



INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

(51) International Patent Classification ⁶ : C12N 5/00	A1	(11) International Publication Number: WO 95/08621 (43) International Publication Date: 30 March 1995 (30.03.95)
(21) International Application Number: PCT/US94/10736 (22) International Filing Date: 21 September 1994 (21.09.94) (30) Priority Data: 124,676 22 September 1993 (22.09.93) US (71) Applicant: CREATIVE BIOMOLECULES, INC. [US/US]; 45 South Street, Hopkinton, MA 01748 (US). (72) Inventors: MARSHALL, Paul, G.; 35 North Main Street, Berkley, MA 02779 (US). GUERTIN, Patrick, M.; 44 Miscoe Road, Mendon, MA 01756 (US). (74) Agent: JOHNSTON, Madeline, I.; Testa, Hurwitz & Thibeault, Exchange Place, 53 State Street, Boston, MA 02109 (US).		(81) Designated States: AM, AU, BB, BG, BR, BY, CA, CN, CZ, EE, FI, GE, HU, JP, KG, KP, KR, KZ, LK, LR, LT, LV, MD, MG, MN, NO, NZ, PL, RO, RU, SI, SK, TJ, TT, UA, UZ, VN, European patent (AT, BE, CH, DE, DK, ES, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, ML, MR, NE, SN, TD, TG), ARIPO patent (KE, MW, SD, SZ). Published <i>With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.</i>
(54) Title: LIPID-MODIFIED SERUM FREE MEDIA (57) Abstract Methods are provided for producing serum-free, animal cell culture media which have enhanced cell culture properties, such as cell growth, cell viability and protein production properties. The cell culture properties of standard serum-free media formulations, which include compositions such as inorganic salts, glucose, fatty acids, amino acids and vitamins, can be enhanced according to the methods of the invention, by providing the media with an additive. The additive in one embodiment can be produced by the acid or base catalyzed hydrolysis of a phosphoglyceride.		

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- 1 -

LIPID-MODIFIED SERUM FREE MEDIAField of Invention

The present invention relates generally to ex vivo cell cultivation, and more particularly, to cell
5 culture media and methods for their production.

Background of the Invention

When a cell is removed from its original tissue or
10 organism and placed in culture, the medium must provide all the environmental conditions that the cell has been exposed to in vivo; only then will it be able to survive, to proliferate, and to differentiate or maintain its differentiated phenotype. Thus,
15 extracellular medium must meet the essential requirements for survival and growth (i.e. must provide nutritional, hormonal, and stromal factors). Where cells are cultured as part of a system to produce biologicals, e.g., for the production of vaccines or
20 recombinant protein, the culturing medium also should not interfere with the cell's production of the desired biological. Among the biological fluids that have proved successful for culturing cells outside the body, serum has gained the most widespread significance.

25

However, there are a number of disadvantages with serum-containing media, particularly as part of a recombinant biologicals production system, including difficulties with sterilization, inconsistent variation
30 between serum batches, the presence of extraneous serum

- 2 -

constituents including indefinable and potentially cytotoxic constituents from which the biological product of interest must be purified, and risk of contaminants. Accordingly, the art long has sought to
5 create non-serum culture media (including "serum-free" and "chemically-defined" media) which provide the necessary environmental conditions for cell growth and viability. Defining components necessary for a non-serum culturing media has been an on-going effort in
10 the art. In many non-serum culture media, cell growth often is slower, and cell density and saturation levels, as well as cell viability, may be diminished. In addition, media formulations found useful for cell growth often are not optimal for recombinant protein
15 production, requiring a change in culture media when protein production is to be induced, or a compromise in desired protein production levels.

Mizrahi and Lazar (Cytotechnology (1988) 1:199-214)
20 describes the general state of the art of serum-free, chemically defined media for ex vivo mammalian cell cultivation.

Along with an energy source, a nitrogen source,
25 vitamins, inorganic salts, nucleic acid precursors, and oxygen, fats (lipids) and fat-soluble components are critical elements for non-serum containing culture medium. Among the lipids found to be useful in serum-free or chemically defined media are
30 phospholipids commonly associated with the cell membrane, and various lipid and phospholipid precursors. Goodwin, et al (1990) Nature 347: 209-210 describes lipid supplements useful in serum-free media. Bromke, et al (1986) J. Microbiol, Methods, 6:55-59
35 describe the use of cholesterol and oleic acid in serum-

- 3 -

free media. Imagawa et al., (1989) PNAS 86:4122-4126 describe the use of the lipid precursors dilinoleoyl phosphatidic acid and phosphatidyl serine to stimulate growth of epithelial cells in a serum-free medium.

- 5 Bashir et al. (1992) In Vitro Cell Dev. Biol. 28A:663-668 describes the use of the lipid precursors phosphatidic acid and lysophosphatidic acid to stimulate the growth of kidney cells in serum-free media.

10

International application PCT/US90/03430, published April 15, 1990, discloses a serum-free cell culture medium for enhanced cell growth that includes the phospholipid precursors choline, ethanolamine,

- 15 phosphatidyl choline and phosphatidyl ethanolamine.

Miyazaki et al. (1991) Res. Exp. Med. 191:77-83

describe the use of phosphatidyl ethanolamine and phosphatidyl choline, as well as the phospholipid precursors ethanolamine and phosphoethanolamine, to

- 20 prolong survival of rat hepatocytes in culture. Kovar (1987) Folia Biologica 33:377-384 describes the use of dipalmitoyl lecithin, cholesterol and linoleic acid to promote the growth of hybridomas in serum-free media.

- 25 It is an object of the invention to provide an improvement in serum-free cell culture media. Another object is to provide a cell culture media, and methods for its production, having improved cell cultivation properties, including enhanced cell growth and cell
- 30 viability, and enhanced production of recombinant biologicals. Another object of this invention is to provide serum-free media having enhanced cell culture properties and which can be utilized in a wide range of cell culture systems for the recombinant production of
- 35 biologicals, including proteins, using any of a number

- 4 -

of cell lines and recombinant protein expression systems known in the art. Still another object of the invention is to provide a media formulation having particular utility for the recombinant production of
5 bone morphogenic proteins. It is a further object of the invention to provide methods for consistently and reproducibly producing such serum-free media having improved cell cultivation properties.

10 These and other objects and features of the invention will be apparent from the description, figures, and claims which follow.

Summary of the Invention

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An improvement in serum-free (non-serum containing) cell culture media having enhanced cell cultivation properties, including enhanced cell growth, cell viability and recombinant biologicals production, now
20 has been discovered. The serum-free media formulations of the invention have particular utility in animal cell culturing systems for the production of biologicals of interest, including recombinant proteins. As described herein, the serum-free media formulations of the
25 invention include the elements provided in standard serum-free media formulations, including an energy source such as glucose, inorganic salts, fat soluble components, a nitrogen source and vitamins. In addition, and as disclosed herein, the serum-free media
30 formulations of the invention also include an additive which unexpectedly has a synergistic effect, when combined with a standard formulation, to enhance significantly the overall cell culture properties of the medium.

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- 5 -

As used herein "serum-free" means a culture medium formulated in the absence of serum, and includes both the media formulations defined in the art as "serum-free" media (which may otherwise be protein supplemented), "protein-free" media, (no protein supplementation) and "chemically defined" (ultra-pure with or without small molecular constituents, genetically engineered peptides or proteins).

10 In one embodiment, to improve the cell culture properties of the media, the serum-free media are provided with an additive having the formula:

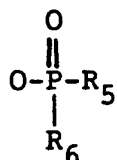


20 wherein one of R_1 or R_2 is OH, and another of R_1 and R_2 is OH or:



wherein R_4 is a C_1 - C_{26} hydrocarbon, preferably a C_{10} - C_{24} fatty acid. Additionally, R_3 is

30



35

wherein one of R_5 and R_6 is O^- , and the other is an alcohol, and the phosphate is esterified to the hydroxyl group of the alcohol. Examples of common alcohol moieties for one of R_5 and R_6 include serine, ethanolamine, choline, inositol and other alcohols commonly associated with membrane phospholipids.

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- 6 -

In another embodiment, the additive comprises a lysophosphatidyl ester formed as a degradation product of a membrane phosphoglyceride ester.

5 As used herein, "phosphoglyceride ester" is a substituted glycerol, wherein the hydroxyl groups at C₁ and C₂ each are esterified to the carboxyl group of a fatty acid, and the C₃ hydroxyl group is esterified to a phosphate (PO₃²⁻).

10

Exemplary phosphoglyceride esters include phosphatidyl ethanolamine, phosphatidylcholine, phosphatidylserine and phosphatidylinositol and other phospholipids that comprise part of a cell membrane.

15

As used herein, a lysophosphatidyl ester is a phosphoglyceride ester hydrolyzed to release at least one of its two fatty acid chain components, (e.g., a C₁-C₂₆ hydrocarbon). Exemplary lysophosphatidyl esters
20 useful as additives in the serum formulations of the invention include lysophosphatidyl ethanolamine, lysophosphatidyl choline, lysophosphatidyl serine and other lysophosphatidyl ester hydrolysis products of phospholipids which comprise part of a cell membrane.

25

In another embodiment, the media formulations are made by adding to the media hydrolysis products of a phosphoglyceride ester composition. Exemplary phosphoglyceride ester compositions include
30 compositions containing phosphoglyceride esters typically associated with a cell membrane, including, without limitation, phosphatidyl choline, phosphatidyl ethanolamine, phosphatidyl serine, and phosphatidyl inositol.

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- 7 -

The additive may be formed, in one embodiment, by the hydrolysis of one or more commercially available phosphoglyceride esters, including commercially available phosphatidyl serine, phosphatidyl ethanolamine, phosphatidyl choline, or phosphatidyl inositol compositions. The phosphoglyceride esters may be hydrolyzed by an acid hydrolysis or by base saponification using standard methodologies well known in the art. The phosphoglyceride may be hydrolyzed in one embodiment by heating the phosphoglyceride ester in an acid or base solution to temperature greater than 40°C. In another preferred embodiment, the ester is heated to a temperature within the range of 45-90°C, preferably 50-70°C, for at least 15 minutes, preferably at least 20 minutes. The concentration of the acid or base in the solution may be adjusted to between about 0.05 M to 10 M of the acid or base. In another embodiment the phosphoglyceride ester is acid hydrolyzed in a solution having a pH in the range of about 0.2-4, preferably in the pH range of 0.2-2. In another embodiment, the phosphoglyceride ester is base saponified in a solution having a pH in the range of about 8-14, preferably in the pH range of 10-14. The acid hydrolysis can be conducted using acids such as hydrochloric acid, sulfuric acid, phosphoric acid or nitric acid, either in an aqueous solvent or in an organic solvent, such as ethanol. The base saponification of the phosphoglyceride ester may be conducted using a base such as sodium hydroxide, potassium hydroxide or ammonium hydroxide, which also may be water or alcohol-based.

When the hydrolysis products of a phospholipid composition are added to standard serum-free culture media, the resulting media demonstrate unexpected

- 8 -

enhanced cell culture properties, including enhanced cell growth, cell viability and recombinant biologicals production, as compared with the media formulated without the additive. Inclusion of the additive has an
5 unexpected synergistic effect, when combined with a standard formulation, to enhance the ex vivo culture properties of the cells.

In another embodiment of the invention, a
10 preferred media formulation is a formulation that includes the hydrolysis products of one or more phospholipid compositions, and a full complement of amino acids.

15 In still another embodiment of the invention, a preferred media formulation is one that includes the hydrolysis products of one or more phospholipid compositions, and a phospholipid precursor.

20 The improved cell culture media are reproducible and easy to make, and consistently demonstrate enhanced cell growth and recombinant protein production properties. The improved media thus allow the artisan to use one media formulation for both cell culturing
25 and recombinant biologicals production, including recombinant protein production, without a need to change formulations or otherwise compromise recombinant protein production yields.

30 The improved cell culture formulations of the invention have utility for both vertebrate and invertebrate cell culture systems, including, without limitation, human, bovine, equine, primate, and other mammalian cells, non-mammalian cells, such as xenopus
35 cell systems, and drosophila, and other insect

- 9 -

(invertebrate) cells, as well as biosynthetic hybrid cell lines, including, for example, hybridomas. The improved cell culture formulation also has utility for the production of a variety of biologicals, including
5 natural-sourced molecules, vaccines and recombinant proteins, such as antibodies, growth factors, members of the blood cascade, cytokines and morphogenic proteins, including true bone morphogens such as those described in U.S. Patent No. 5,011,691.

10

The cell culture media formulations of the invention have particular utility in the culturing of chinese hamster ovary (CHO) cell lines, for the recombinant productions of true tissue morphogens such
15 as OP1 and other natural-sourced, recombinantly produced or biosynthetic tissue morphogens known in the art and described, for example, in 5,011,691.

The foregoing and other objects, features and
20 advantages of the present invention will be made more apparent from the following detailed description of the invention.

Brief Description of the Figures

The foregoing and other objects and features of this invention, as well as the invention itself, may be
5 more fully understood from the following description, when read together with the accompanying drawings, in which

Figure 1 is a graph of viable cell density versus
10 day of culture for CHO-1 cells;

Figure 2 is a graph of viable cell density versus day of culture for CHO-1 cells;

15 Figure 3 is a graph of OP-1 production versus day of culture by CHO-1 cells;

Figure 4 is a graph of viable cell density versus day of culture for CHO-2 cells; and
20

Figure 5 is a graph of OP-1 production versus day of culture for CHO-2 cells.

- 11 -

Detailed Description

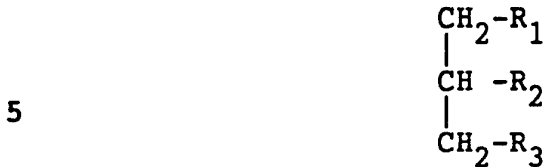
The invention provides an improvement in serum-free cell culture media having enhanced cell growth, cell viability and/or biologicals production properties. The improved media comprise a membrane lipid phosphoglyceride ester degradation product. To date, the art has sought to add lipids and lipid precursors to enhance the cell cultivation properties of serum-free medium. It now has surprisingly been discovered that phospholipid degradation products have unexpected synergistic effects, when combined with standard formulation, to produce media with enhanced cell culture properties. The additive is prepared easily as described herein and can be used in a range of media for a variety of cells and cell culture systems.

Presented below is a detailed description of the improvement in the cell culture media, and methods for their production, as well as general considerations for culture media formulation in which the additive is useful, detailed descriptions on how to make and use the additive and cell culture media of the invention, and several non-limiting examples demonstrating the utility of the improved media for enhancing cell growth, cell viability and recombinant protein production of cultured cells.

A. Serum-Free Cell Culture Media Additive and Methods for its Production.

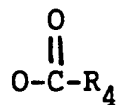
In one embodiment, a serum-free medium having improved cell cultivation properties is provided by including in the medium an additive having the formula:

- 12 -



wherein one of R_1 or R_2 is OH, and another of R_1 or R_2 is OH or:

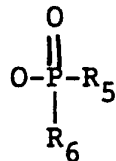
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15

wherein R_4 is a C_1 - C_{26} hydrocarbon. The hydrocarbon preferably is long chain (C_{10} - C_{24}) fatty acid having a structure found in membrane lipids. Exemplary fatty acids found in cell membranes include saturated fatty acids such as lauric acid (C_{12}), myristic acid (C_{14}), palmitic acid (C_{16}), steric acid (C_{18}), arachidic acid (C_{20}), lignoceric acid (C_{24}), and unsaturated fatty acids such as palmitoleic acid (C_{16}), oleic acid (C_{18}), linoleic acid (C_{18}), linolenic acid (C_{18}), and arachidonic acid (C_{20}). Additionally, in the additive, R_3 is

30



wherein one of R_5 and R_6 is O^- , and the other is an alcohol esterified to the phosphate group through its hydroxyl moiety. In a preferred embodiment, one of R_5 and R_6 is an alcohol normally associated with membrane phospholipids. Exemplary alcohols include serine, ethanolamine, choline and inositol. The additives of this formula are referred to herein as lysophosphoglyceride esters.

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- 13 -

In another embodiment, the additive comprises one or more hydrolysis products of a phospholipid ester, preferably a phospholipid ester that normally is associated with a cell membrane including, for example, 5 phosphatidyl choline (PC), phosphatidyl ethanolamine (PEA), phosphatidyl inositol (PI), phosphatidyl serine (PS) and the like.

The additive may be formed in one embodiment by 10 the acid hydrolysis or base saponification of a phosphoglyceride ester as described in detail below. Alternatively, the additive may be formed by enzymatic hydrolysis of a phosphoglyceride ester, e.g., using a phospholipase available in the art such as phospholipase 15 A₂, a phospholipase which selectively hydrolyzes the 2-acyl bond of 3-n-phosphoglycerides. Uthe et al., Can. J. Biochem., 49:776 (1971).

20 A.1. Acid Hydrolysis

Phosphoglyceride esters can be acid hydrolyzed using standard methodologies well known in the art to produce additives useful in the media formulations of the invention. Currently preferred is to hydrolyze the 25 phosphoglyceride ester by heating it in an acid solution to a temperature of at least about 40°C, preferably in the range of about 45°-90°C, most preferably 50-70°C, for at least 10 minutes, preferably at least 20 minutes. Heating the solution for longer 30 than 90 minutes does not appear to provide additional efficacy. As will be appreciated by those having ordinary skill in the art, hydrolysis also will occur at room temperature, but this requires an extended period of time (e.g., at least several days, sometimes 35 greater than 7-14 days, and complete hydrolysis can not

- 14 -

be guaranteed by this means). The currently most preferred methodology is to heat the solution for between about 30-50 minutes at a temperature within the range of about 60°-65°C. The acidic solution may be
5 created by exposing the phosphoglyceride ester to a solution having a pH within the range of about 0.2-4, preferably in the range of 0.2-2. In one embodiment the acidic solution is made by adding an acid to a final concentration of between about 0.5 M to 10 M of
10 the acid in the solution. Acids such as hydrochloric acid, sulfuric acid, phosphoric acid or nitric acid may be utilized to advantage. The hydrolysis may be conducted in aqueous solvents, or in an alcohol such as ethanol. For example, a medium having enhanced cell
15 culture properties can be prepared in one embodiment by adding phosphatidyl choline to a volume of 200 proof ethanol and about a 1:10 volume of 1 N (1M) HCl. The solution then may be heated on a water bath for 30 to 45 minutes at 60°C to 65°C. Subsequently, an
20 approximately 1:1000 volume of the hydrolyzed phosphatidyl choline solution per liter of medium is added directly to the serum-free medium, to produce a medium having improved cell culture properties.

25 A.2. Base Saponification.

The phosphoglyceride ester also may be hydrolyzed by base saponification, e.g., by heating the phosphoglyceride ester in a basic solution at a
30 temperature greater than 40°C for at least about 10 minutes, preferably at least 20 minutes. Heating the solution for more than 90 minutes does not appear to provide additional efficacy. Currently preferred is heating to a temperature within the range of 50-70°C
35 for 30-50 minutes. The basic solution may be created

- 15 -

by exposing the phosphatidyl ester to a solution having a pH within the range of about 8-14, preferably 10-14. In one embodiment, the basic solution may be formulated by adjusting the solution to a concentration of 0.05 M
5 to 10.0 M of a base such as sodium hydroxide, potassium hydroxide or ammonium hydroxide.

The hydrolyzed phospholipid then can be added to the medium as is, and the mixture sterilized as
10 described below.

B. Cell Culture Media-General Considerations

The cell culture properties of a wide range of
15 serum-free media formulations known in the art may be improved by addition of the additive disclosed herein. As will be appreciated by those having ordinary skill in the art, many individualized media formulations have been developed over the years to maximize cell growth,
20 cell viability and/or biologicals production for a given cultured cell or cell line. These media compositions differ from one another, for example, in the number, type and concentration of growth factors, antibiotics and amino acid complements added to the
25 media, and may also include components such as protease inhibitors, and one or more anti-foaming agents, particularly where cells are grown in suspension. Other individualized components include additives that enhance recombinant protein production. For example,
30 where the host cell was developed for gene amplification of a recombinant protein, the selectable marker DHFR (dihydrofolate reductase) typically comprises part of the transfected host cell as a selectable marker, and methotrexate is included in the
35 culture medium. Insulin or other growth factors such as IGF that play a role in the energy source cascade also often are included to enhance cell growth.

- 16 -

The additive disclosed herein is anticipated to have a synergistic effect on individualized media formulations as well as in standard formulations.

5

All serum-free media formulations include essential components which permit cell growth, including (1) an energy source, typically glucose or glutamine, or other sugars such as fructose, galactose, 10 mannose, and the like; (2) a nitrogen source, typically obtained via inclusion of one or more amino acids; and (3) vitamins, which are cofactors in enzyme reactions. Also essential are a wide range of inorganic salts, including Na^+ , K^+ , Ca^{2+} , Mg^{2+} , Cl^- , HPO_3^{2-} , and the 15 like, and fats and fat soluble components, including fatty acids (preferably conjugated), cholesterol, phospholipids and their precursors. Components of a standard serum-free media formulation are listed in Table 1 and include the components found in "DMEM/F- 20 12", available from media manufacturers, such as Grand Island Biological Co. (GIBCO), Grand Island, N.Y. For a review of animal cell culture media, see Mizrahi and Lazar, Cytotechnology, 1:199-214 (1988). Media considerations for insect cell cultures are disclosed 25 in Goodwin, R.H. (1990) Nature 347:209-210. An exemplary defined serum-free medium individualized for hybridoma cultivation is described by Kovar, J. (1987) Folia Biologia 33:377-384. Imagawa et al. (1989) PNAS 86:4122-4126 describe individualized media developed to 30 maximize cell growth in mouse mammary epithelial cells, and Miyazaki et al. (1991) Res. Exp. Med. 191:77-83 describe media individualized to enhance rat hepatocyte cell viability in vitro.

Table 1: Standard Media Components

	<u>Inorganic Salts:</u>
	CaCl ₂ (anhyd.)
5	CuSO ₄ ·5H ₂ O
	Fe(NO ₃) ₃ ·9H ₂ O
	FeSO ₄ ·7H ₂ O
	KCl
	MgCl ₂
10	MgSO ₄
	NaCl
	NaH ₂ PO ₄ ·H ₂ O
	Na ₂ HPO ₄
	ZnSO ₄ ·7H ₂ O
15	
	<u>Vitamins</u>
	Biotin
	D-Ca pantothenate
	Choline chloride
20	Folic acid
	i-Inositol
	Niacinamide
	Pyridoxal·HCl
	Pyridoxine·HCl
25	Riboflavin
	Thiamine·HCl
	Thymidine
	Vitamin B ₁₂
30	
	<u>Other Components:</u>
	D-Glucose
	Na hypoxanthine
	Linoleic acid
	Lipoic acid
35	Phenol red
	Putrescine·2HCl
	Sodium pyruvate
	Amino acids

- 18 -

Thus, the invention provides serum-free media with improved cell culture properties, which include standard serum-free medium components as well as an additive, formed, in one embodiment, by the acid or base hydrolysis of a phosphoglyceride ester. The serum-free medium can be purchased from a media manufacturer or created de novo in the laboratory. The proportions of the components of the serum-free medium can be adjusted and optimized for the particular cell line or protein expression system utilized. To enhance the cell cultivation properties of the medium, in addition to the additive, the serum-free medium also may be supplemented with other components, such as a fatty acids, amino acids, or phospholipid precursors.

The medium components typically can be added in any order and the final combination sterilized using a standard methodology, e.g., by filter sterilization.

The serum-free media of the invention may be utilized to improve the cell cultivation properties of a wide range of cells, both vertebrate and invertebrate, as well as the biologicals production properties of these cells. The serum-free media is anticipated to have particular utility in cell culture systems designed for vaccine or recombinant protein production. Thus, the serum-free media of the invention may be used to improve the cell culture properties in many different cell culture systems.

The invention will be understood further from the following nonlimiting examples.

- 19 -

C. ExamplesExample 1

5 In this example, the effect of an additive,
hydrolyzed phosphatidyl choline, on the cell culture
properties of a serum-free medium was examined. The
serum-free medium additive was produced by the
hydrolysis of a phosphatidyl choline composition
10 purchased from Sigma Co., St. Louis, MO. The
composition is derived from hen egg and its fatty acid
chain components are a mixture primarily of palmitic,
stearic, linoleic and oleic acids. Solutions of
hydrolyzed phosphatidyl choline were generated by both
15 an acid catalyzed hydrolysis and by base
saponification. Acid hydrolysis was implemented by
dissolving 30 mg phosphatidyl choline in 1 ml ethanol.
100 μ l of 1 N (1M) HCl then was added, and the solution
heated in a water bath at 60° to 70°C for 30 minutes,
20 to produce the additive (PC(A)). The base
saponification was implemented by adding 30 mg of
phosphatidyl choline to 1 mL ethanol and 100 μ L
1 N (1M) NaOH. The solution then was heated to 60°-
70°C in a water bath for 30 minutes, to form the
25 additive (PC(B)).

The effect of the two hydrolyzed solutions on cell
growth, cell viability and protein production was
examined as follows. CHO-1 cells, chinese hamster
30 ovary cells adapted to express the recombinant
osteogenic protein OP-1, were cultured in the presence
and absence of either PC(A) or PC(B). The osteogenic
protein OP1, is a bone morphogen capable of inducing
the cascade of events that result in true tissue
35 regeneration, including endochondral bone formation.

- 20 -

Osteogenic protein is described, for example, in U.S. Patent No. 4,968,590, and U.S. Patent No. 5,011,691. Stable OP1 transfectants were created essentially as described in international application PCT /US90/05903
5 (WO91/05802), published May 2, 1991, the disclosure of which is incorporated herein by reference. Briefly, DNA encoding OP-1 and DHFR were co-transfected into CHO cells and the cells induced to grow in the presence of methotrexate. Stable cell lines were generated for
10 selected gene-amplified clones.

In the experiment, CHO-1 cells were grown in a serum-free medium, "Medium 1", under three different conditions. In a control run, CHO-1 cells were
15 cultured in Medium 1 with a phosphatidyl choline (PC) composition (Sigma Co.) added at a concentration of 30 mg/L. In a second run, CHO-1 cells were grown in Medium 1 in the presence of 1 ml of PC(A) per liter of media. In a third run, CHO-1 cells were cultured in
20 Medium 1 in the presence 1 ml of PC(B) per L of media. "Medium 1" in all cases is a standard media composition, e.g., DMEM/F12, described in Table I and available from GIBCO, transfected for OP1 expression to which, for example, methotrexate has been added.

25

The results are presented in Figure 1 where numbers of viable cells/mL is plotted as a function of day of culture. Cell growth in the presence of Medium 1 alone is indicated by solid squares, growth in the presence
30 of Medium 1 plus PC(A) is indicated by open squares; and growth in the presence of Medium 1 plus PC(B) is indicated by solid diamonds. Viable cell density was measured with Trypan blue stain and a hemocytometer. As can be seen in the figure, the addition of
35 hydrolyzed phosphatidyl choline to Medium 1 increased

- 21 -

maximum viable cell density 20%, i.e., from a maximum of 900,000 cells/ml, using non-hydrolyzed phosphatidyl choline (PC), to a maximum density of 1,100,000 cells/ml, with medium including either PC(A) or PC(B).

5 Additionally, the slope of the lines in Figure 1 indicates that CHO-1 cell growth rate was faster in cells provided with PC(A) or PC(B) as compared with the control cells with PC.

10 Example 2

In this example, the effect of acid hydrolyzed phosphatidyl choline, PC(A) in combination with a second, different medium, "Medium 2", was evaluated.

15 Medium 2 is a variant of Medium 1, also adapted for OP-1 expression, e.g., including, for example, methotrexate, as well as the phospholipid precursor ethanolamine, and additional amino acids to double the total amino acid contribution.

20

The cell growth and OP-1 expression of CHO-1 cells in Medium 1 supplemented with PC (30 mg/L) was compared with that of CHO-1 cells in Medium 2 supplemented with PC(A) (30 mg/L) as described in Example 1. The results
25 presented in Figure 2 demonstrate that the density of viable cells is greater for CHO-1 cells grown in Medium 2 plus PC(A) (open squares) as compared with the density of cells grown in the control medium, e.g., Medium 1 plus non-hydrolyzed phosphatidyl choline (PC,
30 solid squares).

Similarly, in Figure 3, OP-1 production increased 2-3 fold in serum-free Medium 2 plus PC(A) as compared with the control Medium 1. (In the figure, Medium 1
35 plus PC is represented by a solid bar and Medium 2 plus

- 22 -

PC(A) is represented by an open bar.) In a separate control experiment, Medium 2 alone, without the addition of hydrolyzed phosphatidyl choline, was found to have only a marginally beneficial effect on the cell culture properties, as compared with Medium 1 alone. The improved cell culture properties demonstrated in Figures 2 and 3, therefore, are due primarily to the presence of the hydrolyzed phosphatidyl choline in the medium.

10

The addition of hydrolyzed phospholipid to Medium 1 or Medium 2 therefore, synergistically enhances the cell culture properties of the medium.

15

Example 3

In this example, the effect of hydrolyzed phosphatidyl choline, PC(A), on growth and expression of a second cell line, "CHO-2" cells, in serum-free Medium 2 was examined. CHO-2 cells are different, independently created chinese hamster ovary cell line adapted to express the osteogenic protein OP-1 and adapted to grow in methotrexate. Cells were grown in Medium 1 plus non-hydrolyzed phosphatidyl choline, PC, (30 mg/L) as a control, and in Medium 2 plus PC(A), 30 mg/L, prepared as for Example 2.

As illustrated in Figures 4 and 5, both viable cell growth and recombinant protein expression of CHO-2 cells were enhanced in the presence of Medium 2 plus PC(A). Figure 4 is a graph of viable cell density versus day of culture. As illustrated in Figure 4, cell growth and cell density of CHO-2 cells is 30-45% greater in Medium 2 plus PC(A) (open squares) as

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- 23 -

compared with the control medium (solid squares). As illustrated in Figure 5, OP-1 production was 2-3 times greater in Medium 2 plus PC(A) as compared with the control medium (control is represented by a solid bar, and Medium 2 plus PC(A) is represented by an open bar in the figure.) In a separate control experiment, the enhanced cell culture properties were found to be attributable primarily to the addition of hydrolyzed phosphatidyl choline to the medium, as CHO-2 cells grown in Medium 2, alone, demonstrated only marginally beneficial effects as compared with CHO-2 cells grown in Medium 1 alone.

Thus, the addition of hydrolyzed phospholipid to Medium 2 synergistically enhances the overall cell culture properties of the medium in different cell lines.

Example 4

20

The effect of hydrolyzed phosphatidyl choline PC(A) on the growth of CHO-3 cells, a third chinese hamster ovary cell line, transfected to express a synthetic DNA encoding human insulin growth factor-1, IGF-1, was examined in this example. The cDNA for human IFG-1 is described, for example, in Jansen et al., Nature, 306:609-611 (1983).

In the experiment, CHO-3 cells were grown in a serum-free medium, Medium 3 alone (no hydrolyzed phospholipid, control) or in the presence of hydrolyzed phosphatidyl choline (PC(A), 30 mg/L) and additional amino acids. "Medium 3" is a variant of Medium 1, adapted for IGF-1 expression and including linoleic acid. Cells were grown in suspension at 37 C, and counted on day 4 and day 7 as for Examples 1-3 above, e.g., with a hemocytometer.

- 24 -

The presence of hydrolyzed phosphatidyl choline in the medium was found to greatly improve the cell growth of CHO-3 cells in culture. The viable cell density in the control (Medium 3 alone) was 600,000/ml after
5 4 days and 740,000/ml after 7 days. By contrast, the viable cell density of CHO-3 cells grown in the same medium supplemented with hydrolyzed phosphatidyl choline increased to 710,000/ml after 4 days and 1,080,000/ml after 7 days, an improvement of 25%.

10

Thus, the addition of hydrolyzed phospholipid to a medium synergistically enhances the overall cell culture properties of the medium in cell lines adapted to express different biologicals, including different
15 recombinant proteins.

Other Embodiments

The invention may be embodied in other specific
20 forms without departing from the spirit or essential characteristics thereof. The present embodiments are therefore to be considered in all respects as illustrative and not restrictive, the scope of the invention being indicated by the appended claims rather
25 than by the foregoing description, and all changes which come within the meaning and range of equivalency of the claims are therefore intended to be embraced therein.

- 25 -

What is claimed is:

1. A serum-free cell culture medium having enhanced ex vivo cell cultivation properties, said medium
5 comprising an additive having the formula:



wherein:

- 15 one of R₁ and R₂ is OH;
 another of R₁ and R₂ is selected from the
 group consisting of OH and



wherein R₄ is a C₁-C₂₆ hydrocarbon; and

25

R₃ is



35

wherein one of R₅ and R₆ is O⁻, and the other is an alcohol.

2. The medium of claim 1 wherein said alcohol is part
of phosphate ester group of a cell membrane
phospholipid.

40

3. The medium of claim 1 wherein said alcohol is
selected from the group consisting of serine,
ethanolamine, choline and inositol.

- 26 -

4. The medium of claim 1 wherein said additive is a reaction product of a base saponification or acid hydrolysis of a phosphoglyceride ester.
- 5 5. The medium of claim 4 wherein said phosphoglyceride ester is a cell membrane phospholipid.
6. The medium of claim 5 wherein said phosphoglyceride ester is selected from the group consisting of a
10 phosphatidyl serine, phosphatidyl ethanolamine, phosphatidyl choline, and phosphatidyl inositol.
7. The medium of claim 1 wherein R_4 is a long chain fatty acid.
- 15 8. The medium of claim 1 wherein R_4 is a C_{10} - C_{24} hydrocarbon.
9. The medium of claim 6 wherein said hydrocarbon is a
20 fatty acid component of a cell membrane phospholipid.
10. The medium of claim 1 further comprising a phospholipid precursor.
- 25 11. The medium of claim 10 wherein said precursor comprises ethanolamine.
12. The medium of claim 1 further comprising one or
more amino acids.
- 30 13. A serum-free cell culture medium having enhanced cell culture properties, said medium comprising a hydrolysis product of a phosphoglyceride ester.

- 27 -

14. The medium of claim 13 wherein said phosphoglyceride ester is a cell membrane phospholipid.

15. The medium of claim 13 wherein said phosphoglyceride is selected from the group consisting of phosphatidyl serine, phosphatidyl choline, phosphatidyl ethanolamine, and phosphatidyl inositol.

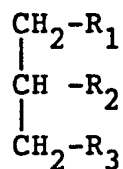
16. The medium of claim 13 wherein said hydrolysis product is a lysophosphoglyceride ester.

17. A method of improving the cell cultivation or biologicals production properties of a serum-free cell culture medium comprising the step of:

15

adding to a serum-free cell culture medium an additive having the formula:

20



25

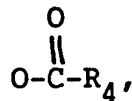
wherein:

one of R_1 or R_2 is OH;

30

another of R_1 or R_2 is selected from the group consisting of OH and

35

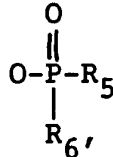


- 28 -

wherein R_4 is a C_1-C_{26} hydrocarbon; and
 wherein

R_3 is

5



10

wherein one of R_5 and R_6 is O^- , and the other
 is an alcohol.

15 18. The method of claim 17 wherein said alcohol
 comprises part of a phosphate ester group of a cell
 membrane phospholipid.

20 19. The method of claim 17 wherein the alcohol is
 selected from the group consisting of serine,
 ethanolamine, choline and inositol.

20. The method of claim 17 wherein R_4 is a $C_{10}-C_{24}$
 hydrocarbon.

25

21. The method of claim 17 wherein R_4 is a fatty acid
 component of a cell membrane phospholipid.

22. The method of claim 17 wherein the medium further
 30 comprises a phospholipid precursor.

23. The method of claim 22 wherein said precursor
 comprises ethanolamine.

35 24. The method of claim 17 wherein the medium further
 comprises one or more amino acids.

- 29 -

25. The method of claim 17 for increasing the growth rate of cultured cells exposed to said medium.

26. The method of claim 17 for enhancing the viability
5 of cultured cells exposed to said medium.

27. The method of claim 17 capable of enhancing the recombinant protein production of cultured cells exposed to said medium.

10

28. The method of claim 17, further comprising, prior to adding said additive to said medium, the step of:

15 hydrolyzing a phosphoglyceride ester selected from the group consisting of a phosphatidyl serine, phosphatidyl ethanolamine, phosphatidyl choline, and phosphatidyl inositol, thereby to produce the additive.

20 29. The method of claim 28 wherein said hydrolyzing step comprises an acid hydrolysis.

25 30. The method of claim 29 wherein said acid hydrolysis comprises exposing said phosphoglyceride ester to a solution having pH in the range of about 0.2-4.

31. The method of claim 30 wherein said solution has a pH in the range of 3-5.

30 32. The method of claim 29 wherein the acid solution comprises a concentration of about 0.05 M to 10 M of an acid selected from the group consisting of hydrochloric acid, sulfuric acid, phosphoric acid and nitric acid.

- 30 -

33. The method of claim 28 wherein said hydrolyzing step comprises a base saponification.
34. The method of claim 33 wherein said base
5 saponification comprises exposing said phosphoglyceride ester to a solution having a pH in the range of about 8-16.
35. The method of claim 34 wherein said solution has a
10 pH in the range of 8-14.
36. The method of claim 33 wherein the basic solution comprises a concentration of 0.05 M to 10.0 M of a base selected from the group consisting of sodium hydroxide,
15 potassium hydroxide and ammonium hydroxide.
37. The method of claim 29 or 34 wherein the solution comprises ethanol.
- 20 38. The method of claim 28 wherein said phosphoglyceride ester is heated to a temperature greater than 40°C.
39. The method of claim 38 wherein said
25 phosphoglyceride ester is heated to a temperature in the the range of 45°C-90°C.
40. The method of claim 39 wherein said phosphoglyceride ester is heated to a temperature in
30 the range of about 50°C-65°C.
41. The method of claim 38 wherein said phosphoglyceride ester is heated for at least 15 minutes.

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- 31 -

42. A method of improving the cell culture properties of a serum-free cell culture medium comprising the steps of:

- 5 (i) hydrolyzing a phosphoglyceride ester thereby to form a solution comprising a hydrolyzed phosphoglyceride ester ; and
 (ii) adding the solution, obtained in step (i), to the serum-free medium.

10

43. The method of claim 42 wherein the phosphoglyceride ester hydrolyzed in step (i) is selected from the group consisting of a phosphatidyl serine, phosphatidyl ethanolamine, phosphatidyl choline, and phosphatidyl
15 inositol.

44. The method of claim 42 wherein the hydrolysis conducted in step (i) is acid catalyzed.

20 45. The method of claim 42 wherein step (i) comprises:

 heating the phosphoglyceride ester to a temperature in the range of about 45-90°C for at least 20 minutes in an solution having a pH in the range of 0.2-4.

25

46. The method of claim 42 wherein the hydrolysis conducted in step (i) is base catalyzed.

30 47. The method of claim 45 wherein step (i) comprises:

 heating the phosphoglyceride ester to a temperature in the range of about 45-90°C for at least 20 minutes in a solution having a pH in the range of 8-14.

- 32 -

48. A method of enhancing the cell cultivation or recombinant biologicals production properties of a cell cultured ex vivo, the method comprising the step of cultivating said cell in the serum-free medium of claim
5 1 or 12.

49. The method of claim 48 for enhancing the growth rate of said cell.

10 50. The method of claim 48 for enhancing the viability of said cell.

51. The method of claim 48 for enhancing the production of a recombinant protein in said cell.

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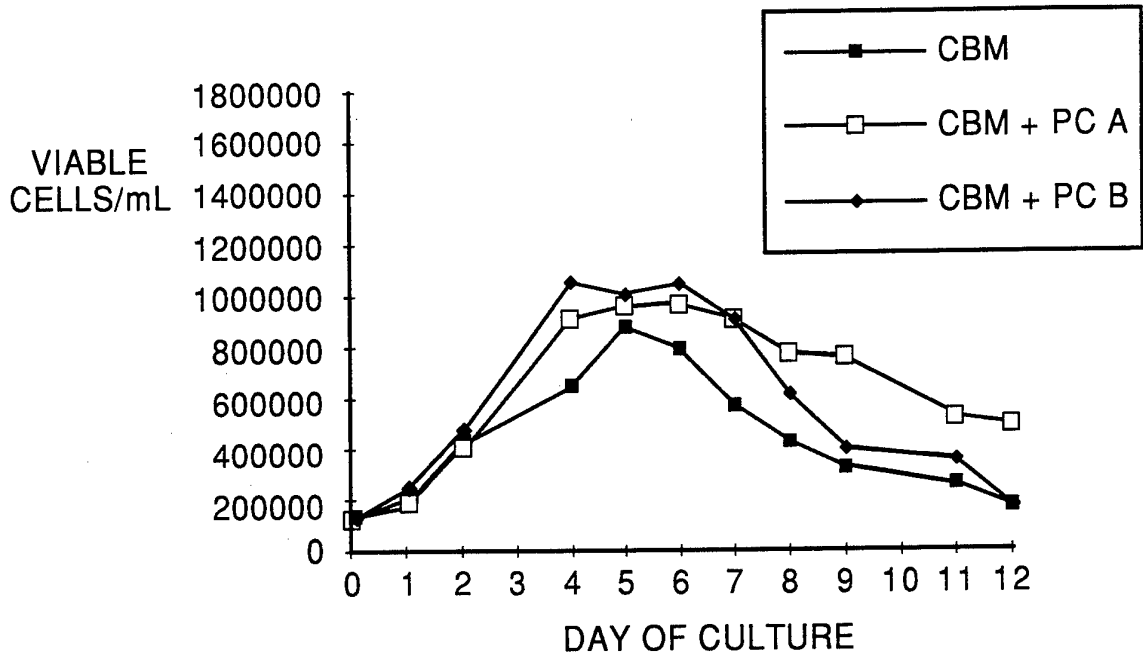


FIG. 1

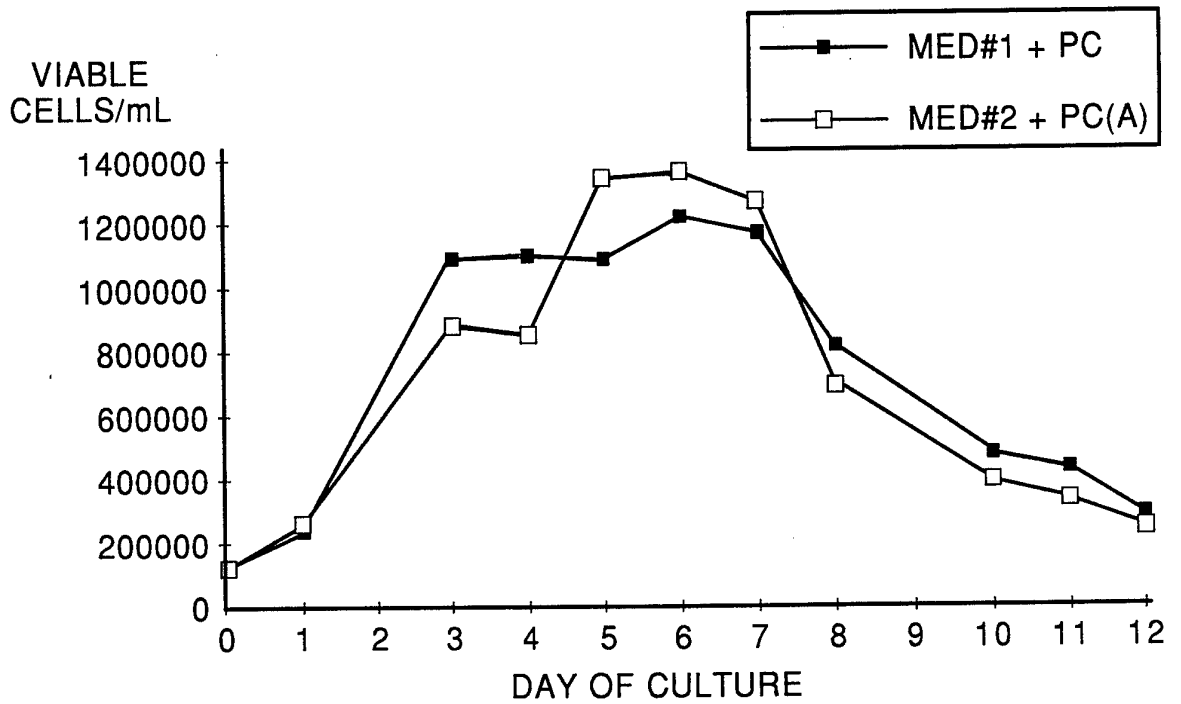


FIG. 2

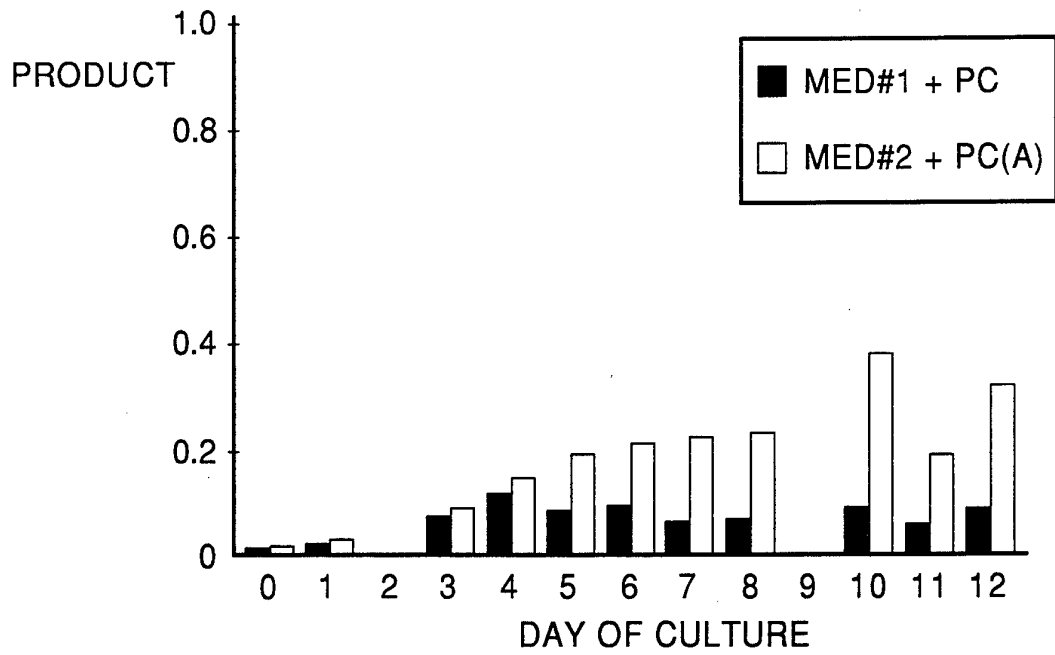


FIG. 3

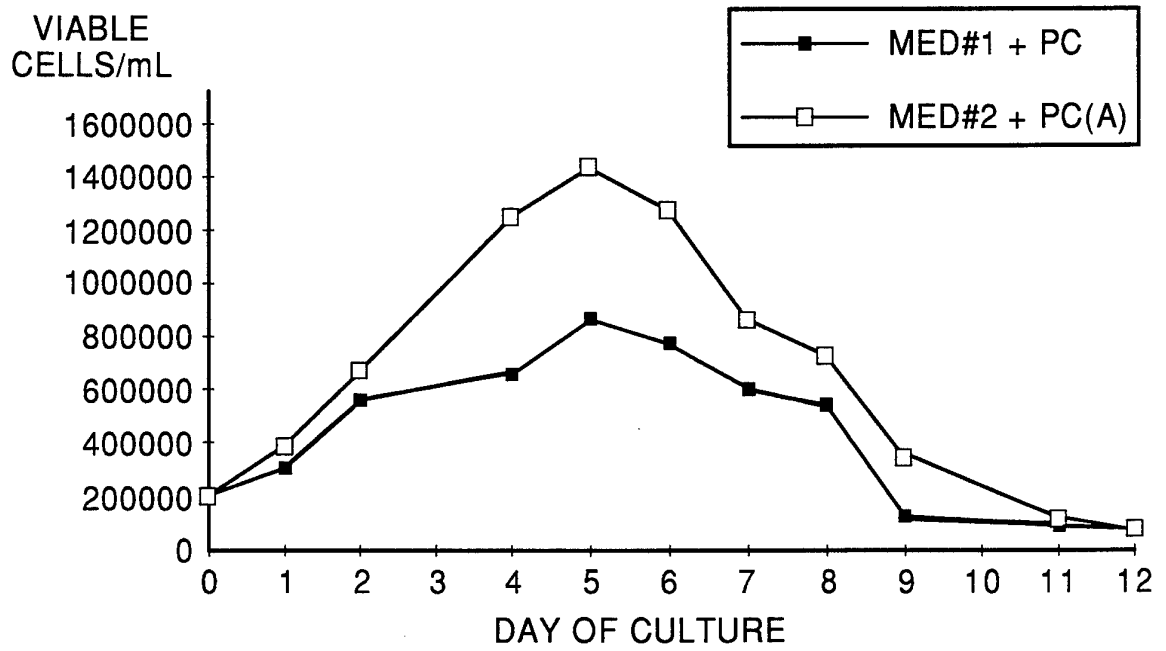


FIG. 4

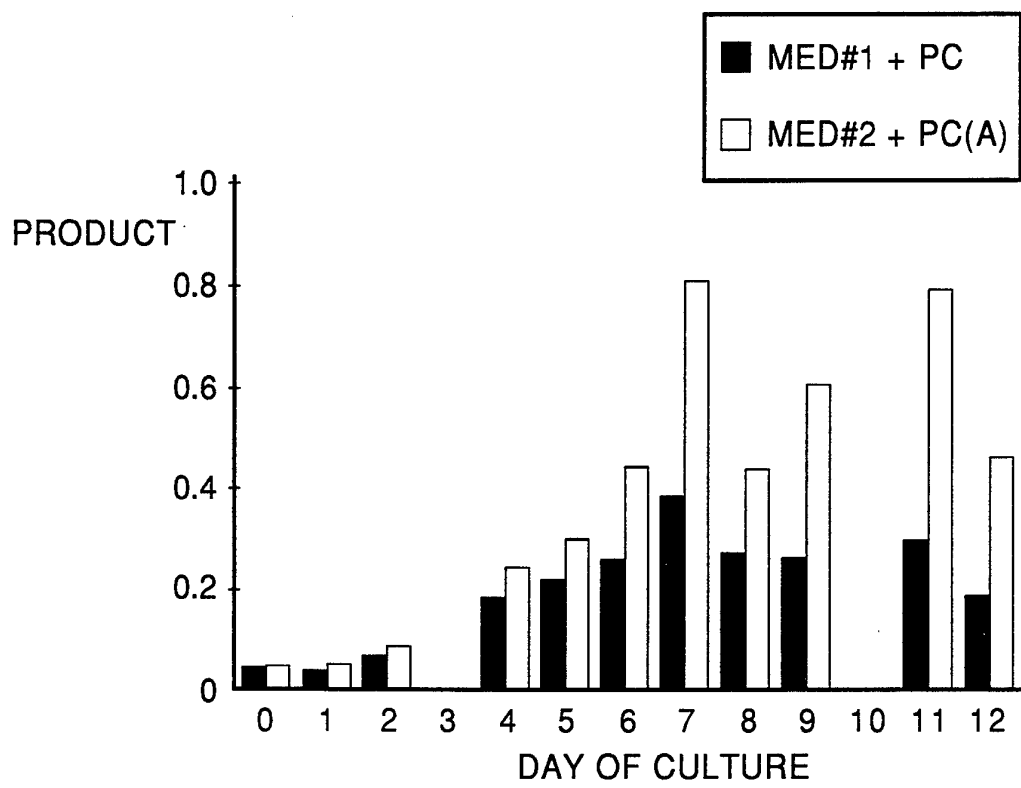


FIG. 5

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 94/10736

<p>A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N5/00</p> <p>According to International Patent Classification (IPC) or to both national classification and IPC</p>											
<p>B. FIELDS SEARCHED</p> <p>Minimum documentation searched (classification system followed by classification symbols) IPC 6 C12N</p> <p>Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched</p> <p>Electronic data base consulted during the international search (name of data base and, where practical, search terms used)</p>											
<p>C. DOCUMENTS CONSIDERED TO BE RELEVANT</p> <table border="1"> <thead> <tr> <th>Category *</th> <th>Citation of document, with indication, where appropriate, of the relevant passages</th> <th>Relevant to claim No.</th> </tr> </thead> <tbody> <tr> <td>X</td> <td>WO,A,90 07007 (THE UNITED STATES OF AMERICA) 28 June 1990</td> <td>1-9, 12-21, 24-27, 48-51</td> </tr> <tr> <td>Y</td> <td>see page 4, line 4 - line 10; tables 1,3 see page 7, line 17 - page 8, line 13; example 2 --- -/--</td> <td>10,11, 22,23, 28-47</td> </tr> </tbody> </table>			Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.	X	WO,A,90 07007 (THE UNITED STATES OF AMERICA) 28 June 1990	1-9, 12-21, 24-27, 48-51	Y	see page 4, line 4 - line 10; tables 1,3 see page 7, line 17 - page 8, line 13; example 2 --- -/--	10,11, 22,23, 28-47
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.									
X	WO,A,90 07007 (THE UNITED STATES OF AMERICA) 28 June 1990	1-9, 12-21, 24-27, 48-51									
Y	see page 4, line 4 - line 10; tables 1,3 see page 7, line 17 - page 8, line 13; example 2 --- -/--	10,11, 22,23, 28-47									
<p><input checked="" type="checkbox"/> Further documents are listed in the continuation of box C. <input checked="" type="checkbox"/> Patent family members are listed in annex.</p>											
<p>* Special categories of cited documents :</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p>											
<p>Date of the actual completion of the international search</p> <p>3 February 1995</p>		<p>Date of mailing of the international search report</p> <p>15 -02- 1995</p>									
<p>Name and mailing address of the ISA</p> <p>European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+ 31-70) 340-2040, Tx. 31 651 epo nl, Fax (+ 31-70) 340-3016</p>		<p>Authorized officer</p> <p>Ryckebosch, A</p>									

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 94/10736

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>CHEMICAL ABSTRACTS, vol. 115, no. 15, 14 October 1991, Columbus, Ohio, US; abstract no. 154361y, M. MIYAZAKI ET AL. 'EXTENDING EFFECTS OF PHOSPHOLIPIDS, CHOLESTEROL, AND ETHANOLAMINES ON SURVIVAL OF ADULT RAT HEPATOCYTES IN SERUM-FREE PRIMARY CULTURE.' page 472 ; cited in the application see abstract & RES. EXP. MED., vol.191, no.2 pages 77 - 83</p>	10,11, 22,23
Y	<p>--- CHEMICAL ABSTRACTS, vol. 106, no. 17, 27 April 1987, Columbus, Ohio, US; abstract no. 134714b, J.G. ALVAREZ ET AL. 'ALKALINE HYDROLYSIS OF PHOSPHOGLYCERIDES ON THIN LAYER PLATES IN SITU.' page 349 ; see abstract & J. LIQ. CHROMATOGR., vol.9, no.16, 27 April 1987 pages 3495 - 3504</p>	28-47
Y	<p>--- CHEMICAL ABSTRACTS, vol. 115, no. 21, 25 November 1991, Columbus, Ohio, US; abstract no. 230554f, page 799 ; see abstract & JP,A,3 123 493 (NIPPON OIL AND FATS CO., LTD.) 27 May 1991</p>	28-47
Y	<p>--- CHEMICAL ABSTRACTS, vol. 109, no. 25, 19 December 1988, Columbus, Ohio, US; abstract no. 226289g, page 436 ; see abstract & JP,A,6 333 387 (Q.P. CORP.) 13 February 1988</p> <p>--- -/--</p>	28-47

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 94/10736

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	<p>BIOLOGICAL ABSTRACTS, vol. 83, no. 8 1987, Philadelphia, PA, US; abstract no. 78066, L. FACCI ET AL. 'DISSOCIATION OF THE STELLATE MORPHOLOGY FROM INTRACELLULAR CYCLIC AMP LEVELS IN CULTURED RAT BRAIN ASTROGLIAL CELLS: EFFECTS OF GANGLIOSIDE Gm1 AND LYSOPHOSPHATIDYLSERINE.' page AB-763 ; see abstract & J. NEUROCHEM., vol.48, no.2, 1987 pages 566 - 573</p> <p style="text-align: center;">---</p>	1-9
X	<p>BIOLOGICAL ABSTRACTS, vol. 96, no. 12 December 1993, Philadelphia, PA, US; abstract no. 136808, S. HOMMA ET AL. 'VITAMIN D-BINDING PROTEIN (GROUP-SPECIFIC COMPONENT) IS THE ROLE SERUM PROTEIN REQUIRED FOR MACROPHAGE ACTIVATION AFTER TREATMENT OF PERITONEAL CELLS WITH LYSOPHOSPHATIDYLCHOLINE.' page AB-574 ; see abstract & IMMUNOL. CELL BIOL., vol.71, no.4 pages 249 - 257</p> <p style="text-align: center;">-----</p>	1-9

INTERNATIONAL SEARCH REPORT

Information on patent family members

International application No.
PCT/US 94/10736

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO-A-9007007	28-06-90	AU-B- 621972	26-03-92
		AU-A- 4815990	10-07-90
		CA-A- 2005494	14-06-90
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		US-A- 5342777	30-08-94

JP-A-3123493	27-05-91	NONE	

JP-A-6333387	02-12-94	NONE	
