(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(19) World Intellectual Property Organization
International Bureau

(43) International Publication Date
11 June 2015 (11.06.2015)

WIPO | PCT

(10) International Publication Number
WO 2015/083121 A1

(51) International Patent Classification:
G01N 33/574 (2006.01)

(21) International Application Number:
PCT/IB2014/066602

(22) International Filing Date:
4 December 2014 (04.12.2014)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:
13195770.6 4 December 2013 (04.12.2013) EP

(71) Applicants: **SILICON BIOSYSTEMS S.P.A.** [IT/IT]; Via dei Lapidari, 12, I-40129 Bologna (IT). **FRAUNHOFER-GESELLSCHAFT ZUR FOERDERUNG DER ANGEWANDTEN FORSCHUNG E.V.** [DE/DE]; Hansastrasse 27 C, 80686 Muenchen (DE).

(72) Inventors: **KLEIN, Christoph Andreas**; Eichenstrasse 12a, 93049 Regensburg (DE). **POLZER, Bernhard Michael**; Selma-Lagerlöf-Strasse 36, 81829 Muenchen (DE). **MANARESI, Nicolò**; Via Santo Stefano, 132, I-40125 Bologna (IT).

(74) Agents: **BOSIA, Alessandra** et al.; c/o STUDIO TORTA S.p.A., Via Viotti, 9, I-10121 Torino (IT).

(81) Designated States *(unless otherwise indicated, for every kind of national protection available)*: AE, AG, AL, AM, AO, AT, AU, AZ, BA, BB, BG, BH, BN, BR, BW, BY, BZ, CA, CH, CL, CN, CO, CR, CU, CZ, DE, DK, DM, DO, DZ, EC, EE, EG, ES, FI, GB, GD, GE, GH, GM, GT, HN, HR, HU, ID, IL, IN, IR, IS, JP, KE, KG, KN, KP, KR, KZ, LA, LC, LK, LR, LS, LU, LY, MA, MD, ME, MG, MK, MN, MW, MX, MY, MZ, NA, NG, NI, NO, NZ, OM, PA, PE, PG, PH, PL, PT, QA, RO, RS, RU, RW, SA, SC, SD, SE, SG, SK, SL, SM, ST, SV, SY, TH, TJ, TM, TN, TR, TT, TZ, UA, UG, US, UZ, VC, VN, ZA, ZM, ZW.

(84) Designated States *(unless otherwise indicated, for every kind of regional protection available)*: ARIPO (BW, GH, GM, KE, LR, LS, MW, MZ, NA, RW, SD, SL, ST, SZ, TZ, UG, ZM, ZW), Eurasian (AM, AZ, BY, KG, KZ, RU, TJ, TM), European (AL, AT, BE, BG, CH, CY, CZ, DE, DK, EE, ES, FI, FR, GB, GR, HR, HU, IE, IS, IT, LT, LU, LV, MC, MK, MT, NL, NO, PL, PT, RO, RS, SE, SI, SK, SM, TR), OAPI (BF, BJ, CF, CG, CI, CM, GA, GN, GQ, GW, KM, ML, MR, NE, SN, TD, TG).

Published:

— *with international search report (Art. 21(3))*

— *with sequence listing part of description (Rule 5.2(a))*

(54) Title: METHOD AND KIT FOR DETERMINING THE GENOME INTEGRITY AND/OR THE QUALITY OF A LIBRARY OF DNA SEQUENCES OBTAINED BY DETERMINISTIC RESTRICTION SITE WHOLE GENOME AMPLIFICATION

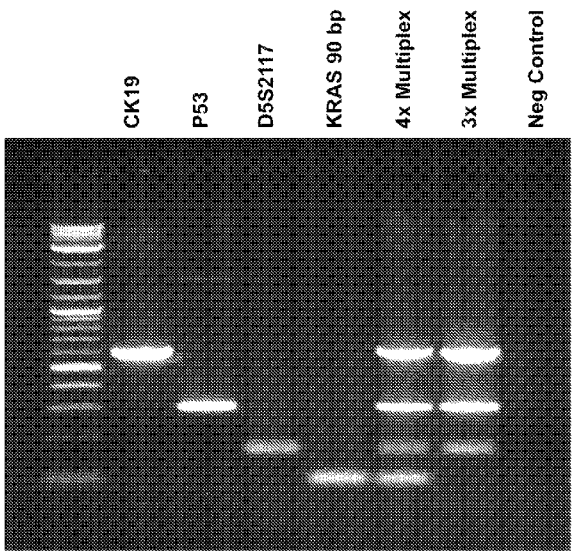

FIG. 1

(57) Abstract: The present invention relates to a method for determining the integrity of the genome of a sample and/or the quality of a library of DNA sequences obtained by deterministic restriction site whole genome amplification (DRS-WGA) of the genome of the sample comprising the steps of : (a) providing the library of DNA sequences; (b) amplifying the library of DNA sequences by PGR using at least one first primer pair, one second primer pair and one third primer pair, the primer pairs each hybridising to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located respectively on a first, second and third chromosome arm, the step of amplifying giving rise to a first, second and third PCR product each from 50 bp to 1000 bp and each having a different size from the others; (c) detecting the first, second and third PCR products; (d) correlating the presence of the first, second and third PCR products with the integrity of the genome of the sample and/or the quality of the library of DNA sequences.

WO 2015/083121 A1

METHOD AND KIT FOR DETERMINING THE GENOME INTEGRITY AND/OR THE QUALITY OF A LIBRARY OF DNA SEQUENCES OBTAINED BY DETERMINISTIC RESTRICTION SITE WHOLE GENOME AMPLIFICATION

* * *

The present invention relates to a method and a kit for determining the integrity of the genome of a sample, in particular a single cell, and/or the quality of a library of DNA sequences obtained by deterministic restriction site whole genome amplification (DRS-WGA) of the genome of the sample.

State of the art

Whole Genome Amplification (WGA) permits detection of somatic mutations and copy alterations in DNA of limited starting material, such as in the case of single circulating tumour cells (CTC) of cancer patients or in preimplantation diagnostics.

For the diagnostic use of WGA for single cell analysis, quality of genomic DNA (i.e. genome integrity) of the single cell sample of interest plays a major role for successful molecular analysis after WGA.

In particular, CTCs have been described as being frequently apoptotic (Mehes, G., et al., Circulating breast cancer cells are frequently apoptotic. Am J Pathol, 2001. 159(1): p.17-20).

Moreover, during caspase-mediated apoptosis genomic

DNA is fragmented into small pieces of 180 bp to 200 bp length (Wyllie, AH., Glucocorticoid-induced thymocyte apoptosis is associated with endogenous endonuclease activation. Nature, 1980. 284(5756): p.555-6).

It is thus important to assess the Genome Integrity status of a single cell, as this can be linked to the biological status of the cell itself, and give clinically relevant information on the overall status of a cancer patient, which goes beyond the information provided by just counting the CTCs and complements the molecular characterization of those CTCs.

Besides, DNA crosslinking and/or fragmentation occur with chemical treatment (e.g. fixation) applied on patient-derived cells and tissues for needed sample conservation after biopsy.

To predict performance of molecular assays for single cell analysis and evaluation of resulting data derived from such samples, assessing the genomic integrity of single cells is of paramount importance.

Available single cell WGA kits assess quality of whole genome amplification by measuring the concentration of the WGA product only. As protocols for these methods include at least one random step during the procedure of single cell DNA amplification, specific assays to evaluate genome integrity of the input sample (in general a single cell)

such as apoptotic or non-apoptotic status, or the quality of the output of the WGA product, such as the suitability for further genetic analysis, are difficult.

A specific kind of WGA is deterministic restriction site whole genome amplification (hereinafter referred to as DRS-WGA). DRS-WGA, which is known from EP1109938 and is commercialised as Ampli1™ by Silicon Biosystems Spa, is based on specific restriction digestion of double stranded DNA at MseI sites (TTAA) and ligation of a universal adaptor for amplification.

DRS-WGA has been shown to be better for the amplification of single cells (see for example: Lee YS, et al: Comparison of whole genome amplification methods for further quantitative analysis with microarray-based comparative genomic hybridization. Taiwan J Obstet Gynecol. 2008, 47(1):32-41) and also more tolerant to DNA degradation due to fixative treatment (see for example: Stoecklein N.H. et al: SCOMP is Superior to Degenerated Oligonucleotide Primed-PCR for Global Amplification of Minute Amounts of DNA from Microdissected Archival Samples. American Journal of Pathology 2002, Vol. 161, No. 1; Arneson N. et al.: Comparison of Whole Genome Amplification methods for analysis of DNA extracted from microdissected early breast lesions in formalin-fixed paraffin-embedded tissue. ISRN Oncol. 2012; 2012;710692).

To date there are no specific assays to evaluate the genome integrity of an input sample from the DRS-WGA product or the quality of the DRS-WGA product obtained.

A need is therefore felt to develop methods and kits allowing to determine the genome integrity of an input sample and/or the quality of the DRS-WGA product obtained and permitting to predict performance of molecular assays downstream of DRS-WGA for single cell analysis and evaluation of resulting data.

An object of the present invention is therefore to provide a method for determining the integrity of the genome of a sample and/or the quality of a library of DNA sequences obtained by DRS-WGA that provides robust and reliable results and allows in particular to assess the biological status of a cell/cells of the sample and/or predict the performance of molecular assays downstream of the DRS-WGA.

This object is achieved by the present invention as it relates to a method as defined in claim 1.

It is a further object of the present invention to provide a kit as defined in claim 8.

Definitions

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which

this invention pertains.

By the term "sample", there is intended a sample comprising at least one particle of a biological entity, said particle comprising at least a DNA sequence representing the genome or a substantial subset of the genome of that biological entity. By way of non-limiting example, said entity may be a human, said at least one particle may be a set of 5 cells or less, a single cell, or a single-cell nucleus, or a haploid germ cell, or a chromosome.

By the term "genome" there is intended the entire genome or said substantial subset of the genome.

By the term "integrity" of the genome there is intended the absence of DNA damages such as double strand breaks, or nicks or similar conditions which may hamper the replication of the genome or its normal functionality.

By the term "quality" of a library of DNA sequences there is intended the suitability of the library of DNA sequences to be used for the genetic characterization of certain features such as, by way of non-limiting example, the presence of point mutations, deletions, insertions, copy number variations.

Brief description of the drawings

Figure 1 shows an agarose gel picture of single marker PCRs, and of a 4-multiplex assay and a 3-multiplex assay

according to preferred embodiments of the present invention;

Figure 2 shows an agarose gel picture of PCRs on PIK3CA hotspot 1 and hotspot2 (M=size marker, 01 = breast13, 02 = breast15, 03 = breast17, 04 = breast20, 05 = prostate14, 06 = prostate16, 07 = prostate24, 08= melanoma13, 09 = melanoma14, 10 = melanoma16);

Figure 3 shows an agarose gel picture of 8 samples tested by the preferred 4-multiplex assay of Figure 1.

Figure 4 shows a histogram of the distribution of the Genome Integrity Index among single Circulating Tumour Cells (CTCs) and single leukocytes (WBCs) from breast cancer patients.

Detailed description of the invention

The method according to the present invention for determining the integrity of the genome of a sample and/or the quality of a library of DNA sequences obtained by deterministic restriction site whole genome amplification (DRS-WGA) of the genome of the sample comprises steps (a) to (d).

By allowing the determination of the integrity of the genome of a sample and/or the quality of a library of sequences obtained by DRS-WGA of the genome of the sample, the method also allows to assess the biological status of of a cell/cells of the sample and/or predict the success

rate of genetic analysis assays on the library of DNA sequences.

The sample preferably consists of 5 cells or less, more preferably the sample is a single cell. The single cell is preferably a circulating tumour cell (CTC), a circulating fetal cell, a circulating endothelial cell (CEC), an oocyte, a cumulus cell, a sperm, a blastomere, or a trophectoderm cell.

In step (a), the library of DNA sequences is provided.

In step (b), the library of DNA sequences is amplified by PCR using at least one first primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp, preferably from 1000 bp to 2000 bp, and corresponding to a sequence of the genome located on a first chromosome arm, the step of amplifying giving rise to a first PCR product from 50 bp to 1000 bp.

Preferably, the DNA sequence of the library to which the first primer pair hybridises encompasses the D5S2117 region of chromosome 5q.

More preferably, the forward primer of the first primer pair is SEQ ID NO:4 and the reverse primer of the first primer pair is SEQ ID NO:3.

In step (c) the first PCR product is detected. Agarose gel electrophoresis may be used to separate and detect the PCR product as well as other methods known in the art.

In step (d) the presence of the first PCR product is correlated with the integrity of the genome of the sample and/or the quality of the library of DNA sequences.

Advantageously, in step (b) at least one second primer pair is used which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp, more preferably from 1000 bp to 2000 bp, and corresponding to a sequence of the genome located on a second chromosome arm other than the first chromosome arm, the step of amplifying giving rise to a second PCR product from 50 bp to 1000 bp having a size other than the first PCR product.

The second PCR product is also detected in step (c) and the presence of the second PCR product is also correlated with the integrity of the genome of the sample and/or the quality of the library of DNA sequences in step (d).

Preferably, the DNA sequence of the library to which the second primer pair hybridises encompasses exons 2 and 3 of the TRP53 gene.

More preferably, the forward primer of the second primer pair is SEQ ID NO:6 and the reverse primer of second primer pair is SEQ ID NO:5.

Advantageously, in step (b) at least one third primer pair is used which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp, more

preferably from 1000 bp to 2000 bp, and corresponding to a sequence of the genome located on a third chromosome arm other than the first and second chromosome arms, the step of amplifying giving rise to a third PCR product from 50 bp to 1000 bp having a size other than the first and second PCR products.

The third PCR product is also detected in step (c) and the presence of the third PCR product is also correlated with the integrity of the genome of the sample and/or the quality of the library of DNA sequences in step (d).

Preferably, the DNA sequence of the library to which the third primer pair hybridises encompasses the KRT19 pseudo-gene 1 (indicated for short as CK19).

More preferably, the forward primer of the third primer pair is SEQ ID NO:8 and the reverse primer of the third primer pair is SEQ ID NO:7.

Even more preferably, in step (b) at least one fourth primer pair is used which hybridises to a DNA sequence of the library having a length from 80 bp to 300 bp, the step of amplifying giving rise to a fourth PCR product from 50 bp to 200 bp having a size other than the first, the second and the third PCR products.

When step (b) uses at least one fourth primer pair, the fourth PCR product is also detected in step (c) and the presence of the fourth PCR product is also correlated with

the integrity of the genome of the sample and/or the quality of the library of DNA sequences in step (d).

Preferably, the DNA sequence of the library to which the fourth primer pair hybridises encompasses codons 12 and 13 of the KRAS gene.

Even more preferably, the forward primer of the fourth primer pair is SEQ ID NO:17 and the reverse primer of the fourth primer pair is SEQ ID NO:18.

According to the present invention there is also provided a kit for determining the integrity of the genome of a sample and/or the quality of a library of DNA sequences obtained by deterministic restriction site whole genome amplification (DRS-WGA) of the genome of the sample comprising:

- at least one first primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located on a first chromosome arm,

- at least one second primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located on a second chromosome arm other than the first chromosome arm,

- at least one third primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to

5000 bp and corresponding to a sequence of the genome located on a third chromosome arm other than the first and second chromosome arms, wherein the first, second and third primer pairs give rise, when amplified by PCR, to PCR products having different size from one another.

Preferably, the kit further comprises at least one fourth primer pair which hybridises to a DNA sequence of the library having a length from 80 bp to 300 bp, wherein the first, second, third and fourth primer pairs give rise, when amplified by PCR, to PCR products having different size from one another.

More preferably, the kit comprises the first, second, third and fourth primer pairs.

The kit may be used for predicting the success of a genetic analysis assay after amplification of single cell genomic DNA by DRS-WGA. Preferably the genetic analysis assay is Sanger sequencing for mutation analysis, assessment of specific copy number changes, quantitative PCR for gene amplification, metaphase Comparative Genomic Hybridisation (CGH), or array Comparative Genomic Hybridisation (CGH).

For gene specific assays, such as Sanger sequencing for mutation analysis, samples with at least 1-2 out of 4 PCR products can be used. Positivity to at least 3 out of 4 PCR products is predictive of successful genome-wide

analysis with metaphase CGH. For array CGH it is advisable to use samples with 4 out of 4 positive PCR products.

The kit may also be used to determine the integrity of the genome of a sample and thus determine the biological status of the cell/cells of the sample. The sample preferably consists of 5 cells or less, more preferably the sample is a single cell.

As a matter of fact, if there is a large number of cells in the sample, there will be, overall, enough template copies for the first, second, third (and optionally fourth) primer pairs to amplify respectively the first, second, third (and optionally fourth) PCR products even if the DNA of the cells of the sample is highly fragmented. The result is the loss of the discrimination power of the method.

On the other hand, it has been experimentally determined that, even in the case of very damaged DNA cell samples, 5 cells is an amount that allows reliable results with the method, i.e allows to obtain the PCR products of step (d) without them being a results of excess template copies.

Further, for the analysis of very damaged DNA cell samples (for which single cells could result in PCR products not being amplified), the use of a number of cells greater than one, i.e. two to five, allows to quantify

genome integrity thus increasing the power of resolution of the method.

Examples

In brief, several primer pairs designed on MseI fragments located on different chromosomal locations and with varying fragment lengths were tested. Three primer pairs that predict successful whole genome analysis of single cell products with high specificity and sensitivity were selected (Example 1). A fourth primer pair of a shorter MseI fragment was added to indicate successful DRS-WGA of low quality cells, e.g. apoptotic CTCs (Example 2). In the examples, PCR products are also referred to as "markers" and PCR amplification reactions including several primer pairs are also referred to as "multiplex assays". In particular, PCR amplification reactions including several primer pairs designed to determine the integrity of the genome of a sample and/or the quality of the library of DNA sequences obtained by deterministic restriction site whole genome amplification (DRS-WGA) of the genome of the sample are also referred to as "quality control assays" or "QC assays".

Example 1

Two alternative marker combinations were shown to predict the success of metaphase comparative genomic hybridization (CGH) after amplification of single cell

genomic DNA with DRS-WGA (Resolution 10-20 Mb) on both cancer cell samples and samples of diploid cells with normal karyotype.

A. Characteristics of the 8 tested PCR markers

PCRs on 8 different MseI-fragments covering an MseI-fragment length from 239-1936 bp were tested (Table 1). Sequences located on 7 different chromosome arms were selected to minimize the chance of a negative assay result because of genomic DNA loss in a single cancer cell.

Pipetting scheme (1x)	1.0 µl	Buffer +dNTPs (10mM MgCl, 100mM Tris (pH 8.5), 500 mM KCl, 1mM dNTPs)
	0.5 µl	Primer 3′ (8 µM)
	0.5 µl	Primer 5′ (8 µM)
	0.25 µl	BSA (for molecular biology)
	7.25 µl	PCR-H_2O
	0.1 µl	Taq polymerase (5U/µl)
	0.5 µl	Ampli1 product (test sample)

Thermal profile

Step 1	94.0°C	2 min
Step 2	Annealing Temp	30 s
Step 3	72.0°C	2 min
Step 4	94.0°C	15 s
Step 5	Annealing Temp	30 s
Step 6	72.0°C	20 s
	14 additional cycles (steps 4-6)	
Step 7	94.0°C	15 s
Step 8	Annealing Temp	30 s
Step 9	72.0°C	30 s
	24 additional cycles (steps 7-9)	
Step 10	72.0°C	2 min
Step 11	4°C	forever

Table 1. Features of the 8 selected PCR primer pairs tested for the QC assay to assess DRS-WGA quality

Primer name	SEQ ID NO	Sequence	Annealing temperature	Chromosome	Mse-fragment length	PCR-fragment length
BCR-TT-R	1	TCAGCCTCAGGACTCTTGTG	61°C	22q	1936 bp	323 bp
BCR-TT-F	2	CGTGGACAACTACGGAGTTG	61°C	22q	1936 bp	323 bp
D5S2117-R	3	ACTGAGTCCTCCAACCATGG	58°C	5q	1376 bp	140 bp*
D5S2117-F	4	CCAGGTGAGAACCTAGTCAG	58°C	5q	1376 bp	140 bp*
TRP53-Ex2/3-R	5	CAGCCCAACCCTTGTCCTTA	58°C	17p	1374 bp	299 bp
TRP53-Ex2/3-F	6	GAAGCGTCTCATGCTGGATC	58°C	17p	1374 bp	299 bp

* This primer pair amplifies a microsatellite sequence. Therefore the actual fragment length can vary slightly between alleles of one patient and between different patients.

Continued

Primer name	SEQ ID NO	Sequence	Annealing temperature	Chromosome	Mse-fragment length	PCR-fragment length
CK19-R	7	TTCATGCTCAGCTGTGACTG	58°C	6q	1146 bp	614 bp
CK19-F	8	GAAGATCCGCGACTGGTAC	58°C	6q	1146 bp	614 bp
IGF2R-R	9	GGATCTTTGGTACCACTCATG	58°C	6q	647 bp	217 bp
IGF2R-F	10	GCCACTGTCGAAGTCTGCA	58°C	6q	647 bp	217 bp
RUFY2-R	11	CAGCTAGGAACTCCAGGAAT CA	64°C	10q	458 bp	104 bp
RUFY2-F	12	GTTGAGGGCTTCATCAACAC CCA	64°C	10q	458 bp	104 bp
SMYD1-R	13	CTTTTCCCTGAAGGTCTTAG	55°C	2p	287 bp	163 bp
SMYD1-F	14	GGGTGACCTGCTTGACATC	55°C	2p	287 bp	163 bp
PHACTR2-R	15	TGTGAGAAAGACTTGGAGTT	58°C	6q	239 bp	205 bp
PHACTR2-F	16	ACTGAACAGAGCAGGTCTAC	58°C	6q	239 bp	205 bp

B. Selection of PCR markers for the QC assay

To select the best possible combination of PCR markers, 72 single cell genomes from different types of human cancers (24 breast disseminated cancer cells (DCCs) 24 prostate DCCs, 24 melanoma DCCs) were re-amplified. For each of the three groups, 12 cells were included which resulted in successful metaphase hybridization in a previous CGH experiment and 12 cells which resulted in a failed metaphase hybridization in a previous CGH experiment. Specific PCRs for all selected markers were performed and the assay was evaluated for accuracy in predicting the outcome of metaphase CGH.

As single markers, PCRs on the long Mse-fragments D5S2117, TRP53-Ex2/3 and KRT19 pseudogene 1 (hereinafter also referred to as CK19) provided best separation between amplified genomes with successful and failed metaphase CGH. An assay of these three fragments showed high assay accuracy in predicting the success of metaphase CGH. The addition of the PCR on the shortest Mse-fragment PHACTR2 slightly increased the assay accuracy in the collective of 72 DCC genomes.

In summary, two alternative QC assays for the QC kit for Ampli1 were developed (Table 2).

Alternative 1: QC assay with 3 markers (D5S2117, TRP53-Ex2/3 and CK19).

Alternative 2: QC assay with 4 markers (D5S2117, TRP53-Ex2/3, CK19 and PHACTR2).

Table 2. Assay accuracy of two alternative marker combinations for the QC assay (72 DCC samples)

Statistical measurement	Alternative 1		Alternative 2	
	2/3 PCRs +	3/3 PCRs +	3/4 PCRs +	4/4 PCRs +
True +	35	29	35	29
False -	1	7	1	7
True -	34	36	35	36
False +	2	0	1	0
Sensitivity	0.97	0.81	0.97	0.81
Specificity	0.94	1.0	0.97	1.0
Positive predictive value	0.95	1.0	0.97	1.0
Negative predictive value	0.97	0.84	0.97	0.84

If possible, only samples that are of the highest quality and that are positive for all selected markers (for both alternative assays 100% specificity) should be used. However, the trade-off for applying this high standard of

quality control is a high rate of false negatives (7/36 = 19.4%). If the number of test samples with the highest quality standard is limited, DRS-WGA amplified genomes with 2/3 or 3/4 positive PCRs still predict a high success rate for metaphase CGH (specificity 0.94 for assay with 3 markers and 0.97 for assay with 4 markers, respectively).

C. Testing of PCR markers on a set of 100 diploid cells with normal karyotype

To further test the set of PCR markers for the QC assay, 100 single cell genomes from diploid cells with normal karyotypes were re-amplified. All samples were tested for the 8 different PCR markers. Additionally, metaphase CGH success was checked for 22 genomes with predicted good quality and 10 genomes with predicted bad quality. The results of the statistical evaluation of assay accuracy are shown in Table 3.

Table 3. Assay accuracy of two alternative marker combinations for the Ampli1 QC kit (32 normal samples)

Statistical measurement	Alternative 1		Alternative 2	
	2/3 PCRs +	3/3 PCRs +	3/4 PCRs +	4/4 PCRs +
True +	22	21	22	21
False -	0	1	0	1
True -	10	10	10	10

False +	0	0	0	0
Sensitivity	1.0	0.95	1.0	0.95
Specificity	1.0	1.0	1.0	1.0
Positive predictive value	1.0	1.0	1.0	1.0
Negative predictive value	1.0	0.91	1.0	0.91

D. Further validation of selected QC markers on DRS-WGA amplified samples

As the tested samples in B and C were taken from an existing biobank of single cell genomes, they were amplified with DRS-WGA customized reagents of the related labs (provided by different suppliers). To validate the performance of the proposed QC assay with samples amplified by Ampli1 kit (as provided by Silicon Biosystems), single mononuclear PBLs and cell pools of a healthy donor and single cells and cell pools of the breast cancer cell line SKBR3 were isolated. The markers proposed in B and C for the QC assay were used to predict quality of the amplified genomes. Then, metaphase CGH experiments for a set of samples (5 single cells, 1 cell pool and 1 Ampli1 negative control) for diploid blood cells as well as SKBR3 cells

were performed to validate the predictive accuracy of the QC assay.

SKBR: 10/11 single cells and both cell pools showed 4/4 positive bands marker PCRs

1/11 cells was negative for all tested markers

negative control clean in all tested PCRs

PBL: 11/11 single cells and both cell pools showed 4/4 positive bands marker PCRs

negative control clean in all tested PCRs

Example 2

As other downstream analyses, e.g. Sanger sequencing for specific fragments, are not dependent on a quantitatively and qualitatively very high amplification of the single cell DNA, a fourth fragment was included. Experiments were carried out to test whether a successful amplification of this sequence only is enough to predict success of Sanger sequencing of comparable Mse fragments (here PIK3CA hotspots 1 and 2).

A. Protocol for 4-Multiplex assay

In order to have compatibility with the already established 3-multiplex assay, new primers were designed for the KRAS Mse-fragment encompassing the frequently mutated nucleotides encoding for codons 12 and 13. These primers amplify a PCR fragment of 91 bp length clearly

distinguishable from the other three bands (D5S2117, CK19 and TP53-Exon2/3, see Figure 1).

The following pipetting scheme and thermal profile were used adding the following KRAS primers in a concentration of 4 µM to the primer mix.

KRAS91bp-F ATAAGGCCTGCTGAAAATGAC (SEQ ID NO:17)

KRAS91bp-R CTGAATTAGCTGTATCGTCAAGG (SEQ ID NO:18)

Pipetting scheme (1x)	1.0 µl	Ampli1™ PCR Reaction Buffer (20mM $MgCl_2$ included)
	0.2 µl	dNTPs (10mM)
	1.0 µl	Primer mix (8 primers, each 4 µM)
	0.2 µl	BSA (20 mg/ml)
	6.5 µl	PCR-H_2O
	0.1 µl	Ampli1™ Taq polymerase (5U/µl)
	1.0 µl	Ampli1™ product (test sample)

Thermal profile

Step 1	95.0°C	4 min
Step 2	95.0°C	30 s
Step 3	58.0°C	30 s
Step 4	72.0°C	90 s
	32 cycles (steps 2-4)	
Step 5	72.0°C	7 min
Step 6	4.0°C	forever

5 μl of each PCR product were loaded on a 1.2% agarose gel. The results were checked by comparing the obtained amplicons bp length with the expected ones, as shown in table 4.

Table 4

PCR product identification			
Target	Primer marker	Chromosome	Amplicon length (bp)
A	KRAS	12p	91
B	D5S2117	5q	108-166
C	TRP53	17p	299
D	KRT19 pseudo-gene 1 (CK19)	6q	614

As already noted above, marker B maps on a polymorphic region, therefore the PCR products may be:

homozygous, and show one band of a bp comprised in the described range (108-166);

heterozygous and show two bands of different bp comprised in the described range (108-166).

Figure 3 shows an example of an agarose gel electrophoresis of PCR products obtained from different samples by using the above disclosed 4 multiplex assay.

Sample 1 in Figure 3 for example displays 3/4 positive markers. Marker B heterozygosity or homozigosity (one or

two bands) must be counted as one in the evaluation of the positivity for marker B. Samples 2 and 8 for example display 4/4 positive markers.

B. Comparison of 4-Multiplex assay with results of single fragment PCR reactions and established 3-Multiplex

72 DRS-WGA libraries of breast, prostate and melanoma DCC that had already been tested by the 3-multiplex assay of Example 1 were selected. Unfortunately the sample Breast 01 was lost due to a fissure in the reaction tube. All other 71 DRS-WGA products were additionally tested with KRAS 91bp single PCR and the new 4-multiplex assay. In summary, 53/71 samples showed the expected band at 91bp in single marker PCR, and all of them but one (Breast 24) showed the same band in the 4-multiplex assay. All the other bands for D5S2117, CK19 and TP53-Exon2/3 were detected in the same samples than with 3-multiplex assay. Moreover, the KRAS 91bp amplicon was detected in all 38 samples that showed one or more bands in the currently used 3-multiplex. Additionally, 15 samples were identified that were negative for D5S2117, CK19 and TP53-Exon2/3 but positive for KRAS91bp, only. Finally 18 samples were negative for all four tested amplicons.

C. PIK3CA sequencing of ten selected samples

Out of the 15 samples which showed the KRAS 91bp amplicon only, ten samples were selected for PIK3CA

sequence analysis (4 breast DCC samples, 3 prostate DCC samples, and 3 melanoma DCC samples). PIK3CA PCRs were performed for HS1 and HS2 fragments, using *Ampli1*™ PIK3CA Seq Kit (Silicon Biosystems SpA, Italy), according to the manufacturer instructions. After PCR all samples were loaded on a 1.5% agarose gel and amplicons visualized after electrophoresis. Strong bands were obtained only for HS1 in four DCC samples (prostate DCC 16, melanoma DCCs 13, 14, and 16) and for HS2 in five DCC samples (prostate DCCs 14, and 16, melanoma DCCs 13, 14, and 16). Additionally, weak but visible bands were detected for an additional two (breast DCC 13, prostate DCC 24) and three samples (breast DCCs 13, and 15, prostate DCC 24), respectively (Figure 2). One sample (melanoma DCC 16) showed a strong smear for DNA fragment ≥ 1kb, which was also visible in all previous gels of this sample. Nevertheless, the PCR amplicon was purified for all samples and sequenced.

Sanger sequencing for PIK3CA mutational hotspots resulted in very strong and clean sequences for the samples showing strong amplification in gel electrophoresis (see Figure 2), including sample melanoma DCC 16. Most of the other samples with lower amplicon concentration showed high background noise, although sequences could easily be edited for two additional samples for HS1 (breast DCC 13, prostate DCC 24) and four additional samples for HS2 (breast DCCs

13, 15, and 20, prostate DCC 24). For the remaining samples background signal was too high for a secure editing of the complete sequence (breast DCCs 15, 17, and 20, prostate DCC 14 for HS1; breast DCC 15 for HS2).

On a collective of CTCs and White Blood Cells (WBCs) harvested with DEPArray™ (Silicon Biosystems SpA) from breast cancer patients peripheral blood enriched with CellSearch® (Jannsen Diagnostics LLC), the Genome Integrity was assessed by analysing the quality of Ampli1™ WGA product according to the invention. The resulting number of PCR products detected following the multiplex PCR reaction (Genome Integrity Index or GII) was found to be significantly skewed, as shown in Figure 4, toward lower values of GII in CTCs with respect to normal WBCs collected from the same patients, undergoing the same process of enrichment sorting and Ampli1™ WGA.

By way of explanation, a GII with a value of 0 corresponds to a situation in which none of the first, second, third or fourth PCR products is amplified, a GII with a value of 1 corresponds to a situation in which only the fourth PCR product is amplified, a GII with a value of 2 corresponds to a situation in which one among the first, second and third PCR products and the fourth PCR product are amplified, a GII with a value of 3 corresponds to a situation in which two among the first, second and third

PCR products and the fourth PCR product are amplified, and a GII with a value of 4 corresponds to a situation in which the first, second, third and fourth PCR products are amplified.

The GII was further used to assess the success rate of targeted Sanger sequencing for PIK3CA exon 9 and exon 20 mutation hotspots, of a custom qPCR assay to determine the amplification of ERBB2 gene, and of array CGH. Table 5 shows a synopsis of the results.

Table 5

Molecular assay	Analyzed cells	Genome Integrity Index (GII)					P Value
		GII 0	GII 1	GII 2	GII 3	GII 4	Chi-square
PIK3CA HS1	n=383	7/23 (30.4%)	14/25 (56.0%)	48/62 (77.4%)	102/117 (87.2%)	146/156 (93.6%)	<0.0001
PIK3CA HS2	n=383	8/23 (34.8%)	18/25 (72.0%)	55/62 (88.7%)	109/117 (93.2%)	149/156 (95.5%)	<0.0001
PIK3CA complete	n=383	4/23 (17.4%)	12/25 (48.0%)	45/62 (72.6%)	97/117 (82.9%)	141/156 (90.4%)	<0.0001
HER2 qPCR	n=351	3/12 (25.0%)	8/18 (50.0%)	41/61 (67.2%)	95/112 (84.8%)	136/148 (91.9%)	<0.0001
aCGH	n=50	Not assessed	Not assessed	4/5 (80.0%)	7/9 (77.8%)	36/36 (100%)	0.016

The above examples show that the method according to the present invention allows to determine the integrity of the genome of a sample and the quality of a library of DNA sequences obtained by DRS-WGA of the genome of the sample with robust and reliable results and allows in particular to predict the performance of molecular assays, such as metaphase and array comparative genomic hybridization (CGH), Sanger sequencing and qPCR, downstream of the DRS-WGA.

From an analysis of the features of the method and kit of the present invention, the resulting advantages are apparent.

In particular, in virtue of the fact that DRS-WGA is based on specific restriction digestion of double stranded DNA at MseI site (TTAA) and ligation of one universal adaptor for amplification, in principle all fragments generated during amplification that represent the WGA library are known. Thus, by the identification of specific MseI fragments having a length of > 1000 bp in a single cell WGA library, the genomic integrity of the DNA of an isolated cell can be measured, and thus the quality of the sample for different kinds of successful molecular analysis determined.

Since the first, second and third primer pairs have been specifically selected in target regions of the genome

corresponding to long amplicons of the DRS-WGA DNA digestion enzyme, MseI, which are more difficult to amplify in case of DNA fragmentation, their amplification is indicative of good overall success of the DRS-WGA on a given sample.

Further, in virtue of the fact that the first, second and third primer pairs are designed on different chromosome arms, the method allows to assess the integrity of the genome of a sample and/or the quality of the library of DNA sequences obtained over the broadest region of the genome.

Further, in virtue of the fact that the first, second, third and fourth PCR products have different size, it is possible to use the first, second, third and fourth primer pairs together in a multiplex reaction.

Moreover, in virtue of the fact that the fourth primer pair hybridises to a DNA sequence of the library having a length from 80 bp to 300 bp, it is possible to predict the performance of molecular assays downstream of the DRS-WGA also for low quality cells, for example apoptotic CTCs. In fact there is a significant difference in performance between the success rate in e.g. targeted sanger sequencing of the PIK3CA exon 9 (30%) and exon 20 (34%) or qPCR for Her2 CNV analysis (25%) of cells for which not even this fourth PCR product is amplified and those cells for which at least this is amplifiable, for which the success rate

roughly doubles (56%, 72%, 50% respectively). The two populations would otherwise be indistinguishable when using only the 3 primer pairs hybridizing to long amplicons of the DRS-WGA.

Finally, it is clear that modifications and variants to the method and kit disclosed and shown may be made without because of this departing from the scope of protection of the appended claims.

In particular, the method may be multiplexed by using further pairs of primers which do not interfere with the PCR amplification with the first, second, third and possibly fourth primer.

CLAIMS

1. A method for determining the integrity of the genome of a sample and/or the quality of a library of DNA sequences obtained by deterministic restriction site whole genome amplification (DRS-WGA) of the genome of the sample comprising the steps of:

(a) providing the library of DNA sequences,

(b) amplifying the library of DNA sequences by PCR using:

- at least one first primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located on a first chromosome arm,

- at least one second primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located on a second chromosome arm other than the first chromosome arm, and

- at least one third primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located on a third chromosome arm other than the first and second chromosome arms,

the step of amplifying giving rise to a first PCR product from 50 bp to 1000 bp, a second PCR product from 50

bp to 1000 bp having a size other than the first PCR product, and a third PCR product from 50 bp to 1000 bp having a size other than the first and second PCR products;

(c) detecting the first, second and third PCR products;

(d) correlating the presence of the first, second and third PCR products with the integrity of the genome of the sample and/or the quality of the library of DNA sequences.

2. The method according to claim 1, wherein the forward primer of the at least one first primer pair is SEQ ID NO:4 and the reverse primer of the at least one first primer pair is SEQ ID NO:3.

3. The method according to claim 1 or 2, wherein the forward primer of the at least one second primer pair is SEQ ID NO:6 and the reverse primer of the at least one second primer pair is SEQ ID NO:5.

4. The method according to any of claims 1 to 3, wherein the forward primer of the at least one third primer pair is SEQ ID NO:8 and the reverse primer of the at least one third primer pair is SEQ ID NO:7.

5. The method according to any of claims 2 to 4, wherein step (b) further uses at least one fourth primer pair which hybridises to a DNA sequence of the library having a length from 80 bp to 300 bp, the step of amplifying giving rise to a fourth PCR product from 50 bp

to 200 bp having a size other than the first, the second and the third PCR products;

- step (c) further comprises detecting the fourth PCR product;

- step (d) further comprises correlating the presence of the fourth PCR product with the integrity of the genome of the sample and/or the quality of the library of DNA sequences.

6. The method according to claim 5, wherein the DNA sequence of the library to which the at least one fourth primer pair hybridises encompasses Codon 12/13 of the KRAS gene.

7. The method according to claim 6, wherein the forward primer of the at least one fourth primer pair is SEQ ID NO:17 and the reverse primer of the at least one first primer pair is SEQ ID NO:18.

8. A kit for determining the integrity of the genome of a sample and/or the quality of a library of DNA sequences obtained by deterministic restriction site whole genome amplification (DRS-WGA) of the genome of the sample comprising:

- at least one first primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located on a first chromosome arm,

- at least one second primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located on a second chromosome arm other than the first chromosome arm, and

- at least one third primer pair which hybridises to a DNA sequence of the library having a length from 1000 bp to 5000 bp and corresponding to a sequence of the genome located on a third chromosome arm other than the first and second chromosome arms,

wherein the first, second and third primer pairs give rise, when amplified by PCR, to PCR products having different size from one another.

9. The kit according to claim 8, further comprising at least one fourth primer pair which hybridises to a DNA sequence of the library having a length from 80 bp to 300 bp,

wherein the first, second, third and fourth primer pairs give rise, when amplified by PCR, to PCR products having different size from one another.

10. Use of the kit according to claim 8 or 9 in predicting the success of a genetic analysis assay after amplification of single cell genomic DNA by DRS-WGA.

11. Use according to claim 10, wherein the genetic analysis assay is selected from the group consisting of

Sanger sequencing for mutation analysis, assessment of specific copy number changes, quantitative PCR for gene amplification, metaphase Comparative Genomic Hybridisation (CGH), and array Comparative Genomic Hybridisation (CGH).

12. Kit according to claim 8 or 9 for use in determining the integrity of the genome of a sample.

13. Kit for use according to claim 12, wherein the sample consists of 5 cells or less.

14. Kit for use according to claim 13, wherein the sample is a single cell.

15. Kit for use according to claim 13 or 14, wherein determining the integrity of the genome of the sample allows to determine the biological status of the cell/cells of the sample.

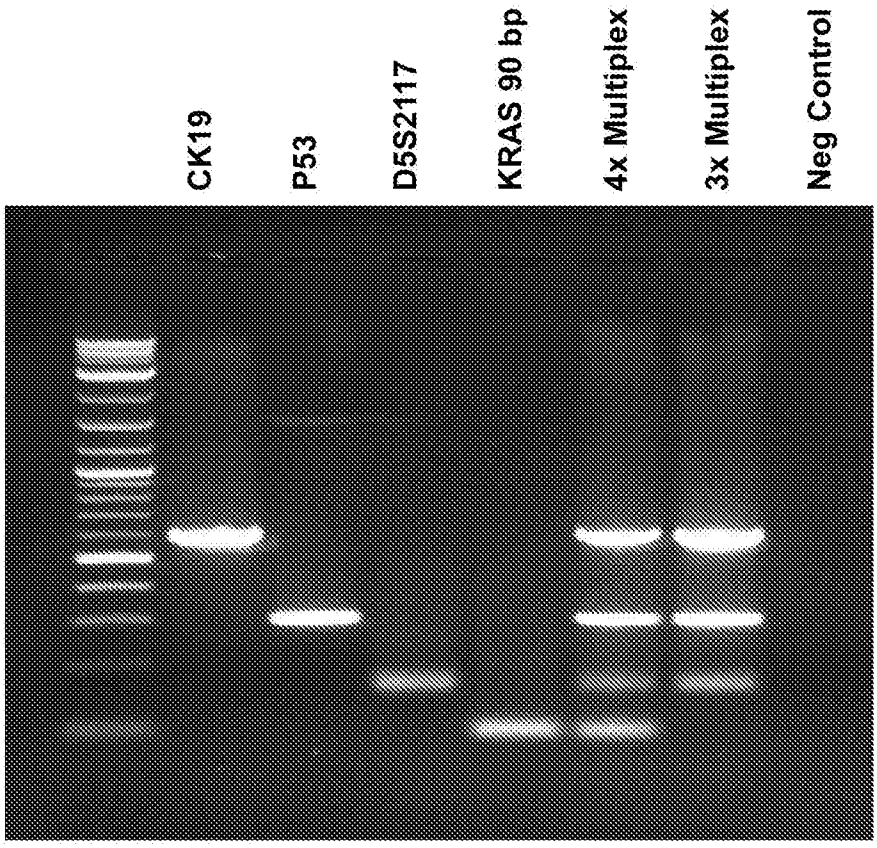
CK19
P53
D5S2117
KRAS 90 bp
4x Multiplex
3x Multiplex
Neg Control

FIG. 1

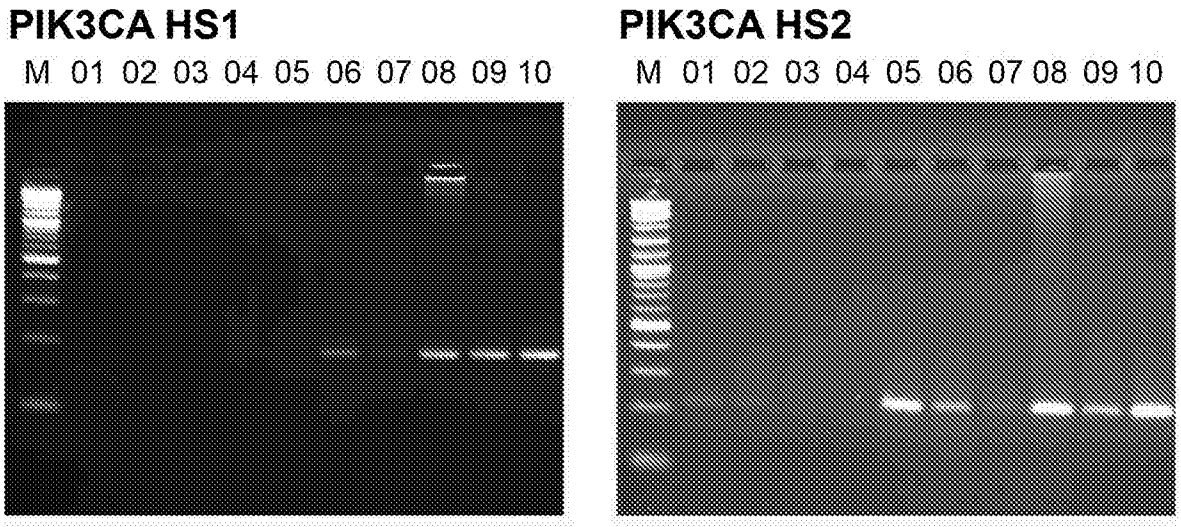
PIK3CA HS1
M 01 02 03 04 05 06 07 08 09 10
PIK3CA HS2
M 01 02 03 04 05 06 07 08 09 10

FIG. 2

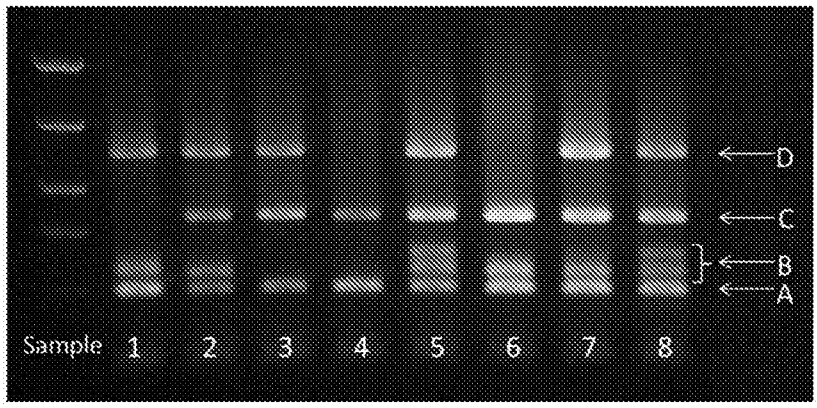
D
C
B
A
Sample 1 2 3 4 5 6 7 8

FIG. 3

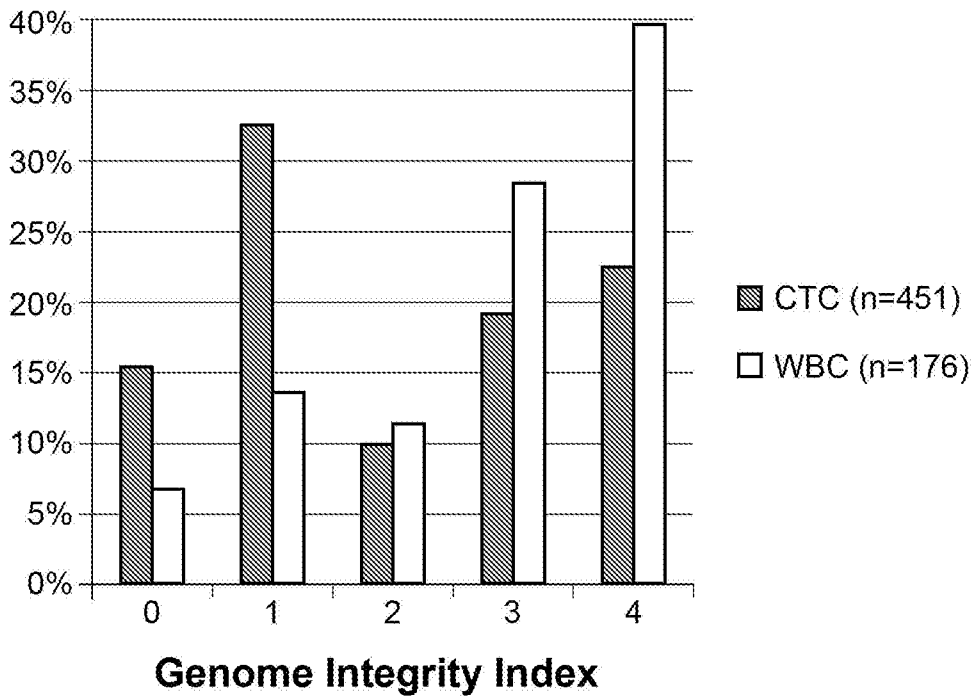
40%
35%
30%
25%
20%
15%
10%
5%
0%
0 1 2 3 4
Genome Integrity Index
CTC (n=451)
WBC (n=176)

FIG. 4

INTERNATIONAL SEARCH REPORT

International application No
PCT/IB2014/066602

A. CLASSIFICATION OF SUBJECT MATTER
INV. G01N33/574
ADD.

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)
G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
EPO-Internal, WPI Data

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	E H VAN BEERS ET AL: "A multiplex PCR predictor for aCGH success of FFPE samples", BRITISH JOURNAL OF CANCER, vol. 94, no. 2, 30 January 2006 (2006-01-30), pages 333-337, XP055098207, ISSN: 0007-0920, DOI: 10.1038/sj.bjc.6602889	1,8-15
A	abstract page 33, column 1 - column 2 page 335, column 1 - column 2	2-7

-/--

[X] Further documents are listed in the continuation of Box C.

[X] See patent family annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

"E" earlier application or patent but published on or after the international filing date

"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)

"O" document referring to an oral disclosure, use, exhibition or other means

"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone

"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art

"&" document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
5 February 2015	13/02/2015

Name and mailing address of the ISA/	Authorized officer
European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Fax: (+31-70) 340-3016	Celler, Jakub

2

Form PCT/ISA/210 (second sheet) (April 2005)

C(Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 2010/093465 A1 (CARIS MPI INC [US]; VON HOFF DANIEL D [US]; LOESCH DAVID M [US]; ALARC) 19 August 2010 (2010-08-19) abstract paragraphs [0007], [0014], [0015], [0349]; claims 6,7,11	1-15
Y	LEE Y S ET AL: "Comparison of Whole Genome Amplification Methods for Further Quantitative Analysis with Microarray-based Comparative Genomic Hybridization", TAIWANESE JOURNAL OF OBSTETRICS AND GYNECOLOGY, ELSEVIER (SINGAPORE) PTE LTD, HONG KONG BRANCH, HONG KONG, HK, vol. 47, no. 1, 1 March 2008 (2008-03-01), pages 32-41, XP025480394, ISSN: 1028-4559, DOI: 10.1016/S1028-4559(08)60052-2 [retrieved on 2008-03-01]	1,8-15
A	abstract page 33, column 2, paragraph t page 34, column 1, line t	2-7
Y	BUFFART TINEKE E ET AL: "DNA quality assessment for array CGH by isothermal whole genome amplification", CELLULAR ONCOLOGY, IOS PRESS, LONDON, GB, vol. 29, no. 4, 1 January 2007 (2007-01-01), pages 351-359, XP009175704, ISSN: 1570-5870	1,8-15
A	abstract page 351, column 2	2-7

2

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No
PCT/IB2014/066602

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 2010093465 A1	19-08-2010	CA 2751888 A1	19-08-2010
		CN 102439454 A	02-05-2012
		EP 2396661 A1	21-12-2011
		JP 2012517238 A	02-08-2012
		WO 2010093465 A1	19-08-2010