to 20 years. Persons of ordinary skill in the art can easily estimate repetition rates for dosing based on measured residence times and concentrations of the targetable construct or complex in bodily fluids or tissues. Administration of the present invention could be intravenous, intraarterial, intraperitoneal, intramuscular, subcutaneous, intrapleural, intrathecal, intracavitary, by perfusion through a catheter or by direct intralesional injection. This may be administered once or more times daily, once or more times weekly, once or more times monthly, and once or more times annually.

[00181] The description above describes multiple aspects and embodiments of the invention. The patent application specifically contemplates all combinations and permutations of the aspects and embodiments.

EXAMPLES

[00182] The invention now being generally described, will be more readily understood by reference to the following examples, which are included merely for purposes of illustration of certain aspects and embodiments of the present invention, and is not intended to limit the invention.

Example 1 – NKG2D-binding domains bind to NKG2DNKG2D-binding domains bind to purified recombinant NKG2D

[00183] The nucleic acid sequences of human, mouse or cynomolgus NKG2D ectodomains were fused with nucleic acid sequences encoding human IgG1 Fc domains and introduced into mammalian cells to be expressed. After purification, NKG2D-Fc fusion proteins were adsorbed to wells of microplates. After blocking the wells with bovine serum albumin to prevent non-specific binding, NKG2D-binding domains were titrated and added to the wells pre-adsorbed with NKG2D-Fc fusion proteins. Primary antibody binding was detected using a secondary antibody which was conjugated to horseradish peroxidase and specifically recognizes a human kappa light chain to avoid Fc cross-reactivity. 3,3',5,5'-Tetramethylbenzidine (TMB), a substrate for horseradish peroxidase, was added to the wells to visualize the binding signal, whose absorbance was measured at 450 nM and corrected at 540 nM. An NKG2D-binding domain clone, an isotype control or a positive control (selected from SEQ ID NOs:45-48, or anti-mouse NKG2D clones MI-6 and CX-5 available at eBioscience) was added to each well.

[00184] The isotype control showed minimal binding to recombinant NKG2D-Fc proteins, while the positive control bound strongest to the recombinant antigens. NKG2D-binding domains produced by all clones demonstrated binding across human, mouse, and cynomolgus recombinant NKG2D-Fc proteins, although with varying affinities from clone to clone. Generally, each anti-NKG2D clone bound to human (FIG. 3) and cynomolgus (FIG. 4) recombinant NKG2D-Fc with similar affinity, but with lower affinity to mouse (FIG. 5) recombinant NKG2D-Fc.

NKG2D-binding domains bind to cells expressing NKG2D

[00185] EL4 mouse lymphoma cell lines were engineered to express human or mouse NKG2D - CD3 zeta signaling domain chimeric antigen receptors. An NKG2D-binding clone, an isotype control or a positive control was used at a 100 nM concentration to stain extracellular NKG2D expressed on the EL4 cells. The antibody binding was detected using fluorophore-conjugated anti-human IgG secondary antibodies. Cells were analyzed by flow cytometry, and fold-over-background (FOB) was calculated using the mean fluorescence intensity (MFI) of NKG2D-expressing cells compared to parental EL4 cells.

[00186] NKG2D-binding domains produced by all clones bound to EL4 cells expressing human and mouse NKG2D. Positive control antibodies (selected from SEQ ID NO: 45-48, or anti-mouse NKG2D clones MI-6 and CX-5 available at eBioscience) gave the best FOB

binding signal. The NKG2D-binding affinity for each clone was similar between cells expressing human NKG2D (FIG. 6) and mouse (FIG. 7) NKG2D.

Example 2 – NKG2D-binding domains block natural ligand binding to NKG2D

5 Competition With ULBP-6

[00187] Recombinant human NKG2D-Fc proteins were adsorbed to wells of a microplate, and the wells were blocked with bovine serum albumin reduce non-specific binding. A saturating concentration of ULBP-6-His-biotin was added to the wells, followed by addition of the NKG2D-binding domain clones. After a 2-hour incubation, wells were
10 washed and ULBP-6-His-biotin that remained bound to the NKG2D-Fc coated wells was detected by streptavidin-conjugated to horseradish peroxidase and TMB substrate. Absorbance was measured at 450 nM and corrected at 540 nM. After subtracting background, specific binding of NKG2D-binding domains to the NKG2D-Fc proteins was calculated from the percentage of ULBP-6-His-biotin that was blocked from binding to the NKG2D-Fc
15 proteins in wells. The positive control antibody (selected from SEQ ID NOs:45-48) and various NKG2D-binding domains blocked ULBP-6 binding to NKG2D, while isotype control showed little competition with ULBP-6 (FIG. 8).

Competition With MICA

[00188] Recombinant human MICA-Fc proteins were adsorbed to wells of a microplate,
20 and the wells were blocked with bovine serum albumin to reduce non-specific binding. NKG2D-Fc-biotin was added to wells followed by NKG2D-binding domains. After incubation and washing, NKG2D-Fc-biotin that remained bound to MICA-Fc coated wells was detected using streptavidin-HRP and TMB substrate. Absorbance was measured at 450 nM and corrected at 540 nM. After subtracting background, specific binding of NKG2D-
25 binding domains to the NKG2D-Fc proteins was calculated from the percentage of NKG2D-Fc-biotin that was blocked from binding to the MICA-Fc coated wells. The positive control antibody (selected from SEQ ID NOs:45-48) and various NKG2D-binding domains blocked MICA binding to NKG2D, while isotype control showed little competition with MICA (FIG. 9).

30 Competition With Rae-1 delta

[00189] Recombinant mouse Rae-1delta-Fc (purchased from R&D Systems) was adsorbed to wells of a microplate, and the wells were blocked with bovine serum albumin to reduce non-specific binding. Mouse NKG2D-Fc-biotin was added to the wells followed by

NKG2D-binding domains. After incubation and washing, NKG2D-Fc-biotin that remained bound to Rae-1delta-Fc coated wells was detected using streptavidin-HRP and TMB substrate. Absorbance was measured at 450 nM and corrected at 540 nM. After subtracting background, specific binding of NKG2D-binding domains to the NKG2D-Fc proteins was calculated from the percentage of NKG2D-Fc-biotin that was blocked from binding to the Rae-1delta-Fc coated wells. The positive control (selected from SEQ ID NOs:45-48, or anti-mouse NKG2D clones MI-6 and CX-5 available at eBioscience) and various NKG2D-binding domain clones blocked Rae-1delta binding to mouse NKG2D, while the isotype control antibody showed little competition with Rae-1delta (FIG. 10).

Example 3 – NKG2D-binding domain clones activate NKG2D

[00190] Nucleic acid sequences of human and mouse NKG2D were fused to nucleic acid sequences encoding a CD3 zeta signaling domain to obtain chimeric antigen receptor (CAR) constructs. The NKG2D-CAR constructs were then cloned into a retrovirus vector using Gibson assembly and transfected into expi293 cells for retrovirus production. EL4 cells were infected with viruses containing NKG2D-CAR together with 8 µg/mL polybrene. 24 hours after infection, the expression levels of NKG2D-CAR in the EL4 cells were analyzed by flow cytometry, and clones which express high levels of the NKG2D-CAR on the cell surface were selected.

[00191] To determine whether NKG2D-binding domains activate NKG2D, they were adsorbed to wells of a microplate, and NKG2D-CAR EL4 cells were cultured on the antibody fragment-coated wells for 4 hours in the presence of brefeldin-A and monensin. Intracellular TNFα production, an indicator for NKG2D activation, was assayed by flow cytometry. The percentage of TNFα-positive cells was normalized to the cells treated with the positive control. All NKG2D-binding domains activated both human NKG2D (FIG. 11) and mouse NKG2D (FIG. 12).

Example 4 – NKG2D-binding domains activate NK cells

Primary human NK cells

[00192] Peripheral blood mononuclear cells (PBMCs) were isolated from human peripheral blood buffy coats using density gradient centrifugation. NK cells (CD3⁻ CD56⁺) were isolated using negative selection with magnetic beads from PBMCs, and the purity of the isolated NK cells was typically >95%. Isolated NK cells were then cultured in media containing 100 ng/mL IL-2 for 24-48 hours before they were transferred to the wells of a

microplate to which the NKG2D-binding domains were adsorbed, and cultured in the media containing fluorophore-conjugated anti-CD107a antibody, brefeldin-A, and monensin. Following culture, NK cells were assayed by flow cytometry using fluorophore-conjugated antibodies against CD3, CD56 and IFN- γ . CD107a and IFN- γ staining were analyzed in CD3⁻ CD56⁺ cells to assess NK cell activation. The increase in CD107a/IFN- γ double-positive cells is indicative of better NK cell activation through engagement of two activating receptors rather than one receptor. NKG2D-binding domains and the positive control (selected from SEQ ID NOs:45-48) showed a higher percentage of NK cells becoming CD107a⁺ and IFN- γ ⁺ than the isotype control (FIG. 13 & FIG. 14 represent data from two independent experiments, each using a different donor's PBMC for NK cell preparation).

Primary mouse NK cells

[00193] Spleens were obtained from C57Bl/6 mice and crushed through a 70 μ m cell strainer to obtain single cell suspension. Cells were pelleted and resuspended in ACK lysis buffer (purchased from Thermo Fisher Scientific #A1049201; 155mM ammonium chloride, 10mM potassium bicarbonate, 0.01mM EDTA) to remove red blood cells. The remaining cells were cultured with 100 ng/mL hIL-2 for 72 hours before being harvested and prepared for NK cell isolation. NK cells (CD3⁻NK1.1⁺) were then isolated from spleen cells using a negative depletion technique with magnetic beads with typically >90% purity. Purified NK cells were cultured in media containing 100 ng/mL mL-15 for 48 hours before they were transferred to the wells of a microplate to which the NKG2D-binding domains were adsorbed, and cultured in the media containing fluorophore-conjugated anti-CD107a antibody, brefeldin-A, and monensin. Following culture in NKG2D-binding domain-coated wells, NK cells were assayed by flow cytometry using fluorophore-conjugated antibodies against CD3, NK1.1 and IFN- γ . CD107a and IFN- γ staining were analyzed in CD3⁻ NK1.1⁺ cells to assess NK cell activation. The increase in CD107a/IFN- γ double-positive cells is indicative of better NK cell activation through engagement of two activating receptors rather than one receptor. NKG2D-binding domains and the positive control (selected from anti-mouse NKG2D clones MI-6 and CX-5 available at eBioscience) showed a higher percentage of NK cells becoming CD107a⁺ and IFN- γ ⁺ than the isotype control (FIG. 15 & FIG. 16 represent data from two independent experiments, each using a different mouse for NK cell preparation).

Example 5 – NKG2D-binding domains enable cytotoxicity of target tumor cells

[00194] Human and mouse primary NK cell activation assays demonstrate increased cytotoxicity markers on NK cells after incubation with NKG2D-binding domains. To address whether this translates into increased tumor cell lysis, a cell-based assay was utilized where each NKG2D-binding domain was developed into a monospecific antibody. The Fc region was used as one targeting arm, while the Fab region (NKG2D-binding domain) acted as another targeting arm to activate NK cells. THP-1 cells, which are of human origin and express high levels of Fc receptors, were used as a tumor target and a Perkin Elmer DELFIA Cytotoxicity Kit was used. THP-1 cells were labeled with BATDA reagent, and resuspended at 10^5 /mL in culture media. Labeled THP-1 cells were then combined with NKG2D antibodies and isolated mouse NK cells in wells of a microtiter plate at 37 °C for 3 hours. After incubation, 20 μ l of the culture supernatant was removed, mixed with 200 μ l of Europium solution and incubated with shaking for 15 minutes in the dark. Fluorescence was measured over time by a PheraStar plate reader equipped with a time-resolved fluorescence module (Excitation 337nm, Emission 620nm) and specific lysis was calculated according to the kit instructions.

[00195] The positive control, ULBP-6 - a natural ligand for NKG2D, showed increased specific lysis of THP-1 target cells by mouse NK cells. NKG2D antibodies also increased specific lysis of THP-1 target cells, while isotype control antibody showed reduced specific lysis. The dotted line indicates specific lysis of THP-1 cells by mouse NK cells without antibody added (FIG. 17).

Example 6 – NKG2D antibodies show high thermostability

[00196] Melting temperatures of NKG2D-binding domains were assayed using differential scanning fluorimetry. The extrapolated apparent melting temperatures are high relative to typical IgG1 antibodies (FIG. 18).

Example 7 – Multi-specific binding proteins display enhanced ability to activate NK cells

[00197] Peripheral blood mononuclear cells (PBMCs) were isolated from human peripheral blood buffy coats using density gradient centrifugation. NK cells ($CD3^- CD56^+$) were isolated using negative selection with magnetic beads from PBMCs, and the purity of the isolated NK cells was typically >95%. Isolated NK cells were then cultured in media containing 100 ng/mL IL-2 for 24-48 hours before they were transferred to the wells of a

microplate to which multi-specific and bispecific binding proteins were adsorbed respectively, and cultured in the media containing fluorophore-conjugated anti-CD107a antibody, brefeldin-A, and monensin. Following culture, NK cells were assayed by flow cytometry using fluorophore-conjugated antibodies against CD3, CD56 and IFN- γ . CD107a and IFN- γ staining were analyzed in CD3⁺ CD56⁺ cells to assess NK cell activation. The increase in CD107a/IFN- γ double-positive cells is indicative of better NK cell activation. AL2.2 is a multi-specific binding protein containing HER2-binding domain (trastuzumab), NKG2D-binding domain (ULBP-6) and a human IgG1 Fc domain. It was made through a controlled Fab-arm exchange reaction (cFAE) starting from trastuzumab homodimer and ULBP-6-Fc homodimer (*see* Labrijn *et al.*, Nature Protocols 9, 2450-2463). SC2.2 is single chain protein including an scFv derived from trastuzumab, and ULBP-6 (SEQ ID NO:93).

SEQ ID NO:93

MAAAAI PALLLCLPLL FLLFGWSRARRDDPHSLCYDITVIPKFRPGPRWCAVQGGQVD
 EKTF LHYDCGNKTVTPVSPLGKKLNVTMAWKAQNPVLREVVDILTEQLLDIQLENY
 TPKEPLTLQARMSCEQKAEGHSSGSWQFSIDGQTFLFDSEKRMWTTVHPGARKMK
 EKWENDKDVAMSFHYISMGDCIGWLEDFLMGMDSTLEPSAGAPLAMSSGTTQLRA
 TATTLILCCLLIILPCFILPGI

[00198] Analysis of CD107a and IFN- γ staining indicated that isotype control IgG showed no activation of NK cells, while a higher percentage of NK cells becoming CD107a⁺ and IFN- γ ⁺ after stimulation with a multi-specific binding protein compared with a bispecific protein, demonstrating stronger NK cell activation through engagement of two activating receptors (NKG2D and CD16) rather than just one (NKG2D) (FIG. 19). This increase in NK cell activation is expected to translate into more potent tumor cell killing.

Example 8 – Multi-specific binding proteins display enhanced cytotoxicity towards target tumor cells

Primary human NK cell cytotoxicity assay

[00199] Peripheral blood mononuclear cells (PBMCs) were isolated from human peripheral blood buffy coats using density gradient centrifugation. NK cells (CD3⁺ CD56⁺) were isolated using negative selection with magnetic beads from PBMCs, and the purity of the isolated NK cells was typically >95%. NK cells were then cultured overnight in media containing 100ng/mL IL-2 before used in cytotoxicity assays. The following day NK cells were resuspended at 5x10⁵/mL in fresh culture media. Human breast cancer cell SkBr-3 cells

were labeled with BATDA reagent according to Perkin Elmer DELFIA Cytotoxicity Kit and resuspended at 5×10^4 /mL in culture media. Various dilution of the multi-specific binding proteins were made into culture media. NK cells, the labeled SkBr-3 cells and the multi-specific binding proteins were then combined in wells of a microtiter plate and incubated at 37°C for 3 hours. After incubation, 20 μ l of the culture supernatant was removed, mixed with 200 μ l of Europium solution and incubated with shaking for 15 minutes in the dark.

Fluorescence was measured over time by a PheraStar plate reader equipped with a time-resolved fluorescence module (Excitation 337nm, Emission 620nm) and specific lysis was calculated according to the kit instructions. AL0.2 is a multi-specific binding protein containing HER2-binding domain (trastuzumab), NKG2D-binding domain (selected from SEQ ID NO: 1-44)) and a human IgG1 Fc domain. It was made through a controlled Fab-arm exchange reaction (cFAE) starting from trastuzumab homodimer and anti-NKG2D homodimer. AL0.2si is based on AL0.2 and contains an additional D265A mutation in Fc domain which abrogates CD16 binding. Trastuzumab-si is based on Trastuzumab and contains an additional D265A mutation in Fc domain which abrogates CD16 binding. AL2.2 is a multi-specific binding protein containing HER2-binding domain (trastuzumab), NKG2D-binding domain (ULBP-6) and a human IgG1 Fc domain. SC2.2 is single chain protein including an scFv derived from trastuzumab, and ULBP-6.

[00200] AL0.2 showed enhanced lysis of SkBr-3 target cells by human NK cells than trastuzumab in a dose dependent manner, with a p value of 0.0311 in EC50 (FIG. 20). AL0.2si (FIG. 21) and trastuzumab-si (FIG. 22) showed reduction in both potency and maximum specific lysis of SkBr-3 cells compared to AL0.2, with a p-value of 0.0002, and 0.0001 in EC50, respectively (FIGs. 21-22). In addition, AL0.2 showed enhanced lysis of SkBr-3 cells than AL2.2 in a dose-dependent manner (FIG. 23). Isotype control IgG showed no increase in specific lysis at any of the concentrations tested. Together the data have demonstrated that multi-specific binding proteins engaging 2 activating receptors on NK cells and one tumor antigen, induce more potent killing of tumor cells by human NK cells compared to bispecific proteins engaging one activating receptor on NK cells and one tumor antigen.

Primary mouse NK cell cytotoxicity assay

[00201] Spleens were obtained from C57Bl/6 mice and crushed through a 70 μ m cell strainer to obtain single cell suspension. Cells were pelleted and resuspended in ACK lysis buffer (purchased from Thermo Fisher Scientific #A1049201; 155mM ammonium chloride,

10mM potassium bicarbonate, 0.01mM EDTA) to remove red blood cells. The remaining cells were cultured with 100 ng/mL hIL-2 for 72 hours before being harvested and prepared for NK cell isolation. NK cells (CD3⁻NK1.1⁺) were then isolated from spleen cells using a negative depletion technique with magnetic beads with typically >90% purity. Purified NK

5 cells were cultured in media containing 100 ng/mL mIL-15 for 48 hours before resuspended in culture media at 10⁶/mL for cytotoxic assays. RMA-HER2-dTomato, a mouse tumor cell line engineered to express HER2 and dTomato, and its control counterpart, RMA cells expressing zsGreen were used as targets. They were resuspended at 2x10⁵/mL in culture media and seeded into wells of a micro plate at 1:1 ratio. Dilutions of multi-specific protein

10 were made into culture media, and added to the RMA cells together with the NK cells. After incubation overnight at 37 °C with 5% CO₂, the percentage of RMA-HER2-dTomato and RMA-zsGreen cells were determined by flow cytometry using the fluorescent reporter to identify the two cells types. Specific target cell death = (1 - ((% RMA-Ca2T-dTomato cells in treatment group * % RMA-zsGreen cells in control group) / (% RMA-Ca2T-dTomato cells in

15 control group * % RMA-zsGreen cells in treatment group))) * 100%.

[00202] AL2.2 is more potent in redirecting NK cell responses to tumor targets than SC2.2 (FIG. 25) and Trastuzumab (FIG. 24). Control protein showed little impact on specific target death. These data demonstrate the multi-specific binding proteins engaging 2 activating receptors on NK cells and one tumor antigen, induce more potent killing of tumor cells by

20 mouse NK cells compared to bispecific proteins engaging one activating receptor on NK cells and one tumor antigen.

Example 9 – Multi-specific binding proteins bind to NKG2D

[00203] EL4 mouse lymphoma cell lines were engineered to express human NKG2D trispecific binding proteins (TriNKETs) that each contain an NKG2D-binding domain, a

25 HER2-binding domain, and an Fc domain that binds to CD16 as shown in FIG. 1, were tested for their affinity to extracellular NKG2D expressed on EL4 cells. The binding of the multi-specific binding proteins to NKG2D was detected using fluorophore-conjugated anti-human IgG secondary antibodies. Cells were analyzed by flow cytometry, and fold-over-background (FOB) was calculated using the mean fluorescence intensity (MFI) of NKG2D-expressing

30 cells compared to parental EL4 cells.

[00204] TriNKETs tested include HER2-TriNKET-C26 (ADI-28226 and a HER2-binding domain), and HER2-TriNKET-F04 (ADI-29404 and a HER2-binding domain). The HER2-

binding domain used in the tested molecules was composed of a heavy chain variable domain and a light chain variable domain of Trastuzumab.

[00205] The data show that a HER2 targeting TriNKETs of the present disclosure bind to NKG2D (FIG. 26).

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Example 10 – Multi-specific binding proteins bind to human tumor antigen

Trispecific-binding proteins bind to HER2

[00206] Human cancer cell lines expressing HER2 were used to assay the binding of HER2 targeting TriNKETs to the tumor associated antigen. Renal cell carcinoma cell line 786-O expresses low levels of HER2. TriNKETs and optionally the parental anti-HER2 monoclonal antibody (Trastuzumab) were incubated with the cells, and the binding was detected using fluorophore-conjugated anti-human IgG secondary antibodies. Cells were analyzed by flow cytometry, and fold-over-background (FOB) was calculated using the mean fluorescence intensity (MFI) from TriNKETs and Trastuzumab normalized to secondary antibody controls. HER2-TriNKET-C26, and HER2-TriNKET-F04 show comparable levels of binding to HER2 expressed on 786-O cells as compared with Trastuzumab (FIG. 27A).

[00207] RMA cells transduced with human HER2 were used to test binding to cell expressed human HER2 by HER2 targeting TriNKETs. TriNKETs were diluted to 20 µg/mL, and binding was detected using a fluorophore conjugated anti-human IgG secondary antibody. Cells were analyzed by flow cytometry, binding to cell expressed HER2 was compared to isotype stained and unstained cell populations. FIG. 27B and FIG. 27C show binding profiles of TriNKETs containing two distinct NKG2D binding domains (the binding profile of C26.2 TriNKET with HER2-binding site shown in FIG. 27B; the binding profile of F04.2 TriNKET with HER2-binding site shown in FIG. 27C), but with the same HER-binding domain. Both TriNKETs show similar level of binding to cell surface HER2 on RMA cells.

Example 11 – Multi-specific binding proteins activate NK cells

[00208] Peripheral blood mononuclear cells (PBMCs) were isolated from human peripheral blood buffy coats using density gradient centrifugation. NK cells (CD3⁻ CD56⁺) were isolated using negative selection with magnetic beads from PBMCs, and the purity of the isolated NK cells was typically >90%. Isolated NK cells were cultured in media

containing 100 ng/mL IL-2 for activation or rested overnight without cytokine. IL-2-activated NK cells were used within 24-48 hours after activation.

[00209] Human cancer cells expressing a tumor antigen were harvested and resuspended in culture media at 2×10^6 /mL. Monoclonal antibodies or TriNKETs targeting the tumor antigen were diluted in culture media. Activated NK cells were harvested, washed, and resuspended at 2×10^6 /mL in culture media. Cancer cells were then mixed with monoclonal antibodies/TriNKETs and activated NK cells in the presence of IL-2. Brefeldin-A and monensin were also added to the mixed culture to block protein transport out of the cell for intracellular cytokine staining. Fluorophore-conjugated anti-CD107a was added to the mixed culture and the culture was incubated for 4 hours before samples were prepared for FACS analysis using fluorophore-conjugated antibodies against CD3, CD56 and IFN- γ . CD107a and IFN- γ staining was analyzed in CD3⁺ CD56⁺ cells to assess NK cell activation. The increase in CD107a/IFN- γ double-positive cells is indicative of better NK cell activation through engagement of two activating receptors rather than one receptor.

[00210] TriNKETs mediate activation of human NK cells co-cultured with HER2-expressing SkBr-3 cells (FIG. 28A), Colo201 cells (FIG. 28B), and HCC1954 cells (FIG. 28C) respectively as indicated by an increase of CD107a degranulation and IFN- γ production. SkBr-3 cells and HCC1954 cells have high levels of surface HER2 expression, and Colo201 has medium HER2 expression. Compared to the monoclonal antibody trastuzumab, TriNKETs show superior activation of human NK cells in the presence of human cancer cells. NK cells alone, NK cells plus SkBr-3 cells are used as negative controls.

[00211] TriNKETs (C26-TriNKET-HER2 and F04-TriNKET-HER2) mediate activation of human NK cells co-cultured with CD33-expressing human AML Mv4-11 cells showed an increase of CD107a degranulation and IFN- γ production. Compared to the monoclonal anti-CD33 antibody, TriNKETs (C26-TriNKET-HER2 and F04-TriNKET-HER2) showed superior activation of human NK cells in the presence of human cancer cells expressing HER2 (FIGs. 28A-28C).

Example 12 – Trispecific-binding proteins enable cytotoxicity of target cancer cells

[00212] Peripheral blood mononuclear cells (PBMCs) were isolated from human peripheral blood buffy coats using density gradient centrifugation. NK cells (CD3⁺ CD56⁺) were isolated using negative selection with magnetic beads from PBMCs, and the purity of the isolated NK cells was typically >90%. Isolated NK cells were cultured in media

containing 100 ng/mL IL-2 for activation or rested overnight without cytokine. IL-2-activated or rested NK cells were used the following day in cytotoxicity assays.

[00213] In order to test the ability of human NK cells to lyse cancer cells in the presence of TriNKETs, a cyto Tox 96 non-radioactive cytotoxicity assay from Promega (G1780) was used according to the manufacturer's instructions. Briefly, human cancer cells expressing a tumor antigen were harvested, washed, and resuspended in culture media at $1-2 \times 10^5$ /mL. Rested and/or activated NK cells were harvested, washed, and resuspended at $10^5-2.0 \times 10^6$ /mL in the same culture media as that of the cancer cells. In each well of a 96 well plate, 50 μ l of the cancer cell suspension was mixed with 50 μ l of NK cell suspension with or without TriNKETs targeting the tumor antigen expressed on the cancer cells. After incubation at 37 °C with 5% CO₂ for 3 hours and 15 minutes, 10x lysis buffer was added to wells containing only cancer cells, and to wells containing only media for the maximum lysis and negative reagent controls, respectively. The plate was then placed back into the incubator for an additional 45 minutes to reach a total of 4 hours incubation. Cells were then pelleted, and the culture supernatant was transferred to a new 96 well plate and mixed with a substrate for development. The new plate was incubated for 30 minutes at room temperature, and the absorbance was read at 492 nm on a SpectraMax i3x. Percentage of specific lysis of the cancer cells was calculated as follows: % Specific lysis = ((experimental lysis – spontaneous lysis from NK cells alone – spontaneous lysis from cancer cells alone) / (Maximum lysis – negative reagent control)) x 100%.

[00214] TriNKETs enhance NK cell cytotoxicity against targets with low surface expression compared to the cytotoxic activity of trastuzumab, an anti-HER2 monoclonal antibody. Rested human NK cells were mixed with high HER2-expressing SkBr tumor cells and low HER2-expressing 786-O cancer cells, and TriNKETs' ability to enhance the cytotoxic activity of rested human NK cells against the high and low HER2-expressing cancer cells in a dose-responsive manner was assayed. Dotted lines in FIG. 29A and FIG. 29B indicate the cytotoxic activity of rested NK cells against the cancer cells in the absence of TriNKETs. As shown in FIG. 29B, upon mixing activated human NK cells with low HER2-expressing 786-O cells and TriNKET (*e.g.*, CD26-TriNKET and F04-TriNKET, which includes a binding domain for HER2), dose-responsive cytotoxic activity of activated human NK cells against the cancer cells was observed.

Example 13 – Synergistic activation of human NK cells by cross-linking NKG2D and CD16

Primary human NK cell activation assay

[00215] Peripheral blood mononuclear cells (PBMCs) were isolated from peripheral
 5 human blood buffy coats using density gradient centrifugation. NK cells were purified from
 PBMCs using negative magnetic beads (StemCell # 17955). NK cells were >90% CD3⁻
 CD56⁺ as determined by flow cytometry. Cells were then expanded 48 hours in media
 containing 100 ng/mL hIL-2 (Peprotech #200-02) before use in activation assays. Antibodies
 were coated onto a 96-well flat-bottom plate at a concentration of 2 µg/ml (anti-CD16,
 10 Biolegend # 302013) and 5 µg/mL (anti-NKG2D, R&D #MAB139) in 100 µl sterile PBS
 overnight at 4 °C followed by washing the wells thoroughly to remove excess antibody. For
 the assessment of degranulation IL-2-activated NK cells were resuspended at 5×10⁵ cells/ml
 in culture media supplemented with 100 ng/mL hIL2 and 1 µg/mL APC-conjugated anti-
 CD107a mAb (Biolegend # 328619). 1×10⁵ cells/well were then added onto antibody coated
 15 plates. The protein transport inhibitors Brefeldin A (BFA, Biolegend # 420601) and
 Monensin (Biolegend # 420701) were added at a final dilution of 1:1000 and 1:270
 respectively. Plated cells were incubated for 4 hours at 37 °C in 5% CO₂. For intracellular
 staining of IFN-γ NK cells were labeled with anti-CD3 (Biolegend #300452) and anti-CD56
 mAb (Biolegend # 318328) and subsequently fixed and permeabilized and labeled with anti-
 20 IFN-γ mAb (Biolegend # 506507). NK cells were analyzed for expression of CD107a and
 IFN-γ by flow cytometry after gating on live CD56⁺CD3⁻ cells.

[00216] To investigate the relative potency of receptor combination, crosslinking of
 NKG2D or CD16 and co-crosslinking of both receptors by plate-bound stimulation was
 performed. As shown in Figure 30 (FIGs. 30A-30C), combined stimulation of CD16 and
 25 NKG2D resulted in highly elevated levels of CD107a (degranulation) (FIG. 30A) and/or
 IFN-γ production (FIG. 30B). Dotted lines represent an additive effect of individual
 stimulations of each receptor.

[00217] CD107a levels and intracellular IFN-γ production of IL-2-activated NK cells
 were analyzed after 4 hours of plate-bound stimulation with anti-CD16, anti-NKG2D or a
 30 combination of both monoclonal antibodies. Graphs indicate the mean (n = 2) ± SD. FIG.
 19A demonstrates levels of CD107a; FIG. 30B demonstrates levels of IFNγ; FIG. 30C
 demonstrates levels of CD107a and IFNγ. Data shown in FIGs. 30A-30C are representative
 of five independent experiments using five different healthy donors.

[00218] CD107a degranulation and intracellular IFN- γ production of IL-2-activated NK cells were analyzed after 4 hours of plate-bound stimulation with trastuzumab, anti-NKG2D, or a TriNKET derived from the binding domains of trastuzumab and the anti-NKG2D antibody (FIG. 31). In all cases antibodies tested were of the human IgG1 isotype. Graphs
5 indicate the mean ($n = 2$) $\pm$ SD.

Example 14 – Properties of the TriNKETs

Assessment of TriNKET binding to cell-expressed human NKG2D

[00219] EL4 cells transduced with human NKG2D were used to test binding to cell-
10 expressed human NKG2D. TriNKETs were diluted to 20 μ g/mL, and then diluted serially. The mAb or TriNKET dilutions were used to stain cells, and binding of the TriNKET or mAb was detected using a fluorophore-conjugated anti-human IgG secondary antibody. Cells were analyzed by flow cytometry, binding MFI was normalized to secondary antibody controls to obtain fold over background values.

Assessment of TriNKET binding to cell-expressed human cancer antigens

[00220] Human cancer cell lines expressing HER2 were used to assess tumor antigen
binding of TriNKETs derived from different NKG2D targeting clones. The human renal cell carcinoma cell line 786-O expresses low levels of HER2 and was used to assess TriNKET
binding to cell-expressed HER2. TriNKETs were diluted to 20 μ g/mL, and were incubated
20 with the respective cells. Binding of the TriNKET was detected using a fluorophore-conjugated anti-human IgG secondary antibody. Cells were analyzed by flow cytometry, binding MFI to cell expressing HER2 was normalized to secondary antibody controls to obtain fold over background values.

Determination of antibody binding capacity of human HER2-positive cancer cell lines

[00221] Antibody binding capacity (ABC) of HER2-positive human cancer cell lines was
25 measured. The Quantum Simply Cellular kit from Bangs Lab was used (#815), and the manufacturer instructions were followed for the preparation of antibody labeled beads. Briefly, each of the four populations of beads were stained with a saturating amount of anti-HER2 antibody, and the cell populations were also stained with a saturating amount of the
30 same antibody. Sample data was acquired for each bead population, as well as the cell populations. The QuickCal worksheet, provided with the kit, was used for the generation of a standard curve and extrapolation of ABC values for each of the cell lines.

Activation of primary NK cells by TriNKETs

[00222] PBMCs were isolated from human peripheral blood buffy coats using density gradient centrifugation. Isolated PBMCs were washed and prepared for NK cell isolation. NK cells were isolated using a negative selection technique with magnetic beads; the purity of isolated NK cells was typically >90% CD3-CD56+. Isolated NK cells were cultured in media containing 100ng/mL IL-2 for activation or rested overnight without cytokine. IL-2-activated NK cells were used 24-48 hours later; rested NK cells were always used the day after purification.

[00223] Human cancer cell lines expressing a cancer target of interest were harvested from culture, and cells were adjusted to 2×10^6 /mL. Monoclonal antibodies or TriNKETs targeting the cancer target of interest were diluted in culture media. Rested and/or activated NK cells were harvested from culture, cells were washed, and were resuspended at 2×10^6 /mL in culture media. IL-2, and fluorophore-conjugated anti-CD107a were added to the NK cells for the activation culture. Brefeldin-A and monensin were diluted into culture media to block protein transport out of the cell for intracellular cytokine staining. Into a 96-well plate 50 μ l of tumor targets, mAbs/TriNKETs, BFA/monensin, and NK cells were added for a total culture volume of 200 μ l. The plate was cultured for 4 hours before samples were prepared for FACS analysis.

[00224] Following the 4 hour activation culture, cells were prepared for analysis by flow cytometry using fluorophore-conjugated antibodies against CD3, CD56 and IFN γ . CD107a and IFN γ staining was analyzed in CD3-CD56+ populations to assess NK cell activation.

Primary human NK cell cytotoxicity assay

[00225] PBMCs were isolated from human peripheral blood buffy coats using density gradient centrifugation. Isolated PBMCs were washed and prepared for NK cell isolation. NK cells were isolated using a negative selection technique with magnetic beads, purity of isolated NK cells was typically >90% CD3-CD56+. Isolated NK cells were cultured in media containing 100 ng/mL IL-2 or were rested overnight without cytokine. IL-2-activated or rested NK cells were used the following day in cytotoxicity assays.

Cyto Tox 96 LHD release assay:

[00226] The ability of human NK cells to lyse tumor cells was measured with or without the addition of TriNKETs using the cyto Tox 96 non-radioactive cytotoxicity assay from Promega (G1780). Human cancer cell lines expressing a cancer target of interest were

harvested from culture, cells were washed with PBS, and were resuspended in growth media at $1-2 \times 10^5$ /mL for use as target cells. 50 μ l of the target cell suspension were added to each well. Monoclonal antibodies or TriNKETs targeting a cancer antigen of interest were diluted in culture media, 50 μ l of diluted mAb or TriNKET were added to each well. Rested and/or
 5 activated NK cells were harvested from culture, cells were washed, and were resuspended at $10^5-2.0 \times 10^6$ /mL in culture media depending on the desired E:T ratio. 50 μ l of NK cells were added to each well of the plate to make a total of 150 μ l culture volume. The plate was incubated at 37 °C with 5% CO₂ for 3 hours and 15 minutes. After the incubation, 10x lysis buffer was added to wells of target cells alone, and to wells containing media alone, for
 10 maximum lysis and volume controls. The plate was then placed back into the incubator for an additional 45 minutes, to make to total of 4 hours of incubation before development.

[00227] After incubation, the plate was removed from the incubator and the cells were pelleted by centrifugation at 200g for 5 minutes. 50 μ l of culture supernatant were transferred to a clean microplate and 50 μ l of substrate solution were added to each well. The
 15 plate was protected from the light and incubated for 30 minutes at room temperature. 50 μ l of stop solution were added to each well, and absorbance was read at 492nm on a SpectraMax i3x. % Specific lysis was calculated as follows: % Specific lysis = ((Experimental release – Spontaneous release from effector – Spontaneous release from target) / (Maximum release – Spontaneous release)) * 100%.

20 *DELFLIA cytotoxicity assay:*

[00228] Human cancer cell lines expressing a target of interest were harvested from culture, cells were washed with PBS, and were resuspended in growth media at 10^6 /mL for labeling with BATDA reagent (Perkin Elmer AD0116). Manufacturer instructions were followed for labeling of the target cells. After labeling cells were washed 3x with PBS, and
 25 were resuspended at $0.5-1.0 \times 10^5$ /mL in culture media. To prepare the background wells an aliquot of the labeled cells was put aside, and the cells were spun out of the media. 100 μ l of the media were carefully added to wells in triplicate to avoid disturbing the pelleted cells. 100 μ l of BATDA labeled cells were added to each well of the 96-well plate. Wells were saved for spontaneous release from target cells, and wells were prepared for max lysis of
 30 target cells by addition of 1% Triton-X. Monoclonal antibodies or TriNKETs against the tumor target of interest were diluted in culture media and 50 μ l of diluted mAb or TriNKET were added to each well. Rested and/or activated NK cells were harvested from culture, cells were washed, and were resuspended at $10^5-2.0 \times 10^6$ /mL in culture media depending on the

desired E:T ratio. 50 µl of NK cells were added to each well of the plate to make a total of 200 µl culture volume. The plate was incubated at 37 °C with 5% CO₂ for 2-3 hours before developing the assay.

[00229] After culturing for 2-3 hours, the plate was removed from the incubator and the
 5 cells were pelleted by centrifugation at 200g for 5 minutes. 20 µl of culture supernatant was transferred to a clean microplate provided from the manufacturer, 200 µl of room temperature europium solution was added to each well. The plate was protected from the light and incubated on a plate shaker at 250rpm for 15 minutes. Plate was read using either Victor 3 or SpectraMax i3X instruments. % Specific lysis was calculated as follows: % Specific lysis =
 10 $((\text{Experimental release} - \text{Spontaneous release}) / (\text{Maximum release} - \text{Spontaneous release})) * 100\%$.

Long term human PBMC cytotoxicity assay

[00230] SkBr-3 target cells were labeled with BacMam 3.0 NucLight Green (#4622) to allow for tracking of the target cells. The manufacturer's protocol was followed for labeling
 15 of SkBr-3 target cells. Annexin V Red (Essen Bioscience #4641) was diluted and prepared according to the manufacturer's instructions. Monoclonal antibodies or TriNKETs were diluted into culture media. 50 µl of mAbs or TriNKETs, Annexin V, and rested NK cells were added to wells of a 96 well plate already containing labeled SkBr-3 cells; 50 µl of complete culture media was added for a total of 200 µl culture volume.

20 [00231] Image collection was setup on the IncuCyte S3. Images for the phase, green, and red channels were collected every hour, with 2 images per well. Image analysis was done using the IncuCyte S3 software. Masks for the green and red channels were created to count the number of tumor cells, and annexin V-positive cells respectively. To calculate the %
 25 annexin V-positive Mv4-11 target cells the following formula was used. % Annexin V-positive SkBr-3 cells = $((\text{overlap object count}) / (\text{green object count})) * 100\%$.

Comparing a TriNKET that targets HER+ cancer Cells with SC2.2

[00232] A TriNKET targeting HER2 is more effective than Trastuzumab at reducing SkBr-3 cell number, and only 60% of the cells from time zero were left after 60 hours. A TriNKET of the present disclosure that targets HER2 expressing tumor/cancer cells is more
 30 effective than SC2.2 — a single chain bispecific molecule built from an scFv derived from trastuzumab linked to ULBP-6, a ligand for NKG2D. SC2.2 binds HER2+ cancer cells and NKG2D+ NK cells simultaneously. Therefore, effectiveness of SC2.2 in reducing HER2+

cancer cell number was investigated. *In vitro* activation and cytotoxicity assays demonstrated that SC2.2 was effective in activating and killing NK cells. However, SC2.2 failed to demonstrate efficacy in the RMA/S-HER2 subcutaneous tumor model. The efficacy of SC2.2 was also tested *in vivo* using an RMA/S-HER2 overexpressing syngeneic mouse model. In this mouse model, SC2.2 failed to demonstrate control of tumor growth compared to vehicle control. Thus, although SC2.2 was able to activate and kill NK cells, and binds to HER2+ cancer cells, these properties were insufficient to effectively control HER2+ tumor growth.

Assessment of SC2.2 serum half-life in C57Bl/6 mice

[00233] To determine the serum half-life of SC2.2 in C57Bl/6 mice, SC2.2 was labeled with a fluorescent tag to track its concentration in vivo. SC2.2 was labeled with IRDye 800CW (Licor #929-70020). The labeled protein was injected intravenously into 3 C57Bl/6 mice, blood was taken from each mouse at the indicated time points. After collection blood was centrifuged at 1000g for 15 minutes and serum was collected from each sample and stored at 4C until all time points were collected.

[00234] Serum was imaged using an Odyssey CLx infrared imaging system, the fluorescent signal from the 800 channel was quantified using Image J software. Image intensities were normalized to the first time point, and the data was fit to a biphasic decay equation. In this experimental system the beta half-life of SC2.2 was calculated to be around 7 hours.

***In vivo* testing of SC2.2 against RMA/S-HER2 subcutaneous tumors**

[00235] An *in vivo* study was designed according to FIG. 37 to test the efficacy of SC2.2 against subcutaneous RMA/S-HER2 tumors. 10^6 RMA/S cells transduced with human HER2 were injected subcutaneously into the flank of 20 C57Bl/6 mice. Starting day 2 after tumor innoculation SC2.2 was dosed daily via IP injection. SC2.2 was dosed at a high and a low concentrations along with a vehicle control. Starting day 4 after tumor innoculation tumors were measured Monday, Wednesday, and Friday for the duration of the study. Tumor volume was calculated using the following formula: Tumor volume = Length x width x height.

Antibody binding capacity of human HER2-positive cancer cell lines

[00236] Table 10 shows the results of HER2 surface quantification. SkBr-3 and HCC1954 cells were identified to have high (+++) levels of surface HER2. ZR-75-1 and Colo201

showed medium levels (++) of surface HER2, and 786-O showed the lowest level of HER2 (+).

[00237] Table 10: ABC of HER2-positive cancer cell lines

Cell Line	HER2 expression	ABC
786-0	Low	28,162
Colo201	Medium	273,568
ZR-75-1	Medium	281,026
SkBr-3	High	6,820,532
HCC1954	High	10,569,869

5 Primary human NK cells are activated by TriNKETs in co-culture with human cancer lines expressing varying levels of HER2

[00238] FIGs. 28A – 28C show that TriNKETs and trastuzumab were able to activate primary human NK cells in co-culture with HER2-positive human tumor cells, indicated by an increase in CD107a degranulation and IFN γ cytokine production. Compared to the monoclonal antibody trastuzumab, both TriNKETs (HER2-TriNKET-C26 and HER2-TriNKET-F04) showed superior activation of human NK cells with a variety of human HER2 cancer cells.

[00239] FIG. 28A shows that human NK cells are activated by TriNKETs when cultured with SkBr-3 cells. FIG. 28B shows that human NK cells are activated by TriNKETs when cultured with Colo201 cells. FIG. 28C shows that human NK cell are activated by TriNKETs when cultured with HCC1954 cells.

TriNKETs enhance cytotoxicity of rested and IL-2-activated human NK cells

[00240] FIGs. 32A – 32C show TriNKET enhancement of cytotoxic activity using IL-2-activated and rested human NK cells. FIG. 32A shows percent specific lysis of SkBr-3 tumor cells by rested human NK cells. FIG. 32B shows percent specific lysis of SkBr-3 tumor cells by IL-2-activated human NK cells. IL-2-activated and rested NK cell populations came from the same donor. Compared to trastuzumab, TriNKETs more potently direct responses against SkBr-3 cells by either activated or rested NK cell populations. FIG. 32C shows percent specific lysis of HER2-expressing NCI-H661 lung cancer cells by rested human NK cells.

Two TriNKETs with different NKG2D-binding domains are able to induce higher maximal lysis of NCI-H661 HER2+ cancer cells compared to the monoclonal antibody Trastuzumab.

TriNKETs enhance NK cell cytotoxicity against targets with low surface expression

[00241] Effects of TriNKETs against targets cells with low HER2 surface expression was investigated. FIGs. 29A-29B show TriNKETs provide a greater advantage against HER2-medium and low cancers compared to trastuzumab. FIG. 29A shows activated human NK cell killing of HER2-high SkBr-3 tumor cells. FIG. 29B shows human NK cell killing of HER2-low 786-O tumor cells. TriNKETs provide a greater advantage compared to trastuzumab against cancer cells with low HER2 expression..

10 **The advantage of TriNKETs in treating cancers with high expression of FcR, or in tumor microenvironments with high levels of FcR**

[00242] Monoclonal antibody therapy has been approved for the treatment of many cancer types, including both hematological and solid tumors. While the use of monoclonal antibodies in cancer treatment has improved patient outcomes, there are still limitations.

15 Mechanistic studies have demonstrated monoclonal antibodies exert their effects on tumor growth through multiple mechanisms including ADCC, CDC, phagocytosis, and signal blockade amongst others.

[00243] Most notably, ADCC is thought to be a major mechanism through which monoclonal antibodies exert their effect. ADCC relies on antibody Fc engagement of the low-affinity FcγRIII (CD16) on the surface of natural killer cells, which mediate direct lysis of the tumor cell. Amongst FcγR, CD16 has the lowest affinity for IgG Fc, FcγRI (CD64) is the high-affinity FcR, and binds about 1000 times stronger to IgG Fc than CD16.

[00244] CD64 is normally expressed on many hematopoietic lineages such as the myeloid lineage, and can be expressed on tumors derived from these cell types, such as acute myeloid leukemia (AML). Immune cells infiltrating into the tumor, such as MDSCs and monocytes, also express CD64 and are known to infiltrate the tumor microenvironment. Expression of CD64 by the tumor or in the tumor microenvironment can have a detrimental effect on monoclonal antibody therapy. Expression of CD64 in the tumor microenvironment makes it difficult for these antibodies to engage CD16 on the surface of NK cells, as the antibodies prefer to bind the high-affinity receptor. Through targeting two activating receptors on the surface of NK cells, TriNKETs may be able to overcome the detrimental effect of CD64 expression on monoclonal antibody therapy.

Killing of normal myeloid and normal B cells in PBMC cultures: TriNKETs provide better safety profile through less on-target off-tumor side effects

[00245] Natural killer cells and CD8 T cells are both able to directly lyse tumor cells, although the mechanisms through which NK cells and CD8 T cell recognize normal self from tumor cells differ. The activity of NK cells is regulated by the balance of signals from activating (NCRs, NKG2D, CD16, *etc.*) and inhibitory (KIRs, NKG2A, *etc.*) receptors. The balance of these activating and inhibitory signals allow NK cells to determine healthy self-cells from stressed, virally infected, or transformed self-cells. This “built-in” mechanism of self-tolerance, will help protect normal healthy tissue from NK cell responses. To extend this principle, the self-tolerance of NK cells will allow TriNKETs to target antigens expressed both on self and tumor without off tumor side effects, or with an increased therapeutic window.

[00246] Unlike natural killer cells, T cells require recognition of a specific peptide presented by MHC molecules for activation and effector functions. T cells have been the primary target of immunotherapy, and many strategies have been developed to redirect T cell responses against the tumor. T cell bispecifics, checkpoint inhibitors, and CAR-T cells have all been approved by the FDA, but often suffer from dose-limiting toxicities. T cell bispecifics and CAR-T cells work around the TCR-MHC recognition system by using binding domains to target antigens on the surface of tumor cells, and using engineered signaling domains to transduce the activation signals into the effector cell. Although effective at eliciting an anti-tumor immune response these therapies are often coupled with cytokine release syndrome (CRS), and on-target off-tumor side effects. TriNKETs are unique in this context as they will not “override” the natural systems of NK cell activation and inhibition. Instead, TriNKETs are designed to sway the balance, and provide additional activation signals to the NK cells, while maintaining NK tolerance to healthy self.

[00247] PBMCs were isolated from whole blood by density gradient centrifugation. Any contaminating red blood cells were lysed by incubation in ACK lysis buffer. PBMCs were washed 3x in PBS, and total PBMCs were counted. PBMCs were adjusted to 10^6 /mL in primary cell culture media. 1mL of PBMCs were seeded into wells of a 24 well plate, the indicated TriNKETs or mAbs were added to the PBMC cultures at 10 μ g/mL. Cells were cultured overnight at 37°C with 5% CO₂. The following day (24 hours later) PBMCs were harvested from culture and prepared for FACS analysis. The percentage of CD45+; CD19+ B cells and CD45+; CD33+; CD11b+ myeloid cells was analyzed over the different treatment groups.

[00248] FIGs. 33A & 33B shows B cells from a health donor are sensitive to TriNKET mediated lysis, FIGs. 33C & 33D show that autologous myeloid cells are protected from TriNKET mediated NK cell responses, and, therefore, are resistant to TriNKET lysis. PBMCs treated with TriNKETs targeting CD20 showed reduced frequency of CD19+ B cells with the CD45+ lymphocyte population (FIG. 33A), but no effect in CD45+, CD3-, CD56- lymphocyte population (FIG. 33B). In these cultures the frequency of CD45+, CD33+, CD11b+ myeloid cells (FIG. 33C), or the frequency of CD45+, CD33+, CD11b+ myeloid cells (FIG. 33D) were unchanged.

TriNKETs mediate hPBMC killing of SkBr-3 tumor cells in long-term co-cultures

Primary human PBMC cytotoxicity assay

[00249] FIG. 34 shows long term killing of SkBr-3 cells in culture with human PBMCs. When cultured alone SkBr-3 cells proliferate and almost double in 60 hours. When human PBMCs are added to SkBr-3 cells in culture the rate of proliferation is slowed, and when an isotype control TriNKET targeting CD33 is added proliferation is also slowed, but to a lesser extent. When cultures are treated with Trastuzumab, SkBr-3 no longer proliferate and, after 60 hours, only 80% of the cells from time zero are left. As SkBr-3 cells are sensitive to HER2 signal blockade, the effect on SkBr-3 cell growth could be mediated by HER2 signal blockade or through Fc effector functions such as ADCC.

Example 15 – Cytotoxic activity of rested human NK cells mediated by TriNKETs, monoclonal antibodies, or bispecific antibodies against HER2-positive cells

[00250] PBMCs were isolated from human peripheral blood buffy coats using density gradient centrifugation. Isolated PBMCs were washed and prepared for NK cell isolation. NK cells were isolated using a negative selection technique with magnetic beads; the purity of the isolated NK cells was typically >90% CD3-CD56+. Isolated NK cells were cultured in media containing 100 ng/mL IL-2 or were rested overnight without cytokine. IL-2-activated or rested NK cells were used the following day in cytotoxicity assays.

DELFIa cytotoxicity assay:

[00251] Human cancer cell lines expressing a target of interest were harvested from culture, cells were washed with HBS, and were resuspended in growth media at 10^6 /mL for labeling with BATDA reagent (Perkin Elmer AD0116). Manufacturer instructions were

followed for labeling of the target cells. After labeling, cells were washed 3x with HBS, and were resuspended at $0.5-1.0 \times 10^5/\text{mL}$ in culture media. To prepare the background wells an aliquot of the labeled cells was put aside, and the cells were spun out of the media. 100 μl of the media was carefully added to wells in triplicate to avoid disturbing the pelleted cells. 100 μl of BATDA labeled cells were added to each well of the 96-well plate. Wells were saved for spontaneous release from target cells, and wells were prepared for maximal lysis of target cells by addition of 1% Triton-X. Monoclonal antibodies or TriNKETs against the tumor target of interest were diluted in culture media and 50 μl of diluted mAb or TriNKET was added to each well. Rested and/or activated NK cells were harvested from culture, the cells were washed and were resuspended at $10^5-2.0 \times 10^6/\text{mL}$ in culture media depending on the desired E:T ratio. 50 μl of NK cells were added to each well of the plate to make a total 200 μl culture volume. The plate was incubated at 37°C with 5% CO_2 for 2-3 hours before developing the assay.

[00252] After culturing for 2-3 hours, the plate was removed from the incubator and the cells were pelleted by centrifugation at 200g for 5 minutes. 20 μl of culture supernatant was transferred to a clean microplate provided from the manufacturer and 200 μl of room temperature europium solution was added to each well. The plate was protected from the light and incubated on a plate shaker at 250rpm for 15 minutes. The plate was read using either Victor 3 or SpectraMax i3X instruments. % Specific lysis was calculated as follows: % Specific lysis = ((Experimental release – Spontaneous release) / (Maximum release – Spontaneous release)) * 100%.

Combination of monoclonal antibody and bispecific NK cell engager does not recapitulate TriNKET activity

[00253] FIG. 35 shows the cytotoxic activity of rested human NK cells mediated by TriNKETs, monoclonal antibodies, or bispecific antibodies against the HER2-positive Colo-201 cell line. A TriNKET (ADI-29404 (F04)) targeting HER2-induced maximum lysis of Colo-201 cells by rested human NK cells. The D265A mutation was introduced into the CH2 domain of the TriNKET to abrogate FcR binding. The HER2-TriNKET (ADI-29404 (F04))-D265A failed to mediate lysis of Colo-201 cells, demonstrating the importance of dual targeting of CD16 and NKG2D on NK cells. To further demonstrate the importance of dual targeting on NK cells, the monoclonal antibody Trastuzumab was used to target HER2 and mediate ADCC by NK cells, Trastuzumab alone was able to increase NK cell lysis of Colo-201 cells, but maximum lysis achieved by Trastuzumab alone was about 4x lower compared

to the TriNKET. To understand the importance of having CD16 and NKG2D targeting on the same molecule, TriNKET (ADI-29404 (F04)) activity was compared to the activity of a bispecific antibody targeting HER2 and NKG2D, combined with Trastuzumab. When used at equimolar concentrations the combination of bispecific and Trastuzumab was not able to mediate maximal lysis of Colo-201 cells by rested human NK cells. The failure of Trastuzumab + bispecific combination demonstrates the importance of containing the trispecific-binding of TriNKETs in one molecule.

Example 16 – Bridging assay

[00254] RMA cells transduced with human HER2 were used to test simultaneous binding to HER2 and NKG2D by HER2 targeting TriNKETs. The TriNKETs were used to stain surface HER2 at 20 µg/mL. Binding of the TriNKET was then detected using biotinylated recombinant human NKG2D-Fc. Bound NKG2D-Fc was then detected using streptavidin-APC. Cells were analyzed by flow cytometry, and TriNKET-bridging was compared to isotype stained and unstained cell populations. FIG. 38A shows that TriNKET-C26 that includes a binding domain for HER2, bridges hNKG2D-Fc to RMA-HER2 cells, and FIG. 38B shows TriNKET-F04 that includes a binding domain for HER2, bridges hNKG2D-Fc to RMA-HER2 cells.

INCORPORATION BY REFERENCE

[00255] The entire disclosure of each of the patent documents and scientific articles referred to herein is incorporated by reference for all purposes.

EQUIVALENTS

[00256] The invention may be embodied in other specific forms without departing from the spirit or essential characteristics thereof. The foregoing embodiments are therefore to be considered in all respects illustrative rather than limiting the invention described herein. Scope of the invention is thus indicated by the appended claims rather than by the foregoing description, and all changes that come within the meaning and range of equivalency of the claims are intended to be embraced therein.

CLAIMSWHAT IS CLAIMED IS:

1. A protein comprising:
 - (a) a first antigen-binding site that binds NKG2D;
 - (b) a second antigen-binding site that binds HER2; and
 - (c) an antibody Fc domain or a portion thereof sufficient to bind CD16, or a third antigen-binding site that binds CD16.
2. The protein of claim 1, wherein the first antigen-binding site binds to NKG2D in humans, non-human primates, and rodents.
3. The protein of claim 1 or 2, wherein the first antigen-binding site comprises a heavy chain variable domain and a light chain variable domain.
4. A protein according to claim 3, wherein the heavy chain variable domain and the light chain variable domain are present on the same polypeptide.
5. The protein according to any one of claims 3-4, wherein the second antigen-binding site comprises a heavy chain variable domain and a light chain variable domain.
6. The protein according to claim 5, wherein the heavy chain variable domain and the light chain variable domain of the second antigen-binding site are present on the same polypeptide.
7. The protein according to claim 5 or 6, wherein the light chain variable domain of the first antigen-binding site has an amino acid sequence identical to the amino acid sequence of the light chain variable domain of the second antigen-binding site.
8. The protein according to any one of the preceding claims, wherein the first antigen-binding site comprises a heavy chain variable domain at least 90% identical to SEQ ID NO:1.
9. The protein according to any of claims 1-7, wherein the first antigen-binding site comprises a heavy chain variable domain at least 90% identical to SEQ ID NO:41 and a light chain variable domain at least 90% identical to SEQ ID NO:42.

10. The protein according to any of claims 1-7, wherein the first antigen-binding site comprises a heavy chain variable domain at least 90% identical to SEQ ID NO:43 and a light chain variable domain at least 90% identical to SEQ ID NO:44.
11. The protein according to any of claims 1-7, wherein the first antigen-binding site comprises a heavy chain variable domain at least 90% identical to SEQ ID NO:45 and a light chain variable domain at least 90% identical to SEQ ID NO:46.
12. The protein according to any of claims 1-7, wherein the first antigen-binding site comprises a heavy chain variable domain at least 90% identical to SEQ ID NO:47 and a light chain variable domain at least 90% identical to SEQ ID NO:48.
13. The protein according to any of claims 1-7, wherein the first antigen-binding site comprises a heavy chain variable domain at least 90% identical to SEQ ID NO:94 and a light chain variable domain at least 90% identical to SEQ ID NO:95.
14. The protein according to any of claims 1-7, wherein the first antigen-binding site comprises a heavy chain variable domain at least 90% identical to SEQ ID NO:102 and a light chain variable domain at least 90% identical to SEQ ID NO:103.
15. The protein of claim 1 or 2, wherein the first antigen-binding site is a single-domain antibody.
16. The protein of claim 15, wherein the single-domain antibody is a V_HH fragment or a V_{NAR} fragment.
17. The protein according to any one of claims 1-2 or 15-16, wherein the second antigen-binding site comprises a heavy chain variable domain and a light chain variable domain.
18. The protein according to claim 17, wherein the heavy chain variable domain and the light chain variable domain of the second antigen-binding site are present on the same polypeptide.
19. A protein according to any of the preceding claims, wherein the heavy chain variable domain of the second antigen-binding site comprises an amino acid sequence at least 90% identical to SEQ ID NO:49 and the light chain variable domain of the second antigen-binding site comprises an amino acid sequence at least 90% identical to SEQ ID NO:53.

20. The protein according to any of the preceding claims, wherein the heavy chain variable domain of the second antigen-binding site comprises an amino acid sequence including:

a heavy chain CDR1 sequence identical to the amino acid sequence of SEQ ID NO:50;

a heavy chain CDR2 sequence identical to the amino acid sequence of SEQ ID NO:51; and

a heavy chain CDR3 sequence identical to the amino acid sequence of SEQ ID NO:52.

21. The protein according to claim 20, wherein the light chain variable domain of the second antigen-binding site comprises an amino acid sequence including:

a light chain CDR1 sequence identical to the amino acid sequence of SEQ ID NO:54;

a light chain CDR2 sequence identical to the amino acid sequence of SEQ ID NO:55;
and

a light chain CDR3 sequence identical to the amino acid sequence of SEQ ID NO:56.

22. A protein according to any one of claims 1-18, wherein the heavy chain variable domain of the second antigen-binding site comprises an amino acid sequence at least 90% identical to SEQ ID NO:57 and the light chain variable domain of the second antigen-binding site comprises an amino acid sequence at least 90% identical to SEQ ID NO:58.

23. The protein according to any one of claims 1-18 or 22, wherein the heavy chain variable domain of the second antigen-binding site comprises an amino acid sequence including:

a heavy chain CDR1 sequence identical to the amino acid sequence of SEQ ID NO:77;

a heavy chain CDR2 sequence identical to the amino acid sequence of SEQ ID NO:78; and

a heavy chain CDR3 sequence identical to the amino acid sequence of SEQ ID NO:79.

24. The protein according to claim 23, wherein the light chain variable domain of the second antigen-binding site comprises an amino acid sequence including:

a light chain CDR1 sequence identical to the amino acid sequence of SEQ ID NO:80;

a light chain CDR2 sequence identical to the amino acid sequence of SEQ ID NO:81;

and

a light chain CDR3 sequence identical to the amino acid sequence of SEQ ID NO:82.

25. A protein according to any one of claims 1-18, wherein the heavy chain variable domain of the second antigen-binding site comprises an amino acid sequence at least 90% identical to SEQ ID NO:59 and the light chain variable domain of the second antigen-binding site comprises an amino acid sequence at least 90% identical to SEQ ID NO:60.

26. The protein according to any one of claims 1-18 or 25, wherein the heavy chain variable domain of the second antigen-binding site comprises an amino acid sequence including:

a heavy chain CDR1 sequence identical to the amino acid sequence of SEQ ID NO:83;

a heavy chain CDR2 sequence identical to the amino acid sequence of SEQ ID NO:84; and

a heavy chain CDR3 sequence identical to the amino acid sequence of SEQ ID NO:85.

27. The protein according to any one of claim 26, wherein the light chain variable domain of the second antigen-binding site comprises an amino acid sequence including:

a light chain CDR1 sequence identical to the amino acid sequence of SEQ ID NO:86;

a light chain CDR2 sequence identical to the amino acid sequence of SEQ ID NO:87;

and

a light chain CDR3 sequence identical to the amino acid sequence of SEQ ID NO:88.

28. The protein according to any one of claims 1-4 or 8-16, wherein the second antigen-binding site is a single-domain antibody.
29. The protein of claim 28, wherein the second antigen-binding site is a V_HH fragment or a V_{NAR} fragment.
30. A protein according to any one of the preceding claims, wherein the protein comprises a portion of an antibody Fc domain sufficient to bind CD16, wherein the antibody Fc domain comprises hinge and CH2 domains.
31. The protein according to claim 30, wherein the antibody Fc domain comprises hinge and CH2 domains of a human IgG1 antibody.
32. The protein according to claim 30 or 31, wherein the Fc domain comprises an amino acid sequence at least 90% identical to amino acids 234-332 of a human IgG1 antibody.
33. The protein according to any one of claims 30-32, wherein the Fc domain comprises amino acid sequence at least 90% identical to the Fc domain of human IgG1 and differs at one or more positions selected from the group consisting of Q347, Y349, T350, L351, S354, E356, E357, K360, Q362, S364, T366, L368, K370, N390, K392, T394, D399, S400, D401, F405, Y407, K409, T411, K439.
34. A formulation comprising a protein according to any one of the preceding claims and a pharmaceutically acceptable carrier.
35. A cell comprising one or more nucleic acids expressing a protein according to any one of claims 1-33.
36. A method of directly and/or indirectly enhancing tumor cell death, the method comprising exposing a tumor and natural killer cells to a protein according to any one of claims 1-33.
37. A method of treating cancer, wherein the method comprises administering a protein according to any one of claims 1-33 or a formulation according to claim 34 to a patient.
38. The method of claim 37, wherein the cancer is selected from the group consisting of breast, ovarian, esophageal, bladder and gastric cancer, salivary duct carcinoma,

adenocarcinoma of the lung and aggressive forms of uterine cancer, such as uterine serous endometrial carcinoma.

FIG. 1

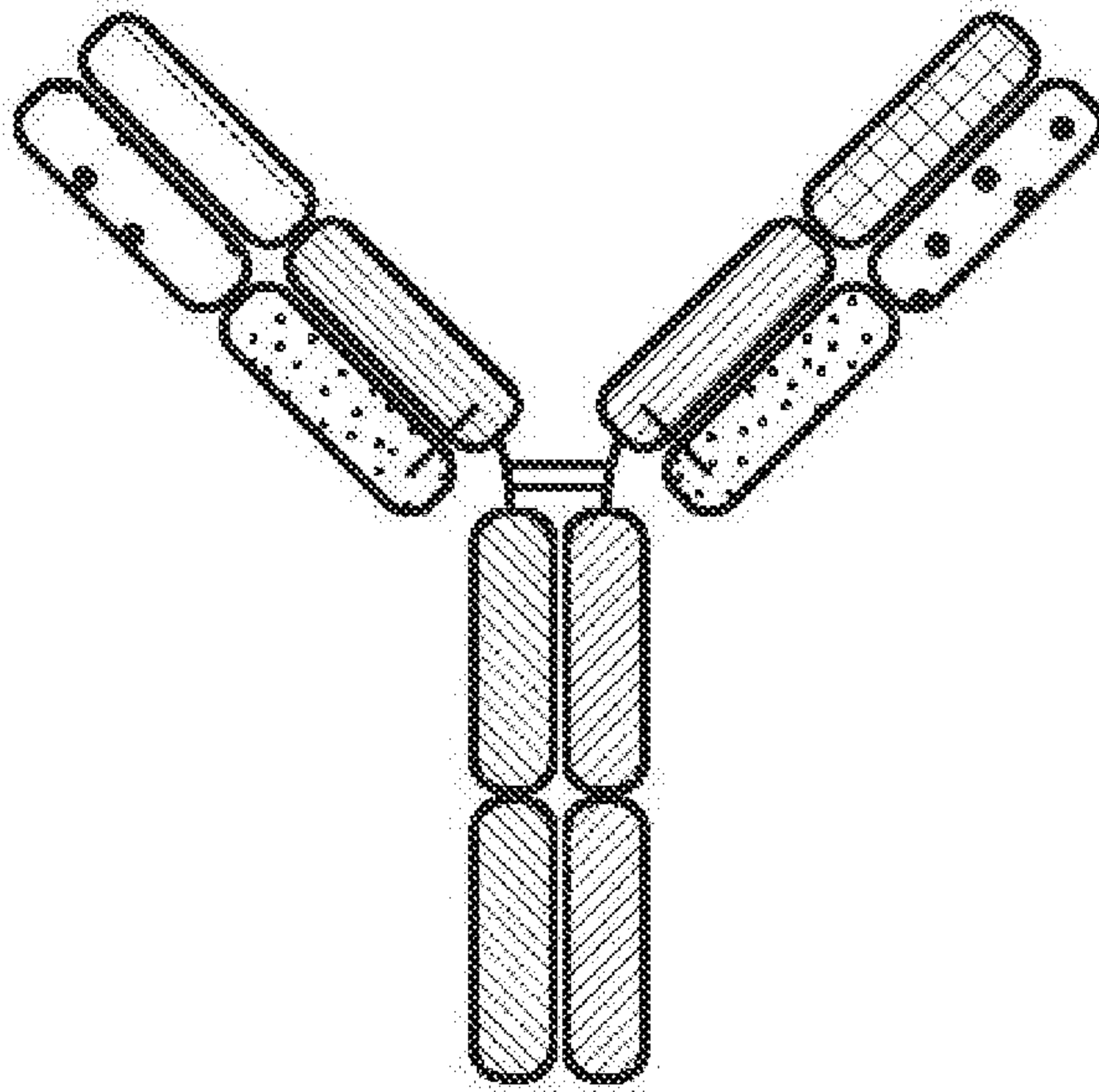


FIG. 2

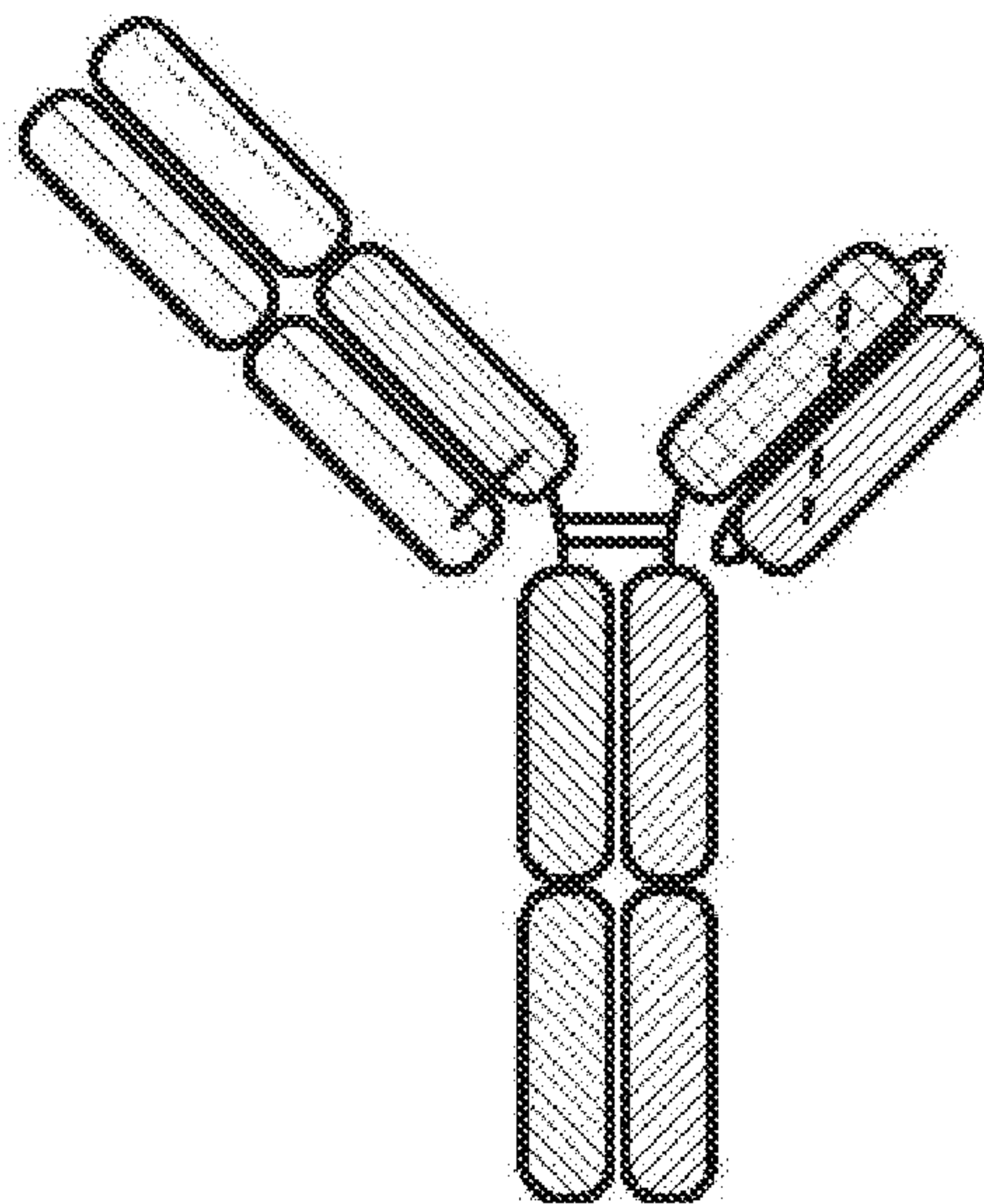


FIG. 3

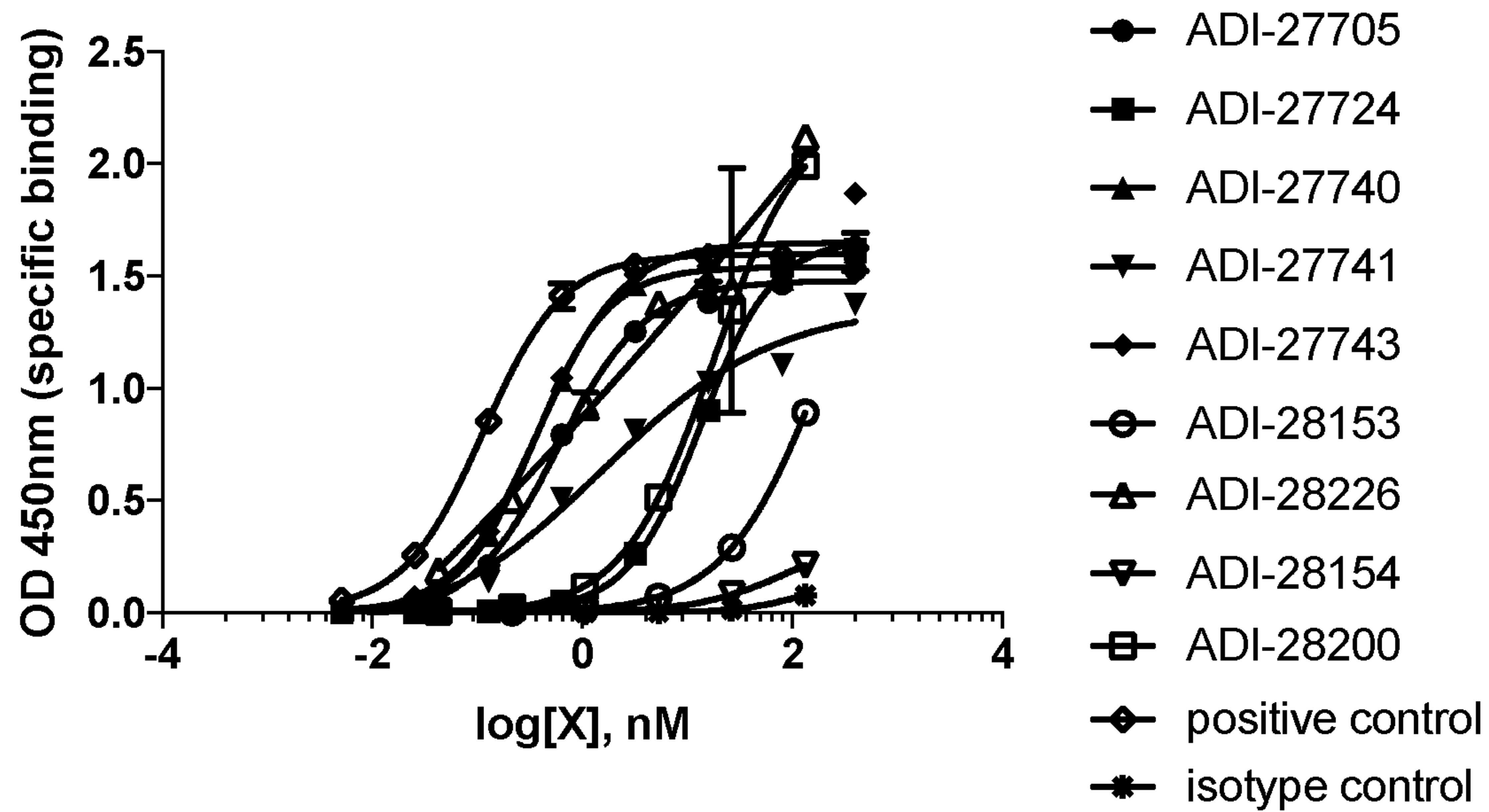


FIG. 4

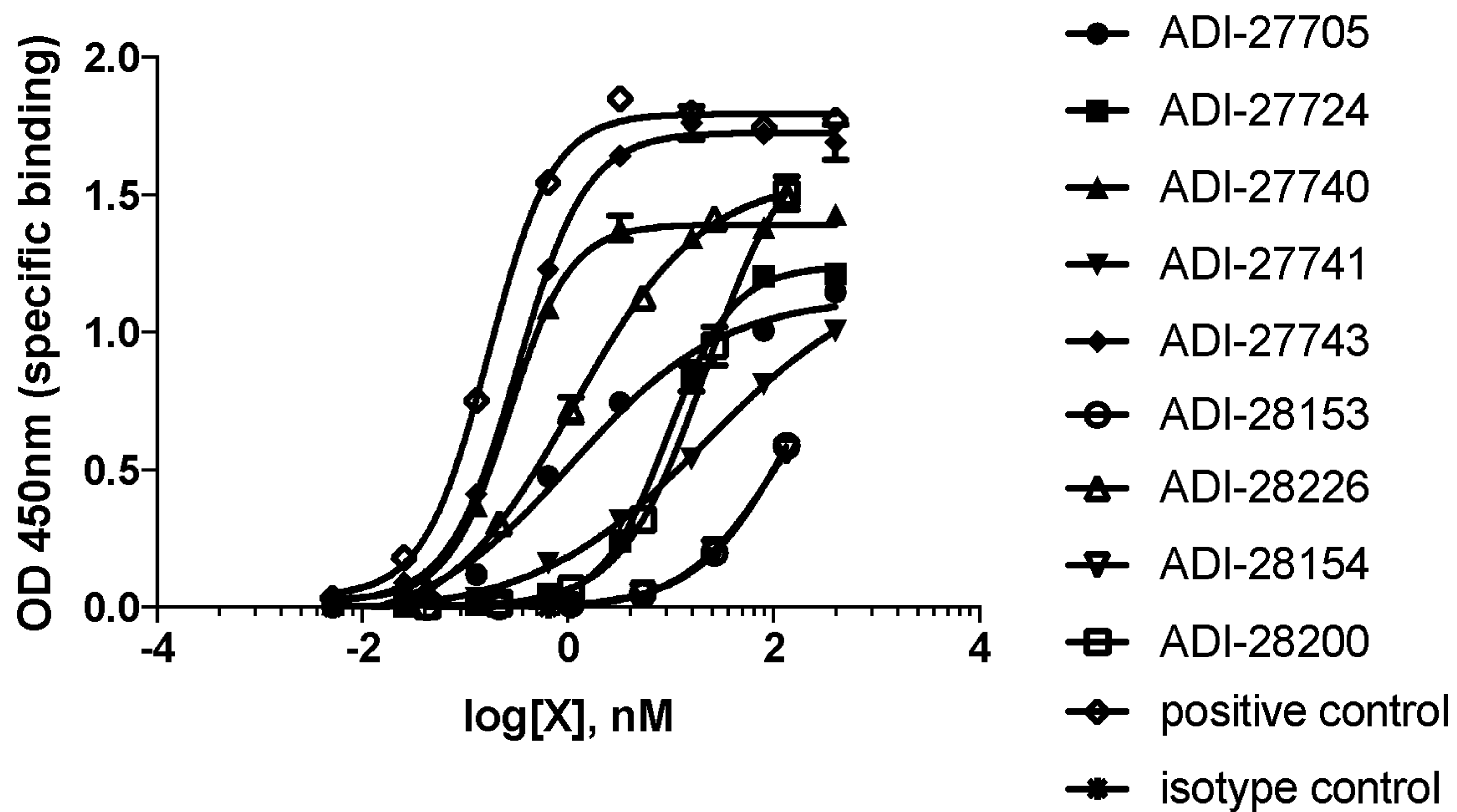


FIG. 5

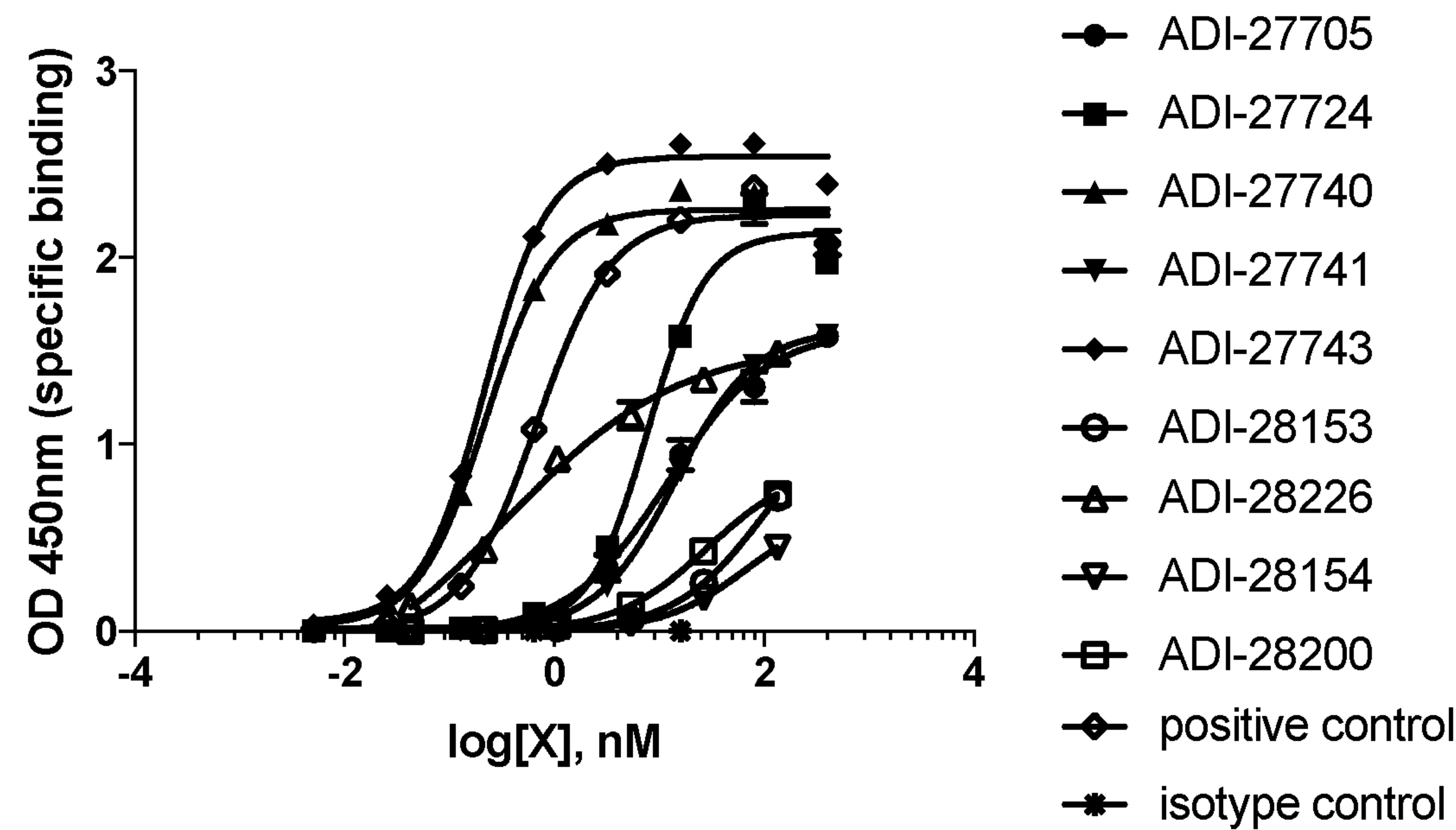


FIG. 6

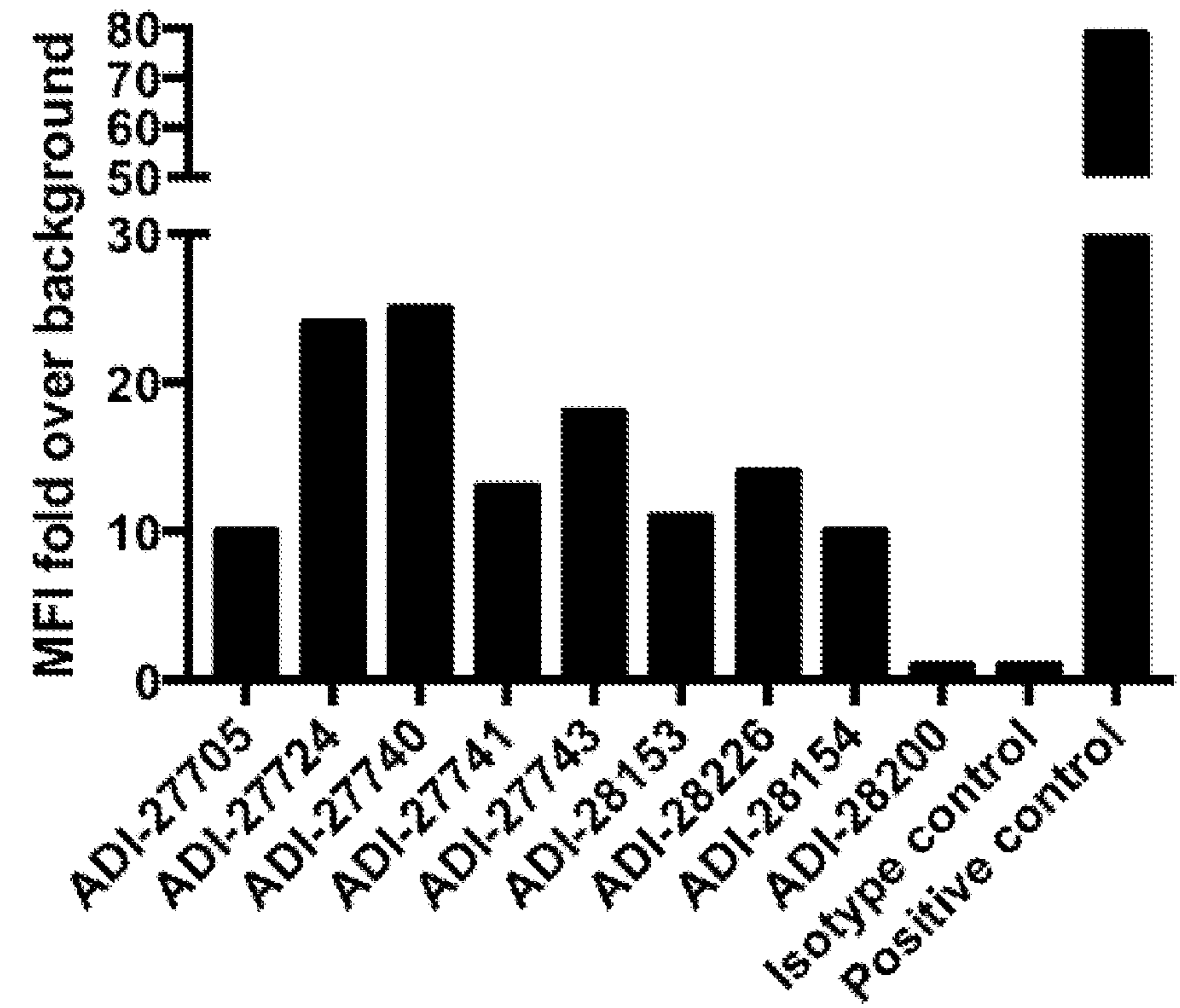


FIG. 7

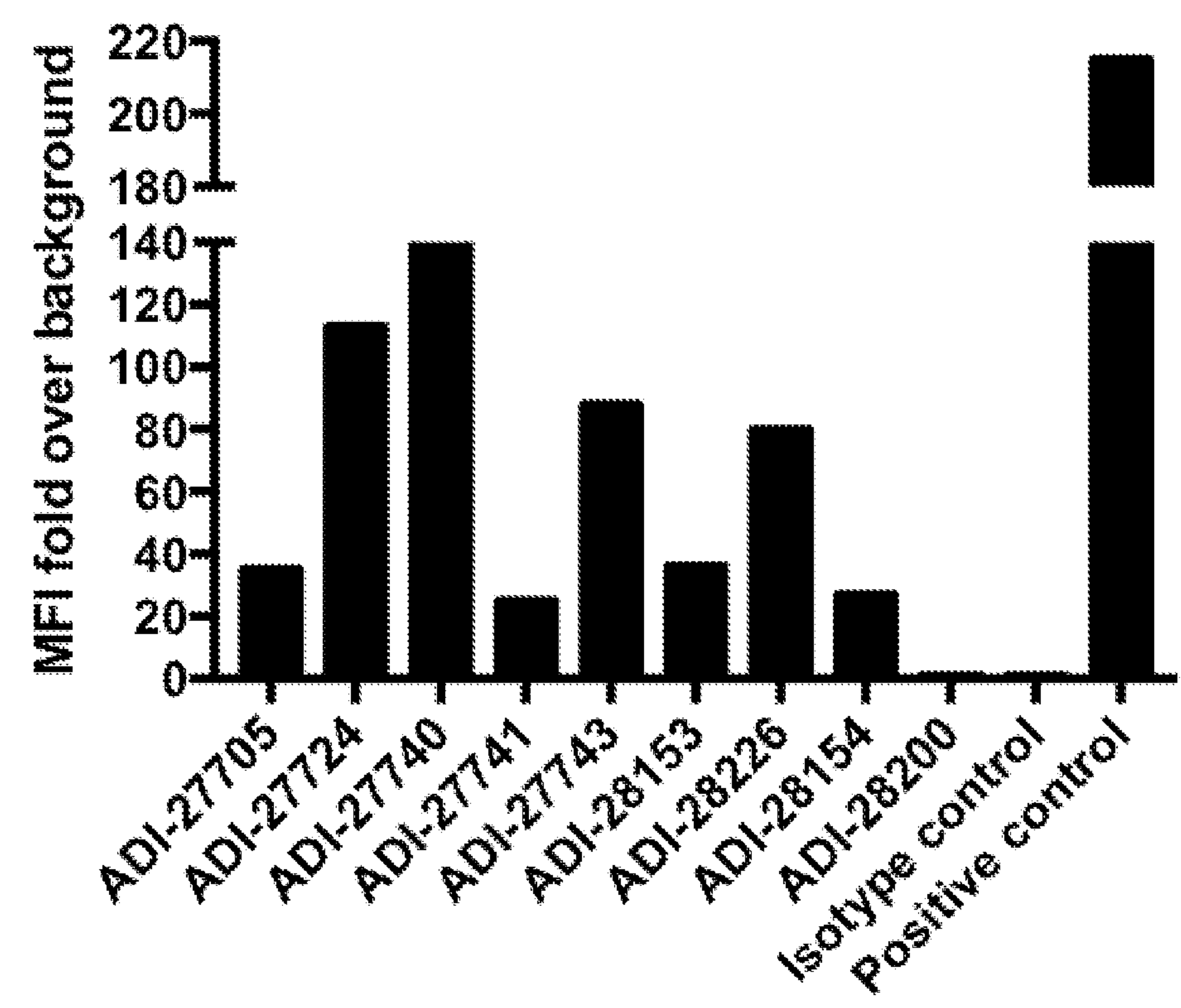


FIG. 8

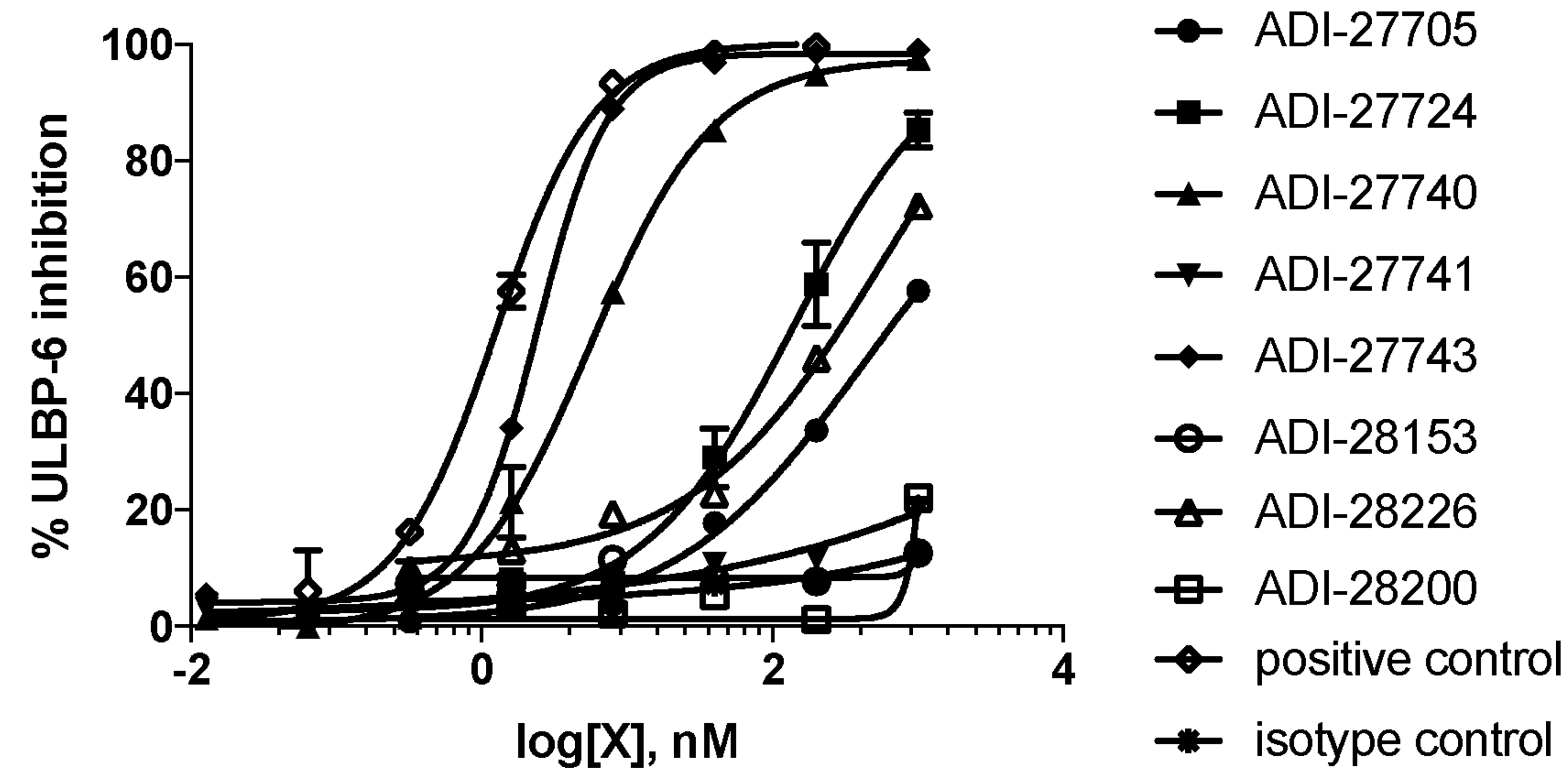


FIG. 9

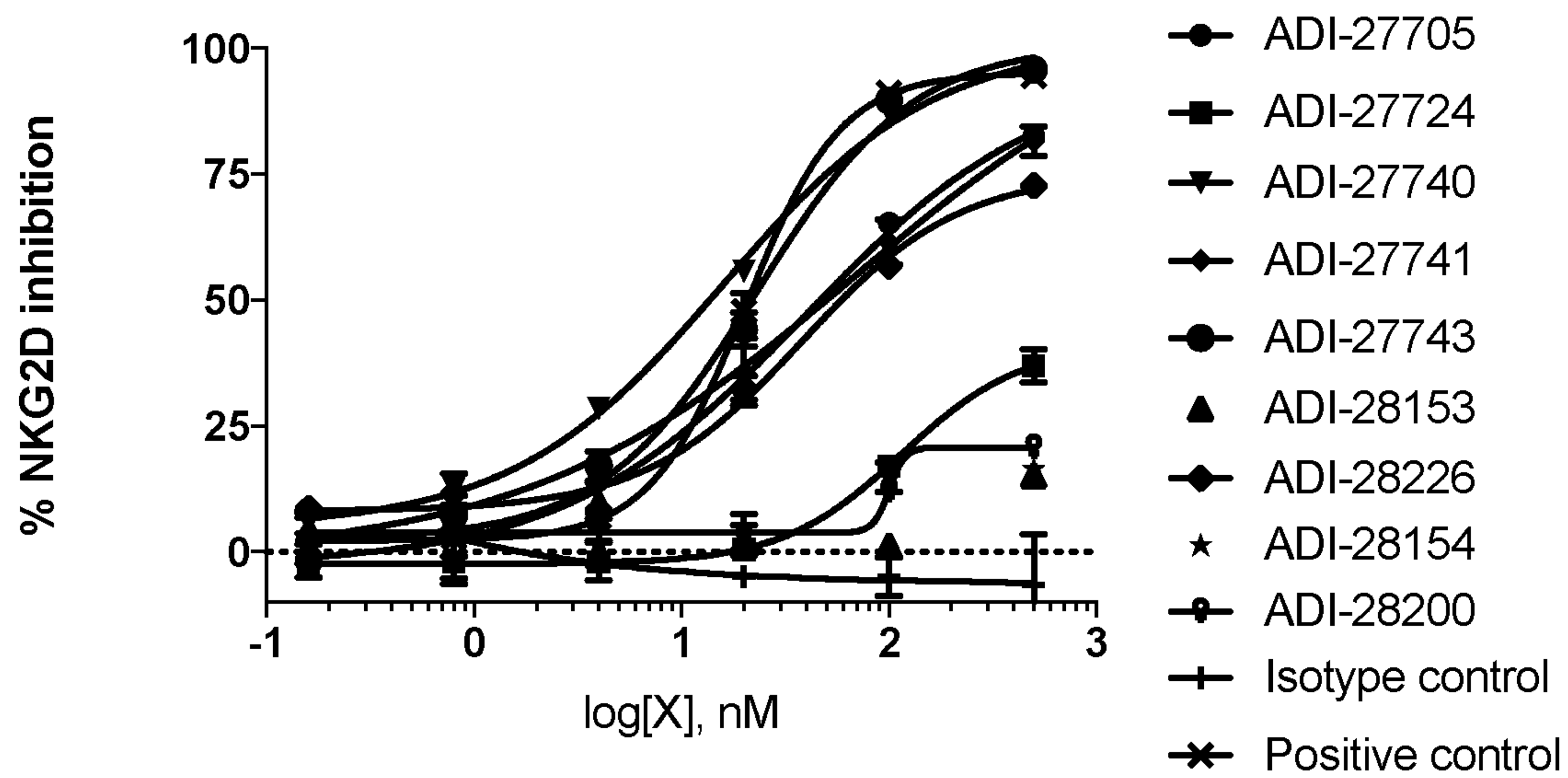


FIG. 10

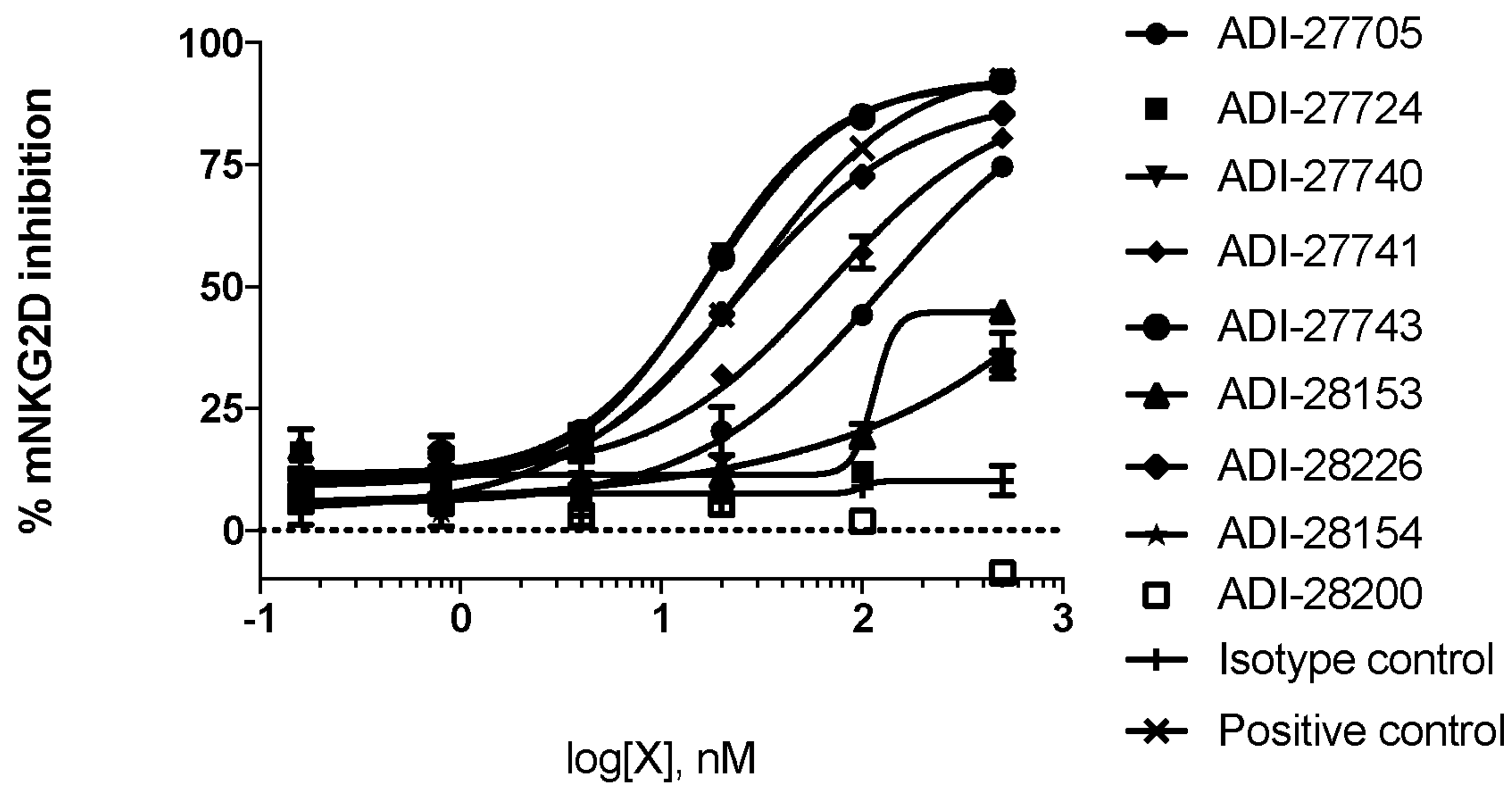


FIG. 11

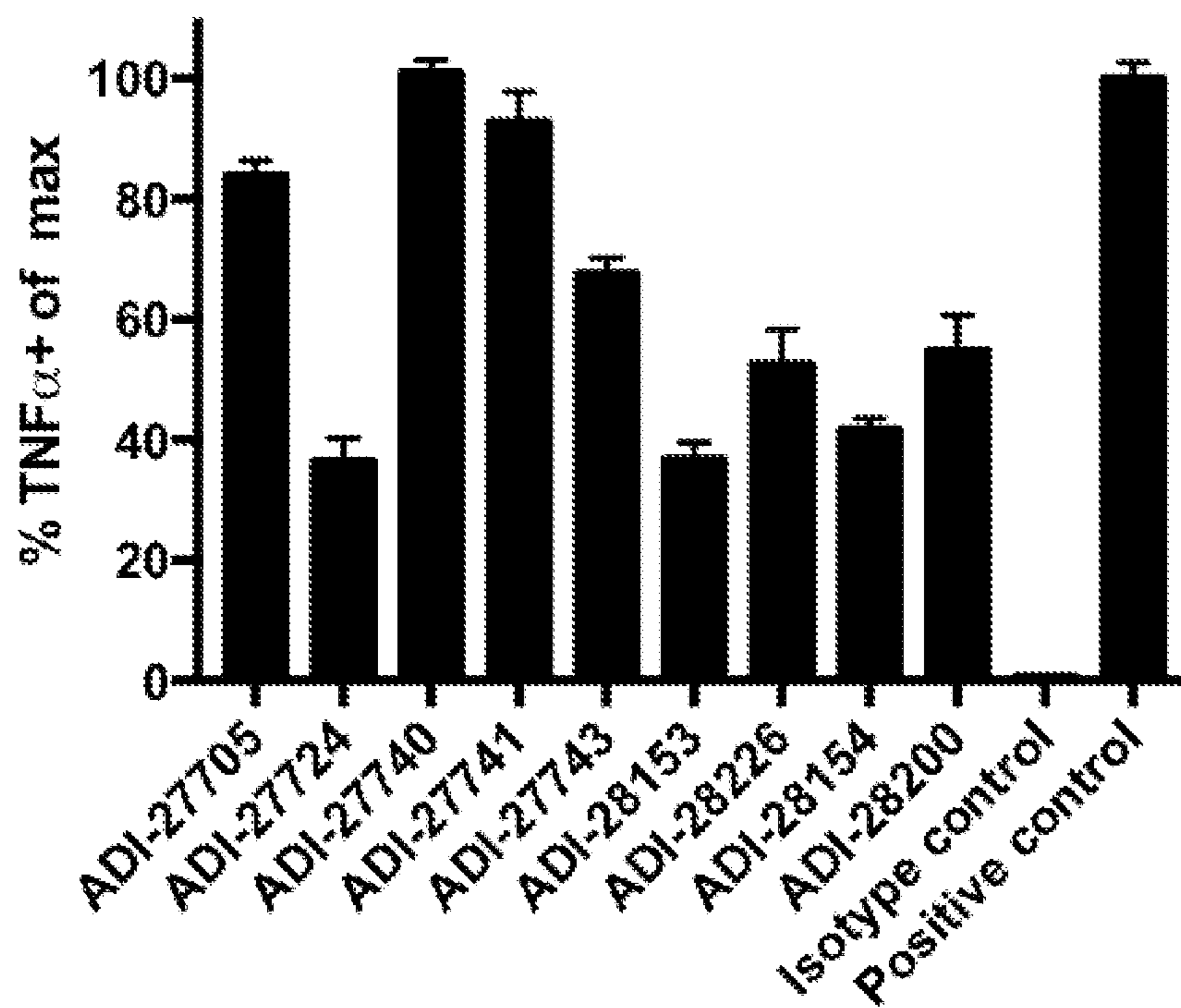


FIG. 12

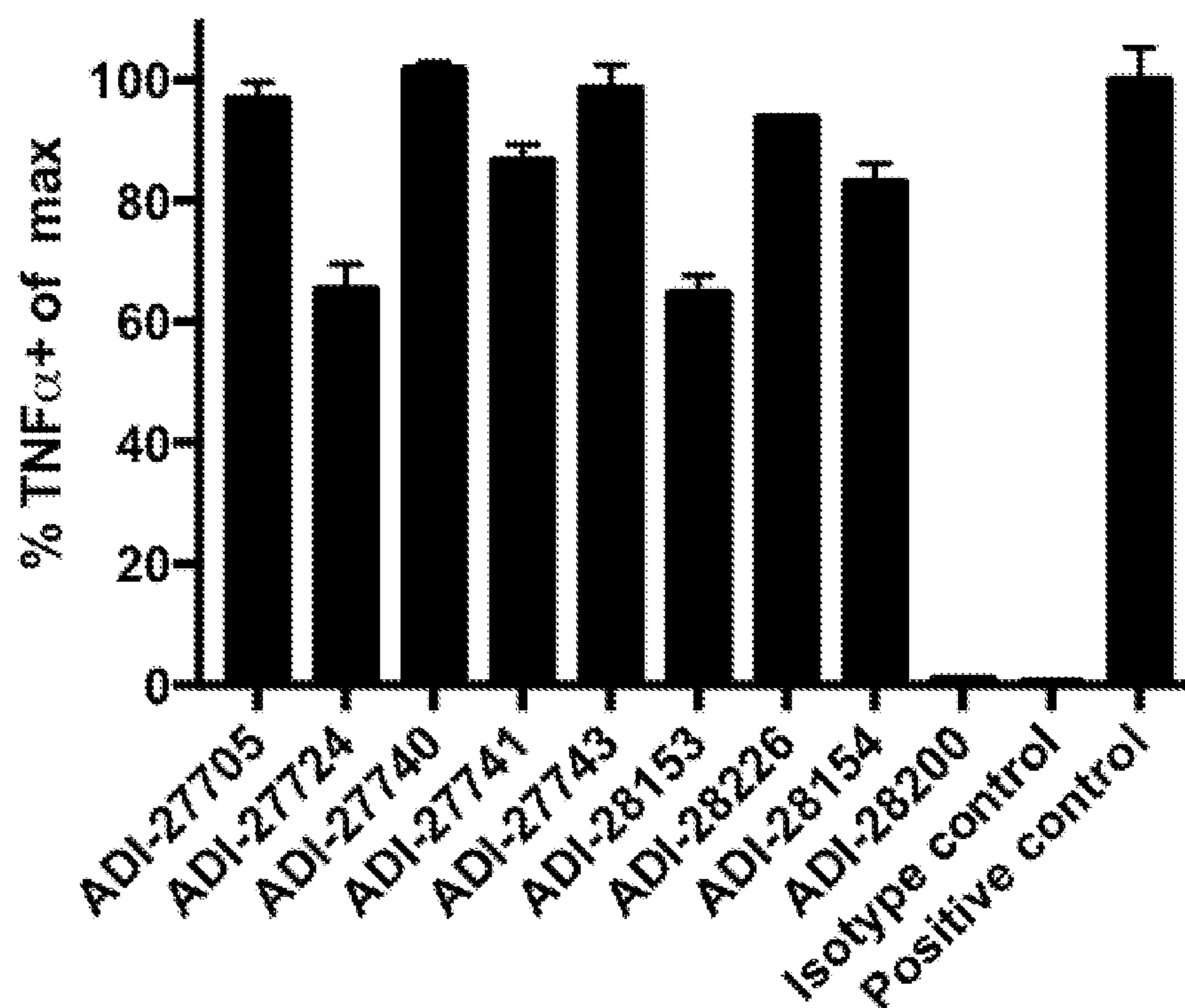


FIG. 13

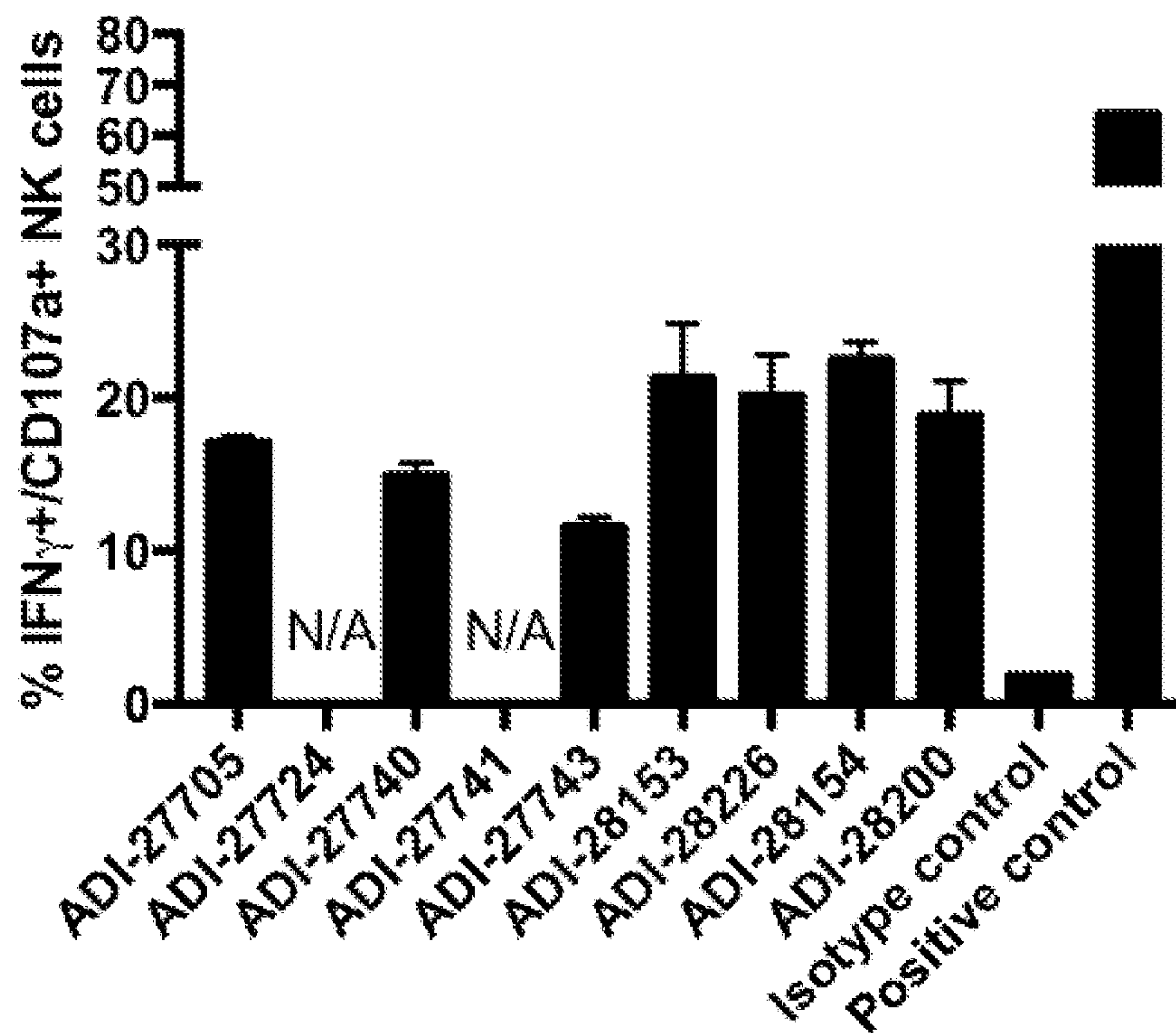


FIG. 14

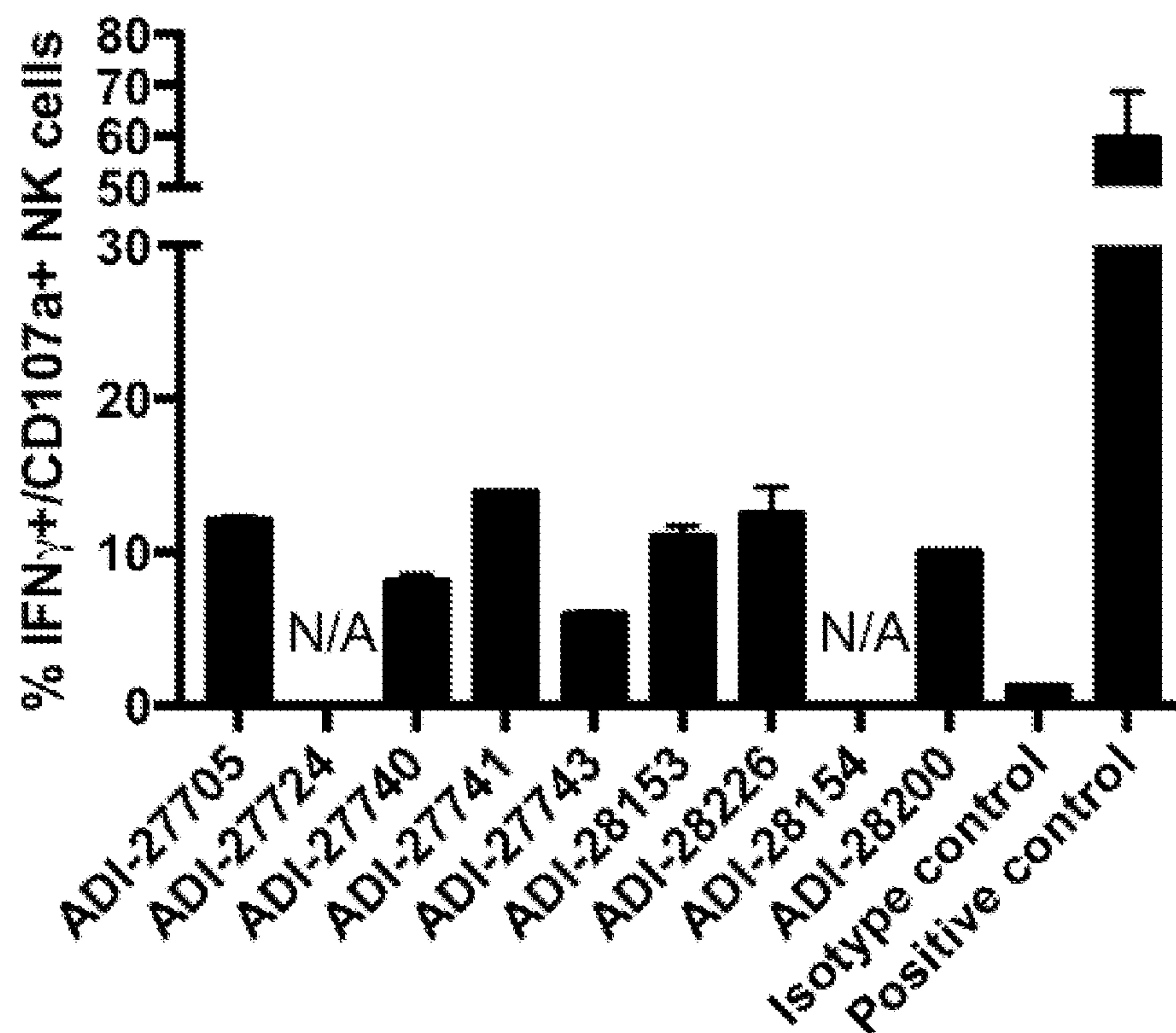


FIG. 15

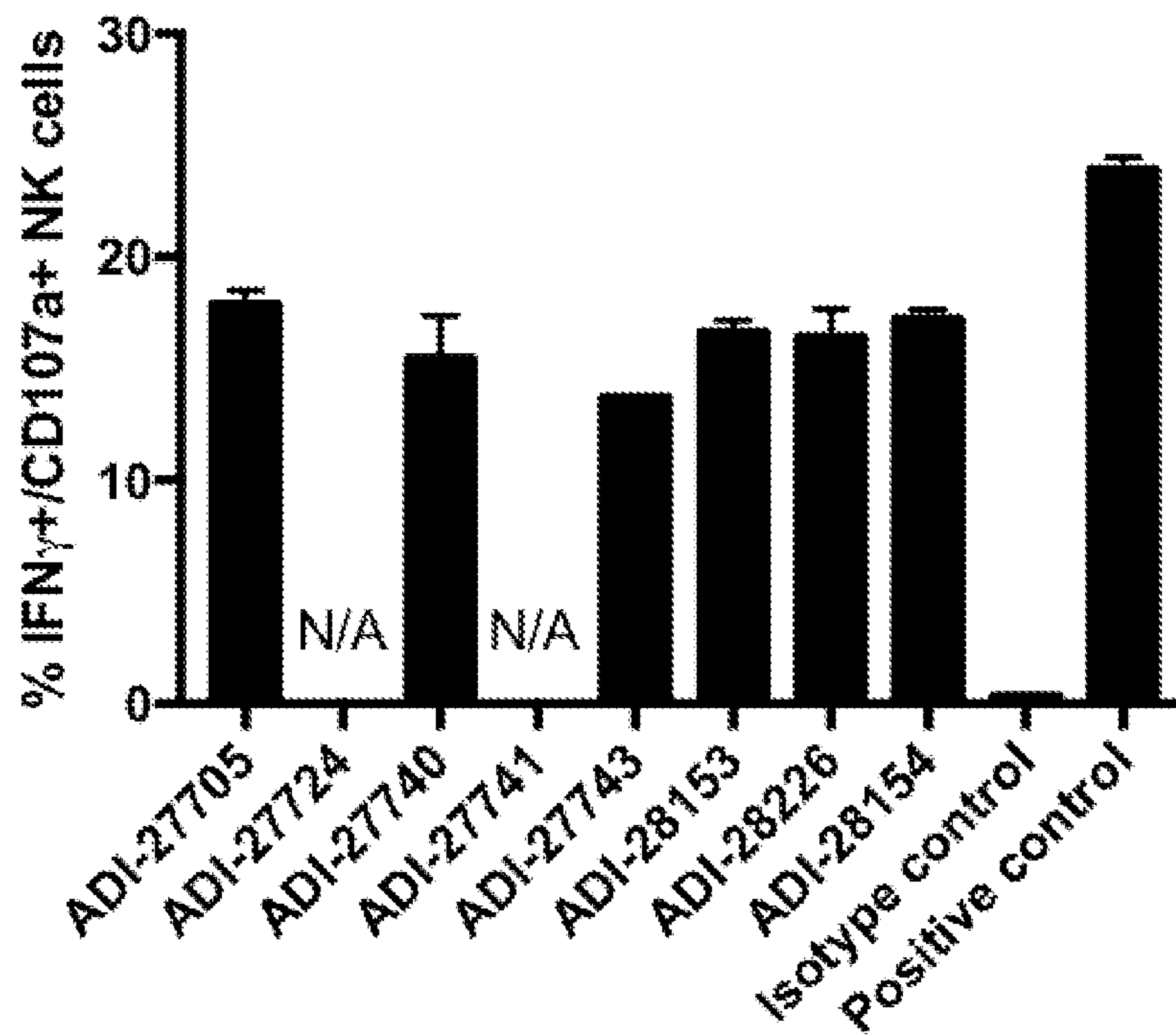


FIG. 16

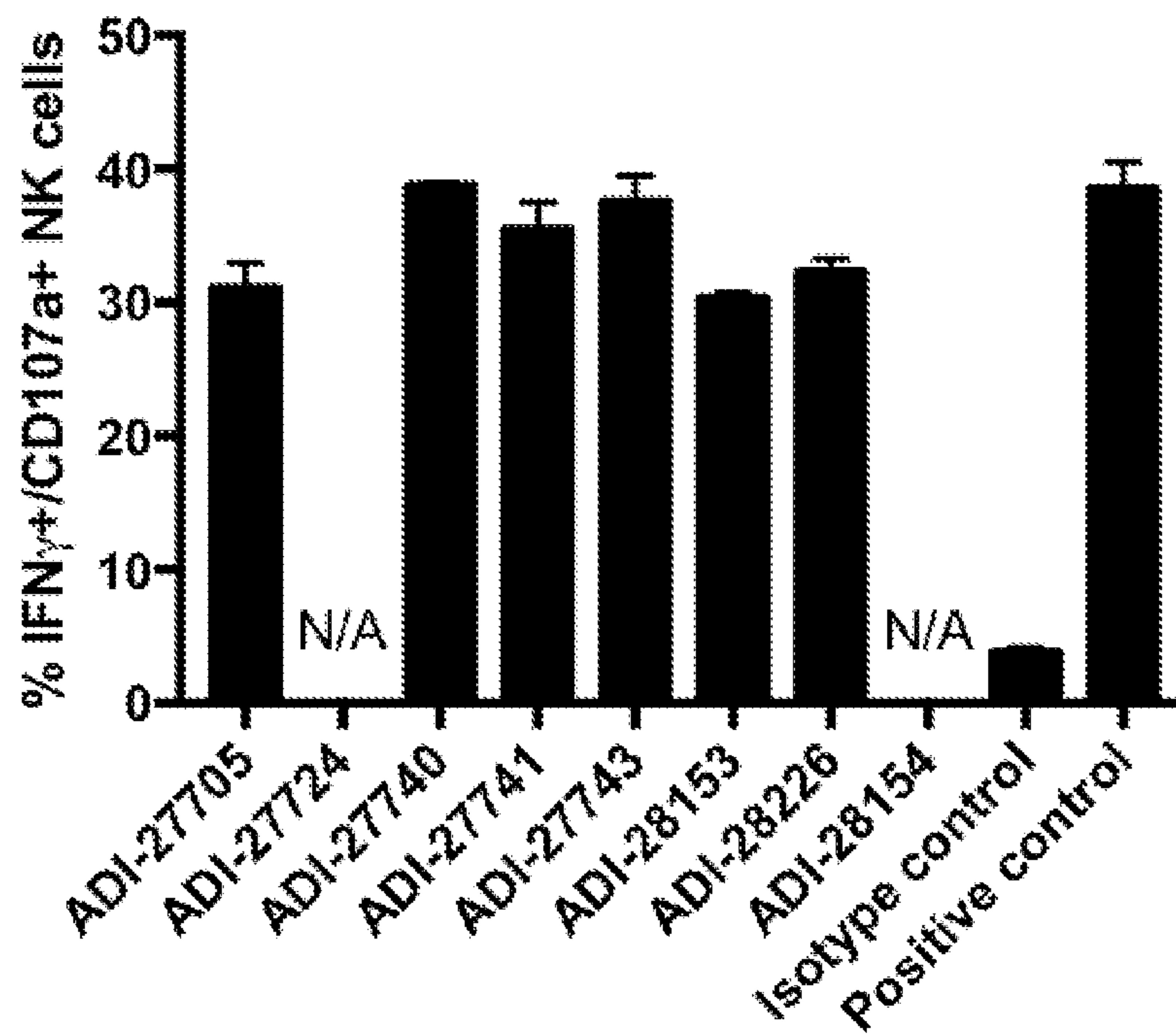


FIG. 17

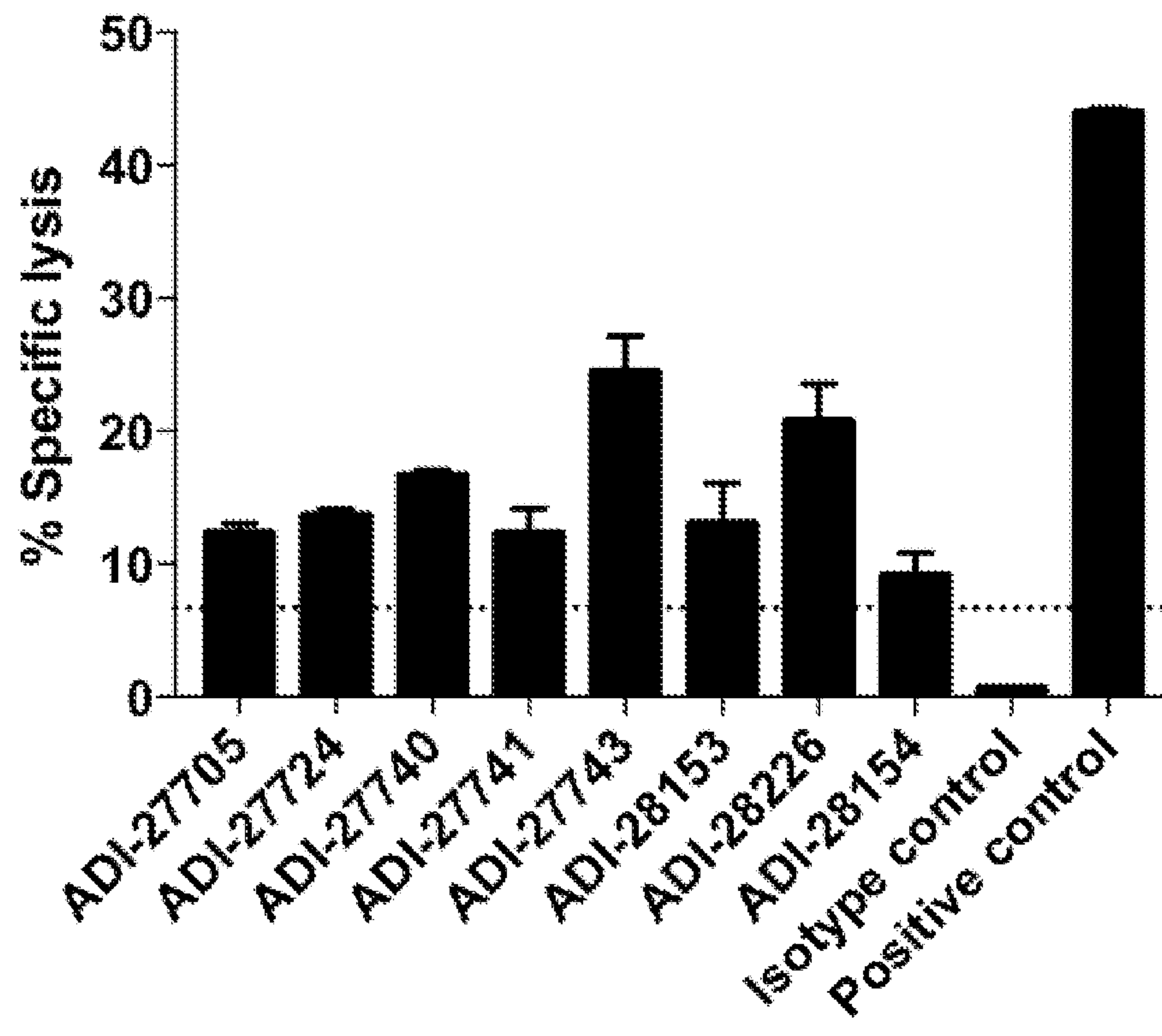


FIG. 18

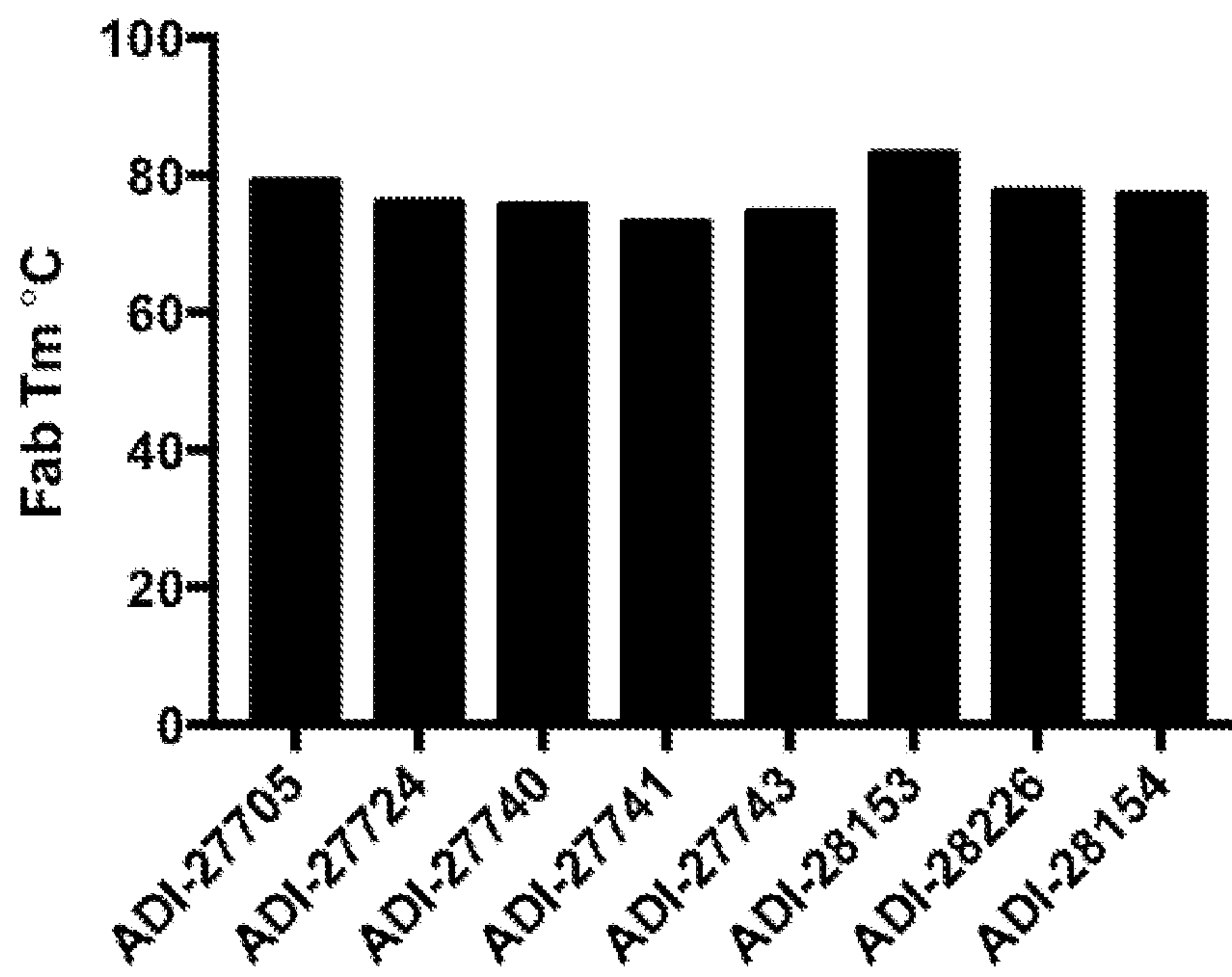


FIG. 19

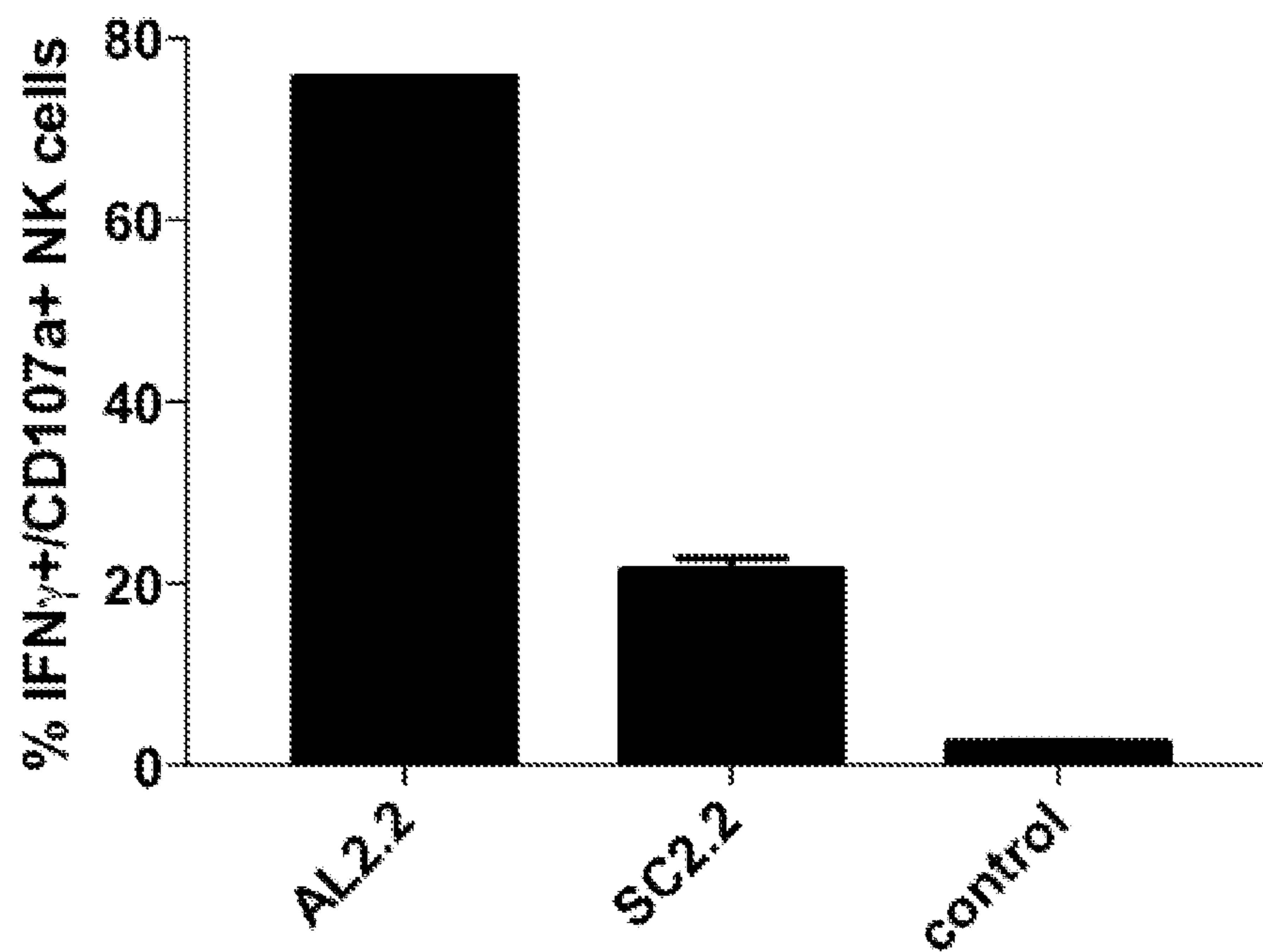


FIG. 20

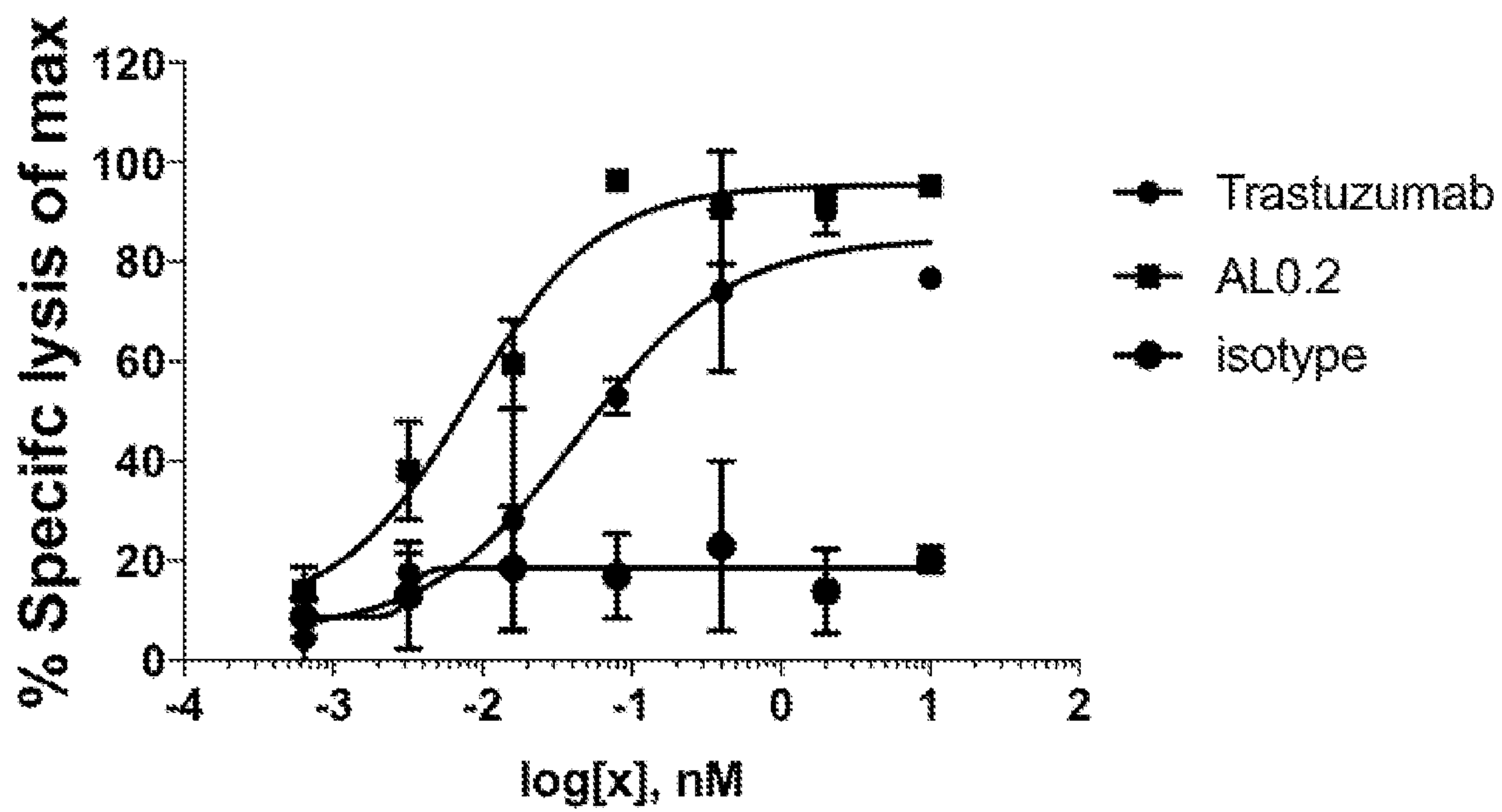


FIG. 21

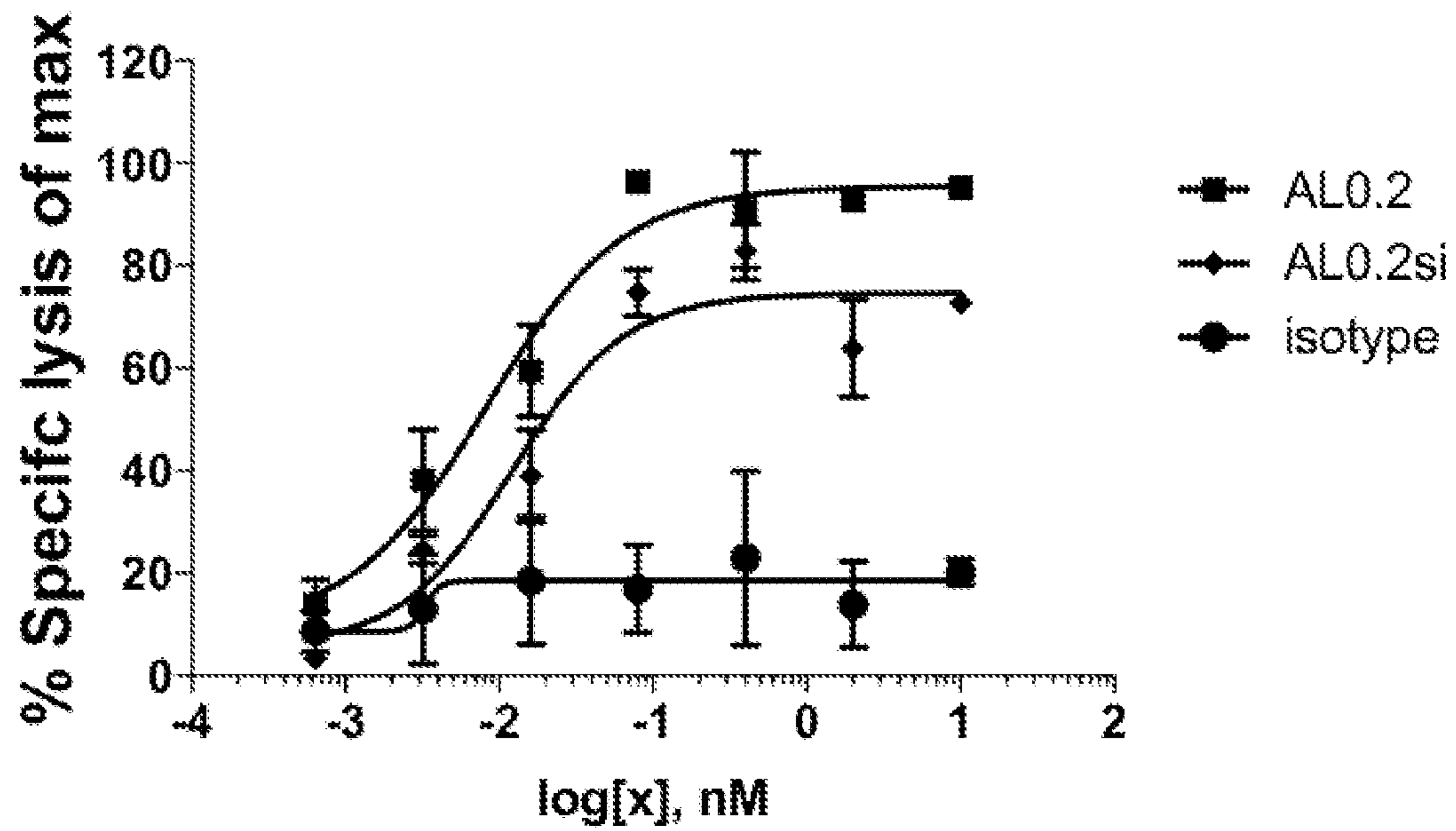


FIG. 22

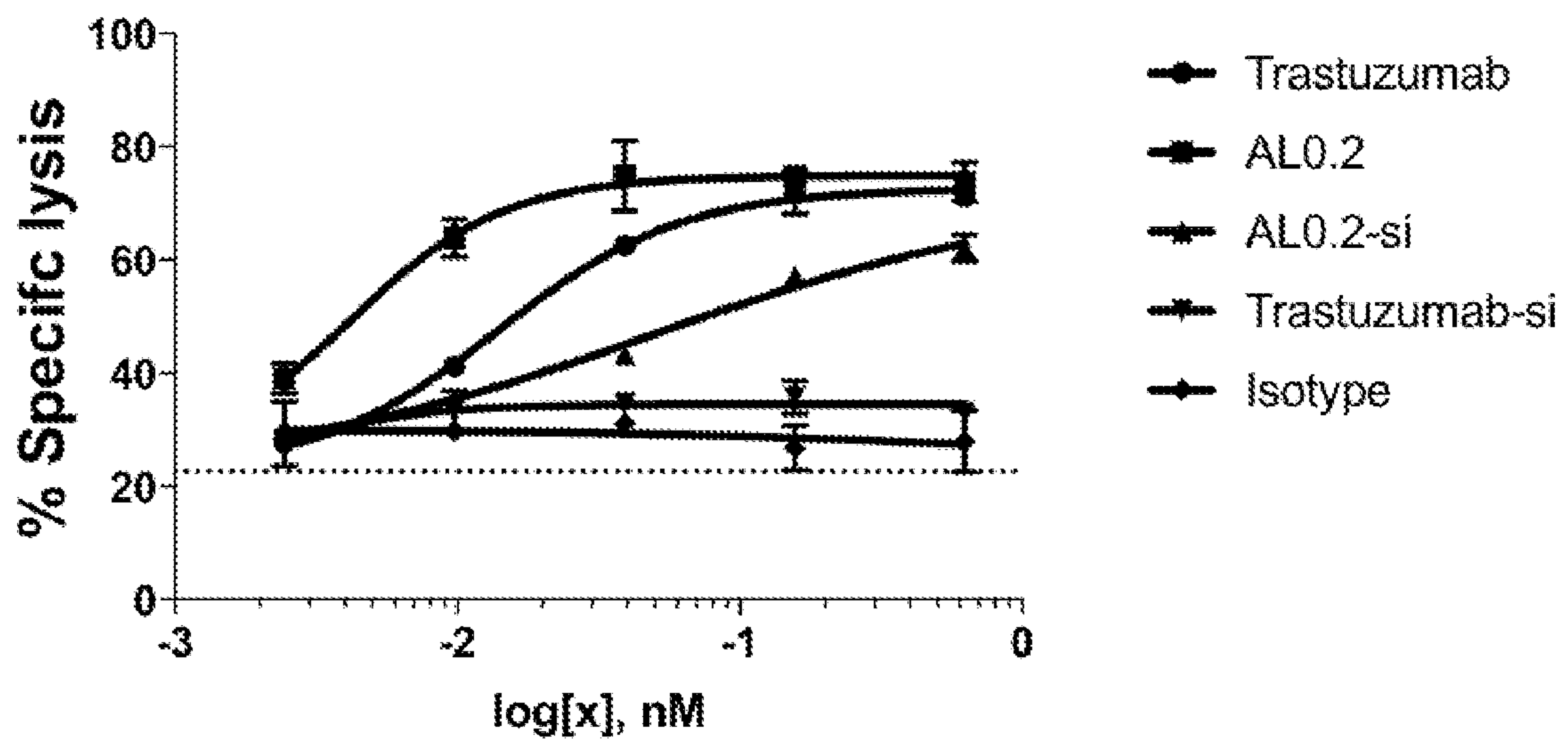


FIG. 23

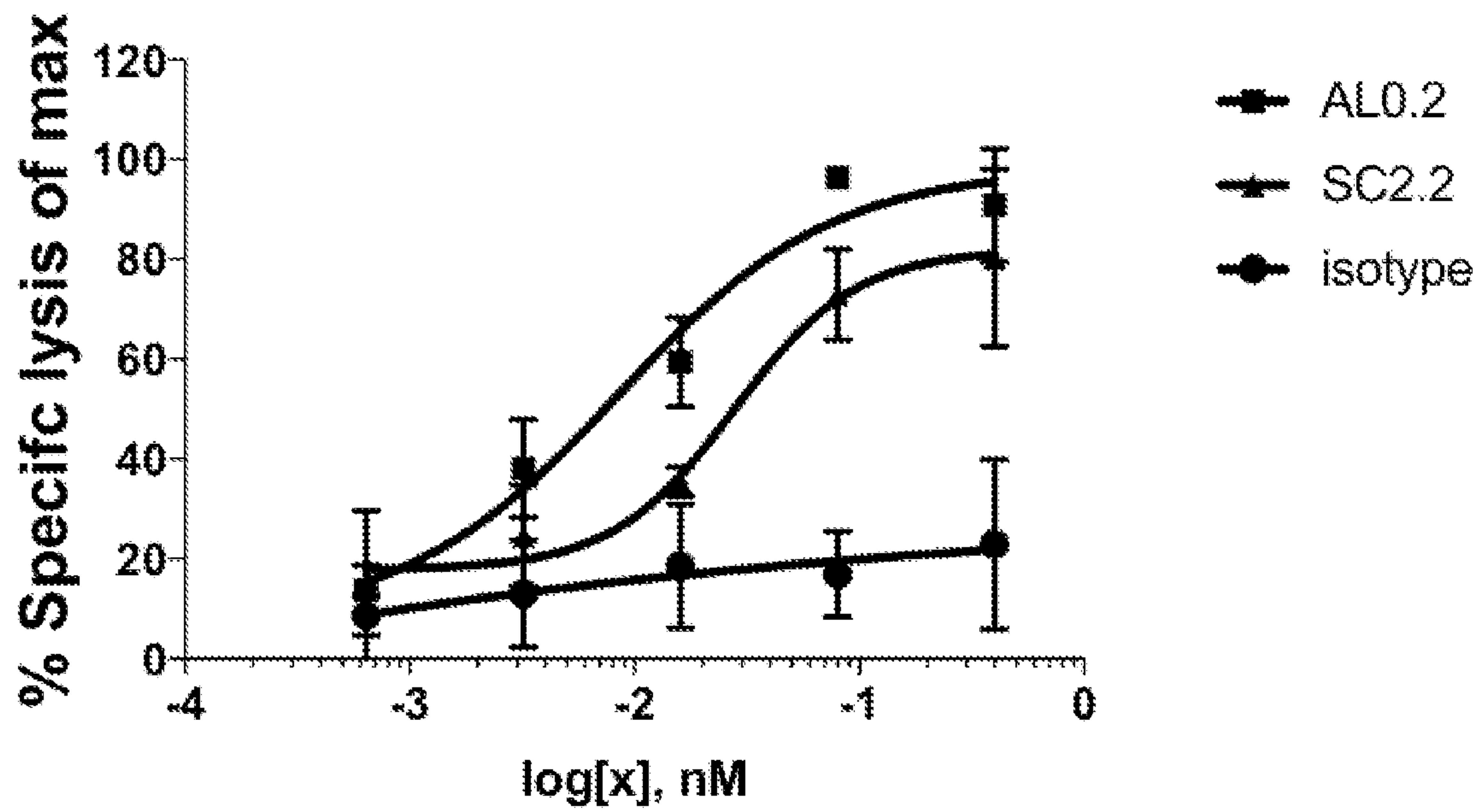


FIG. 24

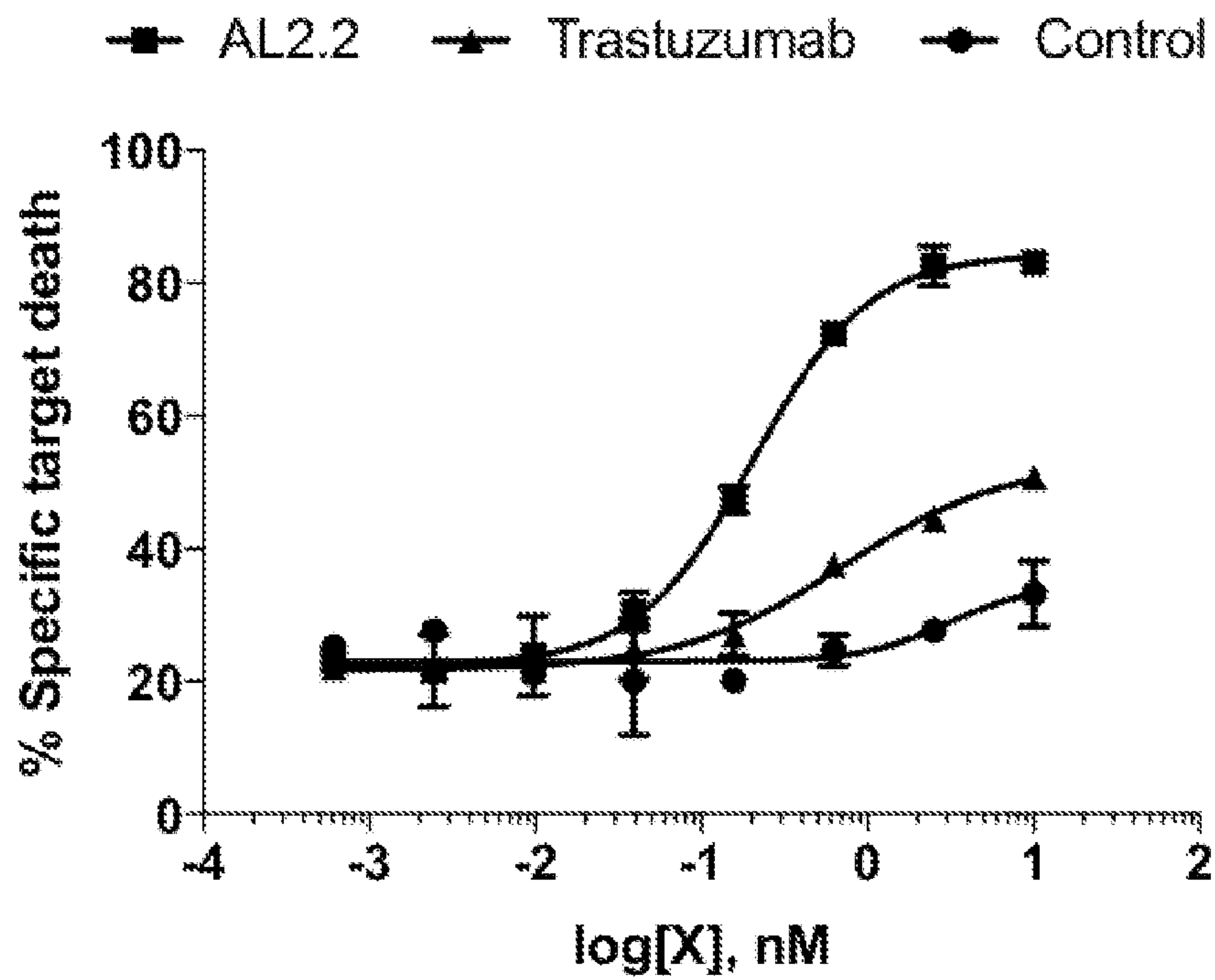


FIG. 25

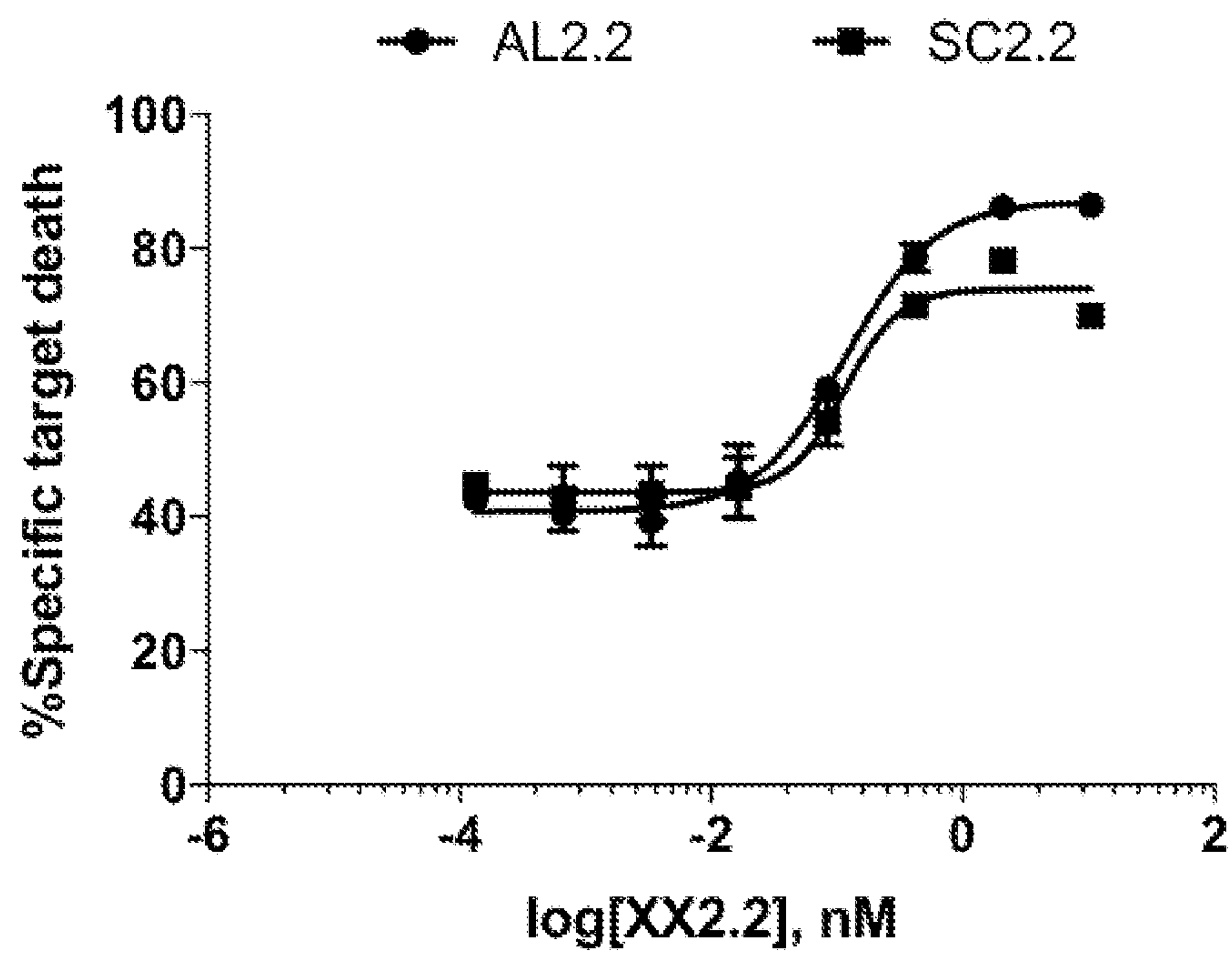


FIG. 26

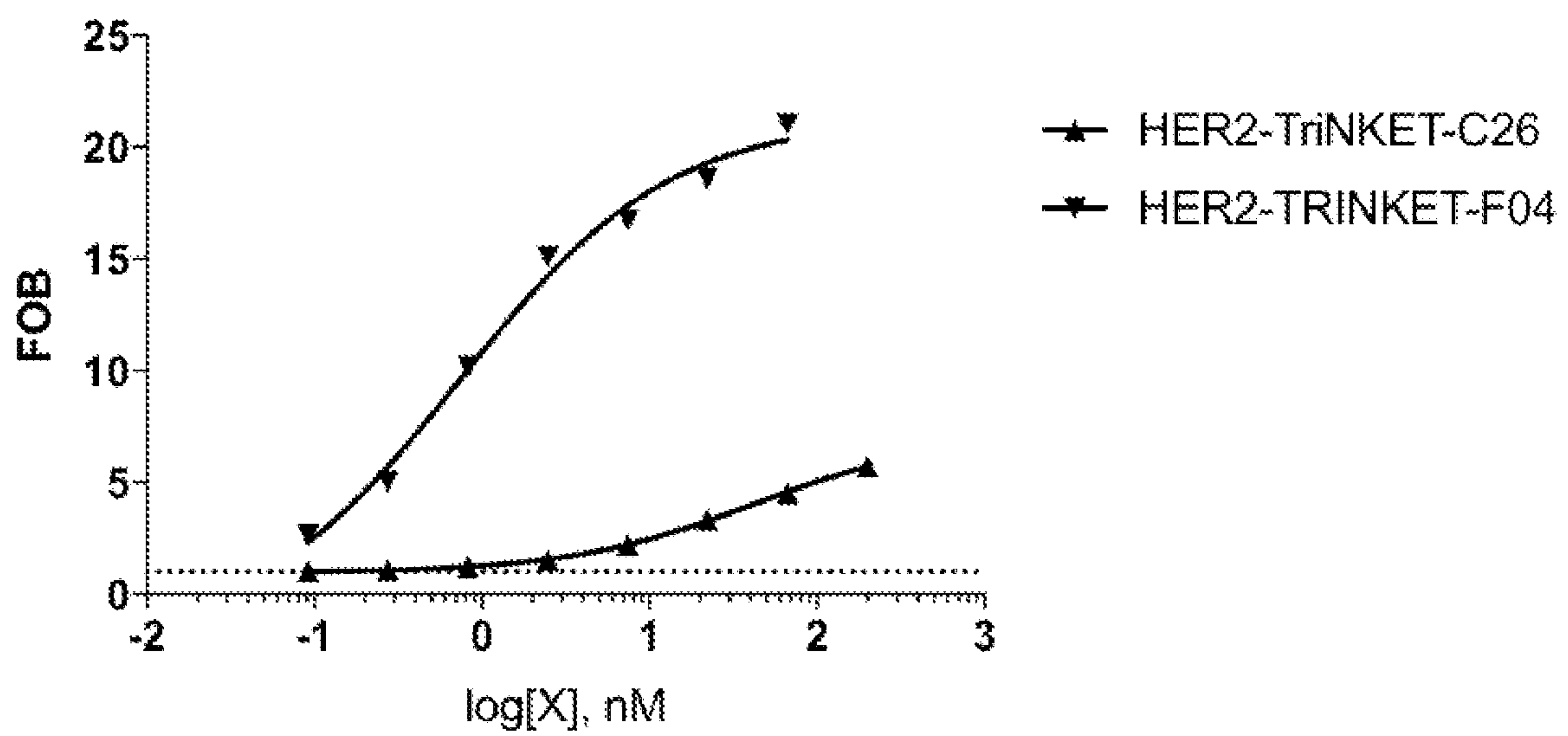


FIG. 27A

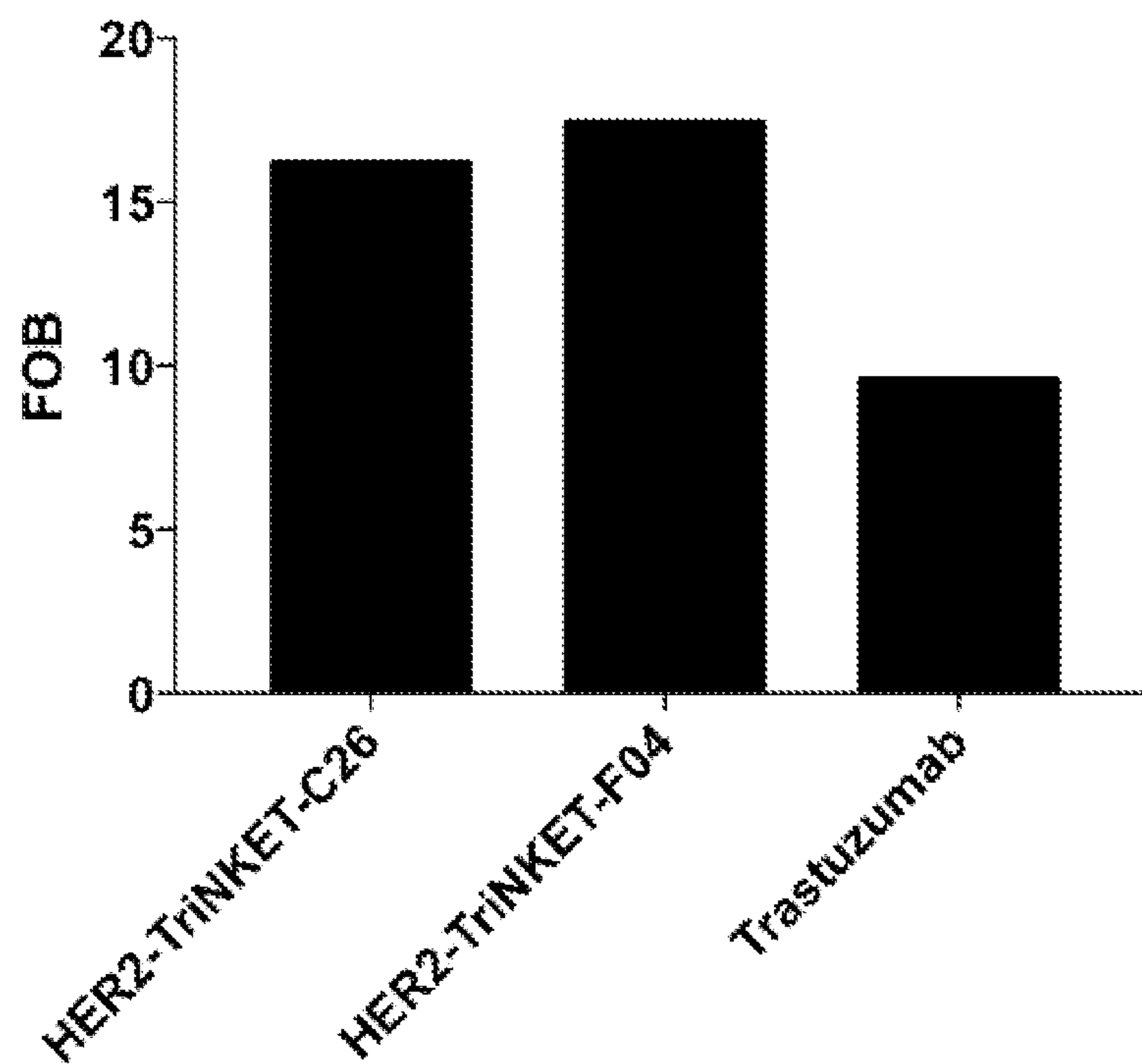


FIG. 27B

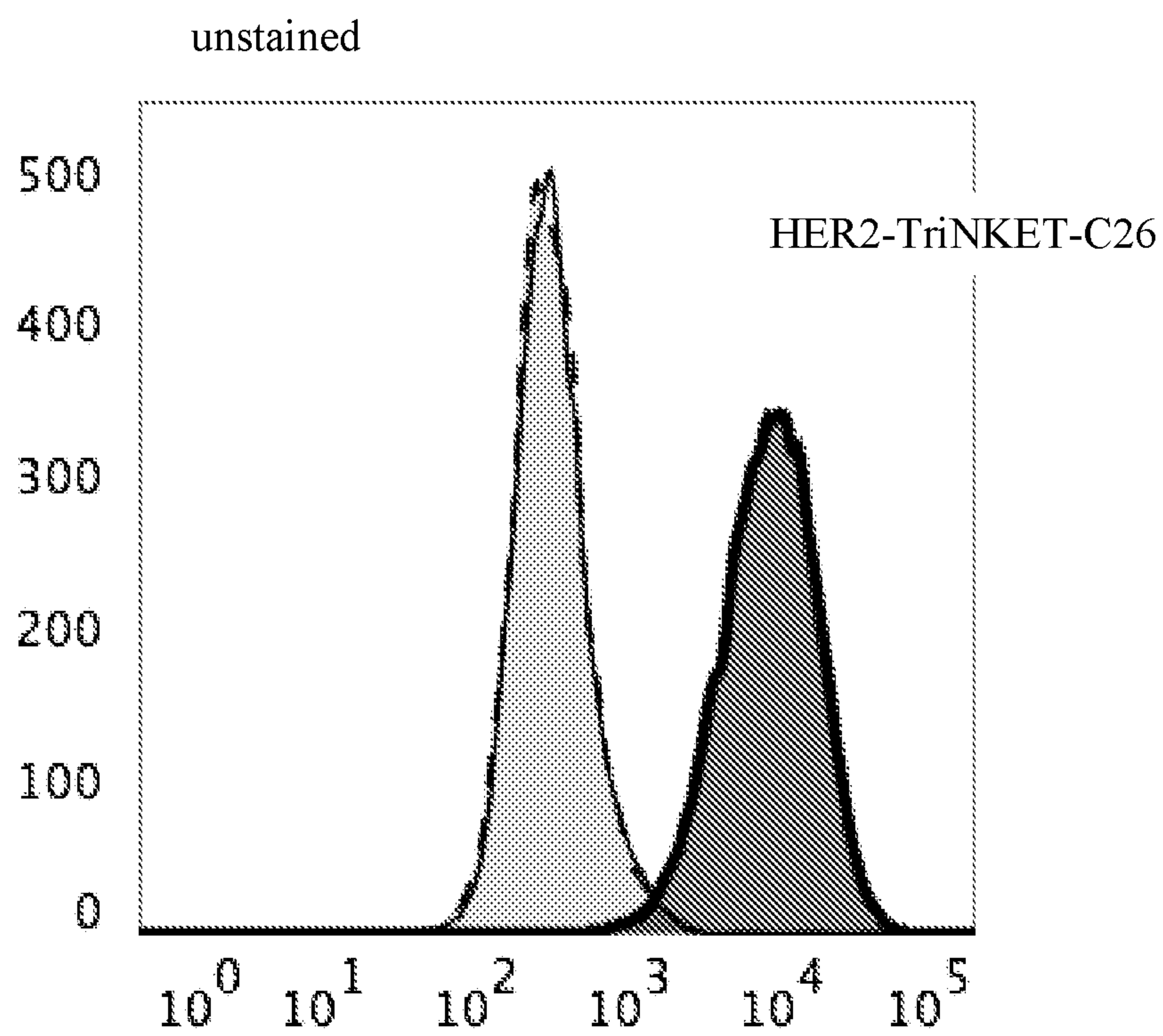


FIG. 27C

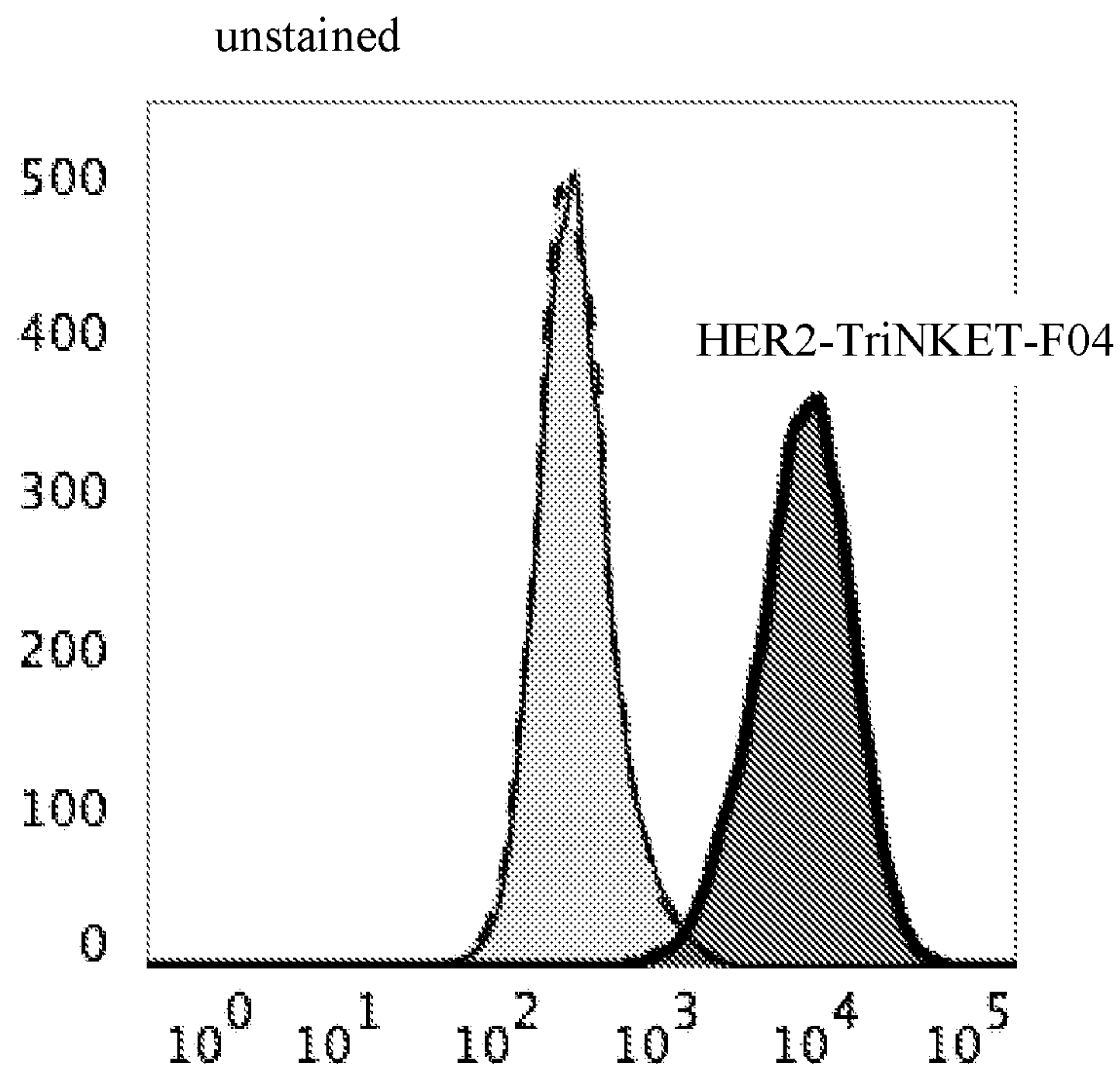


FIG. 28A

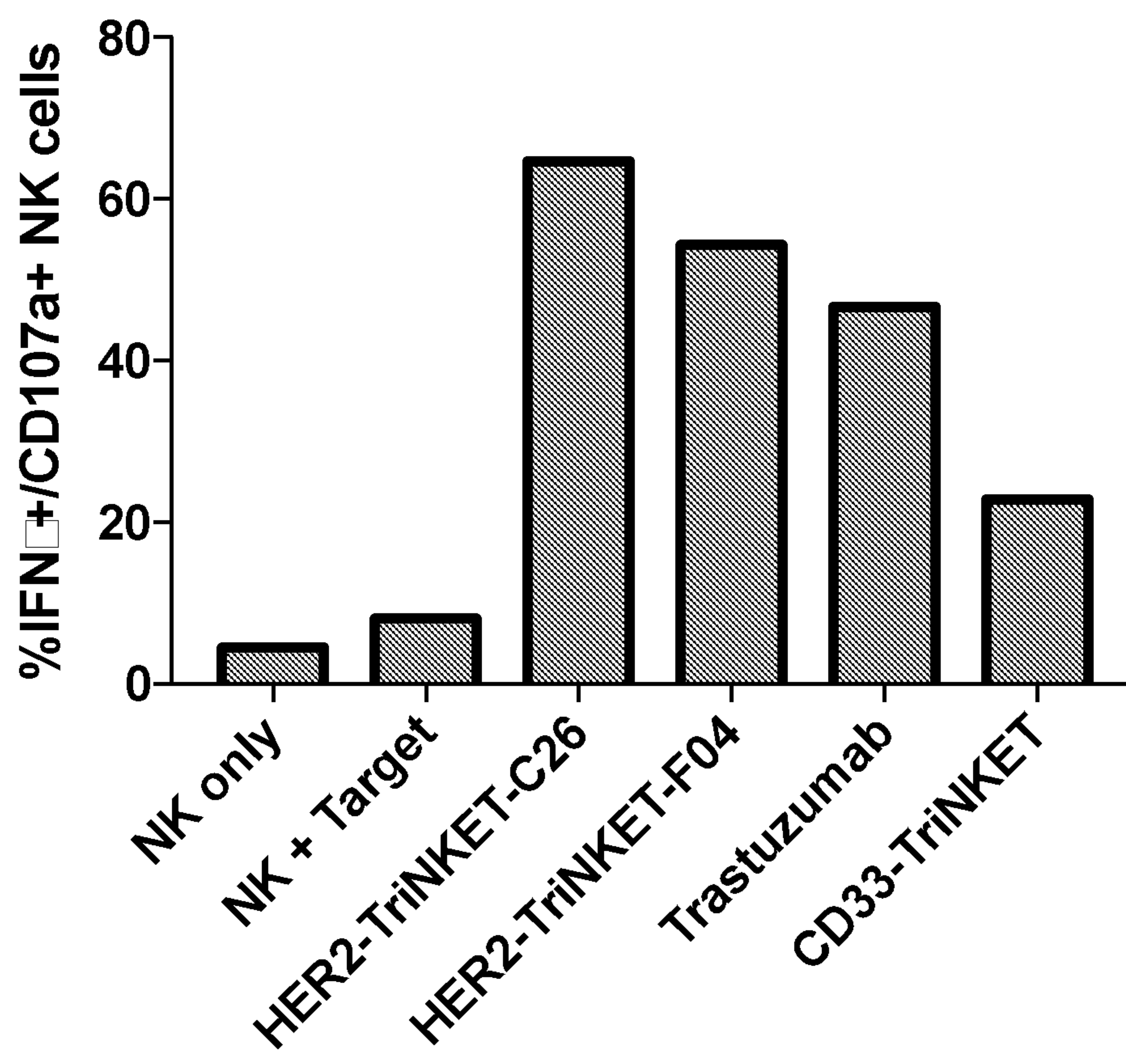


FIG. 28B

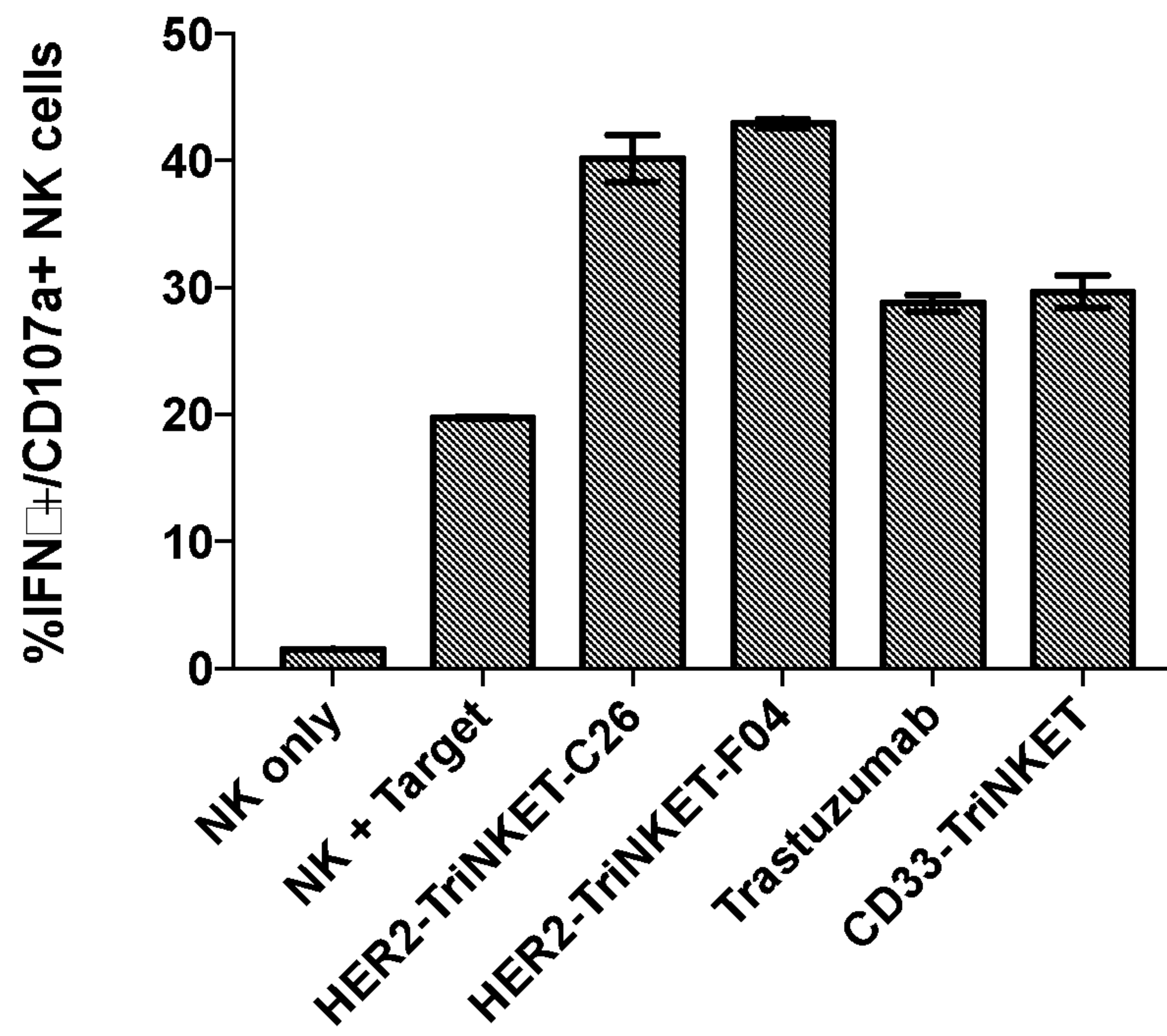


FIG. 28C

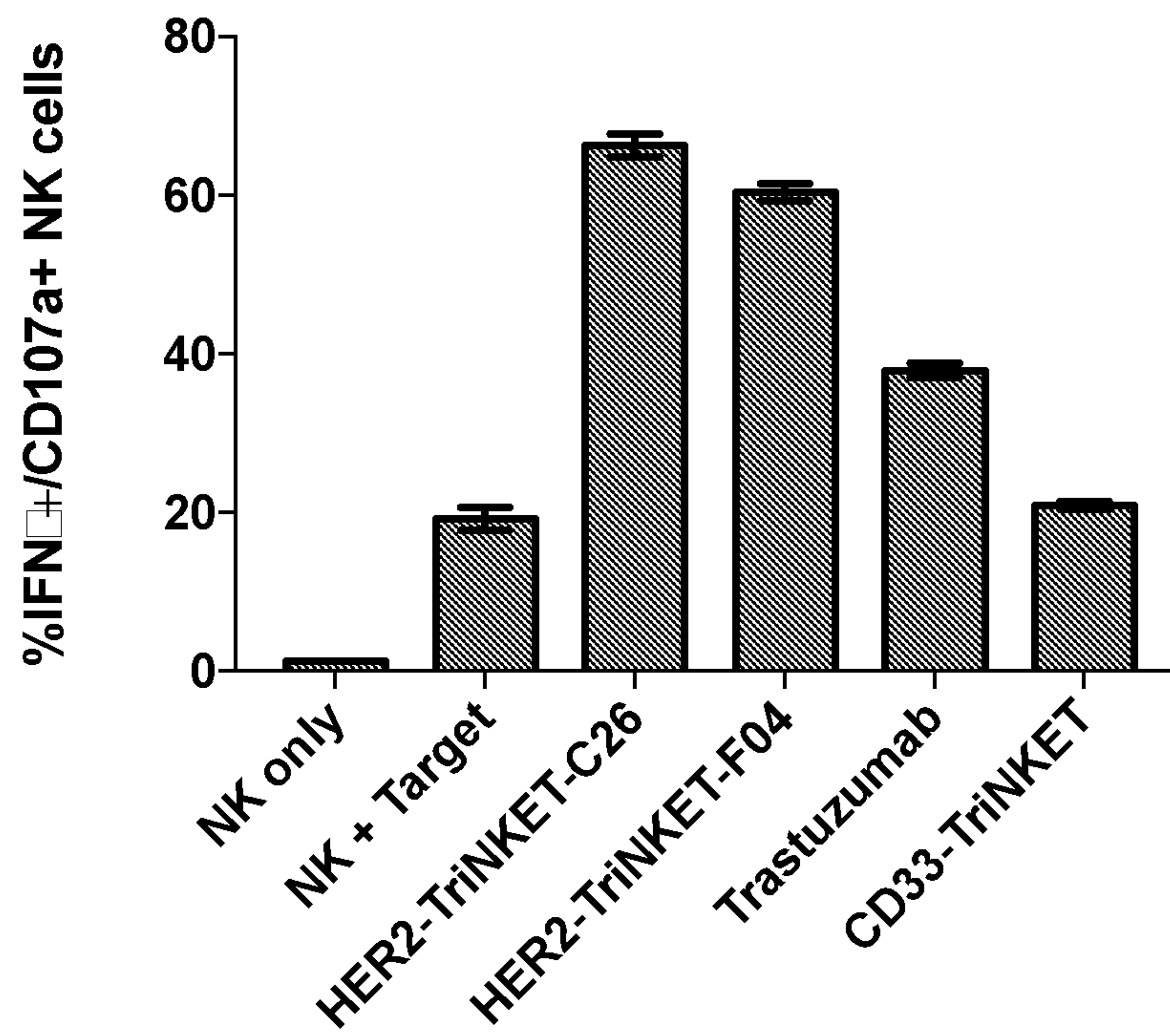


FIG. 29A

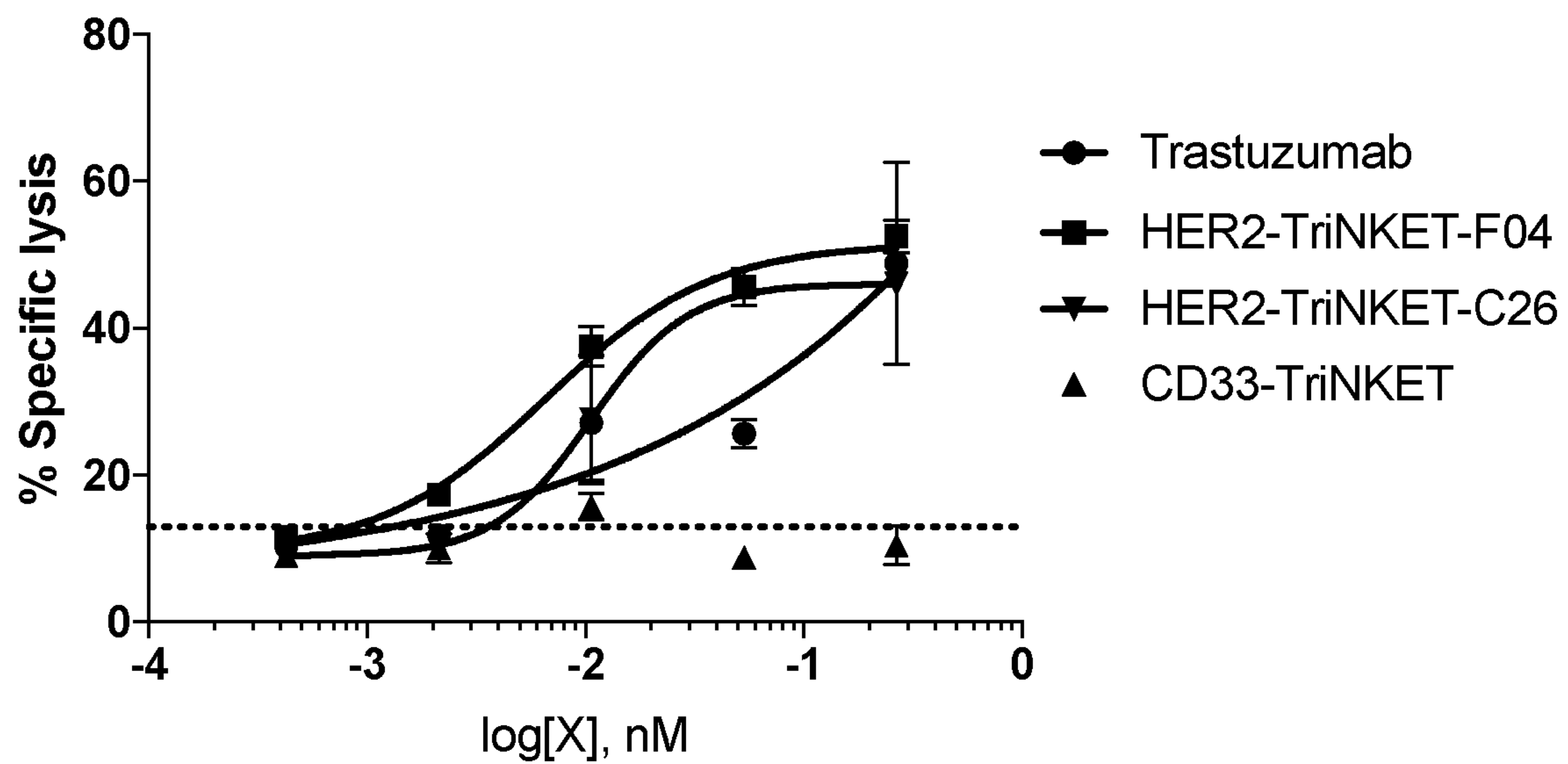


FIG. 29B

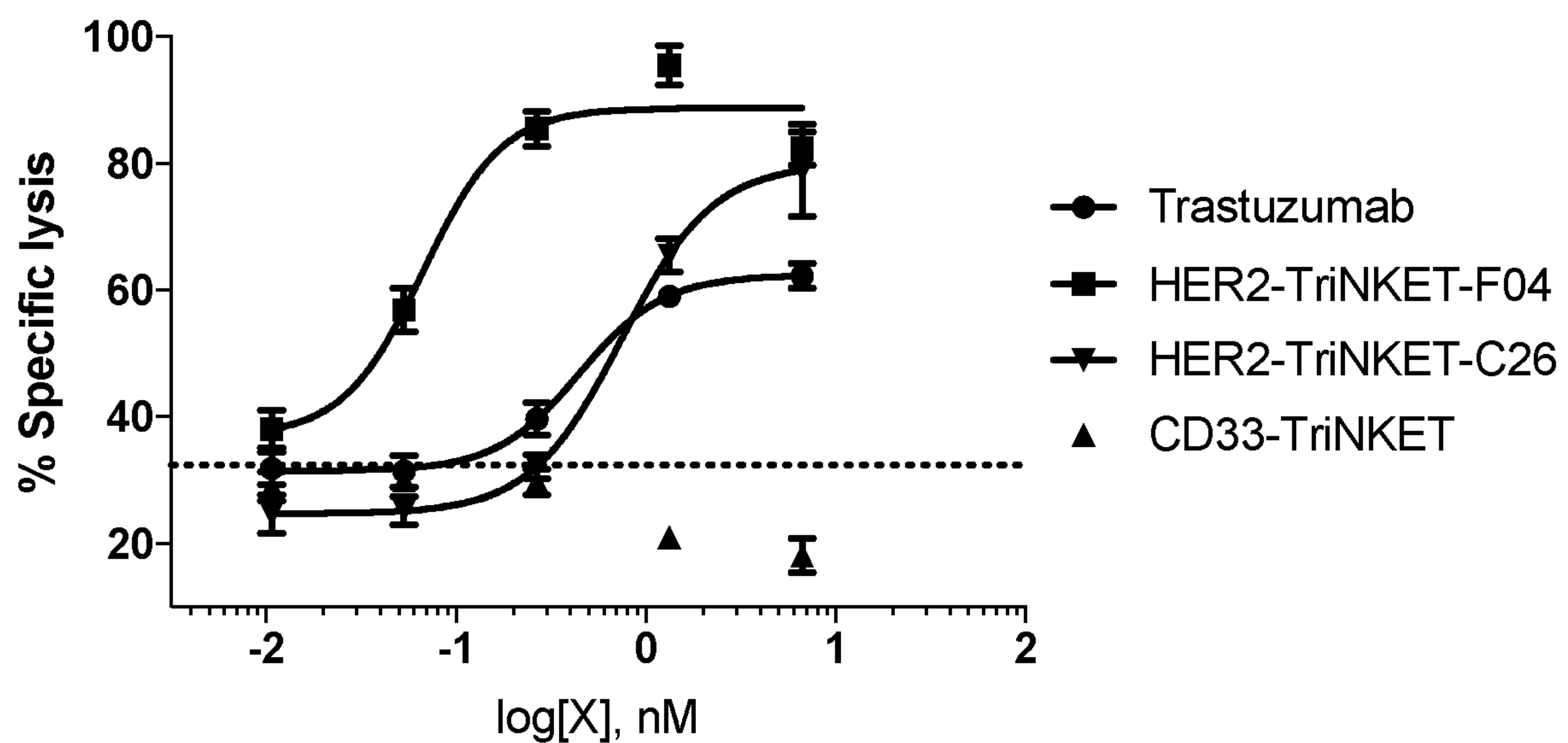


FIG. 30A

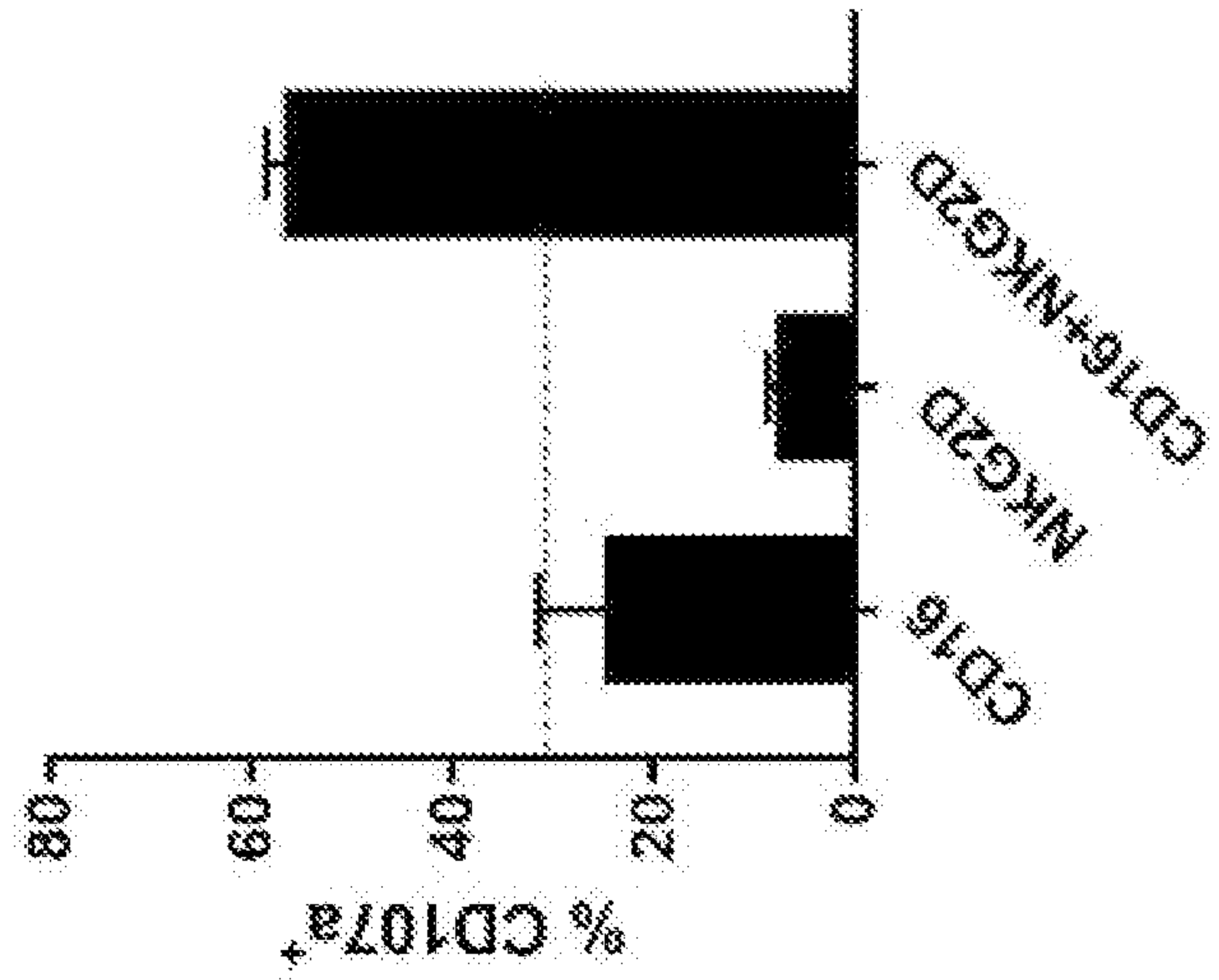


FIG. 30B

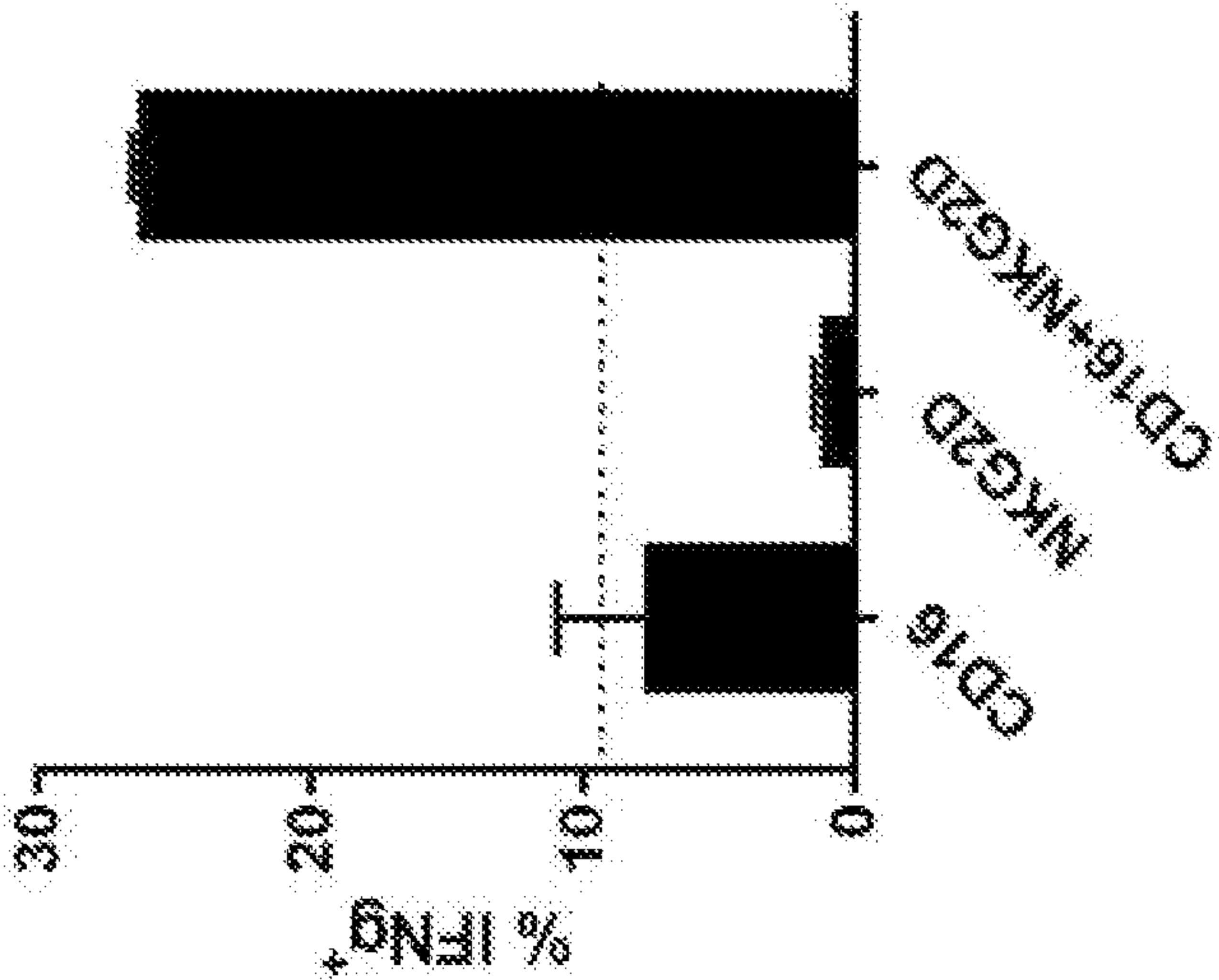


FIG. 30C

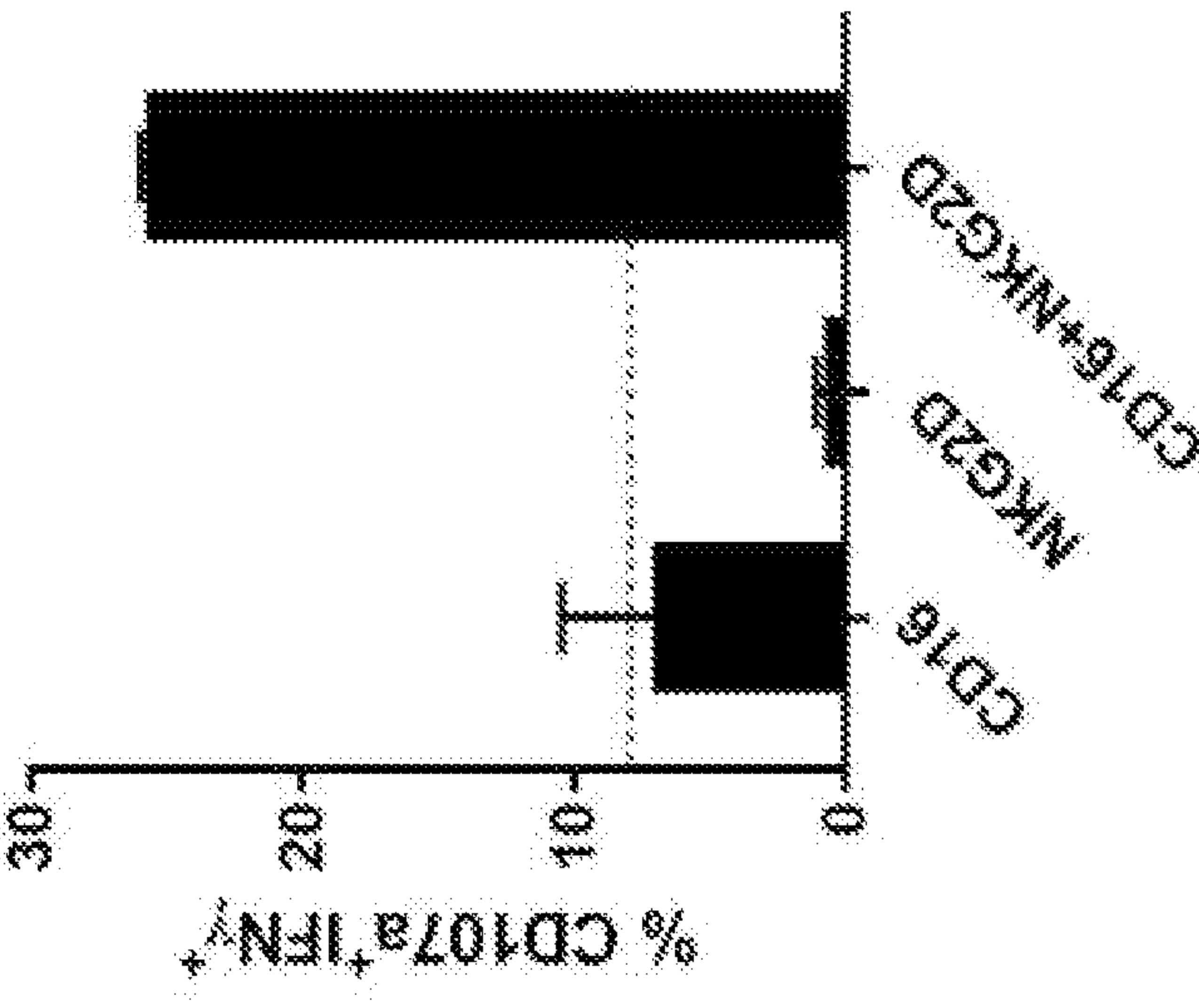


FIG. 31

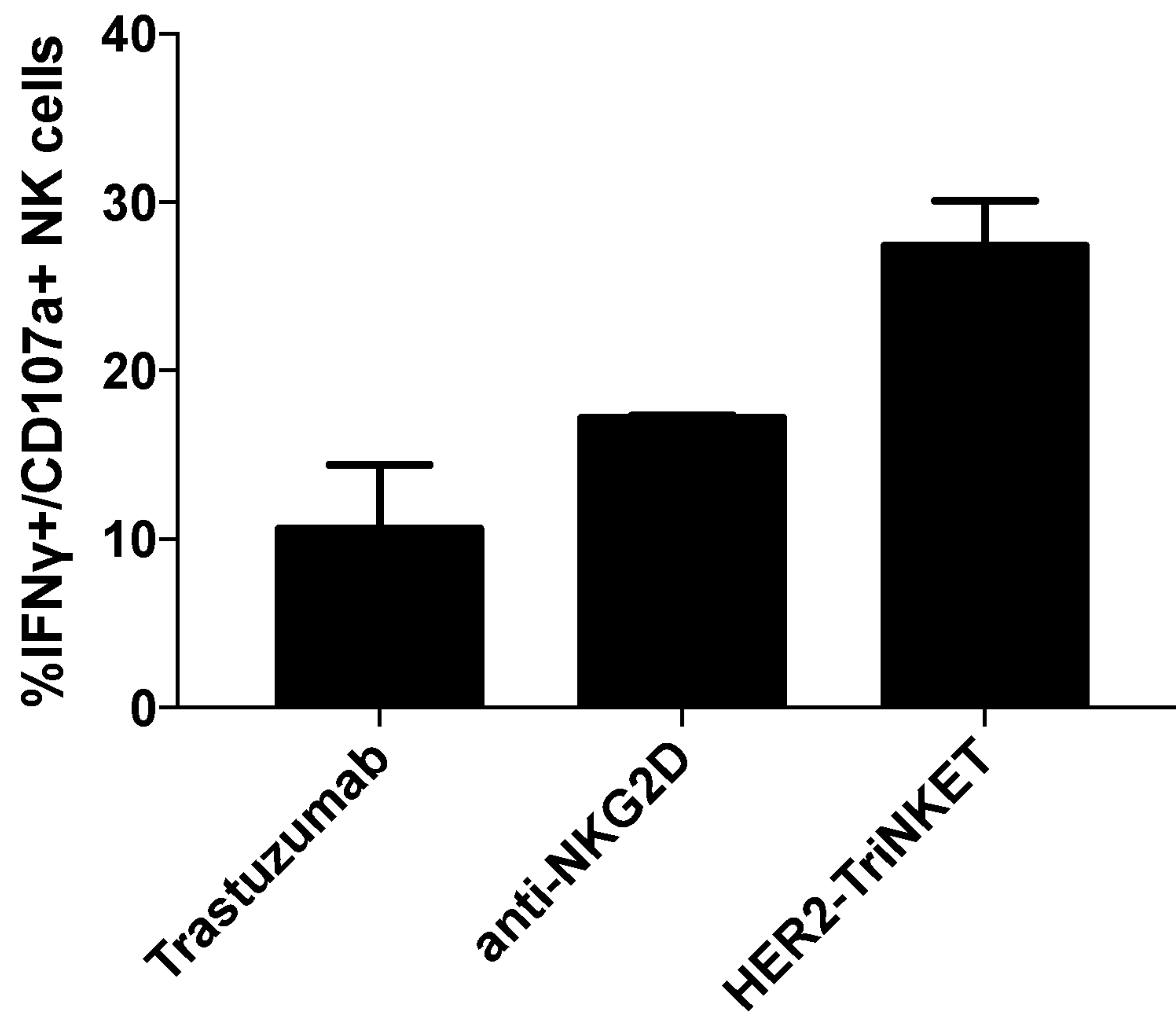


FIG. 32A

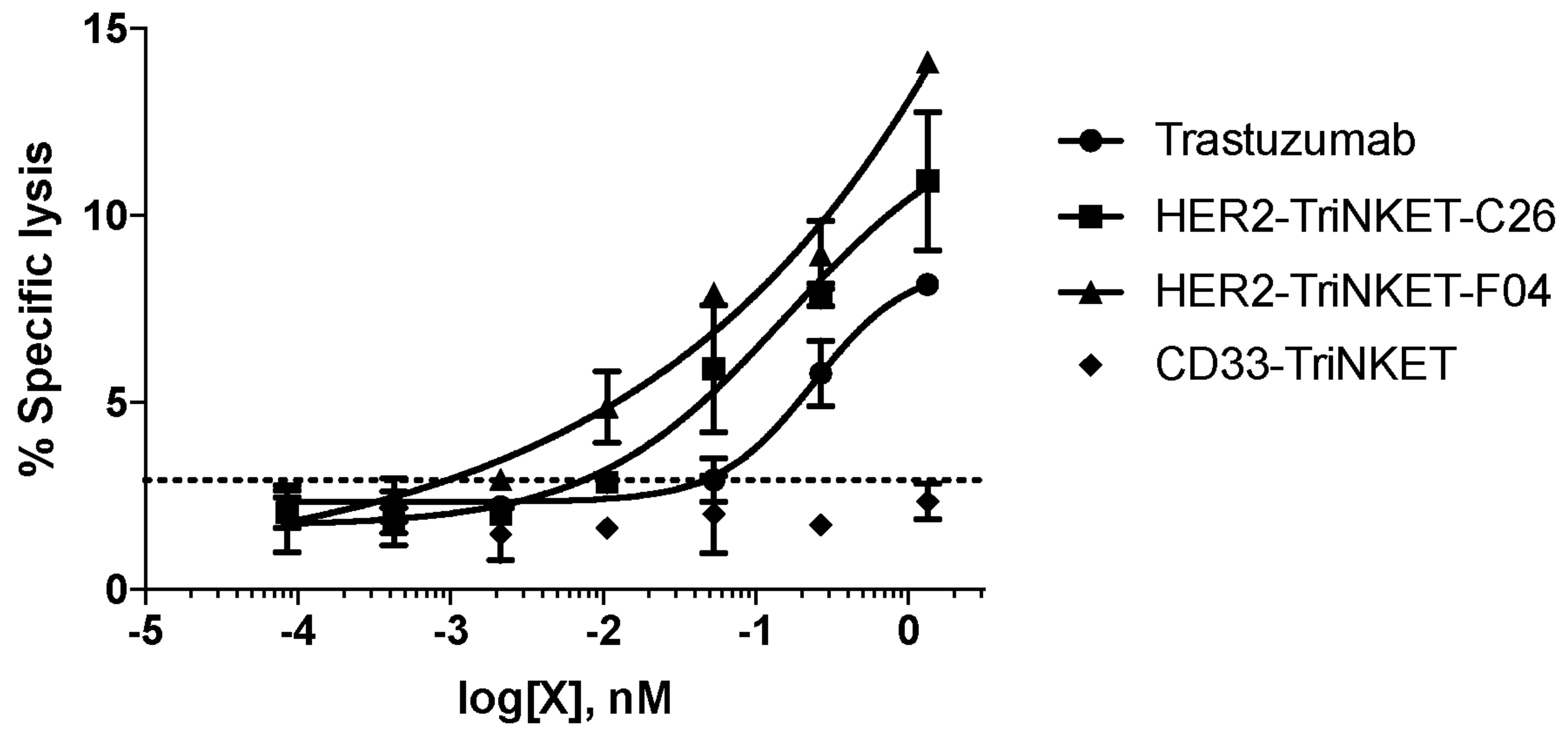


FIG. 32B

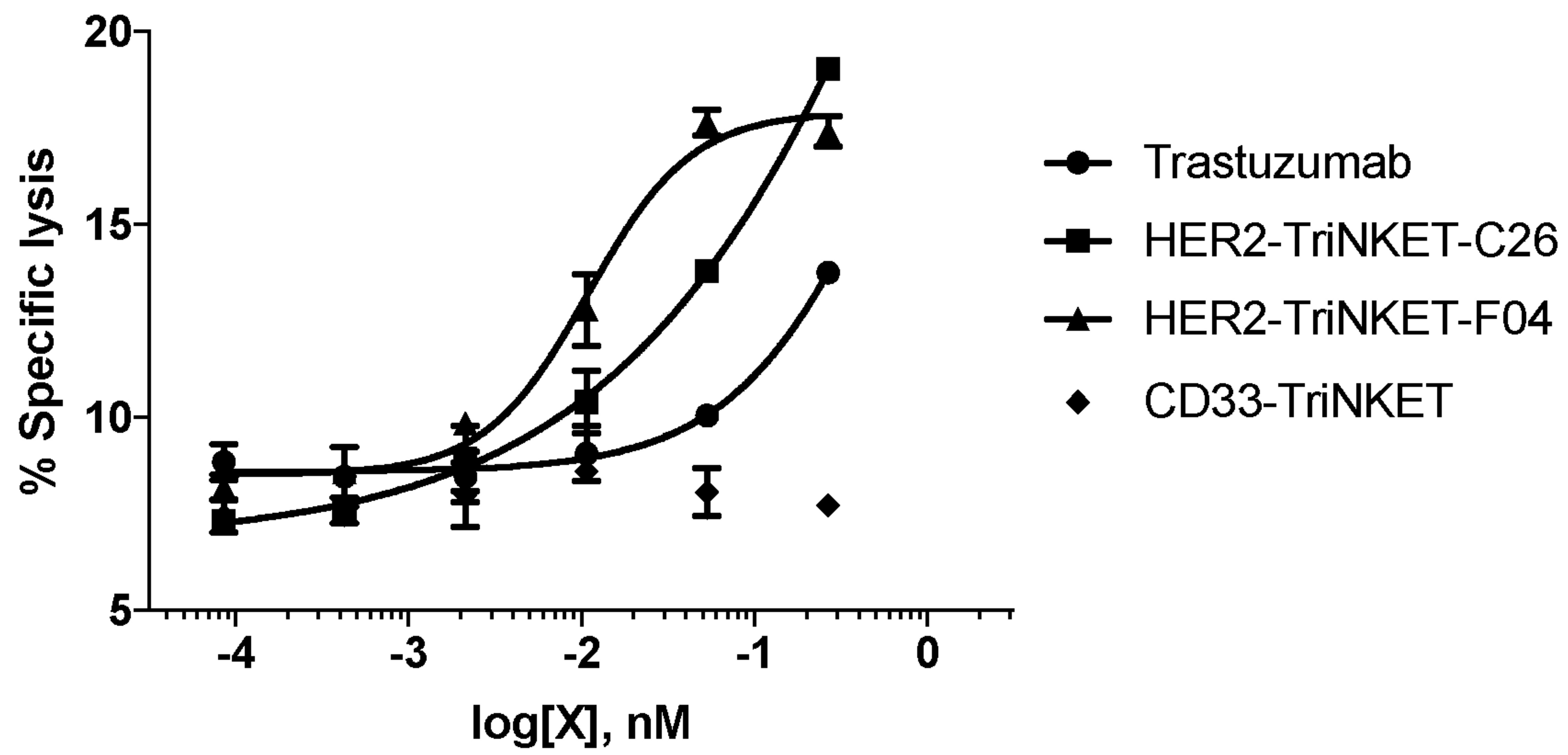


FIG. 32C

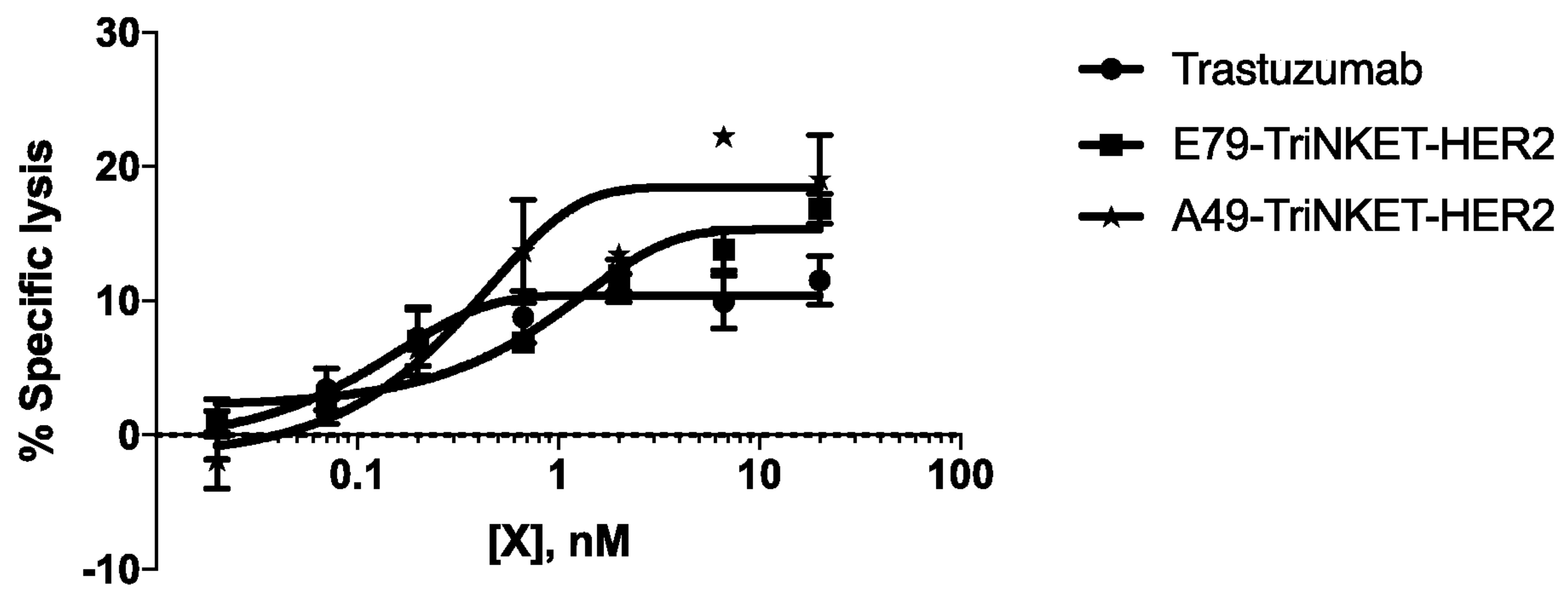


FIG. 33B

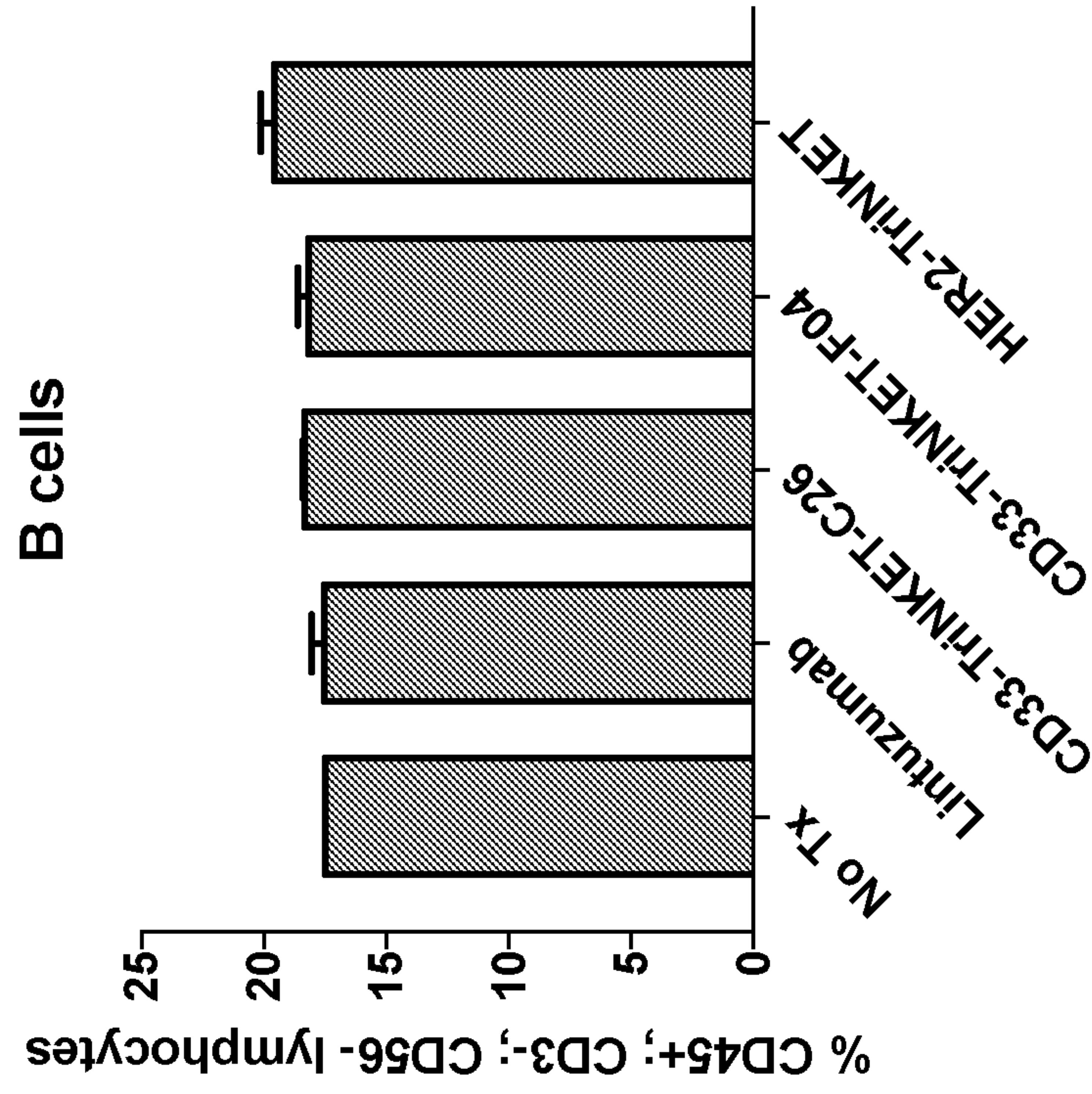


FIG. 33A

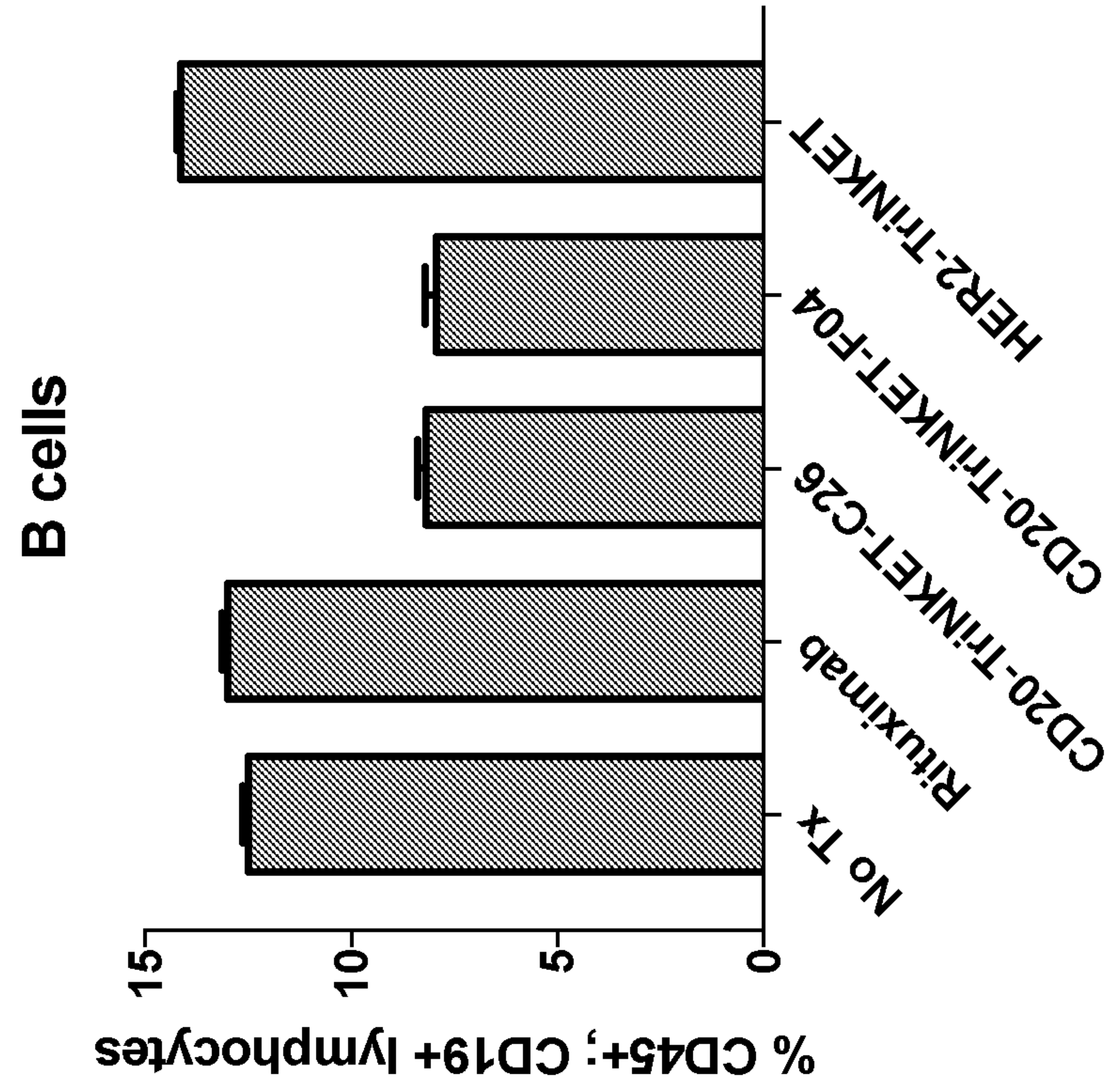


FIG. 33D

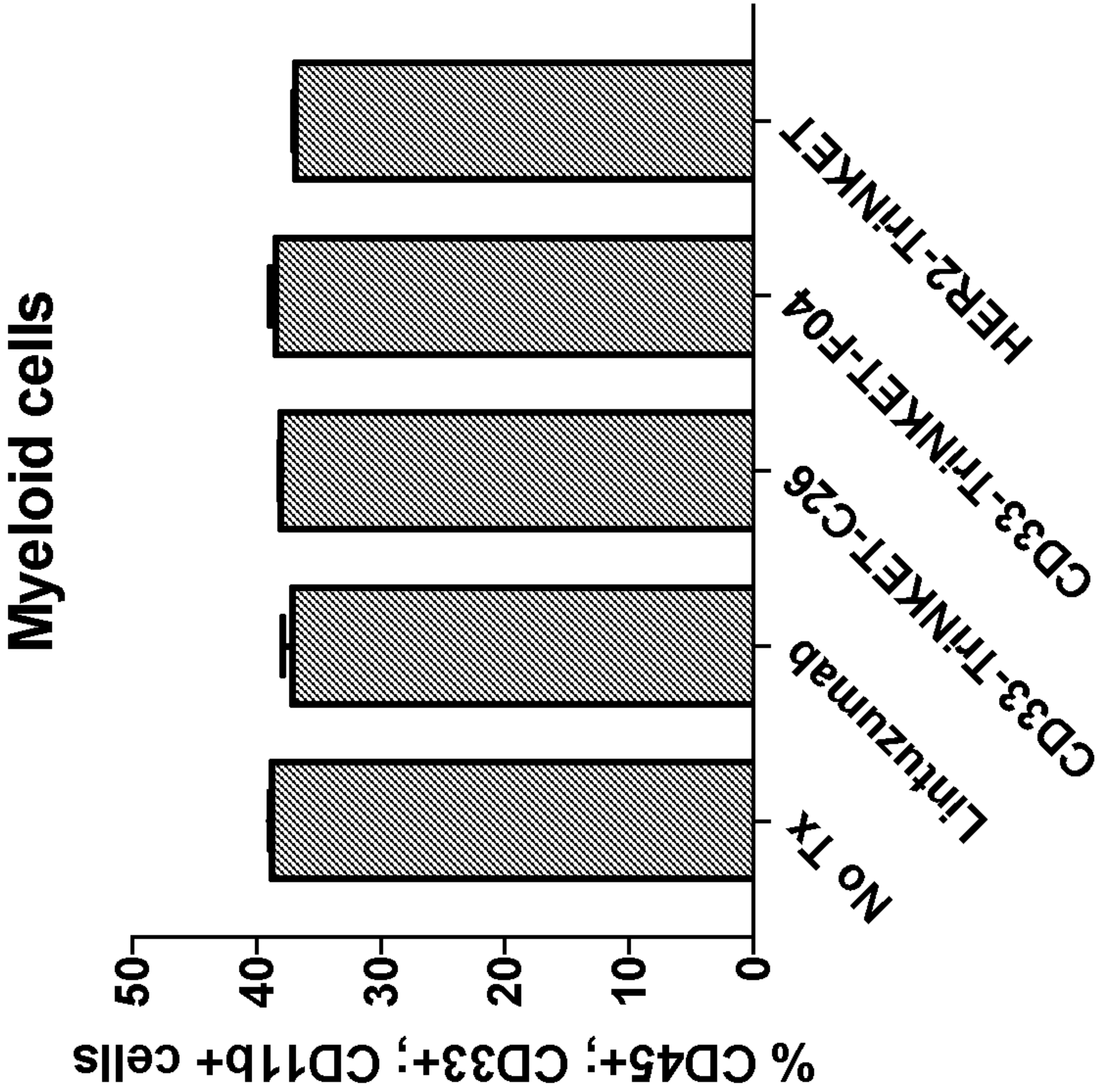


FIG. 33C

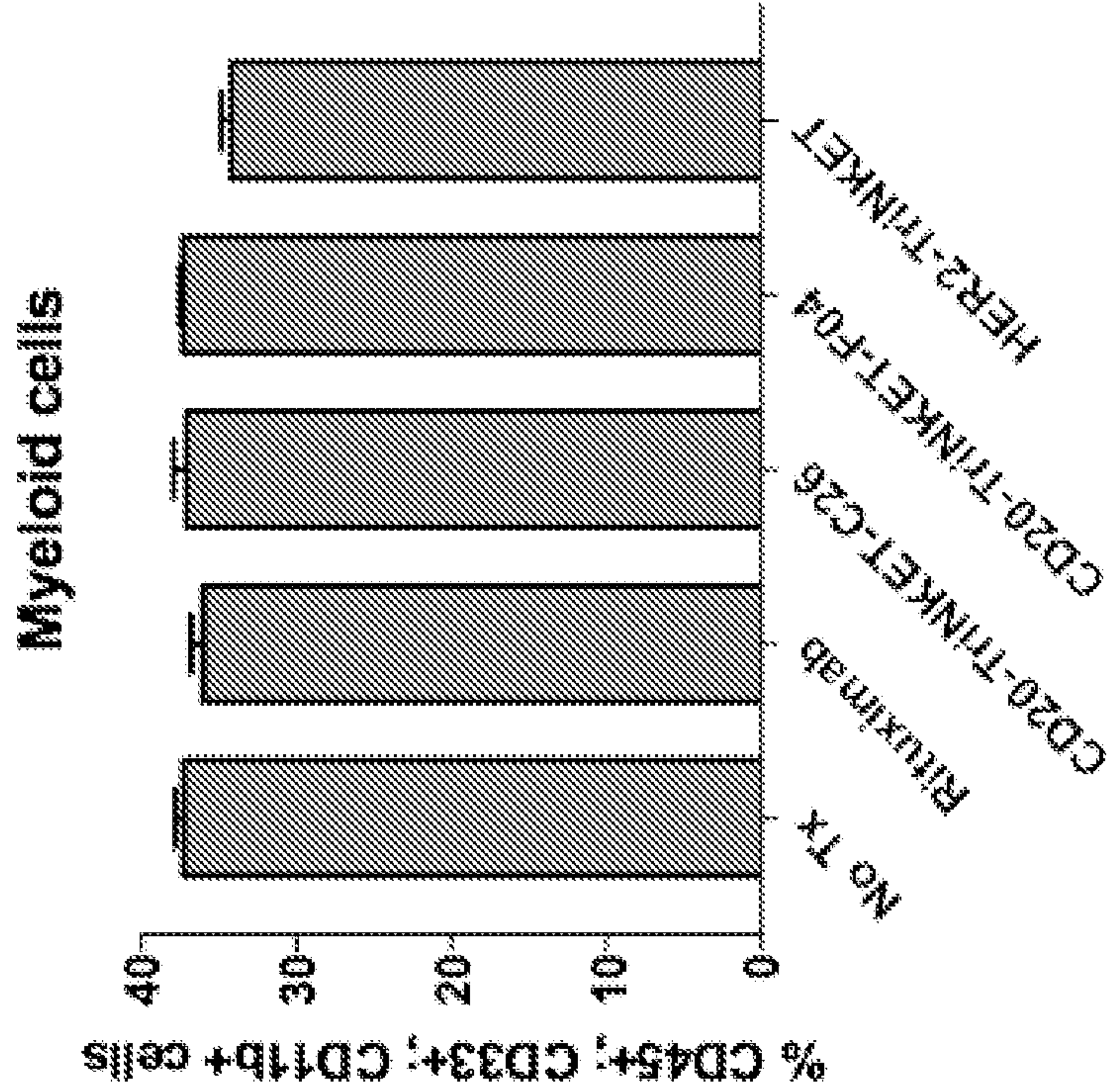


FIG. 34

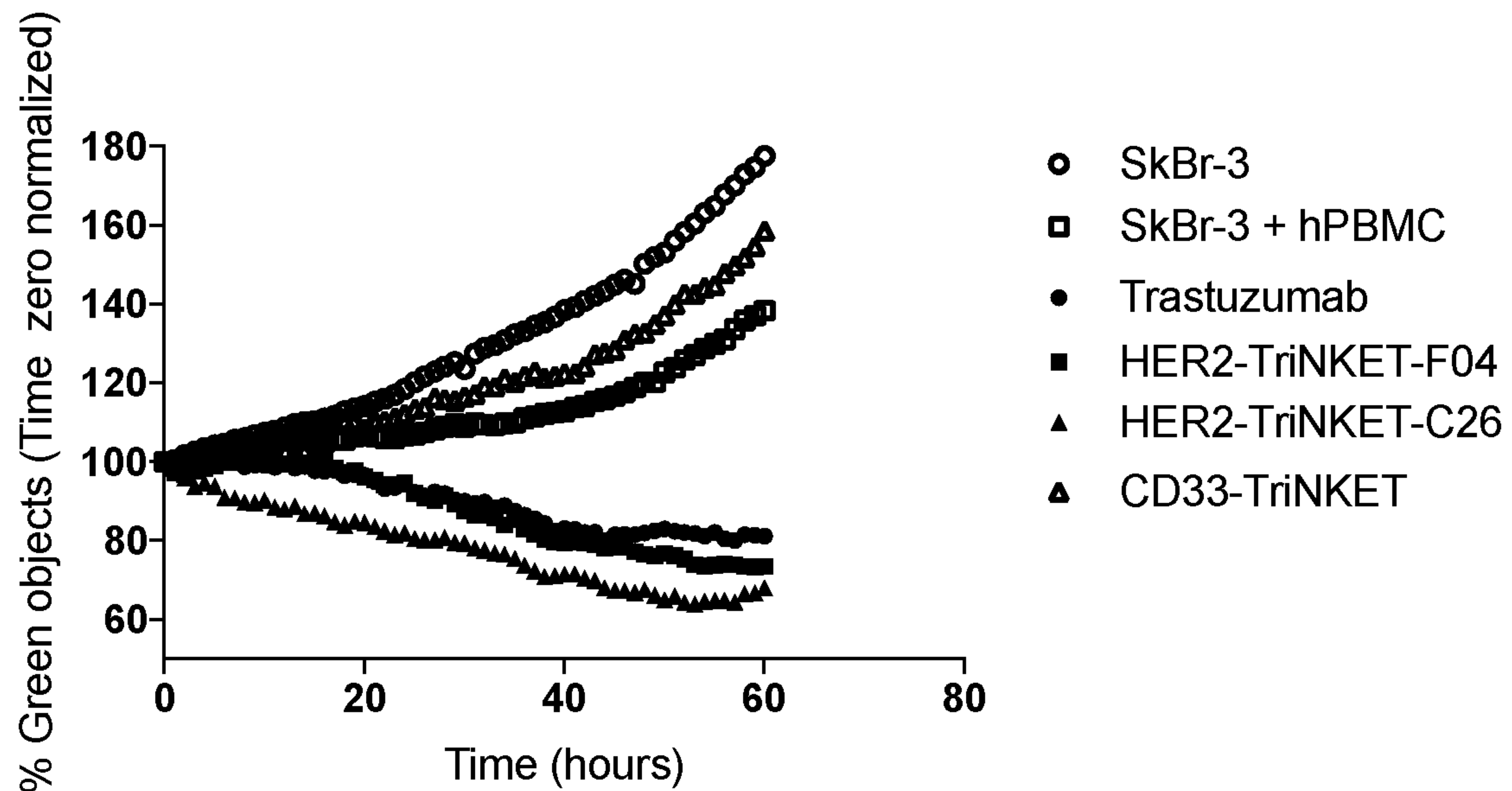
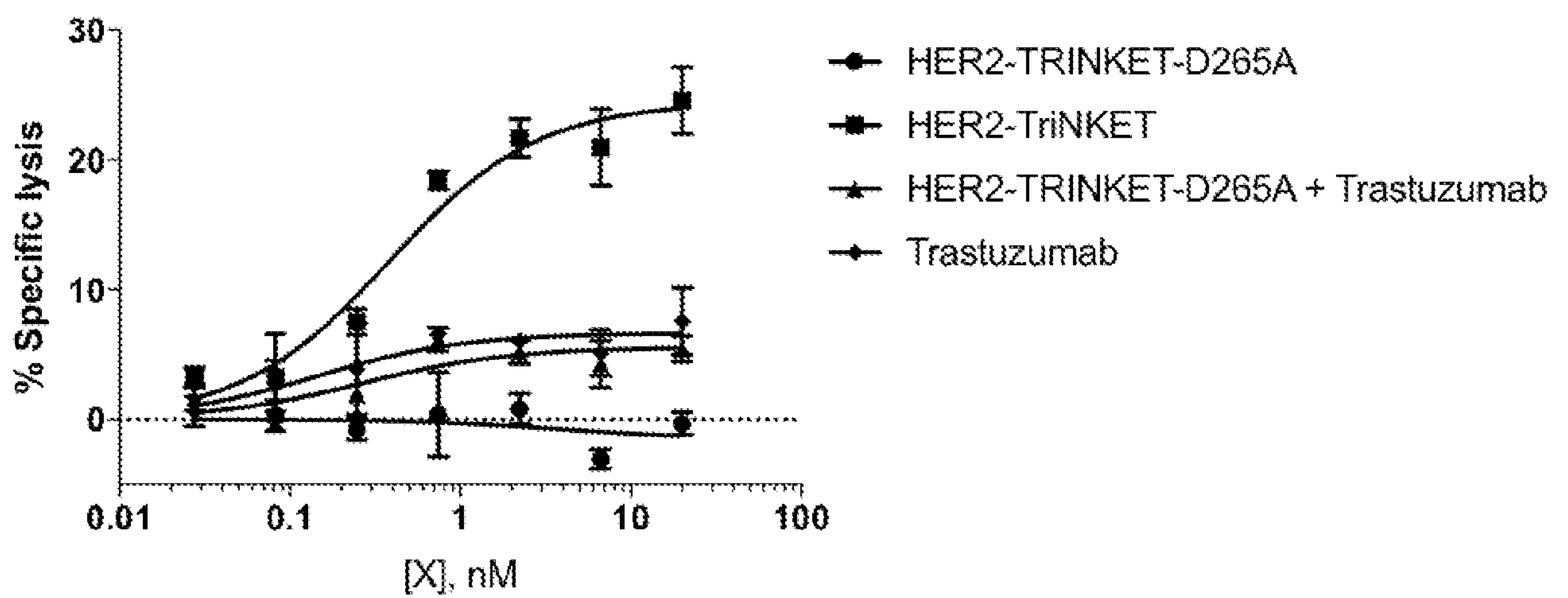


FIG. 35



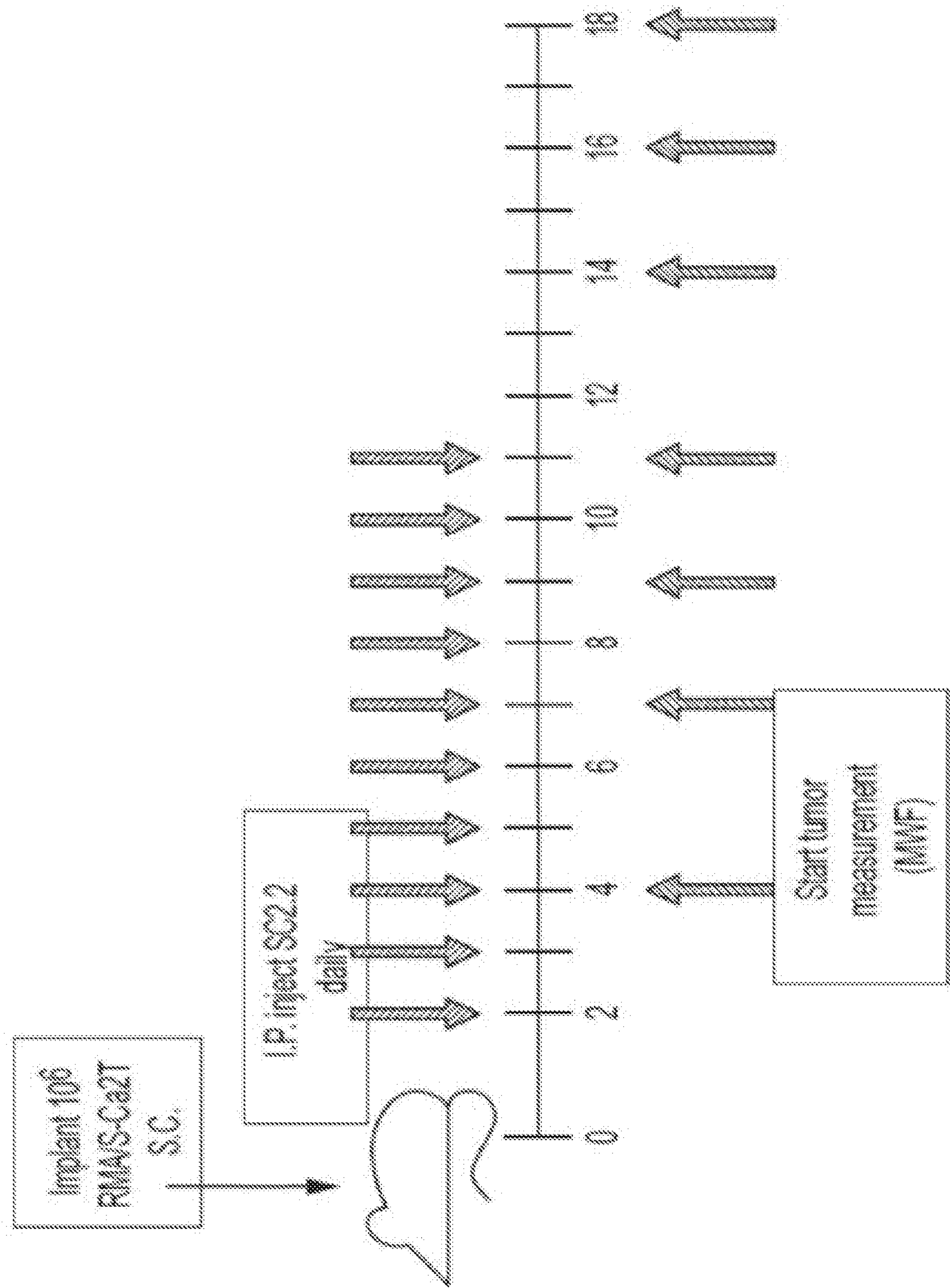


FIG. 36

FIG. 37

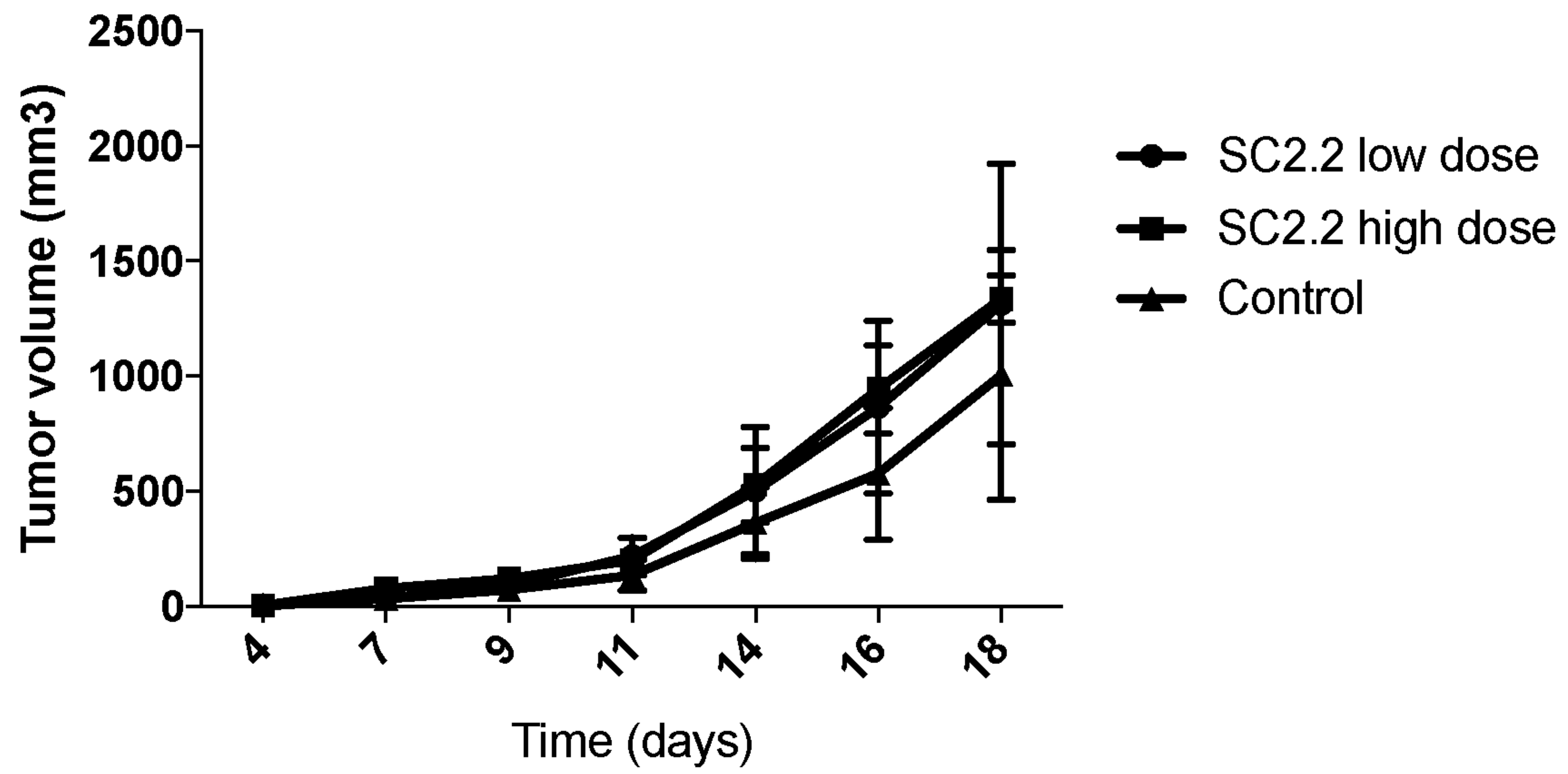


FIG. 38A

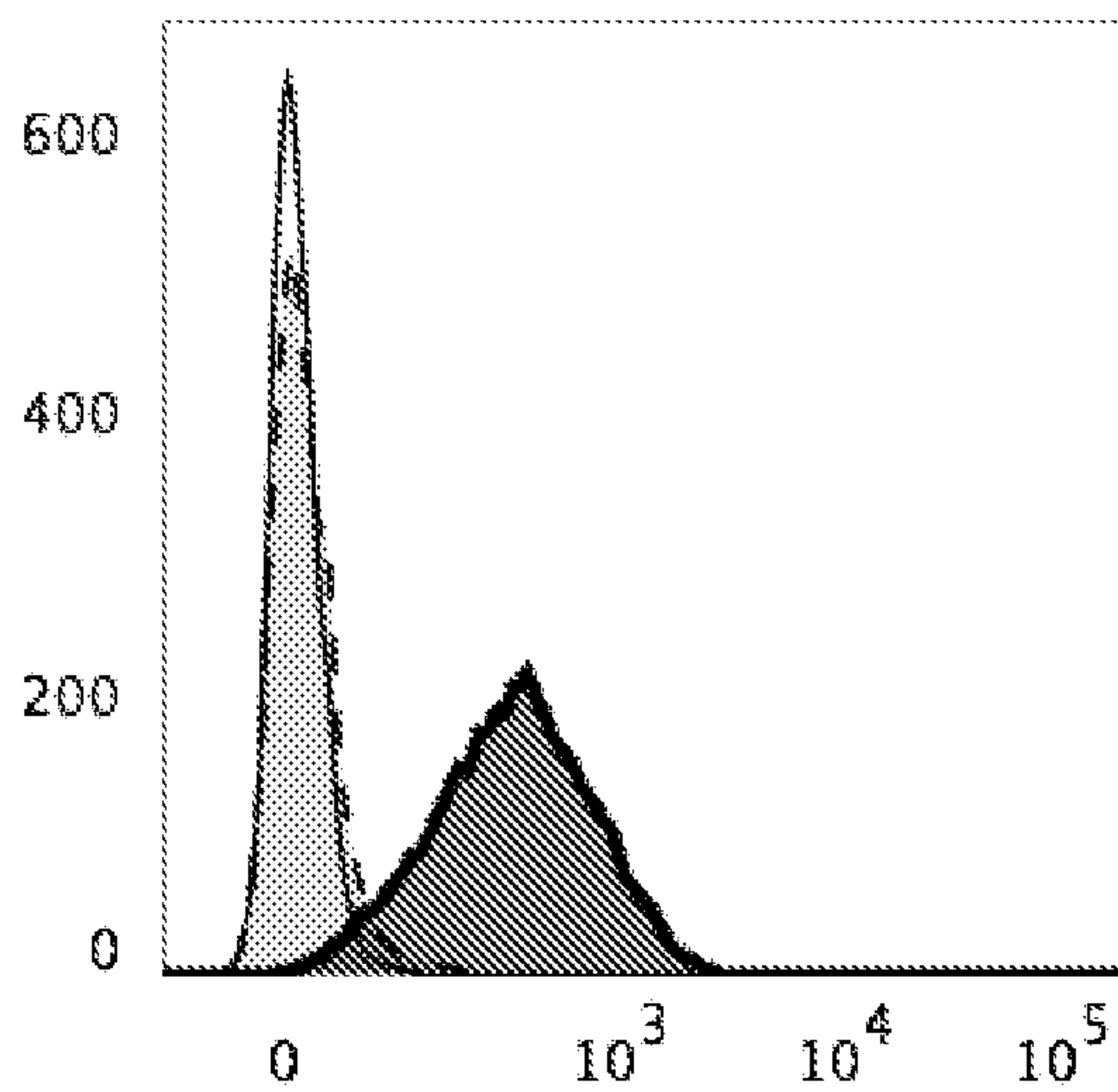


FIG. 38B

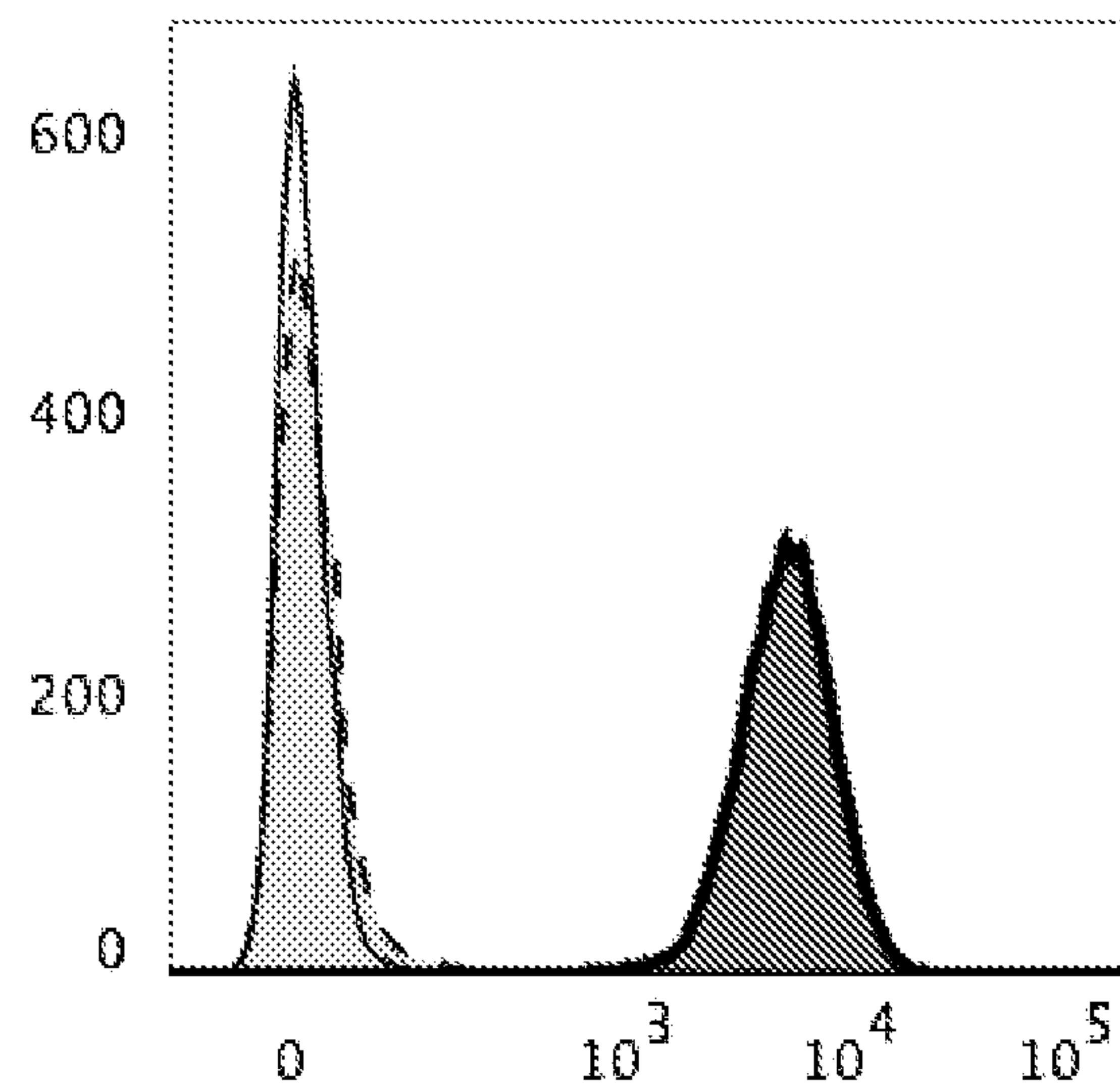


FIG. 39

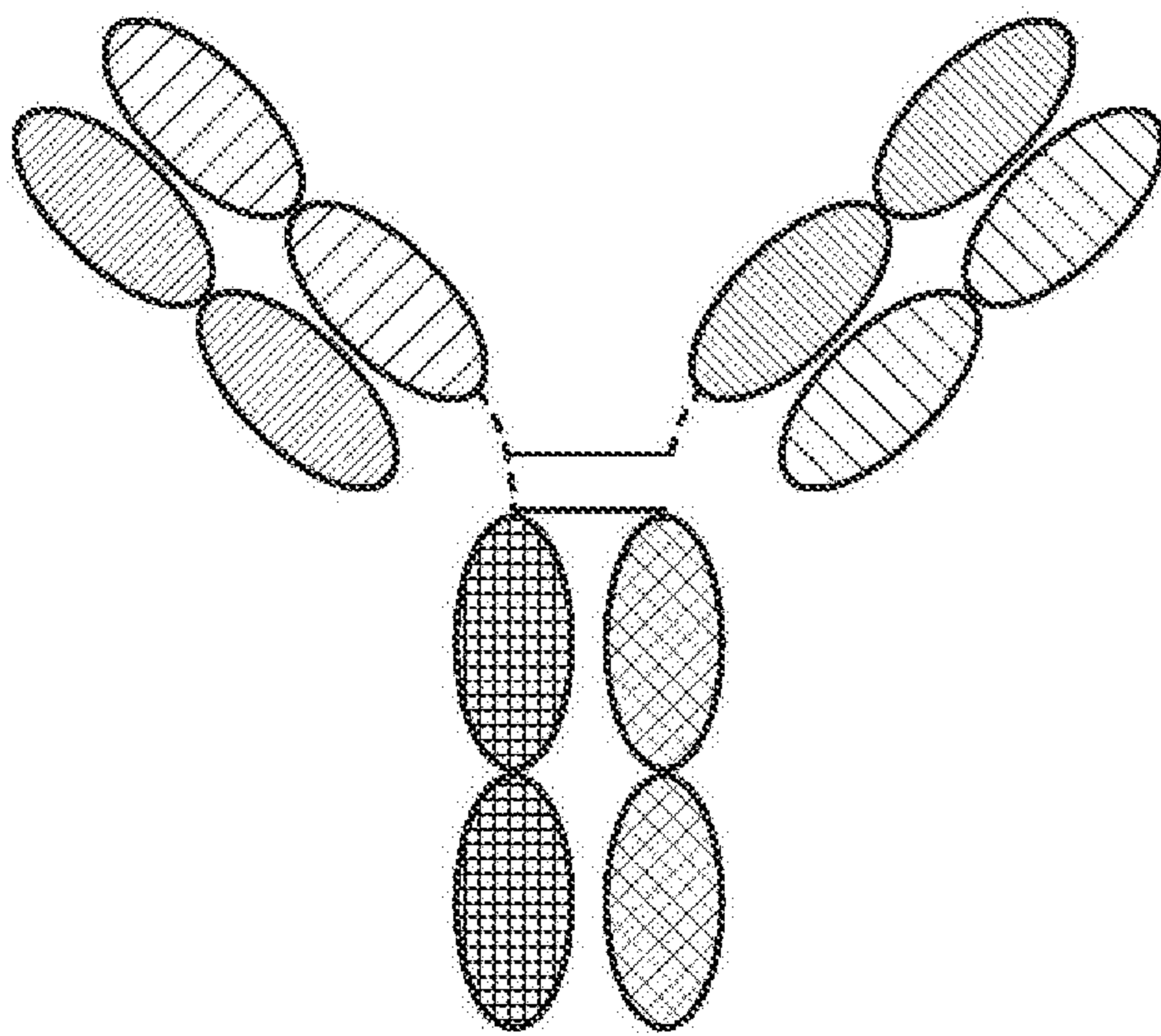


FIG. 40

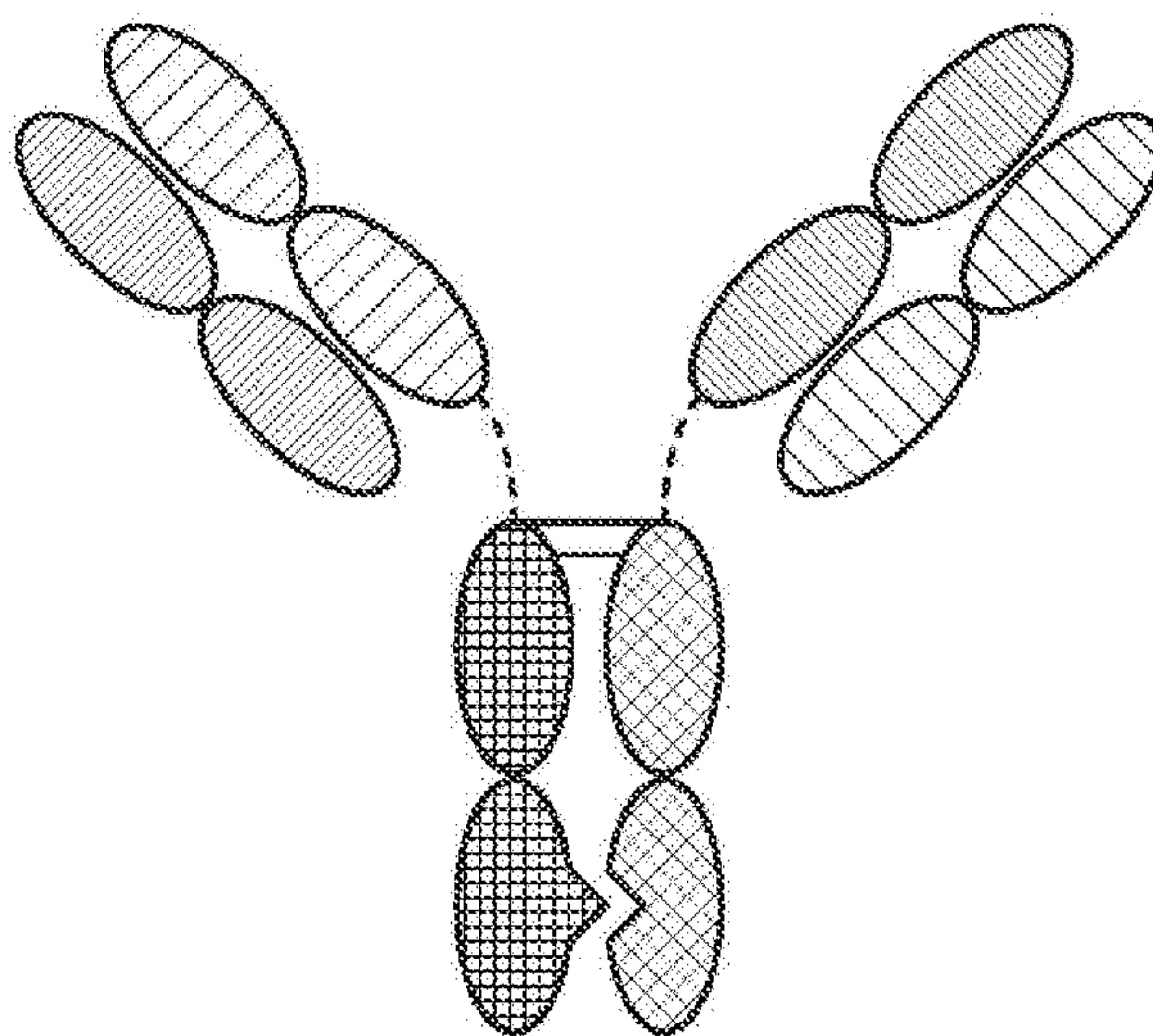


FIG. 41

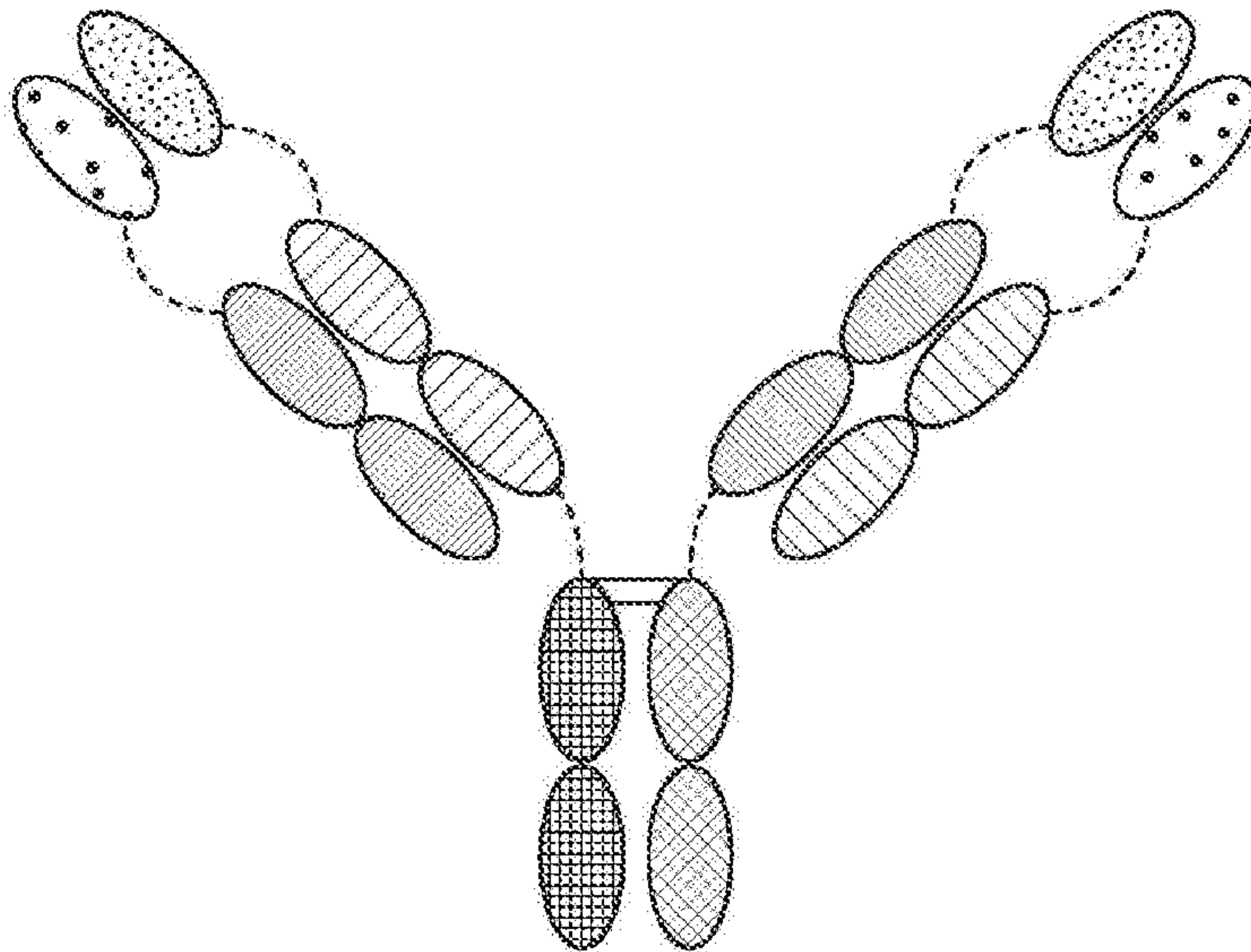


FIG. 42

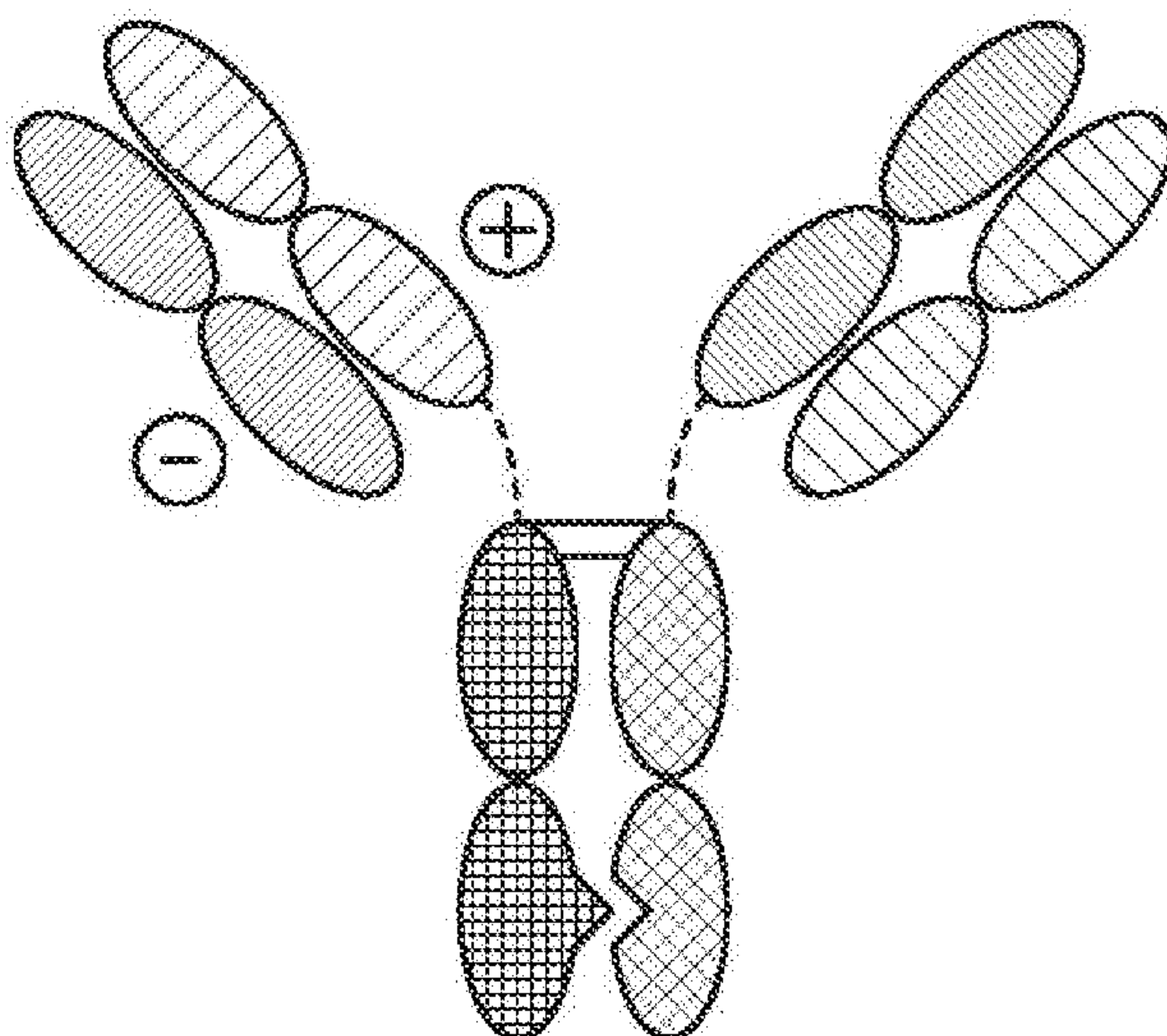


FIG. 43

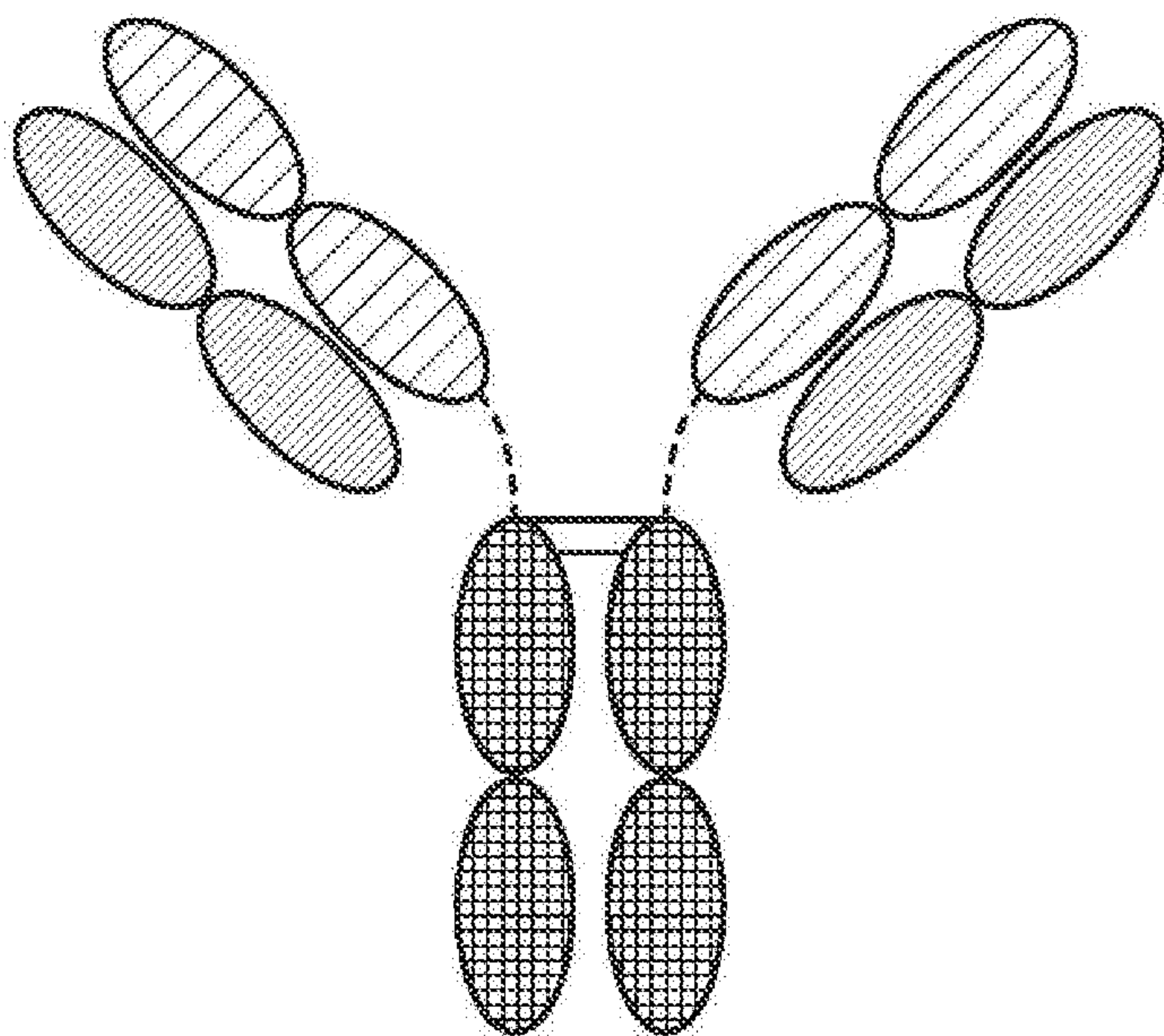


FIG. 44

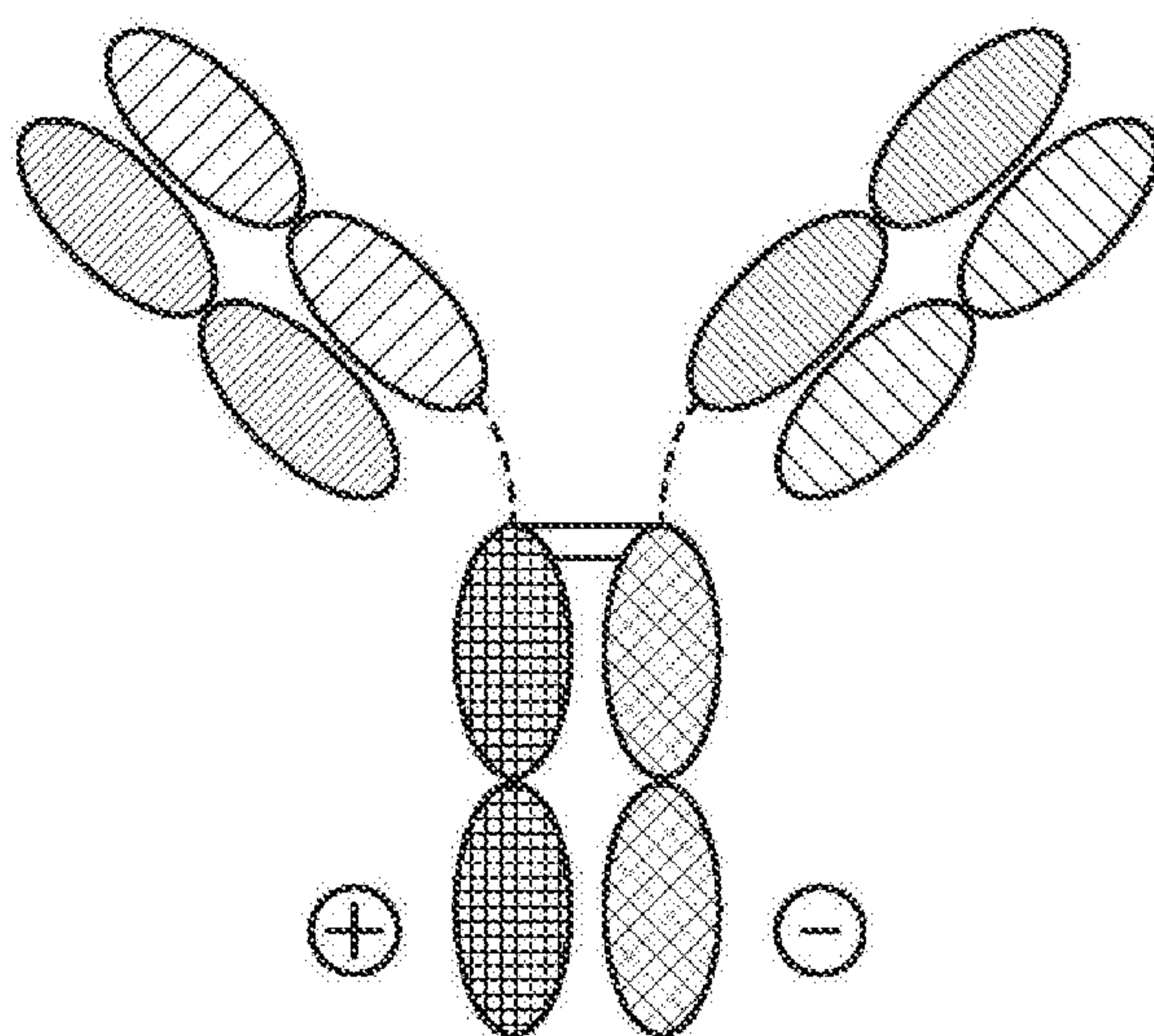


FIG. 45

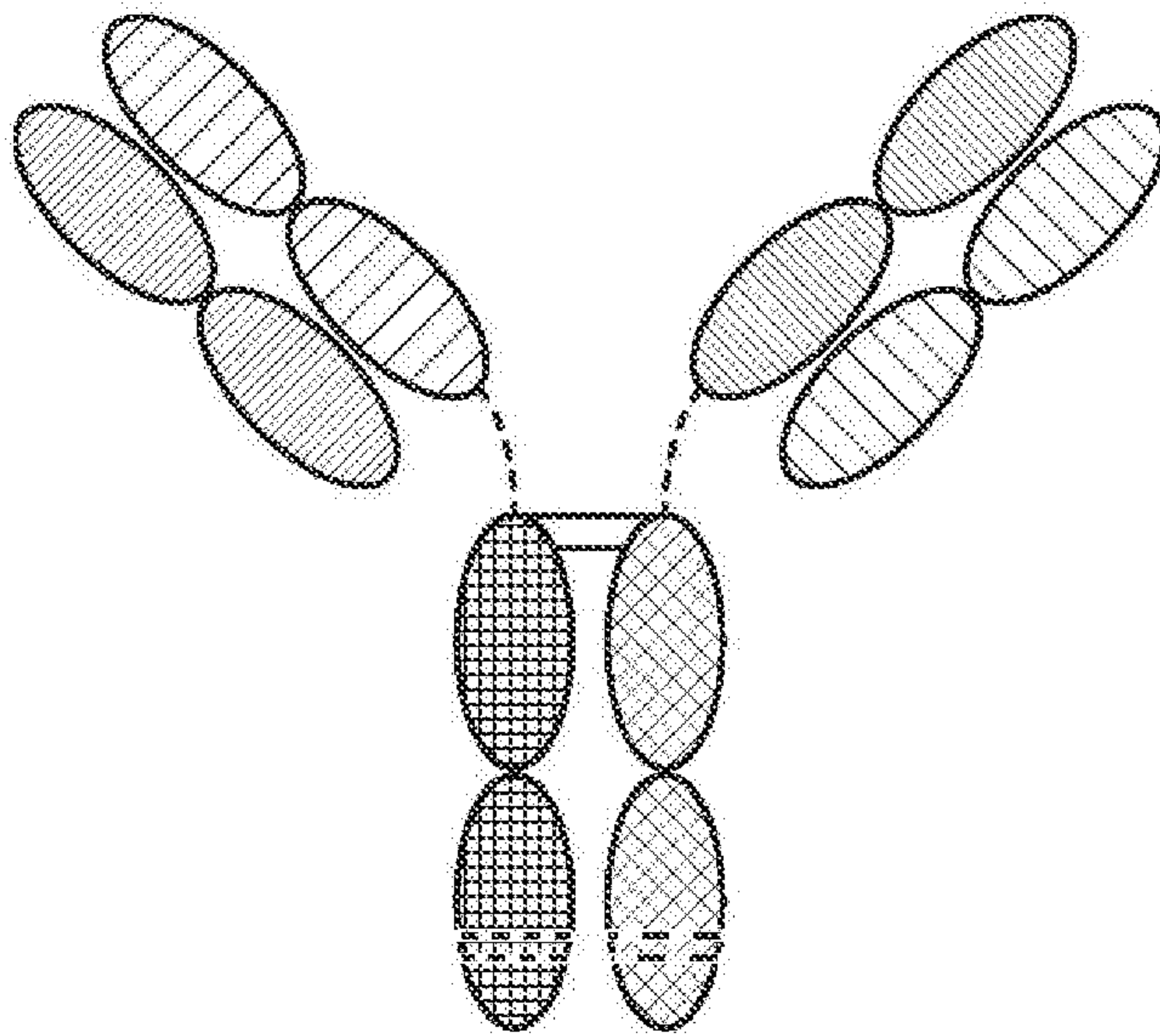


FIG. 46

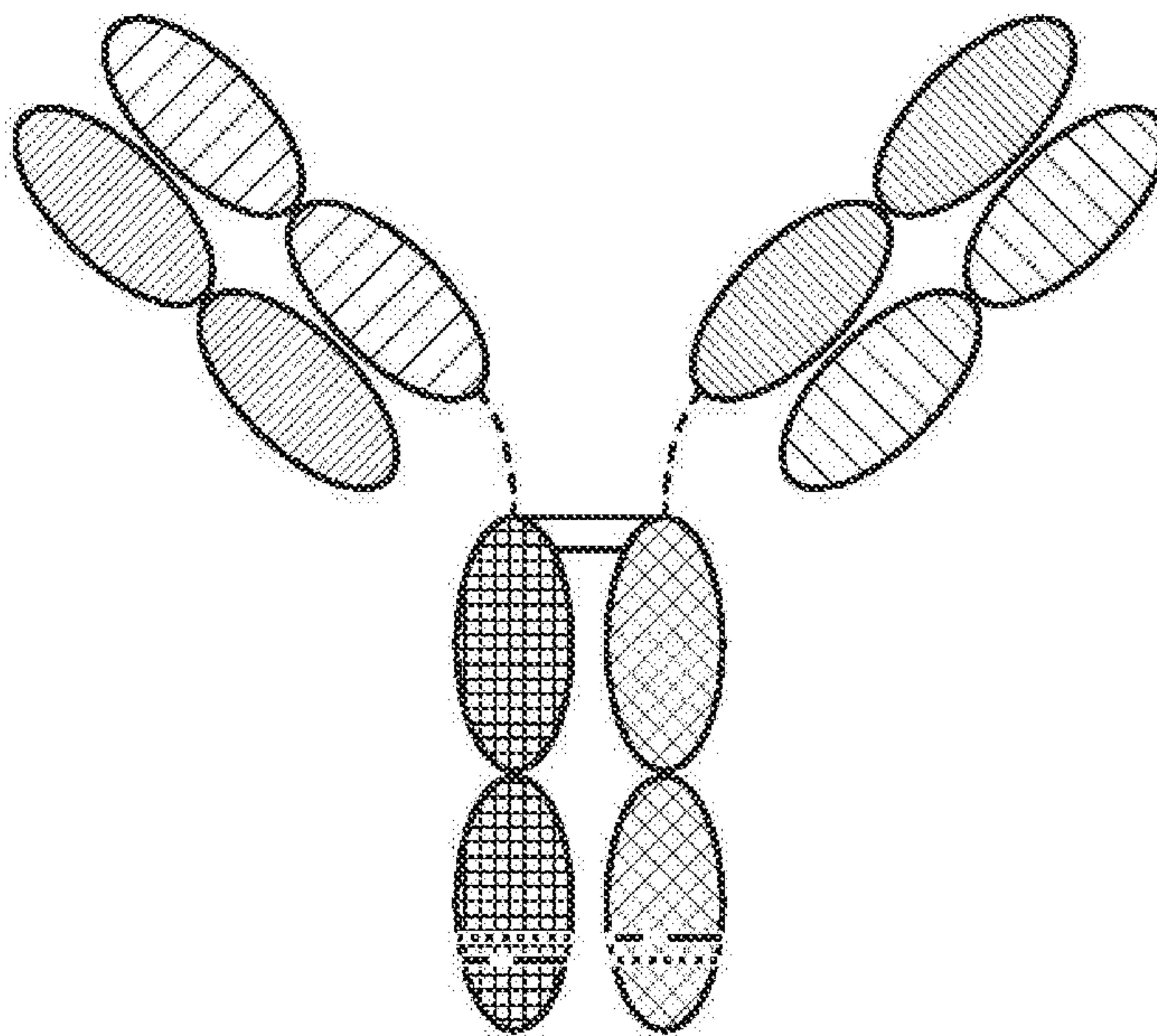


FIG. 47

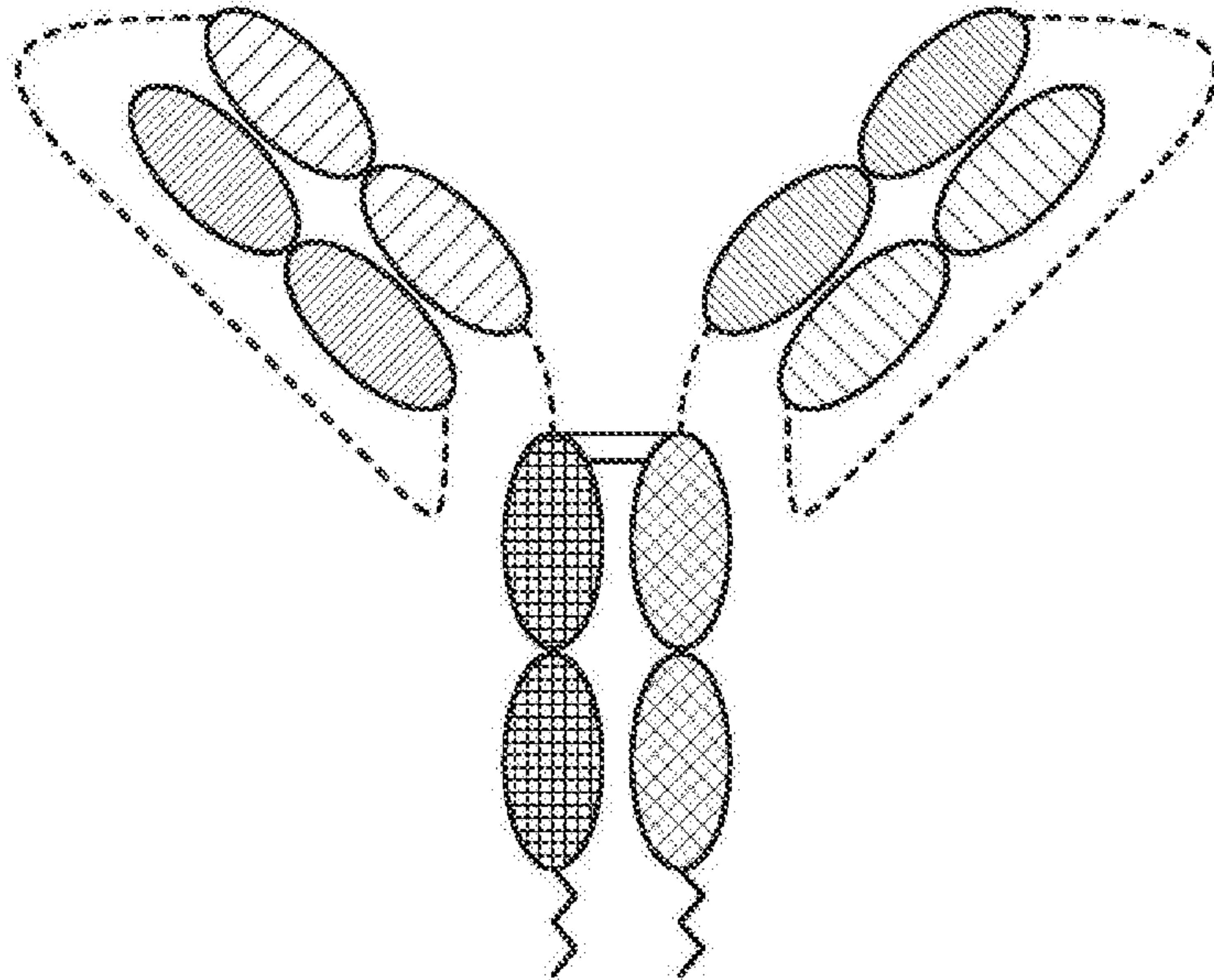


FIG. 48

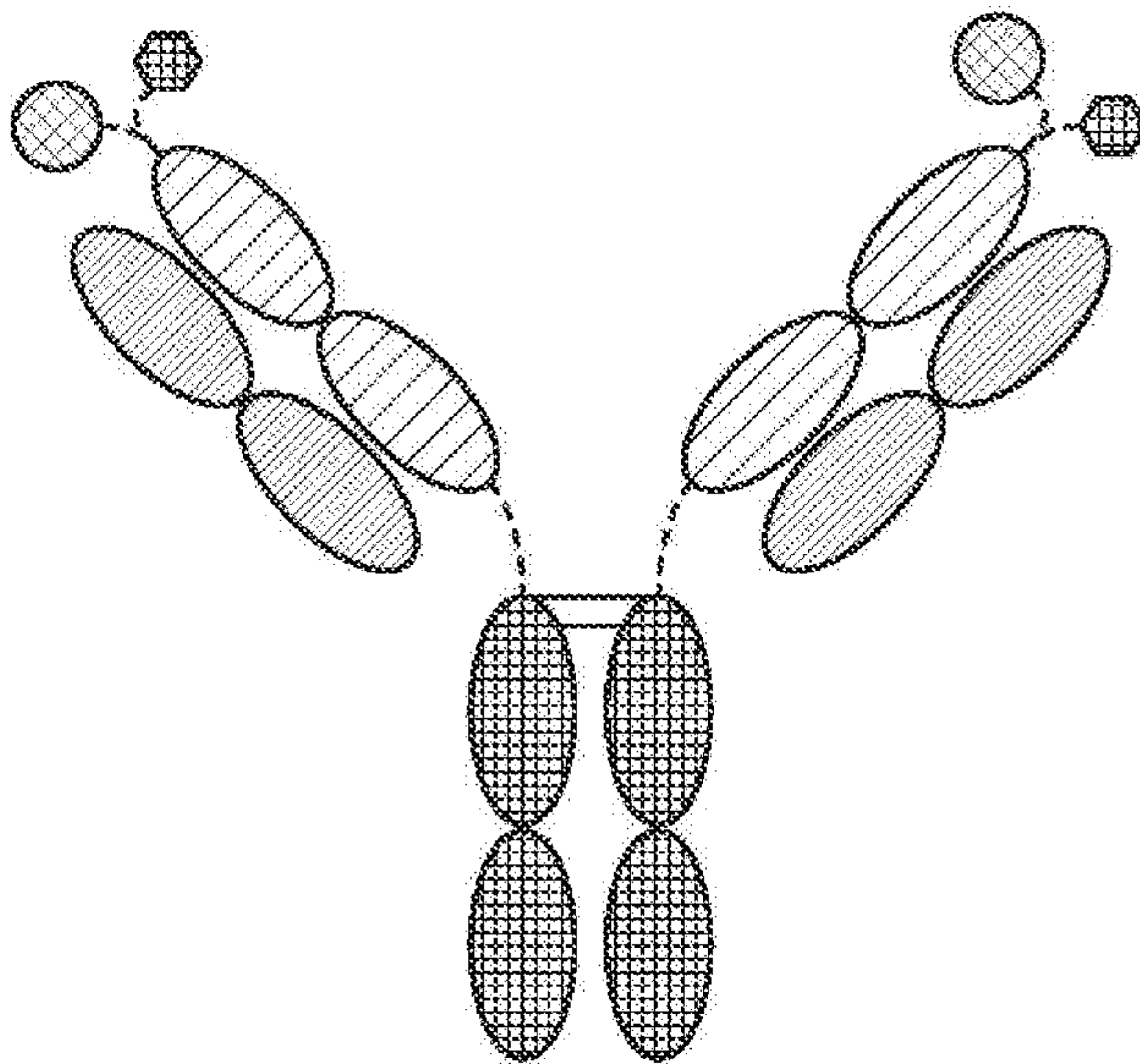


FIG. 49A

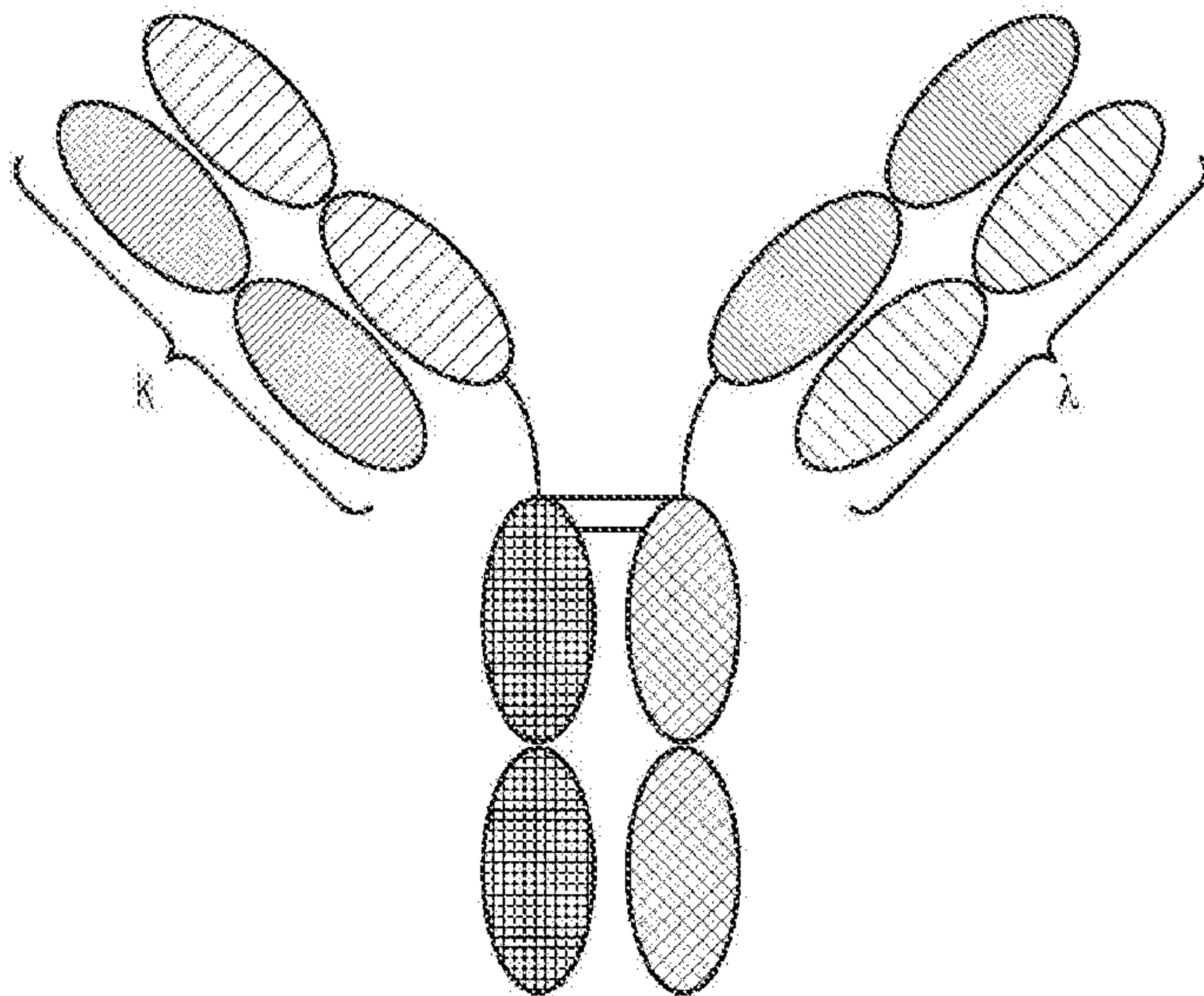


FIG. 49B

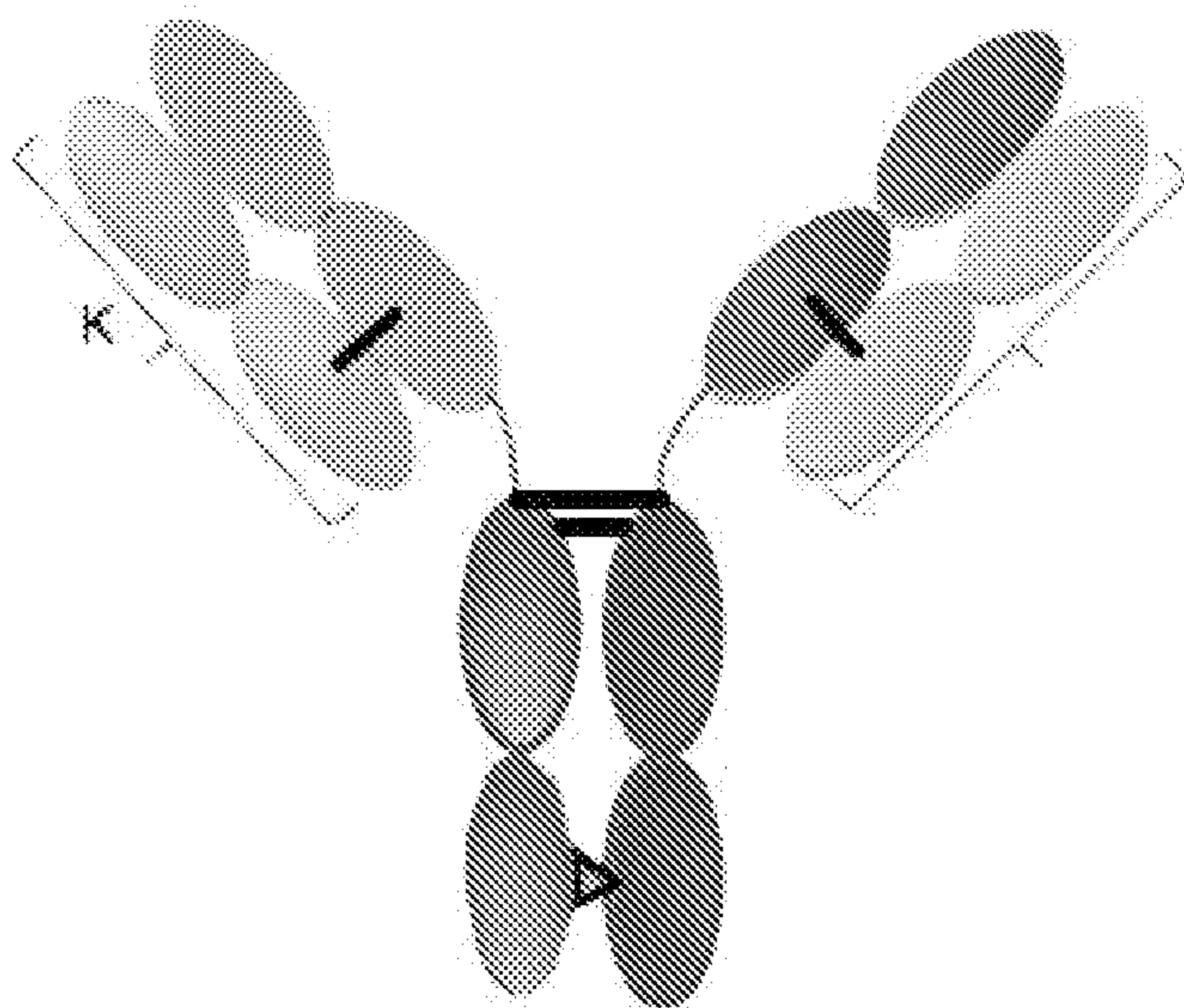


FIG. 50

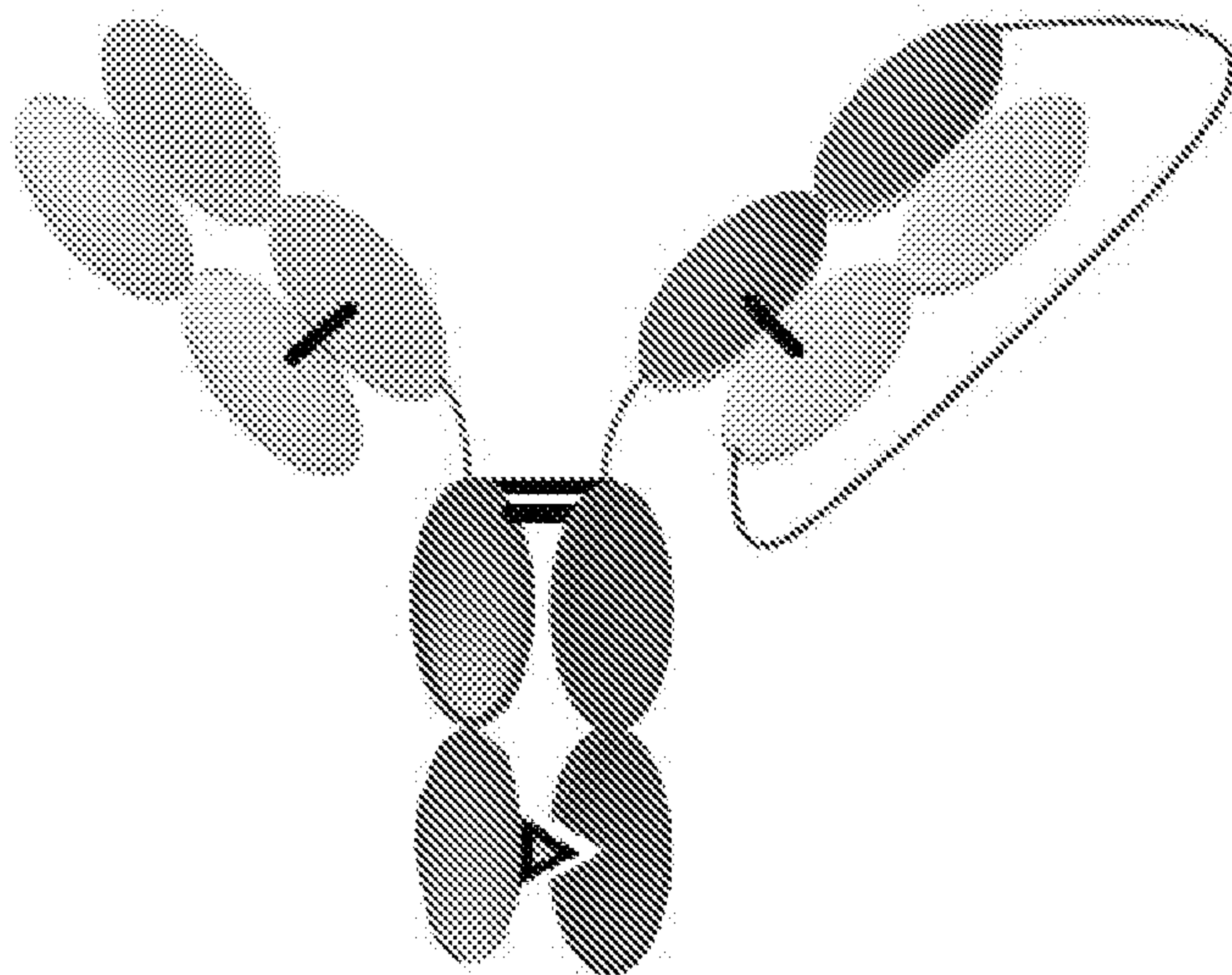


FIG. 51

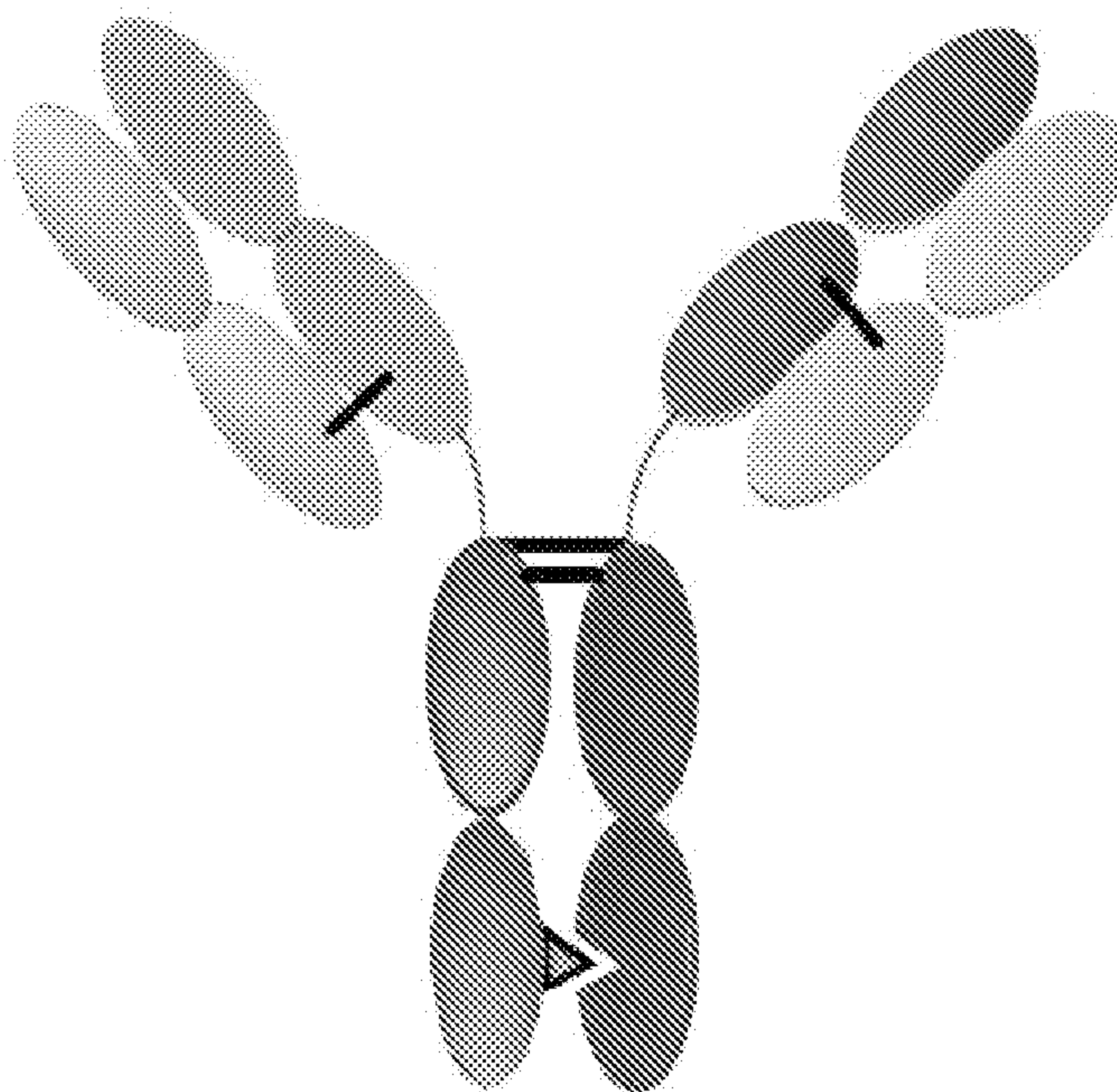


FIG. 52

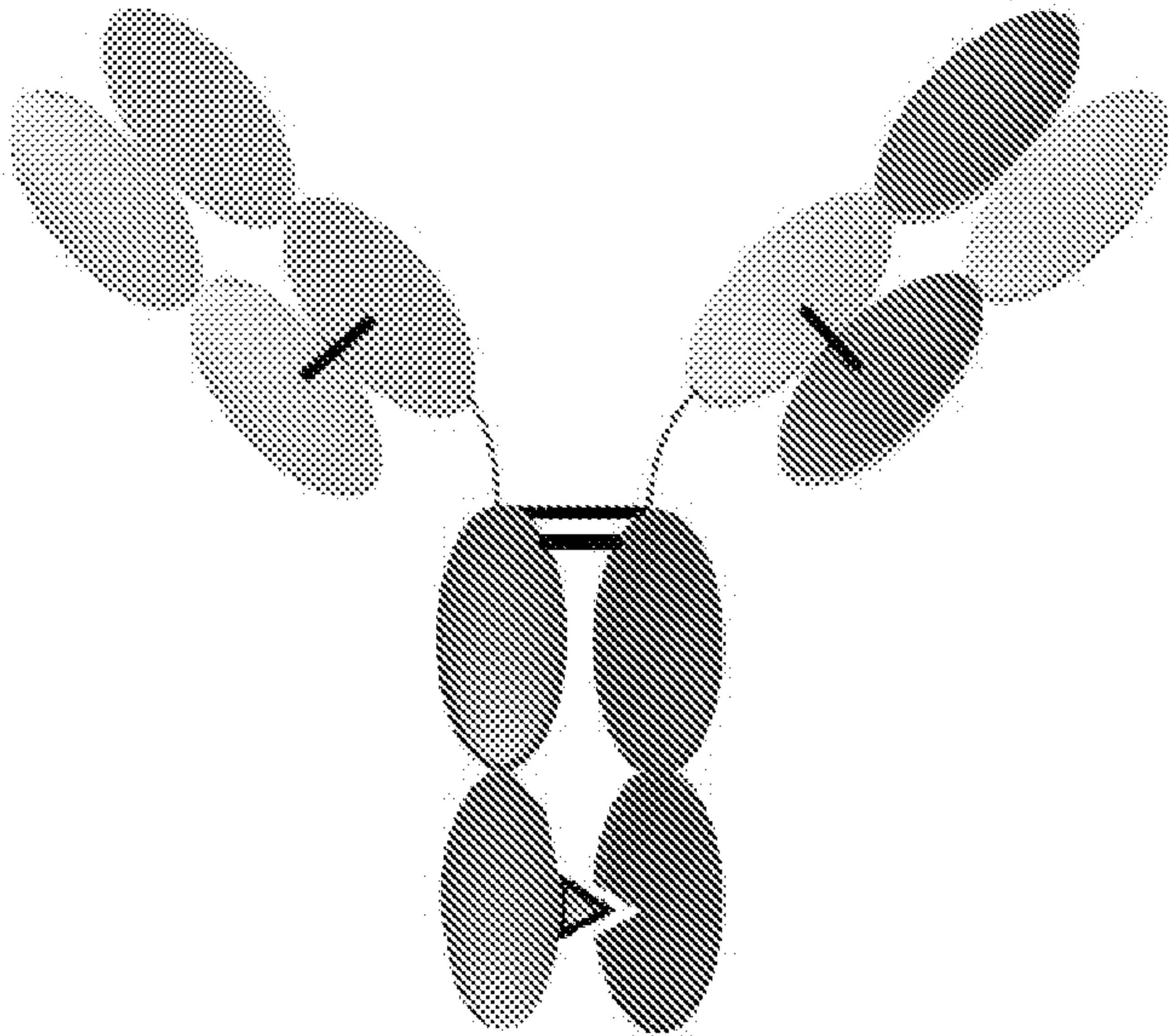
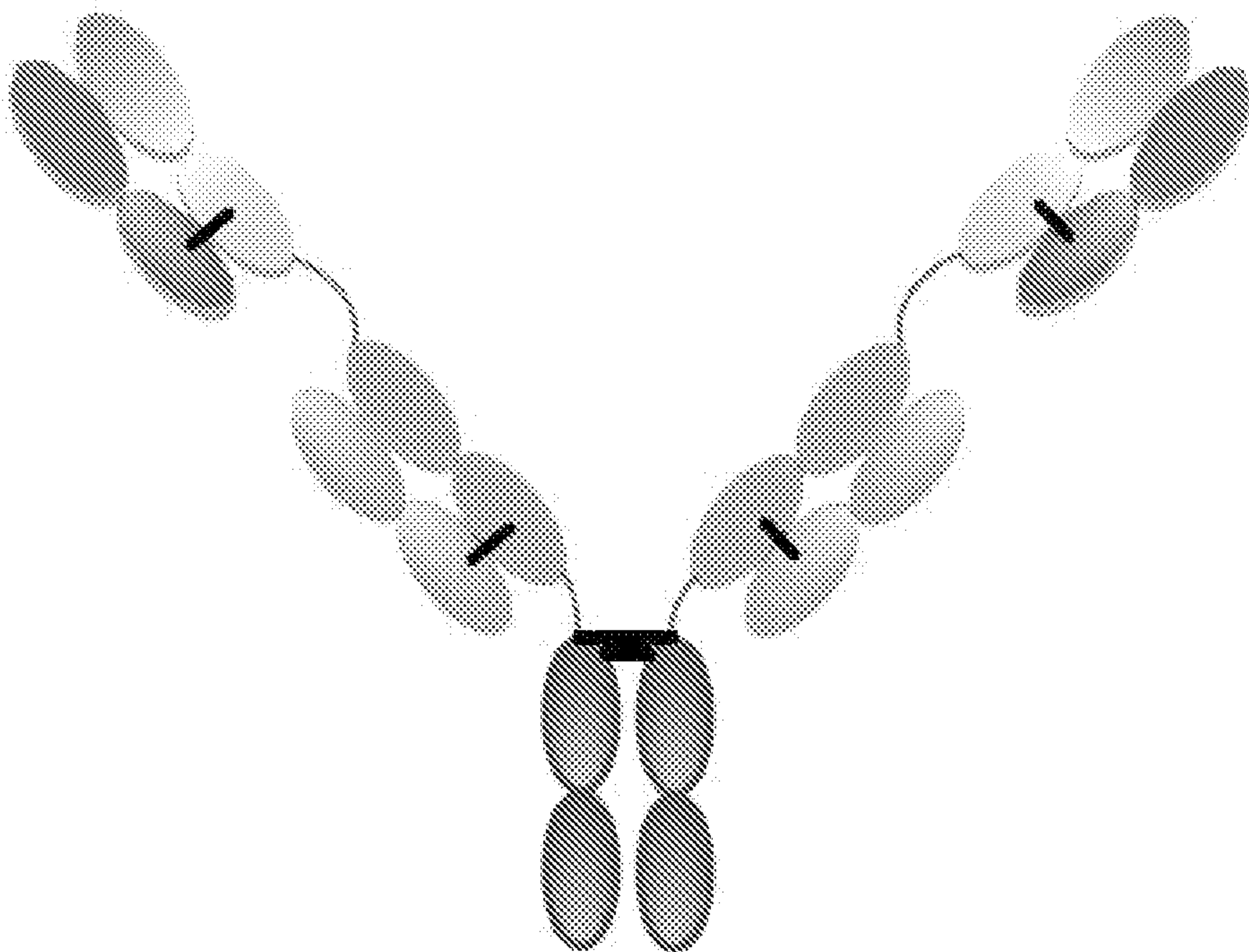


FIG. 53



WO 2018/152518
INTERNATIONAL SEARCH REPORT

PCT/US2018/018771

International application No.
PCT/US 18/18771

A. CLASSIFICATION OF SUBJECT MATTER IPC(8) - A61K 39/00; A61K 39/395; C12N 5/16; C12P 21/04; C12P 21/08; C07K 16/00 (2018.01) CPC - A61K 2039/505; C07K 16/2866; C07K 16/24; C07K 16/28; C07K 2319/00; C07K 2317/24; G01N 33/57492; C07K 2317/56; C07K 2317/569; C07K 2317/60												
According to International Patent Classification (IPC) or to both national classification and IPC												
B. FIELDS SEARCHED												
Minimum documentation searched (classification system followed by classification symbols) See Search History Document												
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched See Search History Document												
Electronic data base consulted during the international search (name of data base and, where practicable, search terms used) See Search History Document												
C. DOCUMENTS CONSIDERED TO BE RELEVANT												
Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.										
X	US 2015/0210765 A1 (ZYNGENIA, INC.) 30 July 2015 (30.07.2015), para [0010], [0041], [0047], [0088], [0114], [0145], [0156], [0223], [0236], [0273], [0432], [0492], and [0802]	1										
Y	US 2010/0272718 A1 (URSO et al.) 28 October 2010 (28.10.2010), Abstract, para [0001], [0006], [0021], [0035], [0036], [0037], [0039], [0041], [0066], [0067], [0070], [0074], [0099], [0120], [0122], [0124], [0126], [0127], [0158], and [0178]	1-4, 15-16										
Y	US 2016/0122432 A1 [INSERM (INSTITUT NATIONAL DE LA SANTE ET DE LA RECHERCHE MEDICALE) et al.] 05 May 2016 (05.05.2016), Abstract, para [0002], [0004], [0006], [0014], [0015], [0018], [0019], [0050], and [0086]	1-4, 15-16										
A	PETRICEVIC et al., Trastuzumab mediates antibody-dependent cell-mediated cytotoxicity and phagocytosis to the same extent in both adjuvant and metastatic HER2/neu breast cancer patients. J Transl Med. 2013, Vol. 11:307. PDF File: pg 1-11. Entire documentation, especially Abstract; pg 2, para 2 and para 3; pg 4, Fig 1 and Fig 2; and pg 8, col 1, para 1	1-4, 15-16										
A	RAAB et al., Fc-Optimized NKG2D-Fc Constructs Induce NK Cell Antibody-Dependent Cellular Cytotoxicity against Breast Cancer Cells Independently of HER2/neu Expression Status. J Immunol. 2014, Vol. 193(8), p. 4261-72. Entire documentation, especially Abstract; and pg 4262, col 1, top para	1-4, 15-16										
A	ROMEE et al., NK cell CD16 surface expression and function is regulated by a disintegrin and metalloprotease-17 (ADAM17). Blood. 2013, Vol. 121(18), p. 3599-608. Entire documentation, especially Abstract	1-4, 15-16										
☒ Further documents are listed in the continuation of Box C. ☐ See patent family annex.												
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Date of the actual completion of the international search 05 April 2018	Date of mailing of the international search report 25 APR 2018											
Name and mailing address of the ISA/US Mail Stop PCT, Attn: ISA/US, Commissioner for Patents P.O. Box 1450, Alexandria, Virginia 22313-1450 Facsimile No. 571-273-8300	Authorized officer: Lee W. Young PCT Helpdesk: 571-272-4300 PCT OSP: 571-272-7774											

FIG. 1

