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(54) Titre : PROCEDE DE PRODUCTION DE L-LYSINE
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(57) Abrégé/Abstract:
The L-lysine-producing ability and the L-lysine-producing speed are improved in a coryneform bacterium harboring an aspartokinase in which feedback inhibition by L-lysine and L-threonine is substantially desensitized, by successively enhancing DNA coding for a dihydrodipicolinate reductase, DNA coding for a dihydrodipicolinate synthase, DNA coding for a diaminopimelate decarboxylase, and DNA coding for a diaminopimelate dehydrogenase.

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Abstract

The L-lysine-producing ability and the L-lysine-producing speed are improved in a coryneform bacterium harboring an aspartokinase in which feedback inhibition by L-lysine and L-threonine is substantially desensitized, by successively enhancing DNA coding for a dihydrodipicolinate reductase, DNA coding for a dihydrodipicolinate synthase, DNA coding for a diaminopimelate decarboxylase, and DNA coding for a diaminopimelate dehydrogenase.

METHOD OF PRODUCING L-LYSINE

Technical Field

The present invention relates to a method for producing L-lysine by cultivating a microorganism obtained by modifying a coryneform bacterium used for fermentative production of amino acid or the like by means of a technique based on genetic engineering.

Background Art

L-Lysine, which is used as a fodder additive, is usually produced by a fermentative method by using an L-lysine-producing mutant strain belonging to the coryneform bacteria. Various L-lysine-producing bacteria known at present are those created by artificial mutation starting from wild type strains belonging to the coryneform bacteria.

As for the coryneform bacteria, there are disclosed a vector plasmid which is autonomously replicable in bacterial cells and has a drug resistance marker gene (see United States Patent No. 4,514,502), and a method for introducing a gene into bacterial cells (for example, Japanese Patent Laid-open No. 2-207791). There is also disclosed a possibility for breeding an L-threonine- or L-isoleucine-producing bacterium by using

the techniques as described above (see United States Patent Nos. 4,452,890 and 4,442,208). As for breeding of an L-lysine-producing bacterium, a technique is known, in which a gene participating in L-lysine biosynthesis is incorporated into a vector plasmid to amplify the gene in bacterial cells (for example, Japanese Patent Laid-open No. 56-160997).

Known genes for L-lysine biosynthesis include, for example, a dihydrodipicolinate reductase gene (Japanese Patent Laid-open No. 7-75578) and a diaminopimelate dehydrogenase gene (Ishino, S. et al., Nucleic Acids Res., 15, 3917 (1987)) in which a gene participating in L-lysine biosynthesis is cloned, as well as a phosphoenolpyruvate carboxylase gene (Japanese Patent Laid-open No. 60-87788), a dihydrodipicolinate synthase gene (Japanese Patent Publication No. 6-55149), and a diaminopimelate decarboxylase gene (Japanese Patent Laid-open No. 60-62994) in which amplification of a gene affects L-lysine productivity.

As for enzymes participating in L-lysine biosynthesis, a case is known for an enzyme which undergoes feedback inhibition when used as a wild type. In this case, L-lysine productivity is improved by introducing an enzyme gene having such mutation that the feedback inhibition is desensitized. Those known as such a gene specifically include, for example, an aspartokinase gene (International Publication Pamphlet

of WO 94/25605).

As described above, certain successful results have been obtained by means of amplification of genes for the L-lysine biosynthesis system, or introduction of mutant genes. For example, a coryneform bacterium, which harbors a mutant aspartokinase gene with desensitized concerted inhibition by lysine and threonine, produces a considerable amount of L-lysine (about 25 g/L). However, this bacterium suffers decrease in growth speed as compared with a bacterium harboring no mutant aspartokinase gene. It is also reported that L-lysine productivity is improved by further introducing a dihydrodipicolinate synthase gene in addition to a mutant aspartokinase gene (Applied and Environmental Microbiology, 57(6), 1746-1752 (1991)). However, such a bacterium suffers further decrease in growth speed.

As for the dihydrodipicolinate reductase gene, it has been demonstrated that the activity of dihydrodipicolinate reductase is increased in a coryneform bacterium into which the gene has been introduced, however, no report is included for the influence on L-lysine productivity (Japanese Patent Laid-open No. 7-75578).

In the present circumstances, no case is known for the coryneform bacteria, in which anyone has succeeded in remarkable improvement in L-lysine yield without restraining growth by combining a plurality of genes for

L-lysine biosynthesis. No case has been reported in which growth is intended to be improved by enhancing a gene for L-lysine biosynthesis as well.

Disclosure of the Invention

An object of the present invention is to improve the L-lysine-producing ability and the growth speed of a coryneform bacterium by using genetic materials of DNA sequences each coding for aspartokinase (hereinafter referred to as "AK", provided that a gene coding for an AK protein is hereinafter referred to as "lysC", if necessary), dihydrodipicolinate reductase (hereinafter referred to as "DDPR", provided that a gene coding for a DDPR protein is hereinafter referred to as "dapB", if necessary), dihydrodipicolinate synthase (hereinafter abbreviate as "DDPS", provided that a gene coding for a DDPS protein is hereinafter referred to as "dapA", if necessary), diaminopimelate decarboxylase (hereinafter referred to as "DDC", provided that a gene coding for a DDC protein is hereinafter referred to as "lysA", if necessary), and diaminopimelate dehydrogenase (hereinafter referred to as "DDH", provided that a gene coding for a DDH protein is hereinafter referred to as "ddh", if necessary) which are important enzymes for L-lysine biosynthesis in cells of coryneform bacteria.

When an objective substance is produced

fermentatively by using a microorganism, the production speed, as well as the yield of the objective substance relative to an introduced material, is an extremely important factor. An objective substance may be produced remarkably inexpensively by increasing the production speed per a unit of fermentation equipment. Accordingly, it is industrially extremely important that the fermentative yield and the production speed are compatible with each other. The present invention proposes a solution for the problem as described above in order to fermentatively produce L-lysine by using a coryneform bacterium.

The principle of the present invention is based on the fact that the growth of a coryneform bacterium can be improved, and the L-lysine-producing speed thereof can be improved by making enhancement while combining dapB with mutant lysC (hereinafter simply referred to as "mutant lysC", if necessary) coding for mutant AK (hereinafter simply referred to as "mutant type AK", if necessary) in which concerted inhibition by lysine and threonine is desensitized, as compared with a case in which lysC is enhanced singly, and that the L-lysine-producing speed can be further improved in a stepwise manner by successively enhancing dapA, lysA, and ddh.

The present invention provides a method for producing L-lysine comprising the steps of cultivating a coryneform bacterium *Corynebacterium glutamicum* in an appropriate medium, to produce and accumulate L-lysine in a culture of the bacterium, and collecting L-lysine from the culture, wherein the coryneform bacterium harbors an aspartokinase in which feedback inhibition by L-lysine and L-threonine is desensitized as compared with a wild

type aspartokinase, and wherein the intracellular activities of dihydrodipicolinate reductase and dihydrodipicolinate synthase are upregulated by increasing the copy number of, and/or using a strong promoter for a DNA sequence from *Corynebacterium glutamicum* coding for a dihydrodipicolinate reductase and a DNA sequence from *Corynebacterium glutamicum* coding for a dihydrodipicolinate synthase, and wherein the intracellular activity of diaminopimelate decarboxylase is further upregulated by increasing the copy number of, and/or using a strong promoter for a DNA sequence from *Corynebacterium glutamicum* coding for a diaminopimelate decarboxylase.

The aspartokinase is a mutant aspartokinase in which a 279th alanine residue as counted from its N-terminal is changed into an amino acid residue other than alanine and other than acidic amino acid in its α-subunit as shown in SEQ ID NO: 5 in the Sequence Listing, and a 30th alanine residue is changed into an amino acid residue other than alanine and other than acidic amino acid in its β-subunit as shown in SEQ ID NO: 7 in the Sequence Listing.

In another aspect, the present invention provides a coryneform bacterium harbouring an aspartokinase in which feedback inhibition by L-lysine and L-threonine is substantially desensitized, and comprising enhanced DNA coding for a dihydrodipicolinate reductase. The present invention provides a coryneform bacterium further comprising enhanced DNA coding for a dihydrodipicolinate synthase in the aforementioned coryneform bacterium. The present invention provides a coryneform bacterium further comprising enhanced DNA coding for a diaminopimelate decarboxylase in the aforementioned coryneform bacterium, in addition to the three DNA's described above. The present invention provides a

coryneform bacterium further comprising enhanced DNA coding for a diaminopimelate dehydrogenase in the aforementioned coryneform bacterium, in addition to the four DNA's described above.

In still another aspect, the present invention provides a method for producing L-lysine comprising the steps of cultivating any one of the coryneform bacteria described above in an appropriate medium, producing and accumulating L-lysine in a culture of the bacterium, and collecting L-lysine from the culture.

The coryneform bacteria referred to in the present invention are a group of microorganisms as defined in Bergey's Manual of Determinative Bacteriology, 8th ed., p. 599 (1974), which are aerobic Gram-positive rods having no acid resistance and no spore-forming ability. The coryneform bacteria include bacteria belonging to the genus Corynebacterium, bacteria belonging to the genus Brevibacterium having been hitherto classified into the genus Brevibacterium but united as bacteria belonging to the genus Corynebacterium at present, and bacteria belonging to the genus Brevibacterium closely relative to bacteria belonging to the genus Corynebacterium.

The present invention will be explained in detail below.

<1> Preparation of genes for L-lysine biosynthesis used for the present invention

The genes for L-lysine biosynthesis used in the present invention are obtained respectively by preparing chromosomal DNA from a bacterium as a DNA donor, constructing a chromosomal DNA library by using a plasmid vector or the like, selecting a strain harboring a desired gene, and recovering, from the selected strain, recombinant DNA into which the gene has been inserted. The DNA donor for the gene for L-lysine biosynthesis used in the present invention is not specifically limited provided that the desired gene for L-lysine biosynthesis expresses an enzyme protein which functions in cells of coryneform bacteria. However, the DNA donor is preferably a coryneform bacterium.

All of the genes of lysC, dapA, and dapB originating from coryneform bacteria have known sequences. Accordingly, they can be obtained by performing amplification in accordance with the polymerase chain reaction method (PCR; see White, T. J. et al., Trends Genet., 5, 185 (1989)).

Each of the genes for L-lysine biosynthesis used in the present invention is obtainable in accordance with certain methods as exemplified below.

(1) Preparation of mutant lysC

A DNA fragment containing mutant lysC can be

prepared from a mutant strain in which synergistic feedback inhibition on the AK activity by L-lysine and L-threonine is substantially desensitized (International Publication Pamphlet of WO 94/25605). Such a mutant strain can be obtained, for example, from a group of cells originating from a wild type strain of a coryneform bacterium subjected to a mutation treatment by applying an ordinary mutation treatment such as ultraviolet irradiation and treatment with a mutating agent such as N-methyl-N'-nitro-N-nitrosoguanidine. The AK activity can be measured by using a method described by Miyajima, R. et al. in The Journal of Biochemistry (1968), 63(2), 139-148. The most preferred as such a mutant strain is represented by an L-lysine-producing bacterium AJ3445 (FERM P-1944) derived by a mutation treatment from a wild type strain of Brevibacterium lactofermentum ATCC 13869 (having its changed present name of Corynebacterium glutamicum).

Alternatively, mutant lysC is also obtainable by an in vitro mutation treatment of plasmid DNA containing wild type lysC. In another aspect, information is specifically known on mutation to desensitize synergistic feedback inhibition on AK by L-lysine and L-threonine (International Publication Pamphlet of WO 94/25605). Accordingly, mutant lysC can be also prepared from wild type lysC on the basis of the information in accordance with, for example, the site-

directed mutagenesis method.

A fragment comprising lysC can be isolated from a coryneform bacterium by preparing chromosomal DNA in accordance with, for example, a method of Saito and Miura (H. Saito and K. Miura, Biochem. Biophys. Acta, 72, 619 (1963)), and amplifying lysC in accordance with the polymerase chain reaction method (PCR; see White, T. J. et al., Trends Genet., 5, 185 (1989)).

DNA primers are exemplified by single strand DNA's of 23-mer and 21-mer having nucleotide sequences shown in SEQ ID NOs: 1 and 2 in Sequence Listing in order to amplify, for example, a region of about 1,643 bp coding for lysC based on a sequence known for Corynebacterium glutamicum (see Molecular Microbiology (1991), 5(5), 1197-1204; Mol. Gen. Genet. (1990), 224, 317-324). DNA can be synthesized in accordance with an ordinary method by using DNA synthesizer model 380B produced by Applied Biosystems and using the phosphoamidite method (see Tetrahedron Letters (1981), 22, 1859). PCR can be performed by using DNA Thermal Cycler Model PJ2000 produced by Takara Shuzo, and using Taq DNA polymerase in accordance with a method designated by the supplier.

It is preferred that lysC amplified by PCR is ligated with vector DNA autonomously replicable in cells of E. coli and/or coryneform bacteria to prepare recombinant DNA, and the recombinant DNA is introduced into cells of E. coli beforehand. Such provision makes

following operations easy. The vector autonomously replicable in cells of E. coli is preferably a plasmid vector which is preferably autonomously replicable in cells of a host, including, for example, pUC19, pUC18, pBR322, pHSG299, pHSG399, pHSG398, and RSF1010.

When a DNA fragment having an ability to allow a plasmid to be autonomously replicable in coryneform bacteria is inserted into these vectors, they can be used as a so-called shuttle vector autonomously replicable in both E. coli and coryneform bacteria.

Such a shuttle vector includes the followings. Microorganisms harboring each of vectors and deposition numbers in international deposition facilities are shown in parentheses.

pHC4: Escherichia coli AJ12617 (FERM BP-3532)

pAJ655: Escherichia coli AJ11882 (FERM BP-136)

Corynebacterium glutamicum SR8201 (ATCC 39135)

pAJ1844: Escherichia coli AJ11883 (FERM BP-137)

Corynebacterium glutamicum SR8202 (ATCC 39136)

pAJ611: Escherichia coli AJ11884 (FERM BP-138)

pAJ3148: Corynebacterium glutamicum SR8203 (ATCC 39137)

pAJ440: Bacillus subtilis AJ11901 (FERM BP-140)

These vectors are obtainable from the deposited microorganisms as follows. Cells collected at a logarithmic growth phase were lysed by using lysozyme and SDS, followed by separation from a lysate by centrifugation at 30,000 x g to obtain a supernatant to

which polyethylene glycol is added, followed by fractionation and purification by means of cesium chloride-ethidium bromide equilibrium density gradient centrifugation.

E. coli can be transformed by introducing a plasmid in accordance with, for example, a method of D. M. Morrison (Methods in Enzymology, 68, 326 (1979)) or a method in which recipient cells are treated with calcium chloride to increase permeability for DNA (Mandel, M. and Higa, A., J. Mol. Biol., 53, 159 (1970)).

Wild type lysC is obtained when lysC is isolated from an AK wild type strain, while mutant lysC is obtained when lysC is isolated from an AK mutant strain in accordance with the method as described above.

An example of a nucleotide sequence of a DNA fragment containing wild type lysC is shown in SEQ ID NO: 3 in Sequence Listing. An amino acid sequence of α-subunit of a wild type AK protein is deduced from the nucleotide sequence, which is shown in SEQ ID NO: 4 in Sequence Listing together with the DNA sequence. Only the amino acid sequence is shown in SEQ ID NO: 5. An amino acid sequence of β-subunit of the wild type AK protein is deduced from the nucleotide sequence of DNA, which is shown in SEQ ID NO: 6 in Sequence Listing together with the DNA. Only the amino acid sequence is shown in SEQ ID NO: 7. In each of the subunits, GTG is used as an initiation codon, and a corresponding amino

acid is represented by methionine. However, this representation refers to methionine, valine, or formylmethionine.

The mutant lysC used in the present invention is not specifically limited provided that it codes for AK in which synergistic feedback inhibition by L-lysine and L-threonine is desensitized. However, the mutant lysC is exemplified by one including mutation in which a 279th alanine residue as counted from the N-terminal is changed into an amino acid residue other than alanine and other than acidic amino acid in the α-subunit, and a 30th alanine residue is changed into an amino acid residue other than alanine and other than acidic amino acid in the β-subunit in the amino acid sequence of the wild type AK. The amino acid sequence of the wild type AK specifically includes the amino acid sequence shown in SEQ ID NO: 5 in Sequence Listing as the α-subunit, and the amino acid sequence shown in SEQ ID NO: 7 in Sequence Listing as the β-subunit.

Those preferred as the amino acid residue other than alanine and other than acidic amino acid include threonine, arginine, cyteine, phenylanaline, proline, serine, tyrosine, and valine residues.

The codon corresponding to an amino acid residue to be substituted is not specifically limited for its type provided that it codes for the amino acid residue. It is assumed that the amino acid sequence of possessed

wild type AK may slightly differ depending on the difference in bacterial species and bacterial strains. AK's, which have mutation based on, for example, substitution, deletion, or insertion of one or more amino acid residues at one or more positions irrelevant to the enzyme activity as described above, can be also used for the present invention. Other AK's, which have mutation based on, for example, substitution, deletion, or insertion of other one or more amino acid residues, can be also used provided that no influence is substantially exerted on the AK activity, and on the desensitization of synergistic feedback inhibition by L-lysine and L-threonine.

An AJ12691 strain obtained by introducing a mutant lysC plasmid p399AK9B into an AJ12036 strain (FERM BP-734)as a wild type strain of Brevibacterium lactofermentum has been deposited on April 10, 1992 under a deposition number of FERM P-12918 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan), transferred to international deposition based on the Budapest Treaty on February 10, 1995, and deposited under a deposition number of FERM BP-4999.

(2) Preparation of dapB

A DNA fragment containing dapB can be prepared from chromosome of a coryneform bacterium by means of PCR. The DNA donor is not specifically limited, however, it is exemplified by Brevibacterium lactofermentum ATCC 13869 strain.

A DNA sequence coding for DDPR is known for Brevibacterium lactofermentum (Journal of Bacteriology, 175(9), 2743-2749 (1993)), on the basis of which DNA primers for PCR can be prepared. Such DNA primers are specifically exemplified by DNA's of 23-mers respectively having nucleotide sequences depicted in SEQ ID NOs: 8 and 9 in Sequence Listing. Synthesis of DNA, PCR, and preparation of a plasmid containing obtained dapB can be performed in the same manner as those for lysC described above.

A nucleotide sequence of a DNA fragment containing dapB and an amino acid sequence deduced from the nucleotide sequence are illustrated in SEQ ID NO: 10. Only the amino acid sequence is shown in SEQ ID NO: 11. In addition to DNA fragments coding for this amino acid sequence, the present invention can equivalently use DNA fragments coding for amino acid sequences substantially the same as the amino acid sequence shown in SEQ ID NO: 11, namely amino acid sequences having mutation based on, for example, substitution, deletion, or insertion of one or more amino acids provided that there is no

substantial influence on the DDPR activity.

A transformant strain AJ13107 obtained by introducing a plasmid pCRDAPB containing dapB obtained in Example described later on into E. coli JM109 strain has been internationally deposited since May 26, 1995 under a deposition number of FERM BP-5114 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) based on the Budapest Treaty.

(3) Preparation of dapA

A DNA fragment containing dapA can be prepared from chromosome of a coryneform bacterium by means of PCR. The DNA donor is not specifically limited, however, it is exemplified by Brevibacterium lactofermentum ATCC 13869 strain.

A DNA sequence coding for DDPS is known for Corynebacterium glutamicum (see Nucleic Acids Research, 18(21), 6421 (1990); EMBL accession No. X53993), on the basis of which DNA primers for PCR can be prepared. Such DNA primers are specifically exemplified by DNA's of 23-mers respectively having nucleotide sequences depicted in SEQ ID NOs: 12 and 13 in Sequence Listing. Synthesis of DNA, PCR, and preparation of a plasmid containing obtained dapA can be performed in the same

manner as those for lysC described above.

A nucleotide sequence of a DNA fragment containing dapA and an amino acid sequence deduced from the nucleotide sequence are exemplified in SEQ ID NO: 14. Only the amino acid sequence is shown in SEQ ID NO: 15. In addition to DNA fragments coding for this amino acid sequence, the present invention can equivalently use DNA fragments coding for amino acid sequences substantially the same as the amino acid sequence shown in SEQ ID NO: 15, namely amino acid sequences having mutation based on, for example, substitution, deletion, or insertion of one or more amino acids provided that there is no substantial influence on the DDPS activity.

A transformant strain AJ13106 obtained by introducing a plasmid pCRDAPA containing dapA obtained in Example described later on into E. coli JM109 strain has been internationally deposited since May 26, 1995 under a deposition number of FERM BP-5113 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) based on the Budapest Treaty.

(4) Preparation of lysA

A DNA fragment containing lysA can be prepared from chromosome of a coryneform bacterium by means of PCR.

The DNA donor is not specifically limited, however, it is exemplified by Brevibacterium lactofermentum ATCC 13869 strain.

In the coryneform bacteria, lysA forms an operon together with argS (arginyl-tRNA synthase gene), and lysA exists downstream from argS. Expression of lysA is regulated by a promoter existing upstream from argS (see Journal of Bacteriology, Nov., 7356-7362 (1993)). DNA sequences of these genes are known for Corynebacterium glutamicum (see Molecular Microbiology, 4(11), 1819-1830 (1990); Molecular and General Genetics, 212, 112-119 (1988)), on the basis of which DNA primers for PCR can be prepared. Such DNA primers are specifically exemplified by DNA's of 23-mers respectively having nucleotide sequences shown in SEQ ID NO: 16 in Sequence Listing (corresponding to nucleotide numbers 11 to 33 in a nucleotide sequence described in Molecular Microbiology, 4(11), 1819-1830 (1990)) and SEQ ID NO: 17 (corresponding to nucleotide numbers 1370 to 1392 in a nucleotide sequence described in Molecular and General Genetics, 212, 112-119 (1988)). Synthesis of DNA, PCR, and preparation of a plasmid containing obtained lysA can be performed in the same manner as those for lysC described above.

In Example described later on, a DNA fragment containing a promoter, argS, and lysA was used in order to enhance lysA. However, argS is not essential for the

present invention. It is allowable to use a DNA fragment in which lysA is ligated just downstream from a promoter.

A nucleotide sequence of a DNA fragment containing argS and lysA, and an amino acid sequence deduced to be encoded by the nucleotide sequence are exemplified in SEQ ID NO: 18. An example of an amino acid sequence encoded by argS is shown in SEQ ID NO: 19, and an example of an amino acid sequence encoded by lysA is shown in SEQ ID NO: 20. In addition to DNA fragments coding for these amino acid sequences, the present invention can equivalently use DNA fragments coding for amino acid sequences substantially the same as the amino acid sequence shown in SEQ ID NO: 20, namely amino acid sequences having mutation based on, for example, substitution, deletion, or insertion of one or more amino acids provided that there is no substantial influence on the DDC activity.

(5) Preparation of ddh

A DNA fragment containing ddh can be prepared from chromosome of a coryneform bacterium by means of PCR. The DNA donor is not specifically limited, however, it is exemplified by Brevibacterium lactofermentum ATCC 13869 strain.

A DDH gene is known for Corynebacterium glutamicum (Ishino, S. et al., Nucleic Acids Res., 15, 3917

(1987)), on the basis of which primers for PCR can be prepared. Such DNA primers are specifically exemplified by DNA's of 20-mers respectively having nucleotide sequences depicted in SEQ ID NOs: 21 and 22 in Sequence Listing. Synthesis of DNA, PCR, and preparation of a plasmid containing obtained ddh can be performed in the same manner as those for lysC described above.

A nucleotide sequence of a DNA fragment containing ddh and an amino acid sequence deduced from the nucleotide sequence are illustrated in SEQ ID NO: 23. Only the amino acid sequence is shown in SEQ ID NO: 24. In addition to DNA fragments coding for this amino acid sequence, the present invention can equivalently use DNA fragments coding for amino acid sequences substantially the same as the amino acid sequence shown in SEQ ID NO: 24, namely amino acid sequences having mutation based on, for example, substitution, deletion, or insertion of one or more amino acids provided that there is no substantial influence on the DDH activity.

<2> Recombinant DNA and coryneform bacterium of the present invention

The coryneform bacterium of the present invention harbors an aspartokinase (mutant AK) in which feedback inhibition by L-lysine and L-threonine is substantially desensitized, wherein DNA (dapB) coding for a dihydrodipicolinate reductase is enhanced. In a preferred embodiment, the coryneform bacterium of the present invention is a coryneform bacterium in which DNA (dapA) coding for dihydrodipicolinate synthase is

further enhanced. In a more preferred embodiment, the coryneform bacterium of the present invention is a coryneform bacterium in which DNA (lysA) coding for diaminopimelate decarboxylase is further enhanced. In a more preferred embodiment, the coryneform bacterium of the present invention is a coryneform bacterium in which DNA (ddh) coding for diaminopimelate dehydrogenase is further enhanced.

The term "enhance" DNA herein refers to the fact that the intracellular activity of an enzyme encoded by the DNA is raised by, for example, increasing the copy number of a gene, using a strong promoter, using a gene coding for an enzyme having a high specific activity, or combining these means.

The coryneform bacterium harboring the mutant AK may be those which produce the mutant aspartokinase as a result of mutation, or those which are transformed by introducing mutant lysC.

Examples of the coryneform bacterium used to introduce the DNA described above include, for example, the following lysine-producing wild type strains:

Corynebacterium acetoacidophilum ATCC 13870;

Corynebacterium acetoglutamicum ATCC 15806;

Corynebacterium callunae ATCC 15991;

Corynebacterium glutamicum ATCC 13032;

(Brevibacterium divaricatum) ATCC 14020;

(Brevibacterium lactofermentum) ATCC 13869;

(Corynebacterium lilium) ATCC 15990;

(Brevibacterium flavum) ATCC 14067;

Corynebacterium melassecola ATCC 17965;

Brevibacterium saccharolyticum ATCC 14066;

Brevibacterium immariophilum ATCC 14068;

Brevibacterium roseum ATCC 13825;

Brevibacterium thiogenitalis ATCC 19240;

Microbacterium ammoniaphilum ATCC 15354;

Corynebacterium thermoaminogenes AJ12340 (FERM BP-1539).

Other than the bacterial strains described above, those usable as a host include, for example, mutant strains having an L-lysine-producing ability derived from the aforementioned strains. Such artificial mutant strains includes the followings: S-(2-aminoethyl)-cysteine (hereinafter abbreviated as "AEC") resistant mutant strains (Brevibacterium lactofermentum AJ11082 (NRRL B-1147), Japanese Patent Publication Nos. 56-1914, 56-1915, 57-14157, 57-14158, 57-30474, 58-10075, 59-4993, 61-35840, 62-24074, 62-36673, 5-11958, 7-112437, and 7-112438); mutant strains which require amino acid such as L-homoserine for their growth (Japanese Patent Publication Nos. 48-28078 and 56-6499); mutant strains which exhibit resistance to AEC and require amino acids such as L-leucine, L-homoserine, L-proline, L-serine, L-arginine, L-alanine, and L-valine (United States Patent Nos. 3,708,395 and 3,825,472); L-lysine-producing mutant strains which exhibit resistance to DL-α-amino-ϵ-caprolactam, α-amino-lauryllactam, aspartate-analog, sulfa drug, quinoid, and N-lauroylleucine; L-lysine-producing mutant strains which exhibit resistance to inhibitors of oxyaloacetate decarboxylase or respiratory system enzymes (Japanese Patent Laid-open Nos. 50-53588, 50-31093, 52-102498, 53-9394, 53-86089, 55-9783, 55-

9759, 56-32995 and 56-39778, and Japanese Patent Publication Nos. 53-43591 and 53-1833); L-lysine-producing mutant strains which require inositol or acetic acid (Japanese Patent Laid-open Nos. 55-9784 and 56-8692); L-lysine-producing mutant strains which exhibit sensitivity to fluoropyruvic acid or temperature not less than 34 °C (Japanese Patent Laid-open Nos. 55-9783 and 53-86090); and producing mutant strains belonging to the genus Brevibacterium or Corynebacterium which exhibit resistance to ethylene glycol and produce L-lysine (United States Patent No. 4,411,997).

In a specified embodiment, in order to enhance the genes for L-lysine biosynthesis in the host as described above, the genes are introduced into the host by using a plasmid vector, transposon or phage vector or the like. Upon the introduction, it is expected to make enhancement to some extent even by using a low copy type vector. However, it is preferred to use a multiple copy type vector. Such a vector includes, for example, plasmid vectors, pAJ655, pAJ1844, pAJ611, pAJ3148, and pAJ440 described above. Besides, transposons derived from coryneform bacteria are described in International Publication Pamphlets of WO02/02627 and WO93/18151, European Patent Publication No. 445385, Japanese Patent Laid-open No. 6-46867, Vertes, A. A. et al., Mol. Microbiol., 11, 739-746 (1994), Bonamy, C., et al., Mol. Microbiol., 14, 571-581 (1994), Vertes, A. A. et al., Mol. Gen. Genet., 245, 397-405 (1994), Jagar, W. et al., FEMS Microbiology Letters, 126, 1-6 (1995), Japanese Patent Laid-open No. 7-107976, Japanese Patent Laid-open

No. 7-327680 and the like.

In the present invention, it is not indispensable that the mutant lysC is necessarily enhanced. It is allowable to use those which have mutation on lysC on chromosomal DNA, or in which the mutant lysC is incorporated into chromosomal DNA. Alternatively, the mutant lysC may be introduced by using a plasmid vector. On the other hand, dapA, dapB, lysA, and ddh are preferably enhanced in order to efficiently produce L-lysine.

Each of the genes of lysC, dapA, dapB, lysA, and ddh may be successively introduced into the host by using different vectors respectively. Alternatively, two, three, four, or five species of the genes may be introduced together by using a single vector. When different vectors are used, the genes may be introduced in any order, however, it is preferred to use vectors which have a stable sharing and harboring mechanism in the host, and which are capable of co-existing with each other.

A coryneform bacterium harboring the mutant AK and further comprising enhanced dapB is obtained, for example, by introducing, into a host coryneform bacterium, a recombinant DNA containing mutant lysC and dapB autonomously replicable in cells of coryneform bacteria.

A coryneform bacterium further comprising enhanced dapA in addition to mutant lysC and dapB is obtained, for example, by introducing, into a host coryneform bacterium, a recombinant DNA containing mutant lysC,

dapB, and dapA autonomously replicable in cells of coryneform bacteria.

A coryneform bacterium further comprising enhanced lysA in addition to mutant lysC, dapB, and dapA is obtained, for example, by introducing, into a host coryneform bacterium, a recombinant DNA containing mutant lysC, dapB, dapA, and lysA autonomously replicable in cells of coryneform bacteria.

A coryneform bacterium further comprising enhanced ddh in addition to mutant lysC, dapB, dapA, and lysA is obtained, for example, by introducing, into a host coryneform bacterium, a recombinant DNA containing mutant lysC, dapB, dapA, lysA, and ddh autonomously replicable in cells of coryneform bacteria.

The above-mentioned recombinant DNAs can be obtained, for example, by inserting each of the genes participating in L-lysine biosynthesis into a vector such as plasmid vector, transposon or phage vector as described above.

In the case in which a plasmid is used as a vector, the recombinant DNA can be introduced into the host in accordance with an electric pulse method (Sugimoto et al., Japanese Patent Laid-open No. 2-207791). Amplification of a gene using transposon can be performed by introducing a plasmid which carrying a transposon into the host cell and inducing transposition of the transposon.

<3> Method for producing L-lysine

L-Lysine can be efficiently produced by cultivating, in an appropriate medium, the coryneform bacterium comprising the enhanced genes for L-lysine

biosynthesis as described above, producing and accumulating L-lysine in a culture of the bacterium, and collecting L-lysine from the culture.

The medium to be used is exemplified by an ordinary medium containing a carbon source, a nitrogen source, inorganic ions, and optionally other organic components.

As the carbon source, it is possible to use sugars such as glucose, fructose, sucrose, molasses, and starch hydrolysate; and organic acids such as fumaric acid, citric acid, and succinic acid.

As the nitrogen source, it is possible to use inorganic ammonium salts such as ammonium sulfate, ammonium chloride, and ammonium phosphate; organic nitrogen such as soybean hydrolysate; ammonia gas; and aqueous ammonia.

As organic trace nutrient sources, it is desirable to contain required substances such as vitamin B_1 and L-homoserine or yeast extract or the like in appropriate amounts. Other than the above, potassium phosphate, magnesium sulfate, iron ion, manganese ion and so on are added in small amounts, if necessary.

Cultivation is preferably carried out under an aerobic condition for about 30 to 90 hours. The cultivation temperature is preferably controlled at 25 °C to 37 °C, and pH is preferably controlled at 5 to 8 during cultivation. Inorganic or organic, acidic or alkaline substances, or ammonia gas or the like can be used for pH adjustment. L-lysine can be collected from a culture by combining an ordinary ion exchange resin method, a precipitation method, and other known methods.

Brief Description of the Drawings

Fig. 1 illustrates a process of construction of plasmids p399AKYB and p399AK9B comprising mutant lysC.

Fig. 2 illustrates a process of construction of a plasmid pDPRB comprising dapB and Brevi.-ori.

Fig. 3 illustrates a process of construction of a plasmid pDPSB comprising dapA and Brevi.-ori.

Fig. 4 illustrates a process of construction of a plasmid p299LYSA comprising lysA.

Fig. 5 illustrates a process of construction of a plasmid pLYSAB comprising lysA and Brevi.-ori.

Fig. 6 illustrates a process of construction of a plasmid pPK4D comprising ddh and Brevi.-ori.

Fig. 7 illustrates a process of construction of a plasmid pCRCAB comprising lysC, dapA and Brevi.-ori.

Fig. 8 illustrates a process of construction of a plasmid pCB comprising mutant lysC, dapB, and Brevi.-ori.

Fig. 9 illustrates a process of construction of a plasmid pAB comprising dapA, dapB and Brevi.-ori.

Fig. 10 illustrates a process of construction of a plasmid p399DL comprising ddh and lysA.

Fig. 11 illustrates a process of construction of a plasmid pDL comprising ddh, lysA and Brevi.-ori.

Fig. 12 illustrates a process of construction of a plasmid pCAB comprising mutant lysC, dapA, dapB, and Brevi.-ori.

Fig. 13 illustrates a process of construction of a plasmid pCABL comprising mutant lysC, dapA, dapB, lysA, and Brevi.-ori.

Fig. 14 illustrates a process of construction of a plasmid pCABDL comprising mutant lysC, dapA, dapB, ddh, lysA, and Brevi.-ori.

Description of Preferred Embodiments

The present invention will be more specifically explained below with reference to Examples.

Example 1: Preparation of Wild Type lysC Gene and Mutant lysC Gene from Brevibacterium lactofermentum

<1> Preparation of wild type and mutant lysC's and preparation of plasmids containing them

A strain of Brevibacterium lactofermentum ATCC 13869, and an L-lysine-producing mutant strain AJ3445 (FERM P-1944) obtained from the ATCC 13869 strain by a mutation treatment were used as chromosomal DNA donors. The AJ3445 strain had been subjected to mutation so that lysC was changed to involve substantial desensitization

from concerted inhibition by lysine and threonine (Journal of Biochemistry, 68, 701-710 (1970)).

A DNA fragment containing lysC was amplified from chromosomal DNA in accordance with the PCR method (polymerase chain reaction; see White, T. J. et al., Trends Genet., 5, 185 (1989)). As for DNA primers used for amplification, single strand DNA's of 23-mer and 21-mer having nucleotide sequences shown in SEQ ID NOs: 1 and 2 were synthesized in order to amplify a region of about 1,643 bp coding for lysC on the basis of a sequence known for Corynebacterium glutamicum (see Molecular Microbiology (1991), 5(5), 1197-1204; and Mol. Gen. Genet. (1990), 224, 317-324). DNA was synthesized in accordance with an ordinary method by using DNA synthesizer model 380B produced by Applied Biosystems and using the phosphoamidite method (see Tetrahedron Letters (1981), 22, 1859).

The gene was amplified by PCR by using DNA Thermal Cycler Model PJ2000 produced by Takara Shuzo, and using Taq DNA polymerase in accordance with a method designated by the supplier. An amplified gene fragment of 1,643 kb was confirmed by agarose gel electrophoresis. After that, the fragment excised from the gel was purified in accordance with an ordinary method, and it was digested with restriction enzymes NruI (produced by Takara Shuzo) and EcoRI (produced by Takara Shuzo).

pHSG399 (see Takeshita, S. et al., Gene (1987), 61, 63-74) was used as a cloning vector for the gene fragment. pHSG399 was digested with restriction enzymes SmaI (produced by Takara Shuzo) and EcoRI, and it was ligated with the amplified lysC fragment. DNA was ligated by using DNA ligation kit (produced by Takara Shuzo) in accordance with a designated method. Thus plasmids were prepared, in which the lysC fragments amplified from chromosomes of Brevibacterium lactofermentum were ligated with pHSG399 respectively. A plasmid comprising lysC from ATCC 13869 (wild type strain) was designated as p399AKY, and a plasmid comprising lysC from AJ3463 (L-lysine-producing bacterium) was designated as p399AK9.

A DNA fragment (hereinafter referred to as "Brevi.-ori") having an ability to make a plasmid autonomously replicable in bacteria belonging to the genus Corynebacterium was introduced into p399AKY and p399AK9 respectively to prepare plasmids carrying lysC autonomously replicable in bacteria belonging to the genus Corynebacterium. Brevi.-ori was prepared from a plasmid vector pHK4 containing Brevi.-ori and autonomously replicable in cells of both Escherichia coli and bacteria belonging to the genus Corynebacterium. pHK4 was constructed by digesting pHC4 with KpnI (produced by Takara Shuzo) and BamHI (produced by Takara Shuzo), extracting a Brevi.-ori fragment, and

ligating it with pHSG298 having been also digested with KpnI and BamHI (see Japanese Patent Laid-open No. 5-7491). pHK4 gives kanamycin resistance to a host. Escherichia coli harboring pHK4 was designated as Escherichia coli AJ13136, and deposited on August 1, 1995 under a deposition number of FERM BP-5186 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan).

pHK4 was digested with restriction enzymes KpnI and BamHI, and cleaved edges were blunt-ended. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated BamHI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only BamHI. This plasmid was digested with BamHI, and the generated Brevi.-ori DNA fragment was ligated with p399AKY and p399AK9 having been also digested with BamHI respectively to prepare plasmids each containing the lysC gene autonomously replicable in bacteria belonging to the genus Corynebacterium.

A plasmid containing the wild type lysC gene originating from p399AKY was designated as p399AKYB, and

a plasmid containing the mutant lysC gene originating from p399AK9 was designated as p399AK9B. The process of construction of p399AK9B and p399AKYB is shown in Fig. 1. A strain AJ12691 obtained by introducing the mutant lysC plasmid p399AK9B into a wild type strain of Brevibacterium lactofermentum (AJ12036 strain, FERM BP-734) was deposited on April 10, 1992 under a deposition number of FERM P-12918 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan), transferred to international deposition based on the Budapest Treaty on February 10, 1995, and deposited under a deposition number of FERM BP-4999.

<2> Determination of nucleotide sequences of wild type lysC and mutant lysC from Brevibacterium lactofermentum

The plasmid p399AKY containing the wild type lysC and the plasmid p399AK9 containing the mutant lysC were prepared from the respective transformants to determine nucleotide sequences of the wild type and mutant lysC's. Nucleotide sequence determination was performed in accordance with a method of Sanger et al. (for example, F. Sanger et al., Proc. Natl. Acad. Sci., 74, 5463 (1977)).

The nucleotide sequence of wild type lysC encoded

by p399AKY is shown in SEQ ID NO: 3 in Sequence Listing. On the other hand, the nucleotide sequence of mutant lysC encoded by p399AK9 had only mutation of one nucleotide such that 1051th G was changed into A in SEQ ID NO: 3 as compared with wild type lysC. It is known that lysC of Corynebacterium glutamicum has two subunits (α, β) encoded in an identical reading frame on an identical DNA strand (see Kalinowski, J. et al., Molecular Microbiology (1991) 5(5), 1197-1204). Judging from homology, it is assumed that the gene sequenced herein also has two subunits (α, β) encoded in an identical reading frame on an identical DNA strand.

An amino acid sequence of the α-subunit of the wild type AK protein deduced from the nucleotide sequence of DNA is shown in SEQ ID NO: 4 together with the DNA sequence. Only the amino acid sequence is shown in SEQ ID NO: 5. An amino acid sequence of the β-subunit of the wild type AK protein deduced from the nucleotide sequence of DNA is shown in SEQ ID NO: 6 together with DNA. Only the amino acid sequence is shown in SEQ ID NO: 7. In each of the subunits, GTG is used as an initiation codon, and a corresponding amino acid is represented by methionine. However, this representation refers to methionine, valine, or formylmethionine.

On the other hand, mutation on the sequence of mutant lysC means occurrence of amino acid residue substitution such that a 279th alanine residue of the α-

subunit is changed into a threonine residue, and a 30th alanine residue of the β-subunit is changed into a threonine residue in the amino acid sequence of the wild type AK protein (SEQ ID NOs: 5, 7).

Example 2: Preparation of dapB from Brevibacterium lactofermentum

<1> Preparation of dapB and construction of plasmid containing dapB

A wild type strain of Brevibacterium lactofermentum ATCC 13869 was used as a chromosomal DNA donor. Chromosomal DNA was prepared from the ATCC 13869 strain in accordance with an ordinary method. A DNA fragment containing dapB was amplified from the chromosomal DNA in accordance with PCR. As for DNA primers used for amplification, DNA's of 23-mers having nucleotide sequences depicted in SEQ ID NOs: 8 and 9 in Sequence Listing respectively were synthesized in order to amplify a region of about 2.0 kb coding for DDPR on the basis of a sequence known for Brevibacterium lactofermentum (see Journal of Bacteriology, 157(9), 2743-2749 (1993)). Synthesis of DNA and PCR were performed in the same manner as described in Example 1. pCR-Script (produced by Invitrogen) was used as a cloning vector for the amplified gene fragment of 2,001 bp, which was ligated with the amplified dapB fragment.

Thus a plasmid was constructed, in which the dapB fragment of 2,001 bp amplified from chromosome of Brevibacterium lactofermentum was ligated with pCR-Script. The plasmid obtained as described above, which had dapB originating from ATCC 13869, was designated as pCRDAPB. A transformant strain AJ13107 obtained by introducing pCRDAPB into E. coli JM109 strain has been internationally deposited since May 26, 1995 under a deposition number of FERM BP-5114 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) based on the Budapest Treaty.

A fragment of 1,101 bp containing a structural gene of DDPR was extracted by digesting pCRDAPB with EcoRV and SphI. This fragment was ligated with pHSG399 having been digested with HincII and SphI to prepare a plasmid. The prepared plasmid was designated as p399DPR.

Brevi.-ori was introduced into the prepared p399DPR to construct a plasmid carrying dapB autonomously replicable in coryneform bacteria. pHK4 was digested with a restriction enzyme KpnI (produced by Takara Shuzo), and cleaved edges were blunt-ended. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a

phosphorylated BamHI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only BamHI. This plasmid was digested with BamHI, and the generated Brevi.-ori DNA fragment was ligated with p399DPR having been also digested with BamHI to prepare a plasmid containing dapB autonomously replicable in coryneform bacteria. The prepared plasmid was designated as pDPRB. The process of construction of pDPRB is shown in Fig. 2.

<2> Determination of nucleotide sequence of dapB from Brevibacterium lactofermentum

Plasmid DNA was prepared from the AJ13107 strain harboring p399DPR, and its nucleotide sequence was determined in the same manner as described in Example 1. A determined nucleotide sequence and an amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 10. Only the amino acid sequence is shown in SEQ ID NO: 11.

Example 3: Preparation of dapA from Brevibacterium lactofermentum

<1> Preparation of dapA and construction of plasmid containing dapA

A wild type strain of Brevibacterium lactofermentum

ATCC 13869 was used as a chromosomal DNA donor. Chromosomal DNA was prepared from the ATCC 13869 strain in accordance with an ordinary method. A DNA fragment containing dapA was amplified from the chromosomal DNA in accordance with PCR. As for DNA primers used for amplification, DNA's of 20-mers having nucleotide sequences shown in SEQ ID NOs: 12 and 13 in Sequence Listing respectively were synthesized in order to amplify a region of about 1.5 kb coding for DDPS on the basis of a sequence known for Corynebacterium glutamicum (see Nucleic Acids Research, 18(21), 6421 (1990); EMBL accession No. X53993). Synthesis of DNA and PCR were performed in the same manner as described in Example 1. pCR1000 (produced by Invitrogen, see Bio/Technology, 9, 657-663 (1991)) was used as a cloning vector for the amplified gene fragment of 1,411 bp, which was ligated with the amplified dapA fragment. Ligation of DNA was performed by using DNA ligation kit (produced by Takara Shuzo) in accordance with a designated method. Thus a plasmid was constructed, in which the dapA fragment of 1,411 bp amplified from chromosome of Brevibacterium lactofermentum was ligated with pCR1000. The plasmid obtained as described above, which had dapA originating from ATCC 13869, was designated as pCRDAPA.

A transformant strain AJ13106 obtained by introducing pCRDAPA into E. coli JM109 strain has been internationally deposited since May 26, 1995 under a

deposition number of FERM BP-5113 in National Institute of Bioscience and Human Technology of Agency of Industrial Science and Technology of Ministry of International Trade and Industry (postal code: 305, 1-3, Higashi 1-chome, Tsukuba-shi, Ibaraki-ken, Japan) based on the Budapest Treaty.

Brevi.-ori was introduced into the prepared pCRDAPA to construct a plasmid carrying dapA autonomously replicable in coryneform bacteria. pHK4 was digested with restriction enzymes KpnI and BamHI (produced by Takara Shuzo), and cleaved edges were blunt-ended. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated SmaI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only SmaI. This plasmid was digested with SmaI, and the generated Brevi.-ori DNA fragment was ligated with pCRDAPA having been also digested with SmaI to prepare a plasmid containing dapA autonomously replicable in coryneform bacteria. This plasmid was designated as pDPSB. The process of construction of pDPSB(Km^r) is shown in Fig. 3.

<2> Determination of nucleotide sequence of dapA from

Brevibacterium lactofermentum

Plasmid DNA was prepared from the AJ13106 strain harboring pCRDAPA, and its nucleotide sequence was determined in the same manner as described in Example 1. A determined nucleotide sequence and an amino acid sequence deduced from the nucleotide sequence are shown in SEQ ID NO: 14. Only the amino acid sequence is shown in SEQ ID NO: 15.

Example 4: Preparation of lysA from Brevibacterium lactofermentum

<1> Preparation of lysA and construction of plasmid containing lysA

A wild type strain of Brevibacterium lactofermentum ATCC 13869 was used as a chromosomal DNA donor. Chromosomal DNA was prepared from the ATCC 13869 strain in accordance with an ordinary method. A DNA fragment containing argS, lysA, and a promoter of an operon containing them was amplified from the chromosomal DNA in accordance with PCR. As for DNA primers used for amplification, synthetic DNA's of 23-mers having nucleotide sequences depicted in SEQ ID NOs: 16 and 17 in Sequence Listing respectively were used in order to amplify a region of about 3.6 kb coding for arginyl-tRNA synthase and DDC on the basis of a sequence known for Corynebacterium glutamicum (see Molecular Microbiology,

4(11), 1819-1830 (1990); Molecular and General Genetics, 212, 112-119 (1988)). Synthesis of DNA and PCR were performed in the same manner as described in Example 1. pHSG399 was used as a cloning vector for the amplified gene fragment of 3,579 bp. pHSG399 was digested with a restriction enzyme SmaI (produced by Takara Shuzo), which was ligated with the DNA fragment containing amplified lysA. A plasmid obtained as described above, which had lysA originating from ATCC 13869, was designated as p399LYSA.

A DNA fragment containing lysA was extracted by digesting p399LYSA with KpnI (produced by Takara Shuzo) and BamHI (produced by Takara Shuzo). This DNA fragment was ligated with pHSG299 having been digested with KpnI and BamHI. An obtained plasmid was designated as p299LYSA. The process of construction of p299LYSA is shown in Fig. 4.

Brevi.-ori was introduced into the obtained p299LYSA to construct a plasmid carrying lysA autonomously replicable in coryneform bacteria. pHK4 was digested with restriction enzymes KpnI and BamHI, and cleaved edges were blunt-ended. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated KpnI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to

the Brevi.-ori portion might be excised from pHK4 by digestion with only KpnI. This plasmid was digested with KpnI, and the generated Brevi.-ori DNA fragment was ligated with p299LYSA having been also digested with KpnI to prepare a plasmid containing lysA autonomously replicable in coryneform bacteria. The prepared plasmid was designated as pLYSAB. The process of construction of pLYSAB is shown in Fig. 5.

<2> Determination of nucleotide sequence of lysA from Brevibacterium lactofermentum

Plasmid DNA of p299LYSA was prepared, and its nucleotide sequence was determined in the same manner as described in Example 1. A determined nucleotide sequence and an amino acid sequence deduced to be encoded by the nucleotide sequence are shown in SEQ ID NO: 18. Concerning the nucleotide sequence, an amino acid sequence encoded by argS and an amino acid sequence encoded by lysA are shown in SEQ ID NOs: 19 and 20 respectively.

Example 5: Preparation of ddh from Brevibacterium lactofermentum

A ddh gene was obtained by amplifying the ddh gene from chromosomal DNA of Brevibacterium lactofermentum ATCC 13869 in accordance with the PCR method by using

two oligonucleotide primers (SEQ ID NOs: 21, 22) prepared on the basis of a known nucleotide sequence of a ddh gene of Corynebacterium glutamicum (Ishino, S. et al., Nucleic Acids Res., 15, 3917 (1987)). An obtained amplified DNA fragment was digested with EcoT22I and AvaI, and cleaved edges were blunt-ended. After that, the fragment was inserted into a SmaI site of pMW119 to obtain a plasmid pDDH.

Next, pDDH was digested with SalI and EcoRI, followed by blunt end formation. After that, an obtained fragment was ligated with pUC18 having been digested with SmaI. A plasmid thus obtained was designated as pUC18DDH.

Brevi.-ori was introduced into pUC18DDH to construct a plasmid carrying ddh autonomously replicable in coryneform bacteria. pHK4 was digested with restriction enzymes KpnI and BamHI, and cleaved edges were blunt-ended. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated PstI linker (produced by Takara Shuzo) was ligated so that it was inserted into a PstI site of pHSG299. A plasmid constructed as described above was designated as pPK4. Next, pUC18DDH was digested with XbaI and KpnI, and a generated fragment was ligated with pPK4 having been digested with KpnI and XbaI. Thus a plasmid containing ddh

autonomously replicable in coryneform bacteria was constructed. This plasmid was designated as pPK4D. The process of construction of pPK4D is shown in Fig. 6.

Example 6: Construction of Plasmid Comprising Combination of Mutant lysC and dapA

A plasmid comprising mutant lysC, dapA, and replication origin of coryneform bacteria was constructed from the plasmid pCRDAPA comprising dapA and the plasmid p399AK9B comprising mutant lysC and Brevi.-ori. p399AK9B was completely degraded with SalI, and then it was blunt-ended, with which an EcoRI linker was ligated to construct a plasmid in which the SalI site was modified into an EcoRI site. The obtained plasmid was designated as p399AK9BSE. The mutant lysC and Brevi.-ori were excised as one fragment by partially degrading p399AK9BSE with EcoRI. This fragment was ligated with pCRDAPA having been digested with EcoRI. An obtained plasmid was designated as pCRCAB. This plasmid is autonomously replicable in E. coli and coryneform bacteria, and it gives kanamycin resistance to a host, the plasmid comprising a combination of mutant lysC and dapA. The process of construction of pCRCAB is shown in Fig. 7.

Example 7: Construction of Plasmid Comprising Combination of Mutant lysC and dapB

A plasmid comprising mutant lysC and dapB was constructed from the plasmid p399AK9 having mutant lysC and the plasmid p399DPR having dapB. A fragment of 1,101 bp containing a structural gene of DDPR was extracted by digesting p399DPR with EcoRV and SphI. This fragment was ligated with p399AK9 having been digested with SalI and then blunt-ended and having been further digested with SphI to construct a plasmid comprising a combination of mutant lysC and dapB. This plasmid was designated as p399AKDDPR.

Next, Brevi.-ori was introduced into the obtained p399AKDDPR. The plasmid pHK4 containing Brevi.-ori was digested with a restriction enzyme KpnI (produced by Takara Shuzo), and cleaved edges were blunt-ended. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated BamHI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only BamHI. This plasmid was digested with BamHI, and the generated Brevi.-ori DNA fragment was ligated with p399AKDDPR having been also digested with BamHI to construct a

plasmid containing mutant lysC and dapB autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pCB. The process of construction of pCB is shown in Fig. 8.

Example 8: Construction of Plasmid Comprising Combination of dapA and dapB

The plasmid pCRDAPA comprising dapA was digested with KpnI and EcoRI to extract a DNA fragment containing dapA which was ligated with the vector plasmid pHSG399 having been digested with KpnI and EcoRI. An obtained plasmid was designated as p399DPS.

On the other hand, the plasmid pCRDAPB comprising dapB was digested with SacII and EcoRI to extract a DNA fragment of 2.0 kb containing a region coding for DDPR which was ligated with p399DPS having been digested with SacII and EcoRI to construct a plasmid comprising a combination of dapA and dapB. The obtained plasmid was designated as p399AB.

Next, Brevi.-ori was introduced into p399AB. pHK4 containing Brevi.-ori was digested with a restriction enzyme BamHI (produced by Takara Shuzo), and cleaved edges were blunt-ended. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated KpnI linker

(produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only KpnI. This plasmid was digested with KpnI, and the generated Brevi.-ori DNA fragment was ligated with p399AB having been also digested with KpnI to construct a plasmid containing dapA and dapB autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pAB. The process of construction of pAB is shown in Fig. 9.

Example 9: Construction of Plasmid Comprising Combination of ddh and lysA

The plasmid pUC18DDH comprising ddh was digested with EcoRI and XbaI to extract a DNA fragment containing ddh. This ddh fragment was ligated with the plasmid p399LYSA comprising lysA having been digested with BamHI and XbaI with cleaved edges having been blunt-ended after the digestion. An obtained plasmid was designated as p399DL. The process of construction of p399DL is shown in Fig. 10.

Next, Brevi.-ori was introduced into p399DL. pHK4 was digested with XbaI and BamHI, and cleaved edges were blunt-ended. After the blunt end formation, a phosphorylated XbaI linker was ligated to make modification so that the DNA fragment corresponding to

the Brevi.-ori portion might be excised from pHK4 by digestion with only XbaI. This plasmid was digested with XbaI, and the generated Brevi.-ori DNA fragment was ligated with p399DL having been also digested with XbaI to construct a plasmid containing ddh and lysA autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pDL. The process of construction of pDL is shown in Fig. 11.

Example 10: Construction of Plasmid Comprising Combination of Mutant lysC, dapA, and dapB

p399DPS was degraded with EcoRI and SphI to form blunt ends followed by extraction of a dapA gene fragment. This fragment was ligated with the p399AK9 having been digested with SalI and blunt-ended to construct a plasmid p399CA in which mutant lysC and dapA co-existed.

The plasmid pCRDAPB comprising dapB was digested with EcoRI and blunt-ended, followed by digestion with SacI to extract a DNA fragment of 2.0 kb comprising dapB. The plasmid p399CA comprising dapA and mutant lysC was digested with SpeI and blunt-ended, which was thereafter digested with SacI and ligated with the extracted dapB fragment to obtain a plasmid comprising mutant lysC, dapA, and dapB. This plasmid was designated as p399CAB.

Next, Brevi.-ori was introduced into p399CAB. The plasmid pHK4 comprising Brevi.-ori was digested with a restriction enzyme BamHI (produced by Takara Shuzo), and cleaved edges were blunt-ended. Blunt end formation was performed by using DNA Blunting kit (produced by Takara Shuzo) in accordance with a designated method. After the blunt end formation, a phosphorylated KpnI linker (produced by Takara Shuzo) was ligated to make modification so that the DNA fragment corresponding to the Brevi.-ori portion might be excised from pHK4 by digestion with only KpnI. This plasmid was digested with KpnI, and the generated Brevi.-ori DNA fragment was ligated with p399CAB having been also digested with KpnI to construct a plasmid comprising a combination of mutant lysC, dapA, and dapB autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pCAB. The process of construction of pCAB is shown in Fig. 12.

Example 11: Construction of Plasmid Comprising Combination of Mutant lysC, dapA, dapB, and lysA

The plasmid p299LYSA comprising lysA was digested with KpnI and BamHI and blunt-ended, and then a lysA gene fragment was extracted. This fragment was ligated with pCAB having been digested with HpaI (produced by Takara Shuzo) and blunt-ended to construct a plasmid

comprising a combination of mutant lysC, dapA, dapB, and lysA autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pCABL. The process of construction of pCABL is shown in Fig. 13. It is noted that the lysA gene fragment is inserted into a HpaI site in a DNA fragment containing the dapB gene in pCABL, however, the HpaI site is located upstream from a promoter for the dapB gene (nucleotide numbers 611 to 616 in SEQ ID NO: 10), and the dapB gene is not decoupled.

Example 12: Construction of Plasmid Comprising Combination of Mutant lysC, dapA, dapB, ddh, and lysA

pHSG299 was digested with XbaI and KpnI, which was ligated with p399DL comprising ddh and lysA having been digested with XbaI and KpnI. A constructed plasmid was designated as p299DL. p299DL was digested with XbaI and KpnI and blunt-ended. After the blunt end formation, a DNA fragment comprising ddh and lysA was extracted. This DNA fragment was ligated with the plasmid pCAB comprising the combination of mutant lysC, dapA, and dapB having been digested with HpaI and blunt-ended to construct a plasmid comprising a combination of mutant lysC, dapA, dapB, lysA and ddh autonomously replicable in coryneform bacteria. The constructed plasmid was designated as pCABDL. The process of construction of

pCABDL is shown in Fig. 14.

Example 13: Introduction of Plasmids Comprising Genes for L-Lysine Biosynthesis into L-Lysine-Producing Bacterium of Brevibacterium lactofermentum

The plasmids comprising the genes for L-lysine biosynthesis constructed as described above, namely p399AK9B(Cm^r), pDPSB(Km^r), pDPRB(Cm^r), pLYSAB(Cm^r), pPK4D(Cm^r), pCRCAB(Km^r), pAB(Cm^r), pCB(Cm^r), pDL(Cm^r), pCAB(Cm^r), pCABL(Cm^r), and pCABDL(Cm^r) were introduced into an L-lysine-producing bacterium AJ11082 (NRRL B-11470) of Brevibacterium lactofermentum respectively. AJ11082 strain has a property of AEC resistance. The plasmids were introduced in accordance with an electric pulse method (Sugimoto et al., Japanese Patent Laid-open No. 2-207791). Transformants were selected based on drug resistance markers possessed by the respective plasmids. Transformants were selected on a complete medium containing 5 µg/ml of chloramphenicol when a plasmid comprising a chloramphenicol resistance gene was introduced, or transformants were selected on a complete medium containing 25 µg/ml of kanamycin when a plasmid comprising a kanamycin resistance gene was introduced.

Example 14: Production of L-Lysine

Each of the transformants obtained in Example 13 was cultivated in an L-lysine-producing medium to evaluate its L-lysine productivity. The L-lysine-producing medium had the following composition.

[L-Lysine-producing medium]

The following components other than calcium carbonate (per 1 L) were dissolved to make adjustment at pH 8.0 with KOH. The medium was sterilized at 115 °C for 15 minutes, to which calcium carbonate (50 g) having been separately sterilized in hot air in a dry state was thereafter added.

Glucose	100 g
$(NH_4)_2SO_4$	55 g
KH_2PO_4	1 g
$MgSO_4 \cdot 7H_2O$	1 g
Biotin	500 µg
Thiamin	2000 µg
$FeSO_4 \cdot 7H_2O$	0.01 g
$MnSO_4 \cdot 7H_2O$	0.01 g
Nicotinamide	5 mg
Protein hydrolysate (Mamenou)	30 ml
Calcium carbonate	50 g

Each of the various types of the transformants and the parent strain was inoculated to the medium having the composition described above to perform cultivation at 31.5 °C with reciprocating shaking. The amount of

produced L-lysine after 40 or 72 hours of cultivation, and the growth after 72 hours (OD_{562}) are shown in Table 1. In the table, lysC* represents mutant lysC. The growth was quantitatively determined by measuring OD at 560 nm after 101-fold dilution.

Table 1

Accumulation of L-Lysine after Cultivation for 40 or 72 Hours

Bacterial strain /plasmid	Introduced gene	Amount of produced L-lysine(g/L) after 40 hrs	Amount of produced L-lysine(g/L) after 72 hrs	Growth (OD_{562}/101)
AJ11082		22.0	29.8	0.450
AJ11082/p399AK9B	lysC*	16.8	34.5	0.398
AJ11082/pDPSB	dapA	18.7	33.8	0.410
AJ11082/pDPRB	dapB	19.9	29.9	0.445
AJ11082/pLYSAB	lysA	19.8	32.5	0.356
AJ11082/pPK4D	ddh	19.0	33.4	0.330
AJ11082/pCRCAB	lysC*, dapA	19.7	36.5	0.360
AJ11082/pAB	dapA, dapB	19.0	34.8	0.390
AJ11082/pCB	lysC*, dapB	23.3	35.0	0.440
AJ11082/pDL	ddh, lysA	23.3	31.6	0.440
AJ11082/pCAB	lysC*, dapA, dapB	23.0	45.0	0.425
AJ11082/pCABL	lysC*, dapA, dapB, lysA	26.2	46.5	0.379
AJ11082/pCABDL	lysC*, dapA, dapB, lysA, ddh	26.5	47.0	0.409

As shown in Table 1, when mutant lysC, dapA, or dapB was enhanced singly, the amount of produced L-lysine was larger than or equivalent to that produced by the parent strain after 72 hours of cultivation,

however, the amount of produced L-lysine was smaller than that produced by the parent strain after 40 hours of cultivation. Namely, the L-lysine-producing speed was lowered in cultivation for a short period.

Similarly, when mutant lysC and dapA, or dapA and dapB were enhanced in combination, the amount of produced L-lysine was larger than that produced by the parent strain after 72 hours of cultivation, however, the amount of produced L-lysine was smaller than that produced by the parent strain after 40 hours of cultivation. Thus the L-lysine-producing speed was lowered.

On the other hand, when lysA or ddh was enhanced singly, or when lysA and ddh were enhanced in combination, the amount of produced L-lysine was larger than that produced by the parent strain after 40 hours of cultivation, however, the amount of produced L-lysine was consequently smaller than that produced by the parent strain after the long period of cultivation because of decrease in growth.

On the contrary, in the case of the strain in which dapB was enhanced together with mutant lysC, the growth was improved, the L-lysine-producing speed was successfully restored in the short period of cultivation, and the accumulated amount of L-lysine was also improved in the long period of cultivation. In the case of the strain in which three of mutant lysC, dapA,

and dapB were simultaneously enhanced, the L-lysine productivity was further improved. Both of the L-lysine-producing speed and the amount of accumulated L-lysine were improved in a stepwise manner by successively enhancing lysA and ddh.

Industrial Applicability

According to the present invention, the L-lysine-producing ability of coryneform bacteria can be improved, and the growth speed can be also improved.

The L-lysine-producing speed can be improved, and the productivity can be also improved in coryneform L-lysine-producing bacteria by enhancing dapB together with mutant lysC. The L-lysine-producing speed and the productivity can be further improved by successively enhancing dapA, lysA, and ddh in addition to the aforementioned genes.

SEQUENCE LISTING

(1) GENERAL INFORMATION:
(i) APPLICANT: AJINOMOTO CO., INC.
(ii) TITLE OF INVENTION: METHOD OF PRODUCING L-LYSINE
(iii) NUMBER OF SEQUENCES: 24
(iv) CORRESPONDENCE ADDRESS:
(A) ADDRESSEE:
(B) STREET:
(C) CITY:
(E) COUNTRY:
(F) ZIP:
(v) COMPUTER READABLE FORM:
(A) MEDIUM TYPE: Floppy disk
(B) COMPUTER: IBM PC compatible
(C) OPERATING SYSTEM: PC-DOS/MS-DOS
(D) SOFTWARE: PatentIn Release #1.0, Version #1.30
(vi) CURRENT APPLICATION DATA:
(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION:
(vii) PRIOR APPLICATION DATA:
(A) APPLICATION NUMBER: JP 7-140614
(B) FILING DATE: 07-JUL-1995
(viii) ATTORNEY/AGENT INFORMATION:
(A) NAME:
(B) REGISTRATION NUMBER:
(ix) TELECOMMUNICATION INFORMATION:
(A) TELEPHONE:
(B) TELEFAX:

(2) INFORMATION FOR SEQ ID NO:1:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: NO
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:
TCGCGAAGTA GCACCTGTCA CTT 23

(2) INFORMATION FOR SEQ ID NO:2:
(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 21 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other..synthetic DNA
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: YES
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
ACGGAATTCA ATCTTACGGC C 21

(2) INFORMATION FOR SEQ ID NO:3:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1643 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)
(iv) ANTI-SENSE: NO
(vi) ORIGINAL SOURCE:
(A) ORGANISM: Brevibacterium lactofermentum
(B) STRAIN: ATCC 13869
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

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GTAACTGTCA GCACGTAGAT CGAAAGGTGC ACAAAGGTGG CCCTGGTCGT ACAGAAATAT 240
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ACCAAGAAGG CTGGAAATGA TGTCGTGGTT GTCTGCTCCG CAATGGGAGA CACCACGGAT 360
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GTGTGTGAGA TTTACTCGGA CGTTGACGGT GTGTATACCG CTGACCCGCG CATCGTTCCT 780
AATGCACAGA AGCTGGAAAA GCTCAGCTTC GAAGAAATGC TGGAACTTGC TGCTGTTGGC 840
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GCAGAAATCA ACATTGACAT GGTTCTGCAG AACGTCTCCT CTGTGGAAGA CGGCACCACC 1140
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(2) INFORMATION FOR SEQ ID NO:4:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1643 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Brevibacterium lactofermentum
(B) STRAIN: ATCC 13869

(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 217..1482

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

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                                         Met Ala Leu Val Val Gln
                                          1               5
AAA TAT GGC GGT TCC TCG CTT GAG AGT GCG GAA CGC ATT AGA AAC GTC       282
Lys Tyr Gly Gly Ser Ser Leu Glu Ser Ala Glu Arg Ile Arg Asn Val
            10                  15                  20
GCT GAA CGG ATC GTT GCC ACC AAG AAG GCT GGA AAT GAT GTC GTG GTT       330
Ala Glu Arg Ile Val Ala Thr Lys Lys Ala Gly Asn Asp Val Val Val
        25                  30                  35
GTC TGC TCC GCA ATG GGA GAC ACC ACG GAT GAA CTT CTA GAA CTT GCA       378
Val Cys Ser Ala Met Gly Asp Thr Thr Asp Glu Leu Leu Glu Leu Ala
    40                  45                  50
GCG GCA GTG AAT CCC GTT CCG CCA GCT CGT GAA ATG GAT ATG CTC CTG       426
Ala Ala Val Asn Pro Val Pro Pro Ala Arg Glu Met Asp Met Leu Leu
55                  60                  65                  70
ACT GCT GGT GAG CGT ATT TCT AAC GCT CTC GTC GCC ATG GCT ATT GAG       474
Thr Ala Gly Glu Arg Ile Ser Asn Ala Leu Val Ala Met Ala Ile Glu
                75                  80                  85
TCC CTT GGC GCA GAA GCT CAA TCT TTC ACT GGC TCT CAG GCT GGT GTG       522
Ser Leu Gly Ala Glu Ala Gln Ser Phe Thr Gly Ser Gln Ala Gly Val
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Glu Glu Met Leu Glu Leu Ala Ala Val Gly Ser Lys Ile Leu Val Leu
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(2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 421 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

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65                  70                  75                  80
Val Ala Met Ala Ile Glu Ser Leu Gly Ala Glu Ala Gln Ser Phe Thr
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Gly Ser Gln Ala Gly Val Leu Thr Thr Glu Arg His Gly Asn Ala Arg
            100                 105                 110
Ile Val Asp Val Thr Pro Gly Arg Val Arg Glu Ala Leu Asp Glu Gly
        115                 120                 125
Lys Ile Cys Ile Val Ala Gly Phe Gln Gly Val Asn Lys Glu Thr Arg
    130                 135                 140
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145 150 155 160
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Ser Lys Ile Leu Val Leu Arg Ser Val Glu Tyr Ala Arg Ala Phe Asn
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Asp Gly Thr Thr Asp Ile Thr Phe Thr Cys Pro Arg Ala Asp Gly Arg
305 310 315 320
Arg Ala Met Glu Ile Leu Lys Lys Leu Gln Val Gln Gly Asn Trp Thr
325 330 335
Asn Val Leu Tyr Asp Asp Gln Val Gly Lys Val Ser Leu Val Gly Ala
340 345 350
Gly Met Lys Ser His Pro Gly Val Thr Ala Glu Phe Met Glu Ala Leu
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(2) INFORMATION FOR SEQ ID NO:6:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1643 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)
(iv) ANTI-SENSE: NO
(vi) ORIGINAL SOURCE:

(A) ORGANISM: Brevibacterium lactofermentum
(B) STRAIN: ATCC 13869
(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 964..1482
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

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GTGTGTGAGA TTTACTCGGA CGTTGACGGT GTGTATACCG CTGACCCGCG CATCGTTCCT     780
AATGCACAGA AGCTGGAAAA GCTCAGCTTC GAAGAAATGC TGGAACTTGC TGCTGTTGGC     840
TCCAAGATTT TGGTGCTGCG CAGTGTTGAA TACGCTCGTG CATTCAATGT GCCACTTCGC     900
GTACGCTCGT CTTATAGTAA TGATCCCGGC ACTTTGATTG CCGGCTCTAT GGAGGATATT     960
CCT  GTG GAA GAA GCA GTC CTT ACC GGT GTC GCA ACC GAC AAG TCC GAA      1008
     Met Glu Glu Ala Val Leu Thr Gly Val Ala Thr Asp Lys Ser Glu
       1               5                  10                  15
GCC AAA GTA ACC GTT CTG GGT ATT TCC GAT AAG CCA GGC GAG GCT GCC      1056
Ala Lys Val Thr Val Leu Gly Ile Ser Asp Lys Pro Gly Glu Ala Ala
                   20                  25                  30
AAG GTT TTC CGT GCG TTG GCT GAT GCA GAA ATC AAC ATT GAC ATG GTT      1104
Lys Val Phe Arg Ala Leu Ala Asp Ala Glu Ile Asn Ile Asp Met Val
               35                  40                  45
CTG CAG AAC GTC TCC TCT GTG GAA GAC GGC ACC ACC GAC ATC ACG TTC      1152
Leu Gln Asn Val Ser Ser Val Glu Asp Gly Thr Thr Asp Ile Thr Phe
           50                  55                  60
ACC TGC CCT CGC GCT GAC GGA CGC CGT GCG ATG GAG ATC TTG AAG AAG      1200
Thr Cys Pro Arg Ala Asp Gly Arg Arg Ala Met Glu Ile Leu Lys Lys
       65                  70                  75
CTT CAG GTT CAG GGC AAC TGG ACC AAT GTG CTT TAC GAC GAC CAG GTC      1248
Leu Gln Val Gln Gly Asn Trp Thr Asn Val Leu Tyr Asp Asp Gln Val
   80                  85                  90                  95
GGC AAA GTC TCC CTC GTG GGT GCT GGC ATG AAG TCT CAC CCA GGT GTT      1296
Gly Lys Val Ser Leu Val Gly Ala Gly Met Lys Ser His Pro Gly Val
               100                 105                 110
ACC GCA GAG TTC ATG GAA GCT CTG CGC GAT GTC AAC GTG AAC ATC GAA      1344
Thr Ala Glu Phe Met Glu Ala Leu Arg Asp Val Asn Val Asn Ile Glu
```

```
                115                 120                 125
TTG ATT TCC ACC TCT GAG ATC CGC ATT TCC GTG CTG ATC CGT GAA GAT      1392
Leu Ile Ser Thr Ser Glu Ile Arg Ile Ser Val Leu Ile Arg Glu Asp
        130                 135                 140
GAT CTG GAT GCT GCT GCA CGT GCA TTG CAT GAG CAG TTC CAG CTG GGC      1440
Asp Leu Asp Ala Ala Ala Arg Ala Leu His Glu Gln Phe Gln Leu Gly
    145                 150                 155
GGC GAA GAC GAA GCC GTC GTT TAT GCA GGC ACC GGA CGC TAAAGTTTTAA      1490
Gly Glu Asp Glu Ala Val Val Tyr Ala Gly Thr Gly Arg
160                 165                 170
AGGAGTAGTT TTACAATGAC CACCATCGCA GTTGTTGGTG CAACCGGCCA GGTCGGCCAG    1550
GTTATGCGCA CCCTTTTGGA AGAGCGCAAT TTCCCAGCTG ACACTGTTCG TTTCTTTGCT    1610
TCCCCGCGTT CCGCAGGCCG TAAGATTGAA TTC                                 1643
```

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 172 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

```
Met Glu Glu Ala Val Leu Thr Gly Val Ala Thr Asp Lys Ser Glu Ala
  1               5                  10                  15
Lys Val Thr Val Leu Gly Ile Ser Asp Lys Pro Gly Glu Ala Ala Lys
            20                  25                  30
Val Phe Arg Ala Leu Ala Asp Ala Glu Ile Asn Ile Asp Met Val Leu
        35                  40                  45
Gln Asn Val Ser Ser Val Glu Asp Gly Thr Thr Asp Ile Thr Phe Thr
    50                  55                  60
Cys Pro Arg Ala Asp Gly Arg Arg Ala Met Glu Ile Leu Lys Lys Leu
65                  70                  75                  80
Gln Val Gln Gly Asn Trp Thr Asn Val Leu Tyr Asp Asp Gln Val Gly
                85                  90                  95
Lys Val Ser Leu Val Gly Ala Gly Met Lys Ser His Pro Gly Val Thr
            100                 105                 110
Ala Glu Phe Met Glu Ala Leu Arg Asp Val Asn Val Asn Ile Glu Leu
        115                 120                 125
Ile Ser Thr Ser Glu Ile Arg Ile Ser Val Leu Ile Arg Glu Asp Asp
    130                 135                 140
Leu Asp Ala Ala Ala Arg Ala Leu His Glu Gln Phe Gln Leu Gly Gly
145                 150                 155                 160
Glu Asp Glu Ala Val Val Tyr Ala Gly Thr Gly Arg
                165                 170
```

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: NO
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:
GGATCCCCAA TCGATACCTG GAA 23

(2) INFORMATION FOR SEQ ID NO:9:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: YES
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
CGGTTCATCG CCAAGTTTTT CTT 23

(2) INFORMATION FOR SEQ ID NO:10:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 2001 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)
(iv) ANTI-SENSE: NO
(vi) ORIGINAL SOURCE:
(A) ORGANISM: Brevibacterium lactofermentum
(B) STRAIN: ATCC 13869
(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 730..1473
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

```
GGATCCCCAA TCGATACCTG GAACGACAAC CTGATCAGGA TATCCAATGC CTTGAATATT      60
GACGTTGAGG AAGGAATCAC CAGCCATCTC AACTGGAAGA CCTGACGCCT GCTGAATTGG     120
ATCAGTGGCC CAATCGACCC ACCAACCAGG TTGGCTATTA CCGGCGATAT CAAAAACAAC     180
TCGCGTGAAC GTTTCGTGCT CGGCAACGCG GATGCCAGCG ATCGACATAT CGGAGTCACC     240
AACTTGAGCC TGCTGCTTCT GATCCATCGA CGGGGAACCC AACGGCGGCA AAGCAGTGGG     300
GGAAGGGGAG TTGGTGGACT CTGAATCAGT GGGCTCTGAA GTGGTAGGCG ACGGGGCAGC     360
ATCTGAAGGC GTGCGAGTTG TGGTGACCGG GTTAGCGGTT TCAGTTTCTG TCACAACTGG     420
```

```
AGCAGGACTA GCAGAGGTTG TAGGCGTTGA GCCGCTTCCA TCACAAGCAC TTAAAAGTAA      480
AGAGGCGGAA ACCACAAGCG CCAAGGAACT ACCTGCGGAA CGGGCGGTGA AGGGCAACTT      540
AAGTCTCATA TTTCAAACAT AGTTCCACCT GTGTGATTAA TCTCCAGAAC GGAACAAACT      600
GATGAACAAT CGTTAACAAC ACAGACCAAA ACGGTCAGTT AGGTATGGAT ATCAGCACCT      660
TCTGAATGGG TACGTCTAGA CTGGTGGGCG TTTGAAAAAC TCTTCGCCCC ACGAAAATGA      720
AGGAGCATA ATG GGA ATC AAG GTT GGC GTT CTC GGA GCC AAA GGC CGT            768
                Met Gly Ile Lys Val Gly Val Leu Gly Ala Lys Gly Arg
                  1               5                  10
GTT GGT CAA ACT ATT GTG GCA GCA GTC AAT GAG TCC GAC GAT CTG GAG         816
Val Gly Gln Thr Ile Val Ala Ala Val Asn Glu Ser Asp Asp Leu Glu
     15                  20                  25
CTT GTT GCA GAG ATC GGC GTC GAC GAT GAT TTG AGC CTT CTG GTA GAC         864
Leu Val Ala Glu Ile Gly Val Asp Asp Asp Leu Ser Leu Leu Val Asp
 30                  35                  40                  45
AAC GGC GCT GAA GTT GTC GTT GAC TTC ACC ACT CCT AAC GCT GTG ATG         912
Asn Gly Ala Glu Val Val Val Asp Phe Thr Thr Pro Asn Ala Val Met
                 50                  55                  60
GGC AAC CTG GAG TTC TGC ATC AAC AAC GGC ATT TCT GCG GTT GTT GGA         960
Gly Asn Leu Glu Phe Cys Ile Asn Asn Gly Ile Ser Ala Val Val Gly
               65                  70                  75
ACC ACG GGC TTC GAT GAT GCT CGT TTG GAG CAG GTT CGC GCC TGG CTT        1008
Thr Thr Gly Phe Asp Asp Ala Arg Leu Glu Gln Val Arg Ala Trp Leu
             80                  85                  90
GAA GGA AAA GAC AAT GTC GGT GTT CTG ATC GCA CCT AAC TTT GCT ATC        1056
Glu Gly Lys Asp Asn Val Gly Val Leu Ile Ala Pro Asn Phe Ala Ile
     95                 100                 105
TCT GCG GTG TTG ACC ATG GTC TTT TCC AAG CAG GCT GCC CGC TTC TTC        1104
Ser Ala Val Leu Thr Met Val Phe Ser Lys Gln Ala Ala Arg Phe Phe
110                 115                 120                 125
GAA TCA GCT GAA GTT ATT GAG CTG CAC CAC CCC AAC AAG CTG GAT GCA        1152
Glu Ser Ala Glu Val Ile Glu Leu His His Pro Asn Lys Leu Asp Ala
                130                 135                 140
CCT TCA GGC ACC GCG ATC CAC ACT GCT CAG GGC ATT GCT GCG GCA CGC        1200
Pro Ser Gly Thr Ala Ile His Thr Ala Gln Gly Ile Ala Ala Ala Arg
              145                 150                 155
AAA GAA GCA GGC ATG GAC GCA CAG CCA GAT GCG ACC GAG CAG GCA CTT        1248
Lys Glu Ala Gly Met Asp Ala Gln Pro Asp Ala Thr Glu Gln Ala Leu
            160                 165                 170
GAG GGT TCC CGT GGC GCA AGC GTA GAT GGA ATC CCA GTT CAC GCA GTC        1296
Glu Gly Ser Arg Gly Ala Ser Val Asp Gly Ile Pro Val His Ala Val
     175                 180                 185
CGC ATG TCC GGC ATG GTT GCT CAC GAG CAA GTT ATC TTT GGC ACC CAG        1344
Arg Met Ser Gly Met Val Ala His Glu Gln Val Ile Phe Gly Thr Gln
190                 195                 200                 205
GGT CAG ACC TTG ACC ATC AAG CAG GAC TCC TAT GAT CGC AAC TCA TTT        1392
```

```
Gly Gln Thr Leu Thr Ile Lys Gln Asp Ser Tyr Asp Arg Asn Ser Phe
                210                 215                 220
GCA CCA GGT GTC TTG GTG GGT GTG CGC AAC ATT GCA CAG CAC CCA GGC      1440
Ala Pro Gly Val Leu Val Gly Val Arg Asn Ile Ala Gln His Pro Gly
            225                 230                 235
CTA GTC GTA GGA CTT GAG CAT TAC CTA GGC CTG TAAAGGCTCA TTTCAGCAGC    1493
Leu Val Val Gly Leu Glu His Tyr Leu Gly Leu
        240                 245
GGGTGGAATT TTTTAAAAGG AGCGTTTAAA GGCTGTGGCC GAACAAGTTA AATTGAGCGT    1553
GGAGTTGATA GCGTGCAGTT CTTTTACTCC ACCCGCTGAT GTTGAGTGGT CAACTGATGT    1613
TGAGGGCGCG GAAGCACTCG TCGAGTTTGC GGGTCGTGCC TGCTACGAAA CTTTTGATAA    1673
GCCGAACCCT CGAACTGCTT CCAATGCTGC GTATCTGCGC CACATCATGG AAGTGGGGCA    1733
CACTGCTTTG CTTGAGCATG CCAATGCCAC GATGTATATC CGAGGCATTT CTCGGTCCGC    1793
GACCCATGAA TTGGTCCGAC ACCGCCATTT TTCCTTCTCT CAACTGTCTC AGCGTTTCGT    1853
GCACAGCGGA GAATCGGAAG TAGTGGTGCC CACTCTCATC GATGAAGATC CGCAGTTGCG    1913
TGAACTTTTC ATGCACGCCA TGGATGAGTC TCGGTTCGCT TTCAATGAGC TGCTTAATGC    1973
GCTGGAAGAA AAACTTGGCG ATGAACCG                                       2001
```

(2) INFORMATION FOR SEQ ID NO:11:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 248 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

```
Met Gly Ile Lys Val Gly Val Leu Gly Ala Lys Gly Arg Val Gly Gln
 1               5                  10                  15
Thr Ile Val Ala Ala Val Asn Glu Ser Asp Asp Leu Glu Leu Val Ala
            20                  25                  30
Glu Ile Gly Val Asp Asp Asp Leu Ser Leu Leu Val Asp Asn Gly Ala
        35                  40                  45
Glu Val Val Val Asp Phe Thr Thr Pro Asn Ala Val Met Gly Asn Leu
    50                  55                  60
Glu Phe Cys Ile Asn Asn Gly Ile Ser Ala Val Val Gly Thr Thr Gly
65                  70                  75                  80
Phe Asp Asp Ala Arg Leu Glu Gln Val Arg Ala Trp Leu Glu Gly Lys
                85                  90                  95
Asp Asn Val Gly Val Leu Ile Ala Pro Asn Phe Ala Ile Ser Ala Val
            100                 105                 110
Leu Thr Met Val Phe Ser Lys Gln Ala Ala Arg Phe Phe Glu Ser Ala
        115                 120                 125
Glu Val Ile Glu Leu His His Pro Asn Lys Leu Asp Ala Pro Ser Gly
    130                 135                 140
Thr Ala Ile His Thr Ala Gln Gly Ile Ala Ala Ala Arg Lys Glu Ala
145                 150                 155                 160
```

Gly Met Asp Ala Gln Pro Asp Ala Thr Glu Gln Ala Leu Glu Gly Ser
165 170 175
Arg Gly Ala Ser Val Asp Gly Ile Pro Val His Ala Val Arg Met Ser
180 185 190
Gly Met Val Ala His Glu Gln Val Ile Phe Gly Thr Gln Gly Gln Thr
195 200 205
Leu Thr Ile Lys Gln Asp Ser Tyr Asp Arg Asn Ser Phe Ala Pro Gly
210 215 220
Val Leu Val Gly Val Arg Asn Ile Ala Gln His Pro Gly Leu Val Val
225 230 235 240
Gly Leu Glu His Tyr Leu Gly Leu
245

(2) INFORMATION FOR SEQ ID NO:12:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: NO
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:
GTCGACGGAT CGCAAATGGC AAC 23

(2) INFORMATION FOR SEQ ID NO:13:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 23 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: YES
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:
GGATCCTTGA GCACCTTGCG CAG 23

(2) INFORMATION FOR SEQ ID NO:14:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1411 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)
(iv) ANTI-SENSE: NO

(vi) ORIGINAL SOURCE:
(A) ORGANISM: Brevibacterium lactofermentum
(B) STRAIN: ATCC 13869
(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 311..1213
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

```
CTCTCGATAT CGAGAGAGAA GCAGCGCCAC GGTTTTTCGG TGATTTTGAG ATTGAAACTT      60
TGGCAGACGG ATCGCAAATG GCAACAAGCC CGTATGTCAT GGACTTTTAA CGCAAAGCTC     120
ACACCCACGA GCTAAAAATT CATATAGTTA AGACAACATT TTTGGCTGTA AAAGACAGCC     180
GTAAAAACCT CTTGCTCATG TCAATTGTTC TTATCGGAAT GTGGCTTGGG CGATTGTTAT     240
GCAAAAGTTG TTAGGTTTTT TGCGGGGTTG TTTAACCCCC AAATGAGGGA AGAAGGTAAC     300
CTTGAACTCT ATG AGC ACA GGT TTA ACA GCT AAG ACC GGA GTA GAG CAC         349
          Met Ser Thr Gly Leu Thr Ala Lys Thr Gly Val Glu His
            1               5                  10
TTC GGC ACC GTT GGA GTA GCA ATG GTT ACT CCA TTC ACG GAA TCC GGA       397
Phe Gly Thr Val Gly Val Ala Met Val Thr Pro Phe Thr Glu Ser Gly
     15                  20                  25
GAC ATC GAT ATC GCT GCT GGC CGC GAA GTC GCG GCT TAT TTG GTT GAT       445
Asp Ile Asp Ile Ala Ala Gly Arg Glu Val Ala Ala Tyr Leu Val Asp
 30                  35                  40                  45
AAG GGC TTG GAT TCT TTG GTT CTC GCG GGC ACC ACT GGT GAA TCC CCA       493
Lys Gly Leu Asp Ser Leu Val Leu Ala Gly Thr Thr Gly Glu Ser Pro
                 50                  55                  60
ACG ACA ACC GCC GCT GAA AAA CTA GAA CTG CTC AAG GCC GTT CGT GAG       541
Thr Thr Thr Ala Ala Glu Lys Leu Glu Leu Leu Lys Ala Val Arg Glu
            65                  70                  75
GAA GTT GGG GAT CGG GCG AAC GTC ATC GCC GGT GTC GGA ACC AAC AAC       589
Glu Val Gly Asp Arg Ala Asn Val Ile Ala Gly Val Gly Thr Asn Asn
        80                  85                  90
ACG CGG ACA TCT GTG GAA CTT GCG GAA GCT GCT GCT TCT GCT GGC GCA       637
Thr Arg Thr Ser Val Glu Leu Ala Glu Ala Ala Ala Ser Ala Gly Ala
     95                  100                 105
GAC GGC CTT TTA GTT GTA ACT CCT TAT TAC TCC AAG CCG AGC CAA GAG       685
Asp Gly Leu Leu Val Val Thr Pro Tyr Tyr Ser Lys Pro Ser Gln Glu
110                 115                 120                 125
GGA TTG CTG GCG CAC TTC GGT GCA ATT GCT GCA GCA ACA GAG GTT CCA       733
Gly Leu Leu Ala His Phe Gly Ala Ile Ala Ala Ala Thr Glu Val Pro
                130                 135                 140
ATT TGT CTC TAT GAC ATT CCT GGT CGG TCA GGT ATT CCA ATT GAG TCT       781
Ile Cys Leu Tyr Asp Ile Pro Gly Arg Ser Gly Ile Pro Ile Glu Ser
            145                 150                 155
GAT ACC ATG AGA CGC CTG AGT GAA TTA CCT ACG ATT TTG GCG GTC AAG       829
Asp Thr Met Arg Arg Leu Ser Glu Leu Pro Thr Ile Leu Ala Val Lys
        160                 165                 170
```

```
GAC GCC AAG GGT GAC CTC GTT GCA GCC ACG TCA TTG ATC AAA GAA ACG        877
Asp Ala Lys Gly Asp Leu Val Ala Ala Thr Ser Leu Ile Lys Glu Thr
    175                 180                 185
GGA CTT GCC TGG TAT TCA GGC GAT GAC CCA CTA AAC CTT GTT TGG CTT        925
Gly Leu Ala Trp Tyr Ser Gly Asp Asp Pro Leu Asn Leu Val Trp Leu
190                 195                 200                 205
GCT TTG GGC GGA TCA GGT TTC ATT TCC GTA ATT GGA CAT GCA GCC CCC        973
Ala Leu Gly Gly Ser Gly Phe Ile Ser Val Ile Gly His Ala Ala Pro
                210                 215                 220
ACA GCA TTA CGT GAG TTG TAC ACA AGC TTC GAG GAA GGC GAC CTC GTC       1021
Thr Ala Leu Arg Glu Leu Tyr Thr Ser Phe Glu Glu Gly Asp Leu Val
            225                 230                 235
CGT GCG CGG GAA ATC AAC GCC AAA CTA TCA CCG CTG GTA GCT GCC CAA       1069
Arg Ala Arg Glu Ile Asn Ala Lys Leu Ser Pro Leu Val Ala Ala Gln
        240                 245                 250
GGT CGC TTG GGT GGA GTC AGC TTG GCA AAA GCT GCT CTG CGT CTG CAG       1117
Gly Arg Leu Gly Gly Val Ser Leu Ala Lys Ala Ala Leu Arg Leu Gln
    255                 260                 265
GGC ATC AAC GTA GGA GAT CCT CGA CTT CCA ATT ATG GCT CCA AAT GAG       1165
Gly Ile Asn Val Gly Asp Pro Arg Leu Pro Ile Met Ala Pro Asn Glu
270                 275                 280                 285
CAG GAA CTT GAG GCT CTC CGA GAA GAC ATG AAA AAA GCT GGA GTT CTA       1213
Gln Glu Leu Glu Ala Leu Arg Glu Asp Met Lys Lys Ala Gly Val Leu
                290                 295                 300
TAAATATGAA TGATTCCCGA AATCGCGGCC GGAAGGTTAC CCGCAAGGCG GCCCACCAGA     1273
AGCTGGTCAG GAAAACCATC TGGATACCCC TGTCTTTCAG GCACCAGATG CTTCCTCTAA     1333
CCAGAGCGCT GTAAAAGCTG AGACCGCCGG AAACGACAAT CGGGATGCTG CGCAAGGTGC     1393
TCAAGGATCC CAACATTC                                                   1411
```

(2) INFORMATION FOR SEQ ID NO:15:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 301 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

```
Met Ser Thr Gly Leu Thr Ala Lys Thr Gly Val Glu His Phe Gly Thr
  1               5                  10                  15
Val Gly Val Ala Met Val Thr Pro Phe Thr Glu Ser Gly Asp Ile Asp
            20                  25                  30
Ile Ala Ala Gly Arg Glu Val Ala Ala Tyr Leu Val Asp Lys Gly Leu
        35                  40                  45
Asp Ser Leu Val Leu Ala Gly Thr Thr Gly Glu Ser Pro Thr Thr Thr
    50                  55                  60
Ala Ala Glu Lys Leu Glu Leu Leu Lys Ala Val Arg Glu Glu Val Gly
```

65 70 75 80
Asp Arg Ala Asn Val Ile Ala Gly Val Gly Thr Asn Asn Thr Arg Thr
85 90 95
Ser Val Glu Leu Ala Glu Ala Ala Ala Ser Ala Gly Ala Asp Gly Leu
100 105 110
Leu Val Val Thr Pro Tyr Tyr Ser Lys Pro Ser Gln Glu Gly Leu Leu
115 120 125
Ala His Phe Gly Ala Ile Ala Ala Ala Thr Glu Val Pro Ile Cys Leu
130 135 140
Tyr Asp Ile Pro Gly Arg Ser Gly Ile Pro Ile Glu Ser Asp Thr Met
145 150 155 160
Arg Arg Leu Ser Glu Leu Pro Thr Ile Leu Ala Val Lys Asp Ala Lys
165 170 175
Gly Asp Leu Val Ala Ala Thr Ser Leu Ile Lys Glu Thr Gly Leu Ala
180 185 190
Trp Tyr Ser Gly Asp Asp Pro Leu Asn Leu Val Trp Leu Ala Leu Gly
195 200 205
Gly Ser Gly Phe Ile Ser Val Ile Gly His Ala Ala Pro Thr Ala Leu
210 215 220
Arg Glu Leu Tyr Thr Ser Phe Glu Glu Gly Asp Leu Val Arg Ala Arg
225 230 235 240
Glu Ile Asn Ala Lys Leu Ser Pro Leu Val Ala Ala Gln Gly Arg Leu
245 250 255
Gly Gly Val Ser Leu Ala Lys Ala Ala Leu Arg Leu Gln Gly Ile Asn
260 265 270
Val Gly Asp Pro Arg Leu Pro Ile Met Ala Pro Asn Glu Gln Glu Leu
275 280 285
Glu Ala Leu Arg Glu Asp Met Lys Lys Ala Gly Val Leu
290 295 300

(2) INFORMATION FOR SEQ ID NO:16:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 bases

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(A) DESCRIPTION: /desc = "synthetic DNA"

(iv) ANTI-SENSE: NO

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

GTGGAGCCGA CCATTCCGCG AGG 23

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 23 bases

(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: YES
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

CCAAAACCGC CCTCCACGGC GAA 23

(2) INFORMATION FOR SEQ ID NO:18:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 3579 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)
(iv) ANTI-SENSE: NO
(vi) ORIGINAL SOURCE:
(A) ORGANISM: Brevibacterium lactofermentum
(B) STRAIN: ATCC 13869
(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 533..2182
(ix) FEATURE:
(A) NAME/KEY: CDS
(B) LOCATION: 2188..3522
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GTGGAGCCGA CCATTCCGCG AGGCTGCACT GCAACGAGGT CGTAGTTTTG GTACATGGCT 60
TCTGGCCAGT TCATGGATTG GCTGCCGAAG AAGCTATAGG CATCGCACCA GGGCCACCGA 120
GTTACCGAAG ATGGTGCCGT GCTTTTCGCC TTGGGCAGGG ACCTTGACAA AGCCCACGCT 180
GATATCGCCA AGTGAGGGAT CAGAATAGTG CATGGGCACG TCGATGCTGC CACATTGAGC 240
GGAGGCAATA TCTACCTGAG GTGGGCATTC TTCCCAGCGG ATGTTTTCTT GCGCTGCTGC 300
AGTGGGCATT GATACCAAAA AGGGGCTAAG CGCAGTCGAG GCGGCAAGAA CTGCTACTAC 360
CCTTTTTATT GTCGAACGGG GCATTACGGC TCCAAGGACG TTTGTTTTCT GGGTCAGTTA 420
CCCCAAAAAG CATATACAGA GACCAATGAT TTTTCATTAA AAAGGCAGGG ATTTGTTATA 480
AGTATGGGTC GTATTCTGTG CGACGGGTGT ACCTCGGCTA GAATTTCTCC CC ATG 535

Met
1

ACA CCA GCT GAT CTC GCA ACA TTG ATT AAA GAG ACC GCG GTA GAG GTT 583
Thr Pro Ala Asp Leu Ala Thr Leu Ile Lys Glu Thr Ala Val Glu Val
5 10 15
TTG ACC TCC CGC GAG CTC GAT ACT TCT GTT CTT CCG GAG CAG GTA GTT 631
Leu Thr Ser Arg Glu Leu Asp Thr Ser Val Leu Pro Glu Gln Val Val
20 25 30
GTG GAG CGT CCG CGT AAC CCA GAG CAC GGC GAT TAC GCC ACC AAC ATT 679

```
Val Glu Arg Pro Arg Asn Pro Glu His Gly Asp Tyr Ala Thr Asn Ile
     35                  40                  45
GCA TTG CAG GTG GCT AAA AAG GTC GGT CAG AAC CCT CGG GAT TTG GCT      727
Ala Leu Gln Val Ala Lys Lys Val Gly Gln Asn Pro Arg Asp Leu Ala
 50                  55                  60                  65
ACC TGG CTG GCA GAG GCA TTG GCT GCA GAT GAC GCC ATT GAT TCT GCT      775
Thr Trp Leu Ala Glu Ala Leu Ala Ala Asp Asp Ala Ile Asp Ser Ala
                 70                  75                  80
GAA ATT GCT GGC CCA GGC TTT TTG AAC ATT CGC CTT GCT GCA GCA GCA      823
Glu Ile Ala Gly Pro Gly Phe Leu Asn Ile Arg Leu Ala Ala Ala Ala
             85                  90                  95
CAG GGT GAA ATT GTG GCC AAG ATT CTG GCA CAG GGC GAG ACT TTC GGA      871
Gln Gly Glu Ile Val Ala Lys Ile Leu Ala Gln Gly Glu Thr Phe Gly
        100                 105                 110
AAC TCC GAT CAC CTT TCC CAC TTG GAC GTG AAC CTC GAG TTC GTT TCT      919
Asn Ser Asp His Leu Ser His Leu Asp Val Asn Leu Glu Phe Val Ser
    115                 120                 125
GCA AAC CCA ACC GGA CCT ATT CAC CTT GGC GGA ACC CGC TGG GCT GCC      967
Ala Asn Pro Thr Gly Pro Ile His Leu Gly Gly Thr Arg Trp Ala Ala
130                 135                 140                 145
GTG GGT GAC TCT TTG GGT CGT GTG CTG GAG GCT TCC GGC GCG AAA GTG     1015
Val Gly Asp Ser Leu Gly Arg Val Leu Glu Ala Ser Gly Ala Lys Val
                150                 155                 160
ACC CGC GAA TAC TAC TTC AAC GAT CAC GGT CGC CAG ATC GAT CGT TTC     1063
Thr Arg Glu Tyr Tyr Phe Asn Asp His Gly Arg Gln Ile Asp Arg Phe
            165                 170                 175
GCT TTG TCC CTT CTT GCA GCG GCG AAG GGC GAG CCA ACG CCA GAA GAC     1111
Ala Leu Ser Leu Leu Ala Ala Ala Lys Gly Glu Pro Thr Pro Glu Asp
        180                 185                 190
GGT TAT GGC GGC GAA TAC ATT AAG GAA ATT GCG GAG GCA ATC GTC GAA     1159
Gly Tyr Gly Gly Glu Tyr Ile Lys Glu Ile Ala Glu Ala Ile Val Glu
    195                 200                 205
AAG CAT CCT GAA GCG TTG GCT TTG GAG CCT GCC GCA ACC CAG GAG CTT     1207
Lys His Pro Glu Ala Leu Ala Leu Glu Pro Ala Ala Thr Gln Glu Leu
210                 215                 220                 225
TTC CGC GCT GAA GGC GTG GAG ATG ATG TTC GAG CAC ATC AAA TCT TCC     1255
Phe Arg Ala Glu Gly Val Glu Met Met Phe Glu His Ile Lys Ser Ser
                230                 235                 240
CTG CAT GAG TTC GGC ACC GAT TTC GAT GTC TAC TAC CAC GAG AAC TCC     1303
Leu His Glu Phe Gly Thr Asp Phe Asp Val Tyr Tyr His Glu Asn Ser
            245                 250                 255
CTG TTC GAG TCC GGT GCG GTG GAC AAG GCC GTG CAG GTG CTG AAG GAC     1351
Leu Phe Glu Ser Gly Ala Val Asp Lys Ala Val Gln Val Leu Lys Asp
        260                 265                 270
AAC GGC AAC CTG TAC GAA AAC GAG GGC GCT TGG TGG CTG CGT TCC ACC     1399
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Asn Gly Asn Leu Tyr Glu Asn Glu Gly Ala Trp Trp Leu Arg Ser Thr
    275                 280                 285
GAA TTC GGC GAT GAC AAA GAC CGC GTG GTG ATC AAG TCT GAC GGC GAC      1447
Glu Phe Gly Asp Asp Lys Asp Arg Val Val Ile Lys Ser Asp Gly Asp
290                 295                 300                 305
GCA GCC TAC ATC GCT GGC GAT ATC GCG TAC GTG GCT GAT AAG TTC TCC      1495
Ala Ala Tyr Ile Ala Gly Asp Ile Ala Tyr Val Ala Asp Lys Phe Ser
                310                 315                 320
CGC GGA CAC AAC CTA AAC ATC TAC ATG TTG GGT GCT GAC CAC CAT GGT      1543
Arg Gly His Asn Leu Asn Ile Tyr Met Leu Gly Ala Asp His His Gly
            325                 330                 335
TAC ATC GCG CGC CTG AAG GCA GCG GCG GCG GCA CTT GGC TAC AAG CCA      1591
Tyr Ile Ala Arg Leu Lys Ala Ala Ala Ala Ala Leu Gly Tyr Lys Pro
        340                 345                 350
GAA GGC GTT GAA GTC CTG ATT GGC CAG ATG GTG AAC CTG CTT CGC GAC      1639
Glu Gly Val Glu Val Leu Ile Gly Gln Met Val Asn Leu Leu Arg Asp
    355                 360                 365
GGC AAG GCA GTG CGT ATG TCC AAG CGT GCA GGC ACC GTG GTC ACC CTA      1687
Gly Lys Ala Val Arg Met Ser Lys Arg Ala Gly Thr Val Val Thr Leu
370                 375                 380                 385
GAT GAC CTC GTT GAA GCA ATC GGC ATC GAT GCG GCG CGT TAC TCC CTG      1735
Asp Asp Leu Val Glu Ala Ile Gly Ile Asp Ala Ala Arg Tyr Ser Leu
                390                 395                 400
ATC CGT TCC TCC GTG GAT TCT TCC CTG GAT ATC GAT CTC GGC CTG TGG      1783
Ile Arg Ser Ser Val Asp Ser Ser Leu Asp Ile Asp Leu Gly Leu Trp
            405                 410                 415
GAA TCC CAG TCC TCC GAC AAC CCT GTG TAC TAC GTG CAG TAC GGA CAC      1831
Glu Ser Gln Ser Ser Asp Asn Pro Val Tyr Tyr Val Gln Tyr Gly His
        420                 425                 430
GCT CGT CTG TGC TCC ATC GCG CGC AAG GCA GAG ACC TTG GGT GTC ACC      1879
Ala Arg Leu Cys Ser Ile Ala Arg Lys Ala Glu Thr Leu Gly Val Thr
    435                 440                 445
GAG GAA GGC GCA GAC CTA TCT CTA CTG ACC CAC GAC CGC GAA GGC GAT      1927
Glu Glu Gly Ala Asp Leu Ser Leu Leu Thr His Asp Arg Glu Gly Asp
450                 455                 460                 465
CTC ATC CGC ACA CTC GGA GAG TTC CCA GCA GTG GTG AAG GCT GCC GCT      1975
Leu Ile Arg Thr Leu Gly Glu Phe Pro Ala Val Val Lys Ala Ala Ala
                470                 475                 480
GAC CTA CGT GAA CCA CAC CGC ATT GCC CGC TAT GCT GAG GAA TTA GCT      2023
Asp Leu Arg Glu Pro His Arg Ile Ala Arg Tyr Ala Glu Glu Leu Ala
            485                 490                 495
GGA ACT TTC CAC CGC TTC TAC GAT TCC TGC CAC ATC CTT CCA AAG GTT      2071
Gly Thr Phe His Arg Phe Tyr Asp Ser Cys His Ile Leu Pro Lys Val
        500                 505                 510
GAT GAG GAT ACG GCA CCA ATC CAC ACA GCA CGT CTG GCA CTT GCA GCA      2119
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Asp Glu Asp Thr Ala Pro Ile His Thr Ala Arg Leu Ala Leu Ala Ala
    515                 520                 525
GCA ACC CGC CAG ACC CTC GCT AAC GCC CTG CAC CTG GTT GGC GTT TCC      2167
Ala Thr Arg Gln Thr Leu Ala Asn Ala Leu His Leu Val Gly Val Ser
530                 535                 540                 545
GCA CCG GAG AAG ATG TAACA ATG GCT ACA GTT GAA AAT TTC AAT GAA      2214
Ala Pro Glu Lys Met       Met Ala Thr Val Glu Asn Phe Asn Glu
                550         1                   5
CTT CCC GCA CAC GTA TGG CCA CGC AAT GCC GTG CGC CAA GAA GAC GGC      2262
Leu Pro Ala His Val Trp Pro Arg Asn Ala Val Arg Gln Glu Asp Gly
 10                  15                  20                  25
GTT GTC ACC GTC GCT GGT GTG CCT CTG CCT GAC CTC GCT GAA GAA TAC      2310
Val Val Thr Val Ala Gly Val Pro Leu Pro Asp Leu Ala Glu Glu Tyr
                30                  35                  40
GGA ACC CCA CTG TTC GTA GTC GAC GAG GAC GAT TTC CGT TCC CGC TGT      2358
Gly Thr Pro Leu Phe Val Val Asp Glu Asp Asp Phe Arg Ser Arg Cys
            45                  50                  55
CGC GAC ATG GCT ACC GCA TTC GGT GGA CCA GGC AAT GTG CAC TAC GCA      2406
Arg Asp Met Ala Thr Ala Phe Gly Gly Pro Gly Asn Val His Tyr Ala
        60                  65                  70
TCT AAA GCG TTC CTG ACC AAG ACC ATT GCA CGT TGG GTT GAT GAA GAG      2454
Ser Lys Ala Phe Leu Thr Lys Thr Ile Ala Arg Trp Val Asp Glu Glu
    75                  80                  85
GGG CTG GCA CTG GAC ATT GCA TCC ATC AAC GAA CTG GGC ATT GCC CTG      2502
Gly Leu Ala Leu Asp Ile Ala Ser Ile Asn Glu Leu Gly Ile Ala Leu
 90                  95                  100                 105
GCC GCT GGT TTC CCC GCC AGC CGT ATC ACC GCG CAC GGC AAC AAC AAA      2550
Ala Ala Gly Phe Pro Ala Ser Arg Ile Thr Ala His Gly Asn Asn Lys
                110                 115                 120
GGC GTA GAG TTC CTG CGC GCG TTG GTT CAA AAC GGT GTG GGA CAC GTG      2598
Gly Val Glu Phe Leu Arg Ala Leu Val Gln Asn Gly Val Gly His Val
            125                 130                 135
GTG CTG GAC TCC GCA CAG GAA CTA GAA CTG TTG GAT TAC GTT GCC GCT      2646
Val Leu Asp Ser Ala Gln Glu Leu Glu Leu Leu Asp Tyr Val Ala Ala
        140                 145                 150
GGT GAA GGC AAG ATT CAG GAC GTG TTG ATC CGC GTA AAG CCA GGC ATC      2694
Gly Glu Gly Lys Ile Gln Asp Val Leu Ile Arg Val Lys Pro Gly Ile
    155                 160                 165
GAA GCA CAC ACC CAC GAG TTC ATC GCC ACT AGC CAC GAA GAC CAG AAG      2742
Glu Ala His Thr His Glu Phe Ile Ala Thr Ser His Glu Asp Gln Lys
170                 175                 180                 185
TTC GGA TTC TCC CTG GCA TCC GGT TCC GCA TTC GAA GCA GCA AAA GCC      2790
Phe Gly Phe Ser Leu Ala Ser Gly Ser Ala Phe Glu Ala Ala Lys Ala
                190                 195                 200
GCC AAC AAC GCA GAA AAC CTG AAC CTG GTT GGC CTG CAC TGC CAC GTT      2838
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Ala Asn Asn Ala Glu Asn Leu Asn Leu Val Gly Leu His Cys His Val
                205                 210                 215
GGT TCC CAG GTG TTC GAC GCC GAA GGC TTC AAG CTG GCA GCA GAA CGC      2886
Gly Ser Gln Val Phe Asp Ala Glu Gly Phe Lys Leu Ala Ala Glu Arg
        220                 225                 230
GTG TTG GGC CTG TAC TCA CAG ATC CAC AGC GAA CTG GGC GTT GCC CTT      2934
Val Leu Gly Leu Tyr Ser Gln Ile His Ser Glu Leu Gly Val Ala Leu
    235                 240                 245
CCT GAA CTG GAT CTC GGT GGC GGA TAC GGC ATT GCC TAT ACC GCA GCT      2982
Pro Glu Leu Asp Leu Gly Gly Gly Tyr Gly Ile Ala Tyr Thr Ala Ala
250                 255                 260                 265
GAA GAA CCA CTC AAC GTC GCA GAA GTT GCC TCC GAC CTG CTC ACC GCA      3030
Glu Glu Pro Leu Asn Val Ala Glu Val Ala Ser Asp Leu Leu Thr Ala
                270                 275                 280
GTC GGA AAA ATG GCA GCG GAA CTA GGC ATC GAC GCA CCA ACC GTG CTT      3078
Val Gly Lys Met Ala Ala Glu Leu Gly Ile Asp Ala Pro Thr Val Leu
            285                 290                 295
GTT GAG CCC GGC CGC GCT ATC GCA GGC CCC TCC ACC GTG ACC ATC TAC      3126
Val Glu Pro Gly Arg Ala Ile Ala Gly Pro Ser Thr Val Thr Ile Tyr
        300                 305                 310
GAA GTC GGC ACC ACC AAA GAC GTC CAC GTA GAC GAC GAC AAA ACC CGC      3174
Glu Val Gly Thr Thr Lys Asp Val His Val Asp Asp Asp Lys Thr Arg
    315                 320                 325
CGT TAC ATC GCC GTG GAC GGA GGC ATG TCC GAC AAC ATC CGC CCA GCA      3222
Arg Tyr Ile Ala Val Asp Gly Gly Met Ser Asp Asn Ile Arg Pro Ala
330                 335                 340                 345
CTC TAC GGC TCC GAA TAC GAC GCC CGC GTA GTA TCC CGC TTC GCC GAA      3270
Leu Tyr Gly Ser Glu Tyr Asp Ala Arg Val Val Ser Arg Phe Ala Glu
                350                 355                 360
GGA GAC CCA GTA AGC ACC CGC ATC GTG GGC TCC CAC TGC GAA TCC GGC      3318
Gly Asp Pro Val Ser Thr Arg Ile Val Gly Ser His Cys Glu Ser Gly
            365                 370                 375
GAT ATC CTG ATC AAC GAT GAA ATC TAC CCA TCT GAC ATC ACC AGC GGC      3366
Asp Ile Leu Ile Asn Asp Glu Ile Tyr Pro Ser Asp Ile Thr Ser Gly
        380                 385                 390
GAC TTC CTT GCA CTC GCA GCC ACC GGC GCA TAC TGC TAC GCC ATG AGC      3414
Asp Phe Leu Ala Leu Ala Ala Thr Gly Ala Tyr Cys Tyr Ala Met Ser
    395                 400                 405
TCC CGC TAC AAC GCC TTC ACA CGG CCC GCC GTC GTG TCC GTC CGC GCT      3462
Ser Arg Tyr Asn Ala Phe Thr Arg Pro Ala Val Val Ser Val Arg Ala
410                 415                 420                 425
GGC AGC TCC CGC CTC ATG CTG CGC CGC GAA ACG CTC GAC GAC ATC CTC      3510
Gly Ser Ser Arg Leu Met Leu Arg Arg Glu Thr Leu Asp Asp Ile Leu
                430                 435                 440
TCA CTA GAG GCA TAACGCTTTT CGACGCCTGA CCCCGCCCTT CACCTTCGCC          3562
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Ser Leu Glu Ala
445
GTGGAGGGCG GTTTTGG 3579

(2) INFORMATION FOR SEQ ID NO:19:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 550 amino acids
(B) TYPE: amino acid
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: protein
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

Met Thr Pro Ala Asp Leu Ala Thr Leu Ile Lys Glu Thr Ala Val Glu
1 5 10 15
Val Leu Thr Ser Arg Glu Leu Asp Thr Ser Val Leu Pro Glu Gln Val
20 25 30
Val Val Glu Arg Pro Arg Asn Pro Glu His Gly Asp Tyr Ala Thr Asn
35 40 45
Ile Ala Leu Gln Val Ala Lys Lys Val Gly Gln Asn Pro Arg Asp Leu
50 55 60
Ala Thr Trp Leu Ala Glu Ala Leu Ala Ala Asp Asp Ala Ile Asp Ser
65 70 75 80
Ala Glu Ile Ala Gly Pro Gly Phe Leu Asn Ile Arg Leu Ala Ala Ala
85 90 95
Ala Gln Gly Glu Ile Val Ala Lys Ile Leu Ala Gln Gly Glu Thr Phe
100 105 110
Gly Asn Ser Asp His Leu Ser His Leu Asp Val Asn Leu Glu Phe Val
115 120 125
Ser Ala Asn Pro Thr Gly Pro Ile His Leu Gly Gly Thr Arg Trp Ala
130 135 140
Ala Val Gly Asp Ser Leu Gly Arg Val Leu Glu Ala Ser Gly Ala Lys
145 150 155 160
Val Thr Arg Glu Tyr Tyr Phe Asn Asp His Gly Arg Gln Ile Asp Arg
165 170 175
Phe Ala Leu Ser Leu Leu Ala Ala Ala Lys Gly Glu Pro Thr Pro Glu
180 185 190
Asp Gly Tyr Gly Gly Glu Tyr Ile Lys Glu Ile Ala Glu Ala Ile Val
195 200 205
Glu Lys His Pro Glu Ala Leu Ala Leu Glu Pro Ala Ala Thr Gln Glu
210 215 220
Leu Phe Arg Ala Glu Gly Val Glu Met Met Phe Glu His Ile Lys Ser
225 230 235 240
Ser Leu His Glu Phe Gly Thr Asp Phe Asp Val Tyr Tyr His Glu Asn
245 250 255
Ser Leu Phe Glu Ser Gly Ala Val Asp Lys Ala Val Gln Val Leu Lys
260 265 270

Asp Asn Gly Asn Leu Tyr Glu Asn Glu Gly Ala Trp Trp Leu Arg Ser
275 280 285
Thr Glu Phe Gly Asp Asp Lys Asp Arg Val Val Ile Lys Ser Asp Gly
290 295 300
Asp Ala Ala Tyr Ile Ala Gly Asp Ile Ala Tyr Val Ala Asp Lys Phe
305 310 315 320
Ser Arg Gly His Asn Leu Asn Ile Tyr Met Leu Gly Ala Asp His His
325 330 335
Gly Tyr Ile Ala Arg Leu Lys Ala Ala Ala Ala Ala Leu Gly Tyr Lys
340 345 350
Pro Glu Gly Val Glu Val Leu Ile Gly Gln Met Val Asn Leu Leu Arg
355 360 365
Asp Gly Lys Ala Val Arg Met Ser Lys Arg Ala Gly Thr Val Val Thr
370 375 380
Leu Asp Asp Leu Val Glu Ala Ile Gly Ile Asp Ala Ala Arg Tyr Ser
385 390 395 400
Leu Ile Arg Ser Ser Val Asp Ser Ser Leu Asp Ile Asp Leu Gly Leu
405 410 415
Trp Glu Ser Gln Ser Ser Asp Asn Pro Val Tyr Tyr Val Gln Tyr Gly
420 425 430
His Ala Arg Leu Cys Ser Ile Ala Arg Lys Ala Glu Thr Leu Gly Val
435 440 445
Thr Glu Glu Gly Ala Asp Leu Ser Leu Leu Thr His Asp Arg Glu Gly
450 455 460
Asp Leu Ile Arg Thr Leu Gly Glu Phe Pro Ala Val Val Lys Ala Ala
465 470 475 480
Ala Asp Leu Arg Glu Pro His Arg Ile Ala Arg Tyr Ala Glu Glu Leu
485 490 495
Ala Gly Thr Phe His Arg Phe Tyr Asp Ser Cys His Ile Leu Pro Lys
500 505 510
Val Asp Glu Asp Thr Ala Pro Ile His Thr Ala Arg Leu Ala Leu Ala
515 520 525
Ala Ala Thr Arg Gln Thr Leu Ala Asn Ala Leu His Leu Val Gly Val
530 535 540
Ser Ala Pro Glu Lys Met
545 550

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 445 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

Met Ala Thr Val Glu Asn Phe Asn Glu Leu Pro Ala His Val Trp Pro

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  1               5                  10                  15
Arg Asn Ala Val Arg Gln Glu Asp Gly Val Val Thr Val Ala Gly Val
            20                  25                  30
Pro Leu Pro Asp Leu Ala Glu Glu Tyr Gly Thr Pro Leu Phe Val Val
        35                  40                  45
Asp Glu Asp Asp Phe Arg Ser Arg Cys Arg Asp Met Ala Thr Ala Phe
    50                  55                  60
Gly Gly Pro Gly Asn Val His Tyr Ala Ser Lys Ala Phe Leu Thr Lys
65                  70                  75                  80
Thr Ile Ala Arg Trp Val Asp Glu Glu Gly Leu Ala Leu Asp Ile Ala
                85                  90                  95
Ser Ile Asn Glu Leu Gly Ile Ala Leu Ala Ala Gly Phe Pro Ala Ser
            100                 105                 110
Arg Ile Thr Ala His Gly Asn Asn Lys Gly Val Glu Phe Leu Arg Ala
        115                 120                 125
Leu Val Gln Asn Gly Val Gly His Val Val Leu Asp Ser Ala Gln Glu
    130                 135                 140
Leu Glu Leu Leu Asp Tyr Val Ala Ala Gly Glu Gly Lys Ile Gln Asp
145                 150                 155                 160
Val Leu Ile Arg Val Lys Pro Gly Ile Glu Ala His Thr His Glu Phe
                165                 170                 175
Ile Ala Thr Ser His Glu Asp Gln Lys Phe Gly Phe Ser Leu Ala Ser
            180                 185                 190
Gly Ser Ala Phe Glu Ala Ala Lys Ala Ala Asn Asn Ala Glu Asn Leu
        195                 200                 205
Asn Leu Val Gly Leu His Cys His Val Gly Ser Gln Val Phe Asp Ala
    210                 215                 220
Glu Gly Phe Lys Leu Ala Ala Glu Arg Val Leu Gly Leu Tyr Ser Gln
225                 230                 235                 240
Ile His Ser Glu Leu Gly Val Ala Leu Pro Glu Leu Asp Leu Gly Gly
                245                 250                 255
Gly Tyr Gly Ile Ala Tyr Thr Ala Ala Glu Glu Pro Leu Asn Val Ala
            260                 265                 270
Glu Val Ala Ser Asp Leu Leu Thr Ala Val Gly Lys Met Ala Ala Glu
        275                 280                 285
Leu Gly Ile Asp Ala Pro Thr Val Leu Val Glu Pro Gly Arg Ala Ile
    290                 295                 300
Ala Gly Pro Ser Thr Val Thr Ile Tyr Glu Val Gly Thr Thr Lys Asp
305                 310                 315                 320
Val His Val Asp Asp Asp Lys Thr Arg Arg Tyr Ile Ala Val Asp Gly
                325                 330                 335
Gly Met Ser Asp Asn Ile Arg Pro Ala Leu Tyr Gly Ser Glu Tyr Asp
            340                 345                 350
Ala Arg Val Val Ser Arg Phe Ala Glu Gly Asp Pro Val Ser Thr Arg
        355                 360                 365
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Ile Val Gly Ser His Cys Glu Ser Gly Asp Ile Leu Ile Asn Asp Glu
370 375 380
Ile Tyr Pro Ser Asp Ile Thr Ser Gly Asp Phe Leu Ala Leu Ala Ala
385 390 395 400
Thr Gly Ala Tyr Cys Tyr Ala Met Ser Ser Arg Tyr Asn Ala Phe Thr
405 410 415
Arg Pro Ala Val Val Ser Val Arg Ala Gly Ser Ser Arg Leu Met Leu
420 425 430
Arg Arg Glu Thr Leu Asp Asp Ile Leu Ser Leu Glu Ala
435 440 445

(2) INFORMATION FOR SEQ ID NO:21:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: NO
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:
CATCTAAGTA TGCATCTCGG 20

(2) INFORMATION FOR SEQ ID NO:22:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 20 bases
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: other nucleic acid
(A) DESCRIPTION: /desc = "synthetic DNA"
(iv) ANTI-SENSE: YES
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:
TGCCCCTCGA GCTAAATTAG 20

(2) INFORMATION FOR SEQ ID NO:23:
(i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 1034 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear
(ii) MOLECULE TYPE: DNA (genomic)
(iv) ANTI-SENSE: NO
(vi) ORIGINAL SOURCE:
(A) ORGANISM: Brevibacterium lactofermentum

(B) STRAIN: ATCC 13869

(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 61..1020

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:

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ATGCATCTCG GTAAGCTCGA CCAGGACAGT GCCACCACAA TTTTGGAGGA TTACAAGAAC      60
ATG ACC AAC ATC CGC GTA GCT ATC GTG GGC TAC GGA AAC CTG GGA CGC        108
Met Thr Asn Ile Arg Val Ala Ile Val Gly Tyr Gly Asn Leu Gly Arg
 1               5                  10                  15
AGC GTC GAA AAG CTT ATT GCC AAG CAG CCC GAC ATG GAC CTT GTA GGA        156
Ser Val Glu Lys Leu Ile Ala Lys Gln Pro Asp Met Asp Leu Val Gly
            20                  25                  30
ATC TTC TCG CGC CGG GCC ACC CTC GAC ACA AAG ACG CCA GTC TTT GAT        204
Ile Phe Ser Arg Arg Ala Thr Leu Asp Thr Lys Thr Pro Val Phe Asp
        35                  40                  45
GTC GCC GAC GTG GAC AAG CAC GCC GAC GAC GTG GAC GTG CTG TTC CTG        252
Val Ala Asp Val Asp Lys His Ala Asp Asp Val Asp Val Leu Phe Leu
    50                  55                  60
TGC ATG GGC TCC GCC ACC GAC ATC CCT GAG CAG GCA CCA AAG TTC GCG        300
Cys Met Gly Ser Ala Thr Asp Ile Pro Glu Gln Ala Pro Lys Phe Ala
65                  70                  75                  80
CAG TTC GCC TGC ACC GTA GAC ACC TAC GAC AAC CAC CGC GAC ATC CCA        348
Gln Phe Ala Cys Thr Val Asp Thr Tyr Asp Asn His Arg Asp Ile Pro
                85                  90                  95
CGC CAC CGC CAG GTC ATG AAC GAA GCC GCC ACC GCA GCC GGC AAC GTT        396
Arg His Arg Gln Val Met Asn Glu Ala Ala Thr Ala Ala Gly Asn Val
            100                 105                 110
GCA CTG GTC TCT ACC GGC TGG GAT CCA GGA ATG TTC TCC ATC AAC CGC        444
Ala Leu Val Ser Thr Gly Trp Asp Pro Gly Met Phe Ser Ile Asn Arg
        115                 120                 125
GTC TAC GCA GCG GCA GTC TTA GCC GAG CAC CAG CAG CAC ACC TTC TGG        492
Val Tyr Ala Ala Ala Val Leu Ala Glu His Gln Gln His Thr Phe Trp
    130                 135                 140
GGC CCA GGT TTG TCA CAG GGC CAC TCC GAT GCT TTG CGA CGC ATC CCT        540
Gly Pro Gly Leu Ser Gln Gly His Ser Asp Ala Leu Arg Arg Ile Pro
145                 150                 155                 160
GGC GTT CAA AAG GCA GTC CAG TAC ACC CTC CCA TCC GAA GAC GCC CTG        588
Gly Val Gln Lys Ala Val Gln Tyr Thr Leu Pro Ser Glu Asp Ala Leu
                165                 170                 175
GAA AAG GCC CGC CGC GGC GAA GCC GGC GAC CTT ACC GGA AAG CAA ACC        636
Glu Lys Ala Arg Arg Gly Glu Ala Gly Asp Leu Thr Gly Lys Gln Thr
            180                 185                 190
CAC AAG CGC CAA TGC TTC GTG GTT GCC GAC GCG GCC GAT CAC GAG CGC        684
His Lys Arg Gln Cys Phe Val Val Ala Asp Ala Ala Asp His Glu Arg
        195                 200                 205
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ATC GAA AAC GAC ATC CGC ACC ATG CCT GAT TAC TTC GTT GGC TAC GAA 732
Ile Glu Asn Asp Ile Arg Thr Met Pro Asp Tyr Phe Val Gly Tyr Glu
210 215 220

GTC GAA GTC AAC TTC ATC GAC GAA GCA ACC TTC GAC TCC GAG CAC ACC 780
Val Glu Val Asn Phe Ile Asp Glu Ala Thr Phe Asp Ser Glu His Thr
225 230 235 240

GGC ATG CCA CAC GGT GGC CAC GTG ATT ACC ACC GGC GAC ACC GGT GGC 828
Gly Met Pro His Gly Gly His Val Ile Thr Thr Gly Asp Thr Gly Gly
245 250 255

TTC AAC CAC ACC GTG GAA TAC ATC CTC AAG CTG GAC CGA AAC CCA GAT 876
Phe Asn His Thr Val Glu Tyr Ile Leu Lys Leu Asp Arg Asn Pro Asp
260 265 270

TTC ACC GCT TCC TCA CAG ATC GCT TTC GGT CGC GCA GCT CAC CGC ATG 924
Phe Thr Ala Ser Ser Gln Ile Ala Phe Gly Arg Ala Ala His Arg Met
275 280 285

AAG CAG CAG GGC CAA AGC GGA GCT TTC ACC GTC CTC GAA GTT GCT CCA 972
Lys Gln Gln Gly Gln Ser Gly Ala Phe Thr Val Leu Glu Val Ala Pro
290 295 300

TAC CTG CTC TCC CCA GAG AAC TTG GAC GAT CTG ATC GCA CGC GAC GTC 1020
Tyr Leu Leu Ser Pro Glu Asn Leu Asp Asp Leu Ile Ala Arg Asp Val
305 310 315 320

TAATTTAGCT CGAG 1034

(2) INFORMATION FOR SEQ ID NO:24:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 320 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

Met Thr Asn Ile Arg Val Ala Ile Val Gly Tyr Gly Asn Leu Gly Arg
1 5 10 15

Ser Val Glu Lys Leu Ile Ala Lys Gln Pro Asp Met Asp Leu Val Gly
20 25 30

Ile Phe Ser Arg Arg Ala Thr Leu Asp Thr Lys Thr Pro Val Phe Asp
35 40 45

Val Ala Asp Val Asp Lys His Ala Asp Asp Val Asp Val Leu Phe Leu
50 55 60

Cys Met Gly Ser Ala Thr Asp Ile Pro Glu Gln Ala Pro Lys Phe Ala
65 70 75 80

Gln Phe Ala Cys Thr Val Asp Thr Tyr Asp Asn His Arg Asp Ile Pro
85 90 95

Arg His Arg Gln Val Met Asn Glu Ala Ala Thr Ala Ala Gly Asn Val
100 105 110

Ala Leu Val Ser Thr Gly Trp Asp Pro Gly Met Phe Ser Ile Asn Arg

115 120 125
Val Tyr Ala Ala Ala Val Leu Ala Glu His Gln Gln His Thr Phe Trp
130 135 140
Gly Pro Gly Leu Ser Gln Gly His Ser Asp Ala Leu Arg Arg Ile Pro
145 150 155 160
Gly Val Gln Lys Ala Val Gln Tyr Thr Leu Pro Ser Glu Asp Ala Leu
165 170 175
Glu Lys Ala Arg Arg Gly Glu Ala Gly Asp Leu Thr Gly Lys Gln Thr
180 185 190
His Lys Arg Gln Cys Phe Val Val Ala Asp Ala Ala Asp His Glu Arg
195 200 205
Ile Glu Asn Asp Ile Arg Thr Met Pro Asp Tyr Phe Val Gly Tyr Glu
210 215 220
Val Glu Val Asn Phe Ile Asp Glu Ala Thr Phe Asp Ser Glu His Thr
225 230 235 240
Gly Met Pro His Gly Gly His Val Ile Thr Thr Gly Asp Thr Gly Gly
245 250 255
Phe Asn His Thr Val Glu Tyr Ile Leu Lys Leu Asp Arg Asn Pro Asp
260 265 270
Phe Thr Ala Ser Ser Gln Ile Ala Phe Gly Arg Ala Ala His Arg Met
275 280 285
Lys Gln Gln Gly Gln Ser Gly Ala Phe Thr Val Leu Glu Val Ala Pro
290 295 300
Tyr Leu Leu Ser Pro Glu Asn Leu Asp Asp Leu Ile Ala Arg Asp Val
305 310 315 320

What is claimed is:

1. A method for producing L-lysine comprising the steps of cultivating a coryneform bacterium *Corynebacterium glutamicum* in an appropriate medium, to produce and accumulate L-lysine in a culture of the bacterium, and collecting L-lysine from the culture, wherein the coryneform bacterium harbors an aspartokinase wherein said aspartokinase is a mutant aspartokinase in which a 279^{th} alanine residue as counted from its N-terminal is changed into an amino acid residue other than alanine and other than acidic amino acid in its α-subunit as shown in SEQ ID NO: 5 in the Sequence Listing, and a 30^{th} alanine residue is changed into an amino acid residue other than alanine and other than acidic amino acid in its β-subunit as shown in SEQ ID NO: 7 in the Sequence Listing, and in which feedback inhibition by L-lysine and L-threonine is desensitized as compared with a wild type aspartokinase, and wherein the intracellular activities of dihydrodipicolinate reductase and dihydrodipicolinate synthase are upregulated by increasing the copy number of, and/or using a strong promoter for a DNA sequence from *Corynebacterium glutamicum* coding for a dihydrodipicolinate reductase and a DNA sequence from *Corynebacterium glutamicum* coding for a dihydrodipicolinate synthase, and wherein the intracellular activity of diaminopimelate decarboxylase is further upregulated by increasing the copy number of, and/or using a strong promoter for a DNA sequence from *Corynebacterium glutamicum* coding for a diaminopimelate decarboxylase.

2. The method according to claim 1, where the

intracellular activity of diaminopimelate dehydrogenase is further raised by increasing the copy number of, and/or using a strong promoter for a DNA sequence from *Corynebacterium glutamicum* coding for a diaminopimelate dehydrogenase.

3. The method according to claim 1, wherein the corynebacterium is transformed by introduction of a recombinant DNA autonomously replicable in cells of coryneform bacteria, comprising a DNA sequence from *Corynebacterium glutamicum coding* for an aspartokinase in which feedback inhibition by L-lysine and L-threonine is desensitized as compared with a wild type aspartokinase, and a DNA sequence from *Corynebacterium glutamicum* coding for a dihydrodipicolinate reductase, and a DNA sequence from *Corynebacterium glutamicum* coding for a dihydrodipicolinate synthase, and wherein the recombinant DNA further comprises a DNA sequence from *Corynebacterium glutamicum* coding for a diaminopimelate decarboxylase.

4. The method according to claim 3, wherein the recombinant DNA further comprises a DNA sequence from *Corynebacterium glutamicum* coding for a diaminopimelate dehydrogenase.

5. The method according to claim 1, wherein said DNA sequence coding for the dihydrodipicolinate reductase codes for an amino acid sequence depicted in SEQ ID NO: 11 in the Sequence Listing.

6. The method according to claim 1, where said DNA sequence coding for the dihydrodipicolinate synthase

codes for an amino acid sequence depicted in SEQ ID NO: 15 in Sequence Listing.

7. The method according to claim 2, wherein said DNA sequence coding for the diaminopimelate decarboxylase codes for an amino acid sequence depicted in SEQ ID NO: 20 in Sequence Listing.

8. The method according to claim 2, wherein said DNA sequence coding for the diaminopimelate dehydrogenase codes for an amino acid sequence depicted in SEQ ID NO: 24 in Sequence Listing.

FIG. 1

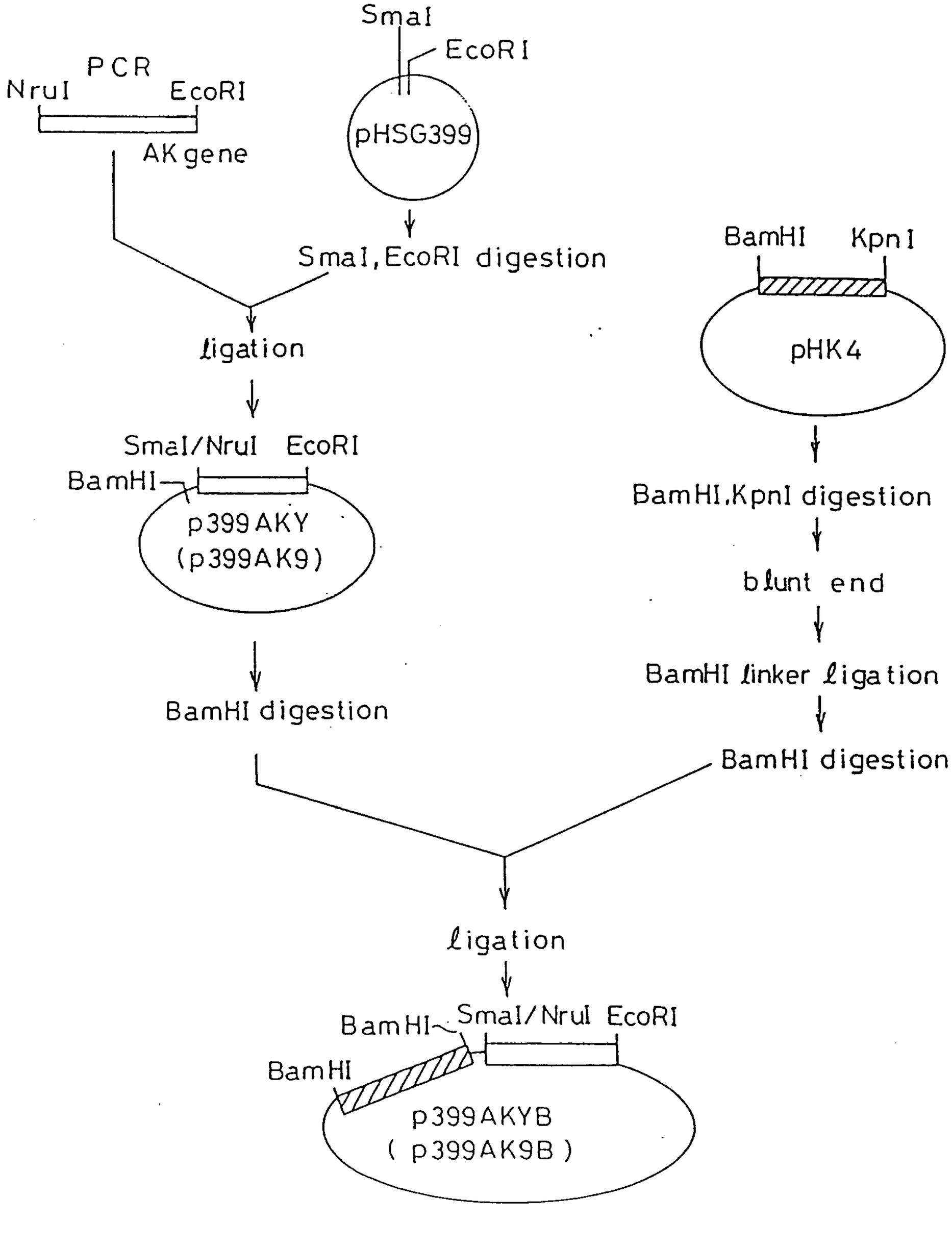

SmaI
EcoRI
pHSG399
PCR
NruI
EcoRI
AK gene
SmaI, EcoRI digestion
ligation
SmaI/NruI EcoRI
BamHI
p399AKY
(p399AK9)
BamHI digestion
BamHI KpnI
pHK4
BamHI, KpnI digestion
blunt end
BamHI linker ligation
BamHI digestion
ligation
BamHI
SmaI/NruI EcoRI
BamHI
p399AKYB
(p399AK9B)

FIG. 2

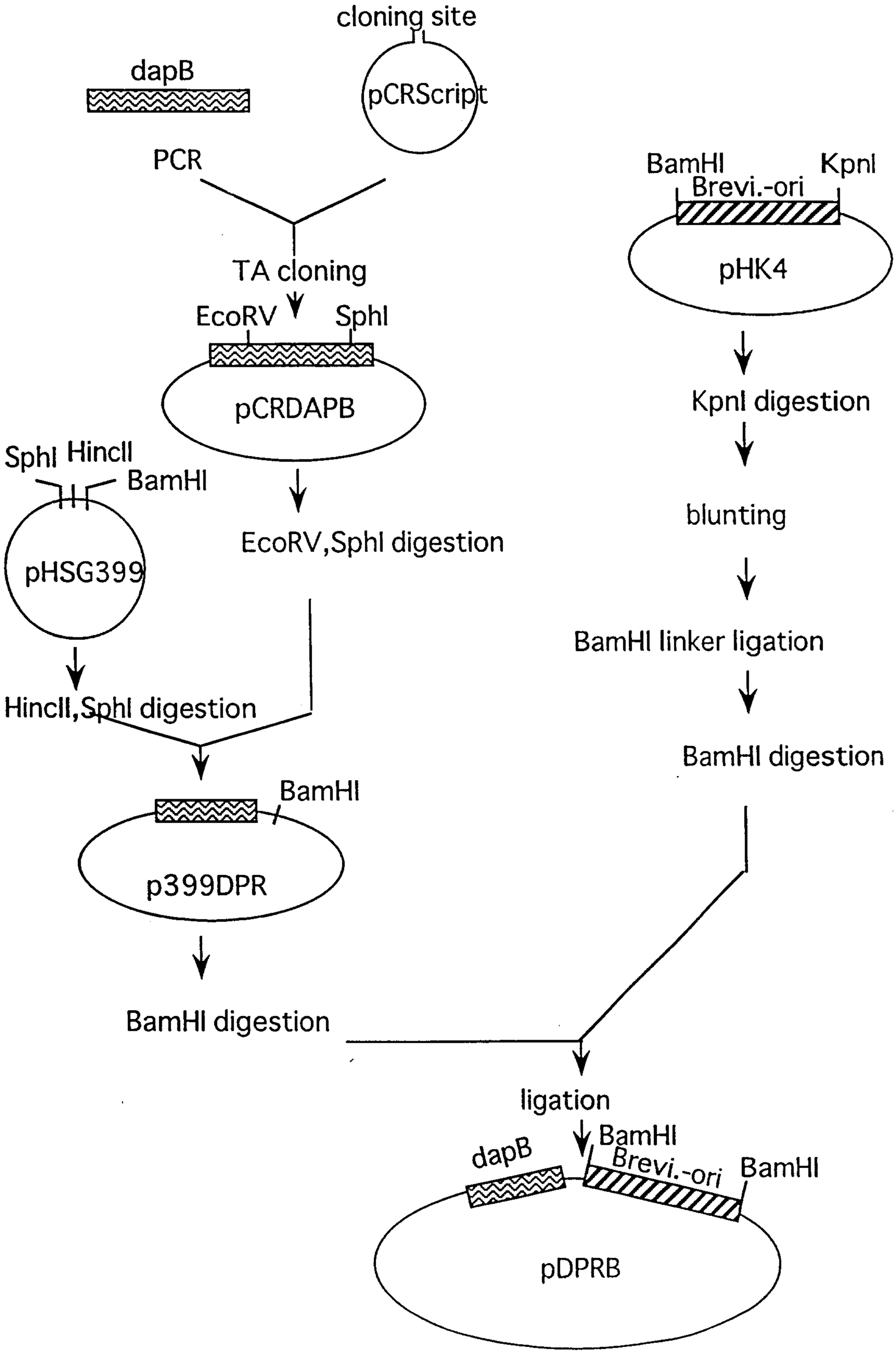

cloning site
dapB
pCRScript
PCR
TA cloning
EcoRV
SphI
pCRDAPB
SphI
HincII
BamHI
pHSG399
EcoRV,SphI digestion
HincII,SphI digestion
BamHI
p399DPR
BamHI digestion
BamHI
KpnI
Brevi.-ori
pHK4
KpnI digestion
blunting
BamHI linker ligation
BamHI digestion
ligation
dapB
BamHI
Brevi.-ori
BamHI
pDPRB

FIG. 3

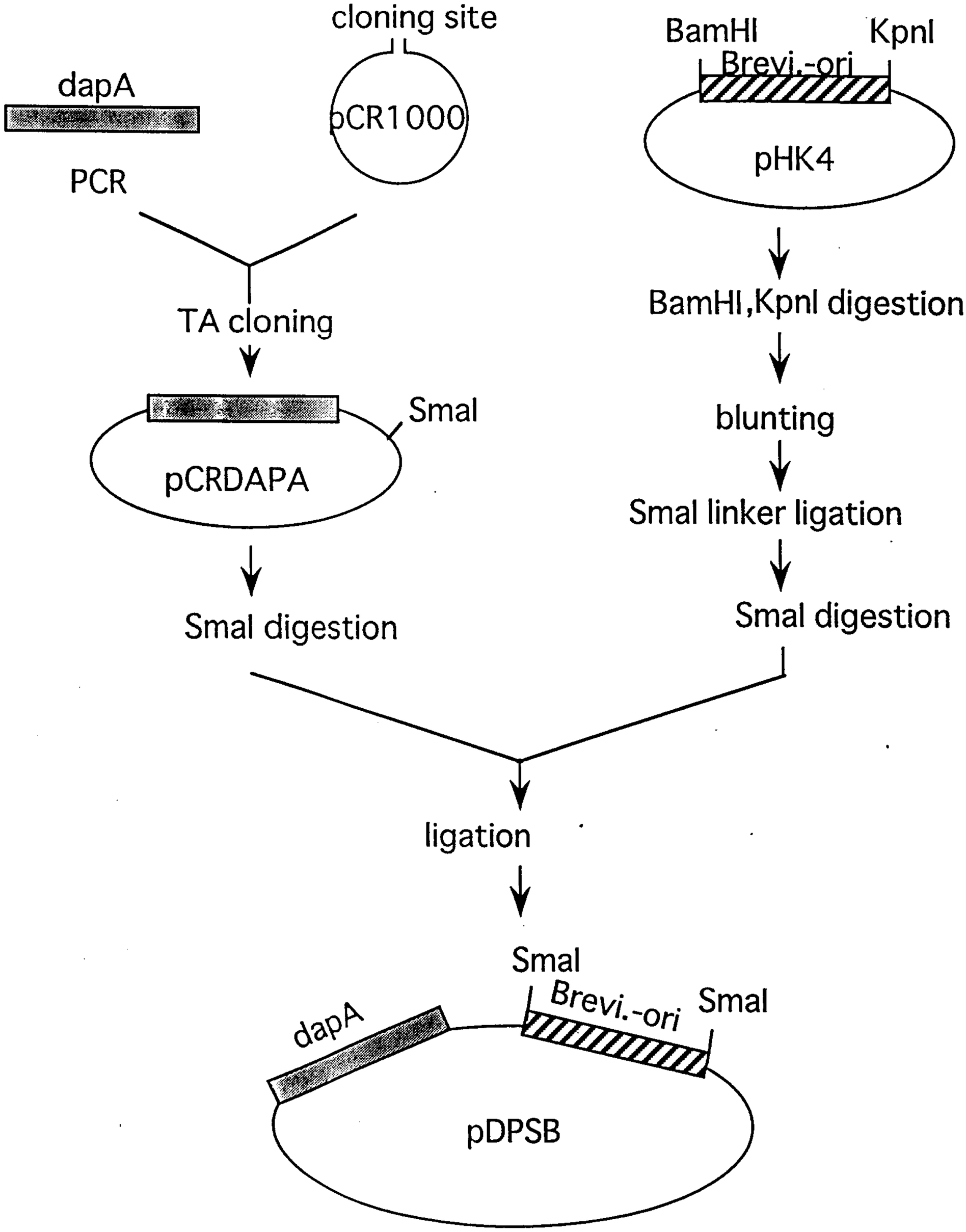

cloning site
dapA
pCR1000
PCR
TA cloning
SmaI
pCRDAPA
SmaI digestion
BamHI
KpnI
Brevi.-ori
pHK4
BamHI,KpnI digestion
blunting
SmaI linker ligation
SmaI digestion
ligation
SmaI
dapA
Brevi.-ori
SmaI
pDPSB

FIG. 4

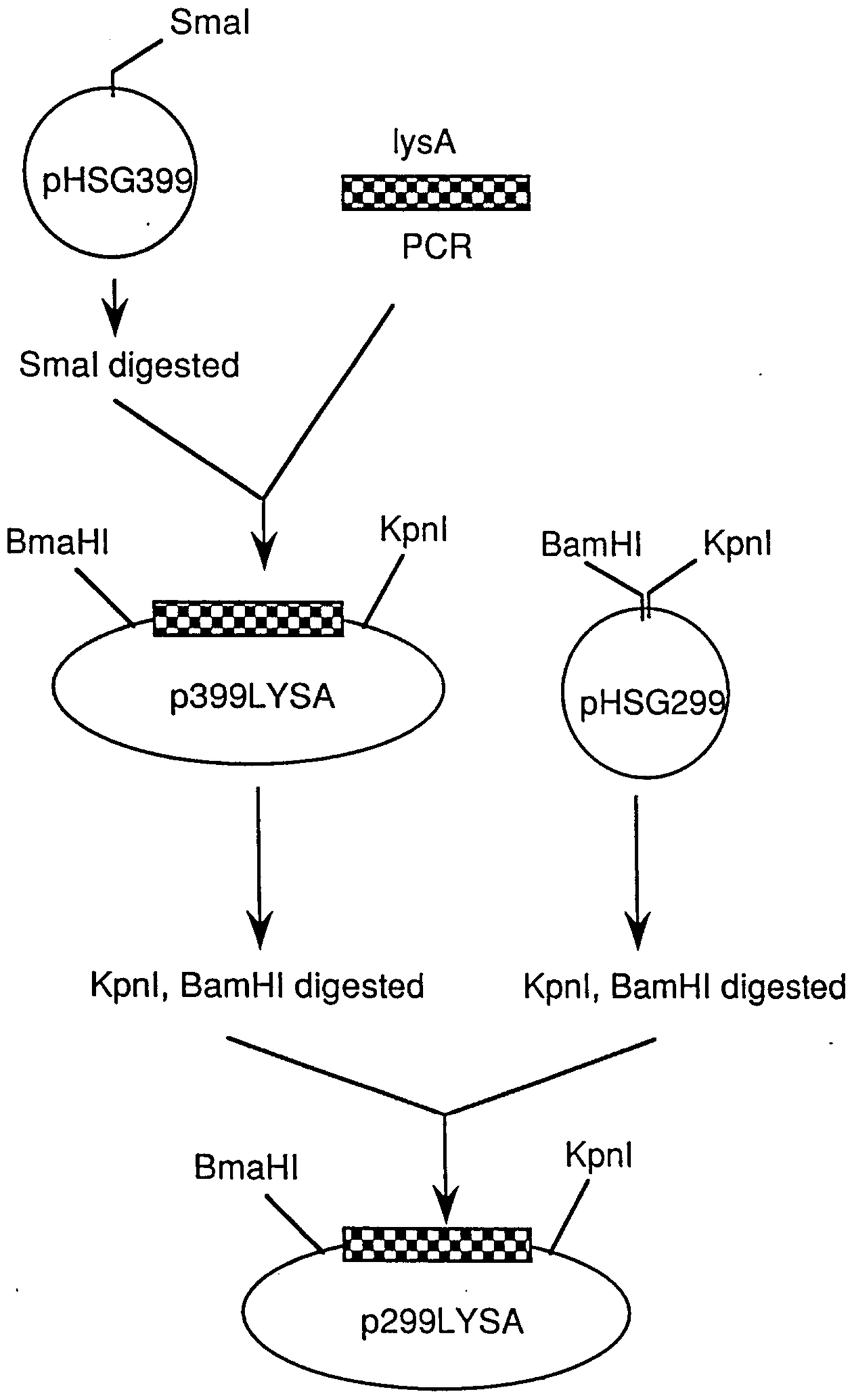

SmaI
pHSG399
lysA
PCR
SmaI digested
BmaHI
KpnI
p399LYSA
BamHI
KpnI
pHSG299
KpnI, BamHI digested
KpnI, BamHI digested
BmaHI
KpnI
p299LYSA

FIG. 5

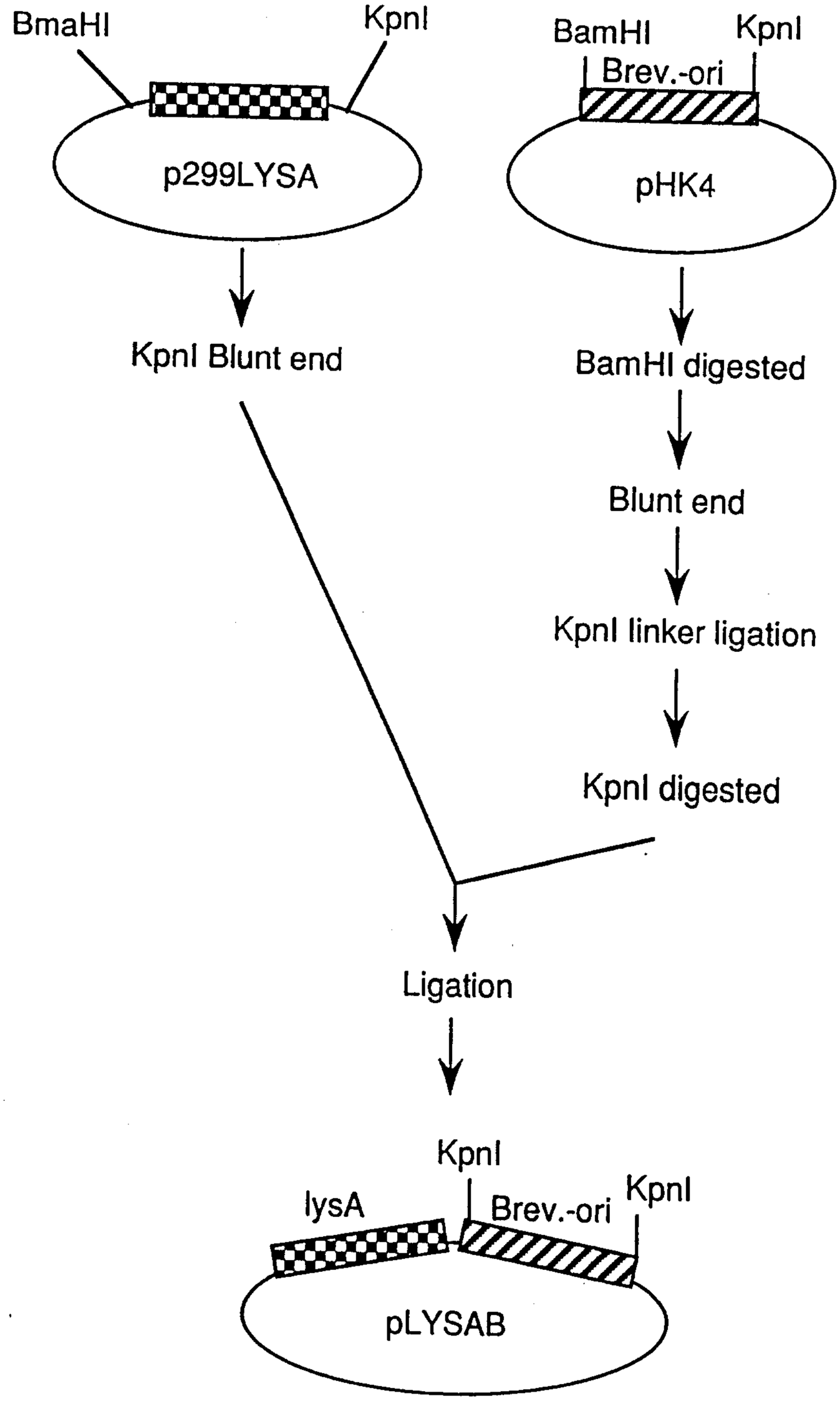

BmaHI
KpnI
p299LYSA
KpnI Blunt end
BamHI
KpnI
Brev.-ori
pHK4
BamHI digested
Blunt end
KpnI linker ligation
KpnI digested
Ligation
KpnI
lysA
Brev.-ori
KpnI
pLYSAB

FIG. 6

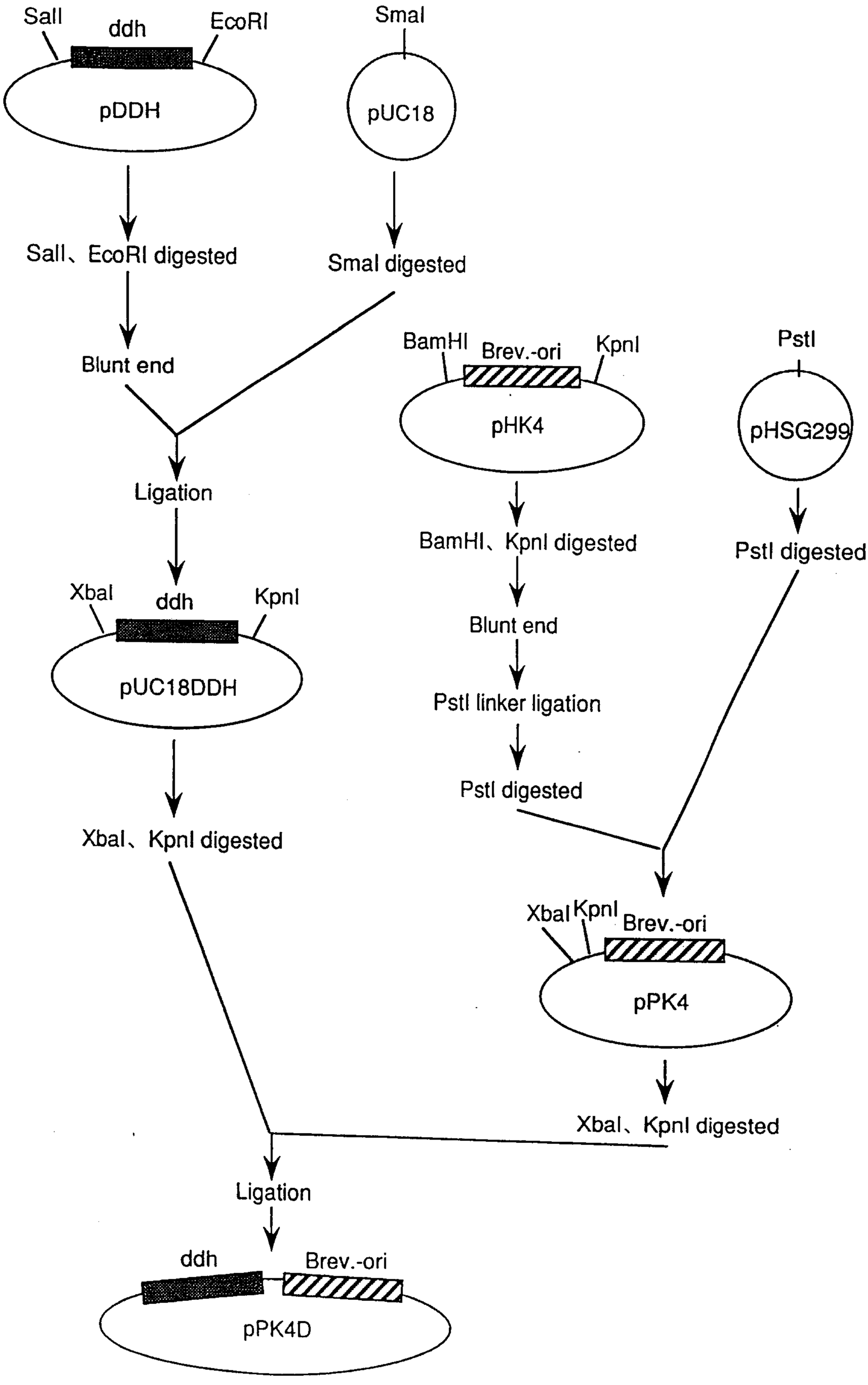

SalI
ddh
EcoRI
pDDH
SmaI
pUC18
SalI、EcoRI digested
SmaI digested
Blunt end
Ligation
BamHI
Brev.-ori
KpnI
pHK4
PstI
pHSG299
BamHI、KpnI digested
PstI digested
Blunt end
PstI linker ligation
PstI digested
XbaI
ddh
KpnI
pUC18DDH
XbaI、KpnI digested
XbaI
KpnI
Brev.-ori
pPK4
XbaI、KpnI digested
Ligation
ddh
Brev.-ori
pPK4D

FIG. 7

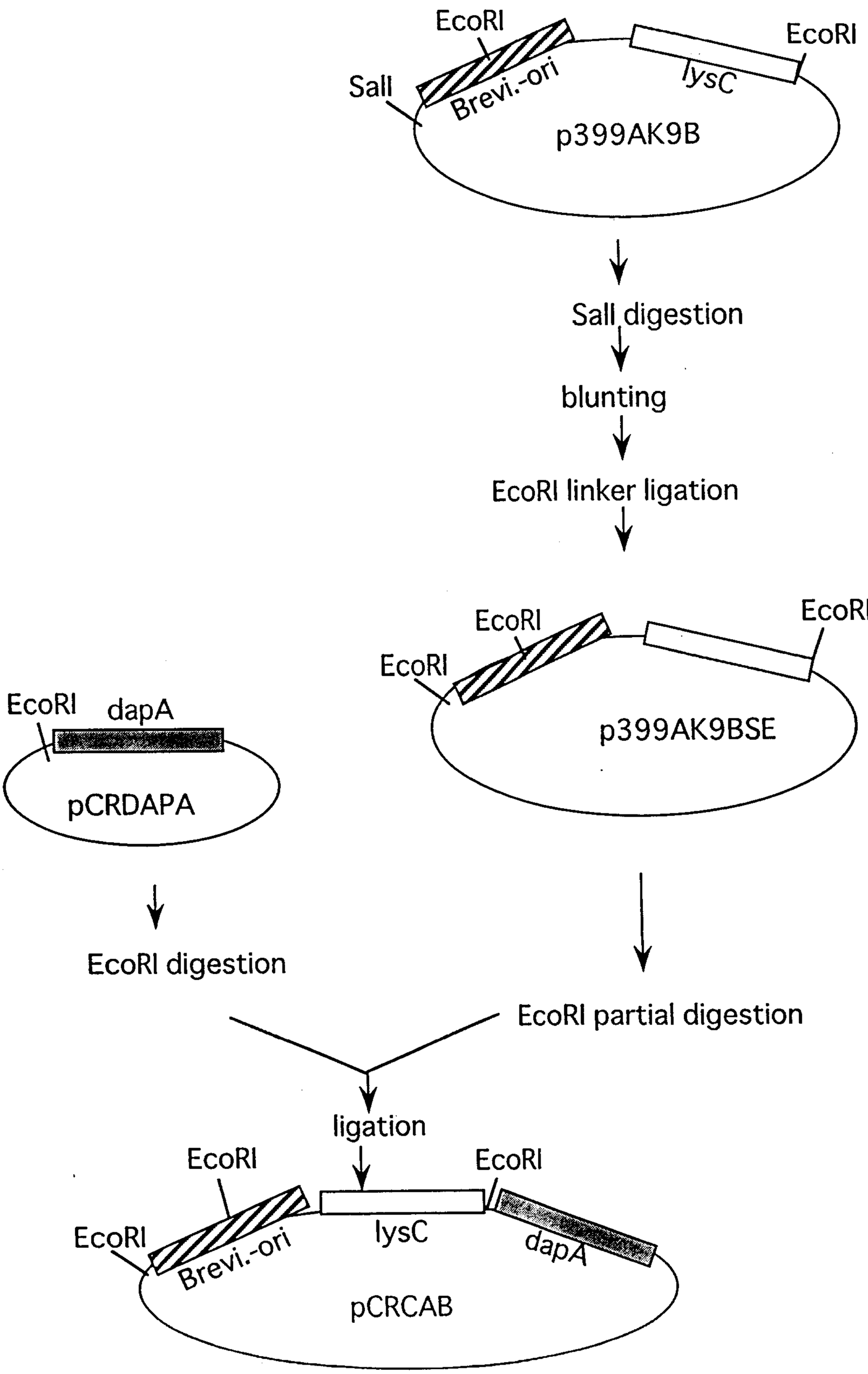

EcoRI
EcoRI
SalI
Brevi.-ori
lysC
p399AK9B
SalI digestion
blunting
EcoRI linker ligation
EcoRI
EcoRI
EcoRI
EcoRI
dapA
p399AK9BSE
pCRDAPA
EcoRI digestion
EcoRI partial digestion
ligation
EcoRI
EcoRI
EcoRI
Brevi.-ori
lysC
dapA
pCRCAB

FIG. 8

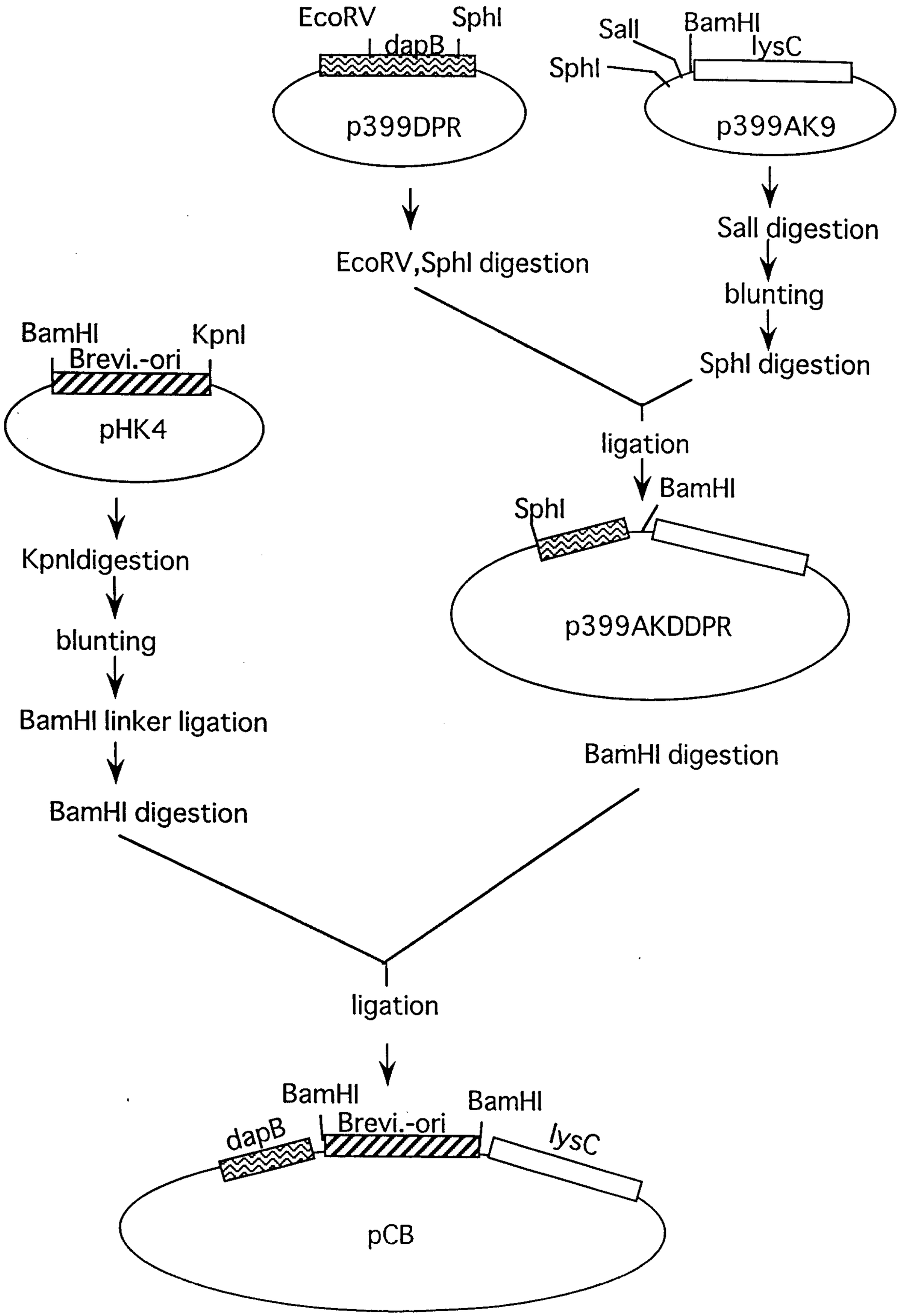

EcoRV
SphI
dapB
p399DPR
EcoRV,SphI digestion
SalI
BamHI
lysC
SphI
p399AK9
SalI digestion
blunting
SphI digestion
ligation
BamHI
SphI
p399AKDDPR
BamHI
KpnI
Brevi.-ori
pHK4
KpnIdigestion
blunting
BamHI linker ligation
BamHI digestion
BamHI digestion
ligation
BamHI
BamHI
dapB
Brevi.-ori
lysC
pCB

FIG. 9

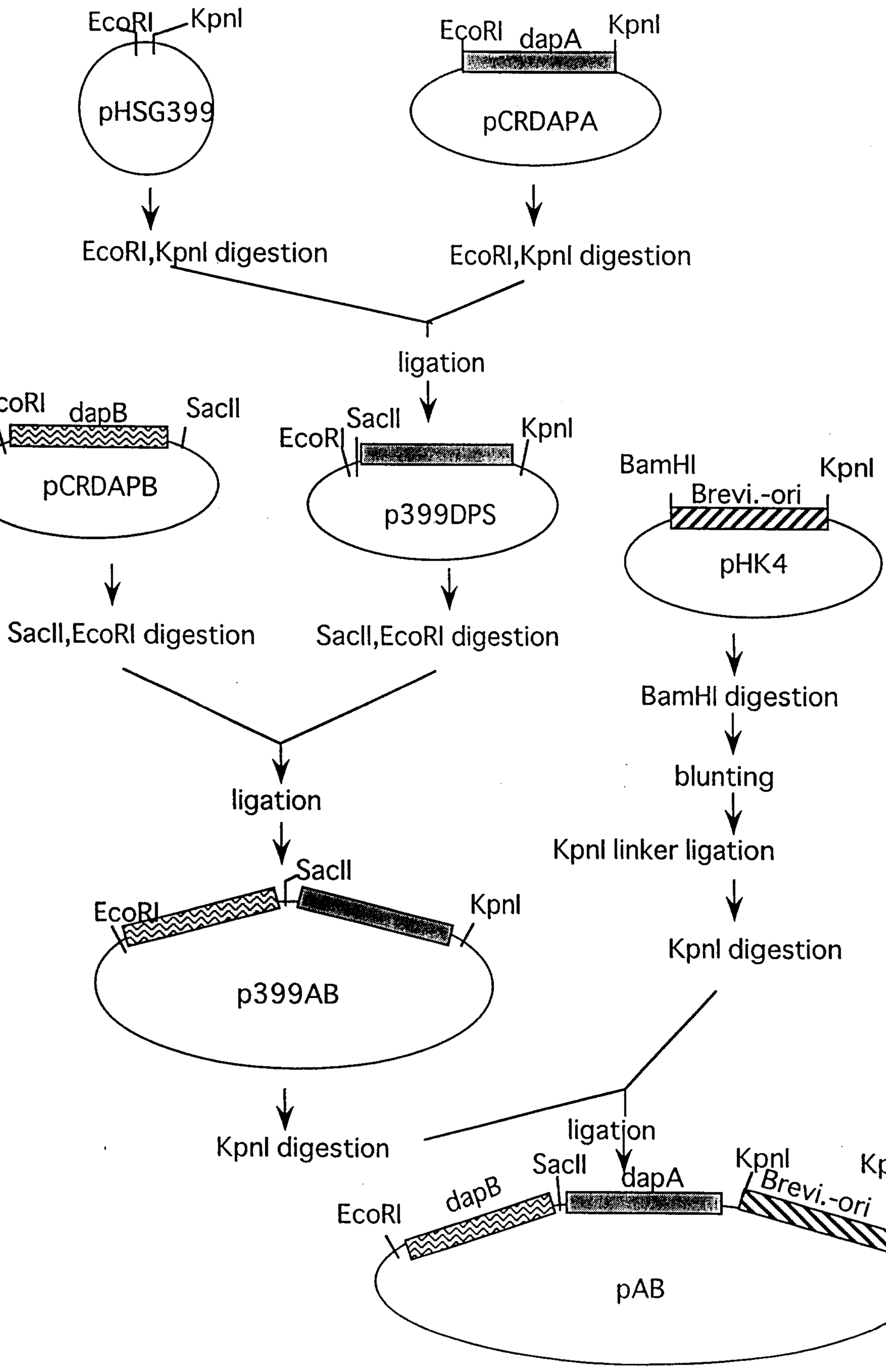

EcoRI
KpnI
pHSG399
EcoRI dapA KpnI
pCRDAPA
EcoRI,KpnI digestion
EcoRI,KpnI digestion
ligation
EcoRI dapB SacII
pCRDAPB
SacII
EcoRI
KpnI
p399DPS
BamHI
KpnI
Brevi.-ori
pHK4
SacII,EcoRI digestion
SacII,EcoRI digestion
BamHI digestion
ligation
blunting
SacII
EcoRI
KpnI
p399AB
KpnI linker ligation
KpnI digestion
KpnI digestion
ligation
KpnI
KpnI
SacII
dapA
EcoRI
dapB
Brevi.-ori
pAB

FIG. 10

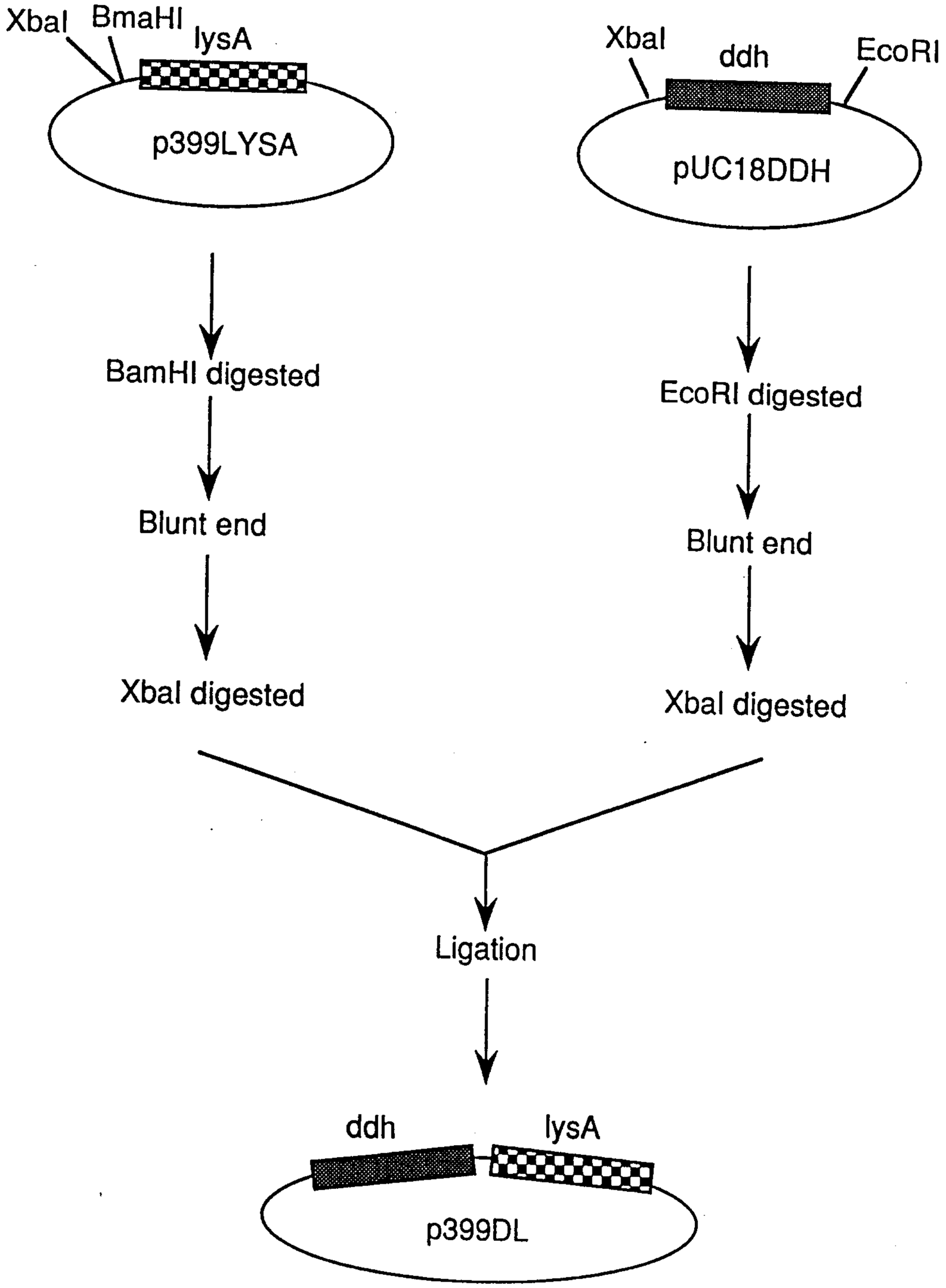

XbaI
BmaHI
lysA
p399LYSA
XbaI
ddh
EcoRI
pUC18DDH
BamHI digested
EcoRI digested
Blunt end
Blunt end
XbaI digested
XbaI digested
Ligation
ddh
lysA
p399DL

FIG. 11

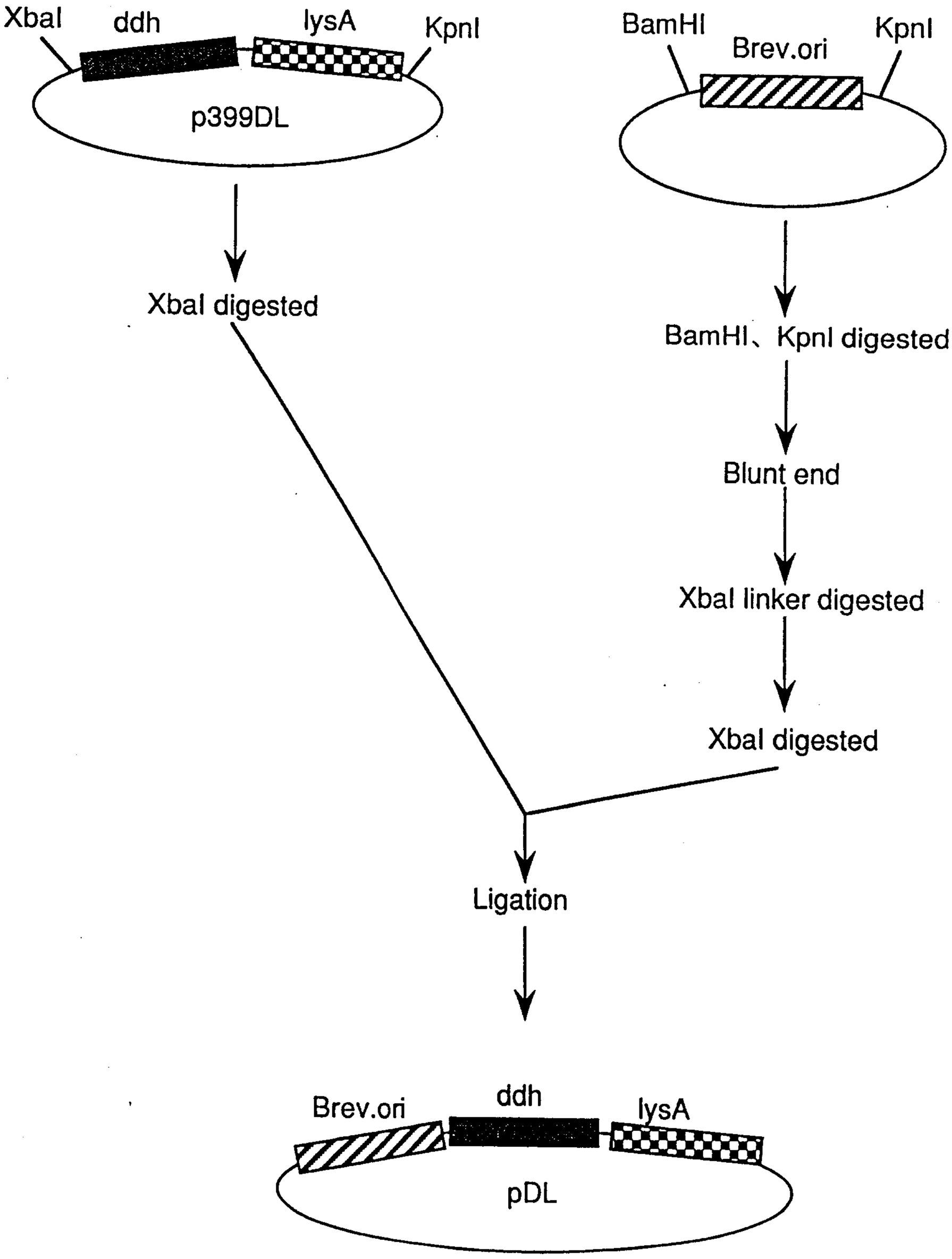

XbaI
ddh
lysA
KpnI
p399DL
XbaI digested
BamHI
Brev.ori
KpnI
BamHI、KpnI digested
Blunt end
XbaI linker digested
XbaI digested
Ligation
Brev.ori
ddh
lysA
pDL

FIG. 12

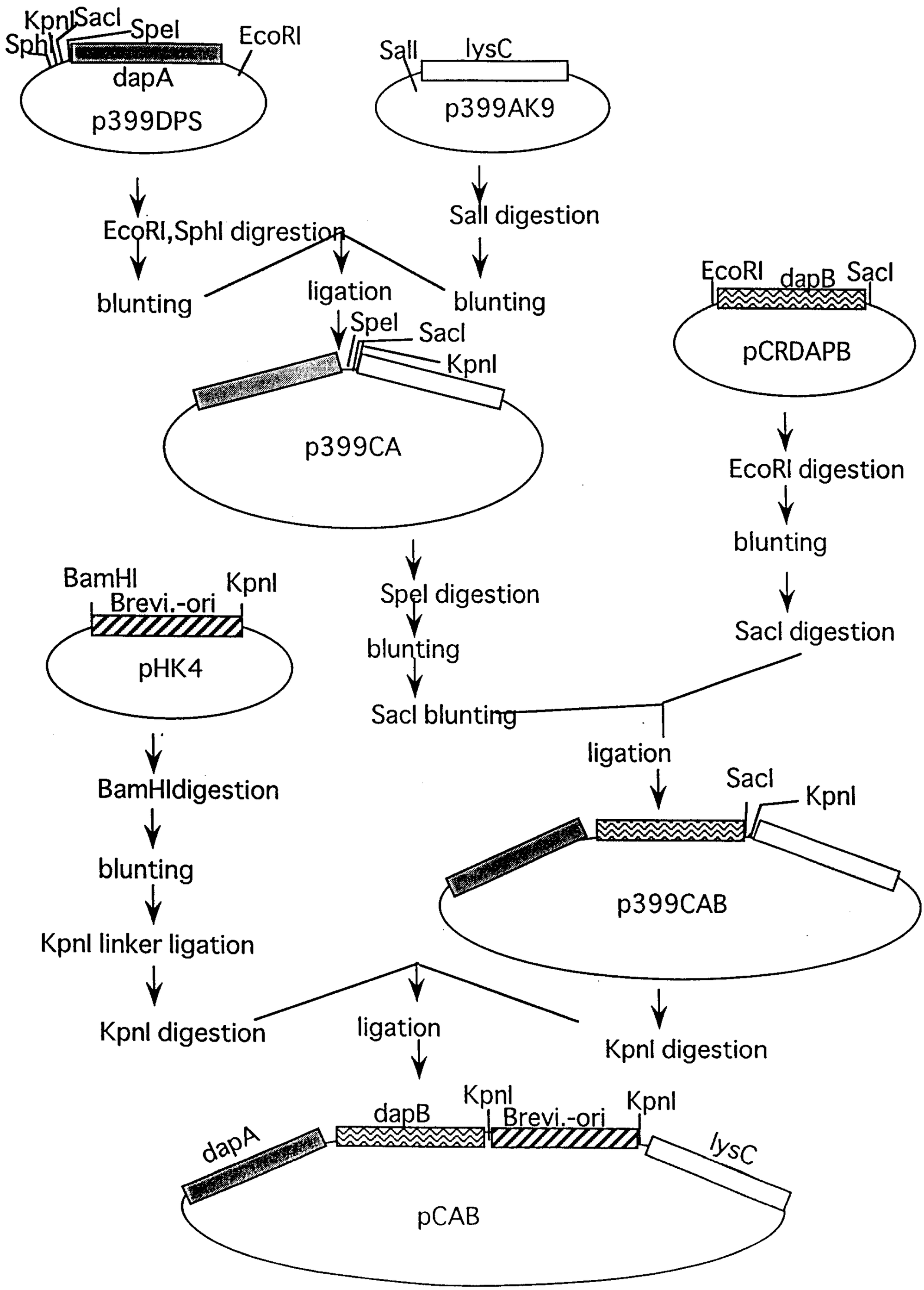

KpnI
SacI
SpeI
SphI
EcoRI
dapA
p399DPS
SalI
lysC
p399AK9
EcoRI,SphI digrestion
SalI digestion
blunting
ligation
blunting
SpeI
SacI
KpnI
p399CA
EcoRI
dapB
SacI
pCRDAPB
EcoRI digestion
blunting
SacI digestion
BamHI
KpnI
Brevi.-ori
pHK4
SpeI digestion
blunting
SacI blunting
ligation
SacI
KpnI
p399CAB
BamHIdigestion
blunting
KpnI linker ligation
KpnI digestion
ligation
KpnI digestion
KpnI
KpnI
dapB
Brevi.-ori
dapA
lysC
pCAB

FIG. 13

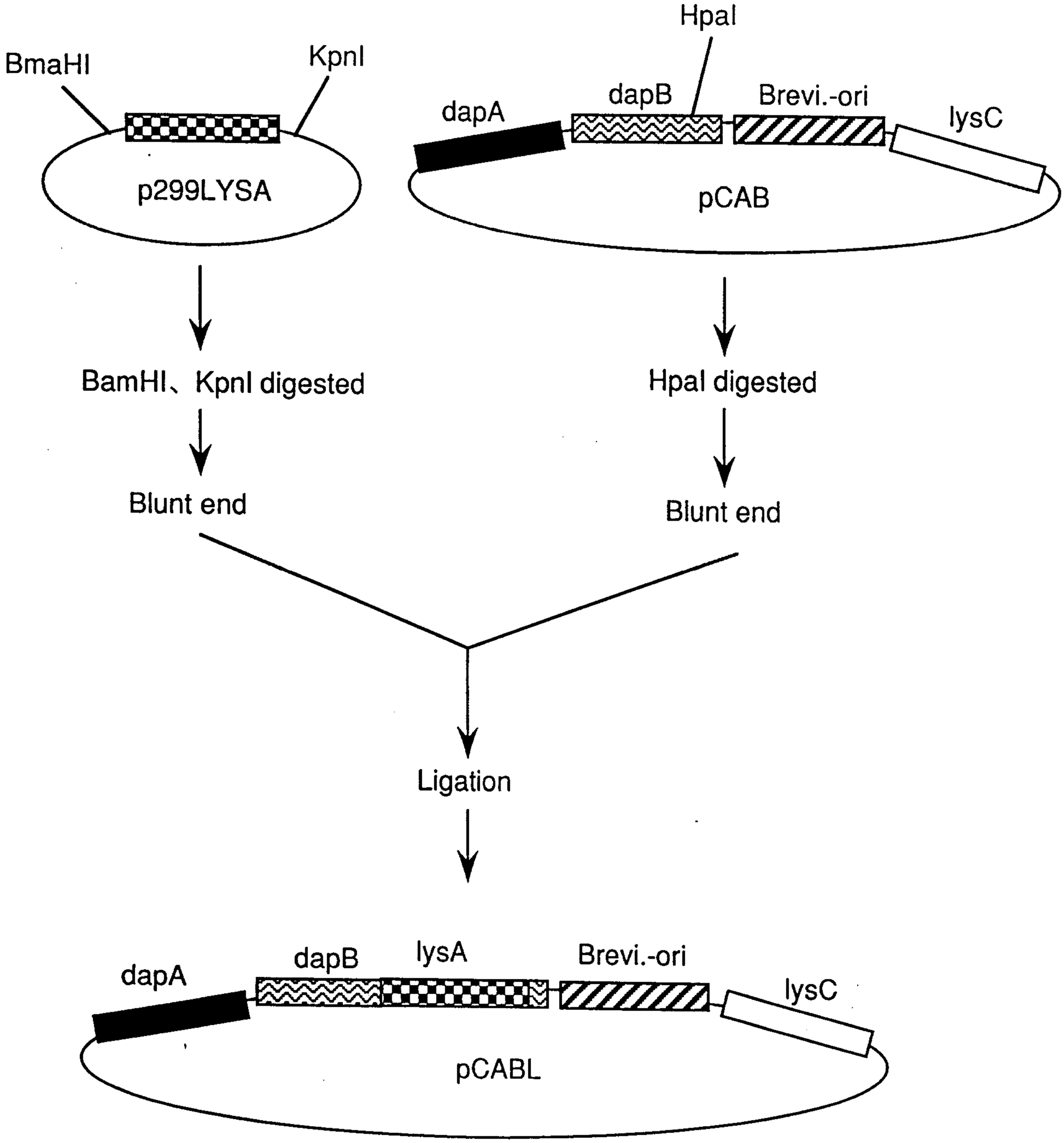
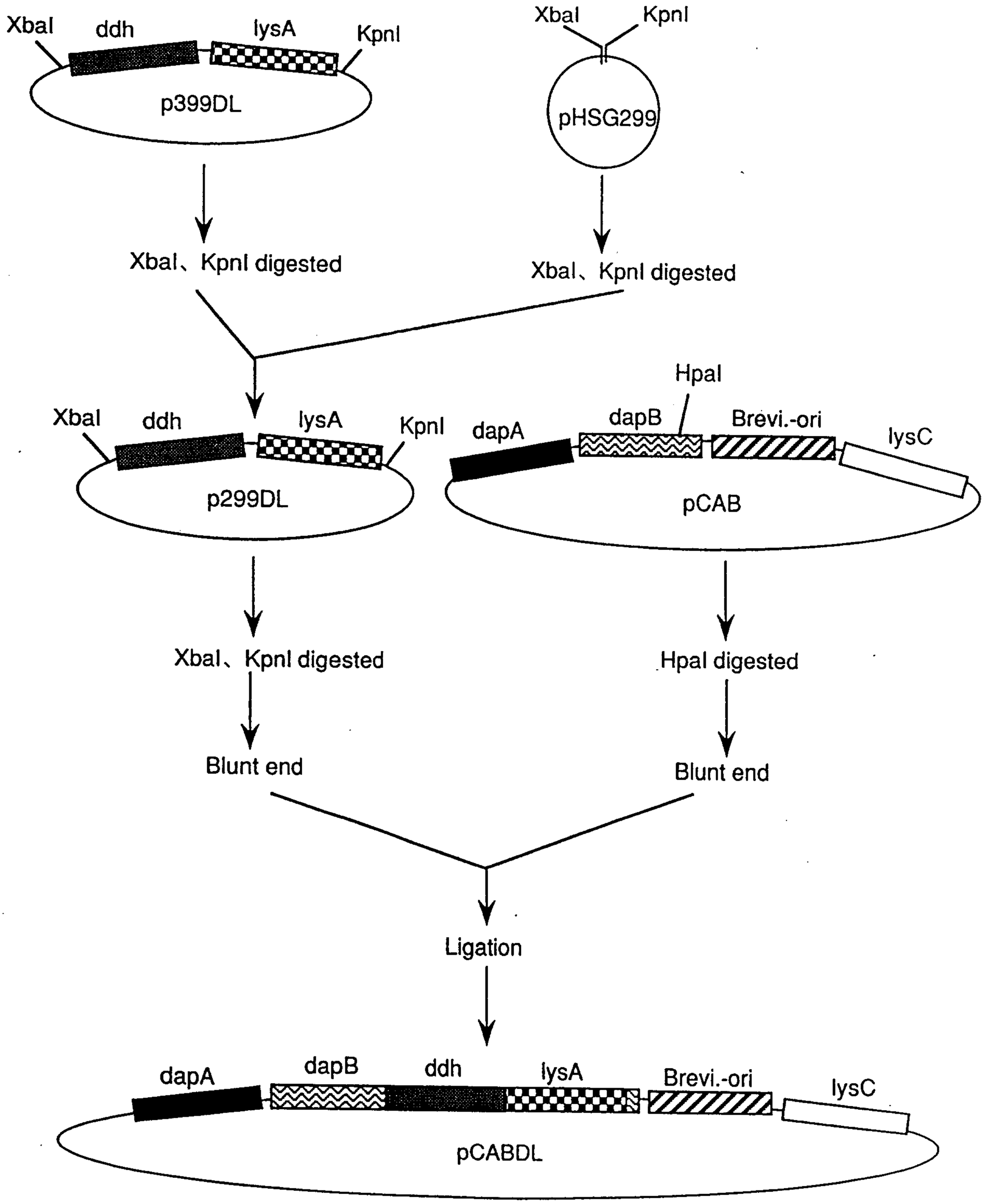

BmaHI
KpnI
p299LYSA
HpaI
dapA
dapB
Brevi.-ori
lysC
pCAB
BamHI、KpnI digested
HpaI digested
Blunt end
Blunt end
Ligation
dapA
dapB
lysA
Brevi.-ori
lysC
pCABL

FIG. 14


XbaI
ddh
lysA
KpnI
p399DL
XbaI、KpnI digested
XbaI
KpnI
pHSG299
XbaI、KpnI digested
XbaI
ddh
lysA
KpnI
p299DL
HpaI
dapA
dapB
Brevi.-ori
lysC
pCAB
XbaI、KpnI digested
HpaI digested
Blunt end
Blunt end
Ligation
dapA
dapB
ddh
lysA
Brevi.-ori
lysC
pCABDL